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(54) **DEVELOPMENT OF MUTATIONS USEFUL FOR ATTENUATING DENGUE VIRUSES AND CHIMERIC DENGUE VIRUSES**

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**A61K 39/12** (2006.01)

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USPC ..... **424/218.1**

(58) **Field of Classification Search**  
None  
See application file for complete search history.

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(57) **ABSTRACT**

A menu of mutations was developed that is useful in fine-tuning the attenuation and growth characteristics of dengue virus vaccines.

**13 Claims, 12 Drawing Sheets**

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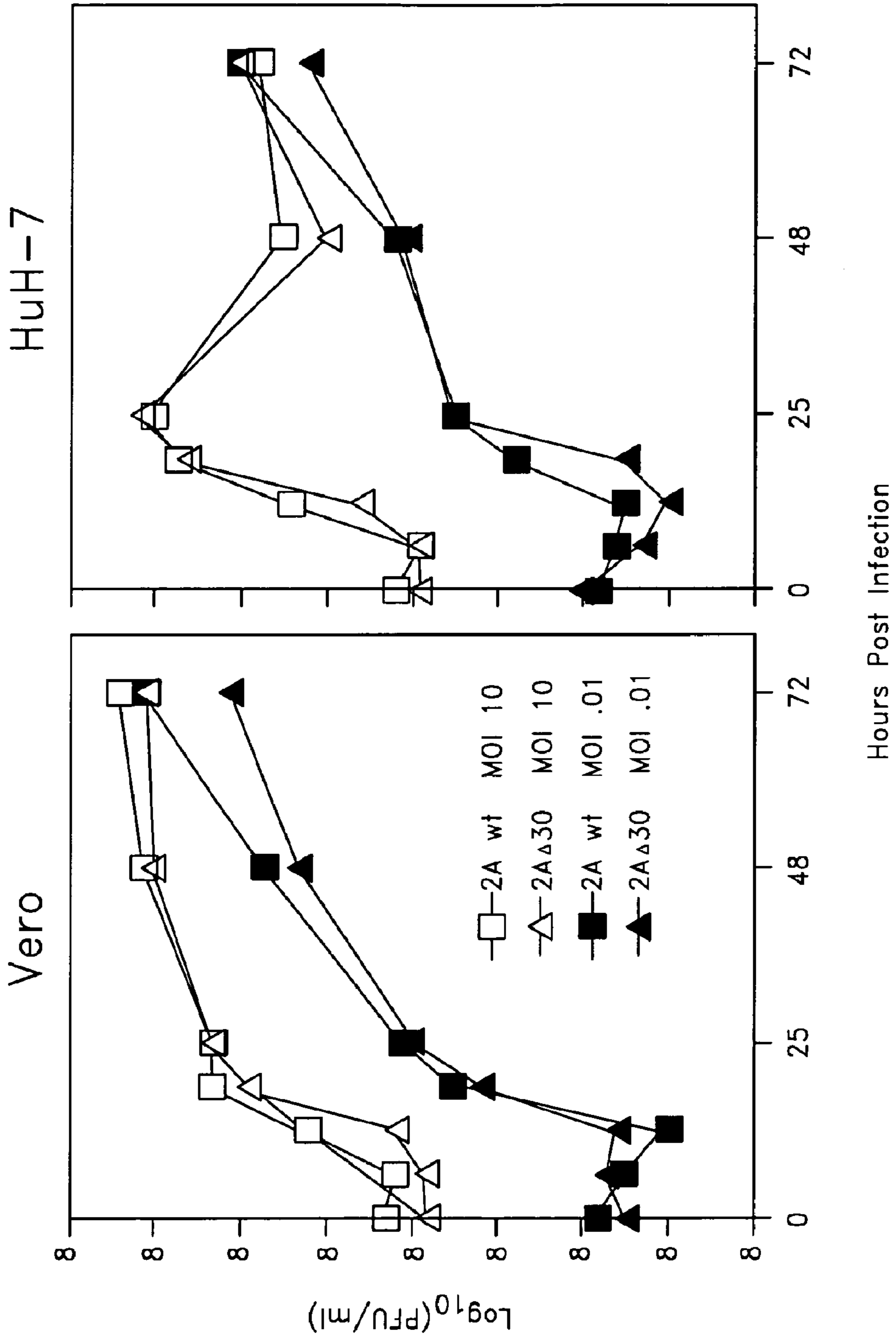


FIG. 1

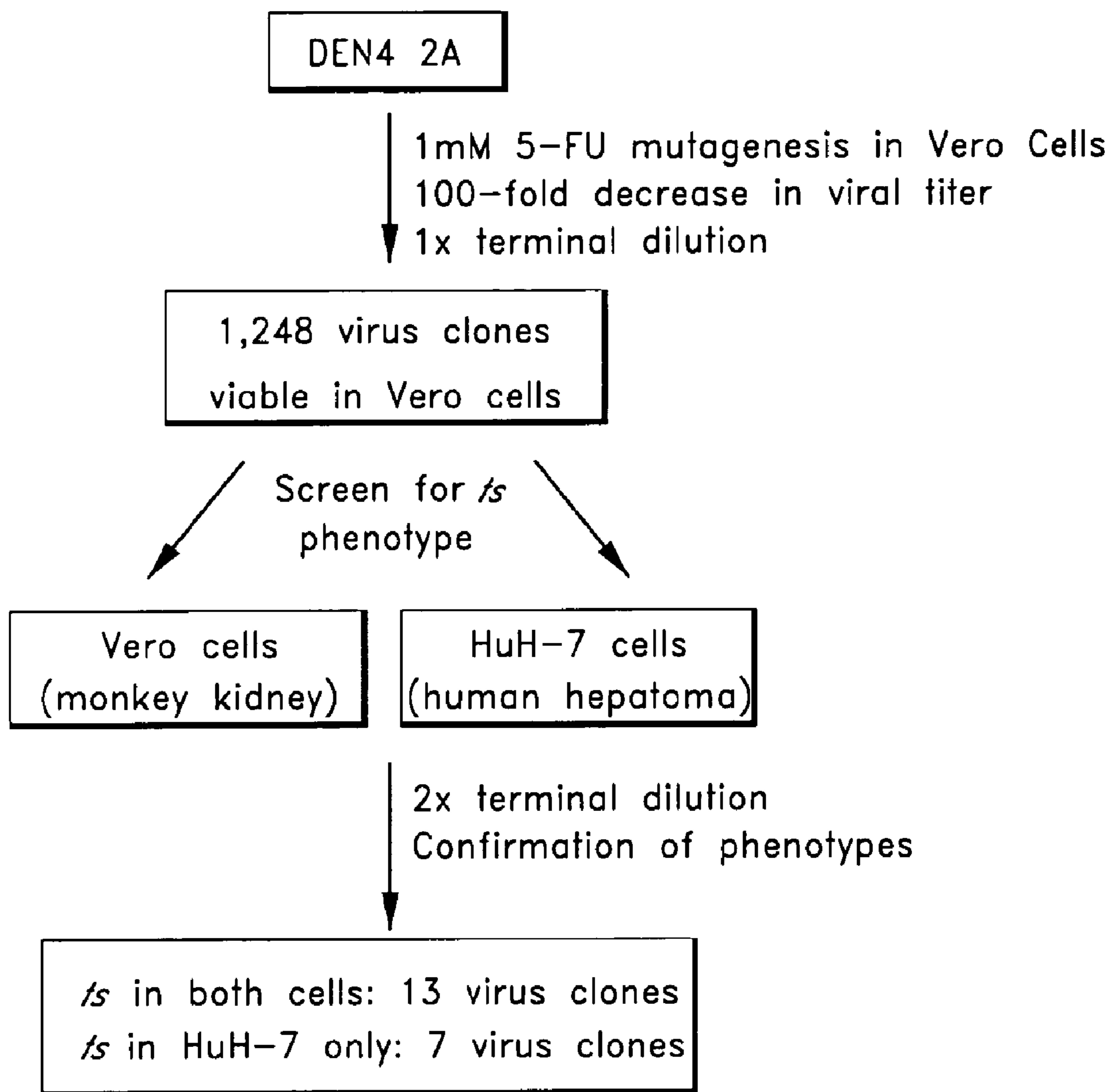


FIG. 2

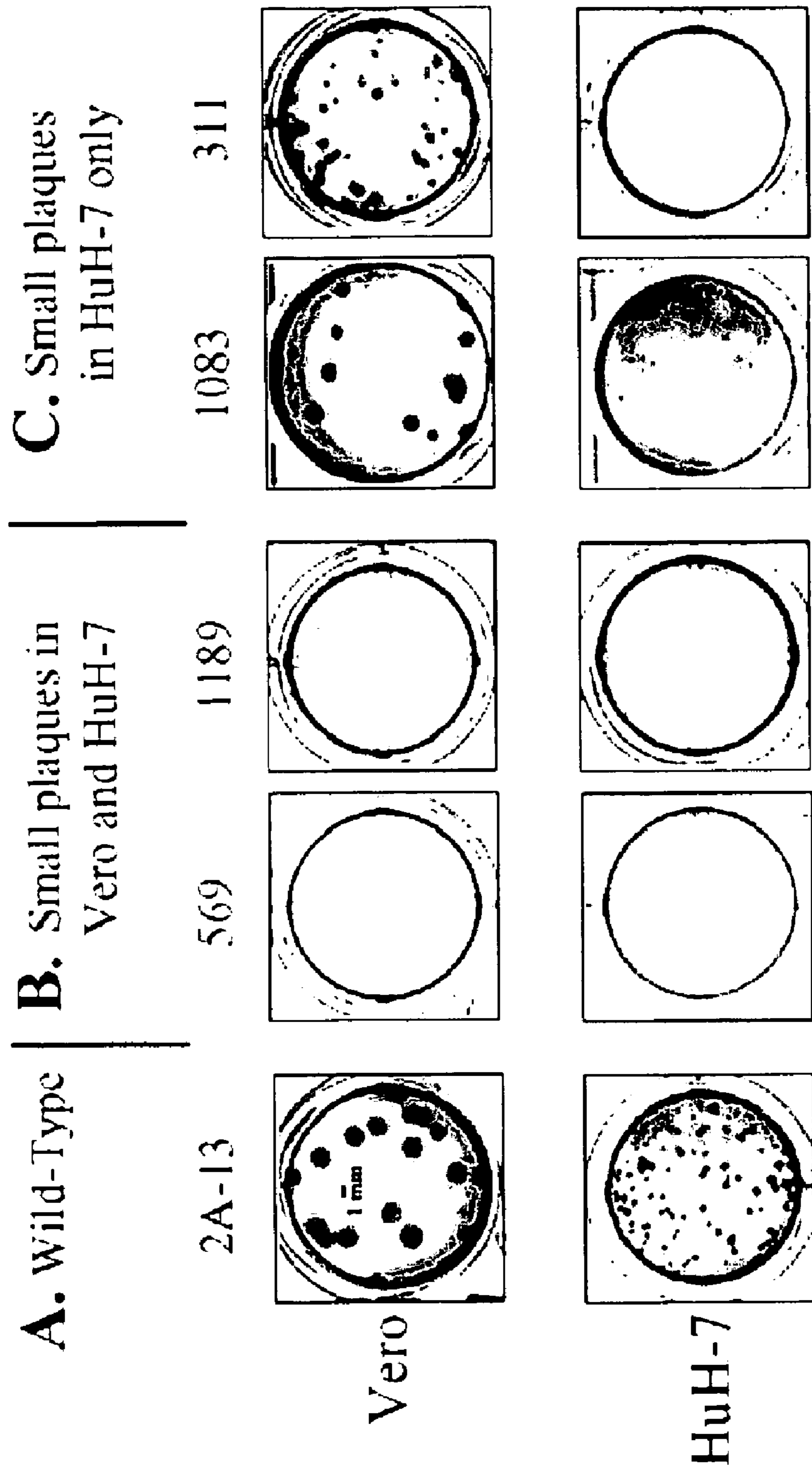


FIG. 3

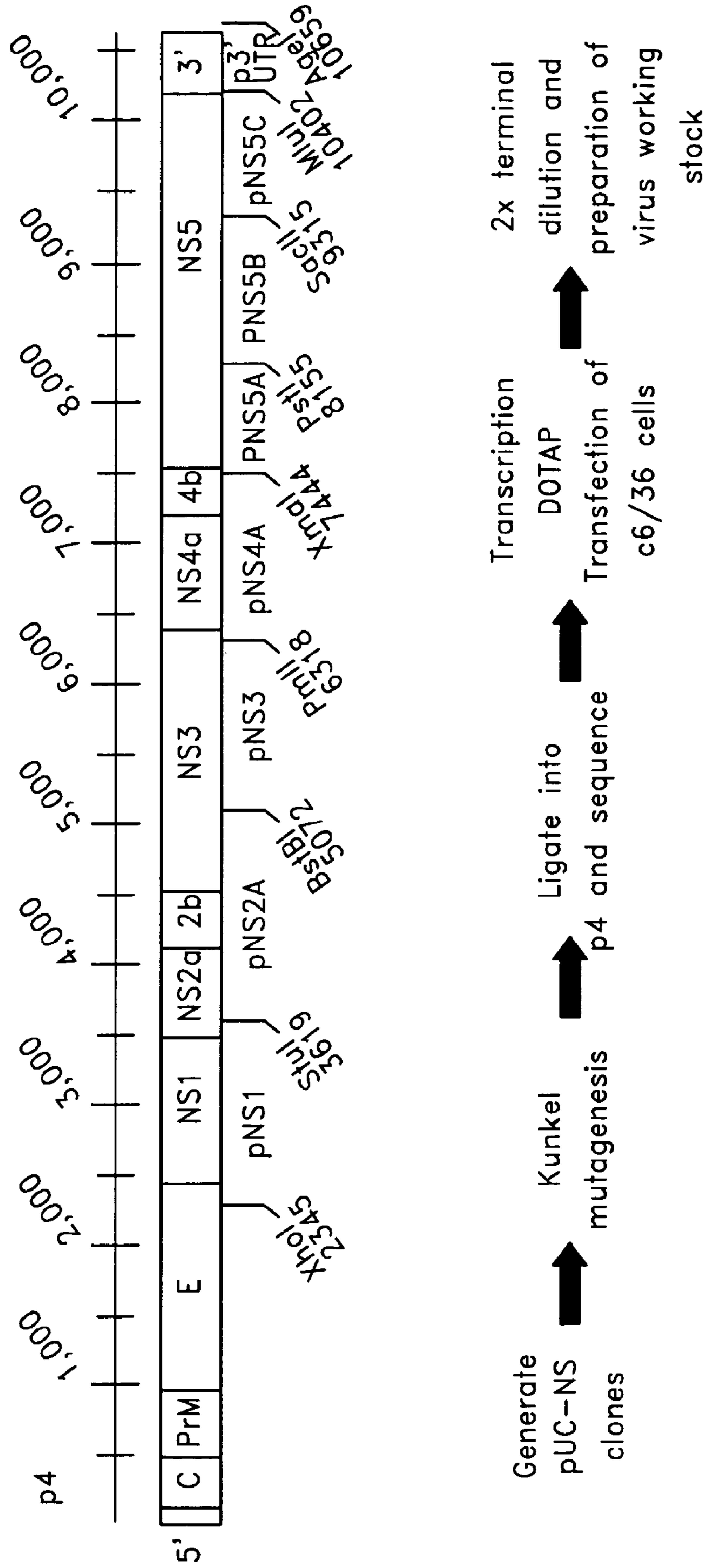


FIG. 4



1 GTGTTGETLG EKGKQQLNSL DRKEFEEYKR SGILEVDRTE AKSALKDGSK  
51 IKHAVSRGSS KIRWIVERGM VKPKGKVVDL GCGRGGWSYY MATLKNVTEV  
101 KGYTKGGPGH EEPIPNATYG WNLVKLHSGV DVFKPTEQV DTLLQDIGES  
151 SSNFTEEGR TLRVLKMEP WLSSKPEFCI KVLNPYMPTV IEELEKLQRK  
201 HGNLVRCPL SRNSTHEMYW VSGASGNIYS SVNTSKMLL NRFTTRERKP  
251 TYEKDVLGA GTRSVSTETE KPDMTIIGRR LQRLQEEHKE TWHYDQENPY  
301 RTWAYHGSYE APSTGSASSM VNGVKLLTK PNDVIPMVTQ LAMDTTTFFG  
351 QQRVFEKVD TRTEQFKPGT RMVMTTIANW LWALLKKKN FRLCTRBFFI  
401 SKVRSNAAG AVFQEEGGWT SASEAVNDSR FWBLVDKERA LHQEGKCBSC  
451 VYNNMGKREK KLGEFGRAGK SRAIWYMWLG ARFLEALG FLNEDHWFGR  
501 ENSWSGVEGE GLHRLGVILE EIDKKDGLM YADDTAGNDT RITEDDLQNE  
551 ELITEQMAPH HKILAKAIFK LTYQNKVVKV LRPTPRGAVM DIISRKDQRG  
601 SGQVGTYGLN TFTNNEVQLI RQMEREGVIT QDDMQPKGL KERVEKWLKE  
651 CGVDRLLKRNA ISGDDCVVKP LDERFGTSLI FLNDMGKVRK DIPQWBPSSK  
701 WKNWQEVVFC SHFHKIFMK DGRSLVVPCR NQDELIGRAR ISQGAGWSLR  
751 ETACLGKAYA QMWSLMYFHR RDLRLASNAI CSAVPTWFFP TSRTTWSIHA  
801 HHQMTTEDM LKVNRVWIE DNPNMTDKTP VHSWBDIPYL GKREDLWCGS  
851 LIGLSSRATW AKNIHTAITQ VRNLIGKEEY VDYMVPVKRY SAPSESBQVL

SAM

Importin-  
binding +  
NLS

Polymerase

FIG. 5

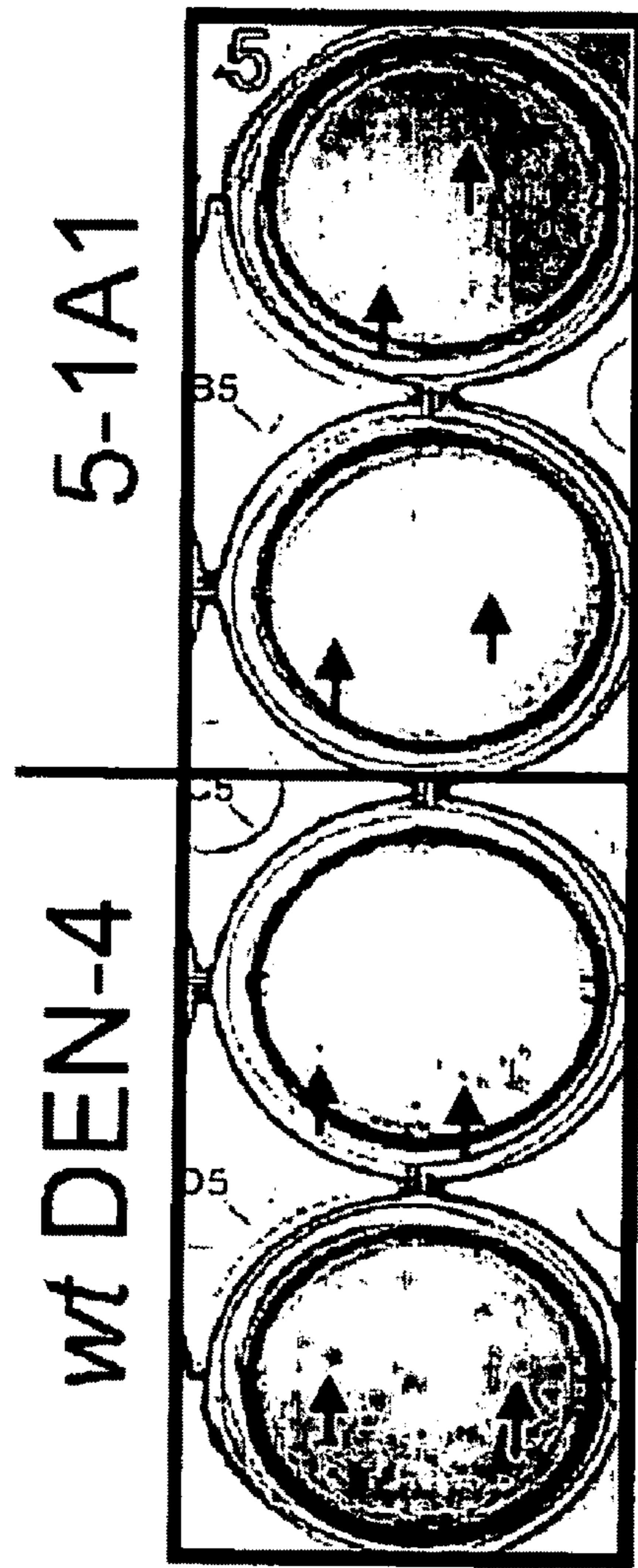


FIG. 6

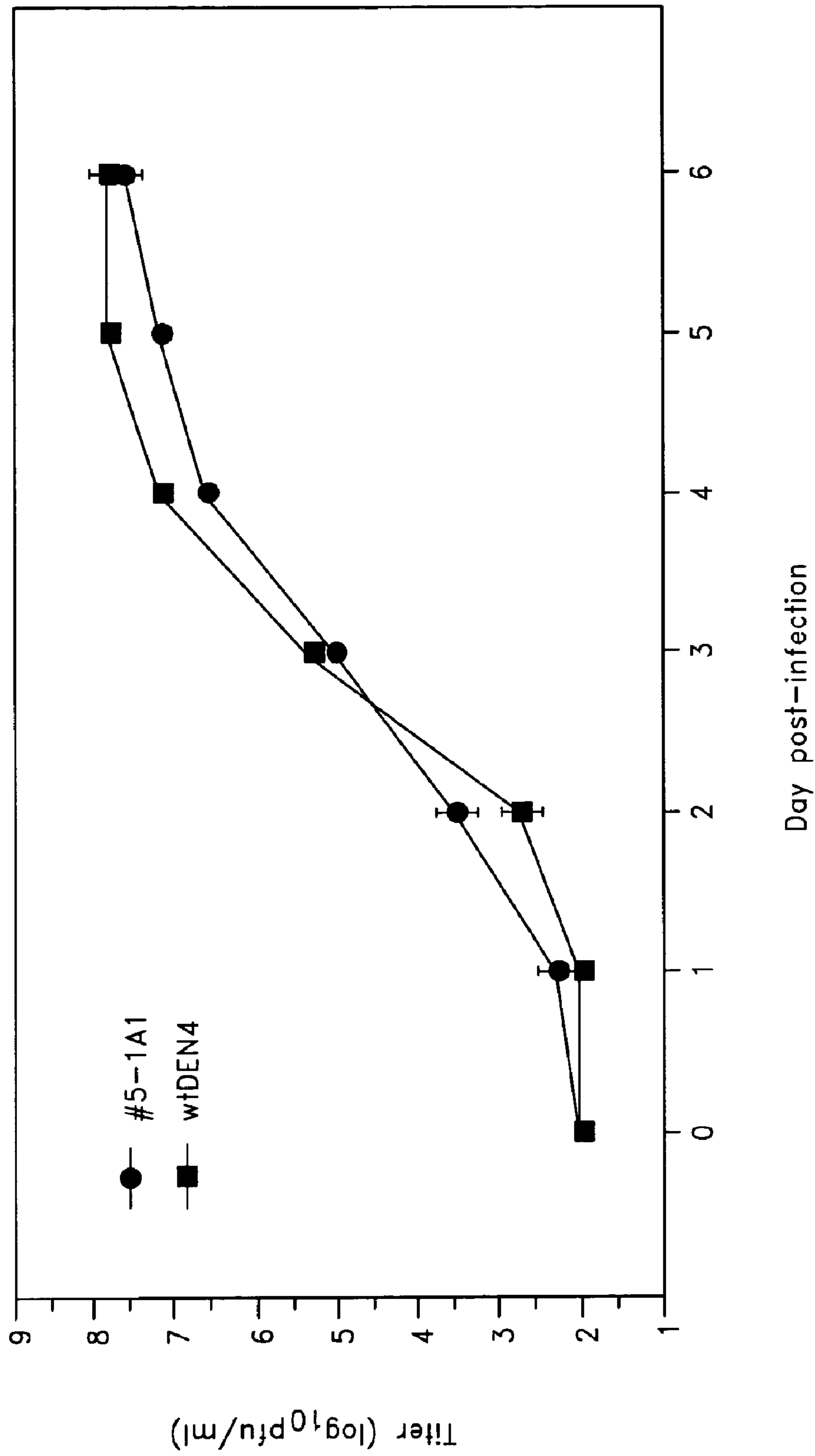


FIG. 7

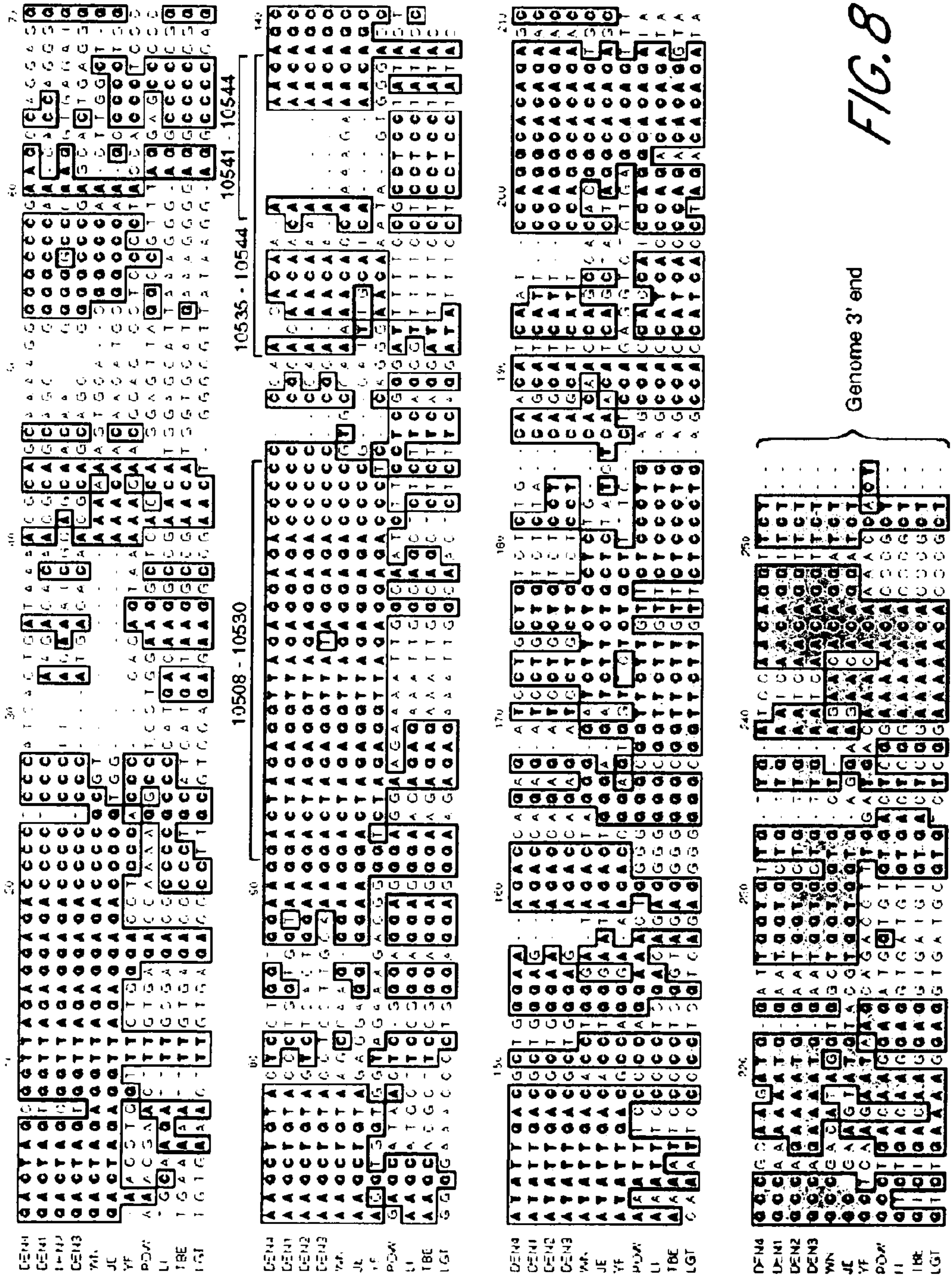


FIG. 8

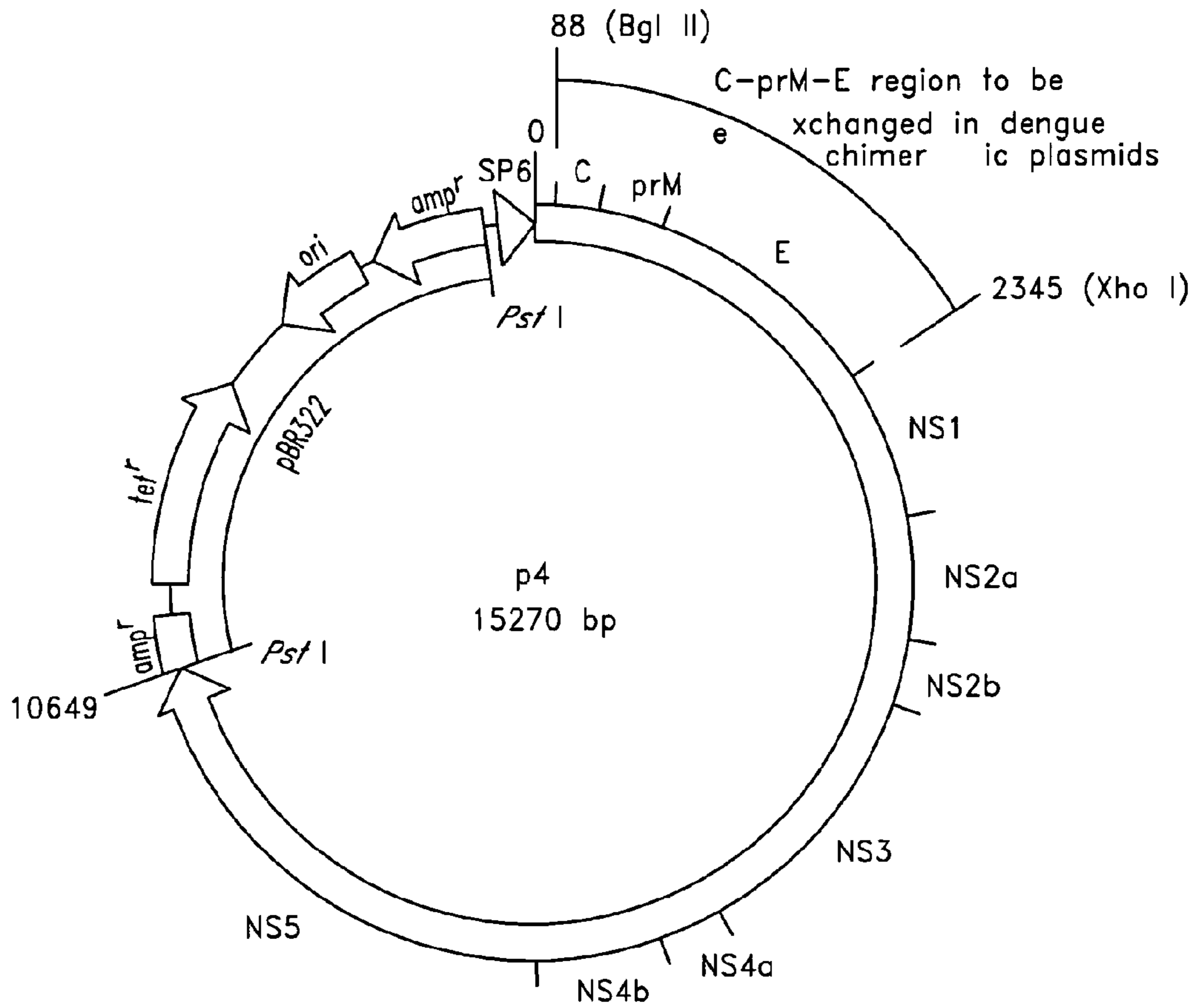


FIG. 9

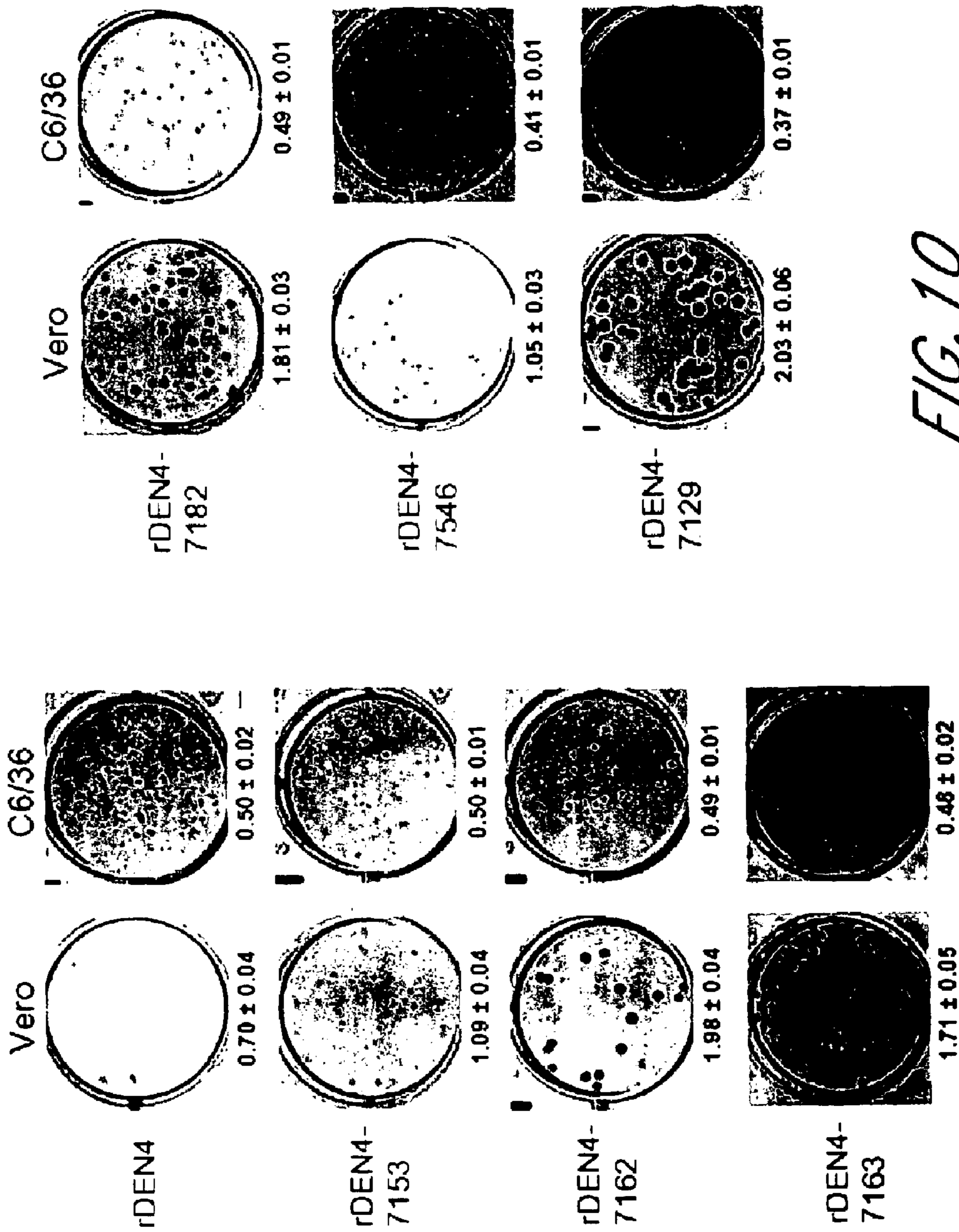


FIG. 10

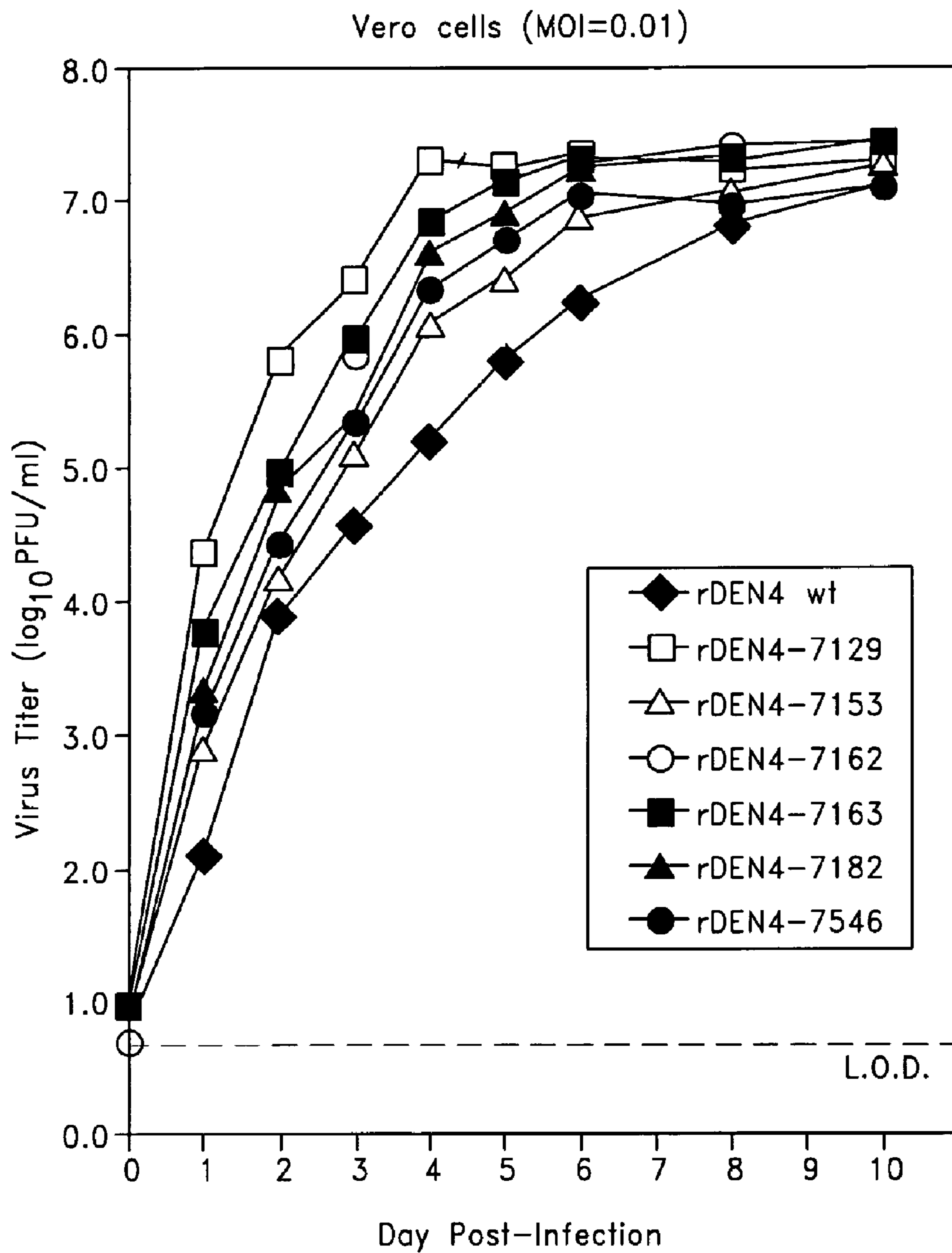
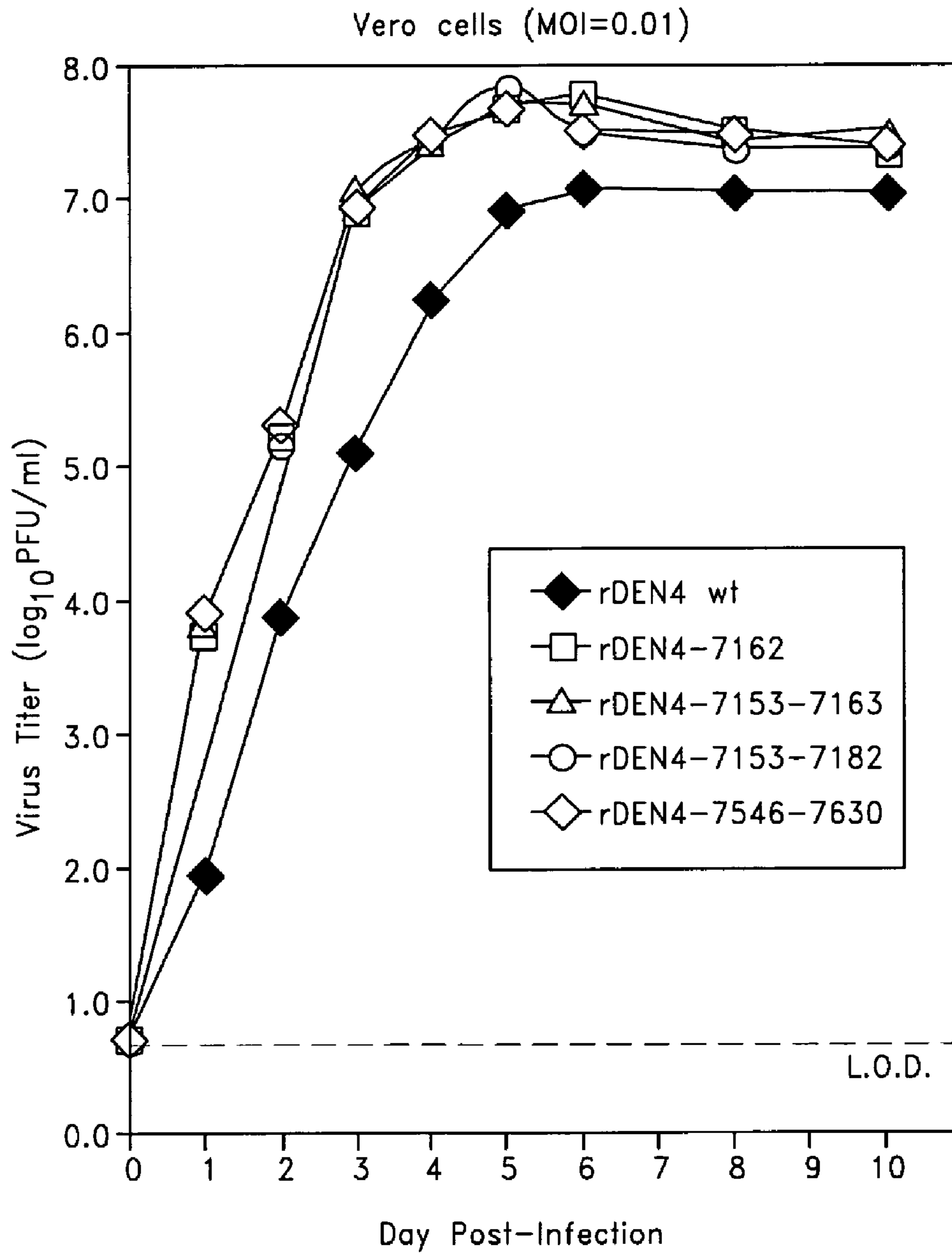


FIG. 11



*FIG. 12*



**DEVELOPMENT OF MUTATIONS USEFUL  
FOR ATTENUATING DENGUE VIRUSES AND  
CHIMERIC DENGUE VIRUSES**

**Matter enclosed in heavy brackets [ ] appears in the original patent but forms no part of this reissue specification; matter printed in italics indicates the additions made by reissue.**

RELATED APPLICATIONS

This application is a continuation and claims the benefit of priority of International Application No. PCT/JUS02/16308 filed May 22, 2002, designating the United States of America and published in English as WO 02/095075 on Nov. 28, 2002, which claims the benefit of priority of U.S. Provisional Application No. 60/293,049 filed May 22, 2001, both of which are hereby expressly incorporated by reference in their entireties.

FIELD OF THE INVENTION

A menu of mutations was developed that is useful in fine-tuning the attenuation and growth characteristics of dengue virus vaccines.

BACKGROUND OF THE INVENTION

Dengue virus is a positive-sense RNA virus belonging to the Flavivirus genus of the family Flaviviridae. Dengue virus is widely distributed throughout the tropical and semitropical regions of the world and is transmitted to humans by mosquito vectors. Dengue virus is a leading cause of hospitalization and death in children in at least eight tropical Asian countries (WHO, 1997. *Dengue haemorrhagic fever: diagnosis, treatment prevention and control*—2nd ed. Geneva: WHO). There are four serotypes of dengue virus (DEN-1, DEN-2, DEN-3, and DEN-4) which annually cause an estimated 50-100 million cases of dengue fever and 500,000 cases of the more severe form of dengue virus infection, dengue hemorrhagic fever/dengue shock syndrome (DHF/DSS) (Gubler, D. J. & Meltzer, M. 1999 *Adv Virus Res* 53:35-70). DHF/DSS is seen predominately in children and adults experiencing a second dengue virus infection with a serotype different than that of their first dengue virus infection and in primary infection of infants who still have circulating dengue-specific maternal antibody (Burke, D. S. et al. 1988 *Am J Trop Med Hyg* 38:172-80; Halstead, S. B. et al. 1969 *Am J Trop Med Hyg* 18:997-1021; Thein S. et al. 1997 *Am J Trop Med Hyg* 56:566-72). A vaccine is needed to lessen the disease burden caused by dengue virus, but none is licensed. Because of the association of more severe disease with secondary dengue virus infection, a successful vaccine must induce immunity to all four serotypes. Immunity is primarily mediated by neutralizing antibody directed against the envelope E glycoprotein, a virion structural protein. Infection with one serotype induces long-lived homotypic immunity and a short-lived heterotypic immunity (Sabin, A. 1955 *Amer J Trop Med Hyg* 4:198-207). Therefore, the goal of immunization is to induce a long-lived neutralizing antibody response against DEN-1, DEN-2, DEN-3, and DEN-4, which can best be achieved economically using live attenuated virus vaccines. This is a reasonable goal since a live attenuated vaccine has already been developed for the related yellow fever virus, another mosquito-borne flavivirus present in tropical and semitropical regions of the world (Monath, T. P.

& Heinz, F. X. 1996 in: Fields B. N. et al. eds. *Fields Virology* Philadelphia: Lippincott-Ravan Publishers, 961-1034).

Several live attenuated dengue vaccine candidates have been developed and evaluated in humans or non-human primates. The first live attenuated dengue vaccine candidates were host range mutants developed by serial passage of wild type dengue viruses in the brains of mice and selection of mutants attenuated for humans (Kimura, R. & Hotta, S. 1944 *Japanese J Bacteriology* 1:96-99; Sabin, A. B. & Schlesinger, R. W. 1945 *Science* 101:640; Wisseman, C. L. Jr. et al. 1963 *Am J Trop Med* 12:620-623). Although these candidate vaccine viruses were immunogenic in humans, their poor growth in cell culture discouraged further development. Additional live attenuated DEN-1, DEN-2, DEN-3, and DEN-4 vaccine candidates have been developed by serial passage in tissue culture (Angsubhakorn, S. et al. 1994 *Southeast Asian J Trop Med Public Health* 25:554-9; Bancroft, W. H. et al. 1981 *Infect Immun* 31:698-703; Bhamarapravati, N. et al. 1987 *Bull World Health Organ* 65:189-95; Eckels, K. H. et al. 1984 *Am J Trop Med Hyg* 33:684-9; Hoke, C. H. Jr. et al. 1990 *Am J Trop Med Hyg* 43:219-26; Kanesa-thasan, N. et al. 2001 *Vaccine* 19:3179-88) or by chemical mutagenesis (McKee, K. T. Jr. et al. 1987 *Am J Trop Med Hyg* 36:435-42). It has proven very difficult to achieve a satisfactory balance between attenuation and immunogenicity for each of the four serotypes of dengue virus using these approaches and to formulate a tetravalent vaccine that is safe and satisfactorily immunogenic against each of the four dengue viruses (Kanesa-thasan, N. et al. 2001 *Vaccine* 19:3179-88; Bhamarapravati, N. & Sutee, Y. 2000 *Vaccine* 18 Suppl 2: 44-7).

Two major advances utilizing recombinant DNA technology have recently made it possible to develop additional promising live attenuated dengue virus vaccine candidates. First, methods have been developed to recover infectious dengue virus from cells transfected with RNA transcripts derived from a full-length cDNA clone of the dengue virus genome, thus making it possible to derive infectious viruses bearing attenuating mutations which have been introduced into the cDNA clone by site-directed mutagenesis (Lai, C. J. et al. 1991 *PNAS USA* 88:5139-43). Second, it is possible to produce antigenic chimeric viruses in which the structural protein coding region of the full-length cDNA clone of dengue virus is replaced by that of a different dengue virus serotype or from a more divergent flavivirus (Bray, M. & Lai, C. J. 1991 *PNAS USA* 88: 10342-6; Chen, W. et al. 1995 *J Virol* 69:5186-90; Huang, C. Y. et al. 2000 *J Virol* 74:3020-8; Pletnev, A. G. & Men, R. 1998 *PNAS USA* 95:1746-51). These techniques have been used to construct intertypic chimeric dengue viruses which have been shown to be effective in protecting monkeys against homologous dengue virus challenge (Bray, M. et al. 1996 *J Virol* 70:4162-6). Despite these advances, there is a need to develop attenuated antigenic dengue virus vaccines that specify a satisfactory balance between attenuation and immunogenicity for humans.

SUMMARY OF THE INVENTION

The invention provides mutations that confer temperature sensitivity in Vero cells or human liver cells, host-cell restriction in mosquito or human liver cells, host-cell adaptation for improved replication in Vero cells, or attenuation in mice, which mutations are useful in fine tuning the attenuation and growth characteristics of dengue virus vaccines.

BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1 shows growth of wt DEN4 2A and vaccine candidate, 2AΔ30, in Vero and HuH-7 cells. Vero (A) or HuH-7 (B)

## 3

cells were infected with DEN4 2A or 2AΔ30 at a multiplicity of infection (MOI) of 10 or 0.01. Confluent cell monolayers in 25-mm tissue culture flasks were washed and overlaid with a 1.5 ml inoculum containing the indicated virus. After a two hour incubation at 37° C., cells were washed three times in PBS and 7 ml of culture media supplemented with 2% FBS was added. A 1 ml aliquot of tissue culture medium was removed, replaced with fresh medium, and designated the 0 hour time-point. At the indicated time points post-infection, samples of tissue culture media were removed and frozen at -70° C. The level of viral replication was assayed by plaque titration in Vero cells. Briefly, serial ten-fold dilutions of cell culture media samples were inoculated onto confluent Vero cell monolayers in 24-well plates in duplicate and overlaid with OptiMEM containing 0.8% methylcellulose. After five days, plaques were visualized by immunoperoxidase staining as described in Example 1.

FIG. 2 shows generation of temperature-sensitive (ts) DEN4 viruses by 5-fluorouracil (5-FU) chemical mutagenesis. The wild-type DEN4 2A virus was derived from a cDNA clone of DEN4 strain 814669 (Dominica, 1981). Vero cells were infected with DEN4 2A and overlaid with culture media containing 1 mM 5-fluorouracil (5-FU) which resulted in a reduction of approximately 100-fold in viral replication when compared to untreated controls. Viral progeny from the 1 mM 5-FU-treated cultures were subjected to a single round of terminal dilutions generating 1,248 biologically cloned viruses which were screened for ts phenotypes by assessing virus replication at 35° C. and 39° C. in Vero and HuH-7 cells. Virus clones which demonstrated a 100-fold or greater reduction in titer at 39° C. were terminally diluted an additional two times and amplified in Vero cells. Temperature-sensitive phenotypes of the 3× biologically cloned viruses were confirmed by evaluating efficiency of plaque formation (EOP) in the indicated cells as described in Example 1.

FIG. 3 shows plaque size phenotypes of representative 5-FU mutant DEN4 viruses. Serial ten-fold dilutions of wild-type DEN4 2A-13 (A), 5-FU mutant viruses #569 and #1189 (B), and 5-FU mutant viruses #1083 and #311 (C) were inoculated onto confluent Vero and HuH-7 cell monolayers in 24-well plates. After incubation at 35° C. for two hours, monolayers were overlaid with 0.8% methylcellulose culture media. Following incubation at 35° C. for five days, plaques were visualized by immunoperoxidase staining. Viruses which had a plaque size that was ≤1 mm (approximately ≤50% the size of wt DEN4 2A-13) at the permissive temperature of 35° C. were designated as having the small-plaque (sp) phenotype. Mutant viruses #569 and #1189 (B) were sp in both Vero and HuH-7 cells, and #311 and #1083 (C) were sp in only HuH-7 cells.

FIG. 4 shows generation of recombinant DEN4 viruses. (A), The p4 cDNA clone is represented which was constructed from the 2A cDNA clone (derived from DEN4 814669) by site-directed mutagenesis. Restriction enzyme sites were introduced or removed to facilitate subsequent cloning of DEN4 recombinants bearing introduced attenuating mutations. Restriction enzyme sites are shown and define fragments of the genome that were sub-cloned into modified pUC-119 vectors for site-directed mutagenesis to introduce mutations identified in the 5-FU mutant viruses. (B), An outline of the methods used to generate rDEN4 viruses is also represented and described in Example 1.

FIG. 5 shows amino acid sequence of the rDEN4 NS5 gene (SEQ ID NO: 1). Eighty underlined amino acid pairs were mutagenized to alanine pairs; 32 pairs in boldface represent mutant viruses that could be recovered in either Vero or C6/36 cells; pairs in normal type represent mutant viruses that could

## 4

not be recovered in either Vero or C6/36 cells. Boxed regions indicate putative functional domains, including an S-adenosylmethionine utilizing methyltransferase domain (SAM), an importin-β binding domain adjacent to a nuclear localization sequence (importin-β-binding+NLS) and an RNA-dependent RNA polymerase domain (Polymerase).

FIG. 6 shows plaque size of mutant 5-1A1 in C6/36 cells. Note that 5-1A1 has a small plaque phenotype in C6/36 cells relative to that of the wild type virus.

FIG. 7 shows growth of wild type rDEN4 and 5-1A1 in C6/36 cells. Cells were inoculated in triplicate with each virus at an MOI of 0.01, and the amount of virus present in the supernatants that were harvested on the indicated days was determined by plaque enumeration in Vero cells. The titers are expressed as log<sub>10</sub> PFU/ml ± standard error.

FIG. 8 shows nucleotide alignment of the 3' UTR of mosquito-borne and tick-borne flaviviruses. cDNA sequences are shown 5' to 3' and represent a portion of the UTR corresponding to DEN4 nucleotides 10417 to 10649 (3' genome end). Nucleotide numbering represents the position in the alignment. Regions deleted or swapped are indicated using the nucleotide numbering of DEN4. GenBank accession numbers for mosquito-borne viruses: DEN4 (SEQ ID NO: 2): AF326825, DEN1 (SEQ ID NO: 3): U88535, DEN2 (SEQ ID NO: 4): AF038403, DEN3 (SEQ ID NO: 5): M93130, West Nile virus (WN) (SEQ ID NO: 6): M12294, Japanese encephalitis virus (JE) (SEQ ID NO: 7): AF315119, Yellow fever virus (YF) (SEQ ID NO: 8): U17067; GenBank accession numbers for tick-borne viruses: Powassan virus (POW) (SEQ ID NO: 9): L06436, Louping Ill virus (LI) (SEQ ID NO: 10): Y07863, Tick-borne encephalitis virus (TBE) (SEQ ID NO: 11): U27495, and Langat virus (LGT) (SEQ ID NO: 12): AF253419.

FIG. 9 shows genetic map of plasmid p4. Dengue cDNA is shown as bold line, with the C-prM-E region exchanged during construction of chimeric dengue virus cDNAs indicated.

FIG. 10 shows plaque size phenotypes of rDEN4 viruses encoding Vero adaptation mutations. Serial three-fold dilutions of the indicated viruses were inoculated onto confluent Vero and C6/36 cell monolayers in 6-well plates. After incubation at 37° C. (Vero) or 32° C. (C6/36) for two hours, monolayers were overlaid with 0.8% methylcellulose culture media. Following incubation for five days, plaques were visualized by immunoperoxidase staining. Values below each well are the average plaque size in mm standard error. For each of the virus-infected wells, 36 plaques were measured on the digital image of the 6-well plate on Adobe Photoshop at 300% view.

FIG. 11 shows growth curve in Vero cells of rDEN4 viruses encoding single Vero adaptation mutations. Vero cells were infected with the indicated viruses at an MOI of 0.01. Confluent cell monolayers in 25-cm<sup>2</sup> tissue culture flasks were washed and overlaid with a 1.5 ml inoculum containing the indicated virus. After a two hour incubation at 37° C., cells were washed three times in PBS and 5 ml of culture medium supplemented with 2% FBS was added. A 1 ml aliquot of tissue culture medium was removed, replaced with fresh medium, and designated the 0 hour time-point. At the indicated time points post-infection, samples of tissue culture medium were removed, clarified, and frozen at -70° C. The level of virus replication was assayed by plaque titration in Vero cells. Briefly, serial ten-fold dilutions of cell culture media samples were inoculated onto confluent Vero cell monolayers in 24-well plates in duplicate and overlaid with Opti-MEM containing 0.8% methylcellulose. After five days,

plaques were visualized by immunoperoxidase staining as described in Example 1. Limit of detection (L.O.D.) is  $\geq 0.7 \log_{10}$  PFU/ml.

FIG. 12 shows growth curve in Vero cells of rDEN4 viruses encoding combined Vero cell adaptation mutations. Vero cells were infected with the indicated viruses at an MOI of 0.01. Confluent cell monolayers in 25-cm<sup>2</sup> tissue culture flasks were washed and overlaid with a 1.5 ml inoculum containing the indicated virus. After a two hour incubation at 37° C., cells were washed three times in PBS and 5 ml of culture medium supplemented with 2% FBS was added. A 1 ml aliquot of tissue culture medium was removed, replaced with fresh medium, and designated the 0 hour time-point. At the indicated time points post-infection, samples of tissue culture medium were removed, clarified, and frozen at -70° C. The level of virus replication was assayed by plaque titration in Vero cells. Limit of detection (L.O.D.) is  $\geq 0.7 \log_{10}$  PFU/ml.

#### BRIEF DESCRIPTION OF THE TABLES

Table 1. Susceptibility of mice to intracerebral DEN4 infection is age-dependent.

Table 2. Temperature-sensitive (ts) and mouse brain attenuation (att) phenotypes of 5-FU mutant DEN4 viruses.

Table 3. Nucleotide and amino acid differences of the 5-FU mutant viruses which are ts in both Vero and HuH-7 cells.

Table 4. Nucleotide and amino acid differences of the 5-FU mutant viruses which are ts in only HuH-7 cells.

Table 5. Mutations which are represented in multiple 5-FU mutant DEN4 viruses.

Table 6. Addition of ts mutation 4995 to rDEN4 $\Delta$ 30 confers a ts phenotype and further attenuates its replication in suckling mouse brain.

Table 7. Temperature-sensitive (ts) and mouse brain attenuation (att) phenotypes of 5-FU DEN4 mutant viruses which exhibit a small plaque (sp) phenotype.

Table 8. Viruses with both ts and sp phenotypes are more restricted in replication in mouse brain than those with only a ts phenotype.

Table 9. Nucleotide and amino acid differences of the 5-FU mutant DEN4 viruses which produce small plaques in both Vero and HuH-7 cells.

Table 10. Nucleotide and amino acid differences of the 5-FU mutant DEN4 viruses which produce small plaques in only HuH-7 cells.

Table 11. Putative Vero cell adaptation mutations derived from the full set of 5-FU mutant viruses.

Table 12. Mutagenic oligonucleotides used to generate recombinant DEN4 viruses containing single 5-FU mutations.

Table 13. sp, ts and mouse attenuation phenotypes of rDEN4 mutant viruses encoding single mutations identified in six sp 5-FU mutant viruses.

Table 14. Phenotypes of rDEN4 mutant viruses encoding single mutations identified in 10 5-FU mutant viruses that are ts in both Vero and HuH-7 cells.

Table 15. sp, ts and mouse attenuation phenotypes of rDEN4 mutant viruses encoding single mutations identified in 3 HuH-7 cell-specific ts 5-FU mutant viruses.

Table 16. Temperature-sensitive (ts) and mouse brain attenuation (att) phenotypes of additional rDEN4 viruses encoding single 5-FU mutations.

Table 17. Growth of wt DEN-4 2A-13 in SCID mice transplanted with HuH-7 cells.

Table 18. Combination of ts mutations, NS3 4995 and NS5 7849, in rDEN4 results in an additive ts phenotype.

Table 19. The 5-FU mutations are compatible with the  $\Delta$ 30 mutation for replication in the brain of suckling mice.

Table 20. Temperature-sensitive and mouse brain attenuation phenotypes of viruses bearing charge-cluster-to-alanine mutations in the NS5 gene of DEN4.

Table 21. SCID-HuH-7 attenuation phenotypes of viruses bearing charge-cluster-to-alanine mutations in the NS5 gene of DEN4.

Table 22. Combination of paired charge-cluster-to-alanine mutations into double-pair mutant viruses.

Table 23. Temperature-sensitive and mouse brain attenuation phenotypes of double charge-cluster-to-alanine mutants of the NS5 gene of rDEN4.

Table 24. SCID-HuH-7 attenuation phenotypes of double charge-cluster-to-alanine mutants of the NS5 gene of rDEN4.

Table 25. Phenotypes (temperature sensitivity, plaque size and replication in mouse brain and SCID-HuH-7 mice) of wt DEN4 and viruses containing the  $\Delta$ 30 and 7129 mutations.

Table 26. The 5-fluorouracil 5-1A1 small plaque mutant demonstrates a restriction of midgut infection following oral infection of *Aedes aegypti* mosquitoes.

Table 27. The 5-fluorouracil 5-1A1 small plaque mutant demonstrates a restriction of infection following intrathoracic inoculation of *Toxorhynchites splendens* mosquitoes.

Table 28. Mutagenesis primers for the deletion or swap of sequences in DEN4 showing conserved differences from tick-borne flaviviruses.

Table 29. Virus titer and plaque size of 3' UTR mutant viruses in Vero and C6/36 cells.

Table 30. Infectivity of wt DEN4 and 3' UTR mutants for *Toxorhynchites splendens* via intrathoracic inoculation.

Table 31. Infectivity of 3' UTR swap mutant viruses for *Aedes aegypti* fed on an infectious bloodmeal.

Table 32. Putative Vero cell adaptation mutations derived from the set of 5-FU mutant viruses and other DEN4 viruses passaged in Vero cells.

Table 33. Sequence analysis of rDEN2/4 $\Delta$ 30 clone 27(p4)-2-2A2.

Table 34. Sequence analysis of rDEN2/4 $\Delta$ 30 clone 27(p3)-2-1A1.

Table 35. Recombinant virus rDEN2/4 $\Delta$ 30 bearing Vero adaptation mutations can be recovered and titered on Vero cells.

Table 36. Putative Vero cell adaptation mutations of dengue type 4 virus and the corresponding wildtype amino acid residue in other dengue viruses.

Table 37. Mutations known to attenuate dengue type 4 virus and the corresponding wildtype amino acid residue in other dengue virus.

#### BRIEF DESCRIPTION OF THE APPENDICES

Appendix 1. Sequence of recombinant dengue type 4 virus strain 2A (amino acid sequence SEQ ID NO: 13 and nucleotide sequence SEQ ID NO: 14).

Appendix 2. Sequence of recombinant dengue type 4 virus strain rDEN4 (amino acid sequence SEQ ID NO: 15 and nucleotide sequence SEQ ID NO: 16).

Appendix 3. Sequence of recombinant dengue type 2 chimeric virus strain rDEN2/4 $\Delta$ 30 (amino acid sequence SEQ ID NO: 17 and nucleotide sequence SEQ ID NO: 18).

Appendix 4. Alignment of dengue virus polyproteins. DEN4 (SEQ ID NO: 19); DEN1-WP (SEQ ID NO: 20); DEN2-NGC (SEQ ID NO: 21); DEN3-H87 (SEQ ID NO: 22).

## DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENT

To assemble a collection of useful mutations for incorporation in recombinant live dengue virus vaccines, site-directed and random mutagenesis techniques were used to introduce mutations into the dengue virus genome. The resulting mutant viruses were screened for several valuable phenotypes, including temperature sensitivity in Vero cells or human liver cells, host cell restriction in mosquito cells or human liver cells, host-cell adaptation for improved replication in Vero cells, and attenuation in mice. The genetic basis for each observed phenotype was determined by direct sequence analysis of the virus genome. Mutations identified through these sequencing efforts have been further evaluated by their re-introduction, singly, or in combination, into recombinant dengue virus and characterization of the resulting phenotypes. In this manner, a menu of mutations was developed that is useful in fine-tuning the attenuation and growth characteristics of dengue virus vaccines.

## EXAMPLE 1

## Chemical Mutagenesis of Dengue Virus Type 4 Yields Temperature-Sensitive and Attenuated Mutant Viruses

A recombinant live attenuated dengue virus type 4 (DEN4) vaccine candidate, 2AΔ30, was found previously to be generally well-tolerated in humans, but a rash and an elevation of liver enzymes in the serum occurred in some vaccinees. 2AΔ30, a non-temperature-sensitive (ts) virus, contains a 30 nucleotide deletion in the 3' untranslated region (UTR) of the viral genome. In the present study, chemical mutagenesis of DEN4 has been utilized to generate attenuating mutations which may be useful to further attenuate the incompletely attenuated 2AΔ30 candidate vaccine. Wild-type DEN4 2A virus was grown in Vero cells in the presence of 5-fluorouracil, and, from a panel of 1,248 clones that were isolated in Vero cells, twenty ts mutant viruses were identified which were ts in both Vero and HuH-7 cells (n=13) or in HuH-7 cells only (n=7). Each of the twenty ts mutations possessed an attenuation (att) phenotype as indicated by restricted replication in the brains of seven day old mice. The complete nucleotide sequence of the 20 ts mutant viruses identified nucleotide substitutions in structural and non-structural genes as well as in the 5' and 3' UTR with more than one change occurring, in general, per mutant virus. A ts mutation in the NS3 protein (nucleotide position 4,995) was introduced into a recombinant DEN4 virus possessing the Δ30 deletion creating the rDEN4Δ30-4995 recombinant virus which was found to be ts and to be more attenuated than rDEN4Δ30 in the brains of mice. A menu of attenuating mutations is being assembled that should be useful in generating satisfactorily attenuated recombinant dengue vaccine viruses and in increasing our understanding of the pathogenesis of dengue virus.

The mosquito-borne dengue (DEN) viruses (serotypes 1 to 4) are members of the Flavivirus genus and contain a single-stranded positive-sense RNA genome of approximately 10,600 nucleotides (nt) (Monath, T. P. & Heinz, F. X. 1996 in: Fields Virology B. N. Fields, et al. Eds. pp. 961-1034 Lippincott-Raven Publishers, Philadelphia). The genome organization of DEN viruses is 5'-UTR-C-prM-E-NS1-NS2A-NS2B-NS3-NS4A-NS4B-NS5-UTR-3' (UTR-untranslated region, C-capsid, PrM-pre-membrane, E-envelope, NS-non-structural) (Chang, G.-J. 1997 in: Dengue and dengue hemor-

rhagic fever D. J. Gubler & G. Kuno, eds. pp. 175-198 CAB International, New York; Rice, C. M. 1996 in: Fields Virology B. N. Fields et al. Eds. pp. 931-959 Lippincott-Raven Publishers, Philadelphia). A single viral polypeptide is co-translationally processed by viral and cellular proteases generating three structural proteins (C, M, and E) and seven NS proteins. The disease burden associated with DEN virus infection has increased over the past several decades in tropical and semi-tropical countries. Annually, there are an estimated 50-100 million cases of dengue fever (DF) and 500,000 cases of the more severe and potentially lethal dengue hemorrhagic fever/dengue shock syndrome (DHF/DSS) (Gubler, D. J. & Meltzer, M. 1999 *Adv Virus Res* 53:35-70).

The site of viral replication in DEN virus-infected humans and the pathogenesis of DF and DHF/DSS are still incompletely understood (Innis, B. L. 1995 in: Exotic viral infections J. S. Porterfield, ed. pp. 103-146 Chapman and Hall, London). In humans, DEN virus infects lymphocytes (Kurane, I. et al. 1990 *Arch Virol* 110:91-101; Theofilopoulos, A. N. et al. 1976 *J Immunol* 117:953-61), macrophages (Halstead, S. B. et al. 1977 *J Exp Med* 146:218-29; Scott, R. M. et al. 1980 *J Infect Dis* 141:1-6), dendritic cells (Libraty, D. H. et al. 2001 *J Virol* 75:3501-8; Wu, S. J. et al. 2000 *Nat Med* 6:816-20), and hepatocytes (Lin, Y. L. et al. 2000 *J Med Virol* 60:425-31; Marianneau, P. et al. 1996 *J Gen Virol* 77:2547-54). The liver is clearly involved in DEN virus infection of humans, as indicated by the occurrence of transient elevations in serum alanine aminotransferase (ALT) and aspartate aminotransferase (AST) levels in the majority of dengue virus-infected patients and by the presence of hepatomegaly in some patients (Kalayanarooj, S. et al. 1997 *J Infect Dis* 176:313-21; Kuo, C. H. et al. 1992 *Am J Trop Med Hyg* 47:265-70; Mohan, B. et al. 2000 *J Trop Pediatr* 46:40-3; Wahid, S. F. et al. 2000 *Southeast Asian J Trop Med Public Health* 31:259-63). DEN virus antigen-positive hepatocytes are seen surrounding areas of necrosis in the liver of fatal cases (Couvelard, A. et al. 1999 *Hum Pathol* 30:1106-10; Huerre, M. R. et al. 2001 *Virchows Arch* 438:107-15), and dengue virus sequences were identified in such cases using RT-PCR (Rosen, L. et al. 1999 *Am J Trop Med Hyg* 61:720-4). Of potential importance to the etiology of severe dengue virus infection, three studies have demonstrated that the mean levels of serum ALT/AST were significantly increased in patients with DHF/DSS versus those with DF (Kalayanarooj, S. et al. 1997 *J Infect Dis* 176:313-21; Mohan, B. et al. 2000 *J Trop Pediatr* 46:40-3; Wahid, S. F. et al. 2000 *Southeast Asian J Trop Med Public Health* 31:259-63).

A vaccine for DEN viruses is not presently licensed. Since previous infection with one dengue virus serotype can increase the risk for DHF/DSS following infection with a different serotype (Burke, D. S. et al. 1988 *Am J Trop Med Hyg* 38:172-80; Halstead, S. B. et al. 1969 *Am J Trop Med Hyg* 18:997-1021; Thein, S. et al. 1997 *Am J Trop Med Hyg* 56:566-72), it is clear that a dengue virus vaccine will need to protect against each of the four dengue virus serotypes, namely DEN1, DEN2, DEN3, and DEN4. Several strategies are currently being actively pursued in the development of a live attenuated tetravalent DEN virus vaccine (Bancroft, W. H. et al. 1984 *J Infect Dis* 149:1005-10; Bhamarapravati, N. & Sutee, Y. 2000 *Vaccine* 18:44-7; Guirakhoo, F. et al. 2000 *J Virol* 74:5477-85; Huang, C. Y. et al. 2000 *J Virol* 74:3020-8). Recently, we demonstrated that a live attenuated DEN4 vaccine candidate, 2AΔ30, was attenuated and immunogenic in a group of 20 human volunteers (see Example 8). This recombinant DEN4 virus contains a 30 nt deletion in the 3' UTR which removes nucleotides 10,478-10,507 and was restricted in replication in rhesus monkeys. Levels of viremia in humans

were low or undetectable, and virus recovered from the vaccinees retained the  $\Delta 30$  mutation. An asymptomatic rash was reported in 50% of patients. The only laboratory abnormality observed was an asymptomatic, transient rise in the serum ALT level in 5 of 20 vaccinees. All vaccinees developed serum-neutralizing antibody against DEN4 virus (mean titer: 1:580). Importantly, 2A $\Delta 30$  was not transmitted to mosquitoes fed on vaccinees and has restricted growth properties in mosquitoes (Troyer, J. M. et al. 2001 *Am J Trop Med Hyg* 65:414-9). The presence of a rash and of the elevated ALT levels suggests that the 2A $\Delta 30$  vaccine candidate is slightly under-attenuated in humans. Because of the overall set of desirable properties conferred by the  $\Delta 30$  mutation, chimeric vaccine candidates are being constructed which contain the structural genes of dengue virus type 1, 2, and 3 and the DEN4 attenuated backbone bearing the genetically stable  $\Delta 30$  mutation.

Although the initial findings indicate the utility of the 2A $\Delta 30$  vaccine candidate, many previous attempts to develop live attenuated dengue virus vaccines have yielded vaccine candidates that were either over- or under-attenuated in humans (Eckels, K. H. et al. 1984 *Am J Trop Med Hyg* 33:684-9; Bhamarapravati, N. & Yoksan, S. 1997 in: *Dengue and dengue hemorrhagic fever* D. J. Gubler & G. Kuno eds. pp. 367-377 CAB International, New York; Innis, B. L. et al. 1988 *J Infect Dis* 158:876-80; McKee, K. T., Jr. et al. 1987 *Am J Trop Med Hyg* 36:435-42). Therefore, we developed a menu of point mutations which confer temperature-sensitive (ts) and attenuation (att) phenotypes upon DEN4. These mutations are envisioned as being useful to attenuate DEN4 viruses to different degrees and therefore as having purpose in fine-tuning the level of attenuation of vaccine candidates such as 2A $\Delta 30$ . Addition of such mutations to 2A $\Delta 30$  or to other dengue virus vaccine candidates is envisioned as resulting in the generation of a vaccine candidate that exhibits a satisfactory balance between attenuation and immunogenicity for humans.

In the present example, chemical mutagenesis of DEN4 has been utilized to identify point mutations which confer the ts phenotype, since such viruses often are attenuated in humans. Additionally, because of the reported involvement of the liver in natural dengue infection and the elevated ALT levels in a subset of 2A $\Delta 30$  vaccinees, mutagenized DEN4 viruses were also evaluated for ts phenotype in HuH-7 liver cells derived from a human hepatoma. Here, we describe the identification of 20 DEN4 ts mutant viruses each of which replicates efficiently in Vero cells, the proposed substrate for vaccine manufacture, and each of which is attenuated in mice. Finally, the feasibility of modifying the attenuation phenotype of the 2A $\Delta 30$  vaccine candidate by introduction of a point mutation in NS3 is demonstrated.

Cells and viruses. WHO Vero cells (African green monkey kidney cells) were maintained in MEM (Life Technologies, Grand Island, N.Y.) supplemented with 10% fetal bovine serum (FBS) (Summit Biotechnologies, Fort Collins, Colo.), 2 mM L-glutamine (Life Technologies), and 0.05 mg/ml gentamicin (Life Technologies). HuH-7 cells (human hepatoma cells) (Nakabayashi, H. et al. 1982 *Cancer Res* 42:3858-63) were maintained in D-MEM/F-12 (Life Technologies) supplemented with 10% FBS, 1 mM L-glutamine and 0.05 mg/ml gentamicin. C6/36 cells (*Aedes albopictus* mosquito cells) were maintained in complete MEM as described above supplemented with 2 mM non-essential amino acids (Life Technologies).

The wild type (wt) DEN4 2A virus was derived from a cDNA clone of DEN4 strain 814669 (Dominica, 1981) (Men, R. et al. 1996 *J Virol* 70:3930-7). Sequence of the cDNA of

DEN 4 2A virus is presented in Appendix 1. The full-length 2A cDNA clone has undergone several subsequent modifications to improve its ability to be genetically manipulated. As previously described, a translationally-silent XhoI restriction enzyme site was engineered near the end of the E region at nucleotide 2348 to create clone 2A-XhoI (Bray, M. & Lai, C. J. 1991 *PNAS USA* 88:10342-6). The viral coding sequence of the 2A-XhoI cDNA clone was further modified using site-directed mutagenesis to create clone p4: a unique BbvCI restriction site was introduced near the C-prM junction (nucleotides 447-452); an extra XbaI restriction site was ablated by mutation of nucleotide 7730; and a unique SacII restriction site was created in the NS5 region (nucleotides 9318-9320). Each of these engineered mutations is translationally silent and does not change the amino acid sequence of the viral polypeptide. Also, several mutations were made in the vector region of clone p4 to introduce or ablate additional restriction sites. The cDNA clone p4 $\Delta 30$  was generated by introducing the  $\Delta 30$  mutation into clone p4. This was accomplished by replacing the MluI-KpnI fragment of p4 (nucleotides 10403-10654) with that derived from plasmid 2A $\Delta 30$  containing the 30 nucleotide deletion. The cDNA clones p4 and p4 $\Delta 30$  were subsequently used to generate recombinant viruses rDEN4 (Appendix 2) and rDEN4 $\Delta 30$ , respectively. (The GenBank accession number for rDEN4 is AF326825 and the accession for rDEN4 $\Delta 30$  is AF326827).

Chemical mutagenesis of DEN4. Confluent monolayers of Vero cells were infected with wt DEN4 2A at a multiplicity of infection (MOI) of 0.01 and incubated for 2 hours at 32° C. Infected cells were then overlaid with MEM supplemented with 2% FBS and 5-fluorouracil (5-FU) (Sigma, St. Louis, Mo.) at concentrations ranging from 10 mM to 10 nM. After incubation at 32° C. for five days, cell culture medium was harvested, clarified by centrifugation, and frozen at -70° C. Clarified supernatants were then assayed for virus titer by plaque titration in Vero cells. Serial ten-fold dilutions of the clarified supernatant were prepared in Opti-MEM I (Life Technologies) and inoculated onto confluent Vero cell monolayers in 24-well plates. After incubation at 35° C. for two hours, monolayers were overlaid with 0.8% methylcellulose (EM Science, Gibbstown, N.J.) in Opti-MEM I supplemented with 2% FBS, gentamicin, and L-glutamine. Following incubation at 35° C. for five days, plaques were visualized by immunoperoxidase staining. Vero cell monolayers were fixed in 80% methanol for 30 minutes and washed for 10 minutes with antibody buffer which consists of 3.5% (w/v) nonfat dry milk (Nestle, Solon, Ohio) in phosphate buffered saline (PBS). Cells were then incubated for one hour at 37° C. with an anti-DEN4 rabbit polyclonal antibody preparation (PRNT<sub>50</sub> of >1:2000) diluted 1:1,000 in antibody buffer. After one wash with antibody buffer, cells were incubated for one hour with peroxidase-labeled goat-anti-rabbit IgG (KPL, Gaithersburg, Md.) diluted 1:500 in antibody buffer. Monolayers were washed with PBS, allowed to dry briefly, overlaid with peroxidase substrate (KPL), and plaques were counted.

Virus yields in cultures treated with 1 mM 5-FU were reduced 100-fold compared to untreated cultures, and the virus present in the supernatant from the 1 mM 5-FU-treated culture was terminally diluted to derive clones for phenotypic characterization. Briefly, 96 well plates of Vero cells were inoculated with the 5-FU-treated virus at an MOI that yielded 10 or fewer virus-positive wells per plate. After a five-day incubation at 35° C., cell culture media from the 96 well plates were temporarily transferred to 96 well plates lacking cells, and the positive cultures were identified by immunoperoxidase staining of the infected-cell monolayers. Virus from each positive well was transferred to confluent Vero cell

monolayers in 12 well plates for amplification. Cell culture medium was harvested from individual wells five or six days later, clarified by centrifugation, aliquoted to 96 deep-well polypropylene plates (Beckman, Fullerton, Calif.) and frozen at  $-70^{\circ}\text{C}$ . A total of 1,248 virus clones were prepared from the 1 mM 5-FU-treated cultures. Two wt virus clones, 2A-1 and 2A-13, were generated in the same manner from the 5-FU untreated control cultures.

Screening of clones for ts and att phenotypes. The 1,248 virus clones were screened for ts phenotype by assessing virus replication at  $35^{\circ}\text{C}$ . and  $39^{\circ}\text{C}$ . in Vero and HuH-7 cells. Cell monolayers in 96 well plates were inoculated with serial ten-fold dilutions of virus in L-15 media (Quality Biologicals, Gaithersburg, Md.) supplemented with 2% FBS, L-glutamine and gentamicin. Cells were incubated at the indicated temperatures for five days in temperature-controlled water baths, and presence of virus was determined by immunoperoxidase staining as described above. Virus clones which demonstrated a 100-fold or greater reduction in titer at  $39^{\circ}\text{C}$ . were terminally diluted an additional two times and amplified in Vero cells. The efficiency of plaque formation (EOP) at permissive and restrictive temperatures of each triply biologically cloned virus suspension was determined as follows. Plaque titration in Vero and HuH-7 cells was performed as described above except virus-infected monolayers were overlaid with 0.8% methylcellulose in L-15 medium supplemented with 5% FBS, gentamicin, and L-glutamine. After incubation of replicate plates for five days at 35, 37, 38, or  $39^{\circ}\text{C}$ . in temperature-controlled water baths, plaques were visualized by immunoperoxidase staining and counted.

The replication of DEN4 5-FU ts mutant viruses was evaluated in Swiss Webster suckling mice (Taconic Farms, Germantown, N.Y.). Groups of six one-week-old mice were inoculated intracranially with 104 PFU of virus diluted in 30  $\mu\text{l}$  Opti-MEM I. Five days later, mice were sacrificed and brains were removed and individually homogenized in a 10% suspension of phosphate-buffered Hank's balanced salt solution containing 7.5% sucrose, 5 mM sodium glutamate, 0.05 mg/ml ciprofloxacin, 0.06 mg/ml clindamycin, and 0.0025 mg/ml amphotericin B. Clarified supernatants were frozen at  $-70^{\circ}\text{C}$ . and subsequently virus titer was determined by titration in Vero cells, and plaques were stained by the immunoperoxidase method described above.

Sequence analysis of viral genomes. The nucleotide sequence of the 5-FU-mutagenized DEN4 viruses was determined. Briefly, genomic viral RNA was isolated from virus clones with the QIAamp viral RNA mini kit (Qiagen, Valencia, Calif.) and reverse transcription was performed using the SuperScript First Strand Synthesis System for RT-PCR (Life Technologies) and random hexamer primers. Advantage cDNA polymerase (Clontech, Palo Alto, Calif.) was used to generate overlapping PCR fragments of approximately 2,000 nt which were purified by HighPure PCR Product Purification System (Roche Diagnostics, Indianapolis, Ind.). DEN-specific primers were used in Big-Dye terminator cycle sequencing reactions (Applied Biosystems, Foster City, Calif.) and reactions were analyzed on a 3100 genetic analyzer (Applied Biosystems). Primers were designed to sequence both strands of the PCR product from which consensus sequences were assembled.

The nucleotide sequence of the 5' and 3' regions of the viral genome were determined as above after circularization of the RNA genome. The 5' cap nucleoside of the viral RNA was excised using tobacco acid pyrophosphatase (Epicentre Technologies, Madison, Wis.) and the genome was circularized by RNA ligase (Epicentre Technologies). A RT-PCR fragment

was generated which overlapped the ligation junction (5' and 3' ends) and was sequenced as described above.

Generation of recombinant DEN4 viruses. The mutation at nt position 4,995 in NS3 was introduced into the p4 cDNA construct by site-directed mutagenesis (Kunkel, T. A. 1985 PNAS USA 82:488-92). The StuI-BstBI (nt 3,619-5,072) fragment of p4 was sub-cloned into a modified pUC119 vector. The U>C mutation at nt position 4,995 was engineered by site-directed mutagenesis into the p4 fragment, cloned back into the p4 cDNA construct, and the presence of the mutation was confirmed by sequence analysis. The  $\Delta 30$  mutation was introduced into the 3' UTR of the p4-4995 cDNA clone by replacing the MluI-KpnI fragment with that derived from the p4 $\Delta 30$  cDNA clone, and the presence of the deletion was confirmed by sequence analysis. Full length RNA transcripts were prepared from the above cDNA clones by in vitro transcription. Briefly, transcription consisted of a 50  $\mu\text{l}$  reaction mixture containing 1  $\mu\text{g}$  linearized plasmid, 60 U SP6 polymerase (New England Biolabs (NEB), Beverly, Mass.), 1 $\times$  RNA polymerase buffer (40 mM Tris-HCl, pH 7.9, 6 mM  $\text{MgCl}_2$ , 2 mM spermidine, 10 mM dithiothreitol), 0.5 mM m7G(5')ppp(5')G cap analog (NEB), 1 mM each nucleotide triphosphate, 1 U pyrophosphatase (NEB), and 80 U RNase inhibitor (Roche, Indianapolis, Ind.). This reaction mixture was incubated at  $40^{\circ}\text{C}$ . for 90 min and the resulting transcripts were purified using RNeasy mini kit (Qiagen, Valencia, Calif.).

For transfection of C6/36 cells, RNA transcripts were combined with DOTAP liposomal transfection reagent (Roche) in HEPES-buffered saline (pH 7.6) and added to cell monolayers in 6 well plates. After incubation at  $32^{\circ}\text{C}$ . for 12-18 hours, the cell culture media were removed and replaced with MEM supplemented with 5% FBS, L-glutamine, gentamicin and non-essential amino acids. Cell monolayers were incubated for an additional 5 to 7 days and cell culture media were harvested, clarified by centrifugation, and assayed for the presence of virus by plaque titration in Vero cells. Recovered viruses were terminally diluted twice as described above, and virus suspensions for further analysis were prepared in Vero cells.

In vitro (tissue culture) and in vivo replication of wt DEN4 and DEN4 $\Delta 30$ . The level of replication of both wt DEN4 2A and the vaccine candidate, 2A $\Delta 30$ , was evaluated in Vero (monkey kidney) and HuH-7 (human hepatoma) cells (FIG. 1), the latter of which has recently been found to efficiently support the replication of DEN2 virus (Lin, Y. L. et al. 2000 J Med Virol 60:425-31). The pattern of replication of wt DEN4 2A and 2A $\Delta 30$  was similar in both cell lines. Viral titers from cultures infected with 2A $\Delta 30$  at an MOI of 0.01 were slightly reduced compared to wt DEN4 2A at 72 hours, but at later time points their level of replication was equivalent. The efficient replication of both DEN4 viruses in each cell line indicated that these continuous lines of cells would be useful for characterization of the ts phenotype of the 1248 potential mutant viruses.

The level of replication of DEN4 virus administered intracerebrally to Swiss Webster mice was first determined to assess whether mice could be used to efficiently evaluate and quantitate the attenuation phenotype of a large set of mutant viruses. Since the susceptibility of mice to DEN infection is age dependent (Cole, G. A. & Wisseman, C. L. Jr. 1969 Am J Epidemiol 89:669-80; Cole, G. A. et al. 1973 J Comp Pathol 83:243-52), mice aged 7 to 21 days were infected with 2A-13 (a clone of DEN4 wild type virus—see below), rDEN4 or rDEN4 $\Delta 30$ , and after five days the brain of each mouse was removed, and the level of viral replication was quantitated by plaque assay (Table 1). The results indicated that the two wt

DEN4 viruses and the rDEN4 $\Delta$ 30 vaccine candidate replicated to high titer ( $>6.0 \log_{10}$  PFU/g brain) in 7-day old mice and that the mean viral titers were similar among the three viruses. These results demonstrated the feasibility of using 7-day old mice to screen a large set of mutant viruses, and the high level of replication of wild type and vaccine candidate permits one to quantitate the magnitude of the restriction of replication specified by an attenuating mutation over a 10,000-fold range.

Generation and in vitro characterization of 1DEN4 5-FU mutant viruses. A panel of 1,248 DEN4 virus clones was generated from a 5-FU-mutagenized suspension of wt DEN4 2A as described above (FIG. 2). Each clone was tested in Vero and HuH-7 cells for the ts phenotype at 39° C., and putative ts mutant viruses were subjected to two additional rounds of biological cloning by terminal dilution, and the ts phenotype of each further cloned virus population was examined in more detail by determining their efficiency of plating (EOP) at permissive temperature (35° C.) and at various restrictive temperatures (Table 2). One virus (clone 2A-13) without a ts phenotype, which was passaged in an identical fashion as the ts mutant viruses, served as the virus to which each of the ts mutant viruses was directly compared for both the ts and att phenotypes.

Thirteen 5-FU mutant viruses were identified which have a ts phenotype in both Vero and HuH-7 cells, and seven mutant viruses were ts only in HuH-7 cells (Table 2). Mutant viruses which were ts in Vero cells but not in HuH-7 cells were not identified. Temperature-sensitivity was defined as a  $\geq 2.5$  or  $\leq 3.5 \log_{10}$  PFU/ml reduction in virus titer in Vero or HuH-7 cells, respectively, at an indicated temperature when compared to the permissive temperature of 35° C. Wild type DEN4 2A was found to have approximately a 0.5 and 1.5  $\log_{10}$  PFU/ml reduction in virus titer in Vero or HuH-7 cells at 39° C., respectively. The  $\Delta$ 30 deletion did not confer a ts phenotype in Vero or HuH-7 cells and exhibited only a slight reduction in virus titer ( $2.2 \log_{10}$  PFU/ml) at 39° C. in HuH-7 cells, which was less than 10-fold greater than the reduction of wt DEN4 2A at that temperature. Several 5-FU mutant viruses had a greater than 10,000-fold reduction in virus titer at 39° C. in both Vero and HuH-7 cells. A complete shut-off in viral replication at 39° C. in HuH-7 cells was observed in five virus clones (#571, 605, 631, 967, and 992) which were not ts in Vero cells. Mutations that selectively restrict replication in HuH-7 liver cells may be particularly useful in controlling the replication of dengue virus vaccine candidates in the liver of vaccinees.

Replication of DEN4 5-FU mutant viruses in suckling mice. The level of replication of each of the 20 ts DEN4 mutant viruses in mouse brain was determined (Table 2). The titers obtained were compared to that of the two wt viruses, 2A-13 and rDEN4, which each replicated to a level of greater than  $10^6$  PFU/g of brain tissue, and to that of the 2A $\Delta$ 30 mutant, which conferred only a limited  $0.5 \log_{10}$  PFU/g reduction in mean virus titer compared to the wt controls. The observed reduction in the level of rDEN4 $\Delta$ 30 replication was consistent among 11 separate experiments. Interestingly, the rDEN4 $\Delta$ 30 virus, which was attenuated in both rhesus monkeys and humans (Example 8), was only slightly restricted in replication in mouse brain. Varying levels of restriction of replication were observed among the mutant viruses ranging from a 10-fold (#473) to over 6,000-fold (#686) reduction. Mutant viruses with ts phenotypes in both Vero and HuH-7 cells, as well as in HuH-7 cells alone, were found to have significant att phenotypes. Five of 13 5-FU mutant viruses with ts phenotypes in both Vero and HuH-7 cells and five of seven mutant viruses with ts phenotypes in HuH-7 cells alone

had greater than a 100-fold reduction in virus replication. There appeared to be no direct correlation between the magnitude of the reduction in replication at restrictive temperature in tissue culture and the level of attenuation in vivo. The similar level of temperature sensitivity and replication of the rDEN4 wt and clone 2A-13 in mouse brain indicated that observed differences in replication between the ts mutant viruses and clone 2A-13 was not simply a function of passage in Vero cells, but reflects the sequence differences between these viruses.

Sequence analysis of DEN4 5-FU mutant viruses. To determine the genetic basis of the observed ts and att phenotypes, the complete nucleotide sequence of each ts mutant and of clone 2A-13 was determined and summarized in Table 3 (ts in Vero and HuH-7 cells) and Table 4 (ts in only HuH-7 cells).

The only type of mutation identified in the 20 mutant viruses sequenced was a nucleotide substitution (no deletions or insertions occurred), and these were present in each of the coding regions except C and NS4A. Three mutant viruses (#239, 489, and 773) contained only a single missense point mutation in NS3 at nt position 4,995 resulting in a Ser to Pro amino acid (a.a.) change at a.a. position 1,632. For #773, this was the sole mutation present (Table 3). The non-coding mutations in coding regions are not considered to be significant. The 17 additional mutant viruses had multiple mutations (two to five) in a coding region or in an UTR which could potentially confer the observed ts or att phenotypes. Five of the 17 mutant viruses with multiple mutations (#473, 718, 759, 816, and 938) also encoded the point mutation at nt position 4,995. The presence of the 4,995 mutation was found in only DEN4 mutant viruses with ts phenotypes in both Vero and HuH-7 cells.

The sequence analysis indicated that 10 mutant viruses which were ts in Vero and HuH-7 cells and three mutant viruses which were ts in only HuH-7 cells contained mutations in only the 5' and 3' UTR and/or in a nonstructural protein. These mutations are especially suitable for inclusion in chimeric dengue virus vaccine candidates in which the structural genes derive from a DEN1, DEN2, or DEN3 serotype and the remaining coding and non-coding regions come from an attenuated DEN4 vector. Mutations identified in 5-FU DEN4 mutant viruses which were ts in only HuH-7 cells (Table 4) may potentially be utilized in vaccine candidates, such as rDEN4 $\Delta$ 30, to selectively control the replication and pathogenesis of DEN4 in the liver. These combined results from the sequence analysis of 5-FU mutant viruses demonstrate the utility of chemical mutagenesis as a means of introducing attenuating mutations into the dengue virus genome.

The presence of a point mutation at nt position 4,995 in eight separate mutant viruses was described above. Five additional point mutations were also represented in multiple viruses including nt changes at position 1,455 in E, 7,162, 7,163 and 7,564 in NS4B, and 10,275 in the 3' UTR (Table 5). The significance of the occurrence of these "sister" mutations in multiple viruses is discussed in Example 6. Interestingly, the wild-type, parallel-passaged virus, 2A-13, also contained a single mutation at the 7,163 nt position in NS4B.

Introduction of a ts mutation into rDEN4 and rDEN4 $\Delta$ 30. The presence of a single nucleotide substitution (U>C mutation at nt position 4,995 in NS3) in three separate mutant viruses (clones 239, 489, and 773) indicated that this mutation specified the ts and att phenotypes in each of the three mutant viruses. This mutation was cloned into cDNA construct of p4 and p4 $\Delta$ 30 and recombinant viruses were recovered and designated rDEN4-4995 and rDEN4 $\Delta$ 30-4995, respectively. These recombinant viruses were tested for ts and att phenotypes as described above (Table 6). As expected,

introduction of mutation 4995 into rDEN4 wt resulted in a significant ts phenotype at 39° C. in both Vero and HuH-7 cells. rDEN4-4995 grew to nearly wild-type levels at the permissive temperature, 35° C., in both cell types, but demonstrated a greater than 10,000-fold reduction at 39° C. (shut-off temperature) in both Vero and HuH-7 cells. The addition of the 4995 mutation to rDEN4Δ30 yields a recombinant virus, rDEN4Δ30-4995, that exhibits the same level of temperature sensitivity as rDEN4-4995 (Table 6).

The rDEN4 viruses encoding the 4995 mutation were next tested for replication in the brains of suckling mice (Table 6). The 4995 mutation conferred an att phenotype upon both rDEN4 and rDEN4Δ30. There was an approximately 1,000-fold reduction in virus replication compared to that of wt virus. The combination of point mutation 4995 and the Δ30 deletion did not appear to result in an additive reduction of virus replication. These results confirmed that the 4995 point mutation indeed specifies the ts and att phenotypes. Importantly, the utility of modifying tissue culture and in vivo phenotypes of the rDEN4Δ30 vaccine candidate by introduction of additional mutations was also demonstrated.

Discussion. Herein we teach how to prepare a tetravalent, live-attenuated dengue virus vaccine using rDEN4Δ30 as the DEN4 component and three antigenic chimeric viruses expressing the structural proteins (C, prM, and E) of DEN1, DEN2, and DEN3 from the attenuated rDEN4Δ30 vector (Example 8). DEN4 virus rDEN4Δ30 containing the Δ30 deletion mutation in the 3' UTR manifests restricted replication in humans while retaining immunogenicity. Since rDEN4Δ30 retains a low level of residual virulence for humans despite this restricted replication, the present study was initiated to generate additional attenuating mutations that are envisioned as being useful to further attenuate rDEN4Δ30 or other dengue viruses and that are envisioned as being incorporated into any of the three antigenic chimeric viruses or other dengue viruses as needed. Temperature-sensitive mutants of dengue viruses (Bharnarapravati, N. & Yoksan, S. 1997 in: *Dengue and Dengue Hemorrhagic Fever* D. J. Gubler & G. Kuno eds. pp. 367-377 CAB International, New York; Eckels, K. H. et al. 1980 *Infect Immun* 27:175-80) as well as other viruses (Skiadopoulos, M. H. et al. 1998 *J Virol* 72:1762-8; Whitehead, S. S. et al. 1999 *J Virol* 73:871-7) manifest restricted replication in vivo. We have generated a panel of 20 ts DEN4 mutant viruses, determined their genomic sequence, and assessed their in vivo attenuation phenotypes. The 20 ts DEN4 mutant viruses were generated by growth in the presence of 5-FU and were first selected for viability in Vero cells, the substrate planned for use in the manufacture of these vaccines, to ensure that the mutant viruses can be grown efficiently in a suitable substrate.

Two classes of mutant viruses were obtained; those ts in both Vero and HuH-7 cells (n=13) or those ts in only HuH-7 cells (n=7). The viruses exhibited a range in their level of temperature sensitivity from a 100- to 1,000,000-fold reduction in replication at the restrictive temperature of 39° C. Since our DEN4 vaccine candidate retains a low level of virulence for the liver and other findings support the ability of dengue viruses to infect hepatocytes (Lin, Y. L. et al. 2000 *J Med Virol* 60:425-31; Marianneau, P. et al. 1997 *J Virol* 71:3244-9) and cause liver pathology (Couvelard, A. et al. 1999 *Hum Pathol* 30:1106-10; Huerre, M. R. et al. 2001 *Virchows Arch* 438:107-15), we sought to develop mutations that would selectively restrict replication of dengue 4 virus in liver cells. Toward this end, we identified seven mutant viruses which have a HuH-7 cell-specific ts phenotype. The mutations present in these viruses are the first reported in DEN viruses that confer restricted replication in liver cells

and are envisioned as being useful in limiting virus replication and pathogenesis in the liver of vaccine recipients. The contribution of individual mutations identified in the HuH-7 cell-specific ts viruses to the observed phenotypes is envisioned as being assessed by introduction of the individual mutations into recombinant DEN4 viruses.

Recent evidence has indicated that the magnitude of the viremia in DEN-infected patients positively correlates with disease severity, i.e., the higher the titer of viremia the more severe the disease (Murgue, B. et al. 2000 *J Med Virol* 60:432-8; Vaughn, D. W. et al. 2000 *J Infect Dis* 181:2-9). This indicates that mutations that significantly restrict replication of vaccine candidates in vivo are the foundation of a safe and attenuated vaccine. Evaluation of DEN virus vaccine candidates for in vivo attenuation is complicated by the lack of a suitable animal model which accurately mimics the disease caused by dengue viruses in humans. In the absence of such a model, the replication of the panel of 5-FU mutant viruses in the brains of Swiss Webster suckling mice was assessed as a means to identify an in vivo attenuation phenotype since this animal model is well-suited for the evaluation of a large set of mutant viruses. Each of the 20 ts mutant viruses exhibited an att phenotype, manifesting a 10- to 6,000-fold reduction in replication in the brain of mice as compared to wt DEN4 virus (Table 2). This indicates that there is a correlation between the presence of the ts phenotype in tissue culture and attenuation of the mutant in vivo confirming the utility of selecting viruses with this marker as vaccine candidates. However, there was no correlation between the level of temperature sensitivity and the level of restriction in vivo. Furthermore, Sabin observed a dissociation between mouse neurovirulence and attenuation in humans by generating an effective live attenuated virus vaccine against DEN by passage of virus in mouse brain. This research actually resulted in a highly mouse-neurotropic DEN virus which, paradoxically, was significantly attenuated in humans (Sabin, A. B. 1952 *Am J Trop Med Hyg* 1:30-50). Despite this, attenuation for the suckling mouse brain has been reported for other live-attenuated DEN virus vaccine candidates including the DEN2 PDK-53 vaccine strain which is non-lethal in mice and DEN-2 PR-159/S-1 vaccine strain which was significantly attenuated compared to its parental wild-type virus (Bharnarapravati, N. & Yoksan, S. 1997 in: *Dengue and Dengue Hemorrhagic Fever* D. J. Gubler & G. Kuno eds. pp. 367-377 CAB International, New York; Butrapet, S. et al. 2000 *J Virol* 74:3011-9; Eckels, K. H. et al. 1980 *Infect Immun* 27:175-80; Innis, B. L. et al. 1988 *J Infect Dis* 158:876-80). Replication in rhesus monkeys has been reported to be predictive of attenuation for humans (Innis, B. L. et al. 1988 *J Infect Dis* 158:876-80). Recently, murine models of DEN virus infection have been developed using SCID mice transplanted with human macrophage (Lin, Y. L. et al. 1998 *J Virol* 72:9729-37) or liver cell lines (An, J. et al. 1999 *Virology* 263:70-7), but these mice have not as yet been used to assess att phenotypes of candidate vaccine viruses. Mutant viruses or recombinant viruses bearing one or more of these mutations described herein are envisioned as being tested for replication in rhesus monkeys (or other suitable animal model) as predictive for attenuation in humans.

The chemical mutagenesis of DEN4 virus and sequence analysis of resulting viruses described here has resulted in the identification of a large number of point mutations resulting in amino acid substitutions in all genes except C and NS4A as well as point mutations in the 5' and 3' UTR (Tables 3 and 4). This approach of whole-genome mutagenesis has the benefit of identifying mutations dispersed throughout the entire genome which are pre-selected for viability in the Vero cell



substrate. Ten 5-FU mutant viruses which were ts in Vero and HuH-7 cells and three viruses which were selectively ts in HuH-7 cells contained only mutations outside of the genes encoding the structural proteins, i.e., in the 5' and 3' UTR or NS genes. These mutations along with the  $\Delta 30$  deletion in the 3' UTR are particularly suited for inclusion in antigenic, chimeric vaccines which consist of an attenuated DEN4 vector bearing the wild-type structural genes (C, prM, E) of the other DEN virus serotypes. Use of this strategy has several advantages. Each antigenic chimeric virus that possesses structural proteins from a wild-type virus along with attenuating mutations in their UTRs or NS genes should maintain its infectivity for humans, which is mediated largely by the E protein, and, therefore, each vaccine component should be immunogenic (Huang, C. Y. et al. 2000 J Virol 74:3020-8). The replicative machinery of the tetravalent vaccine strains would share the same attenuating mutations in the NS genes or in the UTR which should attenuate each vaccine component to a similar degree and thereby minimize interference or complementation among the four vaccine viruses. In addition, wild-type E protein would be expected to most efficiently induce neutralizing antibodies against each individual DEN virus.

Sequence analysis of dengue viruses (Blok, J. et al. 1992 Virology 187:573-90; Lee, E. et al. 1997 Virology 232:281-90; Puri, B. et al. 1997 J Gen Virol 78:2287-91) and yellow fever viruses (Dunster, L. M. et al. 1999 Virology 261:309-18; Holbrook, M. R. et al. 2000 Virus Res 69:31-9) previously generated by serial passage in tissue culture have mutations throughout much of the genome, a pattern we have observed in the present study. Recent analysis of the DEN2 PDK-53 vaccine strain has identified the important mutations involved in attenuation which were located in non-structural regions including the 5' UTR, NS1 and NS3 (Butrapet, S. et al. 2000 J Virol 74:3011-9). This DEN2 vaccine strain has been used to generate a chimeric virus with DEN1 C-prM-E genes (Huang, C. Y. et al. 2000 J Virol 74:3020-8). In separate studies, the sequence of the DEN1 vaccine strain 45AZ5 PDK-27 was determined and compared to parental viruses, but the mutations responsible for attenuation have not yet been identified (Puri, B. et al. 1997 J Gen Virol 78:2287-91).

Several amino acid substitutions were identified in more than one ts 5-FU mutant virus (Table 5). Lee et al. have previously reported finding repeated mutations in separate DEN3 virus clones after serial passage in Vero cells (Lee, E. et al. 1997 Virology 232:281-90). A mutation (K>N) identified in E at a.a. position 202 in a single DEN3 passage series was also found in our 5-FU mutant virus #1012 (K>E). Mutations observed in the 5-FU sister mutant viruses are envisioned as representing adaptive changes that confer an increased efficiency of DEN4 replication in Vero cells. Such mutations are envisioned as being beneficial for inclusion in a live-attenuated DEN virus vaccine by increasing the yield of vaccine virus during manufacture. Interestingly, three distinct amino acid substitutions were found in NS4B of the 5-FU sister mutant viruses. The exact function of this gene is unknown, but previous studies of live-attenuated yellow fever vaccines (Jennings, A. D. et al. 1994 J Infect Dis 169:512-8; Wang, E. et al. 1995 J Gen Virol 76:2749-55) and Japanese encephalitis vaccines (Ni, H. et al. 1995 J Gen Virol 76:409-13) have identified mutations in NS4B associated with attenuation phenotypes.

The mutation at nt position 4995 of NS3 (S1632P) was present as the only significant mutation identified in three 5-FU mutant viruses (#239, #489, and #773). This mutation was introduced into a recombinant DEN4 virus and found to confer a ts and att phenotype (Table 6). These observations

clearly identify the 4995 mutation as an attenuating mutation. Analysis of a sequence alignment (Chang, G.-J. 1997 in: Dengue and Dengue Hemorrhagic Fever D. J. Gubler & G. Kuno, eds. pp. 175-198 CAB International, New York) of the four dengue viruses indicated that the Ser at a.a. position 1632 is conserved in DEN1 and DEN2, while DEN3 contains an Asn at this position indicating that the mutation is predicted to be useful in modifying the phenotypes of the other DEN virus serotypes. The NS3 protein is 618 a.a. in length and contains both serine protease and helicase activities (Bazan, J. F. & Fletterick, R. J. 1989 Virology 171:637-9; Brinkworth, R. I. et al. 1999 J Gen Virol 80:1167-77; Valle, R. P. & Falgout, B. 1998 J Virol 72:624-32). The 4995 mutation results in a change at a.a. position 158 in NS3 which is located in the N-terminal region containing the protease domain. Amino acid position 158 is located two a.a. residues away from an NS3 conserved region designated homology box four. This domain has been identified in members of the flavivirus family and is believed to be a critical determinant of the NS3 protease substrate specificity (Bazan, J. F. & Fletterick, R. J. 1989 Virology 171:637-9; Brinkworth, R. I. et al. 1999 J Gen Virol 80:1167-77). However, the exact mechanism which results in the phenotype associated with the 4995 mutation has not yet been identified. The identification of the 4995 mutation as an attenuating mutation permits a prediction of its usefulness for the further attenuation of rDEN4 $\Delta 30$ .

We have determined the contribution of individual 5-FU mutations to the observed phenotypes by introduction of the mutations into recombinant DEN4 viruses as was demonstrated herein for the 4995 mutation (see Example 3). In addition, combination of individual mutations with each other or with the  $\Delta 30$  mutation is useful to further modify the attenuation phenotype of DEN4 virus candidate vaccines. The introduction of the 4995 mutation into rDEN4 $\Delta 30$  described herein rendered the rDEN4 $\Delta 30$ -4995 double mutant ts and 1000-fold more attenuated for the mouse brain than rDEN4 $\Delta 30$ . This observation has demonstrated the feasibility of modifying both tissue culture and in vivo phenotypes of this and other dengue virus vaccine candidates. Once the mutations responsible for the HuH-7 cell-specific ts phenotype are identified as described above and introduced into the rDEN4 $\Delta 30$  vaccine candidate, we envision confirming that these mutations attenuate rDEN4 $\Delta 30$  vaccine virus for the liver of humans. A menu of attenuating mutations is envisioned as being assembled that is predicted to be useful in generating satisfactorily attenuated recombinant dengue vaccine viruses and in increasing our understanding of the pathogenesis of dengue virus (see Example 7).

## EXAMPLE 2

### Chemical Mutagenesis of DEN4 Virus Results in Small-Plaque Mutant Viruses with Temperature-Sensitive and Attenuation Phenotypes

Mutations that restrict replication of dengue virus have been sought for the generation of recombinant live-attenuated dengue virus vaccines. Dengue virus type 4 (DEN4) was previously grown in Vero cells in the presence of 5-fluorouracil, and the characterization of 1,248 mutagenized, Vero cell-passaged clones identified 20 temperature-sensitive (ts) mutant viruses that were attenuated (att) in suckling mouse brain (Example 1). The present investigation has extended these studies by identifying an additional 22 DEN4 mutant viruses which have a small-plaque size (sp) phenotype in Vero cells and/or the liver cell line, HuH-7. Five mutant viruses have a sp phenotype in both Vero and HuH-7 cells, three of

which are also ts. Seventeen mutant viruses have a sp phenotype in only HuH-7 cells, thirteen of which are also ts. Each of the sp viruses was growth restricted in the suckling mouse brain, exhibiting a wide range of reduction in replication (9- to 100,000-fold). Complete nucleotide sequence was determined for the 22 DEN4 sp mutant viruses, and nucleotide substitutions were found in the 3' untranslated region (UTR) as well as in all coding regions except NS4A. Identical mutations have been identified in multiple virus clones indicating that they are involved in the adaptation of DEN4 virus to efficient growth in Vero cells.

The DEN viruses cause more disease and death of humans than any other arbovirus, and more than 2.5 billion people live in regions with endemic dengue infection (Gubler, D. J. 1998 *Clin Microbiol Rev* 11:480-96). Annually, there are an estimated 50-100 million cases of dengue fever (DF) and 500,000 cases of the more severe and potentially lethal dengue hemorrhagic fever/dengue shock syndrome (DHF/DSS) (Gubler, D. J. & Meltzer, M. 1999 *Adv Virus Res* 53:35-70). Dengue fever is an acute infection characterized by fever, retro-orbital headache, myalgia, and rash. At the time of defervescence during DF, a more severe complication of DEN virus infection, DHF/DSS, may occur which is characterized by a second febrile period, hemorrhagic manifestations, hepatomegaly, thrombocytopenia, and hemoconcentration, which may lead to potentially life-threatening shock (Gubler, D. J. 1998 *Clin Microbiol Rev* 11:480-96).

The sites of DEN virus replication in humans and their importance and relationship to the pathogenesis of DF and DHF/DSS are still incompletely understood (Innis, B. L. 1995 in: *Exotic Viral Infections* J. S. Porterfield, ed. pp. 103-146 Chapman and Hall, London). In addition to replication in lymphoid cells, it has become evident that the liver is involved in DEN infection of humans. Transient elevations in serum alanine aminotransferase (ALT) and aspartate aminotransferase (AST) levels are observed in the majority of DEN virus-infected patients and hepatomegaly is observed in some patients (Kalayanarooj, S. et al. 1997 *J Infect Dis* 176:313-21; Kuo, C. H. et al. 1992 *Am J Trop Med Hyg* 47:265-70; Mohan, B. et al. 2000 *J Trop Pediatr* 46:40-3; Wahid, S. F. et al. 2000 *Southeast Asian J Trop Med Public Health* 31:259-63). DEN virus antigen-positive hepatocytes are seen surrounding areas of necrosis in the liver of fatal cases (Couvellard, A. et al. 1999 *Hum Pathol* 30:1106-10; Huerre, M. R. et al. 2001 *Virchows Arch* 438:107-15), from which dengue virus sequences were identified using RT-PCR (Rosen, L. et al. 1999 *Am J Trop Med Hyg* 61:720-4). Of potential importance to the etiology of severe dengue virus infection, three studies have demonstrated that the mean levels of serum ALT and AST were significantly increased in patients with DHF/DSS compared to those with DF (Kalayanarooj, S. et al. 1997 *J Infect Dis* 176:313-21; Mohan, B. et al. 2000 *J Trop Pediatr* 46:40-3; Wahid, S. F. et al. 2000 *Southeast Asian J Trop Med Public Health* 31:259-63). As expected, elevation of serum liver enzymes has previously been observed in clinical trials of DEN virus vaccine candidates (Example 8; Eckels, K. H. et al. 1984 *Am J Trop Med Hyg* 33:684-9; Edelman, R. et al. 1994 *J Infect Dis* 170:1448-55; Kanesa-thasan, N. et al. 2001 *Vaccine* 19:3179-3188; Vaughn, D. W. et al. 1996 *Vaccine* 14:329-36).

Based on the increasing disease burden associated with DEN virus infection over the past several decades, a vaccine which confers protection against the four dengue virus serotypes is needed, but none is presently licensed. Because of the increased risk for severe DHF/DSS associated with secondary infection with a heterologous DEN virus serotype (Burke, D. S. et al. 1988 *Am J Trop Med Hyg* 38:172-80; Halstead, S.

B. et al. 1977 *J Exp Med* 146:218-29; Thein, S. et al. 1997 *Am J Trop Med Hyg* 56:566-72), an effective vaccine must confer simultaneous protection against each of the four DEN virus serotypes. Several approaches are presently being pursued to develop a tetravalent vaccine against the dengue viruses (Bancroft, W. H. et al. 1984 *J Infect Dis* 149:1005-10; Bhamarapavati, N. & Sutee, Y. 2000 *Vaccine* 18:44-7; Butrapet, S. et al. 2000 *J Virol* 74:3011-9; Guirakhoo, F. et al. 2000 *J Virol* 74:5477-85; Huang, C. Y. et al. 2000 *J Virol* 74:3020-8; Kanesa-thasan, N. et al. 2001 *Vaccine* 19:3179-3188). One such approach, a live-attenuated DEN4 vaccine candidate, termed 2AΔ30, was both attenuated and immunogenic in a cohort of 20 volunteers (Example 8). The recombinant 2AΔ30 virus contains a 30 nt deletion in the 3' UTR which removes nucleotides 10,478-10,507 and was found to produce a low or undetectable level of viremia in vaccinees at a dose of 10<sup>5</sup> PFU/vaccinee. An asymptomatic rash was reported in 50% of volunteers, and the only laboratory abnormality observed was an asymptomatic, transient rise in the serum ALT level in 5 of the 20 vaccinees. All 2AΔ30 vaccinees developed serum neutralizing antibodies against DEN4 virus (mean titer: 1:580), and 2AΔ30 was not transmitted to mosquitoes that fed experimentally on vaccinees (Troyer, J. M. et al. 2001 *Am J Trop Med Hyg* 65:414-9). Because of the desirable properties conferred by the Δ30 mutation, chimeric vaccine candidates are being constructed which contain the structural genes of DEN virus type 1, 2, and 3, in the attenuated DEN4 background bearing the genetically stable Δ30 mutation. Attenuating mutations outside of the structural genes are particularly attractive for inclusion in antigenic chimeric vaccine candidates because they will not affect the infectivity or immunogenicity conferred by the major mediator of humoral immunity to DEN viruses, the envelope (E) protein.

The presence of rash and elevated ALT levels suggests that the 2AΔ30 vaccine candidate may be slightly under-attenuated in humans. Similarly, many previous attempts to develop live attenuated dengue virus vaccines have yielded vaccine candidates that were either over- or under-attenuated in humans, some of which also induced elevation of serum ALT and AST levels (Bhamarapavati, N. & Yoksan, S. 1997 in: *Dengue and Dengue Hemorrhagic Fever* D. J. Gubler & G. Kuno eds. pp. 367-377 CAB International, New York; Eckels, K. H. et al. 1984 *Am J Trop Med Hyg* 33:684-9; Innis, B. L. et al. 1988 *J Infect Dis* 158:876-80; Kanesa-thasan, N. et al. 2001 *Vaccine* 19:3179-3188; McKee, K. T., Jr. et al. 1987 *Am J Trop Med Hyg* 36:435-42). Therefore, we have developed a menu of point mutations conferring temperature-sensitive (ts), small-plaque (sp), and attenuation (att) phenotypes capable of attenuating DEN4 viruses to a varying degree (Example 1). We have previously described 20 mutant viruses that exhibit a ts, but not sp, phenotype in Vero cells or HuH-7 liver cells and that show attenuated replication in mouse brain (Example 1). Addition of such mutations to 2AΔ30 or to other dengue virus vaccine candidates is envisioned as yielding vaccine candidates that exhibit a more satisfactory balance between attenuation and immunogenicity.

In the present Example, we have extended our analysis of the panel of 1,248 DEN4 virus clones previously generated by mutagenesis with 5-fluorouracil (5-FU) (Example 1), by identifying a set of 22 sp mutant viruses, some of which also have a ts phenotype. Small plaque mutant viruses were sought since such viruses are often attenuated in humans (Bhamarapavati, N. & Yoksan, S. 1997 in: *Dengue and Dengue Hemorrhagic Fever* D. J. Gubler & G. Kuno eds. pp. 367-377 CAB International, New York; Butrapet, S. et al. 2000 *J Virol* 74:3011-9; Crowe, J. E. Jr. et al. 1994 *Vaccine* 12:783-790;

Crowe, J. E. Jr. et al. 1994 *Vaccine* 12:691-699; Eckels, K. H. et al. 1980 *Infect Immun* 27:175-80; Innis, B. L. et al. 1988 *J Infect Dis* 158:876-80; Murphy, B. R. & Chanock, R. M. 2001 in: *Fields Virology* D. M. Knipe, et al. Eds. Vol. 1, pp. 435-468 Lippincott Williams & Wilkins, Philadelphia; Takemoto, K. K. 1966 *Prog Med Virol* 8:314-48). Because natural infection with dengue viruses and vaccination with 2AΔ30 may be associated with liver toxicity in humans, we identified mutant viruses with restricted replication in human liver cells. Accordingly, viruses were screened for plaque size and temperature-sensitivity in the human hepatoma cell line, HuH-7, as well as in Vero cells. Here we describe the ts phenotype, nucleotide sequence, and growth properties in suckling mice of 22 sp DEN4 mutant virus clones.

Cells and viruses. WHO Vero cells (African green monkey kidney cells) and HuH-7 cells (human hepatoma cells) (Nakabayashi, H. et al. 1982 *Cancer Res* 42:3858-63) were maintained as described in Example 1. DEN4 2A virus is a wild type virus derived from a cDNA clone of DEN4 strain 814669 (Dominica, 1981) (Lai, C. J. et al. 1991 *PNAS USA* 88:5139-43; Mackow, E. et al. 1987 *Virology* 159:217-28). The nucleotide sequence of DEN4 2A, the parent of the 5-FU mutant viruses, was previously assigned GenBank accession number AF375822 (Example 1). The DEN4 vaccine candidate, 2AΔ30, (Example 8) contains a 30 nt deletion in the 3' untranslated region (UTR) which removes nucleotides 10,478-10,507 (Men, R. et al. 1996 *J Virol* 70:3930-7). The cDNA clones p4, a modified derivative of the DEN4 2A cDNA clone, and p4Δ30 were used to generate recombinant wild type and attenuated viruses, rDEN4 and rDEN4Δ30, respectively (Example 8). GenBank accession numbers were previously assigned as follows (virus: accession number): DEN4 strain 814669: AF326573; 2AΔ30: AF326826; rDEN4: AF326825; rDEN4Δ30: AF326827.

Generation and biological cloning of mutant viruses with a sp phenotype. The generation of 1,248 virus clones from a pool of 5-fluorouracil-mutagenized DEN4 2A has been previously described (Example 1). Briefly, monolayers of Vero cells were infected with DEN4 2A at a multiplicity of infection (MOI) of 0.01 and overlaid with MEM supplemented with 2% FBS and 1 mM 5-fluorouracil (5-FU) (Sigma, St. Louis, Mo.), which reduced replication of DEN4 2A 100-fold. Vero cells in 96-well plates were inoculated with the 5-FU treated virus suspension, and virus clones were harvested from plates receiving terminally-diluted virus. A total of 1,248 virus clones were generated from the cultures treated with 1 mM 5-FU. Two virus clones, 2A-1 and 2A-13, were generated in the same manner from control cultures not treated with 5-FU and served as parallel-passaged control viruses with a wild type phenotype.

Evaluation of in vitro plaque size and temperature sensitivity. The 1,248 5-FU-mutagenized virus clones were screened for temperature sensitivity by assessing virus replication at 35° C. (permissive temperature) and 39° C. (restrictive temperature) in Vero and HuH-7 cells. Cell monolayers in 96-well plates were inoculated with serial ten-fold dilutions of virus and replicate plates were incubated at 35° C. and 39° C. for five days in temperature-controlled water baths. Virus replication was determined by immunoperoxidase staining as previously described (Example 1). A collection of 193 5-FU virus clones demonstrated a 100-fold or greater reduction in titer at 39° C. in either cell line, and these presumptive ts viruses were further characterized. The efficiency of plaque formation (EOP) at permissive and restrictive temperatures and the plaque size of each of the 193 virus clones were determined as follows. Serial ten-fold dilutions of virus suspension were inoculated onto confluent Vero cell and HuH-7

cell monolayers in replicate 24-well plates. After incubation at 35° C. for two hours, monolayers were overlaid with 0.8% methylcellulose (EM Science, Gibbstown, N.J.) in L-15 medium (Quality Biologicals, Gaithersburg, Md.) supplemented with 2% FBS, gentamicin, and L-glutamine. After incubation of replicate plates for five days at 35, 37, 38, or 39° C. in temperature-controlled water baths, plaques were visualized by immunoperoxidase staining and counted as previously described. Plaque size of each of the 193 viruses was evaluated at the permissive temperature (35° C.) and compared to that of DEN4 2A-13 parallel-passaged control virus with a wild type plaque size. Mutant viruses incubated at the permissive temperature of 35° C. which had a plaque size  $\leq 1$  mm or  $\leq 0.4$  mm (approximately  $\leq 50\%$  the size of wild type DEN4 2A-13) in Vero or HuH-7 cells, respectively, were designated as having a sp phenotype. The level of temperature sensitivity and plaque size of each virus was confirmed in at least two separate experiments. Seventy-five viruses which were confirmed to have a putative ts and/or sp phenotype were biologically cloned an additional two times and phenotypes were re-assessed. Twenty-two of the 75 terminally diluted viruses were found to have a sp phenotype. Sixteen of the 22 sp mutant viruses were also found to have a ts phenotype as defined by a 2.5 or 3.5  $\log_{10}$  PFU/ml reduction in virus titer in Vero or HuH-7 cells, respectively, at restrictive temperature compared to the permissive temperature of 35° C. as previously described (Example 1). Twenty of the 75 terminally-diluted viruses were found to have a ts phenotype without a sp phenotype and were previously described (Example 1). The remainder of the 75 viruses did not meet either criteria for a ts or sp mutant virus.

Evaluation of sp mutant viruses for restricted replication in suckling mice. Animal experiments were carried out in accordance with the regulations and guidelines of the National Institutes of Health, Bethesda, Md. Growth of DEN4 5-FU mutant viruses was determined in Swiss Webster suckling mice (Taconic Farms, Germantown, N.Y.). Groups of six seven-day-old mice were inoculated intracerebrally with  $10^4$  PFU of virus in 30  $\mu$ l Opti-MEM I (Invitrogen) and the brain of each mouse was removed five days later and individually analyzed as previously described (Example 1). Clarified supernatants of 10% suspensions of mouse brain were frozen at -70° C., and the virus titer was determined by plaque assay in Vero cells.

Determination of the complete genomic sequence of the sp mutant viruses. The nucleotide sequence of the 5-FU-mutagenized DEN4 viruses was determined as described in Example 8. Briefly, genomic RNA was isolated from virus clones and cDNA was prepared by reverse transcription and served as template for the generation of overlapping PCR fragments. A panel of primers was designed to sequence both strands of the PCR product from which consensus sequences were assembled and analyzed. The nucleotide sequence of the 5' and 3' regions of the virus genome was determined after circularization of the RNA genome as described in Example 8.

Identification of DEN4 5-fluorouracil mutant viruses with a sp phenotype. The generation of a panel of 1,248 virus clones from a wild type DEN4 2A virus suspension mutagenized by 5-FU has been described previously (Example 1). In the present study twenty-two mutant viruses with a sp phenotype were identified. The plaque size of representative mutant viruses is illustrated in FIG. 3. The plaque size of DEN4 2A-13 virus (a parallel-passaged virus with a wild type phenotype derived from control cultures not treated with 5-FU) was consistently smaller in HuH-7 cells than that observed in Vero cells (FIG. 3A). Mutant viruses #569 and #1189 (FIG.

3B) were sp in both Vero and HuH-7 cells. In contrast, 5-FU mutant virus clones #311 and #1083 (FIG. 3C) were sp in only HuH-7 cells, suggesting a liver cell-specific defect in replication within this phenotypic group. As indicated in Table 7, five mutant viruses were found to have a sp phenotype in both Vero and HuH-7 cells while 17 viruses had a sp phenotype in only HuH-7 cells. Each 5-FU mutant virus clone was compared for a sp or ts phenotype with three control viruses, 2A-13, wild type rDEN4, and rDEN4Δ30. The recombinant viruses, rDEN4 and rDEN4Δ30, each had a plaque size in Vero and HuH-7 cells similar to that of DEN4 2A-13 indicating that the Δ30 mutation does not confer a sp phenotype (Table 7).

Most of the sp 5-FU mutant viruses also had a ts phenotype in Vero and/or HuH-7 cells (Table 7) since mutant viruses were initially screened for temperature sensitivity. Temperature-sensitivity was defined as a 2.5 or 3.5 log<sub>10</sub> PFU/ml reduction in virus titer in Vero or HuH-7 cells, respectively, at restrictive temperature compared to the permissive temperature of 35° C. as previously defined (Example 1). Three mutant viruses (#574, #1269 and #1189) were sp and ts in both Vero and HuH-7 cells, while nine mutant viruses (#506-326 in Table 7) were found to be ts in both cell types but sp only in HuH-7 cells. Four viruses (#1104, 952, 738, and 1083) were found to have a wild type phenotype in Vero cells but were both sp and ts in HuH-7 cells. These four mutant viruses each had a 6,000- to 600,000-fold reduction in virus titer at 39° C. in HuH-7 cells with only a 6- to 40-fold reduction at 39° C. in Vero cells. Finally, sp mutant viruses were identified which did not have a ts phenotype in either cell line; two of these viruses (#569 and #761) were sp in both Vero and HuH-7 cells and four viruses (#1096-1012) were sp in only HuH-7 cells (Table 7). As described previously, the Δ30 mutation did not confer temperature-sensitivity in either cell line (Example 1).

The sp 5-FU mutant viruses have restricted replication in suckling mouse brain. The 22 sp DEN4 5-FU mutant viruses were evaluated for their ability to replicate in the brain of one-week-old suckling mice. As a marker for in vivo attenuation, their level of replication was compared with that of the parallel-passaged control virus with a wild type phenotype, 2A-13 (Table 7). Nineteen of 22 sp mutant viruses had a greater than 100-fold reduction in virus replication in the brain of suckling mice compared to 2A-13 and nine viruses had a reduction of greater than 10,000-fold.

The five mutant viruses which were sp in both Vero and HuH-7 cells were 5,000-fold to 100,000-fold restricted in replication compared to 2A-13. Two of these mutant viruses, #569 and #761, were not ts in either cell line but had a reduction in virus titer of greater than 10,000-fold in mouse brain, indicating that the sp phenotype in both Vero and HuH-7 cells is an important surrogate marker for attenuated replication in suckling mouse brain. 5-FU mutant viruses which were sp in only HuH-7 cells had a more variable range of replication in mouse brain. Three viruses had a mean reduction in virus titer of less than 10-fold when compared to 2A-13 virus. However, 8 of 13 viruses which were ts in Vero and/or HuH-7 cells but sp in only HuH-7 cells had a greater than 5,000-fold reduction in virus replication. The results of the in vivo replication analysis of the previously described 20 ts 5-FU mutant viruses (Example 1) and the 22 sp mutant viruses are summarized in Table 8. Mutant viruses with both a sp and ts phenotype were found to have a significantly greater level of attenuation in the brain of suckling mice when compared to viruses with only a ts phenotype.

Sequence analysis of the sp 5-FU mutant viruses. To initiate an analysis of the genetic basis of the ts, sp, or att

phenotype of the 22 sp mutant viruses, the complete nucleotide sequence of each virus genome was determined and is summarized in Table 9 (sp in Vero and HuH-7 cells) and Table 10 (sp in only HuH-7 cells). All identified mutations were nucleotide substitutions, as deletions or insertions were not observed. Point mutations were distributed throughout the genome, including the 3' UTR as well as in all coding regions. Because all 5-FU mutant viruses were found to have at least two mutations (two to six), the observed phenotypes cannot be directly attributed to a specific mutation. The majority of sp viruses also contained translationally silent point mutations (none to four) in the structural or non-structural coding regions. However, these silent mutations are not expected to contribute to the observed phenotypes. Six of the 22 sp mutant viruses (Tables 9 and 10) were found to have mutations in only the NS genes and/or the 3' UTR, indicating that the sp phenotype can be conferred by mutations outside of the structural genes.

Presence of identical mutations in multiple 5-FU mutant viruses. Analysis of the complete nucleotide sequence data for the 5-FU mutant viruses identified several repeated mutations which were present in two or more viruses. Such mutations were also identified previously during our analysis of twenty 5-FU mutant viruses with a ts but not sp phenotype (Example 1). Because these mutations occurred in viruses together with additional mutations, the contribution of the repeated mutations to the observed sp, ts, and att phenotypes remains empirical. Table 11 lists the repeated mutations found among the 20 ts (not sp) mutant viruses described previously (Example 1) and the 22 sp mutant viruses described here. Repeated mutations were identified in the following genes: two in E, two in NS3, five in NS4B, one in NS5, and two in the 3' UTR. Interestingly, within a thirty nucleotide region of NS4B (nt 7153-7182), there were five different nucleotide substitutions which were found in sixteen viruses. Also at nt 7,546 in NS4B, an amino acid substitution (Ala→Val) was found in 10 different 5-FU mutant viruses. The significance of these repeated mutations in NS4B as well as in other DEN4 genomic regions remains empirical, but a reasonable explanation for this phenomenon is that these mutations are involved in adaptation of DEN4 virus for efficient growth in Vero cells, as further discussed in Example 6.

Discussion. As part of a molecular genetic vaccine strategy, we have developed attenuating mutations that are envisioned as being useful in the development of a live attenuated tetravalent dengue virus vaccine. Specifically, mutations which restrict replication of the vaccine virus in human liver cells were generated since there was some residual virulence of the rDEN4Δ30 vaccine candidate for the liver of humans. Mutant viruses with a sp phenotype were sought in both Vero cells and HuH-7 human liver cells, in order to identify host-range mutant viruses that were specifically restricted in replication in HuH-7 cells (sp in HuH-7 but not in Vero). Such mutations are envisioned as being useful in limiting replication of a candidate vaccine in the liver of vaccinees while preserving both efficient replication in Vero cells and immunogenicity in vivo.

Several observations from the present study indicate that sp mutations confer an att phenotype in vivo. This is not surprising since attenuation in suckling mouse brain has been reported for live DEN virus vaccine candidates possessing sp phenotypes, including the DEN2 PDK-53 and DEN2 PR-159/S-1 vaccine strains (Bhamarapavati, N. & Yoksan, S. 1997 in: *Dengue and Dengue Hemorrhagic Fever* D. J. Gubler & G. Kuno eds. pp. 367-377 CAB International, New York; Butrapet, S. et al. 2000 *J Virol* 74:3011-9; Eckels, K. H. et al.

1980 *Infect Immun* 27:175-80; Innis, B. L. et al. 1988 *J Infect Dis* 158:876-80). Each of 22 DEN4 5-FU mutant viruses with a sp phenotype (some of which were also ts) in either Vero or HuH-7 cells manifested restricted replication in the brains of mice. Six 5-FU mutant viruses with a sp phenotype in the absence of a ts phenotype were more attenuated in the brains of suckling mice than mutant viruses with solely a ts phenotype (Example 1), indicating that the sp phenotype specifies a greater level of attenuation for mouse brain than does the ts phenotype. Mutant viruses with both a ts and sp phenotype had an even greater reduction in replication, further indicating that the attenuation conferred by the ts and sp phenotypes can be additive. Importantly, seventeen of the 22 sp mutant viruses were host-range sp mutant viruses, being sp only in HuH-7 cells. Since such mutations are envisioned as being useful in restricting the replication of a DEN4 virus in human liver cells, we used nucleotide sequence analysis to determine the genetic basis of the sp phenotype.

Analysis of the complete genomic sequence of the 22 sp DEN4 viruses revealed substitutions in the 3' UTR as well as coding mutations in all genes except NS4A. It was first noted that several specific mutations were present in two or more of the 22 sp DEN4 mutant viruses and that many of these same mutations were also previously identified among the set of 20 ts DEN4 mutant viruses (Example 1). Since flaviviruses can rapidly accumulate mutations during passage in tissue culture (Dunster, L. M. et al. 1999 *Virology* 261:309-18; Mandl, C. W. et al. 2001 *J Virol* 75:5627-37), many of these over-represented mutations, previously referred to as putative Vero cell adaptation mutations (Example 1), likely promote efficient replication in Vero cells and were selected unintentionally during the biological cloning of the mutant viruses. The effect of these mutations on DEN virus replication in Vero cells, the proposed substrate for vaccine manufacture, is discussed in Example 6.

The sp mutations identified among the 5-FU mutant viruses are envisioned as being useful in several different approaches for the development of DEN virus vaccine strains. As described above for the generation of antigenic chimeric viruses, one or more sp attenuating mutations are envisioned as being added to the attenuated DEN4 $\Delta$ 30 genetic background to supplement the att phenotype of the  $\Delta$ 30 mutation. A second approach is to introduce a sp attenuating mutation, with or without  $\Delta$ 30, into infectious cDNA clones of the other three DEN serotypes. The ability to transfer mutations among genetically-related viruses and maintain similar att phenotypes has been previously demonstrated (Skiadopoulos, M. H. et al. 1999 *Virology* 260: 125-35). These distinct strategies are envisioned as being useful as separate or complementary approaches to the construction of a tetravalent DEN virus vaccine, underlining the importance of the identification of a large panel of att mutations within the DEN viruses.

### EXAMPLE 3

#### Recombinant DEN4 Viruses Containing Mutations Identified in 5-FU Mutant Viruses Show Restricted Replication in Suckling Mouse Brain and in SCID Mice Transplanted with Human Liver Cells

Data was presented in Examples 1 and 2 that summarizes the generation, characterization and sequence analysis of 42 attenuated mutant DEN4 viruses. For three of the mutant viruses (#239, 489, and 773) with a single missense mutation at nt position 4995 in NS3, it was clear that the identified mutation specified the ts and att phenotypes. This conclusion was confirmed in Example 1 by tissue culture and in vivo

characterization of rDEN4-4995, a recombinant virus into which the 4995 mutation had been introduced by site-directed mutagenesis. In this analysis, rDEN4-4995 exhibited the same level of temperature sensitivity and attenuation as 5-FU mutant viruses #239, 489, and 773. The individual mutation(s) in the remaining 5-FU mutant viruses that specify the observed phenotypes remains to be identified, since most of these viruses possess more than one nucleotide substitution. We have conducted an analysis to identify the mutations in a subset of the other 39 mutant viruses that specify the ts, sp, and att phenotypes by introduction of each mutation into the wt DEN4 cDNA (p4) and evaluation of the phenotypes of the resulting recombinant DEN4 viruses bearing the individual mutations. Previous studies of a DEN2 virus vaccine candidate (Butrapet, S. et al. 2000 *J Virol* 74:3011-9) as well as other virus vaccines (Whitehead, S. S. et al. 1999 *J Virol* 73:871-7) have demonstrated the utility of this approach for the identification of the genetic basis of attenuation.

As described in Examples 1 and 2, 19 5-FU mutant viruses were identified which were found to contain coding mutations in only the NS genes and/or nucleotide substitutions in the 5' or 3' UTR which would facilitate the generation of antigenic chimeric viruses. In the present example, the genetic basis of the observed sp, ts, and mouse brain att phenotypes was identified for these 19 viruses using reverse genetics to generate recombinant DEN4 (rDEN4) viruses containing individual mutations identified in the panel of DEN4 mutant viruses. In addition, the 19 5-FU mutant viruses were evaluated for replication in a novel small animal model for DEN4 virus replication, SCID mice transplanted with HuH-7 cells (SCID-HuH-7), and the genetic basis of the att viruses was identified using mutant rDEN4 viruses. Also presented are findings describing the generation and characterization of a recombinant virus containing two of the identified attenuating mutations as well as combination of select 5-FU mutations with the 430 mutation.

Generation of rDEN4 viruses containing 5-FU mutations. The methods used for the generation of rDEN4 viruses are outlined in FIG. 4 and are similar to those described in Example 1. Briefly, the p4 cDNA was digested with the appropriate restriction enzymes and the resulting fragments were subcloned into a modified pUC 119 vector. For Kunkel mutagenesis, single-stranded DNA preparations of the pUC-NS vectors were made, and primers were designed to individually introduce mutations that were present in the 5-FU mutant viruses. The sequences of the 41 mutagenic oligonucleotides used to generate the single-mutation recombinant viruses are presented in Table 12. Primers were designed to co-introduce or co-ablate a translationally-silent restriction enzyme site in the cDNA, which greatly facilitates the screening and identification of cDNA clones possessing the mutant sequence. Fragments containing the introduced mutations were cloned back into p4, and nucleotide sequence analysis confirmed the presence of the nucleotide changes. A total of 33 rDEN4 viruses was generated which contained each of the individual mutations present in the 19 5-FU mutant viruses containing only coding mutations in the NS genes and/or nucleotide substitutions in the 5' or 3' UTR. An additional 8 rDEN4 viruses were generated from mutations identified in the remaining panel of 42 5-FU mutant viruses.

A cDNA clone was also generated which combined the mutations identified at nt position 4995 in NS3 and 7849 in NS5. The 7849 mutation was introduced into the p4-4995 cDNA clone by replacing the XmaI-PstI fragment with that derived from the p4-7849 cDNA clone. The presence of both mutations was confirmed by sequence analysis. The  $\Delta$ 30 mutation was introduced into the 3' UTR of the individual

mutant cDNA clones by replacing the MluI-KpnI fragment with that derived from the p4Δ30 cDNA clone, and the presence of the deletion was confirmed by sequence analysis.

Recombinant viruses were recovered by transfection of Vero or C6/36 cells with RNA transcripts derived from the mutant cDNA clones as described in Example 1. Recovered viruses were terminally diluted twice and working stocks of viruses were prepared in Vero cells. Each of the mutant cDNA clones was recovered after transfection as expected since the 5-FU mutant viruses containing these mutations were viable.

Characterization of ts and att phenotypes of the rDEN4 viruses containing introduced mutations. Of the 19 5-FU mutant viruses with mutations in only NS genes and/or the 5' or 3' UTR, six had an sp phenotype (Table 13), ten had a ts phenotype in Vero and HuH-7 cells (Table 14), and three had a ts phenotype in only HuH-7 cells (Table 15). For the six sp 5-FU mutant viruses, #738, 922, 1081, 1083, 1136, and 1189, seventeen mutations identified by sequence analysis resulted in a coding change or a nucleotide change in the UTR and each was engineered into an individual DEN4 cDNA clone. Virus containing each defined mutation was successfully recovered and propagated and was tested for efficiency of plaque formation in Vero and HuH-7 cells at various temperatures, plaque size phenotype, and growth properties in suckling mice using methods previously described in Examples 1 and 2.

Table 13 lists the phenotypes of the six sp 5-FU mutant parent viruses and those of the 17 rDEN4 viruses encoding single mutations present in the parent virus. For example, 5-FU mutant #1189 (parent), which was ts and sp in both cell lines and had an almost 10,000-fold reduction in replication in suckling mouse brain, contained 4 coding mutations at nt position 3303 in NS1, 4812 and 5097 in NS3, and 7182 in NS4B. Analysis of the four rDEN4 viruses containing each of these mutations indicated that rDEN4-5097 had a ts, sp, and att phenotype while rDEN4-3303, rDEN4-4812, and rDEN4-7182 had no discernible phenotypes, indicating that the mutation at nt 5097 was responsible for the phenotype observed in the 5-FU parent, #1189. Thus, analysis of the relative contributions of the four mutations present in the 5-FU mutant #1189 to its attenuation phenotype provides the framework for a similar analysis of the remaining 5-FU mutant viruses. This analysis specifically demonstrates the methods used to identify mutations contributing to the observed phenotype. The ts, sp, and att phenotypes of 5-FU parent viruses #738, 922, 1081, and 1083, were similarly attributed to single mutations 3540, 4306, 2650, and 10634, respectively. However, two separate mutations (3771 and 4891) contributed to the phenotypes of 5-FU mutant virus #1136.

Table 14 lists the genetic basis of the ts and mouse brain attenuation for the ten 5-FU mutant viruses with ts phenotypes in both Vero and HuH-7 cells. As described in Example 1, the 4995 mutation which is the only mutation present in three 5-FU mutant viruses, #239, #489, and #773, was found to confer a ts and att phenotype, confirming the genetic basis for the phenotypes exhibited by these viruses. In three separate experiments, the rDEN4-4995 virus was found to have an approximately 1,000-fold decrease in replication in the brains of suckling mice when compared to that of wild-type virus (Table 6 and 14). The 4995 mutation is also present in 5-FU mutant viruses #473, #759, and #816, each of which has additional mutations. The ts and att phenotypes observed in these viruses can be attributed to the 4995 mutation since the additional mutations did not show discernible phenotypes. Interestingly, 5-FU mutant virus #938 has the 4995 mutation and an additional mutation at nt 3442 in NS1 with both mutations independently conferring restricted replication in

mouse brain. The remaining three 5-FU parent viruses in Table 14, #173, #509, and #1033, were found to each contain a single mutation responsible for the att phenotype: 7849, 8092, and 4907, respectively.

Three 5-FU mutant viruses, #686, #992, and #1175 with HuH-7 cell-specific ts phenotypes are listed in Table 15. Mutations in NS3 (5695) and NS5 (10186) were found to confer the phenotypes observed for parent virus #992 and #1175. Interestingly, two mutations in NS2A, 3575 and 4062, were found to result in a synergistic increase in the level of attenuation. Both individual mutations had an approximately 100-fold decrease in virus replication in the brain while the parent virus with both mutations had an almost 10,000-fold reduction. Table 16 lists two additional mutations with an att phenotype, 4896 and 6259 in NS3.

Replication of DEN4 viruses in SCID mice transplanted with HuH-7 cells. Since DEN viruses replicate poorly in the liver of mice and corresponding studies are impractical to conduct in non-human primates, an animal model that evaluates the in vivo level of replication of DEN virus in liver cells was developed based on a recent report examining the replication of DEN virus in SCID mice transplanted with a continuous cell line of human liver tumor cells (An, J. et al. 1999 Virology 263:70-7). SCID mice transplanted with human continuous cell lines, primary cells, or organized tissues have similarly been used to study the replication of other viruses which lack a suitable small animal model (Mosier, D. E. 2000 Virology 271:215-9). In our study, SCID mice were transplanted with HuH-7 cells since DEN4 virus replicated efficiently in these cells in tissue culture and since these were the cells used to define the host-range phenotype. These studies are envisioned as addressing the utility of examining DEN virus infection in SCID mouse-xenograft models for vaccine development (An, J. et al. 1999 Virology 263:70-7; Lin, Y. L. et al. 1998 J Virol 72:9729-37).

To further examine the in vivo growth properties of the 19 5-FU mutant DEN4 viruses with mutations in only the NS genes and/or the 3' UTR and selected corresponding rDEN4 mutant viruses, replication was assessed in SCID mice transplanted with HuH-7 cells (SCID-HuH-7). For analysis of DEN4 virus replication in SCID-HuH-7 mice, four to six week-old SCID mice (Tac:Icr:Ha(ICR)-Prkdc<sup>scid</sup>) (Taconic Farms) were injected intraperitoneally with 10<sup>7</sup> HuH-7 cells suspended in 200 μl phosphate-buffered saline (PBS). In preparation for transplantation, HuH-7 cells were propagated in cell culture as described above and harvested by trypsinization at approximately 80% confluence. Cells were washed twice in PBS, counted, resuspended in an appropriate volume of PBS, and injected into the peritoneum of mice. Tumors were detected in the peritoneum five to six weeks after transplantation, and only mice with apparent tumors were used for inoculation. Mice were infected by direct inoculation into the tumor with 10<sup>4</sup> PFU of virus in 50 μl Opti-MEM I. Mice were monitored daily for seven days and serum for virus titration was obtained by tail-nicking on day 6 and 7. Approximately 400 μl blood was collected in a serum separator tube (Sarstedt, Germany), centrifuged, and serum was aliquoted and stored at -70°C. The virus titer was determined by plaque assay in Vero cells. Seven days after infection, most mice developed morbidity and all mice were sacrificed. Tumors were excised and weighed to confirm uniformity of the experimental groups.

Preliminary experiments indicated that SCID-HuH-7 mice inoculated with DEN4 2A-13 directly into the tumor developed viremia with maximum levels (up to 8.0 log<sub>10</sub>PFU/ml serum) achieved on day 5 (Table 17). Virus could also be detected in brain, liver, and tumor homogenates.

The level of viremia in SCID-HuH-7 mice infected with parental 5-FU or rDEN4 mutant viruses was compared with that of the parallel-passaged control virus, 2A-13, or rDEN4, respectively. Results of 4 separate experiments indicated that the vaccine candidate, rDEN4 $\Delta$ 30, had an almost 10-fold reduction in virus replication compared to wild type rDEN4 (Table 13) which reflects the apparent attenuation of the rDEN4 $\Delta$ 30 vaccine candidate in humans (Example 8). Results in Tables 13 to 15 indicate that three 5-FU mutant viruses had a greater than 100-fold reduction in viremia in the SCID-HuH-7 mice compared to wild type 2A-13 virus: #1081, #1083, and #1189. The common phenotype among these viruses was a sp phenotype in HuH-7 cells. Analysis of the genetic basis of the att phenotype in these parent 5-FU mutant viruses identified three individual mutations in NS1, NS3, and the 3' UTR which conferred at least a 100-fold reduction in viremia. Specifically, rDEN4-2650 (NS1), rDEN4-5097 (NS3), and rDEN4-10634 (3' UTR) manifested a 2.2, 3.6, and 4.3 log<sub>10</sub>PFU/ml reduction in peak titer of viremia compared to rDEN4, respectively. These mutations also conferred the att phenotype in suckling mouse brain. 5-FU mutant virus #738 and #509 had a reduction in viremia in the SCID-HuH-7 mice compared to wild type 2A-13 of 1.9 and 1.5 log<sub>10</sub>PFU/ml, respectively, and the genetic basis for these phenotypes is envisioned as being assessed on an empirical basis.

This analysis of the genetic basis of the phenotypes specified by the mutations in the 5-FU mutant viruses that manifested restricted replication in SCID-HuH-7 mice indicated that (1) three separate mutations conferred the att phenotype; (2) these mutations were located in two proteins, NS1 and NS3, and in the 3' UTR; (3) these three mutations were fully responsible for each of the cell culture (ts or sp) and in vivo (attenuation in mouse brain and SCID-HuH-7 mice) phenotypes of the parent viruses; and (4) two of the three mutations specify the host-range sp phenotype (sp on HuH-7 only) and therefore are envisioned as being useful in a vaccine virus. Although the relevance of such SCID-transplant models to virus replication and disease in humans is unknown, the identification of three novel mutations which restrict DEN4 virus replication in SCID-HuH-7 mice is envisioned as facilitating an examination of the correlation between the att phenotype in SCID-HuH-7 mice with that in rhesus monkeys or humans. Such mutations, specifically the host-range sp mutations, are envisioned as being useful in conjunction with the  $\Delta$ 30 or other mutation to decrease the residual virulence of rDEN4 $\Delta$ 30 or other dengue virus for the human liver, and studies are envisioned as being conducted to construct such rDEN4 viruses and evaluate them in monkeys and humans (Example 8).

Combination of two 5-FU mutations results in an additive ts phenotype. The ability to combine individual mutations in rDEN4 virus as a means to modulate the phenotype of the resulting double mutant virus is a major advantage of using recombinant cDNA technology to generate or modify dengue virus vaccine candidates. Addition of multiple ts and att mutations to recombinant vaccine viruses is envisioned as improving the phenotypic stability of the double recombinant due to the decreased possibility of co-reversion of the two mutations to wild-type virulence (Crowe, J. E. Jr. et al. 1994a Vaccine 12:783-790; Skiadopoulos, M. H. et al. 1998 J Virol 72:1762-8; Subbarao, E. K. et al. 1995 J Virol 69:5969-5977; Whitehead, S. S. et al. 1999 J Virol 73:871-7). The mutations identified at nt position 4995 in NS3 and 7849 in NS5 were combined in a single p4 cDNA clone and a recombinant virus, designated rDEN4-4995-7849, was recovered and evaluated for its ts and att phenotypes (Table 18). rDEN4-4995-7849

was more ts than either recombinant virus containing the individual mutations (Table 18), indicating the additive effect of the two ts mutations. The rDEN4-4995-7849 virus had a greater than 10,000-fold reduction in replication in the brains of suckling mice. The reduction in replication of the double mutant virus was only slightly increased over that of rDEN4-7849, however, a difference in the level of replication between rDEN4-4995-7849 and rDEN4-7849 would be difficult to detect since the level of replication of both viruses was close to the lower limit of detection (2.0 log<sub>10</sub>PFU/g brain).

Combination of selected 5-FU mutations with the 430 mutation confers increased attenuation of rDEN4 $\Delta$ 30 for the brains of suckling mice. To define the effect of adding individual mutations to the attenuated rDEN4 $\Delta$ 30 background, five combinations have been constructed: rDEN4 $\Delta$ 30-2650, rDEN4 $\Delta$ 30-4995, rDEN4 $\Delta$ 30-5097, rDEN4 $\Delta$ 30-8092, and rDEN4 $\Delta$ 30-10634. Addition of such missense mutations with various ts, sp, and att phenotypes is envisioned as serving to decrease the reactogenicity of rDEN4 $\Delta$ 30 while maintaining sufficient immunogenicity.

The  $\Delta$ 30 mutation was introduced into the 3' UTR of the individual mutant cDNA clones by replacing the MluI-KpnI fragment with that derived from the p4 $\Delta$ 30 cDNA clone, and the presence of the deletion was confirmed by sequence analysis. Recombinant viruses were recovered by transfection in C6/36 cells for each rDEN4 virus. However, upon terminal dilution and passage, the rDEN4 $\Delta$ 30-5097 virus was found to not grow to a sufficient titer in Vero cells and was not pursued further. This is an example of a cDNA in which the 5-FU mutation and the  $\Delta$ 30 mutation are not compatible for efficient replication in cell culture. To begin the process of evaluating the in vivo phenotypes of the other four viruses which replicated efficiently in cell culture, rDEN4 viruses containing the individual mutations and the corresponding rDEN4 $\Delta$ 30 combinations were tested together for levels of replication in suckling mouse brain. The results in Table 19 indicate that addition of each of the mutations confers an increased level of attenuation in growth upon the rDEN4 $\Delta$ 30 virus, similar to the level conferred by the individual 5-FU mutation. No synergistic effect in attenuation was observed between the missense mutations and  $\Delta$ 30. These results indicate that the missense mutations at nucleotides 2650, 4995, 8092, and 10634 are compatible with  $\Delta$ 30 for growth in cell culture and in vivo and can further attenuate the rDEN4 $\Delta$ 30 virus in mouse brain. Further studies in SCID-HuH-7 mice, rhesus monkeys, and humans are envisioned as establishing the effect of the combination of individual mutations and  $\Delta$ 30 upon attenuation and immunogenicity (Example 8).

By identifying the specific mutations in the 5-FU mutant viruses which confer the observed phenotypes, a menu of defined ts, sp, and att mutations is envisioned as being assembled (see Example 7). Numerous combinations of two or more of these mutations are envisioned as being selected with or without the  $\Delta$ 30 mutation. Such mutations and their combinations are envisioned as being useful for the construction of recombinant viruses with various levels of in vivo attenuation, thus facilitating the generation of candidate vaccines with acceptable levels of attenuation, immunogenicity, and genetic stability.

#### EXAMPLE 4

##### Generation of DEN4 Mutant Viruses with Temperature-Sensitive and Mouse Attenuation Phenotypes Through Charge-Cluster-to-Alanine Mutagenesis

The previous Examples described the creation of a panel of DEN4 mutant viruses with ts, sp, and att phenotypes obtained

through 5-FU mutagenesis. As indicated in these Examples, the attenuating mutations identified in the 5-FU mutant viruses are envisioned as having several uses including (1) fine tuning the level of attenuation of existing dengue virus vaccine candidates and (2) generation of new vaccine candidates by combination of two or more of these attenuating mutations. In the current example, we created a second panel of mutant viruses through charge-cluster-to-alanine mutagenesis of the NS5 gene of DEN4 and examined the resulting mutant viruses for the ts, sp, and att phenotypes as described in Examples 1 and 2. The charge-cluster-to-alanine mutant viruses recovered demonstrated a range of phenotypes including ts in Vero cells alone, ts in HuH-7 cells alone, ts in both cell types, att in suckling mouse brains, and att in SCID-HuH-7 mice.

The usefulness of mutant viruses expressing these phenotypes has already been described, however charge-cluster-to-alanine mutant viruses possess some additional desirable characteristics. First, the relevant mutations are envisioned as being designed for use in the genes encoding the non-structural proteins of DEN4, and therefore are envisioned as being useful to attenuate DEN1, DEN2, and DEN3 antigenic chimeric recombinants possessing a DEN4 vector background. Second, the phenotype is usually specified by three or more nucleotide changes, rendering the likelihood of reversion of the mutant sequence to that of the wild type sequence less than for a single point mutation, such as mutations identified in the panel of 5-FU mutant viruses. Finally, charge-cluster-to-alanine attenuating mutations are envisioned as being easily combinable among themselves or with other attenuating mutations to modify the attenuation phenotype of DEN4 vaccine candidates or of DEN1, DEN2, and DEN3 antigenic chimeric recombinant viruses possessing a DEN4 vector background.

**Charge-Cluster-to-Alanine-Mutagenesis.** The cDNA p4, from which recombinant wild type and mutant viruses were generated, has been described in Examples 1, 2, and 3 and in FIG. 4. Charge-cluster-to-alanine mutagenesis (Muylaert, I. R. et al. 1997 *J Virol* 71:291-8), in which pairs of charged amino acids are replaced with alanine residues, was used to individually mutagenize the coding sequence for 80 pairs of contiguous charged amino acids in the DEN4 NS5 gene. Subclones suitable for mutagenesis were derived from the full length DEN4 plasmid (p4) by digestion with XmaI/PstI (pNS5A), PstI/SacII (pNS5B) or SacII/MluI (pNS5C) at the nucleotide positions indicated in FIG. 4. These fragments were then subcloned and Kunkel mutagenesis was conducted as described in Examples 1 and 3. To create each mutation, oligonucleotides were designed to change the sequence of individual pairs of codons to GCAGCX (SEQ ID NO: 69), thereby replacing them with two alanine codons (GCX) and also creating a BbvI restriction site (GCAGC) (SEQ ID NO: 70). The BbvI site was added to facilitate screening of cDNAs and recombinant viruses for the presence of the mutant sequence. Restriction enzyme fragments bearing the alanine mutations were cloned back into the full-length p4 plasmid as described in Examples 1 and 3.

Initial evaluation of the phenotype of the 32 charge-cluster-to-alanine mutant viruses revealed a range in restriction of replication in suckling mouse brain and SCID-HuH-7 mice. To determine whether attenuation could be enhanced by combining mutations, double mutant viruses carrying two pairs of charge-cluster-to-alanine mutations were created by swapping appropriate fragments carrying one pair of mutations into a previously-mutagenized p4 cDNA carrying a second pair of mutations in a different fragment using conventional cloning techniques.

**Transcription and Transfection.** 5'-capped transcripts were synthesized in vitro from mutagenized cDNA templates using AmpliCap SP6 RNA polymerase (Epicentre, Madison, Wis.). Transfection mixtures, consisting of 1 µg of transcript in 60 µl of HEPES/saline plus 12 µl of dioleoyl trimethylammonium propane (DOTAP) (Roche Diagnostics Corp., Indianapolis, Ind.), were added, along with 1 ml Virus production-serum free medium (VP-SFM) to subconfluent monolayers of Vero cells in 6-well plates. Transfected monolayers were incubated at 35° C. for approximately 18 hr, cell culture medium was removed and replaced with 2 ml VP-SFM, and cell monolayers were incubated at 35° C. After 5 to 6 days, cell culture medium was collected, and the presence of virus was determined by titration in Vero cells followed by immunoperoxidase staining as previously described. Recovered virus was amplified by an additional passage in Vero cells, and virus suspensions were combined with SPG (sucrose—phosphate—glutamate) stabilizer (final concentration: 218 mM sucrose, 6 mM L-glutamic acid, 3.8 mM potassium phosphate, monobasic, and 7.2 mM potassium phosphate, dibasic, pH 7.2), aliquoted, frozen on dry ice, and stored at -70° C.

cDNA constructs not yielding virus after transfection of Vero cells were used to transfect C6/36 cells as follows. Transfection mixtures, as described above, were added, along with 1 ml of MEM containing 10% fetal bovine serum (FBS), 2 mM L-glutamine, 2 mM non-essential amino acids, and 0.05 mg/ml gentamicin, to monolayers of C6/36 cells. Transfected cell monolayers were incubated at 32° C. for 18 hr, cell culture medium was removed and replaced with 2 ml fresh medium, and cell monolayers were incubated at 32° C. After 5 to 6 days, cell culture media were then used to infect Vero cells and incubated for 5-6 days, at which time cell culture media were collected, frozen and titered as described above.

Recovered viruses were biologically cloned by two rounds of terminal dilution in Vero cells followed by an additional amplification in Vero cells. Briefly, virus was initially diluted to a concentration of approximately 20 PFU/ml in VP-SFM and then subjected to a series of two-fold dilutions across a 96-well plate. Virus dilutions were used to infect Vero cell monolayers in a 96-well plate and incubated for 5 to 6 days at 35° C. Following incubation, cell culture media were removed and temporarily stored at 4° C., and the virus-positive cell monolayers were identified by immunoperoxidase staining. Terminal dilution was achieved when ≤25% of cell monolayers were positive for virus. Cell culture medium from a positive monolayer at the terminal dilution was subjected to an additional round of terminal dilution. Following the second terminal dilution, virus was amplified in Vero cells (75 cm<sup>2</sup> flask), collected and frozen as previously described.

**Assays for temperature-sensitivity and mouse attenuation.** Assay of the level of temperature sensitivity of the charge-cluster-to-alanine mutant viruses in Vero and HuH-7 cells and their level of replication in the brain of suckling mice were conducted as described in Example 1 and assay of the level of replication in SCID-HuH-7 mice was conducted as described in Example 3.

Charge-cluster-to-alanine mutant viruses are viable and show temperature-sensitive and mouse attenuation phenotypes. Of 80 full-length DEN4 cDNA constructs containing a single pair of charge-to-alanine mutations, virus was recovered from 32 in either Vero or C6/36 cells (FIG. 5). The level of temperature sensitivity of wt rDEN4, rDEN4Δ30, and the



32 mutant viruses is summarized in Table 20. One mutant virus (645-646) was ts in Vero but not HuH-7 cells and 7 mutant viruses were ts in HuH-7 but not Vero cells. Such mutants whose temperature sensitivity is host-cell dependent are referred to as temperature-sensitive, host-range (tshr) mutants. Thirteen mutant viruses were ts in both cell types, and 11 mutant viruses were not ts on either cell type. Thus a total of 21 mutant viruses were ts with 8 mutant viruses exhibiting an tshr specificity. None of the mutant viruses showed a small plaque phenotype at permissive temperature. Mutant viruses showed a wide range (0 to 10,000-fold) of restricted replication in suckling mouse brain (Table 20). Fourteen mutant viruses were attenuated in suckling mouse brain, arbitrarily defined as a  $\geq 1.5 \log_{10}$ -unit reduction in virus titer. There was no correlation between attenuation in mouse brain and temperature sensitivity in either Vero cells (Kendall Rank correlation:  $P=0.77$ ) or HuH-7 cells (Kendall Rank correlation:  $P=0.06$ ).

Thirteen mutant viruses that either showed an att phenotype in suckling mouse brain or whose unmutated charged amino acid pair was highly conserved among the four DEN serotypes (see Example 7) were assayed for att in SCID-HuH-7 mice (Table 21). Three of these mutant viruses showed  $>100$ -fold decrease in replication relative to wild type DEN4. Overall, mean log reduction from wild type in suckling mice did not show significant correlation with mean log reduction in SCID-HuH-7 mice (Spearman rank correlation,  $N=13$ ,  $P=0.06$ ). However, mutant virus 200-201 was unusual in that it showed a high level of restriction in SCID-HuH-7 mice but little restriction in suckling mouse brain. When virus 200-201 was removed from the analysis, restriction of replication in suckling and SCID-HuH-7 mice showed a significant correlation (Spearman rank correlation,  $N=12$ ,  $P=0.02$ ).

Combining charge-cluster-to-alanine mutations present in two viruses into one virus can enhance its ts and att phenotypes. Six paired mutations were combined into fourteen double-pair mutant viruses, of which six could be recovered in Vero or C6/36 cells (Table 22). All of the individual paired mutations used in double-pair mutant viruses were ts on HuH-7 cells, none was ts in Vero cells, and for all combinations at least one mutation pair conferred an att phenotype in suckling mouse brain. Evaluation of four of the double-pair mutant viruses (Table 23) revealed that combining charge-cluster-to-alanine mutation pairs invariably resulted in the acquisition of a ts phenotype in Vero cells (4 out of 4 viruses) and often resulted in a lowered shutoff temperature in HuH-7 cells (3 out of 4 viruses). In half of the viruses assayed, combination of charge-cluster-to-alanine mutation pairs resulted in enhanced restriction of replication (10-fold greater than either component mutation) in suckling mouse brain (Table 23) and in SCID-HuH-7 mice (Table 24).

Summary. The major usefulness of the charge-cluster-to-alanine mutations stems from their design: they are located in the DEN4 non-structural gene region and therefore are envisioned as being useful to attenuate DEN4 itself as well as antigenic chimeric viruses possessing the DEN4 NS gene region. Furthermore, they are predicted to be phenotypically more stable than the single-nucleotide substitution mutant viruses such as the 5-FU mutant viruses. Finally, combinations of mutations are envisioned as being created in order to fine-tune attenuation and to further stabilize attenuation phenotypes.

### Identification and Characterization of DEN4 Mutant Viruses Restricted in Replication in Mosquitoes

#### SECTION 1. Identification of Viruses Showing Restriction of Replication in Mosquitoes.

In Examples 1 and 4, DEN4 mutant viruses were generated through 5-FU mutagenesis and charge-cluster-to-alanine mutagenesis, respectively, in order to identify mutations that confer ts, sp and att phenotypes. Another highly desirable phenotype of a dengue virus vaccine is restricted growth in the mosquito host. A dengue virus vaccine candidate should not be transmissible from humans to mosquitoes in order to prevent both the introduction of a dengue virus into an environment in which it is currently not endemic and to prevent the possible loss of the attenuation phenotype during prolonged replication in an individual mosquito host. Loss of the attenuation phenotype could also occur following sustained transmission between humans and mosquitoes. Recently, loss of attenuation of a live attenuated poliovirus vaccine was seen following sustained transmission among humans (CDC 2000 MMWR 49:1094).

In the present example, a panel of 1248 DEN4 mutant viruses generated through 5-FU mutagenesis and 32 DEN4 mutant viruses generated through charge-cluster-to-alanine mutagenesis were assayed for restricted growth in mosquito cells. This is a useful preliminary assay for restriction in vivo, since restriction in cultured mosquito cells is often, though not always, associated with poor infectivity for mosquitoes (Huang, C. Y. et al. 2000 J Virol 74:3020-8). Mutant viruses that showed restriction in mosquito cells and robust growth in Vero cells (the substrate for vaccine development, as discussed in Example 6) were targeted for further characterization.

Generation and characterization of the 5-1A1 mutant. The generation and isolation of the panel of 1248 5-FU mutant viruses and the panel of 32 charge-cluster-to-alanine mutant viruses have been described in Examples 1, 2, and 4. Vero and C6/36 cells were maintained as described in Example 1.

Each of the 1248 5-FU mutant viruses and 32 charge-cluster-to-alanine mutant viruses was titered in C6/36 cell monolayers in 24-well plates at 32° C. and 5% CO<sub>2</sub>. After 5 days, plaques were immunostained with anti-DEN4 rabbit polyclonal antibody and counted as described in the preceding Examples. Mutant viruses were assayed for one of two phenotypes indicating restricted growth in mosquito cells: either sp in C6/36 cells relative to Vero cells or a  $\geq 3.5 \log_{10}$  PFU/ml decrease in titer between Vero and C6/36 cells at the permissive temperature for each cell type. Two mutant viruses, one generated by 5-FU mutagenesis (#5) and one generated by charge-cluster-to-alanine mutagenesis (rDEN4-356,357), showed reduced plaque size in C6/36 cells. After three terminal dilutions, the 5-FU mutant #5, designated 5-1A1, maintained the reduced plaque size phenotype. Additionally, recombinant virus rDEN4-7546, tested for Vero cell adaptation (discussed in detail in Example 6) also showed reduced plaque size in C6/36 (FIG. 10).

The multicycle growth kinetics of both 5-1A1 and the recombinant wild type rDEN4 in C6/36 cells were determined as described in Example 1. Briefly, cells were infected in triplicate at a multiplicity of infection of 0.01 and samples were harvested at 24-hr intervals. Samples were flash frozen and titered in a single assay in Vero cell monolayers.

Oral infection of mosquitoes. *Aedes aegypti* is one of the primary vectors of dengue virus (Gubler, D. J. 1998 Clin Microbiol Rev 11:480-96). This species was reared at 26° C.

and 80% relative humidity (RH) with a 16 hr daylight cycle. Adults were allowed continuous access to a cotton pad soaked in a 10% sucrose solution. Five to ten day old female *Ae. aegypti* which had been deprived of a sugar source for 48 hr were fed a bloodmeal consisting of equal volumes of washed human red blood cells, 10% sucrose solution, and dengue virus suspension. The infected blood meal was prepared immediately prior to feeding and offered to mosquitoes in a water-jacketed feeder covered in stretched parafilm and pre-heated to 38° C. (Rutledge, L. C. et al. 1964 *Mosquito News* 24:407-419). Mosquitoes that took a full bloodmeal within 45 min were transferred to a new container by aspirator and maintained as described above. After 21 days, mosquitoes were stored at -20° C. until dissection.

Intrathoracic inoculation of mosquitoes. The large, non-haematophagous mosquito *Toxorhynchites splendens* is a sensitive host for determining the infectivity of dengue virus. This species was reared at 24° C. and 75% RH with a 12 hr daylight cycle. Larvae and pupae were fed on appropriately sized *Aedes* larvae; adults were allowed continuous access to a cotton pad soaked in a 10% sucrose solution. Groups of one to ten day old adult *T. splendens* of both sexes were immobilized by immersion of their container in an icewater bath and inoculated intrathoracically with undiluted virus and serial tenfold dilutions of virus in 1×PBS. Virus was inoculated in a 0.22 µl dose using a Harvard Apparatus microinjector (Medical Systems Corp, Greenvale N.Y.) and a calibrated glass needle (technique is a modification of the method described in Rosen and Gubler, 1974).

Detection of viral antigen in body and head tissues by immunofluorescence assay (IFA). Head and midgut preparations of *Aedes aegypti* and head preparations of *Toxorhynchites splendens* were made on glass slides as described in Sumanochitrapon et al. (Sumanochitrapon, W. et al. 1998 *Am J Trop Med Hyg* 58:283-6). Slides were fixed in acetone for 20 min, and placed at 4° C. until processed by IFA. The primary antibody, hyperimmune mouse ascites fluid specific for DEN-4 (HMAF), was diluted 1/100 in PBS-Tween 20 (0.05%). Slides were incubated at 37° C. in a humid chamber for 30 min, and subsequently rinsed in PBS-Tween 20. The secondary antibody, FITC conjugated goat anti-mouse IgG (KPL, Gaithersburg, Md.), was diluted 1/200 in PBS-Tween 20 with 0.002% Evan's Blue. Slides were viewed on an Olympus BX60 microscope. The infectious dose required to infect 50% of mosquitoes (ID<sub>50</sub>) was determined by the method of Reed and Muench (Reed, L. J. & Muench, H. 1938 *Am J Hyg* 27:493-497). For *Aedes aegypti* infections, two OID<sub>50</sub> (oral infectious dose 50) values were calculated for each virus: the OID<sub>50</sub> required to produce an infection in the midgut, with or without dissemination to the head, and the OID<sub>50</sub> required to produce disseminated infection. For *T. splendens* one MID<sub>50</sub> (mosquito infectious dose 50) value was calculated.

Statistical Analysis. The percentage of mosquitoes infected by different viruses were compared using logistic regression analysis (Statview, Abacus Inc.).

Mutations restricting growth of DEN4 in mosquito cells but not Vero cells are rare. Out of 1280 mutant viruses initially assayed, only two, #5 and rDEN4-356,357, showed reduced plaque size in C6/36 cells and normal plaque size in Vero cells. One additional virus, rDEN4-7546 (described in Example 6), with reduced plaque size in C6/36 was detected in subsequent assays. Mutant virus #5 was cloned by three successive terminal dilutions and designated 5-1A1; rDEN4-7546 and rDEN4-356,357 had already been twice-terminally diluted when they were tested in C6/36 cells. Virus 5-1A1 has been extensively characterized and its phenotypes are

described in detail in the following section. rDEN4-356,357 and rDEN4-7546 are envisioned as being characterized in a similar fashion.

Plaque size and growth kinetics of 5-1A1. 5-1A1 replicated to 6.7 log<sub>10</sub>PFU/ml in Vero cells with normal plaque size and replicated to 7.6 log<sub>10</sub>PFU/ml in C6/36 cells with small plaque size (FIG. 6, Table 25). In comparison, wild type DEN4 used as a concurrent control replicated to 7.3 log<sub>10</sub>PFU/ml in Vero cells, 8.3 log<sub>10</sub>PFU/ml in C6/36 cells, and showed normal plaque size in both cell types (FIG. 6, Table 25). The growth kinetics of 5-1A1 was compared to that of wild type DEN4 by infecting C6/36 cells at an MOI of 0.01 and monitoring the production of infectious virus. The kinetics and magnitude of replication of 5-1A1 in C6/36 cells was comparable to that of wild type DEN4 (FIG. 7).

5-1A1 is restricted in its ability to infect mosquitoes. 5-1A1 was evaluated for its ability to infect *Aedes aegypti* mosquitoes through an artificial bloodmeal (Table 26). In this assay the ability to infect the midgut of the mosquito and the ability for a midgut infection to disseminate to the head are measured separately. The oral infectious dose 50 (OID<sub>50</sub>) of wild type DEN4 for the midgut was 3.3 log<sub>10</sub>PFU; the OID<sub>50</sub> of wild type DEN4 for a disseminated infection was 3.9 log<sub>10</sub>PFU. In contrast, 5-1A1 never infected 50% of mosquitoes at the doses used. In order to calculate the OID<sub>50</sub> for midgut infections by 5-1A1, it was assumed that at a 10-fold higher dose, 100% of 25 mosquitoes would have become infected. Using this assumption, the conservative estimate of the OID<sub>50</sub> for midgut infections by 5-1A1 was ≥3.9 log<sub>10</sub> PFU. Because 5-1A1 produced only 3 disseminated infections, we did not attempt to calculate an OID<sub>50</sub> for this category. 5-1A1 was significantly restricted in its ability to infect the midgut relative to wild type DEN4 (logistic regression, N=150, P<0.001). Additionally, 5-1A1 produced very few disseminated infections, but because of low numbers this result was not amenable to statistical analysis.

5-1A1 was also significantly restricted in its ability to infect *T. splendens* mosquitoes following intrathoracic inoculation (Table 27). The MID<sub>50</sub> of wild type DEN4 was 2.3 log<sub>10</sub> PFU whereas the MID<sub>50</sub> of 5-1A1 was estimated to be >3.0 log<sub>10</sub> PFU (logistic regression, N=36, P<0.01).

5-1A1 does not show a ts or an att phenotype. 5-1A1 was tested for temperature sensitivity in Vero and HuH-7 cells and for attenuation in suckling mouse brains as described in Example 1. The mutant virus was not temperature sensitive, as defined in Example 1, and was not attenuated in suckling mouse brain (Table 25).

Identification and confirmation of the mutation responsible for the phenotype of 5-1A1. The nucleotide sequence of the entire genome of 5-1A1 was determined as described in Example 1. Sequencing of 5-1A1 revealed three changes from the wild type sequence: two translationally-silent point mutations at positions 7359 and 9047, and one coding point mutation (C to U) at position 7129 in the NS4B gene which resulted in a proline to leucine substitution.

To formally confirm the effect of the C7129U mutation, the mutation was inserted into the cDNA p4, which has been described in Examples 1, 2, and 3 and in FIG. 4, using Kunkel mutagenesis as described in Examples 1 and 3. The mutagenized cDNA was transcribed and transfected as described in Example 3, and the resulting virus, after two terminal dilutions, was designated rDEN4-7129-1A. Like 5-1A1, rDEN4-7129-1A showed normal plaque size and titer in Vero cells and reduced plaque size and normal titer in C6/36 cells (Table 25). rDEN4-7129-1A was not ts on either Vero or HuH-7 cells and was not att in suckling mouse brain. Additionally, rDEN4-7129-1A did not show the SCID-

HuH-7 att phenotype described in Example 3 (Table 25). The ability of rDEN4-7129-1A to infect mosquitoes is envisioned as being tested in both *Ae. aegypti* and *Tx. splendens*.

To test the compatibility of the C7129U mutation and the  $\Delta$ 30 deletion, the C7129U mutation was inserted into rDEN4 $\Delta$ 30 using previously described techniques. The resulting virus, designated rDEN4 $\Delta$ 30-7129, is envisioned as being tested for the phenotypes listed in Table 25.

In summary, three mutant viruses, 5-1A1, rDEN4-356, 357 and rDEN4-7546, showed a particular combination of phenotypes characterized by normal plaque size and replication to high titers in Vero cells and small plaque size but unrestricted growth in mosquito cells. 5-1A1 was further characterized and lacked temperature sensitivity in either Vero or HuH-7 cells and showed normal levels of replication in mouse brain and in SCID-HuH-7 mice and restricted infectivity for both *Ae. aegypti* and *Tx. splendens* mosquitoes. In comparison to wild type rDEN4, the 5-1A1 mutant had one coding mutation: a point mutation (C to U) at nucleotide 7129 in NS4B resulting in a replacement of Pro with Leu. Because 5-1A1 contains only a single missense mutation, the phenotype of this mutant virus can be attributed to the effect of the mutation at position 7129. These results indicate that the 7129 mutation is responsible for the phenotype of decreased infectivity for mosquitoes and is predicted to be useful to restrict replication of vaccine candidates in mosquitoes. To formally confirm this, we have inserted the 7129 mutation into a recombinant DEN4 virus. The resulting virus, designated rDEN4-7129-1A, shows an absence of ts and att phenotypes similar to 5-1A1. It is envisioned as being tested for mosquito infectivity.

The 7129 mutation is a valuable point mutation to include in a DEN4 vaccine candidate and into each of the dengue virus antigenic chimeric vaccine candidates since its biological activity is host specific, i.e., it is restricted in replication in mosquitoes but not in mammals. Moreover, as discussed in Example 6, the 7129 mutation has also been shown to enhance replication in Vero cells. Thus, its insertion into a vaccine candidate is envisioned as enhancing vaccine production in tissue culture without affecting the biological properties specified by other attenuating mutations. It is also envisioned as providing a useful safeguard against mosquito transmission of a dengue virus vaccine.

#### SECTION II. Design of Mutations to Restrict Replication in Mosquitoes

In Section 1 of Example 5, we screened a large panel of mutant viruses carrying both random mutations (generated with 5-fluorouracil) and specific mutations (generated through charge-cluster-to-alanine mutagenesis) for restricted growth in C6/36 cells, a proxy measure for restriction in mosquitoes. However, in neither case were mutations designed for the specific purpose of restricting replication in mosquitoes. In this section, we identified nucleotide sequences in the 3' UTR that show conserved differences between the mosquito-transmitted and tick-transmitted flaviviruses. We then altered those sequences in the DEN4 cDNA p4 by either deleting them altogether or exchanging them with the homologous sequence of the tick-transmitted Langat virus. The resulting viruses were assayed for reduced plaque size and titer in both Vero and C6/36 cells and for infectivity for *Ae. aegypti* and *Tx. splendens*.

Identification and modification of particular 3' UTR sequences showing conserved differences between vectors. Several studies (Olsthoom, R. C. & Bol, J. F. 2001 RNA 7:1370-7; Proutski, V. et al. 1997 Nucleic Acids Res 25:1194-202) have identified conserved differences in the nucleotide sequences of the 3' UTR of mosquito-transmitted and tick-

transmitted flaviviruses. Such differences are concentrated in the 3' terminal core region, the approximately 400 3' terminal nucleotides. It has been suggested that these sequences may have a vector-specific function (Proutski, V. et al. 1997 Nucleic Acids Res 25:1194-202). While such a function has not been identified, it may nonetheless be possible to disrupt vector infectivity by deleting or otherwise altering these nucleotides.

To identify target sequences for this type of alteration, we constructed an alignment of the 3' UTR nucleotide sequences of seven mosquito-transmitted flaviviruses and four tick-transmitted flaviviruses (FIG. 8). From this alignment, we identified several sequences that showed conserved differences between the mosquito-transmitted flaviviruses and tick-transmitted flaviviruses. We then designed primers to alter these sequences in the wt DEN4 cDNA p4 (FIG. 4) in one of two ways: 1) deletion of the nucleotides ( $\Delta$ ) or 2) replacement of the nucleotides with the homologous sequence from the tick-transmitted flavivirus Langat (swap). Langat was chosen as the template for swapped nucleotides because it is naturally attenuated (Pletnev, A. G. 2001 Virology 282:288-300), and therefore unlikely to enhance the virulence of rDEN4 virus derived from the modified cDNA. The DEN4 sequences altered and the mutagenesis primers used to do so are listed in Table 28. Nucleotides 10508-10530 correspond to the CS2 region identified in previous studies (Proutski, V. et al. 1997 Nucleic Acids Res 25:1194-202).

Mutagenesis of p4, transcription and transfection were conducted as previously described in Section I of this Example. All five of the engineered viruses were recovered, and all were subjected to two rounds of terminal dilution as previously described.

Evaluation of phenotypes: cell culture. Viruses were titered in Vero and C6/36 cells as previously described, and the results are listed in Table 29. All of the viruses replicated to  $>5.0 \log_{10}$  PFU/ml; one of them (rDEN4 $\Delta$ 10508-10530) replicated to  $>8.0 \log_{10}$  PFU/ml. Only one of the viruses (rDEN4 $\Delta$ 10535-10544) was small plaque in C6/36 cells; this virus showed wild-type plaque size in Vero cells. Interestingly, another virus (rDEN4swap10508-10539) showed wild type plaque size in C6/36 cells but was sp in Vero cells.

Evaluation of phenotypes: mosquito infectivity. To date one of the five viruses has been tested for infectivity via intrathoracic inoculation in *Tx. splendens*, using previously described methods. Virus rDEN4 $\Delta$ 10508-10530 was dramatically restricted in infectivity relative to the wild type (Table 30). So few mosquitoes were infected that it was not possible to calculate an MID<sub>50</sub> for this virus.

One of the five viruses has been tested for infectivity of *Ae. aegypti* fed on an infectious bloodmeal using previously described methods. rDEN4swap10535-10544 (Table 31) caused significantly fewer midgut infections than wild type rDEN4, but the percentage of disseminated infections did not differ between rDEN4swap10535-10544 and wild type rDEN4. All of the viruses are envisioned as being tested for mosquito infectivity using both methods.

Summary. In this example we have outlined two different strategies for preventing mosquito transmission of a dengue vaccine. First, several small substitution mutations, including two point mutations and one paired charge-to-alanine substitution, have been shown to restrict the replication of DEN4 in mosquito C6/36 cells in cell culture, and one of these mutations (C7129U) has been shown to restrict the ability of DEN4 virus to infect mosquitoes. Second, we have created a variety of deletion and substitution mutations in regions of the DEN4 3' UTR that show conserved differences between mosquito-transmitted and tick-transmitted flaviviruses. One of

these viruses is sp in C6/36 cells and at least two of these viruses show some degree of restriction of mosquito infectivity. By design, the nucleotide sequences in which these mutations were made are highly conserved among the four dengue serotypes and among mosquito-transmitted flaviviruses in general, indicating that they are portable to other vaccine candidates for mosquito-borne flaviviruses. All of the mutations discussed in this Example 11e outside the structural genes and so are envisioned as being useful in constructing antigenic-chimeric vaccine candidates.

## EXAMPLE 6

## Adaptation Mutations which Enhance the Replication of DEN4 and DEN4 Chimeric Viruses in Vero Cells.

Vero cells are a highly characterized substrate that should be suitable for the manufacture of live attenuated flavivirus vaccines, such as dengue virus and tick-borne encephalitis virus. In addition, Vero cells can also be used to grow flaviviruses to high titer for the preparation of an inactivated virus vaccine. Optimal sequences for the efficient growth of dengue viruses in Vero cells have not been identified, but it is well known that flaviviruses accumulate mutations during passage in various cell cultures (Dunster, L. M. et al. 1999 *Virology* 261:309-18; Theiler, M. & Smith, H. H. 1937 *J Exp Med* 65:787-800). Inclusion of specific sequences in live attenuated viruses that enhance their replication in Vero cells and increase the number of doses of vaccine produced per unit substrate would greatly facilitate their manufacture. Similarly, inclusion of Vero cell growth-promoting sequences in wild type viruses used for the preparation of an inactivated virus vaccine would also greatly facilitate the manufacture of the vaccine. The present example identifies mutations that occur following passage of DEN4 virus and DEN2/4 chimeric viruses in Vero cells. Data derived from five sources provided information for this analysis making it possible to generate a list of Vero cell growth-promoting sequences.

Presence of identical mutations in multiple 5-FU mutant viruses. First, as described in Examples 1 and 2, the genomes of 42 dengue virus clones isolated from a 5-FU mutagenized stock of virus were completely sequenced. If mutations that enhance replication occurred during the passage of these 42 mutant viruses in Vero cells, then such mutations should reveal themselves by representation in more than one clone. Analysis of the 42 sequences revealed the occurrence of specific missense mutations in coding regions or nucleotide substitutions in UTRs in multiple clones that are not present in the 2A parental virus genome (Tables 11 and 32). These mutations, many of which occur within a 400 nucleotide section of the NS4B coding region, represent Vero cell-adaptation mutations. One mutation, such as the 4995 mutation, present in eight viruses was found to specify both ts and att phenotypes (Examples 1 and 3). In contrast, the 7163 mutation, present in six viruses, does not specify a ts or att phenotype (Table 13) and thus is an example of a specific Vero cell growth-promoting mutation.

Presence of Vero cell adaptation mutations in other DEN4 viruses and DEN2/4 antigenic chimeric viruses. Second, the 2A-13 dengue virus that was used as a parallel passaged wild type control during the 5-FU experiments described in Example I was grown and cloned in Vero cells in the absence of 5-FU in a manner identical to that of the 5-FU treated viruses. Sequence analysis of this 5-FU untreated virus, designated 2A-13-1A1, revealed that the virus genome contained a mutation at nucleotide 7163 (Example 1 and Table 32), identical to the missense mutation previously identified in 6 of the 5-FU mutant viruses (Tables 11 and 32). This indicates

that growth and passage of DEN4 virus in Vero cells is sufficient to acquire this specific mutation, i.e. mutagenesis with 5-FU is not required. Thus, information from two separate sources indicates that the 7163 mutation appeared in separate Vero cell passaged viruses, thereby strengthening the interpretation that this mutation is growth promoting.

Third, following passage of the 2AΔ30 and rDEN4Δ30 in Vero cells, sequence analysis revealed the presence of a mutation at nucleotides 7153 and 7163, respectively. These two mutations were also previously identified among the 5-FU treated viruses (Table 32). Again, identical mutations appeared following independent passage of virus in Vero cells, corroborating the hypothesis that these mutations confer a growth advantage in Vero cells.

Fourth, an antigenic chimeric dengue virus vaccine candidate was generated that expressed the structural proteins C, prM, and E from DEN2 on a DEN4 wild type genetic background or an attenuated Δ30 genetic background. To construct this virus, the C, prM and E region of wild type cDNA plasmid p4 was replaced with a similar region from DEN2 virus strain NGC (FIG. 10). Specifically, nucleotides between restriction sites BglII (nt 88) and XhoI (nt 2345) of p4 were replaced with those derived from dengue type 2 virus. RNA transcripts synthesized from the resulting p4-D2 plasmid were transfected into Vero cells and rDEN2/4 virus was recovered. A further attenuated version of this chimeric virus containing the Δ30 mutation, rDEN2/4Δ30, was recovered in C6/36 mosquito cells following transfection of cells with RNA transcripts derived p4Δ30-D2. However, rDEN2/4Δ30 could not be recovered directly in Vero cells. The rDEN2/4Δ30 mutant virus recovered in C6/36 cells replicated to very low levels in Vero cells (<1.0 log<sub>10</sub>PFU/ml) but grew to high titer in C6/36 cells (>6.0 log<sub>10</sub>PFU/ml). Genomic sequence of the C6/36-derived virus matched the predicted cDNA sequence and is shown in Appendix 3. Nevertheless, when C6/36-derived rDEN2/4Δ30 was serially passaged 3 to 4 times in Vero cells, a virus population adapted for growth in Vero cells emerged. Virus from this Vero cell-adapted preparation was cloned and amplified in Vero cells to a titer >6.0 log<sub>10</sub>PFU/ml. The genomic sequence was determined for 2 independent virus clones and compared to the predicted cDNA sequence (Table 33 and 34). Each cloned virus contains a mutation in a non-structural gene which coincides closely in location or sequence with a mutation previously identified among the panel of 5-FU mutagenized viruses. The other mutations in these two clones also might confer a growth advantage in Vero cells. Importantly, the mutations identified in Tables 33 and 34 are absolutely required for replication in Vero cells, and it would not be possible to produce the rDEN2/4Δ30 vaccine candidate in Vero cells without the growth-promoting mutations identified in Tables 33 and 34.

Fifth, sequence analysis of the dengue 4 wild-type virus strain 814669 (GenBank accession no. AF326573) following passage in Vero cells identified a mutation in the NS5 region at nucleotide 7630 which had previously been identified among the panel of 5-FU mutagenized viruses (Table 32). This mutation at nucleotide 7630 was introduced into recombinant virus rDEN4 by site-directed mutagenesis as described in Table 16. The resulting virus, rDEN4-7630, was not temperature sensitive when tested at 39° C., indicating that mutation 7630 does not contribute to temperature sensitivity.

Characterization of rDEN2/4Δ30 chimeric viruses containing single and multiple Vero cell adaptation mutations. The generation of chimeric virus rDEN2/4Δ30 provided a unique opportunity for evaluating the capacity of individual mutations to promote increased growth in Vero cells. Because

rDEN2/4Δ30 replicates to very low titer in Vero cells, yet can be efficiently generated in C6/36 mosquito cells, recombinant virus bearing putative Vero-cell adapting mutations were first generated in C6/36 cells and then virus titers were determined in both C6/36 and Vero cells. As shown in Table 35, addition of a single mutation to rDEN2/4Δ30 resulted in a greater than 1000-fold increase in titer in Vero cells, confirming the Vero cell adaptation phenotype conferred by these mutations. However, the combination of two separate mutations into a single virus did not increase the titer in Vero cells beyond the level observed for viruses bearing a single adaptation mutation. Inclusion of either the 7182 or 7630 mutation in the cDNA of rDEN2/4Δ30 allowed the virus to be recovered directly in Vero cells, circumventing the need to recover the virus in C6/36 cells.

Characterization of the growth properties of rDEN4 viruses containing single and multiple defined Vero cell adaptation mutations. To confirm the ability of Vero cell adaptation mutations to enhance growth of DEN4 viruses, site-directed mutagenesis was used to generate rDEN4 viruses encoding selected individual mutations as described in Examples 1 and 3. Five mutations in NS4B (7153, 7162, 7163, 7182, and 7546) from the list of repeated mutations in the 5-FU mutant viruses (Table 32) were introduced singly into the p4 cDNA clone. In addition, the mosquito-restricted, rDEN4-7129 virus was evaluated for enhanced growth in Vero cells since the location of this mutation is in the same region of NS4B. Each virus, including wild-type rDEN4, was recovered, terminally diluted, and propagated in C6/36 cells to prevent introduction of additional Vero cell adaptation mutations, however, because of its restricted growth in C6/36 cells, rDEN4-7129 was propagated only in Vero cells.

Plaque size was evaluated for each mutant rDEN4 virus in Vero cells and C6/36 cells and compared to wild-type rDEN4. Six-well plates of each cell were inoculated with dilutions of virus and plaques were visualized five days later. Representative plaques are illustrated in FIG. 10 and demonstrate that the presence of a Vero cell adaptation mutation does indeed confer increased virus cell to cell spread and growth specifically in Vero cells. In C6/36 cells, average plaque size was approximately 0.50 mm for both wild-type rDEN4 and each mutant virus (except for rDEN4-7546 and rDEN4-7129 which were smaller than wild-type; see Example 5). However, rDEN4 viruses expressing mutation 7162, 7163, 7182, and 7129 had a greater than two-fold increase in plaque size in Vero cells compared to wild-type rDEN4 virus. A smaller but consistent increase in plaque size was observed for rDEN4-7153 and rDEN4-7546.

Growth kinetics and virus yield in Vero cells was assessed for the same panel of rDEN4 viruses. Vero cells were infected at an MOI of 0.01 and samples were removed daily for 10 days, titered on Vero cells, and plaques were visualized. The results in FIG. 11 indicate that the presence of a Vero cell adaptation mutation increased the kinetics of virus growth, but had only a marginal effect on the peak virus yield. At day four post-infection, wild-type rDEN4 grew to 5.2 log<sub>10</sub>PFU/ml while the level of replication in rDEN4-7129-infected cells was 100-fold higher. The rest of the mutant rDEN4 viruses had an increased yield at day four ranging from 0.9 (rDEN4-7153) to 1.6 (rDEN4-7162 and -7163) log<sub>10</sub>PFU/ml. Interestingly, enhanced kinetics of virus growth correlated with increased plaque size in Vero cells. The peak virus yield was reached by day 6 post-infection for rDEN4-7129, -7162, -7163, and -7182 while wild-type rDEN4 did not reach peak titer until day 10. However, the peak virus yield was only slightly higher in rDEN4 viruses expressing Vero cell adaptation mutations.

In an effort to further enhance rDEN4 replication, especially the peak virus yield, combinations of selected Vero cell adaptation mutations were introduced into the rDEN4 background. Three viruses with dual mutations were generated: rDEN4-7153-7163, rDEN4-7153-7182, and rDEN4-7546-7630 and tested in a Vero cell time course infection as described above along with rDEN4 and rDEN4-7162 as a positive control (FIG. 12). The viruses expressing combined mutations grew in a nearly identical manner to rDEN4-7162 indicating that these selected combinations did not enhance the kinetics or peak virus yield. Additional combinations of these and other Vero cell adaptation mutations are envisioned as increasing peak virus yield.

Discussion. Some of the growth promoting mutations listed in Table 32 are also found in homologous regions of DEN1, DEN2, and DEN3 and are envisioned as serving to promote the replication of these viruses in Vero cells. Specifically, the growth promoting mutations indicated in Table 32 that are present in a DEN4 virus are envisioned as being useful for importation into homologous regions of other flaviviruses, such as DEN1, DEN2 and DEN3. Examples of such conserved regions are shown in Appendix 4 and are listed in Table 36. The nucleotides for both mutation 7129 and 7182 are conserved in all four dengue virus serotypes. It is also interesting to note that mutation 7129 not only increases growth in Vero cells (FIG. 10), but it also forms small plaques in mosquito cells (FIG. 6, Table 25). Lee et al. previously passaged DEN3 virus in Vero cells and performed limited sequence analysis of only the structural gene regions of the resulting viruses (Lee, E. et al. 1997 Virology 232:281-90). From this analysis a menu of Vero adaptation mutations was assembled. Although none of these mutations correspond to the Vero adaptation mutations identified in this Example, a single mutation at amino acid position 202 in DEN3 corresponds to mutation 1542 identified in 5-FU mutant virus #1012. The current Example emphasizes the importance in this type of study of determining the sequence of the entire viral genome.

Vero cell growth optimized viruses are envisioned as having usefulness in the following areas. First, the yield of a live attenuated vaccine virus in Vero cells is predicted to be augmented. The live attenuated vaccine candidate is conveniently a DEN4 or other dengue virus or a DEN1/4, DEN2/4, or DEN3/4 antigenic chimeric virus, or a chimeric virus of another flavivirus based on the DEN4 background. The increased yield of vaccine virus is envisioned as decreasing the cost of vaccine manufacture. Second, Vero cell adaptation mutations that are attenuating mutations, such as the 4995 mutation, are envisioned as being stable during the multiple passage and amplification of virus in Vero cell cultures that is required for production of a large number of vaccine doses. Third, Vero cell adaptation mutations are actually required for the growth of the rDEN2/4Δ30 vaccine candidate in Vero cells. Fourth, the increase in yield of a DEN wild type or an attenuated virus is envisioned as making it economically feasible to manufacture an inactivated virus vaccine. Fifth, the presence of the Vero cell growth promoting mutations in the DEN4 vector of the rDEN1/4, rDEN2/4, and rDEN3/4 antigenic chimeric viruses or other flavivirus chimeric viruses based on DEN4 is envisioned as permitting the viruses to grow to a high titer and as thereby being useful in the manufacture of a inactivated virus vaccine. Sixth, the insertion of Vero cell growth promoting mutations into cDNAs such as rDEN2/4Δ30 is envisioned as permitting recovery of virus directly in Vero cells, for which there are qualified master cell banks for manufacture, rather than in C6/36 cells for which qualified cell banks are not available. And seventh, insertion

of the 7129 and 7182 mutations into DEN1, DEN2, or DEN3 wt viruses is envisioned as increasing their ability to replicate efficiently and be recovered from cDNA in Vero cells.

## EXAMPLE 7

## Assembly of a List of Attenuating Mutations

The data presented in these examples permits the assembly of a list of attenuating mutations that is summarized in Table 37. This list contains individual mutations identified in Tables 13-16, 20, and 21 that are known to independently specify an attenuation phenotype. Mutation 7129 is also included since it is derived from virus 5-1A1 shown to be attenuated in mosquitoes. We envision using various combinations of mutations from this list to generate viruses with sets of desirable properties such as restricted growth in the liver or in the brain as taught in Example 3 (Table 18) and Example 4 (Tables 23 and 24). These mutations are also combinable with other previously described attenuating mutations such as the  $\Delta 30$  mutation, as taught in Example 1 (Table 6) and Example 3 (Table 19) to produce recombinant viruses that are satisfactorily attenuated and immunogenic. Mutations listed in Table 37 are also envisioned as being combined with other previously described attenuating mutations such as other deletion mutations or other point mutations (Blok, J. et al. 1992 *Virology* 187:573-90; Butrapet, S. et al. 2000 *J Virol* 74:3011-9; Men, R. et al. 1996 *J Virol* 70:3930-7; Puri, B. et al. 1997 *J Gen Virol* 78:2287-91).

The possibility of importing an attenuating mutation present in one paramyxovirus into a homologous region of a second paramyxovirus has recently been described (Durbin, A. P. et al. 1999 *Virology* 261:319-30; Skiadopoulos, M. H. et al. 1999 *Virology* 260:125-35). Such an importation confers an att phenotype to the second virus or, alternatively, further attenuates the virus for growth in vivo. Similarly we envision importing an attenuating mutation present in one flavivirus to a homologous region of a second flavivirus which would confer an att phenotype to the second flavivirus or, alternatively, would further attenuate the virus for growth in vivo. Specifically, the attenuating mutations indicated in Table 37 are envisioned as being useful for importation into homologous regions of other flaviviruses. Examples of such homologous regions are indicated in Appendix 4 for the mutations listed in Table 37.

## EXAMPLE 8

## Evaluation of Dengue Virus Vaccine In Humans And Rhesus Monkeys

The present example evaluates the attenuation for humans and rhesus monkeys (as an animal model) of a DEN-4 mutant bearing a 30 nucleotide deletion ( $\Delta 30$ ) that was introduced into its 3' untranslated region by site-directed mutagenesis and that was found previously to be attenuated for rhesus monkeys (Men, R. et al. 1996 *J Virol* 70:3930-7), as representative of the evaluation of any dengue virus vaccine for attenuation in humans and rhesus monkeys (as an animal model).

Viruses and cells. The wild type (wt) DEN-4 virus strain 814669 (Dominica, 1981), originally isolated in *Aedes pseudoscutellaris* (AP61) cells, was previously plaque-purified in LLC-MK2 cells and amplified in C6/36 cells as described (Mackow, E. et al. 1987 *Virology* 159:217-28). For further amplification, the C6/36 suspension was passaged 2 times in Vero (WHO) cells maintained in MEM-E (Life Technologies,

Grand Island, N.Y.) supplemented with 10% FBS. Viruses derived from RNA transfection or used for clinical lot development were grown in Vero (WHO) cells maintained in serum-free media, VP-SFM (Life Technologies).

5 Construction of DEN-4 deletion mutants. A 30 nucleotide (nt) deletion was previously introduced into the 3' untranslated region of the 2A cDNA clone of wt DEN-4 strain 814669 as described (Men, R. et al. 1996 *J Virol* 70:3930-7). This deletion removes nucleotides 10478-10507, and was originally designated 3'd 172-143, signifying the location of the deletion relative to the 3' end of the viral genome. In the current example, this deletion is referred to as  $\Delta 30$ . The full-length 2A cDNA clone has undergone several subsequent modifications to improve its ability to be genetically manipulated. As previously described, a translationally-silent XhoI restriction enzyme site was engineered near the end of the E region at nucleotide 2348 to create clone 2A-XhoI (Bray, M. & Lai, C. J. 1991 *PNAS USA* 88:10342-6). In this example, the viral coding sequence of the 2A-XhoI cDNA clone was further modified using site-directed mutagenesis to create clone p4: a unique BbvCI restriction site was introduced near the C-prM junction (nucleotides 447-452); an extra XbaI restriction site was ablated by mutation of nucleotide 7730; and a unique SacII restriction site was created in the NS5 region (nucleotides 9318-9320). Each of these engineered mutations is translationally silent and does not change the amino acid sequence of the viral polypeptide. Also, several mutations were made in the vector region of clone p4 to introduce or ablate additional restriction sites. The cDNA clone p4 $\Delta 30$  was generated by introducing the  $\Delta 30$  mutation into clone p4. This was accomplished by replacing the MluI-KpnI fragment of p4 (nucleotides 10403-10654) with that derived from plasmid 2A $\Delta 30$  containing the 30 nucleotide deletion. The cDNA clones p4 and p4 $\Delta 30$  were subsequently used to generate recombinant viruses rDEN4 and rDEN4 $\Delta 30$ , respectively.

Generation of viruses. Full-length RNA transcripts were synthesized from cDNA clones 2A and 2A $\Delta 30$  using SP6 RNA polymerase as previously described (Lai, C. J. et al. 1991 *PNAS USA* 88:5139-43; Men, R. et al. 1996 *J Virol* 70:3930-7). The reaction to generate full-length RNA transcripts from cDNA clones p4 and p4 $\Delta 30$  was modified and consisted of a 50  $\mu$ l reaction mixture containing 1  $\mu$ g linearized plasmid, 60 U SP6 polymerase (New England Biolabs (NEB), Beverly, Mass.), 1 $\times$ RNA polymerase buffer (40 mM Tris-HCl, pH 7.9, 6 mM MgCl<sub>2</sub>, 2 mM spermidine, 10 mM dithiothreitol), 0.5 mM m<sup>7</sup>G(5')ppp(5')G cap analog (NEB), 1 mM each nucleotide triphosphate, 1 U pyrophosphatase (NEB), and 80 U RNase inhibitor (Roche, Indianapolis, IN). This reaction mixture was incubated at 40° C. for 90 min and the resulting transcripts were purified using RNeasy mini kit (Qiagen, Valencia, Calif.). For transfection of Vero cells, purified transcripts (1  $\mu$ g) were mixed with 12  $\mu$ l DOTAP liposome reagent (Roche) in saline containing 20 mM HEPES buffer (pH 7.6) and added to cell monolayer cultures in a 6-well plate. After 5-17 days, tissue culture medium was harvested, clarified by centrifugation, and virus was amplified in Vero cells. The presence of virus was confirmed by plaque titration. It should be noted that during the course of transfection and amplification of 2A $\Delta 30$  to create the vaccine lot, the virus underwent a total of 6 passages entirely in Vero cells. The remaining viruses, rDEN4 and rDEN4 $\Delta 30$  were passaged 5 times in Vero cells to generate the virus suspension used for sequence analysis and studies in rhesus monkeys.

65 Vaccine Production. An aliquot of clarified tissue culture fluid containing vaccine candidate 2A $\Delta 30$  was submitted to DynCorp (Rockville, Md.) for amplification of virus in Vero

cells and production of a vaccine lot. For vaccine production, 2AΔ30 infected tissue culture supernatant was harvested, SPG buffer added (final concentration: 218 mM sucrose, 6 mM L-glutamic acid, 3.8 mM potassium phosphate, monobasic, and 7.2 mM potassium phosphate, dibasic, pH 7.2), and the virus suspension was clarified by low speed centrifugation. To degrade residual Vero cell DNA, the vaccine suspension was treated with Benzonase endonuclease (American International Chemical, Natick, Mass.), 100 U/ml and incubated for 1 hr at 37° C., followed by high-speed centrifugation (17,000× g, 16 hr). The resulting virus pellet was gently rinsed with MEM-E, resuspended in MEM-E containing SPG, sonicated, distributed into heat-sealed ampoules, and stored frozen at -70° C. Final container safety testing confirmed microbial sterility, tissue culture purity, and animal safety. The 2AΔ30 vaccine lot (designated DEN4-9) has a titer of 7.48 log<sub>10</sub>PFU/ml, with a single dose of 5.0 log<sub>10</sub>PFU/ml containing <1 pg/ml Vero cell DNA and <0.001 U/ml Benzonase endonuclease.

Sequence of cDNA clones and viral genomes. The nucleotide sequence of the viral genome region of cDNA plasmids 2A and p4 was determined on a 310 genetic analyzer (Applied Biosystems, Foster City, Calif.) using vector-specific and DEN-4-specific primers in BigDye terminator cycle sequencing reactions (Applied Biosystems). The nucleotide sequence of the genomes of the parental wt DEN-4 strain 814669 and of recombinant viruses 2A wt, 2AΔ30 (vaccine lot), rDEN4, and rDEN4Δ30 was also determined. Viral RNA was extracted from virus preparations and serum samples using the QIAamp Viral RNA mini kit (Qiagen). Reverse transcription (RT) was performed using random hexamers and the SuperScript First-Strand Synthesis System for RT-PCR (Life Technologies). Overlapping PCR fragments of approximately 2000 base pairs were generated using optimized DEN-4 specific primers and Advantage cDNA polymerase (ClonTech, Palo Alto, Calif.). Both strands of purified PCR fragments were sequenced directly using dye-terminator reactions as described above and results were assembled into a consensus sequence. To determine the nucleotide sequence of the viral RNA 5' and 3' regions, the 5' cap nucleoside of the viral RNA was removed with tobacco acid pyrophosphatase (Epicentre, Madison, Wis.) followed by circularization of the RNA using RNA ligase (Epicentre). RT-PCR was performed as described and a cDNA fragment spanning the ligation junction was sequenced using DEN-4 specific primers. GenBank accession numbers have been assigned as follows (virus: accession number): 814669: AF326573, 2AΔ30: AF326826, rDEN4: AF326825, and rDEN4Δ30: AF326827.

Human Vaccine Recipients. 20 normal healthy adult volunteers were recruited by the Johns Hopkins School of Hygiene and Public Health Center for Immunization Research (CIR) located in Baltimore, Md. The clinical protocol was reviewed and approved by the Joint Committee for Clinical Investigation of the Johns Hopkins University School of Medicine and informed consent was obtained from each volunteer. Volunteers were enrolled in the study if they met the following inclusion criteria: 18-45 years of age; no history of chronic illness; a normal physical examination; human immunodeficiency virus antibody negative, hepatitis B surface antigen negative, and hepatitis C antibody negative; no stool occult blood; and normal values for complete blood cell count (CBC) with differential, hematocrit, platelet count, serum creatinine, serum aspartate amino transferase (AST), alanine amino transferase (ALT), alkaline phosphatase, bilirubin, prothrombin time (PT), partial thromboplastin time (PTT), and urinalysis. Female volunteers were required to have a negative urine pregnancy test prior to vaccination and

on the day of vaccination and to agree to use contraception or abstain from sexual intercourse for the duration of the study. Volunteers also lacked serological evidence of prior flavivirus infection as defined by hemagglutination-inhibition antibody titer <1:10 to DEN-1, DEN-2, DEN-3, DEN-4, St. Louis encephalitis virus, Japanese encephalitis virus, or yellow fever virus and a plaque-reduction neutralization titer <1:10 to DEN-4 and yellow fever virus.

Studies in Humans. Volunteers were immunized in three successive cohorts of four, six, and ten volunteers to assess the safety of the vaccine. In this study, an illness was defined as the following: dengue virus infection associated with a platelet count of <90,000/mm<sup>3</sup>; serum ALT >4 times normal; oral temperature >38° C. for >2 successive days; or headache and/or myalgia lasting >2 successive days. Systemic illness was defined as the occurrence of fever >38° C. for >2 consecutive days, or any 2 of the following for at least two consecutive study days: headache, malaise, anorexia, and myalgia/arthralgia. The trials were conducted between October and April, a time of low mosquito prevalence, to reduce the risk of transmission of vaccine virus from the volunteers to the community.

On the day of vaccination, vaccine candidate 2AΔ30 was diluted to 5.3 log<sub>10</sub>PFU/ml in sterile saline for injection, USP, and each volunteer was injected subcutaneously with a 0.5 ml containing 5.0 log<sub>10</sub>PFU of vaccine into the left deltoid region. Volunteers were given a home diary card on which they were to record their temperature twice daily for days 0-5 post-vaccination. The volunteers returned to the clinic each day for examination by a physician and their diary cards were reviewed. The injection site was evaluated for erythema, induration, and tenderness. Clinical signs and symptoms such as headache, rash, petechiae, lymphadenopathy, hepatomegaly, abdominal tenderness, anorexia, nausea, fatigue, myalgia, arthralgia, eye pain, and photophobia were assessed daily. Symptoms were graded as mild (no need for treatment or a change in activity), moderate (treatment needed or change in activity noted, yet still able to continue daily activity) or severe (confined to bed). Blood was drawn for CBC with differential and for virus quantitation on days 0, 2 and 4. Volunteers were admitted to the inpatient unit at the CIR on the sixth day after immunization. The study physician evaluated all volunteers each day by physical examination and interview. The volunteers had their blood pressure, pulse, and temperature recorded four times a day. Blood was drawn each day for CBC with differential and for virus quantitation and every other day for ALT measurement. Volunteers were confined to the inpatient unit until discharge on study day 15. On study days 28 and 42, volunteers returned for physical examination and blood was drawn for virus quantitation (day 28) and for serum antibody measurement (day 28 and 42).

Virus quantitation and amplification. Serum was obtained for detection of viremia and titration of virus in positive specimens. For these purposes 8.5 ml of blood was collected in a serum separator tube and incubated at room temperature for less than 30 min. Serum was decanted into 0.5 ml aliquots, rapidly frozen in a dry ice/ethanol bath and stored at -70° C. Serum aliquots were thawed and serial 10-fold dilutions were inoculated onto Vero cell monolayer cultures in 24-well plates. After one hour incubation at room temperature, the monolayers were overlaid with 0.8% methylcellulose in OptiMEM (Life Technologies) supplemented with 5% fetal bovine serum (FBS). Following incubation at 37° C. for four days, virus plaques were visualized by immunoperoxidase staining. Briefly, cell monolayers were fixed in 80% methanol for 30 min and rinsed with antibody buffer (5% nonfat milk in phosphate buffered saline). Rabbit polyclonal DEN-4 anti-

bodies were diluted 1:1000 in antibody buffer and added to each well followed by a one hr incubation at 37° C. Primary antibody was removed and the cell monolayers were washed twice with antibody buffer. Peroxidase-labelled goat-anti-rabbit IgG (KPL, Gaithersburg, Md.) was diluted 1:500 in antibody buffer and added to each well followed by a one hr incubation at 37° C. Secondary antibody was removed and the wells were washed twice with phosphate buffered saline. Peroxidase substrate (4 chloro-1 -naphthol in H<sub>2</sub>O<sub>2</sub>) was added to each well and visible plaques were counted.

For amplification of virus in serum samples, a 0.3 ml aliquot of serum was inoculated directly onto a single well of a 6-well plate of Vero cell monolayers and incubated at 37° C. for 7 days. Cell culture fluid was then assayed for virus by plaque assay as described above.

Serology. Hemagglutination-inhibition (HAI) assays were performed as previously described (Clarke, D. H. & Casals, J. 1958 *Am J Trop Med Hyg* 7:561-73). Plaque-reduction neutralization titers (PRNT) were determined by a modification of the technique described by Russell (Russell, P. K. et al. 1967 *J Immunol* 99:285-90). Briefly, test sera were heat inactivated (56° C. for 30 min) and serial 2-fold dilutions beginning at 1:10 were made in OptiMEM supplemented with 0.25% human serum albumin. rDEN4Δ30 virus, diluted to a final concentration of 1000 PFU/ml in the same diluent, was added to equal volumes of the diluted serum and mixed well. The virus/serum mixture was incubated at 37° C. for 30 min. Cell culture medium was removed from 90% confluent monolayer cultures of Vero cells on 24-well plates and 50 μl of virus/serum mixture was transferred onto duplicate cell monolayers. Cell monolayers were incubated for 60 min at 37° C. and overlaid with 0.8% methylcellulose in OptiMEM supplemented with 2% FBS. Samples were incubated at 37° C. for 4 days after which plaques were visualized by immunoperoxidase staining as described above, and a 60% plaque-reduction neutralization titer was calculated.

Studies in rhesus monkeys. Evaluation of the replication and immunogenicity of wt virus 814669, and recombinant viruses 2A wt, 2AΔ30 (vaccine lot), rDEN4, and rDEN4Δ30 in juvenile rhesus monkeys was performed as previously described (Men R. et al. 1996 *J Virol* 70:3930-7). Briefly, dengue virus seronegative monkeys were injected subcutaneously with 5.0 log<sub>10</sub> PFU of virus diluted in L-15 medium (Quality Biological, Gaithersburg, Md.) containing SPG buffer. A dose of 1 ml was divided between two injections in each side of the upper shoulder area. Monkeys were observed daily and blood was collected on days 0-10 and 28, and processed for serum, which was stored frozen at -70° C. Titer of virus in serum samples was determined by plaque assay on Vero cells as described above. Neutralizing antibody titers were determined for the day 28 serum samples as described above. A group of monkeys inoculated with either 2AΔ30 (n=4) or wt virus 814669 (n=8) were challenged on day 42 with a single dose of 5.0 log<sub>10</sub> PFU/ml wt virus 814669 and blood was collected for 10 days. Husbandry and care of rhesus monkeys was in accordance with the National Institutes of Health guidelines for the humane use of laboratory animals.

Construction and characterization of DEN-4 wild type and deletion mutant viruses. The nucleotide and deduced amino acid sequences of the previously described wt 814669 virus, the DEN-4 2A wt virus derived from it (designated 2A wt), and the 2AΔ30 vaccine candidate derived from 2A wt virus were first determined. Sequence analysis showed that the wt 814669 virus used in this study had apparently accumulated 2 missense mutations (nucleotides 5826 and 7630) and 3 silent mutations during its passage and amplification since these

mutations were not described in previously published reports of the viral sequence (GenBank accession number M14931) and were not present in the 2A cDNA derived from the virus. Sequence comparison between viruses 2A wt and vaccine lot 2AΔ30 revealed that 2AΔ30 accumulated 2 missense mutations (nucleotides 7153 and 8308) and also confirmed the presence of the Δ30 mutation (nucleotides 10478-10507) as well as an additional deletion of nucleotide 10475, which occurred during the original construction of the Δ30 mutation (Men, R. et al. 1996 *J Virol* 70:3930-7). This sequence analysis revealed significant sequence divergence between the biologically-derived wt 814669 virus and its recombinant 2A wt derivative and between the 2A wt and 2AΔ30 virus. Since the 2A wt and 2AΔ30 viruses differed at nucleotides other than the deletion mutation, the attenuation phenotype previously reported for 2AΔ30 (Men, R. et al. 1996 *J Virol* 70:3930-7) could not be formally ascribed solely to the 430 mutation and may have been specified by the mutations at nucleotides 7153, 8308, 10475, or the Δ30 deletion.

To determine whether the Δ30 mutation was responsible for the observed attenuation of 2AΔ30, a second pair of viruses, one with and one without the Δ30 mutation, were produced for evaluation in monkeys. A new DEN-4 cDNA vector construct, designated p4, was derived from the 2A-XhoI cDNA clone and translationally-silent mutations were introduced to add or ablate several restriction enzyme sites. These sites were added to facilitate the future genetic manipulation of this DEN-4 wt cDNA by the introduction of other attenuating mutations if needed. The sequence of the genomic region of the p4 cDNA plasmid was identical to that of the 2A wt virus except for the engineered restriction site changes and a point mutation at nucleotide 2440 which was introduced during the original mutagenesis of the 2A cDNA plasmid to create the XhoI site (Bray, M. & Lai, C. J. 1991 *PNAS USA* 88:10342-6). The 430 mutation and the neighboring deletion at nucleotide 10475 were co-introduced into the p4 plasmid by replacing a short restriction fragment with one derived from the cDNA clone of 2AΔ30. RNA transcripts derived from the p4 cDNA clone and from its Δ30 derivative each yielded virus (designated rDEN4 wt and rDEN4Δ30, respectively) following transfection of Vero cells. Sequence analysis of the rDEN4 virus revealed that during its passage and amplification in Vero cells it accumulated 2 missense mutations (nucleotides 4353 and 6195), a silent mutation (nucleotide 10157), and a point mutation in the 3' untranslated region (nucleotide 10452). In addition to containing the Δ30 and the accompanying deletion at nucleotide 10475, rDEN4Δ30 had also accumulated a missense mutation (nucleotide 7163) and a silent mutation (nucleotide 7295).

Parental wt 814669 virus and recombinant viruses 2A wt, 2AΔ30, rDEN4, and rDEN4Δ30 each replicate in Vero cells to a titer exceeding 7.0 log<sub>10</sub> PFU/ml, and their replication is not temperature sensitive at 39° C.

Virus replication, immunogenicity, and efficacy in monkeys. Groups of rhesus monkeys were inoculated with wt DEN-4 814669, 2A wt, rDEN4 2AΔ30 and rDEN4Δ30 to assess the level of restriction of replication specified by the Δ30 mutation. Serum samples were collected daily and titer of virus present in the serum was determined by plaque enumeration on Vero cell monolayer cultures. Monkeys inoculated with wt 814669 virus or its recombinant counterparts, 2A wt or rDEN4, were viremic for 3 to 4 days with a mean peak virus titer of nearly 2 log<sub>10</sub> PFU/ml. Monkeys inoculated with virus 2AΔ30 or rDEN4Δ30 had a lower frequency of viremia (83% and 50%, respectively), were viremic for only about 1 day, and the mean peak titer was 10-fold lower. Monkeys inoculated with DEN-4 814669, 2A wt, or rDEN4



viruses developed high levels of neutralizing antibody, with mean titers between 442 and 532, consistent with their presumed wild type phenotype. Monkeys inoculated with 2AΔ30 or rDEN4Δ30 developed a lower level of neutralizing antibody, with mean titers of 198 and 223, respectively. The decrease in neutralizing antibody titer in response to 2AΔ30 and rDEN4Δ30 is consistent with the attenuation phenotype of these viruses. Monkeys inoculated with either 2AΔ30 (n=4) or wt 814669 virus (n=8) were challenged after 42 days with wt virus 814669. Dengue virus was not detected in any serum sample collected for up to 10 days following virus challenge, indicating that these monkeys were completely protected following immunization with either wt virus or vaccine candidate 2AΔ30.

Since DEN-4 814669, 2A wt, and rDEN4 each manifest the same level of replication and immunogenicity in rhesus monkeys, it is reasonable to conclude that the identified sequence differences between these presumptive wild type viruses that arose during passage in tissue culture or during plasmid construction do not significantly affect their level of replication *in vivo*. Similarly, the comparable level of attenuation of 2AΔ30 and rDEN4Δ30 indicates that the mutations shared by these viruses, namely, the Δ30 mutation and its accompanying 10475 deletion mutation, are probably responsible for the attenuation of these viruses rather than their incidental sequence differences.

Clinical Response to immunization with 2AΔ30. The 2AΔ30 vaccine candidate was administered subcutaneously at a dose of 10<sup>5</sup> PFU to 20 seronegative volunteers. Each of the vaccinees was infected and the virus was well tolerated by all vaccinees. Viremia was detected in 70% of the vaccinees, was present only at low titer, and did not extend beyond day 11.

None of the 20 vaccinees reported soreness or swelling at the injection site. Mild erythema (1-3 mm) around the injection site was noted on examination of 8 volunteers 30 minutes post-vaccination which resolved by the next day in 7 of those volunteers and by the third day in the remaining volunteer. Mild tenderness to pressure at the vaccination site was noted in 2 volunteers and lasted a maximum of 48 hours. During physical examination, ten volunteers (50%) were noted to have a very mild dengue-like erythematous macular rash (truncal distribution) which occurred with greatest frequency on day 10. None of the volunteers noted the rash themselves, and it was asymptomatic in each instance. Rash was seen only in vaccinees with detectable viremia. Volunteers did not develop systemic illness. Seven volunteers noted an occasional headache that was described as mild, lasting less than 2 hours, and was not present in any volunteer on two consecutive days. One volunteer reported fever of 38.6° C. and 38.2° C. without accompanying headache, chills, eye pain, photophobia, anorexia, myalgia, or arthralgia as an outpatient the evening of day 3 and day 5, respectively. However, this volunteer was afebrile when evaluated by the study staff on the morning of days 3, 4, 5 and 6. All other temperature measurements recorded by the volunteer or study staff were normal. Although tourniquet tests were not performed, two volunteers were noted to have petechiae at the site of the blood pressure cuff after a blood pressure measurement was performed (one on day 6, the other on days 7 and 10). Both of these volunteers had normal platelet counts at that time and throughout the study.

Significant hematological abnormalities were not seen in any vaccinee. Three vaccinees with presumed benign ethnic neutropenia manifested an absolute neutrophil count (ANC) below 1500/mm<sup>3</sup>. These three volunteers had baseline ANC's which were significantly lower than the remaining 17 volun-

teers and which did not decrease disproportionately to the other volunteers. Two of the three volunteers who became neutropenic never had detectable viremia. A mild increase in ALT levels was noted in 4 volunteers, and a more significant increase in ALT level (up to 238 IU/L) was noted in one volunteer. These ALT elevations were transient, were not associated with hepatomegaly, and were completely asymptomatic in each of the 5 volunteers. Elevated ALT values returned to normal by day 26 post-vaccination. The volunteer with the high ALT value was also noted to have an accompanying mild elevation in AST on day 14 (10<sup>4</sup> IU/L) which also returned to baseline by day 26 post-vaccination. This volunteer did not have an associated increase in LDH, bilirubin, or alkaline phosphatase levels.

Serologic response of humans to immunization with 2AΔ30. Each of the twenty vaccinees developed a significant rise in serum neutralizing antibody titer against DEN-4 by day 28. The level of serum neutralizing antibody was similar in viremic (1:662) and non-viremic vaccinees (1:426). The DEN-4 neutralizing antibody titers of both groups had not changed significantly by day 42.

Genetic stability of the Δ30 mutation. RT-PCR and sequence analysis of viral RNA isolated from serum samples (n=6) collected from volunteers 6 to 10 days post-vaccination confirmed the presence of the Δ30 mutation and neighboring deletion at nucleotide 10475.

## EXAMPLE 9

### Pharmaceutical Compositions

Live attenuated dengue virus vaccines, using replicated virus of the invention, are used for preventing or treating dengue virus infection. Additionally, inactivated dengue virus vaccines are provided by inactivating virus of the invention using known methods, such as, but not limited to, formalin or β-propiolactone treatment. Live attenuated or inactivated viruses containing the mutations described above form the basis of an improved vaccine for the prevention or treatment of dengue infection in humans.

Pharmaceutical compositions of the present invention comprise live attenuated or inactivated dengue viruses, optionally further comprising sterile aqueous or non-aqueous solutions, suspensions, and emulsions. The composition can further comprise auxiliary agents or excipients, as known in the art. See, e.g., Berkow et al. eds. 1987 *The Merck Manual*, 15th edition, Merck and Co., Rahway, N.J.; Goodman et al. eds. 1990 *Goodman and Gilman's The Pharmacological Basis of Therapeutics*, 8th edition, Pergamon Press, Inc., Elmsford, N.Y.; Avery's *Drug Treatment: Principles and Practice of Clinical Pharmacology and Therapeutics*, 3rd edition, ADIS Press, LTD., Williams and Wilkins, Baltimore, Md. 1987; Osol, A. ed. 1980 *Remington's Pharmaceutical Sciences* Mack Publishing Co, Easton, Pa. pp. 1324-1341; Katzung, ed. 1992 *Basic and Clinical Pharmacology* Fifth Edition, Appleton and Lange, Norwalk, Conn.

A virus vaccine composition of the present invention can comprise from about 10<sup>2</sup>-10<sup>9</sup> plaque forming units (PFU)/ml, or any range or value therein, where the virus is attenuated. A vaccine composition comprising an inactivated virus can comprise an amount of virus corresponding to about 0.1 to 50 μg of E protein/ml, or any range or value therein.

The agents may be administered using techniques well known to those in the art. Preferably, agents are formulated and administered systemically. Suitable routes may include oral, rectal, transmucosal, or intestinal administration; parenteral delivery, including intramuscular, subcutaneous,

intramedullary injections, as well as intrathecal, direct intraventricular, intravenous, intraperitoneal, intradermal, intranasal, or intraocular injections, just to name a few. For injection, the agents of the invention may be formulated in aqueous solutions, preferably in physiologically compatible buffers such as saline, phosphate buffered saline, Tris buffered saline, Hank's buffered saline, growth media such as Eagle's Minimum Essential Medium (MEM), and the like.

When a vaccine composition of the present invention is used for administration to an individual, it can further comprise salts, buffers, adjuvants, or other substances which are desirable for improving the efficacy of the composition. Adjuvants useful with the invention include, but are not limited to: (1) aluminum salts (alum), such as aluminum hydroxide, aluminum phosphate, aluminum sulfate, etc.; (2) oil-in-water emulsion formulations (with or without other specific immunostimulating agents such as muramyl peptides or bacterial cell wall components), such as for example (a) MF59 (International Publication No. WO 90/14837), containing 5% Squalene, 0.5% Tween 80, and 0.5% Span 85 (optionally containing various amounts of MTP-PE, although not required) formulated into submicron particles using a microfluidizer such as Model 110Y microfluidizer (Microfluidics, Newton, Mass.), (b) SAF, containing 10% Squalene, 0.4% Tween 80, 5% pluronic-blocked polymer L121, and thr-MDP either microfluidized into a submicron emulsion or vortexed to generate a larger particle size emulsion, and (c) Ribi™ adjuvant system (RAS), (Ribi Immunochem, Hamilton, Mont.) containing 2% Squalene, 0.2% Tween 80, and one or more bacterial cell wall components from the group consisting of monophosphorylipid A (MPL), trehalose dimycolate (TDM), and cell wall skeleton (CWS), preferably MPL+CWS (Detox™); (3) saponin adjuvants, such as Stimulon™ (Cambridge Bioscience, Worcester, Mass.) may be used or particle generated therefrom such as ISCOMs (immunostimulating complexes); (4) Complete Freund's Adjuvant (CFA) and Incomplete Freund's Adjuvant (IFA); (5) cytokines, such as interleukins (IL-1, IL-2, etc.), macrophage colony stimulating factor (M-CSF), tumor necrosis factor (TNF), etc.; (6) mucosal adjuvants such as those derived from cholera toxin (CT), pertussis toxin (PT), E. coli heat labile toxin (LT), and mutants thereof (see, e.g., International Publication Nos. WO 95/17211, WO 93/13202, and WO 97/02348); and (7) other substances that act as immunostimulating agents to enhance the effectiveness of the composition.

The pharmacologically active compounds of this invention can be processed in accordance with conventional methods of galenic pharmacy to produce medicinal agents for administration to patients, e.g., mammals including humans.

The compounds of this invention can be employed in admixture with conventional excipients, i.e., pharmaceutically acceptable organic or inorganic carrier substances suitable for parenteral, enteral (e.g., oral) or topical application, which do not deleteriously react with the active compounds. Suitable pharmaceutically acceptable carriers include but are not limited to water, salt solutions, alcohols, gum arabic, vegetable oils, benzyl alcohols, polyethylene glycols, gelatin, carbohydrates such as lactose, amylose or starch, magnesium stearate, talc, silicic acid, viscous paraffin, perfume oil, fatty acid monoglycerides and diglycerides, pentaerythritol fatty acid esters, hydroxy methylcellulose, polyvinyl pyrrolidone, etc. The pharmaceutical preparations can be sterilized and if desired mixed with auxiliary agents, e.g., lubricants, preservatives, stabilizers, wetting agents, emulsifiers, salts for influencing osmotic pressure, buffers, coloring, flavoring and/or aromatic substances and the like which do not deleteriously

react with the active compounds. They can also be combined where desired with other active agents, e.g., vitamins.

For parenteral application, particularly suitable are injectable, sterile solutions, preferably oily or aqueous solutions, as well as suspensions, emulsions, or implants, including suppositories. Ampoules are convenient unit dosages.

For enteral application, particularly suitable are tablets, dragees, liquids, drops, suppositories, or capsules. A syrup, elixir, or the like can be used wherein a sweetened vehicle is employed.

For topical application, there are employed as non-sprayable forms, viscous to semi-solid or solid forms comprising a carrier compatible with topical application and having a dynamic viscosity preferably greater than water. Suitable formulations include but are not limited to solutions, suspensions, emulsions, creams, ointments, powders, liniments, salves, aerosols, etc., which are, if desired, sterilized or mixed with auxiliary agents, e.g., preservatives, stabilizers, wetting agents, buffers or salts for influencing osmotic pressure, etc.

For topical application, also suitable are sprayable aerosol preparations wherein the active ingredient, preferably in combination with a solid or liquid inert carrier material, is packaged in a squeeze bottle or in admixture with a pressurized volatile, normally gaseous propellant, e.g., a freon.

The vaccine can also contain variable but small quantities of endotoxin, free formaldehyde, and preservative, which have been found safe and not contributing to the reactogenicity of the vaccines for humans.

#### EXAMPLE 10

##### Pharmaceutical Purposes

The administration of the vaccine composition may be for either a "prophylactic" or "therapeutic" purpose. When provided prophylactically, the compositions are provided before any symptom of dengue viral infection becomes manifest. The prophylactic administration of the composition serves to prevent or attenuate any subsequent infection. When provided therapeutically, the live attenuated or inactivated viral vaccine is provided upon the detection of a symptom of actual infection. The therapeutic administration of the compound(s) serves to attenuate any actual infection. See, e.g., Berkow et al. eds. 0.1987 The Merck Manual, 15th edition, Merck and Co., Rahway, N.J.; Goodman et al. eds. 1990 Goodman and Gilman's The Pharmacological Basis of Therapeutics, 8th edition, Pergamon Press, Inc., Elmsford, N.Y.; Avery's Drug Treatment: Principles and Practice of Clinical Pharmacology and Therapeutics, 3rd edition, ADIS Press, LTD., Williams and Wilkins, Baltimore, Md. 1987; Katzung, ed. 1992 Basic and Clinical Pharmacology, Fifth Edition, Appleton and Lange, Norwalk, Conn.

A live attenuated or inactivated vaccine composition of the present invention may thus be provided either before the onset of infection (so as to prevent or attenuate an anticipated infection) or after the initiation of an actual infection.

The vaccines of the invention can be formulated according to known methods to prepare pharmaceutically useful compositions, whereby live attenuated or inactivated viruses are combined in a mixture with a pharmaceutically acceptable vehicle. A composition is said to be a "pharmacologically acceptable vehicle" if its administration can be tolerated by a recipient patient. Suitable vehicles are well known to those in the art, e.g., in Osol, A. ed. 1980 Remington's Pharmaceutical Sciences Mack Publishing Co, Easton, Pa. pp. 1324-1341.

For purposes of administration, a vaccine composition of the present invention is administered to a human recipient in





TABLE 3-continued

Nucleotide and amino acid differences of the 5-FU mutant viruses which are ts in both Vero and HuH-7 cells.							
Virus	Mutations in UTR or coding region that result in an amino acid substitution				Mutations in coding region that do not result in an amino acid substitution		
	Nucleotide position	Gene/region	Nucleotide change	Amino Acid change <sup>b</sup>	Nucleotide position	Gene	Nucleotide change
938 <sup>a</sup>	3442	NS1	A > G	E1114G	747	prM	U > C
	4995	NS3	U > C	S1632P	4196	NS2b	U > C
	10275	3' UTR	A > U	n/a	6155	NS3	G > A
1033 <sup>a</sup>	4907	NS3	A > U	L1602F	548	prM	C > U
	8730	NS5	A > C	N2877H			
	9977	NS5	G > A	M3292I			

<sup>a</sup>Viruses that contain mutation(s) resulting in an a.a. substitution in only a NS gene(s) and/or nucleotide substitutions in the UTRs are indicated; i.e. no a.a. substitutions are present in the structural proteins (C-prM-E).

<sup>b</sup>Amino acid position in DEN4 polyprotein beginning with the methionine residue of the C protein (nt 102-104) as residue #1. Wild-type amino acid on left of amino acid position; mutant amino acid on right.

TABLE 4

Nucleotide and amino acid differences of the 5-FU mutant viruses which are ts in only HuH-7 cells.							
Virus	Mutations in UTR or coding region that result in an amino acid substitution				Mutations in coding region that do not result in an amino acid substitution		
	Nucleotide position	Gene/region	Nucleotide change	Amino acid change <sup>b</sup>	Nucleotide position	Gene	Nucleotide change
571	586	prM	U > C	V162A	6413	NS4A	U > C
	7163	NS4B	A > U	L2354F			
	7947	NS5	G > A	G2616R			
605	1455	E	G > U	V452F	none		
	7546	NS4B	C > U	A2482V			
631	595	prM	A > G	K165R	1175	E	G > A
	6259	NS3	U > C	V2053A	5174	NS3	A > G
	7546	NS4B	C > U	A2482V			
686 <sup>a</sup>	3575	NS2A	G > A	M1158I	4604	NS3	A > G
	4062	NS2A	A > G	T1321A	7937	NS5	A > G
	7163	NS4B	A > U	L2354F			
967	2094	E	G > C	A665P	4616	NS3	C > U
	2416	E	U > C	V772A			
	7162	NS4B	U > C	L2354S			
	7881	NS5	G > A	G2594S			
992 <sup>a</sup>	5695	NS3	A > G	D1865G	3542	NS2A	A > G
	7162	NS4B	U > C	L2354S			
1175 <sup>a</sup>	7153	NS4B	U > C	V2351A	6167	NS3	U > C
	10186	NS5	U > C	I3362T	10184	NS5	G > A
	10275	3' UTR	A > U	n/a			

<sup>a</sup>Viruses that contain mutation(s) resulting in an a.a. substitution in only a NS gene(s) and/or nucleotide substitutions in the UTRs are indicated; i.e. no a.a. substitutions are present in the structural proteins.

<sup>b</sup>Amino acid position in DEN4 polyprotein beginning with the methionine residue of the C protein (nt 102-104) as residue #1. Wild-type amino acid on left of amino acid position; mutant amino acid on right.

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TABLE 5

Mutations which are represented in multiple 5-FU mutant DEN4 viruses.				
Nucleotide position	Gene/region	Nucleotide change	Amino acid change	Number of viruses with "sister" mutations
1455	E	G > U	val > phe	2
4995	NS3	U > C	ser > pro	8
7162	NS4B	U > C	leu > ser	2
7163	NS4B	A > U or C	leu > phe	3
7546	NS4B	C > U	ala > val	3
10275	3' UTR	A > U	n/a <sup>a</sup>	2

65

<sup>a</sup>not applicable









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TABLE 11

Putative Vero cell adaptation mutations derived from the full set of 5-FU mutant viruses.				
5-FU mutant viruses				
Nucleotide position	Gene/region (a.a. #) <sup>b</sup>	Nucleotide change	Amino acid change	No. of viruses with the mutation
1455	E (452)	G > U	Val > Phe	5
2280	E (727)	U > C	Phe > Leu	2
4891	NS3 (1597)	U > C	Ile > Thr	2
4995	NS3 (1599)	U > C	Ser > Pro	8
7153	NS4B (2351)	U > C	Val > Ala	3
7162	NS4B (2354)	U > C	Leu > Ser	4
7163	NS4B (2354)	A > U or C	Leu > Phe	7
7182	NS4B (2361)	G > A	Gly > Ser	2

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TABLE 11-continued

Putative Vero cell adaptation mutations derived from the full set of 5-FU mutant viruses.				
5-FU mutant viruses				
Nucleotide position	Gene/region (a.a. #) <sup>b</sup>	Nucleotide change	Amino acid change	No. of viruses with the mutation
10 7546	NS4B (2482)	C > U	Ala > Val	10
7630	NS5 (2510)	A > G	Lys > Arg	1
10275	3' UTR	A > U	n/a <sup>a</sup>	6
10279	3' UTR	A > C	n/a	4

<sup>a</sup>not applicable15 <sup>b</sup>Amino acid position in DEN4 polyprotein beginning with the methionine residue of the C protein (nt 102-104) as residue #1.

TABLE 12

Mutagenic oligonucleotides used to generate recombinant DEN4 viruses containing single 5-FU mutations.							
SEQ ID NO.	Recombinant virus (rDEN4-)	Nucleotide change	Amino acid change	Gene	pUC clone	RE site <sup>a</sup>	Oligonucleotide <sup>b</sup>
23	40	U > C	n/a	5' UTR	pUC-NheI	BsaWI	CAGTTCCAAA <u>cc</u> CGGAAGCTTG
24	2650	A > G	Asn > Ser	NS1	pUC-NS1	BsiWI	CCAACGAGCTA <u>cc</u> TACGTTCTCTGGG
25	3303	A > G	Arg > Gly	NS1	pUC-NS1	StyI	GATTGTGACCAT <u>gg</u> GGCCCATCTTTG
26	3442	A > G	Glu > Gly	NS1	pUC-NS1	BlpI	GGAGATTAGGCC <u>gc</u> TGAG <u>gc</u> GgtAAAGAAGAG
27	3540	G > A	Glu > Lys	NS2A	pUC-NS1	BsmI	GTTTGTGGAA <u>aa</u> AATGt <u>c</u> TGAGGAGAA
28	3575	G > A	Met > Ile	NS2A	pUC-NS1	SspI	CTAGGAAACACAT <u>aa</u> TATTAGTTTGTGG
29	3702	G > A	Ala > Thr	NS2A	pUC-NS2A	BglI	CAGATCCACCTA <u>aa</u> CCATaATGGCAGTG
30	3771	A > G	Arg > Gly	NS2A	pUC-NS2A	AvaI	GGAAACTCAC <u>cc</u> TC <u>gg</u> GAGAGACAGC
31	4059	A > G	Ile > Val	NS2A	pUC-NS2A	BstEII	TTGGGTAGAG <u>gc</u> Ta <u>cc</u> CGCACTCATCC
32	4062	A > G	Thr > Ala	NS2A	pUC-NS2A	BsrBI	GTAGAAATA <u>gg</u> C <u>cc</u> GCTCTCATCCTAG
33	4266	A > G	Ser > Gly	NS2B	pUC-NS2A	SnaBI	GGCGGCTTACGTaATG <u>gc</u> GaGGTAGCTCAGC
34	4306	A > G	Asn > Ser	NS2B	pUC-NS2A	AlwNI	CTAGAGAAGGCaGCTt <u>ct</u> GTGCAGTGG
35	4480	U > C	Val > Ala	NS2B	pUC-NS2A	MscI	CCTTGGC <u>cc</u> ATTCCAGcaACAATGAC
36	4812	G > A	Val > Ile	NS3	pUC-NS2A	ApoI	GACGTTCA <u>aa</u> TttTaGCCATAGAACC
37	4891	U > C	Ile > Thr	NS3	pUC-NS2A	KasI	CTGGAGAAA <u>cc</u> GG <u>gc</u> GCcGTAACATTAG
38	4896	G > U	Ala > Ser	NS3	pUC-NS2A	BstEII	GAAATTGGAt <u>cc</u> GTAAC <u>cc</u> TTAGATTTC
39	4907	A > U	Leu > Phe	NS3	pUC-NS2A	AclI	GGAGCAGTA <u>Ac</u> G <u>TT</u> tGATTTCAAACCC
40	4995	U > C	Ser > Pro	NS3	pUC-NS2A	BsaJI	GTACCAAA <u>cc</u> CtGGgGATTACGTC
41	5097	G > A	Asp > Asn	NS3	pUC-NS3	BspHI	GATTAACAT <u>t</u> cATG <u>a</u> ACTTACACCC
42	5695	A > G	Asp > Gly	NS3	pUC-NS3	BanI	GGAAAACCTTTG <u>gc</u> AC <u>cc</u> GAGTATCC
43	5872	C > U	Thr > Ile	NS3	pUC-NS3	BsrFI	TCCAGTGAt <u>a</u> CC <u>gc</u> GtAGCGTGCTC
44	6106	A > G	Glu > Gly	NS3	pUC-NS3	MscI	GCCTGAGAGGt <u>G</u> gcCAAAGGAAG
45	6259	U > C	Val > Ala	NS3	pUC-NS3	BglIII	ACATGGAGGcaG <u>A</u> gAT <u>c</u> TGGACTAGA
46	7153	U > C	Val > Ala	NS4B	pUC-NS4A	MscI	AAAGCATG <u>gc</u> CcAAGGATGCTGTC
47	7162	U > C	Leu > Ser	NS4B	pUC-NS4A	BlpI	GCATAATGGAGAC <u>cc</u> tAAGCATGACTAAGG
48	7163	A > C	Leu > Phe	NS4B	pUC-NS4A	ApaLI	TTATTGCATAgT <u>G</u> cACgAAAAGCATG

TABLE 12-continued

Mutagenic oligonucleotides used to generate recombinant DEN4 viruses containing single 5-FU mutations.							
SEQ ID NO.	Recombinant virus (rDEN4-)	Nucleotide change	Amino acid change	Gene	pUC clone	RE site <sup>a</sup>	Oligonucleotide <sup>b</sup>
49	7174	C > U	Ala > Val	NS4B	pUC-NS4A	BsaAI	GGGCCTATTATT <u>TA</u> <del>CGTA</del> ATGGAC
50	7182	G > A	Gly > Ser	NS4B	pUC-NS4A	n/a	CTGCAATCCTGGTgaTATTATTGC
51	7546	C > U	Ala > Val	NS4B	pUC-NS5A	AccII	CTCATAAAGAA <u>AcG</u> ttCAAACCT
52	7630	A > G	Lys > Arg	NS5	pUC-NS5A	HgaI	CATTAGACAG <u>Acg</u> cGAGTTTGAAG
53	7849	A > U	Asn > Ile	NS5	pUC-NS5A	HgaI	TGGCGAC <u>c</u> CTCAAGAtaGTGACTGAAG
54	8020	A > U	Asn > Ile	NS5	pUC-NS5A	ClaI	GAGTCATCa <u>TCg</u> AtaCCAACAATAG
55	8092	A > G	Glu > Gly	NS5	pUC-NS5A	EcoRI	CTTCAAAACCTG <u>gc</u> TTCTGCATCAAAG
56	8281	U > C	Leu > Ser	NS5	pUC-NS5B	XmnI	CAAAGATGTTGagcAACAGGTTCAACAAC
57	8730	A > C	Asn > His	NS5	pUC-NS5B	AvaI	GGAAAGAAGAAAc <u>AcCCg</u> AGACTGTGC
58	8872	A > G	Lys > Arg	NS5	pUC-NS5B	PvuI	GGGAAGTGGT <u>cGAt</u> cgAGAAAGGGC
59	9977	G > A	Met > Ile	NS5	pUC-NS5C	SfcI	CCAGTGGATt <u>Act</u> ACaGAAGATATGCTC
60	10186	U > C	Ile > Thr	NS5	pUC-NS5C	AgeI	CAGGAACCTG <u>Ac</u> CGgtAAAGAGGAATACG
61	10275	A > U	n/a	3' UTR	pUC-NS5C	n/a	CTGTAATTACCAACAtCAAACACCAAAG
62	10279	A > C	n/a	3' UTR	pUC-NS5C	n/a	CCAACAACAA <u>c</u> CACCAAAGGCTATTG
63	10634	U > C	n/a	3' UTR	pUC-3' UTR	n/a	GGATTGGTGTGT <u>c</u> GATCCAACAGG

<sup>a</sup>Primers were engineered which introduced (underline) or ablated (hatched line) translationally-silent restriction enzyme sites.

<sup>b</sup>Lowercase letters indicate nt changes and bold letters indicate the site of the 5-FU mutation, which in some oligonucleotides differs from the original nucleotide substitution change in order to create a unique restriction enzyme site. The change preserves the codon for the amino acid substitution.

TABLE 13

sp, ts and mouse attenuation phenotypes of rDEN4 mutant viruses encoding single mutations identified in six sp 5-FU mutant viruses.											
5-FU mutant virus		Gene/region containing mutation	Mean virus titer (log <sub>10</sub> PFU/ml) at indicated temp (° C.)						Replication in suckling mice <sup>b</sup>		
Virus			Vero cells			HuH-7 cells			Mean virus titer ± SE (log <sub>10</sub> PFU/g brain)	Mean log <sub>10</sub> -unit reduction from value for wt <sup>c</sup>	
			35	39	Δ <sup>a</sup>	35	39	Δ	n		
	2A-13		7.6	7.1	0.5	7.8	6.6	1.2	30	6.5 ± 0.1	—
	rDEN4		7.6	6.8	0.8	8.0	6.7	1.3	54	5.8 ± 0.1	—
	rDEN4Δ30		7.6	6.9	0.7	7.7	5.6	2.1	30	5.6 ± 0.1	0.2
738	parent		6.5	5.7	0.8	<sup>x</sup> 6.9	<u>3.1</u> <sup>e</sup>	3.8	12	4.4 ± 0.4	<b>2.3</b>
	rDEN4-3540	NS2A	6.9	5.1	1.8	7.4	<u>3.7</u>	3.7	12	4.1 ± 0.3	<b>1.7</b>
	rDEN4-7162	NS4B	7.2	6.8	0.4	7.4	6.6	0.8	8	5.6 ± 0.3	0.3
922	parent		7.3	<u>3.8</u>	3.5	<sup>x</sup> 7.4	<u>3.0</u>	4.4	12	3.5 ± 0.1	<b>2.9</b>
	rDEN4-4306	NS2B	<sup>x</sup> 5.0	<u>2.2</u>	2.8	<sup>x</sup> 5.6	<u>&lt;1.6</u>	>4.0	12	1.7 ± 0.1	<b>4.1</b>
	rDEN4-5872	NS3	5.7	<u>2.5</u>	3.2	<sup>x</sup> 6.5	<u>&lt;1.6</u>	>4.9	12	4.5 ± 0.3	1.3
	rDEN4-7163	NS4B	7.8	7.2	0.6	8.0	7.4	0.6	6	6.2 ± 0.2	(+)0.1
	rDEN4-10279	3' UTR	6.9	5.7	1.2	7.7	5.7	2.0	6	4.8 ± 0.2	0.7
1081	parent		6.9	<u>3.9</u>	3.0	<sup>x</sup> 5.8	<u>1.9</u>	3.9	12	2.6 ± 0.2	<b>3.9</b>
	rDEN4-2650	NS1	5.1	3.0	2.1	<sup>x</sup> 5.5	2.8	2.7	12	3.0 ± 0.3	<b>2.8</b>
	rDEN4-7163	NS4B	7.8	7.2	0.6	8.0	7.4	0.6	6	6.2 ± 0.2	(+)0.1
1083	parent		7.4	5.8	1.6	<sup>x</sup> 7.4	<u>&lt;1.6</u>	≥5.8	12	4.5 ± 0.4	<b>2.0</b>
	rDEN4-3702	NS2A	6.8	5.6	1.2	7.6	4.7	2.9	18	4.9 ± 0.3	0.9
	rDEN4-7153	NS4B	7.7	7.2	0.5	8.0	6.9	1.1	6	5.7 ± 0.1	0.2
	rDEN4-10634	3' UTR	4.9	<u>1.6</u>	3.3	<sup>x</sup> 5.7	<u>&lt;1.6</u>	≥4.1	12	2.4 ± 0.3	<b>3.4</b>
1136	parent		5.1	<u>&lt;1.6</u>	≥3.5	<sup>x</sup> 5.7	<u>&lt;1.6</u>	>4.1	6	2.9 ± 0.3	<b>4.5</b>
	rDEN4-3771	NS2A	7.0	4.6	2.4	<sup>x</sup> 7.6	3.7	3.9	12	2.6 ± 0.4	<b>3.2</b>
	rDEN4-4891	NS3	7.1	<u>&lt;1.6</u>	>5.5	<sup>x</sup> 7.4	<u>&lt;1.6</u>	>5.8	12	2.5 ± 0.3	<b>3.5</b>
	rDEN4-10275	3' UTR	6.9	5.8	1.1	7.1	5.2	1.9	6	5.0 ± 0.3	0.5

TABLE 13-continued

sp, ts and mouse attenuation phenotypes of rDEN4 mutant viruses encoding single mutations identified in six sp 5-FU mutant viruses.											
1189	parent		<sup>x</sup> 6.3	<u>3.8</u>	2.5	<sup>x</sup> 5.5	<u>&lt;1.6</u>	>3.9	12	3.2 ± 0.4	<b>3.7</b>
	rDEN4-3303	NS1	6.1	4.8	1.3	6.6	3.9	2.7	8	5.7 ± 0.4	0.2
	rDEN4-4812	NS3	7.0	6.3	0.7	7.1	6.3	0.8	12	4.8 ± 0.2	1.0
	rDEN4-5097	NS3	<sup>x</sup> 5.0	<u>&lt;1.6</u>	>3.4	<sup>x</sup> 4.6	<u>&lt;1.6</u>	>3.0	12	1.8 ± 0.1	<b>4.0</b>
	rDEN4-7182	NS4B	7.7	6.9	0.8	7.8	6.8	1.0	6	6.2 ± 0.1	(+)0.1
Replication in HuH-7-SCID mice <sup>d</sup>											
	5-FU mutant virus	Virus	Gene/region containing mutation		n	Mean peak virus titer ± SE (log <sub>10</sub> PFU/ml serum)		Mean log <sub>10</sub> -unit reduction from value for wt <sup>c</sup>			
		2A-13			29	6.8 ± 0.2		—			
		rDEN4			32	6.3 ± 0.2		—			
		rDEN4Δ30			18	5.4 ± 0.2		0.9			
	738	parent			9	5.4 ± 0.7		1.9			
		rDEN4-3540	NS2A		5	6.1 ± 0.3		(+)0.1			
		rDEN4-7162	NS4B		5	6.8 ± 0.6		0.3			
	922	parent			6	6.2 ± 0.2		0.4			
		rDEN4-4306	NS2B		5	5.2 ± 0.6		1.1			
		rDEN4-5872	NS3		5	6.2 ± 0.5		0.1			
		rDEN4-7163	NS4B		6	5.8 ± 0.6		(+)0.2			
		rDEN4-10279	3' UTR		4	6.7 ± 0.2		0.4			
	1081	parent			4	4.2 ± 0.5		<b>2.4</b>			
		rDEN4-2650	NS1		6	4.7 ± 0.5		<b>2.2</b>			
		rDEN4-7163	NS4B		6	5.8 ± 0.6		(+)0.2			
	1083	parent			9	4.4 ± 0.3		<b>2.9</b>			
		rDEN4-3702	NS2A		7	6.3 ± 0.3		0.2			
		rDEN4-7153	NS4B		4	5.9 ± 0.7		0.1			
		rDEN4-10634	3' UTR		7	3.3 ± 0.4		<b>3.6</b>			
	1136	parent			7	4.5 ± 0.4		1.2			
		rDEN4-3771	NS2A		4	6.4 ± 0.2		(+)0.1			
		rDEN4-4891	NS3		6	6.0 ± 0.5		0.3			
		rDEN4-10275	3' UTR		4	6.7 ± 0.3		0.4			
	1189	parent			13	2.3 ± 0.3		<b>3.8</b>			
		rDEN4-3303	NS1		4	6.3 ± 0.3		0.8			
		rDEN4-4812	NS3		5	6.1 ± 0.5		(+)0.5			
		rDEN4-5097	NS3		8	1.9 ± 0.1		<b>4.3</b>			
		rDEN4-7182	NS4B		6	6.3 ± 0.3		(+)0.7			

<sup>a</sup>Reduction in mean virus titer (log<sub>10</sub>PFU/ml) at 39° C. compared to permissive temperature (35° C.).  
<sup>b</sup>Groups of 6 suckling mice were inoculated i.c. with 10<sup>4</sup> PFU of virus. Brains were removed 5 days later, homogenized, and titered in Vero cells.  
<sup>c</sup>Comparison of mean virus titers of mice inoculated with mutant virus and concurrent DEN4 control. Bold denotes ≥50- or 100-fold decrease in replication in suckling or SCID-HuH-7 mice, respectively.  
<sup>d</sup>Groups of HuH-7-SCID mice were inoculated directly into the tumor with 10<sup>4</sup> PFU virus. Serum was collected on day 6 and 7 and titered in Vero cells.  
<sup>e</sup>Underlined values indicate a 2.5 or 3.5 log<sub>10</sub>PFU/ml reduction in titer in Vero cells or HuH-7 cells, respectively, at indicated temp when compared to permissive temp (35° C.).  
<sup>f</sup>Small plaque size at 35° C.; small plaques have a diameter of <1.0 mm compared to wild type plaque diameter of 1.5-2.0 mm in Vero cells, or a diameter of <0.4 mm compared to wild type plaque diameter of 0.75 to 1.0 mm in HuH-7 cells.

TABLE 14

Phenotypes of rDEN4 mutant viruses encoding single mutations identified in 10 5-FU mutant viruses that are in both Vero and HuH-7 cells.												
5-FU mutant viruses	rDEN4- Mutation (nt position)	Gene/region	Mean virus titer (log <sub>10</sub> PFU/ml) at indicated temp (° C.)									
			Vero cells					HuH-7 cells				
			35	37	39	39	Δ <sup>a</sup>	35	37	38	39	Δ
239, 489	parent		7.6	6.8	5.6	<u>3.3<sup>e</sup></u>	4.3	7.6	6.7	4.7	<u>2.5</u>	5.1
773	4995 <sup>f</sup>	NS3	5.7	4.9	3.6	<u>&lt;1.6</u>	>4.1	6.4	5.7	4.0	<u>&lt;1.6</u>	>4.8
473	parent		6.7	6.3	5.4	<u>2.0</u>	4.7	7.2	6.7	<u>3.7</u>	<u>1.9</u>	5.3
	4480	NS2B	6.7	6.3	6.0	<u>5.7</u>	1.0	7.6	7.2	6.0	<u>5.2</u>	2.4
	4995 <sup>f</sup>	NS3	5.7	4.9	3.6	<u>&lt;1.6</u>	>4.1	6.4	5.7	4.0	<u>&lt;1.6</u>	>4.8
759	parent		7.2	6.9	6.4	<u>4.7</u>	2.5	7.5	6.8	6.3	<u>3.1</u>	4.4
	4995 <sup>f</sup>	NS3	5.7	4.9	3.6	<u>&lt;1.6</u>	>4.1	6.4	5.7	4.0	<u>&lt;1.6</u>	>4.8
	8020	NS5	7.1	6.6	6.7	<u>5.9</u>	1.2	7.4	7.1	6.1	<u>5.4</u>	2.0
816	parent		6.8	6.4	5.8	<u>3.9</u>	2.9	7.5	6.2	5.5	<u>3.1</u>	4.4
	4995 <sup>f</sup>	NS3	5.7	4.9	3.6	<u>&lt;1.6</u>	>4.1	6.4	5.7	4.0	<u>&lt;1.6</u>	>4.8
	7174	NS4B	6.9	7.1	6.9	<u>6.1</u>	0.8	7.5	7.2	7.1	<u>5.6</u>	1.9
938	parent		7.1	6.5	5.6	<u>3.1</u>	4.0	7.2	6.4	5.6	<u>3.1</u>	4.1
	3442	NS1	5.1	3.6	4.3	<u>2.1</u>	3.0	5.9	4.9	3.9	<u>&lt;1.6</u>	4.3
	4995 <sup>f</sup>	NS3	5.7	4.9	3.6	<u>&lt;1.6</u>	>4.1	6.4	5.7	4.0	<u>&lt;1.6</u>	>4.8
	10275	3' UTR	6.9	6.4	6.4	<u>5.8</u>	1.1	7.1	6.8	7.1	<u>5.2</u>	1.9

TABLE 14-continued

Phenotypes of rDEN4 mutant viruses encoding single mutations identified in 10 5-FU mutant viruses that are in both Vero and HuH-7 cells.												
5-FU mutant viruses	rDEN4-Mutation (nt position)	Gene/region	Replication in 7-day mice <sup>b</sup>					Replication in HuH-7-SCID mice <sup>d</sup>				
			n	Mean log <sub>10</sub> reduction from wt <sup>c</sup> (log <sub>10</sub> PFU/g brain)				n	Mean log <sub>10</sub> reduction from wt <sup>c</sup> (log <sub>10</sub> PFU/ml serum)			
173	parent		7.0	6.1	<u>3.2</u>	<u>2.9</u>	4.1	7.0	<u>3.2</u>	<u>3.0</u>	<u>2.1</u>	4.9
	7163	NS4B	7.8	7.7	7.6	7.2	0.6	8.0	7.7	7.5	7.4	0.6
	7849	NS5	7.0	6.7	3.7	2.1	4.9	7.7	5.5	3.6	2.4	5.3
	8872	NS5	7.0	6.3	6.4	<u>4.4</u>	2.6	7.4	6.4	5.1	<u>2.9</u>	4.5
509	parent		6.2	5.8	5.5	<u>3.4</u>	2.8	6.5	6.1	4.5	<u>&lt;1.6</u>	>4.9
	4266	NS2B	5.9	6.1	6.1	5.2	0.7	6.7	6.1	5.7	5.3	1.4
	8092	NS5	5.0 <sup>x</sup>	4.6	4.6	<u>&lt;1.6</u>	>3.4	5.6 <sup>x</sup>	4.8	4.4	<u>&lt;1.6</u>	>4.0
1033	parent		6.7	6.0	5.9	<u>4.1</u>	2.6	6.9	5.6	4.7	<u>&lt;1.6</u>	>5.3
	4907	NS3	6.7	6.0	5.8	<u>4.0</u>	2.7	7.1	6.1	6.8	<u>2.3</u>	4.8
	8730	NS5	7.0	6.7	6.6	6.7	0.3	7.6	7.0	7.2	6.6	1.0
	9977	NS5	5.6	5.5	4.6	4.1	1.5	6.4	6.1	6.2	4.6	1.8

<sup>a</sup>Reduction in mean virus titer (log<sub>10</sub>PFU/ml) at 39° C. compared to permissive temperature (35° C.).

<sup>b</sup>Groups of 6 suckling mice were inoculated i.c. with 10<sup>4</sup> PFU of virus. Brains were removed 5 days later, homogenized, and titered in Vero cells.

<sup>c</sup>Comparison of mean virus titers of mice inoculated with mutant virus and concurrent DEN4 control. Bold denotes ≥50- or ≥100-fold decrease in replication in suckling or SCID-HuH-7 mice, respectively.

<sup>d</sup>Groups of HuH-7-SCID mice were inoculated directly into the tumor with 10<sup>4</sup> PFU virus. Serum was collected on day 6 and 7 and titered in Vero cells.

<sup>e</sup>Underlined values indicate a 2.5 or 3.5 log<sub>10</sub>PFU/ml reduction in titer in Vero cells or HuH-7 cells, respectively, at indicated temp when compared to permissive temp (35° C.).

<sup>f</sup>Data represents the results from a single rDEN4-4995 virus.

<sup>x</sup>Small plaque size at 35° C.; small plaques have a diameter of <1.0 mm compared to wild type plaque diameter of 1.5-2.0 mm in Vero cells, or a diameter of <0.4 mm compared to wild type plaque diameter of 0.75 to 1.0 mm in HuH-7 cells.

TABLE 15

sp, ts and mouse attenuation phenotypes of rDEN4 mutant viruses encoding single mutations identified in 3 HuH-7 cell-specific ts 5-FU mutant viruses.																
5-FU mutant viruses	rDEN4-Mutation (nt position)	Gene/region	Mean virus titer (log <sub>10</sub> PFU/ml) at indicated temp (° C.)										Replication in 7-day mice <sup>b</sup>		Replication in HuH-7-SCID mice <sup>b</sup>	
			Vero cells					HuH-7 cells					Mean log <sub>10</sub> reduction from wt <sup>c</sup>		Mean log <sub>10</sub> reduction from wt <sup>c</sup>	
			35	37	39	39	Δ <sup>a</sup>	35	37	38	39	Δ	n	(log <sub>10</sub> PFU/g brain) <sub>10</sub>	n	(log <sub>10</sub> PFU/ml serum)
686	parent		7.0	6.7	6.7	6.4	0.6	7.3	6.8	6.4	<u>2.2</u>	5.1	12	3.8	6	1.2
	3575	NS2A	6.9	6.9	7.1	7.0	0.1	7.9	6.8	6.9	4.9	3.0	12	2.3		nd <sup>e</sup>

TABLE 15-continued

sp, ts and mouse attenuation phenotypes of rDEN4 mutant viruses encoding single mutations identified in 3 HuH-7 cell-specific ts 5-FU mutant viruses.

rDEN4-5-FU mutant viruses		Mutation (nt position)		Mean virus titer (log <sub>10</sub> PFU/ml) at indicated temp (° C.)										Replication in 7-day mice <sup>b</sup>	Replication in HuH-7-SCID mice <sup>b</sup>		
		Gene/region		Vero cells					HuH-7 cells					Mean log <sub>10</sub> reduction from wt <sup>c</sup> (log <sub>10</sub> PFU/g brain) <sub>10</sub>	Mean log <sub>10</sub> reduction from wt <sup>c</sup> (log <sub>10</sub> PFU/ml serum)		
				35	37	39	39	<u>Δ<sup>a</sup></u>	35	37	38	39	<u>Δ</u>	n	n		
	4062		NS2A	6.8	6.6	6.3	4.7	2.1	6.9	6.8	7.0	<u>&lt;1.6</u>	>5.3	12	2.2	nd	
	7163		NS4B	7.8	7.7	7.6	7.2	0.6	8.0	7.7	7.5	7.4	0.6	6	(+)0.1	nd	
992	parent			7.3	7.1	6.8	5.9	1.4	7.4	6.9	5.0	<u>&lt;1.6</u>	>5.8	6	2.7	7	1.3
	5695		NS3	5.6	4.7	4.7	3.8	1.8	6.3	5.1	3.7	<u>&lt;1.6</u>	>4.7	6	2.8		nd
	7162		NS4B	7.2	7.3	6.6	6.8	0.4	7.4	7.3	7.3	6.6	0.8	8	0.3		nd
1175	parent			7.4	7.1	6.9	5.3	2.1	7.6	6.5	4.7	<u>3.3</u>	4.3	12	1.7	5	1.0
	7153		NS4B	7.7	7.7	7.6	7.2	0.5	8.0	7.8	7.5	6.9	1.1	6	0.2		nd
	10186		NS5	4.3	3.7	2.4	<u>&lt;1.6</u>	>2.7	5.1	<u>&lt;1.6</u>	<u>&lt;1.6</u>	<u>&lt;1.6</u>	>3.5	6	3.4		nd
	10275		3' UTR	6.9	6.4	6.4	<u>5.8</u>	1.1	7.1	6.8	7.1	5.2	1.9	6	0.5		nd

<sup>a</sup>Reduction in titer (log<sub>10</sub>PFU/ml) at 39° C. compared to permissive temperature (35° C.).

<sup>b</sup>Groups of 6 suckling mice were inoculated i.c. with 10<sup>4</sup> PFU virus. Brains were removed 5 days later, homogenized, and titered in Vero cells.

<sup>c</sup>Determined by comparing mean viral titers of mice inoculated with mutant virus and concurrent 2A-13 or rDEN4 wt control.

<sup>d</sup>Underlined values indicate a 2.5 or 3.5 log<sub>10</sub>PFU/ml reduction in titer in Vero cells or HuH-7 cells, respectively, at indicated temp when compared to permissive temp (35° C.).

TABLE 16

Temperature-sensitive (ts) and mouse brain attenuation (att) phenotypes of additional rDEN4 viruses encoding single 5-FU mutations.

5-FU mutant virus		Gene/region containing mutation	Mean virus titer (log <sub>10</sub> PFU/ml) at indicated temp (° C.)										Replication in suckling mice <sup>b</sup>			
			Vero cells					HuH-7 cells					Gene/region containing mutation	n	Mean virus titer ± SE (log <sub>10</sub> PFU/g brain)	Mean log <sub>10</sub> -unit reduction from value for wt <sup>c</sup>
virus		Virus	mutation	35	37	38	39	<u>Δ<sup>a</sup></u>	35	37	38	39	<u>Δ</u>			
695	rDEN4-40		5' UTR	7.4	7.2	6.7	6.2	1.2	7.6	7.5	7.1	5.8	1.8			
718	rDEN4-4059		NS2A	7.0	6.7	6.4	6.2	0.8	7.7	7.1	7.0	6.6	1.1			
311	rDEN4-4896		NS3	7.0	6.1	5.9	<u>4.2</u>	2.8	6.9 <sup>x</sup>	6.0	5.6	<u>3.3</u>	3.6			
695	rDEN4-6106		NS3	6.8	6.3	5.9	<u>3.9</u>	2.9	7.1	6.0	5.2	<u>3.4</u>	3.7			
631	rDEN4-6259		NS3	7.0	6.1	5.8	5.0	2.0	7.5	6.6	5.7	4.2	3.3			
695 <sup>e</sup>	rDEN4-7546		NS4B	7.5	7.6	7.4	6.6	0.9	7.7	7.6	7.3	5.7	2.0			
718	rDEN4-7630		NS5	7.0	6.9	6.9	6.4	0.6	7.4	7.4	7.2	6.8	0.6			
718	rDEN4-8281		NS5	6.4	6.6	6.7	5.4	1.0	7.6	7.6	7.0	5.1	2.5			

5-FU mutant virus		Gene/region containing mutation	n	Mean virus titer ± SE (log <sub>10</sub> PFU/g brain)	Mean log <sub>10</sub> -unit reduction from value for wt <sup>c</sup>
695	rDEN4-40	5' UTR		nd <sup>f</sup>	nd
718	rDEN4-4059	NS2A		nd	nd
311	rDEN4-4896	NS3	6	4.1 ± 0.4	2.0**
695	rDEN4-6106	NS3		nd	nd
631	rDEN4-6259	NS3	6	2.2 ± 0.2	3.9**
695 <sup>e</sup>	rDEN4-7546	NS4B		nd	nd
718	rDEN4-7630	NS5	6	5.0 ± 0.3	0.5
718	rDEN4-8281	NS5	6	5.0 ± 0.5	1.1

<sup>a</sup>Reduction in titer (log<sub>10</sub>PFU/ml) at 39° C. compared to titer at permissive temperature (35° C.).

<sup>b</sup>6 mice were inoculated i.c. with 10<sup>4</sup> PFU virus in 30 μl inoculum. Brains were removed 5 days later, homogenized, and titered on Vero cells. Limit of detection is 2.0 log<sub>10</sub>PFU/g.

<sup>c</sup>Determined by comparing mean viral titers of mice inoculated with sample virus and wt rDEN4 control.

<sup>d</sup>Underlined values indicate a 2.5 or 3.5 log<sub>10</sub>PFU/ml reduction in titer in Vero cells or HuH-7 cells, respectively, at indicated temperature when compared to permissive temperature (35° C.).

<sup>e</sup>The 7546 mutation is also present in nine other 5-FU mutant viruses.

<sup>x</sup>Small plaque size at 35° C.; small plaques have a diameter of <0.4 mm compared to wt plaque diameter of 0.75 to 1.0 mm in HuH-7 cells.

<sup>f</sup>not determined

\*\*The att phenotype is defined as a reduction of >1.5 log<sub>10</sub>PFU/g compared to wt virus.

TABLE 17

Growth of wt DEN-4 2A-13 in SCID mice transplanted with HuH-7 cells. <sup>a</sup>							5	
Dose (log <sub>10</sub> PFU/ml)	Mouse #	Virus titer						10
		log <sub>10</sub> PFU/ml serum		log <sub>10</sub> PFU/g tissue				
		day 3	day 5	Brain	Liver	Tumor		
4	87	2.7	5.9	2.0	6.9	8.0	10	
	88	2.0	5.9	3.8	3.3	8.0		
	89	<1.7	6.2	2.7	3.6	8.0		
	90	1.7	3.5	3.2	3.0	7.0		
5	84	<1.7	7.2	3.2	4.0	7.0	15	
	85	1.7	6.6	3.6	6.3	5.8		
6	91	4.4	8.3	6.0	7.3	8.0	15	
	92	4.2	7.7	3.3	6.9	7.3		
	93	4.0	6.6	3.3	5.7	8.4		
	94	4.3	8.1	5.8	7.8	7.5		

<sup>a</sup>SCID mice were injected i.p. with 10<sup>7</sup>HuH-7 human hepatoma cells. Approximately 8 weeks later, groups of tumor-bearing SCID-HuH-7 mice were inoculated with virus directly into the tumor. Serum and tissues were collected on day 5, processed, and titered in Vero cells. 20

TABLE 18

Combination of ts mutations, NS3 4995 and NS5 7849, in rDEN4 results in an additive ts phenotype.												
Virus	Mean virus titer (log <sub>10</sub> PFU/ml)										Replication in suckling mice <sup>b</sup>	
	at indicated temp (° C.)										Mean virus titer ± SE (log <sub>10</sub> PFU/g brain)	Mean log <sub>10</sub> reduction from wt <sup>c</sup>
	Vero cells					HuH-7 cells						
	35	37	38	39	Δ <sup>d</sup>	35	37	38	39	Δ		
2A-13 wt	7.1	7.1	6.9	6.8	0.3	7.4	7.3	6.7	6.4	1.0	6.9 ± 0.09	—
rDEN4 wt	7.0	6.8	6.6	6.4	0.6	7.5	7.3	6.7	6.4	1.1	6.5 ± 0.11	—
rDEN4Δ30	7.0	6.7	6.2	6.2	0.8	7.5	7.0	6.5	5.1	2.4	5.9 ± 0.21	0.6
rDEN4-4995	5.7	4.9	3.6	<1.6 <sup>d</sup>	>4.1	6.4	5.7	4.0	<1.6	>4.8	3.4 ± 0.10	3.1
rDEN4-7849	7.0	6.7	3.7	2.1	4.9	7.7	5.5	3.6	2.4	5.3	2.6 ± 0.29	3.9
rDEN4-4995-7849	5.9	2.8	<1.6	<1.6	>4.3	5.6	2.4	<1.6	<1.6	>4.0	2.3 ± 0.20	4.2

<sup>a</sup>Reduction in titer (log<sub>10</sub>PFU/ml) at 39° C. compared to titer at permissive temperature (35° C.).

<sup>b</sup>Groups of 6 suckling mice were inoculated i.c. with 10<sup>4</sup> PFU virus. Brains were removed 5 days later, homogenized, and titered in Vero cells. The limit of detection is 2.0 log<sub>10</sub>PFU/g.

<sup>c</sup>Determined by comparing mean viral titers of mice inoculated with sample virus and rDEN4 wt control.

<sup>d</sup>Underlined values indicate a 2.5 or 3.5 log<sub>10</sub>PFU/ml reduction in titer in Vero cells or HuH-7 cells, respectively, at indicated temperature when compared to permissive temperature.

TABLE 19

The 5-FU mutations are compatible with the Δ30 mutation for replication in the brain of suckling mice.				50
Virus	No. of mice/group	Mean virus titer ± SE (log <sub>10</sub> PFU/g brain) <sup>a</sup>	Mean log <sub>10</sub> -unit reduction from wt <sup>b</sup>	
rDEN4	12	6.0 ± 0.1	—	55
rDEN4Δ30	12	5.3 ± 0.1	0.7	
rDEN4-2650 <sup>c</sup>	12	3.7 ± 0.2	2.3	
rDEN4Δ30-2650	12	3.9 ± 0.1	2.1	
rDEN4-4995 <sup>d</sup>	6	3.5 ± 0.2	2.5	
rDEN4Δ30-4995	6	2.7 ± 0.4	3.3	60
rDEN4-8092 <sup>d</sup>	12	2.0 ± 0.1	4.0	
rDEN4Δ30-8092	6	3.2 ± 0.2	2.8	
rDEN4-10634 <sup>c</sup>	12	3.8 ± 0.1	2.2	
rDEN4Δ30-10634	12	3.6 ± 0.1	2.4	

<sup>a</sup>Groups of 6 suckling mice were inoculated i.c. with 10<sup>4</sup> PFU of virus. Brains were removed 5 days later, homogenized, and titered in Vero cells.

<sup>b</sup>Comparison of mean virus titers of mice inoculated with mutant virus and rDEN4 control.

<sup>c</sup>Mutation restricts growth in both mouse brain and HuH-7-SCID mice. 65

<sup>d</sup>Mutation restricts growth in mouse brain only. The 8092 mutation has not been tested in SCID-HuH7 mice.

TABLE 20

Temperature-sensitive and mouse brain attenuation phenotypes of viruses bearing charge-cluster-to-alanine mutations in the NS5 gene of DEN4.

Mutation <sup>a</sup>	Changed AAPair	# nt changed	Mean virus titer (log <sub>10</sub> PFU/ml at indicated temperature (° C.) <sup>b</sup> )									
			Vero Cells					HuH-7 Cells				
			35	37	38	39	Δ <sup>c</sup>	35	37	38	39	Δ
wt (rDEN4)	n/a	0	8.1	8.1	7.9	7.6	0.5	8.3	8.0	7.5	7.5	0.8
deletion	n/a	30	6.3	6.1	6.1	5.7	0.6	6.9	6.3	5.9	4.7	2.2
(rDEN4Δ30)												
21-22	D R	4	7.2	6.8	6.7	6.1	1.1	7.6	7.1	7.0	4.7	2.9
22-23	R K	4	7.0	7.8	6.9	3.7	3.3	7.6	7.6	6.5	<1.7	>5.9
23-24	K E	3	6.7	6.6	6.0	6.5	0.2	7.1	7.3	5.6	<1.7	>5.4
26-27	E E	3	7.8	7.6	6.8	4.0	3.8	8.4	8.2	7.3	4.9	3.5
46-47	K D	3	7.4	7.4	7.3	7.0	0.4	7.8	7.8	7.3	6.8	1.0
157-158	E E	3	6.5	7.2	5.1	5.1	1.4	7.6	7.4	5.9	<1.7	>5.9
200-201	K H	4	5.3	4.6	5.3	4.1	1.2	5.6	4.9	3.7	<1.7	>3.9
246-247	R H	5	6.9	5.8	5.7	5.4	1.5	6.4	6.1	6.1	5.5	0.9
253-254	E K	4	7.1	6.9	6.8	7.0	0.1	7.9	7.5	7.6	6.8	1.1
356-357	K E	3	7.7	7.6	7.0	7.0	0.7	8.0	7.3	6.4	<1.7	>6.3
387-388	K K	5	7.7	6.1	7.0	<1.7	>6.0	7.0	6.3	7.0	<1.7	>5.3
388-389	K K	5	5.1	4.5	<1.7	<1.7	>3.4	6.1	5.0	<1.7	<1.7	>4.4
396-397	R E	4	7.0	7.3	6.5	5.5	1.5	7.5	7.6	7.5	<1.7	>5.8
397-398	E E	2	7.0	7.1	7.0	3.0	4.0	8.0	7.6	7.0	<1.7	>6.3
436-437	D K	4	4.5	3.3	3.0	2.0	2.5	5.7	4.5	<1.7	<1.7	>4.0
500-501	R E	3	6.6	6.3	5.7	2.3	4.3	7.1	6.5	<1.7	<1.7	>5.4
520-521	E E	3	5.6	4.7	4.3	<1.7	>3.9	6.7	5.7	<1.7	<1.7	>5.0
523-524	D K	4	6.6	6.3	6.3	5.8	0.8	7.1	6.6	<1.7	<1.7	>5.4
524-525	K K	5	7.1	6.9	6.9	6.6	0.5	7.8	7.4	7.0	5.3	2.5
525-526	K D	4	7.8	7.1	7.6	6.8	1.0	7.9	7.7	8.0	6.9	1.0
596-597	K D	3	4.6	4.0	2.6	<1.7	>2.9	5.7	4.9	4.0	<1.7	>4.0
641-642	K E	4	7.3	6.9	6.9	5.2	2.1	7.8	7.5	7.2	6.9	0.9
642-643	E R	3	6.8	6.1	4.0	3.3	3.5	7.5	7.1	6.6	3.0	4.5
645-646	E K	4	6.3	5.3	5.9	3.1	3.2	6.4	5.8	5.5	4.5	1.9
649-650	K E	3	6.9	6.8	6.9	6.3	0.6	7.1	7.3	7.5	7.0	0.1
654-655	D R	4	6.3	5.7	<1.7	<1.7	>4.6	7.0	7.1	4.6	<1.7	>5.3
750-751	R E	3	7.1	7.1	6.9	5.7	1.4	7.8	6.9	6.5	5.6	2.2
808-809	E D	3	4.6	4.1	<1.7	<1.7	>2.9	5.2	<1.7	<1.7	<1.7	>3.5
820-821	E D	2	6.3	6.3	5.6	<1.7	>4.6	6.9	6.0	5.7	<1.7	>5.2
827-828	D K	4	6.9	6.3	6.3	5.9	1.0	7.5	6.9	5.0	<1.7	>5.8
877-878	K E	3	7.6	7.3	7.0	7.0	0.6	7.9	7.9	7.3	5.8	2.1
878-879	E E	3	7.6	7.3	7.3	7.1	0.5	8.1	8.1	7.9	6.6	1.5

Replication in suckling mice <sup>d</sup>					
Mutation <sup>a</sup>	Changed AA Pair	# nt changed	n	Mean titer ± SE (log <sub>10</sub> PFU/g brain)	Mean log reduction from wt <sup>c</sup>
wt (rDEN4)	n/a	0	48	6.0 ± 0.16	—
deletion	n/a	30	42	5.4 ± 0.22	0.6
(rDEN4Δ30)					
21-22	D R	4	6	5.0 ± 0.50	0.6
22-23	R K	4	6	2.6 ± 0.19	<b>2.9</b>
23-24	K E	3	18	4.7 ± 0.09	<b>1.5</b>
26-27	E E	3	6	5.7 ± 0.30	+0.1
46-47	K D	3	6	5.4 ± 0.42	0.5
157-158	E E	3	6	2.8 ± 0.31	<b>2.7</b>
200-201	K H	4	12	5.5 ± 0.45	0.8
246-247	R H	5	6	6.1 ± 0.17	+0.5
253-254	E K	4	6	6.2 ± 0.13	+0.6
356-357	K E	3	6	3.5 ± 0.58	<b>2.0</b>
387-388	K K	5	6	3.1 ± 0.33	<b>2.4</b>
388-389	K K	5	6	5.0 ± 0.23	1.4
396-397	R E	4	18	5.4 ± 0.35	1.1
397-398	E E	2	6	6.0 ± 0.22	0.8
436-437	D K	4	12	2.3 ± 0.14	<b>3.9</b>
500-501	R E	3	6	6.9 ± 0.49	+0.7
520-521	E E	3	6	5.2 ± 0.48	0.2
523-524	D K	4	6	4.2 ± 0.47	1.3
524-525	K K	5	6	3.4 ± 0.54	<b>2.1</b>
525-526	K D	4	6	3.7 ± 0.64	<b>1.8</b>
596-597	K D	3	6	5.9 ± 0.14	0.5
641-642	K E	4	6	4.7 ± 0.45	1.2
642-643	E R	3	12	2.6 ± 0.15	<b>3.6</b>
645-646	E K	4	6	5.4 ± 0.51	0.2
649-650	K E	3	12	6.4 ± 0.20	+0.2
654-655	D R	4	12	1.8 ± 0.10	<b>4.0</b>

TABLE 20-continued

Temperature-sensitive and mouse brain attenuation phenotypes of viruses bearing charge-cluster-to-alanine mutations in the NS5 gene of DEN4.					
750-751	R E	3	6	6.0 ± 0.18	0.7
808-809	E D	3	6	1.8 ± 0.05	3.1
820-821	E D	2	6	5n5 ± 0.33	1.2
827-828	D K	4	6	3.6 ± 0.76	<b>2.3</b>
877-878	K E	3	12	4.4 ± 0.65	<b>1.8</b>
878-879	E E	3	12	2.4 ± 0.10	<b>3.8</b>

<sup>a</sup>Positions of the amino acid pair mutated to an alanine pair; numbering starts at the amino terminus of the NS5 protein.

<sup>b</sup>Underlined values indicate a 2.5 or 3.5 log<sub>10</sub> PFU/ml reduction in titer in Vero or HuH-7 cells, respectively, at the indicated temperatures when compared to permissive temperature (35° C.).

<sup>c</sup>Reduction in titer (log<sub>10</sub> PFU/ml) at 39° C. compared to permissive temperature (35° C.).

<sup>d</sup>Groups of six mice were inoculated i.c. with 4.0 log<sub>10</sub> PFU virus in a 30 µl inoculum. The brain was removed 5 days later, homogenized, and titered in Vero cells.

<sup>e</sup>Determined by comparing mean viral titers in mice inoculated with sample virus and concurrent wt controls (n = 6). The attenuation phenotype is defined as a reduction of ≥1.5 log<sub>10</sub> PFU/g compared to wt virus; reductions of ≥1.5 are listed in boldface.

TABLE 21

SCID-HuH-7 attenuation phenotypes of viruses bearing charge-cluster-to-alanine mutations in the NS5 gene of DEN4.				
Replication in SCID-HuH-7 mice <sup>b</sup>				
Mutation <sup>a</sup>	AA changed	n	Mean peak virus titer ± SE (log <sub>10</sub> PFU/ml serum)	Mean log reduction from wt <sup>c</sup>
wt	na	21	5.4 ± 0.4	—
Δ30	na	4	3.7 ± 0.6	<b>2.5</b>
23-24	KE	19	4.7 ± 0.5	1.3
157-158	EE	6	4.6 ± 0.6	1.3
200-201	KH	12	3.7 ± 0.2	<b>2.6</b>
356-357	KE	10	6.3 ± 0.7	(-) 1.1
396-397	RE	12	4.4 ± 1.3	1.2
397-398	EE	6	6.0 ± 0.5	(-) 0.1
436-437	DK	6	3.6 ± 0.2	<b>2.6</b>
500-501	RE	8	5.1 ± 0.4	1.1
523-524	DK	5	5.3 ± 0.7	0.6
750-751	RE	8	5.1 ± 0.4	1.1
808-809	ED	8	3.2 ± 0.4	<b>3.0</b>
827-828	DK	5	2.9 ± 0.2	1.6
878-879	EE	5	4.4 ± 0.7	1.5

<sup>a</sup>Positions of the amino acid pair changed to a pair of alanines; numbering starts at the amino terminus of the NS5 protein.

<sup>b</sup>Groups of SCID-HuH-7 mice were inoculated directly into the tumor with 10<sup>4</sup> PFU virus. Serum was collected on days 6 and 7 and titered in Vero cells.

<sup>c</sup>Comparison of mean virus titers of mice inoculated with mutant virus and concurrent DEN4 control. Bold denotes a ≥100-fold decrease in replication. A (-) sign indicates an increase in replication relative to wt.

TABLE 22

Combination of paired charge-cluster-to-alanine mutations into double-pair mutant viruses.			
Mutation Pair 1	Mutation Pair 2	Recovered	
23-24	200-201	Yes	25
23-24	356-357	Yes	
23-24	396-397	Yes	
23-24	523-524	Yes	
23-24	827-828	No	30
157-158	200-201	No	
157-158	356-357	No	
157-158	396-397	No	
157-158	523-524	Yes	35
157-158	827-828	No	
827-828	200-201	No	
827-828	356-357	No	
827-828	396-397	Yes	40
827-828	523-524	No	

TABLE 23

Temperature-sensitive and mouse brain attenuation phenotypes of double charge-cluster-to-alanine mutants of the NS5 gene of rDEN4.												
Mean virus titer (log <sub>10</sub> PFU/ml) at indicated temperature (° C.) <sup>b</sup>												
Mutation <sup>a</sup>	Charged AA Pair	# nt changed	Vero Cells					HuH-7 cells				
			35	37	38	39	Δ <sup>c</sup>	35	37	38	39	Δ
wt	n/a	0	8.1	8.1	7.9	7.6	0.5	8.3	8.0	7.5	7.5	0.8
Δ30	n/a	30	6.3	6.1	6.1	5.7	0.6	6.9	6.3	5.9	4.7	2.2
23-24	KE	3	6.7	6.6	6.0	6.5	0.2	7.1	7.3	5.6	<u>&lt;1.7</u>	>5.4
200-201	KH	4	5.3	4.6	5.3	4.1	1.2	5.6	4.9	3.7	<u>&lt;1.7</u>	>3.9
23-24; 200-201	KE, KH	7	7.1	6.5	6.6	<u>&lt;1.7</u>	>5.4	7.8	7.3	<u>&lt;1.7</u>	<u>&lt;1.7</u>	>6.1
23-24	KE	3	6.7	6.6	6.0	6.5	0.2	7.1	7.3	5.6	<u>&lt;1.7</u>	>5.4
356-357	KE	3	7.7	7.6	7.0	7.0	0.7	8.0	7.3	6.4	<u>&lt;1.7</u>	>6.3
23-24; 356-357	KE, KE	6										
23-24	KE	3	6.7	6.6	6.0	6.5	0.2	7.1	7.3	5.6	<u>&lt;1.7</u>	>5.4
396-397	RE	4	7.0	7.3	6.5	5.5	1.5	7.5	7.6	7.5	<u>&lt;1.7</u>	>5.8
23-24; 396-397	KE, RE	7	6.3	4.9	<u>&lt;1.7</u>	<u>&lt;1.7</u>	>4.6	7.1	6.0	5.6	<u>&lt;1.7</u>	>5.4
157-158	EE	3	6.5	7.2	5.1	5.1	1.4	7.6	7.4	5.9	<u>&lt;1.7</u>	>5.9
396-397	RE	4	7.0	7.3	6.5	5.5	1.5	7.5	7.6	7.5	<u>&lt;1.7</u>	>5.8



TABLE 23-continued

Temperature-sensitive and mouse brain attenuation phenotypes of double charge-cluster-to-alanine mutants of the NS5 gene of rDEN4.												
157-158; 396-397	E E, R E	7										
157-158	E E	3	6.5	7.2	5.1	5.1	1.4	7.6	7.4	5.9	<u>&lt;1.7</u>	>5.9
523-524	D K	4	6.6	6.3	6.3	5.8	0.8	7.1	6.6	<u>&lt;1.7</u>	<u>&lt;1.7</u>	>5.4
157-158; 523-524	E E, D K	7	5.6	3.9	<u>&lt;1.7</u>	<u>&lt;1.7</u>	>3.9	6.3	4.1	<u>&lt;1.7</u>	<u>&lt;1.7</u>	>4.6
396-397	R E	4	7.0	7.3	6.5	5.5	1.5	7.5	7.6	7.5	<u>&lt;1.7</u>	>5.8
827-828	D K	4	6.9	6.3	6.3	5.9	1.0	7.5	6.9	5.0	<u>&lt;1.7</u>	>5.8
396-397; 827-828	R E, D K	8	7.0	6.5	6.0	<u>&lt;1.7</u>	5.3	>6.7	5.7	<u>&lt;1.7</u>	<u>&lt;1.7</u>	>5.0

Replication in suckling mice <sup>d</sup>					
Mutation <sup>a</sup>	Charged AA Pair	# nt changed	n	Mean virus titer ± SE (log <sub>10</sub> PFU/g brain)	Mean log reduction from wt <sup>e</sup>
wt	n/a	0	48	6.0 ± 0.16	—
Δ30	n/a	30	42	5.4 ± 0.22	0.6
23-24	K E	3	18	4.7 ± 0.09	<b>1.5</b>
200-201	K H	4	12	5.5 ± 0.45	0.8
23-24; 200-201	K E, K H	7	6	5.8 ± 0.16	0.6
23-24	K E	3	18	4.7 ± 0.09	<b>1.5</b>
356-357	K E	3	6	3.5 ± 0.58	<b>2.0</b>
23-24; 356-357	K E, K E	6			
23-24	K E	3	18	4.7 ± 0.09	<b>1.5</b>
396-397	R E	4	18	5.4 ± 0.35	1.1
23-24; 396-397	K E, R E	7	6	3.7 ± 0.44	<b>2.7</b>
157-158	E E	3	6	2.8 ± 0.31	<b>2.7</b>
396-397	R E	4	18	5.4 ± 0.35	1.1
157-158; 396-397	E E, R E	7	6	2.0 ± 0.12	<b>4.8</b>
157-158	E E	3	6	2.8 ± 0.31	<b>2.7</b>
523-524	D K	4	6	4.2 ± 0.47	1.3
157-158; 523-524	E E, D K	7			
396-397	R E	4	6	4.8 ± 0.54	<b>1.6</b>
827-828	D K	4	6	3.6 ± 0.76	<b>2.3</b>
396-397; 827-828	R E, D K	8	6	4.7 ± 0.10	1.2

<sup>a</sup>Positions of the amino acid pair mutated to an alanine pair; numbering starts at the amino terminus of the NS5 protein.

<sup>b</sup>Underlined values indicate a 2.5 or 3.5 log<sub>10</sub>PFU/ml reduction in titer in Vero or HuH-7 cells respectively, at the indicated temperatures when compared to permissive temperature (35° C.).

<sup>c</sup>Reduction in titer (log<sub>10</sub>PFU/ml) at 39° C. compared to permissive temperature (35° C.).

<sup>d</sup>Groups of six suckling mice were inoculated i.e. with 4.0 log<sub>10</sub>PFU virus in a 30 μl inoculum. Brains were removed 5 days later, homogenized, and titered in Vero cells.

<sup>e</sup>Determined by comparing mean viral titers in mice inoculated with sample virus and concurrent wt controls (n = 6); reductions ≥1.5 are listed in boldface.

TABLE 24

SCID-HuH-7 attenuation phenotypes of double charge-cluster-to-alanine mutants of the NS5 gene of rDEN4.				
Replication in SCID-HuH-7 mice <sup>b</sup>				
Mutation <sup>a</sup>	Charged AA Pair	n	Mean peak virus titer ± SE (log <sub>10</sub> PFU/ml serum)	Mean log reduction from wt <sup>c</sup>
wt	n/a	21	5.4 ± 0.4	—
Δ30	n/a	4	3.7 ± 0.6	<b>2.5</b>
23-24	K E	19	4.7 ± 0.5	1.3
200-201	K H	12	3.7 ± 0.2	<b>2.6</b>
23-24; 200-201	K E, K H	13	3.4 ± 0.1	<b>2.9</b>
23-24	K E	19	4.7 ± 0.5	1.3
356-357	K E	10	6.3 ± 0.7	(+) 1.1
23-24; 356-357	K E, K E	4	3.6 ± 0.3	<b>2.3</b>
23-24	K E	19	4.7 ± 0.5	1.3
396-397	R E	12	4.4 ± 1.3	1.2
23-24; 396-397	K E, R E	10	3.4 ± 0.5	<b>3.3</b>
157-158	E E	6	4.6 ± 0.6	1.3
396-397	R E	12	4.4 ± 1.3	1.2
157-158; 396-397	E E, R E	6	2.2 ± 0.2	<b>3.6</b>
157-158	E E	6	4.6 ± 0.6	1.3
523-524	D K	5	5.3 ± 0.7	0.6

TABLE 24-continued

SCID-HuH-7 attenuation phenotypes of double charge-cluster-to-alanine mutants of the NS5 gene of rDEN4.				
Replication in SCID-HuH-7 mice <sup>b</sup>				
Mutation <sup>a</sup>	Charged AA Pair	n	Mean peak virus titer ± SE (log <sub>10</sub> PFU/ml serum)	Mean log reduction from wt <sup>c</sup>
157-158; 523-524	E E, D K	3	5.1 ± 0.6	0.8
396-397	R E	12	4.4 ± 1.3	1.2
827-828	D K	5	2.9 ± 0.2	1.6
396-397; 827-828	R E, D K	4	4.1 ± 0.7	0.4

<sup>a</sup>Positions of the amino acid pair mutated to an alanine pair; numbering starts at the amino terminus of the NS5 protein.

<sup>b</sup>Groups of SCID-HuH-7 mice were inoculated directly into the tumor with 10<sup>4</sup> PFU of virus. Serum was collected on days 6 and 7 and titered in Vero cells.

<sup>c</sup>Comparison of mean virus titers of mice inoculated with mutant virus and concurrent DEN4 control. Bold denotes a ≥100-fold decrease in replication. A (+) sign indicates an increase in replication relative to wt.

TABLE 25

Phenotypes (temperature sensitivity, plaque size and replication in mouse brain and SCID-HuH-7 mice) of wt DEN4 and viruses containing the Δ30 and 7129 mutations.															
Virus ID	Mutation <sup>a</sup>	Mean virus titer (log <sub>10</sub> PFU/ml) at indicated temperature (° C.)							Replication in suckling mouse brain <sup>c</sup>			Replication in SCID-HuH-7 mice <sup>e</sup>			
		VERO			HUH7			C6/36	n	Mean virus titer ± SE (log <sub>10</sub> PFU/g brain)	Mean log reduction from wt <sup>d</sup>	n	Mean peak virus titer ± SE (log <sub>10</sub> PFU/ml serum) <sup>f</sup>	Mean log reduction from wt <sup>d</sup>	
		35	39	Δ <sup>b</sup>	35	39	Δ	32							
		35	39	Δ	35	39	Δ	32	n	Mean virus titer ± SE (log <sub>10</sub> PFU/g brain)	Mean log reduction from wt <sup>d</sup>	n	Mean peak virus titer ± SE (log <sub>10</sub> PFU/ml serum) <sup>f</sup>	Mean log reduction from wt <sup>d</sup>	
1-TD-1A	wt	7.3	6.8	0.5	8	6.8	1.2	8.3	36	6.1 ± 0.21	—	21	5.4 ± 0.4	—	
p4Δ30	Δ30	6.6	6.5	0.1	7.4	6.4	1.0		42	5.4 ± 0.22	0.6	4	3.7 ± 0.6	2.5	
5-1A1	C7129U	6.7	6.5	0.2	7.5	6	1.5	7.6*	6	6.2 ± 0.30	0.0				
rDEN4-7129-1A	C7129U	7.3	7.0	0.3	7.6	6.3	1.3	7.5*	6	7.2 ± 0.12	(-) 0.4	4	5.4 ± 0.8	(-) 0.8	
rDEN4Δ30-7129	C7129U + Δ30	7.0						7.1*							

<sup>a</sup>Position and identity of the mutated nucleotides.

<sup>b</sup>Reduction in titer (log<sub>10</sub> PFU/ml) at 39° C. compared to permissive temperature (35° C.).

<sup>c</sup>Groups of six suckling mice were inoculated i.c. with 4.0 log<sub>10</sub> PFU virus in a 30 μl inoculum. The brain was removed 5 days later, homogenized, and titered in Vero cells.

<sup>d</sup>Determined by comparing mean viral titers in mice inoculated with sample virus and concurrent wt controls (n = 6). The attenuation phenotype is defined as a ≥50- or ≥100-fold decrease in replication in suckling or SCID-HuH-7 mice, respectively. A (-) sign indicates an increase in replication relative to the wt control.

<sup>e</sup>Groups of SCID-HuH-7 mice were inoculated directly into the tumor with 10<sup>4</sup> PFU virus. Serum was collected on days 6 and 7 and titered in Vero cells.

\*Small plaque size.

TABLE 26

The 5-fluorouracil 5-1A1 small plaque mutant demonstrates a restriction of midgut infection following oral infection of <i>Aedes aegypti</i> mosquitoes.					
Virus tested	Dose ingested (log <sub>10</sub> PFU) <sup>a</sup>	No. mosquitoes tested	Midgut-only infection <sup>b</sup>	Disseminated infection <sup>c</sup>	Total no. infected <sup>d,e</sup>
wtDEN4	4.5	19	1 (5%)	17 (89%)	18 (95%)
(2A-13)	3.5	26	9 (35%)	7 (27%)	16 (62%)
	2.5	28	1 (4%)	0	1 (4%)
				OID <sub>50</sub> = 3.9	OID <sub>50</sub> = 3.3
5-1A1	3.5	34	4 (12%)	2 (6%)	6 (18%)
	2.5	9	0	1 (11%)	1 (11%)
	1.5	23	0	0	0
					OID <sub>50</sub> ≥ 3.9

<sup>a</sup> Amount of virus ingested, assuming a 2 μl bloodmeal.

<sup>b</sup> Number (percentage) of mosquitoes with detectable dengue virus antigen in midgut tissue, but no detectable dengue virus antigen in head; mosquitoes were assayed 21 days post-feed, and dengue virus antigen was identified by IFA.

<sup>c</sup> Number (percentage) of mosquitoes with detectable dengue virus antigen in both midgut and head tissue.

<sup>d</sup> Total number (percentage) of mosquitoes with detectable dengue virus antigen.

<sup>e</sup> The proportion of total infections caused by wild type DEN4 was significantly higher than the proportion caused by 5-1A1 (logistic regression, N = 426, P < 0.0001). There were too few disseminated infection caused by 5-1A1 to permit statistical analysis.

TABLE 27

The 5-fluorouracil 5-1A1 small plaque mutant demonstrates a restriction of infection following intrathoracic inoculation of <i>Toxorhynchites splendens</i> mosquitoes.			
Virus tested	Dose ingested (log <sub>10</sub> PFU) <sup>a</sup>	No. mosquitoes tested	No (%) infected <sup>c</sup>
wtDEN4	4.0	5	5 (100)
(2A-13)	3.0	4	4 (100)
	2.0	4	1 (25)
			MID <sub>50</sub> = 2.3 log <sub>10</sub> PFU
5-1A1	3.0	9	0
	2.0	7	1 (14)
	1.0	7	0
			MID <sub>50</sub> > 3.0 log <sub>10</sub> PFU

<sup>a</sup> Amount of virus inoculated in a 0.22 μl inoculum.

<sup>b</sup> Number (percentage) of mosquitoes with detectable dengue virus antigen in head tissue; mosquitoes were assayed 14 days post-inoculation, and dengue virus antigen was identified by IFA.

<sup>c</sup> The proportion of infections caused by wild type DEN4 was significantly higher than the proportion caused by 5-1A1 (logistic regression, N = 36, P < 0.01).

TABLE 28

Mutagenesis primers for the deletion or swap of sequences in DEN4 showing conserved differences from tick-borne flaviviruses.				
DEN4 nucleotides <sup>1</sup>	Type of mutation <sup>2</sup>	Mutagenesis Primer <sup>3</sup>	SEQ ID NO	
10508-10530	Δ	CTGGTGGGAGCCCAACACAAAAAC	64	55
10508-10530	swap	CTGGTGGGAGGAGAGAGAAA TTGGCAACTCCCCAACACAAAAAC	65	55
10535-10544	Δ	AGACCCCCCAAGCATATTGAC	66	65

TABLE 28-continued

Mutagenesis primers for the deletion or swap of sequences in DEN4 showing conserved differences from tick-borne flaviviruses.			
DEN4 nucleotides <sup>1</sup>	Type of mutation <sup>2</sup>	Mutagenesis Primer <sup>3</sup>	SEQ ID NO
10535-10544	swap	AGACCCCCCAATATTTCTCCTC CTATAGCATATTGAC	67
10541-10544	Δ	CCCAACACAAAGCATATTGAC	68

<sup>1</sup>Nucleotides numbered 5' to 3', in the opposite direction from FIG. 5.3

<sup>2</sup>Δ: deletion of specified DEN4 nucleotides; swap: exchange of specified DEN4 nucleotides with homologous sequence from Langat

<sup>3</sup>no swap mutation was made for nucleotides 10541-10544

TABLE 29

Virus titer and plaque size of 3' UTR mutant viruses in Vero and C6/36 cells.				
Virus	Vero		C6/36	
	Titer (log <sub>10</sub> PFU/ml)	Plaque size <sup>1</sup>	Titer (log <sub>10</sub> PFU/ml)	Plaque size
rDEN4Δ10508-10530	8.1	wt	7.5	wt
rDEN4swap10508-10530	5.4	sp	6.6	wt
rDEN4Δ10535-10544	5.8	wt	7.0	sp
rDEN4swap10535-10544	7.0	wt	7.3	wt
rDEN4Δ10541-10544	6.4	wt	>7.0	wt

<sup>1</sup>Plaque size is designated as equivalent to wild type (wt) or ≥50% of wild type (sp) on the designated cell type.

TABLE 30

Infectivity of wt DEN4 and 3' UTR mutants for <i>Toxorhynchites splendens</i> via intrathoracic inoculation.				
Virus	Dose (log <sub>10</sub> PFU) <sup>a</sup>	No. mosquitoes tested	% Infected <sup>b</sup>	MID <sub>50</sub> (log <sub>10</sub> PFU)
rDEN4 wt	3.3	6	83	2.3
	2.3	7	57	
	1.3	6	0	
	0.3	6	0	
rDEN4Δ10508-10530	4.4	8	0	
	3.4	9	11	
	2.4	4	0	

<sup>a</sup>Amount of virus inoculated in a 0.22 μl inoculum.

<sup>b</sup>Percentage of mosquitoes with detectable dengue virus antigen in head tissue; mosquitoes were assayed 14 days post-inoculation, and dengue virus antigen was identified by IFA

TABLE 31

Infectivity of 3' UTR swap mutant viruses for <i>Aedes aegypti</i> fed on an infectious bloodmeal.				
Virus Tested	Dose ingested (log <sub>10</sub> PFU) <sup>a</sup>	No. Mosquitoes Tested	Total No. Infected <sup>b,c</sup>	Disseminated Infections <sup>c,d</sup>
rDEN4	3.8	18	11 (61%)	4 (22%)
	2.8	15	5 (33%)	1 (6%)
	1.8	15	0	0
rDEN4swap10535-10544	3.8	25	5 (20%)	2 (8%)
	2.8	25	0	0
	1.8	20	0	0

<sup>a</sup>Amount of virus ingested, assuming a 2 μl bloodmeal.

<sup>b</sup>Number (%) of mosquitoes with detectable dengue virus antigen in the midgut tissue; mosquitoes were assayed either 14 d post-feed and dengue virus antigen was identified by IFA.

<sup>c</sup>At a dose of 3.8 log<sub>10</sub>PFU, rDEN4swap10535-10544 infected significantly fewer mosquitoes at the midgut than wt rDEN4 (Fisher's exact test, N = 43, P < 0.01), although disseminated infections were not significantly different (Fisher's exact test, N = 43, P = 0.38).

<sup>d</sup>Number (%) of mosquitoes with detectable dengue virus antigen in the head tissue.

TABLE 32

Putative Vero cell adaptation mutations derived from the set of 5-FU mutant viruses and other DEN4 viruses passaged in Vero cells.							
Nucleotide position	Gene/region (a.a. #) <sup>b</sup>	Nucleotide change	Amino acid change	No. of viruses with the mutation	Other DEN viruses passaged in Vero cells		
					Virus	Nucleotide change	Amino acid change
1455	E (452)	G > U	val > phe	5			
2280 <sup>1,2,3</sup>	E (727)	U > C	phe > leu	2			
4891 <sup>2,3</sup>	NS3 (1597)	U > C	ile > thr	2			
4995 <sup>1,2</sup>	NS3 (1599)	U > C	ser > pro	8			
7153	NS4B (2351)	U > C	val > ala	3	2AΔ30	U > C	val > ala
7162	NS4B (2354)	U > C	leu > ser	4	2A-1	U > C	leu > ser
7163	NS4B (2354)	A > U or C	leu > phe	7	rDEN4Δ30	A > U	leu > phe
					2A-13-1A1	A > U	leu > phe
7182 <sup>1,2,3</sup>	NS4B (2361)	G > A	gly > ser	2			
7546	NS4B (2482)	C > U	ala > val	10			
7630 <sup>3</sup>	NS5 (2510)	A > G	lys > arg	1	814669	A > G	lys > arg
10275	3' UTR	A > U	n/a <sup>c</sup>	6			
10279	3' UTR	A > C	n/a	4			

<sup>a</sup>Conservation with DEN1, DEN2, or DEN3 is designated by superscript. Lack of conservation is designated by no superscript.

<sup>b</sup>Amino acid position in DEN4 polyprotein beginning with the methionine residue of the C protein (nt 102-104) as residue #1.

<sup>c</sup>not applicable

87

TABLE 33

Sequence analysis of rDEN2/4A30 clone 27(p4)-2-2A2.			
Nucleotide	Gene	Mutation	
		Nucleotide	Amino acid
743	M anchor	G > A	Gly > Glu
1493	E	C > U	Ser > Phe
7544*	NS4B	C > U	Ala > Val

\*Same as DEN4 nucleotide position 7546

TABLE 34

Sequence analysis of rDEN2/4A30 clone 27(p3)-2-1A1.			
Nucleotide	Gene	Mutation	
		Nucleotide	Amino acid
1345	E	U > C	Tyr > His
4885*	NS3	G > A	Glu > Lys
8297	NS5	G > A	Arg > Lys

\*Codon adjacent to 5-FU mutation 4891

TABLE 35

Recombinant virus rDEN2/4A30 bearing Vero adaptation mutations can be recovery and titered on Vero cells.			
Virus	Virus titer in indicated cell line <sup>1</sup> (log <sub>10</sub> PFU/ml)		Virus titer following recovery in Vero cells (log <sub>10</sub> PFU/ml)
	C6/36	Vero	
rDEN2/4A30 wt	5.2	1.7	<0.7
rDEN2/4A30-7153	5.4	5.2	<0.7
rDEN2/4A30-7162	5.4	5.3	nd <sup>2</sup>
rDEN2/4A30-7182	4.7	4.9	2.3
rDEN2/4A30-7630	5.3	4.8	1.3

88

TABLE 35-continued

Recombinant virus rDEN2/4A30 bearing Vero adaptation mutations can be recovery and titered on Vero cells.			
Virus	Virus titer in indicated cell line <sup>1</sup> (log <sub>10</sub> PFU/ml)		Virus titer following recovery in Vero cells (log <sub>10</sub> PFU/ml)
	C6/36	Vero	
10 rDEN2/4A30-7153-7163	5.1	4.7	nd
rDEN2/4A30-7153-7182	4.1	3.2	nd
rDEN2/4A30-7546-7630	5.2	5.2	nd

<sup>1</sup>Virus recovered following transfection of C6/36 mosquito cells was terminally diluted once in C6/36 cells and titered simultaneously in C6/36 cells and Vero cells.  
<sup>2</sup>not determined

TABLE 36

Putative Vero cell adaptation mutations of dengue type 4 virus and the corresponding wildtype amino acid residue in other dengue viruses.						
Mutation	Amino acid position <sup>a</sup>	Mutant residue	Amino acid in indicated wt dengue virus <sup>b</sup>			
			DEN4	DEN1	DEN2	DEN3
25 1455	452	F	V	I	A	A
2280	727	L	<u>F</u> <sup>c</sup>	<u>F</u>	<u>F</u>	<u>F</u>
4891	1597	T	<u>I</u>	<u>V</u>	<u>I</u>	<u>I</u>
4995	1632	P	<u>S</u>	<u>S</u>	<u>S</u>	<u>N</u>
7129	2343	L	<u>P</u>	<u>P</u>	<u>P</u>	<u>P</u>
7153	2351	A	<u>V</u>	<u>F</u>	<u>F</u>	<u>L</u>
30 7162	2354	S	L	V	V	V
7163	2354	F	L	V	V	V
7182	2361	S	<u>G</u>	<u>G</u>	<u>G</u>	<u>G</u>
7546	2482	V	A	L	T	V
7630	2510	R	K	S	S	K

<sup>a</sup>Amino acid position is given for the polyprotein of DEN4<sup>b</sup>DEN4 = rDEN4 (GenBank AF326825); DEN1 = Western pacific (GenBank DVU88535); DEN2 = New Guinea C (GenBank AF038403); DEN3 = H87 (GenBank M93130)<sup>c</sup>Underlined nucleotides are shared between DEN4 and one or more additional DEN types.

TABLE 37

Mutations known to attenuate dengue type 4 virus and the corresponding wildtype amino acid residue in other dengue viruses.							
Mutation	Amino acid position <sup>a</sup>	Mutant residue	Amino acid in indicated wt dengue virus <sup>b</sup>				
			DEN4	DEN1	DEN2	DEN3	
5-FU mutations	2650	850	S	<u>N</u> <sup>d</sup>	<u>N</u>	<u>N</u>	<u>N</u>
	3442	1114	G	<u>E</u>	<u>E</u>	<u>E</u>	<u>E</u>
	3540	1147	K	<u>E</u>	<u>E</u>	<u>E</u>	<u>E</u>
	3575	1158	I	<u>M</u>	L	A	<u>M</u>
	3771	1224	G	<u>R</u>	R	K	<u>R</u>
	4062	1321	A	<u>T</u>	L	A	<u>T</u>
	4306	1402	S	N	E	D	D
	4891	1597	T	<u>I</u>	V	<u>I</u>	<u>I</u>
	4896	1599	S	<u>A</u>	<u>A</u>	<u>A</u>	<u>A</u>
	4907	1602	F	<u>L</u>	<u>L</u>	<u>L</u>	<u>L</u>
	4995	1632	P	<u>S</u>	<u>S</u>	<u>S</u>	<u>N</u>
	5097	1666	N	<u>D</u>	<u>D</u>	<u>D</u>	<u>D</u>
	5695	1865	G	<u>D</u>	<u>D</u>	<u>D</u>	<u>D</u>
	6259	2053	A	<u>V</u>	<u>V</u>	<u>V</u>	<u>V</u>
	7129 <sup>c</sup>	2343	L	<u>P</u>	<u>P</u>	<u>P</u>	<u>P</u>
	7849	2583	I	<u>N</u>	K	<u>N</u>	K
	8092	2664	G	E	Q	Q	Q
10186	3362	T	<u>I</u>	<u>I</u>	<u>I</u>	<u>I</u>	
10634	3' UTR	—	—	—	—	—	
Charge-cluster-to-alanine mutations	22, 23	2509, 2510	AA	<u>RK</u>	KS	KS	<u>RK</u>
	23, 24	2510, 2511	AA	<u>KE</u>	SE	SE	<u>KE</u>
	157, 158	2644, 2645	AA	<u>EE</u>	EE	EA	<u>EE</u>
	200, 201	2687, 2688	AA	<u>KH</u>	<u>KH</u>	KY	<u>KH</u>
	356, 357	2843, 2844	AA	<u>KE</u>	<u>KE</u>	<u>KE</u>	<u>KE</u>

TABLE 37-continued

Mutations known to attenuate dengue type 4 virus and the corresponding wildtype amino acid residue in other dengue virus.							
Mutation	Amino acid position <sup>a</sup>	Mutant residue	Amino acid in indicated wt dengue virus <sup>b</sup>				
			DEN4	DEN1	DEN2	DEN3	
387, 388	2874, 2875	AA	<u>KK</u>	RN	<u>KK</u>	RN	
436, 437	2923, 2924	AA	<u>DK</u>	HR	<u>DK</u>	<u>DK</u>	
524, 525	3011, 3012	AA	<u>KK</u>	KI	<u>KK</u>	KI	
525, 526	3012, 3013	AA	<u>KD</u>	IP	<u>KE</u>	IP	
642, 643	3129, 3130	AA	<u>ER</u>	<u>ER</u>	IA	KK	
654, 655	3141, 3142	AA	DR	ER	ER	ER	
808, 809	3295, 3296	AA	<u>ED</u>	<u>ED</u>	<u>ED</u>	<u>ED</u>	
827, 828	3314, 3315	AA	<u>DK</u>	<u>DK</u>	<u>DK</u>	<u>DK</u>	
877, 878	3364, 3365	AA	KE	NE	NE	NE	
878, 879	3365, 3366	AA	<u>EE</u>	EN	<u>EE</u>	<u>EE</u>	

<sup>a</sup>Amino acid position is given for the polyprotein of DEN4

<sup>b</sup>DEN4 = rDEN4 (GenBank AF326825); DEN1 = Western pacific (GenBank U88535); DEN2 = New Guinea C (GenBank AF038403); DEN3 = H87 (GenBank M93130)

<sup>c</sup>This mutation results in decreased replication of DEN4 in mosquitoes.

<sup>d</sup>Underlined nucleotides are shared between DEN4 and one or more additional DEN types.

While the present invention has been described in some detail for purposes of clarity and understanding, one skilled in the art will appreciate that various changes in form and detail can be made without departing from the true scope of the invention. All figures, tables, and appendices, as well as patents, applications, and publications, referred to above, are hereby incorporated by reference. 25

## SEQUENCE LISTING

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<212> TYPE: PRT

<213> ORGANISM: Dengue 4 virus

<400> SEQUENCE: 1

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Ile Leu Glu Val Asp Arg Thr Glu Ala Lys Ser Ala Leu Lys Asp Gly
 35             40             45
Ser Lys Ile Lys His Ala Val Ser Arg Gly Ser Ser Lys Ile Arg Trp
 50             55             60
Ile Val Glu Arg Gly Met Val Lys Pro Lys Gly Lys Val Val Asp Leu
 65             70             75             80
Gly Cys Gly Arg Gly Gly Trp Ser Tyr Tyr Met Ala Thr Leu Lys Asn
 85             90             95
Val Thr Glu Val Lys Gly Tyr Thr Lys Gly Gly Pro Gly His Glu Glu
 100            105            110
Pro Ile Pro Met Ala Thr Tyr Gly Trp Asn Leu Val Lys Leu His Ser
 115            120            125
Gly Val Asp Val Phe Tyr Lys Pro Thr Glu Gln Val Asp Thr Leu Leu
 130            135            140
Cys Asp Ile Gly Glu Ser Ser Ser Asn Pro Thr Ile Glu Glu Gly Arg
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Thr Leu Arg Val Leu Lys Met Val Glu Pro Trp Leu Ser Ser Lys Pro
 165            170            175

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 Glu Leu Glu Lys Leu Gln Arg Lys His Gly Gly Asn Leu Val Arg Cys  
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 Pro Leu Ser Arg Asn Ser Thr His Glu Met Tyr Trp Val Ser Gly Ala  
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 Ser Gly Asn Ile Val Ser Ser Val Asn Thr Thr Ser Lys Met Leu Leu  
 225 230 235 240  
 Asn Arg Phe Thr Thr Arg His Arg Lys Pro Thr Tyr Glu Lys Asp Val  
 245 250 255  
 Asp Leu Gly Ala Gly Thr Arg Ser Val Ser Thr Glu Thr Glu Lys Pro  
 260 265 270  
 Asp Met Thr Ile Ile Gly Arg Arg Leu Gln Arg Leu Gln Glu Glu His  
 275 280 285  
 Lys Glu Thr Trp His Tyr Asp Gln Glu Asn Pro Tyr Arg Thr Trp Ala  
 290 295 300  
 Tyr His Gly Ser Tyr Glu Ala Pro Ser Thr Gly Ser Ala Ser Ser Met  
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 340 345 350  
 Arg Val Phe Lys Glu Lys Val Asp Thr Arg Thr Pro Gln Pro Lys Pro  
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 Gly Thr Arg Met Val Met Thr Thr Thr Ala Asn Trp Leu Trp Ala Leu  
 370 375 380  
 Leu Gly Lys Lys Lys Asn Pro Arg Leu Cys Thr Arg Glu Glu Phe Ile  
 385 390 395 400  
 Ser Lys Val Arg Ser Asn Ala Ala Ile Gly Ala Val Phe Gln Glu Glu  
 405 410 415  
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 435 440 445  
 Ser Cys Val Tyr Asn Met Met Gly Lys Arg Glu Lys Lys Leu Gly Glu  
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 His Arg Leu Gly Tyr Ile Leu Glu Glu Ile Asp Lys Lys Asp Gly Asp  
 515 520 525  
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 530 535 540  
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Lys Arg Met Ala Ile Ser Gly Asp Asp Cys Val Val Lys Pro Leu Asp 660 665 670		
Glu Arg Phe Gly Thr Ser Leu Leu Phe Leu Asn Asp Met Gly Lys Val 675 680 685		
Arg Lys Asp Ile Pro Gln Trp Glu Pro Ser Lys Gly Trp Lys Asn Trp 690 695 700		
Gln Glu Val Pro Phe Cys Ser His His Phe His Lys Ile Phe Met Lys 705 710 715 720		
Asp Gly Arg Ser Leu Val Val Pro Cys Arg Asn Gln Asp Glu Leu Ile 725 730 735		
Gly Arg Ala Arg Ile Ser Gln Gly Ala Gly Trp Ser Leu Arg Glu Thr 740 745 750		
Ala Cys Leu Gly Lys Ala Tyr Ala Gln Met Trp Ser Leu Met Tyr Phe 755 760 765		
His Arg Arg Asp Leu Arg Leu Ala Ser Met Ala Ile Cys Ser Ala Val 770 775 780		
Pro Thr Glu Trp Phe Pro Thr Ser Arg Thr Thr Trp Ser Ile His Ala 785 790 795 800		
His His Gln Trp Met Thr Thr Glu Asp Met Leu Lys Val Trp Asn Arg 805 810 815		
Val Trp Ile Glu Asp Asn Pro Asn Met Thr Asp Lys Thr Pro Val His 820 825 830		
Ser Trp Glu Asp Ile Pro Tyr Leu Gly Lys Arg Glu Asp Leu Trp Cys 835 840 845		
Gly Ser Leu Ile Gly Leu Ser Ser Arg Ala Thr Trp Ala Lys Asn Ile 850 855 860		
His Thr Ala Ile Thr Gln Val Arg Asn Leu Ile Gly Lys Glu Glu Tyr 865 870 875 880		
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 aaaacagcat attgacgctg ggaaagacca gagatcctgc tgtctctca gcatcattcc 180  
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 taagccaagg gaaggactag aggttagagg agaccccggt ccaaaaacac caaaagaaac 120  
 agcatattga cacctgggat agactagggg atcttctgct ctgcacaacc agccacacgg 180  
 cacagtgcgc cgacataggt ggctggtggt gctagaacac aggatct 227

&lt;210&gt; SEQ ID NO 7

&lt;211&gt; LENGTH: 229

&lt;212&gt; TYPE: DNA

&lt;213&gt; ORGANISM: Japanese encephalitis virus

&lt;400&gt; SEQUENCE: 7

gactagaggt tagaggagac cccgtggaaa caacaacatg cggcccaagc ccctcgaag 60  
 ctgtagagga ggtggaagga ctagaggta gaggagacc cgcatttgca tcaaacagca 120  
 tattgacacc tgggaataga ctgggagatc ttctgctcta tctcaacatc agtactagg 180  
 cacagagcgc cgaagtatgt acgtggtggt gaggaagaac acaggatct 229

&lt;210&gt; SEQ ID NO 8

&lt;211&gt; LENGTH: 241



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<212> TYPE: DNA  
 <213> ORGANISM: Yellow fever virus

<400> SEQUENCE: 8

aacctggttt ctgggacctc ccaccccaga gtaaaaagaa cggagcctcc gctaccaccc 60  
 tcccacgtgg tggtagaaag acggggctca gaggttagag gagaccctcc agggaacaaa 120  
 tagtgggacc atattgacgc cagggaaaga ccggagtggg tctctgcttt tctccagag 180  
 gtctgtgagc acagtttgct caagaataag cagacctttg gatgacaaac acaaaaccac 240  
 t 241

<210> SEQ ID NO 9  
 <211> LENGTH: 249  
 <212> TYPE: DNA  
 <213> ORGANISM: Powassan virus

<400> SEQUENCE: 9

aaacgaactt tgtgagacca aaaggcctcc tggaaaggctc accaggagtt aggccgttta 60  
 ggagcccccg agcataactc gggaggaggg aggaagaaaa ttggcaatct tctcgggat 120  
 ttttccgctt cctatactaa atttccccca ggaaactggg ggggcggttc ttgttctccc 180  
 tgagccacca ccatccaggc acagatagcc tgacaaggag atgggtgtgtg actcggaaaa 240  
 acaccgct 249

<210> SEQ ID NO 10  
 <211> LENGTH: 250  
 <212> TYPE: DNA  
 <213> ORGANISM: Louping ill virus

<400> SEQUENCE: 10

tgcaagattt tgcgagacc cccgccccat gacaaggccg aacatggagc attaaagga 60  
 ggcccccgga agcatgcttc cgggaggagg gaagagagaa attggcagct ctcttcaggg 120  
 ttttctctcc tctatacca aatttcccc tgcacagagg gggggcggtt cttgttctcc 180  
 ctgagccacc atcaccaga cacagatagt ctgacaagga ggtgatgtgt gactcggaaa 240  
 aacaccgct 250

<210> SEQ ID NO 11  
 <211> LENGTH: 250  
 <212> TYPE: DNA  
 <213> ORGANISM: Tick-borne encephalitis virus

<400> SEQUENCE: 11

tgaaaaattt tgtgagacc cctgcatcat gataaggccg aacatggtgc atgaaagggg 60  
 aggcccccgga aagcacgctt ccgggaggag ggaagagaga aattggcagc tctcttcagg 120  
 attttctctc tctataca aaattcccc tggtagagg gggggcggtt cttgttctcc 180  
 ctgagccacc atcaccaga cacagtagt ctgacaagga ggtgatgtgt gactcggaaa 240  
 aacaccgct 250

<210> SEQ ID NO 12  
 <211> LENGTH: 247  
 <212> TYPE: DNA  
 <213> ORGANISM: Langat virus

<400> SEQUENCE: 12

tgtgaaactt tgtgagacc cttgcgtcca gagaaggccg aactgggcgt tataaggagg 60

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ccccagggg gaaaccctg ggaggagga agagagaaat tggcaactct cttcaggata 120
tttctctctc ctatacaaaa ttcccctcg tcagaggggg ggcgggttctt gttctccctg 180
agccaccatc acctagacac agatagtctg aaaaggaggt gatgcgtgtc tcggaaaaac 240
accgct 247

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&lt;210&gt; SEQ ID NO 13

&lt;211&gt; LENGTH: 3387

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Dengue 4 virus strain 2A

&lt;400&gt; SEQUENCE: 13

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Met Asn Gln Arg Lys Lys Val Val Arg Pro Pro Phe Asn Met Leu Lys
 1          5          10          15
Arg Glu Arg Asn Arg Val Ser Thr Pro Gln Gly Leu Val Lys Arg Phe
 20          25          30
Ser Thr Gly Leu Phe Ser Gly Lys Gly Pro Leu Arg Met Val Leu Ala
 35          40          45
Phe Ile Thr Phe Leu Arg Val Leu Ser Ile Pro Pro Thr Ala Gly Ile
 50          55          60
Leu Lys Arg Trp Gly Gln Leu Lys Lys Asn Lys Ala Ile Lys Ile Leu
 65          70          75          80
Ile Gly Phe Arg Lys Glu Ile Gly Arg Met Leu Asn Ile Leu Asn Gly
 85          90          95
Arg Lys Arg Ser Thr Ile Thr Leu Leu Cys Leu Ile Pro Thr Val Met
 100         105         110
Ala Phe Ser Leu Ser Thr Arg Asp Gly Glu Pro Leu Met Ile Val Ala
 115         120         125
Lys His Glu Arg Gly Arg Pro Leu Leu Phe Lys Thr Thr Glu Gly Ile
 130         135         140
Asn Lys Cys Thr Leu Ile Ala Met Asp Leu Gly Glu Met Cys Glu Asp
 145         150         155         160
Thr Val Thr Tyr Lys Cys Pro Leu Leu Val Asn Thr Glu Pro Glu Asp
 165         170         175
Ile Asp Cys Trp Cys Asn Leu Thr Ser Thr Trp Val Met Tyr Gly Thr
 180         185         190
Cys Thr Gln Ser Gly Glu Arg Arg Arg Glu Lys Arg Ser Val Ala Leu
 195         200         205
Thr Pro His Ser Gly Met Gly Leu Glu Thr Arg Ala Glu Thr Trp Met
 210         215         220
Ser Ser Glu Gly Ala Trp Lys His Ala Gln Arg Val Glu Ser Trp Ile
 225         230         235         240
Leu Arg Asn Pro Gly Phe Ala Leu Leu Ala Gly Phe Met Ala Tyr Met
 245         250         255
Ile Gly Gln Thr Gly Ile Gln Arg Thr Val Phe Phe Val Leu Met Met
 260         265         270
Leu Val Ala Pro Ser Tyr Gly Met Arg Cys Val Gly Val Gly Asn Arg
 275         280         285
Asp Phe Val Glu Gly Val Ser Gly Gly Ala Trp Val Asp Leu Val Leu
 290         295         300
Glu His Gly Gly Cys Val Thr Thr Met Ala Gln Gly Lys Pro Thr Leu
 305         310         315         320
Asp Phe Glu Leu Thr Lys Thr Thr Ala Lys Glu Val Ala Leu Leu Arg
 325         330         335

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Thr	Tyr	Cys	Ile	Glu	Ala	Ser	Ile	Ser	Asn	Ile	Thr	Thr	Ala	Thr	Arg
			340					345					350		
Cys	Pro	Thr	Gln	Gly	Glu	Pro	Tyr	Leu	Lys	Glu	Glu	Gln	Asp	Gln	Gln
		355					360					365			
Tyr	Ile	Cys	Arg	Arg	Asp	Val	Val	Asp	Arg	Gly	Trp	Gly	Asn	Gly	Cys
	370					375					380				
Gly	Leu	Phe	Gly	Lys	Gly	Gly	Val	Val	Thr	Cys	Ala	Lys	Phe	Ser	Cys
385					390					395					400
Ser	Gly	Lys	Ile	Thr	Gly	Asn	Leu	Val	Gln	Ile	Glu	Asn	Leu	Glu	Tyr
				405					410					415	
Thr	Val	Val	Val	Thr	Val	His	Asn	Gly	Asp	Thr	His	Ala	Val	Gly	Asn
			420					425					430		
Asp	Thr	Ser	Asn	His	Gly	Val	Thr	Ala	Met	Ile	Thr	Pro	Arg	Ser	Pro
		435					440					445			
Ser	Val	Glu	Val	Lys	Leu	Pro	Asp	Tyr	Gly	Glu	Leu	Thr	Leu	Asp	Cys
	450					455					460				
Glu	Pro	Arg	Ser	Gly	Ile	Asp	Phe	Asn	Glu	Met	Ile	Leu	Met	Lys	Met
465					470					475					480
Lys	Lys	Lys	Thr	Trp	Leu	Val	His	Lys	Gln	Trp	Phe	Leu	Asp	Leu	Pro
				485					490					495	
Leu	Pro	Trp	Thr	Ala	Gly	Ala	Asp	Thr	Ser	Glu	Val	His	Trp	Asn	Tyr
			500					505					510		
Lys	Glu	Arg	Met	Val	Thr	Phe	Lys	Val	Pro	His	Ala	Lys	Arg	Gln	Asp
		515					520					525			
Val	Thr	Val	Leu	Gly	Ser	Gln	Glu	Gly	Ala	Met	His	Ser	Ala	Leu	Ala
	530					535					540				
Gly	Ala	Thr	Glu	Val	Asp	Ser	Gly	Asp	Gly	Asn	His	Met	Phe	Ala	Gly
545					550					555					560
His	Leu	Lys	Cys	Lys	Val	Arg	Met	Glu	Lys	Leu	Arg	Ile	Lys	Gly	Met
			565						570					575	
Ser	Tyr	Thr	Met	Cys	Ser	Gly	Lys	Phe	Ser	Ile	Asp	Lys	Glu	Met	Ala
			580					585					590		
Glu	Thr	Gln	His	Gly	Thr	Thr	Val	Val	Lys	Val	Lys	Tyr	Glu	Gly	Ala
		595					600					605			
Gly	Ala	Pro	Cys	Lys	Val	Pro	Ile	Glu	Ile	Arg	Asp	Val	Asn	Lys	Glu
	610					615					620				
Lys	Val	Val	Gly	Arg	Ile	Ile	Ser	Ser	Thr	Pro	Leu	Ala	Glu	Asn	Thr
625					630					635					640
Asn	Ser	Val	Thr	Asn	Ile	Glu	Leu	Glu	Pro	Pro	Phe	Gly	Asp	Ser	Tyr
				645					650					655	
Ile	Val	Ile	Gly	Val	Gly	Asn	Ser	Ala	Leu	Thr	Leu	His	Trp	Phe	Arg
			660					665					670		
Lys	Gly	Ser	Ser	Ile	Gly	Lys	Met	Phe	Glu	Ser	Thr	Tyr	Arg	Gly	Ala
		675					680					685			
Lys	Arg	Met	Ala	Ile	Leu	Gly	Glu	Thr	Ala	Trp	Asp	Phe	Gly	Ser	Val
	690					695					700				
Gly	Gly	Leu	Phe	Thr	Ser	Leu	Gly	Lys	Ala	Val	His	Gln	Val	Phe	Gly
705					710					715					720
Ser	Val	Tyr	Thr	Thr	Met	Phe	Gly	Gly	Val	Ser	Trp	Met	Ile	Arg	Ile
				725					730					735	
Leu	Ile	Gly	Phe	Leu	Val	Leu	Trp	Ile	Gly	Thr	Asn	Ser	Arg	Asn	Thr
			740					745					750		
Ser	Met	Ala	Met	Thr	Cys	Ile	Ala	Val	Gly	Gly	Ile	Thr	Leu	Phe	Leu

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755				760				765							
Gly	Phe	Thr	Val	Gln	Ala	Asp	Met	Gly	Cys	Val	Val	Ser	Trp	Ser	Gly
770						775					780				
Lys	Glu	Leu	Lys	Cys	Gly	Ser	Gly	Ile	Phe	Val	Val	Asp	Asn	Val	His
785					790					795				800	
Thr	Trp	Thr	Glu	Gln	Tyr	Lys	Phe	Gln	Pro	Glu	Ser	Pro	Ala	Arg	Leu
				805					810					815	
Ala	Ser	Ala	Ile	Leu	Asn	Ala	His	Lys	Asp	Gly	Val	Cys	Gly	Ile	Arg
			820					825					830		
Ser	Thr	Thr	Arg	Leu	Glu	Asn	Val	Met	Trp	Lys	Gln	Ile	Thr	Asn	Glu
			835				840					845			
Leu	Asn	Tyr	Val	Leu	Trp	Glu	Gly	Gly	His	Asp	Leu	Thr	Val	Val	Ala
	850					855					860				
Gly	Asp	Val	Lys	Gly	Val	Leu	Thr	Lys	Gly	Lys	Arg	Ala	Leu	Thr	Pro
865					870					875					880
Pro	Val	Ser	Asp	Leu	Lys	Tyr	Ser	Trp	Lys	Thr	Trp	Gly	Lys	Ala	Lys
				885					890					895	
Ile	Phe	Thr	Pro	Glu	Ala	Arg	Asn	Ser	Thr	Phe	Leu	Ile	Asp	Gly	Pro
			900						905				910		
Asp	Thr	Ser	Glu	Cys	Pro	Asn	Glu	Arg	Arg	Ala	Trp	Asn	Ser	Leu	Glu
			915				920						925		
Val	Glu	Asp	Tyr	Gly	Phe	Gly	Met	Phe	Thr	Thr	Asn	Ile	Trp	Met	Lys
	930					935						940			
Phe	Arg	Glu	Gly	Ser	Ser	Glu	Val	Cys	Asp	His	Arg	Leu	Met	Ser	Ala
945					950					955					960
Ala	Ile	Lys	Asp	Gln	Lys	Ala	Val	His	Ala	Asp	Met	Gly	Tyr	Trp	Ile
				965					970					975	
Glu	Ser	Ser	Lys	Asn	Gln	Thr	Trp	Gln	Ile	Glu	Lys	Ala	Ser	Leu	Ile
			980						985				990		
Glu	Val	Lys	Thr	Cys	Leu	Trp	Pro	Lys	Thr	His	Thr	Leu	Trp	Ser	Asn
		995					1000						1005		
Gly	Val	Leu	Glu	Ser	Gln	Met	Leu	Ile	Pro	Lys	Ser	Tyr	Ala	Gly	Pro
	1010					1015							1020		
Phe	Ser	Gln	His	Asn	Tyr	Arg	Gln	Gly	Tyr	Ala	Thr	Gln	Thr	Val	Gly
1025					1030					1035					1040
Pro	Trp	His	Leu	Gly	Lys	Leu	Glu	Ile	Asp	Phe	Gly	Glu	Cys	Pro	Gly
				1045					1050					1055	
Thr	Thr	Val	Thr	Ile	Gln	Glu	Asp	Cys	Asp	His	Arg	Gly	Pro	Ser	Leu
			1060						1065				1070		
Arg	Thr	Thr	Thr	Ala	Ser	Gly	Lys	Leu	Val	Thr	Gln	Trp	Cys	Cys	Arg
			1075				1080						1085		
Ser	Cys	Thr	Met	Pro	Pro	Leu	Arg	Phe	Leu	Gly	Glu	Asp	Gly	Cys	Trp
	1090					1095						1100			
Tyr	Gly	Met	Glu	Ile	Arg	Pro	Leu	Ser	Glu	Lys	Glu	Glu	Asn	Met	Val
1105					1110					1115					1120
Lys	Ser	Gln	Val	Thr	Ala	Gly	Gln	Gly	Thr	Ser	Glu	Thr	Phe	Ser	Met
				1125					1130					1135	
Gly	Leu	Leu	Cys	Leu	Thr	Leu	Phe	Val	Glu	Glu	Cys	Leu	Arg	Arg	Arg
			1140						1145				1150		
Val	Thr	Arg	Lys	His	Met	Ile	Leu	Val	Val	Val	Ile	Thr	Leu	Cys	Ala
			1155				1160						1165		
Ile	Ile	Leu	Gly	Gly	Leu	Thr	Trp	Met	Asp	Leu	Leu	Arg	Ala	Leu	Ile
						1175						1180			

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Met Leu Gly Asp Thr Met Ser Gly Arg Ile Gly Gly Gln Ile His Leu  
 1185 1190 1195 1200  
 Ala Ile Met Ala Val Phe Lys Met Ser Pro Gly Tyr Val Leu Gly Val  
 1205 1210 1215  
 Phe Leu Arg Lys Leu Thr Ser Arg Glu Thr Ala Leu Met Val Ile Gly  
 1220 1225 1230  
 Met Ala Met Thr Thr Val Leu Ser Ile Pro His Asp Leu Met Glu Leu  
 1235 1240 1245  
 Ile Asp Gly Ile Ser Leu Gly Leu Ile Leu Leu Lys Ile Val Thr Gln  
 1250 1255 1260  
 Phe Asp Asn Thr Gln Val Gly Thr Leu Ala Leu Ser Leu Thr Phe Ile  
 1265 1270 1275 1280  
 Arg Ser Thr Met Pro Leu Val Met Ala Trp Arg Thr Ile Met Ala Val  
 1285 1290 1295  
 Leu Phe Val Val Thr Leu Ile Pro Leu Cys Arg Thr Ser Cys Leu Gln  
 1300 1305 1310  
 Lys Gln Ser His Trp Val Glu Ile Thr Ala Leu Ile Leu Gly Ala Gln  
 1315 1320 1325  
 Ala Leu Pro Val Tyr Leu Met Thr Leu Met Lys Gly Ala Ser Arg Arg  
 1330 1335 1340  
 Ser Trp Pro Leu Asn Glu Gly Ile Met Ala Val Gly Leu Val Ser Leu  
 1345 1350 1355 1360  
 Leu Gly Ser Ala Leu Leu Lys Asn Asp Val Pro Leu Ala Gly Pro Met  
 1365 1370 1375  
 Val Ala Gly Gly Leu Leu Leu Ala Ala Tyr Val Met Ser Gly Ser Ser  
 1380 1385 1390  
 Ala Asp Leu Ser Leu Glu Lys Ala Ala Asn Val Gln Trp Asp Glu Met  
 1395 1400 1405  
 Ala Asp Ile Thr Gly Ser Ser Pro Ile Ile Glu Val Lys Gln Asp Glu  
 1410 1415 1420  
 Asp Gly Ser Phe Ser Ile Arg Asp Val Glu Glu Thr Asn Met Ile Thr  
 1425 1430 1435 1440  
 Leu Leu Val Lys Leu Ala Leu Ile Thr Val Ser Gly Leu Tyr Pro Leu  
 1445 1450 1455  
 Ala Ile Pro Val Thr Met Thr Leu Trp Tyr Met Trp Gln Val Lys Thr  
 1460 1465 1470  
 Gln Arg Ser Gly Ala Leu Trp Asp Val Pro Ser Pro Ala Ala Thr Lys  
 1475 1480 1485  
 Lys Ala Ala Leu Ser Glu Gly Val Tyr Arg Ile Met Gln Arg Gly Leu  
 1490 1495 1500  
 Phe Gly Lys Thr Gln Val Gly Val Gly Ile His Met Glu Gly Val Phe  
 1505 1510 1515 1520  
 His Thr Met Trp His Val Thr Arg Gly Ser Val Ile Cys His Glu Thr  
 1525 1530 1535  
 Gly Arg Leu Glu Pro Ser Trp Ala Asp Val Arg Asn Asp Met Ile Ser  
 1540 1545 1550  
 Tyr Gly Gly Gly Trp Arg Leu Gly Asp Lys Trp Asp Lys Glu Glu Asp  
 1555 1560 1565  
 Val Gln Val Leu Ala Ile Glu Pro Gly Lys Asn Pro Lys His Val Gln  
 1570 1575 1580  
 Thr Lys Pro Gly Leu Phe Lys Thr Leu Thr Gly Glu Ile Gly Ala Val  
 1585 1590 1595 1600

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Thr Leu Asp Phe Lys Pro Gly Thr Ser Gly Ser Pro Ile Ile Asn Arg  
 1605 1610 1615  
 Lys Gly Lys Val Ile Gly Leu Tyr Gly Asn Gly Val Val Thr Lys Ser  
 1620 1625 1630  
 Gly Asp Tyr Val Ser Ala Ile Thr Gln Ala Glu Arg Ile Gly Glu Pro  
 1635 1640 1645  
 Asp Tyr Glu Val Asp Glu Asp Ile Phe Arg Lys Lys Arg Leu Thr Ile  
 1650 1655 1660  
 Met Asp Leu His Pro Gly Ala Gly Lys Thr Lys Arg Ile Leu Pro Ser  
 1665 1670 1675 1680  
 Ile Val Arg Glu Ala Leu Lys Arg Arg Leu Arg Thr Leu Ile Leu Ala  
 1685 1690 1695  
 Pro Thr Arg Val Val Ala Ala Glu Met Glu Glu Ala Leu Arg Gly Leu  
 1700 1705 1710  
 Pro Ile Arg Tyr Gln Thr Pro Ala Val Lys Ser Glu His Thr Gly Arg  
 1715 1720 1725  
 Glu Ile Val Asp Leu Met Cys His Ala Thr Phe Thr Thr Arg Leu Leu  
 1730 1735 1740  
 Ser Ser Thr Arg Val Pro Asn Tyr Asn Leu Ile Val Met Asp Glu Ala  
 1745 1750 1755 1760  
 His Phe Thr Asp Pro Ser Ser Val Ala Ala Arg Gly Tyr Ile Ser Thr  
 1765 1770 1775  
 Arg Val Glu Met Gly Glu Ala Ala Ala Ile Phe Met Thr Ala Thr Pro  
 1780 1785 1790  
 Pro Gly Ala Thr Asp Pro Phe Pro Gln Ser Asn Ser Pro Ile Glu Asp  
 1795 1800 1805  
 Ile Glu Arg Glu Ile Pro Glu Arg Ser Trp Asn Thr Gly Phe Asp Trp  
 1810 1815 1820  
 Ile Thr Asp Tyr Gln Gly Lys Thr Val Trp Phe Val Pro Ser Ile Lys  
 1825 1830 1835 1840  
 Ala Gly Asn Asp Ile Ala Asn Cys Leu Arg Lys Ser Gly Lys Lys Val  
 1845 1850 1855  
 Ile Gln Leu Ser Arg Lys Thr Phe Asp Thr Glu Tyr Pro Lys Thr Lys  
 1860 1865 1870  
 Leu Thr Asp Trp Asp Phe Val Val Thr Thr Asp Ile Ser Glu Met Gly  
 1875 1880 1885  
 Ala Asn Phe Arg Ala Gly Arg Val Ile Asp Pro Arg Arg Cys Leu Lys  
 1890 1895 1900  
 Pro Val Ile Leu Pro Asp Gly Pro Glu Arg Val Ile Leu Ala Gly Pro  
 1905 1910 1915 1920  
 Ile Pro Val Thr Pro Ala Ser Ala Ala Gln Arg Arg Gly Arg Ile Gly  
 1925 1930 1935  
 Arg Asn Pro Ala Gln Glu Asp Asp Gln Tyr Val Phe Ser Gly Asp Pro  
 1940 1945 1950  
 Leu Lys Asn Asp Glu Asp His Ala His Trp Thr Glu Ala Lys Met Leu  
 1955 1960 1965  
 Leu Asp Asn Ile Tyr Thr Pro Glu Gly Ile Ile Pro Thr Leu Phe Gly  
 1970 1975 1980  
 Pro Glu Arg Glu Lys Thr Gln Ala Ile Asp Gly Glu Phe Arg Leu Arg  
 1985 1990 1995 2000  
 Gly Glu Gln Arg Lys Thr Phe Val Glu Leu Met Arg Arg Gly Asp Leu  
 2005 2010 2015  
 Pro Val Trp Leu Ser Tyr Lys Val Ala Ser Ala Gly Ile Ser Tyr Lys

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2020			2025			2030									
Asp	Arg	Glu	Trp	Cys	Phe	Thr	Gly	Glu	Arg	Asn	Asn	Gln	Ile	Leu	Glu
	2035						2040					2045			
Glu	Asn	Met	Glu	Val	Glu	Ile	Trp	Thr	Arg	Glu	Gly	Glu	Lys	Lys	Lys
	2050						2055					2060			
Leu	Arg	Pro	Arg	Trp	Leu	Asp	Ala	Arg	Val	Tyr	Ala	Asp	Pro	Met	Ala
	2065					2070				2075					2080
Leu	Lys	Asp	Phe	Lys	Glu	Phe	Ala	Ser	Gly	Arg	Lys	Ser	Ile	Thr	Leu
				2085						2090					2095
Asp	Ile	Leu	Thr	Glu	Ile	Ala	Ser	Leu	Pro	Thr	Tyr	Leu	Ser	Ser	Arg
				2100						2105					2110
Ala	Lys	Leu	Ala	Leu	Asp	Asn	Ile	Val	Met	Leu	His	Thr	Thr	Glu	Arg
				2115						2120					2125
Gly	Gly	Arg	Ala	Tyr	Gln	His	Ala	Leu	Asn	Glu	Leu	Pro	Glu	Ser	Leu
	2130						2135						2140		
Glu	Thr	Leu	Met	Leu	Val	Ala	Leu	Leu	Gly	Ala	Met	Thr	Ala	Gly	Ile
	2145						2150			2155					2160
Phe	Leu	Phe	Phe	Met	Gln	Gly	Lys	Gly	Ile	Gly	Lys	Leu	Ser	Met	Gly
				2165						2170					2175
Leu	Ile	Thr	Ile	Ala	Val	Ala	Ser	Gly	Leu	Leu	Trp	Val	Ala	Glu	Ile
				2180						2185					2190
Gln	Pro	Gln	Trp	Ile	Ala	Ala	Ser	Ile	Ile	Leu	Glu	Phe	Phe	Leu	Met
				2195						2200					2205
Val	Leu	Leu	Ile	Pro	Glu	Pro	Glu	Lys	Gln	Arg	Thr	Pro	Gln	Asp	Asn
	2210						2215								2220
Gln	Leu	Ile	Tyr	Val	Ile	Leu	Thr	Ile	Leu	Thr	Ile	Ile	Gly	Leu	Ile
	2225						2230			2235					2240
Ala	Ala	Asn	Glu	Met	Gly	Leu	Ile	Glu	Lys	Thr	Lys	Thr	Asp	Phe	Gly
				2245						2250					2255
Phe	Tyr	Gln	Val	Lys	Thr	Glu	Thr	Thr	Ile	Leu	Asp	Val	Asp	Leu	Arg
				2260						2265					2270
Pro	Ala	Ser	Ala	Trp	Thr	Leu	Tyr	Ala	Val	Ala	Thr	Thr	Ile	Leu	Thr
				2275						2280					2285
Pro	Met	Leu	Arg	His	Thr	Ile	Glu	Asn	Thr	Ser	Ala	Asn	Leu	Ser	Leu
	2290						2295						2300		
Ala	Ala	Ile	Ala	Asn	Gln	Ala	Ala	Val	Leu	Met	Gly	Leu	Gly	Lys	Gly
	2305						2310						2315		2320
Trp	Pro	Leu	His	Arg	Met	Asp	Leu	Gly	Val	Pro	Leu	Leu	Ala	Met	Gly
				2325						2330					2335
Cys	Tyr	Ser	Gln	Val	Asn	Pro	Thr	Thr	Leu	Thr	Ala	Ser	Leu	Val	Met
				2340						2345					2350
Leu	Leu	Val	His	Tyr	Ala	Ile	Ile	Gly	Pro	Gly	Leu	Gln	Ala	Lys	Ala
				2355						2360					2365
Thr	Arg	Glu	Ala	Gln	Lys	Arg	Thr	Ala	Ala	Gly	Ile	Met	Lys	Asn	Pro
	2370						2375						2380		
Thr	Val	Asp	Gly	Ile	Thr	Val	Ile	Asp	Leu	Glu	Pro	Ile	Ser	Tyr	Asp
	2385						2390			2395					2400
Pro	Lys	Phe	Glu	Lys	Gln	Leu	Gly	Gln	Val	Met	Leu	Leu	Val	Leu	Cys
				2405						2410					2415
Ala	Gly	Gln	Leu	Leu	Leu	Met	Arg	Thr	Thr	Trp	Ala	Phe	Cys	Glu	Val
				2420						2425					2430
Leu	Thr	Leu	Ala	Thr	Gly	Pro	Ile	Leu	Thr	Leu	Trp	Glu	Gly	Asn	Pro
				2435						2440					2445

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Gly Arg Phe Trp Asn Thr Thr Ile Ala Val Ser Thr Ala Asn Ile Phe  
 2450 2455 2460

Arg Gly Ser Tyr Leu Ala Gly Ala Gly Leu Ala Phe Ser Leu Ile Lys  
 2465 2470 2475 2480

Asn Ala Gln Thr Pro Arg Arg Gly Thr Gly Thr Thr Gly Glu Thr Leu  
 2485 2490 2495

Gly Glu Lys Trp Lys Arg Gln Leu Asn Ser Leu Asp Arg Lys Glu Phe  
 2500 2505 2510

Glu Glu Tyr Lys Arg Ser Gly Ile Leu Glu Val Asp Arg Thr Glu Ala  
 2515 2520 2525

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Tyr Met Ala Thr Leu Lys Asn Val Thr Glu Val Lys Gly Tyr Thr Lys  
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Gly Gly Pro Gly His Glu Glu Pro Ile Pro Met Ala Thr Tyr Gly Trp  
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Asn Leu Val Lys Leu His Ser Gly Val Asp Val Phe Tyr Lys Pro Thr  
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Glu Gln Val Asp Thr Leu Leu Cys Asp Ile Gly Glu Ser Ser Ser Asn  
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Pro Thr Ile Glu Glu Gly Arg Thr Leu Arg Val Leu Lys Met Val Glu  
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Pro Trp Leu Ser Ser Lys Pro Glu Phe Cys Ile Lys Val Leu Asn Pro  
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Tyr Met Pro Thr Val Ile Glu Glu Leu Glu Lys Leu Gln Arg Lys His  
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Gly Gly Asn Leu Val Arg Cys Pro Leu Ser Arg Asn Ser Thr His Glu  
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Met Tyr Trp Val Ser Gly Ala Ser Gly Asn Ile Val Ser Ser Val Asn  
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Thr Thr Ser Lys Met Leu Leu Asn Arg Phe Thr Thr Arg His Arg Lys  
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Ser Thr Glu Thr Glu Lys Pro Asp Met Thr Ile Ile Gly Arg Arg Leu  
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Gln Arg Leu Gln Glu Glu His Lys Glu Thr Trp His Tyr Asp Gln Glu  
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Asn Pro Tyr Arg Thr Trp Ala Tyr His Gly Ser Tyr Glu Ala Pro Ser  
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Arg Thr Pro Gln Pro Lys Pro Gly Thr Arg Met Val Met Thr Thr Thr  
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Gly	Ala	Val	Phe	Gln	Glu	Glu	Gln	Gly	Trp	Thr	Ser	Ala	Ser	Glu	Ala	2900	2905	2910	
Val	Asn	Asp	Ser	Arg	Phe	Trp	Glu	Leu	Val	Asp	Lys	Glu	Arg	Ala	Leu	2915	2920	2925	
His	Gln	Glu	Gly	Lys	Cys	Glu	Ser	Cys	Val	Tyr	Asn	Met	Met	Gly	Lys	2930	2935	2940	
Arg	Glu	Lys	Lys	Leu	Gly	Glu	Phe	Gly	Arg	Ala	Lys	Gly	Ser	Arg	Ala	2945	2950	2955	2960
Ile	Trp	Tyr	Met	Trp	Leu	Gly	Ala	Arg	Phe	Leu	Glu	Phe	Glu	Ala	Leu	2965	2970	2975	
Gly	Phe	Leu	Asn	Glu	Asp	His	Trp	Phe	Gly	Arg	Glu	Asn	Ser	Trp	Ser	2980	2985	2990	
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Thr	Glu	Gln	Met	Ala	Pro	His	His	Lys	Ile	Leu	Ala	Lys	Ala	Ile	Phe	3045	3050	3055	
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Met	Ala	Ile	Cys	Ser	Ala	Val	Pro	Thr	Glu	Trp	Phe	Pro	Thr	Ser	Arg	3265	3270	3275	3280
Thr	Thr	Trp	Ser	Ile	His	Ala	His	His	Gln	Trp	Met	Thr	Thr	Glu	Asp				

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Thr	Asp Lys Thr Pro Val His Ser Trp Glu Asp Ile Pro Tyr Leu Gly				
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Lys	Arg Glu Asp Leu Trp Cys Gly Ser Leu Ile Gly Leu Ser Ser Arg				
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Ala	Thr Trp Ala Lys Asn Ile His Thr Ala Ile Thr Gln Val Arg Asn				
	3345		3350		3355
Leu	Ile Gly Lys Glu Glu Tyr Val Asp Tyr Met Pro Val Met Lys Arg				
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Tyr	Ser Ala Pro Ser Glu Ser Glu Gly Val Leu				
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&lt;210&gt; SEQ ID NO 14

&lt;211&gt; LENGTH: 10649

&lt;212&gt; TYPE: DNA

&lt;213&gt; ORGANISM: Dengue 4 virus strain 2A

&lt;400&gt; SEQUENCE: 14

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aggagataga caagaaggat ggagacctaa tgtatgctga tgacacagca ggctgggaca 9180
caagaatcac tgaggatgac cttcaaaatg aggaactgat cacggaacag atggctcccc 9240
accacaagat cctagccaaa gccattttca aactaaccta tcaaaacaaa gtggtgaaag 9300
tcctcagacc cacaccgaga ggagcgggta tggatatcat atccaggaaa gaccaaagag 9360
gtagtggaca agttggaaca tatggtttga acacattcac caacatgga gttcaactca 9420
tccgccaat ggaagctgaa ggagtcatca cacaagatga catgcagaac ccaaaaggg 9480
tgaagaaaag agttgagaaa tggctgaaag agtgtggtgt cgacaggtta aagaggatgg 9540
caatcagtg agacgattgc gtggtgaagc ccctagatga gaggtttggc acttcctcc 9600
tcttcttga cgacatggga aaggtgagga aagacattcc gcagtggga ccatctaagg 9660
gatggaaaaa ctggcaagag gttccttttt gctcccacca ctttcacaag atctttatga 9720
aggatggccg ctcactagt gttccatgta gaaaccagga tgaactgata gggagagcca 9780
gaatctcgca gggagctgga tggagcttaa gagaacagc ctgectgggc aaagcttacg 9840
cccagatgtg gtcgcttatg tacttccaca gaagggatct gcgtttagcc tccatggcca 9900
tatgctcagc agttccaacg gaatggtttc caacaagcag aacaacatgg tcaatccacg 9960
ctcatcacca gtggatgacc actgaagata tgctcaaagt gtggaacaga gtgtggatag 10020
aagacaacc taatatgact gacaagactc cagtccattc gtgggaagat ataccttacc 10080
tagggaaaag agaggatttg tgggtgggat ccctgattgg actttcttcc agagccacct 10140
ggcgaagaa cattcacacg gccataacc aggtcaggaa cctgatcgga aaagaggaat 10200
acgtggatta catgccagta atgaaaagat acagtgctcc ttcagagagt gaaggagttc 10260
tgtaattacc aacaacaaac accaaaggct attgaagtca ggccacttgt gccacggttt 10320
gagcaaacg tgctgctgt agctccgcca ataatgggag gcgtaataat cccagggag 10380
gccatgcgcc acggaagctg tacgcgtggc atattggact agcggttaga ggagaccct 10440
cccatcactg acaaacgca gcaaaagggg gccgaagcc aggaggaagc tgtactcctg 10500
gtggaaggac tagaggttag aggagacccc cccaacacaa aaacagcata ttgacgctgg 10560
gaaagaccag agatcctgct gtctctgcaa catcaatcca ggcacagagc gccgcaagat 10620
ggattggtgt tgttgatcca acaggttct 10649

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&lt;210&gt; SEQ ID NO 15

&lt;211&gt; LENGTH: 3387

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Recombinant Dengue 4 virus strain rDEN4

&lt;400&gt; SEQUENCE: 15

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Met Asn Gln Arg Lys Lys Val Val Arg Pro Pro Phe Asn Met Leu Lys
1           5           10           15

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Arg	Glu	Arg	Asn	Arg	Val	Ser	Thr	Pro	Gln	Gly	Leu	Val	Lys	Arg	Phe
			20					25					30		
Ser	Thr	Gly	Leu	Phe	Ser	Gly	Lys	Gly	Pro	Leu	Arg	Met	Val	Leu	Ala
		35					40					45			
Phe	Ile	Thr	Phe	Leu	Arg	Val	Leu	Ser	Ile	Pro	Pro	Thr	Ala	Gly	Ile
	50					55					60				
Leu	Lys	Arg	Trp	Gly	Gln	Leu	Lys	Lys	Asn	Lys	Ala	Ile	Lys	Ile	Leu
65					70					75					80
Ile	Gly	Phe	Arg	Lys	Glu	Ile	Gly	Arg	Met	Leu	Asn	Ile	Leu	Asn	Gly
				85					90					95	
Arg	Lys	Arg	Ser	Thr	Ile	Thr	Leu	Leu	Cys	Leu	Ile	Pro	Thr	Val	Met
			100					105					110		
Ala	Phe	Ser	Leu	Ser	Thr	Arg	Asp	Gly	Glu	Pro	Leu	Met	Ile	Val	Ala
		115					120					125			
Lys	His	Glu	Arg	Gly	Arg	Pro	Leu	Leu	Phe	Lys	Thr	Thr	Glu	Gly	Ile
	130					135					140				
Asn	Lys	Cys	Thr	Leu	Ile	Ala	Met	Asp	Leu	Gly	Glu	Met	Cys	Glu	Asp
145					150					155					160
Thr	Val	Thr	Tyr	Lys	Cys	Pro	Leu	Leu	Val	Asn	Thr	Glu	Pro	Glu	Asp
				165						170				175	
Ile	Asp	Cys	Trp	Cys	Asn	Leu	Thr	Ser	Thr	Trp	Val	Met	Tyr	Gly	Thr
			180					185					190		
Cys	Thr	Gln	Ser	Gly	Glu	Arg	Arg	Arg	Glu	Lys	Arg	Ser	Val	Ala	Leu
		195					200					205			
Thr	Pro	His	Ser	Gly	Met	Gly	Leu	Glu	Thr	Arg	Ala	Glu	Thr	Trp	Met
		210				215					220				
Ser	Ser	Glu	Gly	Ala	Trp	Lys	His	Ala	Gln	Arg	Val	Glu	Ser	Trp	Ile
225					230					235					240
Leu	Arg	Asn	Pro	Gly	Phe	Ala	Leu	Leu	Ala	Gly	Phe	Met	Ala	Tyr	Met
				245					250					255	
Ile	Gly	Gln	Thr	Gly	Ile	Gln	Arg	Thr	Val	Phe	Phe	Val	Leu	Met	Met
			260					265					270		
Leu	Val	Ala	Pro	Ser	Tyr	Gly	Met	Arg	Cys	Val	Gly	Val	Gly	Asn	Arg
		275					280					285			
Asp	Phe	Val	Glu	Gly	Val	Ser	Gly	Gly	Ala	Trp	Val	Asp	Leu	Val	Leu
	290					295					300				
Glu	His	Gly	Gly	Cys	Val	Thr	Thr	Met	Ala	Gln	Gly	Lys	Pro	Thr	Leu
305					310					315					320
Asp	Phe	Glu	Leu	Thr	Lys	Thr	Thr	Ala	Lys	Glu	Val	Ala	Leu	Leu	Arg
				325					330					335	
Thr	Tyr	Cys	Ile	Glu	Ala	Ser	Ile	Ser	Asn	Ile	Thr	Thr	Ala	Thr	Arg
			340					345					350		
Cys	Pro	Thr	Gln	Gly	Glu	Pro	Tyr	Leu	Lys	Glu	Glu	Gln	Asp	Gln	Gln
		355					360					365			
Tyr	Ile	Cys	Arg	Arg	Asp	Val	Val	Asp	Arg	Gly	Trp	Gly	Asn	Gly	Cys
	370					375					380				
Gly	Leu	Phe	Gly	Lys	Gly	Gly	Val	Val	Thr	Cys	Ala	Lys	Phe	Ser	Cys
385					390					395					400
Ser	Gly	Lys	Ile	Thr	Gly	Asn	Leu	Val	Gln	Ile	Glu	Asn	Leu	Glu	Tyr
				405					410					415	
Thr	Val	Val	Val	Thr	Val	His	Asn	Gly	Asp	Thr	His	Ala	Val	Gly	Asn
			420					425					430		
Asp	Thr	Ser	Asn	His	Gly	Val	Thr	Ala	Met	Ile	Thr	Pro	Arg	Ser	Pro

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435	440	445
Ser Val Glu Val Lys Leu Pro Asp Tyr Gly Glu Leu Thr Leu Asp Cys 450 455 460		
Glu Pro Arg Ser Gly Ile Asp Phe Asn Glu Met Ile Leu Met Lys Met 465 470 475 480		
Lys Lys Lys Thr Trp Leu Val His Lys Gln Trp Phe Leu Asp Leu Pro 485 490 495		
Leu Pro Trp Thr Ala Gly Ala Asp Thr Ser Glu Val His Trp Asn Tyr 500 505 510		
Lys Glu Arg Met Val Thr Phe Lys Val Pro His Ala Lys Arg Gln Asp 515 520 525		
Val Thr Val Leu Gly Ser Gln Glu Gly Ala Met His Ser Ala Leu Ala 530 535 540		
Gly Ala Thr Glu Val Asp Ser Gly Asp Gly Asn His Met Phe Ala Gly 545 550 555 560		
His Leu Lys Cys Lys Val Arg Met Glu Lys Leu Arg Ile Lys Gly Met 565 570 575		
Ser Tyr Thr Met Cys Ser Gly Lys Phe Ser Ile Asp Lys Glu Met Ala 580 585 590		
Glu Thr Gln His Gly Thr Thr Val Val Lys Val Lys Tyr Glu Gly Ala 595 600 605		
Gly Ala Pro Cys Lys Val Pro Ile Glu Ile Arg Asp Val Asn Lys Glu 610 615 620		
Lys Val Val Gly Arg Ile Ile Ser Ser Thr Pro Leu Ala Glu Asn Thr 625 630 635 640		
Asn Ser Val Thr Asn Ile Glu Leu Glu Pro Pro Phe Gly Asp Ser Tyr 645 650 655		
Ile Val Ile Gly Val Gly Asn Ser Ala Leu Thr Leu His Trp Phe Arg 660 665 670		
Lys Gly Ser Ser Ile Gly Lys Met Phe Glu Ser Thr Tyr Arg Gly Ala 675 680 685		
Lys Arg Met Ala Ile Leu Gly Glu Thr Ala Trp Asp Phe Gly Ser Val 690 695 700		
Gly Gly Leu Phe Thr Ser Leu Gly Lys Ala Val His Gln Val Phe Gly 705 710 715 720		
Ser Val Tyr Thr Thr Met Phe Gly Gly Val Ser Trp Met Ile Arg Ile 725 730 735		
Leu Ile Gly Phe Leu Val Leu Trp Ile Gly Thr Asn Ser Arg Asn Thr 740 745 750		
Ser Met Ala Met Thr Cys Ile Ala Val Gly Gly Ile Thr Leu Phe Leu 755 760 765		
Gly Phe Thr Val Gln Ala Asp Met Gly Cys Val Ala Ser Trp Ser Gly 770 775 780		
Lys Glu Leu Lys Cys Gly Ser Gly Ile Phe Val Val Asp Asn Val His 785 790 795 800		
Thr Trp Thr Glu Gln Tyr Lys Phe Gln Pro Glu Ser Pro Ala Arg Leu 805 810 815		
Ala Ser Ala Ile Leu Asn Ala His Lys Asp Gly Val Cys Gly Ile Arg 820 825 830		
Ser Thr Thr Arg Leu Glu Asn Val Met Trp Lys Gln Ile Thr Asn Glu 835 840 845		
Leu Asn Tyr Val Leu Trp Glu Gly Gly His Asp Leu Thr Val Val Ala 850 855 860		



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Gly Asp Val Lys Gly Val Leu Thr Lys Gly Lys Arg Ala Leu Thr Pro  
 865 870 875 880  
 Pro Val Ser Asp Leu Lys Tyr Ser Trp Lys Thr Trp Gly Lys Ala Lys  
 885 890 895  
 Ile Phe Thr Pro Glu Ala Arg Asn Ser Thr Phe Leu Ile Asp Gly Pro  
 900 905 910  
 Asp Thr Ser Glu Cys Pro Asn Glu Arg Arg Ala Trp Asn Ser Leu Glu  
 915 920 925  
 Val Glu Asp Tyr Gly Phe Gly Met Phe Thr Thr Asn Ile Trp Met Lys  
 930 935 940  
 Phe Arg Glu Gly Ser Ser Glu Val Cys Asp His Arg Leu Met Ser Ala  
 945 950 955 960  
 Ala Ile Lys Asp Gln Lys Ala Val His Ala Asp Met Gly Tyr Trp Ile  
 965 970 975  
 Glu Ser Ser Lys Asn Gln Thr Trp Gln Ile Glu Lys Ala Ser Leu Ile  
 980 985 990  
 Glu Val Lys Thr Cys Leu Trp Pro Lys Thr His Thr Leu Trp Ser Asn  
 995 1000 1005  
 Gly Val Leu Glu Ser Gln Met Leu Ile Pro Lys Ser Tyr Ala Gly Pro  
 1010 1015 1020  
 Phe Ser Gln His Asn Tyr Arg Gln Gly Tyr Ala Thr Gln Thr Val Gly  
 1025 1030 1035 1040  
 Pro Trp His Leu Gly Lys Leu Glu Ile Asp Phe Gly Glu Cys Pro Gly  
 1045 1050 1055  
 Thr Thr Val Thr Ile Gln Glu Asp Cys Asp His Arg Gly Pro Ser Leu  
 1060 1065 1070  
 Arg Thr Thr Thr Ala Ser Gly Lys Leu Val Thr Gln Trp Cys Cys Arg  
 1075 1080 1085  
 Ser Cys Thr Met Pro Pro Leu Arg Phe Leu Gly Glu Asp Gly Cys Trp  
 1090 1095 1100  
 Tyr Gly Met Glu Ile Arg Pro Leu Ser Glu Lys Glu Glu Asn Met Val  
 1105 1110 1115 1120  
 Lys Ser Gln Val Thr Ala Gly Gln Gly Thr Ser Glu Thr Phe Ser Met  
 1125 1130 1135  
 Gly Leu Leu Cys Leu Thr Leu Phe Val Glu Glu Cys Leu Arg Arg Arg  
 1140 1145 1150  
 Val Thr Arg Lys His Met Ile Leu Val Val Val Ile Thr Leu Cys Ala  
 1155 1160 1165  
 Ile Ile Leu Gly Gly Leu Thr Trp Met Asp Leu Leu Arg Ala Leu Ile  
 1170 1175 1180  
 Met Leu Gly Asp Thr Met Ser Gly Arg Ile Gly Gly Gln Ile His Leu  
 1185 1190 1195 1200  
 Ala Ile Met Ala Val Phe Lys Met Ser Pro Gly Tyr Val Leu Gly Val  
 1205 1210 1215  
 Phe Leu Arg Lys Leu Thr Ser Arg Glu Thr Ala Leu Met Val Ile Gly  
 1220 1225 1230  
 Met Ala Met Thr Thr Val Leu Ser Ile Pro His Asp Leu Met Glu Leu  
 1235 1240 1245  
 Ile Asp Gly Ile Ser Leu Gly Leu Ile Leu Leu Lys Ile Val Thr Gln  
 1250 1255 1260  
 Phe Asp Asn Thr Gln Val Gly Thr Leu Ala Leu Ser Leu Thr Phe Ile  
 1265 1270 1275 1280

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Arg	Ser	Thr	Met	Pro	Leu	Val	Met	Ala	Trp	Arg	Thr	Ile	Met	Ala	Val	1285	1290	1295	
Leu	Phe	Val	Val	Thr	Leu	Ile	Pro	Leu	Cys	Arg	Thr	Ser	Cys	Leu	Gln	1300	1305	1310	
Lys	Gln	Ser	His	Trp	Val	Glu	Ile	Thr	Ala	Leu	Ile	Leu	Gly	Ala	Gln	1315	1320	1325	
Ala	Leu	Pro	Val	Tyr	Leu	Met	Thr	Leu	Met	Lys	Gly	Ala	Ser	Arg	Arg	1330	1335	1340	
Ser	Trp	Pro	Leu	Asn	Glu	Gly	Ile	Met	Ala	Val	Gly	Leu	Val	Ser	Leu	1345	1350	1355	1360
Leu	Gly	Ser	Ala	Leu	Leu	Lys	Asn	Asp	Val	Pro	Leu	Ala	Gly	Pro	Met	1365	1370	1375	
Val	Ala	Gly	Gly	Leu	Leu	Leu	Ala	Ala	Tyr	Val	Met	Ser	Gly	Ser	Ser	1380	1385	1390	
Ala	Asp	Leu	Ser	Leu	Glu	Lys	Ala	Ala	Asn	Val	Gln	Trp	Asp	Glu	Met	1395	1400	1405	
Ala	Asp	Ile	Thr	Gly	Ser	Ser	Pro	Ile	Val	Glu	Val	Lys	Gln	Asp	Glu	1410	1415	1420	
Asp	Gly	Ser	Phe	Ser	Ile	Arg	Asp	Val	Glu	Glu	Thr	Asn	Met	Ile	Thr	1425	1430	1435	1440
Leu	Leu	Val	Lys	Leu	Ala	Leu	Ile	Thr	Val	Ser	Gly	Leu	Tyr	Pro	Leu	1445	1450	1455	
Ala	Ile	Pro	Val	Thr	Met	Thr	Leu	Trp	Tyr	Met	Trp	Gln	Val	Lys	Thr	1460	1465	1470	
Gln	Arg	Ser	Gly	Ala	Leu	Trp	Asp	Val	Pro	Ser	Pro	Ala	Ala	Thr	Lys	1475	1480	1485	
Lys	Ala	Ala	Leu	Ser	Glu	Gly	Val	Tyr	Arg	Ile	Met	Gln	Arg	Gly	Leu	1490	1495	1500	
Phe	Gly	Lys	Thr	Gln	Val	Gly	Val	Gly	Ile	His	Met	Glu	Gly	Val	Phe	1505	1510	1515	1520
His	Thr	Met	Trp	His	Val	Thr	Arg	Gly	Ser	Val	Ile	Cys	His	Glu	Thr	1525	1530	1535	
Gly	Arg	Leu	Glu	Pro	Ser	Trp	Ala	Asp	Val	Arg	Asn	Asp	Met	Ile	Ser	1540	1545	1550	
Tyr	Gly	Gly	Gly	Trp	Arg	Leu	Gly	Asp	Lys	Trp	Asp	Lys	Glu	Glu	Asp	1555	1560	1565	
Val	Gln	Val	Leu	Ala	Ile	Glu	Pro	Gly	Lys	Asn	Pro	Lys	His	Val	Gln	1570	1575	1580	
Thr	Lys	Pro	Gly	Leu	Phe	Lys	Thr	Leu	Thr	Gly	Glu	Ile	Gly	Ala	Val	1585	1590	1595	1600
Thr	Leu	Asp	Phe	Lys	Pro	Gly	Thr	Ser	Gly	Ser	Pro	Ile	Ile	Asn	Arg	1605	1610	1615	
Lys	Gly	Lys	Val	Ile	Gly	Leu	Tyr	Gly	Asn	Gly	Val	Val	Thr	Lys	Ser	1620	1625	1630	
Gly	Asp	Tyr	Val	Ser	Ala	Ile	Thr	Gln	Ala	Glu	Arg	Ile	Gly	Glu	Pro	1635	1640	1645	
Asp	Tyr	Glu	Val	Asp	Glu	Asp	Ile	Phe	Arg	Lys	Lys	Arg	Leu	Thr	Ile	1650	1655	1660	
Met	Asp	Leu	His	Pro	Gly	Ala	Gly	Lys	Thr	Lys	Arg	Ile	Leu	Pro	Ser	1665	1670	1675	1680
Ile	Val	Arg	Glu	Ala	Leu	Lys	Arg	Arg	Leu	Arg	Thr	Leu	Ile	Leu	Ala	1685	1690	1695	
Pro	Thr	Arg	Val	Val	Ala	Ala	Glu	Met	Glu	Glu	Ala	Leu	Arg	Gly	Leu				

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1700	1705	1710
Pro Ile Arg Tyr Gln Thr 1715	Pro Ala Val Lys Ser 1720	Glu His Thr Gly Arg 1725
Glu Ile Val Asp Leu Met 1730	Cys His Ala Thr Phe 1735	Thr Thr Arg Leu Leu 1740
Ser Ser Thr Arg Val 1745	Pro Asn Tyr Asn Leu 1750	Ile Val Met Asp Glu Ala 1755 1760
His Phe Thr Asp 1765	Pro Ser Ser Val Ala 1770	Ala Arg Gly Tyr Ile Ser Thr 1775
Arg Val Glu Met Gly 1780	Glu Ala Ala Ala Ile 1785	Phe Met Thr Ala Thr Pro 1790
Pro Gly Ala Thr Asp 1795	Pro Phe Pro Gln Ser 1800	Asn Ser Pro Ile Glu Asp 1805
Ile Glu Arg Glu Ile 1810	Pro Glu Arg Ser Trp 1815	Asn Thr Gly Phe Asp Trp 1820
Ile Thr Asp Tyr Gln 1825	Gly Lys Thr Val Trp 1830	Phe Val Pro Ser Ile Lys 1835 1840
Ala Gly Asn Asp Ile 1845	Ala Asn Cys Leu Arg 1850	Lys Ser Gly Lys Lys Val 1855
Ile Gln Leu Ser Arg 1860	Lys Thr Phe Asp Thr 1865	Glu Tyr Pro Lys Thr Lys 1870
Leu Thr Asp Trp Asp 1875	Phe Val Val Thr Thr 1880	Asp Ile Ser Glu Met Gly 1885
Ala Asn Phe Arg Ala 1890	Gly Arg Val Ile Asp 1895	Pro Arg Arg Cys Leu Lys 1900
Pro Val Ile Leu Pro 1905	Asp Gly Pro Glu Arg 1910	Val Ile Leu Ala Gly Pro 1915 1920
Ile Pro Val Thr 1925	Pro Ala Ser Ala Ala 1930	Gln Arg Arg Gly Arg Ile Gly 1935
Arg Asn Pro Ala Gln 1940	Glu Asp Asp Gln Tyr 1945	Val Phe Ser Gly Asp Pro 1950
Leu Lys Asn Asp Glu 1955	Asp His Ala His Trp 1960	Thr Glu Ala Lys Met Leu 1965
Leu Asp Asn Ile Tyr 1970	Thr Pro Glu Gly Ile 1975	Ile Ile Pro Thr Leu Phe Gly 1980
Pro Glu Arg Glu Lys 1985	Thr Gln Ala Ile Asp 1990	Gly Glu Phe Arg Leu Arg 1995 2000
Gly Glu Gln Arg Lys 2005	Thr Phe Val Glu Leu 2010	Met Arg Arg Gly Asp Leu 2015
Pro Val Trp Leu Ser 2020	Tyr Lys Val Ala Ser 2025	Ala Gly Ile Ser Tyr Glu 2030
Asp Arg Glu Trp Cys 2035	Phe Thr Gly Glu Arg 2040	Asn Asn Gln Ile Leu Glu 2045
Glu Asn Met Glu Val 2050	Glu Ile Trp Thr Arg 2055	Glu Gly Glu Lys Lys Lys 2060
Leu Arg Pro Arg Trp 2065	Leu Asp Ala Arg Val 2070	Tyr Ala Asp Pro Met Ala 2075 2080
Leu Lys Asp Phe Lys 2085	Glu Phe Ala Ser Gly 2090	Arg Lys Ser Ile Thr Leu 2095
Asp Ile Leu Thr Glu 2100	Ile Ala Ser Leu Pro 2105	Thr Tyr Leu Ser Ser Arg 2110
Ala Lys Leu Ala Leu 2115	Asp Asn Ile Val Met 2120	Leu His Thr Thr Glu Arg 2125

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Gly Gly Arg Ala Tyr Gln His Ala Leu Asn Glu Leu Pro Glu Ser Leu  
 2130 2135 2140

Glu Thr Leu Met Leu Val Ala Leu Leu Gly Ala Met Thr Ala Gly Ile  
 2145 2150 2155 2160

Phe Leu Phe Phe Met Gln Gly Lys Gly Ile Gly Lys Leu Ser Met Gly  
 2165 2170 2175

Leu Ile Thr Ile Ala Val Ala Ser Gly Leu Leu Trp Val Ala Glu Ile  
 2180 2185 2190

Gln Pro Gln Trp Ile Ala Ala Ser Ile Ile Leu Glu Phe Phe Leu Met  
 2195 2200 2205

Val Leu Leu Ile Pro Glu Pro Glu Lys Gln Arg Thr Pro Gln Asp Asn  
 2210 2215 2220

Gln Leu Ile Tyr Val Ile Leu Thr Ile Leu Thr Ile Ile Gly Leu Ile  
 2225 2230 2235 2240

Ala Ala Asn Glu Met Gly Leu Ile Glu Lys Thr Lys Thr Asp Phe Gly  
 2245 2250 2255

Phe Tyr Gln Val Lys Thr Glu Thr Thr Ile Leu Asp Val Asp Leu Arg  
 2260 2265 2270

Pro Ala Ser Ala Trp Thr Leu Tyr Ala Val Ala Thr Thr Ile Leu Thr  
 2275 2280 2285

Pro Met Leu Arg His Thr Ile Glu Asn Thr Ser Ala Asn Leu Ser Leu  
 2290 2295 2300

Ala Ala Ile Ala Asn Gln Ala Ala Val Leu Met Gly Leu Gly Lys Gly  
 2305 2310 2315 2320

Trp Pro Leu His Arg Met Asp Leu Gly Val Pro Leu Leu Ala Met Gly  
 2325 2330 2335

Cys Tyr Ser Gln Val Asn Pro Thr Thr Leu Thr Ala Ser Leu Val Met  
 2340 2345 2350

Leu Leu Val His Tyr Ala Ile Ile Gly Pro Gly Leu Gln Ala Lys Ala  
 2355 2360 2365

Thr Arg Glu Ala Gln Lys Arg Thr Ala Ala Gly Ile Met Lys Asn Pro  
 2370 2375 2380

Thr Val Asp Gly Ile Thr Val Ile Asp Leu Glu Pro Ile Ser Tyr Asp  
 2385 2390 2395 2400

Pro Lys Phe Glu Lys Gln Leu Gly Gln Val Met Leu Leu Val Leu Cys  
 2405 2410 2415

Ala Gly Gln Leu Leu Leu Met Arg Thr Thr Trp Ala Phe Cys Glu Val  
 2420 2425 2430

Leu Thr Leu Ala Thr Gly Pro Ile Leu Thr Leu Trp Glu Gly Asn Pro  
 2435 2440 2445

Gly Arg Phe Trp Asn Thr Thr Ile Ala Val Ser Thr Ala Asn Ile Phe  
 2450 2455 2460

Arg Gly Ser Tyr Leu Ala Gly Ala Gly Leu Ala Phe Ser Leu Ile Lys  
 2465 2470 2475 2480

Asn Ala Gln Thr Pro Arg Arg Gly Thr Gly Thr Thr Gly Glu Thr Leu  
 2485 2490 2495

Gly Glu Lys Trp Lys Arg Gln Leu Asn Ser Leu Asp Arg Lys Glu Phe  
 2500 2505 2510

Glu Glu Tyr Lys Arg Ser Gly Ile Leu Glu Val Asp Arg Thr Glu Ala  
 2515 2520 2525

Lys Ser Ala Leu Lys Asp Gly Ser Lys Ile Lys His Ala Val Ser Arg  
 2530 2535 2540

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Gly Ser Ser Lys Ile Arg Trp Ile Val Glu Arg Gly Met Val Lys Pro  
 2545 2550 2555 2560  
 Lys Gly Lys Val Val Asp Leu Gly Cys Gly Arg Gly Gly Trp Ser Tyr  
 2565 2570 2575  
 Tyr Met Ala Thr Leu Lys Asn Val Thr Glu Val Lys Gly Tyr Thr Lys  
 2580 2585 2590  
 Gly Gly Pro Gly His Glu Glu Pro Ile Pro Met Ala Thr Tyr Gly Trp  
 2595 2600 2605  
 Asn Leu Val Lys Leu His Ser Gly Val Asp Val Phe Tyr Lys Pro Thr  
 2610 2615 2620  
 Glu Gln Val Asp Thr Leu Leu Cys Asp Ile Gly Glu Ser Ser Ser Asn  
 2625 2630 2635 2640  
 Pro Thr Ile Glu Glu Gly Arg Thr Leu Arg Val Leu Lys Met Val Glu  
 2645 2650 2655  
 Pro Trp Leu Ser Ser Lys Pro Glu Phe Cys Ile Lys Val Leu Asn Pro  
 2660 2665 2670  
 Tyr Met Pro Thr Val Ile Glu Glu Leu Glu Lys Leu Gln Arg Lys His  
 2675 2680 2685  
 Gly Gly Asn Leu Val Arg Cys Pro Leu Ser Arg Asn Ser Thr His Glu  
 2690 2695 2700  
 Met Tyr Trp Val Ser Gly Ala Ser Gly Asn Ile Val Ser Ser Val Asn  
 2705 2710 2715 2720  
 Thr Thr Ser Lys Met Leu Leu Asn Arg Phe Thr Thr Arg His Arg Lys  
 2725 2730 2735  
 Pro Thr Tyr Glu Lys Asp Val Asp Leu Gly Ala Gly Thr Arg Ser Val  
 2740 2745 2750  
 Ser Thr Glu Thr Glu Lys Pro Asp Met Thr Ile Ile Gly Arg Arg Leu  
 2755 2760 2765  
 Gln Arg Leu Gln Glu Glu His Lys Glu Thr Trp His Tyr Asp Gln Glu  
 2770 2775 2780  
 Asn Pro Tyr Arg Thr Trp Ala Tyr His Gly Ser Tyr Glu Ala Pro Ser  
 2785 2790 2795 2800  
 Thr Gly Ser Ala Ser Ser Met Val Asn Gly Val Val Lys Leu Leu Thr  
 2805 2810 2815  
 Lys Pro Trp Asp Val Ile Pro Met Val Thr Gln Leu Ala Met Thr Asp  
 2820 2825 2830  
 Thr Thr Pro Phe Gly Gln Gln Arg Val Phe Lys Glu Lys Val Asp Thr  
 2835 2840 2845  
 Arg Thr Pro Gln Pro Lys Pro Gly Thr Arg Met Val Met Thr Thr Thr  
 2850 2855 2860  
 Ala Asn Trp Leu Trp Ala Leu Leu Gly Lys Lys Lys Asn Pro Arg Leu  
 2865 2870 2875 2880  
 Cys Thr Arg Glu Glu Phe Ile Ser Lys Val Arg Ser Asn Ala Ala Ile  
 2885 2890 2895  
 Gly Ala Val Phe Gln Glu Glu Gln Gly Trp Thr Ser Ala Ser Glu Ala  
 2900 2905 2910  
 Val Asn Asp Ser Arg Phe Trp Glu Leu Val Asp Lys Glu Arg Ala Leu  
 2915 2920 2925  
 His Gln Glu Gly Lys Cys Glu Ser Cys Val Tyr Asn Met Met Gly Lys  
 2930 2935 2940  
 Arg Glu Lys Lys Leu Gly Glu Phe Gly Arg Ala Lys Gly Ser Arg Ala  
 2945 2950 2955 2960  
 Ile Trp Tyr Met Trp Leu Gly Ala Arg Phe Leu Glu Phe Glu Ala Leu

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2965					2970					2975					
Gly	Phe	Leu	Asn	Glu	Asp	His	Trp	Phe	Gly	Arg	Glu	Asn	Ser	Trp	Ser
			2980					2985					2990		
Gly	Val	Glu	Gly	Glu	Gly	Leu	His	Arg	Leu	Gly	Tyr	Ile	Leu	Glu	Glu
		2995					3000					3005			
Ile	Asp	Lys	Lys	Asp	Gly	Asp	Leu	Met	Tyr	Ala	Asp	Asp	Thr	Ala	Gly
	3010					3015					3020				
Trp	Asp	Thr	Arg	Ile	Thr	Glu	Asp	Asp	Leu	Gln	Asn	Glu	Glu	Leu	Ile
3025					3030					3035					3040
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ggattggtgt tgttgatcca acaggttct 10649

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&lt;210&gt; SEQ ID NO 17

&lt;211&gt; LENGTH: 3388

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Recombinant Dengue rDEN2/4d30

&lt;400&gt; SEQUENCE: 17

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Met Asn Asn Gln Arg Lys Lys Ala Arg Asn Thr Pro Phe Asn Met Leu
 1             5             10             15
Lys Arg Glu Arg Asn Arg Val Ser Thr Val Gln Gln Leu Thr Lys Arg
          20             25             30
Phe Ser Leu Gly Met Leu Gln Gly Arg Gly Pro Leu Lys Leu Phe Met
          35             40             45
Ala Leu Val Ala Phe Leu Arg Phe Leu Thr Ile Pro Pro Thr Ala Gly
          50             55             60
Ile Leu Lys Arg Trp Gly Thr Ile Lys Lys Ser Lys Ala Ile Asn Val
65             70             75             80
Leu Arg Gly Phe Arg Lys Glu Ile Gly Arg Met Leu Asn Ile Leu Asn
          85             90             95
Arg Arg Arg Arg Thr Ala Gly Met Ile Ile Met Leu Ile Pro Thr Val
          100            105            110
Met Ala Phe His Leu Thr Thr Arg Asn Gly Glu Pro His Met Ile Val

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115					120					125					
Ser	Arg	Gln	Glu	Lys	Gly	Lys	Ser	Leu	Leu	Phe	Lys	Thr	Glu	Asp	Gly
	130					135					140				
Val	Asn	Met	Cys	Thr	Leu	Met	Ala	Met	Asp	Leu	Gly	Glu	Leu	Cys	Glu
145					150					155					160
Asp	Thr	Ile	Thr	Tyr	Lys	Cys	Pro	Leu	Leu	Arg	Gln	Asn	Glu	Pro	Glu
				165					170					175	
Asp	Ile	Asp	Cys	Trp	Cys	Asn	Ser	Thr	Ser	Thr	Trp	Val	Thr	Tyr	Gly
			180					185					190		
Thr	Cys	Thr	Thr	Thr	Gly	Glu	His	Arg	Arg	Glu	Lys	Arg	Ser	Val	Ala
		195					200					205			
Leu	Val	Pro	His	Val	Gly	Met	Gly	Leu	Glu	Thr	Arg	Thr	Glu	Thr	Trp
	210					215					220				
Met	Ser	Ser	Glu	Gly	Ala	Trp	Lys	His	Ala	Gln	Arg	Ile	Glu	Thr	Trp
225					230					235					240
Ile	Leu	Arg	His	Pro	Gly	Phe	Thr	Ile	Met	Ala	Ala	Ile	Leu	Ala	Tyr
				245					250					255	
Thr	Ile	Gly	Thr	Thr	His	Phe	Gln	Arg	Ala	Leu	Ile	Phe	Ile	Leu	Leu
		260						265					270		
Thr	Ala	Val	Ala	Pro	Ser	Met	Thr	Met	Arg	Cys	Ile	Gly	Ile	Ser	Asn
		275					280					285			
Arg	Asp	Phe	Val	Glu	Gly	Val	Ser	Gly	Gly	Ser	Trp	Val	Asp	Ile	Val
	290					295					300				
Leu	Glu	His	Gly	Ser	Cys	Val	Thr	Thr	Met	Ala	Lys	Asn	Lys	Pro	Thr
305					310					315					320
Leu	Asp	Phe	Glu	Leu	Ile	Lys	Thr	Glu	Ala	Lys	Gln	Pro	Ala	Thr	Leu
				325					330					335	
Arg	Lys	Tyr	Cys	Ile	Glu	Ala	Lys	Leu	Thr	Asn	Thr	Thr	Thr	Glu	Ser
			340					345						350	
Arg	Cys	Pro	Thr	Gln	Gly	Glu	Pro	Ser	Leu	Asn	Glu	Glu	Gln	Asp	Lys
		355					360					365			
Arg	Phe	Val	Cys	Lys	His	Ser	Met	Val	Asp	Arg	Gly	Trp	Gly	Asn	Gly
	370					375					380				
Cys	Gly	Leu	Phe	Gly	Lys	Gly	Gly	Ile	Val	Thr	Cys	Ala	Met	Phe	Thr
385					390					395					400
Cys	Lys	Lys	Asn	Met	Glu	Gly	Lys	Val	Val	Gln	Pro	Glu	Asn	Leu	Glu
				405					410					415	
Tyr	Thr	Ile	Val	Ile	Thr	Pro	His	Ser	Gly	Glu	Glu	His	Ala	Val	Gly
			420					425					430		
Asn	Asp	Thr	Gly	Lys	His	Gly	Lys	Glu	Ile	Lys	Ile	Thr	Pro	Gln	Ser
		435					440					445			
Ser	Ile	Thr	Glu	Ala	Glu	Leu	Thr	Gly	Tyr	Gly	Thr	Val	Thr	Met	Glu
	450					455					460				
Cys	Ser	Pro	Arg	Thr	Gly	Leu	Asp	Phe	Asn	Glu	Met	Val	Leu	Leu	Gln
465					470					475					480
Met	Glu	Asn	Lys	Ala	Trp	Leu	Val	His	Arg	Gln	Trp	Phe	Leu	Asp	Leu
				485					490					495	
Pro	Leu	Pro	Trp	Leu	Pro	Gly	Ala	Asp	Thr	Gln	Gly	Ser	Asn	Trp	Ile
			500					505					510		
Gln	Lys	Glu	Thr	Leu	Val	Thr	Phe	Lys	Asn	Pro	His	Ala	Lys	Lys	Gln
		515					520					525			
Asp	Val	Val	Val	Leu	Gly	Ser	Gln	Glu	Gly	Ala	Met	His	Thr	Ala	Leu
	530					535					540				

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Thr Gly Ala Thr Glu Ile Gln Met Ser Ser Gly Asn Leu Leu Phe Thr  
 545 550 555 560  
 Gly His Leu Lys Cys Arg Leu Arg Met Asp Lys Leu Gln Leu Lys Gly  
 565 570 575  
 Met Ser Tyr Ser Met Cys Thr Gly Lys Phe Lys Val Val Lys Glu Ile  
 580 585 590  
 Ala Glu Thr Gln His Gly Thr Ile Val Ile Arg Val Gln Tyr Glu Gly  
 595 600 605  
 Asp Gly Ser Pro Cys Lys Ile Pro Phe Glu Ile Met Asp Leu Glu Lys  
 610 615 620  
 Arg His Val Leu Gly Arg Leu Ile Thr Val Asn Pro Ile Val Thr Glu  
 625 630 635 640  
 Lys Asp Ser Pro Val Asn Ile Glu Ala Glu Pro Pro Phe Gly Asp Ser  
 645 650 655  
 Tyr Ile Ile Ile Gly Val Glu Pro Gly Gln Leu Lys Leu Asn Trp Phe  
 660 665 670  
 Lys Lys Gly Ser Ser Ile Gly Gln Met Phe Glu Thr Thr Met Arg Gly  
 675 680 685  
 Ala Lys Arg Met Ala Ile Leu Gly Asp Thr Ala Trp Asp Phe Gly Ser  
 690 695 700  
 Leu Gly Gly Val Phe Thr Ser Ile Gly Lys Ala Leu His Gln Val Phe  
 705 710 715 720  
 Gly Ala Ile Tyr Gly Ala Ala Phe Ser Gly Val Ser Trp Thr Met Lys  
 725 730 735  
 Ile Leu Ile Gly Val Ile Ile Thr Trp Ile Gly Met Asn Ser Arg Asn  
 740 745 750  
 Thr Ser Met Ala Met Thr Cys Ile Ala Val Gly Gly Ile Thr Leu Phe  
 755 760 765  
 Leu Gly Phe Thr Val Gln Ala Asp Met Gly Cys Val Ala Ser Trp Ser  
 770 775 780  
 Gly Lys Glu Leu Lys Cys Gly Ser Gly Ile Phe Val Val Asp Asn Val  
 785 790 795 800  
 His Thr Trp Thr Glu Gln Tyr Lys Phe Gln Pro Glu Ser Pro Ala Arg  
 805 810 815  
 Leu Ala Ser Ala Ile Leu Asn Ala His Lys Asp Gly Val Cys Gly Ile  
 820 825 830  
 Arg Ser Thr Thr Arg Leu Glu Asn Val Met Trp Lys Gln Ile Thr Asn  
 835 840 845  
 Glu Leu Asn Tyr Val Leu Trp Glu Gly Gly His Asp Leu Thr Val Val  
 850 855 860  
 Ala Gly Asp Val Lys Gly Val Leu Thr Lys Gly Lys Arg Ala Leu Thr  
 865 870 875 880  
 Pro Pro Val Ser Asp Leu Lys Tyr Ser Trp Lys Thr Trp Gly Lys Ala  
 885 890 895  
 Lys Ile Phe Thr Pro Glu Ala Arg Asn Ser Thr Phe Leu Ile Asp Gly  
 900 905 910  
 Pro Asp Thr Ser Glu Cys Pro Asn Glu Arg Arg Ala Trp Asn Ser Leu  
 915 920 925  
 Glu Val Glu Asp Tyr Gly Phe Gly Met Phe Thr Thr Asn Ile Trp Met  
 930 935 940  
 Lys Phe Arg Glu Gly Ser Ser Glu Val Cys Asp His Arg Leu Met Ser  
 945 950 955 960

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Ala	Ala	Ile	Lys	Asp	Gln	Lys	Ala	Val	His	Ala	Asp	Met	Gly	Tyr	Trp	965	970	975	
Ile	Glu	Ser	Ser	Lys	Asn	Gln	Thr	Trp	Gln	Ile	Glu	Lys	Ala	Ser	Leu	980	985	990	
Ile	Glu	Val	Lys	Thr	Cys	Leu	Trp	Pro	Lys	Thr	His	Thr	Leu	Trp	Ser	995	1000	1005	
Asn	Gly	Val	Leu	Glu	Ser	Gln	Met	Leu	Ile	Pro	Lys	Ser	Tyr	Ala	Gly	1010	1015	1020	
Pro	Phe	Ser	Gln	His	Asn	Tyr	Arg	Gln	Gly	Tyr	Ala	Thr	Gln	Thr	Val	1025	1030	1035	1040
Gly	Pro	Trp	His	Leu	Gly	Lys	Leu	Glu	Ile	Asp	Phe	Gly	Glu	Cys	Pro	1045	1050	1055	
Gly	Thr	Thr	Val	Thr	Ile	Gln	Glu	Asp	Cys	Asp	His	Arg	Gly	Pro	Ser	1060	1065	1070	
Leu	Arg	Thr	Thr	Thr	Ala	Ser	Gly	Lys	Leu	Val	Thr	Gln	Trp	Cys	Cys	1075	1080	1085	
Arg	Ser	Cys	Thr	Met	Pro	Pro	Leu	Arg	Phe	Leu	Gly	Glu	Asp	Gly	Cys	1090	1095	1100	
Trp	Tyr	Gly	Met	Glu	Ile	Arg	Pro	Leu	Ser	Glu	Lys	Glu	Glu	Asn	Met	1105	1110	1115	1120
Val	Lys	Ser	Gln	Val	Thr	Ala	Gly	Gln	Gly	Thr	Ser	Glu	Thr	Phe	Ser	1125	1130	1135	
Met	Gly	Leu	Leu	Cys	Leu	Thr	Leu	Phe	Val	Glu	Glu	Cys	Leu	Arg	Arg	1140	1145	1150	
Arg	Val	Thr	Arg	Lys	His	Met	Ile	Leu	Val	Val	Val	Ile	Thr	Leu	Cys	1155	1160	1165	
Ala	Ile	Ile	Leu	Gly	Gly	Leu	Thr	Trp	Met	Asp	Leu	Leu	Arg	Ala	Leu	1170	1175	1180	
Ile	Met	Leu	Gly	Asp	Thr	Met	Ser	Gly	Arg	Ile	Gly	Gly	Gln	Ile	His	1185	1190	1195	1200
Leu	Ala	Ile	Met	Ala	Val	Phe	Lys	Met	Ser	Pro	Gly	Tyr	Val	Leu	Gly	1205	1210	1215	
Val	Phe	Leu	Arg	Lys	Leu	Thr	Ser	Arg	Glu	Thr	Ala	Leu	Met	Val	Ile	1220	1225	1230	
Gly	Met	Ala	Met	Thr	Thr	Val	Leu	Ser	Ile	Pro	His	Asp	Leu	Met	Glu	1235	1240	1245	
Leu	Ile	Asp	Gly	Ile	Ser	Leu	Gly	Leu	Ile	Leu	Leu	Lys	Ile	Val	Thr	1250	1255	1260	
Gln	Phe	Asp	Asn	Thr	Gln	Val	Gly	Thr	Leu	Ala	Leu	Ser	Leu	Thr	Phe	1265	1270	1275	1280
Ile	Arg	Ser	Thr	Met	Pro	Leu	Val	Met	Ala	Trp	Arg	Thr	Ile	Met	Ala	1285	1290	1295	
Val	Leu	Phe	Val	Val	Thr	Leu	Ile	Pro	Leu	Cys	Arg	Thr	Ser	Cys	Leu	1300	1305	1310	
Gln	Lys	Gln	Ser	His	Trp	Val	Glu	Ile	Thr	Ala	Leu	Ile	Leu	Gly	Ala	1315	1320	1325	
Gln	Ala	Leu	Pro	Val	Tyr	Leu	Met	Thr	Leu	Met	Lys	Gly	Ala	Ser	Arg	1330	1335	1340	
Arg	Ser	Trp	Pro	Leu	Asn	Glu	Gly	Ile	Met	Ala	Val	Gly	Leu	Val	Ser	1345	1350	1355	1360
Leu	Leu	Gly	Ser	Ala	Leu	Leu	Lys	Asn	Asp	Val	Pro	Leu	Ala	Gly	Pro	1365	1370	1375	
Met	Val	Ala	Gly	Gly	Leu	Leu	Leu	Ala	Ala	Tyr	Val	Met	Ser	Gly	Ser				

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1380				1385				1390							
Ser	Ala	Asp	Leu	Ser	Leu	Glu	Lys	Ala	Ala	Asn	Val	Gln	Trp	Asp	Glu
		1395					1400							1405	
Met	Ala	Asp	Ile	Thr	Gly	Ser	Ser	Pro	Ile	Val	Glu	Val	Lys	Gln	Asp
		1410				1415					1420				
Glu	Asp	Gly	Ser	Phe	Ser	Ile	Arg	Asp	Val	Glu	Glu	Thr	Asn	Met	Ile
		1425				1430				1435				1440	
Thr	Leu	Leu	Val	Lys	Leu	Ala	Leu	Ile	Thr	Val	Ser	Gly	Leu	Tyr	Pro
						1445				1450				1455	
Leu	Ala	Ile	Pro	Val	Thr	Met	Thr	Leu	Trp	Tyr	Met	Trp	Gln	Val	Lys
			1460							1465				1470	
Thr	Gln	Arg	Ser	Gly	Ala	Leu	Trp	Asp	Val	Pro	Ser	Pro	Ala	Ala	Thr
		1475					1480							1485	
Lys	Lys	Ala	Ala	Leu	Ser	Glu	Gly	Val	Tyr	Arg	Ile	Met	Gln	Arg	Gly
		1490				1495					1500				
Leu	Phe	Gly	Lys	Thr	Gln	Val	Gly	Val	Gly	Ile	His	Met	Glu	Gly	Val
		1505				1510				1515				1520	
Phe	His	Thr	Met	Trp	His	Val	Thr	Arg	Gly	Ser	Val	Ile	Cys	His	Glu
			1525							1530				1535	
Thr	Gly	Arg	Leu	Glu	Pro	Ser	Trp	Ala	Asp	Val	Arg	Asn	Asp	Met	Ile
			1540							1545				1550	
Ser	Tyr	Gly	Gly	Gly	Trp	Arg	Leu	Gly	Asp	Lys	Trp	Asp	Lys	Glu	Glu
		1555					1560							1565	
Asp	Val	Gln	Val	Leu	Ala	Ile	Glu	Pro	Gly	Lys	Asn	Pro	Lys	His	Val
		1570				1575					1580				
Gln	Thr	Lys	Pro	Gly	Leu	Phe	Lys	Thr	Leu	Thr	Gly	Glu	Ile	Gly	Ala
		1585				1590				1595				1600	
Val	Thr	Leu	Asp	Phe	Lys	Pro	Gly	Thr	Ser	Gly	Ser	Pro	Ile	Ile	Asn
			1605							1610				1615	
Arg	Lys	Gly	Lys	Val	Ile	Gly	Leu	Tyr	Gly	Asn	Gly	Val	Val	Thr	Lys
			1620							1625				1630	
Ser	Gly	Asp	Tyr	Val	Ser	Ala	Ile	Thr	Gln	Ala	Glu	Arg	Ile	Gly	Glu
		1635					1640							1645	
Pro	Asp	Tyr	Glu	Val	Asp	Glu	Asp	Ile	Phe	Arg	Lys	Lys	Arg	Leu	Thr
		1650				1655					1660				
Ile	Met	Asp	Leu	His	Pro	Gly	Ala	Gly	Lys	Thr	Lys	Arg	Ile	Leu	Pro
		1665				1670				1675				1680	
Ser	Ile	Val	Arg	Glu	Ala	Leu	Lys	Arg	Arg	Leu	Arg	Thr	Leu	Ile	Leu
			1685							1690				1695	
Ala	Pro	Thr	Arg	Val	Val	Ala	Ala	Glu	Met	Glu	Glu	Ala	Leu	Arg	Gly
			1700							1705				1710	
Leu	Pro	Ile	Arg	Tyr	Gln	Thr	Pro	Ala	Val	Lys	Ser	Glu	His	Thr	Gly
		1715					1720							1725	
Arg	Glu	Ile	Val	Asp	Leu	Met	Cys	His	Ala	Thr	Phe	Thr	Thr	Arg	Leu
		1730				1735					1740				
Leu	Ser	Ser	Thr	Arg	Val	Pro	Asn	Tyr	Asn	Leu	Ile	Val	Met	Asp	Glu
		1745				1750				1755				1760	
Ala	His	Phe	Thr	Asp	Pro	Ser	Ser	Val	Ala	Ala	Arg	Gly	Tyr	Ile	Ser
			1765							1770				1775	
Thr	Arg	Val	Glu	Met	Gly	Glu	Ala	Ala	Ala	Ile	Phe	Met	Thr	Ala	Thr
			1780				1785							1790	
Pro	Pro	Gly	Ala	Thr	Asp	Pro	Phe	Pro	Gln	Ser	Asn	Ser	Pro	Ile	Glu
		1795					1800							1805	

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Asp Ile Glu Arg Glu Ile Pro Glu Arg Ser Trp Asn Thr Gly Phe Asp  
 1810 1815 1820

Trp Ile Thr Asp Tyr Gln Gly Lys Thr Val Trp Phe Val Pro Ser Ile  
 1825 1830 1835 1840

Lys Ala Gly Asn Asp Ile Ala Asn Cys Leu Arg Lys Ser Gly Lys Lys  
 1845 1850 1855

Val Ile Gln Leu Ser Arg Lys Thr Phe Asp Thr Glu Tyr Pro Lys Thr  
 1860 1865 1870

Lys Leu Thr Asp Trp Asp Phe Val Val Thr Thr Asp Ile Ser Glu Met  
 1875 1880 1885

Gly Ala Asn Phe Arg Ala Gly Arg Val Ile Asp Pro Arg Arg Cys Leu  
 1890 1895 1900

Lys Pro Val Ile Leu Pro Asp Gly Pro Glu Arg Val Ile Leu Ala Gly  
 1905 1910 1915 1920

Pro Ile Pro Val Thr Pro Ala Ser Ala Ala Gln Arg Arg Gly Arg Ile  
 1925 1930 1935

Gly Arg Asn Pro Ala Gln Glu Asp Asp Gln Tyr Val Phe Ser Gly Asp  
 1940 1945 1950

Pro Leu Lys Asn Asp Glu Asp His Ala His Trp Thr Glu Ala Lys Met  
 1955 1960 1965

Leu Leu Asp Asn Ile Tyr Thr Pro Glu Gly Ile Ile Pro Thr Leu Phe  
 1970 1975 1980

Gly Pro Glu Arg Glu Lys Thr Gln Ala Ile Asp Gly Glu Phe Arg Leu  
 1985 1990 1995 2000

Arg Gly Glu Gln Arg Lys Thr Phe Val Glu Leu Met Arg Arg Gly Asp  
 2005 2010 2015

Leu Pro Val Trp Leu Ser Tyr Lys Val Ala Ser Ala Gly Ile Ser Tyr  
 2020 2025 2030

Glu Asp Arg Glu Trp Cys Phe Thr Gly Glu Arg Asn Asn Gln Ile Leu  
 2035 2040 2045

Glu Glu Asn Met Glu Val Glu Ile Trp Thr Arg Glu Gly Glu Lys Lys  
 2050 2055 2060

Lys Leu Arg Pro Arg Trp Leu Asp Ala Arg Val Tyr Ala Asp Pro Met  
 2065 2070 2075 2080

Ala Leu Lys Asp Phe Lys Glu Phe Ala Ser Gly Arg Lys Ser Ile Thr  
 2085 2090 2095

Leu Asp Ile Leu Thr Glu Ile Ala Ser Leu Pro Thr Tyr Leu Ser Ser  
 2100 2105 2110

Arg Ala Lys Leu Ala Leu Asp Asn Ile Val Met Leu His Thr Thr Glu  
 2115 2120 2125

Arg Gly Gly Arg Ala Tyr Gln His Ala Leu Asn Glu Leu Pro Glu Ser  
 2130 2135 2140

Leu Glu Thr Leu Met Leu Val Ala Leu Leu Gly Ala Met Thr Ala Gly  
 2145 2150 2155 2160

Ile Phe Leu Phe Phe Met Gln Gly Lys Gly Ile Gly Lys Leu Ser Met  
 2165 2170 2175

Gly Leu Ile Thr Ile Ala Val Ala Ser Gly Leu Leu Trp Val Ala Glu  
 2180 2185 2190

Ile Gln Pro Gln Trp Ile Ala Ala Ser Ile Ile Leu Glu Phe Phe Leu  
 2195 2200 2205

Met Val Leu Leu Ile Pro Glu Pro Glu Lys Gln Arg Thr Pro Gln Asp  
 2210 2215 2220



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Asn	Gln	Leu	Ile	Tyr	Val	Ile	Leu	Thr	Ile	Leu	Thr	Ile	Ile	Gly	Leu	2225	2230	2235	2240
Ile	Ala	Ala	Asn	Glu	Met	Gly	Leu	Ile	Glu	Lys	Thr	Lys	Thr	Asp	Phe	2245	2250	2255	
Gly	Phe	Tyr	Gln	Val	Lys	Thr	Glu	Thr	Thr	Ile	Leu	Asp	Val	Asp	Leu	2260	2265	2270	
Arg	Pro	Ala	Ser	Ala	Trp	Thr	Leu	Tyr	Ala	Val	Ala	Thr	Thr	Ile	Leu	2275	2280	2285	
Thr	Pro	Met	Leu	Arg	His	Thr	Ile	Glu	Asn	Thr	Ser	Ala	Asn	Leu	Ser	2290	2295	2300	
Leu	Ala	Ala	Ile	Ala	Asn	Gln	Ala	Ala	Val	Leu	Met	Gly	Leu	Gly	Lys	2305	2310	2315	2320
Gly	Trp	Pro	Leu	His	Arg	Met	Asp	Leu	Gly	Val	Pro	Leu	Leu	Ala	Met	2325	2330	2335	
Gly	Cys	Tyr	Ser	Gln	Val	Asn	Pro	Thr	Thr	Leu	Thr	Ala	Ser	Leu	Val	2340	2345	2350	
Met	Leu	Leu	Val	His	Tyr	Ala	Ile	Ile	Gly	Pro	Gly	Leu	Gln	Ala	Lys	2355	2360	2365	
Ala	Thr	Arg	Glu	Ala	Gln	Lys	Arg	Thr	Ala	Ala	Gly	Ile	Met	Lys	Asn	2370	2375	2380	
Pro	Thr	Val	Asp	Gly	Ile	Thr	Val	Ile	Asp	Leu	Glu	Pro	Ile	Ser	Tyr	2385	2390	2395	2400
Asp	Pro	Lys	Phe	Glu	Lys	Gln	Leu	Gly	Gln	Val	Met	Leu	Leu	Val	Leu	2405	2410	2415	
Cys	Ala	Gly	Gln	Leu	Leu	Leu	Met	Arg	Thr	Thr	Trp	Ala	Phe	Cys	Glu	2420	2425	2430	
Val	Leu	Thr	Leu	Ala	Thr	Gly	Pro	Ile	Leu	Thr	Leu	Trp	Glu	Gly	Asn	2435	2440	2445	
Pro	Gly	Arg	Phe	Trp	Asn	Thr	Thr	Ile	Ala	Val	Ser	Thr	Ala	Asn	Ile	2450	2455	2460	
Phe	Arg	Gly	Ser	Tyr	Leu	Ala	Gly	Ala	Gly	Leu	Ala	Phe	Ser	Leu	Ile	2465	2470	2475	2480
Lys	Asn	Ala	Gln	Thr	Pro	Arg	Arg	Gly	Thr	Gly	Thr	Thr	Gly	Glu	Thr	2485	2490	2495	
Leu	Gly	Glu	Lys	Trp	Lys	Arg	Gln	Leu	Asn	Ser	Leu	Asp	Arg	Lys	Glu	2500	2505	2510	
Phe	Glu	Glu	Tyr	Lys	Arg	Ser	Gly	Ile	Leu	Glu	Val	Asp	Arg	Thr	Glu	2515	2520	2525	
Ala	Lys	Ser	Ala	Leu	Lys	Asp	Gly	Ser	Lys	Ile	Lys	His	Ala	Val	Ser	2530	2535	2540	
Arg	Gly	Ser	Ser	Lys	Ile	Arg	Trp	Ile	Val	Glu	Arg	Gly	Met	Val	Lys	2545	2550	2555	2560
Pro	Lys	Gly	Lys	Val	Val	Asp	Leu	Gly	Cys	Gly	Arg	Gly	Gly	Trp	Ser	2565	2570	2575	
Tyr	Tyr	Met	Ala	Thr	Leu	Lys	Asn	Val	Thr	Glu	Val	Lys	Gly	Tyr	Thr	2580	2585	2590	
Lys	Gly	Gly	Pro	Gly	His	Glu	Glu	Pro	Ile	Pro	Met	Ala	Thr	Tyr	Gly	2595	2600	2605	
Trp	Asn	Leu	Val	Lys	Leu	His	Ser	Gly	Val	Asp	Val	Phe	Tyr	Lys	Pro	2610	2615	2620	
Thr	Glu	Gln	Val	Asp	Thr	Leu	Leu	Cys	Asp	Ile	Gly	Glu	Ser	Ser	Ser	2625	2630	2635	2640
Asn	Pro	Thr	Ile	Glu	Glu	Gly	Arg	Thr	Leu	Arg	Val	Leu	Lys	Met	Val				

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2645					2650					2655					
Glu	Pro	Trp	Leu	Ser	Ser	Lys	Pro	Glu	Phe	Cys	Ile	Lys	Val	Leu	Asn
			2660					2665					2670		
Pro	Tyr	Met	Pro	Thr	Val	Ile	Glu	Glu	Leu	Glu	Lys	Leu	Gln	Arg	Lys
		2675					2680						2685		
His	Gly	Gly	Asn	Leu	Val	Arg	Cys	Pro	Leu	Ser	Arg	Asn	Ser	Thr	His
	2690					2695					2700				
Glu	Met	Tyr	Trp	Val	Ser	Gly	Ala	Ser	Gly	Asn	Ile	Val	Ser	Ser	Val
2705					2710					2715					2720
Asn	Thr	Thr	Ser	Lys	Met	Leu	Leu	Asn	Arg	Phe	Thr	Thr	Arg	His	Arg
				2725					2730					2735	
Lys	Pro	Thr	Tyr	Glu	Lys	Asp	Val	Asp	Leu	Gly	Ala	Gly	Thr	Arg	Ser
			2740					2745						2750	
Val	Ser	Thr	Glu	Thr	Glu	Lys	Pro	Asp	Met	Thr	Ile	Ile	Gly	Arg	Arg
		2755					2760						2765		
Leu	Gln	Arg	Leu	Gln	Glu	Glu	His	Lys	Glu	Thr	Trp	His	Tyr	Asp	Gln
	2770					2775					2780				
Glu	Asn	Pro	Tyr	Arg	Thr	Trp	Ala	Tyr	His	Gly	Ser	Tyr	Glu	Ala	Pro
2785					2790					2795					2800
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&lt;212&gt; TYPE: DNA

&lt;213&gt; ORGANISM: Recombinant dengue virus rDEN2/4d30

&lt;400&gt; SEQUENCE: 18

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aaagaaagag	ttgagaaatg	gctgaaagag	tgtggtgtcg	acaggttaaa	gaggatggca	9540
atcagtggag	acgattgcgt	ggtgaagccc	ctagatgaga	ggtttggcac	ttccctcctc	9600
ttcttgaacg	acatgggaaa	ggtgaggaaa	gacattccgc	agtgggaacc	atctaagggg	9660
tggaaaaact	ggcaagaggt	tcctttttgc	tcccaccact	ttcacaagat	ctttatgaag	9720
gatggccgct	cactagttgt	tccatgtaga	aaccaggatg	aactgatagg	gagagccaga	9780

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atctcgcagg gagctggatg gagcttaaga gaaacagcct gcctgggcaa agcttacgcc 9840
cagatgtggg cgcttatgta cttccacaga agggatctgc gtttagcctc catggccata 9900
tgctcagcag ttccaacgga atggtttcca acaagcagaa caacatggtc aatccacgct 9960
catcaccagt ggatgaccac tgaagatatg ctcaaagtgt ggaacagagt gtggatagaa 10020
gacaacccta atatgactga caagactcca gtccattcgt gggaagatat accttaccta 10080
gggaaaagag aggatttgtg gtgtggatcc ctgattggac tttcttcag agccacctgg 10140
gcgaagaaca ttcacacggc cataaccag gtcaggaacc tgatcggaaa agaggaatac 10200
gtggattaca tgccagtaat gaaaagatac agtgctcctt cagagagtga aggagtctg 10260
taattaccaa caacaaacac caaaggctat tgaagtcagg ccacttgtgc cacggtttga 10320
gcaaaccgtg ctgcctgtag ctccgccaat aatgggaggc gtaataatcc ccaggagggc 10380
catgcgccac ggaagctgta cgcgtggcat attggactag cggttagagg agaccctcc 10440
catcactgac aaaacgcagc aaaagggggc ccaagactag aggttagagg agaccccccc 10500
aacacaaaaa cagcatattg acgctgggaa agaccagaga tcctgctgtc tctgcaacat 10560
caatccaggc acagagcgcc gcaagatgga ttggtgttgt tgatccaaca ggttct 10616

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&lt;210&gt; SEQ ID NO 19

&lt;211&gt; LENGTH: 3387

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Dengue 4 virus

&lt;400&gt; SEQUENCE: 19

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Met Asn Gln Arg Lys Lys Val Val Arg Pro Pro Phe Asn Met Leu Lys
 1          5          10          15
Arg Glu Arg Asn Arg Val Ser Thr Pro Gln Gly Leu Val Lys Arg Phe
          20          25          30
Ser Thr Gly Leu Phe Ser Gly Lys Gly Pro Leu Arg Met Val Leu Ala
          35          40          45
Phe Ile Thr Phe Leu Arg Val Leu Ser Ile Pro Pro Thr Ala Gly Ile
          50          55          60
Leu Lys Arg Trp Gly Gln Leu Lys Lys Asn Lys Ala Ile Lys Ile Leu
65          70          75          80
Ile Gly Phe Arg Lys Glu Ile Gly Arg Met Leu Asn Ile Leu Asn Gly
          85          90          95
Arg Lys Arg Ser Thr Ile Thr Leu Leu Cys Leu Ile Pro Thr Val Met
          100          105          110
Ala Phe Ser Leu Ser Thr Arg Asp Gly Glu Pro Leu Met Ile Val Ala
          115          120          125
Lys His Glu Arg Gly Arg Pro Leu Leu Phe Lys Thr Thr Glu Gly Ile
          130          135          140
Asn Lys Cys Thr Leu Ile Ala Met Asp Leu Gly Glu Met Cys Glu Asp
145          150          155          160
Thr Val Thr Tyr Lys Cys Pro Leu Leu Val Asn Thr Glu Pro Glu Asp
          165          170          175
Ile Asp Cys Trp Cys Asn Leu Thr Ser Thr Trp Val Met Tyr Gly Thr
          180          185          190
Cys Thr Gln Ser Gly Glu Arg Arg Arg Glu Lys Arg Ser Val Ala Leu
          195          200          205
Thr Pro His Ser Gly Met Gly Leu Glu Thr Arg Ala Glu Thr Trp Met
          210          215          220
Ser Ser Glu Gly Ala Trp Lys His Ala Gln Arg Val Glu Ser Trp Ile

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225	230	235	240
Leu Arg Asn Pro Gly Phe Ala Leu Leu Ala Gly Phe Met Ala Tyr Met 245 250 255			
Ile Gly Gln Thr Gly Ile Gln Arg Thr Val Phe Phe Val Leu Met Met 260 265 270			
Leu Val Ala Pro Ser Tyr Gly Met Arg Cys Val Gly Val Gly Asn Arg 275 280 285			
Asp Phe Val Glu Gly Val Ser Gly Gly Ala Trp Val Asp Leu Val Leu 290 295 300			
Glu His Gly Gly Cys Val Thr Thr Met Ala Gln Gly Lys Pro Thr Leu 305 310 315 320			
Asp Phe Glu Leu Thr Lys Thr Thr Ala Lys Glu Val Ala Leu Leu Arg 325 330 335			
Thr Tyr Cys Ile Glu Ala Ser Ile Ser Asn Ile Thr Thr Ala Thr Arg 340 345 350			
Cys Pro Thr Gln Gly Glu Pro Tyr Leu Lys Glu Glu Gln Asp Gln Gln 355 360 365			
Tyr Ile Cys Arg Arg Asp Val Val Asp Arg Gly Trp Gly Asn Gly Cys 370 375 380			
Gly Leu Phe Gly Lys Gly Gly Val Val Thr Cys Ala Lys Phe Ser Cys 385 390 395 400			
Ser Gly Lys Ile Thr Gly Asn Leu Val Gln Ile Glu Asn Leu Glu Tyr 405 410 415			
Thr Val Val Val Thr Val His Asn Gly Asp Thr His Ala Val Gly Asn 420 425 430			
Asp Thr Ser Asn His Gly Val Thr Ala Met Ile Thr Pro Arg Ser Pro 435 440 445			
Ser Val Glu Val Lys Leu Pro Asp Tyr Gly Glu Leu Thr Leu Asp Cys 450 455 460			
Glu Pro Arg Ser Gly Ile Asp Phe Asn Glu Met Ile Leu Met Lys Met 465 470 475 480			
Lys Lys Lys Thr Trp Leu Val His Lys Gln Trp Phe Leu Asp Leu Pro 485 490 495			
Leu Pro Trp Thr Ala Gly Ala Asp Thr Ser Glu Val His Trp Asn Tyr 500 505 510			
Lys Glu Arg Met Val Thr Phe Lys Val Pro His Ala Lys Arg Gln Asp 515 520 525			
Val Thr Val Leu Gly Ser Gln Glu Gly Ala Met His Ser Ala Leu Ala 530 535 540			
Gly Ala Thr Glu Val Asp Ser Gly Asp Gly Asn His Met Phe Ala Gly 545 550 555 560			
His Leu Lys Cys Lys Val Arg Met Glu Lys Leu Arg Ile Lys Gly Met 565 570 575			
Ser Tyr Thr Met Cys Ser Gly Lys Phe Ser Ile Asp Lys Glu Met Ala 580 585 590			
Glu Thr Gln His Gly Thr Thr Val Val Lys Val Lys Tyr Glu Gly Ala 595 600 605			
Gly Ala Pro Cys Lys Val Pro Ile Glu Ile Arg Asp Val Asn Lys Glu 610 615 620			
Lys Val Val Gly Arg Ile Ile Ser Ser Thr Pro Leu Ala Glu Asn Thr 625 630 635 640			
Asn Ser Val Thr Asn Ile Glu Leu Glu Pro Pro Phe Gly Asp Ser Tyr 645 650 655			

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Ile Val Ile Gly Val Gly Asn Ser Ala Leu Thr Leu His Trp Phe Arg  
 660 665 670  
 Lys Gly Ser Ser Ile Gly Lys Met Phe Glu Ser Thr Tyr Arg Gly Ala  
 675 680 685  
 Lys Arg Met Ala Ile Leu Gly Glu Thr Ala Trp Asp Phe Gly Ser Val  
 690 695 700  
 Gly Gly Leu Phe Thr Ser Leu Gly Lys Ala Val His Gln Val Phe Gly  
 705 710 715 720  
 Ser Val Tyr Thr Thr Met Phe Gly Gly Val Ser Trp Met Ile Arg Ile  
 725 730 735  
 Leu Ile Gly Phe Leu Val Leu Trp Ile Gly Thr Asn Ser Arg Asn Thr  
 740 745 750  
 Ser Met Ala Met Thr Cys Ile Ala Val Gly Gly Ile Thr Leu Phe Leu  
 755 760 765  
 Gly Phe Thr Val Gln Ala Asp Met Gly Cys Val Ala Ser Trp Ser Gly  
 770 775 780  
 Lys Glu Leu Lys Cys Gly Ser Gly Ile Phe Val Val Asp Asn Val His  
 785 790 795 800  
 Thr Trp Thr Glu Gln Tyr Lys Phe Gln Pro Glu Ser Pro Ala Arg Leu  
 805 810 815  
 Ala Ser Ala Ile Leu Asn Ala His Lys Asp Gly Val Cys Gly Ile Arg  
 820 825 830  
 Ser Thr Thr Arg Leu Glu Asn Val Met Trp Lys Gln Ile Thr Asn Glu  
 835 840 845  
 Leu Asn Tyr Val Leu Trp Glu Gly Gly His Asp Leu Thr Val Val Ala  
 850 855 860  
 Gly Asp Val Lys Gly Val Leu Thr Lys Gly Lys Arg Ala Leu Thr Pro  
 865 870 875 880  
 Pro Val Ser Asp Leu Lys Tyr Ser Trp Lys Thr Trp Gly Lys Ala Lys  
 885 890 895  
 Ile Phe Thr Pro Glu Ala Arg Asn Ser Thr Phe Leu Ile Asp Gly Pro  
 900 905 910  
 Asp Thr Ser Glu Cys Pro Asn Glu Arg Arg Ala Trp Asn Ser Leu Glu  
 915 920 925  
 Val Glu Asp Tyr Gly Phe Gly Met Phe Thr Thr Asn Ile Trp Met Lys  
 930 935 940  
 Phe Arg Glu Gly Ser Ser Glu Val Cys Asp His Arg Leu Met Ser Ala  
 945 950 955 960  
 Ala Ile Lys Asp Gln Lys Ala Val His Ala Asp Met Gly Tyr Trp Ile  
 965 970 975  
 Glu Ser Ser Lys Asn Gln Thr Trp Gln Ile Glu Lys Ala Ser Leu Ile  
 980 985 990  
 Glu Val Lys Thr Cys Leu Trp Pro Lys Thr His Thr Leu Trp Ser Asn  
 995 1000 1005  
 Gly Val Leu Glu Ser Gln Met Leu Ile Pro Lys Ser Tyr Ala Gly Pro  
 1010 1015 1020  
 Phe Ser Gln His Asn Tyr Arg Gln Gly Tyr Ala Thr Gln Thr Val Gly  
 1025 1030 1035 1040  
 Pro Trp His Leu Gly Lys Leu Glu Ile Asp Phe Gly Glu Cys Pro Gly  
 1045 1050 1055  
 Thr Thr Val Thr Ile Gln Glu Asp Cys Asp His Arg Gly Pro Ser Leu  
 1060 1065 1070

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Arg Thr Thr Thr Ala Ser Gly Lys Leu Val Thr Gln Trp Cys Cys Arg  
1075 1080 1085

Ser Cys Thr Met Pro Pro Leu Arg Phe Leu Gly Glu Asp Gly Cys Trp  
1090 1095 1100

Tyr Gly Met Glu Ile Arg Pro Leu Ser Glu Lys Glu Glu Asn Met Val  
1105 1110 1115 1120

Lys Ser Gln Val Thr Ala Gly Gln Gly Thr Ser Glu Thr Phe Ser Met  
1125 1130 1135

Gly Leu Leu Cys Leu Thr Leu Phe Val Glu Glu Cys Leu Arg Arg Arg  
1140 1145 1150

Val Thr Arg Lys His Met Ile Leu Val Val Val Ile Thr Leu Cys Ala  
1155 1160 1165

Ile Ile Leu Gly Gly Leu Thr Trp Met Asp Leu Leu Arg Ala Leu Ile  
1170 1175 1180

Met Leu Gly Asp Thr Met Ser Gly Arg Ile Gly Gly Gln Ile His Leu  
1185 1190 1195 1200

Ala Ile Met Ala Val Phe Lys Met Ser Pro Gly Tyr Val Leu Gly Val  
1205 1210 1215

Phe Leu Arg Lys Leu Thr Ser Arg Glu Thr Ala Leu Met Val Ile Gly  
1220 1225 1230

Met Ala Met Thr Thr Val Leu Ser Ile Pro His Asp Leu Met Glu Leu  
1235 1240 1245

Ile Asp Gly Ile Ser Leu Gly Leu Ile Leu Leu Lys Ile Val Thr Gln  
1250 1255 1260

Phe Asp Asn Thr Gln Val Gly Thr Leu Ala Leu Ser Leu Thr Phe Ile  
1265 1270 1275 1280

Arg Ser Thr Met Pro Leu Val Met Ala Trp Arg Thr Ile Met Ala Val  
1285 1290 1295

Leu Phe Val Val Thr Leu Ile Pro Leu Cys Arg Thr Ser Cys Leu Gln  
1300 1305 1310

Lys Gln Ser His Trp Val Glu Ile Thr Ala Leu Ile Leu Gly Ala Gln  
1315 1320 1325

Ala Leu Pro Val Tyr Leu Met Thr Leu Met Lys Gly Ala Ser Arg Arg  
1330 1335 1340

Ser Trp Pro Leu Asn Glu Gly Ile Met Ala Val Gly Leu Val Ser Leu  
1345 1350 1355 1360

Leu Gly Ser Ala Leu Leu Lys Asn Asp Val Pro Leu Ala Gly Pro Met  
1365 1370 1375

Val Ala Gly Gly Leu Leu Leu Ala Ala Tyr Val Met Ser Gly Ser Ser  
1380 1385 1390

Ala Asp Leu Ser Leu Glu Lys Ala Ala Asn Val Gln Trp Asp Glu Met  
1395 1400 1405

Ala Asp Ile Thr Gly Ser Ser Pro Ile Ile Glu Val Lys Gln Asp Glu  
1410 1415 1420

Asp Gly Ser Phe Ser Ile Arg Asp Val Glu Glu Thr Asn Met Ile Thr  
1425 1430 1435 1440

Leu Leu Val Lys Leu Ala Leu Ile Thr Val Ser Gly Leu Tyr Pro Leu  
1445 1450 1455

Ala Ile Pro Val Thr Met Thr Leu Trp Tyr Met Trp Gln Val Lys Thr  
1460 1465 1470

Gln Arg Ser Gly Ala Leu Trp Asp Val Pro Ser Pro Ala Ala Thr Lys  
1475 1480 1485

Lys Ala Ala Leu Ser Glu Gly Val Tyr Arg Ile Met Gln Arg Gly Leu

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1490	1495	1500
Phe Gly Lys Thr Gln Val Gly Val Gly Ile His Met Glu Gly Val Phe 1505	1510	1515
His Thr Met Trp His Val Thr Arg Gly Ser Val Ile Cys His Glu Thr 1525	1530	1535
Gly Arg Leu Glu Pro Ser Trp Ala Asp Val Arg Asn Asp Met Ile Ser 1540	1545	1550
Tyr Gly Gly Gly Trp Arg Leu Gly Asp Lys Trp Asp Lys Glu Glu Asp 1555	1560	1565
Val Gln Val Leu Ala Ile Glu Pro Gly Lys Asn Pro Lys His Val Gln 1570	1575	1580
Thr Lys Pro Gly Leu Phe Lys Thr Leu Thr Gly Glu Ile Gly Ala Val 1585	1590	1595
Thr Leu Asp Phe Lys Pro Gly Thr Ser Gly Ser Pro Ile Ile Asn Arg 1605	1610	1615
Lys Gly Lys Val Ile Gly Leu Tyr Gly Asn Gly Val Val Thr Lys Ser 1620	1625	1630
Gly Asp Tyr Val Ser Ala Ile Thr Gln Ala Glu Arg Ile Gly Glu Pro 1635	1640	1645
Asp Tyr Glu Val Asp Glu Asp Ile Phe Arg Lys Lys Arg Leu Thr Ile 1650	1655	1660
Met Asp Leu His Pro Gly Ala Gly Lys Thr Lys Arg Ile Leu Pro Ser 1665	1670	1675
Ile Val Arg Glu Ala Leu Lys Arg Arg Leu Arg Thr Leu Ile Leu Ala 1685	1690	1695
Pro Thr Arg Val Val Ala Ala Glu Met Glu Glu Ala Leu Arg Gly Leu 1700	1705	1710
Pro Ile Arg Tyr Gln Thr Pro Ala Val Lys Ser Glu His Thr Gly Arg 1715	1720	1725
Glu Ile Val Asp Leu Met Cys His Ala Thr Phe Thr Thr Arg Leu Leu 1730	1735	1740
Ser Ser Thr Arg Val Pro Asn Tyr Asn Leu Ile Val Met Asp Glu Ala 1745	1750	1755
His Phe Thr Asp Pro Ser Ser Val Ala Ala Arg Gly Tyr Ile Ser Thr 1765	1770	1775
Arg Val Glu Met Gly Glu Ala Ala Ala Ile Phe Met Thr Ala Thr Pro 1780	1785	1790
Pro Gly Ala Thr Asp Pro Phe Pro Gln Ser Asn Ser Pro Ile Glu Asp 1795	1800	1805
Ile Glu Arg Glu Ile Pro Glu Arg Ser Trp Asn Thr Gly Phe Asp Trp 1810	1815	1820
Ile Thr Asp Tyr Gln Gly Lys Thr Val Trp Phe Val Pro Ser Ile Lys 1825	1830	1835
Ala Gly Asn Asp Ile Ala Asn Cys Leu Arg Lys Ser Gly Lys Lys Val 1845	1850	1855
Ile Gln Leu Ser Arg Lys Thr Phe Asp Thr Glu Tyr Pro Lys Thr Lys 1860	1865	1870
Leu Thr Asp Trp Asp Phe Val Val Thr Thr Asp Ile Ser Glu Met Gly 1875	1880	1885
Ala Asn Phe Arg Ala Gly Arg Val Ile Asp Pro Arg Arg Cys Leu Lys 1890	1895	1900
Pro Val Ile Leu Pro Asp Gly Pro Glu Arg Val Ile Leu Ala Gly Pro 1905	1910	1915
		1920

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Ile Pro Val Thr Pro Ala Ser Ala Ala Gln Arg Arg Gly Arg Ile Gly  
 1925 1930 1935  
 Arg Asn Pro Ala Gln Glu Asp Asp Gln Tyr Val Phe Ser Gly Asp Pro  
 1940 1945 1950  
 Leu Lys Asn Asp Glu Asp His Ala His Trp Thr Glu Ala Lys Met Leu  
 1955 1960 1965  
 Leu Asp Asn Ile Tyr Thr Pro Glu Gly Ile Ile Pro Thr Leu Phe Gly  
 1970 1975 1980  
 Pro Glu Arg Glu Lys Thr Gln Ala Ile Asp Gly Glu Phe Arg Leu Arg  
 1985 1990 1995 2000  
 Gly Glu Gln Arg Lys Thr Phe Val Glu Leu Met Arg Arg Gly Asp Leu  
 2005 2010 2015  
 Pro Val Trp Leu Ser Tyr Lys Val Ala Ser Ala Gly Ile Ser Tyr Lys  
 2020 2025 2030  
 Asp Arg Glu Trp Cys Phe Thr Gly Glu Arg Asn Asn Gln Ile Leu Glu  
 2035 2040 2045  
 Glu Asn Met Glu Val Glu Ile Trp Thr Arg Glu Gly Glu Lys Lys Lys  
 2050 2055 2060  
 Leu Arg Pro Arg Trp Leu Asp Ala Arg Val Tyr Ala Asp Pro Met Ala  
 2065 2070 2075 2080  
 Leu Lys Asp Phe Lys Glu Phe Ala Ser Gly Arg Lys Ser Ile Thr Leu  
 2085 2090 2095  
 Asp Ile Leu Thr Glu Ile Ala Ser Leu Pro Thr Tyr Leu Ser Ser Arg  
 2100 2105 2110  
 Ala Lys Leu Ala Leu Asp Asn Ile Val Met Leu His Thr Thr Glu Arg  
 2115 2120 2125  
 Gly Gly Arg Ala Tyr Gln His Ala Leu Asn Glu Leu Pro Glu Ser Leu  
 2130 2135 2140  
 Glu Thr Leu Met Leu Val Ala Leu Leu Gly Ala Met Thr Ala Gly Ile  
 2145 2150 2155 2160  
 Phe Leu Phe Phe Met Gln Gly Lys Gly Ile Gly Lys Leu Ser Met Gly  
 2165 2170 2175  
 Leu Ile Thr Ile Ala Val Ala Ser Gly Leu Leu Trp Val Ala Glu Ile  
 2180 2185 2190  
 Gln Pro Gln Trp Ile Ala Ala Ser Ile Ile Leu Glu Phe Phe Leu Met  
 2195 2200 2205  
 Val Leu Leu Ile Pro Glu Pro Glu Lys Gln Arg Thr Pro Gln Asp Asn  
 2210 2215 2220  
 Gln Leu Ile Tyr Val Ile Leu Thr Ile Leu Thr Ile Ile Gly Leu Ile  
 2225 2230 2235 2240  
 Ala Ala Asn Glu Met Gly Leu Ile Glu Lys Thr Lys Thr Asp Phe Gly  
 2245 2250 2255  
 Phe Tyr Gln Val Lys Thr Glu Thr Thr Ile Leu Asp Val Asp Leu Arg  
 2260 2265 2270  
 Pro Ala Ser Ala Trp Thr Leu Tyr Ala Val Ala Thr Thr Ile Leu Thr  
 2275 2280 2285  
 Pro Met Leu Arg His Thr Ile Glu Asn Thr Ser Ala Asn Leu Ser Leu  
 2290 2295 2300  
 Ala Ala Ile Ala Asn Gln Ala Ala Val Leu Met Gly Leu Gly Lys Gly  
 2305 2310 2315 2320  
 Trp Pro Leu His Arg Met Asp Leu Gly Val Pro Leu Leu Ala Met Gly  
 2325 2330 2335

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Cys Tyr Ser Gln Val Asn Pro Thr Thr Leu Thr Ala Ser Leu Val Met  
2340 2345 2350

Leu Leu Val His Tyr Ala Ile Ile Gly Pro Gly Leu Gln Ala Lys Ala  
2355 2360 2365

Thr Arg Glu Ala Gln Lys Arg Thr Ala Ala Gly Ile Met Lys Asn Pro  
2370 2375 2380

Thr Val Asp Gly Ile Thr Val Ile Asp Leu Glu Pro Ile Ser Tyr Asp  
2385 2390 2395 2400

Pro Lys Phe Glu Lys Gln Leu Gly Gln Val Met Leu Leu Val Leu Cys  
2405 2410 2415

Ala Gly Gln Leu Leu Leu Met Arg Thr Thr Trp Ala Phe Cys Glu Val  
2420 2425 2430

Leu Thr Leu Ala Thr Gly Pro Ile Leu Thr Leu Trp Glu Gly Asn Pro  
2435 2440 2445

Gly Arg Phe Trp Asn Thr Thr Ile Ala Val Ser Thr Ala Asn Ile Phe  
2450 2455 2460

Arg Gly Ser Tyr Leu Ala Gly Ala Gly Leu Ala Phe Ser Leu Ile Lys  
2465 2470 2475 2480

Asn Ala Gln Thr Pro Arg Arg Gly Thr Gly Thr Thr Gly Glu Thr Leu  
2485 2490 2495

Gly Glu Lys Trp Lys Arg Gln Leu Asn Ser Leu Asp Arg Lys Glu Phe  
2500 2505 2510

Glu Glu Tyr Lys Arg Ser Gly Ile Leu Glu Val Asp Arg Thr Glu Ala  
2515 2520 2525

Lys Ser Ala Leu Lys Asp Gly Ser Lys Ile Lys His Ala Val Ser Arg  
2530 2535 2540

Gly Ser Ser Lys Ile Arg Trp Ile Val Glu Arg Gly Met Val Lys Pro  
2545 2550 2555 2560

Lys Gly Lys Val Val Asp Leu Gly Cys Gly Arg Gly Gly Trp Ser Tyr  
2565 2570 2575

Tyr Met Ala Thr Leu Lys Asn Val Thr Glu Val Lys Gly Tyr Thr Lys  
2580 2585 2590

Gly Gly Pro Gly His Glu Glu Pro Ile Pro Met Ala Thr Tyr Gly Trp  
2595 2600 2605

Asn Leu Val Lys Leu His Ser Gly Val Asp Val Phe Tyr Lys Pro Thr  
2610 2615 2620

Glu Gln Val Asp Thr Leu Leu Cys Asp Ile Gly Glu Ser Ser Ser Asn  
2625 2630 2635

Pro Thr Ile Glu Glu Gly Arg Thr Leu Arg Val Leu Lys Met Val Glu  
2645 2650 2655

Pro Trp Leu Ser Ser Lys Pro Glu Phe Cys Ile Lys Val Leu Asn Pro  
2660 2665 2670

Tyr Met Pro Thr Val Ile Glu Glu Leu Glu Lys Leu Gln Arg Lys His  
2675 2680 2685

Gly Gly Asn Leu Val Arg Cys Pro Leu Ser Arg Asn Ser Thr His Glu  
2690 2695 2700

Met Tyr Trp Val Ser Gly Ala Ser Gly Asn Ile Val Ser Ser Val Asn  
2705 2710 2715 2720

Thr Thr Ser Lys Met Leu Leu Asn Arg Phe Thr Thr Arg His Arg Lys  
2725 2730 2735

Pro Thr Tyr Glu Lys Asp Val Asp Leu Gly Ala Gly Thr Arg Ser Val  
2740 2745 2750

Ser Thr Glu Thr Glu Lys Pro Asp Met Thr Ile Ile Gly Arg Arg Leu

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2755			2760			2765		
Gln Arg Leu	Gln Glu	Glu His Lys	Glu Thr Trp	His Tyr Asp	Gln Glu			
2770		2775		2780				
Asn Pro Tyr	Arg Thr Trp	Ala Tyr His	Gly Ser Tyr	Glu Ala Pro	Ser			
2785		2790		2795				2800
Thr Gly Ser	Ala Ser Ser	Met Val Asn	Gly Val Val	Lys Leu Leu	Thr			
	2805		2810		2815			
Lys Pro Trp	Asp Val Ile	Pro Met Val	Thr Gln Leu	Ala Met Thr	Asp			
	2820		2825		2830			
Thr Thr Pro	Phe Gly Gln	Gln Arg Val	Phe Lys Glu	Lys Val Asp	Thr			
	2835		2840		2845			
Arg Thr Pro	Gln Pro Lys	Pro Gly Thr	Arg Met Val	Met Thr Thr	Thr			
	2850		2855		2860			
Ala Asn Trp	Leu Trp Ala	Leu Leu Gly	Lys Lys Lys	Asn Pro Arg	Leu			
2865		2870		2875				2880
Cys Thr Arg	Glu Glu Phe	Ile Ser Lys	Val Arg Ser	Asn Ala Ala	Ile			
	2885		2890		2895			
Gly Ala Val	Phe Gln Glu	Glu Gln Gly	Trp Thr Ser	Ala Ser Glu	Ala			
	2900		2905		2910			
Val Asn Asp	Ser Arg Phe	Trp Glu Leu	Val Asp Lys	Glu Arg Ala	Leu			
	2915		2920		2925			
His Gln Glu	Gly Lys Cys	Glu Ser Cys	Val Tyr Asn	Met Met Gly	Lys			
	2930		2935		2940			
Arg Glu Lys	Lys Leu Gly	Glu Phe Gly	Arg Ala Lys	Gly Ser Arg	Ala			
	2945		2950		2955			2960
Ile Trp Tyr	Met Trp Leu	Gly Ala Arg	Phe Leu Glu	Phe Glu Ala	Leu			
	2965		2970		2975			
Gly Phe Leu	Asn Glu Asp	His Trp Phe	Gly Arg Glu	Asn Ser Trp	Ser			
	2980		2985		2990			
Gly Val Glu	Gly Glu Gly	Leu His Arg	Leu Gly Tyr	Ile Leu Glu	Glu			
	2995		3000		3005			
Ile Asp Lys	Lys Asp Gly	Asp Leu Met	Tyr Ala Asp	Asp Thr Ala	Gly			
	3010		3015		3020			
Trp Asp Thr	Arg Ile Thr	Glu Asp Asp	Leu Gln Asn	Glu Glu Leu	Ile			
	3025		3030		3035			3040
Thr Glu Gln	Met Ala Pro	His His Lys	Ile Leu Ala	Lys Ala Ile	Phe			
	3045		3050		3055			
Lys Leu Thr	Tyr Gln Asn	Lys Val Val	Lys Val Leu	Arg Pro Thr	Pro			
	3060		3065		3070			
Arg Gly Ala	Val Met Asp	Ile Ile Ser	Arg Lys Asp	Gln Arg Gly	Ser			
	3075		3080		3085			
Gly Gln Val	Gly Thr Tyr	Gly Leu Asn	Thr Phe Thr	Asn Met Glu	Val			
	3090		3095		3100			
Gln Leu Ile	Arg Gln Met	Glu Ala Glu	Gly Val Ile	Thr Gln Asp	Asp			
	3105		3110		3115			3120
Met Gln Asn	Pro Lys Gly	Leu Lys Glu	Arg Val Glu	Lys Trp Leu	Lys			
	3125		3130		3135			
Glu Cys Gly	Val Asp Arg	Leu Lys Arg	Met Ala Ile	Ser Gly Asp	Asp			
	3140		3145		3150			
Cys Val Val	Lys Pro Leu	Asp Glu Arg	Phe Gly Thr	Ser Leu Leu	Phe			
	3155		3160		3165			
Leu Asn Asp	Met Gly Lys	Val Arg Lys	Asp Ile Pro	Gln Trp Glu	Pro			
	3170		3175		3180			

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Ser Lys Gly Trp Lys Asn Trp Gln Glu Val Pro Phe Cys Ser His His  
 3185 3190 3195 3200  
 Phe His Lys Ile Phe Met Lys Asp Gly Arg Ser Leu Val Val Pro Cys  
 3205 3210 3215  
 Arg Asn Gln Asp Glu Leu Ile Gly Arg Ala Arg Ile Ser Gln Gly Ala  
 3220 3225 3230  
 Gly Trp Ser Leu Arg Glu Thr Ala Cys Leu Gly Lys Ala Tyr Ala Gln  
 3235 3240 3245  
 Met Trp Ser Leu Met Tyr Phe His Arg Arg Asp Leu Arg Leu Ala Ser  
 3250 3255 3260  
 Met Ala Ile Cys Ser Ala Val Pro Thr Glu Trp Phe Pro Thr Ser Arg  
 3265 3270 3275 3280  
 Thr Thr Trp Ser Ile His Ala His His Gln Trp Met Thr Thr Glu Asp  
 3285 3290 3295  
 Met Leu Lys Val Trp Asn Arg Val Trp Ile Glu Asp Asn Pro Asn Met  
 3300 3305 3310  
 Thr Asp Lys Thr Pro Val His Ser Trp Glu Asp Ile Pro Tyr Leu Gly  
 3315 3320 3325  
 Lys Arg Glu Asp Leu Trp Cys Gly Ser Leu Ile Gly Leu Ser Ser Arg  
 3330 3335 3340  
 Ala Thr Trp Ala Lys Asn Ile His Thr Ala Ile Thr Gln Val Arg Asn  
 3345 3350 3355 3360  
 Leu Ile Gly Lys Glu Glu Tyr Val Asp Tyr Met Pro Val Met Lys Arg  
 3365 3370 3375  
 Tyr Ser Ala Pro Ser Glu Ser Glu Gly Val Leu  
 3380 3385

&lt;210&gt; SEQ ID NO 20

&lt;211&gt; LENGTH: 3392

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Dengue 1 virus strain WP

&lt;400&gt; SEQUENCE: 20

Met Asn Asn Gln Arg Lys Lys Thr Gly Arg Pro Ser Phe Asn Met Leu  
 1 5 10 15  
 Lys Arg Ala Arg Asn Arg Val Ser Thr Val Ser Gln Leu Ala Lys Arg  
 20 25 30  
 Phe Ser Lys Gly Leu Leu Ser Gly Gln Gly Pro Met Lys Leu Val Met  
 35 40 45  
 Ala Phe Ile Ala Phe Leu Arg Phe Leu Ala Ile Pro Pro Thr Ala Gly  
 50 55 60  
 Ile Leu Ala Arg Trp Gly Ser Phe Lys Lys Asn Gly Ala Ile Lys Val  
 65 70 75 80  
 Leu Arg Gly Phe Lys Lys Glu Ile Ser Asn Met Leu Asn Ile Met Asn  
 85 90 95  
 Arg Arg Lys Arg Ser Val Thr Met Leu Leu Met Leu Leu Pro Thr Ala  
 100 105 110  
 Leu Ala Phe His Leu Thr Thr Arg Gly Gly Glu Pro His Met Ile Val  
 115 120 125  
 Ser Lys Gln Glu Arg Gly Lys Ser Leu Leu Phe Lys Thr Ser Ala Gly  
 130 135 140  
 Val Asn Met Cys Thr Leu Ile Ala Met Asp Leu Gly Glu Leu Cys Glu  
 145 150 155 160  
 Asp Thr Met Thr Tyr Lys Cys Pro Arg Ile Thr Glu Thr Glu Pro Asp



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165					170					175					
Asp	Val	Asp	Cys	Trp	Cys	Asn	Ala	Thr	Glu	Thr	Trp	Val	Thr	Tyr	Gly
			180					185					190		
Thr	Cys	Ser	Gln	Thr	Gly	Glu	His	Arg	Arg	Asp	Lys	Arg	Ser	Val	Ala
		195					200					205			
Leu	Ala	Pro	His	Val	Gly	Leu	Gly	Leu	Glu	Thr	Arg	Thr	Glu	Thr	Trp
	210					215					220				
Met	Ser	Ser	Glu	Gly	Ala	Trp	Lys	Gln	Ile	Gln	Lys	Val	Glu	Thr	Trp
225					230					235					240
Ala	Leu	Arg	His	Pro	Gly	Phe	Thr	Val	Ile	Ala	Leu	Phe	Leu	Ala	His
			245						250					255	
Ala	Ile	Gly	Thr	Ser	Ile	Thr	Gln	Lys	Gly	Ile	Ile	Phe	Ile	Leu	Leu
		260						265					270		
Met	Leu	Val	Thr	Pro	Ser	Met	Ala	Met	Arg	Cys	Val	Gly	Ile	Gly	Asn
		275					280					285			
Arg	Asp	Phe	Val	Glu	Gly	Leu	Ser	Gly	Ala	Thr	Trp	Val	Asp	Val	Val
	290					295					300				
Leu	Glu	His	Gly	Ser	Cys	Val	Thr	Thr	Met	Ala	Lys	Asp	Lys	Pro	Thr
305					310					315					320
Leu	Asp	Ile	Glu	Leu	Leu	Lys	Thr	Glu	Val	Thr	Asn	Pro	Ala	Val	Leu
			325						330					335	
Arg	Lys	Leu	Cys	Ile	Glu	Ala	Lys	Ile	Ser	Asn	Thr	Thr	Thr	Asp	Ser
			340					345						350	
Arg	Cys	Pro	Thr	Gln	Gly	Glu	Ala	Thr	Leu	Val	Glu	Glu	Gln	Asp	Thr
		355					360					365			
Asn	Phe	Val	Cys	Arg	Arg	Thr	Phe	Val	Asp	Arg	Gly	Trp	Gly	Asn	Gly
	370					375					380				
Cys	Gly	Leu	Phe	Gly	Lys	Gly	Ser	Leu	Ile	Thr	Cys	Ala	Lys	Phe	Lys
385					390					395					400
Cys	Val	Thr	Lys	Leu	Glu	Gly	Lys	Ile	Val	Gln	Tyr	Glu	Asn	Leu	Lys
			405						410					415	
Tyr	Ser	Val	Ile	Val	Thr	Val	His	Thr	Gly	Asp	Gln	His	Gln	Val	Gly
			420					425					430		
Asn	Glu	Thr	Thr	Glu	His	Gly	Thr	Thr	Ala	Thr	Ile	Thr	Pro	Gln	Ala
		435					440					445			
Pro	Thr	Ser	Glu	Ile	Gln	Leu	Thr	Asp	Tyr	Gly	Ala	Leu	Thr	Leu	Asp
			450			455					460				
Cys	Ser	Pro	Arg	Thr	Gly	Leu	Asp	Phe	Asn	Glu	Met	Val	Leu	Leu	Thr
465					470					475					480
Met	Glu	Lys	Lys	Ser	Trp	Leu	Val	His	Lys	Gln	Trp	Phe	Leu	Asp	Leu
				485					490					495	
Pro	Leu	Pro	Trp	Thr	Ser	Gly	Ala	Ser	Thr	Ser	Gln	Glu	Thr	Trp	Asn
			500					505					510		
Arg	Gln	Asp	Leu	Leu	Val	Thr	Phe	Lys	Thr	Ala	His	Ala	Lys	Lys	Gln
		515					520					525			
Glu	Val	Val	Val	Leu	Gly	Ser	Gln	Glu	Gly	Ala	Met	His	Thr	Ala	Leu
		530				535					540				
Thr	Gly	Ala	Thr	Glu	Ile	Gln	Thr	Ser	Gly	Thr	Thr	Thr	Ile	Phe	Ala
545					550					555					560
Gly	His	Leu	Lys	Cys	Arg	Leu	Lys	Met	Asp	Lys	Leu	Thr	Leu	Lys	Gly
				565					570					575	
Met	Ser	Tyr	Val	Met	Cys	Thr	Gly	Ser	Phe	Lys	Leu	Glu	Lys	Glu	Val
			580					585					590		

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Ala Glu Thr Gln His Gly Thr Val Leu Val Gln Val Lys Tyr Glu Gly  
595 600 605

Thr Asp Ala Pro Cys Lys Ile Pro Phe Ser Ser Gln Asp Glu Lys Gly  
610 615 620

Val Thr Gln Asn Gly Arg Leu Ile Thr Ala Asn Pro Ile Val Thr Asp  
625 630 635 640

Lys Glu Lys Pro Val Asn Ile Glu Ala Glu Pro Pro Phe Gly Glu Ser  
645 650 655

Tyr Ile Val Val Gly Ala Gly Glu Lys Ala Leu Lys Leu Ser Trp Phe  
660 665 670

Lys Lys Gly Ser Ser Ile Gly Lys Met Phe Glu Ala Thr Ala Arg Gly  
675 680 685

Ala Arg Arg Met Ala Ile Leu Gly Asp Thr Ala Trp Asp Phe Gly Ser  
690 695 700

Ile Gly Gly Val Phe Thr Ser Val Gly Lys Leu Ile His Gln Ile Phe  
705 710 715 720

Gly Thr Ala Tyr Gly Val Leu Phe Ser Gly Val Ser Trp Thr Met Lys  
725 730 735

Ile Gly Ile Gly Ile Leu Leu Thr Trp Leu Gly Leu Asn Ser Arg Ser  
740 745 750

Thr Ser Leu Ser Met Thr Cys Ile Ala Val Gly Met Val Thr Leu Tyr  
755 760 765

Leu Gly Val Met Val Gln Ala Asp Ser Gly Cys Val Ile Asn Trp Lys  
770 775 780

Gly Arg Glu Leu Lys Cys Gly Ser Gly Ile Phe Val Thr Asn Glu Val  
785 790 795 800

His Thr Trp Thr Glu Gln Tyr Lys Phe Gln Ala Asp Ser Pro Lys Arg  
805 810 815

Leu Ser Ala Ala Ile Gly Lys Ala Trp Glu Glu Gly Val Cys Gly Ile  
820 825 830

Arg Ser Ala Thr Arg Leu Glu Asn Ile Met Trp Lys Gln Ile Ser Asn  
835 840 845

Glu Leu Asn His Ile Leu Leu Glu Asn Asp Met Lys Phe Thr Val Val  
850 855 860

Val Gly Asp Val Ser Gly Ile Leu Ala Gln Gly Lys Lys Met Ile Arg  
865 870 875 880

Pro Gln Pro Met Glu His Lys Tyr Ser Trp Lys Ser Trp Gly Lys Ala  
885 890 895

Lys Ile Ile Gly Ala Asp Val Gln Asn Thr Thr Phe Ile Ile Asp Gly  
900 905 910

Pro Asn Thr Pro Glu Cys Pro Asp Asn Gln Arg Ala Trp Asn Ile Trp  
915 920 925

Glu Val Glu Asp Tyr Gly Phe Gly Ile Phe Thr Thr Asn Ile Trp Leu  
930 935 940

Lys Leu Arg Asp Ser Tyr Thr Gln Val Cys Asp His Arg Leu Met Ser  
945 950 955 960

Ala Ala Ile Lys Asp Ser Lys Ala Val His Ala Asp Met Gly Tyr Trp  
965 970 975

Ile Glu Ser Glu Lys Asn Glu Thr Trp Lys Leu Ala Arg Ala Ser Phe  
980 985 990

Ile Glu Val Lys Thr Cys Ile Trp Pro Lys Ser His Thr Leu Trp Ser  
995 1000 1005

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Asn 1010	Gly	Val	Leu	Glu	Ser	Glu	Met	Ile	Ile	Pro	Lys	Ile	Tyr	Gly	Gly	
Pro	Ile	Ser	Gln	His	Asn	Tyr	Arg	Pro	Gly	Tyr	Phe	Thr	Gln	Thr	Ala	
1025					1030					1035					1040	
Gly	Pro	Trp	His	Leu	Gly	Lys	Leu	Glu	Leu	Asp	Phe	Asp	Leu	Cys	Glu	
				1045					1050					1055		
Gly	Thr	Thr	Val	Val	Val	Asp	Glu	His	Cys	Gly	Asn	Arg	Gly	Pro	Ser	
			1060						1065					1070		
Leu	Arg	Thr	Thr	Thr	Val	Thr	Gly	Lys	Thr	Ile	His	Glu	Trp	Cys	Cys	
		1075					1080					1085				
Arg	Ser	Cys	Thr	Leu	Pro	Pro	Leu	Arg	Phe	Lys	Gly	Glu	Asp	Gly	Cys	
1090							1095					1100				
Trp	Tyr	Gly	Met	Glu	Ile	Arg	Pro	Val	Lys	Glu	Lys	Glu	Glu	Asn	Leu	
1105					1110					1115					1120	
Val	Lys	Ser	Met	Val	Ser	Ala	Gly	Ser	Gly	Glu	Val	Asp	Ser	Phe	Ser	
				1125						1130					1135	
Leu	Gly	Leu	Leu	Cys	Ile	Ser	Ile	Met	Ile	Glu	Glu	Val	Met	Arg	Ser	
			1140					1145					1150			
Arg	Trp	Ser	Arg	Lys	Met	Leu	Met	Thr	Gly	Thr	Leu	Ala	Val	Phe	Leu	
		1155						1160					1165			
Leu	Leu	Thr	Met	Gly	Gln	Leu	Thr	Trp	Asn	Asp	Leu	Ile	Arg	Leu	Cys	
		1170					1175				1180					
Ile	Met	Val	Gly	Ala	Asn	Ala	Ser	Asp	Lys	Met	Gly	Met	Gly	Thr	Thr	
1185					1190						1195				1200	
Tyr	Leu	Ala	Leu	Met	Ala	Thr	Phe	Arg	Met	Arg	Pro	Met	Phe	Ala	Val	
				1205						1210				1215		
Gly	Leu	Leu	Phe	Arg	Arg	Leu	Thr	Ser	Arg	Glu	Val	Leu	Leu	Leu	Thr	
			1220					1225						1230		
Val	Gly	Leu	Ser	Leu	Val	Ala	Ser	Val	Glu	Leu	Pro	Asn	Ser	Leu	Glu	
		1235					1240					1245				
Glu	Leu	Gly	Asp	Gly	Leu	Ala	Met	Gly	Ile	Met	Met	Leu	Lys	Leu	Leu	
		1250				1255					1260					
Thr	Asp	Phe	Gln	Ser	His	Gln	Leu	Trp	Ala	Thr	Leu	Leu	Ser	Leu	Thr	
1265					1270					1275					1280	
Phe	Val	Lys	Thr	Thr	Phe	Ser	Leu	His	Tyr	Ala	Trp	Lys	Thr	Met	Ala	
				1285						1290				1295		
Met	Ile	Leu	Ser	Ile	Val	Ser	Leu	Phe	Pro	Leu	Cys	Leu	Ser	Thr	Thr	
			1300					1305						1310		
Ser	Gln	Lys	Thr	Thr	Trp	Leu	Pro	Val	Leu	Leu	Gly	Ser	Leu	Gly	Cys	
		1315					1320					1325				
Lys	Pro	Leu	Thr	Met	Phe	Leu	Ile	Thr	Glu	Asn	Lys	Ile	Trp	Gly	Arg	
1330						1335						1340				
Lys	Ser	Trp	Pro	Leu	Asn	Glu	Gly	Ile	Met	Ala	Val	Gly	Ile	Val	Ser	
1345					1350					1355					1360	
Ile	Leu	Leu	Ser	Ser	Leu	Leu	Lys	Asn	Asp	Val	Pro	Leu	Ala	Gly	Pro	
				1365					1370					1375		
Leu	Ile	Ala	Gly	Gly	Met	Leu	Ile	Ala	Cys	Tyr	Val	Ile	Ser	Gly	Ser	
			1380					1385						1390		
Ser	Ala	Asp	Leu	Ser	Leu	Glu	Lys	Ala	Ala	Glu	Val	Ser	Trp	Glu	Glu	
		1395					1400						1405			
Glu	Ala	Glu	His	Ser	Gly	Ala	Ser	His	Asn	Ile	Leu	Val	Glu	Val	Gln	
1410						1415					1420					
Asp	Asp	Gly	Thr	Met	Lys	Ile	Lys	Asp	Glu	Glu	Arg	Asp	Asp	Thr	Leu	

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1425	1430	1435	1440
Thr Ile Leu Leu Lys Ala Thr Leu Leu Ala Ile Ser Gly Val Tyr Pro 1445		1450	1455
Met Ser Ile Pro Ala Thr Leu Phe Val Trp Tyr Phe Trp Gln Lys Lys 1460		1465	1470
Lys Gln Arg Ser Gly Val Leu Trp Asp Thr Pro Ser Pro Pro Glu Val 1475		1480	1485
Glu Arg Ala Val Leu Asp Asp Gly Ile Tyr Arg Ile Leu Gln Arg Gly 1490		1495	1500
Leu Leu Gly Arg Ser Gln Val Gly Val Gly Val Phe Gln Glu Gly Val 1505		1510	1515
Phe His Thr Met Trp His Val Thr Arg Gly Ala Val Leu Met Tyr Gln 1525		1530	1535
Gly Lys Arg Leu Glu Pro Ser Trp Ala Ser Val Lys Lys Asp Leu Ile 1540		1545	1550
Ser Tyr Gly Gly Gly Trp Arg Phe Gln Gly Ser Trp Asn Ala Gly Glu 1555		1560	1565
Glu Val Gln Val Ile Ala Val Glu Pro Gly Lys Asn Pro Lys Asn Val 1570		1575	1580
Gln Thr Ala Pro Gly Thr Phe Lys Thr Pro Glu Gly Glu Val Gly Ala 1585		1590	1595
Ile Ala Leu Asp Phe Lys Pro Gly Thr Ser Gly Ser Pro Ile Val Asn 1605		1610	1615
Arg Glu Gly Lys Ile Val Gly Leu Tyr Gly Asn Gly Val Val Thr Thr 1620		1625	1630
Ser Gly Thr Tyr Val Ser Ala Ile Ala Gln Ala Lys Ala Ser Gln Glu 1635		1640	1645
Gly Pro Leu Pro Glu Ile Glu Asp Glu Val Phe Arg Lys Arg Asn Leu 1650		1655	1660
Thr Ile Met Asp Leu His Pro Gly Ser Gly Lys Thr Arg Arg Tyr Leu 1665		1670	1675
Pro Ala Ile Val Arg Glu Ala Ile Arg Arg Asn Val Arg Thr Leu Val 1685		1690	1695
Leu Ala Pro Thr Arg Val Val Ala Ser Glu Met Ala Glu Ala Leu Lys 1700		1705	1710
Gly Met Pro Ile Arg Tyr Gln Thr Thr Ala Val Lys Ser Glu His Thr 1715		1720	1725
Gly Lys Glu Ile Val Asp Leu Met Cys His Ala Thr Phe Thr Met Arg 1730		1735	1740
Leu Leu Ser Pro Val Arg Val Pro Asn Tyr Asn Met Ile Ile Met Asp 1745		1750	1755
Glu Ala His Phe Thr Asp Pro Ala Ser Ile Ala Ala Arg Gly Tyr Ile 1765		1770	1775
Ser Thr Arg Val Gly Met Gly Glu Ala Ala Ala Ile Phe Met Thr Ala 1780		1785	1790
Thr Pro Pro Gly Ser Val Glu Ala Phe Pro Gln Ser Asn Ala Val Ile 1795		1800	1805
Gln Asp Glu Glu Arg Asp Ile Pro Glu Arg Ser Trp Asn Ser Gly Tyr 1810		1815	1820
Asp Trp Ile Thr Asp Phe Pro Gly Lys Thr Val Trp Phe Val Pro Ser 1825		1830	1835
Ile Lys Ser Gly Asn Asp Ile Ala Asn Cys Leu Arg Lys Asn Gly Lys 1845		1850	1855

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Arg Val Val Gln Leu Ser Arg Lys Thr Phe Asp Thr Glu Tyr Gln Lys  
 1860 1865 1870  
 Thr Lys Asn Asn Asp Trp Asp Tyr Val Val Thr Thr Asp Ile Ser Glu  
 1875 1880 1885  
 Met Gly Ala Asn Phe Arg Ala Asp Arg Val Ile Asp Pro Arg Arg Cys  
 1890 1895 1900  
 Leu Lys Pro Val Ile Leu Lys Asp Gly Pro Glu Arg Val Ile Leu Ala  
 1905 1910 1915 1920  
 Gly Pro Met Pro Val Thr Val Ala Ser Ala Ala Gln Arg Arg Gly Arg  
 1925 1930 1935  
 Ile Gly Arg Asn Gln Asn Lys Glu Gly Asp Gln Tyr Ile Tyr Met Gly  
 1940 1945 1950  
 Gln Pro Leu Asn Asn Asp Glu Asp His Ala His Trp Thr Glu Ala Lys  
 1955 1960 1965  
 Met Leu Leu Asp Asn Ile Asn Thr Pro Glu Gly Ile Ile Pro Ala Leu  
 1970 1975 1980  
 Phe Glu Pro Glu Arg Glu Lys Ser Ala Ala Ile Asp Gly Glu Tyr Arg  
 1985 1990 1995 2000  
 Leu Arg Gly Glu Ala Arg Lys Thr Phe Val Glu Leu Met Arg Arg Gly  
 2005 2010 2015  
 Asp Leu Pro Val Trp Leu Ser Tyr Lys Val Ala Ser Glu Gly Phe Gln  
 2020 2025 2030  
 Tyr Ser Asp Arg Arg Trp Cys Phe Asp Gly Glu Arg Asn Asn Gln Val  
 2035 2040 2045  
 Leu Glu Glu Asn Met Asp Val Glu Ile Trp Thr Lys Glu Gly Glu Arg  
 2050 2055 2060  
 Lys Lys Leu Arg Pro Arg Trp Leu Asp Ala Arg Thr Tyr Ser Asp Pro  
 2065 2070 2075 2080  
 Leu Ala Leu Arg Glu Phe Lys Glu Phe Ala Ala Gly Arg Arg Ser Val  
 2085 2090 2095  
 Ser Gly Asp Leu Ile Leu Glu Ile Gly Lys Leu Pro Gln His Leu Thr  
 2100 2105 2110  
 Gln Arg Ala Gln Asn Ala Leu Asp Asn Leu Val Met Leu His Asn Ser  
 2115 2120 2125  
 Glu Gln Gly Gly Lys Ala Tyr Arg His Ala Met Glu Glu Leu Pro Asp  
 2130 2135 2140  
 Thr Ile Glu Thr Leu Met Leu Leu Ala Leu Ile Ala Val Leu Thr Gly  
 2145 2150 2155 2160  
 Gly Val Thr Leu Phe Phe Leu Ser Gly Arg Gly Leu Gly Lys Thr Ser  
 2165 2170 2175  
 Ile Gly Leu Leu Cys Val Ile Ala Ser Ser Ala Leu Leu Trp Met Ala  
 2180 2185 2190  
 Ser Val Glu Pro His Trp Ile Ala Ala Ser Ile Ile Leu Glu Phe Phe  
 2195 2200 2205  
 Leu Met Val Leu Leu Ile Pro Glu Pro Asp Arg Gln Arg Thr Pro Gln  
 2210 2215 2220  
 Asp Asn Gln Leu Ala Tyr Val Val Ile Gly Leu Leu Phe Met Ile Leu  
 2225 2230 2235 2240  
 Thr Ala Ala Ala Asn Glu Met Gly Leu Leu Glu Thr Thr Lys Lys Asp  
 2245 2250 2255  
 Leu Gly Ile Gly His Ala Ala Ala Glu Asn His His His Ala Ala Met  
 2260 2265 2270

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Leu Asp Val Asp Leu His Pro Ala Ser Ala Trp Thr Leu Tyr Ala Val  
 2275 2280 2285  
 Ala Thr Thr Ile Ile Thr Pro Met Met Arg His Thr Ile Glu Asn Thr  
 2290 2295 2300  
 Thr Ala Asn Ile Ser Leu Thr Ala Ile Ala Asn Gln Ala Ala Ile Leu  
 2305 2310 2315 2320  
 Met Gly Leu Asp Lys Gly Trp Pro Ile Ser Lys Met Asp Ile Gly Val  
 2325 2330 2335  
 Pro Leu Leu Ala Leu Gly Cys Tyr Ser Gln Val Asn Pro Leu Thr Leu  
 2340 2345 2350  
 Thr Ala Ala Val Phe Met Leu Val Ala His Tyr Ala Ile Ile Gly Pro  
 2355 2360 2365  
 Gly Leu Gln Ala Lys Ala Thr Arg Glu Ala Gln Lys Arg Thr Ala Ala  
 2370 2375 2380  
 Gly Ile Met Lys Asn Pro Thr Val Asp Gly Ile Val Ala Ile Asp Leu  
 2385 2390 2395 2400  
 Asp Pro Val Val Tyr Asp Ala Lys Phe Glu Lys Gln Leu Gly Gln Ile  
 2405 2410 2415  
 Met Leu Leu Ile Leu Cys Thr Ser Gln Ile Leu Leu Met Arg Thr Thr  
 2420 2425 2430  
 Trp Ala Leu Cys Glu Ser Ile Thr Leu Ala Thr Gly Pro Leu Thr Thr  
 2435 2440 2445  
 Leu Trp Glu Gly Ser Pro Gly Lys Phe Trp Asn Thr Thr Ile Ala Val  
 2450 2455 2460  
 Ser Met Ala Asn Ile Phe Arg Gly Ser Tyr Leu Ala Gly Ala Gly Leu  
 2465 2470 2475 2480  
 Ala Phe Ser Leu Met Lys Ser Leu Gly Gly Gly Arg Arg Gly Thr Gly  
 2485 2490 2495  
 Ala Gln Gly Glu Thr Leu Gly Glu Lys Trp Lys Arg Gln Leu Asn Gln  
 2500 2505 2510  
 Leu Ser Lys Ser Glu Phe Asn Thr Tyr Lys Arg Ser Gly Ile Ile Glu  
 2515 2520 2525  
 Val Asp Arg Ser Glu Ala Lys Glu Gly Leu Lys Arg Gly Glu Pro Thr  
 2530 2535 2540  
 Lys His Ala Val Ser Arg Gly Thr Ala Lys Leu Arg Trp Phe Val Glu  
 2545 2550 2555 2560  
 Arg Asn Leu Val Lys Pro Glu Gly Lys Val Ile Asp Leu Gly Cys Gly  
 2565 2570 2575  
 Arg Gly Gly Trp Ser Tyr Tyr Cys Ala Gly Leu Lys Lys Val Thr Glu  
 2580 2585 2590  
 Val Lys Gly Tyr Thr Lys Gly Gly Pro Gly His Glu Glu Pro Ile Pro  
 2595 2600 2605  
 Met Ala Thr Tyr Gly Trp Asn Leu Val Lys Leu Tyr Ser Gly Lys Asp  
 2610 2615 2620  
 Val Phe Phe Thr Pro Pro Glu Lys Cys Asp Thr Leu Leu Cys Asp Ile  
 2625 2630 2635 2640  
 Gly Glu Ser Ser Pro Asn Pro Thr Ile Glu Glu Gly Arg Thr Leu Arg  
 2645 2650 2655  
 Val Leu Lys Met Val Glu Pro Trp Leu Arg Gly Asn Gln Phe Cys Ile  
 2660 2665 2670  
 Lys Ile Leu Asn Pro Tyr Met Pro Ser Val Val Glu Thr Leu Glu Gln  
 2675 2680 2685  
 Met Gln Arg Lys His Gly Gly Met Leu Val Arg Asn Pro Leu Ser Arg

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2690	2695	2700
Asn Ser Thr His Glu Met Tyr Trp Val Ser Cys Gly Thr Gly Asn Ile 2705	2710	2715 2720
Val Ser Ala Val Asn Met Thr Ser Arg Met Leu Leu Asn Arg Phe Thr 2725	2730	2735
Met Ala His Arg Lys Pro Thr Tyr Glu Arg Asp Val Asp Leu Gly Ala 2740	2745	2750
Gly Thr Arg His Val Ala Val Glu Pro Glu Val Ala Asn Leu Asp Ile 2755	2760	2765
Ile Gly Gln Arg Ile Glu Asn Ile Lys Asn Gly His Lys Ser Thr Trp 2770	2775	2780
His Tyr Asp Glu Asp Asn Pro Tyr Lys Thr Trp Ala Tyr His Gly Ser 2785	2790	2795 2800
Tyr Glu Val Lys Pro Ser Gly Ser Ala Ser Ser Met Val Asn Gly Val 2805	2810	2815
Val Arg Leu Leu Thr Lys Pro Trp Asp Val Ile Pro Met Val Thr Gln 2820	2825	2830
Ile Ala Met Thr Asp Thr Thr Pro Phe Gly Gln Gln Arg Val Phe Lys 2835	2840	2845
Glu Lys Val Asp Thr Arg Thr Pro Lys Ala Lys Arg Gly Thr Ala Gln 2850	2855	2860
Ile Met Glu Val Thr Ala Arg Trp Leu Trp Gly Phe Leu Ser Arg Asn 2865	2870	2875 2880
Lys Lys Pro Arg Ile Cys Thr Arg Glu Glu Phe Thr Arg Lys Val Arg 2885	2890	2895
Ser Asn Ala Ala Ile Gly Ala Val Phe Val Asp Glu Asn Gln Trp Asn 2900	2905	2910
Ser Ala Lys Glu Ala Val Glu Asp Glu Arg Phe Trp Asp Leu Val His 2915	2920	2925
Arg Glu Arg Glu Leu His Lys Gln Gly Lys Cys Ala Thr Cys Val Tyr 2930	2935	2940
Asn Met Met Gly Lys Arg Glu Lys Lys Leu Gly Glu Phe Gly Lys Ala 2945	2950	2955 2960
Lys Gly Ser Arg Ala Ile Trp Tyr Met Trp Leu Gly Ala Arg Phe Leu 2965	2970	2975
Glu Phe Glu Ala Leu Gly Phe Met Asn Glu Asp His Trp Phe Ser Arg 2980	2985	2990
Glu Asn Ser Leu Ser Gly Val Glu Gly Glu Gly Leu His Lys Leu Gly 2995	3000	3005
Tyr Ile Leu Arg Asp Ile Ser Lys Ile Pro Gly Gly Asn Met Tyr Ala 3010	3015	3020
Asp Asp Thr Ala Gly Trp Asp Thr Arg Ile Thr Glu Asp Asp Leu Gln 3025	3030	3035 3040
Asn Glu Ala Lys Ile Thr Asp Ile Met Glu Pro Glu His Ala Leu Leu 3045	3050	3055
Ala Thr Ser Ile Phe Lys Leu Thr Tyr Gln Asn Lys Val Val Arg Val 3060	3065	3070
Gln Arg Pro Ala Lys Asn Gly Thr Val Met Asp Val Ile Ser Arg Arg 3075	3080	3085
Asp Gln Arg Gly Ser Gly Gln Val Gly Thr Tyr Gly Leu Asn Thr Phe 3090	3095	3100
Thr Asn Met Glu Ala Gln Leu Ile Arg Gln Met Glu Ser Glu Gly Ile 3105	3110	3115 3120

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Phe Ser Pro Ser Glu Leu Glu Thr Pro Asn Leu Ala Glu Arg Val Leu  
 3125 3130 3135  
 Asp Trp Leu Lys Lys His Gly Thr Glu Arg Leu Lys Arg Met Ala Ile  
 3140 3145 3150  
 Ser Gly Asp Asp Cys Val Val Lys Pro Ile Asp Asp Arg Phe Ala Thr  
 3155 3160 3165  
 Ala Leu Thr Ala Leu Asn Asp Met Gly Lys Val Arg Lys Asp Ile Pro  
 3170 3175 3180  
 Gln Trp Glu Pro Ser Lys Gly Trp Asn Asp Trp Gln Gln Val Pro Phe  
 3185 3190 3195 3200  
 Cys Ser His His Phe His Gln Leu Ile Met Lys Asp Gly Arg Glu Ile  
 3205 3210 3215  
 Val Val Pro Cys Arg Asn Gln Asp Glu Leu Val Gly Arg Ala Arg Val  
 3220 3225 3230  
 Ser Gln Gly Ala Gly Trp Ser Leu Arg Glu Thr Ala Cys Leu Gly Lys  
 3235 3240 3245  
 Ser Tyr Ala Gln Met Trp Gln Leu Met Tyr Phe His Arg Arg Asp Leu  
 3250 3255 3260  
 Arg Leu Ala Ala Asn Ala Ile Cys Ser Ala Val Pro Val Asp Trp Val  
 3265 3270 3275 3280  
 Pro Thr Ser Arg Thr Thr Trp Ser Ile His Ala His His Gln Trp Met  
 3285 3290 3295  
 Thr Thr Glu Asp Met Leu Ser Val Trp Asn Arg Val Trp Ile Glu Glu  
 3300 3305 3310  
 Asn Pro Trp Met Glu Asp Lys Thr His Val Ser Ser Trp Glu Asp Val  
 3315 3320 3325  
 Pro Tyr Leu Gly Lys Arg Glu Asp Arg Trp Cys Gly Ser Leu Ile Gly  
 3330 3335 3340  
 Leu Thr Ala Arg Ala Thr Trp Ala Thr Asn Ile Gln Val Ala Ile Asn  
 3345 3350 3355 3360  
 Gln Val Arg Arg Leu Ile Gly Asn Glu Asn Tyr Leu Asp Phe Met Thr  
 3365 3370 3375  
 Ser Met Lys Arg Phe Lys Asn Glu Ser Asp Pro Glu Gly Ala Leu Trp  
 3380 3385 3390

&lt;210&gt; SEQ ID NO 21

&lt;211&gt; LENGTH: 3391

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Dengue 2 virus strain NGC

&lt;400&gt; SEQUENCE: 21

Met Asn Asn Gln Arg Lys Lys Ala Arg Asn Thr Pro Phe Asn Met Leu  
 1 5 10 15  
 Lys Arg Glu Arg Asn Arg Val Ser Thr Val Gln Gln Leu Thr Lys Arg  
 20 25 30  
 Phe Ser Leu Gly Met Leu Gln Gly Arg Gly Pro Leu Lys Leu Phe Met  
 35 40 45  
 Ala Leu Val Ala Phe Leu Arg Phe Leu Thr Ile Pro Pro Thr Ala Gly  
 50 55 60  
 Ile Leu Lys Arg Trp Gly Thr Ile Lys Lys Ser Lys Ala Ile Asn Val  
 65 70 75 80  
 Leu Arg Gly Phe Arg Lys Glu Ile Gly Arg Met Leu Asn Ile Leu Asn  
 85 90 95  
 Arg Arg Arg Arg Thr Ala Gly Met Ile Ile Met Leu Ile Pro Thr Val



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100					105					110					
Met	Ala	Phe	His	Leu	Thr	Thr	Arg	Asn	Gly	Glu	Pro	His	Met	Ile	Val
	115						120					125			
Ser	Arg	Gln	Glu	Lys	Gly	Lys	Ser	Leu	Leu	Phe	Lys	Thr	Glu	Asp	Gly
	130					135					140				
Val	Asn	Met	Cys	Thr	Leu	Met	Ala	Met	Asp	Leu	Gly	Glu	Leu	Cys	Glu
145					150					155					160
Asp	Thr	Ile	Thr	Tyr	Lys	Cys	Pro	Phe	Leu	Arg	Gln	Asn	Glu	Pro	Glu
				165					170					175	
Asp	Ile	Asp	Cys	Trp	Cys	Asn	Ser	Thr	Ser	Thr	Trp	Val	Thr	Tyr	Gly
			180					185					190		
Thr	Cys	Thr	Thr	Thr	Gly	Glu	His	Arg	Arg	Glu	Lys	Arg	Ser	Val	Ala
							200					205			
Leu	Val	Pro	His	Val	Gly	Met	Gly	Leu	Glu	Thr	Arg	Thr	Glu	Thr	Trp
	210					215					220				
Met	Ser	Ser	Glu	Gly	Ala	Trp	Lys	His	Ala	Gln	Arg	Ile	Glu	Thr	Trp
225					230					235					240
Ile	Leu	Arg	His	Pro	Gly	Phe	Thr	Ile	Met	Ala	Ala	Ile	Leu	Ala	Tyr
				245					250					255	
Thr	Ile	Gly	Thr	Thr	His	Phe	Gln	Arg	Ala	Leu	Ile	Phe	Ile	Leu	Leu
				260				265					270		
Thr	Ala	Val	Ala	Pro	Ser	Met	Thr	Met	Arg	Cys	Ile	Gly	Ile	Ser	Asn
		275					280					285			
Arg	Asp	Phe	Val	Glu	Gly	Val	Ser	Gly	Gly	Ser	Trp	Val	Asp	Ile	Val
	290					295					300				
Leu	Glu	His	Gly	Ser	Cys	Val	Thr	Thr	Met	Ala	Lys	Asn	Lys	Pro	Thr
305					310					315					320
Leu	Asp	Phe	Glu	Leu	Ile	Lys	Thr	Glu	Ala	Lys	Gln	Pro	Ala	Thr	Leu
				325					330					335	
Arg	Lys	Tyr	Cys	Ile	Glu	Ala	Lys	Leu	Thr	Asn	Thr	Thr	Thr	Asp	Ser
			340					345						350	
Arg	Cys	Pro	Thr	Gln	Gly	Glu	Pro	Ser	Leu	Asn	Glu	Glu	Gln	Asp	Lys
		355					360					365			
Arg	Phe	Val	Cys	Lys	His	Ser	Met	Val	Asp	Arg	Gly	Trp	Gly	Asn	Gly
	370					375					380				
Cys	Gly	Leu	Phe	Gly	Lys	Gly	Gly	Ile	Val	Thr	Cys	Ala	Met	Phe	Thr
385					390					395					400
Cys	Lys	Lys	Asn	Met	Lys	Gly	Lys	Val	Val	Gln	Pro	Glu	Asn	Leu	Glu
			405						410					415	
Tyr	Thr	Ile	Val	Ile	Thr	Pro	His	Ser	Gly	Glu	Glu	His	Ala	Val	Gly
			420					425					430		
Asn	Asp	Thr	Gly	Lys	His	Gly	Lys	Glu	Ile	Lys	Ile	Thr	Pro	Gln	Ser
		435					440					445			
Ser	Ile	Thr	Glu	Ala	Glu	Leu	Thr	Gly	Tyr	Gly	Thr	Val	Thr	Met	Glu
	450					455					460				
Cys	Ser	Pro	Arg	Thr	Gly	Leu	Asp	Phe	Asn	Glu	Met	Val	Leu	Leu	Gln
465					470					475					480
Met	Glu	Asn	Lys	Ala	Trp	Leu	Val	His	Arg	Gln	Trp	Phe	Leu	Asp	Leu
				485					490					495	
Pro	Leu	Pro	Trp	Leu	Pro	Gly	Ala	Asp	Thr	Gln	Gly	Ser	Asn	Trp	Ile
			500					505					510		
Gln	Lys	Glu	Thr	Leu	Val	Thr	Phe	Lys	Asn	Pro	His	Ala	Lys	Lys	Gln
		515					520					525			

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Asp Val Val Val Leu Gly Ser Gln Glu Gly Ala Met His Thr Ala Leu  
 530 535 540

Thr Gly Ala Thr Glu Ile Gln Met Ser Ser Gly Asn Leu Leu Phe Thr  
 545 550 555 560

Gly His Leu Lys Cys Arg Leu Arg Met Asp Lys Leu Gln Leu Lys Gly  
 565 570 575

Met Ser Tyr Ser Met Cys Thr Gly Lys Phe Lys Val Val Lys Glu Ile  
 580 585 590

Ala Glu Thr Gln His Gly Thr Ile Val Ile Arg Val Gln Tyr Glu Gly  
 595 600 605

Asp Gly Ser Pro Cys Lys Ile Pro Phe Glu Ile Met Asp Leu Glu Lys  
 610 615 620

Arg His Val Leu Gly Arg Leu Ile Thr Val Asn Pro Ile Val Thr Glu  
 625 630 635 640

Lys Asp Ser Pro Val Asn Ile Glu Ala Glu Pro Pro Phe Gly Asp Ser  
 645 650 655

Tyr Ile Ile Ile Gly Val Glu Pro Gly Gln Leu Lys Leu Asn Trp Phe  
 660 665 670

Lys Lys Gly Ser Ser Ile Gly Gln Met Ile Glu Thr Thr Met Arg Gly  
 675 680 685

Ala Lys Arg Met Ala Ile Leu Gly Asp Thr Ala Trp Asp Phe Gly Ser  
 690 695 700

Leu Gly Gly Val Phe Thr Ser Ile Gly Lys Ala Leu His Gln Val Phe  
 705 710 715 720

Gly Ala Ile Tyr Gly Ala Ala Phe Ser Gly Val Ser Trp Thr Met Lys  
 725 730 735

Ile Leu Ile Gly Val Ile Ile Thr Trp Ile Gly Met Asn Ser Arg Ser  
 740 745 750

Thr Ser Leu Ser Val Ser Leu Val Leu Val Gly Val Val Thr Leu Tyr  
 755 760 765

Leu Gly Val Met Val Gln Ala Asp Ser Gly Cys Val Val Ser Trp Lys  
 770 775 780

Asn Lys Glu Leu Lys Cys Gly Ser Gly Ile Phe Ile Thr Asp Asn Val  
 785 790 795 800

His Thr Trp Thr Glu Gln Tyr Lys Phe Gln Pro Glu Ser Pro Ser Lys  
 805 810 815

Leu Ala Ser Ala Ile Gln Lys Ala His Glu Glu Gly Ile Cys Gly Ile  
 820 825 830

Arg Ser Val Thr Arg Leu Glu Asn Leu Met Trp Lys Gln Ile Thr Pro  
 835 840 845

Glu Leu Asn His Ile Leu Ser Glu Asn Glu Val Lys Leu Thr Ile Met  
 850 855 860

Thr Gly Asp Ile Lys Gly Ile Met Gln Ala Gly Lys Arg Ser Leu Gln  
 865 870 875 880

Pro Gln Pro Thr Glu Leu Lys Tyr Ser Trp Lys Thr Trp Gly Lys Ala  
 885 890 895

Lys Met Leu Ser Thr Glu Ser His Asn Gln Thr Phe Leu Ile Asp Gly  
 900 905 910

Pro Glu Thr Ala Glu Cys Pro Asn Thr Asn Arg Ala Trp Asn Ser Leu  
 915 920 925

Glu Val Glu Asp Tyr Gly Phe Gly Val Phe Thr Thr Asn Ile Trp Leu  
 930 935 940

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Lys	Leu	Arg	Glu	Lys	Gln	Asp	Val	Phe	Cys	Asp	Ser	Lys	Leu	Met	Ser	945	950	955	960
Ala	Ala	Ile	Lys	Asp	Asn	Arg	Ala	Val	His	Ala	Asp	Met	Gly	Tyr	Trp	965	970	975	
Ile	Glu	Ser	Ala	Leu	Asn	Asp	Thr	Trp	Lys	Ile	Glu	Lys	Ala	Ser	Phe	980	985	990	
Ile	Glu	Val	Lys	Ser	Cys	His	Trp	Pro	Lys	Ser	His	Thr	Leu	Trp	Ser	995	1000	1005	
Asn	Gly	Val	Leu	Glu	Ser	Glu	Met	Ile	Ile	Pro	Lys	Asn	Phe	Ala	Gly	1010	1015	1020	
Pro	Val	Ser	Gln	His	Asn	Tyr	Arg	Pro	Gly	Tyr	His	Thr	Gln	Thr	Ala	1025	1030	1035	1040
Gly	Pro	Trp	His	Leu	Gly	Lys	Leu	Glu	Met	Asp	Phe	Asp	Phe	Cys	Glu	1045	1050	1055	
Gly	Thr	Thr	Val	Val	Val	Thr	Glu	Asp	Cys	Gly	Asn	Arg	Gly	Pro	Ser	1060	1065	1070	
Leu	Arg	Thr	Thr	Thr	Ala	Ser	Gly	Lys	Leu	Ile	Thr	Glu	Trp	Cys	Cys	1075	1080	1085	
Arg	Ser	Cys	Thr	Leu	Pro	Pro	Leu	Arg	Tyr	Arg	Gly	Glu	Asp	Gly	Cys	1090	1095	1100	
Trp	Tyr	Gly	Met	Glu	Ile	Arg	Pro	Leu	Lys	Glu	Lys	Glu	Glu	Asn	Leu	1105	1110	1115	1120
Val	Asn	Ser	Leu	Val	Thr	Ala	Gly	His	Gly	Gln	Ile	Asp	Asn	Phe	Ser	1125	1130	1135	
Leu	Gly	Val	Leu	Gly	Met	Ala	Leu	Phe	Leu	Glu	Glu	Met	Leu	Arg	Thr	1140	1145	1150	
Arg	Val	Gly	Thr	Lys	His	Ala	Ile	Leu	Leu	Val	Ala	Val	Ser	Phe	Val	1155	1160	1165	
Thr	Leu	Ile	Thr	Gly	Asn	Met	Ser	Phe	Arg	Asp	Leu	Gly	Arg	Val	Met	1170	1175	1180	
Val	Met	Val	Gly	Ala	Thr	Met	Thr	Asp	Asp	Ile	Gly	Met	Gly	Val	Thr	1185	1190	1195	1200
Tyr	Leu	Ala	Leu	Leu	Ala	Ala	Phe	Lys	Val	Arg	Pro	Thr	Phe	Ala	Ala	1205	1210	1215	
Gly	Leu	Leu	Leu	Arg	Lys	Leu	Thr	Ser	Lys	Glu	Leu	Met	Met	Thr	Thr	1220	1225	1230	
Ile	Gly	Ile	Val	Leu	Leu	Ser	Gln	Ser	Thr	Ile	Pro	Glu	Thr	Ile	Leu	1235	1240	1245	
Glu	Leu	Thr	Asp	Ala	Leu	Ala	Leu	Gly	Met	Met	Val	Leu	Lys	Met	Val	1250	1255	1260	
Arg	Lys	Met	Glu	Lys	Tyr	Gln	Leu	Ala	Val	Thr	Ile	Met	Ala	Ile	Leu	1265	1270	1275	1280
Cys	Val	Pro	Asn	Ala	Val	Ile	Leu	Gln	Asn	Ala	Trp	Lys	Val	Ser	Cys	1285	1290	1295	
Thr	Ile	Leu	Ala	Val	Val	Ser	Val	Ser	Pro	Leu	Phe	Leu	Thr	Ser	Ser	1300	1305	1310	
Gln	Gln	Lys	Ala	Asp	Trp	Ile	Pro	Leu	Ala	Leu	Thr	Ile	Lys	Gly	Leu	1315	1320	1325	
Asn	Pro	Thr	Ala	Ile	Phe	Leu	Thr	Thr	Leu	Ser	Arg	Thr	Asn	Lys	Lys	1330	1335	1340	
Arg	Ser	Trp	Pro	Leu	Asn	Glu	Ala	Ile	Met	Ala	Val	Gly	Met	Val	Ser	1345	1350	1355	1360
Ile	Leu	Ala	Ser	Ser	Leu	Leu	Lys	Asn	Asp	Ile	Pro	Met	Thr	Gly	Pro				

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1365	1370	1375
Leu Val Ala Gly Gly Leu Leu Thr Val Cys Tyr Val Leu Thr Gly Arg 1380	1385	1390
Ser Ala Asp Leu Glu Leu Glu Arg Ala Ala Asp Val Lys Trp Glu Asp 1395	1400	1405
Gln Ala Glu Ile Ser Gly Ser Ser Pro Ile Leu Ser Ile Thr Ile Ser 1410	1415	1420
Glu Asp Gly Ser Met Ser Ile Lys Asn Glu Glu Glu Glu Gln Thr Leu 1425	1430	1435
Thr Ile Leu Ile Arg Thr Gly Leu Leu Val Ile Ser Gly Leu Phe Pro 1445	1450	1455
Val Ser Ile Pro Ile Thr Ala Ala Ala Trp Tyr Leu Trp Glu Val Lys 1460	1465	1470
Lys Gln Arg Ala Gly Val Leu Trp Asp Val Pro Ser Pro Pro Pro Val 1475	1480	1485
Gly Lys Ala Glu Leu Glu Asp Gly Ala Tyr Arg Ile Lys Gln Lys Gly 1490	1495	1500
Ile Leu Gly Tyr Ser Gln Ile Gly Ala Gly Val Tyr Lys Glu Gly Thr 1505	1510	1515
Phe His Thr Met Trp His Val Thr Arg Gly Ala Val Leu Met His Lys 1525	1530	1535
Gly Lys Arg Ile Glu Pro Ser Trp Ala Asp Val Lys Lys Asp Leu Ile 1540	1545	1550
Ser Tyr Gly Gly Gly Trp Lys Leu Glu Gly Glu Trp Lys Glu Gly Glu 1555	1560	1565
Glu Val Gln Val Leu Ala Leu Glu Pro Gly Lys Asn Pro Arg Ala Val 1570	1575	1580
Gln Thr Lys Pro Gly Leu Phe Lys Thr Asn Ala Gly Thr Ile Gly Ala 1585	1590	1595
Val Ser Leu Asp Phe Ser Pro Gly Thr Ser Gly Ser Pro Ile Ile Asp 1605	1610	1615
Lys Lys Gly Lys Val Val Gly Leu Tyr Gly Asn Gly Val Val Thr Arg 1620	1625	1630
Ser Gly Ala Tyr Val Ser Ala Ile Ala Gln Thr Glu Lys Ser Ile Glu 1635	1640	1645
Asp Asn Pro Glu Ile Glu Asp Asp Ile Phe Arg Lys Arg Lys Leu Thr 1650	1655	1660
Ile Met Asp Leu His Pro Gly Ala Gly Lys Thr Lys Arg Tyr Leu Pro 1665	1670	1675
Ala Ile Val Arg Glu Ala Ile Lys Arg Gly Leu Arg Thr Leu Ile Leu 1685	1690	1695
Ala Pro Thr Arg Val Val Ala Ala Glu Met Glu Glu Ala Leu Arg Gly 1700	1705	1710
Leu Pro Ile Arg Tyr Gln Thr Pro Ala Ile Arg Ala Glu His Thr Gly 1715	1720	1725
Arg Glu Ile Val Asp Leu Met Cys His Ala Thr Phe Thr Met Arg Leu 1730	1735	1740
Leu Ser Pro Val Arg Val Pro Asn Tyr Asn Leu Ile Ile Met Asp Glu 1745	1750	1755
Ala His Phe Thr Asp Pro Ala Ser Ile Ala Ala Arg Gly Tyr Ile Ser 1765	1770	1775
Thr Arg Val Glu Met Gly Glu Ala Ala Gly Ile Phe Met Thr Ala Thr 1780	1785	1790

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Pro Pro Gly Ser Arg Asp Pro Phe Pro Gln Ser Asn Ala Pro Ile Met  
 1795 1800 1805

Asp Glu Glu Arg Glu Ile Pro Glu Arg Ser Trp Ser Ser Gly His Glu  
 1810 1815 1820

Trp Val Thr Asp Phe Lys Gly Lys Thr Val Trp Phe Val Pro Ser Ile  
 1825 1830 1835 1840

Lys Ala Gly Asn Asp Ile Ala Ala Cys Leu Arg Lys Asn Gly Lys Lys  
 1845 1850 1855

Val Ile Gln Leu Ser Arg Lys Thr Phe Asp Ser Glu Tyr Val Lys Thr  
 1860 1865 1870

Arg Thr Asn Asp Trp Asp Phe Val Val Thr Thr Asp Ile Ser Glu Met  
 1875 1880 1885

Gly Ala Asn Phe Lys Ala Glu Arg Val Ile Asp Pro Arg Arg Cys Met  
 1890 1895 1900

Lys Pro Val Ile Leu Thr Asp Gly Glu Glu Arg Val Ile Leu Ala Gly  
 1905 1910 1915 1920

Pro Met Pro Val Thr His Ser Ser Ala Ala Gln Arg Arg Gly Arg Ile  
 1925 1930 1935

Gly Arg Asn Pro Lys Asn Glu Asn Asp Gln Tyr Ile Tyr Met Gly Glu  
 1940 1945 1950

Pro Leu Glu Asn Asp Glu Asp Cys Ala His Trp Lys Glu Ala Lys Met  
 1955 1960 1965

Leu Leu Asp Asn Ile Asn Thr Pro Glu Gly Ile Ile Pro Ser Met Phe  
 1970 1975 1980

Glu Pro Glu Arg Glu Lys Val Asp Ala Ile Asp Gly Glu Tyr Arg Leu  
 1985 1990 1995 2000

Arg Gly Glu Ala Arg Lys Thr Phe Val Asp Leu Met Arg Arg Gly Asp  
 2005 2010 2015

Leu Pro Val Trp Leu Ala Tyr Arg Val Ala Ala Glu Gly Ile Asn Tyr  
 2020 2025 2030

Ala Asp Arg Arg Trp Cys Phe Asp Gly Ile Lys Asn Asn Gln Ile Leu  
 2035 2040 2045

Glu Glu Asn Val Glu Val Glu Ile Trp Thr Lys Glu Gly Glu Arg Lys  
 2050 2055 2060

Lys Leu Lys Pro Arg Trp Leu Asp Ala Arg Ile Tyr Ser Asp Pro Leu  
 2065 2070 2075 2080

Thr Leu Lys Glu Phe Lys Glu Phe Ala Ala Gly Arg Lys Ser Leu Thr  
 2085 2090 2095

Leu Asn Leu Ile Thr Glu Met Gly Arg Leu Pro Thr Phe Met Thr Gln  
 2100 2105 2110

Lys Ala Arg Asp Ala Leu Asp Asn Leu Ala Val Leu His Thr Ala Glu  
 2115 2120 2125

Ala Gly Gly Arg Ala Tyr Asn His Ala Leu Ser Glu Leu Pro Glu Thr  
 2130 2135 2140

Leu Glu Thr Leu Leu Leu Leu Thr Leu Leu Ala Thr Val Thr Gly Gly  
 2145 2150 2155 2160

Ile Phe Leu Phe Leu Met Ser Gly Arg Gly Ile Gly Lys Met Thr Leu  
 2165 2170 2175

Gly Met Cys Cys Ile Ile Thr Ala Ser Ile Leu Leu Trp Tyr Ala Gln  
 2180 2185 2190

Ile Gln Pro His Trp Ile Ala Ala Ser Ile Ile Leu Glu Phe Phe Leu  
 2195 2200 2205

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Ile Val Leu Leu Ile Pro Glu Pro Glu Lys Gln Arg Thr Pro Gln Asp 2210 2215 2220
Asn Gln Leu Thr Tyr Val Val Ile Ala Ile Leu Thr Val Val Ala Ala 2225 2230 2235 2240
Thr Met Ala Asn Glu Met Gly Phe Leu Glu Lys Thr Lys Lys Asp Leu 2245 2250 2255
Gly Leu Gly Ser Ile Thr Thr Gln Gln Pro Glu Ser Asn Ile Leu Asp 2260 2265 2270
Ile Asp Leu Arg Pro Ala Ser Ala Trp Thr Leu Tyr Ala Val Ala Thr 2275 2280 2285
Thr Phe Val Thr Pro Met Leu Arg His Ser Ile Glu Asn Ser Ser Val 2290 2295 2300
Asn Val Ser Leu Thr Ala Ile Ala Asn Gln Ala Thr Val Leu Met Gly 2305 2310 2315 2320
Leu Gly Lys Gly Trp Pro Leu Ser Lys Met Asp Ile Gly Val Pro Leu 2325 2330 2335
Leu Ala Ile Gly Cys Tyr Ser Gln Val Asn Pro Ile Thr Leu Thr Ala 2340 2345 2350
Ala Leu Phe Leu Leu Val Ala His Tyr Ala Ile Ile Gly Pro Gly Leu 2355 2360 2365
Gln Ala Lys Ala Thr Arg Glu Ala Gln Lys Arg Ala Ala Ala Gly Ile 2370 2375 2380
Met Lys Asn Pro Thr Val Asp Gly Ile Thr Val Ile Asp Leu Asp Pro 2385 2390 2395 2400
Ile Pro Tyr Asp Pro Lys Phe Glu Lys Gln Leu Gly Gln Val Met Leu 2405 2410 2415
Leu Val Leu Cys Val Thr Gln Val Leu Met Met Arg Thr Thr Trp Ala 2420 2425 2430
Leu Cys Glu Ala Leu Thr Leu Ala Thr Gly Pro Ile Ser Thr Leu Trp 2435 2440 2445
Glu Gly Asn Pro Gly Arg Phe Trp Asn Thr Thr Ile Ala Val Ser Met 2450 2455 2460
Ala Asn Ile Phe Arg Gly Ser Tyr Leu Ala Gly Ala Gly Leu Leu Phe 2465 2470 2475 2480
Ser Ile Met Lys Asn Thr Thr Asn Thr Arg Arg Gly Thr Gly Asn Ile 2485 2490 2495
Gly Glu Thr Leu Gly Glu Lys Trp Lys Ser Arg Leu Asn Ala Leu Gly 2500 2505 2510
Lys Ser Glu Phe Gln Ile Tyr Lys Lys Ser Gly Ile Gln Glu Val Asp 2515 2520 2525
Arg Thr Leu Ala Lys Glu Gly Ile Lys Arg Gly Glu Thr Asp His His 2530 2535 2540
Ala Val Ser Arg Gly Ser Ala Lys Leu Arg Trp Phe Val Glu Arg Asn 2545 2550 2555 2560
Met Val Thr Pro Glu Gly Lys Val Val Asp Leu Gly Cys Gly Arg Gly 2565 2570 2575
Gly Trp Ser Tyr Tyr Cys Gly Gly Leu Lys Asn Val Arg Glu Val Lys 2580 2585 2590
Gly Leu Thr Lys Gly Gly Pro Gly His Glu Glu Pro Ile Pro Met Ser 2595 2600 2605
Thr Tyr Gly Trp Asn Leu Val Arg Leu Gln Ser Gly Val Asp Val Phe 2610 2615 2620
Phe Thr Pro Pro Glu Lys Cys Asp Thr Leu Leu Cys Asp Ile Gly Glu

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2625	2630	2635	2640
Ser Ser Pro Asn Pro Thr Val Glu Ala Gly Arg Thr Leu Arg Val Leu	2645	2650	2655
Asn Leu Val Glu Asn Trp Leu Asn Asn Asn Thr Gln Phe Cys Ile Lys	2660	2665	2670
Val Leu Asn Pro Tyr Met Pro Ser Val Ile Glu Lys Met Glu Ala Leu	2675	2680	2685
Gln Arg Lys Tyr Gly Gly Ala Leu Val Arg Asn Pro Leu Ser Arg Asn	2690	2695	2700
Ser Thr His Glu Met Tyr Trp Val Ser Asn Ala Ser Gly Asn Ile Val	2705	2710	2715
Ser Ser Val Asn Met Ile Ser Arg Met Leu Ile Asn Arg Phe Thr Met	2725	2730	2735
Arg His Lys Lys Ala Thr Tyr Glu Pro Asp Val Asp Leu Gly Ser Gly	2740	2745	2750
Thr Arg Asn Ile Gly Ile Glu Ser Glu Ile Pro Asn Leu Asp Ile Ile	2755	2760	2765
Gly Lys Arg Ile Glu Lys Ile Lys Gln Glu His Glu Thr Ser Trp His	2770	2775	2780
Tyr Asp Gln Asp His Pro Tyr Lys Thr Trp Ala Tyr His Gly Ser Tyr	2785	2790	2795
Glu Thr Lys Gln Thr Gly Ser Ala Ser Ser Met Val Asn Gly Val Val	2805	2810	2815
Arg Leu Leu Thr Lys Pro Trp Asp Val Val Pro Met Val Thr Gln Met	2820	2825	2830
Ala Met Thr Asp Thr Thr Pro Phe Gly Gln Gln Arg Val Phe Lys Glu	2835	2840	2845
Lys Val Asp Thr Arg Thr Gln Glu Pro Lys Glu Gly Thr Lys Lys Leu	2850	2855	2860
Met Lys Ile Thr Ala Glu Trp Leu Trp Lys Glu Leu Gly Lys Lys Lys	2865	2870	2875
Thr Pro Arg Met Cys Thr Arg Glu Glu Phe Thr Arg Lys Val Arg Ser	2885	2890	2895
Asn Ala Ala Leu Gly Ala Ile Phe Thr Asp Glu Asn Lys Trp Lys Ser	2900	2905	2910
Ala Arg Glu Ala Val Glu Asp Ser Arg Phe Trp Glu Leu Val Asp Lys	2915	2920	2925
Glu Arg Asn Leu His Leu Glu Gly Lys Cys Glu Thr Cys Val Tyr Asn	2930	2935	2940
Met Met Gly Lys Arg Glu Lys Lys Leu Gly Glu Phe Gly Lys Ala Lys	2945	2950	2955
Gly Ser Arg Ala Ile Trp Tyr Met Trp Leu Gly Ala Arg Phe Leu Glu	2965	2970	2975
Phe Glu Ala Leu Gly Phe Leu Asn Glu Asp His Trp Phe Ser Arg Glu	2980	2985	2990
Asn Ser Leu Ser Gly Val Glu Gly Glu Gly Leu His Lys Leu Gly Tyr	2995	3000	3005
Ile Leu Arg Asp Val Ser Lys Lys Glu Gly Gly Ala Met Tyr Ala Asp	3010	3015	3020
Asp Thr Ala Gly Trp Asp Thr Arg Ile Thr Leu Glu Asp Leu Lys Asn	3025	3030	3035
Glu Glu Met Val Thr Asn His Met Glu Gly Glu His Lys Lys Leu Ala	3045	3050	3055

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Glu Ala Ile Phe Lys Leu Thr Tyr Gln Asn Lys Val Val Arg Val Gln  
 3060 3065 3070  
 Arg Pro Thr Pro Arg Gly Thr Val Met Asp Ile Ile Ser Arg Arg Asp  
 3075 3080 3085  
 Gln Arg Gly Ser Gly Gln Val Gly Thr Tyr Gly Leu Asn Thr Phe Thr  
 3090 3095 3100  
 Asn Met Glu Ala Gln Leu Ile Arg Gln Met Glu Gly Glu Gly Val Phe  
 3105 3110 3115 3120  
 Lys Ser Ile Gln His Leu Thr Val Thr Glu Glu Ile Ala Val Gln Asn  
 3125 3130 3135  
 Trp Leu Ala Arg Val Gly Arg Glu Arg Leu Ser Arg Met Ala Ile Ser  
 3140 3145 3150  
 Gly Asp Asp Cys Val Val Lys Pro Leu Asp Asp Arg Phe Ala Ser Ala  
 3155 3160 3165  
 Leu Thr Ala Leu Asn Asp Met Gly Lys Val Arg Lys Asp Ile Gln Gln  
 3170 3175 3180  
 Trp Glu Pro Ser Arg Gly Trp Asn Asp Trp Thr Gln Val Pro Phe Cys  
 3185 3190 3195 3200  
 Ser His His Phe His Glu Leu Ile Met Lys Asp Gly Arg Val Leu Val  
 3205 3210 3215  
 Val Pro Cys Arg Asn Gln Asp Glu Leu Ile Gly Arg Ala Arg Ile Ser  
 3220 3225 3230  
 Gln Gly Ala Gly Trp Ser Leu Arg Glu Thr Ala Cys Leu Gly Lys Ser  
 3235 3240 3245  
 Tyr Ala Gln Met Trp Ser Leu Met Tyr Phe His Arg Arg Asp Leu Arg  
 3250 3255 3260  
 Leu Ala Ala Asn Ala Ile Cys Ser Ala Val Pro Ser His Trp Val Pro  
 3265 3270 3275 3280  
 Thr Ser Arg Thr Thr Trp Ser Ile His Ala Lys His Glu Trp Met Thr  
 3285 3290 3295  
 Thr Glu Asp Met Leu Thr Val Trp Asn Arg Val Trp Ile Gln Glu Asn  
 3300 3305 3310  
 Pro Trp Met Glu Asp Lys Thr Pro Val Glu Ser Trp Glu Glu Ile Pro  
 3315 3320 3325  
 Tyr Leu Gly Lys Arg Glu Asp Gln Trp Cys Gly Ser Leu Ile Gly Leu  
 3330 3335 3340  
 Thr Ser Arg Ala Thr Trp Ala Lys Asn Ile Gln Thr Ala Ile Asn Gln  
 3345 3350 3355 3360  
 Val Arg Ser Leu Ile Gly Asn Glu Glu Tyr Thr Asp Tyr Met Pro Ser  
 3365 3370 3375  
 Met Lys Arg Phe Arg Arg Glu Glu Glu Glu Ala Gly Val Leu Trp  
 3380 3385 3390

<210> SEQ ID NO 22  
 <211> LENGTH: 3390  
 <212> TYPE: PRT  
 <213> ORGANISM: Dengue 3 virus strain H87

<400> SEQUENCE: 22

Met Asn Asn Gln Arg Lys Lys Thr Gly Lys Pro Ser Ile Asn Met Leu  
 1 5 10 15  
 Lys Arg Val Arg Asn Arg Val Ser Thr Gly Ser Gln Leu Ala Lys Arg  
 20 25 30  
 Phe Ser Arg Gly Leu Leu Asn Gly Gln Gly Pro Met Lys Leu Val Met



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35	40	45
Ala Phe Ile Ala Phe Leu Arg Phe Leu Ala Ile Pro Pro Thr Ala Gly 50 55 60		
Val Leu Ala Arg Trp Gly Thr Phe Lys Lys Ser Gly Ala Ile Lys Val 65 70 75 80		
Leu Lys Gly Phe Lys Lys Glu Ile Ser Asn Met Leu Ser Ile Ile Asn 85 90 95		
Lys Arg Lys Lys Thr Ser Leu Cys Leu Met Met Met Leu Pro Ala Thr 100 105 110		
Leu Ala Phe His Leu Thr Ser Arg Asp Gly Glu Pro Arg Met Ile Val 115 120 125		
Gly Lys Asn Glu Arg Gly Lys Ser Leu Leu Phe Lys Thr Ala Ser Gly 130 135 140		
Ile Asn Met Cys Thr Leu Ile Ala Met Asp Leu Gly Glu Met Cys Asp 145 150 155 160		
Asp Thr Val Thr Tyr Lys Cys Pro His Ile Thr Glu Val Glu Pro Glu 165 170 175		
Asp Ile Asp Cys Trp Cys Asn Leu Thr Ser Thr Trp Val Thr Tyr Gly 180 185 190		
Thr Cys Asn Gln Ala Gly Glu His Arg Arg Asp Lys Arg Ser Val Ala 195 200 205		
Leu Ala Pro His Val Gly Met Gly Leu Asp Thr Arg Thr Gln Thr Trp 210 215 220		
Met Ser Ala Glu Gly Ala Trp Arg Gln Val Glu Lys Val Glu Thr Trp 225 230 235 240		
Ala Leu Arg His Pro Gly Phe Thr Ile Leu Ala Leu Phe Leu Ala His 245 250 255		
Tyr Ile Gly Thr Ser Leu Thr Gln Lys Val Val Ile Phe Ile Leu Leu 260 265 270		
Met Leu Val Thr Pro Ser Met Thr Met Arg Cys Val Gly Val Gly Asn 275 280 285		
Arg Asp Phe Val Glu Gly Leu Ser Gly Ala Thr Trp Val Asp Val Val 290 295 300		
Leu Glu His Gly Gly Cys Val Thr Thr Met Ala Lys Asn Lys Pro Thr 305 310 315 320		
Leu Asp Ile Glu Leu Gln Lys Thr Glu Ala Thr Gln Leu Ala Thr Leu 325 330 335		
Arg Lys Leu Cys Ile Glu Gly Lys Ile Thr Asn Ile Thr Thr Asp Ser 340 345 350		
Arg Cys Pro Thr Gln Gly Glu Ala Ile Leu Pro Glu Glu Gln Asp Gln 355 360 365		
Asn Tyr Val Cys Lys His Thr Tyr Val Asp Arg Gly Trp Gly Asn Gly 370 375 380		
Cys Gly Leu Phe Gly Lys Gly Ser Leu Val Thr Cys Ala Lys Phe Gln 385 390 395 400		
Cys Leu Glu Ser Ile Glu Gly Lys Val Val Gln His Glu Asn Leu Lys 405 410 415		
Tyr Thr Val Ile Ile Thr Val His Thr Gly Asp Gln His Gln Val Gly 420 425 430		
Asn Glu Thr Gln Gly Val Thr Ala Glu Ile Thr Ser Gln Ala Ser Thr 435 440 445		
Ala Glu Ala Ile Leu Pro Glu Tyr Gly Thr Leu Gly Leu Glu Cys Ser 450 455 460		

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Pro Arg Thr Gly Leu Asp Phe Asn Glu Met Ile Leu Leu Thr Met Lys  
 465 470 475 480  
 Asn Lys Ala Trp Met Val His Arg Gln Trp Phe Phe Asp Leu Pro Leu  
 485 490 495  
 Pro Trp Thr Ser Gly Ala Thr Thr Lys Thr Pro Thr Trp Asn Arg Lys  
 500 505 510  
 Glu Leu Leu Val Thr Phe Lys Asn Ala His Ala Lys Lys Gln Glu Val  
 515 520 525  
 Val Val Leu Gly Ser Gln Glu Gly Ala Met His Thr Ala Leu Thr Gly  
 530 535 540  
 Ala Thr Glu Ile Gln Thr Ser Gly Gly Thr Ser Ile Phe Ala Gly His  
 545 550 555 560  
 Leu Lys Cys Arg Leu Lys Met Asp Lys Leu Lys Leu Lys Gly Met Ser  
 565 570 575  
 Tyr Ala Met Cys Leu Asn Thr Phe Val Leu Lys Lys Glu Val Ser Glu  
 580 585 590  
 Thr Gln His Gly Thr Ile Leu Ile Lys Val Glu Tyr Lys Gly Glu Asp  
 595 600 605  
 Ala Pro Cys Lys Ile Pro Phe Ser Thr Glu Asp Gly Gln Gly Lys Ala  
 610 615 620  
 His Asn Gly Arg Leu Ile Thr Ala Asn Pro Val Val Thr Lys Lys Glu  
 625 630 635 640  
 Glu Pro Val Asn Ile Glu Ala Glu Pro Pro Phe Gly Glu Ser Asn Ile  
 645 650 655  
 Val Ile Gly Ile Gly Asp Lys Ala Leu Lys Ile Asn Trp Tyr Arg Lys  
 660 665 670  
 Gly Ser Ser Ile Gly Lys Met Phe Glu Ala Thr Ala Arg Gly Ala Arg  
 675 680 685  
 Arg Met Ala Ile Leu Gly Asp Thr Ala Trp Asp Phe Gly Ser Val Gly  
 690 695 700  
 Gly Val Leu Asn Ser Leu Gly Lys Met Val His Gln Ile Phe Gly Ser  
 705 710 715 720  
 Ala Tyr Thr Ala Leu Phe Ser Gly Val Ser Trp Ile Met Lys Ile Gly  
 725 730 735  
 Ile Gly Val Leu Leu Thr Trp Ile Gly Leu Asn Ser Lys Asn Thr Ser  
 740 745 750  
 Met Ser Phe Ser Cys Ile Ala Ile Gly Ile Ile Thr Leu Tyr Leu Gly  
 755 760 765  
 Val Val Val Gln Ala Asp Met Gly Cys Val Ile Asn Trp Lys Gly Lys  
 770 775 780  
 Glu Leu Lys Cys Gly Ser Gly Ile Phe Val Thr Asn Glu Val His Thr  
 785 790 795 800  
 Trp Thr Glu Gln Tyr Lys Phe Gln Ala Asp Ser Pro Lys Arg Val Ala  
 805 810 815  
 Thr Ala Ile Ala Gly Ala Trp Glu Asn Gly Val Cys Gly Ile Arg Ser  
 820 825 830  
 Thr Thr Arg Met Glu Asn Leu Leu Trp Lys Gln Ile Ala Asn Glu Leu  
 835 840 845  
 Asn Tyr Ile Leu Trp Glu Asn Asp Ile Lys Leu Thr Val Val Val Gly  
 850 855 860  
 Asp Ile Thr Gly Val Leu Glu Gln Gly Lys Arg Thr Leu Thr Pro Gln  
 865 870 875 880

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Pro	Met	Glu	Leu	Lys	Tyr	Ser	Trp	Lys	Thr	Trp	Gly	Leu	Ala	Lys	Ile	885	890	895	
Val	Thr	Ala	Glu	Thr	Gln	Asn	Ser	Ser	Phe	Ile	Ile	Asp	Gly	Pro	Ser	900	905	910	
Thr	Pro	Glu	Cys	Pro	Ser	Ala	Ser	Arg	Ala	Trp	Asn	Val	Trp	Glu	Val	915	920	925	
Glu	Asp	Tyr	Gly	Phe	Gly	Val	Phe	Thr	Thr	Asn	Ile	Trp	Leu	Lys	Leu	930	935	940	
Arg	Glu	Val	Tyr	Thr	Gln	Leu	Cys	Asp	His	Arg	Leu	Met	Ser	Ala	Ala	945	950	955	960
Val	Lys	Asp	Glu	Arg	Ala	Val	His	Ala	Asp	Met	Gly	Tyr	Trp	Ile	Glu	965	970	975	
Ser	Gln	Lys	Asn	Gly	Ser	Trp	Lys	Leu	Glu	Lys	Ala	Ser	Leu	Ile	Glu	980	985	990	
Val	Lys	Thr	Cys	Thr	Trp	Pro	Lys	Ser	His	Thr	Leu	Trp	Ser	Asn	Gly	995	1000	1005	
Val	Leu	Glu	Ser	Asp	Met	Ile	Ile	Pro	Lys	Ser	Leu	Ala	Gly	Pro	Ile	1010	1015	1020	
Ser	Gln	His	Asn	His	Arg	Pro	Gly	Tyr	His	Thr	Gln	Thr	Ala	Gly	Pro	1025	1030	1035	1040
Trp	His	Leu	Gly	Lys	Leu	Glu	Leu	Asp	Phe	Asn	Tyr	Cys	Glu	Gly	Thr	1045	1050	1055	
Thr	Val	Val	Ile	Ser	Glu	Asn	Cys	Gly	Thr	Arg	Gly	Pro	Ser	Leu	Arg	1060	1065	1070	
Thr	Thr	Thr	Val	Ser	Gly	Lys	Leu	Ile	His	Glu	Trp	Cys	Cys	Arg	Ser	1075	1080	1085	
Cys	Thr	Leu	Pro	Pro	Leu	Arg	Tyr	Met	Gly	Glu	Asp	Gly	Cys	Trp	Tyr	1090	1095	1100	
Gly	Met	Glu	Ile	Arg	Pro	Ile	Asn	Glu	Lys	Glu	Glu	Asn	Met	Val	Lys	1105	1110	1115	1120
Ser	Leu	Ala	Ser	Ala	Gly	Ser	Gly	Lys	Val	Asp	Asn	Phe	Thr	Met	Gly	1125	1130	1135	
Val	Leu	Cys	Leu	Ala	Ile	Leu	Phe	Glu	Glu	Val	Met	Arg	Gly	Lys	Phe	1140	1145	1150	
Gly	Lys	Lys	His	Met	Ile	Ala	Gly	Val	Leu	Phe	Thr	Phe	Val	Leu	Leu	1155	1160	1165	
Leu	Ser	Gly	Gln	Ile	Thr	Trp	Arg	Gly	Met	Ala	His	Thr	Leu	Ile	Met	1170	1175	1180	
Ile	Gly	Ser	Asn	Ala	Ser	Asp	Arg	Met	Gly	Met	Gly	Val	Thr	Tyr	Leu	1185	1190	1195	1200
Ala	Leu	Ile	Ala	Thr	Phe	Lys	Ile	Gln	Pro	Phe	Leu	Ala	Leu	Gly	Phe	1205	1210	1215	
Phe	Leu	Arg	Lys	Leu	Thr	Ser	Arg	Glu	Asn	Leu	Leu	Leu	Gly	Val	Gly	1220	1225	1230	
Leu	Ala	Met	Ala	Ala	Thr	Leu	Arg	Leu	Pro	Glu	Asp	Ile	Glu	Gln	Met	1235	1240	1245	
Ala	Asn	Gly	Ile	Ala	Leu	Gly	Leu	Met	Ala	Leu	Lys	Leu	Ile	Thr	Gln	1250	1255	1260	
Phe	Glu	Thr	Tyr	Gln	Leu	Trp	Thr	Ala	Leu	Val	Ser	Leu	Thr	Cys	Ser	1265	1270	1275	1280
Asn	Thr	Ile	Phe	Thr	Leu	Thr	Val	Ala	Trp	Arg	Thr	Ala	Thr	Leu	Ile	1285	1290	1295	
Leu	Ala	Gly	Ile	Ser	Leu	Leu	Pro	Val	Cys	Gln	Ser	Ser	Ser	Met	Arg				

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1300			1305			1310		
Lys Thr Asp Trp Leu Pro Met Thr Val Ala Ala Met Gly Val Pro Pro	1315	1320	1325					
Leu Pro Leu Phe Ile Phe Ser Leu Lys Asp Thr Leu Lys Arg Arg Ser	1330	1335	1340					
Trp Pro Leu Asn Glu Gly Val Met Ala Val Gly Leu Val Ser Ile Leu	1345	1350	1355					1360
Ala Ser Ser Leu Leu Arg Asn Asp Val Pro Met Ala Gly Pro Leu Val	1365	1370	1375					
Ala Gly Gly Leu Leu Ile Ala Cys Tyr Val Ile Thr Gly Thr Ser Ala	1380	1385	1390					
Asp Leu Thr Val Glu Lys Ala Ala Asp Val Thr Trp Glu Glu Glu Ala	1395	1400	1405					
Glu Gln Thr Gly Val Ser His Asn Leu Met Ile Thr Val Asp Asp Asp	1410	1415	1420					
Gly Thr Met Arg Ile Lys Asp Asp Glu Thr Glu Asn Ile Leu Thr Val	1425	1430	1435					1440
Leu Leu Lys Thr Ala Leu Leu Ile Val Ser Gly Ile Phe Pro Tyr Ser	1445	1450	1455					
Ile Pro Ala Thr Met Leu Val Trp His Thr Trp Gln Lys Gln Thr Gln	1460	1465	1470					
Arg Ser Gly Val Leu Trp Asp Val Pro Ser Pro Pro Glu Thr Gln Lys	1475	1480	1485					
Ala Glu Leu Glu Glu Gly Val Tyr Arg Ile Lys Gln Gln Gly Ile Phe	1490	1495	1500					
Gly Lys Thr Gln Val Gly Val Gly Val Gln Lys Glu Gly Val Phe His	1505	1510	1515					1520
Thr Met Trp His Val Thr Arg Gly Ala Val Leu Thr His Asn Gly Lys	1525	1530	1535					
Arg Leu Glu Pro Asn Trp Ala Ser Val Lys Lys Asp Leu Ile Ser Tyr	1540	1545	1550					
Gly Gly Gly Trp Arg Leu Ser Ala Gln Trp Gln Lys Gly Glu Glu Val	1555	1560	1565					
Gln Val Ile Ala Val Glu Pro Gly Lys Asn Pro Lys Asn Phe Gln Thr	1570	1575	1580					
Met Pro Gly Ile Phe Gln Thr Thr Thr Gly Glu Ile Gly Ala Ile Ala	1585	1590	1595					1600
Leu Asp Phe Lys Pro Gly Thr Ser Gly Ser Pro Ile Ile Asn Arg Glu	1605	1610	1615					
Gly Lys Val Val Gly Leu Tyr Gly Asn Gly Val Val Thr Lys Asn Gly	1620	1625	1630					
Gly Tyr Val Ser Gly Ile Ala Gln Thr Asn Ala Glu Pro Asp Gly Pro	1635	1640	1645					
Thr Pro Glu Leu Glu Glu Glu Met Phe Lys Lys Arg Asn Leu Thr Ile	1650	1655	1660					
Met Asp Leu His Pro Gly Ser Gly Lys Thr Arg Lys Tyr Leu Pro Ala	1665	1670	1675					1680
Ile Val Arg Glu Ala Ile Lys Arg Arg Leu Arg Thr Leu Ile Leu Ala	1685	1690	1695					
Pro Thr Arg Val Val Ala Ala Glu Met Glu Glu Ala Met Lys Gly Leu	1700	1705	1710					
Pro Ile Arg Tyr Gln Thr Thr Ala Thr Lys Ser Glu His Thr Gly Arg	1715	1720	1725					

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Glu Ile Val Asp Leu Met Cys His Ala Thr Phe Thr Met Arg Leu Leu  
 1730 1735 1740  
 Ser Pro Val Arg Val Pro Asn Tyr Asn Leu Ile Ile Met Asp Glu Ala  
 1745 1750 1755 1760  
 His Phe Thr Asp Pro Ala Ser Ile Ala Ala Arg Gly Tyr Ile Ser Thr  
 1765 1770 1775  
 Arg Val Gly Met Gly Glu Ala Ala Ala Ile Phe Met Thr Ala Thr Pro  
 1780 1785 1790  
 Pro Gly Thr Ala Asp Ala Phe Pro Gln Ser Asn Ala Pro Ile Gln Asp  
 1795 1800 1805  
 Glu Glu Arg Asp Ile Pro Glu Arg Ser Trp Asn Ser Gly Asn Glu Trp  
 1810 1815 1820  
 Ile Thr Asp Phe Val Gly Lys Thr Val Trp Phe Val Pro Ser Ile Lys  
 1825 1830 1835 1840  
 Ala Gly Asn Val Ile Ala Asn Cys Leu Arg Lys Asn Gly Lys Lys Val  
 1845 1850 1855  
 Ile Gln Leu Ser Arg Lys Thr Phe Asp Thr Glu Tyr Gln Lys Thr Lys  
 1860 1865 1870  
 Leu Asn Asp Trp Asp Phe Val Val Thr Thr Asp Ile Ser Glu Met Gly  
 1875 1880 1885  
 Ala Asn Phe Ile Ala Asp Arg Val Ile Asp Pro Arg Arg Cys Leu Lys  
 1890 1895 1900  
 Pro Val Ile Leu Thr Asp Gly Pro Glu Arg Val Ile Leu Ala Gly Pro  
 1905 1910 1915 1920  
 Met Pro Val Thr Val Ala Ser Ala Ala Gln Arg Arg Gly Arg Val Gly  
 1925 1930 1935  
 Arg Asn Pro Gln Lys Glu Asn Asp Gln Tyr Ile Phe Met Gly Gln Pro  
 1940 1945 1950  
 Leu Asn Lys Asp Glu Asp His Ala His Trp Thr Glu Ala Lys Met Leu  
 1955 1960 1965  
 Leu Asp Asn Ile Asn Thr Pro Glu Gly Ile Ile Pro Ala Leu Phe Glu  
 1970 1975 1980  
 Pro Glu Arg Glu Lys Ser Ala Ala Ile Asp Gly Glu Tyr Arg Leu Lys  
 1985 1990 1995 2000  
 Gly Glu Ser Arg Lys Thr Phe Val Glu Leu Met Arg Arg Gly Asp Leu  
 2005 2010 2015  
 Pro Val Trp Leu Ala His Lys Val Ala Ser Glu Gly Ile Lys Tyr Thr  
 2020 2025 2030  
 Asp Arg Lys Trp Cys Phe Asp Gly Glu Arg Asn Asn Gln Ile Leu Glu  
 2035 2040 2045  
 Glu Asn Met Asp Val Glu Ile Trp Thr Lys Glu Gly Glu Lys Lys Lys  
 2050 2055 2060  
 Leu Arg Pro Arg Trp Leu Asp Ala Arg Thr Tyr Ser Asp Pro Leu Ala  
 2065 2070 2075 2080  
 Leu Lys Glu Phe Lys Asp Phe Ala Ala Gly Arg Lys Ser Ile Ala Leu  
 2085 2090 2095  
 Asp Leu Val Thr Glu Ile Gly Arg Val Pro Ser His Leu Ala His Arg  
 2100 2105 2110  
 Thr Arg Asn Ala Leu Asp Asn Leu Val Met Leu His Thr Ser Glu His  
 2115 2120 2125  
 Gly Gly Arg Ala Tyr Arg His Ala Val Glu Glu Leu Pro Glu Thr Met  
 2130 2135 2140

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Glu	Thr	Leu	Leu	Leu	Leu	Gly	Leu	Met	Ile	Leu	Leu	Thr	Gly	Gly	Ala	2145	2150	2155	2160
Met	Leu	Phe	Leu	Ile	Ser	Gly	Lys	Gly	Ile	Gly	Lys	Thr	Ser	Ile	Gly	2165	2170	2175	
Leu	Ile	Cys	Val	Ile	Ala	Ser	Ser	Gly	Met	Leu	Trp	Met	Ala	Asp	Val	2180	2185	2190	
Pro	Leu	Gln	Trp	Ile	Ala	Ser	Ala	Ile	Val	Leu	Glu	Phe	Phe	Met	Met	2195	2200	2205	
Val	Leu	Leu	Ile	Pro	Glu	Pro	Glu	Lys	Gln	Arg	Thr	Pro	Gln	Asp	Asn	2210	2215	2220	
Gln	Leu	Ala	Tyr	Val	Val	Ile	Gly	Ile	Leu	Thr	Leu	Ala	Ala	Ile	Val	2225	2230	2235	2240
Ala	Ala	Asn	Glu	Met	Gly	Leu	Leu	Glu	Thr	Thr	Lys	Arg	Asp	Leu	Gly	2245	2250	2255	
Met	Ser	Lys	Glu	Pro	Gly	Val	Val	Ser	Pro	Thr	Ser	Tyr	Leu	Asp	Val	2260	2265	2270	
Asp	Leu	His	Pro	Ala	Ser	Ala	Trp	Thr	Leu	Tyr	Ala	Val	Ala	Thr	Thr	2275	2280	2285	
Val	Ile	Thr	Pro	Met	Leu	Arg	His	Thr	Ile	Glu	Asn	Ser	Thr	Ala	Asn	2290	2295	2300	
Val	Ser	Leu	Ala	Ala	Ile	Ala	Asn	Gln	Ala	Val	Val	Leu	Met	Gly	Leu	2305	2310	2315	2320
Asp	Lys	Gly	Trp	Pro	Ile	Ser	Lys	Met	Asp	Leu	Gly	Val	Pro	Leu	Leu	2325	2330	2335	
Ala	Leu	Gly	Cys	Tyr	Ser	Gln	Val	Asn	Pro	Leu	Thr	Leu	Ile	Ala	Ala	2340	2345	2350	
Val	Leu	Leu	Leu	Val	Thr	His	Tyr	Ala	Ile	Ile	Gly	Pro	Gly	Leu	Gln	2355	2360	2365	
Ala	Lys	Ala	Thr	Arg	Glu	Ala	Gln	Lys	Arg	Thr	Ala	Ala	Gly	Ile	Met	2370	2375	2380	
Lys	Asn	Pro	Thr	Val	Asp	Gly	Ile	Met	Thr	Ile	Asp	Leu	Asp	Pro	Val	2385	2390	2395	2400
Ile	Tyr	Asp	Ser	Lys	Phe	Glu	Lys	Gln	Leu	Gly	Gln	Val	Met	Leu	Leu	2405	2410	2415	
Val	Leu	Cys	Ala	Val	Gln	Leu	Leu	Leu	Met	Arg	Thr	Ser	Trp	Ala	Leu	2420	2425	2430	
Cys	Glu	Val	Leu	Thr	Leu	Ala	Thr	Gly	Pro	Ile	Thr	Thr	Leu	Trp	Glu	2435	2440	2445	
Gly	Ser	Pro	Gly	Lys	Phe	Trp	Asn	Thr	Thr	Ile	Ala	Val	Ser	Met	Ala	2450	2455	2460	
Asn	Ile	Phe	Arg	Gly	Ser	Tyr	Leu	Ala	Gly	Ala	Gly	Leu	Ala	Leu	Ser	2465	2470	2475	2480
Ile	Met	Lys	Ser	Val	Gly	Thr	Gly	Lys	Arg	Gly	Thr	Gly	Ser	Gln	Gly	2485	2490	2495	
Glu	Thr	Leu	Gly	Glu	Lys	Trp	Lys	Lys	Lys	Leu	Asn	Gln	Leu	Ser	Arg	2500	2505	2510	
Lys	Glu	Phe	Asp	Leu	Tyr	Lys	Lys	Ser	Gly	Ile	Thr	Glu	Val	Asp	Arg	2515	2520	2525	
Thr	Glu	Ala	Lys	Glu	Gly	Leu	Lys	Arg	Gly	Glu	Ile	Thr	His	His	Ala	2530	2535	2540	
Val	Ser	Arg	Gly	Ser	Ala	Lys	Leu	Gln	Trp	Phe	Val	Glu	Arg	Asn	Met	2545	2550	2555	2560
Val	Ile	Pro	Glu	Gly	Arg	Val	Ile	Asp	Leu	Gly	Cys	Gly	Arg	Gly	Gly				

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2565	2570	2575
Trp Ser Tyr Tyr Cys Ala Gly Leu Lys Lys Val Thr Glu Val Arg Gly 2580 2585 2590		
Tyr Thr Lys Gly Gly Pro Gly His Glu Glu Pro Val Pro Met Ser Thr 2595 2600 2605		
Tyr Gly Trp Asn Ile Val Lys Leu Met Ser Gly Lys Asp Val Phe Tyr 2610 2615 2620		
Leu Pro Pro Glu Lys Cys Asp Thr Leu Leu Cys Asp Ile Gly Glu Ser 2625 2630 2635 2640		
Ser Pro Ser Pro Thr Val Glu Glu Ser Arg Thr Ile Arg Val Leu Lys 2645 2650 2655		
Met Val Glu Pro Trp Leu Lys Asn Asn Gln Phe Cys Ile Lys Val Leu 2660 2665 2670		
Asn Pro Tyr Met Pro Thr Val Ile Glu His Leu Glu Arg Leu Gln Arg 2675 2680 2685		
Lys His Gly Gly Met Leu Val Arg Asn Pro Leu Ser Arg Asn Ser Thr 2690 2695 2700		
His Glu Met Tyr Trp Ile Ser Asn Gly Thr Gly Asn Ile Val Ser Ser 2705 2710 2715 2720		
Val Asn Met Val Ser Arg Leu Leu Leu Asn Arg Phe Thr Met Thr His 2725 2730 2735		
Arg Arg Pro Thr Ile Glu Lys Asp Val Asp Leu Gly Ala Gly Thr Arg 2740 2745 2750		
His Val Asn Ala Glu Pro Glu Thr Pro Asn Met Asp Val Ile Gly Glu 2755 2760 2765		
Arg Ile Lys Arg Ile Lys Glu Glu His Ser Ser Thr Trp His Tyr Asp 2770 2775 2780		
Asp Glu Asn Pro Tyr Lys Thr Trp Ala Tyr His Gly Ser Tyr Glu Val 2785 2790 2795 2800		
Lys Ala Thr Gly Ser Ala Ser Ser Met Ile Asn Gly Val Val Lys Leu 2805 2810 2815		
Leu Thr Lys Pro Trp Asp Val Val Pro Met Val Thr Gln Met Ala Met 2820 2825 2830		
Thr Asp Thr Thr Pro Phe Gly Gln Gln Arg Val Phe Lys Glu Lys Val 2835 2840 2845		
Asp Thr Arg Thr Pro Arg Pro Met Pro Gly Thr Arg Lys Val Met Glu 2850 2855 2860		
Ile Thr Ala Glu Trp Leu Trp Arg Thr Leu Gly Arg Asn Lys Arg Pro 2865 2870 2875 2880		
Arg Leu Cys Thr Arg Glu Glu Phe Thr Lys Lys Val Arg Thr Asn Ala 2885 2890 2895		
Ala Met Gly Ala Val Phe Thr Glu Glu Asn Gln Trp Asp Ser Ala Arg 2900 2905 2910		
Ala Ala Val Glu Asp Glu Glu Phe Trp Lys Leu Val Asp Arg Glu Arg 2915 2920 2925		
Glu Leu His Lys Leu Gly Lys Cys Gly Ser Cys Val Tyr Asn Met Met 2930 2935 2940		
Gly Lys Arg Glu Lys Lys Leu Gly Glu Phe Gly Lys Ala Lys Gly Ser 2945 2950 2955 2960		
Arg Ala Ile Trp Tyr Met Trp Leu Gly Ala Arg Tyr Leu Glu Phe Glu 2965 2970 2975		
Ala Leu Gly Phe Leu Asn Glu Asp His Trp Phe Ser Arg Glu Asn Ser 2980 2985 2990		

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Tyr Ser Gly Val Glu Gly Glu Gly Leu His Lys Leu Gly Tyr Ile Leu  
 2995 3000 3005  
 Arg Asp Ile Ser Lys Ile Pro Gly Gly Ala Met Tyr Ala Asp Asp Thr  
 3010 3015 3020  
 Ala Gly Trp Asp Thr Arg Ile Thr Glu Asp Asp Leu His Asn Glu Glu  
 3025 3030 3035 3040  
 Lys Ile Thr Gln Gln Met Asp Pro Glu His Arg Gln Leu Ala Asn Ala  
 3045 3050 3055  
 Ile Phe Lys Leu Thr Tyr Gln Asn Lys Val Val Lys Val Gln Arg Pro  
 3060 3065 3070  
 Thr Pro Lys Gly Thr Val Met Asp Ile Ile Ser Arg Lys Asp Gln Arg  
 3075 3080 3085  
 Gly Ser Gly Gln Val Gly Thr Tyr Gly Leu Asn Thr Phe Thr Asn Met  
 3090 3095 3100  
 Glu Ala Gln Leu Ile Arg Gln Met Glu Gly Glu Gly Val Leu Ser Lys  
 3105 3110 3115 3120  
 Ala Asp Leu Glu Asn Pro His Pro Leu Glu Lys Lys Ile Thr Gln Trp  
 3125 3130 3135  
 Leu Glu Thr Lys Gly Val Glu Arg Leu Lys Arg Met Ala Ile Ser Gly  
 3140 3145 3150  
 Asp Asp Cys Val Val Lys Pro Ile Asp Asp Arg Phe Ala Asn Ala Leu  
 3155 3160 3165  
 Leu Ala Leu Asn Asp Met Gly Lys Val Arg Lys Asp Ile Pro Gln Trp  
 3170 3175 3180  
 Gln Pro Ser Lys Gly Trp His Asp Trp Gln Gln Val Pro Phe Cys Ser  
 3185 3190 3195 3200  
 His His Phe His Glu Leu Ile Met Lys Asp Gly Arg Lys Leu Val Val  
 3205 3210 3215  
 Pro Cys Arg Pro Gln Asp Glu Leu Ile Gly Arg Ala Arg Ile Ser Gln  
 3220 3225 3230  
 Gly Ala Gly Trp Ser Leu Arg Glu Thr Ala Cys Leu Gly Lys Ala Tyr  
 3235 3240 3245  
 Ala Gln Met Trp Thr Leu Met Tyr Phe His Arg Arg Asp Leu Arg Leu  
 3250 3255 3260  
 Ala Ser Asn Ala Ile Cys Ser Ala Val Pro Val His Trp Val Pro Thr  
 3265 3270 3275 3280  
 Ser Arg Thr Thr Trp Ser Ile His Ala His His Gln Trp Met Thr Thr  
 3285 3290 3295  
 Glu Asp Met Leu Thr Val Trp Asn Arg Val Trp Ile Glu Asp Asn Pro  
 3300 3305 3310  
 Trp Met Glu Asp Lys Thr Pro Val Thr Thr Trp Glu Asp Val Pro Tyr  
 3315 3320 3325  
 Leu Gly Lys Arg Glu Asp Gln Trp Cys Gly Ser Leu Ile Gly Leu Thr  
 3330 3335 3340  
 Ser Arg Ala Thr Trp Ala Gln Asn Ile Leu Thr Ala Ile Gln Gln Val  
 3345 3350 3355 3360  
 Arg Ser Leu Ile Gly Asn Glu Glu Phe Leu Asp Tyr Met Pro Ser Met  
 3365 3370 3375  
 Lys Arg Phe Arg Lys Glu Glu Glu Ser Glu Gly Ala Ile Trp  
 3380 3385 3390

&lt;210&gt; SEQ ID NO 23

&lt;211&gt; LENGTH: 21



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<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide

<400> SEQUENCE: 23

cagttccaaa ccggaagctt g 21

<210> SEQ ID NO 24  
<211> LENGTH: 27  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide

<400> SEQUENCE: 24

ccaacgagct atcgtacgtt ctctggg 27

<210> SEQ ID NO 25  
<211> LENGTH: 27  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide

<400> SEQUENCE: 25

gattgtgacc atggcggccc atctttg 27

<210> SEQ ID NO 26  
<211> LENGTH: 31  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide

<400> SEQUENCE: 26

ggagattagg ccgctgagcg gtaaagaaga g 31

<210> SEQ ID NO 27  
<211> LENGTH: 26  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide

<400> SEQUENCE: 27

gtttgtggaa aaatgtctga ggagaa 26

<210> SEQ ID NO 28  
<211> LENGTH: 27  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide

<400> SEQUENCE: 28

ctaggaaaca cataatatta gttgtgg 27

<210> SEQ ID NO 29  
<211> LENGTH: 27  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide

<400> SEQUENCE: 29

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cagatccacc taaccataat ggcagtg 27

<210> SEQ ID NO 30  
<211> LENGTH: 25  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide

<400> SEQUENCE: 30

ggaaactcac ctcgaggag acagc 25

<210> SEQ ID NO 31  
<211> LENGTH: 26  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide

<400> SEQUENCE: 31

ttgggtagag gtcaccgcac tcatcc 26

<210> SEQ ID NO 32  
<211> LENGTH: 25  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide

<400> SEQUENCE: 32

gtagaaatag ccgctctcat cctag 25

<210> SEQ ID NO 33  
<211> LENGTH: 30  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide

<400> SEQUENCE: 33

ggcggcttac gtaatgggag gtagctcagc 30

<210> SEQ ID NO 34  
<211> LENGTH: 27  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide

<400> SEQUENCE: 34

ctagagaagg cagcttctgt gcagtgg 27

<210> SEQ ID NO 35  
<211> LENGTH: 25  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide

<400> SEQUENCE: 35

ccttgccat tccagcaaca atgac 25

<210> SEQ ID NO 36  
<211> LENGTH: 26  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence

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<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide

<400> SEQUENCE: 36

gacgttcaaa ttttagccat agaacc 26

<210> SEQ ID NO 37  
<211> LENGTH: 27  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide

<400> SEQUENCE: 37

ctggagaaac gggcgccgta acattag 27

<210> SEQ ID NO 38  
<211> LENGTH: 27  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide

<400> SEQUENCE: 38

gaaattggat cggtaacctt agatttc 27

<210> SEQ ID NO 39  
<211> LENGTH: 27  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide

<400> SEQUENCE: 39

ggagcagtaa cgtttgattt caaaccc 27

<210> SEQ ID NO 40  
<211> LENGTH: 24  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide

<400> SEQUENCE: 40

gttaccaaac ctggggatta cgtc 24

<210> SEQ ID NO 41  
<211> LENGTH: 25  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide

<400> SEQUENCE: 41

gattaactat catgaactta caccc 25

<210> SEQ ID NO 42  
<211> LENGTH: 25  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide

<400> SEQUENCE: 42

ggaaaacctt tggcaccgag tatcc 25

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<210> SEQ ID NO 43  
<211> LENGTH: 26  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide

<400> SEQUENCE: 43

tccagtgata ccggctagcg ctgctc 26

<210> SEQ ID NO 44  
<211> LENGTH: 23  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide

<400> SEQUENCE: 44

gcctcagagg tggccaaagg aag 23

<210> SEQ ID NO 45  
<211> LENGTH: 26  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide

<400> SEQUENCE: 45

acatggaggc agagatctgg actaga 26

<210> SEQ ID NO 46  
<211> LENGTH: 23  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide

<400> SEQUENCE: 46

aaagcatggc caaggatgct gtc 23

<210> SEQ ID NO 47  
<211> LENGTH: 28  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide

<400> SEQUENCE: 47

gcataatgga cgctaagcat gactaagg 28

<210> SEQ ID NO 48  
<211> LENGTH: 26  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide

<400> SEQUENCE: 48

ttattgcata gtgcacgaaa agcatg 26

<210> SEQ ID NO 49  
<211> LENGTH: 23  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide

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<400> SEQUENCE: 49  
gggcctatta ttacgtaatg gac 23

<210> SEQ ID NO 50  
<211> LENGTH: 24  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide

<400> SEQUENCE: 50  
ctgcaatcct ggtgatatta ttgc 24

<210> SEQ ID NO 51  
<211> LENGTH: 23  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide

<400> SEQUENCE: 51  
ctcataaaga acgttcaaac cct 23

<210> SEQ ID NO 52  
<211> LENGTH: 24  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide

<400> SEQUENCE: 52  
cattagacag acgcgagttt gaag 24

<210> SEQ ID NO 53  
<211> LENGTH: 27  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide

<400> SEQUENCE: 53  
tggcgacgct caagatagtg actgaag 27

<210> SEQ ID NO 54  
<211> LENGTH: 25  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide

<400> SEQUENCE: 54  
gagtcacat cgataccaac aatag 25

<210> SEQ ID NO 55  
<211> LENGTH: 27  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide

<400> SEQUENCE: 55  
cttcaaaacc tggcttctgc atcaaag 27

<210> SEQ ID NO 56

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<211> LENGTH: 28  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide

<400> SEQUENCE: 56

caaagatggt gagcaacagg ttcacaac 28

<210> SEQ ID NO 57  
<211> LENGTH: 27  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide

<400> SEQUENCE: 57

ggaaagaaga aacacccgag actgtgc 27

<210> SEQ ID NO 58  
<211> LENGTH: 25  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide

<400> SEQUENCE: 58

gggaactggt cgatcgagaa agggc 25

<210> SEQ ID NO 59  
<211> LENGTH: 28  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide

<400> SEQUENCE: 59

ccagtggatt actacagaag atatgctc 28

<210> SEQ ID NO 60  
<211> LENGTH: 29  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide

<400> SEQUENCE: 60

caggaacctg accggtaaag aggaatcgc 29

<210> SEQ ID NO 61  
<211> LENGTH: 28  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide

<400> SEQUENCE: 61

ctgtaattac caacatcaaa caccaaag 28

<210> SEQ ID NO 62  
<211> LENGTH: 26  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide

<400> SEQUENCE: 62

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ccaacaacaa ccaccaaagg ctattg 26

<210> SEQ ID NO 63  
<211> LENGTH: 25  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Oligonucleotide  
  
<400> SEQUENCE: 63

ggattggtgt tgcgatcca acagg 25

<210> SEQ ID NO 64  
<211> LENGTH: 24  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Primer  
  
<400> SEQUENCE: 64

ctggtggaag cccaacacaa aaac 24

<210> SEQ ID NO 65  
<211> LENGTH: 45  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Primer  
  
<400> SEQUENCE: 65

ctggtggaag gaagagagaa attggcaact cccaacaca aaaac 45

<210> SEQ ID NO 66  
<211> LENGTH: 22  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Primer  
  
<400> SEQUENCE: 66

agaccccccc aagcatattg ac 22

<210> SEQ ID NO 67  
<211> LENGTH: 39  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Primer  
  
<400> SEQUENCE: 67

agaccccccc aatatttcct cctcctatag catattgac 39

<210> SEQ ID NO 68  
<211> LENGTH: 21  
<212> TYPE: DNA  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: Primer  
  
<400> SEQUENCE: 68

ccaacacaa agcatattga c 21

<210> SEQ ID NO 69  
<211> LENGTH: 6  
<212> TYPE: DNA

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<213> ORGANISM: Artificial Sequence  
 <220> FEATURE:  
 <223> OTHER INFORMATION: Oligonucleotide  
 <220> FEATURE:  
 <221> NAME/KEY: misc\_feature  
 <222> LOCATION: (1)...(6)  
 <223> OTHER INFORMATION: n = A,T,C or G

<400> SEQUENCE: 69

gcagcn

6

<210> SEQ ID NO 70  
 <211> LENGTH: 5  
 <212> TYPE: DNA  
 <213> ORGANISM: Artificial Sequence  
 <220> FEATURE:  
 <223> OTHER INFORMATION: Oligonucleotide

<400> SEQUENCE: 70

gcagc

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What is claimed is:

1. A temperature-sensitive, host-range restricted mutant  
 flavivirus, designated mutant 200, 201, wherein said virus  
 comprises charge-cluster-to-alanine mutations at amino acids  
 2687 and 2688 of the NS5 gene, where amino acid position is  
 given for the polyprotein of dengue virus type 4.

2. The flavivirus of claim 1, further comprising the  $\Delta$ 30  
 mutation.

3. The flavivirus of claim 1, wherein the flavivirus is a  
 dengue virus type 1.

4. The flavivirus of claim 1, wherein the flavivirus is a  
 dengue virus type 2.

5. The flavivirus of claim 1, wherein the flavivirus is a  
 dengue virus type 3.

6. The flavivirus of claim 1, wherein the flavivirus is a  
 dengue virus type 4.

7. The flavivirus of claim 1, wherein the flavivirus is a  
 chimeric virus.

8. The chimeric virus of claim 7 having a dengue 1 back-  
 bone.

9. The chimeric virus of claim 7 having a dengue 2 back-  
 bone.

10. The chimeric virus of claim 7 having a dengue 3 back-  
 bone.

11. The chimeric virus of claim 7 having a dengue 4 back-  
 bone.

12. The flavivirus of claim 1, wherein the phenotype is  
 temperature sensitivity in Vero cells or the human liver cell  
 line HuH-7.

13. A pharmaceutical composition comprising a pharma-  
 cologically acceptable vehicle and a flavivirus according to  
 any of claims 1-12.

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