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(54) **ASPERGILLUS FUMIGATUS CELLULOLYTIC ENZYME COMPOSITIONS AND USES THEREOF**

9/2471 (2013.01); C12N 15/80 (2013.01); C12P 9/00 (2013.01); C12P 2203/00 (2013.01)

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(58) **Field of Classification Search**
None
See application file for complete search history.

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(73) Assignee: **Novozymes, Inc.**, Davis, CA (US)

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(*) Notice: Subject to any disclaimer, the term of this patent is extended or adjusted under 35 U.S.C. 154(b) by 0 days. days.

This patent is subject to a terminal disclaimer.

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(21) Appl. No.: **15/489,035**

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Related U.S. Application Data

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(60) Provisional application No. 61/526,805, filed on Aug. 24, 2011.

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(57) **ABSTRACT**

The present invention relates to recombinant *Trichoderma* host cells producing *Aspergillus fumigatus* cellulolytic enzyme compositions and methods of producing and using the compositions.

17 Claims, 8 Drawing Sheets

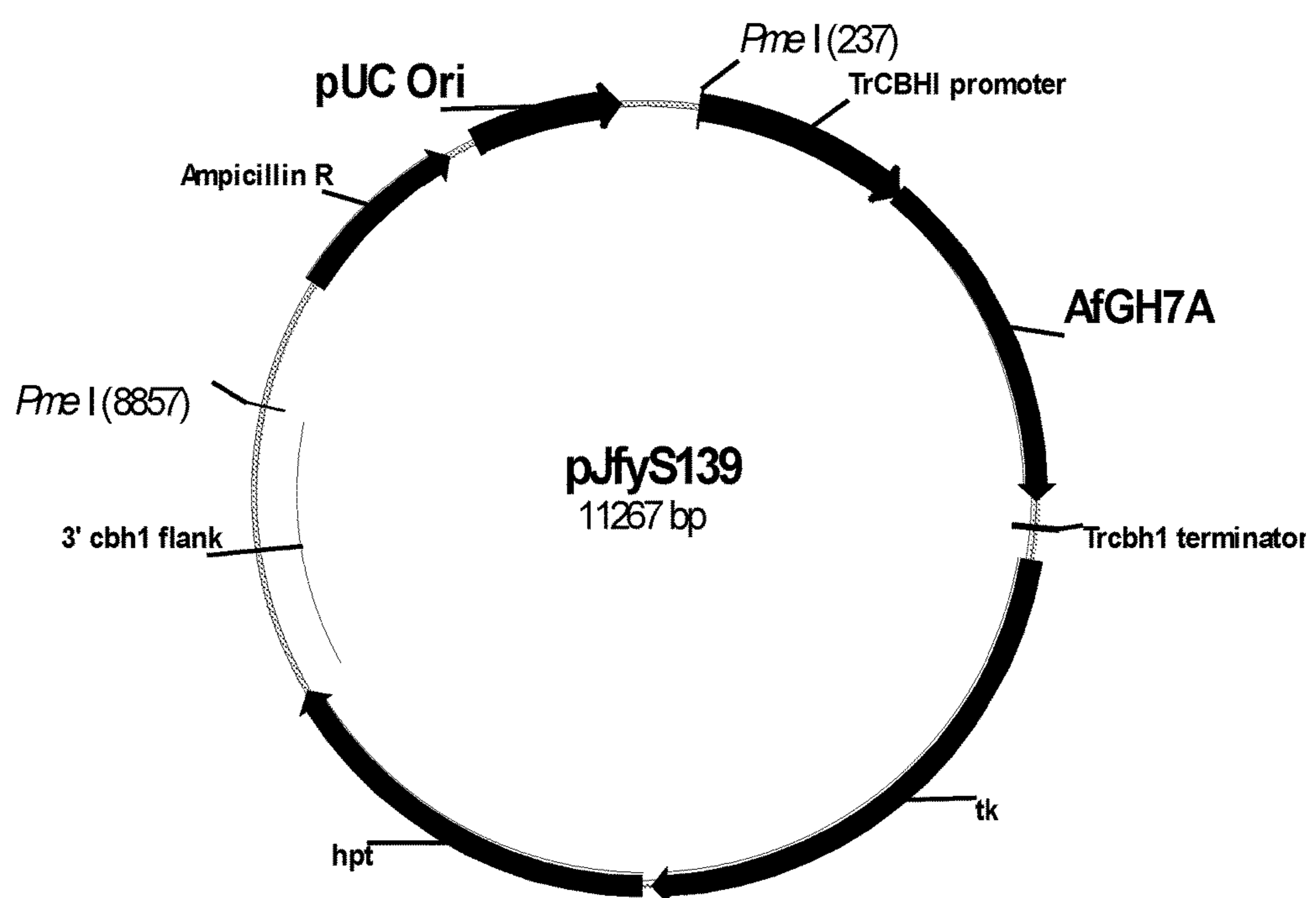


Fig. 1

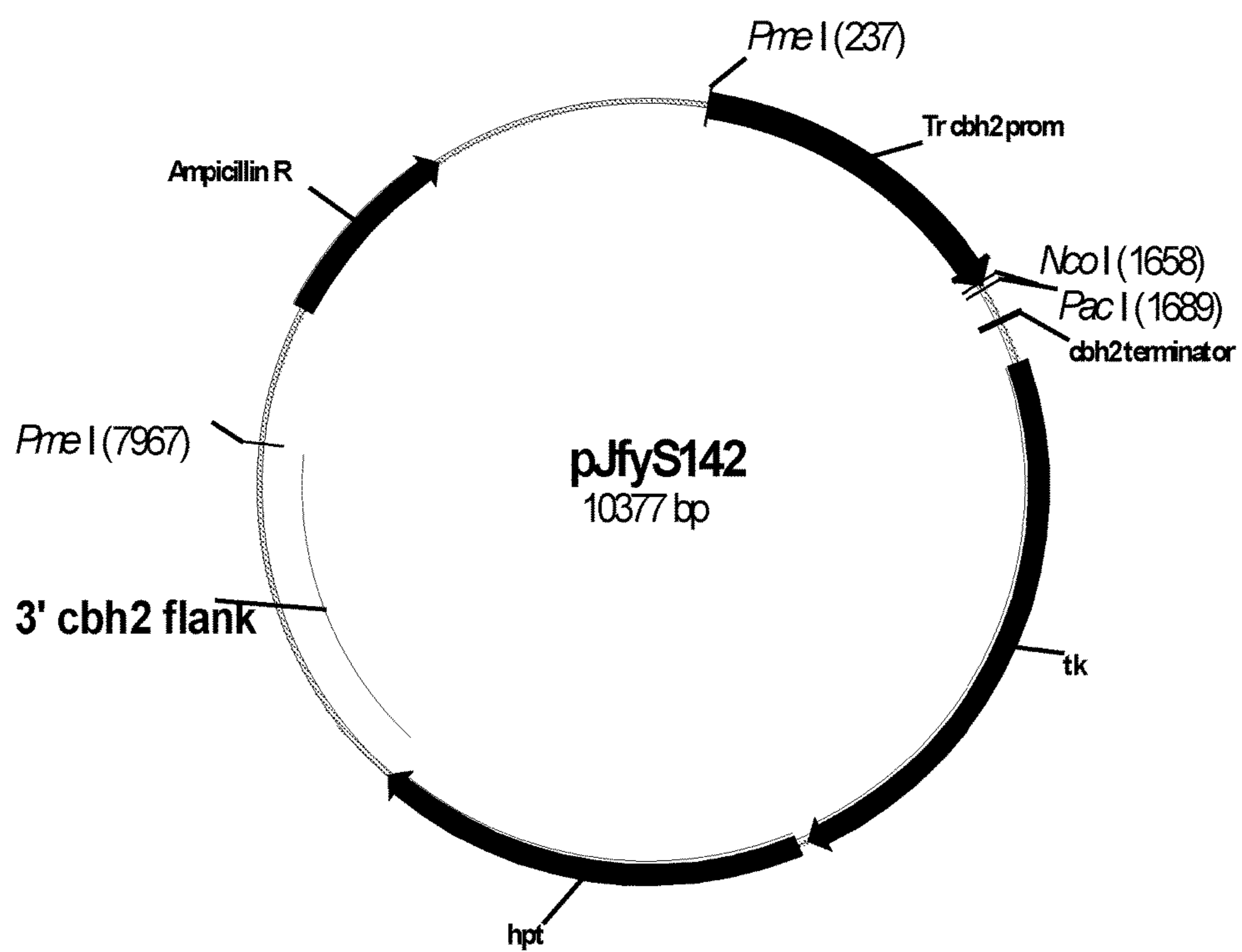


Fig. 2

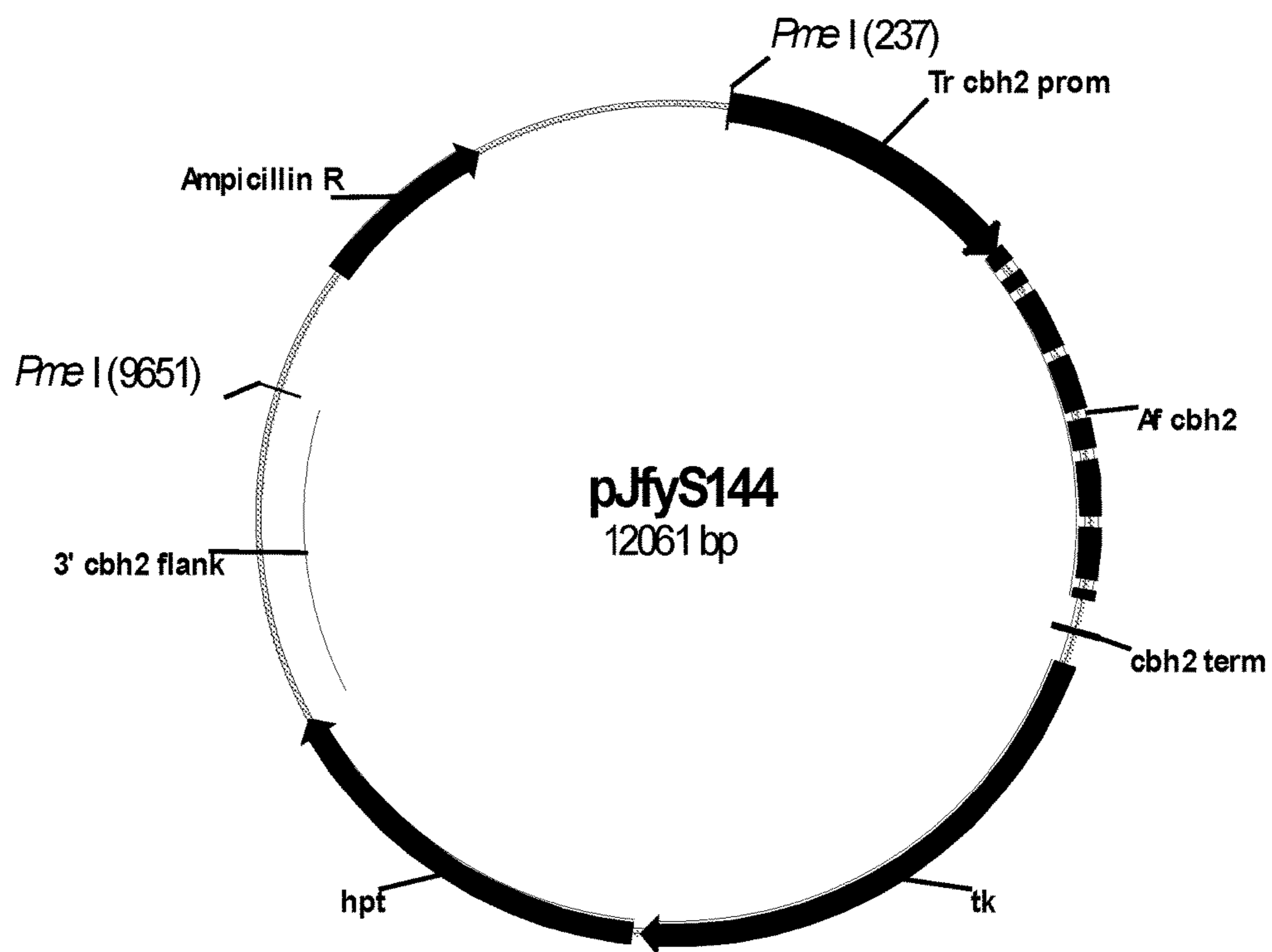


Fig. 3

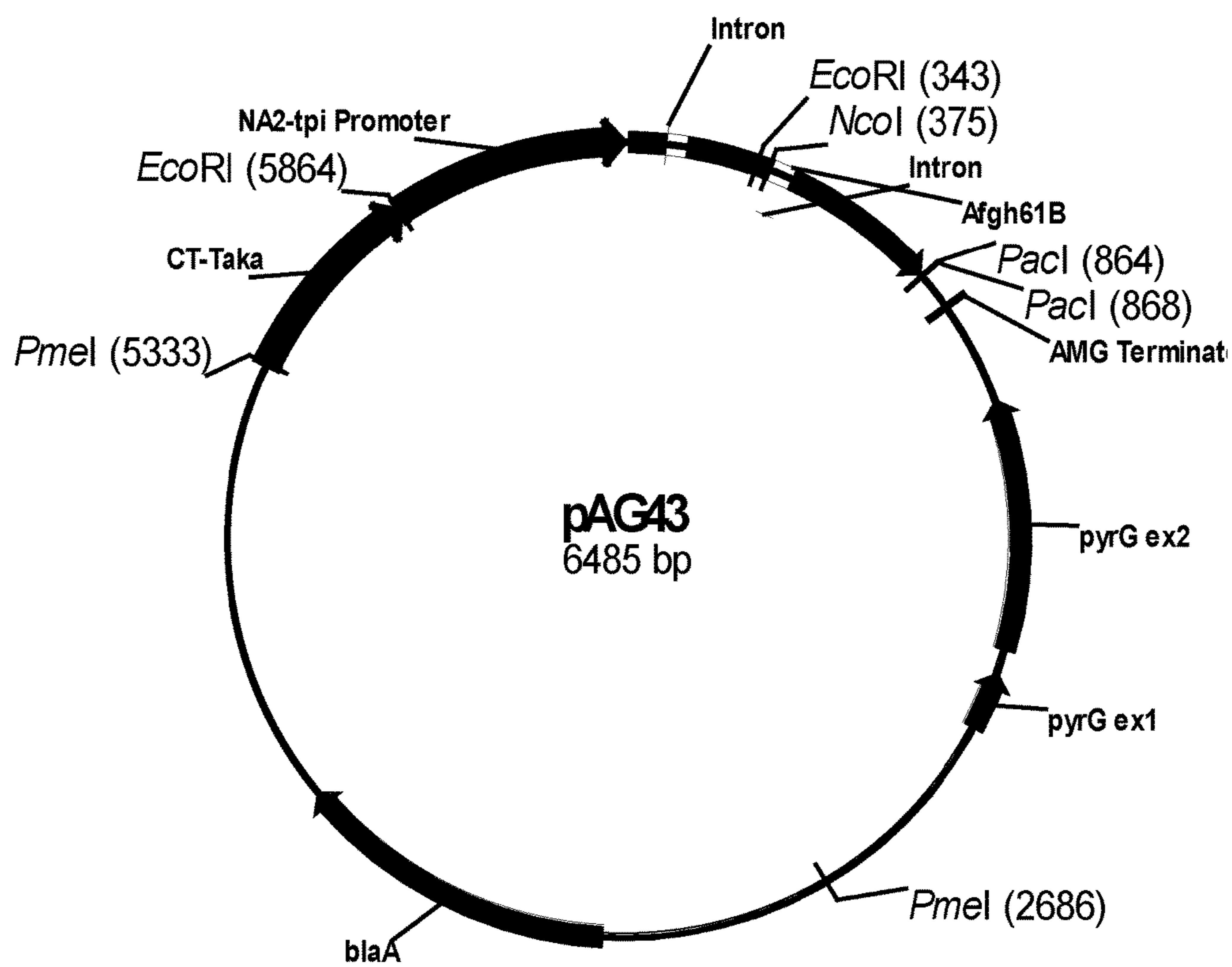


Fig. 4

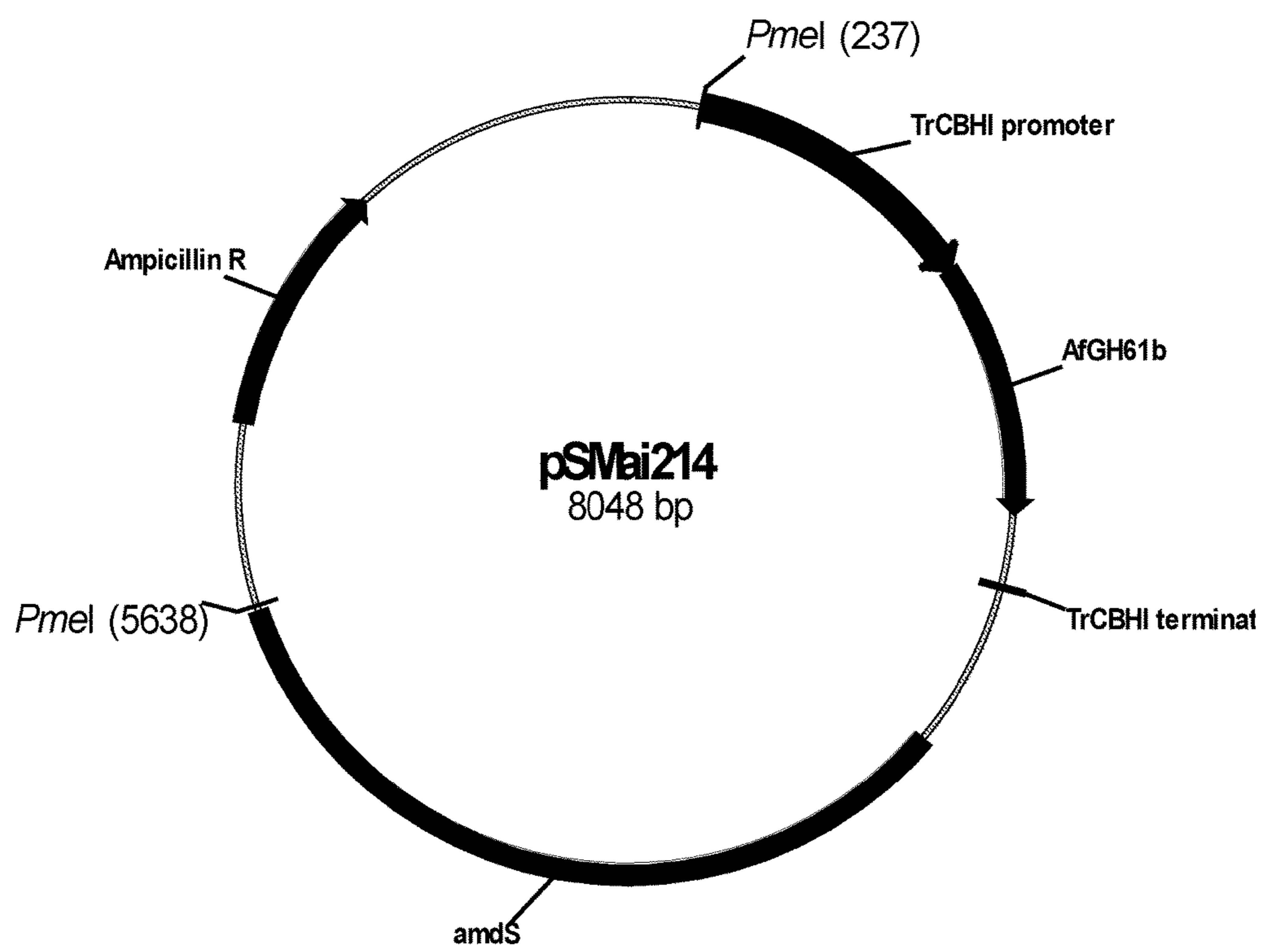


Fig. 5

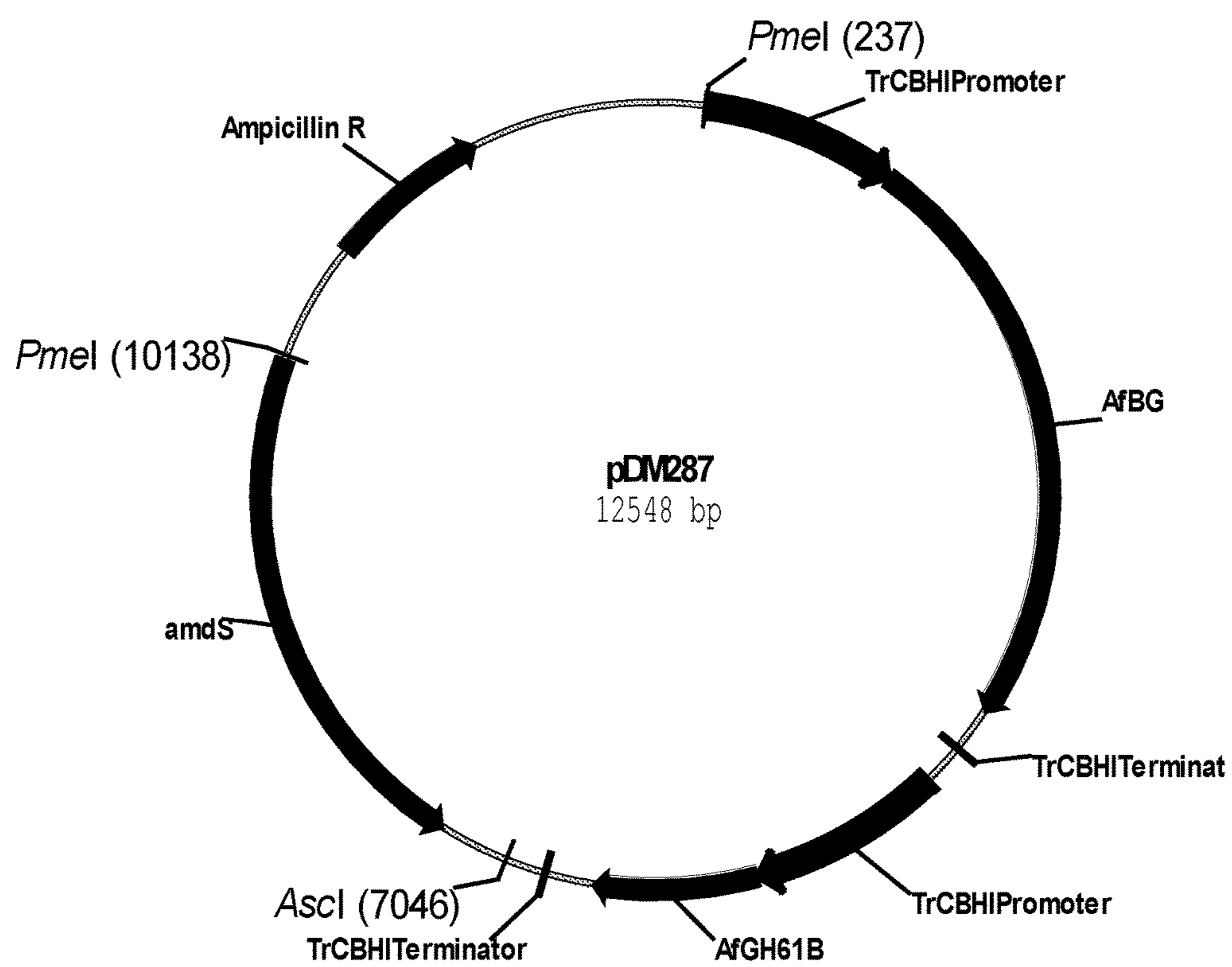


Fig. 6

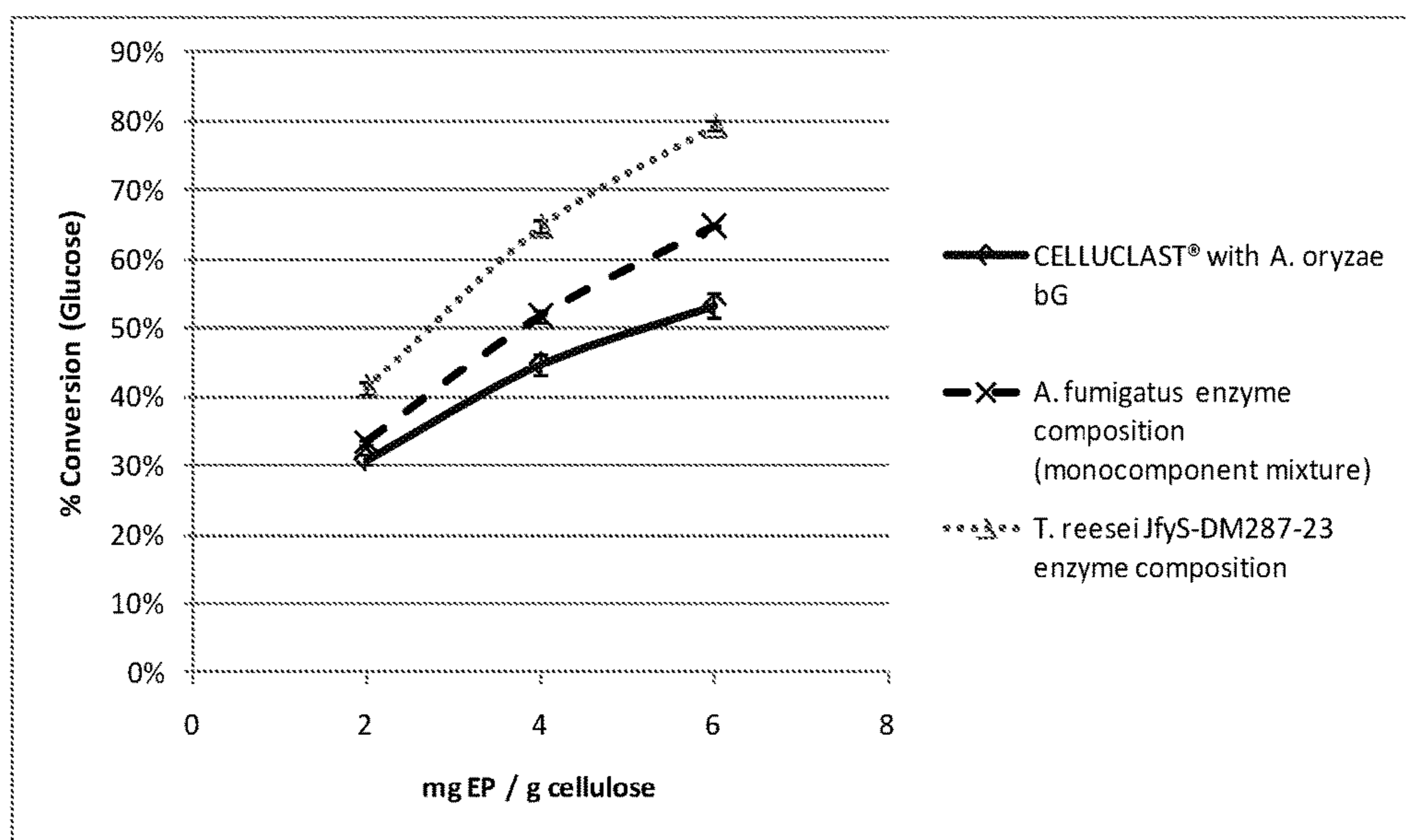


Fig. 7

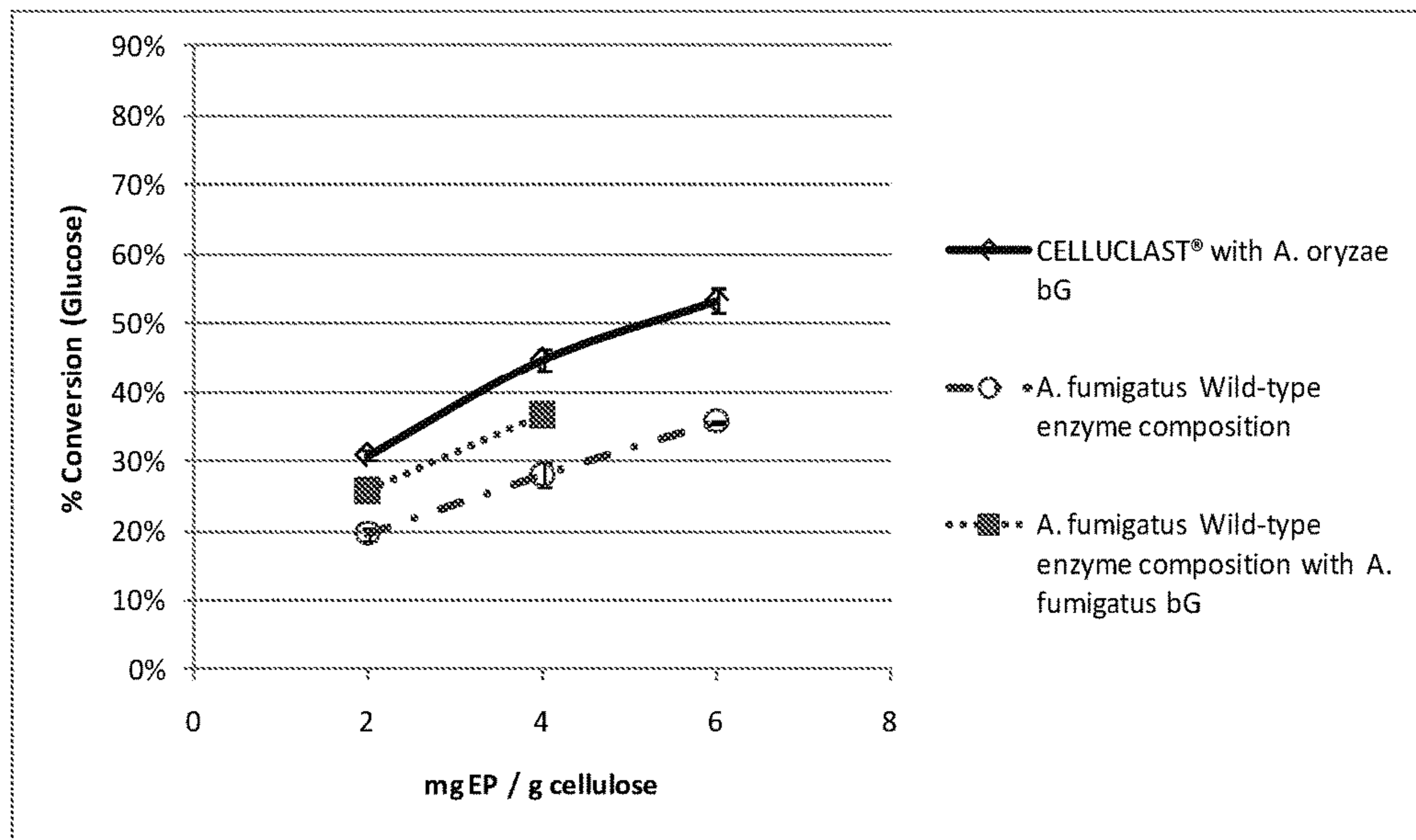


Fig. 8

**ASPERGILLUS FUMIGATUS CELLULOLYTIC
ENZYME COMPOSITIONS AND USES
THEREOF**

CROSS-REFERENCE TO RELATED
APPLICATIONS

This application is a divisional application of U.S. application Ser. No. 14/238,688 filed Feb. 12, 2014, now U.S. Pat. No. 9,663,772, which is a 35 U.S.C. § 371 national application of PCT/US2012/052161 filed Aug. 23, 2012, which claims priority or the benefit under 35 U.S.C. § 119 of U.S. Provisional Application No. 61/526,805 filed on Aug. 24, 2011, the contents of which are fully incorporated herein by reference.

STATEMENT AS TO RIGHTS TO INVENTIONS
MADE UNDER FEDERALLY SPONSORED
RESEARCH AND DEVELOPMENT

This invention was made with Government support under Cooperative Agreement DE-FC36-08GO18080 awarded by the Department of Energy. The government has certain rights in this invention.

REFERENCE TO A SEQUENCE LISTING

This application contains a Sequence Listing in computer readable form, which is incorporated herein by reference.

BACKGROUND OF THE INVENTION

Field of the Invention

The present invention relates to recombinant *Trichoderma* host cells producing *Aspergillus fumigatus* cellulolytic enzyme compositions and methods of producing and using the compositions.

Description of the Related Art

Cellulose is a polymer of glucose linked by beta-1,4-bonds. Many microorganisms produce enzymes that hydrolyze beta-linked glucans. These enzymes include endoglucanases, cellobiohydrolases, and beta-glucosidases. Endoglucanases digest the cellulose polymer at random locations, opening it to attack by cellobiohydrolases. Cellobiohydrolases sequentially release molecules of cellobiose from the ends of the cellulose polymer. Cellobiose is a water-soluble beta-1,4-linked dimer of glucose. Beta-glucosidases hydrolyze cellobiose to glucose.

The conversion of lignocellulosic feedstocks into ethanol has the advantages of the ready availability of large amounts of feedstock, the desirability of avoiding burning or land filling the materials, and the cleanliness of the ethanol fuel. Wood, agricultural residues, herbaceous crops, and municipal solid wastes have been considered as feedstocks for ethanol production. These materials primarily consist of cellulose, hemicellulose, and lignin. Once the cellulose is converted to glucose, the glucose is easily fermented by yeast into ethanol.

One of the most commercially-utilized cellulolytic enzyme systems is that of *Trichoderma reesei* (teleomorph *Hypocrea jecorina*), which secretes endoglucanases that cleave cellulose chains internally, exocellobiohydrolases that degrade cellulose chains processively from the ends, releasing cellobiose, and beta-glucosidases that hydrolyze cellobiose to glucose. *T. reesei* as a cellulolytic enzyme production system has several advantages such as high yields of secreted proteins, productive under large-scale

fermentation, a very active and well understood base cellulolytic enzyme system, and a sequenced and well annotated genome. However, disadvantages of the *T. reesei* cellulolytic enzyme system are that other organisms produce cellulolytic enzymes that are individually superior; hemicellulose-degrading enzymes are insufficient for hemicellulose-rich substrates, and beta-glucosidases are limiting under high solids hydrolysis.

Aspergillus fumigatus reportedly produces several cellulases (Nierman et al., 2005, *Nature* 438: 1151-1156). WO 2011/057140 discloses an *Aspergillus fumigatus* cellobiohydrolase I and gene thereof. WO 2011/057140 discloses an *Aspergillus fumigatus* cellobiohydrolase II and gene thereof. WO 2005/047499 discloses an *Aspergillus fumigatus* beta-glucosidase and gene thereof. WO 2010/138754 discloses an *Aspergillus fumigatus* GH61 polypeptide having cellulolytic enhancing activity and gene thereof. WO 2011/057140 discloses an *Aspergillus fumigatus* endoglucanase II and gene thereof. WO 2006/078256 discloses *Aspergillus fumigatus* GH10 xylanases. WO 2011/057140 discloses an *Aspergillus fumigatus* beta-xylosidase and gene thereof.

There is a need in the art for new cellulolytic enzyme systems that can be produced in commercial quantities and can deconstruct cellulosic material more efficiently.

The present invention provides recombinant *Trichoderma* host cells encoding *Aspergillus fumigatus* cellulolytic enzyme compositions and methods of producing and using the compositions.

SUMMARY OF THE INVENTION

The present invention relates to recombinant *Trichoderma* host cells, comprising polynucleotides encoding (i) an *Aspergillus fumigatus* cellobiohydrolase I; (ii) an *Aspergillus fumigatus* cellobiohydrolase II; (iii) an *Aspergillus fumigatus* beta-glucosidase; and (iv) an *Aspergillus fumigatus* GH61 polypeptide having cellulolytic enhancing activity; or homologs thereof.

The present invention also relates to methods of producing an enzyme composition, comprising: (a) cultivating a *Trichoderma* host cell of the present invention under conditions conducive for production of the enzyme composition; and optionally (b) recovering the enzyme composition.

The present invention also relates to enzyme compositions comprising a recovered fermentation broth of a recombinant *Trichoderma* host cell of the present invention.

The present invention also relates to processes for degrading a cellulosic material, comprising: treating the cellulosic material with an enzyme composition comprising a recovered fermentation broth of the present invention.

The present invention also relates to processes for producing a fermentation product, comprising: (a) saccharifying a cellulosic material with an enzyme composition comprising a recovered fermentation broth of the present invention; (b) fermenting the saccharified cellulosic material with one or more (e.g., several) fermenting microorganisms to produce the fermentation product; and (c) recovering the fermentation product from the fermentation.

The present invention further relates to processes of fermenting a cellulosic material, comprising: fermenting the cellulosic material with one or more (e.g., several) fermenting microorganisms, wherein the cellulosic material is saccharified with an enzyme composition comprising a recovered fermentation broth of the present invention.

BRIEF DESCRIPTION OF THE FIGURES

FIG. 1 shows a restriction map of plasmid pJfyS139.
FIG. 2 shows a restriction map of plasmid pJfyS142.

FIG. 3 shows a restriction map of plasmid pJfyS144.

FIG. 4 shows a restriction map of plasmid pAG43.

FIG. 5 shows a restriction map of plasmid pSMai214.

FIG. 6 shows a restriction map of plasmid pDM287.

FIG. 7 shows a comparison of an enzyme composition of a *Trichoderma reesei* strain expressing an *Aspergillus fumigatus* cellobiohydrolase I, *A. fumigatus* cellobiohydrolase II, *A. fumigatus* beta-glucosidase, and *A. fumigatus* GH61B polypeptide with a *Trichoderma reesei*-based cellulase composition or an *Aspergillus fumigatus* enzyme composition in the hydrolysis of pretreated corn stover.

FIG. 8 shows a comparison of an *Aspergillus fumigatus* wild-type enzyme composition with a *Trichoderma reesei*-based cellulase composition in the hydrolysis of pretreated corn stover.

DEFINITIONS

Acetylxylylan esterase: The term “acetylxylylan esterase” means a carboxylesterase (EC 3.1.1.72) that catalyzes the hydrolysis of acetyl groups from polymeric xylan, acetylated xylose, acetylated glucose, alpha-naphthyl acetate, and p-nitrophenyl acetate. For purposes of the present invention, acetylxylylan esterase activity is determined using 0.5 mM p-nitrophenylacetate as substrate in 50 mM sodium acetate pH 5.0 containing 0.01% TWEEN™ 20 (polyoxyethylene sorbitan monolaurate). One unit of acetylxylylan esterase is defined as the amount of enzyme capable of releasing 1 μmole of p-nitrophenolate anion per minute at pH 5, 25° C.

Allelic variant: The term “allelic variant” means any of two or more (e.g., several) alternative forms of a gene occupying the same chromosomal locus. Allelic variation arises naturally through mutation, and may result in polymorphism within populations. Gene mutations can be silent (no change in the encoded polypeptide) or may encode polypeptides having altered amino acid sequences. An allelic variant of a polypeptide is a polypeptide encoded by an allelic variant of a gene.

Alpha-L-arabinofuranosidase: The term “alpha-L-arabinofuranosidase” means an alpha-L-arabinofuranoside arabinofuranohydrolase (EC 3.2.1.55) that catalyzes the hydrolysis of terminal non-reducing alpha-L-arabinofuranoside residues in alpha-L-arabinosides. The enzyme acts on alpha-L-arabinofuranosides, alpha-L-arabinans containing (1,3)- and/or (1,5)-linkages, arabinoxylans, and arabinogalactans. Alpha-L-arabinofuranosidase is also known as arabinosidase, alpha-arabinosidase, alpha-L-arabinosidase, alpha-arabinofuranosidase, polysaccharide alpha-L-arabinofuranosidase, alpha-L-arabinofuranoside hydrolase, L-arabinosidase, or alpha-L-arabinanase. For purposes of the present invention, alpha-L-arabinofuranosidase activity is determined using 5 mg of medium viscosity wheat arabinoxylan (Megazyme International Ireland, Ltd., Bray, Co. Wicklow, Ireland) per ml of 100 mM sodium acetate pH 5 in a total volume of 200 μl for 30 minutes at 40° C. followed by arabinose analysis by AMINEX® HPX-87H column chromatography (Bio-Rad Laboratories, Inc., Hercules, Calif., USA).

Alpha-glucuronidase: The term “alpha-glucuronidase” means an alpha-D-glucuronate glucuronohydrolase (EC 3.2.1.139) that catalyzes the hydrolysis of an alpha-D-glucuronoside to D-glucuronate and an alcohol. For purposes of the present invention, alpha-glucuronidase activity is determined according to de Vries, 1998, *J. Bacteriol.* 180: 243-249. One unit of alpha-glucuronidase equals the amount of enzyme capable of releasing 1 μmole of glucuronic or 4-O-methylglucuronic acid per minute at pH 5, 40° C.

Aspartic protease: The term “aspartic protease” means a protease that uses an aspartate residue(s) for catalyzing the hydrolysis of peptide bonds in peptides and proteins. Aspartic proteases are a family of protease enzymes that use an aspartate residue for catalytic hydrolysis of their peptide substrates. In general, they have two highly-conserved aspartates in the active site and are optimally active at acidic pH (Szecsi, 1992, *Scand. J. Clin. Lab. In vest. Suppl.* 210: 5-22). For purposes of the present invention, aspartic protease activity is determined according to the procedure described by Aikawa et al., 2001, *J. Biochem.* 129: 791-794.

Beta-glucosidase: The term “beta-glucosidase” means a beta-D-glucoside glucohydrolase (E.C. 3.2.1.21) that catalyzes the hydrolysis of terminal non-reducing beta-D-glucose residues with the release of beta-D-glucose. For purposes of the present invention, beta-glucosidase activity is determined using p-nitrophenyl-beta-D-glucopyranoside as substrate according to the procedure of Venturi et al., 2002, *Extracellular beta-D-glucosidase from Chaetomium thermophilum var. coprophilum: production, purification and some biochemical properties*, *J. Basic Microbiol.* 42: 55-66. One unit of beta-glucosidase is defined as 1.0 μmole of p-nitrophenolate anion produced per minute at 25° C., pH 4.8 from 1 mM p-nitrophenyl-beta-D-glucopyranoside as substrate in 50 mM sodium citrate containing 0.01% TWEEN® 20 (polyoxyethylene sorbitan monolaurate).

Beta-xylosidase: The term “beta-xylosidase” means a beta-D-xyloside xylohydrolase (E.C. 3.2.1.37) that catalyzes the exo-hydrolysis of short beta (1→4)-xylooligosaccharides to remove successive D-xylose residues from non-reducing termini. For purposes of the present invention, one unit of beta-xylosidase is defined as 1.0 μmole of p-nitrophenolate anion produced per minute at 40° C., pH 5 from 1 mM p-nitrophenyl-beta-D-xyloside as substrate in 100 mM sodium citrate containing 0.01% TWEEN® 20.

cDNA: The term “cDNA” means a DNA molecule that can be prepared by reverse transcription from a mature, spliced, mRNA molecule obtained from a eukaryotic or prokaryotic cell. cDNA lacks intron sequences that may be present in the corresponding genomic DNA. The initial, primary RNA transcript is a precursor to mRNA that is processed through a series of steps, including splicing, before appearing as mature spliced mRNA.

Cellobiohydrolase: The term “cellobiohydrolase” means a 1,4-beta-D-glucan cellobiohydrolase (E.C. 3.2.1.91 and E.C. 3.2.1.176) that catalyzes the hydrolysis of 1,4-beta-D-glucosidic linkages in cellulose, cellooligosaccharides, or any beta-1,4-linked glucose containing polymer, releasing cellobiose from the reducing or non-reducing ends of the chain (Teeri, 1997, *Crystalline cellulose degradation: New insight into the function of cellobiohydrolases*, *Trends in Biotechnology* 15: 160-167; Teeri et al., 1998, *Trichoderma reesei cellobiohydrolases: why so efficient on crystalline cellulose?*, *Biochem. Soc. Trans.* 26: 173-178). Cellobiohydrolase activity is determined according to the procedures described by Lever et al., 1972, *Anal. Biochem.* 47: 273-279; van Tilbeurgh et al., 1982, *FEBS Letters*, 149: 152-156; van Tilbeurgh and Claeysens, 1985, *FEBS Letters*, 187: 283-288; and Tomme et al., 1988, *Eur. J. Biochem.* 170: 575-581. In the present invention, the Tomme et al. method can be used to determine cellobiohydrolase activity.

Cellulolytic enzyme or cellulase: The term “cellulolytic enzyme” or “cellulase” means one or more (e.g., several) enzymes that hydrolyze a cellulosic material. Such enzymes include endoglucanase(s), cellobiohydrolase(s), beta-glucosidase(s), or combinations thereof. The two basic approaches for measuring cellulolytic activity include: (1)

measuring the total cellulolytic activity, and (2) measuring the individual cellulolytic activities (endoglucanases, cellobiohydrolases, and beta-glucosidases) as reviewed in Zhang et al., Outlook for cellulase improvement: Screening and selection strategies, 2006, *Biotechnology Advances* 24: 452-481. Total cellulolytic activity is usually measured using insoluble substrates, including Whatman No 1 filter paper, microcrystalline cellulose, bacterial cellulose, algal cellulose, cotton, pretreated lignocellulose, etc. The most common total cellulolytic activity assay is the filter paper assay using Whatman No 1 filter paper as the substrate. The assay was established by the International Union of Pure and Applied Chemistry (IUPAC) (Ghose, 1987, Measurement of cellulase activities, *Pure Appl. Chem.* 59: 257-68).

For purposes of the present invention, cellulolytic enzyme activity is determined by measuring the increase in hydrolysis of a cellulosic material by cellulolytic enzyme(s) under the following conditions: 1-50 mg of cellulolytic enzyme protein/g of cellulose in PCS (or other pretreated cellulosic material) for 3-7 days at a suitable temperature, e.g., 50° C., 55° C., or 60° C., compared to a control hydrolysis without addition of cellulolytic enzyme protein. Typical conditions are 1 ml reactions, washed or unwashed PCS, 5% insoluble solids, 50 mM sodium acetate pH 5, 1 mM MnSO₄, 50° C., 55° C., or 60° C., 72 hours, sugar analysis by AMINEX® HPX-87H column (Bio-Rad Laboratories, Inc., Hercules, Calif., USA).

Cellulosic material: The term "cellulosic material" means any material containing cellulose. The predominant polysaccharide in the primary cell wall of biomass is cellulose, the second most abundant is hemicellulose, and the third is pectin. The secondary cell wall, produced after the cell has stopped growing, also contains polysaccharides and is strengthened by polymeric lignin covalently cross-linked to hemicellulose. Cellulose is a homopolymer of anhydrocellobiose and thus a linear beta-(1-4)-D-glucan, while hemicelluloses include a variety of compounds, such as xylans, xyloglucans, arabinoxylans, and mannans in complex branched structures with a spectrum of substituents. Although generally polymorphous, cellulose is found in plant tissue primarily as an insoluble crystalline matrix of parallel glucan chains. Hemicelluloses usually hydrogen bond to cellulose, as well as to other hemicelluloses, which help stabilize the cell wall matrix.

Cellulose is generally found, for example, in the stems, leaves, hulls, husks, and cobs of plants or leaves, branches, and wood of trees. The cellulosic material can be, but is not limited to, agricultural residue, herbaceous material (including energy crops), municipal solid waste, pulp and paper mill residue, waste paper, and wood (including forestry residue) (see, for example, Wiseloge et al., 1995, in Handbook on Bioethanol (Charles E. Wyman, editor), pp. 105-118, Taylor & Francis, Washington D.C.; Wyman, 1994, *Bioresource Technology* 50: 3-16; Lynd, 1990, *Applied Biochemistry and Biotechnology* 24/25: 695-719; Mosier et al., 1999, Recent Progress in Bioconversion of Lignocelluloses, in *Advances in Biochemical Engineering/Biotechnology*, T. Scheper, managing editor, Volume 65, pp. 23-40, Springer-Verlag, New York). It is understood herein that the cellulose may be in the form of lignocellulose, a plant cell wall material containing lignin, cellulose, and hemicellulose in a mixed matrix. In a preferred aspect, the cellulosic material is any biomass material. In another preferred aspect, the cellulosic material is lignocellulose, which comprises cellulose, hemicelluloses, and lignin.

In one aspect, the cellulosic material is agricultural residue. In another aspect, the cellulosic material is herbaceous

material (including energy crops). In another aspect, the cellulosic material is municipal solid waste. In another aspect, the cellulosic material is pulp and paper mill residue. In another aspect, the cellulosic material is waste paper. In another aspect, the cellulosic material is wood (including forestry residue).

In another aspect, the cellulosic material is arundo. In another aspect, the cellulosic material is bagasse. In another aspect, the cellulosic material is bamboo. In another aspect, the cellulosic material is corn cob. In another aspect, the cellulosic material is corn fiber. In another aspect, the cellulosic material is corn stover. In another aspect, the cellulosic material is miscanthus. In another aspect, the cellulosic material is orange peel. In another aspect, the cellulosic material is rice straw. In another aspect, the cellulosic material is switchgrass. In another aspect, the cellulosic material is wheat straw.

In another aspect, the cellulosic material is aspen. In another aspect, the cellulosic material is eucalyptus. In another aspect, the cellulosic material is fir. In another aspect, the cellulosic material is pine. In another aspect, the cellulosic material is poplar. In another aspect, the cellulosic material is spruce. In another aspect, the cellulosic material is willow.

In another aspect, the cellulosic material is algal cellulose. In another aspect, the cellulosic material is bacterial cellulose. In another aspect, the cellulosic material is cotton linter. In another aspect, the cellulosic material is filter paper. In another aspect, the cellulosic material is microcrystalline cellulose. In another aspect, the cellulosic material is phosphoric-acid treated cellulose.

In another aspect, the cellulosic material is an aquatic biomass. As used herein the term "aquatic biomass" means biomass produced in an aquatic environment by a photosynthesis process. The aquatic biomass can be algae, emergent plants, floating-leaf plants, or submerged plants.

The cellulosic material may be used as is or may be subjected to pretreatment, using conventional methods known in the art, as described herein. In a preferred aspect, the cellulosic material is pretreated.

Coding sequence: The term "coding sequence" means a polynucleotide, which directly specifies the amino acid sequence of a polypeptide. The boundaries of the coding sequence are generally determined by an open reading frame, which begins with a start codon such as ATG, GTG, or TTG and ends with a stop codon such as TAA, TAG, or TGA. The coding sequence may be a genomic DNA, cDNA, synthetic DNA, or a combination thereof.

Control sequences: The term "control sequences" means nucleic acid sequences necessary for expression of a polynucleotide encoding a polypeptide. Each control sequence may be native (i.e., from the same gene) or foreign (i.e., from a different gene) to the polynucleotide encoding the polypeptide or native or foreign to each other. Such control sequences include, but are not limited to, a leader, polyadenylation sequence, propeptide sequence, promoter, signal peptide sequence, and transcription terminator. At a minimum, the control sequences include a promoter, and transcriptional and translational stop signals. The control sequences may be provided with linkers for the purpose of introducing specific restriction sites facilitating ligation of the control sequences with the coding region of the polynucleotide encoding a polypeptide.

Endoglucanase: The term "endoglucanase" means an endo-1,4-(1,3;1,4)-beta-D-glucan 4-glucanohydrolase (E.C. 3.2.1.4) that catalyzes endohydrolysis of 1,4-beta-D-glycosidic linkages in cellulose, cellulose derivatives (such as

carboxymethyl cellulose and hydroxyethyl cellulose), lichenin, beta-1,4 bonds in mixed beta-1,3 glucans such as cereal beta-D-glucans or xyloglucans, and other plant material containing cellulosic components. Endoglucanase activity can be determined by measuring reduction in substrate viscosity or increase in reducing ends determined by a reducing sugar assay (Zhang et al., 2006, *Biotechnology Advances* 24: 452-481). For purposes of the present invention, endoglucanase activity is determined using carboxymethyl cellulose (CMC) as substrate according to the procedure of Ghose, 1987, *Pure and Appl. Chem.* 59: 257-268, at pH 5, 40° C.

Expression: The term “expression” includes any step involved in the production of a polypeptide including, but not limited to, transcription, post-transcriptional modification, translation, post-translational modification, and secretion.

Expression vector: The term “expression vector” means a linear or circular DNA molecule that comprises a polynucleotide encoding a polypeptide and is operably linked to control sequences that provide for its expression.

Family 61 glycoside hydrolase: The term “Family 61 glycoside hydrolase” or “Family GH61” or “GH61” means a polypeptide falling into the glycoside hydrolase Family 61 according to Henrissat B., 1991, A classification of glycosyl hydrolases based on amino-acid sequence similarities, *Biochem. J.* 280: 309-316, and Henrissat B., and Bairoch A., 1996, Updating the sequence-based classification of glycosyl hydrolases, *Biochem. J.* 316: 695-696. The enzymes in this family were originally classified as a glycoside hydrolase family based on measurement of very weak endo-1,4-beta-D-glucanase activity in one family member. The structure and mode of action of these enzymes are non-canonical and they cannot be considered as bona fide glycosidases. However, they are kept in the CAZy classification on the basis of their capacity to enhance the breakdown of lignocellulose when used in conjunction with a cellulase or a mixture of cellulases.

Feruloyl esterase: The term “feruloyl esterase” means a 4-hydroxy-3-methoxycinnamoyl-sugar hydrolase (EC 3.1.1.73) that catalyzes the hydrolysis of 4-hydroxy-3-methoxycinnamoyl (feruloyl) groups from esterified sugar, which is usually arabinose in natural biomass substrates, to produce ferulate (4-hydroxy-3-methoxycinnamate). Feruloyl esterase is also known as ferulic acid esterase, hydroxycinnamoyl esterase, FAE-III, cinnamoyl ester hydrolase, FAEA, cinnAE, FAE-I, or FAE-II. For purposes of the present invention, feruloyl esterase activity is determined using 0.5 mM p-nitrophenylferulate as substrate in 50 mM sodium acetate pH 5.0. One unit of feruloyl esterase equals the amount of enzyme capable of releasing 1 μmole of p-nitrophenolate anion per minute at pH 5, 25° C.

Flanking: The term “flanking” means DNA sequences extending on either side of a specific DNA sequence, locus, or gene. The flanking DNA is immediately adjacent to another DNA sequence, locus, or gene that is to be integrated into the genome of a filamentous fungal cell.

Fragment: The term “fragment” means a polypeptide having one or more (e.g., several) amino acids absent from the amino and/or carboxyl terminus of a mature polypeptide main; wherein the fragment has enzyme activity. In one aspect, a fragment contains at least 85%, e.g., at least 90% or at least 95% of the amino acid residues of the mature polypeptide of an enzyme.

Hemicellulolytic enzyme or hemicellulase: The term “hemicellulolytic enzyme” or “hemicellulase” means one or more (e.g., several) enzymes that hydrolyze a hemicellulosic

material. See, for example, Shallom, D. and Shoham, Y. Microbial hemicellulases. *Current Opinion In Microbiology*, 2003, 6(3): 219-228). Hemicellulases are key components in the degradation of plant biomass. Examples of hemicellulases include, but are not limited to, an acetylmannan esterase, an acetylxylan esterase, an arabinanase, an arabinofuranosidase, a coumaric acid esterase, a feruloyl esterase, a galactosidase, a glucuronidase, a glucuronoyl esterase, a mannanase, a mannosidase, a xylanase, and a xylosidase. The substrates of these enzymes, the hemicelluloses, are a heterogeneous group of branched and linear polysaccharides that are bound via hydrogen bonds to the cellulose microfibrils in the plant cell wall, crosslinking them into a robust network. Hemicelluloses are also covalently attached to lignin, forming together with cellulose a highly complex structure. The variable structure and organization of hemicelluloses require the concerted action of many enzymes for its complete degradation. The catalytic modules of hemicellulases are either glycoside hydrolases (GHs) that hydrolyze glycosidic bonds, or carbohydrate esterases (CEs), which hydrolyze ester linkages of acetate or ferulic acid side groups. These catalytic modules, based on homology of their primary sequence, can be assigned into GH and CE families. Some families, with an overall similar fold, can be further grouped into clans, marked alphabetically (e.g., GH-A). A most informative and updated classification of these and other carbohydrate active enzymes is available in the Carbohydrate-Active Enzymes (CAZy) database. Hemicellulolytic enzyme activities can be measured according to Ghose and Bisaria, 1987, *Pure & Appl. Chem.* 59: 1739-1752, at a suitable temperature, e.g., 50° C., 55° C., or 60° C., and pH, e.g., 5.0 or 5.5.

High stringency conditions: The term “high stringency conditions” means for probes of at least 100 nucleotides in length, prehybridization and hybridization at 42° C. in 5×SSPE, 0.3% SDS, 200 micrograms/ml sheared and denatured salmon sperm DNA, and 50% formamide, following standard Southern blotting procedures for 12 to 24 hours. The carrier material is finally washed three times each for 15 minutes using 2×SSC, 0.2% SDS at 65° C.

Homologous 3' or 5' region: The term “homologous 3' region” means a fragment of DNA that is identical in sequence or has a sequence identity of at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% to a region in the genome and when combined with a homologous 5' region can target integration of a piece of DNA to a specific site in the genome by homologous recombination. The term “homologous 5' region” means a fragment of DNA that is identical in sequence to a region in the genome and when combined with a homologous 3' region can target integration of a piece of DNA to a specific site in the genome by homologous recombination. The homologous 5' and 3' regions must be linked in the genome which means they are on the same chromosome and within at least 200 kb of one another.

Homologous flanking region: The term “homologous flanking region” means a fragment of DNA that is identical or has a sequence identity of at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% to a region in the genome and

is located immediately upstream or downstream of a specific site in the genome into which extracellular DNA is targeted for integration.

Homologous repeat: The term “homologous repeat” means a fragment of DNA that is repeated at least twice in the recombinant DNA introduced into a host cell and which can facilitate the loss of the DNA, i.e., selectable marker that is inserted between two homologous repeats, by homologous recombination. A homologous repeat is also known as a direct repeat.

Host cell: The term “host cell” means any cell type that is susceptible to transformation, transfection, transduction, or the like with a nucleic acid construct or expression vector comprising a polynucleotide encoding a polypeptide. The term “host cell” encompasses any progeny of a parent cell that is not identical to the parent cell due to mutations that occur during replication.

Isolated: The term “isolated” means a substance in a form or environment that does not occur in nature. Non-limiting examples of isolated substances include (1) any non-naturally occurring substance, (2) any substance including, but not limited to, any enzyme, variant, nucleic acid, protein, peptide or cofactor, that is at least partially removed from one or more or all of the naturally occurring constituents with which it is associated in nature; (3) any substance modified by the hand of man relative to that substance found in nature; or (4) any substance modified by increasing the amount of the substance relative to other components with which it is naturally associated (e.g., recombinant production in a host cell; multiple copies of a gene encoding the substance; and use of a stronger promoter than the promoter naturally associated with the gene encoding the substance).

Low stringency conditions: The term “low stringency conditions” means for probes of at least 100 nucleotides in length, prehybridization and hybridization at 42° C. in 5×SSPE, 0.3% SDS, 200 micrograms/ml sheared and denatured salmon sperm DNA, and 25% formamide, following standard Southern blotting procedures for 12 to 24 hours. The carrier material is finally washed three times each for 15 minutes using 2×SSC, 0.2% SDS at 50° C.

Mature polypeptide: The term “mature polypeptide” means a polypeptide in its final form following translation and any post-translational modifications, such as N-terminal processing, C-terminal truncation, glycosylation, phosphorylation, etc. In one aspect, the mature polypeptide of an *A. fumigatus* cellobiohydrolase I is amino acids 27 to 532 of SEQ ID NO: 2 based on the SignalP program (Nielsen et al., 1997, *Protein Engineering* 10: 1-6) that predicts amino acids 1 to 26 of SEQ ID NO: 2 are a signal peptide. In another aspect, the mature polypeptide of an *A. fumigatus* cellobiohydrolase II is amino acids 20 to 454 of SEQ ID NO: 4 based on the SignalP program that predicts amino acids 1 to 19 of SEQ ID NO: 4 are a signal peptide. In another aspect, the mature polypeptide of an *A. fumigatus* beta-glucosidase is amino acids 20 to 863 of SEQ ID NO: 6 based on the SignalP program that predicts amino acids 1 to 19 of SEQ ID NO: 6 are a signal peptide. In another aspect, the mature polypeptide of an *A. fumigatus* GH61 polypeptide is amino acids 22 to 250 of SEQ ID NO: 8 based on the SignalP program that predicts amino acids 1 to 21 of SEQ ID NO: 8 are a signal peptide. In another aspect, the mature polypeptide of an *A. fumigatus* endoglucanase I is amino acids 19 to 407 of SEQ ID NO: 10 based on the SignalP program that predicts amino acids 1 to 18 of SEQ ID NO: 10 are a signal peptide. In another aspect, the mature polypeptide of an *A. fumigatus* endoglucanase II is amino acids 19 to 329 of SEQ ID NO: 12 based on the SignalP program that predicts amino

acids 1 to 18 of SEQ ID NO: 12 are a signal peptide. In another aspect, the mature polypeptide of an *A. fumigatus* xylanase I is amino acids 18 to 364 of SEQ ID NO: 14 based on the SignalP program that predicts amino acids 1 to 17 of SEQ ID NO: 14 are a signal peptide. In another aspect, the mature polypeptide of an *A. fumigatus* xylanase II is amino acids 20 to 323 of SEQ ID NO: 16 based on the SignalP program that predicts amino acids 1 to 19 of SEQ ID NO: 16 are a signal peptide. In another aspect, the mature polypeptide of an *A. fumigatus* xylanase III is amino acids 20 to 397 of SEQ ID NO: 18 based on the SignalP program that predicts amino acids 1 to 19 of SEQ ID NO: 18 are a signal peptide. In another aspect, the mature polypeptide of an *A. fumigatus* beta-xylosidase is amino acids 21 to 792 of SEQ ID NO: 20 based on the SignalP program that predicts amino acids 1 to 20 of SEQ ID NO: 20 are a signal peptide. In another aspect, the mature polypeptide of an *A. fumigatus* swollenin is amino acids 18 to 470 of SEQ ID NO: 22 based on the SignalP program that predicts amino acids 1 to 17 of SEQ ID NO: 22 are a signal peptide.

In another aspect, the mature polypeptide of a *T. reesei* cellobiohydrolase I is amino acids 18 to 514 of SEQ ID NO: 24 based on the SignalP program that predicts amino acids 1 to 17 of SEQ ID NO: 24 are a signal peptide. In another aspect, the mature polypeptide of a *T. reesei* cellobiohydrolase II is amino acids 19 to 471 of SEQ ID NO: 26 based on the SignalP program that predicts amino acids 1 to 18 of SEQ ID NO: 26 are a signal peptide. In another aspect, the mature polypeptide of a *T. reesei* beta-glucosidase is amino acids 20 to 744 of SEQ ID NO: 28 based on the SignalP program that predicts amino acids 1 to 19 of SEQ ID NO: 28 are a signal peptide. In another aspect, the mature polypeptide of a *T. reesei* endoglucanase I is amino acids 23 to 459 of SEQ ID NO: 30 based on the SignalP program that predicts amino acids 1 to 22 of SEQ ID NO: 30 are a signal peptide. In another aspect, the mature polypeptide of a *T. reesei* endoglucanase II is amino acids 22 to 418 of SEQ ID NO: 32 based on the SignalP program that predicts amino acids 1 to 21 of SEQ ID NO: 32 are a signal peptide. In another aspect, the mature polypeptide of a *T. reesei* xylanase I is amino acids 20 to 229 of SEQ ID NO: 34 based on the SignalP program that predicts amino acids 1 to 19 of SEQ ID NO: 34 are a signal peptide. In another aspect, the mature polypeptide of a *T. reesei* xylanase II is amino acids 20 to 223 of SEQ ID NO: 36 based on the SignalP program that predicts amino acids 1 to 19 of SEQ ID NO: 36 are a signal peptide. In another aspect, the mature polypeptide of a *T. reesei* xylanase III is amino acids 17 to 347 of SEQ ID NO: 38 based on the SignalP program that predicts amino acids 1 to 16 of SEQ ID NO: 38 are a signal peptide. In another aspect, the mature polypeptide of a *T. reesei* beta-xylosidase is amino acids 21 to 797 of SEQ ID NO: 40 based on the SignalP program that predicts amino acids 1 to 20 of SEQ ID NO: 40 are a signal peptide. In another aspect, the mature polypeptide of a *T. reesei* swollenin is amino acids 19 to 493 of SEQ ID NO: 42 based on the SignalP program that predicts amino acids 1 to 18 of SEQ ID NO: 42 are a signal peptide. It is known in the art that a host cell may produce a mixture of two or more different mature polypeptides (i.e., with a different C-terminal and/or N-terminal amino acid) expressed by the same polynucleotide.

Mature polypeptide coding sequence: The term “mature polypeptide coding sequence” means a polynucleotide that encodes a mature polypeptide having enzyme activity. In one aspect, the mature polypeptide coding sequence of an *A. fumigatus* cellobiohydrolase I is nucleotides 79 to 1596 of SEQ ID NO: 1 or the cDNA sequence thereof based on the

SignalP program (Nielsen et al., 1997, supra) that predicts nucleotides 1 to 78 of SEQ ID NO: 1 encode a signal peptide. In another aspect, the mature polypeptide coding sequence of an *A. fumigatus* cellobiohydrolase II is nucleotides 58 to 1700 of SEQ ID NO: 3 or the cDNA sequence thereof based on the SignalP program that predicts nucleotides 1 to 57 of SEQ ID NO: 3 encode a signal peptide. In another aspect, the mature polypeptide coding sequence of an *A. fumigatus* beta-glucosidase is nucleotides 58 to 2580 of SEQ ID NO: 5 or the cDNA sequence thereof based on the SignalP program that predicts nucleotides 1 to 57 of SEQ ID NO: 5 encode a signal peptide. In another aspect, the mature polypeptide coding sequence of an *A. fumigatus* GH61 polypeptide is nucleotides 64 to 859 of SEQ ID NO: 7 or the cDNA sequence thereof based on the SignalP program that predicts nucleotides 1 to 63 of SEQ ID NO: 7 encode a signal peptide. In another aspect, the mature polypeptide coding sequence of an *A. fumigatus* endoglucanase I is nucleotides 55 to 1221 of SEQ ID NO: 9 or the cDNA sequence thereof based on the SignalP program that predicts nucleotides 1 to 54 of SEQ ID NO: 9 encode a signal peptide. In another aspect, the mature polypeptide coding sequence of an *A. fumigatus* endoglucanase II is nucleotides 55 to 1248 of SEQ ID NO: 11 or the cDNA sequence thereof based on the SignalP program that predicts nucleotides 1 to 54 of SEQ ID NO: 11 encode a signal peptide. In another aspect, the mature polypeptide coding sequence of an *A. fumigatus* xylanase I is nucleotides 52 to 1145 of SEQ ID NO: 13 or the cDNA sequence thereof based on the SignalP program that predicts nucleotides 1 to 51 of SEQ ID NO: 13 encode a signal peptide. In another aspect, the mature polypeptide coding sequence of an *A. fumigatus* xylanase II is nucleotides 58 to 1400 of SEQ ID NO: 15 or the cDNA sequence thereof based on the SignalP program that predicts nucleotides 1 to 57 of SEQ ID NO: 15 encode a signal peptide. In another aspect, the mature polypeptide coding sequence of an *A. fumigatus* xylanase III is nucleotides 107 to 1415 of SEQ ID NO: 17 or the cDNA sequence thereof based on the SignalP program that predicts nucleotides 1 to 106 of SEQ ID NO: 17 encode a signal peptide. In another aspect, the mature polypeptide coding sequence of an *A. fumigatus* beta-xylosidase is nucleotides 61 to 2373 of SEQ ID NO: 19 or the cDNA sequence thereof based on the SignalP program that predicts nucleotides 1 to 60 of SEQ ID NO: 19 encode a signal peptide. In another aspect, the mature polypeptide coding sequence of an *A. fumigatus* swollenin is nucleotides 52 to 1657 of SEQ ID NO: 21 or the cDNA sequence thereof based on the SignalP program that predicts nucleotides 1 to 51 of SEQ ID NO: 21 encode a signal peptide.

In another aspect, the mature polypeptide coding sequence of a *T. reesei* cellobiohydrolase I is nucleotides 52 to 1545 of SEQ ID NO: 23 or the cDNA sequence thereof based on the SignalP program that predicts nucleotides 1 to 51 of SEQ ID NO: 23 encode a signal peptide. In another aspect, the mature polypeptide coding sequence of a *T. reesei* cellobiohydrolase II is nucleotides 55 to 1608 of SEQ ID NO: 25 or the cDNA sequence thereof based on the SignalP program that predicts nucleotides 1 to 54 of SEQ ID NO: 25 encode a signal peptide. In another aspect, the mature polypeptide coding sequence of a *T. reesei* beta-glucosidase is nucleotides 58 to 2612 of SEQ ID NO: 27 or the cDNA sequence thereof based on the SignalP program that predicts nucleotides 1 to 57 of SEQ ID NO: 27 encode a signal peptide. In another aspect, the mature polypeptide coding sequence of a *T. reesei* endoglucanase I is nucleotides 67 to 1374 of SEQ ID NO: 29 or the cDNA sequence thereof

based on the SignalP program that predicts nucleotides 1 to 66 of SEQ ID NO: 29 encode a signal peptide. In another aspect, the mature polypeptide coding sequence of a *T. reesei* endoglucanase II is nucleotides 64 to 1254 of SEQ ID NO: 31 or the cDNA sequence thereof based on the SignalP program that predicts nucleotides 1 to 63 of SEQ ID NO: 31 encode a signal peptide. In another aspect, the mature polypeptide coding sequence of a *T. reesei* xylanase I is nucleotides 58 to 749 of SEQ ID NO: 33 or the cDNA sequence thereof based on the SignalP program that predicts nucleotides 1 to 57 of SEQ ID NO: 33 encode a signal peptide. In another aspect, the mature polypeptide coding sequence of a *T. reesei* xylanase II is nucleotides 58 to 778 of SEQ ID NO: 35 or the cDNA sequence thereof based on the SignalP program that predicts nucleotides 1 to 57 of SEQ ID NO: 35 encode a signal peptide. In another aspect, the mature polypeptide coding sequence of a *T. reesei* xylanase III is nucleotides 49 to 1349 of SEQ ID NO: 37 or the cDNA sequence thereof based on the SignalP program that predicts nucleotides 1 to 48 of SEQ ID NO: 37 encode a signal peptide. In another aspect, the mature polypeptide coding sequence of a *T. reesei* beta-xylosidase is nucleotides 61 to 2391 of SEQ ID NO: 39 or the cDNA sequence thereof based on the SignalP program that predicts nucleotides 1 to 60 of SEQ ID NO: 39 encode a signal peptide. In another aspect, the mature polypeptide coding sequence of a *T. reesei* swollenin is nucleotides 55 to 2776 of SEQ ID NO: 41 or the cDNA sequence thereof based on the SignalP program that predicts nucleotides 1 to 54 of SEQ ID NO: 41 encode a signal peptide.

Medium stringency conditions: The term "medium stringency conditions" means for probes of at least 100 nucleotides in length, prehybridization and hybridization at 42° C. in 5×SSPE, 0.3% SDS, 200 micrograms/ml sheared and denatured salmon sperm DNA, and 35% formamide, following standard Southern blotting procedures for 12 to 24 hours. The carrier material is finally washed three times each for 15 minutes using 2×SSC, 0.2% SDS at 55° C.

Medium-high stringency conditions: The term "medium-high stringency conditions" means for probes of at least 100 nucleotides in length, prehybridization and hybridization at 42° C. in 5×SSPE, 0.3% SDS, 200 micrograms/ml sheared and denatured salmon sperm DNA, and 35% formamide, following standard Southern blotting procedures for 12 to 24 hours. The carrier material is finally washed three times each for 15 minutes using 2×SSC, 0.2% SDS at 60° C.

Nucleic acid construct: The term "nucleic acid construct" means a nucleic acid molecule, either single- or double-stranded, which is isolated from a naturally occurring gene or is modified to contain segments of nucleic acids in a manner that would not otherwise exist in nature or which is synthetic, which comprises one or more (e.g., several) control sequences.

Operably linked: The term "operably linked" means a configuration in which a control sequence is placed at an appropriate position relative to the coding sequence of a polynucleotide such that the control sequence directs expression of the coding sequence.

Polypeptide having cellulolytic enhancing activity: The term "polypeptide having cellulolytic enhancing activity" means a GH61 polypeptide that catalyzes the enhancement of the hydrolysis of a cellulosic material by enzyme having cellulolytic activity. For purposes of the present invention, cellulolytic enhancing activity is determined by measuring the increase in reducing sugars or the increase of the total of cellobiose and glucose from the hydrolysis of a cellulosic material by cellulolytic enzyme under the following condi-

tions: 1-50 mg of total protein/g of cellulose in PCS, wherein total protein is comprised of 50-99.5% w/w cellulolytic enzyme protein and 0.5-50% w/w protein of a GH61 polypeptide having cellulolytic enhancing activity for 1-7 days at a suitable temperature, e.g., 50° C., 55° C., or 60° C., and pH, e.g., 5.0 or 5.5, compared to a control hydrolysis with equal total protein loading without cellulolytic enhancing activity (1-50 mg of cellulolytic protein/g of cellulose in PCS). In a preferred aspect, a mixture of CELLUCLAST® 1.5 L (Novozymes A/S, Bagsvaerd, Denmark) in the presence of 2-3% of total protein weight *Aspergillus oryzae* beta-glucosidase (recombinantly produced in *Aspergillus oryzae* according to WO 02/095014) or 2-3% of total protein weight *Aspergillus fumigatus* beta-glucosidase (recombinantly produced in *Aspergillus oryzae* as described in WO 2002/095014) of cellulase protein loading is used as the source of the cellulolytic activity.

The GH61 polypeptides having cellulolytic enhancing activity enhance the hydrolysis of a cellulosic material catalyzed by enzyme having cellulolytic activity by reducing the amount of cellulolytic enzyme required to reach the same degree of hydrolysis preferably at least 1.01-fold, e.g., at least 1.05-fold, at least 1.10-fold, at least 1.25-fold, at least 1.5-fold, at least 2-fold, at least 3-fold, at least 4-fold, at least 5-fold, at least 10-fold, or at least 20-fold.

Pretreated corn stover: The term "PCS" or "Pretreated Corn Stover" means a cellulosic material derived from corn stover by treatment with heat and dilute sulfuric acid, alkaline pretreatment, or neutral pretreatment.

Sequence identity: The relatedness between two amino acid sequences or between two nucleotide sequences is described by the parameter "sequence identity".

For purposes of the present invention, the sequence identity between two amino acid sequences is determined using the Needleman-Wunsch algorithm (Needleman and Wunsch, 1970, *J. Mol. Biol.* 48: 443-453) as implemented in the Needle program of the EMBOSS package (EMBOSS: The European Molecular Biology Open Software Suite, Rice et al., 2000, *Trends Genet.* 16: 276-277), preferably version 5.0.0 or later. The parameters used are gap open penalty of 10, gap extension penalty of 0.5, and the EBLOSUM62 (EMBOSS version of BLOSUM62) substitution matrix. The output of Needle labeled "longest identity" (obtained using the -nobrief option) is used as the percent identity and is calculated as follows:

$$\frac{(\text{Identical Residues} \times 100)}{(\text{Length of Alignment} - \text{Total Number of Gaps in Alignment})}$$

For purposes of the present invention, the sequence identity between two deoxyribonucleotide sequences is determined using the Needleman-Wunsch algorithm (Needleman and Wunsch, 1970, supra) as implemented in the Needle program of the EMBOSS package (EMBOSS: The European Molecular Biology Open Software Suite, Rice et al., 2000, supra), preferably version 5.0.0 or later. The parameters used are gap open penalty of 10, gap extension penalty of 0.5, and the EDNAFULL (EMBOSS version of NCBI NUC4.4) substitution matrix. The output of Needle labeled "longest identity" (obtained using the -nobrief option) is used as the percent identity and is calculated as follows:

$$\frac{(\text{Identical Deoxyribonucleotides} \times 100)}{(\text{Length of Alignment} - \text{Total Number of Gaps in Alignment})}$$

Subsequence: The term "subsequence" means a polynucleotide having one or more (e.g., several) nucleotides absent from the 5' and/or 3' end of a mature polypeptide coding sequence; wherein the subsequence encodes a frag-

ment having enzyme activity. In one aspect, a subsequence contains at least 85%, e.g., at least 90% or at least 95% of the nucleotides of the mature polypeptide coding sequence of an enzyme.

5 Subtilisin-like serine protease: The term "subtilisin-like serine protease" means a protease with a substrate specificity similar to subtilisin that uses a serine residue for catalyzing the hydrolysis of peptide bonds in peptides and proteins. Subtilisin-like proteases (subtilases) are serine proteases characterized by a catalytic triad of the three amino acids aspartate, histidine, and serine. The arrangement of these catalytic residues is shared with the prototypical subtilisin from *Bacillus licheniformis* (Siezen and Leunissen, 1997, *Protein Science* 6: 501-523). Subtilisin-like serine protease activity can be determined using a synthetic substrate, N-succinyl-L-Ala-L-Ala-L-Pro-L-Phe-p-nitroanilide (AAPF) (Bachem AG, Bubendorf, Switzerland) in 100 mM NaCl-100 mM MOPS pH 7.0 at 50° C. for 3 hours and then the absorbance at 405 nm is measured.

20 Targeted integration: The term "targeted integration" means the stable integration of extracellular DNA at a defined genomic locus.

Transformant: The term "transformant" means a cell which has taken up extracellular DNA (foreign, artificial or modified) and expresses the gene(s) contained therein.

25 Transformation: The term "transformation" means the introduction of extracellular DNA into a cell, i.e., the genetic alteration of a cell resulting from the direct uptake, incorporation and expression of exogenous genetic material (exogenous DNA) from its surroundings and taken up through the cell membrane(s).

Transformation efficiency: The term "transformation efficiency" means the efficiency by which cells can take up the extracellular DNA and express the gene(s) contained therein, which is calculated by dividing the number of positive transformants expressing the gene(s) by the amount of DNA used during a transformation procedure.

35 Trypsin-like serine protease: The term "trypsin-like serine protease" means a protease with a substrate specificity similar to trypsin that uses a serine residue for catalyzing the hydrolysis of peptide bonds in peptides and proteins. For purposes of the present invention, trypsin-like serine protease activity is determined according to the procedure described by Dienes et al., 2007, *Enzyme and Microbial Technology* 40: 1087-1094.

40 Variant: The term "variant" means a polypeptide having enzyme activity comprising an alteration, i.e., a substitution, insertion, and/or deletion, at one or more (e.g., several) positions. A substitution means replacement of the amino acid occupying a position with a different amino acid; a deletion means removal of the amino acid occupying a position; and an insertion means adding an amino acid adjacent to and immediately following the amino acid occupying a position.

55 Very high stringency conditions: The term "very high stringency conditions" means for probes of at least 100 nucleotides in length, prehybridization and hybridization at 42° C. in 5×SSPE, 0.3% SDS, 200 micrograms/ml sheared and denatured salmon sperm DNA, and 50% formamide, following standard Southern blotting procedures for 12 to 24 hours. The carrier material is finally washed three times each for 15 minutes using 2×SSC, 0.2% SDS at 70° C.

65 Very low stringency conditions: The term "very low stringency conditions" means for probes of at least 100 nucleotides in length, prehybridization and hybridization at 42° C. in 5×SSPE, 0.3% SDS, 200 micrograms/ml sheared and denatured salmon sperm DNA, and 25% formamide,

following standard Southern blotting procedures for 12 to 24 hours. The carrier material is finally washed three times each for 15 minutes using 2×SSC, 0.2% SDS at 45° C.

Xylan-containing material: The term “xylan-containing material” means any material comprising a plant cell wall polysaccharide containing a backbone of beta-(1-4)-linked xylose residues. Xylans of terrestrial plants are heteropolymers possessing a beta-(1-4)-D-xylopyranose backbone, which is branched by short carbohydrate chains. They comprise D-glucuronic acid or its 4-O-methyl ether, L-arabinose, and/or various oligosaccharides, composed of D-xylose, L-arabinose, D- or L-galactose, and D-glucose. Xylan-type polysaccharides can be divided into homoxylans and heteroxylans, which include glucuronoxylans, (arabino)glucuronoxylans, (glucurono)arabinoxylans, arabinoxylans, and complex heteroxylans. See, for example, Ebringerova et al., 2005, *Adv. Polym. Sci.* 186: 1-67.

In the processes of the present invention, any material containing xylan may be used. In a preferred aspect, the xylan-containing material is lignocellulose.

Xylan degrading activity or xylanolytic activity: The term “xylan degrading activity” or “xylanolytic activity” means a biological activity that hydrolyzes xylan-containing material. The two basic approaches for measuring xylanolytic activity include: (1) measuring the total xylanolytic activity, and (2) measuring the individual xylanolytic activities (e.g., endoxylanases, beta-xylosidases, arabinofuranosidases, alpha-glucuronidases, acetylxyylan esterases, feruloyl esterases, and alpha-glucuronyl esterases). Recent progress in assays of xylanolytic enzymes was summarized in several publications including Biely and Puchard, Recent progress in the assays of xylanolytic enzymes, 2006, *Journal of the Science of Food and Agriculture* 86(11): 1636-1647; Spanikova and Biely, 2006, Glucuronoyl esterase—Novel carbohydrate esterase produced by *Schizophyllum commune*, *FEBS Letters* 580(19): 4597-4601; Herrmann, Vrsanska, Jurickova, Hirsch, Biely, and Kubicek, 1997, The beta-D-xylosidase of *Trichoderma reesei* is a multifunctional beta-D-xylan xylohydrolase, *Biochemical Journal* 321: 375-381.

Total xylan degrading activity can be measured by determining the reducing sugars formed from various types of xylan, including, for example, oat spelt, beechwood, and larchwood xylans, or by photometric determination of dyed xylan fragments released from various covalently dyed xylans. The most common total xylanolytic activity assay is based on production of reducing sugars from polymeric 4-O-methyl glucuronoxylan as described in Bailey, Biely, Poutanen, 1992, Interlaboratory testing of methods for assay of xylanase activity, *Journal of Biotechnology* 23(3): 257-270. Xylanase activity can also be determined with 0.2% AZCL-arabinoxylan as substrate in 0.01% TRITON® X-100 (4-(1,1,3,3-tetramethylbutyl)phenyl-polyethylene glycol) and 200 mM sodium phosphate buffer pH 6 at 37° C. One unit of xylanase activity is defined as 1.0 μmole of azurine produced per minute at 37° C., pH 6 from 0.2% AZCL-arabinoxylan as substrate in 200 mM sodium phosphate pH 6 buffer.

For purposes of the present invention, xylan degrading activity is determined by measuring the increase in hydrolysis of birchwood xylan (Sigma Chemical Co., Inc., St. Louis, Mo., USA) by xylan-degrading enzyme(s) under the following typical conditions: 1 ml reactions, 5 mg/ml substrate (total solids), 5 mg of xylanolytic protein/g of substrate, 50 mM sodium acetate pH 5, 50° C., 24 hours, sugar analysis using p-hydroxybenzoic acid hydrazide (PHBAH)

assay as described by Lever, 1972, A new reaction for colorimetric determination of carbohydrates, *Anal. Biochem* 47: 273-279.

Xylanase: The term “xylanase” means a 1,4-beta-D-xylan-xylohydrolase (E.C. 3.2.1.8) that catalyzes the endohydrolysis of 1,4-beta-D-xylosidic linkages in xylans. For purposes of the present invention, xylanase activity is determined with 0.2% AZCL-arabinoxylan as substrate in 0.01% TRITON® X-100 and 200 mM sodium phosphate buffer pH 6 at 37° C. One unit of xylanase activity is defined as 1.0 μmole of azurine produced per minute at 37° C., pH 6 from 0.2% AZCL-arabinoxylan as substrate in 200 mM sodium phosphate pH 6 buffer.

DETAILED DESCRIPTION OF THE INVENTION

The present invention relates to recombinant *Trichoderma* host cells, comprising polynucleotides encoding (i) an *Aspergillus fumigatus* cellobiohydrolase I; (ii) an *Aspergillus fumigatus* cellobiohydrolase II; (iii) an *Aspergillus fumigatus* beta-glucosidase; and (iv) an *Aspergillus fumigatus* GH61 polypeptide having cellulolytic enhancing activity; or homologs thereof. In one aspect, the recombinant *Trichoderma* host cells further comprise one or more (e.g., several) polynucleotides encoding one or more (e.g., several) enzymes selected from the group consisting of: (i) an *Aspergillus fumigatus* endoglucanase I; (ii) an *Aspergillus fumigatus* endoglucanase II; (iii) an *Aspergillus fumigatus* xylanase; (iv) an *Aspergillus fumigatus* beta-xylosidase; and (v) an *Aspergillus fumigatus* swollenin; or homologs thereof.

The recombinant *Trichoderma* host cells of the present invention unexpectedly produce enzyme compositions of *Aspergillus fumigatus* cellulolytic enzymes that are more efficient in the deconstruction of cellulosic material than a native cellulolytic enzyme composition produced by *T. reesei* or *A. fumigatus*.

Aspergillus fumigatus Cellulases and Hemicellulases

In the present invention, any *Aspergillus fumigatus* cellobiohydrolase I, *Aspergillus fumigatus* cellobiohydrolase II, *Aspergillus fumigatus* beta-glucosidase, *Aspergillus fumigatus* GH61 polypeptide having cellulolytic enhancing activity, *Aspergillus fumigatus* endoglucanase I, *Aspergillus fumigatus* endoglucanase II, *Aspergillus fumigatus* xylanase, *Aspergillus fumigatus* beta-xylosidase, and *Aspergillus fumigatus* swollenin, or homologs thereof, may be used.

In one aspect, the *Aspergillus fumigatus* cellobiohydrolase I or a homolog thereof is selected from the group consisting of: (i) a cellobiohydrolase I comprising or consisting of the mature polypeptide of SEQ ID NO: 2; (ii) a cellobiohydrolase I comprising or consisting of an amino acid sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide of SEQ ID NO: 2; (iii) a cellobiohydrolase I encoded by a polynucleotide comprising or consisting of a nucleotide sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide coding sequence of SEQ ID NO: 1; and (iv) a cellobiohydrolase I encoded by a polynucleotide that hybrid-

a xylanase comprising or consisting of the mature polypeptide of SEQ ID NO: 14, SEQ ID NO: 16, or SEQ ID NO: 18; (ii) a xylanase comprising or consisting of an amino acid sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide of SEQ ID NO: 14, SEQ ID NO: 16, or SEQ ID NO: 18; (iii) a xylanase encoded by a polynucleotide comprising or consisting of a nucleotide sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide coding sequence of SEQ ID NO: 13, SEQ ID NO: 15, or SEQ ID NO: 17; and (iv) a xylanase encoded by a polynucleotide that hybridizes under at least high stringency conditions, e.g., very high stringency conditions, with the mature polypeptide coding sequence of SEQ ID NO: 13, SEQ ID NO: 15, or SEQ ID NO: 17; or the full-length complement thereof.

In another aspect, the *Aspergillus fumigatus* beta-xylosidase or a homolog thereof is selected from the group consisting of: (i) a beta-xylosidase comprising or consisting of the mature polypeptide of SEQ ID NO: 20; (ii) a beta-xylosidase comprising or consisting of an amino acid sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide of SEQ ID NO: 20; (iii) a beta-xylosidase encoded by a polynucleotide comprising or consisting of a nucleotide sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide coding sequence of SEQ ID NO: 19; and (iv) a beta-xylosidase encoded by a polynucleotide that hybridizes under at least high stringency conditions, e.g., very high stringency conditions, with the mature polypeptide coding sequence of SEQ ID NO: 19 or the full-length complement thereof.

In another aspect, the *Aspergillus fumigatus* swollenin or a homolog thereof is selected from the group consisting of: (i) a swollenin comprising or consisting of the mature polypeptide of SEQ ID NO: 22; (ii) a swollenin comprising or consisting of an amino acid sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide of SEQ ID NO: 22; (iii) a swollenin encoded by a polynucleotide comprising or consisting of a nucleotide sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence

identity to the mature polypeptide coding sequence of SEQ ID NO: 21; and (iv) a swollenin encoded by a polynucleotide that hybridizes under at least high stringency conditions, e.g., very high stringency conditions, with the mature polypeptide coding sequence of SEQ ID NO: 21 or the full-length complement thereof.

The polynucleotide of SEQ ID NO: 1, 3, 5, 7, 9, 11, 13, 15, 17, 19, or 21, or a subsequence thereof, as well as the polypeptide of SEQ ID NO: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, or 22, or a fragment thereof, may be used to design nucleic acid probes to identify and clone DNA encoding enzymes according to methods well known in the art. In particular, such probes can be used for hybridization with the genomic DNA or cDNA of a cell of interest, following standard Southern blotting procedures, in order to identify and isolate the corresponding gene therein. Such probes can be considerably shorter than the entire sequence, but should be at least 15, e.g., at least 25, at least 35, or at least 70 nucleotides in length. Preferably, the nucleic acid probe is at least 100 nucleotides in length, e.g., at least 200 nucleotides, at least 300 nucleotides, at least 400 nucleotides, at least 500 nucleotides, at least 600 nucleotides, at least 700 nucleotides, at least 800 nucleotides, or at least 900 nucleotides in length. Both DNA and RNA probes can be used. The probes are typically labeled for detecting the corresponding gene (for example, with ^{32}P , ^3H , ^{35}S , biotin, or avidin). Such probes are encompassed by the present invention.

A genomic DNA or cDNA library may be screened for DNA that hybridizes with the probes described above and encodes an enzyme. Genomic or other DNA may be separated by agarose or polyacrylamide gel electrophoresis, or other separation techniques. DNA from the libraries or the separated DNA may be transferred to and immobilized on nitrocellulose or other suitable carrier material. In order to identify a clone or DNA that hybridizes with SEQ ID NO: 1, 3, 5, 7, 9, 11, 13, 15, 17, 19, or 21, or a subsequence thereof, the carrier material is used in a Southern blot.

For purposes of the present invention, hybridization indicates that the polynucleotide hybridizes to a labeled nucleic acid probe corresponding to (i) SEQ ID NO: 1, 3, 5, 7, 9, 11, 13, 15, 17, 19, or 21; (ii) the mature polypeptide coding sequence of SEQ ID NO: 1, 3, 5, 7, 9, 11, 13, 15, 17, 19, or 21; (iii) the cDNA sequence thereof; (iv) the full-length complement thereof; or (v) a subsequence thereof; under very low to very high stringency conditions. Molecules to which the nucleic acid probe hybridizes under these conditions can be detected using, for example, X-ray film or any other detection means known in the art.

In one aspect, the nucleic acid probe is SEQ ID NO: 1, 3, 5, 7, 9, 11, 13, 15, 17, 19, or 21, or the mature polypeptide coding sequence thereof. In another aspect, the nucleic acid probe is a polynucleotide that encodes the polypeptide of SEQ ID NO: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, or 22; the mature polypeptide thereof; or a fragment thereof.

The techniques used to isolate or clone a polynucleotide are known in the art and include isolation from genomic DNA or cDNA, or a combination thereof. The cloning of the polynucleotides from genomic DNA can be effected, e.g., by using the well known polymerase chain reaction (PCR) or antibody screening of expression libraries to detect cloned DNA fragments with shared structural features. See, e.g., Innis et al., 1990, PCR: A Guide to Methods and Application, Academic Press, New York. Other nucleic acid amplification procedures such as ligase chain reaction (LCR), ligation activated transcription (LAT) and polynucleotide-based amplification (NASBA) may be used. The polynucleotides may be cloned from a strain of *Aspergillus fumigatus*

and thus, for example, may be an allelic or species variant of the polypeptide encoding region of the polynucleotide.

A chemically modified or protein engineered mutant of an *A. fumigatus* enzyme above (or protein) may also be used.

An *A. fumigatus* enzyme may also be hybrid enzyme in which a region of the *A. fumigatus* enzyme is fused at the N-terminus or the C-terminus of a region of another enzyme.

An *A. fumigatus* enzyme may further be a fusion polypeptide or cleavable fusion polypeptide in which another enzyme is fused at the N-terminus or the C-terminus of the *A. fumigatus* enzyme. A fusion polypeptide is produced by fusing a polynucleotide encoding another enzyme to a polynucleotide encoding the *A. fumigatus* enzyme. Techniques for producing fusion polypeptides are known in the art, and include ligating the coding sequences encoding the polypeptides so that they are in frame and that expression of the fusion polypeptide is under control of the same promoter(s) and terminator. Fusion polypeptides may also be constructed using intein technology in which fusion polypeptides are created post-translationally (Cooper et al., 1993, *EMBO J.* 12: 2575-2583; Dawson et al., 1994, *Science* 266: 776-779).

A fusion polypeptide can further comprise a cleavage site between the two polypeptides. Upon secretion of the fusion protein, the site is cleaved releasing the two polypeptides. Examples of cleavage sites include, but are not limited to, the sites disclosed in Martin et al., 2003, *J. Ind. Microbiol. Biotechnol.* 3: 568-576; Svetina et al., 2000, *J. Biotechnol.* 76: 245-251; Rasmussen-Wilson et al., 1997, *Appl. Environ. Microbiol.* 63: 3488-3493; Ward et al., 1995, *Biotechnology* 13: 498-503; and Contreras et al., 1991, *Biotechnology* 9: 378-381; Eaton et al., 1986, *Biochemistry* 25: 505-512; Collins-Racie et al., 1995, *Biotechnology* 13: 982-987; Carter et al., 1989, *Proteins: Structure, Function, and Genetics* 6: 240-248; and Stevens, 2003, *Drug Discovery World* 4: 35-48.

The *Aspergillus fumigatus* enzyme may be further one or more (e.g., several) *A. fumigatus* enzymes selected from the group consisting of an acetylmannan esterase, an acetylxylo-esterase, an arabinanase, an arabinofuranosidase, a coumaric acid esterase, a feruloyl esterase, a galactosidase, a glucuronidase, a glucuronoyl esterase, a mannanase, and a mannosidase.

The *Aspergillus fumigatus* enzyme may be even further one or more (e.g., several) *A. fumigatus* enzymes selected from the group consisting of an esterase, an expansin, a laccase, a ligninolytic enzyme, a pectinase, a peroxidase, and a protease.

Trichoderma Host Cells

The *Trichoderma* host cell may be any *Trichoderma* cell useful in the recombinant production of an enzyme or protein. For example, the *Trichoderma* cell may be a *Trichoderma harzianum*, *Trichoderma koningii*, *Trichoderma longibrachiatum*, *Trichoderma reesei*, or *Trichoderma viride* cell. In one aspect, the *Trichoderma* cell is a *Trichoderma harzianum* cell. In another aspect, the *Trichoderma* cell is a *Trichoderma koningii* cell. In another aspect, the *Trichoderma* cell is a *Trichoderma longibrachiatum* cell. In another aspect, the *Trichoderma* cell is a *Trichoderma reesei* cell. In another aspect, the *Trichoderma* cell is a *Trichoderma viride* cell.

In another aspect, the *Trichoderma reesei* cell is *Trichoderma reesei* RutC30. In another aspect, the *Trichoderma reesei* cell is *Trichoderma reesei* TV10. In another aspect, the *Trichoderma reesei* cell is a mutant of *Trichoderma reesei* RutC30. In another aspect, the *Trichoderma reesei* cell is mutant of *Trichoderma reesei*

TV10. In another aspect, the *Trichoderma reesei* cell is a morphological mutant of *Trichoderma reesei*. See, for example, WO 97/26330, which is incorporated herein by reference in its entirety.

The *Trichoderma* cell may be transformed by a process involving protoplast formation, transformation of the protoplasts, and regeneration of the cell wall in a manner known per se. Suitable procedures for transformation of *Trichoderma* host cells are described in EP 238023, Yelton et al., 1984, *Proc. Natl. Acad. Sci. USA* 81: 1470-1474, and Christensen et al., 1988, *Bio/Technology* 6: 1419-1422.

One or more (e.g., several) native cellulase and/or hemicellulase genes may be inactivated in the *Trichoderma* host cell by disrupting or deleting the genes, or a portion thereof, which results in the mutant cell producing less or none of the cellulase and/or hemicellulase than the parent cell when cultivated under the same conditions. In one aspect, the one or more (e.g., several) cellulase genes encode enzymes selected from the group consisting of cellobiohydrolase I, cellobiohydrolase II, endoglucanase I, endoglucanase II, beta-glucosidase, and swollenin. In another aspect, the one or more (e.g., several) hemicellulase genes encode enzymes selected from the group consisting of xylanase I, xylanase II, xylanase III, and beta-xylosidase. In another aspect, the one or more (e.g., several) hemicellulase genes encode enzymes selected from the group consisting of an acetylmannan esterase, an acetylxylo-esterase, an arabinanase, an arabinofuranosidase, a coumaric acid esterase, a feruloyl esterase, a galactosidase, a glucuronidase, a glucuronoyl esterase, a mannanase, and a mannosidase.

The mutant cell may be constructed by reducing or eliminating expression of a polynucleotide encoding a *Trichoderma* cellulase or hemicellulase using methods well known in the art, for example, insertions, disruptions, replacements, or deletions. In a preferred aspect, the polynucleotide is inactivated. The polynucleotide to be modified or inactivated may be, for example, the coding region or a part thereof essential for activity, or a regulatory element required for expression of the coding region. An example of such a regulatory or control sequence may be a promoter sequence or a functional part thereof, i.e., a part that is sufficient for affecting expression of the polynucleotide. Other control sequences for possible modification include, but are not limited to, a leader, polyadenylation sequence, propeptide sequence, signal peptide sequence, transcription terminator, and transcriptional activator.

Modification or inactivation of the polynucleotide may be performed by subjecting the parent cell to mutagenesis and selecting for mutant cells in which expression of the polynucleotide has been reduced or eliminated. The mutagenesis, which may be specific or random, may be performed, for example, by use of a suitable physical or chemical mutagenizing agent, by use of a suitable oligonucleotide, or by subjecting the DNA sequence to PCR generated mutagenesis. Furthermore, the mutagenesis may be performed by use of any combination of these mutagenizing agents.

Examples of a physical or chemical mutagenizing agent suitable for the present purpose include ultraviolet (UV) irradiation, hydroxylamine, N-methyl-N'-nitro-N-nitrosoguanidine (MNNG), O-methyl hydroxylamine, nitrous acid, ethyl methane sulphonate (EMS), sodium bisulphite, formic acid, and nucleotide analogues.

When such agents are used, the mutagenesis is typically performed by incubating the parent cell to be mutagenized in the presence of the mutagenizing agent of choice under suitable conditions, and screening and/or selecting for mutant cells exhibiting reduced or no expression of the gene.

Modification or inactivation of the polynucleotide may also be accomplished by insertion, substitution, or deletion of one or more (e.g., several) nucleotides in the gene or a regulatory element required for transcription or translation thereof. For example, nucleotides may be inserted or removed so as to result in the introduction of a stop codon, the removal of the start codon, or a change in the open reading frame. Such modification or inactivation may be accomplished by site-directed mutagenesis or PCR generated mutagenesis in accordance with methods known in the art. Although, in principle, the modification may be performed in vivo, i.e., directly on the cell expressing the polynucleotide to be modified, it is preferred that the modification be performed in vitro as exemplified below.

An example of a convenient way to eliminate or reduce expression of a polynucleotide is based on techniques of gene replacement, gene deletion, or gene disruption. For example, in the gene disruption method, a nucleic acid sequence corresponding to the endogenous polynucleotide is mutagenized in vitro to produce a defective nucleic acid sequence that is then transformed into the parent cell to produce a defective gene. By homologous recombination, the defective nucleic acid sequence replaces the endogenous polynucleotide. It may be desirable that the defective polynucleotide also encodes a marker that may be used for selection of transformants in which the polynucleotide has been modified or destroyed. In an aspect, the polynucleotide is disrupted with a selectable marker such as those described herein.

Modification or inactivation of the polynucleotide may also be accomplished by inhibiting expression of an enzyme encoded by the polynucleotide in a cell by administering to the cell or expressing in the cell a double-stranded RNA (dsRNA) molecule, wherein the dsRNA comprises a subsequence of a polynucleotide encoding the enzyme. In a preferred aspect, the dsRNA is about 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25 or more duplex nucleotides in length.

The dsRNA is preferably a small interfering RNA (siRNA) or a micro RNA (miRNA). In a preferred aspect, the dsRNA is small interfering RNA for inhibiting transcription. In another preferred aspect, the dsRNA is micro RNA for inhibiting translation. In another aspect, the double-stranded RNA (dsRNA) molecules comprise a portion of the mature polypeptide coding sequence of SEQ ID NO: 23, SEQ ID NO: 25, SEQ ID NO: 27, SEQ ID NO: 29, SEQ ID NO: 31, SEQ ID NO: 33, SEQ ID NO: 35, SEQ ID NO: 37, SEQ ID NO: 39, and/or SEQ ID NO: 41 for inhibiting expression of the polypeptide in a cell. While the present invention is not limited by any particular mechanism of action, the dsRNA can enter a cell and cause the degradation of a single-stranded RNA (ssRNA) of similar or identical sequences, including endogenous mRNAs. When a cell is exposed to dsRNA, mRNA from the homologous gene is selectively degraded by a process called RNA interference (RNAi).

The dsRNAs can be used in gene-silencing to selectively degrade RNA using a dsRNAi of the present invention. The process may be practiced in vitro, ex vivo or in vivo. In one aspect, the dsRNA molecules can be used to generate a loss-of-function mutation in a cell, an organ or an animal. Methods for making and using dsRNA molecules to selectively degrade RNA are well known in the art; see, for example, U.S. Pat. Nos. 6,489,127; 6,506,559; 6,511,824; and 6,515,109.

In one aspect, the *Trichoderma* cellobiohydrolase I or a homolog thereof is selected from the group consisting of: (i) a cellobiohydrolase I comprising or consisting of the mature

polypeptide of SEQ ID NO: 24; (ii) a cellobiohydrolase I comprising or consisting of an amino acid sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide of SEQ ID NO: 24; (iii) a cellobiohydrolase I encoded by a polynucleotide comprising or consisting of a nucleotide sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide coding sequence of SEQ ID NO: 23; and (iv) a cellobiohydrolase I encoded by a polynucleotide that hybridizes under at least high stringency conditions, e.g., very high stringency conditions, with the mature polypeptide coding sequence of SEQ ID NO: 23 or the full-length complement thereof.

In another aspect, the *Trichoderma* cellobiohydrolase II or a homolog thereof is selected from the group consisting of: (i) a cellobiohydrolase II comprising or consisting of the mature polypeptide of SEQ ID NO: 26; (ii) a cellobiohydrolase II comprising or consisting of an amino acid sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide of SEQ ID NO: 26; (iii) a cellobiohydrolase II encoded by a polynucleotide comprising or consisting of a nucleotide sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide coding sequence of SEQ ID NO: 25; and (iv) a cellobiohydrolase II encoded by a polynucleotide that hybridizes under at least high stringency conditions, e.g., very high stringency conditions, with the mature polypeptide coding sequence of SEQ ID NO: 25 or the full-length complement thereof.

In another aspect, the *Trichoderma* beta-glucosidase or a homolog thereof is selected from the group consisting of: (i) a beta-glucosidase comprising or consisting of the mature polypeptide of SEQ ID NO: 28; (ii) a beta-glucosidase comprising or consisting of an amino acid sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide of SEQ ID NO: 28; (iii) a beta-glucosidase encoded by a polynucleotide comprising or consisting of a nucleotide sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide coding sequence of SEQ ID NO: 27; and (iv) a beta-glucosidase encoded by a polynucleotide that hybridizes under at least high stringency conditions, e.g., very high stringency conditions, with the mature polypeptide coding sequence of SEQ ID NO: 27 or the full-length complement thereof.

least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide coding sequence of SEQ ID NO: 39; and (iv) a beta-xylosidase encoded by a polynucleotide that hybridizes under at least high stringency conditions, e.g., very high stringency conditions, with the mature polypeptide coding sequence of SEQ ID NO: 39 or the full-length complement thereof.

In another aspect, the *Trichoderma* swollenin or a homolog thereof is selected from the group consisting of: (i) a swollenin comprising or consisting of the mature polypeptide of SEQ ID NO: 42; (ii) a swollenin comprising or consisting of an amino acid sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide of SEQ ID NO: 42; (iii) a swollenin encoded by a polynucleotide comprising or consisting of a nucleotide sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide coding sequence of SEQ ID NO: 41; and (iv) a swollenin encoded by a polynucleotide that hybridizes under at least high stringency conditions, e.g., very high stringency conditions, with the mature polypeptide coding sequence of SEQ ID NO: 41 or the full-length complement thereof.

In one aspect, a *Trichoderma* cellobiohydrolase I gene is inactivated. In another aspect, a *Trichoderma* cellobiohydrolase II gene is inactivated. In another aspect, a *Trichoderma* endoglucanase I gene is inactivated. In another aspect, a *Trichoderma* endoglucanase II gene is inactivated. In another aspect, a *Trichoderma* beta-glucosidase gene is inactivated. In another aspect, a *Trichoderma* xylanase gene is inactivated. In another aspect, a *Trichoderma* beta-xylosidase gene is inactivated. In another aspect, a *Trichoderma* swollenin gene is inactivated.

In another aspect, a *Trichoderma* cellobiohydrolase I gene and a *Trichoderma* cellobiohydrolase II gene are inactivated.

In another aspect, two or more (e.g., several) of the genes selected from the group consisting of cellobiohydrolase I, cellobiohydrolase II, endoglucanase I, endoglucanase II, beta-glucosidase, xylanase I, xylanase II, xylanase III, beta-xylosidase, and swollenin genes are inactivated. In another aspect, three or more (e.g., several) of the genes selected from the group consisting of cellobiohydrolase I, cellobiohydrolase II, endoglucanase I, endoglucanase II, beta-glucosidase, xylanase I, xylanase II, xylanase III, beta-xylosidase, and swollenin genes are inactivated. In another aspect, four or more (e.g., several) of the genes selected from the group consisting of cellobiohydrolase I, cellobiohydrolase II, endoglucanase I, endoglucanase II, beta-glucosidase, xylanase I, xylanase II, xylanase III, beta-xylosidase, and swollenin genes are inactivated. In another aspect, five or more (e.g., several) of the genes selected from the group consisting of cellobiohydrolase I, cellobiohydrolase II, endoglucanase I, endoglucanase II, beta-glucosidase, xylanase I, xylanase II, xylanase III, beta-xylosidase, and swollenin genes are inactivated. In another aspect, six or more (e.g., several) of the genes selected from the group

consisting of cellobiohydrolase I, cellobiohydrolase II, endoglucanase I, endoglucanase II, beta-glucosidase, xylanase I, xylanase II, xylanase III, beta-xylosidase, and swollenin genes are inactivated. In another aspect, seven or more (e.g., several) of the genes selected from the group consisting of cellobiohydrolase I, cellobiohydrolase II, endoglucanase I, endoglucanase II, beta-glucosidase, xylanase I, xylanase II, xylanase III, beta-xylosidase, and swollenin genes are inactivated. In another aspect, eight or more (e.g., several) of the genes selected from the group consisting of cellobiohydrolase I, cellobiohydrolase II, endoglucanase I, endoglucanase II, beta-glucosidase, xylanase I, xylanase II, xylanase III, beta-xylosidase, and swollenin genes are inactivated. In another aspect, nine or more (e.g., several) of the genes selected from the group consisting of cellobiohydrolase I, cellobiohydrolase II, endoglucanase I, endoglucanase II, beta-glucosidase, xylanase I, xylanase II, xylanase III, beta-xylosidase, and swollenin genes are inactivated.

In another aspect, the cellobiohydrolase I, cellobiohydrolase II, endoglucanase I, endoglucanase II, beta-glucosidase, and beta-xylosidase genes are inactivated. In another aspect, the cellobiohydrolase I, cellobiohydrolase II, endoglucanase I, endoglucanase II, beta-glucosidase, xylanase I, xylanase II, xylanase III, beta-xylosidase, and swollenin genes are inactivated.

In another aspect, one or more (e.g., several) protease genes are inactivated. In another aspect, the one or more (e.g., several) protease genes are subtilisin-like serine protease, aspartic protease, and trypsin-like serine protease genes as described in WO 2011/075677, which is incorporated herein by reference in its entirety.

In each of the aspects above, the *Trichoderma* enzyme is a *Trichoderma harzianum*, *Trichoderma koningii*, *Trichoderma longibrachiatum*, *Trichoderma reesei*, or *Trichoderma viride* enzyme. In another aspect, the *Trichoderma* enzyme is a *Trichoderma harzianum* enzyme. In another aspect, the *Trichoderma* enzyme is a *Trichoderma koningii* enzyme. In another aspect, the *Trichoderma* enzyme is a *Trichoderma longibrachiatum* enzyme. In another aspect, the *Trichoderma* enzyme is a *Trichoderma reesei* cell. In another aspect, the *Trichoderma* enzyme is a *Trichoderma viride* enzyme.

Nucleic Acid Constructs

Nucleic acid constructs comprising a polynucleotide encoding an *Aspergillus fumigatus* enzyme or protein can be constructed by operably linking one or more (e.g., several) control sequences to the polynucleotide to direct the expression of the coding sequence in a *Trichoderma* host cell under conditions compatible with the control sequences. Manipulation of the polynucleotide prior to its insertion into a vector may be desirable or necessary depending on the expression vector. The techniques for modifying polynucleotides utilizing recombinant DNA methods are well known in the art.

The control sequence may be a promoter, a polynucleotide that is recognized by a *Trichoderma* host cell for expression of a polynucleotide encoding an enzyme or protein. The promoter contains transcriptional control sequences that mediate the expression of the polypeptide. The promoter may be any polynucleotide that shows transcriptional activity in the host cell including mutant, truncated, and hybrid promoters, and may be obtained from genes encoding extracellular or intracellular polypeptides either homologous or heterologous to the host cell.

Examples of suitable promoters for directing transcription of the nucleic acid constructs in a *Trichoderma* host cell are promoters obtained from the genes for *Aspergillus nidulans*

acetamidase, *Aspergillus niger* neutral alpha-amylase, *Aspergillus niger* acid stable alpha-amylase, *Aspergillus niger* or *Aspergillus awamori* glucoamylase (glaA), *Aspergillus oryzae* TAKA amylase, *Aspergillus oryzae* alkaline protease, *Aspergillus oryzae* triose phosphate isomerase, *Fusarium oxysporum* trypsin-like protease (WO 96/00787), *Fusarium venenatum* amyloglucosidase (WO 00/56900), *Fusarium venenatum* Daria (WO 00/56900), *Fusarium venenatum* Quinn (WO 00/56900), *Rhizomucor miehei* lipase, *Rhizomucor miehei* aspartic proteinase, *Trichoderma reesei* beta-glucosidase, *Trichoderma reesei* cellobiohydrolase I, *Trichoderma reesei* cellobiohydrolase II, *Trichoderma reesei* endoglucanase I, *Trichoderma reesei* endoglucanase II, *Trichoderma reesei* endoglucanase III, *Trichoderma reesei* endoglucanase V, *Trichoderma reesei* xylanase I, *Trichoderma reesei* xylanase II, *Trichoderma reesei* xylanase III, *Trichoderma reesei* beta-xylosidase, and *Trichoderma reesei* translation elongation factor, as well as the NA2-tpi promoter (a modified promoter from an *Aspergillus* neutral alpha-amylase gene in which the untranslated leader has been replaced by an untranslated leader from an *Aspergillus* triose phosphate isomerase gene; non-limiting examples include modified promoters from an *Aspergillus niger* neutral alpha-amylase gene in which the untranslated leader has been replaced by an untranslated leader from an *Aspergillus nidulans* or *Aspergillus oryzae* triose phosphate isomerase gene); and mutant, truncated, and hybrid promoters thereof. Other promoters are described in U.S. Pat. No. 6,011,147, which is incorporated herein in its entirety.

The control sequence may also be a transcription terminator, which is recognized by a *Trichoderma* host cell to terminate transcription. The terminator is operably linked to the 3'-terminus of the polynucleotide encoding the polypeptide. Any terminator that is functional in the host cell may be used in the present invention.

Preferred terminators for *Trichoderma* host cells are obtained from the genes for *Aspergillus nidulans* anthranilate synthase, *Aspergillus niger* glucoamylase, *Aspergillus niger* alpha-glucosidase, *Aspergillus oryzae* TAKA amylase, *Fusarium oxysporum* trypsin-like protease, *Trichoderma reesei* beta-glucosidase, *Trichoderma reesei* cellobiohydrolase I, *Trichoderma reesei* cellobiohydrolase II, *Trichoderma reesei* endoglucanase I, *Trichoderma reesei* endoglucanase II, *Trichoderma reesei* endoglucanase III, *Trichoderma reesei* endoglucanase V, *Trichoderma reesei* xylanase I, *Trichoderma reesei* xylanase II, *Trichoderma reesei* xylanase III, *Trichoderma reesei* beta-xylosidase, and *Trichoderma reesei* translation elongation factor.

The control sequence may also be a leader, a nontranslated region of an mRNA that is important for translation by a *Trichoderma* host cell. The leader is operably linked to the 5'-terminus of the polynucleotide encoding the polypeptide. Any leader that is functional in the host cell may be used.

Preferred leaders for *Trichoderma* host cells are obtained from the genes for *Aspergillus oryzae* TAKA amylase and *Aspergillus nidulans* triose phosphate isomerase.

The control sequence may also be a polyadenylation sequence, a sequence operably linked to the 3'-terminus of the polynucleotide and, when transcribed, is recognized by a *Trichoderma* host cell as a signal to add polyadenosine residues to transcribed mRNA. Any polyadenylation sequence that is functional in the host cell may be used.

Preferred polyadenylation sequences for *Trichoderma* host cells are obtained from the genes for *Aspergillus nidulans* anthranilate synthase, *Aspergillus niger* glucoamylase, *Aspergillus niger* alpha-glucosidase, *Aspergillus oryzae* TAKA amylase, *Fusarium oxysporum* trypsin-like protease,

Trichoderma reesei cellobiohydrolase I, *Trichoderma reesei* cellobiohydrolase II, and *Trichoderma reesei* endoglucanase V.

The control sequence may also be a signal peptide coding region that encodes a signal peptide linked to the N-terminus of a polypeptide and directs the polypeptide into a cell's secretory pathway. The 5'-end of the coding sequence of the polynucleotide may inherently contain a signal peptide coding sequence naturally linked in translation reading frame with the segment of the coding sequence that encodes the polypeptide. Alternatively, the 5'-end of the coding sequence may contain a signal peptide coding sequence that is foreign to the coding sequence. A foreign signal peptide coding sequence may be required where the coding sequence does not naturally contain a signal peptide coding sequence. Alternatively, a foreign signal peptide coding sequence may simply replace the natural signal peptide coding sequence in order to enhance secretion of the polypeptide. However, any signal peptide coding sequence that directs the expressed polypeptide into the secretory pathway of a host cell may be used.

Effective signal peptide coding sequences for *Trichoderma* host cells are the signal peptide coding sequences obtained from the genes for *Aspergillus niger* neutral amylase, *Aspergillus niger* glucoamylase, *Aspergillus oryzae* TAKA amylase, *Humicola insolens* cellulase, *Humicola insolens* endoglucanase V, *Humicola lanuginosa* lipase, *Rhizomucor miehei* aspartic proteinase, *Trichoderma reesei* cellobiohydrolase I, *Trichoderma reesei* cellobiohydrolase II, *Trichoderma reesei* endoglucanase I, *Trichoderma reesei* endoglucanase II, *Trichoderma reesei* endoglucanase III, and *Trichoderma reesei* endoglucanase V.

The control sequence may also be a propeptide coding sequence that encodes a propeptide positioned at the N-terminus of a polypeptide. The resultant polypeptide is known as a proenzyme or propolypeptide (or a zymogen in some cases). A propolypeptide is generally inactive and can be converted to an active polypeptide by catalytic or autocatalytic cleavage of the propeptide from the propolypeptide. The propeptide coding sequence may be obtained from the genes for *Myceliophthora thermophila* laccase (WO 95/33836) and *Rhizomucor miehei* aspartic proteinase.

Where both signal peptide and propeptide sequences are present, the propeptide sequence is positioned next to the N-terminus of a polypeptide and the signal peptide sequence is positioned next to the N-terminus of the propeptide sequence.

It may also be desirable to add regulatory sequences that regulate expression of the polypeptide relative to the growth of a *Trichoderma* host cell. Examples of regulatory sequences are those that cause expression of the gene to be turned on or off in response to a chemical or physical stimulus, including the presence of a regulatory compound. Regulatory sequences include the *Aspergillus niger* glucoamylase promoter, *Aspergillus oryzae* TAKA alpha-amylase promoter, *Aspergillus oryzae* glucoamylase promoter, *Trichoderma reesei* cellobiohydrolase I promoter, and *Trichoderma reesei* cellobiohydrolase II promoter. Other examples of regulatory sequences are those that allow for gene amplification. In eukaryotic systems, these regulatory sequences include the dihydrofolate reductase gene that is amplified in the presence of methotrexate, and the metallothionein genes that are amplified with heavy metals. In these cases, the polynucleotide encoding the polypeptide would be operably linked with the regulatory sequence.

Expression Vectors

Recombinant expression vectors can be constructed comprising a polynucleotide encoding an *Aspergillus fumigatus* enzyme or protein, a promoter, a terminator, and transcriptional and translational stop signals. The various nucleotide and control sequences may be joined together to produce a recombinant expression vector that may include one or more (e.g., several) convenient restriction sites to allow for insertion or substitution of the polynucleotide encoding the polypeptide at such sites. Alternatively, the polynucleotide may be expressed by inserting the polynucleotide or a nucleic acid construct comprising the polynucleotide into an appropriate vector for expression. In creating the expression vector, the coding sequence is located in the vector so that the coding sequence is operably linked with the appropriate control sequences for expression.

The recombinant expression vector may be any vector (e.g., a plasmid or virus) that can be conveniently subjected to recombinant DNA procedures and can bring about expression of the polynucleotide. The choice of the vector will typically depend on the compatibility of the vector with the host cell into which the vector is to be introduced. The vector may be a linear or closed circular plasmid.

The vector may be an autonomously replicating vector, i.e., a vector that exists as an extrachromosomal entity, the replication of which is independent of chromosomal replication, e.g., a plasmid, an extrachromosomal element, a minichromosome, or an artificial chromosome. The vector may contain any means for assuring self-replication. Alternatively, the vector may be one that, when introduced into the host cell, is integrated into the genome and replicated together with the chromosome(s) into which it has been integrated. Furthermore, a single vector or plasmid or two or more vectors or plasmids that together contain the total DNA to be introduced into the genome of the host cell, or a transposon, may be used.

The vector preferably contains one or more (e.g., several) selectable markers that permit easy selection of transformed, transfected, transduced, or the like cells. A selectable marker is a gene the product of which provides for biocide or viral resistance, resistance to heavy metals, prototrophy to auxotrophs, and the like.

Examples of selectable markers for use in a *Trichoderma* host cell include, but are not limited to, *adeA* (phosphoribosylaminoimidazole-succinocarboxamide synthase), *adeB* (phosphoribosylaminoimidazole synthase), *amdS* (acetamidase), *argB* (ornithine carbamoyltransferase), *bar* (phosphinothricin acetyltransferase), *hph* (hygromycin phosphotransferase), *niaD* (nitrate reductase), *pyrG* (orotidine-5'-phosphate decarboxylase), *sC* (sulfate adenylyltransferase), and *trpC* (anthranilate synthase), as well as equivalents thereof. Preferred for use in an *Aspergillus* cell are *Aspergillus nidulans* or *Aspergillus oryzae* *amdS* and *pyrG* genes and a *Streptomyces hygrosopicus* *bar* gene. Preferred for use in a *Trichoderma* cell are *adeA*, *adeB*, *amdS*, *hph*, and *pyrG* genes. Examples of bacterial selectable markers are markers that confer antibiotic resistance such as ampicillin, chloramphenicol, kanamycin, neomycin, spectinomycin, or tetracycline resistance.

The selectable marker may be a dual selectable marker system as described in WO 2010/039889 A2, which is incorporated herein by reference in its entirety. In one aspect, the selectable marker is a *hph*-*tk* dual selectable marker system.

The vector preferably contains an element(s) that permits integration of the vector into the host cell's genome or autonomous replication of the vector in the cell independent of the genome.

For integration into the host cell genome, the vector may rely on the polynucleotide's sequence encoding the polypeptide or any other element of the vector for integration into the genome by homologous or non-homologous recombination. Alternatively, the vector may contain additional polynucleotides for directing integration by homologous recombination into the genome of the host cell at a precise location(s) in the chromosome(s). To increase the likelihood of integration at a precise location, the integrational elements should contain a sufficient number of nucleic acids, such as 100 to 10,000 base pairs, 400 to 10,000 base pairs, and 800 to 10,000 base pairs, which have a high degree of sequence identity to the corresponding target sequence to enhance the probability of homologous recombination. The integrational elements may be any sequence that is homologous with the target sequence in the genome of the host cell. Furthermore, the integrational elements may be non-encoding or encoding polynucleotides. On the other hand, the vector may be integrated into the genome of the host cell by non-homologous recombination.

For autonomous replication, the vector may further comprise an origin of replication enabling the vector to replicate autonomously in a *Trichoderma* host cell. The origin of replication may be any plasmid replicator mediating autonomous replication that functions in a cell. The term "origin of replication" or "plasmid replicator" means a polynucleotide that enables a plasmid or vector to replicate in vivo.

Examples of origins of replication useful in a *Trichoderma* host cell are AMA1 and ANS1 (Gems et al., 1991, *Gene* 98: 61-67; Cullen et al., 1987, *Nucleic Acids Res.* 15: 9163-9175; WO 00/24883). Isolation of the AMA1 gene and construction of plasmids or vectors comprising the gene can be accomplished according to the methods disclosed in WO 00/24883.

More than one copy of a polynucleotide may be inserted into a *Trichoderma* host cell to increase production of a polypeptide. An increase in the copy number of the polynucleotide can be obtained by integrating at least one additional copy of the sequence into the host cell genome or by including an amplifiable selectable marker gene with the polynucleotide where cells containing amplified copies of the selectable marker gene, and thereby additional copies of the polynucleotide, can be selected for by cultivating the cells in the presence of the appropriate selectable agent.

The procedures used to ligate the elements described above to construct the recombinant expression vectors are well known to one skilled in the art (see, e.g., Sambrook et al., 1989, supra).

Methods of Production

The present invention also relates to methods of producing an enzyme composition comprising *Aspergillus fumigatus* cellulases and/or hemicellulases, comprising (a) cultivating a *Trichoderma* recombinant host cell of the present invention under conditions conducive for production of the enzyme composition; and (b) recovering the enzyme composition.

The *Trichoderma* host cells are cultivated in a nutrient medium suitable for production of the enzyme composition using methods known in the art. For example, the cell may be cultivated by shake flask cultivation, or small-scale or large-scale fermentation (including continuous, batch, fed-batch, or solid state fermentations) in laboratory or industrial fermentors in a suitable medium and under conditions

allowing the enzyme(s) to be expressed and/or isolated. The cultivation takes place in a suitable nutrient medium comprising carbon and nitrogen sources and inorganic salts, using procedures known in the art. Suitable media are available from commercial suppliers or may be prepared according to published compositions (e.g., in catalogues of the American Type Culture Collection).

The *A. fumigatus* cellulases and/or hemicellulases may be detected using methods known in the art that are specific for the enzyme. These detection methods include, but are not limited to, use of specific antibodies, formation of an enzyme product, or disappearance of an enzyme substrate. For example, an enzyme assay may be used to determine activity.

The *A. fumigatus* cellulases and/or hemicellulases may be recovered using methods known in the art. For example, the enzyme may be recovered from the nutrient medium by conventional procedures including, but not limited to, collection, centrifugation, filtration, extraction, spray-drying, evaporation, or precipitation. In one aspect, the whole fermentation broth is recovered.

The *A. fumigatus* cellulases and/or hemicellulases may be purified by a variety of procedures known in the art including, but not limited to, chromatography (e.g., ion exchange, affinity, hydrophobic, chromatofocusing, and size exclusion), electrophoretic procedures (e.g., preparative isoelectric focusing), differential solubility (e.g., ammonium sulfate precipitation), SDS-PAGE, or extraction (see, e.g., *Protein Purification*, Janson and Ryden, editors, VCH Publishers, New York, 1989) to obtain substantially pure polypeptides.

The present invention also relates to enzyme composition comprising a recovered fermentation broth of a recombinant *Trichoderma* host cell of the present invention. In one aspect, the enzyme composition may have one or more components of the fermentation broth removed. In another aspect, the enzyme composition may have no components of the fermentation broth removed.

The enzyme composition comprises (i) an *Aspergillus fumigatus* cellobiohydrolase I; (ii) an *Aspergillus fumigatus* cellobiohydrolase II; (iii) an *Aspergillus fumigatus* beta-glucosidase; and (iv) an *Aspergillus fumigatus* GH61 polypeptide having cellulolytic enhancing activity, or homologs thereof, as described herein. In one aspect, the enzyme composition further comprises one or more (e.g., several) enzymes selected from the group consisting of: (i) an *Aspergillus fumigatus* endoglucanase I; (ii) an *Aspergillus fumigatus* endoglucanase II; (iii) an *Aspergillus fumigatus* xylanase; (iv) an *Aspergillus fumigatus* beta-xylosidase; (v) an *Aspergillus fumigatus* swollenin; or (vi) combinations thereof; or a homolog or homologs thereof, as described herein. In another aspect, the enzyme composition further comprises an *Aspergillus fumigatus* endoglucanase I. In another aspect, the enzyme composition further comprises an *Aspergillus fumigatus* endoglucanase II. In another aspect, the enzyme composition further comprises an *Aspergillus fumigatus* xylanase. In another aspect, the enzyme composition further comprises an *Aspergillus fumigatus* beta-xylosidase. In another aspect, the enzyme composition further comprises an *Aspergillus fumigatus* swollenin.

In another aspect, the enzyme composition further comprises a *Trichoderma* endoglucanase I. In another aspect, the enzyme composition further comprises a *Trichoderma reesei* endoglucanase I. In another aspect, the enzyme composition further comprises a *Trichoderma reesei* Cel7B endoglucanase I (GENBANK™ accession no. M15665). In another aspect, the *Trichoderma reesei* endoglucanase I is native to

the host cell. In another aspect, the *Trichoderma reesei* endoglucanase I is the mature polypeptide of SEQ ID NO: 30.

In another aspect, the enzyme composition further comprises a *Trichoderma* endoglucanase II. In another aspect, the enzyme composition further comprises a *Trichoderma reesei* endoglucanase II. In another aspect, the enzyme composition further comprises a *Trichoderma reesei* Cel5A endoglucanase II (GENBANK™ accession no. M19373). In another aspect, the *Trichoderma reesei* endoglucanase II is native to the host cell. In another aspect, the *Trichoderma reesei* endoglucanase I is the mature polypeptide of SEQ ID NO: 32.

The enzyme composition may further comprise one or more (e.g., several) enzymes selected from the group consisting of a cellulase, a GH61 polypeptide having cellulolytic enhancing activity, a hemicellulase, an esterase, an expansin, a laccase, a ligninolytic enzyme, a pectinase, a peroxidase, a protease, and a swollenin. In another aspect, the cellulase is preferably one or more (e.g., several) enzymes selected from the group consisting of an endoglucanase, a cellobiohydrolase, and a beta-glucosidase. In another aspect, the hemicellulase is preferably one or more (e.g., several) enzymes selected from the group consisting of an acetylmannan esterase, an acetylxylan esterase, an arabinanase, an arabinofuranosidase, a coumaric acid esterase, a feruloyl esterase, a galactosidase, a glucuronidase, a glucuronoyl esterase, a mannanase, a mannosidase, a xylanase, and a xylosidase. The enzyme(s) may be native or foreign to the *Trichoderma* host cell.

The term "fermentation broth" as used herein refers to a preparation produced by cellular fermentation that undergoes no or minimal recovery and/or purification. For example, fermentation broths are produced when microbial cultures are grown to saturation, incubated under carbon-limiting conditions to allow protein synthesis (e.g., expression of enzymes by host cells) and secretion into cell culture medium. The fermentation broth can contain unfractionated or fractionated contents of the fermentation materials derived at the end of the fermentation. Typically, the fermentation broth is unfractionated and comprises the spent culture medium and cell debris present after the microbial cells (e.g., filamentous fungal cells) are removed, e.g., by centrifugation. In some embodiments, the fermentation broth contains spent cell culture medium, extracellular enzymes, and viable and/or nonviable microbial cells.

In an embodiment, the fermentation broth formulation and cell compositions comprise a first organic acid component comprising at least one 1-5 carbon organic acid and/or a salt thereof and a second organic acid component comprising at least one 6 or more carbon organic acid and/or a salt thereof. In a specific embodiment, the first organic acid component is acetic acid, formic acid, propionic acid, a salt thereof, or a mixture of two or more of the foregoing and the second organic acid component is benzoic acid, cyclohexanecarboxylic acid, 4-methylvaleric acid, phenylacetic acid, a salt thereof, or a mixture of two or more of the foregoing.

In one aspect, the composition contains an organic acid(s), and optionally further contains killed cells and/or cell debris. In one embodiment, the killed cells and/or cell debris are removed from a cell-killed whole broth to provide a composition that is free of these components.

The fermentation broth formulations or cell compositions may further comprise a preservative and/or anti-microbial (e.g., bacteriostatic) agent, including, but not limited to, sorbitol, sodium chloride, potassium sorbate, and others known in the art.

The cell-killed whole broth or composition may contain the unfractionated contents of the fermentation materials derived at the end of the fermentation. Typically, the cell-killed whole broth or composition contains the spent culture medium and cell debris present after the microbial cells (e.g., filamentous fungal cells) are grown to saturation, incubated under carbon-limiting conditions to allow protein synthesis. In some embodiments, the cell-killed whole broth or composition contains the spent cell culture medium, extracellular enzymes, and killed filamentous fungal cells. In some embodiments, the microbial cells present in the cell-killed whole broth or composition can be permeabilized and/or lysed using methods known in the art.

A whole broth or cell composition as described herein is typically a liquid, but may contain insoluble components, such as killed cells, cell debris, culture media components, and/or insoluble enzyme(s). In some embodiments, insoluble components may be removed to provide a clarified liquid composition.

The whole broth formulations and cell compositions of the present invention may be produced by a method described in WO 90/15861 or WO 2010/096673.

Uses

The present invention is also directed to the following processes for using an enzyme composition comprising a recovered fermentation broth of the present invention.

The present invention also relates to processes for degrading a cellulosic material, comprising: treating the cellulosic material with an enzyme composition comprising a recovered fermentation broth of the present invention. In one aspect, the processes further comprise recovering the degraded or converted cellulosic material. Soluble products of degradation or conversion of the cellulosic material can be separated from insoluble cellulosic material using a method known in the art such as, for example, centrifugation, filtration, or gravity settling.

The present invention also relates to processes of producing a fermentation product, comprising: (a) saccharifying a cellulosic material with an enzyme composition comprising a recovered fermentation broth of the present invention; (b) fermenting the saccharified cellulosic material with one or more (e.g., several) fermenting microorganisms to produce the fermentation product; and (c) recovering the fermentation product from the fermentation.

The present invention also relates to processes of fermenting a cellulosic material, comprising: fermenting the cellulosic material with one or more (e.g., several) fermenting microorganisms, wherein the cellulosic material is saccharified with an enzyme composition comprising a recovered fermentation broth of the present invention. In one aspect, the fermenting of the cellulosic material produces a fermentation product. In another aspect, the processes further comprise recovering the fermentation product from the fermentation.

The processes of the present invention can be used to saccharify the cellulosic material to fermentable sugars and to convert the fermentable sugars to many useful fermentation products, e.g., fuel, potable ethanol, and/or platform chemicals (e.g., acids, alcohols, ketones, gases, and the like). The production of a desired fermentation product from the cellulosic material typically involves pretreatment, enzymatic hydrolysis (saccharification), and fermentation.

The processing of the cellulosic material according to the present invention can be accomplished using methods conventional in the art. Moreover, the processes of the present

invention can be implemented using any conventional biomass processing apparatus configured to operate in accordance with the invention.

Hydrolysis (saccharification) and fermentation, separate or simultaneous, include, but are not limited to, separate hydrolysis and fermentation (SHF); simultaneous saccharification and fermentation (SSF); simultaneous saccharification and co-fermentation (SSCF); hybrid hydrolysis and fermentation (HHF); separate hydrolysis and co-fermentation (SHCF); hybrid hydrolysis and co-fermentation (HHCF); and direct microbial conversion (DMC), also sometimes called consolidated bioprocessing (CBP). SHF uses separate process steps to first enzymatically hydrolyze the cellulosic material to fermentable sugars, e.g., glucose, cellobiose, and pentose monomers, and then ferment the fermentable sugars to ethanol. In SSF, the enzymatic hydrolysis of the cellulosic material and the fermentation of sugars to ethanol are combined in one step (Philippidis, G. P., 1996, *Cellulose bioconversion technology*, in *Handbook on Bioethanol: Production and Utilization*, Wyman, C. E., ed., Taylor & Francis, Washington, D.C., 179-212). SSCF involves the co-fermentation of multiple sugars (Sheehan, J., and Himmel, M., 1999, *Enzymes, energy and the environment: A strategic perspective on the U.S. Department of Energy's research and development activities for bioethanol*, *Biotechnol. Prog.* 15: 817-827). HHF involves a separate hydrolysis step, and in addition a simultaneous saccharification and hydrolysis step, which can be carried out in the same reactor. The steps in an HHF process can be carried out at different temperatures, i.e., high temperature enzymatic saccharification followed by SSF at a lower temperature that the fermentation strain can tolerate. DMC combines all three processes (enzyme production, hydrolysis, and fermentation) in one or more (e.g., several) steps where the same organism is used to produce the enzymes for conversion of the cellulosic material to fermentable sugars and to convert the fermentable sugars into a final product (Lynd, L. R., Weimer, P. J., van Zyl, W. H., and Pretorius, I. S., 2002, *Microbial cellulose utilization: Fundamentals and biotechnology*, *Microbiol. Mol. Biol. Reviews* 66: 506-577). It is understood herein that any method known in the art comprising pretreatment, enzymatic hydrolysis (saccharification), fermentation, or a combination thereof, can be used in the practicing the processes of the present invention.

A conventional apparatus can include a fed-batch stirred reactor, a batch stirred reactor, a continuous flow stirred reactor with ultrafiltration, and/or a continuous plug-flow column reactor (Fernanda de Castilhos Corazza, Flávio Faria de Moraes, Gisella Maria Zanin and Ivo Neitzel, 2003, *Optimal control in fed-batch reactor for the cellobiose hydrolysis*, *Acta Scientiarum. Technology* 25: 33-38; Guskov, A. V., and Sinitsyn, A. P., 1985, *Kinetics of the enzymatic hydrolysis of cellulose: 1. A mathematical model for a batch reactor process*, *Enz. Microb. Technol.* 7: 346-352), an attrition reactor (Ryu, S. K., and Lee, J. M., 1983, *Bioconversion of waste cellulose by using an attrition bioreactor*, *Biotechnol. Bioeng.* 25: 53-65), or a reactor with intensive stirring induced by an electromagnetic field (Guskov, A. V., Sinitsyn, A. P., Davydkin, I. Y., Davydkin, V. Y., Protas, O. V., 1996, *Enhancement of enzymatic cellulose hydrolysis using a novel type of bioreactor with intensive stirring induced by electromagnetic field*, *Appl. Biochem. Biotechnol.* 56: 141-153). Additional reactor types include fluidized bed, upflow blanket, immobilized, and extruder type reactors for hydrolysis and/or fermentation.

Pretreatment.

In practicing the processes of the present invention, any pretreatment process known in the art can be used to disrupt plant cell wall components of the cellulosic material (Chandra et al., 2007, Substrate pretreatment: The key to effective enzymatic hydrolysis of lignocellulosics?, *Adv. Biochem. Engin./Biotechnol.* 108: 67-93; Galbe and Zacchi, 2007, Pretreatment of lignocellulosic materials for efficient bioethanol production, *Adv. Biochem. Engin./Biotechnol.* 108: 41-65; Hendriks and Zeeman, 2009, Pretreatments to enhance the digestibility of lignocellulosic biomass, *Biore-source Technol.* 100: 10-18; Mosier et al., 2005, Features of promising technologies for pretreatment of lignocellulosic biomass, *Biore-source Technol.* 96: 673-686; Taherzadeh and Karimi, 2008, Pretreatment of lignocellulosic wastes to improve ethanol and biogas production: A review, *Int. J. of Mol. Sci.* 9: 1621-1651; Yang and Wyman, 2008, Pretreatment: the key to unlocking low-cost cellulosic ethanol, *Biofuels Bioproducts and Biorefining-Biofpr.* 2: 26-40).

The cellulosic material can also be subjected to particle size reduction, sieving, pre-soaking, wetting, washing, and/or conditioning prior to pretreatment using methods known in the art.

Conventional pretreatments include, but are not limited to, steam pretreatment (with or without explosion), dilute acid pretreatment, hot water pretreatment, alkaline pretreatment, lime pretreatment, wet oxidation, wet explosion, ammonia fiber explosion, organosolv pretreatment, and biological pretreatment. Additional pretreatments include ammonia percolation, ultrasound, electroporation, microwave, supercritical CO₂, supercritical H₂O, ozone, ionic liquid, and gamma irradiation pretreatments.

The cellulosic material can be pretreated before hydrolysis and/or fermentation. Pretreatment is preferably performed prior to the hydrolysis. Alternatively, the pretreatment can be carried out simultaneously with enzyme hydrolysis to release fermentable sugars, such as glucose, xylose, and/or cellobiose. In most cases the pretreatment step itself results in some conversion of biomass to fermentable sugars (even in absence of enzymes).

Steam Pretreatment. In steam pretreatment, the cellulosic material is heated to disrupt the plant cell wall components, including lignin, hemicellulose, and cellulose to make the cellulose and other fractions, e.g., hemicellulose, accessible to enzymes. The cellulosic material is passed to or through a reaction vessel where steam is injected to increase the temperature to the required temperature and pressure and is retained therein for the desired reaction time. Steam pretreatment is preferably performed at 140-250° C., e.g., 160-200° C. or 170-190° C., where the optimal temperature range depends on addition of a chemical catalyst. Residence time for the steam pretreatment is preferably 1-60 minutes, e.g., 1-30 minutes, 1-20 minutes, 3-12 minutes, or 4-10 minutes, where the optimal residence time depends on temperature range and addition of a chemical catalyst. Steam pretreatment allows for relatively high solids loadings, so that the cellulosic material is generally only moist during the pretreatment. The steam pretreatment is often combined with an explosive discharge of the material after the pretreatment, which is known as steam explosion, that is, rapid flashing to atmospheric pressure and turbulent flow of the material to increase the accessible surface area by fragmentation (Duff and Murray, 1996, *Biore-source Technology* 855: 1-33; Galbe and Zacchi, 2002, *Appl. Microbiol. Biotechnol.* 59: 618-628; U.S. Patent Application No. 20020164730). During steam pretreatment, hemicellulose acetyl groups are cleaved and the resulting acid autocatalyzes partial hydro-

lysis of the hemicellulose to monosaccharides and oligosaccharides. Lignin is removed to only a limited extent.

Chemical Pretreatment: The term "chemical treatment" refers to any chemical pretreatment that promotes the separation and/or release of cellulose, hemicellulose, and/or lignin. Such a pretreatment can convert crystalline cellulose to amorphous cellulose. Examples of suitable chemical pretreatment processes include, for example, dilute acid pretreatment, lime pretreatment, wet oxidation, ammonia fiber/freeze explosion (AFEX), ammonia percolation (APR), ionic liquid, and organosolv pretreatments.

A catalyst such as H₂SO₄ or SO₂ (typically 0.3 to 5% w/w) is often added prior to steam pretreatment, which decreases the time and temperature, increases the recovery, and improves enzymatic hydrolysis (Ballesteros et al., 2006, *Appl. Biochem. Biotechnol.* 129-132: 496-508; Varga et al., 2004, *Appl. Biochem. Biotechnol.* 113-116: 509-523; Sasser et al., 2006, *Enzyme Microb. Technol.* 39: 756-762). In dilute acid pretreatment, the cellulosic material is mixed with dilute acid, typically H₂SO₄, and water to form a slurry, heated by steam to the desired temperature, and after a residence time flashed to atmospheric pressure. The dilute acid pretreatment can be performed with a number of reactor designs, e.g., plug-flow reactors, counter-current reactors, or continuous counter-current shrinking bed reactors (Duff and Murray, 1996, supra; Schell et al., 2004, *Biore-source Technol.* 91: 179-188; Lee et al., 1999, *Adv. Biochem. Eng. Biotechnol.* 65: 93-115).

Several methods of pretreatment under alkaline conditions can also be used. These alkaline pretreatments include, but are not limited to, sodium hydroxide, lime, wet oxidation, ammonia percolation (APR), and ammonia fiber/freeze explosion (AFEX).

Lime pretreatment is performed with calcium oxide or calcium hydroxide at temperatures of 85-150° C. and residence times from 1 hour to several days (Wyman et al., 2005, *Biore-source Technol.* 96: 1959-1966; Mosier et al., 2005, *Biore-source Technol.* 96: 673-686). WO 2006/110891, WO 2006/110899, WO 2006/110900, and WO 2006/110901 disclose pretreatment methods using ammonia.

Wet oxidation is a thermal pretreatment performed typically at 180-200° C. for 5-15 minutes with addition of an oxidative agent such as hydrogen peroxide or over-pressure of oxygen (Schmidt and Thomsen, 1998, *Biore-source Technol.* 64: 139-151; Palonen et al., 2004, *Appl. Biochem. Biotechnol.* 117: 1-17; Varga et al., 2004, *Biotechnol. Bioeng.* 88: 567-574; Martin et al., 2006, *J. Chem. Technol. Biotechnol.* 81: 1669-1677). The pretreatment is performed preferably at 1-40% dry matter, e.g., 2-30% dry matter or 5-20% dry matter, and often the initial pH is increased by the addition of alkali such as sodium carbonate.

A modification of the wet oxidation pretreatment method, known as wet explosion (combination of wet oxidation and steam explosion) can handle dry matter up to 30%. In wet explosion, the oxidizing agent is introduced during pretreatment after a certain residence time. The pretreatment is then ended by flashing to atmospheric pressure (WO 2006/032282).

Ammonia fiber explosion (AFEX) involves treating the cellulosic material with liquid or gaseous ammonia at moderate temperatures such as 90-150° C. and high pressure such as 17-20 bar for 5-10 minutes, where the dry matter content can be as high as 60% (Gollapalli et al., 2002, *Appl. Biochem. Biotechnol.* 98: 23-35; Chundawat et al., 2007, *Biotechnol. Bioeng.* 96: 219-231; Alizadeh et al., 2005, *Appl. Biochem. Biotechnol.* 121: 1133-1141; Teymouri et al., 2005, *Biore-source Technol.* 96: 2014-2018). During AFEX

pretreatment cellulose and hemicelluloses remain relatively intact. Lignin-carbohydrate complexes are cleaved.

Organosolv pretreatment delignifies the cellulosic material by extraction using aqueous ethanol (40-60% ethanol) at 160-200° C. for 30-60 minutes (Pan et al., 2005, *Biotechnol. Bioeng.* 90: 473-481; Pan et al., 2006, *Biotechnol. Bioeng.* 94: 851-861; Kurabi et al., 2005, *Appl. Biochem. Biotechnol.* 121: 219-230). Sulphuric acid is usually added as a catalyst. In organosolv pretreatment, the majority of hemicellulose and lignin is removed.

Other examples of suitable pretreatment methods are described by Schell et al., 2003, *Appl. Biochem. and Biotechnol. Vol.* 105-108, p. 69-85, and Mosier et al., 2005, *Bioresource Technology* 96: 673-686, and U.S. Published Application 2002/0164730.

In one aspect, the chemical pretreatment is preferably carried out as a dilute acid treatment, and more preferably as a continuous dilute acid treatment. The acid is typically sulfuric acid, but other acids can also be used, such as acetic acid, citric acid, nitric acid, phosphoric acid, tartaric acid, succinic acid, hydrogen chloride, or mixtures thereof. Mild acid treatment is conducted in the pH range of preferably 1-5, e.g., 1-4 or 1-2.5. In one aspect, the acid concentration is in the range from preferably 0.01 to 10 wt % acid, e.g., 0.05 to 5 wt % acid or 0.1 to 2 wt % acid. The acid is contacted with the cellulosic material and held at a temperature in the range of preferably 140-200° C., e.g., 165-190° C., for periods ranging from 1 to 60 minutes.

In another aspect, pretreatment takes place in an aqueous slurry. In preferred aspects, the cellulosic material is present during pretreatment in amounts preferably between 10-80 wt %, e.g., 20-70 wt % or 30-60 wt %, such as around 40 wt %. The pretreated cellulosic material can be unwashed or washed using any method known in the art, e.g., washed with water.

Mechanical Pretreatment or Physical Pretreatment: The term "mechanical pretreatment" or "physical pretreatment" refers to any pretreatment that promotes size reduction of particles. For example, such pretreatment can involve various types of grinding or milling (e.g., dry milling, wet milling, or vibratory ball milling).

The cellulosic material can be pretreated both physically (mechanically) and chemically. Mechanical or physical pretreatment can be coupled with steaming/steam explosion, hydrothermolysis, dilute or mild acid treatment, high temperature, high pressure treatment, irradiation (e.g., microwave irradiation), or combinations thereof. In one aspect, high pressure means pressure in the range of preferably about 100 to about 400 psi, e.g., about 150 to about 250 psi. In another aspect, high temperature means temperatures in the range of about 100 to about 300° C., e.g., about 140 to about 200° C. In a preferred aspect, mechanical or physical pretreatment is performed in a batch-process using a steam gun hydrolyzer system that uses high pressure and high temperature as defined above, e.g., a Sunds Hydrolyzer available from Sunds Defibrator AB, Sweden. The physical and chemical pretreatments can be carried out sequentially or simultaneously, as desired.

Accordingly, in a preferred aspect, the cellulosic material is subjected to physical (mechanical) or chemical pretreatment, or any combination thereof, to promote the separation and/or release of cellulose, hemicellulose, and/or lignin.

Biological Pretreatment: The term "biological pretreatment" refers to any biological pretreatment that promotes the separation and/or release of cellulose, hemicellulose, and/or lignin from the cellulosic material. Biological pretreatment techniques can involve applying lignin-solubiliz-

ing microorganisms and/or enzymes (see, for example, Hsu, T.-A., 1996, Pretreatment of biomass, in *Handbook on Bioethanol: Production and Utilization*, Wyman, C. E., ed., Taylor & Francis, Washington, D.C., 179-212; Ghosh and Singh, 1993, Physicochemical and biological treatments for enzymatic/microbial conversion of cellulosic biomass, *Adv. Appl. Microbiol.* 39: 295-333; McMillan, J. D., 1994, Pretreating lignocellulosic biomass: a review, in *Enzymatic Conversion of Biomass for Fuels Production*, Himmel, M. E., Baker, J. O., and Overend, R. P., eds., ACS Symposium Series 566, American Chemical Society, Washington, D.C., chapter 15; Gong, C. S., Cao, N. J., Du, J., and Tsao, G. T., 1999, Ethanol production from renewable resources, in *Advances in Biochemical Engineering/Biotechnology*, Scheper, T., ed., Springer-Verlag Berlin Heidelberg, Germany, 65: 207-241; Olsson and Hahn-Hagerdal, 1996, Fermentation of lignocellulosic hydrolysates for ethanol production, *Enz. Microb. Tech.* 18: 312-331; and Vallander and Eriksson, 1990, Production of ethanol from lignocellulosic materials: State of the art, *Adv. Biochem. Eng./Biotechnol.* 42: 63-95).

Saccharification.

In the hydrolysis step, also known as saccharification, the cellulosic material, e.g., pretreated, is hydrolyzed to break down cellulose and/or hemicellulose to fermentable sugars, such as glucose, cellobiose, xylose, xylulose, arabinose, mannose, galactose, and/or soluble oligosaccharides. The hydrolysis is performed enzymatically by an enzyme composition comprising a recovered fermentation broth of the present invention.

Enzymatic hydrolysis is preferably carried out in a suitable aqueous environment under conditions that can be readily determined by one skilled in the art. In one aspect, hydrolysis is performed under conditions suitable for the activity of the enzyme(s), i.e., optimal for the enzyme(s). The hydrolysis can be carried out as a fed batch or continuous process where the cellulosic material is fed gradually to, for example, an enzyme containing hydrolysis solution.

The saccharification is generally performed in stirred-tank reactors or fermentors under controlled pH, temperature, and mixing conditions. Suitable process time, temperature and pH conditions can readily be determined by one skilled in the art. For example, the saccharification can last up to 200 hours, but is typically performed for preferably about 12 to about 120 hours, e.g., about 16 to about 72 hours or about 24 to about 48 hours. The temperature is in the range of preferably about 25° C. to about 70° C., e.g., about 30° C. to about 65° C., about 40° C. to about 60° C., or about 50° C. to about 55° C. The pH is in the range of preferably about 3 to about 8, e.g., about 3.5 to about 7, about 4 to about 6, or about 5.0 to about 5.5. The dry solids content is in the range of preferably about 5 to about 50 wt %, e.g., about 10 to about 40 wt % or about 20 to about 30 wt %.

In the processes of the present invention, the enzyme composition comprising a recovered fermentation broth of the present invention can be added prior to or during fermentation, e.g., during saccharification or during or after propagation of the fermenting microorganism(s).

The enzyme composition comprising a recovered fermentation broth of the present invention may be in any form suitable for use, such as, for example, a crude fermentation broth with or without cells removed, a cell lysate with or without cellular debris, a semi-purified or purified enzyme preparation, or a *Trichoderma* host cell as a source of the enzymes. The enzyme composition may be a dry powder or granulate, a non-dusting granulate, a liquid, a stabilized liquid, or a stabilized protected enzyme. Liquid enzyme

preparations may, for instance, be stabilized by adding stabilizers such as a sugar, a sugar alcohol or another polyol, and/or lactic acid or another organic acid according to established processes.

The optimum amount of *Aspergillus fumigatus* cellulases or hemicellulases depends on several factors including, but not limited to, the mixture of component cellulolytic and/or hemicellulolytic enzymes, the cellulosic material, the concentration of cellulosic material, the pretreatment(s) of the cellulosic material, temperature, time, pH, and inclusion of fermenting organism (e.g., yeast for Simultaneous Saccharification and Fermentation).

In one aspect, an effective amount of cellulolytic or hemicellulolytic enzyme to the cellulosic material is about 0.01 to about 50.0 mg, e.g., about 0.01 to about 40 mg, about 0.01 to about 30 mg, about 0.01 to about 20 mg, about 0.01 to about 10 mg, about 0.01 to about 5 mg, about 0.025 to about 1.5 mg, about 0.05 to about 1.25 mg, about 0.075 to about 1.25 mg, about 0.1 to about 1.25 mg, about 0.15 to about 1.25 mg, or about 0.25 to about 1.0 mg per g of the cellulosic material.

In another aspect, the GH61 polypeptide having cellulolytic enhancing activity is used in the presence of a soluble activating divalent metal cation according to WO 2008/151043, e.g., manganese sulfate.

In another aspect, a GH61 polypeptide having cellulolytic enhancing activity is used in the presence of a dioxy compound, a bicyclic compound, a heterocyclic compound, a nitrogen-containing compound, a quinone compound, a sulfur-containing compound, or a liquor obtained from a pretreated cellulosic material such as pretreated corn stover (PCS).

The dioxy compound may include any suitable compound containing two or more oxygen atoms. In some aspects, the dioxy compounds contain a substituted aryl moiety as described herein. The dioxy compounds may comprise one or more (e.g., several) hydroxyl and/or hydroxyl derivatives, but also include substituted aryl moieties lacking hydroxyl and hydroxyl derivatives. Non-limiting examples of the dioxy compounds include pyrocatechol or catechol; caffeic acid; 3,4-dihydroxybenzoic acid; 4-tert-butyl-5-methoxy-1,2-benzenediol; pyrogallol; gallic acid; methyl-3,4,5-trihydroxybenzoate; 2,3,4-trihydroxybenzophenone; 2,6-dimethoxyphenol; sinapinic acid; 3,5-dihydroxybenzoic acid; 4-chloro-1,2-benzenediol; 4-nitro-1,2-benzenediol; tannic acid; ethyl gallate; methyl glycolate; dihydroxyfumaric acid; 2-butyne-1,4-diol; (croconic acid; 1,3-propanediol; tartaric acid; 2,4-pentanediol; 3-ethoxy-1,2-propanediol; 2,4,4'-trihydroxybenzophenone; cis-2-butene-1,4-diol; 3,4-dihydroxy-3-cyclobutene-1,2-dione; dihydroxyacetone; acrolein acetal; methyl-4-hydroxybenzoate; 4-hydroxybenzoic acid; and methyl-3,5-dimethoxy-4-hydroxybenzoate; or a salt or solvate thereof.

The bicyclic compound may include any suitable substituted fused ring system as described herein. The compounds may comprise one or more (e.g., several) additional rings, and are not limited to a specific number of rings unless otherwise stated. In one aspect, the bicyclic compound is a flavonoid. In another aspect, the bicyclic compound is an optionally substituted isoflavonoid. In another aspect, the bicyclic compound is an optionally substituted flavylum ion, such as an optionally substituted anthocyanidin or optionally substituted anthocyanin, or derivative thereof. Non-limiting examples of the bicyclic compounds include epicatechin; quercetin; myricetin; taxifolin; kaempferol;

morin; acacetin; naringenin; isorhamnetin; apigenin; cyanidin; cyanin; kuromanin; keracyanin; or a salt or solvate thereof.

The heterocyclic compound may be any suitable compound, such as an optionally substituted aromatic or non-aromatic ring comprising a heteroatom, as described herein. In one aspect, the heterocyclic is a compound comprising an optionally substituted heterocycloalkyl moiety or an optionally substituted heteroaryl moiety. In another aspect, the optionally substituted heterocycloalkyl moiety or optionally substituted heteroaryl moiety is an optionally substituted 5-membered heterocycloalkyl or an optionally substituted 5-membered heteroaryl moiety. In another aspect, the optionally substituted heterocycloalkyl or optionally substituted heteroaryl moiety is an optionally substituted moiety selected from pyrazolyl, furanyl, imidazolyl, isoxazolyl, oxadiazolyl, oxazolyl, pyrrolyl, pyridyl, pyrimidyl, pyridazinyl, thiazolyl, triazolyl, thienyl, dihydrothienopyrazolyl, thianaphthenyl, carbazolyl, benzimidazolyl, benzothienyl, benzofuranyl, indolyl, quinolinyl, benzotriazolyl, benzothiazolyl, benzooxazolyl, benzimidazolyl, isoquinolinyl, isoindolyl, acridinyl, benzoisazolyl, dimethylhydantoin, pyrazinyl, tetrahydrofuranyl, pyrrolinyl, pyrrolidinyl, morpholinyl, indolyl, diazepinyl, azepinyl, thiepinyl, piperidinyl, and oxepinyl. In another aspect, the optionally substituted heterocycloalkyl moiety or optionally substituted heteroaryl moiety is an optionally substituted furanyl. Non-limiting examples of the heterocyclic compounds include (1,2-dihydroxyethyl)-3,4-dihydroxyfuran-2(5H)-one; 4-hydroxy-5-methyl-3-furanone; 5-hydroxy-2(5H)-furanone; [1,2-dihydroxyethyl]furan-2,3,4(5H)-trione; α -hydroxy- γ -butyrolactone; ribonic γ -lactone; aldohexuronic acid γ -lactone; gluconic acid δ -lactone; 4-hydroxycoumarin; dihydrobenzofuran; 5-(hydroxymethyl) furfural; furoin; 2(5H)-furanone; 5,6-dihydro-2H-pyran-2-one; and 5,6-dihydro-4-hydroxy-6-methyl-2H-pyran-2-one; or a salt or solvate thereof.

The nitrogen-containing compound may be any suitable compound with one or more (e.g., several) nitrogen atoms. In one aspect, the nitrogen-containing compound comprises an amine, imine, hydroxylamine, or nitroxide moiety. Non-limiting examples of the nitrogen-containing compounds include acetone oxime; violuric acid; pyridine-2-aldoxime; 2-aminophenol; 1,2-benzenediamine; 2,2,6,6-tetramethyl-1-piperidinyloxy; 5,6,7,8-tetrahydrobiopterin; 6,7-dimethyl-5,6,7,8-tetrahydropterine; and maleamic acid; or a salt or solvate thereof.

The quinone compound may be any suitable compound comprising a quinone moiety as described herein. Non-limiting examples of the quinone compounds include 1,4-benzoquinone; 1,4-naphthoquinone; 2-hydroxy-1,4-naphthoquinone; 2,3-dimethoxy-5-methyl-1,4-benzoquinone or coenzyme Q₀; 2,3,5,6-tetramethyl-1,4-benzoquinone or duroquinone; 1,4-dihydroxyanthraquinone; 3-hydroxy-1-methyl-5,6-indolinedione or adrenochrome; 4-tert-butyl-5-methoxy-1,2-benzoquinone; pyrroloquinoline quinone; or a salt or solvate thereof.

The sulfur-containing compound may be any suitable compound comprising one or more (e.g., several) sulfur atoms. In one aspect, the sulfur-containing comprises a moiety selected from thionyl, thioether, sulfinyl, sulfonyl, sulfamide, sulfonamide, sulfonic acid, and sulfonic ester. Non-limiting examples of the sulfur-containing compounds include ethanethiol; 2-propanethiol; 2-propene-1-thiol; 2-mercaptoethanesulfonic acid; benzenethiol; benzene-1,2-dithiol; cysteine; methionine; glutathione; cystine; or a salt or solvate thereof.

In one aspect, an effective amount of such a compound described above to cellulosic material as a molar ratio to glucosyl units of cellulose is about 10^{-6} to about 10, e.g., about 10^{-6} to about 7.5, about 10^{-6} to about 5, about 10^{-6} to about 2.5, about 10^{-6} to about 1, about 10^{-5} to about 1, about 10^{-5} to about 10^{-1} , about 10^{-4} to about 10^{-1} , about 10^{-3} to about 10^{-1} , or about 10^{-3} to about 10^{-2} . In another aspect, an effective amount of such a compound described above is about 0.1 μ M to about 1 M, e.g., about 0.5 μ M to about 0.75 M, about 0.75 μ M to about 0.5 M, about 1 μ M to about 0.25 M, about 1 μ M to about 0.1 M, about 5 μ M to about 50 mM, about 10 μ M to about 25 mM, about 50 μ M to about 25 mM, about 10 μ M to about 10 mM, about 5 μ M to about 5 mM, or about 0.1 mM to about 1 mM.

The term "liquor" means the solution phase, either aqueous, organic, or a combination thereof, arising from treatment of a lignocellulose and/or hemicellulose material in a slurry, or monosaccharides thereof, e.g., xylose, arabinose, mannose, etc., under conditions as described herein, and the soluble contents thereof. A liquor for cellulolytic enhancement of a GH61 polypeptide can be produced by treating a lignocellulose or hemicellulose material (or feedstock) by applying heat and/or pressure, optionally in the presence of a catalyst, e.g., acid, optionally in the presence of an organic solvent, and optionally in combination with physical disruption of the material, and then separating the solution from the residual solids. Such conditions determine the degree of cellulolytic enhancement obtainable through the combination of liquor and a GH61 polypeptide during hydrolysis of a cellulosic substrate by a cellulase preparation. The liquor can be separated from the treated material using a method standard in the art, such as filtration, sedimentation, or centrifugation.

In one aspect, an effective amount of the liquor to cellulose is about 10^{-6} to about 10 g per g of cellulose, e.g., about 10^{-6} to about 7.5 g, about 10^{-6} to about 5, about 10^{-6} to about 2.5 g, about 10^{-6} to about 1 g, about 10^{-5} to about 1 g, about 10^{-5} to about 10^{-1} g, about 10^{-4} to about 10^{-1} g, about 10^{-3} to about 10^{-1} g, or about 10^{-3} to about 10^{-2} g per g of cellulose.

Fermentation.

The fermentable sugars obtained from the hydrolyzed cellulosic material can be fermented by one or more (e.g., several) fermenting microorganisms capable of fermenting the sugars directly or indirectly into a desired fermentation product. "Fermentation" or "fermentation process" refers to any fermentation process or any process comprising a fermentation step. Fermentation processes also include fermentation processes used in the consumable alcohol industry (e.g., beer and wine), dairy industry (e.g., fermented dairy products), leather industry, and tobacco industry. The fermentation conditions depend on the desired fermentation product and fermenting organism and can easily be determined by one skilled in the art.

In the fermentation step, sugars, released from the cellulosic material as a result of the pretreatment and enzymatic hydrolysis steps, are fermented to a product, e.g., ethanol, by a fermenting organism, such as yeast. Hydrolysis (saccharification) and fermentation can be separate or simultaneous, as described herein.

Any suitable hydrolyzed cellulosic material can be used in the fermentation step in practicing the present invention. The material is generally selected based on the desired fermentation product, i.e., the substance to be obtained from the fermentation, and the process employed, as is well known in the art.

The term "fermentation medium" is understood herein to refer to a medium before the fermenting microorganism(s) is(are) added, such as, a medium resulting from a saccharification process, as well as a medium used in a simultaneous saccharification and fermentation process (SSF).

"Fermenting microorganism" refers to any microorganism, including bacterial and fungal organisms, suitable for use in a desired fermentation process to produce a fermentation product. The fermenting organism can be hexose and/or pentose fermenting organisms, or a combination thereof. Both hexose and pentose fermenting organisms are well known in the art. Suitable fermenting microorganisms are able to ferment, i.e., convert, sugars, such as glucose, xylose, xylulose, arabinose, maltose, mannose, galactose, and/or oligosaccharides, directly or indirectly into the desired fermentation product. Examples of bacterial and fungal fermenting organisms producing ethanol are described by Lin et al., 2006, *Appl. Microbiol. Biotechnol.* 69: 627-642.

Examples of fermenting microorganisms that can ferment hexose sugars include bacterial and fungal organisms, such as yeast. Preferred yeast includes strains of *Candida*, *Kluyveromyces*, and *Saccharomyces*, e.g., *Candida sonorensis*, *Kluyveromyces marxianus*, and *Saccharomyces cerevisiae*.

Examples of fermenting organisms that can ferment pentose sugars in their native state include bacterial and fungal organisms, such as some yeast. Preferred xylose fermenting yeast include strains of *Candida*, preferably *C. sheatae* or *C. sonorensis*; and strains of *Pichia*, preferably *P. stipitis*, such as *P. stipitis* CBS 5773. Preferred pentose fermenting yeast include strains of *Pachysolen*, preferably *P. tannophilus*. Organisms not capable of fermenting pentose sugars, such as xylose and arabinose, may be genetically modified to do so by methods known in the art.

Examples of bacteria that can efficiently ferment hexose and pentose to ethanol include, for example, *Bacillus coagulans*, *Clostridium acetobutylicum*, *Clostridium thermocellum*, *Clostridium phytofermentans*, *Geobacillus* sp., *Thermoanaerobacter saccharolyticum*, and *Zymomonas mobilis* (Philippidis, 1996, supra).

Other fermenting organisms include strains of *Bacillus*, such as *Bacillus coagulans*; *Candida*, such as *C. sonorensis*, *C. methanosorbosa*, *C. diddensiae*, *C. parapsilosis*, *C. naeododendra*, *C. blankii*, *C. entomophilia*, *C. brassicae*, *C. pseudotropicalis*, *C. boidinii*, *C. utilis*, and *C. scheidtiae*; *Clostridium*, such as *C. acetobutylicum*, *C. thermocellum*, and *C. phytofermentans*; *E. coli*, especially *E. coli* strains that have been genetically modified to improve the yield of ethanol; *Geobacillus* sp.; *Hansenula*, such as *Hansenula anomala*; *Klebsiella*, such as *K. oxytoca*; *Kluyveromyces*, such as *K. marxianus*, *K. lactis*, *K. thermotolerans*, and *K. fragilis*; *Schizosaccharomyces*, such as *S. pombe*; *Thermoanaerobacter*, such as *Thermoanaerobacter saccharolyticum*; and *Zymomonas*, such as *Zymomonas mobilis*.

In a preferred aspect, the yeast is a *Bretannomyces*. In a more preferred aspect, the yeast is *Bretannomyces clausenii*. In another preferred aspect, the yeast is a *Candida*. In another more preferred aspect, the yeast is *Candida sonorensis*. In another more preferred aspect, the yeast is *Candida boidinii*. In another more preferred aspect, the yeast is *Candida blankii*. In another more preferred aspect, the yeast is *Candida brassicae*. In another more preferred aspect, the yeast is *Candida diddensii*. In another more preferred aspect, the yeast is *Candida entomophiliia*. In another more preferred aspect, the yeast is *Candida pseudotropicalis*. In another more preferred aspect, the yeast is *Candida scheidtiae*.

tae. In another more preferred aspect, the yeast is *Candida utilis*. In another preferred aspect, the yeast is a *Clavispora*. In another more preferred aspect, the yeast is *Clavispora lusitaniae*. In another more preferred aspect, the yeast is *Clavispora opuntiae*. In another preferred aspect, the yeast is a *Kluyveromyces*. In another more preferred aspect, the yeast is *Kluyveromyces fragilis*. In another more preferred aspect, the yeast is *Kluyveromyces marxianus*. In another more preferred aspect, the yeast is *Kluyveromyces thermotolerans*. In another preferred aspect, the yeast is a *Pachysolen*. In another more preferred aspect, the yeast is *Pachysolen tannophilus*. In another preferred aspect, the yeast is a *Pichia*. In another more preferred aspect, the yeast is a *Pichia stipitis*. In another preferred aspect, the yeast is a *Saccharomyces* spp. In another more preferred aspect, the yeast is *Saccharomyces cerevisiae*. In another more preferred aspect, the yeast is *Saccharomyces distaticus*. In another more preferred aspect, the yeast is *Saccharomyces uvarum*.

In a preferred aspect, the bacterium is a *Bacillus*. In a more preferred aspect, the bacterium is *Bacillus coagulans*. In another preferred aspect, the bacterium is a *Clostridium*. In another more preferred aspect, the bacterium is *Clostridium acetobutylicum*. In another more preferred aspect, the bacterium is *Clostridium phytofermentans*. In another more preferred aspect, the bacterium is *Clostridium thermocellum*. In another more preferred aspect, the bacterium is *Geobacillus* sp. In another more preferred aspect, the bacterium is a *Thermoanaerobacter*. In another more preferred aspect, the bacterium is *Thermoanaerobacter saccharolyticum*. In another preferred aspect, the bacterium is a *Zymomonas*. In another more preferred aspect, the bacterium is *Zymomonas mobilis*.

Commercially available yeast suitable for ethanol production include, e.g., BIOFERM™ AFT and XR (NABC—North American Bioproducts Corporation, GA, USA), ETHANOL RED™ yeast (Fermentis/Lesaffre, USA), FALI™ (Fleischmann's Yeast, USA), FERMIOL™ (DSM Specialties), GERT STRAND™ (Gert Strand AB, Sweden), and SUPERSTART™ and THERMOSACCT™ fresh yeast (Ethanol Technology, WI, USA).

In a preferred aspect, the fermenting microorganism has been genetically modified to provide the ability to ferment pentose sugars, such as xylose utilizing, arabinose utilizing, and xylose and arabinose co-utilizing microorganisms.

The cloning of heterologous genes into various fermenting microorganisms has led to the construction of organisms capable of converting hexoses and pentoses to ethanol (co-fermentation) (Chen and Ho, 1993, Cloning and improving the expression of *Pichia stipitis* xylose reductase gene in *Saccharomyces cerevisiae*, *Appl. Biochem. Biotechnol.* 39-40: 135-147; Ho et al., 1998, Genetically engineered *Saccharomyces* yeast capable of effectively cofermenting glucose and xylose, *Appl. Environ. Microbiol.* 64: 1852-1859; Kotter and Ciriacy, 1993, Xylose fermentation by *Saccharomyces cerevisiae*, *Appl. Microbiol. Biotechnol.* 38: 776-783; Walfridsson et al., 1995, Xylose-metabolizing *Saccharomyces cerevisiae* strains overexpressing the TKL1 and TAL1 genes encoding the pentose phosphate pathway enzymes transketolase and transaldolase, *Appl. Environ. Microbiol.* 61: 4184-4190; Kuyper et al., 2004, Minimal metabolic engineering of *Saccharomyces cerevisiae* for efficient anaerobic xylose fermentation: a proof of principle, *FEMS Yeast Research* 4: 655-664; Beall et al., 1991, Parametric studies of ethanol production from xylose and other sugars by recombinant *Escherichia coli*, *Biotech. Bioeng.* 38: 296-303; Ingram et al., 1998, Metabolic engineering of

bacteria for ethanol production, *Biotechnol. Bioeng.* 58: 204-214; Zhang et al., 1995, Metabolic engineering of a pentose metabolism pathway in ethanologenic *Zymomonas mobilis*, *Science* 267: 240-243; Deanda et al., 1996, Development of an arabinose-fermenting *Zymomonas mobilis* strain by metabolic pathway engineering, *Appl. Environ. Microbiol.* 62: 4465-4470; WO 2003/062430, xylose isomerase).

In a preferred aspect, the genetically modified fermenting microorganism is *Candida sonorensis*. In another preferred aspect, the genetically modified fermenting microorganism is *Escherichia coli*. In another preferred aspect, the genetically modified fermenting microorganism is *Klebsiella oxytoca*. In another preferred aspect, the genetically modified fermenting microorganism is *Kluyveromyces marxianus*. In another preferred aspect, the genetically modified fermenting microorganism is *Saccharomyces cerevisiae*. In another preferred aspect, the genetically modified fermenting microorganism is *Zymomonas mobilis*.

It is well known in the art that the organisms described above can also be used to produce other substances, as described herein.

The fermenting microorganism is typically added to the degraded cellulosic material or hydrolysate and the fermentation is performed for about 8 to about 96 hours, e.g., about 24 to about 60 hours. The temperature is typically between about 26° C. to about 60° C., e.g., about 32° C. or 50° C., and about pH 3 to about pH 8, e.g., pH 4-5, 6, or 7.

In one aspect, the yeast and/or another microorganism are applied to the degraded cellulosic material and the fermentation is performed for about 12 to about 96 hours, such as typically 24-60 hours. In another aspect, the temperature is preferably between about 20° C. to about 60° C., e.g., about 25° C. to about 50° C., about 32° C. to about 50° C., or about 32° C. to about 50° C., and the pH is generally from about pH 3 to about pH 7, e.g., about pH 4 to about pH 7. However, some fermenting organisms, e.g., bacteria, have higher fermentation temperature optima. Yeast or another microorganism is preferably applied in amounts of approximately 10⁵ to 10¹², preferably from approximately 10⁷ to 10¹⁰, especially approximately 2×10⁸ viable cell count per ml of fermentation broth. Further guidance in respect of using yeast for fermentation can be found in, e.g., "The Alcohol Textbook" (Editors K. Jacques, T. P. Lyons and D. R. Kelsall, Nottingham University Press, United Kingdom 1999), which is hereby incorporated by reference.

A fermentation stimulator can be used in combination with any of the processes described herein to further improve the fermentation process, and in particular, the performance of the fermenting microorganism, such as, rate enhancement and ethanol yield. A "fermentation stimulator" refers to stimulators for growth of the fermenting microorganisms, in particular, yeast. Preferred fermentation stimulators for growth include vitamins and minerals. Examples of vitamins include multivitamins, biotin, pantothenate, nicotinic acid, meso-inositol, thiamine, pyridoxine, paraaminobenzoic acid, folic acid, riboflavin, and Vitamins A, B, C, D, and E. See, for example, Alfenore et al., Improving ethanol production and viability of *Saccharomyces cerevisiae* by a vitamin feeding strategy during fed-batch process, Springer-Verlag (2002), which is hereby incorporated by reference. Examples of minerals include minerals and mineral salts that can supply nutrients comprising P, K, Mg, S, Ca, Fe, Zn, Mn, and Cu.

Fermentation Products:

A fermentation product can be any substance derived from the fermentation. The fermentation product can be,

without limitation, an alcohol (e.g., arabinitol, n-butanol, isobutanol, ethanol, glycerol, methanol, ethylene glycol, 1,3-propanediol [propylene glycol], butanediol, glycerin, sorbitol, and xylitol); an alkane (e.g., pentane, hexane, heptane, octane, nonane, decane, undecane, and dodecane),
5 a cycloalkane (e.g., cyclopentane, cyclohexane, cycloheptane, and cyclooctane), an alkene (e.g. pentene, hexene, heptene, and octene); an amino acid (e.g., aspartic acid, glutamic acid, glycine, lysine, serine, and threonine); a gas (e.g., methane, hydrogen (H₂), carbon dioxide (CO₂), and
10 carbon monoxide (CO)); isoprene; a ketone (e.g., acetone); an organic acid (e.g., acetic acid, acetic acid, adipic acid, ascorbic acid, citric acid, 2,5-diketo-D-gluconic acid, formic acid, fumaric acid, glucaric acid, gluconic acid, glucuronic acid, glutaric acid, 3-hydroxypropionic acid, itaconic acid,
15 lactic acid, malic acid, malonic acid, oxalic acid, oxaloacetic acid, propionic acid, succinic acid, and xylonic acid); and polyketide. The fermentation product can also be protein as a high value product.

In a preferred aspect, the fermentation product is an alcohol. It will be understood that the term "alcohol" encompasses a substance that contains one or more (e.g., several) hydroxyl moieties. In a more preferred aspect, the alcohol is n-butanol. In another more preferred aspect, the alcohol is isobutanol. In another more preferred aspect, the alcohol is ethanol. In another more preferred aspect, the alcohol is methanol. In another more preferred aspect, the alcohol is arabinitol. In another more preferred aspect, the alcohol is butanediol. In another more preferred aspect, the alcohol is ethylene glycol. In another more preferred aspect, the alcohol is glycerin. In another more preferred aspect, the alcohol is glycerol. In another more preferred aspect, the alcohol is 1,3-propanediol. In another more preferred aspect, the alcohol is sorbitol. In another more preferred aspect, the alcohol is xylitol. See, for example, Gong, C. S., Cao, N. J., Du, J., and Tsao, G. T., 1999, Ethanol production from renewable resources, in *Advances in Biochemical Engineering/Biotechnology*, Scheper, T., ed., Springer-Verlag Berlin Heidelberg, Germany, 65: 207-241; Silveira, M. M., and Jonas, R., 2002, The biotechnological production of sorbitol, *Appl. Microbiol. Biotechnol.* 59: 400-408; Nigam, P., and Singh, D., 1995, Processes for fermentative production of xylitol—a sugar substitute, *Process Biochemistry* 30 (2): 117-124; Ezeji, T. C., Qureshi, N. and Blaschek, H. P., 2003, Production of acetone, butanol and ethanol by *Clostridium beijerinckii* BA101 and in situ recovery by gas stripping, *World Journal of Microbiology and Biotechnology* 19 (6): 595-603.

In another preferred aspect, the fermentation product is an alkane. The alkane can be an unbranched or a branched
50 alkane. In another more preferred aspect, the alkane is pentane. In another more preferred aspect, the alkane is hexane. In another more preferred aspect, the alkane is heptane. In another more preferred aspect, the alkane is octane. In another more preferred aspect, the alkane is nonane. In another more preferred aspect, the alkane is decane. In another more preferred aspect, the alkane is undecane. In another more preferred aspect, the alkane is dodecane.

In another preferred aspect, the fermentation product is a cycloalkane. In another more preferred aspect, the cycloalkane is cyclopentane. In another more preferred aspect, the cycloalkane is cyclohexane. In another more preferred aspect, the cycloalkane is cycloheptane. In another more preferred aspect, the cycloalkane is cyclooctane.

In another preferred aspect, the fermentation product is an alkene. The alkene can be an unbranched or a branched

alkene. In another more preferred aspect, the alkene is pentene. In another more preferred aspect, the alkene is hexene. In another more preferred aspect, the alkene is heptene. In another more preferred aspect, the alkene is octene.

In another preferred aspect, the fermentation product is an amino acid. In another more preferred aspect, the organic acid is aspartic acid. In another more preferred aspect, the amino acid is glutamic acid. In another more preferred aspect, the amino acid is glycine. In another more preferred aspect, the amino acid is lysine. In another more preferred aspect, the amino acid is serine. In another more preferred aspect, the amino acid is threonine. See, for example, Richard, A., and Margaritis, A., 2004, Empirical modeling of batch fermentation kinetics for poly(glutamic acid) production and other microbial biopolymers, *Biotechnology and Bioengineering* 87 (4): 501-515.

In another preferred aspect, the fermentation product is a gas. In another more preferred aspect, the gas is methane. In another more preferred aspect, the gas is H₂. In another more preferred aspect, the gas is CO₂. In another more preferred aspect, the gas is CO. See, for example, Kataoka, N., A. Miya, and K. Kiriya, 1997, Studies on hydrogen production by continuous culture system of hydrogen-producing anaerobic bacteria, *Water Science and Technology* 36 (6-7): 41-47; and Gunaseelan V. N. in *Biomass and Bioenergy*, Vol. 13 (1-2), pp. 83-114, 1997, Anaerobic digestion of biomass for methane production: A review.

In another preferred aspect, the fermentation product is isoprene.

In another preferred aspect, the fermentation product is a ketone. It will be understood that the term "ketone" encompasses a substance that contains one or more (e.g., several) ketone moieties. In another more preferred aspect, the ketone is acetone. See, for example, Qureshi and Blaschek, 2003, supra.

In another preferred aspect, the fermentation product is an organic acid. In another more preferred aspect, the organic acid is acetic acid. In another more preferred aspect, the organic acid is acetic acid. In another more preferred aspect, the organic acid is adipic acid. In another more preferred aspect, the organic acid is ascorbic acid. In another more preferred aspect, the organic acid is citric acid. In another more preferred aspect, the organic acid is 2,5-diketo-D-gluconic acid. In another more preferred aspect, the organic acid is formic acid. In another more preferred aspect, the organic acid is fumaric acid. In another more preferred aspect, the organic acid is glucaric acid. In another more preferred aspect, the organic acid is gluconic acid. In another more preferred aspect, the organic acid is glucuronic acid. In another more preferred aspect, the organic acid is glutaric acid. In another preferred aspect, the organic acid is 3-hydroxypropionic acid. In another more preferred aspect, the organic acid is itaconic acid. In another more preferred aspect, the organic acid is lactic acid. In another more preferred aspect, the organic acid is malic acid. In another more preferred aspect, the organic acid is malonic acid. In another more preferred aspect, the organic acid is oxalic acid. In another more preferred aspect, the organic acid is propionic acid. In another more preferred aspect, the organic acid is succinic acid. In another more preferred aspect, the organic acid is xylonic acid. See, for example, Chen, R., and Lee, Y. Y., 1997, Membrane-mediated extractive fermentation for lactic acid production from cellulosic biomass, *Appl. Biochem. Biotechnol.* 63-65: 435-448.

In another preferred aspect, the fermentation product is polyketide.

Recovery.

The fermentation product(s) can be optionally recovered from the fermentation medium using any method known in the art including, but not limited to, chromatography, electrophoretic procedures, differential solubility, distillation, or extraction. For example, alcohol is separated from the fermented cellulosic material and purified by conventional methods of distillation. Ethanol with a purity of up to about 96 vol. % can be obtained, which can be used as, for example, fuel ethanol, drinking ethanol, i.e., potable neutral spirits, or industrial ethanol.

The present invention is further described by the following examples that should not be construed as limiting the scope of the invention.

EXAMPLES

Strains

Trichoderma reesei strain 981-O-8 (D4) is a mutagenized strain of *Trichoderma reesei* RutC30 (ATCC 56765; Montencourt and Eveleigh, 1979, *Adv. Chem. Ser.* 181: 289-301).

Trichoderma reesei strain AgJg115-104-7B1 (PCT/US2010/061105, WO 2011/075677) is a ku70- derivative of *T. reesei* strain 981-O-8 (D4).

Media and Buffer Solutions

2XYT plus ampicillin plates were composed of 16 g of tryptone, 10 g of yeast extract, 5 g of sodium chloride, 15 g of Bacto agar, and deionized water to 1 liter. One ml of a 100 mg/ml solution of ampicillin was added after the autoclaved medium was cooled to 55° C.

SOC medium was composed of 20 g of Bacto-tryptone, 5 g of Bacto yeast extract, 0.5 g of NaCl, 2.5 ml of 1 M KCl, and deionized water to 1 liter. The pH was adjusted to 7.0 with 10 N NaOH before autoclaving. Then 20 ml of sterile 1 M glucose was added immediately before use.

LB plates were composed of 10 g of tryptone, 5 g of yeast extract, 5 g of NaCl, 15 g of Bacto agar, and deionized water to 1 liter.

COVE salt solution was composed of 26 g of KCl, 26 g of MgSO₄·7H₂O, 76 g of KH₂PO₄, 50 ml of COVE trace metals solution, and deionized water to 1 liter.

COVE trace metals solution was composed of 0.04 g of NaB₄O₇·10H₂O, 0.4 g of CuSO₄·5H₂O, 1.2 g of FeSO₄·7H₂O, 0.7 g of MnSO₄·H₂O, 0.8 g of Na₂MoO₄·2H₂O, 10 g of ZnSO₄·7H₂O, and deionized water to 1 liter.

COVE plates were composed of 342.3 g of sucrose, 20 ml of COVE salt solution, 10 ml of 1 M acetamide, 10 ml of 1.5 M CsCl, 25 g of Noble agar (Difco), and deionized water to 1 liter.

COVE2 plates were composed of 30 g of sucrose, 20 ml of COVE salt solution, 10 ml of 1 M acetamide, 25 g of Noble agar (Difco), and deionized water to 1 liter.

Trichoderma trace metals solution was composed of 216 g of FeCl₃·6H₂O, 58 g of ZnSO₄·7H₂O, 27 g of MnSO₄·H₂O, 10 g of CuSO₄·5H₂O, 2.4 g of H₃BO₃, 336 g of citric acid, and deionized water to 1 liter.

CIM medium was composed of 20 g of cellulose, 10 g of corn steep solids, 1.45 g of (NH₄)₂SO₄, 2.08 g of KH₂PO₄, 0.28 g of CaCl₂, 0.42 g of MgSO₄·7H₂O, 0.42 ml of *Trichoderma* trace metals solution, 1-2 drops of antifoam, and deionized water to 1 liter; pH adjusted to 6.0.

YP medium was composed of 10 g of yeast extract, 20 g of Bacto peptone, and deionized water to 1 liter.

YPG medium was composed of 4 g of yeast extract, 1 g of K₂HPO₄, 0.5 g of MgSO₄, 15.0 g of glucose, and deionized water to 1 liter (pH 6.0).

PEG buffer was composed of 500 g of polyethylene glycol 4000 (PEG 4000), 10 mM CaCl₂, 10 mM Tris-HCl pH 7.5, and deionized water to 1 liter; filter sterilized.

PDA plates were composed of 39 g of Potato Dextrose Agar (Difco) and deionized water to 1 liter.

PDA overlay medium was composed of 39 g of Potato Dextrose Agar (Difco), 2.44 g of uridine, and deionized water to 1 liter. The previously autoclaved medium was melted in a microwave and then cooled to 55° C. before use.

STC was composed of 1 M sorbitol, 10 mM CaCl₂, and 10 mM Tris-HCl, pH 7.5; filter sterilized.

TE buffer was composed of 1 M Tris pH 8.0 and 0.5 M EDTA pH 8.0.

20×SSC was composed of 175.3 g of NaCl, 88.2 g of sodium citrate, and deionized water to 1 liter.

TrMM-G medium was composed of 20 ml of COVE salt solution, 6 g of (NH₄)₂SO₄, 0.6 g of CaCl₂, 25 g of Nobel agar (Difco), 20 g of glucose, and deionized water to 1 liter.

NZY+ medium was composed of 5 g of NaCl, 3 g of MgSO₄·7H₂O, 5 g of yeast extract, 10 g of NZ amine, 1.2 g of MgCl₂, 4 g of glucose, and deionized water to 1 liter.

NNCYP07-PCS medium was composed of 5.0 g of NaNO₃, 3.0 g of NH₄Cl, 2.0 g of MES (free acid), 2.5 g of citric acid, 0.2 g of CaCl₂·2H₂O, 1.0 g of Bacto Peptone, 5.0 g of yeast extract, 0.2 g of MgSO₄·7H₂O, 4.0 g of K₂HPO₄, 1.0 ml of COVE trace metals solution, 80.0 g of pretreated corn stover (PCS), and deionized water to 1 liter.

Example 1: Construction of a *Trichoderma reesei* cbhl-*Aspergillus fumigatus* cbhl Replacement Construct pJfyS139

The *Aspergillus fumigatus* cellobiohydrolase I (cbhl) coding sequence (SEQ ID NO: 1 [DNA sequence] and SEQ ID NO: 2 [deduced amino acid sequence]) was amplified from pEJG93 (WO 2011/057140) using the gene-specific forward and reverse primers shown below. The region in italics represents vector homology to the site of insertion for an IN-FUSION® reaction and the underlined portion is an introduced Pac I site.

Forward primer:

5'-cgcggaactgcccaccATGCTGGCCTCCACCTTCTCCTACC-3' (SEQ ID NO: 43)

Reverse primer:

5'-ctttcgccacggagcttaattaaCTACAGGCACTGAGAGTAAT AATCA-3' (SEQ ID NO: 44)

The amplification reaction was composed of 20 ng of pEJG93, 200 μM dNTP's, 0.4 μM primers, 1× HERCULASE® Reaction Buffer (Stratagene, La Jolla, Calif., USA), and 1.875 units of HERCULASE® Hot Start High-Fidelity DNA Polymerase (Stratagene, La Jolla, Calif., USA) in a final volume of 50 μl. The amplification reaction was incubated in an EPPENDORF® MASTERCYCLER® 5333 eppgradient S (Eppendorf Scientific, Inc., Westbury, N.Y., USA) programmed for 1 cycle at 95° C. for 2 minutes; 30 cycles each at 95° C. for 30 seconds, 55° C. for 30 seconds, and 72° C. for 1 minute; and 1 cycle at 72° C. for 7 minutes. The PCR products were separated by 1% agarose gel electrophoresis using 40 mM Tris base, 20 mM sodium acetate, 1 mM disodium EDTA (TAE) buffer where a 1.6 kb fragment was excised from the gel and extracted using a MINELUTE® Gel Extraction Kit (QIAGEN Inc., Valencia, Calif., USA) according to the manufacturer's protocol.

The 1.6 kb PCR product was inserted into Nco I/Pac I-digested pSMai155 (WO 05/074647) using an IN-FUSION® Advantage PCR Cloning Kit (Clontech, Palo Alto, Calif., USA) according to the manufacturer's protocol. The IN-FUSION® reaction was composed of 1× IN-FUSION® Reaction Buffer (Clontech, Palo Alto, Calif., USA), 125 ng of Nco I/Pac I-digested pSMai155, 100 ng of the 1.6 kb PCR product, and 1 µl of IN-FUSION® Enzyme (Clontech, Palo Alto, Calif., USA) in a 10 µl reaction volume. The reaction was incubated for 15 minutes at 37° C. followed by 15 minutes at 50° C. After the incubation period 40 µl of TE buffer were added to the reaction. A 2 µl aliquot was used to transform ONE SHOT® TOP10 competent cells (Invitrogen, Carlsbad, Calif., USA) according to the manufacturer's protocol. The cells were heat shocked at 42° C. for 30 seconds and 250 µl of SOC medium were added. The tubes were incubated at 37° C., 200 rpm for 1 hour and 250 µl were plated onto 150 mm diameter 2XYT plus ampicillin plates and incubated at 37° C. overnight. The resulting transformants were screened by sequencing and one clone containing the insert with no PCR errors was identified and designated pJfyS139-A. Plasmid pJfyS139-A was used for insertion of the Herpes simplex virus thymidine kinase (tk) gene.

The Herpes simplex virus tk coding sequence (SEQ ID NO: 45 [DNA sequence] and SEQ ID NO: 46 [deduced amino acid sequence]) was liberated from pJfyS1579-8-6 (WO 2010/039840) by digesting the plasmid with Bgl II and Bam HI. The digestion was subjected to 1% agarose gel electrophoresis using TAE buffer where a 2.3 kb band was excised from the gel and extracted using a MINELUTE® Gel Extraction Kit. The tk gene cassette was inserted into Bam HI-digested, calf intestine phosphatase-treated pJfyS139-A using a QUICK LIGATION™ Kit (New England Biolabs, Inc., Ipswich, Mass. USA) according to the manufacturer's protocol. The ligation reaction was composed of 50 ng of the Bam HI-digested, calf intestine phosphatase-treated pJfyS139-A, 50 ng of the 2.3 kb tk gene insert, 1× QUICK LIGATION™ Buffer (New England Biolabs, Inc., Ipswich, Mass. USA), and 5 units of QUICK LIGASE™ (New England Biolabs, Inc., Ipswich, Mass. USA) in a final volume of 20 µl. The reaction was incubated at room temperature for 5 minutes and 2 µl of the reaction were used to transform ONE SHOT® TOP10 competent cells according to the manufacturer's protocol. The cells were heat shocked at 42° C. for 30 seconds and 250 µl of SOC medium were added. The tubes were incubated at 37° C., 200 rpm for 1 hour and 250 µl were plated onto 150 mm diameter 2XYT plus ampicillin plates and incubated at 37° C. overnight. The resulting transformants were screened by restriction digestion analysis with Xma I to determine the presence and orientation of the insert and a clone containing the insert was identified and designated pJfyS139-B. Plasmid pJfyS139-B was used for insertion of a *T. reesei* 3' cbhl gene flanking sequence.

The 3' cbhl gene flanking sequence was amplified from *T. reesei* RutC30 genomic DNA using the forward and reverse primers below. The underlined portion represents an introduced Not I site for cloning.

Forward Primer:

(SEQ ID NO: 47)
5'-ttagactgcgggccgcGTGGCGAAAGCCTGACGCACCGGTAGAT-3'

Reverse Primer:

(SEQ ID NO: 48)
5'-agtagttagcgggccgcACGGCACGGTTAAGCAGGGTCTTGC-3'

Trichoderma reesei RutC30 was grown in 50 ml of YP medium supplemented with 2% glucose (w/v) in a 250 ml

baffled shake flask at 28° C. for 2 days with agitation at 200 rpm. Mycelia were harvested by filtration using MIRACLOTH® (Calbiochem, La Jolla, Calif., USA), washed twice in deionized water, and frozen under liquid nitrogen. Frozen mycelia were ground by mortar and pestle to a fine powder. Total DNA was isolated using a DNEASY® Plant Maxi Kit (QIAGEN Inc., Valencia, Calif., USA) with the lytic incubation extended to 2 hours.

The amplification reaction was composed of 150 ng of *T. reesei* RutC30 genomic DNA, 200 µM dNTP's, 0.4 µM primers, 1× HERCULASE® Reaction Buffer, and 1.875 units of HERCULASE® Hot Start High-Fidelity DNA Polymerase in a final volume of 50 µl. The amplification reaction was incubated in an EPPENDORF® MASTERCYCLER® 5333 epgradient S programmed for 1 cycle at 95° C. for 2 minutes; 30 cycles each at 95° C. for 30 seconds, 60° C. for 30 seconds, and 72° C. for 1 minute 30 seconds; and 1 cycle at 72° C. for 7 minutes.

The PCR reaction was subjected to a MINELUTE® Nucleotide Removal Kit (QIAGEN Inc., Valencia, Calif., USA) according to the manufacturer's protocol. The resulting PCR mixture was digested with Not I and the digested PCR products were separated by 1% agarose gel electrophoresis using TAE buffer. A 1.3 kb fragment containing the 3' cbhl gene flanking sequence was excised from the gel and extracted using a MINELUTE® Gel Extraction Kit. The 1.3 kb fragment was inserted into Not I-linearized, calf intestine phosphatase-treated pJfyS139-B using a QUICK LIGATION™ Kit. The QUICK LIGATION™ reaction was composed of 100 ng of the Not I-linearized, calf intestine phosphatase-treated pJfyS139-B, 20 ng of the 1.3 kb fragment, 1× QUICK LIGATION™ Buffer, and 5 units of QUICK LIGASE™ in a final volume of 20 µl. The reaction was incubated at room temperature for 5 minutes and 2 µl of the reaction was used to transform ONE SHOT® TOP10 competent cells according to the manufacturer's protocol. The cells were heat shocked at 42° C. for 30 seconds and 250 µl of SOC medium were added. The tubes were incubated at 37° C., 200 rpm for 1 hour and 250 µl were plated onto 150 mm diameter 2XYT plus ampicillin plates and incubated at 37° C. overnight. The resulting transformants were screened by restriction digestion analysis with Xma I to determine the presence and orientation of the insert and positive clones were sequenced. A clone containing the 3' cbhl gene flanking sequence with no PCR errors was designated pJfyS139 (FIG. 1). Plasmid pJfyS139 was used as the vector to replace the *T. reesei* cbhl gene.

Example 2: *Trichoderma reesei* Protoplast Generation and Transformation

Protoplast preparation and transformation were performed using a modified protocol by Penttila et al., 1987, *Gene* 61: 155-164. Briefly, *Trichoderma reesei* strain AgJg115-104-7B1 (PCT/US2010/061105, WO 2011/075677) was cultivated in 25 ml of YP medium supplemented with 2% (w/v) glucose and 10 mM uridine at 27° C. for 17 hours with gentle agitation at 90 rpm. Mycelia were collected by filtration using a Vacuum Driven Disposable Filtration System (Millipore, Bedford, Mass., USA) and washed twice with deionized water and twice with 1.2 M sorbitol. Protoplasts were generated by suspending the washed mycelia in 20 ml of 1.2 M sorbitol containing 15 mg of GLUCANEX® 200 G (Novozymes A/S, Bagsvaerd, Denmark) per ml and 0.36 units of chitinase (Sigma Chemical Co., St. Louis, Mo., USA) per ml for 15-25 minutes at 34° C. with gentle shaking at 90 rpm. Protoplasts were

collected by centrifuging for 7 minutes at 400×g and washed twice with cold 1.2 M sorbitol. The protoplasts were counted using a haemocytometer and re-suspended to a final concentration of 1×10^8 protoplasts per ml in STC. Excess protoplasts were stored in a Cryo 1° C. Freezing Container (Nalgene, Rochester, N.Y., USA) at -80° C.

Approximately 100 µg of a transforming plasmid described in the following Examples were digested with Pme I. The digestion reaction was purified by 1% agarose gel electrophoresis using TAE buffer. A DNA band was excised from the gel and extracted using a QIAQUICK® Gel Extraction Kit (QIAGEN Inc., Valencia, Calif., USA). The resulting purified DNA was added to 100 µl of the protoplast solution and mixed gently. PEG buffer (250 µl) was added, mixed, and incubated at 34° C. for 30 minutes. STC (3 ml) was then added, mixed, and spread onto PDA plates supplemented with 1 M sucrose. After incubation at 28° C. for 16 hours, 20 ml of an overlay PDA medium supplemented with 35 µg of hygromycin B per ml were added to each plate. The plates were incubated at 28° C. for 4-7 days.

Example 3: Replacement of the Native
Trichoderma reesei cbhl Gene with the *Aspergillus fumigatus* cbhl Coding Sequence

In order to replace the *Trichoderma reesei* native cbhl gene (SEQ ID NO: 23 [DNA sequence] and SEQ ID NO: 24 [deduced amino acid sequence]) with the *Aspergillus fumigatus* cbhl coding sequence (SEQ ID NO: 1 [DNA sequence] and SEQ ID NO: 2 [deduced amino acid sequence]), *Trichoderma reesei* ku70- strain AgJg115-104-7B1 (PCT/US2010/061105, WO 2011/075677) was transformed with 4×2 µg of Pme I-linearized pJfyS139 (Example 1) according to the procedure described in Example 2. Seven transformants were obtained and each one was picked and transferred to a PDA plate and incubated for 7 days at 28° C. Genomic DNA was isolated from the transformants according to the procedure described in Example 1 and each transformant submitted to Southern analysis.

For Southern analysis, 2 µg of genomic DNA was digested with 33 units of Bgl II in a 50 µl reaction volume and subjected to 1% agarose electrophoresis in TAE buffer. The DNA in the gel was depurinated with one 10 minute wash in 0.25 N HCl, denatured with two 15 minute washes in 0.5 N NaOH-1.5 M NaCl, neutralized with one 30 minute wash in 1 M Tris pH 8-1.5 M NaCl, and incubated in 20×SSC for 5 minutes. The DNA was transferred to a NYTRAN® Supercharge membrane (Whatman, Inc., Florham Park, N.J., USA) using a TURBOBLOTTER™ System (Whatman, Inc., Florham Park, N.J., USA) according to the manufacturer's protocol. The DNA was UV crosslinked to the membrane using a STRATALINKER™ UV Crosslinker (Stratagene, La Jolla, Calif., USA) and prehybridized for 1 hour at 42° C. in 20 ml of DIG Easy Hyb (Roche Diagnostics Corporation, Indianapolis, Ind., USA).

A probe hybridizing to the 3' cbhl gene flanking sequence was generated using a PCR Dig Probe Synthesis Kit (Roche Diagnostics Corporation, Indianapolis, Ind., USA) according to the manufacturer's instructions with the forward and reverse primers shown below. The PCR reaction was composed of 1× HERCULASE® Reaction Buffer, 400 nM of each primer, 200 µM DIG-labeled dUTP-containing dNTPs, 20 ng of pJfyS139, and 1.5 units of HERCULASE® Hot Start High-Fidelity DNA Polymerase. The amplification reaction was incubated in an EPPENDORF® MASTERCYCLER® 5333 egradient S programmed for 1 cycle at 95° C. for 2 minutes; 25 cycles each at 95° C. for 30 seconds, 55° C. for 30 seconds, and 72° C. for 40 seconds; and 1 cycle at 72° C. for 7 minutes.

Forward primer: (SEQ ID NO: 49)
5'-AAAAACAAACATCCCGTTCATAAC-3'

Reverse primer: (SEQ ID NO: 50)
5'-AACAAAGGTTTACCGGTTTCGAAAAG-3'

The probe was purified by 1% agarose gel electrophoresis using TAE buffer where a 0.5 kb band corresponding to the probe was excised from the gel and extracted using a MINELUTE® Gel Extraction Kit. The probe was boiled for 5 minutes, chilled on ice for 2 minutes, and added to 10 ml of DIG Easy Hyb to produce the hybridization solution. Hybridization was performed at 42° C. for 15-17 hours. The membrane was then washed under low stringency conditions in 2×SSC plus 0.1% SDS for 5 minutes at room temperature followed by two high stringency washes in 0.5×SSC plus 0.1% SDS for 15 minutes each at 65° C. The probe-target hybrids were detected by chemiluminescent assay (Roche Diagnostics, Indianapolis, Ind., USA) according to the manufacturer's instructions. Southern analysis indicated that 3 of the 7 transformants contained the replacement cassette at the cbhl locus and one transformant, *T. reesei* JfyS139-8, was chosen for curing the hpt and tk markers.

A fresh plate of spores was generated by transferring spores of a 7 day old PDA plate grown at 28° C. to a PDA plate and incubating for 7 days at 28° C. Spores were collected in 10 ml of 0.01% TWEEN® 20 using a sterile spreader. The concentration of spores was determined using a hemocytometer and 10^5 spores were spread onto 150 mm plates containing TrMM-G medium supplemented with 1 µM 5-fluoro-2'-deoxyuridine (FdU).

Three hundred FdU-resistant spore isolates were obtained and DNA was extracted from 2 of the spore isolates as described above. The isolates were analyzed by Southern analysis as described above and the results indicated that both spore isolates had excised the hpt/tk region between the homologous repeats of the replacement cassette. One strain designated *T. reesei* JfyS139-8A was chosen for replacing the cbhl gene.

Example 4: Construction of an Empty *Trichoderma reesei* cbhl Replacement Construct pJfyS142

To generate a construct to replace the *Trichoderma reesei* cbhl gene (SEQ ID NO: 25 [DNA sequence] and SEQ ID NO: 26 [deduced amino acid sequence]) with the *Aspergillus fumigatus* cbhl coding sequence (SEQ ID NO: 3 [DNA sequence] and SEQ ID NO: 4 [deduced amino acid sequence]), the *T. reesei* cbhl promoter was first amplified from *T. reesei* RutC30 genomic DNA using the gene-specific forward and reverse primers shown below. The region in italics represents vector homology to the site of insertion in an IN-FUSION® reaction. *T. reesei* RutC30 genomic DNA was prepared according to the procedure described in Example

Forward primer: (SEQ ID NO: 51)
5'-acgaattgtttaaacgtcgacCCAAGTATCCAGAGGTGTATGGAAAT
ATCAGAT-3'

Reverse primer: (SEQ ID NO: 52)
5'-cgcgtagatctgcccgcattGGTGCAATACACAGAGGGTGTATCTT-3'

The amplification reaction was composed of 20 ng of *T. reesei* RutC30 genomic DNA, 200 µM dNTP's, 0.4 µM primers, 1× HERCULASE® Reaction Buffer, and 1.875

units of HERCULASE® Hot Start High-Fidelity DNA Polymerase in a final volume of 50 µl. The amplification reaction was incubated in an EPPENDORF® MASTERCYCLER® programmed for 1 cycle at 95° C. for 2 minutes; 25 cycles each at 95° C. for 30 seconds, 55° C. for 30 seconds, and 72° C. for 1 minute 30 seconds; and 1 cycle at 72° C. for 7 minutes. The PCR products were separated by 1% agarose gel electrophoresis using TAE buffer where a 1.6 kb fragment was excised from the gel and extracted using a MINELUTE® Gel Extraction Kit.

The 1.6 kb PCR product was inserted into Nco I/Sal I-digested pSMai155 (WO 05/074647) using an IN-FUSION® Advantage PCR Cloning Kit according to the manufacturer's protocol. The IN-FUSION® reaction was composed of 1× IN-FUSION® Reaction Buffer, 125 ng of the Nco I/Sal I-digested pSMai155, 100 ng of the 1.6 kb PCR product, and 1 µl of IN-FUSION® Enzyme in a 10 µl reaction volume. The reaction was incubated for 15 minutes at 37° C. and 15 minutes at 50° C. After the incubation period 40 µl of TE were added to the reaction. A 2 µl aliquot was used to transform ONE SHOT® TOP10 competent cells according to the manufacturer's protocol. The cells were heat shocked at 42° C. for 30 seconds and 250 µl of SOC medium were added. The tubes were incubated at 37° C., 200 rpm for 1 hour and 250 µl were plated onto 150 mm diameter 2XYT plus ampicillin plates and incubated at 37° C. overnight. The resulting transformants were screened by restriction digestion analysis with Pci I and positive clones sequenced to ensure the absence of PCR errors. One clone containing the insert with no PCR errors was identified and designated pJfyS142-A. Plasmid pJfyS142-A was used to insert the *T. reesei* cbhll terminator.

The cbhll terminator was amplified from *T. reesei* RutC30 genomic DNA using the gene-specific forward and reverse primers shown below. The region in italics represents vector homology to the site of insertion in an IN-FUSION® reaction.

Forward primer:

(SEQ ID NO: 53)

5' - *atctacgcgtactagttaattaa*GGCTTTCGTGACCGGGCTTCAA

ACA-3'

Reverse primer:

(SEQ ID NO: 54)

5' - *gcgggcgttactagtgatcc*ACTCGGAGTTGTTATACGCTAC

TCG-3'

The amplification reaction was composed of 150 ng of *T. reesei* RutC30 genomic DNA, 200 µM dNTP's, 0.4 µM primers, 1× HERCULASE® Reaction Buffer, and 1.875 units of HERCULASE® Hot Start High-Fidelity DNA Polymerase in a final volume of 50 µl. The amplification reaction was incubated in an EPPENDORF® MASTERCYCLER® programmed for 1 cycle at 95° C. for 2 minutes; 25 cycles each at 95° C. for 30 seconds, 54° C. for 30 seconds, and 72° C. for 50 seconds; and 1 cycle at 72° C. for 7 minutes. PCR products were separated by 1% agarose gel electrophoresis using TAE buffer where a 0.3 kb fragment was excised from the gel and extracted using a MINELUTE® Gel Extraction Kit.

The 0.3 kb PCR product was inserted into Pac I/Bam HI-digested pJfyS142-A using an IN-FUSION® Advantage PCR Cloning Kit according to the manufacturer's protocol. The IN-FUSION® reaction was composed of 1× IN-FUSION® Reaction Buffer, 150 ng of the PacI/Bam HI-

digested pJfyS142-A, 50 ng of the 0.3 kb PCR product, and 1 µl of IN-FUSION® Enzyme in a 10 µl reaction volume. The reaction was incubated for 15 minutes at 37° C. and 15 minutes at 50° C. After the incubation period 40 µl of TE were added to the reaction. A 2 µl aliquot was used to transform ONE SHOT® TOP10 competent cells according to the manufacturer's protocol. The cells were heat shocked at 42° C. for 30 seconds and 250 µl of SOC medium were added. The tubes were incubated at 37° C., 200 rpm for 1 hour and 250 µl were plated onto 150 mm diameter 2XYT plus ampicillin plates and incubated at 37° C. overnight. The transformants were screened by sequence analysis to identify positive clones and to ensure the absence of PCR errors. One clone containing the insert with no PCR errors was identified and designated pJfyS142-B. Plasmid pJfyS142-B was used for insertion of the Herpes simplex tk gene.

The Herpes simplex tk gene was liberated from pJfyS1579-8-6 (WO 2010/039840) by digesting the plasmid with Bgl II and Bam HI. The digestion was submitted to 1% agarose gel electrophoresis using TAE buffer where a 2.3 kb band was excised from the gel and extracted using a MINELUTE® Gel Extraction Kit. The tk cassette was inserted into Bam HI-digested, calf Intestine phosphatase-dephosphorylated pJfyS142-B using a QUICK LIGATION™ Kit according to the manufacturer's protocol. The ligation reaction was composed of 50 ng of the Bam HI-digested, calf Intestine phosphatase-dephosphorylated pJfyS142-B, 50 ng of the 2.3 kb tk gene insert, 1× QUICK LIGATION™ Buffer, and 5 units of QUICK LIGASE™ in a 20 µl ligation volume. The reaction was incubated at room temperature for 5 minutes and 2 µl of the reaction was used to transform ONE SHOT® TOP10 competent cells according to the manufacturer's protocol. The cells were heat shocked at 42° C. for 30 seconds and 250 µl of SOC medium were added. The tubes were incubated at 37° C., 200 rpm for 1 hour and 250 µl were plated onto 150 mm diameter 2XYT plus ampicillin plates and incubated at 37° C. overnight. The resulting transformants were screened by restriction digestion analysis with Xma I and Bam HI to determine the presence and orientation of the insert and a clone containing the insert was identified and designated pJfyS142-C. Plasmid pJfyS142-C was used for insertion of the *T. reesei* 3' cbhll gene flanking sequence.

The 3' cbhll gene flanking sequence was amplified from *T. reesei* RutC30 genomic DNA using the forward and reverse primers shown below. The region in italics represents vector homology to the site of insertion in an IN-FUSION® reaction.

Forward primer:

(SEQ ID NO: 55)

5' - *atccatcacactggcggccgc*GCTTCAAACAATGATGTGCGAT

GGT-3'

Reverse primer:

(SEQ ID NO: 56)

5' - *gatgcatgctcgagcggccgc*TACCTTGGCAGCCCTACGAGA

GAG-3'

The amplification reaction was composed of 150 ng of *T. reesei* RutC30 genomic DNA, 200 µM dNTP's, 0.4 µM primers, 1× HERCULASE® Reaction Buffer, and 1.875 units of HERCULASE® Hot Start High-Fidelity DNA Polymerase in a final volume of 50 µl. The amplification reaction was incubated in an EPPENDORF® MASTERCYCLER® programmed for 1 cycle at 95° C. for 2 minutes; 30

57

cycles each at 95° C. for 30 seconds, 56° C. for 30 seconds, and 72° C. for 1 minute 50 seconds; and 1 cycle at 72° C. for 7 minutes. The PCR reaction was subjected to 1% agarose gel electrophoresis using TAE buffer where a 1.5 kb band was excised from the gel and extracted using a MIN-ELUTE® Gel Extraction Kit. The 3' *cbhll* gene flanking sequence was inserted into Not I-linearized pJfyS142-C using an IN-FUSION® Advantage PCR Cloning Kit according to the manufacturer's protocol. The IN-FUSION® reaction was composed of 1× IN-FUSION® Reaction Buffer, 150 ng of pJfyS142-C, 80 ng of the 1.5 kb PCR product, and 1 µl of IN-FUSION® Enzyme in a 10 µl reaction volume. The reaction was incubated for 15 minutes at 37° C. and 15 minutes at 50° C. After the incubation period 40 µl of TE were added to the reaction. A 2 µl aliquot was used to transform ONE SHOT® TOP10 competent cells according to the manufacturer's protocol. The cells were heat shocked at 42° C. for 30 seconds and 250 µl of SOC medium were added. The tubes were incubated at 37° C., 200 rpm for 1 hour and 250 µl were plated onto 150 mm diameter 2XYT plus ampicillin plates and incubated at 37° C. overnight. The resulting transformants were screened by restriction digestion analysis with Bgl II and positive clones were sequenced to ensure the absence of PCR errors. One clone containing the insert with no PCR errors was identified and designated pJfyS142 (FIG. 2). Plasmid pJfyS142 was used to insert the *A. fumigatus* *cbhll* coding sequence.

Example 5: Construction of a *Trichoderma reesei* *cbhll-Aspergillus fumigatus* *cbhll* Replacement Construct pJfyS144

The *Aspergillus fumigatus* *cbhll* coding sequence (SEQ ID NO: 3 [DNA sequence] and SEQ ID NO: 4 [deduced amino acid sequence]) was amplified from pAIlO33 (WO 2011/057140) using the forward and reverse primers shown below. The region in italics represents vector homology to the site of insertion for an IN-FUSION® reaction.

Forward primer: (SEQ ID NO: 57)
5' - *ctctgtgtattgcacc*ATGAAGCACCTTGCATCTCCATCG-3'

Reverse primer: (SEQ ID NO: 58)
5' - *ccggtcacgaaagcc*TTAATTAAAAGGACGGGTTAGCGTT-3'

The amplification reaction was composed of 20 ng of pAIlO33, 200 µM dNTP's, 0.4 µM primers, 1 mM HERCULASE® Reaction Buffer, and 1.875 units of HERCULASE® Hot Start High-Fidelity DNA Polymerase in a final volume of 50 µl. The amplification reaction was incubated in an EPPENDORF® MASTERCYCLER® programmed for 1 cycle at 95° C. for 2 minutes; 30 cycles each at 95° C. for 30 seconds, 55° C. for 30 seconds, and 72° C. for 2 minutes; and 1 cycle at 72° C. for 7 minutes.

The PCR reaction was subjected to 1% agarose gel electrophoresis using TAE buffer where a 1.7 kb band was excised from the gel and extracted using a MINELUTE® Gel Extraction Kit. The 1.7 kb PCR product was inserted into Nco I/Pac I-digested pJfyS142 (Example 4) using an IN-FUSION® Advantage PCR Cloning Kit according to the manufacturer's protocol. The IN-FUSION® reaction was composed of 1× IN-FUSION® Reaction Buffer, 120 ng of the Nco II Pac I-digested pJfyS142, 70 ng of the 1.7 kb PCR product, and 1 µl of IN-FUSION® Enzyme in a 10 µl reaction volume. The reaction was incubated for 15 minutes

58

at 37° C. and 15 minutes at 50° C. After the incubation period 40 µl of TE were added to the reaction. A 2 µl aliquot was used to transform ONE SHOT® TOP10 competent cells according to the manufacturer's protocol. The cells were heat shocked at 42° C. for 30 seconds and 250 µl of SOC medium were added. The tubes were incubated at 37° C., 200 rpm for 1 hour and 250 µl were plated onto 150 mm diameter 2XYT plus ampicillin plates and incubated at 37° C. overnight. The resulting transformants were sequenced to ensure the absence of PCR errors and determine the presence of the insert. One clone with error-free sequence was identified and designated pJfyS144 (FIG. 3). Plasmid pJfyS144 was used to replace the native *cbhll* gene with the *cbhll* coding sequence from *A. fumigatus*.

Example 6: Replacement of the Native *Trichoderma reesei* *cbhll* Gene with the *Aspergillus fumigatus* *cbhll* Coding Sequence

In order to replace the native *T. reesei* *cbhll* gene (SEQ ID NO: 25 [DNA sequence] and SEQ ID NO: 26 [deduced amino acid sequence]) with the *Aspergillus fumigatus* *cbhll* coding sequence (SEQ ID NO: 3 [DNA sequence] and SEQ ID NO: 4 [deduced amino acid sequence]), *Trichoderma reesei* JfyS139-8A (Example 3) was transformed according to the procedure described in Example 2 with 2 µg of Pme I-linearized and gel purified pJfyS144 (Example 5). Seven transformants were obtained and each one was picked and transferred to a PDA plate and incubated for 7 days at 28° C. A fungal spore PCR method described below was used to screen for transformants bearing gene replacement using the forward primer shown below annealing to a region upstream of the 5' *cbhll* gene flanking sequence beyond the region of integration, and the reverse primer shown below annealing in the *A. fumigatus* *cbhll* coding sequence.

Forward primer: (SEQ ID NO: 59)
5' - AGCCACATGCCGCATATTGACAAAG-3'

Reverse primer: (SEQ ID NO: 60)
5' - AGGGATTCAAGTGTGCTACAGGCTGC-3'

A 1.8 kb PCR product would be generated only upon the occurrence of a precise gene replacement at the *cbhll* locus. If the cassette had integrated elsewhere in the genome, no amplification would result.

A small amount of spores from each transformant was suspended in 25 µl of TE buffer and heated on high in a microwave oven for 1 minute. Each microwaved spore suspension was used as a template in the PCR reaction. The reaction was composed of 1 µl of the microwaved spore suspension, 1 µl of a 10 mM dNTPs, 12.5 µl of 2× ADVANTAGE® GC-Melt LA Buffer (Clontech, Mountain View, Calif., USA), 25 pmol of forward primer, 25 pmol of reverse primer, 1.25 units of ADVANTAGE® GC Genomic LA Polymerase Mix (Clontech, Mountain View, Calif., USA), and 9.25 µl of water. The reaction was incubated in an EPPENDORF® MASTERCYCLER® 5333 epgradient S programmed for 1 cycle at 95° C. for 10 minutes; 35 cycles each at 95° C. for 30 seconds, 56° C. for 30 seconds, and 72° C. for 1 minute 40 seconds; 1 cycle at 72° C. for 7 minutes; and a 4° C. hold. The PCR reactions were subjected to 1% agarose gel electrophoresis using TAE buffer. The spore PCR indicated that four of the seven transformants contained the replacement cassette at the targeted locus and

three of them were submitted to Southern analysis to confirm the replacement cassette was in a single copy.

Genomic DNA was isolated from the three transformants according to the procedure described in Example 1 and each transformant submitted to Southern analysis. For Southern analysis, 2 μ g of genomic DNA was digested with 50 units of Dra I in a 50 μ l reaction volume and subjected to 1% agarose electrophoresis in TAE buffer. The DNA in the gel was depurinated with one 10 minute wash in 0.25 N HCl, denatured with two 15 minute washes in 0.5 N NaOH-1.5 M NaCl, neutralized with one 30 minute wash in 1 M Tris pH 8-1.5 M NaCl, and incubated in 20 \times SSC for 5 minutes. The DNA was transferred to a NYTRAN[®] Supercharge membrane. The DNA was UV crosslinked to the membrane using a STRATALINKER[™] UV crosslinker and prehybridized for 1 hour at 42 $^{\circ}$ C. in 20 ml of DIG Easy Hyb.

A probe hybridizing to the 3' cbhll gene flanking sequence was generated using a PCR Dig Probe Synthesis Kit according to the manufacturer's instructions with the forward and reverse primers indicated below. The PCR reaction was composed of 1 \times HERCULASE[®] Reaction Buffer, 400 nM each primer, 200 μ M DIG-labeled dUTP-containing dNTPs, 150 ng of *T. reesei* RutC30 genomic DNA, and 1.5 units of HERCULASE[®] Hot Start High-Fidelity DNA Polymerase. The reaction was incubated in an EPPENDORF[®] MASTERCYCLER[®] 5333 egradient S programmed for 1 cycle at 95 $^{\circ}$ C. for 2 minutes; 30 cycles each at 95 $^{\circ}$ C. for 30 seconds, 51 $^{\circ}$ C. for 30 seconds, and 72 $^{\circ}$ C. for 40 seconds; and 1 cycle at 72 $^{\circ}$ C. for 7 minutes.

Forward primer: (SEQ ID NO: 61)
5' -AAAAACAAACATCCCGTTCATAAC-3'

Reverse primer: (SEQ ID NO: 62)
5' -ACAAGGTTTACCGGTTTCGAAAAG-3'

The probe was purified by 1% agarose gel electrophoresis using TAE buffer where a 0.5 kb band corresponding to the probe was excised from the gel and extracted using a QIAQUICK[®] Gel Extraction Kit. The probe was boiled for 5 minutes, chilled on ice for 2 minutes, and added to 10 ml of DIG Easy Hyb to produce the hybridization solution. Hybridization was performed at 42 $^{\circ}$ C. for approximately 17 hours. The membrane was then washed under low stringency conditions in 2 \times SSC plus 0.1% SDS for 5 minutes at room temperature followed by two high stringency washes in 0.5 \times SSC plus 0.1% SDS for 15 minutes each at 65 $^{\circ}$ C. The probe-target hybrids were detected by chemiluminescent assay (Roche Diagnostics, Indianapolis, Ind., USA) according to the manufacturer's instructions. Southern analysis indicated that the three transformants contained the replacement cassette at the cbhll locus and all three (designated JfyS139/144-5, -6, and -10) were chosen for curing the hpt and tk markers.

A fresh plate of spores for each transformant was generated by transferring a plug of a 7 day old culture grown on a PDA plate at 28 $^{\circ}$ C. to a new PDA plate and incubating for 7 days at 28 $^{\circ}$ C. Spores were collected in 10 ml of 0.01% TWEEN[®] 20 using a sterile spreader. The concentration of spores was determined using a hemacytometer and 10⁵ and 10⁴ spores were spread onto 150 mm plates containing TrMM-G medium supplemented with 1 μ M FdU.

Approximately 500 FdU-resistant spore isolates for each transformant were obtained from the plate containing 10⁵ spores and approximately 100 FdU-resistant spore isolates

for each transformant from the plate containing 10⁴ spores. Eight spore isolates were picked for strains JfyS139/144-5 and -6 and four were picked for strain JfyS139/144-10. Each isolate 1 to 8 from primary transformant 5 was designated JfyS139/144-5A to -5H. Isolates 1 to 8 from primary transformant 6 were designated JfyS139/144-6A to 6H. Isolates from primary transformant 10 were designated JfyS139/144-10A to 10D for isolates 1 to 4. Spore PCR was conducted as described above, using the forward and reverse primers shown below, to confirm the hpt and tk markers had been correctly excised.

Forward primer: (SEQ ID NO: 63)
5' -GTTAAGCATACAATTGAACGAGAATGG-3'

Reverse primer: (SEQ ID NO: 64)
5' -GATGATATAATGGAGCAAATAAGGG-3'

The PCR reactions were performed as described above with the following cycling parameters: 1 cycle at 95 $^{\circ}$ C. for 2 minutes; 30 cycles each at 95 $^{\circ}$ C. for 30 seconds, 55 $^{\circ}$ C. for 30 seconds, and 72 $^{\circ}$ C. for 6 minutes seconds; and 1 cycle at 72 $^{\circ}$ C. for 7 minutes.

The primers annealed to the 5' (forward) and 3' (reverse) flanking sequences used for the cbhll gene replacement. Strains from which the hpt/tk cassette had been correctly excised would display a 3.5 kb fragment while those with the markers intact would display an 8 kb fragment. The PCR screen indicated that all of the spore isolates had correctly excised the hpt/tk cassette.

DNA was extracted from the A and B spore isolates from each primary transformant and submitted to Southern analysis as described above. The Southern analysis confirmed that each spore isolate had correctly excised the hpt/tk cassette. Spore isolate *T. reesei* JfyS139/144-10B was chosen to represent the strain containing both the *T. reesei* cbhl and cbhll genes replaced with the respective homologs from *Aspergillus fumigatus*.

Example 7: Generation of *Trichoderma reesei* Ku70 Gene Repair Plasmid pTH239

Four DNA segments were combined using an IN-FUSION[®] Advantage PCR Cloning Kit to generate a construct to replace the disrupted *Trichoderma reesei* ku70 coding sequence with the native *Trichoderma reesei* ku70 coding sequence (SEQ ID NO: 65 [DNA sequence] and SEQ ID NO: 66 [deduced amino acid sequence]). The ampicillin resistance marker region including the prokaryotic origin of replication was amplified from pJfyS139-B (Example 1) using the sequence-specific forward and reverse primers shown below (SEQ ID NOs: 67 and 68). The *T. reesei* ku70 gene upstream sequence (consisting of 989 bp from upstream of the ku70 coding sequence and the first 1010 bp of the ku70 coding sequence) was amplified from *T. reesei* 981-O-8 genomic DNA using the sequence-specific forward and reverse primers shown below (SEQ ID NOs: 69 and 70). The *T. reesei* ku70 gene downstream sequence (consisting of a 500 bp segment repeated from the 3' end of the 1010 bp segment of the ku70 coding sequence amplified in the upstream PCR product, and a 1067 bp segment containing the remainder of the ku70 coding sequence, and 461 bp from downstream of the ku70 coding sequence) was amplified from *T. reesei* 981-O-8 genomic DNA using the sequence-specific forward and reverse primers shown below (SEQ ID

61

NOs: 71 and 72). *T. reesei* 981-O-8 genomic DNA was prepared according to the procedure described in Example 1.

Forward primer:

(SEQ ID NO: 67)
5' -GTGTGCGGCCGCTCGAGCATGCATGTTTAAACAGCTTGGCACTGGCC
GTCGTTTT-3'

Reverse primer:

(SEQ ID NO: 68)
5' -ATCAGCCCCGAGACGGCGCCGCGTTTAAACAATTCTAATCATGGTC
ATAGCTGT-3'

Forward primer:

(SEQ ID NO: 69)
5' -CATGATTACGAATTGTTTAAACGCGCGCCGCTCTCGGGGCTGATCTT
GTCGAGGA-3'

Reverse primer:

(SEQ ID NO: 70)
5' -GGCGGCCGTTACTAGTGGATCCAGCCCTTGACAGTGATCTTGAGTCC
AGGTGCAA-3'

Forward primer:

(SEQ ID NO: 71)
5' -TGCAGATATCCATCACACTGGCGGCCGAGTTTCCATGTCCAACGTG
TTGTTTTGCGC-3'

Reverse primer:

(SEQ ID NO: 72)
5' -GCCAGTGCCAAGCTGTTTAAACATGCATGCTCGAGCGGCCGCACAG
CCCTCTCCTCG-3'

For amplification of the ampicillin resistance marker and prokaryotic origin of replication region, the reaction was composed of 100 ng of *T. reesei* 981-O-8 genomic DNA, 200 μ M dNTPs, 1 μ M of each primer (SEQ ID NO: 67 and 70), 1 \times PHUSION[®] High-Fidelity Hot Start DNA Polymerase Buffer (New England Biolabs, Inc., Ipswich, Mass., USA), and 1.0 unit of PHUSION[®] High-Fidelity Hot Start DNA Polymerase (New England Biolabs, Inc., Ipswich, Mass., USA) in a final volume of 50 μ l. The amplification reaction was incubated in an EPPENDORF[®] MASTERCYCLER[®] 5333 egradient S programmed for 1 cycle at 98 $^{\circ}$ C. for 30 seconds; 30 cycles each at 98 $^{\circ}$ C. for 10 seconds, 55 $^{\circ}$ C. for 30 seconds, and 72 $^{\circ}$ C. for 1 minute 30 seconds; and 1 cycle at 72 $^{\circ}$ C. for 7 minutes. The PCR product was separated by 1% agarose gel electrophoresis using TAE buffer where a 2.692 kb fragment was excised from the gels and extracted using a MINELUTE[®] Gel Extraction Kit.

For amplification of the ku70 gene upstream sequence or downstream sequence, the reactions were composed of 100 ng of pJfyS139-B, 200 μ M dNTPs, 1 μ M of each primer (SEQ ID NOs: 69 and 70 or 71 and 72, respectively), 1 \times PHUSION[®] High-Fidelity Hot Start DNA Polymerase Buffer, and 1.0 unit of PHUSION[®] High-Fidelity Hot Start DNA Polymerase in a final volume of 50 μ l. The amplification reactions were incubated in an EPPENDORF[®] MASTERCYCLER[®] 5333 egradient S programmed for 1 cycle at 98 $^{\circ}$ C. for 30 seconds; 30 cycles each at 98 $^{\circ}$ C. for 10 seconds, 55 $^{\circ}$ C. for 30 seconds, and 72 $^{\circ}$ C. for 1 minute 30 seconds; and 1 cycle at 72 $^{\circ}$ C. for 7 minutes. The PCR products were separated by 1% agarose gel electrophoresis using TAE buffer where 1.999 kb and 2.028 kb fragments were separately excised from the gels and extracted using a MINELUTE[®] Gel Extraction Kit.

62

The fourth DNA segment was generated from a restriction enzyme digestion of pJfyS139-B with Not I and Bam HI. The reaction was composed of 5 μ g of pJfyS139-B, 10 units of Not I, 20 units of Bam HI, and 20 μ l of Restriction Enzyme Buffer 2 (New England Biolabs, Inc., Ipswich, Mass., USA) in a total volume of 50 μ l. The reaction was incubated for 1 hour at 37 $^{\circ}$ C. and then separated by 1% agarose gel electrophoresis using TAE buffer where a 4.400 kb fragment was excised from the gel and extracted using a MINELUTE[®] Gel Extraction Kit.

The three PCR products of 2,028 bp, 1,999 bp and 2,692 bp were inserted into Not I and Bam HI-digested pJfyS139-B using an IN-FUSION[®] Advantage PCR Cloning Kit according to the manufacturer's protocol. The IN-FUSION[®] reaction was composed of 1 \times IN-FUSION[®] Reaction Buffer, 50 ng of the Not I/Bam HI-digested pJfyS139-B, 50 ng of the 1.999 kb ku70 gene upstream PCR product, 50 ng of the 2.028 kb ku70 gene downstream PCR product, 50 ng of the 2.692 kb ampicillin resistance marker and prokaryotic origin of replication PCR product, and 1 μ l of IN-FUSION[®] Enzyme in a 10 μ l reaction volume. The reaction was incubated for 15 minutes at 37 $^{\circ}$ C. followed by 15 minutes at 50 $^{\circ}$ C. After the incubation period 40 μ l of TE were added to the reaction. A 3 μ l aliquot was used to transform *E. coli* XL10 GOLD[®] competent cells (Stratagene, La Jolla, Calif., USA) according to the manufacturer's protocol. The cells were heat shocked at 42 $^{\circ}$ C. for 30 seconds and then 500 μ l of NZY+ medium, pre-heated to 42 $^{\circ}$ C., were added. The tubes were incubated at 37 $^{\circ}$ C. with shaking at 200 rpm for 40 minutes and then plated onto 150 mm diameter 2XYT plus ampicillin plates and incubated at 37 $^{\circ}$ C. overnight. The resulting transformants were screened by restriction digestion analysis with Hind III and Xba I and positive clones sequenced to ensure the absence of PCR errors. One clone containing the insert with no PCR errors was identified and designated pTH239.

Example 8: Repair of the Ku70 Gene in the *A. fumigatus* cbh1 and cbh2 Replacement Strain JfyS139/144-10B

The native *Trichoderma reesei* ku70 gene was repaired in strain *T. reesei* JfyS139/144-10B (Example 6) in order to facilitate strain manipulation steps requiring the function of the ku70 gene in non-homologous end-joining. *T. reesei* JfyS129/144-10B was transformed with 23 \times 2 μ g of Pme I-linearized pTH239 (Example 7) according to the procedure described in Example 2. Nineteen transformants were obtained and each one was separately transferred to a PDA plate and incubated for 7 days at 28 $^{\circ}$ C.

All nineteen transformants were screened by PCR to confirm homologous integration of the pTH239 Pme I fragment at the disrupted ku70 gene locus. For each of the transformants a sterile inoculation loop was used to collect spores from a 7 day old PDA plate. The spores were transferred to a tube containing 25 μ l of 1 mM EDTA-10 mM Tris buffer and microwaved on high for 1 minute. A 1 μ l aliquot of the microwaved spore mixture was added directly to the PCR reaction as template DNA. A set of PCR primers shown below were designed to amplify across the disrupted region of the ku70 coding sequence to distinguish between the host genome with the disruption in the ku70 coding sequence (848 bp) and the pTH239 targeted strain of interest (606 bp). The PCR reaction was composed of 1 \times ADVANTAGE[®] Genomic LA Polymerase Reaction Buffer (Clontech, Mountain View, Calif., USA), 400 nM of each primer, 200 μ M dNTPs, 1 μ l of microwaved TE-spore

mixture (described above), and 1.0 unit of ADVANTAGE® Genomic LA Polymerase (Clontech, Mountain View, Calif., USA). The amplification reaction was incubated in an EPPENDORF® MASTERCYCLER® 5333 egradient S programmed for 1 cycle at 95° C. for 10 minutes; 30 cycles each at 95° C. for 30 seconds, 55° C. for 30 seconds, and 72° C. for 60 seconds; and 1 cycle at 72° C. for 7 minutes.

Forward primer: (SEQ ID NO: 73)
5'-CAATGACGATCCGCACGCGT-3'

Reverse primer: (SEQ ID NO: 74)
5'-CAATGACGATCCGCACGCGT-3'

Only one of the nineteen transformants (#19) was positive for the 606 bp PCR product and negative for the 848 bp PCR product indicative of a strain containing the pTH239 PmeI fragment homologously integrated at the ku70 locus.

Spores from the 7 day old PDA plate of transformant #19 were collected in 10 ml of 0.01% TWEEN® 20 using a sterile spreader. The concentration of spores was determined using a hemocytometer and 10⁶ spores were spread onto 150 mm plates containing TrMM-G medium supplemented with 1 μM 5-fluoro-2'-deoxyuridine (FdU) and cultured for 5 days at 28° C. Twenty-two FdU-resistant spore isolates were obtained and transferred to PDA plates and cultivated at 28° C. for five days.

All twenty-two spore isolates (#19A-V) were screened by PCR for excision of the hpt/tk marker region present between the homologous repeats of the ku70 coding sequence within the repair cassette. For each of the spore isolates a sterile inoculating loop was used to collect spores from a 7 day old PDA plate. The spores were transferred to a tube containing 25 μl of 1 mM EDTA-10 mM Tris buffer and microwaved on high for 1 minute. A 1 μl aliquot of the spore mixture was added directly to the PCR reaction as template genomic DNA. A set of PCR primers shown below were designed to amplify across the hpt/tk region to distinguish between the presence (6 kb) or absence (1.1 kb) of the hpt/tk region. The PCR reaction was composed of 1× ADVANTAGE® Genomic LA Polymerase Reaction Buffer, 400 nM of each primer (below), 200 μM dNTPs, 1 μl of microwaved TE-spore mixture (described above), and 1.0 unit of ADVANTAGE® Genomic LA Polymerase. The amplification reaction was incubated in an EPPENDORF® MASTERCYCLER® 5333 egradient S programmed for 1 cycle at 95° C. for 10 minutes; 30 cycles each at 95° C. for 30 seconds, 50° C. for 30 seconds, and 72° C. for 6 minutes; and 1 cycle at 72° C. for 7 minutes.

Forward primer: (SEQ ID NO: 75)
5'-GACTCTTTTCTCCCATCT-3'

Reverse primer: (SEQ ID NO: 76)
5'-GAGGAGCAGAAGAAGCTCCG-3'

All twenty-two spore isolates were negative for the 6 kb PCR product corresponding to the hpt/tk marker region.

Spores from the 7 day old PDA plates of isolates #19A and #19L were collected in 10 ml of 0.01% TWEEN® 20 using a sterile spreader. The concentration of spores was determined using a hemocytometer and 10³, 10², and 10¹ spores were spread onto 150 mm PDA plates containing 1 M sucrose and cultured for 3 days at 28° C. Ten spore isolates

were selected from the PDA plates for both strains #19A and #19L and transferred to fresh PDA plates and placed at 28° C.

Genomic DNA was extracted from 6 spore isolates of both #19L and #19A, according to the procedure described in Example 1 and submitted to Southern analysis.

For Southern analysis, 2 μg of genomic DNA was digested with (1) 5 units and 10 units, respectively, of Asc I and Xho I or (2) 5 units and 25 units, respectively, of Asc I and Apa I in a 50 μl reaction volume and subjected to 1% agarose electrophoresis using TAE buffer. The DNA in the gel was depurinated with one 10 minute wash in 0.25 N HCl, denatured with two 15 minute washes in 0.5 N NaOH-1.5 M NaCl, neutralized with one 30 minute wash in 1 M Tris pH 8-1.5 M NaCl, and incubated in 20×SSC for 5 minutes. The DNA was transferred to a NYTRAN® Supercharge membrane using a TURBOBLOTTER™ System according to the manufacturer's protocol. The DNA was UV crosslinked to the membrane using a STRATALINKER™ UV Crosslinker and prehybridized for 1 hour at 42° C. in 20 ml of DIG Easy Hyb.

A probe hybridizing to the 3' end of the ku70 coding sequence was generated using a PCR Dig Probe Synthesis Kit (Roche Diagnostics Corporation, Indianapolis, Ind., USA) according to the manufacturer's instructions with the forward and reverse primers shown below. In order to generate a pure template for the probe PCR reaction, the 3' end of the ku70 coding sequence was amplified from *T. reesei* 981-O-8 genomic DNA. The PCR reaction was composed of 1× PHUSION® High-Fidelity Hot Start DNA Polymerase Buffer, 1 μM of each primer, 200 μM dNTPs, 165 ng of *T. reesei* 981-O-8 genomic DNA, and 1.0 unit of PHUSION® High-Fidelity Hot Start DNA Polymerase. The amplification reaction was incubated in an EPPENDORF® MASTERCYCLER® 5333 egradient S programmed for 1 cycle at 98° C. for 30 seconds; 35 cycles each at 98° C. for 10 seconds, 60° C. for 30 seconds, and 72° C. for 15 seconds; and 1 cycle at 72° C. for 10 minutes.

Forward primer: (SEQ ID NO: 77)
5'-gcatatataaaccactcaagta-3'

Reverse primer: (SEQ ID NO: 78)
5'-attatcttgaccggcgccagg-3'

The 0.5 kb probe template was purified by 1% agarose gel electrophoresis using TAE buffer and excised from the gel and extracted using a MINELUTE® Gel Extraction Kit. The purified PCR product was used to generate a DIG-labeled probe as specified by the manufacturer's instructions using the primers and amplification conditions specified above. The 0.5 kb DIG-labeled probe was purified by 1% agarose gel electrophoresis using TAE buffer and excised from the gel and extracted using a MINELUTE® Gel Extraction Kit. The probe was boiled for 5 minutes, chilled on ice for 2 minutes, and added to 10 ml of DIG Easy Hyb to produce the hybridization solution. Hybridization was performed at 42° C. for 15-17 hours. The membrane was then washed under low stringency conditions in 2×SSC plus 0.1% SDS for 5 minutes at room temperature followed by two high stringency washes in 0.5×SSC plus 0.1% SDS for 15 minutes each at 65° C. The probe-target hybrids were detected by chemiluminescent assay (Roche Diagnostics, Indianapolis, Ind., USA) according to the manufacturer's instructions. Southern analysis indicated that all spore isolates contained

the repair/replacement cassette at the ku70 locus and were cured of the hpt and tk markers. One strain designated *T. reesei* 981-O-8.5#10B+Ku70#19L3 was chosen for further transformations.

Example 9: Cloning of an *Aspergillus fumigatus* GH61B Polypeptide Gene

A tblastn search (Altschul et al., 1997, *Nucleic Acids Res.* 25: 3389-3402) of the *Aspergillus fumigatus* partial genome sequence (The Institute for Genomic Research, Rockville, Md., USA) was performed using as query several known GH61 polypeptides including the *Thermoascus aurantiacus* GH61A polypeptide (GeneSeqP Accession Number AEC05922). Several genes were identified as putative GH61 Family homologs based upon a high degree of similarity to the query sequences at the amino acid level. One genomic region of approximately 850 bp with greater than 70% sequence identity to the *Thermoascus aurantiacus* GH61A polypeptide sequence at the amino acid level was chosen for further study.

A. fumigatus NN051616 was grown and harvested as described in U.S. Pat. No. 7,244,605. Frozen mycelia were ground, by mortar and pestle, to a fine powder and genomic DNA was isolated using a DNEASY® Plant Maxi Kit according to manufacturer's instructions.

Two synthetic oligonucleotide primers shown below were designed to PCR amplify the *A. fumigatus* Family GH61B polypeptide coding sequence from the genomic DNA. An IN-FUSION® Cloning Kit (Clontech, Palo Alto, Calif., USA) was used to clone the fragment directly into the expression vector pAILo2 (WO 2004/099228), without the need for restriction digestion and ligation.

Forward primer:

(SEQ ID NO: 79)
5'-ACTGGATTTACCATGACTTTGTCCAAGATCACTTCCA-3'

Reverse primer:

(SEQ ID NO: 80)
5'-TCACCTCTAGTTAATTAAGCGTTGAACAGTGCAGGACCAG-3'

Bold letters represent coding sequence. The remaining sequences are homologous to insertion sites of pAILo2.

Fifty picomoles of each of the primers above were used in a PCR reaction composed of 204 ng of *A. fumigatus* genomic DNA, 1×Pfx Amplification Buffer (Invitrogen, Carlsbad, Calif., USA), 1.5 µl of a 10 mM blend of dATP, dTTP, dGTP, and dCTP, 2.5 units of PLATINUM® Pfx DNA Polymerase (Invitrogen Corp., Carlsbad, Calif., USA), and 1 µl of 50 mM MgSO₄ in a final volume of 50 µl. The amplification was performed using an EPPENDORF® MASTERCYCLER® 5333 egradient S programmed for 1 cycle at 94° C. for 3 minutes; and 30 cycles each at 94° C. for 30 seconds, 56° C. for 30 seconds, and 72° C. for 1 minutes. The heat block was then held at 72° C. for 15 minutes followed by a 4° C. soak cycle. The reaction products were isolated by 1.0% agarose gel electrophoresis using TAE buffer where an approximately 850 bp product band was excised from the gel and purified using a MIN-ELUTE® Gel Extraction Kit according to the manufacturer's instructions.

The 850 bp fragment was then cloned into pAILo2 using an IN-FUSION® Cloning Kit. Plasmid pAILo2 was digested with Nco I and Pac I. The plasmid fragment was purified by gel electrophoresis as above and a QIAQUICK® Gel Purification Kit. The gene fragment and the digested

vector were combined together in a reaction described below resulting in the expression plasmid pAG43 (FIG. 4) in which transcription of the *A. fumigatus* GH61B polypeptide coding sequence was under the control of the NA2-tpi promoter.

5 The NA2-tpi promoter is a modified promoter from the *Aspergillus niger* neutral alpha-amylase gene in which the untranslated leader has been replaced by an untranslated leader from the *Aspergillus nidulans* triose phosphate isomerase gene. The recombination reaction (20 µl) was composed of 1× IN-FUSION® Reaction Buffer, 1×BSA (Clontech, Palo Alto, Calif., USA), 1 µl of IN-FUSION® Enzyme (diluted 1:10), 166 ng of pAILo2 digested with Nco I and Pac I, and 110 ng of the *A. fumigatus* GH61B polypeptide purified PCR product. The reaction was incubated at 37° C. for 15 minutes followed by 15 minutes at 50° C. The reaction was diluted with 40 µl of 10 mM Tris-0.1 M EDTA buffer and 2.5 µl of the diluted reaction was used to transform *E. coli* XL10 SOLOPACK® Gold Competent Cells (Stratagene, La Jolla, Calif., USA). An *E. coli* transformant containing pAG43 (GH61B polypeptide coding sequence) was identified by restriction enzyme digestion and plasmid DNA was prepared using a BIOROBOT® 9600 (QIAGEN Inc., Valencia, Calif., USA).

25 DNA sequencing of the 862 bp PCR fragment was performed with an Applied Biosystems Model 377 XL Automated DNA Sequencer (Applied Biosystems, Carlsbad, Calif., USA) using dye-terminator chemistry (Giesecke et al., 1992, *Journal of Virology Methods* 38: 47-60) and primer walking strategy. The following vector specific primers were used for sequencing:

pAILo2 5 Seq:

(SEQ ID NO: 81)
5'-TGTCCCTTGTTCGATGCG 3'

pAILo2 3 Seq:

(SEQ ID NO: 82)
5'-CACATGACTTGGCTTCC 3'

40 Nucleotide sequence data were scrutinized for quality and all sequences were compared to each other with assistance of PHRED/PHRAP software (University of Washington, Seattle, Wash., USA).

45 A gene model for the *A. fumigatus* sequence was constructed based on similarity of the encoded protein to the *Thermoascus aurantiacus* GH61A protein (GeneSeqP Accession Number AEC05922). The nucleotide sequence and deduced amino acid sequence of the *A. fumigatus* GH61B polypeptide coding sequence are shown in SEQ ID NO: 7 and SEQ ID NO: 8, respectively. The genomic fragment encodes a polypeptide of 250 amino acids, interrupted by 2 introns of 53 and 56 bp. The % G+C content of the coding sequence and the mature coding sequence are 53.9% and 57%, respectively. Using the SignalP software program (Nielsen et al., 1997, *Protein Engineering* 10: 1-6), a signal peptide of 21 residues was predicted. The predicted mature protein contains 221 amino acids with a predicted molecular mass of 23.39 kDa.

Example 10: Construction of pSMai214 for Expression of the *Aspergillus fumigatus* GH61B Polypeptide

65 The *Aspergillus fumigatus* GH61B polypeptide coding sequence was amplified from plasmid pAG43 (Example 9) using the gene-specific forward and reverse primers shown

below. The region in italics represents vector homology to the site of insertion for an IN-FUSION® reaction.

Forward primer: (SEQ ID NO: 83)
5' - *GGACTGCGCACCATGACTTTGTCCAAGATCACTTCCA* - 3'

Reverse primer: (SEQ ID NO: 84)
5' - *GCCACGGAGCTTAATTAATTAAGCGTTGAACAGTGACG* - 3'

Fifty picomoles of each of the primers above were used in a PCR reaction composed of 10 ng of pAG43 DNA, 1×Pfx Amplification Buffer, 1.5 µl of a 10 mM blend of dATP, dTTP, dGTP, and dCTP, 2.5 units of PLATINUM® Pfx DNA Polymerase, and 1 µl of 50 mM MgSO₄ in a final volume of 50 µl. The amplification was performed using an EPPENDORF® MASTERCYCLER® 5333 egradient S programmed for 1 cycle at 98° C. for 3 minutes; and 30 cycles each at 98° C. for 30 seconds, 56° C. for 30 seconds, and 72° C. for 1 minute. The heat block was then held at 72° C. for 15 minutes. The PCR products were separated by 1% agarose gel electrophoresis using TAE buffer where an approximately 0.9 kb fragment was excised from the gel and extracted using a MINELUTE® Gel Extraction Kit according to the manufacturer's protocol.

Plasmid pMJ09 (WO 2005/047499) was digested with Nco I and Pac I, isolated by 1.0% agarose gel electrophoresis in 1 mM disodium EDTA-50 mM Tris base-50 mM boric acid (TBE) buffer, excised from the gel, and extracted using a QIAQUICK® Gel Extraction Kit according to the manufacturer's instructions.

The 0.9 kb PCR product was inserted into the gel-purified Nco I/Pac I digested pMJ09 using an IN-FUSION® PCR Cloning Kit (Clontech, Palo Alto, Calif., USA) according to the manufacturer's protocol. The IN-FUSION® reaction was composed of 1× IN-FUSION® Reaction Buffer, 100 ng of the gel-purified Nco I/Pac I digested pMJ09, 37 ng of the 0.9 kb PCR product, 2 µl of 500 µg/ml BSA, and 1 µl of IN-FUSION® Enzyme in a 20 µl reaction volume. The reaction was incubated for 15 minutes at 37° C. and 15 minutes at 50° C. After the incubation period 30 µl of TE buffer were added to the reaction. A 2.5 µl aliquot was used to transform SOLOPACK® Gold Supercompetent Cells according to the manufacturer's protocol. Transformants were screened by sequencing and one clone containing the insert with no PCR errors was identified and designated pSMai214 (FIG. 5). Plasmid pSMai214 can be digested with Pme I to generate an approximately 5.4 kb fragment for *T. reesei* transformation. The 5.4 kb fragment contains the expression cassette composed of the *T. reesei* Cel7A cellobiohydrolase I gene promoter, *A. fumigatus* GH61B polypeptide coding sequence, *T. reesei* Cel7A cellobiohydrolase I gene terminator, and *Aspergillus nidulans* acetamidase (amdS) gene.

Example 11: Construction of a Tandem Construct pDM287 for Expression of Both *Aspergillus fumigatus* CEL3A Beta-Glucosidase and *Aspergillus fumigatus* GH61B Polypeptide

An *A. fumigatus* GH61B polypeptide expression cassette was amplified from plasmid pSMai214 using the gene-specific forward and reverse primers shown below. The region in italics represents vector homology to the site of insertion for an IN-FUSION® reaction.

Forward primer: (SEQ ID NO: 85)
5' - *CGCGGTAGTGGCGCGGTTCGACCGAATGTAGGATTGTT* - 3'

Reverse primer: (SEQ ID NO: 86)
5' - *TTACCAATTGGCGCGCCACTACCGCGTTCGAGAAGA* - 3'

Fifty picomoles of each of the primers above were used in a PCR reaction composed of 25 ng of pSMai214 DNA, 1× PHUSION™ High-Fidelity Hot Start DNA Polymerase Buffer, 1 µl of a 10 mM blend of dATP, dTTP, dGTP, and dCTP, and 1 unit of PHUSION™ High-Fidelity Hot Start DNA Polymerase in a final volume of 50 µl. The amplification was performed using an EPPENDORF® MASTERCYCLER® 5333 egradient S programmed for 1 cycle at 98° C. for 30 seconds; 35 cycles each at 98° C. for 10 seconds, 60° C. for 30 seconds, and 72° C. for 1 minute 30 seconds; and 1 cycle at 72° C. for 10 minutes. PCR products were separated by 0.8% agarose gel electrophoresis using TAE buffer where an approximately 2.33 kb fragment was excised from the gel and extracted using a NUCLEOSPIN® Extract II Kit (Macherey-Nagel, Inc., Bethlehem, Pa., USA) according to the manufacturer's protocol.

The approximately 2.3 kb PCR product was inserted into Asc I-digested pEJG107 (WO 2005/047499) using an IN-FUSION® Advantage PCR Cloning Kit according to the manufacturer's protocol. Plasmid pEJG107 comprises an *Aspergillus fumigatus* CEL3A beta-glucosidase encoding sequence (SEQ ID NO: 5 [DNA sequence] and SEQ ID NO: 6 [deduced amino acid sequence]). The IN-FUSION® reaction was composed of 1× IN-FUSION® Reaction Buffer, 125 ng of the Asc I-digested pEJG107, 90 ng of the 2.33 kb PCR product, and 1 µl of IN-FUSION® Enzyme in a 10 µl reaction volume. The reaction was incubated for 15 minutes at 37° C. followed by 15 minutes at 50° C. After the incubation period 40 µl of TE were added to the reaction. A 2 µl aliquot was used to transform ONE SHOT® TOP10 competent cells according to the manufacturer's protocol. The *E. coli* transformation reactions were spread onto 2XYT plus ampicillin plates. The transformants were screened by sequencing and one clone containing the insert with no PCR errors was identified and designated pDM287 (FIG. 6). Plasmid pDM287 can be digested with Pme I to generate an approximately 9.9 kb fragment for *T. reesei* transformation. The 9.9 kb fragment contains two expression cassettes composed of (1) the *T. reesei* Cel7A cellobiohydrolase I gene promoter, *A. fumigatus* CEL3A beta-glucosidase coding sequence, and *T. reesei* Cel7A cellobiohydrolase I gene terminator; and (2) the *T. reesei* Cel7A cellobiohydrolase I gene promoter, *A. fumigatus* GH61B polypeptide coding sequence, and *T. reesei* Cel7A cellobiohydrolase I gene terminator. The 9.9 kb fragment also contains the *Aspergillus nidulans* acetamidase (amdS) gene.

Example 12: Expression of *Aspergillus fumigatus* Beta-Glucosidase and GH61B Polypeptide in Ku70+ *Aspergillus fumigatus* cbhl and Cbh2 Replacement Strain *T. reesei* 981-O-8.5#10B+Ku70#19L3

In order to express the *A. fumigatus* beta-glucosidase and *A. fumigatus* GH61B polypeptide, protoplasts of *Trichoderma reesei* strain 981-O-8.5#10B+Ku70#19L3 (described in Example 8) were generated as described in Example 2 and transformed with 2 µg of Pme I linearized pDM287. For transformation 100 µl of protoplasts were

transferred to a 14 ml polypropylene tube to which 2 µg of Pme I linearized, gel-purified pDM287 was added. Two hundred and fifty µl of PEG buffer were added and the tubes mixed gently by inverting 6 times. The tubes were incubated at 34° C. for 30 minutes after which 3 ml of STC were added. The contents of the tube were split and plated onto 2 separate 150 mm diameter COVE plates and incubated at 28° C. for 11 days. Transformants were picked using a sterile 1 µl inoculation loop and transferred to a fresh 75 mm diameter COVE2+10 mM uridine plate and incubated for 6 days at 28° C.

Transformants were each grown in shake flasks by inoculating 25 ml of CIM medium in a 125 ml polycarbonate non-baffled shake flask with spores collected using a 10 µl inoculation loop. The flasks were incubated at 28° C. with shaking at 200 rpm for 5 days. The shake flasks were harvested by pouring the entire culture into a 50 ml conical bottom tube and centrifuging the samples for 10 minutes at 2500 rpm in a SORVALL® Legend RT+ swing-bucket centrifuge (Thermo Fisher Scientific, Waltham, Mass., USA). Ten ml of each supernatant were transferred to a 15 ml conical bottom tube. Five µl of supernatant were combined with 5 µl of Laemelli sample buffer (Bio-Rad Laboratories, Hercules, Calif., USA) with 5% beta-mercaptoethanol (Sigma Aldrich, St. Louis, Mo., USA) in a 0.2 ml microcentrifuge tube and boiled for 5 minutes at 95° C. in an EPPENDORF® MASTERCYCLER® 5333 egradient S. Samples were analyzed by SDS-PAGE using a CRITERION® 8-16% Tris-HCl Gel (Bio-Rad Laboratories, Hercules, Calif., USA) according to the manufacturer's instructions using 10 µl of PRECISION PLUS™ All Blue Protein Standards (Bio-Rad Laboratories, Hercules, Calif., USA). Gels were stained and de-stained using BIO-SAFE™ Coomassie (Bio-Rad Laboratories, Hercules, Calif., USA). One strain (*T. reesei* JfyS-DM287-23) was selected based on high expression of the *A. fumigatus* beta-glucosidase and *A. fumigatus* GH61B polypeptide. Ten ml of *T. reesei* JfyS-DM287-23 shake flask broth was sterilized using a sterile 30 ml syringe and a MILLEX® GP 0.22 µm syringe filter (Millipore, Bedford, Mass., USA), transferred to a new 15 ml conical tube, and stored at -20° C. Protein concentration was determined using a Microplate BCA™ Protein Assay Kit (Thermo Fischer Scientific, Waltham, Mass., USA) in which bovine serum albumin was used as a protein standard.

Example 13: Pretreated Corn Stover Hydrolysis Assay

Corn stover was pretreated at the U.S. Department of Energy National Renewable Energy Laboratory (NREL) using 1.4 wt % sulfuric acid at 165° C. and 107 psi for 8 minutes. The water-insoluble solids in the pretreated corn stover (PCS) contained 56.5% cellulose, 4.6% hemicelluloses, and 28.4% lignin. Cellulose and hemicellulose were determined by a two-stage sulfuric acid hydrolysis with subsequent analysis of sugars by high performance liquid chromatography using NREL Standard Analytical Procedure #002. Lignin was determined gravimetrically after hydrolyzing the cellulose and hemicellulose fractions with sulfuric acid using NREL Standard Analytical Procedure #003.

Milled unwashed PCS (dry weight 32.35%) was prepared by milling whole slurry PCS in a Cosmos ICMG 40 wet multi-utility grinder (EssEmm Corporation, Tamil Nadu, India).

The hydrolysis of PCS was conducted using 2.2 ml deep-well plates (Axygen, Union City, Calif., USA) in a

total reaction volume of 1.0 ml. The hydrolysis was performed with 50 mg of insoluble PCS solids per ml of 50 mM sodium acetate pH 5.0 buffer containing 1 mM manganese sulfate and various protein loadings of various enzyme compositions (expressed as mg protein per gram of cellulose). Enzyme compositions were prepared and then added simultaneously to all wells in a volume ranging from 50 µl to 200 µl, for a final volume of 1 ml in each reaction. The plates were then sealed using an ALPS-300™ plate heat sealer (Abgene, Epsom, United Kingdom), mixed thoroughly, and incubated at a specific temperature for 72 hours. All experiments reported were performed in triplicate.

Following hydrolysis, samples were filtered using a 0.45 µm MULTISCREEN® 96-well filter plate (Millipore, Bedford, Mass., USA) and filtrates analyzed for sugar content as described below. When not used immediately, filtered aliquots were frozen at -20° C. The sugar concentrations of samples diluted in 0.005 M H₂SO₄ were measured using a 4.6×250 mm AMINEX® HPX-87H column (Bio-Rad Laboratories, Inc., Hercules, Calif., USA) by elution with 0.05% w/w benzoic acid-0.005 M H₂SO₄ at 65° C. at a flow rate of 0.6 ml per minute, and quantitation by integration of the glucose, cellobiose, and xylose signals from refractive index detection (CHEMSTATION®, AGILENT® 1100 HPLC, Agilent Technologies, Santa Clara, Calif., USA) calibrated by pure sugar samples. The resultant glucose and cellobiose equivalents were used to calculate the percentage of cellulose conversion for each reaction.

Glucose, cellobiose, and xylose were measured individually. Measured sugar concentrations were adjusted for the appropriate dilution factor. In case of unwashed PCS, the net concentrations of enzymatically-produced sugars were determined by adjusting the measured sugar concentrations for corresponding background sugar concentrations in unwashed PCS at zero time points. All HPLC data processing was performed using MICROSOFT EXCEL™ software (Microsoft, Richland, Wash., USA).

The degree of cellulose conversion to glucose was calculated using the following equation: % conversion=(glucose concentration/glucose concentration in a limit digest)×100. In order to calculate % conversion, a 100% conversion point was set based on a cellulase control (100 mg of *Trichoderma reesei* cellulase per gram cellulose), and all values were divided by this number and then multiplied by 100. Triplicate data points were averaged and standard deviation was calculated.

Example 14: Preparation of *Aspergillus oryzae* Beta-Glucosidase

Aspergillus oryzae beta-glucosidase (SEQ ID NO: 87 [DNA sequence] and SEQ ID NO: 88 [deduced amino acid sequence]) was prepared recombinantly according to WO 02/095014 using *Aspergillus oryzae* as a host.

The filtered broth of *Aspergillus oryzae* beta-glucosidase was concentrated and buffer exchanged using a tangential flow concentrator (Pall Filtron, Northborough, Mass., USA) equipped with a 10 kDa polyethersulfone membrane (Pall Filtron, Northborough, Mass., USA) with 20 mM Tris-HCl pH 8.0. The buffer exchanged sample was loaded onto a MonoQ® column (GE Healthcare, Piscataway, N.J., USA) equilibrated with 20 mM Tris-HCl pH 8.0, and the bound protein was eluted with a linear gradient from 0 to 1000 mM sodium chloride. Protein fractions were pooled and concentrated using 10 kDa MW-CO Amicon Ultra centrifuge concentrator (Millipore, Bedford, Mass., USA) into 20 mM Tris-HCl pH 8.0. Protein concentration was determined

using a Microplate BCA™ Protein Assay Kit with bovine serum albumin as a protein standard.

Example 15: Preparation of *Aspergillus fumigatus*
Enzyme Composition

Preparation of *Aspergillus fumigatus* Cel7A cellobiohydrolase I. The *Aspergillus fumigatus* Cel7A cellobiohydrolase I (SEQ ID NO: 1 [DNA sequence] and SEQ ID NO: 2 [deduced amino acid sequence]) was prepared recombinantly in *Aspergillus oryzae* as described in WO 2011/057140. The *Aspergillus fumigatus* Cel7A cellobiohydrolase I was purified according to WO 2011/057140.

Preparation of *Aspergillus fumigatus* cellobiohydrolase II. The *Aspergillus fumigatus* GH6A cellobiohydrolase II (SEQ ID NO: 3 [DNA sequence] and SEQ ID NO: 4 [deduced amino acid sequence]) was prepared recombinantly in *Aspergillus oryzae* as described in WO 2011/057140. The filtered broth of *Aspergillus fumigatus* GH6A cellobiohydrolase II was buffer exchanged into 20 mM Tris pH 8.0 using a 400 ml SEPHADEX™ G-25 column (GE Healthcare, United Kingdom) according to the manufacturer's instructions. Fractions were collected, pooled, and adjusted to 1.2 M ammonium sulphate-20 mM Tris pH 8.0. The equilibrated protein was loaded onto a PHENYL SEPHAROSE™ 6 Fast Flow column (high sub) (GE Healthcare, Piscataway, N.J., USA) equilibrated in 20 mM Tris pH 8.0 with 1.2 M ammonium sulphate, and bound proteins were eluted with 20 mM Tris pH 8.0 with no ammonium sulphate. The fractions were pooled.

Preparation of *Aspergillus fumigatus* strain GH5 endoglucanase II. The *Aspergillus fumigatus* GH5 endoglucanase II (SEQ ID NO: 11 [DNA sequence] and SEQ ID NO: 12 [deduced amino acid sequence]) was prepared recombinantly in *Aspergillus oryzae* as described in WO 2011/057140. The *Aspergillus fumigatus* GH5 endoglucanase II was purified according to WO 2011/057140.

Preparation of *Aspergillus fumigatus* GH61B polypeptide having cellulolytic enhancing activity. The *Aspergillus fumigatus* GH61B polypeptide having cellulolytic enhancing activity (SEQ ID NO: 7 [DNA sequence] and SEQ ID NO: 8 [deduced amino acid sequence]) was prepared recombinantly in *Aspergillus oryzae* as described in WO 2011/057140. The *Aspergillus fumigatus* GH61B polypeptide having cellulolytic enhancing activity was purified according to WO 2011/057140.

Preparation of *Aspergillus fumigatus* GH10 xylanase. The *Aspergillus fumigatus* GH10 xylanase (xyn3) (SEQ ID NO: 17 [DNA sequence] and SEQ ID NO: 18 [deduced amino acid sequence]) was prepared recombinantly according to WO 2006/078256 using *Aspergillus oryzae* BECh2 (WO 2000/39322) as a host. The filtered broth of *Aspergillus fumigatus* NN055679 GH10 xylanase (xyn3) was desalted and buffer-exchanged into 50 mM sodium acetate pH 5.0 using a HIPREP® 26/10 Desalting Column according to the manufacturer's instructions.

Preparation of *Aspergillus fumigatus* Cel3A beta-glucosidase. *Aspergillus fumigatus* Cel3A beta-glucosidase (SEQ ID NO: 5 [DNA sequence] and SEQ ID NO: 6 [deduced amino acid sequence]) was recombinantly prepared according to WO 2005/047499 using *Aspergillus oryzae* as a host. The filtered broth of *Aspergillus fumigatus* Cel3A beta-glucosidase was concentrated and buffer exchanged using a tangential flow concentrator equipped with a 10 kDa polyethersulfone membrane with 20 mM Tris-HCl pH 8.5. The sample was loaded onto a Q SEPHAROSE® High Performance column (GE Healthcare, Piscataway, N.J.,

USA) equilibrated in 20 mM Tris pH 8.5, and bound proteins were eluted with a linear gradient from 0-600 mM sodium chloride. The fractions were concentrated and loaded onto a SUPERDEX® 75 HR 26/60 column equilibrated with 20 mM Tris-150 mM sodium chloride pH 8.5. Alternatively, the filtered broth was adjusted to pH 8.0 with 20% sodium acetate, which made the solution turbid. To remove the turbidity, the solution was centrifuged at 20,000×g for 20 minutes, and the supernatant was filtered through a 0.2 µm filtration unit (Nalgene, Rochester, N.Y., USA). The filtrate was diluted with deionized water to reach the same conductivity as 50 mM Tris/HCl, pH 8.0. The adjusted enzyme solution was applied to a Q SEPHAROSE® Fast Flow column (GE Healthcare, Piscataway, N.J., USA) equilibrated in 50 mM Tris-HCl, pH 8.0 and eluted with a linear gradient from 0 to 500 mM sodium chloride. Fractions were pooled and treated with 1% (w/v) activated charcoal to remove color from the beta-glucosidase pool. The charcoal was removed by filtration of the suspension through a 0.2 µm filtration unit (Nalgene, Rochester, N.Y., USA). The filtrate was adjusted to pH 5.0 with 20% acetic acid and diluted 10 times with deionized water. The adjusted filtrate was applied to SP SEPHAROSE® Fast Flow column (GE Healthcare, Piscataway, N.J., USA) equilibrated in 10 mM succinic acid pH 5.0 and eluted with a linear gradient from 0 to 500 mM sodium chloride. Fractions were analyzed by SDS-PAGE. Fractions, where only one band was seen on a Coomassie stained SDS-PAGE gel, were pooled as the purified product and used for further experiments.

Preparation of *Aspergillus fumigatus* strain GH3 beta-xylosidase. The *Aspergillus fumigatus* GH3 beta-xylosidase (SEQ ID NO: 19 [DNA sequence] and SEQ ID NO: 20 [deduced amino acid sequence]) was prepared recombinantly in *Aspergillus oryzae* as described in WO 2011/057140. The *Aspergillus fumigatus* GH3 beta-xylosidase was purified according to WO 2011/057140.

The protein concentration for each of the monocomponents described above was determined using a Microplate BCA™ Protein Assay Kit in which bovine serum albumin was used as a protein standard. An *Aspergillus fumigatus* enzyme composition was composed of each monocomponent, prepared as described above, as follows: 37% *Aspergillus fumigatus* Cel7A cellobiohydrolase I, 25% *Aspergillus fumigatus* Cel6A cellobiohydrolase II, 10% *Aspergillus fumigatus* Cel5A endoglucanase II, 15% *Aspergillus fumigatus* GH61B polypeptide having cellulolytic enhancing activity, 5% *Aspergillus fumigatus* GH10 xylanase (xyn3), 5% *Aspergillus fumigatus* beta-glucosidase, and 3% *Aspergillus fumigatus* beta-xylosidase. The *Aspergillus fumigatus* enzyme composition is designated herein as "*Aspergillus fumigatus* enzyme composition (monocomponent mixture)".

Example 16: Expression of *Aspergillus fumigatus*
Wild-Type Composition

Aspergillus fumigatus strain NN051616 (EXT2007-00107) was inoculated onto a PDA plate and incubated for 4 days at 45° C. in the darkness. Several mycelia-PDA plugs were inoculated into 500 ml shake flasks containing 100 ml of NNCYP07-PCS medium. The flasks were incubated for 5 days at 45° C. with shaking at 160 rpm. The culture broth was filtered using a 0.45 µm DURAPORE® Membrane (Millipore, Bedford, Mass., USA).

A 3 ml volume of the filtrate was desalted and buffer exchanged into 50 mM sodium acetate pH 5.0 using an ECONO-PAC® 10-DG desalting column (Bio-Rad Laboratories, Inc., Hercules, Calif., USA) according to the manu-

facturer's instructions. Protein concentration was determined by SDS-PAGE densitometry using a CRITERION® 8-16% Tris-HCl Gel, staining with GELCODE™ Blue Stain Reagent (Pierce, Rockford, Ill., USA), and software analysis by ImageJ (National Institutes of Health, Bethesda, Md., USA).

The *Aspergillus fumigatus* enzyme composition is designated herein as "*Aspergillus fumigatus* wild-type enzyme composition".

Example 17: PCS Hydrolysis Assay Comparing the Enzyme Composition of the *T. reesei* Strain Expressing *Aspergillus fumigatus* Enzymes with a *Trichoderma reesei*-Based Cellulase Composition and an *Aspergillus fumigatus* Enzyme Composition

The enzyme composition of *T. reesei* JfyS-DM287-23 expressing *Aspergillus fumigatus* cellobiohydrolase I, *A. fumigatus* cellobiohydrolase II, *A. fumigatus* beta-glucosidase, and *A. fumigatus* GH61B polypeptide was evaluated relative to a *Trichoderma reesei*-based cellulase composition (CELLUCLAST™ 1.5L FG; Novozymes A/S, Bagsværd, Denmark replaced with 5% *Aspergillus oryzae* beta-glucosidase), and the *Aspergillus fumigatus* enzyme composition (monocomponent mixture) (Example 15) using milled unwashed PCS at 50° C. The results were compared with the results for the *Trichoderma reesei*-based cellulase composition and the *Aspergillus fumigatus* enzyme composition (monocomponent mixture). All compositions were used at 2.0, 4.0, and 6.0 mg protein per g cellulose.

The assay was performed as described in Example 13. The 1 ml reactions with 5% milled unwashed PCS were conducted for 72 hours in 50 mM sodium acetate pH 5.0 buffer containing 1 mM manganese sulfate. All reactions were performed in triplicate and involved single mixing at the beginning of hydrolysis.

The results shown in FIG. 7 demonstrated that the enzyme composition of *T. reesei* JfyS-DM-23 expressing *A. fumigatus* cellobiohydrolase I, *A. fumigatus* cellobiohydrolase II, *A. fumigatus* beta-glucosidase, and *A. fumigatus* GH61B polypeptide and the *Aspergillus fumigatus* enzyme composition (monocomponent mixture) produced significantly higher hydrolysis than the *Trichoderma reesei*-based cellulase composition at all three loadings. In addition, the enzyme composition of *T. reesei* JfyS-DM-23 expressing *A. fumigatus* cellobiohydrolase I, *A. fumigatus* cellobiohydrolase II, *A. fumigatus* beta-glucosidase, and *A. fumigatus* GH61B polypeptide produced higher hydrolysis than the *Aspergillus fumigatus* enzyme composition (monocomponent mixture) at all enzyme loadings.

Example 18: PCS Hydrolysis Assay Comparing an *Aspergillus fumigatus* Wild-Type Enzyme Composition with a *Trichoderma reesei*-Based Cellulase Composition

The *Aspergillus fumigatus* wild-type enzyme composition (Example 16) was compared to the *Trichoderma reesei*-based cellulase composition (CELLUCLAST™ 1.5L FG; Novozymes A/S, Bagsværd, Denmark replaced with 5% *Aspergillus oryzae* beta-glucosidase) using milled unwashed PCS at 50° C. In addition, the *Aspergillus fumigatus* wild-type enzyme composition was replaced with 5% *Aspergillus fumigatus* beta-glucosidase. The *Aspergillus fumigatus* wild-type enzyme composition and *Trichoderma reesei*-based cellulase composition were used at 2.0, 4.0, and 6.0 mg protein per g cellulose, and the *Aspergillus fumigatus* wild-

type enzyme composition replaced with 5% *Aspergillus fumigatus* beta-glucosidase was used at 2.0 and 4.0 mg protein per g cellulose.

The assay was performed as described in Example 13. The 1 ml reactions with 5% milled unwashed PCS were conducted for 72 hours in 50 mM sodium acetate pH 5.0 buffer containing 1 mM manganese sulfate. All reactions were performed in triplicate and involved single mixing at the beginning of hydrolysis.

The results shown in FIG. 8 demonstrated that the *Trichoderma reesei*-based cellulase composition had significantly higher hydrolysis than the *Aspergillus fumigatus* wild-type composition and the *Aspergillus fumigatus* wild-type composition replaced with 5% *Aspergillus fumigatus* beta-glucosidase. The replacement of the *Aspergillus fumigatus* wild-type composition with *Aspergillus fumigatus* beta-glucosidase significantly increased cellulose hydrolysis above that of the *Aspergillus fumigatus* wild-type composition alone but both mixtures produced significantly less hydrolysis than the *Trichoderma reesei*-based cellulase composition.

The present invention is further described by the following numbered paragraphs:

[1] A recombinant *Trichoderma* host cell, comprising polynucleotides encoding: (i) an *Aspergillus fumigatus* cellobiohydrolase I; (ii) an *Aspergillus fumigatus* cellobiohydrolase II; (iii) an *Aspergillus fumigatus* beta-glucosidase; and (iv) an *Aspergillus fumigatus* GH61 polypeptide having cellulolytic enhancing activity; or homologs thereof.

[2] The recombinant *Trichoderma* host cell of paragraph 1, wherein the *Aspergillus fumigatus* cellobiohydrolase I or homolog thereof is selected from the group consisting of: (i) a cellobiohydrolase I comprising or consisting of the mature polypeptide of SEQ ID NO: 2; (ii) a cellobiohydrolase I comprising or consisting of an amino acid sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide of SEQ ID NO: 2; (iii) a cellobiohydrolase I encoded by a polynucleotide comprising or consisting of a nucleotide sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide coding sequence of SEQ ID NO: 1; and (iv) a cellobiohydrolase I encoded by a polynucleotide that hybridizes under at least high stringency conditions, e.g., at least very high stringency conditions, with the mature polypeptide coding sequence of SEQ ID NO: 1 or the full-length complement thereof.

[3] The recombinant *Trichoderma* host cell of paragraph 1 or 2, wherein the *Aspergillus fumigatus* cellobiohydrolase II or homolog thereof is selected from the group consisting of: (i) a cellobiohydrolase II comprising or consisting of the mature polypeptide of SEQ ID NO: 4; (ii) a cellobiohydrolase II comprising or consisting of an amino acid sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide of SEQ ID NO: 4; (iii) a cellobiohydrolase II encoded by a

polynucleotide comprising or consisting of a nucleotide sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide coding sequence of SEQ ID NO: 3; and (iv) a cellobiohydrolase II encoded by a polynucleotide that hybridizes under at least high stringency conditions, e.g., at least very high stringency conditions, with the mature polypeptide coding sequence of SEQ ID NO: 3 or the full-length complement thereof.

[4] The recombinant *Trichoderma* host cell of any of paragraphs 1-3, wherein the *Aspergillus fumigatus* beta-glucosidase or homolog thereof is selected from the group consisting of: (i) a beta-glucosidase comprising or consisting of the mature polypeptide of SEQ ID NO: 6; (ii) a beta-glucosidase comprising or consisting of an amino acid sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide of SEQ ID NO: 6; (iii) a beta-glucosidase encoded by a polynucleotide comprising or consisting of a nucleotide sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide coding sequence of SEQ ID NO: 5; and (iv) a beta-glucosidase encoded by a polynucleotide that hybridizes under at least high stringency conditions, e.g., at least very high stringency conditions, with the mature polypeptide coding sequence of SEQ ID NO: 5 or the full-length complement thereof.

[5] The recombinant *Trichoderma* host cell of any of paragraphs 1-4, wherein the *Aspergillus fumigatus* GH61 polypeptide having cellulolytic enhancing activity or homolog thereof is selected from the group consisting of: (i) a GH61 polypeptide having cellulolytic enhancing activity comprising or consisting of the mature polypeptide of SEQ ID NO: 8; (ii) a GH61 polypeptide having cellulolytic enhancing activity comprising or consisting of an amino acid sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, sequence identity to the mature polypeptide of SEQ ID NO: 8; (iii) a GH61 polypeptide having cellulolytic enhancing activity encoded by a polynucleotide comprising or consisting of a nucleotide sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide coding sequence of SEQ ID NO: 7; and (iv) a GH61 polypeptide having cellulolytic enhancing activity encoded by a polynucleotide that hybridizes under at least high stringency conditions, e.g., at least very

high stringency conditions, with the mature polypeptide coding sequence of SEQ ID NO: 7 or the full-length complement thereof.

[6] The recombinant *Trichoderma* host cell of any of paragraphs 1-5, which further comprises one or more polynucleotides encoding one or more enzymes selected from the group consisting of: (i) an *Aspergillus fumigatus* endoglucanase I; (ii) an *Aspergillus fumigatus* endoglucanase II; (iii) an *Aspergillus fumigatus* xylanase; (iv) an *Aspergillus fumigatus* beta-xylosidase; and (v) an *Aspergillus fumigatus* swollenin.

[7] The recombinant *Trichoderma* host cell of paragraph 6, wherein the *Aspergillus fumigatus* endoglucanase I or homolog thereof is selected from the group consisting of: (i) an endoglucanase I comprising or consisting of the mature polypeptide of SEQ ID NO: 10; (ii) an endoglucanase I comprising or consisting of an amino acid sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide of SEQ ID NO: 10; (iii) an endoglucanase I encoded by a polynucleotide comprising or consisting of a nucleotide sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide coding sequence of SEQ ID NO: 9; and (iv) an endoglucanase I encoded by a polynucleotide that hybridizes under at least high stringency conditions, e.g., at least very high stringency conditions, with the mature polypeptide coding sequence of SEQ ID NO: 9 or the full-length complement thereof.

[8] The recombinant *Trichoderma* host cell of paragraph 6 or 7, wherein the *Aspergillus fumigatus* endoglucanase II or homolog thereof is selected from the group consisting of: (i) an *Aspergillus fumigatus* endoglucanase II comprising or consisting of the mature polypeptide of SEQ ID NO: 12; (ii) an endoglucanase II comprising or consisting of an amino acid sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide of SEQ ID NO: 12; (iii) an endoglucanase II encoded by a polynucleotide comprising or consisting of a nucleotide sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide coding sequence of SEQ ID NO: 11; and (iv) an endoglucanase II encoded by a polynucleotide that hybridizes under at least high stringency conditions, e.g., at least very high stringency conditions, with the mature polypeptide coding sequence of SEQ ID NO: 11 or the full-length complement thereof.

[9] The recombinant *Trichoderma* host cell of any of paragraphs 6-8, wherein the *Aspergillus fumigatus* xylanase or homolog thereof is selected from the group consisting of: (i) an *Aspergillus fumigatus* xylanase comprising or consisting of the mature polypeptide of SEQ ID NO: 14, SEQ ID

NO: 16, or SEQ ID NO: 18; (ii) a xylanase comprising or consisting of an amino acid sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide of SEQ ID NO: 14, SEQ ID NO: 16, or SEQ ID NO: 18; (iii) a xylanase encoded by a polynucleotide comprising or consisting of a nucleotide sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide coding sequence of SEQ ID NO: 13, SEQ ID NO: 15, or SEQ ID NO: 17; and (iv) a xylanase encoded by a polynucleotide that hybridizes under at least high stringency conditions, e.g., at least very high stringency conditions, with the mature polypeptide coding sequence of SEQ ID NO: 13, SEQ ID NO: 15, or SEQ ID NO: 17; or the full-length complement thereof.

[10] The recombinant *Trichoderma* host cell of any of paragraphs 6-9, wherein the *Aspergillus fumigatus* beta-xylosidase or homolog thereof is selected from the group consisting of: (i) a beta-xylosidase comprising or consisting of the mature polypeptide of SEQ ID NO: 20; (ii) a beta-xylosidase comprising or consisting of an amino acid sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide of SEQ ID NO: 20; (iii) a beta-xylosidase encoded by a polynucleotide comprising or consisting of a nucleotide sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide coding sequence of SEQ ID NO: 19; and (iv) a beta-xylosidase encoded by a polynucleotide that hybridizes under at least high stringency conditions, e.g., at least very high stringency conditions, with the mature polypeptide coding sequence of SEQ ID NO: 19 or the full-length complement thereof.

[11] The recombinant *Trichoderma* host cell of any of paragraphs 6-10, wherein the *Aspergillus fumigatus* swollenin or homolog thereof is selected from the group consisting of: (i) a swollenin comprising or consisting of the mature polypeptide of SEQ ID NO: 22; (ii) a swollenin comprising or consisting of an amino acid sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide of SEQ ID NO: 22; (iii) a swollenin encoded by a polynucleotide comprising or consisting of a nucleotide sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least

96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide coding sequence of SEQ ID NO: 21; and (iv) a swollenin encoded by a polynucleotide that hybridizes under at least high stringency conditions, e.g., at least very high stringency conditions, with the mature polypeptide coding sequence of SEQ ID NO: 21 or the full-length complement thereof.

[12] The recombinant *Trichoderma* host cell of any of paragraphs 1-11, which is selected from the group consisting of *Trichoderma harzianum*, *Trichoderma koningii*, *Trichoderma longibrachiatum*, *Trichoderma reesei*, and *Trichoderma viride*.

[13] The recombinant *Trichoderma* host cell of any of paragraphs 1-11, which is *Trichoderma reesei*.

[14] The recombinant *Trichoderma* host cell of any of paragraphs 1-13, wherein one or more of the cellulase genes, one or more of hemicellulase genes, or a combination thereof, endogenous to the *Trichoderma* host cell have been inactivated.

[15] The recombinant *Trichoderma* host cell of paragraph 14, wherein a *Trichoderma* cellobiohydrolase I gene or a homolog thereof has been inactivated.

[16] The recombinant *Trichoderma* host cell of paragraph 15, wherein the *Trichoderma* cellobiohydrolase I gene encodes the mature polypeptide of SEQ ID NO: 24 or a homolog thereof.

[17] The recombinant *Trichoderma* host cell of paragraph 16, wherein the *Trichoderma* cellobiohydrolase I gene homolog is selected from the group consisting of: (i) a cellobiohydrolase I comprising or consisting of an amino acid sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide of SEQ ID NO: 24; (ii) a cellobiohydrolase I encoded by a polynucleotide comprising or consisting of a nucleotide sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide coding sequence of SEQ ID NO: 23; and (iii) a cellobiohydrolase I encoded by a polynucleotide that hybridizes under at least high stringency conditions, e.g., at least very high stringency conditions, with the mature polypeptide coding sequence of SEQ ID NO: 23 or the full-length complement thereof.

[18] The recombinant *Trichoderma* host cell of any of paragraphs 14-17, wherein a *Trichoderma* cellobiohydrolase II gene or a homolog thereof has been inactivated.

[19] The recombinant *Trichoderma* host cell of paragraph 18, wherein the *Trichoderma* cellobiohydrolase II gene encodes the mature polypeptide of SEQ ID NO: 26 or a homolog thereof.

[20] The recombinant *Trichoderma* host cell of paragraph 19, wherein the *Trichoderma* cellobiohydrolase II gene homolog is selected from the group consisting of: (i) a cellobiohydrolase II comprising or consisting of an amino acid sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature

polypeptide of SEQ ID NO: 26; (ii) a cellobiohydrolase II encoded by a polynucleotide comprising or consisting of a nucleotide sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide coding sequence of SEQ ID NO: 25; and (iii) a cellobiohydrolase II encoded by a polynucleotide that hybridizes under at least high stringency conditions, e.g., at least very high stringency conditions, with the mature polypeptide coding sequence of SEQ ID NO: 25 or the full-length complement thereof.

[21] The recombinant *Trichoderma* host cell of any of paragraphs 14-20, wherein a *Trichoderma* beta-glucosidase gene or a homolog thereof has been inactivated.

[22] The recombinant *Trichoderma* host cell of paragraph 21, wherein the *Trichoderma* beta-glucosidase gene encodes the mature polypeptide of SEQ ID NO: 28 or a homolog thereof.

[23] The recombinant *Trichoderma* host cell of paragraph 22, wherein the *Trichoderma* beta-glucosidase gene homolog is selected from the group consisting of: (i) a beta-glucosidase comprising or consisting of an amino acid sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide of SEQ ID NO: 28; (ii) a beta-glucosidase encoded by a polynucleotide comprising or consisting of a nucleotide sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide coding sequence of SEQ ID NO: 27; and (iii) a beta-glucosidase encoded by a polynucleotide that hybridizes under at least high stringency conditions, e.g., at least very high stringency conditions, with the mature polypeptide coding sequence of SEQ ID NO: 27 or the full-length complement thereof.

[24] The recombinant *Trichoderma* host cell of any of paragraphs 14-23, wherein a *Trichoderma* endoglucanase I gene or a homolog thereof has been inactivated.

[25] The recombinant *Trichoderma* host cell of paragraph 24, wherein the *Trichoderma* endoglucanase I gene encodes the mature polypeptide of SEQ ID NO: 30 or a homolog thereof.

[26] The recombinant *Trichoderma* host cell of paragraph 25, wherein the *Trichoderma* endoglucanase I gene homolog is selected from the group consisting of: (i) an endoglucanase I comprising or consisting of an amino acid sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide of SEQ ID NO: 30; (ii) an endoglucanase I encoded by a polynucleotide comprising or consisting of a nucleotide sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least

89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide coding sequence of SEQ ID NO: 29; and (iii) an endoglucanase I encoded by a polynucleotide that hybridizes under at least high stringency conditions, e.g., at least very high stringency conditions, with the mature polypeptide coding sequence of SEQ ID NO: 29 or the full-length complement thereof.

[27] The recombinant *Trichoderma* host cell of any of paragraphs 14-26, wherein a *Trichoderma* endoglucanase II gene or a homolog thereof has been inactivated.

[28] The recombinant *Trichoderma* host cell of paragraph 27, wherein the *Trichoderma* endoglucanase II gene encodes the mature polypeptide of SEQ ID NO: 32 or a homolog thereof.

[29] The recombinant *Trichoderma* host cell of paragraph 28, wherein the *Trichoderma* endoglucanase II gene homolog is selected from the group consisting of: (i) an endoglucanase II comprising or consisting of an amino acid sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide of SEQ ID NO: 32; (ii) an endoglucanase II encoded by a polynucleotide comprising or consisting of a nucleotide sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide coding sequence of SEQ ID NO: 31; and (iii) an endoglucanase II encoded by a polynucleotide that hybridizes under at least high stringency conditions, e.g., at least very high stringency conditions, with the mature polypeptide coding sequence of SEQ ID NO: 31 or the full-length complement thereof.

[30] The recombinant *Trichoderma* host cell of any of paragraphs 14-29, wherein a *Trichoderma* xylanase I gene or a homolog thereof has been inactivated.

[31] The recombinant *Trichoderma* host cell of paragraph 30, wherein the *Trichoderma* xylanase I gene encodes the mature polypeptide of SEQ ID NO: 34 or a homolog thereof.

[32] The recombinant *Trichoderma* host cell of paragraph 31, wherein the *Trichoderma* beta xylanase I gene homolog is selected from the group consisting of: (i) a xylanase I comprising or consisting of an amino acid sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide of SEQ ID NO: 34; (ii) a xylanase I encoded by a polynucleotide comprising or consisting of a nucleotide sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide coding sequence of SEQ ID NO: 33; and (iii) a xylanase I encoded by a polynucleotide that hybridizes under at least high stringency conditions, e.g., at least very high stringency conditions, with the

mature polypeptide coding sequence of SEQ ID NO: 33 or the full-length complement thereof.

[33] The recombinant *Trichoderma* host cell of any of paragraphs 14-32, wherein a *Trichoderma* xylanase II gene or a homolog thereof has been inactivated.

[34] The recombinant *Trichoderma* host cell of paragraph 33, wherein the *Trichoderma* xylanase II gene encodes the mature polypeptide of SEQ ID NO: 36 or a homolog thereof.

[35] The recombinant *Trichoderma* host cell of paragraph 34, wherein the *Trichoderma* xylanase II gene homolog is selected from the group consisting of: (i) a xylanase II comprising or consisting of an amino acid sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide of SEQ ID NO: 36; (ii) a xylanase II encoded by a polynucleotide comprising or consisting of a nucleotide sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide coding sequence of SEQ ID NO: 35; and (iii) a xylanase II encoded by a polynucleotide that hybridizes under at least high stringency conditions, e.g., at least very high stringency conditions, with the mature polypeptide coding sequence of SEQ ID NO: 35 or the full-length complement thereof.

[36] The recombinant *Trichoderma* host cell of any of paragraphs 14-35, wherein a *Trichoderma* xylanase III gene or a homolog thereof has been inactivated.

[37] The recombinant *Trichoderma* host cell of paragraph 36, wherein the *Trichoderma* xylanase III gene encodes the mature polypeptide of SEQ ID NO: 38 or a homolog thereof.

[38] The recombinant *Trichoderma* host cell of paragraph 37, wherein the *Trichoderma* xylanase III gene homolog is selected from the group consisting of: (i) a xylanase III comprising or consisting of an amino acid sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide of SEQ ID NO: 38; (ii) a xylanase III encoded by a polynucleotide comprising or consisting of a nucleotide sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide coding sequence of SEQ ID NO: 37; and (iii) a xylanase III encoded by a polynucleotide that hybridizes under at least high stringency conditions, e.g., at least very high stringency conditions, with the mature polypeptide coding sequence of SEQ ID NO: 37 or the full-length complement thereof.

[39] The recombinant *Trichoderma* host cell of any of paragraphs 14-38, wherein a *Trichoderma* beta-xylosidase gene or a homolog thereof has been inactivated.

[40] The recombinant *Trichoderma* host cell of paragraph 39, wherein the *Trichoderma* beta-xylosidase gene encodes the mature polypeptide of SEQ ID NO: 40 or a homolog thereof.

[41] The recombinant *Trichoderma* host cell of paragraph 40, wherein the *Trichoderma* beta-xylosidase gene homolog is selected from the group consisting of: (i) a beta-xylosidase comprising or consisting of an amino acid sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide of SEQ ID NO: 40; (ii) a beta-xylosidase encoded by a polynucleotide comprising or consisting of a nucleotide sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide coding sequence of SEQ ID NO: 39; and (iii) a beta-xylosidase encoded by a polynucleotide that hybridizes under at least high stringency conditions, e.g., at least very high stringency conditions, with the mature polypeptide coding sequence of SEQ ID NO: 39 or the full-length complement thereof.

[42] The recombinant *Trichoderma* host cell of any of paragraphs 14-41, wherein a *Trichoderma* swollenin gene or a homolog thereof has been inactivated.

[43] The recombinant *Trichoderma* host cell of paragraph 42, wherein the *Trichoderma* swollenin gene encodes the mature polypeptide of SEQ ID NO: 42 or a homolog thereof.

[44] The recombinant *Trichoderma* host cell of paragraph 43, wherein the *Trichoderma* swollenin gene homolog is selected from the group consisting of: (i) a swollenin comprising or consisting of an amino acid sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide of SEQ ID NO: 42; (ii) a swollenin encoded by a polynucleotide comprising or consisting of a nucleotide sequence having at least 70%, e.g., at least 75%, at least 80%, at least 81%, at least 82%, at least 83%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% sequence identity to the mature polypeptide coding sequence of SEQ ID NO: 41; and (iii) a swollenin encoded by a polynucleotide that hybridizes under at least high stringency conditions, e.g., at least very high stringency conditions, with the mature polypeptide coding sequence of SEQ ID NO: 41 or the full-length complement thereof.

[45] A method of producing an enzyme composition, comprising: cultivating the host cell of any of paragraphs 1-44 under conditions conducive for production of the enzyme composition.

[46] The method of paragraph 45, further comprising recovering the enzyme composition.

[47] An enzyme composition comprising a recovered fermentation broth of the recombinant *Trichoderma* host cell of any of paragraphs 1-44.

[48] The enzyme composition of paragraph 47, which has one or more components of the fermentation broth removed.

[49] The enzyme composition of paragraph 47, which has no components of the fermentation broth removed.

[50] A process for degrading a cellulosic material, comprising: treating the cellulosic material with the enzyme composition of any of paragraphs 47-49.

[51] The process of paragraph 50, wherein the cellulosic material is pretreated.

[52] The process of paragraph 50 or 51, further comprising recovering the degraded cellulosic material.

[53] The process of paragraph 52, wherein the degraded cellulosic material is a sugar.

[54] The process of paragraph 53, wherein the sugar is selected from the group consisting of glucose, xylose, mannose, galactose, and arabinose.

[55] A process for producing a fermentation product, comprising: (a) saccharifying a cellulosic material with the enzyme composition of any of paragraphs 46-48; (b) fermenting the saccharified cellulosic material with one or more fermenting microorganisms to produce the fermentation product; and (c) recovering the fermentation product from the fermentation.

[56] The process of paragraph 55, wherein the cellulosic material is pretreated.

[57] The process of paragraph 55 or 56, wherein steps (a) and (b) are performed simultaneously in a simultaneous saccharification and fermentation.

[58] The process of any of paragraphs 55-57, wherein the fermentation product is an alcohol, an alkane, a cycloalkane, an alkene, an amino acid, a gas, isoprene, a ketone, an organic acid, or polyketide.

[59] A process of fermenting a cellulosic material, comprising: fermenting the cellulosic material with one or more fermenting microorganisms, wherein the cellulosic material is saccharified with the enzyme composition of any of paragraphs 47-49.

[60] The process of paragraph 59, wherein the fermenting of the cellulosic material produces a fermentation product.

[61] The process of paragraph 60, further comprising recovering the fermentation product from the fermentation.

[62] The process of paragraph 60 or 61, wherein the fermentation product is an alcohol, an alkane, a cycloalkane, an alkene, an amino acid, a gas, isoprene, a ketone, an organic acid, or polyketide.

[63] The process of any of paragraphs 59-62, wherein the cellulosic material is pretreated before saccharification.

[64] The enzyme composition of paragraphs 47-49, further comprising a *Trichoderma* endoglucanase I, a *Trichoderma* endoglucanase II, or a *Trichoderma* endoglucanase I and a *Trichoderma* endoglucanase II.

[65] The enzyme composition of paragraph 64, wherein the *Trichoderma* endoglucanase I is a *Trichoderma reesei* endoglucanase I.

[66] The enzyme composition of paragraph 64, wherein the *Trichoderma* endoglucanase II is a *Trichoderma reesei* endoglucanase II.

The invention described and claimed herein is not to be limited in scope by the specific aspects herein disclosed, since these aspects are intended as illustrations of several aspects of the invention. Any equivalent aspects are intended to be within the scope of this invention. Indeed, various modifications of the invention in addition to those shown and described herein will become apparent to those skilled in the art from the foregoing description. Such modifications are also intended to fall within the scope of the appended claims. In the case of conflict, the present disclosure including definitions will control.

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115         120         125
Pro Lys Ala Ser Ala Val Ala Glu Val Pro Ser Phe Val Trp Leu Asp
130         135         140

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Val	Ala	Ala	Lys	Val	Pro	Thr	Met	Gly	Thr	Tyr	Leu	Ala	Asp	Ile	Gln
145					150					155					160
Ala	Lys	Asn	Lys	Ala	Gly	Ala	Asn	Pro	Pro	Ile	Ala	Gly	Ile	Phe	Val
				165					170					175	
Val	Tyr	Asp	Leu	Pro	Asp	Arg	Asp	Cys	Ala	Ala	Leu	Ala	Ser	Asn	Gly
			180					185					190		
Glu	Tyr	Ser	Ile	Ala	Asn	Asn	Gly	Val	Ala	Asn	Tyr	Lys	Ala	Tyr	Ile
		195					200					205			
Asp	Ala	Ile	Arg	Ala	Gln	Leu	Val	Lys	Tyr	Ser	Asp	Val	His	Thr	Ile
	210					215					220				
Leu	Val	Ile	Glu	Pro	Asp	Ser	Leu	Ala	Asn	Leu	Val	Thr	Asn	Leu	Asn
225					230					235					240
Val	Ala	Lys	Cys	Ala	Asn	Ala	Gln	Ser	Ala	Tyr	Leu	Glu	Cys	Val	Asp
				245					250					255	
Tyr	Ala	Leu	Lys	Gln	Leu	Asn	Leu	Pro	Asn	Val	Ala	Met	Tyr	Leu	Asp
			260					265					270		
Ala	Gly	His	Ala	Gly	Trp	Leu	Gly	Trp	Pro	Ala	Asn	Leu	Gly	Pro	Ala
		275					280					285			
Ala	Thr	Leu	Phe	Ala	Lys	Val	Tyr	Thr	Asp	Ala	Gly	Ser	Pro	Ala	Ala
	290					295					300				
Val	Arg	Gly	Leu	Ala	Thr	Asn	Val	Ala	Asn	Tyr	Asn	Ala	Trp	Ser	Leu
305					310					315					320
Ser	Thr	Cys	Pro	Ser	Tyr	Thr	Gln	Gly	Asp	Pro	Asn	Cys	Asp	Glu	Lys
				325					330					335	
Lys	Tyr	Ile	Asn	Ala	Met	Ala	Pro	Leu	Leu	Lys	Glu	Ala	Gly	Phe	Asp
			340					345					350		
Ala	His	Phe	Ile	Met	Asp	Thr	Ser	Arg	Asn	Gly	Val	Gln	Pro	Thr	Lys
		355					360					365			
Gln	Asn	Ala	Trp	Gly	Asp	Trp	Cys	Asn	Val	Ile	Gly	Thr	Gly	Phe	Gly
370						375					380				
Val	Arg	Pro	Ser	Thr	Asn	Thr	Gly	Asp	Pro	Leu	Gln	Asp	Ala	Phe	Val
385					390					395					400
Trp	Ile	Lys	Pro	Gly	Gly	Glu	Ser	Asp	Gly	Thr	Ser	Asn	Ser	Thr	Ser
				405					410					415	
Pro	Arg	Tyr	Asp	Ala	His	Cys	Gly	Tyr	Ser	Asp	Ala	Leu	Gln	Pro	Ala
			420					425					430		
Pro	Glu	Ala	Gly	Thr	Trp	Phe	Gln	Ala	Tyr	Phe	Glu	Gln	Leu	Leu	Thr
		435					440					445			
Asn	Ala	Asn	Pro	Ser	Phe										
450															

<210> SEQ ID NO 5
 <211> LENGTH: 3060
 <212> TYPE: DNA
 <213> ORGANISM: Aspergillus fumigatus

<400> SEQUENCE: 5

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gtttgtgatg ctttcccgtc attgtttcgg atatagttga caatagtcac ggaaataatc	120
aggaattggc tttctctcca ccattctacc cttcgccttg ggctgatggc cagggagagt	180
gggcagatgc ccatcgacgc gccgtcgaga tcgtttctca gatgacactg gccgagaagg	240
ttaaccttac aacgggtact ggggtgggttg cgactttttt gttgacagtg agctttcttc	300
actgaccatc tacacagatg ggaaatggac cgatgcgtcg gtcaaaccgg cagcgttccc	360

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aggtaagctt	gcaattctgc	aacaacgtgc	aagtgtagtt	gctaaaacgc	ggtgggtgcag	420
acttgggtatc	aactgggggtc	tttgtggcca	ggattcccct	ttgggtatcc	gtttctgtga	480
gctatacccg	cggagtcttt	cagtccttgt	attatgtgct	gatgattgtc	tctgtatagc	540
tgacctcaac	tccgccttcc	ctgctgggtac	taatgtcgcc	gcgacatggg	acaagacact	600
cgctacctt	cgtggcaagg	ccatgggtga	ggaattcaac	gacaagggcg	tggacatttt	660
gctggggcct	gctgctggtc	ctctcggcaa	ataccgggac	ggcggcagaa	tctgggaagg	720
cttctctct	gatccgggtc	tactgggtgt	acttttcgcc	gaaactatca	agggtatcca	780
agacgcgggt	gtgattgcta	ctgccaagca	ttacattctg	aatgaacagg	agcatttccg	840
acaggttggc	gaggcccagg	gatatggtta	caacatcacg	gagacgatca	gctccaacgt	900
ggatgacaag	accatgcacg	agttgtacct	ttggtgagta	gttgacactg	caaatgagga	960
ccttgattga	tttgactgac	ctggaatgca	ggccctttgc	agatgctgtg	cgcggttaaga	1020
ttttccgtag	acttgacctc	gcgacgaaga	aatcgctgac	gaacctatcg	agctggcggt	1080
ggcgtgtca	tgtgttctca	caatcaaata	aacaacagct	acggttgtca	aaacagtcaa	1140
actctcaaca	agctcctcaa	ggctgagctg	ggcttccaag	gcttcgtcat	gagtgactgg	1200
agcgtcacc	acagcgggtg	cggcgtgcc	ctcgtgggt	tggatatgtc	gatgcctgga	1260
gacatttct	tcgacgacgg	actctccttc	tggggcacga	acctaactgt	cagtgttctt	1320
aacggcaccg	ttccagcctg	gcgtgtcgat	gacatggctg	ttcgtatcat	gaccgcgtac	1380
tacaaggttg	gtcgtgaccg	tcttcgtatt	ccccctaact	tcagctcctg	gaccgggat	1440
gagtacggct	gggagcattc	tgtgtctcc	gagggagcct	ggaccaaggt	gaacgacttc	1500
gtcaatgtgc	agcgcagtca	ctctcagatc	atccgtgaga	ttggtgccgc	tagtacagtg	1560
ctcttgaaga	acacgggtgc	tcttcctttg	accggcaagg	aggttaaagt	gggtgttctc	1620
ggtgaagacg	ctggttccaa	cccgtgggggt	gctaaccgct	gccccgaccg	cggtgtgat	1680
aacggcactc	ttgctatggc	ctggggtagt	ggtactgcca	acttcctta	ccttgtcacc	1740
cccgagcagg	ctatccagcg	agaggctatc	agcaacggcg	gcaatgtctt	tgtgtgact	1800
gataacgggg	ctctcagcca	gatggcagat	gttgcactc	aatccagggt	agtgcgggct	1860
cttagaaaaa	gaacgttctc	tgaatgaagt	tttttaacca	ttgcgaacag	cgtgtctttg	1920
gtgtttgtca	acgccgactc	tggagagggt	ttcatcagtg	tcgacggcaa	cgagggtgac	1980
cgaaaaaatc	tactctgtg	gaagaacggc	gaggccgtca	ttgacactgt	tgtcagccac	2040
tgcaacaaca	cgattgtgggt	tattcacagt	gttgggcccg	tcttgatcga	ccggtgggtat	2100
gataacocca	acgtcactgc	catcatctgg	gccgcttgc	ccggtcagga	gagtggcaac	2160
tccctgggtc	acgtgctcta	tggccgcgtc	aaccccagcg	ccaagacccc	gttcacctgg	2220
ggcaagactc	gggagtctta	cggggctccc	ttgctcaccg	agcctaacia	tggcaatgggt	2280
gctccccagg	atgatttcaa	cgagggcgtc	ttcattgact	accgtcactt	tgacaagcgc	2340
aatgagacc	ccatttatga	gtttggccat	ggcttgagct	acaccacctt	tggttactct	2400
caccttcggg	ttcaggccct	caatagttcg	agttcggcat	atgtcccgc	tagcggagag	2460
accaagcctg	cgccaacctc	tgggtgagatc	ggtagtccg	ccgactacct	gtatcccag	2520
ggtctcaaaa	gaattaccac	gtttatttac	ccttggctca	actcgaccga	cctcgaggat	2580
tcttctgacg	accegaacta	cggctgggag	gactcggagt	acattcccga	aggcgtagg	2640
gatgggtctc	ctcaaccctt	cctgaaggct	ggcggcgtc	ctgggtgtaa	ccctaccctt	2700

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tatcaggatc ttgtaggt gtcggccacc ataaccaaca ctggtaacgt cgccggttat 2760
gaagtcctc aattggtgag tgaccgcgcat gttccttgcg ttgcaatttg gctaactcgc 2820
ttctagtatg tttcactggg cggaccgaac gagcctcggg tcgttctcgc caagttcgac 2880
cgaatcttcc tggctcctgg ggagcaaaag gtttggaacca cgactcttaa ccgctcgtgat 2940
ctcgccaatt gggatgtgga ggctcaggac tgggtcatca caaagtaccc caagaaagtg 3000
cacgtcggca gctcctcgcg taagctgcct ctgagagcgc ctctgccccg tgtctactag 3060
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<210> SEQ ID NO 6
<211> LENGTH: 863
<212> TYPE: PRT
<213> ORGANISM: Aspergillus fumigatus
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<400> SEQUENCE: 6
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Met Arg Phe Gly Trp Leu Glu Val Ala Ala Leu Thr Ala Ala Ser Val
1          5          10          15
Ala Asn Ala Gln Glu Leu Ala Phe Ser Pro Pro Phe Tyr Pro Ser Pro
20          25          30
Trp Ala Asp Gly Gln Gly Glu Trp Ala Asp Ala His Arg Arg Ala Val
35          40          45
Glu Ile Val Ser Gln Met Thr Leu Ala Glu Lys Val Asn Leu Thr Thr
50          55          60
Gly Thr Gly Trp Glu Met Asp Arg Cys Val Gly Gln Thr Gly Ser Val
65          70          75          80
Pro Arg Leu Gly Ile Asn Trp Gly Leu Cys Gly Gln Asp Ser Pro Leu
85          90          95
Gly Ile Arg Phe Ser Asp Leu Asn Ser Ala Phe Pro Ala Gly Thr Asn
100         105         110
Val Ala Ala Thr Trp Asp Lys Thr Leu Ala Tyr Leu Arg Gly Lys Ala
115         120         125
Met Gly Glu Glu Phe Asn Asp Lys Gly Val Asp Ile Leu Leu Gly Pro
130         135         140
Ala Ala Gly Pro Leu Gly Lys Tyr Pro Asp Gly Gly Arg Ile Trp Glu
145         150         155         160
Gly Phe Ser Pro Asp Pro Val Leu Thr Gly Val Leu Phe Ala Glu Thr
165         170         175
Ile Lys Gly Ile Gln Asp Ala Gly Val Ile Ala Thr Ala Lys His Tyr
180         185         190
Ile Leu Asn Glu Gln Glu His Phe Arg Gln Val Gly Glu Ala Gln Gly
195         200         205
Tyr Gly Tyr Asn Ile Thr Glu Thr Ile Ser Ser Asn Val Asp Asp Lys
210         215         220
Thr Met His Glu Leu Tyr Leu Trp Pro Phe Ala Asp Ala Val Arg Ala
225         230         235         240
Gly Val Gly Ala Val Met Cys Ser Tyr Asn Gln Ile Asn Asn Ser Tyr
245         250         255
Gly Cys Gln Asn Ser Gln Thr Leu Asn Lys Leu Leu Lys Ala Glu Leu
260         265         270
Gly Phe Gln Gly Phe Val Met Ser Asp Trp Ser Ala His His Ser Gly
275         280         285
Val Gly Ala Ala Leu Ala Gly Leu Asp Met Ser Met Pro Gly Asp Ile
290         295         300
Ser Phe Asp Asp Gly Leu Ser Phe Trp Gly Thr Asn Leu Thr Val Ser
305         310         315         320
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Val Leu Asn Gly Thr Val Pro Ala Trp Arg Val Asp Asp Met Ala Val
 325 330 335
 Arg Ile Met Thr Ala Tyr Tyr Lys Val Gly Arg Asp Arg Leu Arg Ile
 340 345 350
 Pro Pro Asn Phe Ser Ser Trp Thr Arg Asp Glu Tyr Gly Trp Glu His
 355 360 365
 Ser Ala Val Ser Glu Gly Ala Trp Thr Lys Val Asn Asp Phe Val Asn
 370 375 380
 Val Gln Arg Ser His Ser Gln Ile Ile Arg Glu Ile Gly Ala Ala Ser
 385 390 395 400
 Thr Val Leu Leu Lys Asn Thr Gly Ala Leu Pro Leu Thr Gly Lys Glu
 405 410 415
 Val Lys Val Gly Val Leu Gly Glu Asp Ala Gly Ser Asn Pro Trp Gly
 420 425 430
 Ala Asn Gly Cys Pro Asp Arg Gly Cys Asp Asn Gly Thr Leu Ala Met
 435 440 445
 Ala Trp Gly Ser Gly Thr Ala Asn Phe Pro Tyr Leu Val Thr Pro Glu
 450 455 460
 Gln Ala Ile Gln Arg Glu Val Ile Ser Asn Gly Gly Asn Val Phe Ala
 465 470 475 480
 Val Thr Asp Asn Gly Ala Leu Ser Gln Met Ala Asp Val Ala Ser Gln
 485 490 495
 Ser Ser Val Ser Leu Val Phe Val Asn Ala Asp Ser Gly Glu Gly Phe
 500 505 510
 Ile Ser Val Asp Gly Asn Glu Gly Asp Arg Lys Asn Leu Thr Leu Trp
 515 520 525
 Lys Asn Gly Glu Ala Val Ile Asp Thr Val Val Ser His Cys Asn Asn
 530 535 540
 Thr Ile Val Val Ile His Ser Val Gly Pro Val Leu Ile Asp Arg Trp
 545 550 555 560
 Tyr Asp Asn Pro Asn Val Thr Ala Ile Ile Trp Ala Gly Leu Pro Gly
 565 570 575
 Gln Glu Ser Gly Asn Ser Leu Val Asp Val Leu Tyr Gly Arg Val Asn
 580 585 590
 Pro Ser Ala Lys Thr Pro Phe Thr Trp Gly Lys Thr Arg Glu Ser Tyr
 595 600 605
 Gly Ala Pro Leu Leu Thr Glu Pro Asn Asn Gly Asn Gly Ala Pro Gln
 610 615 620
 Asp Asp Phe Asn Glu Gly Val Phe Ile Asp Tyr Arg His Phe Asp Lys
 625 630 635 640
 Arg Asn Glu Thr Pro Ile Tyr Glu Phe Gly His Gly Leu Ser Tyr Thr
 645 650 655
 Thr Phe Gly Tyr Ser His Leu Arg Val Gln Ala Leu Asn Ser Ser Ser
 660 665 670
 Ser Ala Tyr Val Pro Thr Ser Gly Glu Thr Lys Pro Ala Pro Thr Tyr
 675 680 685
 Gly Glu Ile Gly Ser Ala Ala Asp Tyr Leu Tyr Pro Glu Gly Leu Lys
 690 695 700
 Arg Ile Thr Lys Phe Ile Tyr Pro Trp Leu Asn Ser Thr Asp Leu Glu
 705 710 715 720
 Asp Ser Ser Asp Asp Pro Asn Tyr Gly Trp Glu Asp Ser Glu Tyr Ile
 725 730 735

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Pro Glu Gly Ala Arg Asp Gly Ser Pro Gln Pro Leu Leu Lys Ala Gly
740 745 750

Gly Ala Pro Gly Gly Asn Pro Thr Leu Tyr Gln Asp Leu Val Arg Val
755 760 765

Ser Ala Thr Ile Thr Asn Thr Gly Asn Val Ala Gly Tyr Glu Val Pro
770 775 780

Gln Leu Tyr Val Ser Leu Gly Gly Pro Asn Glu Pro Arg Val Val Leu
785 790 795 800

Arg Lys Phe Asp Arg Ile Phe Leu Ala Pro Gly Glu Gln Lys Val Trp
805 810 815

Thr Thr Thr Leu Asn Arg Arg Asp Leu Ala Asn Trp Asp Val Glu Ala
820 825 830

Gln Asp Trp Val Ile Thr Lys Tyr Pro Lys Lys Val His Val Gly Ser
835 840 845

Ser Ser Arg Lys Leu Pro Leu Arg Ala Pro Leu Pro Arg Val Tyr
850 855 860

<210> SEQ ID NO 7
<211> LENGTH: 862
<212> TYPE: DNA
<213> ORGANISM: Aspergillus fumigatus

<400> SEQUENCE: 7

atgactttgt ccaagatcac ttccattgct ggccttctgg cctcagcgtc tctcgtggct 60
ggccacggct ttgtttctgg cattgttgct gatgggaaat agtatgtgct tgaaccacac 120
aatgacagc tgcaacagct aacttctatt ccagttacgg agggtagctt gttaaccaat 180
accctacat gagcaaccct cccgacacca ttgcctggtc caccaccgc accgacctcg 240
gctttgtgga cggcaccggc taccagtctc cggatattat ctgccacaga gacgcaaaga 300
atggcaagtt gaccgcaacc gttgcagccg gttcacagat cgaattccag tggacgacgt 360
ggccagagtc tcaccatgga ccggtacgac gccgaagaga agagaacata ttgtgaccag 420
ataggctaac atagcatagt tgattactta cctcgcctca tgcaacggcg actgtgccac 480
cgtggacaag accaccctga agtttgtcaa gatcgccgct caaggcttga tgcagggctc 540
caaccacct ggtgtttggg ctgatgatga aatgatcgcc aacaacaaca cggccacagt 600
gaccattcct gcctcctatg cccccggaaa ctacgtcctt cgccacgaga tcatcgcctt 660
tactctgcg ggtaacctga acggcgcgca gaactacccc cagtgtttca acatccaaat 720
caccgggtggc ggcagtgtc agggatctgg caccgtggc acgtccctgt acaagaatac 780
tgatcctggc atcaagtttg acatctactc ggatctgagc ggtggatacc ctattcctgg 840
tcctgcactg ttcaacgctt aa 862

<210> SEQ ID NO 8
<211> LENGTH: 250
<212> TYPE: PRT
<213> ORGANISM: Aspergillus fumigatus

<400> SEQUENCE: 8

Met Thr Leu Ser Lys Ile Thr Ser Ile Ala Gly Leu Leu Ala Ser Ala
1 5 10 15

Ser Leu Val Ala Gly His Gly Phe Val Ser Gly Ile Val Ala Asp Gly
20 25 30

Lys Tyr Tyr Gly Gly Tyr Leu Val Asn Gln Tyr Pro Tyr Met Ser Asn
35 40 45

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Pro	Pro	Asp	Thr	Ile	Ala	Trp	Ser	Thr	Thr	Ala	Thr	Asp	Leu	Gly	Phe
	50					55					60				
Val	Asp	Gly	Thr	Gly	Tyr	Gln	Ser	Pro	Asp	Ile	Ile	Cys	His	Arg	Asp
65					70					75					80
Ala	Lys	Asn	Gly	Lys	Leu	Thr	Ala	Thr	Val	Ala	Ala	Gly	Ser	Gln	Ile
				85					90					95	
Glu	Phe	Gln	Trp	Thr	Thr	Trp	Pro	Glu	Ser	His	His	Gly	Pro	Leu	Ile
			100					105					110		
Thr	Tyr	Leu	Ala	Pro	Cys	Asn	Gly	Asp	Cys	Ala	Thr	Val	Asp	Lys	Thr
		115					120					125			
Thr	Leu	Lys	Phe	Val	Lys	Ile	Ala	Ala	Gln	Gly	Leu	Ile	Asp	Gly	Ser
	130					135					140				
Asn	Pro	Pro	Gly	Val	Trp	Ala	Asp	Asp	Glu	Met	Ile	Ala	Asn	Asn	Asn
145					150					155					160
Thr	Ala	Thr	Val	Thr	Ile	Pro	Ala	Ser	Tyr	Ala	Pro	Gly	Asn	Tyr	Val
				165					170					175	
Leu	Arg	His	Glu	Ile	Ile	Ala	Leu	His	Ser	Ala	Gly	Asn	Leu	Asn	Gly
			180					185					190		
Ala	Gln	Asn	Tyr	Pro	Gln	Cys	Phe	Asn	Ile	Gln	Ile	Thr	Gly	Gly	Gly
		195					200						205		
Ser	Ala	Gln	Gly	Ser	Gly	Thr	Ala	Gly	Thr	Ser	Leu	Tyr	Lys	Asn	Thr
	210					215					220				
Asp	Pro	Gly	Ile	Lys	Phe	Asp	Ile	Tyr	Ser	Asp	Leu	Ser	Gly	Gly	Tyr
225					230					235					240
Pro	Ile	Pro	Gly	Pro	Ala	Leu	Phe	Asn	Ala						
				245					250						

<210> SEQ ID NO 9

<211> LENGTH: 1254

<212> TYPE: DNA

<213> ORGANISM: Aspergillus fumigatus

<400> SEQUENCE: 9

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actggattta ccatggetca aacactggca gccgcctcgc tggttctcgt ccccttgtg      60
actgocgagc agatcgggtc catcgctgag aaccaccocg agctcaaaac atacaggtgc      120
ggttctcagg ctggctgctg tgcacaaagc acctcagtgg ttcttgacat caacgcacac      180
tggatacacc aaatgggagc ccaaactgca tgcaccacga gtagtggcct cgaccctct      240
ctctgccccg acaaggtgac ctgcagccag aactgcgtag tggaaggcat caccgactac      300
agtagcttcg gcggtgcagaa ctcaggcgac gccataaacc tccgccaata ccaagtacaa      360
aacggccaga tcaagacact gaggccgcgc gtgtacctcc tcgcccagga tggcatcaac      420
tacagcaagc tgcagctcct caaccaagag ttcaccttcg acgttgacgc ctccaagctc      480
ccctgcggca tgaacggcgc cctctacctc tccgaaatgg acgcctcagg cggccgcagc      540
gccctcaacc ccgctggtgc aacctacggc acaggctact gcgacgcgca gtgcttcaac      600
cccggtcctt ggatcaacgg cgaggccaac accctcggcg cagggtgcctg ctgccaggag      660
atggacctct gggaggccaa ctcgcgctcc acaattttct ccccgaccc gtgcaccaca      720
gccggcctgt acgcgtgcac cggcgccgaa tgctactcca tctgcgacgg gtatggctgc      780
acctacaacc cctacgagct cggcgcaaag gattactatg gctacggtct caccggtgac      840
accgccaagc cgataaccgt cgtcatgcag ttcgtgacgg ccgataacac cgcgacagga      900
actctggccg agatccgcag gctgtacgtg caggagggtg tggatgatcg caattcggcc      960

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gtcgccatga cggaggcttt ctgctcgtcg tcgaggacgt tcgaggcgct gggtagggtg 1020
cagcgtatgg gcgaggctct ggggaggggc atggtgacctg tgttcagtat ctgggatgat 1080
ccgagcctgt ggatgcattg gcttgatagt gacggtgccg ggccgtgccc tagcactgag 1140
ggagatcctg ctttcatcca ggctaactat cccaatacgg cggtgacttt ttcgaaggtc 1200
aggtgggggg atattgacag tacctactct gtttagttaa ttaactagag gtga 1254

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<210> SEQ ID NO 10
<211> LENGTH: 407
<212> TYPE: PRT
<213> ORGANISM: Aspergillus fumigatus

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<400> SEQUENCE: 10

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Met Ala Gln Thr Leu Ala Ala Ala Ser Leu Val Leu Val Pro Leu Val
1          5          10          15
Thr Ala Gln Gln Ile Gly Ser Ile Ala Glu Asn His Pro Glu Leu Lys
20          25          30
Thr Tyr Arg Cys Gly Ser Gln Ala Gly Cys Val Ala Gln Ser Thr Ser
35          40          45
Val Val Leu Asp Ile Asn Ala His Trp Ile His Gln Met Gly Ala Gln
50          55          60
Thr Ser Cys Thr Thr Ser Ser Gly Leu Asp Pro Ser Leu Cys Pro Asp
65          70          75          80
Lys Val Thr Cys Ser Gln Asn Cys Val Val Glu Gly Ile Thr Asp Tyr
85          90          95
Ser Ser Phe Gly Val Gln Asn Ser Gly Asp Ala Ile Thr Leu Arg Gln
100         105         110
Tyr Gln Val Gln Asn Gly Gln Ile Lys Thr Leu Arg Pro Arg Val Tyr
115        120        125
Leu Leu Ala Glu Asp Gly Ile Asn Tyr Ser Lys Leu Gln Leu Leu Asn
130        135        140
Gln Glu Phe Thr Phe Asp Val Asp Ala Ser Lys Leu Pro Cys Gly Met
145        150        155        160
Asn Gly Ala Leu Tyr Leu Ser Glu Met Asp Ala Ser Gly Gly Arg Ser
165        170        175
Ala Leu Asn Pro Ala Gly Ala Thr Tyr Gly Thr Gly Tyr Cys Asp Ala
180        185        190
Gln Cys Phe Asn Pro Gly Pro Trp Ile Asn Gly Glu Ala Asn Thr Leu
195        200        205
Gly Ala Gly Ala Cys Cys Gln Glu Met Asp Leu Trp Glu Ala Asn Ser
210        215        220
Arg Ser Thr Ile Phe Ser Pro His Pro Cys Thr Thr Ala Gly Leu Tyr
225        230        235        240
Ala Cys Thr Gly Ala Glu Cys Tyr Ser Ile Cys Asp Gly Tyr Gly Cys
245        250        255
Thr Tyr Asn Pro Tyr Glu Leu Gly Ala Lys Asp Tyr Tyr Gly Tyr Gly
260        265        270
Leu Thr Val Asp Thr Ala Lys Pro Ile Thr Val Val Met Gln Phe Val
275        280        285
Thr Ala Asp Asn Thr Ala Thr Gly Thr Leu Ala Glu Ile Arg Arg Leu
290        295        300
Tyr Val Gln Glu Gly Met Val Ile Gly Asn Ser Ala Val Ala Met Thr
305        310        315        320
Glu Ala Phe Cys Ser Ser Ser Arg Thr Phe Glu Ala Leu Gly Gly Leu

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	325		330		335										
Gln	Arg	Met	Gly	Glu	Ala	Leu	Gly	Arg	Gly	Met	Val	Pro	Val	Phe	Ser
			340					345					350		
Ile	Trp	Asp	Asp	Pro	Ser	Leu	Trp	Met	His	Trp	Leu	Asp	Ser	Asp	Gly
		355					360				365				
Ala	Gly	Pro	Cys	Gly	Ser	Thr	Glu	Gly	Asp	Pro	Ala	Phe	Ile	Gln	Ala
		370				375					380				
Asn	Tyr	Pro	Asn	Thr	Ala	Val	Thr	Phe	Ser	Lys	Val	Arg	Trp	Gly	Asp
385					390					395					400
Ile	Asp	Ser	Thr	Tyr	Ser	Val									
					405										

<210> SEQ ID NO 11
 <211> LENGTH: 1263
 <212> TYPE: DNA
 <213> ORGANISM: Aspergillus fumigatus

<400> SEQUENCE: 11

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actggattta ccatgaaatt cggtagcatt gtgctcattg ctgctgcggc aggettcgcg      60
gtggctgctc ctgcaaagag agcttcggta tttcaatggt ggttcctgt ggtaaagtgt      120
gattaaagg aactaacat actgcagggt tcggaagcaa tgagtctgga gcagagtttg      180
gcgaaaatac cattcctggc tcttatgtat gttgtgcatac tgagagaagt atactgctgc      240
tgacaacatc aaggggaaag aattcatctt cccggaccct tctacaatca gcacattgat      300
cgggaagggc atgaacatct tccggattca attcctcatg gagagactgg tgccaagctc      360
tatgacaggc tcctataacg aggagtacct tgccaatctg acatcggttg gtttgagcag      420
cagcatgttg gactgtatga ggctgactcg accaggttgt ggacgctgtc accaagggcag      480
gatcttatgc tattttggac ccacacaact ttggcagata gtgagtaatg cccggcatac      540
tgtggacttg ttctaagcc actcagcaat ggtcagatta tctccagcac cgacgacttc      600
aagaccttct ggcagaatct ggctggaaag ttcaagtcca acaatctcgt catctttgat      660
actagtatgg ctaactcatt tggttctgat gagcttcaact gaccccgat gttagacaat      720
gagtatcacg acatggacca gacactggta ctgaacctca accaggccgc tatcaacggg      780
atccgcgctg caggagccac ctgcgaatac atctttgtgg agggcaactc ctggaccggc      840
gcctggacct gggccgacgt caatgacaac ctgaaggctc tgaccgacc ccaggataag      900
atcgtctacg agatgcacca gtatctcgac tcggatggat ccggcaccgc ggagagctgc      960
gtgtctacca cgattggtaa ggagcggggt tcggccgcaa caaagtggct caaggataac     1020
ggcaagggtg gcatcattgg tgagtctgct ggtggcgtca atgatcagt cccgaccgct     1080
atctcaggaa tgctggagta cttggctcag aacacagacg tgtggaagg agctctctgg     1140
tgggcggctg gccctggtg gggaaactat atgttcaaca tggagcctcc gagcgggtgca     1200
gcttatgtgg gcatgttga catcttggag cctacctgg gttgattaat taactagagg     1260
tga                                                                 1263
    
```

<210> SEQ ID NO 12
 <211> LENGTH: 329
 <212> TYPE: PRT
 <213> ORGANISM: Aspergillus fumigatus

<400> SEQUENCE: 12

Met	Lys	Phe	Gly	Ser	Ile	Val	Leu	Ile	Ala	Ala	Ala	Ala	Gly	Phe	Ala
1				5					10					15	

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Val Ala Ala Pro Ala Lys Arg Ala Ser Val Phe Gln Trp Phe Gly Ser
 20 25 30

Asn Glu Ser Gly Ala Glu Phe Gly Glu Asn Thr Ile Pro Gly Ser Tyr
 35 40 45

Gly Lys Glu Phe Ile Phe Pro Asp Pro Ser Thr Ile Ser Thr Leu Ile
 50 55 60

Gly Lys Gly Met Asn Ile Phe Arg Ile Gln Phe Leu Met Glu Arg Leu
 65 70 75 80

Val Pro Ser Ser Met Thr Gly Ser Tyr Asn Glu Glu Tyr Leu Ala Asn
 85 90 95

Leu Thr Ser Val Val Asp Ala Val Thr Lys Ala Gly Ser Tyr Ala Ile
 100 105 110

Leu Asp Pro His Asn Phe Gly Arg Tyr Asn Gly Gln Ile Ile Ser Ser
 115 120 125

Thr Asp Asp Phe Lys Thr Phe Trp Gln Asn Leu Ala Gly Lys Phe Lys
 130 135 140

Ser Asn Asn Leu Val Ile Phe Asp Thr Asn Asn Glu Tyr His Asp Met
 145 150 155 160

Asp Gln Thr Leu Val Leu Asn Leu Asn Gln Ala Ala Ile Asn Gly Ile
 165 170 175

Arg Ala Ala Gly Ala Thr Ser Gln Tyr Ile Phe Val Glu Gly Asn Ser
 180 185 190

Trp Thr Gly Ala Trp Thr Trp Ala Asp Val Asn Asp Asn Leu Lys Ala
 195 200 205

Leu Thr Asp Pro Gln Asp Lys Ile Val Tyr Glu Met His Gln Tyr Leu
 210 215 220

Asp Ser Asp Gly Ser Gly Thr Ala Glu Ser Cys Val Ser Thr Thr Ile
 225 230 235 240

Gly Lys Glu Arg Val Ser Ala Ala Thr Lys Trp Leu Lys Asp Asn Gly
 245 250 255

Lys Val Gly Ile Ile Gly Glu Phe Ala Gly Gly Val Asn Asp Gln Cys
 260 265 270

Arg Thr Ala Ile Ser Gly Met Leu Glu Tyr Leu Ala Gln Asn Thr Asp
 275 280 285

Val Trp Lys Gly Ala Leu Trp Trp Ala Ala Gly Pro Trp Trp Gly Asn
 290 295 300

Tyr Met Phe Asn Met Glu Pro Pro Ser Gly Ala Ala Tyr Val Gly Met
 305 310 315 320

Leu Asp Ile Leu Glu Pro Tyr Leu Gly
 325

<210> SEQ ID NO 13
 <211> LENGTH: 1145
 <212> TYPE: DNA
 <213> ORGANISM: Aspergillus fumigatus
 <400> SEQUENCE: 13

atgcgtttct cccttgccgc caccgctctt ctcgctggcc tggccacggc agcgccttcg 60
 agcaacaaga acaacgtcaa tcttgataag cttgctcggc gtaatggcat gctttggttc 120
 ggcaactgcag ccgatatccc tggtaacctca gaaacaaccg acaagcctta tctgagcatc 180
 ctgcgcaagc agttcggcga aatgacaccc gcaaacgcat tgaaggtgag ccagagtgat 240
 agtacacctc atctcgtgtc ggcgctgacc agacgatggt attcacaatag ttcattgtata 300

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ccgagcccga gcagaatgtc ttcaacttca ctcaagggga ctacttcatg gacttggccg 360
atcactatgg tcacgccgtg cgctgccata acctcgctcg ggccagccaa gtgtccgact 420
gggtcacctc caggaactgg accgccacag aactcaaaga agtgatgaag aaccacatat 480
tcaagaccgt ccaacatttt ggcaagcgct gctacgcgtg ggacgtcgtc aatgaagcta 540
ttaatgggga cgggaccttt tctccagtg tgtggtacga cacaattggc gaggaatact 600
tctaccttgc attccagtat gccaggaag ccttggcgca gattcacgcc aaccagggtca 660
agctttacta taacgactat ggcaattgaga accccggccc caaggcagat gctgttctga 720
agctagtcgc cgagttgagg aagcggggca ttcgcattga cggagtcggt ctcgagtccc 780
acttcatcgt cggcgagact ccttcgctgg ctgaccagct cgccaccaag aaggcttata 840
tcgaggccgg acttgaggtc gccatcaccg aacttgacgt ccgcttttct caggccccgt 900
tctacaccgc cgaggcccaa aagcagcagg ctgccgacta ctatgctagc gtcgccagtt 960
gcaagcatgc cggaccgcgc tgtgttggtg ttgtagtctg ggatttcgat gacgcctact 1020
cgtggattcc gggtagcttc gagggacagg gtggcgctcg tctatataat gagacactcg 1080
aggtgaagcc ggccttctat gctgctgccg aggcgttggg gaacaagccc tgcactgtat 1140
gctag 1145

```

<210> SEQ ID NO 14

<211> LENGTH: 364

<212> TYPE: PRT

<213> ORGANISM: Aspergillus fumigatus

<400> SEQUENCE: 14

```

Met Arg Phe Ser Leu Ala Ala Thr Ala Leu Leu Ala Gly Leu Ala Thr
1           5           10          15
Ala Ala Pro Ser Ser Asn Lys Asn Asn Val Asn Leu Asp Lys Leu Ala
20          25          30
Arg Arg Asn Gly Met Leu Trp Phe Gly Thr Ala Ala Asp Ile Pro Gly
35          40          45
Thr Ser Glu Thr Thr Asp Lys Pro Tyr Leu Ser Ile Leu Arg Lys Gln
50          55          60
Phe Gly Glu Met Thr Pro Ala Asn Ala Leu Lys Val Ser Gln Ser Asp
65          70          75          80
Phe Met Tyr Thr Glu Pro Glu Gln Asn Val Phe Asn Phe Thr Gln Gly
85          90          95
Asp Tyr Phe Met Asp Leu Ala Asp His Tyr Gly His Ala Val Arg Cys
100         105         110
His Asn Leu Val Trp Ala Ser Gln Val Ser Asp Trp Val Thr Ser Arg
115         120         125
Asn Trp Thr Ala Thr Glu Leu Lys Glu Val Met Lys Asn His Ile Phe
130         135         140
Lys Thr Val Gln His Phe Gly Lys Arg Cys Tyr Ala Trp Asp Val Val
145         150         155         160
Asn Glu Ala Ile Asn Gly Asp Gly Thr Phe Ser Ser Ser Val Trp Tyr
165         170         175
Asp Thr Ile Gly Glu Glu Tyr Phe Tyr Leu Ala Phe Gln Tyr Ala Gln
180         185         190
Glu Ala Leu Ala Gln Ile His Ala Asn Gln Val Lys Leu Tyr Tyr Asn
195         200         205
Asp Tyr Gly Ile Glu Asn Pro Gly Pro Lys Ala Asp Ala Val Leu Lys
210         215         220

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Leu Val Ala Glu Leu Arg Lys Arg Gly Ile Arg Ile Asp Gly Val Gly
 225 230 235 240

Leu Glu Ser His Phe Ile Val Gly Glu Thr Pro Ser Leu Ala Asp Gln
 245 250 255

Leu Ala Thr Lys Lys Ala Tyr Ile Glu Ala Gly Leu Glu Val Ala Ile
 260 265 270

Thr Glu Leu Asp Val Arg Phe Ser Gln Ala Pro Phe Tyr Thr Ala Glu
 275 280 285

Ala Gln Lys Gln Gln Ala Ala Asp Tyr Tyr Ala Ser Val Ala Ser Cys
 290 295 300

Lys His Ala Gly Pro Arg Cys Val Gly Val Val Val Trp Asp Phe Asp
 305 310 315 320

Asp Ala Tyr Ser Trp Ile Pro Gly Thr Phe Glu Gly Gln Gly Gly Ala
 325 330 335

Cys Leu Tyr Asn Glu Thr Leu Glu Val Lys Pro Ala Phe Tyr Ala Ala
 340 345 350

Ala Glu Ala Leu Glu Asn Lys Pro Cys Thr Val Cys
 355 360

<210> SEQ ID NO 15
 <211> LENGTH: 1400
 <212> TYPE: DNA
 <213> ORGANISM: Aspergillus fumigatus

<400> SEQUENCE: 15

atggtcgtcc tcagcaagct cgtcagcagc attctctttg tctccctggt ttcggcgggc 60
 gtgatcgacg aacgccaggc agccggcatc aaccaggcgt ttacctcca tggcaagaag 120
 tactttggca ccgccagtga ccaagctctg ctccagaagt cgcagaatga ggccattgtg 180
 cgcaaagact ttggccagct gacgccggag aatagcatga agtgggatgc gactgagcgt 240
 aggtctctcg gccactgtgg ctgacgttaa cttgttgaca tgactgtctg tgtagcatcg 300
 caaggaagat tcaacttcgc tgggtgctgat ttcttggtat gcaatctgct catctcggtc 360
 gagctcctgc tgaaggacaa taaataggtc aactatgcaa aacagaatgg caagaaggtc 420
 cgcggacaca ccttaggtat tcatgcgccc tcacggcatt tcgaggatac agccaagctg 480
 acagtgtagt ctggcactcc caactcccgt cctgggtgtc ggctatcagc gacaaaaaca 540
 ccctgacctc ggtgctgaag aaccacatca ccaccgtcat gaccgggtac aagggccaga 600
 tctacgctg ggtatatttc cctctatccc acacaatgcc agccccagct aatagctgca 660
 aaggacgtcg tcaacgagat cttcaacgag gacggctccc tccgcgacag cgtcttctcc 720
 cgcgtgctgg gcgaggactt tgtgctgatt gccttcgaga cggcgcgctc tgtggatccc 780
 tcggcgaagc tgtacatcaa cgattacaag taagcttggt gttttgtcga gagatgtact 840
 ccgtcctgga tctgaccatc acagtctcga ctccggctagc tatggcaaaa ccaggggat 900
 ggtgagatat gtcaagaagt ggctggctgc gggcattcct atcgatggaa tcggtgagca 960
 caggctcggg agctgtgtgt gatgattgta cgctgactct tcctgaaggc actcaaacc 1020
 accttggtgc ggggtgcttcg tccagcgtca aaggataagt ctcttggtt ttcttgctta 1080
 cgtaacgctg accccccgtg tacagcattg actgctcttg cgtcttccgg cgtctctgag 1140
 gtcgccatta ccgagctgga tatcgccggg gcgagctccc aggactacgt caatgtatgt 1200
 ctctgattg ccagtggcag ggtcatcgat actaatagaa acaggtcgtc aagcatgcc 1260
 tggatgtccc caagtgtgtg ggaatcaccg tctggggggg gtcggacagg gactcgtggc 1320

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gctccggtc gtctccgctg ctgttcgaca gcaactacca gcccaaggcg gcgtataatg 1380
 ccatcattgc tgctctctga 1400

<210> SEQ ID NO 16
 <211> LENGTH: 323
 <212> TYPE: PRT
 <213> ORGANISM: Aspergillus fumigatus

<400> SEQUENCE: 16

Met Val Val Leu Ser Lys Leu Val Ser Ser Ile Leu Phe Val Ser Leu
 1 5 10 15
 Val Ser Ala Gly Val Ile Asp Glu Arg Gln Ala Ala Gly Ile Asn Gln
 20 25 30
 Ala Phe Thr Ser His Gly Lys Lys Tyr Phe Gly Thr Ala Ser Asp Gln
 35 40 45
 Ala Leu Leu Gln Lys Ser Gln Asn Glu Ala Ile Val Arg Lys Asp Phe
 50 55 60
 Gly Gln Leu Thr Pro Glu Asn Ser Met Lys Trp Asp Ala Thr Glu Ala
 65 70 75 80
 Ser Gln Gly Arg Phe Asn Phe Ala Gly Ala Asp Phe Leu Val Asn Tyr
 85 90 95
 Ala Lys Gln Asn Gly Lys Lys Val Arg Gly His Thr Leu Trp His Ser
 100 105 110
 Gln Leu Pro Ser Trp Val Ser Ala Ile Ser Asp Lys Asn Thr Leu Thr
 115 120 125
 Ser Val Leu Lys Asn His Ile Thr Thr Val Met Thr Arg Tyr Lys Gly
 130 135 140
 Gln Ile Tyr Ala Trp Asp Val Val Asn Glu Ile Phe Asn Glu Asp Gly
 145 150 155 160
 Ser Leu Arg Asp Ser Val Phe Ser Arg Val Leu Gly Glu Asp Phe Val
 165 170 175
 Arg Ile Ala Phe Glu Thr Ala Arg Ser Val Asp Pro Ser Ala Lys Leu
 180 185 190
 Tyr Ile Asn Asp Tyr Lys Leu Asp Ser Ala Ser Tyr Gly Lys Thr Gln
 195 200 205
 Gly Met Val Arg Tyr Val Lys Lys Trp Leu Ala Ala Gly Ile Pro Ile
 210 215 220
 Asp Gly Ile Gly Gln Thr His Leu Gly Ala Gly Ala Ser Ser Ser Val
 225 230 235 240
 Lys Gly Ala Leu Thr Ala Leu Ala Ser Ser Gly Val Ser Glu Val Ala
 245 250 255
 Ile Thr Glu Leu Asp Ile Ala Gly Ala Ser Ser Gln Asp Tyr Val Asn
 260 265 270
 Val Val Lys Ala Cys Leu Asp Val Pro Lys Cys Val Gly Ile Thr Val
 275 280 285
 Trp Gly Val Ser Asp Arg Asp Ser Trp Arg Ser Gly Ser Ser Pro Leu
 290 295 300
 Leu Phe Asp Ser Asn Tyr Gln Pro Lys Ala Ala Tyr Asn Ala Ile Ile
 305 310 315 320
 Ala Ala Leu

<210> SEQ ID NO 17
 <211> LENGTH: 1415
 <212> TYPE: DNA

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<213> ORGANISM: *Aspergillus fumigatus*

<400> SEQUENCE: 17

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atggtccatc tatcttcatt ggcagcagcc ctggctgctc tgcctctgta tgtttaccca    60
ctcacgagag gaggaacagc tttgacattg ctatagtgta tatggagctg gcctgaacac    120
agcagccaaa gccaaaggac taaagtactt tggttccgcc acggacaatc cagagctcac    180
ggactctgcg tatgtcgcgc aactgagcaa caccgatgat tttgggtcaa tcacaccg    240
aaactccatg aaggtttget tacgtctgcc tccctggagc attgcctcaa aagctaattg    300
gttgttttgt ttggatagtg ggatgccacc gagccttctc agaattcttt ttcgttcgca    360
aatggagacg ccgtgggtcaa tctggcgaac aagaatggcc agctgatgcg atgccatact    420
ctggtctggc acagtcagct accgaactgg ggtatgtaa cgtcttctct attctcaaat    480
actctctaac agttgacagt ctctagcggg tcatggacca atgcgaccct tttggcggcc    540
atgaagaatc ataccaccaa tgtggttact cactacaagg ggaagtgcta cgctgggat    600
gttgatcaatg aaggtttgtt gctccatcta tctcaatag ttcttttgaa actgacaagc    660
ctgtcaatct agcctgaac gaggacggta ctttccgtaa ctctgtcttc taccagatca    720
tcggcccagc atacattcct attgcgttcg ccacggctgc tgccgcagat cccgacgtga    780
aactctacta caacgactac aacattgaat actcaggcgc caaagcgact gctgcgcaga    840
atatcgtcaa gatgatcaag gcctacggcg cgaagatcga cggcgtcggc ctccaggcac    900
actttatcgt cggcagcact ccgagtcaat cggatctgac gaccgtcttg aagggtaca    960
ctgctctcgg cgttgaggty gcctataccg aacttgacat ccgcatgcag ctgccctcga   1020
ccgccgcaaa gctggcccag cagtccactg acttccaagg cgtggccgca gcatgcgtta   1080
gcaccactgg ctgctggtgt gtcactatct gggactggac cgacaagtac tctgggtcc   1140
ccagcgtggt ccaaggctac ggcgccccat tgccttggga tgagaactat gtgaagaagc   1200
cagcgtacga tggcctgatg ggggtcttg gagcaagcgg ctccggcacc acaacgacca   1260
ctactactac ttctactacg acaggaggta cggaccctac tggagtgcgt cagaaatggg   1320
gacagtgtgg cggatttggc tggaccgggc caacaacttg tgtcagtggg accacttgcc   1380
aaaagctgaa tgactgttac tcacagtgcc tgtaa                                1415

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<210> SEQ ID NO 18

<211> LENGTH: 397

<212> TYPE: PRT

<213> ORGANISM: *Aspergillus fumigatus*

<400> SEQUENCE: 18

```

Met Val His Leu Ser Ser Leu Ala Ala Ala Leu Ala Ala Leu Pro Leu
1           5           10          15
Val Tyr Gly Ala Gly Leu Asn Thr Ala Ala Lys Ala Lys Gly Leu Lys
20          25          30
Tyr Phe Gly Ser Ala Thr Asp Asn Pro Glu Leu Thr Asp Ser Ala Tyr
35          40          45
Val Ala Gln Leu Ser Asn Thr Asp Asp Phe Gly Gln Ile Thr Pro Gly
50          55          60
Asn Ser Met Lys Trp Asp Ala Thr Glu Pro Ser Gln Asn Ser Phe Ser
65          70          75          80
Phe Ala Asn Gly Asp Ala Val Val Asn Leu Ala Asn Lys Asn Gly Gln
85          90          95
Leu Met Arg Cys His Thr Leu Val Trp His Ser Gln Leu Pro Asn Trp

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100			105			110									
Val	Ser	Ser	Gly	Ser	Trp	Thr	Asn	Ala	Thr	Leu	Leu	Ala	Ala	Met	Lys
		115					120					125			
Asn	His	Ile	Thr	Asn	Val	Val	Thr	His	Tyr	Lys	Gly	Lys	Cys	Tyr	Ala
	130					135					140				
Trp	Asp	Val	Val	Asn	Glu	Ala	Leu	Asn	Glu	Asp	Gly	Thr	Phe	Arg	Asn
145				150						155					160
Ser	Val	Phe	Tyr	Gln	Ile	Ile	Gly	Pro	Ala	Tyr	Ile	Pro	Ile	Ala	Phe
				165					170					175	
Ala	Thr	Ala	Ala	Ala	Ala	Asp	Pro	Asp	Val	Lys	Leu	Tyr	Tyr	Asn	Asp
		180						185						190	
Tyr	Asn	Ile	Glu	Tyr	Ser	Gly	Ala	Lys	Ala	Thr	Ala	Ala	Gln	Asn	Ile
	195						200					205			
Val	Lys	Met	Ile	Lys	Ala	Tyr	Gly	Ala	Lys	Ile	Asp	Gly	Val	Gly	Leu
	210					215					220				
Gln	Ala	His	Phe	Ile	Val	Gly	Ser	Thr	Pro	Ser	Gln	Ser	Asp	Leu	Thr
225					230					235					240
Thr	Val	Leu	Lys	Gly	Tyr	Thr	Ala	Leu	Gly	Val	Glu	Val	Ala	Tyr	Thr
				245					250					255	
Glu	Leu	Asp	Ile	Arg	Met	Gln	Leu	Pro	Ser	Thr	Ala	Ala	Lys	Leu	Ala
			260					265					270		
Gln	Gln	Ser	Thr	Asp	Phe	Gln	Gly	Val	Ala	Ala	Ala	Cys	Val	Ser	Thr
		275					280					285			
Thr	Gly	Cys	Val	Gly	Val	Thr	Ile	Trp	Asp	Trp	Thr	Asp	Lys	Tyr	Ser
	290					295					300				
Trp	Val	Pro	Ser	Val	Phe	Gln	Gly	Tyr	Gly	Ala	Pro	Leu	Pro	Trp	Asp
305					310					315					320
Glu	Asn	Tyr	Val	Lys	Lys	Pro	Ala	Tyr	Asp	Gly	Leu	Met	Ala	Gly	Leu
				325					330				335		
Gly	Ala	Ser	Gly	Ser	Gly	Thr	Thr	Thr	Thr	Thr	Thr	Thr	Thr	Ser	Thr
			340					345						350	
Thr	Thr	Gly	Gly	Thr	Asp	Pro	Thr	Gly	Val	Ala	Gln	Lys	Trp	Gly	Gln
		355					360					365			
Cys	Gly	Gly	Ile	Gly	Trp	Thr	Gly	Pro	Thr	Thr	Cys	Val	Ser	Gly	Thr
	370					375					380				
Thr	Cys	Gln	Lys	Leu	Asn	Asp	Trp	Tyr	Ser	Gln	Cys	Leu			
385					390					395					

<210> SEQ ID NO 19
 <211> LENGTH: 2376
 <212> TYPE: DNA
 <213> ORGANISM: Aspergillus fumigatus

<400> SEQUENCE: 19

```

atggcgggttg ccaaatctat tgctgccgtg ctggtagcac tgttgectgg tgcgcttgct    60
caggcgaata caagctatgt tgattacaat gtggaggcga atccggatct caccctcag    120
tcggtcgcta cgattgacct gtcctttccc gactgcgaga atggaccgct cagcaagact    180
ctcgtttgcg acacgtcggc tcggccgcat gaccgagctg ctgccctggt ttccatgttc    240
accttogagg agctggtgaa caacacaggc aactagacc ctggtgttcc aagacttgg    300
ctccctccgt accaagtatg gagcgaggct ctccatggac ttgaccgcgc caacttcaca    360
aacgagggag agtacagctg ggccacctcg ttccccatgc ctatcctgac aatgtcggcc    420
ttgaaccgaa ccctgatcaa ccagatcgcg accatcatcg caactcaagg acgagctttc    480
    
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aataacggtg ggcggtatgg gctggacgtg tacgccccga atataaatgc attcagatcg 540
gctatgtggg gaagaggtca agagaccccc ggagaagacg cttactgcct ggcacgggcg 600
tatgcgtagc agtatatcac tggcatccag ggtggtgttg atccggaaca cctcaagttg 660
gtggccactg ccaaacta tgccgggctac gatcttgaga actgggacgg tcaactccgt 720
ttgggcaacg atatgaacat tacacagcag gaactttccg aatactacac ccctcagttc 780
cttgttgcag ccagagacgc caaagtgcac agtgcacatg gtcctacaa cgcggtaaat 840
ggggtgcccc gctgcgcaaa ctggttcttc ctccagaccc tcctccgtga cacattcggc 900
ttcgtcgagg atggttatgt atccagcgcac tgcgactcgg cgtacaatgt ctggaacccg 960
cacgagtttg cggccaacat cacggggggc gctgcagact ctatccgggc ggggacggac 1020
attgattgcg gcactactta tcaatactat ttcggcgaag cctttgacga gcaagaggtc 1080
accctgagc aaatcgaaag aggtgtgatc cgctgtaca gcaacttggg gcgtctcggc 1140
tatttcgatg gcaatggaag cgtgtatcgg gacctgacgt ggaatgatgt cgtgaccacg 1200
gatgcctgga atatctcata cgaagccgct gtagaaggca ttgtcctact gaagaacgat 1260
ggaaccttgc ctctcgcaa gtcgggtccgc agtgttgcac tgattgggac ctggatgaat 1320
gtgacgactc agcttcaggg caactacttt ggaccggcgc cttatctgat tagtccgttg 1380
aatgccttcc agaattctga cttcgacgtg aactacgctt tcggcacgaa catttcatcc 1440
cactccacag atgggttttc cgaggcgttg tctgctgca agaaatccga cgtcatcata 1500
ttcgcgggcg ggattgacaa cactttggaa gcagaagcca tggatcgcat gaatataca 1560
tggccccgca atcagctaca gctcatcgac cagttgagcc aactcggcaa accgctgatc 1620
gtcctccaga tgggcgggcg ccaagtgcac tcctcctcgc tcaagtcaa caagaatgtc 1680
aactccctga tctggggtgg ataccccgga caatccggcg ggcaggctct cctagacatc 1740
atcacccgca agcgcgcccc cgccggccga ctcggtgca cgcagtacc ggccgaatac 1800
gcaaccagc tccccgccac cgacatgagc ctgcccctc acggcaataa tccccgccag 1860
acctacatgt ggtacaccgg ccccccgtc tacgagtttg gccacgggct cttctacacg 1920
acctccacg cctccctccc tggcaccggc aaggacaaga cctccttcaa catccaagac 1980
ctcctcacgc agcgcacatc gggcttcgca aacgtcgagc aaatgccttt gctcaacttc 2040
accgtgacga tcaccaatac cggcaaggtc gcttccgact aactgctat gctcttcgcg 2100
aacaccaccg cgggacctgc tccatacccg aacaagtggc tcgtcggctt cgaccggctg 2160
gcgagcctgg aaccgcacag gtgcgagact atgaccatcc ccgtgactat cgacagcgtg 2220
gctcgtagcg atgaggccgg caatcggggt ctctaccogg gaaagtacga gttggccctg 2280
aacaatgagc ggtcgggtgt ccttcagttt gtgctgacag gccgagaggc tgtgattttc 2340
aagtggcctg tagagcagca gcagatttcg tctgcg 2376

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<210> SEQ ID NO 20

<211> LENGTH: 792

<212> TYPE: PRT

<213> ORGANISM: *Aspergillus fumigatus*

<400> SEQUENCE: 20

```

Met Ala Val Ala Lys Ser Ile Ala Ala Val Leu Val Ala Leu Leu Pro
1           5           10           15

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Gly Ala Leu Ala Gln Ala Asn Thr Ser Tyr Val Asp Tyr Asn Val Glu
20           25           30

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Ala	Asn	Pro	Asp	Leu	Thr	Pro	Gln	Ser	Val	Ala	Thr	Ile	Asp	Leu	Ser
		35					40					45			
Phe	Pro	Asp	Cys	Glu	Asn	Gly	Pro	Leu	Ser	Lys	Thr	Leu	Val	Cys	Asp
	50					55					60				
Thr	Ser	Ala	Arg	Pro	His	Asp	Arg	Ala	Ala	Ala	Leu	Val	Ser	Met	Phe
65					70					75					80
Thr	Phe	Glu	Glu	Leu	Val	Asn	Asn	Thr	Gly	Asn	Thr	Ser	Pro	Gly	Val
				85					90					95	
Pro	Arg	Leu	Gly	Leu	Pro	Pro	Tyr	Gln	Val	Trp	Ser	Glu	Ala	Leu	His
			100					105					110		
Gly	Leu	Asp	Arg	Ala	Asn	Phe	Thr	Asn	Glu	Gly	Glu	Tyr	Ser	Trp	Ala
		115					120					125			
Thr	Ser	Phe	Pro	Met	Pro	Ile	Leu	Thr	Met	Ser	Ala	Leu	Asn	Arg	Thr
	130					135					140				
Leu	Ile	Asn	Gln	Ile	Ala	Thr	Ile	Ile	Ala	Thr	Gln	Gly	Arg	Ala	Phe
145					150					155					160
Asn	Asn	Val	Gly	Arg	Tyr	Gly	Leu	Asp	Val	Tyr	Ala	Pro	Asn	Ile	Asn
				165					170					175	
Ala	Phe	Arg	Ser	Ala	Met	Trp	Gly	Arg	Gly	Gln	Glu	Thr	Pro	Gly	Glu
			180					185					190		
Asp	Ala	Tyr	Cys	Leu	Ala	Ser	Ala	Tyr	Ala	Tyr	Glu	Tyr	Ile	Thr	Gly
		195					200					205			
Ile	Gln	Gly	Gly	Val	Asp	Pro	Glu	His	Leu	Lys	Leu	Val	Ala	Thr	Ala
	210					215					220				
Lys	His	Tyr	Ala	Gly	Tyr	Asp	Leu	Glu	Asn	Trp	Asp	Gly	His	Ser	Arg
225					230					235					240
Leu	Gly	Asn	Asp	Met	Asn	Ile	Thr	Gln	Gln	Glu	Leu	Ser	Glu	Tyr	Tyr
				245					250					255	
Thr	Pro	Gln	Phe	Leu	Val	Ala	Ala	Arg	Asp	Ala	Lys	Val	His	Ser	Val
			260					265					270		
Met	Cys	Ser	Tyr	Asn	Ala	Val	Asn	Gly	Val	Pro	Ser	Cys	Ala	Asn	Ser
		275					280					285			
Phe	Phe	Leu	Gln	Thr	Leu	Leu	Arg	Asp	Thr	Phe	Gly	Phe	Val	Glu	Asp
	290					295					300				
Gly	Tyr	Val	Ser	Ser	Asp	Cys	Asp	Ser	Ala	Tyr	Asn	Val	Trp	Asn	Pro
305					310					315					320
His	Glu	Phe	Ala	Ala	Asn	Ile	Thr	Gly	Ala	Ala	Ala	Asp	Ser	Ile	Arg
				325					330					335	
Ala	Gly	Thr	Asp	Ile	Asp	Cys	Gly	Thr	Thr	Tyr	Gln	Tyr	Tyr	Phe	Gly
			340					345						350	
Glu	Ala	Phe	Asp	Glu	Gln	Glu	Val	Thr	Arg	Ala	Glu	Ile	Glu	Arg	Gly
		355					360					365			
Val	Ile	Arg	Leu	Tyr	Ser	Asn	Leu	Val	Arg	Leu	Gly	Tyr	Phe	Asp	Gly
	370					375					380				
Asn	Gly	Ser	Val	Tyr	Arg	Asp	Leu	Thr	Trp	Asn	Asp	Val	Val	Thr	Thr
385					390					395					400
Asp	Ala	Trp	Asn	Ile	Ser	Tyr	Glu	Ala	Ala	Val	Glu	Gly	Ile	Val	Leu
				405					410					415	
Leu	Lys	Asn	Asp	Gly	Thr	Leu	Pro	Leu	Ala	Lys	Ser	Val	Arg	Ser	Val
			420					425					430		
Ala	Leu	Ile	Gly	Pro	Trp	Met	Asn	Val	Thr	Thr	Gln	Leu	Gln	Gly	Asn
		435					440					445			
Tyr	Phe	Gly	Pro	Ala	Pro	Tyr	Leu	Ile	Ser	Pro	Leu	Asn	Ala	Phe	Gln

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450	455	460
Asn Ser Asp Phe Asp Val Asn Tyr Ala Phe Gly Thr Asn Ile Ser Ser 465	470	475 480
His Ser Thr Asp Gly Phe Ser Glu Ala Leu Ser Ala Ala Lys Lys Ser 485	490	495
Asp Val Ile Ile Phe Ala Gly Gly Ile Asp Asn Thr Leu Glu Ala Glu 500	505	510
Ala Met Asp Arg Met Asn Ile Thr Trp Pro Gly Asn Gln Leu Gln Leu 515	520	525
Ile Asp Gln Leu Ser Gln Leu Gly Lys Pro Leu Ile Val Leu Gln Met 530	535	540
Gly Gly Gly Gln Val Asp Ser Ser Ser Leu Lys Ser Asn Lys Asn Val 545	550	555 560
Asn Ser Leu Ile Trp Gly Gly Tyr Pro Gly Gln Ser Gly Gly Gln Ala 565	570	575
Leu Leu Asp Ile Ile Thr Gly Lys Arg Ala Pro Ala Gly Arg Leu Val 580	585	590
Val Thr Gln Tyr Pro Ala Glu Tyr Ala Thr Gln Phe Pro Ala Thr Asp 595	600	605
Met Ser Leu Arg Pro His Gly Asn Asn Pro Gly Gln Thr Tyr Met Trp 610	615	620
Tyr Thr Gly Thr Pro Val Tyr Glu Phe Gly His Gly Leu Phe Tyr Thr 625	630	635 640
Thr Phe His Ala Ser Leu Pro Gly Thr Gly Lys Asp Lys Thr Ser Phe 645	650	655
Asn Ile Gln Asp Leu Leu Thr Gln Pro His Pro Gly Phe Ala Asn Val 660	665	670
Glu Gln Met Pro Leu Leu Asn Phe Thr Val Thr Ile Thr Asn Thr Gly 675	680	685
Lys Val Ala Ser Asp Tyr Thr Ala Met Leu Phe Ala Asn Thr Thr Ala 690	695	700
Gly Pro Ala Pro Tyr Pro Asn Lys Trp Leu Val Gly Phe Asp Arg Leu 705	710	715 720
Ala Ser Leu Glu Pro His Arg Ser Gln Thr Met Thr Ile Pro Val Thr 725	730	735
Ile Asp Ser Val Ala Arg Thr Asp Glu Ala Gly Asn Arg Val Leu Tyr 740	745	750
Pro Gly Lys Tyr Glu Leu Ala Leu Asn Asn Glu Arg Ser Val Val Leu 755	760	765
Gln Phe Val Leu Thr Gly Arg Glu Ala Val Ile Phe Lys Trp Pro Val 770	775	780
Glu Gln Gln Gln Ile Ser Ser Ala 785	790	

<210> SEQ ID NO 21
 <211> LENGTH: 1660
 <212> TYPE: DNA
 <213> ORGANISM: Aspergillus fumigatus

<400> SEQUENCE: 21

atgactcttc tatttggtat atttcttgcc cggettgcgg tggctgccgc ccagagcagc	60
tgcgtctcgg cttggtgagt cttcctcatg ctagagcctg cattttgcta agagtcaggg	120
gtcaatgtgg cggattttct tataccgggt ctacctgctg tcagagtggg aatacctgcg	180

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ccgagattaa ctcgtactac taccagtgc taccagcaac agctgtgagt gttatcccc 240
ccccccccc agtggtttgt actggtcata acgcagagta gacttcaact accacagtga 300
catctccctc tagctctggg tctgtctctg ccaacacgctc aggttataca acgacgactg 360
tcttgcccag tcccacagca acacagaacc cttatcctcc ggccaatgcc agtcatgtg 420
gatcgtgggc tctcgttgat aacgtttgct gcccatacta ctgtctctcg gataatgaat 480
ccgagtctcg caccagcagc tgtaccggcg gctgcggcag tcctgattcc tccatgtgca 540
agtccgggac gatgtgatg tgtgcacctt cgaagcgtga gcacggctga tacaaccaca 600
gggggagagc aacacaccgt aacttccaac gaggactggc attacagcgt gagcaattct 660
catcctgcct cggattcata ctgacgactg cagcgtcaa cacactttgg tctgaccagc 720
ggtggcgctt gcgggttcgg tctgtacggc ctctgcacca agaacagtgt caccgagcgc 780
tggacagacc caatgctggg ctctacctgc gacgccttct gcaccgcta tccgctctc 840
tgcaaagacc ccgctaactg cactctccgc gggaactttg ctgccccgaa cggagactac 900
tacactcagg taagcacaca cacaggttct ccaagcgaca actcctaagt caatagttct 960
ggccttccct ggctgcatcc ggcaatcccc acaactacct ctcttgccgg gagtgttctg 1020
aggtcatccg caccaagccc gacgggacag actacgccgt cggcgagagc ggatataccg 1080
accccatagt cctcgaggta gtagacagct gtccctgcaa tgccaattca aaatggtgct 1140
gtaagcgcac ccccccccc ccccttcccc ttttttcccc gccttccaac ggacaaaatc 1200
tcacaagacc agggggcagc gggcgagacc actgcggcga aatcgacttc aagtacggct 1260
gtcctctgcc ggaagatcc catcacatgg acctctccga tattgccatg ggccgactcc 1320
agggaaaagg cagtctcgc gacgggtgta ttccatccg ctaccggcgt gtccccctgc 1380
ccaagccggg gaacatctac atctggcttc atgatggggc gggaccgat tattttgcgc 1440
tgagcgcggg caataccaac gggcccggct ccgtaatcag tatcgaagtc caggctggtg 1500
gctccggggc atgggttgcc ctcgagcacg atccgaacta caccagtagc cggccgcagg 1560
agcggtatgg ctcttggtc ctcccgcagg gcaagggacc cactactgtg cctgtttcga 1620
tgaggctgac ggcgccaat ggttaattaa ctagaggtga 1660

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<210> SEQ ID NO 22

<211> LENGTH: 470

<212> TYPE: PRT

<213> ORGANISM: Aspergillus fumigatus

<400> SEQUENCE: 22

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Met Thr Leu Leu Phe Gly Ile Phe Leu Ala Arg Leu Ala Val Ala Ala
1           5           10           15
Ala Gln Ser Ser Cys Val Ser Ala Trp Gly Gln Cys Gly Gly Ile Ser
20           25           30
Tyr Thr Gly Ser Thr Cys Cys Gln Ser Gly Asn Thr Cys Ala Glu Ile
35           40           45
Asn Ser Tyr Tyr Tyr Gln Cys Ile Pro Ala Thr Ala Val Ser Thr Ser
50           55           60
Thr Thr Thr Val Thr Ser Pro Ser Ser Ser Gly Ser Ser Ser Ala Asn
65           70           75           80
Thr Ser Gly Tyr Thr Thr Thr Thr Val Leu Pro Ser Pro Thr Ala Thr
85           90           95
Gln Asn Pro Tyr Pro Pro Ala Asn Ala Ser Ser Cys Gly Ser Trp Ala
100          105          110

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Leu Val Asp Asn Val Cys Cys Pro Tyr Tyr Cys Leu Ser Asp Asn Glu
 115 120 125
 Ser Glu Ser Cys Thr Ser Ser Cys Thr Gly Gly Cys Gly Ser Pro Asp
 130 135 140
 Ser Ser Met Cys Lys Ser Gly Thr Met Trp Gly Glu Gln His Thr Val
 145 150 155 160
 Thr Ser Asn Glu Asp Trp His Tyr Ser Arg Ser Thr His Phe Gly Leu
 165 170 175
 Thr Ser Gly Gly Ala Cys Gly Phe Gly Leu Tyr Gly Leu Cys Thr Lys
 180 185 190
 Asn Ser Val Thr Ala Ser Trp Thr Asp Pro Met Leu Gly Ser Thr Cys
 195 200 205
 Asp Ala Phe Cys Thr Ala Tyr Pro Leu Leu Cys Lys Asp Pro Ala Asn
 210 215 220
 Val Thr Leu Arg Gly Asn Phe Ala Ala Pro Asn Gly Asp Tyr Tyr Thr
 225 230 235 240
 Gln Phe Trp Pro Ser Leu Ala Ala Ser Gly Asn Pro Asp Asn Tyr Leu
 245 250 255
 Ser Cys Gly Glu Cys Phe Glu Val Ile Arg Thr Lys Pro Asp Gly Thr
 260 265 270
 Asp Tyr Ala Val Gly Glu Ser Gly Tyr Thr Asp Pro Ile Val Leu Glu
 275 280 285
 Val Val Asp Ser Cys Pro Cys Asn Ala Asn Ser Lys Trp Cys Cys Lys
 290 295 300
 Arg Thr Pro Pro Pro Pro Leu Pro Leu Phe Ser Pro Pro Ser Asn Gly
 305 310 315 320
 Gln Asn Leu Thr Arg Pro Gly Gly Ser Gly Ala Asp His Cys Gly Glu
 325 330 335
 Ile Asp Phe Lys Tyr Gly Cys Pro Leu Pro Glu Gly Ser His His Met
 340 345 350
 Asp Leu Ser Asp Ile Ala Met Gly Arg Leu Gln Gly Asn Gly Ser Leu
 355 360 365
 Ala Asp Gly Val Ile Pro Ile Arg Tyr Arg Arg Val Pro Cys Pro Lys
 370 375 380
 Pro Gly Asn Ile Tyr Ile Trp Leu His Asp Gly Ala Gly Pro Tyr Tyr
 385 390 395 400
 Phe Ala Leu Ser Ala Val Asn Thr Asn Gly Pro Gly Ser Val Ile Ser
 405 410 415
 Ile Glu Val Gln Ala Gly Gly Ser Gly Pro Trp Val Ala Leu Glu His
 420 425 430
 Asp Pro Asn Tyr Thr Ser Ser Arg Pro Gln Glu Arg Tyr Gly Ser Trp
 435 440 445
 Val Leu Pro Gln Gly Lys Gly Pro Ile Thr Val Pro Val Ser Met Arg
 450 455 460
 Leu Thr Ala Pro Asn Gly
 465 470

<210> SEQ ID NO 23

<211> LENGTH: 1545

<212> TYPE: DNA

<213> ORGANISM: Trichoderma reesei

<400> SEQUENCE: 23

atgtatcgga agttggcgt catctcggcc ttcttgcca cagctcgtgc tcagtcggcc 60

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tgcactctcc aatcggagac tcacccgcct ctgacatggc agaaatgctc gtctggtggc 120
acgtgcactc aacagacagg ctccgtgggc atcgacgcca actggcgctg gactcacgct 180
acgaacagca gcacgaactg ctacgatggc aacacttggg gctcgaccct atgtcctgac 240
aacgagacct gcgcgaagaa ctgctgtctg gacggtgccg cctacgcgctc cacgtacgga 300
gttaccacga gcggtaacag cctctccatt ggctttgtca cccagtctgc gcagaagaac 360
gttggcgctc gcctttacct tatggcgagc gacacgacct accaggaatt caccctgctt 420
ggcaacgagt tctctttcga tgttgatggt tcgcagctgc cgtgcggtt gaacggagct 480
ctctacttcg tgtccatgga cgcggatggt ggctgagca agtatccac caacaccgct 540
ggcgccaagt acggcacggg gtactgtgac agccagtgtc cccgcgatct gaagttcatc 600
aatggccagg ccaacgttga gggctgggag ccgtcatcca acaacgcgaa cacgggcatt 660
ggaggacacg gaagctgctg ctctgagatg gatatctggg aggccaactc catctccgag 720
gctcttacc cccacccttg cacgactgtc ggccaggaga tctgcgaggg tgatgggtgc 780
ggcggaactt actccgataa cagatatggc ggcacttgcg atcccgatgg ctgcgactgg 840
aaccataacc gcctgggcaa caccagcttc tacggccctg gctcaagctt taccctcgat 900
accaccaaga aattgaccgt tgtcaccag ttcgagacgt cgggtgccat caaccgatac 960
tatgtccaga atggcgtcac tttccagcag cccaacgccg agcttggtag ttactctggc 1020
aacgagctca acgatgatta ctgcacagct gaggaggcag aattcggcgg atcctctttc 1080
tcagacaagg gggcctgac tcagttcaag aaggctacct ctggcggcat ggttctggtc 1140
atgagtctgt gggatgatta ctacgccaac atgctgtggc tggactccac ctaccgaca 1200
aacgagacct cctccacacc cggtgccgtg cgcggaagct gctccaccag ctccggtgtc 1260
cctgctcagg tcgaatctca gtctcccaac gccaaaggta ccttctccaa catcaagttc 1320
ggacccattg gcagcaccgg caaccctagc ggcggcaacc ctcccggcgg aaaccgcct 1380
ggcaccacca ccaccgccg cccagccact accactggaa gctctcccgg acctaccag 1440
tctcactacg gccagtgcgg cggatttggc tacagcggcc ccacggtctg cgccagcggc 1500
acaacttgcc aggtcctgaa cccttactac tctcagtgcc tgtaa 1545

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<210> SEQ ID NO 24

<211> LENGTH: 514

<212> TYPE: PRT

<213> ORGANISM: Trichoderma reesei

<400> SEQUENCE: 24

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Met Tyr Arg Lys Leu Ala Val Ile Ser Ala Phe Leu Ala Thr Ala Arg
1           5           10           15

Ala Gln Ser Ala Cys Thr Leu Gln Ser Glu Thr His Pro Pro Leu Thr
20           25           30

Trp Gln Lys Cys Ser Ser Gly Gly Thr Cys Thr Gln Gln Thr Gly Ser
35           40           45

Val Val Ile Asp Ala Asn Trp Arg Trp Thr His Ala Thr Asn Ser Ser
50           55           60

Thr Asn Cys Tyr Asp Gly Asn Thr Trp Ser Ser Thr Leu Cys Pro Asp
65           70           75           80

Asn Glu Thr Cys Ala Lys Asn Cys Cys Leu Asp Gly Ala Ala Tyr Ala
85           90           95

Ser Thr Tyr Gly Val Thr Thr Ser Gly Asn Ser Leu Ser Ile Gly Phe
100          105          110

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Val	Thr	Gln	Ser	Ala	Gln	Lys	Asn	Val	Gly	Ala	Arg	Leu	Tyr	Leu	Met
		115					120					125			
Ala	Ser	Asp	Thr	Thr	Tyr	Gln	Glu	Phe	Thr	Leu	Leu	Gly	Asn	Glu	Phe
	130					135					140				
Ser	Phe	Asp	Val	Asp	Val	Ser	Gln	Leu	Pro	Cys	Gly	Leu	Asn	Gly	Ala
145					150					155					160
Leu	Tyr	Phe	Val	Ser	Met	Asp	Ala	Asp	Gly	Gly	Val	Ser	Lys	Tyr	Pro
				165					170					175	
Thr	Asn	Thr	Ala	Gly	Ala	Lys	Tyr	Gly	Thr	Gly	Tyr	Cys	Asp	Ser	Gln
			180					185					190		
Cys	Pro	Arg	Asp	Leu	Lys	Phe	Ile	Asn	Gly	Gln	Ala	Asn	Val	Glu	Gly
		195					200					205			
Trp	Glu	Pro	Ser	Ser	Asn	Asn	Ala	Asn	Thr	Gly	Ile	Gly	Gly	His	Gly
	210					215					220				
Ser	Cys	Cys	Ser	Glu	Met	Asp	Ile	Trp	Glu	Ala	Asn	Ser	Ile	Ser	Glu
225					230					235					240
Ala	Leu	Thr	Pro	His	Pro	Cys	Thr	Thr	Val	Gly	Gln	Glu	Ile	Cys	Glu
				245					250					255	
Gly	Asp	Gly	Cys	Gly	Gly	Thr	Tyr	Ser	Asp	Asn	Arg	Tyr	Gly	Gly	Thr
			260					265					270		
Cys	Asp	Pro	Asp	Gly	Cys	Asp	Trp	Asn	Pro	Tyr	Arg	Leu	Gly	Asn	Thr
		275					280					285			
Ser	Phe	Tyr	Gly	Pro	Gly	Ser	Ser	Phe	Thr	Leu	Asp	Thr	Thr	Lys	Lys
	290					295					300				
Leu	Thr	Val	Val	Thr	Gln	Phe	Glu	Thr	Ser	Gly	Ala	Ile	Asn	Arg	Tyr
305					310					315					320
Tyr	Val	Gln	Asn	Gly	Val	Thr	Phe	Gln	Gln	Pro	Asn	Ala	Glu	Leu	Gly
				325					330					335	
Ser	Tyr	Ser	Gly	Asn	Glu	Leu	Asn	Asp	Asp	Tyr	Cys	Thr	Ala	Glu	Glu
			340					345					350		
Ala	Glu	Phe	Gly	Gly	Ser	Ser	Phe	Ser	Asp	Lys	Gly	Gly	Leu	Thr	Gln
		355					360					365			
Phe	Lys	Lys	Ala	Thr	Ser	Gly	Gly	Met	Val	Leu	Val	Met	Ser	Leu	Trp
	370					375					380				
Asp	Asp	Tyr	Tyr	Ala	Asn	Met	Leu	Trp	Leu	Asp	Ser	Thr	Tyr	Pro	Thr
385					390					395					400
Asn	Glu	Thr	Ser	Ser	Thr	Pro	Gly	Ala	Val	Arg	Gly	Ser	Cys	Ser	Thr
				405					410					415	
Ser	Ser	Gly	Val	Pro	Ala	Gln	Val	Glu	Ser	Gln	Ser	Pro	Asn	Ala	Lys
			420					425					430		
Val	Thr	Phe	Ser	Asn	Ile	Lys	Phe	Gly	Pro	Ile	Gly	Ser	Thr	Gly	Asn
		435					440					445			
Pro	Ser	Gly	Gly	Asn	Pro	Pro	Gly	Gly	Asn	Pro	Pro	Gly	Thr	Thr	Thr
	450					455					460				
Thr	Arg	Arg	Pro	Ala	Thr	Thr	Thr	Gly	Ser	Ser	Pro	Gly	Pro	Thr	Gln
465					470					475					480
Ser	His	Tyr	Gly	Gln	Cys	Gly	Gly	Ile	Gly	Tyr	Ser	Gly	Pro	Thr	Val
				485					490					495	
Cys	Ala	Ser	Gly	Thr	Thr	Cys	Gln	Val	Leu	Asn	Pro	Tyr	Tyr	Ser	Gln
			500					505					510		
Cys	Leu														

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<211> LENGTH: 1611

<212> TYPE: DNA

<213> ORGANISM: *Trichoderma reesei*

<400> SEQUENCE: 25

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atgattgtcg gcattctcac cacgctggct acgctggcca cactcgcagc tagtgtgcct    60
ctagaggagc ggcaagcttg ctcaagcgtc tggttaattat gtgaaccctc tcaagagacc    120
caaatactga gatatgtcaa ggggccaatg tggtagccag aattggtcgg gtccgacttg    180
ctgtgcttcc ggaagcacat gcgtctactc caacgactat tactcccagt gtcttcccgg    240
cgctgcaagc tcaagctcgt ccacgcgcgc cgcgtcgacg acttctcgag tatcccccac    300
aacatcccgg tcgagctccg cgacgcctcc acctggttct actactacca gagtacctcc    360
agtcggatcg ggaaccgcta cgtattcagg caaccctttt gttggggta ctccttgggc    420
caatgcatat tacgcctctg aagttagcag cctcgctatt cctagcttga ctggagccat    480
ggcactgct gcagcagctg tcgcaaaggt tccctctttt atgtggctgt aggtcctccc    540
ggaaccaagg caatctgtta ctgaaggctc atcattcact gcagagatac tcttgacaag    600
acccctctca tggagcaaac cttggccgac atccgcaccg ccaacaagaa tggcggtaac    660
tatgccggac agtttgtggt gtatgacttg ccgcatcgcg attgcgctgc ccttgccctg    720
aatggcgaat actctattgc cgatggtggc gtcgccaaat ataagaacta tatcgacacc    780
atcgtcaaaa ttgtcgtgga atattccgat atccggaccc tccctggtat tggatgagt    840
ttaaacacct gcctcccccc ccccttccct tcccttcccg ccggcatctt gtcggtgtgc    900
taactattgt tccctcttcc agagcctgac tctcttgcca acctggtgac caacctcggc    960
actccaaagt gtgccaatgc tcagtcagcc taccttgagt gcatcaacta cgccgtcaca   1020
cagctgaacc ttccaaatgt tgcgatgat ttggacgctg gccatgcagg atggcttggc    1080
tggccggcaa accaagaccg ggccgctcag ctatttgcaa atgtttaca gaatgcatcg   1140
tctccgagag ctcttcgagg attggcaacc aatgtcgcga actacaacgg gtggaacatt   1200
accagcccc catcgtacac gcaaggcaac gctgtctaca acgagaagct gtacatccac   1260
gctattggac gtcttcttgc caatcacggc tgggtccaaag ccttcttcat cactgatcaa   1320
ggtcgatcgg gaaagcagcc taccggacag caacagtggg gagactggtg caatgtgatc   1380
ggcaaccggat ttggtattcg cccatccgca aacctgggg actcgttgct ggattcgttt   1440
gtctgggtca agccaggcgg cgagtgtgac ggcaccagcg acagcagtgc gccacgattt   1500
gactcccact gtgcgctccc agatgccttg caaccggcgc ctcaagctgg tgcttggttc   1560
caagcctact ttgtgcagct tctcaciaac gcaaaccat cgttcctgta a           1611

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<210> SEQ ID NO 26

<211> LENGTH: 471

<212> TYPE: PRT

<213> ORGANISM: *Trichoderma reesei*

<400> SEQUENCE: 26

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Met Ile Val Gly Ile Leu Thr Thr Leu Ala Thr Leu Ala Thr Leu Ala
1           5           10          15

Ala Ser Val Pro Leu Glu Glu Arg Gln Ala Cys Ser Ser Val Trp Gly
20          25          30

Gln Cys Gly Gly Gln Asn Trp Ser Gly Pro Thr Cys Cys Ala Ser Gly
35          40          45

Ser Thr Cys Val Tyr Ser Asn Asp Tyr Tyr Ser Gln Cys Leu Pro Gly
50          55          60

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Ala Ala Ser Ser Ser Ser Ser Thr Arg Ala Ala Ser Thr Thr Ser Arg
 65 70 75 80
 Val Ser Pro Thr Thr Ser Arg Ser Ser Ser Ala Thr Pro Pro Pro Gly
 85 90 95
 Ser Thr Thr Thr Arg Val Pro Pro Val Gly Ser Gly Thr Ala Thr Tyr
 100 105 110
 Ser Gly Asn Pro Phe Val Gly Val Thr Pro Trp Ala Asn Ala Tyr Tyr
 115 120 125
 Ala Ser Glu Val Ser Ser Leu Ala Ile Pro Ser Leu Thr Gly Ala Met
 130 135 140
 Ala Thr Ala Ala Ala Ala Val Ala Lys Val Pro Ser Phe Met Trp Leu
 145 150 155 160
 Asp Thr Leu Asp Lys Thr Pro Leu Met Glu Gln Thr Leu Ala Asp Ile
 165 170 175
 Arg Thr Ala Asn Lys Asn Gly Gly Asn Tyr Ala Gly Gln Phe Val Val
 180 185 190
 Tyr Asp Leu Pro Asp Arg Asp Cys Ala Ala Leu Ala Ser Asn Gly Glu
 195 200 205
 Tyr Ser Ile Ala Asp Gly Gly Val Ala Lys Tyr Lys Asn Tyr Ile Asp
 210 215 220
 Thr Ile Arg Gln Ile Val Val Glu Tyr Ser Asp Ile Arg Thr Leu Leu
 225 230 235 240
 Val Ile Glu Pro Asp Ser Leu Ala Asn Leu Val Thr Asn Leu Gly Thr
 245 250 255
 Pro Lys Cys Ala Asn Ala Gln Ser Ala Tyr Leu Glu Cys Ile Asn Tyr
 260 265 270
 Ala Val Thr Gln Leu Asn Leu Pro Asn Val Ala Met Tyr Leu Asp Ala
 275 280 285
 Gly His Ala Gly Trp Leu Gly Trp Pro Ala Asn Gln Asp Pro Ala Ala
 290 295 300
 Gln Leu Phe Ala Asn Val Tyr Lys Asn Ala Ser Ser Pro Arg Ala Leu
 305 310 315 320
 Arg Gly Leu Ala Thr Asn Val Ala Asn Tyr Asn Gly Trp Asn Ile Thr
 325 330 335
 Ser Pro Pro Ser Tyr Thr Gln Gly Asn Ala Val Tyr Asn Glu Lys Leu
 340 345 350
 Tyr Ile His Ala Ile Gly Arg Leu Leu Ala Asn His Gly Trp Ser Asn
 355 360 365
 Ala Phe Phe Ile Thr Asp Gln Gly Arg Ser Gly Lys Gln Pro Thr Gly
 370 375 380
 Gln Gln Gln Trp Gly Asp Trp Cys Asn Val Ile Gly Thr Gly Phe Gly
 385 390 395 400
 Ile Arg Pro Ser Ala Asn Thr Gly Asp Ser Leu Leu Asp Ser Phe Val
 405 410 415
 Trp Val Lys Pro Gly Gly Glu Cys Asp Gly Thr Ser Asp Ser Ser Ala
 420 425 430
 Pro Arg Phe Asp Ser His Cys Ala Leu Pro Asp Ala Leu Gln Pro Ala
 435 440 445
 Pro Gln Ala Gly Ala Trp Phe Gln Ala Tyr Phe Val Gln Leu Leu Thr
 450 455 460
 Asn Ala Asn Pro Ser Phe Leu
 465 470

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<210> SEQ ID NO 27
 <211> LENGTH: 2615
 <212> TYPE: DNA
 <213> ORGANISM: Trichoderma reesei

<400> SEQUENCE: 27

atgggatttg gccgcaatgc tgccgagccc gagtgtttct gcaacgttat ccaggagatt	60
tgcgcttgcc caagagggag ttgacgggga gagtcccaac tggttccttc agtaacgcca	120
ccctggcaga ctatataact tgtggacaag actctgcttt gttgagttct tcctaccagt	180
cttgaccaag accattctgt tgagcccaat cagaaatgcg ttaccgaaca gcagctgcgc	240
tggcacttgc cactgggccc tttgctaggg cagacagtca gtatagctgg tccatactgg	300
gatgtatatg taccctggag acaccatgct gactcttgaa tcaaggtagc tcaacatcgg	360
gggcctcggc tgaggcagtt gtacctctg cagggactcc atggggaacc gcgtacgaca	420
aggcgaaggc cgcattggca aagctcaatc tccaagataa ggtcggcatc gtgagcggtg	480
tcggctggaa cggcggtcct tgcgctggaa acacatctcc ggcctccaag atcagctatc	540
catcgctatg ccttcaagac ggaccctcgt gtgttcgata ctcgacaggc agcacagcct	600
ttacgcggg cgttcaagcg gcctcgacgt gggatgtcaa tttgatccgc gaacgtggac	660
agttcatcgg tgaggaggtg aaggcctcgg ggattcatgt catacttggc cctgtggctg	720
ggcgcctggg aaagactccg cagggcggtc gcaactggga gggcttcggg gtcgatccat	780
atctcacggg cattgccatg ggtcaaacca tcaacggcat ccagtcggta ggcgtgcagg	840
cgacagcgaa gcactatata ctcaacgagc aggagctcaa tcgagaaacc atttcgagca	900
accagatga ccgaactctc catgagctgt atacttggcc atttgccgac gcggttcagg	960
ccaatgtcgc ttctgtcatg tgctcgtaca acaaggtaa taccacctgg gcctgcgagg	1020
atcagtacac gctgcagact gtgctgaaag accagctggg gttcccaggc tatgtcatga	1080
cggactggaa cgcacagcac acgactgtcc aaagcgcgaa ttctgggctt gacatgtcaa	1140
tgcttgccac agacttcaac ggtaacaatc ggctctgggg tccagctctc accaatgcgg	1200
taaatagcaa tcaggtcccc acgagcagag tcgacgatat ggtgactcgt atcctcgccg	1260
catggtactt gacaggccag gaccaggcag gctatccgtc gttcaacatc agcagaaatg	1320
ttcaaggaaa ccacaagacc aatgtcaggg caattgccag ggacggcatc gttctgctca	1380
agaatgacgc caacatcctg ccgctcaaga agcccgtag cattgccgtc gttggatctg	1440
ccgcaatcat tggttaaccac gccagaaact cgcctcgtg caacgacaaa ggctgcgacg	1500
acggggcctt gggcatgggt tggggttccg gcgccgtcaa ctatccgtac ttcgtcgcgc	1560
cctacgatgc catcaatacc agagcgtctt cgcagggcac ccaggttacc ttgagcaaca	1620
ccgacaacac gtcctcaggc gcatctgcag caagaggaaa ggacgtcggc atcgtcttca	1680
tcaccgccga ctcgggtgaa ggctacatca ccgtggaggg caacgcgggc gatcgcaaca	1740
acctggatcc gtggcacaac ggcaatgccc tgggtccaggc ggtggccggg gccaacagca	1800
acgtcattgt tgttgtccac tccgttggcg ccatcattct ggagcagatt cttgctcttc	1860
cgcaggtaaa ggccgttgtc tgggcgggtc ttccttctca ggagagcggc aatgcgctcg	1920
tcgacgtgct gtggggagat gtcagccctt ctggcaagct ggtgtacacc attgcgaaga	1980
gccccaatga ctataaact cgcacgttt ccggcggcag tgacagcttc agcgagggac	2040
tgttcatcga ctataagcac ttcgacgacg ccaatatcac gccgcggtag gagttcggct	2100
atggactgtg taagtttgct aacctgaaca atctattaga caggttgact gacggatgac	2160

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tgtggaatga tagcttacac caagttcaac tactcaccgcc tctccgtctt gtcgaccgcc 2220
aagtctgggc ctgcgactgg ggccggttg cggggaggcc cgagtgatct gttccagaat 2280
gtcgcgacag tcaccgttga catcgcaaac tctggccaag tgactggtgc cgaggtagcc 2340
cagctgtaca tcacctacc atcttcagca cccaggaccc ctccgaagca gctgcgagge 2400
tttgccaagc tgaacctcac gctggtcag agcggaacag caacgttcaa catccgacga 2460
cgagatctca gctactggga cacggcttcg cagaaatggg tggtgccgtc ggggtcgttt 2520
ggcatcagcg tgggagcgag cagccgggat atcaggctga cgagcactct gtcggtagcg 2580
tagcgcgagg aggggtgaagg cggttgacct gtgac 2615

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<210> SEQ ID NO 28

<211> LENGTH: 744

<212> TYPE: PRT

<213> ORGANISM: Trichoderma reesei

<400> SEQUENCE: 28

```

Met Arg Tyr Arg Thr Ala Ala Ala Leu Ala Leu Ala Thr Gly Pro Phe
1          5          10          15
Ala Arg Ala Asp Ser His Ser Thr Ser Gly Ala Ser Ala Glu Ala Val
20          25          30
Val Pro Pro Ala Gly Thr Pro Trp Gly Thr Ala Tyr Asp Lys Ala Lys
35          40          45
Ala Ala Leu Ala Lys Leu Asn Leu Gln Asp Lys Val Gly Ile Val Ser
50          55          60
Gly Val Gly Trp Asn Gly Gly Pro Cys Val Gly Asn Thr Ser Pro Ala
65          70          75          80
Ser Lys Ile Ser Tyr Pro Ser Leu Cys Leu Gln Asp Gly Pro Leu Gly
85          90          95
Val Arg Tyr Ser Thr Gly Ser Thr Ala Phe Thr Pro Gly Val Gln Ala
100         105         110
Ala Ser Thr Trp Asp Val Asn Leu Ile Arg Glu Arg Gly Gln Phe Ile
115         120         125
Gly Glu Glu Val Lys Ala Ser Gly Ile His Val Ile Leu Gly Pro Val
130         135         140
Ala Gly Pro Leu Gly Lys Thr Pro Gln Gly Gly Arg Asn Trp Glu Gly
145         150         155         160
Phe Gly Val Asp Pro Tyr Leu Thr Gly Ile Ala Met Gly Gln Thr Ile
165         170         175
Asn Gly Ile Gln Ser Val Gly Val Gln Ala Thr Ala Lys His Tyr Ile
180         185         190
Leu Asn Glu Gln Glu Leu Asn Arg Glu Thr Ile Ser Ser Asn Pro Asp
195         200         205
Asp Arg Thr Leu His Glu Leu Tyr Thr Trp Pro Phe Ala Asp Ala Val
210         215         220
Gln Ala Asn Val Ala Ser Val Met Cys Ser Tyr Asn Lys Val Asn Thr
225         230         235         240
Thr Trp Ala Cys Glu Asp Gln Tyr Thr Leu Gln Thr Val Leu Lys Asp
245         250         255
Gln Leu Gly Phe Pro Gly Tyr Val Met Thr Asp Trp Asn Ala Gln His
260         265         270
Thr Thr Val Gln Ser Ala Asn Ser Gly Leu Asp Met Ser Met Pro Gly
275         280         285

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Thr	Asp	Phe	Asn	Gly	Asn	Asn	Arg	Leu	Trp	Gly	Pro	Ala	Leu	Thr	Asn
290						295					300				
Ala	Val	Asn	Ser	Asn	Gln	Val	Pro	Thr	Ser	Arg	Val	Asp	Asp	Met	Val
305					310					315					320
Thr	Arg	Ile	Leu	Ala	Ala	Trp	Tyr	Leu	Thr	Gly	Gln	Asp	Gln	Ala	Gly
				325					330					335	
Tyr	Pro	Ser	Phe	Asn	Ile	Ser	Arg	Asn	Val	Gln	Gly	Asn	His	Lys	Thr
			340					345					350		
Asn	Val	Arg	Ala	Ile	Ala	Arg	Asp	Gly	Ile	Val	Leu	Leu	Lys	Asn	Asp
		355					360					365			
Ala	Asn	Ile	Leu	Pro	Leu	Lys	Lys	Pro	Ala	Ser	Ile	Ala	Val	Val	Gly
370						375					380				
Ser	Ala	Ala	Ile	Ile	Gly	Asn	His	Ala	Arg	Asn	Ser	Pro	Ser	Cys	Asn
385					390					395					400
Asp	Lys	Gly	Cys	Asp	Asp	Gly	Ala	Leu	Gly	Met	Gly	Trp	Gly	Ser	Gly
				405					410					415	
Ala	Val	Asn	Tyr	Pro	Tyr	Phe	Val	Ala	Pro	Tyr	Asp	Ala	Ile	Asn	Thr
			420					425					430		
Arg	Ala	Ser	Ser	Gln	Gly	Thr	Gln	Val	Thr	Leu	Ser	Asn	Thr	Asp	Asn
		435					440					445			
Thr	Ser	Ser	Gly	Ala	Ser	Ala	Ala	Arg	Gly	Lys	Asp	Val	Ala	Ile	Val
450						455					460				
Phe	Ile	Thr	Ala	Asp	Ser	Gly	Glu	Gly	Tyr	Ile	Thr	Val	Glu	Gly	Asn
465					470					475					480
Ala	Gly	Asp	Arg	Asn	Asn	Leu	Asp	Pro	Trp	His	Asn	Gly	Asn	Ala	Leu
				485					490					495	
Val	Gln	Ala	Val	Ala	Gly	Ala	Asn	Ser	Asn	Val	Ile	Val	Val	Val	His
			500					505					510		
Ser	Val	Gly	Ala	Ile	Ile	Leu	Glu	Gln	Ile	Leu	Ala	Leu	Pro	Gln	Val
	515						520					525			
Lys	Ala	Val	Val	Trp	Ala	Gly	Leu	Pro	Ser	Gln	Glu	Ser	Gly	Asn	Ala
530						535					540				
Leu	Val	Asp	Val	Leu	Trp	Gly	Asp	Val	Ser	Pro	Ser	Gly	Lys	Leu	Val
545					550					555					560
Tyr	Thr	Ile	Ala	Lys	Ser	Pro	Asn	Asp	Tyr	Asn	Thr	Arg	Ile	Val	Ser
				565					570					575	
Gly	Gly	Ser	Asp	Ser	Phe	Ser	Glu	Gly	Leu	Phe	Ile	Asp	Tyr	Lys	His
			580					585					590		
Phe	Asp	Asp	Ala	Asn	Ile	Thr	Pro	Arg	Tyr	Glu	Phe	Gly	Tyr	Gly	Leu
		595					600					605			
Ser	Tyr	Thr	Lys	Phe	Asn	Tyr	Ser	Arg	Leu	Ser	Val	Leu	Ser	Thr	Ala
	610					615					620				
Lys	Ser	Gly	Pro	Ala	Thr	Gly	Ala	Val	Val	Pro	Gly	Gly	Pro	Ser	Asp
625					630					635					640
Leu	Phe	Gln	Asn	Val	Ala	Thr	Val	Thr	Val	Asp	Ile	Ala	Asn	Ser	Gly
				645					650					655	
Gln	Val	Thr	Gly	Ala	Glu	Val	Ala	Gln	Leu	Tyr	Ile	Thr	Tyr	Pro	Ser
			660					665					670		
Ser	Ala	Pro	Arg	Thr	Pro	Pro	Lys	Gln	Leu	Arg	Gly	Phe	Ala	Lys	Leu
		675					680					685			
Asn	Leu	Thr	Pro	Gly	Gln	Ser	Gly	Thr	Ala	Thr	Phe	Asn	Ile	Arg	Arg
690						695					700				
Arg	Asp	Leu	Ser	Tyr	Trp	Asp	Thr	Ala	Ser	Gln	Lys	Trp	Val	Val	Pro

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705	710	715	720
Ser Gly Ser Phe Gly Ile Ser Val Gly Ala Ser Ser Arg Asp Ile Arg			
	725	730	735
Leu Thr Ser Thr Leu Ser Val Ala			
	740		
<p><210> SEQ ID NO 29 <211> LENGTH: 1377 <212> TYPE: DNA <213> ORGANISM: Trichoderma reesei</p>			
<p><400> SEQUENCE: 29</p>			
atggcgccct cagttacact gccggtgacc acggccatcc tggccattgc cgggctcgtc			60
gccgcccagc aaccgggtac cagcaccccc gaggtccatc ccaagttgac aacctacaag			120
tgtacaaaagt cgggggggtg cgtggcccag gacacctcgg tggtccttga ctggaactac			180
cgctggatgc acgacgcaaa ctacaactcg tgcaccgtca acggcggcgt caacaccacg			240
ctctgccctg acgaggcgac ctgtggcaag aactgcttca tcgagggcgt cgactacgcc			300
gcctcgggcg tcacgacctc gggcagcagc ctacccatga accagtacat gcccagcagc			360
tctggcggct acagcagcgt ctctcctcgg ctgtatctcc tggactctga cggtgagtac			420
gtgatgctga agctcaacgg ccaggagctg agcttcgacg tcgacctctc tgctctgccg			480
tgtggagaga acggctcgt ctacctgtct cagatggacg agaacggggg cgccaaccag			540
tataaacagg cgggtgccaa ctacgggagc ggctactgcg atgctcagtg ccccgccag			600
acatggagga acggcacct caaactagc caccaggcgt tctgctgcaa cgagatggat			660
atcctggagg gcaactcgag ggccaatgcc ttgaccctc actcttgac ggccacggcc			720
tggcactctg cgggttgagg cttcaacccc tatggcagcg gctacaaaag ctactacggc			780
cccggagata cggttgacac ctccaagacc ttcaccatca tcaccagtt caacacggac			840
aacggctcgc cctcgggcaa ccttgtgagc atcacccgca agtaccagca aaacggcgtc			900
gacatcccca gcgccagcc cggcggcgac accatctcgt cctgcccgtc cgccctcagcc			960
tacggcggcc tcgccaccat gggcaaggcc ctgagcagcg gcatggtgct cgtgttcagc			1020
atttgaacg acaacagcca gtacatgaac tggctcgaca gcggcaacgc cggcccctgc			1080
agcagcaccg agggcaaccc atccaacatc ctggccaaca accccaacac gcacgtcgtc			1140
ttctccaaca tccgctgggg agacattggg tctactacga actcgactgc gccccgccc			1200
ccgcctcgtt ccagcagcag gttttcgact acacggagga gctcgacgac ttcgagcagc			1260
ccgagctgca cgcagactca ctgggggagc tgcggtggca ttgggtacag cgggtgcaag			1320
acgtgcacgt cgggcactac gtgccagtat agcaacgact actactcgca atgcctt			1377
<p><210> SEQ ID NO 30 <211> LENGTH: 459 <212> TYPE: PRT <213> ORGANISM: Trichoderma reesei</p>			
<p><400> SEQUENCE: 30</p>			
Met Ala Pro Ser Val Thr Leu Pro Leu Thr Thr Ala Ile Leu Ala Ile			
1	5	10	15
Ala Arg Leu Val Ala Ala Gln Gln Pro Gly Thr Ser Thr Pro Glu Val			
	20	25	30
His Pro Lys Leu Thr Thr Tyr Lys Cys Thr Lys Ser Gly Gly Cys Val			
	35	40	45

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<210> SEQ ID NO 31
 <211> LENGTH: 1254
 <212> TYPE: DNA
 <213> ORGANISM: Trichoderma reesei

<400> SEQUENCE: 31

```

atgaacaagt cegtggctcc attgctgctt gcagcgtcca tactatatgg cggcgccgctc   60
gcacagcaga ctgtctgggg ccagtgtgga ggtattggtt ggagcggacc tacgaattgt   120
gctcctgggt cagcttgctt gaccctcaat ccttattatg cgcaatgtat tccgggagcc   180
actactatca ccacttcgac cgggccacca tccgggccaa ccaccaccac cagggctacc   240
tcaacaagct catcaactcc acccagcagc tctggggctc gatttgccgg cgttaacatc   300
gcgggttttg actttggctg taccacagat ggcacttgcg ttacctcgaa ggtttatcct   360
ccgttgaaga acttcaccgg ctcaaacaac taccctgatg gcatcggcca gatgcagcac   420
ttcgtcaacg aggacgggat gactatcttc cgttacctcg tcggatggca gtacctcgtc   480
aacaacaatt tgggcgga tcttgattcc acgagcattt ccaagtatga tcagcttggt   540
caggggtgcc tgtctctggg cgcatactgc atcgtcgaca tccacaatta tgctcgatgg   600
aacggtggga tcatttgtca gggcgccct actaatgctc aattcacgag cctttggctg   660
cagttggcat caaagtacgc atctcagtcg aggggtgtgt tcggcatcat gaatgagccc   720
cacgacgtga acatcaacac ctgggctgcc acggtccaag aggttgtaac cgcaatccgc   780
aacgctggtg ctacgtcgca attcatctct ttgcctggaa atgattggca atctgctggg   840
gctttcatat ccgatggcag tgcagccgcc ctgtctcaag tcacgaacct ggatgggtca   900
acaacgaatc tgatttttga cgtgcacaaa tacttggact cagacaactc cggctactcac   960
gccgaatgta ctacaaataa cattgacggc gccttttctc cgcttgccac ttggctccga  1020
cagaacaatc gccaggctat cctgacagaa accggtgtgt gcaacgttca gtctgcata  1080
caagacatgt gccagcaaat ccaatatctc aaccagaact cagatgteta tcttggtat  1140
gttggtggg gtgccggatc atttgatagc acgtatgtcc tgacggaaac accgactagc  1200
agtggtaact catggacgga cacatccttg gtcagctcgt gtctcgcaag aaag      1254

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<210> SEQ ID NO 32
 <211> LENGTH: 418
 <212> TYPE: PRT
 <213> ORGANISM: Trichoderma reesei

<400> SEQUENCE: 32

```

Met Asn Lys Ser Val Ala Pro Leu Leu Leu Ala Ala Ser Ile Leu Tyr
 1           5           10           15

Gly Gly Ala Val Ala Gln Gln Thr Val Trp Gly Gln Cys Gly Gly Ile
          20           25           30

Gly Trp Ser Gly Pro Thr Asn Cys Ala Pro Gly Ser Ala Cys Ser Thr
 35           40           45

Leu Asn Pro Tyr Tyr Ala Gln Cys Ile Pro Gly Ala Thr Thr Ile Thr
 50           55           60

Thr Ser Thr Arg Pro Pro Ser Gly Pro Thr Thr Thr Thr Arg Ala Thr
 65           70           75           80

Ser Thr Ser Ser Ser Thr Pro Pro Thr Ser Ser Gly Val Arg Phe Ala
          85           90           95

Gly Val Asn Ile Ala Gly Phe Asp Phe Gly Cys Thr Thr Asp Gly Thr
          100          105          110

Cys Val Thr Ser Lys Val Tyr Pro Pro Leu Lys Asn Phe Thr Gly Ser

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115	120	125
Asn Asn Tyr Pro Asp Gly Ile Gly Gln Met Gln His Phe Val Asn Glu 130 135 140		
Asp Gly Met Thr Ile Phe Arg Leu Pro Val Gly Trp Gln Tyr Leu Val 145 150 155 160		
Asn Asn Asn Leu Gly Gly Asn Leu Asp Ser Thr Ser Ile Ser Lys Tyr 165 170 175		
Asp Gln Leu Val Gln Gly Cys Leu Ser Leu Gly Ala Tyr Cys Ile Val 180 185 190		
Asp Ile His Asn Tyr Ala Arg Trp Asn Gly Gly Ile Ile Gly Gln Gly 195 200 205		
Gly Pro Thr Asn Ala Gln Phe Thr Ser Leu Trp Ser Gln Leu Ala Ser 210 215 220		
Lys Tyr Ala Ser Gln Ser Arg Val Trp Phe Gly Ile Met Asn Glu Pro 225 230 235 240		
His Asp Val Asn Ile Asn Thr Trp Ala Ala Thr Val Gln Glu Val Val 245 250 255		
Thr Ala Ile Arg Asn Ala Gly Ala Thr Ser Gln Phe Ile Ser Leu Pro 260 265 270		
Gly Asn Asp Trp Gln Ser Ala Gly Ala Phe Ile Ser Asp Gly Ser Ala 275 280 285		
Ala Ala Leu Ser Gln Val Thr Asn Pro Asp Gly Ser Thr Thr Asn Leu 290 295 300		
Ile Phe Asp Val His Lys Tyr Leu Asp Ser Asp Asn Ser Gly Thr His 305 310 315 320		
Ala Glu Cys Thr Thr Asn Asn Ile Asp Gly Ala Phe Ser Pro Leu Ala 325 330 335		
Thr Trp Leu Arg Gln Asn Asn Arg Gln Ala Ile Leu Thr Glu Thr Gly 340 345 350		
Gly Gly Asn Val Gln Ser Cys Ile Gln Asp Met Cys Gln Gln Ile Gln 355 360 365		
Tyr Leu Asn Gln Asn Ser Asp Val Tyr Leu Gly Tyr Val Gly Trp Gly 370 375 380		
Ala Gly Ser Phe Asp Ser Thr Tyr Val Leu Thr Glu Thr Pro Thr Ser 385 390 395 400		
Ser Gly Asn Ser Trp Thr Asp Thr Ser Leu Val Ser Ser Cys Leu Ala 405 410 415		

Arg Lys

<210> SEQ ID NO 33
 <211> LENGTH: 752
 <212> TYPE: DNA
 <213> ORGANISM: Trichoderma reesei

<400> SEQUENCE: 33

atgggttgcc tttccagcct catctgcgct ctaccagca tcgccagtac tctggcgatg 60
 cccacaggcc tcgagcctga gaggcgtgac aacgtcacag agcgtggcat gtacgacttt 120
 gttcttgagg ctcaaatga tcatcgccgt cgtgctagca tcaactacga ccaaaactac 180
 caaactggcg gacaagtcag ctattcgcct tccaacactg gcttctcagt gaactggaac 240
 actcaagatg actttgttgt gggcgttggt tggacgactg gatcttctgc gtaggaggac 300
 tcctcatcat tctgcacttt gaaagcatct tctgaccaa agcttctctt agtccatca 360
 actttggcgg ctcttttagt gtcaacagcg gaactggcct gctttccgct tatggctgga 420

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gcaccaaccc actggttgag tactacatca tggaggacaa ccacaactac ccagcacagg 480
gtaccgtcaa ggggaaccgtc accagcgacg gagccactta caccatctgg gagaataccc 540
gtgtcaacga gccttccatc cagggcacag cgaccttcaa ccagtacatt tccgtgcgga 600
actcgcccag gaccagcgga actggttactg tgcagaacca cttcaatgct tgggcctcgc 660
ttggcctgca ccttgggcag atgaactacc aggttgctgc tgtcgaaggc tggggtggtgta 720
gtggttctgc ctcacagagt gtcagcaact ag 752

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<210> SEQ ID NO 34
<211> LENGTH: 229
<212> TYPE: PRT
<213> ORGANISM: Trichoderma reesei

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```

<400> SEQUENCE: 34

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```

Met Val Ala Phe Ser Ser Leu Ile Cys Ala Leu Thr Ser Ile Ala Ser
1           5           10           15
Thr Leu Ala Met Pro Thr Gly Leu Glu Pro Glu Ser Ser Val Asn Val
20           25           30
Thr Glu Arg Gly Met Tyr Asp Phe Val Leu Gly Ala His Asn Asp His
35           40           45
Arg Arg Arg Ala Ser Ile Asn Tyr Asp Gln Asn Tyr Gln Thr Gly Gly
50           55           60
Gln Val Ser Tyr Ser Pro Ser Asn Thr Gly Phe Ser Val Asn Trp Asn
65           70           75           80
Thr Gln Asp Asp Phe Val Val Gly Val Gly Trp Thr Thr Gly Ser Ser
85           90           95
Ala Pro Ile Asn Phe Gly Gly Ser Phe Ser Val Asn Ser Gly Thr Gly
100          105          110
Leu Leu Ser Val Tyr Gly Trp Ser Thr Asn Pro Leu Val Glu Tyr Tyr
115          120          125
Ile Met Glu Asp Asn His Asn Tyr Pro Ala Gln Gly Thr Val Lys Gly
130          135          140
Thr Val Thr Ser Asp Gly Ala Thr Tyr Thr Ile Trp Glu Asn Thr Arg
145          150          155          160
Val Asn Glu Pro Ser Ile Gln Gly Thr Ala Thr Phe Asn Gln Tyr Ile
165          170          175
Ser Val Arg Asn Ser Pro Arg Thr Ser Gly Thr Val Thr Val Gln Asn
180          185          190
His Phe Asn Ala Trp Ala Ser Leu Gly Leu His Leu Gly Gln Met Asn
195          200          205
Tyr Gln Val Val Ala Val Glu Gly Trp Gly Gly Ser Gly Ser Ala Ser
210          215          220
Gln Ser Val Ser Asn
225

```

```

<210> SEQ ID NO 35
<211> LENGTH: 796
<212> TYPE: DNA
<213> ORGANISM: Trichoderma reesei

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<400> SEQUENCE: 35

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```

caagaagaca tcaacatggt ctccttcacc tcctcctcgc cggcgctcgc cgccatctcg 60
ggcgtcttgg ccgctcccgc cgccgaggtc gaatccgtgg ctgtggagaa gcgccagacg 120
attcagcccg gcacgggcta caacaacggc tacttctact cgtactggaa cgatggccac 180

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ggcggcgtga cgtacaccaa tggccccggc gggcagttct ccgtcaactg gtccaactcg 240
ggcaactttg tcggcgccaa gggatggcag cccggcacca agaacaagta agactaccta 300
ctcttaccct ctttgaccaa cacagcacia cacaatacaa cacatgtgac taccaatcat 360
ggaatcggat ctaacagctg tgttttcaaa aaaaagggtc atcaacttct cgggcagcta 420
caaccccaac ggcaacagct acctctccgt gtacggctgg tcccgcaacc ccctgatcga 480
gtactacatc gtcgagaact ttggcaccta caaccctcc acggggcgcca ccaagctggg 540
cgaggtcacc tccgacggca gcgtctacga catttaccgc acgcagcgcg tcaaccagcc 600
gtccatcacc gccaccgcca ccttttacca gtactggctc gtccgcccga accaccgctc 660
gagcggctcc gtcaacacgg cgaaccactt caacgcgtgg gctcagcaag gcctgacgct 720
cgggacgatg gattaccaga ttgttgccgt ggagggttac tttagctctg gctctgcttc 780
catcacctgc agctaa 796

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<210> SEQ ID NO 36
<211> LENGTH: 223
<212> TYPE: PRT
<213> ORGANISM: Trichoderma reesei

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<400> SEQUENCE: 36

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```

Met Val Ser Phe Thr Ser Leu Leu Ala Gly Val Ala Ala Ile Ser Gly
1           5           10          15
Val Leu Ala Ala Pro Ala Ala Glu Val Glu Ser Val Ala Val Glu Lys
20          25          30
Arg Gln Thr Ile Gln Pro Gly Thr Gly Tyr Asn Asn Gly Tyr Phe Tyr
35          40          45
Ser Tyr Trp Asn Asp Gly His Gly Gly Val Thr Tyr Thr Asn Gly Pro
50          55          60
Gly Gly Gln Phe Ser Val Asn Trp Ser Asn Ser Gly Asn Phe Val Gly
65          70          75          80
Gly Lys Gly Trp Gln Pro Gly Thr Lys Asn Lys Val Ile Asn Phe Ser
85          90          95
Gly Ser Tyr Asn Pro Asn Gly Asn Ser Tyr Leu Ser Val Tyr Gly Trp
100         105         110
Ser Arg Asn Pro Leu Ile Glu Tyr Tyr Ile Val Glu Asn Phe Gly Thr
115         120         125
Tyr Asn Pro Ser Thr Gly Ala Thr Lys Leu Gly Glu Val Thr Ser Asp
130         135         140
Gly Ser Val Tyr Asp Ile Tyr Arg Thr Gln Arg Val Asn Gln Pro Ser
145         150         155         160
Ile Ile Gly Thr Ala Thr Phe Tyr Gln Tyr Trp Ser Val Arg Arg Asn
165         170         175
His Arg Ser Ser Gly Ser Val Asn Thr Ala Asn His Phe Asn Ala Trp
180         185         190
Ala Gln Gln Gly Leu Thr Leu Gly Thr Met Asp Tyr Gln Ile Val Ala
195         200         205
Val Glu Gly Tyr Phe Ser Ser Gly Ser Ala Ser Ile Thr Val Ser
210         215         220

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<210> SEQ ID NO 37
<211> LENGTH: 1352
<212> TYPE: DNA
<213> ORGANISM: Trichoderma reesei

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-continued

<400> SEQUENCE: 37

```

atgaaagcaa acgtcatctt gtgcctcctg gccccctgg tcgcccgtct ccccaccgaa    60
accatccacc tcgaccccga gctcgccgct ctccgcgcca acctcaccga gcgaacagcc    120
gacctctggg accgccaagc ctctcaaagc atcgaccagc tcatcaagag aaaaggcaag    180
ctctactttg gcaccgccac cgaccgcggc ctctccaac gggaaaagaa cggggccatc    240
atccaggcag acctcggcca ggtgacgccc gagaacagca tgaagtggca gtcgctcgag    300
aacaaccaag gccagctgaa ctggggagac gccgactatc tcgtcaactt tgcccagcaa    360
aacggcaagt cgatacgcgg ccacactctg atctggcact cgcagctgcc tgcgtgggtg    420
aacaatatca acaacgcgga tactctgegg caagtcatcc gcacccatgt ctctactgtg    480
gttgggcggt acaagggcaa gattcgtgct tgggtgagtt ttgaacacca catgccctt    540
ttcttagtcc gctcctctc ctcttggaac ttctcacagt tatagccgta tacaacattc    600
gacaggaaat ttaggatgac aactactgac tgacttgtgt gtgtgatggc gataggacgt    660
gttcaatgaa atcttcaacg aggatggaac gctgcgctct tcagtctttt ccaggctcct    720
cggcgaggag tttgtctcga ttgcctttcg tgctgctcga gatgctgacc cttctgcccg    780
tctttacatc aacgactaca atctcgaccg cgccaactat ggcaaggcca acggggtgaa    840
gacttacgtc tccaagtgga tctctcaagg agttcccatt gacggtattg gtgagccacg    900
accctaataa gtccccatt agagtctctt tctagagcca aggcttgaag ccattcaggg    960
actgacacga gagccttctc tacaggaagc cagtcccata tcagcggcgg cggaggctct  1020
ggtacgctgg gtgcgctcca gcagctggca acggtaccgg tcaccgagct ggccattacc  1080
gagctggaca ttcagggggc accgacgacg gattacaccc aagttgttca agcatgcctg  1140
agcgtctcca agtgcgctcg catcaccgtg tggggcatca gtgacaaggt aagttgcttc  1200
ccctgtctgt gcttatcaac tgtaagcagc aacaactgat gctgtctgtc tttacctagg  1260
actcgtggcg tgccagcacc aaccctcttc tgtttgacgc aaacttcaac cccaagccgg  1320
catataacag cattggtggc atcttacaat ag                                1352

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<210> SEQ ID NO 38

<211> LENGTH: 347

<212> TYPE: PRT

<213> ORGANISM: Trichoderma reesei

<400> SEQUENCE: 38

```

Met Lys Ala Asn Val Ile Leu Cys Leu Leu Ala Pro Leu Val Ala Ala
1           5           10           15

Leu Pro Thr Glu Thr Ile His Leu Asp Pro Glu Leu Ala Ala Leu Arg
          20           25           30

Ala Asn Leu Thr Glu Arg Thr Ala Asp Leu Trp Asp Arg Gln Ala Ser
          35           40           45

Gln Ser Ile Asp Gln Leu Ile Lys Arg Lys Gly Lys Leu Tyr Phe Gly
          50           55           60

Thr Ala Thr Asp Arg Gly Leu Leu Gln Arg Glu Lys Asn Ala Ala Ile
65           70           75           80

Ile Gln Ala Asp Leu Gly Gln Val Thr Pro Glu Asn Ser Met Lys Trp
          85           90           95

Gln Ser Leu Glu Asn Asn Gln Gly Gln Leu Asn Trp Gly Asp Ala Asp
          100          105          110

Tyr Leu Val Asn Phe Ala Gln Gln Asn Gly Lys Ser Ile Arg Gly His
          115          120          125

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Thr Leu Ile Trp His Ser Gln Leu Pro Ala Trp Val Asn Asn Ile Asn
 130 135 140

Asn Ala Asp Thr Leu Arg Gln Val Ile Arg Thr His Val Ser Thr Val
 145 150 155 160

Val Gly Arg Tyr Lys Gly Lys Ile Arg Ala Trp Asp Val Val Asn Glu
 165 170 175

Ile Phe Asn Glu Asp Gly Thr Leu Arg Ser Ser Val Phe Ser Arg Leu
 180 185 190

Leu Gly Glu Glu Phe Val Ser Ile Ala Phe Arg Ala Ala Arg Asp Ala
 195 200 205

Asp Pro Ser Ala Arg Leu Tyr Ile Asn Asp Tyr Asn Leu Asp Arg Ala
 210 215 220

Asn Tyr Gly Lys Val Asn Gly Leu Lys Thr Tyr Val Ser Lys Trp Ile
 225 230 235 240

Ser Gln Gly Val Pro Ile Asp Gly Ile Gly Ser Gln Ser His Leu Ser
 245 250 255

Gly Gly Gly Gly Ser Gly Thr Leu Gly Ala Leu Gln Gln Leu Ala Thr
 260 265 270

Val Pro Val Thr Glu Leu Ala Ile Thr Glu Leu Asp Ile Gln Gly Ala
 275 280 285

Pro Thr Thr Asp Tyr Thr Gln Val Val Gln Ala Cys Leu Ser Val Ser
 290 295 300

Lys Cys Val Gly Ile Thr Val Trp Gly Ile Ser Asp Lys Asp Ser Trp
 305 310 315 320

Arg Ala Ser Thr Asn Pro Leu Leu Phe Asp Ala Asn Phe Asn Pro Lys
 325 330 335

Pro Ala Tyr Asn Ser Ile Val Gly Ile Leu Gln
 340 345

<210> SEQ ID NO 39
 <211> LENGTH: 2564
 <212> TYPE: DNA
 <213> ORGANISM: Trichoderma reesei

<400> SEQUENCE: 39

ggacagccgg acgcaatggt gaataacgca gctcttctcg ccgcctgtc ggctctctg 60
 cccacggccc tggcgagaaa caatcaaaca tacgccaact actctgctca gggccagcct 120
 gatctctacc ccgagacact tgccacgctc aactctctgt tccccgactg cgaacatggc 180
 cccctcaaga acaatctcgt ctgtgactca tcggccggt atgtagagcg agcccaggcc 240
 ctcatctcgc tcttcaccct cgaggagctc attctcaaca cgcaaaactc gggccccggc 300
 gtgcctcgcc tgggtcttcc gaactaccaa gtctggaatg aggctctgca cggcttgac 360
 cgcgccaact tcgccaccaa gggcggccag ttcgaatggg cgacctcgtt ccccatgccc 420
 atcctcacta cggcggccct caaccgcaca ttgatccacc agattgccga catcatctcg 480
 acccaagctc gagcattcag caacagcggc cgttacggtc tcgacgtcta tgcgccaaac 540
 gtcaatggct tccgaagccc cctctggggc cgtggccagg agacgcccgg cgaagacgcc 600
 ttttctca gctccgcta tacttacgag tacatcacgg gcatccaggg tggcgtcgac 660
 cctgagcacc tcaaggttgc cgccacgggt aagcactttg ccggatacga cctcgagaac 720
 tggaaacaacc agtcccgtct cggtttcgac gccatcataa ctacgagga cctctccgaa 780
 tactaacctc cccagttcct cgctgcccgc cgttatgcaa agtcacgcag cttgatgtgc 840

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gcatacaact ccgtaacg cgtgcccagc tgtgccaaca gcttcttctt gcagacgctt 900
ttgcgcgaga gctggggctt ccccgaatgg ggatacgtct cgtccgattg cgatgccgtc 960
tacaacgttt tcaaccctca tgactacgcc agcaaccagt cgtcagccgc cgccagctca 1020
ctgcgagccg gcaccgatat cgactgcggt cagacttacc cgtggcacct caacgagctc 1080
tttgtggccg gcgaagtctc ccgcccggag atcgagcggg ccgtaacccg tctgtacgcc 1140
aacctcgtcc gtctcggata cttcgacaag aagaaccagt accgctcgtc cggttggaag 1200
gatgtcgtca agactgatgc ctggaacatc tcgtacgagg ctgctgttga gggcatcgtc 1260
ctgctcaaga acgatggcac tctccctctg tccaagaagg tgcgcagcat tgctctgatc 1320
ggaccatggg ccaatgccac aacccaaatg caaggcaact actatggccc tgccccatac 1380
ctcatcagcc ctctggaagc tgctaagaag gccggctatc acgtcaactt tgaactcggc 1440
acagagatcg ccggcaacag caccactggc ttgccaagg ccattgctgc cgccaagaag 1500
tcggatgcca tcatctacct cgggtgaatt gacaacacca ttgaacagga gggcgctgac 1560
cgcacggaca ttgcttggcc cggtaatcag ctggatctca tcaagcagct cagcgaggtc 1620
ggcaaacccc ttgtcgtcct gcaaatgggc ggtggtcagg tagactcatc ctgctcaag 1680
agcaacaaga aggtcaactc cctcgtctgg ggcggatata ccggccagtc gggaggcgtt 1740
gccctcttcg acattctctc tggcaagcgt gctcctgccg gccgactggt caccactcag 1800
taccggctg agtatgttca ccaattcccc cagaatgaca tgaacctccg acccgatgga 1860
aagtcaaacc ctggacagac ttacatctgg tacaccggca aaccgcteta cgagtttggc 1920
agtgtctct tctacaccac cttcaaggag actctcgcca gccaccccaa gagcctcaag 1980
ttcaaacct catcgatcct ctctgctcct caccocggat acacttacag cgagcagatt 2040
cccgtcttca ccttcgagcc caacatcaag aactcgggca agacggagtc cccatatacg 2100
gccatgctgt ttgttcgcac aagcaacgct ggcccagccc cgtaccgaa caagtggctc 2160
gtcggattcg accgacttgc cgacatcaag cctggtcact cttccaagct cagcatcccc 2220
atccctgtca gtgctctcgc ccgtgttgat tctcacggaa accggattgt ataccocggc 2280
aagtatgagc tagccttgaa caccgacgag tctgtgaagc ttgagtttga gttggtggga 2340
gaagaggtaa cgattgagaa ctggccggtg gaggagcaac agatcaagga tgctacacct 2400
gacgcataag ggttttaatg atgttggtat gacaaacggg tagagtagtt aatgatggaa 2460
taggaagagg ccatagtttt ctgtttgcaa accatttttg ccattgcaa aaaaaaaaaa 2520
aaaaaaaaaa aaaaaaaaaa aaaaaaaaaa aaaaaaaaaa aaaa 2564

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<210> SEQ ID NO 40

<211> LENGTH: 780

<212> TYPE: PRT

<213> ORGANISM: Trichoderma reesei

<400> SEQUENCE: 40

```

Met Val Asn Asn Ala Ala Leu Leu Ala Ala Leu Ser Ala Leu Leu Pro
1           5           10           15

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Thr Ala Leu Ala Gln Asn Asn Gln Thr Tyr Ala Asn Tyr Ser Ala Gln
20           25           30

```

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Gly Gln Pro Asp Leu Tyr Pro Glu Thr Leu Ala Thr Leu Thr Leu Ser
35           40           45

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Phe Pro Asp Cys Glu His Gly Pro Leu Lys Asn Asn Leu Val Cys Asp
50           55           60

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Ser Ser Ala Gly Tyr Val Glu Arg Ala Gln Ala Leu Ile Ser Leu Phe

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65	70	75	80
Thr Leu Glu Glu Leu Ile Leu Asn Thr Gln Asn Ser Gly Pro Gly Val 85 90 95			
Pro Arg Leu Gly Leu Pro Asn Tyr Gln Val Trp Asn Glu Ala Leu His 100 105 110			
Gly Leu Asp Arg Ala Asn Phe Ala Thr Lys Gly Gly Gln Phe Glu Trp 115 120 125			
Ala Thr Ser Phe Pro Met Pro Ile Leu Thr Thr Ala Ala Leu Asn Arg 130 135 140			
Thr Leu Ile His Gln Ile Ala Asp Ile Ile Ser Thr Gln Ala Arg Ala 145 150 155 160			
Phe Ser Asn Ser Gly Arg Tyr Gly Leu Asp Val Tyr Ala Pro Asn Val 165 170 175			
Asn Gly Phe Arg Ser Pro Leu Trp Gly Arg Gly Gln Glu Thr Pro Gly 180 185 190			
Glu Asp Ala Phe Phe Leu Ser Ser Ala Tyr Thr Tyr Glu Tyr Ile Thr 195 200 205			
Gly Ile Gln Gly Gly Val Asp Pro Glu His Leu Lys Val Ala Ala Thr 210 215 220			
Val Lys His Phe Ala Gly Tyr Asp Leu Glu Asn Trp Asn Asn Gln Ser 225 230 235 240			
Arg Leu Gly Phe Asp Ala Ile Ile Thr Gln Gln Asp Leu Ser Glu Tyr 245 250 255			
Tyr Thr Pro Gln Phe Leu Ala Ala Ala Arg Tyr Ala Lys Ser Arg Ser 260 265 270			
Leu Met Cys Ala Tyr Asn Ser Val Asn Gly Val Pro Ser Cys Ala Asn 275 280 285			
Ser Phe Phe Leu Gln Thr Leu Leu Arg Glu Ser Trp Gly Phe Pro Glu 290 295 300			
Trp Gly Tyr Val Ser Ser Asp Cys Asp Ala Val Tyr Asn Val Phe Asn 305 310 315 320			
Pro His Asp Tyr Ala Ser Asn Gln Ser Ser Ala Ala Ala Ser Ser Leu 325 330 335			
Arg Ala Gly Thr Asp Ile Asp Cys Gly Gln Thr Tyr Pro Trp His Leu 340 345 350			
Asn Glu Ser Phe Val Ala Gly Glu Val Ser Arg Gly Glu Ile Glu Arg 355 360 365			
Ser Val Thr Arg Leu Tyr Ala Asn Leu Val Arg Leu Gly Tyr Phe Asp 370 375 380			
Lys Lys Asn Gln Tyr Arg Ser Leu Gly Trp Lys Asp Val Val Lys Thr 385 390 395 400			
Asp Ala Trp Asn Ile Ser Tyr Glu Ala Ala Val Glu Gly Ile Val Leu 405 410 415			
Leu Lys Asn Asp Gly Thr Leu Pro Leu Ser Lys Lys Val Arg Ser Ile 420 425 430			
Ala Leu Ile Gly Pro Trp Ala Asn Ala Thr Thr Gln Met Gln Gly Asn 435 440 445			
Tyr Tyr Gly Pro Ala Pro Tyr Leu Ile Ser Pro Leu Glu Ala Ala Lys 450 455 460			
Lys Ala Gly Tyr His Val Asn Phe Glu Leu Gly Thr Glu Ile Ala Gly 465 470 475 480			
Asn Ser Thr Thr Gly Phe Ala Lys Ala Ile Ala Ala Ala Lys Lys Ser 485 490 495			

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Asp Ala Ile Ile Tyr Leu Gly Gly Ile Asp Asn Thr Ile Glu Gln Glu
 500 505 510
 Gly Ala Asp Arg Thr Asp Ile Ala Trp Pro Gly Asn Gln Leu Asp Leu
 515 520 525
 Ile Lys Gln Leu Ser Glu Val Gly Lys Pro Leu Val Val Leu Gln Met
 530 535 540
 Gly Gly Gly Gln Val Asp Ser Ser Ser Leu Lys Ser Asn Lys Lys Val
 545 550 555 560
 Asn Ser Leu Val Trp Gly Gly Tyr Pro Gly Gln Ser Gly Gly Val Ala
 565 570 575
 Leu Phe Asp Ile Leu Ser Gly Lys Arg Ala Pro Ala Gly Arg Leu Val
 580 585 590
 Thr Thr Gln Tyr Pro Ala Glu Tyr Val His Gln Phe Pro Gln Asn Asp
 595 600 605
 Met Asn Leu Arg Pro Asp Gly Lys Ser Asn Pro Gly Gln Thr Tyr Ile
 610 615 620
 Trp Tyr Thr Gly Lys Pro Val Tyr Glu Phe Gly Ser Gly Leu Phe Tyr
 625 630 635 640
 Thr Thr Phe Lys Glu Thr Leu Ala Ser His Pro Lys Ser Leu Lys Phe
 645 650 655
 Asn Thr Ser Ser Ile Leu Ser Ala Pro His Pro Gly Tyr Thr Tyr Ser
 660 665 670
 Glu Gln Ile Pro Val Phe Thr Phe Glu Ala Asn Ile Lys Asn Ser Gly
 675 680 685
 Lys Thr Glu Ser Pro Tyr Thr Ala Met Leu Phe Val Arg Thr Ser Asn
 690 695 700
 Ala Gly Pro Ala Pro Tyr Pro Asn Lys Trp Leu Val Gly Phe Asp Arg
 705 710 715 720
 Leu Ala Asp Ile Lys Pro Gly His Ser Ser Lys Leu Ser Ile Pro Ile
 725 730 735
 Pro Val Ser Ala Leu Ala Arg Val Asp Ser His Gly Asn Arg Ile Val
 740 745 750
 Tyr Pro Gly Lys Tyr Glu Leu Ala Leu Asn Thr Asp Glu Ser Val Lys
 755 760 765
 Leu Glu Phe Glu Leu Val Gly Glu Glu Val Thr Ile
 770 775 780

<210> SEQ ID NO 41

<211> LENGTH: 2779

<212> TYPE: DNA

<213> ORGANISM: Trichoderma reesei

<400> SEQUENCE: 41

atgtccgcgt ctagtgattt atatgtagaa tgatcacaat tcatgtaact gcgttttcgc 60
 acatgcaaaa agccctaacg tgagactgag ccacttctta gttttcgtat catgtcagtt 120
 gcaaggttac accacaatgc agctcaacga gacaacgctg ccagcccata ataatggata 180
 gctggttgta gagagattaa gaagagaatg ctgtttcaga aggaagacta tatcatagca 240
 gctgctacat ttccctcttt ccctctttcc atcccttaat agatacgtac ccttgcaatt 300
 ggccgtttcg gaagagcttt tctgcttate ctaaccacct acgccagaat accgggtggaa 360
 taatcagtgc tcaatagggg aacccccaac tgcagcatat aagcctatta acaagacgtc 420
 ccaacatgca ttttccttca gtccgcagca ggctatagag agtaggcaat ttacacacca 480

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cttttagcct	ctgcacatat	ctcaccacat	ttgcattacg	gcatccacta	ttacaaccac	540
ttggcacctg	atggccttgc	tctaccata	tcggttttta	cgttccgctg	tggtcagtcg	600
ttaaatccgt	ggtaggagcag	aacgaccagc	ttctcgtatc	gggaactccg	cttatccgat	660
accctcagtc	gaacccttcg	tgatactcag	cctaaataac	atcgcatcgt	agcagacaac	720
ttcagtaatt	tttgtggggt	aatggtcaga	tcgctcctct	tatatataaa	gcagaggtta	780
gtgggctaag	gaaattcgtg	gttcgcttat	agtagagctg	tcagttgcc	ttcccgaact	840
gttagacggg	atggctggta	agcttatcct	cgtggctcta	gcaagccttg	tatcactctc	900
tattcagcag	aattgcgag	cattattgta	agagtgttga	gcgtgttgag	taccatctgt	960
atcgttgcta	acgtaggctt	ttagtggcca	atgtggagge	ataggggtgt	ccggcaccac	1020
atgttgcggt	gctggcgccc	agtgcagttt	tgtcaatgac	tggtactccc	agtgccttgc	1080
gtcaaccgta	tgagctccga	tccgggccc	caatatcttc	taactccaga	ctgtacaggg	1140
cggaaacccc	ccaaacgga	caacttcctc	tagcttggtt	tcacggacgt	cgtcagcatc	1200
ctcatccgtc	ggctcgtctt	cacccggcgg	caactcacca	actggcagtg	cttccaccta	1260
cacaaccaca	gatacagcta	ccgtggctcc	tcattcgcag	tctccttacc	ccagcattgc	1320
cgcacccagt	tgccgatcgt	ggaccctcgt	ggataatggt	tgctgccc	catattgtgc	1380
taatgatgac	acatccgagt	catgctcagg	ctgcggtacc	tgactacgc	cgccctcggc	1440
ggactgcaaa	tccggaacca	tgtatccaga	ggccatcac	gtatccagca	acgagagctg	1500
gcactacagt	gtaagatgac	caacgctggg	gtatctaate	ctttgtcttc	ctcggcgtgc	1560
tgacctgga	gcatttagag	atcaaccac	tttggcctaa	cgagcggcgg	ggcctgtggc	1620
tttggcctgt	acggtctctg	cacaaagggc	agtgttacag	ccagctggac	ggatcccatg	1680
cttggcgcga	cgtgtgacgc	ttttgtaca	gcgtatcccc	tgctttgcaa	agaccctacc	1740
ggcactaccc	ttcgtggcaa	cttcgcagct	ccaaacggcg	attactacac	ccaagtggg	1800
gaccccgaga	ggcaatcatt	ttctggtgta	gtattcactg	acagtgcgat	agttctggtc	1860
ctcgttgcca	ggagccctcg	ataactacct	gtcctgcggc	gagtgcattg	agctgataca	1920
aacaaagccc	gatgggaccg	attatgctgt	cggagaagcc	ggctacacgg	atccaattac	1980
tctcgagatt	gtggacagct	gcccgtgcag	cgcaactcc	aagtgggtgt	gtcagagagc	2040
cccgtccatc	ccgtccattg	tactacatgc	gccaaaccgaa	tggccctggc	taacatctcg	2100
caggtgggcc	gggcccgat	cattgcccag	agatcgactt	caaatacggc	tgctcctctc	2160
ctgctgacag	cattcatctc	gacctgtcag	acattgccat	gggcccgttg	cagggcaatg	2220
gatcactaac	caatggcgtc	atcccgactc	gatatagaag	agtccaatgc	cccaaagttg	2280
ggaacgccta	catttggtct	cgaaatggcg	gagggcctta	ctatcttctc	ctcacggcag	2340
tcaacaccaa	cggaccgggc	tcagtcacca	aaatcgagat	caagggcgca	gacaccgaca	2400
actgggttgc	cttgggtccat	gacccaaact	atacagtag	ccgcccacaa	gaacgctatg	2460
gcagttgggt	aatcccacag	ggatcagggc	cctttaactt	gcctgttgga	attcgtctga	2520
ctagcccaac	gggggaacag	attgtgaatg	aacaggccat	caagacattc	actcctccgg	2580
ccacaggtga	ccccaatctt	tactacattg	acattgggtg	gcagtttagc	cagaattgat	2640
ggcaagcatt	gggcaatggg	cttcttgcgt	tgggacaatg	atgtaggcta	gattctcaat	2700
gcttcaagta	tgtgggtgac	gtcttcgtgt	gtatagatag	gtatgctgtt	cacttaaata	2760
cacatccttt	ggtacgttg					2779

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<210> SEQ ID NO 42
<211> LENGTH: 493
<212> TYPE: PRT
<213> ORGANISM: Trichoderma reesei

<400> SEQUENCE: 42

Met Ala Gly Lys Leu Ile Leu Val Ala Leu Ala Ser Leu Val Ser Leu
1          5          10          15

Ser Ile Gln Gln Asn Cys Ala Ala Leu Phe Gly Gln Cys Gly Gly Ile
20          25          30

Gly Trp Ser Gly Thr Thr Cys Cys Val Ala Gly Ala Gln Cys Ser Phe
35          40          45

Val Asn Asp Trp Tyr Ser Gln Cys Leu Ala Ser Thr Gly Gly Asn Pro
50          55          60

Pro Asn Gly Thr Thr Ser Ser Ser Leu Val Ser Arg Thr Ser Ser Ala
65          70          75          80

Ser Ser Ser Val Gly Ser Ser Ser Pro Gly Gly Asn Ser Pro Thr Gly
85          90          95

Ser Ala Ser Thr Tyr Thr Thr Thr Asp Thr Ala Thr Val Ala Pro His
100         105         110

Ser Gln Ser Pro Tyr Pro Ser Ile Ala Ala Ser Ser Cys Gly Ser Trp
115         120         125

Thr Leu Val Asp Asn Val Cys Cys Pro Ser Tyr Cys Ala Asn Asp Asp
130         135         140

Thr Ser Glu Ser Cys Ser Gly Cys Gly Thr Cys Thr Thr Pro Pro Ser
145         150         155         160

Ala Asp Cys Lys Ser Gly Thr Met Tyr Pro Glu Val His His Val Ser
165         170         175

Ser Asn Glu Ser Trp His Tyr Ser Arg Ser Thr His Phe Gly Leu Thr
180         185         190

Ser Gly Gly Ala Cys Gly Phe Gly Leu Tyr Gly Leu Cys Thr Lys Gly
195         200         205

Ser Val Thr Ala Ser Trp Thr Asp Pro Met Leu Gly Ala Thr Cys Asp
210         215         220

Ala Phe Cys Thr Ala Tyr Pro Leu Leu Cys Lys Asp Pro Thr Gly Thr
225         230         235         240

Thr Leu Arg Gly Asn Phe Ala Ala Pro Asn Gly Asp Tyr Tyr Thr Gln
245         250         255

Phe Trp Ser Ser Leu Pro Gly Ala Leu Asp Asn Tyr Leu Ser Cys Gly
260         265         270

Glu Cys Ile Glu Leu Ile Gln Thr Lys Pro Asp Gly Thr Asp Tyr Ala
275         280         285

Val Gly Glu Ala Gly Tyr Thr Asp Pro Ile Thr Leu Glu Ile Val Asp
290         295         300

Ser Cys Pro Cys Ser Ala Asn Ser Lys Trp Cys Cys Gly Pro Gly Ala
305         310         315         320

Asp His Cys Gly Glu Ile Asp Phe Lys Tyr Gly Cys Pro Leu Pro Ala
325         330         335

Asp Ser Ile His Leu Asp Leu Ser Asp Ile Ala Met Gly Arg Leu Gln
340         345         350

Gly Asn Gly Ser Leu Thr Asn Gly Val Ile Pro Thr Arg Tyr Arg Arg
355         360         365

Val Gln Cys Pro Lys Val Gly Asn Ala Tyr Ile Trp Leu Arg Asn Gly
370         375         380

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Gly Gly Pro Tyr Tyr Phe Ala Leu Thr Ala Val Asn Thr Asn Gly Pro
 385 390 395 400

Gly Ser Val Thr Lys Ile Glu Ile Lys Gly Ala Asp Thr Asp Asn Trp
 405 410 415

Val Ala Leu Val His Asp Pro Asn Tyr Thr Ser Ser Arg Pro Gln Glu
 420 425 430

Arg Tyr Gly Ser Trp Val Ile Pro Gln Gly Ser Gly Pro Phe Asn Leu
 435 440 445

Pro Val Gly Ile Arg Leu Thr Ser Pro Thr Gly Glu Gln Ile Val Asn
 450 455 460

Glu Gln Ala Ile Lys Thr Phe Thr Pro Pro Ala Thr Gly Asp Pro Asn
 465 470 475 480

Phe Tyr Tyr Ile Asp Ile Gly Val Gln Phe Ser Gln Asn
 485 490

<210> SEQ ID NO 43
 <211> LENGTH: 40
 <212> TYPE: DNA
 <213> ORGANISM: Aspergillus fumigatus

<400> SEQUENCE: 43

cgcgactgc gcaccatgct ggccctcacc ttctcctacc 40

<210> SEQ ID NO 44
 <211> LENGTH: 48
 <212> TYPE: DNA
 <213> ORGANISM: Aspergillus fumigatus

<400> SEQUENCE: 44

ctttcgccac ggagcttaat taactacagg cactgagagt aataatca 48

<210> SEQ ID NO 45
 <211> LENGTH: 1131
 <212> TYPE: DNA
 <213> ORGANISM: Herpes simplex virus

<400> SEQUENCE: 45

atggcttcgt acccggcca tcaacacgcg tctgcgttcg accaggctgc gcggttctcgc 60

ggccatagca accgacgtac ggcggttgcgc cctcgccggc agcaagaagc cacggaagtc 120

cgcccggagc agaaaatgcc cacgctactg cgggtttata tagacggtcc ccacgggatg 180

gggaaaacca ccaccacgca actgctggtg gccttgggtt cgcgcgacga tatcgtctac 240

gtaccggagc cgatgactta ctggcgggtg ctgggggctt ccgagacaat cgccaacatc 300

tacaccacac aacaccgct cgaccagggt gagatatcgg ccggggacgc ggcgggtgta 360

atgacaagcg cccagataac aatgggcatg ccttatgccc tgaccgacgc cgttctggct 420

cctcatatcg ggggggaggc tgggagctca catgccccgc ccccgccct caccctcatc 480

ttcgaccgce atcccatcgc cgccctcctg tgctaccggc ccgcgcggtta ccttatgggc 540

agcatgacc cccaggccgt gctggcgctt gtggccctca tcccggcgac cttgcccggc 600

accaacatcg tgcttggggc ccttccggag gacagacaca tcgaccgctt ggccaaacgc 660

cagcgcctcg gcgagcggct ggacctggct atgctggctg cgattcgccg cgtttacggg 720

ctacttgcca atacggtgcg gtatctgcag tgcggcgggt cgtggcggtga ggactgggga 780

cagctttcgg ggacggccgt gccgccccag ggtgccgagc cccagagcaa cgcgggcccc 840

cgaccccata tcggggacac gttatctacc ctgtttcggg gccccgagtt gctggcccc 900

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aacggcgacc tgtataacgt gtttgctggt gccttgagcg tcttgccaa acgctccgt 960
tccatgcacg tctttatcct ggattacgac caatcgcccg ccggctgccg ggacgccctg 1020
ctgcaactta cctccgggat ggtccagacc cacgtcacca cccccggctc cataccgacg 1080
atatgcgacc tggcgcgcac gtttgcccgg gagatggggg aggctaactg a 1131

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<210> SEQ ID NO 46

<211> LENGTH: 376

<212> TYPE: PRT

<213> ORGANISM: Herpes simplex virus

<400> SEQUENCE: 46

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Met Ala Ser Tyr Pro Gly His Gln His Ala Ser Ala Phe Asp Gln Ala
1          5          10          15
Ala Arg Ser Arg Gly His Ser Asn Arg Arg Thr Ala Leu Arg Pro Arg
20          25          30
Arg Gln Gln Glu Ala Thr Glu Val Arg Pro Glu Gln Lys Met Pro Thr
35          40          45
Leu Leu Arg Val Tyr Ile Asp Gly Pro His Gly Met Gly Lys Thr Thr
50          55          60
Thr Thr Gln Leu Leu Val Ala Leu Gly Ser Arg Asp Asp Ile Val Tyr
65          70          75          80
Val Pro Glu Pro Met Thr Tyr Trp Arg Val Leu Gly Ala Ser Glu Thr
85          90          95
Ile Ala Asn Ile Tyr Thr Thr Gln His Arg Leu Asp Gln Gly Glu Ile
100         105         110
Ser Ala Gly Asp Ala Ala Val Val Met Thr Ser Ala Gln Ile Thr Met
115         120         125
Gly Met Pro Tyr Ala Val Thr Asp Ala Val Leu Ala Pro His Ile Gly
130         135         140
Gly Glu Ala Gly Ser Ser His Ala Pro Pro Pro Ala Leu Thr Leu Ile
145         150         155         160
Phe Asp Arg His Pro Ile Ala Ala Leu Leu Cys Tyr Pro Ala Ala Arg
165         170         175
Tyr Leu Met Gly Ser Met Thr Pro Gln Ala Val Leu Ala Phe Val Ala
180         185         190
Leu Ile Pro Pro Thr Leu Pro Gly Thr Asn Ile Val Leu Gly Ala Leu
195         200         205
Pro Glu Asp Arg His Ile Asp Arg Leu Ala Lys Arg Gln Arg Pro Gly
210         215         220
Glu Arg Leu Asp Leu Ala Met Leu Ala Ala Ile Arg Arg Val Tyr Gly
225         230         235         240
Leu Leu Ala Asn Thr Val Arg Tyr Leu Gln Cys Gly Gly Ser Trp Arg
245         250         255
Glu Asp Trp Gly Gln Leu Ser Gly Thr Ala Val Pro Pro Gln Gly Ala
260         265         270
Glu Pro Gln Ser Asn Ala Gly Pro Arg Pro His Ile Gly Asp Thr Leu
275         280         285
Phe Thr Leu Phe Arg Gly Pro Glu Leu Leu Ala Pro Asn Gly Asp Leu
290         295         300
Tyr Asn Val Phe Ala Trp Ala Leu Asp Val Leu Ala Lys Arg Leu Arg
305         310         315         320
Ser Met His Val Phe Ile Leu Asp Tyr Asp Gln Ser Pro Ala Gly Cys
325         330         335

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Arg Asp Ala Leu Leu Gln Leu Thr Ser Gly Met Val Gln Thr His Val
 340 345 350

Thr Thr Pro Gly Ser Ile Pro Thr Ile Cys Asp Leu Ala Arg Thr Phe
 355 360 365

Ala Arg Glu Met Gly Glu Ala Asn
 370 375

<210> SEQ ID NO 47

<211> LENGTH: 43

<212> TYPE: DNA

<213> ORGANISM: Trichoderma reesei

<400> SEQUENCE: 47

ttagactgcg gccgcgtggc gaaagcctga cgcaccggta gat 43

<210> SEQ ID NO 48

<211> LENGTH: 41

<212> TYPE: DNA

<213> ORGANISM: Trichoderma reesei

<400> SEQUENCE: 48

agtagttagc ggccgcacgg cacggttaag cagggctctt c 41

<210> SEQ ID NO 49

<211> LENGTH: 25

<212> TYPE: DNA

<213> ORGANISM: Trichoderma reesei

<400> SEQUENCE: 49

aaaaa caaaa catcccgttc ataac 25

<210> SEQ ID NO 50

<211> LENGTH: 25

<212> TYPE: DNA

<213> ORGANISM: Trichoderma reesei

<400> SEQUENCE: 50

aacaaggttt accggtttcg aaaag 25

<210> SEQ ID NO 51

<211> LENGTH: 54

<212> TYPE: DNA

<213> ORGANISM: Trichoderma reesei

<400> SEQUENCE: 51

acgaattggt taaacgtcga cccaagtatc cagagggtgta tggaaatc agat 54

<210> SEQ ID NO 52

<211> LENGTH: 44

<212> TYPE: DNA

<213> ORGANISM: Trichoderma reesei

<400> SEQUENCE: 52

cgcgtagatc tgcggccatg gtgcaatata cagagggtga tctt 44

<210> SEQ ID NO 53

<211> LENGTH: 48

<212> TYPE: DNA

<213> ORGANISM: Trichoderma reesei

<400> SEQUENCE: 53

atctacgcgt actagttaat taaggctttc gtgaccgggc ttcaaaca 48

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<210> SEQ ID NO 54
 <211> LENGTH: 46
 <212> TYPE: DNA
 <213> ORGANISM: *Trichoderma reesei*

 <400> SEQUENCE: 54

 gcggccgcta ctagtggatc cactcggagt tgttatacgc tactcg 46

<210> SEQ ID NO 55
 <211> LENGTH: 46
 <212> TYPE: DNA
 <213> ORGANISM: *Trichoderma reesei*

 <400> SEQUENCE: 55

 atccatcaca ctggcggccg cgcttcaaac aatgatgtgc gatggt 46

<210> SEQ ID NO 56
 <211> LENGTH: 46
 <212> TYPE: DNA
 <213> ORGANISM: *Trichoderma reesei*

 <400> SEQUENCE: 56

 gatgcatgct cgagcggccg cctaccttgg cagccctacg agagag 46

<210> SEQ ID NO 57
 <211> LENGTH: 41
 <212> TYPE: DNA
 <213> ORGANISM: *Aspergillus fumigatus*

 <400> SEQUENCE: 57

 ctctgtgtat tgcacatga agcaccttgc atcttccatc g 41

<210> SEQ ID NO 58
 <211> LENGTH: 40
 <212> TYPE: DNA
 <213> ORGANISM: *Aspergillus fumigatus*

 <400> SEQUENCE: 58

 ccggtcacga aagccttaat taaaaggacg ggtagcgtt 40

<210> SEQ ID NO 59
 <211> LENGTH: 25
 <212> TYPE: DNA
 <213> ORGANISM: *Aspergillus fumigatus*

 <400> SEQUENCE: 59

 agccacatgc cgcatttga caaag 25

<210> SEQ ID NO 60
 <211> LENGTH: 25
 <212> TYPE: DNA
 <213> ORGANISM: *Aspergillus fumigatus*

 <400> SEQUENCE: 60

 agggattcag tgtgctacag gctgc 25

<210> SEQ ID NO 61
 <211> LENGTH: 25
 <212> TYPE: DNA
 <213> ORGANISM: *Aspergillus fumigatus*

 <400> SEQUENCE: 61

 aaaaaacaaa catcccgttc ataac 25

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<210> SEQ ID NO 62
 <211> LENGTH: 25
 <212> TYPE: DNA
 <213> ORGANISM: *Aspergillus fumigatus*

 <400> SEQUENCE: 62
 aacaaggttt accggtttcg aaaag 25

<210> SEQ ID NO 63
 <211> LENGTH: 27
 <212> TYPE: DNA
 <213> ORGANISM: *Aspergillus fumigatus*

 <400> SEQUENCE: 63
 gttaagcata caattgaacg agaatgg 27

<210> SEQ ID NO 64
 <211> LENGTH: 25
 <212> TYPE: DNA
 <213> ORGANISM: *Aspergillus fumigatus*

 <400> SEQUENCE: 64
 gatgatataa tggagcaaat aaggg 25

<210> SEQ ID NO 65
 <211> LENGTH: 2077
 <212> TYPE: DNA
 <213> ORGANISM: *Trichoderma reesei*

 <400> SEQUENCE: 65
 atgacggatg cacaaaagaa ttggaggaga gacgaaaacg acgaggacga tgaagcagag 60
 caggagctcg atgaggctgt aagtcgccgc gactcgcac tggctcgaca agcgtcgtct 120
 gacactcttt tctccatct agagcctcaa ggcgcagaaa gatgcaattc ttctagccat 180
 tgaagtcagt ccgtcgatgc ttgagcctcc gccagtctcc agctctagga aagctgatcg 240
 ggacagcccc gttcaagctg cgctgaaatg cgccccccac ctgatggagc agcgcacatc 300
 ctccaacccc aaagacatga tgggaatcct cctctttggg acagaaaaga ccaagttccg 360
 ggacgacaat ggccgcagtg ggctcgggta tccgaattgc tacctcttta tggacctcga 420
 cattccggca gctgaagacg tcaaagcgtt gaaggcgtg accgaggacg aagacgaaga 480
 cgaagtgctg aagcccgccca ccaccgacac agtttccatg tccaacgtgt tgttttgccg 540
 caaccagata ttcaccacaa agggggccaa ctttggcagc cggcgacttt tcattgtgac 600
 ggacaatgac gatccgcacg cgctcggaca ggccggcagg tctgctgccg ctgttcgggc 660
 aaaggacttg tacgatctgg gcatcacgat cgacttgttt ccaatcacca caggagactc 720
 caagtttgat ctacgcaaat ttacgatgt aagctatatt tcttcgtttc ttcgctctaa 780
 aatcaccac cctccgtcgt gacatagact gacaaggaac taggatattg tctatcgca 840
 cccgaatgcc gaggccaatc gcaccgaagt gcgagcctca aaatcgggag atggactgtc 900
 tcttctcaac tcgctcattt caaacatcaa ttccaagcag acgccaagc gagcattggt 960
 ccatctgcca tttgagattg cacctggact caagatcact gtcaagggtt acaacattgt 1020
 gcatcggcaa acgcccggca gaacgtgcta catctggctg gaaggggaga aggctcagat 1080
 tgcaacaggg gaaacgacgc gagttgcaga ggattctgcc agaacagtcg aaaagcaaga 1140
 gataaaaaag gcctacaagt ttggtggcga atacgtatac tttacgcccg aggagcagaa 1200

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gaagctccgg gattttggcg cgcccacgat ccggatcatt ggattcaaga agcgcagcat 1260
gattcccgtc tgggccagcg tcaagaagtc gacctttatc tttcccagcg aagaggatta 1320
catcggatcg acacgcgtct tttcagccct atggcagaag cttctaaagg atgacaagat 1380
cggcctcgct tgggtcgtgc ttcgatctaa cgcgcagccc atgtttgccg ctctgattcc 1440
atcaagagag cagtccgaag acgacgcggg gacaccatat ctaccagctg gcctgtggct 1500
gtatcctctc cctacggctg acgacctgcg agatataaat gtcgaacgaa agctcgactg 1560
ctcggaggac ctaaaaacca aatgagagt cattgtacaa cagctcaatc tccccaggg 1620
catatataac cactcaagt acccgaacc ggctctgcaa tggcactaca agatcctcca 1680
gaccctcgcc ttggaggagg agatgccgga agaaccgaa gacttgacgg agcccaaaaa 1740
caaggcgata agcaaacgcg tcggagggta cttggaggag tggccgaga ctctgaaaga 1800
cgaggcggac agggccactc gatccaggtc cttgaagcga gagattgaag atgatgcccc 1860
ggagcgcgcc gcaaagcaga gaaaggtagc tggagagcgg cccagcggat cgaatcttag 1920
catggcgcag cttagggatg ccattgagag cgggagcatc tcgaagatga cagtggcaca 1980
gctgaaggat gtcgctggcg ccagaggact cagcacgggt ggtaagaagg ctgatttgct 2040
ggagcggata gagcagtggg ttgaggagaa cagctga 2077

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<210> SEQ ID NO 66

<211> LENGTH: 649

<212> TYPE: PRT

<213> ORGANISM: Trichoderma reesei

<400> SEQUENCE: 66

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Met Thr Asp Ala Gln Lys Asn Trp Arg Arg Asp Glu Asn Asp Glu Asp
1          5          10          15
Asp Glu Ala Glu Gln Glu Leu Asp Glu Ala Val Ser Arg Arg Asp Leu
20          25          30
Lys Ala Gln Lys Asp Ala Ile Leu Leu Ala Ile Glu Val Ser Pro Ser
35          40          45
Met Leu Glu Pro Pro Pro Val Ser Ser Ser Arg Lys Ala Asp Arg Asp
50          55          60
Ser Pro Val Gln Ala Ala Leu Lys Cys Ala Arg His Leu Met Glu Gln
65          70          75          80
Arg Ile Ile Ser Asn Pro Lys Asp Met Met Gly Ile Leu Leu Phe Gly
85          90          95
Thr Glu Lys Thr Lys Phe Arg Asp Asp Asn Gly Arg Ser Gly Leu Gly
100         105         110
Tyr Pro Asn Cys Tyr Leu Phe Met Asp Leu Asp Ile Pro Ala Ala Glu
115         120         125
Asp Val Lys Ala Leu Lys Ala Leu Thr Glu Asp Glu Asp Glu Asp Glu
130         135         140
Val Leu Lys Pro Ala Thr Thr Asp Thr Val Ser Met Ser Asn Val Leu
145         150         155         160
Phe Cys Ala Asn Gln Ile Phe Thr Thr Lys Ala Ala Asn Phe Gly Ser
165         170         175
Arg Arg Leu Phe Ile Val Thr Asp Asn Asp Asp Pro His Ala Ser Asp
180         185         190
Lys Ala Ala Arg Ser Ala Ala Ala Val Arg Ala Lys Asp Leu Tyr Asp
195         200         205

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Leu Gly Ile Thr Ile Asp Leu Phe Pro Ile Thr Thr Gly Asp Ser Lys
210 215 220

Phe Asp Leu Ser Lys Phe Tyr Asp Asp Ile Val Tyr Arg Asp Pro Asn
225 230 235 240

Ala Glu Ala Asn Arg Thr Glu Val Arg Ala Ser Lys Ser Gly Asp Gly
245 250 255

Leu Ser Leu Leu Asn Ser Leu Ile Ser Asn Ile Asn Ser Lys Gln Thr
260 265 270

Pro Lys Arg Ala Leu Phe His Leu Pro Phe Glu Ile Ala Pro Gly Leu
275 280 285

Lys Ile Thr Val Lys Gly Tyr Asn Ile Val His Arg Gln Thr Pro Ala
290 295 300

Arg Thr Cys Tyr Ile Trp Leu Glu Gly Glu Lys Ala Gln Ile Ala Thr
305 310 315 320

Gly Glu Thr Thr Arg Val Ala Glu Asp Ser Ala Arg Thr Val Glu Lys
325 330 335

Gln Glu Ile Lys Lys Ala Tyr Lys Phe Gly Gly Glu Tyr Val Tyr Phe
340 345 350

Thr Pro Glu Glu Gln Lys Lys Leu Arg Asp Phe Gly Ala Pro Thr Ile
355 360 365

Arg Ile Ile Gly Phe Lys Lys Arg Ser Met Ile Pro Val Trp Ala Ser
370 375 380

Val Lys Lys Ser Thr Phe Ile Phe Pro Ser Glu Glu Asp Tyr Ile Gly
385 390 395 400

Ser Thr Arg Val Phe Ser Ala Leu Trp Gln Lys Leu Leu Lys Asp Asp
405 410 415

Lys Ile Gly Leu Ala Trp Cys Val Leu Arg Ser Asn Ala Gln Pro Met
420 425 430

Phe Ala Ala Leu Ile Pro Ser Arg Glu Gln Ser Glu Asp Asp Ala Gly
435 440 445

Thr Pro Tyr Leu Pro Ala Gly Leu Trp Leu Tyr Pro Leu Pro Thr Ala
450 455 460

Asp Asp Leu Arg Asp Ile Asn Val Glu Arg Lys Leu Asp Cys Ser Glu
465 470 475 480

Asp Leu Lys Thr Lys Met Arg Val Ile Val Gln Gln Leu Asn Leu Pro
485 490 495

Lys Gly Ile Tyr Asn Pro Leu Lys Tyr Pro Asn Pro Ala Leu Gln Trp
500 505 510

His Tyr Lys Ile Leu Gln Thr Leu Ala Leu Glu Glu Glu Met Pro Glu
515 520 525

Glu Pro Glu Asp Leu Thr Glu Pro Lys Asn Lys Ala Ile Ser Lys Arg
530 535 540

Val Gly Gly Tyr Leu Glu Glu Trp Ser Glu Thr Leu Lys Asp Glu Ala
545 550 555 560

Asp Arg Ala Thr Arg Ser Arg Ser Leu Lys Arg Glu Ile Glu Asp Asp
565 570 575

Ala Pro Glu Arg Pro Ala Lys Gln Arg Lys Val Ala Gly Glu Arg Pro
580 585 590

Ser Gly Ser Asn Leu Ser Met Ala Gln Leu Arg Asp Ala Ile Glu Ser
595 600 605

Gly Ser Ile Ser Lys Met Thr Val Ala Gln Leu Lys Asp Val Ala Gly
610 615 620

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Ala Arg Gly Leu Ser Thr Gly Gly Lys Lys Ala Asp Leu Leu Glu Arg
 625 630 635 640

Ile Glu Gln Trp Val Glu Glu Asn Ser
 645

<210> SEQ ID NO 67
 <211> LENGTH: 55
 <212> TYPE: DNA
 <213> ORGANISM: Trichoderma reesei

<400> SEQUENCE: 67

gtgtgcgggcc gctcgagcat gcatgtttaa acagcttggc actggccgtc gtttt 55

<210> SEQ ID NO 68
 <211> LENGTH: 55
 <212> TYPE: DNA
 <213> ORGANISM: Trichoderma reesei

<400> SEQUENCE: 68

atcagccccg agacggcgcc gcgtttaaac aattcgtaat catggtcata gctgt 55

<210> SEQ ID NO 69
 <211> LENGTH: 55
 <212> TYPE: DNA
 <213> ORGANISM: Trichoderma reesei

<400> SEQUENCE: 69

catgattacg aattgtttaa acgcgggcgcc gtctcggggc tgatcttgtc gagga 55

<210> SEQ ID NO 70
 <211> LENGTH: 55
 <212> TYPE: DNA
 <213> ORGANISM: Trichoderma reesei

<400> SEQUENCE: 70

ggcgccggtt actagtggat ccagcccttg acagtgatct tgagtccagg tgcaa 55

<210> SEQ ID NO 71
 <211> LENGTH: 58
 <212> TYPE: DNA
 <213> ORGANISM: Trichoderma reesei

<400> SEQUENCE: 71

tgcatatc catcacactg gcgcccgag tttccatgtc caacgtgttg ttttgcgc 58

<210> SEQ ID NO 72
 <211> LENGTH: 58
 <212> TYPE: DNA
 <213> ORGANISM: Trichoderma reesei

<400> SEQUENCE: 72

gccagtgcc aagtgtttaa acatgcatgc tcgagcggcc gcacacgccc tctcctcg 58

<210> SEQ ID NO 73
 <211> LENGTH: 20
 <212> TYPE: DNA
 <213> ORGANISM: Trichoderma reesei

<400> SEQUENCE: 73

caatgacgat ccgcacgct 20

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<210> SEQ ID NO 74
 <211> LENGTH: 20
 <212> TYPE: DNA
 <213> ORGANISM: *Trichoderma reesei*

 <400> SEQUENCE: 74

 caatgacgat cgcacgcgt 20

<210> SEQ ID NO 75
 <211> LENGTH: 20
 <212> TYPE: DNA
 <213> ORGANISM: *Trichoderma reesei*

 <400> SEQUENCE: 75

 gacactcttt tctcccatct 20

<210> SEQ ID NO 76
 <211> LENGTH: 20
 <212> TYPE: DNA
 <213> ORGANISM: *Trichoderma reesei*

 <400> SEQUENCE: 76

 gaggagcaga agaagctccg 20

<210> SEQ ID NO 77
 <211> LENGTH: 22
 <212> TYPE: DNA
 <213> ORGANISM: *Trichoderma reesei*

 <400> SEQUENCE: 77

 gcatatataa cccactcaag ta 22

<210> SEQ ID NO 78
 <211> LENGTH: 22
 <212> TYPE: DNA
 <213> ORGANISM: *Trichoderma reesei*

 <400> SEQUENCE: 78

 attatcttgg accggccgca gg 22

<210> SEQ ID NO 79
 <211> LENGTH: 37
 <212> TYPE: DNA
 <213> ORGANISM: *Aspergillus fumigatus*

 <400> SEQUENCE: 79

 actggattta ccatgacttt gtccaagatc acttcca 37

<210> SEQ ID NO 80
 <211> LENGTH: 40
 <212> TYPE: DNA
 <213> ORGANISM: *Aspergillus fumigatus*

 <400> SEQUENCE: 80

 tcacctctag ttaattaagc gttgaacagt gcaggaccag 40

<210> SEQ ID NO 81
 <211> LENGTH: 17
 <212> TYPE: DNA
 <213> ORGANISM: *Aspergillus fumigatus*

 <400> SEQUENCE: 81

 tgtcccttgt cgatgcg 17

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<210> SEQ ID NO 82
 <211> LENGTH: 17
 <212> TYPE: DNA
 <213> ORGANISM: *Aspergillus fumigatus*

<400> SEQUENCE: 82

cacatgactt ggcttcc 17

<210> SEQ ID NO 83
 <211> LENGTH: 37
 <212> TYPE: DNA
 <213> ORGANISM: *Aspergillus fumigatus*

<400> SEQUENCE: 83

ggactgcgca ccatgacttt gtccaagatc acttcca 37

<210> SEQ ID NO 84
 <211> LENGTH: 38
 <212> TYPE: DNA
 <213> ORGANISM: *Aspergillus fumigatus*

<400> SEQUENCE: 84

gccacggagc ttaattaatt aagcgttgaa cagtgcag 38

<210> SEQ ID NO 85
 <211> LENGTH: 37
 <212> TYPE: DNA
 <213> ORGANISM: *Aspergillus fumigatus*

<400> SEQUENCE: 85

cgcggtagtg gcgcggtcga ccgaatgtag gattggt 37

<210> SEQ ID NO 86
 <211> LENGTH: 36
 <212> TYPE: DNA
 <213> ORGANISM: *Aspergillus fumigatus*

<400> SEQUENCE: 86

ttaccaattg gcgcgccact accgcgttcg agaaga 36

What is claimed is:

1. An enzyme composition comprising a recovered fermentation broth of a recombinant *Trichoderma* host cell, wherein the recombinant *Trichoderma* host cell comprises polynucleotides encoding: (i) an *Aspergillus fumigatus* cellobiohydrolase I; (ii) an *Aspergillus fumigatus* cellobiohydrolase II; (iii) an *Aspergillus fumigatus* beta-glucosidase; (iv) an *Aspergillus fumigatus* GH61 polypeptide having cellulolytic enhancing activity; (v) an *Aspergillus fumigatus* xylanase; and (vi) an *Aspergillus fumigatus* beta-xylosidase; or homologs thereof;

wherein the *Aspergillus fumigatus* cellobiohydrolase I or homolog thereof is selected from the group consisting of:

- (i) a cellobiohydrolase I comprising the mature polypeptide of SEQ ID NO: 2;
- (ii) a cellobiohydrolase I comprising an amino acid sequence having at least 95% sequence identity to the mature polypeptide of SEQ ID NO: 2;
- (iii) a cellobiohydrolase I encoded by a polynucleotide comprising a nucleotide sequence having at least 95% sequence identity to the mature polypeptide coding sequence of SEQ ID NO: 1; and
- (iv) a cellobiohydrolase I encoded by a polynucleotide that hybridizes under very high stringency conditions

with the mature polypeptide coding sequence of SEQ ID NO: 1 or the full-length complement thereof; wherein the *Aspergillus fumigatus* cellobiohydrolase II or homolog thereof is selected from the group consisting of:

- (i) a cellobiohydrolase II comprising the mature polypeptide of SEQ ID NO: 4;
- (ii) a cellobiohydrolase II comprising an amino acid sequence having at least 95% sequence identity to the mature polypeptide of SEQ ID NO: 4;
- (iii) a cellobiohydrolase II encoded by a polynucleotide comprising a nucleotide sequence having at least 95% sequence identity to the mature polypeptide coding sequence of SEQ ID NO: 3; and
- (iv) a cellobiohydrolase II encoded by a polynucleotide that hybridizes under very high stringency conditions with the mature polypeptide coding sequence of SEQ ID NO: 3 or the full-length complement thereof;

wherein the *Aspergillus fumigatus* beta-glucosidase or homolog thereof is selected from the group consisting of:

- (i) a beta-glucosidase comprising the mature polypeptide of SEQ ID NO: 6;

- (ii) a beta-glucosidase comprising an amino acid sequence having at least 95% sequence identity to the mature polypeptide of SEQ ID NO: 6;
- (iii) a beta-glucosidase encoded by a polynucleotide comprising a nucleotide sequence having at least 95% sequence identity to the mature polypeptide coding sequence of SEQ ID NO: 5; and
- (iv) a beta-glucosidase encoded by a polynucleotide that hybridizes under very high stringency conditions with the mature polypeptide coding sequence of SEQ ID NO: 5 or the full-length complement thereof;
- wherein the *Aspergillus fumigatus* GH61 polypeptide having cellulolytic enhancing activity or homolog thereof is selected from the group consisting of:
- (i) a GH61 polypeptide having cellulolytic enhancing activity comprising the mature polypeptide of SEQ ID NO: 8;
- (ii) a GH61 polypeptide having cellulolytic enhancing activity comprising an amino acid sequence having at least 95% sequence identity to the mature polypeptide of SEQ ID NO: 8;
- (iii) a GH61 polypeptide having cellulolytic enhancing activity encoded by a polynucleotide comprising a nucleotide sequence having at least 95% sequence identity to the mature polypeptide coding sequence of SEQ ID NO: 7; and
- (iv) a GH61 polypeptide having cellulolytic enhancing activity encoded by a polynucleotide that hybridizes under very high stringency conditions with the mature polypeptide coding sequence of SEQ ID NO: 7 or the full-length complement thereof;
- wherein the *Aspergillus fumigatus* xylanase or homolog thereof is selected from the group consisting of:
- (i) an *Aspergillus fumigatus* xylanase comprising the mature polypeptide of SEQ ID NO: 14, SEQ ID NO: 16, or SEQ ID NO: 18;
- (ii) a xylanase comprising an amino acid sequence having at least 95% sequence identity to the mature polypeptide of SEQ ID NO: 14, SEQ ID NO: 16, or SEQ ID NO: 18;
- (iii) a xylanase encoded by a polynucleotide comprising a nucleotide sequence having at least 95% sequence identity to the mature polypeptide coding sequence of SEQ ID NO: 13, SEQ ID NO: 15, or SEQ ID NO: 17; and
- (iv) a xylanase encoded by a polynucleotide that hybridizes under very high stringency conditions with the mature polypeptide coding sequence of SEQ ID NO: 13, SEQ ID NO: 15, or SEQ ID NO: 17; or the full-length complement thereof;
- wherein the *Aspergillus fumigatus* beta-xylosidase or homolog thereof is selected from the group consisting of:
- (i) a beta-xylosidase comprising the mature polypeptide of SEQ ID NO: 20;
- (ii) a beta-xylosidase comprising an amino acid sequence having at least 95% sequence identity to the mature polypeptide of SEQ ID NO: 20;
- (iii) a beta-xylosidase encoded by a polynucleotide comprising a nucleotide sequence having at least 95% sequence identity to the mature polypeptide coding sequence of SEQ ID NO: 19; and
- (iv) a beta-xylosidase encoded by a polynucleotide that hybridizes under very high stringency conditions with the mature polypeptide coding sequence of SEQ ID NO: 19 or the full-length complement thereof; and

wherein very high stringency conditions are defined as prehybridization and hybridization at 42° C. in 5×SSPE, 0.3% SDS, 200 micrograms/ml sheared and denatured salmon sperm DNA, and 50% formamide, and washing three times each for 15 minutes using 2×SSC, 0.2% SDS at 70° C.

2. The enzyme composition of claim 1, wherein the recombinant *Trichoderma* host cell further comprises one or more polynucleotides encoding one or more enzymes selected from the group consisting of: (i) an *Aspergillus fumigatus* endoglucanase I; (ii) an *Aspergillus fumigatus* endoglucanase II; and (iii) an *Aspergillus fumigatus* swollenin;

wherein the *Aspergillus fumigatus* endoglucanase I or homolog thereof is selected from the group consisting of:

- (i) an endoglucanase I comprising the mature polypeptide of SEQ ID NO: 10;
- (ii) an endoglucanase I comprising an amino acid sequence having at least 95% sequence identity to the mature polypeptide of SEQ ID NO: 10;
- (iii) an endoglucanase I encoded by a polynucleotide comprising a nucleotide sequence having at least 95% sequence identity to the mature polypeptide coding sequence of SEQ ID NO: 9; and
- (iv) an endoglucanase I encoded by a polynucleotide that hybridizes under very high stringency conditions with the mature polypeptide coding sequence of SEQ ID NO: 9 or the full-length complement thereof;

wherein the *Aspergillus fumigatus* endoglucanase II or homolog thereof is selected from the group consisting of:

- (i) an *Aspergillus fumigatus* endoglucanase II comprising the mature polypeptide of SEQ ID NO: 12;
- (ii) an endoglucanase II comprising an amino acid sequence having at least 95% sequence identity to the mature polypeptide of SEQ ID NO: 12;
- (iii) an endoglucanase II encoded by a polynucleotide comprising a nucleotide sequence having at least 95% sequence identity to the mature polypeptide coding sequence of SEQ ID NO: 11; and
- (iv) an endoglucanase II encoded by a polynucleotide that hybridizes under very high stringency conditions with the mature polypeptide coding sequence of SEQ ID NO: 11 or the full-length complement thereof;

(i) wherein the *Aspergillus fumigatus* swollenin or homolog thereof is selected from the group consisting of:

- (i) a swollenin comprising the mature polypeptide of SEQ ID NO: 22;
- (ii) a swollenin comprising an amino acid sequence having at least 95% sequence identity to the mature polypeptide of SEQ ID NO: 22;
- (iii) a swollenin encoded by a polynucleotide comprising a nucleotide sequence having at least 95% sequence identity to the mature polypeptide coding sequence of SEQ ID NO: 21; and
- (iv) a swollenin encoded by a polynucleotide that hybridizes under very high stringency conditions with the mature polypeptide coding sequence of SEQ ID NO: 21 or the full-length complement thereof; and

wherein very high stringency conditions are defined as prehybridization and hybridization at 42° C. in 5×SSPE, 0.3% SDS, 200 micrograms/ml sheared and denatured salmon sperm DNA, and 50% formamide, and washing three times each for 15 minutes using 2×SSC, 0.2% SDS at 70° C.

191

3. The enzyme composition of claim 1, wherein the recombinant *Trichoderma* host cell is selected from the group consisting of *Trichoderma harzianum*, *Trichoderma koningii*, *Trichoderma longibrachiatum*, *Trichoderma reesei*, and *Trichoderma viride*.

4. The enzyme composition of claim 1, wherein the recombinant *Trichoderma* host cell is *Trichoderma reesei*.

5. A process for degrading a cellulosic material, comprising: treating the cellulosic material with the enzyme composition of claim 1, and optionally recovering the degraded cellulosic material.

6. The process of claim 5, wherein the cellulosic material is pretreated.

7. The process of claim 5, wherein the degraded cellulosic material is a sugar.

8. The process of claim 7, wherein the sugar is selected from the group consisting of glucose, xylose, mannose, galactose, and arabinose.

9. A process for producing a fermentation product, comprising:

- (a) saccharifying a cellulosic material with the enzyme composition of claim 1;
- (b) fermenting the saccharified cellulosic material with one or more fermenting microorganisms to produce the fermentation product; and

192

(c) recovering the fermentation product from the fermentation.

10. The process of claim 9, wherein the cellulosic material is pretreated.

11. The process of claim 9, wherein steps (a) and (b) are performed simultaneously in a simultaneous saccharification and fermentation.

12. The process of claim 9, wherein the fermentation product is an alcohol, an alkane, a cycloalkane, an alkene, an amino acid, a gas, isoprene, a ketone, an organic acid, or polyketide.

13. A process of fermenting a cellulosic material, comprising: fermenting the cellulosic material with one or more fermenting microorganisms, wherein the cellulosic material is saccharified with the enzyme composition of claim 1.

14. The process of claim 13, wherein the cellulosic material is pretreated before saccharification.

15. The process of claim 13, wherein the fermenting of the cellulosic material produces a fermentation product.

16. The process of claim 15, further comprising recovering the fermentation product from the fermentation.

17. The process of claim 16, wherein the fermentation product is an alcohol, an alkane, a cycloalkane, an alkene, an amino acid, a gas, isoprene, a ketone, an organic acid, or polyketide.

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