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Lehmann

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(54) **CONSENSUS PHYTASES**
 (75) Inventor: **Martin Lehmann**, Grenzach-Wyhlen
 (DE)
 (73) Assignee: **DSM IP Assets B.V.**, TE Heerlen (NL)
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 15/00; C12N 5/00; C12N 1/20

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See application file for complete search history.

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Primary Examiner—Rebecca E. Prouty
Assistant Examiner—Delia M. Ramirez
 (74) *Attorney, Agent, or Firm*—Bryan Cave LLP

(57) ABSTRACT

A process for obtaining a consensus protein from a group of amino acid sequences of a defined protein family, proteins and polynucleotides so obtained, and compositions containing such proteins.

5 Claims, 25 Drawing Sheets

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50

<i>A. terreus</i> 9A-1	KhsDCNSVDh GYQCFPELSH kWGLYAPYFS LQDESPFP1D VPEDChITFV
<i>A. terreus</i> cbs	NhsDCTSVDr GYQCFPELSH kWGLYAPYFS LQDESPFP1D VPDDChITFV
<i>A. niger</i> var. <i>awamori</i>	NqsTCDTVdQ GYQCFSETSH LWGQYAPFFS LANESAISPD VPAGCrVTFA
<i>A. niger</i> T213	NqsSCDTVdQ GYQCFSETSH LWGQYAPFFS LANESVISPD VPAGCrVTFA
<i>A. niger</i> NRRL3135	NqsSCDTVdQ GYQCFSETSH LWGQYAPFFS LANESVISPE VPAGCrVTFA
<i>A. fumigatus</i> 13073	GSkSCDTVd1 GYQCSPATSH LWGQYSPFFS LEDE1SVSSK LPKDCrITLV
<i>A. fumigatus</i> 32722	GSkSCDTVd1 GYQCSPATSH LWGQYSPFFS LEDE1SVSSK LPKDCrITLV
<i>A. fumigatus</i> 58128	GSkSCDTVd1 GYQCSPATSH LWGQYSPFFS LEDE1SVSSK LPKDCrITLV
<i>A. fumigatus</i> 26906	GSkSCDTVd1 GYQCSPATSH LWGQYSPFFS LEDE1SVSSK LPKDCrITLV
<i>A. fumigatus</i> 32239	GSkACDTVE1 GYQCSPGTSH LWGQYSPFFS LEDE1SVSSD LPKDCrVTFV
<i>A. nidulans</i>	QNHSCNTADG GYQCFPNVSH VWGQYSPYFS IEQESAISeD VPHGceVTFV
<i>T. thermophilus</i>	DSHSCNTVEG GYQCrPEISH sWGQYSPFFS LADQSEISPD VPQNCKITFV
<i>M. thermophila</i>	ESRPCDTpD1 GFQCgTAISH FWGQYSPYFS VpSE1DaS.. IPDDCeVTFA

FIG. 1A

Consensus NSHSCDTVDG GYQCFPEISH LWGQYSPYFS LEDESAISPD
VPDDC-VTFV

Consensus phytase NSHSCDTVDG GYQCFPEISH LWGQYSPYFS LEDESAISPD
VPDDCRVTFV

51

100

A. terreus 9A-1 QVLARHGARs PThSKtKAYA AtIAAIQKSA TaFpGKYAFL
QSYNYSLDSE

A. terreus cbs QVLARHGARs PTDSKtKAYA AtIAAIQKNA TaLpGKYAFL
KSYNYSMGSE

A. niger var. *awamori* QVLSRHGARY PTESKgKkYS ALIEEIQQNV TtFDGKYAFL
KTYNYSLGAD

A. niger T213 QVLSRHGARY PTESKgKkYS ALIEEIQQNV TtFDGKYAFL
KTYNYSLGAD

A. niger NRRL3135 QVLSRHGARY PTDSKgKkYS ALIEEIQQNA TtFDGKYAFL
KTYNYSLGAD

A. fumigatus 13073 QVLSRHGARY PTSSKsKkYK kLVTAIQaNA TdFKGKFAFL
KTYNYTLGAD

A. fumigatus 32722 QVLSRHGARY PTSSKsKkYK kLVTAIQaNA TdFKGKFAFL
KTYNYTLGAD

A. fumigatus 58128 QVLSRHGARY PTSSKsKkYK kLVTAIQaNA TdFKGKFAFL
KTYNYTLGAD

A. fumigatus 26906 QVLSRHGARY PTSSKsKkYK kLVTAIQaNA TdFKGKFAFL
KTYNYTLGAD

A. fumigatus 32239 QVLSRHGARY PTASKsKkYK kLVTAIQKNA TeFKGKFAFL
ETNYNYTLGAD

FIG. 1B

<i>A. nidulans</i>	QVLSRHGARY PTESKsKAYS GLIEAIQKNA TsFwGQYAFL ESYNYTLGAD
<i>T. thermophilus</i>	QLLSRHGARY PTSSKtELYS QLISrIQKTA TaYKGyYAFL KDYrYqLGAN
<i>M. thermophila</i>	QVLSRHGARa PtlKRaaSYv DLIDrIHhGA IsYgPgYEFL RTYDYTLGAD
Consensus	QVLSRHGARY PTSSK-KAYS ALIEAIQKNA T-FKGKYAFL KTINYTLGAD
Consensus phytase	QVLSRHGARY PTSSKSKAYS ALIEAIQKNA TAFKGKYAFL KTINYTLGAD

101

150

<i>A. terreus</i> 9A-1	ELTPFGrNQL rDlGaQFYeR YNALTRhInP FVRATDASRV hESAekFVEG
<i>A. terreus</i> cbs	NLTPFGrNQL qDlGaQFYRR YDTLTRhInP FVRAADSSRV hESAekFVEG
<i>A. niger</i> var. <i>awamori</i>	DLTPFGEQEL VNSGIKfYQR YESLTRNIIP FIRSSGSSRV IASGEKFIEG
<i>A. niger</i> T213	DLTPFGEQEL VNSGIKfYQR YESLTRNIIP FIRSSGSSRV IASGEKFIEG
<i>A. niger</i> NRRL3135	DLTPFGEQEL VNSGIKfYQR YESLTRNIVP FIRSSGSSRV IASGKKFIEG
<i>A. fumigatus</i> 13073	DLTPFGEQQL VNSGIKfYQR YKALARSVVP FIRASGSDRV IASGEKFIEG
<i>A. fumigatus</i> 32722	DLTPFGEQQL VNSGIKfYQR YKALARSVVP FIRASGSDRV IASGEKFIEG

FIG. 1C

<i>A. fumigatus</i> 58128	DLTPFGEQQL VNSGIKFYQR YKALARSVVP FIRASGSDRV IASGEKFIEG
<i>A. fumigatus</i> 26906	DLTAFGEQQL VNSGIKFYQR YKALARSVVP FIRASGSDRV IASGEKFIEG
<i>A. fumigatus</i> 32239	DLTPFGEQQM VNSGIKFYQK YKALAgSVVP FIRSSGSDRV IASGEKFIEG
<i>A. nidulans</i>	DLTiFGENQM VDSGaKFYRR YKNLARKnTP FIRASGSDRV VASAEKFING
<i>T. thermophilus</i>	DLTPFGENQM IQlGIKFYnH YKSLARNaVP FVRCSGSDRV IASGr1FIEG
<i>M. thermophila</i>	ELTRtGQQQM VNSGIKFYRR YRALARKsIP FVRTAGqDRV VhSAENFTQG
Consensus	DLTPFGENQM VNSGIKFYRR YKALARK-VP FVRASGSDRV IASAEKFIEG
Consensus phytase	DLTPFGENQM VNSGIKFYRR YKALARKIVP FIRASGSDRV IASAEKFIEGAA

FIG. 1D

151

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<i>A. terreus</i> 9A-1	FQTARqDDHh ANpHQSPPrV DVaIPEGSAY NNTLEHSICT
AFES...STV	
<i>A. terreus</i> cbs	FQNARqGDPH ANpHQSPPrV DVVIPEGTAY NNTLEHSICT
AFEA...STV	
<i>A. niger</i> var. <i>awamori</i>	FQSTKLkDPr AqpgQSSPkI DVVISEASSs NNTLDPGTCT
VFED...SEL	
<i>A. niger</i> T213	FQSTKLkDPr AqpgQSSPkI DVVISEASSs NNTLDPGTCT
VFED...SEL	
<i>A. niger</i> NRRL3135	FQSTKLkDPr AqpgQSSPkI DVVISEASSs NNTLDPGTCT
VFED...SEL	
<i>A. fumigatus</i> 13073	FQqAKLADPG A.TNRAAPAI SVIIPeSETF NNTLDHGVCT
kFEA...SQL	
<i>A. fumigatus</i> 32722	FQqAKLADPG A.TNRAAPAI SVIIPeSETF NNTLDHGVCT
kFEA...SQL	
<i>A. fumigatus</i> 58128	FQqAKLADPG A.TNRAAPAI SVIIPeSETF NNTLDHGVCT
kFEA...SQL	
<i>A. fumigatus</i> 26906	FQqAKLADPG A.TNRAAPAI SVIIPeSETF NNTLDHGVCT
kFEA...SQL	
<i>A. fumigatus</i> 32239	FQqANVADPG A.TNRAAPVI SVIIPeSETY NNTLDHsvCT
NFEA...SEL	
<i>A. nidulans</i>	FRKAQLhDHG S..gQATPVV NVIIPeIdGF NNTLDHSTCV
SFEN...DEr	
<i>T. thermophilus</i>	FQSAKvLDPh SDkHDAPPTI NVIIeEGPSY NNTLDtGSCP
VFED...SSg	
<i>M. thermophila</i>	FHSAlLADRG STvRPTlPyd mVVIPEtAGa NNTLHNDlCT
AFEEgpySTI	

FIG. 1E

Consensus FQSAKLADPG S-PHQASPVI NVIIPEGSGY NNTLDHGTCT
 AFED---SEL

Consensus phytase FQSAKLADPG SQPHQASPVI DVIIPEGSGY NNTLDHGTCT
 AFED...SEL

201

250

A. terreus 9A-1 GDDAVANFTA VFAPAIaQRL EADLPGVqLS TDDVnLMAM
 CPFETVSlTD

A. terreus cbs GDAAADNFTA VFAPAIakRL EADLPGVqLS ADDVnLMAM
 CPFETVSlTD

A. niger var. *awamori* ADTVEANFTA TFAPSIRQRL ENDLSGVTLT DTEVTyLMDM
 CSFDTISStST

A. niger T213 ADTVEANFTA TFAPSIRQRL ENDLSGVTLT DTEVTyLMDM
 CSFDTISStST

A. niger NRRL3135 ADTVEANFTA TFVPSIRQRL ENDLSGVTLT DTEVTyLMDM
 CSFDTISStST

A. fumigatus 13073 GDEVAANFTA lFAPDIRARa EkHLPGVTLT DEDVVsLMDM
 CSFDTVARTS

A. fumigatus 32722 GDEVAANFTA lFAPDIRARa EkHLPGVTLT DEDVVsLMDM
 CSFDTVARTS

A. fumigatus 58128 GDEVAANFTA lFAPDIRARa EkHLPGVTLT DEDVVsLMDM
 CSFDTVARTS

A. fumigatus 26906 GDEVAANFTA lFAPDIRARa KkHLPGVTLT DEDVVsLMDM
 CSFDTVARTS

A. fumigatus 32239 GDEVEANFTA lFAPAIRARI EkHLPGVqLT DDDVVsLMDM
 CSFDTVARTA

FIG. 1F

<i>A. nidulans</i>	ADEiEANFTA IMGPPiRkRL ENDLPGIKLT NENViYLM DM
CSFDTMARTA	
<i>T. thermophilus</i>	GHDAQEKFAk qFAPAiLEKI KDHLPGVDLA vSDVpyLMDL
CPFETLARNh	
<i>M. thermophila</i>	GDDAQDTYlS TFAGPiRARV NANLPGANLT DADTVaLMDL
CPFETVAsSS	
Consensus	GDDAEANFTA TFAPAIRARL EADLPGVTLT DEDVV-LMDM
CPFETVARTS	
Consensus phytase	GDDVEANFTA LFAPAIRARL EADLPGVTLT DEDVVYLMDM
CPFETVARTS	
	251
300	
<i>A. terreus</i> 9A-1DAhTLSPFC DLFTAtEWtq YNYLlSLDKY
YGYGGGNPLG	
<i>A. terreus</i> cbsDAhTLSPFC DLFTAaEWtq YNYLlSLDKY
YGYGGGNPLG	
<i>A. niger</i> var. <i>awamori</i>vDTKLSPFC DLFTHdEWih YDYLQSLkKY
YGHGAGNPLG	
<i>A. niger</i> T213vDTKLSPFC DLFTHdEWih YDYLRSLkKY
YGHGAGNPLG	
<i>A. niger</i> NRRL3135vDTKLSPFC DLFTHdEWin YDYLQSLkKY
YGHGAGNPLG	
<i>A. fumigatus</i> 13073DASQLSPFC QLFTHnEWkk YNYLQSLGKY
YGYGAGNPLG	
<i>A. fumigatus</i> 32722DASQLSPFC QLFTHnEWkk YNYLQSLGKY
YGYGAGNPLG	

FIG. 1G

A. fumigatus 58128DASQLSPFC QLFTHnEWkk YNYLQSLGKY
YGYGAGNPLG

A. fumigatus 26906DASQLSPFC QLFTHnEWkk YNYLQSLGKY
YGYGAGNPLG

A. fumigatus 32239DASELSPFC AIFTHnEWkk YDYLQSLGKY
YGYGAGNPLG

A. nidulansHGTELSRFC AIFTEkEWlq YDYLQSLSKY
YGYGAGSPLG

T. thermophilusTDT.LSPFC ALsTQeEWqa YDYYQSLGKY
YGnGGGNPLG

M. thermophila sdpatadagg gNGrpLSPFC rLFSEsEWra YDYLQSVGKW
YGYGPGNPLG

Consensus ----- -DATELSRFC ALFTE-EW-- YDYLQSLGKY
YGYGAGNPLG

Consensus phytaseDATELSRFC ALFTHDEWRQ YDYLQSLGKY
YGYGAGNPLG

FIG. 1H

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<i>A. terreus</i> 9A-1	PVQGVGWaNE	LMARLTRAPV	HDHTCVNNTL	DASPATFPLN
ATLYADFSHD				
<i>A. terreus</i> cbs	PVQGVGWaNE	LIARLTRSPV	HDHTCVNNTL	DANPATFPLN
ATLYADFSHD				
<i>A. niger</i> var. <i>awamori</i>	PTQGVGYaNE	LIARLTHSPV	HDDTSSNHTL	DSNPATFPLN
STLYADFSHD				
<i>A. niger</i> T213	PTQGVGYaNE	LIARLTHSPV	HDDTSSNHTL	DSNPATFPLN
STLYADFSHD				
<i>A. niger</i> NRRL3135	PTQGVGYaNE	LIARLTHSPV	HDDTSSNHTL	DSSPATFPLN
STLYADFSHD				
<i>A. fumigatus</i> 13073	PAQGIGFtNE	LIARLTRSPV	QDHTSTNsTL	vSNPATFPLN
ATMYVDFSHD				
<i>A. fumigatus</i> 32722	PAQGIGFtNE	LIARLTRSPV	QDHTSTNsTL	vSNPATFPLN
ATMYVDFSHD				
<i>A. fumigatus</i> 58128	PAQGIGFtNE	LIARLTRSPV	QDHTSTNsTL	vSNPATFPLN
ATMYVDFSHD				
<i>A. fumigatus</i> 26906	PAQGIGFtNE	LIARLTRSPV	QDHTSTNsTL	vSNPATFPLN
ATMYVDFSHD				
<i>A. fumigatus</i> 32239	PAQGIGFtNE	LIARLTNSPV	QDHTSTNsTL	DSDPATFPLN
ATLYVDFSHD				
<i>A. nidulans</i>	PAQGIGFtNE	LIARLTQSPV	QDNTSTNHTL	DSNPATFPLD
rKLYADFSHD				
<i>T. thermophilus</i>	PAQGVGFvNE	LIARMTHSPV	QDYTTVNHTL	DSNPATFPLN
ATLYADFSHD				
<i>M. thermophila</i>	PTQGVGFvNE	LLARLAgvPV	RDgTSTNRTL	DGDPrTFPLG
rPLYADFSHD				

FIG. 11

Consensus PAQGVGF-NE LIARLTHSPV QDHTSTNHTL DSNPATFPLN
ATLYADFSHD

Consensus phytase PAQGVGFANE LIARLTRSPV QDHTSTNHTL DSNPATFPLN
ATLYADFSHD

351

400

A. terreus 9A-1 SNLVSIFWAL GLYNGTAPLS qTSVESVSQT DGYAAAWTVP
FAARAYVEMM

A. terreus cbs SNLVSIFWAL GLYNGTkPLS qTTVEDITrT DGYAAAWTVP
FAARAYIEMM

A. niger var. *awamori* NGIISILFAL GLYNGTkPLS TTTVENITQT DGFSSAWTVP
FASRLYVEMM

A. niger T213 NGIISILFAL GLYNGTkPLS TTTVENITQT DGFSSAWTVP
FASRLYVEMM

A. niger NRRL3135 NGIISILFAL GLYNGTkPLS TTTVENITQT DGFSSAWTVP
FASRLYVEMM

A. fumigatus 13073 NSMVSIFFAL GLYNGTEPLS rTSVESaKEl DGYSASWVVP
FGARAYFetM

A. fumigatus 32722 NSMVSIFFAL GLYNGTGPLS rTSVESaKEl DGYSASWVVP
FGARAYFetM

A. fumigatus 58128 NSMVSIFFAL GLYNGTEPLS rTSVESaKEl DGYSASWVVP
FGARAYFetM

A. fumigatus 26906 NSMVSIFFAL GLYNGTEPLS rTSVESaKEl DGYSASWVVP
FGARAYFetM

A. fumigatus 32239 NGMIPIFFAM GLYNGTEPLS qTSeESTKES NGYSASWAVP
FGARAYFetM

FIG. 1J

<i>A. nidulans</i>	NSMISIFFAM GLYNGTQPLS mDSVESIQEm DGYAASWTVP
FGARAYFELM	
<i>T. thermophilus</i>	NTMTSIFaAL GLYNGTAKLS TTEIKSIEET DGYSAAWTVP
FGGRAYIEMM	
<i>M. thermophila</i>	NDMMGVLgAL GaYDGVPPLD KTArrDpEEl GGYAASWAVP
FAARiYVEKM	
Consensus	NSMISIFFAL GLYNGTAPLS TTSVESIEET DGYAASWTVP
FGARAYVEMM	
Consensus phytase	NSMISIFFAL GLYNGTAPLS TTSVESIEET DGYSASWTVP
FGARAYVEMM	
	401
450	
<i>A. terreus</i> 9A-1	QC..... .RAEKE PLVRVLVNDR VMPLHGCPD
KLGRCKrDAF	
<i>A. terreus</i> cbs	QC..... .RAEKQ PLVRVLVNDR VMPLHGCAVD
NLGRCKrDDF	
<i>A. niger</i> var. <i>awamori</i>	QC..... .QAEQE PLVRVLVNDR VVPLHGCPID
aLGRCTrDSF	
<i>A. niger</i> T213	QC..... .QAEQE PLVRVLVNDR VVPLHGCPID
aLGRCTrDSF	
<i>A. niger</i> NRRL3135	QC..... .QAEQE PLVRVLVNDR VVPLHGCPVD
aLGRCTrDSF	
<i>A. fumigatus</i> 13073	QC..... .KSEKE PLVRALINDR VVPLHGCDVD
KLGRCKLNDF	
<i>A. fumigatus</i> 32722	QC..... .KSEKE PLVRALINDR VVPLHGCDVD
KLGRCKLNDF	

FIG. 1K

<i>A. fumigatus</i> 58128	QC..... .KSEKE SLVRALINDR VVPLHGCDVD
KLGRCKLNDF	
<i>A. fumigatus</i> 26906	QC..... .KSEKE PLVRALINDR VVPLHGCDVD
KLGRCKLNDF	
<i>A. fumigatus</i> 32239	QC..... .KSEKE PLVRALINDR VVPLHGCAVD
KLGRCKLKDF	
<i>A. nidulans</i>	QC..... .E.KKE PLVRVLVNDR VVPLHGCAVD
KFGRCTLDDW	
<i>T. thermophilus</i>	QC..... .DDSDE PVVRVLVNDR VVPLHGCEVD
SLGRCKrDDF	
<i>M. thermophila</i>	RCsggggggg ggegrQEKDE eMVRVLVNDR VMTLkGCGAD
ErGMCTLErF	
Consensus	QC----- ----QAEKE PLVRVLVNDR VVPLHGCAVD
KLGRCKLDDF	
Consensus phytase	QC..... .QAEKE PLVRVLVNDR VVPLHGCAVD
KLGRCKRDDF	

FIG. 1L

	451	471
<i>A. terreus</i> 9A-1	VAGLSFAQAG	GNWADCF~~~ ~
<i>A. terreus</i> cbs	VEGLSFARAG	
	GNWAECF~~~ ~	
<i>A. niger</i> var. <i>awamori</i>	VrGLSFARSG	GDWAECsA~~ ~
<i>A. niger</i> T213	VrGLSFARSG	GDWAECFA~~ ~
<i>A. niger</i> NRRL3135	VrGLSFARSG	
	GDWAECFA~~ ~	
<i>A. fumigatus</i> 13073	VKGLSWARSG	GNWGECSFS~~ ~
<i>A. fumigatus</i> 32722	VKGLSWARSG	GNWGECSFS~~ ~
<i>A. fumigatus</i> 58128	VKGLSWARSG	GNWGECSFS~~ ~
<i>A. fumigatus</i> 26906	VKGLSWARSG	GNWGECSFS~~ ~
<i>A. fumigatus</i> 32239	VKGLSWARSG	
	GNSEQSFS~~ ~	
<i>A. nidulans</i>	VEGLNFARSG	GNWkTCFTl~ ~
<i>T. thermophilus</i>	VrGLSFARqG	GNWEGCYAas e
<i>M. thermophila</i>	IESMAFARGN	GKWDlCFA~~ ~
Consensus	VEGLSFARSG	GNWAECFA-- -
Consensus phytase	VEGLSFARSG	GNWAECFA.. .

FIG. 1M

CP-1

Eco RI M G V F V V L L S I A T L F G S T

TATATGAATTCATGGGCGTGTTTCGTCGTGCTACTGTCCATTGCCACCTTGTTTCGGTTCCA

1 -----+-----+-----+-----+-----+-----+-----+

60

ATATACTTAAGTACCCGCACAAGCAGCACGATGACAGGTAACGGTGAACAAGCCAAGGT

S G T A L G P R G N S H S C D T V D G G

CATCCGGTACCGCCTTGGGTCCTCGTGGTAATTCTCACTCTTGTGACACTGTTGACGGTG

61 -----+-----+-----+-----+-----+-----+-----+

120

GTAGGCCATGGCGGAACCCAGGAGCACCATTAAGAGTGAGAACACTGTGACAACTGCCAC

CP-2

CP-3

Y Q C F P E I S H L W G Q Y S P Y F S L

GTTACCAATGTTTCCCAGAAATTTCTCACTTGTGGGGTCAATACTCTCCATACTTCTCTT

121 -----+-----+-----+-----+-----+-----+-----+

180

CAATGGTTACAAAGGGTCTTTAAAGAGTGAACACCCCAGTTATGAGAGGTATGAAGAGAA

E D E S A I S P D V P D D C R V T F V Q

TGGAAGACGAATCTGCTATTTCTCCAGACGTTCCAGACGACTGTAGAGTTACTTTCGTTT

181 -----+-----+-----+-----+-----+-----+-----+

240

ACCTTCTGCTTAGACGATAAAGAGGTCTGCAAGGTCTGCTGACATCTCAATGAAAGCAAG

CP-4

CP-5

V L S R H G A R Y P T S S K S K A Y S A

AAGTTTGTCTAGACACGGTGCTAGATACCCAACCTTCTTCTAAGTCTAAGGCTTACTCTG

FIG. 2A

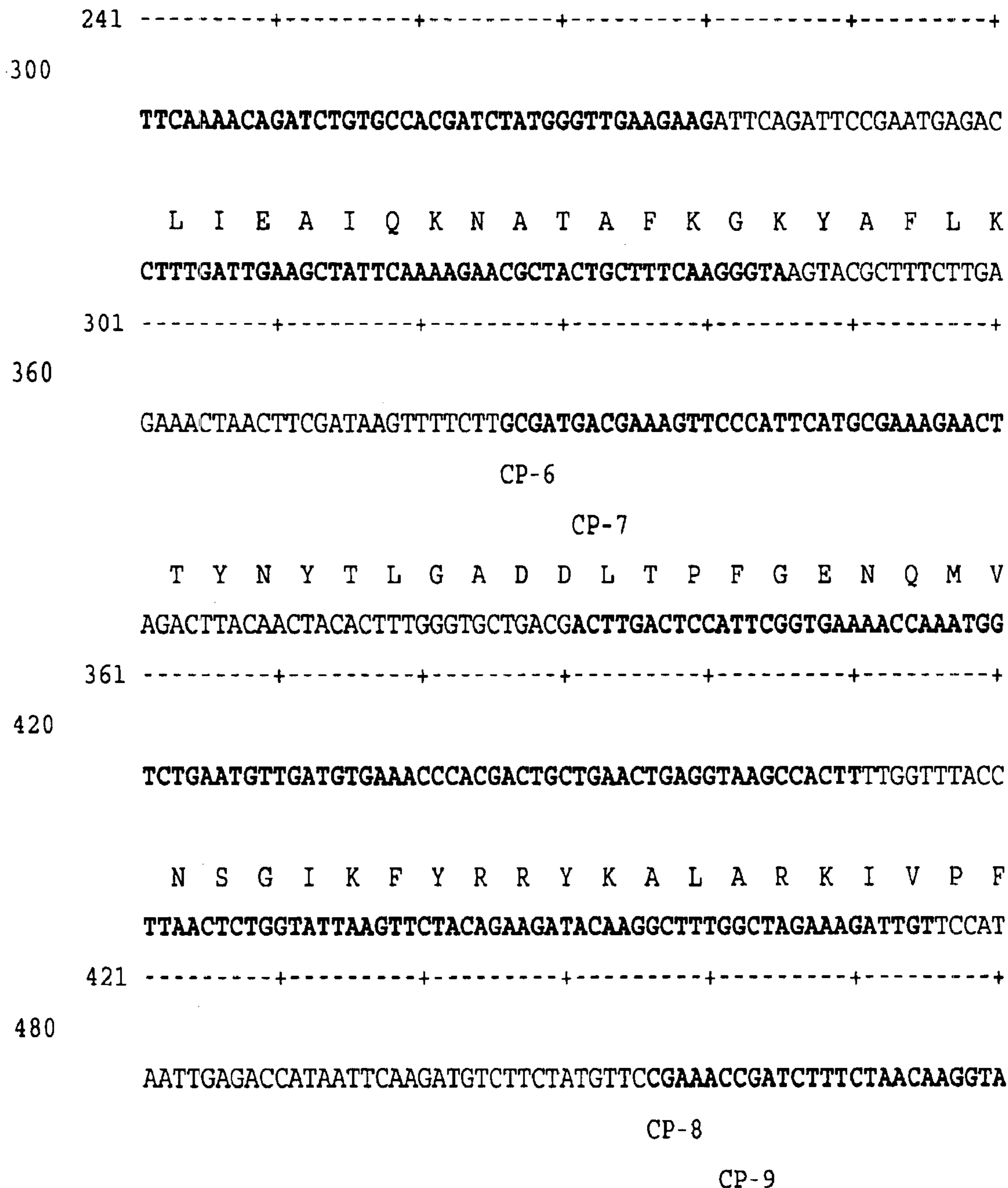


FIG. 2B

I R A S G S D R V I A S A E K F I E G F
TCATTAGAGCTTCTGGTTCTGACAGAGTTATTGCTTCTGCTGAAAAGTTCATTGAAGGTT
481 -----+-----+-----+-----+-----+-----+
540
AGTAATCTCGAAGACCAAGACTGTCTCAATAACGAAGACGACTTTTCAAGTAACTTCCAA

Q S A K L A D P G S Q P H Q A S P V I D
TCCAATCTGCTAAGTTGGCTGACCCAGGTTCTCAACCACACCAAGCTTCTCCAGTTATTG
541 -----+-----+-----+-----+-----+-----+
600
AGGTTAGACGATTCAACCGACTGGGTCCAAGAGTTGGTGTGGTTCGAAGAGGTCAATAAC
CP-10
CP-11
V I I P E G S G Y N N T L D H G T C T A
ACGTTATTATTCCAGAAGGaTCcGGTTACAACAACACTTTGGACCACGGTACTTGTACTG
601 -----+-----+-----+-----+-----+-----+
660
TGCAATAATAAGGTCTTCCTAGgCCAATGTTGTTGTGAAACCTGGTGCCATGAACATGAC

FIG. 2C

F E D S E L G D D V E A N F T A L F A P
 CTTTCGAAGACTCTGAATTGGGTGACGACGTTGAAGCTAACTTCACTGCTTTGTTGCTC
 661 -----+-----+-----+-----+-----+-----+-----+
 720
 GAAAGCTTCTGAGACTTAACCCACTGCTGCAACTTCGATTGAAGTGACGAAACAAGCGAG
 CP-12
 A I R A R L E A D L P G V T L T D E D V
 CAGCTATTAGAGCTAGATTGGAAGCTGACTTGCCAGGTGTTACTTTGACTGACGAAGACG
 721 -----+-----+-----+-----+-----+-----+-----+
 780
 GTCGATAATCTCGATCTAACCTTCGACTGAACGGTCCACAATGAAACTGACTGCTTCTGC
 CP-13
 V Y L M D M C P F E T V A R T S D A T E
 TTGTTTACTTGATGGACATGTGTCCATTCGAAACTGTTGCTAGAACTTCTGACGCTACTG
 781 -----+-----+-----+-----+-----+-----+-----+
 840
 AACAAATGAACTACCTGTACACAGGTAAGCTTTGACAACGATCTTGAAGACTGCGATGAC
 L S P F C A L F T H D E W R Q Y D Y L Q
 AATTGTCTCCATTCTGTGCTTTGTTCACTCACGACGAATGGAGACAATACGACTACTTGC
 841 -----+-----+-----+-----+-----+-----+-----+
 900
 TTAACAGAGGTAAGACACGAAACAAGTGAGTGCTGCTTACCTCTGTTATGCTGATGAACG
 CP-14
 CP-15
 S L G K Y Y G Y G A G N P L G P A Q G V

FIG. 2D

AATCTTTGGGTAAGTACTACGGTTACGGTGCTGGTAACCCATTGGGTCCAGCTCAAGGTG
 901 -----+-----+-----+-----+-----+-----+-----+
 960
 TTAGAAACCCATTCATGATGCCAATGCCACGACCATTGGGTAACCCAGGTCGAGTTCCAC

 G F A N E L I A R L T R S P V Q D H T S
 TTGGTTTCGCTAACGAATTGATTGCTAGATTGACTAGATCTCCAGTTCAAGACCACACTT
 961 -----+-----+-----+-----+-----+-----+-----+
 1020
 AACCAAAGCGATTGCTTAACTAACGATCTAACTGATCTAGAGGTCAAGTTCTGGTGTGAA

 CP-16
 CP-17
 T N H T L D S N P A T F P L N A T L Y A
 CTACTAACCACTTTGGACTCTAACCCAGCTACTTCCCATTGAACGCTACTTTGTACG
 1021 -----+-----+-----+-----+-----+-----+-----+
 1080
 GATGATTGGTGTGAAACCTGAGATTGGGTGATGAAAGGGTAACTTGCATGAAACATGC

 D F S H D N S M I S I F F A L G L Y N G
 CTGACTTCTCTCACGACAACCTCTATGATTTCTATTTTCTTCGCTTTGGGTTTGTACAACG
 1081 -----+-----+-----+-----+-----+-----+-----+
 1140
 GACTGAAGAGAGTGCTGTTGAGATACTAAAGATAAAGAAGCGAAACCCAAACATGTTGC

 CP-18
 CP-19
 T A P L S T T S V E S I E E T D G Y S A
 GTACTGCTCCATTGTCTACTACTTCTGTTGAATCTATTGAAGAACTGACGGTTACTCTG

FIG. 2E

1141 -----+-----+-----+-----+-----+-----+
1200
CATGACGAGGTAACAGATGATGAAGACAACTTAGATAACTTCTTTGACTGCCAATGAGAC

S W T V P F G A R A Y V E M M Q C Q A E
CTTCTTGGACTGTTCCATTCGGTGCTAGAGCTTACGTTGAAATGATGCAATGTCAAGCTG
1201 -----+-----+-----+-----+-----+-----+
1260
GAAGAACCTGACAAGGTAAGCCACGATCTCGAATGCAACTTTACTACGTTACAGTTCGAC

CP-20
CP-21
K E P L V R V L V N D R V V P L H G C A
AAAAGGAACCATTGGTTAGAGTTTTGGTTAACGACAGAGTTGTTCCATTGCACGGTTGTG
1261 -----+-----+-----+-----+-----+-----+
1320
TTTTCCTTGGTAACCAATCTCAAACCAATGCTGTCTCAACAAGGTAACGTGCCAACAC

FIG. 2F

V D K L G R C K R D D F V E G L S F A R
CTGTTGACAAGTTGGGTAGATGTAAGAGAGACGACTTCGTTGAAGGTTTGTCTTTCGCTA
1321 -----+-----+-----+-----+-----+-----+-----+
1380
GACAACTGTTCAACCCATCTACATTCTCTCTGCTGAAGCAACTTCCAAACAGAAAGCGAT
CP-22
S G G N W A E C F A * Eco RI
GATCTGGTGGTAACTGGGCTGAATGTTTCGCTTAAGAATTCATATA
1381 -----+-----+-----+-----+----- 1426
CTAGACCACCATTGACCCGACTTACAAAGCGAATTCTTAAGTATAT

FIG. 2G

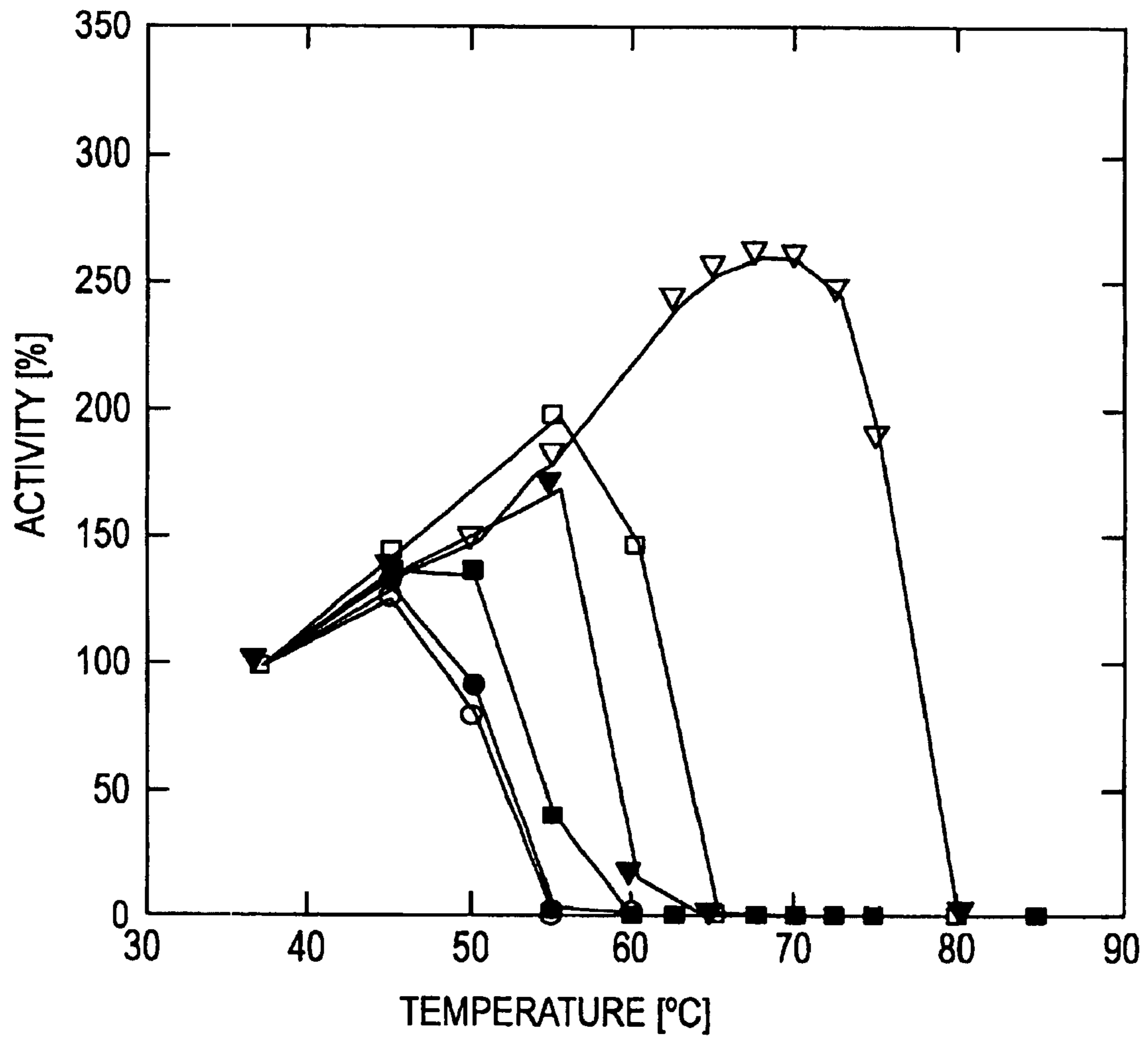


FIG. 3

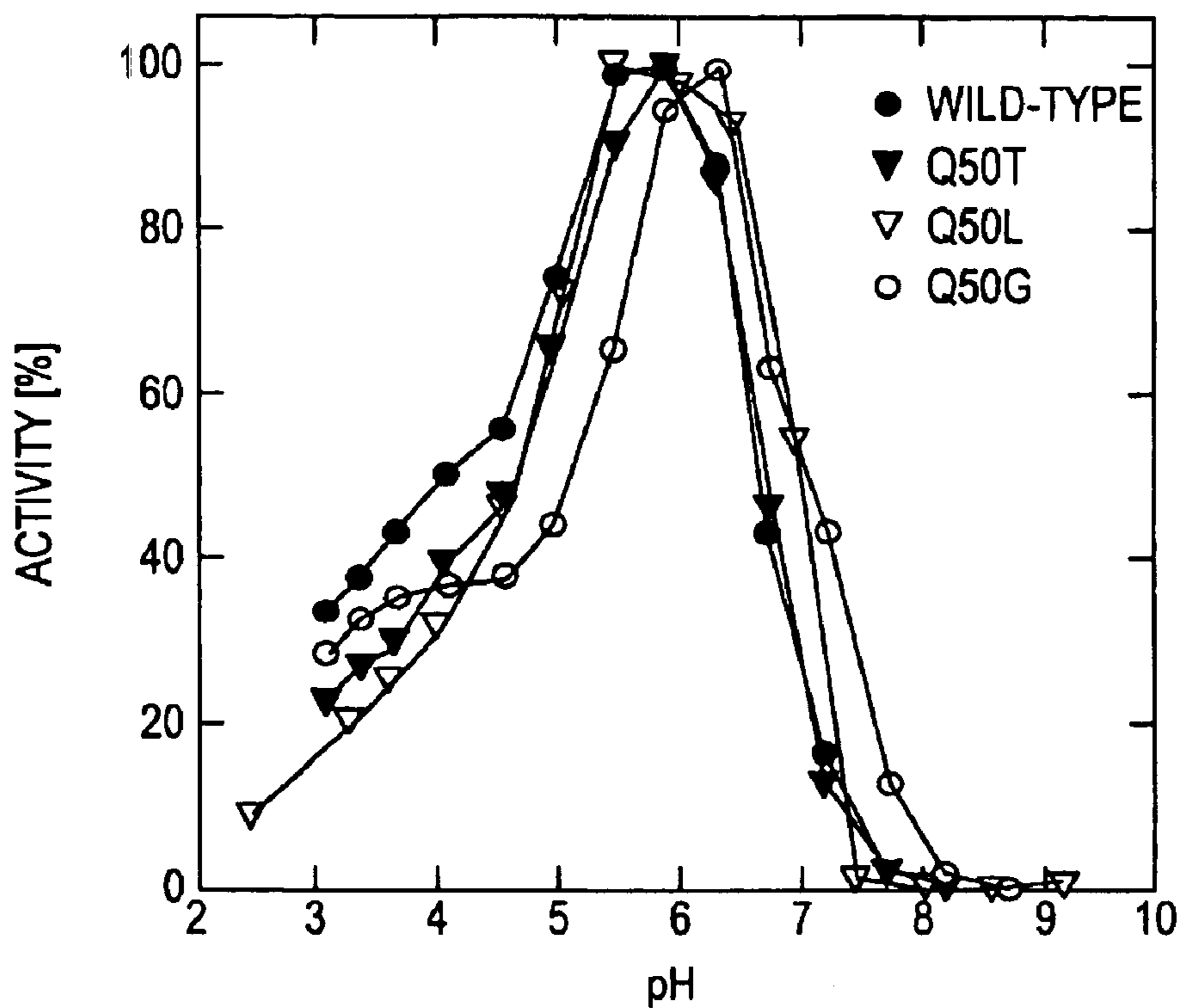


FIG. 4A

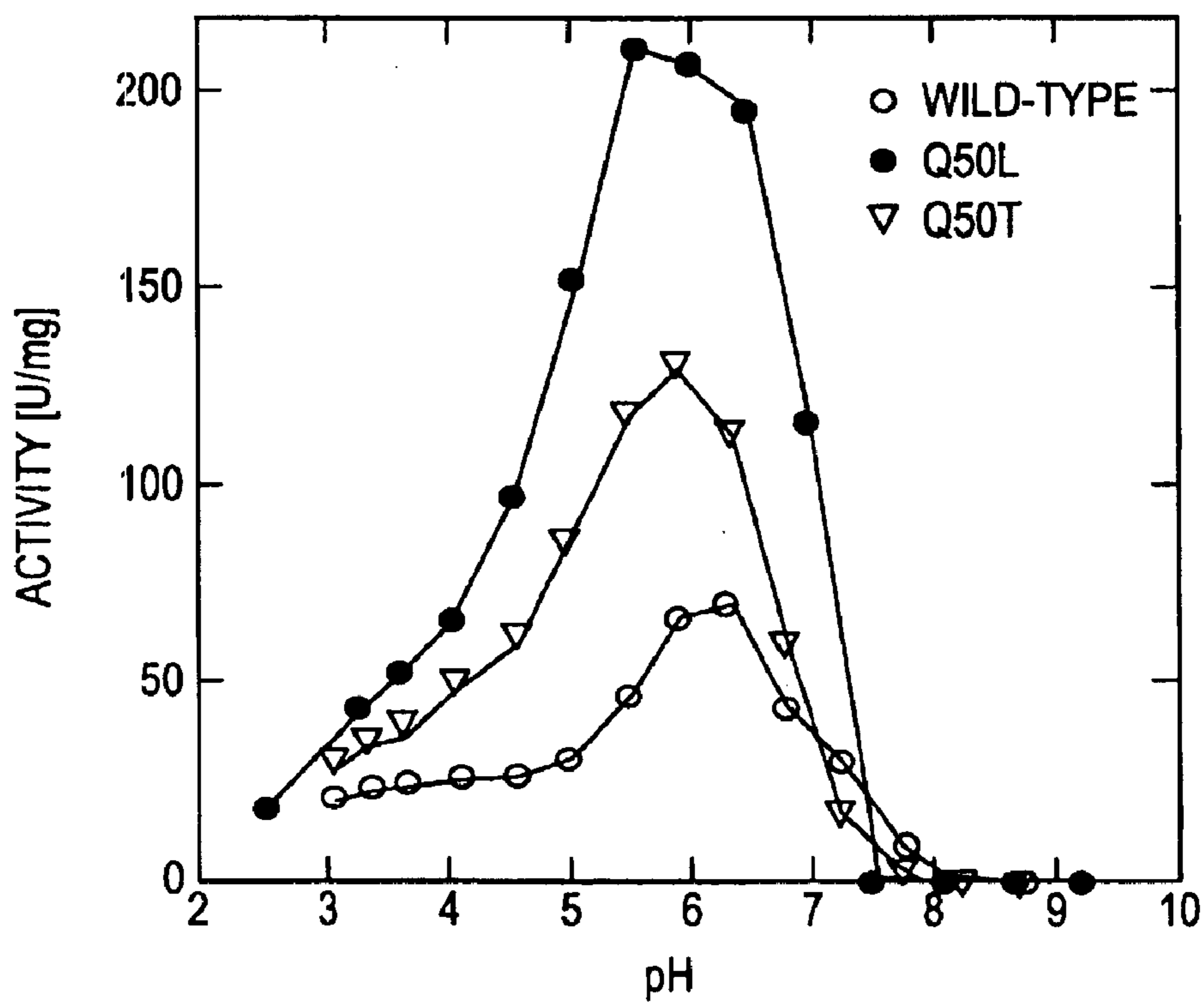


FIG. 4B

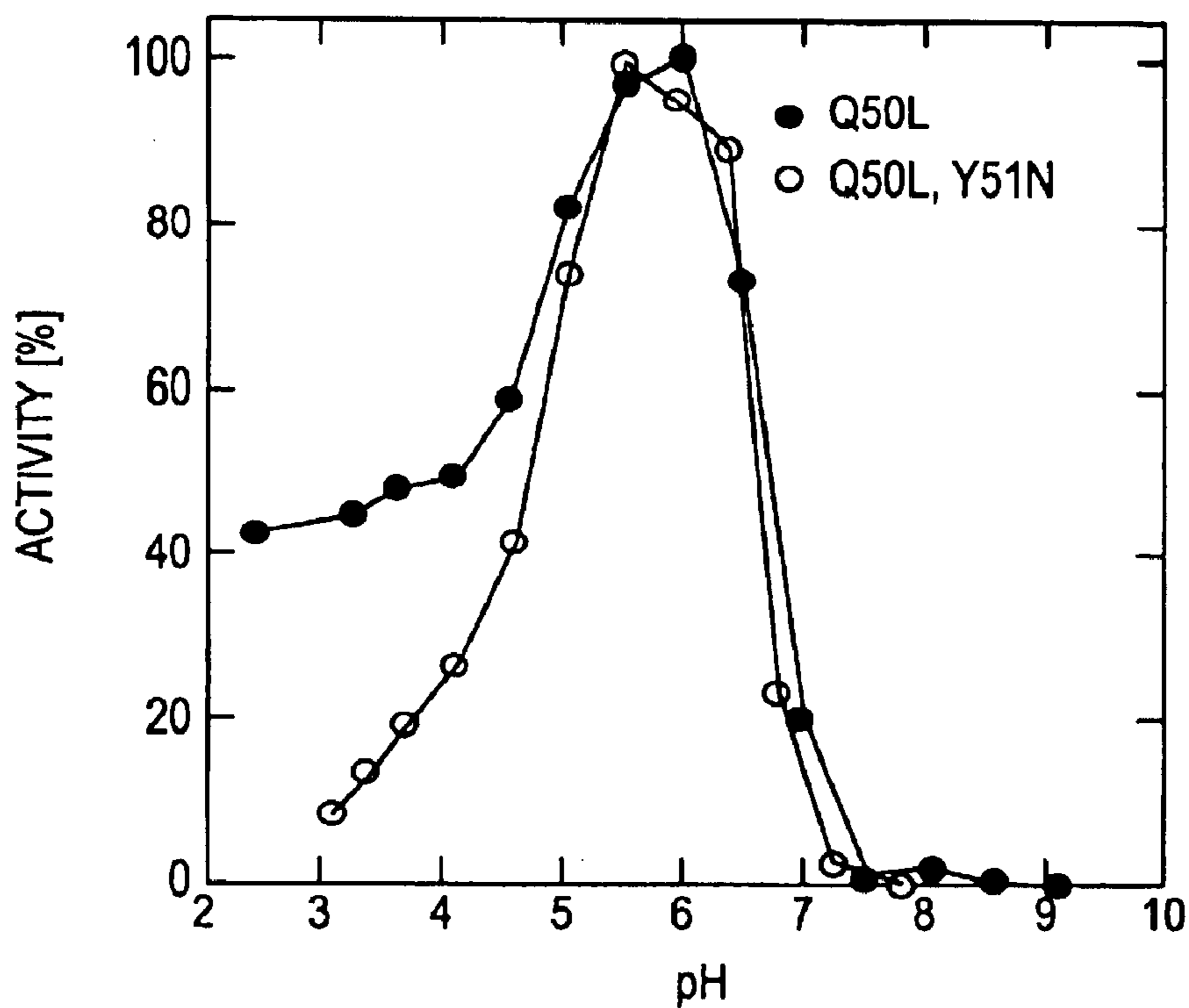


FIG. 5A

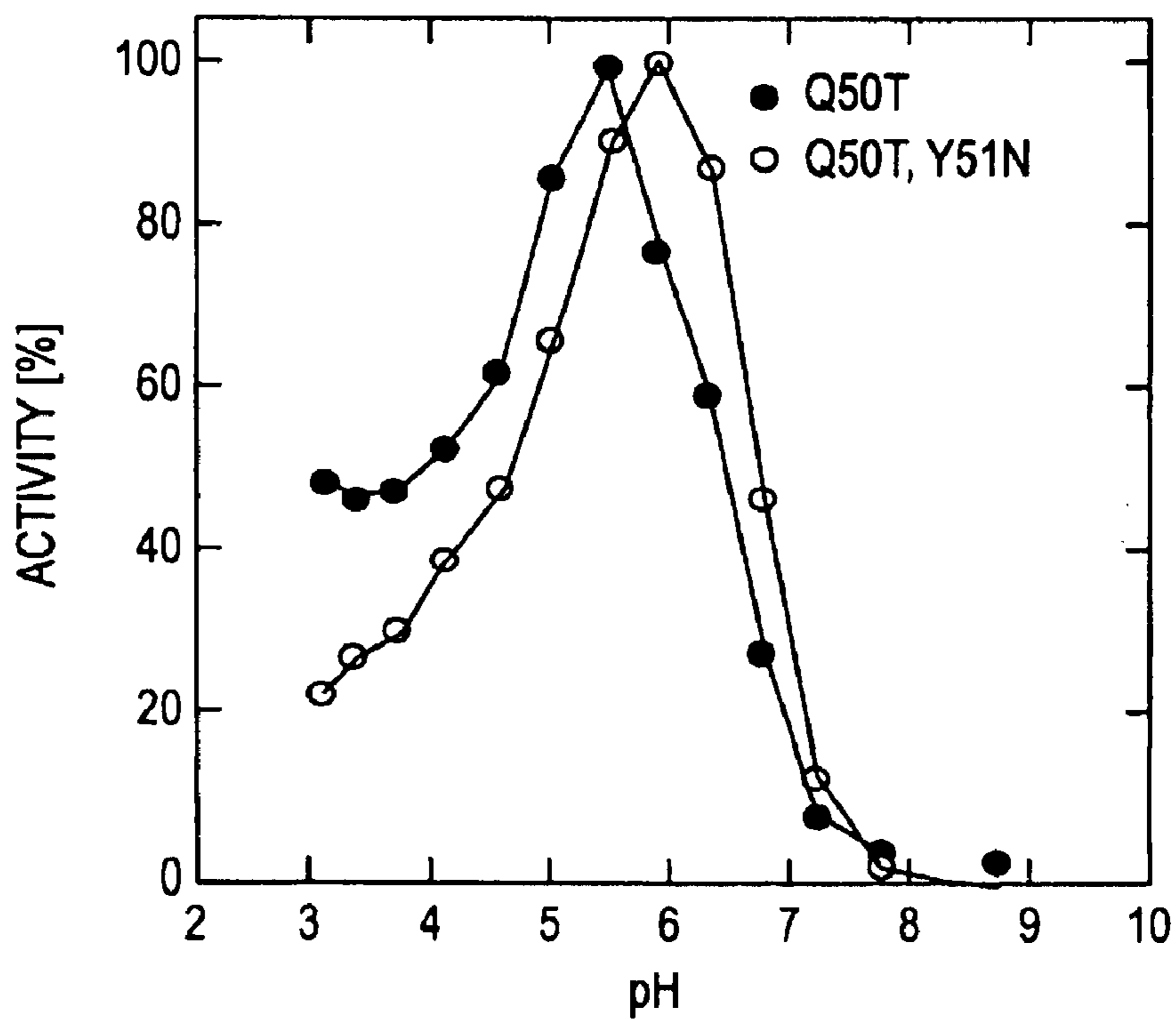


FIG. 5B

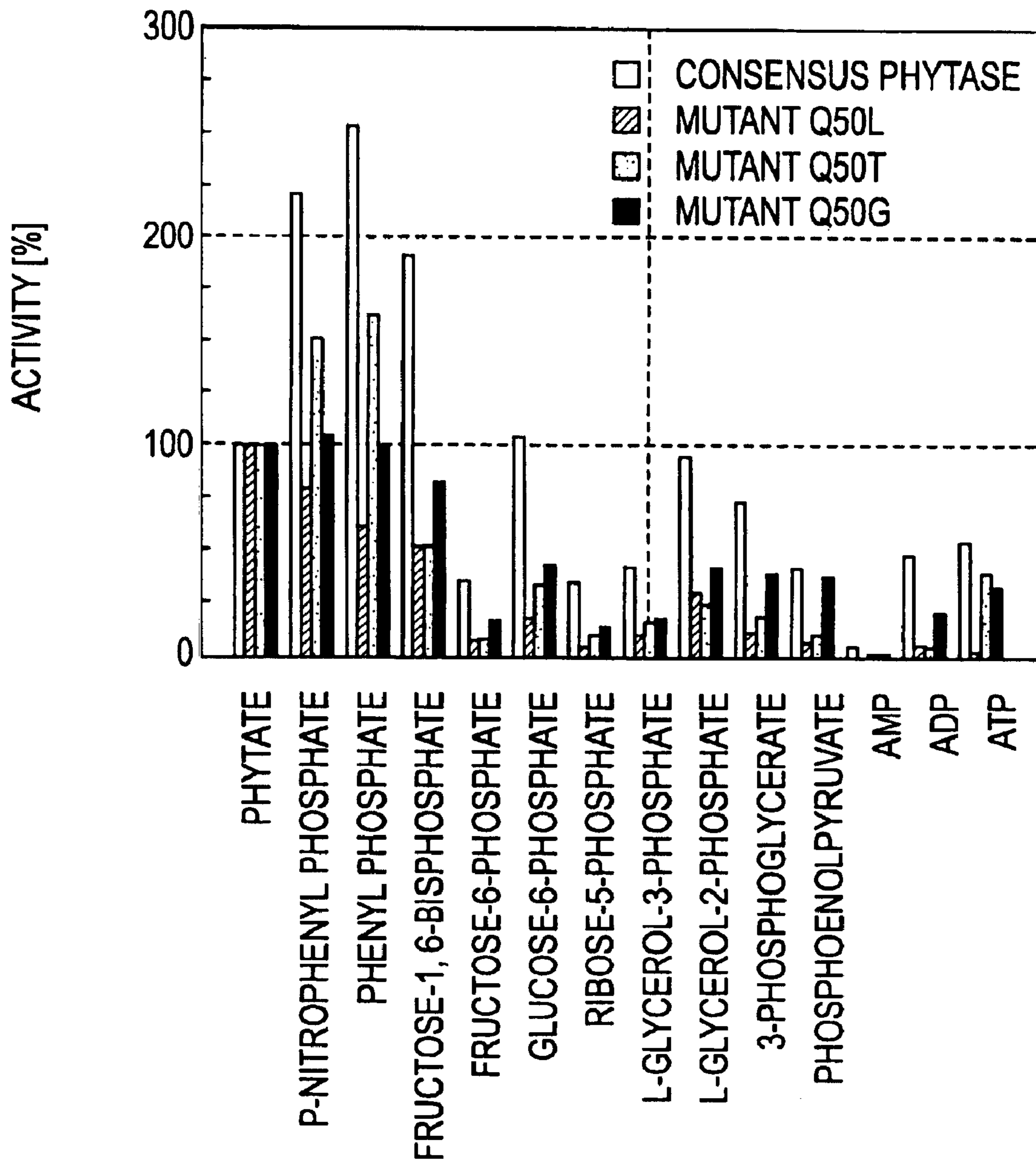


FIG. 6

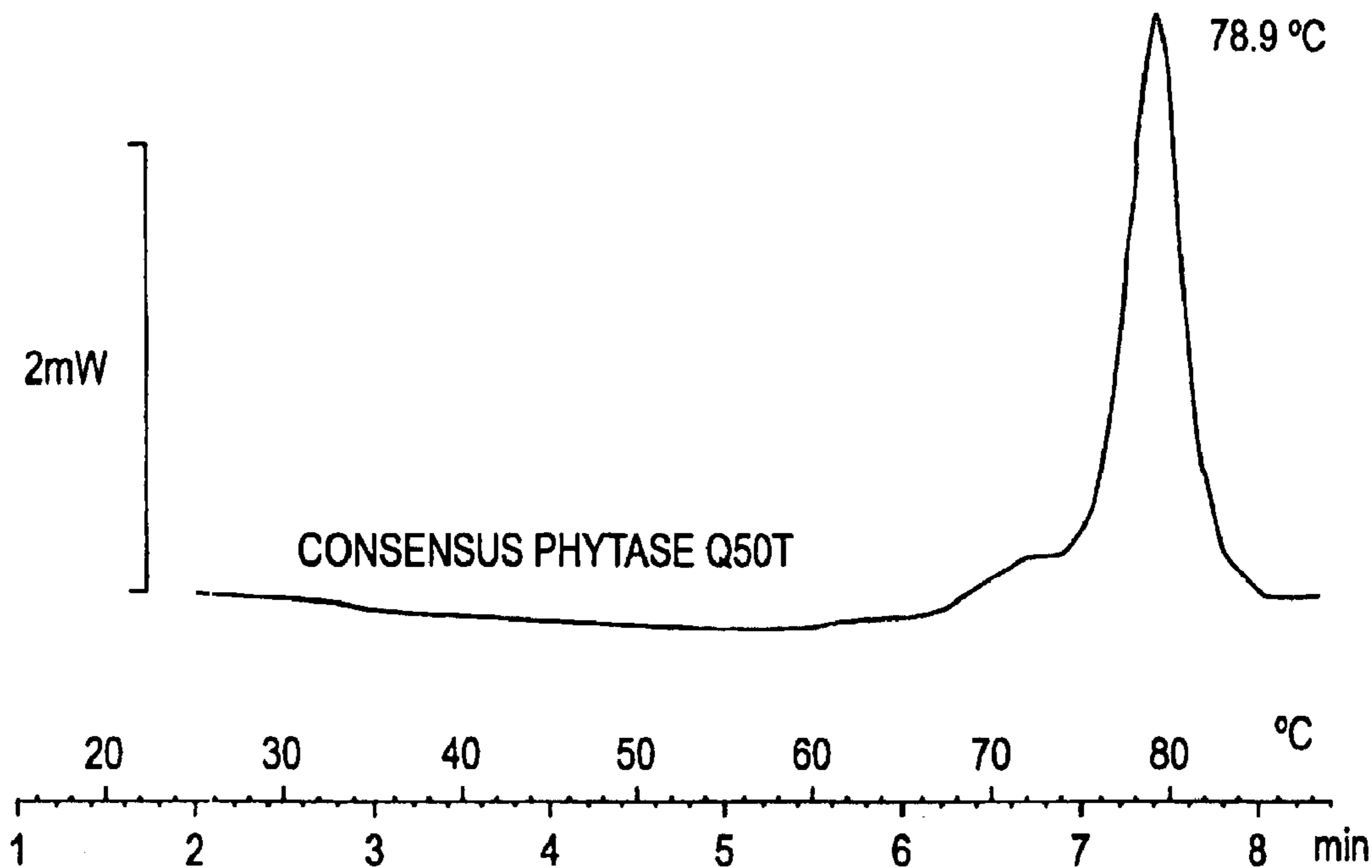


FIG. 7A

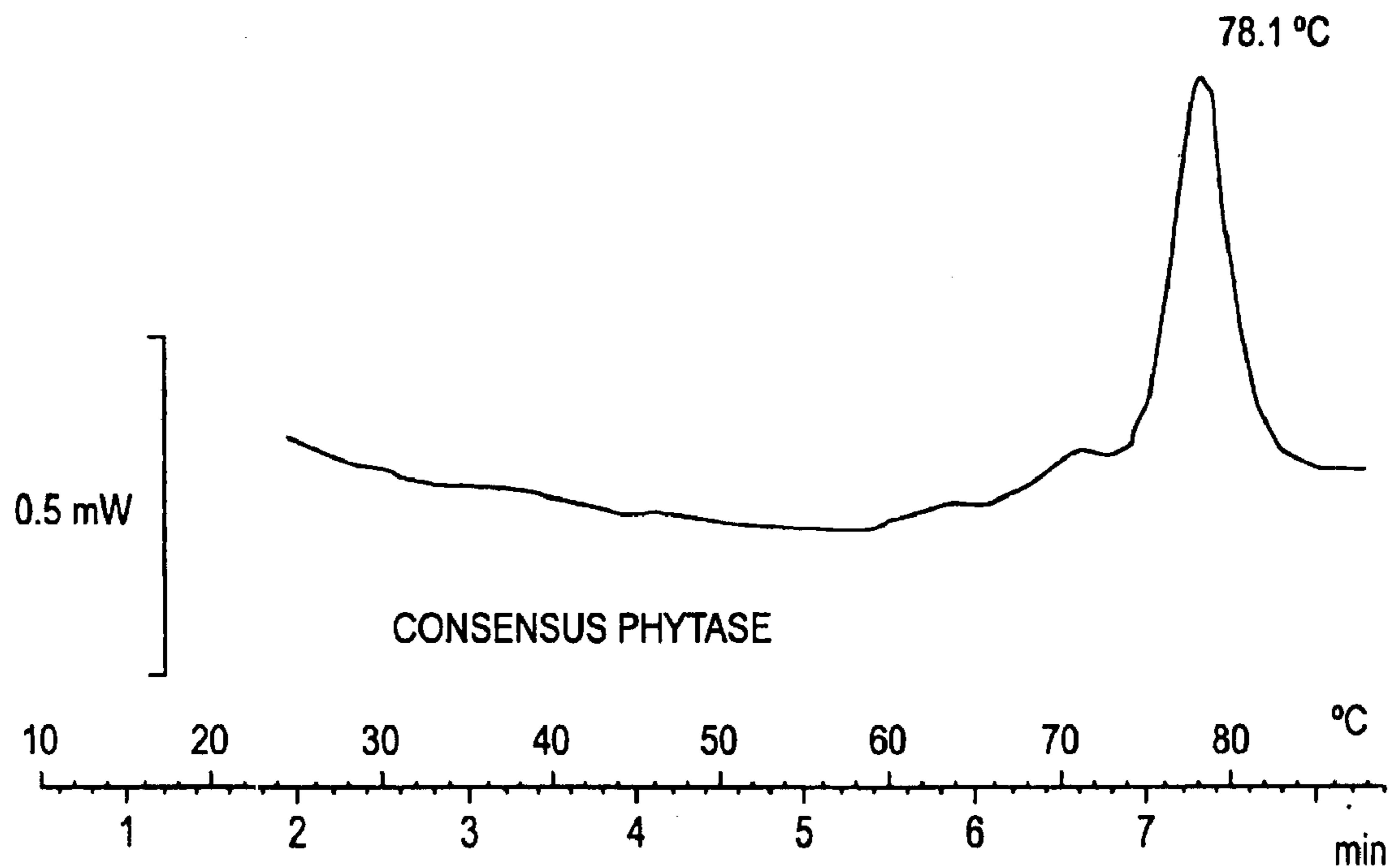


FIG. 7B

CONSENSUS PHYTASES

Matter enclosed in heavy brackets [] appears in the original patent but forms no part of this reissue specification; matter printed in italics indicates the additions made by reissue.

This is a divisional of U.S. application Ser. No. 09/121,425, filed Jul. 23, 1998, now U.S. Pat. No. 6,153,418.

BACKGROUND OF THE INVENTION

Phytases (myo-inositol hexakisphosphate phosphohydrolases; EC3.1.3.8) are enzymes that hydrolyze phytate (myo-inositol hexakisphosphate) to myo-inositol and inorganic phosphate and are known to be valuable feed additives.

A phytase was first described in rice bran in 1907 [Suzuki et al., Bull. Coll. Agr. Tokio Imp. Univ. 7, 495 (1907)] and phytases from *Aspergillus* species in 1911 [Dox and Golden, J. Biol. Chem. 10, 183-186 (1911)]. Phytases have also been found in wheat bran, plant seeds, animal intestines and in microorganisms [Howsen and Davis, Enzyme Microb. Technol. 5, 377-382 (1983), Lambrechts et al., Biotech. Lett. 14, 61-66 (1992), Shieh and Ware, Appl. Microbiol. 16, 1348-1351 (1968)].

The cloning and expression of the phytase from *Aspergillus niger* (ficcum) has been described by Van Hartingsveldt et al., in *Gene*, 127, 87-94 (1993) and in European Patent Application, Publication No. (EP) 420 358 and from *Aspergillus niger* var. *awamori* by Piddington et al., in *Gene* 133, 55-62 (1993).

Cloning, expression and purification of phytases with improved properties have been disclosed in EP 684 313. However, since there is a still ongoing need for further improved phytases, especially with respect to the thermostability, it is an object of the present invention to provide the following process which is, however, not only applicable to phytases.

SUMMARY OF THE INVENTION

The invention herein is a process for the preparation of a consensus protein, especially a phytase. The invention is also directed to a consensus phytase and to a DNA sequence encoding the consensus phytase. As is well known, a consensus protein is a new protein whose sequence is created from sequence information obtained from at least three other proteins having a similar biological activity. The object in preparing a consensus protein is to obtain a single protein which combines the advantageous properties of the original proteins.

The process is characterized by the following steps:

- a) at least three preferably four amino acid sequences of a defined protein family are aligned by any standard alignment program known in the art;
- b) amino acids at the same position according to such alignment are compared regarding their evolutionary similarity by any standard program known in the art, whereas the degree of similarity provided by such a program which defines the least similarity of the amino acids that is used for the determination of an amino acid of corresponding positions is set to a less stringent number and the parameters are set in such a way that it is possible for the program to determine from only 2 identical amino acids at a corresponding position an amino acid for the consensus protein; however, if among the compared amino acid sequences are

sequences that show a much higher degree of similarity to each other than to the residual sequences, the sequences are represented by their consensus sequence determined as defined in the same way as to the present process for the consensus sequence of the consensus protein or a vote weight of 1 divided by the number of such sequences is assigned to every of those sequences.

- c) in case no common amino acid at a defined position can be identified by the program, any of the amino acids of all sequences used for the comparison, preferably the most frequent amino acid of all such sequences is selected or an amino acid is selected on the basis of the consideration given in Example 2.
- d) once the consensus sequence has been defined, such sequence is back-translated into a DNA sequence, preferably using a codon frequency table of the organism in which expression should take place;
- e) the DNA sequence is synthesized by methods known in the art and used either integrated into a suitable expression vector or by itself to transform an appropriate host cell;
- f) the transformed host cell is grown under suitable culture conditions and the consensus protein is isolated from the host cell or its culture medium by methods known in the art.

In a preferred embodiment of this process step b) can also be defined as follows: b) amino acids at the same position according to such an alignment are compared regarding their evolutionary similarity by any standard program known in the art, whereas the degree of similarity provided by such program is set at the lowest possible value and the amino acid which is the most similar for at least half of the sequences used for the comparison is selected for the corresponding position in the amino acid sequence of the consensus protein.

Thus the claimed invention is a process for obtaining a consensus protein from a group of amino acid sequences of a defined protein family, which comprises:

- a) aligning a group consisting of three to one hundred, but preferably three or four amino acid sequences from a defined protein family;
- b) comparing the evolutionary similarity of amino acids which occupy a position in the aligned sequences to select a consensus amino acid for said position using a system which is so organized that if two amino acids which occupy said position are identical, then the identical amino acid is selected as the consensus amino acid for said position, unless three or more other amino acids at said position have a higher degree of structural similarity to each other than to the identical amino acid, in which case the amino acid which has the highest degree of evolutionary similarity to the other amino acids is selected as the consensus amino acid for said position, with the proviso that if a set of amino acid sequences exists within the group of step a) such that the amino acid sequences within the set have more evolutionary similarity to each other than to any of the amino acid sequences of the group which are not part of the set, then the amino acids which occupy said position in members of the set will have a vote weight of one divided by the number of amino acid sequences in the set where the amino acids which occupy said position in amino acid sequences which are not in the set will have a vote weight of one, and repeating the procedure for each position in the aligned group of amino acid sequences;

- c) if no consensus amino acid for said position is obtained by the method of step b), then any amino acid at said position is selected as the consensus sequence, preferably the most frequent amino acid;
- d) combining the consensus amino acids obtained in steps b) and c) obtain a consensus amino acid sequence;
- e) translating the consensus amino acid sequence into a DNA sequence, preferably using a codon frequency table specific to whichever host organism has been selected for expressing the DNA sequence;
- f) obtaining the DNA sequence and using said DNA sequence to express a protein which is the consensus protein of the defined protein family.

The present invention is also directed to new phytases, preferably phytases having the amino acid sequence depicted in FIG. 2 and variants and muteins thereof. In addition, the invention includes polynucleotides which encode such new phytases.

A BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1: Calculation of the consensus phytase sequence from the alignment of nearly all known fungal phytase amino acid sequences. The letters represent the amino acid residues in the one-letter code. The following sequences were used for the alignment: phyA from *Aspergillus terreus* 9A-1 (Mitchell et al., 1997; from amino acid (aa) 27) (*SEQ ID NO: 1*), phyA from *Aspergillus terreus* cbs116.46 (van Loon et al., 1997; from aa 27) (*SEQ ID NO: 2*), phyA from *Aspergillus niger* var. *awamori* (Piddington et al., 1993; from aa 27) (*SEQ ID NO: 3*), phyA from *Aspergillus niger* T213[.] (from aa 27) (*SEQ ID NO: 4*), phyA from *Aspergillus niger* strain NRRL3135 (van Hartingsveldt et al., 1993; from aa 27) (*SEQ ID NO: 5*), phyA from *Aspergillus fumigatus* ATCC 13073 (Pasamontes et al., 1997b; from aa 25) (*SEQ ID NO: 6*), phyA from *Aspergillus fumigatus* ATCC 32722 (van Loon et al., 1997; from aa 27) (*SEQ ID NO: 7*), phyA from *Aspergillus fumigatus* ATCC 58128 (van Loon et al., 1997; from aa 27) (*SEQ ID NO: 8*), phyA from *Aspergillus fumigatus* ATCC 26906 (van Loon et al., 1997; from aa 27) (*SEQ ID NO: 9*), phyA from *Aspergillus fumigatus* ATCC 32239 (van Loon et al., 1997; from 30) (*SEQ ID NO: 10*), phyA from *Aspergillus nidulans* (Pasamontes et al., 1997a; from aa 25) (*SEQ ID NO: 11*), phyA from *Talaromyces thermophilus* (Pasamontes et al., 1997a; from aa 24) (*SEQ ID NO: 12*), and phyA from *Myceliophthora thermophila* (Mitchell et al., 1997; from aa 19) (*SEQ ID NO: 13*). The alignment was calculated using the program PILEUP. The location of the gaps was refined by hand. Capitalized amino acid residues in the alignment at a given position belong to the amino acid coalition that establish the consensus residue. In bold, beneath the calculated consensus sequence (*SEQ ID NO: 14*), the amino acid sequence of the finally constructed fungal consensus phytase (Fcp) is shown (*SEQ ID NO: 15*). The gaps in the calculated consensus sequence were filled by hand according to principals stated in Example 2.

FIG. 2: DNA sequence of the fungal consensus phytase gene (fcp) (*SEQ ID NO: 16*) and of the primers synthesized for gene construction. The calculated amino acid sequence (FIG. 1) was converted into a DNA sequence using the program BACKTRANSLATE (Devereux et al., 1984), and the codon frequency table of highly expressed yeast genes (GCG program package, 9.0). The signal peptide of the phytase from *A. terreus* cbs was fused to the N-terminus. The bold bases represent the sequences of the oligonucleotides used to generate the gene. The names of the respective

oligonucleotides are noted above or below the sequence. The underlined bases represent the start and stop codons of the gene. The bases written in italics show the two introduced Eco RI sites. *The amino acid sequence of the encoded polypeptide (SEQ ID NO: 17) is also shown.*

FIG. 3: Temperature optimum of fungal consensus phytase and other phytases used to calculate the consensus sequence. For the determination of the temperature optimum, the phytase standard assay was performed at a series of temperatures between 37 and 85° C. The phytases used were purified according Example 5y, ▽; fungal consensus phytase; ▲, *A. fumigatus* 13073 phytase; □, *A. niger* NRRL3135 phytase; ○, *A. nidulans* phytase; ■, *A. terreus* 9A-1 phytase; ●, *A. terreus* cbs phytase.

FIG. 4: The pH-dependent activity profile of fungal consensus phytase and of the mutant Q50L, Q50T, and Q50G. The phytase activity was determined using the standard assay in appropriate buffers (see Example 9) at different pH-values. Plot: a) shows a comparison of fungal consensus phytase (●) to the mutants Q50L (▽), Q50T (▲), and Q50G (○) in percent activity. Plot b) shows a comparison of fungal consensus phytase (○) to mutant Q50L (●) and Q50T (▽) using the specific activity of the purified enzymes expressed in *H. polymorpha*.

FIG. 5: The pH-dependent activity profile of the mutants Q50L, Y51N and Q50T, Y51N in comparison to the mutants Q50T and Q50L of fungal consensus phytase. The phytase activity was determined using the standard assay in appropriate buffers (see Example 9) at different pH-values. Graph a) shows the influence of the mutation Y51N (●) on mutant Q50L (○). Graph b) shows the influence of the same mutation (●) on mutant Q50T (○).

FIG. 6: Substrate specificity of fungal consensus phytase and its mutants Q50L, Q50T, and Q50G. The bars represent the relative activity in comparison to the activity with phytic acid (100%) with a variety of known natural and synthetic phosphorylated compounds.

FIG. 7: Differential scanning calorimetry (DSC) of fungal consensus phytase and its mutant Q50T. The protein samples were concentrated to carry 50–60 mg/ml and extensively dialyzed against 10 mM sodium acetate, pH 5. A constant heating rate of 10° C./min was applied up to 90° C. DSC of consensus phytase Q50T (upper graph) yielded in a melting temperature of 78.9° C., which is nearly identical to the melting point of fungal consensus phytase (78.1° C., lower graph).

DETAILED DESCRIPTION OF THE INVENTION

A preferred embodiment of this whole process can be seen in a process in which a sequence is chosen from a number of highly homologous sequences and only those amino acid residues are replaced which clearly differ from a consensus, sequence of this protein family calculated under moderately stringent conditions, while at all positions of the alignment where the method is not able to determine an amino acid under moderately stringent conditions the amino acids of the preferred sequence are taken.

It is furthermore an object of the present invention to provide such a process, wherein the program used for the comparison of amino acids at a defined position regarding their evolutionary similarity is the program "PRETTY". It is more specifically an object of the present invention to provide such a process, wherein the defined protein family is the family of phytases, especially wherein the phytases are of fungal origin.

It is furthermore an object of the present invention to provide such processes, wherein the host cell is of eukaryotic, especially fungal, preferably *Aspergillus* or yeast, preferably *Saccharomyces* or *Hansenula* origin. It is also an object of the present invention to provide a consensus protein obtainable by such a process. A preferred consensus protein obtained by the present process is of the defined protein family of phytases. The especially preferred consensus phytase is created based on phytase sequences from:

Aspergillus terreus 9A-1, aa 27 (Mitchell et al., 1997);
Aspergillus terreus cbs116.46, aa 27 (van Loon et al., 1997);
Aspergillus niger var. *awamori*, aa 27 (Piddington et al., 1993);
Aspergillus niger T213, aa 27;
Aspergillus niger strain NRRL3135 aa 27 (van Hartingsveldt et al., 1993);
Aspergillus fumigatus ATCC 13073, aa 26 (Pasamontes et al., 1997);
Aspergillus fumigatus ATCC 32722, aa 26 (van Loon et al., 1997);
Aspergillus fumigatus ATCC 58128, aa 26 (van Loon et al., 1997);
Aspergillus fumigatus ATCC 26906, aa 26 (van Loon et al., 1997);
Aspergillus fumigatus ATCC 32239, aa 30 (van Loon et al., 1997);
Aspergillus nidulans, aa 25 (Pasamontes et al., 1997a);
Talaromyces thermophilus ATCC 20186, aa 24 (Pasamontes et al., 1997a); and
Myceliophthora thermophila, aa 19 (Mitchell et al., 1997).

Therefore the preferred group of amino acid sequences used in the process of this invention is the amino acid sequences encoding the phrases of the above fungi.

The preferred phytase of the invention is a consensus protein whose sequence is created based on the sequences of the twelve phytases shown in Table 3, below, but which is not highly homologous to any of the twelve phytases in that the consensus phytase is not more than about 80% identical to any of the twelve phytases. The present invention is particularly directed to a consensus phytase which has the amino acid sequence shown in FIG. 2 or a variant or mutein thereof. The consensus phytase of FIG. 2 is not highly homologous to any of the twelve phytases which were used to create its sequence, as can be seen from the sequence comparison results shown in Table 3. Another consensus phytase of this invention has the sequence shown in FIG. 1 as consensus phytase (bottom line in boldface type) or a variant or mutein thereof.

A "variant" of the consensus phytase with amino acid sequence shown in FIG. 1 or preferably FIG. 2 is the consensus phytase of Figure or preferably FIG. 2 in which at one or more positions amino acids have been deleted, added or replaced by one or more other amino acids with the proviso that the resulting sequence provides for a phytase whose basic properties like enzymatic activity (type of and specific activity), thermostability, activity in a certain pH-range (pH-stability) have not significantly been changed. "Significantly" means in this context that a skilled person would say that the properties of the variant may still be different but would not be unobvious over the ones of the consensus phytase with the amino acid sequence of FIG. 1 or FIG. 2 itself.

A mutein refers in the context of the present invention to replacements of the amino acid in the amino acid sequence

of the consensus protein shown in FIG. 1 or preferably FIG. 2 which lead to consensus proteins with further improved properties, e.g., activity. Such muteins can be defined and prepared on the basis of the teachings given in European Patent Application number 97810175.6, e.g., Q50L, Q50T, Q50G, Q50L-Y51N, or Q50T-Y51N. "Q50L" means in this context that a position 50 of the amino acid sequence the amino acid Q has been replaced by amino acid L. Therefore specific muteins of this invention include a mutein which has the amino acid sequence of FIG. 2 except that Q at position 50 has been replaced by L, T, or G, and two muteins which have the amino acid sequence of FIG. 1 except that Q at position 50 has been replaced by T or L and Y at position 51 has been replaced by N.

Polynucleotides which encode the consensus phytase of this invention, i.e., a phytase with the amino acid sequence of FIG. 1 or preferably FIG. 2 or variants and muteins thereof, especially the specific muteins listed above, are also part of this invention. Such polynucleotides may be obtained by known methods, for example by backtranslation of the mutein's amino acid sequence and PCR synthesis of the corresponding polynucleotide as described below.

In addition, a food, feed, premix or pharmaceutical composition comprising a consensus protein as defined above is also an object of the present invention. Food, feed, and premix compositions, preferably for domestic livestock, are well known to a skilled person, as are pharmaceutical compositions. Such pharmaceutical compositions are likely to be veterinary compositions formulated for oral ingestion, such as pills and the like.

In this context "at least three preferably four amino acid sequences of such defined protein family" means that three, four, five, six to 12, 20, 50, 100 or even more sequences can be used for the alignment and the comparison to create the amino acid sequence of the consensus protein. Amino acid sequences may be obtained from known sources such as publications or databases, or may be deduced by translation of DNA sequences which are publicly available, or may be determined by known techniques for sequencing an isolated protein or obtaining and sequencing a gene encoding a protein and translating the DNA sequence. "Sequences of a defined protein family" means that such sequences fold into a three dimensional structure, wherein the α -helixes, the β -sheets and-turns are at the same position so that such structures are, as called by the skilled person, superimposable. Furthermore these sequences characterize proteins which show the same type of biological activity, e.g., a defined enzyme class such as the phytases. As known in the art, the three dimensional structure of one of such sequences is sufficient to allow the modelling of the structure of the other sequences of such a family. An example, how this can be effected, is given in the Reference Example of the present case.

Aligning amino acid sequences is a well known process whereby two or more amino acids are lined up in such a way to maximize the intern amino acid sequences which they have in common.

"Evolutionary similarity" in the context of the present invention refers to a schema which classifies amino acids regarding their structural similarity which allows that one amino acid can be replaced by another amino acid with a minimal influence on the overall structure, as this is done e.g. by programs, like "PRETTY", known in the art. The phrase "the degree of similarity provided by such a program . . . is set to less stringent number" means in the context of the present invention that values for the parameters which determine the degree of similarity in the pro-

gram used in the practice of the present invention are chosen in a way to allow the program to define a common amino acid for a maximum of positions of the whole amino acid sequence, e.g. in case of the program PRETTY a value of 2 or 3 for the THRESHOLD and a value of 2 for the PLURALITY can be chosen.

A consensus amino acid is an amino acid chosen to occupy a given position in the consensus protein obtained by this method. A system which is organized to select consensus amino acids as described above may be a computer program, or a combination of one or more computer programs with "by hand" analysis and calculation. A set of amino acid sequences existing within the group of amino acid sequences from which the consensus sequence is prepared means a set of such sequences which are more similar to each other than to other members of the group, based on the evolutionary similarity analysis performed above. An example of such a group is a species where a set within the group would be members of a particular strain. Furthermore, "a vote weight of one divided by the number of such sequence means in the context of the present invention that the sequences which define a group of sequences with a higher degree of similarity as the other sequences used for the determination of the consensus sequence only contribute to such determination with a factor which is equal to one divided by a number of all sequences of this group. Thus an amino acid occupying a particular position in the aligned sequences will, if it is a member of a set not have a comparison value of equal weight with the other amino acids (e.g. one) but will have a lower weight depending on the size of the set which it is in, as the weight is one divided by the number of amino acid sequences in the set.

When a consensus amino acid is obtained for each position of the aligned amino acid sequences, then these consensus amino acids are "lined up" to obtain the amino acid sequence of the consensus protein.

As mentioned before should the program not allow selection of the most similar amino acid, the most frequent amino acid is selected, should the latter be impossible the skilled person will select an amino acid from all the sequences used for the comparison which is known in the art for its property to improve the thermostability in proteins as discussed, e.g., by:

Janecek, S. (1993), *Process. Biochem.* 28, 435-445 or Fersht, A. R. & Serrano, L. (1993), *Curr. Opin. Struct. Biol.* 3, 75-83.

Alber, T. (1989), *Annu. Rev. Biochem.* 58, 765-798 or Matthews, B. W. (1987), *Biochemistry* 26, 6885-6888.

Matthews, B. W. (1991), *Curr. Opin. Struct. Biol.* 1, 17-21.

The stability of an enzyme is a critical factor for many industrial applications. Therefore, a lot of attempts, more or less successful, have been made to improve the stability, preferably the thermostability, or enzymes be rational (van den Burg et al., 1998) or irrational approaches (Akanuma et al., 1998). The forces influencing the thermostability of a protein are the same those that are responsible for the proper folding of a peptide strand (hydrophobic interactions, van der Waals interactions, H-bonds, salt bridges, conformational strain (Matthews, 1993). Furthermore, as shown by Matthews et al. (1987), the free energy of the unfolded state has also an influence on the stability of a protein. Enhancing of protein stability means to increase the number and strength of favorable interactions and to decrease the number and strength of unfavorable interactions. It has been possible to introduce disulfide linkages (Sauer et al., 1986)

to replace glycine with alanine residues or to increase the proline content in order to reduce the free energy of the unfolded state (Margarit et al., 1992; Matthews, 1987a). Other groups concentrated on the importance of additional H-bonds or salt bridges for the stability of a protein (Blaber et al., 1993) or tried to fill cavities in the protein interior to increase the buried hydrophobic surface area and the van der Waals interactions (Karpusas et al., 1989). Furthermore, the stabilization of secondary structure elements, especially α -helices, for example, by improved helix capping, was also investigated (Munoz & Serrano, 1995).

However, there is no fast and promising strategy to identify amino acid replacements which will increase the stability, preferably the thermal stability of a protein. Commonly, the 3D structure of a protein is required to find locations in the molecule where an amino acid replacement possibly will stabilize the protein's folded state. Alternative ways to circumvent this problem are either to search for a homologous protein in a thermo- or hydrothermophile organism or to detect stability-increasing amino acid replacements by a random mutagenesis approach. This latter possibility succeeds in only 10^3 to 10^4 mutations and is restricted to enzymes for which fast screening procedure is available (Arase et al., 1993; Risse et al., 1992). For all these approaches, success was variable and unpredictable and, if successful, the thermostability enhancements nearly always were rather small.

Here we present an alternative way to improve the thermostability of a protein. Imanaka et al. (1986) were among the first to use the comparisons of homologous proteins to enhance the stability of a protein. They used a comparison of proteases from thermophilic with homologous ones of mesophilic organisms to enhance the stability of a mesophilic protease. Serrano et al. (1993) used the comparison of the amino acid sequences of two homologous mesophilic RNases to construct a more thermostable Rnase. They mutated individually all of the residues that differ between the two and combined the mutations that increase the stability in a multiple mutant. Pantoliano et al. (1989) and in particular, Steipe et al. (1994) suggested that the most frequent amino acid at every position of an alignment of homologous proteins contribute to the largest amount to the stability of a protein. Steipe et al. (1994) proved this for a variable domain of an immunoglobulin, whereas Pantoliano et al. (1989) looked for positions in the primary sequence of subtilisin in which the sequence of the enzyme chosen to be improved for higher stability was singularly divergent. Their approach resulted in the replacement M50F which increased the T_m of subtilisin by 1.8° C.

Steipe et al. (1994) proved on a variable domain of immunoglobulin that it is possible to predict a stabilizing mutation with better than 60% success rate just by using a statistical method which determines the most frequent amino acid residue at a certain position of this domain. It was also suggested that this method would provide useful results not only for stabilization of variable domains of antibodies but also for domains of other proteins. However, it was never mentioned that this method could be extended to the entire protein. Furthermore, nothing is said about the program which was used to calculate the frequency of amino acid residues at a distinct position or whether scoring matrices were used as in the present case.

It is therefore an object of the present invention to provide a process for the preparation of a consensus protein comprising a process to calculate an amino acid residue for nearly all positions of a so-called consensus protein and to synthesize a complete gene from this sequence that could be expressed in a pro- or eukaryotic expression system.

DNA sequences from which amino acid sequences may be derived for making consensus proteins of the present invention, can be constructed starting from genomic or cDNA sequences coding for proteins, e.g. phytases known in the state of the art [for sequence information see references mentioned above, e.g. EP 684 313 or sequence data bases, for example like Genbank (Intelligenetics, California, USA), European Bioinformatics Institute (Hinston Hall, Cambridge, GB), NBRF (Georgetown University, Medical Centre, Washington D.C., USA) and Vecbase (University of Wisconsin, Biotechnology Centre, Madison, Wis., USA) or disclosed in the figures by methods of in vitro mutagenesis [see e.g. Sambrook et al., Molecular Cloning, Cold Spring Harbor Laboratory Press, New York]. A widely used strategy for such "site directed mutagenesis", as originally outlined by Hurchinson and Edgell [J. Virol. 8, 181 (1971)], involves the annealing of a synthetic oligonucleotide carrying the desired nucleotide substitution to a target region of a single-stranded DNA sequence wherein the mutation should be introduced [for review see Smith, Annu. Rev. Genet. 19, 423 (1985) and for improved methods see references 2-6 in Stanssen et al. Nucl. Acid Res., 17, 4441-4454 (1989)].

Another possibility of mutating a given DNA sequence which is also preferred for the practice of the present invention is the mutagenesis using the polymerase chain reaction (PCR). DNA as starting material can be isolated by methods known in the art and described e.g. in Sambrook et al. (Molecular Cloning) from the respective strains. For strain information see, e.g., EP 684 313 or any depository authority indicated below. *Aspergillus niger* [ATCC 9142], *Myceliophthora thermophila* [ATCC 48102], *Talaromyces thermophilus* [ATCC 20186], and *Aspergillus fumigatus* [ATCC 34625] have been redeposited, according to the conditions of the Budapest Treaty at the American Type Culture Cell Collection under the following accession numbers: ATCC 74337, ATCC 74340, ATCC 74338 and ATCC 74339, respectively. Amino acid sequences may be obtained by known methods from these DNA sequences for use in the process of this invention to obtain a consensus protein. It is however, understood that DNA encoding a consensus protein in accordance with the present invention can also be prepared in a synthetic manner as described, e.g. in EP 747 483 or the examples by methods known in the art.

Once complete DNA sequences of the present invention have been obtained (for example by synthesis based on backtranslation of a consensus protein obtained in accordance with the invention) they can be integrated into vectors by methods known in the art and described e.g. in Sambrook et al. (s.a.) to overexpress the encoded polypeptide in appropriate host systems. However, a skilled person knows that also the DNA sequences themselves can be used to transform the suitable host systems of the invention to get overexpression of the encoded polypeptide. Appropriate host systems are for example fungi, like *Aspergilli*, e.g. *Aspergillus niger* [ATCC 9142] or *Aspergillus ficuum* [NRRL 3135] or like *Trichoderma*, e.g. *Trichoderma reesei* or yeasts, like *Saccharomyces*, e.g. *Saccharomyces cerevisiae* or *Pichia*, like *Pichia pastoris*, or *Hansenula polymorpha*, e.g. *H. polymorpha* (DSM5215) plants, as described, e.g. by Pen et al., Bio/Technology 11, 811-814 (1994), skilled person knows that such microorganisms are available from depository authorities, e.g. the American Type Culture Collection (ATCC), the Centraalbureau voor Schimmelcultures (CBS) or the Deutsche Sammlung für Mikroorganismen and Zellkulturen GmbH (DSM) or any other depository authority as listed in the Journal "Industrial Property" [(1991) 11 pages 29-40]. Bacteria which can be

used are e.g. *E. coli*, Bacilli as, e.g. *Bacillus subtilis* or *Streptomyces*, e.g. *Streptomyces lividans* (see e.g. Anné and Mallaert in FEMS Microbiol. Letters 114, 121 (1993)). *E. coli*, which could be used are *E. coli* K12 strains e.g. M15 [described as DZ 291 by Villarejo et al. in J. Bacteriol. 120, 466-474 (1974)], HB 101 [ATCC No. 33694] or *E. coli* SG13009 [Gottesman et al., J. Bacteriol. 148, 265-273 (1981)].

Vectors which can be used for expression in fungi are known in the art and described e.g. in EP 420 358, or by Cullen et al. [Bio/Technology 5, 369-376 (1987)] or Ward in Molecular Industrial Mycology, Systems and Applications for Filamentous Fungi, Marcel Dekker, New York (1991), Upshall et al. [Bio/Technology 5, 1301-1304 (1987)], Gwynne et al. [Bio/Technology 5, 71-79 (1987)], Punt et al. [J. Biotechnol. 17, 19-34 (1991)] and for yeast by Sreekrishna et al. [J. Basic Microbiol. 28, 265-278 (1988), Biochemistry 28, 4117-4125 (1989)], Hitzemann et al. [Nature 293, 717-722 (1981)] or in EP 183 070, EP 183 071, EP 248 227, EP 263 311. Suitable vectors which can be used for expression in *E. coli* are mentioned, e.g. by Sambrook et al. [s.a.] or by Fiers et al. in Proc. 8th Int. Biotechnology Symposium" [Soc. Franc. de Microbiol., Paris (Durand et al., eds.), pp. 680-697 (1988)] or by Bujard et al. in Methods in Enzymology, eds. Wu and Grossmann, Academic Press, Inc. Vol. 155, 416-433 (1987) and Stüber et al. in Immunological Methods, eds. Lefkovits and Pernis, Academic Press, Inc., Vol. IV, 121-152 (1990). Vectors which could be used for expression in Bacilli are known in the art and described, e.g. in EP 405 370, Proc. Natl. Acad. Sci. USA 81, 439 (1984) by Yansura and Henner, Meth. Enzymol. 185, 199-228 (1990) or EP 207 459. Vectors which can be used for the expression in *H. polymorpha* are known in the art and described, e.g. in Gellissen et al., Biotechnology 9, 291-295 (1991).

Either such vectors already carry regulatory elements, e.g., promoters, or the DNA sequences of the present invention can be engineered to contain such elements. Suitable promoter elements which can be used are known in the art and are, e.g. for *Trichoderma reesei* the *cbh1*-[Haarki et al., Biotechnology 7, 596-600 (1989)] or the *pki1*-promoter [Schindler et al., Gene 130, 271-275 (1993)], for *Aspergillus oryzae* the *amy*-promoter [Christensen et al., Abstr. 19th Lunteren Lectures on Molecular Genetics F23 (1987), Christensen et al., Biotechnology 6, 1419-1422 (1988), Tada et al., Mol. Gen. Genet. 229, 301 (1991)], for *Aspergillus niger* the *glaA*-[Cullen et al., Bio/Technology 5, 369-376 (1987), Gwynne et al., Bio/Technology 5, 713-719 (1987), Ward in Molecular Industrial Mycology, Systems and Applications for Filamentous Fungi, Marcel Dekker, New York, 83-106 (1991)], *alcA*-[Gwynne et al., Bio/Technology 5, 718-719 (1987)], *suc1*-[Boddy et al., Curr. Genet. 24, 60-66 (1993)], *aphA*-[MacRae et al., Gene 71, 339-348 (1988), MacRae et al., Gene 132, 193-198 (1993)], *tpiA*-[McKnight et al., Cell 46, 143-147 (1986), Upshall et al., Bio/Technology 5, 1301-1304 (1987)], *gpdA*-[Punt et al., Gene 69, 49-57 (1988), Punt et al., J. Biotechnol. 17, 19-37 (1991)] and the *pkiA*-promoter [de Graaff et al., Curr. Genet. 22, 21-27 (1992)]. Suitable promoter elements which could be used for expression in yeast are known in the art and are, e.g. the *pho5*-promoter [Vogel et al., Mol. Cell. Biol., 2050-2057 (1989); Rudolf and Hinnen, Proc. Natl. Acad. Sci. 84, 1340-1344 (1987)] or the *gap*-promoter for expression in *Saccharomyces cerevisiae* and for *Pichia pastoris*, e.g. the *aox1*-promoter [Koutz et al., Yeast 5, 167-177 (1989); Sreekrishna et al., J. Basic Microbiol. 28, 265-278 (1988)], or the FMD promoter [Hollenberg et al.,

EPA No. 0299108] or MOX-promoter [Ledebøer et al., Nucleic Acids Res. 13, 3063–3082 (1985)] for *H. polymorpha*.

Accordingly, vectors comprising DNA sequences of the present invention, preferably for the expression of said DNA sequences in bacteria or a fungal or a yeast host and such transformed bacteria or fungal or yeast hosts are also an object of the present invention.

It is also an object of the present invention to provide a system which allows for high expression of proteins, preferably phytases like the consensus phytase of the present invention in *Hansenula* characterized therein that the codons of the encoding DNA sequence of such a protein have been selected on the basis of a codon frequency table of the organism used for expression, e.g. yeast as in the present case (see e.g. in Example 3) and optionally the codons for the signal sequence have been selected in a manner as described for the specific case in Example 3. That means that a codon frequency table is prepared on the basis of the codons used in the DNA sequences which encode the amino acid sequences of the defined protein family. Then the codons for the design of the DNA sequence of the signal sequence are selected from a codon frequency table of the host cell used for expression whereby always codons of comparable frequency in both tables are used.

Once such DNA sequences have been expressed in an appropriate host cell in a suitable medium, the encoded protein can be isolated either from the medium in the case the protein is secreted into the medium or from the host organism in case such protein is present intracellularly by methods known in the art of protein purification or described in case of a phytase, e.g. in EP 420 358. Accordingly a process for the preparation of a consensus protein (i.e. a polypeptide) of the present invention characterized in that transformed bacteria or a host cell as described above is cultured under suitable culture conditions and the consensus protein is recovered therefrom and a consensus protein produced by such a process or a consensus protein encoded by a DNA sequence of the present invention are also an object of the present invention.

Once obtained, the consensus proteins (i.e. polypeptides), preferably phytases, of the present invention can be characterized regarding their properties which make them useful in agriculture. Any assay known in the art may be used such as those described, e.g., by Simons et al. [Br. J. Nutr. 64, 525–540 (1990)], Schöner et al. [J. Anim. Physiol. a. Anim. Nutr. 66, 248–255 (1991)], Vogt [Arch. Geflügelk. 56, 93–98 (1992)], Jongbloed et al. [J. Anim. Sci., 70, 1159–1168 (1992)], Perney et al. [Poultry Sci. 72, 2106–2114 (1993)], Farrell et al., [J. Anim. Physiol. a. Anim. Nutr. 69, 278–283 (1993)], Broz et al., [Br. Poultry Sci. 35, 273–280 (1994)] and Düngelhof et al. [Animal Feed Sci. Technol. 49, 1–10 (1994)].

In general the consensus phytases of the present invention can be used without being limited to a specific field of application, e.g., in case of phytases for the conversion of inositol polyphosphates, like phytate to inositol and inorganic phosphate.

Furthermore the consensus phytases of the present invention can be used in a process for the preparation of a pharmaceutical composition or compound food or feeds wherein the components of such a composition are mixed with one or more consensus phytases of the present invention. Accordingly compound food or feeds or pharmaceutical compositions comprising one or more consensus phytases of the present invention are also an object of the present invention. A skilled person is familiar with their

process of preparation. Such pharmaceutical compositions or compound food or feeds can further comprise additives or components generally used for such purpose and known in the state of the art.

It is furthermore an object of the present invention to provide a process for the reduction of levels of phytate in animal manure characterized in that an animal is fed such a feed composition in an amount effective in converting phytate contained in the feedstuff to inositol and inorganic phosphate.

The Examples which follow further elucidate the invention but are not intended to limit it in any way.

EXAMPLES

Reference Example

Homology Modeling of *A. fumigatus* and *A. terreus* cbs116.46 Phytase

The amino acid sequences of *A. fumigatus* and *A. terreus* cbs116.46 phytase were compared with the sequence of *A. niger* NRRL 3135 phytase (see FIG. 1) for which the three-dimensional structure had been determined by X-ray crystallography.

A multiple amino acid sequence alignment of *A. niger* NRRL 3135 phytase, *A. fumigatus* phytase and *A. terreus* cbs116.46 phytase was calculated with the program “PILEUP” (Prog. Menu for the Wisconsin Package, version 8, September 1994, Genetics Computer Group, 575 Science Drive, Madison Wis., USA 53711). The three-dimensional models of *A. fumigatus* phytase and *A. terreus* cbs116.46 phytase were built by using the structure of *A. niger* NRRL 3135 phytase as template and exchanging the amino acids of *A. niger* NRRL 3135 phytase according to the sequence alignment to amino acids of *A. fumigatus* and *A. terreus* cbs116.46 phytases, respectively. Model construction and energy optimization were performed by using the program Moloc (Gerber and Müller, 1995). C-alpha positions were kept fixed except for new insertions/deletions and in loop regions distant from the active site.

Only small differences of the modelled structures to the original crystal structure could be observed in external loops. Furthermore the different substrate molecules that mainly occur on the degradation pathway of phytic acid (myo-inositol-hexakisphosphate) by *Pseudomonas* sp. bacterium phytase and, as far as determined, by *A. niger* NRRL 3135 phytase (Cosgrove, 1980) were constructed and forged into the active site cavity of each phytase structure. Each of these substrates was oriented in a hypothetical binding mode proposed for histidine acid phosphatases (Van Etten, 1982). The scissile phosphate group was oriented towards the catalytically essential His 59 to form the covalent phosphoenzyme intermediate. The oxygen of the substrate phosphoester bond which will be protonated by Asp 339 after cleavage was orientated towards the proton donor. Conformational relaxation of the remaining structural part of the substrates as well as the surrounding active site residues was performed by energy optimization with the program Moloc.

Based on the structure models the residues pointing into the active site cavity were identified. More than half (60%) of these positions were identical between these three phytases, whereas only few positions were not conserved (see FIG. 1). This observation could be extended to four additional phytase sequences (*A. nidulans*, *A. terreus* 9A1, *Talaromyces thermophilus*, *Myceliophthora thermophila*).

Example 1

Alignment of the Amino Acid Sequence of the Fungal Phytases

The alignment was calculated using the program PILEUP from the Sequence Analysis Package Release 9.0 (Devereux

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et al., 1984) with the standard parameter (gap creation penalty 12, gap extension penalty 4). The location of the gaps was refined using a text editor. Amino acid sequences encoded by the following genes (see FIG. 1) without the signal sequence were used for the performance of the alignment starting with the amino acid (aa) mentioned below:

phyA gene from *Aspergillus terreus* 9A-1, aa 27 (Mitchell et al., 1997)

phyA gene from *Aspergillus terreus* cbs116.46, aa 27 (van Loon et al., 1997)

phyA gene from *Aspergillus niger* var. *awamori*, aa 27 (Piddington et al., 1993)

phyA gene from *Aspergillus niger* T213, aa 27

phyA gene from *Aspergillus niger* strain NRRL3135, aa 27 (van Hartingsveldt et al., 1993)

phyA gene from *Aspergillus fumigatus* ATCC 13073, aa 26 (Pasamontes et al., 1997)

phyA gene from *Aspergillus fumigatus* ATCC 32722, aa 26 (van Loon et al., 1997)

phyA gene from *Aspergillus fumigatus* ATCC 58128, aa 26 (van Loon et al., 1997)

phyA gene from *Aspergillus fumigatus* ATCC 26906, aa 26 (van Loon et al., 1997)

phyA gene from *Aspergillus fumigatus* ATCC 32239, aa 30 (van Loon et al., 1997)

phyA gene from *Aspergillus nidulans*, aa 25 (Pasamontes et al., 1997a)

phyA gene from *Talaromyces thermophilus* ATCC 20186, aa 24 (Pasamontes et al., 1997a)

phyA gene from *Myceliophthora thermophila*, aa 19 (Mitchell et al., 1997)

Table 2 shows the homology of the phytase sequences mentioned above.

TABLE 2

	% identity						
	<i>A. terreus</i> 9A-1	<i>A. terreus</i> cbs 116.46	<i>A. niger</i> NRRL 3135	<i>A. fumigatus</i> 13073	<i>A. nidulans</i>	<i>T. thermophilus</i>	<i>M. thermophila</i>
<i>A. terreus</i> 9A-1		89.1	62.0	60.6	59.3	58.3	48.6
<i>A. terreus</i> cbs	90.7		63.6	62.0	61.2	59.7	49.1
<i>A. niger</i> NRRL 3135	67.3	68.9		66.8	64.2	62.5	49.4
<i>A. fumigatus</i> 13073	66.1	67.2	71.1		68.0	62.6	53.0
<i>A. nidulans</i>	65.0	66.7	69.0	73.3		60.5	52.5
<i>T. thermophilus</i>	63.8	64.5	68.9	68.1	67.4		49.8
<i>M. thermophila</i>	53.7	54.6	57.6	61.0	59.9	57.8	
	% similarity						

Table 2: Homology of the fungal phytases. The amino acid sequences of the phytases used in the alignment were compared by the program GAP (GCG program package, 9; Devereux et al., 1984) using the standard parameters. The comparison was restricted to the part of the sequence that was also used for the alignment (see legend to FIG. 1) lacking the signal peptide which was rather divergent. The numbers above and beneath the diagonal represent the amino acid identities and similarities, respectively.

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Example 2

Calculation of the Amino Acid Sequence of Fungal Consensus Phytases

Using the refined alignment of Example 1 as input, the consensus sequence was calculated by the program PRETTY from the Sequence Analysis Package Release 9.0 (Devereux et al., 1984). PRETTY prints sequences with their columns aligned and can display a consensus sequence for the alignment. A vote weight that pays regard to the similarity between the amino acid sequences of the phytases aligned were assigned to all sequences. The vote weight was set such as the combined impact of all phytases from one sequence subgroup (same species of origin but different strains), e.g. the amino acid sequences of all phytases from *A. fumigatus*, on the election was set one, that means that each sequence contributes with a value of 1 divided by the number of strain sequences (see Table 1). By this means, it was possible to prevent that very similar amino acid sequences, e.g. of the phytases from different *A. fumigatus* strains, dominate the calculated consensus sequence.

TABLE 1

<i>Aspergillus terreus</i> 9A-1 phytase:	0.50
<i>Aspergillus terreus</i> cba116.46 phytase:	0.50
<i>Aspergillus niger</i> var. <i>awamori</i> phytase:	0.3333
<i>Aspergillus niger</i> T213 phytase:	0.3333
<i>Aspergillus niger</i> NRRL3135 phytase:	0.3333
<i>Aspergillus fumigatus</i> ATCC 13073 phytase:	0.20
<i>Aspergillus fumigatus</i> ATCC 32722 phytase:	0.20
<i>Aspergillus fumigatus</i> ATCC 58128 phytase:	0.20
<i>Aspergillus fumigatus</i> ATCC 26906 phytase:	0.20
<i>Aspergillus fumigatus</i> ATCC 32239 phytase:	0.20
<i>Aspergillus nidulans</i> phytase:	1.00
<i>Talaromyces thermophilus</i> ATCC 20186 phytase:	1.00
<i>Myceliophthora thermophila</i> phytase:	1.00

Table 1: Vote weights of the amino acid sequences of the fungal phytases used. The table shows the vote weights used to calculate the consensus sequence of the fungal phytases.

The program PRETTY was started with the following parameters: The plurality defining the number of votes below which there is no consensus was set on 2.0. The threshold, which determines the scoring matrix value below which an amino acid residue may not vote for a coalition of residues, was set on 2. PRETTY used the PrettyPep.Cmp consensus scoring matrix for peptides.

Ten positions of the alignment (position 46, 66, 82, 138, 162, 236, 276, 279, 280, 308; FIG. 1), for which the program was not able to determine a consensus residue, were filled by

hand according to the following rules: if a most frequent residue existed, this residue was chosen (138, 236, 280); if a prevalent group of chemically similar or equivalent residues occurred, the most frequent or, if not available, one residue of this group was selected (46, 66, 82, 162, 276, 308). If there was either a prevalent residue nor a prevalent group, one of the occurring residues was chosen according to common assumption on their influence on the protein stability (279). Eight other positions (132, 170, 204, 211, 275, 317, 384, 447; FIG. 1) were not filled with the amino acid residue selected by the program but normally with amino acids that occur with the same frequency as the residues that were chosen by the program. In most cases, the slight underrating of the three *A. niger* sequences (sum of the vote weights: 0.99) was eliminated by this corrections.

Table 3 shows the homology of the calculated fungal consensus phytase amino acid sequence to the phytase sequences used for the calculation.

TABLE 3

Phytase	Identity [%]	Similarity [%]
<i>A. niger</i> T213	76.6	79.6
<i>A. niger</i> var. <i>awamori</i>	76.6	79.6
<i>A. niger</i> NRRL3135	76.6	79.4
<i>A. nidulans</i>	77.4	81.5
<i>A. terreus</i> 9A-1	70.7	74.8
<i>A. terreus</i> cbs116.46	72.1	75.9
<i>A. fumigatus</i> 13073	80.0	83.9
<i>A. fumigatus</i> 32239	78.2	82.3
<i>T. thermophilus</i>	72.7	76.8
<i>M. thermophila</i>	58.3	64.5

Table 3: Homology of the amino acid sequence of fungal consensus phytase to the phytases used for its calculation. The amino acid sequences of all phytases were compared with the fungal consensus phytase sequence using the program GAP (GCG program package, 9.0). Again, the comparison was restricted to that part of the sequence that was used in the alignment.

Example 3

Conversion of the Fungal Consensus Phytase Amino acid Sequence to a DNA Sequence

The first 26 amino acid residues of *A. terreus* cbs116.46 phytase were used as signal peptide and, therefore, fused to the N-terminus of all consensus phytases. For this stretch, we used a special method to calculate the corresponding DNA sequence. Purvis et al. (1987) proposed that the incorporation of rare codons in a gene has an influence on the folding efficiency of the protein. Therefore, at least the distribution of rare codons in the signal sequence of *A. terreus* cbs116.46, which was used for the fungal consensus phytase and which is very important for secretion of the protein, but converted into the *S. cerevisiae* codon usage, was transfected into the new signal sequence generated for expression in *S. cerevisiae*. For the remaining parts of the protein, we used the codon frequency table of highly expressed *S. cerevisiae* genes, obtained from the GCG program package, to translate the calculated amino acid sequence into a DNA sequence. The resulting sequence of the fcp gene are shown in FIG. 2.

Example 4

Construction and Cloning of the Fungal Consensus Phytase Genes

The calculated DNA sequence of fungal consensus phytase was divided into oligonucleotides of 85 bp, alter-

nately using the sequence of the sense and the anti-sense strand. Every oligonucleotide overlaps 20 bp with its previous and its following oligonucleotide of the opposite strand. The location of all primers, purchased by Microsynth, Balgach (Switzerland) and obtained in a PAGE-purified form, is indicated in FIG. 2. In three PCR reactions, the synthesized oligonucleotides were composed to the entire gene. For the PCR, the High Fidelity Kit from Boehringer Mannheim (Boehringer Mannheim, Mannheim, Germany) and the thermo cycler The Protokol™ from AMS Biotechnology (Europe) Ltd. (Lugano, Switzerland) were used.

Oligonucleotide CP-1 to CP-10 (Mix 1, FIG. 2) were mixed to a concentration of 0.2 pMol/μl per each oligonucleotide. A second oligonucleotide mixture (Mix 2) was prepared with CP-9 to CP-22 (0.2 pMol/μl per each oligonucleotide). Additionally, four short primers were used in the PCR reactions:

		(SEQ ID NO:18)
	Eco RI	
CP-a:	5'-TAT ATG AAT TCA TGG GCG TGT TCG TC-3'	
		(SEQ ID NO:19)
CP-b:	5'-TGA AAA GTT CAT TGA AGG TTT C-3'	
		(SEQ ID NO:20)
CP-c:	5'-TCT TCG AAA GCA GTA CAA GTA C-3'	
		(SEQ ID NO:21)
	Eco RI	
CP-e:	5'-TAT ATG AAT TCT TAA GCG AAA C-3'	

PCR reaction a: 10 μl Mix 1 (2.0 pmol of each oligonucleotide)

2 μl nucleotides (10 mM each nucleotide)

2 μl primer CP-a (10 pmol/μl)

2 μl primer CP-c (10 pmol/μl)

10[.] .0 μl PCR buffer

0.75 μl polymerase mixture

73.25 μl H₂O

PCR reaction b: 10 μl Mix 2 (2.0 pmol of each oligonucleotide)

2 μl nucleotides (10 mM each nucleotide)

2 μl primer CP-b (10 pmol/μl)

2 μl primer CP-e (10 pmol/μl)

10[.] .0 μl PCR buffer

0.75 μl polymerase mixture (2.6 U)

73.25 μl H₂O

Reaction conditions for PCR reaction a and b:

step 1 2 min—45° C.

step 2 30 sec—72° C.

step 3 30 sec—94° C.

step 4 30 sec—52° C.

step 5 1 min—72° C.

Step 3 to 5 were repeated 40-times.

The PCR products (670 and 905 bp) were purified by an agarose gel electrophoresis (Q.9% agarose) and a following gel extraction (QIAEX II Gel Extraction Kit, Qiagen, Hilden, Germany). The purified DNA fragments were used for the PCR reaction c.

PCR reaction c: 6 μl PCR product of reaction a (≈50 ng)

6 μl PCR product of reaction b (≈50 ng)

2 μl primer CP-a (10 pmol/μl)

2 μl primer CP-e (10 pmol/μl)

10,0 μ l PCR buffer
 0.75 μ l polymerase mixture (2.6 U)
 73.25 μ l H₂O

Reaction conditions for PCR reaction c:

step 1 2 min—94° C.
 step 2 30 sec—94° C.
 step 3 30 sec—55° C.
 step 4 1 min—72° C.

Step 2 to 4 were repeated 31 times.

The resulting PCR product (1.4 kb) was purified as mentioned above, digested with Eco RI, and ligated in an Eco RI-digested and dephosphorylated pBsk(-)-vector (Stratagene, La Jolla, Calif., USA). 1 μ l of the ligation mixture was used to transform *E. coli* XL-1 competent cells (Stratagene, La Jolla, Calif., USA). All standard procedures were carried out as described by Sambrook et al. (1987). The constructed fungal consensus phytase gene (*fc*p) was verified by sequencing (plasmid pBsk⁻-*fc*p).

Example 5

Expression of the Fungal Consensus Phytase Gene *fc*p and its Variants in *Saccharomyces cerevisiae* and Their Purification from Culture Supernatant

A fungal consensus phytase gene was isolated from the plasmid pBsk⁻-*fc*p ligated into the Eco RI sites of the expression cassette of the *Saccharomyces cerevisiae* expression vector pYES2 (Invitrogen, San Diego, Calif., USA) or subcloned between the shortened GAPFL (glyceraldehyde-3-phosphate dehydrogenase) promoter and the *pho5* terminator as described by Janes et al. (1990). The correct orientation of the gene was checked by PCR. Transformation of *S. cerevisiae* strains, e.g. INVSc1 (Invitrogen, San Diego, Calif., USA) was done according to Hinnen et al. (1978). Single colonies harboring the phytase gene under the control of the GAPFL promoter were picked and cultivated in 5 ml selection medium (SD-uracil, Sherman et al., 1986) at 30° C. under vigorous shaking (250 rpm) for one day. The preculture was then added to 500 ml YPD medium (Sherman et al., 1986) and grown under the same conditions. Induction of the *gall* promoter was done according to manufacturer's instruction. After four days of incubation cell broth was centrifuged (7000 rpm, GS3 rotor, 15 min, 5° C.) to remove the cells and the supernatant was concentrated by way of ultrafiltration in Amicon 8400 cells (PM30 membranes) and ultrafree-15 centrifugal filter device (Biomax-30K, Millipore, Bedford, Mass., USA). The concentrate (10 ml) was desalted on a 40 ml Sephadex G25 Superfine column (Pharmacia Biotech, Freiburg, Germany), with 10 mM sodium acetate, pH 5.0, serving as elution buffer. The desalted sample was brought to 2 M (NH₄)₂SO₄ and directly loaded onto a 1 ml Butyl Sepharose 4 Fast Flow hydrophobic interaction chromatography column (Pharmacia Biotech, Freiburg, Germany) which was eluted with a linear gradient from 2 M to 0 M (NH₄)₂SO₄ in 10 mM sodium acetate, pH 5.0. Phytase was eluted in the break-through concentrated and loaded on a 120 ml Sephacryl S-300 gel permeation chromatography column. (Pharmacia Biotech, Freiburg, Germany). Fungal consensus phytase and fungal consensus phytase 7 eluted as a homogeneous symmetrical peak and was shown by SDS-PAGE to be approx. 95% pure.

Example 6

Expression of the Fungal Consensus Phytase Genes *fc*p and its Variants in *Hansenula polymorpha*

The phytase expression vectors, used to transform *H. polymorpha*, was constructed by inserting the Eco RI frag-

ment of pBsk⁻-*fc*p encoding the consensus phytase or a variant into the multiple cloning site of the *H. polymorpha* expression vector pFPMT121, which is based on an *ura3* selection marker and the FMD promoter. The 5' end of the *fc*p gene is fused to the FMD promoter, the 3' end to the MOX terminator (Gellissen et al., 1996; EP 0299 108 B). The resulting expression vector are designated pFPMT*fc*p and pBsk⁻-*fc*p7.

The constructed plasmids were propagated in *E. coli*. Plasmid DNA was purified using standard state of the art procedures. The expression plasmids were transformed into the *H. polymorpha* strain RP11 deficient in orotidine-5'-phosphate decarboxylase (*ura3*) using the procedure for preparation of competent cells and for transformation of yeast as described in Gellissen et al. (1996). Each transformation mixture was plated on YNB (0.14% w/v Difco YNB and 0.5% ammonium sulfate) containing 2% glucose and 1.8% agar and incubated at 37° C. After 4 to 5 days individual transformant colonies were picked and grown in the liquid medium described above for 2 days at 37° C. Subsequently, an aliquot of this culture was used to inoculate fresh vials with YNB-medium containing 2% glucose. After seven further passages in selective medium, the expression vector integrates into the yeast genome in multimeric form. Subsequently, mitotically stable transformants were obtained by two additional cultivation steps in 3 ml non-selective liquid medium (YPD, 2% glucose, 10 g yeast extract, and 20 g peptone). In order to obtain genetically homogeneous recombinant strains an aliquot from the last stabilization culture was plated on a selective plate. Single colonies were isolated for analysis of phytase expression in YNB containing 2% glycerol instead of glucose to derepress the *find* promoter. Purification of the fungal consensus phytases was done as described in Example 5.

Example 7

Expression of the Fungal Consensus Genes *fc*p and its Variants in *Aspergillus niger*

Plasmid pBsk⁻-*fc*p or the corresponding plasmid of a variant of the *fc*p gene were used as template for the introduction of a Bsp HI-site upstream of the start codon of the genes and an Eco RV-site downstream of the stop codon. The Expand™ High Fidelity PCR Kit (Boehringer Mannheim, Mannheim, Germany) was used with the following primers:

Primer Asp-1 (SEQ ID NO:22):

Bsp HI
 5'-TAT ATC ATG AGC GTG TTC GTC GTG CTA CTG TTC-3'

Primer Asp-2 for cloning of *fc*p and *fc*p7

(SEQ ID NO:23):
 3'-ACC CGA CTT ACA AAG CGA ATT CTA TAG ATA TAT-5'
 Eco RV

The reaction was performed as described by the supplier. The PCR-amplified *fc*p gene had a new Bsp HI site at the start codon, introduced by primer Asp-1, which resulted in a replacement of the second amino acid residue glycine by serine. Subsequently, the DNA-fragment was digested with Bsp HI and Eco RV and ligated into the Nco I site downstream of the glucoamylase promoter of *Aspergillus niger* (*glaA*) and the Eco RV site upstream of the *Aspergillus nidulans* tryptophan C terminator (*trpC*) (Mullaney et al., 1985). After this cloning step, the genes were sequenced to detect possible failures introduced by PCR. The resulting expression plasmids which basically corresponds to the

pGLAC vector as described in Example 9 of EP 684 313, contained the orotidine-5'-phosphate decarboxylase gene (pyr4) of *Neurospora crassa* as a selection marker. Transformation of *Aspergillus niger* and expression of the consensus phytase genes was done as described in EP 684 313. The fungal consensus phytases were purified as described in Example 5.

Example 8

Construction of Muteins of Fungal Consensus Phytase

To construct muteins for expression in *A. niger*, *S. cerevisiae*, or *H. polymorpha*, the corresponding expression plasmid containing the fungal consensus phytase gene was used as template for site-directed mutagenesis. Mutations were introduced using the "quick exchange site-directed mutagenesis kit" from Stratagene (La Jolla, Calif., USA) following the manufacturer's protocol and using the corresponding primers. All mutations made and the corresponding primers are summarized in Table 4. Clones harboring the desired mutation were identified by DNA sequence analysis as known in the art. The mutated phytase were verified by sequencing of the complete gene.

TABLE 4

mutation	Primer set
Q50L (SEQ ID NO:24)	5'-CAC TTG TGG GGT TTG TAC AGT CCA TAC TTC TC-3'
(SEQ ID NO:25)	5'-GAG AAG TAT GGA CTG TAC AAA CCC CAC AAG TG-3'
Q50T (SEQ ID NO:26)	5'-CAC TTG TGG GGT ACC TAC TCT CCA TAC TTC TC-3'
(SEQ ID NO:27)	5'-GA GAA GTA TGG AGA GTA GGT ACC CCA CAA GTG-3'
Q50G (SEQ ID NO:28)	5'-CAC TTG TGG GGT GGT TAC TCT CCA TAC TTC TC-3'
(SEQ ID NO:29)	5'-GA GAA GTA TGG AGA GTA ACC ACC CCA CAA GTG-3'
Q50T-Y51N (SEQ ID NO:30)	5'-CAC TTG TGG GGT ACC AAC TCT CCA TAC TTC TC-3'
(SEQ ID NO:31)	5'-GA GAA GTA TGG AGA GTT GGT ACC CCA CAA GTG-3'
Q50L-Y51N (SEQ ID NO:32)	5'-CAC TTG TGG GGT CTC AAC TCT CCA TAC TTC TC-3'
(SEQ ID NO:33)	5'-GA GAA GTA TGG AGA GTT GAG ACC CCA CAA GTG-3'

Table 4: Primers used for the introduction of single mutations into fungal consensus phytase. For the introduction of each mutation, two primers containing the desired mutation were required (see Example 8). The changed triplets are highlighted in bold letters.

Example 9

Determination of the Phytase Activity and of the Temperature Optimum of the Consensus Phytase and its Variants

Phytase activity was determined basically as described by Mitchell et al. (1997). The activity was measured in a assay mixture containing 0.5% phytic acid (≈ 5 mM), 200 mM sodium acetate, pH 5.0. After 15 min incubation at 37° C., the reaction was stopped by addition of an equal volume of

15% trichloroacetic acid. The liberated phosphate was quantified by mixing 100 μ l of the assay mixture with 900 μ l H₂O and 1 ml of 0.6 M H₂SO₄, 2% ascorbic acid and 0.5% ammonium molybdate. Standard solutions of potassium phosphate were used as reference. One unit of enzyme activity was defined as the amount of enzyme that releases 1 μ mol phosphate per minute at 37° C. The protein concentration was determined using the enzyme extinction coefficient at 280 nm calculated according to Pace et al. (1995): fungal consensus phytase, 1.101; fungal consensus phytase 7, 1.068. In case of pH-optimum curves, purified enzymes were diluted in 10 mM sodium acetate, pH 5.0. Incubations were started by mixing aliquots of the diluted protein with an equal volume of 1% phytic acid (≈ 10 mM) in a series of different buffers; 0.4 M glycine/HCl, pH 2.5; 0.4 M acetate/NaOH, pH 3.0, 3.5, 4.0, 4.5, 5.0, 5.5; 0.4 M imidazole/HCl, pH 6.0, 6.5; 0.4 M Tris/HCl pH 7.0, 7.5, 8.0, 8.5, 9.0. Control experiments showed that pH was only slightly affected by the mixing step. Incubations were performed for 15 min at 37° C. as described above.

For determination of the substrate specificities of the phytases, phytic acid in the assay mixture was replaced by 5 mM concentrations of the respective phosphate compounds. The activity tests were performed as described above.

For determination of the temperature optimum, enzyme (100 μ l) and substrate solution (100 μ l) were pre-incubated for 5 min at the given temperature. The reaction was started by addition of the substrate solution to the enzyme. After 15 min incubation, the reaction was stopped with trichloroacetic acid and the amount of phosphate released was determined.

The pH-optimum of the original fungal consensus phytase was around pH 6.0–6.5 (70 U/mg). By introduction of the Q50T mutation, the pH-optimum shifted, to pH 6.0 (130 U/mg), while the replacement by a leucine at the same position resulted in a maximum activity around pH 5.5 (212 U/mg). The exchange Q50G resulted in a pH-optimum of the activity above pH 6.0 (see FIG. 4). The exchange of tyrosine at position 51 with asparagine resulted in a relative increase of the activity below pH 5.0 (see FIG. 5). Especially

by the Q50L mutation, the specificity for phytate of fungal consensus phytase was drastically increased (see FIG. 6).

The temperature optimum of fungal consensus phytase (70° C.) was 15–25° C. higher than the temperature optimum of the wild-type phytases (45–55° C.) which were used to calculate the consensus sequence (see Table 5 and FIG. 3).

TABLE 5

phytase	temperature optimum	T _m
Consensus phytase	70° C.	78.0° C.
<i>A. niger</i> NRRL3135	55° C.	63.3° C.
<i>A. fumigatus</i> 13073	55° C.	62.5° C.
<i>A. terreus</i> 9A-1	49° C.	57.5° C.
<i>A. terreus</i> cbs	45° C.	58.5° C.
<i>A. nidulans</i>	45° C.	55.7° C.
<i>M. thermophila</i>	55° C.	—

Table 5: Temperature optimum and T_m-value of fungal consensus phytase and of the phytases from *A. fumigatus*, *A. niger*, *A. nidulans*, and *M. thermophila*. The temperature optima were taken from FIG. 3. ^a The T_m-values were determined by differential scanning calorimetry as described in Example 10 and shown in FIG. 7.

Determination of the Melting point by Differential Scanning Calorimetry (DSC)

In order to determine the unfolding temperature of the fungal consensus phytases, differential scanning calorimetry was applied as previously published by Brugger et al. (1997). Solutions of 50–60 mg/ml homogeneous phytase were used for the tests. A constant heating rate of 10° C./min was applied up to 90° C.

The determined melting points clearly show the strongly improved thermostability of the fungal consensus phytase in comparison to the wild-type phytases (see Table 5 and FIG. 7). FIG. 7 shows the melting profile of fungal consensus phytase and its mutant Q50T. Its common melting point was determined between 78 to 79° C.

SEQUENCE LISTING

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Asp Glu Ser Pro Phe Pro Leu Asp Val Pro Glu Asp Cys His Ile Thr
35          40          45
Phe Val Gln Val Leu Ala Arg His Gly Ala Arg Ser Pro Thr His Ser
50          55          60
Lys Thr Lys Ala Tyr Ala Ala Thr Ile Ala Ala Ile Gln Lys Ser Ala
65          70          75          80
Thr Ala Phe Pro Gly Lys Tyr Ala Phe Leu Gln Ser Tyr Asn Tyr Ser
85          90          95
Leu Asp Ser Glu Glu Leu Thr Pro Phe Gly Arg Asn Gln Leu Arg Asp
100         105         110
Leu Gly Ala Gln Phe Tyr Glu Arg Tyr Asn Ala Leu Thr Arg His Ile
115         120         125
Asn Pro Phe Val Arg Ala Thr Asp Ala Ser Arg Val His Glu Ser Ala
130         135         140
Glu Lys Phe Val Glu Gly Phe Gln Thr Ala Arg Gln Asp Asp His His
145         150         155         160
Ala Asn Pro His Gln Pro Ser Pro Arg Val Asp Val Ala Ile Pro Glu
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Gly Ser Ala Tyr Asn Asn Thr Leu Glu His Ser Leu Cys Thr Ala Phe
180         185         190

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 Gln Leu Ser Thr Asp Asp Val Val Asn Leu Met Ala Met Cys Pro Phe
 225 230 235 240
 Glu Thr Val Ser Leu Thr Asp Asp Ala His Thr Leu Ser Pro Phe Cys
 245 250 255
 Asp Leu Phe Thr Ala Thr Glu Trp Thr Gln Tyr Asn Tyr Leu Leu Ser
 260 265 270
 Leu Asp Lys Tyr Tyr Gly Tyr Gly Gly Asn Pro Leu Gly Pro Val
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 340 345 350
 Ala Pro Leu Ser Gln Thr Ser Val Glu Ser Val Ser Gln Thr Asp Gly
 355 360 365
 Tyr Ala Ala Ala Trp Thr Val Pro Phe Ala Ala Arg Ala Tyr Val Glu
 370 375 380
 Met Met Gln Cys Arg Ala Glu Lys Glu Pro Leu Val Arg Val Leu Val
 385 390 395 400
 Asn Asp Arg Val Met Pro Leu His Gly Cys Pro Thr Asp Lys Leu Gly
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 50 55 60
 Lys Thr Lys Ala Tyr Ala Ala Thr Ile Ala Ala Ile Gln Lys Asn Ala
 65 70 75 80
 Thr Ala Leu Pro Gly Lys Tyr Ala Phe Leu Lys Ser Tyr Asn Tyr Ser
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 35 40 45
 Phe Ala Gln Val Leu Ser Arg His Gly Ala Arg Tyr Pro Thr Glu Ser

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Leu	Gly	Ala	Asp	Asp	Leu	Thr	Pro	Phe	Gly	Glu	Gln	Glu	Leu	Val	Asn					
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Ile	Pro	Phe	Ile	Arg	Ser	Ser	Gly	Ser	Ser	Arg	Val	Ile	Ala	Ser	Gly					
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 Ser Gly Ile Lys Phe Tyr Gln Arg Tyr Glu Ser Leu Thr Arg Asn Ile
 115 120 125
 Ile Pro Phe Ile Arg Ser Ser Gly Ser Ser Arg Val Ile Ala Ser Gly
 130 135 140
 Glu Lys Phe Ile Glu Gly Phe Gln Ser Thr Lys Leu Lys Asp Pro Arg
 145 150 155 160
 Ala Gln Pro Gly Gln Ser Ser Pro Lys Ile Asp Val Val Ile Ser Glu
 165 170 175
 Ala Ser Ser Ser Asn Asn Thr Leu Asp Pro Gly Thr Cys Thr Val Phe
 180 185 190
 Glu Asp Ser Glu Leu Ala Asp Thr Val Glu Ala Asn Phe Thr Ala Thr
 195 200 205
 Phe Ala Pro Ser Ile Arg Gln Arg Leu Glu Asn Asp Leu Ser Gly Val
 210 215 220
 Thr Leu Thr Asp Thr Glu Val Thr Tyr Leu Met Asp Met Cys Ser Phe
 225 230 235 240
 Asp Thr Ile Ser Thr Ser Thr Val Asp Thr Lys Leu Ser Pro Phe Cys
 245 250 255
 Asp Leu Phe Thr His Asp Glu Trp Ile His Tyr Asp Tyr Leu Arg Ser
 260 265 270
 Leu Lys Lys Tyr Tyr Gly His Gly Ala Gly Asn Pro Leu Gly Pro Thr
 275 280 285
 Gln Gly Val Gly Tyr Ala Asn Glu Leu Ile Ala Arg Leu Thr His Ser
 290 295 300
 Pro Val His Asp Asp Thr Ser Ser Asn His Thr Leu Asp Ser Asn Pro
 305 310 315 320
 Ala Thr Phe Pro Leu Asn Ser Thr Leu Tyr Ala Asp Phe Ser His Asp
 325 330 335
 Asn Gly Ile Ile Ser Ile Leu Phe Ala Leu Gly Leu Tyr Asn Gly Thr
 340 345 350
 Lys Pro Leu Ser Thr Thr Thr Val Glu Asn Ile Thr Gln Thr Asp Gly
 355 360 365
 Phe Ser Ser Ala Trp Thr Val Pro Phe Ala Ser Arg Leu Tyr Val Glu
 370 375 380
 Met Met Gln Cys Gln Ala Glu Gln Glu Pro Leu Val Arg Val Leu Val
 385 390 395 400
 Asn Asp Arg Val Val Pro Leu His Gly Cys Pro Ile Asp Ala Leu Gly

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	405		410		415
Arg Cys Thr Arg Asp Ser Phe Val Arg Gly Leu Ser Phe Ala Arg Ser					
	420		425		430
Gly Gly Asp Trp Ala Glu Cys Phe Ala					
	435		440		

<210> SEQ ID NO 5
 <211> LENGTH: 441
 <212> TYPE: PRT
 <213> ORGANISM: Aspergillus niger

<400> SEQUENCE: 5

Asn Gln Ser Ser Cys Asp Thr Val Asp Gln Gly Tyr Gln Cys Phe Ser					
1	5		10		15
Glu Thr Ser His Leu Trp Gly Gln Tyr Ala Pro Phe Phe Ser Leu Ala					
	20		25		30
Asn Glu Ser Val Ile Ser Pro Glu Val Pro Ala Gly Cys Arg Val Thr					
	35		40		45
Phe Ala Gln Val Leu Ser Arg His Gly Ala Arg Tyr Pro Thr Asp Ser					
	50		55		60
Lys Gly Lys Lys Tyr Ser Ala Leu Ile Glu Glu Ile Gln Gln Asn Ala					
65	70		75		80
Thr Thr Phe Asp Gly Lys Tyr Ala Phe Leu Lys Thr Tyr Asn Tyr Ser					
	85		90		95
Leu Gly Ala Asp Asp Leu Thr Pro Phe Gly Glu Gln Glu Leu Val Asn					
	100		105		110
Ser Gly Ile Lys Phe Tyr Gln Arg Tyr Glu Ser Leu Thr Arg Asn Ile					
	115		120		125
Val Pro Phe Ile Arg Ser Ser Gly Ser Ser Arg Val Ile Ala Ser Gly					
	130		135		140
Lys Lys Phe Ile Glu Gly Phe Gln Ser Thr Lys Leu Lys Asp Pro Arg					
145	150		155		160
Ala Gln Pro Gly Gln Ser Ser Pro Lys Ile Asp Val Val Ile Ser Glu					
	165		170		175
Ala Ser Ser Ser Asn Asn Thr Leu Asp Pro Gly Thr Cys Thr Val Phe					
	180		185		190
Glu Asp Ser Glu Leu Ala Asp Thr Val Glu Ala Asn Phe Thr Ala Thr					
	195		200		205
Phe Val Pro Ser Ile Arg Gln Arg Leu Glu Asn Asp Leu Ser Gly Val					
	210		215		220
Thr Leu Thr Asp Thr Glu Val Thr Tyr Leu Met Asp Met Cys Ser Phe					
225	230		235		240
Asp Thr Ile Ser Thr Ser Thr Val Asp Thr Lys Leu Ser Pro Phe Cys					
	245		250		255
Asp Leu Phe Thr His Asp Glu Trp Ile Asn Tyr Asp Tyr Leu Gln Ser					
	260		265		270
Leu Lys Lys Tyr Tyr Gly His Gly Ala Gly Asn Pro Leu Gly Pro Thr					
	275		280		285
Gln Gly Val Gly Tyr Ala Asn Glu Leu Ile Ala Arg Leu Thr His Ser					
	290		295		300
Pro Val His Asp Asp Thr Ser Ser Asn His Thr Leu Asp Ser Ser Pro					
305	310		315		320
Ala Thr Phe Pro Leu Asn Ser Thr Leu Tyr Ala Asp Phe Ser His Asp					
	325		330		335

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Asn Gly Ile Ile Ser Ile Leu Phe Ala Leu Gly Leu Tyr Asn Gly Thr
 340 345 350

Lys Pro Leu Ser Thr Thr Thr Val Glu Asn Ile Thr Gln Thr Asp Gly
 355 360 365

Phe Ser Ser Ala Trp Thr Val Pro Phe Ala Ser Arg Leu Tyr Val Glu
 370 375 380

Met Met Gln Cys Gln Ala Glu Gln Glu Pro Leu Val Arg Val Leu Val
 385 390 395 400

Asn Asp Arg Val Val Pro Leu His Gly Cys Pro Val Asp Ala Leu Gly
 405 410 415

Arg Cys Thr Arg Asp Ser Phe Val Arg Gly Leu Ser Phe Ala Arg Ser
 420 425 430

Gly Gly Asp Trp Ala Glu Cys Phe Ala
 435 440

<210> SEQ ID NO 6

<211> LENGTH: 440

<212> TYPE: PRT

<213> ORGANISM: Aspergillus fumigatus

<400> SEQUENCE: 6

Gly Ser Lys Ser Cys Asp Thr Val Asp Leu Gly Tyr Gln Cys Ser Pro
 1 5 10 15

Ala Thr Ser His Leu Trp Gly Gln Tyr Ser Pro Phe Phe Ser Leu Glu
 20 25 30

Asp Glu Leu Ser Val Ser Ser Lys Leu Pro Lys Asp Cys Arg Ile Thr
 35 40 45

Leu Val Gln Val Leu Ser Arg His Gly Ala Arg Tyr Pro Thr Ser Ser
 50 55 60

Lys Ser Lys Lys Tyr Lys Lys Leu Val Thr Ala Ile Gln Ala Asn Ala
 65 70 75 80

Thr Asp Phe Lys Gly Lys Phe Ala Phe Leu Lys Thr Tyr Asn Tyr Thr
 85 90 95

Leu Gly Ala Asp Asp Leu Thr Pro Phe Gly Glu Gln Gln Leu Val Asn
 100 105 110

Ser Gly Ile Lys Phe Tyr Gln Arg Tyr Lys Ala Leu Ala Arg Ser Val
 115 120 125

Val Pro Phe Ile Arg Ala Ser Gly Ser Asp Arg Val Ile Ala Ser Gly
 130 135 140

Glu Lys Phe Ile Glu Gly Phe Gln Gln Ala Lys Leu Ala Asp Pro Gly
 145 150 155 160

Ala Thr Asn Arg Ala Ala Pro Ala Ile Ser Val Ile Ile Pro Glu Ser
 165 170 175

Glu Thr Phe Asn Asn Thr Leu Asp His Gly Val Cys Thr Lys Phe Glu
 180 185 190

Ala Ser Gln Leu Gly Asp Glu Val Ala Ala Asn Phe Thr Ala Leu Phe
 195 200 205

Ala Pro Asp Ile Arg Ala Arg Ala Glu Lys His Leu Pro Gly Val Thr
 210 215 220

Leu Thr Asp Glu Asp Val Val Ser Leu Met Asp Met Cys Ser Phe Asp
 225 230 235 240

Thr Val Ala Arg Thr Ser Asp Ala Ser Gln Leu Ser Pro Phe Cys Gln
 245 250 255

Leu Phe Thr His Asn Glu Trp Lys Lys Tyr Asn Tyr Leu Gln Ser Leu
 260 265 270

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Gly Lys Tyr Tyr Gly Tyr Gly Ala Gly Asn Pro Leu Gly Pro Ala Gln
 275 280 285
 Gly Ile Gly Phe Thr Asn Glu Leu Ile Ala Arg Leu Thr Arg Ser Pro
 290 295 300
 Val Gln Asp His Thr Ser Thr Asn Ser Thr Leu Val Ser Asn Pro Ala
 305 310 315 320
 Thr Phe Pro Leu Asn Ala Thr Met Tyr Val Asp Phe Ser His Asp Asn
 325 330 335
 Ser Met Val Ser Ile Phe Phe Ala Leu Gly Leu Tyr Asn Gly Thr Glu
 340 345 350
 Pro Leu Ser Arg Thr Ser Val Glu Ser Ala Lys Glu Leu Asp Gly Tyr
 355 360 365
 Ser Ala Ser Trp Val Val Pro Phe Gly Ala Arg Ala Tyr Phe Glu Thr
 370 375 380
 Met Gln Cys Lys Ser Glu Lys Glu Pro Leu Val Arg Ala Leu Ile Asn
 385 390 395 400
 Asp Arg Val Val Pro Leu His Gly Cys Asp Val Asp Lys Leu Gly Arg
 405 410 415
 Cys Lys Leu Asn Asp Phe Val Lys Gly Leu Ser Trp Ala Arg Ser Gly
 420 425 430
 Gly Asn Trp Gly Glu Cys Phe Ser
 435 440

<210> SEQ ID NO 7
 <211> LENGTH: 440
 <212> TYPE: PRT
 <213> ORGANISM: Aspergillus fumigatus

<400> SEQUENCE: 7

Gly Ser Lys Ser Cys Asp Thr Val Asp Leu Gly Tyr Gln Cys Ser Pro
 1 5 10 15
 Ala Thr Ser His Leu Trp Gly Gln Tyr Ser Pro Phe Phe Ser Leu Glu
 20 25 30
 Asp Glu Leu Ser Val Ser Ser Lys Leu Pro Lys Asp Cys Arg Ile Thr
 35 40 45
 Leu Val Gln Val Leu Ser Arg His Gly Ala Arg Tyr Pro Thr Ser Ser
 50 55 60
 Lys Ser Lys Lys Tyr Lys Lys Leu Val Thr Ala Ile Gln Ala Asn Ala
 65 70 75 80
 Thr Asp Phe Lys Gly Lys Phe Ala Phe Leu Lys Thr Tyr Asn Tyr Thr
 85 90 95
 Leu Gly Ala Asp Asp Leu Thr Pro Phe Gly Glu Gln Gln Leu Val Asn
 100 105 110
 Ser Gly Ile Lys Phe Tyr Gln Arg Tyr Lys Ala Leu Ala Arg Ser Val
 115 120 125
 Val Pro Phe Ile Arg Ala Ser Gly Ser Asp Arg Val Ile Ala Ser Gly
 130 135 140
 Glu Lys Phe Ile Glu Gly Phe Gln Gln Ala Lys Leu Ala Asp Pro Gly
 145 150 155 160
 Ala Thr Asn Arg Ala Ala Pro Ala Ile Ser Val Ile Ile Pro Glu Ser
 165 170 175
 Glu Thr Phe Asn Asn Thr Leu Asp His Gly Val Cys Thr Lys Phe Glu
 180 185 190
 Ala Ser Gln Leu Gly Asp Glu Val Ala Ala Asn Phe Thr Ala Leu Phe

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195				200				205							
Ala	Pro	Asp	Ile	Arg	Ala	Arg	Ala	Glu	Lys	His	Leu	Pro	Gly	Val	Thr
	210					215					220				
Leu	Thr	Asp	Glu	Asp	Val	Val	Ser	Leu	Met	Asp	Met	Cys	Ser	Phe	Asp
	225				230					235					240
Thr	Val	Ala	Arg	Thr	Ser	Asp	Ala	Ser	Gln	Leu	Ser	Pro	Phe	Cys	Gln
				245					250					255	
Leu	Phe	Thr	His	Asn	Glu	Trp	Lys	Lys	Tyr	Asn	Tyr	Leu	Gln	Ser	Leu
			260						265				270		
Gly	Lys	Tyr	Tyr	Gly	Tyr	Gly	Ala	Gly	Asn	Pro	Leu	Gly	Pro	Ala	Gln
		275					280					285			
Gly	Ile	Gly	Phe	Thr	Asn	Glu	Leu	Ile	Ala	Arg	Leu	Thr	Arg	Ser	Pro
	290					295					300				
Val	Gln	Asp	His	Thr	Ser	Thr	Asn	Ser	Thr	Leu	Val	Ser	Asn	Pro	Ala
	305					310				315				320	
Thr	Phe	Pro	Leu	Asn	Ala	Thr	Met	Tyr	Val	Asp	Phe	Ser	His	Asp	Asn
				325					330					335	
Ser	Met	Val	Ser	Ile	Phe	Phe	Ala	Leu	Gly	Leu	Tyr	Asn	Gly	Thr	Gly
			340					345					350		
Pro	Leu	Ser	Arg	Thr	Ser	Val	Glu	Ser	Ala	Lys	Glu	Leu	Asp	Gly	Tyr
		355					360					365			
Ser	Ala	Ser	Trp	Val	Val	Pro	Phe	Gly	Ala	Arg	Ala	Tyr	Phe	Glu	Thr
	370					375					380				
Met	Gln	Cys	Lys	Ser	Glu	Lys	Glu	Pro	Leu	Val	Arg	Ala	Leu	Ile	Asn
	385				390					395					400
Asp	Arg	Val	Val	Pro	Leu	His	Gly	Cys	Asp	Val	Asp	Lys	Leu	Gly	Arg
				405					410					415	
Cys	Lys	Leu	Asn	Asp	Phe	Val	Lys	Gly	Leu	Ser	Trp	Ala	Arg	Ser	Gly
			420					425				430			
Gly	Asn	Trp	Gly	Glu	Cys	Phe	Ser								
	435						440								

<210> SEQ ID NO 8

<211> LENGTH: 440

<212> TYPE: PRT

<213> ORGANISM: Aspergillus fumigatus

<400> SEQUENCE: 8

Gly	Ser	Lys	Ser	Cys	Asp	Thr	Val	Asp	Leu	Gly	Tyr	Gln	Cys	Ser	Pro
1				5					10					15	
Ala	Thr	Ser	His	Leu	Trp	Gly	Gln	Tyr	Ser	Pro	Phe	Phe	Ser	Leu	Glu
			20					25					30		
Asp	Glu	Leu	Ser	Val	Ser	Ser	Lys	Leu	Pro	Lys	Asp	Cys	Arg	Ile	Thr
		35					40					45			
Leu	Val	Gln	Val	Leu	Ser	Arg	His	Gly	Ala	Arg	Tyr	Pro	Thr	Ser	Ser
	50					55					60				
Lys	Ser	Lys	Lys	Tyr	Lys	Lys	Leu	Val	Thr	Ala	Ile	Gln	Ala	Asn	Ala
	65				70					75					80
Thr	Asp	Phe	Lys	Gly	Lys	Phe	Ala	Phe	Leu	Lys	Thr	Tyr	Asn	Tyr	Thr
			85						90					95	
Leu	Gly	Ala	Asp	Asp	Leu	Thr	Pro	Phe	Gly	Glu	Gln	Gln	Leu	Val	Asn
			100					105					110		
Ser	Gly	Ile	Lys	Phe	Tyr	Gln	Arg	Tyr	Lys	Ala	Leu	Ala	Arg	Ser	Val
		115					120					125			

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Val	Pro	Phe	Ile	Arg	Ala	Ser	Gly	Ser	Asp	Arg	Val	Ile	Ala	Ser	Gly
	130					135					140				
Glu	Lys	Phe	Ile	Glu	Gly	Phe	Gln	Gln	Ala	Lys	Leu	Ala	Asp	Pro	Gly
	145				150					155					160
Ala	Thr	Asn	Arg	Ala	Ala	Pro	Ala	Ile	Ser	Val	Ile	Ile	Pro	Glu	Ser
				165					170					175	
Glu	Thr	Phe	Asn	Asn	Thr	Leu	Asp	His	Gly	Val	Cys	Thr	Lys	Phe	Glu
			180					185					190		
Ala	Ser	Gln	Leu	Gly	Asp	Glu	Val	Ala	Ala	Asn	Phe	Thr	Ala	Leu	Phe
		195					200					205			
Ala	Pro	Asp	Ile	Arg	Ala	Arg	Ala	Glu	Lys	His	Leu	Pro	Gly	Val	Thr
	210					215					220				
Leu	Thr	Asp	Glu	Asp	Val	Val	Ser	Leu	Met	Asp	Met	Cys	Ser	Phe	Asp
	225				230					235					240
Thr	Val	Ala	Arg	Thr	Ser	Asp	Ala	Ser	Gln	Leu	Ser	Pro	Phe	Cys	Gln
				245					250					255	
Leu	Phe	Thr	His	Asn	Glu	Trp	Lys	Lys	Tyr	Asn	Tyr	Leu	Gln	Ser	Leu
			260					265					270		
Gly	Lys	Tyr	Tyr	Gly	Tyr	Gly	Ala	Gly	Asn	Pro	Leu	Gly	Pro	Ala	Gln
		275					280					285			
Gly	Ile	Gly	Phe	Thr	Asn	Glu	Leu	Ile	Ala	Arg	Leu	Thr	Arg	Ser	Pro
	290					295					300				
Val	Gln	Asp	His	Thr	Ser	Thr	Asn	Ser	Thr	Leu	Val	Ser	Asn	Pro	Ala
	305				310					315					320
Thr	Phe	Pro	Leu	Asn	Ala	Thr	Met	Tyr	Val	Asp	Phe	Ser	His	Asp	Asn
				325					330					335	
Ser	Met	Val	Ser	Ile	Phe	Phe	Ala	Leu	Gly	Leu	Tyr	Asn	Gly	Thr	Glu
			340					345					350		
Pro	Leu	Ser	Arg	Thr	Ser	Val	Glu	Ser	Ala	Lys	Glu	Leu	Asp	Gly	Tyr
			355				360					365			
Ser	Ala	Ser	Trp	Val	Val	Pro	Phe	Gly	Ala	Arg	Ala	Tyr	Phe	Glu	Thr
	370					375					380				
Met	Gln	Cys	Lys	Ser	Glu	Lys	Glu	Ser	Leu	Val	Arg	Ala	Leu	Ile	Asn
	385				390					395					400
Asp	Arg	Val	Val	Pro	Leu	His	Gly	Cys	Asp	Val	Asp	Lys	Leu	Gly	Arg
				405					410					415	
Cys	Lys	Leu	Asn	Asp	Phe	Val	Lys	Gly	Leu	Ser	Trp	Ala	Arg	Ser	Gly
			420					425					430		
Gly	Asn	Trp	Gly	Glu	Cys	Phe	Ser								
	435						440								

<210> SEQ ID NO 9

<211> LENGTH: 440

<212> TYPE: PRT

<213> ORGANISM: Aspergillus fumigatus

<400> SEQUENCE: 9

Gly	Ser	Lys	Ser	Cys	Asp	Thr	Val	Asp	Leu	Gly	Tyr	Gln	Cys	Ser	Pro
1				5					10					15	
Ala	Thr	Ser	His	Leu	Trp	Gly	Gln	Tyr	Ser	Pro	Phe	Phe	Ser	Leu	Glu
			20					25					30		
Asp	Glu	Leu	Ser	Val	Ser	Ser	Lys	Leu	Pro	Lys	Asp	Cys	Arg	Ile	Thr
	35						40					45			
Leu	Val	Gln	Val	Leu	Ser	Arg	His	Gly	Ala	Arg	Tyr	Pro	Thr	Ser	Ser
	50					55					60				

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Lys Ser Lys Lys Tyr Lys Lys Leu Val Thr Ala Ile Gln Ala Asn Ala
 65 70 75 80
 Thr Asp Phe Lys Gly Lys Phe Ala Phe Leu Lys Thr Tyr Asn Tyr Thr
 85 90 95
 Leu Gly Ala Asp Asp Leu Thr Ala Phe Gly Glu Gln Gln Leu Val Asn
 100 105 110
 Ser Gly Ile Lys Phe Tyr Gln Arg Tyr Lys Ala Leu Ala Arg Ser Val
 115 120 125
 Val Pro Phe Ile Arg Ala Ser Gly Ser Asp Arg Val Ile Ala Ser Gly
 130 135 140
 Glu Lys Phe Ile Glu Gly Phe Gln Gln Ala Lys Leu Ala Asp Pro Gly
 145 150 155 160
 Ala Thr Asn Arg Ala Ala Pro Ala Ile Ser Val Ile Ile Pro Glu Ser
 165 170 175
 Glu Thr Phe Asn Asn Thr Leu Asp His Gly Val Cys Thr Lys Phe Glu
 180 185 190
 Ala Ser Gln Leu Gly Asp Glu Val Ala Ala Asn Phe Thr Ala Leu Phe
 195 200 205
 Ala Pro Asp Ile Arg Ala Arg Ala Lys Lys His Leu Pro Gly Val Thr
 210 215 220
 Leu Thr Asp Glu Asp Val Val Ser Leu Met Asp Met Cys Ser Phe Asp
 225 230 235 240
 Thr Val Ala Arg Thr Ser Asp Ala Ser Gln Leu Ser Pro Phe Cys Gln
 245 250 255
 Leu Phe Thr His Asn Glu Trp Lys Lys Tyr Asn Tyr Leu Gln Ser Leu
 260 265 270
 Gly Lys Tyr Tyr Gly Tyr Gly Ala Gly Asn Pro Leu Gly Pro Ala Gln
 275 280 285
 Gly Ile Gly Phe Thr Asn Glu Leu Ile Ala Arg Leu Thr Arg Ser Pro
 290 295 300
 Val Gln Asp His Thr Ser Thr Asn Ser Thr Leu Val Ser Asn Pro Ala
 305 310 315 320
 Thr Phe Pro Leu Asn Ala Thr Met Tyr Val Asp Phe Ser His Asp Asn
 325 330 335
 Ser Met Val Ser Ile Phe Phe Ala Leu Gly Leu Tyr Asn Gly Thr Glu
 340 345 350
 Pro Leu Ser Arg Thr Ser Val Glu Ser Ala Lys Glu Leu Asp Gly Tyr
 355 360 365
 Ser Ala Ser Trp Val Val Pro Phe Gly Ala Arg Ala Tyr Phe Glu Thr
 370 375 380
 Met Gln Cys Lys Ser Glu Lys Glu Pro Leu Val Arg Ala Leu Ile Asn
 385 390 395 400
 Asp Arg Val Val Pro Leu His Gly Cys Asp Val Asp Lys Leu Gly Arg
 405 410 415
 Cys Lys Leu Asn Asp Phe Val Lys Gly Leu Ser Trp Ala Arg Ser Gly
 420 425 430
 Gly Asn Trp Gly Glu Cys Phe Ser
 435 440

<210> SEQ ID NO 10

<211> LENGTH: 440

<212> TYPE: PRT

<213> ORGANISM: Aspergillus fumigatus

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<400> SEQUENCE: 10

Gly	Ser	Lys	Ala	Cys	Asp	Thr	Val	Glu	Leu	Gly	Tyr	Gln	Cys	Ser	Pro	1	5	10	15
Gly	Thr	Ser	His	Leu	Trp	Gly	Gln	Tyr	Ser	Pro	Phe	Phe	Ser	Leu	Glu	20	25	30	
Asp	Glu	Leu	Ser	Val	Ser	Ser	Asp	Leu	Pro	Lys	Asp	Cys	Arg	Val	Thr	35	40	45	
Phe	Val	Gln	Val	Leu	Ser	Arg	His	Gly	Ala	Arg	Tyr	Pro	Thr	Ala	Ser	50	55	60	
Lys	Ser	Lys	Lys	Tyr	Lys	Lys	Leu	Val	Thr	Ala	Ile	Gln	Lys	Asn	Ala	65	70	75	80
Thr	Glu	Phe	Lys	Gly	Lys	Phe	Ala	Phe	Leu	Glu	Thr	Tyr	Asn	Tyr	Thr	85	90	95	
Leu	Gly	Ala	Asp	Asp	Leu	Thr	Pro	Phe	Gly	Glu	Gln	Gln	Met	Val	Asn	100	105	110	
Ser	Gly	Ile	Lys	Phe	Tyr	Gln	Lys	Tyr	Lys	Ala	Leu	Ala	Gly	Ser	Val	115	120	125	
Val	Pro	Phe	Ile	Arg	Ser	Ser	Gly	Ser	Asp	Arg	Val	Ile	Ala	Ser	Gly	130	135	140	
Glu	Lys	Phe	Ile	Glu	Gly	Phe	Gln	Gln	Ala	Asn	Val	Ala	Asp	Pro	Gly	145	150	155	160
Ala	Thr	Asn	Arg	Ala	Ala	Pro	Val	Ile	Ser	Val	Ile	Ile	Pro	Glu	Ser	165	170	175	
Glu	Thr	Tyr	Asn	Asn	Thr	Leu	Asp	His	Ser	Val	Cys	Thr	Asn	Phe	Glu	180	185	190	
Ala	Ser	Glu	Leu	Gly	Asp	Glu	Val	Glu	Ala	Asn	Phe	Thr	Ala	Leu	Phe	195	200	205	
Ala	Pro	Ala	Ile	Arg	Ala	Arg	Ile	Glu	Lys	His	Leu	Pro	Gly	Val	Gln	210	215	220	
Leu	Thr	Asp	Asp	Asp	Val	Val	Ser	Leu	Met	Asp	Met	Cys	Ser	Phe	Asp	225	230	235	240
Thr	Val	Ala	Arg	Thr	Ala	Asp	Ala	Ser	Glu	Leu	Ser	Pro	Phe	Cys	Ala	245	250	255	
Ile	Phe	Thr	His	Asn	Glu	Trp	Lys	Lys	Tyr	Asp	Tyr	Leu	Gln	Ser	Leu	260	265	270	
Gly	Lys	Tyr	Tyr	Gly	Tyr	Gly	Ala	Gly	Asn	Pro	Leu	Gly	Pro	Ala	Gln	275	280	285	
Gly	Ile	Gly	Phe	Thr	Asn	Glu	Leu	Ile	Ala	Arg	Leu	Thr	Asn	Ser	Pro	290	295	300	
Val	Gln	Asp	His	Thr	Ser	Thr	Asn	Ser	Thr	Leu	Asp	Ser	Asp	Pro	Ala	305	310	315	320
Thr	Phe	Pro	Leu	Asn	Ala	Thr	Ile	Tyr	Val	Asp	Phe	Ser	His	Asp	Asn	325	330	335	
Gly	Met	Ile	Pro	Ile	Phe	Phe	Ala	Met	Gly	Leu	Tyr	Asn	Gly	Thr	Glu	340	345	350	
Pro	Leu	Ser	Gln	Thr	Ser	Glu	Glu	Ser	Thr	Lys	Glu	Ser	Asn	Gly	Tyr	355	360	365	
Ser	Ala	Ser	Trp	Ala	Val	Pro	Phe	Gly	Ala	Arg	Ala	Tyr	Phe	Glu	Thr	370	375	380	
Met	Gln	Cys	Lys	Ser	Glu	Lys	Glu	Pro	Leu	Val	Arg	Ala	Leu	Ile	Asn	385	390	395	400
Asp	Arg	Val	Val	Pro	Leu	His	Gly	Cys	Ala	Val	Asp	Lys	Leu	Gly	Arg	405	410	415	

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Cys Lys Leu Lys Asp Phe Val Lys Gly Leu Ser Trp Ala Arg Ser Gly
 420 425 430

Gly Asn Ser Glu Gln Ser Phe Ser
 435 440

<210> SEQ ID NO 11

<211> LENGTH: 439

<212> TYPE: PRT

<213> ORGANISM: Aspergillus nidulans

<400> SEQUENCE: 11

Gln Asn His Ser Cys Asn Thr Ala Asp Gly Gly Tyr Gln Cys Phe Pro
 1 5 10 15

Asn Val Ser His Val Trp Gly Gln Tyr Ser Pro Tyr Phe Ser Ile Glu
 20 25 30

Gln Glu Ser Ala Ile Ser Glu Asp Val Pro His Gly Cys Glu Val Thr
 35 40 45

Phe Val Gln Val Leu Ser Arg His Gly Ala Arg Tyr Pro Thr Glu Ser
 50 55 60

Lys Ser Lys Ala Tyr Ser Gly Leu Ile Glu Ala Ile Gln Lys Asn Ala
 65 70 75 80

Thr Ser Phe Trp Gly Gln Tyr Ala Phe Leu Glu Ser Tyr Asn Tyr Thr
 85 90 95

Leu Gly Ala Asp Asp Leu Thr Ile Phe Gly Glu Asn Gln Met Val Asp
 100 105 110

Ser Gly Ala Lys Phe Tyr Arg Arg Tyr Lys Asn Leu Ala Arg Lys Asn
 115 120 125

Thr Pro Phe Ile Arg Ala Ser Gly Ser Asp Arg Val Val Ala Ser Ala
 130 135 140

Glu Lys Phe Ile Asn Gly Phe Arg Lys Ala Gln Leu His Asp His Gly
 145 150 155 160

Ser Gly Gln Ala Thr Pro Val Val Asn Val Ile Ile Pro Glu Ile Asp
 165 170 175

Gly Phe Asn Asn Thr Leu Asp His Ser Thr Cys Val Ser Phe Glu Asn
 180 185 190

Asp Glu Arg Ala Asp Glu Ile Glu Ala Asn Phe Thr Ala Ile Met Gly
 195 200 205

Pro Pro Ile Arg Lys Arg Leu Glu Asn Asp Leu Pro Gly Ile Lys Leu
 210 215 220

Thr Asn Glu Asn Val Ile Tyr Leu Met Asp Met Cys Ser Phe Asp Thr
 225 230 235 240

Met Ala Arg Thr Ala His Gly Thr Glu Leu Ser Pro Phe Cys Ala Ile
 245 250 255

Phe Thr Glu Lys Glu Trp Leu Gln Tyr Asp Tyr Leu Gln Ser Leu Ser
 260 265 270

Lys Tyr Tyr Gly Tyr Gly Ala Gly Ser Pro Leu Gly Pro Ala Gln Gly
 275 280 285

Ile Gly Phe Thr Asn Glu Leu Ile Ala Arg Leu Thr Gln Ser Pro Val
 290 295 300

Gln Asp Asn Thr Ser Thr Asn His Thr Leu Asp Ser Asn Pro Ala Thr
 305 310 315 320

Phe Pro Leu Asp Arg Lys Leu Tyr Ala Asp Phe Ser His Asp Asn Ser
 325 330 335

Met Ile Ser Ile Phe Phe Ala Met Gly Leu Tyr Asn Gly Thr Gln Pro

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340				345				350							
Leu	Ser	Met	Asp	Ser	Val	Glu	Ser	Ile	Gln	Glu	Met	Asp	Gly	Tyr	Ala
		355					360					365			
Ala	Ser	Trp	Thr	Val	Pro	Phe	Gly	Ala	Arg	Ala	Tyr	Phe	Glu	Leu	Met
	370					375				380					
Gln	Cys	Glu	Lys	Lys	Glu	Pro	Leu	Val	Arg	Val	Leu	Val	Asn	Asp	Arg
385					390					395					400
Val	Val	Pro	Leu	His	Gly	Cys	Ala	Val	Asp	Lys	Phe	Gly	Arg	Cys	Thr
				405					410					415	
Leu	Asp	Asp	Trp	Val	Glu	Gly	Leu	Asn	Phe	Ala	Arg	Ser	Gly	Gly	Asn
			420					425					430		
Trp	Lys	Thr	Cys	Phe	Thr	Leu									
		435													

<210> SEQ ID NO 12

<211> LENGTH: 443

<212> TYPE: PRT

<213> ORGANISM: Talaromyces thermophilus

<400> SEQUENCE: 12

Asp	Ser	His	Ser	Cys	Asn	Thr	Val	Glu	Gly	Gly	Tyr	Gln	Cys	Arg	Pro
1				5					10					15	
Glu	Ile	Ser	His	Ser	Trp	Gly	Gln	Tyr	Ser	Pro	Phe	Phe	Ser	Leu	Ala
			20					25					30		
Asp	Gln	Ser	Glu	Ile	Ser	Pro	Asp	Val	Pro	Gln	Asn	Cys	Lys	Ile	Thr
		35					40					45			
Phe	Val	Gln	Leu	Leu	Ser	Arg	His	Gly	Ala	Arg	Tyr	Pro	Thr	Ser	Ser
	50					55					60				
Lys	Thr	Glu	Leu	Tyr	Ser	Gln	Leu	Ile	Ser	Arg	Ile	Gln	Lys	Thr	Ala
65					70					75					80
Thr	Ala	Tyr	Lys	Gly	Tyr	Tyr	Ala	Phe	Leu	Lys	Asp	Tyr	Arg	Tyr	Gln
				85					90					95	
Leu	Gly	Ala	Asn	Asp	Leu	Thr	Pro	Phe	Gly	Glu	Asn	Gln	Met	Ile	Gln
			100					105					110		
Leu	Gly	Ile	Lys	Phe	Tyr	Asn	His	Tyr	Lys	Ser	Leu	Ala	Arg	Asn	Ala
		115					120					125			
Val	Pro	Phe	Val	Arg	Cys	Ser	Gly	Ser	Asp	Arg	Val	Ile	Ala	Ser	Gly
	130					135					140				
Arg	Leu	Phe	Ile	Glu	Gly	Phe	Gln	Ser	Ala	Lys	Val	Leu	Asp	Pro	His
145					150					155					160
Ser	Asp	Lys	His	Asp	Ala	Pro	Pro	Thr	Ile	Asn	Val	Ile	Ile	Glu	Glu
				165					170					175	
Gly	Pro	Ser	Tyr	Asn	Asn	Thr	Leu	Asp	Thr	Gly	Ser	Cys	Pro	Val	Phe
			180					185					190		
Glu	Asp	Ser	Ser	Gly	Gly	His	Asp	Ala	Gln	Glu	Lys	Phe	Ala	Lys	Gln
		195					200					205			
Phe	Ala	Pro	Ala	Ile	Leu	Glu	Lys	Ile	Lys	Asp	His	Leu	Pro	Gly	Val
	210					215					220				
Asp	Leu	Ala	Val	Ser	Asp	Val	Pro	Tyr	Leu	Met	Asp	Leu	Cys	Pro	Phe
225					230					235					240
Glu	Thr	Leu	Ala	Arg	Asn	His	Thr	Asp	Thr	Leu	Ser	Pro	Phe	Cys	Ala
				245					250					255	
Leu	Ser	Thr	Gln	Glu	Glu	Trp	Gln	Ala	Tyr	Asp	Tyr	Tyr	Gln	Ser	Leu
			260					265					270		

-continued

Gly Lys Tyr Tyr Gly Asn Gly Gly Gly Asn Pro Leu Gly Pro Ala Gln
 275 280 285

 Gly Val Gly Phe Val Asn Glu Leu Ile Ala Arg Met Thr His Ser Pro
 290 295 300

 Val Gln Asp Tyr Thr Thr Val Asn His Thr Leu Asp Ser Asn Pro Ala
 305 310 315 320

 Thr Phe Pro Leu Asn Ala Thr Leu Tyr Ala Asp Phe Ser His Asp Asn
 325 330 335

 Thr Met Thr Ser Ile Phe Ala Ala Leu Gly Leu Tyr Asn Gly Thr Ala
 340 345 350

 Lys Leu Ser Thr Thr Glu Ile Lys Ser Ile Glu Glu Thr Asp Gly Tyr
 355 360 365

 Ser Ala Ala Trp Thr Val Pro Phe Gly Gly Arg Ala Tyr Ile Glu Met
 370 375 380

 Met Gln Cys Asp Asp Ser Asp Glu Pro Val Val Arg Val Leu Val Asn
 385 390 395 400

 Asp Arg Val Val Pro Leu His Gly Cys Glu Val Asp Ser Leu Gly Arg
 405 410 415

 Cys Lys Arg Asp Asp Phe Val Arg Gly Leu Ser Phe Ala Arg Gln Gly
 420 425 430

 Gly Asn Trp Glu Gly Cys Tyr Ala Ala Ser Glu
 435 440

<210> SEQ ID NO 13

<211> LENGTH: 466

<212> TYPE: PRT

<213> ORGANISM: Myceliophthora thermophila

<400> SEQUENCE: 13

Glu Ser Arg Pro Cys Asp Thr Pro Asp Leu Gly Phe Gln Cys Gly Thr
 1 5 10 15

 Ala Ile Ser His Phe Trp Gly Gln Tyr Ser Pro Tyr Phe Ser Val Pro
 20 25 30

 Ser Glu Leu Asp Ala Ser Ile Pro Asp Asp Cys Glu Val Thr Phe Ala
 35 40 45

 Gln Val Leu Ser Arg His Gly Ala Arg Ala Pro Thr Leu Lys Arg Ala
 50 55 60

 Ala Ser Tyr Val Asp Leu Ile Asp Arg Ile His His Gly Ala Ile Ser
 65 70 75 80

 Tyr Gly Pro Gly Tyr Glu Phe Leu Arg Thr Tyr Asp Tyr Thr Leu Gly
 85 90 95

 Ala Asp Glu Leu Thr Arg Thr Gly Gln Gln Gln Met Val Asn Ser Gly
 100 105 110

 Ile Lys Phe Tyr Arg Arg Tyr Arg Ala Leu Ala Arg Lys Ser Ile Pro
 115 120 125

 Phe Val Arg Thr Ala Gly Gln Asp Arg Val Val His Ser Ala Glu Asn
 130 135 140

 Phe Thr Gln Gly Phe His Ser Ala Leu Leu Ala Asp Arg Gly Ser Thr
 145 150 155 160

 Val Arg Pro Thr Leu Pro Tyr Asp Met Val Val Ile Pro Glu Thr Ala
 165 170 175

 Gly Ala Asn Asn Thr Leu His Asn Asp Leu Cys Thr Ala Phe Glu Glu
 180 185 190

 Gly Pro Tyr Ser Thr Ile Gly Asp Asp Ala Gln Asp Thr Tyr Leu Ser
 195 200 205

-continued

Thr Phe Ala Gly Pro Ile Thr Ala Arg Val Asn Ala Asn Leu Pro Gly
 210 215 220
 Ala Asn Leu Thr Asp Ala Asp Thr Val Ala Leu Met Asp Leu Cys Pro
 225 230 235 240
 Phe Glu Thr Val Ala Ser Ser Ser Ser Asp Pro Ala Thr Ala Asp Ala
 245 250 255
 Gly Gly Gly Asn Gly Arg Pro Leu Ser Pro Phe Cys Arg Leu Phe Ser
 260 265 270
 Glu Ser Glu Trp Arg Ala Tyr Asp Tyr Leu Gln Ser Val Gly Lys Trp
 275 280 285
 Tyr Gly Tyr Gly Pro Gly Asn Pro Leu Gly Pro Thr Gln Gly Val Gly
 290 295 300
 Phe Val Asn Glu Leu Leu Ala Arg Leu Ala Gly Val Pro Val Arg Asp
 305 310 315 320
 Gly Thr Ser Thr Asn Arg Thr Leu Asp Gly Asp Pro Arg Thr Phe Pro
 325 330 335
 Leu Gly Arg Pro Leu Tyr Ala Asp Phe Ser His Asp Asn Asp Met Met
 340 345 350
 Gly Val Leu Gly Ala Leu Gly Ala Tyr Asp Gly Val Pro Pro Leu Asp
 355 360 365
 Lys Thr Ala Arg Arg Asp Pro Glu Glu Leu Gly Gly Tyr Ala Ala Ser
 370 375 380
 Trp Ala Val Pro Phe Ala Ala Arg Ile Tyr Val Glu Lys Met Arg Cys
 385 390 395 400
 Ser Gly Gly Gly Gly Gly Gly Gly Gly Gly Glu Gly Arg Gln Glu Lys
 405 410 415
 Asp Glu Glu Met Val Arg Val Leu Val Asn Asp Arg Val Met Thr Leu
 420 425 430
 Lys Gly Cys Gly Ala Asp Glu Arg Gly Met Cys Thr Leu Glu Arg Phe
 435 440 445
 Ile Glu Ser Met Ala Phe Ala Arg Gly Asn Gly Lys Trp Asp Leu Cys
 450 455 460
 Phe Ala
 465

<210> SEQ ID NO 14

<211> LENGTH: 441

<212> TYPE: PRT

<213> ORGANISM: Artificial

<220> FEATURE:

<223> OTHER INFORMATION: Calculated consensus sequence.

<220> FEATURE:

<221> NAME/KEY: MISC_FEATURE

<222> LOCATION: (1)..(441)

<223> OTHER INFORMATION: X is position for which program was not able to determine a consensus residue.

<400> SEQUENCE: 14

Asn Ser His Ser Cys Asp Thr Val Asp Gly Gly Tyr Gln Cys Phe Pro
 1 5 10 15
 Glu Ile Ser His Leu Trp Gly Gln Tyr Ser Pro Tyr Phe Ser Leu Glu
 20 25 30
 Asp Glu Ser Ala Ile Ser Pro Asp Val Pro Asp Asp Cys Xaa Val Thr
 35 40 45
 Phe Val Gln Val Leu Ser Arg His Gly Ala Arg Tyr Pro Thr Ser Ser
 50 55 60

-continued

Lys Xaa Lys Ala Tyr Ser Ala Leu Ile Glu Ala Ile Gln Lys Asn Ala
 65 70 75 80
 Thr Xaa Phe Lys Gly Lys Tyr Ala Phe Leu Lys Thr Tyr Asn Tyr Thr
 85 90 95
 Leu Gly Ala Asp Asp Leu Thr Pro Phe Gly Glu Asn Gln Met Val Asn
 100 105 110
 Ser Gly Ile Lys Phe Tyr Arg Arg Tyr Lys Ala Leu Ala Arg Lys Xaa
 115 120 125
 Val Pro Phe Val Arg Ala Ser Gly Ser Asp Arg Val Ile Ala Ser Ala
 130 135 140
 Glu Lys Phe Ile Glu Gly Phe Gln Ser Ala Lys Leu Ala Asp Pro Gly
 145 150 155 160
 Ser Xaa Pro His Gln Ala Ser Pro Val Ile Asn Val Ile Ile Pro Glu
 165 170 175
 Gly Ser Gly Tyr Asn Asn Thr Leu Asp His Gly Thr Cys Thr Ala Phe
 180 185 190
 Glu Asp Ser Glu Leu Gly Asp Asp Ala Glu Ala Asn Phe Thr Ala Thr
 195 200 205
 Phe Ala Pro Ala Ile Arg Ala Arg Leu Glu Ala Asp Leu Pro Gly Val
 210 215 220
 Thr Leu Thr Asp Glu Asp Val Val Xaa Leu Met Asp Met Cys Pro Phe
 225 230 235 240
 Glu Thr Val Ala Arg Thr Ser Asp Ala Thr Glu Leu Ser Pro Phe Cys
 245 250 255
 Ala Leu Phe Thr Glu Xaa Glu Trp Xaa Xaa Tyr Asp Tyr Leu Gln Ser
 260 265 270
 Leu Gly Lys Tyr Tyr Gly Tyr Gly Ala Gly Asn Pro Leu Gly Pro Ala
 275 280 285
 Gln Gly Val Gly Phe Xaa Asn Glu Leu Ile Ala Arg Leu Thr His Ser
 290 295 300
 Pro Val Gln Asp His Thr Ser Thr Asn His Thr Leu Asp Ser Asn Pro
 305 310 315 320
 Ala Thr Phe Pro Leu Asn Ala Thr Leu Tyr Ala Asp Phe Ser His Asp
 325 330 335
 Asn Ser Met Ile Ser Ile Phe Phe Ala Leu Gly Leu Tyr Asn Gly Thr
 340 345 350
 Ala Pro Leu Ser Thr Thr Ser Val Glu Ser Ile Glu Glu Thr Asp Gly
 355 360 365
 Tyr Ala Ala Ser Trp Thr Val Pro Phe Gly Ala Arg Ala Tyr Val Glu
 370 375 380
 Met Met Gln Cys Gln Ala Glu Lys Glu Pro Leu Val Arg Val Leu Val
 385 390 395 400
 Asn Asp Arg Val Val Pro Leu His Gly Cys Ala Val Asp Lys Leu Gly
 405 410 415
 Arg Cys Lys Leu Asp Asp Phe Val Glu Gly Leu Ser Phe Ala Arg Ser
 420 425 430
 Gly Gly Asn Trp Ala Glu Cys Phe Ala
 435 440

<210> SEQ ID NO 15

<211> LENGTH: 441

<212> TYPE: PRT

<213> ORGANISM: Artificial

<220> FEATURE:

<223> OTHER INFORMATION: Constructed consensus phytase sequence.

-continued

<400> SEQUENCE: 15

Asn Ser His Ser Cys Asp Thr Val Asp Gly Gly Tyr Gln Cys Phe Pro
 1 5 10 15
 Glu Ile Ser His Leu Trp Gly Gln Tyr Ser Pro Tyr Phe Ser Leu Glu
 20 25 30
 Asp Glu Ser Ala Ile Ser Pro Asp Val Pro Asp Asp Cys Arg Val Thr
 35 40 45
 Phe Val Gln Val Leu Ser Arg His Gly Ala Arg Tyr Pro Thr Ser Ser
 50 55 60
 Lys Ser Lys Ala Tyr Ser Ala Leu Ile Glu Ala Ile Gln Lys Asn Ala
 65 70 75 80
 Thr Ala Phe Lys Gly Lys Tyr Ala Phe Leu Lys Thr Tyr Asn Tyr Thr
 85 90 95
 Leu Gly Ala Asp Asp Leu Thr Pro Phe Gly Glu Asn Gln Met Val Asn
 100 105 110
 Ser Gly Ile Lys Phe Tyr Arg Arg Tyr Lys Ala Leu Ala Arg Lys Ile
 115 120 125
 Val Pro Phe Ile Arg Ala Ser Gly Ser Asp Arg Val Ile Ala Ser Ala
 130 135 140
 Glu Lys Phe Ile Glu Gly Phe Gln Ser Ala Lys Leu Ala Asp Pro Gly
 145 150 155 160
 Ser Gln Pro His Gln Ala Ser Pro Val Ile Asp Val Ile Ile Pro Glu
 165 170 175
 Gly Ser Gly Tyr Asn Asn Thr Leu Asp His Gly Thr Cys Thr Ala Phe
 180 185 190
 Glu Asp Ser Glu Leu Gly Asp Asp Val Glu Ala Asn Phe Thr Ala Leu
 195 200 205
 Phe Ala Pro Ala Ile Arg Ala Arg Leu Glu Ala Asp Leu Pro Gly Val
 210 215 220
 Thr Leu Thr Asp Glu Asp Val Val Tyr Leu Met Asp Met Cys Pro Phe
 225 230 235 240
 Glu Thr Val Ala Arg Thr Ser Asp Ala Thr Glu Leu Ser Pro Phe Cys
 245 250 255
 Ala Leu Phe Thr His Asp Glu Trp Arg Gln Tyr Asp Tyr Leu Gln Ser
 260 265 270
 Leu Gly Lys Tyr Tyr Gly Tyr Gly Ala Gly Asn Pro Leu Gly Pro Ala
 275 280 285
 Gln Gly Val Gly Phe Ala Asn Glu Leu Ile Ala Arg Leu Thr Arg Ser
 290 295 300
 Pro Val Gln Asp His Thr Ser Thr Asn His Thr Leu Asp Ser Asn Pro
 305 310 315 320
 Ala Thr Phe Pro Leu Asn Ala Thr Leu Tyr Ala Asp Phe Ser His Asp
 325 330 335
 Asn Ser Met Ile Ser Ile Phe Phe Ala Leu Gly Leu Tyr Asn Gly Thr
 340 345 350
 Ala Pro Leu Ser Thr Thr Ser Val Glu Ser Ile Glu Glu Thr Asp Gly
 355 360 365
 Tyr Ser Ala Ser Trp Thr Val Pro Phe Gly Ala Arg Ala Tyr Val Glu
 370 375 380
 Met Met Gln Cys Gln Ala Glu Lys Glu Pro Leu Val Arg Val Leu Val
 385 390 395 400
 Asn Asp Arg Val Val Pro Leu His Gly Cys Ala Val Asp Lys Leu Gly

-continued

405	410	415	
Arg Cys Lys Arg Asp Asp Phe Val Glu Gly Leu Ser Phe Ala Arg Ser			
420	425	430	
Gly Gly Asn Trp Ala Glu Cys Phe Ala			
435	440		

<210> SEQ ID NO 16
 <211> LENGTH: 1426
 <212> TYPE: DNA
 <213> ORGANISM: Artificial
 <220> FEATURE:
 <223> OTHER INFORMATION: DNA sequence of the fungal consensus gene (fcp).

<400> SEQUENCE: 16

tatatgaatt catgggcgtg ttcgctgtgc tactgtccat tgccaccttg ttcggttcca	60
catccggtac cgccttgggt cctcgtggta attctcactc ttgtgacact gttgacgggtg	120
gttaccaatg tttcccagaa atttctcact tgtgggggtca atactctcca tactttctctt	180
tggaagacga atctgctatt tctccagacg ttccagacga ctgtagagtt actttcgttc	240
aagttttgtc tagacacggg gctagatacc caacttcttc taagtctaag gcttactctg	300
ctttgattga agctattcaa aagaacgcta ctgctttcaa gggtaagtac gctttcttga	360
agacttacia ctacactttg ggtgctgacg acttgactcc attcgggtgaa aaccaaattg	420
ttaactctgg tattaagttc tacagaagat acaaggcttt ggctagaaag attggtccat	480
tcattagagc ttctggttct gacagagtta ttgcttctgc tgaaaagttc attgaaggtt	540
tccaatctgc taagttggct gaccaggtt ctcaaccaca ccaagcttct ccagttattg	600
acgttattat tccagaagga tccggttaca acaacacttt ggaccacggg acttgtactg	660
ctttcgaaga ctctgaattg ggtgacgacg ttgaagctaa cttcactgct ttgttcgctc	720
cagctattag agctagattg gaagctgact tgccaggtgt tactttgact gacgaagacg	780
ttgtttactt gatggacatg tgtccattcg aaactgttgc tagaacttct gacgctactg	840
aattgtctcc attctgtgct ttgttctc acgacgaatg gagacaatac gactacttgc	900
aatctttggg taagtactac ggttacgggt ctggttaacc attgggtcca gctcaagggtg	960
ttggtttcgc taacgaattg attgctagat tgactagatc tccagttcaa gaccacactt	1020
ctactaacca cactttggac tctaaccag ctactttccc attgaacgct actttgtacg	1080
ctgacttctc tcacgacaac tctatgattt ctattttctt cgctttgggt ttgtacaacg	1140
gtactgctcc attgtctact acttctgttg aatctattga agaaactgac gggtactctg	1200
cttcttggac tgttccattc ggtgctagag cttacgttga aatgatgcaa tgtcaagctg	1260
aaaaggaacc attggttaga gttttgggta acgacagagt tgttccattg cacggttgtg	1320
ctgttgacaa gttgggtaga tgtaagagag acgacttctg tgaaggtttg tctttcgcta	1380
gatctggtgg taactgggct gaatgtttcg cttagaatt catata	1426

<210> SEQ ID NO 17
 <211> LENGTH: 467
 <212> TYPE: PRT
 <213> ORGANISM: Artificial
 <220> FEATURE:
 <223> OTHER INFORMATION: Constructed consensus phytase sequence with signal peptide of phytase from *A. terreus* cbs fused to N-terminus.

<400> SEQUENCE: 17

Met Gly Val Phe Val Val Leu Leu Ser Ile Ala Thr Leu Phe Gly Ser

-continued

1	5	10	15
Thr Ser Gly	Thr Ala Leu Gly	Pro Arg Gly Asn Ser His	Ser Cys Asp
	20	25	30
Thr Val Asp	Gly Gly Tyr Gln Cys Phe	Pro Glu Ile Ser His Leu Trp	
	35	40	45
Gly Gln Tyr	Ser Pro Tyr Phe Ser Leu Glu Asp	Glu Ser Ala Ile Ser	
	50	55	60
Pro Asp Val	Pro Asp Asp Cys Arg Val Thr Phe Val Gln Val Leu Ser		
	65	70	75
Arg His Gly	Ala Arg Tyr Pro Thr Ser Ser Lys Ser Lys Ala Tyr Ser		
	85	90	95
Ala Leu Ile	Glu Ala Ile Gln Lys Asn Ala Thr Ala Phe Lys Gly Lys		
	100	105	110
Tyr Ala Phe	Leu Lys Thr Tyr Asn Tyr Thr Leu Gly Ala Asp Asp Leu		
	115	120	125
Thr Pro Phe	Gly Glu Asn Gln Met Val Asn Ser Gly Ile Lys Phe Tyr		
	130	135	140
Arg Arg Tyr	Lys Ala Leu Ala Arg Lys Ile Val Pro Phe Ile Arg Ala		
	145	150	155
Ser Gly Ser	Asp Arg Val Ile Ala Ser Ala Glu Lys Phe Ile Glu Gly		
	165	170	175
Phe Gln Ser	Ala Lys Leu Ala Asp Pro Gly Ser Gln Pro His Gln Ala		
	180	185	190
Ser Pro Val	Ile Asp Val Ile Ile Pro Glu Gly Ser Gly Tyr Asn Asn		
	195	200	205
Thr Leu Asp	His Gly Thr Cys Thr Ala Phe Glu Asp Ser Glu Leu Gly		
	210	215	220
Asp Asp Val	Glu Ala Asn Phe Thr Ala Leu Phe Ala Pro Ala Ile Arg		
	225	230	235
Ala Arg Leu	Glu Ala Asp Leu Pro Gly Val Thr Leu Thr Asp Glu Asp		
	245	250	255
Val Val Tyr	Leu Met Asp Met Cys Pro Phe Glu Thr Val Ala Arg Thr		
	260	265	270
Ser Asp Ala	Thr Glu Leu Ser Pro Phe Cys Ala Leu Phe Thr His Asp		
	275	280	285
Glu Trp Arg	Gln Tyr Asp Tyr Leu Gln Ser Leu Gly Lys Tyr Tyr Gly		
	290	295	300
Tyr Gly Ala	Gly Asn Pro Leu Gly Pro Ala Gln Gly Val Gly Phe Ala		
	305	310	315
Asn Glu Leu	Ile Ala Arg Leu Thr Arg Ser Pro Val Gln Asp His Thr		
	325	330	335
Ser Thr Asn	His Thr Leu Asp Ser Asn Pro Ala Thr Phe Pro Leu Asn		
	340	345	350
Ala Thr Leu	Tyr Ala Asp Phe Ser His Asp Asn Ser Met Ile Ser Ile		
	355	360	365
Phe Phe Ala	Leu Gly Leu Tyr Asn Gly Thr Ala Pro Leu Ser Thr Thr		
	370	375	380
Ser Val Glu	Ser Ile Glu Glu Thr Asp Gly Tyr Ser Ala Ser Trp Thr		
	385	390	395
Val Pro Phe	Gly Ala Arg Ala Tyr Val Glu Met Met Gln Cys Gln Ala		
	405	410	415
Glu Lys Glu	Pro Leu Val Arg Val Leu Val Asn Asp Arg Val Val Pro		
	420	425	430

-continued

Leu His Gly Cys Ala Val Asp Lys Leu Gly Arg Cys Lys Arg Asp Asp
 435 440 445

Phe Val Glu Gly Leu Ser Phe Ala Arg Ser Gly Gly Asn Trp Ala Glu
 450 455 460

Cys Phe Ala
 465

<210> SEQ ID NO 18
 <211> LENGTH: 26
 <212> TYPE: DNA
 <213> ORGANISM: Artificial
 <220> FEATURE:
 <223> OTHER INFORMATION: PCR primer.

<400> SEQUENCE: 18

tatatgaatt catggcgtg ttcgtc 26

<210> SEQ ID NO 19
 <211> LENGTH: 22
 <212> TYPE: DNA
 <213> ORGANISM: Artificial
 <220> FEATURE:
 <223> OTHER INFORMATION: PCR primer.

<400> SEQUENCE: 19

tgaaaagttc attgaagtt tc 22

<210> SEQ ID NO 20
 <211> LENGTH: 22
 <212> TYPE: DNA
 <213> ORGANISM: Artificial
 <220> FEATURE:
 <223> OTHER INFORMATION: PCR primer.

<400> SEQUENCE: 20

tcttcgaaag cagtacaagt ac 22

<210> SEQ ID NO 21
 <211> LENGTH: 22
 <212> TYPE: DNA
 <213> ORGANISM: Artificial
 <220> FEATURE:
 <223> OTHER INFORMATION: PCR primer.

<400> SEQUENCE: 21

tatatgaatt cttagcgaa ac 22

<210> SEQ ID NO 22
 <211> LENGTH: 33
 <212> TYPE: DNA
 <213> ORGANISM: Artificial
 <220> FEATURE:
 <223> OTHER INFORMATION: PCR primer.

<400> SEQUENCE: 22

tatatcatga gcgtgttcgt cgtgctactg ttc 33

<210> SEQ ID NO 23
 <211> LENGTH: 33
 <212> TYPE: DNA
 <213> ORGANISM: Artificial
 <220> FEATURE:
 <223> OTHER INFORMATION: PCR primer.

<400> SEQUENCE: 23

-continued

acccgactta caaagcgaat tctatagata tat 33

<210> SEQ ID NO 24
<211> LENGTH: 32
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Primer for site-directed mutagenesis.

<400> SEQUENCE: 24

cacttgtaggg gttgtacag tccatacttc tc 32

<210> SEQ ID NO 25
<211> LENGTH: 32
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Primer for site-directed mutagenesis.

<400> SEQUENCE: 25

gagaagtatg gactgtacaa accccacaag tg 32

<210> SEQ ID NO 26
<211> LENGTH: 32
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Primer for site-directed mutagenesis.

<400> SEQUENCE: 26

cacttgtaggg gtacctactc tccatacttc tc 32

<210> SEQ ID NO 27
<211> LENGTH: 32
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Primer for site-directed mutagenesis.

<400> SEQUENCE: 27

gagaagtatg gagagtaggt accccacaag tg 32

<210> SEQ ID NO 28
<211> LENGTH: 32
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Primer for site-directed mutagenesis.

<400> SEQUENCE: 28

cacttgtaggg gtggttactc tccatacttc tc 32

<210> SEQ ID NO 29
<211> LENGTH: 32
<212> TYPE: DNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: Primer for site-directed mutagenesis.

<400> SEQUENCE: 29

gagaagtatg gagagtaacc accccacaag tg 32

<210> SEQ ID NO 30
<211> LENGTH: 32
<212> TYPE: DNA

-continued

<213> ORGANISM: Artificial
 <220> FEATURE:
 <223> OTHER INFORMATION: Primer for site-directed mutagenesis.

<400> SEQUENCE: 30

cacttgtagg gtaccaactc tccatacttc tc 32

<210> SEQ ID NO 31
 <211> LENGTH: 32
 <212> TYPE: DNA
 <213> ORGANISM: Artificial
 <220> FEATURE:
 <223> OTHER INFORMATION: Primer for site-directed mutagenesis.

<400> SEQUENCE: 31

gagaagtatg gagagttggt accccacaag tg 32

<210> SEQ ID NO 32
 <211> LENGTH: 32
 <212> TYPE: DNA
 <213> ORGANISM: Artificial
 <220> FEATURE:
 <223> OTHER INFORMATION: Primer for site-directed mutagenesis.

<400> SEQUENCE: 32

cacttgtagg gtctcaactc tccatacttc tc 32

<210> SEQ ID NO 33
 <211> LENGTH: 32
 <212> TYPE: DNA
 <213> ORGANISM: Artificial
 <220> FEATURE:
 <223> OTHER INFORMATION: Primer for site-directed mutagenesis.

<400> SEQUENCE: 33

gagaagtatg gagagttgag accccacaag tg 32

What is claimed is:

1. A *purified* polynucleotide encoding a consensus protein of [SEQ ID NO:2] SEQ ID NO:17.

2. A *purified* polynucleotide encoding a consensus protein of [SEQ ID NO:1] SEQ ID NO:15.

3. A *purified* polynucleotide which encodes a consensus protein having the amino acid sequence of [SEQ ID NO:2] SEQ ID NO:17 except that Q at position 50 has been replaced by L, T, or G.

40

4. A *purified* polynucleotide which encodes a consensus protein having the amino acid sequence of [SEQ ID NO:2] SEQ ID NO: 17 except that Q at position 50 has been replaced by T and Y at position 51 has been replaced by N.

45

5. A *purified* polynucleotide which encodes a consensus protein having the amino acid sequence of [SEQ ID NO:2] SEQ ID NO:17 except that Q at position 50 has been replaced by L and Y at position 51 has been replaced by N.

* * * * *