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(54) **METHODS OF PRODUCING 6-CARBON CHEMICALS USING 2,6-DIAMINOPIMELATE AS PRECURSOR TO 2-AMINOPIMELATE**

(71) Applicant: **INVISTA North America S.á r.l.**,  
Wilmington, DE (US)

(72) Inventors: **Alex Van Eck Conradie**, Eaglescliffe  
(GB); **Adriana Leonora Botes**,  
Rosedale East (GB)

(73) Assignee: **INVISTA NORTH AMERICA S.A.R.L.**,  
Wilmington, DE (US)

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*Primary Examiner* — Gregory Listvoyb

(74) *Attorney, Agent, or Firm* — Finnegan, Henderson, Farabow, Garrett & Dunner, LLP; Carla A. Mouta-Bellum

(57) **ABSTRACT**

This document describes biochemical pathways for producing 2-aminopimelate from 2,6-diaminopimelate, and methods for converting 2-aminopimelate to one or more of adipic acid, adipate semialdehyde, caprolactam, 6-aminohexanoic acid, 6-hexanoic acid, hexamethylenediamine, or 1,6-hexanediol by decarboxylating 2-aminopimelate into a six carbon chain aliphatic backbone and enzymatically forming one or two terminal functional groups, comprised of carboxyl, amine or hydroxyl group, in the backbone.

**34 Claims, 30 Drawing Sheets**

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FIG. 1

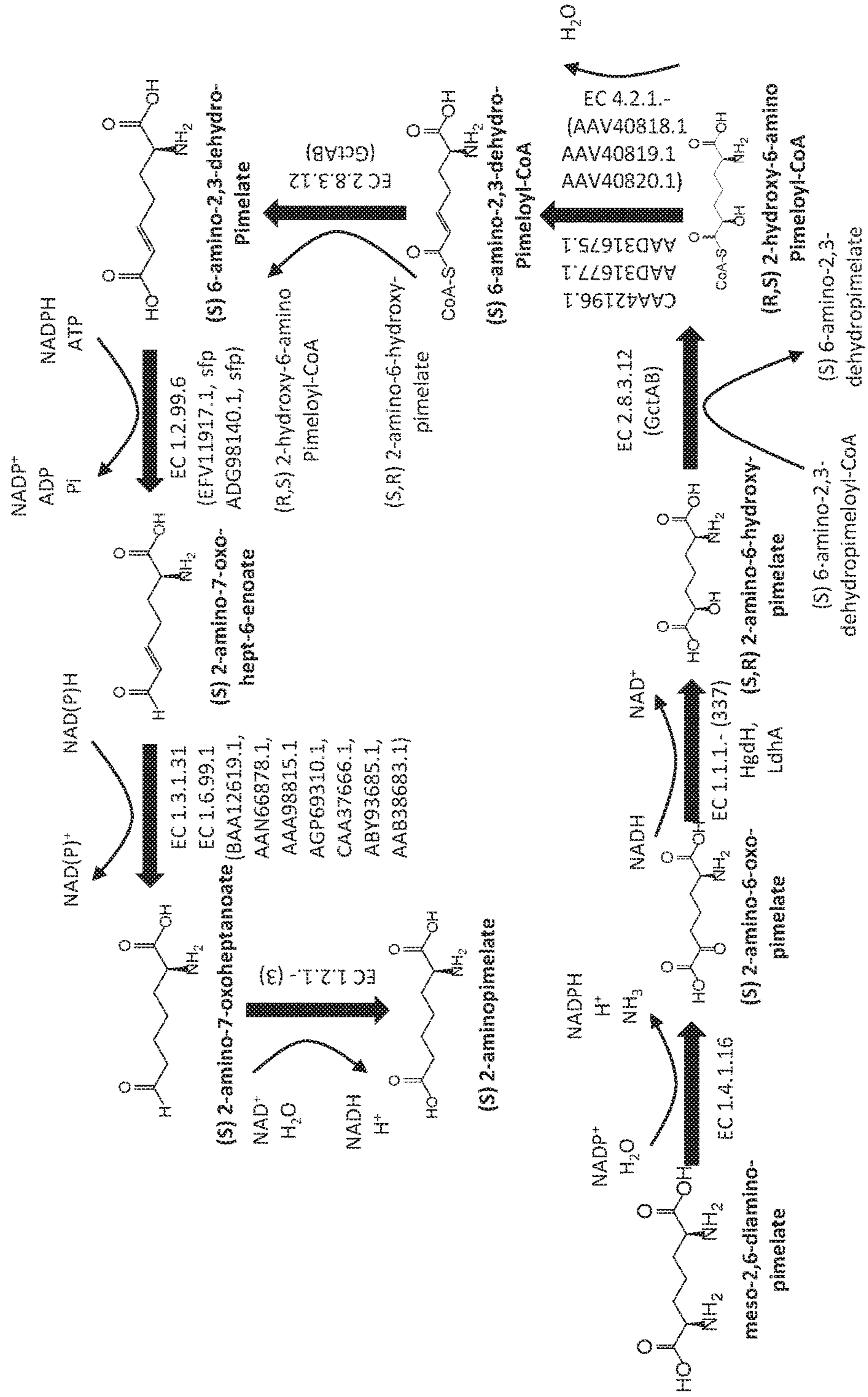


FIG. 2

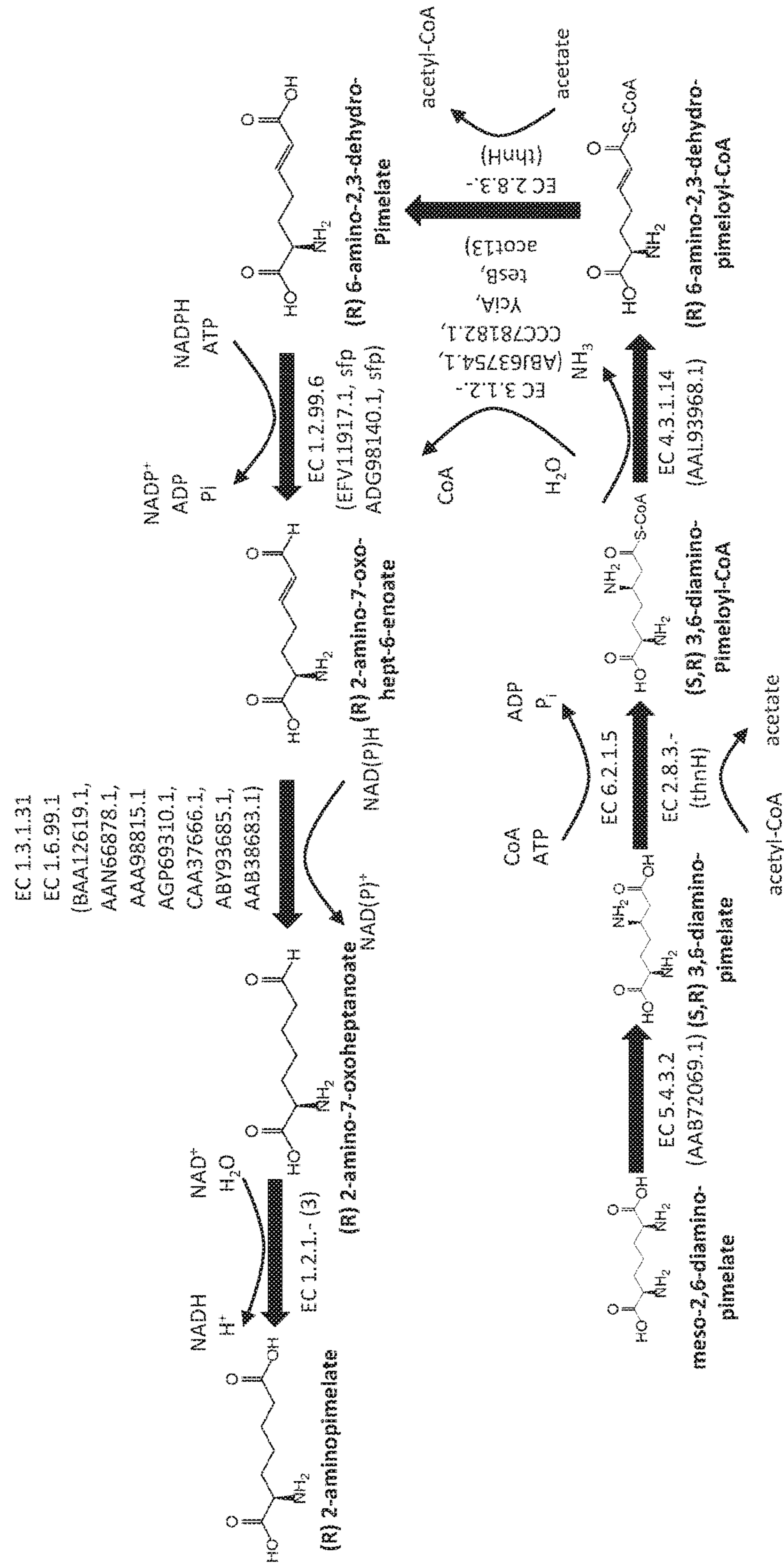


FIG. 3

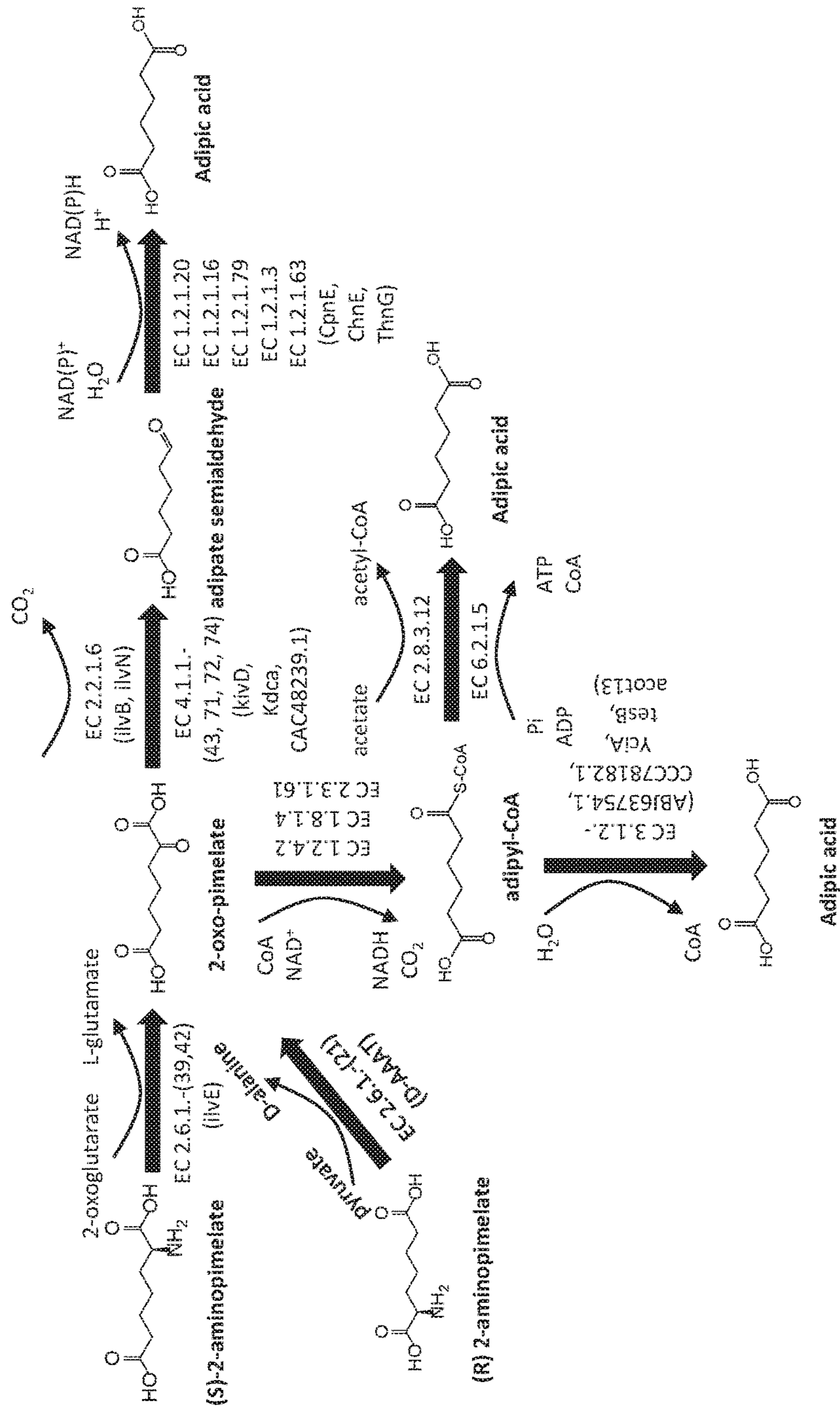


FIG. 4

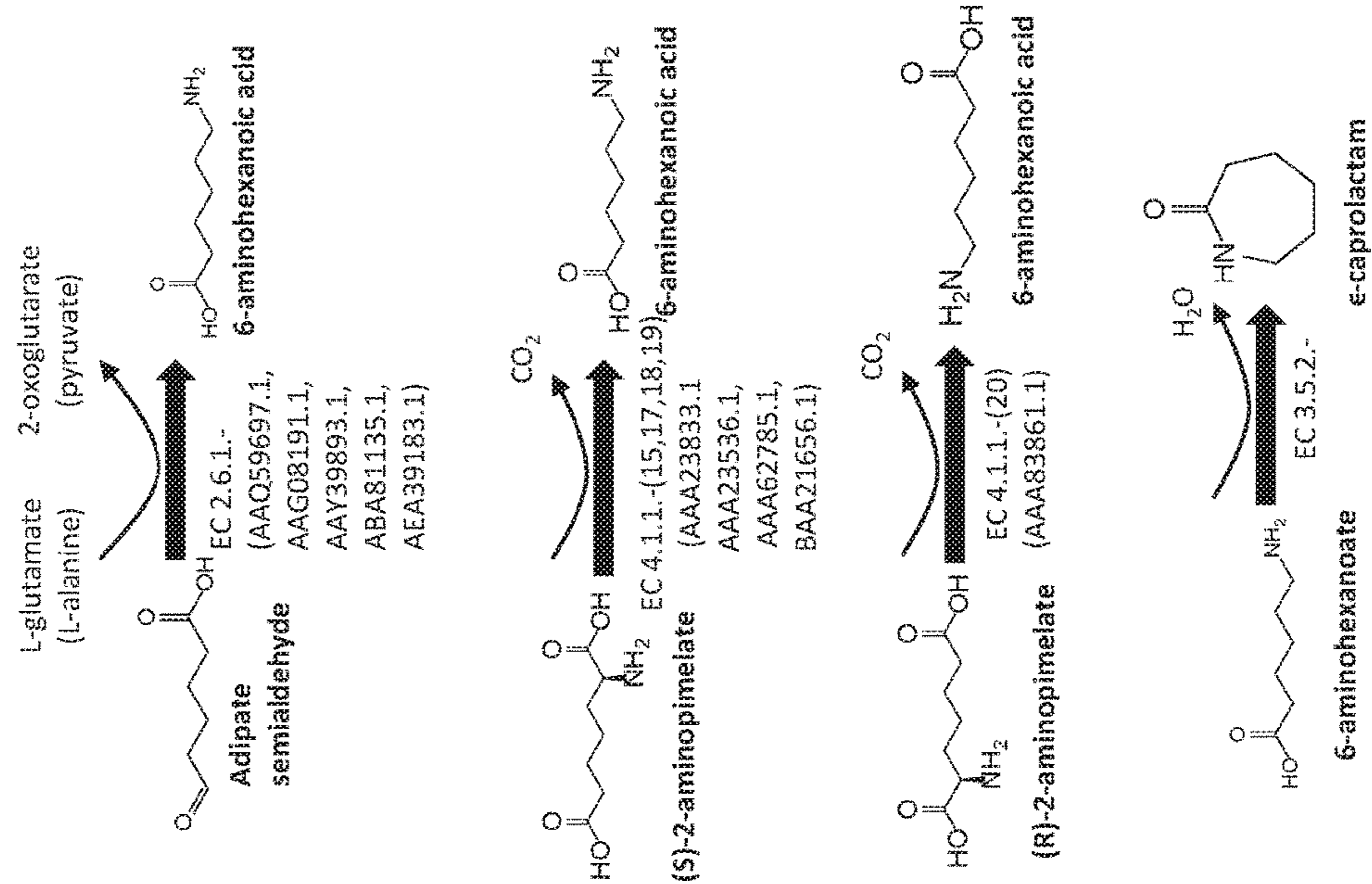


FIG. 5

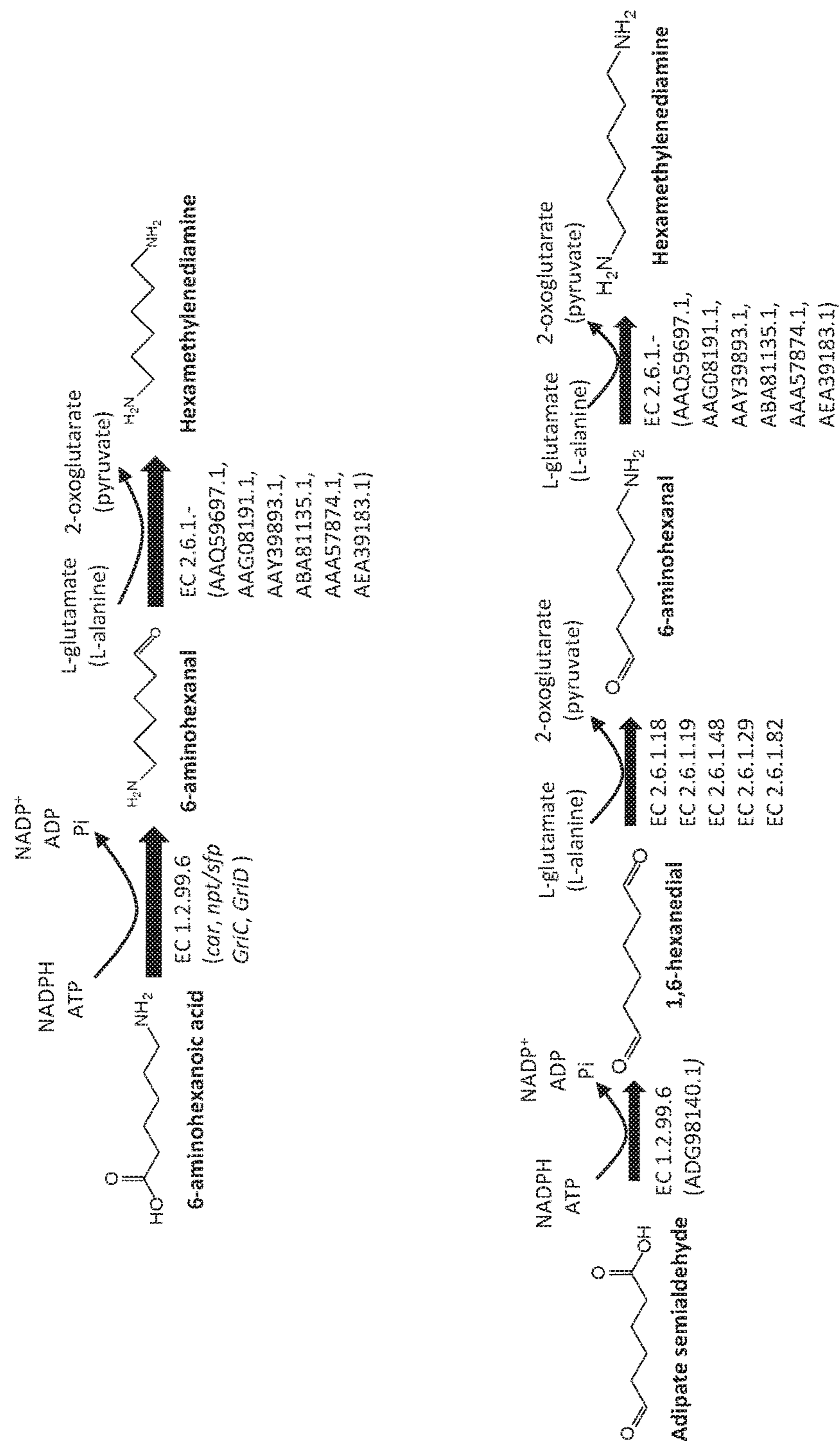


FIG. 6

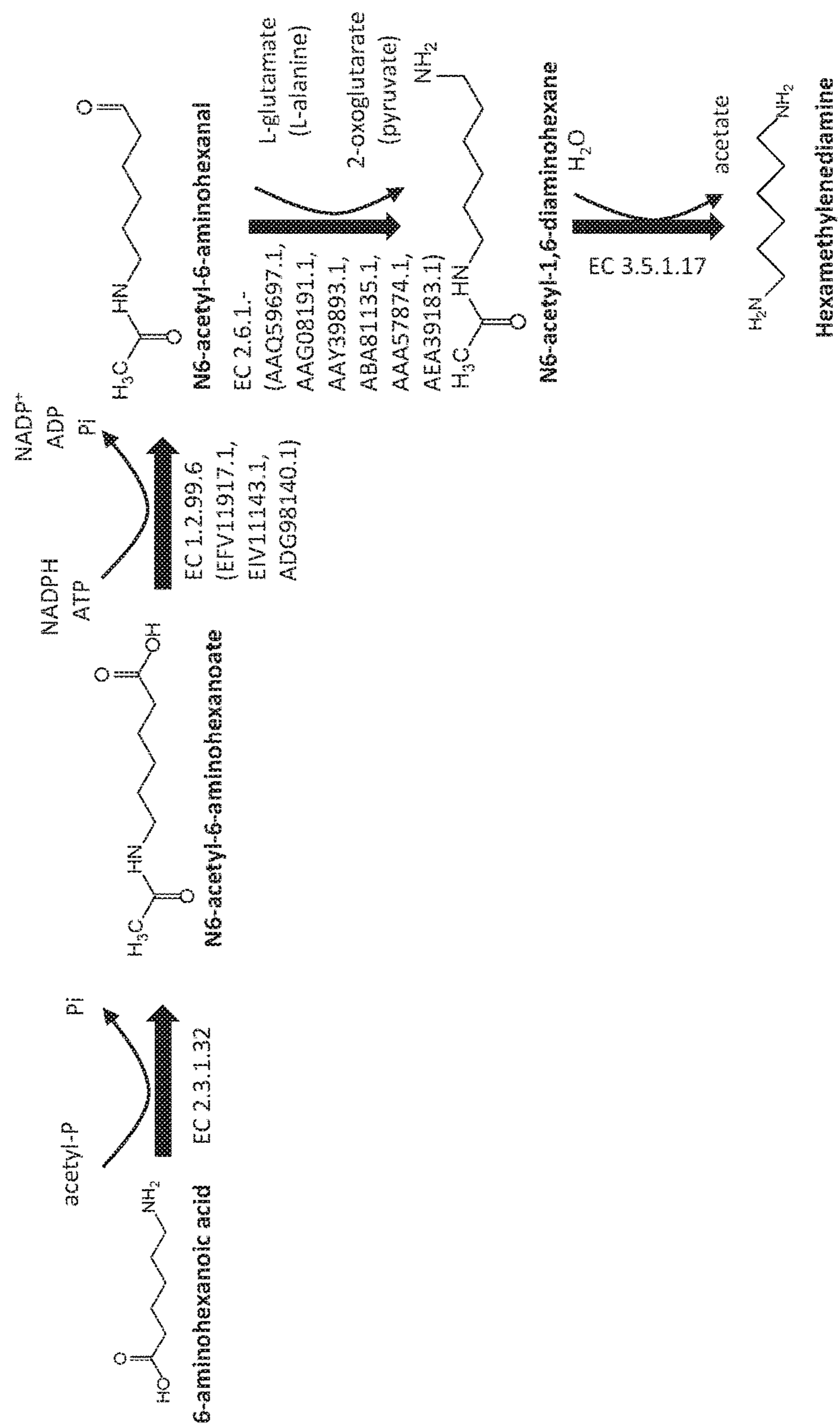




FIG. 7

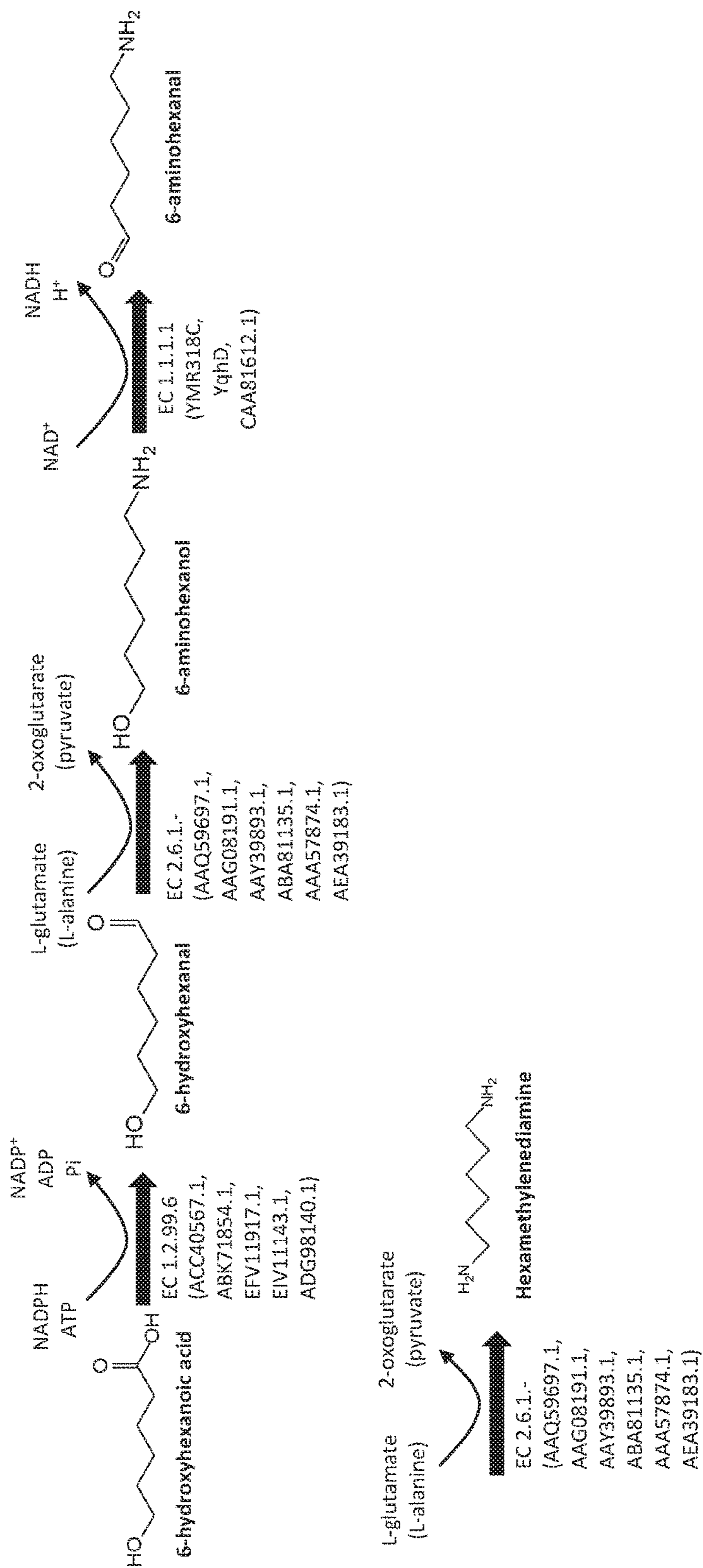


FIG. 8

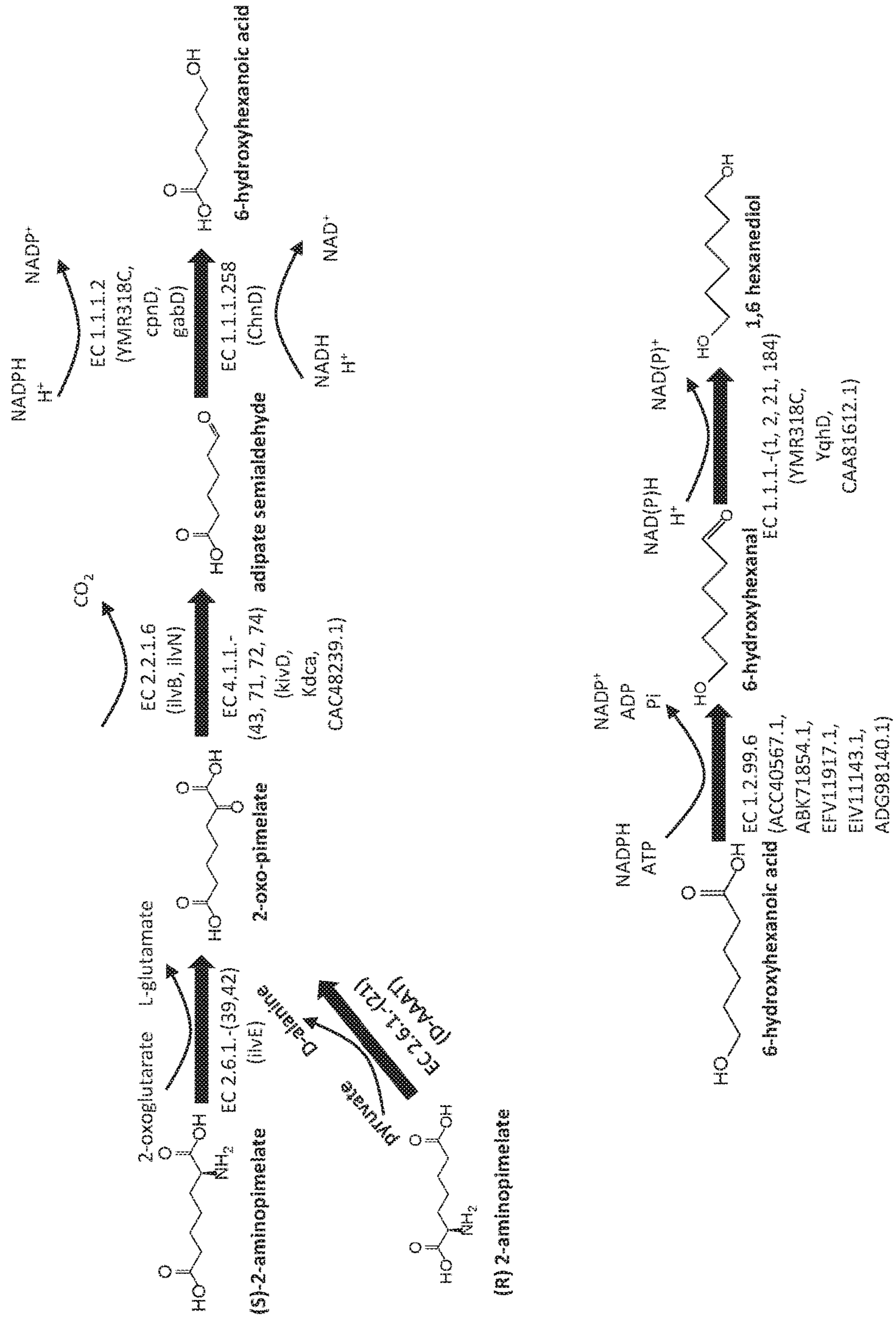


FIG. 9

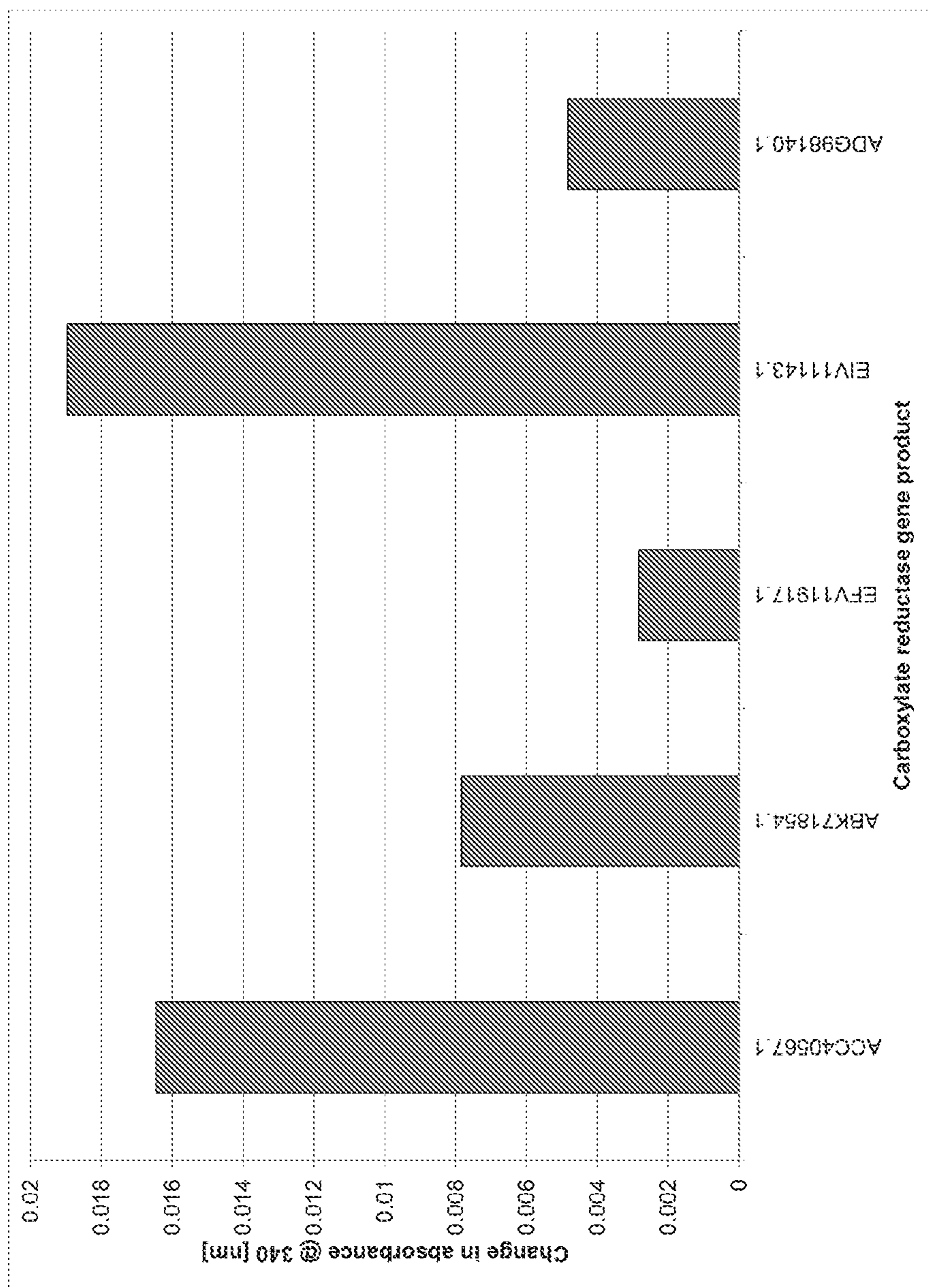


FIG. 10

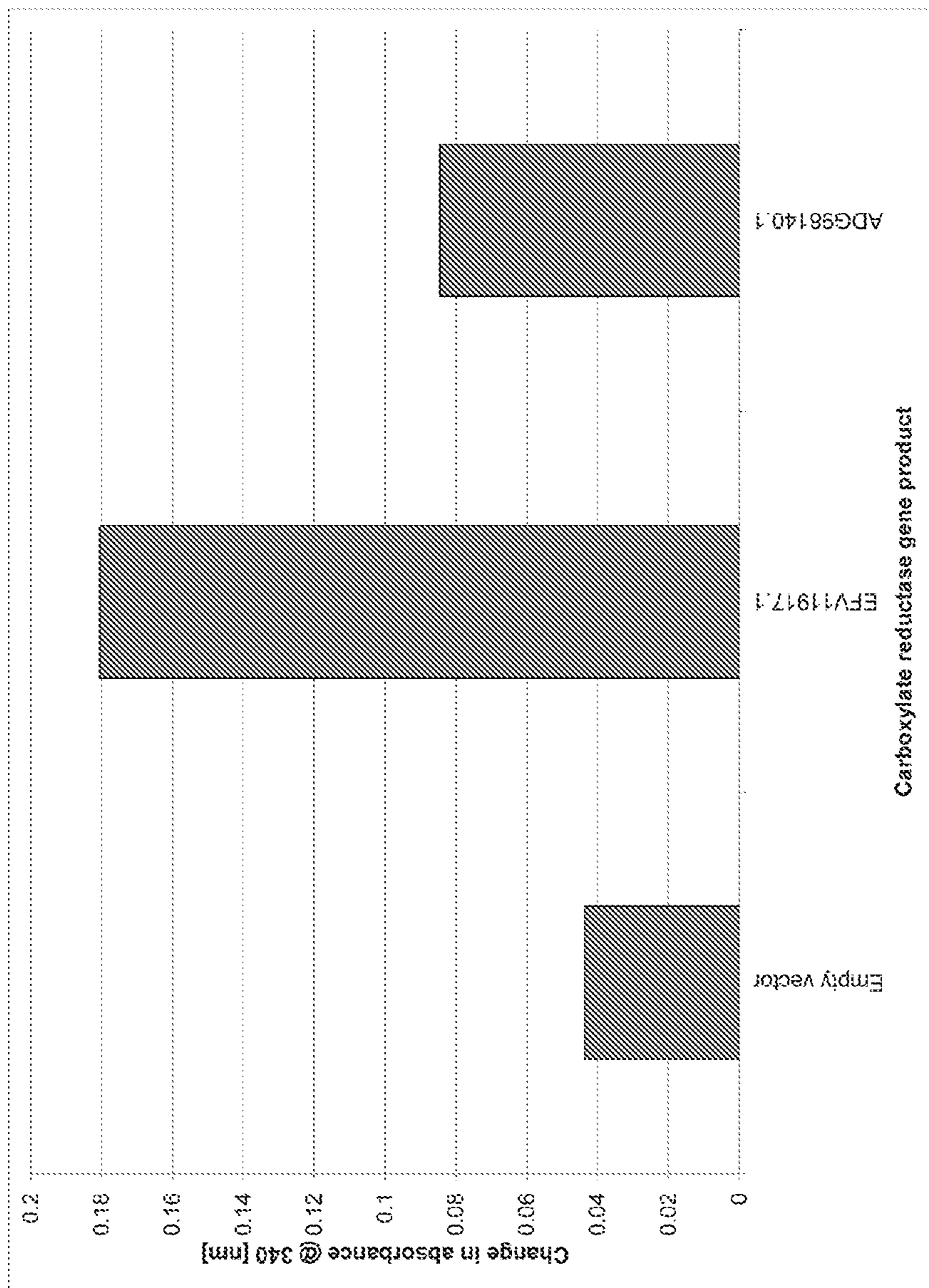


FIG. 11

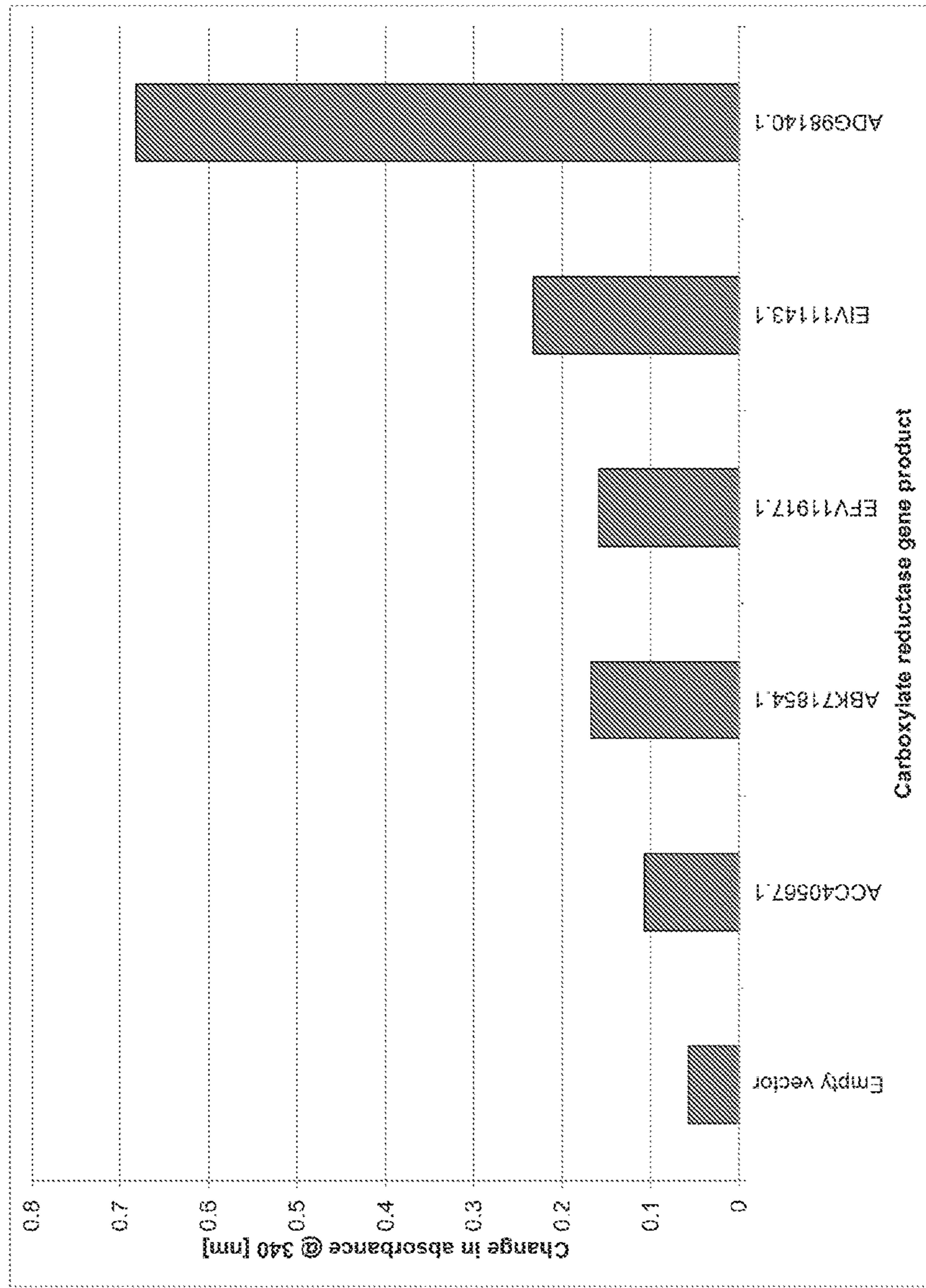


FIG. 12

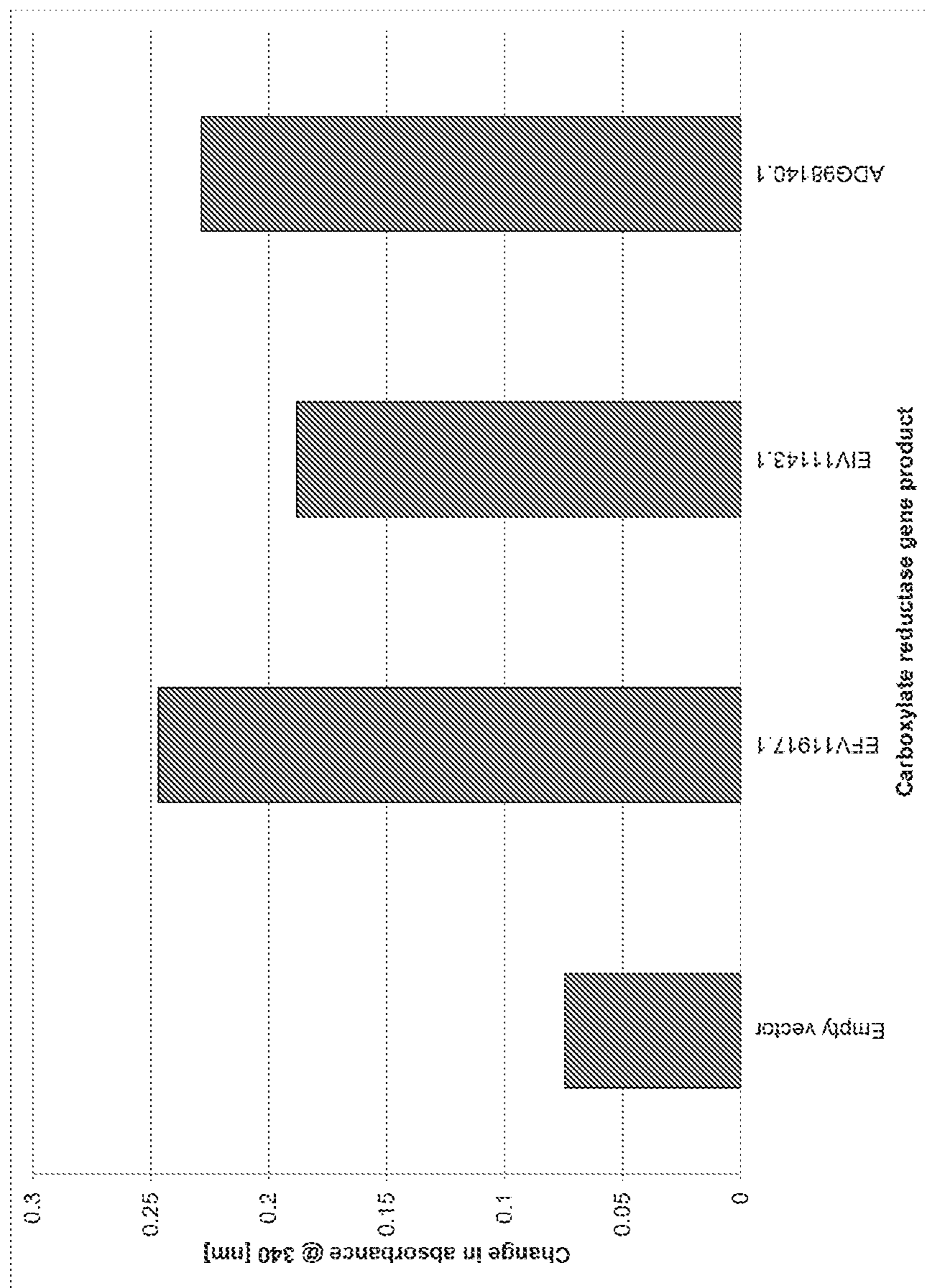


FIG. 13

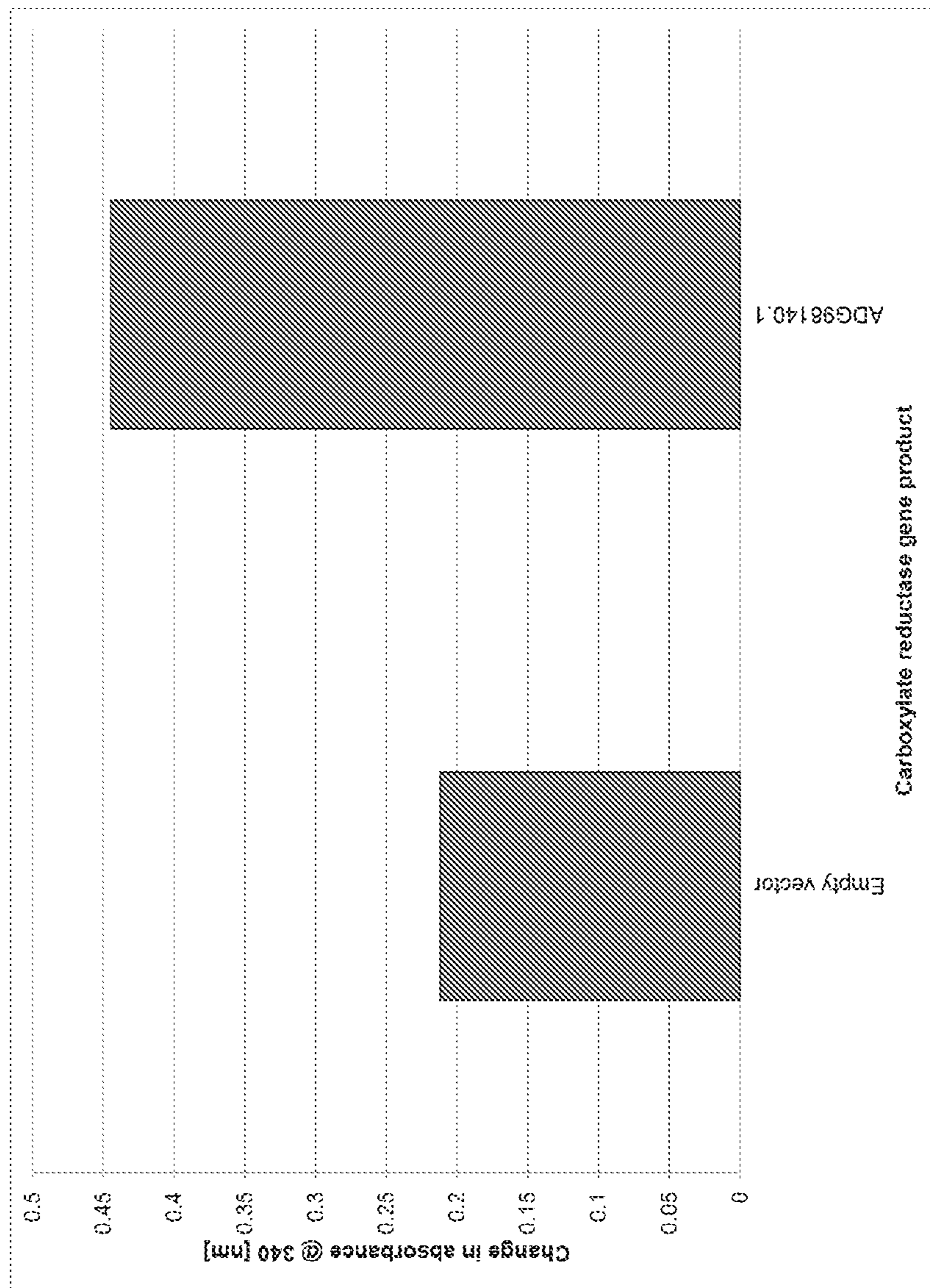


FIG. 14

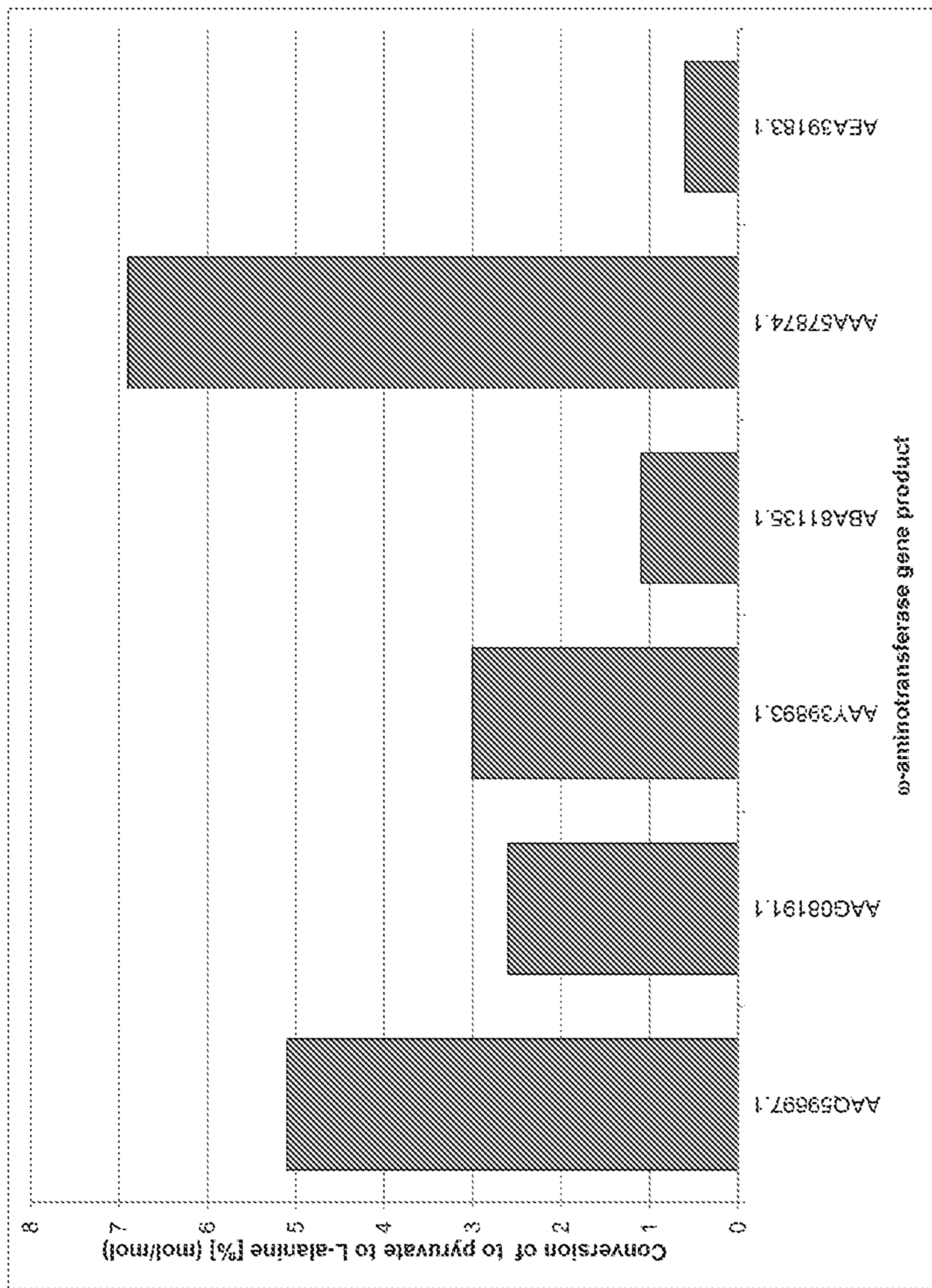




FIG. 15

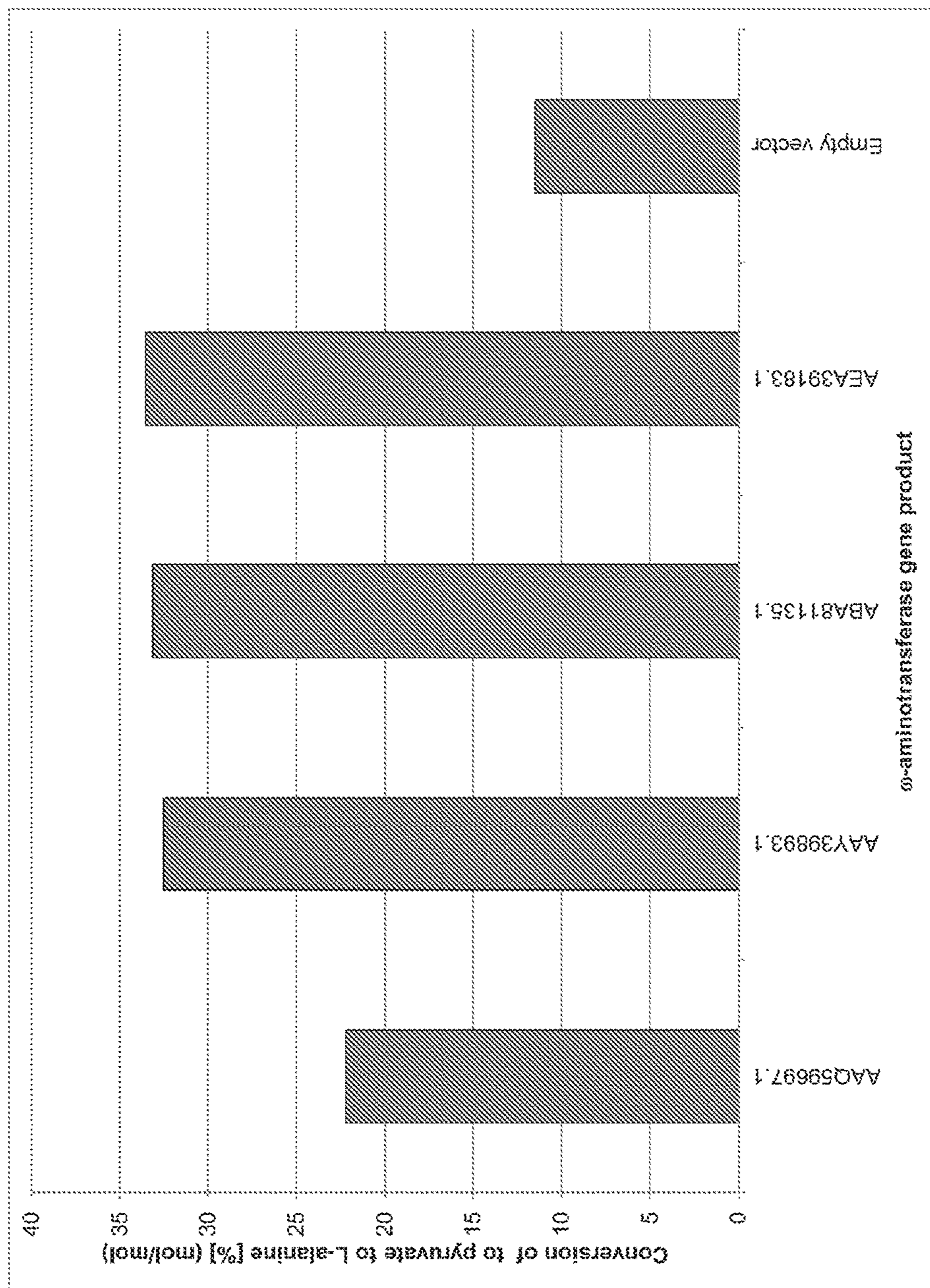


FIG. 16

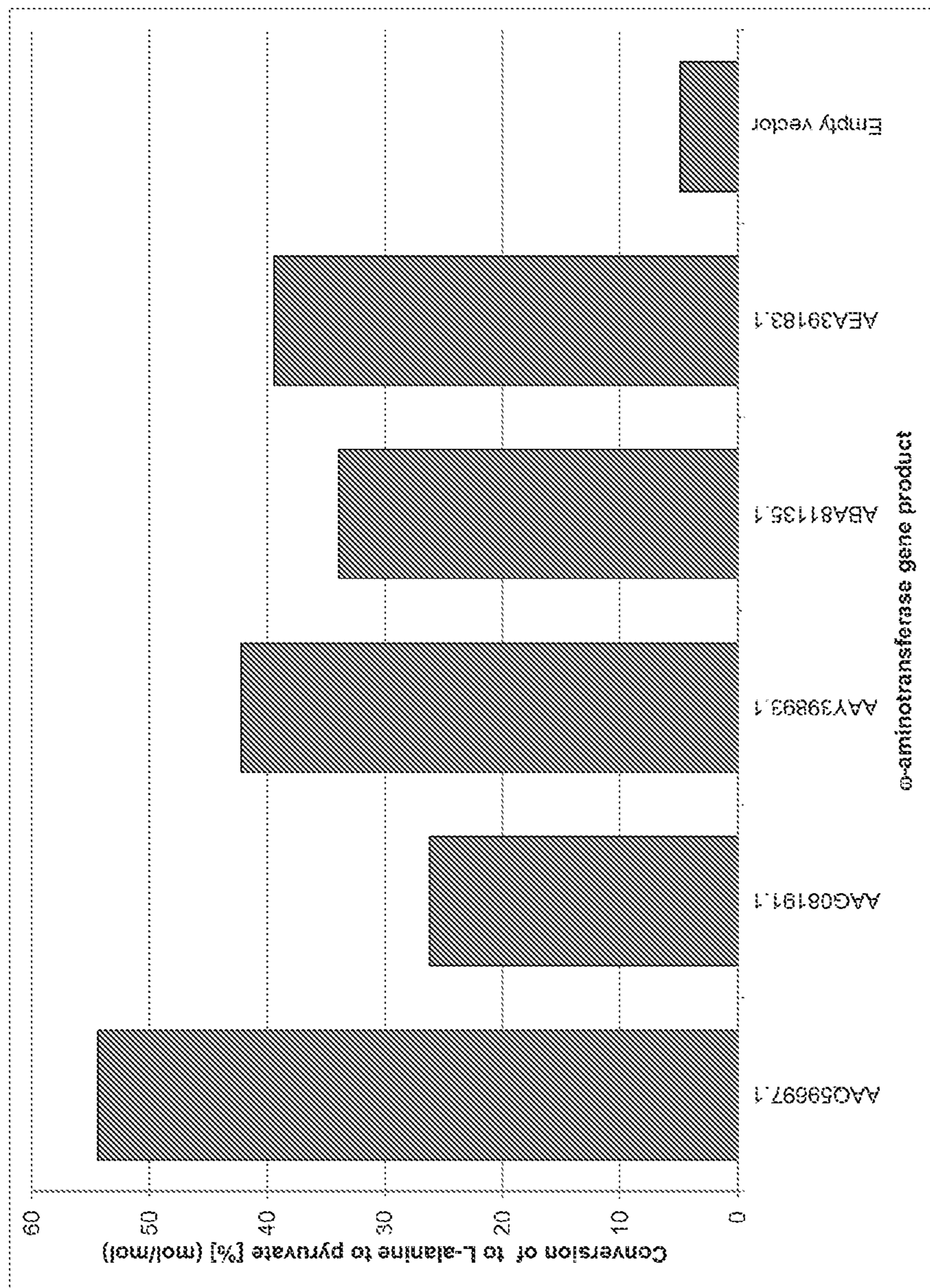


FIG. 17

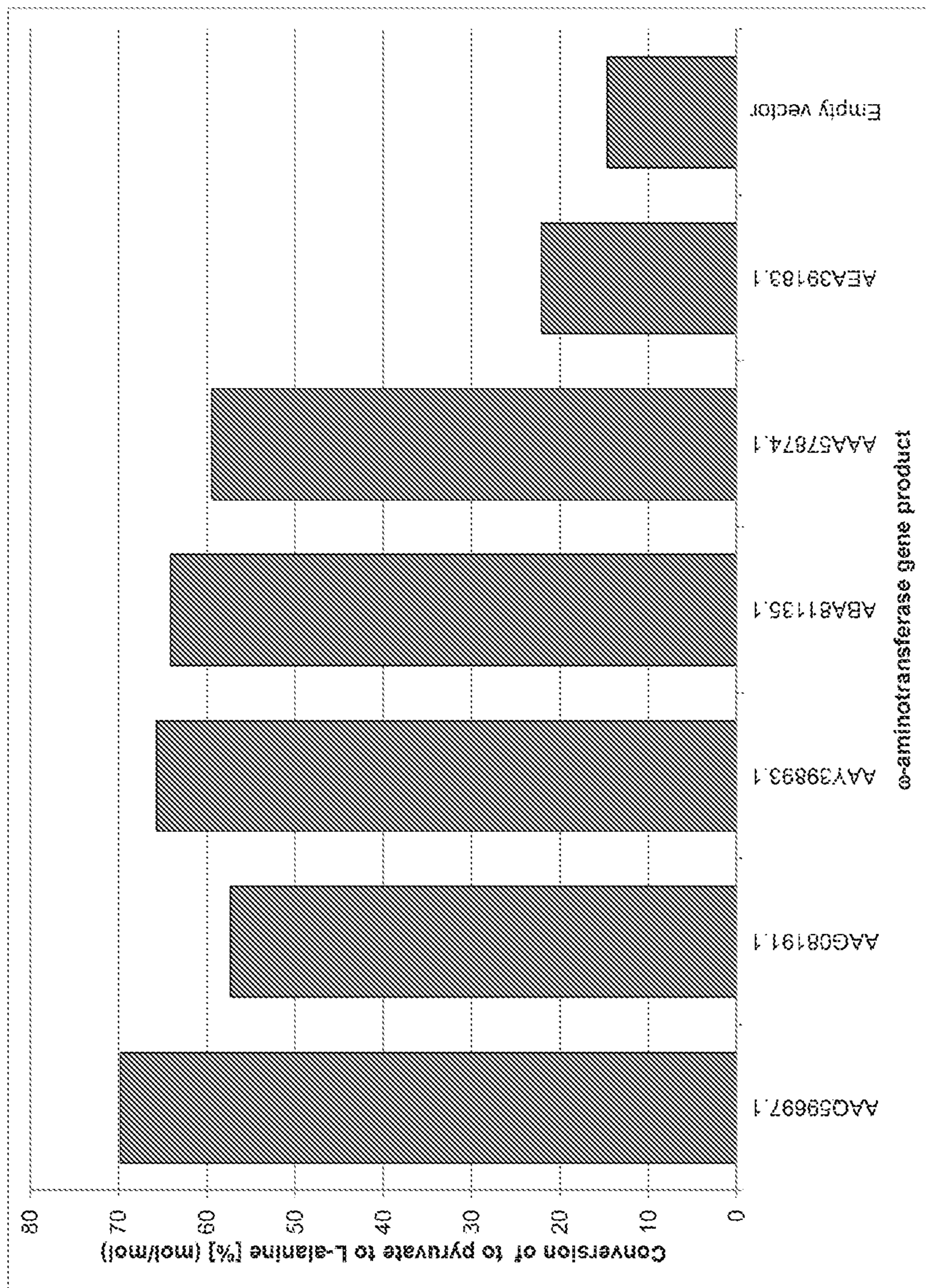


FIG. 18

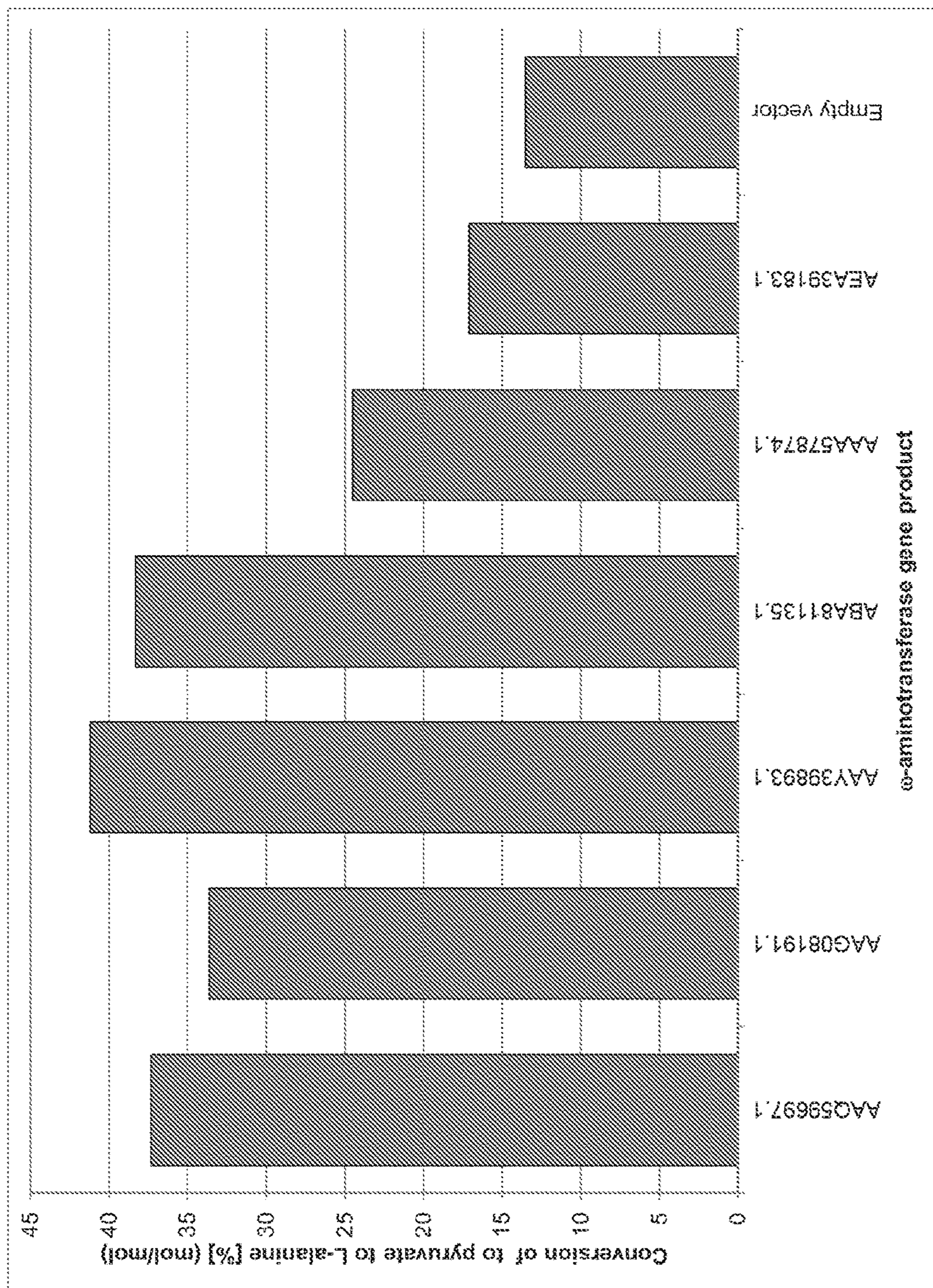


FIG. 19

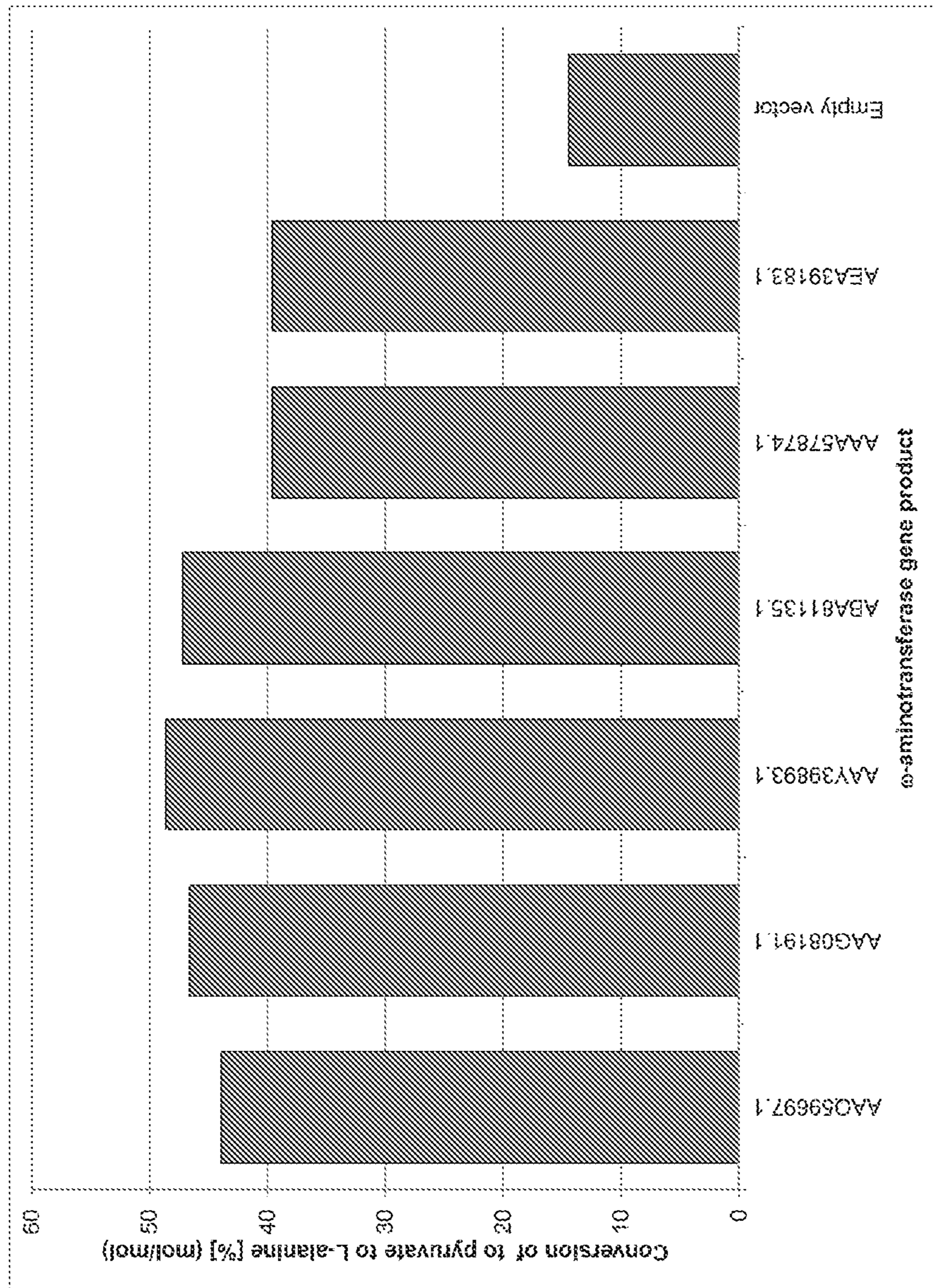




FIG. 20B

SEQ ID NO	Organism	GENBANK reference	Amino acid sequence
4	<i>Mycobacterium smegmatis</i>	ABK71854.1	MTSDVHDAIDGVTETALDDEQSTRIAELYATDPEFAAAAFLPAVVDAAHKPGLRLAEL QTLFTGYGDRPALGYRARELATEDEGRTVTRLLPRFDTLTYAQVWSRVQAVAAALRHNF QPIYPGDAVATIGFASPDYLTDLVCAYLGLVSPQLQHNAPVSRSLAPILAEVEPRILTVS AEYLDLAVESVRDVNSVSQLVFDHHPVEVDDHRDALARAREQLAGKGIIVTTLDIAIDEG AGLPAEPIYADHDQRILAMILYTSGSTGAPKGMYTEAMVARLWIMSFITGDPPTVINVN FMPINHLGGRIPISTAVQNGGTSYFVPESDMSTLFEDLALVRPTELGLVPRVADMLYQHH LATVDRRLVTOGADDELTAEKQAGAEELREQVLGGRVITGFVSTAPLAAEMRAFLDITLGAH VDGYGLTETGAVTRDGVIVRPPVIDYKLIIDVPELGYFSTDKPYPRGELLVRSQILTPGY KRPEVTASVFDKRDGYYHTGDVMAETAPDHLVYVDRRNVIKLAQGEFVAVANIEAVF5GA ALVRQIFVYGNSESRFLLAVVPTPEALEQYDPAALKAALADSLQRTARDAELOSIEVPA DFIVETERFSAANGLLSGVKKLRPNLKDQRYQRLEQMYADIAATQANQLRELRAAATQ FVIDTLTQAAAATILGTSEVASDAHFTDLGGDSLSALTLSNLLSDFGFEVPGTIVNPA TNLAQLAQHIEAQRTAGDRRPSFTTVHGADATEIRAASELTDKFIDAEITLRAAPGLPKVT TEPRTVLLSGANGWLGFRFLTQWLERLAPVGGTLITVGRDDAAARARLTQAYDIDPEL SRRFAELADRRHLRVVAGDIGDPNLGLTPEIWHRLAAEVDLVVHPAALVNHVLPYRQLFGP NVVGTAEVVKLALTERIKPVTYLSVAMGIPDFEEDGDIRTVSPRPLDGGYANGYGN SKWAGEVLLREAHDLCLPVAIFRSDMILAHPRYRQGVNPFDMFTRLLSLLITGVAPRS FYIGDGERPRAHYPGLTVDVFAEAVTTLGAQQREGVSYDVMPHDDGISLDVFDWLR AGHPIDRVDDYDQWVRRFETALTALPEKRRRAQTIVLPLLHAFRAPQAPLRGAPETEVFHA AVRTAKVGGDIPHLDEALIEKIRDLREFGLI

FIG. 20C

SEQ ID NO	Organism	GENBANK reference	Amino acid sequence
5	<i>Segniliparus rugosus</i>	EFV11917.1	<p>MGDGEERAKRFFQRIQELSATDPOFAAAAPDPVAVEAVSDPSLSFTRYLDILMRGYAERP            ALAHRVAGAYETISYGELWARVGAIAAAWQADGLAPGDFVATVGTSPDYVAVDLAAARS            GLVSPLOAGASLAQLVILEETEPKVLAASASSIEGAVACALAAPSVQRLVVFDRGPD            ASESADERRGALADAEQELARAGRAVVVETLADLAARGEALPEALPEPAEGEDPLALL            IYTSSTGAPKGMYSQRLVSQLWGRTPVPGMPNLSLHYPMLSHSYGRAVLGALSAGG            TAHFTANSDLSTLFEDIALARPTFLALVPRVCEMLFQESQRQGDVAELRERVIGGRLIVA            VCGSAPLSPEMRAFMEEVGLFPLLDGYGSTEALGVMRNGIQRPPVIDYKLVDPPELGYR            TTDKPYPRGELCIRSTSLISGYKRPETAEVFDAAQGYKTDGVMMAEIAPDHLVYVDRSK            NVLKLQQGEFVAVANKLEAAAYGTSPYVKOIFVYGNSEKSFLLAVVVPNAEVLGARDQEEAK            PLIAASLQKIAKEAGLQSYEVRDELIEETPTQNGLLSEVGLKLLRPKPKARYGEALEA            RYDEIAHGOADELRALRDGAGQRPVTVVRAAAVAISGSEGAEVGPEANFADLGGDSLSA            LSLANLLHDVFEVEVPRRIIGPTASLAGIAKHEAERAGASAPTAASVHGAGATIRAS            ELTLEKFLPEDLLAAAKGLPAADQVRTVLLTGANGWLGFRFLALEQLERLARGQDGGKLI            CLVRGKDAARRRIEETLGTDPALAAARFAELAEGRLEVVPGDVGEKFGLDAAWDRLA            EEVDVIVHPAALVNHVLPYHQLFGPNVWGTAEHRLAITAKRKPVTYLSLTVAAAGVEPS            SFEEDGDIRAVVPERPLGDDYANGYNSKWAGEVLLREAHHELVLPLVAVFRSDMILAHTR            YTGQLNVPDQFTRLVLSLLATGIAPKSFYQQGAAGERQRAHYDGIPIVDFTAETITLGAE            PSWFDDGGAGFRSFDVFNPHHDGVLDFEVDWLEAGHPISRIDDKHEWFAFETAVRGLP            EAQRQHSLLPLLRAYSFPHPVDDGVSYPYTKFQGVAAQAQVGSDDHDPHGLKALIVKYAD            DLKALGLL</p>



FIG. 20D

SEQ ID NO	Organism	GENBANK reference	Amino acid sequence
6	<i>Mycobacterium massiliense</i>	E1V1.1.143.1	MTNETNPQQEQLSRRRESLRESDPQFRAAQPDPAAVEQVLRPGLHLSSEAIALMTGYAER PALGERARELVIDQDGRITLRLPRFDITTYGELWRSRTTSVAAAWHHDATHFPVKAGDLVA TLGFTSIDYTVLDLAIMILGGVAVPLQTSAPASQWTTILAEAEPTLAVSIELIGAAAMES VRATPSIKQVVVFDYTPVEDDQREAFEAASTQLAGTGIALETLDAVIARGAALPAAPLYA PSAGDDPLALIYTSSTGAPKGMHSENIWRWIREDMAGTENILPMIGLNFMPMSHI MGRGTLTSTLSTGGTYFAASSDMSTLFEDMELRPTALALVPRVCDMVFORFQTEVDRR LASGDTASAEAAEVKADIRDNLFGGRVSAVMVGSAPLSEELGFEIESCFELNITDGYG STEAGMVFVRDGIQRPVIDYKLVDPVPELGYFSTDKPMPRGELLKTDGMFLGYKRPV TASVFDADGFMVTDIVAEIADHNDIEHRRNNVVKLSQGEFVAVATLEAEYANSPVVHQ IYVYSSERSYLLAVVFTPEAVAAAKGDAALKTTIADSLQDIKAEIQLSYEVPRDFI IEPQFTQNGSLTGIAKLARPNIKAHYGPRLEQMYAEIAEQQAELRALHGVDPDKPAL ETVLKAQAQLLGVSSAEAAADAHFTDLGGDSLSALSFDLLRDFAVEVVPVGVVSAAND LGGVAKFVDEQRHSGGTRPTAETVHGAGHTEIRAADLTLDKFEIATLHAAPSLPKAAGI PHTVLLTGSNGYLGHYLALEWLERLDKDKUVIVRGKNAEAAVGRLEEFDTGDTTELL AHFSLADKHLVLAGDIGDNLGIDADTWQRLADTVDIVHPAALVNHVLPVYNQLFGPN VVGTAEHKLAFTTKIKPVTYLSTVAVAAVVDPTTFDEESDIRUSAVRPIDDDGYANGY NAKWAGEVLLREAHDLGGLPVAVFRSDMILAHSRVTGQLNVPDQFTRLSLIATGIAPG SFYQAQTTGERPLAHYDGLPGDFTAEAITTLGTQVPEGSEGFVYDCVNPHADGSLDNF VDWLIEAGYPIARIDNYTEWTRFDTAIRGLSEKQKQHSLLPLLHAFEQPSAAENHGVVP AKRFQHAVQAAGIGPVGQDGTDDIPHLRRLIVKYAKDLEQLGLL

FIG. 20E

SEQ ID NO	Organism	GENBANK reference	Amino acid sequence
7	<i>Seghiiiparus rotundus</i>	ADG98140.1	MTQSHIQGPOASAAHSLARRAAELLATDPOAAAATLPDPEVVRQATRPGLRAERVDAIL SGYADRALGQRFQTVKDPITGRSSVELLPTFDITTYRELPERATAIASDLAHHHPQAPA KPGDFLASIGFISVDYVAIDIAIGVFAGLTAVPLQTGATLATLTAITAEAPTLLFAASIEH LPTAVDAVLATPSVRRLLVFDYRAGSDEDEAEEAAKRIADAGSSVLVDVDEVIARGK SAPKAPLPATDAGDQSLIIVTSGSTGTPKGMYPERNVAHFVGGVWAAAFDEDAAPP VPAINITFLPSHVASRLSMFTLARGGLMHFVAKSDLSLTFEDLKLARPTNLFVPRVV EMLYQHYQSELDRRGVQDGTREAEAVKDLRTGLGGRILTAGFGSAPLSAELAGFIESL LQIHLVDGYGSTEAGPVWRDGYLVKPPVTDYKLIIDVPELGYFSTDSPPHPRGELAIKTQTI LPGYYKRPETTAEVEDEGGYLTGDVAQIGPEQFAVYDRKKNVILKLSGGEFFVTLAKLEA AYSSSPLVRQLFVYGSSESYLLAVIVPTPDALKKFGVGEAAKAAALGESLQKIADEGLQ SYEVRPDIETDPFTVENGLSDARKSLRPKLKEHYGERLEAMYKELADGGQANELRDIR RGVQQRPTLETVRRAAAAMLGASAAEKPDAAHFDLGGDSLSALTSFNFLHDLFEVDVPPV GVVSAANTLGSVAEHIDAOIAGGRAPPTFATVHGKSTTIKASDLTKDFIDEQTEAA KHLKPADPPTVLLTGANGWLRFLALEWLERLAPAGGKLTIVRGKDAQAQAKARLDA YESGDKLAGHYQDLAATLLEVLADGDFSEPRGLDEATWNRLADEVDFISHPGALVNHVL PYNQLFGPNVAGVAEIIKLAITTRIKPVTYLVAVAAAGVEPSALDEDEDGDIRTVSAERSV DEGYANGYNSKWGGEVLLREAHDRDTGLPVRFRSDMILAHQKTYTQGVNATDQFTRELVQS LLATGLAPKSYELDAQNRQRAHYDGPVDFTAESITLGGDGLGYSYVNFNPHRDG VGLDEFVDWLEAGHPTRIDDDYDQWLSRFETSRLRGLPEKRSQASVLPLLHAFARPPAV DGSFRNTVFRTEVQKAKIGAEHDIPHLGKALVLYADDEIKQLGLL
8	<i>Chromobacterium violaceum</i>	AAQ59697.1	MQKORTTSQWRELDAAHHLHFFDTASLNQAGARVMTRGEVYVWDSSEGNKIIDGMAGLW CVNVGYGRKDFAEARRQMEELPFYNTFFKTHPAAVVELSLLAEVTPAGFDRVYFTNSG SESVDTMRMVRRYWVQKPEKTLIGRWNGYHGSTIGGASLGGMKYMIHQGDLPPIGM AHIEQPWWYKHGKDMTPDEFVVAARWLEEKLEIGADKVAAFVGEPIQGAGGVVPPAT YWPEIERICRYDVLVVADEVICGFGRTGEWFGHQHFGFOPDLFTAAGLSSGYLPIGAV FVGRVAEGLIAGGDFNHGFTYSGHPVCAVAHANVAALRDEGIVQRVKKDIDGPIYMQKRW RETFSRFEHVDDVGVGMVQAFLLVKNKAKRELFPDFGEIGTLCRDIFFRNLIIMRACGD HIVSAPPLVMTRAEVDEMLAVERCLEEFEEQTLKARGLA
9	<i>Pseudomonas aeruginosa</i>	AAQ08191.1	MINARLHATSPIGDADLVRADQAHYMHVGHVDFDHRVNGSENIAGDGAYIVDTAGNRYLD AVGGMWCTNIGLGRMARTVAEQTRLLAYSNPFCDMANPRAIELCRKLAELAPGDLDHV FLTTGGSTAVDTAIRLMHYQNCGRKRAKHHVTRINAYHGSTFLGMSLGGKSADRPAEF DFLDERIHLACPYRYRAPEGLGEAEFLDGLVDEFERKILELADRVGAFISEPVFGSGG VIVPPAGYHRRMWELCORYDVLVYSDEVVTSFGRGLGHFFASQAVFGVQPDILTAKGLTS GYQFLGACIFSRRIWEVIAEPDKGRCFSHGFTYSGHPVACAAALKNIEHREGLLAHAD EVGRYFEERLQSLRDLPIVGDVGRMIRFMACEVFEVADKASKALFPESLNIGEWVHLRAQKR GLLVRPIVHLNVMSPPULITREQVDTVVRLRESIEETVEDLVRAGHR

FIG. 20F

SEQ ID NO	Organism	GENBANK reference	Amino acid sequence
10	<i>Pseudomonas syringae</i>	AA539893.1	MSANNPQTLEWQALSSSEHLLAPFSYKQIKKPKRIITRAEGVYLVWSEGNKILDGMSGL WCVAIGYGREELADAASKQMRLEPYNLFQTAHPPVLELAKASDIAPGEMNHVFFFTGS GSENDTMLRMVRYWALKGQPNKKTIIISRVNGYHGSTVAGASLGGMTYMHHEQGDLPFG VYHIPQYWFEGGDMTPDEFGIWAAEQLEKILELGVENVGAFIAEPIOGAGGVIVPPD SYWPKIKEIISRYDILFAADEVICGFGRTSEWFGSDPYGLRPMMTIAKGLTSGYVPMGG LIVRDEIVAVLINEGGDFNHGFTYSGHPVAAAVALENIRILREEKIVERVSETAPYLQKR LRELSDPHLVGEVRGVLGALIELVKDKTTREYTDKGAGMICRTECFDNGLIMRAVGD MHAAPLVISFAQIDELVEKARTCLDELAVLQG
11	<i>Rhodobacter sphaeroides</i>	ABA81135.1	MTRNDATNAAGAVGAAMRDHILLPADEMAGKLSAQVPLTHAEGIVVHTEDGRRLLDGP GMWCAQVGYGRREIVDAMAHQAMVLPYASPWYMATSPAARLAEKIATLTPGDENRIEFTT GGSTAVDSALRSEFYNNVLRPQKRIIYRVDYGHGSTALLTAACRTGRTGNWPNFDIAQD RISFLSPNPRHAGNRSQAEFLDLVQEFEDRIESLGPDTIAFLAEPILASGGVIIPPA GYHARFKAICEKHILYISDEVVTGFCRGEWFASEKVFVWPDHITFAKGVTSYVPLG GLAISEAVLARISGENAKGWFNGYYSNQPVACAAALANIELMEREGIVDQAREMADY FAAALASLRDLPGVAETRSVGLVGVQCCLDPTRADGTAEDKFTLKIDERCFCFELGLIVR PLGDLCVISPLHSRAQIDEMVAIMRQAITVSAAHGLTAKPAAV
12	<i>Escherichia coli</i>	AAA57874.1	MNRLPSSASALACSAPHAENLIEKRTLDEHEMKALNREVIEWFKEHVNPGEFLEYSKSYTAG GDYGAWEWQAGSLNTLVDQGGERIDCLGGFGIFNVGHRNPVVSVAQNQLAKQLHSQE LLDPLRAMLAKTLAALTPGKLYSFFCNSGTSVEAALKAKAYQSPRKFTRIATSGAF HGKSLGALSATAKSTFRKPMPLLPGRHVDFGNHEAMRTALNECKKTGDDVAIVLEPI QGGGVILPPPGYLTAVRKLCEFGALMILDEVOTGMGRITGKMFACHEENVQFDILCLAK ALGGVMPIGATIAEEVSVLFDNPFLLHTTFFGGNPLACAAALATINVLLEONLPAQAE QKGDMLLDGFRQLAREYDVLQEARGGKMLMAIEFVDNEIGNYFASEMFRQRVLVAGTLN NAKTIRIEPPLTLTIEQCELVKAAKALAAAMRVSEEA
13	<i>Vibrio Fluvialis</i>	AEA39183.1	MNKPQSWEARAETSLYGFDDMPSLHQRGTVVVTHGEGPIYEVNRRYLDANSGLWNMV AGFDHKGLIDAAKAQYERFPQYHAFGRMSDQTVMLSEKLEVSFPDSSGRVFTNSGSEA NDTMVKMLWFLHAAEGKPKRKLTRWNAYHGVTAVSASMTGKPYNSVFGPLPFGVHLT CPHYWRYGEEGETEEQFVARLARELEETIQREGADTIAGFFAEPVMGAGGVIPPAKGYFQ AILPILRYDIPVISDEVICGFGRTGNTWGCVTYDFTPDAISSKNLTAGFFPMGAVILG PELSKRELTAEAIIEEPPHGFTASGHPVGCALAKAIDVVMNEGLAENVRRLAPRFEERL KHIAERPNIYGRGIGFMWALEAVKDKASKTFFDGNLSVSERIANTCTDILGLICRPLGQS VVLCPFFILTEAQMDEMFDKLEKALDKVFAEVA
14	<i>Bacillus subtilis</i>	CAA44858.1	MKIYGYMDRPLSQEENEFMSFISPEKREKCRFRFYHKEDAHRTLLGDVLRVSRQYQ LDKSDIRFSTQYEGKPCIPDLPDAHFNISHSGRWVICA FDSQPIGIDIEKTKPISLEIAK RFFSKTEYSDLLARKDDEQTDYFYHLWSMKESFIKQEGKLSLPLDSFVRLHQDGCQVSI ELPDSHSPCYIKTYEVDPGYKMAVCAAHDPEDPEDITMVSYEEL

FIG. 20G

SEQ ID NO	Organism	GENBANK reference	Amino acid sequence
15	<i>Nocardia</i> sp. NRRL 5646	AB183656.1	MIEILPAGVESAELEYPEDLKAHPAEHEHIAKSVEKRRRDFIGARHCARLALAEELGEP PVAIGKGERGAPWPRGVVGSLSLTHCDGYRAAAVAHMKMRFRSIGIDAEFHATLPEGVLDVSV SLPPERWIKTTDSALHLDRLLFCAKEATYKAWWPLTARWLGFGEFAHITFEIEDGSADSG NGTFHSELLVPGQTNDGGTIPLLSFDGRWELJADGFFILTAIAYA
16	<i>Bacillus subtilis</i>	BAA12619.1	MARKLFTPIIKDMTLKNRIVMSPMCMYSSHEKDGKLTFFHMAHYISRAIGQVGLIIEA SAVNPQGRITDQDLGIWSDHEHIEGFALTEQYKEQSGKIGQLAHAGRKALEGDFHAPS AIAFDEQSATPVEMSAEKVETVQEFKQAAARAKEAGFDVIEIHAAGYLHIEFLSPLSN HRTDEYGGSPENRYRFLREHDEVKQWWDGGLFVRSASDYTDKGLDIADHIGFAKWKIKE QGVLDLDCSSGALVHADINVPFGYQVFAEKIREQADMATGAVGMITDGSMAEEILQNGR ADLIFIGRELLRDPFFARTAAQLNTEIPAPVQYERGW
17	<i>Pseudomonas putida</i>	AAN56878.1	MSALFEPYTKDVTLRNRIAPPMCCYMAEDGMINDWHHVHLGAGLARGGAGLLVEATAV APEGRITPGCAGIWSDAHAQAFVPPVQAIAKAA GSVPGIQAHAHAGRKAASANRPFWEGLDHHIA ADDAARGWETAPSAIAFGAHLPKVPREMTLDDIARVKQDFVDAARRARDAGFEWIELHFA HGVLGQSFSEHSNKRTDAYGGSDNRSRFLLETLAAVREVWPNLPLTARFVLEYGDR DEQTEESIELAPRFKAGGLDLSVSVGFTIPDNIPWGPAPMGPFAERVREAKLPVTS AWGFGTQLAEALQANQLDLSVSGRAHLADPHWAYFAAKELGVEKASWTLPAHYAHWLE RYR
18	<i>Kluyveromyces fragilis</i>	AAA98815.1	MSFMNFERPLADTDFIKPIKINTEIKHRVVMPPALTRMRALHPGNVFPNDWAVEYRQR SQYPTMIITEGAFSAQGGYDNPAGVWSEELAQWIKFKAIHDKNSFVWVQLWVLR QAFADNLARDGLRYSASDEVYMGEDKERAIRSNPQHGITKDEIKQYIRDYDAKKC IDAGADGVEIHSANGYLLNQFLDPIPNKRTDEYGGSIENRFRVLEVDVAVDGAERT SIRFSPYGVFTMSGGSDPVLVAGFAYVLAELEKRAKRAYVLDVPRVTSPPQPEF EGWYKGGTMEFVYVWKGNVLRVGNVALDPPDAAITDSKNPNTUIGYGRAFIANPDLVERL EKGLPLNQYDRPSFYKMSAEGYIDYPTTYEEAVAKGYKK
19	<i>Lactobacillus casei</i>	AGP69310.1	MSGYHFKPFFKHKQHTLKNRIVIPPMITRLSFDGTVTRDEIRVYQQRAGGVGMRIITG TANVWALGKGFEGELSVADDRFIPGLSKLAAAMKTGGTKAILQIFSAGRMSNSKILRGEQ PVSASAVAAPRAGYETPRALTSAEIEATHDFGQAVRRAILAGFDGIELHGANTYLIQQF YSPNSNRRTDWGGDRKMRPLAVVHEAKVIAIADRPFLGGRYRISPEELEECPGITL DDTLALIDALKQTKIDYLVHVSQSDVWRTSLRNFEDTAIMNEQIRDHVAGAFVIVVGGIK TPADAEKAAESFDLVAIGHEMIREPHVWQKVLVDHDEKAIYQIAPADLLEELGIAPTFLDF IESISGGAKGYPLTTAQSVTSSNVTDQ
20	<i>Saccharomyces pastorianus</i>	CAA37666.1	MSFYDFKPOALGDTNLFKPKIGNNEILLHRAVIPPPLTRMRALHPGNIPNRDWAVEYTTQ KARPGTMIITEGAFISQAGGYDNPAGVWSEELQWIKFKAIHDKNSFVWVQLWVLR WAAFPDNLARDGLRYSASDNVFMDAEQEAKAKKANPQHSITKDEIKQYKEYVQAAKN SIAAGADGVEIHSANGYLLNQFLDPIPNKRTDEYGGSIENRFRVLEVDVAVDGAERT VGLRSPYGVFNMSGGAEITGIVACVAVVAGELEKRAKRAYVLDVPRVTSPPQPEF GEGEYEGGSDVYVSIWKGPVIRAGNFAHPEVREEVKDKRTLIGYGRFFISNPDLVDR LEKGLPLNKYDRDTEYQMSAHGYIDYPTTYEEALKEGWDKK

FIG. 20H

SEQ ID NO	Organism	GENBANK reference	Amino acid sequence
21	<i>Thermoanaerobacter pseudethanolicus</i>	ABY93685.1	MSILHMPKIKKIDTIKINRIMVSPMCMYSASTDGMFNDWHIVHYATRAIGGVGLIMQEATA VESRGRITDHDLGWVNDQVKELKIVDICKANGAVMGIQLAHAGRKCNISSYEDVVGPS IKAGDRYKLPRELSVEEKIVKAFGEAAKRAKLANAGYDVVEIHAAGYLIHEFLSPLSNK RKDEYGNSEINRARFLIEVIDEVRKNWPNKPIFVRSADDYMEGGINIDMMVEYINMIK DKVDLIDVSSGGLNVDINILYPGYQVYAETIKKRCNIKTSAVGLTTQELAEELISNER ADLVALGRELLRNPYVVLHTYTSKEDWPKQYERAFKK
22	<i>Enterobacter cloacae</i>	AA838683.1	MSAEKLTPLKVGAVTAPNRVFMAPLTRLRSIEPGDIPTPLMGEYRQRASAGLIIEAT QISQAQKGYAGAPGLHSPECIAAWKKITAGVHAEDGRIAVQLWHTGRISHSSIQPGGOAP VSASALNANRTRSLRDEINGNAIRVDTTTPRALEDEIPGIVNDFROAVANAREAGFDLVE LHSAHGYLHQFLSPSSNQRTDQYGGSVENRRLVLEVDVAVCNESADRIGIRVSPIGT FQNVNNGPNEEADALYIEELAKRGIAYLHMSETDLAGKPYSEAFRQKVRERFHGVIIIG AGAYTAEKAEDLIGKGLIDAVAFGRDYIANPDLVARLCKKAEINPQRPESFYGGGAEGYT DYPST
23	<i>Fusobacterium nucleatum subsp. nucleatum</i>	AA193968.1	MKSLRLRMSSDAHYGNLVGARMLOLQFDVATELLIQLDGGDEGLFKAYDSVEFMAPV FAGDYIEAGEIVNVGSSRRKMVFEARKVIVPRPDISDSDAADVLAEPVVCRAVTCVTP KDKQRGKK
24	<i>Acidaminococcus fermentans</i>	CAA42196.1	MSIYTLGIDVGSTASKCILKDGKEIVAKSLVAVGTGTSGPARSISEVLENAHMKKEDMA FTLATGYGRNSLEGIADKQMSSELSCHAMGASFIWPNVHTVIDGGQDVKVHVENGMTMN FQVNDKCAAGTGRFLDVMANILEVKVSDLAELGAKTKRVAISSTCTVFAESEVISQLSK GTDKIDHAGIHRVSVSRVIGLANRVGIVKDVVMTGGVAQNYGVRGALFEGLGVEIKTSP LAQYNGALGAALVAYKKAOK
25	<i>Clostridium symbiosum</i>	AA031677.1 & AA031675.1	MSINALLDEFKVAATPKQQLAEYKAQKKVIGVLPYPAPEELVYAAGMVPMGIWGSNNK TISRKEYCATFYCTIAQLALEMLLDGTMQDLDGHIPTICDTRPMSQNFVAMGDKMA VIFLAQPQNFEDFGLOFSDQYTNVKKLEKAVAGKEITNEAIQDAIKVYKSRARRKF VELASAHCDVITPTKRSVILKSFVMEKPEYIEKLEELNAELEKLPVCDWQGTQVVTSGI ICDNPKLEIFEENNAIAADDVGHESKSRVDAPEDEADALMALAKQFANMDYDVLVLYD PKSTENRRRGEFANMVKESGAQGLVLFMQQFCDFEEMEPYLLKALNNAIPHIKLIGIDQ QMRDFGQASTAIOAFADVLEMQK
			MSGIYTLGIDVGSTASKCIVLKDGEIVAKSLIDVGAGTSGPQRAIEAVLNEAGMKKEDM AYTLATGYGRNLSMDGIADKQMSSELSCHAKGATFLFPNVHTVIDGGQDVKVHVENGAM TNFQVNDKCAAGTGRFLDVMARVLEVKVEDLGRLGAMSRKVKVGSISSTCTVFAESEVISQL AMGTDKCDIIDGIHRVVAHRVTGLAHRIGVVPVDMTGGVAQNEGVYKALQDELGCPINT SPLTOYNGALGAALLAWQAASRRQSN

FIG. 20I

SEQ ID NO	Organism	GENBANK reference	Amino acid sequence
26	<i>Bacillus subtilis</i>	AA72059.1	MNKKWYKPKRHWKEIELWQVPEEKWVNDWLVQLTHTVRLDLDLKRKVINLTDEEEGVRS TKTIPLNITPYASLMDPDNPRCPVRMQSVPLSEEMHKTKYDLEDPLHEDESPVPLGLTH RYPDRVFLVLTNQCSCMYCRYCTRRRFRSGQIGMGVPRKQLDAAIAYIRETPEIRDCLSGG DGLINDQILEYILKELRSHIPLEVRIGTRAPVVFQRTDHLCEILKRYHPVWLNTHF NTSIEMTEESVEACEKLVNAGVPGVQAVLAGINDSVPMKLMHDLVKIRVRPYIYQ CDLSEGIGHFRAPVSKGLEIEGLRHTSGYAVPTFVDAPGGGKIALQPNVYLSQSPD KVILRNFEGVHTSYPEPENYIPNQADAYFESVFPETADKKKEPIGLSAIFADKEVSTPEN VDRIKRREAYIANPEHETLDRRREKRDQKKEKFLAQOKKQKETECCGGDSS MYTMGLDIGSTASKGVILKNGEDIVASETISSTGTTGSPRVLEKLYGKTGLAREDIKVV VVTGYGRMNVSDADKQISELSCHARGVNFIPETRTIIDIGGQDAKVKLDNNGRLLNPL MNDKCAAGTGRFLDVMAKIHVEVSELSISMSNSQNEVSISSTCTVFAESEVISHSENA KIEDIVAGIHTSVAKRVSSLVKRVQVQVNVVGGVARNSGIVRAMAREINTEIIVPDIP QLTGALGAALYAFDEAKESQKEVKNI
27	<i>Peptoclostridium difficile</i>	AAV40818.1	MSEKKEARVINDLLAEQYANAFKAKEGRPVGWSTSVFPQELAEVFDLNVLYPENQAAG VAAKGSLCEIAESKGSYDLCAYARTNFGLENGGCEALDMPAPDFLLCCNNICNOV IKWYENISRELDIPLIMIDTTFINNEDEVTSRDIYKACFEAAIKLEISGKFKDPKFF EEVMKISAEENGRWLKYSMSLIPADSSPDMNGFDLFTYMAVVCARGKKTTEAFKLLIEE LEDNMTGKSSFRGEERYIMMEGIPCPWYGYKMKTLAKFGVNMVTSVYPHAWALQVEV NDLDGMAYAVSTMFNVNLDRTMTRKRVDSLVGKCDGAFYHMNRSCKLSLIQYEMQORRA AEETGLPYAGFDGDDQADPRAFNAQFETRIQGLVEVMEERKLNIRGEI
28	<i>Peptoclostridium difficile</i>	AAV40819.1 & AAV40820.1	MEAILS KMKEVVENPNAAVKRYKSETGKKAIGCFVYCEEIHAAGMLPVGWGGQTEL DLAKQYFPAFACSIMQSCLEYGLKAYDELGVIPIGMCDTULCGQNWKSAPVPHIKYIS LVHPQNRKLEAGVKYLISEYKGVKRELEECGYEIEEAKIHESIEVYNEHRKTMRDFVEV AYKHSNTIKPSIRSLVIKSGFFMRKEEHELVKDLIAKLNAMPPEEVCSGKKVLLTGILAD SKDILDLEDNNISVADDLAQETRQFRDVPAGDDALERLARQWSNIEGCSLAYDPKKK RGSLLVDEVKDKDIDGVIFCMMKFCDEEYDPLVRKDIEDSGIPTLYVEIDQQQTQNEQ ARTRIQTFAEEMMSLA
29	<i>Escherichia coli</i>	AAA23833.1	MDQKLTDFRSELDLDRFGAKAISTIAESKRFPLEMRDDVAFQIINDELYLDGNARQNI ATFCQTWDDENVHKLMDLSINKNWDKKEEYQSAADLRQVNMVADLWHFAPKNGQAVG TNTIGSSEACMLGGMAMKWRWRKRMEAAAGKPTDKPNLVCGPVQICWHKFAFYWDVREI PMRPGQLFMDPKRMIEACDENTIGVVTFGTGTYTGNVEFPQPLHDALDKFQADTIDIDM HIDAASGGFLAPFVAPDIVWDFRPLPRVKSASGKFGFLAFLGCGVWVWRDEEALPQELV FNVDYLGQGIQTFAINFSPAGQVIAQYEFRLRGREGYTKVQNASYQVAAYLADEIAKL GPYEFICTGRPDEGIPAVCFKLDGDEDPGYTLYDLSERLRLRGVQVPAFTLGGEATDIVV MRIMCRRGFEMDFAEELLEEDYKASLYLSDHPKLGIAQQNSFKHT

FIG. 20J

SEQ ID NO	Organism	GENBANK reference	Amino acid sequence
30	<i>Escherichia coli</i>	AAA23536.1	MNVIAILNHMGVYFKEEPIREHRALERLNFQIWPNDRODLLKIENNARLCGVIFDWD KYNELECEEISKMNENLPIYAFANTYSTLDVSLNLRQLQISFFEYALGAAEDIAKIKQT TDEYNTLPLLTALKFKVREGKYTFCTPGHMGGTAFQKSPVGSLSFYDFGPNIMKSDI SISVSELGSLDHSQPHKEAEQYARVFNADRSYMTNGTSTANKVGMYSAPAGSTILI DRNCHKSLTHLMMMSDVTPIYFRPNAYGILGGIPOSEFQAHATIAKRKYETPNATWPVH AVITNSTYDGLLYNTDFIKKTLQVSIHFDVSAWVPTNFSPYEGKCGMSSGGRVEGKVIY ETQSTHKLLAASFQASMIHVKGDVNEETNEAYMMHTTTSPHYGIVASTETAAMMMKGNM GKRLNGSHERAIKFRKEIKRLRTESDGVWFFDWWQPDHIDTTECWPLRSDSVWHGFKNID NEHMYLDPHKVTLTPGMKDGTMDFGIPASIVAKYLDHGHVVEKTPYNNLLFLFSIG IDKTKALLRALDFKRAFQDNLRVKNMLPSLYREDPEFVYENMRIQELAQNIHKLVVHH NLPDUMYRAFEVLPMTVMITPYAAFQKELHGMTEEVYDEMVGRINANMILPYPPGVPLVM PGEMITEESRPVLEELQMLCEIGAHYPGEETDIHGAYRQADGRYTVKVLKEESKK MSKLIASDSCPDCFTTQRECIYNESRNIDVAAIVLSLNDVTCGKLEIDATGYGIPV FIATENQERVPAEYLPRISSGVFENCESTRREFYGRQLETAASHYETQLRPPFFRALVDYVN QGNSAFDCPGHQGGEFFRRHPAGNQFVEYFGEALFRADLCNADVAMGDLLIHEGAPCIAQ QHAAKVFNAKTYFVLNGTSSNKVNLNALLTPGDVLFDRNNHKSNNHHGALLQAGATPY YLETARNPYGFIGGIDAHCFEESYRELIAEVAPORAKEARFRLAVIQLGTYDGTIYNA RQVVDKIGHLCDYILFDSAWVGYEQFIPMADCSPLILDINENDPGLVTSVHKQOAGF SQTQIHKKDSHIKGCQRYVPHKRMNNAFMHASTSPFPLFAALNINAKMHGVSGRNIM WMDCVVNGINARKLINDCQHIRPFVPELVGKPKWQSYETAQIAVDLRFQFVPGEHWHS FEGYAENQVFDPCKLLITTPGIDARNGEYAFGVPATILANFLRENGVVPEKCDNSIL FLLTPAEDMAKIQQLVALLVREKLESAPLAELPSIKQHEERYAGYTLRQLCQEMH DLYARHNVKQLOKEMFRKEHFRVSMNPNQEAANYALRGEVELVRLPDAEGRIAAEGALPY PPGVLCVVPGEIWWGGAVLYFSALLEGINLLPGFAPELQQVYIEEHDGRKQVWVCYIKPR DAQSTLLKGEKL
31	<i>Escherichia coli</i>	AAA62785.1	MINIIMGPHGVFYKDEPIKELESALVAQGFQIWPQNSVDLLKFIENPRICGVIFDWD EYSLDLCSDINQLNEYLPLYARINTHSTMDSVQDMRIMALWFFEYALGQAEIAIRMRQY TDEYLDNITPPFTKALFTYVKERKTYCTPGHMGGTAYQKSPVGSLSFYDFGPNIMKSDI SISVTELGSLDHTGPHLEAEYIARTFGAEQSYVTNGTSTSNKIVGMYAAPSGSTLLI DRNCHKSLAHLMMNDVVPVWVKPTRNALGILGGIPRREFTRDSIEEKVAATTQAQWVPH AVITNSTYDGLLYNTDWIKQLDPSIHFDVSAWVPTNFSPYEGKCGMSSGGRVAGKVIY ETQSTHKMLAALSQASLHIKGEYDEEAFNEAFMMHTTTSYFYFIVASVETAAAMLGNP GKRLNRSVERALHFRKEVQRLREESDGVWFFDWWQPDHIDTTECWPLRSDSVWHGFKNID ADHMFLDPVVKVTLTPGMDEQGNMSEEGIPAAALVAKFLDERGIVVEKTPYNNLLFLFSIG IDKTKAMGLRLGLTEFKRSYDNLNRIKNNMLPDLAEDPDFYRNMRIQDLAQGIHKLRKH DLPGLMLRAFDTLPEMIMTPHQAWQROIKGEVETIALEQLVGRVSNMILPYPPGVPLLM PGEMLTRESRTVLDLFLMLCSVGGHYPGFETDIHGAKQDEDDGYRVRVVKMAG
32	<i>Escherichia coli</i>	BAA21656.1	MINIIMGPHGVFYKDEPIKELESALVAQGFQIWPQNSVDLLKFIENPRICGVIFDWD EYSLDLCSDINQLNEYLPLYARINTHSTMDSVQDMRIMALWFFEYALGQAEIAIRMRQY TDEYLDNITPPFTKALFTYVKERKTYCTPGHMGGTAYQKSPVGSLSFYDFGPNIMKSDI SISVTELGSLDHTGPHLEAEYIARTFGAEQSYVTNGTSTSNKIVGMYAAPSGSTLLI DRNCHKSLAHLMMNDVVPVWVKPTRNALGILGGIPRREFTRDSIEEKVAATTQAQWVPH AVITNSTYDGLLYNTDWIKQLDPSIHFDVSAWVPTNFSPYEGKCGMSSGGRVAGKVIY ETQSTHKMLAALSQASLHIKGEYDEEAFNEAFMMHTTTSYFYFIVASVETAAAMLGNP GKRLNRSVERALHFRKEVQRLREESDGVWFFDWWQPDHIDTTECWPLRSDSVWHGFKNID ADHMFLDPVVKVTLTPGMDEQGNMSEEGIPAAALVAKFLDERGIVVEKTPYNNLLFLFSIG IDKTKAMGLRLGLTEFKRSYDNLNRIKNNMLPDLAEDPDFYRNMRIQDLAQGIHKLRKH DLPGLMLRAFDTLPEMIMTPHQAWQROIKGEVETIALEQLVGRVSNMILPYPPGVPLLM PGEMLTRESRTVLDLFLMLCSVGGHYPGFETDIHGAKQDEDDGYRVRVVKMAG

FIG. 20K

SEQ ID NO	Organism	GENBANK reference	Amino acid sequence
33	<i>Escherichia coli</i>	AAA83861.1	MPHSLFSTDTDLTAENLLRLPAEFGCPVWVYDAQIIRRQIAALKQFDVVVRFQAQKACSNIH ILRLMREQGVKVDVSLGEIERALAAAGYNPQTHPDDIVFTADVIDQATLERSVSELIQIPVN AGSVDMLDQLGQVSPGHRVWLRVNPFGFHGHSQKNTGGENSKHGIWYTDLPAALDVIOR HHLQLVGIHMHIGSGVDYAHLEQVCGAMVRQVIEFGQDLQASAGGGLSVPYQQGEEAVD TEHYGLWNAAREQIARHLGHPVKLEIEPGRFVAQSGVLITQVRSVKOMGSRHFVLDVA GFNDLMPAMYGSYHHISALAADGRSLEHAPTIVETVAGPLCESGDVFTQOEGGNVETRA LPEVKAGDYLVLHDTGAYGASMSSNYSRPLPEVLEFDNGQARLIRRRQTEIEELALELI
34	<i>Salmonella typhimurium</i>	CAC48239.1	MQNPYTVADYLLDRLAGCGGIGHLFGVPGDYNLQFLDHDVIDHPTLRWVGCANELNAAYAAD GYARMSGAGALLTTFGVGELSAINGIAGSYAEYVPLHIVGAPCSAAQQRGELMHHTLGD GDFFRHYRMSQAISAASAILDEQNAACFEIDRVLGEMLAARRPGYIMLPADVAKKTAIPPT QALALPVHEACQGVETAFRYHARQCLMNSRRIALADFLAGFGLRPLLQRWMAETPIAH ATLLMGKGLFDEQHPNFVGTYSAGASKEVROAIEDADRVCVGTFRFVDTLTAFTQQLP AERTLEIQPYASRIGETWVFNLPMAQAVSTLRELCLECAFAPPPTRSAGQPVRIKKGELTQ ESFWQTLQQYLKPGDIIIVDQGTAAFGAAALSPLDGAEVVLPPLWGSIGYSLPAAFGAQT ACPDRRVIIIGDGAQLTIQEMGSMILRDGQAPVILLNNDGYTVERAIHGAAQRYNDIA SWNWTQIPPALNAAQQAECWRVTOAIQLAEVLERLARPORLSFIEVMLPKADLPELLRTV TRALEARNGG



## 1

**METHODS OF PRODUCING 6-CARBON  
CHEMICALS USING  
2,6-DIAMINOPIMELATE AS PRECURSOR  
TO 2-AMINOPIMELATE**

CROSS-REFERENCE TO RELATED  
APPLICATIONS

This application claims priority to U.S. Application Ser. No. 61/993,532, filed on May 15, 2015, the disclosure of which is incorporated by reference in its entirety.

TECHNICAL FIELD

Disclosed herein are methods for biosynthesizing 2-aminopimelate in a recombinant host from 2,6-diaminopimelate using one or more of a polypeptide having 2-hydroxyacyl-CoA dehydratase activity, a polypeptide having mutase activity, a polypeptide having ammonia lyase activity, and a polypeptide having enoate reductase activity. The biosynthesized 2-aminopimelate can be enzymatically converted to a product selected from the group consisting of adipic acid, adipate semialdehyde, 6-aminohexanoic acid, 6-hydroxyhexanoic acid, caprolactam, hexamethylenediamine, and 1,6-hexanediol using, for example, one or more of a polypeptide having  $\alpha$ -oxoacid decarboxylase activity classified under EC 4.1.1.-, a polypeptide having  $\alpha$ -aminoacid decarboxylase activity classified under EC 4.1.1.-, a polypeptide having synthase activity, and a polypeptide having the activity of a dehydrogenase complex; and one or more optional polypeptides having an activity such as aldehyde dehydrogenase activity, alcohol dehydrogenase activity, CoA-transferase activity, carboxylate reductase activity,  $\alpha$ -aminotransferase activity, thioesterase activity, hydrolase activity,  $\omega$ -transaminase activity, N-acetyltransferase activity, or deacylase activity, and combinations thereof.

BACKGROUND

Nylons are polyamides which are sometimes synthesized by the condensation polymerisation of a diamine with a dicarboxylic acid. Similarly, nylons may be produced by the condensation polymerisation of lactams. A ubiquitous nylon is nylon 6,6, which is produced by reaction of hexamethylenediamine (HMD) and adipic acid. Nylon 6 is produced by a ring opening polymerisation of caprolactam. Therefore, adipic acid, hexamethylenediamine, and caprolactam are important intermediates in the production of nylons (Anton & Baird, Polyamides Fibers, Encyclopedia of Polymer Science and Technology, 2001).

Industrially, adipic acid and caprolactam are produced via air oxidation of cyclohexane. The air oxidation of cyclohexane produces, in a series of steps, a mixture of cyclohexanone (K) and cyclohexanol (A), designated as KA oil. Nitric acid oxidation of KA oil produces adipic acid (Musser, Adipic acid, Ullmann's Encyclopedia of Industrial Chemistry, 2000). Caprolactam is produced from cyclohexanone via its oxime and subsequent acid rearrangement (Fuchs, Kieczka and Moran, Caprolactam, Ullmann's Encyclopedia of Industrial Chemistry, 2000).

Industrially, hexamethylenediamine (HMD) is produced by hydrocyanation of C6 Building Block to adiponitrile, followed by hydrogenation to HMD (Herzog and Smiley, Hexamethylenediamine, Ullmann's Encyclopedia of Industrial Chemistry, 2012).

Given a reliance on petrochemical feedstocks; biotechnology offers an alternative approach via biocatalysis. Bio-

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catalysis is the use of biological catalysts, such as enzymes, to perform biochemical transformations of organic compounds.

Both bioderived feedstocks and petrochemical feedstocks are viable starting materials for the biocatalysis processes.

Accordingly, against this background, it is clear that there is a need for sustainable methods for producing adipic acid, caprolactam, 6-aminohexanoic acid, hexamethylenediamine and 1,6-hexanediol (hereafter "C6 building blocks") wherein the methods are biocatalyst-based (Jang et al., *Biotechnology & Bioengineering*, 2012, 109(10), 2437-2459).

However, no wild-type prokaryote or eukaryote naturally overproduces or excretes C6 building blocks to the extracellular environment. Nevertheless, the metabolism of adipic acid and caprolactam has been reported (Ramsay et al., *Appl. Environ. Microbiol.*, 1986, 52(1), 152-156; Kulkarni and Kanekar, *Current Microbiology*, 1998, 37, 191-194).

The dicarboxylic acid, adipic acid, is converted efficiently as a carbon source by a number of bacteria and yeasts via  $\beta$ -oxidation into central metabolites.  $\beta$ -oxidation of adipate to 3-oxoadipate facilitates further catabolism via, for example, the ortho-cleavage pathway associated with aromatic substrate degradation. The catabolism of 3-oxoadipyl-CoA to acetyl-CoA and succinyl-CoA by several bacteria and fungi has been characterised comprehensively (Harwood and Parales, *Annual Review of Microbiology*, 1996, 50, 553-590). Both adipate and 6-aminohexanoic acid are intermediates in the catabolism of caprolactam, finally degraded via 3-oxoadipyl-CoA to central metabolites.

Potential metabolic pathways have been suggested for producing adipic acid from biomass-sugar: (1) biochemically from glucose to cis,cis muconic acid via the ortho-cleavage aromatic degradation pathway, followed by chemical catalysis to adipic acid; (2) a reversible adipic acid degradation pathway via the condensation of succinyl-CoA and acetyl-CoA and (3) combining  $\beta$ -oxidation, fatty acid synthase, and  $\omega$ -oxidation. However, no information using these strategies has been reported (Jang et al., *Biotechnology & Bioengineering*, 2012, 109(10), 2437-2459).

An optimality principle states that microorganisms regulate their biochemical networks to support maximum biomass growth. Beyond the need for expressing heterologous pathways in a host organism, directing carbon flux towards C6 building blocks that serve as carbon sources rather than as biomass growth constituents, contradicts the optimality principle. For example, transferring the 1-butanol pathway from *Clostridium* species into other production strains has often fallen short by an order of magnitude compared to the production performance of native producers (Shen et al., *Appl. Environ. Microbiol.*, 2011, 77(9), 2905-2915).

The efficient synthesis of a six or seven carbon aliphatic backbone as central precursor is a key consideration in synthesizing C6 building blocks prior to forming terminal functional groups, such as carboxyl, amine or hydroxyl groups, on the C6 aliphatic backbone.

SUMMARY

This document is based, at least in part, on the discovery that it is possible to construct biochemical pathways for producing a seven carbon chain aliphatic backbone as a central precursor, which can be decarboxylated to a six carbon aliphatic backbone in which one or two functional groups, i.e., carboxyl, amine or hydroxyl, can be formed, leading to the synthesis of adipic acid, adipate semialdehyde, 6-aminohexanoic acid, 6-hydroxyhexanoate, hexam-

ethylenediamine, caprolactam, or 1,6-hexanediol (hereafter “C6 building blocks”). Adipic acid and adipate, 6-hydroxyhexanoic acid and 6-hydroxyhexanoate, and 6-aminohexanoic acid and 6-aminohexanoate are used interchangeably herein to refer to the compound in any of its neutral or ionized forms, including any salt forms thereof. It is understood by those skilled in the art that the specific form will depend on pH. These pathways, metabolic engineering, and cultivation strategies described herein use meso-2,6 diaminopimelate as a central metabolite, which can be enzymatically converted to (S) 2-aminopimelate or (R) 2-aminopimelate.

In the face of an optimality principle, surprisingly it has been discovered that appropriate non-natural pathways, feedstocks, host microorganisms, attenuation strategies to the host’s biochemical network and cultivation strategies may be combined to efficiently produce one or more C6 building blocks.

In one aspect, this document features a method of biosynthesizing 2-aminopimelate in a recombinant host. The method includes enzymatically converting 2,6-diaminopimelate to 2-aminopimelate in the host using at least one polypeptide having an activity selected from the group consisting of 2-hydroxyacyl-CoA dehydratase activity, mutase activity, ammonia lyase activity, and enoate reductase activity. In some embodiments, the method can include enzymatically converting 2,6-diaminopimelate to (S) 2-aminopimelate. In some embodiments, the method can include enzymatically converting 2,6-diaminopimelate to (R) 2-aminopimelate. The method can include using a polypeptide having 2-hydroxyacyl-CoA dehydratase activity and a polypeptide having enoate reductase activity to enzymatically convert 2,6-diaminopimelate to 2-aminopimelate. The polypeptide having 2-hydroxyacyl-CoA dehydratase activity can have at least 70%, at least 80%, or at least 90% sequence identity to the amino acid sequence set forth in SEQ ID NO: 25 or SEQ ID NO: 28. The polypeptide having enoate reductase activity can have at least 70%, at least 80%, or at least 90% sequence identity to the amino acid sequence set forth in any one of SEQ ID NOs: 16-22. The method can include using a polypeptide having mutase activity, a polypeptide having ammonia lyase activity, a said polypeptide having enoate reductase activity to enzymatically convert 2,6-diaminopimelate to 2-aminopimelate. The polypeptide having ammonia lyase activity can have at least 70%, at least 80%, or at least 90% sequence identity to the amino acid sequence set forth in SEQ ID NO: 23. The polypeptide having mutase activity has at least 70%, at least 80%, or at least 90% sequence identity to the amino acid sequence set forth in SEQ ID NO: 26.

The method disclosed can further include using at least one polypeptide having an activity selected from the group consisting of diaminopimelate dehydrogenase activity, 2-hydroxycarboxylate dehydrogenase activity, CoA-transferase activity, 2-hydroxyacid dehydratase activity, and carboxylate reductase activity to enzymatically convert 2,6-diaminopimelate to 2-aminopimelate. The methods disclosed can further include using at least one polypeptide having an activity selected from the group consisting of CoA ligase activity, CoA-transferase activity, carboxylate reductase activity, and aldehyde dehydrogenase activity to enzymatically convert 2,6-diaminopimelate to 2-aminopimelate.

In some embodiments, the central precursor comprises a C7 aliphatic backbone such as (S)-2-aminopimelate or (R)-2-aminopimelate, for enzymatic conversion to one or more C6 building blocks. Such C7 aliphatic backbones can be

formed from a lysine biosynthesis precursor such as meso-2,6 diaminopimelate. See FIG. 1 and FIG. 2.

In some embodiments, a terminal carboxyl group can be enzymatically formed using a thioesterase, a CoA-transferase or CoA-ligase, or an aldehyde dehydrogenase. See FIG. 3.

In some embodiments, a terminal amine group can be enzymatically formed using an (R) alpha-aminodecarboxylase (classified, for example, under EC 4.1.1.- such as EC 4.1.1.20), (S) alpha-aminodecarboxylase (classified, for example, under EC 4.1.1.- such as EC 4.1.1.15, EC 4.1.1.17 or EC 4.1.1.18) or a transaminase (classified, for example, under EC 2.6.1.-). See FIG. 4, FIG. 5, FIG. 6, and FIG. 7.

In some embodiments, a terminal hydroxyl group can be enzymatically formed using a NADPH-specific or NADH-specific alcohol dehydrogenase. See FIG. 8.

In some embodiments, the principal carbon source fed to the fermentation derived from a biological feedstock or a non-biological feedstock

In some embodiments, the biological feedstock can be or can derive from, monosaccharides, disaccharides, lignocellulose, hemicellulose, cellulose, lignin, levulinic acid and formic acid, triglycerides, glycerol, fatty acids, agricultural waste, condensed distillers’ solubles, or municipal waste.

In some embodiments, the non-biological feedstock can be or can derive from natural gas, syngas, CO<sub>2</sub>/H<sub>2</sub>, methanol, ethanol, benzoate, non-volatile residue (NVR) or a caustic wash waste stream from cyclohexane oxidation processes, or terephthalic acid/isophthalic acid mixture waste streams.

In some embodiments, the host microorganism is a prokaryote. For example, the prokaryote can be from the bacterial genus *Escherichia* such as *Escherichia coli*; from the bacterial genus *Clostridia* such as *Clostridium ljungdahlii*, *Clostridium autoethanogenum* or *Clostridium kluyveri*; from the bacterial genus *Corynebacteria* such as *Corynebacterium glutamicum*; from the bacterial genus *Cupriavidus* such as *Cupriavidus necator* or *Cupriavidus metallidurans*; from the bacterial genus *Pseudomonas* such as *Pseudomonas fluorescens*, *Pseudomonas putida* or *Pseudomonas oleovorans*; from the bacterial genus *Delftia* such as *Delftia acidovorans*; from the bacterial genus *Bacillus* such as *Bacillus subtilis*; from the bacterial genus *Lactobacillus* such as *Lactobacillus delbrueckii*; or from the bacterial genus *Lactococcus* such as *Lactococcus lactis*. Such prokaryotes also can be a source of genes to construct recombinant host cells described herein that are capable of producing one or more C6 building blocks.

In some embodiments, the host microorganism is a eukaryote (e.g., a fungus such as a yeast). For example, the eukaryote can be from the fungus genus *Aspergillus* such as *Aspergillus niger*; from the yeast genus *Saccharomyces* such as *Saccharomyces cerevisiae*; from the yeast genus *Pichia* such as *Pichia pastoris*; from the yeast genus *Yarrowia* such as *Yarrowia lipolytica*; from the yeast genus *Issatchenkia* such as *Issatchenkia orientalis*; from the yeast genus *Debaryomyces* such as *Debaryomyces hansenii*; from the yeast genus *Arxula* such as *Arxula adeninivorans*; or from the yeast genus *Kluyveromyces* such as *Kluyveromyces lactis*. Such eukaryotes also can be a source of genes to construct recombinant host cells described herein that are capable of producing one or more C6 building blocks.

The reactions of the pathways described herein can be performed in one or more cell (e.g., host cell) strains (a) naturally expressing one or more relevant enzymes, (b) genetically engineered to express one or more relevant enzymes, or (c) naturally expressing one or more relevant

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enzymes and genetically engineered to express one or more relevant enzymes. Alternatively, relevant enzymes can be extracted from any of the above types of host cells and used in a purified or semi-purified form. Extracted enzymes can optionally be immobilized to a solid substrate such as the floors and/or walls of appropriate reaction vessels. Moreover, such extracts include lysates (e.g. cell lysates) that can be used as sources of relevant enzymes. In the methods provided by the document, all the steps can be performed in cells (e.g., host cells), all the steps can be performed using extracted enzymes, or some of the steps can be performed in cells and others can be performed using extracted enzymes.

Many of the enzymes described herein catalyze reversible reactions, and the reaction of interest may be the reverse of the described reaction. The schematic pathways shown in FIGS. 1-8 illustrate the reaction of interest for each of the intermediates.

In some embodiments, the host microorganism's tolerance to high concentrations of a C6 building block is improved through continuous cultivation in a selective environment.

In some embodiments, the host microorganism's biochemical network is attenuated or augmented to (1) ensure the intracellular availability of oxaloacetate, (2) create an NADPH imbalance that may only be balanced via the formation of one or more C6 building blocks, (3) prevent degradation of central metabolites or central precursors leading to and including C6 building blocks and (4) ensure efficient efflux from the cell.

In some embodiments, the cultivation strategy entails either achieving an aerobic or micro-aerobic cultivation condition.

In some embodiments, the cultivation strategy entails nutrient limitation either via nitrogen, phosphate or oxygen limitation.

In some embodiments, the cultivation strategy entails preventing the incorporation of fatty acids into lipid bodies or other carbon storage units.

In some embodiments, one or more C6 building blocks are produced by a single type of microorganism, e.g., a recombinant host containing one or more exogenous nucleic acids, using, for example, a fermentation strategy.

In some aspects, the methods disclosed further comprising enzymatically converting 2-aminopimelate to a product selected from the group consisting of adipic acid, adipate semialdehyde, 6-aminohexanoic acid, 6-hydroxyhexanoic acid, caprolactam, hexamethylenediamine, and 1,6-hexanediol. The method includes enzymatically converting 2-aminopimelate to one or more of said products using (i) at least one polypeptide having an activity selected from the group consisting of  $\alpha$ -oxoacid decarboxylase activity classified under EC 4.1.1.-,  $\alpha$ -aminoacid decarboxylase activity classified under EC 4.1.1.-, synthase activity, and activity of a dehydrogenase complex; and (ii) one or more optional polypeptides having an activity selected from the group consisting of aldehyde dehydrogenase activity, alcohol dehydrogenase activity, CoA-transferase activity, carboxylate reductase activity,  $\alpha$ -aminotransferase activity, thioesterase activity, hydrolase activity,  $\omega$ -transaminase activity, N-acetyltransferase activity, and deacylase activity. The polypeptide having  $\alpha$ -oxoacid decarboxylase activity can be classified under EC 4.1.1.43, EC 4.1.1.71, EC 4.1.1.72 or EC 4.1.1.74. The polypeptide having  $\alpha$ -aminoacid decarboxylase activity can be classified under EC 4.1.1.15, EC 4.1.1.17, EC 4.1.1.18, EC 4.1.1.19. The polypeptide having synthase activity is classified under EC 2.2.1.6, or the

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polypeptide having the activity of a dehydrogenase complex comprises activities can be classified under EC 1.2.4.2, EC 1.8.1.4 and EC 2.3.1.61.

For example, the methods disclosed herein further can include enzymatically converting 2-aminopimelate to adipic acid using at least one polypeptide having an activity selected from the group consisting of  $\alpha$ -aminotransferase activity, 2-oxoacid decarboxylase activity, synthase activity, dehydrogenase complex activity, thioesterase activity, CoA-transferase activity, CoA-ligase activity, and aldehyde dehydrogenase activity.

For example, the methods disclosed herein further can include enzymatically converting 2-aminopimelate to adipate semialdehyde using at least one polypeptide having an activity selected from the group consisting of  $\alpha$ -aminotransferase activity, 2-oxoacid decarboxylase activity, and synthase activity.

For example, the methods disclosed herein further can include enzymatically converting 2-aminopimelate to 6-aminohexanoic acid using a polypeptide having  $\alpha$ -aminoacid decarboxylase activity.

For example, the methods disclosed herein further can include enzymatically converting adipate semialdehyde to 6-aminohexanoic from using a  $\omega$ -transaminase. The methods can further include biosynthesizing caprolactam from 6-aminohexanoic acid using a polypeptide having the activity of a hydrolase.

For example, the methods disclosed herein further can include enzymatically converting 6-aminohexanoic acid to hexamethylenediamine from using at least one polypeptide having an activity selected from the group consisting of carboxylate reductase activity, N-acetyltransferase activity,  $\omega$ -transaminase activity, and deacylase activity.

For example, the method further can include enzymatically converting adipate semialdehyde to hexamethylenediamine using at least one polypeptide having an activity selected from the group consisting of carboxylate reductase activity and  $\omega$ -transaminase activity.

For example, the methods disclosed herein further can include enzymatically converting 2-aminopimelate to 6-hydroxyhexanoic acid using at least one polypeptide having an activity selected from the group consisting of  $\alpha$ -aminotransferase activity, 2-oxoacid decarboxylase activity, synthase activity, and alcohol dehydrogenase activity.

For example, the methods disclosed herein further can include enzymatically converting 6-hydroxyhexanoic acid to hexamethylenediamine using at least one polypeptide having an activity selected from the group consisting of carboxylate reductase activity,  $\omega$ -transaminase activity, and alcohol dehydrogenase activity.

For example, the methods disclosed herein further can include enzymatically converting 6-hydroxyhexanoic acid to 1,6-hexanediol using a polypeptide having carboxylate reductase activity and a polypeptide having alcohol dehydrogenase activity.

The polypeptide having 2-oxoacid decarboxylase activity can have at least 70% sequence identity to the amino acid sequence set forth in SEQ ID NO:34, the polypeptide having  $\alpha$ -aminoacid decarboxylase activity can have at least 70% sequence identity to the amino acid sequence set forth in any one of SEQ ID NOs: 29-34.

The polypeptide having carboxylate reductase activity can have at least 70% sequence identity to the amino acid sequence set forth in any one of SEQ ID NOs: 3-7.

The polypeptide having  $\omega$ -transaminase activity can have at least 70% sequence identity to the amino acid sequence set forth in any one of SEQ ID NOs: 8-13.

The polypeptide having thioesterase activity can have at least 70% sequence identity to the amino acid sequence set forth in SEQ ID NO: 1 or SEQ ID NO: 2.

In some embodiments, the host comprises one or more of the following: the intracellular concentration of oxaloacetate for biosynthesis of a C6 building block is increased in the host by overexpressing recombinant genes forming oxaloacetate; wherein an imbalance in NADPH is generated that can be balanced via the formation of a C6 building block; wherein an exogenous lysine biosynthesis pathway synthesizing lysine from 2-oxoglutarate via 2-oxoadipate is introduced in a host using the meso 2,6 diaminopimelate pathway for lysine synthesis; wherein an exogenous lysine biosynthesis pathway synthesizing lysine from oxaloacetate to meso 2,6 diaminopimelate is introduced in a host using the 2-oxoadipate pathway for lysine synthesis; wherein endogenous degradation pathways of central metabolites and central precursors leading to and including C6 building blocks are attenuated in the host; or wherein the efflux of a C6 building block across the cell membrane to the extracellular media is enhanced or amplified by genetically engineering structural modifications to the cell membrane or increasing any associated transporter activity for a C6 building block.

This document also features a recombinant host that includes at least one exogenous nucleic acid encoding at least one polypeptide having an activity selected from the group consisting of 2-hydroxyacyl-CoA dehydratase activity, mutase activity, ammonia lyase activity, and enoate reductase activity, said host producing 2-aminopimelate from 2,6-diaminopimelate. For example, the recombinant host can include a polypeptide having exogenous 2-hydroxyacyl-CoA dehydratase activity and a polypeptide having enoate reductase activity. For example, the recombinant host can include a polypeptide having mutase activity, a polypeptide having ammonia lyase activity, and a polypeptide having enoate reductase activity. The polypeptide having enoate reductase activity can have at least 70% sequence identity to the amino acid sequence set forth in any one of SEQ ID NOs: 16-22. The polypeptide having 2-hydroxyacyl-CoA dehydratase activity can have at least 70% sequence identity to the amino acid sequence set forth in SEQ ID NO: 25 or SEQ ID NO: 28. The polypeptide having mutase activity can have at least 70% sequence identity to the amino acid sequence set forth in SEQ ID NO: 26. The polypeptide having ammonia lyase activity can have at least 70% sequence identity to the amino acid sequence set forth in SEQ ID NO: 23.

The host can further include at least one or more exogenous polypeptides having an activity selected from the group consisting of a) diaminopimelate dehydrogenase activity, 2-hydroxycarboxylate dehydrogenase activity, CoA-transferase activity, 2-hydroxyacid dehydratase activity, and carboxylate reductase activity; or b) CoA ligase activity, CoA-transferase activity, carboxylate reductase activity, and aldehyde dehydrogenase activity.

The host can further include at least one or more exogenous polypeptides having an activity selected from the group consisting of  $\alpha$ -oxoacid decarboxylase activity classified under EC 4.1.1.-,  $\alpha$ -aminoacid decarboxylase activity classified under EC 4.1.1.-, synthase activity, and activity of a dehydrogenase complex.

The host can further include at least one or more exogenous polypeptides having an activity selected from the group consisting of aldehyde dehydrogenase activity, alcohol dehydrogenase activity, CoA-transferase activity, carboxylate reductase activity,  $\alpha$ -aminotransferase activity,

thioesterase activity, hydrolase activity,  $\omega$ -transaminase activity, N-acetyltransferase activity, and deacylase activity, the host producing a product selected from the group consisting of adipic acid, adipate semialdehyde, 6-aminohexanoic acid, 6-hydroxyhexanoic acid, caprolactam, hexamethylenediamine, and 1,6-hexanediol.

The host can further include at least one or more exogenous polypeptides having an activity selected from the group consisting of  $\alpha$ -aminotransferase activity, 2-oxoacid decarboxylase activity, activity of a dehydrogenase complex, thioesterase activity, CoA-transferase activity, CoA-ligase activity, and aldehyde dehydrogenase activity, the host producing adipic acid.

The host can further include at least one or more exogenous polypeptides having an activity selected from the group consisting of  $\alpha$ -aminotransferase activity, 2-oxoacid decarboxylase activity, synthase activity, the host producing adipate semialdehyde.

The host can further include at least one or more exogenous polypeptides having an  $\alpha$ -aminoacid decarboxylase activity, the host producing 6-aminohexanoic acid.

A recombinant host producing 6-aminohexanoic acid can include an exogenous polypeptide having  $\omega$ -transaminase activity. A recombinant host producing 6-aminohexanoic acid further can include an exogenous polypeptide having hydrolase activity, the host producing caprolactam. The host can further include one or more of an exogenous polypeptide having carboxylate reductase activity, N-acetyltransferase activity,  $\omega$ -transaminase activity, or deacylase activity, the host producing hexamethylenediamine.

The host cell can further include at least one exogenous polypeptide having carboxylate reductase activity and/or at least one exogenous polypeptide having  $\omega$ -transaminase activity, the host producing hexamethylenediamine.

The host cell can further include at least one exogenous polypeptide having an activity selected from the group consisting of  $\alpha$ -aminotransferase activity,  $\alpha$ -oxoacid decarboxylase activity, alcohol dehydrogenase activity, or synthase activity, the host producing 6-hydroxyhexanoic acid.

The host cell can further include at least one exogenous polypeptide having an activity selected from the group consisting of carboxylate reductase activity,  $\omega$ -transaminase activity, and alcohol dehydrogenase activity, the host producing hexamethylenediamine.

The host cell can further include at an exogenous polypeptide having carboxylate reductase activity and/or an exogenous polypeptide having alcohol dehydrogenase activity, the host producing 1,6-hexanediol.

In one aspect, this document features a method for producing a bioderived 6-carbon compound. The method for producing a bioderived 6-carbon compound can include culturing or growing a recombinant host as described herein under conditions and for a sufficient period of time to produce the bioderived 6-carbon compound, wherein, optionally, the bioderived 6-carbon compound is selected from the group consisting of adipic acid, adipate semialdehyde, 6-aminohexanoic acid, 6-hydroxyhexanoic acid, caprolactam, hexamethylenediamine, 1,6-hexanediol, and combinations thereof.

In one aspect, this document features composition comprising a bioderived 6-carbon compound as described herein and a compound other than the bioderived 6-carbon compound, wherein the bioderived 6-carbon compound is selected from the group consisting of adipic acid, adipate semialdehyde, 6-aminohexanoic acid, 6-hydroxyhexanoic acid, caprolactam, hexamethylenediamine, 1,6-hexanediol,

and combinations thereof. For example, the bioderived 6-carbon compound is a cellular portion of a host cell or an organism.

This document also features a biobased polymer comprising the bioderived adipic acid, adipate semialdehyde, 6-aminohexanoic acid, 6-hydroxyhexanoic acid, caprolactam, hexamethylenediamine, 1,6-hexanediol, and combinations thereof.

This document also features a biobased resin comprising the bioderived adipic acid, adipate semialdehyde, 6-aminohexanoic acid, 6-hydroxyhexanoic acid, caprolactam, hexamethylenediamine, 1,6-hexanediol, and combinations thereof, as well as a molded product obtained by molding a biobased resin.

In another aspect, this document features a process for producing a biobased polymer that includes chemically reacting the bioderived adipic acid, adipate semialdehyde, 6-aminohexanoic acid, 6-hydroxyhexanoic acid, caprolactam, hexamethylenediamine, 1,6-hexanediol, with itself or another compound in a polymer producing reaction.

In another aspect, this document features a process for producing a biobased resin that includes chemically reacting the bioderived adipic acid, adipate semialdehyde, 6-aminohexanoic acid, 6-hydroxyhexanoic acid, caprolactam, hexamethylenediamine, 1,6-hexanediol, with itself or another compound in a resin producing reaction.

Any of the recombinant hosts described herein further can include attenuation of one or more of the following enzymes: a polyhydroxyalkanoate synthase, an acetyl-CoA thioesterase, a phosphotransacetylase forming acetate, an acetate kinase, a lactate dehydrogenase, a menaquinol-fumarate oxidoreductase, an alcohol dehydrogenase forming ethanol, a triose phosphate isomerase, a pyruvate decarboxylase, a glucose-6-phosphate isomerase, NADH-consuming transhydrogenase, an NADH-specific glutamate dehydrogenase, a NADH/NADPH-utilizing glutamate dehydrogenase, a pimeloyl-CoA dehydrogenase; an acyl-CoA dehydrogenase accepting C6 building blocks and central precursors as substrates; a butyryl-CoA dehydrogenase; or an adipyl-CoA synthetase.

Any of the recombinant hosts described herein further can overexpress one or more genes encoding: 2-hydroxyacyl-CoA dehydratase; a mutase; a CoA-ligase; an ammonia lyase; an acetyl-CoA synthetase; an enoate reductase; a 6-phosphogluconate dehydrogenase; a transketolase; a puridine nucleotide transhydrogenase; a glyceraldehyde-3P-dehydrogenase; a malic enzyme; a glucose-6-phosphate dehydrogenase; a glucose dehydrogenase; a fructose 1,6 diphosphatase; a L-alanine dehydrogenase; a L-glutamate dehydrogenase; a formate dehydrogenase; a L-glutamine synthetase; a diamine transporter; a dicarboxylate transporter; diaminopimelate dehydrogenase; 2-hydroxycarboxylate dehydrogenase, 2-hydroxyacid dehydratase, carboxylate reductase and/or a multidrug transporter.

Also, described herein is a biochemical network comprising a dehydrogenase, a CoA-transferase, a dehydratase, a reductase, a mutase, a CoA-ligase, an ammonia lyase, or a thioesterase and meso-2,6-diaminopimelate, wherein the dehydrogenase, the CoA-transferase, the dehydratase, the reductase, the mutase, the CoA-ligase, the ammonia lyase, or the thioesterase enzymatically converts the meso-2,6-diaminopimelate to 2-aminopimelate. The biochemical network can further include an  $\alpha$ -aminotransferase, wherein the aminotransferase enzymatically converts 2-aminopimelate to 2-oxo-pimelate. The biochemical network can further include a decarboxylase, a synthase, or a dehydrogenase complex, wherein the decarboxylase, the synthase, or the

dehydrogenase complex enzymatically converts 2-oxo-pimelate to adipyl-CoA or adipate semialdehyde. The biochemical network can further include a dehydrogenase, a CoA transferase, a CoA ligase, or a thioesterase, wherein the dehydrogenase, the CoA transferase, the CoA ligase, or the thioesterase enzymatically convert adipyl-CoA or adipate semialdehyde to adipic acid.

Also, described herein is a biochemical network comprising a dehydrogenase, a CoA-transferase, a dehydratase, a reductase, a mutase, a CoA-ligase, an ammonia lyase, or a thioesterase and meso-2,6-diaminopimelate, wherein the dehydrogenase, the CoA-transferase, the dehydratase, the reductase, the mutase, the CoA-ligase, the ammonia lyase, or the thioesterase enzymatically converts the meso-2,6-diaminopimelate to 2-aminopimelate. The biochemical network can further include a decarboxylase, wherein the decarboxylase enzymatically converts 2-aminopimelate to 6-aminohexanoic acid. The biochemical network can further include a hydrolase, a reductase (e.g., a carboxylate reductase), a transaminase, an N-acetyltransferase, or a deacetylase, wherein the hydrolase, the reductase, the transaminase, the N-acetyltransferase, or the deacetylase enzymatically convert 6-aminohexanoic acid into at least one of caprolactam or hexamethylenediamine.

Also, described herein is a biochemical network comprising a dehydrogenase, a CoA-transferase, a dehydratase, a reductase, a mutase, a CoA-ligase, an ammonia lyase, or a thioesterase and meso-2,6-diaminopimelate, wherein the dehydrogenase, the CoA-transferase, the dehydratase, the reductase, the mutase, the CoA-ligase, the ammonia lyase, or the thioesterase enzymatically converts the meso-2,6-diaminopimelate to 2-aminopimelate. The biochemical network can further include an aminotransferase, a synthase, a decarboxylase, or a dehydrogenase wherein the aminotransferase, the synthase, the decarboxylase, or the dehydrogenase enzymatically converts 2-aminopimelate to 6-hydroxyhexanoic acid. The biochemical network can further include a reductase (e.g., a carboxylate reductase), a transaminase, or an alcohol dehydrogenase, wherein the reductase, the transaminase, or the alcohol dehydrogenase enzymatically convert 6-hydroxyhexanoic acid into at least one of hexamethylenediamine and 1,6-hexanediol.

Also, described herein is a means for obtaining 2-aminopimelate using at least one of a dehydrogenase, a CoA-transferase, a dehydratase, a reductase, a mutase, a CoA-ligase, an ammonia lyase, or a thioesterase. The means can further include means for converting 2-aminopimelate to at least one of adipic acid, 6-aminohexanoic acid, caprolactam, hexamethylenediamine, 6-hydroxyhexanoic acid, and 1,6-hexanediol. The means can include a decarboxylase, a synthase, a dehydrogenase complex, a dehydrogenase, a CoA-transferase, a dehydratase, a reductase, a mutase, a CoA-ligase, a lyase, a thioesterase, an aminotransferase, a hydrolase, a transaminase, or an N-acetyltransferase.

Also described herein is (i) step for obtaining 2-aminopimelate using a dehydrogenase, a CoA-transferase, a dehydratase, a reductase, a mutase, a CoA-ligase, an ammonia lyase, or a thioesterase (ii) a step for obtaining adipic acid using a decarboxylase, a synthase, or a dehydrogenase complex; (iii) a step for obtaining 6-aminohexanoic acid using a decarboxylase; and (iv) a step for obtaining 6-hydroxyhexanoic acid using a at least one of a aminotransferase, a synthase, a decarboxylase, or a dehydrogenase.

In another aspect, this document features a composition comprising 2-aminopimelate and decarboxylase, a synthase, or a dehydrogenase complex. The composition can be cellular. The composition can further include a dehydrogenase,

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a CoA-transferase, a CoA-dehydratase, a dehydratase, a reductase, a mutase, a CoA-ligase, a lyase, a thioesterase, an aminotransferase, a hydrolase, a transaminase, or an N-acetyltransferase and at least one of adipic acid, 6-amino-  
5 hexanoic acid, caprolactam, hexamethylenediamine, 6-hydroxyhexanoic acid, and 1,6-hexanediol. The composition can be cellular.

Unless otherwise defined, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention pertains. Although methods and materials similar or equivalent to those described herein can be used to practice the invention, suitable methods and materials are described below. All publications, patent applications, patents, and other references mentioned herein are incorporated by reference in their entirety. In case of conflict, the present specification, including definitions, will control. In addition, the materials, methods, and examples are illustrative only and not intended to be limiting.

The details of one or more embodiments of the invention are set forth in the accompanying drawings and the description below. Other features, objects, and advantages of the invention will be apparent from the description and drawings, and from the claims. The word "comprising" in the claims may be replaced by "consisting essentially of" or with "consisting of," according to standard practice in patent law.

## DESCRIPTION OF DRAWINGS

FIG. 1 is a schematic of an exemplary biochemical pathway leading to biosynthesis of (S) 2-aminopimelate using meso-2,6-diaminopimelate as a central metabolite.

FIG. 2 is a schematic of an exemplary biochemical pathway leading to the biosynthesis of (R) 2-aminopimelate using meso-2,6-diaminopimelate as a central metabolite.

FIG. 3 is a schematic of exemplary biochemical pathways leading to adipic acid using either (S) 2-aminopimelate or (R) 2-aminopimelate as a central precursor.

FIG. 4 is a schematic of exemplary biochemical pathways leading to 6-aminohexanoic acid using either (S) 2-aminopimelate, (R) 2-aminopimelate or adipate semialdehyde as a central precursor. FIG. 4 also contains a schematic of an exemplary biochemical pathway to caprolactam from 6-aminohexanoic acid.

FIG. 5 is a schematic of exemplary biochemical pathways leading to hexamethylenediamine using 6-aminohexanoic acid or adipate semialdehyde as a central precursor.

FIG. 6 is a schematic of an exemplary biochemical pathway leading to hexamethylenediamine using 6-aminohexanoic acid as a central precursor.

FIG. 7 is a schematic of an exemplary biochemical pathway leading to hexamethylenediamine using 6-hydroxyhexanoic acid as a central precursor.

FIG. 8 is a schematic of (i) exemplary biochemical pathways leading to 6-hydroxyhexanoic acid using either (S) 2-aminopimelate or (R) 2-aminopimelate as a central precursor and (ii) exemplary biochemical pathways leading to 1,6-hexanediol using 6-hydroxyhexanoic acid as a central precursor.

FIG. 9 is a bar graph summarizing the change in absorbance at 340 nm after 20 minutes, which is a measure of the consumption of NADPH and activity of carboxylate reductases relative to the enzyme only controls (no substrate).

FIG. 10 is a bar graph of the change in absorbance at 340 nm after 20 minutes, which is a measure of the consumption

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of NADPH and the activity of carboxylate reductases for converting adipate to adipate semialdehyde relative to the empty vector control.

FIG. 11 is a bar graph of the change in absorbance at 340 nm after 20 minutes, which is a measure of the consumption of NADPH and the activity of carboxylate reductases for converting 6-hydroxyhexanoate to 6-hydroxyhexanal relative to the empty vector control.

FIG. 12 is a bar graph of the change in absorbance at 340 nm after 20 minutes, which is a measure of the consumption of NADPH and the activity of carboxylate reductases for converting N6-acetyl-6-aminohexanoate to N6-acetyl-6-aminohexanal relative to the empty vector control.

FIG. 13 is a bar graph of the change in absorbance at 340 nm after 20 minutes, which is a measure of the consumption of NADPH and activity of carboxylate reductases for converting adipate semialdehyde to hexanediol relative to the empty vector control.

FIG. 14 is a bar graph summarizing the percent conversion after 4 hours of pyruvate to L-alanine (mol/mol) as a measure of the  $\omega$ -transaminase activity of the enzyme only controls (no substrate).

FIG. 15 is a bar graph of the percent conversion after 24 hours of pyruvate to L-alanine (mol/mol) as a measure of the  $\omega$ -transaminase activity for converting 6-aminohexanoate to adipate semialdehyde relative to the empty vector control.

FIG. 16 is a bar graph of the percent conversion after 4 hours of L-alanine to pyruvate (mol/mol) as a measure of the  $\omega$ -transaminase activity for converting adipate semialdehyde to 6-aminohexanoate relative to the empty vector control.

FIG. 17 is a bar graph of the percent conversion after 4 hours of pyruvate to L-alanine (mol/mol) as a measure of the  $\omega$ -transaminase activity for converting hexamethylenediamine to 6-aminohexanal relative to the empty vector control.

FIG. 18 is a bar graph of the percent conversion after 4 hours of pyruvate to L-alanine (mol/mol) as a measure of the  $\omega$ -transaminase activity for converting N6-acetyl-1,6-diaminohexane to N6-acetyl-6-aminohexanal relative to the empty vector control.

FIG. 19 is a bar graph of the percent conversion after 4 hours of pyruvate to L-alanine (mol/mol) as a measure of the  $\omega$ -transaminase activity for converting 6-aminohexanol to 6-oxohexanol relative to the empty vector control.

FIGS. 20A-20K contains the amino acid sequences of a *Lactobacillus brevis* thioesterase (see GenBank Accession No. ABJ63754.1, SEQ ID NO: 1), an *Lactobacillus plantarum* thioesterase (see GenBank Accession No. CCC78182.1, SEQ ID NO: 2), *Mycobacterium marinum* carboxylate reductase (see Genbank Accession No. ACC40567.1, SEQ ID NO: 3), a *Mycobacterium smegmatis* carboxylate reductase (see Genbank Accession No. ABK71854.1, SEQ ID NO: 4), a *Segniliparus rugosus* carboxylate reductase (see Genbank Accession No. EFV11917.1, SEQ ID NO: 5), a *Mycobacterium massiliense* carboxylate reductase (see Genbank Accession No. EIV11143.1, SEQ ID NO: 6), a *Segniliparus rotundus* carboxylate reductase (see Genbank Accession No. ADG98140.1, SEQ ID NO: 7), a *Chromobacterium violaceum*  $\omega$ -transaminase (see Genbank Accession No. AAQ59697.1, SEQ ID NO: 8), a *Pseudomonas aeruginosa*  $\omega$ -transaminase (see Genbank Accession No. AAG08191.1, SEQ ID NO: 9), a *Pseudomonas syringae*  $\omega$ -transaminase (see Genbank Accession No. AAY39893.1, SEQ ID NO: 10), a *Rhodobacter sphaeroides*  $\omega$ -transaminase (see Genbank Accession No. ABA81135.1, SEQ ID NO: 11), an

*Escherichia coli*  $\omega$ -transaminase (see Genbank Accession No. AAA57874.1, SEQ ID NO: 12), a *Vibrio fluvialis*  $\omega$ -transaminase (see Genbank Accession No. AEA39183.1, SEQ ID NO: 13), a *Bacillus subtilis* phosphopantetheinyl transferase (see Genbank Accession No. CAA44858.1, SEQ ID NO:14), a *Nocardia* sp. NRRL 5646 phosphopantetheinyl transferase (see Genbank Accession No. ABI83656.1, SEQ ID NO:15), a *Bacillus subtilis* enoate reductase (see Genbank Accession No. BAA12619.1, SEQ ID NO: 16), a *Pseudomonas putida* enoate reductase (see Genbank Accession No. AAN66878.1, SEQ ID NO: 17), a *Kluyveromyces lactis* enoate reductase (see Genbank Accession No. AAA98815.1, SEQ ID NO: 18), a *Lactobacillus casei* enoate reductase (see Genbank Accession No. AGP69310.1, SEQ ID NO: 19), a *Saccharomyces pastorianus* enoate reductase (see Genbank Accession No. CAA37666.1, SEQ ID NO: 20), a *Thermoanaerobacter pseudethanolicus* enoate reductase (see Genbank Accession No. ABY93685.1, SEQ ID NO: 21), an *Enterobacter cloacae* enoate reductase (see Genbank Accession No. AAB38683.1, SEQ ID NO: 22), a *Fusobacterium nucleatum* ammonia lyase (see Genbank Accession No. AAL93968.1, SEQ ID NO: 23), an *Acidaminococcus fermentans* 2-hydroxyglutaryl-CoA dehydratase activator (see Genbank Accession No. CAA42196.1, SEQ ID NO: 24), a *Clostridium symbiosum* 2-hydroxyglutaryl-CoA dehydratase (see Genbank Accession No. AAD31677.1 & AAD31675.1, SEQ ID NO: 25), a *Bacillus subtilis* aminomutase (see Genbank Accession No. AAB72069.1, SEQ ID NO: 26), a *Peptoclostridium difficile* 2-Hydroxyisocaproyl-CoA dehydratase activator (see Genbank Accession No. AAV40818.1, SEQ ID NO: 27), a *Peptoclostridium difficile* 2-Hydroxyisocaproyl-CoA dehydratase (see Genbank Accession No. AAV40819.1 & AAV40820.1, SEQ ID NO: 28), an *Escherichia coli* glutamate decarboxylase (see Genbank Accession No. AAA23833.1, SEQ ID NO: 29), an *Escherichia coli* lysine decarboxylase (see Genbank Accession No. AAA23536.1, SEQ ID NO: 30), an *Escherichia coli* ornithine decarboxylase (see Genbank Accession No. AAA62785.1, SEQ ID NO: 31), an *Escherichia coli* lysine decarboxylase (see Genbank Accession No. BAA21656.1, SEQ ID NO: 32), an *Escherichia coli* diaminopimelate decarboxylase (see Genbank Accession No. AAA83861.1, SEQ ID NO: 33), and a *Salmonella typhimurium* indole-3-pyruvate decarboxylase (see Genbank Accession NO. CAC48239.1, SEQ ID: 34).

#### DETAILED DESCRIPTION

Described herein are enzymes, non-natural pathways, cultivation strategies, feedstocks, host microorganisms and attenuations to the host's biochemical network, which generates a seven carbon chain aliphatic backbone from central metabolites which can be decarboxylated to a six carbon aliphatic backbone into which one or two terminal functional groups may be formed leading to the synthesis of adipic acid, adipate semialdehyde, caprolactam, 6-amino-hexanoic acid, 6-hydroxyhexanoic acid, hexamethylenediamine or 1,6-hexanediol (referred to as "C6 building blocks" herein). As used herein, the term "central precursor" is used to denote any metabolite in any metabolic pathway shown herein leading to the synthesis of one or more C6 building blocks. The term "central metabolite" is used herein to denote a metabolite that is produced in all microorganisms to support growth.

Host microorganisms described herein can include endogenous pathways that can be manipulated such that one or more C6 building blocks can be produced. In an endogenous

pathway, the host microorganism naturally expresses all of the enzymes catalyzing the reactions within the pathway. A host microorganism containing an engineered pathway does not naturally express all of the enzymes catalyzing the reactions within the pathway but has been engineered such that all of the enzymes within the pathway are expressed in the host. Within an engineered pathway, the enzymes can be from a single source, i.e., from one species, or can be from multiple sources, i.e., different species or genera. Nucleic acids encoding the enzymes described herein have been identified from various organisms and are readily available in publicly available databases such as GenBank or EMBL. Engineered hosts can naturally express none or some (e.g., one or more, two or more, three or more, four or more, five or more, or six or more) of the enzymes of the pathways described herein. Thus, a pathway within an engineered host can include all exogenous enzymes, or can include both endogenous and exogenous enzymes. Endogenous genes of the engineered hosts also can be disrupted to prevent the formation of undesirable metabolites or prevent the loss of intermediates in the pathway through other enzymes acting on such intermediates. Engineered hosts can be referred to as recombinant hosts or recombinant host cells. Thus, as described herein recombinant hosts can include nucleic acids encoding one or more of a dehydrogenase, decarboxylase, reductase, dehydratase, CoA-transferase, CoA-ligase, thioesterase, hydrolase, ammonia lyase, mutase, synthase, aminotransferase, or transaminase as described in more detail below.

The term "exogenous" as used herein with reference to a nucleic acid (or a protein) and a host refers to a nucleic acid that does not occur in (and cannot be obtained from) a cell of that particular type as it is found in nature or a protein encoded by such a nucleic acid. Thus, a non-naturally-occurring nucleic acid is considered to be exogenous to a host once in the host. It is important to note that non-naturally-occurring nucleic acids can contain nucleic acid subsequences or fragments of nucleic acid sequences that are found in nature provided the nucleic acid as a whole does not exist in nature. For example, a nucleic acid molecule containing a genomic DNA sequence within an expression vector is non-naturally-occurring nucleic acid, and thus is exogenous to a host cell once introduced into the host, since that nucleic acid molecule as a whole (genomic DNA plus vector DNA) does not exist in nature. Thus, any vector, autonomously replicating plasmid, or virus (e.g., retrovirus, adenovirus, or herpes virus) that as a whole does not exist in nature is considered to be non-naturally-occurring nucleic acid. It follows that genomic DNA fragments produced by PCR or restriction endonuclease treatment as well as cDNAs are considered to be non-naturally-occurring nucleic acid since they exist as separate molecules not found in nature. It also follows that any nucleic acid containing a promoter sequence and polypeptide-encoding sequence (e.g., cDNA or genomic DNA) in an arrangement not found in nature is non-naturally-occurring nucleic acid. A nucleic acid that is naturally-occurring can be exogenous to a particular host microorganism. For example, an entire chromosome isolated from a cell of yeast x is an exogenous nucleic acid with respect to a cell of yeast y once that chromosome is introduced into a cell of yeast y.

In contrast, the term "endogenous" as used herein with reference to a nucleic acid (e.g., a gene) (or a protein) and a host refers to a nucleic acid (or protein) that does occur in (and can be obtained from) that particular host as it is found in nature. Moreover, a cell "endogenously expressing" a nucleic acid (or protein) expresses that nucleic acid (or

protein) as does a host of the same particular type as it is found in nature. Moreover, a host “endogenously producing” or that “endogenously produces” a nucleic acid, protein, or other compound produces that nucleic acid, protein, or compound as does a host of the same particular type as it is found in nature.

In some embodiments, depending on the host and the compounds produced by the host, one or more of the following polypeptides having 2-hydroxyacyl-CoA dehydratase activity, mutase activity, ammonia lyase activity, and enoate reductase activity may be expressed in the host in addition to one or more of: a polypeptide having  $\alpha$ -oxoacid decarboxylase activity, a polypeptide having  $\alpha$ -aminoacid decarboxylase activity, a polypeptide having synthase activity, a polypeptide having the activity of a dehydrogenase complex, a polypeptide having diaminopimelate dehydrogenase activity, a polypeptide having (R)-2-hydroxyisocaproate dehydrogenase activity, a polypeptide having (R)-2-hydroxyglutarate dehydrogenase activity, a polypeptide having glutaconate CoA-transferase activity, a polypeptide having 2-hydroxyisocaproyl-CoA dehydratase activity, a polypeptide having (R)-2-hydroxyglutryl-CoA dehydratase activity, a polypeptide having carboxylate reductase activity, a polypeptide having aldehyde dehydrogenase activity, a polypeptide having lysine 2, 3-aminomutase activity, a polypeptide having succinate-CoA ligase activity, a polypeptide having 3-aminobutyryl-CoA ammonia lyase activity, a polypeptide having thioesterase activity, a polypeptide having CoA-transferase activity, a polypeptide having alpha-aminotransferase activity, a polypeptide having branch-chain-2-oxoacid decarboxylase activity, a polypeptide having acetolactate synthase activity, a polypeptide having aldehyde dehydrogenase activity, a polypeptide having hydrolase activity, a polypeptide having  $\omega$ -transaminase activity, a polypeptide having N-acetyltransferase activity, a polypeptide having lysine N-acetyltransferase activity, or a polypeptide having alcohol dehydrogenase activity. In recombinant hosts expressing a carboxylate reductase, a phosphopantetheinyl transferase also can be expressed as it enhances activity of the carboxylate reductase.

For example, a recombinant host can include at least one exogenous polypeptide having an activity selected from the group consisting of 2-hydroxyacyl-CoA dehydratase activity, mutase activity, ammonia lyase activity, and enoate reductase activity and produce 2-aminopimelate from 2,6-diaminopimelate.

For example, a host can include an exogenous polypeptide having 2-hydroxyacyl-CoA dehydratase activity and an exogenous polypeptide having enoate reductase activity and produce 2-aminopimelate (e.g., (S)-aminopimelate). Such a host further can include at least one polypeptide having an activity selected from the group consisting of diaminopimelate dehydrogenase activity, 2-hydroxycarboxylate dehydrogenase activity, CoA-transferase activity, 2-hydroxyacid dehydratase activity, and carboxylate reductase activity. See, e.g., FIG. 1.

For example, a recombinant host can include (i) an exogenous polypeptide having diaminopimelate dehydrogenase activity classified, for example, under EC 1.4.1.16, (ii) an exogenous polypeptide having 2-hydroxyisocaproate dehydrogenase activity or an exogenous polypeptide having (R)-2-hydroxyglutarate dehydrogenase activity classified, for example, under EC 1.1.1.- such as EC 1.1.1.337, (iii) an exogenous polypeptide having glutaconate CoA-transferase activity classified, for example, under EC 2.8.3.12, (iv) an exogenous polypeptide having 2-hydroxyisocaproyl-CoA dehydratase activity or a polypeptide having 2-hydroxyglu-

tryl-CoA dehydratase activity classified, for example, under EC 4.2.1.-, (v) an exogenous polypeptide having carboxylate reductase activity classified, for example, under EC 1.2.99.6, (vi) an exogenous polypeptide having enoate reductase activity classified, for example, under EC 1.3.1.31 or EC 1.3.99.1, (vii) or an exogenous polypeptide having aldehyde dehydrogenase activity classified, for example, under EC 1.2.1.- such as EC 1.2.1.3 and produce (S) 2-aminopimelate. See, FIG. 1.

For example, a recombinant host can include an exogenous polypeptide having mutase activity, an exogenous polypeptide having ammonia lyase activity, and an exogenous polypeptide having enoate reductase activity and produce 2-aminopimelate (e.g., (R)-aminopimelate). Such a host further can include at least one polypeptide having an activity selected from the group consisting of CoA ligase activity, CoA-transferase activity, carboxylate reductase activity, and aldehyde dehydrogenase activity. See, FIG. 2.

For example, a recombinant host can include (i) an exogenous polypeptide having lysine 2,3-aminomutase activity classified, for example, under EC 5.4.3.2, (ii) an exogenous polypeptide having succinate-CoA ligase activity classified, for example, under EC 6.2.1.5 or a polypeptide having CoA-transferase activity classified, for example, under EC 2.8.3.-, (iii) an exogenous polypeptide having 3-aminobutyryl-CoA ammonia lyase activity classified, for example, under EC 4.3.1.14, (iv) an exogenous polypeptide having thioesterase activity classified, for example, under EC 3.1.2.- or polypeptide having CoA-transferase activity classified, for example, under EC 2.8.3.-, (v) an exogenous polypeptide having carboxylate reductase activity classified, for example, under EC 1.2.99.6, (vi) an exogenous polypeptide having enoate reductase activity classified, for example, under EC 1.3.1.31 or EC 1.6.99.1 or (vii) a polypeptide having aldehyde dehydrogenase activity classified, for example, under EC 1.2.1.- such as EC 1.2.1.3 and produce (R) 2-aminopimelate.

A recombinant host producing 2-aminopimelate also can include at least one exogenous polypeptide having an activity selected from the group consisting of  $\alpha$ -oxoacid decarboxylase activity classified under EC 4.1.1.-,  $\alpha$ -aminoacid decarboxylase activity classified under EC 4.1.1.-, synthase activity, and activity of a dehydrogenase complex. See, e.g., FIG. 3 and FIG. 4.

In some embodiments, a recombinant host producing 2-aminopimelate can include an exogenous polypeptide having 2-oxoacid decarboxylase activity classified, for example, under EC 4.1.1.- such as EC 4.1.1.43, EC 4.1.1.71, EC 4.1.1.72 or EC 4.1.1.74 or an exogenous polypeptide having acetolactate synthase activity classified, for example, under EC 2.2.1.6 and produce adipic acid. See, FIG. 3.

For example, a recombinant host producing 2-aminopimelate can include (i) an exogenous polypeptide having 2-oxoacid decarboxylase activity classified, for example, under EC 4.1.1.- such as EC 4.1.1.43, EC 4.1.1.71, EC 4.1.1.72 or EC 4.1.1.74 or an exogenous polypeptide having acetolactate synthase activity classified, for example, under EC 2.2.1.6 and (ii) an exogenous polypeptide having  $\alpha$ -aminotransferase activity classified, for example, under EC 2.6.1.- such as EC 2.6.1.39, EC 2.6.1.42 or EC 2.6.1.21 and produce adipate semialdehyde or adipic acid. See, FIG. 3.

For example, a recombinant host producing 2-aminopimelate can include (i) an exogenous polypeptide having 2-oxoacid decarboxylase activity classified, for example, under EC 4.1.1.- such as EC 4.1.1.43, EC 4.1.1.71, EC 4.1.1.72 or EC 4.1.1.74 or an exogenous polypeptide having acetolactate synthase activity classified, for example, under



EC 2.2.1.6, (ii) an exogenous polypeptide having  $\alpha$ -amino-transferase activity classified, for example, under EC 2.6.1.- such as EC 2.6.1.39, EC 2.6.1.42 or EC 2.6.1.21, and (iii) an exogenous polypeptide having aldehyde dehydrogenase activity classified, for example, under EC 1.2.1.- such as EC 1.2.1.3, EC 1.2.1.16, EC 1.2.1.20, EC 1.2.1.63 or EC 1.2.1.79 and produce adipic acid. See, FIG. 3.

In some embodiments, a recombinant host producing 2-aminopimelate can include an exogenous polypeptide having acetolactate synthase activity classified, for example, under EC 2.2.1.6 and produce adipic acid. See, FIG. 3.

For example, a recombinant host producing 2-aminopimelate can include (i) an exogenous polypeptide having acetolactate synthase activity classified, for example, under EC 2.2.1.6 and an exogenous polypeptide having  $\alpha$ -amino-transferase activity classified, for example, under EC 2.6.1.- such as EC 2.6.1.39, EC 2.6.1.42 or EC 2.6.1.21, and produce adipic acid. See, FIG. 3.

For example, a recombinant host producing 2-aminopimelate can include an exogenous polypeptide having acetolactate synthase activity classified, for example, under EC 2.2.1.6, an exogenous polypeptide having alpha-amino-transferase activity classified, for example, under EC 2.6.1.- such as EC 2.6.1.39, EC 2.6.1.42 or EC 2.6.1.21, and an exogenous polypeptide having aldehyde dehydrogenase activity classified, for example, under EC 1.2.1.- such as EC 1.2.1.3, EC 1.2.1.16, EC 1.2.1.20, EC 1.2.1.63 or EC 1.2.1.79 and produce adipic acid. See, FIG. 3.

In some embodiments, a recombinant host producing 2-aminopimelate can include an exogenous dehydrogenase complex comprised of enzyme activities classified, for example, EC 1.2.4.2, EC 1.8.1.4 or EC 2.3.1.61 and produce adipic acid. See, FIG. 3.

For example, a recombinant host producing 2-aminopimelate can include (i) an exogenous dehydrogenase complex comprised of enzyme activities classified, for example, EC 1.2.4.2, EC 1.8.1.4 or EC 2.3.1.61 and (ii) an exogenous polypeptide having  $\alpha$ -aminotransferase activity classified, for example, under EC 2.6.1.- such as EC 2.6.1.39, EC 2.6.1.42 or EC 2.6.1.21 and produce adipic acid. See, FIG. 3.

For example, a recombinant host producing 2-aminopimelate can include an exogenous dehydrogenase complex comprised of enzyme activities classified, for example, EC 1.2.4.2, EC 1.8.1.4 or EC 2.3.1.61 and an exogenous polypeptide having thioesterase activity classified, for example, under EC 3.1.2.- and produce adipic acid. See, FIG. 3.

For example, a recombinant host producing 2-aminopimelate can include an exogenous dehydrogenase complex and an exogenous polypeptide having glutaconate CoA-transferase activity classified, for example, under EC 2.8.3.12 or an exogenous polypeptide having succinate CoA-ligase activity classified, for example, under EC 6.2.1.5 and produce adipic acid. See, FIG. 3.

For example, a recombinant host producing 2-aminopimelate can include (i) an exogenous dehydrogenase complex comprised of enzyme activities classified, for example, EC 1.2.4.2, EC 1.8.1.4 or EC 2.3.1.61, (ii) an exogenous polypeptide having alpha-aminotransferase activity classified, for example, under EC 2.6.1.- such as EC 2.6.1.39, EC 2.6.1.42 or EC 2.6.1.21, and (iii) an exogenous polypeptide having thioesterase activity classified, for example, under EC 3.1.2.-, a polypeptide having CoA-ligase activity classified, for example, under EC 6.2.1.5 or a

polypeptide having CoA-transferase activity classified, for example, under EC 2.8.3.12 and produce adipic acid. See, FIG. 3.

For example, a recombinant host producing 2-aminopimelate can include an exogenous dehydrogenase complex, an exogenous polypeptide having alpha-aminotransferase activity, and an exogenous polypeptide having glutaconate CoA-transferase activity or an exogenous polypeptide having succinate CoA-ligase activity and produce adipic acid. See, FIG. 3.

In some embodiments, a recombinant host producing (S)-2-aminopimelate can include a polypeptide having decarboxylase activity classified, for example, under EC 4.1.1.- such as EC 4.1.1.15, EC 4.1.1.17, EC 4.1.1.18, EC 4.1.1.19 and produce 6-aminohexanoic acid, which can be converted to caprolactam using an exogenous polypeptide having amidohydrolase activity (classified, for example, under EC 3.5.2.-). See, FIG. 4.

In some embodiments, a recombinant host producing (R)-2-aminopimelate can include a polypeptide having decarboxylase activity classified, for example, under EC 4.1.1.- such as EC 4.1.1.20 and produce 6-aminohexanoic acid from (R)-2-aminopimelate, which can be converted to caprolactam using an exogenous polypeptide having hydrolase activity (classified, for example, under EC 3.5.2.-). See, FIG. 4.

A recombinant host producing 2-aminopimelate can include (i) an exogenous polypeptide having  $\alpha$ -aminotransferase activity classified, for example, under EC 2.6.1.- such as EC 2.6.1.21, EC 2.6.1.39 or EC 2.6.1.42 (ii) an exogenous polypeptide having decarboxylase activity classified, for example, under EC 4.1.1.- such as EC 4.1.1.43, EC 4.1.1.71, EC 4.1.1.71 or EC 4.1.1.74 or a polypeptide having acetolactate synthase activity classified, for example, under EC 2.2.1.6 and (iii) an exogenous polypeptide having  $\omega$ -transaminase activity classified, for example, under EC 2.6.1.- such as EC 2.6.1.18, EC 2.6.1.19, EC 2.6.1.48, EC 2.6.1.29, or EC 2.6.1.82 and produce 6-aminohexanoic acid. See, FIG. 4.

A recombinant host producing 6-aminohexanoic acid can further include (i) an exogenous polypeptide having carboxylate reductase activity classified, for example, under EC 1.2.99.6 (ii) an exogenous polypeptide having  $\omega$ -transaminase activity classified, for example, under EC 2.6.1.- such as EC 2.6.1.18, EC 2.6.1.19, EC 2.6.1.48, EC 2.6.1.29, or EC 2.6.1.82 and produce hexamethylenediamine. See, FIG. 5.

A recombinant host producing 2-aminopimelate can include (i) an exogenous polypeptide having  $\alpha$ -aminotransferase activity classified, for example, under EC 2.6.1.39 or EC 2.6.1.42, (ii) classified, for example, under EC 4.1.1.- such as EC 4.1.1.43, EC 4.1.1.71, EC 4.1.1.71 or EC 4.1.1.74 or a polypeptide having acetolactate synthase activity classified, for example, under EC 2.2.1.6, (iii) a polypeptide having carboxylate reductase activity classified, for example, under EC 1.2.99.6 and (iv) exogenous polypeptide having  $\omega$ -transaminase activity classified, for example, under EC 2.6.1.- such as EC 2.6.1.18, EC 2.6.1.19, EC 2.6.1.48, EC 2.6.1.29, or EC 2.6.1.82 and produce hexamethylenediamine. See, FIG. 5.

A recombinant host producing 6-aminohexanoic acid can further include (i) an exogenous polypeptide having N-acetyltransferase activity classified, for example, under EC 2.3.1.32 (ii) a polypeptide having carboxylate reductase activity classified, for example, under EC 1.2.99.6, (iii) a polypeptide having  $\omega$ -transaminase activity classified, for example, under EC 2.6.1.- such as EC 2.6.1.18, EC 2.6.1.19,

or EC 2.6.1.48, EC 2.6.1.29, or EC 2.6.1.82 and (iv) and a polypeptide having deacylase activity classified, for example, under EC 3.5.1.17 and produce hexamethylenediamine. See, FIG. 6.

In some embodiments, a recombinant host can include a polypeptide having  $\alpha$ -aminotransferase activity classified, for example, under EC 2.6.1.- such as EC 2.6.1.39, EC 2.6.1.42 or EC 2.6.1.21 and produce 6-hydroxyhexanoic acid. See, FIG. 7.

For example, a recombinant host can include (i) a polypeptide having  $\alpha$ -aminotransferase activity classified, for example, under EC 2.6.1.- such as EC 2.6.1.39, EC 2.6.1.42 or EC 2.6.1.21 and (ii) an exogenous polypeptide having 2-oxoacid decarboxylase activity classified, for example, under EC 4.1.1.- such as EC 4.1.1.43, EC 4.1.1.71, EC 4.1.1.72 or EC 4.1.1.74 or an exogenous polypeptide having acetolactate synthase activity classified, for example, under EC 2.2.1.6 and produce 6-hydroxyhexanoic acid. See, FIG. 8.

For example, a recombinant host can include (i) a polypeptide having  $\alpha$ -aminotransferase activity classified, for example, under EC 2.6.1.- such as EC 2.6.1.39, EC 2.6.1.42 or EC 2.6.1.21 and (ii) an exogenous polypeptide having 2-oxoacid decarboxylase activity classified, for example, under EC 4.1.1.- such as EC 4.1.1.43, EC 4.1.1.71, EC 4.1.1.72 or EC 4.1.1.74 or an exogenous polypeptide having acetolactate synthase activity classified, for example, under EC 2.2.1.6, (iii) and a polypeptide having alcohol dehydrogenase activity classified, for example, under EC 1.1.1.- such as EC 1.1.1.2 or EC 1.1.1.258 and produce 6-hydroxyhexanoic acid. See, FIG. 8.

A recombinant host producing 6-hydroxyhexanoic acid can further include (i) a polypeptide having carboxylate reductase activity classified, for example, under EC 1.2.99.6, (ii) a polypeptide having  $\omega$ -transaminase activity classified, for example, under EC 2.6.1.- such as EC 2.6.1.18, EC 2.6.1.19, or EC 2.6.1.48, EC 2.6.1.29, or EC 2.6.1.82, and (iii) a polypeptide having alcohol dehydrogenase activity classified, for example, under EC 1.1.1.- such as EC 1.1.1.1 and produce hexamethylenediamine. See, FIG. 7.

A recombinant host producing 6-hydroxyhexanoic acid can further include (i) an exogenous polypeptide having carboxylate reductase activity classified, for example, under EC 1.2.99.6 and (ii) an exogenous polypeptide having alcohol dehydrogenase activity classified, for example, under EC 1.1.1.- such as EC 1.1.1.1, EC 1.1.1.2, EC 1.1.1.21 or EC 1.1.1.184 and produce 1,6 hexanediol. See, FIG. 8.

Any of the enzymes described herein that can be used for production of one or more C6 building blocks can have at least 70% sequence identity (homology) (e.g., at least 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or 100%) to the amino acid sequence of the corresponding wild-type enzyme. It will be appreciated that the sequence identity can be determined on the basis of the mature enzyme (e.g., with any signal sequence removed) or on the basis of the immature enzyme (e.g., with any signal sequence included). It also will be appreciated that the initial methionine residue may or may not be present on any of the enzyme sequences described herein.

Any of the enzymes described herein that can be used for production of one or more C6 building blocks can have at least 70% sequence identity (homology) (e.g., at least 75%, 80%, 85%, 90%, 95%, 97%, 98%, 99%, or 100%) to the amino acid sequence of the corresponding wild-type enzyme. For example, a thioesterase described herein can have at least 70% sequence identity (homology) (e.g., at least 75%, 80%, 85%, 90%, 95%, 97%, 98%, 99%, or 100%)

to the amino acid sequence of a *Lactobacillus brevis* thioesterase (see GenBank Accession No. ABJ63754.1, SEQ ID NO: 1) or to the amino acid sequence of a *Lactobacillus plantarum* thioesterase (see GenBank Accession No. CCC78182.1, SEQ ID NO: 2). See FIG. 20A.

For example, a carboxylate reductase described herein can have at least 70% sequence identity (homology) (e.g., at least 75%, 80%, 85%, 90%, 95%, 97%, 98%, 99%, or 100%) to the amino acid sequence of a *Mycobacterium marinum* (see Genbank Accession No. ACC40567.1, SEQ ID NO: 3), a *Mycobacterium smegmatis* (see Genbank Accession No. ABK71854.1, SEQ ID NO: 4), a *Segniliparus rugosus* (see Genbank Accession No. EFV11917.1, SEQ ID NO: 5), a *Mycobacterium massiliense* (see Genbank Accession No. EIV11143.1, SEQ ID NO: 6), or a *Segniliparus rotundus* (see Genbank Accession No. ADG98140.1, SEQ ID NO: 7) carboxylate reductase. See, FIGS. 20A-20E.

For example, a  $\omega$ -transaminase described herein can have at least 70% sequence identity (homology) (e.g., at least 75%, 80%, 85%, 90%, 95%, 97%, 98%, 99%, or 100%) to the amino acid sequence of a *Chromobacterium violaceum* (see Genbank Accession No. AAQ59697.1, SEQ ID NO: 8), a *Pseudomonas aeruginosa* (see Genbank Accession No. AAG08191.1, SEQ ID NO: 9), a *Pseudomonas syringae* (see Genbank Accession No. AAY39893.1, SEQ ID NO: 10), a *Rhodobacter sphaeroides* (see Genbank Accession No. ABA81135.1, SEQ ID NO: 11), an *Escherichia coli* (see Genbank Accession No. AAA57874.1, SEQ ID NO: 12), or a *Vibrio fluvialis* (see Genbank Accession No. AEA39183.1, SEQ ID NO: 13)  $\omega$ -transaminase. Some of these  $\omega$ -transaminases are diamine  $\omega$ -transaminases. See, FIG. 20E and FIG. 20F.

For example, a phosphopantetheinyl transferase described herein can have at least 70% sequence identity (homology) (e.g., at least 75%, 80%, 85%, 90%, 95%, 97%, 98%, 99%, or 100%) to the amino acid sequence of a *Bacillus subtilis* phosphopantetheinyl transferase (see Genbank Accession No. CAA44858.1, SEQ ID NO: 14) or a *Nocardia* sp. NRRL 5646 phosphopantetheinyl transferase (see Genbank Accession No. ABI83656.1, SEQ ID NO: 15). See, FIG. 20F and FIG. 20G.

For example, an enoate reductase described herein can have at least 70% sequence identity (homology) (e.g., at least 75%, 80%, 85%, 90%, 95%, 97%, 98%, 99%, or 100%) to the amino acid sequence of a *Bacillus subtilis* enoate reductase (see Genbank Accession No. BAA12619.1, SEQ ID NO: 16), a *Pseudomonas putida* enoate reductase (see Genbank Accession No. AAN66878.1, SEQ ID NO: 17), a *Kluyveromyces lactis* enoate reductase (see Genbank Accession No. AAA98815.1, SEQ ID NO: 18), a *Lactobacillus casei* enoate reductase (see Genbank Accession No. AGP69310.1, SEQ ID NO: 19), a *Saccharomyces pastorianus* enoate reductase (see Genbank Accession No. CAA37666.1, SEQ ID NO: 20), a *Thermoanaerobacter pseudethanolicus* enoate reductase (see Genbank Accession No. ABY93685.1, SEQ ID NO: 21), a *Enterobacter cloacae* enoate reductase (see Genbank Accession No. AAB38683.1, SEQ ID NO: 22).

For example, an ammonia lyase described herein can have at least 70% sequence identity (homology) (e.g., at least 75%, 80%, 85%, 90%, 95%, 97%, 98%, 99%, or 100%) to the amino acid sequence of a *Fusobacterium nucleatum* ammonia lyase (see Genbank Accession No. AAL93968.1, SEQ ID NO: 23).

For example, a dehydratase activator described herein can have at least 70% sequence identity (homology) (e.g., at least 75%, 80%, 85%, 90%, 95%, 97%, 98%, 99%, or 100%)

to the amino acid sequence of an *Acidaminococcus fermentans* 2-hydroxyglutaryl-CoA dehydratase activator (see Genbank Accession No. CAA42196.1, SEQ ID NO: 24) or a *Peptoclostridium difficile* 2-Hydroxyisocaproyl-CoA dehydratase activator (see Genbank Accession No. AAV40818.1, SEQ ID NO: 27).

For example, a 2-hydroxyacyl-CoA dehydratase described herein can have at least 70% sequence identity (homology) (e.g., at least 75%, 80%, 85%, 90%, 95%, 97%, 98%, 99%, or 100%) to the amino acid sequence of a *Clostridium symbiosum* 2-hydroxyglutaryl-CoA dehydratase (see Genbank Accession No. AAD31677.1 & AAD31675.1, SEQ ID NO: 25), or a *Peptoclostridium difficile* 2-Hydroxyisocaproyl-CoA dehydratase (see Genbank Accession No. AAV40819.1 & AAV40820.1, SEQ ID NO: 28).

For example, an aminomutase described herein can have at least 70% sequence identity (homology) (e.g., at least 75%, 80%, 85%, 90%, 95%, 97%, 98%, 99%, or 100%) to the amino acid sequence of a *Bacillus subtilis* aminomutase (see Genbank Accession No. AAB72069.1, SEQ ID NO: 26).

For example, a decarboxylase described herein can have at least 70% sequence identity (homology) (e.g., at least 75%, 80%, 85%, 90%, 95%, 97%, 98%, 99%, or 100%) to the amino acid sequence of an *Escherichia coli* glutamate decarboxylase (see Genbank Accession No. AAA23833.1, SEQ ID NO: 29), an *Escherichia coli* lysine decarboxylase (see Genbank Accession No. AAA23536.1, SEQ ID NO: 30), an *Escherichia coli* ornithine decarboxylase (see Genbank Accession No. AAA62785.1, SEQ ID NO: 31), an *Escherichia coli* lysine decarboxylase (see Genbank Accession No. BAA21656.1, SEQ ID NO: 32), an *Escherichia coli* diaminopimelate decarboxylase (see Genbank Accession No. AAA83861.1, SEQ ID NO: 33), a *Salmonella typhimurium* indole-3-pyruvate decarboxylase (see Genbank Accession No. CAC48239.1, SEQ ID NO: 34).

The percent identity (homology) between two amino acid sequences can be determined as follows. First, the amino acid sequences are aligned using the BLAST 2 Sequences (BL2seq) program from the stand-alone version of BLASTZ containing BLASTP version 2.0.14. This stand-alone version of BLASTZ can be obtained from Fish & Richardson's web site (e.g., [www.fr.com/blast/](http://www.fr.com/blast/)) or the U.S. government's National Center for Biotechnology Information web site ([www.ncbi.nlm.nih.gov](http://www.ncbi.nlm.nih.gov)). Instructions explaining how to use the BL2seq program can be found in the readme file accompanying BLASTZ. BL2seq performs a comparison between two amino acid sequences using the BLASTP algorithm. To compare two amino acid sequences, the options of BL2seq are set as follows: -i is set to a file containing the first amino acid sequence to be compared (e.g., C:\seq1.txt); -j is set to a file containing the second amino acid sequence to be compared (e.g., C:\seq2.txt); -p is set to blastp; -o is set to any desired file name (e.g., C:\output.txt); and all other options are left at their default setting. For example, the following command can be used to generate an output file containing a comparison between two amino acid sequences: C:\BL2seq -i c:\seq1.txt -j c:\seq2.txt -p blastp -o c:\output.txt. If the two compared sequences share homology (identity), then the designated output file will present those regions of homology as aligned sequences. If the two compared sequences do not share homology (identity), then the designated output file will not present aligned sequences. Similar procedures can be following for nucleic acid sequences except that blastn is used.

Once aligned, the number of matches is determined by counting the number of positions where an identical amino acid residue is presented in both sequences. The percent identity (homology) is determined by dividing the number of matches by the length of the full-length polypeptide amino acid sequence followed by multiplying the resulting value by 100. It is noted that the percent identity (homology) value is rounded to the nearest tenth. For example, 78.11, 78.12, 78.13, and 78.14 is rounded down to 78.1, while 78.15, 78.16, 78.17, 78.18, and 78.19 is rounded up to 78.2. It also is noted that the length value will always be an integer.

It will be appreciated that a number of nucleic acids can encode a polypeptide having a particular amino acid sequence. The degeneracy of the genetic code is well known to the art; i.e., for many amino acids, there is more than one nucleotide triplet that serves as the codon for the amino acid. For example, codons in the coding sequence for a given enzyme can be modified such that optimal expression in a particular species (e.g., bacteria or fungus) is obtained, using appropriate codon bias tables for that species.

Functional fragments of any of the enzymes described herein can also be used in the methods of the document. The term "functional fragment" as used herein refers to a peptide fragment of a protein that has at least 25% (e.g., at least: 30%; 40%; 50%; 60%; 70%; 75%; 80%; 85%; 90%; 95%; 98%; 99%; 100%; or even greater than 100%) of the activity of the corresponding mature, full-length, wild-type protein. The functional fragment can generally, but not always, be comprised of a continuous region of the protein, wherein the region has functional activity.

This document also provides (i) functional variants of the enzymes used in the methods of the document and (ii) functional variants of the functional fragments described above. Functional variants of the enzymes and functional fragments can contain additions, deletions, or substitutions relative to the corresponding wild-type sequences. Enzymes with substitutions will generally have not more than 50 (e.g., not more than one, two, three, four, five, six, seven, eight, nine, ten, 12, 15, 20, 25, 30, 35, 40, or 50) amino acid substitutions (e.g., conservative substitutions). This applies to any of the enzymes described herein and functional fragments. A conservative substitution is a substitution of one amino acid for another with similar characteristics. Conservative substitutions include substitutions within the following groups: valine, alanine and glycine; leucine, valine, and isoleucine; aspartic acid and glutamic acid; asparagine and glutamine; serine, cysteine, and threonine; lysine and arginine; and phenylalanine and tyrosine. The nonpolar hydrophobic amino acids include alanine, leucine, isoleucine, valine, proline, phenylalanine, tryptophan and methionine. The polar neutral amino acids include glycine, serine, threonine, cysteine, tyrosine, asparagine and glutamine. The positively charged (basic) amino acids include arginine, lysine and histidine. The negatively charged (acidic) amino acids include aspartic acid and glutamic acid. Any substitution of one member of the above-mentioned polar, basic or acidic groups by another member of the same group can be deemed a conservative substitution. By contrast, a nonconservative substitution is a substitution of one amino acid for another with dissimilar characteristics.

Deletion variants can lack one, two, three, four, five, six, seven, eight, nine, ten, 11, 12, 13, 14, 15, 16, 17, 18, 19, or 20 amino acid segments (of two or more amino acids) or non-contiguous single amino acids. Additions (addition variants) include fusion proteins containing: (a) any of the enzymes described herein or a fragment thereof; and (b) internal or terminal (C or N) irrelevant or heterologous

amino acid sequences. In the context of such fusion proteins, the term “heterologous amino acid sequences” refers to an amino acid sequence other than (a). A heterologous sequence can be, for example a sequence used for purification of the recombinant protein (e.g., FLAG, polyhistidine (e.g., hexahistidine), hemagglutinin (HA), glutathione-S-transferase (GST), or maltosebinding protein (MBP)). Heterologous sequences also can be proteins useful as detectable markers, for example, luciferase, green fluorescent protein (GFP), or chloramphenicol acetyl transferase (CAT). In some embodiments, the fusion protein contains a signal sequence from another protein. In certain host cells (e.g., yeast host cells), expression and/or secretion of the target protein can be increased through use of a heterologous signal sequence. In some embodiments, the fusion protein can contain a carrier (e.g., KLH) useful, e.g., in eliciting an immune response for antibody generation) or ER or Golgi apparatus retention signals. Heterologous sequences can be of varying length and in some cases can be a longer sequences than the full-length target proteins to which the heterologous sequences are attached

In addition, the production of one or more C6 building blocks can be performed in vitro using the isolated enzymes described herein, using a lysate (e.g., a cell lysate) from a host microorganism as a source of the enzymes, or using a plurality of lysates from different host microorganisms as the source of the enzymes.

The reactions of the pathways described herein can be performed in one or more host strains (a) naturally expressing one or more relevant enzymes, (b) genetically engineered to express one or more relevant enzymes, or (c) naturally expressing one or more relevant enzymes and genetically engineered to express one or more relevant enzymes. Alternatively, relevant enzymes can be extracted from of the above types of host cells and used in a purified or semi-purified form. Moreover, such extracts include lysates (e.g. cell lysates) that can be used as sources of relevant enzymes. In the methods provided by the document, all the steps can be performed in host cells, all the steps can be performed using extracted enzymes, or some of the steps can be performed in cells and others can be performed using extracted enzymes.

In addition, the production of one or more C6 building blocks can be performed in vitro using the isolated enzymes described herein, using a lysate (e.g., a cell lysate) from a host microorganism as a source of the enzymes, or using a plurality of lysates from different host microorganisms as the source of the enzymes.

Enzymes Generating the C7 Aliphatic Backbone for Conversion to C6 Building Blocks

In some embodiments, (S)-2-amino-6-oxopimelate in FIG. 1 is substituted with the central precursor N-Acetyl-L-2-amino-6-oxopimelate.

In some embodiments, (S)-2-amino-6-oxopimelate in FIG. 1 is substituted with the central precursor N-Succinyl-2-L-amino-6-oxoheptanedioate.

In some embodiments, the C7 aliphatic backbone can be enzymatically formed from meso-2,6-diaminopimelate using one or more of a dehydrogenase, a CoA-transferase, a dehydratase, a reductase, a mutase, a CoA-ligase, an ammonia lyase and a thioesterase. See, e.g., FIGS. 1 and 2.

In some embodiments, the dehydrogenase is a diaminopimelate dehydrogenase classified, for example, under EC 1.4.1.16.

In some embodiments, the dehydrogenase is a (R)-2-hydroxyisocaproate dehydrogenase such as the gene product of LdhA or a 2-hydroxyglutarate dehydrogenase such as the gene product of HgdH.

In some embodiments, the CoA-transferase is a glutamate CoA-transferase, classified, for example, under EC 2.8.3.12, such as the gene product of GctAB or a pimelate CoA-transferase classified, for example, under EC 2.8.3.- such as the gene product of thnH.

In some embodiments, the CoA-ligase is a succinate CoA-ligase classified, for example, under EC 6.2.1.5.

In some embodiments, the dehydratase is a 2-hydroxyisocaproyl-CoA dehydratase such as SEQ ID NO: 28 or a 2-hydroxyglutaryl-CoA dehydratase such as SEQ ID NO: 25.

In some embodiments, the thioesterase is classified, for example, under EC 3.1.2.-, such as that encoded by YciA, tesB, acot13, SEQ ID NO: 1 or SEQ ID NO: 2.

In some embodiments, the reductase is a carboxylate reductase classified, for example, under EC 1.2.99.6 such as the gene products of car & npt, GriC & GriD or SEQ ID NO: 5, 7.

In some embodiments, the reductase is an enoate reductase (old yellow enzyme) classified, for example, under EC 1.3.1.31 or EC 1.6.99.1 such as the gene product of SEQ ID NO: 16-22.

In some embodiments, the dehydrogenase is an aldehyde dehydrogenase classified, for example, under EC 1.2.1.- such as EC 1.2.1.3.

In some embodiments, the mutase is a lysine 2,3-amino-mutase classified, for example, under EC 5.4.3.2 such as SEQ ID NO: 26.

In some embodiments, the ammonia lyase is a 3-butyryl-CoA ammonia lyase classified, for example, under EC 4.3.1.14 such as SEQ ID NO: 23.

Enzymes Generating the Terminal Carboxyl Groups in the Biosynthesis of C6 Building Blocks

As depicted in FIG. 1, FIG. 2, and FIG. 3, a terminal carboxyl group can be enzymatically formed using an aldehyde dehydrogenase, a thioesterase, a CoA-transferase, or a CoA-ligase.

In some embodiments, the first terminal carboxyl group leading to the synthesis of adipic acid is enzymatically formed by an aldehyde dehydrogenase classified, for example, under EC 1.2.1.3 (Guerrillot & Vandecasteele, Eur. J. Biochem., 1977, 81, 185-192). See, e.g., FIG. 3.

In some embodiments, the second terminal carboxyl group leading to the synthesis of a C6 building block is enzymatically formed by an acyl-CoA hydrolase or thioesterase classified under EC 3.1.2.-, such as the gene product of YciA, tesB, Acot13, SEQ ID NO: 1 or SEQ ID NO: 2 (see, for example, Cantu et al., Protein Science, 2010, 19, 1281-1295; Zhuang et al., Biochemistry, 2008, 47(9), 2789-2796; or Naggert et al., Journal of Biological Chemistry, 1991, 266(17), 11044-11050, Jing et al., BMC Biochemistry, 2011, 12, 44). See, e.g., FIG. 3.

In some embodiments, the second terminal carboxyl group leading to the synthesis of a C6 building block is enzymatically formed by a CoA-transferase such as a glutamate CoA-transferase classified, for example, under EC 2.8.3.12. See, e.g., FIG. 3.

In some embodiments, the second terminal carboxyl group leading to the synthesis of a C6 building block is enzymatically formed by a reversible CoA-ligase such as succinate CoA-ligase classified under EC 6.2.1.5. See, e.g., FIG. 3.

In some embodiments, the second terminal carboxyl group leading to the synthesis of adipic acid is enzymatically formed by an aldehyde dehydrogenase classified, for example, under EC 1.2.1.63, such as the gene product of ChnE (Iwaki et al., Appl. Environ. Microbiol., 1999, 65(11), 5158-5162). See, e.g., FIG. 3.

Enzymes Generating the Terminal Amine Groups in the Biosynthesis of C6 Building Blocks

As depicted in FIG. 5, FIG. 6, and FIG. 7 a terminal amine group can be enzymatically formed using a  $\omega$ -transaminase.

In some embodiments, a terminal amine group is enzymatically formed by a  $\omega$ -transaminase classified, for example, under EC 2.6.1.-, e.g., EC 2.6.1.18, EC 2.6.1.19, EC 2.6.1.29, EC 2.6.1.48, or EC 2.6.1.82 such as that obtained from *Chromobacterium violaceum* (Genbank Accession No. AAQ59697.1), *Pseudomonas aeruginosa* (Genbank Accession No. AAG08191.1), *Pseudomonas syringae* (Genbank Accession No. AAY39893.1), *Rhodobacter sphaeroides* (Genbank Accession No. ABA81135.1), *Vibrio fluvialis* (Genbank Accession No. AEA39183.1), *Streptomyces griseus*, or *Clostridium viride*. See, FIG. 3.

An additional  $\omega$ -transaminase that can be used in the methods and hosts described herein is from *Escherichia coli* (Genbank Accession No. AAA57874.1). Some of the  $\omega$ -transaminases classified, for example, under EC 2.6.1.29 or EC 2.6.1.82 are diamine  $\omega$ -transaminases.

In some embodiments, the first terminal amine group leading to the synthesis of 6-aminohexanoic acid is enzymatically formed by a  $\omega$ -transaminase classified under EC 2.6.1.18, such as that obtained from *Vibrio fluvialis* or *Chromobacterium violaceum*, EC 2.6.1.19, such as that obtained from *Streptomyces griseus*, or EC 2.6.1.48, such as that obtained from *Clostridium viride*.

The reversible  $\omega$ -transaminase from *Chromobacterium violaceum* has demonstrated analogous activity accepting 6-aminohexanoic acid as amino donor, thus forming the first terminal amine group in adipate semialdehyde (Kaulmann et al., Enzyme and Microbial Technology, 2007, 41, 628-637).

The reversible 4-aminobutyrate: 2-oxoglutarate transaminase from *Streptomyces griseus* has demonstrated analogous activity for the conversion of 6-aminohexanoic acid to adipate semialdehyde (Yonaha et al., Eur. J. Biochem., 1985, 146, 101-106).

The reversible 5-aminovalerate transaminase from *Clostridium viride* has demonstrated analogous activity for the conversion of 6-aminohexanoic acid to adipate semialdehyde (Barker et al., The Journal of Biological Chemistry, 1987, 262(19), 8994-9003).

In some embodiments, the second terminal amine group leading to the synthesis of hexamethylenediamine is enzymatically formed by a transaminase classified under EC 2.6.1.29 or classified under EC 2.6.1.82, such as the gene product of YgjG.

The gene product of ygjG accepts a broad range of diamine carbon chain length substrates, such as putrescine, cadaverine and spermidine (Samsonova et al., BMC Microbiology, 2003, 3:2).

The diamine transaminase from *E. coli* strain B has demonstrated activity for 1,6 diaminohexane (Kim, The Journal of Chemistry, 1963, 239(3), 783-786)

Enzymes Generating the Terminal Hydroxyl Groups in the Biosynthesis of C6 Building Blocks

As depicted in FIG. 8, the terminal hydroxyl group can be enzymatically forming using an alcohol dehydrogenase.

In some embodiments, the first terminal hydroxyl group leading to the synthesis of 1,6 hexanediol is enzymatically

formed by an alcohol dehydrogenase classified, for example, under EC 1.1.1.2 such as the gene product of YMR318C or an alcohol dehydrogenase classified, for example, under EC 1.1.1.258 such as the gene product of ChnD.

In some embodiments, the second terminal hydroxyl group leading to the synthesis of 1,6 hexanediol is enzymatically formed by an alcohol dehydrogenase classified under EC 1.1.1.- (e.g., 1, 2, 21, or 184).

Biochemical Pathways

10 Pathways to (S) 2-aminopimelate and (R) 2-aminopimelate as Precursor Leading to Central Precursors to C6 Building Blocks

In some embodiments, (S) 2-aminopimelate is synthesized from the central metabolite, meso-2,6-diaminopimelate, by conversion of meso-2,6-diaminopimelate to (S)-2-amino-6-oxopimelate by a diaminopimelate dehydrogenase (classified for example under EC 1.4.1.16); followed by conversion of (S)-2-amino-6-oxopimelate to (S,R) 2-amino-6-hydroxypimelate by a (R)-2-hydroxyisocaproate dehydrogenase (classified for example under EC 1.1.1.337) such as the gene product of LdhA or a (R) 2-hydroxyglutarate dehydrogenase such as the gene product of HgdH; followed by conversion of (S,R) 2-amino-6-hydroxypimelate to (R,S) 2-hydroxy-6-aminopimeloyl-CoA by a glutaconate CoA-transferase (classified, for example, under EC 2.8.3.12) such as the gene product of GctAB; followed by conversion of (R,S) 2-hydroxy-6-aminopimeloyl-CoA to (S) 6-amino-2,3-dehydropimeloyl-CoA by a 2-hydroxyisocaproyl-CoA dehydratase such as SEQ ID NO: 28 activated SEQ ID NO: 27 or (R)-2-hydroxyglutryl-CoA dehydratase such as SEQ ID NO: 25 activated by SEQ ID NO: 24; followed by conversion of (S) 6-amino-2,3-dehydropimeloyl-CoA to (S) 6-amino-2,3-dehydropimelate by a glutaconate CoA-transferase (classified, for example, under EC 2.8.3.12); followed by conversion to (S) 2-amino-7-oxohept-6-enoate by a carboxylate reductase classified, for example, under EC 1.2.99.6) such as the gene product of car & npt, GriC & GriD or a carboxylate reductase such as SEQ ID NO: 5, 7; followed by conversion to (S) 2-amino-7-oxoheptanoate by an enoate reductase (classified, for example, under EC 1.3.1.31 or EC 1.6.99.1) such as the gene product of SEQ ID NO: 16-22; followed by conversion to (S) 2-aminopimelate by an aldehyde dehydrogenase (classified, for example, under EC 1.2.1.3). See FIG. 1.

In some embodiments, (S)-2-amino-6-oxopimelate in FIG. 1 is substituted with the central precursor N-Acetyl-L-2-amino-6-oxopimelate.

In some embodiments, (S)-2-amino-6-oxopimelate in FIG. 1 is substituted with the central precursor N-Succinyl-L-2-amino-6-oxoheptanedioate.

In some embodiments, (R) 2-aminopimelate is synthesized from the central metabolite, meso-2,6-diaminopimelate, by conversion of meso-2,6-diaminopimelate to (S,R) 3,6 diaminopimelate by a lysine 2,3-aminomutase (classified, for example, under EC 5.4.3.2) such SEQ ID NO: 26; followed by conversion of (S,R) 3,6 diaminopimelate to (S,R) 3,6 diaminopimeloyl-CoA by a succinate-CoA ligase (classified, for example, under EC 6.2.1.5); followed by conversion of (S,R) 3,6 diaminopimeloyl-CoA to (R) 6-amino-2,3-dehydropimeloyl-CoA by a 3-aminobutyryl-CoA ammonia lyase (classified, for example, under EC 4.3.1.14) such as SEQ ID NO: 23; followed by the conversion of (R) 6-amino-2,3-dehydropimeloyl-CoA to (R) 6-amino-2,3-dehydropimelate by a thioesterase (classified, for example, under EC 3.1.2.-) such as SEQ ID NO: 1-2 or the gene product of YciA, tesB or acot13 or by a CoA-transferase (classified, for example, under EC 2.8.3.-) such

as the gene product of *thnH*; followed by conversion to (R) 2-amino-7-oxohept-6-enoate by a carboxylate reductase (classified, for example, under EC 1.2.99.6) such as the gene product of *car* & *npt*, *GriC* & *GriD* or the carboxylate reductase SEQ ID NO: 5,7; followed by conversion to (R) 2-amino-7-oxoheptanoate by an enoate reductase (classified, for example, under EC 1.3.1.31) such as SEQ ID NO: 16-22; followed by conversion to (R) 2-aminopimelate by an aldehyde dehydrogenase (classified, for example, under EC 1.2.1.3). See FIG. 2.

Pathways Using (S) 2-aminopimelate or (R) 2-aminopimelate as Central Precursor to Adipic Acid

In some embodiments, adipic acid is synthesized from the central precursor (S) 2-aminopimelate or (R) 2-aminopimelate by conversion of (S) 2-aminopimelate to 2-oxopimelate by an L-specific alpha-aminotransferase (classified under EC 2.6.1.- such as EC 2.6.1.39 or EC 2.6.1.42) such as the gene product of *ilvE* or by conversion of (R) 2-aminopimelate to 2-oxopimelate by a D-specific alpha-aminotransferase (classified under EC 2.6.1.- such as EC 2.6.1.21) such as the gene product of *D-AAAT*; followed by conversion of 2-oxopimelate to adipate semialdehyde by a branch-chain-2-oxoacid decarboxylase (classified, for example, under EC 4.1.1.- such as EC 4.1.1.43, EC 4.1.1.71, EC 4.1.1.72 or EC 4.1.1.74) such as SEQ ID NO: 34 or the gene product of *kivD* or *kdca* or an acetolactate synthase (classified, for example, under EC 2.2.1.6) such as the gene product of *ilvB* & *ilvN*; followed by conversion of adipate semialdehyde to adipic acid by an aldehyde dehydrogenase (classified, for example, under EC 1.2.1.- such as EC 1.2.1.3, EC 1.2.1.16, EC 1.2.1.20, EC 1.2.1.63, EC 1.2.1.79) such as the gene product of *ChnE*, *CpnE* or *ThnG*. See FIG. 3.

In some embodiments, 2-oxopimelate obtained as described above is converted to adpyl-CoA by a dehydrogenase complex (classified, for example, under EC 1.2.4.2, EC 1.8.1.4, and EC 2.3.1.61); followed by conversion to adipic acid by a thioesterase (classified, for example, under EC 3.1.2.-) such as SEQ ID NO: 1-2 or the gene product of *YciA*, *tesB* or *acot13* or by a glutaconate CoA-transferase (classified under, for example, EC 2.8.3.12) or a reversible succinate CoA-ligase (classified, for example, under EC 6.2.1.5). See FIG. 3.

Pathway Using (R) 2-aminopimelate or (S) 2-aminopimelate as Central Precursor to 6-aminohexanoate and  $\epsilon$ -caprolactam

In some embodiments, 6-aminohexanoic acid is synthesized from the central precursor (S) 2-aminopimelate, by conversion of (S) 2-aminopimelate to 6-aminohexanoic acid by a decarboxylase (classified, for example, under EC 4.1.1.- such as EC 4.1.1.15, EC 4.1.1.17, EC 4.1.1.18 or EC 4.1.1.19) such as SEQ ID NO: 29-32. See FIG. 4.

In some embodiments, 6-aminohexanoic acid is synthesized from the central precursor (R) 2-aminopimelate by conversion of (R) 2-aminopimelate to 6-aminohexanoic acid by a decarboxylase (classified, for example, under EC 4.1.1.- such as EC 4.1.1.20) such as SEQ ID NO: 33. See FIG. 4.

In some embodiments,  $\epsilon$ -caprolactam is synthesized from the central precursor hexanoic acid by conversion of 6-aminohexanoic acid to  $\epsilon$ -caprolactam by a hydrolase (classified, for example, under EC 3.5.2.-). See FIG. 4.

In some embodiments, 6-aminohexanoic acid is synthesized from the central precursor (S) 2-aminopimelate or (R) 2-aminopimelate by conversion of (S) 2-aminopimelate to 2-oxopimelate by an L-specific alpha-aminotransferase (classified under EC 2.6.1.- such as EC 2.6.1.39 or EC 2.6.1.42) such as the gene product of *ilvE* or by conversion

of (R) 2-aminopimelate to 2-oxopimelate by a D-specific alpha-aminotransferase (classified under EC 2.6.1.- such as EC 2.6.1.21) such as the gene product of *D-AAAT*; followed by conversion of 2-oxopimelate to adipate semialdehyde by a branch-chain-2-oxoacid decarboxylase (classified, for example, under EC 4.1.1.- such as EC 4.1.1.43, EC 4.1.1.71, EC 4.1.1.72 or EC 4.1.1.74) such as SEQ ID NO: 34 or the gene product of *kivD* or *kdca* or an acetolactate synthase (classified, for example, under EC 2.2.1.6) such as the gene product of *ilvB* & *ilvN*; followed by conversion of adipate semialdehyde to 6-aminohexanoic acid by an  $\omega$ -transaminase (classified, for example, under EC 2.6.1.- such as EC 2.6.1.18, EC 2.6.1.19, or EC 2.6.1.48, EC 2.6.1.29, or EC 2.6.1.82) such as SEQ ID NO 8-13. See FIGS. 1, 2 and 4.

Pathway Using 6-aminohexanoic Acid as Central Precursor to Hexamethylenediamine

In some embodiments, hexamethylenediamine is synthesized from the central precursor, 6-aminohexanoic acid, by conversion of 6-aminohexanoic acid to 6-aminohexanal by a carboxylate reductase (classified under, for example, EC 1.2.99.6) such as the gene product of *car* alongside the gene product of *npt* or the gene product of *GriC* & *GriD* (Suzuki et al., *J. Antibiot.*, 2007, 60(6), 380-387); followed by conversion of 6-aminohexanal to hexamethylenediamine by a  $\omega$ -transaminase (classified, for example, under EC 2.6.1.18, EC 2.6.1.19, 2.6.1.48, EC 2.6.1.29, or EC 2.6.1.82) such as SEQ ID NO: 8-13. See FIG. 5.

The carboxylate reductase encoded by the gene product of *car* and enhancer *npt* has broad substrate specificity, including terminal difunctional C4 and C5 carboxylic acids (Venkatasubramanian et al., *Enzyme and Microbial Technology*, 2008, 42, 130-137).

In some embodiments, 6-aminohexanoic acid is synthesized from the central precursor (S) 2-aminopimelate or (R) 2-aminopimelate by conversion of (S) 2-aminopimelate to 2-oxopimelate by an L-specific alpha-aminotransferase (classified under EC 2.6.1.- such as EC 2.6.1.39 or EC 2.6.1.42) such as the gene product of *ilvE* or by conversion of (R) 2-aminopimelate to 2-oxopimelate by a D-specific alpha-aminotransferase (classified under EC 2.6.1.- such as EC 2.6.1.21) such as the gene product of *D-AAAT*; followed by conversion of 2-oxopimelate to adipate semialdehyde by a branch-chain-2-oxoacid decarboxylase (classified, for example, under EC 4.1.1.- such as EC 4.1.1.43, EC 4.1.1.71, EC 4.1.1.72 or EC 4.1.1.74) such as SEQ ID NO: 34 or the gene product of *kivD* or *kdca* or an acetolactate synthase (classified, for example, under EC 2.2.1.6) such as the gene product of *ilvB* & *ilvN*; followed by conversion of adipate semialdehyde to 1,6 hexanedial by a carboxylate reductase (classified, for example, under EC 1.2.99.6) such as SEQ ID NO: 7; followed by conversion of 1,6-hexanedial to 6-aminohexanal by an  $\omega$ -transaminase (classified, for example, under EC 2.6.1.- such as EC 2.6.1.18, EC 2.6.1.19, or EC 2.6.1.48, EC 2.6.1.29, or EC 2.6.1.82); followed by conversion of 6-aminohexanal to hexamethylenediamine by a  $\omega$ -transaminase (classified, for example, under EC 2.6.1.- such as EC 2.6.1.18, EC 2.6.1.19, or EC 2.6.1.48, EC 2.6.1.29, or EC 2.6.1.82) such as SEQ ID NO: 8-13. See FIGS. 1, 2 and 5.

In some embodiments, hexamethylenediamine is synthesized from the central precursor, 6-aminohexanoic acid, by conversion of 6-aminohexanoic acid to N6-acetyl-6-aminohexanoic acid by a N-acetyltransferase classified, for example, under EC 2.3.1.32; followed by conversion of N6-acetyl-6-aminohexanoic acid to N6-acetyl-6-aminohexanal by a carboxylate reductase classified, for example, under EC 1.2.99.6 such as SEQ ID NO: 5-7 or the gene

product of GriC & GriD (Suzuki et al., J. Antibiot., 2007, 60(6), 380-387); followed by conversion of N6-acetyl-6-aminohexanal to N6-acetyl-1,6-diaminohexane by a  $\omega$ -transaminase (classified, for example, under EC 2.6.1.18, EC 2.6.1.19, EC 2.6.1.29, EC 2.6.1.48 or EC 2.6.1.82) such as SEQ ID NO: 8-13; followed by conversion of N6-acetyl-1,6-diaminohexane to hexamethylenediamine by a deacetylase (classified, for example, under EC 3.5.1.17). See FIG. 6.

Pathway Using 6-hydroxyhexanoic Acid as Central Precursor to Hexamethylenediamine

In some embodiments, hexamethylenediamine is synthesized from the central precursor, 6-hydroxyhexanoic acid, by conversion of 6-hydroxyhexanoic acid to 6-hydroxyhexanal by a carboxylate reductase (classified, for example, under EC 1.2.99.6) such as SEQ ID NO: 3-7 or the gene product of car alongside the gene product of npt or the gene product of GriC & GriD (Suzuki et al., J. Antibiot., 2007, 60(6), 380-387); followed by conversion of 6-hydroxyhexanal to 1-amino-6-hydroxy-hexane by a transaminase (classified, for example, under EC 2.6.1.18, EC 2.6.1.19, EC 2.6.1.29, EC 2.6.1.48 or EC 2.6.1.82) such as SEQ ID NO: 8-13; followed by conversion of 1-amino-6-hydroxy-hexane to 6-aminohexanal by an alcohol dehydrogenase classified, for example, under EC 1.1.1.1 encoded by YMR318C, YqhD or the protein having GenBank Accession No. CAA81612.1 (from *Geobacillus stearothermophilus*); followed by conversion of 6-aminohexanal to hexamethylenediamine by a  $\omega$ -transaminase (classified, for example, under EC 2.6.1.18, EC 2.6.1.19, 2.6.1.48, EC 2.6.1.29, or EC 2.6.1.82) such as SEQ ID NO: 8-13. See FIG. 7

Pathways Using (R) 2-aminopimelate or (S) 2-aminopimelate as Central Precursor to 1,6-hexanediol

In some embodiments, adipic acid is synthesized from the central precursor (S) 2-aminopimelate or (R) 2-aminopimelate by conversion of (S) 2-aminopimelate to 2-oxopimelate by an L-specific alpha-aminotransferase (classified under EC 2.6.1.- such as EC 2.6.1.39 or EC 2.6.1.42) such as the gene product of ilvE or by conversion of (R) 2-aminopimelate to 2-oxopimelate by a D-specific alpha-aminotransferase (classified under EC 2.6.1.- such as EC 2.6.1.21) such as the gene product of D-AAAT; followed by conversion of 2-oxopimelate to adipate semialdehyde by a branch-chain-2-oxoacid decarboxylase (classified, for example, under EC 4.1.1.-such as EC 4.1.1.43, EC 4.1.1.71, EC 4.1.1.72 or EC 4.1.1.74) such as SEQ ID NO: 34 or the gene product of kivD or kdca or an acetolactate synthase (classified, for example, under EC 2.2.1.6) such as the gene product of ilvB & ilvN; followed by conversion of adipate semialdehyde to 6-hydroxyhexanoic acid by an alcohol dehydrogenase (classified, for example, under EC 1.1.1.- such as EC 1.1.1.2 or EC 1.1.1.258) such as encoded by YMR318C, ChnD, cpnD or gabD. See, FIG. 8.

In some embodiments, 1,6 hexanediol is synthesized from the central precursor 6-hydroxyhexanoic acid by conversion of 6-hydroxyhexanoic acid to 6-hydroxyhexanal by a carboxylate reductase (classified, for example, under EC 1.2.99.6) such as SEQ ID NO: 3-7; followed by conversion of 6-hydroxyhexanal to 1,6 hexanediol by an alcohol dehydrogenase (classified, for example, under EC 1.1.1.- such as EC 1.1.1.1, EC 1.1.1.2, EC 1.1.1.21, or EC 1.1.1.184) such as encoded by YMR318C, YqhD or CAA81612.1 (Liu et al., Microbiology, 2009, 155, 2078-2085).

Cultivation Strategy

In some embodiments, one or more C6 building blocks are biosynthesized in a recombinant host using anaerobic, aerobic or micro-aerobic cultivation conditions. In some

embodiments, the cultivation strategy entails nutrient limitation such as nitrogen, phosphate or oxygen limitation.

In some embodiments in which (S) 2-aminopimelate is produced as a central precursor, a cultivation strategy entails either achieving an anaerobic or micro-aerobic cultivation condition.

In some embodiments in which (R) 2-aminopimelate is produced as a central precursor, a cultivation strategy entails either achieving an anaerobic, aerobic or micro-aerobic cultivation condition.

In some embodiments, a cell retention strategy using, for example, ceramic hollow fiber membranes is employed to achieve and maintain a high cell density during either fed-batch or continuous fermentation.

In some embodiments, the cultivation strategy entails culturing under conditions of nutrient limitation either via nitrogen, phosphate or oxygen limitation.

In some embodiments, the principal carbon source fed to the fermentation in the synthesis of one or more C6 building blocks can derive from biological or non-biological feedstocks.

In some embodiments, the biological feedstock can be or can derive from, monosaccharides, disaccharides, lignocellulose, hemicellulose, cellulose, lignin, levulinic acid and formic acid, triglycerides, glycerol, fatty acids, agricultural waste, condensed distillers' solubles, or municipal waste.

The efficient catabolism of crude glycerol stemming from the production of biodiesel has been demonstrated in several microorganisms such as *Escherichia coli*, *Cupriavidus necator*, *Pseudomonas oleovorans*, *Pseudomonas putida* and *Yarrowia lipolytica* (Lee et al., *Appl. Biochem. Biotechnol.*, 2012, 166:1801-1813; Yang et al., *Biotechnology for Biofuels*, 2012, 5:13; Meijnen et al., *Appl. Microbiol. Biotechnol.*, 2011, 90:885-893).

The efficient catabolism of lignocellulosic-derived levulinic acid has been demonstrated in several organisms such as *Cupriavidus necator* and *Pseudomonas putida* in the synthesis of 3-hydroxyvalerate via the precursor propanoyl-CoA (Jaremko and Yu, 2011, supra; Martin and Prather, *J. Biotechnol.*, 2009, 139:61-67).

The efficient catabolism of lignin-derived aromatic compounds such as benzoate analogues has been demonstrated in several microorganisms such as *Pseudomonas putida*, *Cupriavidus necator* (Bugg et al., *Current Opinion in Biotechnology*, 2011, 22, 394-400; Pérez-Pantoja et al., *FEMS Microbiol. Rev.*, 2008, 32, 736-794).

The efficient utilization of agricultural waste, such as olive mill waste water has been demonstrated in several microorganisms, including *Yarrowia lipolytica* (Papanikolaou et al., *Bioresour. Technol.*, 2008, 99(7):2419-2428).

The efficient utilization of fermentable sugars such as monosaccharides and disaccharides derived from cellulosic, hemicellulosic, cane and beet molasses, cassava, corn and other agricultural sources has been demonstrated for several microorganism such as *Escherichia coli*, *Corynebacterium glutamicum* and *Lactobacillus delbrueckii* and *Lactococcus lactis* (see, e.g., Hermann et al., *J. Biotechnol.*, 2003, 104: 155-172; Wee et al., *Food Technol. Biotechnol.*, 2006, 44(2):163-172; Ohashi et al., *J. Bioscience and Bioengineering*, 1999, 87(5):647-654).

The efficient utilization of furfural, derived from a variety of agricultural lignocellulosic sources, has been demonstrated for *Cupriavidus necator* (Li et al., *Biodegradation*, 2011, 22:1215-1225).

In some embodiments, the non-biological feedstock can be or can derive from natural gas, syngas, CO<sub>2</sub>/H<sub>2</sub>, methanol, ethanol, benzoate, non-volatile residue (NVR) or a

caustic wash waste stream from cyclohexane oxidation processes, or terephthalic acid/isophthalic acid mixture waste streams.

The efficient catabolism of methanol has been demonstrated for the methylotrophic yeast *Pichia pastoris*.

The efficient catabolism of ethanol has been demonstrated for *Clostridium kluyveri* (Seedorf et al., *Proc. Natl. Acad. Sci. USA*, 2008, 105(6) 2128-2133).

The efficient catabolism of CO<sub>2</sub> and H<sub>2</sub>, which may be derived from natural gas and other chemical and petrochemical sources, has been demonstrated for *Cupriavidus necator* (Prybylski et al., *Energy, Sustainability and Society*, 2012, 2:11).

The efficient catabolism of syngas has been demonstrated for numerous microorganisms, such as *Clostridium ljungdahlii* and *Clostridium autoethanogenum* (Köpke et al., *Applied and Environmental Microbiology*, 2011, 77(15): 5467-5475).

The efficient catabolism of the non-volatile residue waste stream from cyclohexane processes has been demonstrated for numerous microorganisms, such as *Delftia acidovorans* and *Cupriavidus necator* (Ramsay et al., *Applied and Environmental Microbiology*, 1986, 52(1):152-156).

In some embodiments, the host microorganism is a prokaryote. For example, the prokaryote can be a bacterium from the genus *Escherichia* such as *Escherichia coli*; from the genus *Clostridia* such as *Clostridium ljungdahlii*, *Clostridium autoethanogenum* or *Clostridium kluyveri*; from the genus *Corynebacteria* such as *Corynebacterium glutamicum*; from the genus *Cupriavidus* such as *Cupriavidus necator* or *Cupriavidus metallidurans*; from the genus *Pseudomonas* such as *Pseudomonas fluorescens*, *Pseudomonas putida* or *Pseudomonas oleovorans*; from the genus *Delftia* such as *Delftia acidovorans*; from the genus *Bacillus* such as *Bacillus subtilis*; from the genus *Lactobacillus* such as *Lactobacillus delbrueckii*; or from the genus *Lactococcus* such as *Lactococcus lactis*. Such prokaryotes also can be a source of genes to construct recombinant host cells described herein that are capable of producing one or more C6 building blocks.

In some embodiments, the host microorganism is a eukaryote. For example, the eukaryote can be a filamentous fungus, e.g., one from the genus *Aspergillus* such as *Aspergillus niger*. Alternatively, the eukaryote can be a yeast, e.g., one from the genus *Saccharomyces* such as *Saccharomyces cerevisiae*; from the genus *Pichia* such as *Pichia pastoris*; or from the genus *Yarrowia* such as *Yarrowia lipolytica*; from the genus *Issatchenkia* such as *Issatchenkia orientalis*; from the genus *Debaryomyces* such as *Debaryomyces hansenii*; from the genus *Arxula* such as *Arxula adenoinivorans*; or from the genus *Kluyveromyces* such as *Kluyveromyces lactis*. Such eukaryotes also can be a source of genes to construct recombinant host cells described herein that are capable of producing one or more C6 building blocks.

#### Metabolic Engineering

The present document provides methods involving less than all the steps described for all the above pathways. Such methods can involve, for example, one, two, three, four, five, six, seven, eight, nine, ten, or more of such steps. Where less than all the steps are included in such a method, the first step can be any one of the steps listed.

Furthermore, recombinant hosts described herein can include any combination of the above enzymes such that one or more of the steps, e.g., one, two, three, four, five, six, seven, eight, nine, ten, or more of such steps, can be performed within a recombinant host. This document provides host cells of any of the genera and species listed and

genetically engineered to express one or more (e.g., two, three, four, five, six, seven, eight, nine, 10, 11, 12 or more) recombinant forms of any of the enzymes recited in the document. Thus, for example, the host cells can contain exogenous nucleic acids encoding enzymes catalyzing one or more of the steps of any of the pathways described herein.

In addition, this document recognizes that where enzymes have been described as accepting CoA-activated substrates, analogous enzyme activities associated with [acp]-bound substrates exist that are not necessarily in the same enzyme class.

Also, this document recognizes that where enzymes have been described accepting (R)-enantiomers of substrate, analogous enzyme activities associated with (S)-enantiomer substrates exist that are not necessarily in the same enzyme class.

This document also recognizes that where an enzyme is shown to accept a particular co-factor, such as NADPH, or co-substrate, such as acetyl-CoA, many enzymes are promiscuous in terms of accepting a number of different co-factors or co-substrates in catalyzing a particular enzyme activity. Also, this document recognizes that where enzymes have high specificity for e.g., a particular co-factor such as NADH, an enzyme with similar or identical activity that has high specificity for the co-factor NADPH may be in a different enzyme class.

In some embodiments, the enzymes in the pathways outlined in section 4.5 are the result of enzyme engineering via non-direct or rational enzyme design approaches with aims of improving activity, improving specificity, reducing feedback inhibition, reducing repression, improving enzyme solubility, changing stereo-specificity, or changing co-factor specificity.

In some embodiments, the enzymes in the pathways outlined in section 4.5 are gene dosed (i.e., overexpressed by having a plurality of copies of the gene in the host organism), into the resulting genetically modified organism via episomal or chromosomal integration approaches.

In some embodiments, genome-scale system biology techniques such as Flux Balance Analysis are utilized to devise genome scale attenuation or knockout strategies for directing carbon flux to a C6 building block.

Attenuation strategies include, but are not limited to; the use of transposons, homologous recombination (double cross-over approach), mutagenesis, enzyme inhibitors and RNA interference (RNAi).

In some embodiments, fluxomic, metabolomic and transcriptomic data are utilized to inform or support genome-scale system biology techniques, thereby devising genome scale attenuation or knockout strategies in directing carbon flux to a C6 building block.

In some embodiments, the host microorganism's tolerance to high concentrations of a C6 building block is improved through continuous cultivation in a selective environment.

In some embodiments, the host microorganism's biochemical network is attenuated or augmented to (1) ensure the intracellular availability of oxaloacetate, (2) create an NADPH imbalance that may only be balanced via the formation of one or more C6 building blocks, (3) prevent degradation of central metabolites, central precursors leading to and including C6 building blocks and (4) ensure efficient efflux from the cell.

In some embodiments, the anaplerotic reactions from glycolysis leading into the Krebs cycle to augment oxaloacetate are overexpressed in the host.



In some embodiments where the host microorganism uses the lysine biosynthesis pathway via meso-2,6-diaminopimelate, the genes encoding the synthesis of lysine from 2-oxoglutarate via 2-oxoadipate are gene dosed into the host organisms.

In some embodiments where the host microorganism uses the lysine biosynthesis pathway via 2-oxoadipate, the genes encoding the synthesis of lysine via meso-2,6-diaminopimelate are gene dosed into the host organisms.

In some embodiments, where pathways require excess NADPH co-factor in the synthesis of a C6 building block, a puridine nucleotide transhydrogenase gene such as UdhA is overexpressed in the host organisms (Brigham et al., *Advanced Biofuels and Bioproducts*, 2012, Chapter 39, 1065-1090).

In some embodiments, where pathways require excess NADPH co-factor in the synthesis of a C6 building block, a glyceraldehyde-3P-dehydrogenase gene such as GapN is overexpressed in the host organisms (Brigham et al., 2012, supra).

In some embodiments, where pathways require excess NADPH co-factor in the synthesis of a C6 building block, a gene encoding a malic enzyme, such as maeA or maeB, is overexpressed in the host (Brigham et al., 2012, supra).

In some embodiments, where pathways require excess NADPH co-factor in the synthesis of a C6 building block, a gene encoding a glucose-6-phosphate dehydrogenase such as zwf is overexpressed in the host (Lim et al., *Journal of Bioscience and Bioengineering*, 2002, 93(6), 543-549).

In some embodiments, where pathways require excess NADPH co-factor in the synthesis of a C6 building block, a gene encoding a fructose 1,6 diphosphatase such as fbp is overexpressed in the host (Becker et al., *Journal of Biotechnology*, 2007, 132, 99-109).

In some embodiments, where pathways require excess NADPH co-factor in the synthesis of a C6 building block, an endogenous gene encoding a triose phosphate isomerase (EC 5.3.1.1) is attenuated.

In some embodiments, where pathways require excess NADPH co-factor in the synthesis of a C6 building block, an endogenous gene encoding a glucose dehydrogenase such as the gene product of gdh is overexpressed in the host (Sato et al., *Journal of Bioscience and Bioengineering*, 2003, 95(4), 335-341).

In some embodiments, endogenous genes encoding enzymes facilitating the conversion of NADPH to NADH are attenuated, such as the NADH generation cycle that may be generated via inter-conversion of a glutamate dehydrogenase in EC 1.4.1.2 (NADH-specific) and EC 1.4.1.4 (NADPH-specific).

In some embodiments, an endogenous gene encoding a glutamate dehydrogenase (EC 1.4.1.3) that can utilize both NADH and NADPH as co-factors is attenuated.

In some embodiments using hosts that naturally accumulate polyhydroxyalkanoates, one or more endogenous genes encoding polymer synthase enzymes can be attenuated in the host strain.

In some embodiments,  $\beta$ -oxidation enzymes degrading central metabolites and central precursors leading to and including C6 building blocks are attenuated.

In some embodiments, enzymes activating C6 building blocks via Coenzyme A esterification such as CoA-ligases are attenuated.

In some embodiments, the efflux of a C6 building block across the cell membrane to the extracellular media is enhanced or amplified by genetically engineering structural

modifications to the cell membrane or increasing any associated transporter activity for a C6 building block.

Producing C6 Building Blocks Using a Recombinant Host

Typically, one or more C6 building blocks can be produced by providing a host microorganism and culturing the provided microorganism with a culture medium containing a suitable carbon source as described above. In general, the culture media and/or culture conditions can be such that the microorganisms grow to an adequate density and produce a C6 building block efficiently. For large-scale production processes, any method can be used such as those described elsewhere (Manual of Industrial Microbiology and Biotechnology, 2<sup>nd</sup> Edition, Editors: A. L. Demain and J. E. Davies, ASM Press; and Principles of Fermentation Technology, P. F. Stanbury and A. Whitaker, Pergamon). Briefly, a large tank (e.g., a 100 gallon, 200 gallon, 500 gallon, or more tank) containing an appropriate culture medium is inoculated with a particular microorganism. After inoculation, the microorganism is incubated to allow biomass to be produced. Once a desired biomass is reached, the broth containing the microorganisms can be transferred to a second tank. This second tank can be any size. For example, the second tank can be larger, smaller, or the same size as the first tank. Typically, the second tank is larger than the first such that additional culture medium can be added to the broth from the first tank. In addition, the culture medium within this second tank can be the same as, or different from, that used in the first tank.

Once transferred, the microorganisms can be incubated to allow for the production of a C6 building block. Once produced, any method can be used to isolate C6 building blocks. For example, C6 building blocks can be recovered selectively from the fermentation broth via adsorption processes. In the case of adipic acid and 6-aminoheptanoic acid, the resulting eluate can be further concentrated via evaporation, crystallized via evaporative and/or cooling crystallization, and the crystals recovered via centrifugation. In the case of hexamethylenediamine and 1,6-hexanediol, distillation may be employed to achieve the desired product purity.

## EXAMPLES

### Example 1

#### Enzyme Activity of $\omega$ -transaminase Using Adipate Semialdehyde as Substrate and Forming 6-aminohexanoate

A nucleotide sequence encoding a His-tag was added to the genes from *Chromobacterium violaceum*, *Pseudomonas aeruginosa*, *Pseudomonas syringae*, *Rhodobacter sphaeroides*, and *Vibrio Fluvialis* encoding the  $\omega$ -transaminases of SEQ ID NOs: 8, 9, 10, 11 and 13, respectively (see FIG. 20E and FIG. 20F) such that N-terminal HIS tagged  $\omega$ -transaminases could be produced. Each of the resulting modified genes was cloned into a pET21a expression vector under control of the T7 promoter and each expression vector was transformed into a BL21[DE3] *E. coli* host. The resulting recombinant *E. coli* strains were cultivated at 37° C. in a 250 mL shake flask culture containing 50 mL LB media and antibiotic selection pressure, with shaking at 230 rpm. Each culture was induced overnight at 16° C. using 1 mM IPTG.

The pellet from each induced shake flask culture was harvested via centrifugation. Each pellet was resuspended and lysed via sonication. The cell debris was separated from

the supernatant via centrifugation and the cell free extract was used immediately in enzyme activity assays.

Enzyme activity assays in the reverse direction (i.e., 6-aminohexanoate to adipate semialdehyde) were performed in a buffer composed of a final concentration of 50 mM HEPES buffer (pH=7.5), 10 mM 6-aminohexanoate, 10 mM pyruvate and 100  $\mu$ M pyridoxyl 5' phosphate. Each enzyme activity assay reaction was initiated by adding cell free extract of the  $\omega$ -transaminase gene product or the empty vector control to the assay buffer containing the 6-aminohexanoate and incubated at 25° C. for 24 h, with shaking at 250 rpm. The formation of L-alanine from pyruvate was quantified via RP-HPLC.

Each enzyme only control without 6-aminohexanoate demonstrated low base line conversion of pyruvate to L-alanine See FIG. 14. The gene product of SEQ ID NO 8, SEQ ID NO 10, SEQ ID NO 11 and SEQ ID NO 13 accepted 6-aminohexanoate as substrate as confirmed against the empty vector control. See FIG. 15.

Enzyme activity in the forward direction (i.e., adipate semialdehyde to 6-aminohexanoate) was confirmed for the transaminases of SEQ ID NO 8, SEQ ID NO 9, SEQ ID NO 10, SEQ ID NO 11 and SEQ ID NO 13. Enzyme activity assays were performed in a buffer composed of a final concentration of 50 mM HEPES buffer (pH=7.5), 10 mM adipate semialdehyde, 10 mM L-alanine and 100  $\mu$ M pyridoxyl 5' phosphate. Each enzyme activity assay reaction was initiated by adding a cell free extract of the  $\omega$ -transaminase gene product or the empty vector control to the assay buffer containing the adipate semialdehyde and incubated at 25° C. for 4 h, with shaking at 250 rpm. The formation of pyruvate was quantified via RP-HPLC.

The gene product of SEQ ID NO 8, SEQ ID NO 9, SEQ ID NO 10, SEQ ID NO 11 and SEQ ID NO 13 accepted adipate semialdehyde as substrate as confirmed against the empty vector control. See FIG. 16. The reversibility of the  $\omega$ -transaminase activity was confirmed, demonstrating that the  $\omega$ -transaminases of SEQ ID NO 8, SEQ ID NO 9, SEQ ID NO 10, SEQ ID NO 11 and SEQ ID NO 13 accepted adipate semialdehyde as substrate and synthesized 6-aminohexanoate as a reaction product.

#### Example 2

##### Enzyme Activity of Carboxylate Reductase Using 6-hydroxyhexanoate as Substrate and Forming 6-hydroxyhexanal

A nucleotide sequence encoding a His-tag was added to the genes from *Mycobacterium marinum*, *Mycobacterium smegmatis*, *Mycobacterium smegmatis*, *Segniliparus rugosus*, *Mycobacterium massiliense*, and *Segniliparus rotundus* that encode the carboxylate reductases of SEQ ID NOs: 3-7, respectively (see FIGS. 20A-20E) such that N-terminal HIS tagged carboxylate reductases could be produced. Each of the modified genes was cloned into a pET Duet expression vector alongside a sfp gene encoding a His-tagged phosphopantetheine transferase from *Bacillus subtilis*, both under control of the T7 promoter. Each expression vector was transformed into a BL21[DE3] *E. coli* host. Each resulting recombinant *E. coli* strain was cultivated at 37° C. in a 250 mL shake flask culture containing 50 mL LB media and antibiotic selection pressure, with shaking at 230 rpm. Each culture was induced overnight at 37° C. using an auto-induction media.

The pellet from each induced shake flask culture was harvested via centrifugation. Each pellet was resuspended

and lysed via sonication. The cell debris was separated from the supernatant via centrifugation. The carboxylate reductases and phosphopantetheine transferase were purified from the supernatant using Ni-affinity chromatography, diluted 10-fold into 50 mM HEPES buffer (pH=7.5) and concentrated via ultrafiltration.

Enzyme activity (i.e., 6-hydroxyhexanoate to 6-hydroxyhexanal) assays were performed in triplicate in a buffer composed of a final concentration of 50 mM HEPES buffer (pH=7.5), 2 mM 6-hydroxyhexanal, 10 mM MgCl<sub>2</sub>, 1 mM ATP, and 1 mM NADPH. Each enzyme activity assay reaction was initiated by adding purified carboxylate reductase and phosphopantetheine transferase or the empty vector control to the assay buffer containing the 6-hydroxyhexanoate and then incubated at room temperature for 20 min. The consumption of NADPH was monitored by absorbance at 340 nm. Each enzyme only control without 6-hydroxyhexanoate demonstrated low base line consumption of NADPH. See FIG. 9.

The gene products of SEQ ID NO 3-7, enhanced by the gene product of sfp, accepted 6-hydroxyhexanoate as substrate as confirmed against the empty vector control (see FIG. 11), and synthesized 6-hydroxyhexanal.

#### Example 3

##### Enzyme Activity of $\omega$ -transaminase for 6-aminohexanol, Forming 6-oxohexanol

A nucleotide sequence encoding an N-terminal His-tag was added to the *Chromobacterium violaceum*, *Pseudomonas aeruginosa*, *Pseudomonas syringae*, *Rhodobacter sphaeroides*, *Escherichia coli*, and *Vibrio fluvialis* genes encoding the  $\omega$ transaminases of SEQ ID NOs: 8-13, respectively (see FIG. 20E and FIG. 20F) such that N-terminal HIS tagged  $\omega$ -transaminases could be produced. The modified genes were cloned into a pET21a expression vector under the T7 promoter. Each expression vector was transformed into a BL21[DE3] *E. coli* host. Each resulting recombinant *E. coli* strain were cultivated at 37° C. in a 250 mL shake flask culture containing 50 mL LB media and antibiotic selection pressure, with shaking at 230 rpm. Each culture was induced overnight at 16° C. using 1 mM IPTG.

The pellet from each induced shake flask culture was harvested via centrifugation. Each pellet was resuspended and lysed via sonication. The cell debris was separated from the supernatant via centrifugation and the cell free extract was used immediately in enzyme activity assays.

Enzyme activity assays in the reverse direction (i.e., 6-aminohexanol to 6-oxohexanol) were performed in a buffer composed of a final concentration of 50 mM HEPES buffer (pH=7.5), 10 mM 6-aminohexanol, 10 mM pyruvate, and 100  $\mu$ M pyridoxyl 5' phosphate. Each enzyme activity assay reaction was initiated by adding cell free extract of the  $\omega$ -transaminase gene product or the empty vector control to the assay buffer containing the 6-aminohexanol and then incubated at 25° C. for 4 h, with shaking at 250 rpm. The formation of L-alanine was quantified via RP-HPLC.

Each enzyme only control without 6-aminohexanol had low base line conversion of pyruvate to L-alanine See FIG. 14.

The gene products of SEQ ID NO 8-13 accepted 6-aminohexanol as substrate as confirmed against the empty vector control (see FIG. 19) and synthesized 6-oxohexanol as reaction product. Given the reversibility of the  $\omega$ -transaminase activity (see Example 1), it can be concluded that the

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gene products of SEQ ID 8-13 accept 6-aminohexanol as substrate and form 6-oxohexanol.

## Example 4

Enzyme Activity of  $\omega$ -transaminase Using Hexamethylenediamine as Substrate and Forming 6-aminohexanal

A nucleotide sequence encoding an N-terminal His-tag was added to the *Chromobacterium violaceum*, *Pseudomonas aeruginosa*, *Pseudomonas syringae*, *Rhodobacter sphaeroides*, *Escherichia coli*, and *Vibrio fluvialis* genes encoding the  $\omega$ -transaminases of SEQ ID NOs: 8-13, respectively (see FIG. 20E and 20F) such that N-terminal HIS tagged  $\omega$ -transaminases could be produced. The modified genes were cloned into a pET21a expression vector under the T7 promoter. Each expression vector was transformed into a BL21[DE3] *E. coli* host. Each resulting recombinant *E. coli* strain were cultivated at 37° C. in a 250 mL shake flask culture containing 50 mL LB media and antibiotic selection pressure, with shaking at 230 rpm. Each culture was induced overnight at 16° C. using 1 mM IPTG.

The pellet from each induced shake flask culture was harvested via centrifugation. Each pellet was resuspended and lysed via sonication. The cell debris was separated from the supernatant via centrifugation and the cell free extract was used immediately in enzyme activity assays.

Enzyme activity assays in the reverse direction (i.e., hexamethylenediamine to 6-aminohexanal) were performed in a buffer composed of a final concentration of 50 mM HEPES buffer (pH=7.5), 10 mM hexamethylenediamine, 10 mM pyruvate, and 100  $\mu$ M pyridoxyl 5' phosphate. Each enzyme activity assay reaction was initiated by adding cell free extract of the  $\omega$ -transaminase gene product or the empty vector control to the assay buffer containing the hexamethylenediamine and then incubated at 25° C. for 4 h, with shaking at 250 rpm. The formation of L-alanine was quantified via RP-HPLC.

Each enzyme only control without hexamethylenediamine had low base line conversion of pyruvate to L-alanine. See FIG. 14.

The gene products of SEQ ID NO 8-13 accepted hexamethylenediamine as substrate as confirmed against the empty vector control (see FIG. 17) and synthesized 6-aminohexanal as reaction product. Given the reversibility of the  $\omega$ -transaminase activity (see Example 1), it can be concluded that the gene products of SEQ ID NOs: 8-13 accept 6-aminohexanal as substrate and form hexamethylenediamine.

## Example 5

Enzyme Activity of Carboxylate Reductase for N6-acetyl-6-aminohexanoate, Forming N6-acetyl-6-aminohexanal

The activity of each of the N-terminal His-tagged carboxylate reductases of SEQ ID NOs: 5-7 (see Example 2, and FIGS. 20C-20E) for converting N6-acetyl-6-aminohexanoate to N6-acetyl-6-aminohexanal was assayed in triplicate in a buffer composed of a final concentration of 50 mM HEPES buffer (pH=7.5), 2 mM N6-acetyl-6-aminohexanoate, 10 mM MgCl<sub>2</sub>, 1 mM ATP, and 1 mM NADPH. The assays were initiated by adding purified carboxylate reductase and phosphopantetheine transferase or the empty vector control to the assay buffer containing the N6-acetyl-

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6-aminohexanoate then incubated at room temperature for 20 min. The consumption of NADPH was monitored by absorbance at 340 nm. Each enzyme only control without N6-acetyl-6-aminohexanoate demonstrated low base line consumption of NADPH. See FIG. 9.

The gene products of SEQ ID NO 5-7, enhanced by the gene product of *sfp*, accepted N6-acetyl-6-aminohexanoate as substrate as confirmed against the empty vector control (see FIG. 12), and synthesized N6-acetyl-6-aminohexanal.

## Example 6

Enzyme Activity of  $\omega$ -transaminase using N6-acetyl-1,6-diaminohexane, and Forming N6-acetyl-6-aminohexanal

The activity of the N-terminal His-tagged  $\omega$ -transaminases of SEQ ID NOs: 8-13 (see Example 4, and FIG. 20E and FIG. 20F) for converting N6-acetyl-1,6-diaminohexane to N6-acetyl-6-aminohexanal was assayed using a buffer composed of a final concentration of 50 mM HEPES buffer (pH=7.5), 10 mM N6-acetyl-1,6-diaminohexane, 10 mM pyruvate and 100  $\mu$ M pyridoxyl 5' phosphate. Each enzyme activity assay reaction was initiated by adding a cell free extract of the  $\omega$ -transaminase or the empty vector control to the assay buffer containing the N6-acetyl-1,6-diaminohexane then incubated at 25° C. for 4 h, with shaking at 250 rpm. The formation of L-alanine was quantified via RP-HPLC.

Each enzyme only control without N6-acetyl-1,6-diaminohexane demonstrated low base line conversion of pyruvate to L-alanine See FIG. 14.

The gene product of SEQ ID NO 8-13 accepted N6-acetyl-1,6-diaminohexane as substrate as confirmed against the empty vector control (see FIG. 18) and synthesized N6-acetyl-6-aminohexanal as reaction product.

Given the reversibility of the  $\omega$ -transaminase activity (see example 1), the gene products of SEQ ID 8-13 accept N6-acetyl-6-aminohexanal as substrate forming N6-acetyl-1,6-diaminohexane.

## Example 7

Enzyme Activity of Carboxylate Reductase Using Adipate Semialdehyde as Substrate and Forming Hexanedial

The N-terminal His-tagged carboxylate reductase of SEQ ID NO 7 (see Example 2 and FIG. 20E) was assayed using adipate semialdehyde as substrate. The enzyme activity assay was performed in triplicate in a buffer composed of a final concentration of 50 mM HEPES buffer (pH=7.5), 2 mM adipate semialdehyde, 10 mM MgCl<sub>2</sub>, 1 mM ATP and 1 mM NADPH. The enzyme activity assay reaction was initiated by adding purified carboxylate reductase and phosphopantetheine transferase or the empty vector control to the assay buffer containing the adipate semialdehyde and then incubated at room temperature for 20 min. The consumption of NADPH was monitored by absorbance at 340 nm. The enzyme only control without adipate semialdehyde demonstrated low base line consumption of NADPH. See FIG. 9.

The gene product of SEQ ID NO 7, enhanced by the gene product of *sfp*, accepted adipate semialdehyde as substrate as confirmed against the empty vector control (see FIG. 13) and synthesized hexanedial.

## OTHER EMBODIMENTS

It is to be understood that while the invention has been described in conjunction with the detailed description

thereof, the foregoing description is intended to illustrate and not limit the scope of the invention, which is defined by the scope of the appended claims. Other aspects, advantages, and modifications are within the scope of the following claims.

## SEQUENCE LISTING

<160> NUMBER OF SEQ ID NOS: 34

<210> SEQ ID NO 1

<211> LENGTH: 246

<212> TYPE: PRT

<213> ORGANISM: *Lactobacillus brevis*

<400> SEQUENCE: 1

Met Ala Ala Asn Glu Phe Ser Glu Thr His Arg Val Val Tyr Tyr Glu  
1 5 10 15

Ala Asp Asp Thr Gly Gln Leu Thr Leu Ala Met Leu Ile Asn Leu Phe  
20 25 30

Val Leu Val Ser Glu Asp Gln Asn Asp Ala Leu Gly Leu Ser Thr Ala  
35 40 45

Phe Val Gln Ser His Gly Val Gly Trp Val Val Thr Gln Tyr His Leu  
50 55 60

His Ile Asp Glu Leu Pro Arg Thr Gly Ala Gln Val Thr Ile Lys Thr  
65 70 75 80

Arg Ala Thr Ala Tyr Asn Arg Tyr Phe Ala Tyr Arg Glu Tyr Trp Leu  
85 90 95

Leu Asp Asp Ala Gly Gln Val Leu Ala Tyr Gly Glu Gly Ile Trp Val  
100 105 110

Thr Met Ser Tyr Ala Thr Arg Lys Ile Thr Thr Ile Pro Ala Glu Val  
115 120 125

Met Ala Pro Tyr His Ser Glu Glu Gln Thr Arg Leu Pro Arg Leu Pro  
130 135 140

Arg Pro Asp His Phe Asp Glu Ala Val Asn Gln Thr Leu Lys Pro Tyr  
145 150 155 160

Thr Val Arg Tyr Phe Asp Ile Asp Gly Asn Gly His Val Asn Asn Ala  
165 170 175

His Tyr Phe Asp Trp Met Leu Asp Val Leu Pro Ala Thr Phe Leu Arg  
180 185 190

Ala His His Pro Thr Asp Val Lys Ile Arg Phe Glu Asn Glu Val Gln  
195 200 205

Tyr Gly His Gln Val Thr Ser Glu Leu Ser Gln Ala Ala Ala Leu Thr  
210 215 220

Thr Gln His Met Ile Lys Val Gly Asp Leu Thr Ala Val Lys Ala Thr  
225 230 235 240

Ile Gln Trp Asp Asn Arg  
245

<210> SEQ ID NO 2

<211> LENGTH: 261

<212> TYPE: PRT

<213> ORGANISM: *Lactobacillus plantarum*

<400> SEQUENCE: 2

Met Ala Thr Leu Gly Ala Asn Ala Ser Leu Tyr Ser Glu Gln His Arg  
1 5 10 15

Ile Thr Tyr Tyr Glu Cys Asp Arg Thr Gly Arg Ala Thr Leu Thr Thr  
20 25 30

-continued

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Leu Ile Asp Ile Ala Val Leu Ala Ser Glu Asp Gln Ser Asp Ala Leu  
           35                                  40                                  45  
 Gly Leu Thr Thr Glu Met Val Gln Ser His Gly Val Gly Trp Val Val  
       50                                  55                                  60  
 Thr Gln Tyr Ala Ile Asp Ile Thr Arg Met Pro Arg Gln Asp Glu Val  
   65                                  70                                  75                                  80  
 Val Thr Ile Ala Val Arg Gly Ser Ala Tyr Asn Pro Tyr Phe Ala Tyr  
                   85                                  90                                  95  
 Arg Glu Phe Trp Ile Arg Asp Ala Asp Gly Gln Gln Leu Ala Tyr Ile  
                   100                                  105                                  110  
 Thr Ser Ile Trp Val Met Met Ser Gln Thr Thr Arg Arg Ile Val Lys  
           115                                  120                                  125  
 Ile Leu Pro Glu Leu Val Ala Pro Tyr Gln Ser Glu Val Val Lys Arg  
       130                                  135                                  140  
 Ile Pro Arg Leu Pro Arg Pro Ile Ser Phe Glu Ala Thr Asp Thr Thr  
   145                                  150                                  155                                  160  
 Ile Thr Lys Pro Tyr His Val Arg Phe Phe Asp Ile Asp Pro Asn Arg  
                   165                                  170                                  175  
 His Val Asn Asn Ala His Tyr Phe Asp Trp Leu Val Asp Thr Leu Pro  
                   180                                  185                                  190  
 Ala Thr Phe Leu Leu Gln His Asp Leu Val His Val Asp Val Arg Tyr  
                   195                                  200                                  205  
 Glu Asn Glu Val Lys Tyr Gly Gln Thr Val Thr Ala His Ala Asn Ile  
       210                                  215                                  220  
 Leu Pro Ser Glu Val Ala Asp Gln Val Thr Thr Ser His Leu Ile Glu  
   225                                  230                                  235                                  240  
 Val Asp Asp Glu Lys Cys Cys Glu Val Thr Ile Gln Trp Arg Thr Leu  
                   245                                  250                                  255  
 Pro Glu Pro Ile Gln  
                   260

<210> SEQ ID NO 3  
 <211> LENGTH: 1174  
 <212> TYPE: PRT  
 <213> ORGANISM: Mycobacterium marinum

<400> SEQUENCE: 3

Met Ser Pro Ile Thr Arg Glu Glu Arg Leu Glu Arg Arg Ile Gln Asp  
   1                  5                                  10                                  15  
 Leu Tyr Ala Asn Asp Pro Gln Phe Ala Ala Ala Lys Pro Ala Thr Ala  
                   20                                  25                                  30  
 Ile Thr Ala Ala Ile Glu Arg Pro Gly Leu Pro Leu Pro Gln Ile Ile  
       35                                  40                                  45  
 Glu Thr Val Met Thr Gly Tyr Ala Asp Arg Pro Ala Leu Ala Gln Arg  
   50                                  55                                  60  
 Ser Val Glu Phe Val Thr Asp Ala Gly Thr Gly His Thr Thr Leu Arg  
   65                                  70                                  75                                  80  
 Leu Leu Pro His Phe Glu Thr Ile Ser Tyr Gly Glu Leu Trp Asp Arg  
                   85                                  90                                  95  
 Ile Ser Ala Leu Ala Asp Val Leu Ser Thr Glu Gln Thr Val Lys Pro  
                   100                                  105                                  110  
 Gly Asp Arg Val Cys Leu Leu Gly Phe Asn Ser Val Asp Tyr Ala Thr  
                   115                                  120                                  125  
 Ile Asp Met Thr Leu Ala Arg Leu Gly Ala Val Ala Val Pro Leu Gln  
   130                                  135                                  140

-continued

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Thr Ser Ala Ala Ile Thr Gln Leu Gln Pro Ile Val Ala Glu Thr Gln  
 145 150 155 160  
 Pro Thr Met Ile Ala Ala Ser Val Asp Ala Leu Ala Asp Ala Thr Glu  
 165 170 175  
 Leu Ala Leu Ser Gly Gln Thr Ala Thr Arg Val Leu Val Phe Asp His  
 180 185 190  
 His Arg Gln Val Asp Ala His Arg Ala Ala Val Glu Ser Ala Arg Glu  
 195 200 205  
 Arg Leu Ala Gly Ser Ala Val Val Glu Thr Leu Ala Glu Ala Ile Ala  
 210 215 220  
 Arg Gly Asp Val Pro Arg Gly Ala Ser Ala Gly Ser Ala Pro Gly Thr  
 225 230 235 240  
 Asp Val Ser Asp Asp Ser Leu Ala Leu Leu Ile Tyr Thr Ser Gly Ser  
 245 250 255  
 Thr Gly Ala Pro Lys Gly Ala Met Tyr Pro Arg Arg Asn Val Ala Thr  
 260 265 270  
 Phe Trp Arg Lys Arg Thr Trp Phe Glu Gly Gly Tyr Glu Pro Ser Ile  
 275 280 285  
 Thr Leu Asn Phe Met Pro Met Ser His Val Met Gly Arg Gln Ile Leu  
 290 295 300  
 Tyr Gly Thr Leu Cys Asn Gly Gly Thr Ala Tyr Phe Val Ala Lys Ser  
 305 310 315 320  
 Asp Leu Ser Thr Leu Phe Glu Asp Leu Ala Leu Val Arg Pro Thr Glu  
 325 330 335  
 Leu Thr Phe Val Pro Arg Val Trp Asp Met Val Phe Asp Glu Phe Gln  
 340 345 350  
 Ser Glu Val Asp Arg Arg Leu Val Asp Gly Ala Asp Arg Val Ala Leu  
 355 360 365  
 Glu Ala Gln Val Lys Ala Glu Ile Arg Asn Asp Val Leu Gly Gly Arg  
 370 375 380  
 Tyr Thr Ser Ala Leu Thr Gly Ser Ala Pro Ile Ser Asp Glu Met Lys  
 385 390 395 400  
 Ala Trp Val Glu Glu Leu Leu Asp Met His Leu Val Glu Gly Tyr Gly  
 405 410 415  
 Ser Thr Glu Ala Gly Met Ile Leu Ile Asp Gly Ala Ile Arg Arg Pro  
 420 425 430  
 Ala Val Leu Asp Tyr Lys Leu Val Asp Val Pro Asp Leu Gly Tyr Phe  
 435 440 445  
 Leu Thr Asp Arg Pro His Pro Arg Gly Glu Leu Leu Val Lys Thr Asp  
 450 455 460  
 Ser Leu Phe Pro Gly Tyr Tyr Gln Arg Ala Glu Val Thr Ala Asp Val  
 465 470 475 480  
 Phe Asp Ala Asp Gly Phe Tyr Arg Thr Gly Asp Ile Met Ala Glu Val  
 485 490 495  
 Gly Pro Glu Gln Phe Val Tyr Leu Asp Arg Arg Asn Asn Val Leu Lys  
 500 505 510  
 Leu Ser Gln Gly Glu Phe Val Thr Val Ser Lys Leu Glu Ala Val Phe  
 515 520 525  
 Gly Asp Ser Pro Leu Val Arg Gln Ile Tyr Ile Tyr Gly Asn Ser Ala  
 530 535 540  
 Arg Ala Tyr Leu Leu Ala Val Ile Val Pro Thr Gln Glu Ala Leu Asp  
 545 550 555 560

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Ala	Val	Pro	Val	Glu	Glu	Leu	Lys	Ala	Arg	Leu	Gly	Asp	Ser	Leu	Gln
				565					570					575	
Glu	Val	Ala	Lys	Ala	Ala	Gly	Leu	Gln	Ser	Tyr	Glu	Ile	Pro	Arg	Asp
			580					585					590		
Phe	Ile	Ile	Glu	Thr	Thr	Pro	Trp	Thr	Leu	Glu	Asn	Gly	Leu	Leu	Thr
		595					600					605			
Gly	Ile	Arg	Lys	Leu	Ala	Arg	Pro	Gln	Leu	Lys	Lys	His	Tyr	Gly	Glu
	610					615					620				
Leu	Leu	Glu	Gln	Ile	Tyr	Thr	Asp	Leu	Ala	His	Gly	Gln	Ala	Asp	Glu
625					630					635					640
Leu	Arg	Ser	Leu	Arg	Gln	Ser	Gly	Ala	Asp	Ala	Pro	Val	Leu	Val	Thr
				645					650					655	
Val	Cys	Arg	Ala	Ala	Ala	Ala	Leu	Leu	Gly	Gly	Ser	Ala	Ser	Asp	Val
			660					665						670	
Gln	Pro	Asp	Ala	His	Phe	Thr	Asp	Leu	Gly	Gly	Asp	Ser	Leu	Ser	Ala
		675					680					685			
Leu	Ser	Phe	Thr	Asn	Leu	Leu	His	Glu	Ile	Phe	Asp	Ile	Glu	Val	Pro
690						695					700				
Val	Gly	Val	Ile	Val	Ser	Pro	Ala	Asn	Asp	Leu	Gln	Ala	Leu	Ala	Asp
705					710					715					720
Tyr	Val	Glu	Ala	Ala	Arg	Lys	Pro	Gly	Ser	Ser	Arg	Pro	Thr	Phe	Ala
				725					730					735	
Ser	Val	His	Gly	Ala	Ser	Asn	Gly	Gln	Val	Thr	Glu	Val	His	Ala	Gly
			740					745					750		
Asp	Leu	Ser	Leu	Asp	Lys	Phe	Ile	Asp	Ala	Ala	Thr	Leu	Ala	Glu	Ala
		755					760					765			
Pro	Arg	Leu	Pro	Ala	Ala	Asn	Thr	Gln	Val	Arg	Thr	Val	Leu	Leu	Thr
		770				775					780				
Gly	Ala	Thr	Gly	Phe	Leu	Gly	Arg	Tyr	Leu	Ala	Leu	Glu	Trp	Leu	Glu
785					790					795					800
Arg	Met	Asp	Leu	Val	Asp	Gly	Lys	Leu	Ile	Cys	Leu	Val	Arg	Ala	Lys
				805					810					815	
Ser	Asp	Thr	Glu	Ala	Arg	Ala	Arg	Leu	Asp	Lys	Thr	Phe	Asp	Ser	Gly
			820					825					830		
Asp	Pro	Glu	Leu	Leu	Ala	His	Tyr	Arg	Ala	Leu	Ala	Gly	Asp	His	Leu
		835					840					845			
Glu	Val	Leu	Ala	Gly	Asp	Lys	Gly	Glu	Ala	Asp	Leu	Gly	Leu	Asp	Arg
850						855					860				
Gln	Thr	Trp	Gln	Arg	Leu	Ala	Asp	Thr	Val	Asp	Leu	Ile	Val	Asp	Pro
865					870					875					880
Ala	Ala	Leu	Val	Asn	His	Val	Leu	Pro	Tyr	Ser	Gln	Leu	Phe	Gly	Pro
				885					890					895	
Asn	Ala	Leu	Gly	Thr	Ala	Glu	Leu	Leu	Arg	Leu	Ala	Leu	Thr	Ser	Lys
			900					905					910		
Ile	Lys	Pro	Tyr	Ser	Tyr	Thr	Ser	Thr	Ile	Gly	Val	Ala	Asp	Gln	Ile
		915						920				925			
Pro	Pro	Ser	Ala	Phe	Thr	Glu	Asp	Ala	Asp	Ile	Arg	Val	Ile	Ser	Ala
						935					940				
Thr	Arg	Ala	Val	Asp	Asp	Ser	Tyr	Ala	Asn	Gly	Tyr	Ser	Asn	Ser	Lys
945					950					955					960
Trp	Ala	Gly	Glu	Val	Leu	Leu	Arg	Glu	Ala	His	Asp	Leu	Cys	Gly	Leu
				965					970					975	
Pro	Val	Ala	Val	Phe	Arg	Cys	Asp	Met	Ile	Leu	Ala	Asp	Thr	Thr	Trp

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980	985	990
Ala Gly Gln Leu Asn Val Pro Asp Met Phe Thr Arg Met Ile Leu Ser		
995	1000	1005
Leu Ala Ala Thr Gly Ile Ala Pro Gly Ser Phe Tyr Glu Leu Ala Ala		
1010	1015	1020
Asp Gly Ala Arg Gln Arg Ala His Tyr Asp Gly Leu Pro Val Glu Phe		
1025	1030	1035
1040		
Ile Ala Glu Ala Ile Ser Thr Leu Gly Ala Gln Ser Gln Asp Gly Phe		
1045	1050	1055
His Thr Tyr His Val Met Asn Pro Tyr Asp Asp Gly Ile Gly Leu Asp		
1060	1065	1070
Glu Phe Val Asp Trp Leu Asn Glu Ser Gly Cys Pro Ile Gln Arg Ile		
1075	1080	1085
Ala Asp Tyr Gly Asp Trp Leu Gln Arg Phe Glu Thr Ala Leu Arg Ala		
1090	1095	1100
Leu Pro Asp Arg Gln Arg His Ser Ser Leu Leu Pro Leu Leu His Asn		
1105	1110	1115
1120		
Tyr Arg Gln Pro Glu Arg Pro Val Arg Gly Ser Ile Ala Pro Thr Asp		
1125	1130	1135
Arg Phe Arg Ala Ala Val Gln Glu Ala Lys Ile Gly Pro Asp Lys Asp		
1140	1145	1150
Ile Pro His Val Gly Ala Pro Ile Ile Val Lys Tyr Val Ser Asp Leu		
1155	1160	1165
Arg Leu Leu Gly Leu Leu		
1170		
<210> SEQ ID NO 4		
<211> LENGTH: 1173		
<212> TYPE: PRT		
<213> ORGANISM: Mycobacterium smegmatis		
<400> SEQUENCE: 4		
Met Thr Ser Asp Val His Asp Ala Thr Asp Gly Val Thr Glu Thr Ala		
1	5	10
15		
Leu Asp Asp Glu Gln Ser Thr Arg Arg Ile Ala Glu Leu Tyr Ala Thr		
20	25	30
Asp Pro Glu Phe Ala Ala Ala Ala Pro Leu Pro Ala Val Val Asp Ala		
35	40	45
Ala His Lys Pro Gly Leu Arg Leu Ala Glu Ile Leu Gln Thr Leu Phe		
50	55	60
Thr Gly Tyr Gly Asp Arg Pro Ala Leu Gly Tyr Arg Ala Arg Glu Leu		
65	70	75
80		
Ala Thr Asp Glu Gly Gly Arg Thr Val Thr Arg Leu Leu Pro Arg Phe		
85	90	95
Asp Thr Leu Thr Tyr Ala Gln Val Trp Ser Arg Val Gln Ala Val Ala		
100	105	110
Ala Ala Leu Arg His Asn Phe Ala Gln Pro Ile Tyr Pro Gly Asp Ala		
115	120	125
Val Ala Thr Ile Gly Phe Ala Ser Pro Asp Tyr Leu Thr Leu Asp Leu		
130	135	140
Val Cys Ala Tyr Leu Gly Leu Val Ser Val Pro Leu Gln His Asn Ala		
145	150	155
160		
Pro Val Ser Arg Leu Ala Pro Ile Leu Ala Glu Val Glu Pro Arg Ile		
165	170	175



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Leu Thr Val Ser Ala Glu Tyr Leu Asp Leu Ala Val Glu Ser Val Arg  
 180 185 190  
 Asp Val Asn Ser Val Ser Gln Leu Val Val Phe Asp His His Pro Glu  
 195 200 205  
 Val Asp Asp His Arg Asp Ala Leu Ala Arg Ala Arg Glu Gln Leu Ala  
 210 215 220  
 Gly Lys Gly Ile Ala Val Thr Thr Leu Asp Ala Ile Ala Asp Glu Gly  
 225 230 235 240  
 Ala Gly Leu Pro Ala Glu Pro Ile Tyr Thr Ala Asp His Asp Gln Arg  
 245 250 255  
 Leu Ala Met Ile Leu Tyr Thr Ser Gly Ser Thr Gly Ala Pro Lys Gly  
 260 265 270  
 Ala Met Tyr Thr Glu Ala Met Val Ala Arg Leu Trp Thr Met Ser Phe  
 275 280 285  
 Ile Thr Gly Asp Pro Thr Pro Val Ile Asn Val Asn Phe Met Pro Leu  
 290 295 300  
 Asn His Leu Gly Gly Arg Ile Pro Ile Ser Thr Ala Val Gln Asn Gly  
 305 310 315 320  
 Gly Thr Ser Tyr Phe Val Pro Glu Ser Asp Met Ser Thr Leu Phe Glu  
 325 330 335  
 Asp Leu Ala Leu Val Arg Pro Thr Glu Leu Gly Leu Val Pro Arg Val  
 340 345 350  
 Ala Asp Met Leu Tyr Gln His His Leu Ala Thr Val Asp Arg Leu Val  
 355 360 365  
 Thr Gln Gly Ala Asp Glu Leu Thr Ala Glu Lys Gln Ala Gly Ala Glu  
 370 375 380  
 Leu Arg Glu Gln Val Leu Gly Gly Arg Val Ile Thr Gly Phe Val Ser  
 385 390 395 400  
 Thr Ala Pro Leu Ala Ala Glu Met Arg Ala Phe Leu Asp Ile Thr Leu  
 405 410 415  
 Gly Ala His Ile Val Asp Gly Tyr Gly Leu Thr Glu Thr Gly Ala Val  
 420 425 430  
 Thr Arg Asp Gly Val Ile Val Arg Pro Pro Val Ile Asp Tyr Lys Leu  
 435 440 445  
 Ile Asp Val Pro Glu Leu Gly Tyr Phe Ser Thr Asp Lys Pro Tyr Pro  
 450 455 460  
 Arg Gly Glu Leu Leu Val Arg Ser Gln Thr Leu Thr Pro Gly Tyr Tyr  
 465 470 475 480  
 Lys Arg Pro Glu Val Thr Ala Ser Val Phe Asp Arg Asp Gly Tyr Tyr  
 485 490 495  
 His Thr Gly Asp Val Met Ala Glu Thr Ala Pro Asp His Leu Val Tyr  
 500 505 510  
 Val Asp Arg Arg Asn Asn Val Leu Lys Leu Ala Gln Gly Glu Phe Val  
 515 520 525  
 Ala Val Ala Asn Leu Glu Ala Val Phe Ser Gly Ala Ala Leu Val Arg  
 530 535 540  
 Gln Ile Phe Val Tyr Gly Asn Ser Glu Arg Ser Phe Leu Leu Ala Val  
 545 550 555 560  
 Val Val Pro Thr Pro Glu Ala Leu Glu Gln Tyr Asp Pro Ala Ala Leu  
 565 570 575  
 Lys Ala Ala Leu Ala Asp Ser Leu Gln Arg Thr Ala Arg Asp Ala Glu  
 580 585 590  
 Leu Gln Ser Tyr Glu Val Pro Ala Asp Phe Ile Val Glu Thr Glu Pro

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595			600			605									
Phe	Ser	Ala	Ala	Asn	Gly	Leu	Leu	Ser	Gly	Val	Gly	Lys	Leu	Leu	Arg
610						615					620				
Pro	Asn	Leu	Lys	Asp	Arg	Tyr	Gly	Gln	Arg	Leu	Glu	Gln	Met	Tyr	Ala
625				630						635					640
Asp	Ile	Ala	Ala	Thr	Gln	Ala	Asn	Gln	Leu	Arg	Glu	Leu	Arg	Arg	Ala
				645						650				655	
Ala	Ala	Thr	Gln	Pro	Val	Ile	Asp	Thr	Leu	Thr	Gln	Ala	Ala	Ala	Thr
			660					665					670		
Ile	Leu	Gly	Thr	Gly	Ser	Glu	Val	Ala	Ser	Asp	Ala	His	Phe	Thr	Asp
		675						680					685		
Leu	Gly	Gly	Asp	Ser	Leu	Ser	Ala	Leu	Thr	Leu	Ser	Asn	Leu	Leu	Ser
	690						695				700				
Asp	Phe	Phe	Gly	Phe	Glu	Val	Pro	Val	Gly	Thr	Ile	Val	Asn	Pro	Ala
705					710					715					720
Thr	Asn	Leu	Ala	Gln	Leu	Ala	Gln	His	Ile	Glu	Ala	Gln	Arg	Thr	Ala
				725						730				735	
Gly	Asp	Arg	Arg	Pro	Ser	Phe	Thr	Thr	Val	His	Gly	Ala	Asp	Ala	Thr
				740				745					750		
Glu	Ile	Arg	Ala	Ser	Glu	Leu	Thr	Leu	Asp	Lys	Phe	Ile	Asp	Ala	Glu
		755						760					765		
Thr	Leu	Arg	Ala	Ala	Pro	Gly	Leu	Pro	Lys	Val	Thr	Thr	Glu	Pro	Arg
	770						775				780				
Thr	Val	Leu	Leu	Ser	Gly	Ala	Asn	Gly	Trp	Leu	Gly	Arg	Phe	Leu	Thr
785					790					795					800
Leu	Gln	Trp	Leu	Glu	Arg	Leu	Ala	Pro	Val	Gly	Gly	Thr	Leu	Ile	Thr
				805						810				815	
Ile	Val	Arg	Gly	Arg	Asp	Asp	Ala	Ala	Ala	Arg	Ala	Arg	Leu	Thr	Gln
			820					825					830		
Ala	Tyr	Asp	Thr	Asp	Pro	Glu	Leu	Ser	Arg	Arg	Phe	Ala	Glu	Leu	Ala
		835						840					845		
Asp	Arg	His	Leu	Arg	Val	Val	Ala	Gly	Asp	Ile	Gly	Asp	Pro	Asn	Leu
	850						855				860				
Gly	Leu	Thr	Pro	Glu	Ile	Trp	His	Arg	Leu	Ala	Ala	Glu	Val	Asp	Leu
865					870					875					880
Val	Val	His	Pro	Ala	Ala	Leu	Val	Asn	His	Val	Leu	Pro	Tyr	Arg	Gln
				885						890				895	
Leu	Phe	Gly	Pro	Asn	Val	Val	Gly	Thr	Ala	Glu	Val	Ile	Lys	Leu	Ala
				900				905					910		
Leu	Thr	Glu	Arg	Ile	Lys	Pro	Val	Thr	Tyr	Leu	Ser	Thr	Val	Ser	Val
		915						920					925		
Ala	Met	Gly	Ile	Pro	Asp	Phe	Glu	Glu	Asp	Gly	Asp	Ile	Arg	Thr	Val
	930						935				940				
Ser	Pro	Val	Arg	Pro	Leu	Asp	Gly	Gly	Tyr	Ala	Asn	Gly	Tyr	Gly	Asn
945					950					955					960
Ser	Lys	Trp	Ala	Gly	Glu	Val	Leu	Leu	Arg	Glu	Ala	His	Asp	Leu	Cys
				965						970				975	
Gly	Leu	Pro	Val	Ala	Thr	Phe	Arg	Ser	Asp	Met	Ile	Leu	Ala	His	Pro
				980				985					990		
Arg	Tyr	Arg	Gly	Gln	Val	Asn	Val	Pro	Asp	Met	Phe	Thr	Arg	Leu	Leu
		995					1000						1005		
Leu	Ser	Leu	Leu	Ile	Thr	Gly	Val	Ala	Pro	Arg	Ser	Phe	Tyr	Ile	Gly
	1010						1015				1020				

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Asp Gly Glu Arg Pro Arg Ala His Tyr Pro Gly Leu Thr Val Asp Phe  
 1025 1030 1035 1040  
 Val Ala Glu Ala Val Thr Thr Leu Gly Ala Gln Gln Arg Glu Gly Tyr  
 1045 1050 1055  
 Val Ser Tyr Asp Val Met Asn Pro His Asp Asp Gly Ile Ser Leu Asp  
 1060 1065 1070  
 Val Phe Val Asp Trp Leu Ile Arg Ala Gly His Pro Ile Asp Arg Val  
 1075 1080 1085  
 Asp Asp Tyr Asp Asp Trp Val Arg Arg Phe Glu Thr Ala Leu Thr Ala  
 1090 1095 1100  
 Leu Pro Glu Lys Arg Arg Ala Gln Thr Val Leu Pro Leu Leu His Ala  
 1105 1110 1115 1120  
 Phe Arg Ala Pro Gln Ala Pro Leu Arg Gly Ala Pro Glu Pro Thr Glu  
 1125 1130 1135  
 Val Phe His Ala Ala Val Arg Thr Ala Lys Val Gly Pro Gly Asp Ile  
 1140 1145 1150  
 Pro His Leu Asp Glu Ala Leu Ile Asp Lys Tyr Ile Arg Asp Leu Arg  
 1155 1160 1165  
 Glu Phe Gly Leu Ile  
 1170

<210> SEQ ID NO 5  
 <211> LENGTH: 1148  
 <212> TYPE: PRT  
 <213> ORGANISM: Segniliparus rugosus

<400> SEQUENCE: 5

Met Gly Asp Gly Glu Glu Arg Ala Lys Arg Phe Phe Gln Arg Ile Gly  
 1 5 10 15  
 Glu Leu Ser Ala Thr Asp Pro Gln Phe Ala Ala Ala Ala Pro Asp Pro  
 20 25 30  
 Ala Val Val Glu Ala Val Ser Asp Pro Ser Leu Ser Phe Thr Arg Tyr  
 35 40 45  
 Leu Asp Thr Leu Met Arg Gly Tyr Ala Glu Arg Pro Ala Leu Ala His  
 50 55 60  
 Arg Val Gly Ala Gly Tyr Glu Thr Ile Ser Tyr Gly Glu Leu Trp Ala  
 65 70 75 80  
 Arg Val Gly Ala Ile Ala Ala Ala Trp Gln Ala Asp Gly Leu Ala Pro  
 85 90 95  
 Gly Asp Phe Val Ala Thr Val Gly Phe Thr Ser Pro Asp Tyr Val Ala  
 100 105 110  
 Val Asp Leu Ala Ala Ala Arg Ser Gly Leu Val Ser Val Pro Leu Gln  
 115 120 125  
 Ala Gly Ala Ser Leu Ala Gln Leu Val Gly Ile Leu Glu Glu Thr Glu  
 130 135 140  
 Pro Lys Val Leu Ala Ala Ser Ala Ser Ser Leu Glu Gly Ala Val Ala  
 145 150 155 160  
 Cys Ala Leu Ala Ala Pro Ser Val Gln Arg Leu Val Val Phe Asp Leu  
 165 170 175  
 Arg Gly Pro Asp Ala Ser Glu Ser Ala Ala Asp Glu Arg Arg Gly Ala  
 180 185 190  
 Leu Ala Asp Ala Glu Glu Gln Leu Ala Arg Ala Gly Arg Ala Val Val  
 195 200 205  
 Val Glu Thr Leu Ala Asp Leu Ala Ala Arg Gly Glu Ala Leu Pro Glu

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210					215					220					
Ala	Pro	Leu	Phe	Glu	Pro	Ala	Glu	Gly	Glu	Asp	Pro	Leu	Ala	Leu	Leu
225					230					235					240
Ile	Tyr	Thr	Ser	Gly	Ser	Thr	Gly	Ala	Pro	Lys	Gly	Ala	Met	Tyr	Ser
				245					250					255	
Gln	Arg	Leu	Val	Ser	Gln	Leu	Trp	Gly	Arg	Thr	Pro	Val	Val	Pro	Gly
			260					265					270		
Met	Pro	Asn	Ile	Ser	Leu	His	Tyr	Met	Pro	Leu	Ser	His	Ser	Tyr	Gly
		275					280					285			
Arg	Ala	Val	Leu	Ala	Gly	Ala	Leu	Ser	Ala	Gly	Gly	Thr	Ala	His	Phe
	290					295					300				
Thr	Ala	Asn	Ser	Asp	Leu	Ser	Thr	Leu	Phe	Glu	Asp	Ile	Ala	Leu	Ala
305					310					315					320
Arg	Pro	Thr	Phe	Leu	Ala	Leu	Val	Pro	Arg	Val	Cys	Glu	Met	Leu	Phe
			325						330					335	
Gln	Glu	Ser	Gln	Arg	Gly	Gln	Asp	Val	Ala	Glu	Leu	Arg	Glu	Arg	Val
			340					345					350		
Leu	Gly	Gly	Arg	Leu	Leu	Val	Ala	Val	Cys	Gly	Ser	Ala	Pro	Leu	Ser
		355					360					365			
Pro	Glu	Met	Arg	Ala	Phe	Met	Glu	Glu	Val	Leu	Gly	Phe	Pro	Leu	Leu
	370					375					380				
Asp	Gly	Tyr	Gly	Ser	Thr	Glu	Ala	Leu	Gly	Val	Met	Arg	Asn	Gly	Ile
385					390					395					400
Ile	Gln	Arg	Pro	Pro	Val	Ile	Asp	Tyr	Lys	Leu	Val	Asp	Val	Pro	Glu
			405						410					415	
Leu	Gly	Tyr	Arg	Thr	Thr	Asp	Lys	Pro	Tyr	Pro	Arg	Gly	Glu	Leu	Cys
			420					425					430		
Ile	Arg	Ser	Thr	Ser	Leu	Ile	Ser	Gly	Tyr	Tyr	Lys	Arg	Pro	Glu	Ile
	435						440					445			
Thr	Ala	Glu	Val	Phe	Asp	Ala	Gln	Gly	Tyr	Tyr	Lys	Thr	Gly	Asp	Val
	450					455					460				
Met	Ala	Glu	Ile	Ala	Pro	Asp	His	Leu	Val	Tyr	Val	Asp	Arg	Ser	Lys
465					470					475					480
Asn	Val	Leu	Lys	Leu	Ser	Gln	Gly	Glu	Phe	Val	Ala	Val	Ala	Lys	Leu
			485						490					495	
Glu	Ala	Ala	Tyr	Gly	Thr	Ser	Pro	Tyr	Val	Lys	Gln	Ile	Phe	Val	Tyr
			500					505					510		
Gly	Asn	Ser	Glu	Arg	Ser	Phe	Leu	Leu	Ala	Val	Val	Val	Pro	Asn	Ala
		515					520					525			
Glu	Val	Leu	Gly	Ala	Arg	Asp	Gln	Glu	Glu	Ala	Lys	Pro	Leu	Ile	Ala
	530					535					540				
Ala	Ser	Leu	Gln	Lys	Ile	Ala	Lys	Glu	Ala	Gly	Leu	Gln	Ser	Tyr	Glu
545					550					555					560
Val	Pro	Arg	Asp	Phe	Leu	Ile	Glu	Thr	Glu	Pro	Phe	Thr	Thr	Gln	Asn
			565						570					575	
Gly	Leu	Leu	Ser	Glu	Val	Gly	Lys	Leu	Leu	Arg	Pro	Lys	Leu	Lys	Ala
			580					585					590		
Arg	Tyr	Gly	Glu	Ala	Leu	Glu	Ala	Arg	Tyr	Asp	Glu	Ile	Ala	His	Gly
		595					600					605			
Gln	Ala	Asp	Glu	Leu	Arg	Ala	Leu	Arg	Asp	Gly	Ala	Gly	Gln	Arg	Pro
	610					615					620				
Val	Val	Glu	Thr	Val	Val	Arg	Ala	Ala	Val	Ala	Ile	Ser	Gly	Ser	Glu
625					630					635					640

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Gly Ala Glu Val Gly Pro Glu Ala Asn Phe Ala Asp Leu Gly Gly Asp  
 645 650 655  
 Ser Leu Ser Ala Leu Ser Leu Ala Asn Leu Leu His Asp Val Phe Glu  
 660 665 670  
 Val Glu Val Pro Val Arg Ile Ile Ile Gly Pro Thr Ala Ser Leu Ala  
 675 680 685  
 Gly Ile Ala Lys His Ile Glu Ala Glu Arg Ala Gly Ala Ser Ala Pro  
 690 695 700  
 Thr Ala Ala Ser Val His Gly Ala Gly Ala Thr Arg Ile Arg Ala Ser  
 705 710 715 720  
 Glu Leu Thr Leu Glu Lys Phe Leu Pro Glu Asp Leu Leu Ala Ala Ala  
 725 730 735  
 Lys Gly Leu Pro Ala Ala Asp Gln Val Arg Thr Val Leu Leu Thr Gly  
 740 745 750  
 Ala Asn Gly Trp Leu Gly Arg Phe Leu Ala Leu Glu Gln Leu Glu Arg  
 755 760 765  
 Leu Ala Arg Ser Gly Gln Asp Gly Gly Lys Leu Ile Cys Leu Val Arg  
 770 775 780  
 Gly Lys Asp Ala Ala Ala Ala Arg Arg Arg Ile Glu Glu Thr Leu Gly  
 785 790 800  
 Thr Asp Pro Ala Leu Ala Ala Arg Phe Ala Glu Leu Ala Glu Gly Arg  
 805 810 815  
 Leu Glu Val Val Pro Gly Asp Val Gly Glu Pro Lys Phe Gly Leu Asp  
 820 825 830  
 Asp Ala Ala Trp Asp Arg Leu Ala Glu Glu Val Asp Val Ile Val His  
 835 840 845  
 Pro Ala Ala Leu Val Asn His Val Leu Pro Tyr His Gln Leu Phe Gly  
 850 855 860  
 Pro Asn Val Val Gly Thr Ala Glu Ile Ile Arg Leu Ala Ile Thr Ala  
 865 870 875 880  
 Lys Arg Lys Pro Val Thr Tyr Leu Ser Thr Val Ala Val Ala Ala Gly  
 885 890 895  
 Val Glu Pro Ser Ser Phe Glu Glu Asp Gly Asp Ile Arg Ala Val Val  
 900 905 910  
 Pro Glu Arg Pro Leu Gly Asp Gly Tyr Ala Asn Gly Tyr Gly Asn Ser  
 915 920 925  
 Lys Trp Ala Gly Glu Val Leu Leu Arg Glu Ala His Glu Leu Val Gly  
 930 935 940  
 Leu Pro Val Ala Val Phe Arg Ser Asp Met Ile Leu Ala His Thr Arg  
 945 950 955 960  
 Tyr Thr Gly Gln Leu Asn Val Pro Asp Gln Phe Thr Arg Leu Val Leu  
 965 970 975  
 Ser Leu Leu Ala Thr Gly Ile Ala Pro Lys Ser Phe Tyr Gln Gln Gly  
 980 985 990  
 Ala Ala Gly Glu Arg Gln Arg Ala His Tyr Asp Gly Ile Pro Val Asp  
 995 1000 1005  
 Phe Thr Ala Glu Ala Ile Thr Thr Leu Gly Ala Glu Pro Ser Trp Phe  
 1010 1015 1020  
 Asp Gly Gly Ala Gly Phe Arg Ser Phe Asp Val Phe Asn Pro His His  
 1025 1030 1035 1040  
 Asp Gly Val Gly Leu Asp Glu Phe Val Asp Trp Leu Ile Glu Ala Gly  
 1045 1050 1055

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His Pro Ile Ser Arg Ile Asp Asp His Lys Glu Trp Phe Ala Arg Phe  
 1060 1065 1070

Glu Thr Ala Val Arg Gly Leu Pro Glu Ala Gln Arg Gln His Ser Leu  
 1075 1080 1085

Leu Pro Leu Leu Arg Ala Tyr Ser Phe Pro His Pro Pro Val Asp Gly  
 1090 1095 1100

Ser Val Tyr Pro Thr Gly Lys Phe Gln Gly Ala Val Lys Ala Ala Gln  
 1105 1110 1115 1120

Val Gly Ser Asp His Asp Val Pro His Leu Gly Lys Ala Leu Ile Val  
 1125 1130 1135

Lys Tyr Ala Asp Asp Leu Lys Ala Leu Gly Leu Leu  
 1140 1145

<210> SEQ ID NO 6  
 <211> LENGTH: 1185  
 <212> TYPE: PRT  
 <213> ORGANISM: Mycobacterium massiliense

<400> SEQUENCE: 6

Met Thr Asn Glu Thr Asn Pro Gln Gln Glu Gln Leu Ser Arg Arg Ile  
 1 5 10 15

Glu Ser Leu Arg Glu Ser Asp Pro Gln Phe Arg Ala Ala Gln Pro Asp  
 20 25 30

Pro Ala Val Ala Glu Gln Val Leu Arg Pro Gly Leu His Leu Ser Glu  
 35 40 45

Ala Ile Ala Ala Leu Met Thr Gly Tyr Ala Glu Arg Pro Ala Leu Gly  
 50 55 60

Glu Arg Ala Arg Glu Leu Val Ile Asp Gln Asp Gly Arg Thr Thr Leu  
 65 70 75 80

Arg Leu Leu Pro Arg Phe Asp Thr Thr Thr Tyr Gly Glu Leu Trp Ser  
 85 90 95

Arg Thr Thr Ser Val Ala Ala Ala Trp His His Asp Ala Thr His Pro  
 100 105 110

Val Lys Ala Gly Asp Leu Val Ala Thr Leu Gly Phe Thr Ser Ile Asp  
 115 120 125

Tyr Thr Val Leu Asp Leu Ala Ile Met Ile Leu Gly Gly Val Ala Val  
 130 135 140

Pro Leu Gln Thr Ser Ala Pro Ala Ser Gln Trp Thr Thr Ile Leu Ala  
 145 150 155 160

Glu Ala Glu Pro Asn Thr Leu Ala Val Ser Ile Glu Leu Ile Gly Ala  
 165 170 175

Ala Met Glu Ser Val Arg Ala Thr Pro Ser Ile Lys Gln Val Val Val  
 180 185 190

Phe Asp Tyr Thr Pro Glu Val Asp Asp Gln Arg Glu Ala Phe Glu Ala  
 195 200 205

Ala Ser Thr Gln Leu Ala Gly Thr Gly Ile Ala Leu Glu Thr Leu Asp  
 210 215 220

Ala Val Ile Ala Arg Gly Ala Ala Leu Pro Ala Ala Pro Leu Tyr Ala  
 225 230 235 240

Pro Ser Ala Gly Asp Asp Pro Leu Ala Leu Leu Ile Tyr Thr Ser Gly  
 245 250 255

Ser Thr Gly Ala Pro Lys Gly Ala Met His Ser Glu Asn Ile Val Arg  
 260 265 270

Arg Trp Trp Ile Arg Glu Asp Val Met Ala Gly Thr Glu Asn Leu Pro  
 275 280 285

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Met Ile Gly Leu Asn Phe Met Pro Met Ser His Ile Met Gly Arg Gly  
 290 295 300  
 Thr Leu Thr Ser Thr Leu Ser Thr Gly Gly Thr Gly Tyr Phe Ala Ala  
 305 310 315 320  
 Ser Ser Asp Met Ser Thr Leu Phe Glu Asp Met Glu Leu Ile Arg Pro  
 325 330 335  
 Thr Ala Leu Ala Leu Val Pro Arg Val Cys Asp Met Val Phe Gln Arg  
 340 345 350  
 Phe Gln Thr Glu Val Asp Arg Arg Leu Ala Ser Gly Asp Thr Ala Ser  
 355 360 365  
 Ala Glu Ala Val Ala Ala Glu Val Lys Ala Asp Ile Arg Asp Asn Leu  
 370 375 380  
 Phe Gly Gly Arg Val Ser Ala Val Met Val Gly Ser Ala Pro Leu Ser  
 385 390 395 400  
 Glu Glu Leu Gly Glu Phe Ile Glu Ser Cys Phe Glu Leu Asn Leu Thr  
 405 410 415  
 Asp Gly Tyr Gly Ser Thr Glu Ala Gly Met Val Phe Arg Asp Gly Ile  
 420 425 430  
 Val Gln Arg Pro Pro Val Ile Asp Tyr Lys Leu Val Asp Val Pro Glu  
 435 440 445  
 Leu Gly Tyr Phe Ser Thr Asp Lys Pro His Pro Arg Gly Glu Leu Leu  
 450 455 460  
 Leu Lys Thr Asp Gly Met Phe Leu Gly Tyr Tyr Lys Arg Pro Glu Val  
 465 470 475 480  
 Thr Ala Ser Val Phe Asp Ala Asp Gly Phe Tyr Met Thr Gly Asp Ile  
 485 490 495  
 Val Ala Glu Leu Ala His Asp Asn Ile Glu Ile Ile Asp Arg Arg Asn  
 500 505 510  
 Asn Val Leu Lys Leu Ser Gln Gly Glu Phe Val Ala Val Ala Thr Leu  
 515 520 525  
 Glu Ala Glu Tyr Ala Asn Ser Pro Val Val His Gln Ile Tyr Val Tyr  
 530 535 540  
 Gly Ser Ser Glu Arg Ser Tyr Leu Leu Ala Val Val Val Pro Thr Pro  
 545 550 555 560  
 Glu Ala Val Ala Ala Ala Lys Gly Asp Ala Ala Ala Leu Lys Thr Thr  
 565 570 575  
 Ile Ala Asp Ser Leu Gln Asp Ile Ala Lys Glu Ile Gln Leu Gln Ser  
 580 585 590  
 Tyr Glu Val Pro Arg Asp Phe Ile Ile Glu Pro Gln Pro Phe Thr Gln  
 595 600 605  
 Gly Asn Gly Leu Leu Thr Gly Ile Ala Lys Leu Ala Arg Pro Asn Leu  
 610 615 620  
 Lys Ala His Tyr Gly Pro Arg Leu Glu Gln Met Tyr Ala Glu Ile Ala  
 625 630 635 640  
 Glu Gln Gln Ala Ala Glu Leu Arg Ala Leu His Gly Val Asp Pro Asp  
 645 650 655  
 Lys Pro Ala Leu Glu Thr Val Leu Lys Ala Ala Gln Ala Leu Leu Gly  
 660 665 670  
 Val Ser Ser Ala Glu Leu Ala Ala Asp Ala His Phe Thr Asp Leu Gly  
 675 680 685  
 Gly Asp Ser Leu Ser Ala Leu Ser Phe Ser Asp Leu Leu Arg Asp Ile  
 690 695 700

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Phe	Ala	Val	Glu	Val	Pro	Val	Gly	Val	Ile	Val	Ser	Ala	Ala	Asn	Asp	705	710	715	720
Leu	Gly	Gly	Val	Ala	Lys	Phe	Val	Asp	Glu	Gln	Arg	His	Ser	Gly	Gly	725	730	735	
Thr	Arg	Pro	Thr	Ala	Glu	Thr	Val	His	Gly	Ala	Gly	His	Thr	Glu	Ile	740	745	750	
Arg	Ala	Ala	Asp	Leu	Thr	Leu	Asp	Lys	Phe	Ile	Asp	Glu	Ala	Thr	Leu	755	760	765	
His	Ala	Ala	Pro	Ser	Leu	Pro	Lys	Ala	Ala	Gly	Ile	Pro	His	Thr	Val	770	775	780	
Leu	Leu	Thr	Gly	Ser	Asn	Gly	Tyr	Leu	Gly	His	Tyr	Leu	Ala	Leu	Glu	785	790	795	800
Trp	Leu	Glu	Arg	Leu	Asp	Lys	Thr	Asp	Gly	Lys	Leu	Ile	Val	Ile	Val	805	810	815	
Arg	Gly	Lys	Asn	Ala	Glu	Ala	Ala	Tyr	Gly	Arg	Leu	Glu	Glu	Ala	Phe	820	825	830	
Asp	Thr	Gly	Asp	Thr	Glu	Leu	Leu	Ala	His	Phe	Arg	Ser	Leu	Ala	Asp	835	840	845	
Lys	His	Leu	Glu	Val	Leu	Ala	Gly	Asp	Ile	Gly	Asp	Pro	Asn	Leu	Gly	850	855	860	
Leu	Asp	Ala	Asp	Thr	Trp	Gln	Arg	Leu	Ala	Asp	Thr	Val	Asp	Val	Ile	865	870	875	880
Val	His	Pro	Ala	Ala	Leu	Val	Asn	His	Val	Leu	Pro	Tyr	Asn	Gln	Leu	885	890	895	
Phe	Gly	Pro	Asn	Val	Val	Gly	Thr	Ala	Glu	Ile	Ile	Lys	Leu	Ala	Ile	900	905	910	
Thr	Thr	Lys	Ile	Lys	Pro	Val	Thr	Tyr	Leu	Ser	Thr	Val	Ala	Val	Ala	915	920	925	
Ala	Tyr	Val	Asp	Pro	Thr	Thr	Phe	Asp	Glu	Glu	Ser	Asp	Ile	Arg	Leu	930	935	940	
Ile	Ser	Ala	Val	Arg	Pro	Ile	Asp	Asp	Gly	Tyr	Ala	Asn	Gly	Tyr	Gly	945	950	955	960
Asn	Ala	Lys	Trp	Ala	Gly	Glu	Val	Leu	Leu	Arg	Glu	Ala	His	Asp	Leu	965	970	975	
Cys	Gly	Leu	Pro	Val	Ala	Val	Phe	Arg	Ser	Asp	Met	Ile	Leu	Ala	His	980	985	990	
Ser	Arg	Tyr	Thr	Gly	Gln	Leu	Asn	Val	Pro	Asp	Gln	Phe	Thr	Arg	Leu	995	1000	1005	
Ile	Leu	Ser	Leu	Ile	Ala	Thr	Gly	Ile	Ala	Pro	Gly	Ser	Phe	Tyr	Gln	1010	1015	1020	
Ala	Gln	Thr	Thr	Gly	Glu	Arg	Pro	Leu	Ala	His	Tyr	Asp	Gly	Leu	Pro	1025	1030	1035	1040
Gly	Asp	Phe	Thr	Ala	Glu	Ala	Ile	Thr	Thr	Leu	Gly	Thr	Gln	Val	Pro	1045	1050	1055	
Glu	Gly	Ser	Glu	Gly	Phe	Val	Thr	Tyr	Asp	Cys	Val	Asn	Pro	His	Ala	1060	1065	1070	
Asp	Gly	Ile	Ser	Leu	Asp	Asn	Phe	Val	Asp	Trp	Leu	Ile	Glu	Ala	Gly	1075	1080	1085	
Tyr	Pro	Ile	Ala	Arg	Ile	Asp	Asn	Tyr	Thr	Glu	Trp	Phe	Thr	Arg	Phe	1090	1095	1100	
Asp	Thr	Ala	Ile	Arg	Gly	Leu	Ser	Glu	Lys	Gln	Lys	Gln	His	Ser	Leu	1105	1110	1115	1120
Leu	Pro	Leu	Leu	His	Ala	Phe	Glu	Gln	Pro	Ser	Ala	Ala	Glu	Asn	His				



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1125					1130					1135					
Gly	Val	Val	Pro	Ala	Lys	Arg	Phe	Gln	His	Ala	Val	Gln	Ala	Ala	Gly
			1140					1145					1150		
Ile	Gly	Pro	Val	Gly	Gln	Asp	Gly	Thr	Thr	Asp	Ile	Pro	His	Leu	Ser
		1155					1160					1165			
Arg	Arg	Leu	Ile	Val	Lys	Tyr	Ala	Lys	Asp	Leu	Glu	Gln	Leu	Gly	Leu
		1170				1175					1180				
Leu															
1185															
<210> SEQ ID NO 7															
<211> LENGTH: 1186															
<212> TYPE: PRT															
<213> ORGANISM: Segniliparus rotundus															
<400> SEQUENCE: 7															
Met	Thr	Gln	Ser	His	Thr	Gln	Gly	Pro	Gln	Ala	Ser	Ala	Ala	His	Ser
1				5					10					15	
Arg	Leu	Ala	Arg	Arg	Ala	Ala	Glu	Leu	Leu	Ala	Thr	Asp	Pro	Gln	Ala
		20					25					30			
Ala	Ala	Thr	Leu	Pro	Asp	Pro	Glu	Val	Val	Arg	Gln	Ala	Thr	Arg	Pro
		35					40				45				
Gly	Leu	Arg	Leu	Ala	Glu	Arg	Val	Asp	Ala	Ile	Leu	Ser	Gly	Tyr	Ala
	50					55					60				
Asp	Arg	Pro	Ala	Leu	Gly	Gln	Arg	Ser	Phe	Gln	Thr	Val	Lys	Asp	Pro
65				70					75					80	
Ile	Thr	Gly	Arg	Ser	Ser	Val	Glu	Leu	Leu	Pro	Thr	Phe	Asp	Thr	Ile
			85						90					95	
Thr	Tyr	Arg	Glu	Leu	Arg	Glu	Arg	Ala	Thr	Ala	Ile	Ala	Ser	Asp	Leu
			100					105					110		
Ala	His	His	Pro	Gln	Ala	Pro	Ala	Lys	Pro	Gly	Asp	Phe	Leu	Ala	Ser
		115					120					125			
Ile	Gly	Phe	Ile	Ser	Val	Asp	Tyr	Val	Ala	Ile	Asp	Ile	Ala	Gly	Val
	130					135					140				
Phe	Ala	Gly	Leu	Thr	Ala	Val	Pro	Leu	Gln	Thr	Gly	Ala	Thr	Leu	Ala
145				150					155					160	
Thr	Leu	Thr	Ala	Ile	Thr	Ala	Glu	Thr	Ala	Pro	Thr	Leu	Phe	Ala	Ala
			165					170					175		
Ser	Ile	Glu	His	Leu	Pro	Thr	Ala	Val	Asp	Ala	Val	Leu	Ala	Thr	Pro
			180					185					190		
Ser	Val	Arg	Arg	Leu	Leu	Val	Phe	Asp	Tyr	Arg	Ala	Gly	Ser	Asp	Glu
		195					200					205			
Asp	Arg	Glu	Ala	Val	Glu	Ala	Ala	Lys	Arg	Lys	Ile	Ala	Asp	Ala	Gly
	210					215					220				
Ser	Ser	Val	Leu	Val	Asp	Val	Leu	Asp	Glu	Val	Ile	Ala	Arg	Gly	Lys
225				230					235					240	
Ser	Ala	Pro	Lys	Ala	Pro	Leu	Pro	Pro	Ala	Thr	Asp	Ala	Gly	Asp	Asp
			245						250				255		
Ser	Leu	Ser	Leu	Leu	Ile	Tyr	Thr	Ser	Gly	Ser	Thr	Gly	Thr	Pro	Lys
			260					265					270		
Gly	Ala	Met	Tyr	Pro	Glu	Arg	Asn	Val	Ala	His	Phe	Trp	Gly	Gly	Val
		275					280					285			
Trp	Ala	Ala	Ala	Phe	Asp	Glu	Asp	Ala	Ala	Pro	Pro	Val	Pro	Ala	Ile
	290					295						300			

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Asn	Ile	Thr	Phe	Leu	Pro	Leu	Ser	His	Val	Ala	Ser	Arg	Leu	Ser	Leu	305	310	315	320
Met	Pro	Thr	Leu	Ala	Arg	Gly	Gly	Leu	Met	His	Phe	Val	Ala	Lys	Ser	325	330	335	
Asp	Leu	Ser	Thr	Leu	Phe	Glu	Asp	Leu	Lys	Leu	Ala	Arg	Pro	Thr	Asn	340	345	350	
Leu	Phe	Leu	Val	Pro	Arg	Val	Val	Glu	Met	Leu	Tyr	Gln	His	Tyr	Gln	355	360	365	
Ser	Glu	Leu	Asp	Arg	Arg	Gly	Val	Gln	Asp	Gly	Thr	Arg	Glu	Ala	Glu	370	375	380	
Ala	Val	Lys	Asp	Asp	Leu	Arg	Thr	Gly	Leu	Leu	Gly	Gly	Arg	Ile	Leu	385	390	395	400
Thr	Ala	Gly	Phe	Gly	Ser	Ala	Pro	Leu	Ser	Ala	Glu	Leu	Ala	Gly	Phe	405	410	415	
Ile	Glu	Ser	Leu	Leu	Gln	Ile	His	Leu	Val	Asp	Gly	Tyr	Gly	Ser	Thr	420	425	430	
Glu	Ala	Gly	Pro	Val	Trp	Arg	Asp	Gly	Tyr	Leu	Val	Lys	Pro	Pro	Val	435	440	445	
Thr	Asp	Tyr	Lys	Leu	Ile	Asp	Val	Pro	Glu	Leu	Gly	Tyr	Phe	Ser	Thr	450	455	460	
Asp	Ser	Pro	His	Pro	Arg	Gly	Glu	Leu	Ala	Ile	Lys	Thr	Gln	Thr	Ile	465	470	475	480
Leu	Pro	Gly	Tyr	Tyr	Lys	Arg	Pro	Glu	Thr	Thr	Ala	Glu	Val	Phe	Asp	485	490	495	
Glu	Asp	Gly	Phe	Tyr	Leu	Thr	Gly	Asp	Val	Val	Ala	Gln	Ile	Gly	Pro	500	505	510	
Glu	Gln	Phe	Ala	Tyr	Val	Asp	Arg	Arg	Lys	Asn	Val	Leu	Lys	Leu	Ser	515	520	525	
Gln	Gly	Glu	Phe	Val	Thr	Leu	Ala	Lys	Leu	Glu	Ala	Ala	Tyr	Ser	Ser	530	535	540	
Ser	Pro	Leu	Val	Arg	Gln	Leu	Phe	Val	Tyr	Gly	Ser	Ser	Glu	Arg	Ser	545	550	555	560
Tyr	Leu	Leu	Ala	Val	Ile	Val	Pro	Thr	Pro	Asp	Ala	Leu	Lys	Lys	Phe	565	570	575	
Gly	Val	Gly	Glu	Ala	Ala	Lys	Ala	Ala	Leu	Gly	Glu	Ser	Leu	Gln	Lys	580	585	590	
Ile	Ala	Arg	Asp	Glu	Gly	Leu	Gln	Ser	Tyr	Glu	Val	Pro	Arg	Asp	Phe	595	600	605	
Ile	Ile	Glu	Thr	Asp	Pro	Phe	Thr	Val	Glu	Asn	Gly	Leu	Leu	Ser	Asp	610	615	620	
Ala	Arg	Lys	Ser	Leu	Arg	Pro	Lys	Leu	Lys	Glu	His	Tyr	Gly	Glu	Arg	625	630	635	640
Leu	Glu	Ala	Met	Tyr	Lys	Glu	Leu	Ala	Asp	Gly	Gln	Ala	Asn	Glu	Leu	645	650	655	
Arg	Asp	Ile	Arg	Arg	Gly	Val	Gln	Gln	Arg	Pro	Thr	Leu	Glu	Thr	Val	660	665	670	
Arg	Arg	Ala	Ala	Ala	Ala	Met	Leu	Gly	Ala	Ser	Ala	Ala	Glu	Ile	Lys	675	680	685	
Pro	Asp	Ala	His	Phe	Thr	Asp	Leu	Gly	Gly	Asp	Ser	Leu	Ser	Ala	Leu	690	695	700	
Thr	Phe	Ser	Asn	Phe	Leu	His	Asp	Leu	Phe	Glu	Val	Asp	Val	Pro	Val	705	710	715	720
Gly	Val	Ile	Val	Ser	Ala	Ala	Asn	Thr	Leu	Gly	Ser	Val	Ala	Glu	His				

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725					730					735					
Ile	Asp	Ala	Gln	Leu	Ala	Gly	Gly	Arg	Ala	Arg	Pro	Thr	Phe	Ala	Thr
			740					745					750		
Val	His	Gly	Lys	Gly	Ser	Thr	Thr	Ile	Lys	Ala	Ser	Asp	Leu	Thr	Leu
		755					760					765			
Asp	Lys	Phe	Ile	Asp	Glu	Gln	Thr	Leu	Glu	Ala	Ala	Lys	His	Leu	Pro
	770					775					780				
Lys	Pro	Ala	Asp	Pro	Pro	Arg	Thr	Val	Leu	Leu	Thr	Gly	Ala	Asn	Gly
	785					790					795				800
Trp	Leu	Gly	Arg	Phe	Leu	Ala	Leu	Glu	Trp	Leu	Glu	Arg	Leu	Ala	Pro
				805					810					815	
Ala	Gly	Gly	Lys	Leu	Ile	Thr	Ile	Val	Arg	Gly	Lys	Asp	Ala	Ala	Gln
			820					825					830		
Ala	Lys	Ala	Arg	Leu	Asp	Ala	Ala	Tyr	Glu	Ser	Gly	Asp	Pro	Lys	Leu
		835					840					845			
Ala	Gly	His	Tyr	Gln	Asp	Leu	Ala	Ala	Thr	Thr	Leu	Glu	Val	Leu	Ala
		850				855						860			
Gly	Asp	Phe	Ser	Glu	Pro	Arg	Leu	Gly	Leu	Asp	Glu	Ala	Thr	Trp	Asn
				870							875				880
Arg	Leu	Ala	Asp	Glu	Val	Asp	Phe	Ile	Ser	His	Pro	Gly	Ala	Leu	Val
				885					890					895	
Asn	His	Val	Leu	Pro	Tyr	Asn	Gln	Leu	Phe	Gly	Pro	Asn	Val	Ala	Gly
			900					905					910		
Val	Ala	Glu	Ile	Ile	Lys	Leu	Ala	Ile	Thr	Thr	Arg	Ile	Lys	Pro	Val
		915					920					925			
Thr	Tyr	Leu	Ser	Thr	Val	Ala	Val	Ala	Ala	Gly	Val	Glu	Pro	Ser	Ala
		930				935						940			
Leu	Asp	Glu	Asp	Gly	Asp	Ile	Arg	Thr	Val	Ser	Ala	Glu	Arg	Ser	Val
				950							955				960
Asp	Glu	Gly	Tyr	Ala	Asn	Gly	Tyr	Gly	Asn	Ser	Lys	Trp	Gly	Gly	Glu
				965					970					975	
Val	Leu	Leu	Arg	Glu	Ala	His	Asp	Arg	Thr	Gly	Leu	Pro	Val	Arg	Val
			980					985					990		
Phe	Arg	Ser	Asp	Met	Ile	Leu	Ala	His	Gln	Lys	Tyr	Thr	Gly	Gln	Val
		995					1000						1005		
Asn	Ala	Thr	Asp	Gln	Phe	Thr	Arg	Leu	Val	Gln	Ser	Leu	Leu	Ala	Thr
		1010				1015						1020			
Gly	Leu	Ala	Pro	Lys	Ser	Phe	Tyr	Glu	Leu	Asp	Ala	Gln	Gly	Asn	Arg
				1025		1030					1035				1040
Gln	Arg	Ala	His	Tyr	Asp	Gly	Ile	Pro	Val	Asp	Phe	Thr	Ala	Glu	Ser
				1045					1050					1055	
Ile	Thr	Thr	Leu	Gly	Gly	Asp	Gly	Leu	Glu	Gly	Tyr	Arg	Ser	Tyr	Asn
			1060					1065						1070	
Val	Phe	Asn	Pro	His	Arg	Asp	Gly	Val	Gly	Leu	Asp	Glu	Phe	Val	Asp
		1075					1080					1085			
Trp	Leu	Ile	Glu	Ala	Gly	His	Pro	Ile	Thr	Arg	Ile	Asp	Asp	Tyr	Asp
		1090				1095						1100			
Gln	Trp	Leu	Ser	Arg	Phe	Glu	Thr	Ser	Leu	Arg	Gly	Leu	Pro	Glu	Ser
				1105		1110					1115				1120
Lys	Arg	Gln	Ala	Ser	Val	Leu	Pro	Leu	Leu	His	Ala	Phe	Ala	Arg	Pro
				1125					1130					1135	
Gly	Pro	Ala	Val	Asp	Gly	Ser	Pro	Phe	Arg	Asn	Thr	Val	Phe	Arg	Thr
			1140					1145						1150	

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Asp Val Gln Lys Ala Lys Ile Gly Ala Glu His Asp Ile Pro His Leu  
 1155 1160 1165

Gly Lys Ala Leu Val Leu Lys Tyr Ala Asp Asp Ile Lys Gln Leu Gly  
 1170 1175 1180

Leu Leu  
 1185

<210> SEQ ID NO 8  
 <211> LENGTH: 459  
 <212> TYPE: PRT  
 <213> ORGANISM: Chromobacterium violaceum

<400> SEQUENCE: 8

Met Gln Lys Gln Arg Thr Thr Ser Gln Trp Arg Glu Leu Asp Ala Ala  
 1 5 10 15

His His Leu His Pro Phe Thr Asp Thr Ala Ser Leu Asn Gln Ala Gly  
 20 25 30

Ala Arg Val Met Thr Arg Gly Glu Gly Val Tyr Leu Trp Asp Ser Glu  
 35 40 45

Gly Asn Lys Ile Ile Asp Gly Met Ala Gly Leu Trp Cys Val Asn Val  
 50 55 60

Gly Tyr Gly Arg Lys Asp Phe Ala Glu Ala Ala Arg Arg Gln Met Glu  
 65 70 75 80

Glu Leu Pro Phe Tyr Asn Thr Phe Phe Lys Thr Thr His Pro Ala Val  
 85 90 95

Val Glu Leu Ser Ser Leu Leu Ala Glu Val Thr Pro Ala Gly Phe Asp  
 100 105 110

Arg Val Phe Tyr Thr Asn Ser Gly Ser Glu Ser Val Asp Thr Met Ile  
 115 120 125

Arg Met Val Arg Arg Tyr Trp Asp Val Gln Gly Lys Pro Glu Lys Lys  
 130 135 140

Thr Leu Ile Gly Arg Trp Asn Gly Tyr His Gly Ser Thr Ile Gly Gly  
 145 150 155 160

Ala Ser Leu Gly Gly Met Lys Tyr Met His Glu Gln Gly Asp Leu Pro  
 165 170 175

Ile Pro Gly Met Ala His Ile Glu Gln Pro Trp Trp Tyr Lys His Gly  
 180 185 190

Lys Asp Met Thr Pro Asp Glu Phe Gly Val Val Ala Ala Arg Trp Leu  
 195 200 205

Glu Glu Lys Ile Leu Glu Ile Gly Ala Asp Lys Val Ala Ala Phe Val  
 210 215 220

Gly Glu Pro Ile Gln Gly Ala Gly Gly Val Ile Val Pro Pro Ala Thr  
 225 230 235 240

Tyr Trp Pro Glu Ile Glu Arg Ile Cys Arg Lys Tyr Asp Val Leu Leu  
 245 250 255

Val Ala Asp Glu Val Ile Cys Gly Phe Gly Arg Thr Gly Glu Trp Phe  
 260 265 270

Gly His Gln His Phe Gly Phe Gln Pro Asp Leu Phe Thr Ala Ala Lys  
 275 280 285

Gly Leu Ser Ser Gly Tyr Leu Pro Ile Gly Ala Val Phe Val Gly Lys  
 290 295 300

Arg Val Ala Glu Gly Leu Ile Ala Gly Gly Asp Phe Asn His Gly Phe  
 305 310 315 320

Thr Tyr Ser Gly His Pro Val Cys Ala Ala Val Ala His Ala Asn Val





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Lys Thr Ile Ile Ser Arg Val Asn Gly Tyr His Gly Ser Thr Val Ala  
 145 150 155 160  
 Gly Ala Ser Leu Gly Gly Met Thr Tyr Met His Glu Gln Gly Asp Leu  
 165 170 175  
 Pro Ile Pro Gly Val Val His Ile Pro Gln Pro Tyr Trp Phe Gly Glu  
 180 185 190  
 Gly Gly Asp Met Thr Pro Asp Glu Phe Gly Ile Trp Ala Ala Glu Gln  
 195 200 205  
 Leu Glu Lys Lys Ile Leu Glu Leu Gly Val Glu Asn Val Gly Ala Phe  
 210 215 220  
 Ile Ala Glu Pro Ile Gln Gly Ala Gly Gly Val Ile Val Pro Pro Asp  
 225 230 235 240  
 Ser Tyr Trp Pro Lys Ile Lys Glu Ile Leu Ser Arg Tyr Asp Ile Leu  
 245 250 255  
 Phe Ala Ala Asp Glu Val Ile Cys Gly Phe Gly Arg Thr Ser Glu Trp  
 260 265 270  
 Phe Gly Ser Asp Phe Tyr Gly Leu Arg Pro Asp Met Met Thr Ile Ala  
 275 280 285  
 Lys Gly Leu Thr Ser Gly Tyr Val Pro Met Gly Gly Leu Ile Val Arg  
 290 295 300  
 Asp Glu Ile Val Ala Val Leu Asn Glu Gly Gly Asp Phe Asn His Gly  
 305 310 315 320  
 Phe Thr Tyr Ser Gly His Pro Val Ala Ala Val Ala Leu Glu Asn  
 325 330 335  
 Ile Arg Ile Leu Arg Glu Glu Lys Ile Val Glu Arg Val Arg Ser Glu  
 340 345 350  
 Thr Ala Pro Tyr Leu Gln Lys Arg Leu Arg Glu Leu Ser Asp His Pro  
 355 360 365  
 Leu Val Gly Glu Val Arg Gly Val Gly Leu Leu Gly Ala Ile Glu Leu  
 370 375 380  
 Val Lys Asp Lys Thr Thr Arg Glu Arg Tyr Thr Asp Lys Gly Ala Gly  
 385 390 395 400  
 Met Ile Cys Arg Thr Phe Cys Phe Asp Asn Gly Leu Ile Met Arg Ala  
 405 410 415  
 Val Gly Asp Thr Met Ile Ile Ala Pro Pro Leu Val Ile Ser Phe Ala  
 420 425 430  
 Gln Ile Asp Glu Leu Val Glu Lys Ala Arg Thr Cys Leu Asp Leu Thr  
 435 440 445  
 Leu Ala Val Leu Gln Gly  
 450

&lt;210&gt; SEQ ID NO 11

&lt;211&gt; LENGTH: 467

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Rhodobacter sphaeroides

&lt;400&gt; SEQUENCE: 11

Met Thr Arg Asn Asp Ala Thr Asn Ala Ala Gly Ala Val Gly Ala Ala  
 1 5 10 15  
 Met Arg Asp His Ile Leu Leu Pro Ala Gln Glu Met Ala Lys Leu Gly  
 20 25 30  
 Lys Ser Ala Gln Pro Val Leu Thr His Ala Glu Gly Ile Tyr Val His  
 35 40 45  
 Thr Glu Asp Gly Arg Arg Leu Ile Asp Gly Pro Ala Gly Met Trp Cys

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50	55	60
Ala Gln Val Gly Tyr Gly Arg Arg Glu Ile Val Asp Ala Met Ala His 65 70 75 80		
Gln Ala Met Val Leu Pro Tyr Ala Ser Pro Trp Tyr Met Ala Thr Ser 85 90 95		
Pro Ala Ala Arg Leu Ala Glu Lys Ile Ala Thr Leu Thr Pro Gly Asp 100 105 110		
Leu Asn Arg Ile Phe Phe Thr Thr Gly Gly Ser Thr Ala Val Asp Ser 115 120 125		
Ala Leu Arg Phe Ser Glu Phe Tyr Asn Asn Val Leu Gly Arg Pro Gln 130 135 140		
Lys Lys Arg Ile Ile Val Arg Tyr Asp Gly Tyr His Gly Ser Thr Ala 145 150 155 160		
Leu Thr Ala Ala Cys Thr Gly Arg Thr Gly Asn Trp Pro Asn Phe Asp 165 170 175		
Ile Ala Gln Asp Arg Ile Ser Phe Leu Ser Ser Pro Asn Pro Arg His 180 185 190		
Ala Gly Asn Arg Ser Gln Glu Ala Phe Leu Asp Asp Leu Val Gln Glu 195 200 205		
Phe Glu Asp Arg Ile Glu Ser Leu Gly Pro Asp Thr Ile Ala Ala Phe 210 215 220		
Leu Ala Glu Pro Ile Leu Ala Ser Gly Gly Val Ile Ile Pro Pro Ala 225 230 235 240		
Gly Tyr His Ala Arg Phe Lys Ala Ile Cys Glu Lys His Asp Ile Leu 245 250 255		
Tyr Ile Ser Asp Glu Val Val Thr Gly Phe Gly Arg Cys Gly Glu Trp 260 265 270		
Phe Ala Ser Glu Lys Val Phe Gly Val Val Pro Asp Ile Ile Thr Phe 275 280 285		
Ala Lys Gly Val Thr Ser Gly Tyr Val Pro Leu Gly Gly Leu Ala Ile 290 295 300		
Ser Glu Ala Val Leu Ala Arg Ile Ser Gly Glu Asn Ala Lys Gly Ser 305 310 315 320		
Trp Phe Thr Asn Gly Tyr Thr Tyr Ser Asn Gln Pro Val Ala Cys Ala 325 330 335		
Ala Ala Leu Ala Asn Ile Glu Leu Met Glu Arg Glu Gly Ile Val Asp 340 345 350		
Gln Ala Arg Glu Met Ala Asp Tyr Phe Ala Ala Ala Leu Ala Ser Leu 355 360 365		
Arg Asp Leu Pro Gly Val Ala Glu Thr Arg Ser Val Gly Leu Val Gly 370 375 380		
Cys Val Gln Cys Leu Leu Asp Pro Thr Arg Ala Asp Gly Thr Ala Glu 385 390 395 400		
Asp Lys Ala Phe Thr Leu Lys Ile Asp Glu Arg Cys Phe Glu Leu Gly 405 410 415		
Leu Ile Val Arg Pro Leu Gly Asp Leu Cys Val Ile Ser Pro Pro Leu 420 425 430		
Ile Ile Ser Arg Ala Gln Ile Asp Glu Met Val Ala Ile Met Arg Gln 435 440 445		
Ala Ile Thr Glu Val Ser Ala Ala His Gly Leu Thr Ala Lys Glu Pro 450 455 460		
Ala Ala Val 465		



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<210> SEQ ID NO 12  
 <211> LENGTH: 459  
 <212> TYPE: PRT  
 <213> ORGANISM: Escherichia coli  
  
 <400> SEQUENCE: 12

Met Asn Arg Leu Pro Ser Ser Ala Ser Ala Leu Ala Cys Ser Ala His  
 1 5 10 15  
 Ala Leu Asn Leu Ile Glu Lys Arg Thr Leu Asp His Glu Glu Met Lys  
 20 25 30  
 Ala Leu Asn Arg Glu Val Ile Glu Tyr Phe Lys Glu His Val Asn Pro  
 35 40 45  
 Gly Phe Leu Glu Tyr Arg Lys Ser Val Thr Ala Gly Gly Asp Tyr Gly  
 50 55 60  
 Ala Val Glu Trp Gln Ala Gly Ser Leu Asn Thr Leu Val Asp Thr Gln  
 65 70 75 80  
 Gly Gln Glu Phe Ile Asp Cys Leu Gly Gly Phe Gly Ile Phe Asn Val  
 85 90 95  
 Gly His Arg Asn Pro Val Val Val Ser Ala Val Gln Asn Gln Leu Ala  
 100 105 110  
 Lys Gln Pro Leu His Ser Gln Glu Leu Leu Asp Pro Leu Arg Ala Met  
 115 120 125  
 Leu Ala Lys Thr Leu Ala Ala Leu Thr Pro Gly Lys Leu Lys Tyr Ser  
 130 135 140  
 Phe Phe Cys Asn Ser Gly Thr Glu Ser Val Glu Ala Ala Leu Lys Leu  
 145 150 155 160  
 Ala Lys Ala Tyr Gln Ser Pro Arg Gly Lys Phe Thr Phe Ile Ala Thr  
 165 170 175  
 Ser Gly Ala Phe His Gly Lys Ser Leu Gly Ala Leu Ser Ala Thr Ala  
 180 185 190  
 Lys Ser Thr Phe Arg Lys Pro Phe Met Pro Leu Leu Pro Gly Phe Arg  
 195 200 205  
 His Val Pro Phe Gly Asn Ile Glu Ala Met Arg Thr Ala Leu Asn Glu  
 210 215 220  
 Cys Lys Lys Thr Gly Asp Asp Val Ala Ala Val Ile Leu Glu Pro Ile  
 225 230 235 240  
 Gln Gly Glu Gly Gly Val Ile Leu Pro Pro Pro Gly Tyr Leu Thr Ala  
 245 250 255  
 Val Arg Lys Leu Cys Asp Glu Phe Gly Ala Leu Met Ile Leu Asp Glu  
 260 265 270  
 Val Gln Thr Gly Met Gly Arg Thr Gly Lys Met Phe Ala Cys Glu His  
 275 280 285  
 Glu Asn Val Gln Pro Asp Ile Leu Cys Leu Ala Lys Ala Leu Gly Gly  
 290 295 300  
 Gly Val Met Pro Ile Gly Ala Thr Ile Ala Thr Glu Glu Val Phe Ser  
 305 310 315 320  
 Val Leu Phe Asp Asn Pro Phe Leu His Thr Thr Thr Phe Gly Gly Asn  
 325 330 335  
 Pro Leu Ala Cys Ala Ala Ala Leu Ala Thr Ile Asn Val Leu Leu Glu  
 340 345 350  
 Gln Asn Leu Pro Ala Gln Ala Glu Gln Lys Gly Asp Met Leu Leu Asp  
 355 360 365  
 Gly Phe Arg Gln Leu Ala Arg Glu Tyr Pro Asp Leu Val Gln Glu Ala



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Ala Gly Phe Phe Pro Met Gly Ala Val Ile Leu Gly Pro Glu Leu Ser  
 290 295 300

Lys Arg Leu Glu Thr Ala Ile Glu Ala Ile Glu Glu Phe Pro His Gly  
 305 310 315 320

Phe Thr Ala Ser Gly His Pro Val Gly Cys Ala Ile Ala Leu Lys Ala  
 325 330 335

Ile Asp Val Val Met Asn Glu Gly Leu Ala Glu Asn Val Arg Arg Leu  
 340 345 350

Ala Pro Arg Phe Glu Glu Arg Leu Lys His Ile Ala Glu Arg Pro Asn  
 355 360 365

Ile Gly Glu Tyr Arg Gly Ile Gly Phe Met Trp Ala Leu Glu Ala Val  
 370 375 380

Lys Asp Lys Ala Ser Lys Thr Pro Phe Asp Gly Asn Leu Ser Val Ser  
 385 390 395 400

Glu Arg Ile Ala Asn Thr Cys Thr Asp Leu Gly Leu Ile Cys Arg Pro  
 405 410 415

Leu Gly Gln Ser Val Val Leu Cys Pro Pro Phe Ile Leu Thr Glu Ala  
 420 425 430

Gln Met Asp Glu Met Phe Asp Lys Leu Glu Lys Ala Leu Asp Lys Val  
 435 440 445

Phe Ala Glu Val Ala  
 450

<210> SEQ ID NO 14  
 <211> LENGTH: 224  
 <212> TYPE: PRT  
 <213> ORGANISM: Bacillus subtilis

<400> SEQUENCE: 14

Met Lys Ile Tyr Gly Ile Tyr Met Asp Arg Pro Leu Ser Gln Glu Glu  
 1 5 10 15

Asn Glu Arg Phe Met Ser Phe Ile Ser Pro Glu Lys Arg Glu Lys Cys  
 20 25 30

Arg Arg Phe Tyr His Lys Glu Asp Ala His Arg Thr Leu Leu Gly Asp  
 35 40 45

Val Leu Val Arg Ser Val Ile Ser Arg Gln Tyr Gln Leu Asp Lys Ser  
 50 55 60

Asp Ile Arg Phe Ser Thr Gln Glu Tyr Gly Lys Pro Cys Ile Pro Asp  
 65 70 75 80

Leu Pro Asp Ala His Phe Asn Ile Ser His Ser Gly Arg Trp Val Ile  
 85 90 95

Cys Ala Phe Asp Ser Gln Pro Ile Gly Ile Asp Ile Glu Lys Thr Lys  
 100 105 110

Pro Ile Ser Leu Glu Ile Ala Lys Arg Phe Phe Ser Lys Thr Glu Tyr  
 115 120 125

Ser Asp Leu Leu Ala Lys Asp Lys Asp Glu Gln Thr Asp Tyr Phe Tyr  
 130 135 140

His Leu Trp Ser Met Lys Glu Ser Phe Ile Lys Gln Glu Gly Lys Gly  
 145 150 155 160

Leu Ser Leu Pro Leu Asp Ser Phe Ser Val Arg Leu His Gln Asp Gly  
 165 170 175

Gln Val Ser Ile Glu Leu Pro Asp Ser His Ser Pro Cys Tyr Ile Lys  
 180 185 190

Thr Tyr Glu Val Asp Pro Gly Tyr Lys Met Ala Val Cys Ala Ala His  
 195 200 205

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Pro Asp Phe Pro Glu Asp Ile Thr Met Val Ser Tyr Glu Glu Leu Leu  
 210 215 220

<210> SEQ ID NO 15  
 <211> LENGTH: 222  
 <212> TYPE: PRT  
 <213> ORGANISM: Nocardia sp. NRRL 5646

<400> SEQUENCE: 15

Met Ile Glu Thr Ile Leu Pro Ala Gly Val Glu Ser Ala Glu Leu Leu  
 1 5 10 15  
 Glu Tyr Pro Glu Asp Leu Lys Ala His Pro Ala Glu Glu His Leu Ile  
 20 25 30  
 Ala Lys Ser Val Glu Lys Arg Arg Arg Asp Phe Ile Gly Ala Arg His  
 35 40 45  
 Cys Ala Arg Leu Ala Leu Ala Glu Leu Gly Glu Pro Pro Val Ala Ile  
 50 55 60  
 Gly Lys Gly Glu Arg Gly Ala Pro Ile Trp Pro Arg Gly Val Val Gly  
 65 70 75 80  
 Ser Leu Thr His Cys Asp Gly Tyr Arg Ala Ala Ala Val Ala His Lys  
 85 90 95  
 Met Arg Phe Arg Ser Ile Gly Ile Asp Ala Glu Pro His Ala Thr Leu  
 100 105 110  
 Pro Glu Gly Val Leu Asp Ser Val Ser Leu Pro Pro Glu Arg Glu Trp  
 115 120 125  
 Leu Lys Thr Thr Asp Ser Ala Leu His Leu Asp Arg Leu Leu Phe Cys  
 130 135 140  
 Ala Lys Glu Ala Thr Tyr Lys Ala Trp Trp Pro Leu Thr Ala Arg Trp  
 145 150 155 160  
 Leu Gly Phe Glu Glu Ala His Ile Thr Phe Glu Ile Glu Asp Gly Ser  
 165 170 175  
 Ala Asp Ser Gly Asn Gly Thr Phe His Ser Glu Leu Leu Val Pro Gly  
 180 185 190  
 Gln Thr Asn Asp Gly Gly Thr Pro Leu Leu Ser Phe Asp Gly Arg Trp  
 195 200 205  
 Leu Ile Ala Asp Gly Phe Ile Leu Thr Ala Ile Ala Tyr Ala  
 210 215 220

<210> SEQ ID NO 16  
 <211> LENGTH: 338  
 <212> TYPE: PRT  
 <213> ORGANISM: Bacillus subtilis

<400> SEQUENCE: 16

Met Ala Arg Lys Leu Phe Thr Pro Ile Thr Ile Lys Asp Met Thr Leu  
 1 5 10 15  
 Lys Asn Arg Ile Val Met Ser Pro Met Cys Met Tyr Ser Ser His Glu  
 20 25 30  
 Lys Asp Gly Lys Leu Thr Pro Phe His Met Ala His Tyr Ile Ser Arg  
 35 40 45  
 Ala Ile Gly Gln Val Gly Leu Ile Ile Val Glu Ala Ser Ala Val Asn  
 50 55 60  
 Pro Gln Gly Arg Ile Thr Asp Gln Asp Leu Gly Ile Trp Ser Asp Glu  
 65 70 75 80  
 His Ile Glu Gly Phe Ala Lys Leu Thr Glu Gln Val Lys Glu Gln Gly  
 85 90 95

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Ser Lys Ile Gly Ile Gln Leu Ala His Ala Gly Arg Lys Ala Glu Leu  
 100 105 110  
 Glu Gly Asp Ile Phe Ala Pro Ser Ala Ile Ala Phe Asp Glu Gln Ser  
 115 120 125  
 Ala Thr Pro Val Glu Met Ser Ala Glu Lys Val Lys Glu Thr Val Gln  
 130 135 140  
 Glu Phe Lys Gln Ala Ala Ala Arg Ala Lys Glu Ala Gly Phe Asp Val  
 145 150 155 160  
 Ile Glu Ile His Ala Ala His Gly Tyr Leu Ile His Glu Phe Leu Ser  
 165 170 175  
 Pro Leu Ser Asn His Arg Thr Asp Glu Tyr Gly Gly Ser Pro Glu Asn  
 180 185 190  
 Arg Tyr Arg Phe Leu Arg Glu Ile Ile Asp Glu Val Lys Gln Val Trp  
 195 200 205  
 Asp Gly Pro Leu Phe Val Arg Val Ser Ala Ser Asp Tyr Thr Asp Lys  
 210 215 220  
 Gly Leu Asp Ile Ala Asp His Ile Gly Phe Ala Lys Trp Met Lys Glu  
 225 230 235 240  
 Gln Gly Val Asp Leu Ile Asp Cys Ser Ser Gly Ala Leu Val His Ala  
 245 250 255  
 Asp Ile Asn Val Phe Pro Gly Tyr Gln Val Ser Phe Ala Glu Lys Ile  
 260 265 270  
 Arg Glu Gln Ala Asp Met Ala Thr Gly Ala Val Gly Met Ile Thr Asp  
 275 280 285  
 Gly Ser Met Ala Glu Glu Ile Leu Gln Asn Gly Arg Ala Asp Leu Ile  
 290 295 300  
 Phe Ile Gly Arg Glu Leu Leu Arg Asp Pro Phe Phe Ala Arg Thr Ala  
 305 310 315 320  
 Ala Lys Gln Leu Asn Thr Glu Ile Pro Ala Pro Val Gln Tyr Glu Arg  
 325 330 335

Gly Trp

&lt;210&gt; SEQ ID NO 17

&lt;211&gt; LENGTH: 363

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Pseudomonas putida

&lt;400&gt; SEQUENCE: 17

Met Ser Ala Leu Phe Glu Pro Tyr Thr Leu Lys Asp Val Thr Leu Arg  
 1 5 10 15  
 Asn Arg Ile Ala Ile Pro Pro Met Cys Gln Tyr Met Ala Glu Asp Gly  
 20 25 30  
 Met Ile Asn Asp Trp His His Val His Leu Ala Gly Leu Ala Arg Gly  
 35 40 45  
 Gly Ala Gly Leu Leu Val Val Glu Ala Thr Ala Val Ala Pro Glu Gly  
 50 55 60  
 Arg Ile Thr Pro Gly Cys Ala Gly Ile Trp Ser Asp Ala His Ala Gln  
 65 70 75 80  
 Ala Phe Val Pro Val Val Gln Ala Ile Lys Ala Ala Gly Ser Val Pro  
 85 90 95  
 Gly Ile Gln Ile Ala His Ala Gly Arg Lys Ala Ser Ala Asn Arg Pro  
 100 105 110  
 Trp Glu Gly Asp Asp His Ile Ala Ala Asp Asp Ala Arg Gly Trp Glu  
 115 120 125

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Thr Ile Ala Pro Ser Ala Ile Ala Phe Gly Ala His Leu Pro Lys Val  
 130 135 140  
 Pro Arg Glu Met Thr Leu Asp Asp Ile Ala Arg Val Lys Gln Asp Phe  
 145 150 155 160  
 Val Asp Ala Ala Arg Arg Ala Arg Asp Ala Gly Phe Glu Trp Ile Glu  
 165 170 175  
 Leu His Phe Ala His Gly Tyr Leu Gly Gln Ser Phe Phe Ser Glu His  
 180 185 190  
 Ser Asn Lys Arg Thr Asp Ala Tyr Gly Gly Ser Phe Asp Asn Arg Ser  
 195 200 205  
 Arg Phe Leu Leu Glu Thr Leu Ala Ala Val Arg Glu Val Trp Pro Glu  
 210 215 220  
 Asn Leu Pro Leu Thr Ala Arg Phe Gly Val Leu Glu Tyr Asp Gly Arg  
 225 230 235 240  
 Asp Glu Gln Thr Leu Glu Glu Ser Ile Glu Leu Ala Arg Arg Phe Lys  
 245 250 255  
 Ala Gly Gly Leu Asp Leu Leu Ser Val Ser Val Gly Phe Thr Ile Pro  
 260 265 270  
 Asp Thr Asn Ile Pro Trp Gly Pro Ala Phe Met Gly Pro Ile Ala Glu  
 275 280 285  
 Arg Val Arg Arg Glu Ala Lys Leu Pro Val Thr Ser Ala Trp Gly Phe  
 290 295 300  
 Gly Thr Pro Gln Leu Ala Glu Ala Ala Leu Gln Ala Asn Gln Leu Asp  
 305 310 315 320  
 Leu Val Ser Val Gly Arg Ala His Leu Ala Asp Pro His Trp Ala Tyr  
 325 330 335  
 Phe Ala Ala Lys Glu Leu Gly Val Glu Lys Ala Ser Trp Thr Leu Pro  
 340 345 350  
 Ala Pro Tyr Ala His Trp Leu Glu Arg Tyr Arg  
 355 360

&lt;210&gt; SEQ ID NO 18

&lt;211&gt; LENGTH: 398

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Kluyveromyces lactis

&lt;400&gt; SEQUENCE: 18

Met Ser Phe Met Asn Phe Glu Pro Lys Pro Leu Ala Asp Thr Asp Ile  
 1 5 10 15  
 Phe Lys Pro Ile Lys Ile Gly Asn Thr Glu Leu Lys His Arg Val Val  
 20 25 30  
 Met Pro Ala Leu Thr Arg Met Arg Ala Leu His Pro Gly Asn Val Pro  
 35 40 45  
 Asn Pro Asp Trp Ala Val Glu Tyr Tyr Arg Gln Arg Ser Gln Tyr Pro  
 50 55 60  
 Gly Thr Met Ile Ile Thr Glu Gly Ala Phe Pro Ser Ala Gln Ser Gly  
 65 70 75 80  
 Gly Tyr Asp Asn Ala Pro Gly Val Trp Ser Glu Glu Gln Leu Ala Gln  
 85 90 95  
 Trp Arg Lys Ile Phe Lys Ala Ile His Asp Asn Lys Ser Phe Val Trp  
 100 105 110  
 Val Gln Leu Trp Val Leu Gly Arg Gln Ala Phe Ala Asp Asn Leu Ala  
 115 120 125  
 Arg Asp Gly Leu Arg Tyr Asp Ser Ala Ser Asp Glu Val Tyr Met Gly

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130	135	140
Glu Asp Glu Lys Glu Arg Ala Ile Arg Ser Asn Asn Pro Gln His Gly 145 150 155 160		
Ile Thr Lys Asp Glu Ile Lys Gln Tyr Ile Arg Asp Tyr Val Asp Ala 165 170 175		
Ala Lys Lys Cys Ile Asp Ala Gly Ala Asp Gly Val Glu Ile His Ser 180 185 190		
Ala Asn Gly Tyr Leu Leu Asn Gln Phe Leu Asp Pro Ile Ser Asn Lys 195 200 205		
Arg Thr Asp Glu Tyr Gly Gly Ser Ile Glu Asn Arg Ala Arg Phe Val 210 215 220		
Leu Glu Val Val Asp Ala Val Val Asp Ala Val Gly Ala Glu Arg Thr 225 230 235 240		
Ser Ile Arg Phe Ser Pro Tyr Gly Val Phe Gly Thr Met Ser Gly Gly 245 250 255		
Ser Asp Pro Val Leu Val Ala Gln Phe Ala Tyr Val Leu Ala Glu Leu 260 265 270		
Glu Lys Arg Ala Lys Ala Gly Lys Arg Leu Ala Tyr Val Asp Leu Val 275 280 285		
Glu Pro Arg Val Thr Ser Pro Phe Gln Pro Glu Phe Glu Gly Trp Tyr 290 295 300		
Lys Gly Gly Thr Asn Glu Phe Val Tyr Ser Val Trp Lys Gly Asn Val 305 310 315 320		
Leu Arg Val Gly Asn Tyr Ala Leu Asp Pro Asp Ala Ala Ile Thr Asp 325 330 335		
Ser Lys Asn Pro Asn Thr Leu Ile Gly Tyr Gly Arg Ala Phe Ile Ala 340 345 350		
Asn Pro Asp Leu Val Glu Arg Leu Glu Lys Gly Leu Pro Leu Asn Gln 355 360 365		
Tyr Asp Arg Pro Ser Phe Tyr Lys Met Ser Ala Glu Gly Tyr Ile Asp 370 375 380		
Tyr Pro Thr Tyr Glu Glu Ala Val Ala Lys Gly Tyr Lys Lys 385 390 395		

&lt;210&gt; SEQ ID NO 19

&lt;211&gt; LENGTH: 387

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Lactobacillus casei

&lt;400&gt; SEQUENCE: 19

Met Ser Gly Tyr His Phe Leu Lys Pro Phe Thr Phe Lys His Gln Thr 1 5 10 15
Ile Thr Leu Lys Asn Arg Ile Val Ile Pro Pro Met Thr Thr Arg Leu 20 25 30
Ser Phe Glu Asp Gly Thr Val Thr Arg Asp Glu Ile Arg Tyr Tyr Gln 35 40 45
Gln Arg Ala Gly Gly Val Gly Met Phe Ile Thr Gly Thr Ala Asn Val 50 55 60
Asn Ala Leu Gly Lys Gly Phe Glu Gly Glu Leu Ser Val Ala Asp Asp 65 70 75 80
Arg Phe Ile Pro Gly Leu Ser Lys Leu Ala Ala Ala Met Lys Thr Gly 85 90 95
Gly Thr Lys Ala Ile Leu Gln Ile Phe Ser Ala Gly Arg Met Ser Asn 100 105 110





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Glu Trp Thr Lys Ile Phe Asn Ala Ile His Glu Lys Lys Ser Phe Val  
 100 105 110  
 Trp Val Gln Leu Trp Val Leu Gly Trp Ala Ala Phe Pro Asp Asn Leu  
 115 120 125  
 Ala Arg Asp Gly Leu Arg Tyr Asp Ser Ala Ser Asp Asn Val Phe Met  
 130 135 140  
 Asp Ala Glu Gln Glu Ala Lys Ala Lys Lys Ala Asn Asn Pro Gln His  
 145 150 155 160  
 Ser Leu Thr Lys Asp Glu Ile Lys Gln Tyr Ile Lys Glu Tyr Val Gln  
 165 170 175  
 Ala Ala Lys Asn Ser Ile Ala Ala Gly Ala Asp Gly Val Glu Ile His  
 180 185 190  
 Ser Ala Asn Gly Tyr Leu Leu Asn Gln Phe Leu Asp Pro His Ser Asn  
 195 200 205  
 Thr Arg Thr Asp Glu Tyr Gly Gly Ser Ile Glu Asn Arg Ala Arg Phe  
 210 215 220  
 Thr Leu Glu Val Val Asp Ala Leu Val Glu Ala Ile Gly His Glu Lys  
 225 230 235 240  
 Val Gly Leu Arg Leu Ser Pro Tyr Gly Val Phe Asn Ser Met Ser Gly  
 245 250 255  
 Gly Ala Glu Thr Gly Ile Val Ala Gln Tyr Ala Tyr Val Ala Gly Glu  
 260 265 270  
 Leu Glu Lys Arg Ala Lys Ala Gly Lys Arg Leu Ala Phe Val His Leu  
 275 280 285  
 Val Glu Pro Arg Val Thr Asn Pro Phe Leu Thr Glu Gly Glu Gly Glu  
 290 295 300  
 Tyr Glu Gly Gly Ser Asn Asp Phe Val Tyr Ser Ile Trp Lys Gly Pro  
 305 310 315 320  
 Val Ile Arg Ala Gly Asn Phe Ala Leu His Pro Glu Val Val Arg Glu  
 325 330 335  
 Glu Val Lys Asp Lys Arg Thr Leu Ile Gly Tyr Gly Arg Phe Phe Ile  
 340 345 350  
 Ser Asn Pro Asp Leu Val Asp Arg Leu Glu Lys Gly Leu Pro Leu Asn  
 355 360 365  
 Lys Tyr Asp Arg Asp Thr Phe Tyr Gln Met Ser Ala His Gly Tyr Ile  
 370 375 380  
 Asp Tyr Pro Thr Tyr Glu Glu Ala Leu Lys Leu Gly Trp Asp Lys Lys  
 385 390 395 400

&lt;210&gt; SEQ ID NO 21

&lt;211&gt; LENGTH: 337

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Thermoanaerobacter pseudethanolicus

&lt;400&gt; SEQUENCE: 21

Met Ser Ile Leu His Met Pro Leu Lys Ile Lys Asp Ile Thr Ile Lys  
 1 5 10 15  
 Asn Arg Ile Met Met Ser Pro Met Cys Met Tyr Ser Ala Ser Thr Asp  
 20 25 30  
 Gly Met Pro Asn Asp Trp His Ile Val His Tyr Ala Thr Arg Ala Ile  
 35 40 45  
 Gly Gly Val Gly Leu Ile Met Gln Glu Ala Thr Ala Val Glu Ser Arg  
 50 55 60  
 Gly Arg Ile Thr Asp His Asp Leu Gly Ile Trp Asn Asp Glu Gln Val

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65	70	75	80
Lys Glu Leu Lys Lys Ile Val Asp Ile Cys Lys Ala Asn Gly Ala Val 85 90 95			
Met Gly Ile Gln Leu Ala His Ala Gly Arg Lys Cys Asn Ile Ser Tyr 100 105 110			
Glu Asp Val Val Gly Pro Ser Pro Ile Lys Ala Gly Asp Arg Tyr Lys 115 120 125			
Leu Pro Arg Glu Leu Ser Val Glu Glu Ile Lys Ser Ile Val Lys Ala 130 135 140			
Phe Gly Glu Ala Ala Lys Arg Ala Asn Leu Ala Gly Tyr Asp Val Val 145 150 155 160			
Glu Ile His Ala Ala His Gly Tyr Leu Ile His Glu Phe Leu Ser Pro 165 170 175			
Leu Ser Asn Lys Arg Lys Asp Glu Tyr Gly Asn Ser Ile Glu Asn Arg 180 185 190			
Ala Arg Phe Leu Ile Glu Val Ile Asp Glu Val Arg Lys Asn Trp Pro 195 200 205			
Glu Asn Lys Pro Ile Phe Val Arg Val Ser Ala Asp Asp Tyr Met Glu 210 215 220			
Gly Gly Ile Asn Ile Asp Met Met Val Glu Tyr Ile Asn Met Ile Lys 225 230 235 240			
Asp Lys Val Asp Leu Ile Asp Val Ser Ser Gly Gly Leu Leu Asn Val 245 250 255			
Asp Ile Asn Leu Tyr Pro Gly Tyr Gln Val Lys Tyr Ala Glu Thr Ile 260 265 270			
Lys Lys Arg Cys Asn Ile Lys Thr Ser Ala Val Gly Leu Ile Thr Thr 275 280 285			
Gln Glu Leu Ala Glu Glu Ile Leu Ser Asn Glu Arg Ala Asp Leu Val 290 295 300			
Ala Leu Gly Arg Glu Leu Leu Arg Asn Pro Tyr Trp Val Leu His Thr 305 310 315 320			
Tyr Thr Ser Lys Glu Asp Trp Pro Lys Gln Tyr Glu Arg Ala Phe Lys 325 330 335			

Lys

&lt;210&gt; SEQ ID NO 22

&lt;211&gt; LENGTH: 365

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Enterobacter cloacae

&lt;400&gt; SEQUENCE: 22

Met Ser Ala Glu Lys Leu Phe Thr Pro Leu Lys Val Gly Ala Val Thr 1 5 10 15
Ala Pro Asn Arg Val Phe Met Ala Pro Leu Thr Arg Leu Arg Ser Ile 20 25 30
Glu Pro Gly Asp Ile Pro Thr Pro Leu Met Gly Glu Tyr Tyr Arg Gln 35 40 45
Arg Ala Ser Ala Gly Leu Ile Ile Ser Glu Ala Thr Gln Ile Ser Ala 50 55 60
Gln Ala Lys Gly Tyr Ala Gly Ala Pro Gly Leu His Ser Pro Glu Gln 65 70 75 80
Ile Ala Ala Trp Lys Lys Ile Thr Ala Gly Val His Ala Glu Asp Gly 85 90 95
Arg Ile Ala Val Gln Leu Trp His Thr Gly Arg Ile Ser His Ser Ser

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100				105				110							
Ile	Gln	Pro	Gly	Gly	Gln	Ala	Pro	Val	Ser	Ala	Ser	Ala	Leu	Asn	Ala
		115					120					125			
Asn	Thr	Arg	Thr	Ser	Leu	Arg	Asp	Glu	Asn	Gly	Asn	Ala	Ile	Arg	Val
	130					135					140				
Asp	Thr	Thr	Thr	Pro	Arg	Ala	Leu	Glu	Leu	Asp	Glu	Ile	Pro	Gly	Ile
	145				150					155				160	
Val	Asn	Asp	Phe	Arg	Gln	Ala	Val	Ala	Asn	Ala	Arg	Glu	Ala	Gly	Phe
			165						170					175	
Asp	Leu	Val	Glu	Leu	His	Ser	Ala	His	Gly	Tyr	Leu	Leu	His	Gln	Phe
		180							185				190		
Leu	Ser	Pro	Ser	Ser	Asn	Gln	Arg	Thr	Asp	Gln	Tyr	Gly	Gly	Ser	Val
		195					200					205			
Glu	Asn	Arg	Ala	Arg	Leu	Val	Leu	Glu	Val	Val	Asp	Ala	Val	Cys	Asn
	210					215					220				
Glu	Trp	Ser	Ala	Asp	Arg	Ile	Gly	Ile	Arg	Val	Ser	Pro	Ile	Gly	Thr
	225				230					235				240	
Phe	Gln	Asn	Val	Asp	Asn	Gly	Pro	Asn	Glu	Glu	Ala	Asp	Ala	Leu	Tyr
			245						250					255	
Leu	Ile	Glu	Glu	Leu	Ala	Lys	Arg	Gly	Ile	Ala	Tyr	Leu	His	Met	Ser
		260							265				270		
Glu	Thr	Asp	Leu	Ala	Gly	Gly	Lys	Pro	Tyr	Ser	Glu	Ala	Phe	Arg	Gln
		275					280					285			
Lys	Val	Arg	Glu	Arg	Phe	His	Gly	Val	Ile	Ile	Gly	Ala	Gly	Ala	Tyr
	290					295					300				
Thr	Ala	Glu	Lys	Ala	Glu	Asp	Leu	Ile	Gly	Lys	Gly	Leu	Ile	Asp	Ala
	305				310					315				320	
Val	Ala	Phe	Gly	Arg	Asp	Tyr	Ile	Ala	Asn	Pro	Asp	Leu	Val	Ala	Arg
			325						330					335	
Leu	Gln	Lys	Lys	Ala	Glu	Leu	Asn	Pro	Gln	Arg	Pro	Glu	Ser	Phe	Tyr
		340					345					350			
Gly	Gly	Gly	Ala	Glu	Gly	Tyr	Thr	Asp	Tyr	Pro	Ser	Leu			
		355					360					365			

&lt;210&gt; SEQ ID NO 23

&lt;211&gt; LENGTH: 128

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Fusobacterium nucleatum subsp. nucleatum

&lt;400&gt; SEQUENCE: 23

Met	Lys	Ser	Leu	Ile	Arg	Leu	Arg	Met	Ser	Ser	His	Asp	Ala	His	Tyr
	1				5				10					15	
Gly	Gly	Asn	Leu	Val	Asp	Gly	Ala	Arg	Met	Leu	Gln	Leu	Phe	Gly	Asp
		20							25				30		
Val	Ala	Thr	Glu	Leu	Leu	Ile	Gln	Leu	Asp	Gly	Asp	Glu	Gly	Leu	Phe
		35					40						45		
Lys	Ala	Tyr	Asp	Ser	Val	Glu	Phe	Met	Ala	Pro	Val	Phe	Ala	Gly	Asp
	50					55				60					
Tyr	Ile	Glu	Ala	Glu	Gly	Glu	Ile	Val	Asn	Val	Gly	Asn	Ser	Ser	Arg
	65				70				75					80	
Lys	Met	Val	Phe	Glu	Ala	Arg	Lys	Val	Ile	Val	Pro	Arg	Pro	Asp	Ile
			85						90					95	
Ser	Asp	Ser	Ala	Ala	Asp	Val	Leu	Ala	Glu	Pro	Ile	Val	Val	Cys	Arg
			100				105						110		

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Ala Thr Gly Thr Cys Val Thr Pro Lys Asp Lys Gln Arg Gly Lys Lys  
 115 120 125

<210> SEQ ID NO 24  
 <211> LENGTH: 260  
 <212> TYPE: PRT  
 <213> ORGANISM: Acidaminococcus fermentans

<400> SEQUENCE: 24

Met Ser Ile Tyr Thr Leu Gly Ile Asp Val Gly Ser Thr Ala Ser Lys  
 1 5 10 15  
 Cys Ile Ile Leu Lys Asp Gly Lys Glu Ile Val Ala Lys Ser Leu Val  
 20 25 30  
 Ala Val Gly Thr Gly Thr Ser Gly Pro Ala Arg Ser Ile Ser Glu Val  
 35 40 45  
 Leu Glu Asn Ala His Met Lys Lys Glu Asp Met Ala Phe Thr Leu Ala  
 50 55 60  
 Thr Gly Tyr Gly Arg Asn Ser Leu Glu Gly Ile Ala Asp Lys Gln Met  
 65 70 75 80  
 Ser Glu Leu Ser Cys His Ala Met Gly Ala Ser Phe Ile Trp Pro Asn  
 85 90 95  
 Val His Thr Val Ile Asp Ile Gly Gly Gln Asp Val Lys Val Ile His  
 100 105 110  
 Val Glu Asn Gly Thr Met Thr Asn Phe Gln Met Asn Asp Lys Cys Ala  
 115 120 125  
 Ala Gly Thr Gly Arg Phe Leu Asp Val Met Ala Asn Ile Leu Glu Val  
 130 135 140  
 Lys Val Ser Asp Leu Ala Glu Leu Gly Ala Lys Ser Thr Lys Arg Val  
 145 150 155 160  
 Ala Ile Ser Ser Thr Cys Thr Val Phe Ala Glu Ser Glu Val Ile Ser  
 165 170 175  
 Gln Leu Ser Lys Gly Thr Asp Lys Ile Asp Ile Ile Ala Gly Ile His  
 180 185 190  
 Arg Ser Val Ala Ser Arg Val Ile Gly Leu Ala Asn Arg Val Gly Ile  
 195 200 205  
 Val Lys Asp Val Val Met Thr Gly Gly Val Ala Gln Asn Tyr Gly Val  
 210 215 220  
 Arg Gly Ala Leu Glu Glu Gly Leu Gly Val Glu Ile Lys Thr Ser Pro  
 225 230 235 240  
 Leu Ala Gln Tyr Asn Gly Ala Leu Gly Ala Ala Leu Tyr Ala Tyr Lys  
 245 250 255  
 Lys Ala Ala Lys  
 260

<210> SEQ ID NO 25  
 <211> LENGTH: 650  
 <212> TYPE: PRT  
 <213> ORGANISM: Clostridium symbiosum

<400> SEQUENCE: 25

Met Ser Ile Asn Ala Leu Leu Asp Glu Phe Lys Val Lys Ala Ala Thr  
 1 5 10 15  
 Pro Lys Gln Gln Leu Ala Glu Tyr Lys Ala Gln Gly Lys Lys Val Ile  
 20 25 30  
 Gly Val Leu Pro Tyr Tyr Ala Pro Glu Glu Leu Val Tyr Ala Ala Gly  
 35 40 45

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Met	Val	Pro	Met	Gly	Ile	Trp	Gly	Ser	Asn	Asn	Lys	Thr	Ile	Ser	Arg
50						55					60				
Ala	Lys	Glu	Tyr	Cys	Ala	Thr	Phe	Tyr	Cys	Thr	Ile	Ala	Gln	Leu	Ala
65					70					75					80
Leu	Glu	Met	Leu	Leu	Asp	Gly	Thr	Met	Asp	Gln	Leu	Asp	Gly	Ile	Ile
				85					90					95	
Thr	Pro	Thr	Ile	Cys	Asp	Thr	Leu	Arg	Pro	Met	Ser	Gln	Asn	Phe	Arg
			100					105					110		
Val	Ala	Met	Gly	Asp	Lys	Met	Ala	Val	Ile	Phe	Leu	Ala	Gln	Pro	Gln
		115					120					125			
Asn	Arg	Phe	Glu	Asp	Phe	Gly	Leu	Gln	Phe	Ser	Val	Asp	Gln	Tyr	Thr
130						135					140				
Asn	Val	Lys	Lys	Glu	Leu	Glu	Lys	Val	Ala	Gly	Lys	Glu	Ile	Thr	Asn
145					150					155					160
Glu	Ala	Ile	Gln	Asp	Ala	Ile	Lys	Val	Tyr	Asn	Lys	Ser	Arg	Ala	Ala
				165					170					175	
Arg	Arg	Lys	Phe	Val	Glu	Leu	Ala	Ser	Ala	His	Cys	Asp	Val	Ile	Thr
			180					185					190		
Pro	Thr	Lys	Arg	Ser	Ala	Val	Leu	Lys	Ser	Phe	Phe	Phe	Met	Glu	Lys
		195					200						205		
Pro	Glu	Tyr	Ile	Glu	Lys	Leu	Glu	Glu	Leu	Asn	Ala	Glu	Leu	Glu	Lys
	210					215					220				
Leu	Pro	Val	Cys	Asp	Trp	Gln	Gly	Thr	Lys	Val	Val	Thr	Ser	Gly	Ile
225						230					235				240
Ile	Cys	Asp	Asn	Pro	Lys	Leu	Leu	Glu	Ile	Phe	Glu	Glu	Asn	Asn	Ile
				245					250					255	
Ala	Ile	Ala	Ala	Asp	Asp	Val	Gly	His	Glu	Ser	Arg	Ser	Phe	Arg	Val
			260					265					270		
Asp	Ala	Pro	Glu	Asp	Glu	Ala	Asp	Ala	Leu	Met	Ala	Leu	Ala	Lys	Gln
		275					280					285			
Phe	Ala	Asn	Met	Asp	Tyr	Asp	Val	Leu	Leu	Tyr	Asp	Pro	Lys	Ser	Thr
	290					295					300				
Glu	Asn	Arg	Arg	Gly	Glu	Phe	Ile	Ala	Asn	Met	Val	Lys	Glu	Ser	Gly
305					310					315					320
Ala	Gln	Gly	Leu	Val	Leu	Phe	Met	Gln	Gln	Phe	Cys	Asp	Pro	Glu	Glu
				325					330					335	
Met	Glu	Tyr	Pro	Tyr	Leu	Lys	Lys	Ala	Leu	Asn	Asn	Ala	Gly	Ile	Pro
			340					345						350	
His	Ile	Lys	Leu	Gly	Ile	Asp	Gln	Gln	Met	Arg	Asp	Phe	Gly	Gln	Ala
		355					360					365			
Ser	Thr	Ala	Ile	Gln	Ala	Phe	Ala	Asp	Val	Leu	Glu	Met	Gln	Lys	Met
	370					375					380				
Ser	Gly	Ile	Tyr	Thr	Leu	Gly	Ile	Asp	Val	Gly	Ser	Thr	Ala	Ser	Lys
385					390					395					400
Cys	Ile	Val	Leu	Lys	Asp	Gly	Lys	Glu	Ile	Val	Ala	Lys	Ser	Leu	Ile
				405					410					415	
Asp	Val	Gly	Ala	Gly	Thr	Ser	Gly	Pro	Gln	Arg	Ala	Ile	Glu	Ala	Val
			420					425					430		
Leu	Asn	Glu	Ala	Gly	Met	Lys	Lys	Glu	Asp	Met	Ala	Tyr	Thr	Leu	Ala
		435					440					445			
Thr	Gly	Tyr	Gly	Arg	Thr	Ser	Leu	Met	Asp	Gly	Ile	Ala	Asp	Lys	Gln
	450					455					460				
Met	Ser	Glu	Leu	Ser	Cys	His	Ala	Lys	Gly	Ala	Thr	Phe	Leu	Phe	Pro

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465		470		475		480
Asn Val His Thr	Val Ile Asp Ile Gly Gly Gln Asp Val Lys Val Leu	485		490		495
His Ile Asp Asn Gly Ala Met Thr Asn Phe Gln Met Asn Asp Lys Cys	500		505		510	
Ala Ala Gly Thr Gly Arg Phe Leu Asp Val Met Ala Arg Val Leu Glu	515		520		525	
Val Lys Val Glu Asp Leu Gly Arg Leu Gly Ala Met Ser Arg Lys Lys	530		535		540	
Val Gly Ile Ser Ser Thr Cys Thr Val Phe Ala Glu Ser Glu Val Ile	545		550		555	560
Ser Gln Leu Ala Met Gly Thr Asp Lys Cys Asp Ile Ile Asp Gly Ile	565		570		575	
His Arg Ser Val Ala His Arg Val Thr Gly Leu Ala His Arg Ile Gly	580		585		590	
Val Val Pro Asp Val Val Met Thr Gly Gly Val Ala Gln Asn Glu Gly	595		600		605	
Val Val Lys Ala Leu Gln Asp Glu Leu Gly Cys Pro Ile Asn Thr Ser	610		615		620	
Pro Leu Thr Gln Tyr Asn Gly Ala Leu Gly Ala Ala Leu Leu Ala Trp	625		630		635	640
Gln Ala Ala Ser Arg Arg Gln Ser Asn Ser	645		650			

&lt;210&gt; SEQ ID NO 26

&lt;211&gt; LENGTH: 471

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Bacillus subtilis

&lt;400&gt; SEQUENCE: 26

Met Lys Asn Lys Trp Tyr Lys Pro Lys Arg His Trp Lys Glu Ile Glu	1	5	10	15
Leu Trp Lys Asp Val Pro Glu Glu Lys Trp Asn Asp Trp Leu Trp Gln	20	25	30	
Leu Thr His Thr Val Arg Thr Leu Asp Asp Leu Lys Lys Val Ile Asn	35	40	45	
Leu Thr Glu Asp Glu Glu Glu Gly Val Arg Ile Ser Thr Lys Thr Ile	50	55	60	
Pro Leu Asn Ile Thr Pro Tyr Tyr Ala Ser Leu Met Asp Pro Asp Asn	65	70	75	80
Pro Arg Cys Pro Val Arg Met Gln Ser Val Pro Leu Ser Glu Glu Met	85	90	95	
His Lys Thr Lys Tyr Asp Leu Glu Asp Pro Leu His Glu Asp Glu Asp	100	105	110	
Ser Pro Val Pro Gly Leu Thr His Arg Tyr Pro Asp Arg Val Leu Phe	115	120	125	
Leu Val Thr Asn Gln Cys Ser Met Tyr Cys Arg Tyr Cys Thr Arg Arg	130	135	140	
Arg Phe Ser Gly Gln Ile Gly Met Gly Val Pro Lys Lys Gln Leu Asp	145	150	155	160
Ala Ala Ile Ala Tyr Ile Arg Glu Thr Pro Glu Ile Arg Asp Cys Leu	165	170	175	
Ile Ser Gly Gly Asp Gly Leu Leu Ile Asn Asp Gln Ile Leu Glu Tyr	180	185	190	

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Ile Leu Lys Glu Leu Arg Ser Ile Pro His Leu Glu Val Ile Arg Ile  
 195 200 205  
 Gly Thr Arg Ala Pro Val Val Phe Pro Gln Arg Ile Thr Asp His Leu  
 210 215 220  
 Cys Glu Ile Leu Lys Lys Tyr His Pro Val Trp Leu Asn Thr His Phe  
 225 230 235 240  
 Asn Thr Ser Ile Glu Met Thr Glu Glu Ser Val Glu Ala Cys Glu Lys  
 245 250 255  
 Leu Val Asn Ala Gly Val Pro Val Gly Asn Gln Ala Val Val Leu Ala  
 260 265 270  
 Gly Ile Asn Asp Ser Val Pro Ile Met Lys Lys Leu Met His Asp Leu  
 275 280 285  
 Val Lys Ile Arg Val Arg Pro Tyr Tyr Ile Tyr Gln Cys Asp Leu Ser  
 290 295 300  
 Glu Gly Ile Gly His Phe Arg Ala Pro Val Ser Lys Gly Leu Glu Ile  
 305 310 315 320  
 Ile Glu Gly Leu Arg Gly His Thr Ser Gly Tyr Ala Val Pro Thr Phe  
 325 330 335  
 Val Val Asp Ala Pro Gly Gly Gly Gly Lys Ile Ala Leu Gln Pro Asn  
 340 345 350  
 Tyr Val Leu Ser Gln Ser Pro Asp Lys Val Ile Leu Arg Asn Phe Glu  
 355 360 365  
 Gly Val Ile Thr Ser Tyr Pro Glu Pro Glu Asn Tyr Ile Pro Asn Gln  
 370 375 380  
 Ala Asp Ala Tyr Phe Glu Ser Val Phe Pro Glu Thr Ala Asp Lys Lys  
 385 390 395 400  
 Glu Pro Ile Gly Leu Ser Ala Ile Phe Ala Asp Lys Glu Val Ser Phe  
 405 410 415  
 Thr Pro Glu Asn Val Asp Arg Ile Lys Arg Arg Glu Ala Tyr Ile Ala  
 420 425 430  
 Asn Pro Glu His Glu Thr Leu Lys Asp Arg Arg Glu Lys Arg Asp Gln  
 435 440 445  
 Leu Lys Glu Lys Lys Phe Leu Ala Gln Gln Lys Lys Gln Lys Glu Thr  
 450 455 460  
 Glu Cys Gly Gly Asp Ser Ser  
 465 470

<210> SEQ ID NO 27  
 <211> LENGTH: 266  
 <212> TYPE: PRT  
 <213> ORGANISM: Peptoclostridium difficile

<400> SEQUENCE: 27

Met Tyr Thr Met Gly Leu Asp Ile Gly Ser Thr Ala Ser Lys Gly Val  
 1 5 10 15  
 Ile Leu Lys Asn Gly Glu Asp Ile Val Ala Ser Glu Thr Ile Ser Ser  
 20 25 30  
 Gly Thr Gly Thr Thr Gly Pro Ser Arg Val Leu Glu Lys Leu Tyr Gly  
 35 40 45  
 Lys Thr Gly Leu Ala Arg Glu Asp Ile Lys Lys Val Val Val Thr Gly  
 50 55 60  
 Tyr Gly Arg Met Asn Tyr Ser Asp Ala Asp Lys Gln Ile Ser Glu Leu  
 65 70 75 80  
 Ser Cys His Ala Arg Gly Val Asn Phe Ile Ile Pro Glu Thr Arg Thr  
 85 90 95

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Ile Ile Asp Ile Gly Gly Gln Asp Ala Lys Val Leu Lys Leu Asp Asn  
 100 105 110  
 Asn Gly Arg Leu Leu Asn Phe Leu Met Asn Asp Lys Cys Ala Ala Gly  
 115 120 125  
 Thr Gly Arg Phe Leu Asp Val Met Ala Lys Ile Ile Glu Val Asp Val  
 130 135 140  
 Ser Glu Leu Gly Ser Ile Ser Met Asn Ser Gln Asn Glu Val Ser Ile  
 145 150 155 160  
 Ser Ser Thr Cys Thr Val Phe Ala Glu Ser Glu Val Ile Ser His Leu  
 165 170 175  
 Ser Glu Asn Ala Lys Ile Glu Asp Ile Val Ala Gly Ile His Thr Ser  
 180 185 190  
 Val Ala Lys Arg Val Ser Ser Leu Val Lys Arg Ile Gly Val Gln Arg  
 195 200 205  
 Asn Val Val Met Val Gly Gly Val Ala Arg Asn Ser Gly Ile Val Arg  
 210 215 220  
 Ala Met Ala Arg Glu Ile Asn Thr Glu Ile Ile Val Pro Asp Ile Pro  
 225 230 235 240  
 Gln Leu Thr Gly Ala Leu Gly Ala Ala Leu Tyr Ala Phe Asp Glu Ala  
 245 250 255  
 Lys Glu Ser Gln Lys Glu Val Lys Asn Ile  
 260 265

&lt;210&gt; SEQ ID NO 28

&lt;211&gt; LENGTH: 783

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Peptoclostridium difficile

&lt;400&gt; SEQUENCE: 28

Met Ser Glu Lys Lys Glu Ala Arg Val Val Ile Asn Asp Leu Leu Ala  
 1 5 10 15  
 Glu Gln Tyr Ala Asn Ala Phe Lys Ala Lys Glu Glu Gly Arg Pro Val  
 20 25 30  
 Gly Trp Ser Thr Ser Val Phe Pro Gln Glu Leu Ala Glu Val Phe Asp  
 35 40 45  
 Leu Asn Val Leu Tyr Pro Glu Asn Gln Ala Ala Gly Val Ala Ala Lys  
 50 55 60  
 Lys Gly Ser Leu Glu Leu Cys Glu Ile Ala Glu Ser Lys Gly Tyr Ser  
 65 70 75 80  
 Ile Asp Leu Cys Ala Tyr Ala Arg Thr Asn Phe Gly Leu Leu Glu Asn  
 85 90 95  
 Gly Gly Cys Glu Ala Leu Asp Met Pro Ala Pro Asp Phe Leu Leu Cys  
 100 105 110  
 Cys Asn Asn Ile Cys Asn Gln Val Ile Lys Trp Tyr Glu Asn Ile Ser  
 115 120 125  
 Arg Glu Leu Asp Ile Pro Leu Ile Met Ile Asp Thr Thr Phe Asn Asn  
 130 135 140  
 Glu Asp Glu Val Thr Gln Ser Arg Ile Asp Tyr Ile Lys Ala Gln Phe  
 145 150 155 160  
 Glu Glu Ala Ile Lys Gln Leu Glu Ile Ile Ser Gly Lys Lys Phe Asp  
 165 170 175  
 Pro Lys Lys Phe Glu Glu Val Met Lys Ile Ser Ala Glu Asn Gly Arg  
 180 185 190  
 Leu Trp Lys Tyr Ser Met Ser Leu Pro Ala Asp Ser Ser Pro Ser Pro



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195					200					205					
Met	Asn	Gly	Phe	Asp	Leu	Phe	Thr	Tyr	Met	Ala	Val	Ile	Val	Cys	Ala
	210					215					220				
Arg	Gly	Lys	Lys	Glu	Thr	Thr	Glu	Ala	Phe	Lys	Leu	Leu	Ile	Glu	Glu
	225					230					235				240
Leu	Glu	Asp	Asn	Met	Lys	Thr	Gly	Lys	Ser	Ser	Phe	Arg	Gly	Glu	Glu
				245					250					255	
Lys	Tyr	Arg	Ile	Met	Met	Glu	Gly	Ile	Pro	Cys	Trp	Pro	Tyr	Ile	Gly
			260					265					270		
Tyr	Lys	Met	Lys	Thr	Leu	Ala	Lys	Phe	Gly	Val	Asn	Met	Thr	Gly	Ser
		275					280					285			
Val	Tyr	Pro	His	Ala	Trp	Ala	Leu	Gln	Tyr	Glu	Val	Asn	Asp	Leu	Asp
	290					295					300				
Gly	Met	Ala	Val	Ala	Tyr	Ser	Thr	Met	Phe	Asn	Asn	Val	Asn	Leu	Asp
	305					310					315				320
Arg	Met	Thr	Lys	Tyr	Arg	Val	Asp	Ser	Leu	Val	Glu	Gly	Lys	Cys	Asp
				325					330					335	
Gly	Ala	Phe	Tyr	His	Met	Asn	Arg	Ser	Cys	Lys	Leu	Met	Ser	Leu	Ile
			340					345					350		
Gln	Tyr	Glu	Met	Gln	Arg	Arg	Ala	Ala	Glu	Glu	Thr	Gly	Leu	Pro	Tyr
		355					360					365			
Ala	Gly	Phe	Asp	Gly	Asp	Gln	Ala	Asp	Pro	Arg	Ala	Phe	Thr	Asn	Ala
	370					375					380				
Gln	Phe	Glu	Thr	Arg	Ile	Gln	Gly	Leu	Val	Glu	Val	Met	Glu	Glu	Arg
	385					390					395				400
Lys	Lys	Leu	Asn	Arg	Gly	Glu	Ile	Met	Glu	Ala	Ile	Leu	Ser	Lys	Met
			405						410					415	
Lys	Glu	Val	Val	Glu	Asn	Pro	Asn	Ala	Ala	Val	Lys	Lys	Tyr	Lys	Ser
			420					425					430		
Glu	Thr	Gly	Lys	Lys	Ala	Ile	Gly	Cys	Phe	Pro	Val	Tyr	Cys	Pro	Glu
		435					440					445			
Glu	Ile	Ile	His	Ala	Ala	Gly	Met	Leu	Pro	Val	Gly	Ile	Trp	Gly	Gly
	450					455					460				
Gln	Thr	Glu	Leu	Asp	Leu	Ala	Lys	Gln	Tyr	Phe	Pro	Ala	Phe	Ala	Cys
	465					470					475				480
Ser	Ile	Met	Gln	Ser	Cys	Leu	Glu	Tyr	Gly	Leu	Lys	Gly	Ala	Tyr	Asp
			485						490					495	
Glu	Leu	Ser	Gly	Val	Ile	Ile	Pro	Gly	Met	Cys	Asp	Thr	Leu	Ile	Cys
			500					505					510		
Leu	Gly	Gln	Asn	Trp	Lys	Ser	Ala	Val	Pro	His	Ile	Lys	Tyr	Ile	Ser
		515					520					525			
Leu	Val	His	Pro	Gln	Asn	Arg	Lys	Leu	Glu	Ala	Gly	Val	Lys	Tyr	Leu
	530					535					540				
Ile	Ser	Glu	Tyr	Lys	Gly	Val	Lys	Arg	Glu	Leu	Glu	Glu	Ile	Cys	Gly
	545					550					555				560
Tyr	Glu	Ile	Glu	Glu	Ala	Lys	Ile	His	Glu	Ser	Ile	Glu	Val	Tyr	Asn
			565						570					575	
Glu	His	Arg	Lys	Thr	Met	Arg	Asp	Phe	Val	Glu	Val	Ala	Tyr	Lys	His
			580					585					590		
Ser	Asn	Thr	Ile	Lys	Pro	Ser	Ile	Arg	Ser	Leu	Val	Ile	Lys	Ser	Gly
		595					600					605			
Phe	Phe	Met	Arg	Lys	Glu	Glu	His	Thr	Glu	Leu	Val	Lys	Asp	Leu	Ile
	610					615					620				

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Ala Lys Leu Asn Ala Met Pro Glu Glu Val Cys Ser Gly Lys Lys Val  
625 630 635 640

Leu Leu Thr Gly Ile Leu Ala Asp Ser Lys Asp Ile Leu Asp Ile Leu  
645 650 655

Glu Asp Asn Asn Ile Ser Val Val Ala Asp Asp Leu Ala Gln Glu Thr  
660 665 670

Arg Gln Phe Arg Thr Asp Val Pro Ala Gly Asp Asp Ala Leu Glu Arg  
675 680 685

Leu Ala Arg Gln Trp Ser Asn Ile Glu Gly Cys Ser Leu Ala Tyr Asp  
690 695 700

Pro Lys Lys Lys Arg Gly Ser Leu Ile Val Asp Glu Val Lys Lys Lys  
705 710 715 720

Asp Ile Asp Gly Val Ile Phe Cys Met Met Lys Phe Cys Asp Pro Glu  
725 730 735

Glu Tyr Asp Tyr Pro Leu Val Arg Lys Asp Ile Glu Asp Ser Gly Ile  
740 745 750

Pro Thr Leu Tyr Val Glu Ile Asp Gln Gln Thr Gln Asn Asn Glu Gln  
755 760 765

Ala Arg Thr Arg Ile Gln Thr Phe Ala Glu Met Met Ser Leu Ala  
770 775 780

<210> SEQ ID NO 29  
<211> LENGTH: 466  
<212> TYPE: PRT  
<213> ORGANISM: Escherichia coli

<400> SEQUENCE: 29

Met Asp Gln Lys Leu Leu Thr Asp Phe Arg Ser Glu Leu Leu Asp Ser  
1 5 10 15

Arg Phe Gly Ala Lys Ala Ile Ser Thr Ile Ala Glu Ser Lys Arg Phe  
20 25 30

Pro Leu His Glu Met Arg Asp Asp Val Ala Phe Gln Ile Ile Asn Asp  
35 40 45

Glu Leu Tyr Leu Asp Gly Asn Ala Arg Gln Asn Leu Ala Thr Phe Cys  
50 55 60

Gln Thr Trp Asp Asp Glu Asn Val His Lys Leu Met Asp Leu Ser Ile  
65 70 75 80

Asn Lys Asn Trp Ile Asp Lys Glu Glu Tyr Pro Gln Ser Ala Ala Ile  
85 90 95

Asp Leu Arg Cys Val Asn Met Val Ala Asp Leu Trp His Ala Pro Ala  
100 105 110

Pro Lys Asn Gly Gln Ala Val Gly Thr Asn Thr Ile Gly Ser Ser Glu  
115 120 125

Ala Cys Met Leu Gly Gly Met Ala Met Lys Trp Arg Trp Arg Lys Arg  
130 135 140

Met Glu Ala Ala Gly Lys Pro Thr Asp Lys Pro Asn Leu Val Cys Gly  
145 150 155 160

Pro Val Gln Ile Cys Trp His Lys Phe Ala Arg Tyr Trp Asp Val Glu  
165 170 175

Leu Arg Glu Ile Pro Met Arg Pro Gly Gln Leu Phe Met Asp Pro Lys  
180 185 190

Arg Met Ile Glu Ala Cys Asp Glu Asn Thr Ile Gly Val Val Pro Thr  
195 200 205

Phe Gly Val Thr Tyr Thr Gly Asn Tyr Glu Phe Pro Gln Pro Leu His

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210	215	220
Asp Ala Leu Asp Lys Phe Gln Ala Asp Thr Gly Ile Asp Ile Asp Met 225 230 235 240		
His Ile Asp Ala Ala Ser Gly Gly Phe Leu Ala Pro Phe Val Ala Pro 245 250 255		
Asp Ile Val Trp Asp Phe Arg Leu Pro Arg Val Lys Ser Ile Ser Ala 260 265 270		
Ser Gly His Lys Phe Gly Leu Ala Pro Leu Gly Cys Gly Trp Val Ile 275 280 285		
Trp Arg Asp Glu Glu Ala Leu Pro Gln Glu Leu Val Phe Asn Val Asp 290 295 300		
Tyr Leu Gly Gly Gln Ile Gly Thr Phe Ala Ile Asn Phe Ser Arg Pro 305 310 315 320		
Ala Gly Gln Val Ile Ala Gln Tyr Tyr Glu Phe Leu Arg Leu Gly Arg 325 330 335		
Glu Gly Tyr Thr Lys Val Gln Asn Ala Ser Tyr Gln Val Ala Ala Tyr 340 345 350		
Leu Ala Asp Glu Ile Ala Lys Leu Gly Pro Tyr Glu Phe Ile Cys Thr 355 360 365		
Gly Arg Pro Asp Glu Gly Ile Pro Ala Val Cys Phe Lys Leu Lys Asp 370 375 380		
Gly Glu Asp Pro Gly Tyr Thr Leu Tyr Asp Leu Ser Glu Arg Leu Arg 385 390 395 400		
Leu Arg Gly Trp Gln Val Pro Ala Phe Thr Leu Gly Gly Glu Ala Thr 405 410 415		
Asp Ile Val Val Met Arg Ile Met Cys Arg Arg Gly Phe Glu Met Asp 420 425 430		
Phe Ala Glu Leu Leu Leu Glu Asp Tyr Lys Ala Ser Leu Lys Tyr Leu 435 440 445		
Ser Asp His Pro Lys Leu Gln Gly Ile Ala Gln Gln Asn Ser Phe Lys 450 455 460		
His Thr 465		

<210> SEQ ID NO 30  
 <211> LENGTH: 715  
 <212> TYPE: PRT  
 <213> ORGANISM: Escherichia coli

<400> SEQUENCE: 30

Met Asn Val Ile Ala Ile Leu Asn His Met Gly Val Tyr Phe Lys Glu 1 5 10 15
Glu Pro Ile Arg Glu Leu His Arg Ala Leu Glu Arg Leu Asn Phe Gln 20 25 30
Ile Val Tyr Pro Asn Asp Arg Asp Asp Leu Leu Lys Leu Ile Glu Asn 35 40 45
Asn Ala Arg Leu Cys Gly Val Ile Phe Asp Trp Asp Lys Tyr Asn Leu 50 55 60
Glu Leu Cys Glu Glu Ile Ser Lys Met Asn Glu Asn Leu Pro Leu Tyr 65 70 75 80
Ala Phe Ala Asn Thr Tyr Ser Thr Leu Asp Val Ser Leu Asn Asp Leu 85 90 95
Arg Leu Gln Ile Ser Phe Phe Glu Tyr Ala Leu Gly Ala Ala Glu Asp 100 105 110

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Ile Ala Asn Lys Ile Lys Gln Thr Thr Asp Glu Tyr Ile Asn Thr Ile  
 115 120 125  
 Leu Pro Pro Leu Thr Lys Ala Leu Phe Lys Tyr Val Arg Glu Gly Lys  
 130 135 140  
 Tyr Thr Phe Cys Thr Pro Gly His Met Gly Gly Thr Ala Phe Gln Lys  
 145 150 155 160  
 Ser Pro Val Gly Ser Leu Phe Tyr Asp Phe Phe Gly Pro Asn Thr Met  
 165 170 175  
 Lys Ser Asp Ile Ser Ile Ser Val Ser Glu Leu Gly Ser Leu Leu Asp  
 180 185 190  
 His Ser Gly Pro His Lys Glu Ala Glu Gln Tyr Ile Ala Arg Val Phe  
 195 200 205  
 Asn Ala Asp Arg Ser Tyr Met Val Thr Asn Gly Thr Ser Thr Ala Asn  
 210 215 220  
 Lys Ile Val Gly Met Tyr Ser Ala Pro Ala Gly Ser Thr Ile Leu Ile  
 225 230 235 240  
 Asp Arg Asn Cys His Lys Ser Leu Thr His Leu Met Met Met Ser Asp  
 245 250 255  
 Val Thr Pro Ile Tyr Phe Arg Pro Thr Arg Asn Ala Tyr Gly Ile Leu  
 260 265 270  
 Gly Gly Ile Pro Gln Ser Glu Phe Gln His Ala Thr Ile Ala Lys Arg  
 275 280 285  
 Val Lys Glu Thr Pro Asn Ala Thr Trp Pro Val His Ala Val Ile Thr  
 290 295 300  
 Asn Ser Thr Tyr Asp Gly Leu Leu Tyr Asn Thr Asp Phe Ile Lys Lys  
 305 310 315 320  
 Thr Leu Asp Val Lys Ser Ile His Phe Asp Ser Ala Trp Val Pro Tyr  
 325 330 335  
 Thr Asn Phe Ser Pro Ile Tyr Glu Gly Lys Cys Gly Met Ser Gly Gly  
 340 345 350  
 Arg Val Glu Gly Lys Val Ile Tyr Glu Thr Gln Ser Thr His Lys Leu  
 355 360 365  
 Leu Ala Ala Phe Ser Gln Ala Ser Met Ile His Val Lys Gly Asp Val  
 370 375 380  
 Asn Glu Glu Thr Phe Asn Glu Ala Tyr Met Met His Thr Thr Thr Ser  
 385 390 395 400  
 Pro His Tyr Gly Ile Val Ala Ser Thr Glu Thr Ala Ala Ala Met Met  
 405 410 415  
 Lys Gly Asn Ala Gly Lys Arg Leu Ile Asn Gly Ser Ile Glu Arg Ala  
 420 425 430  
 Ile Lys Phe Arg Lys Glu Ile Lys Arg Leu Arg Thr Glu Ser Asp Gly  
 435 440 445  
 Trp Phe Phe Asp Val Trp Gln Pro Asp His Ile Asp Thr Thr Glu Cys  
 450 455 460  
 Trp Pro Leu Arg Ser Asp Ser Thr Trp His Gly Phe Lys Asn Ile Asp  
 465 470 475 480  
 Asn Glu His Met Tyr Leu Asp Pro Ile Lys Val Thr Leu Leu Thr Pro  
 485 490 495  
 Gly Met Glu Lys Asp Gly Thr Met Ser Asp Phe Gly Ile Pro Ala Ser  
 500 505 510  
 Ile Val Ala Lys Tyr Leu Asp Glu His Gly Ile Val Val Glu Lys Thr  
 515 520 525  
 Gly Pro Tyr Asn Leu Leu Phe Leu Phe Ser Ile Gly Ile Asp Lys Thr

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530	535	540
Lys Ala Leu Ser Leu Leu Arg Ala Leu Thr Asp Phe Lys Arg Ala Phe 545	550	555 560
Asp Leu Asn Leu Arg Val Lys Asn Met Leu Pro Ser Leu Tyr Arg Glu 565	570	575
Asp Pro Glu Phe Tyr Glu Asn Met Arg Ile Gln Glu Leu Ala Gln Asn 580	585	590
Ile His Lys Leu Ile Val His His Asn Leu Pro Asp Leu Met Tyr Arg 595	600	605
Ala Phe Glu Val Leu Pro Thr Met Val Met Thr Pro Tyr Ala Ala Phe 610	615	620
Gln Lys Glu Leu His Gly Met Thr Glu Glu Val Tyr Leu Asp Glu Met 625	630	635 640
Val Gly Arg Ile Asn Ala Asn Met Ile Leu Pro Tyr Pro Pro Gly Val 645	650	655
Pro Leu Val Met Pro Gly Glu Met Ile Thr Glu Glu Ser Arg Pro Val 660	665	670
Leu Glu Phe Leu Gln Met Leu Cys Glu Ile Gly Ala His Tyr Pro Gly 675	680	685
Phe Glu Thr Asp Ile His Gly Ala Tyr Arg Gln Ala Asp Gly Arg Tyr 690	695	700
Thr Val Lys Val Leu Lys Glu Glu Ser Lys Lys 705	710	715
<210> SEQ ID NO 31		
<211> LENGTH: 732		
<212> TYPE: PRT		
<213> ORGANISM: Escherichia coli		
<400> SEQUENCE: 31		
Met Ser Lys Leu Lys Ile Ala Val Ser Asp Ser Cys Pro Asp Cys Phe 1	5	10 15
Thr Thr Gln Arg Glu Cys Ile Tyr Ile Asn Glu Ser Arg Asn Ile Asp 20	25	30
Val Ala Ala Ile Val Leu Ser Leu Asn Asp Val Thr Cys Gly Lys Leu 35	40	45
Asp Glu Ile Asp Ala Thr Gly Tyr Gly Ile Pro Val Phe Ile Ala Thr 50	55	60
Glu Asn Gln Glu Arg Val Pro Ala Glu Tyr Leu Pro Arg Ile Ser Gly 65	70	75 80
Val Phe Glu Asn Cys Glu Ser Arg Arg Glu Phe Tyr Gly Arg Gln Leu 85	90	95
Glu Thr Ala Ala Ser His Tyr Glu Thr Gln Leu Arg Pro Pro Phe Phe 100	105	110
Arg Ala Leu Val Asp Tyr Val Asn Gln Gly Asn Ser Ala Phe Asp Cys 115	120	125
Pro Gly His Gln Gly Gly Glu Phe Phe Arg Arg His Pro Ala Gly Asn 130	135	140
Gln Phe Val Glu Tyr Phe Gly Glu Ala Leu Phe Arg Ala Asp Leu Cys 145	150	155 160
Asn Ala Asp Val Ala Met Gly Asp Leu Leu Ile His Glu Gly Ala Pro 165	170	175
Cys Ile Ala Gln Gln His Ala Ala Lys Val Phe Asn Ala Asp Lys Thr 180	185	190

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Tyr Phe Val Leu Asn Gly Thr Ser Ser Ser Asn Lys Val Val Leu Asn  
 195 200 205  
 Ala Leu Leu Thr Pro Gly Asp Leu Val Leu Phe Asp Arg Asn Asn His  
 210 215 220  
 Lys Ser Asn His His Gly Ala Leu Leu Gln Ala Gly Ala Thr Pro Val  
 225 230 235 240  
 Tyr Leu Glu Thr Ala Arg Asn Pro Tyr Gly Phe Ile Gly Gly Ile Asp  
 245 250 255  
 Ala His Cys Phe Glu Glu Ser Tyr Leu Arg Glu Leu Ile Ala Glu Val  
 260 265 270  
 Ala Pro Gln Arg Ala Lys Glu Ala Arg Pro Phe Arg Leu Ala Val Ile  
 275 280 285  
 Gln Leu Gly Thr Tyr Asp Gly Thr Ile Tyr Asn Ala Arg Gln Val Val  
 290 295 300  
 Asp Lys Ile Gly His Leu Cys Asp Tyr Ile Leu Phe Asp Ser Ala Trp  
 305 310 315 320  
 Val Gly Tyr Glu Gln Phe Ile Pro Met Met Ala Asp Cys Ser Pro Leu  
 325 330 335  
 Leu Leu Asp Leu Asn Glu Asn Asp Pro Gly Ile Leu Val Thr Gln Ser  
 340 345 350  
 Val His Lys Gln Gln Ala Gly Phe Ser Gln Thr Ser Gln Ile His Lys  
 355 360 365  
 Lys Asp Ser His Ile Lys Gly Gln Gln Arg Tyr Val Pro His Lys Arg  
 370 375 380  
 Met Asn Asn Ala Phe Met Met His Ala Ser Thr Ser Pro Phe Tyr Pro  
 385 390 395 400  
 Leu Phe Ala Ala Leu Asn Ile Asn Ala Lys Met His Glu Gly Val Ser  
 405 410 415  
 Gly Arg Asn Met Trp Met Asp Cys Val Val Asn Gly Ile Asn Ala Arg  
 420 425 430  
 Lys Leu Ile Leu Asp Asn Cys Gln His Ile Arg Pro Phe Val Pro Glu  
 435 440 445  
 Leu Val Asp Gly Lys Pro Trp Gln Ser Tyr Glu Thr Ala Gln Ile Ala  
 450 455 460  
 Val Asp Leu Arg Phe Phe Gln Phe Val Pro Gly Glu His Trp His Ser  
 465 470 475 480  
 Phe Glu Gly Tyr Ala Glu Asn Gln Tyr Phe Val Asp Pro Cys Lys Leu  
 485 490 495  
 Leu Leu Thr Thr Pro Gly Ile Asp Ala Arg Asn Gly Glu Tyr Glu Ala  
 500 505 510  
 Phe Gly Val Pro Ala Thr Ile Leu Ala Asn Phe Leu Arg Glu Asn Gly  
 515 520 525  
 Val Val Pro Glu Lys Cys Asp Leu Asn Ser Ile Leu Phe Leu Leu Thr  
 530 535 540  
 Pro Ala Glu Asp Met Ala Lys Leu Gln Gln Leu Val Ala Leu Leu Val  
 545 550 555 560  
 Arg Phe Glu Lys Leu Leu Glu Ser Asp Ala Pro Leu Ala Glu Val Leu  
 565 570 575  
 Pro Ser Ile Tyr Lys Gln His Glu Glu Arg Tyr Ala Gly Tyr Thr Leu  
 580 585 590  
 Arg Gln Leu Cys Gln Glu Met His Asp Leu Tyr Ala Arg His Asn Val  
 595 600 605  
 Lys Gln Leu Gln Lys Glu Met Phe Arg Lys Glu His Phe Pro Arg Val



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Val	Val	Pro	Val	Trp	Leu	Lys	Pro	Thr	Arg	Asn	Ala	Leu	Gly	Ile	Leu
			260					265					270		
Gly	Gly	Ile	Pro	Arg	Arg	Glu	Phe	Thr	Arg	Asp	Ser	Ile	Glu	Glu	Lys
		275					280					285			
Val	Ala	Ala	Thr	Thr	Gln	Ala	Gln	Trp	Pro	Val	His	Ala	Val	Ile	Thr
	290					295					300				
Asn	Ser	Thr	Tyr	Asp	Gly	Leu	Leu	Tyr	Asn	Thr	Asp	Trp	Ile	Lys	Gln
305					310					315					320
Thr	Leu	Asp	Val	Pro	Ser	Ile	His	Phe	Asp	Ser	Ala	Trp	Val	Pro	Tyr
				325					330					335	
Thr	His	Phe	His	Pro	Ile	Tyr	Gln	Gly	Lys	Ser	Gly	Met	Ser	Gly	Glu
			340					345					350		
Arg	Val	Ala	Gly	Lys	Val	Ile	Phe	Glu	Thr	Gln	Ser	Thr	His	Lys	Met
		355					360						365		
Leu	Ala	Ala	Leu	Ser	Gln	Ala	Ser	Leu	Ile	His	Ile	Lys	Gly	Glu	Tyr
	370					375					380				
Asp	Glu	Glu	Ala	Phe	Asn	Glu	Ala	Phe	Met	Met	His	Thr	Thr	Thr	Ser
385					390					395					400
Pro	Ser	Tyr	Pro	Ile	Val	Ala	Ser	Val	Glu	Thr	Ala	Ala	Ala	Met	Leu
				405					410					415	
Arg	Gly	Asn	Pro	Gly	Lys	Arg	Leu	Ile	Asn	Arg	Ser	Val	Glu	Arg	Ala
			420					425					430		
Leu	His	Phe	Arg	Lys	Glu	Val	Gln	Arg	Leu	Arg	Glu	Glu	Ser	Asp	Gly
		435					440					445			
Trp	Phe	Phe	Asp	Ile	Trp	Gln	Pro	Pro	Gln	Val	Asp	Glu	Ala	Glu	Cys
	450					455					460				
Trp	Pro	Val	Ala	Pro	Gly	Glu	Gln	Trp	His	Gly	Phe	Asn	Asp	Ala	Asp
465					470					475					480
Ala	Asp	His	Met	Phe	Leu	Asp	Pro	Val	Lys	Val	Thr	Ile	Leu	Thr	Pro
				485					490					495	
Gly	Met	Asp	Glu	Gln	Gly	Asn	Met	Ser	Glu	Glu	Gly	Ile	Pro	Ala	Ala
			500					505					510		
Leu	Val	Ala	Lys	Phe	Leu	Asp	Glu	Arg	Gly	Ile	Val	Val	Glu	Lys	Thr
		515					520					525			
Gly	Pro	Tyr	Asn	Leu	Leu	Phe	Leu	Phe	Ser	Ile	Gly	Ile	Asp	Lys	Thr
	530					535					540				
Lys	Ala	Met	Gly	Leu	Leu	Arg	Gly	Leu	Thr	Glu	Phe	Lys	Arg	Ser	Tyr
545					550					555					560
Asp	Leu	Asn	Leu	Arg	Ile	Lys	Asn	Met	Leu	Pro	Asp	Leu	Tyr	Ala	Glu
			565						570					575	
Asp	Pro	Asp	Phe	Tyr	Arg	Asn	Met	Arg	Ile	Gln	Asp	Leu	Ala	Gln	Gly
			580					585					590		
Ile	His	Lys	Leu	Ile	Arg	Lys	His	Asp	Leu	Pro	Gly	Leu	Met	Leu	Arg
		595					600					605			
Ala	Phe	Asp	Thr	Leu	Pro	Glu	Met	Ile	Met	Thr	Pro	His	Gln	Ala	Trp
	610					615					620				
Gln	Arg	Gln	Ile	Lys	Gly	Glu	Val	Glu	Thr	Ile	Ala	Leu	Glu	Gln	Leu
625					630					635					640
Val	Gly	Arg	Val	Ser	Ala	Asn	Met	Ile	Leu	Pro	Tyr	Pro	Pro	Gly	Val
				645					650					655	
Pro	Leu	Leu	Met	Pro	Gly	Glu	Met	Leu	Thr	Lys	Glu	Ser	Arg	Thr	Val
			660					665					670		
Leu	Asp	Phe	Leu	Leu	Met	Leu	Cys	Ser	Val	Gly	Gln	His	Tyr	Pro	Gly





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Val Ala Gly Pro Leu Cys Glu Ser Gly Asp Val Phe Thr Gln Gln Glu  
                   340                                  345                                  350

Gly Gly Asn Val Glu Thr Arg Ala Leu Pro Glu Val Lys Ala Gly Asp  
                   355                                  360                                  365

Tyr Leu Val Leu His Asp Thr Gly Ala Tyr Gly Ala Ser Met Ser Ser  
           370                                  375                                  380

Asn Tyr Asn Ser Arg Pro Leu Leu Pro Glu Val Leu Phe Asp Asn Gly  
 385                                  390                                  395                                  400

Gln Ala Arg Leu Ile Arg Arg Arg Gln Thr Ile Glu Glu Leu Leu Ala  
                                   405                                  410                                  415

Leu Glu Leu Leu  
           420

<210> SEQ ID NO 34  
 <211> LENGTH: 550  
 <212> TYPE: PRT  
 <213> ORGANISM: Salmonella typhimurium

<400> SEQUENCE: 34

Met Gln Asn Pro Tyr Thr Val Ala Asp Tyr Leu Leu Asp Arg Leu Ala  
   1                  5                                  10                                  15

Gly Cys Gly Ile Gly His Leu Phe Gly Val Pro Gly Asp Tyr Asn Leu  
                   20                                  25                                  30

Gln Phe Leu Asp His Val Ile Asp His Pro Thr Leu Arg Trp Val Gly  
           35                                  40                                  45

Cys Ala Asn Glu Leu Asn Ala Ala Tyr Ala Ala Asp Gly Tyr Ala Arg  
   50                                  55                                  60

Met Ser Gly Ala Gly Ala Leu Leu Thr Thr Phe Gly Val Gly Glu Leu  
   65                                  70                                  75                                  80

Ser Ala Ile Asn Gly Ile Ala Gly Ser Tyr Ala Glu Tyr Val Pro Val  
                   85                                  90                                  95

Leu His Ile Val Gly Ala Pro Cys Ser Ala Ala Gln Gln Arg Gly Glu  
           100                                  105                                  110

Leu Met His His Thr Leu Gly Asp Gly Asp Phe Arg His Phe Tyr Arg  
           115                                  120                                  125

Met Ser Gln Ala Ile Ser Ala Ala Ser Ala Ile Leu Asp Glu Gln Asn  
   130                                  135                                  140

Ala Cys Phe Glu Ile Asp Arg Val Leu Gly Glu Met Leu Ala Ala Arg  
   145                                  150                                  155                                  160

Arg Pro Gly Tyr Ile Met Leu Pro Ala Asp Val Ala Lys Lys Thr Ala  
           165                                  170                                  175

Ile Pro Pro Thr Gln Ala Leu Ala Leu Pro Val His Glu Ala Gln Ser  
           180                                  185                                  190

Gly Val Glu Thr Ala Phe Arg Tyr His Ala Arg Gln Cys Leu Met Asn  
           195                                  200                                  205

Ser Arg Arg Ile Ala Leu Leu Ala Asp Phe Leu Ala Gly Arg Phe Gly  
   210                                  215                                  220

Leu Arg Pro Leu Leu Gln Arg Trp Met Ala Glu Thr Pro Ile Ala His  
   225                                  230                                  235                                  240

Ala Thr Leu Leu Met Gly Lys Gly Leu Phe Asp Glu Gln His Pro Asn  
           245                                  250                                  255

Phe Val Gly Thr Tyr Ser Ala Gly Ala Ser Ser Lys Glu Val Arg Gln  
           260                                  265                                  270

Ala Ile Glu Asp Ala Asp Arg Val Ile Cys Val Gly Thr Arg Phe Val  
           275                                  280                                  285

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Asp Thr Leu Thr Ala Gly Phe Thr Gln Gln Leu Pro Ala Glu Arg Thr  
 290 295 300  
 Leu Glu Ile Gln Pro Tyr Ala Ser Arg Ile Gly Glu Thr Trp Phe Asn  
 305 310 315 320  
 Leu Pro Met Ala Gln Ala Val Ser Thr Leu Arg Glu Leu Cys Leu Glu  
 325 330 335  
 Cys Ala Phe Ala Pro Pro Pro Thr Arg Ser Ala Gly Gln Pro Val Arg  
 340 345 350  
 Ile Asp Lys Gly Glu Leu Thr Gln Glu Ser Phe Trp Gln Thr Leu Gln  
 355 360 365  
 Gln Tyr Leu Lys Pro Gly Asp Ile Ile Leu Val Asp Gln Gly Thr Ala  
 370 375 380  
 Ala Phe Gly Ala Ala Ala Leu Ser Leu Pro Asp Gly Ala Glu Val Val  
 385 390 395 400  
 Leu Gln Pro Leu Trp Gly Ser Ile Gly Tyr Ser Leu Pro Ala Ala Phe  
 405 410 415  
 Gly Ala Gln Thr Ala Cys Pro Asp Arg Arg Val Ile Leu Ile Ile Gly  
 420 425 430  
 Asp Gly Ala Ala Gln Leu Thr Ile Gln Glu Met Gly Ser Met Leu Arg  
 435 440 445  
 Asp Gly Gln Ala Pro Val Ile Leu Leu Leu Asn Asn Asp Gly Tyr Thr  
 450 455 460  
 Val Glu Arg Ala Ile His Gly Ala Ala Gln Arg Tyr Asn Asp Ile Ala  
 465 470 475 480  
 Ser Trp Asn Trp Thr Gln Ile Pro Pro Ala Leu Asn Ala Ala Gln Gln  
 485 490 495  
 Ala Glu Cys Trp Arg Val Thr Gln Ala Ile Gln Leu Ala Glu Val Leu  
 500 505 510  
 Glu Arg Leu Ala Arg Pro Gln Arg Leu Ser Phe Ile Glu Val Met Leu  
 515 520 525  
 Pro Lys Ala Asp Leu Pro Glu Leu Leu Arg Thr Val Thr Arg Ala Leu  
 530 535 540  
 Glu Ala Arg Asn Gly Gly  
 545 550

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What is claimed is:

1. A method of biosynthesizing 2-aminopimelate in a recombinant host, the method comprising enzymatically converting 2,6-diaminopimelate to 2-aminopimelate using at least one polypeptide having an activity selected from 2-hydroxyacyl-CoA dehydratase activity, mutase activity, ammonia lyase activity, and enoate reductase activity, wherein said polypeptide having enoate reductase activity has at least 85% sequence identity to the amino acid sequence set forth in any one of SEQ ID NOs: 16-22, wherein said polypeptide having 2-hydroxyacyl-CoA dehydratase activity has at least 85% sequence identity to the amino acid sequence set forth in SEQ ID NO: 25 or SEQ ID NO: 28, wherein said polypeptide having mutase activity has at least 85% sequence identity to the amino acid sequence set forth in SEQ ID NO: 26, and wherein said polypeptide having ammonia lyase activity has at least 85% sequence identity to the amino acid sequence set forth in SEQ ID NO: 23, the method optionally further comprising using at least one polypeptide having an activity selected from diaminopimelate dehydrogenase activity, 2-hydroxy-

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carboxylate dehydrogenase activity, CoA-transferase activity, and carboxylate reductase activity to enzymatically convert 2,6-diaminopimelate to 2-aminopimelate, wherein said polypeptide having diaminopimelate dehydrogenase activity is classified under EC 1.4.1.16, wherein said polypeptide having 2-hydroxycarboxylate dehydrogenase activity is classified under EC 1.1.1.337, wherein said polypeptide having CoA-transferase activity is classified under EC 2.8.3, and wherein said polypeptide having carboxylate reductase activity has at least 85% sequence identity to the amino acid sequence set forth in any one of SEQ ID NOs: 3-7.

2. The method of claim 1, wherein (S) 2-aminopimelate is biosynthesized.

3. The method of claim 1, said method comprising: using said polypeptide having 2-hydroxyacyl-CoA dehydratase activity and said polypeptide having enoate reductase activity to enzymatically convert 2,6-diaminopimelate to 2-aminopimelate; or using said polypeptide having mutase activity, said polypeptide having ammonia lyase activity, and said polypeptide having

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peptide having enoate reductase activity to enzymatically convert 2,6-diaminopimelate to 2-aminopimelate.

4. The method of claim 1, wherein (R) 2-aminopimelate is biosynthesized.

5. The method of claim 1, said method further comprising using at least one polypeptide having an activity selected from CoA ligase activity, CoA-transferase activity, carboxylate reductase activity, and aldehyde dehydrogenase activity to enzymatically convert 2,6-diaminopimelate to 2-aminopimelate, wherein said polypeptide having CoA ligase activity is classified under EC 6.2.1.5, wherein said polypeptide having CoA-transferase activity is classified under EC 2.8.3, wherein said polypeptide having carboxylate reductase activity has at least 85% sequence identity to the amino acid sequence set forth in any one of SEQ ID NOs: 3-7, and wherein said aldehyde dehydrogenase is classified under EC 1.2.1.-.

6. The method of claim 1, wherein the host is subjected to a cultivation strategy under aerobic or micro-aerobic cultivation conditions.

7. The method of claim 1, wherein the host is cultured under conditions of nitrogen, phosphate or oxygen limitation.

8. The method of claim 1, wherein the host is retained using a ceramic membrane to maintain a high cell density during fermentation.

9. The method of claim 1, further comprising a principal carbon source fed to the fermentation derived from a biological feedstock.

10. The method of claim 9, wherein the biological feedstock is or derives from monosaccharides, disaccharides, lignocellulose, hemicellulose, cellulose, lignin, levulinic acid and formic acid, triglycerides, glycerol, fatty acids, agricultural waste, condensed distillers' solubles, or municipal waste.

11. The method of claim 1, further comprising a principal carbon source fed to the fermentation derived from a non-biological feedstock.

12. The method of claim 11, wherein the non-biological feedstock is, or derives from, natural gas, syngas, CO<sub>2</sub>/H<sub>2</sub>, methanol, ethanol, benzoate, non-volatile residue (NVR) or a caustic wash waste stream from cyclohexane oxidation processes, or terephthalic acid/isophthalic acid mixture waste streams.

13. The method of claim 1, wherein the host is a prokaryote selected from *Escherichia*, *Clostridia*, *Corynebacteria*, *Cupriavidus*, *Pseudomonas*, *Delftia*, *Bacillus*, *Lactobacillus*, *Lactococcus*, and *Rhodococcus*, or a eukaryote selected from *Aspergillus*, *Saccharomyces*, *Pichia*, *Yarrowia*, *Issatchenkia*, *Debaryomyces*, *Arxula*, and *Kluyveromyces*.

14. The method of claim 1, wherein the host exhibits tolerance to high concentrations of a C6 building block, and wherein the tolerance to high concentrations of a C6 building block is improved through continuous cultivation in a selective environment.

15. The method of claim 1, wherein the host comprises one or more of the following: the intracellular concentration of oxaloacetate for biosynthesis of a C6 building block is increased in the host by overexpressing recombinant genes forming oxaloacetate; wherein an imbalance in NADPH is generated that can be balanced via the formation of a C6 building block; wherein an exogenous lysine biosynthesis pathway synthesizing lysine from 2-oxoglutarate via 2-oxoadipate is introduced in a host using the meso 2,6 diamino-

oxaloacetate to meso 2,6 diaminopimelate is introduced in a host using the 2-oxoadipate pathway for lysine synthesis; wherein endogenous degradation pathways of central metabolites and central precursors leading to and including C6 building blocks are attenuated in the host; or wherein the efflux of a C6 building block across the cell membrane to the extracellular media is enhanced or amplified by genetically engineering structural modifications to the cell membrane or increasing any associated transporter activity for a C6 building block.

16. A recombinant host cell comprising at least one exogenous nucleic acid encoding at least one polypeptide having an activity selected from 2-hydroxyacyl-CoA dehydratase activity, mutase activity, ammonia lyase activity, and enoate reductase activity, wherein said polypeptide having enoate reductase activity has at least 85% sequence identity to the amino acid sequence set forth in any one of SEQ ID NOs: 16-22, said polypeptide having 2-hydroxyacyl-CoA dehydratase activity has at least 85% sequence identity to the amino acid sequence set forth in SEQ ID NO: 25 or SEQ ID NO: 28, said polypeptide having mutase activity has at least 85% sequence identity to the amino acid sequence set forth in SEQ ID NO: 26, said polypeptide having ammonia lyase activity has at least 85% sequence identity to the amino acid sequence set forth in SEQ ID NO: 23, said host producing 2-aminopimelate from 2,6-diaminopimelate,

the host optionally further comprising one or more exogenous polypeptides having an activity selected from aldehyde dehydrogenase activity, alcohol dehydrogenase activity, CoA-transferase activity, carboxylate reductase activity,  $\alpha$ -aminotransferase activity, thioesterase activity, hydrolase activity,  $\omega$ -transaminase activity, N-acetyltransferase activity, and deacylase activity, the host producing a product selected from adipic acid, adipate semialdehyde, 6-aminohexanoic acid, 6-hydroxyhexanoic acid, caprolactam, hexamethylenediamine, and 1,6-hexanediol, wherein said polypeptide having aldehyde dehydrogenase activity is classified under EC 1.2.1, wherein said polypeptide having alcohol dehydrogenase activity is classified under EC 1.1.1, wherein said polypeptide having CoA-transferase activity is classified under EC 2.8.3, wherein said polypeptide having carboxylate reductase activity has at least 85% sequence identity to the amino acid sequence set forth in any one of SEQ ID NOs: 3-7 wherein said polypeptide having  $\alpha$ -aminotransferase activity classified under EC 2.6.1, wherein said polypeptide having thioesterase activity has at least 85% sequence identity to the amino acid sequence set forth in SEQ ID NO: 1 or SEQ ID NO: 2, wherein said polypeptide having hydrolase activity classified under EC 3.5.2, wherein said polypeptide having  $\omega$ -transaminase activity has at least 85% sequence identity to the amino acid sequence set forth in any one of SEQ ID NOs: 8-13, wherein said polypeptide having N-acetyltransferase activity is classified under EC 2.3.1.32 and wherein said polypeptide having deacylase activity is classified under EC 3.5.1.17.

17. The host of claim 16, said host comprising said polypeptide having exogenous 2-hydroxyacyl-CoA dehydratase activity and said polypeptide having enoate reductase activity.

18. The host of claim 16, said host comprising said polypeptide having mutase activity, said polypeptide having ammonia lyase activity, and said polypeptide having enoate reductase activity.

19. The host of claim 16, said host further comprising at least one polypeptide having an activity selected from a) diaminopimelate dehydrogenase activity, 2-hydroxycarboxylate dehydrogenase activity, CoA-transferase activity, and carboxylate reductase activity, wherein said polypeptide having diaminopimelate dehydrogenase activity is classified under EC 1.4.1.16, wherein said polypeptide having 2-hydroxycarboxylate dehydrogenase activity is classified under EC 1.1.1.337, wherein said polypeptide having CoA-transferase activity is classified under EC 2.8.3, and wherein said polypeptide having carboxylate reductase activity has at least 85% sequence identity to the amino acid sequence set forth in any one of SEQ ID NOs: 3-7; or b) CoA ligase activity, CoA-transferase activity, carboxylate reductase activity, and aldehyde dehydrogenase activity, wherein said polypeptide having CoA ligase activity is classified under EC 6.2.1.5, wherein said polypeptide having CoA-transferase activity is classified under EC 2.8.3, wherein said polypeptide having carboxylate reductase activity has at least 85% sequence identity to the amino acid sequence set forth in any one of SEQ ID NOs: 3-7 and wherein said polypeptide having aldehyde dehydrogenase activity is classified under EC 1.2.1.

20. The host of claim 16, further comprising at least one exogenous polypeptide having an activity selected from 2-oxoacid decarboxylase activity classified under EC 4.1.1,  $\alpha$ -aminoacid decarboxylase activity classified under EC 4.1.1, synthase activity, and activity of a dehydrogenase complex, wherein the polypeptide having 2-oxoacid decarboxylase activity is classified under EC 4.1.1.43, EC 4.1.1.71, EC 4.1.1.72, or EC 4.1.1.74 and has at least 85% sequence identity to the amino acid sequence set forth in SEQ ID NO: 34; the polypeptide having  $\alpha$ -aminoacid decarboxylase activity is classified under EC 4.1.1.15, EC 4.1.1.17, or EC 4.1.1.18, or EC 4.1.1.19 and has at least 85% sequence identity to the amino acid sequence set forth in any one of SEQ ID NOs: 29-34; the polypeptide having synthase activity is classified under EC 2.2.1.6, or the polypeptide having the activity of a dehydrogenase complex comprises activities classified under EC 1.2.4.2 EC 1.8.1.4, or EC 2.3.1.61.

21. The recombinant host of claim 19, wherein the host cell further comprises at least one exogenous polypeptide having an activity selected from  $\alpha$ -aminotransferase activity, 2-oxoacid decarboxylase activity, activity of a dehydrogenase complex, thioesterase activity, CoA-transferase activity, CoA-ligase activity, and aldehyde dehydrogenase activity, the host producing adipic acid, wherein said polypeptide having  $\alpha$ -aminotransferase activity is classified under EC 2.6.1, wherein said polypeptide having 2-oxoacid decarboxylase activity has at least 85% sequence identity to the amino acid sequence set forth in SEQ ID NO: 34, wherein said polypeptide having the activity of a dehydrogenase complex is classified under EC 1.2.4.2, EC 1.8.1.4, or EC 2.3.1.61, wherein said polypeptide having thioesterase activity has at least 85% sequence identity to the amino acid sequence set forth in SEQ ID NO: 1 or SEQ ID NO: 2, wherein said polypeptide having CoA-transferase activity is classified under EC 2.8.3, wherein said polypeptide having CoA-ligase activity is classified under EC 6.2.1.5, and wherein said polypeptide having aldehyde dehydrogenase activity is classified under EC 1.2.1,

the host cell optionally further comprising at least one exogenous polypeptide having an activity selected from  $\alpha$ -aminotransferase activity, 2-oxoacid decarboxylase activity, and synthase activity, the host producing adipate semialdehyde, wherein said polypeptide

having  $\alpha$ -aminotransferase activity is classified under EC 2.6.1-, wherein said polypeptide having 2-oxoacid decarboxylase activity has at least 85% sequence identity to the amino acid sequence set forth in SEQ ID NO: 34, and wherein said polypeptide having synthase activity is classified under EC 2.2.1.6.

22. The recombinant host of claim 16, wherein the host cell further comprises:

an exogenous polypeptide having  $\alpha$ -aminoacid decarboxylase activity that has at least 85% sequence identity to the amino acid sequence set forth in any one of SEQ ID NOs: 29-34, the host producing 6-amino-hexanoic acid; or

an exogenous  $\omega$ -transaminase that has at least 85% sequence identity to the amino acid sequence set forth in any one of SEQ ID NOs: 8-13, the host producing 6-aminohexanoic acid;

the host cell optionally further comprising an exogenous polypeptide having hydrolase activity, wherein said polypeptide having hydrolase activity is classified under EC 3.5.2, the host producing caprolactam.

23. The recombinant host of claim 22, wherein the host cell further comprises one or more of an exogenous polypeptide having carboxylate reductase activity, N-acetyltransferase activity,  $\omega$ -transaminase activity, or deacylase activity, the host producing hexamethylenediamine, wherein said polypeptide having carboxylate reductase activity has at least 85% sequence identity to the amino acid sequence set forth in any one of SEQ ID NOs: 3-7, wherein said polypeptide having N-acetyltransferase activity is classified under EC 2.3.1.32, wherein said polypeptide having  $\omega$ -transaminase activity has at least 85% sequence identity to the amino acid sequence set forth in any one of SEQ ID NOs: 8-13, and wherein said polypeptide having deacylase activity is classified under EC 3.5.1.17.

24. The recombinant host of claim 21, wherein the host cell further comprises at least one exogenous polypeptide having carboxylate reductase activity and/or at least one exogenous polypeptide having r-transaminase activity, the host producing hexamethylenediamine, wherein said polypeptide having carboxylate reductase activity has at least 85% sequence identity to the amino acid sequence set forth in any one of SEQ ID NOs: 3-7 and wherein said polypeptide having  $\omega$ -transaminase activity has at least 85% sequence identity to the amino acid sequence set forth in any one of SEQ ID NOs: 8-13.

25. The recombinant host of claim 16, wherein the host cell further comprises at least one exogenous polypeptide having an activity selected from  $\alpha$ -aminotransferase activity, 2-oxoacid decarboxylase activity, alcohol dehydrogenase activity, and synthase activity, the host producing 6-hydroxyhexanoic acid, wherein said polypeptide having  $\alpha$ -aminotransferase activity is classified under EC 2.6.1, wherein said polypeptide having 2-oxoacid decarboxylase activity has at least 85% sequence identity to the amino acid sequence set forth in SEQ ID NO: 34, wherein said polypeptide having alcohol dehydrogenase activity is classified under EC 1.1.1, and wherein said polypeptide having synthase activity is classified under EC 2.2.1.6, the host cell optionally further comprising:

at least one exogenous polypeptide having an activity selected from carboxylate reductase activity,  $\omega$ -transaminase activity, and alcohol dehydrogenase activity, the host producing hexamethylenediamine, wherein said polypeptide having carboxylate reductase activity has at least 85% sequence identity to the amino acid sequence set forth in any one of SEQ ID NOs: 3-7,

wherein said polypeptide having cg-transaminase activity has at least 85% sequence identity to the amino acid sequence set forth in any one of SEQ ID NOs: 8-13 and wherein said polypeptide having alcohol dehydrogenase activity is classified under EC 1.1.1; or  
 an exogenous polypeptide having carboxylate reductase activity and/or an exogenous polypeptide having alcohol dehydrogenase activity, the host producing 1,6-hexanediol, wherein said polypeptide having carboxylate reductase activity has at least 85% sequence identity to the amino acid sequence set forth in any one of SEQ ID NOs: 3-7 and wherein said polypeptide having alcohol dehydrogenase activity is classified under EC 1.1.1.

**26.** A method of biosynthesizing 2-aminopimelate in a recombinant host, the method comprising enzymatically converting 2,6-diaminopimelate to 2-aminopimelate using at least one polypeptide selected from a polypeptide having the activity of an enoate reductase, classified under EC 1.3.1.31 or EC 1.6.99.1, or having at least 85% sequence identity to the amino acid sequence set forth in any one of SEQ ID NOs: 16-22; a polypeptide having the activity of a 2-hydroxyacyl-CoA dehydratase, classified under EC 4.2.1, or having at least 85% sequence identity to the amino acid sequence set forth in SEQ ID NO: 25 or SEQ ID NO: 28; a polypeptide having the activity of a mutase, classified under EC 5.4.3.2, or having at least 85% sequence identity to the amino acid sequence set forth in SEQ ID NO: 26; and a polypeptide having the activity of an ammonia lyase, classified under EC 4.3.1.14, or having at least 85% sequence identity to the amino acid sequence set forth in SEQ ID NO: 23, the method optionally further comprising using at least one polypeptide selected from a polypeptide having the activity of a diaminopimelate dehydrogenase or classified under EC 1.4.1.16; a polypeptide having the activity of a CoA-transferase, classified under EC 2.8.3, classified under EC 2.8.3.12, or having at least 85% sequence identity to the amino acid sequence of the gene product of thnH or gctAB; and a polypeptide having the activity of a carboxylate reductase, classified under EC 1.2.99.6, having at least 85% sequence identity to the amino acid sequence set forth in any one of SEQ ID NOs: 5-7, having at least 85% sequence identity to the amino acid sequence of the gene product of car and npt, or having at least 85% sequence identity to the amino acid sequence of the gene product of griC and griD.

**27.** The method of claim **26**, wherein (S) 2-aminopimelate is biosynthesized.

**28.** The method of claim **26**, said method comprising: using said polypeptide having the activity of a 2-hydroxyacyl-CoA dehydratase, classified under EC 4.2.1, or having at least 85% sequence identity to the amino acid sequence set forth in SEQ ID NO: 25 or SEQ ID NO: 28 and said polypeptide having the activity of an enoate reductase, classified under EC 1.3.1.31 or EC 1.6.99.1, or having at least 85% sequence identity to the amino acid sequence set forth in any one of SEQ ID NOs: 16-22 to enzymatically convert 2,6-diaminopimelate to 2-aminopimelate; or

using said a polypeptide having the activity of a mutase, classified under EC 5.4.3.2, or having at least 85% sequence identity to the amino acid sequence set forth in SEQ ID NO: 26, said polypeptide having the activity of an ammonia lyase, classified under EC 4.3.1.14, or having at least 85% sequence identity to the amino acid sequence set forth in SEQ ID NO: 23, and said polypeptide having the activity of an enoate reductase, classified under EC 1.3.1.31 or EC 1.6.99.1, or having

at least 85% sequence identity to the amino acid sequence set forth in any one of SEQ ID NOs: 16-22 to enzymatically convert 2,6-diaminopimelate to 2-aminopimelate.

**29.** The method of claim **26**, wherein (R) 2-aminopimelate is biosynthesized.

**30.** The method of claim **26**, said method further comprising using at least one polypeptide selected a polypeptide having the activity of a CoA ligase, having the activity of a succinate-CoA ligase, or classified under EC 6.2.1.5, a polypeptide having the activity of a CoA-transferase, having the activity of a glutaconate CoA-transferase, classified under EC 2.8.3, classified under EC 2.8.3.12, having at least 85% sequence identity to the amino acid sequence of the gene product of thnH, or having at least 85% sequence identity to the amino acid sequence of the gene product of gctAB; a polypeptide having the activity of a carboxylate reductase, classified under EC 1.2.99.6, having at least 85% sequence identity to the amino acid sequence set forth in any one of SEQ ID NOs: 5-7, having at least 85% sequence identity to the amino acid sequence of the gene product of car and npt, or having at least 85% sequence identity to the amino acid sequence of the gene product of griC and griD, and a polypeptide having the activity of an aldehyde dehydrogenase, classified under EC 1.2.1, or classified under EC 1.2.1.3, EC 1.2.1.16, EC 1.2.1.20, EC 1.2.1.63, or EC 1.2.1.79 to enzymatically convert 2,6-diaminopimelate to 2-aminopimelate.

**31.** The method of claim **26**, wherein the host comprises one or more of the following: the intracellular concentration of oxaloacetate for biosynthesis of a C6 building block is increased in the host by overexpressing recombinant genes forming oxaloacetate; wherein an imbalance in NADPH is generated that can be balanced via the formation of a C6 building block; wherein an exogenous lysine biosynthesis pathway synthesizing lysine from 2-oxoglutarate via 2-oxoadipate is introduced in a host using the meso 2,6 diaminopimelate pathway for lysine synthesis; wherein an exogenous lysine biosynthesis pathway synthesizing lysine from oxaloacetate to meso 2,6 diaminopimelate is introduced in a host using the 2-oxoadipate pathway for lysine synthesis; wherein endogenous degradation pathways of central metabolites and central precursors leading to and including C6 building blocks are attenuated in the host; or wherein the efflux of a C6 building block across the cell membrane to the extracellular media is enhanced or amplified by genetically engineering structural modifications to the cell membrane or increasing any associated transporter activity for a C6 building block.

**32.** The method of claim **26**, said method comprising enzymatically converting 2,6-diaminopimelate to 2-aminopimelate using at least one polypeptide selected from: a polypeptide classified under EC 1.3.1.31 or EC 1.6.99.1; a polypeptide classified under EC 4.2.1; a polypeptide classified under EC 5.4.3.2; and a polypeptide classified under EC 4.3.1.14, the method optionally further comprising using at least one polypeptide selected from: a polypeptide classified under EC 1.4.1.16, a polypeptide classified under EC 2.8.3, and a polypeptide classified under EC 1.2.99.6.

**33.** The method of claim **26**, said method comprising: using said polypeptide classified under EC 4.2.1 and said polypeptide classified under EC 1.3.1.31 or EC 1.6.99.1 to enzymatically convert 2,6-diaminopimelate to 2-aminopimelate; or using said polypeptide classified under EC 5.4.3.2, said polypeptide classified under EC 4.3.1.14, and said

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polypeptide classified under EC 1.3.1.31 or EC 1.6.99.1 to enzymatically convert 2,6-diaminopimelate to 2-aminopimelate.

**34.** The method of claim **26**, said method further comprising using at least one polypeptide selected from a 5 polypeptide classified under EC 6.2.1.5; a polypeptide classified under EC 2.8.3; a polypeptide classified under EC 1.2.99.6, and a polypeptide classified under EC 1.2.1 to enzymatically convert 2,6-diaminopimelate to 2-aminopimelate. 10

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UNITED STATES PATENT AND TRADEMARK OFFICE  
**CERTIFICATE OF CORRECTION**

PATENT NO. : 9,745,607 B2  
APPLICATION NO. : 14/714164  
DATED : August 29, 2017  
INVENTOR(S) : Alex Van Eck Conradie et al.

Page 1 of 1

It is certified that error appears in the above-identified patent and that said Letters Patent is hereby corrected as shown below:

On the Title Page

Item (71), Line 1, "INVISTA North America S.á r.l.," should read --INVISTA North America S.á.r.l.--.

Item (74) Attorney, Agent, or Firm, "Finnegan Henderson, Farabow, Garrett & Dunner, LLP; Carla A. Mouta-Bellum" should read --William J. Simmons; Carla A. Mouta-Bellum, Ph.D.--.

In the Claims

Claim 20, Column 137, Line 30, "EC 4.1.1.43." should read as --EC 4.1.1.43.--.

Claim 20, Column 137, Line 31, "4.1.1.71." should read as --4.1.1.71.--.

Claim 20, Column 137, Line 40, "EC 1.2.4.2 EC 1.8.1.4," should read as --EC 1.2.4.2, EC 1.8.1.4.--.

Claim 24, Column 138, Line 39, "r-transaminase" should read as -- $\omega$ - transaminase--.

Claim 25, Column 139, Line 1, "cg-transaminase" should read as -- $\omega$ - transaminase--.

Signed and Sealed this  
Nineteenth Day of December, 2017



Joseph Matal

*Performing the Functions and Duties of the  
Under Secretary of Commerce for Intellectual Property and  
Director of the United States Patent and Trademark Office*