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Wnek et al.

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(54) **CONTROLLED CROSS-LINKING
PROCESSING OF PROTEINS**

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US 2016/0046663 A1 Feb. 18, 2016

Related U.S. Application Data

(60) Division of application No. 13/430,562, filed on Mar.
26, 2012, now Pat. No. 9,040,665, which is a
continuation-in-part of application No. 12/571,043,
filed on Sep. 30, 2009, now Pat. No. 8,318,903.

(60) Provisional application No. 61/467,923, filed on Mar.
25, 2011, provisional application No. 61/194,685,
filed on Sep. 30, 2008.

(51) **Int. Cl.**
C07K 1/107 (2006.01)
C07K 1/36 (2006.01)
A61L 27/22 (2006.01)
C07K 14/78 (2006.01)
D01F 1/10 (2006.01)
D01F 4/00 (2006.01)

(52) **U.S. Cl.**
CPC **C07K 1/1075** (2013.01); **A61L 27/22**
(2013.01); **C07K 1/107** (2013.01); **C07K 1/36**
(2013.01); **C07K 14/78** (2013.01); **D01F 1/10**
(2013.01); **D01F 4/00** (2013.01)

(58) **Field of Classification Search**
None
See application file for complete search history.

(56) **References Cited**

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Huang et al., "A review on polymer nanofibers by electrospinning
and their applications in nanocomposites", *Composites Science and
Technology* 63 (2003) 2223-2253.*

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Primary Examiner — Suzanne M Noakes

(74) *Attorney, Agent, or Firm* — Benesch, Friedlander,
Coplan & Aronoff, LLP

(57) **ABSTRACT**

A method of forming a cross-linked protein structures
includes preparing a solution of protein dissolved in a
benign solvent and forming an intermediate protein structure
from the solution. The intermediate protein structure can be
cross-linked by providing for a specific ratio of chemical
cross-linking agents to form the cross-linked protein struc-
ture. The solution can be prepared by adding a cross-linker
of N-hydroxysuccinimide (NHS) and 1-ethyl-3-(3-dimeth-
ylaminopropyl)carbodiimide hydrochloride (EDC) at a ratio
of two-to-one of NHS to EDC to alcohol. PBS buffer (20×)
can be added to the solution until the volume ratio of PBS
buffer (20×) to alcohol is about one-to-one. About 16 percent
by weight of protein can be dissolved in the solution. The
solution can be electrospun to form an intermediate protein
structure. After a period of time, the protein structure can be
cross-linked to form the cross-linked protein structure.

13 Claims, 28 Drawing Sheets

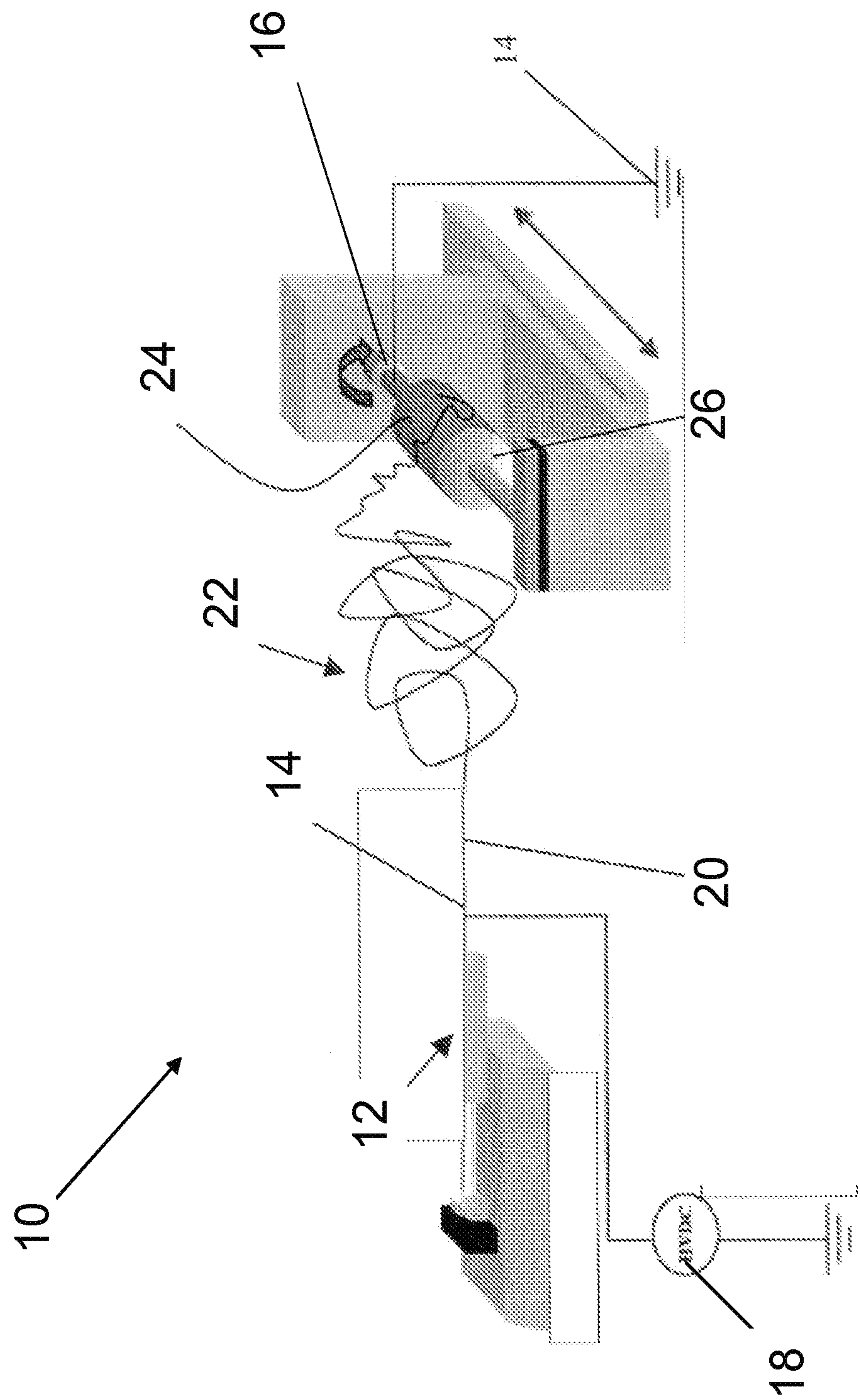


Figure 1

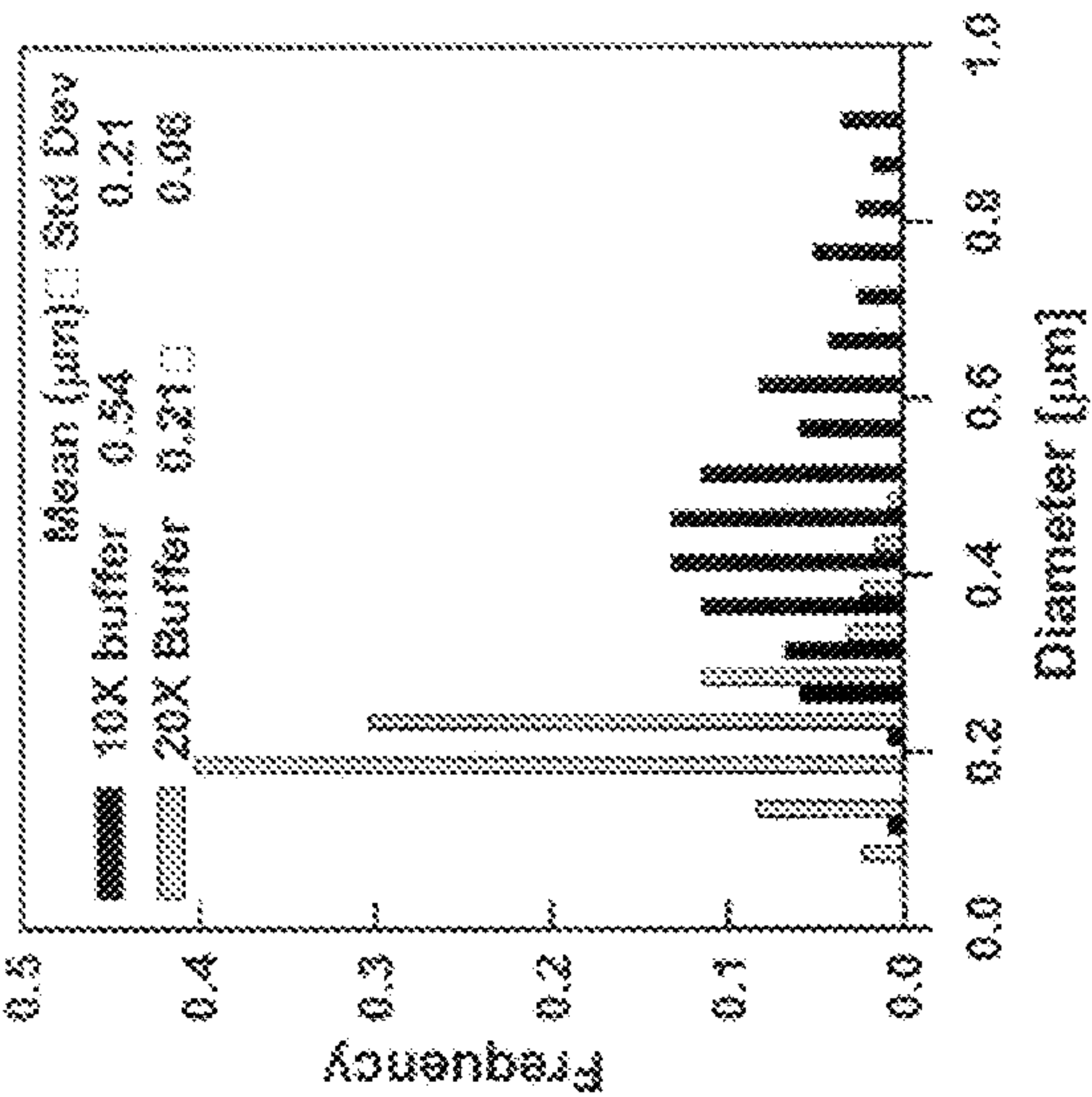


Figure 2A

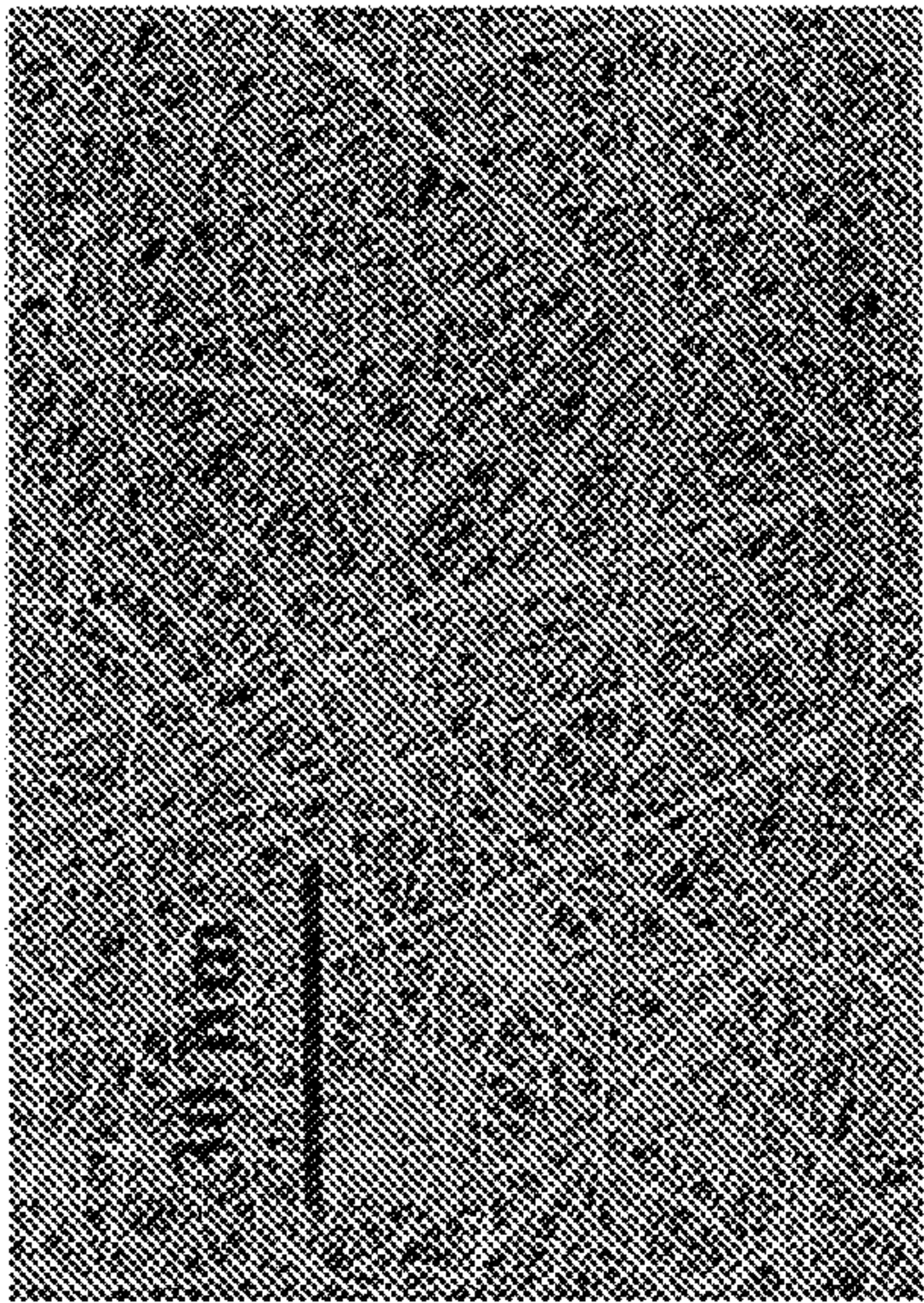


Figure 2B

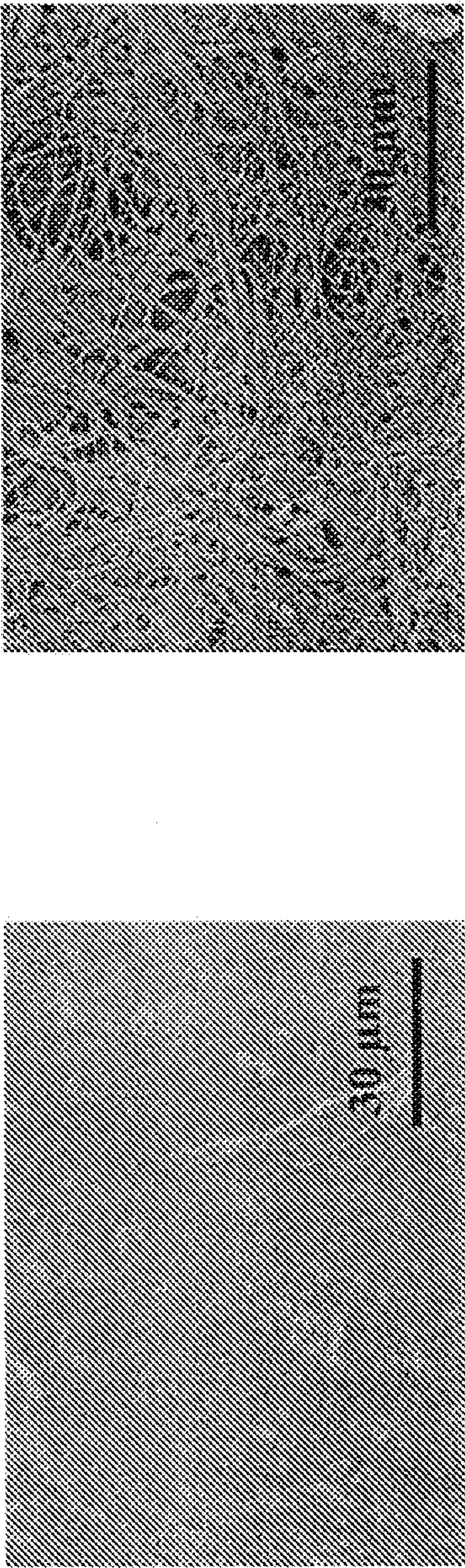


Figure 3A

Figure 3B

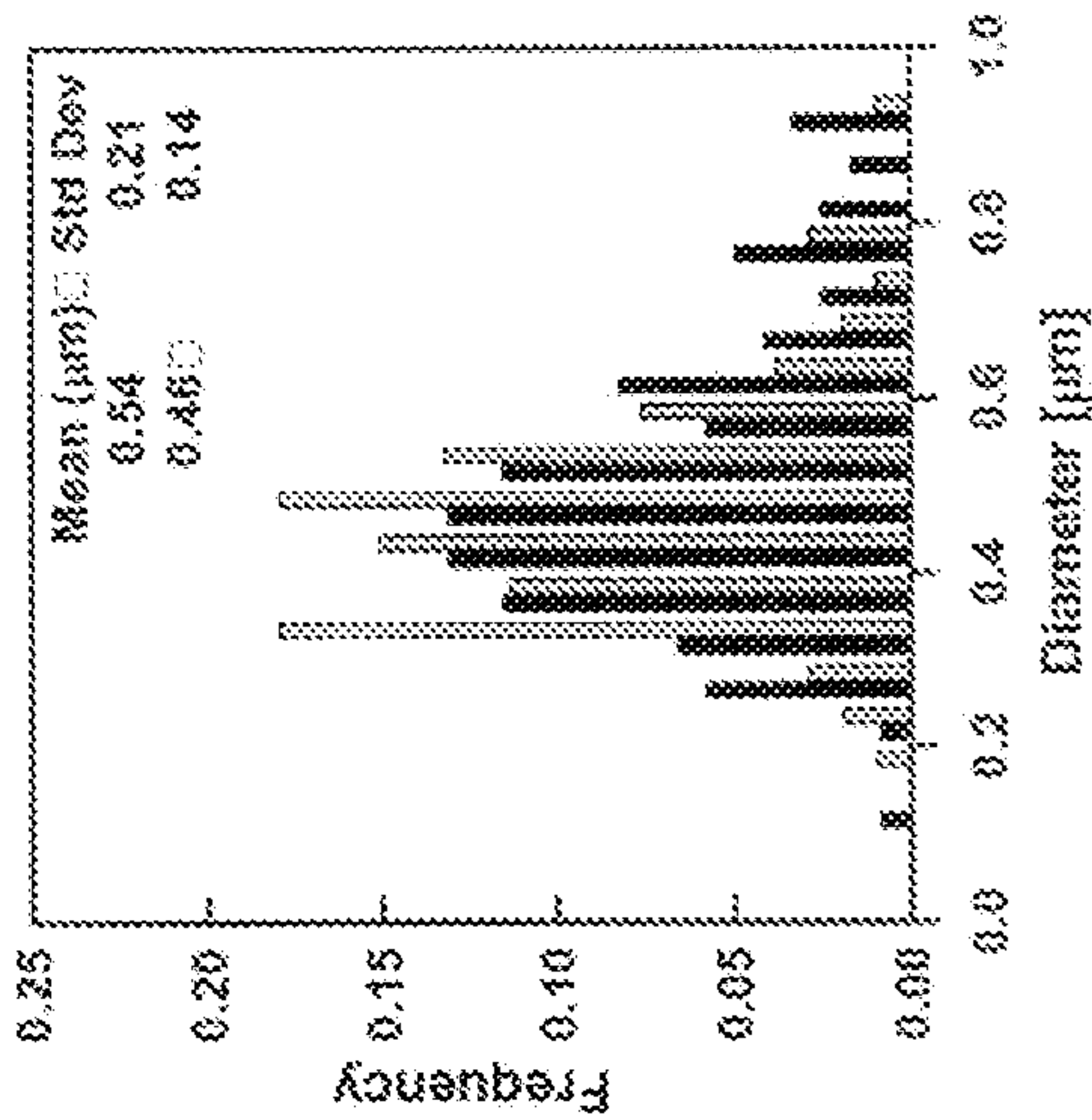


Figure 3C

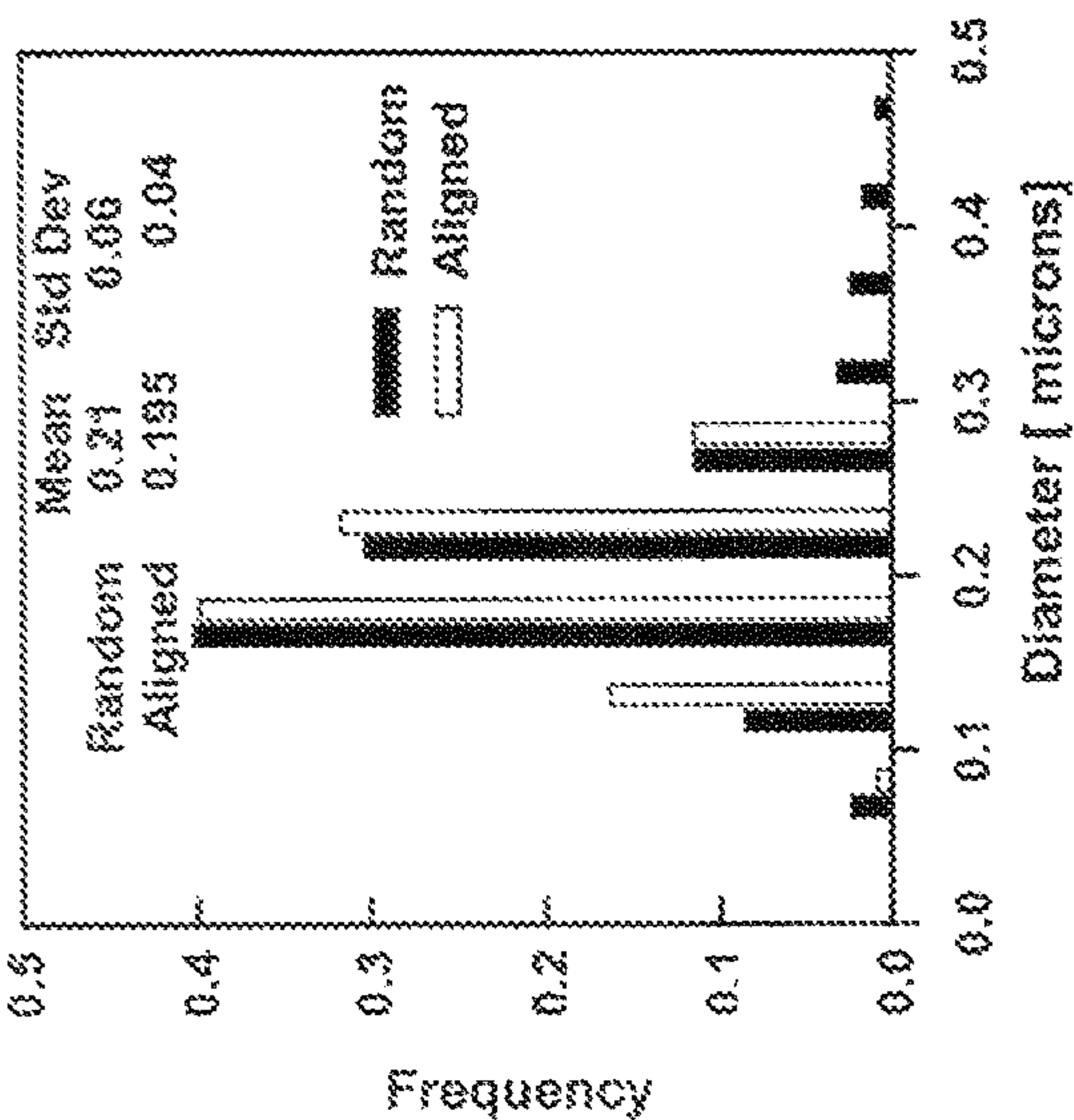


Figure 4B

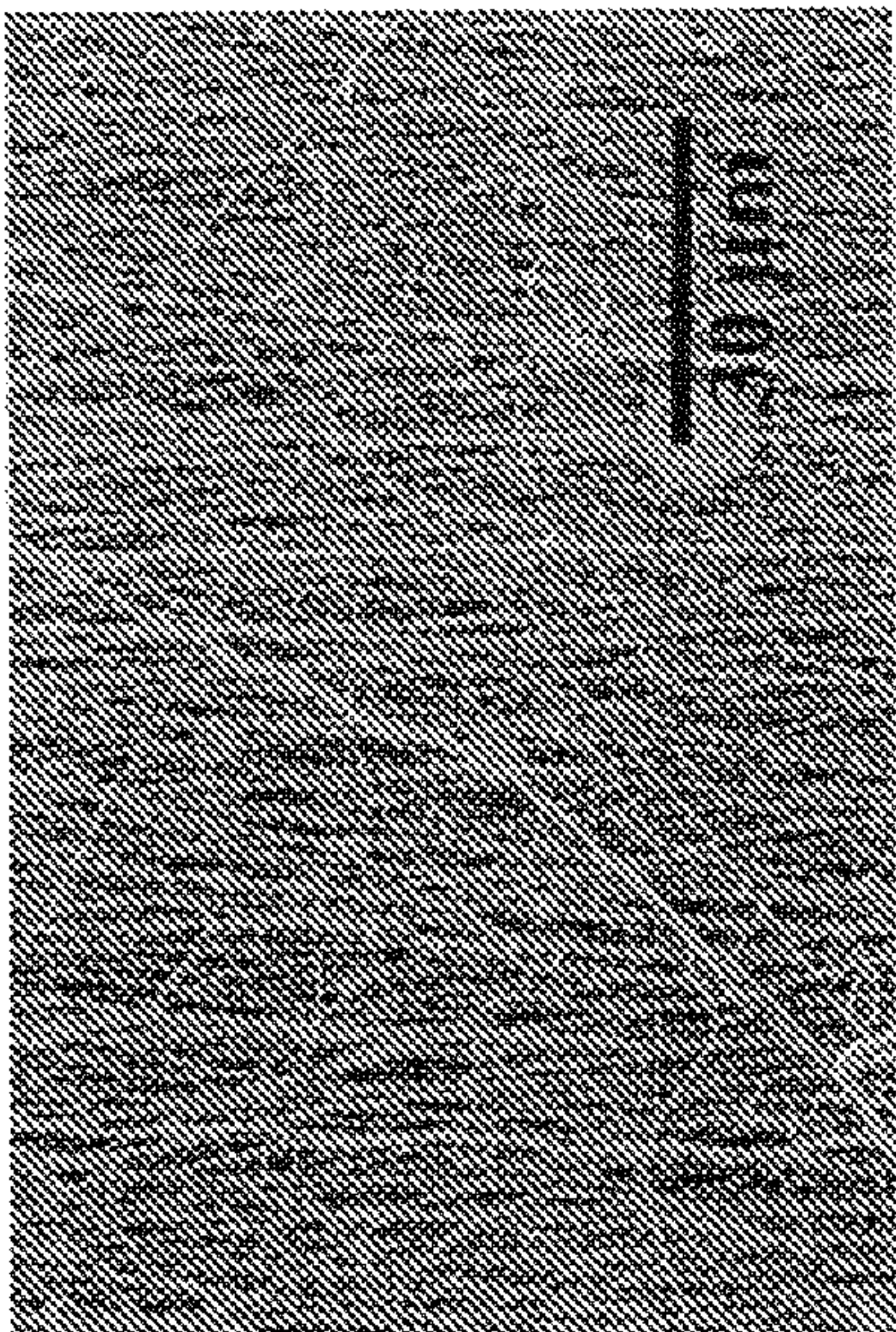


Figure 4A

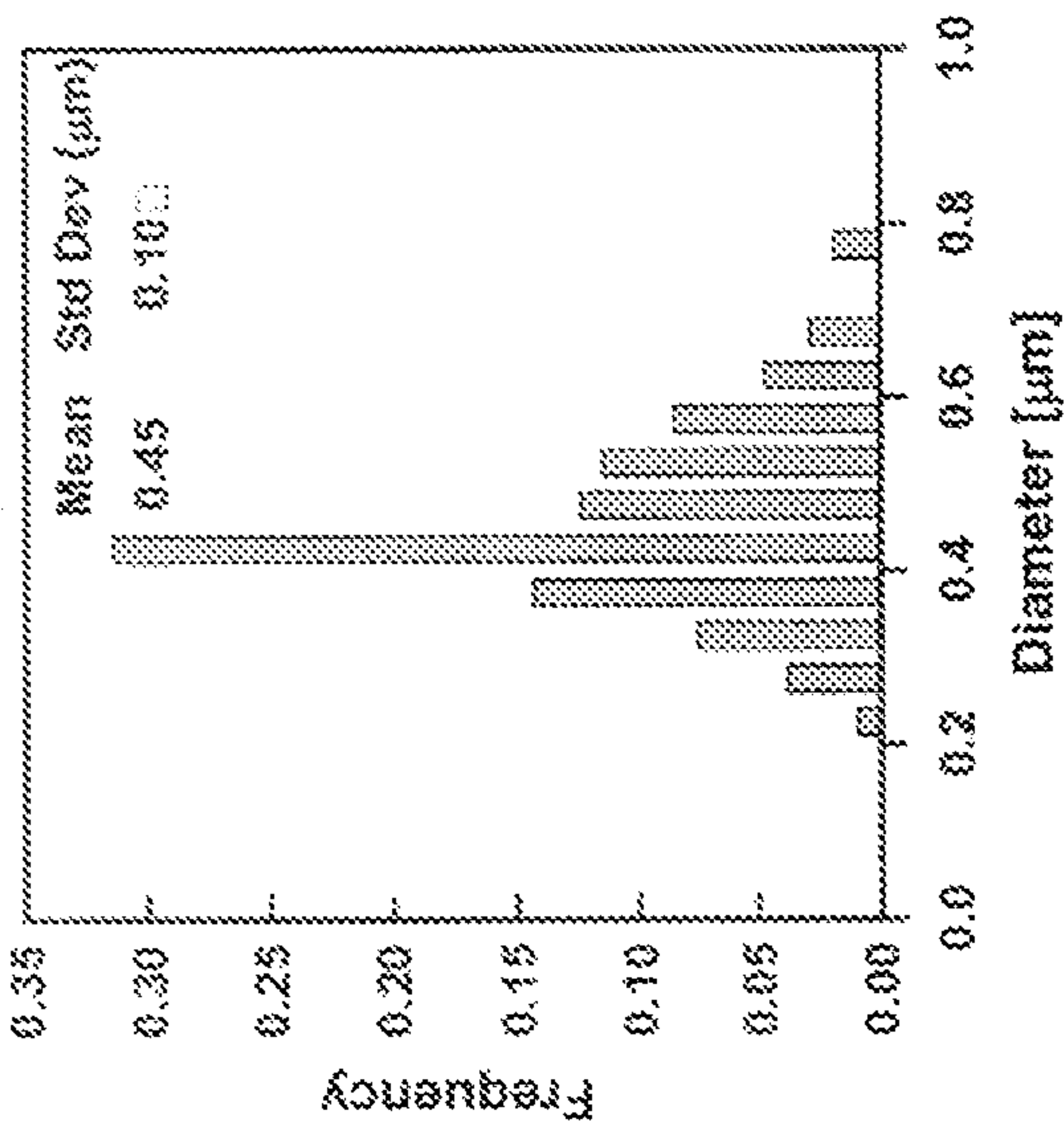


Figure 5B

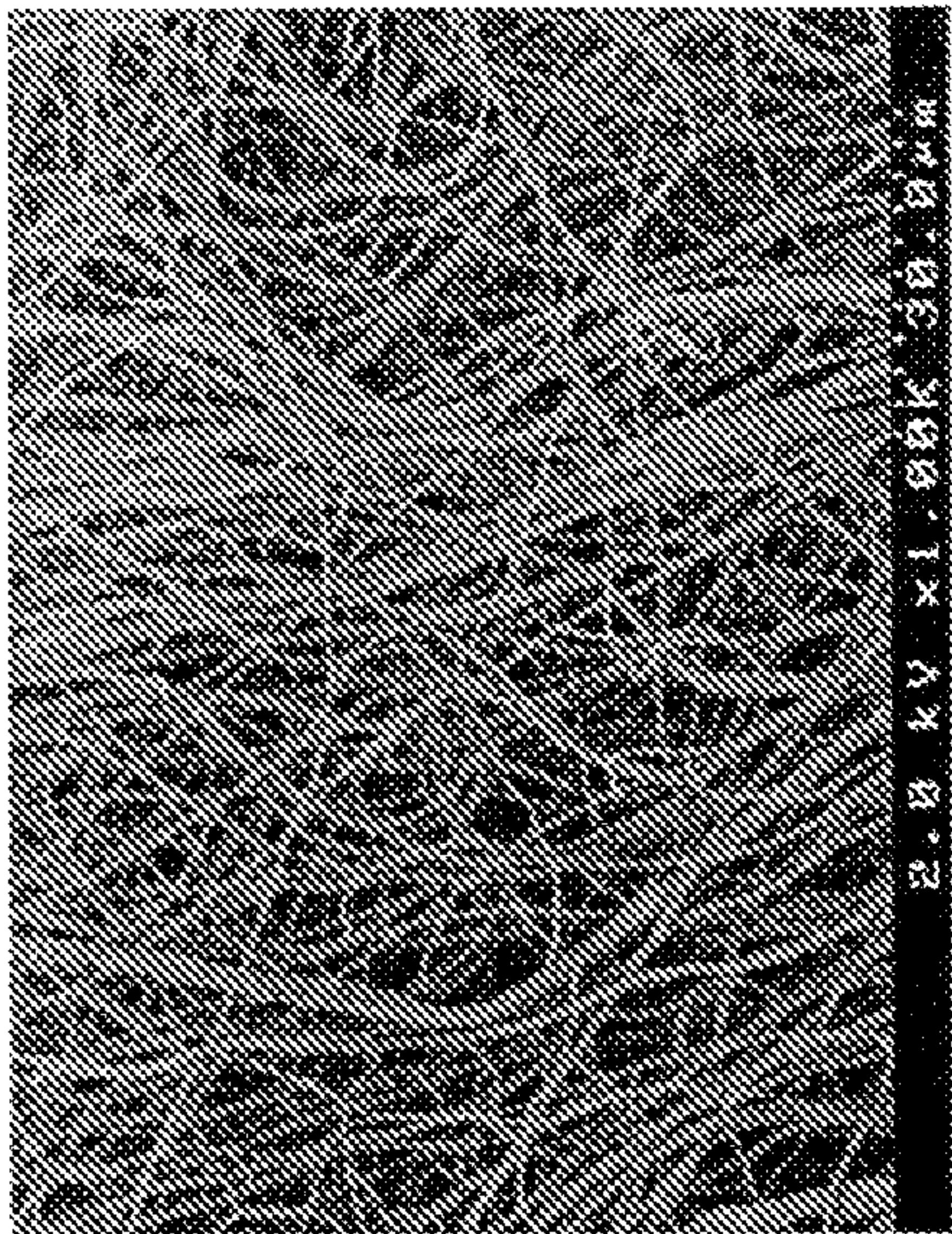


Figure 5A

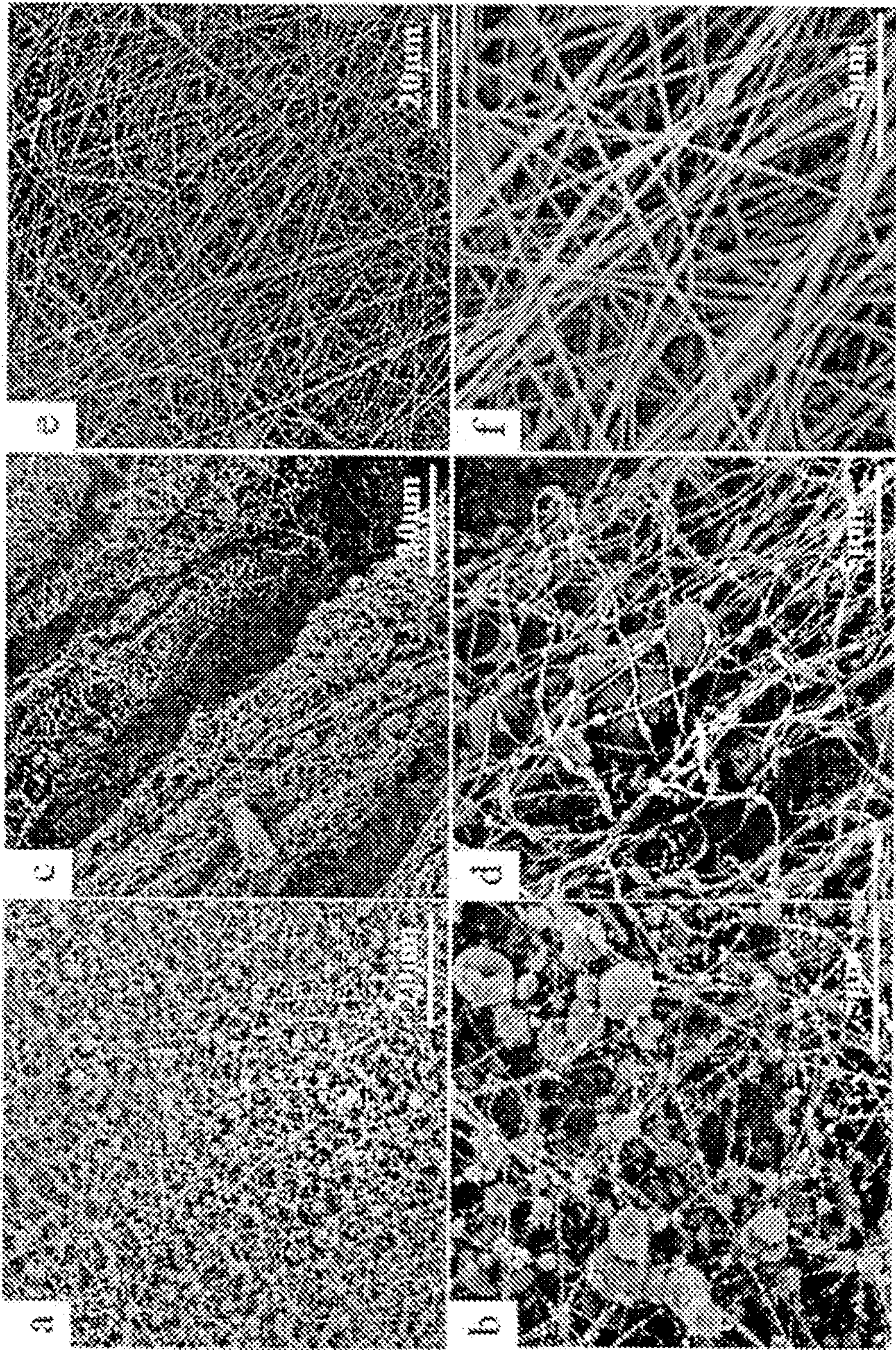


Figure 6

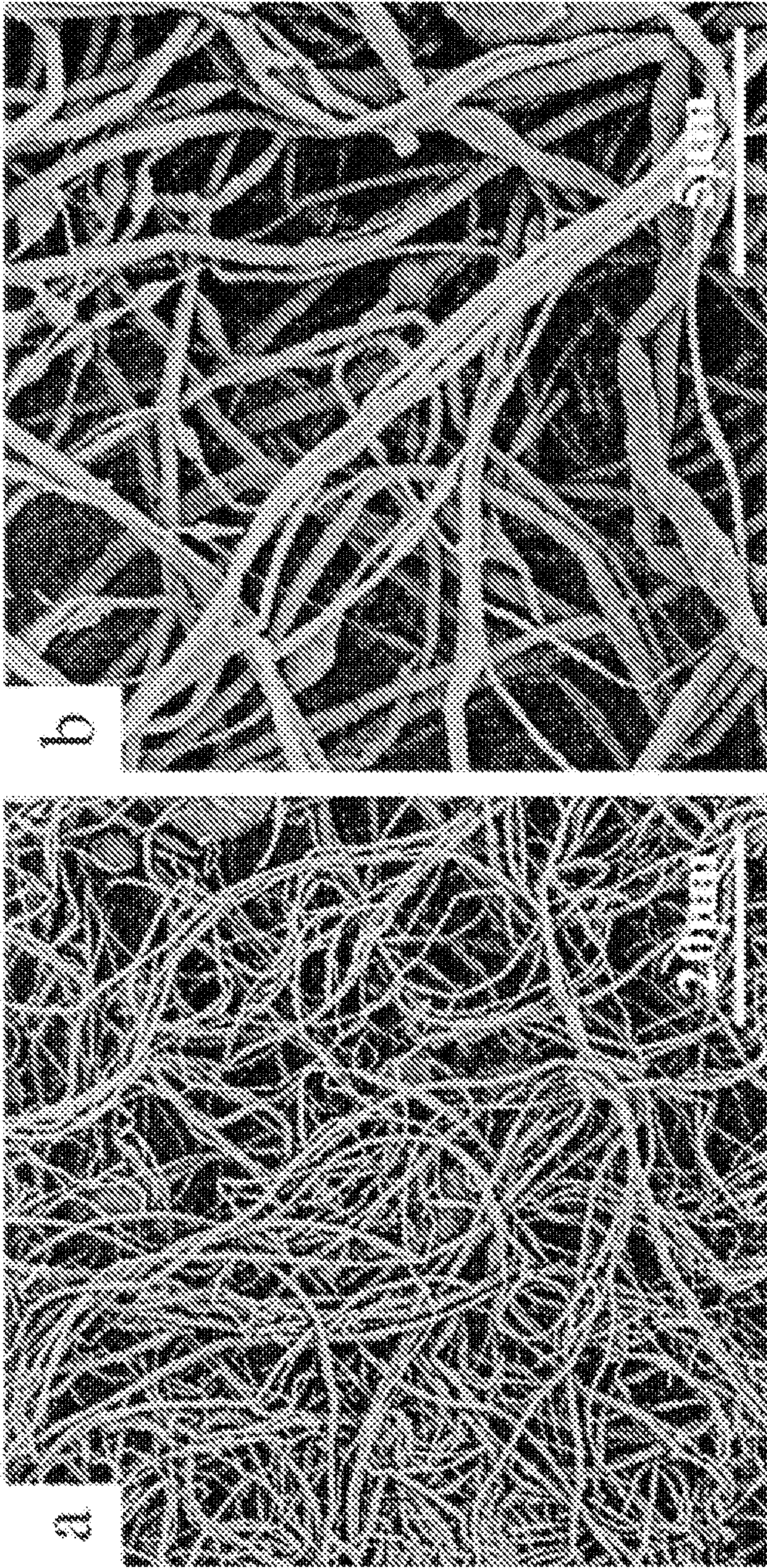
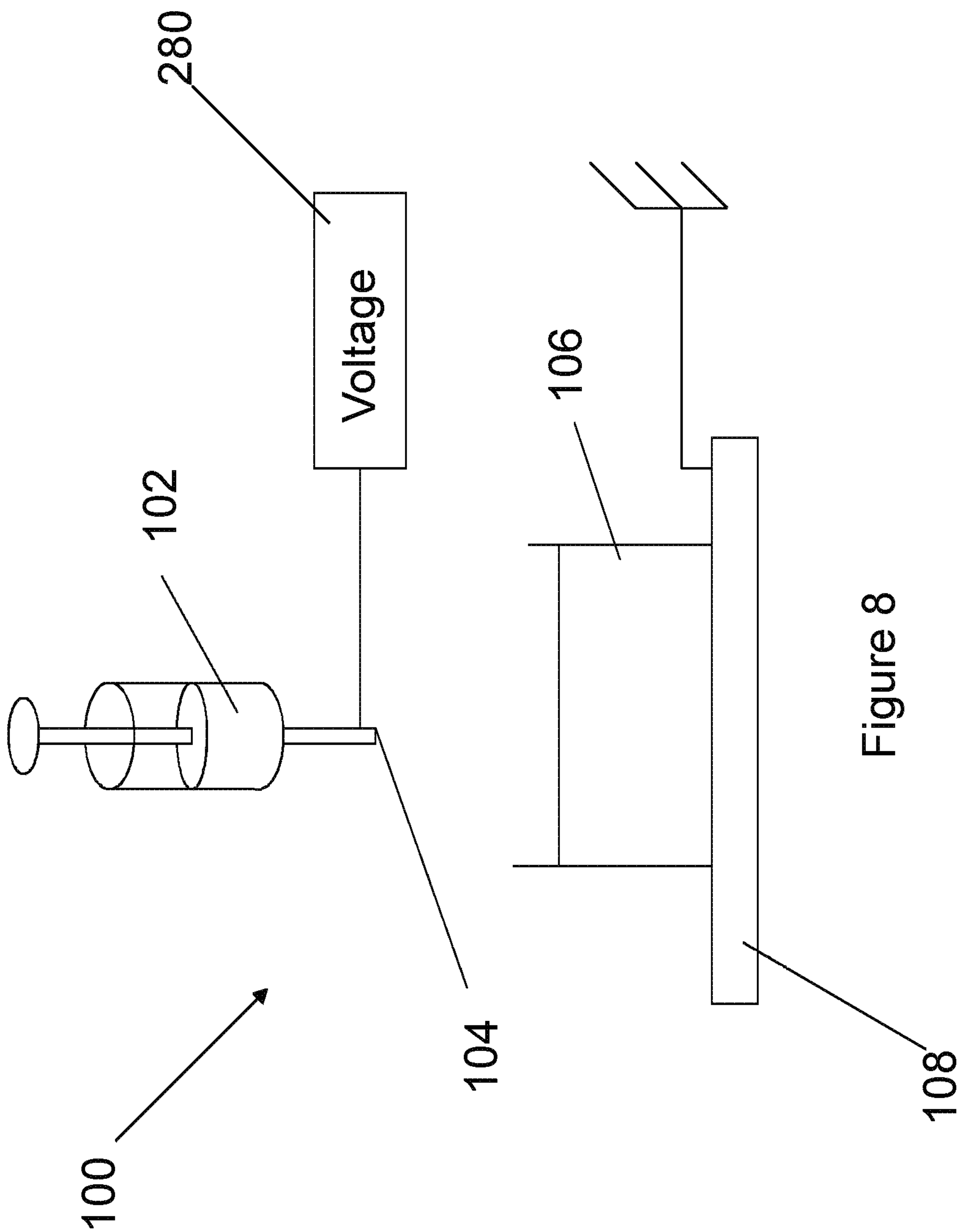


Figure 7



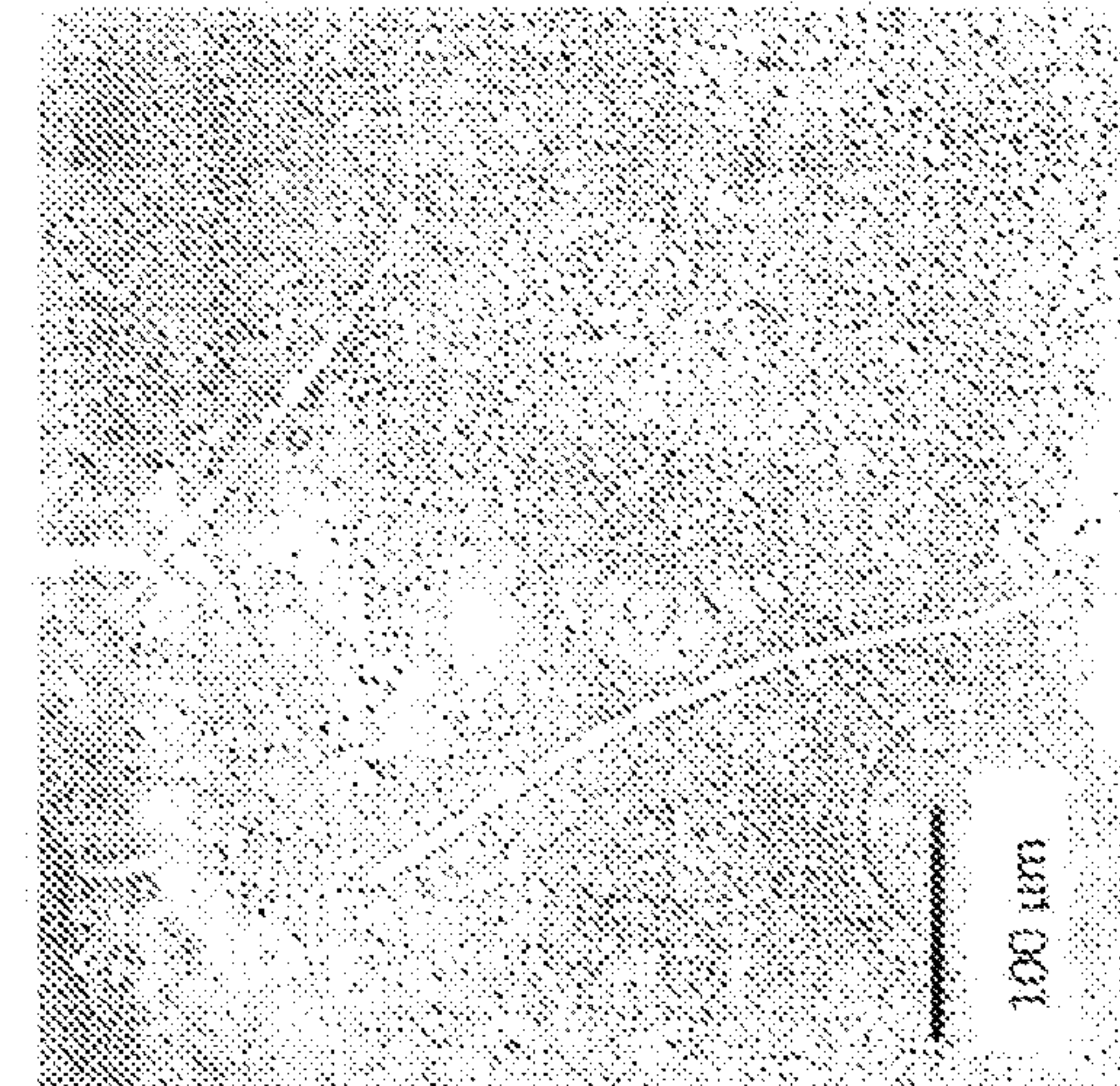


Figure 10

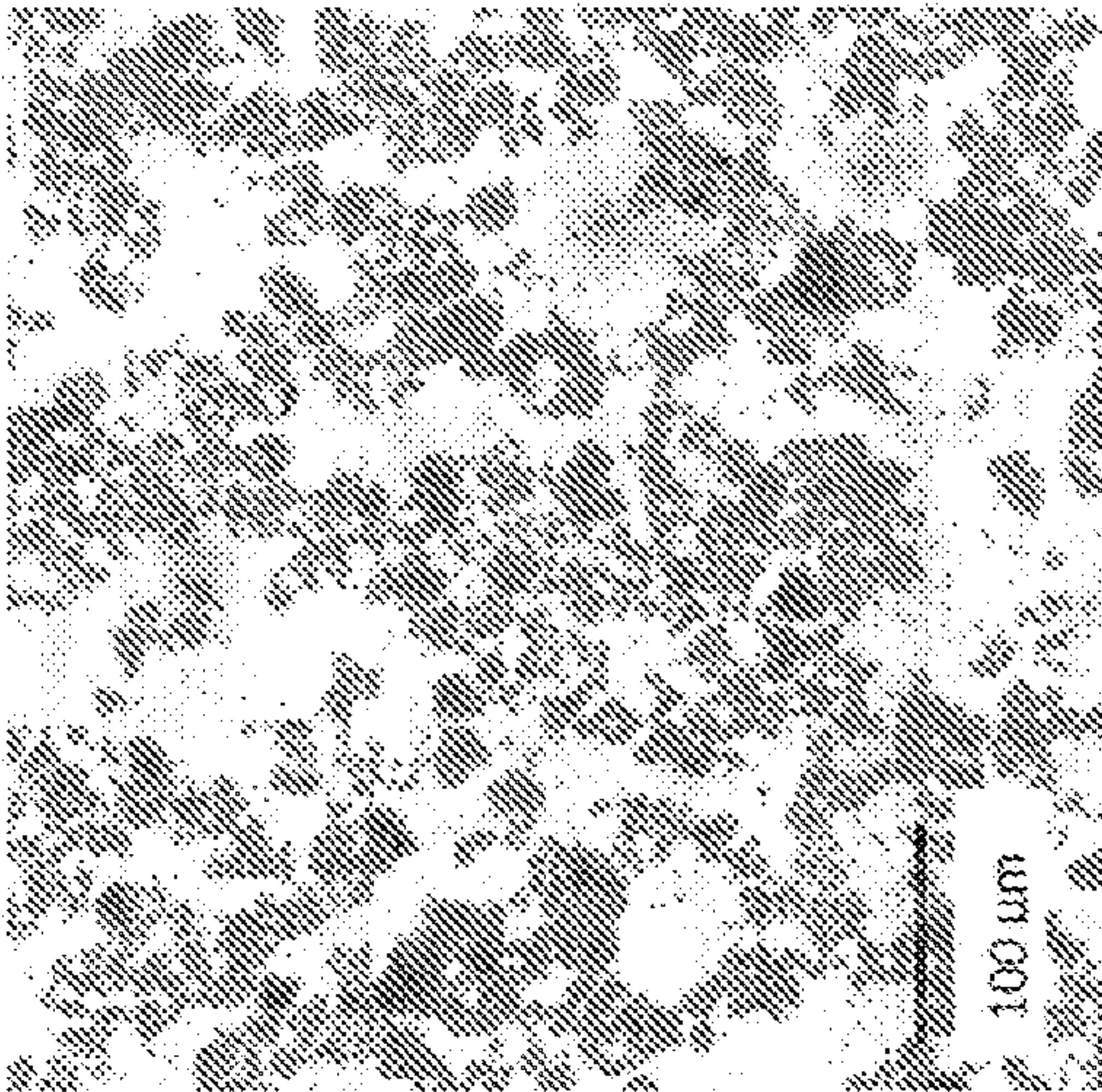


Figure 11

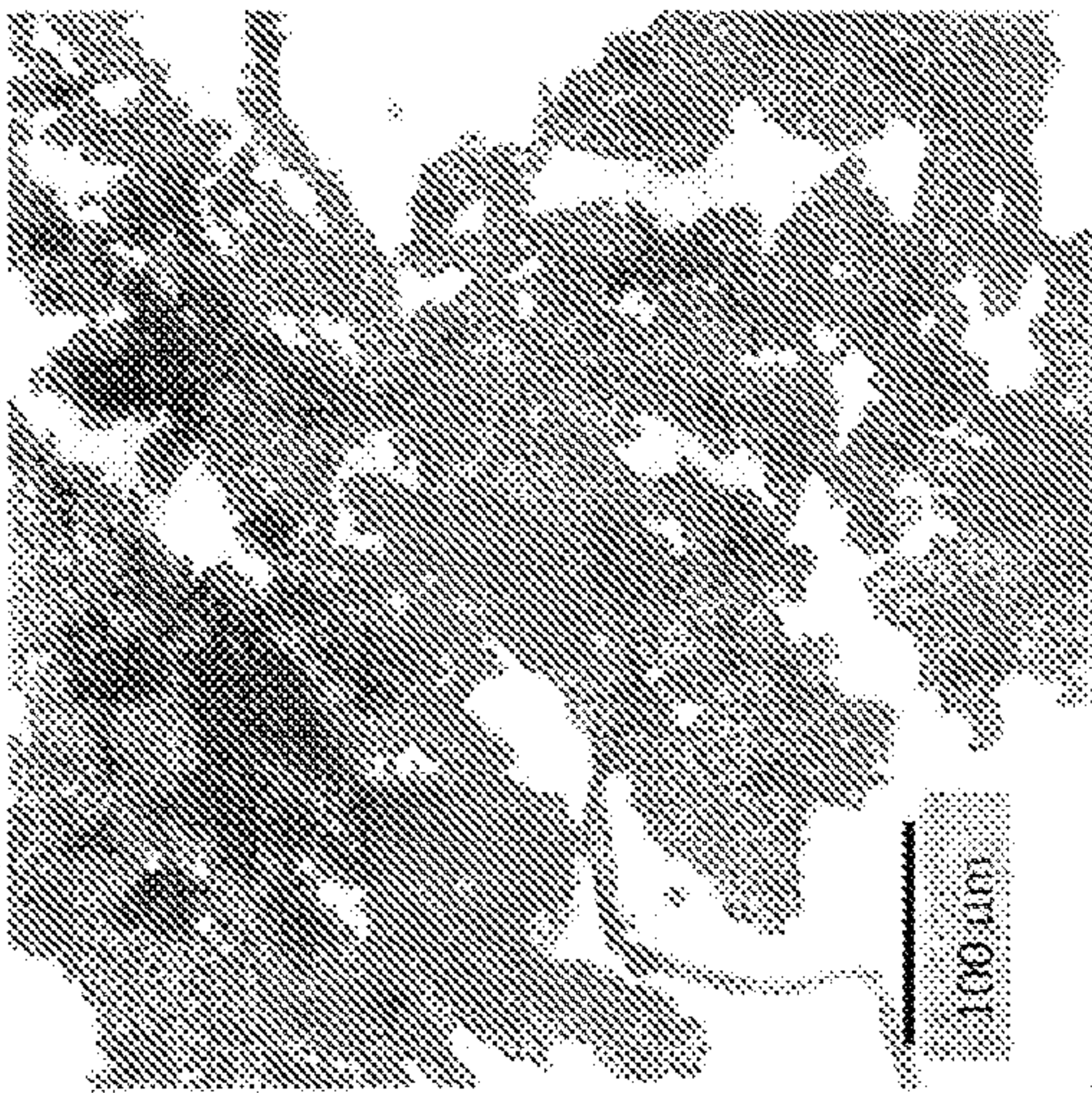


Figure 9

Figure 12A

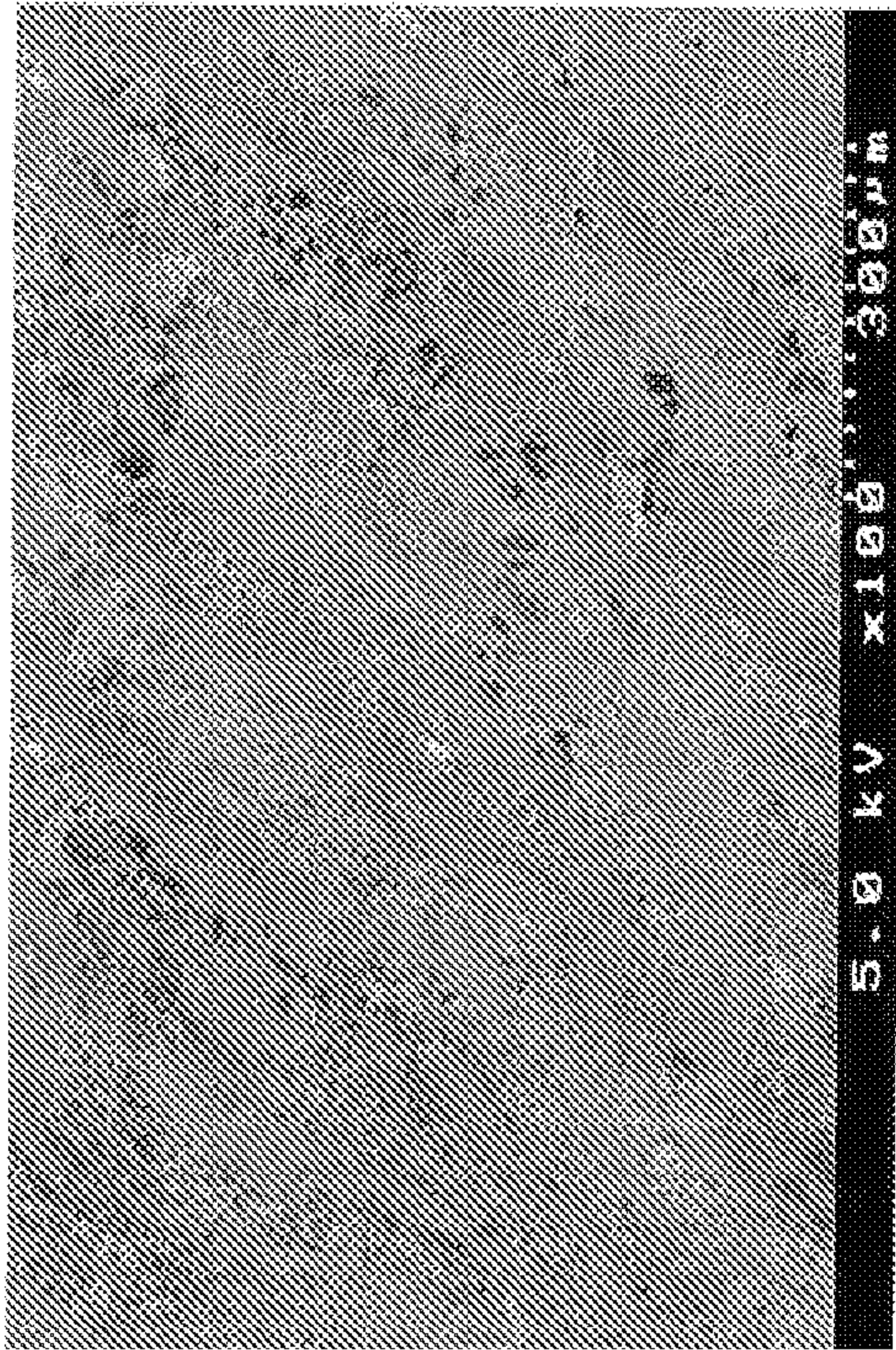
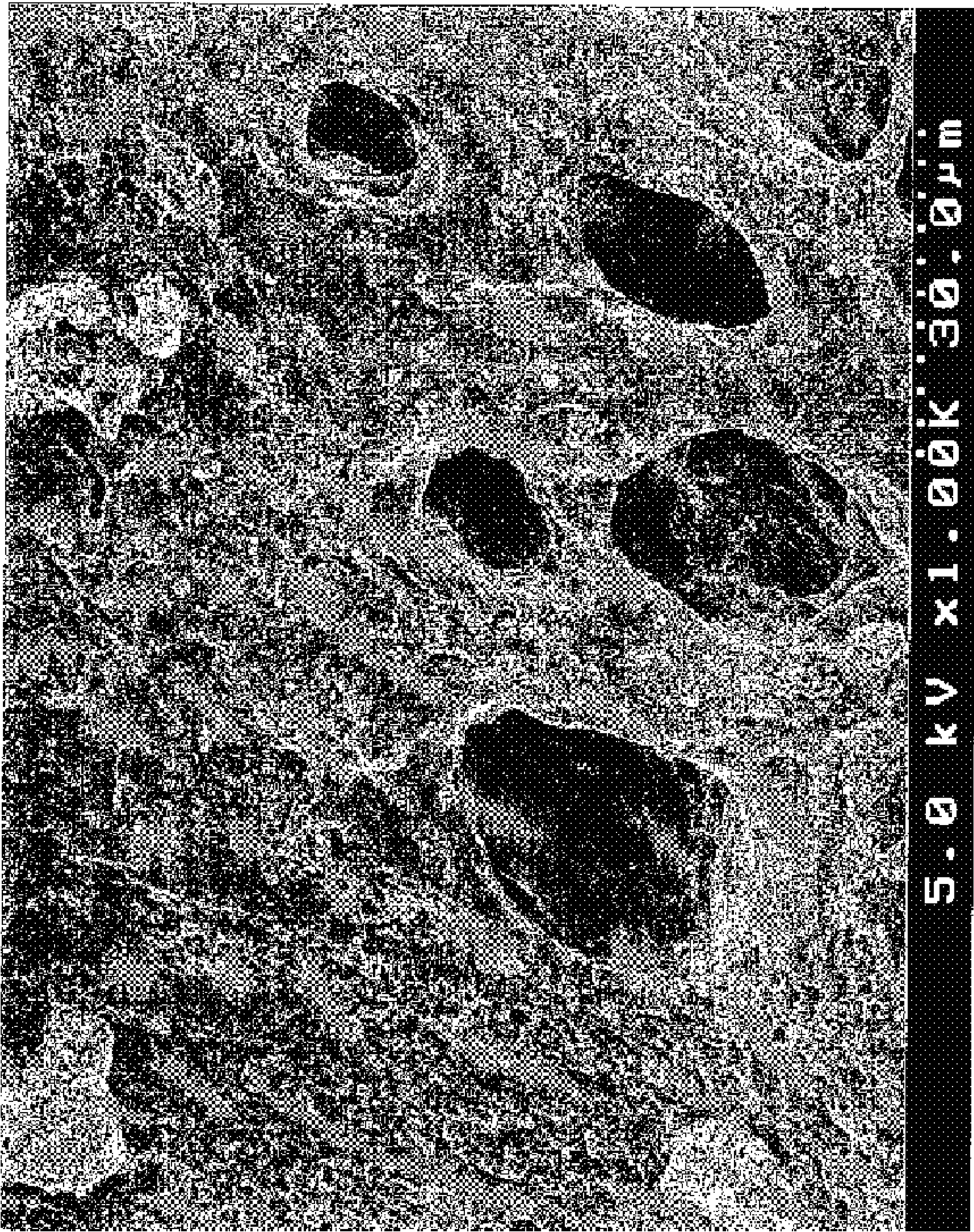


Figure 12B



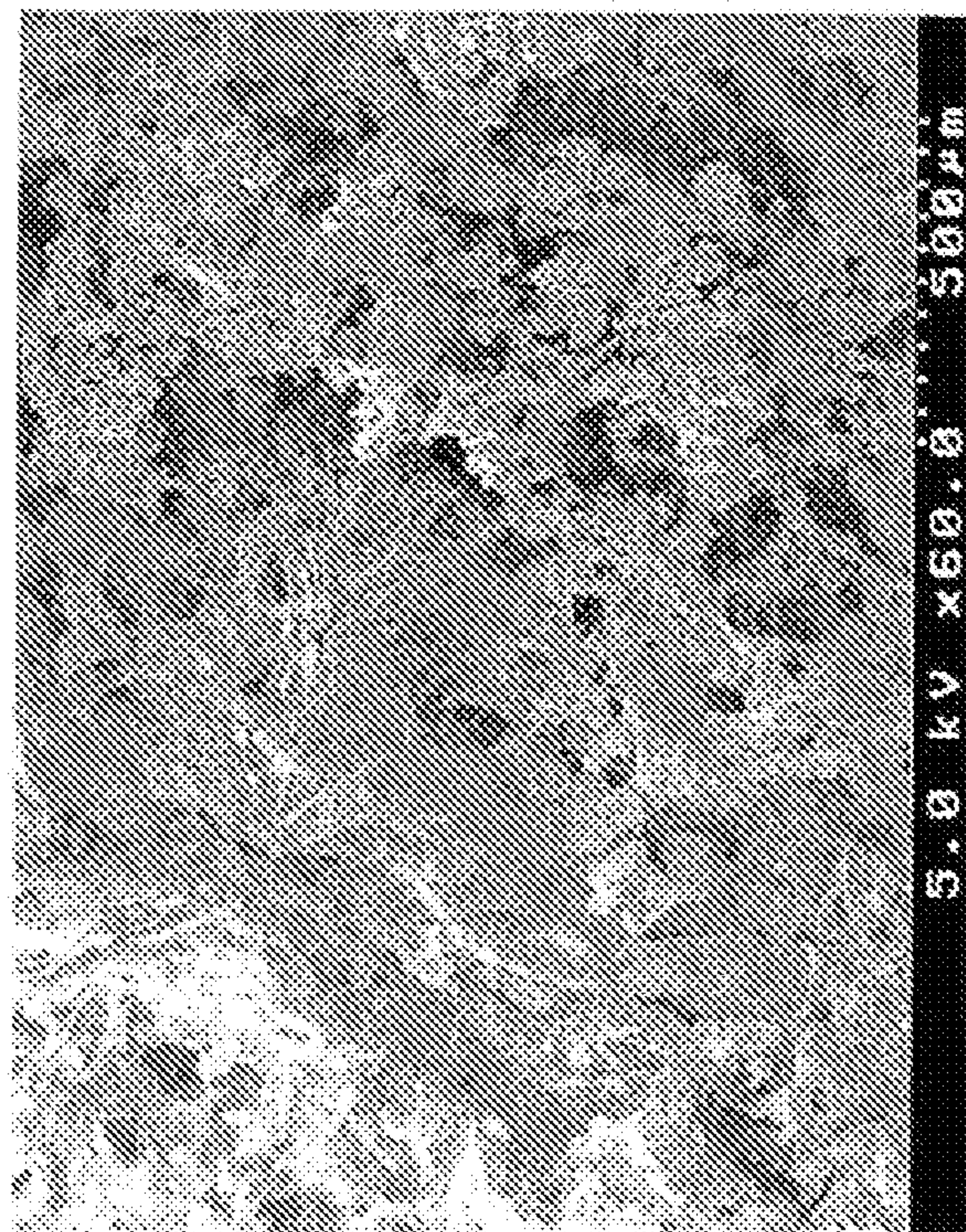


Figure 13B

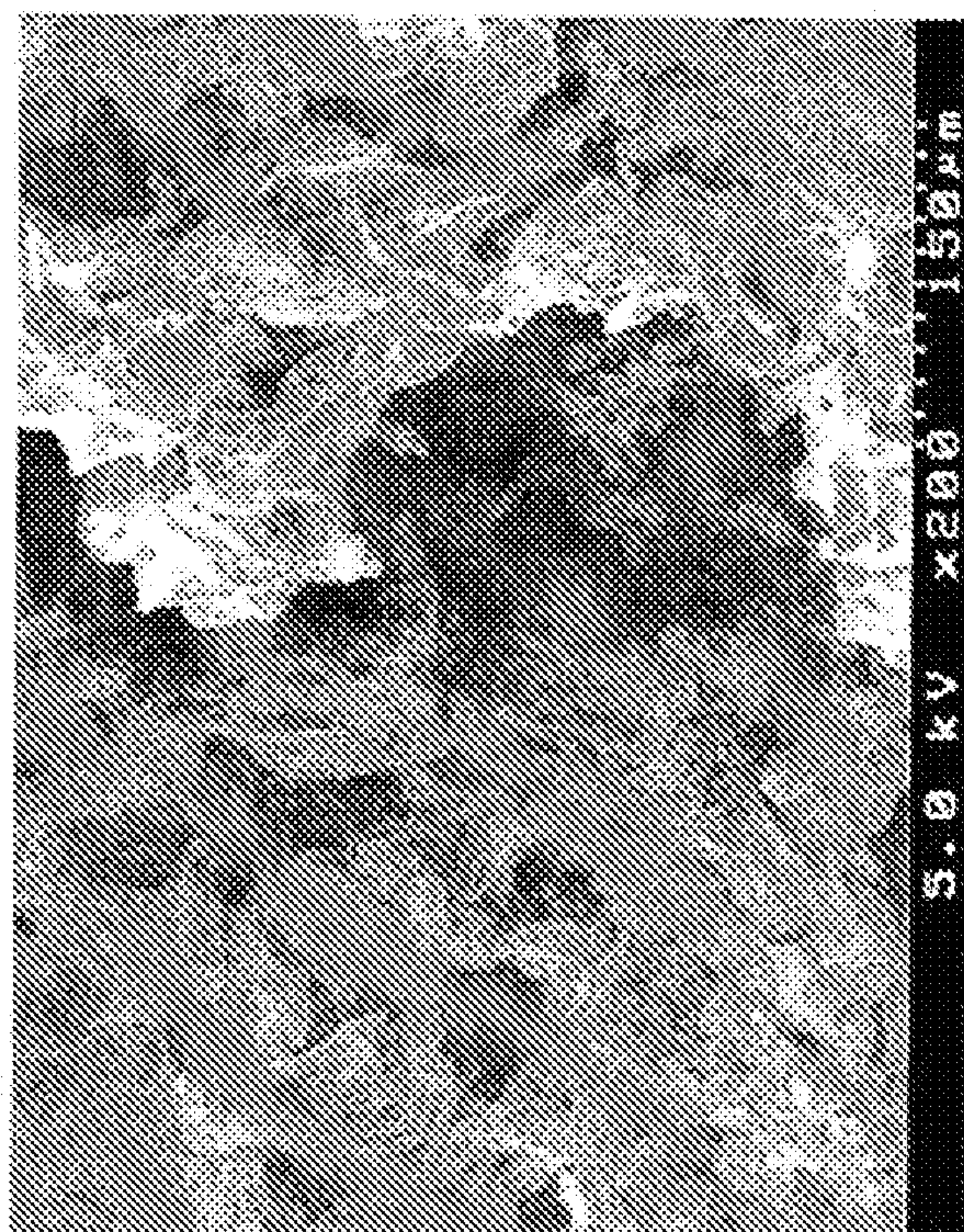


Figure 13A

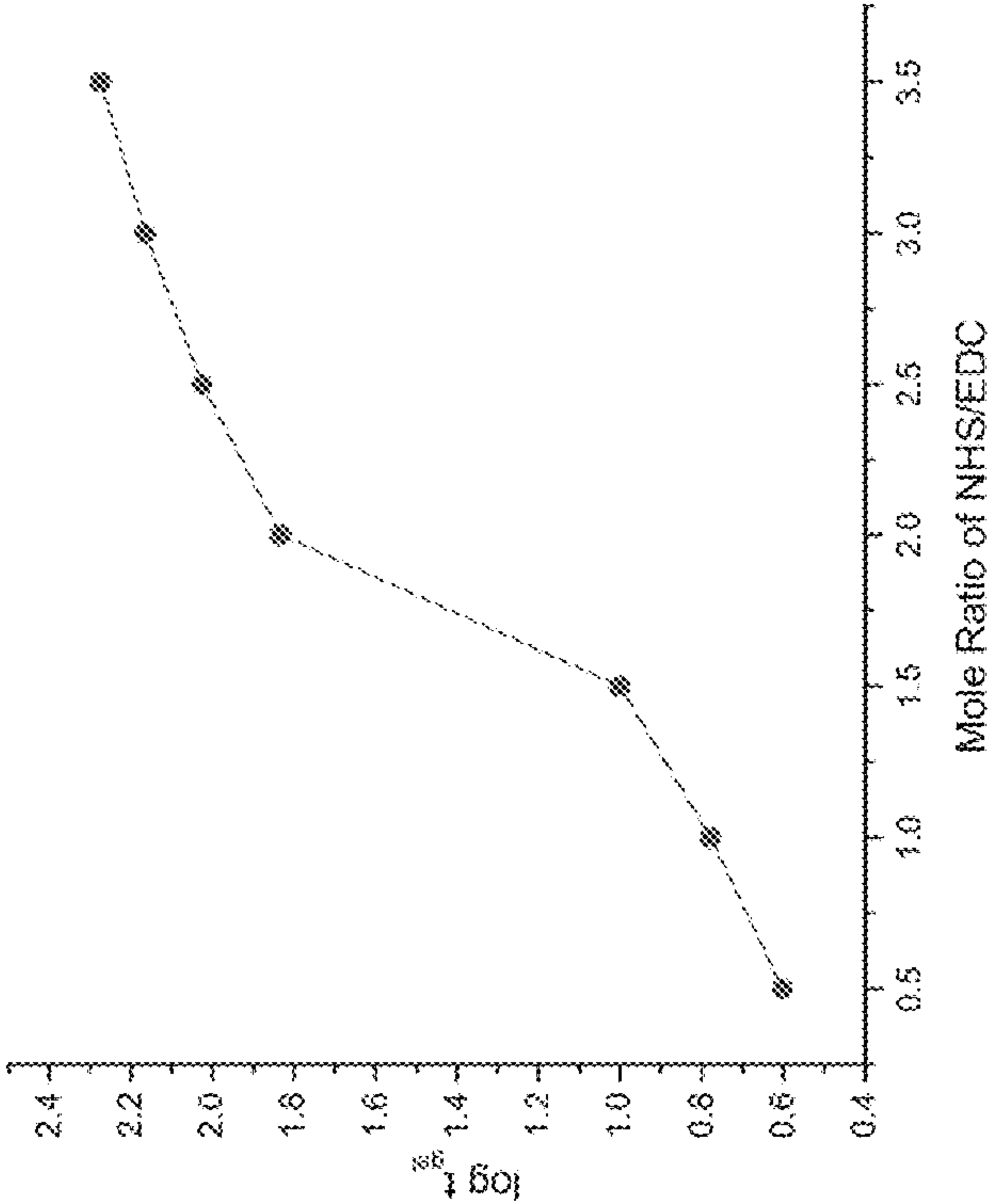


Figure 14A

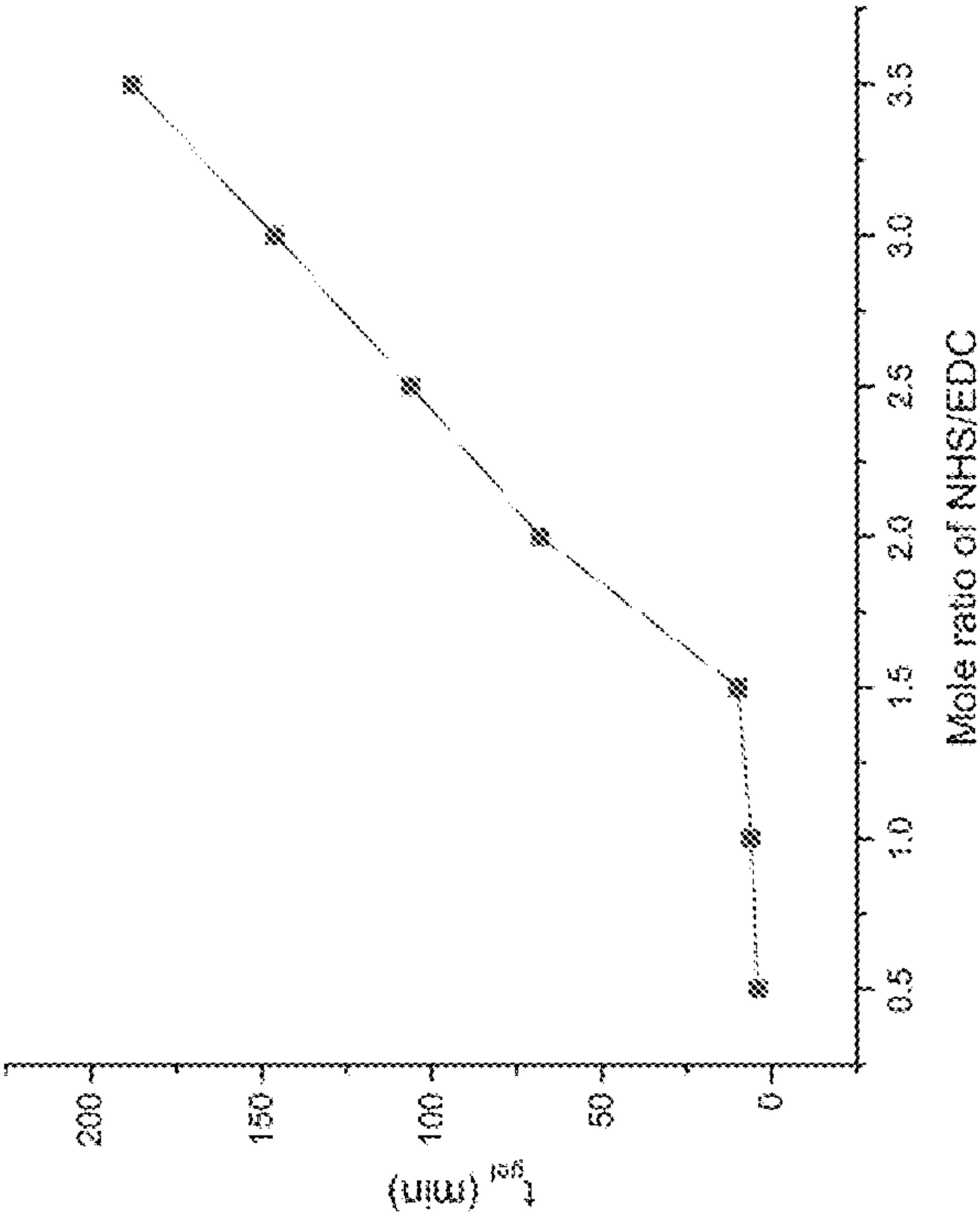


Figure 14B



Figure 15A



Figure 15B

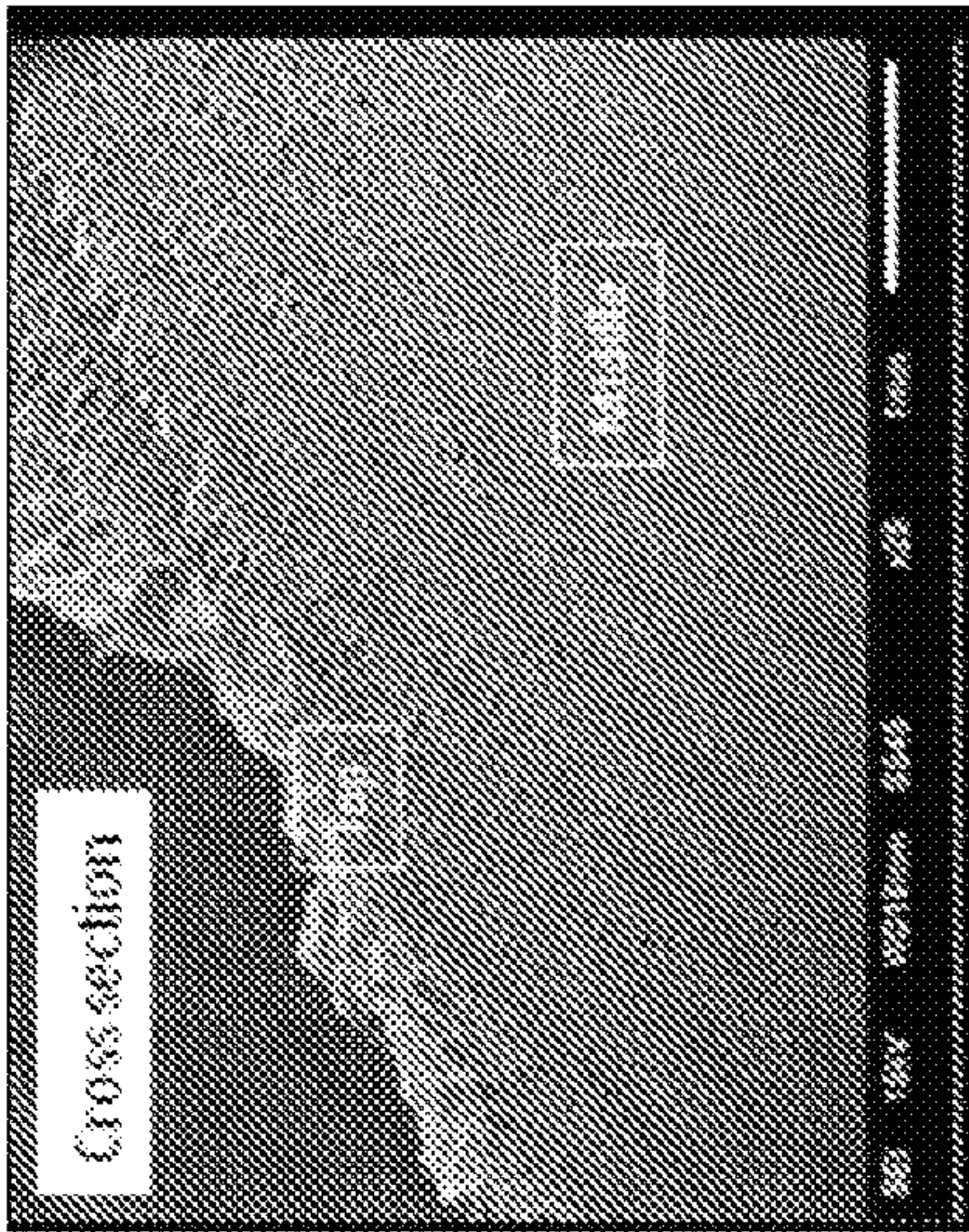


Figure 16A

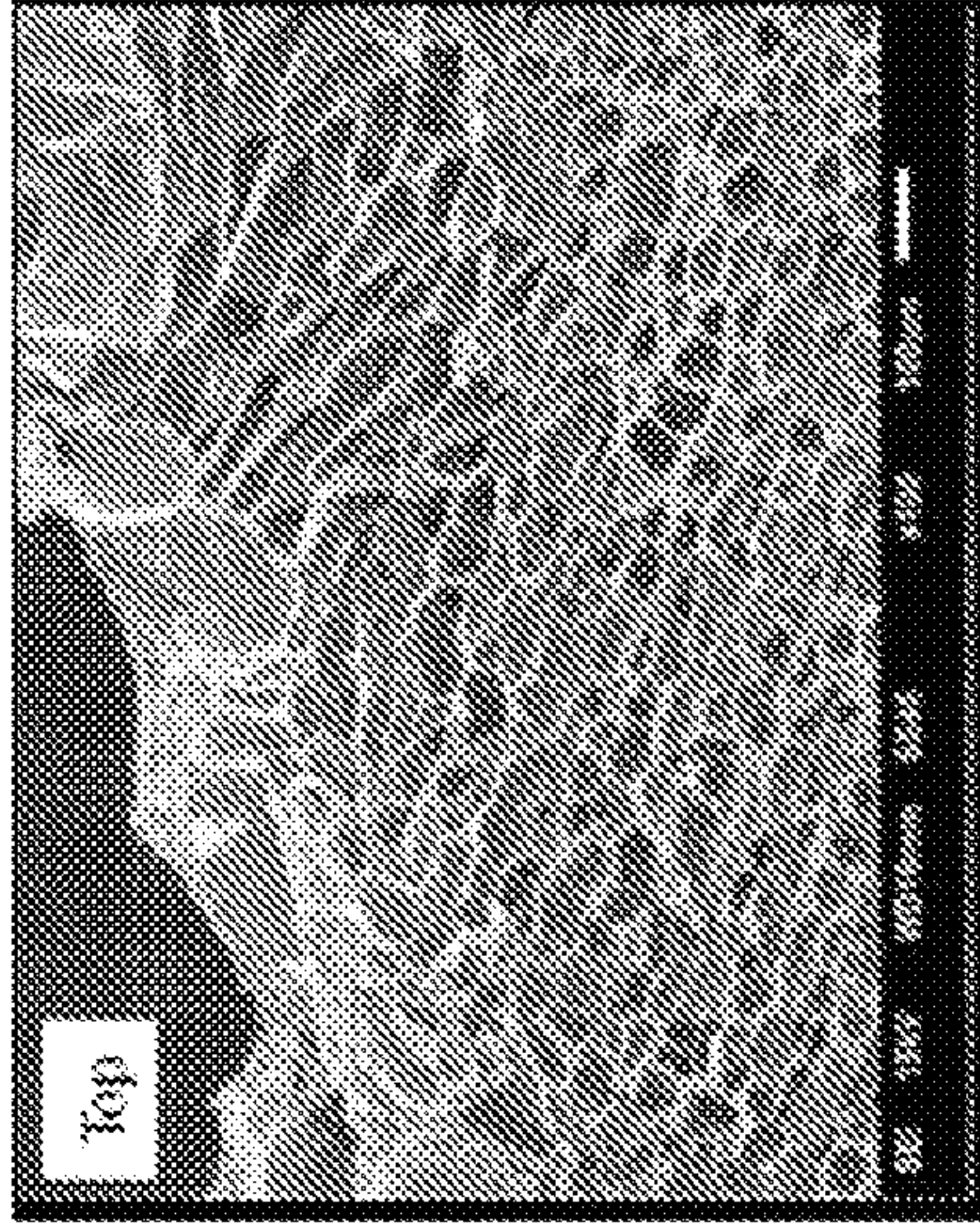


Figure 16B

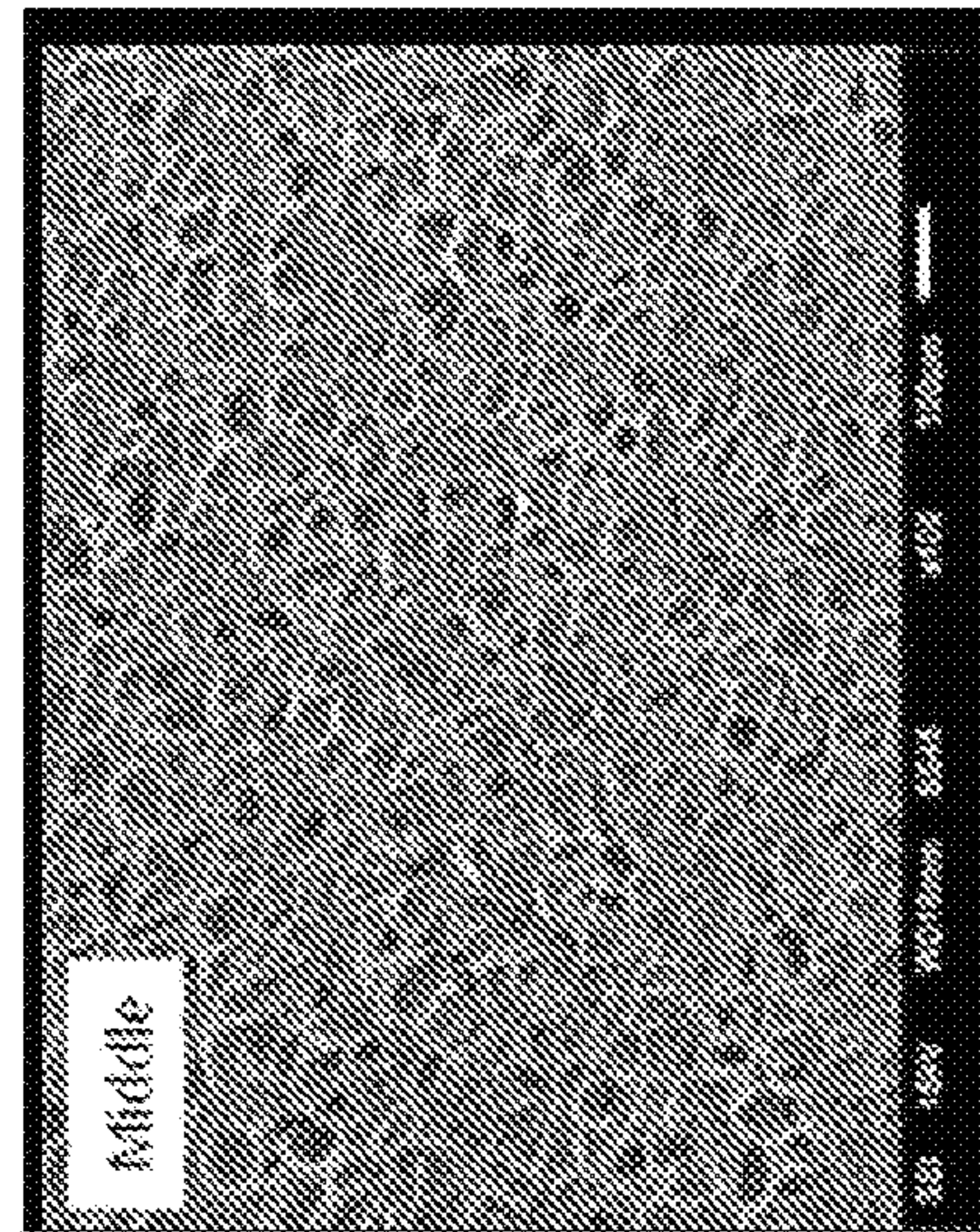


Figure 16C

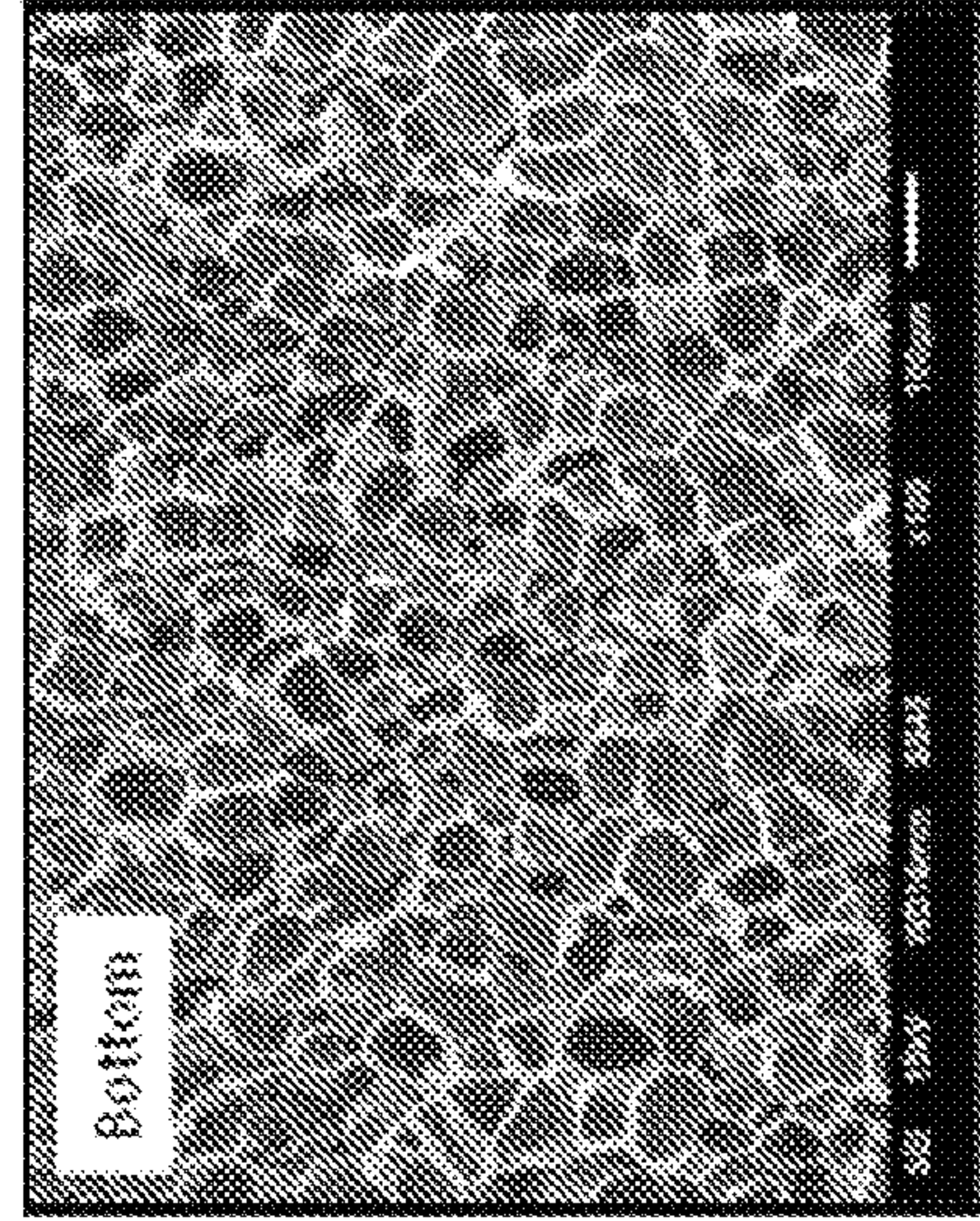


Figure 16D

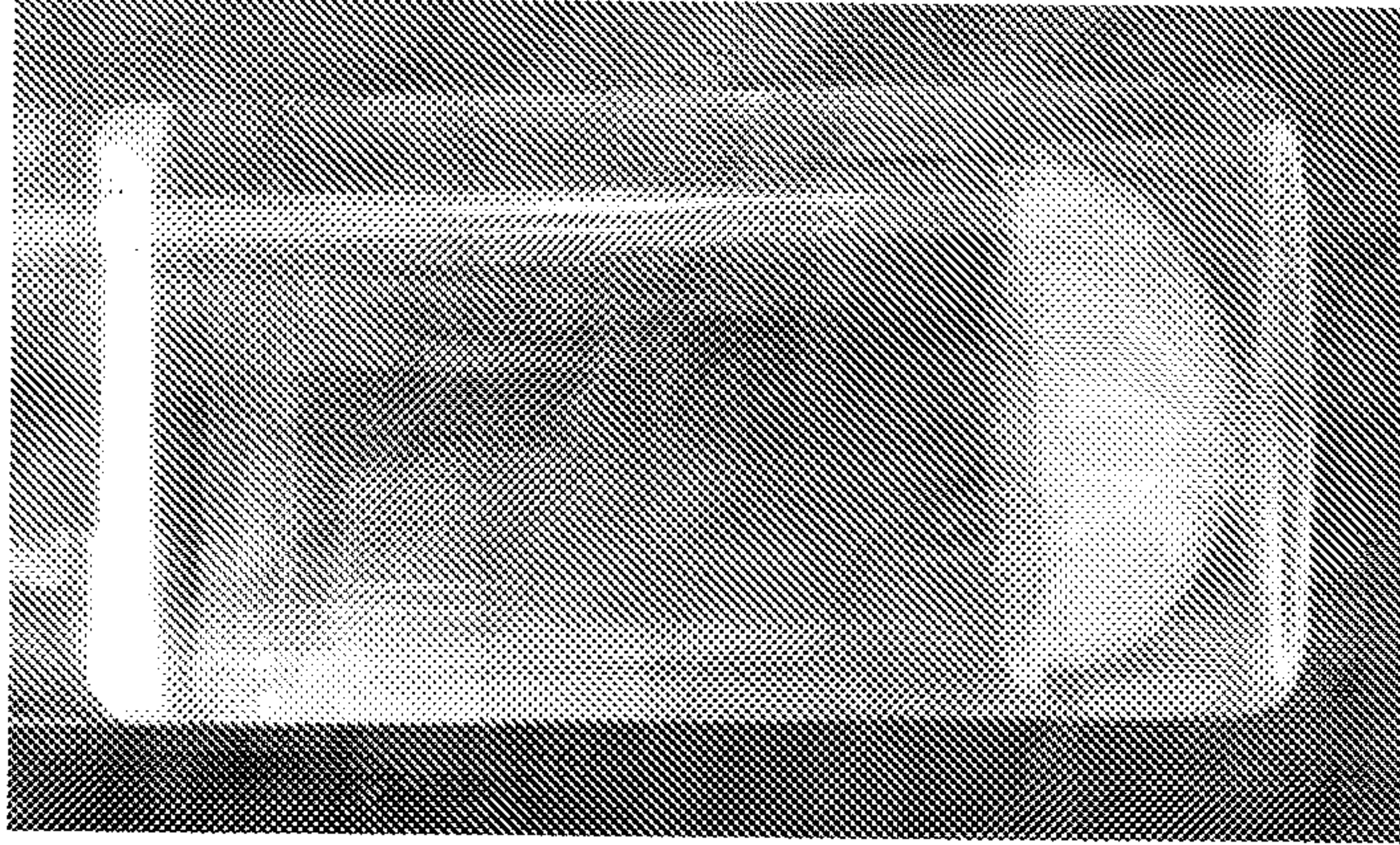


Figure 17B



Figure 17A

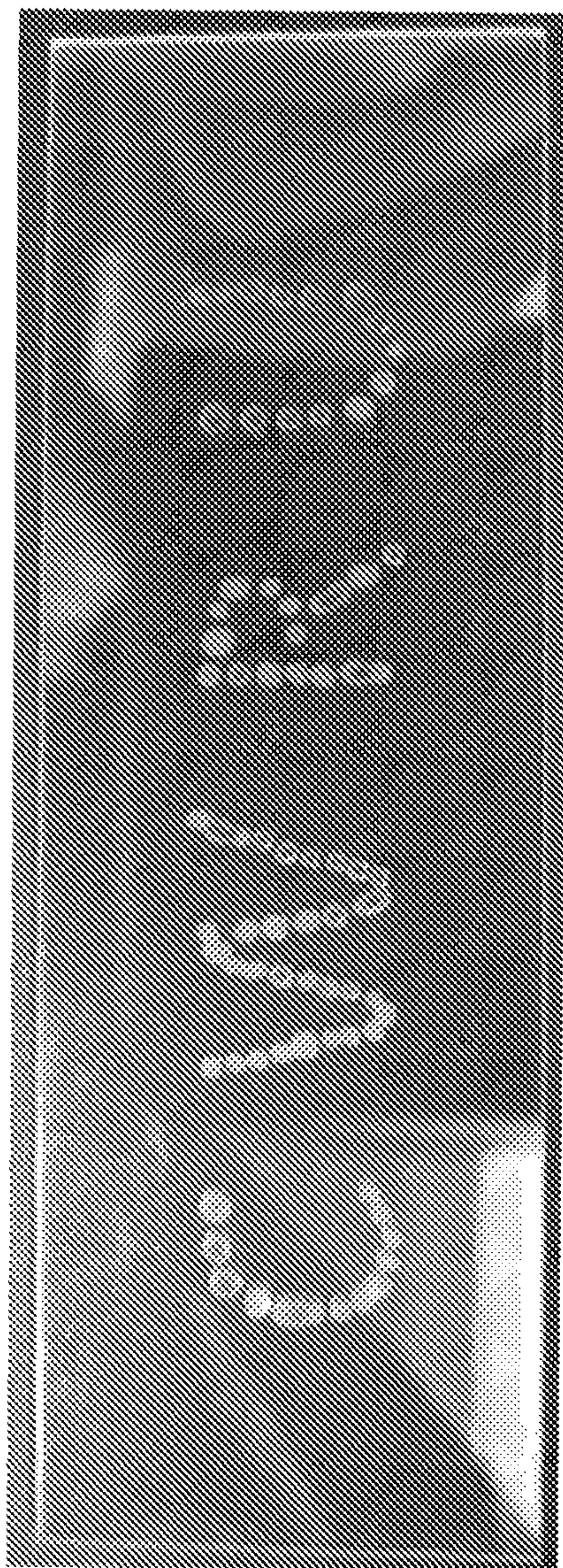


Figure 18

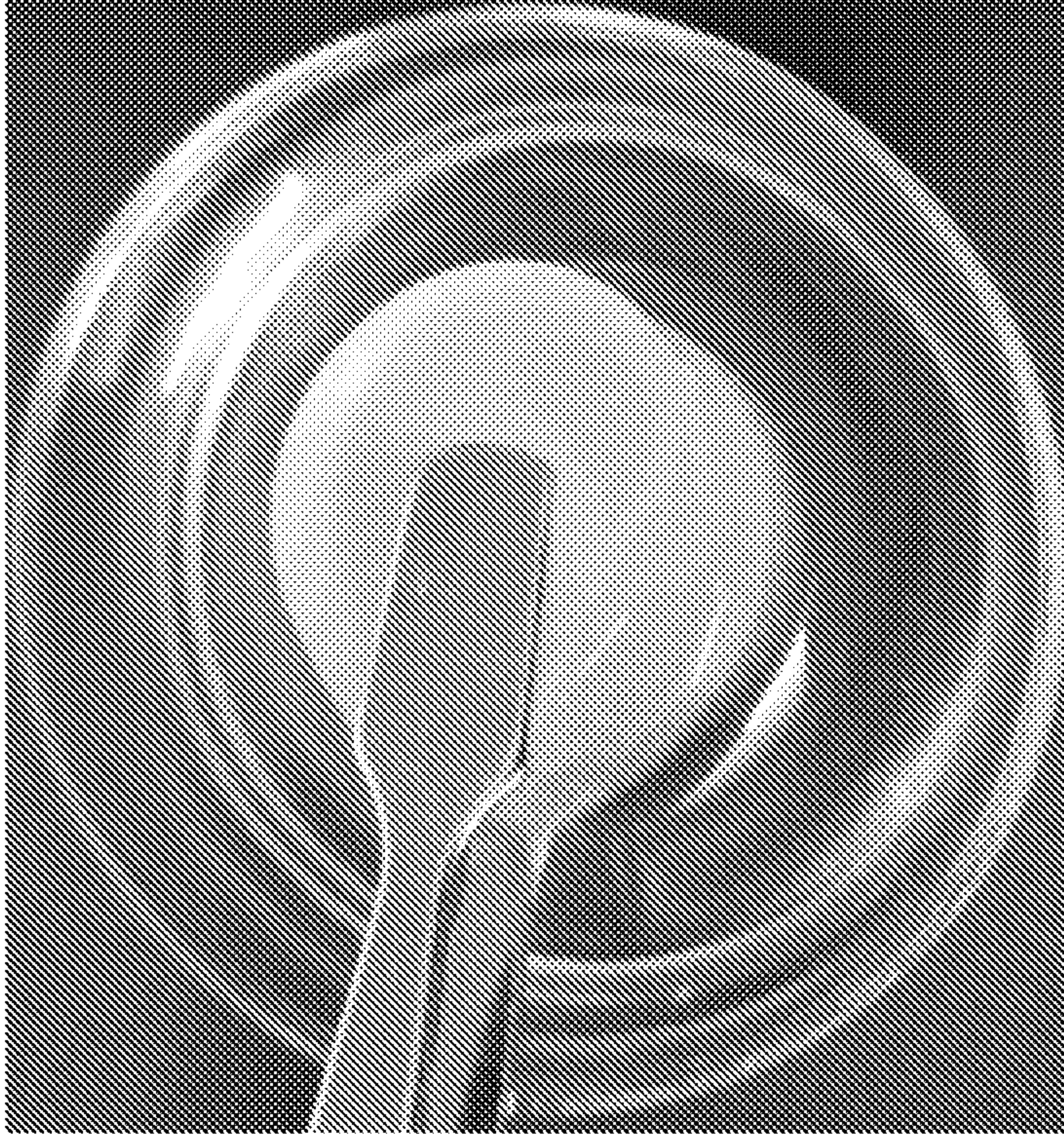


Figure 19A



Figure 19B

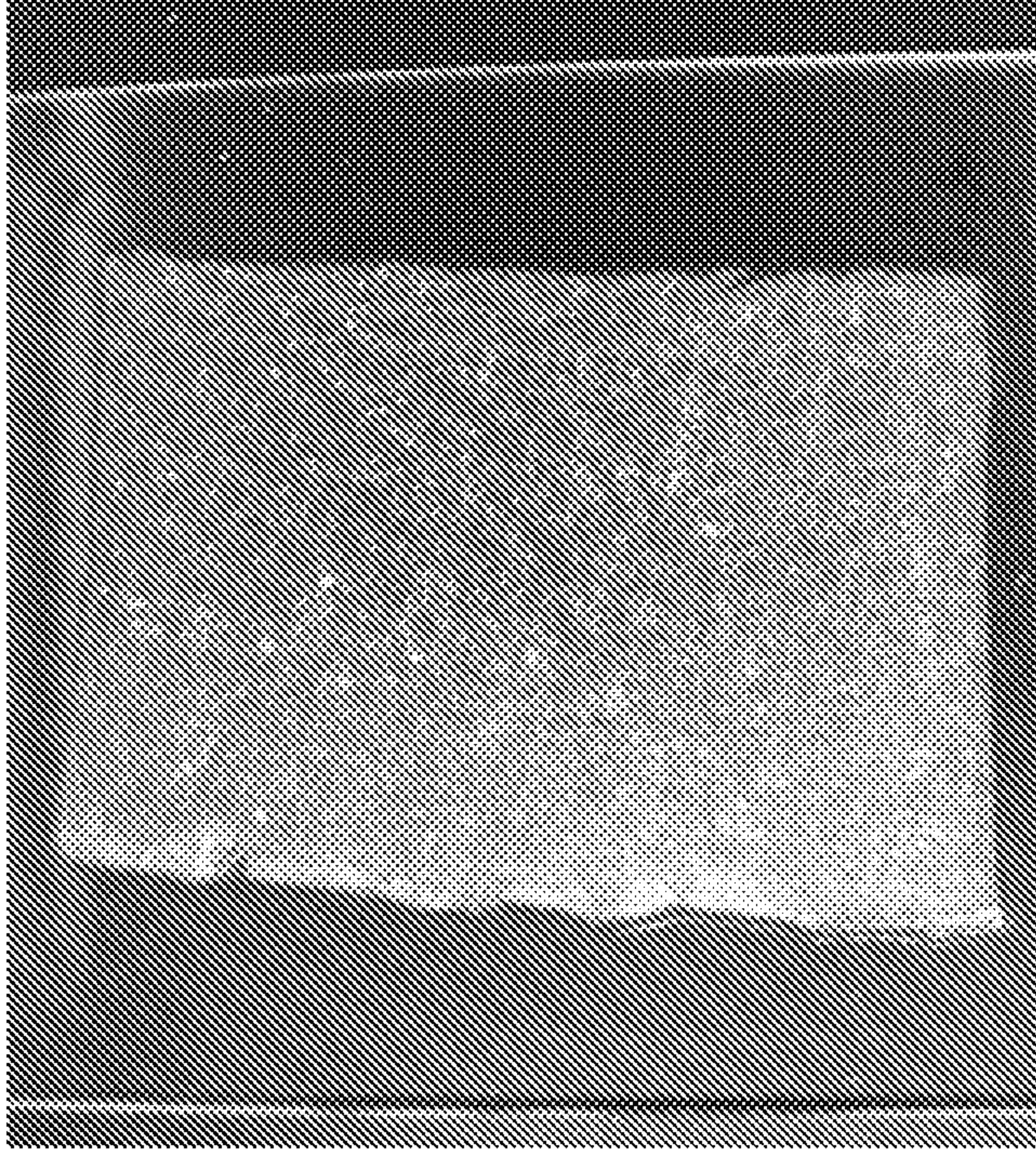


Figure 20B

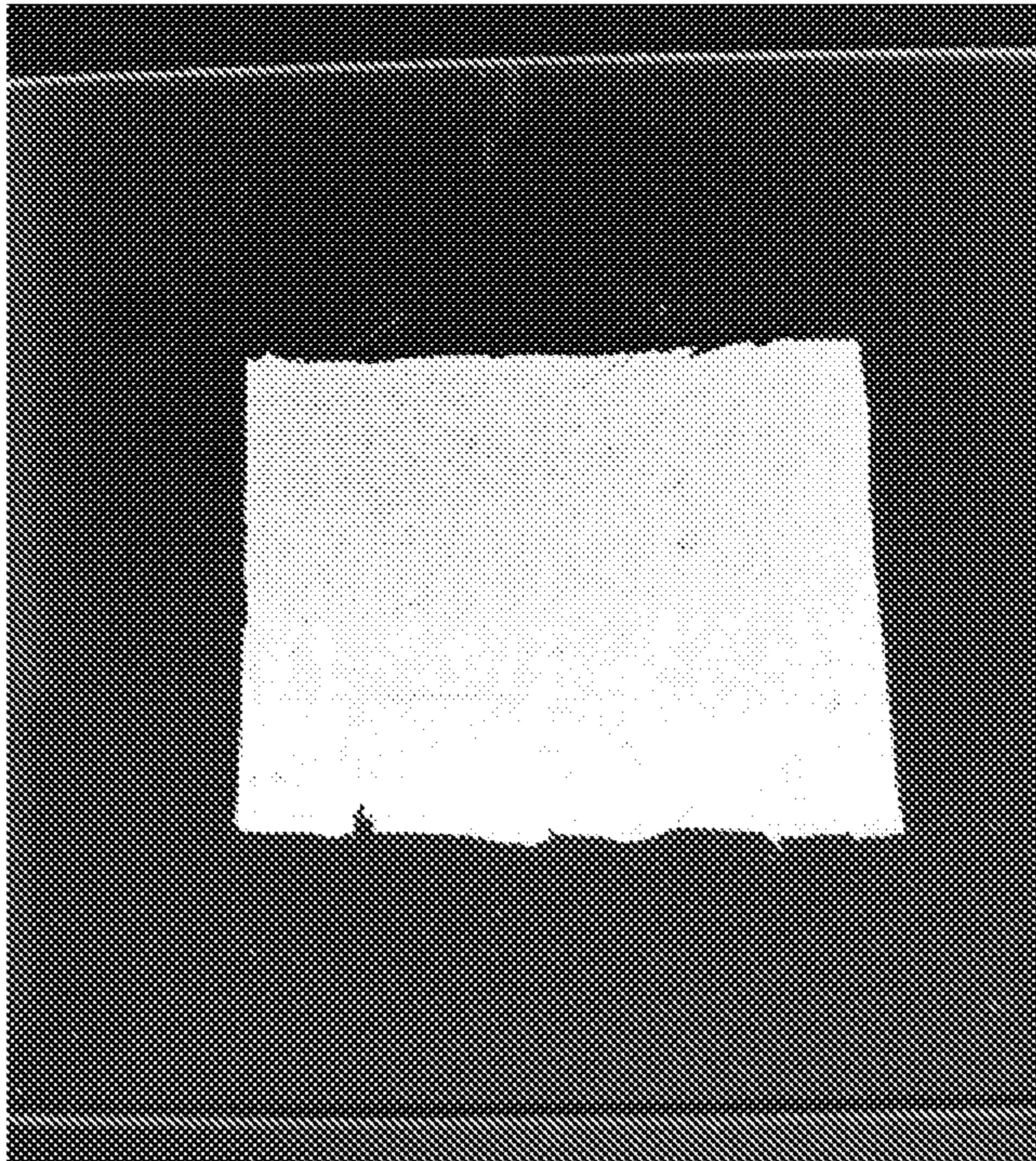


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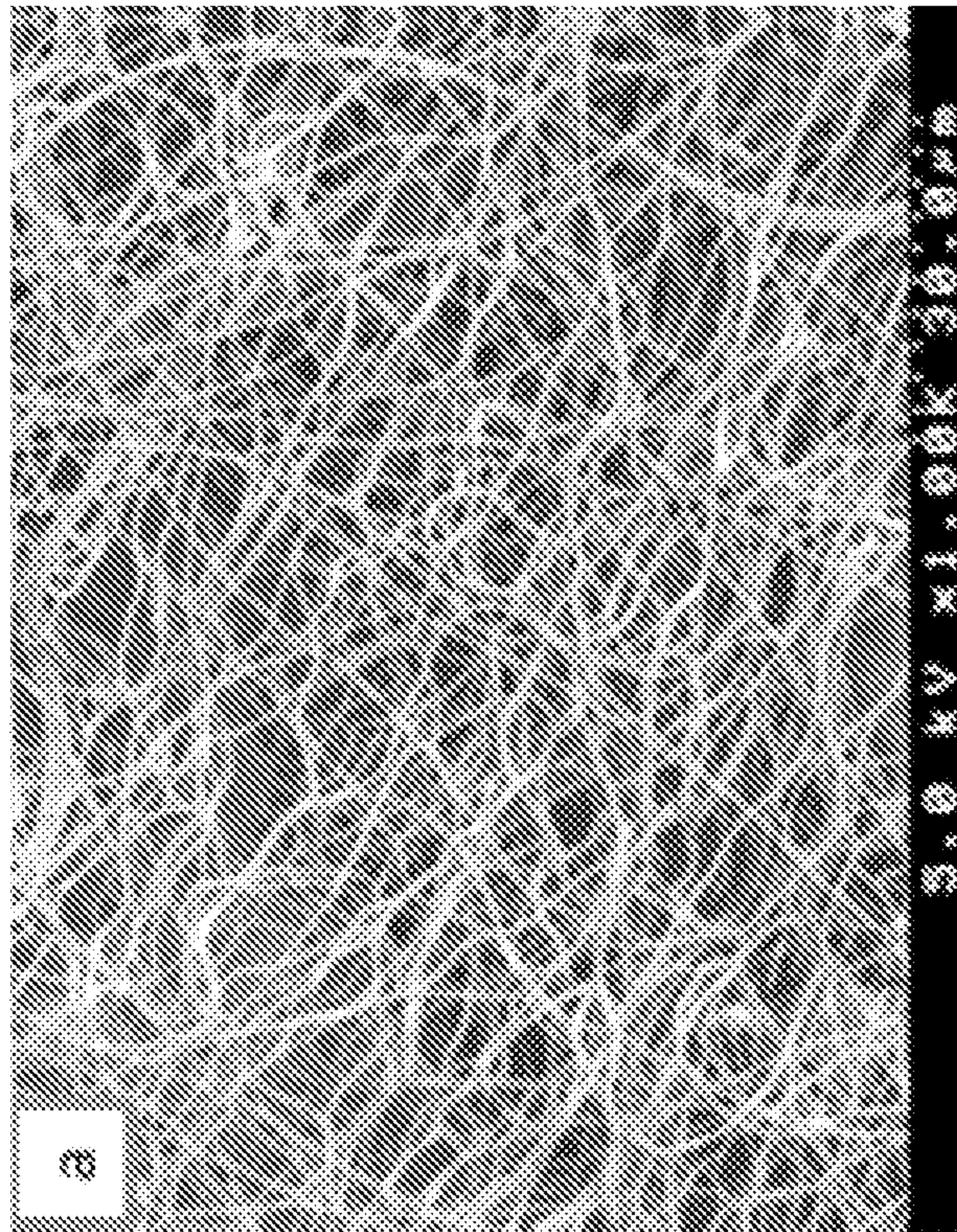


Figure 21A

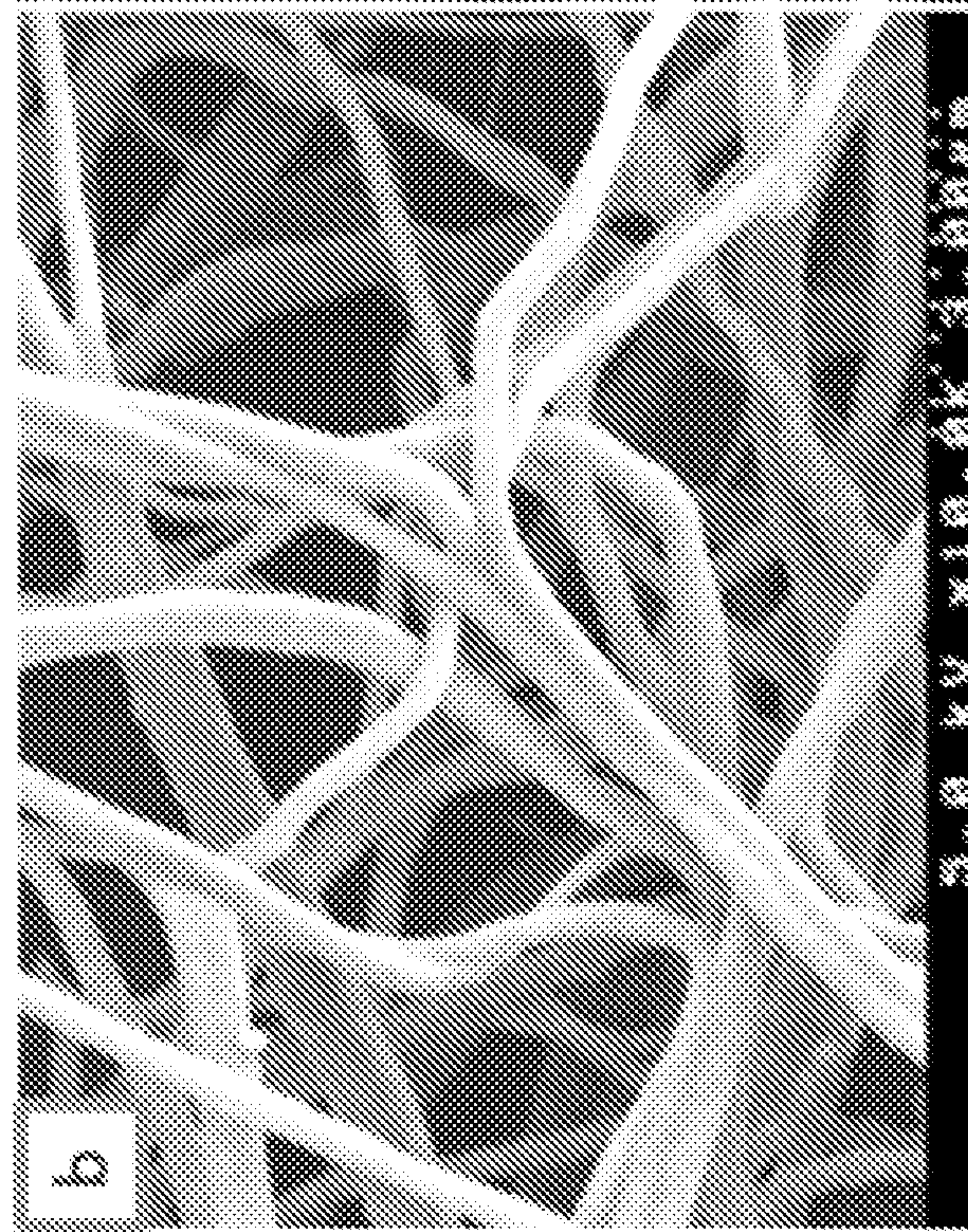


Figure 21B

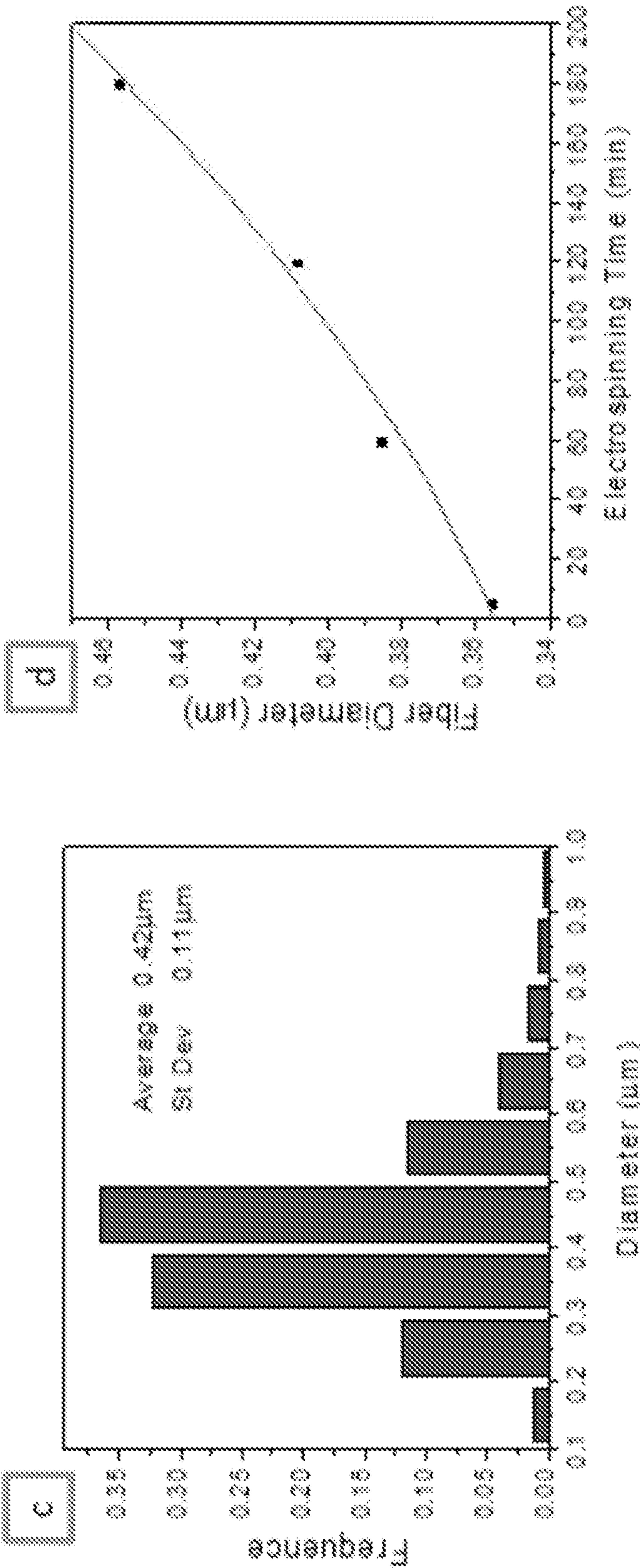


Figure 21C

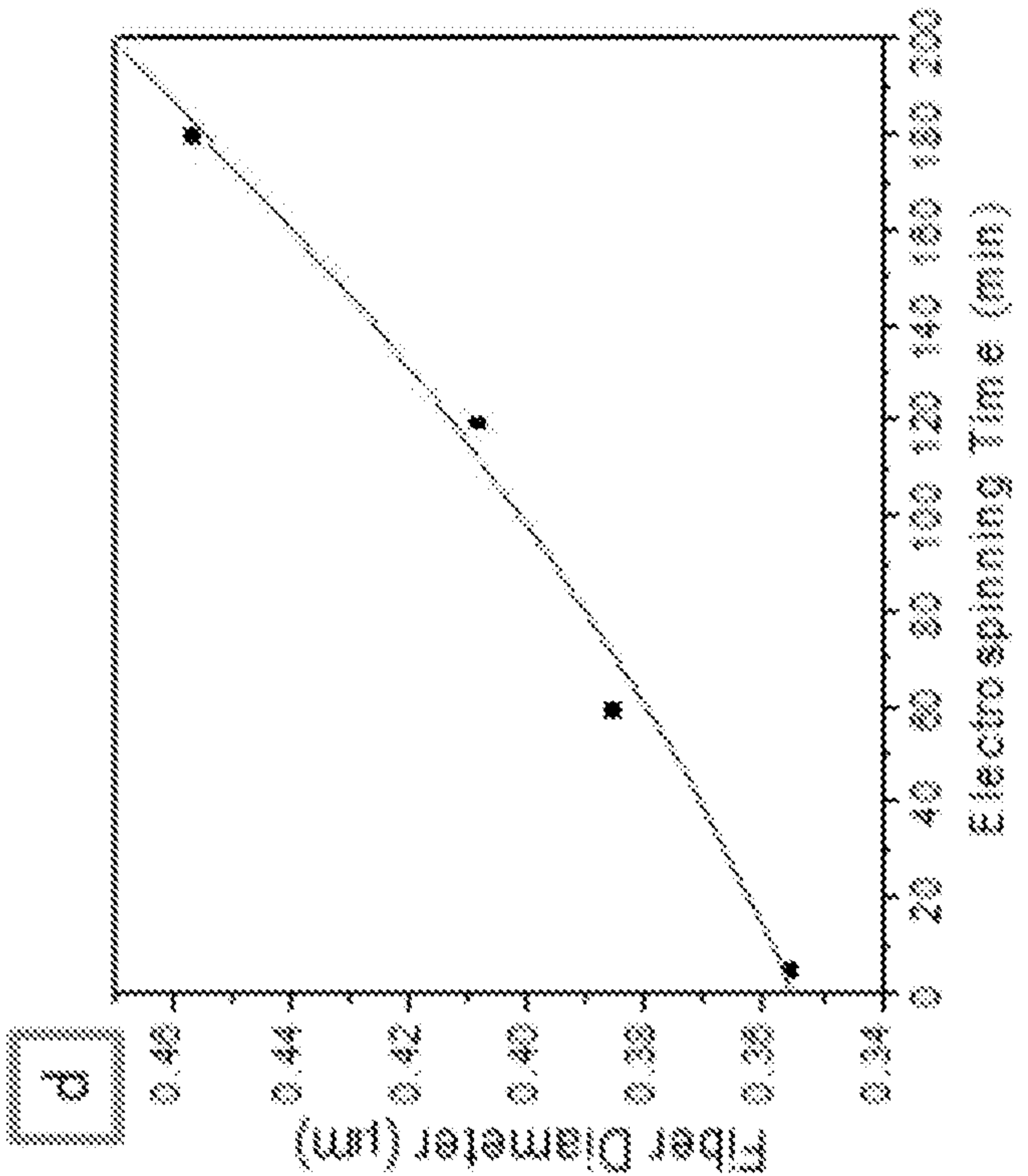


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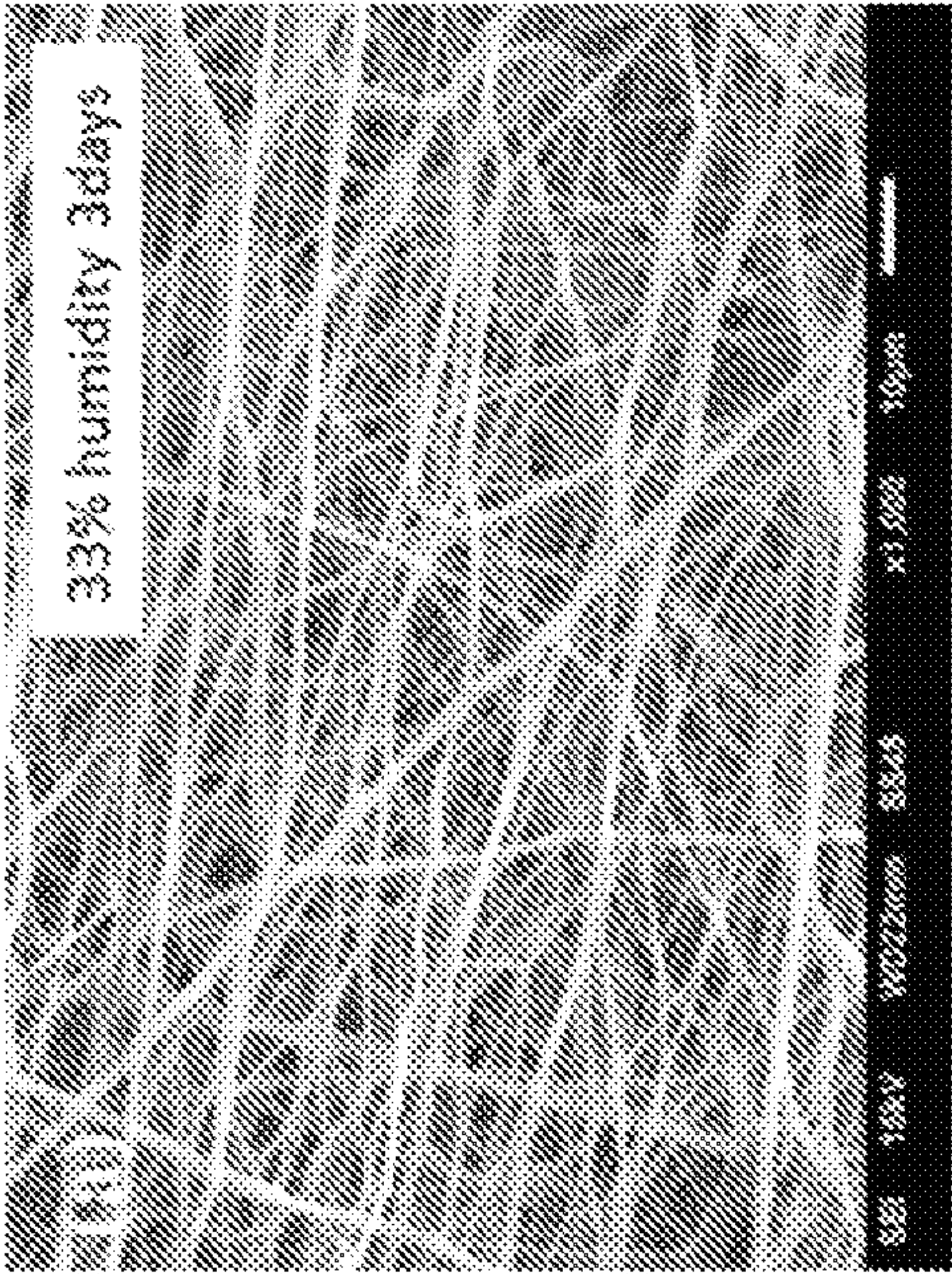


Figure 22A

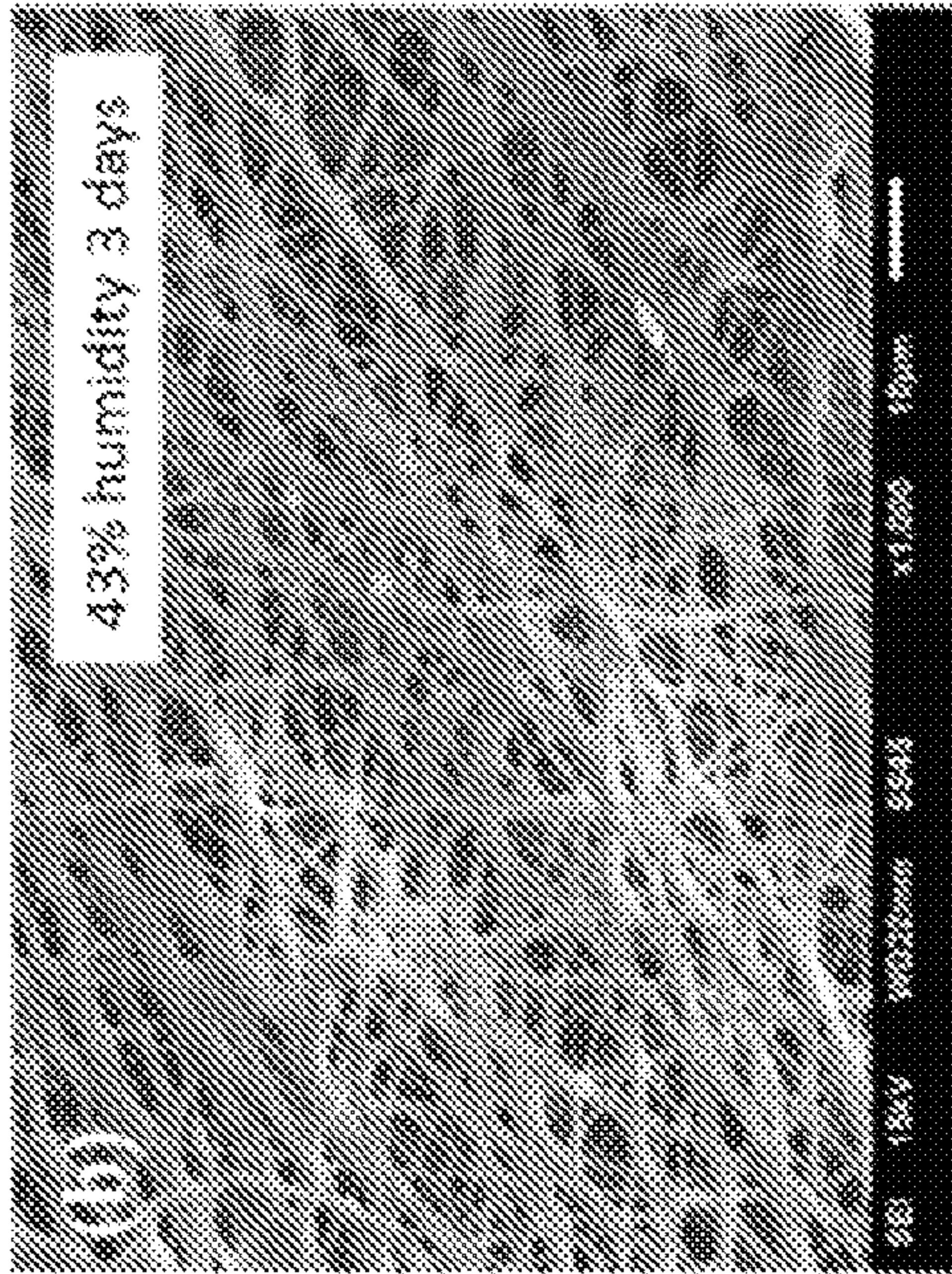


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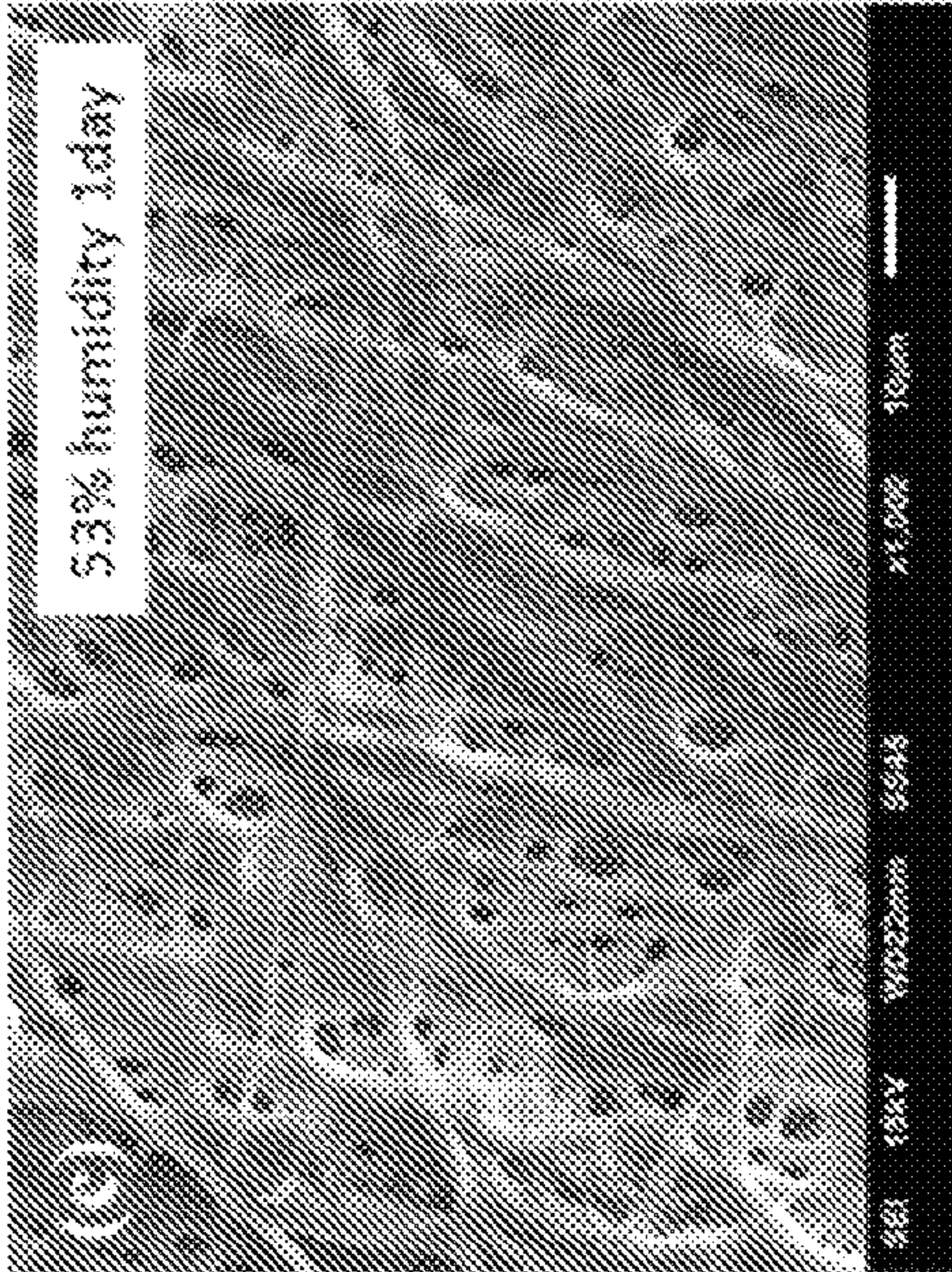


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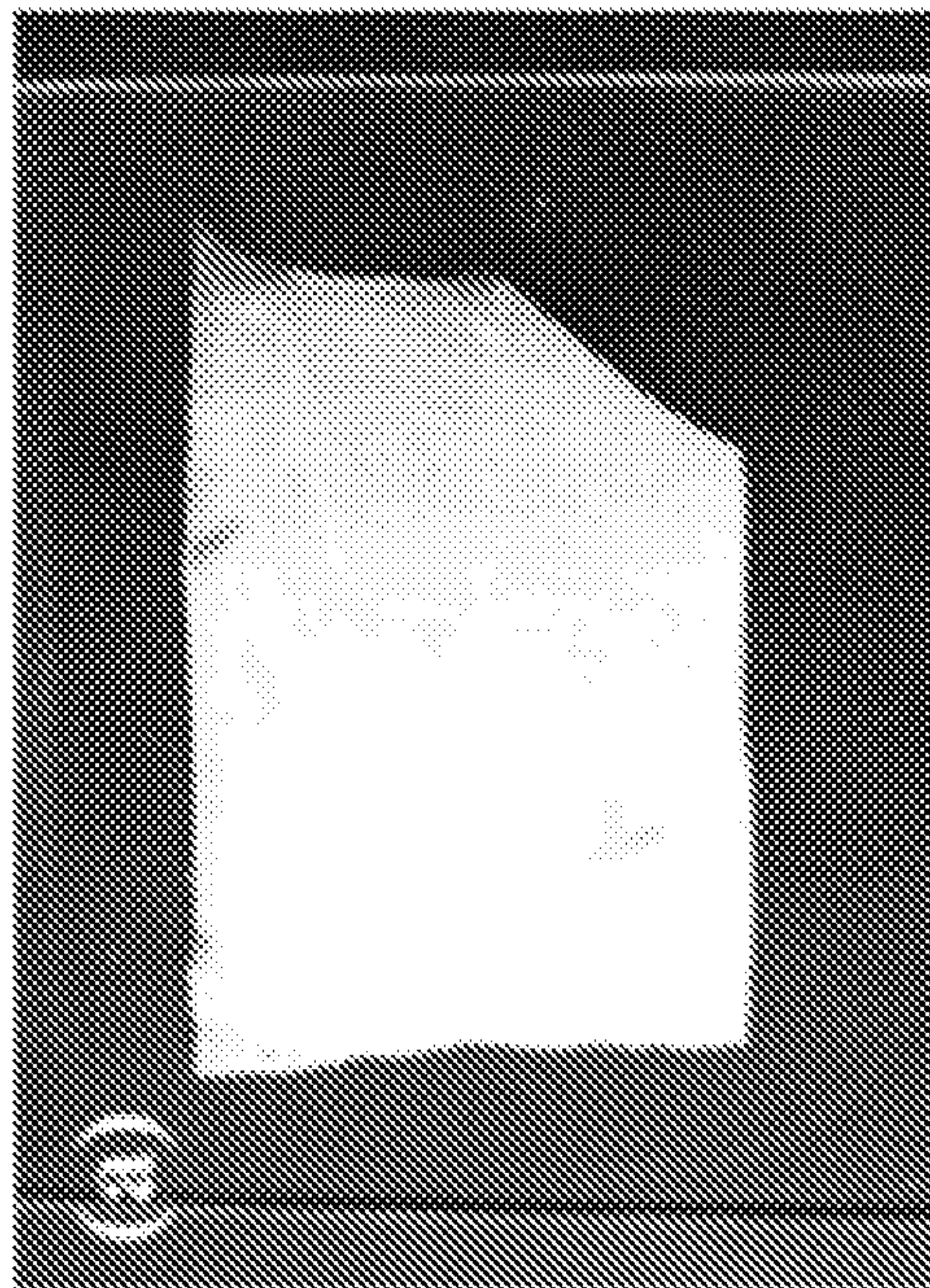


Figure 23A



Figure 23B

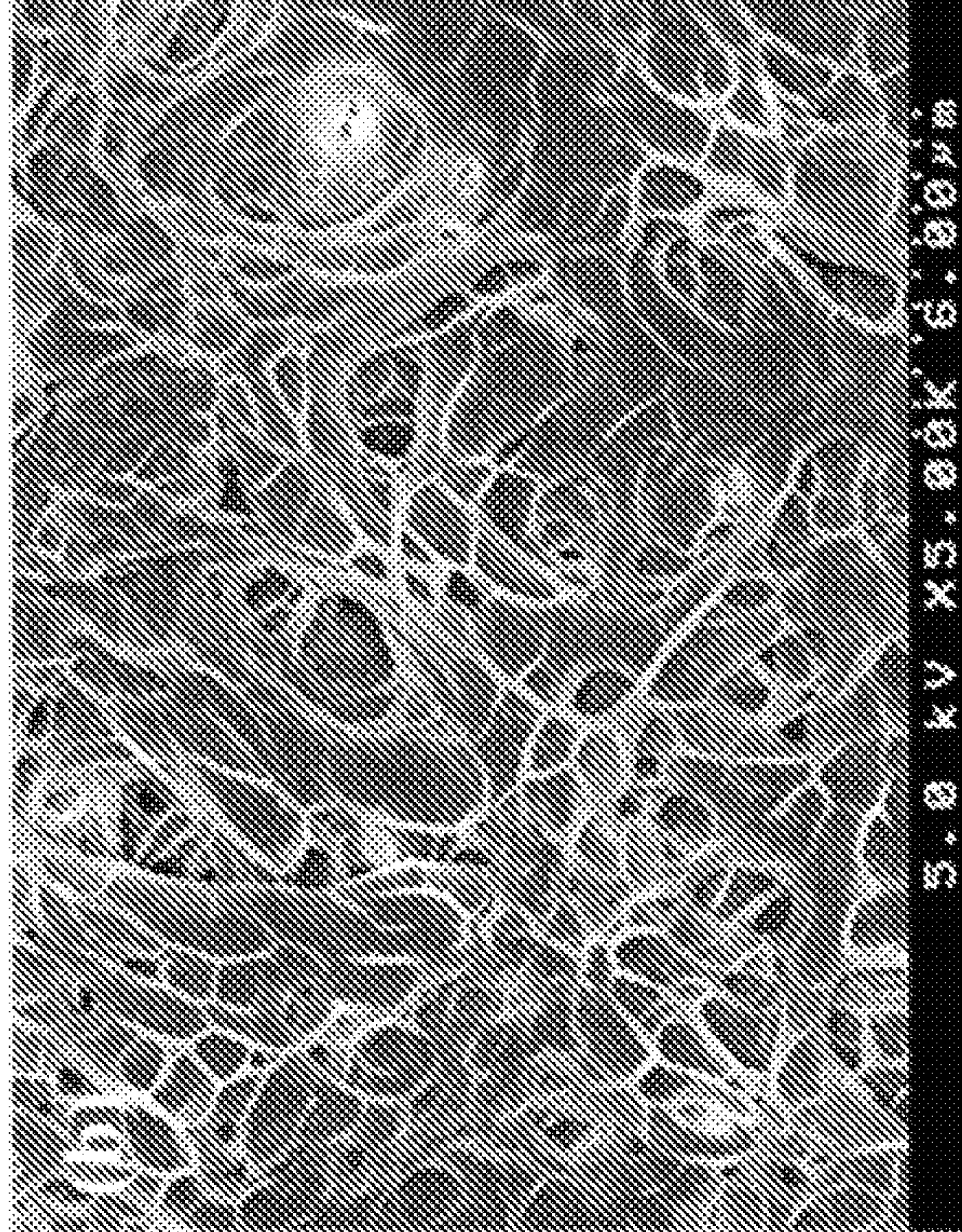


Figure 24B

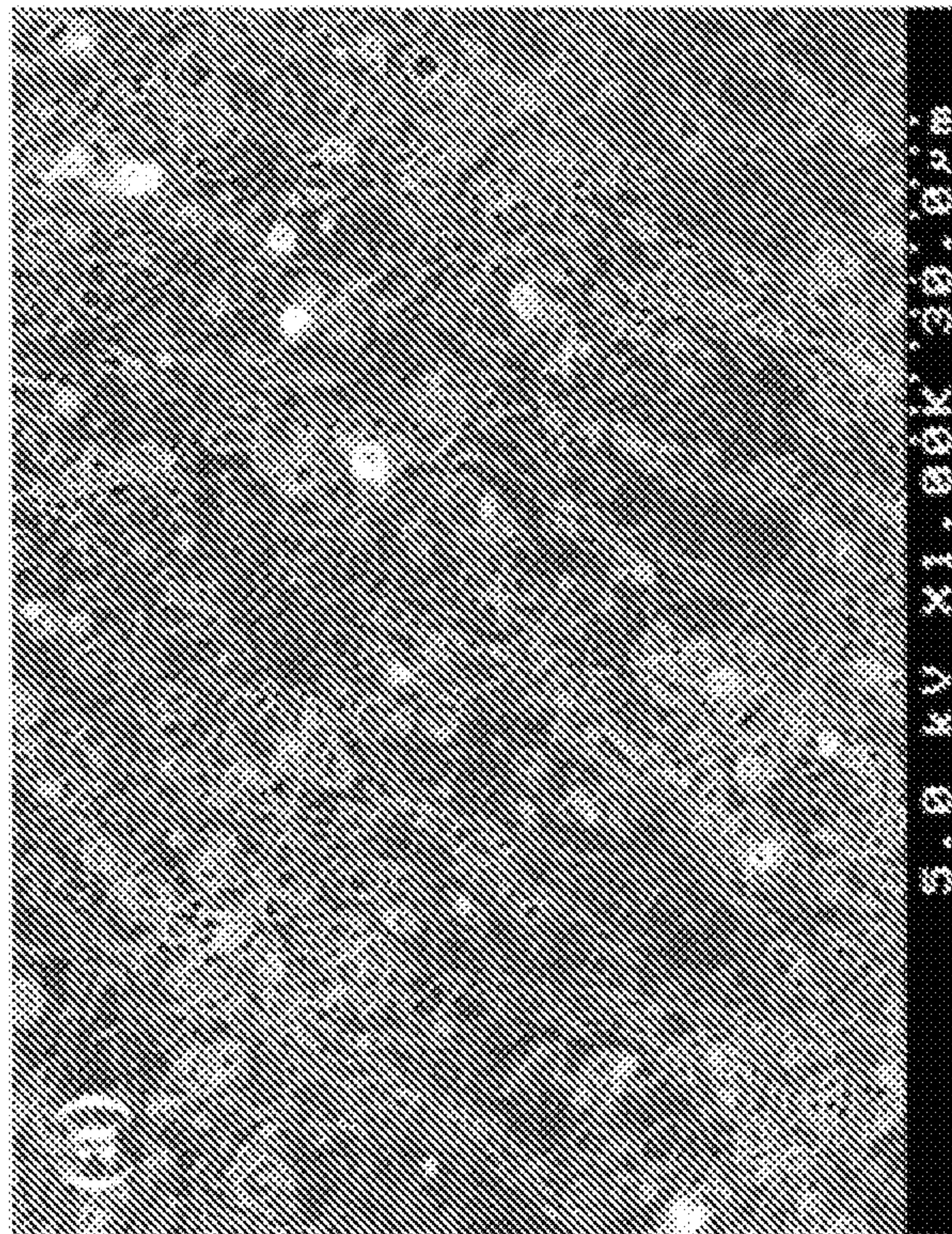


Figure 24A

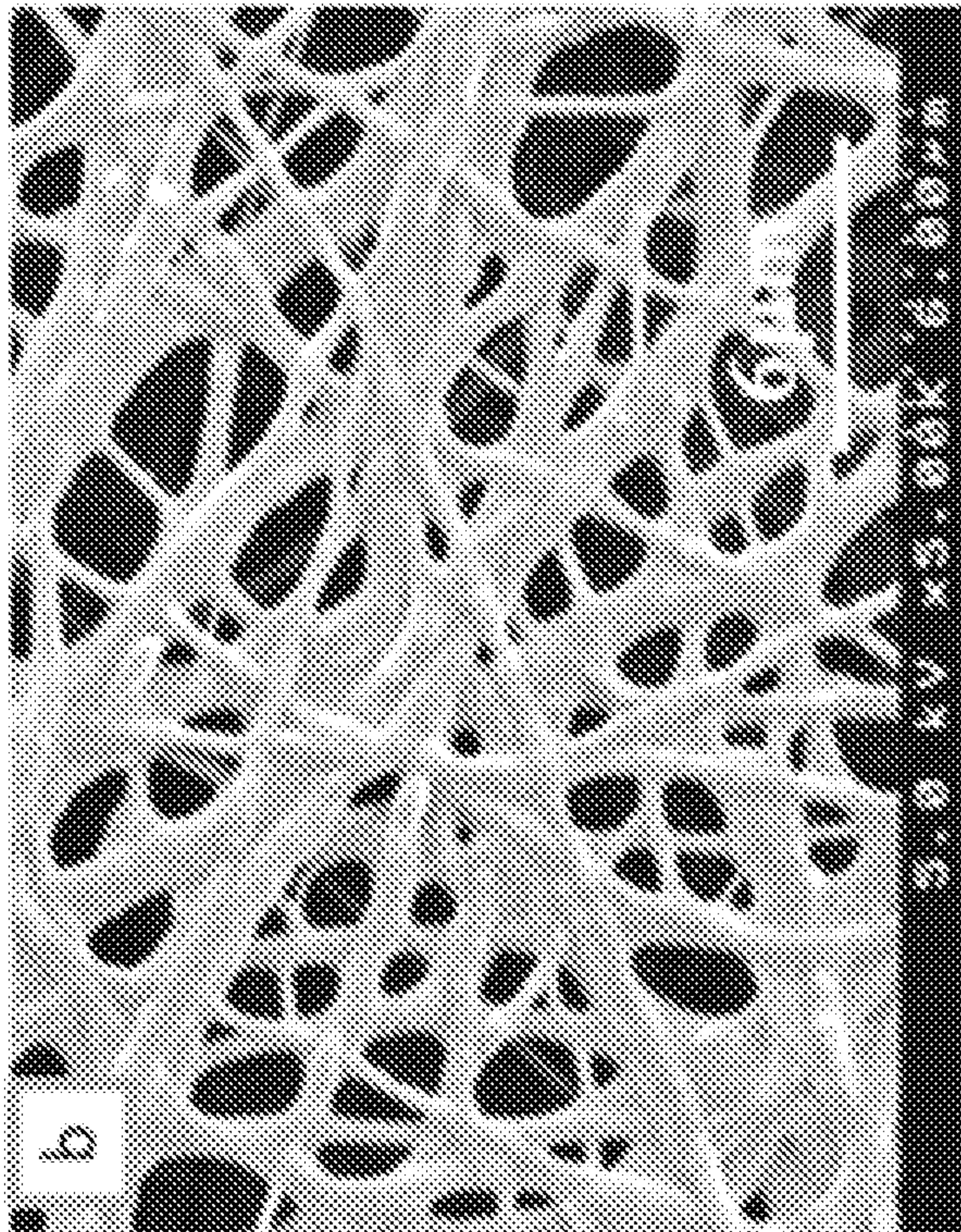


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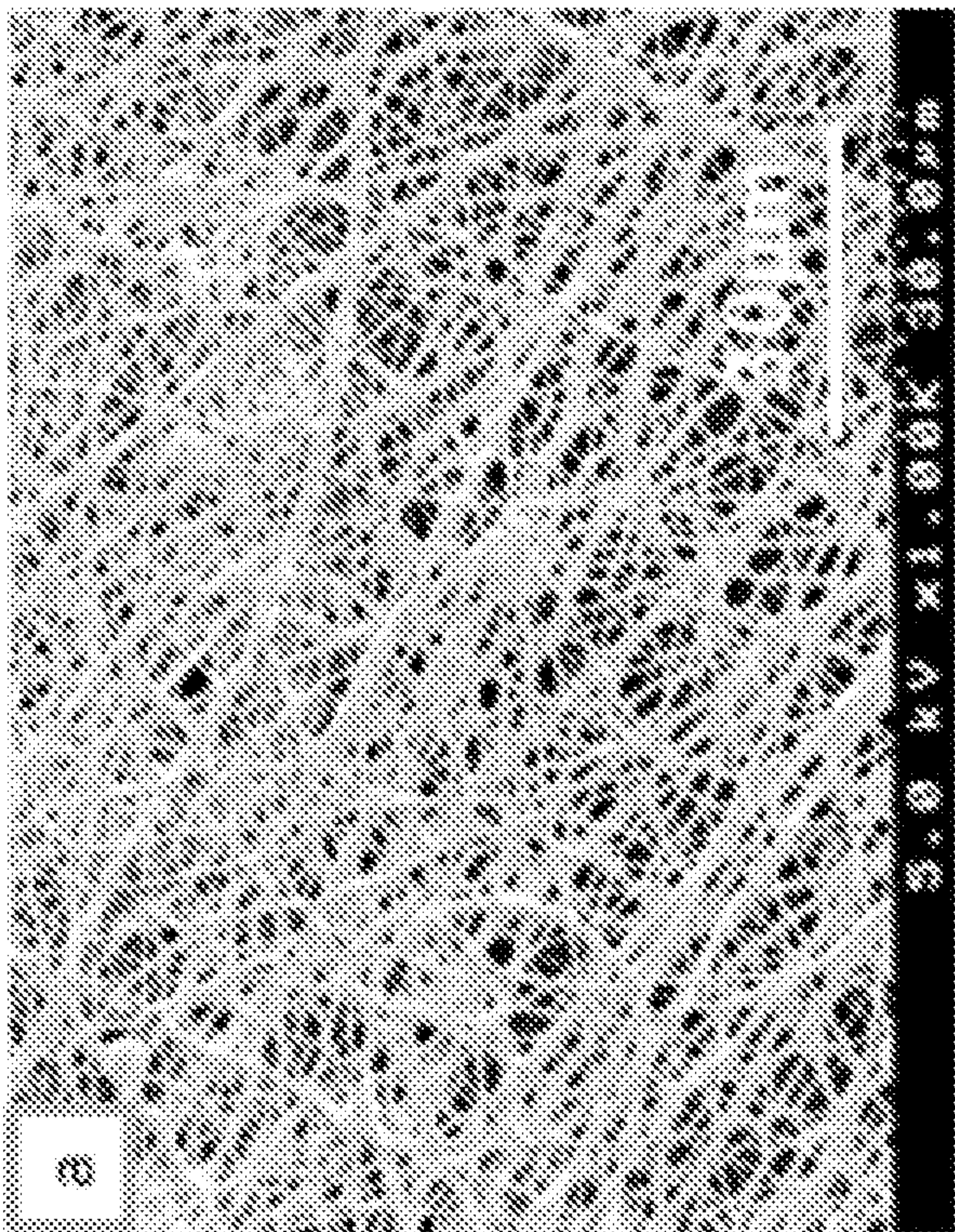


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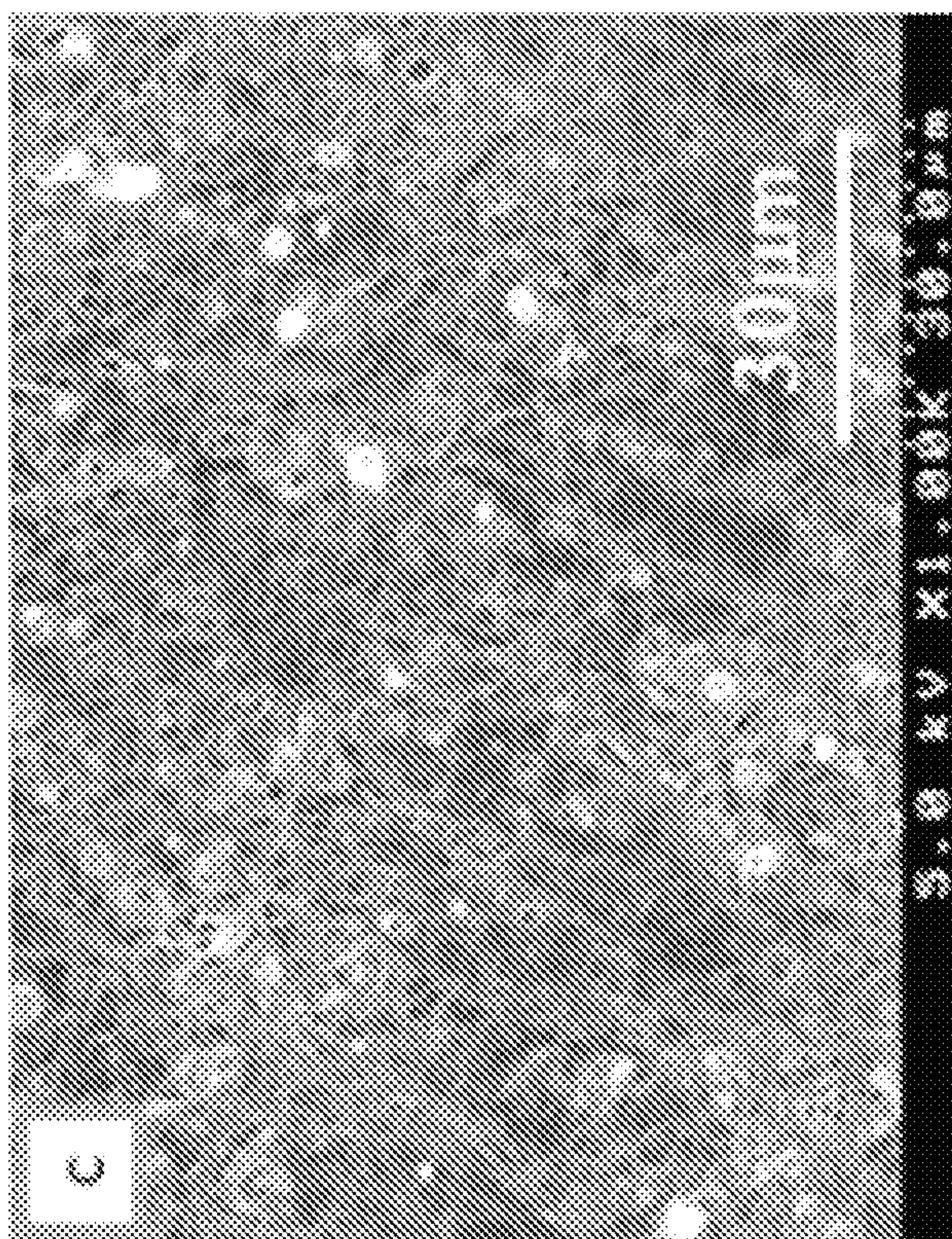


Figure 25C

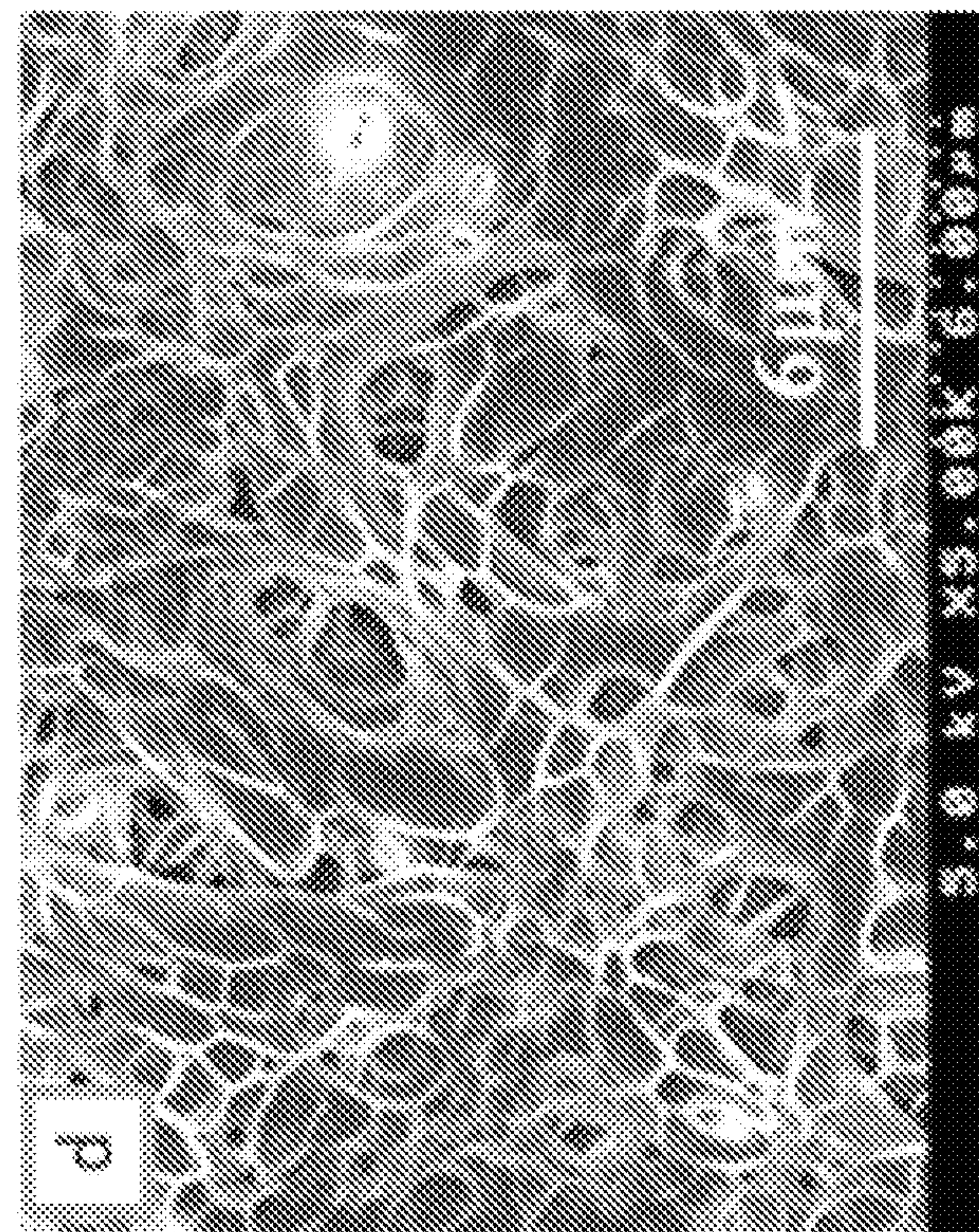


Figure 25D

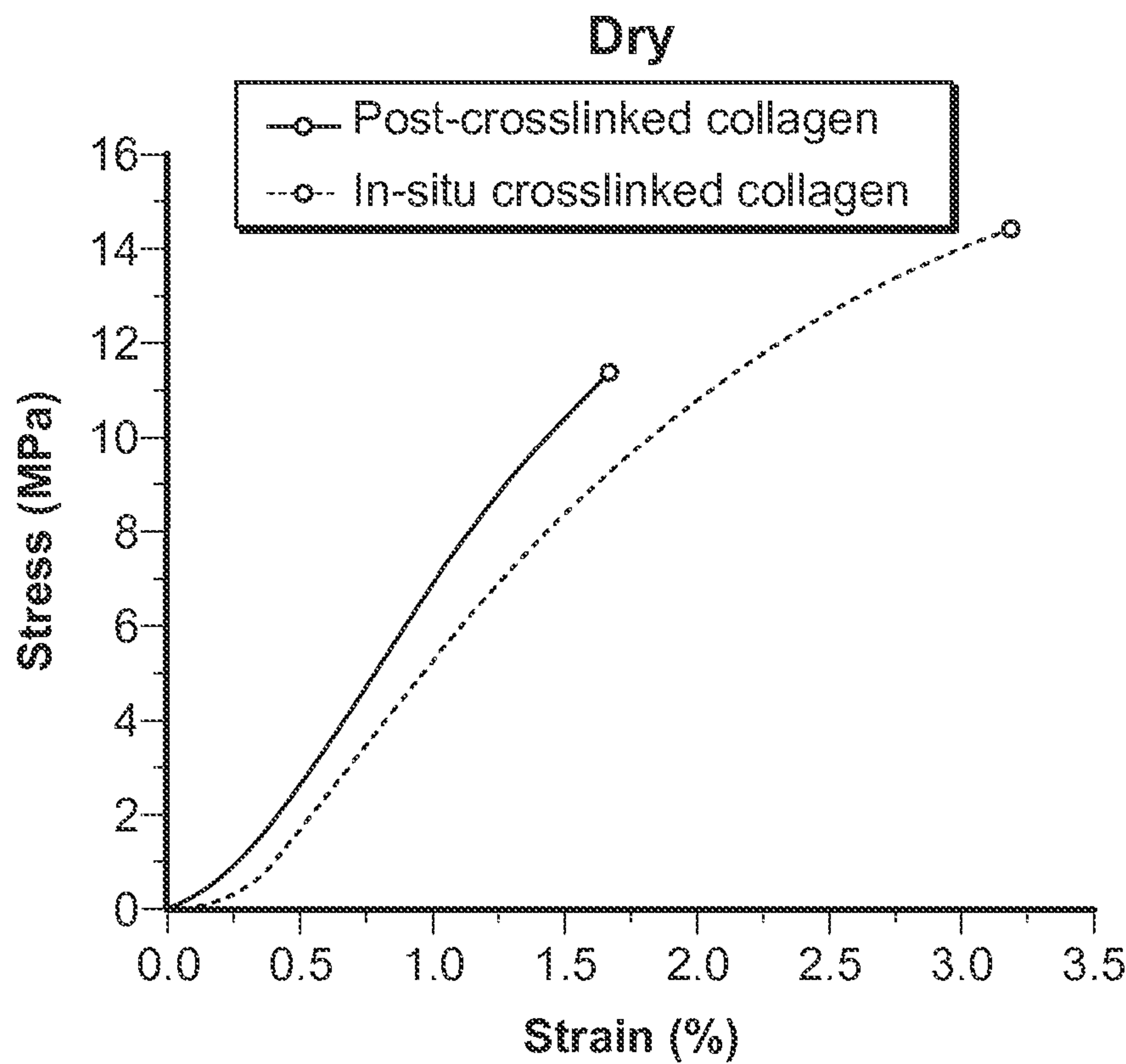


Figure 26

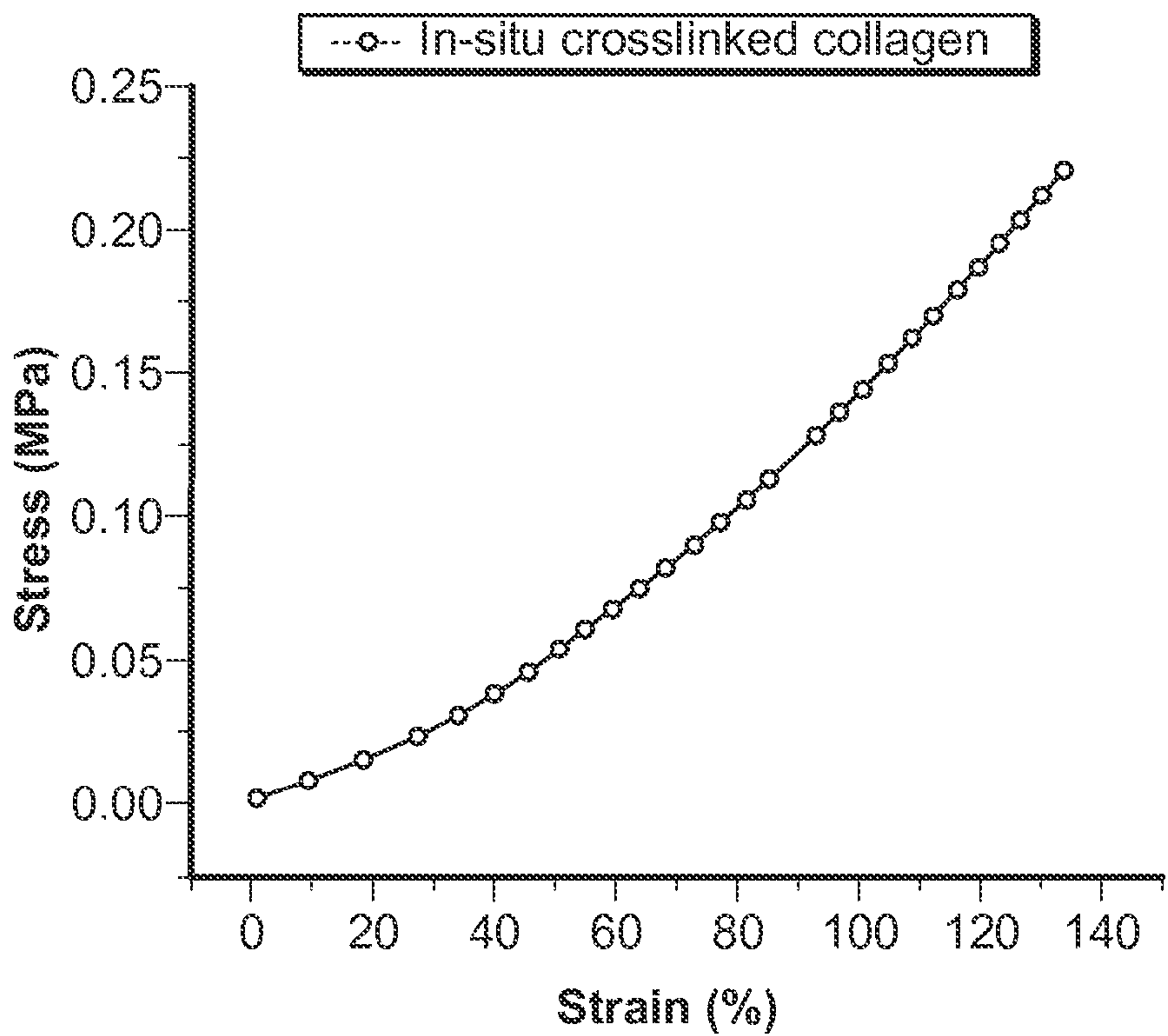


Figure 27

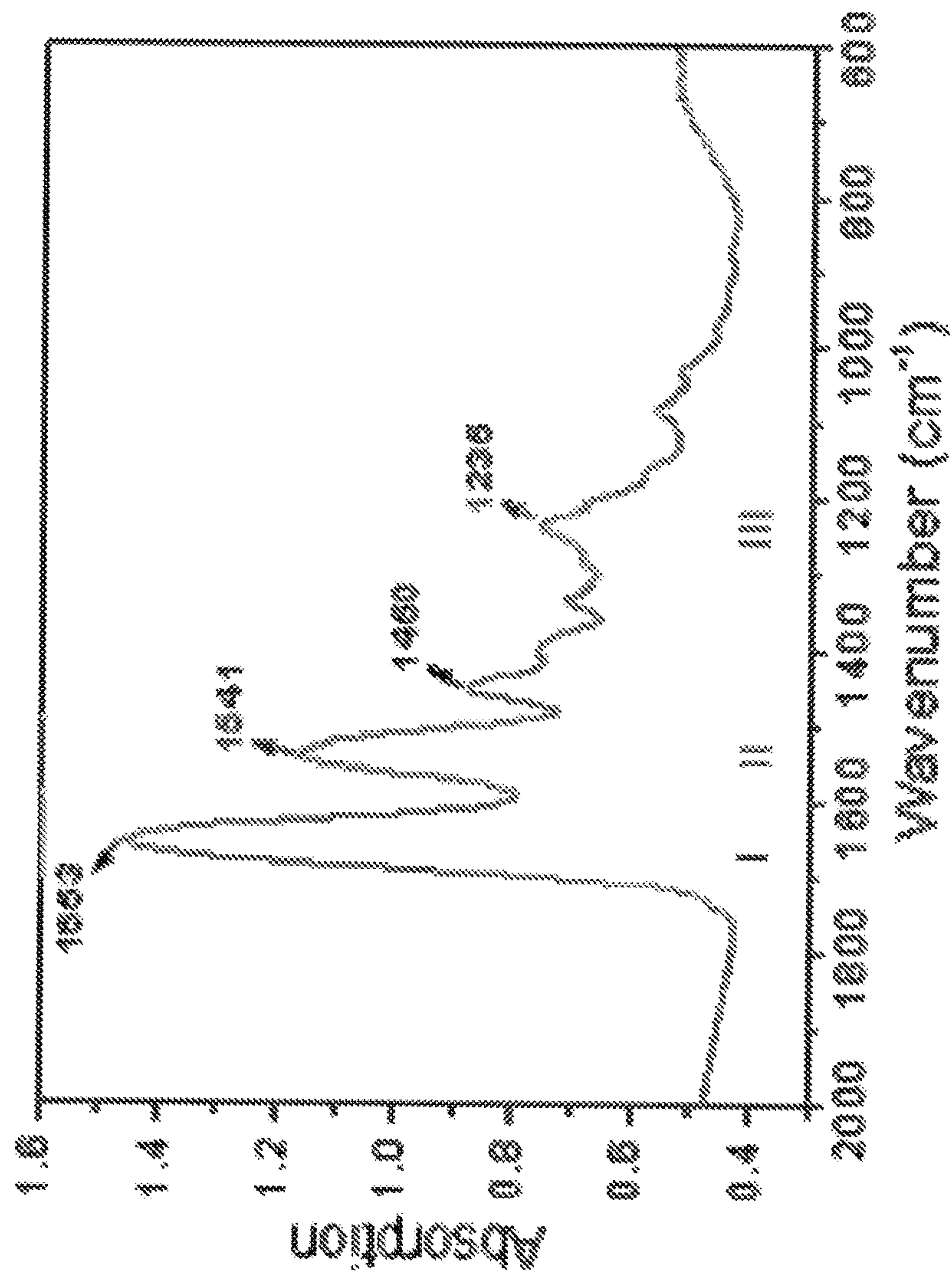


Figure 28

CONTROLLED CROSS-LINKING PROCESSING OF PROTEINS

PRIORITY CLAIM

This application is a division application of U.S. patent application Ser. No. 13/430,562, titled "Controlled Cross-Linking Processing of Proteins," filed on Mar. 26, 2012, which is a continuation-in-part of U.S. patent application Ser. No. 12/571,043, titled "Benign Solvents for Forming Protein Structures," filed on Sep. 30, 2009, which claims priority to U.S. Provisional Patent Application Ser. No. 61/194,685, titled "Electrospinning of Fiber Scaffolds," filed on Sep. 30, 2008. U.S. patent application Ser. No. 13/430,562 further claims priority to and the full benefit of U.S. Provisional Patent Application Ser. No. 61/467,923, titled "Kinetically Controlled Cross-Linking Processing of Proteins," filed on Mar. 25, 2011. This application claims priority to and the full benefit of all the foregoing referenced patent applications, which are incorporated by reference as if fully rewritten herein.

STATEMENT OF GOVERNMENT INTEREST

This invention was made with government support under Grant Number AR039750 awarded by the National Institutes of Health, National Institute of Arthritis and under Grant Number EY019406 awarded by the National Institute of Health, National Eye Institute. The government has certain rights in the invention.

TECHNICAL FIELD

The disclosed material relates generally to forming protein structures from a solution of protein dissolved in a benign solvent and cross-linking such protein structures.

BACKGROUND

Products and devices constructed from man-made materials can be implanted into or applied onto a human body to treat injuries, diseases, and other conditions of the human body. The materials chosen for such products or devices can be important for the product or device to successfully treat conditions of the human body. For instance, the compatibility of a material with the human body can determine if the product or device can be positioned on or in the human body. Products or devices can be made from synthetic material. However, if the synthetic material is dissimilar to human tissue, the success of the product or device can be limited. Products and devices constructed from naturally occurring materials such as proteins can provide biocompatible products or devices for implantation into or applying onto the human body to treat conditions of the human body.

SUMMARY

A method of forming a final protein structures includes preparing a solution of protein dissolved in a benign solvent and forming an intermediate protein structure from the solution. The intermediate protein structure is cross-linked by providing for a specific ratio of chemical cross-linking agents to form the final protein structure.

In another method, a cross-linked protein structure is formed. A solution is prepared by adding a cross-linker of N-hydroxysuccinimide (NHS) and 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide hydrochloride (EDC) at a ratio

of two-to-one of NHS to EDC to alcohol. PBS buffer (20×) is added to the solution until the volume ratio of PBS buffer (20×) to alcohol is about one-to-one. About 16 percent by weight of protein is dissolved in the solution. The solution is electrospun to form an intermediate protein structure. After a period of time, the protein structure is cross-linked to form the cross-linked protein structure.

BRIEF DESCRIPTION OF THE DRAWINGS

Objects and advantages together with the operation of the invention may be better understood by reference to the detailed description taken in connection with the following illustrations, wherein:

FIG. 1 is a schematic illustration of apparatus for electrospinning protein fibers from a protein solution;

FIG. 2A is a chart showing electrospun fiber diameter distribution;

FIG. 2B is an SEM image of an electrospun fiber scaffold;

FIG. 3A is an SEM image of an electrospun fiber scaffold;

FIG. 3B is an SEM image of an electrospun fiber scaffold;

FIG. 3C is a chart showing electrospun fiber diameter distribution;

FIG. 4A is an SEM image of an electrospun fiber scaffold;

FIG. 4B is a chart showing electrospun fiber diameter distribution;

FIG. 5A is an SEM image of an electrospun fiber scaffold;

FIG. 5B is a chart showing electrospun fiber diameter distribution;

FIG. 6 is multiple SEM images of electrospun fiber scaffolds;

FIG. 7 is an SEM image of an electrospun fiber scaffold;

FIG. 8 is a schematic illustration of apparatus for electrospinning or electrospraying protein agglomerations from a protein solution;

FIG. 9 is an SEM image of cross-linked agglomerations;

FIG. 10 is an SEM image of cross-linked agglomerations;

FIG. 11 is an SEM image of cross-linked agglomerations;

FIG. 12A is an SEM image of a cross-linked protein film;

FIG. 12B is an SEM image of a cross-linked protein film;

FIG. 13A is an SEM image of a protein structure with an open network of pores; and

FIG. 13B is an SEM image of a protein structure with an open network of pores.

FIG. 14A is a graph illustrating the relationship between gelation time and mole ratio of NHS to EDC based on rheological results.

FIG. 14B is a graph illustrating the relationship between the log of gelation time and mole ratio of NHS to EDC based on rheological results.

FIG. 15A is a photograph of hydrated in-situ cross-linked collagen gel.

FIG. 15B is a photograph of lyophilized in-situ cross-linked collagen gel.

FIG. 16A is a scanning electron microscope (SEM) image of a cross-section of lyophilized in-situ cross-linked collagen gel.

FIG. 16B is an SEM image of a top portion of FIG. 3a.

FIG. 16C is an SEM image of a middle portion of FIG. 3a.

FIG. 16D is an SEM image of a bottom portion of FIG. 3a.

FIG. 17A is a photograph of in-situ cross-linked collagen gel formed as tube-like structures.

FIG. 17B is a photograph of in-situ cross-linked collagen gel formed as a hemispheric-like structure.

FIG. 18 is a photograph of collagen gel used to print onto a glass slide.

FIG. 19A is a photograph of in-situ cross-linked collagen hydrogels with PCL-Pac mat embedding.

FIG. 19B is a photograph of in-situ cross-linked collagen hydrogels with PLEA-beta carotene macro beads.

FIG. 20A is a photograph of an electrospun in-situ cross-linked collagen fibrous scaffold in a dry state.

FIG. 20B is a photograph of an electrospun in-situ cross-linked collagen fibrous scaffold in a hydrated state.

FIG. 21A is a SEM image of electrospun in-situ cross-linked collagen fibers.

FIG. 21B is a SEM image of electrospun in-situ cross-linked collagen fibers.

FIG. 21C is a histogram of the electrospun in-situ cross-linked collagen fibers of FIG. 21A.

FIG. 21D is a graph illustrating the relationship between average fiber diameter and electrospinning time.

FIG. 22A is an SEM image of in-situ cross-linked collagen fibers exposed to 33% relative humidity for 3 days.

FIG. 22B is an SEM image of in-situ cross-linked collagen fibers exposed to 43% relative humidity for 3 days.

FIG. 22C is an SEM image of in-situ cross-linked collagen fibers exposed to 53% relative humidity for 1 day.

FIG. 23A is a photograph of an electrospun cross-linked collagen fibrous scaffold in a dry state.

FIG. 23B is a photograph of an electrospun cross-linked collagen fibrous scaffold in a hydrated state.

FIG. 24A is an SEM image of electrospun in-situ cross-linked collagen fibrous scaffolds after water treatment.

FIG. 24B is an SEM image of electrospun in-situ cross-linked collagen fibrous scaffolds after water treatment.

FIG. 25A is an SEM image of in-situ cross-linked collagen fibers three days after electrospinning with no water treatment.

FIG. 25B is an SEM image of in-situ cross-linked collagen fibers three days after electrospinning with no water treatment.

FIG. 25C is an SEM image of in-situ cross-linked collagen fibers provided with a water treatment three days after electrospinning.

FIG. 25D is an SEM image of in-situ cross-linked collagen fibers provided with a water treatment three days after electrospinning.

FIG. 26 is a graph illustrating the relationship between stress and strain for post-cross-linked collagen and in-situ cross-linked collagen in a dry state.

FIG. 27 is a chart illustrating the relationship between stress and strain for in-situ cross-linked collagen in a hydrated state.

FIG. 28 is a FTIR spectrum of water treated in-situ cross-linked collagen membranes.

DETAILED DESCRIPTION

The apparatuses and methods disclosed in this document are described in detail by way of examples and with reference to the figures. It will be appreciated that modifications to disclosed and described examples, arrangements, configurations, components, elements, apparatuses, methods, materials, etc. can be made and may be desired for a specific application. In this disclosure, any identification of specific shapes, materials, techniques, arrangements, etc. are either related to a specific example presented or are merely a general description of such a shape, material, technique, arrangement, etc. Identifications of specific details or examples are not intended to be and should not be construed as mandatory or limiting unless specifically designated as such. Selected examples of apparatuses and methods for

forming biocompatible protein structures from a solution of protein dissolved in a benign solvent and cross-linking such protein structures are hereinafter disclosed and described in detail with reference made to FIGS. 1-28.

Naturally occurring materials are good candidates for products and devices that are intended for use with biological material such as human and animal tissue. One category of materials that can be compatible with the biological material is natural polymers such as proteins. Examples of such biocompatible proteins include, but are not limited to, collagens, gelatin, elastin, fibrinogen, silk, and other suitable proteins. Such proteins can be used to form protein structures for implantation into or application onto a human body. Other materials that are generally biocompatible are polysaccharides such as hyaluronic acid, chitosan, and derivatives of starch and cellulose such as hydroxypropyl methyl cellulose phthalate, deoxyribonucleic acid (DNA), and ribonucleic acid (RNA).

One example of a protein structure that can be useful in forming products and devices for the human body is a scaffold or porous mat formed from protein fibers. Protein fibers can be used to construct a scaffold or porous mat structure that mimics an extra cellular matrix (ECM) of human tissue. Natural ECM generally has an open and porous structure. As will be described herein, fibers formed from proteins and joined into a matrix can simulate such an open and porous structure. Such a protein structure can be used in tissue engineering or wound care as a substrate for growing cells and/or tissue.

In another example, proteins can be used to form structures such as, for example, generally spherical agglomerates. Such agglomerates can be formed in a variety of sizes, ranging from submicron diameters to several hundred micrometers in diameter. Because of the compatibility of proteins with human tissue, protein agglomerates can be successfully implanted in or passed through the human body to affect treatment of a medical condition. For example, protein agglomerates can function as a component in a drug delivery system. A drug or other useful chemical compound can be attached to or inserted into a protein agglomerate. The protein agglomerate can then be passed through the human body, including through the blood stream, to a desired location where the drug can be released. In another example, protein agglomerates can function as structural or supportive components in the human body. For instance, protein agglomerates can be used in cosmetic medicine. Protein agglomerates can be injected under the skin to support the skin and smooth out wrinkles.

One method of forming a protein structure begins with dissolving a protein such as collagen in a solvent. Once dissolved, the protein can be extracted from the solvent and organized into a protein structure. One common solvent is 1,1,1,3,3,3 hexafluoro-2-propanol (HFP). However, any protein structure produced using such a solvent can have limited usefulness because of health concerns. For example, the United States Food and Drug Administration (FDA) has strict guidelines as to the amount of HFP allowed in a device or product intended for use with the human body. Because of strict FDA guidelines and general health concerns, using a solvent with benign characteristics for dissolving proteins or other biocompatible materials can yield biocompatible structures for implantation into or application onto a human body. Generally, a benign solvent is a solvent that either reduces health risks to a human body or is of minimal risk to the health of a human body.

One example of a benign solvent for dissolving protein comprises water, alcohol, and salt. The protein can be a Type

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I collagen, the alcohol can be ethanol, and the salt can be sodium chloride (NaCl). The association between water molecules, salt, and alcohol creates a complex structure in which proteins such as collagen are substantially soluble. Collagen is insoluble in most solvents because of interpeptide interaction. Collagen is substantially soluble in suitable water-alcohol-salt benign solvents because the properties of the solvents screen interpeptide interaction that usually results in insolubility of collagen. For example, the electrostatic interaction between the salt and the carbonyl group of the hydrophilic part of collagen and the hydrophobic interaction between the hydrocarbon chain of ethanol and the hydrophobic part of collagen can screen such interpeptide interaction. In general, any molecule or complex that exhibits a hydrophilic part and a hydrophobic part spaced by approximately the same distance as the hydrophilic part and hydrophobic part of the collagen molecule can dissolve collagen.

Although examples described herein include Type I collagen, it will be understood that all collagens—Type II, Type III, and so on—can be used in forming a protein structure for use with human tissue.

Generally, in suitable water-alcohol-salt solvents, the ratio of water to alcohol can range from a volume ratio of about 99:1 to about 1:99, the salt concentration can range from near 0 moles per liter (M) to the maximum salt concentration soluble in water, and the amount of protein by weight (as compared to the solvent) can range from near 0 percent to about 25 percent. In one example, the benign solvent comprises about a one-to-one ratio of water to ethanol and a salt concentration of about 3 M NaCl. Collagen is dissolved in such a solvent until the solution reaches about 16 percent collagen by weight. In another example, the solution comprises semed S (principally collagen type I with a ca. 5 percent collagen type III) dissolved in a solvent comprising phosphate buffered saline (PBS) buffer and ethanol, where the buffer to ethanol ratio of about one-to-one by volume. The saline concentration in the PBS buffer can range from 5× to 20×. The collagen concentration can be for example about 16 percent as compared to the total weight of the PBS/ethanol solvent. In yet another example, the protein dissolved in the solvent can be gelatin. The solvent can comprise a PBS buffer with a salt concentration of 10× mixed with ethanol at a one-to-one ratio by volume. Gelatin can be dissolved until the amount of gelatin by weight is about 16 percent by weight.

When protein has been dissolved in a suitable water-alcohol-salt solvent to form a protein solution, suitable processing methods can be used to extract protein from the solution and form protein structures. As previously discussed, such protein structures can be implanted into or applied onto the human body to affect treatment of a condition. Examples of suitable processing methods include, but are not limited to, electrospinning, electrospraying, and gravitational feed methods.

In one example, electrospinning can be used to form a protein structure. An example of apparatus 10 for forming a protein structure by electrospinning protein dissolved in a benign water-alcohol-salt solution is schematically shown in FIG. 1. The electrospinning method can include placing the protein solution in a syringe 12. The syringe can include a metal needle 14. The protein solution in the syringe 12 can be charged by the application of an electrical potential between the metal needle 14 and a ground target 16 spaced a distance away from the metal needle 14. The electrical potential can be applied by charging the metal needle 14 with a voltage from a power supply 18. The electrical

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potential can be increased until the electrostatic forces in the protein solution overcome the surface tension at the tip of the metal needle 14. As this surface tension is overcome, a fine jet 20 of solution containing entangled protein chains can be drawn out of the metal needle 14. As the fine jet 20 travels through the air, at least a portion of the solvent evaporates, resulting in a protein fiber 22 that dries as it travels through the air. The dry protein fiber 22 can be collected on a surface 24 that is in contact with the ground target 16. As shown in FIG. 1, the surface 24 can be on a rotating cylinder 26. It will be understood that the electrical potential can be created using a direct current (DC) power supply or an alternating current (AC) power supply.

In one example, the protein solution can be placed in a 5 milliliter (ml) syringe equipped with a 21 gauge blunt needle. The syringe can be placed in a syringe pump. A rotating drum can be placed approximately 10 centimeters (cm) from the tip of the needle. The pump rate can be set to about 1 milliliter per hour (ml/h) and the electrical potential can be set to about 20 kilovolts (kV). The result of such a setup can include the formation of a scaffold or mat on the rotating drum containing randomly oriented fibers or quasi-aligned fibers. The electrospinning process parameters, such as flow rate, potential field, and needle-to-collector distance can be adjusted to produce a variety of results or to optimize the stability of the fine jet of solution during electrospinning.

In another example, a scaffold or mat can be electrospun from a solution of about 16 percent by weight of gelatin dissolved in a PBS (10×) and ethanol solution with a volume ratio of about one-to-one. A flow rate of about 1 ml/h, a potential field of about 20 kV, and a needle-to-collector distance of about 10 cm can produce a stable jet of gelatin drawn from the gelatin solution.

Electrospinning proteins such as collagen and gelatin can result in the spinning of fibers as shown in FIG. 1. Such fibers can be highly aligned or oriented when mats and scaffolds are formed. In one example, electrospinning may be used to draw out protein fibers and such fibers can be generally arranged in a matrix. Once the fibers are arranged in a matrix, the fibers can be cross-linked to mimic the structure of ECM. Once cross-linked, the formed mat or scaffold can be a stable non-water-soluble protein structure that is biocompatible with the human body and thus implantable into or applicable onto the human body.

As the mat or scaffold is being formed by electrospinning, the fibers can be arranged so that fibers overlay one another and are in contact with one another. While in such an arrangement, the physical structure of the mat or scaffold can be enhanced by cross-linking the protein fibers. In one example, end groups such as aldehyde, carbodiimide, or epoxy can facilitate the cross-linking of the protein fibers of the mat or scaffold. A carbodiimide such as EDC can cross-link collagen using NHS as a catalyst. An electrospun collagen mat can be immersed in a 200 mM EDC and NHS ethanol solution for approximately 4 hours to cross-link collagen fibers. Once cross-linked, the collagen mat or scaffold can be placed in a PBS and salt solution similar to the buffer described above. Such a step can remove any non-cross-linked collagen from the mat or scaffold. The mat and scaffold, which can mimic the extra cellular matrix of human tissue can now be used as a substrate to grow cells or tissue, or can be used as a covering for an open wound to promote growth of tissue of the wound.

Examples of methods for forming a protein mat or scaffold by electrospinning can include adjusting the protein's solubility in the benign solvent; adjusting the evaporation rate of the solvent; adjusting the viscosity of the solution; or

adjusting the surface tension of the solution. In one example, the solubility of collagen is enhanced by the addition of a salt to a water and ethanol mixture with a generally neutral pH level. When about 5 percent by weight of NaCl is added to a water and ethanol mixture for an about 16 percent by weight collagen solution, substantially all collagen dissolves. In one example, the salt composition of a PBS buffer solution can be about 80 percent NaCl by weight, about 17.4 percent sodium phosphate anhydrate by weight, and about 2.4 percent potassium phosphate anhydrate by weight. In another example, collagen may be dissolved in a PBS buffer where the total salt concentration exceeds 5 percent by weight. The evaporation rate of the solvent can be increased by increasing the amount of alcohol as compared to water in the protein solution.

As will be understood, the pH level, temperature, type of collagen, and type and concentration of salt all influence the structure of collagen in the protein solution. For example, at low collagen concentrations and a pH level of about 7.4, the transition temperature of crystalline polymer to random coil polymer is about 45 degrees Celsius. The transition temperature can be independent of salt concentration for potassium chloride (KCl) and NaCl. There is a progressive decrease in precipitation of collagen, that is to say that collagen becomes more soluble as more salt is added. Addition of salt results in destabilization of the precipitated collagen while the ionic strength increases with salt additions. Collagen solubility can increase even if it appears that the crystalline structure of collagen is maintained upon addition of salt.

Alcohol affects the solubility of collagen in the buffer and ethanol solution. Alcohol and collagen interaction is moderated by hydrocarbon chain length, with alcohol disrupting internal hydrophobic interactions in the collagen. With increased alcohol concentration, there is a progressive increase in molar destabilization of the crystalline collagen precipitated in an alcohol and potassium acetate buffer mixture at an acidic pH, for example, a pH of about 4.8. For single collagen molecules, structural stability is primarily a function of interpeptide hydrogen bonding and chain rigidity.

The addition of salt promotes the solubility of collagens. Hydrogen bonding between the hydrophilic part of collagen and water molecules can be too weak to break the interpeptide interaction, and the stronger electrostatic forces induced by salt in aqueous media may be necessary. The combination of both electrostatic and hydrophobic forces appears to interact strongly enough with the collagen chain to substantially dissolve the collagen in a mixture of ethanol and PBS buffer with an about one-to-one ratio when a salt concentration is at least about 5× in the buffer.

In addition to dissolving proteins such as collagens, the buffer and ethanol binary solvent can further facilitate the electrospinning process. The salt in the buffer as well as the alcohol can assist in overcoming the high surface tension of water that can partially inhibit spinnability of water based polymeric solution. In addition, the salt increases the charge density in the protein solution, which can facilitate the formation of a stable Taylor cone. The low evaporation rate of water, which can inhibit the formation of fibers during electrospinning, can be compensated for by the high evaporation rate of alcohol.

Electrospinning of collagen solutions with about a one-to-one volume ratio of ethanol to PBS (10× or 20×) can be stable and create fiber mats or scaffolds that exhibit relatively consistent fiber diameters. The increase of salt concentration in the PBS buffer can decrease the fiber diameter,

and higher salt concentration can result in greater elongation of the electrospun jet due to higher density of repulsive charges in the Taylor cone. As is shown in the chart of FIG. 2A, increasing the salt concentration from 10× to 20× decreases the average fiber diameter from 540 micrometers to 210 micrometers but also significantly reduces the standard deviation of the fiber diameter distribution (from 210 micrometers to 60 micrometers). FIG. 2B is a scanning electron microscope (SEM) image of collagen fibers electrospun from PBS (20×) and ethanol.

As will be understood, cross-linking of protein mats or scaffolds facilitates the use of electrospun mats or scaffolds for regenerative or tissue engineering and wound care, because cross-linking promotes stability of the collagen mat or scaffold. In addition to mimicking the ECM of human tissue to promote cell or tissue growth, when collagens with hemostatic properties are used, application of a mat or scaffold over a new or existing wound can arrest blood flow from the wound and promote clotting.

Cross-linking can be facilitated by the presence of carboxyl groups on the hydrophilic part of collagens. FIGS. 3A and 3B illustrate cross-linking of fiber mats with EDC and NHS as a catalyst. Mats are immersed in an ethanol solution comprising EDC and NHS for four hours. The mats can then be immersed in a buffer solution containing the same salt concentration as the one the collagen was electrospun from to remove un-cross-linked fibers.

For the collagen mats shown in FIGS. 3A and 3B, the collagen fibers of the mat were cross-linked with EDC and NHS. The fibers were electrospun from a PBS (10×) and ethanol solution. FIG. 3A is an SEM image of a self-standing mat and FIG. 3B is a framed mat. FIG. 3C is a chart of the diameter distribution of cross-linked fibers electrospun from a PBS (10×) and ethanol mixture. As seen in FIG. 3B, the cross-linked mat may retain a porous and open structure upon cross-linking.

A collagen mat can be soaked in an ethanol solution such that the mat shrinks to form a film-like surface (for example, as shown in FIG. 3A). If shrinkage is not desired, frames can be placed on each side of the mat and clipped together to prevent the mat from shrinking when it is immersed in the ethanol solution (for example, as shown in FIG. 3B). Such frames can be constructed of material that is easy to remove, such as Teflon. The fiber diameter distribution does not significantly change between non-cross-linked and cross-linked collagen when a frame is used. Therefore, the frame can efficiently prevent fiber shrinkage when immersed in ethanol.

The architectural structure of protein mats or scaffolds can be important depending on the intended application of the mat or scaffold. For example, mats and scaffolds can be used to simulate types of human tissue. Aligned fibers may be useful in simulating a variety of tissue types including ligaments, nerves, cardiac tissues, and the like. The alignment of electrospun fibers may be controlled by the rotational speed of the rotating cylinder 26 shown in FIG. 1. If the speed of the cylinder matches or is faster than the speed of the jet of protein solution exiting the syringe, the protein fibers may be drawn out of the syringe in the loop direction of the cylinder. The orientation of protein fibers in the mat can be characterized by Herman's orientation function, which is:

$$F = (3 \langle \cos^2 \theta \rangle - 1) / 2$$

where θ is the angle of the protein fibers compared to the loop direction of the drum.

An optimally aligned fiber mat (that is to say, a mat where all the fibers all aligned in the same direction) will have a Herman's orientation function equal to 1. An optimally random configured fiber mat (that is to say, a mat where all the fibers are randomly aligned) will have a Herman's orientation function equal to -0.5 . FIG. 4A shows an SEM image of a protein fiber mat, where the speed of the rotating drum matched the speed of the jet of protein solution. The mat was electrospun from a PBS (20×) and ethanol solution and has a Herman's orientation function equal to about 0.93. The mat shown has relatively highly oriented fibers. Because oriented fibers can be mechanically drawn, the fibers can have smaller diameters than randomly oriented fibers electrospun under similar conditions. However, as shown in FIG. 4B, for this particular mat, where the speed of the rotating drum matched the speed of the jet of protein solution, the drawn and aligned fibers do not show significantly smaller diameters than random fibers.

As previously discussed, gelatin may be electrospun from binary solutions and electrospinning conditions disclosed herein. For example, when a PBS (10×) and ethanol solution is used for dissolving gelatin at about 16 percent by weight, similar results are obtained as compared to collagen fibers. In addition, the gelatin can be cross-linked in a similar manner and under similar conditions as described for collagens. FIG. 5A shows a gelatin fiber mat and FIG. 5B shows a chart of the diameter distribution of the gelatin fibers.

The concentration of collagen in the solution may affect electrospinning. Collagens readily dissolve in a solvent comprising a one-to-one ratio between PBS (20×) and ethanol. Generally, collagen solutions ranging from about 4 percent by weight to about 25 percent by weight can be electrospun. By controlling the concentration of collagen, different morphologies and fiber diameters can result. FIG. 6 includes several SEM images of morphologies resulting from electrospinning different concentrations of collagen dissolved in solutions. FIGS. 6a and 6b are different magnifications of a fiber mat electrospun from a solution with about 4 percent collagen by weight. As may be seen, generally the diameter of the fibers is inconsistent because the viscosity of the solution is low and does not generally form continuous fibers during electrospinning. FIGS. 6c and 6d are different magnifications of a fiber mat electrospun from a solution with about 10 percent collagen by weight. As the concentration of collagen is increased, the diameter of the fibers becomes more consistent because continuous fibers are more readily generated by a solution with about 10 percent collagen by weight. FIGS. 6e and 6f are different magnifications of a fiber mat electrospun from a solution with about 16 percent collagen by weight. As can be seen, submicron fibers of generally consistent diameter are formed.

The concentration of salt and ethanol can affect the solubility of collagens in water. Collagen can be generally insoluble at about 16 percent by weight in either PBS (20×) or ethanol. However, when a small amount of ethanol is added into PBS (20×) buffer to form a PBS (20×) to ethanol volume ratio of about nine-to-one, the collagen substantially dissolves into this mixture. By adding more ethanol into PBS (20×) buffer (that is, the volume ratio decreases from about nine-to-one to about seven-to-three to about one-to-one) there is generally no affect on the solubility of collagen. The collagen remains substantially soluble. However, when the PBS (20×) to ethanol volume ratio is reduced to three-to-seven, collagen is generally no longer soluble. Furthermore, the salt concentration affects the solubility of collagen when the water to ethanol volume ratio is held constant at

about one-to-one. The salt concentration in 5×, 10× and 20×PBS buffer is sufficient to substantially dissolve collagen in the mixture solution.

The addition of salt and ethanol to the protein solution can facilitate the electrospinning of the polymer solution. As salt increases the conductivity and ethanol decreases the boiling point, concentrations of salt and ethanol affect the electrospinnability of solutions that are capable of dissolving collagen with PBS (20×) to ethanol ratio varying from about nine-to-one to about one-to-one. As seen in FIGS. 7a-7b, fibers may be formed from electrospinning collagen with PBS (20×) to ethanol volume ratios of about seven-to-three. In addition, collagen solutions with a PBS (20×) to ethanol volume ratio of about one-to-one demonstrate good electrospinnability and a stable Taylor cone. Such a solution may be electrospun to form fibers and a mat as thick as about 150 microns.

In one example, the protein solution can include a cross-linking agent so that cross-linking of protein fibers occurs as the protein fibers are being electrospun. This reduces the formation and cross-linking of protein fibers to one general step. In such an example, the protein solution includes protein, water, alcohol, salt, and a cross-linking agent. A protein solution is formed by dissolving about 16 percent by weight of collagen in a solvent. The solvent comprises PBS buffer (20×) and alcohol. Prior to forming the solvent a cross-linker is added to the alcohol. The cross-linker can be about 200 mMoles of EDC and NHS at a ratio by weight of about one-to-one. The collagen solution can be deposited in a syringe equipped with a metal needle as previously described. The protein solution is subjected to an electrical potential and electrospun to form a jet of protein solution and form a protein structure such as a mat or scaffold. In one example, a voltage of about 20 KV can be applied to the metal needle and the pump rate can be about 0.5 milliliters per hour. A rotating drum can be positioned about 10 centimeters from the needle to collect the electrospun mat.

Protein dissolved in benign solvents as described herein can be used to form protein agglomerates such as generally spherical particles or beads. An apparatus 100 for forming protein agglomerate is schematically shown in FIG. 8. Protein agglomerates can be formed using methods that include electrospinning, electrospraying, and gravitational feed methods. The apparatus 100 includes a syringe 102 equipped with a metal needle 104. The syringe 102 is suspended over a receptacle 106, and the receptacle 106 is positioned on a metal plate 108, which is grounded. A protein solution comprising protein dissolved in a water-alcohol-salt solvent as described herein is placed in the syringe 102. Similar to previous descriptions, an electrical potential can be applied to charge the protein solution by applying a voltage from a power supply 110 to the metal needle 104. A solution of a cross-linking agent such as EDC dissolved in a solvent such as ethanol can be placed in the receptacle 106.

For electrospinning, the electrical potential can be increased to grow the electrostatic forces and overcome the surface tension at a tip of the needle 104. As this surface tension is overcome, a fine jet of protein solution containing entangled protein chains can be drawn out of the needle 104. As the fine jet travels through the air, the solvent evaporates leaving a dry protein fiber that engages the surface of the cross-linking solution in the receptacle 106. The impact of the protein fiber's engagement with the surface of the cross-linking solution fractures the fiber into relatively short sections. Upon entering the cross-linking solutions, each short section of protein fiber draws inward and cross-links

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with itself, resulting in a generally spherical protein agglomerate or bead. In one example, the protein solvent comprises about 16 percent collagen by weight dissolved in a solvent of about one-to-one ratio by volume of PBS buffer (20×) to ethanol. A flow rate of about 1 ml/h is applied to the protein solution in the syringe **102**, a voltage of about 25 kV is applied to the metal needle **104**, and the metal plate **108** is spaced about 20 cm from the tip of the metal needle **104**. An cross-linking solution of EDC dissolved in ethanol is placed in the receptacle **106**. Such parameters form cross-linked protein agglomerates as shown in FIG. **9**. Such protein agglomerates can be, for example, more than 100 micrometers in diameter.

For electrospraying, the tip of the needle **104** and the grounded plate **108** can be placed closer together as compared to the described electrospinning method. Such positioning can result in the protein solution exiting the needle **104** and forming droplets of solution prior to entering the cross-linking solution in the receptacle **106**. Such droplets internally cross-link once entering the cross-linking solution and form spherical protein agglomerates or beads. Protein agglomerates formed by electrospraying are shown in FIG. **10**. Such protein agglomerates can be, for example, approximately 2 to 3 micrometers in diameter.

For a gravitation feed method, no electrical potential is needed. Gravity is used to draw beads of protein solution from the needle **104**. The beads fall into the cross-linking solution and internally cross-link forming generally spherical protein agglomerates. Alternatively, each bead can break up into smaller beads upon impact with the surface of the cross-linking solution. Protein agglomerates formed by the gravitational feed method are shown in FIG. **11**. Such protein agglomerates can be, for example, approximately 20 to 30 micrometers in diameter.

Parameters such as the distance between the tip of the needle **104** and the metal plate **108**, flow rate of protein solution from the needle **104**, voltage applied to the needle **104**, concentration of protein in the protein solution, concentration of salt in the protein solution, and ratio of alcohol to water in the protein solution can affect the size of protein agglomerates or beads. However, for comparatively similar parameters, electrospraying can produce the smallest protein agglomerates, gravity feed can produce protein agglomerates larger than electrospraying, and electrospinning can produce protein agglomerates larger than the gravitational feed method.

Protein dissolved in benign solvents as described herein can be used to form porous protein films, scaffolds and gels. In one example, a protein solution can be deposited in a receptacle so that the protein solution covers the bottom of the receptacle. A solution that includes a cross-linking agent such as EDC in ethanol is poured over the protein solution. In one example, the cross-linking solution comprises about 0.2 millimoles of EDC. The receptacle can be hermetically covered for a period of time, for example about 24 hours. Evaporation of the solution results in a protein film forming on the bottom of the receptacle. Some salt crystals may be present on the surface of the film. Such salt crystals can be removed by washing the film with deionized water, which can leach out the salt. Once the salt is leached out, the film is left with a porous structure that includes numerous pores that intersect forming a protein structure with an open network of pores. FIGS. **12A** and **12B** show porous films formed from the described method. The porous structure of the film can include pores that range from submicron in size to over 30 micrometers in size.

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Such a film with intersection pores can be suitable as a scaffold for cell repopulation or tissue growth. It will be understood that the intersecting pores mimic the ECM structure of human tissue and provide expanded surfaces on which cells and tissue can grow.

An example of another method of forming a protein structure with an open network of pores is hereafter described. A protein solution as described herein is prepared in a receptacle and stirred. As the protein solution is stirred, a cross-linking solution including a cross-linking agent such as EDC is deposited in the receptacle. Stirring continues until a protein cross-links and forms a gel in the receptacle. Once cross-linked the protein gel can be rinsed with deionized water to remove salts and alcohol from the gel. The gel is quenched in liquid nitrogen and frozen. The gel is placed in a vacuum chamber and water in or on the gel sublimates or otherwise evaporates. Such a method results in a low density protein scaffold with foam-like properties and an open network of pores. FIGS. **13A** and **13B** show a scaffold with an open network of pores formed from the described method. The pores as shown range in size from about 10 micrometers to about 50 micrometers. The size of the pores can be controlled by varying the protein content in the protein solution and the buffer to alcohol ratios.

Additional methods of forming cross-linked protein structures can be achieved by combining the formation of protein fibers and a delayed post-cross-linking treatment in a single step. In one example, a method of forming cross-linked proteins is an in-situ method of kinetically controlling cross-linking resulting in versatile processing of collagen and other proteins. This is to say that the formation of protein fibers is achieved such that a cross-linking process of the fibers is delayed. Such a method enables protein fibers to cross-link after the formation of such fibers, without the need for a post-production process. Such versatility in processing provides for methods that can be designed to produce structures, products, devices, etc. for specific purposes.

One such method includes adding NHS and EDC to solutions of protein dissolved in ethanol and PBS. The time it takes for a cross-linking protein solution to become hydrogel (i.e., gelation time (t_{gel}) can be dependent on the mole ratio of NHS to EDC. Gelation time can be defined as the time at which the shear storage (G') and loss (G'') moduli are approximately equal. The shear storage and loss moduli can be determined rheologically in dynamic oscillatory shear experiments. Gelation time as a function of mole ratios of NHS to EDC can be determined by using an AR-2000ex Rheometer. FIG. **14A** illustrates the relationship between gelation time and mole ratio of NHS to EDC based on rheological results, where the concentration of EDC is fixed at 200 mM. FIG. **14B** illustrates the relationship between the log of gelation time and mole ratio of NHS to EDC based on rheological results. As is shown in FIGS. **14A** and **14B**, the gelation time shows a substantial increase beginning at a NHS/EDC ratio of 1.5, with a generally linear relationship with NHS/EDC molar ratios of two-to-one or greater than two-to-one. For NHS/EDC mole ratios below two-to-one, protein solutions become gel relatively quickly. The addition of excess NHS (i.e., increasing the NHS to EDC ratio) can postpone or delay the cross-linking reactions.

In one example, increasing the ratio of NHS to EDC can increase the delay in cross-linking reactions from minutes to two to three hours or more. Such a delay can be used to control cross-linking by delaying the cross-linking so as to provide for the versatile processing of proteins such as collagen. During such a delay in cross-linking, the solution

of protein dissolved in a benign solvent can be processed into a variety of shapes and morphologies. For example, electrospinning of protein fibers from the benign solvent became possible, with crosslinking occurring in-situ afterward.

In one example a collagen hydrogel can be formed using the following method. A collagen solution can be formed by dissolving about 16 percent by weight of collagen in a solvent such as PBS buffer (20×) and alcohol with volume ratio of about one to one. Optionally, prior to adding the buffer, a bio-friendly cross-linker can be added to the alcohol. The cross-linker can be about 200 mM of EDC and 400 mM of NHS. A cross-linked collagen hydrogel can be formed in about 2 hours. The collagen hydrogel can be rinsing in water to remove alcohol and salts after gel formation. FIGS. 15A and 15B depict collagen hydrogel formed at least partially by the foregoing method. FIG. 15A depicts hydrated collagen gel, and FIG. 15B depicts lyophilized collagen gel.

Collagen foam can be formed by lyophilizing the in-situ cross-linked hydrogel. Such a lyophilized collagen gel can have a sponge-like structure that can include pore sizes ranging from approximately several microns to approximately several tens of microns. FIG. 16A is a scanning electron microscope (SEM) image of a cross section of lyophilized in-situ cross-linked collagen gel; FIG. 16B is an SEM image of a top portion of FIG. 16A; FIG. 16C is an SEM image of a middle portion of FIG. 16A; and FIG. 16D is an SEM image of a bottom portion of FIG. 16A. Such a collagen porous structure can benefit cell culturing and proliferation. Such a collagen porous structure can also improve collagen gel performance in tissue engineering.

Collagen gels can be fabricated in different shapes, sizes and configurations. In one example, collagen hydrogel can be fabricated as a tube-like structure. One method of forming such a tube-like collagen hydrogel is to pour or otherwise deposit the above-described collagen solution into a mold prior to the onset of gelation. As shown in FIG. 17A, collagen hydrogels can be formed into tube-like structures of varying lengths and thicknesses. The structures shown in FIG. 17A are immersed in water. In another example, collagen gels can be fabricated as a hemispheric-like structure via in-situ cross-linking methods. Such a hemispheric-like structure is shown in FIG. 17B. The structure shown in FIG. 17B is immersed in water. Such a hemispheric-like structure can be applicable in the field of ocular medication. For example, collagen gels can be fabricated as a contact lens-like shaped structure.

Furthermore, before the onset of gelation the collagen hydrogel can still be liquid-like in nature. In such a liquid-like state, collagen hydrogel can be used for a number of different procedures such as, for example, printing or coating. The collagen hydrogel can be used as an "ink" that is used to print on a glass slide. As shown in FIG. 18, collagen hydrogel can be useful as an "ink" to print on a glass slide.

In another example, drugs or medications can be incorporated into collagen gels for sustained drug delivery. Various drugs can be loaded into the collagen solution prior to cross-linking so as to entrap or embed the drug after gel formation. In addition to drugs and medications, many polymeric materials can be entrapped or embedded into the collagen gel. Many different drugs can be loaded into embedded polymer materials and/or collagen gel. The drugs can be released simultaneously with tunable release profiles. In one example, electrospun poly(caprolactone) (PCL) containing photodynamic therapy drug Pc4 and electrosprayed PLGA macrobeads with beta-carotene can be successfully

embedded into in-situ cross-linked collagen hydrogels. Such an arrangement is shown in FIGS. 19A and 19B. FIG. 19A depicts in-situ cross-linked collagen hydrogels with PCL-Pc4 mat embedding. FIG. 19B depicts in-situ cross-linked collagen hydrogels with PLGA-beta carotene macrobeads. The collagen hydrogel samples of both FIG. 19A and FIG. 19B are immersed in water.

The delay in cross-linking reactions due to NHS/EDC ratios provides a period of time for a number of processes prior to the collagen becoming cross-linked. Such a delay provides for a variety of methods that incorporate additional processing during the formation of collagen hydrogels. The incorporation of addition processing allows for collagen hydrogels to be designed to serve any number of functions and purposes.

In one example, an electrospun in-situ cross-linked collagen fibrous scaffold can be fabricated by using electrospinning techniques. Such techniques can begin with a benign solvent. In one example, a collagen solution can be prepared by dissolving about 16 percent by weight of Type I collagen in a solvent such as PBS buffer (20×) and alcohol with volume ratio of about one to one. Prior to adding the buffer, the crosslinker can optionally be added to the alcohol. The crosslinker can be, for example, about 400 mM of NHS and about 200 mM of EDC.

As similar described with regard to FIG. 1, the mixed collagen solution can be loaded to a 5 ml BD syringe with a 21 gauge blunt needle, which can then be placed in a syringe pump. The process parameters (such as flow rate, potential field, and needle-collector distance) can be varied to optimize the stability of the electrostatic jet. Electrospinning can be carried out at 20 KV with a pump rate of 0.5 ml/h. A drum rotating at 5 m/s is placed 12 cm far from the needle in order to collect the electrospun fibers. Electrospun collagen fibrous scaffolds fabricated with such a process are shown in FIGS. 20A and 20B. FIG. 20A depicts an electrospun in-situ cross-linked collagen fibrous scaffold in a dry state. FIG. 20B depicts an electrospun in-situ cross-linked collagen fibrous scaffold in a hydrated state immersed in water.

Such an in-situ cross-linking electrospinning method can generate generally cylindrical and continuous collagen nanofibers in random arrays with a generally porous structure. FIGS. 21A and 21B depict such electrospun in-situ cross-linked collagen fibers at two different magnifications. FIG. 21C is a histogram of the diameter of electrospun in-situ cross-linked collagen fibers, and FIG. 21D is a chart of average fiber diameter and a function electrospinning time. The average fiber diameter is approximately 0.42 ± 0.11 micrometers. The diameter of in-situ cross-linked collagen fibers increased from 0.35 to 0.46 micrometers during approximately three hours of electrospinning (as shown in FIG. 21D), which suggests that the onset of cross-linking occurs during that time. The average fiber diameter of 0.42 micrometers is about twice the diameter of collagen fibers generated from ethanol/PBS without cross-linking. The addition of EDC into the collagen solution can increased the viscosity, even though macroscopic gelation was postponed by the presence of excess amount of NHS. A higher viscosity of collagen solution can result in the formation of larger fibers. Similar results were also found in rheological experiments. During testing the storage modulus (G') of in-situ crosslinking collagen solution increased until the dynamic oscillatory shear testing finished, which indicates that the viscosity of solution gradually increased after mixing the collagen and the crosslinkers together.

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Humidity levels can be an important in-situ cross-linked collagen nanofiber scaffolds were tested at relative humidities of 33%, 43% and 53% at room temperature. SEM images of electrospun, in-situ crosslinked collagen fibers tested at these different humidity levels for certain periods of time are shown in FIGS. 22A-C. FIG. 22A depicts a collagen fiber scaffold stored at 33% humidity for three days. FIG. 22B depicts a collagen fiber scaffold stored at 43% humidity for three days. FIG. 22C depicts a collagen fiber scaffold stored at 53% humidity for 1 day. As is shown, relative humidity can be an important parameter to control the fiber morphology of in-situ cross-linked collagen after electrospinning. A collagen fiber scaffold stored at 33% humidity for three days (FIG. 22A) includes individual fiber features. A collagen fiber scaffold stored at 53% for one day (FIG. 22C) includes some melting of fibers. A higher humidity can plasticize the collagen before extensive crosslinking by incorporated EDC and NHS, while lower humidity may limit the motilities of the crosslinkers and collagen chains in the fibers.

Post-crosslinking of collagen fiber scaffolds was also performed by immersing the collagen scaffolds electrospun from ethanol/PBS without crosslinkers into 200 mM each of EDC and NHS in ethanol for 4 hours. After the post-crosslinking process, the resulting collagen scaffolds shrank to 40% of the original dimensions when immersed in the water, as shown in FIGS. 23A and 23B. In-situ cross-linked collagen scaffolds can swell up to two-fold of the original size when they were hydrated (see FIGS. 20A and 20B). The water-treated, in-situ cross-linked collagen scaffolds include a porous structure, as shown in FIGS. 24A and 24B. EDC and NHS are more uniformly distributed in collagen fibers when using the in-situ method described herein and cross-linking reactions can occur throughout the fibers, which can result in more homogeneously crosslinked collagen scaffolds.

Another example of a collagen fibrous scaffold that is relatively dense and water-insoluble obtained after electrospinning is depicted in FIGS. 25A and 25B. The collagen fibrous scaffold depicted was in-situ cross-linked three days after electrospinning and prior to any water treatment. FIGS. 25A and 25B depict a relatively strong network with a significant extent of interlinked fibers. Such a condition can be a result of inter-molecular and intra-molecular covalent bonds and bonding between the fiber junctions. Using the methods disclosed herein, in-situ cross-linked collagen scaffolds generally preserve their shape and also swell to some extent when hydrated (as shown in FIGS. 20A and 20B). In addition, the fibrous structure of in-situ cross-linked collagen mats produced by the methods disclosed herein can largely be conserved after water-treatment. FIGS. 25C and 25D depict in-situ cross-linked collagen fibers three days after electrospinning and after water treatment. Such behavior can be desirable and beneficial for cell culture and tissue engineering applications. The advantages of in-situ cross-linked collagen scaffolds as described herein can be ascribed to the efficiency and homogeneity of cross-linking when EDC/NHS is incorporated within the fibers during electrospinning. With regard to post-cross-linking, the cross-linking reaction is mainly concentrated on the surface of collagen fibers because of the slow diffusion of EDC and NHS into the fibers, which is increasingly retarded as cross-linking continues at or near the surface.

Uniaxial tensile stress tests of in-situ cross-linked collagen scaffolds were performed on a dynamic mechanical analyzer (DMA-Q800, TA Instruments Inc.). Representative plots of the stress-strain curves of dry scaffolds are shown in

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FIG. 26 and hydrated scaffolds in FIG. 27. Respective tensile properties in terms of peak stress, strain at break, and tangential modulus for dry scaffolds are summarized in Table 1 below. The tensile results indicate that dry in-situ cross-linked collagen has greater peak stress and strain compared to those of post-cross-linked samples. Both high cross-link content and largely fibrous texture of dry in-situ cross-linked collagen scaffolds can be responsible for such differences.

TABLE 1

Tensile properties of post and in-situ cross-linked collagen membranes			
	Peak Stress (MPa)	Strain at break (%)	Tangential Modulus (MPa)
In-situ cross-linked collagen	14.53 ± 1.61	3.28 ± 0.12	654.29 ± 1.29
Post cross-linked collagen	11.33 ± 4.77	1.66 ± 0.09	778.40 ± 1.48

For hydrated in-situ cross-linked scaffolds, the peak stress is about 0.22±0.02 MPa and strain at break is about 134.5±10.0 percent.

FTIR experiments have been conducted to explore the collagen inner structure of electrospun in-situ cross-linked scaffolds. In a typical FTIR spectrum of collagen, the amide I absorption arises predominantly from protein amide C=O stretching vibrations, the amide II absorption is made up of amide N—H bending vibrations and C—N stretching vibrations; the amide III peak is complex, consisting of components from C—N stretching and N—H in plane bending from amide linkages, as well as absorptions originating from wagging vibrations from CH₂ groups from the glycine backbone and proline side-chains. In FIG. 24, the indicative bands at 1653, 1541 and 1235 cm⁻¹, which are characteristic of the amide I, II and III absorptions, respectively, are the preliminary indication that the unique triple-helical structure of native collagen presents in the in-situ cross-linked scaffolds. This result is also confirmed by the value of IR absorption ratio (0.86) between the 1235 (amide III) and 1450 bands, which is larger than 0.59.

Regarding the mechanism of in-situ cross-linking by EDC/NHS, the amine-reactive O-acylisourea intermediate, which is formed by the reaction between EDC and carboxyl groups of collagen molecules, is susceptible to hydrolysis, making it unstable and short-lived in aqueous solution. The addition of small amount of NHS can stabilize the amine-reactive intermediate by converting it to an amine-reactive NHS ester, thus increasing the efficiency of EDC-mediated coupling reactions. The amine-reactive NHS ester can be more stable but less active than the amine-reactive EDC ester. Therefore, the addition of excess NHS can convert too many amine-reactive EDC esters to the less active NHS esters, which results in decreasing EDC activity and finally postpones the full cross-linking reaction. Also, excess NHS can compete with amines for NHS esters, the former leading to “NHS exchange” and delaying formation of amide bonds. The disclosed in-situ cross-linking technique can also have applicability with other biomaterials including polypeptides and proteins and polysaccharides, to produce versatile structures for tissue engineering.

Collagen and other protein-based gels can be formed by filling cavities of microfluidic devices followed by cross-linking. Such a method can be useful for 3-D cell culturing. Also, the cross-linked gel can be removed from the device forming effectively a “microfluidically printed” structure.

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The delayed cross-linking chemistry can be useful in functionalizing collagen and other protein for later reactions different from the carboxylic acid/amine coupling that occurs in the systems and methods described above. For example, it may be possible to functionalize collagen with, for example, acrylates by reaction of methyl acrylate and collagen amine units in the presence of EDC/NHS. The collagen could then be cross-linked later, for example by UV irradiation. One advantage is the possibility of selective cross-linking depending upon which regions are irradiated.

The foregoing description of examples has been presented for purposes of illustration and description. It is not intended to be exhaustive or limiting to the forms described. Numerous modifications are possible in light of the above teachings. Some of those modifications have been discussed, and others will be understood by those skilled in the art. The examples were chosen and described in order to best illustrate principles of various examples as are suited to particular uses contemplated. The scope is, of course, not limited to the examples set forth herein, but can be employed in any number of applications and equivalent devices by those of ordinary skill in the art.

What is claimed is:

1. A method of forming a protein structure comprising:
forming a solution comprising water, alcohol, salt, collagen, and a cross-linker;
forming a plurality of collagen fibers from the solution;
and
after the passage of a period of time, crosslinking at least a portion of the plurality of collagen fibers to form a protein structure.

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2. The method of claim 1, wherein the cross-linker is 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide hydrochloride (EDC) and N-hydroxysuccinimide (NHS).

3. The method of claim 2, wherein the molar ratio of NHS to EDC is greater than about 1.5:1.

4. The method of claim 3, wherein a concentration of EDC is about 200 nM and a concentration of NHS is about 400 nM.

5. The method of claim 3, wherein the salt is a PBS buffer (20×) and a volume ratio of PBS buffer (20×) to alcohol is about 1:1.

6. The method of claim 5, wherein the collagen is dissolved in the solution.

7. The method of claim 1, wherein the plurality of collagen fibers are formed by an electrospinning method.

8. The method of claim 7, wherein the electrospinning method results in the plurality of collagen fibers generally aligned after electrospinning.

9. The method of claim 1, wherein the protein structure is placed in an environment where the relative humidity is between about 40% and about 60%.

10. The method of claim 1, wherein the protein structure is immersed in a solution of salt, alcohol and water.

11. The method of claim 2, wherein the molar ratio of NHS to EDC is about 1:1.

12. The method of claim 6, wherein the collagen dissolved in the solution is less than about 25 percent by weight.

13. The method of claim 12, wherein the collagen dissolved in the solution is less than about 16 percent by weight.

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