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# Raucher et al.

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# (54) INHIBITION OF METASTASIS BY CELL PENETRATING PEPTIDES

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- (\*) Notice: Subject to any disclaimer, the term of this patent is extended or adjusted under 35

U.S.C. 154(b) by 673 days.

This patent is subject to a terminal dis-

claimer.

- (21) Appl. No.: 12/789,236
- (22) Filed: May 27, 2010

### Related U.S. Application Data

- (63) Continuation-in-part of application No. 12/422,975, filed on Apr. 13, 2009, now abandoned, which is a continuation-in-part of application No. 12/162,283, filed as application No. PCT/US2007/061240 on Jan. 29, 2007, now Pat. No. 8,252,740.
- (60) Provisional application No. 61/044,398, filed on Apr. 11, 2008, provisional application No. 60/762,919, filed on Jan. 27, 2006.
- (51) Int. Cl. A61K 38/00 (2006.01)

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#### (57) ABSTRACT

A compound including a cell penetrating peptide (CPP) and elastin-like polypeptide (ELP), and a method for use thereof, are useful for inhibiting the proliferation of cancer.

#### 8 Claims, 12 Drawing Sheets

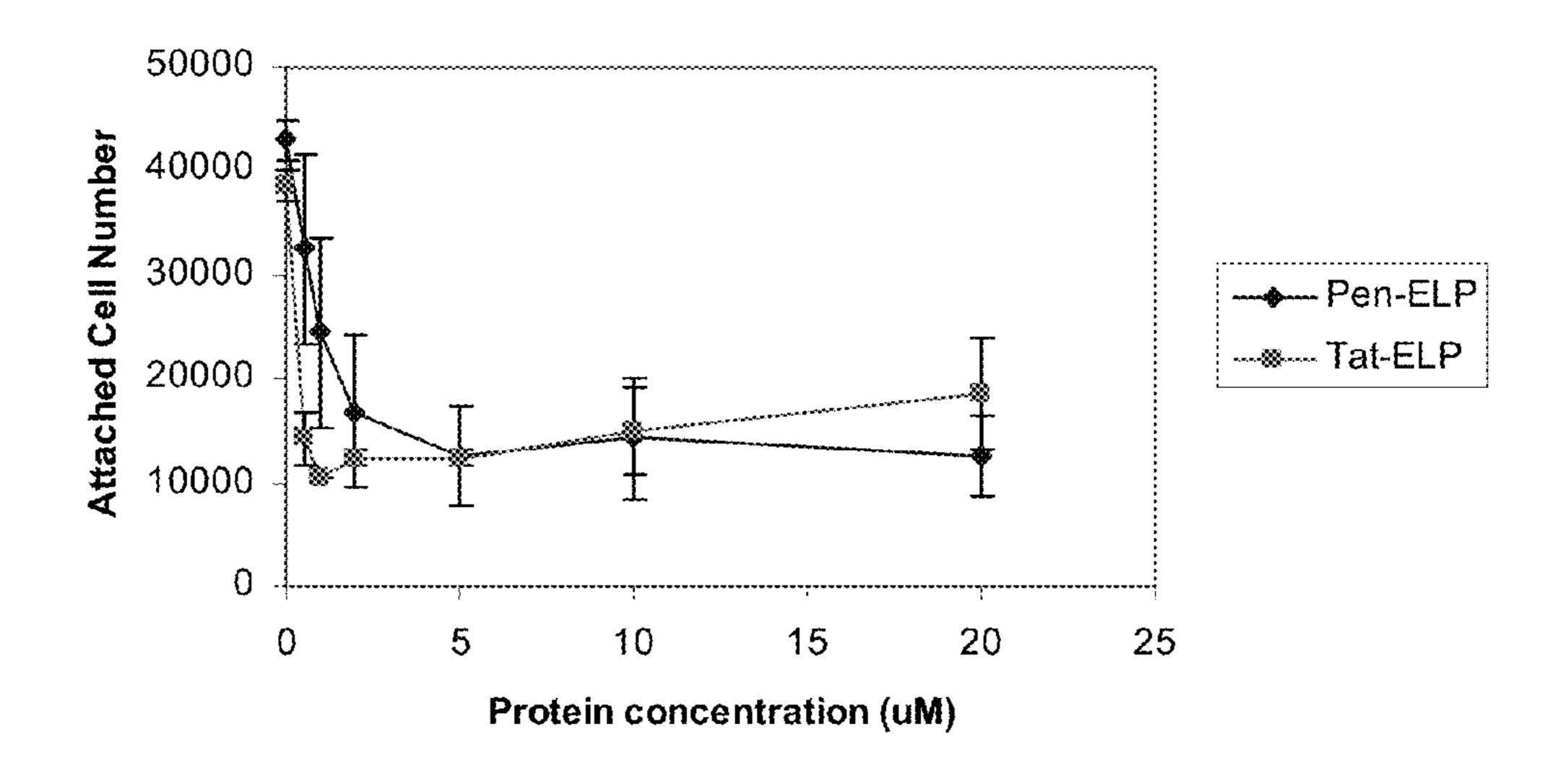


Figure 1.

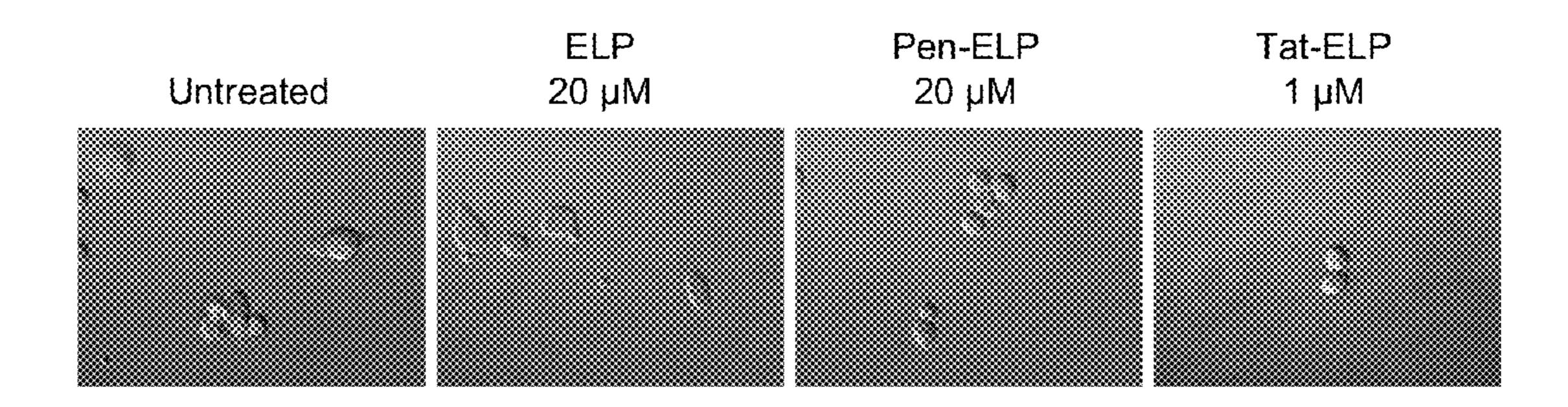


Figure 2.

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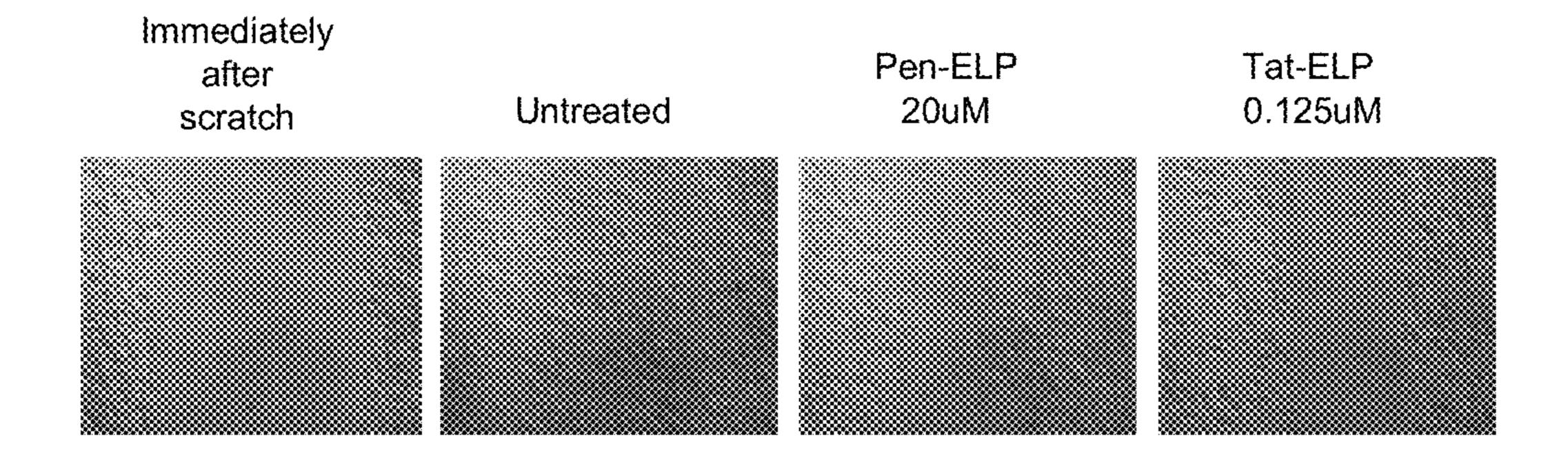


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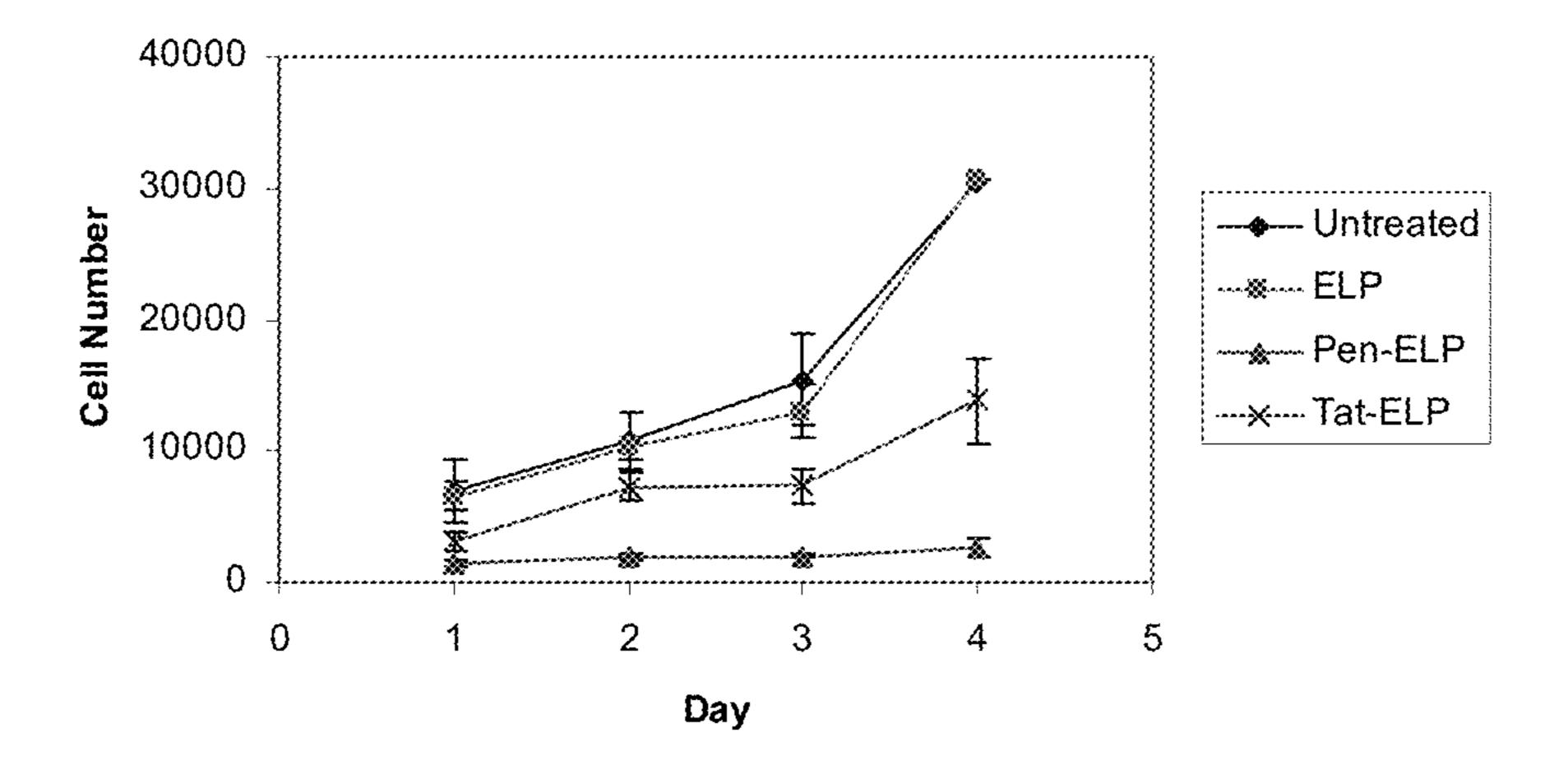


Figure 4.

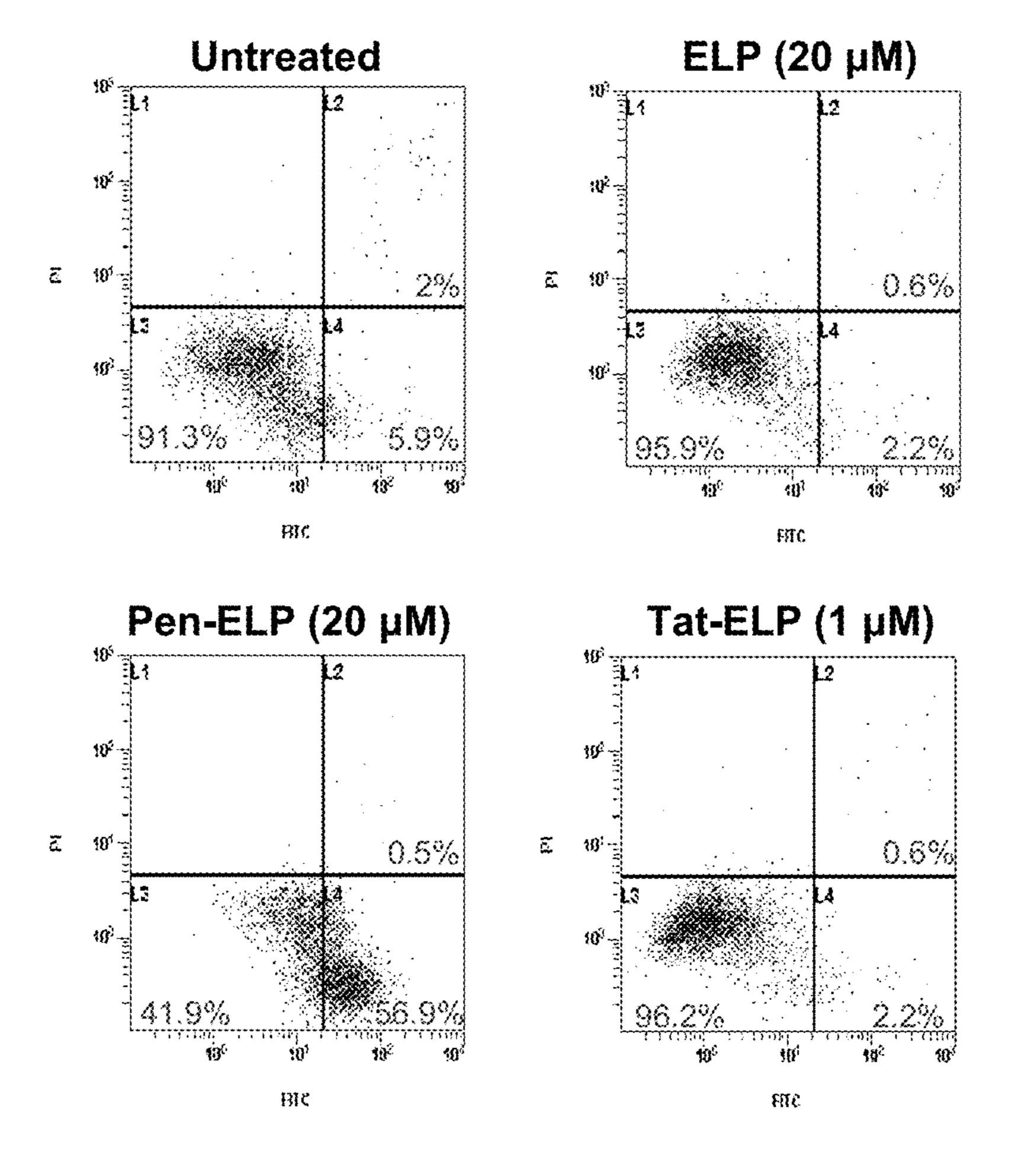


Figure 5.

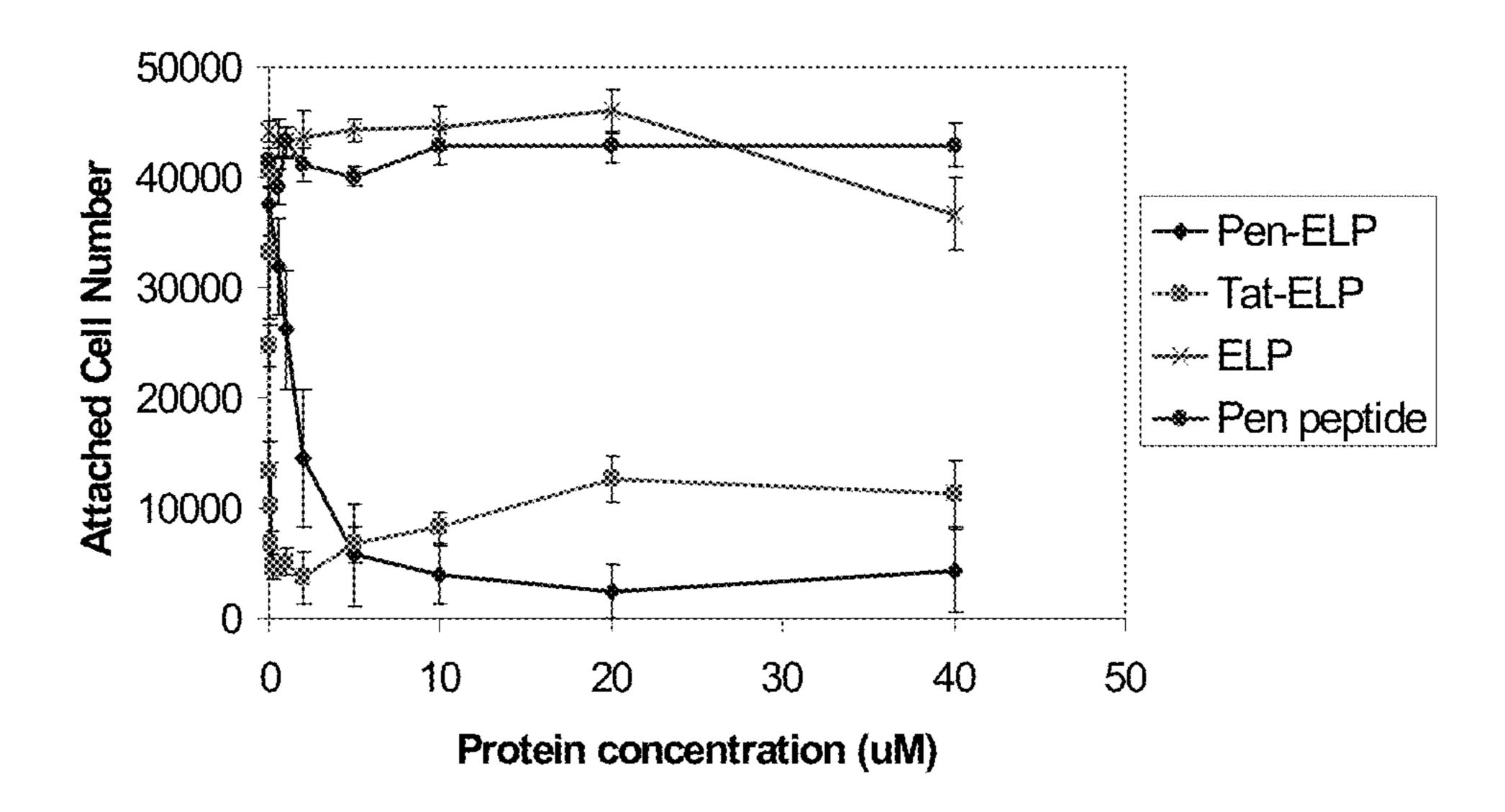


Figure 6.

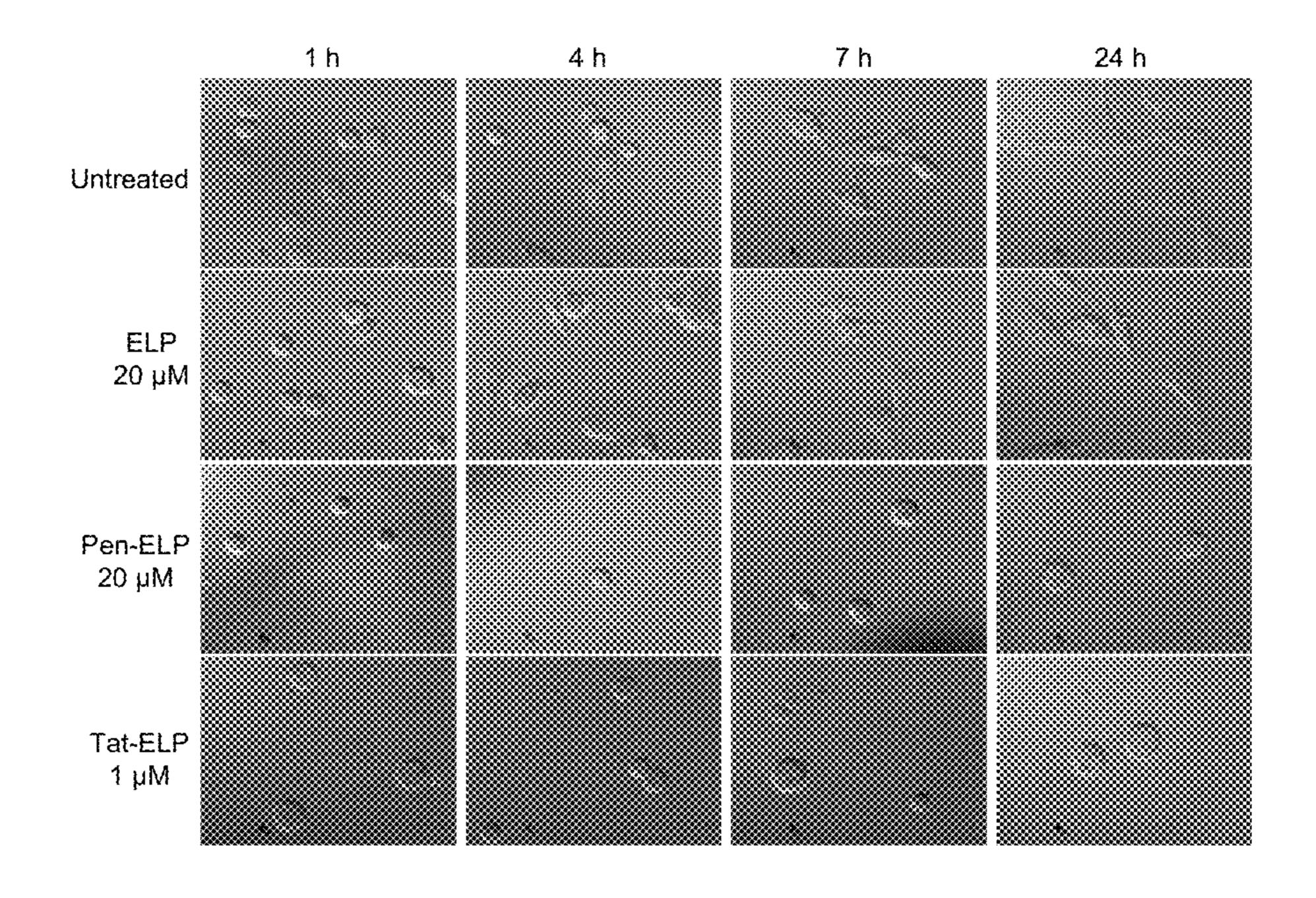


Figure 7.

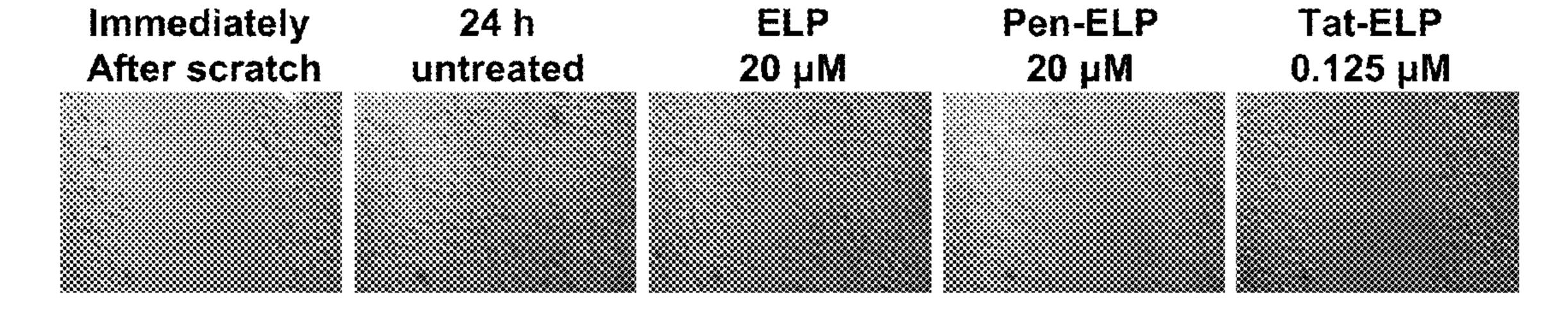


Figure 8.

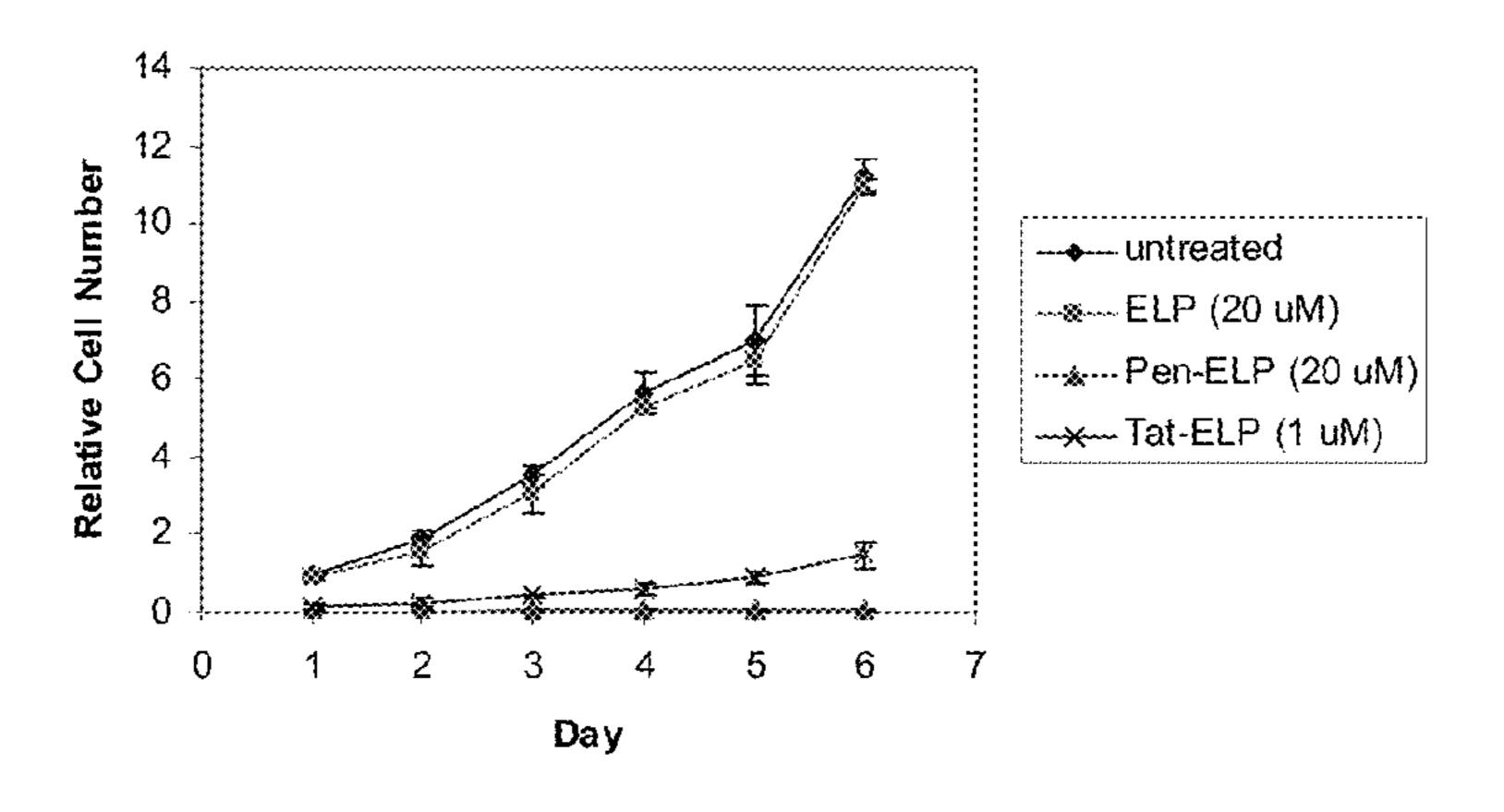


Figure 9.

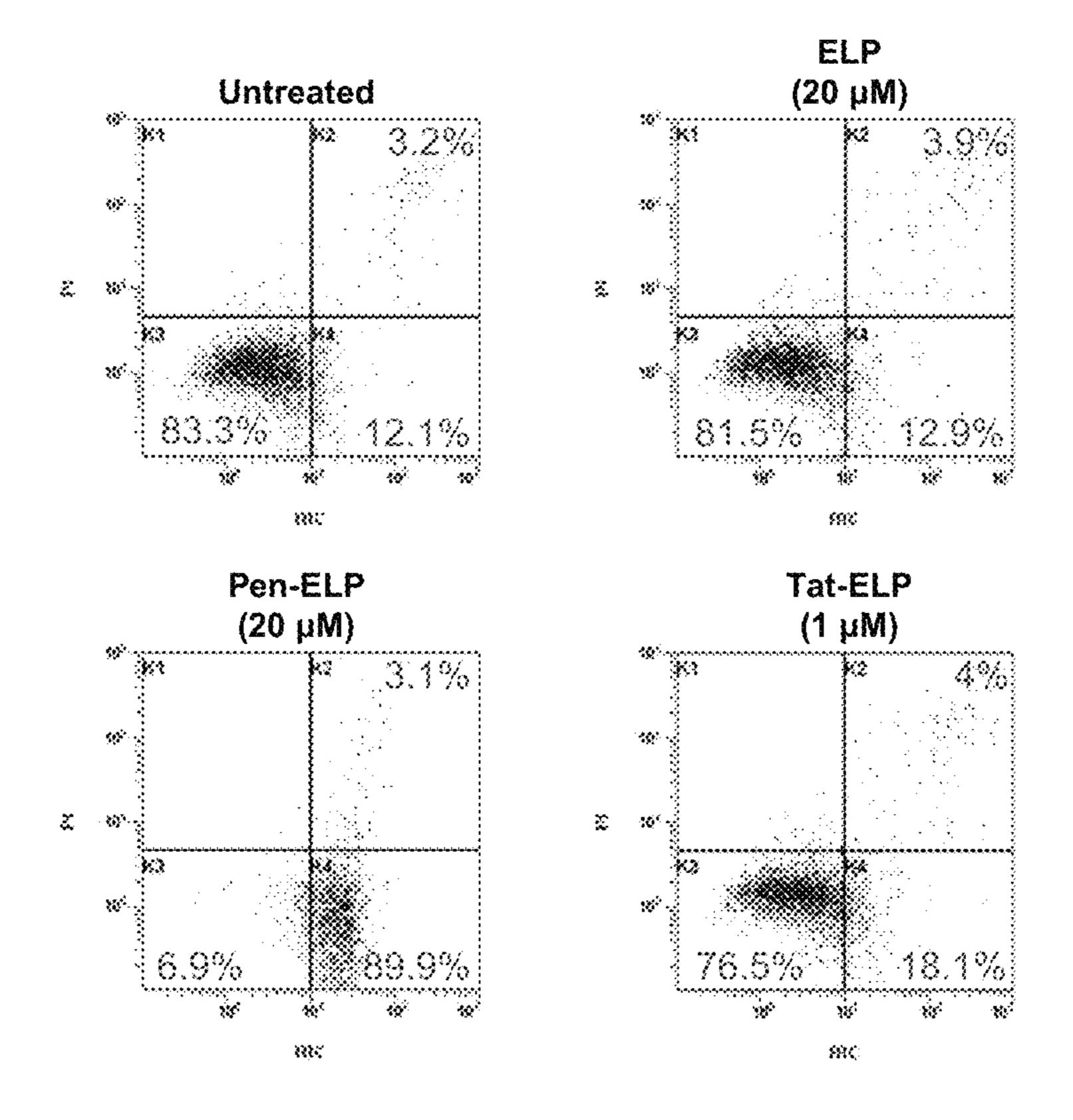


Figure 10.

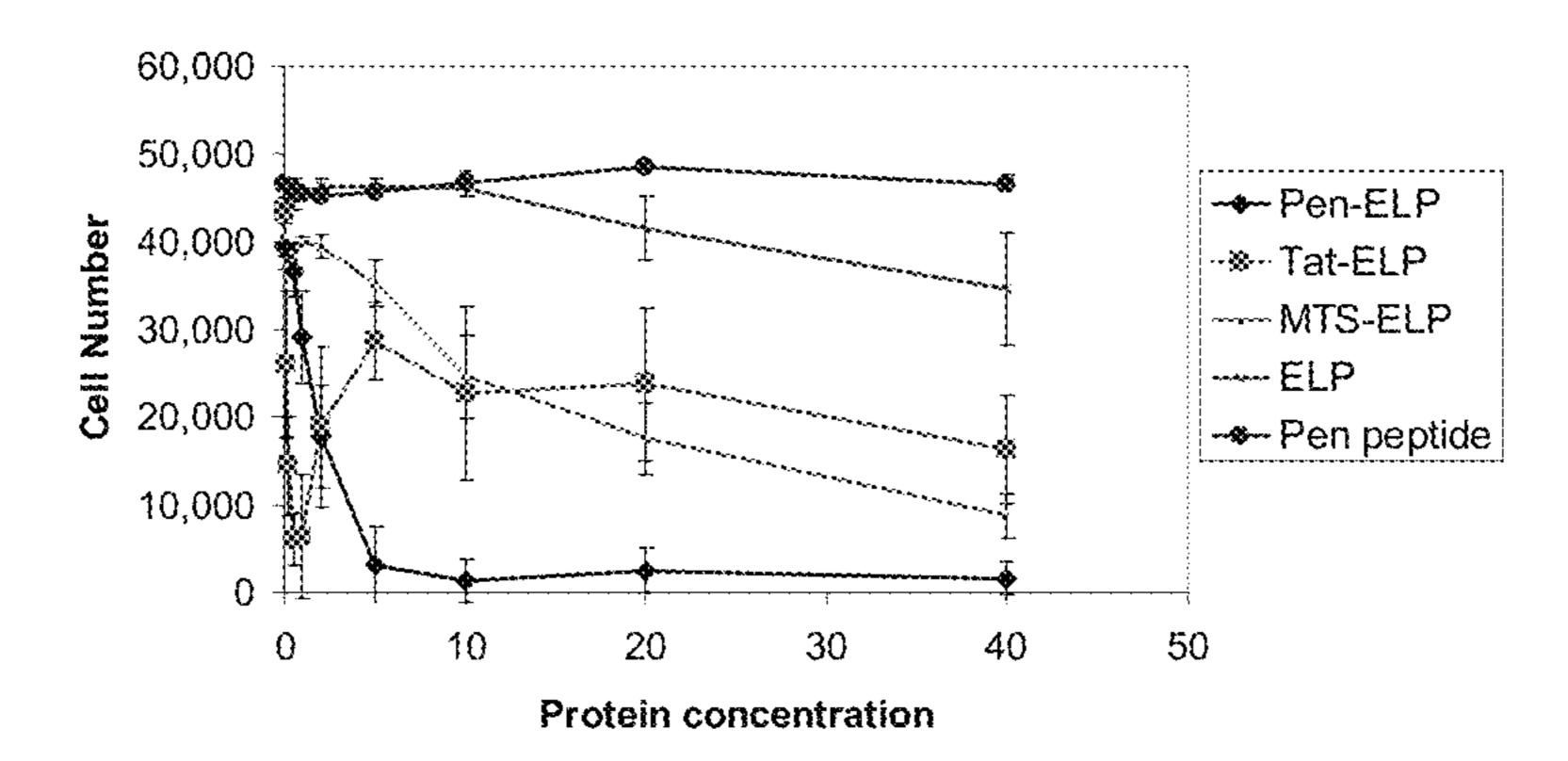


Figure 11.

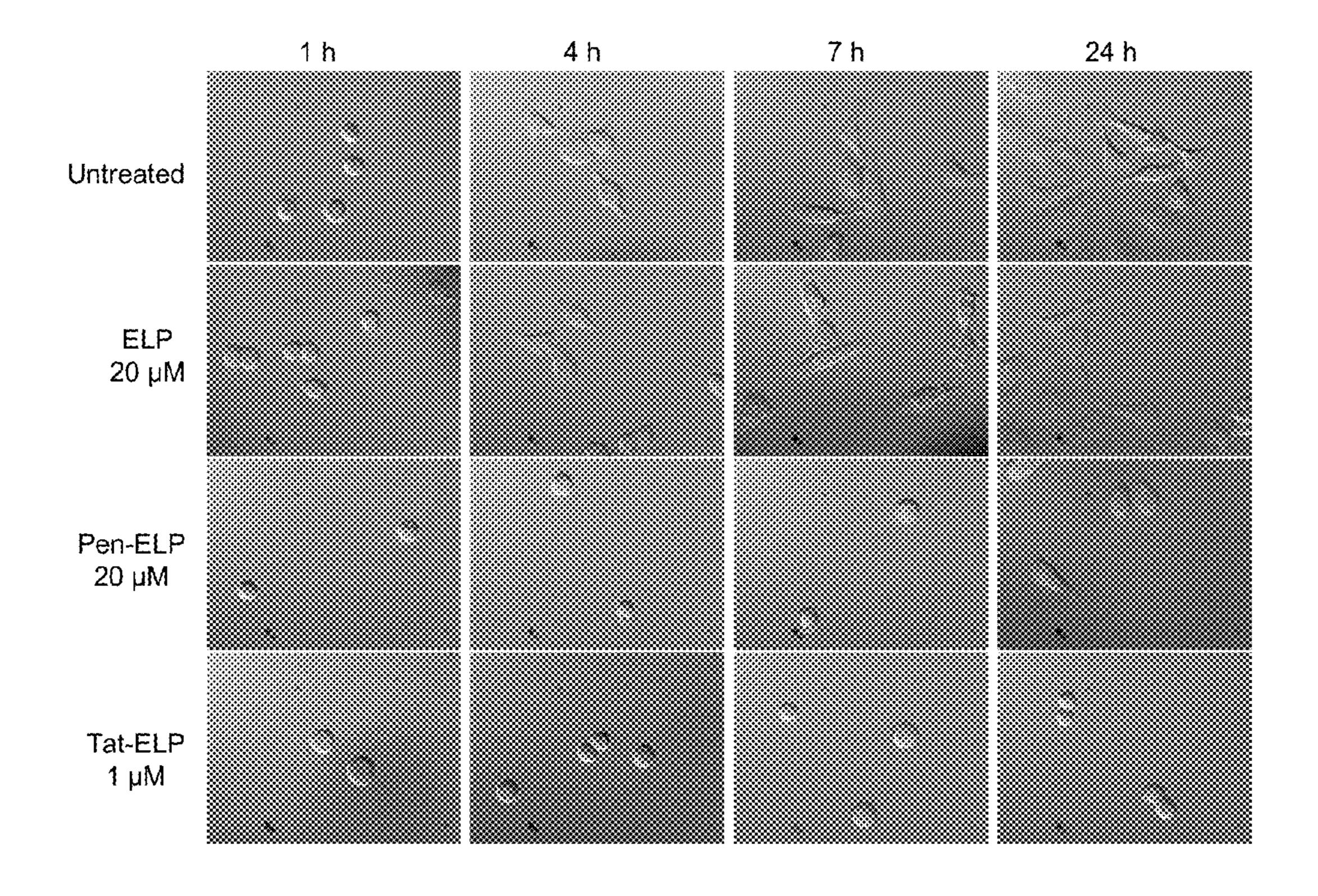


Figure 12.

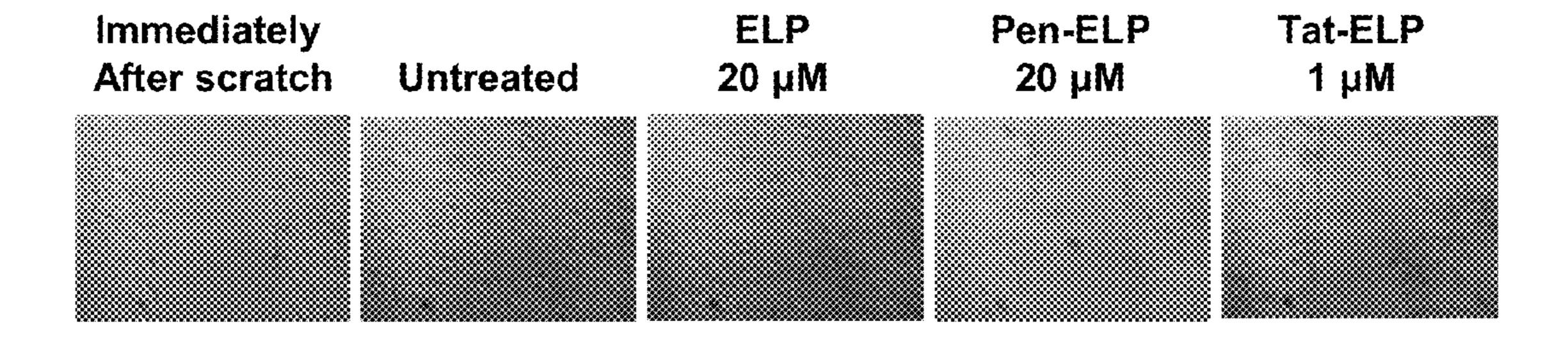


Figure 13.

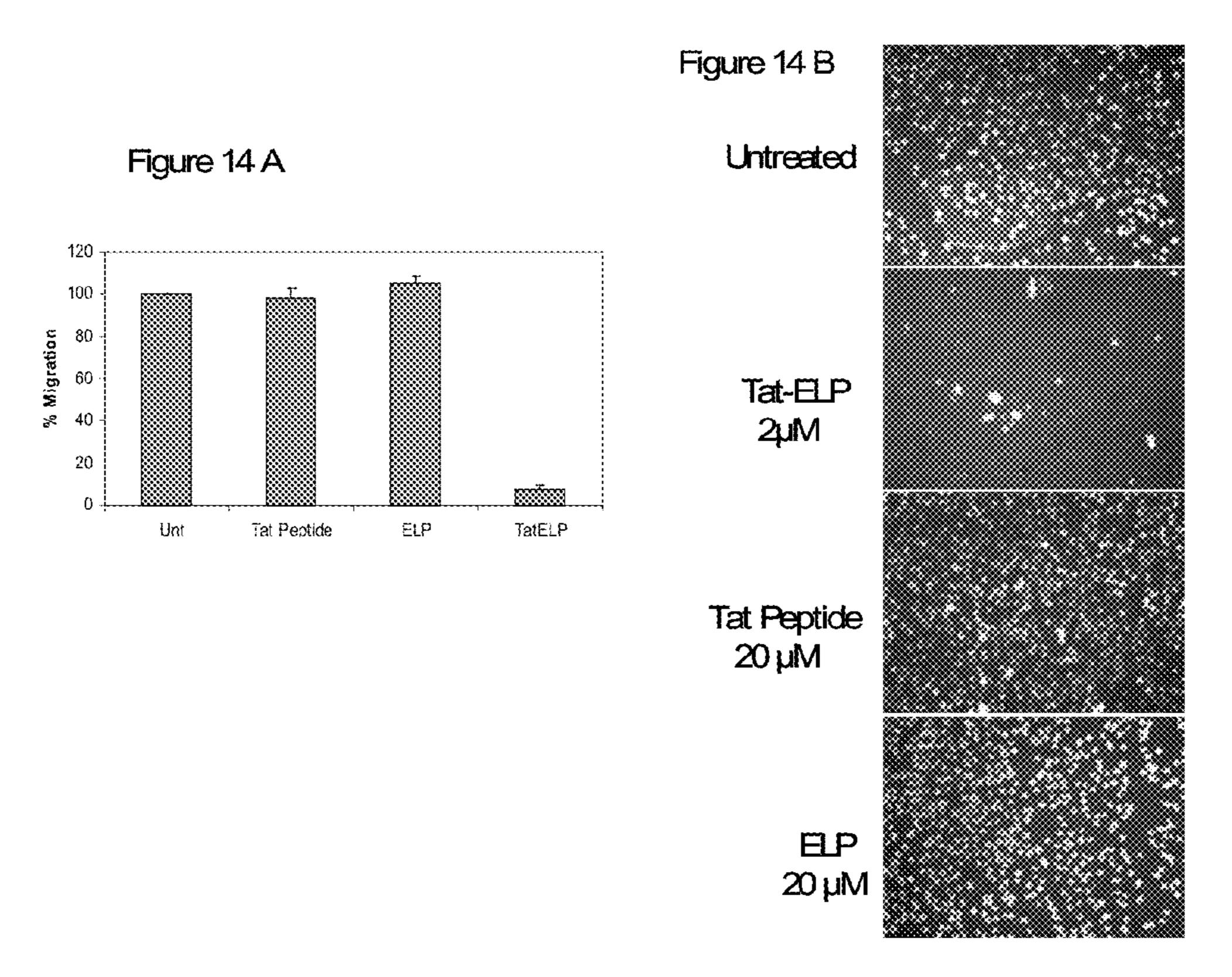


Figure 14.

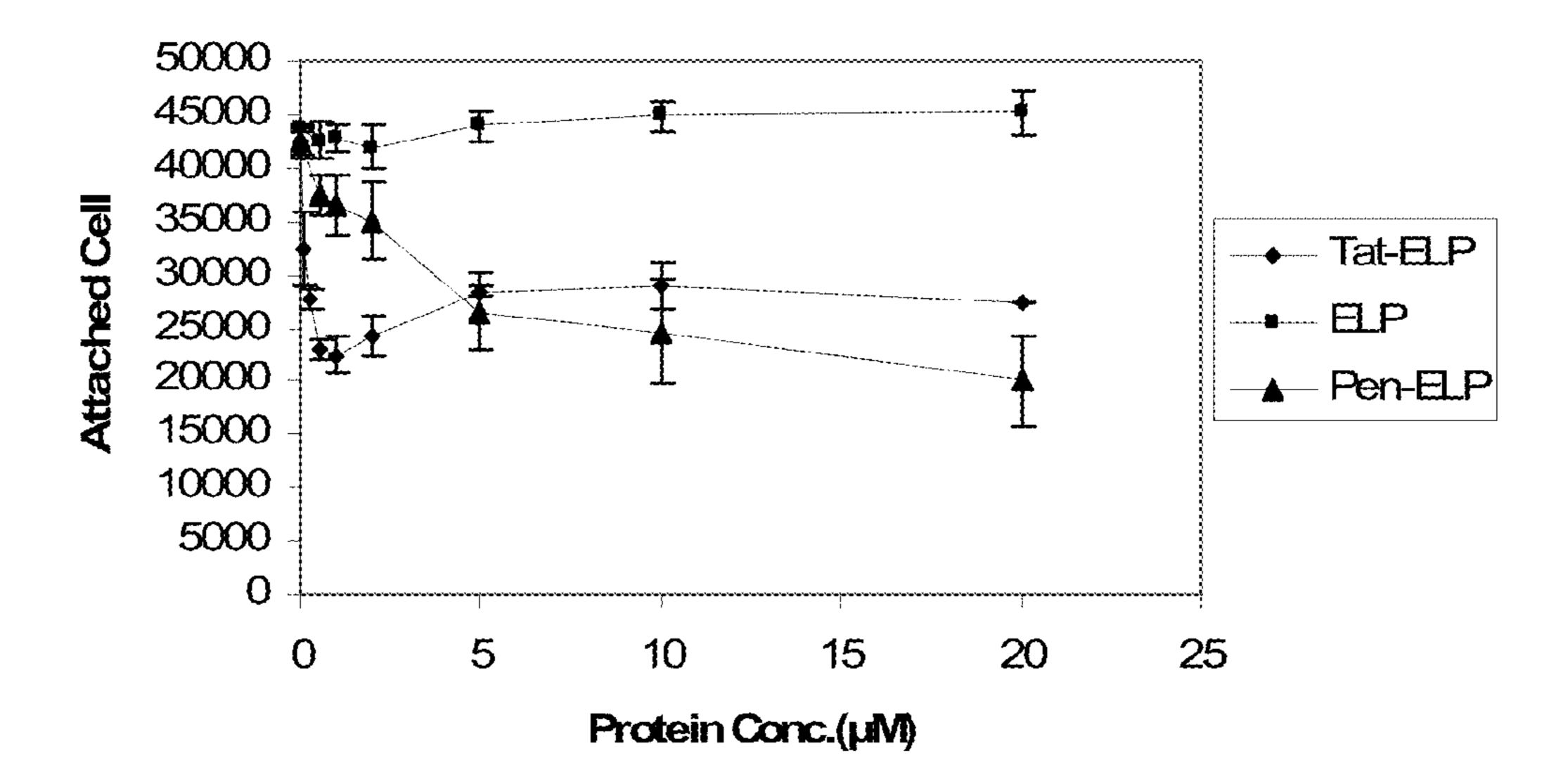


Figure 15.

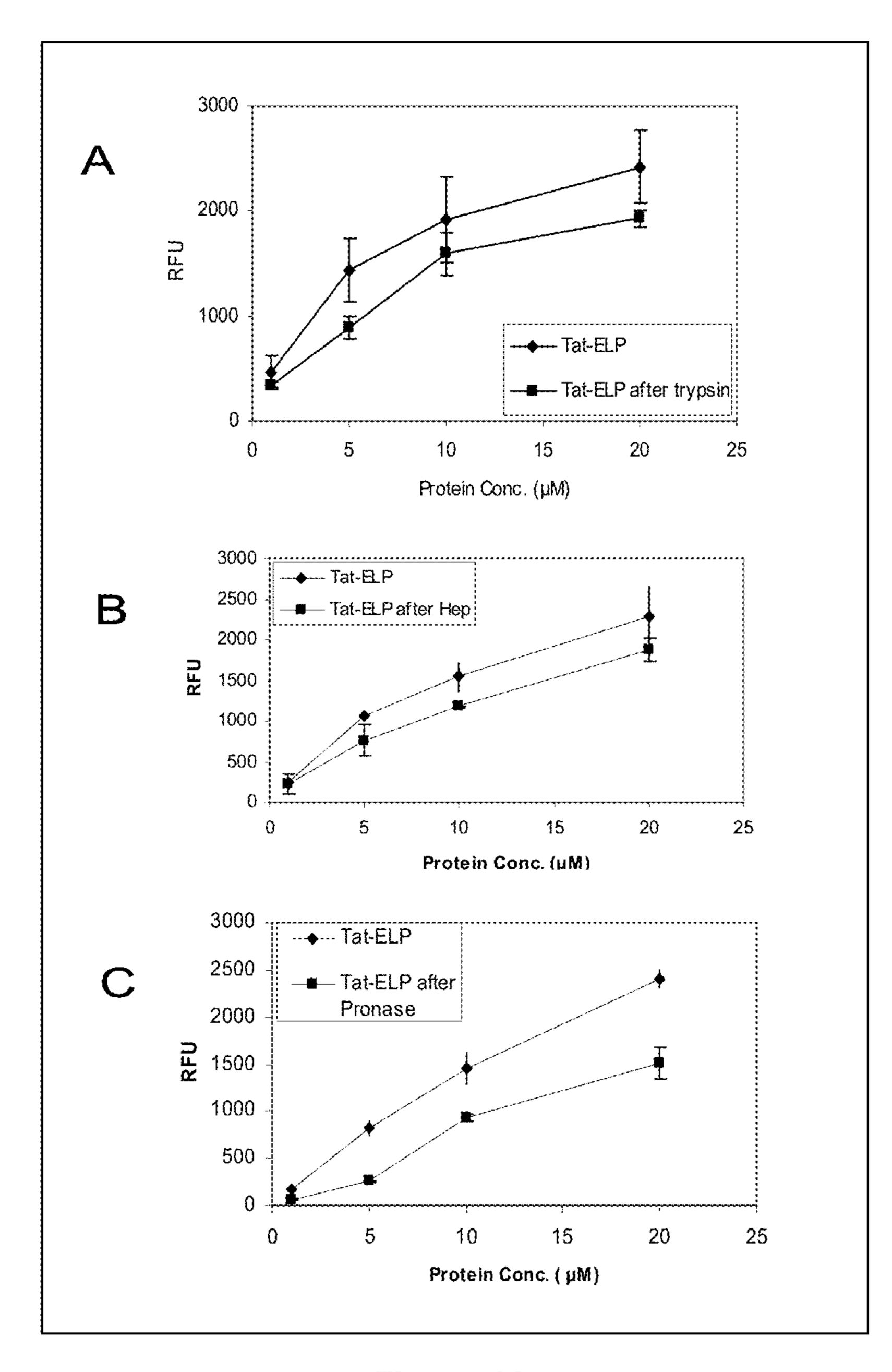


Figure 16.

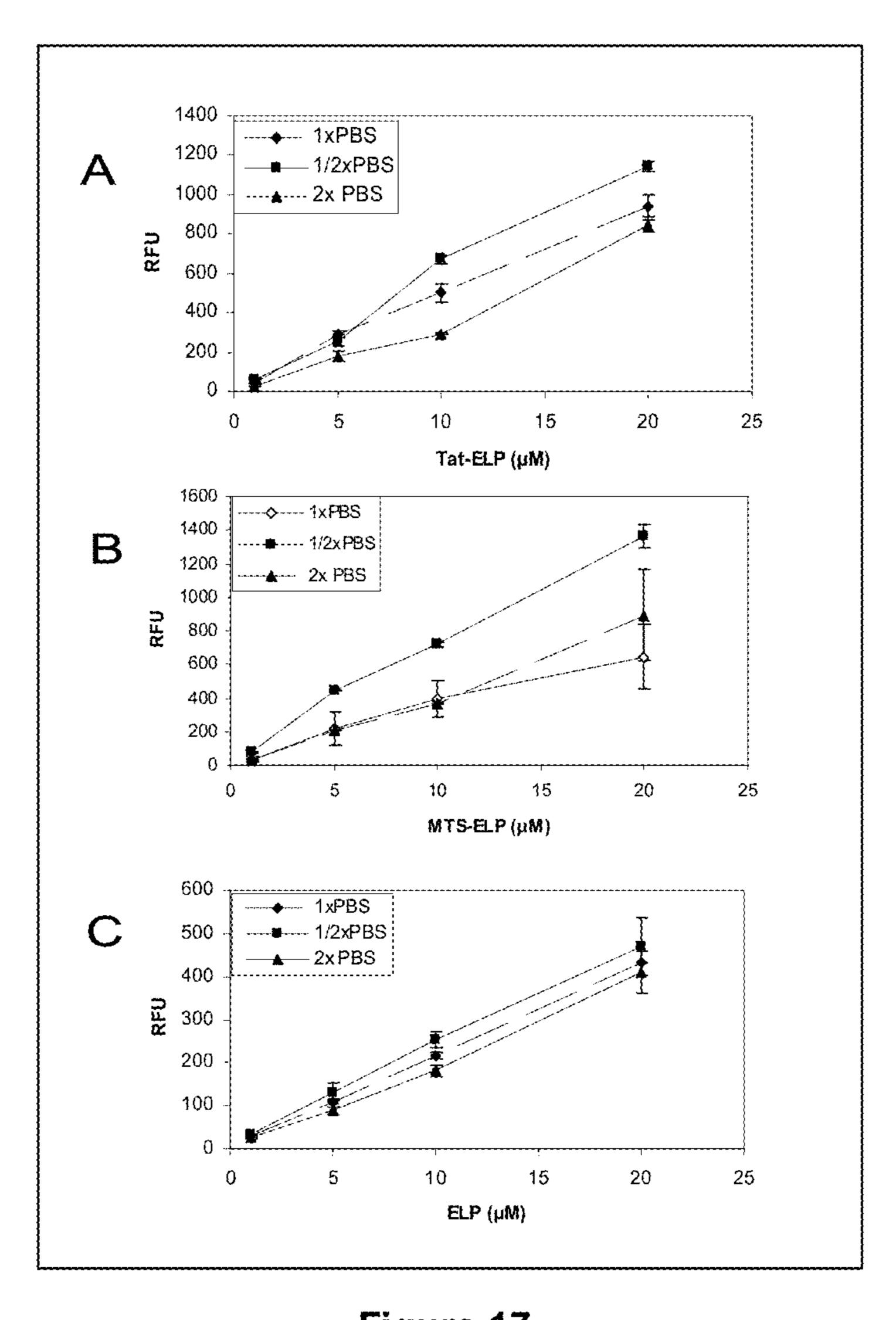


Figure 17.

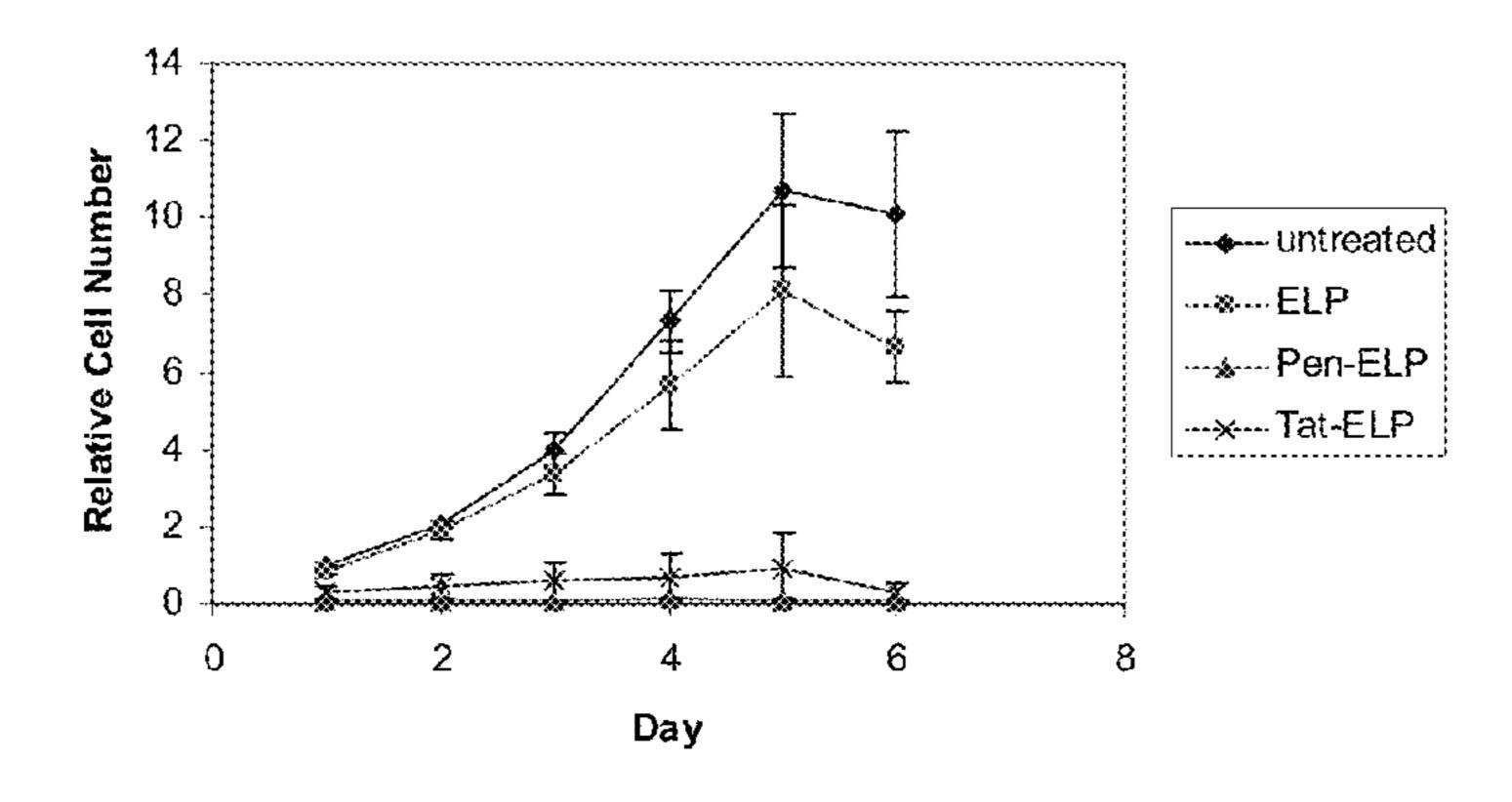


Figure 18.

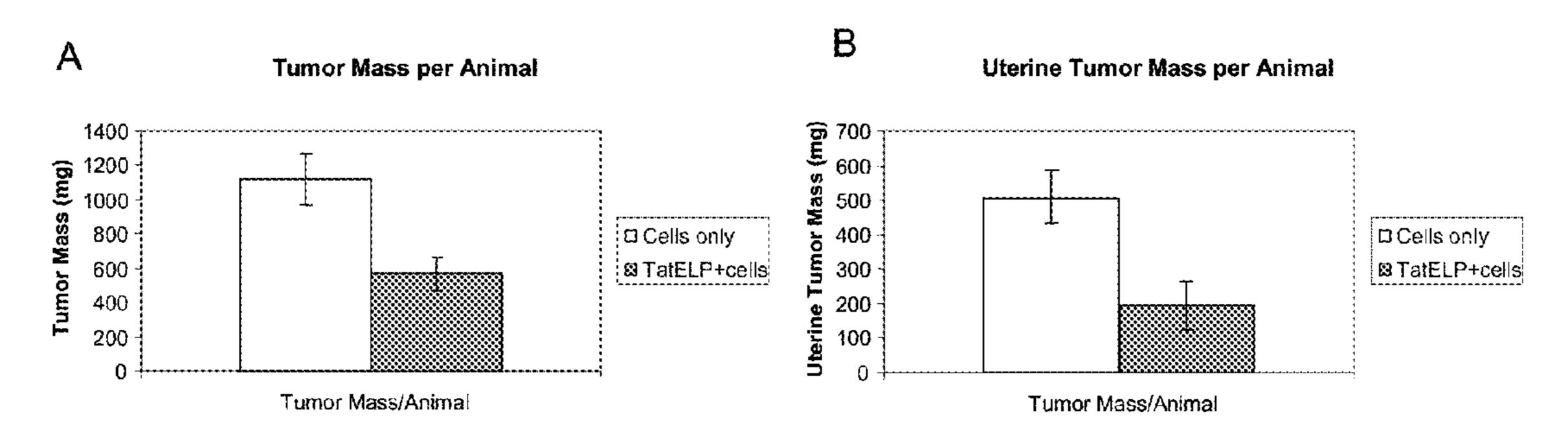


Figure 19.

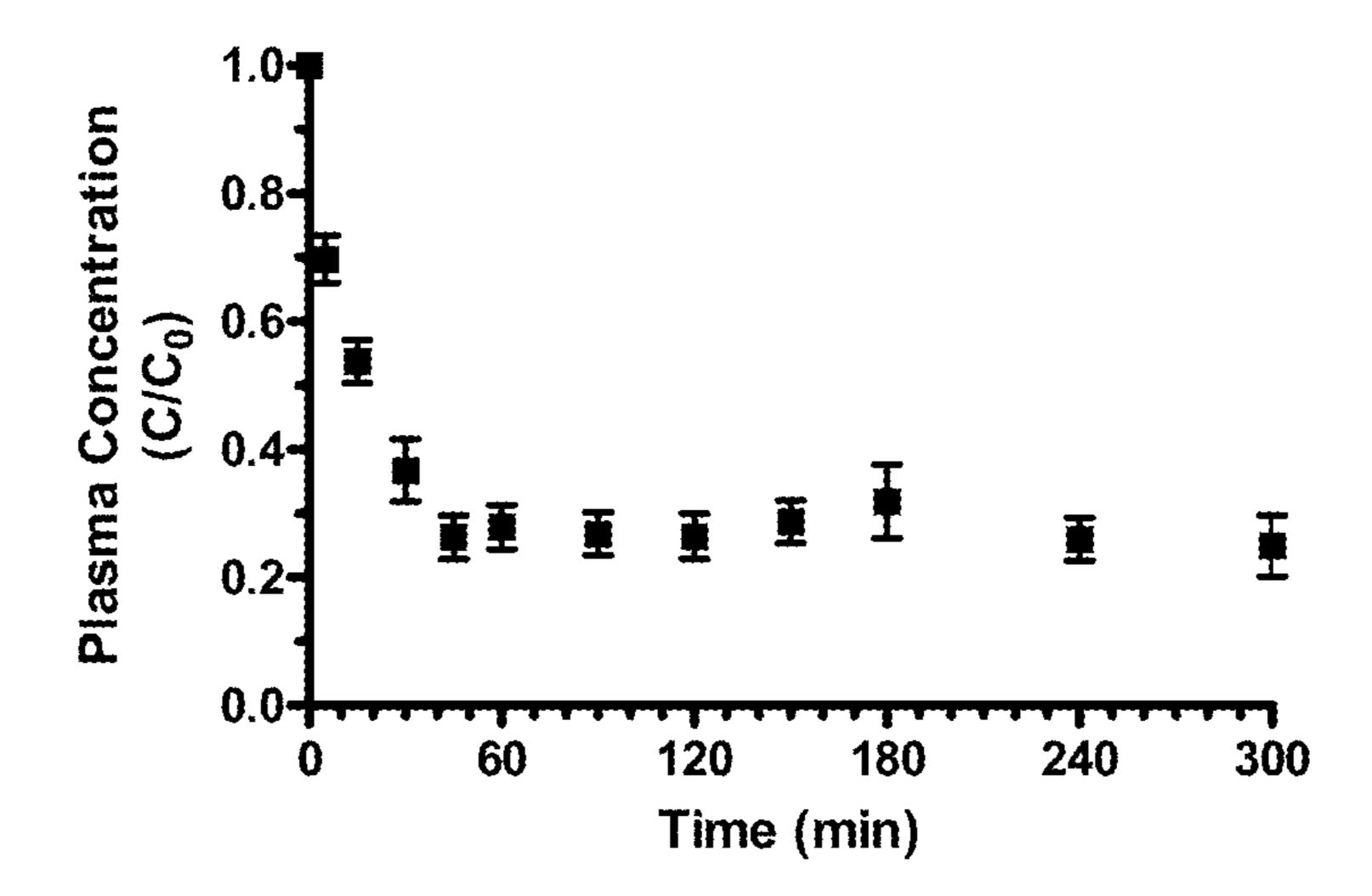


Figure 20.

# INHIBITION OF METASTASIS BY CELL PENETRATING PEPTIDES

#### PRIOR APPLICATIONS

This application is a continuation-in-part of U.S. patent application Ser. No. 12/422,975 filed on Apr. 13, 2009, now abandoned, which claims benefit to U.S. Patent Application No. 61/044,398, filed on Apr. 11, 2008; and this application is a continuation-in-part of U.S. patent application Ser. No. 12/162,283, filed Jun. 24, 2009, now issued as U.S. Pat. No. 8,252,740, which is a national phase application of International Patent Application No. PCT/US07/61240, filed on Jan. 29, 2007, which claims benefit to U.S. Patent Application No. 60/762,919, filed on Jan. 27, 2006. The content of patent application Ser. Nos. 12/422,975; 61/044,398; 12/162,283; PCT/US07/61240; and 60/762,919 are incorporated herein by reference.

# INTRODUCTION AND SUMMARY OF THE INVENTION

Embodiments of the present invention include a compound having a cell penetrating peptide (CPP) fused to an elastin-like polypeptide (ELP), and a method of using the compound as an anti-metastases agent.

Tumor cell metastasis is a complex, multi-step process that is a major cause of death and morbidity amongst cancer patients. Cell adhesion plays a critical role in the develop- 30 ment of metastatic cancer, and it is mediated by interactions between receptors on the cell surface and ligands of the extracellular matrix or other surfaces. Therefore, inhibition of the cell adhesion process appears to be an effective method of preventing metastasis.

To prevent cell adhesion the present inventors developed, as part of the present invention, genetically engineered polypeptides with the potential to inhibit metastases. Embodiments of the present invention include the cell 40 penetrating peptides (CPP) Tat or penetratin (Pen), fused with elastin-like polypeptide (CPP-ELP) inhibited adhesion, spreading, invasion and migration of SKOV-3 ovarian cancer cells, SK-MEL-2 melanoma cells, and MDA-MB-231 breast cancer cells. Additionally, examples of the present invention include the administration of Tat-ELP for antimetastatic treatment methods.

Accordingly, the polypeptides and other embodiments of the present invention are useful and needed as a therapeutic intervention in cancer metastasis.

Metastasis is the direct cause of mortality in most cancer patients. Therefore, efforts to understand and prevent the metastatic process are of tremendous clinical importance. While several approaches are being pursued to target cancer cell growth, relatively few focus specifically on preventing metastasis with drugs that can be safely administered on a long-term basis. Given the importance of metastasis as the major cause of increased morbidity and eventual mortality in cancer patients, the development of agents, such as the anti-adhesive polypeptide described here, that prevent or significantly delay metastasis without excessive collateral toxicity to other organs, would offer tremendous potential clinical benefit.

Since the adhesive interaction between tumor cells and host cells or extracellular matrix (ECM) plays a crucial role

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in metastatic formation (1-3), inhibition of the cell adhesion process appears to be an effective method of preventing metastasis. Rapid progress has been made in structural and functional analysis of cellular adhesive molecules involved in cell-cell or cell-ECM interactions. Several studies have suggested that synthetic peptides derived from adhesion molecules that are present in extracellular matrices or basement membranes can modulate the mechanism involved in metastasizing tumor cells (4, 5). It has been shown that peptides such as YIGSR, comprising residues 929-933 on the  $\beta_1$  chain of laminin, and the RGDS sequence in the central cell-binding domain of fibronectin can reduce formation of human melanoma tumors in nude mice (6, 7). However, most of these peptides as well some cytokines or anti-cancer drugs have very short half lives in the circulation, which results in a decrease in therapeutic and biological effect in vivo. Therefore, an increase in the half-life of a drug in circulation without increasing its toxicity may lead to 20 improved biological effect. Previous studies have shown that conjugation of RGD and YIGSR containing peptides with various drug carriers such as polyethylene glycol, poly (carboxymethylmethacrylamide), carboxymethyl chitin, and bovine serum albumin increased the inhibition of experimental and spontaneous tumor metastases (8, 9). Although bioconjugation of peptides with polymeric modifiers improved the plasma clearance and body distribution, most of these polymers are limited in their clinical application. Therefore, further improvements that increase the therapeutic effect and decrease side effects are needed.

The present inventors have found that the cell penetrating peptides of the present invention, such as Tat or penetratin, fused with elastin-like polypeptide (CPP-ELP), inhibited ovarian cancer, breast cancer and melanoma cell adhesion, spreading, invasion and migration. The polypeptides of the present invention have great potential as a therapeutic intervention in cancer metastasis. There are several advantages of these novel ELP-based polypeptides over existing antiadhesion polymers. First, while classical approaches rely on chemical synthesis of anti-adhesive peptides and chemical conjugation of anti-adhesive peptides to carriers, we produce an anti-adhesive peptide using simple molecular biology techniques. The coding sequence for ELP may be 45 modified by addition of the cell penetrating peptide (CPP) or any other peptide with anti-metastatic properties. Second, ELPs consist of Val-Pro-Gly-Xaa-Gly (VPGXG (SEQ ID NO: 37)) repeated units, and they are attractive from a molecular design perspective for targeted drug delivery because they are genetically encoded, which provides control over the ELP sequence and molecular weight (MW) to an extent that is impossible with synthetic polymer analogs. Control of macromolecular chain length and polydispersity is important because it controls the residence time of the drug-polymer conjugate in systemic circulation (10, 11) (please see Section I, FIG. 20 for preliminary results of plasma clearance). Finally, an additional advantage of ELPbased genetically encoded polypeptides over synthetic polymer carriers is that they are thermally responsive. Therefore, they may be expressed and purified from  $E.\ coli$  by a simple process called thermal cycling, which easily produces a large quantity of the purified polypeptide (12, 13).

Cell penetrating peptides are known for their ability to mediate cellular uptake of large proteins and macromolecules (reviewed in (14-16)). Also, the penetratin peptide intracellular delivery system has been patented (U.S. Pat. No. 6,844,324, to Zhang et al.).

## BRIEF DESCRIPTION OF THE SEQUENCE LISTING

SEQ ID NO: 1 is a Tat cell penetrating polypeptide.

SEQ ID NO: 2 is a Penetratin cell penetrating polypeptide.

SEQ ID NO: 3 is a Bac cell penetrating polypeptide.

SEQ ID NO: 4 is a SynB1 cell penetrating polypeptide.

SEQ ID NO: 5 is a Syn B1-NLS cell penetrating polypeptide.

SEQ ID NO: 6 is a poly-arginine cell penetrating polypeptide including seven (7) arginines.

SEQ ID NO: 7 is a poly-arginine cell penetrating polypeptide including eight (8) arginines.

SEQ ID NO: 8 is a poly-arginine cell penetrating polypeptide including nine (9) arginines.

SEQ ID NO: 9 is a poly-arginine cell penetrating polypeptide including ten (10) arginines.

SEQ ID NO: 10 is a poly-arginine cell penetrating polypeptide including eleven (11) arginines.

SEQ ID NO: 11 is a VP22 cell penetrating polypeptide.

SEQ ID NO: 12 is a Transportan cell penetrating polypeptide.

SEQ ID NO: 13 is a MAP cell penetrating polypeptide. 25

SEQ ID NO: 14 is a pVEC cell penetrating polypeptide.

SEQ ID NO: 15 is a MTS cell penetrating polypeptide.

SEQ ID NO: 16 is a hCT-derived cell penetrating polypeptide.

SEQ ID NO: 17 is a MPG cell penetrating polypeptide. SEQ ID NO: 18 is a Buforin 2 cell penetrating polypeptide.

SEQ ID NO: 19 is a PEP-1 cell penetrating polypeptide. SEQ ID NO: 20 is a Magainin 2 cell penetrating polypeptide.

SEQ ID NO: 21 is an embodiment of an elastin-like polypeptide that includes repeating units of the amino acid sequence VPGXG (SEQ ID NO: 37), where each X is independently selected from valine, glycine, and alanine 40 such that the Xs are provided in a 5:3:2 ratio.

SEQ ID NO: 22 is an embodiment of an elastin-like polypeptide that includes repeating units of the amino acid sequence VPGXG (SEQ ID NO: 37), where each X is independently selected from valine, glycine, and alanine 45 such that the Xs are provided in a 1:7:8 ratio.

SEQ ID NO: 23 is an embodiment of an amino acid comprising a Tat cell penetrating polypeptide and an elastinlike polypeptide.

SEQ ID NO: 24 is another embodiment of an amino acid 50 including a Tat cell penetrating polypeptide and an elastinlike polypeptide.

SEQ ID NO: 25 is an embodiment of an amino acid including a Penetratin cell penetrating polypeptide and an elastin-like polypeptide.

SEQ ID NO: 26 is another embodiment of an amino acid including a Penetratin cell penetrating polypeptide and an elastin-like polypeptide.

SEQ ID NO: 27 is an embodiment of an amino acid including an MTS cell penetrating polypeptide and an 60 cells were harvested and counted. Results represent the elastin-like polypeptide.

SEQ ID NO: 28 is another embodiment of an amino acid including an MTS cell penetrating polypeptide and an elastin-like polypeptide.

SEQ ID NO: 29 is an embodiment of an amino acid 65 including a Bac-7 cell penetrating polypeptide and an elastin-like polypeptide.

SEQ ID NO: 30 is another embodiment of an amino acid including a Bac-7 cell penetrating polypeptide and an elastin-like polypeptide.

SEQ ID NO: 31 is an embodiment of an amino acid including a Transportan cell penetrating polypeptide and an elastin-like polypeptide.

SEQ ID NO: 32 is another embodiment of an amino acid including a Transportan cell penetrating polypeptide and an elastin-like polypeptide.

SEQ ID NO: 33 is an embodiment of an amino acid including a pVEC cell penetrating polypeptide and an elastin-like polypeptide.

SEQ ID NO: 34 is another embodiment of an amino acid including a pVEC cell penetrating polypeptide and an elas-15 tin-like polypeptide.

SEQ ID NO: 35 is an embodiment of an amino acid including a SynB1 cell penetrating polypeptide and an elastin-like polypeptide.

SEQ ID NO: 36 is another embodiment of an amino acid including a SynB1 cell penetrating polypeptide and an elastin-like polypeptide.

SEQ ID NO: 37 is a VPGXG unit, wherein each X is independently selected from valine, glycine, and alanine.

#### BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1 is a chart showing in vitro attachment inhibition. MDA-MB-231 cells were exposed to polypeptides at the indicated concentration during a 3 h plating period in gelatin-coated dishes. Floating cells were harvested and counted. Results represent the mean±SEM of at least 3 independent experiments.

FIG. 2 is a set of photographs showing the spreading of MDA-MB-231 onto coverslips. Cells were treated in suspension with CPP-ELP at the indicated concentration and plated onto serum coated coverslips. Images were collected 4 h after plating using a Zeiss Axiovert DIC microscope with a 40× oil immersion objective.

FIG. 3 is a set of photographs showing a scratch migration assay. MDA-MB-231 cells were grown to confluence and a scratch was made in the monolayer. Cells were treated with protein and allowed to migrate for 24 h. Migration was measured by collecting DIC images with at 10× magnification.

FIG. 4 a growth curve chart of MDA-MB-231 cells. Cells were treated with the indicated proteins before plating. Daily counts were made using a Coulter counter. Data represents an average of 3 independent experiments; bars, SE.

FIG. 5 is a set of charts showing apoptosis assay MDA-MB-231 cells treated with the indicated protein for 10 min and plated for 5 h. Cells were harvested and stained with FITC-annexin and propidium iodide, and fluorescence levels were determined by flow cytometry. Live cells are unstained by either agent and appear in L3. Apoptotic cells stain with 55 FITC-annexin, but not with propidium iodide, and appear in L4. Cells in L2 stain with both agents and are necrotic.

FIG. 6 is a chart showing in vitro attachment inhibition. SK-MEL-2 cells were exposed to polypeptides at the indicated concentration during a 3 h plating period. Floating mean±SEM of at least 3 independent experiments.

FIG. 7 is a set of photographs showing spreading of SK-MEL-2 onto coverslips. Cells were treated in suspension with CPP-ELP at the indicated concentration and plated onto acid-washed coverslips. Images were collected at the indicated times using a Zeiss Axiovert DIC microscope with a 40× oil immersion objective.

FIG. **8** is a set of photographs showing scratch migration assay. SK-MEL-2 cells were grown to confluence and a scratch was made in the monolayer. Cells were treated with protein and allowed to migrate for 24 h. Migration was measured by collecting DIC images with at 10× magnifica- <sup>5</sup> tion.

FIG. 9 is a chart showing a growth curve of SK-MEL-2 cells. Cells were treated with the indicated proteins before plating. Daily counts were made using a Coulter counter. Data represents an average of 3 independent experiments; bars, SE.

FIG. 10 is a set of photographs showing an apoptosis assay. SK-MEL-2 cells were treated with the indicated protein for 10 min and plated for 5 h. Cells were harvested and stained with FITC-annexin and propidium iodide, and fluorescence levels were determined by flow cytometry. Live cells are unstained by either agent and appear in K3. Apoptotic cells stain with FITC-annexin, but not with propidium iodide, and appear in K4. Cells in K2 stain with both agents and are necrotic.

FIG. 11 is a chart showing in vitro attachment inhibition. SKOV-3 cells were exposed to polypeptides at the indicated concentration during a 3 h plating period. Floating cells were harvested and counted. Results represent the mean±SEM of at least 3 independent experiments.

FIG. 12 is a set of photographs showing the spreading of SKOV-3 onto coverslips. Cells were treated in suspension with CPP-ELP at the indicated concentration and plated onto acid-washed coverslips. Images were collected at the indicated times using a Zeiss Axiovert DIC microscope with a 40× oil immersion objective.

FIG. 13 is a set of photographs showing another scratch migration assay. SKOV-3 cells were grown to confluence and a scratch was made in the monolayer. Cells were treated with protein and allowed to migrate for 24 h. Migration was measured by collecting DIC images at 10× magnification.

FIG. 14 is data related to a Boyden chamber assay. FIG. 14A shows % migration of SKOV-3 cells were incubated with the indicated polypeptides at a fixed concentration for 10 min at 37° C., added to the upper chamber of the boyden chamber insert and allowed to migrate for 24 h. Cells were washed, fixed, stained with hematoxylin and 4 random fields were counted at 20× magnification. FIG. 14B is a set of images after cells were later incubated with Hoechst dye for 10 min. and images were obtained at 10× magnification.

FIG. 15 is a chart that shows in vitro attachment inhibition on vitronectin coated plates. SKOV-3 cells were incubated with the different polypeptides at the indicated concentrations during a 3 h plating period on vitronectin coated plates. Floating cells were harvested and counted. Results represent 50 the mean±SEM of at least 3 independent experiments.

FIG. 16 illustrates a cell surface receptor assay. A. Trypsin treatment. Cells were briefly incubated with trypsin, washed and resuspended in PBS containing fluorescein-labeled polypeptide at various concentrations for 10-15 min. Binding of different polypeptides was measured by immediately analyzing cell fluorescence by flow cytometry. B. Heparanase treatment. Cells were briefly incubated with heparanase, washed and resuspended in PBS containing fluorescein-labeled polypeptide, and cell fluorescence was 60 determined as described above. C. Pronase treatment. Cells were briefly incubated with pronase, washed and resuspended in PBS containing fluorescein-labeled polypeptide, and cell fluorescence was determined as described above.

FIG. 17 is a set of tables showing binding at different ionic 65 strengths. Cells were resuspended in ½x, 1x and 2xPBS and incubated with fluorescene labeled polypeptides in a con-

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centration dependent manner for 10-15 min. and immediately run on flow cytometer. FIG. 17 a shows binding with Tat-ELP. FIG. 17B shows binding with MTS-ELP. FIG. 17C shows binding with ELP.

FIG. 18 is a chart showing a growth curve of SKOV-3 cells. Cells were treated with the indicated proteins before plating. Daily counts were made using a Coulter counter. Data represents an average of 3 independent experiments; bars, SE.

FIG. 19 is a set of charts showing an experimental metastases assay. BALB/C nude mice were given an i.p injection of SKVO-3 cells in PBS and SKOV-3 cells in 500 μM/mL Tat-ELP. The mice were sacrificed 17 days later and tumor mass per animal was recorded. A. Total tumor burden per animal in treated and untreated animals. B. Tumor burden recorded in uterus/fallopian tube of each animal. Bars; SD, n=5.

FIG. 20 is a chart showing mean of normalized plasma concentration-time profile of CPP-ELP after administration of a bolus IV injection. Plasma was diluted for fluorescence measurement of polypeptide concentration. Data are normalized to the initial concentration and plotted as mean±SE from three animals.

# DESCRIPTION OF EMBODIMENTS OF THE INVENTION

The present invention includes a compound for inhibiting proliferation of cancer, including a cell penetrating polypep-tide (CPP) and an elastin-like polypeptide (ELP). In some embodiments, the compound can be administered to a subject to inhibit the proliferation of a cancer in the subject.

As used herein, the term "cell penetrating polypeptide" (CPP) refers to a polypeptide that facilitates transport of the compound through a cell membrane.

As used herein, the term "polypeptide" means any polymer comprising any of the 20 protein amino acids, regardless of its size. Although "protein" is often used in reference to relatively large polypeptides, and "peptide" is often used in reference to small polypeptides, usage of these terms in the art overlaps and varies. The term "polypeptide" as used herein refers to peptides, polypeptides, and proteins, unless otherwise noted.

Cell penetrating peptides can be short polypeptides capable of mediating delivery of molecules across a cell membrane. In some embodiments, CPPs can be comprised of mostly basic amino acids, hydrophobic amino acids, or an amphipathic sequence. Examples of CPPs that can be used in accordance with the present invention include, but are not limited, to those set forth in Table 1.

TABLE 1

5	CPP	Amino Acid Sequence	SEQ ID NO:
	Tat <sup>1</sup>	YGRKKRRQRRR	1
	Penetratin	RQIKIWFQNRRMKWKK	2
	$(Antp)^2$		
	Bac <sup>3</sup>	RRIRPRPPRLPRPRPLPFPRPG	3
`	SynB1	RGGRLSYSRRRFSTSTGR	4
,	SynB1-	RGGRLSYSRRRFSTSTGRWSQPKKKRKV	5
	$NLS^4$		
	Poly-	$(R)_{7-11}$	6, 7,
	arginine		8, 9,
			10
_	VP22	DAATATRGRSAASRPTQRPRAPARSASRPRRPVQ	11
•	Trans-	GWTLNSAGYLLGKINLKALAALAKKIL	12
	portan <sup>5</sup>		

СРР	Amino Acid Sequence	SEQ ID NO:
MAP	KLALKLALKAALKLA	13
pVEC <sup>6</sup>	LLIILRRRIRKQAHAHSK	14
$MTS^7$	AAVALLPAVLLALLAP	15
hCT	LGTYTQDFNKFHTFPQTAIGVGAP	16
derived		
MPG	GALFLGFLGAAGSTMGAWSQPKKKRKV	17
Buforin 2	TRSSRAGLQFPVGRVHRLLRK	18
PEP-1	KETWWETWWTEWSQPKKKRKV	19
Magainin	GIGKFLHSAKKFGKAFVGEIMNS	20

<sup>1</sup>Tat is a cell penetrating peptide derived from the HIV-1 Tat protein (18).

<sup>2</sup>Penetratin (commonly abbreviated Pen or AntP) is the penetratin peptide derived from the *Drosophila* transcription factor Antennapaedia (17).

<sup>3</sup>Bac-7 is an antimicrobial peptide from the Bactenecin-7 family (20).

<sup>4</sup>SynB1-NLS is a version of the SynB1 CPP modified in the present inventors' lab by the addition of a nuclear localization sequence (NLS, underlined amino acids) to allow delivery of the compound not only across the cell membrane, but also into the cell's nucleus.

Transportan is a chimeric peptide in which the first 13 amino acids are derived from galanin and the other 14 amino acids from the wasp venom peptide toxin, mastoparan (21). <sup>6</sup>pVEC is derived from murine Vascular Endothelial Cadherin (22).

<sup>7</sup>MTS is the membrane translocating sequence derived from Kaposi fibroblast growth factor (19).

Embodiments of compounds of the present invention further include an elastin-like polypeptide (ELP) that is 25 fused to the CPP (e.g., fusion protein including an ELP and a CPP). IN some embodiments, the CPP and the ELP are provided as a fusion protein, wherein the CPP is fused directly to the ELP. In some embodiments the CPP and the ELP are provided as a fusion protein, wherein linker comprising one or more amino acids is disposed between the CPP and the ELP.

In some embodiments, the ELP is an approximately 60 kilodalton protein comprising repeated units of the amino acid sequence VPGXG (SEQ ID NO: 37), where each X is 35 independently selected from valine (Val; V), glycine (Gly; G), and alanine (Ala; A).

In some embodiments, the ELP can comprise the amino acid sequence (VPGXG (SEQ ID NO: 37)), WP or (VPGXG (SEQ ID NO: 37)), where n is about 20, 21, 22, 23, 24, 25, 40 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100, 101, 102, 103, 45 104, 105, 106, 107, 108, 109, 110, 111, 112, 113, 114, 115, 116, 117, 118, 119, 120, 121, 122, 123, 124, 125, 126, 127, 128, 129, 130, 131, 132, 133, 134, 135, 135, 137, 138, 139, 140, 141, 142, 143, 144, 145, 146, 147, 148, 149, 150, 151, 152, 153, 154, 155, 156, 157, 158, 159, 160, 161, 162, 163, 50 164, 165, 166, 167, 168, 169, 170, 171, 172, 173, 174, 175, 176, 177, 178, 179, 180, 181, 182, 183, 184, 185, 186, 187, 188, 189, 190, 191, 192, 193, 194, 195, 196, 197, 198, 199, 200, 201, 202, 203, 205, 206, 207, 208, 209, 210, 211, 212, 213, 214, 215, 216, 217, 218, 219, 220, 221, 222, 223, 224, 55 225, 226, 227, 228, 229, 230, 231, 232, 233, 234, 235, 236, 237, 238, 239, 240, 241, 242, 243, 244, or 245.

In some embodiments, the ELP can comprise the amino acid sequence (VPGXG (SEQ ID NO: 37))<sub>n</sub> WP or (VPGXG (SEQ ID NO: 37))<sub>n</sub> where n is an integer of at least about 20, 60 and each X is independently selected from valine, glycine, and alanine.

In some embodiments, the ELP can comprise the amino acid sequence (VPGXG (SEQ ID NO: 37))<sub>n</sub> WP or (VPGXG)<sub>n</sub> where each X is independently selected from 65 valine, glycine, and alanine such that the Xs are Val:Gly:Ala in a 5:3:2 ratio. For exemplary purposes only, to illustrate the

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ratio of Val:Gly:Ala in some embodiments of the composition, where n is 20, ten (10) of the Xs would be selected to be valine, six (6) of the Xs would be selected to be glycine, and four (4) of the Xs would be selected to be alanine.

In some embodiments, the ELP can comprise the amino acid sequence (VPGXG (SEQ ID NO: 37))<sub>n</sub> WP or (VPGXG (SEQ ID NO: 37))<sub>n</sub> where n is about 20, 40, 80, 150, or 160, and where each X is independently selected from valine, glycine, and alanine such that the Xs are Val:Gly:Ala in a 5:3:2 ratio. In some embodiments, the ELP having Xs that are Val:Gly:Ala in a 5:3:2 ratio can comprise the amino acid sequence of SEQ ID NO: 22.

In some embodiments, the ELP can comprise the amino acid sequence (VPGXG (SEQ ID NO: 37))<sub>n</sub> WP or (VPGXG (SEQ ID NO: 37))<sub>n</sub> where each X is independently selected from valine, glycine, and alanine such that the Xs are Val:Gly:Ala in a 3:1:1 ratio. For exemplary purposes only, to illustrate the ratio of Val:Gly:Ala in some embodiments of the composition, where n is 5, three (3) of the Xs would be selected to be valine, one (1) X would be selected to be glycine, and one (1) X would be selected to be alanine.

In some embodiments, the ELP can comprise the amino acid sequence (VPGXG (SEQ ID NO: 37))<sub>n</sub> WP or (VPGXG (SEQ ID NO: 37))<sub>n</sub> where n is about 20, 40, 80, 150, or 160, and where each X is independently selected from valine, glycine, and alanine such that the Xs are Val:Gly:Ala in a 3:1:1 ratio.

In some embodiments, the ELP can comprise the amino acid sequence (VPGXG (SEQ ID NO: 37))<sub>n</sub> WP or (VPGXG (SEQ ID NO: 37))<sub>n</sub> where each X is independently selected from valine, glycine, and alanine such that the Xs are Val:Gly:Ala in a 1:7:8 ratio. For exemplary purposes only, to illustrate the ratio of Val:Gly:Ala in some embodiments of the composition, where n is 16, one (1) X would be selected to be valine, seven (7) of the Xs would be selected to be glycine, and eight (8) of the Xs would be selected to be alanine.

In some embodiments, the ELP can comprise the amino acid sequence (VPGXG (SEQ ID NO: 37))<sub>n</sub> WP or (VPGXG (SEQ ID NO: 37))<sub>n</sub> where n is about 20, 40, 80, 150, or 160, and where each X is independently selected from valine, glycine, and alanine such that the Xs are Val:Gly:Ala in a 1:7:8 ratio. In some embodiments, the ELP having Xs that are Val:Gly:Ala in a 1:7:8 ratio can comprise the amino acid sequence of SEQ ID NO: 23.

In some embodiments, the ELP can be an ELP having the amino acid sequence of SEQ ID NO: 22. In some embodiments, the ELP can be an ELP having the amino acid sequence of SEQ ID NO: 23.

In some embodiments, the ELP is an ELP as described in U.S. Patent Application Publication No. 2005/0255554 of A. Chilkoti, which is incorporated herein by this reference.

In some embodiments, the compound of the presently-disclosed subject matter can include the amino acid sequence of SEQ ID NO: 24. In some embodiments, the compound of the presently-disclosed subject matter can include the amino acid sequence of SEQ ID NO: 25. In some embodiments, the compound of the presently-disclosed subject matter can include the amino acid sequence of SEQ ID NO: 26. In some embodiments, the compound of the presently-disclosed subject matter can include the amino acid sequence of SEQ ID NO: 27. In some embodiments, the compound of the presently-disclosed subject matter can include the amino acid sequence of SEQ ID NO: 28. In some embodiments, the compound of the presently-disclosed subject matter can include the amino acid sequence of SEQ ID NO: 29. In some embodiments, the compound of the presently-disclosed subject matter can include the amino acid sequence of SEQ ID NO: 29. In some embodiments, the compound of the presently-disclosed subject matter can include the amino acid sequence of SEQ ID NO: 29. In some embodiments, the compound of the presently-disclosed subject matter can include the amino acid sequence of SEQ ID NO: 29. In some embodiments, the compound of the presently-disclosed subject matter can include the amino acid sequence of SEQ ID NO: 29. In some embodiments, the compound of the presently-disclosed subject matter can include the amino acid sequence of SEQ ID NO: 29. In some embodiments, the compound of the presently-disclosed subject matter can include the amino acid sequence of SEQ ID NO: 29. In some embodiments, the compound of the presently-disclosed subject matter can include the amino acid sequence of SEQ ID NO: 29. In some embodiments, the compound of the presently-disclosed subject matter can include the amino acid sequence of SEQ ID NO: 29. In some embodiments, the compound of the presently-disclosed subject matter can include the amino acid sequence of SEQ ID NO: 29.

TABLE 2

ently-disclosed subject matter can include the amino acid sequence of SEQ ID NO: 30. In some embodiments, the compound of the presently-disclosed subject matter can include the amino acid sequence of SEQ ID NO: 31. In some embodiments, the compound of the presently-disclosed sub- 5 ject matter can include the amino acid sequence of SEQ ID NO: 32. In some embodiments, the compound of the presently-disclosed subject matter can include the amino acid sequence of SEQ ID NO: 33. In some embodiments, the compound of the presently-disclosed subject matter can include the amino acid sequence of SEQ ID NO: 34. In some embodiments, the compound of the presently-disclosed subject matter can include the amino acid sequence of SEQ ID NO: 35. In some embodiments, the compound of the pres- 15 present invention. This method allows for efficient determiently-disclosed subject matter can include the amino acid sequence of SEQ ID NO: 36. In some embodiments, the compound of the presently-disclosed subject matter can include the amino acid sequence of SEQ ID NO: 37.

As will be recognized by those skilled in the art upon 20 studying the present document, with reference to the specific examples of CPPs and ELPs that can be used in accordance with the presently-disclosed subject matter, one or more amino acids can be added to and/or one or more amino acids can be removed from and/or conservative substations of one 25 or more amino acids can be made as compared to the exemplary sequences set forth herein to obtain additional embodiments of the presently-disclosed subject matter. With regard to removing and/or making a conservative substitution of one or more amino acids relative to the specific 30 examples of CPPs and ELPs as set forth herein, consideration to cell binding efficacy, and aggregation efficacy should be considered.

A "conservative substitution" is a substitution of an amino acid residue with a functionally similar residue. Examples of 35 conservative substitutions include the substitution of one non-polar (hydrophobic) residue such as isoleucine, valine, leucine or methionine for another; the substitution of one charged or polar (hydrophilic) residue for another such as between arginine and lysine, between glutamine and asparagine, between threonine and serine; the substitution of one basic residue such as lysine or arginine for another; or the substitution of one acidic residue, such as aspartic acid or glutamic acid for another; or the substitution of one aromatic residue, such as phenylalanine, tyrosine, or tryptophan for 45 another.

As will be recognized by those skilled in the art upon studying the present document, compounds as described herein can be made using standard molecular biology techniques.

The present inventors have shown, for example, that penetratin-ELP and the Tat-ELP inhibited adhesion, spreading, and migration of MDA-MB-231 breast cancer cells, SKOV-3 ovarian cancer cells and SK-MEL-2 melanoma cells.

The present invention further includes methods of inhibition comprising the use of polypeptides or compounds of the present invention. Table 2, below shows that inhibition of cell adhesion is cell line and CPP-ELP dependent. More specifically, Table 2 indicates that the  $IC_{50}$  is the concentra- 60 tion of each polypeptide needed to prevent attachment of 50% of the plated cells. Tat-ELP, MTS-ELP and Antp-ELP polypeptides were shown to effectively prevent attachment in ovarian, melanoma and breast cancer cell lines, with Tat-ELP being the most efficient in all cell lines. Other 65 polypeptides like Bac-7-ELP, Trans-ELP and pVEC-ELP are tested for their ability to prevent cell adhesion.

	IC <sub>50</sub> of Different Polypeptides (μM)												
Cell Name	Cell Type	Tat- ELP	Antp- ELP		Bac-7- ELP	Trans- ELP	pVEC- ELP						
		0.125 0.015	1.5 1	10 10	nd* nd*	nd* nd*	nd* nd*						
MDA	Breast	0.5	1	nd*	nd*	nd*	nd*						

10 \*nd—not determined

Therefore, the present inventors have designed a system to achieve maximum cell adhesion inhibition for a particular cell line, including those for use with CPP-ELPs of the nation of CPP-ELPs in connection with inhibition of specific cancer cells.

Thus, additional embodiments of the present invention are novel classes of anti-adhesion polypeptides (compounds of the present invention), which are capable of inhibiting adhesion, spreading, and migration of cancer. In some embodiments, the compounds of the present invention are capable of inhibiting adhesion, spreading, and migration of cancer in a subject. Further embodiments of the present invention are methods of inhibiting the progression of tumors comprising administering the compounds of the present invention to a subject.

As used herein, the term "subject" includes both human and animal subjects. Thus, veterinary therapeutic uses are provided in accordance with the presently disclosed subject matter. As such, the presently disclosed subject matter provides for the treatment of mammals such as humans, as well as those mammals of importance due to being endangered, such as Siberian tigers; of economic importance, such as animals raised on farms for consumption by humans; and/or animals of social importance to humans, such as animals kept as pets or in zoos. Examples of such animals include but are not limited to: carnivores such as cats and dogs; swine, including pigs, hogs, and wild boars; ruminants and/or ungulates such as cattle, oxen, sheep, giraffes, deer, goats, bison, and camels; and horses. Also provided is the treatment of birds, including the treatment of those kinds of birds that are endangered and/or kept in zoos, as well as fowl, and more particularly domesticated fowl, i.e., poultry, such as turkeys, chickens, ducks, geese, guinea fowl, and the like, as they are also of economic importance to humans. Thus, also provided is the treatment of livestock, including, but not limited to, domesticated swine, ruminants, ungulates, horses (including race horses), poultry, and the like.

In some embodiments a CPP-ELP can be formulated by purifying the polypeptide from cultured bacterial cells grown in culture flasks or a bioreactor. Once purity of the polypeptide agent is insured, it will be formulated for injection by dissolving it in the appropriate amount of 55 physiological saline to produce an injection of the proper dose and volume for administration, which can vary depending on the administration route used as outlined herein.

Conventional methods, known to those of ordinary skill in the art of medicine, can be used to administer effective amounts of the compound in a suitable formulation to a subject. Suitable methods for administering embodiments of the compound of the present invention in accordance with the methods of the present invention include but are not limited to systemic administration, parenteral administration (including intravascular, intramuscular, intraarterial, intraperitoneal administration), oral delivery, buccal delivery, subcutaneous administration, inhalation, intratracheal installa-

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tion, surgical implantation, transdermal delivery, local injection, and hyper-velocity injection/bombardment.

The particular mode of drug administration used in accordance with the methods of the present invention depends on various factors, including but not limited to the severity of 5 the condition to be treated.

The term "effective amount" is used herein to refer to an amount of the compound sufficient to produce a measurable biological response. Actual dosage levels of the compound in an appropriate formulation can be varied so as to admin- 10 ister an amount of the compound that is effective to achieve the desired response for a particular subject and/or application. The selected dosage level will depend upon a variety of factors including the route of administration, combination with other drugs or treatments, severity of the condition <sup>15</sup> being treated, and the physical condition and prior medical history of the subject being treated. Preferably, a minimal dose is administered, and dose is escalated in the absence of dose-limiting toxicity to a minimally effective amount. Determination and adjustment of an effective dose, as well 20 as evaluation of when and how to make such adjustments, are known to those of ordinary skill in the art of medicine and can be determined in a particular case by one skilled in the art using only routine experimentation. Dosing can being at levels needed to achieve plasma concentration of the 25 compound in substantially the ranges as set forth in Table 2, e.g., equating to a dose of about 1 mg polypeptide/kg of body weight, up to about 200 to about 500 mg polypeptide/ kg of body weight.

#### EXAMPLES

The following Examples are presented for exemplary purposes. Accordingly, they are to be construed as showing embodiments of the present invention and are not to be <sup>35</sup> construed as being limiting thereof.

The Examples demonstrate that the exemplary compounds of the present invention are capable of inhibiting adhesion, spreading, invasion and migration of breast cancer cells, melanoma cells and ovarian cancer cells.

#### Example 1

#### Breast Cancer Metastasis

This Example helps characterize the mechanism of CPP-ELP action:

Extracellular matrix protein adhesion assay. The ability of CPP-ELP to modulate adhesion and spreading of MDA-MB-231 breast cancer cells to a substrate coated with 50 specific extracellular matrix (ECM) molecules (fibronection, laminin, and collagen IV).

Cell surface receptor assay. In order to elucidate the nature of the cell surface molecules involved, the effect of proteolytic digestion of specific cell surface molecules on 55 the ability of CPP-ELP to bind to MDA-MB-231 cells.

Immunoprecipitation of cell surface proteins. Using the results from the cell surface receptor assay, any protease that demonstrates an effect on CPP-ELP binding used in an immunoprecipitation assay to identify the specific proteins 60 released from the membrane that are important in CPP-ELP binding.

Effect of CPP-ELP size on attachment inhibition. Experiments can demonstrate whether or not specific proteins are involved in CPP-ELP inhibition. If no proteins are involved 65 in CPP-ELP inhibition, then it is likely that CPP-ELP works by simply coating the cell surface and blocking many

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important cell to ECM interactions. CPP-ELPs of different molecular weights can be used to determine the size dependence of cell adhesion inhibition. This hypothesis indicates that a larger CPP-ELP may be a more potent inhibitor than a smaller one.

#### Example 2

#### Metastasis of Melanoma Cells

To test the ability of CPP-ELP to reduce the metastatic properties of melanoma cells in vivo the following experiments may be done:

Experimental metastasis assay. Briefly, immunodeficient mice are given an i.v. injection of SK-MEL-2 melanoma cells mixed with various concentrations of CPP-ELP. Two weeks after inoculation of tumor cells, the mice are sacrificed and the number of tumor colonies in the lung, spleen, and kidneys will be recorded.

Spontaneous metastasis assay. In the spontaneous metastasis assay, mice are injected subcutaneously in the hind limb with melanoma cells to form a primary tumor. Polypeptides are administered i.v. on various days after tumor inoculation, and metastasized tumor colonies in the lung, spleen, and kidneys will be counted four weeks later.

#### Example 3

#### Breast Cancer Metastasis

When breast carcinoma remains confined to breast tissue, cure rates exceed 90%. However, cells from a primary tumor can spread to distant tissues via blood vasculature or lymphatics and form secondary tumors, or metastases. As cells spread, long-term survival decreases dramatically depending upon the extent of and the sites of colonization. Metastases in visceral organs and brain are the most life-threatening, and they are the direct cause of mortality in most breast cancer patients, with 5-year survival rates usually less than 20%. Therefore, efforts to understand and prevent the metastatic process of breast cancer cells are of tremendous clinical importance.

In Vitro Cell Attachment and Spreading: The present inventors have shown that immediately after incubation with 45 MCF-7 breast cancer cells, CPP-ELP based polypeptides are localized to the plasma membrane (30). Therefore, without being bound by theory, it appears that the polypeptides of the present invention, with such cell membrane binding properties, may affect the cell's ability to attach to a substrate. In order to investigate CPP-ELPs for the ability to inhibit attachment, an in vitro cell attachment assay was used. MDA-MB-231 human breast cancer cells were incubated in suspension ( $5 \times 10^4$  cells in 1 ml) with the CPP-ELPs for 10 minutes. 24 well tissue culture dishes were coated for 2 h with a 2% gelatin solution, and the cell/protein mixture was then plated and incubated for 3 h to allow cell attachment. After the attachment period, non-adherent cells were collected by removing the media and rinsing the wells gently with PBS. The floating fraction was counted using a Coulter counter, and the results are shown in FIG. 1.

During the 3 h attachment period, more than  $4\times10^4$  of the  $5\times10^4$  untreated MDA-MB-231 cells plated attached to the substrate (as shown at zero polypeptide concentration in FIG. 1). The ELP polypeptide, which has no CPP to facilitate cell binding, showed no inhibition of cell attachment at any concentration tested (not shown). In contrast, Pen-ELP showed a concentration dependent inhibition of MDA-MB-

231 attachment, with complete inhibition observed at 5 µM Pen-ELP. Tat-ELP inhibited cell attachment even more efficiently, with a maximum inhibition occurring at only 1 µM. The inhibition of attachment was not simply a property of the CPP peptide, since the 16 aa penetratin peptide alone had 5 no effect on cell attachment (data not shown). These results show that both the CPP and ELP are required for attachment inhibition.

In addition to inhibiting attachment of the cells to the substrate, the CPP-ELPs also inhibited the spreading of any 10 cells that did attach. MDA-MB-231 cells were incubated as described above and plated on acid-washed coverslips coated with serum proteins. The coverslips were mounted observed using differential interference contrast (DIC) microscopy. FIG. 2 shows that 4 h after plating, control and ELP treated cells began to spread onto the coated coverslips. In contrast, the cells treated with Pen-ELP and Tat-ELP did not spread onto the substrate.

Cell Migration: Since the CPP-ELP molecules showed potent inhibition of cell attachment and spreading, we also assayed for their ability to inhibit cell migration. Migration is an important event in metastasis, in which the tumor cells must invade the layer of endothelial cells to gain access to 25 the vascular circulation (31). We employed an in vitro scratch migration assay to test the ability of CPP-ELP to inhibit cell migration (32). Briefly, MDA-MB-231 cells were grown to confluence on acid-washed coverslips. A linear scratch was made, and the growth medium was 30 replaced with media containing CPP-ELP. Cells were placed in the incubator for 24 h to allow migration of the cells into the newly generated scratch. Coverslips were mounted and cell migration was assessed by collecting DIC images of the scratched area. FIG. 3 shows that the scratch method 35 removed all cells from the area of interest. 24 h later, control cells and ELP treated cells (ELP data not shown) completely filled the cleared area. Pen-ELP significantly reduced the ability of the MDA-MB-231 cells to migrate into the cleared area, and Tat-ELP produced an even stronger inhibition of 40 migration.

Cell Proliferation and Apoptosis: During the attachment and spreading assays, cells treated with CPP-ELP never attached or spread even 24 h after plating. We also examined the cells for a longer time period in order to determine if the 45 cells were ever able to attach and to learn whether the cells can proliferate after attachment. To address this question, we monitored the growth of MDA-MB-231 cells after treatment with CPP-ELP. Cells were treated as described above and plated in serum-coated 24-well plates. Daily cell counts of 50 the attached cells were made using a Coulter counter, and a growth curve (shown in FIG. 4) was generated. Control and ELP treated cells grew normally, making about 2 doublings during the 4 days of the assay. In contrast, cells treated with Pen-ELP showed no increase in cell number over 4 days, and 55 Tat-ELP treated cells showed only a slight increase in number.

Inhibition of cell proliferation by CPP-ELP requires that the cells be treated while in suspension before plating. CPP-ELP shows no toxicity and no inhibition of growth 60 when added to cells which are already attached to substratum (data not shown). These results suggest that inhibition of the proliferation of breast cancer cells by CPP-ELP treatment may be due to inhibition of cell attachment. This observation may be advantageous for the treatment of breast 65 cancer metastasis, where the desired target cells are unattached and circulating.

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The DIC images from the spreading assay revealed the presence of some apoptotic cells after treatment with CPP-ELP. After learning that CPP-ELP treated cells were not proliferating, we used an annexin binding assay to quantitate the extent of apoptosis after plating cells treated with CPP-ELP. MDA-MB-231 cells were treated and plated as above in 6-well plates. 5 h after plating, all cells (floating and attached) were collected by trypsinization, stained with FITC-annexin and propidium iodide (Molecular Probes, Eugene, Oreg.), and analyzed by flow cytometry. One event in the induction of apoptosis is the externalization of phosphatidyl serine to the outer leaflet of the plasma membrane. These cells stain with FITC-annexin, allowing quantitation onto slides 4 h after plating, and the cell morphology was 15 of the amount of apoptotic cells. This assay also employs propidium iodide staining to elucidate the necrotic cells from the apoptotic cells.

> When treated with ELP, cells were unaffected and gave similar results to control cells in the apoptosis assay (FIG. 5). Pen-ELP produced a strong effect on the MDA-MB-231 cells, inducing apoptosis in nearly 60% of the cells within the 5 h tested. In contrast, Tat-ELP had no apoptosisinducing effect. Although Tat-ELP is a much more efficient inhibitor of cell attachment than Pen-ELP, Tat-ELP does not induce the apoptotic response nearly as extensively as Pen-ELP. This suggests that the two polypeptides may have different modes of action. This possibility will be addressed by the in vitro experiments proposed here.

In summary, this example shows that treatment of unattached breast cancer cells with CPP-ELPs inhibits their attachment, spreading, migration, and proliferation. Cells which are already attached are unaffected by CPP-ELP treatment, which is a promising fact for the future application of CPP-ELP

### Example 4

## Melanoma Cancer Metastasis

In Vitro Cell Attachment and Spreading: In order to investigate CPP-ELPs for the ability to inhibit attachment of melanoma cells, an in vitro cell attachment assay similar to the one described above was used, and the results are shown in FIG. **6**.

During the 3 h attachment period, more than  $4 \times 10^4$  of the 5×10<sup>4</sup> untreated SK-MEL-2 cells plated attached to the substrate (as shown at zero polypeptide concentration in FIG. 6). The ELP polypeptide, which has no CPP to facilitate cell binding, showed no inhibition of cell attachment at any concentration tested. In contrast, Pen-ELP showed a concentration dependent inhibition of SK-MEL-2 attachment, with complete inhibition observed at 20 µM Pen-ELP. Tat-ELP inhibited cell attachment even more efficiently, with a maximum inhibition occurring at only 2 μM. The inhibition of attachment was not simply a property of the CPP peptide, since the 16 aa penetratin peptide alone had no effect on cell attachment. These results show that both the CPP and ELP are required for attachment inhibition.

In addition to inhibiting attachment of the cells to the substrate, the CPP-ELPs also inhibited the spreading of any cells that did attach. SK-MEL-2 cells were incubated as described above and plated on acid-washed coverslips, and spreading was assayed using DIC microscopy. FIG. 7 shows that ELP does not affect cell spreading, but cells pretreated with Pen-ELP and Tat-ELP do not spread.

This spreading inhibition is not limited to cells plated on glass coverslips. Preliminary experiments with cells plated

on fibronectin-coated cover-slips show that both Pen-ELP and Tat-ELP can inhibit SK-MEL-2 cell spreading onto a natural ECM substratum.

Cell Migration: Since the CPP-ELP molecules showed potent inhibition of melanoma cell attachment and spreading, we also assayed for their ability to inhibit cell migration using the scratch migration assay described above. FIG. 8 shows that the scratch method removed all cells from the area of interest. 24 h later, control and ELP treated cells almost completely filled the cleared area. Pen-ELP signifi- 10 cantly reduced the ability of the SK-MEL-2 cells to migrate into the cleared area, and Tat-ELP produced an even stronger inhibition of migration.

Cell Proliferation and Apoptosis: As with the SKOV-3 cells above, the SK-MEL-2 cells were not able to proliferate 15 after plating in the presence of Pen-ELP or Tat-ELP (FIG. 9).

The DIC images from the spreading assay revealed the presence of some apoptotic cells after treatment with CPP-ELP. After learning that CPP-ELP treated cells were not proliferating, we used an annexin binding assay to quantitate 20 the extent of apoptosis after plating cells treated with CPP-ELP. SK-MEL-2 cells were treated and plated as above in 6-well plates. 5 h after plating, all cells (floating and attached) were collected by trypsinization, stained with FITC-annexin and propidium iodide (Molecular Probes, 25 Eugene, Oreg.), and analyzed by flow cytometry.

When treated with ELP, cells were unaffected and gave similar results to control cells in the apoptosis assay (FIG. 10). Pen-ELP produced a strong effect on the SK-MEL-2 cells, inducing apoptosis in nearly 90% of the cells within 30 the 5 h tested. Tat-ELP also induced apoptosis, but much less effectively than Pen-ELP. Although Tat-ELP is a much more efficient inhibitor of cell attachment than Pen-ELP, Tat-ELP does not induce the apoptotic response nearly as extensively as Pen-ELP. This suggests that the two polypeptides may have different modes of action. This possibility will be addressed by future in vitro experiments.

#### Example 5

#### Ovarian Cancer Metastasis

In North America and Europe, ovarian cancer is the fourth most common cause of cancer death among women and the prime cause of death among gynecological malignancies. 45 Primary tumors from the ovaries tend to spread throughout the abdominal cavity and to other organs or areas of the body forming metastases. The main route of metastatic dissemination of epithelial ovarian cancer is by exfoliation of the tumor cells, which migrate, implant, and invade throughout 50 the peritoneal cavity (33). The molecular mechanisms underlying this process are not well characterized, but it is clear that the attachment of cancer cells to the surfaces of other organs is one of the crucial steps in the development of metastatic ovarian cancer.

In Vitro Cell Attachment and Spreading: The in vitro cell attachment assay described above was used to assay the ability of the CPP-ELPs to inhibit attachment of SKOV-3 ovarian cancer cells, and the results are shown in FIG. 1.  $5\times10^4$  untreated cells plated attached to the substrate (as shown at zero polypeptide concentration in FIG. 11). The ELP polypeptide, which has no CPP to facilitate cell binding, showed no inhibition of cell attachment at any concentration tested. In contrast, Pen-ELP showed a concentration 65 dependent inhibition of SKOV-3 attachment, with complete inhibition observed at 5 µM Pen-ELP. Tat-ELP inhibited cell

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attachment even more efficiently, with a maximum inhibition occurring at only 0.5 µM. The inhibition of attachment was not simply a property of the CPP peptide, since the 16 aa penetratin peptide alone had no effect on cell attachment. These results show that both the CPP and ELP are required for attachment inhibition.

In addition to inhibiting attachment of the cells to the substrate, the CPP-ELPs also inhibited the spreading of any cells that did attach. SKOV-3 cells were incubated as described above and plated on acid-washed coverslips. The coverslips were mounted onto slides at various time points after plating, and the cell morphology was observed using differential interference contrast (DIC) microscopy. FIG. 12 shows that 1 h after plating, control cells were attached but still rounded. Fewer cells were attached after treatment with Pen-ELP and Tat-ELP, but little morphological difference was apparent at this time point. 4 h after plating, control and ELP treated cells began to spread onto the glass coverslips, and even more extensive spreading was observed at 7 h and 24 h. In contrast, the cells treated with Pen-ELP and Tat-ELP never spread onto the substrate. This spreading inhibition is not limited to cells plated on glass coverslips. Preliminary experiments with cells plated on fibronectin-coated coverslips show that both Pen-ELP and Tat-ELP can inhibit SKOV-3 cell spreading onto a natural ECM substratum.

Cell Migration: Since the CPP-ELP molecules showed potent inhibition of cell attachment and spreading, we also assayed for their ability to inhibit cell migration using the scratch migration assay described above. FIG. 13 shows that the scratch method removed all cells from the area of interest. 24 h later, control and ELP treated cells almost completely filled the cleared area. Pen-ELP significantly reduced the ability of the SKOV-3 cells to migrate into the cleared area, and Tat-ELP produced an even stronger inhi-35 bition of migration.

In order to confirm the role of CPP-ELPs in preventing migration of SKOV-3 cells, a boyden chamber assay was performed (35). This assay involves a membrane with pore size of 8 µm. SKOV-3 cells are plated on one side of the 40 membrane, and a chemoattractant (FBS) is placed on the opposite side of the membrane. The cells are allowed to migrate trough the pores toward the chemoattractant for h, and the cells which penetrated the membrane are quantified by cell counting and independently by Hoechst staining and fluorescence microscopy. As shown in FIG. 14 A, Tat-ELP treated cells did not migrate, while untreated, Tat peptide and ELP treated cells migrated across the membrane when counted under 20× magnification. This was also confirmed by Hoechst staining, which shows the number of cells migrated in FIG. 14 B. These experiments show that CPP-ELPs have a role in inhibiting migration. Serum starvation of cells overnight ruled out the fact that proliferation was a contributing factor in this assay.

Inhibition of SKOV-3 Attachment on Vitronectin: Cell 55 migration is governed by a variety of factors, including cell surface adhesion receptor binding to extracellular matrix (ECM) proteins. One such matrix protein is vitronectin (VN). VN is a widely distributed high molecular weight glycoprotein found in most extracellular matrices and blood During the 3 h attachment period, more than  $4 \times 10^4$  of the 60 plasma that is known to promote cell adhesion and affect cell morphology, migration, differentiation, and cytoskeletal organization. FIG. 15 shows that attachment was inhibited nearly 50% on VN treated plates by different CPP-ELPs, while ELP had no such effect.

> Cell Surface Receptor Assay: Cell migration is governed by a variety of factors, including cell surface adhesion receptor binding to extracellular matrix (ECM) proteins.

One such matrix protein is vitronectin (VN). VN is a widely distributed high molecular weight glycoprotein found in most extracellular matrices and blood plasma that is known to promote cell adhesion and affect cell morphology, migration, differentiation, and cytoskeletal organization. Cell 5 migration is governed by a variety of factors, including cell surface adhesion receptor binding to extracellular matrix (ECM) proteins. One such matrix protein is vitronectin (VN). VN is a widely distributed high molecular weight glycoprotein found in most extracellular matrices and blood 10 plasma that is known to promote cell adhesion and affect cell morphology, migration, differentiation, and cytoskeletal organization. Cell migration is governed by a variety of factors, including cell surface adhesion receptor binding to extracellular matrix (ECM) proteins. One such matrix pro- 15 tein is vitronectin (VN). VN is a widely distributed high molecular weight glycoprotein found in most extracellular matrices and blood plasma that is known to promote cell adhesion and affect cell morphology, migration, differentiation, and cytoskeletal organization. Since the action of 20 CPP-ELPs is mediated via interaction with the various cell surface receptors on the cell membrane, we assumed that treatment of cells with the enzymes mentioned below may provide some information about the binding characteristics of the CPP-ELPs.

Trypsin, Heparanase, and Pronase Treatment: Brief 3 min. incubation of cells with trypsin prior to polypeptide treatment showed nearly 20-40% decrease in Tat-ELP binding as compared to binding in normal cells (FIG. 16 A). Similarly, incubation of cells with heparanase (FIG. 16 B) and pronase 30 (FIG. 16 C) for specified time periods caused ~20-25% and 40-70% decrease in the respective binding of Tat-ELP over the concentration range. Binding of ELP polypeptide did not differ significantly under different enzymatic conditions. These enzymes have been reported to digest various cell 35 surface receptors such as heparin surface proteoglycans and glycoproteins which are involved in attachment and tumor metastases (36, 37). These results show that binding is decreased by the enzyme pretreatment and provide some information about the role of various cell surface receptors 40 and their interaction with CPP-ELPs.

Effect of Ionic Strength on Binding: It is important to explore the selective binding of CPP-ELPs to cell surfaces in greater detail, in particular with regard to ionic strength. Since the nature of the interaction between cell membrane 45 and CPP-ELPs is ionic due to their charged nature, we tested the binding of different polypeptides at different ionic concentrations. The solution's ionic strength was varied by changing the concentration of NaCl in PBS from 0.5 M to 2 M. The binding experiment was performed in each PBS 50 solution and compared to binding in standard PBS solution. It was observed that binding of Tat-ELP was enhanced by ~25-30% in ½ M PBS and reduced by ~18-40% in 2 M PBS when compared with binding of Tat-ELP in 1 M PBS (FIG. 17 A). The binding of uncharged polypeptide ELP (17 C) did 55 not show any significant difference at various PBS concentrations. This experiment demonstrates that the binding of the cationic peptide Tat to the cell membrane depends on the ionic composition of the media, with less ionic environment favoring more binding and vice versa.

Cell Proliferation: During the attachment and spreading assays, cells treated with CPP-ELP never attached or spread even 24 h after plating. We also examined the cells for a longer time period in order to determine if the cells were ever able to attach and to learn whether the cells can 65 proliferate after attachment. To address this question, we monitored the growth of SKOV-3 cells after treatment with

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CPP-ELP. Cells were treated as described above and plated in 6-well plates. Daily cell counts of the attached cells were made using a Coulter counter, and a growth curve (shown in FIG. 18) was generated. Control and ELP treated cells grew normally, making 8 to 10 doublings during the 6 days of the assay. In contrast, cells treated with Pen-ELP and Tat-ELP showed no increase in cell number over 6 days.

Inhibition of cell proliferation by CPP-ELP requires that the cells be treated while in suspension before plating. CPP-ELP shows no toxicity and no inhibition of growth when added to cells which are already attached to substratum (data not shown). These results suggest that inhibition of the proliferation of ovarian cancer cells by CPP-ELP treatment may be due to inhibition of cell attachment. This observation may be advantageous for the treatment of ovarian cancer metastasis, where the desired target cells are unattached and circulating.

Ovarian Peritoneal Metastasis In Vivo: The most probable use for an anti-metastatic agent such as CPP-ELP will be at the time of surgical resection of the primary tumor in order to prevent the spread of micrometastases resulting from tumor cells that were loosened or not removed during the surgery. Therefore, in order to test the ability of CPP-ELP to inhibit metastasis in a clinically relevant setting, we used a 25 rat ovarian metastasis model. In this assay,  $75 \times 10^6$  SKOV-3 cells grown in culture were injected into the peritoneal cavity of athymic nude rats. The cells were premixed with the most efficient attachment inhibitor, Tat-ELP at a concentration of 500 µM, or with PBS control. After injection, metastases were allowed to develop for 17 days. The animals were sacrificed, and the peritoneal metastases were carefully dissected from the attached normal tissue. The weight of each tumor was measured. The tumor burden per animals is reported as the sum of all tumor nodules from the small and large intestine, omentum, spleen, diaphragm, fallopian tubes, and bladder. As shown in the FIG. 19 A, the tumor burden/animal in the Tat-ELP treated group was nearly 50% less than the control group. SKOV-3 ovarian cancer cells have a high tendency to metastasize to the uterus and fallopian tubes. Therefore, the uterine tumor mass per animal was compared in untreated and treated groups. As shown in FIG. 19 B, Tat-ELP treated animals showed nearly 60% reduction in uterine tumor mass as compared to untreated animals.

In vivo Plasma Concentrations of CPP-ELP1-H1: In order to assess the plasma kinetics of the CPP-ELP polypeptide in vivo, preliminary studies were performed in athymic rats. A fluorescently labeled polypeptide (CPP-ELP-fluorescein) was injected as a bolus in anesthetized rats. The plasma concentration of this polypeptide was measured for five hours as shown in FIG. 20. The plasma concentration of the polypeptide declined to about 30% of its initial value during the first 45-60 min after injection, then showed little further decline over the following 4 hours. The apparent volume of distribution for the distribution phase of the curve was 35.5±9.7 ml, which indicates that the volume is over three times the estimated plasma volume (~7% of 150 g rat). The relatively stable concentration of the agent after the first 45 minutes is likely due to the large molecular weight of the 60 polypeptide, but may also be due to significant protein and cellular binding. Further characterization of the pharmacokinetic parameters and binding properties for several CPP-ELP polypeptides are planned in the future.

Because diagnosis of ovarian cancer occurs most often only after the disease has progressed, treatment is limited to surgical resection in combination with chemotherapy. It has been observed that an increased incidence of metastasis

occurs after this surgical manipulation (38). Therefore, the development of an antimetastatic agent that could be administered at or before the time of surgery would reduce the chance of post-surgical metastases forming. Thus, embodiments of the present invention are non-toxic agents given as adjuvant therapy to patients at the time of surgery, and could complement chemotherapy to slow the spread of remaining cancerous cells. Our results have shown that treatment of unattached ovarian cancer cells with CPP-ELPs inhibits their attachment, spreading, migration, invasion and proliferation in vitro and peritoneal metastasis of ovarian cells in vivo. Therefore, CPP-ELP has potential in anti-metastatic therapy to improve the cure rate for surgically resected ovarian tumors.

#### Example 6

# Test Regarding Present Invention and C6 Cell Proliferation

CPP-ELP-H1 is an effective inhibitor of breast cancer cell proliferation in vitro (see above), and its efficacy for breast cancer therapy in vivo is currently being established. However, malignant glioma is a cancer that is much more difficult to treat and with a much lower cure rate than breast cancer. 25 Therefore, developmental therapeutics for malignant glioma is a field that could greatly benefit from the targeted approach applied with ELP technology. In order to examine the antiproliferative effects of the CPP-ELP-H1 polypeptides in glioma cells, C6 cells were exposed for 1 hour to 20 30 μM Bac-ELP1-H1 or Bac-ELP2-H1 at 37° C. or 42° C. one day after cell seeding. The polypeptides were washed away, and the cells were allowed to grow until day 3. Cell number was determined using the MTS assay. The resulting data show that the thermally sensitive peptide Bac-ELP1-H1 did 35 not inhibit cell proliferation when cells were treated at 37° C. However, when cells were treated at 42° C., cell proliferation was inhibited by up to 60%.

The nonthermally responsive control, Bac-ELP2-H1, had no effect on C6 proliferation, and control polypeptides 40 lacking the c-Myc inhibitory sequence (Bac-ELP1 or Bac-ELP2) did not have any effect on cell proliferation. These results suggest that the polypeptides exhibit an antiproliferative effect in C-6 cells, which can be further enhanced by hyperthermia.

Development of the C6 Gliobastoma Model: This study will utilize an intracerebral tumor-bearing rat model of glioma (C6). The rat glioma model is similar to human malignant glioma (glioblastoma multiforme) both histologically and in rapid proliferation. In a previous study, C6 50 gliomas were induced by intracranial injection of a suspension of C6 cells. The tumors were imaged sequentially with 3-D volume measurements generated by means of a clinical magnetic resonance imaging system (CMRI) and commercially available wrist coil. This study demonstrated that 55 gliomas can be reliably grown in rats using the C6 cell line, and MRI imaging is an effective means of monitoring tumor progression.

Heating intracerebral C6 tumors using infrared light: Our previous studies in subcutaneous tumor models have used 60 infrared (IR) light generated by the Laser Systim 540® (Mettler Electronics) to heat the tumors. This method is preferred over more primitive techniques such as water bath immersion because the heat can be applied to a more concentrated area and without physical contact with the 65 animal. In order to test the effectiveness of this method in the glioma model, three representative C6 tumors grown in rat

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brains were used in a heating trial. The tumor core reached the desired hyperthermia temperature within 15 minutes of the start of illumination, and the temperature remained in the desired hyperthermia range for the remainder of an hour. This experiment demonstrates that the use of IR light is a feasible and minimally invasive method of heating intracerebral C6 tumors.

Following the heating period, the rats were exsanguinated and perfused with 4% paraformaldehyde, and the brains were removed, sectioned, and stained with hematoxylin and eosin (H&E). All three animals developed tumors, and the tumors were highly vascularized. Additionally, the tumor tissue displayed rosettes that are characteristic of glioblastoma. The area around the tumor appeared as normal neural tissue, and no acute damage from the hyperthermia treatment was apparent.

Imaging Intratumor Distribution of the Therapeutic Peptide Carrier: Previous studies have shown that ELP accumulation in tumor vasculature or interstitium can be 20 increased with focused hyperthermia. However, entry of the ELP carrier into the tumor cells, a property necessary for effective drug delivery, has never been demonstrated. The use of CPPs fused to the ELP carrier may enhance its uptake into the tumor cells. To test the ability of the Bac and Tat CPPs to enhance ELP uptake into tumor cells in vivo, rats bearing C6 tumors were intravenously injected with Rhodamine-labeled Bac-ELP1-H1 or Tat-ELP1-H1. One tumor was heated for 60 min. using IR illumination as described above. 1 min prior to euthanasia, high molecular weight (500 kDa) FITC-dextran was injected IV in order to mark the perfused vessels. The tumors were removed, rapidly frozen, and sectioned using a cryomicrotome. Tumor sections were fixed and stained with Hoescht 33342 and imaged using a Nikon fluorescence microscope with a CoolSnap CCD camera. Bac-ELP1-H1 is present not only in the tumor blood vessels, but it also escaped circulation and entered the tumor cells. Below the T<sub>t</sub>, Bac-ELP1-H1 is present in the cytoplasm of the tumor cells. Above the T<sub>t</sub>, the polypeptide can also be detected in the tumor cell nuclei. This is consistent with the localization of the polypeptide in cultured cells. Tat-ELP1-H1 is also able to escape the tumor vasculature and enter the tumor cells. Tat-ELP1-H1 was present in the cell cytoplasm at temperature above and below the  $T_t$ , again consistent with its subcellular localiza-45 tion in vitro. Thus, this Example indicates a direct observation that ELP aided by a CPP can escape the tumor vasculature and enter the tumor cells.

Tumor Size Reduction by the ELP-delivered c-Myc Inhibitory Peptide: In order to assess the ability of the ELP-delivered c-Myc inhibitory peptide to reduce tumor size, rats bearing 2 C6 tumors implanted subcutaneously in the rats' back were treated by IV injection of Bac-ELP1-H1 (130 mg/kg) or saline control, and one tumor was heated for 60 min using the IR heating method. Bac-ELP1-H1 was used in this study because it was found to be the most potent inhibitor of cell proliferation in vitro (unpublished data). Following treatment, the animals were returned to their cages, and tumor size and body weight was monitored for 19 days. In saline treated rats, the tumors proliferated rapidly up to a volume of 4000 mm<sup>3</sup>, and hyperthermia alone had no effect on tumor size. In contrast, the tumors in animals treated with Bac-ELP1-H1 began to shrink 4 days after treatment, and were nearly undetectable at day 19. Both the heated and unheated tumors in the treated animals were eventually cleared after treatment, but the heated tumor was reduced slightly faster. This indicates that at a dose of 130 mg/kg, Bac-ELP-H1 is potent enough to completely elimi-

nate C6 tumor growth. Further studies are underway at lower doses in order to look for a more significant difference between the heated and unheated tumors. No body weight loss, injection site reactions, or gross signs of toxicity were observed.

Rats were sacrificed on Day 19 after implantation, and tumors were removed and weighed. Untreated animals had tumors weighing approximately 2 g, and there was no significant difference between heated and unheated tumors. Animals treated with Bac-ELP1-H1, however, had significantly smaller tumors, with an average weight of less than 300 mg. No significant difference was seen between heated and unheated Bac-ELP1-H1 treated tumors, again supporting the evidence that 130 mg/kg is a sufficient dose to cause complete tumor regression even without hyperthermia.

Evaluation of Toxicity of the Bac-ELP1-H1 Polypeptide: In addition to monitoring tumor size during the above experiment, rats were monitored for weight loss or other signs of acute toxicity due to the Bac-ELP1-H1 treatment. The average body weight of the animals did not differ 20 between saline and Bac-ELP1-H1 treated groups, and no weight loss was observed following Bac-ELP1-H1 treatment. In addition, all major organs were removed and weighed at necropsy on Day 19. No significant difference was seen in organ weights from animals treated with saline 25 control and animals treated with Bac-ELP1-H1. Also, no hair loss or injection site reactions were observed, and no other signs of toxicity were noticed.

Throughout this application, and specifically, below, various references are mentioned. All references are incorporated herein by reference in their entirety and should be considered to be part of this application.

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Various changes in the details, steps and materials that have been described may be made by those skilled in the art within the principles and scope of the invention herein illustrated and defined in the appended claims. Therefore, while the present invention has been shown and described herein in what is believed to be the most practical and preferred embodiment, it is recognized that departures can be made therefrom within the scope of the invention, which is therefore not to be limited to the details disclosed herein but is to be accorded the full scope of the claims so as to embrace any and all equivalent apparatus and methods.

Unless defined otherwise, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which the presently-disclosed subject matter belongs. Although any methods, devices, and materials similar or equivalent to those described herein can be used in the practice or testing of the presently-disclosed subject matter, representative methods, devices, and materials are now described.

Following long-standing patent law convention, the terms "a", "an", and "the" refer to "one or more" when used in this application, including the claims. Thus, for example, refer-30 ence to "a cell" includes a plurality of such cells, and so forth.

Unless otherwise indicated, all numbers expressing quantities of ingredients, properties such as reaction conditions, and so forth used in the specification and claims are to be 34. Massodi, I., Bidwell, G. L., 3rd, and Raucher, D. 35 understood as being modified in all instances by the term "about". Accordingly, unless indicated to the contrary, the numerical parameters set forth in this specification and claims are approximations that can vary depending upon the desired properties sought to be obtained by the presentlydisclosed subject matter.

> As used herein, the term "about," when referring to a value or to an amount of mass, weight, time, volume, concentration or percentage is meant to encompass variations of in some embodiments ±20%, in some embodiments ±10%, in some embodiments ±5%, in some embodiments  $\pm 1\%$ , in some embodiments  $\pm 0.5\%$ , and in some embodiments ±0.1% from the specified amount, as such variations are appropriate to perform the disclosed method.

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Pro 545	Gly	Gly	Gly	Val	Pro 550	Gly	Ala	Gly	Val	Pro 555	Gly	Gly	Gly	Val	Pro 560
Gly	Ala	Gly	Val	Pro	Gly	Gly	Gly	Val	Pro	Gly	Ala	Gly	Val	Pro	Gly

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			565					570					575	
Val Gly	Val	Pro 580	Gly	Ala	Gly	Val	Pro 585	Gly	Gly	Gly	Val	Pro 590	Gly	Ala
Gly Val	Pro 595	_	Gly	Gly	Val	Pro 600	Gly	Ala	Gly	Val	Pro 605	Gly	Gly	Gly
Val Pro	_	Ala	Gly	Val	Pro 615	Gly	Gly	Gly	Val	Pro 620	Gly	Ala	Gly	Val
Pro Gly 625	Gly	Gly	Val	Pro 630	Gly	Ala	Gly	Val	Pro 635	Gly	Gly	Gly	Val	Pro 640
Gly Ala	Gly	Val	Pro 645	Gly	Gly	Gly	Val	Pro 650	Gly	Ala	Gly	Val	Pro 655	Gly
Val Gly	Val	Pro 660	Gly	Ala	Gly	Val	Pro 665	Gly	Gly	Gly	Val	Pro 670	Gly	Ala
Gly Val	Pro 675	_	Gly	Gly	Val	Pro 680	Gly	Ala	Gly	Val	Pro 685	Gly	Gly	Gly
Val Pro	_	Ala	Gly	Val	Pro 695	Gly	Gly	Gly	Val	Pro 700	Gly	Ala	Gly	Val
Pro Gly 705	Gly	Gly	Val	Pro 710	Gly	Ala	Gly	Val	Pro 715	_	Gly	Gly	Val	Pro 720
Gly Ala	Gly	Val	Pro 725	Gly	Gly	Gly	Val	Pro 730	Gly	Ala	Gly	Val	Pro 735	Gly
Val Gly	Val	Pro 740	Gly	Ala	Gly	Val	Pro 745	Gly	Gly	Gly	Val	Pro 750	Gly	Ala
Gly Val	Pro 755	_	Gly	Gly	Val	Pro 760	Gly	Ala	Gly	Val	Pro 765	Gly	Gly	Gly
Val Pro	_	Ala	Gly	Val	Pro 775	Gly	Gly	Gly	Val	Pro 780	Gly	Ala	Gly	Val
Pro Gly 785	Gly	Gly	Val	Pro 790	Gly	Ala	Gly	Val	Pro 795	Gly	Gly	Gly	Val	Pro 800
Gly Ala	Gly	Val	Pro 805	Gly	Gly	Gly	Val	Pro 810	Gly	Ala	Gly	Val	Pro 815	Gly
Trp Pro	Gly	Ser 820	Gly	Gly	Cys									
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Lys Gly Cys Gly Pro Gly Val Gly Val Pro Gly Val Gly Val Pro Gly 25

Gly Gly Val Pro Gly Ala Gly Val Pro Gly Val Gly Val Pro Gly Val
35

Gly Val Pro Gly Val Gly Val Pro Gly Gly Val Pro Gly Ala Gly 50

Val Pro Gly Gly Val Pro Gly Val Gly Val Pro Gly Val Gly Val 65 70 80

Pro Gly Gly Val Pro Gly Ala Gly Val Pro Gly Val Gly Val Pro 85

Gly Val Gly Val Pro Gly Val Gly Val Pro Gly Gly Val Pro Gly

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COILCILIACA

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Ala	Gly	Val 115	Pro	Gly	Gly	Gly	Val 120	Pro	Gly	Val	Gly	Val 125	Pro	Gly	Val
Gly	Val 130	Pro	Gly	Gly	Gly	Val 135	Pro	Gly	Ala	Gly	Val 140	Pro	Gly	Val	Gly
Val 145	Pro	Gly	Val	Gly	Val 150	Pro	Gly	Val	Gly	Val 155	Pro	Gly	Gly	Gly	Val 160
Pro	Gly	Ala	Gly	Val 165	Pro	Gly	Gly	Gly	Val 170		Gly	Val	Gly	Val 175	Pro
Gly	Val	Gly	Val 180	Pro	Gly	Gly	Gly	Val 185	Pro	Gly	Ala	Gly	Val 190	Pro	Gly
Val	Gly	Val 195	Pro	Gly	Val	Gly	Val 200	Pro	Gly	Val	Gly	Val 205	Pro	Gly	Gly
Gly	Val 210	Pro	Gly	Ala	Gly	Val 215	Pro	Gly	Gly	Gly	Val 220	Pro	Gly	Val	Gly
Val 225	Pro	Gly	Val	Gly	Val 230	Pro	Gly	Gly	Gly	Val 235	Pro	Gly	Ala	Gly	Val 240
Pro	Gly	Val	Gly	Val 245	Pro	Gly	Val	Gly	Val 250	Pro	Gly	Val	Gly	Val 255	Pro
Gly	Gly	Gly	Val 260	Pro	Gly	Ala	Gly	Val 265	Pro	Gly	Gly	Gly	Val 270	Pro	Gly
Val	Gly	Val 275	Pro	Gly	Val	Gly	Val 280	Pro	Gly	Gly	Gly	Val 285	Pro	Gly	Ala
Gly	Val 290	Pro	Gly	Val	Gly	Val 295	Pro	Gly	Val	Gly	Val 300	Pro	Gly	Val	Gly
Val 305	Pro	Gly	Gly	Gly			Gly			Val 315	Pro	Gly	Gly	Gly	Val 320
Pro	Gly	Val	Gly	Val 325	Pro	Gly	Val	Gly	Val 330	Pro	Gly	Gly	Gly	Val 335	Pro
Gly	Ala	Gly	Val 340	Pro	Gly	Val	Gly	Val 345	Pro	Gly	Val	Gly	Val 350	Pro	Gly
Val	Gly	Val 355	Pro	Gly	Gly	Gly	Val 360	Pro	Gly	Ala	Gly	Val 365	Pro	Gly	Gly
Gly	Val 370	Pro	Gly	Val	Gly	Val 375	Pro	Gly	Val	Gly	Val 380	Pro	Gly	Gly	Gly
Val 385	Pro	Gly	Ala	Gly	Val 390	Pro	Gly	Val	Gly	Val 395	Pro	Gly	Val	Gly	Val 400
Pro	Gly	Val	Gly	Val 405	Pro	Gly	Gly	Gly	Val 410	Pro	Gly	Ala	Gly	Val 415	Pro
Gly	Gly	Gly	Val 420	Pro	Gly	Val	Gly	Val 425	Pro	Gly	Val	Gly	Val 430	Pro	Gly
Gly	Gly	Val 435	Pro	Gly	Ala	Gly	Val 440	Pro	Gly	Val	Gly	Val 445	Pro	Gly	Val
Gly	Val 450	Pro	Gly	Val	Gly	Val 455	Pro	Gly	Gly	Gly	Val 460	Pro	Gly	Ala	Gly
Val 465	Pro	Gly	Gly	Gly	Val 470	Pro	Gly	Val	Gly	Val 475	Pro	Gly	Val	Gly	Val 480
Pro	Gly	Gly	Gly	Val 485	Pro	Gly	Ala	Gly	Val 490	Pro	Gly	Val	Gly	Val 495	Pro
Gly	Val	Gly	Val 500	Pro	Gly	Val	Gly	Val 505	Pro	Gly	Gly	Gly	Val 510	Pro	Gly
Ala	Gly	Val 515	Pro	Gly	Gly	Gly	Val 520	Pro	Gly	Val	Gly	Val 525	Pro	Gly	Val

Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly Val Pro Gly Val Gly Val Pro Gly Val Gly Val Pro Gly Val Gly Val Pro Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Gly Val Pro Gly Val Gly Val Pro Gly Val Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly Val Pro Gly Val Gly Val Pro Gly Val Gly Val Pro Gly Val Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Gly Val Pro Gly Val Gly Val Pro Gly Val Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly Val Pro Gly Val Gly Val Pro Gly Val Gly Val Pro Gly Val Gly Val Pro Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Gly Val Pro Gly Val Gly Val Pro Gly Val Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly Val Pro Gly Val Gly Val Pro Gly Val Gly Val Pro Gly Val Gly Val Pro Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Val Pro Gly Val Gly Val Pro Gly Val Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly Val Pro Gly Val Gly Val Pro Gly Val Gly Val Pro Gly Val Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Gly Val Pro Gly Trp Pro <210> SEQ ID NO 26 <211> LENGTH: 824 <212> TYPE: PRT <213 > ORGANISM: Artificial Sequence <220> FEATURE: <223> OTHER INFORMATION: Antp-ELP2 <400> SEQUENCE: 26 Met Arg Gln Ile Lys Ile Trp Phe Gln Asn Arg Arg Met Lys Trp Lys Lys Gly Cys Gly Pro Gly Val Gly Val Pro Gly Ala Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Val Pro Gly Ala Gly Val Pro Gly Val Gly Val Pro Gly Ala Gly Val Pro Gly 

Gly	Gly	Val 115	Pro	Gly	Ala	Gly	Val 120	Pro	Gly	Gly	Gly	Val 125	Pro	Gly	Ala
Gly	Val 130	Pro	Gly	Gly	Gly	Val 135	Pro	Gly	Ala	Gly	Val 140	Pro	Gly	Gly	Gly
Val 145	Pro	Gly	Ala	Gly	Val 150	Pro	Gly	Gly	Gly	Val 155	Pro	Gly	Ala	Gly	Val 160
Pro	Gly	Gly	Gly	Val 165	Pro	Gly	Ala	Gly	Val 170	Pro	Gly	Gly	Gly	Val 175	Pro
Gly	Ala	Gly	Val 180	Pro	Gly	Val	Gly	Val 185	Pro	Gly	Ala	Gly	Val 190	Pro	Gly
Gly	Gly	Val 195	Pro	Gly	Ala	Gly	Val 200	Pro	Gly	Gly	Gly	Val 205	Pro	Gly	Ala
Gly	Val 210	Pro	Gly	Gly	Gly	Val 215	Pro	Gly	Ala	Gly	Val 220	Pro	Gly	Gly	Gly
Val 225	Pro	Gly	Ala	Gly	Val 230	Pro	Gly	Gly	Gly	Val 235	Pro	Gly	Ala	Gly	Val 240
Pro	Gly	Gly	Gly	Val 245	Pro	Gly	Ala	Gly	Val 250	Pro	Gly	Gly	Gly	Val 255	Pro
Gly	Ala	Gly	Val 260	Pro	Gly	Val	Gly	Val 265	Pro	Gly	Ala	Gly	Val 270	Pro	Gly
Gly	Gly	Val 275	Pro	Gly	Ala	Gly	Val 280	Pro	Gly	Gly	Gly	Val 285	Pro	Gly	Ala
Gly	Val 290	Pro	Gly	Gly	Gly	Val 295	Pro	Gly	Ala	Gly	Val 300	Pro	Gly	Gly	Gly
Val 305	Pro	Gly	Ala	Gly	Val 310	Pro	Gly	Gly	Gly	Val 315	Pro	Gly	Ala	Gly	Val 320
Pro	Gly	Gly	Gly	Val 325	Pro	Gly	Ala	Gly	Val 330	Pro	Gly	Gly	Gly	Val 335	Pro
Gly	Ala	Gly	Val 340	Pro	Gly	Val	Gly	Val 345	Pro	Gly	Ala	Gly	Val 350	Pro	Gly
Gly	Gly	Val 355	Pro	Gly	Ala	Gly	Val 360	Pro	Gly	Gly	Gly	Val 365	Pro	Gly	Ala
Gly	Val 370	Pro	Gly	Gly	Gly	Val 375	Pro	Gly	Ala	Gly	Val 380	Pro	Gly	Gly	Gly
Val 385	Pro	Gly	Ala	Gly	Val 390	Pro	Gly	Gly	Gly	Val 395	Pro	Gly	Ala	Gly	Val 400
Pro	Gly	Gly	Gly	Val 405	Pro	Gly	Ala	Gly	Val 410	Pro	Gly	Gly	Gly	Val 415	Pro
Gly	Ala	Gly	Val 420	Pro	Gly	Val	Gly	Val 425	Pro	Gly	Ala	Gly	Val 430	Pro	Gly
Gly	Gly	Val 435	Pro	Gly	Ala	Gly	Val 440	Pro	Gly	Gly	Gly	Val 445	Pro	Gly	Ala
Gly	Val 450	Pro	Gly	Gly	Gly	Val 455	Pro	Gly	Ala	Gly	Val 460	Pro	Gly	Gly	Gly
Val 465	Pro	Gly	Ala	Gly	Val 470	Pro	Gly	Gly	Gly	Val 475	Pro	Gly	Ala	Gly	Val 480
Pro	Gly	Gly	Gly	Val 485	Pro	Gly	Ala	Gly	Val 490	Pro	Gly	Gly	Gly	Val 495	Pro
Gly	Ala	Gly	Val 500	Pro	Gly	Val	Gly	Val 505	Pro	Gly	Ala	Gly	Val 510	Pro	Gly
Gly	Gly	Val 515	Pro	Gly	Ala	Gly	Val 520	Pro	Gly	Gly	Gly	Val 525	Pro	Gly	Ala

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Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Val Pro Gly Ala Gly Val Pro Gly Val Gly Val Pro Gly Ala Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Val Pro Gly Ala Gly Val Pro Gly Val Gly Val Pro Gly Ala Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Val Pro Gly Ala Gly Val Pro Gly Val Gly Val Pro Gly Ala Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Val Pro Gly Ala Gly Val Pro Gly Trp Pro <210> SEQ ID NO 27 <211> LENGTH: 778 <212> TYPE: PRT <213 > ORGANISM: Artificial Sequence <220> FEATURE:

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Pro Gly Gly Pro Gly Val Gly Val Gly Val Pro Gly Gly 30

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Val Pro Gly Val Gly Val Pro Gly Gly Val Pro Gly Ala Gly Val 50

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						31					_	con	tin <sup>.</sup>	ued	
Pro 65	Gly	Gly	Gly	Val	Pro 70	Gly	Val	Gly	Val	Pro 75	Gly	Val	Gly	Val	Pro 80
Gly	Gly	Gly	Val	Pro 85	Gly	Ala	Gly	Val	Pro 90	Gly	Val	Gly	Val	Pro 95	Gly
Val	Gly	Val	Pro 100	Gly	Val	Gly	Val	Pro 105	Gly	Gly	Gly	Val	Pro 110	Gly	Ala
Gly	Val	Pro 115	Gly	Gly	Gly	Val	Pro 120	Gly	Val	Gly	Val	Pro 125	Gly	Val	Gly
Val	Pro 130	Gly	Gly	Gly	Val	Pro 135	Gly	Ala	Gly	Val	Pro 140	Gly	Val	Gly	Val
Pro 145	Gly	Val	Gly	Val	Pro 150	Gly	Val	Gly	Val	Pro 155	Gly	Gly	Gly	Val	Pro 160
Gly	Ala	Gly	Val	Pro 165	Gly	Gly	Gly	Val	Pro 170	Gly	Val	Gly	Val	Pro 175	Gly
Val	Gly	Val	Pro 180	Gly	Gly	Gly	Val	Pro 185	Gly	Ala	Gly	Val	Pro 190	Gly	Val
Gly	Val	Pro 195	Gly	Val	Gly	Val	Pro 200	Gly	Val	Gly	Val	Pro 205	Gly	Gly	Gly
Val	Pro 210	Gly	Ala	Gly	Val	Pro 215	Gly	Gly	Gly	Val	Pro 220	Gly	Val	Gly	Val
Pro 225	_	Val	_			_	_	_			_	Ala	_	Val	Pro 240
Gly	Val	Gly	Val	Pro 245	Gly	Val	Gly	Val	Pro 250	Gly	Val	Gly	Val	Pro 255	Gly
Gly	Gly	Val	Pro 260	Gly	Ala	Gly	Val	Pro 265	Gly	Gly	Gly	Val	Pro 270	Gly	Val
Gly	Val	Pro 275	Gly	Val	Gly	Val	Pro 280	Gly	Gly	Gly	Val	Pro 285	Gly	Ala	Gly
Val	Pro 290	Gly	Val	Gly	Val	Pro 295	Gly	Val	Gly	Val	Pro 300	Gly	Val	Gly	Val
Pro 305	Gly	Gly	Gly	Val	Pro 310	Gly	Ala	Gly	Val	Pro 315	Gly	Gly	Gly	Val	Pro 320
Gly	Val	Gly	Val	Pro 325	Gly	Val	Gly	Val	Pro 330	Gly	Gly	Gly	Val	Pro 335	Gly

Ala Gly Val Pro Gly Val Gly Val Pro Gly Val Gly Val Pro Gly Val

Gly Val Pro Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly

Val Pro Gly Val Gly Val Pro Gly Val Gly Val Pro Gly Gly Val

Pro Gly Ala Gly Val Pro Gly Val Gly Val Pro Gly Val Gly Val Pro

Gly Val Gly Val Pro Gly Gly Val Pro Gly Ala Gly Val Pro Gly

Gly Gly Val Pro Gly Val Gly Val Pro Gly Val Gly Val Pro Gly Gly

Gly Val Pro Gly Ala Gly Val Pro Gly Val Gly Val Pro Gly Val Gly

Val Pro Gly Val Gly Val Pro Gly Gly Val Pro Gly Ala Gly Val

Pro Gly Gly Val Pro Gly Val Gly Val Pro Gly Val Gly Val Pro

Gly Gly Val Pro Gly Ala Gly Val Pro Gly Val Gly Val Pro Gly

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Gly	Val	Pro 515	Gly	Gly	Gly	Val	Pro 520	Gly	Val	Gly	Val	Pro 525	Gly	Val	Gly
Val	Pro 530	Gly	Gly	Gly	Val	Pro 535	Gly	Ala	Gly	Val	Pro 540	Gly	Val	Gly	Val
Pro 545	Gly	Val	Gly	Val		_	Val	_		Pro 555	Gly	Gly	Gly	Val	Pro 560
Gly	Ala	Gly	Val	Pro 565	Gly	Gly	Gly	Val	Pro 570	Gly	Val	Gly	Val	Pro 575	Gly
Val	Gly	Val	Pro 580	Gly	Gly	Gly	Val	Pro 585	Gly	Ala	Gly	Val	Pro 590	Gly	Val
Gly	Val	Pro 595	Gly	Val	Gly	Val	Pro 600	Gly	Val	Gly	Val	Pro 605	Gly	Gly	Gly
Val	Pro 610	Gly	Ala	Gly	Val	Pro 615	Gly	Gly	Gly	Val	Pro 620	Gly	Val	Gly	Val
Pro 625	Gly	Val	Gly	Val	Pro 630	Gly	Gly	Gly	Val	Pro 635	Gly	Ala	Gly	Val	Pro 640
Gly	Val	Gly	Val	Pro 645	Gly	Val	Gly	Val	Pro 650	Gly	Val	Gly	Val	Pro 655	Gly
Gly	Gly	Val	Pro 660	Gly	Ala	Gly	Val	Pro 665	Gly	Gly	Gly	Val	Pro 670	Gly	Val
Gly	Val	Pro 675	_	Val	Gly	Val	Pro 680	Gly	Gly	Gly	Val	Pro 685	Gly	Ala	Gly
Val	Pro 690	_		_			Gly		_		Pro 700	_	Val	Gly	Val
Pro 705	Gly	Gly	Gly	Val	Pro 710	Gly	Ala	Gly	Val	Pro 715	Gly	Gly	Gly	Val	Pro 720
Gly	Val	Gly	Val	Pro 725	Gly	Val	Gly	Val	Pro 730	Gly	Gly	Gly	Val	Pro 735	Gly
Ala	Gly	Val	Pro 740	Gly	Val	Gly	Val	Pro 745	Gly	Val	Gly	Val	Pro 750	Gly	Val
Gly	Val	Pro 755	Gly	Gly	Gly	Val	Pro 760	Gly	Ala	Gly	Val	Pro 765	Gly	Gly	Gly
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Gly Val Pro Gly Ala Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly 

Val Pro Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Val 

Pro Gly Ala Gly Val Pro Gly Gly Val Pro Gly Ala Gly Val Pro

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Gly	Gly	Gly	Val	Pro 85	Gly	Ala	Gly	Val	Pro 90	Gly	Gly	Gly	Val	Pro 95	Gly
Ala	Gly	Val	Pro 100	Gly	Val	Gly	Val	Pro 105	Gly	Ala	Gly	Val	Pro 110	Gly	Gly
Gly	Val	Pro 115	Gly	Ala	Gly	Val	Pro 120	Gly	Gly	Gly	Val	Pro 125	Gly	Ala	Gly
Val	Pro 130	Gly	Gly	Gly	Val	Pro 135	Gly	Ala	Gly	Val	Pro 140	Gly	Gly	Gly	Val
Pro 145	Gly	Ala	Gly	Val	Pro 150	Gly	Gly	Gly	Val	Pro 155	Gly	Ala	Gly	Val	Pro 160
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Ala	Gly	Val	Pro 180	Gly	Val	Gly	Val	Pro 185	Gly	Ala	Gly	Val	Pro 190	Gly	Gly
Gly	Val	Pro 195	Gly	Ala	Gly	Val	Pro 200	Gly	Gly	Gly	Val	Pro 205	Gly	Ala	Gly
Val	Pro 210	Gly	Gly	Gly	Val	Pro 215	Gly	Ala	Gly	Val	Pro 220	Gly	Gly	Gly	Val
Pro 225	Gly	Ala	Gly	Val	Pro 230	Gly	Gly	Gly	Val	Pro 235	Gly	Ala	Gly	Val	Pro 240
Gly	Gly	Gly	Val	Pro 245	Gly	Ala	Gly	Val	Pro 250	Gly	Gly	Gly	Val	Pro 255	Gly
Ala	Gly	Val	Pro 260	Gly	Val	Gly	Val	Pro 265	Gly	Ala	Gly	Val	Pro 270	Gly	Gly
Gly	Val	Pro 275	Gly	Ala	Gly	Val	Pro 280	Gly	Gly	Gly	Val	Pro 285	Gly	Ala	Gly
Val	Pro 290	Gly	Gly	Gly	Val	Pro 295	Gly	Ala	Gly	Val	Pro 300	Gly	Gly	Gly	Val
Pro 305	Gly	Ala	Gly	Val	Pro 310	Gly	Gly	Gly	Val	Pro 315	Gly	Ala	Gly	Val	Pro 320
Gly	Gly	Gly	Val	Pro 325	Gly	Ala	Gly	Val	Pro 330	Gly	Gly	Gly	Val	Pro 335	Gly
Ala	Gly	Val	Pro 340	Gly	Val	Gly	Val	Pro 345	Gly	Ala	Gly	Val	Pro 350	Gly	Gly
Gly	Val	Pro 355	Gly	Ala	Gly	Val	Pro 360	Gly	Gly	Gly	Val	Pro 365	Gly	Ala	Gly
Val	Pro 370	Gly	Gly	Gly	Val	Pro 375	Gly	Ala	Gly	Val	Pro 380	Gly	Gly	Gly	Val
Pro 385	Gly	Ala	Gly	Val	Pro 390	Gly	Gly	Gly	Val	Pro 395	Gly	Ala	Gly	Val	Pro 400
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Ala	Gly	Val	Pro 420	Gly	Val	Gly	Val	Pro 425	Gly	Ala	Gly	Val	Pro 430	Gly	Gly
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Val	Pro 450	Gly	Gly	Gly	Val	Pro 455	Gly	Ala	Gly	Val	Pro 460	Gly	Gly	Gly	Val
Pro 465	Gly	Ala	Gly	Val	Pro 470	Gly	Gly	Gly	Val	Pro 475	Gly	Ala	Gly	Val	Pro 480
Gly	Gly	Gly	Val	Pro 485	Gly	Ala	Gly	Val	Pro 490	Gly	Gly	Gly	Val	Pro 495	Gly

510

63

Ala Gly Val Pro Gly Val Gly Val Pro Gly Ala Gly Val Pro Gly Gly

505

Gly Val Pro Gly Ala Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly 515

500

Val Pro Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Val 530

Pro Gly Ala Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly Val Pro 545 550 550

Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Gly Val Pro Gly 575

Ala Gly Val Pro Gly Val Gly Val Pro Gly Ala Gly Val Pro Gly Gly 580 585

Gly Val Pro Gly Ala Gly Val Pro Gly Gly Val Pro Gly Ala Gly 595 600

Val Pro Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Val 610 620

Pro Gly Ala Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly Val Pro 625 630 635

Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Gly Val Pro Gly 645 650

Ala Gly Val Pro Gly Val Gly Val Pro Gly Ala Gly Val Pro Gly Gly 660 670

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Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Gly Val Pro Gly 735

Ala Gly Val Pro Gly Val Gly Val Pro Gly Ala Gly Val Pro Gly Gly 740 750

Gly Val Pro Gly Ala Gly Val Pro Gly Gly Val Pro Gly Ala Gly 755 760 765

Val Pro Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Val 770 780

Pro Gly Ala Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly Val Pro 795 795

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Ala Gly Val Pro Gly Trp Pro Gly Ser Gly Gly Cys 820

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<213 > ORGANISM: Artificial Sequence

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Arg Pro Leu Pro Phe Pro Arg Pro Gly Gly Gly Pro Gly Val Gly Val 20 25 30

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Gly 65	Gly	Val	Pro	Gly	Ala 70	Gly	Val	Pro	Gly	Gly 75	Gly	Val	Pro	Gly	Val 80
Gly	Val	Pro	Gly	Val 85	Gly	Val	Pro	Gly	Gly 90	Gly	Val	Pro	Gly	Ala 95	Gly
Val	Pro	Gly	Val 100	Gly	Val	Pro	Gly	Val 105	Gly	Val	Pro	Gly	Val 110	Gly	Val
Pro	Gly	Gly 115	_	Val	Pro	Gly	Ala 120	Gly	Val	Pro	Gly	Gly 125	Gly	Val	Pro
Gly	Val 130	Gly	Val	Pro	Gly	Val 135	Gly	Val	Pro	Gly	Gly 140	Gly	Val	Pro	Gly
Ala 145	Gly	Val	Pro	Gly	Val 150	Gly	Val	Pro	Gly	Val 155	Gly	Val	Pro	Gly	Val 160
Gly	Val	Pro	_		_			_		_		Pro	_		Gly
Val	Pro	Gly	Val 180	Gly	Val	Pro	Gly	Val 185	Gly	Val	Pro	Gly	Gly 190	Gly	Val
Pro	Gly	Ala 195	Gly	Val	Pro	Gly	Val 200	Gly	Val	Pro	Gly	Val 205	Gly	Val	Pro
Gly	Val 210	Gly	Val	Pro	Gly	Gly 215	Gly	Val	Pro	Gly	Ala 220	Gly	Val	Pro	Gly
Gly 225	Gly	Val	Pro	Gly	Val 230	Gly	Val	Pro	Gly	Val 235	Gly	Val	Pro	Gly	Gly 240
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Val	Pro	Gly	Val 260	Gly	Val	Pro	Gly	_	Gly		Pro	Gly	Ala 270	Gly	Val
Pro	Gly	Gly 275	_	Val	Pro	Gly	Val 280	Gly	Val	Pro	Gly	Val 285	Gly	Val	Pro
Gly	Gly 290	Gly	Val	Pro	Gly	Ala 295	Gly	Val	Pro	Gly	Val 300	Gly	Val	Pro	Gly
	Gly			_		_			_	<u>-</u>	_	Val		_	Ala 320
Gly	Val	Pro	Gly	Gly 325	Gly	Val	Pro	Gly	Val 330	Gly	Val	Pro	Gly	Val 335	Gly
Val	Pro	Gly	Gly 340	Gly	Val	Pro	Gly	Ala 345	Gly	Val	Pro	Gly	Val 350	Gly	Val
Pro	Gly	Val 355	Gly	Val	Pro	Gly	Val 360	Gly	Val	Pro	Gly	Gly 365	Gly	Val	Pro
Gly	Ala 370	Gly	Val	Pro	Gly	Gly 375	Gly	Val	Pro	Gly	Val 380	Gly	Val	Pro	Gly
Val 385	Gly	Val	Pro	Gly	Gly 390	Gly	Val	Pro	Gly	Ala 395	Gly	Val	Pro	Gly	Val 400
Gly	Val	Pro	Gly	Val 405	Gly	Val	Pro	Gly	Val 410	Gly	Val	Pro	Gly	Gly 415	Gly
Val	Pro	Gly	Ala 420	Gly	Val	Pro	Gly	Gly 425	Gly	Val	Pro	Gly	Val 430	Gly	Val
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Pro	Gly	Ala 115	Gly	Val	Pro	Gly	Gly 120	Gly	Val	Pro	Gly	Ala 125	Gly	Val	Pro
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Val Pro Gly Val Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly Val Pro Gly Val Gly Val Pro Gly Val Gly Val Pro Gly Val Gly Val Pro Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Gly Val Pro Gly Val Gly Val Pro Gly Val Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly Val Pro Gly Val Gly Val Pro Gly Val Gly Val Pro Gly Val Gly Val Pro Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Val Pro Gly Val Gly Val Pro Gly Val Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly Val Pro Gly Val Gly Val Pro Gly Val Gly Val Pro Gly Val Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Gly Val Pro Gly Val Gly Val Pro Gly Val Gly Val Pro Gly Gly Val Pro Gly Ala Gly Val Pro Gly Val Gly Val Pro Gly Val Gly Val Pro Gly Val Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Val Pro Gly Val Gly Val Pro Gly Val Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly Val Pro Gly Val Gly Val Pro Gly Val Gly Val Pro Gly Val Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Val Pro Gly Val Gly Val Pro Gly Val Gly Val Pro Gly Gly Val Pro Gly Ala Gly Val Pro Gly Val Gly Val Pro Gly Val Gly Val Pro Gly Val Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Gly Val Pro Gly Val Gly Val Pro Gly Val Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly Val Pro Gly Val Gly Val Pro Gly Val Gly Val Pro Gly Val Gly Val Pro Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Gly Val Pro Gly Val Gly Val Pro Gly Val Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly Val Pro Gly Val Gly Val Pro Gly Val Gly Val Pro Gly Val Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly Val Pro Gly Gly Gly Val Pro Gly Trp Pro Gly Ser Gly Gly Cys

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Gly	Val 370	Pro	Gly	Gly	Gly	Val 375		Gly	Ala	Gly	Val 380	Pro	Gly	Gly	Gly
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Gly	Ala	Gly	Val 500	Pro	Gly	Gly	Gly	Val 505	Pro	Gly	Ala	Gly	Val 510	Pro	Gly
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Gly Val Pro Gly Ala Gly Val Pro Gly Gly Gly Val Pro Gly Ala Gly

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We claim:

1. A method of inhibiting the metastasis of cancer, comprising:

administering an effective amount of a compound comprising a cell penetrating peptide (CPP) and an elastin-like protein (ELP) to a subject having one or more cancer cells, the compound binding to an exterior surface of the one or more cancer cells thereby inhibiting adhesion, spreading, invasion and migration of the one or more cancer cells to a metastatic site; wherein the compound is not conjugated to a therapeutic agent,

the CPP is selected from Tat, Penetratin, Bac-7, Trans- 50 cells are circulating cells.

portan, pVEC, MTS, and combinations thereof,

4. The method of claim

the ELP comprises the sequence (VPGXG (SEQ ID NO: 37))<sub>n</sub>, where n is an integer from about 30 to about 200 and each X is independently selected from valine (Val; V), glycine (Gly; G), and alanine (Ala; A), and

the cancer is selected from the group consisting of breast cancer, ovarian cancer, melanoma, and combinations thereof.

2. A method of inhibiting the metastasis of cancer, comprising:

administering an effective amount of a compound comprising a cell penetrating peptide (CPP) and an elastin-like protein (ELP) to a subject having one or more cancer cells, the compound binding to an exterior surface of the one or more cancer cells thereby inhib- 65 iting adhesion, spreading, invasion and migration of the one or more cancer cells to a metastatic site; wherein

the compound is not conjugated to a therapeutic agent, the CPP is selected from Tat, Bac-7, Transportan, pVEC, MTS, and combinations thereof,

the ELP comprises the sequence (VPGXG (SEQ ID NO: 37))<sub>n</sub>, where n is an integer from about 30 to about 200 and each X is independently selected from valine (Val; V), glycine (Gly; G), and alanine (Ala; A), and

the cancer is selected from the group consisting of breast cancer, ovarian cancer, melanoma, and combinations thereof.

- 3. The method of claim 1 wherein the one or more cancer cells are circulating cells.
- 4. The method of claim 2 wherein the one or more cancer cells are circulating cells.
- 5. A method of inhibiting the metastasis of cancer, comprising:

administering an effective amount of a compound consisting of a cell penetrating peptide (CPP) and an elastin-like protein (ELP) to a subject having one or more cancer cells, the compound binding to an exterior surface of the one or more cancer cells thereby inhibiting adhesion, spreading, invasion and migration of the one or more cancer cells to a metastatic site; wherein

the CPP is selected from Tat, Penetratin, Bac-7, Transportan, pVEC, MTS, and combinations thereof,

the ELP comprises the sequence (VPGXG (SEQ ID NO: 37))<sub>n</sub>, where n is an integer from about 30 to about 200 and each X is independently selected from valine (Val; V), glycine (Gly; G), and alanine (Ala; A), and

the cancer is selected from the group consisting of breast cancer, ovarian cancer, melanoma, and combinations thereof.

- 6. The method of claim 5 wherein the one or more cancer cells are circulating cells.
- 7. A method of inhibiting the metastasis of cancer, comprising:

administering an effective amount of a compound consisting of a cell penetrating peptide (CPP) and an elastin-like protein (ELP) to a subject having one or 10 more cancer cells, the compound binding to an exterior surface of the one or more cancer cells thereby inhibiting adhesion, spreading, invasion and migration of the one or more cancer cells to a metastatic site; wherein

the CPP is selected from Tat, Bac-7, Transportan, pVEC, 15 MTS, and combinations thereof,

the ELP comprises the sequence (VPGXG (SEQ ID NO: 37))<sub>n</sub>, where n is an integer from about 30 to about 200 and each X is independently selected from valine (Val; V), glycine (Gly; G), and alanine (Ala; A), and

the cancer is selected from the group consisting of breast cancer, ovarian cancer, melanoma, and combinations thereof.

8. The method of claim 7 wherein the one or more cancer cells are circulating cells.

\* \* \* \* \*