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(54) **MYOCARDIAL GRAFTS AND CELLULAR COMPOSITIONS**

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(56) **References Cited**

U.S. PATENT DOCUMENTS

4,419,446 A	12/1983	Howley et al.	
4,518,584 A	5/1985	Mark et al.	
5,145,676 A	9/1992	Fahey, III et al.	
5,215,895 A	6/1993	Bennett et al.	
5,278,287 A	1/1994	Rollins et al.	
5,306,709 A	4/1994	Gewirtz	
5,346,686 A	9/1994	Lyle et al.	
6,015,671 A *	1/2000	Field	435/6

OTHER PUBLICATIONS

Acsadi, G. et al., "Direct Gene Transfer and Expression into Rat Heart in Vivo", *The New Biologist*, vol. 3, No. 1, 1991, pp. 71-81.

Brunner, A. et al., "Site-Directed Mutagenesis of Cysteine Residues in the Pro Region of the Transforming Growth Factor β 1 Precursor", *J. Biol. Chem.* vol. 264, No. 23, Aug. 15, 1989, pp. 13660-13664.

Delcarpio, J. et al., "Morphological Characterization of Cardiomyocytes Isolated from a Transplantable Cardiac Tumor Derived from Transgenic Mouse Atria (AT-1 Cells)", *Circulation Research*, vol. 69, No. 6, Dec. 1991, pp. 1591-1600.

Dhawan, J. et al., "systemic Delivery of Human Growth Hormone by Injection of Genetically Engineered Myoblasts", *Science*, vol. 254, 1991, pp. 1509-1512.

Fields, L. "Atrial Natriuretic Factor-SV40 T Antigen Transgenes Produce Tumors and Cardiac Arrhythmias in Mice", *Science*, vol. 239, Feb. 26, 1988, pp. 1029-1033.

Gussoni, E. et al., "Normal Dystrophin Transcripts Detected in Duchenne Muscular Dystrophy Patients after Myoblast Transplantation", *Nature*, vol. 356, Apr. 2, 1992, pp. 435-438.

Jacobson, S. et al., "Evidence for Functional Sodium and Calcium Ion Channels in the Membrane of Cultured Cardiomyocytes of the Adult Rat", *Canadian Journal of Physiology*, vol. 61, 1983, pp. 1312-1316.

Janse, M. et al., "The Border Zone in Myocardial Ischemia. An Electrophysiological, Metabolic and Histochemical Correlation in the Pig Heart", *Circulation Research*, vol. 44, 1979, pp. 576-588.

Jiao, S. et al., "Long-Term Correction of Rat Model of Parkinson's Disease by Gene Therapy", *Nature*, vol. 362, Apr. 1, 1993, pp. 450-453.

Katz, E. et al., "Cardiomyocyte Proliferation in Mice Expressing α -Cardiac Myosin Heavy Chain-SV40 T-Antigen Transgenes", *Amer. J. Phys.*, vol. 262 (Heart and Circulatory Physiology 31), 1992, pp. 1867-1876.

Kitsis, R. et al., "Hormonal Modulation of a Gene Injected into a Rat Heart in vivo", *Proc.Natl.Acad.Sci.USA*, vol. 88, May 1991, pp. 4138-4142.

Koh, G. et al., "Long-Term Survival of AT-1 Cardiomyocyte Grafts in Syngeneic Myocardium", *Amer. Physiol. So.*, 1993, pp. 1727-1733.

Koh, G. et al., "Differentiation and Long-Term Survival of C2C12 Myoblast Grafts in Heart", *J. Clin. Invest.*, vol. 92, Sep. 1993.

Lallemant, Y. and Brulet, P., "An in situ Assessment of the Routes of Extents of Colonisation of the Mouse Embryo by Embryonic Stem Cells and Their Descendants", *Development*, vol. 110, 1990, pp. 1241-1248.

Loewy, A. et al., " β -Galactosidase Expressing Recombinant Pseudorabies Virus for Light and Electron Microscopic Study of Transneuronally Labeled CNS Neurons", *Brain Research* 555, 1991, pp. 346-352.

MacDonald, R. et al., "Transgenic Progeny Inherit Tissue-Specific Expression of Rat Elastase I Genes", *DNA*, vol. 5, No. 5, 1986, pp. 393-401.

Nadal-Ginard, B., "Commitment, Fusion and Biochemical Differentiation of a Myogenic Cell Line in the Absence of DNA Synthesis", *Cell*, vol. 15, Nov. 1978, pp. 855-864.

Rockman, H. et al., "Segregation of Atrial-Specific and Inducible Expression of an ANF Transgene in an in vivo Murine Model of Cardiac Hypertrophy", *Proc.Natl. Acad. Sci.USA*, vol. 88, Sep. 1991, pp. 8277-8281.

Samuel, S. et al., "Autocrine Induction of Tumor Protease Production and Invasion by a Metallothionein-Regulated TGF- β 1", *EMBO Journal*, vol. 11, No. 4, 1992, pp. 1599-1605.

(List continued on next page.)

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(57) **ABSTRACT**

Described are preferred myocardial grafts of skeletal myoblasts or cardiomyocytes, and cellular compositions and methods useful in obtaining the grafts. The myocardial grafts are stable and can be used, for example, to deliver recombinant proteins directly to the heart.

13 Claims, 1 Drawing Sheet

OTHER PUBLICATIONS

Spear, J. et al., "Cellular Electrophysiologic Characteristics of Chronically Infarcted Myocardium in Dogs Susceptible to Sustained Ventricular Tachyarrhythmias", *J Am Coll Cardiol*, 1983:I(4), pp. 1099-1110.

Steinhelper, M. et al., "Hypotension in Transgenic Mice Expressing Atrial Natriuretic Factor Fusion Genes", *Hypertension*, vol. 16, No. 3, 1990, pp. 301-307.

Steinhelper, M. et al., Cardiac Tumors and Dysrhythmias in Transgenic Mice, *Toxicologic Pathology* 18, 1990, pp. 464-469.

Steinhelper, M. et al., "Proliferation in Vivo and in Culture of Differentiated Adult Atrial Cardiomyocytes from Transgenic Mice", *Amer. J. Phys.* 259 (Heart and Circulatory Physiology 28): 1990, pp. 1826-1834.

Thompson, L., "Fetal Transplants Show Promise", *Science*, vol. 257, 1992, pp. 868-870.

Yaffe, D. et al., "Serial Passaging and Differentiation of Myogenic Cells Isolated from Dystrophic Mouse Muscle", *Nature*, vol. 270, 1977, pp. 725-727.

Shen et al., *Int. J. Dev. Biol.*, vol. 36, 1992, pp. 465-476.*

* cited by examiner

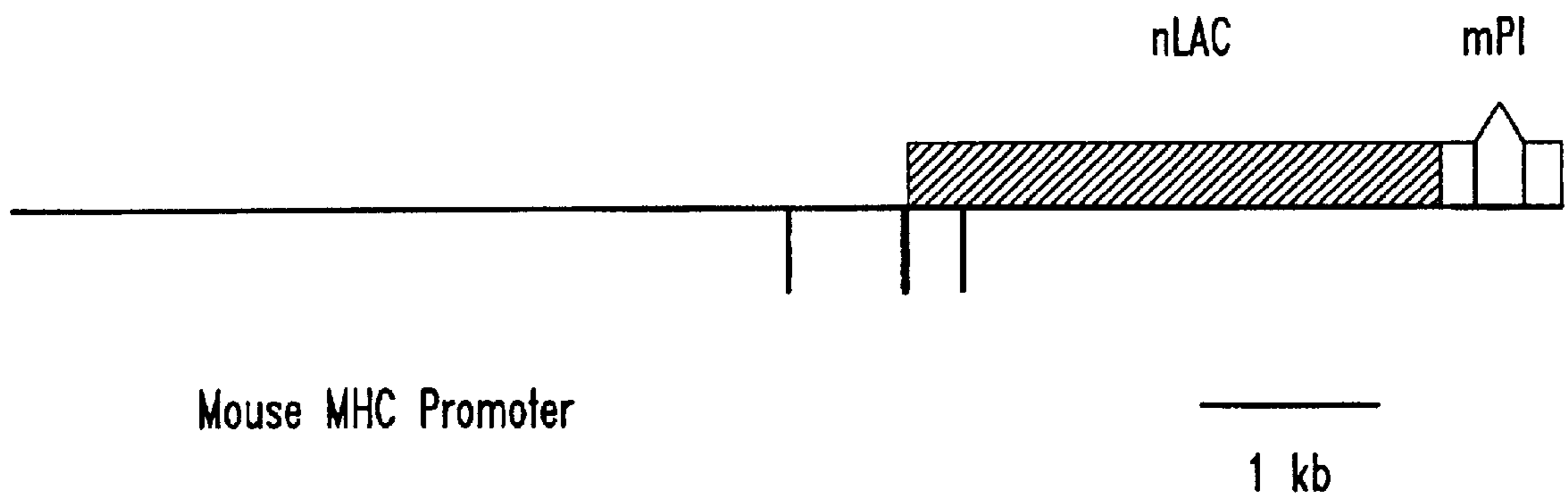


Fig. 1

MYOCARDIAL GRAFTS AND CELLULAR COMPOSITIONS

Matter enclosed in heavy brackets [] appears in the original patent but forms no part of this reissue specification; matter printed in italics indicates the additions made by reissue.

[This application is a continuation of application Ser. No. 08/477,783, filed Jun. 7, 1995, now U.S. Pat. No. 5,733,767.]

REFERENCE TO RELATED APPLICATIONS

This is a continuation of U.S. patent application Ser. No. 08/477,783 filed Jun. 7, 1995, now issued as U.S. Pat. No. 5,733,727, which is a divisional of U.S. patent application Ser. No. 08/153,664 filed Nov. 16, 1993, now issued as U.S. Pat. No. 5,602,301.

This invention was made with government support under grant number HL45453 from the National Institute of Health. The government has certain rights in the invention.

BACKGROUND OF THE INVENTION

The present invention resides generally in the field of cardiology, and more particularly relates to stable myocardial grafts and methods and cellular compositions useful for achieving such grafts.

As further background, organ transplantation has been widely used to replace diseased, nonfunctional tissue. More recently, cellular transplantation to augment deficiencies in host tissue function has emerged as a potential therapeutic paradigm. One example of this approach is the well publicized use of fetal tissue in individuals with Parkinsonism (reviewed in (1), see reference list, *infra.*), where dopamine secretion from transplanted cells alleviates the deficiency in patients. In other studies, transplanted myoblasts from-
unaffected siblings fused with endogenous myotubes in Duchenne's patients; importantly the grafted myotubes expressed wild-type dystrophin (2).

Despite their relevance in other areas, there earlier studies do not describe any cellular transplantation technology which can be successfully applied to the heart, where the ability to replace damaged myocardium would have obvious clinical relevance. Additionally, the use of intra-cardiac grafts to target the long-term expression of angiogenic factors and ionotropic peptides would be of therapeutic value for individuals with myocardial ischemia or congestive heart failure, respectively.

In light of this background there is a need for the development of cellular transplantation technology in the heart. Desirably, such technology would not only provide stable grafts in the heart but also enable the delivery of useful recombinant proteins or other molecules directly to the heart. The present invention addresses these needs.

SUMMARY OF THE INVENTION

The applicant has established cellular grafts in the myocardium which are viable long-term. Cardiomyocytes and skeletal myoblasts have been grafted directly into the myocardium of syngeneic animals. Viable grafts were detected at least one-half year post-implantation (the latest time point assayed). The presence of the grafts was not accompanied by overt cardiac arrhythmia, and the majority of the grafts were juxtaposed directly to the host myocardium and not encapsulated. It has thus been discovered that the myocardium can serve as a stable platform for cellular transplants. These

transplants can be used for the local delivery of recombinant molecules to the heart and/or for replacing diseased tissue to supplement myocardial function.

Accordingly, one preferred embodiment of the invention provides a myocardial graft in an animal which includes a stable graft of skeletal myoblasts or cardiomyocytes incorporated in myocardial tissue of the animal.

Another preferred embodiment of the invention provides a method for forming a stable myocardial graft in an animal. The inventive method includes the step of introducing skeletal myoblasts or cardiomyocytes in myocardial tissue of the animal so as to form a stable myocardial graft. The cells can be conveniently introduced, for example, by injection.

Another preferred embodiment of the invention provides a method for delivering a recombinant molecule to myocardial tissue of an animal. This method includes the step of establishing a stable graft of skeletal myoblasts or cardiomyocytes incorporated in myocardial tissue of the animal, wherein the myoblasts or cardiomyocytes deliver the recombinant molecule to the myocardial tissue. In this embodiment the myoblasts or cardiomyocytes will carry transgenes encoding the recombinant molecule.

Another preferred embodiment of the invention provides a cellular composition comprising a substantially homogeneous population of non-immortalized cardiomyocytes. This and other cell populations can be obtained utilizing a preferred inventive method that includes (i) transfecting embryonic stem cells to introduce a marker gene enabling selection of one cell lineage from other cell lineages resulting from differentiation of the stem cells, (iii) causing the stem cells to differentiate, and (iv) selecting said one cell lineage based on the marker gene. The cells used in and resulting from such methods also form a part of the present invention.

Still another preferred embodiment of the invention provides a nonhuman animal having a stable graft of skeletal myoblasts or cardiomyocytes incorporated in myocardial tissue of the animal.

The invention thus provides myocardial grafts, methods and cellular compositions useful for forming myocardial grafts, and animals which have the myocardial grafts. The grafts will find use both as a vehicle for delivering therapeutic substances such as recombinant proteins and other molecules, and as a means for replacing diseased tissue to supplement myocardial function. Cellular compositions of the invention can be used directly to prepare grafts, and will also be useful in screening drug substance effects on cardiomyocytes and for expressing and obtaining recombinant proteins. Grafted animals can be used, for example, to screen the effects of recombinant molecules on the heart.

These and other objects and advantages of the invention will be apparent from the following description.

BRIEF DESCRIPTION OF THE FIGURE

FIG. 1 is a schematic diagram illustrating DNA used to generate MHC-nLAC transgenic mice in Example 3, *infra.*

DESCRIPTION OF THE PREFERRED EMBODIMENT

For the purpose of promoting an understanding of the principles of the invention, reference will now be made to certain embodiments thereof and specific language will be used to describe the same. It will nevertheless be understood that no limitation of the scope of the invention is thereby intended, such alterations, further modifications and

applications of the principles of the invention as illustrated herein being contemplated as would normally occur to one skilled in the art to which the invention relates.

As indicated above, the present invention provides stable myocardial grafts of skeletal myoblasts and/or cardiomyocytes. In this regard, as used herein the term "stable myocardial graft" is intended to mean a myocardial graft whose cells are viable for a period of at least about 2 weeks. Surprisingly, such stable grafts have been readily achieved in accordance with the invention, with preferred grafts having cells viable for six months or more. Myocardial grafts of the invention can thus provide for long-term delivery of recombinant proteins or other molecules to the heart and/or long-term supplementation of myocardial tissue.

The skeletal myoblasts and cardiomyocytes used in the invention can be obtained or isolated from any suitable source. Skeletal myoblasts, including for example C2C12 skeletal myoblasts, are available from public depositories such as the American Type Culture Collection (ATCC) (Rockville, Md.). Skeletal myoblasts can also be isolated from skeletal muscle using techniques well known to the art and literature. Cardiomyocytes useful for the invention can be obtained using techniques described in the literature (3) or using methods described more particularly in the Examples below. Briefly, one such method involves digestion of heart tissues to obtain cardiomyocytes.

Another method involves using an appropriate marker to select specific cell lineages, such as cardiomyocytes, from other cell lineages resulting from the differentiation of embryonic stem cells (totipotent cell lines derived from the inner cell mass of blastocysts as described in (22)). The preferred method involves a positive selection scheme. Thus, a marker gene, such as a gene conferring antibiotic resistance (e.g. neomycin or hygromycin), is introduced into the stem cells under appropriate control such that expression of the gene occurs only in the desired cell lineage. For example, the marker gene can be under the control of a promoter which is active only in the desired cell lineage. Upon differentiation of the stem cells, the desired lineage is then selected based upon the marker, e.g. by contacting the mixed differentiated cells with the appropriate antibiotic to which the desired lineage has been conferred resistance. Cell lines other than the desired line will thus be killed, and a substantially pure, homogeneous population of the desired line can be recovered. In more preferred methods, two markers are introduced into the parent stem cells, one allowing selection of transfected stem cells from non-transfected cells, and one allowing selection of the desired cell lineage from other lineages. A double positive selection scheme can thus be used where each selectable marker confers antibiotic resistance. Using this selection methodology, populations comprised about 90% and even about 95–100% of the desired cell lineage can be obtained, as demonstrated in the Examples below.

To obtain grafts of the invention, the skeletal myoblasts or cardiomyocytes will be introduced into the myocardial tissue of a living animal such as a mammal. The cells can be introduced in any suitable manner, but it is preferred that the mode of introduction be as non-invasive as possible. Thus, delivery of the cells by injection, catheterization or similar means will be more desired.

The resulting graft-bearing animals have exhibited normal sinus rhythms, indicating that the graft per se, as well as the graft-host myocardium border zone, does not induce arrhythmias. This is in stark contrast to the remodeling that

frequently occurs following infarcts in humans; the border zone of the infarct may give rise to circus loops which result in clinically significant arrhythmias (4, 5).

Grafts of the invention can be proliferative or non-proliferative. For example, the AT-1 grafts established in the specific Examples below are proliferative. On the other hand, the skeletal myoblast-derived grafts formed in the Examples are non-proliferative, with the absence of tritiated thymidine uptake demonstrating that the formation of stable intra-cardiac grafts was not dependent upon sustained cell proliferation.

Preferred grafts will be characterized by the presence of direct intracellular coupling and the formation of gap junctions between host and grafted cells. Moreover, such grafts will not cause immune response in the host, and will exhibit terminal differentiation of grafted cells and a non-tumorigenic nature.

Grafts of the invention are useful inter alia to deliver therapeutic proteins and the like via secretion from grafted cells, and to replace diseased or damaged tissue to supplement myocardial function. As examples of therapeutic protein deliveries, grafts may express angiogenic factors (as exemplified by basic and acidic Fibroblast Growth Factor; Transforming Growth Factor-Beta. Vascular Endothelial Growth Factor and Hepatocyte Growth Factor) to induce neovascularization. Similarly, grafts expressing neurotrophic agents near an infarcted region may be used to ameliorate the arrhythmogenesis associated with the border zone. These and many other candidate substances for targeted delivery to the heart will be apparent to those skilled in the area.

To promote a further understanding of the invention and its principles and advantages, the following specific Examples are provided. It will be understood that these Examples are illustrative, and not limiting, in nature.

EXAMPLE 1

Generation of Stable AT-1 Cardiomyocyte Grafts

A. Method

AT-1 Cell Culture and Myocardial Grafting Protocol

AT-1 cardiomyocytes were isolated from subcutaneous tumors by sequential collagenase digestion and cultured in PC-1 medium (Ventrex, Coon Rapids Minn.) containing 10% fetal calf serum as previously described in (6). Cells were labeled with 10 μ M 8-chloromethyl-4,4-difluoro-1,3,5,7,-tetramethyl-4-bora-3a,4a-diazaindecene (BODIPY, Molecular Probes, Eugene Oreg.) for 30 min at 37° C. to facilitate localization of the injection site. Immediately before injection, cells were harvested with trypsin and collagenase, washed three times with serum-free PC-1 medium and directly injected into the ventricular myocardium of syngeneic B6D2/F1 mice (Jackson Laboratories, Bar Harbor Mass.) under open heart surgery as described in (7). Cells (4–10 \times 10) were injected in a volume of 2–3 μ l using a plastic syringe fitted with a 30 gauge needle.

Histology

Hearts were removed following cervical dislocation and cryoprotected in 30% sucrose, embedded and sectioned at 10 μ m with a cryomicrotome as described (8). For hematoxylin and eosin (H and E) staining, sections were post fixed in acetone:methanol (1:1) and stained according to manufacturer's specifications (Sigma Diagnostics, St. Louis Mo.). For immuno-histology, unfixed sections were reacted with polyclonal rabbit anti-T-Ag antibodies (either 161-T, see (3) or 162-T) followed by horseradish peroxidase-conjugated goat anti-rabbit antisera (Boehringer Mannheim,

Indianapolis Ind.), and visualized by diaminobenzidine reaction with nickel enhancement as described in (9). Monoclonal antibodies against the common leukocyte antigen (CD45; antibody M1/9.3HL, Boehringer Mannheim) and against the macrophage Mac-1 antigen (CD11b; antibody M1/70HL, Boehringer Mannheim) were used to monitor intra-cardiac graft rejection. The Mac-1 antibody has 75–90% cross reactivity with lymphocytes. After treatment with primary antibody, sections were incubated with horseradish peroxidase-conjugated rabbit anti-rat antisera (Boehringer Mannheim), and visualized by diaminobenzidine reaction with nickel enhancement. For [³H]-thymidine incorporation, mice were given a single bolus injection of isotope (400 μ Ci at 28 Ci/mM, Amersham, Arlington Heights Ill.) and eighteen hours later sacrificed by cervical dislocation. The heart was removed, cryoprotected in 30% sucrose, embedded and sectioned with a cryomicrotome. Sections were post-fixed in methanol:acetone (1:1), stained with H and E, and a thin layer of photographic emulsion (Ilford L4, Polysciences, Warrington Pa.) diluted 1:1 with distilled water was applied. Sections were exposed for 5–7 days at 4° C., and developed in Kodak D-19 at 20° C. for 4 minutes, washed with distilled water for 1 minute, fixed in 30% sodium thiosulfate for 10 minutes, and washed in distilled water.

Electron Microscopy (EM)

Tissue blocks were fixed in 2% glutaraldehyde in 0.1M cacodylate buffer (pH 7.4) and post-fixed in 2% osmium tetroxide (Stevens Metallurgical Corp., New York N.Y.). All other EM chemicals were obtained from Ladd Research Industries, Inc. (Burlington Vt.). Tissue was stained en bloc with 2% uranyl acetate in pH 5.2 maleate buffer (0.05 M), dehydrated, and embedded in Ladd LX-112. Grafts were located using 1 μ m sections stained with toluidine blue. After trimming, the block was thin sectioned, and stained with uranyl acetate and lead citrate. Specimens were viewed on a Phillips 400 transmission electron microscope.

Electrocardiogram (ECG) Analyses

For surface ECG records, mice were anesthetized (2.5% Avertin, 0.015 ml/g body weight, IP, Fluka Chemicals, Lake Ronkonkoma N.Y.), surface electrodes were placed in the standard lead 1 position, and ECGs were recorded with a Narco Biosystems (Houston Tex.) high gain amplifier coupled to an A/D converter (Coulbourn Instruments, Lehigh Valley Pa.).

Plasma Enzyme Assay (PEA)

For lactate dehydrogenase (LDH) isoform assay, plasma was isolated by retro-orbital sinus bleeds under anesthesia (2.5% Avertin, 0.015 ml/g body weight, intraperitoneally (IP)). Plasma was fractionated on 1% agarose gels (CK Isoenzyme electrophoresis system. CIBA-Corning Diagnostics, Corning N.Y.) and the LDH isoforms visualized by a TNBT-Formazan histochemical assay (LDH Assay Kit, Sigma Diagnostics, St. Louis Mo.).

B. Results

In these studies AT-1 cardiomyocytes (derived from transgenic animals that expressed the T-Ag oncoprotein in the heart) were injected directly into the myocardium of syngeneic mice and the viability of the grafted material was assessed. To facilitate localization of the injection site in preliminary experiments, AT-1 cardiocytes were incubated briefly with BODIPY prior to grafting. BODIPY is a non-toxic glutathione reactive dye which permits fluorescent tracking of living cells. The graft site was easily visualized by fluorescence microscopy using a FITC cube. Subsequent experiments did not utilize BODIPY.

Fifty percent (14/28) of the animals receiving AT-1 cardiomyocyte injections developed intra-cardiac grafts. In

most instances, the grafts were neither encapsulated nor surrounded by scarred myocardium. At the level of light microscopy, grafted AT-1 cardiomyocytes were observed directly juxtaposed with host cardiomyocytes. The identity of the AT-1 cardiomyocytes was confirmed by immunoperoxidase assay using an anti-T-Ag antibody primary antibody (162-T) followed by a horseradish peroxidase conjugated secondary antibody. Specificity of the anti-T-Ag antibody has been established previously (10, 11). Black precipitate was observed over cardiomyocyte nuclei in the graft but not in the host myocardium, confirming that the graft was comprised of AT-1 cardiomyocytes. Similar results were obtained in the absence of primary antibody.

Viable AT-1 cardiomyocytes were observed at least as long as four months post-implantation. During this period, some degree of graft proliferation occurred; [³H]-thymidine incorporation analyses detected DNA synthesis in the grafted cells. Ten percent of the AT-1 cardiomyocyte nuclei were synthesizing DNA as evidenced by isotope incorporation into the nucleus. However, the rate was appreciably less than that observed for cultured AT-1 cardiomyocytes, where 50% of the cells synthesized DNA following a similar [³H]-thymidine pulse. In several instances, the grafted AT-1 cardiomyocytes were localized within the subpericardial space.

Immunohistologic experiments were employed to determine if the intra-cardiac grafts were subject to chronic rejection. Grafts older than one month failed to react with antibodies specific for mouse leukocytes; signals observed in blood vessels located on the same section provided a positive control for the experiment. Similarly, an antibody which detects mouse macrophages and lymphocytes did not react with the intra-cardiac graft; once again positive signal was observed in a blood vessel located on the same section. Collectively, these results indicate the absence of chronic graft rejection by the syngeneic hosts. This result is supported by the observation that cyclosporine treatment (50 mg/kg body weight, administered intraperitoneally daily) did not influence significantly the frequency of intra-cardiac grafting (50% success rate, n=6). Sex of the host animal also did not appear to influence significantly the rate of graft formation (46% success rate in males, n=13; 53% success rate in females, n=15). The frequency of grafting was similar in animals examined at early time points (1–40 days post-grafting, 47%, n=15) as compared to those examined at later time points (40–120 days post-grafting, 54%, n=13). Finally, similar frequencies of intra-cardiac grafting were observed when cells were delivered to either the left ventricular free wall or the apex of the heart.

Electron microscopic analysis of the AT-1 cardiomyocyte grafts confirmed the absence of encapsulation. High power views revealed well-developed junctional complexes between adjacent cells within the graft. Graft cardiomyocytes contained numerous polyribosomes and the dedifferentiated myofibrillar ultrastructure typical of AT-1 tumors in vivo (6). Electron-dense secretory granules were also observed in the AT-1 cardiomyocyte grafts, as would be expected for myocytes of atrial origin. Host cardiomyocytes bordering the grafts had normal ultrastructure with well-formed sarcomeres. Although only a thin basement membrane separated AT-1 and host cardiomyocytes, no junctional connections between these two cell types were observed.

Surface electrocardiograms were performed to determine if the presence of AT-1 cardiomyocyte grafts influenced the autonomic rhythm. No appreciable differences were observed between records from sham animals and those which harbored grafts. In each case, the experimental ani-

mals exhibited normal sinus rhythm, with an anesthetized heart rate of approximately 400 beats per minute. Normal P-QRS coupling was maintained, indicating that the grafted AT-1 cardiomyocytes did not act as an ectopic pacemaker. This latter result is important in light of the observation that AT-1 cardiomyocytes exhibit spontaneous electrical activity both in vivo (12) and in culture (3). The absence of overt arrhythmia also indicated that graft-induced myocardial remodeling was not associated with the generation of significant circus rhythms.

In addition to surface ECG, plasma LDH levels were assessed in mice carrying AT-1 cardiomyocyte grafts. The presence of the cardiac LDH isoform in the circulation is a well established hallmark of myocardial infarction. No cardiac LDH (isoform-1) was apparent in mouse plasma prior to grafting. After the introduction of AT-1 cardiomyocytes, there was a transient appearance of the cardiac isoform in the plasma, which most likely reflected damage to the host myocardium as well as damaged AT-1 cardiomyocytes. A transient increase in plasma skeletal LDH isoform was also observed following grafting surgery, presumably reflecting damage caused by the trans-thoracic incision. The plasma LDH profiles returned to normal by 7 days post-implantation. Thereafter, the plasma LDH profiles remained normal despite the presence of grafts.

EXAMPLE 2

Generation of Stable C1C12 Myoblast Grafts

A. Methods

C2C12 Cell Culture and Myocardial Grafting Protocol

C2C12 myoblasts were obtained from ATCC. Cells were maintained in the undifferentiated state by culturing at low density in high glucose Dulbecco's Modified Eagle Media (DMEM) supplemented with 20% fetal bovine serum, 1% chicken embryo extract, 100 units/ml penicillin and 100 $\mu\text{g}/\text{ml}$ streptomycin. For some studies, myogenic differentiation was induced by culturing in DMEM supplemented with 2% horse serum and antibiotics. Immediately before injection, myoblasts were harvested with trypsin, washed three times with serum free DMEM and directly injected into the ventricular myocardium of adult syngeneic C3Heb/FeJ mice (Jackson Laboratories) under open heart surgery as described in (7). Cells ($4-10 \times 10^4$) were injected in a volume of 2-3 μl using a plastic syringe fitted with a 30 gauge needle.

Histology

Hearts were removed, cryoprotected, embedded and sectioned as in Example 1. H and E staining and [^3H]-thymidine incorporation assays were also conducted as in Example 1. For immunohistology, methanol fixed sections (-20°C ., 10 min.) were reacted with the monoclonal anti-skeletal myosin heavy chain antibody (MY-32, Sigma Chemical Corp.) followed by rhodamine-conjugated sheep anti-mouse IgG F(ab')₂ fragment (Boehringer Mannheim), and visualized by epifluorescence.

Electron Microscope

EM was performed as in Example 1.

Electrocardiogram Analyses

ECG analyses were performed as in Example 1.

Plasma Enzyme Assay

PEA was performed as in Example 1.

B. Results

Several myoblast cell lines are known which, as exemplified by C2C12 cells, have the capacity to differentiate into myotubes in culture (13). C2C12 myoblasts were derived from cultured explants of injured thigh muscle of C3H mice. When maintained in serum-rich media, the myoblasts pro-

liferate rapidly and retain an undifferentiated phenotype. However, when cultured in serum-poor media myogenic differentiation is induced. The C2C12 cells withdraw from the cell cycle and fuse, thereby forming multinucleated myotubes. Myogenic differentiation is also induced, as evidenced by the appearance of numerous muscle-specific gene products. Thus, in this model proliferation and myogenic differentiation are mutually exclusive (14). Myoblast differentiation in vitro is thought to mimic satellite cell mediated myofiber regeneration in vivo.

Myoblasts were injected directly into the myocardium of syngeneic C3Heb/FeJ mice and the viability of the grafted material was assessed. One hundred percent (13/13) of the mice receiving intra-cardiac implants of C2C12 myoblasts developed grafts in the heart. Viable grafts were observed as long as six months post-implantation (this was the last time point assayed) In all instances, the grafted material was not encapsulated. The differentiated status of the grafted C2C12 cells was determined by immunohistological assay with an anti-myosin heavy chain antibody (MY-32). This antibody does not react with myoblasts nor with cardiac myosin heavy chain. Although differentiated C2C12 cells were observed in every heart receiving myoblast injections, the grafting efficiency of individual cells was not determined. As an additional control, hearts bearing AT-1, intra-cardiac grafts (see Example 1) were examined with the MY-32 antibody. No staining was observed, thereby ruling out the possibility that the signal seen in the C2C12 grafts was due to skeletal myosin heavy chain induction in host cardiomyocytes.

Example 1 above demonstrates that AT-1 cardiomyocytes form stable grafts in syngeneic myocardium. However, the observation that these cells retained the capacity for proliferation in vivo raised the possibility that sustained cell division might be required for successful intra-cardiac grafting. The proliferative status of the C2C12 grafts was therefore examined. Virtually no DNA synthesis (as assessed by tritiated thymidine incorporation) was observed, indicating that the majority of the grafted C2C12 cells had indeed withdrawn from the cell cycle. Examination of serial sections indicated that less than 0.1% of the cells in or near the grafts were synthesizing DNA. This result most likely reflects fibroblast proliferation during the remodeling process. As with the AT-1 grafts, immunohistological analyses of C2C12 grafts failed to detect macrophage, inflammatory leukocyte or lymphocyte infiltration at two months post-implantation, indicating the absence of chronic graft rejection by the syngeneic hosts.

At the level of light microscopy, the C2C12 intra-cardiac grafts exhibited cellular heterogeneity with both H and E and MY-32 immunofluorescence staining. Electron microscopic analyses were employed in an effort to further characterize the cellular make-up of the C2C12 grafts. Toluidine-stained 1 μm sections were surveyed at 100 μm intervals to locate graft sites for EM analysis. Once localized, thin sections were prepared from the block. Cells with morphology typical of skeletal myocytes were observed throughout the graft. Abundant mitochondria localized between well developed sarcomeres were readily detected. Prominent Z bands and thick and thin filaments were observed. Occasionally, expanded t-tubules and ruffled cell membranes were detected in the grafted myocytes. In addition to well developed myocytes, a second less differentiated cell type was observed in C2C12 grafts. Most notably, these cells exhibited a large nucleus to cytoplasm ratio, with a prominent band of heterochromatin at the nuclear periphery. Moderate amounts of centrally located heterochromatin

were also detected. Limited rough endoplasmic reticulum and few mitochondria were observed in these cells. Similar ultrastructural characteristics have been ascribed to satellite cells in vivo and in culture (15, 16).

Two studies were initiated to assess any deleterious effects of C2C12 intra-cardiac grafts on host heart function. In the first study surface electrocardiograms failed to detect any appreciable differences between records from control and experimental mice. All animals examined had normal P-QRS coupling, and exhibited normal sinus rhythm with an anesthetized heart rate of approximately 400 beats per minute. These data indicate that the intra-cardiac myoblast grafts did not induce overt cardiac arrhythmias. In the second study plasma LDH levels were monitored in graft-bearing animals. The presence of cardiac LDH isoform in the circulation is a well established hallmark of myocardial infarction. The cardiac-specific LDH isoforms (isoforms 1, 2, and 3) were not observed in plasma prior to grafting. Immediately after grafting, an increase in the cardiac isoforms was observed in plasma, which most likely reflected damage to the host myocardium. A transient increase in the plasma skeletal LDH isoform (isoform 5) was also observed, presumably reflecting damage caused by the trans-thoracic incision. Plasma LDH profiles returned to normal by 7 days post-implantation.

EXAMPLE 3

Generation of Stable Fetal Cardiomyocyte Grafts

A. Methods

Cardiomyocyte Cell Culture and Myocardial Grafting Protocol

Transgenic mice were generated which carry a fusion gene comprised of the α -cardiac myosin heavy chain (MHC) promoter and a modified β galactosidase (nLAC) reporter. To generate the MHC-nLAC transgenic mice, MHC-nLAC insert DNA (see FIG. 1) was purified by absorption onto glass beads, dissolved at a concentration of 5 μ g/ml, and microinjected into the nuclei of one cell inbred C3H3B/FeJ embryos according to established protocols (17). Polymerase Chain Reaction (PCR) analysis was employed to identify founder animals and to monitor transgene segregation. The sense strand primer 5'-GGTGGGGGCTCTTCACCCAGACCTCTCC-3' was localized to the MHC promoter and the antisense strand primer 5'-GCCAGGGTTTTCCAGTCACGACGTTGT-3' was localized to the nLAC reporter. PCR analyses were as described in (18). The MHC promoter consisted of 4.5 kb of 5' flanking sequence and 1 kb of the gene encompassing exons 1 through 3 up to but not including the initiation codon. The nLAC reporter was modified so as to carry both a eukaryotic translation initiation site and the SV40 nuclear localization signal (19). The mPl sequences carried an intron, as well as transcriptional termination and polyadenylation signals from the mouse protamine 1 gene.

For preparations for examination of β galactosidase (β GAL) activity and DAPI epifluorescence, transgenic animals were heparinized (10,000 U/kg IP) prior to sacrifice by cervical dislocation. Hearts were placed in a beaker of gassed (95% O₂, 5% CO₂) KHB buffer (105 mM NaCl, 20 mM NaHCO₃, 3.8 mM KCl, 1 mM KH₂PO₄, 1.2 mM MgSO₄, 0.01 mM CaCl₂, 1 mM mannitol, 10 mM taurine, 10 mM dextrose, 5 mM Na-pyruvate). Hearts were then hung by the aorta and perfused with gassed KHB (0.5 ml/min at 37° C.) containing 2.5 mM EGTA for five minutes, followed by 0.17% collagenase (Type I, Worthington Biochemical, Freehold N.J.) in KHB. Hearts were perfused until flaccid and the ventricles were minced with scissors and isolated

cells obtained by triturating with a Pasteur pipette. After at least one hour of formalin fixation, suspensions were filtered and smeared onto positively charged slides (Superfrost Plus, Fisher, Pittsburgh Pa.), and allowed to dry.

For isolation of single cells for injection, females with 15 day embryos (onset of pregnancy determined by vaginal plugs) were sacrificed by cervical dislocations. Embryos were removed, decapitated, and hearts were harvested under PBS, and ventricles and atria were separated. Transgenic ventricles (identified by cardiac β GAL activity) were digested in 0.1% collagenase (Worthington) in DPBS (Dulbecco's Phosphate Buffered Saline, Sigma) for 45 minutes, and were triturated with a Pasteur pipette in PC-1 medium (Ventrex, Coons Rapids Minn.) with 10% FBS, resulting in a suspension of single cells.

Immediately after isolation, embryonic cardiomyocytes were washed three times with DPBS and directly injected into the ventricular myocardium of syngeneic mice (Jackson Laboratories) under open heart surgery as in Example 1. 1-10 \times 10⁴ cells were injected in a volume of 2-3 μ using a plastic syringe fitted with a 30 gauge needle.

Histology

For H and E, X-GAL, immunohistology and thymidine analyses, hearts were removed following cervical dislocation and cryoprotected in 30% sucrose, embedded and sectioned at 10 μ m with a cryomicrotome as in Example 1. H and E staining, monitoring for intra-cardiac graft rejection, and assay for [³H]-thymidine incorporation were also conducted as in Example 1. To assay β GAL activity, section were hydrated in PBS, post-fixed in acetone:methanol (1:1) and then overlaid with mixture containing 1 mg/ml X-GAL (5-bromo-4-chloro-3-indolyl- β -D-galactoside), 5 mM potassium ferricyanide, 5 mM potassium ferrocyanide and 2 mM magnesium chloride in PBS. Positive staining is indicated by the appearance of a blue chromophore. After treatment with primary antibody, signal was visualized by an avidin-biotin (ABC) kit (Vector Labs, Burlingame Calif.). The heart was processed as described above, and sections were post-fixed in methanol:acetone (1:1), stained with H and E, and coated with a thin layer of photographic emulsion (Ilford L4. Polysciences) diluted 1:1 with distilled water. Sections were exposed, developed, washed, fixed and washed as in Example 1, X-GAL staining of single cell preparations was as described above. For visualization of nuclei in single cell preparations, slides were stained with DAPI in PBS (0.28 μ M, three min. at room temperature, Boehringer Mannheim), washed three times in PBS, and wet-mounted in 2% propyl gallate dissolved in glycerol. To obtain coronal heart sections, mice were sacrificed by cervical dislocation, hearts were harvested and perfused on a Langendorff apparatus with 2% glutaraldehyde in 0.1 M cacodylate buffer (pH 7.4). After immersion fixation overnight in the same buffer, 200 μ m coronal sections were made with a vibratome (Campden, London, United Kingdom). To localize the graft, sections were pooled and stained for β GAL activity with X-GAL as described above.

Electron Microscopy

MHC-nLAC embryonic grafts were localized in coronal heart sections as described above. After trimming, the tissue was post-fixed in 2% osmium tetroxide (Stevens Metallurgical Corp., New York, N.Y.). Tissue was then dehydrated and embedded in Ladd LX-112 (Ladd Research Industries). Grafted areas were further trimmed, thin sectioned, and stained with uranyl acetate and lead citrate. Specimens were viewed on a Phillips 400 transmission electron microscope as in Example 1.

Electrocardiogram Analyses

ECG analyses were conducted as in Example 1.

B. Results

Transgenic mice generated as above carried a fusion gene comprised of the MHC promoter and a nLAC reporter. nLAC carries the SV40 nuclear transport signal, which results in the accumulation of β galactosidase activity in the nucleus of targeted cells. Four transgenic lineages were produced, and two (designated MHC-nLAC-2 and MHC-nLAC-4) were selected for further analyses. To ensure that the MHC-nLAC transgene would provide a suitable cell lineage marker, β galactosidase (β GAL) activity was assessed in transgenic cardiomyocytes. Single cell preparation generated by retrograde collagenase perfusion were examined simultaneously for β GAL activity and DAPI epifluorescence, $99.0 \pm 0.45\%$ (n=400) of the transgenic cardiomyocyte nuclei expressed β GAL, whereas no β GAL activity was detected in noncardiomyocytes. In addition, no nuclear β GAL activity was detected in nontransgenic control cardiomyocytes.

Single cell suspensions were prepared by collagenase digestion of hearts harvested from embryonic day 15 transgenic mice. Greater than 95% of the cardiomyocytes isolated by this technique were viable as evidenced by dye exclusion assay. Cardiomyocytes were delivered to left ventricular free wall of syngeneic nontransgenic animals. Grafted cardiomyocytes were readily and unambiguously identified by virtue of the nuclear β GAL activity encoded by the MHC-nLAC transgene. Grafted cardiomyocytes were frequently observed at sites distal to the point of delivery; it presently is not clear if this distribution of grafted cells reflects cardiomyocyte migration or passive diffusion along dissection planes produced by the injection process. Approximately 50% (7/13) of the animals receiving intra-cardiac injections of embryonic cardiomyocytes developed grafts. This frequency of successful graft formation is likely to increase as cell preparation and implantation protocols are optimized.

Light microscopic analyses of H and E stained sections processed for β GAL activity indicated that grafted cardiomyocytes (blue nuclei) were juxtaposed directly with host cardiomyocytes (purple nuclei). Additional H and E analyses failed to detect significant graft encapsulation. The observed proximity of graft and host cardiomyocytes and absence of encapsulation are prerequisites for successful coupling between the two cell types.

Consecutive sections of a 19 day old intra-cardiac graft were processed for H and E, β GAL activity, and macrophage and leukocyte immunoreactivity. No evidence for graft rejection was observed, despite the fact that the animals were not immune suppressed. As a positive control for the immunohistology, grafts of incompatible MHC haplotype were produced; graft rejection was clearly evident in these hearts. Tritiated thymidine uptake analyses indicated that only 0.6% (n=156) of the β GAL-positive nuclei were synthesizing DNA, although noncardiomyocyte DNA synthesis was apparent. Since the embryonic day 15 donor cells were still mitotically active when grafted (labeling index of ca. 29%), the exceedingly low level of DNA synthesis observed in β GAL positive cells at 19 days post-grafting suggested that the MHC-nLAC embryonic cardiomyocytes had undergone terminal differentiation.

The juxtaposition of graft and host cardiomyocytes observed by light microscopic analyses prompted a determination whether direct intercellular coupling could be detected between the two cell types. The X-GAL reaction product is an electron-dense precipitate which can be

detected by transmission electron microscopy (TEM, see 19). Vibratome sections from glutaraldehyde perfusion-fixed hearts were stained β GAL activity, and graft GAL positive nuclei were readily observed by light microscopic analysis of 1 μ m sections. The X-GAL reaction product had a perinuclear appearance due to a slight degree of nuclear leaching which occurred during the embedding process. The same groups of cardiomyocytes were identified by TEM analysis of a consecutive thin section. Host cardiomyocytes, which were not readily identified in the light micrographs due to the absence of perinuclear β GAL activity, were observed by electron microscopy to be juxtaposed with the grafted cells. Numerous junctional complexes were present between the host and graft cardiomyocytes, indicating a high degree of intercellular coupling. Many examples of intercellular coupling between host and graft cardiomyocytes were observed throughout the grafted regions. Importantly, intercellular connections could be traced from β GAL positive cardiomyocytes through numerous host cells, thus demonstrating that grafted cardiomyocytes could be participating in a functional syncytium.

In addition to documenting the presence of abundant intercellular coupling between grafted and host cardiomyocytes, the TEM analyses revealed that the grafted cardiomyocytes were highly differentiated. Normal characteristics of adult cardiomyocytes were observed including myofibrillae forming complete sarcomeres, numerous junctional complexes between cells and abundant mitochondria. Indeed, aside from the presence of the X-GAL reaction product, grafted cardiomyocytes were indistinguishable from host cells. Further, binucleated, β GAL positive cells could be detected in the intra-cardiac grafts. Because binucleation is a characteristic of adult rodent cardiomyocytes, this observation further supports that the grafted grafts have undergone terminal differentiation.

Surface ECG recordings were employed to determine if the presence of coupled embryonic cardiomyocyte grafts negatively influenced host heart automaticity. ECG traces from graft-bearing animals were indistinguishable from sham operated controls, and exhibited P and QRS complexes typical for mice. There was no evidence for cardiac arrhythmia in graft-bearing animals, despite the presence of a high degree of intercellular coupling between grafted and host cardiomyocytes.

EXAMPLE 4

Preparation of Substantially Pure Cardiomyocyte Culture

Embryonic stem cells were genetically modified in a manner enabling the production of a substantially homogeneous population of non-immortalized cardiomyocytes. The parental ES cell line (D3) was cotransfected with a pGK-HYG (hygromycin) plasmid and a plasmid containing a MHC-neo^r gene. The pGK-HYG plasmid provides selection for transfected ES cells, while the mMHC-neo^r gene facilitates a second round of selection on differentiated cells: incubation in the presence of G418 eliminates non-cardiomyocytes (that is, cells in which the MHC promoter is not active)

Stably transfected ES cells were selected by growth in the presence of hygromycin. The plasmids were linearized and introduced into the stem cells via electroporation at 1180 μ Farad, 220 volts. The transfected cells were maintained in DMEM supplemented with 10% preselected FBS, 0.1 mM β -mercaptoethanol, nonessential amino acids. PenStrep and LIF, and transformants selected by the addition of hygro-

mycin into the medium. Co-transfectants were then identified by PCR analysis specific for both transgenes. The transfections produced a cell line, designated 9A, which carries both transgenes.

Cardiogenesis was induced in 9A ES cells by plating 2×10^6 cells onto uncoated 100 mm bacterial petri dishes in the absence of LIF. After 8 days in culture, numerous patches of cells exhibiting spontaneous contractile activity (i.e. cardiomyocytes) were observed. At this point, G418 was added to the media, and the cells incubated for an additional 9 days. During this treatment it was apparent that many of the non-cardiomyocytes were being killed by the G418. Importantly, the G418 had no discernible effects on the cardiomyocytes, which retained their spontaneous beating activity throughout the course of the experiment.

After a total of 9 days of G418 selection, the surviving cells were dissociated with collagenase and trypsin, and then replated onto fibronectin coated microscope slides. The cells were cultured an additional 24 hours to allow them to recover from passage, and then fixed for immunocytologic analysis. Cells were reacted with MF20, a monoclonal antibody which recognizes sarcomeric myosin. Cells were then individually counted for the presence or absence of sarcomeric myosin, a marker for cardiomyocytes. The results were as follows:

Total number of cells counted:	794
Number of MF20+ cells:	791
Number of MF20- cells:	3*
Percent cardiomyocytes:	99.6%

*The 3 cells which did not stain with MF20 may still have been cardiomyocytes, since this antibody will not react with monomeric myosin.

It was thus demonstrated that the expression of drug (neomycin) resistance under a cardiac-specific promoter enables the selection of an essentially pure population of ES derived cardiomyocytes in culture. Such populations can be used to form myocardial grafts using procedures as discussed in the Examples above.

EXAMPLE 5

Delivery of Protein via Graft

A. Methods

C12C12 Cell Culture and Transfection

C12C12 myoblasts (ATCC) were maintained in the undifferentiated state by culturing at low density in high glucose Dulbecco's Modified Eagle Media (DMEM)¹ supplemented with 20% fetal bovine serum, 1% chicken embryo extract, 100 units/ml penicillin and 100 μ g/ml streptomycin. For some studies, myogenic differentiation was induced by culturing in DMEM supplemented with 2% horse serum and antibiotics.

A fusion gene comprised of the metallothionein (MT) promoter driving a modified Transforming Growth Factor-Beta 1 (TGF- β 1) cDNA was obtained from Samuel and colleagues (20). Transcriptional activity of the metallothionein promoter can be regulated by modulating the heavy metal content of cell culture media. The TGF- β 1 cDNA carried site-directed mutations which resulted in the conversion of Cys²²³ and Cys²²⁵ to serines. This modification (described further in (21)) results in the elaboration of a TGF- β 1 molecule which is unable to form dimers, and consequently is not subject to normal post translational regulation. Cells expressing the modified cDNA constitu-

tively secrete processed, active TGF- β 1 (20). The MT-TGF fusion gene was introduced into C12C12 myoblasts by calcium phosphate transfection; stable transfectants were selected by virtue of co-transfection with an SV40-neo^r transgene. Four independent clones were isolated, and presence of the transgene was confirmed by Southern blot analysis. The relative levels of TGF- β 1 expression in the different clonal cell lines was initially assessed by Northern blot analysis, and one line, designated C2(280), was utilized for subsequent experiments.

Myocardial Grafting Protocol

The grafting protocol was as described in Example 1. Fourteen days post-surgery, graft bearing animals were given heavy metal (25 mM ZnSO₄ in drinking water). Zinc treatment was continued until the terminate of the experiment (1-4 weeks).

Histology

For paraffin sections, hearts were fixed in 10% neutral buffered formalin, dehydrated through graded alcohols, and infiltrated with paraffin. Tissue blocks were then sectioned at 6 μ m. H and E staining was performed directly after sectioning according to manufacturer's specifications (Sigma Diagnostics).

For [³H]-thymidine incorporation, mice were given a bolus and sacrificed as in Example 1. The heart was removed and processed for paraffin embedding as described above. Autoradiography was likewise conducted as in Example 1.

B. Results

Expression of recombinant TGF- β 1 in response to heavy metal induction was examined in C2(280) myoblasts and myotubes. Transgene transcripts (1.8 kb) were readily distinguished from those originating from the endogenous TGF- β 1 gene (2.5 kb) by Northern blot analysis. Addition of heavy metal to the culture media resulted in a marked increase of recombinant TGF- β 1 transcripts in C2(280) myoblasts and myotubes. As indicated above, modified TGF- β 1 expressed by C2(280) cells should have constitutive biological activity. To directly test this, conditioned media from C2(280) myoblasts and myotubes was examined by growth inhibition assay.

C2(280) myoblasts were used to produce intra-cardiac grafts in syngeneic C3Heb/FeJ mice. The presence of grafts was readily detected in H and E stained sections. 100% (n>50) of the animals receiving intra-cardiac injections of C2(280) cells went on to develop grafts. Interestingly, H and E analysis suggested that the C2(280) grafts were somewhat less differentiated as compared to those produced with unmodified C2C12 cells. This result was confirmed by immunohistologic analysis with a monoclonal antibody which recognizes skeletal myosin heavy chain.

C2(280) graft transgene expression was assessed by immunohistology with an anti-TGF- β 1 antibody; TGF- β 1 expression was readily detected in C2(280) grafts. As a negative control, TGF- β 1 expression was assessed in grafts produced by C2C12 myoblasts. As expected, the relative levels of TGF- β 1 expression were markedly reduced in C2C12 grafts as compared to C2(280) grafts.

TGF- β 1 is a well known angiogenic factor. ³H-thymidine incorporation analyses in vascular endothelial cells was therefore assessed to determine if an enhanced angiogenic response occurred in grafts expressing the MT-TGF transgene. DNA synthesis in vascular endothelial cells was readily apparent in C2(280) grafts under administration of a single bolus injection of ³H-thymidine (H-THY). In contrast, vascular endothelial DNA synthesis was markedly reduced in non-transfected C2C12 grafts (Table 1). To rule out the possibility that the angiogenic responses was due

solely to graft mass. ³H-thymidine incorporation was compared between similar size and aged C2C12 and C2(280) grafts (Table 1). A marked increase in the number of vascular endothelial cells synthesizing DNA was apparent in all of the analyses. Finally, the thymidine incorporation assay also revealed that a percentage of the grafted myoblasts continued to proliferate. This observation is consistent with the known inhibitory effect of TGF- β 1 on myodifferentiation, and most likely accounts for the undifferentiated appearance of the C2(280) grafts.

TABLE 1

		TGF- β 1 Delivery and Vascular Endothelial DNA Synthesis	
Time Post Zn Induction		C2C12 TGF- β 1(-)	C2(280) TGF- β 1(+)
1 Week	Total # ³ H-THY + Endothelial Cells	0	8
	Total # of Vessel Sections Counted	35	17
	# Synthetic Cells/ Vessel Section	0.0 \pm 0.00	0.46 \pm 0.036
2 Weeks	Total # ³ H-THY + Endothelial Cells	1	7
	Total # of Vessel Sections Counted	18	21
	# Synthesis Cells/ Vessel Section	0.06 \pm 0.056	0.34 \pm 0.052

While the invention has been illustrated and described in detail in the foregoing description, the same is to be considered as illustrative and not restrictive in character, it being understood that only the preferred embodiments have been described and that all modifications that come within the spirit of the invention are desired to be protected.

REFERENCES

The following references, to the extent that they provide exemplary procedural or other details supplementary to those set forth herein, are specifically incorporated herein by reference.

1. Tompson, L. Fetal transplants show promise. *Science* 257: 868-870, 1992.
2. Gussoni, E., Pavlath, G. K., Lanctot, A. M., Sharma, K. R., Miller, R. G., Steinman, L. and Blau, H. M. Normal dystrophin transcripts detected in Duchenne muscular dystrophy patients after myoblast transplantation. *Nature* 356: 435-438, 1992.
3. Steinhelper, M. E., Lanson, N., Dresdner, K., Delcarpio, J. B., Wit, A., Claycomb, W. C. and Field, L. J. Proliferation in vivo and in culture of differentiated adult atrial cardiomyocytes from transgenic mice. *American Journal of Physiology* 259 (Heart and Circulatory Physiology 28): H1826-H1834, 1990.
4. Janse, M. J., Cinca, J., Morena, H., Fiolet, J. W., Kleber, A. G., deVries, G. P., Becker, A. E. and Durrer, C. The border zone in myocardial ischemia. An electrophysiological, metabolic and histochemical correlation in the pig heart. *Circulation Research* 44: 576-588, 1979.
5. Spear, J. F., Michelson, E. L., and More, E. N. Cellular electrophysiologic characteristics of chronically infarcted myocardium in dogs susceptible to sustained ventricular tachyarrhythmias. *Journal of the American College of Cardiology* 4:1099-1110, 1983.
6. Delcarpio, J. B., Lanson, N. A. Jr., Field, L. J. and Claycomb, W. C. Morphological characterization of car-

diomyocytes isolated from a transplantable cardiac tumor derived from transgenic mouse atria (AT-1 cells). *Circulation Research* 69:1591-1600, 1991.

7. Rockman, H. A., Ross, R. A., Harris, A. N., Knowlton, K. U., Steinhelper, M. E., Field, L. J., Ross, J. Jr. and Chien, K. R. Segregation of atrial-specific and inducible expression of an ANF transgene in an in vivo murine model of cardiac hypertrophy. *Proc. Natl. Acad. Sci. U.S.A.* 88:8277-8281, 1991.
8. Bullock, G. R. and Petrusz, P., in *Techniques in Immunocytochemistry*, Vol. II, Academic Press, New York, 1983.
9. Field, L. J. Atrial natriuretic factor-SV40 T antigen transgenes produce tumors and cardiac arrhythmias in mice. *Science* 239:1029-1033, 1988.
10. Katz, E., Steinhelper, M. E., Daud, A. Delcarpio, J. B., Claycomb, W. C. and Field, L. J. Ventricular cardiomyocyte proliferation in transgenic mice expressing α -Cardiac Myosin Heavy Chain-SV40 T antigen fusion genes. *American Journal of Physiology* 262 (heart and Circulatory Physiology 31):H1867-1876, 1992.
11. Steinhelper, M. E. and Field, L. J. "SV40 large T-Antigen induces myocardiocyte proliferation in transgenic mice", in *The development and regenerative potential of cardiac muscle*. John Oberpriller and Jean Oberpriller, eds. Harwood Academic press, 1990.
12. Steinhelper, M. E. and Field, L. J. Cardiac Tumors and dysrhythmias in transgenic mice. *Toxicologic Pathology* 18: 464-469, 1990.
13. Yaffe, D., and O. Saxel. Serial passaging and differentiation of myogenic cells isolated from dystrophic mouse muscle. *Nature* 270:725-727, 1977.
14. Nadal-Ginard, B. Commitment, fusion, and biochemical differentiation of a myogenic cell line in the absence of DNA synthesis. *Cell* 15:885-864, 1978.
15. Bruni, C. Mitotic activity of muscle satellite cells during the early stages of rhabdomyosarcomas induction with nickel subsulfide. In *Muscle Regeneration*, A. Mauro, editor. Raven Press, New York, N.Y., 1979.
16. Rubin, L. L., C. E. Keller and S. M. Schuetze, Satellite cells in isolated adult muscle fibers in tissue culture. In *Muscle Regeneration*, A. Mauro, editor. Raven Press, New York, N.Y., 1979.
17. B. Hogan, F. Costantini and E. Lacy, in *Manipulating the Mouse Embryo—A Laboratory Manual*, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y., 1986.
18. M. E. Steinhelper, K. L. Cochrane and L. J. Field. Hypotension in transgenic mice expressing atrial natriuretic factor fusion genes. *Hypertension* 16:301-307, 1990.
19. A. D. Loewy, P. C. Bridgman and T. C. Mettenleiter. *Brain Research* 555:346, 1991.
20. S. K. Samuel, et al. Autocrine induction of tumor protease production and invasion by a metallothionein-regulated TGF-beta 1 (Ser223,225). *Embo Journal [JC:emb]* 11(4):1599-1605, 1992.
21. A. M. Brunner, et al. Site-directed mutagenesis of cysteine residues in the pro region of the transforming growth factor beta 1 precursor. Expression and characterization of mutant proteins. *Journal of Biological Chemistry* 264(23):13660-13664, 1989.
22. E. J. Robertson. Embryo-derived stem cell lines, in *Teratocarcinomas and embryonic stem cells: a practical approach*, E. J. Robertson, editor. IRL Press, Washington D.C., 1987.

SEQUENCE LISTING

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28

- What is claimed is:
1. A method of obtaining a population of cells enriched in a first cell lineage, comprising:
 - transfecting embryonic stem cells to introduce a marker gene enabling selection of a first cell lineage from other cell lineages resulting from the differentiation of the stem cells;
 - causing the stem cells to differentiate in vitro; and
 - selecting said one cell lineage based on said marker gene in vitro.
 2. The method of claim 1, comprising:
 - transfecting the stem cells to introduce (i) a first marker gene enabling selection of transfected stem cells from non-transfected stem cells and (ii) a second marker gene enabling selection of said one cell lineage from said other cell lineages;
 - selecting transfected stem cells based on the first marker gene;
 - causing the selected stem cells to differentiate; and
 - selecting said one cell lineage based on said marker gene.
 3. An embryonic stem cell having introduced DNA encoding a marker gene enabling selection of one cell lineage from other cell lineages resulting from the differentiation of the stem cells.
 4. The stem cell of claim 3 which includes introduced DNA encoding (i) a first marker gene enabling selection of embryonic stem cells having the first marker gene from embryonic stem cells not having the first marker gene and (ii) a second marker gene enabling selection of said one cell lineage from said other cell lineages.
 5. The stem cell of claim 3, wherein said marker gene confers antibiotic resistance to cells which express the marker gene.
 6. The stem cell of claim 4, wherein the first and second marker genes each confer antibiotic resistance to cells which express the marker genes.
 7. The stem cell of claim 6, wherein said second marker gene is under the control of a promoter which causes expression of the marker gene only in said one cell lineage.
 8. A method of obtaining a population of cells, comprising:
 - providing an embryonic stem cell having introduced DNA encoding a marker gene enabling selection of one cell lineage from other cell lineages resulting from the differentiation of the stem cells;
 - causing the stem cell to differentiate in culture to form a mixed differentiated cellular population containing said one cell lineage and said other cell lineages;
 - selecting from the cellular population said one cell lineage from said other cell lineages based on said marker gene.
 9. The method of claim 8 in which:
 - said marker gene is under the control of a promoter which causes expression of the marker gene in said one cell lineage but not in said other cell lineages, so as to enable selection of said one cell lineage from said other cell lineages.
 10. The method of claim 9, in which said marker gene confers antibiotic resistance, and wherein said selecting includes contacting the mixed differentiated cellular population with an antibiotic to which cells of said one cell lineage have been conferred resistance by the expression of said first marker gene, thereby killing cells of said other cell lineages but not cells of said one cell lineage which express the marker gene.
 11. The method of claim 10, in which said marker gene confers resistance to neomycin or hygromycin.
 12. A population of cells obtained by a method according to claim 8.
 13. A population of cells obtained by a method according to claim 10.

* * * * *