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- (54) **MODULATION OF CD40 EXPRESSION**
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- (57) **ABSTRACT**

Disclosed herein are antisense compounds and methods for decreasing CD40. Examples of disease conditions that can be ameliorated with the administration of antisense compounds targeted to CD40 include hyperproliferative disorders, graft versus host disease (GVHD), graft rejection, asthma, airway hyperresponsiveness, chronic obstructive pulmonary disease (COPD), multiple sclerosis (MS), systemic lupus erythematosus (SLE), and certain forms of arthritis.

22 Claims, No Drawings**Specification includes a Sequence Listing.**

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MODULATION OF CD40 EXPRESSION

Matter enclosed in heavy brackets [] appears in the original patent but forms no part of this reissue specification; matter printed in italics indicates the additions made by reissue; a claim printed with strikethrough indicates that the claim was canceled, disclaimed, or held invalid by a prior post-patent action or proceeding.

RELATED APPLICATIONS

This application is a 35 U.S.C §371 national phase application of international application serial no. PCT/US2008/012998, filed on Nov. 20, 2008, which is a non-provisional of and claims priority to U.S. patent application Ser. No. 60/989421, filed on Nov. 20, 2007, the disclosure of each of which is incorporated herein by reference in its entirety.

SEQUENCE LISTING

The present application is being filed along with a Sequence Listing in electronic format. The Sequence Listing is provided as a file entitled 33841-513SEQLIST.txt, created Nov. 19, 2008, which is 67 Kb in size. The information in the electronic format of the sequence listing is incorporated herein by reference in its entirety. This sequence listing is identical to the sequence listing filed on Nov. 20, 2007, with the exception of the addition of SEQ ID NO: 237.

FIELD OF THE INVENTION

The present invention provides methods and compositions for lowering levels of CD40 in an animal. Such methods and compositions are useful as anti-inflammatory compounds and anti-tumor compounds.

BACKGROUND OF THE INVENTION

The immune system serves a vital role in protecting the body against infectious agents. It is well established, however, that a number of disease states and/or disorders are a result of either abnormal or undesirable activation of immune responses. Common examples include graft versus host disease (GVHD) and graft rejection, and autoimmune linked diseases such as multiple sclerosis (MS), systemic lupus erythematosus (SLE), and certain forms of arthritis.

In general, an immune response is activated as a result of either tissue injury or infection. Both cases involve the recruitment and activation of a number of immune system effector cells (e.g., B- and T-lymphocytes, macrophages, eosinophils, neutrophils) in a process coordinated through a series of complex cell-cell interactions. A typical scenario by which an immune response is mounted against a foreign protein is as follows: foreign proteins captured by antigen presenting cells (APC's) such as macrophages or dendritic cells are processed and displayed on the cell surface of the APC. Circulating T-helper cells which express an immunoglobulin that recognizes (i.e. binds) the displayed antigen undergo activation by the APC. These activated T-helpers in turn activate appropriate B-cell clones to proliferate and differentiate into plasma cells that produce and secrete humoral antibodies targeted against the foreign antigen. The secreted humoral antibodies are free to circulate and bind to any cells expressing the foreign protein on their cell surface, in effect marking the cell for destruction by other immune

effector cells. In each of the stages described above, direct cell-cell contact between the involved cell types is required in order for activation to occur. (Gruss et al., Leuk. Lymphoma 1989, 24:393). In recent years, a number of cell surface receptors that mediate these cell-cell contact dependent activation events have been identified. Among these cell surface receptors is CD40 and its physiological ligand, CD40 Ligand (CD40L) which is also known as CD154.

CD40 was first characterized as a receptor expressed on B-lymphocytes. It was later found that engagement of B-cell CD40 with CD40L expressed on activated T-cells is essential for T-cell dependent B-cell activation (i.e. proliferation, immunoglobulin secretion, and class switching). It was subsequently revealed that functional CD40 is expressed on a variety of cell types other than B-cells, including macrophages, dendritic cells, thymic epithelial cells, Langerhans cells, and endothelial cells. These studies have led to the current belief that CD40 plays a broad role in immune regulation by mediating interactions of T-cells with B-cells as well as other cell types. In support of this notion, it has been shown that stimulation of CD40 in macrophages and dendritic results is required for T-cell activation during antigen presentation. (Gruss et al., Leuk. Lymphoma, 1997, 24:393). Recent evidence points to a role for CD40 in tissue inflammation as well. Production of the inflammatory mediators IL-12 and nitric oxide by macrophages have been shown to be CD40 dependent. (Buhlmann and Noelle, J. Clin. Immunol., 1996, 16:83). In endothelial cells, stimulation of CD40 by CD40L has been found to induce surface expression of E-selectin, ICAM-1, and VCAM-1, promoting adhesion of leukocytes to sites of inflammation (Buhlmann and Noelle, J. Clin. Immunol., 1996, 16:83); Gruss et al., Leuk. Lymphoma, 1997, 24:393). Finally, a number of reports have documented overexpression of CD40 in epithelial and hematopoietic tumors as well as tumor infiltrating endothelial cells, indicating that CD40 may play a role in tumor growth and/or angiogenesis as well (Gruss et al., Leuk. Lymphoma, 1997, 24:393; Kluth et al., Cancer Res., 1997, 57:891).

Due to the pivotal role that CD40 plays in humoral immunity, the potential exists that therapeutic strategies aimed at downregulating CD40 or interfering with CD40 signaling may provide a novel class of agents useful in treating a number of immune associated disorders, including but not limited to graft-versus-host disease (GVHD), graft rejection, and autoimmune diseases such as multiple sclerosis (MS), systemic lupus erythematosus (SLE), and certain forms of arthritis. Inhibitors of CD40 may also prove useful as anti-inflammatory compounds, and could therefore be useful as treatment for a variety of inflammatory and allergic conditions such as asthma, rheumatoid arthritis, allograft rejections, inflammatory bowel disease, autoimmune encephalomyelitis, thyroiditis, various dermatological conditions, and psoriasis. Recently, both CD40 and CD154 have been shown to be expressed on vascular endothelial cells, vascular smooth muscle cells and macrophages present in atherosclerotic plaques, suggesting that inflammation and immunity contribute to the atherogenic process. That this process involves CD40 signaling is suggested by several studies in mouse models in which disruption of CD154 (by knockout or by monoclonal antibody) reduced the progression or size of atherosclerotic lesions. (Mach et al., Nature, 1998, 394:200-3; Lutgens et al., 1999, Nat. Med. 5:1313-6).

Finally, as more is learned of the association between CD40 overexpression and tumor growth, inhibitors of CD40 may prove useful as anti-tumor agents and inhibitors of other hyperproliferative conditions as well.

Currently, there are no known therapeutic agents which effectively inhibit the synthesis of CD40. To date, strategies aimed at inhibiting CD40 function have involved the use of a variety of agents that disrupt CD40/CD40L binding. These include monoclonal antibodies directed against either CD40 or CD40L, soluble forms of CD40, and synthetic peptides derived from a second CD40 binding protein, A20. The use of neutralizing antibodies against CD40 and/or CD40L in animal models has provided evidence that inhibition of CD40 signaling would have therapeutic benefit for GVHD, allograft rejection, rheumatoid arthritis, SLE, MS, and B-cell lymphoma. (Buhlmann and Noelle, *J. Clin. Immunol.* 1996, 16:83). Clinical investigations were initiated using anti-CD154 monoclonal antibody in patients with lupus nephritis. However, studies were terminated due to the development of thrombotic events. (Boumpas et al., 2003, *Arthritis Rheum.* 2003, 48:719-27).

Due to the problems associated with the use of large proteins as therapeutic agents, there is a long-felt need for additional agents capable of effectively inhibiting CD40 function. Antisense oligonucleotides avoid many of the pitfalls of current agents used to block CD40/CD40L interactions and may therefore prove to be uniquely useful in a number of therapeutic, diagnostic and research applications. U.S. Pat. No. 6,197,584 (Bennett and Cowser) discloses antisense compounds targeted to CD40.

SUMMARY OF THE INVENTION

Provided herein are antisense compounds, compositions, and methods for the treatment and prevention of inflammatory conditions and cancer.

Antisense compounds described herein may be 12 to 30 nucleobases in length targeted to a CD40 nucleic acid. In certain embodiments, the CD40 nucleic acid may be any of the sequences as set forth in GENBANK® Accession No. X60592.1, incorporated herein as SEQ ID NO: 1; GENBANK® Accession No. H50598.1, incorporated herein as SEQ ID NO: 2; GENBANK® Accession No. AA203290.1, incorporated herein as SEQ ID NO: 3; and nucleotides 9797000 to nucleotide 9813000 of GENBANK Accession No. NT_011362.9, incorporated herein as SEQ ID NO: 4, or GENBANK® Accession No. BC064518.1, incorporated herein as SEQ ID NO: 237.

The antisense compound may be 12 to 30 nucleobases in length and may have a nucleobase sequence comprising at least 8 contiguous nucleobases complementary to an equal length portion of an intron region of the CD40 gene, selected from the following regions of SEQ ID NO: 4:

- (a) positions 11250-12685, corresponding to intron 6;
- (b) positions 2943-6367, corresponding to intron 1,
- (c) positions 6447-6780, corresponding to intron 2,
- (d) positions 6907-7157, corresponding to intron 3,
- (e) positions 7305-7673, corresponding to intron 4,
- (f) positions 7768-11187, corresponding to intron 5,
- (g) positions 12773-12877, corresponding to intron 7, or
- (h) positions 12907-13429, corresponding to intron 8,

wherein the remaining part or parts of the antisense compound are at least 70% complementary to the sequence shown in SEQ ID NO: 4. Preferably, the remaining parts of the antisense compound are at least 75%, 80%, 85%, 90%, 95%, 98%, 99%, or, most preferably, 100% complementary to the sequence shown in SEQ ID NO: 4.

Preferably, the antisense compound may comprise at least 8 contiguous nucleobases complementary to an equal length portion of positions 12527 to 12685 of SEQ ID NO: 4, which is a region that can be either part of intron 6, or can be part

of an alternative version of exon 7 when a different splice acceptor site is selected. Preferably, the antisense compound has a nucleobase sequence comprising at least 8 contiguous nucleobases of the nucleobase sequence of SEQ ID NO: 208, wherein the nucleobase sequence of the compound is at least 70% complementary to the sequence shown in SEQ ID NO: 4. Preferably, the antisense compound is at least 75%, 80%, 85%, 90%, 95%, 98%, 99%, or, most preferably, 100% complementary to the sequence shown in SEQ ID NO: 4. More preferably, the antisense compound has the sequence of SEQ ID NO: 208. Even more preferably, the antisense compound is 20 nucleobases in length and consists of the nucleobase sequence of SEQ ID NO: 208. Most preferably, the antisense compound is an antisense oligonucleotide 20 nucleotides in length having the sequence of nucleotides as set forth in SEQ ID NO:208, wherein each cytosine is a 5-methylcytosine, each internucleoside linkage is a phosphorothioate linkage, nucleotides 1-5 and 16-20 are 2'-O-methoxyethyl nucleotides, and nucleotides 6-15 are 2'-deoxynucleotides; most preferably the antisense compound is ISIS 396236.

In an alternative embodiment, the antisense compound may be 12 to 30 nucleobases in length and have a nucleobase sequence comprising at least 8 contiguous nucleobases complementary to an equal length portion of a region of the CD40 gene, corresponding to positions 13662-16001 of SEQ ID NO: 4, which forms part of exon 9 or a region 3' to exon 9, wherein the remaining parts of the antisense compound are at least 70% complementary to the sequence shown in SEQ ID NO: 4. Preferably, the target region of the CD40 gene corresponds to positions 13877-14084, even more preferably to positions 13937-13996, of SEQ ID NO: 4. Preferably, the remaining parts of the antisense compound are at least 75%, 80%, 85%, 90%, 95%, 98%, 99%, or, most preferably, 100% complementary to the sequence shown in SEQ ID NO: 4.

In yet another alternative embodiment, the antisense compound is 12 to 30 nucleobases in length and has a nucleobase sequence complementary to the sequence shown in SEQ ID NO: 1, starting at position 69 or 70 of SEQ ID NO: 1, wherein the nucleobase sequence is at least 95% complementary to the sequence shown in SEQ ID NO: 1. Preferably, the nucleobase sequence is essentially complementary to the sequence shown in SEQ ID NO: 1. More preferably, the nucleobase sequence is selected from the sequences of SEQ ID Nos: 90 and 163. Even more preferably, the antisense compound has a nucleobase sequence of SEQ ID NO: 90. Even more preferably, the antisense compound is 18 or 20 nucleobases in length and consists of the nucleobase sequence of SEQ ID NO: 90 or SEQ ID NO: 163. The antisense compound may be ISIS26163, ISIS396201 or ISIS396278. Preferably, the antisense compound is an antisense oligonucleotide 18 nucleotides in length having the sequence of nucleotides as set forth in SEQ ID NO: 90, wherein each cytosine is a 5-methylcytosine, each internucleoside linkage is a phosphorothioate linkage, nucleotides 1-4 and 15-18 are 2'-O-methoxyethyl nucleotides, and nucleotides 5 to 14 are 2'-deoxynucleotides. Most preferably, the antisense compound is ISIS26163.

An antisense compound according to the invention may comprise a modified oligonucleotide consisting of 12 to 30 linked nucleosides and having a nucleobase sequence comprising at least 12 contiguous nucleobases of a nucleobase sequence selected from among the nucleobase sequences recited in SEQ ID NOs: 5 to 236. Preferably, the compound consists of a single-stranded modified oligonucleotide. Preferably, the nucleobase sequence of the modified oligonucle-

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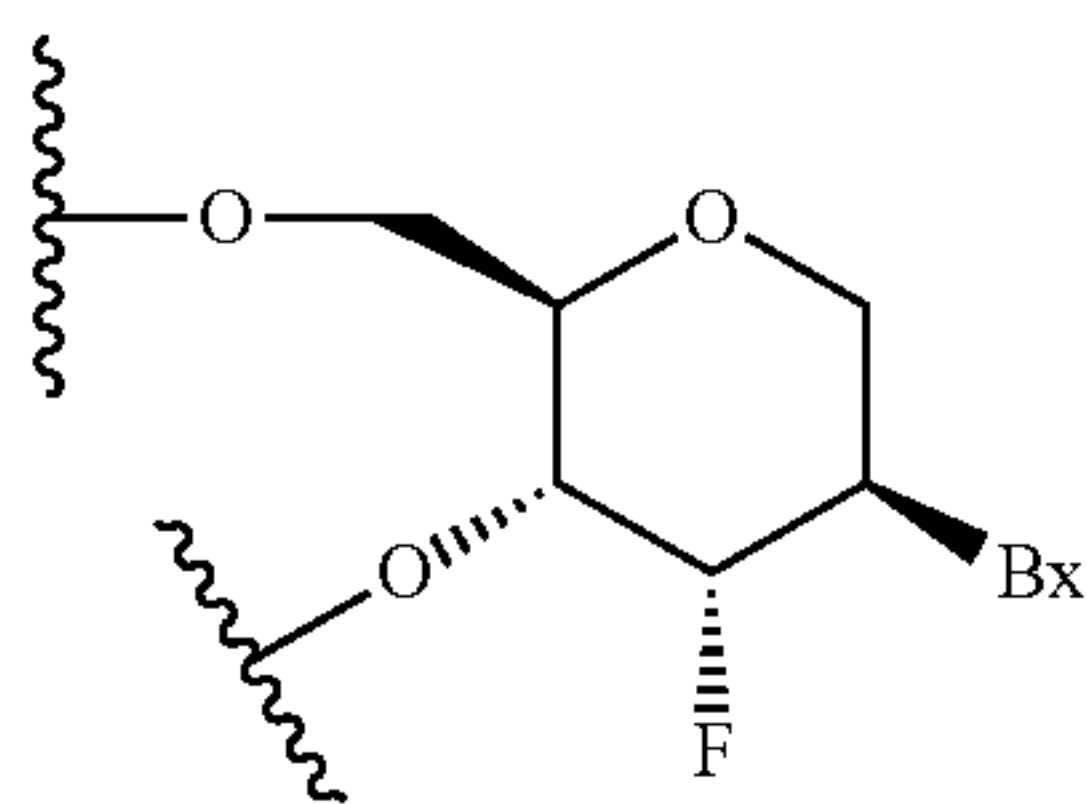
otide is 100% complementary to a nucleobase sequence of SEQ ID NO: 1, SEQ ID NO: 2, SEQ ID NO: 3, SEQ ID NO: 4, or SEQ ID NO: 237.

The antisense compound may comprise linked nucleosides. Preferably, the antisense compound is an antisense oligonucleotide.

The antisense compound may be a single-stranded or double-stranded oligonucleotide. Preferably, the antisense compound is a single-stranded oligonucleotide.

The antisense oligonucleotide may be modified, wherein at least one internucleoside linkage is a modified internucleoside linkage. The internucleoside linkage may be a phosphorothioate internucleoside linkage.

The antisense oligonucleotide may be modified, wherein at least one nucleoside comprises a modified sugar. The modified sugar may be a bicyclic sugar. Preferably, the at least one bicyclic sugar comprises a 4'-CH(CH₃)—O-2' bridge. The modified sugar may comprise a 2'-O-methoxyethyl. The antisense compound may comprise at least one tetrahydropyran modified nucleoside, wherein a tetrahydropyran ring replaces the furanose ring. Preferably, each of the at least one tetrahydropyran modified nucleoside has the structure



wherein Bx is an optionally protected heterocyclic base moiety.

The antisense compound may comprise a modified nucleobase. The modified nucleobase may be a 5-methylcytosine. Preferably, every cytosine is a 5-methylcytosine.

The antisense compound may be a gapmer, for example an oligonucleotide comprising:

- a gap segment consisting of linked deoxynucleosides;
- a 5' wing segment consisting of linked nucleosides;
- a 3' wing segment consisting of linked nucleosides;

wherein the gap segment is positioned between the 5' wing segment and the 3' wing segment and wherein each nucleoside of each wing segment comprises a modified sugar. Preferably, each nucleoside of each wing segment comprises a 2'-O-methoxyethyl sugar; and preferably each internucleoside linkage is a phosphorothioate linkage.

The antisense oligonucleotide may be a 5-10-5 MOE gapmer or a 2-15-3 MOE gapmer. The antisense oligonucleotide may consist of 20 linked nucleosides.

The antisense oligonucleotide may be a 4-10-4 MOE gapmer. The antisense oligonucleotide may consist of 18 linked nucleosides.

Compositions described herein may comprise an oligonucleotide consisting of 12 to 30 linked nucleosides, targeted to a CD40 nucleic acid or a salt thereof and a pharmaceutically acceptable carrier or diluent.

The composition may comprise a single-stranded or double-stranded oligonucleotide.

Another embodiment of the invention is a pharmaceutical composition comprising an antisense compound as described above and a liposome or a lipid based delivery system. Preferably, said liposome is an amphoteric liposome. Preferably, said amphoteric liposome is formed from

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a lipid phase comprising an amphoteric lipid or a mixture of lipid components with amphoteric properties. Said amphoteric liposome may further comprise one or more neutral or zwitterionic lipids. More preferably, said amphoteric liposome is formed from a lipid phase comprising

(a) about 15 mol % POPC, about 45 mol % DOPE, about 20 mol % MoChol, about 20 mol % Chems

(b) about 60 mol % POPC, about 10 mol % DOTAP, about 30 mol % Chems

(c) about 30 mol % POPC, about 10 mol % DOTAP, about 20 mol % Chems, about 40 mol % Chol

(d) about 60 mol % POPC, about 20 mol % HistChol, about 20 mol % Chol.

A further embodiment of the invention is an antisense compound or composition as described above for medical use. Yet a further embodiment of the invention is an antisense compound or a composition as described above for the treatment of cancer or an inflammatory or immune associated condition. The treatment may further comprise administering a second drug, which may be administered separately or concomitantly with the antisense compound of the invention.

Methods described herein may comprise administering to an animal an antisense compound as described above, preferably an antisense compound comprising an oligonucleotide consisting of 12 to 30 linked nucleosides targeted to a CD40 nucleic acid, or a composition comprising said antisense compound. Preferably, the animal is a human.

Administration of the antisense compound and/or the second drug may be by parenteral administration, topical administration, oral administration or aerosol administration. Parenteral administration may be any of subcutaneous or intravenous administration.

DETAILED DESCRIPTION OF THE INVENTION

It is to be understood that both the foregoing general description and the following detailed description are exemplary and explanatory only and are not restrictive of the invention, as claimed. Herein, the use of the singular includes the plural unless specifically stated otherwise. As used herein, the use of "or" means "and/or" unless stated otherwise. Furthermore, the use of the term "including" as well as other forms, such as "includes" and "included", is not limiting. Also, terms such as "element" or "component" encompass both elements and components comprising one unit and elements and components that comprise more than one subunit, unless specifically stated otherwise.

The section headings used herein are for organizational purposes only and are not to be construed as limiting the subject matter described. All documents, or portions of documents, cited in this application, including, but not limited to, patents, patent applications, articles, books, and treatises, are hereby expressly incorporated by reference in their entirety for any purpose.

Definitions

Unless specific definitions are provided, the nomenclature utilized in connection with, and the procedures and techniques of, analytical chemistry, synthetic organic chemistry, and medicinal and pharmaceutical chemistry described herein are those well known and commonly used in the art. Standard techniques may be used for chemical synthesis, and chemical analysis. Where permitted, all patents, applications, published applications and other publications, GENBANK Accession Numbers and associated sequence information obtainable through databases such as National

Center for Biotechnology Information (NCBI) and other data referred to throughout in the disclosure herein are incorporated by reference in their entirety.

Unless otherwise indicated, the following terms have the following meanings:

“2'-O-methoxyethyl” (also 2'-MOE and 2'-O(CH₂)₂—OCH₃) refers to an O-methoxy-ethyl modification of the 2' position of a furosyl ring. A 2'-O-methoxyethyl modified sugar is a modified sugar.

“2'-O-methoxyethyl nucleotide” means a nucleotide comprising a 2'-O-methoxyethyl modified sugar moiety.

“5-methylcytosine” means a cytosine modified with a methyl group attached to the 5' position. A 5-methylcytosine is a modified nucleobase.

“Acceptable safety profile” means a pattern of side effects that is within clinically acceptable limits.

“Active pharmaceutical ingredient” means the substance or substances in a pharmaceutical composition that provides a desired effect.

“Active target region” means a target region to which one or more active antisense compounds is targeted. “Active antisense compounds” means antisense compounds that reduce target nucleic acid levels.

“Administered concomitantly” refers to the co-administration of two agents in any manner in which the pharmacological effects of both are manifest in the patient at the same time. Concomitant administration does not require that both agents be administered in a single pharmaceutical composition, in the same dosage form, or by the same route of administration.

“Administering” means providing a pharmaceutical agent to an individual, and includes, but is not limited to administering by a medical professional and self-administering.

“Antisense compound” means an oligomeric compound that is capable of undergoing hybridization to a target nucleic acid through hydrogen bonding.

“Antisense inhibition” means reduction of target nucleic acid levels in the presence of an antisense compound complementary to a target nucleic acid compared to target nucleic acid levels in the absence of the antisense compound.

“Antisense oligonucleotide” means a single-stranded oligonucleotide having a nucleobase sequence that permits hybridization to a corresponding region or segment of a target nucleic acid.

“Bicyclic sugar” means a furosyl ring modified by the bridging of two non-geminal ring atoms. A bicyclic sugar is a modified sugar.

“Bicyclic nucleic acid” or “BNA” or “bicyclic nucleoside” or “bicyclic nucleotide” refers to a nucleoside or nucleotide wherein the furanose portion of the nucleoside includes a bridge connecting two carbon atoms on the furanose ring, thereby forming a bicyclic ring system. As used herein, unless otherwise indicated, the term “methyleneoxy BNA” alone refers to β-D-methyleneoxy BNA.

“Cap structure” or “terminal cap moiety” means chemical modifications, which have been incorporated at either terminus of an antisense compound.

“Chimeric antisense compounds” means antisense compounds that have at least 2 chemically distinct regions, each position having a plurality of subunits. A “gapmer” means an antisense compound in which an internal position having a plurality of nucleotides that supports RNaseH cleavage is positioned between external regions having one or more nucleotides that are chemically distinct from the nucleosides of the internal region. A “gap segment” means the plurality

of nucleotides that make up the internal region of a gapmer. A “wing segment” means the external region of a gapmer.

“Co-administration” means administration of two or more pharmaceutical agents to an individual. The two or more pharmaceutical agents may be in a single pharmaceutical composition, or may be in separate pharmaceutical compositions. Each of the two or more pharmaceutical agents may be administered through the same or different routes of administration. Co-administration encompasses administration in parallel or sequentially.

“Complementarity” means the capacity for pairing between nucleobases of a first nucleic acid and a second nucleic acid.

“Comply” means the adherence with a recommended therapy by a individual.

“Contiguous nucleobases” means nucleobases immediately adjacent to each other.

“Diluent” means an ingredient in a composition that lacks pharmacological activity, but is pharmaceutically necessary or desirable. For example, in drugs that are injected the diluent may be a liquid, e.g. saline solution.

“Dose” means a specified quantity of a pharmaceutical agent provided in a single administration, or in a specified time period. In certain embodiments, a dose may be administered in two or more boluses, tablets, or injections. For example, in certain embodiments, where subcutaneous administration is desired, the desired dose requires a volume not easily accommodated by a single injection. In such embodiments, two or more injections may be used to achieve the desired dose. In certain embodiments, a dose may be administered in two or more injections to minimize injection site reaction in a individual. In other embodiments, the pharmaceutical agent is administered by infusion over an extended period of time or continuously. Doses may be stated as the amount of pharmaceutical agent per hour, day, week or month.

“Dosage unit” means a form in which a pharmaceutical agent is provided, e.g. pill, tablet, or other dosage unit known in the art. In certain embodiments, a dosage unit is a vial containing lyophilized antisense oligonucleotide. In certain embodiments, a dosage unit is a vial containing reconstituted antisense oligonucleotide.

“Duration” means the period of time during which an activity or event continues. In certain embodiments, the duration of treatment is the period of time during which doses of a pharmaceutical agent are administered.

“Efficacy” means the ability to produce a desired effect.

“CD40 nucleic acid” means any nucleic acid encoding CD40. For example, in certain embodiments, a CD40 nucleic acid includes, without limitation, a DNA sequence encoding CD40, an RNA sequence transcribed from DNA encoding CD40, and an mRNA sequence encoding CD40. “CD40 mRNA” means an mRNA encoding a CD40 protein.

“Fully complementary” means each nucleobase of a first nucleic acid has a complementary nucleobase in a second nucleic acid. In certain embodiments, a first nucleic acid is an antisense compound and a target nucleic acid is a second nucleic acid. In certain such embodiments, an antisense oligonucleotide is a first nucleic acid and a target nucleic acid is a second nucleic acid.

“Gap-widened” means an antisense compound has a gap segment of 12 or more contiguous 2'-deoxyribonucleotides positioned between and immediately adjacent to 5' and 3' wing segments having from one to six nucleotides having modified sugar moieties. “Immediately adjacent” means there are no intervening nucleotides between the immediately adjacent elements.

“Hybridization” means the annealing of complementary nucleic acid molecules. In certain embodiments, complementary nucleic acid molecules include, but are not limited to, an antisense compound and a nucleic acid target. In certain such embodiments, complementary nucleic acid molecules include, but are not limited to, an antisense oligonucleotide and a nucleic acid target

“Individual” means a human or non-human animal selected for treatment or therapy.

“Individual compliance” means adherence to a recommended or prescribed therapy by a individual.

“Injection site reaction” means inflammation or abnormal redness of skin at a site of injection in a individual.

“Internucleoside linkage” refers to the chemical bond between nucleosides.

“Linked nucleosides” means adjacent nucleosides which are bonded together.

“Modified internucleoside linkage” refers to a substitution and/or any change from a naturally occurring internucleoside bond (i.e. a phosphodiester internucleoside bond).

“Modified oligonucleotide” means an oligonucleotide comprising a modified internucleoside linkage, a modified sugar, and/or a modified nucleobase.

“Modified sugar” refers to a substitution and/or any change from a natural sugar.

“Modified nucleobase” means any nucleobase other than adenine, cytosine, guanine, thymidine, or uracil. An “unmodified nucleobase” means the purine bases adenine (A) and guanine (G), and the pyrimidine bases thymine (T), cytosine (C) and uracil (U).

“Modified nucleotide” means a nucleotide having, independently, a modified sugar moiety, modified internucleoside linkage, or modified nucleobase. A “modified nucleoside” means a nucleotide having, independently, a modified sugar moiety or modified nucleobase.

“Modified sugar moiety” means a sugar moiety having any substitution and/or change from a natural sugar moiety.

“Motif” means the pattern of unmodified and modified nucleosides in an antisense compound.

“Naturally occurring internucleoside linkage” means a 3' to 5' phosphodiester linkage.

“Natural sugar moiety” means a sugar moiety found in DNA (2'-H) or RNA (2'-OH).

“Non-complementary nucleobase” or “mismatch” means a nucleobase of a first nucleic acid that is not capable of pairing with the corresponding nucleobase of a second or target nucleic acid.

“Nucleoside” means a nucleobase linked to a sugar.

As used herein the term “nucleoside mimetic” is intended to include those structures used to replace the sugar or the sugar and the base and not necessarily the linkage at one or more positions of an oligomeric compound such as for example nucleoside mimetics having morpholino, cyclohexenyl, cyclohexyl, tetrahydropyranyl, bicyclo or tricyclo sugar mimetics e.g. non furanose sugar units.

“Nucleobase” means a heterocyclic moiety capable of pairing with a base of another nucleic acid.

“Nucleobase sequence” means the order of contiguous nucleobases independent of any sugar, linkage, and/or nucleobase modification.

“Nucleotide” means a nucleoside having a phosphate group covalently linked to the sugar portion of the nucleoside.

The term “nucleotide mimetic” is intended to include those structures used to replace the nucleoside and the linkage at one or more positions of an oligomeric compound

such as for example peptide nucleic acids or morpholinos (morpholinos linked by —N(H)—C(=O)—O— or other non-phosphodiester linkage).

“Oligomeric compound” means a polymer or oligomer of linked monomeric subunits which is capable of hybridizing to at least a region of a nucleic acid molecule.

“Oligonucleotide” means an oligonucleotide in which the internucleoside linkages do not contain a phosphorus atom.

“Oligonucleotide” means a polymer or oligomer of linked nucleosides each of which can be modified or unmodified, independent one from another.

“Parenteral administration,” means administration through injection or infusion. Parenteral administration includes, but is not limited to, subcutaneous administration, intravenous administration, or intramuscular administration.

“Pharmaceutical agent” means a substance that provides a therapeutic benefit when administered to a individual. For example, in certain embodiments, an antisense oligonucleotide targeted to CD40 is pharmaceutical agent.

“Pharmaceutically acceptable salts” means physiologically and pharmaceutically acceptable salts of antisense compounds, i.e., salts that retain the desired biological activity of the parent oligonucleotide and do not impart undesired toxicological effects thereto.

“Pharmaceutical composition” means a mixture of substances suitable for administering to an individual. For example, a pharmaceutical composition may comprise one or more antisense oligonucleotides or a combination of antisense oligonucleotides and non-antisense active agents and a sterile aqueous solution or other pharmaceutically acceptable additive.

“Phosphorothioate linkage” means a linkage between nucleosides where the phosphodiester bond is modified by replacing one of the non-bridging oxygen atoms with a sulfur atom. A phosphorothioate linkage is a modified internucleoside linkage.

“Portion” means a defined number of contiguous (i.e. linked) nucleobases of a nucleic acid. In certain embodiments, a portion is a defined number of contiguous nucleobases of a target nucleic acid. In certain embodiments, a portion is a defined number of contiguous nucleobases of an antisense compound.

“Prodrug” means a therapeutic agent that is prepared in an inactive form that is converted to an active form (i.e., drug) within the body or cells thereof by the action of endogenous enzymes or other chemicals and/or conditions.

“Recommended therapy” means a therapeutic regimen recommended by a medical professional for the treatment, amelioration, or prevention of a disease.

“Side effects” means physiological responses attributable to a treatment other than desired effects. In certain embodiments, side effects include, without limitation, injection site reactions, liver function test abnormalities, renal function abnormalities, liver toxicity, renal toxicity, central nervous system abnormalities, and myopathies. For example, increased aminotransferase levels in serum may indicate liver toxicity or liver function abnormality. For example, increased bilirubin may indicate liver toxicity or liver function abnormality.

“Single-stranded modified oligonucleotide” means a modified oligonucleotide which is not hybridized to a complementary strand.

“Specifically hybridizable” means an antisense compound that hybridizes to a target nucleic acid to induce a desired effect, while exhibiting minimal or no effects on non-target nucleic acids.

The term “sugar surrogate” overlaps with the slightly broader term “nucleoside mimetic” but is intended to indicate replacement of the sugar unit (furanose ring) only. The tetrahydropyranyl rings provided herein are illustrative of an example of a sugar surrogate wherein the furanose sugar group has been replaced with a tetrahydropyranyl ring system.

“Stringent hybridization conditions” means conditions under which a nucleic acid molecule, such as an antisense compound, will hybridize to a target nucleic acid sequence, but to a minimal number of other sequences. Stringent conditions are sequence-dependent and will vary in different circumstances. In the context of this invention, “stringent conditions” under which oligomeric compounds hybridize to a target sequence are determined by the nature and composition of the oligomeric compounds and the assays in which they are being investigated.

“Subcutaneous administration” means administration just below the skin. “Intravenous administration” means administration into a vein.

“Targeted” or “targeted to” means having a nucleobase sequence that will allow specific hybridization of an antisense compound to a target nucleic acid to induce a desired effect. In certain embodiments, a desired effect is reduction of a target nucleic acid. In certain such embodiments, a desired effect is reduction of a CD40 mRNA.

“Targeting” means the process of design and selection of an antisense compound that will specifically hybridize to a target nucleic acid and induce a desired effect.

“Target nucleic acid,” “target RNA,” “target RNA transcript” and “nucleic acid target” all mean a nucleic acid capable of being targeted by antisense compounds. Target nucleic acids may include, but are not limited to, DNA, RNA (including, but not limited to pre-mRNA and mRNA or portions thereof) transcribed from DNA encoding a target, and also miRNA.

“Target region” means a portion of a target nucleic acid to which one or more antisense compounds is targeted.

“Target segment” means the sequence of nucleotides of a target nucleic acid to which an antisense compound is targeted. “5' target site” refers to the 5'-most nucleotide of a target segment. “3' target site” refers to the 3'-most nucleotide of a target segment.

“Therapeutically effective amount” means an amount of a pharmaceutical agent that provides a therapeutic benefit to an individual.

“Unmodified nucleotide” means a nucleotide composed of naturally occurring nucleobases, sugar moieties and internucleoside linkages. In certain embodiments, an unmodified nucleotide is an RNA nucleotide (i.e., β -D-ribonucleosides) or a DNA nucleotide (i.e., β -D-deoxyribonucleoside).

Antisense Compounds

Antisense compounds include, but are not limited to, oligonucleotides, oligonucleosides, oligonucleotide analogs, oligonucleotide mimetics, antisense oligonucleotides, and siRNAs. An oligomeric compound may be “antisense” to a target nucleic acid, meaning that is capable of undergoing hybridization to a target nucleic acid through hydrogen bonding.

In certain embodiments, an antisense compound has a nucleobase sequence that, when written in the 5' to 3' direction, comprises the reverse complement of the target segment of a target nucleic acid to which it is targeted. In certain such embodiments, an antisense oligonucleotide has a nucleobase sequence that, when written in the 5' to 3' direction, comprises the reverse complement of the target segment of a target nucleic acid to which it is targeted.

In certain embodiments, an antisense compound targeted to a CD40 nucleic acid is 12 to 30 subunits in length. In other words, antisense compounds are from 12 to 30 linked subunits. In other embodiments, the antisense compound is 8 to 80, 12 to 50, 15 to 30, 18 to 24, 19 to 22, or 20 linked subunits. In certain such embodiments, the antisense compounds are 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, or 80 linked subunits in length, or a range defined by any two of the above values. In some embodiments the antisense compound is an antisense oligonucleotide, and the linked subunits are nucleotides.

In certain embodiments, a shortened or truncated antisense compound targeted to a CD40 nucleic acid has a single subunit deleted from the 5' end (5' truncation), or alternatively from the 3' end (3' truncation). A shortened or truncated antisense compound targeted to a CD40 nucleic acid may have two subunits deleted from the 5' end, or alternatively may have two subunits deleted from the 3' end, of the antisense compound. Alternatively, the deleted nucleosides may be dispersed throughout the antisense compound, for example, in an antisense compound having one nucleoside deleted from the 5' end and one nucleoside deleted from the 3' end.

When a single additional subunit is present in a lengthened antisense compound, the additional subunit may be located at the 5' or 3' end of the antisense compound. When two or more additional subunits are present, the added subunits may be adjacent to each other, for example, in an antisense compound having two subunits added to the 5' end (5' addition), or alternatively to the 3' end (3' addition), of the antisense compound. Alternatively, the added subunits may be dispersed throughout the antisense compound, for example, in an antisense compound having one subunit added to the 5' end and one subunit added to the 3' end.

It is possible to increase or decrease the length of an antisense compound, such as an antisense oligonucleotide, and/or introduce mismatch bases without eliminating activity. For example, in Woolf et al. (Proc. Natl. Acad. Sci. USA 89:7305-7309, 1992), a series of antisense oligonucleotides 13-25 nucleobases in length were tested for their ability to induce cleavage of a target RNA in an oocyte injection model. Antisense oligonucleotides 25 nucleobases in length with 8 or 11 mismatch bases near the ends of the antisense oligonucleotides were able to direct specific cleavage of the target mRNA, albeit to a lesser extent than the antisense oligonucleotides that contained no mismatches. Similarly, target specific cleavage was achieved using 13 nucleobase antisense oligonucleotides, including those with 1 or 3 mismatches.

Gautschi et al (J. Natl. Cancer Inst. 93:463-471, March 2001) demonstrated the ability of an oligonucleotide having 100% complementarity to the bcl-2 mRNA and having 3 mismatches to the bcl-xL mRNA to reduce the expression of both bcl-2 and bcl-xL in vitro and in vivo. Furthermore, this oligonucleotide demonstrated potent anti-tumor activity in vivo.

Maher and Dolnick (Nuc. Acid. Res. 16:3341-3358, 1988) tested a series of tandem 14 nucleobase antisense oligonucleotides, and a 28 and 42 nucleobase antisense oligonucleotides comprised of the sequence of two or three of the tandem antisense oligonucleotides, respectively, for their ability to arrest translation of human DHFR in a rabbit reticulocyte assay. Each of the three 14 nucleobase antisense

oligonucleotides alone was able to inhibit translation, albeit at a more modest level than the 28 or 42 nucleobase antisense oligonucleotides.

Bhanot et al. (PCT/US2007/068401) provided short antisense compounds, including compounds comprising chemically-modified high-affinity monomers 8 to 16 monomers in length. These short antisense compounds were shown to be useful for reducing target nucleic acids and/or proteins in cells, tissues, and animals with increased potency and improved therapeutic index. Short antisense compounds were effective at lower doses than previously described antisense compounds, allowing for a reduction in toxicity and cost of treatment. In addition, the described short antisense compounds have greater potential for oral dosing.

Antisense Compound Motifs

In certain embodiments, antisense compounds targeted to a CD40 nucleic acid have chemically modified subunits arranged in patterns, or motifs, to confer to the antisense compounds properties such as enhanced the inhibitory activity, increased binding affinity for a target nucleic acid, or resistance to degradation by *in vivo* nucleases.

Chimeric antisense compounds typically contain at least one region modified so as to confer increased resistance to nuclease degradation, increased cellular uptake, increased binding affinity for the target nucleic acid, and/or increased inhibitory activity. A second region of a chimeric antisense compound may optionally serve as a substrate for the cellular endonuclease RNase H, which cleaves the RNA strand of an RNA:DNA duplex.

Antisense compounds having a gapmer motif are considered chimeric antisense compounds. In a gapmer an internal region having a plurality of nucleotides that supports RNaseH cleavage is positioned between external regions having a plurality of nucleotides that are chemically distinct from the nucleosides of the internal region. In the case of an antisense oligonucleotide having a gapmer motif, the gap segment generally serves as the substrate for endonuclease cleavage, while the wing segments comprise modified nucleosides. In a preferred embodiment, the regions of a gapmer are differentiated by the types of sugar moieties comprising each distinct region. The types of sugar moieties that are used to differentiate the regions of a gapmer may in some embodiments include β -D-ribonucleosides, β -D-deoxyribonucleosides, 2'-modified nucleosides (such as 2'-modified nucleosides may include 2'-MOE, and 2'-O—CH₃, among others), and bicyclic sugar modified nucleosides (such as bicyclic sugar modified nucleosides may include those having a 4'-(CH₂)_n-O-2' bridge, where n=1 or n=2). Preferably, each distinct region comprises uniform sugar moieties. The wing-gap-wing motif is frequently described as "X-Y-Z", where "X" represents the length of the 5' wing region, "Y" represents the length of the gap region, and "Z" represents the length of the 3' wing region. Any of the antisense compounds described herein can have a gapmer motif. In some embodiments, X and Z are the same, in other embodiments they are different. In a preferred embodiment, Y is between 8 and 15 nucleotides. X, Y or Z can be any of 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 25, 30 or more nucleotides. Thus, gapmers of the present invention include, but are not limited to, for example 5-10-5, 4-8-4, 4-10-4, 2-15-3, 4-12-3, 4-12-4, 3-14-3, 2-16-2, 1-18-1, 3-10-3, 2-10-2, 1-10-1 or 2-8-2.

In some embodiments, the antisense compound as a "wingmer" motif, having a wing-gap or gap-wing configuration, i.e. an X-Y or Y-Z configuration as described above for the gapmer configuration. Thus, wingmer configurations

of the present invention include, but are not limited to, for example 5-10, 8-4, 4-12, 12-4, 3-14, 16-2, 18-1, 10-3, 2-10, 1-10 or 8-2.

In one embodiment, antisense compounds targeted to a CD40 nucleic acid possess a 5-10-5 gapmer motif.

In some embodiments, an antisense compound targeted to a CD40 nucleic acid has a gap-widened motif. In other embodiments, an antisense oligonucleotide targeted to a CD40 nucleic acid has a gap-widened motif.

In one embodiment, a gap-widened antisense oligonucleotide targeted to a CD40 nucleic acid has a gap segment of fourteen 2'-deoxyribonucleotides positioned between wing segments of three chemically modified nucleosides. In one embodiment, the chemical modification comprises a 2'-sugar modification. In another embodiment, the chemical modification comprises a 2'-MOE sugar modification.

Target Nucleic Acids, Target Regions and Nucleotide Sequences

Nucleotide sequences that encode CD40 include, without limitation, the following: GENBANK Accession No. X60592.1, first deposited with GENBANK® on Apr. 21, 1993 and incorporated herein as SEQ ID NO: 1; GENBANK® Accession No. H50598.1, first deposited with GENBANK® on Sep. 19, 1995, and incorporated herein as SEQ ID NO: 2; GENBANK Accession No. AA203290.1, first deposited with GENBANK® on Jan. 25, 1997, and incorporated herein as SEQ ID NO: 3; and nucleotides 9797000 to 9813000 of GENBANK Accession No. NT_011362.9, first deposited with GENBANK® on Nov. 29, 2000, and incorporated herein as SEQ ID NO: 4, and GENBANK® Accession No. BC064518.1, incorporated herein as SEQ ID NO: 237.

It is understood that the sequence set forth in each SEQ ID NO in the Examples contained herein is independent of any modification to a sugar moiety, an internucleoside linkage, or a nucleobase. As such, antisense compounds defined by a SEQ ID NO may comprise, independently, one or more modifications to a sugar moiety, an internucleoside linkage, or a nucleobase. Antisense compounds described by Isis Number (Isis No) indicate a combination of nucleobase sequence and motif.

In one embodiment, a target region is a structurally defined region of the nucleic acid. For example, a target region may encompass a 3' UTR, a 5' UTR, an exon, an intron, a coding region, a translation initiation region, translation termination region, or other defined nucleic acid region. The structurally defined regions for CD40 can be obtained by accession number from sequence databases such as NCBI and such information is incorporated herein by reference. In other embodiments, a target region may encompass the sequence from a 5' target site of one target segment within the target region to a 3' target site of another target segment within the target region.

Targeting includes determination of at least one target segment to which an antisense compound hybridizes, such that a desired effect occurs. In certain embodiments, the desired effect is a reduction in mRNA target nucleic acid levels. In other embodiments, the desired effect is reduction of levels of protein encoded by the target nucleic acid or a phenotypic change associated with the target nucleic acid.

A target region may contain one or more target segments. Multiple target segments within a target region may be overlapping. Alternatively, they may be non-overlapping. In one embodiment, target segments within a target region are separated by no more than about 300 nucleotides. In other embodiments, target segments within a target region are separated by no more than about, 250, 200, 150, 100, 90, 80,

70, 60, 50, 40, 30, 20, or 10 nucleotides on the target nucleic acid. In another embodiment, target segments within a target region are separated by no more than about 5 nucleotides on the target nucleic acid. In additional embodiments, target segments are contiguous.

Suitable target segments may be found within a 5' UTR, a coding region, a 3' UTR, an intron, or an exon. Target segments containing a start codon or a stop codon are also suitable target segments. A suitable target segment may specifically exclude a certain structurally defined region such as the start codon or stop codon.

The determination of suitable target segments may include a comparison of the sequence of a target nucleic acid to other sequences throughout the genome. For example, the BLAST algorithm may be used to identify regions of similarity amongst different nucleic acids. This comparison can prevent the selection of antisense compound sequences that may hybridize in a non-specific manner to sequences other than a selected target nucleic acid (i.e., non-target or off-target sequences).

There may be variation in activity (e.g., as defined by percent reduction of target nucleic acid levels) of the antisense compounds within an active target region. In one embodiment, reductions in CD40 mRNA levels are indicative of inhibition of CD40 expression. Reductions in levels of a CD40 protein are also indicative of inhibition of target mRNA expression. Further, phenotypic changes are indicative of inhibition of CD40 expression. For example, changes in cell morphology over time or treatment dose as well as changes in levels of cellular components such as proteins, lipids, nucleic acids, hormones, saccharides, or metals is indicative of inhibition of CD40 expression. Reduction of eosinophils is indicative of inhibition of CD40 expression. Measurements of cellular status which include pH, stage of cell cycle, intake or excretion of biological indicators by the cell are also endpoints of interest.

Analysis of the genotype of the cell (measurement of the expression of one or more of the genes of the cell) after treatment is also used as an indicator of the efficacy or potency of the CD40 inhibitors. Hallmark genes, or those genes suspected to be associated with a specific disease state, condition, or phenotype, are measured in both treated and untreated cells.

Genomic Structure, Exons and Introns

Although some eukaryotic mRNA transcripts are directly translated, many contain one or more regions, known as "introns," which are excised from a transcript before it is translated. The remaining (and therefore translated) regions are known as "exons" and are spliced together to form a continuous mRNA sequence. Targeting splice sites, i.e., intron-exon junctions or exon-intron junctions, may also be particularly useful in situations where aberrant splicing is implicated in disease, or where an overproduction of a particular splice product is implicated in disease. Aberrant fusion junctions due to rearrangements or deletions are also preferred target sites. mRNA transcripts produced via the process of splicing of two (or more) mRNAs from different gene sources are known as "fusion transcripts". It is also known that introns can be effectively targeted using antisense compounds targeted to, for example, DNA or pre-mRNA.

It is also known in the art that alternative RNA transcripts can be produced from the same genomic region of DNA. These alternative transcripts are generally known as "variants". More specifically, "pre-mRNA variants" are transcripts produced from the same genomic DNA that differ from other transcripts produced from the same genomic

DNA in either their start or stop position and contain both intronic and exonic sequence.

Upon excision of one or more exon or intron regions, or portions thereof during splicing, pre-mRNA variants produce smaller "mRNA variants". Consequently, mRNA variants are processed pre-mRNA variants and each unique pre-mRNA variant must always produce a unique mRNA variant as a result of splicing. These mRNA variants are also known as "alternative splice variants". If no splicing of the pre-mRNA variant occurs then the pre-mRNA variant is identical to the mRNA variant.

Hybridization

In some embodiments, hybridization occurs between an antisense compound disclosed herein and a CD40 nucleic acid. The most common mechanism of hybridization involves hydrogen bonding (e.g., Watson-Crick, Hoogsteen or reversed Hoogsteen hydrogen bonding) between complementary nucleobases of the nucleic acid molecules.

Hybridization can occur under varying conditions. Stringent conditions are sequence-dependent and are determined by the nature and composition of the nucleic acid molecules to be hybridized.

Methods of determining whether a sequence is specifically hybridizable to a target nucleic acid are well known in the art. In one embodiment, the antisense compounds provided herein are specifically hybridizable with a CD40 nucleic acid.

Complementarity

An antisense compound and a target nucleic acid are complementary to each other when a sufficient number of nucleobases of the antisense compound can hydrogen bond with the corresponding nucleobases of the target nucleic acid, such that a desired effect will occur (e.g., antisense inhibition of a target nucleic acid, such as a CD40 nucleic acid).

Non-complementary nucleobases between an antisense compound and a CD40 nucleic acid may be tolerated provided that the antisense compound remains able to specifically hybridize to a target nucleic acid. Moreover, an antisense compound may hybridize over one or more segments of a CD40 nucleic acid such that intervening or adjacent segments are not involved in the hybridization event (e.g., a loop structure, mismatch or hairpin structure).

In some embodiments, the antisense compounds provided herein are at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 96%, at least 97%, at least 98% or at least 99% complementary to a CD40 nucleic acid. Percent complementarity of an antisense compound with a target nucleic acid can be determined using routine methods. For example, an antisense compound in which 18 of 20 nucleobases of the antisense compound are complementary to a target region, and would therefore specifically hybridize, would represent 90 percent complementarity. In this example, the remaining noncomplementary nucleobases may be clustered or interspersed with complementary nucleobases and need not be contiguous to each other or to complementary nucleobases. As such, an antisense compound which is 18 nucleobases in length having 4 (four) noncomplementary nucleobases which are flanked by two regions of complete complementarity with the target nucleic acid would have 77.8% overall complementarity with the target nucleic acid and would thus fall within the scope of the present invention. Percent complementarity of an antisense compound with a region of a target nucleic acid can be determined routinely using BLAST programs (basic local alignment search tools) and Power-BLAST programs known in the art (Altschul et al., J. Mol.

Biol., 1990, 215, 403 410; Zhang and Madden, Genome Res., 1997, 7, 649 656). Percent homology, sequence identity or complementarity, can be determined by, for example, the Gap program (Wisconsin Sequence Analysis Package, Version 8 for Unix, Genetics Computer Group, University Research Park, Madison Wis.), using default settings, which uses the algorithm of Smith and Waterman (Adv. Appl. Math., 1981, 2, 482 489).

In other embodiments, the antisense compounds provided herein are fully complementary (i.e., 100% complementary) to a target nucleic acid. For example, an antisense compound may be fully complementary to a CD40 nucleic acid, or a target region, or a target segment or target sequence thereof. As used herein, "fully complementary" means each nucleobase of an antisense compound is capable of precise base pairing with the corresponding nucleobases of a target nucleic acid.

The location of a non-complementary nucleobase may be at the 5' end or 3' end of the antisense compound. Alternatively, the non-complementary nucleobase or nucleobases may be at an internal position of the antisense compound. When two or more non-complementary nucleobases are present, they may be contiguous (i.e. linked) or non-contiguous. In one embodiment, a non-complementary nucleobase is located in the wing segment of a gapmer antisense oligonucleotide.

In one embodiment, antisense compounds up to 20 nucleobases in length comprise no more than 4, no more than 3, no more than 2 or no more than 1 non-complementary nucleobase(s) relative to a target nucleic acid, such as a CD40 nucleic acid.

In another embodiment, antisense compounds up to 30 nucleobases in length comprise no more than 6, no more than 5, no more than 4, no more than 3, no more than 2 or no more than 1 non-complementary nucleobase(s) relative to a target nucleic acid, such as a CD40 nucleic acid.

The antisense compounds provided herein also include those which are complementary to a portion of a target nucleic acid. As used herein, "portion" refers to a defined number of contiguous (i.e. linked) nucleobases within a region or segment of a target nucleic acid. A "portion" can also refer to a defined number of contiguous nucleobases of an antisense compound. In one embodiment, the antisense compounds are complementary to at least an 8 nucleobase portion of a target segment. In another embodiment, the antisense compounds are complementary to at least a 12 nucleobase portion of a target segment. In yet another embodiment, the antisense compounds are complementary to at least a 15 nucleobase portion of a target segment. Also contemplated are antisense compounds that are complementary to at least a 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, or more nucleobase portion of a target segment, or a range defined by any two of these values.

Identity

The antisense compounds provided herein may also have a defined percent identity to a particular nucleotide sequence, SEQ ID NO, or compound represented by a specific Isis number. As used herein, an antisense compound is identical to the sequence disclosed herein if it has the same nucleobase pairing ability. For example, a RNA which contains uracil in place of thymidine in a disclosed DNA sequence would be considered identical to the DNA sequence since both uracil and thymidine pair with adenine. Shortened and lengthened versions of the antisense compounds described herein as well as compounds having non-identical bases relative to the antisense compounds provided herein also are contemplated. The non-identical

bases may be adjacent to each other or dispersed throughout the antisense compound. Percent identity of an antisense compound is calculated according to the number of bases that have identical base pairing relative to the sequence to which it is being compared.

In one embodiment, the antisense compounds are at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, 99% or 100% identical to one or more of the antisense compounds or SEQ ID NOs, or a portion thereof, disclosed herein.

Modifications

A nucleoside is a base-sugar combination. The nucleobase (also known as base) portion of the nucleoside is normally a heterocyclic base moiety. Nucleotides are nucleosides that further include a phosphate group covalently linked to the sugar portion of the nucleoside. For those nucleosides that include a pentofuranosyl sugar, the phosphate group can be linked to the 2', 3' or 5' hydroxyl moiety of the sugar. Oligonucleotides are formed through the covalent linkage of adjacent nucleosides to one another, to form a linear polymeric oligonucleotide. Within the oligonucleotide structure, the phosphate groups are commonly referred to as forming the internucleoside linkages of the oligonucleotide.

Modifications to antisense compounds encompass substitutions or changes to internucleoside linkages, sugar moieties, or nucleobases. Modified antisense compounds are often preferred over native forms because of desirable properties such as, for example, enhanced cellular uptake, enhanced affinity for nucleic acid target, increased stability in the presence of nucleases, or increased inhibitory activity.

Chemically modified nucleosides may also be employed to increase the binding affinity of a shortened or truncated antisense oligonucleotide for its target nucleic acid. Consequently, comparable results can often be obtained with shorter antisense compounds that have such chemically modified nucleosides.

Modified Internucleoside Linkages

The naturally occurring internucleoside linkage of RNA and DNA is a 3' to 5' phosphodiester linkage. Antisense compounds having one or more modified, i.e. non-naturally occurring, internucleoside linkages are often selected over antisense compounds having naturally occurring internucleoside linkages because of desirable properties such as, for example, enhanced cellular uptake, enhanced affinity for target nucleic acids, and increased stability in the presence of nucleases.

Oligonucleotides having modified internucleoside linkages include internucleoside linkages that retain a phosphorus atom as well as internucleoside linkages that do not have a phosphorus atom. Representative phosphorus containing internucleoside linkages include, but are not limited to, phosphodiesters, phosphotriesters, methylphosphonates, phosphonoacetates, phosphoramidate, and phosphorothioates or phosphorodithioates. Internucleoside linkages that do not have a phosphorus atom include, amongst others, methylene(methylimino) or MMI linkages, morpholino linkages or amide linkages. In peptide nucleic acids (PNA) the sugar backbone is replaced with an amide containing backbone.

Methods of preparation of phosphorous-containing and non-phosphorous-containing linkages are well known.

Representative United States patents that teach the preparation of phosphorus-containing linkages include, but are not limited to, U.S. Pat. Nos. 3,687,808; 4,469,863; 4,476,301; 5,023,243; 5,177,196; 5,188,897; 5,264,423; 5,276,019; 5,278,302; 5,286,717; 5,321,131; 5,399,676; 5,405,939; 5,453,496; 5,455,233; 5,466,677; 5,476,925; 5,519,126; 5,536,821; 5,541,306; 5,550,111; 5,563,253; 5,571,799; 5,587,361; 5,194,599; 5,565,555; 5,527,899; 5,721,

218; 5,672,697; 5,625,050 and U.S. Pat. No. 6,693,187, each of which is herein incorporated by reference.

Representative United States patents that teach the preparation of non-phosphorous-containing linkages include, but are not limited to, U.S. Pat. Nos. 5,034,506; 5,166,315; 5,185,444; 5,214,134; 5,216,141; 5,235,033; 5,264,562; 5,264,564; 5,405,938; 5,434,257; 5,466,677; 5,470,967; 5,489,677; 5,541,307; 5,561,225; 5,596,086; 5,602,240; 5,610,289; 5,602,240; 5,608,046; 5,610,289; 5,618,704; 5,623,070; 5,663,312; 5,633,360; 5,677,437; 5,792,608; 5,646,269 and 5,677,439, each of which is herein incorporated by reference.

In one embodiment, antisense compounds targeted to a CD40 nucleic acid comprise one or more modified internucleoside linkages. In some embodiments, the modified internucleoside linkages are phosphorothioate linkages. In other embodiments, each internucleoside linkage of an antisense compound is a phosphorothioate internucleoside linkage.

Modified Sugar Moieties

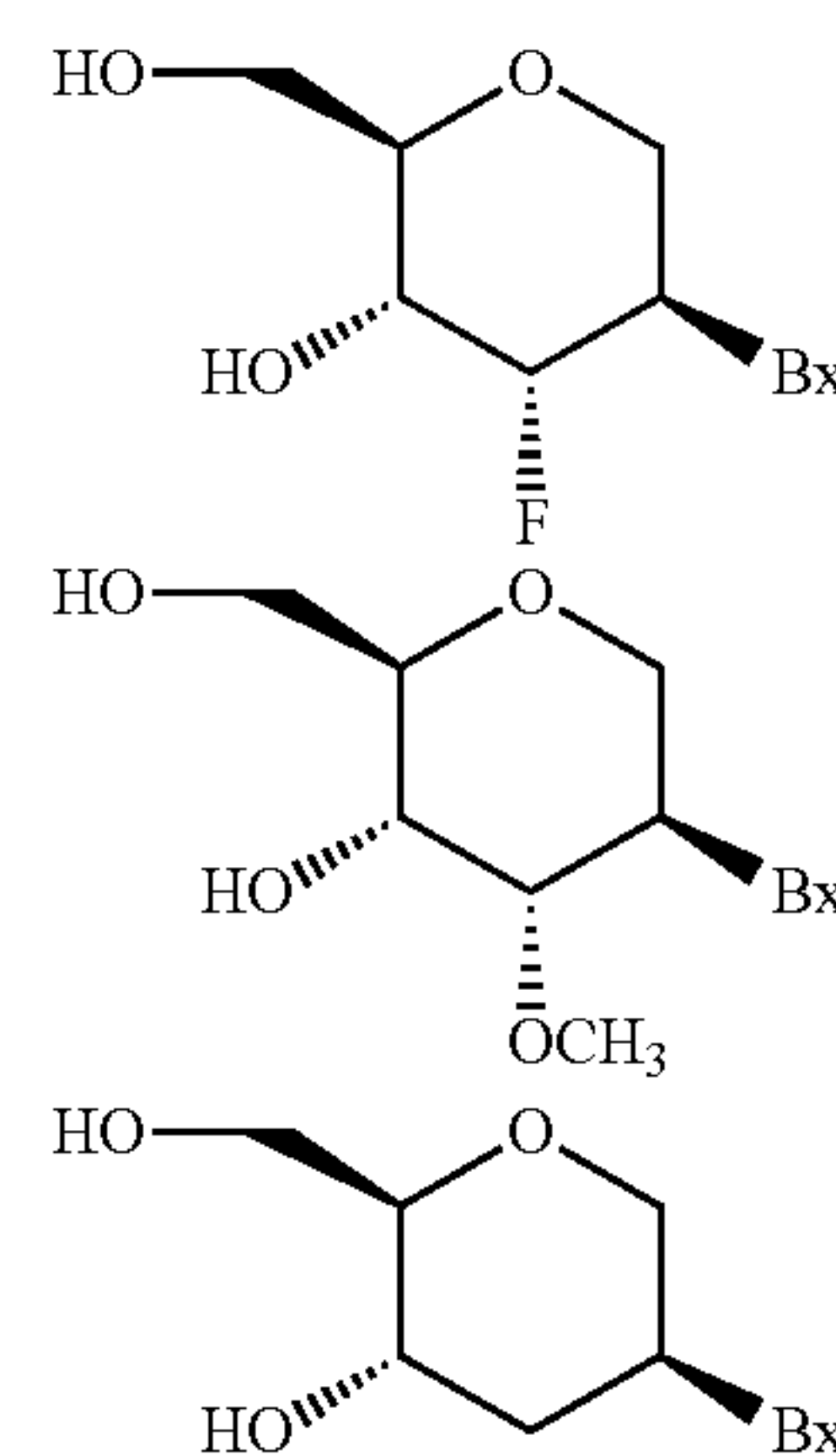
Antisense compounds of the invention can optionally contain one or more nucleosides wherein the sugar group has been modified. Such sugar modified nucleosides may impart enhanced nuclease stability, increased binding affinity or some other beneficial biological property to the antisense compounds. In certain embodiments, nucleosides are modified by modification of the ribofuranose ring. Such modifications include without limitation, addition of substituent groups, bridging of non-geminal ring atoms to form a bicyclic nucleic acid (BNA), as in locked nucleic acids (LNA), replacement of the ribosyl ring oxygen atom with S, N(R), or C(R1)(R)2 (R=H, C1-C12 alkyl or a protecting group) and combinations thereof. Examples of chemically modified sugars include 2'-F-5'-methyl substituted nucleoside (see PCT International Application WO 2008/101157 published on Aug. 21, 2008 for other disclosed 5',2'-bis substituted nucleosides) or replacement of the ribosyl ring oxygen atom with S with further substitution at the 2'-position (see published U.S. Patent Application US2005-0130923, published on Jun. 16, 2005) or alternatively 5'-substitution of a BNA (see PCT International Application WO 2007/134181 Published on Nov. 22, 2007 wherein LNA is substituted with for example a 5'-methyl or a 5'-vinyl group).

Examples of nucleosides having modified sugar moieties include without limitation nucleosides comprising 5'-vinyl, 5'-methyl (R or S), 4'-S, 2'-F, 2'-OCH₃ (known as 2'-OMe) and 2'-O(CH₂)₂OCH₃ (known as 2'MOE) substituent groups. The substituent at the 2' position can also be selected from allyl, amino, azido, thio, O-allyl, O-C1-C10 alkyl, OCF₃, O-CH₂CH₂CH₂NH₂, O(CH₂)₂SCH₃, O(CH₂)₂-O-N(Rm)(Rn) such as 2'-dimethylaminoxyethoxy (2'-O-(CH₂)₂ON(CH₃)₂ or 2'-DMAOE), O(CH₂)₂-O-(CH₂)₂-N(Rm)(Rn) such as 2'-dimethylaminoethoxyethoxy (2'-O-(CH₂)₂-O-(CH₂)₂-N(CH₃)₂ or 2'-DMAEOE and O-CH₂-C(=O)-N(Rm)(Rn), where each Rm and Rn is, independently, H or substituted or unsubstituted C1-C10 alkyl.

Examples of bicyclic nucleic acids (BNAs) include without limitation nucleosides comprising a bridge between the 4' and the 2' ribosyl ring atoms, e.g. a 4'-(CH₂)_n-O-2' bridge, where n=1 or n=2. In certain embodiments, antisense compounds provided herein include one or more BNA nucleosides wherein the bridge comprises one of the formulas: 4'-(CH₂)—O-2' (LNA); 4'-(CH₂)—S-2'; 4'-(CH₂)—O-2' (LNA); 4'-(CH₂)₂—O-2' (ENA); 4'-C(CH₃)₂—O-2' (see PCT/US2008/068922); 4'-CH(CH₃)—O-2' and 4'CH

(CH₂OCH₃)—O-2' (see U.S. Pat. No. 7,399,845, issued on Jul. 15, 2008); 4'-CH₂—N(OCH₃)-2' (see PCT/US2008/064591); 4'-CH₂—O—N(CH₃)-2' (see published U.S. Patent Application US2004-0171570, published Sep. 2, 2004); 4'-CH₂—N(R)—O-2' (see U.S. Pat. No. 7,427,672, issued on Sep. 23, 2008); 4'-CH₂—C(CH₃)-2' and 4'-CH₂—C(=CH₂)-2' (see PCT/US2008/066154); and wherein R is, independently, H, C1-C12 alkyl, or a protecting group. Each of the foregoing BNAs include various stereochemical sugar configurations including for example α-L-ribofuranose and β-D-ribofuranose (See PCT international application PCT/DK98/00393, published on Mar. 25, 1999 as WO99/14226).

In certain embodiments, nucleosides are modified by replacement of the ribosyl ring with a sugar surrogate. Such modification includes without limitation, replacement of the ribosyl ring with a surrogate ring system (sometimes referred to as DNA analogs) such as a morpholino ring, a cyclohexenyl ring, a cyclohexyl ring or a tetrahydropyran ring such as one having one of the formula:



wherein Bx is an optionally protected heterocyclic base moiety.

Many other bicyclo and tricyclo sugar surrogate ring systems are also known in the art that can be used to modify nucleosides for incorporation into antisense compounds (see for example Leumann, C. J., *Bioorg. Med. Chem.* 10, (2002), 841-854). Such ring systems can undergo various additional substitutions to enhance activity.

Methods for the preparations of modified sugars are well known to those skilled in the art. Representative United States patents that teach the preparation of modified sugars include, but are not limited to U.S. Pat. No. 4,981,957; 5,118,800; 5,319,080; 5,359,044; 5,393,878; 5,446,137; 5,466,786; 5,514,785; 5,519,134; 5,567,811; 5,576,427; 5,591,722; 5,597,909; 5,610,300; 5,627,053; 5,639,873; 5,646,265; 5,658,873; 5,670,633; and 5,700,920, each of which is herein incorporated by reference.

In certain embodiments, a 2'-modified nucleoside has a bicyclic sugar moiety. In certain such embodiments, the bicyclic sugar moiety is a D sugar in the alpha configuration. In certain such embodiments, the bicyclic sugar moiety is a D sugar in the beta configuration. In certain such embodiments, the bicyclic sugar moiety is an L sugar in the alpha configuration. In certain such embodiments, the bicyclic sugar moiety is an L sugar in the beta configuration.

In certain embodiments, the bicyclic sugar moiety comprises a bridge group between the 2' and the 4'-carbon atoms. In certain such embodiments, the bridge group comprises from 1 to 8 linked biradical groups. In certain embodiments,

the bicyclic sugar moiety comprises from 1 to 4 linked biradical groups. In certain embodiments, the bicyclic sugar moiety comprises 2 or 3 linked biradical groups. In certain embodiments, the bicyclic sugar moiety comprises 2 linked biradical groups. In certain embodiments, a linked biradical group is selected from —O—, —S—, —N(R1)-, —C(R1)(R2)-, —C(R1)=C(R1)-, —C(R1)=N—, —C(=NR1)-, —Si(R1)(R2)-, —S(=O)₂—, —S(=O)—, —C(=O)— and —C(=S)—; where each R1 and R2 is, independently, H, hydroxyl, C1-C12 alkyl, substituted C1-C12 alkyl, C2-C12 alkenyl, substituted C2-C12 alkenyl, C2-C12 alkynyl, substituted C2-C12 alkynyl, C5-C20 aryl, substituted C5-C20 aryl, a heterocycle radical, a substituted heterocycle radical, heteroaryl, substituted heteroaryl, C5-C7 alicyclic radical, substituted C5-C7 alicyclic radical, halogen, substituted oxy (—O—), amino, substituted amino, azido, carboxyl, substituted carboxyl, acyl, substituted acyl, CN, thiol, substituted thiol, sulfonyl (S(=O)₂—H), substituted sulfonyl, sulfoxyl (S(=O)—H) or substituted sulfoxyl; and each substituent group is, independently, halogen, C1-C12 alkyl, substituted C1-C12 alkyl, C2-C12 alkenyl, substituted C2-C12 alkenyl, C2-C12 alkynyl, substituted C2-C12 alkynyl, amino, substituted amino, acyl, substituted acyl, C1-C12 aminoalkyl, C1-C12 aminoalkoxy, substituted C1-C12 aminoalkyl, substituted C1-C12 aminoalkoxy or a protecting group.

In some embodiments, the bicyclic sugar moiety is bridged between the 2' and 4' carbon atoms with a biradical group selected from —O—(CH₂)_p—, —O—CH₂—, —O—CH₂CH₂—, —O—CH(alkyl)—, —NH—(CH₂)_p—, —N(alkyl)—(CH₂)_p—, —O—CH(alkyl)—, —(CH(alkyl))(CH₂)_p—, —NH—O—(CH₂)_p—, —N(alkyl)—O—(CH₂)_p—, or —O—N(alkyl)—(CH₂)_p—, wherein p is 1, 2, 3, 4 or 5 and each alkyl group can be further substituted. In certain embodiments, p is 1, 2 or 3.

In one aspect, each of said bridges is, independently, —[C(R1)(R2)]_n—, —[C(R1)(R2)]_n—O—, —C(R1R2)—N(R1)—O— or —C(R1R2)—O—N(R1)—. In another aspect, each of said bridges is, independently, 4'-(CH₂)₃-2',4'-(CH₂)₂-2',4'-CH₂—O-2',4'-(CH₂)₂—O-2',4'-CH₂—O—N(R1)-2' and 4'-CH₂—N(R1)-O-2'— wherein each R1 is, independently, H, a protecting group or C1-C12 alkyl.

In nucleotides having modified sugar moieties, the nucleobase moieties (natural, modified or a combination thereof) are maintained for hybridization with an appropriate nucleic acid target.

In one embodiment, antisense compounds targeted to a CD40 nucleic acid comprise one or more nucleotides having modified sugar moieties. In a preferred embodiment, the modified sugar moiety is 2'-MOE. In other embodiments, the 2'-MOE modified nucleotides are arranged in a gapmer motif.

Modified Nucleobases

Nucleobase (or base) modifications or substitutions are structurally distinguishable from, yet functionally interchangeable with, naturally occurring or synthetic unmodified nucleobases. Both natural and modified nucleobases are capable of participating in hydrogen bonding. Such nucleobase modifications may impart nuclease stability, binding affinity or some other beneficial biological property to antisense compounds. Modified nucleobases include synthetic and natural nucleobases such as, for example, 5-methylcytosine (5-me-C). Certain nucleobase substitutions, including 5-methylcytosine substitutions, are particularly useful for increasing the binding affinity of an antisense compound for a target nucleic acid. For example, 5-methylcytosine substitutions have been shown to increase nucleic

acid duplex stability by 0.6-1.2° C. (Sanghvi, Y. S., Crooke, S. T. and Lebleu, B., eds., *Antisense Research and Applications*, CRC Press, Boca Raton, 1993, pp. 276-278).

Additional unmodified nucleobases include 5-hydroxymethyl cytosine, xanthine, hypoxanthine, 2-aminoadenine, 6-methyl and other alkyl derivatives of adenine and guanine, 2-propyl and other alkyl derivatives of adenine and guanine, 2-thiouracil, 2-thiothymine and 2-thiocytosine, 5-halouracil and cytosine, 5-propynyl (—C≡C—CH₃) uracil and cytosine and other alkynyl derivatives of pyrimidine bases, 6-azouracil, cytosine and thymine, 5-uracil (pseudouracil), 4-thiouracil, 8-halo, 8-amino, 8-thiol, 8-thioalkyl, 8-hydroxyl and other 8-substituted adenines and guanines, 5-halo particularly 5-bromo, 5-trifluoromethyl and other 5-substituted uracils and cytosines, 7-methylguanine and 7-methyladenine, 2-F-adenine, 2-amino-adenine, 8-azaguanine and 8-azaadenine, 7-deazaguanine and 7-deazaadenine and 3-deazaguanine and 3-deazaadenine.

Heterocyclic base moieties may also include those in which the purine or pyrimidine base is replaced with other heterocycles, for example 7-deaza-adenine, 7-deazaguanosine, 2-aminopyridine and 2-pyridone. Nucleobases that are particularly useful for increasing the binding affinity of antisense compounds include 5-substituted pyrimidines, 6-azapyrimidines and N-2, N-6 and O-6 substituted purines, including 2 aminopropyladenine, 5-propynyluracil and 5-propynylcytosine.

Methods for the preparations of modified nucleobases are well known to those skilled in the art. Representative United States patents that teach the preparation of modified nucleobases include, but are not limited to U.S. Pat. No. 3,687, 808, as well as U.S. Pat. Nos. 4,845,205; 5,130,302; 5,134, 066; 5,175,273; 5,367,066; 5,432,272; 5,457,187; 5,459, 255; 5,484,908; 5,502,177; 5,525,711; 5,552,540; 5,587, 469; 5,594,121, 5,596,091; 5,614,617; 5,645,985; 5,830, 653; 5,763,588; 6,005,096; 5,750,692 and 5,681,941 each of which is herein incorporated by reference.

In one embodiment, antisense compounds targeted to a CD40 nucleic acid comprise one or more modified nucleobases. In an additional embodiment, gap-widened antisense oligonucleotides targeted to a CD40 nucleic acid comprise one or more modified nucleobases. In some embodiments, the modified nucleobase is 5-methylcytosine. In further embodiments, each cytosine is a 5-methylcytosine.

Conjugated Antisense Compounds

Antisense compounds may be covalently linked to one or more moieties or conjugates which enhance the activity, cellular distribution or cellular uptake of the resulting antisense oligonucleotides. Typical conjugate groups include cholesterol moieties and lipid moieties. Additional conjugate groups include carbohydrates, phospholipids, biotin, phenazine, folate, phenanthridine, anthraquinone, acridine, fluoresceins, rhodamines, coumarins, and dyes.

Antisense compounds can also be modified to have one or more stabilizing groups that are generally attached to one or both termini of antisense compounds to enhance properties such as, for example, nuclease stability. Included in stabilizing groups are cap structures. These terminal modifications protect the antisense compound having terminal nucleic acid from exonuclease degradation, and can help in delivery and/or localization within a cell. The cap can be present at the 5'-terminus (5'-cap), or at the 3'-terminus (3'-cap), or can be present on both termini. Cap structures are well known in the art and include, for example, inverted deoxy abasic caps. Further 3' and 5'-stabilizing groups that can be used to cap one or both ends of an antisense

compound to impart nuclease stability include those disclosed in WO 03/004602 published on Jan. 16, 2003.

Compositions and Methods for Formulating Pharmaceutical Compositions

Antisense oligonucleotides may be admixed with pharmaceutically acceptable active and/or inert substances for the preparation of pharmaceutical compositions or formulations. Compositions and methods for the formulation of pharmaceutical compositions are dependent upon a number of criteria, including, but not limited to, route of administration, extent of disease, or dose to be administered.

Antisense compound targeted to a CD40 nucleic acid can be utilized in pharmaceutical compositions by combining the antisense compound with a suitable pharmaceutically acceptable diluent or carrier. A pharmaceutically acceptable diluent includes for example phosphate-buffered saline (PBS). PBS is a diluent suitable for use in compositions to be delivered parenterally. Accordingly, in one embodiment, employed in the methods described herein is a pharmaceutical composition comprising an antisense compound targeted to a CD40 nucleic acid and a pharmaceutically acceptable diluent. In one embodiment, the pharmaceutically acceptable diluent is PBS. In other embodiments, the antisense compound is an antisense oligonucleotide.

Pharmaceutical compositions comprising antisense compounds encompass any pharmaceutically acceptable salts, esters, or salts of such esters, or any other oligonucleotide which, upon administration to an animal, including a human, is capable of providing (directly or indirectly) the biologically active metabolite or residue thereof. Accordingly, for example, the disclosure is also drawn to pharmaceutically acceptable salts of antisense compounds, prodrugs, pharmaceutically acceptable salts of such prodrugs, and other bioequivalents. Suitable pharmaceutically acceptable salts include, but are not limited to, sodium and potassium salts.

A prodrug can include the incorporation of additional nucleosides at one or both ends of an antisense compound which are cleaved by endogenous nucleases within the body, to form the active antisense compound.

The present invention also includes pharmaceutical compositions and formulations which include the antisense compounds of the invention. The pharmaceutical compositions of the present invention may be administered in a number of ways depending upon whether local or systemic treatment is desired and upon the area to be treated. Administration may be topical (including ophthalmic and to mucous membranes including vaginal and rectal delivery), pulmonary, e.g., by inhalation or insufflation of powders or aerosols, including by nebulizer; intratracheal, intranasal, epidermal and transdermal), oral or parenteral. Parenteral administration includes intravenous, intraarterial, subcutaneous, intraperitoneal or intramuscular injection or infusion; or intracranial, e.g., intrathecal or intraventricular, administration. Oligonucleotides with at least one 2'-O-methoxyethyl modification are believed to be particularly useful for oral administration.

Pharmaceutical compositions and formulations for topical administration may include transdermal patches, ointments, lotions, creams, gels, drops, suppositories, sprays, liquids and powders. Conventional pharmaceutical carriers, aqueous, powder or oily bases, thickeners and the like may be necessary or desirable. Coated condoms, gloves and the like may also be useful.

The pharmaceutical formulations of the present invention, which may conveniently be presented in unit dosage form, may be prepared according to conventional techniques well known in the pharmaceutical industry. Such techniques

include the step of bringing into association the active ingredients with the pharmaceutical carrier(s) or excipient(s). In general, the formulations are prepared by uniformly and intimately bringing into association the active ingredients with liquid carriers or finely divided solid carriers or both, and then, if necessary, shaping the product.

The compositions of the present invention may be formulated into any of many possible dosage forms such as, but not limited to, tablets, capsules, gel capsules, liquid syrups, soft gels, suppositories, and enemas. The compositions of the present invention may also be formulated as suspensions in aqueous, non-aqueous or mixed media. Aqueous suspensions may further contain substances which increase the viscosity of the suspension including, for example, sodium carboxymethylcellulose, sorbitol and/or dextran. The suspension may also contain stabilizers.

Pharmaceutical compositions of the present invention include, but are not limited to, solutions, emulsions, foams and liposome-containing formulations. The pharmaceutical compositions and formulations of the present invention may comprise one or more penetration enhancers, carriers, excipients or other active or inactive ingredients.

Emulsions are typically heterogenous systems of one liquid dispersed in another in the form of droplets usually exceeding 0.1 μm in diameter. Emulsions may contain additional components in addition to the dispersed phases, and the active drug which may be present as a solution in either the aqueous phase, oily phase or itself as a separate phase. Microemulsions are included as an embodiment of the present invention. Emulsions and their uses are well known in the art and are further described in U.S. Pat. No. 6,287,860, which is incorporated herein in its entirety.

Formulations of the present invention include liposomal formulations. As used in the present invention, the term "liposome" means a vesicle composed of amphiphilic lipids arranged in a spherical bilayer or bilayers. Liposomes are unilamellar or multilamellar vesicles which have a membrane formed from a lipophilic material and an aqueous interior that contains the composition to be delivered. Cationic liposomes are positively charged liposomes which are believed to interact with negatively charged DNA molecules to form a stable complex. Liposomes that are pH-sensitive or negatively-charged are believed to entrap DNA rather than complex with it. Both cationic and noncationic liposomes have been used to deliver DNA to cells.

Liposomes also include "sterically stabilized" liposomes, a term which, as used herein, refers to liposomes comprising one or more specialized lipids that, when incorporated into liposomes, result in enhanced circulation lifetimes relative to liposomes lacking such specialized lipids. Examples of sterically stabilized liposomes are those in which part of the vesicle-forming lipid portion of the liposome comprises one or more glycolipids or is derivatized with one or more hydrophilic polymers, such as a polyethylene glycol (PEG) moiety. Liposomes and their uses are further described in U.S. Pat. No. 6,287,860, which is incorporated herein in its entirety.

Other liposomes or lipid based delivery systems known in the art are described for example in WO 05/105152; WO 06/069782; Morrissey et al., *Nature Biotechnology*, 23 (8), 1002-1007, 2005; WO 05/007196; Wheeler et al., *Gene Therapy*, 6 (2), 271-281, 1999; WO 02/34236; Budker et al., *Nature Biotechnology*, 14 (6), 760-764, 1996; U.S. Pat. No. 5,965,434; U.S. Pat. No. 5,635,487; Spagnou et al., *Biochemistry*, 43 (42), 13348-13356, 2004; U.S. Pat. No. 6,756,

054; WO 06/016097 and U.S. Pat. No. 5,785,992; WO 04/035523, each of which is herein incorporated by reference.

In a preferred embodiment of the invention amphoteric liposomes may be used as formulations which include the inventive antisense compounds. Amphoteric liposomes are a class of liposomes having an anionic or neutral charge at pH 7.5 and a cationic charge at pH 4. Reference is made to WO 02/066012 by Panzner et al. which is incorporated herein by reference. The use, selection and manufacturing of amphoteric liposomes for the transfection of cells is further described in WO 05/094783 of Ended et al., WO 07/031,333 of Panzner et al., WO 07/107,304 of Panzner et al. and WO 08/043,575 of Panzner et al. Amphoteric liposomes have an excellent bio-distribution and are well tolerated in animals. They can encapsulate nucleic acid molecules with high efficiency. WO 06/048329 of Panzner et al., which is incorporated herein by reference in its entirety, describes pharmaceutical compositions comprising amphoteric liposomes and oligonucleotides which are adapted to target nucleic acids encoding CD40.

By "amphoteric" is meant herein that the liposomes comprise charged groups of both anionic and cationic character wherein:

(i) at least one of the charged groups has a pKa between 4 and 7.4,

(ii) the cationic charge prevails at pH 4 and

(iii) the anionic charge prevails at pH 7.4;

whereby the liposomes have an isoelectric point of zero net charge between pH 4 and pH 7.4. Amphoteric character, by this definition, is different from "zwitterionic character", because zwitterions do not have a pK in the range mentioned above. In consequence, zwitterions are essentially neutral over a range of pH values. Phosphatidylcholine or phosphatidylethanolamines, for example, are neutral lipids with zwitterionic character.

Amphoteric liposomes may be formed from a lipid phase comprising an amphoteric lipid. In some embodiments said lipid phase may comprise 5 to 30 mol. % of said amphoteric lipid, preferably 10 to 25 mol. %.

Suitable amphoteric lipids are disclosed in WO 02/066489 and WO 03/070735 by Panzner et al. Preferably, said amphoteric lipid is selected from the group consisting of HistChol, HistDG, isoHistSuccDG, Acylcarnosin and HCChol. (A glossary of such abbreviated forms of the names of the lipids referred to herein is included below for ease of reference. A number of such abbreviations are those that are commonly used by those skilled in the art.)

Alternatively, said amphoteric liposomes may be formed from a lipid phase comprising a mixture of lipid components with amphoteric properties. Such amphoteric liposomes may be formed from pH-responsive anionic and/or cationic components, as disclosed for example in WO 02/066012. Cationic lipids sensitive to pH are disclosed in WO 02/066489 and WO 03/070220 and in the references made therein, in particular in Budker, et al. 1996, Nat Biotechnol. 14 (6): 760-4, and can be used in combination with constitutively charged anionic lipids or with anionic lipids that are sensitive to pH.

Alternatively, the cationic charge may be introduced from constitutively charged lipids that are known to those skilled in the art in combination with a pH sensitive anionic lipid. Combinations of constitutively charged anionic and cationic lipids, e.g. DOTAP and DPPG, are not preferred. Thus, in some presently preferred embodiments of the invention, said mixture of lipid components may comprise (i) a stable cationic lipid and a chargeable anionic lipid, (ii) a chargeable

cationic lipid and chargeable anionic lipid or (iii) a stable anionic lipid and a chargeable cationic lipid.

Preferred cationic components include DMTAP, DPTAP, DOTAP, DC-Chol, MoChol, HisChol, DPIM, CHIM, DOME, DDAB, DAC-Chol, TC-Chol, DOTMA, DOGS, (C18)₂Gly⁺ N,N-dioctadecylamido-glycin, CTAB, CPyC, DODAP and DOEPC.

Preferred anionic lipids for use with the invention include DOGSucc, POGSucc, DMGSucc, DPGSucc, DMPS, DPPS, DOPS, POPS, DMPG, DPPG, DOPG, POPG, DMPA, DPPA, DOPA, POPA, CHEMS and CetylP.

Preferably, such an amphoteric mixture of lipids does not constitute more than about 70 mol. % of the lipid phase. In some embodiments, said mixture may constitute not more than 50 mol. % of the lipid phase; preferably said lipid phase comprises about 20 to about 40 mol. % of such a mixture.

In some embodiments, said lipid phase may further comprise a neutral lipid, preferably a neutral phospholipid, such as a phosphatidylcholine. Presently preferred phosphatidylcholines include POPC, natural or hydrogenated soy bean PC, natural or hydrogenated egg PC, DMPC, DPPC, DSPC and DOPC. More preferably, said phosphatidylcholine comprises POPC, non-hydrogenated soy bean PC or non-hydrogenated egg PC.

The lipid phase may comprise at least 15 mol. % of said phosphatidylcholine, preferably at least 20 mol. %. In some embodiments, said lipid phase may comprise no less than about 25 mol. % phosphatidylcholine. Alternatively, said lipid phase may comprise no less than about 40 mol. % phosphatidylcholine.

A presently preferred formulation in accordance with the present invention comprises a liposome having about 60 mol. % POPC, about 10 mol. % DOTAP and about 30 mol. % CHEMS.

In some embodiments said neutral lipid may comprise a phosphatidylethanolamine or a mixture of phosphatidylcholine and phosphatidylethanolamine. Said neutral phosphatidylcholines or phosphatidylethanolamines or mixtures of the two may be present in the lipid phase in the molar amount (mol. %) not constituted by the other components of the lipid phase, but to at least 20 mol. % (the total for the lipid phase being 100 mol. %).

Preferred phosphatidylethanolamines include DOPE, DMPE and DPPE.

In some embodiments said neutral lipid may comprise POPC and DOPE.

Advantageously, said lipid phase may comprise a mixture of anionic and cationic lipids with amphoteric properties, phosphatidylcholine and phosphatidylethanolamine. Amphoteric liposomes formed from such a lipid phase may be serum-stable and therefore suitable for systemic delivery. Preferably said lipid phase comprises MoChol as a cationic lipid and CHEMS or DMG-Succ as an anionic lipid.

Further presently preferred amphoteric liposomes for use as formulations which include antisense compounds of the present invention may be selected from the group consisting of:

(a) about 15 mol. % POPC, about 45 mol. % DOPE, about 20 mol. % MoChol and about 20 mol. % CHEMS;

(b) about 10 mol. % POPC, about 30 mol. % DOPE, about 30 mol. % MoChol and about 30 mol. % CHEMS;

(c) about 10 mol. % POPC, about 30 mol. % DOPE, about 20 mol. % MoChol and about 40 mol. % CHEMS;

(d) about 6 mol. % POPC, about 24 mol. % DOPE, about 47 mol. % MoChol and about 23 mol. % CHEMS.

Alternatively, said lipid phase may comprise a mixture of anionic and cationic lipids with amphoteric properties a

neutral phosphatidylcholine and cholesterol. Such liposomes may also be serum-stable. In some embodiments, said lipid phase may comprise from 30 mol. % to 50 mol. % cholesterol, preferably from about 35 mol. % to about 45 mol. %. Alternatively, said lipid phase may comprise phosphatidylcholine and from 10 mol. % to 25 mol. % cholesterol, preferably from about 15 mol. % to about 25 mol. %.

A presently preferred formulation comprises 10 to 25 mol. % amphoteric lipid, e.g. HistChol, HistDG or Acylcarnosin, 15 to 25 mol. % cholesterol and the remainder being POPC, soy bean PC, egg PC, DMPC, DPPC or DOPC, preferably POPC; for example about 60 mol. % POPC, about 20 mol. % HistChol and about 20 mol. % Chol.

Another presently preferred formulation in accordance with the present invention comprises a liposome including a mix of lipid components with amphoteric properties and having about 30 mol. % POPC, about 10 mol. % DOTAP, about 20 mol. % CHEMS and about 40 mol. % Chol.

The amphoteric liposomes may have a size in the range 50 to 500 nm, preferably 100 to 500 nm, more preferably 150 and 300 nm.

The amphoteric liposome formulations of the present invention may be formulated for use as a colloid in a suitable pharmacologically acceptable vehicle. Vehicles such as water, saline, phosphate buffered saline and the like are well known to those skilled in the art for this purpose.

In some embodiments, the amphoteric liposome formulations of the present invention may be administered at a physiological pH of between about 7 and about 8. To this end, the formulation comprising the antisense compound, excipient and vehicle may be formulated to have a pH in this range.

The amphoteric liposome formulations of the invention may be manufactured using suitable methods that are known to those skilled in the art. Such methods include, but are not limited to, extrusion through membranes of defined pore size, injection of lipid solutions in ethanol into a water phase containing the cargo to be encapsulated, or high pressure homogenisation.

A solution of the oligonucleotide may be contacted with said excipient at a neutral pH, thereby resulting in volume inclusion of a certain percentage of the solution. An high concentrations of the excipient, ranging from about 50 mM to about 150 mM, is preferred to achieve substantial encapsulation of the active agent.

Amphoteric liposomes used as formulations in accordance with the present invention offer the distinct advantage of binding oligonucleotides at or below their isoelectric point, thereby concentrating said active agent at the liposome surface. This process is described in more detail in WO 02/066012.

Irrespective of the actual production process used to make the amphoteric liposome formulations, in some embodiments, non-encapsulated oligonucleotide may be removed from the liposomes after the initial production step in which the liposomes are formed as tight containers. Again, the technical literature and the references included herein describe such methodology in detail and suitable process steps may include, but are not limited to, size exclusion chromatography, sedimentation, dialysis, ultrafiltration and diafiltration.

However, the removal of any non-encapsulated oligonucleotide is not required for performance of the invention, and in some embodiments the composition may comprise free as well as entrapped drug.

In some aspects of the invention the amphoteric liposome formulations which include the inventive antisense com-

pounds may be used as pharmaceutical compositions for the prevention or treatment of an inflammatory, immune or autoimmune disorder of a human or non-human animal such as graft rejection, graft-versus-host disease, multiple sclerosis, systemic lupus erythematosus, rheumatoid arthritis, asthma, inflammatory bowel disease, psoriasis or thyroiditis, Morbus Crohn and Colitis ulcerosa.

Glossary Of Common Abbreviated Lipid Names

10	DMPC Dimyristoylphosphatidylcholine
	DPPC Dipalmitoylphosphatidylcholine
	DSPC Distearoylphosphatidylcholine
	POPC Palmitoyl-oleoylphosphatidylcholine
15	DOPC Dioleoylphosphatidylcholine
	DOPE Dioleoylphosphatidylethanolamine
	DMPE Dimyristoylphosphatidylethanolamine
	DPPE Dipalmitoylphosphatidylethanolamine
	DOPG Dioleoylphosphatidylglycerol
20	POPG Palmitoyl-oleoylphosphatidylglycerol
	DMPG Dimyristoylphosphatidylglycerol
	DPPG Dipalmitoylphosphatidylglycerol
	DMPS Dimyristoylphosphatidylserine
	DPPS Dipalmitoylphosphatidylserine
25	DOPS Dioleoylphosphatidylserine
	POPS Palmitoyl-oleoylphosphatidylserine
	DMPA Dimyristoylphosphatidic acid
	DPPA Dipalmitoylphosphatidic acid
	DOPA Dioleoylphosphatidic acid
30	POPA Palmitoyl-oleoylphosphatidic acid
	CHEMS Cholesterolhemisuccinate
	DC-Chol 3-β-[N—(N',N'-dimethylethane) carbamoyl]cholesterol
	CetylP Cetylphosphate
35	DODAP (1,2-dioleoyloxypropyl)-N,N-dimethylammonium chloride
	DOEPC 1,2-dioleoyl-sn-glycero-3-ethylphosphocholine
	DAC-Chol 3-β-[N—(N',N'-dimethylethane) carbamoyl]cholesterol
40	TC-Chol 3-β-[N—(N',N',N'-trimethylaminoethane)carbamoyl]cholesterol
	DOTMA (1,2-dioleoyloxypropyl)-N,N,N-trimethylammoniumchlorid (Lipofectin®)
	DOGS ((C18) ₂ GlySper ³⁺) N,N-dioctadecylamido-glycylspermine (Transfectam®)
45	CTAB Cetyl-trimethylammoniumbromide,
	CPyC Cetyl-pyridiniumchloride
	DOTAP (1,2-dioleoyloxypropyl)-N,N,N-trimethylammonium salt
50	DMTAP (1,2-dimyristoyloxypropyl)-N,N,N-trimethylammonium salt
	DPTAP (1,2-dipalmitoyloxypropyl)-N,N,N-trimethylammonium salt
	DOTMA (1,2-dioleoyloxypropyl)-N,N,N-trimethylammonium chloride)
55	DORIE (1,2-dioleoyloxypropyl)-3 dimethylhydroxyethyl ammoniumbromide)
	DDAB Dimethyldioctadecylammonium bromide
	DPIM 4-(2,3-bis-palmitoyloxy-propyl)-1-methyl-1H-imidazole
60	CHIM Histaminy1-Cholesterolcarbamate
	MoChol 4-(2-Aminoethyl)-Morpholino-Cholesterol-hemisuccinate
	His Chol Histaminy1-Cholesterolhemisuccinate.
65	HCChol Nα-Histidiny1-Cholesterolcarbamate
	HistChol Nα-Histidiny1-Cholesterol-hemisuccinate.
	AC Acylcarnosine, Stearyl- & Palmitoylcarnosine

HistDG 1,2-Dipalmitoylglycerol-hemisuccinate-N α -Histidiny-l-hemisuccinate, & Distearoyl-, Dimyristoyl-, Dioleoyl- or palmitoyl-oleoylderivatives

IsoHistSuccDG 1,2-Dipalmitoylglycerol-O α -Histidiny-l-N α -hemisuccinat, & Distearoyl-, Dimyristoyl-, Dioleoyl- or palmitoyl-oleoylderivatives

DGSucc 1,2-Dipalmitoylglycerol-3-hemisuccinate & Distearoyl-, Dimyristoyl-Dioleoyl- or palmitoyl-oleoyl-derivatives

The pharmaceutical formulations and compositions of the present invention may also include surfactants. The use of surfactants in drug products, formulations and in emulsions is well known in the art. Surfactants and their uses are further described in U.S. Pat. No. 6,287,860, which is incorporated herein in its entirety. In one embodiment, the present invention employs various penetration enhancers to effect the efficient delivery of nucleic acids, particularly oligonucleotides. In addition to aiding the diffusion of non-lipophilic drugs across cell membranes, penetration enhancers also enhance the permeability of lipophilic drugs. Penetration enhancers may be classified as belonging to one of five broad categories, i.e., surfactants, fatty acids, bile salts, chelating agents, and non-chelating non-surfactants. Penetration enhancers and their uses are further described in U.S. Pat. No. 6,287,860, which is incorporated herein in its entirety.

One of skill in the art will recognize that formulations are routinely designed according to their intended use, i.e. route of administration.

Preferred formulations for topical administration include those in which the oligonucleotides of the invention are in admixture with a topical delivery agent such as lipids, liposomes, fatty acids, fatty acid esters, steroids, chelating agents and surfactants. Preferred lipids and liposomes include neutral (e.g. dioleoylphosphatidyl DOPE ethanolamine, dimyristoylphosphatidyl choline DMPC, distearoylphosphatidyl choline) negative (e.g. dimyristoylphosphatidyl glycerol DMPG); cationic (e.g. dioleoyltetramethylaminopropyl DOTAP and dioleoylphosphatidyl ethanolamine DOTMA) or amphoteric lipids or lipid mixtures wherein a mixture of cationic and anionic lipids displays amphoteric properties. For topical or other administration, oligonucleotides of the invention may be encapsulated within liposomes or may form complexes thereto, in particular to cationic liposomes. Alternatively, oligonucleotides may be complexed to lipids, in particular to cationic lipids. Preferred fatty acids and esters, pharmaceutically acceptable salts thereof, and their uses are further described in U.S. Pat. No. 6,287,860, which is incorporated herein in its entirety. Topical formulations are described in detail in U.S. patent application Ser. No. 09/315,298 filed on May 20, 1999, which is incorporated herein by reference in its entirety.

Compositions and formulations for oral administration include powders or granules, microparticulates, nanoparticulates, suspensions or solutions in water or non-aqueous media, capsules, gel capsules, sachets, tablets or minitabets. Thickeners, flavoring agents, diluents, emulsifiers, dispersing aids or binders may be desirable. Preferred oral formulations are those in which oligonucleotides of the invention are administered in conjunction with one or more penetration enhancers surfactants and chelators. Preferred surfactants include fatty acids and/or esters or salts thereof, bile acids and/or salts thereof. Preferred bile acids/salts and fatty acids and their uses are further described in U.S. Pat. No. 6,287,860, which is incorporated herein in its entirety. Also preferred are combinations of penetration enhancers, for

example, fatty acids/salts in combination with bile acids/salts. A particularly preferred combination is the sodium salt of lauric acid, capric acid and UDCA. Further penetration enhancers include polyoxyethylene-9-lauryl ether, polyoxyethylene-20-cetyl ether. Oligonucleotides of the invention may be delivered orally, in granular form including sprayed dried particles, or complexed to form micro or nanoparticles. Oligonucleotide complexing agents and their uses are further described in U.S. Pat. No. 6,287,860, which is incorporated herein in its entirety. Oral formulations for oligonucleotides and their preparation are described in detail in U.S. application Ser. Nos. 09/108,673 (filed Jul. 1, 1998), 09/315,298 (filed May 20, 1999) and 10/071,822, filed Feb. 8, 2002, each of which is incorporated herein by reference in their entirety.

Compositions and formulations for parenteral, intrathecal or intraventricular administration may include sterile aqueous solutions which may also contain buffers, diluents and other suitable additives such as, but not limited to, penetration enhancers, carrier compounds and other pharmaceutically acceptable carriers or excipients.

Certain embodiments of the invention provide pharmaceutical compositions containing one or more oligomeric compounds and one or more other active agents which function by a non-antisense mechanism, such as for example chemotherapeutic agents or antiinflammatory drugs. Examples of such chemotherapeutic agents include but are not limited to cancer chemotherapeutic drugs such as daunorubicin, daunomycin, dactinomycin, doxorubicin, epirubicin, idarubicin, esorubicin, bleomycin, mafosfamide, ifosfamide, cytosine arabinoside, bis-chloroethylnitrosurea, busulfan, mitomycin C, actinomycin D, mithramycin, prednisone, hydroxyprogesterone, testosterone, tamoxifen, dacarbazine, procarbazine, hexamethylmelamine, pentamethylmelamine, mitoxantrone, amsacrine, chlorambucil, methylcyclohexylnitrosurea, nitrogen mustards, melphalan, cyclophosphamide, 6-mercaptopurine, 6-thioguanine, cytarabine, 5-azacytidine, hydroxyurea, deoxycoformycin, 4-hydroxyperoxycyclophosphoramide, 5-fluorouracil (5-FU), 5-fluorodeoxyuridine (5-FUdR), methotrexate (MTX), colchicine, taxol, vincristine, vinblastine, etoposide (VP-16), trimetrexate, irinotecan, topotecan, gemcitabine, teniposide, cisplatin and diethylstilbestrol (DES). When used with the compounds of the invention, such chemotherapeutic agents may be used individually (e.g., 5-FU and oligonucleotide), sequentially (e.g., 5-FU and oligonucleotide for a period of time followed by MTX and oligonucleotide), or in combination with one or more other such chemotherapeutic agents (e.g., 5-FU, MTX and oligonucleotide, or 5-FU, radiotherapy and oligonucleotide).

Anti-inflammatory drugs, including but not limited to non-steroidal anti-inflammatory drugs and corticosteroids, and antiviral drugs, including but not limited to ribivirin, vidarabine, acyclovir and ganciclovir, may also be combined in compositions of the invention. Combinations of antisense compounds and other non-antisense drugs are also within the scope of this invention. Two or more combined compounds may be used together or sequentially.

In another related embodiment, compositions of the invention may contain one or more antisense compounds, particularly oligonucleotides, targeted to a first nucleic acid and one or more additional antisense compounds targeted to a second nucleic acid target. Alternatively, compositions of the invention may contain two or more antisense compounds targeted to different regions of the same nucleic acid target.

Numerous examples of antisense compounds are known in the art. Two or more combined compounds may be used together or sequentially.

Cell Culture and Antisense Compounds Treatment

The effects of antisense compounds on the level, activity or expression of CD40 nucleic acids can be tested in vitro in a variety of cell types. Cell types used for such analyses are available from commercial vendors (e.g. American Type Culture Collection, Manassas, Va.; Zen-Bio, Inc., Research Triangle Park, N.C.; Clonetics Corporation, Walkersville, Md.) and cells are cultured according to the vendor's instructions using commercially available reagents (e.g. Invitrogen Life Technologies, Carlsbad, Calif.). Illustrative cell types include, but are not limited to, HepG2 cells, Hep3B cells, HuVEC cells, T24, A549, and primary hepatocytes.

In vitro Testing of Antisense Oligonucleotides

Described herein are methods for treatment of cells with antisense oligonucleotides, which can be modified appropriately for treatment with other antisense compounds.

In general, cells are treated with antisense oligonucleotides when the cells reach approximately 60-80% confluency in culture.

One reagent commonly used to introduce antisense oligonucleotides into cultured cells includes the cationic lipid transfection reagent LIPOFECTIN® (Invitrogen, Carlsbad, Calif.). Antisense oligonucleotides are mixed with LIPOFECTIN® in OPTI-MEM® 1 (Invitrogen, Carlsbad, Calif.) to achieve the desired final concentration of antisense oligonucleotide and a LIPOFECTIN® concentration that typically ranges 2 to 12 ug/mL per 100 nM antisense oligonucleotide.

Another reagent used to introduce antisense oligonucleotides into cultured cells includes LIPOFECTAMINE® (Invitrogen, Carlsbad, Calif.). Antisense oligonucleotide is mixed with LIPOFECTAMINE® in OPTI-MEM® 1 reduced serum medium (Invitrogen, Carlsbad, Calif.) to achieve the desired concentration of antisense oligonucleotide and a LIPOFECTAMINE® concentration that typically ranges 2 to 12 ug/mL per 100 nM antisense oligonucleotide.

Cells are treated with antisense oligonucleotides by routine methods. Cells are typically harvested 16-24 hours after antisense oligonucleotide treatment, at which time RNA or protein levels of target nucleic acids are measured by methods known in the art and described herein. In general, when treatments are performed in multiple replicates, the data are presented as the average of the replicate treatments.

The concentration of antisense oligonucleotide used varies from cell line to cell line. Methods to determine the optimal antisense oligonucleotide concentration for a particular cell line are well known in the art. Antisense oligonucleotides are typically used at concentrations ranging from 1 nM to 300 nM.

RNA Isolation

RNA analysis can be performed on total cellular RNA or poly(A)+ mRNA. Methods of RNA isolation are well known in the art. RNA is prepared using methods well known in the art, for example, using the TRIZOL® Reagent (Invitrogen, Carlsbad, Calif.) according to the manufacturer's recommended protocols.

Analysis of Inhibition of Target Levels or Expression

Inhibition of levels or expression of a CD40 nucleic acid can be assayed in a variety of ways known in the art. For example, target nucleic acid levels can be quantitated by, e.g., Northern blot analysis, competitive polymerase chain reaction (PCR), or quantitative real-time PCR. RNA analysis

can be performed on total cellular RNA or poly(A)+ mRNA. Methods of RNA isolation are well known in the art. Northern blot analysis is also routine in the art. Quantitative real-time PCR can be conveniently accomplished using the commercially available ABI PRISM® 7600, 7700, or 7900 Sequence Detection System, available from PE-Applied Biosystems, Foster City, Calif. and used according to manufacturer's instructions.

Quantitative Real-Time PCR Analysis of Target RNA Levels

Quantitation of target RNA levels may be accomplished by quantitative real-time PCR using the ABI PRISM® 7600, 7700, or 7900 Sequence Detection System (PE-Applied Biosystems, Foster City, Calif.) according to manufacturer's instructions. Methods of quantitative real-time PCR are well known in the art.

Prior to real-time PCR, the isolated RNA is subjected to a reverse transcriptase (RT) reaction, which produces complementary DNA (cDNA) that is then used as the substrate for the real-time PCR amplification. The RT and real-time PCR reactions are performed sequentially in the same sample well. RT and real-time PCR reagents are obtained from Invitrogen (Carlsbad, Calif.). RT, real-time-PCR reactions are carried out by methods well known to those skilled in the art.

Gene (or RNA) target quantities obtained by real time PCR are normalized using either the expression level of a gene whose expression is constant, such as GAPDH or by quantifying total RNA using RIBOGREEN® (Invitrogen, Inc. Carlsbad, Calif.). GAPDH expression is quantified by real time PCR, by being run simultaneously with the target, multiplexing, or separately. Total RNA is quantified using RIBOGREEN® RNA quantification reagent (Invitrogen, Inc. Eugene, Oreg.). Methods of RNA quantification by RIBOGREEN® are taught in Jones, L. J., et al, (Analytical Biochemistry, 1998, 265, 368-374). A CYTOFLUOR® 4000 instrument (PE Applied Biosystems) is used to measure RIBOGREEN® fluorescence.

Probes and primers are designed to hybridize to a CD40 nucleic acid. Methods for designing real-time PCR probes and primers are well known in the art, and may include the use of software such as PRIMER EXPRESS® Software (Applied Biosystems, Foster City, Calif.).

Analysis of Protein Levels

Antisense inhibition of CD40 nucleic acids can be assessed by measuring CD40 protein levels. Protein levels of CD40 can be evaluated or quantitated in a variety of ways well known in the art, such as immunoprecipitation, Western blot analysis (immunoblotting), enzyme-linked immunosorbent assay (ELISA), quantitative protein assays, protein activity assays (for example, caspase activity assays), immunohistochemistry, immunocytochemistry or fluorescence-activated cell sorting (FACS). Antibodies directed to a target can be identified and obtained from a variety of sources, such as the MSRS catalog of antibodies (Aerie Corporation, Birmingham, Mich.), or can be prepared via conventional monoclonal or polyclonal antibody generation methods well known in the art. Antibodies useful for the detection of human and rat CD40 are commercially available.

In vivo Testing of Antisense Compounds

Antisense compounds, for example, antisense oligonucleotides, are tested in animals to assess their ability to inhibit expression of CD40 and produce phenotypic changes, such as changes in cell morphology over time or treatment dose as well as changes in levels of cellular components such as proteins, nucleic acids, hormones, cytokines, and eosinophils. Testing may be performed in normal animals, or in experimental disease models. For administration to animals,

antisense oligonucleotides are formulated in a pharmaceutically acceptable diluent, such as phosphate-buffered saline. Administration includes pulmonary administration, aerosol administration, topical administration, and parenteral routes of administration, such as intraperitoneal, intravenous, and subcutaneous. Calculation of antisense oligonucleotide dosage and dosing frequency is within the abilities of those skilled in the art, and depends upon factors such as route of administration and animal body weight. Following a period of treatment with antisense oligonucleotides, RNA is isolated from liver tissue and changes in CD40 nucleic acid expression are measured.

Certain Indications

In certain embodiments, the invention provides methods of treating an individual comprising administering one or more pharmaceutical compositions of the present invention. In certain embodiments, the individual has an inflammatory or hyperproliferative disorder. In certain embodiments the invention provides methods for prophylactically reducing CD40 expression in an individual. Certain embodiments include treating an individual in need thereof by administering to an individual a therapeutically effective amount of an antisense compound targeted to a CD40 nucleic acid.

In one embodiment, administration of a therapeutically effective amount of an antisense compound targeted to a CD40 nucleic acid is accompanied by monitoring of eosinophils in an individual, to determine an individual's response to administration of the antisense compound. An individual's response to administration of the antisense compound is used by a physician to determine the amount and duration of therapeutic intervention.

In one embodiment, administration of an antisense compound targeted to a CD40 nucleic acid results in reduction of CD40 expression by at least 15, 20, 25, 30, 35, 40, 45, 50, 55, 60, 65, 70, 75, 80, 85, 90, 95 or 99%, or a range defined by any two of these values. In some embodiments, administration of a CD40 antisense compound increases the measure by at least 15, 20, 25, 30, 35, 40, 45, 50, 55, 60, 65, 70, 75, 80, 85, 90, 95 or 99%, or a range defined by any two of these values. In some embodiments, administration of a CD40 antisense compound decreases the measure by at least 15, 20, 25, 30, 35, 40, 45, 50, 55, 60, 65, 70, 75, 80, 85, 90, 95 or 99%, or a range defined by any two of these values.

In certain embodiments pharmaceutical composition comprising an antisense compound targeted to CD40 is used for the preparation of a medicament for treating a patient suffering or susceptible to an inflammatory condition or a hyperproliferative disorder.

The formulation of therapeutic compositions and their subsequent administration (dosing) is believed to be within the skill of those in the art. Dosing is dependent on severity and responsiveness of the disease state to be treated, with the course of treatment lasting from several days to several months, or until a cure is effected or a diminution of the disease state is achieved. Optimal dosing schedules can be calculated from measurements of drug accumulation in the body of the patient. Persons of ordinary skill can easily determine optimum dosages, dosing methodologies and repetition rates. Optimum dosages may vary depending on the relative potency of individual oligonucleotides, and can generally be estimated based on EC50s found to be effective in *in vitro* and *in vivo* animal models. In general, dosage is from 0.01 ug to 100 g per kg of body weight, and may be given once or more daily, weekly, monthly or yearly, or even once every 2 to 20 years. Persons of ordinary skill in the art can easily estimate repetition rates for dosing based on measured residence times and concentrations of the drug in

bodily fluids or tissues. Following successful treatment, it may be desirable to have the patient undergo maintenance therapy to prevent the recurrence of the disease state, wherein the oligonucleotide is administered in maintenance doses, ranging from 0.01 ug to 100 g per kg of body weight, once or more daily, to once every 20 years.

Certain Combination Therapies

In certain embodiments, one or more pharmaceutical compositions of the present invention are co-administered with one or more other pharmaceutical agents. In certain embodiments, such one or more other pharmaceutical agents are designed to treat the same disease or condition as the one or more pharmaceutical compositions of the present invention. In certain embodiments, such one or more other pharmaceutical agents are designed to treat a different disease or condition as the one or more pharmaceutical compositions of the present invention. In certain embodiments, such one or more other pharmaceutical agents are designed to treat an undesired effect of one or more pharmaceutical compositions of the present invention. In certain embodiments, one or more pharmaceutical compositions of the present invention are co-administered with another pharmaceutical agent to treat an undesired effect of that other pharmaceutical agent. In certain embodiments, one or more pharmaceutical compositions of the present invention and one or more other pharmaceutical agents are administered at the same time. In certain embodiments, one or more pharmaceutical compositions of the present invention and one or more other pharmaceutical agents are administered at different times. In certain embodiments, one or more pharmaceutical compositions of the present invention and one or more other pharmaceutical agents are prepared together in a single formulation. In certain embodiments, one or more pharmaceutical compositions of the present invention and one or more other pharmaceutical agents are prepared separately.

In certain embodiments, pharmaceutical agents that may be co-administered with a pharmaceutical composition of the present invention include steroids and/or chemotherapeutic agents. In certain such embodiments, pharmaceutical agents that may be co-administered with a pharmaceutical composition of the present invention include, but are not limited to prednisone, corticosteroids, and paclitaxel. In certain such embodiments, the agent is administered prior to administration of a pharmaceutical composition of the present invention. In certain such embodiments, the agent is administered following administration of a pharmaceutical composition of the present invention. In certain such embodiments the agent is administered at the same time as a pharmaceutical composition of the present invention. In certain such embodiments the dose of a co-administered agent is the same as the dose that would be administered if the agent was administered alone. In certain such embodiments the dose of a co-administered agent is lower than the dose that would be administered if the agent was administered alone. In certain such embodiments the dose of a co-administered agent is greater than the dose that would be administered if the agent was administered alone.

In certain embodiments, the co-administration of a second compound enhances the effect of a first compound, such that co-administration of the compounds results in an effect that is greater than the effect of administering the first compound alone. In other embodiments, the co-administration results in effects that are additive of the effects of the compounds when administered alone. In other embodiments, the co-administration results in effects that are supra-additive of the effects of the compounds when administered alone. In some embodiments, the first compound is an antisense compound. In some embodiments, the second compound is an antisense compound.

Nonlimiting Disclosure and Incorporation by Reference

While certain compounds, compositions and methods described herein have been described with specificity in accordance with certain embodiments, the following examples serve only to illustrate the compounds described herein and are not intended to limit the same. Each of the references recited in the present application is incorporated herein by reference in its entirety.

Example 1

Antisense Inhibition of Human CD40 In Vitro

Antisense oligonucleotides targeted to a CD40 nucleic acid were tested for their effects on CD40 mRNA in vitro.

When cultured cells, grown in a 96-well plate, reached 80% confluency, they were treated with 150 nM antisense oligonucleotide. After a treatment period of approximately 24 hours, RNA was isolated from the cells and CD40 mRNA levels were measured by quantitative real-time PCR, as described herein. CD40 mRNA levels were adjusted according to total RNA content as measured by normalization to RIBOGREEN®. Results are presented as percent inhibition of CD40, relative to untreated control cells in Table 1.

The antisense oligonucleotides were designed as 18-mers with phosphorothioate backbones (internucleoside linkages) throughout. "5' target site" indicates the 5'-most nucleotide which the antisense oligonucleotide is targeted to SEQ ID NO: 1 (GENBANK® Accession No X60592.1). Data are averages from three experiments.

TABLE 1

Inhibition of human CD40 mRNA levels by fully phosphorothioate oligodeoxynucleotides							
Oligo ID	Target SEQ ID NO	Target Start Site	Target Stop Site	Target Region	Sequence (5' to 3')	% Inhibition	SEQ ID NO
18623	1	18	35	5' UTR	CCAGGCGGCAGGACCACT	31	5
18624	1	20	37	5' UTR	GACCAGGCGGCAGGACCA	28	6
18625	1	26	43	5' UTR	AGGTGAGACCAGGCGGCA	22	7
18626	1	48	65	AUG	CAGAGGCAGACGAACCAT	0	8
18627	1	49	66	Coding	GCAGAGGCAGACGAACCA	0	9
18628	1	73	90	Coding	GCAAGCAGCCCCAGAGGA	0	10
18629	1	78	95	Coding	GGTCAGCAAGCAGCCCCA	30	11
18630	1	84	101	Coding	GACAGCGGTCAGCAAGCA	0	12
18631	1	88	105	Coding	GATGGACAGCGGTCAGCA	0	13
18632	1	92	109	Coding	TCTGGATGGACAGCGGTC	0	14
18633	1	98	115	Coding	GGTGGTTCTGGATGGACA	0	15
18634	1	101	118	Coding	GTGGGTGGTTCTGGATGG	0	16
18635	1	104	121	Coding	GCAGTGGGTGGTTCTGGA	0	17
18636	1	152	169	Coding	CACAAAGAACAGCACTGA	0	18
18637	1	156	173	Coding	CTGGCACAAAGAAGCA	0	19
18638	1	162	179	Coding	TCCTGGCTGGCACAAAGA	0	20
18639	1	165	182	Coding	CTGTCTGGCTGGCACAA	5	21
18640	1	176	193	Coding	CTCACCAGTTTCTGTCCT	0	22
18641	1	179	196	Coding	TCACTCACCAGTTTCTGT	0	23
18642	1	185	202	Coding	GTGCAGTCACTCACCAGT	0	24
18643	1	190	207	Coding	ACTCTGTGCAGTCACTCA	0	25
18644	1	196	213	Coding	CAGTGAACCTGTGCAGT	5	26
18645	1	205	222	Coding	ATTCCGTTTCAGTGA	0	27
18646	1	211	228	Coding	GAAGGCATTCCGTTTCAG	9	28
18647	1	222	239	Coding	TTCACCGCAAGGAAGGCA	0	29
18648	1	250	267	Coding	CTCTGTTCCAGGTGTCTA	0	30

TABLE 1-continued

Inhibition of human CD40 mRNA levels by fully phosphorothioate oligodeoxynucleotides							
Oligo ID	Target SEQ ID NO	Target Start Site	Target Stop Site	Target Region	Sequence (5' to 3')	% Inhibition	SEQ ID NO
18649	1	267	284	Coding	CTGGTGGCAGTGTGTCTC	0	31
18650	1	286	303	Coding	TGGGGTCGCAGTATTTGT	0	32
18651	1	289	306	Coding	GGTTGGGGTCGCAGTATT	0	33
18652	1	292	309	Coding	CTAGGTTGGGGTCGCAGT	0	34
18653	1	318	335	Coding	GGTGCCCTTCTGCTGGAC	20	35
18654	1	322	339	Coding	CTGAGGTGCCCTTCTGCT	16	36
18655	1	332	349	Coding	GTGTCTGTTTCTGAGGTG	0	37
18656	1	334	351	Coding	TGGTGTCTGTTTCTGAGG	0	38
18657	1	345	362	Coding	ACAGGTGCAGATGGTGTC	0	39
18658	1	348	365	Coding	TTCACAGGTGCAGATGGT	0	40
18659	1	360	377	Coding	GTGCCAGCCTTCTTCACA	6	41
18660	1	364	381	Coding	TACAGTGCCAGCCTTCTT	8	42
18661	1	391	408	Coding	GGACACAGCTCTCACAGG	0	43
18662	1	395	412	Coding	TGCAGGACACAGCTCTCA	0	44
18663	1	401	418	Coding	GAGCGGTGCAGGACACAG	0	45
18664	1	416	433	Coding	AAGCCGGGCGAGCATGAG	0	46
18665	1	432	449	Coding	AATCTGCTTGACCCAAA	6	47
18666	1	446	463	Coding	GAAACCCCTGTAGCAATC	0	48
18667	1	452	469	Coding	GTATCAGAAACCCCTGTA	0	49
18668	1	463	480	Coding	GCTCGCAGATGGTATCAG	0	50
18669	1	468	485	Coding	GCAGGGCTCGCAGATGGT	34	51
18670	1	471	488	Coding	TGGGCAGGGCTCGCAGAT	0	52
18671	1	474	491	Coding	GACTGGGCAGGGCTCGCA	3	53
18672	1	490	507	Coding	CATTGGAGAAGAAGCCGA	0	54
18673	1	497	514	Coding	GATGACACATTGGAGAAG	0	55
18674	1	500	517	Coding	GCAGATGACACATTGGAG	0	56
18675	1	506	523	Coding	TCGAAAGCAGATGACACA	0	57
18676	1	524	541	Coding	GTCCAAGGGTGACATTTT	8	58
18677	1	532	549	Coding	CACAGCTTGTTCCAAGGGT	0	59
18678	1	539	556	Coding	TTGGTCTCACAGCTTGTC	0	60
18679	1	546	563	Coding	CAGGTCTTTGGTCTCACA	7	61
18680	1	558	575	Coding	CTGTTGCACAACCAGGTC	19	62
18681	1	570	587	Coding	GTTTGTGCCTGCCTGTTG	2	63
18682	1	575	592	Coding	GTCTTGTGTTGTGCCTGCC	0	64
18683	1	590	607	Coding	CCACAGACAACATCAGTC	0	65
18684	1	597	614	Coding	CTGGGGACCACAGACAAC	0	66
18685	1	607	624	Coding	TCAGCCGATCCTGGGGAC	0	67

TABLE 1-continued

Inhibition of human CD40 mRNA levels by fully phosphorothioate oligodeoxynucleotides							
Oligo ID	Target SEQ ID NO	Target Start Site	Target Stop Site	Target Region	Sequence (5' to 3')	% Inhibition	SEQ ID NO
18686	1	621	638	Coding	CACCACCAGGGCTCTCAG	23	68
18687	1	626	643	Coding	GGGATCACCACCAGGGCT	0	69
18688	1	657	674	Coding	GAGGATGGCAAACAGGAT	0	70
18689	1	668	685	Coding	ACCAGCACCAAGAGGATG	0	71
18690	1	679	696	Coding	TTTTGATAAAGACCAGCA	0	72
18691	1	703	720	Coding	TATTGGTTGGCTTCTTGG	0	73
18692	1	729	746	Coding	GGGTTCTGCTTGGGGTG	0	74
18693	1	750	767	Coding	GTCGGGAAAATTGATCTC	0	75
18694	1	754	771	Coding	GATCGTCGGGAAAATTGA	0	76
18695	1	765	782	Coding	GGAGCCAGGAAGATCGTC	0	77
18696	1	766	783	Coding	TGGAGCCAGGAAGATCGT	0	78
18697	1	780	797	Coding	TGGAGCAGCAGTGTGGA	0	79
18698	1	796	813	Coding	GTAAAGTCTCTGCACTG	0	80
18699	1	806	823	Coding	TGGCATCCATGTAAAGTC	0	81
18700	1	810	827	Coding	CGGTTGGCATCCATGTAA	0	82
18701	1	834	851	Coding	CTCTTTGCCATCCTCCTG	4	83
18702	1	861	878	Coding	CTGTCTCTCCTGCACTGA	0	84
18703	1	873	890	Stop	GGTGCAGCCTCACTGTCT	0	85
18704	1	910	927	3' UTR	AACTGCCTGTTTGCCAC	34	86
18705	1	954	971	3' UTR	CTTCTGCCTGCACCCCTG	0	87
18706	1	976	993	3' UTR	ACTGACTGGGCATAGCTC	0	88

Example 2

Antisense Inhibition of Human CD40 In Vitro

Antisense oligonucleotides targeted to a CD40 nucleic acid were tested for their effects on CD40 mRNA in vitro. T24 cells at a density of 7000 cells per well in a 96-well plate were treated with 150 nM antisense oligonucleotide. After a treatment period of approximately 24 hours, RNA was isolated from the cells and CD40 mRNA levels were measured by quantitative real-time PCR, as described herein. CD40 mRNA levels were adjusted according to GAPDH content, a housekeeping gene. Results are presented as

percent inhibition of CD40, relative to untreated control cells in Table 2.

The antisense oligonucleotides were designed as 4-10-4 gapmers, where the gap segment comprises 2'-deoxynucleotides and each wing segment comprises 2'-MOE nucleotides and 5-methylcytosine substitutions. The antisense oligonucleotides comprise phosphorothioate backbones (internucleoside linkages) throughout. "5' target site" indicates the 5'-most nucleotide which the antisense oligonucleotide is targeted to SEQ ID NO: 1 (GENBANK® Accession No X60592.1). Data are averages from three experiments. "ND" indicates a value was not determined.

TABLE 2

Inhibition of human CD40 mRNA levels by chimeric oligonucleotides having 4-10-4 MOE wings and deoxy gap							
OligoID	Target SEQ ID NO	Target Start Site	Target Stop Site	Target Region	Sequence (5' to 3')	% Inhibition	SEQ ID NO
19211	1	18	35	5' UTR	CCAGGCGGCAGGACCACT	76	5
19212	1	20	37	5' UTR	GACCAGGCGGCAGGACCA	77	6

TABLE 2-continued

Inhibition of human CD40 mRNA levels by chimeric oligonucleotides having 4-10-4 MOE wings and deoxy gap							
OligoID	Target		Target Stop Site	Target Region	Sequence (5' to 3')	% Inhibition	SEQ ID NO
	SEQ ID NO	Start Site					
19213	1	26	43	5' UTR	AGGTGAGACCAGGCGGCA	81	7
19214	1	48	65	AUG	CAGAGGCAGACGAACCAT	24	8
19215	1	49	66	Coding	GCAGAGGCAGACGAACCA	46	9
19216	1	73	90	Coding	GCAAGCAGCCCCAGAGGA	66	10
19217	1	78	95	Coding	GGTCAGCAAGCAGCCCCA	75	11
19218	1	84	101	Coding	GACAGCGGTCAGCAAGCA	67	12
19219	1	88	105	Coding	GATGGACAGCGGTCAGCA	65	13
19220	1	92	109	Coding	TCTGGATGGACAGCGGTC	79	14
19221	1	98	115	Coding	GGTGGTTCTGGATGGACA	81	15
19222	1	101	118	Coding	GTGGGTGGTTCTGGATGG	58	16
19223	1	104	121	Coding	GCAGTGGGTGGTTCTGGA	74	17
19224	1	152	169	Coding	CACAAAGAACAGCACTGA	40	18
19225	1	156	173	Coding	CTGGCACAAGAAGCAGCA	60	19
19226	1	162	179	Coding	TCCTGGCTGGCACAAGA	10	20
19227	1	165	182	Coding	CTGTCTGGCTGGCACA	24	21
19228	1	176	193	Coding	CTCACCAGTTTCTGTCTCT	22	22
19229	1	179	196	Coding	TCACTCACCAGTTTCTGT	41	23
19230	1	185	202	Coding	GTGCAGTCACTCACCAGT	82	24
19231	1	190	207	Coding	ACTCTGTGCAGTCACTCA	38	25
19232	1	196	213	Coding	CAGTGAACCTGTGCAGT	40	26
19233	1	205	222	Coding	ATTCCGTTTTCAGTGA	56	27
19234	1	211	228	Coding	GAAGGCATTCCGTTTCAG	32	28
19235	1	222	239	Coding	TTCACCGCAAGGAAGGCA	61	29
19236	1	250	267	Coding	CTCTGTTCCAGGTGTCTA	62	30
19237	1	267	284	Coding	CTGGTGGCAGTGTGTCTC	70	31
19238	1	286	303	Coding	TGGGGTCGCAGTATTTGT	0	32
19239	1	289	306	Coding	GGTTGGGGTCGCAGTATT	19	33
19240	1	292	309	Coding	CTAGGTTGGGGTCGCAGT	36	34
19241	1	318	335	Coding	GGTGCCCTTCTGCTGGAC	79	35
19242	1	322	339	Coding	CTGAGGTGCCCTTCTGCT	70	36
19243	1	332	349	Coding	GTGTCTGTTTCTGAGGTG	63	37
19244	1	334	351	Coding	TGGTGTCTGTTTCTGAGG	43	38
19245	1	345	362	Coding	ACAGGTGCAGATGGTGTC	73	39
19246	1	348	365	Coding	TTCACAGGTGCAGATGGT	48	40
19247	1	360	377	Coding	GTGCCAGCCTTCTTCACA	61	41
19248	1	364	381	Coding	TACAGTGCCAGCCTTCTT	47	42

TABLE 2-continued

Inhibition of human CD40 mRNA levels by chimeric oligonucleotides having 4-10-4 MOE wings and deoxy gap							
OligoID	Target		Target Stop Site	Target Region	Sequence (5' to 3')	% Inhibition	SEQ ID NO
	SEQ ID NO	Target Start Site					
19249	1	391	408	Coding	GGACACAGCTCTCACAGG	0	43
19250	1	395	412	Coding	TGCAGGACACAGCTCTCA	52	44
19251	1	401	418	Coding	GAGCGGTGCAGGACACAG	50	45
19252	1	416	433	Coding	AAGCCGGGCGAGCATGAG	32	46
19253	1	432	449	Coding	AATCTGCTTGACCCCAA	0	47
19254	1	446	463	Coding	GAAACCCCTGTAGCAATC	0	48
19255	1	452	469	Coding	GTATCAGAAACCCCTGTA	36	49
19256	1	463	480	Coding	GCTCGCAGATGGTATCAG	65	50
19257	1	468	485	Coding	GCAGGGCTCGCAGATGGT	75	51
19258	1	471	488	Coding	TGGGCAGGGCTCGCAGAT	0	52
19259	1	474	491	Coding	GACTGGGCAGGGCTCGCA	82	53
19260	1	490	507	Coding	CATTGGAGAAGAAGCCGA	41	54
19261	1	497	514	Coding	GATGACACATTGGAGAAG	14	55
19262	1	500	517	Coding	GCAGATGACACATTGGAG	78	56
19263	1	506	523	Coding	TCGAAAGCAGATGACACA	59	57
19264	1	524	541	Coding	GTCCAAGGGTGACATTTT	71	58
19265	1	532	549	Coding	CACAGCTTGTTCCAAGGGT	0	59
19266	1	539	556	Coding	TTGGTCTCACAGCTTGTC	46	60
19267	1	546	563	Coding	CAGGTCTTTGGTCTCACA	64	61
19268	1	558	575	Coding	CTGTTGCACAACCAGGTC	82	62
19269	1	570	587	Coding	GTTTGTGCCTGCCTGTTG	70	63
19270	1	575	592	Coding	GTCTTGTGTTGTCCTGCC	69	64
19271	1	590	607	Coding	CCACAGACAACATCAGTC	11	65
19272	1	597	614	Coding	CTGGGGACCACAGACAAC	9	66
19273	1	607	624	Coding	TCAGCCGATCCTGGGGAC	0	67
19274	1	621	638	Coding	CACCACCAGGGCTCTCAG	23	68
19275	1	626	643	Coding	GGGATCACCACCAGGGCT	58	69
19276	1	657	674	Coding	GAGGATGGCAAACAGGAT	49	70
19277	1	668	685	Coding	ACCAGCACCAAGAGGATG	ND	71
19278	1	679	696	Coding	TTTTGATAAAGACCAGCA	31	72
19279	1	703	720	Coding	TATTGGTTGGCTTCTTGG	49	73
19280	1	729	746	Coding	GGGTTCCCTGCTTGGGGTG	14	74
19281	1	750	767	Coding	GTCGGGAAAATTGATCTC	55	75
19282	1	754	771	Coding	GATCGTCGGGAAAATTGA	0	76
19283	1	765	782	Coding	GGAGCCAGGAAGATCGTC	69	77
19284	1	766	783	Coding	TGGAGCCAGGAAGATCGT	54	78

TABLE 2-continued

Inhibition of human CD40 mRNA levels by chimeric oligonucleotides having 4-10-4 MOE wings and deoxy gap								
OligoID	Target		Target Start Site	Target Stop Site	Target Region	Sequence (5' to 3')	% Inhibition	SEQ ID NO
	SEQ ID NO	Target Start Site						
19285	1	780	797	Coding	TGGAGCAGCAGTGTGGA	15	79	
19286	1	796	813	Coding	GTAAAGTCTCCTGCACTG	31	80	
19287	1	806	823	Coding	TGGCATCCATGTAAAGTC	65	81	
19288	1	810	827	Coding	CGGTTGGCATCCATGTAA	34	82	
19289	1	834	851	Coding	CTCTTTGCCATCCTCCTG	42	83	
19290	1	861	878	Coding	CTGTCTCTCCTGCACTGA	26	84	
19291	1	873	890	Stop	GGTGCAGCCTCACTGTCT	76	85	
19292	1	910	927	3' UTR	AACTGCCTGTTTGCCAC	63	86	
19293	1	954	971	3' UTR	CTTCTGCCTGCACCCCTG	0	87	
19294	1	976	993	3' UTR	ACTGACTGGGCATAGCTC	12	88	

Example 3

Antisense Inhibition of Human CD40

Antisense oligonucleotides targeted to a CD40 nucleic acid were tested for their effects on CD40 mRNA in vitro. T24 cells at a density of 7000 cells per well in a 96-well plate were treated with 100 nM of antisense oligonucleotide. After a treatment period of approximately 24 hours, RNA was isolated from the cells and CD40 mRNA levels were measured by quantitative real-time PCR, as described herein. CD40 mRNA levels were adjusted according to GAPDH

content, a housekeeping gene. Results are presented as percent inhibition of CD40, relative to untreated control cells in Table 3.

The antisense oligonucleotides were designed as 4-10-4 gapmers, where the gap segment comprises 2'-deoxynucleotides and each wing segment comprises 2'-MOE nucleotides. The antisense oligonucleotides comprise phosphorothioate backbones (internucleoside linkages) and 5-methylcytosine substitutions throughout. "5' target site" indicates the 5'-most nucleotide which the antisense oligonucleotide is targeted to SEQ ID NO: 1 (GENBANK Accession No. X60592.1), SEQ ID NO: 2 (GENBANK® Accession No. H50598.1), and SEQ ID NO: 3 (GENBANK® Accession No. AA203290.1).

TABLE 3

Inhibition of human CD40 mRNA levels by chimeric oligonucleotides having 4-10-4 MOE wings and deoxy gap							
Oligo ID	Target		Target Start Site	Target Stop Site	Sequence (5' to 3')	% Inhibition	SEQ ID NO
	SEQ ID NO	Target Start Site					
26162	1	66	83	GCCCCAGAGGACGCACTG	0	89	
26163	1	70	87	AGCAGCCCCAGAGGACGC	98	90	
26164	1	74	91	AGCAAGCAGCCCCAGAGG	47	91	
26165	1	80	97	GCGGTCAGCAAGCAGCCC	54	92	
26167	1	95	112	GGTTCTGGATGGACAGCG	66	93	
26168	1	102	119	AGTGGGTGGTTCTGGATG	26	94	
26169	1	141	158	GCACTGACTGTTTATTAG	43	95	
26170	1	154	171	GGCACAAGAACAGCACT	53	96	
26171	1	164	181	TGTCCTGGCTGGCACAAA	29	97	
26172	1	171	188	CAGTTTCTGTCTGGCTG	48	98	
26173	1	180	197	GTCCTCACCAGTTTCTG	47	99	

TABLE 3-continued

Inhibition of human CD40 mRNA levels by chimeric oligonucleotides having 4-10-4 MOE wings and deoxy gap						
Oligo ID	Target SEQ ID NO	Target Start Site	Target Stop Site	Sequence (5' to 3')	% Inhibition	SEQ ID NO
26174	1	210	227	AAGGCATTCCGTTTCAGT	57	100
26175	1	224	241	CTTTCACCGCAAGGAAGG	34	101
26176	1	250	267	CTCTGTTCCAGGTGTCTA	78	30
26177	1	257	274	TGTGTCTCTCTGTTCCAG	57	102
26178	1	264	281	GTGGCAGTGTGTCTCTCT	0	103
26179	1	314	331	CCCTTCTGCTGGACCCGA	58	104
26180	1	321	338	TGAGGTGCCCTTCTGCTG	69	105
26181	1	329	346	TCTGTTTCTGAGGTGCCC	44	106
26182	1	336	353	GATGGTGTCTGTTTCTGA	12	107
26183	1	364	381	TACAGTGCCAGCCTTCTT	14	42
26184	1	445	462	AAACCCCTGTAGCAATCT	15	108
26185	1	460	477	CGCAGATGGTATCAGAAA	53	109
26186	1	469	486	GGCAGGGCTCGCAGATGG	79	110
26202	1	485	502	GAGAAGAAGCCGACTGGG	0	111
26187	1	487	504	TGGAGAAGAAGCCGACTG	23	112
26204	1	489	506	ATTGGAGAAGAAGCCGAC	0	113
26205	1	491	508	ACATTGGAGAAGAAGCCG	4	114
26206	1	493	510	ACACATTGGAGAAGAAGC	0	115
26207	1	495	512	TGACACATTGGAGAAGAA	46	116
26188	1	496	513	ATGACACATTGGAGAAGA	0	117
26208	1	497	514	GATGACACATTGGAGAAG	0	55
26189	1	503	520	AAAGCAGATGACACATTG	6	118
26209	1	524	541	GTCCAAGGGTGACATTTT	53	58
26210	1	545	562	AGGTCTTTGGTCTCACAG	81	119
26211	1	555	572	TTGCACAACCAGGTCTTT	48	120
26212	1	570	587	GTTTGTGCCTGCCTGTTG	76	63
26213	1	572	589	TTGTTTGTGCCTGCCTGT	50	121
26214	1	574	591	TCTTGTTTGTGCCTGCCT	87	122
26215	1	576	593	AGTCTTGTTTGTGCCTGC	83	123
26216	1	577	594	CAGTCTTGTTTGTGCCTG	80	124
26217	1	578	595	TCAGTCTTGTTTGTGCCT	88	125
26218	1	580	597	CATCAGTCTTGTTTGTGC	52	126
26219	1	590	607	CCACAGACAACATCAGTC	16	65
26220	1	592	609	GACCACAGACAACATCAG	11	127
26221	1	594	611	GGGACCACAGACAACATC	40	128
26222	1	622	639	TCACCACCAGGGCTCTCA	37	129

TABLE 3-continued

Inhibition of human CD40 mRNA levels by chimeric oligonucleotides having 4-10-4 MOE wings and deoxy gap						
Oligo ID	Target SEQ ID NO	Target Start Site	Target Stop Site	Sequence (5' to 3')	% Inhibition	SEQ ID NO
26223	1	624	641	GATCACCACCAGGGCTCT	82	130
26224	1	658	675	AGAGGATGGCAAACAGGA	33	131
26225	1	659	676	AAGAGGATGGCAAACAGG	0	132
26226	1	660	677	CAAGAGGATGGCAAACAG	0	133
26227	1	669	686	GACCAGCACCAAGAGGAT	57	134
26228	1	671	688	AAGACCAGCACCAAGAGG	35	135
26229	1	673	690	TAAAGACCAGCACCAAGA	13	136
26230	1	676	693	TGATAAAGACCAGCACCA	0	137
26231	1	678	695	TTTGATAAAGACCAGCAC	26	138
26232	2	375	392	ACTCTCTTTGCCCATCCT	0	139
26233	2	377	394	CGACTCTCTTTGCCCATC	31	140
26234	2	380	397	ATGCGACTCTCTTTGCC	12	141
26235	2	382	399	AAATGCGACTCTCTTTGC	36	142
26236	2	385	402	CTGAAATGCGACTCTCTT	51	143
26237	2	387	404	AACTGAAATGCGACTCTC	0	144
26238	2	406	423	CTTCACTGTCTCTCCCTG	0	145
26239	2	407	424	CCTTCACTGTCTCTCCCT	56	146
26240	2	409	426	AACCTTCACTGTCTCTCC	0	147
26190	3	520	537	GATCACCACAGGCTCTCA	0	148
26191	3	565	582	TGATAAGACAGCACCAAG	9	149
26192	3	584	601	GGTAGTTCTTGCCACTTT	0	150
26193	3	593	610	GGGCCTATGGGTAGTTCT	0	151
26194	3	617	634	ATTATCTCTGGGTCTGCT	9	152
26195	3	646	663	ACTGACACATTTGAGCAG	0	153
26196	3	654	671	GACTCCCTACTGACACAT	0	154
26197	3	689	706	CAAAGAGCGGTTCTCCAC	0	155
26198	3	696	713	AATTCTCCAAAGAGCGGT	0	156
26199	3	728	745	TCTTGACATCCTTTTCAT	0	157
26200	3	736	753	CCCACCTATCTTGACATC	0	158
26201	3	791	808	AGGCCGAGAGTTCAAAAT	0	159

Example 4

Antisense Inhibition of Human CD40

Antisense oligonucleotides targeted to a CD40 nucleic acid were tested for their effects on CD40 mRNA in vitro. A549 cells at a density of 5000 cells per well in a 96-well plate were treated with 120 nM of antisense oligonucleotide.

60 After a treatment period of approximately 24 hours, RNA was isolated from the cells and CD40 mRNA levels were measured by quantitative real-time PCR, as described herein. CD40 primer probe set LTS37 was used to measure mRNA levels. CD40 mRNA levels were adjusted according to total RNA content as measured by RIBOGREEN®. 65 Results are presented as percent inhibition of CD40, relative to untreated control cells in Table 4.

The antisense oligonucleotides were designed as 4-10-4 gapmers, 5-10-5 gapmers, or 2-15-3 gapmers, where the gap segment comprises 2'-deoxynucleotides and each wing segment comprises 2'-MOE nucleotides. The motif for each compound is indicated by the column labeled "motif." The antisense oligonucleotides comprise phosphorothioate back-

bones (internucleoside linkages) and 5-methylcytosine substitutions throughout. "5' target site" indicates the 5'-most nucleotide which the antisense oligonucleotide is targeted to SEQ ID NO: 1 (GENBANK Accession No. X60592.1) or SEQ ID NO: 4 (nucleotides 9797000 to 9813000 of GENBANK Accession No. NT_011362.9).

TABLE 4

Inhibition of human CD40 mRNA levels by chimeric oligonucleotides having 4-10-4 MOE wings and deoxy gap, 5-10-5 MOE wings and deoxy gap, and 2-15-3 MOE wings and deoxy gap

Oligo ID	Target SEQ ID NO	Target Motif	Target Start Site	Target Stop Site	Sequence (5' to 3')	% Inhibition	SEQ ID NO
26163	4	4-10-4	2914	2931	AGCAGCCCCAGAGGACGC	74	90
396243	4	5-10-5	2728	2747	CCAGCAATTCACCGCGCAGG	0	160
396320	4	2-15-3	2728	2747	CCAGCAATTCACCGCGCAGG	0	160
396199	4	5-10-5	2892	2911	TGCAGAGGCAGACGAACCAT	75	161
396276	4	2-15-3	2892	2911	TGCAGAGGCAGACGAACCAT	55	161
396200	4	5-10-5	2904	2923	CAGAGGACGCACTGCAGAGG	79	162
396277	4	2-15-3	2904	2923	CAGAGGACGCACTGCAGAGG	69	162
396201	4	5-10-5	2913	2932	AAGCAGCCCCAGAGGACGCA	76	163
396278	4	2-15-3	2913	2932	AAGCAGCCCCAGAGGACGCA	78	163
396202	4	5-10-5	2924	2943	CAGCGGTCAGCAAGCAGCCC	68	164
396279	4	2-15-3	2924	2943	CAGCGGTCAGCAAGCAGCCC	88	164
396244	4	5-10-5	2928	2947	CTCACAGCGGTCAGCAAGCA	86	165
396321	4	2-15-3	2928	2947	CTCACAGCGGTCAGCAAGCA	75	165
396245	4	5-10-5	3349	3368	GCTGGCAAGGAGATGATAAC	51	166
396322	4	2-15-3	3349	3368	GCTGGCAAGGAGATGATAAC	54	166
396246	4	5-10-5	3480	3499	AGGTTGGAACACCCAAGATA	69	167
396323	4	2-15-3	3480	3499	AGGTTGGAACACCCAAGATA	78	167
396247	4	5-10-5	3649	3668	GGAGAAACCCCTGGTTTCTC	45	168
396324	4	2-15-3	3649	3668	GGAGAAACCCCTGGTTTCTC	26	168
396248	4	5-10-5	3860	3879	TCATTCCTGCCAGGCTTCA	43	169
396325	4	2-15-3	3860	3879	TCATTCCTGCCAGGCTTCA	39	169
396249	4	5-10-5	3950	3969	TCAGGTGAAAGTGAAAGCTG	68	170
396326	4	2-15-3	3950	3969	TCAGGTGAAAGTGAAAGCTG	69	170
396250	4	5-10-5	4490	4509	TACCATCTTCAAACACATGA	79	171
396327	4	2-15-3	4490	4509	TACCATCTTCAAACACATGA	71	171
396251	4	5-10-5	4604	4623	TTACCCAAAATGGGAAAGGA	86	172
396328	4	2-15-3	4604	4623	TTACCCAAAATGGGAAAGGA	48	172
396252	4	5-10-5	4810	4829	GAAAGAATACATGTATATGG	72	173
396329	4	2-15-3	4810	4829	GAAAGAATACATGTATATGG	10	173
396253	4	5-10-5	4944	4963	AGAGTCAGACAGCTTTAGAC	78	174
396330	4	2-15-3	4944	4963	AGAGTCAGACAGCTTTAGAC	79	174
396254	4	5-10-5	5651	5670	GTACCACCCATGCTATTAAT	79	175

TABLE 4-continued

Inhibition of human CD40 mRNA levels by chimeric oligonucleotides having
4-10-4 MOE wings and deoxy gap, 5-10-5 MOE wings and deoxy gap, and
2-15-3 MOE wings and deoxy gap

Oligo ID	Target SEQ ID NO	Target Motif	Target Start Site	Target Stop Site	Target Sequence (5' to 3')	% Inhibition	SEQ ID NO
396331	4	2-15-3	5651	5670	GTACCACCCATGCTATTAAT	84	175
396255	4	5-10-5	5740	5759	ACAGTGACAGAGTCCAATG	85	176
396332	4	2-15-3	5740	5759	ACAGTGACAGAGTCCAATG	75	176
396256	4	5-10-5	5830	5849	AATGTAAAGCTGGAAGGGTA	52	177
396333	4	2-15-3	5830	5849	AATGTAAAGCTGGAAGGGTA	37	177
396257	4	5-10-5	5964	5983	GGGCTATGTTTAGCACTTGG	79	178
396334	4	2-15-3	5964	5983	GGGCTATGTTTAGCACTTGG	73	178
396258	4	5-10-5	6078	6097	GGGCTTGATGCCTGAGTCAT	73	179
396335	4	2-15-3	6078	6097	GGGCTTGATGCCTGAGTCAT	40	179
396259	4	5-10-5	6251	6270	TGAAGTGCAAGTCAAAACAG	52	180
396336	4	2-15-3	6251	6270	TGAAGTGCAAGTCAAAACAG	44	180
396260	4	5-10-5	6332	6351	GCAATTTGAAGGGATCTTGA	68	181
396337	4	2-15-3	6332	6351	GCAATTTGAAGGGATCTTGA	42	181
396203	4	5-10-5	6374	6393	CATGCAGTGGGTGGTTCTGG	77	182
396280	4	2-15-3	6374	6393	CATGCAGTGGGTGGTTCTGG	83	182
396204	4	5-10-5	6385	6404	GTTTTTCTCTGCATGCAGTG	78	183
396281	4	2-15-3	6385	6404	GTTTTTCTCTGCATGCAGTG	70	183
396205	4	5-10-5	6424	6443	GCTGGCACAAAGAACAGCAC	61	184
396282	4	2-15-3	6424	6443	GCTGGCACAAAGAACAGCAC	65	184
396261	4	5-10-5	6709	6728	CACTAACCACACAATGATCA	85	185
396338	4	2-15-3	6709	6728	CACTAACCACACAATGATCA	62	185
396206	4	5-10-5	6787	6806	TGTGCAGTCACTCACCAGTT	83	186
396283	4	2-15-3	6787	6806	TGTGCAGTCACTCACCAGTT	72	186
396207	4	5-10-5	6838	6857	GTCTAGGAATTCGCTTTCAC	95	187
396284	4	2-15-3	6838	6857	GTCTAGGAATTCGCTTTCAC	85	187
396208	4	5-10-5	6843	6862	CAGGTGTCTAGGAATTCGCT	98	188
396285	4	2-15-3	6843	6862	CAGGTGTCTAGGAATTCGCT	90	188
396209	4	5-10-5	6883	6902	GTCGCAGTATTTGTGCTGGT	84	189
396286	4	2-15-3	6883	6902	GTCGCAGTATTTGTGCTGGT	86	189
396262	4	5-10-5	7154	7173	ACCCGAAGCCCTAGGTCTGA	92	190
396339	4	2-15-3	7154	7173	ACCCGAAGCCCTAGGTCTGA	84	190
396210	4	5-10-5	7158	7177	CTGGACCCGAAGCCCTAGGT	82	191
396287	4	2-15-3	7158	7177	CTGGACCCGAAGCCCTAGGT	90	191
396211	4	5-10-5	7163	7182	TTCTGCTGGACCCGAAGCCC	65	192
396288	4	2-15-3	7163	7182	TTCTGCTGGACCCGAAGCCC	80	192
396212	4	5-10-5	7204	7223	CTTCTTACAGGTGCAGATG	79	193

TABLE 4-continued

Inhibition of human CD40 mRNA levels by chimeric oligonucleotides having
4-10-4 MOE wings and deoxy gap, 5-10-5 MOE wings and deoxy gap, and
2-15-3 MOE wings and deoxy gap

Oligo ID	Target SEQ ID NO	Target Motif	Target Start Site	Target Stop Site	Target Sequence (5' to 3')	% Inhibition	SEQ ID NO
396289	4	2-15-3	7204	7223	CTTCTTCACAGGTGCAGATG	72	193
396263	4	5-10-5	7590	7609	AGCCAGTGGCCAGGCAGGAC	70	194
396340	4	2-15-3	7590	7609	AGCCAGTGGCCAGGCAGGAC	56	194
396214	4	5-10-5	7704	7723	GAAGAAGCCGACTGGGCAGG	76	195
396291	4	2-15-3	7704	7723	GAAGAAGCCGACTGGGCAGG	80	195
396215	4	5-10-5	7709	7728	TTGGAGAAGAAGCCGACTGG	77	196
396292	4	2-15-3	7709	7728	TTGGAGAAGAAGCCGACTGG	80	196
396216	4	5-10-5	7718	7737	GATGACACATTGGAGAAGAA	76	197
396293	4	2-15-3	7718	7737	GATGACACATTGGAGAAGAA	65	197
396264	4	5-10-5	7953	7972	TGTCTATTACCTCAAAGAGA	89	198
396341	4	2-15-3	7953	7972	TGTCTATTACCTCAAAGAGA	72	198
396265	4	5-10-5	8492	8511	ACAGTGTGTTTCAGAGGATTG	82	199
396342	4	2-15-3	8492	8511	ACAGTGTGTTTCAGAGGATTG	67	199
396266	4	5-10-5	9755	9774	ACAATACACTTTACATGTTT	90	200
396343	4	2-15-3	9755	9774	ACAATACACTTTACATGTTT	63	200
396267	4	5-10-5	10414	10433	ATTGTGTCTTTAGAACCAGA	84	201
396344	4	2-15-3	10414	10433	ATTGTGTCTTTAGAACCAGA	59	201
396268	4	5-10-5	10528	10547	GGGCCCTAAAGGATGTAAAA	34	202
396345	4	2-15-3	10528	10547	GGGCCCTAAAGGATGTAAAA	76	202
396217	4	5-10-5	11218	11237	CAGTCTTGTTTGTGCCTGCC	70	203
396294	4	2-15-3	11218	11237	CAGTCTTGTTTGTGCCTGCC	79	203
396269	4	5-10-5	11244	11263	TGTCCAGGACTCACCACAGA	77	204
396346	4	2-15-3	11244	11263	TGTCCAGGACTCACCACAGA	83	204
396270	4	5-10-5	11801	11820	TATGGCACCTTCTTAAATAT	85	205
396347	4	2-15-3	11801	11820	TATGGCACCTTCTTAAATAT	81	205
396271	4	5-10-5	12248	12267	TGCTTTTGGTATAGAAGAGT	86	206
396348	4	2-15-3	12248	12267	TGCTTTTGGTATAGAAGAGT	76	206
396235	4	5-10-5	12526	12545	AAATGTGGCTGGCAGATGTC	79	207
396312	4	2-15-3	12526	12545	AAATGTGGCTGGCAGATGTC	82	207
396236	4	5-10-5	12572	12591	GTCAGAGCTCATCTACATCA	87	208
396313	4	2-15-3	12572	12591	GTCAGAGCTCATCTACATCA	82	208
396237	4	5-10-5	12754	12773	CTGATAAAGACCAGCACCAA	69	209
396314	4	2-15-3	12754	12773	CTGATAAAGACCAGCACCAA	70	209
396238	4	5-10-5	12762	12781	AGGACTCACTGATAAAGACC	69	210
396315	4	2-15-3	12762	12781	AGGACTCACTGATAAAGACC	43	210
396239	4	5-10-5	12982	13001	CAGACTCTGAATCAGTTTTA	78	211

TABLE 4-continued

Inhibition of human CD40 mRNA levels by chimeric oligonucleotides having
4-10-4 MOE wings and deoxy gap, 5-10-5 MOE wings and deoxy gap, and
2-15-3 MOE wings and deoxy gap

Oligo ID	Target SEQ ID NO	Target Motif	Target Start Site	Target Stop Site	Target Sequence (5' to 3')	% Inhibition	SEQ ID NO
396316	4	2-15-3	12982	13001	CAGACTCTGAATCAGTTTTA	70	211
396240	4	5-10-5	13021	13040	CAGTCCCCAATTCTGCTGCC	43	212
396317	4	2-15-3	13021	13040	CAGTCCCCAATTCTGCTGCC	70	212
396241	4	5-10-5	13107	13126	CCAGTGTTAGGCTCTGCCAG	76	213
396318	4	2-15-3	13107	13126	CCAGTGTTAGGCTCTGCCAG	85	213
396242	4	5-10-5	13134	13153	GAATGCCAGGAAAGGAGTGA	69	214
396319	4	2-15-3	13134	13153	GAATGCCAGGAAAGGAGTGA	84	214
396272	4	5-10-5	13171	13190	CAGCCCCAAGGCCCAAAGAT	48	215
396349	4	2-15-3	13171	13190	CAGCCCCAAGGCCCAAAGAT	57	215
396220	4	5-10-5	13491	13510	CTGCACTGGAGCAGCAGTGT	81	216
396297	4	2-15-3	13491	13510	CTGCACTGGAGCAGCAGTGT	74	216
396221	4	5-10-5	13517	13536	ACCGGTTGGCATCCATGTAA	61	217
396298	4	2-15-3	13517	13536	ACCGGTTGGCATCCATGTAA	73	217
396222	4	5-10-5	13525	13544	CCTGGGTGACCGGTTGGCAT	71	218
396299	4	2-15-3	13525	13544	CCTGGGTGACCGGTTGGCAT	82	218
396223	4	5-10-5	13802	13821	CAAGTTGGGAGACTGGATGG	65	219
396300	4	2-15-3	13802	13821	CAAGTTGGGAGACTGGATGG	79	219
396224	4	5-10-5	13810	13829	CTTTAATACAAGTTGGGAGA	68	220
396301	4	2-15-3	13810	13829	CTTTAATACAAGTTGGGAGA	64	220
396225	4	5-10-5	13877	13896	TCGGAAGGTCTGGTGGATAT	65	221
396302	4	2-15-3	13877	13896	TCGGAAGGTCTGGTGGATAT	83	221
396226	4	5-10-5	13896	13915	TGGGCACCAAAGCTGGAT	70	222
396303	4	2-15-3	13896	13915	TGGGCACCAAAGCTGGAT	76	222
396227	4	5-10-5	13937	13956	TATGGCTTCCTGGGCGCAGG	59	223
396304	4	2-15-3	13937	13956	TATGGCTTCCTGGGCGCAGG	74	223
396228	4	5-10-5	13961	13980	AATGCTGCAATGGGCATCTG	78	224
396305	4	2-15-3	13961	13980	AATGCTGCAATGGGCATCTG	83	224
396229	4	5-10-5	13977	13996	GTTCACTATCACAACAATG	84	225
396306	4	2-15-3	13977	13996	GTTCACTATCACAACAATG	67	225
396230	4	5-10-5	13997	14016	CAGTTAAGCAGCTTCCAGTT	84	226
396307	4	2-15-3	13997	14016	CAGTTAAGCAGCTTCCAGTT	85	226
396231	4	5-10-5	14028	14047	AATTTTATTTAGCCAGTCTC	80	227
396308	4	2-15-3	14028	14047	AATTTTATTTAGCCAGTCTC	79	227
396232	4	5-10-5	14046	14065	GTTGTATAAATATATTCTAA	44	228
396309	4	2-15-3	14046	14065	GTTGTATAAATATATTCTAA	25	228
396233	4	5-10-5	14065	14084	ACAGTGTTTTTGAGATTCTG	83	229

TABLE 4-continued

Inhibition of human CD40 mRNA levels by chimeric oligonucleotides having 4-10-4 MOE wings and deoxy gap, 5-10-5 MOE wings and deoxy gap, and 2-15-3 MOE wings and deoxy gap							
Oligo ID	Target SEQ ID NO	Target Motif	Target Start Site	Target Stop Site	Target Sequence (5' to 3')	% Inhibition	SEQ ID NO
396310	4	2-15-3	14065	14084	ACAGTGTTTTTGAGATTCTG	50	229
396273	4	5-10-5	14725	14744	CTCAGGACCCAGAGTGAGGA	37	230
396350	4	2-15-3	14725	14744	CTCAGGACCCAGAGTGAGGA	50	230
396274	4	5-10-5	15073	15092	TGGGTTAAACCTCACCTCGA	59	231
396351	4	2-15-3	15073	15092	TGGGTTAAACCTCACCTCGA	56	231
396275	4	5-10-5	15350	15369	ATTAGGTCCCAAAGTTCCCC	23	232
396352	4	2-15-3	15350	15369	ATTAGGTCCCAAAGTTCCCC	48	232
396234	1	5-10-5	42	61	GGCAGACGAACCATGGCGAG	86	233
396311	1	2-15-3	42	61	GGCAGACGAACCATGGCGAG	82	233
396213	1	5-10-5	435	454	GTAGCAATCTGCTTGACCCC	82	234
396290	1	2-15-3	435	454	GTAGCAATCTGCTTGACCCC	79	234
396218	1	5-10-5	590	609	GACCACAGACAACATCAGTC	89	235
396295	1	2-15-3	590	609	GACCACAGACAACATCAGTC	85	235
396219	1	5-10-5	683	702	CCACCTTTTTGATAAAGACC	65	236
396296	1	2-15-3	683	702	CCACCTTTTTGATAAAGACC	41	236

Example 5

Antisense Inhibition of Human CD40 in HuVEC
Cells, Primer Probe Set LTS37

Several antisense oligonucleotides exhibiting in vitro inhibition of CD40 (see Example 4) were tested at various doses in HuVEC cells. Cells were plated at densities of 5000 cells per well and treated with nM concentrations of antisense oligonucleotide as indicated in Table 5. After a treatment period of approximately 24 hours, RNA was isolated from the cells and CD40 mRNA levels were measured by quantitative real-time PCR, as described herein. Human CD40 primer probe set LTS37 was used to measure mRNA levels. CD40 mRNA levels were adjusted according to total RNA content as measured by RIBOGREEN®. Results are presented as percent inhibition of CD40, relative to untreated control cells. As illustrated in Table 5, CD40 mRNA levels were reduced in a dose-dependent manner.

TABLE 5

Antisense Inhibition of human CD40 in HuVEC cells, Primer Probe Set LTS37								
ISIS No	0.2344 nM	0.4688 nM	0.9375 nM	1.875 nM	3.75 nM	7.5 nM	15.0 nM	30.0 nM
26163	17	35	38	51	62	67	82	89
396236	23	49	59	77	86	92	91	89
396266	35	45	58	74	55	72	57	56
396307	21	45	43	56	80	79	82	82
396218	34	47	52	57	78	82	86	86
396279	34	54	59	49	72	82	88	87

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TABLE 5-continued

Antisense Inhibition of human CD40 in HuVEC cells, Primer Probe Set LTS37								
ISIS No	0.2344 nM	0.4688 nM	0.9375 nM	1.875 nM	3.75 nM	7.5 nM	15.0 nM	30.0 nM
396287	31	48	52	50	64	77	85	86
396264	39	34	49	56	71	84	88	86

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Example 6

Antisense Inhibition of Human CD40 in AGS Cells

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Antisense oligonucleotides exhibiting in vitro inhibition of CD40 (see Example 4) were tested at various doses in AGS cells (human adenocarcinoma cells). Antisense oligonucleotides of SEQ ID No. 90 and SEQ ID No. 208 were designed as 4-10-4 gapmers or 5-10-5 gapmers, respectively, where the gap segment comprises 2'-deoxynucleotides and each wing segment comprises 2'-MOE or 2'OMe nucleotides. The antisense oligonucleotides comprise phosphorothioate backbones (internucleoside linkages) and 5-methylcytosine substitutions throughout.

Cells were plated at densities of 5000 cells per well and treated with nM concentrations of antisense oligonucleotide as indicated in Table 6. After a treatment period of approximately 24 hours, RNA was isolated from the cells and relative CD40 mRNA expression levels were quantified by real time RT-PCR using the QuantiTect™ SYBR® Green RT-PCR kit (Qiagen). CD40 mRNA levels were adjusted

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according to GAPDH content, a housekeeping gene. Results are presented as percent inhibition of CD40, relative to cells treated with a scrambled control oligonucleotide (TCCATTATTAGTCTAGGAA (5-10-5 gapmer, where the gap segment comprises 2'-deoxynucleotides and each wing segment comprises 2'-MOE nucleotides. The oligonucleotide comprises phosphorothioate backbones (internucleoside linkages) and 5-methylcytosine substitutions throughout.

TABLE 6

Antisense Inhibition of human CD40 in AGS cells						
ISIS No	Motif	Wing segment	12.5 nM	25.0 nM	50.0 nM	Seq ID
26163	4-10-4	2'MOE	80	69	90	90
396236	5-10-5	2'MOE	83	86	94	208
—	4-10-4	2'OMe	51	54	66	90
—	5-10-5	2'OMe	60	63	69	208

As illustrated in Table 6, CD40 mRNA levels were reduced in a dose-dependent manner. Antisense oligonucleotides comprising 2'MOE wing segments are more active than those with 2'OMe wing segments.

Example 7

Antisense Inhibition of Murine CD40 In Vitro

Chimeric antisense oligonucleotides having 5-10-5 MOE wings and deoxy gap and 4-10-4 MOE wings and deoxy gap may be designed to target murine CD40. These antisense oligonucleotides can be evaluated for their ability to reduce CD40 mRNA in primary mouse hepatocytes using similar methods as described in the human in vitro study.

For example, primary mouse hepatocytes may be treated with 0.2344 nM, 0.4688 nM, 0.9375 nM, 1.875 nM, 3.75 nM, 7.5 nM, 15.0 nM, and 30.0 nM of antisense oligonucleotides for a period of approximately 24 hours. RNA can be isolated from the cells and CD40 mRNA levels can be measured by quantitative real-time PCR, as described herein. Murine CD40 primer probe sets can be used to measure mRNA levels. CD40 mRNA levels can then be adjusted according to total RNA content as measured by RIBOGREEN®.

Example 8

Antisense Inhibition of Murine CD40 In Vivo

Antisense oligonucleotides showing statistically significant dose-dependent inhibition from an in vitro study can be evaluated for their ability to reduce CD40 mRNA in vivo. Treatment

Antisense oligonucleotide can be evaluated in Balb/c mice and compared to a control group treated with saline. Oligonucleotide or saline would be administered subcutaneously at a dose of 5 mg/kg, 10 mg/kg, 25 mg/kg, or 50 mg/kg twice a week for three weeks. After the treatment period, whole liver can be collected for RNA analysis and protein analysis.

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RNA Analysis

Liver RNA can be isolated for real-time PCR analysis of CD40. It is theorized that an antisense oligonucleotide showing significant dose-dependent inhibition in vitro may show significant dose-dependent inhibition in vivo.

Protein Analysis

Liver CD40 protein may be measured by Western blot.

Example 9

Tolerability of Antisense Compounds in Rodents

Male 6 week old Balb/c mice were dosed subcutaneous 2× per week for 4 weeks with 25 or 50 mg/kg of antisense oligonucleotides Isis 26163 or Isis 396236. Mice were sacrificed 2 days following last administration. Body weights of the animals were monitored throughout the study. After sacrifice liver, spleen and kidney weights and liver enzymes ALT and AST from mouse plasma were determined.

Compared to a saline control treatment body weights of the mice are not affected by antisense oligonucleotides Isis 26163 or Isis 396236. Liver weight and spleen weight displayed a slight increase for Isis 26163 but not for Isis 396236. LFT (liver function test) elevations were small and within the normal range of high dose mouse studies.

Example 10

Antisense Inhibition of Human CD40 In Vitro on T24 Cells—Comparative Data for ISIS 26163 and ISIS19216

Antisense oligonucleotides ISIS 26163 and ISIS19216 targeted to a CD40 nucleic acid were tested for their effects on CD40 mRNA in vitro. The antisense oligonucleotides were designed as 4-10-4 gapmers, where the gap segment comprises 2'-deoxynucleotides and each wing segment comprises 2'-MOE nucleotides. The antisense oligonucleotides comprise phosphorothioate backbones (internucleoside linkages) and 5-methylcytosine substitutions throughout or in the wings, respectively.

T24 cells at a density of 7000 cells per well in a 96-well plate were treated with 100 nM or 150 nM, respectively, of antisense oligonucleotide. After a treatment period of approximately 24 hours, RNA was isolated from the cells and CD40 mRNA levels were measured by quantitative real-time PCR, as described herein. CD40 mRNA levels were adjusted according to GAPDH content, a housekeeping gene.

Results are presented in Table 7 as percent inhibition of CD40, relative to untreated control cells.

TABLE 7

Oligo ID	Target site	nM on T24 cells	% Inhibition	SEQ ID No.
26163	70-87	100	98	90
19216	73-90	150	66	10

Sequence ISIS 26163 shows a superior activity over ISIS19216, which overlaps the sequence 15 nucleobases.

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gactgggcag ggctcgca 18

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caggtctttg gtctcaca 18

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gtcttgttg tgctgcc 18

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ccacagaaa catcagtc 18

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ctggggacca cagacaac 18

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tcagccgatc ctggggac 18

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caccaccagg gctctcag 18

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gggatcacca ccagggct 18

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gaggatggca aacaggat 18

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ttttgataaa gaccagca 18

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tattggttgg cttcttgg 18

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gggttctctgc ttggggtg 18

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 aactgcctgt ttgccac 18

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agcagcccca gaggacgc 18

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gcggtcagca agcagccc 18

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ggttctggat ggacagcg 18

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<400> SEQUENCE: 94

agtgggtggt tctggatg 18

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<220> FEATURE:
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gcactgactg tttattag 18

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<400> SEQUENCE: 96

ggcacaaga acagcact 18

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tgtcctggct ggcacaaa 18

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cagtttctgt cctggctg 18

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gtcactcacc agtttctg 18

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<400> SEQUENCE: 101

ctttcaccgc aaggaagg 18

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<400> SEQUENCE: 102

tgtgtctctc tgttccag 18

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<400> SEQUENCE: 104

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tgaggtgccc ttctgctg 18

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<400> SEQUENCE: 115

acacattgga gaagaagc 18

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<400> SEQUENCE: 116

tgacacattg gagaagaa 18

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<400> SEQUENCE: 117

atgacacatt ggagaaga 18

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<400> SEQUENCE: 118

aaagcagatg acacattg 18

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<400> SEQUENCE: 119

aggtctttgg tctcacag 18

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ttgtttgtgc ctgcctgt 18

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tcttgttgt gcctgcct 18

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agtcttggtt gtgcctgc 18

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cagtcttggt tgtgcctg 18

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<400> SEQUENCE: 125

tcagtcttgt ttgtgcct 18

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catcagtctt gtttgtgc 18

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gaccacagac aacatcag 18

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gggaccacag acaacatc 18

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agaggatggc aaacagga 18

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<400> SEQUENCE: 133

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<400> SEQUENCE: 134

gaccagcacc aagaggat 18

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aagaccagca ccaagagg 18

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tgataaagac cagcacca 18

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tttgataaag accagcac 18

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actctctttg cccatcct 18

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cgactctctt tgcccatc 18

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<223> OTHER INFORMATION: Synthetic Oligonucleotide

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atgcgactct ctttgccc 18

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aaatgcgact ctctttgc 18

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ctgaaatgcg actctctt 18

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<223> OTHER INFORMATION: Synthetic Oligonucleotide

<400> SEQUENCE: 144

aactgaaatg cgactctc 18

<210> SEQ ID NO 145
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<400> SEQUENCE: 145

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<400> SEQUENCE: 147

aaccttcaact gtctctcc 18

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<210> SEQ ID NO 148
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gatcaccaca ggctctca 18

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<400> SEQUENCE: 149

tgataagaca gcaccaag 18

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<223> OTHER INFORMATION: Synthetic Oligonucleotide

<400> SEQUENCE: 150

ggtagttcct gccacttt 18

<210> SEQ ID NO 151
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<400> SEQUENCE: 151

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<210> SEQ ID NO 152
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<400> SEQUENCE: 152

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<223> OTHER INFORMATION: Synthetic Oligonucleotide

<400> SEQUENCE: 153

actgacacat ttgagcag 18

<210> SEQ ID NO 154
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gactccctac tgacacat 18

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<223> OTHER INFORMATION: Synthetic Oligonucleotide

<400> SEQUENCE: 155
caaagagcgg ttctccac 18

<210> SEQ ID NO 156
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<400> SEQUENCE: 156
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<210> SEQ ID NO 157
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<223> OTHER INFORMATION: Synthetic Oligonucleotide

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cccacctatc ttgacatc 18

<210> SEQ ID NO 159
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<400> SEQUENCE: 159
aggccgagag ttcaaaat 18

<210> SEQ ID NO 160
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<212> TYPE: DNA
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 <400> SEQUENCE: 161

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<210> SEQ ID NO 162
 <211> LENGTH: 20
 <212> TYPE: DNA
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 <220> FEATURE:
 <223> OTHER INFORMATION: Synthetic Oligonucleotide

 <400> SEQUENCE: 162

 cagaggacgc actgcagagg 20

<210> SEQ ID NO 163
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<210> SEQ ID NO 164
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 <212> TYPE: DNA
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 <400> SEQUENCE: 164

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<210> SEQ ID NO 165
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 <212> TYPE: DNA
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 <223> OTHER INFORMATION: Synthetic Oligonucleotide

 <400> SEQUENCE: 165

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<210> SEQ ID NO 166
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 <212> TYPE: DNA
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<210> SEQ ID NO 167
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 <212> TYPE: DNA
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 <223> OTHER INFORMATION: Synthetic Oligonucleotide

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aggttgaac acccaagata 20

<210> SEQ ID NO 168
<211> LENGTH: 20
<212> TYPE: DNA
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<220> FEATURE:
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<400> SEQUENCE: 168

ggagaaacc ctggtttctc 20

<210> SEQ ID NO 169
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<223> OTHER INFORMATION: Synthetic Oligonucleotide

<400> SEQUENCE: 170

tcaggtgaaa gtgaaagctg 20

<210> SEQ ID NO 171
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<212> TYPE: DNA
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<223> OTHER INFORMATION: Synthetic Oligonucleotide

<400> SEQUENCE: 171

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<210> SEQ ID NO 172
<211> LENGTH: 20
<212> TYPE: DNA
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<220> FEATURE:
<223> OTHER INFORMATION: Synthetic Oligonucleotide

<400> SEQUENCE: 172

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<210> SEQ ID NO 173
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<220> FEATURE:
<223> OTHER INFORMATION: Synthetic Oligonucleotide

<400> SEQUENCE: 173

gaaagaatac atgtatatgg 20

<210> SEQ ID NO 174
<211> LENGTH: 20
<212> TYPE: DNA
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<220> FEATURE:
<223> OTHER INFORMATION: Synthetic Oligonucleotide

<400> SEQUENCE: 174

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<210> SEQ ID NO 175
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<400> SEQUENCE: 175

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<400> SEQUENCE: 176

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<210> SEQ ID NO 177
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<212> TYPE: DNA
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<220> FEATURE:
<223> OTHER INFORMATION: Synthetic Oligonucleotide

<400> SEQUENCE: 177

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<212> TYPE: DNA
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<220> FEATURE:
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<400> SEQUENCE: 178

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<212> TYPE: DNA
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<223> OTHER INFORMATION: Synthetic Oligonucleotide

<400> SEQUENCE: 179

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<210> SEQ ID NO 180
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<220> FEATURE:
<223> OTHER INFORMATION: Synthetic Oligonucleotide

<400> SEQUENCE: 180

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<210> SEQ ID NO 181
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<400> SEQUENCE: 181

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<210> SEQ ID NO 182
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<400> SEQUENCE: 182

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<210> SEQ ID NO 183
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<212> TYPE: DNA
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<400> SEQUENCE: 183

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<210> SEQ ID NO 184
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<212> TYPE: DNA
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<220> FEATURE:
<223> OTHER INFORMATION: Synthetic Oligonucleotide

<400> SEQUENCE: 184

gctggcaca aagaacagcac 20

<210> SEQ ID NO 185
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<212> TYPE: DNA
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<220> FEATURE:
<223> OTHER INFORMATION: Synthetic Oligonucleotide

<400> SEQUENCE: 185

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<210> SEQ ID NO 186
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<212> TYPE: DNA
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<223> OTHER INFORMATION: Synthetic Oligonucleotide

<400> SEQUENCE: 186

tgtgcagtca ctcaccagtt 20

<210> SEQ ID NO 187
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<400> SEQUENCE: 187

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<212> TYPE: DNA

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<223> OTHER INFORMATION: Synthetic Oligonucleotide

<400> SEQUENCE: 193

cttcttcaca ggtgcagatg

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<400> SEQUENCE: 194

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gaagaagccg actgggcagg 20

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tgtctattac ctcaaagaga 20

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<400> SEQUENCE: 199

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<400> SEQUENCE: 200

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<220> FEATURE:
<223> OTHER INFORMATION: Synthetic Oligonucleotide

<400> SEQUENCE: 201

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<400> SEQUENCE: 202

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<220> FEATURE:
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<400> SEQUENCE: 204

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<210> SEQ ID NO 205
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<400> SEQUENCE: 207

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<220> FEATURE:
<223> OTHER INFORMATION: Synthetic Oligonucleotide

<400> SEQUENCE: 208

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<223> OTHER INFORMATION: Synthetic Oligonucleotide

<400> SEQUENCE: 214

gaatgccagg aaaggagtga 20

<210> SEQ ID NO 215
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<212> TYPE: DNA
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<400> SEQUENCE: 215

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<212> TYPE: DNA
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<220> FEATURE:
<223> OTHER INFORMATION: Synthetic Oligonucleotide

<400> SEQUENCE: 216

ctgcactgga gcagcagtgt 20

<210> SEQ ID NO 217
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<212> TYPE: DNA
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<223> OTHER INFORMATION: Synthetic Oligonucleotide

<400> SEQUENCE: 217

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<223> OTHER INFORMATION: Synthetic Oligonucleotide

<400> SEQUENCE: 220

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<223> OTHER INFORMATION: Synthetic Oligonucleotide

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tgggcaccaa actgctgat 20

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<223> OTHER INFORMATION: Synthetic Oligonucleotide

<400> SEQUENCE: 225

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<210> SEQ ID NO 226
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<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic Oligonucleotide

<400> SEQUENCE: 226

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<223> OTHER INFORMATION: Synthetic Oligonucleotide

<400> SEQUENCE: 227

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<210> SEQ ID NO 228
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<220> FEATURE:
<223> OTHER INFORMATION: Synthetic Oligonucleotide

<400> SEQUENCE: 228

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<212> TYPE: DNA
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<220> FEATURE:
<223> OTHER INFORMATION: Synthetic Oligonucleotide

<400> SEQUENCE: 229

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<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Synthetic Oligonucleotide

<400> SEQUENCE: 235

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<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Synthetic Oligonucleotide

<400> SEQUENCE: 236

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<213> ORGANISM: H. sapiens

<400> SEQUENCE: 237

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tctttgtgcc agccaggaca gaaactggtg agtgactgca cagagtccac tgaaacggaa 240

tgccttcctt gcggtgaaag cgaattccta gacacctgga acagagagac acacttcac 300

cagcaciaat actgcgacc caacctaggg cttcgggtcc agcagaagg cacctcagaa 360

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atccccatca tcttcgggat cctgtttgcc atctcttgg tgctggtctt tatcaaaaag 720

gtggccaaga agccaaccaa taaggcccc caccacaagc aggaaccca ggagatcaat 780

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<210> SEQ ID NO 238
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<213> ORGANISM: Artificial Sequence
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<223> OTHER INFORMATION: Synthetic Oligonucleotide

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<400> SEQUENCE: 238

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The invention claimed is:

1. A modified antisense compound 12 to 30 nucleobases in length and having a nucleobase sequence that is at least 90% complementary to an equal length portion of the human CD40 gene [but not to other sequences throughout the human genome], selected from the following regions of SEQ ID NO: 4:

- (a) positions [11250-12685, corresponding to intron 6] 11801-12591;
- (b) positions 2943-6367, corresponding to intron 1;
- (c) positions 6447-6780, corresponding to intron 2;
- (d) positions 6907-7157, corresponding to intron 3;
- (e) positions 7305-7673, corresponding to intron 4;
- (f) positions 7768-11187, corresponding to intron 5;
- (g) positions 12773-12877, corresponding to intron 7;
- (h) positions 12907-13429, corresponding to intron 8; and
- (i) positions 13662-16001, which forms part of exon 9 or a region 3' to exon 9.

[2. The antisense compound of claim 1, wherein the nucleobase sequence is at least 90% complementary to an equal length portion of positions 12527-12685 of SEQ ID NO: 4.]

3. The antisense compound of claim [2] 1, having a nucleobase sequence comprising at least 8 contiguous nucleobases of the nucleobase sequence of SEQ ID NO: 208, wherein the nucleobase sequence of the compound is at least 95% complementary to the sequence shown in SEQ ID NO: 4.

4. A modified antisense compound 20 nucleobases in length and consisting of the nucleobase sequence of SEQ ID NO: 208.

5. The antisense compound of claim 1, wherein said antisense compound is an antisense oligonucleotide.

6. The antisense compound of claim 5, wherein at least one internucleoside linkage is a modified internucleoside linkage.

7. The antisense compound of claim 6, wherein each internucleoside linkage is a phosphorothioate internucleoside linkage.

8. The antisense compound of claim 5, wherein at least one nucleoside comprises a modified sugar.

9. The antisense compound of claim 8, wherein at least one modified sugar is a bicyclic sugar.

10. The antisense compound of claim 9, wherein the at least one bicyclic sugar comprises a 4'-CH(CH₃)—O-2' bridge.

11. The antisense compound of claim 8, wherein at least one modified sugar comprises a 2'-O-methoxyethyl.

12. The antisense compound of claim 1, wherein at least one said nucleobase is a modified nucleobase.

13. The antisense compound of claim 12, wherein the modified nucleobase is a 5-methylcytosine.

14. The antisense compound of claim 1, wherein the compound is an oligonucleotide comprising:

- a gap segment consisting of linked deoxynucleosides;
- a 5' wing segment consisting of linked nucleosides;
- a 3' wing segment consisting of linked nucleosides;

wherein the gap segment is positioned between the 5' wing segment and the 3' wing segment and wherein each nucleoside of each wing segment comprises a modified sugar.

15. The antisense compound of claim 14, wherein the oligonucleotide comprises:

- a gap segment consisting of ten linked deoxynucleosides;
 - a 5' wing segment consisting of five linked nucleosides;
 - a 3' wing segment consisting of five linked nucleosides;
- wherein the gap segment is positioned between the 5' wing segment and the 3' wing segment, wherein each nucleoside of each wing segment comprises a 2'-O-methoxyethyl sugar;

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and wherein each internucleoside linkage of said antisense compound is a phosphorothioate linkage.

16. The antisense compound of claim **14**, wherein the oligonucleotide comprises:

a gap segment consisting of fifteen linked deoxynucleosides;

a 5' wing segment consisting of two linked nucleosides;

a 3' wing segment consisting of three linked nucleosides;

wherein the gap segment is positioned between the 5' wing segment and the 3' wing segment, wherein each nucleoside of each wing segment comprises a 2'-O-methoxyethyl sugar;

and wherein each internucleoside linkage of said antisense compound is a phosphorothioate linkage.

17. The antisense compound of claim **15** or **16**, wherein every cytosine is a 5-methylcytosine.

18. An antisense oligonucleotide 20 nucleobases in length having the sequence of nucleobases as set forth in SEQ ID NO:208, wherein each cytosine is a 5-methylcytosine, each

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internucleoside linkage is a phosphorothioate linkage, nucleotides 1-5 and 16-20 are 2'-O-methoxyethyl nucleotides, and nucleotides 6-15 are 2'-deoxynucleotides.

19. A composition comprising an antisense compound of claim **1** or **18** or a salt thereof and a pharmaceutically acceptable carrier or diluent.

20. A method comprising administering to an animal an antisense compound of claim **1** or an oligonucleotide of claim **18**.

21. *The antisense compound of claim 1, wherein said antisense compound is 15, 16, 17, 18, 19, 20 or 21 nucleobases in length.*

22. *The antisense compound of claim 21, wherein the nucleobase sequence of the compound is at least 95% complementary to the sequence shown in SEQ ID NO: 4.*

23. *The antisense compound of claim 21, wherein the nucleobase sequence of the compound is 100% complementary to the sequence shown in SEQ ID NO: 4.*

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