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(54) **ENHANCED NUCLEIC ACID CONSTRUCTS
FOR EUKARYOTIC GENE EXPRESSION**

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(57) **ABSTRACT**

The present invention provides polynucleotide vectors for
high expression of heterologous genes, and methods for
constructing such vectors. Some vectors further comprise
novel transposons and transposases that further improve
expression. Further disclosed are vectors that can be used in
a gene transfer system for stably introducing nucleic acids
into the DNA of a cell. The gene transfer systems can be
used in methods, for example, but not limited to, gene
expression, gene therapy, insertional mutagenesis, or gene
discovery.

8 Claims, 5 Drawing Sheets

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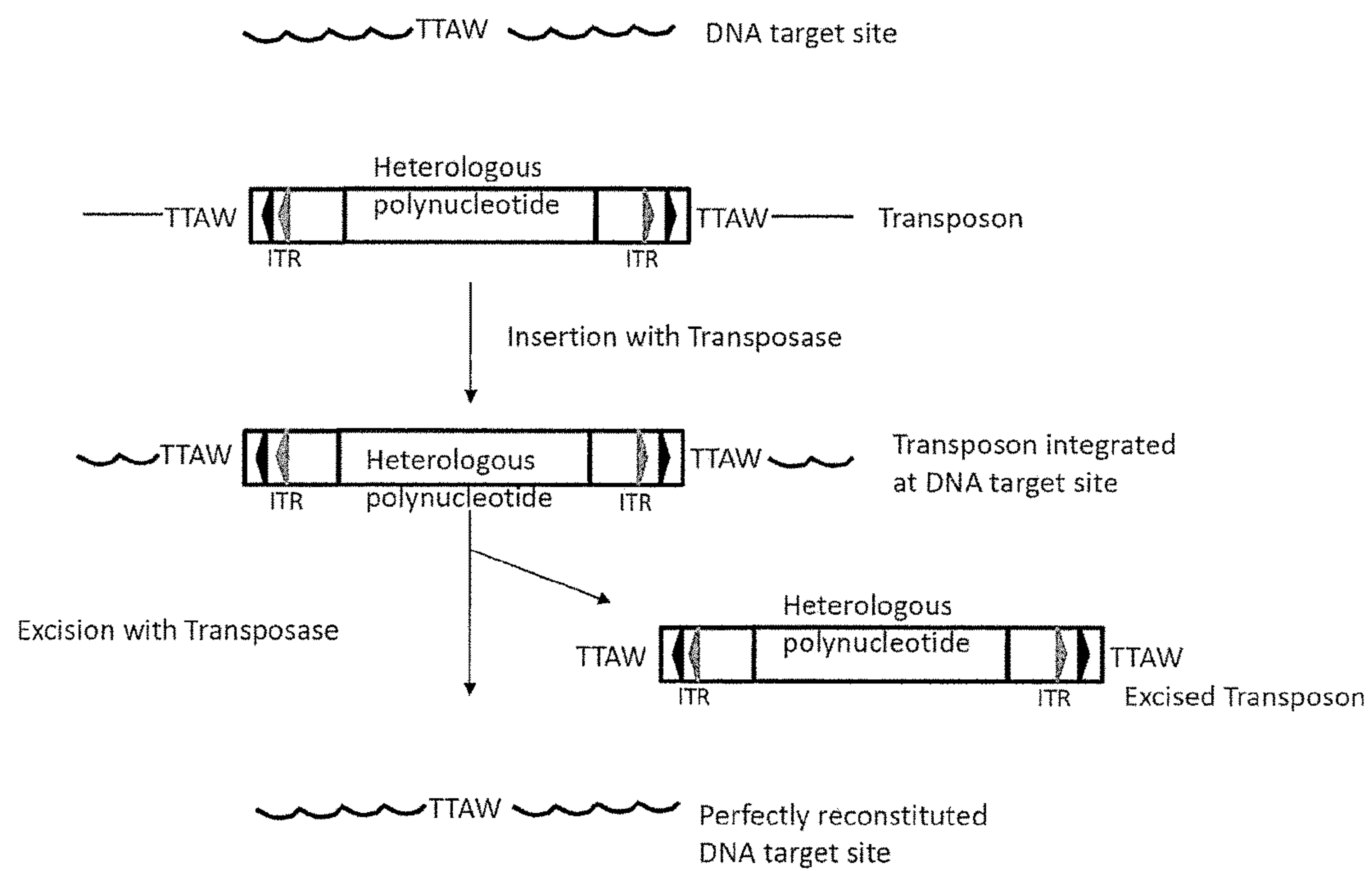


FIGURE 1

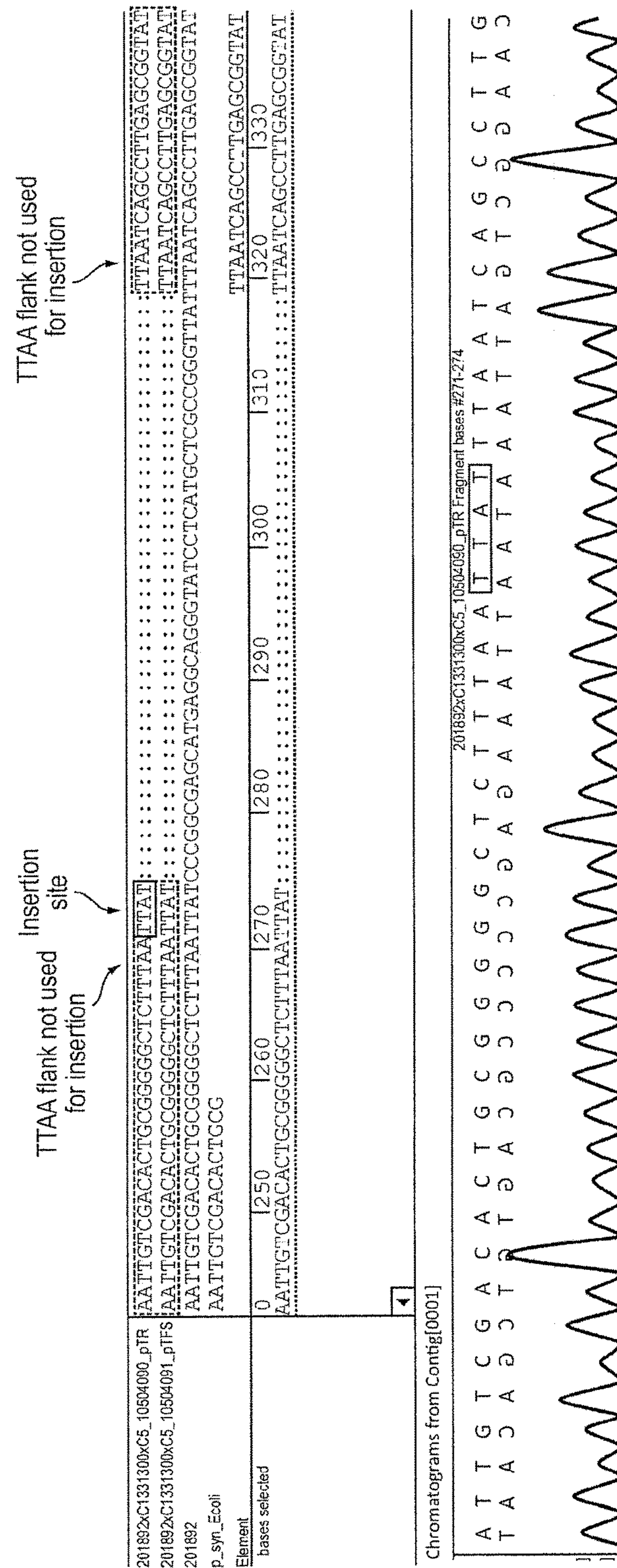


FIGURE 2

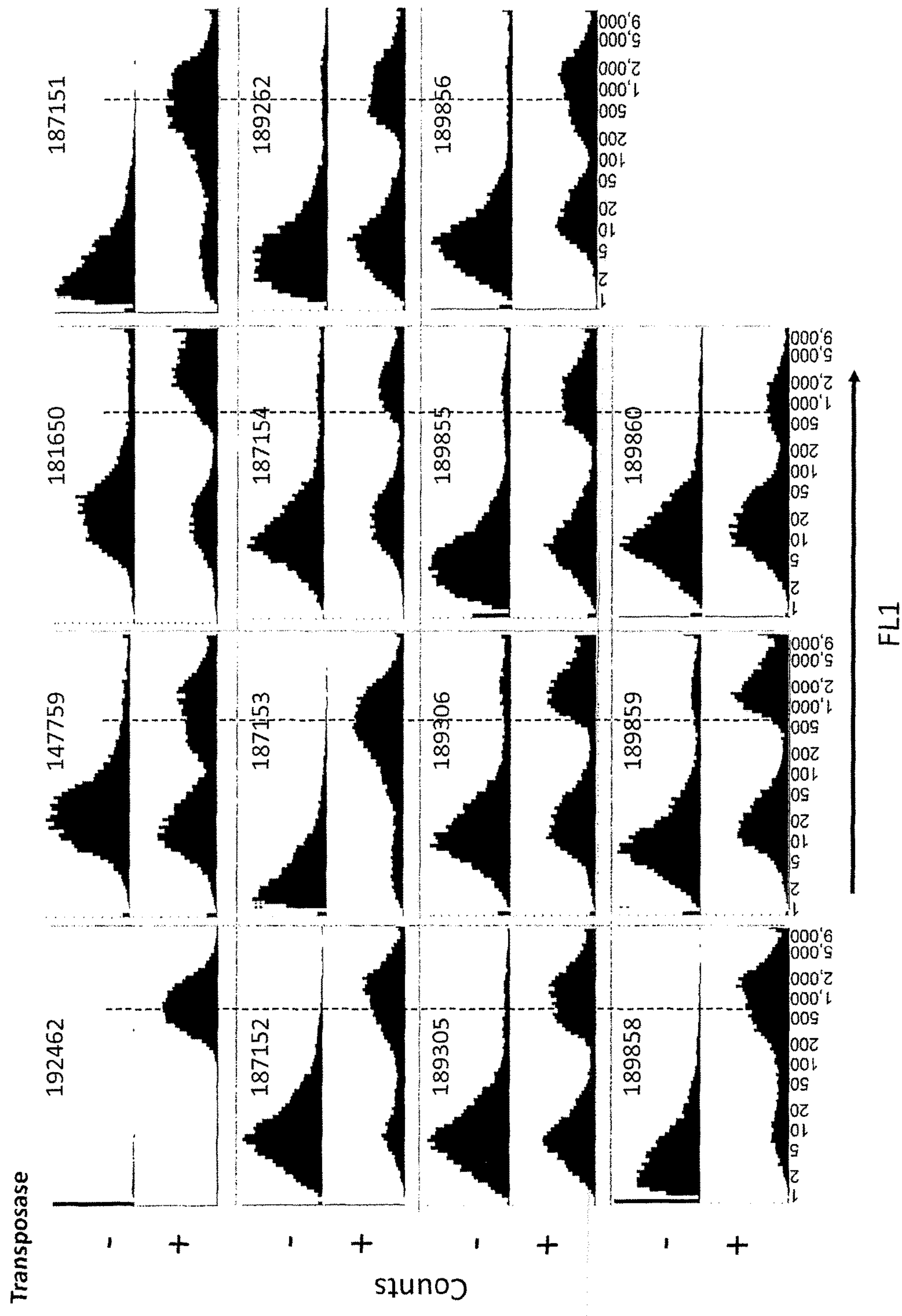


FIGURE 3

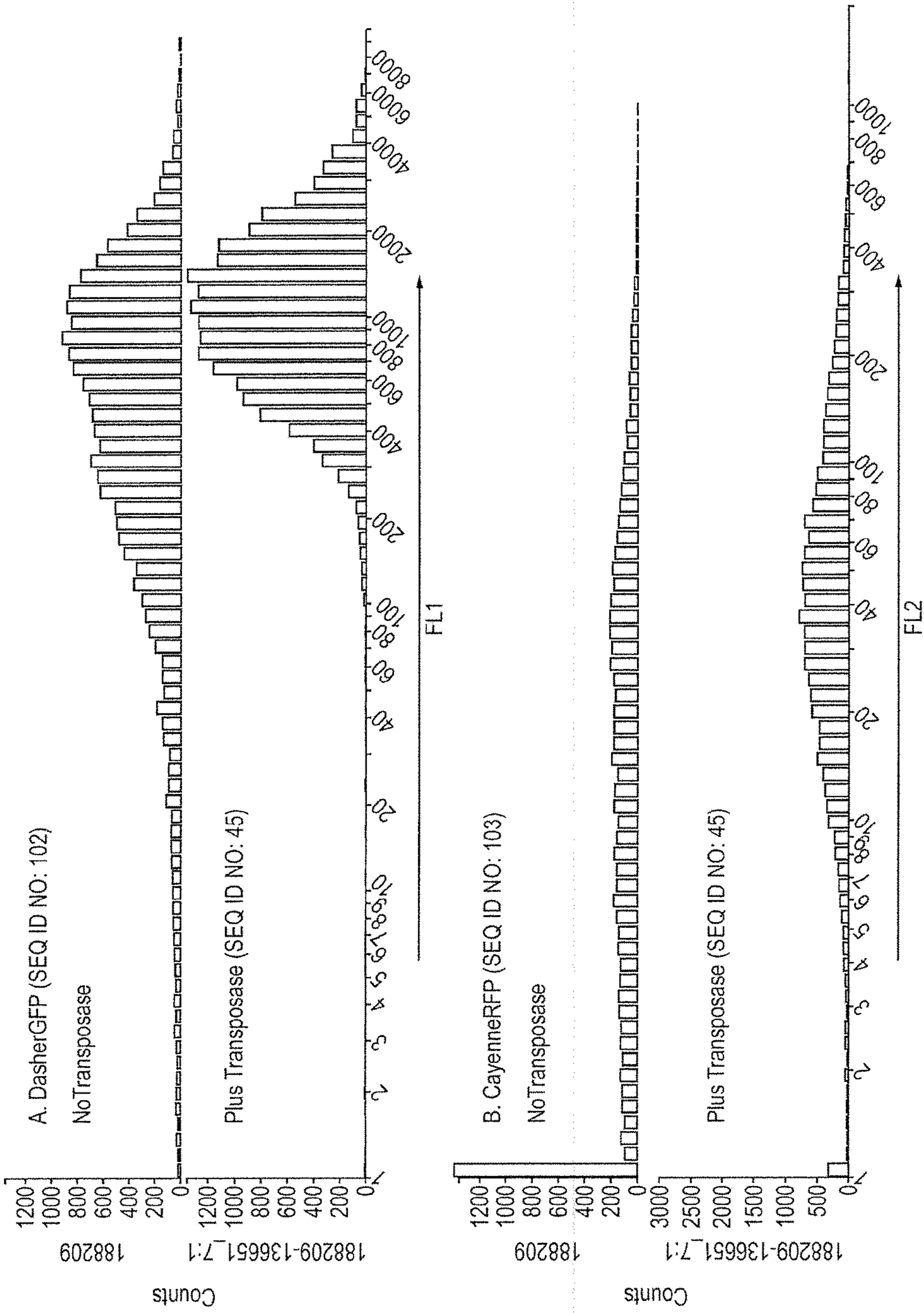
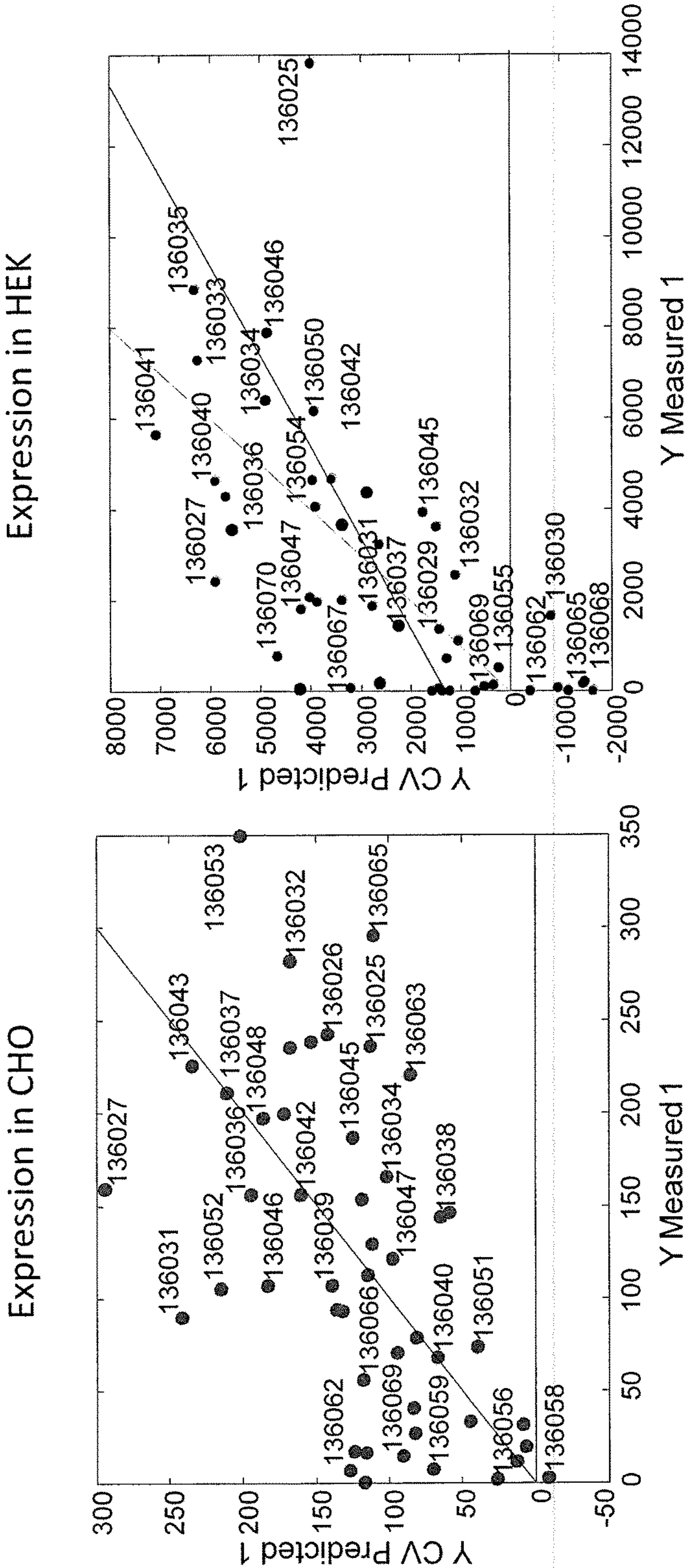


FIGURE 4



ENHANCED NUCLEIC ACID CONSTRUCTS FOR EUKARYOTIC GENE EXPRESSION

CROSS REFERENCE TO RELATED APPLICATIONS

The present application is a continuation of U.S. application Ser. No. 14/683,121 filed Apr. 9, 2015, which claims the benefit of U.S. Provisional Application No. 61/977,474 filed Apr. 9, 2014; 62/003,397 filed May 27, 2014; 62/046,875 filed Sep. 5, 2014; 62/046,705 filed Sep. 5, 2014; 62/069,656 filed Oct. 28, 2014; and 62/120,522 filed Feb. 25, 2015, each incorporated by reference in its entirety for all purposes.

REFERENCE TO A SEQUENCE LISTING

This application includes sequence listing in a txt file named "482321_SEQLIST.txt", created on Jul. 27, 2016 and containing 153,266 bytes, which is hereby incorporated by reference in its entirety for all purposes.

1. FIELD OF THE INVENTION

The field of the present invention relates to configurations of DNA vectors for heterologous gene expression, methods for identifying preferred configurations including those that are able to achieve stable modifications of the genomes of target cells, and the use of transposons and transposases.

2. BACKGROUND OF THE INVENTION

DNA constructs are typically propagated as plasmids. Plasmids are frequently constructed by cloning a first polynucleotide sequence into a vector. The vector generally comprises sequences required for propagation in at least one host cell, but it often also comprises sequences that contribute to the functioning of the first polynucleotide sequence. For example a vector may comprise elements that affect the expression of a polypeptide encoded by the first polynucleotide sequence such as promoters, enhancers, introns, terminators, translational initiation signals, polyadenylation signals, replication elements, RNA processing and export elements, and elements that affect chromatin structure that become operably linked to the first polynucleotide. The process of optimizing a polynucleotide for a specific function often comprises creating a plurality of polynucleotides, cloning them into the same vector to create a first plurality of cloned polynucleotides and measuring a property of some of the cloned polynucleotides.

Because the process of cloning polynucleotides into a single vector is relatively simple, while the process of constructing a vector is more complex and costly, optimization almost always focuses on creating variation in the cloned polynucleotide and very rarely on variations in the vector. Even if the vector sequence is varied, this will typically be done by selecting from a small number of pre-existing vectors rather than by deliberately constructing a new set of vectors. However vectors frequently contain many or even most of the elements that determine the function of the cloned polynucleotide, for example the expression of the polynucleotide in an expression-host. The functional performance of many of these elements may depend on the precise host cell being used, for example some elements that perform well in human cells may perform poorly in rodent cells, the same vector is often used in both.

Furthermore, many available vectors have been constructed by standard restriction site cloning methods and derived from other vectors wherein the functional elements have not been well defined. Consequently many vectors contain "fossil" sequences that are unnecessary for their function but have just been included because of imprecise cloning methods or a lack of understanding of function (for example the fl phage origin of replication, originally incorporated for generation of phagemids which can be found in many vectors that are never used to make phagemids), or they contain sequences that actually compromise function (for example the use of the beta lactamase gene as a selectable marker which exacerbates instability in vectors such as lentiviruses).

Because of the immense size of sequence space, there is no effective way to test all possible permutations of a polymeric biological molecule such as a nucleic acid or protein for a desired property. To test each possible nucleotide base at each position in a vector, rapidly leads to such a large number of molecules to be tested such that no available methods of synthesis or testing are feasible, even for a polymer of modest length. Furthermore, most molecules generated in such a way would lack any measurable level of the desired property. Total sequence space is very large and the functional solutions in this space are sparsely distributed.

There is thus a need in the art for methods to efficiently identify vector components that contribute to performance, and to assess this performance.

Typical methods for introducing DNA into a cell include DNA condensing reagents such as calcium phosphate, polyethylene glycol, lipid-containing reagents such as liposomes, multi-lamellar vesicles as well as virus-mediated strategies. However, such methods can have certain limitations. For example, there are size constraints associated with DNA condensing reagents and virus-mediated strategies. Further, the amount of nucleic acid that can be transfected into a cell is limited in viral strategies. In addition, not all methods facilitate insertion of the delivered nucleic acid into cellular nucleic acid, and while DNA condensing methods and lipid-containing reagents are relatively easy to prepare, the insertion of nucleic acid into viral vectors can be labor intensive. Virus-mediated strategies can be cell-type specific or tissue-type specific, and the use of virus-mediated strategies can create immunologic problems when used in vivo.

Integration of heterologous DNA into a target genome, and the expression levels of genes encoded by the integrated heterologous DNA can be increased by the configuration of DNA elements. The efficiency of integration, the size of the heterologous DNA sequence that can be integrated, and the number of copies of the heterologous DNA sequence that are integrated into each genome can often be further improved by using transposons. Transposons or transposable elements include a short nucleic acid sequence with terminal repeat sequences upstream and downstream. Active transposons can encode enzymes that facilitate the excision and insertion of the nucleic acid into target DNA sequences. A number of transposable elements have been described in the art that facilitate insertion of nucleic acids into the genome of vertebrates. For example, transposable elements discovered from various sources, for example, an engineered transposon from the genome of salmonid fish called sleeping beauty; piggyBac transposon from lepidopteran cells; piggyBac transposon from the bat *Myotis lucifugus*; mariner transposon first discovered in *Drosophila* and; an engineered transposon and transposon inverted repeats from the frog species, *Rana pipiens* called frog prince.

Different transposable elements show different preferences for the genomic sites at which they integrate. For example the piggyBac and piggyBat transposons have a preference for transcriptionally inactive regions. Although this may be an advantage for the “wild” transposon which does not wish to disrupt gene expression in its host and risk killing it, it is a disadvantage for transposons that are being used to maximize gene expression. Thus although a number of transposable elements capable of facilitating insertion of nucleic acids into the eukaryotic genome have been identified in the art, there exists a need for alternative transposable elements and enhanced constructs that facilitate higher expression levels from inserted DNA, either because of higher insertion efficiency or because the genomic insertions are made at more favorable positions within the genome, compared with transposable elements currently described in the art.

3. SUMMARY OF THE INVENTION

We describe novel ways to assess the performance of individual vector elements by analyzing the function of small numbers of vectors. The results of this analysis can then be used to create high performing combinations of the sequence elements. Such maps are used to direct perturbations or modifications of the nucleic acid construct sequences to perturb or modify the activity of the nucleic acid construct in a controlled fashion.

Specific combinations of vector elements are described that contribute to vector performance in mammalian cells, in particular to yielding high levels of expression of polypeptides in either transiently or stably transfected cells. Vector elements include promoters, enhancers, introns, terminators, translational initiation signals, polyadenylation signals, virally derived replication elements, RNA processing and export elements, transposons, transposases and elements that affect chromatin structure.

In some embodiments, heterologous gene expression can be improved when the construct further comprises sequence elements that enhance expression by effects on chromatin structure, or by affecting RNA processing or RNA export including scaffold and matrix attachment regions, introns and post transcriptional responsive elements such as WPRE, HPRE and AGS. In some embodiments, heterologous expression is improved when the construct further comprises sequences that reduce the spread of heterochromatin or the interference between one expression control region and another such as HS4 insulators or their core sequence.

Heterologous gene expression from constructs that stably integrate into the target cell genome can be further improved by incorporating transposon ends: sequence elements that are recognized and transposed by transposases. DNA sequences inserted between a pair of Transposon ends can be excised by a transposase from one DNA molecule and (unless the transposase is integration-deficient) inserted into a second DNA molecule. Two novel transposon-transposase systems are disclosed, one derived from the silkworm *Bombyx mori* and the other from the frog *Xenopus tropicalis*. Each of these comprises sequences that function as transposon ends and that can be used in conjunction with a transposase that recognizes and acts upon those transposon ends, as gene transfer systems for stably introducing nucleic acids into the DNA of a cell. The gene transfer systems of the present invention can be used in methods, for example, but not limited to, heterologous gene expression, gene therapy, insertional mutagenesis, or gene discovery.

In one aspect the invention features a transposon comprising a segment of heterologous DNA flanked by a pair of transposon end sequences or variants, derivatives and fragments of the transposon end sequences such that the transposon retains transposon activity. In one embodiment, the sequence of the transposon ends is derived from the species *Bombyx mori*. In one embodiment, the sequence of the transposon ends is derived from the species *Xenopus tropicalis*.

In some embodiments the invention further comprises a transposase that recognizes the transposon and effects the integration of the heterologous DNA between the transposon ends into the genomic DNA of a target cell. In one embodiment, the transposase has a higher activity for transposon excision compared to activity for transposon integration. In some preferred embodiments, the transposase further comprises a heterologous nuclear localization signal (NLS). In some embodiments, the transposase may further comprise a DNA binding domain. In some embodiments the transposase is encoded in a polynucleotide.

In one embodiment, a polynucleotide encodes a transposase operably linked to a heterologous promoter, wherein the transposase inserts a transposon at the sequence 5'-TTAT-3' within a target polynucleotide. In one embodiment, a polynucleotide encodes a transposase operably linked to a heterologous promoter, wherein the transposase excises a transposon by recognizing the sequence 5'-TTAT-3' adjacent to the transposon inverted terminal repeats. The polynucleotide encoding the transposase is at least 85%, at least 90%, at least 95% or at least 98% identical to SEQ ID NO: 44. In one embodiment the polynucleotide encoding the transposase further encodes a heterologous nuclear localization signal (NLS) expressible fused to the transposase. In one embodiment the polynucleotide encoding the transposase further encodes a DNA binding domain (DBD) expressible as a fusion protein with the transposase. In some embodiments, the transposase is a hyperactive variant of SEQ ID NO: 44. In some embodiments, the transposase is an integration defective variant of SEQ ID NO: 44.

In one embodiment, a polynucleotide encodes a transposase fused to a heterologous NLS and operably linked to a heterologous promoter, wherein the transposase is at least 85%, at least 90%, at least 95% or at least 98% identical to SEQ ID NO: 45. In one embodiment, the polynucleotide encoding the transposase further encodes a DNA binding domain (DBD) expressible as a fusion protein with the transposase. In some embodiments, the transposase is a hyperactive variant of SEQ ID NO:45. In other embodiments, the transposase is an integration defective variant of SEQ ID NO: 45.

In some embodiments, a first polynucleotide comprises a transposon comprising inverted repeats of a piggyBac-like transposon from the species *Xenopus tropicalis* flanking a heterologous polynucleotide, the inverted repeats being flanked by copies of the target sequence 5'-TTAA-3' at each end, such that the transposon can be excised leaving a single copy of the 5'-TTAA-3' target sequence in place of the transposon in the polynucleotide. Some embodiments further comprise a second polynucleotide encoding a transposase such that the transposon, but not the transposase, is excisable from their respective polynucleotides, and integratable into a recipient DNA molecule at a 5'-TTAA-3' target sequence by action of the transposase. In some embodiments the first and second polynucleotides are part of the same molecule, in some embodiments they are different molecules, in some embodiments they are different molecules provided together as part of a kit.

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In some embodiments, a first polynucleotide comprises a transposon comprising inverted repeats of a piggyBac-like transposon flanking a heterologous polynucleotide, the inverted repeats being flanked by copies of the target sequence 5'-TTAT-3' at each end, such that the transposon can be excised leaving a single copy of the 5'-TTAT-3' target sequence in place of the transposon in the polynucleotide. Some embodiments further comprise a second polynucleotide encoding a transposase such that the transposon, but not the transposase, is excisable from their respective polynucleotides, and integratable into a recipient DNA molecule at a 5'-TTAT-3' target sequence by action of the transposase. In some embodiments the first and second polynucleotides are part of the same molecule, in some embodiments they are different molecules, in some embodiments they are different molecules provided together as part of a kit. In some embodiments the piggyBac-like transposon is derived from the species *Bombyx mori*.

In some embodiments a transposon or transposase is modified to increase its integration activity or its excision activity, or to modify its target sequence specificity. This modification may be effected by transfecting into a cell (a) a first polynucleotide comprising a gene encoding a marker whose expression is interrupted by a transposon, and (b) a second polynucleotide encoding a transposase expressible from the polynucleotide, wherein if the transposase has activity for the transposon it transposes the transposon out of the first polynucleotide thereby causing expression of the marker to generate a signal indicating the transposase is active on the transposon. The transposon may have ends comprising SEQ ID NOS. 1 and 2 or a variant of either or both of these having at least 90% sequence identity to SEQ ID NO: 1 or 2, and the transposase has the sequence of SEQ ID NO:44 or a variant showing at least 90% sequence identity thereto. The transposon may have ends comprising SEQ ID NOS. 5 and 6 or a variant of either or both of these having at least 90% sequence identity to SEQ ID NO: 5 or 6, and the transposase has the sequence of SEQ ID NO:45 or a variant showing at least 90% sequence identity thereto.

In one embodiment, a transposon comprises a heterologous polynucleotide inserted between a pair of inverted repeats, where the transposon is capable of transposition by a transposase that is at least 85%, at least 90%, at least 95%, at least 98% identical to SEQ ID NOS: 44. In some preferred embodiments, the transposon is capable of insertion by the transposase at the sequence 5'-TTAT-3' within a target nucleic acid.

In some embodiments, the transposon end comprises at least 16 contiguous nucleotides from SEQ ID NO: 1 and the other transposon end comprises at least 16 contiguous nucleotides from SEQ ID NO: 2. In some embodiments, the transposon end comprises at least 17, at least 18, at least 19, at least 20, at least 22, at least 25, at least 30 contiguous nucleotides from SEQ ID NO: 1 and the other transposon end comprises at least 17, at least 18, at least 19, at least 20, at least 22, at least 25, at least 30 contiguous nucleotides from SEQ ID NO: 2. In some embodiments, each inverted terminal repeat (ITR) is at least 90% identical to SEQ ID NO: 32, in some embodiments, each inverted terminal repeat (ITR) comprises SEQ ID NO: 32. In some embodiments, one transposon end is at least 85%, at least 90%, at least 95%, at least 98% identical to SEQ ID NO: 1 and the other transposon end is at least 85%, at least 90%, at least 95%, at least 98% identical to SEQ ID NO: 2.

In one embodiment, a polynucleotide comprises a transposon comprising inverted terminal repeats of a piggyBac-like transposon flanking a heterologous nucleic acid, the

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inverted repeats being flanked by copies of a target sequence, where the transposon is capable of transposition by a transposase identical to SEQ ID NO: 45 fused to a heterologous nuclear localization signal (NLS). In some embodiments, the transposon end comprises at least 14 contiguous nucleotides from SEQ ID NO: 5 or 7 or 9 and the other transposon end comprises at least 14 contiguous nucleotides from SEQ ID NO: 6 or 8.

In some embodiments, the transposon end comprises at least 15, at least 16, at least 17, at least 18, at least 19, at least 20, at least 22, at least 25, at least 30 contiguous nucleotides from SEQ ID NOS: 5 or 7 or 9 and the other transposon end comprises at least 15, at least 16, at least 17, at least 18, at least 19, at least 20, at least 22, at least 25, at least 30 contiguous nucleotides from SEQ ID NOS: 6 or 8. In one embodiment, each transposon inverted terminal repeat (ITR) comprises SEQ ID NO: 42. In one embodiment, one inverted terminal repeat (ITR) comprises SEQ ID NO: 38 and one ITR comprises SEQ ID NO: 41. In one embodiment, one transposon end is at least 90% identical to SEQ ID NO: 5 and the other transposon end is at least 90% identical to SEQ ID NO: 6.

In some embodiments, the heterologous polynucleotide comprises a promoter. In some embodiments the promoter is an EF1a promoter, a CMV promoter, a GAPDH promoter, a Herpes Simplex Virus thymidine kinase (HSV-TK) promoter, an actin promoter, a PGK promoter or an ubiquitin promoter. In some embodiments the heterologous polynucleotide is in a gene transfer vector; in some embodiments, the heterologous polynucleotide is part of the transposon. In some embodiments, the gene transfer vector further comprises a transposon. In some embodiments, the heterologous polynucleotide further comprises a second promoter. The direction of transcription from the first and second promoters may be the same or different. In some embodiments, the promoter is operably linked to at least one or more of: i) an open reading frame; ii) a selectable marker; iii) a counter-selectable marker, iii) a nucleic acid encoding a regulatory protein; iv) a nucleic acid encoding an inhibitory RNA. In some preferred embodiments, the promoter is active in a eukaryotic cell.

In other embodiments, the heterologous polynucleotide comprises one or more sequence elements that increase expression by enhancing RNA processing or export from the nucleus. The RNA processing or export elements are selected from but not limited to WPRE, HPRE (SEQ ID NO: 104-105), SAR (SEQ ID NOS: 108-111), AGS (SEQ ID NOS: 106-107). In other embodiments, the heterologous polynucleotide comprises a pair of insulators. The insulators are selected from but not limited to SEQ ID NOS: 112-113. In some embodiments, the nucleic acid comprising the gene transfer vector further comprises one or more viral replication sequences, In some embodiments, the nucleic acid comprising the transposon further comprises one or more viral replication sequences, such that the replication sequences are not capable of transposition by the transposase. The viral replication sequences may include the SV40ori, SV40 large T antigen, EBVoriP and EBNA.

In some embodiments, the heterologous polynucleotide operably linked to a promoter comprises two open reading frames (ORFs), wherein the two ORFs are linked by coupling elements selected from IRES or CHYSEL. In some embodiments, IRES elements are selected from but not limited to SEQ ID NOS: 58-100. In some embodiments, CHYSEL elements are selected from but not limited to SEQ ID NO: 101. In some embodiments, IRES are at least 80%, at least 85%, at least 90%, at least 95%, at least 98%

identical to any of SEQ ID NOS: 58-100. In some embodiments, IRES is selected from picornavirus 5' UTR elements. In some embodiments, IRES have at least 80%, at least 85%, at least 90%, at least 95%, at least 98% identical to picornavirus 5'UTR elements. In some embodiments, the two open reading frames encode: i) an antibody heavy chain (HIC); ii) an antibody light chain (LC). In some embodiments, IRESs are used to control ratios of two, three four or more open reading frames (ORFs). In some embodiments, two or more IRESs control expression levels of three ORFs. The IRESs selected may be the same or different. A kit comprising an expression vector with one or more IRESs selected from SEQ ID NOS: 58-100. A kit comprising a nucleic acid panel of IRES sequences selected from SEQ ID NOS: 58-100.

A method for modifying the genomic DNA of a cell comprising: a) Introducing into a cell of a target organism: i) a transposase at least 90% identical to SEQ ID NO: 44; and ii) a transposon comprising transposon ends flanking a nucleic acid heterologous to the transposon ends, wherein the transposase inserts the transposon at a sequence 5'-TTAT-3' in the genome of the cell; b) Isolating the cell with the inserted transposon comprising the heterologous nucleic acid. A method for modifying the genomic DNA of a cell comprising: a) Introducing into a cell of a target organism: i) a transposase at least 90% identical to SEQ ID NO: 45; and ii) a transposon comprising transposon ends flanking a nucleic acid heterologous to the transposon ends; b) Isolating the cell with the inserted transposon comprising the heterologous nucleic acid. In some embodiments, the transposase is at least 85%, at least 95%, at least 98% identical to SEQ ID NO: 44. In some embodiments, the transposase is at least 85%, at least 95%, at least 98% identical to SEQ ID NO: 45. The method further comprises removing the heterologous polynucleotide inserted in the genome by treating the cell with a transposase. The transposase is least 85%, at least 95%, at least 98% identical to SEQ ID NO: 44. In some embodiments, the transposase is at least 85%, at least 95%, at least 98% identical to SEQ ID NO: 45. In some embodiments, the transposase is integration deficient. In some embodiments, the transposase is provided as a nucleic acid encoding the transposase, in other embodiments, the transposase is provided as a protein. In some embodiments, the host cell is obtained from a eukaryote; the cell is from a mammal; the cell is a Chinese Hamster ovary (CHO) cell or a Human embryonic kidney (HEK293) cell. A method for producing protein from a cell, the method comprising i) integrating a transposon encoding the heterologous protein and, ii) obtaining protein from the cell. The transposon comprises a heterologous polynucleotide operably linked to a promoter and comprises two open reading frames (ORFs), wherein the two ORFs are linked by coupling elements selected from IRES or CHYSEL. In some embodiments, a method of producing protein from a cell comprises i) introducing a gene transfer vector comprising a heterologous polynucleotide operably linked to a promoter and comprises two open reading frames (ORFs), wherein the two ORFs are linked by coupling elements selected from IRES or CHYSEL and, ii) obtaining protein from a cell. A method for producing an antibody from a cell, the method comprising: i) Integrating a transposon encoding the heterologous protein comprising heavy and light chains of the antibody linked by coupling elements selected from IRES or CHYSEL and, ii) obtaining antibody from the cell. A method for producing an antibody from a cell, the method comprising: i) Introducing a gene transfer vector encoding the heterologous protein comprising heavy and light chains of

the antibody linked by coupling elements selected from IRES or CHYSEL and, ii) obtaining antibody from the cell.

Other embodiments are a cell line comprising the transposon as described above; a cell line produced by the method of modifying the genomic DNA of a cell as described above; a cell line created by removing the heterologous DNA inserted in the genome comprising treating the cell with a transposase. Other embodiments include a protein made by any of the methods described above. In some embodiments, the protein is an antibody, an antibody fragment or a derivative thereof. A transgenic animal comprising the transposon and; a pharmaceutical composition comprising the transposon and transposase together with a pharmaceutically acceptable carrier, adjuvant or vehicle.

In one embodiment, the transposon nucleic acid sequence comprises a sequence selected from SEQ ID NOS: 1-29. In one embodiment, the transposase nucleic acid sequence encodes an amino acid sequence selected from SEQ ID NOS: 43-56.

In one embodiment, the transposon is capable of inserting into the DNA of a cell.

In other embodiments, the transposon of the above embodiments further comprises a nucleic acid encoding a selectable marker, for example a gene encoding one of glutamine synthase, dihydrofolate reductase, a protein conferring resistance to puromycin, neomycin, hygromycin, zeocin or blasticidin.

In some embodiments, the transposon is inserted in a plasmid. In one embodiment, the transposon further comprises an open reading frame. It is expressly contemplated that the transposon may comprise combinations of any of the sequence elements described above, including promoters, enhancers, introns, terminators, translational initiation signals, polyadenylation signals, RNA processing and export elements and elements that affect chromatin structure. It is further contemplated that plasmids into which transposons are inserted may also comprise combinations of any transposases or virally derived replication elements.

In one embodiment, the invention features a gene transfer system comprising a transposon according to any of the above embodiments; and a *Bombyx mori* transposase. In a further embodiment, the transposase comprises an amino acid sequence corresponding to SEQ ID NOS: 43-44. In one embodiment, the invention features a gene transfer system comprising a transposon according to any of the above embodiments; and a *Xenopus tropicalis* transposase. In a further embodiment, the transposase comprises an amino acid sequence corresponding to SEQ ID NOS: 45-46. In some embodiments, the transposon and transposase are on separate plasmids; in some embodiments the transposon and transposase are on the same plasmid.

In some preferred embodiments, the gene transfer system comprising a transposon and a transposase further comprises IRES sequences (for example those described in SEQ ID NOS: 58-100 such that the relative expression of two open reading frames (ORFs) expressed from a single promoter can be specified based on IRES strength. In a further embodiment, the ORFs encode heavy and light chains of an antibody. In some embodiments, IRES sequences are used as enhancer elements. In some embodiments the IRES works well in combination with a secretion signal. This is an important aspect for secretion of the expressed polypeptides and is of particular importance for secreted expression of heavy and light chains of an antibody in stably transfected Chinese hamster ovary (CHO) and Human embryonic kidney (HEK293) cells.

In some embodiments, the transposon is inserted into the genome of a cell. In some embodiments, the cell is chosen from Chinese hamster ovary (CHO) or Human Embryonic kidney (HEK293) cell lines. In another embodiment, the cell is obtained from an animal. In another embodiment, the cell is from a vertebrate or invertebrate. In a further embodiment, the vertebrate is a mammal. In other embodiments, the present invention also features a cell comprising a transposon of any of the above-described embodiments.

In other embodiments, the present invention features a pharmaceutical composition comprising a *Bombyx mori* transposase and a transposon recognized and transposed by the transposase, together with a pharmaceutically acceptable carrier, adjuvant or vehicle. In other embodiments, the present invention features a pharmaceutical composition comprising a *Xenopus tropicalis* transposase and a transposon recognized and transposed by the transposase, together with a pharmaceutically acceptable carrier, adjuvant or vehicle.

The present invention also features a method for introducing exogenous DNA into a cell comprising contacting a cell with the gene transfer system of the above-described embodiments, thereby introducing exogenous DNA into a cell. In some embodiments, the cell is a eukaryotic cell. In some other embodiments, the cell is from a mammal. In some embodiments, the cell is a stem cell. In other embodiments, the cell is a Chinese hamster ovary (CHO) cell or Human embryonic kidney (HEK293) cell.

The present invention also includes a method for producing protein using the method for introducing exogenous DNA into a cell as described herein above. In some preferred embodiments, the protein is an antibody, an antibody fragment, or a derivative thereof. Other embodiments of the present invention include a cell line comprising a *Bombyx mori* transposon or a *Xenopus tropicalis* transposon; a transgenic animal comprising a *Bombyx mori* transposon or a *Xenopus tropicalis* transposon; or a cell line produced by the method described herein above.

In another embodiment, the present invention features a kit comprising a *Bombyx mori* transposon and a *Bombyx mori* transposase or a *Xenopus tropicalis* transposon and a *Xenopus tropicalis* transposase; and; instructions for introducing DNA into a cell. In another embodiment, the present invention also features a kit comprising: a *Bombyx mori* transposase or a *Xenopus tropicalis* transposase that is integration defective, and instructions for use.

The present invention also includes methods for producing two or more polypeptides within the same cell. In some embodiments this is achieved using translational coupling elements such as IRES elements. One embodiment includes a method for expressing a plurality of polypeptides from a single construct comprising a) a eukaryotic promoter and a plurality of polynucleotides encoding a plurality of polypeptides b) an IRES sequence linking each of the plurality of polynucleotides wherein the plurality of polynucleotide-IRES-polynucleotide are operably linked to a single eukaryotic promoter such that on insertion into a host cell, a plurality of polypeptides are expressed and expression level of each of the polypeptides is determined by the IRES sequence. The method wherein 2, 3, 4, 5, 6 or more IRES sequences link the plurality of polynucleotides is another embodiment. IRES sequences selected may be the same or different sequences.

Some embodiments comprise a polynucleotide comprising: a eukaryotic promoter operably linked to a first polynucleotide encoding a first polypeptide, an IRES sequence and a second polynucleotide encoding a second polypeptide

wherein i) the first polynucleotide, IRES and second polynucleotide are operably linked to a single eukaryotic promoter such that on insertion into a host cell, both polypeptides are expressed, ii) the two polypeptide interact in the formation of a product such that the ratio of expression of the two polypeptides determines the amount of product formed, iii) the relative expression level of the polynucleotides is determined by the IRES sequence, iv) expression of the two polypeptides are operably linked to a regulatory sequence selected from SEQ ID NOS: 104-111. IRES elements selected include hybrid elements selected from SEQ ID NOS: 73-91, 95-97. In one embodiment, the polynucleotide comprises sequences encoding heavy chain and light chain of an antibody. The polynucleotide further comprises regulatory elements, wherein the expression of the two polypeptides is operably linked to a regulatory sequence selected from HPRE (SEQ ID NOS: 104-105), AGS (SEQ ID NOS: 106-107), SAR (108-111).

In some embodiments, a polynucleotide comprises an IRES element selected from SEQ ID NOS: 74-77, 81-91, 93-98, the IRES element is flanked by sequences encoding a heavy chain and light chain of an antibody operably linked to a single eukaryotic promoter. In other embodiments, the IRES element is flanked by sequences encoding a heavy chain or a light chain of an antibody and a reporter protein operably linked to a single eukaryotic promoter. The polynucleotide may comprise 2, 3, 4, 5, 6 or more IRES sequences. The IRES sequences are the same or different sequences. In some embodiments, a polynucleotide comprises an IRES element and regulatory sequences selected from HPRE (SEQ ID NOS: 104-105), AGS (SEQ ID NOS: 106-107), SAR (108-111). The IRES sequence is selected from SEQ ID NOS: 58-100. In some embodiments, the polynucleotide further comprising sequences encoding heavy chain and light chain of an antibody linked by an IRES sequence operably linked to a single eukaryotic promoter. In some embodiments, the polynucleotide may comprise 2, 3, 4, 5, 6 or more IRES sequences. Some embodiments comprise a polynucleotide comprising a first polynucleotide encoding a first polypeptide, an IRES sequence and a second polynucleotide encoding a second polypeptide operably linked to a single eukaryotic promoter, wherein the IRES sequence is selected from one of SEQ ID NOS: 58-100. In some embodiments, the IRES sequence is selected from SEQ ID NOS: 73-91, 95-97.

Some embodiments comprise a polynucleotide comprising sequences encoding heavy and light chains of an antibody linked by an IRES element operably linked to transcription control sequences. In some embodiments the transcription control sequences are flanked by insulators. The transcriptional control sequences are one or more sequences selected from promoters, enhancers, introns, 5'UTRs. In some embodiments introns are selected from SEQ ID NOS: 119, 123) and enhancers are selected from SEQ ID NOS: 116-119. In some embodiments, the polynucleotide further comprises secretion peptides at the amino termini of the polypeptides. In some embodiments, the IRES sequence is operably linked to a secretion peptide (SEQ ID NOS: 114-115). In some embodiments, the IRES sequence is one of SEQ ID NOS: 58-59. In some embodiments, the EF1a promoter is flanked by insulators (SEQ ID NOS: 112-113). In some embodiments, the polynucleotide further comprises sequences that promote integration into a host cell. In some embodiments, the polynucleotide further comprises RNA export elements. In some embodiments, RNA export elements are selected from WPRE. HPRE (SEQ ID NO: 104-105), SAR (SEQ ID NOS: 108-111). AGS (SEQ ID

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NOS: 106-107). In one embodiment, the polypeptide comprising transcriptional control elements comprise a CMV enhancer, an EF1a promoter, a hybrid intron (SEQ ID NO: 119), further comprising insulators (SEQ ID NOS: 112-113). In one embodiment, the polypeptide comprising transcriptional control elements comprise a CMV enhancer, an actin promoter, a hybrid actin intron (SEQ ID NO: 123) sequences and SV40 enhancer (SEQ ID NO: 117) sequence. In one embodiment, the polypeptide comprising transcriptional control elements comprise a CMV enhancer, a GCAPDH promoter, a CMVc intron sequences and SV40 enhancer sequence. In one embodiment, the polypeptide comprising transcriptional control elements comprise a CMV enhancer, a CMV promoter, a SV40 enhancer sequences. In some embodiments, the polynucleotide further comprises comprising viral replication sequences selected from the SV40ori, SV40 large T antigen, EBVoriP and EBNA. Other embodiments comprise a host cell with the polynucleotide of the above embodiments, wherein the host cell is a eukaryotic cell, is from a mammal. A protein made by the methods described herein is another embodiment. In some embodiments, the protein is an antibody, an antibody fragment or a derivative thereof. Some embodiments comprise a pharmaceutical composition comprising the polynucleotides of the above embodiments.

Additional embodiments comprise a polynucleotide comprising sequences encoding heavy chain and light chain of an antibody, each sequence operably linked to transcriptional control sequences. In some embodiments, the two transcriptional control sequences comprise enhancers, promoters and introns. In some embodiments, the transcriptional control sequences are combinations of a CMV enhancer, an actin promoter, hybrid intron (SEQ ID NO: 123) and EF1a promoter and intron EF1a. In some embodiments, the transcriptional control sequences are combinations of a CMV enhancer, an actin promoter, hybrid intron (SEQ ID NO: 123) and CMV promoter with intron CMVc. In some embodiments, the polynucleotide further comprises polyadenylation signals at the 3'-end of the sequence encoding the polypeptide. In some embodiments, each promoter and polyadenylation signal is flanked by insulators. In some embodiments, the polynucleotide further comprises viral replication sequences selected from SV40 ori, SV40 large T antigen, EBV oriP, EBNA.

Other embodiments will be evident to those of ordinary skill in the art from the teachings contained herein in combination with what is known to the art.

4. BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1: shows a transposon comprising a heterologous polynucleotide between flanking transposon ends which each comprise inverted terminal repeats (ITRs) (shown by black and grey arrowheads). The transposon ITRs are adjacent to a direct repeat of its target sequence. For the *Trichoplusia ni* piggyBac and the *Xenopus* transposons of the invention, this target sequence is 5'-TTAA-3'. For the *Bombyx* transposons of the invention this target sequence is 5'-TTAT-3'. The target sequence here is therefore represented as 5'-TTAW-3', where the W is A or T. When the transposon is transposed by the action of a transposase, it is excised from one DNA molecule where it leaves a single copy of the target sequence, and integrated into a second DNA molecule where it duplicates the target sequence such that the transposon remains flanked by the target sequence. The transposase may be provided either in cis (encoded in the same vector) or trans (encoded in a separate polynucle-

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otide or as protein). When the transposase excises the transposon, the original sequence 5'-TTAW-3' is perfectly restored.

FIG. 2: shows sequencing results with the 5'-TTAT-3' target sequence left after integration and excision (shown by arrow) by a transposase from *Bombyx mori*. Chinese hamster ovary (CHO) cells were co-transfected with a transposon comprising transposon ends (SEQ ID NOS: 1 and 2) and a construct encoding a transposase (SEQ ID NO: 44) from *Bombyx mori*; cells were grown for 14 days post puromycin selection as described in Example 6.1.1. DNA was miniprepred from cell lysates and PCRd under standard cycling conditions with a 5 second extension time using nested amplification primers flanking the inverted terminal repeats (ITRs). PCR product was cloned into a cloning vector and transformed into *E. coli*. 16 clones from each of the amplified PCR products were picked and sequenced. All 16 clones showed a single 5'-TTAT-3' scar sequence showing the integration target sequence to be 5'-TTAT-3'.

FIG. 3: shows FACS data for stably transfected Chinese hamster ovary (CHO) cell populations expressing DasherGFP (SEQ ID NO: 102). CHO cells were transfected with gene transfer vectors comprising *Xenopus tropicalis* transposon ends (SEQ ID NOS: 5, 6) flanking a heterologous nucleic acid encoding DasherGFP. The gene transfer vectors comprised different combinations of control elements including promoters and insulator sequences. Co-transfections with an expression vector encoding transposase (SEQ ID NO: 45) were done in parallel. Vectors with *Trichoplusia ni* piggyBac transposon ends (SEQ ID NOS: 30, 31) and hyperactive *Trichoplusia ni* piggyBac transposase (SEQ ID NO: 57) were tested under the same conditions. Cells were grown as described in Example 6.2. DasherGFP expressing cell populations are shown for CHO cells transfected in the absence of transposase (top panel) and after co-transfection with transposase (bottom panel).

FIG. 4: shows FACS data for stably transfected Chinese hamster ovary (CHO) cell populations expressing DasherGFP (SEQ ID NO: 102) and CayenneRFP (SEQ ID NO: 103) linked by an IRES element (SEQ ID NO: 59) and operably linked to a single EF1a promoter. CHO cells were transfected with a gene transfer vector with configuration as shown in Table 13 lines 3 and 4. Co-transfections with an expression vector encoding a transposase (SEQ ID NO: 45) were done in parallel. Cells were grown as described in Example 6.2. DasherGFP expressing cell populations are shown (Panel A) for CHO cells transfected in the absence of transposase (top panel) and with co-transfection with transposase (bottom panel). CayenneRFP expressing cell populations are shown (Panel B) for CHO cells transfected in the absence of transposase (top panel) and after co-transfection with transposase (bottom panel).

FIG. 5: shows two graphs in which the measured values of expression of DasherGFP are shown on the X-axis, where DasherGFP is expressed from vectors with configurations shown in Table 15, expression data is from Table 19. The Y-axis of each graph shows the value predicted for DasherGFP expression from these vectors, using a model constructed, using partial least squares regression, from the sequence data in Table 15 and the expression properties shown in Table 19.

5. DETAILED DESCRIPTION OF THE INVENTION

5.1 Definitions

Use of the singular forms "a," "an," and "the" include plural references unless the context clearly dictates other-

wise. Thus, for example, reference to “a polynucleotide” includes a plurality of polynucleotides, reference to “a substrate” includes a plurality of such substrates, reference to “a variant” includes a plurality of variants, and the like.

Terms such as “connected,” “attached,” “linked,” and “conjugated” are used interchangeably herein and encompass direct as well as indirect connection, attachment, linkage or conjugation unless the context clearly dictates otherwise. Where a range of values is recited, it is to be understood that each intervening integer value, and each fraction thereof, between the recited upper and lower limits of that range is also specifically disclosed, along with each subrange between such values. The upper and lower limits of any range can independently be included in or excluded from the range, and each range where either, neither or both limits are included is also encompassed within the invention. Where a value being discussed has inherent limits, for example where a component can be present at a concentration of from 0 to 100%, or where the pH of an aqueous solution can range from 1 to 14, those inherent limits are specifically disclosed. Where a value is explicitly recited, it is to be understood that values which are about the same quantity or amount as the recited value are also within the scope of the invention. Where a combination is disclosed, each sub combination of the elements of that combination is also specifically disclosed and is within the scope of the invention. Conversely, where different elements or groups of elements are individually disclosed, combinations thereof are also disclosed. Where any element of an invention is disclosed as having a plurality of alternatives, examples of that invention in which each alternative is excluded singly or in any combination with the other alternatives are also hereby disclosed; more than one element of an invention can have such exclusions, and all combinations of elements having such exclusions are hereby disclosed.

Unless defined otherwise herein, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. Singleton, et al., *Dictionary of Microbiology and Molecular Biology*, 2nd Ed., John Wiley and Sons, New York (1994), and Hale & Marham, *The Harper Collins Dictionary of Biology*, Harper Perennial, NY, 1991, provide one of skill with a general dictionary of many of the terms used in this invention. Although any methods and materials similar or equivalent to those described herein can be used in the practice or testing of the present invention, the preferred methods and materials are described. Unless otherwise indicated, nucleic acids are written left to right in 5' to 3' orientation; amino acid sequences are written left to right in amino to carboxy orientation, respectively. The terms defined immediately below are more fully defined by reference to the specification as a whole.

The terms “polynucleotide,” “oligonucleotide,” “nucleic acid” and “nucleic acid molecule” and “gene” are used interchangeably herein to refer to a polymeric form of nucleotides of any length, and may comprise ribonucleotides, deoxyribonucleotides, analogs thereof, or mixtures thereof. This term refers only to the primary structure of the molecule. Thus, the term includes triple-, double- and single-stranded deoxyribonucleic acid (“DNA”), as well as triple-, double- and single-stranded ribonucleic acid (“RNA”). It also includes modified, for example by alkylation, and/or by capping, and unmodified forms of the polynucleotide. More particularly, the terms “polynucleotide,” “oligonucleotide,” “nucleic acid” and “nucleic acid molecule” include polydeoxyribonucleotides (containing 2-deoxy-D-ribose), polyribonucleotides (containing D-ri-

bose), including tRNA, rRNA, hRNA, siRNA and mRNA, whether spliced or unspliced, any other type of polynucleotide which is an N- or C-glycoside of a purine or pyrimidine base, and other polymers containing nonnucleotidic backbones, for example, polyamide (for example, peptide nucleic acids (“PNAs”)) and polymorpholino (commercially available from the Anti-Virals, Inc., Corvallis, Oreg., as Neugene) polymers, and other synthetic sequence-specific nucleic acid polymers providing that the polymers contain nucleobases in a configuration which allows for base pairing and base stacking, such as is found in DNA and RNA. There is no intended distinction in length between the terms “polynucleotide,” “oligonucleotide,” “nucleic acid” and “nucleic acid molecule,” and these terms are used interchangeably herein. These terms refer only to the primary structure of the molecule. Thus, these terms include, for example, 3'-deoxy-2',5'-DNA, oligodeoxyribonucleotide N3' P5' phosphoramidates, 2'-O-alkyl-substituted RNA, double- and single-stranded DNA, as well as double- and single-stranded RNA, and hybrids thereof including for example hybrids between DNA and RNA or between PNAs and DNA or RNA, and also include known types of modifications, for example, labels, alkylation, “caps,” substitution of one or more of the nucleotides with an analog, internucleotide modifications such as, for example, those with uncharged linkages (for example, methyl phosphonates, phosphotriesters, phosphoramidates, carbamates, or the like) with negatively charged linkages (for example, phosphorothioates, phosphorodithioates, or the like), and with positively charged linkages (for example, aminoalkylphosphoramidates, aminoalkylphosphotriesters), those containing pendant moieties, such as, for example, proteins (including enzymes (for example, nucleases), toxins, antibodies, signal peptides, poly-L-lysine, or the like), those with intercalators (for example, acridine, psoralen, or the like), those containing chelates (of, for example, metals, radioactive metals, boron, oxidative metals, or the like), those containing alkylators, those with modified linkages (for example, alpha anomeric nucleic acids, or the like), as well as unmodified forms of the polynucleotide or oligonucleotide.

The terms “polynucleotide,” “oligonucleotide,” “nucleic acid” and “nucleic acid molecule” and “gene” refer to the entire sequence or gene or a fragment thereof. The fragment thereof can be a functional fragment. Where the polynucleotides are to be used to express encoded proteins, nucleotides that can perform that function or which can be modified (for example, reverse transcribed) to perform that function are used. Where the polynucleotides are to be used in a scheme that requires that a complementary strand be formed to a given polynucleotide, nucleotides are used which permit such formation.

As used herein, the terms “nucleoside” and “nucleotide” will include those moieties which contain not only the known purine and pyrimidine bases, but also other heterocyclic bases which have been modified. Such modifications include methylated purines or pyrimidines, acylated purines or pyrimidines, or other heterocycles. Modified nucleosides or nucleotides can also include modifications on the sugar moiety, for example, where one or more of the hydroxyl groups are replaced with halogen, aliphatic groups, or is functionalized as ethers, amines, or the like. The term “nucleotidic unit” is intended to encompass nucleosides and nucleotides.

Standard A-T and G-C base pairs form under conditions which allow the formation of hydrogen bonds between the N3-H and C4-oxy of thymidine and the N1 and C6-NH₂,

respectively, of adenosine and between the C2-oxy, N3 and C4-NH₂, of cytidine and the C2-NH₂, N¹-H and C6-oxy, respectively, of guanosine. Thus, for example, guanosine (2-amino-6-oxy-9-β-D-ribofuranosyl-purine) may be modified to form isoguanosine (2-oxy-6-amino-9-β-D-ribofuranosyl-purine). Such modification results in a nucleoside base which will no longer effectively form a standard base pair with cytosine. However, modification of cytosine (1-β-D-ribofuranosyl-2-oxy-4-amino-pyrimidine) to form isocytosine (1-β-D-ribofuranosyl-2-amino-4-oxy-pyrimidine-) results in a modified nucleotide which will not effectively base pair with guanosine but will form a base pair with isoguanosine (U.S. Pat. No. 5,681,702 to Collins et al., hereby incorporated by reference in its entirety). Isocytosine is available from Sigma Chemical Co. (St. Louis, Mo.); isocytidine may be prepared by the method described by Switzer et al. (1993) *Biochemistry* 32:10489-10496 and references cited therein; 2'-deoxy-5-methyl-isocytidine may be prepared by the method of Tor et al., 1993, *J. Am. Chem. Soc.* 115:4461-4467 and references cited therein; and isoguanine nucleotides may be prepared using the method described by Switzer et al., 1993, *supra*, and Mantsch et al., 1993, *Biochem.* 14:5593-5601, or by the method described in U.S. Pat. No. 5,780,610 to Collins et al., each of which is hereby incorporated by reference in its entirety. Other non-natural base pairs may be synthesized by the method described in Piccirilli et al., 1990, *Nature* 343:33-37, hereby incorporated by reference in its entirety, for the synthesis of 2,6-diaminopyrimidine and its complement (1-methylpyrazolo-[4,3]pyrimidine-5,7-(4H,6H)-dione. Other such modified nucleotidic units which form unique base pairs are known, such as those described in Leach et al. (1992) *J. Am. Chem. Soc.* 114:3675-3683 and Switzer et al., *supra*.

The phrase "DNA sequence" refers to a contiguous nucleic acid sequence. The sequence can be either single stranded or double stranded, DNA or RNA, but double stranded DNA sequences are preferable. The sequence can be an oligonucleotide of 2 to 20 nucleotides in length to a full length genomic sequence of thousands or hundreds of thousands of base pairs.

The term "vector" or "DNA vector" or "gene transfer vector" refers to a polynucleotide sequence that is used to perform a "carrying" function for another polynucleotide. For example vectors are often used to allow a polynucleotide to be propagated within a living cell, or to allow a polynucleotide to be packaged for delivery into a cell, or to allow a polynucleotide to be integrated into the genomic DNA of a cell. A vector may further comprise additional functional elements, for example it may comprise a transposon.

A "gene transfer system" comprises a vector or gene transfer vector, or a polynucleotide cloned into a vector. A gene transfer system may also comprise other features to facilitate the process of gene transfer. For example a gene transfer system may comprise a vector and a lipid or viral packaging mix for enabling a first polynucleotide to enter a cell, or it may comprise a vector that includes a transposon and a second polynucleotide sequence encoding a transposase to enhance productive genomic integration of the transposon.

The term "host" means any prokaryotic or eukaryotic organism that can be a recipient of a nucleic acid. A "host," as the term is used herein, includes prokaryotic or eukaryotic organisms that can be genetically engineered. For examples of such hosts, see Maniatis et al., *Molecular Cloning. A Laboratory Manual*, Cold Spring Harbor Laboratory, Cold

Spring Harbor, N.Y. (1982). As used herein, the terms "host," "host cell," "host system" and "expression host" be used interchangeably.

The term "expression construct" means any double-stranded DNA or double-stranded RNA designed to transcribe an RNA, for example, a construct that contains at least one promoter which is or may be operably linked to a downstream gene, coding region, or polynucleotide sequence (for example, a cDNA or genomic DNA fragment that encodes a polypeptide or protein, or an RNA effector molecule, for example, an antisense RNA, triplex-forming RNA, ribozyme, an artificially selected high affinity RNA ligand (aptamer), a double-stranded RNA, for example, an RNA molecule comprising a stem-loop or hairpin dsRNA, or a bi-finger or multi-finger dsRNA or a microRNA, or any RNA). An "expression construct" includes a double-stranded DNA or RNA comprising one or more promoters, wherein one or more of the promoters is not in fact operably linked to a polynucleotide sequence to be transcribed, but instead is designed for efficient insertion of an operably-linked polynucleotide sequence to be transcribed by the promoter. Transfection or transformation of the expression construct into a recipient cell allows the cell to express an RNA effector molecule, polypeptide, or protein encoded by the expression construct. An expression construct may be a genetically engineered plasmid, virus, recombinant virus, or an artificial chromosome derived from, for example, a bacteriophage, adenovirus, adeno-associated virus, retrovirus, lentivirus, poxvirus, or herpesvirus, or further embodiments described under "expression vector" below. An expression construct can be replicated in a living cell, or it can be made synthetically. For purposes of this application, the terms "expression construct", "expression vector", "vector", and "plasmid" are used interchangeably to demonstrate the application of the invention in a general, illustrative sense, and are not intended to limit the invention to a particular type of expression construct.

The term "expression vector" or "expression construct" means a DNA construct that contains at least one promoter which is or may be operably linked to a downstream gene, coding region, or polynucleotide sequence to be transcribed (for example, a cDNA or genomic DNA fragment that encodes a protein, optionally, operably linked to sequence lying outside a coding region, an antisense RNA coding region, or RNA sequences lying outside a coding region). An expression vector or expression construct may also be a DNA construct comprising one or more promoters, wherein one or more of the promoters is not in fact operably linked to a polynucleotide sequence to be transcribed, but instead is designed for efficient insertion of an operably-linked polynucleotide sequence to be transcribed by the promoter. Transfection or transformation of the expression vector into a recipient cell allows the cell to express RNA encoded by the expression vector. An expression vector may be a genetically engineered plasmid, virus, transposon or artificial chromosome derived from, for example, a bacteriophage, adenovirus, adeno-associated virus, retrovirus, poxvirus, or herpesvirus. Such expression vectors can include sequences from bacteria, viruses or phages. Such vectors include chromosomal, episomal and virus-derived vectors, for example, vectors derived from bacterial plasmids, bacteriophages, yeast episomes, yeast chromosomal elements, and viruses, vectors derived from combinations thereof, such as those derived from plasmid and bacteriophage genetic elements, cosmids and phagemids. Thus, one exemplary vector is a double-stranded DNA phage vector. Another exemplary vector is a double-stranded DNA viral

vector. In one aspect, the invention relates to expression vectors, plasmids, and constructs as described herein, which are isolated and purified so as to be useful for any of a variety of applications, for example, as a reagent for scientific research, for human and/or veterinary use for therapeutic and/or prophylactic pharmaceutical purposes.

An 'isolated' polypeptide or polynucleotide means a polypeptide or polynucleotide that has been either removed from its natural environment, produced using recombinant techniques, or chemically or enzymatically synthesized. Preferably, a polypeptide or polynucleotide of this invention is purified, that is, it is essentially free from any other polypeptide or polynucleotide and associated cellular products or other impurities.

The term "Scar" refers to extra DNA sequences that are left as part of a polynucleotide construct that are an unavoidable consequence of the construction method rather than being incorporated because of their desirable functional properties. For example recombinases, integrases and restriction endonucleases often have recognition sequences that remain within the sequence of a polynucleotide that is constructed using the action of the recombinases, integrases and restriction endonucleases. The term "Scar Size" refers to the length of the extra DNA sequences. For example a scar size of 34 base pairs is left in a construct with a recognition sequence for Cre recombinase, a scar size of 25 base pairs is added on when attB integrase is used. Scars can interfere with the functions of other sequence elements within the construct.

The term "overhang" or "DNA overhang" refers to the single-stranded portion at the end of a double-stranded DNA molecule. Complementary overhangs are those which will base-pair with each other.

The term "Selectable marker" refers to a polynucleotide segment that allows one to select for or against a molecule or a cell that contains it, often under particular conditions. These markers can encode an activity, such as, but not limited to, production of RNA, peptide, or protein, or can provide a binding site for RNA, peptides, proteins, inorganic and organic compounds or compositions. Examples of Selectable markers include but are not limited to: (1) DNA segments that encode products which provide resistance against otherwise toxic compounds (e.g., antibiotics); (2) DNA segments that encode products which are otherwise lacking in the recipient cell (e.g., tRNA genes, auxotrophic markers); (3) DNA segments that encode products which suppress the activity of a gene product; (4) DNA segments that encode products which can be readily identified (e.g., phenotypic markers such as beta-galactosidase, green fluorescent protein (GFP), and cell surface proteins); (5) DNA segments that bind products which are otherwise detrimental to cell survival and/or function; (6) DNA segments that otherwise inhibit the activity of any of the DNA segments described in Nos. 1-5 above (e.g., antisense oligonucleotides); (7) DNA segments that bind products that modify a substrate (e.g. restriction endonucleases); (8) DNA segments that can be used to isolate a desired molecule (e.g. specific protein binding sites); (9) DNA segments that encode a specific nucleotide sequence which can be otherwise non-functional (e.g., for PCR amplification of subpopulations of molecules); and/or (10) DNA segments, which when absent, directly or indirectly confer sensitivity to particular compounds.

The term "Counter Selectable Marker" refers to a polynucleotide sequence that confers a selective disadvantage upon a host cell. Examples of counter-selectable markers include sacB, rpsL, tetAR, pheS, thyA, gata-1, ccdB, kid and

barnase (Bernard, 1995, *Journal/Gene*, 162: 159-160; Bernard et al., 1994, *Journal/Gene*, 148: 71-74; Gabant et al., 1997, *Journal/Biotechniques*, 23: 938-941; Gababt et al., 1998, *Journal/Gene*, 207: 87-92; Gababt et al., 2000, *Journal/Biotechniques*, 28: 784-788; Galvao and de Lorenzo, 2005, *Journal/AppI Environ Microbiol*, 71: 883-892; Hartzog et al., 2005, *Journal/Yeast*, 22:789-798; Knipfer et al., 1997, *Journal/Plasmid*, 37: 129-140; Reytrat et al., 1998, *Journal/Infect Immun*, 66: 4011-4017; Soderholm et al., 2001, *Journal/Biotechniques*, 31: 306-310, 312; Tamura et al., 2005, *Journal Appl Environ Microbiol*, 71: 587-590; Yazynin et al., 1999, *Journal/FEBS Lett*, 452: 351-354). Counter-selectable markers often confer their selective disadvantage in specific contexts. For example they may confer sensitivity to compounds that can be added to the environment of the host cell, or they may kill a host with one genotype but not kill a host with a different genotype. Conditions which do not confer a selective disadvantage upon a cell carrying a counter-selectable marker are described as "permissive". Conditions which do confer a selective disadvantage upon a cell carrying a counter-selectable marker are described as "restrictive".

The term "Recognition sequence" refers to particular DNA sequences which are recognized (and bound by) a protein, DNA, or RNA molecule, including a restriction endonuclease, a modification methylase, and a recombinase. For example, the recognition sequence for Cre recombinase is loxP which is a 34 base pair sequence comprised of two 13 base pair inverted repeats (serving as the recombinase binding sites) flanking an 8 base pair core sequence. See FIG. 1 of Sauer, B., *Current Opinion in Biotechnology* 5:521-527 (1994). Other examples of recognition sequences are the attB, attP, attL, and attR sequences which are recognized by the integrase of bacteriophage lambda. AttB is an approximately 25 base pair sequence containing two 9 base pair core-type Int binding sites and a 7 base pair overlap region. attP is an approximately 240 base pair sequence containing core-type Int binding sites and arm-type Int binding sites as well as sites for auxiliary proteins IHF, FIS, and Xis. See Landy, *Current Opinion in Biotechnology* 3:699-707 (1993). Such sites are also engineered according to the present invention to enhance methods and products.

The term "Recombinase" refers to an enzyme which catalyzes the exchange of DNA segments at specific recombination sites.

The term "Recombinational Cloning" refers to a method described herein, whereby segments of DNA molecules are exchanged, inserted, replaced, substituted or modified, in vitro or in vivo.

The term "Recombination proteins" includes excisive or integrative proteins, enzymes, co-factors or associated proteins that are involved in recombination reactions involving one or more recombination sites. See, Landy (1994), *infra*.

The term "expression system" refers to any in vivo or in vitro biological system that is used to produce one or more polypeptide encoded by a polynucleotide.

The term "annealing temperature" or "melting temperature" or "transition temperature" refers to the temperature at which a pair of nucleic acids is in a state intermediate between being fully annealed and fully melted. The term refers to the behavior of a population of nucleic acids: the "annealing temperature" or "melting temperature" or "transition temperature" is the temperature at which 50% of the molecules are annealed and 50% are separate. Annealing temperatures can be determined experimentally. There are also methods well known in the art for calculating these temperatures.

The term “translation” refers to the process by which a polypeptide is synthesized by a ribosome ‘reading’ the sequence of a polynucleotide.

The term “selectable protein” refers to a protein that provides a physical, chemical or biological method for selecting cells on the basis of how much of the selectable protein is expressed.

The term “coupling element” refers to a DNA sequence that allows the expression of a first polypeptide to be linked to the expression of a second polypeptide. Internal ribosome binding sites (IRES elements) and cis-acting hydrolase elements (CHYSEL elements) are examples of coupling elements.

The phrase “predetermined time period” refers to a specified amount of time. A “predetermined period of time” can be on the order of seconds, minutes, hours, days, weeks, or months. For example, a “predetermined time period” can be between 1 and 59 minutes, or any increment between 1 and 2 hours, or any increment between 2 and 4 hours, or any increment between 4 and 6 hours, or any increment between 6 and 12 hours, or any increment between 12 and 24 hours, or any increment between 1 day and 2 days, or any increment between 2 days and 4 days, and any increment between 4 days and 7 days, and any increment between 1 week and 4 weeks, and any increment between 1 month and 12 months, or any combination of incremental time periods therein.

The term “typeII restriction enzyme” is used herein to refer to any restriction enzyme that cleaves DNA at a defined distance outside its recognition sequence, and whose recognition sequence is non-palindromic.

The terms “ligatable ends” or “compatible ends” are used herein to describe two ends of polynucleotide molecules that are both blunt or that both possess overhangs of the same length and directionality (i.e. both are 5'-overhangs, or both are 3'-overhangs) and with perfectly complementary sequences, such that the DNA ends form standard Watson-Crick base pairs (i.e. C with G and T or U with A) and can be joined by a DNA ligase.

The term “operably linked” refers to functional linkage between two sequences such that one sequence modifies the behavior of the other. For example a first polynucleotide comprising a nucleic acid expression control sequence (such as a promoter, IRES sequence, enhancer or array of transcription factor binding sites) and a second polynucleotide are operably linked if the first polynucleotide affects transcription and/or translation of the second polynucleotide. Similarly a first amino acid sequence comprising a secretion signal or a subcellular localization signal and a second amino acid sequence are operably linked if the first amino acid sequence causes the second amino acid sequence to be secreted or localized to a subcellular location.

A “promoter” means a nucleic acid sequence sufficient to direct transcription of an operably linked nucleic acid molecule. Also included in this definition are those transcription control elements (for example, enhancers) that are sufficient to render promoter-dependent gene expression controllable in a cell type-specific, tissue-specific, or temporal-specific manner, or that are inducible by external signals or agents; such elements, which are well-known to skilled artisans, may be found in a 5' or 3' region of a gene or within an intron. Desirably, a promoter is operably linked to a nucleic acid sequence, for example, a cDNA or a gene sequence, or an effector RNA coding sequence, in such a way as to enable expression of the nucleic acid sequence, or a promoter is

provided in an expression cassette into which a selected nucleic acid sequence to be transcribed can be conveniently inserted.

‘Integration defective’ means a transposase that integrates a transposon at a lower frequency into the host genome than a corresponding wild type transposase.

As used herein, the term “transposon” or “transposable element” refers to a polynucleotide that can be excised from a first polynucleotide, for instance, a vector, and be integrated into a second position in the same polynucleotide, or into a second polynucleotide, for instance, the genomic or extrachromosomal DNA of a cell, by the action of a trans-acting transposase. A transposon comprises a first transposon end and a second transposon end which are polynucleotide sequences recognized by and transposed by a transposase. A transposon usually further comprises a first polynucleotide sequence between the two transposon ends, such that the first polynucleotide sequence is transposed along with the two transposon ends by the action of the transposase. Natural transposons frequently comprise DNA encoding a transposase that acts upon the transposon. The invention provides transposons in which a naturally present sequence encoding a functional transposase has been replaced with a sequence encoding a heterologous polynucleotide, which is transposable by virtue of its juxtaposition between the transposon ends.

As used herein, the term “transposon end” refers to cis-acting nucleotide sequences that are sufficient for recognition by and transposition by a transposase. A pair of transposon ends typically comprises paired perfect or imperfect repeats such that the respective repeats in the members of a pair are reverse complements of each other in the two different transposon ends. These are referred to as inverted terminal repeats (ITR) or terminal inverted repeats (TIR). In piggyBac-like transposons, each transposon end further comprises a target sequence immediately distal to the ITR (distal meaning on the side further from the transposase or heterologous polynucleotide transposed by the ITR). A transposon end may or may not include additional sequence proximal to the ITR that promotes or augments transposition.

As used herein, a “piggyBac-like transposase” means a transposase with at least 20% and preferably at least 30% sequence identity as identified using the TBLASTN algorithm to the piggyBac transposase from *Trichoplusia ni* (SEQ ID NO. 57), and as more fully described in Sakar, A. et. al., (2003). Mol. Gen. Genomics 270: 173-180. “Molecular evolutionary analysis of the widespread piggyBac transposon family and related ‘domesticated’ species”, and further characterized by a DDE-like DDD motif, with aspartate residues at positions corresponding to D268, D346, and D447 of *Trichoplusia ni* piggyBac transposase on maximal alignment. A “piggyBac-like transposon” means a transposon having transposon ends which are the same or at least 80% and preferably at least 90, 95, 96, 97, 98 or 99% identical to the transposon ends of a naturally occurring transposon that encodes a piggyBac-like transposase. A piggyBac-like transposon includes an inverted repeat sequence of approximately 13 bases at each end, immediately adjacent to a sequence corresponding to the target sequence which is duplicated upon transposon integration (the Target Site Duplication or Target Sequence Duplication or TSD). piggyBac-like transposons and transposases have been identified in a wide range of organisms including *Argyrogramma agnate* (GU477713), *Anopheles gambiae* (XP_312615; XP_320414; XP_310729), *Aphis gossypii* (GU329918), *Acyrtosiphon pisum* (XP_001948139), *Agro-*

tis epsilon (GU477714), *Bombyx mori* (BAD 11135), *Ciona intestinalis* (XP_002123602), *Chilo suppressalis* (JX294476), *Drosophila melanogaster* (AAL39784), *Daphnia pulex* (AAM76342), *Helicoverpa armigera* (ABS 18391), *Homo sapiens* (NP_689808), *Heliothis virescens* (ABD76335), *Macdunnoughia crassissima* (EU287451), *Macaca fascicularis* (AB179012), *Mus musculus* (NP_741958), *Pectinophora gossypiella* (GU270322), *Rattus norvegicus* (XP_220453), *Tribolium castaneum* (XP_001814566), *Trichoplusia ni* (AAA87375) and *Xenopus tropicalis* (BAF82026).

A target nucleic acid is a nucleic acid into which a transposon is to be inserted. Such a target can be part of a chromosome, episome or vector. The target nucleic acid for a transposon of the present invention should contain at least one motif recognized by a transposase of the present invention (5'-TTAT-3' or 5'-TTAA-3').

As used herein, a "target site" or "target sequence" for a transposase is a site or sequence in a target DNA molecule into which a transposon can be inserted by a transposase. The piggyBac transposase from *Trichoplusia ni* inserts its transposon into the target sequence 5'-TTAA-3'.

As used herein, the term "transposase" refers to a polypeptide that catalyzes the excision of a transposon from a donor polynucleotide, for example a vector, and (providing the transposase is not integration-deficient) the subsequent integration of the transposon into the genomic or extrachromosomal DNA of a target cell. The transposase binds a transposon end. The transposase may be present as a polypeptide. Alternatively, the transposase is present as a polynucleotide that includes a coding sequence encoding a transposase. The polynucleotide can be RNA, for instance an mRNA encoding the transposase, or DNA, for instance a coding sequence encoding the transposase. When the transposase is present as a coding sequence encoding the transposase, in some aspects of the invention the coding sequence may be present on the same vector that includes the transposon, that is, in cis. In other aspects of the invention, the transposase coding sequence may be present on a second vector, that is, in trans.

"IRES" or "internal ribosome entry site" means a specialized sequence that directly promotes ribosome binding, independent of a cap structure.

"Open Reading Frame" or ORF means a portion of a DNA molecule that, when translated into amino acids, contains no stop codons. The genetic code reads DNA sequences in groups of three base pairs, which means that a double-stranded DNA molecule can read in any of six possible reading frames—three in the forward direction and three in the reverse. A long open reading frame is likely a part of a gene.

Two elements are heterologous to one another if not naturally associated. For example, a nucleic acid sequence encoding a protein linked to a heterologous promoter means a promoter other than that which naturally expresses the protein. A heterologous nucleic acid flanked by transposon ends or ITRs means a heterologous nucleic acid not naturally flanked by those transposon ends or ITRs, such as a nucleic acid encoding a polypeptide other than a transposase, including an antibody heavy or light chain. Heterologous nucleic acids flanked by transposon ends or ITRs can vary in length, for example ranging from 20 base pairs to 20 kilo base pairs or more. A nucleic acid is heterologous to a cell if not normally found in the cell or in a different location (e.g., episomal or different genomic location) than the location naturally present within a cell.

Sequence identity can be determined by aligning sequences using algorithms, such as BESTFIT, FASTA, and TFASTA in the Wisconsin Genetics Software Package Release 7.0, Genetics Computer Group, 575 Science Dr., Madison, Wis.), using default gap parameters, or by inspection, and the best alignment (i.e., resulting in the highest percentage of sequence similarity over a comparison window). Percentage of sequence identity is calculated by comparing two optimally aligned sequences over a window of comparison, determining the number of positions at which the identical residues occurs in both sequences to yield the number of matched positions, dividing the number of matched positions by the total number of matched and mismatched positions not counting gaps in the window of comparison (i.e., the window size), and multiplying the result by 100 to yield the percentage of sequence identity. Unless otherwise indicated the window of comparison between two sequences is defined by the entire length of the shorter of the two sequences.

For purposes of classifying amino acids substitutions as conservative or nonconservative, amino acids are grouped as follows: Group I (hydrophobic sidechains): norleucine, met, ala, val, leu, ile; Group II (neutral hydrophilic side chains): cys, ser, thr; Group III (acidic side chains): asp, glu; Group IV (basic side chains): asn, gin, his, lys, arg; Group V (residues influencing chain orientation): gly, pro; and Group VI (aromatic side chains): tip, tyr, phe. Conservative substitutions involve substitutions between amino acids in the same class. Non-conservative substitutions constitute exchanging a member of one of these classes for a member of another.

The "configuration" of a polynucleotide means the functional sequence elements within the polynucleotide, and the order and direction of those elements. For example Tables 6-18

5.2 Description

5.2.1 Vector Components

The properties of a biological system including natural as well as non-natural systems with respect to any measurable function depends on the interaction between different nucleic acid sequence elements, which may be located at positions throughout the total nucleic acid component of the system, herein referred to as the "nucleic acid construct" of the system. The ability to rationally design a nucleic acid construct with an optimal configuration of elements is advantageous for various applications such as protein synthesis via vector optimization, cell line development and strain engineering. Protein synthesis is a highly dynamic and multi-step process and which plays a central role in synthetic biology, pharmaceutical production and other applications in biotechnology. This importance has led to the development of various parts or genetic control elements able to modulate and precisely control various aspects of protein expression. This capability is not only essential for the successful construction of more complex synthetic biological systems, but also provides tools needed for the tuning of their function for improved performance and reliability.

A limitation with the current state of the art is that effects of many control elements are dependent on the genetic context in which they are used. For example, combining the same promoter with different RBSs and genes can result in very different levels of expression.

While effects of combinations of one or two transcriptional or translational elements have been studied including the genetic context in which they are used, there remains a

need in the art to identify optimal configurations of multiple functional elements. Such elements can include those that influence DNA copy number, sites of DNA integration into chromosomes, RNA transcription rate, RNA degradation, RNA processing, RNA localization, translation initiation rate, and transcriptional termination. Examples of such elements are promoters, enhancers, introns, polyadenylation signals, ribosome binding sites, Kozak sequences, 5' untranslated sequences, origins of replication, nuclear export signals, internal ribosome entry sites and transcriptional terminators. Functional elements may also include those that encode functional polypeptides, such as secretion signals, resistance markers, anchoring peptides, localization signals, fusion tags, affinity tags, chaperonins and proteases. The ability to rationally engineer multiple elements within the DNA content of a host cell or expression system is an important aspect of this invention. In another embodiment, engineering of multiple elements within the DNA content of host cell or expression system along with variables of environmental stress or culture conditions is another aspect of this invention. Such environmental variables can include media components (whether complex or defined media), aeration, temperature, matrix for growth and others.

5.2.2 Genomic Integration

In some embodiments, the nucleic acid construct is a vector with enhanced expression and integration properties. For example, an optimal configuration of vector elements for enhanced transient expression as well as more efficient stable integration and expression was identified by the methods described herein. A mammalian vector construct variant set was generated using multiple combinations of various transposon ends or insulators, enhancers, promoters, introns, 5' untranslated regions (UTRs), 3' untranslated regions (UTRs), RNA export modulating sequence, polyadenylation sequences, terminators and matrix attachment element. The mammalian vector variant set was used to express DasherGFP in Human embryonic kidney (HEK 293) cells and Chinese hamster ovary (CHO) cells. Vector constructs with different promoter combinations were also shown to affect DasherGFP expression. Further optimization of this vector construct for different cell lines using the methods described herein is expressly contemplated. An advantage of the methods described herein is to quickly identify a subset of sequence elements most likely to influence desired activity as well as to facilitate predictable construction of optimal configuration of elements.

In some embodiments, elements that are useful in enhancing performance may include those localized to the genomic DNA of a cell. For example expression may be influenced by the levels of RNA polymerases, chaperonins, proteases, processing enzymes, or other factor encoded by DNA on the cell chromosome. It might also be advantageous to augment the host chromosome with functional elements that influence performance. In some embodiments, a variable for engineering is the site at which a functional gene is integrated into a host cell chromosome.

In some embodiments, the nucleic acid construct is a polynucleotide comprising of elements or combinations of elements arranged in an optimal configuration. In some embodiments, the polynucleotide is linear. In some embodiments, the elements in a nucleic acid construct comprise functional genetic features, for example, promoters, enhancers, introns, polyadenylation signals, origins of replication, and terminators. In some embodiments, the elements in a nucleic acid construct comprise protein encoding elements such as secretion signals, resistance markers, anchoring peptides, localization signals, and fusion tags. In some

embodiments, the plurality of elements comprises three or more elements, six or more elements, between three and twenty elements or between three and hundred elements. In some embodiments, nucleic acid construct variants include substitutions in a single element comprising one or more positions, three or more positions, six or more positions. In some embodiments, nucleic acid variants include substitutions wherein the substitutions are variations in elements and/or presence or absence of elements. In some embodiments, the substitutions include changes in the position of one or more elements. In some embodiments, nucleic acid variants include a change in the order of one or more elements.

5.2.3 Novel Transposons

DNA transposons undergo a 'cut and paste' system of replication in which the elements are physically excised from the one DNA molecule and reinserted in a second. The DNA transposons are characterized by inverted terminal repeats (ITRs) and are mobilized by an element-encoded transposase.

While DNA transposons are widespread and active in a variety of eukaryotes, they have been thought to be positionally inactive in mammalian genomes.

The natural process of horizontal gene transfer can be mimicked under laboratory conditions. In plants, transposons of the Ac/Ds and Spm families have been routinely transfected into heterologous species. In animals, however, a considerable obstacle to the transfer of an active transposon system from one species to another has been that of species-specificity of transposition due to the requirement for factors produced by the natural host.

A number of transposable elements have been described in the art that show no host-restriction in vertebrates, for example, an engineered transposon from the genome of salmonid fish called Sleeping Beauty; piggyBac transposon from lepidopteran cells; piggyBat transposon from the bat *Myotis lucifugus*; mariner transposon first discovered in *Drosophila* and; an engineered transposon and transposon inverted repeats from the frog species, *Rana pipiens* called frog prince; but the efficiency of transposition in cell lines derived from different species is variable. Therefore, it is advantageous to have a palette of different transposons with different host preferences to widen the potential of transposons as genomic tools in vertebrates.

It is currently used for many purposes including genome editing, enhancer trapping, gene discovery and identifying gene function in insects and mammals. The piggyBac transposon/transposase system is particularly useful because of the precision with which the transposon is integrated and excised (see for example "Fraser, M. J. (2001) The TTAA-Specific Family of Transposable Elements: Identification, Functional Characterization, and Utility for Transformation of Insects. Insect Transgenesis: Methods and Applications. A. M. Handler and A. A. James. Boca Raton, Fla., CRC Press: 249-268"; and "US 20070204356 A1: PiggyBac constructs in vertebrates" and references therein). This integration and excision is shown schematically in FIG. 1.

Many sequences with sequence similarity to the piggyBac transposase from *Trichoplusia ni* have been found in the genomes of phylogenetically distinct species from fungi to mammals, but very few have been shown to possess transposase activity (see for example Wu M, et al (2011) *Genetica* 139:149-54. Cloning and characterization of piggyBac-like elements in lepidopteran insects, and references therein). To discover novel transposons and transposases capable of transposing a heterologous polynucleotide into the genome of a host cell, we identified the sequences of 14 putative

transposases (SEQ ID NO: 43-56) by searching public sequence databases for polypeptides with sequence similarity to known active transposases. We then identified their corresponding transposon ends by taking the non-coding region associated with the gene sequence of the transposase. We designed and synthesized a polynucleotide to express each of the 14 transposases under control of the CMV promoter, and a second polynucleotide to express each of the 14 transposases fused to a heterologous nuclear localization signal under control of the CMV promoter. For each transposase we further designed two corresponding transposons. The first transposon comprised a heterologous polynucleotide comprising a puromycin resistance gene under control of a murine PGK promoter and a DasherGFP gene under control of a human EF1a promoter, with the two promoters oriented such that transcription from them is in opposite directions and divergent, surrounded by a pair of transposon ends. Sequences of transposon ends and corresponding transposases are described in Example 6.1.1 and shown in Tables 1 and 2. The second transposon comprised a heterologous polynucleotide comprising a puromycin resistance gene under control of a murine PGK promoter, with a DasherGFP gene translationally coupled to the puromycin-resistance gene through a CHYSEL sequence, surrounded by a pair of transposon ends. Sequences of transposon ends and corresponding transposases are described in Example 6.1.4 and shown in Table 5. We then transfected each transposon into CHO cells, in parallel sets with or without its corresponding transposase. Transposases that increased the reporter fluorescence from their transposons relative to the transposon-alone comprised novel transposon-transposase pairs that could integrate a heterologous polynucleotide into the genome of a cell.

As described in Examples 6.6.1 and 6.1.4 and shown in Tables 1, 2 and 5, of 14 that we identified and synthesized, only 4 showed detectable transposase activity. Thus sequence similarity to the *Trichoplusia ni* piggyBac sequence is insufficient to characterize a sequence as a transposase.

Using this method, we have identified two novel active piggyBac-like transposases together with their transposon ends and the ITR sequences on which they act.

One transposon was identified from the genome of *Xenopus tropicalis* with functional transposon ends being contained within SEQ ID NOS: 5, 6. Two transposases that can recognize and transpose these transposon ends are SEQ ID NOS: 45 and 46. Excision activity has been identified in Txb transposases from *Xenopus* (Hikosaka et. al., Mol. Biol. Evol., 24(12):2648-2656, 2007), but the authors conclude “In the present study, we demonstrated that the Xtr-Urbo2 Tase has excision activity toward the target transposon, although there is no evidence for the integration of the excised target into the genome” Here we have identified transposon ends including ITRs that can be placed at either end of a heterologous polynucleotide sequence to effect the efficient integration of the polynucleotide into genomic DNA by the action of the *Xenopus* transposase. Gene transfer vectors comprising *Xenopus* transposon ends and with optimal configurations of vector elements described herein (Section 5.2.6) show stable genomic integration even in the absence of transposase. In the presence of transposase, the expression from stably integrated transposons is increased 3-70 fold (see for example Table 14).

One transposon was identified from the genome of *Bombyx mori* with the functional transposon ends being contained within SEQ ID NOS: 1, 2. A transposase that can recognize and transpose a transposon comprising these

transposon ends is SEQ ID NOS: 44. We noted that the transposon end sequence associated with SEQ ID NO: 43 (SEQ ID NOS: 121 and 122) were terminated by the canonical 5'-TTAA-3' integration sequence always observed for transposons with significant sequence identity to *Trichoplusia ni* piggyBac. In contrast the transposon end sequence associated with SEQ ID NO: 44 (SEQ ID NOS: 1 and 2) were terminated by 5'-TTAT-3' sequences adjacent to the ITRs. We did not initially know whether this indicated that the transposase really used a novel integration sequence. Previous studies have indicated that the *Trichoplusia ni* piggyBac transposase is unable to transpose a transposon whose ends comprise a target sequence other than 5'-TTAA-3' (Mitra et al (2008) EMBO Journal 27: 1097-1109 “piggyBac can bypass DNA synthesis during cut and paste transposition”). Alternative possibilities for the 5'-TTAT-3' target sequence in the transposon ends within SEQ ID NOS: 1, 2 included sequencing errors or that the transposon had mutated and the transposase was no longer capable of transposing the sequence. Because active piggyBac-like transposons have only been described to use 5'-TTAA-3' integration sequences, we added the sequence 5'-TTAA-3' to both ends of the *Bombyx* transposon when we synthesized it to maximize the chance that we could reconstitute an active transposon (so the transposon sequence was arranged 5'-TTAATTAT-transposon end 1-heterologous polynucleotide-transposon end 2-TTATTTAA-3'). Contrary to the claims of Daimon et al (who describe *Bombyx* transposases as being essentially inactive, see Daimon T et al, 2010. Genome. 53:585-93. “Recent transposition of yabusame, a novel piggyBac-like transposable element in the genome of the silkworm, *Bombyx mori*.”) we found that the *Bombyx* transposon was highly active (see Examples 6.6.1 and Tables 1 and 2), we wished to determine its integration sequence. As shown in FIG. 1, when a piggyBac-like transposon is transposed, it leaves a single copy of its target sequence in the DNA from which it is excised. We therefore sequenced plasmids from which transposons had been excised by transposases. FIG. 2 shows a sequence trace file from a plasmid from which the *Bombyx* transposon has been excised. Both copies of the 5'-TTAA-3' sequence that we placed around the transposon ends are still present; however only a single copy of the 5'-TTAT-3' site remains. We examined 16 independent transposition events from *Bombyx* transposons, and all 16 left only a single perfectly intact copy of the 5'-TTAT-3' integration sequence. In contrast, when we examined 16 independent transposition events from *Xenopus* transposons, all 16 left only a single perfectly intact copy of the 5'-TTAA-3' integration sequence. Both *Xenopus* and *Bombyx* transposons disclosed here are transposed by their respective transposases with the same precision as has been described for *Trichoplusia ni* piggyBac, a precision which is highly advantageous for any genomic modifications that may be desirable to make reversibly. In contrast to *Trichoplusia ni* piggyBac, however, the *Bombyx mori* transposon thus comprises a 5'-TTAT-3' integration sequence, and the *Bombyx mori* transposase can excise and integrate *Bombyx mori* transposons at 5'-TTAT-3' recognition sequences. This is in contrast to all other known transposases with homology to *Trichoplusia ni* piggyBac, all of which recognize and insert transposons at the sequence 5'-TTAA-3'. This difference may be highly advantageous: *Trichoplusia ni* piggyBac has a preference for inserting transposons into transcriptionally inactive DNA. Because 5'-TTAT-3' is a reverse complement of 5'-ATAA-3' which is part of the canonical mammalian polyA signal 5'-aATAAa-3'. Thus the 5'-TTAT-3' insertion site recognized by the

Bombyx mori transposase will occur at almost every polyA signal. PolyA signals are associated with transcriptionally active regions of the chromosome. Thus transposons that insert at 5'-TTAT-3' sites, including the *Bombyx mori* transposons, are likely to yield higher expression levels of the genes they carry than transposons that insert 5'-TTAA-3' sites.

The invention provides a heterologous polynucleotide flanked by inverted repeats, which are in turn flanked by a direct repeat of the target sequence, 5'-TTAT-3'. In other words, from 5' to 3' such polynucleotides comprise a 5'-TTAT-3' target sequence, an ITR, a heterologous polynucleotide not naturally flanked by the ITR and 5'-TTAT-3', a second ITR in reverse orientation to the first ITR, and a second 5'-TTAT-3' target sequence. Transposition of such a transposon by a transposase leaves a single 5'-TTAT-3' motif in the locus previously occupied in a transposon. The transposon is transposed to a second polynucleotide including a 5'-TTAT-3' motif to generate a modified second polynucleotide including the transposon with the same components as when the transposon occupied the first polynucleotide.

The present application discloses a piggyBac-like *Bombyx* transposon (AB162707.1 GI:42600553) comprising transposon ends (each end including an ITR) corresponding to SEQ ID NO. 1 and 2, which has a target sequence corresponding to 5'-TTAT-3'. It also comprises a sequence encoding a transposase (SEQ ID NO. 44). A previously described *Bombyx* transposon (AB159601.1 GI:41016737) comprises one transposon end also identical to SEQ ID NO. 1 and a second transposon end corresponding to SEQ ID NO. 122. SEQ ID NO. 122 is very similar to SEQ ID NO. 2, but has a large insertion shortly before the ITR. Although the ITR sequences for the two transposons are identical (they are both identical to SEQ ID NO. 32), they have different target sequences: the second transposon has a target sequence corresponding to 5'-TTAA-3', providing evidence that no change in ITR sequence is necessary to modify the target sequence specificity. The *Bombyx* transposase (SEQ ID NO: 43), which is associated with the 5'-TTAA-3' target site differs from the 5'-TTAT-3'-associated transposase (SEQ ID NO: 44) by only 4 amino acid changes (D322Y, S473C, A507T, H582R). The transposase (SEQ ID NO: 43), which is associated with the 5'-TTAA-3' target site is less active than the 5'-TTAT-3'-associated transposase (SEQ ID NO: 44) on the transposon with 5'-TTAT-3' ends (see for example Table 5). These results provide evidence that other transposons with 5'-TTAA-3' target duplication sites can be converted to transposases with 5'-TTAT-3' target duplication sites by replacing 5'-TTAA-3' target duplication sites with 5'-TTAT-3'. Such novel transposons can be used either with *Bombyx* transposase (SEQ ID NO:43), which recognizes the 5'-TTAT-3' target sequence, or with a variant of the transposase originally associated with the 5'-TTAA-3' transposon. The high similarity between *Bombyx* 5'-TTAA-3' and 5'-TTAT-3' transposases provides evidence that very few changes to the amino acid sequence of a piggyBac-like transposase may confer altered target sequence specificity. The invention therefore provides transposon transposase transfer systems which can be formed by modification of any piggyBac-like transposon-transposase gene transfer system of which there are many known examples, in which 5'-TTAA-3' target sequences are replaced with 5'-TTAT-3'-target sequences, the ITRs remain the same, and the transposase is the original transposase or a variant thereof resulting from using a low-level mutagenesis to introduce mutations into the transposase. Similarly the invention also

provides transposon transposase transfer systems which can be formed by modification of a 5'-TTAT-3'-active piggyBac-like transposon-transposase gene transfer system such as the *Bombyx* system disclosed herein, in which 5'-TTAT-3' target sequences are replaced with 5'-TTAA-3'-target sequences, the ITRs remain the same, and the transposase is the original transposase or a variant thereof resulting from using a low-level mutagenesis to introduce mutations into the transposase.

Transposases that are active on a new target sequence such as a 5'-TTAT-3' target sequence can be selected, for example by coupling transposon excision to the production of a full coding sequence for a selectable marker. A transposon whose ITRs are flanked by the new target sequences (for example 5'-TTAT-3' target sequences) can be inserted into a selectable marker such that expression of the selectable marker is prevented. For example the transposon may be in the middle of an open reading frame encoding a selectable marker such that the presence of the transposon prevents the translation of a functional version of the selectable marker, or the transposon may be in an intron within an open reading frame encoding a selectable marker, such that the presence of the transposon prevents the splicing of the intron and thus prevents synthesis of the selectable marker. The transposon is placed such that when it is excised, the selectable marker regains functionality. For example the transposon may be placed at the sequence 5'-TTAT-3' within the coding sequence for a selectable marker such as an auxotrophic marker, the precise excision of the transposon from the open reading frame of the selectable marker restores the coding sequence of the selectable marker. A gene encoding the transposon-interrupted selectable marker and a gene encoding a transposase are introduced into a cell. The cell is then subjected to restrictive conditions that require expression of the selectable marker to enable the cell to survive. Expression of the selectable marker in turn depends upon excision of the transposon which in turn requires an active transposase. Cell survival can thus be used to identify active transposases. An example of such a selection scheme has been described by Yusa et al (Yusa et. al., PNAS, vol 108, no. 4, 1531-1536, 2011).

A transposase with modified activity, either for activity on a new target sequence including a 5'-TTAT-3' target sequence, or increased activity on an existing target sequence may be obtained by using variations of the selection scheme outlined above. To create the modified transposase, an existing transposase is used as a starting point, for example any of SEQ ID NOS: 43-57 or any other piggyBac-like transposase. One or more variant transposase sequences are created. These can be created in a variety of different ways, for example the gene may be subjected to random mutagenesis; the gene may be "DNA shuffled" with one or more homologous genes; systematic substitutions may be introduced into the gene including the creation of all possible single amino acid substitutions; substitutions may be introduced based on phylogenetic analysis and other rules for example as described in U.S. Pat. No. 8,635,029 B2. The sequence encoding the one or more variant transposases are operably linked to a promoter such that the transposases may be expressed in a cell. Each variant transposase is introduced into a cell that contains the transposon-interrupted transposable marker, and the cell is then subjected to restrictive conditions for which it requires the active selectable marker to survive. When a transposase is able to excise the transposon the cell will survive, and the gene encoding the active transposase may be recovered from the cell, for example by PCR. The process may be performed in pools of variants: a

more active transposase will create active selectable markers more frequently, and will thus be more highly represented in the population.

A comparable process may be used to increase the transposability of the transposon ends by a transposase. In this case, the transposon may comprise a first active selectable marker. Transposon ends may be selected from any of SEQ ID NOS: 1-31 or 32-42 or from any other piggyBac-like transposon including those associated with the inverted terminal repeats shown in SEQ ID NOS: 125-130. The sequence of one or both transposon ends may be subjected to random or pre-determined sequence changes, including changes to the target sequence, the ITR or to other parts of the transposon ends. The transposon may then be introduced into a first cell that contains a target polynucleotide comprising a second active selectable marker and an active transposase. If the transposase is able to transpose the transposon, some fraction of the transposons will be transposed into the target polynucleotide. The target polynucleotide is purified from the first cell, and introduced into a second cell which is subjected to restrictive conditions for which it requires the first selectable marker and the second selectable marker to survive. The transposon may be recovered, for example by sequencing out from the transposon to identify the flanking sequence, and then amplifying the transposon using PCR. The process may be performed in pools of variants: a more active transposon will create target polynucleotides containing both selectable markers more frequently, and will thus be more highly represented in the population. In this process, the transposon may optionally be present as a reversible interruption in a selectable marker as described for the transposase activity screen. However this is not necessary for the transposon activity screen, since the transposed transposons are detected directly.

These new transposases allow effective insertion of a transposon into a eukaryotic cell, including a mammalian cell such as a Chinese hamster ovary (CHO) cell or a Human embryonic kidney (HEK293) cell.

As well as exemplified transposases having sequences SEQ ID NOS. 43, 44, 45 and 46, the invention provides variants having at least 90, 91, 92, 93, 94, 95, 96, 97, 98 or 99% sequence identity to an exemplified sequence retaining transposase activity. Variations can be conservative or non-conservative substitutions, insertions and deletions. Deletions can be from the N-terminal or C-terminal end or internal. Some variants resulting from alanine scanning mutagenesis contain an alanine substitution at single positions throughout the molecule. Variations retain transposase activity on a transposon in which the exemplified transposase is activity. We have identified active transposases from *Bombyx mori* (SEQ ID NOS: 44) and *Xenopus tropicalis* (SEQ ID NOS: 45 and 46) that show variations in at least 4 residues that serve as a starting point for further studies for identification of hyperactive variants and integration deficient variants that retain excision activity. As used herein a *Bombyx mori* transposase or a *Bombyx* transposase refers to a polypeptide with at least 90% sequence identity with SEQ ID NO: 44 that can recognize and transpose a transposon. In some embodiments the transposon comprises two transposon ends, each of which comprises SEQ ID NO: 32 in inverted orientations in the two transposon ends. As used herein a *Xenopus tropicalis* transposase or a *Xenopus* transposase refers to a polypeptide with at least 90% sequence identity with SEQ ID NO: 45 or SEQ ID NO: 46 that can recognize and transpose a transposon. In some embodiments the transposon comprises two transpo-

son ends each of which comprise SEQ ID NO: 42 in inverted orientations in the two transposon ends.

These new transposases allow effective insertion of a transposon into a eukaryotic cell, including a mammalian cell such as a Chinese hamster ovary (CHO) cell or a Human embryonic kidney (HEK293) cell.

As well as exemplified transposases having sequences SEQ ID NOS. 43, 44, 45 and 46, the invention provides variants having at least 90, 91, 92, 93, 94, 95, 96, 97, 98 or 99% sequence identity to an exemplified sequence retaining transposase activity. Variations can be conservative or non-conservative substitutions, insertions and deletions. Deletions can be from the N-terminal or C-terminal end or internal. Some variants resulting from alanine scanning mutagenesis contain an alanine substitution at single positions throughout the molecule. Variations retain transposase activity on a transposon in which the exemplified transposase is activity. We have identified active variants of transposases from *Bombyx mori* (SEQ ID NOS: 43 and 44) and *Xenopus tropicalis* (SEQ ID NOS: 45 and 46) that show variations in at least 4 residues that serve as a starting point for further studies for identification of hyperactive variants and integration deficient variants that retain excision activity. As used herein a *Bombyx mori* transposase or a *Bombyx* transposase refers to a polypeptide with at least 90% sequence identity with SEQ ID NO: 43 or SEQ ID NO: 44 that can recognize and transpose a transposon. In some embodiments the transposon comprises two transposon ends, each of which comprises SEQ ID NO: 32 in inverted orientations in the two transposon ends. As used herein a *Xenopus tropicalis* transposase or a *Xenopus* transposase refers to a polypeptide with at least 90% sequence identity with SEQ ID NO: 45 or SEQ ID NO: 46 that can recognize and transpose a transposon. In some embodiments the transposon comprises two transposon ends each of which comprise SEQ ID NO: 42 in inverted orientations in the two transposon ends.

The invention also provides variants of exemplified sequences of transposon ends. The *Bombyx* transposase recognizes a transposon with a left sequence corresponding to SEQ ID NO: 1, and a right sequence corresponding to SEQ ID NO: 2. It will excise the transposon from one DNA molecule by cutting the DNA at the 5'-TTAT-3' sequence at the left end of one transposon end to the 5'-TTAT-3' at the right end of the second transposon end, including any heterologous DNA that is placed between them, and insert the excised sequence into a second DNA molecule. Truncated and modified versions of the left and right transposon ends will also function as part of a transposon that can be transposed by the *Bombyx* transposase. For example the left transposon end can be replaced by a shorter sequence corresponding to SEQ ID NO: 3, or the right transposon end can be replaced by a shorter sequence corresponding to SEQ ID NO: 4. It is thus expressly contemplated that modified versions of the transposon end sequences will be tolerated by the transposase, and may even result in increased transposition. In addition, we note that the left and right transposon ends share a 16 bp repeat sequence at their ends (SEQ ID NO: 32) immediately adjacent to the 5'-TTAT-3' insertion site, which is inverted in the orientation in the two ends. That is the left transposon end begins with the sequence 5'-TTATCCCGGCGAGCATGAGG-3' (SEQ ID NO: 33), and the right transposon ends with the reverse complement of this sequence: 5'-CCTCATGCTCGCCGGGTTAT-3' (SEQ ID NO: 34). The perfect conservation of this 16 bp sequence at both ends of the transposon suggests that it is important for function. One embodiment of the invention is

a transposon that comprises a heterologous polynucleotide inserted between SEQ ID NO: 33 and SEQ ID NO: 34. As used herein, a *Bombyx mori* transposon or a *Bombyx* transposon means a transposon comprising a heterologous polynucleotide and any of SEQ ID NOS: 32-34 or SEQ ID NOS: 1-4, or sub-sequences of SEQ ID NOS: 1-4, such that the heterologous polynucleotide can be transposed by a transposase at least 90% identical to SEQ ID NOS: 43 or 44.

The *Xenopus* transposase recognizes a transposon end with a left sequence corresponding to SEQ ID NO: 5, and a right sequence corresponding to SEQ ID NO: 6. It will excise the transposon from one DNA molecule by cutting the DNA at the 5'-TTAA-3' sequence at the left end of one transposon end to the 5'-TTAA-3' at the right end of the second transposon end, including any heterologous DNA that is placed between them, and insert the excised sequence into a second DNA molecule. Truncated and modified versions of the left and right transposon ends will also function as part of a transposon that can be transposed by the *Xenopus tropicalis* transposase. For example the left transposon end can be replaced by a sequence corresponding to SEQ ID NO: 7 or SEQ ID NO: 9, the right transposon end can be replaced by a shorter sequence corresponding to SEQ ID NO: 8. In addition, we note that the left and right transposon ends share an 18 bp almost perfectly repeated sequence at their ends (5'-TTAACCTTTTACTGCCA: SEQ ID NO: 42) that includes the 5'-TTAA-3' insertion site, which sequence is inverted in the orientation in the two ends. That is in SEQ ID NO: 5 and SEQ ID NO: 9 the left transposon end begins with the sequence 5'-TTAACCTTTTACTGCCA-3' (SEQ ID NO: 37), or in SEQ ID NO: 7 the left transposon end begins with the sequence 5'-TTAACCTTTTGCCTGCCA-3' (SEQ ID NO: 38); the right transposon ends with approximately the reverse complement of this sequence: in SEQ ID NO: 6 it ends 5'-TGGCAGTAAAAGGGTTAA-3' (SEQ ID NO: 40), in SEQ ID NO: 8 it ends 5'-TGGCAGT-GAAAGGGTTAA-3' (SEQ ID NO: 41). The near-perfect conservation of this 18 bp sequence at both ends of the transposon suggests that it is important for function. One embodiment of the invention is a transposon that comprises a heterologous polynucleotide inserted between two transposon ends each comprising SEQ ID NO: 42 in inverted orientations in the two transposon ends. In some embodiments one transposon end comprises a sequence selected from SEQ ID NOS: 37-39. In some embodiments one transposon end comprises a sequence selected from SEQ ID NOS: 40-41.

Further studies to identify additional variants of transposon ends and transposase sequences with enhanced activity are expressly contemplated. Generation of hyperactive variants is another aspect of this invention wherein mutational studies identify mutations in the transposase, which give rise to hyperactivity (Yusa et. al., PNAS, vol 108, no. 4, 1531-1536, 2011). Once the individual mutants are identified and verified for their transposition activity in cells, these mutations or combinations of mutations can be combined into one sequence to generate a hyperactive transposase that shows higher rates of transposition than the wild-type transposase. Hyperactive variants also include variants with enhanced integration activity, enhanced excision activity or both. Another aspect of this invention includes integration deficient variants, wherein the transposase shows lower integration activity but enhanced excision activity.

Accordingly, the present invention features *Bombyx* or *Xenopus* transposons and transposases. Another aspect of this invention refers to a transposon, that comprises a heterologous polynucleotide sequence, as described herein,

positioned between at least two ITRs, at least one repeat on either side of the heterologous polynucleotide, wherein these repeats can bind to a transposase protein and wherein the transposon is capable of inserting into DNA of a cell. Accordingly, repeats are preferably sequences that are recognized and bound by the transposase as defined herein.

According to certain preferred embodiments of the present invention, a transposon that is bound by a transposase contains a pair of repeat sequences. In certain preferred embodiments, the first repeat is typically located upstream of the heterologous polynucleotide and the second repeat is typically located downstream of the heterologous polynucleotide. The second repeat represents the same sequence as the first repeat, but shows an inverted orientation compared with the first repeat. That is, considering only one strand of a double-stranded DNA molecule, the second repeat will occur as the reverse complement of the first repeat. In some embodiments these repeats are identical inverted sequences. In some embodiments these inverted repeats may be highly similar but not identical, differing by 1, 2, 3 or 4 nucleotides. These repeats are then termed "inverted repeats" (IRs) or "inverted terminal repeats" (ITRs), due to the fact that each repeat is an inversely repeated copy of the other. In certain embodiments, repeats may occur in a multiple number upstream and downstream of the above mentioned nucleic acid sequence. In certain embodiments, the repeats are short, between 10-20 base pairs, and preferably 14-16 base pairs. In some other embodiments, the transposon ends further comprise additional sequences that may or may not be repeats.

In some embodiments, the *Xenopus* transposon comprises one end comprising at least 14 or 16 or 18 or 20 or 30 or 40 contiguous nucleotides from SEQ ID NO: 5, 7 or 9; in some embodiments, the *Xenopus* transposon comprises one end comprising at least 14 or 16 or 18 or 20 or 30 or 40 contiguous nucleotides from SEQ ID NO: 6 or 8. In some embodiments, the *Xenopus* transposon comprises one end with at least 90% identity to SEQ ID NO: 5 or 7 or 9; in some embodiments, the *Xenopus* transposon comprises one end with at least 90% identity to SEQ ID NO: 6 or 8. In some embodiments of the *Xenopus* transposon, each inverted terminal repeat (ITR) comprises SEQ ID NO: 42. In some embodiments of the *Xenopus* transposon, one ITR comprises a sequence selected from SEQ ID NOS: 37-39. In some embodiments of the *Xenopus* transposon, one ITR comprises a sequence selected from SEQ ID NOS: 40-41.

In some embodiments, the *Bombyx* transposon comprises one end comprising at least 14 or 16 or 18 or 20 or 30 or 40 contiguous nucleotides from SEQ ID NO: 1 or 3; in some embodiments, the *Bombyx* transposon comprises one end comprising at least 14 or 16 or 18 or 20 or 30 or 40 contiguous nucleotides from SEQ ID NO: 2 or 4. In some embodiments, the *Bombyx* transposon comprises one end with at least 90% identity to SEQ ID NO: 1 or 3; in some embodiments, the *Bombyx* transposon comprises one end with at least 90% identity to SEQ ID NO: 2 or 4.

In some embodiments, the *Bombyx* transposon comprises one end comprising at least 16 contiguous nucleotides from SEQ ID NO: 33 and one end comprising at least 16 contiguous nucleotides from SEQ ID NO: 34. In some embodiments, each ITR of the *Bombyx* transposon comprises SEQ ID NO: 32. In these embodiments, SEQ ID NO: 32 is immediately adjacent to the sequence 5'-TTAT-3' or 5'-TTAA-3'.

The ITRs as described herein preferably flank a nucleic acid sequence which is inserted into the DNA of a cell. The nucleic acid sequence can include all or part of an open

reading frame of a gene (i.e., that part of a protein encoding gene), one or more expression control sequences (i.e., regulatory regions in nucleic acid) alone or together with all or part of an open reading frame. Preferred expression control sequences include, but are not limited to promoters, enhancers, introns, polyadenylation sequences, border control elements, locus-control regions; expression enhancers that enhance RNA export from the nucleus, including woodchuck hepatitis post-transcriptional regulatory element (WPRE), hepatitis B virus post-transcriptional regulatory element (HPRE) (for example but not limited to SEQ ID NO: 104-105) and arctic ground squirrel post-transcriptional regulatory element (AGS) (for example but not limited to SEQ ID NOS: 106-107); and elements whose mechanism of action may not be known, such as scaffold attachment region (SAR) sequences (for example but not limited to SEQ ID NOS: 108-111), and insulator sequences that are thought to prevent the spread of condensed chromatin that might otherwise silence expression and prevent interference from a distal enhancer on a promoter, for example HS4 (for example but not limited to SEQ ID NOS: 112-113) (Yusufzai et. al., PNAS, vol. 101, no. 23, 8620-8624, June 2004).

Cells whose genomes contain a *Bombyx mori* transposon or a *Xenopus tropicalis* transposon are an aspect of the invention.

In a preferred embodiment, the nucleic acid sequence comprises a promoter operably linked to an open reading frame. The open reading frame may comprise a selectable marker that enables selection by a demonstrable phenotype, for example a fluorescent reporter. According to certain preferred embodiment, transposons of the present invention can preferably occur as a linear transposon (extending from the 5' end to the 3' end, by convention) that can be used as a linear fragment or circularized, for example in a plasmid.

Activity of transposases may be increased by fusion of nuclear localization signal (NLS) at the N-terminus, C-terminus, both at the N- and C-termini or internal regions of the transposase protein so long as transposase activity is retained. A nuclear localization signal or sequence (NLS) is an amino acid sequence that 'tags' or facilitates interaction of a protein, either directly or indirectly with nuclear transport proteins for import into the cell nucleus. Nuclear localization signals (NLS) used can include, but are not limited to, consensus NLS sequences, viral NLS sequences, cellular NLS sequences, and combinations thereof. In preferred embodiments, the NLS sequences are operably linked to the transposase.

The transposase protein can be introduced into a cell as a protein or as a nucleic acid encoding the transposase, for example as a ribonucleic acid, including mRNA, as DNA, e.g. as extrachromosomal DNA including, but not limited to, episomal DNA, as plasmid DNA, or as viral nucleic acid. Furthermore, the nucleic acid encoding the transposase protein can be transfected into a cell as a nucleic acid vector such as a plasmid, or as a gene expression vector, including a viral vector. The nucleic acid can be circular or linear. A vector, as used herein, refers to a plasmid, a viral vector or a cosmid that can incorporate nucleic acid encoding the transposase protein or the transposon of this invention. DNA encoding the transposase protein can be stably inserted into the genome of the cell or into a vector for constitutive or inducible expression. Where the transposase protein is transfected into the cell or inserted into the vector as nucleic acid, the transposase encoding sequence is preferably operably linked to a heterologous promoter. There are a variety of promoters that could be used including, but not limited to, constitutive promoters, tissue-specific promoters, inducible

promoters, and the like. All DNA or RNA sequences encoding the *Bombyx mori* or *Xenopus tropicalis* transposase proteins are expressly contemplated.

5.2.4 Gene Transfer System

The present invention also features a gene transfer system comprising a *Bombyx mori* transposase and a transposon comprising a heterologous polynucleotide between a left ITR and a right ITR that are recognized and transposed by the transposase. The present invention also features a gene transfer system comprising a *Xenopus tropicalis* transposase and a transposon comprising a heterologous polynucleotide between a left ITR and a right ITR that are recognized and transposed by the transposase. The transposase can be encoded on the same polynucleotide as the transposon in the gene transfer system, or it may be encoded on a second polynucleotide. If the transposase is encoded on the same nucleic acid molecule as the transposon, the transposase is preferably in a part of the molecule that is not transposed. The gene transfer system of this invention, therefore, preferably comprises two components: the transposase as described herein and a transposon as described herein. In combination these two components provide active transposon activity and allow the transposon to be relocated. In use, the transposase binds to the transposon ends and promotes insertion of the intervening nucleic acid sequence into DNA of a cell as defined below.

In some embodiments, a gene transfer vector further comprises sequences encoding the transposase fused to certain protein functional domains. Such protein functional domains can include, but are not limited to, one or more DNA binding domains, one or more nuclear localization signals, one or more flexible hinge regions that can facilitate one or more domain fusions, and combinations thereof. Fusions can be made either to the N-terminus, C-terminus, or internal regions of the transposase protein so long as transposase activity is retained. DNA binding domains used can include, but are not limited to, a helix-turn-helix domain, Zn-finger domain, a leucine zipper domain, or a helix-loop-helix domain. Specific DNA binding domains used can include, but are not limited to, a Gal4 DNA binding domain, a LexA DNA binding domain, or a Zif268 DNA binding domain. Nuclear localization signals (NLS) used can include, but are not limited to, consensus NLS sequences, viral NLS sequences, cellular NLS sequences, and combinations thereof. Flexible hinge regions used can include, but are not limited to, glycine/serine linkers and variants thereof.

In further exemplary embodiments, the gene transfer system comprises a *Bombyx mori* transposon as defined above in combination with a *Bombyx mori* transposase protein (or nucleic acid encoding the *Bombyx mori* transposase protein to provide its activity in a cell). In other embodiments, the gene transfer system comprises a *Xenopus tropicalis* transposon as defined above in combination with a *Xenopus tropicalis* transposase protein (or nucleic acid encoding the *Xenopus tropicalis* transposase protein to provide its activity in a cell). This combination preferably results in the insertion of the nucleic acid sequence into the DNA of the cell. Alternatively, it is possible to insert the *Bombyx mori* or *Xenopus tropicalis* transposon into DNA of a cell through non-homologous recombination through a variety of reproducible mechanisms, and even without the activity of a transposase. In either event the described transposon can be used for gene transfer by using this gene transfer system.

In certain embodiments, a gene transfer vector further comprises a recombination protein, for example a recombinase, an integrase or a transposase including a *Bombyx mori*

transposase or *Xenopus tropicalis* transposase and two or more site specific integration recognition sites to facilitate integration of an expression cassette into the genome of an expression host. In certain embodiments, these integration-facilitating sequences include a 5'-TTAA-3'-target sequence specific insertion element. In some other embodiments, the integration-facilitating sequences include a 5'-TTAT-3'-target sequence specific insertion element. In certain embodiments the integration-facilitating sequences are recognized by an integrase or a transposase, in certain embodiments the integrase is a *Bombyx mori* integrase, in other embodiments the integrase is a *Xenopus tropicalis* integrase. In certain embodiments the gene transfer vector further comprises a gene encoding the integrase. In certain embodiments, an expression vector further comprises Lentiviral LTR (long terminal repeats) or inverted repeats (IR) to facilitate integration of an expression cassette into the genome of an expression host.

In some embodiments, a gene transfer vector has restriction endonuclease sites in the expression cassette between the promoter and terminator sequence that facilitate cloning of heterologous polynucleotides for insertion. In preferred embodiments, these restriction endonuclease sites are type IIs restriction sites. Type IIs restriction endonucleases recognize asymmetric DNA sequences and cleave both DNA strands at fixed positions, typically several base pairs away from the recognition sites. This property makes type IIs restriction endonucleases particularly useful for assembling DNA fragments, where fragments with matching type IIs-generated ends are annealed and ligated, leaving an assembled DNA product without restriction recognition sequence scars at the ligation junctions. Type IIs restriction endonucleases that recognize non-palindromic sequences of 5, 6 or 7 base pairs, are found at an average frequency of one in 512, 2048 or 8192 base pairs respectively. It is therefore, relatively easy to identify type IIs restriction endonucleases that do not cut inside a typical gene-sized DNA fragment or a gRNA fragment.

A gene transfer vector can be constructed to permit cloning using type IIs restriction endonucleases and ligase by incorporating a stuffer, comprising a counter-selectable marker and flanked by type IIs restriction sites, into a vector comprising a selectable marker. It is advantageous if the type IIs restriction sites are chosen such that cleavage of the gene transfer vector with one or more type IIs restriction enzymes yields a linear nucleic acid fragment comprising a selectable marker and with ends that are not compatible with each other. This design allows directional insertion of an insert DNA fragment that has cohesive ends compatible with the linear nucleic acid fragment of the gene transfer vector. The insert DNA fragment may be prepared by annealing a pair of oligonucleotides, or more preferably by PCR amplification and restriction digestion. In preferred embodiments the gene transfer vector ends are also not pseudo-compatible with each other; that is they do not anneal with each other by forming at least one non-standard Watson-Crick base pair (i.e., T or U with G) in a way that can be joined by a DNA ligase with reasonable efficiency.

In certain preferred embodiments, the gene transfer system mediates insertion of the *Bombyx mori* or *Xenopus tropicalis* transposon into the DNA of a variety of cell types and a variety of species by using the *Bombyx mori* or *Xenopus tropicalis* transposase protein. Preferably, such cells include any cell suitable in the present context, including but not limited to animal cells or cells from bacteria, fungi (example, yeast and more) or plants. Preferred animal cells can be vertebrate or invertebrate. For example, pre-

ferred vertebrate cells include cells from mammals including, but not limited to, rodents, such as rats or mice, ungulates, such as cows or goats, sheep, swine or cells from a human. Target cells also include without being limited thereto, lymphocytes, hepatocytes, neural cells, muscle cells, a variety of blood cells, and a variety of cells of an organism, embryonic stem cells, somatic stem cells e.g. hematopoietic cells, embryos, zygotes, sperm cells (some of which are open to be manipulated by an in vitro setting).

In other further exemplary embodiments, such cells, particularly cells derived from a mammal as defined above, can be pluripotent (i.e., a cell whose descendants can differentiate into several restricted cell types, such as hematopoietic stem cells or other stem cells) and totipotent cells (i.e., a cell whose descendants can become any cell type in an organism, e.g., embryonic stem cells). These cells are advantageously used to affirm stable expression of the transposase or to obtain a multiple number of cells already transfected with the components of the gene transfer system. Additionally, cells such as oocytes, eggs, and one or more cells of an embryo may also be considered as targets for stable transfection with the present gene transfer system. In certain preferred embodiments of the invention, the cells are Chinese hamster ovary (CHO) cells or Human embryonic kidney (HEK293) cells.

In other certain exemplary embodiments, the cell DNA that acts as a recipient of the transposon described herein includes any DNA present in a cell (as mentioned above) to be transfected, if the *Bombyx mori* or *Xenopus tropicalis* transposon is in contact with a *Bombyx mori* or *Xenopus tropicalis* transposase protein within the cell. For example, the DNA can be part of the cell genome or it can be extrachromosomal, such as an episome, a plasmid, a circular or linear DNA fragment. Typical targets for insertion are for example, double-stranded DNA.

The components of the gene transfer system described herein, that is the *Bombyx mori* or *Xenopus tropicalis* transposase protein (either as a protein or encoded by a nucleic acid as described herein) and the *Bombyx mori* or *Xenopus tropicalis* transposon can be transfected into a cell, preferably into a cell as defined above, and more preferably into the same cell. Transfection of these components may furthermore occur in subsequent order or in parallel. For example, the *Bombyx mori* or *Xenopus tropicalis* transposase protein or its encoding nucleic acid may be transfected into a cell as defined above prior to, simultaneously with or subsequent to transfection of the *Bombyx mori* or *Xenopus tropicalis* transposon. Alternatively, the transposon may be transfected into a cell as defined above prior to, simultaneously with or subsequent to transfection of the *Bombyx mori* or *Xenopus tropicalis* transposase protein or its encoding nucleic acid. If transfected in parallel, preferably both components are provided in a separated formulation and/or mixed with each other directly prior to administration to avoid transposition prior to transfection. Additionally, administration of at least one component of the gene transfer system may occur repeatedly, for example, by administering at least one, two or multiple doses of this component.

For any of the above transfection reactions, the gene transfer system may be formulated in a suitable manner as known in the art, or as a pharmaceutical composition or kit as described herein. In further preferred embodiments, the components of the gene transfer system may preferably be transfected into one or more cells by techniques such as particle bombardment, electroporation, microinjection, combining the components with lipid-containing vesicles, such as cationic lipid vesicles, DNA condensing reagents

(example, calcium phosphate, polylysine or polyethyleneimine), and inserting the components (that is the nucleic acids thereof into a viral vector and contacting the viral vector with the cell. Where a viral vector is used, the viral vector can include any of a variety of viral vectors known in the art including viral vectors selected from the group consisting of a retroviral vector, an adenovirus vector or an adeno-associated viral vector.

In another embodiment, the nucleic acid encoding the *Bombyx mori* or *Xenopus tropicalis* transposase protein may be RNA or DNA. Similarly, either the nucleic acid encoding the *Bombyx mori* or *Xenopus tropicalis* transposase protein or the transposon of this invention can be transfected into the cell as a linear fragment or as a circularized fragment, preferably as a plasmid or as recombinant viral DNA.

In another embodiment, the nucleic acid encoding the *Bombyx mori* or *Xenopus tropicalis* transposase protein is thereby preferably stably or transiently inserted into the genome of the cell to facilitate temporary or prolonged expression of the *Bombyx mori* or *Xenopus tropicalis* transposase protein in the cell.

5.2.5 Increasing Expression by Selection

Sequences that are integrated at regions of the genome that are highly transcriptionally active may result in high levels of expression of encoded genes. In addition, or alternatively, sequences that are integrated into the genome in multiple copies may result in high levels of expression of encoded genes.

Methods are known in the art to increase the expression of a first polypeptide encoded by a construct (the expression polypeptide) by attempting to link the expression of the first polypeptide to the expression of a second quantitatively selectable polypeptide. For example, Glutamine synthase (GS) is used as a selectable marker that allows selection via glutamine metabolism. Glutamine synthase is the enzyme responsible for the biosynthesis of glutamine from glutamate and ammonia, and is a crucial component of the only pathway for glutamine formation in a mammalian cell. In the absence of glutamine in the growth medium, the GS enzyme is essential for the survival of mammalian cells in culture. Some cell lines, for example mouse myeloma cells do not express sufficient GS to survive without added glutamine. In these cells a transfected GS can function as a selectable marker by permitting growth in a glutamine-free medium. In other cell lines, for example Chinese hamster ovary (CHO) cells express sufficient GS to survive without exogenously added glutamine. These cell lines can be manipulated by genome editing techniques including CRISPR/Cas9 to reduce or eliminate the activity of the GS enzyme. In all of these cases, GS inhibitors such as methionine sulfoximine (MSX) can be used to inhibit a cell's endogenous GS activity. Selection protocols known in the art include introducing a construct comprising sequences encoding a first polypeptide and a glutamine synthase selectable marker, and then treating the cell with inhibitors of glutamine synthase such as methionine sulfoximine. The higher the levels of methionine sulfoximine that are used, the higher the level of glutamine synthase expression is required to allow the cell to synthesize sufficient glutamine to survive. Some of these cells will also show an increased expression of the first polypeptide.

A second system for increasing expression by selection uses the enzyme dihydrofolate reductase (DHFR) which is required for catalyzing the reduction of 5,6-dihydrofolate (DHF) to 5,6,7,8-tetrahydrofolate (THF) and is used as a selectable marker. DHFR confers resistance to methotrexate (MTX). DHFR can be inhibited by higher levels of metho-

trexate. Selection protocols known in the art include introducing a construct comprising sequences encoding a first polypeptide and a DHFR selectable marker into a cell, and then treating the cell with inhibitors of DHFR such as methotrexate. The higher the levels of methotrexate that are used, the higher the level of DHFR expression is required to allow the cell to synthesize sufficient DHFR to survive. Some of these cells will also show an increased expression of the first polypeptide.

The use of transposons and transposases in conjunction with such quantitatively selectable markers has several advantages over non-transposon constructs. One is that linkage between expression of the first polypeptide and the quantitatively selectable marker is better for transposons, because a transposase will integrate the entire sequence that lies between the two transposon ends into the genome. In contrast when heterologous DNA is introduced into the nucleus of a eukaryotic cell, for example a mammalian cell, it is gradually broken into random fragments which may either be integrated into the cell's genome, or degraded. Thus if a construct comprising sequences that encode a first polypeptide and a quantitatively selectable marker is introduced into a population of cells, some cells will integrate the sequences encoding the quantitatively selectable marker but not those encoding the first polypeptide, and vice versa. Selection of cells expressing high levels of selectable marker is thus only somewhat correlated with cells that also express high levels of the first polypeptide. In contrast, because the transposase integrates all of the sequences between the transposon ends, cells expressing high levels of selectable marker are highly likely to also express high levels of the first polypeptide.

A second advantage of transposons and transposases is that they are much more efficient at integrating DNA sequences into the genome. Thus a much higher fraction of the cell population is likely to receive one or more copies of the construct in their genomes, so there will be a correspondingly higher likelihood of good stable expression of both the selectable marker and the first polypeptide.

One embodiment of the present invention is thus a transposon such as described above, that further comprises a sequence encoding a first polypeptide and a selectable marker that can be inhibited by a small molecule inhibitor. In one embodiment the first polypeptide is part of an antibody. Other aspects of the invention include methods for introducing the transposon into the genome of a cell using a transposase, and selecting for high levels of expression of the quantitatively selectable marker. In some embodiments the selectable marker is glutamine synthase, in some embodiments the selectable marker is DHFR.

DNA transposases use a cut-and-paste mechanism for inserting their transposon into a DNA molecule. The number of copies of a transposon that can be integrated into the genome by a transposase is thus limited by the number of copies of the transposon that are present in the cell. The number of nuclear non-integrated copies of a plasmid in a eukaryotic cell can be increased if it contains viral replication sequences. For example in mammalian cells including CHO cells and HEK cells the SV40 origin of replication causes increases in the number of copies of a plasmid especially in the presence of the SV40 large T antigen. Similarly the Epstein-Barr virus origin of replication (OriP) causes increases in the number of copies of a plasmid especially in the presence of the Epstein-Barr virus nuclear antigen 1 (EBNA) and its truncated derivatives. Plasmids comprising a transposon in addition to viral replication sequences such as the SV40 origin of replication or the

Epstein-Barr virus OriP which are not contained within the transposable portion of the transposon will therefore accumulate within the nucleus, providing more substrate copies of the transposon to be integrated into the cell genome. Such plasmids are an aspect of the current invention. The use of

such plasmids to increase the number of copies of a transposon that is integrated into a target cell genome is also an aspect of the current invention. These plasmids may further comprise sequences encoding the SV40 large T antigen or the Epstein-Barr virus nuclear antigen 1 (EBNA).

Plasmids comprising viral replication sequences and transposons may be introduced into cells together with the transposase, or they may be introduced sequentially. Higher numbers of integrated transposons may be selected using quantitatively selectable markers such as DHFR or glutamine synthase.

5.2.6 Gene Transfer Vector Components

The function of sequence elements is dependent upon the context relative to the other sequences within the DNA sequence. An embodiment of the present invention provides a method for constructing a gene transfer vector variant set to improve an expression property of a polynucleotide encoding a polypeptide. In some embodiments the expression property is an amount of the polypeptide expressed, in some embodiments the expression property is an amount of soluble polypeptide expressed, in some embodiments the expression property is an amount of active polypeptide expressed. In the method, a plurality of sequence elements are identified in a gene transfer vector. The plurality of elements are classified by functional grouping, for example the elements are classified as enhancers, promoters, introns, 5' untranslated regions, 3' untranslated regions, RNA export promoting elements, elements that modulate chromatin structure, polyadenylation signals or transcriptional terminators. In addition, if the gene transfer vector will express more than one gene, the elements are further grouped according to the gene to which they are operably linked. A first gene transfer vector variant set comprising a plurality of configurations of the gene transfer vector is selected, such that members of the gene transfer vector set are related to one another by the substitution of one or more sequence elements, with a different element from the same functional group, or by the complete removal of an element of that functional group. For example a second member of the polynucleotide vector set may have the same configuration as a first member, but with a first enhancer element replaced by a second enhancer element, or a second member of the polynucleotide vector set may have the same configuration as a first member, but lack an enhancer element. A functional group in which there is more than one possible element in the gene transfer vector variant set is referred to as a variant group.

The number of variant groups and the number of elements that can be tested at each of those group positions is then calculated, such that each element will be present in a statistically representative fraction of the first gene transfer vector variant set. Additionally, when using search methods like Tabu, Ant optimization or similar techniques, the space can be searched on a sequence by sequence basis by using a memory of the space that has been visited previously and the properties encountered.

In some embodiments, selection of the variant set comprises applying complete factorial design, a 2^k factorial design, a 2^k fractional factorial design, a latin squares approach, a greco-latin squares approach, a Plackett-Burmann design, a Taguchi design, a monte carlo algorithm, a genetic algorithm, combinations thereof or some other sta-

tistical method for Design of Experiment, to the distribution of elements in the gene transfer vector variant set.

A first expression set, comprising a first expression polynucleotide in all or a portion of the first gene transfer vector variant set, is constructed. An expression property of the first expression set is measured. In some embodiments the expression polynucleotide encodes a first polypeptide, and the expression property is an amount of the polypeptide expressed, or an amount of soluble polypeptide expressed, or an amount of active polypeptide expressed. In some embodiments the first expression polynucleotide is the same sequence in all of the sequences of the first expression set.

In some embodiments of the invention the expression property is measured in one of the following expression systems: bacterial expression systems including *Escherichia coli*, *Salmonella* species, *Bacillus* species, *Streptomyces* species, *Pseudomonas* species, *Ralstonia eutropha*, *Chlamydomonas* species; yeast expression systems including *Saccharomyces*, *Pichia*, *Klebsiella* and *Candida* species, *Saccharomyces cerevisiae*, *Pichia pastoris*, *Pichia methanolica*, *Klebsiella lactis*; fungal expression systems including *Cryptosporidium* and *Trichoderma* species, filamentous fungal protein production systems, protozoan expression systems including *Plasmodium falciparum* (the causative agent of malaria), *Leishmania* model organisms including *Caenorhabditis elegans*, *Drosophila melanogaster*, *Xenopus laevis*; plants including soybean, bushbean, maize, cotton, tobacco, *Arabidopsis*, tissue culture expression systems including COS cells, Chinese Hamster Ovary cells and fibroblasts including 3T3 cells, cell lines infected with adenovirus, insect cell lines such as those derived from *Spodoptera* species for growing baculovirus; model organisms for the study of disease and tests of the efficacies of DNA vaccines such as macaques, mice, rats, guinea pigs, sheep, goats and rabbits; in vitro expression systems prepared from extracts of living cells including *E coli* extracts, wheat germ extracts, rabbit reticulocyte lysates; in vitro expression systems prepared by assembly of purified individual components.

Standard techniques may be utilized to measure the expression property value for each respective polynucleotide in the plurality of polynucleotides of the first expression set. For example, standard techniques can be employed using, e.g., immunoassays such as, for example Western blot, immunoprecipitation followed by sodium dodecyl sulfate polyacrylamide gel electrophoresis, (SDS-PAGE), immunocytochemistry, and the like to determine an expression property value of a respective polynucleotide (e.g., an amount of a protein encoded by the respective polynucleotide) in the plurality of polynucleotides present in an expression system. Other methods for detection of specific polypeptides include mass spectroscopy and mass spectroscopy of protein samples that have been treated with one or more site specific proteases to produce polypeptide fragments which can be uniquely identified by mass spectroscopy. One exemplary agent for detecting a protein of interest is an antibody capable of specifically binding to a protein of interest, preferably an antibody detectably labeled, either directly or indirectly.

One of the ways in which an antibody specific for a protein of interest can be detectably labeled is by linking the same to an enzyme and use in an enzyme immunoassay (EIA) (Voller, 1978, "The Enzyme Linked Immunosorbent Assay (ELISA)", Diagnostic Horizons 2:1-7, Microbiological Associates Quarterly Publication, Walkersville, Md.; Voller et al., 1978, J. Clin. Pathol. 31:507-520; Butler, J. E., 1981, Meth. Enzymol. 73:482-523; Maggio (ed.), 1980,

Enzyme Immunoassay, CRC Press, Boca Raton, Fla.; Ishikawa et al., (eds.), 1981, *Enzyme Immunoassay*, Kaku Shoin, Tokyo, each of which is hereby incorporated by reference in its entirety). The enzyme which is bound to the antibody will react with an appropriate substrate, preferably a chromogenic substrate, in such a manner as to produce a chemical moiety which can be detected, for example, by spectrophotometric, fluorimetric or by visual means. Enzymes which can be used to detectably label the antibody include, but are not limited to, malate dehydrogenase, staphylococcal nuclease, delta-5-steroid isomerase, yeast alcohol dehydrogenase, alpha-glycerophosphate dehydrogenase, triose phosphate isomerase, horseradish peroxidase, alkaline phosphatase, asparaginase, glucose oxidase, beta-galactosidase, ribonuclease, urease, catalase, glucose-6-phosphate dehydrogenase, glucoamylase and acetylcholinesterase. The detection can be accomplished by colorimetric methods which employ a chromogenic substrate for the enzyme. Detection can also be accomplished by visual comparison of the extent of enzymatic reaction of a substrate in comparison with similarly prepared standards.

The method of systematic variation of vector elements and analysis of expression is conceptually quite different from previous methods described in the art, and is an aspect of the present invention. These previous methods have used very small numbers of different vector configurations and anecdotal correlations between vectors and expression properties to derive rules for optimal vector design. Such data are very unlikely to provide a basis from which to accurately model the effects of element choices within the vector upon expression. This is because there has been no systematic variation of vector elements, and because elements frequently interact, so that without systematic design in which co-variation of elements is minimized, it is impossible to attribute an effect to a particular element. In contrast the method of the present invention can be performed without assumptions regarding the element preferences of the expression host, or the underlying mechanism of such preference. Instead, the expression system is interrogated with systematically varied sets of sequences and measurements of the expression properties to determine the element configurations that result in desired expression properties. This method may be applied to any expression system as well as to identify an optimal configuration for high expression in multiple systems if a gene transfer vector is to be used in different systems.

An expression property of each of the polynucleotides in the plurality of polynucleotides of the first expression set can be compared to the element configuration in each of the polynucleotides to ascertain a relationship between element configuration and the expression property. Such correlation can also be achieved using pattern classification methods or statistical methods. Examples of pattern classification methods or statistical methods include, but are not limited to linear regression, non-linear regression, logistic regression, multivariate data analysis, classification using a regression tree, partial least squares projection to latent variables, computation of a neural network, computation of a Bayesian model, computation of a generalized additive model, use of a support vector machine, or modeling comprising boosting or adaptive boosting. See, for example, Duda et al., 2001, *Pattern Classification*, Second Edition, John Wiley & Sons, Inc., New York; Hastie, 2003, *The Elements of Statistical Learning*, Springer, New York; and Agresti 1996, *An Introduction to Categorical Data Analysis*, John Wiley & Sons, New York, each of which is hereby incorporated by reference herein for such purpose. Such modeling or correlation

can then be used to assign values for the different elements in the expression system. The design and synthesis of a gene transfer vector variant set and measurement of an expression property of the polynucleotides within a gene transfer vector variant set for the purpose of evaluating different vector element choices within an expression system is an aspect of the present invention.

In some embodiments, the method further comprises the steps of (i) modeling a sequence-activity relationship between (a) one or more substitutions at one or more elements in the gene transfer vector variant set and (b) the expression property measured for all or the portion of the variants in the variant set, and (ii) defining a second gene transfer variant set to comprise variants that include substitutions in the plurality of elements that are selected based on a function of the sequence-activity relationship. In some embodiments the modeling a sequence-activity relationship comprises modeling a plurality of sequence-activity relationships each of which uses only a subset of the available sequence and activity data.

Several methods exist for regression of multivariate data, where predictive relationships between some or all of the independent variables and expression level are determined. Examples of such methods are Partial Least Squares (PLS) and Principal Components Regression (PCR) (Wold et al., 1993, "DNA and peptide sequences and chemical processes multivariately modeled by principal component analysis and partial least-squares projections to latent structures," *Analytica Chimica Acta* 277, 239-253). PLS algorithms, for example, seek to maximize the correlation of the X-data (e.g., codon frequencies) and expression while simultaneously maximizing the X-data variance captured in the model. In doing so, the algorithm determines new orthogonal variables, called latent variables, which are linear combinations of the original variables that best capture the X-data and explain Y variation.

In some embodiments, the sequence-expression relationship has the form:

$$Y=f(w_1x_1, w_2x_2, \dots w_ix_i)$$

where,

Y is a quantitative measure of the expression property;

x_i is a descriptor of a substitution, a combination of substitutions, or a component of one or more substitutions, at one or more positions in the plurality of positions;

w_i is a weight applied to descriptor x_i ; and

$f()$ is a mathematical function.

In some embodiments, the modeling comprises regressing:

$$Y=f(w_1x_1, w_2x_2, \dots w_ix_i).$$

In some instances this regressing comprises linear regression, non-linear regression, logistic regression, or partial least squares projection to latent variables.

A sequence-expression relationship derived from the expression properties of a first expression variant set may be used to design gene transfer vectors to express a second polypeptide with a different amino acid sequence. The use of a sequence-expression relationship to design vectors for the expression of a polypeptide of interest, where the sequence-expression relationship was derived from polynucleotides encoding polypeptides that are not the polypeptide of interest is an aspect of the invention.

In some embodiments, modeling techniques are used to derive sequence-expression relationships. Such modeling techniques include linear and non-linear approaches. Linear and non-linear approaches are differentiated from each other

based on the algebraic relationships used between variables and responses in such approaches. In the system being modeled, the input data (e.g., variables that serve as descriptors of the biopolymer sequence), in turn, can be linearly related to the variables provided or non-linear combinations of the variables. It is therefore possible to perform different combinations of models and data-types: linear input variables can be incorporated into a linear model, non-linear input variables can be incorporated into a linear model and non-linear variables can be incorporated into non-linear models.

In some embodiments, supervised learning techniques are used to identify relationships between vector element configurations in the expression set and measured expression properties. Such supervised learning techniques include, but are not limited to, Bayesian modeling, nonparametric techniques (e.g., Parzen windows, k_n -Nearest-Neighbor algorithms, and fuzzy classification), neural networks (e.g., hopfield network, multilayer neural networks and support vector machines), and machine learning algorithms (e.g., algorithm-independent machine learning). See, for example, Duda et al., *Pattern Classification*, 2nd edition, 2001, John Wiley & Sons, Inc. New York; and Pearl, *Probabilistic Reasoning in Intelligent Systems: Networks of Plausible Inference*, Revised Second Printing, 1988, Morgan Kaufmann, San Francisco. For example, the sequence-expression data can be used to predict the expression property of any sequence given the codon descriptors for a sequence using a neural network. The input for the network is the descriptors and the output is the predicted value of Y. The weights and the activation function can be trained using supervised decision based learning rules. The learning is performed on a subset of variants called the training set and performance of the network is evaluated on a test set.

In some embodiments, unsupervised learning techniques are used to identify relationships between vector element configurations in the expression set and measured expression properties. Such unsupervised learning techniques include, but are not limited to stochastic searches (e.g., simulated annealing, Boltzmann learning, evolutionary methods, principal component analysis, and clustering methods). See, for example, Duda et al., *Pattern Classification*, 2nd edition, 2001, John Wiley & Sons, Inc. New York. For example, the weights in equation B can be adjusted by using monte carlo and genetic algorithms. The optimization of weights for non-linear functions can be complicated and no simple analytical method can provide a good solution in closed form. Genetic algorithms have been successfully used in search spaces of such magnitude. Genetic algorithms and genetic programming techniques can also be used to optimize the function form to best fit the data. For instance, many recombinations of functional forms applied on descriptors of the sequence variants can be applied.

In some embodiments, boosting techniques are used to construct and/or improve models developed using any of the other techniques described herein. A model of the sequence-expression relationship can be described as a functional form whose parameters have been trained for the input data (Y and x_i). Many algorithms/techniques to build models have been described. Algorithms applied on a specific dataset can be weak in that the predictions can be less accurate or “weak” (yielding poor models). Models can be improved using boosting techniques. See, for example, Hastie et al., *The Elements of Statistical Learning*, 2001, Springer, New York. The purpose of boosting is to combine the outputs of many “weak” predictors into a powerful “committee.” In

one embodiment of the invention, boosting is applied using the AdaBoost algorithm. Here, the prediction algorithm is sequentially applied to repeatedly modified versions of the data thereby producing a sequence of models. The predictions from all of these models are combined through a weighted majority vote to produce the final prediction. The data modification at each step consists of applying weights (W_i^b) to each of the i training observations. Initially weights are set to $1/N$, where N is the number of training observation (sequence-activity data). The weights are modified individually in each successive iteration. Training observations that were predicted poorly by a particular model have their weights increased and training observations that were predicted more accurately have their weights decreased. This forces each successive model to concentrate on those training observations that are issued by the previous model. The step of combining the models to produce a “committee” assigns a weight to each model based on the overall prediction error of that model.

The various modeling techniques and algorithms described herein can be adapted to derive relationships between one or more expression properties and the element configuration of a polynucleotide and therefore to make multiple predictions from the same model. Modeling techniques that have been adapted to derive sequence-expression relationships for polynucleotides are within the scope of the present invention. Some of these methods derive linear relationships (for example partial least squares projection to latent structures) and others derive non-linear relationships (for example neural networks). Algorithms that are specialized for mining associations in the data are also useful for designing sequences to be used in the next iteration of sequence space exploration. These modeling techniques can robustly deal with experimental noise in the activity measured for each variant. Often experiments are performed in replicates and for each variant there will be multiple measurement of the same activity. These multiple measurements (replicate values) can be averaged and treated as a single number for every variant while modeling the sequence-expression relationship. The average can be a simple mean or another form of an average such as a geometric or a harmonic mean. In the case of multiple measurements, outliers can be eliminated. In addition, the error estimation for a model derived using any algorithm disclosed herein can incorporate the multiple measurements through calculating the standard deviation of the measurement and comparing the predicted activity from the model with the average and estimate the confidence interval within which the prediction lies. Weights for observations to be used in models can also be derived from the accuracy of measurement, for example, through estimating standard deviation and confidence intervals. This procedure can put less emphasis on variants whose measurements are not accurate. Alternatively, these replicate values can be treated independently.

This will result in duplicating the sequences in the dataset. For example, if sequence variant, represented by descriptor values $\{x_j\}^{i1}$, has been measured in triplicates (Y_{i1}, Y_{i2}, Y_{i3}), the training set for modeling will include descriptor value $\{x_j\}^{i2}$ with activity Y_{i2} and $\{x_j\}^{i3}$ with activity Y_{i3} in addition to $\{x_j\}^{i1}$ with activity Y_{i1} , where $\{x_j\}^{i1} = \{x_j\}^{i2} = \{x_j\}^{i3}$.

The models developed using various algorithms and methods in the previous step can be evaluated by cross validation methods. For example, by randomly leaving data out to build a model and making predictions of data not incorporated into the model is a standard technique for cross validation. In some instances, data may be generated over a period of months. The data can be added incrementally to the

modeling procedure as and when such data becomes available. This can allow for validation of the model with partial or additional datasets, as well as predictions for the properties of gene transfer vector configurations for which activities are still not available. This information may then be used to validate the model.

In one embodiment of the present invention, average values and standard deviations for weight functions can be obtained by omitting a part of the available data. Either individual sequences and their associated expression activities or individual codons can be left out. A sequence-expression relationship can then be constructed from this partial data. This process can be repeated many times, each time the data to leave out is selected randomly. Finally an average and range of values for each weight function is calculated. The weight functions can then also be ranked in order of their importance to activity. The range of values for each weight can provide a measure of the confidence with which the weight is assigned. It can also provide a measure of the importance of the variable in determining the expression property. For example in some instances, the larger the standard deviation for a variable weight, the larger the range of values for that variable that are associated with desirable expression properties.

In some instances the mean value for the variable weight is used to indicate the likely contribution of the element or combination of elements to vector performance. In some instances the mean value for the variable weight minus the standard deviation of the weight is used to indicate the likely contribution of the element or combination of elements to vector performance. In some instances an element or combination of elements is selected if the mean value of the variable weight is above a predetermined value. In some instances an element or combination of elements is selected if the mean value of the weight minus the standard deviation of the weight is above a predetermined value. In some instances the predetermined value is the mean value of all of the variable weights in the model. In some instances the predetermined value is greater than the value of 95% of the variable weights; in some instances the predetermined value is greater than the value of 90% of the variable weights; in some instances the predetermined value is greater than the value of 80% of the variable weights; in some instances the predetermined value is greater than the value of 50% of the variable weights.

In some instances the modeling comprises partial least squares regression and the weight is a regression vector. The regression vector for each variable is used to identify elements that are most favorable for expression in a system. In some instances an element is selected if its regression vector is the highest in the element set, or if it has one of the top 2 or top 3 or top 4 or top 5 or top 6 or top 7 or top 8 or top 9 or top 10 values for regression vectors.

The initial set of data can be small, so models built from it can be inaccurate. Improving the modeled relationship further depends upon obtaining better values for weights whose confidence scores are low. To obtain this data, additional variants designed will provide additional data useful in establishing more precise sequence-expression relationships.

In some embodiments, defining the second variant set comprises adding one or more variants each having an element not present in any variant in the first variant set. In some embodiments, defining the second variant set comprises adding one or more variants each having an element changed in a group not varied in any variant in the first variant set.

Sequence-activity modeling requires an adequate amount of data from variants with statistically distributed element compositions. In some embodiments the first expression set comprises between 5 and 200 gene transfer vectors which each differ from the other members of the set by at least 1 functional element, in preferred embodiments the first expression set comprises at least 10 gene transfer vectors which each differ from the other members of the set by at least 2 functional elements, in some embodiments the first expression set comprises between 10 and 100 gene transfer vectors which each differ from the other members of the set by at least 2 functional elements, in some embodiments the first expression set comprises between 15 and 60 gene transfer vectors which each differ from the other members of the set by at least 2 functional elements.

Examples of such polynucleotide vector sets are shown in Tables 15-18. For two or more functional groups, the polynucleotide vector set is constructed with a first coding polynucleotide that encodes a first expression polypeptide. In some embodiments the first expression polypeptide is a fluorescent protein, or an antibody chain. An expression property is measured for all or a portion of the variants in the variant set. A sequence-activity relationship is modeled between (i) one or more substitutions at one or more elements of the polynucleotide vector set and (ii) the property measured for all or the portion of the variants in the variant set. The variant set is then redefined to comprise variants that include substitutions in the plurality of elements that are selected based on a function of the sequence-activity relationship. In preferred embodiments the variant set comprises between 5 and 200 vector configurations, in preferred embodiments the variant set comprises between 10 and 100 vector configurations, in preferred embodiments the variant set comprises between 15 and 60 vector configurations.

The properties of a biological system including natural as well as non-natural systems with respect to any measured property depends on the interaction between multiple nucleic acid sequence elements, which may be located at positions throughout the polynucleotide. The ability to rationally design a nucleic acid construct with an optimal configuration of elements is advantageous for various applications such as protein synthesis via vector optimization, cell line development and strain engineering. Protein synthesis is a highly dynamic and multi-step process and which plays a central role in synthetic biology, pharmaceutical production and other applications in biotechnology. This importance has led to the development of various parts or genetic control elements able to modulate and precisely control various aspects of protein expression. This capability is not only essential for the successful construction of more complex synthetic biological devices, but also provides tools needed for the tuning of their function for improved performance and reliability.

Many different types of parts capable of controlling transcriptional and translational aspects of the protein synthesis process have been developed. At the transcriptional level, libraries of promoters have been created spanning a wide range of expression levels (Mey et. al., 2007 BMC Biotechnology; Hartner et. al., 2008 Nucleic Acids Research) and efforts have been made to understand potential rules governing promoter structure (Blount et. al., 2012 PLoS One 7; Blazeck et. al., 2013 Biotechnology Journal; Lubliner et. al., 2013 Nucleic Acids Research). At the translational level, libraries of ribosome binding sites (RBSs) have been generated (Mutalik et. al., 2013 Nature Methods) and some rational approaches developed (Salis et.

al., 2009 Nature Biotechnology). Biophysical models of interactions between the ribosome and mRNA have successfully been used to predict relative ribosome initiation strengths and applied in a forward-engineering mode to suggest potential RBS sequences with a desired strength (Salis et. al., 2009 Nature Biotechnology). In addition to RBSs, the speed of translation has been found to be strongly influenced by synonymous codon usage within the gene being expressed. Changes in codon usage have been shown to strongly affect overall expression levels (Welch et. al., 2009 PLoS; Kudla et. al., 2009 Science), influence correct folding of active proteins (Zhang et. al., 2009 Nature Structural and Molecular Biology), and to enable dynamic responses to environmental stresses (Wohlgemuth et. al., 2013 Nucleic Acids Research).

Configurations of polynucleotide vectors with improved expression properties are an aspect of the present invention, including the configurations shown in Tables 6-18.

In preferred embodiments, a gene transfer vector comprises expression elements capable of driving high protein expression, for example a mammalian enhancer selected from amongst the CMV immediate early enhancer (see for example, DQ000968.1 GI:66276969; KF853603.1 GI:576890587), the EF1a enhancer (see for example, J04617.1 GI:181962), the adenoviral major late protein enhancer (see for example, JX173086.1 GI:406679291), the SV40 enhancer (see for example, KM486843.1 GI:731516977; JQ394984.1 GI:41058488); a promoter selected from amongst the EF1a promoter (see for example, J04617.1 GI:181962; AC097023.6 GI:49615137; NM_010106.2 GI:126032328; AY188393.1 GI:30313796) from any mammalian or avian species including but not limited to human, rat, mice, chicken and Chinese hamster, the CMV promoter (see for example, DQ000968.1 GI:66276969; M64943.1 GI:330637), the GAPDH promoter (see for example, J04038.1 GI: 182980) from any mammalian species, the Herpes Simplex Virus thymidine kinase (HSV-TK) promoter (see for example, J04327.1 GI:330219), the actin promoter (see for example, X00182.1 GI:63017) from any mammalian or avian species including but not limited to human, rat, mice, chicken and Chinese hamster, and the ubiquitin promoter (see for example, BC000379.2 GI:33875368); an intron selected from among CMV intron A (see for example, M21295.1 GI:330620), CMV intron B (see for example, M21295.1 GI:330620), CMV intron C (see for example, M21295.1 GI:330620), the EF1a intron (see for example, J04617.1 GI:181962) from any mammalian or avian species including but not limited to human, rat, mice, chicken and Chinese hamster, the actin intron (see for example, X00182.1 GI:63017) from any mammalian or avian species including but not limited to human, rat, mice, chicken and Chinese hamster, the GAPDH intron (see for example, J04038.1 GI: 182980) from any mammalian or avian species including but not limited to human, rat, mice, chicken and Chinese hamster, the adenoviral major late protein intron (see for example, U89672.1 GI:1899166), the PGK promoter (see for example, KC710227.1 GI:501416041) from any mammalian or avian species including but not limited to human, rat, mice, chicken and Chinese hamster; 5' untranslated regions (5' UTRs) from any mammalian or avian species including but not limited to human, rat, mice, chicken and Chinese hamster.

In preferred embodiments the gene transfer vector may comprise selectable markers to enable selection of cells with stably integrated transposons. Examples include genes that confer resistance to puromycin, neomycin, hygromycin,

blasticidin and zeocin. The gene transfer vector may comprise a bacterial resistance marker and a bacterial origin of replication to facilitate manipulation in prokaryotic cells. These prokaryotic elements are preferably contained within the non-transposable portion of the vector.

In preferred embodiments gene transfer vector may comprise other sequence elements that enhance expression of the genes that they encode. Examples include elements that are believed to enhance RNA processing and nuclear export such as woodchuck hepatitis post-transcriptional regulatory element (WPRE), hepatitis B virus post-transcriptional regulatory element (HPRE) (for example but not limited to SEQ ID NO: 104-105) and arctic ground squirrel post-transcriptional regulatory element (AGS) (for example but not limited to SEQ ID NOS: 106-107). Examples also include polyadenylation sequences such as the polyadenylation sequences from BGH (bovine growth hormone) (see for example, M57764.1 GI:163091; KF992215.1 GI:593024220), HGH (human growth hormone) (see for example, M13438.1 GI:183156), the polyadenylation signals from human (see for example, X03145.1 GI:34173) or rabbit (see for example, NM_001082260.2 GI: 129270172; EF186084.1 GI: 122893039) beta globin, viral polyadenylation signals including those from SV40 (see for example, AY122060.1 GI:22001016) or herpes simplex virus (see for example, M38699.1 GI:330309) and terminator sequences from gastrin. Examples also include sequences that are thought to act as insulators by preventing the spread of heterochromatin or promoter interference such as but not limited to the HS4 (SEQ ID NOS: 112) and HS4 core (SEQ ID NOS: 113). In some preferred embodiments, a pair of insulators surround the expressible sequences. Examples also include sequences believed to mediate attachment to the chromatin scaffold such as but not limited to SEQ ID NOS: 108-111. Regardless of their actual mechanism, incorporation of expression enhancing elements into gene transfer vectors and transposons is expressly contemplated. In preferred embodiments, gene transfer vectors comprise transposons.

In some embodiments, the nucleic acid construct is a vector with enhanced expression and integration properties. For example, an optimal configuration of vector elements for enhanced transient expression as well as more efficient stable integration and expression was identified by the methods described herein. A mammalian vector construct variant set was generated using multiple combinations of various transposon ends, insulators, enhancers, promoters, 5' untranslated regions (UTRs), 3' untranslated regions (UTRs), RNA export modulating sequence, polyadenylation sequences, terminators, matrix attachment element and transposases. The mammalian vector variant set was tested for optimal DasherGFP expression in Human embryonic kidney (HEK 293) cells to identify an optimized vector construct. Other optimized vector constructs were identified with optimal configurations of elements listed above that show high expression of DasherGFP in HEK 293 and CHO cell lines as shown in Tables 6-20. Vector configurations with different promoter combinations were also shown to affect DasherGFP expression. Further optimization of this vector construct for different cell lines using the methods described herein is expressly contemplated. An advantage of the methods described herein is to quickly identify a subset of sequence elements most likely to influence desired activity as well as to facilitate predictable construction of optimal configuration of elements.

In some embodiments, two promoters are placed in opposite orientation, each driving an expression cassette such that

transcription from the two promoters diverges. Such a configuration greatly improves expression of transcripts from the expression cassettes.

In some embodiments, elements that are useful in enhancing performance may include those localized to the genomic DNA of a cell. For example expression may be influenced by the levels of RNA polymerases, chaperonins, proteases, processing enzymes, or other factor encoded by DNA on the cell chromosome. It might also be advantageous to augment the host chromosome with functional elements that influence performance. In some embodiments, a variable for engineering is the site at which a functional gene is integrated into a host cell chromosome.

In some embodiments, the nucleic acid construct is a polynucleotide comprising elements or combinations of elements arranged in an optimal configuration. In some embodiments, the polynucleotide is linear. In some embodiments, the elements in a nucleic acid construct comprise functional genetic features, for example, promoters, enhancers, introns, polyadenylation signals, origins of replication, and terminators. In some embodiments, the elements in a nucleic acid construct comprise protein-encoding elements such as secretion signals, resistance markers, anchoring peptides, localization signals, and fusion tags. In some embodiments, the plurality of elements comprises three or more elements, six or more elements, between three and twenty elements or between three and hundred elements. In some embodiments, nucleic acid construct variants include substitutions in a single element comprising one or more positions, three or more positions, six or more positions. In some embodiments, nucleic acid variants include substitutions wherein the substitutions are variations in elements and/or presence or absence of elements. In some embodiments, the substitutions include changes in the position of one or more elements. In some embodiments, nucleic acid variants include a change in the order of one or more elements.

An important aspect of the present invention is that it enables the assessment of the performance of different types of elements: those that affect transcription, those that affect RNA processing, those that affect RNA export from the nucleus of the cell, those that affect integration into the host genome, those that affect replication within the host cell, those that affect translational initiation and those that affect translational elongation. The present invention allows sets of polynucleotide constructs to be designed to test the interactions of these types of elements.

In some embodiments, the configuration of sequence elements in the transposon will result in highly efficient integration into the genome of the target cell. In these instances addition of the transposase may provide only a small improvement in the expression of genes on the transposon, or no improvement at all. The present invention expressly contemplates that under some circumstances, the configuration of sequence elements within the transposon will be sufficient so that the gene transfer system will not need to include the transposase.

5.2.7 Use of Coupling Elements in a Gene Transfer System

Messenger RNA molecules in eukaryotic cells are generally monocistronic, that is, they usually encode a single polypeptide. This is because translation in eukaryotes generally occurs by a process in which the ribosome binds to a structure at the 5' end of the mRNA and then "scans" down the mRNA until it finds an initiation codon (generally AUG) where it begins translation. It then translates the mRNA, producing the encoded polypeptide, until it reaches a termination codon (generally UAA, UAG or UGA) which causes

the ribosome to end translation and dissociate from the mRNA. Certain eukaryotic viruses have evolved mechanisms by which they can express more than one polypeptide from a single mRNA. These include internal ribosome entry sites (IRES), and cis-acting hydrolase element (CHYSEL) sequences.

An IRES provides a structure to which the ribosome can bind that does not need to be at the 5' end of the mRNA. It can therefore direct a ribosome to initiate translation at a second initiation codon within an mRNA, allowing more than 1 polypeptide to be produced from a single mRNA. A CHYSEL sequence causes a translating eukaryotic ribosome to release the growing polypeptide chain that it is synthesizing without dissociating from the mRNA. The ribosome continues translating, and therefore produces a second polypeptide. A single genetic construct can contain more than one IRES or CHYSEL sequence, and it can contain both IRES and CHYSEL sequences, so can therefore encode 2 or 3 or 4 or 5 or 6 or more than 6 polypeptides on a single mRNA.

IRES or CHYSEL sequences can therefore be used as coupling elements, to link the expression of two or more polypeptides. For example the expression of a first polypeptide may be linked to the expression of a selectable protein that provides a physical, chemical or biological method for selecting cells on the basis of how much of the selectable protein is expressed. The use of certain selectable proteins to indicate the status or functionality of a genetic construct within an organism is an aspect of the invention. The combining of selectable proteins with IRES or CHYSEL sites to indicate the status or functionality of a polynucleotide, or to indicate the level of expression of a polynucleotide or polypeptide is another aspect of the invention. IRES sequences are used to simultaneously express two or more proteins from a single promoter.

Another important application of translational coupling sequences, particularly IRES sequences, is in allowing the co-expression of two polypeptide chains that function together, either to catalyze different steps in a metabolic pathway, or as parts of the same molecule. A particularly important example is in the formation of a human antibody; a full human antibody consists of two heavy and two light chains. For antibody production, it is desirable for the heavy and light chains to be expressed in an optimal ratio. Monoclonal antibodies (Mab) are heterotetramers consisting of an equimolar ratio of heavy chain (HC) to light chain (LC) genes encoded on either one or two plasmids. Although the chains are present at equimolar amounts ratio in the final antibody molecule, higher amounts of antibody are generally expressed if the light chain is more highly expressed than the heavy chain. Further, although the optimal ratio may be as high as 5× as much light chain as heavy chain, the exact ratio that gives most assembled antibody tetramer depends on the exact antibody being expressed.

Optimization of the ratio of heavy and light chain is typically accomplished in one of two ways. In the first, polynucleotides encoding the heavy chain and the light chain are carried on two different plasmids and co-transfected into a host cell. In the second, a single plasmid carries both polynucleotides (encoding the heavy chain and the light chain), each with its own promoter and polyadenylation sequence. In the case of co-transfection, individual cells take up different numbers of each plasmid. Because it is only possible to control the average number of each plasmid taken up by each cell, many cells do not end up expressing the optimal ratio of heavy and light chain. This problem is amplified in the case of stable cell lines, because there is the

further variable of integration location which also affects expression levels. Dual promoter constructs overcome these difficulties, and can often be effective. However they can become large and genetically unstable because of repeated sequences: promoters, enhancers, polyadenylation sequences, RNA export sequences such as WPRE and HPRE and matrix attachment regions may all need to be duplicated. This can compromise transfection efficiency and performance; there can also be interference between two eukaryotic promoters in the same construct.

These drawbacks may be overcome by using IRES sequences, providing a set of IRES sequences are available which can produce different levels of expression of the second polypeptide relative to the first. This allows the equivalent of the titration that is currently accomplished by co-transfecting different amounts of the two plasmids. However it has the significant advantage that each cell gets an equal number of copies of the polynucleotides encoding the first polypeptide and the second polypeptide (because they are on the same plasmid), even though different cells may get different numbers of plasmids. Thus even though the amount of each polypeptide may vary from cell to cell, the ratio of the amount of the first polypeptide to the second polypeptide should be much less variable.

The most commonly used IRES in mammalian systems is that from Encephalomyocarditis virus, which includes four amino acids of the N-terminus of the second open reading frame (MATT). There are thus two significant limitations in the currently available tools for co-expression of multiple genes in eukaryotic cells. Firstly, there is no readily available set of IRES sequences that give a known range of ratios of expression between the first and second open reading frame, so it is difficult to control the relative expression of two proteins. Secondly, the most commonly used IRES sequence requires an N-terminal extension of the second protein, which may compromise or modify the function of that protein. There is thus a need in the art for a set of characterized sequences that can be easily incorporated between a first and a second open reading frame to create an optimal expression balance for a downstream function or product of the cell.

We synthesized sequences inspired by the 5' untranslated regions of positive strand RNA viruses, cloned them under control of a single promoter into a vector between a polynucleotide encoding a green fluorescent protein and a polynucleotide encoding a red fluorescent protein, transfected the construct into mammalian cells and measured the expression of red and green fluorescence. Using this test we identified IRES sequences that function in human embryonic kidney (HEK) and Chinese hamster ovary (CHO) cells (SEQ ID NOS: 58-100) and that show different expression levels of the second (IRES-controlled) protein relative to the first. This is of particular importance since Chinese hamster ovary (CHO) cells are the dominant host for industrial monoclonal antibody production because of their capacity for proper protein folding, assembly and appropriate post-translational modifications. Each of these active sequences can be used to search sequence databases for similar sequences, and similar sequences can in turn be tested using the same system. In preferred embodiments, the IRES sequence is selected from bat picornavirus sequences or rodent picornavirus sequences. Very similar sequences have quite large differences in function. One method to improve the function of individual IRESes is to create consensus sequences. Another method is to identify sequences from viral strains that are reported to be most virulent. Another method is to create

libraries of variant sequences and to test members of these libraries using a pair of fluorescent reporter proteins.

In some embodiments, the IRES sequence has at least 80% similarity to SEQ ID NOS: 58-100, or is a chimera of two or more of these sequences. In some other embodiments, the IRES sequence has at least 90% similarity to SEQ ID NOS: 58-100. In some embodiments, the nucleic acid encodes IRES sequences, wherein the nucleic acid has at least at least 80%, at least 90% at least 95%, at least 98% or at least 99% sequence identity to a consensus sequence derived from a set of naturally occurring sequences of the 5' untranslated region of RNA viral genomes; in some embodiments the consensus sequence is not itself identical to any naturally occurring sequence. In some embodiments, an IRES sequence has at least 50%, at least 60%, at least 70%, at least 80%, at least 90% similarity to 5' untranslated regions (UTRs) of the picornavirus family of viruses. In some embodiments, the IRES elements of the invention are incorporated into an expression vector with a single promoter and one or more IRES elements that allow control of the expression ratios of one or more genes. In other embodiments, IRES elements that function as enhancers are incorporated into an expression vector wherein the IRES elements function to enhance expression of one or multiple genes.

Use of IRES sequences to control expression levels of two or more proteins in a gene transfer vector comprising transposons for stable integration with or without transposases identified herein, with particular emphasis on expressing antibodies in Chinese hamster ovary cells (CHO) or Human embryonic kidney (HEK) cells is another important aspect of this invention. For example, expression of heavy and light chains of an antibody can be controlled by selecting an IRES with the appropriate strength.

Another aspect of the invention is the use of IRES sequences in transient expression vectors. Use of IRES elements to control expression levels of two or more proteins in expression vectors comprising elements or combinations of elements arranged in an optimal configuration as described herein (Section 5.2.6) above is another embodiment of the invention. Additional embodiments include use of IRES sequences to control expression levels of two or more polypeptides in expression vectors (with transposons for stable integration and without for transient expression) with viral replication sequences to increase copy number of the plasmid. Use of IRES sequences in any of the vector configurations shown in any of the Tables here shown is an aspect of the invention.

Ratios of expressed gene(s) can be controlled using the various IRES elements identified. This is particularly useful when the gene products are to be expressed in a fixed ratio to get optimal results.

In some preferred embodiments, IRES sequences are selected from SEQ ID NOS: 58-100.

In some embodiments, it is advantageous to include secretory peptides at the amino-terminus of a protein that enables translocation of the protein to the endoplasmic reticulum (ER). This helps not only in the ease of purification but also allows proper folding of complex disulfide bonds and glycosylation. Proper selection of a signal peptide can have dramatic consequences on protein over-expression. A number of efficient secretion peptides have been described, for example interleukin-2, CD5, the immunoglobulin kappa light chain, trypsinogen, serum albumin and prolactin. We have identified secretion peptides (SEQ ID NOS: 114-115) that function well in combination with IRES

sequences. This is especially important for expression of antibodies in a single construct using IRES elements as described herein.

A kit comprising a single expression vector with one or more IRES elements and reagents to facilitate cloning of ORFs into the vector is another aspect of the invention. The kit can additionally include a set of IRES elements as templates, such that the IRES elements can be incorporated into an expression vector of choice.

5.2.8 Additional Applications of the Gene Transfer System

Using the gene transfer system for methods such as gene discovery and/or gene tagging, permits, for example, identification, isolation, and characterization of genes involved with growth and development through the use of transposons as insertional mutagens or identification, isolation and characterization of transcriptional regulatory sequences controlling growth and development.

The gene transfer system of the invention represents a considerable refinement of non-viral DNA-mediated gene transfer. For example, adapting viruses as agents for gene therapy restricts genetic design to the constraints of that virus genome in terms of size, structure and regulation of expression. Non-viral vectors, as described herein, are generated largely from synthetic starting materials and are therefore more easily manufactured than viral vectors. Non-viral reagents are less likely to be immunogenic than viral agents making repeat administration possible. Non-viral vectors are more stable than viral vectors and therefore better suited for pharmaceutical formulation and application than are viral vectors. Additionally, the gene transfer system of the present invention is a non-viral gene transfer system that facilitates insertion into DNA and markedly improves the frequency of stable gene transfer.

An efficient method for using transposon-transposase combinations of the present invention to stably introduce a chimeric antigen receptor (CAR) to redirect the specificity of human T-cells is expressly contemplated and is an important aspect of the invention. For example, redirecting the specificity of T cells for B-lineage antigens and advanced B-cell malignancies by infusion of such tumor-specific T cells (adoptive cell transfer) modified by the transposon-transposase of the present invention is another embodiment. Combining cell-based therapies with gene-based therapies, in which genetically engineered chimeric antigen receptors (CARs) or tumor-specific T-cell receptor genes are expressed in immune effector cells has enormous therapeutic potential. CARs combine intracellular signaling domains with a single-chain variable fragment of an antibody (Ab) into a single chimeric protein. Engineering immune cells (T-cells) to recognize and attack their tumors is a powerful approach especially in treating B-cell malignancies or lymphomas. A chimeric antigen receptor (CAR) recognizes cell-surface tumor-associated antigen independent of human leukocyte antigen (HLA) and employs one or more signaling molecules to activate genetically modified T cells for killing, proliferation, and cytokine production. For example, targeting CD19 has been achieved through the enforced expression of a CAR that recognizes CD19 independent of HLA. In contrast to methods that genetically modify T cells using recombinant retrovirus, a non-viral gene transfer approach using the transposon-transposase system to enforce expression of the introduced CAR is a viable alternative and avoids some of the issues such as preferential integration sites associated with most viruses. To improve therapeutic potential, CAR signaling through CD28 and CD3- to sustain T-cell proliferation and recycle effector functions in vivo is also contemplated.

The present invention further provides an efficient method for producing transgenic animals, including the step of applying the gene transfer system of the present invention to an animal. Transgenic DNA has not been efficiently inserted into chromosomes. Only about one in a million of the foreign DNA molecules are inserted into the cellular genome, generally several cleavage cycles into development. Consequently, most transgenic animals are mosaic (Hackett et al. 'The molecular biology of transgenic fish'; Biochemistry and Molecular Biology of Fishes ((Hochachka & Mommsen, eds.) Vol. 2, pp. 207-240, 1993). As a result, animals raised from embryos into which transgenic DNA has been delivered must be cultured until gametes can be assayed for the presence of inserted foreign DNA. Many transgenic animals fail to express the transgene due to position effects. A simple, reliable procedure that directs early insertion of exogenous DNA into the chromosomes of animals at the one-cell stage is needed. The present system helps to fill this need.

In certain preferred embodiments, the gene transfer system of this invention can readily be used to produce transgenic animals that carry a particular marker or express a particular protein in one or more cells of the animal. Generally, methods for producing transgenic animals are known in the art and incorporation of the gene transfer system of the present invention into these techniques does not require undue experimentation, e.g. there are a variety of methods for producing transgenic animals for research or for protein production including, but not limited to Hackett et al. (1993, *supra*). Other methods for producing transgenic animals are described in the art (e.g. M. Markkula et al. *Rev. Reprod.*, 1, 97-106 (1996); R. T. Wall et al., *J. Dairy Sci.*, 80, 2213-2224 (1997)), J. C. Dalton, et al. (*Adv. Exp. Med. Biol.*, 411, 419-428 (1997)) and H. Lubon et al. (*Transfus. Med. Rev.*, 10, 131-143 (1996)). A transposon including one or more protein encoding nucleic acids to be expressed in the transgenic animal flanked by ITRs can be introduced into a suitable cells, for example, a zygote, embryonic stem cell or adult cell for nuclear transfer together with a transposase, either in protein form or encoded by the same or different nucleic acid than the transposon. The transposon integrates into the genome of the cell. The cell is then propagated to an embryo, and then into a transgenic animal as is conventional transgenesis.

In another embodiment, the present invention features a transgenic animal produced by the methods described herein, preferably by using the gene transfer system presently described. For example, transgenic animals may preferably contain a nucleic acid sequence inserted into the genome of the animal by the gene transfer system, thereby enabling the transgenic animal to produce its gene product, for example, a protein. In transgenic animals this protein is preferably a product for isolation from a cell, for example the inventive protein can be produced in quantity in milk, urine, blood or eggs. Promoters can be used that promote expression in milk, urine, blood or eggs and these promoters include, but are not limited to, casein promoter, the mouse urinary protein promoter, beta-globin promoter and the ovalbumin promoter respectively. Recombinant growth hormone, recombinant insulin, and a variety of other recombinant proteins have been produced using other methods for producing protein in a cell. Nucleic acids encoding these or other proteins can be inserted into the transposon of this invention and transfected into a cell. Expression from a transposon of the present invention can be improved when a transposase protein is present to catalyze integration of the transposon into the DNA of a cell. Where the cell is part of

a tissue or part of transgenic animal, large amounts of recombinant protein can be obtained. Transgenic animals may be selected from vertebrates and invertebrates, for example, fish, birds, mammals including, but not limited to, rodents, such as rats or mice, ungulates, such as cows or goats, sheep, swine or humans.

The present invention furthermore provides a method for gene therapy comprising the step of introducing the gene transfer system into cells as described herein. Therefore, the transposon as described herein preferably comprises a gene to provide a gene therapy to a cell or an organism. Preferably, the gene is placed under the control of a tissue specific promoter or of a ubiquitous promoter or one or more other expression control regions for the expression of a gene in a cell in need of that gene. Presently, a variety of genes are being tested for a variety of gene therapies including, but not limited to, the CFTR gene for cystic fibrosis, adenosine deaminase (ADA) for immune system disorders, factor IX and interleukin-2 (IL-2) for blood cell diseases, alpha-1-antitrypsin for lung disease, and tumor necrosis factors (TNFs) and multiple drug resistance (MDR) proteins for cancer therapies. These and a variety of human or animal specific gene sequences including gene sequences to encode marker proteins and a variety of recombinant proteins are available in the known gene databases such as GenBank.

An advantage of the gene transfer system of the present invention for gene therapy purposes is that it is limited to a much lesser extent by the size of the polynucleotide between the transposon ends than is the case for many other gene transfer systems. There is no known limit on the size of the nucleic acid sequence that can be inserted into DNA of a cell using the transposase proteins of the present invention. In particular preferred embodiments, for gene therapy purposes, but also for other inventive purposes, the gene transfer system may be transfected into cells by a variety of methods including by microinjection, lipid-mediated strategies or by viral-mediated strategies. For example, where microinjection is used, there is very little restraint on the size of the intervening sequence of the transposon of this invention. Similarly, lipid-mediated strategies do not have substantial size limitations. However, other strategies for introducing the gene transfer system into a cell, such as viral-mediated strategies could limit the length of the nucleic acid sequence positioned between the repeats.

Accordingly, in certain exemplary embodiments, the gene transfer system as described herein can be delivered to cells via viruses, including retroviruses (such as lentiviruses), adenoviruses, adeno-associated viruses, herpes viruses, and others. There are several potential combinations of delivery mechanisms that are possible for the transposon portion containing the heterologous polynucleotide flanked by the terminal repeats and the gene encoding the transposase. For example, both the transposon and the transposase gene can be contained together on the same recombinant viral genome; a single infection delivers both parts of the gene transfer system such that expression of the transposase then directs cleavage of the transposon from the recombinant viral genome for subsequent insertion into a cellular chromosome. In another example, the transposase and the transposon can be delivered separately by a combination of viruses and/or non-viral systems such as lipid-containing reagents. In these cases either the transposon and/or the transposase gene can be delivered by a recombinant virus. In every case, the expressed transposase gene directs liberation of the transposon from its carrier DNA (viral genome) for insertion into chromosomal DNA. In certain preferred embodiments of the present invention, transposons may be

utilized for insertional mutagenesis, preferably followed by identification of the mutated gene. DNA transposons, particularly the transposons, have several advantages compared to approaches in the prior art, for example, with respect to viral and retroviral methods. For example, unlike proviral insertions, transposon insertions can be remobilized by supplying the transposase activity in trans. Thus, instead of performing time-consuming microinjections, it is possible according to the present invention to generate transposon insertions at new loci by crossing stocks transgenic for the above mentioned two components of the transposon system, the transposon and the transposase. In a preferred embodiment the gene transfer system is directed to the germline of the experimental animals to mutagenize germ cells. Alternatively, transposase expression can be directed to particular tissues or organs by using a variety of specific promoters. In addition, remobilization of a mutagenic transposon out of its insertion site can be used to isolate revertants and, if transposon excision is associated with a deletion of flanking DNA, the gene transfer system of the present invention may be used to generate deletion mutations. Furthermore, since transposons are composed of DNA, and can be maintained in simple plasmids, gene transfer systems and transposons of the present invention are much safer and easier to work with than highly infectious retroviruses. The transposase activity can be supplied in the form of DNA, mRNA or protein as defined above in the desired experimental phase.

In another embodiment, the present invention also provides an efficient system for gene discovery, for example genome mapping, by introducing a transposon as defined above into a gene using a gene transfer system as described in the present invention. In one example, the transposon in combination with the transposase protein or a nucleic acid encoding the transposase protein is transfected into a cell. In certain preferred embodiments, the transposon preferably comprises a nucleic acid sequence positioned between at least two repeats, wherein the repeats bind to transposase protein and wherein the transposon is inserted into the DNA of the cell in the presence of the transposase protein. In certain preferred embodiments, the nucleic acid sequence includes a marker protein, such as GFP and a restriction endonuclease recognition site. Following insertion, the cell DNA is isolated and digested with the restriction endonuclease. For example, if the endonuclease recognition site is a 6-base recognition site and a restriction endonuclease is used that employs a 6-base recognition sequence, the cell DNA is cut into about 4000-bp fragments on average. These fragments can be either cloned or linkers can be added to the ends of the digested fragments to provide complementary sequence for PCR primers. Where linkers are added, PCR reactions are used to amplify fragments using primers from the linkers and primers binding to the direct repeats of the repeats in the transposon. The amplified fragments are then sequenced and the DNA flanking the direct repeats is used to search computer databases such as GenBank.

In another exemplary embodiment of the present invention, the invention provides a method for mobilizing a nucleic acid sequence in a cell. According to this method the *Bombyx mori* or *Xenopus tropicalis* transposon is inserted into DNA of a cell, as described herein. A protein or nucleic acid encoding the *Bombyx mori* or *Xenopus tropicalis* transposase protein is transfected into the cell and the protein is able to mobilize (i.e. move) the transposon from a first position within the DNA of the cell to a second position within the DNA of the cell. The DNA of the cell is preferably genomic DNA or extrachromosomal DNA. The inventive method allows movement of the transposon from one loca-

tion in the genome to another location in the genome, or for example, from a plasmid in a cell to the genome of that cell.

In other embodiments, the gene transfer system can also be used as part of a method involving RNA-interference techniques. RNA interference (RNAi), is a technique in which exogenous, double-stranded RNAs (dsRNAs), being complementary to mRNA's or genes/gene fragments of the cell, are introduced into this cell to specifically bind to a particular mRNA and/or a gene and thereby diminishing or abolishing gene expression. The technique has proven effective in *Drosophila*, *Caenorhabditis elegans*, plants, and recently, in mammalian cell cultures. To apply this technique in context with the present invention, the transposon preferably contains short hairpin expression cassettes encoding small interfering RNAs (siRNAs), which are complementary to mRNA's and/or genes/gene fragments of the cell. These siRNAs have preferably a length of 20 to 30 nucleic acids, more preferably a length of 20 to 25 nucleic acids and most preferably a length of 21 to 23 nucleic acids. The siRNA may be directed to any mRNA and/or a gene, that encodes any protein as defined above, e.g. an oncogene. This use, particularly the use of transposons for integration of siRNA vectors into the host genome provides a long-term expression of siRNA in vitro or in vivo and thus enables a long-term silencing of specific gene products.

5.2.9 Pharmaceutical Compositions

The present invention further includes pharmaceutical compositions containing either i) a *Bombyx mori* transposase as a protein or encoded by a nucleic acid, and/or a *Bombyx mori* transposon, or a gene transfer system comprising a *Bombyx mori* transposase as a protein or encoded by a nucleic acid, in combination with an active *Bombyx mori* transposon or; ii) a *Xenopus tropicalis* transposase as a protein or encoded by a nucleic acid, and/or a *Xenopus tropicalis* transposon, or a gene transfer system comprising a *Xenopus tropicalis* transposase as a protein or encoded by a nucleic acid, in combination with an active *Xenopus tropicalis* transposon.

The pharmaceutical composition may optionally be provided together with a pharmaceutically acceptable carrier, adjuvant or vehicle. In this context, a pharmaceutically acceptable carrier, adjuvant or vehicle according to the invention refers to a non-toxic carrier, adjuvant or vehicle that does not destroy the pharmacological activity of the component(s) with which it is formulated.

Pharmaceutically acceptable carriers, adjuvants or vehicles that may be used in the compositions of this invention include, but are not limited to, ion exchangers, alumina, aluminum stearate, lecithin, serum proteins, such as human serum albumin, buffer substances such as phosphates, glycine, sorbic acid, potassium sorbate, partial glyceride mixtures of saturated vegetable fatty acids, water, salts or electrolytes, such as protamine sulfate, disodium hydrogen sulfate, disodium hydrogen phosphate, potassium hydrogen phosphate, sodium chloride, zinc salts, colloidal silica, magnesium trisilicate, polyvinyl pyrrolidone, cellulose-based substances, polyethylene glycol, sodium carboxymethyl cellulose, polyacrylates, waxes, polyethylene-polyoxypropylene-block polymers, polyethylene glycol and wool fat.

The pharmaceutical compositions of the present invention may be administered orally, parenterally, by inhalation spray, topically, rectally, nasally, buccally, vaginally or via an implanted reservoir.

The term parenteral as used herein includes subcutaneous, intravenous, intramuscular, intra-articular, intra-synovial, intrasternal, intrathecal, intrahepatic, intralesional and

intracranial injection or infusion techniques. Preferably, the pharmaceutical compositions are administered orally, intraperitoneally or intravenously. Sterile injectable forms of the pharmaceutical compositions of this invention may be aqueous or oleaginous suspension. These suspensions may be formulated according to techniques known in the art using suitable dispersing or wetting agents and suspending agents. The sterile injectable preparation may also be a sterile injectable solution or suspension in a non-toxic parenterally-acceptable diluent or solvent, for example as a solution in 1,3-butanediol. Among the acceptable vehicles and solvents that may be employed are water, Ringer's solution and isotonic sodium chloride solution. In addition, sterile, fixed oils are conventionally employed as a solvent or suspending medium.

For this purpose, any bland fixed oil may be employed including synthetic mono- or di-glycerides. Fatty acids, such as oleic acid and its glyceride derivatives are useful in the preparation of injectables, as are natural pharmaceutically-acceptable oils, such as olive oil or castor oil, especially in their polyoxyethylated versions. These oil solutions or suspensions may also contain a long-chain alcohol diluent or dispersant, such as carboxymethyl cellulose or similar dispersing agents that are commonly used in the formulation of pharmaceutically acceptable dosage forms including emulsions and suspensions. Other commonly used surfactants, such as Tweens, Spans and other emulsifying agents or bioavailability enhancers which are commonly used in the manufacture of pharmaceutically acceptable solid, liquid, or other dosage forms may also be used for the purposes of formulation.

The pharmaceutically acceptable compositions of this invention may be orally administered in any orally acceptable dosage form including, but not limited to, capsules, tablets, aqueous suspensions or solutions. In the case of tablets for oral use, carriers commonly used include lactose and corn starch. Lubricating agents, such as magnesium stearate, are also typically added. For oral administration in a capsule form, useful diluents include lactose and dried cornstarch. When aqueous suspensions are required for oral use, the active ingredient is combined with emulsifying and suspending agents. If desired, certain sweetening, flavoring or coloring agents may also be added.

Alternatively, the pharmaceutically acceptable compositions of this invention may be administered in the form of suppositories for rectal administration. These can be prepared by mixing the gene transfer system or components thereof with a suitable non-irritating excipient that is solid at room temperature but liquid at rectal temperature and therefore will melt in the rectum to release the drug. Such materials include cocoa butter, beeswax and polyethylene glycols. The pharmaceutically acceptable compositions of this invention may also be administered topically, especially when the target of treatment includes areas or organs readily accessible by topical application, including diseases of the eye, the skin, or the lower intestinal tract. Suitable topical formulations are readily prepared for each of these areas or organs.

For topical applications, the pharmaceutically acceptable compositions may be formulated in a suitable ointment containing the gene transfer system or components thereof suspended or dissolved in one or more carriers. Carriers for topical administration of the components of this invention include, but are not limited to, mineral oil, liquid petrolatum, white petrolatum, propylene glycol, polyoxyethylene, polyoxypropylene component, emulsifying wax and water. Alternatively, the pharmaceutically acceptable compositions

can be formulated in a suitable lotion or cream containing the active components suspended or dissolved in one or more pharmaceutically acceptable carriers. Suitable carriers include, but are not limited to, mineral oil, sorbitan monostearate, polysorbate 60, cetyl esters wax, cetaryl alcohol, 2-octyldodecanol, benzyl alcohol and water.

For ophthalmic use, the pharmaceutically acceptable compositions may be formulated as micronized suspensions in isotonic, pH adjusted sterile saline, or, preferably, as solutions in isotonic, pH adjusted sterile saline, either with or without a preservative such as benzylalkonium chloride. Alternatively, for ophthalmic uses, the pharmaceutically acceptable compositions may be formulated in an ointment such as petrolatum.

The pharmaceutically acceptable compositions of this invention may also be administered by nasal aerosol or inhalation. Such compositions are prepared according to techniques well-known in the art of pharmaceutical formulation and may be prepared as solutions in saline, employing benzyl alcohol or other suitable preservatives, absorption promoters to enhance bioavailability, fluorocarbons, and/or other conventional solubilizing or dispersing agents.

The amount of the components of the present invention that may be combined with the carrier materials to produce a composition in a single dosage form will vary depending upon the host treated, the particular mode of administration. It has to be noted that a specific dosage and treatment regimen for any particular patient will depend upon a variety of factors, including the activity of the specific component employed, the age, body weight, general health, sex, diet, time of administration, rate of excretion, drug combination, and the judgment of the treating physician and the severity of the particular disease being treated. The amount of a component of the present invention in the composition will also depend upon the particular component(s) in the composition. The pharmaceutical composition is preferably suitable for the treatment of diseases, particular diseases caused by gene defects such as cystic fibrosis, hypercholesterolemia, hemophilia, immune deficiencies including HIV, Huntington disease, .alpha.-anti-Trypsin deficiency, as well as cancer selected from colon cancer, melanomas, kidney cancer, lymphoma, acute myeloid leukemia (AML), acute lymphoid leukemia (ALL), chronic myeloid leukemia (CML), chronic lymphocytic leukemia (CLL), gastrointestinal tumors, lung cancer, gliomas, thyroid cancer, mamma carcinomas, prostate tumors, hepatomas, diverse virus-induced tumors such as e.g. papilloma virus induced carcinomas (e.g. cervix carcinoma), adeno carcinomas, herpes virus induced tumors (e.g. Burkitt's lymphoma, EBV induced B cell lymphoma), Hepatitis B induced tumors (Hepato cell carcinomas), HTLV-I und HTLV-2 induced lymphoma, lung cancer, pharyngeal cancer, anal carcinoma, glioblastoma, lymphoma, rectum carcinoma, astrocytoma, brain tumors, stomach cancer, retinoblastoma, basalioma, brain metastases, medullo blastoma, vaginal cancer, pancreatic cancer, testis cancer, melanoma, bladder cancer, Hodgkin syndrome, meningeoma, Schneeberger's disease, bronchial carcinoma, pituitary cancer, mycosis fungoides, gullet cancer, breast cancer, neurinoma, spinalioma, Burkitt's lymphoma, laryngeal cancer, thymoma, corpus carcinoma, bone cancer, non-Hodgkin lymphoma, urethra cancer, CUP-syndrome, oligodendroglioma, vulva cancer, intestinal cancer, oesphagus carcinoma, small intestine tumors, craniopharyngeoma, ovarian carcinoma, ovarian cancer, liver cancer, leukemia, or cancers of the skin or the eye; and more.

5.3 Kits

The present invention also features kits comprising a *Bombyx mori* transposase as a protein or encoded by a

nucleic acid, and/or a *Bombyx mori* transposon; or a gene transfer system as described herein comprising a *Bombyx mori* transposase as a protein or encoded by a nucleic acid as described herein, in combination with a *Bombyx mori* transposon; optionally together with a pharmaceutically acceptable carrier, adjuvant or vehicle, and optionally with instructions for use. Any of the components of the inventive kit may be administered and/or transfected into cells in a subsequent order or in parallel, e.g. the *Bombyx mori* transposase protein or its encoding nucleic acid may be administered and/or transfected into a cell as defined above prior to, simultaneously with or subsequent to administration and/or transfection of the *Bombyx mori* transposon. Alternatively, the *Bombyx mori* transposon may be transfected into a cell as defined above prior to, simultaneously with or subsequent to transfection of the *Bombyx mori* transposase protein or its encoding nucleic acid. If transfected in parallel, preferably both components are provided in a separated formulation and/or mixed with each other directly prior to administration to avoid transposition prior to transfection. Additionally, administration and/or transfection of at least one component of the kit may occur in a time staggered mode, e.g. by administering multiple doses of this component.

In addition, the present invention also features kits comprising a *Xenopus tropicalis* transposase as a protein or encoded by a nucleic acid, and/or a *Xenopus laevis* transposon; or a gene transfer system as described herein comprising a *Xenopus tropicalis* transposase as a protein or encoded by a nucleic acid as described herein, in combination with a *Xenopus tropicalis* transposon; optionally together with a pharmaceutically acceptable carrier, adjuvant or vehicle, and optionally with instructions for use. Any of the components of the inventive kit may be administered and/or transfected into cells in a subsequent order or in parallel, e.g. the *Xenopus tropicalis* transposase protein or its encoding nucleic acid may be administered and/or transfected into a cell as defined above prior to, simultaneously with or subsequent to administration and/or transfection of the *Xenopus tropicalis* transposon. Alternatively, the *Xenopus tropicalis* transposon may be transfected into a cell as defined above prior to, simultaneously with or subsequent to transfection of the *Xenopus tropicalis* transposase protein or its encoding nucleic acid. If transfected in parallel, preferably both components are provided in a separated formulation and/or mixed with each other directly prior to administration to avoid transposition prior to transfection. Additionally, administration and/or transfection of at least one component of the kit may occur in a time staggered mode, e.g. by administering multiple doses of this component.

6. EXAMPLES

The following examples are intended to illustrate the methods, compositions and kits disclosed herein and should not be construed as limiting in any way. Various equivalents will be apparent to one skilled in the art from the following examples; such equivalents are also contemplated to be part of the invention disclosed herein.

6.1.1 Stable Integration in Chinese Hamster Ovary (CHO) Cells

In some embodiments, a gene transfer system comprises a transposon and a transposase. The transposon comprises a heterologous expression polynucleotide that includes expression control elements and a sequence encoding a first expressible polypeptide. Cells into which the transposon and

transposase have both been introduced express higher levels of the expressible polypeptide than cells into which only the transposon have been introduced.

Tables 1 and 2 show data obtained from parallel triplicate experiments testing expression from an expression polynucleotide comprising a puromycin resistance gene under control of a murine PGK promoter and a DasherGFP gene under control of a human EF1a promoter, with the two promoters oriented such that transcription from them is in opposite directions and divergent. The expression polynucleotide was inserted between putative pairs of transposon ends to create a series of putative transposons. SEQ IDs of the transposon ends are indicated in the tables. The transposons were then either transfected alone into CHO cells, or co-transfected with a transposase. The ratio of transposon DNA to transposase-encoding DNA is indicated in the tables.

CHO-K1 cells (from ATCC) were grown in F12-K (from ATCC)+10% FBS (from ATCC)+1% Penicillin-streptomycin (from ATCC) at 37° C., 5% CO₂ to 80% confluence. 5E+05 cells were plated in 24-well tissue culture plates and incubated at 37° C., 5% CO₂ for 24 hours prior to transfection, transfections were set up in triplicates. Each transfection used a total of 0.5 µg DNA with Lipofectamine 2000 as per manufacturer's protocol. Media with puromycin was added 72 hours post transfection. Cells were grown for 14 days post puromycin selection with two passages and two changes of media. Fluorescence of the ORF encoding fluorescent reporter DasherGFP (SEQ ID: 102) was measured at Ex/Em of 488/518 nm.

A fluorescence signal that was higher in cells that received the transposon plus the transposase than the transposon alone indicated that the transposase was able to recognize the transposon ends and enhance integration into the genomic DNA, either by integrating more copies of the DNA or by integrating the DNA at places in the genome that were more favorable for expression.

Table 1 shows that expression from a transposon comprising *Bombyx mori* transposon ends SEQ ID NOS: 1 and 2 is increased approximately 5-fold when the transposon is co-transfected with a vector encoding a *Bombyx mori* transposase SEQ ID NO: 44. Tables 1 and 2 show that expression from a transposon comprising *Xenopus tropicalis* transposon ends SEQ ID NOS: 5 and 6 is increased between 50% and 2.5-fold when the transposon is co-transfected with a vector encoding a *Xenopus tropicalis* transposase SEQ ID NO: 45 or 46, when the transposase was fused to a nuclear localization signal.

Identifying and testing hyperactive variants of the identified transposases is also expressly contemplated and is another aspect of this invention.

6.1.2 Insulator Elements Enhance Stable Expression in CHO Cells

In some embodiments, a gene transfer system comprises a polynucleotide that includes expression control elements and a sequence encoding a first expressible polypeptide. Expression of the expressible polypeptide can be increased in some configurations of the gene transfer system by incorporating insulator sequences. In some embodiments the polynucleotide and the insulator sequences are part of a transposon, and expression can be further increased by the action of a transposase.

Table 3 shows data obtained from parallel triplicate experiments testing expression from an expression polynucleotide comprising a puromycin resistance gene under control of a murine PGK promoter and a DasherGFP gene under control of a second promoter as indicated in the table.

The expression polynucleotide was optionally inserted between pairs of insulator sequences as indicated in the table. The resulting polynucleotides were then inserted between pairs of transposon ends as indicated in the table. These transposons were then transfected into CHO cells, either alone or together with a transposase, as indicated in the table. The transposases in this example were fused to a heterologous nuclear localization signal.

CHO-K1 cells (from ATCC) were grown in F12-K (from ATCC)+10% FBS (from ATCC)+1% Penicillin-streptomycin (from ATCC) at 37° C., 5% CO₂ to 80% confluence. 5E+05 cells were plated in 24-well tissue culture plates and incubated at 37° C., 5% CO₂ for 24 hours prior to transfection, transfections were set up in triplicates. Each transfection used a total of 0.5 µg/µl DNA with Roche Extreme Gene 9 reagent (2:1 ratio) as per manufacturer's protocol. Media with 5 µg/µl puromycin was added 72 hours post transfection. Puromycin selection was carried out for 72 hours. Cells were grown for 14 days post puromycin selection with two passages and changes of media. Fluorescence represents expression of the ORF encoding fluorescent reporter DasherGFP from stably integrated transposons and was measured at Ex/Em of 488/518 nm.

Table 3 shows that the presence of HS4 insulators, either the full sequence or just the core sequence, can significantly increase expression from the transposon even in the absence of a transposase (for example compare Table 3 rows 1 and 3 or rows 5 and 9). This effect appears to be influenced by the promoter that is being used to drive expression of the expressible polypeptide: in this vector context the HS4 insulator increases expression from an EF1a promoter on the transposon by 50% to 4-fold, but there is no effect observed when the expressible polypeptide is under the control of the CMV promoter (Table 3 rows 11, 13 and 15). In the presence of the transposase, expression from almost all of the transposons increased, from 50% to more than 10-fold. Increases in expression from the transposon as a result of the insulators and the transposases appeared to be synergistic. Preferred embodiments of gene transfer systems comprise a gene encoding a *Xenopus* or *Bombyx* transposase fused to a nuclear localization signal and a *Xenopus* or *Bombyx* transposon comprising two HS4 or HS4 core insulators.

6.1.3 Transposases can be Provided in Cis or in Trans

In some embodiments, a gene transfer system comprises a transposon and a transposase. The transposon comprises a heterologous polynucleotide that includes expression control elements and a sequence encoding a first expressible polypeptide. In some embodiments the transposase is encoded on a polynucleotide vector that also comprises the transposon.

Table 4 shows data obtained from parallel triplicate experiments testing expression from an expression polynucleotide comprising a puromycin resistance gene under control of a murine PGK promoter and a DasherGFP gene under control of a second promoter as indicated in the table. The DasherGFP gene is followed by an RNA export element, as indicated in the table. The expression polynucleotide was optionally inserted between pairs of insulator sequences as indicated in the table. The resulting polynucleotides were then inserted between pairs of transposon ends as indicated in the table. These transposons were then optionally cloned into vectors that also contained a gene for expression of a transposase, under control of a promoter as indicated (P_Transposase). Some transposases were fused to a nuclear localization signal, as indicated in the table.

Transposons were then transfected into CHO cells, either alone or together with a transposase, as indicated in the table.

CHO-K1 cells (from ATCC) were grown in F12-K (from ATCC)+10% FBS (from ATCC)+1% Penicillin-streptomycin (from ATCC) at 37° C., 5% CO₂ to 80% confluence. 5E+05 cells were plated in 24-well tissue culture plates and incubated at 37° C., 5% CO₂ for 24 hours prior to transfection, transfections were set up in triplicates. Each transfection used a total of 0.5 µg DNA with Roche Extreme Gene 9 reagent (2:1 ratio) as per manufacturer's protocol. Media with 5 µg/µl puromycin was added 72 hours post transfection. Puromycin selection was carried out for 72 hours. Cells were grown for 14 days post puromycin selection with two passages and changes of media. Fluorescence represents expression of the ORF encoding fluorescent reporter DasherGFP from stably integrated transposons and was measured at Ex/Em of 488/518 nm.

Table 4 shows that expression of transposase from the same vector that contained the transposon produced comparable levels of expression improvement to the expression increases obtained by co-transfecting a second plasmid carrying the gene for the transposase. This improvement is observed in many different vector configurations, as shown in the table. Thus expression from a *Bombyx mori* transposon or a *Xenopus tropicalis* transposon can be increased by the action of a transposase that is provided either in cis or in trans.

The transposons described in Table 4 also comprised an RNA export element selected from WPRE and HPRE; they also comprised the expression enhancing element SARI. Preferred embodiments of gene transfer vectors comprise one or more of these elements.

6.1.4 Transposases with Nuclear Localization Signals

In some embodiments, a gene transfer system comprises a transposon and a transposase, where the transposase is fused with a nuclear localization signal. The transposon comprises a heterologous polynucleotide that includes expression control elements and a sequence encoding a first expressible polypeptide. Cells into which the transposon and transposase fused to the nuclear localization signal have both been introduced express higher levels of the expressible polypeptide than cells into which only the transposon have been introduced.

Table 5 shows data obtained from parallel triplicate experiments testing expression from an expression polynucleotide comprising a puromycin resistance gene under control of a murine PGK promoter, with a DasherGFP gene translationally coupled to the puromycin-resistance gene through a CHYSEL sequence. The expression polynucleotide was inserted between putative pairs of transposon ends to create a series of putative transposons. SEQ IDs of the transposon ends are indicated in the tables. The transposons were then either transfected alone into CHO cells, or co-transfected with a transposase fused to a nuclear localization sequence. The ratio of transposon DNA to transposase-encoding DNA is indicated in the tables.

CHO-K1 cells (from ATCC) were grown in F12-K (from ATCC)+10% FBS (from ATCC)+1% Penicillin-streptomycin (from ATCC) at 37° C., 5% CO₂ to 80% confluence. 5E+05 cells were plated in 24-well tissue culture plates and incubated at 37° C., 5% CO₂ for 24 hours prior to transfection, transfections were set up in triplicates. Each transfection used a total of 0.5 µg DNA with Lipofectamine 2000 as per manufacturer's protocol. Media with puromycin was added 72 hours post transfection. Cells were grown for 14 days post puromycin selection with two passages and two

changes of media. Fluorescence of the ORF encoding fluorescent reporter DasherGFP was measured at Ex/Em of 488/518 nm.

A fluorescence signal that was higher in cells that received the transposon plus the transposase fused to the nuclear localization signal than the transposon alone indicated that the transposase fused to the nuclear localization signal was able to recognize the transposon ends and enhance integration into the genomic DNA, either by integrating more copies of the DNA or by integrating the DNA at places in the genome that were more favorable for expression.

Trichoplusia ni piggyBac and *Bombyx mori* and *Xenopus tropicalis* transposases were all active when fused to N-terminus nuclear localization signals (NLS). A preferred embodiment of a gene transfer system comprises a gene encoding a transposase that is fused to a nuclear localization signal.

6.1.5 Stable Integration in Human Embryonic Kidney (HEK 293) Cells

In some embodiments the gene transfer system comprises a transposon and transposase that are used to integrate an expression polynucleotide into the genome of a mammalian cell; in some embodiments the cell is a CHO cell, in some embodiments the cell is an HEK cell.

Table 6 shows data obtained from parallel triplicate experiments testing expression from an expression polynucleotide in a *Xenopus* transposon with ends SEQ ID NO: 9 and 6, comprising a puromycin resistance gene under control of a murine PGK promoter. The expression polynucleotide further comprised a DasherGFP gene operably linked to various promoters, introns, RNA export sequences and polyadenylation sequences as indicated in the table. Transcription from the two promoters was in opposite directions and divergent. The transposons were then either transfected alone into HEK cells, or co-transfected with a gene encoding *Xenopus* transposase SEQ ID NO: 45 fused to a nuclear localization sequence.

HEK 293 cells were grown in EMEM (from ATCC)+10% FBS (from ATCC)+1% Penicillin-streptomycin (from ATCC) at 37° C., 5% CO₂ to 80% confluence. 5E+05 cells were plated in 24-well tissue culture plates and incubated at 37° C., 5% CO₂ for 24 hours prior to transfection, transfections were set up in triplicates. Each transfection used a total of 0.5 µg DNA with Roche Extreme Gene 9 reagent (2:1 ratio) as per manufacturer's protocol. Media with 5 µg/µl puromycin was added 72 hours post transfection. Puromycin selection was carried out for 72 hours and passaged into complete media minus puromycin. Cells were grown for 14 days post puromycin selection with two passages and changes of media. Fluorescence represents expression of the ORF encoding fluorescent reporter DasherGFP from stably integrated transposons and was measured at Ex/Em of 488/518 nm.

All gene transfer vector configurations tested showed improved expression in HEK cells when the transposase gene was co-transfected. Preferred embodiments of a gene transfer vector include all vector configurations shown in Table 6

6.1.6 Expression Levels of Two Polypeptides Using IRES Elements in Transient Expression in HEK293 and CHO Cells

In some embodiments a gene transfer system comprises genes encoding two polypeptides. In some embodiments the two polypeptides are encoded on a single polynucleotide. In some embodiments the two polypeptides interact after they are synthesized. In some embodiments the relative amounts of the two polypeptides expressed by a cell is important for

the functioning of the two polypeptides. In some embodiments the two polypeptides are enzymes in a pathway. In some embodiments the two polypeptides bind together or are subunits of a larger molecule; in some embodiments the two polypeptides are the heavy and light chains of an antibody.

Tables 7-11 show the expression levels observed in HEK and CHO cells for two different polypeptides (in this case two different fluorescent proteins, Dasher GFP and CayenneRFP) encoded on a single gene transfer vector. The genes for the two different proteins were operably linked to a single enhancer, promoter, polyadenylation signal and optionally an intron, as indicated in the tables. Expression of the two genes was operably linked by an IRES element, as indicated in the tables, with the order of elements being DasherGFP-IRES-CayenneRFP.

HEK 293a cells (from ATCC) were grown in EMEM (from ATCC)+10% FBS (from ATCC)+1% Penicillin-streptomycin (from ATCC) at 37° C., 5% CO₂ to 80% confluence, 1E+05 cells were plated in 24-well tissue culture plates and incubated at 37° C., 5% CO₂ for 24 hours prior to transfection, transfections were set up in triplicates. Each transfection used 0.5 µg DNA with Lipofectamine 2000 as per manufacturer's protocol. Cells were harvested 72 hours post transfection. Fluorescence of the two ORFs encoding fluorescent reporters DasherGFP (SEQ ID NO: 102) and CayenneRFP (SEQ ID NO: 103) was measured at Ex/Em of 488/518 nm for DasherGFP and Ex/Em of 525/580 nm for CayenneRFP.

CHO-K1 cells (from ATCC) were grown in F12-K (from ATCC)+10% FBS (from ATCC)+1% Penicillin-streptomycin (from ATCC) at 37° C., 5% CO₂ to 80% confluence. 5E+05 cells were plated in 24-well tissue culture plates and incubated at 37° C., 5% CO₂ for 24 hours prior to transfection, transfections were set up in triplicates. Each transfection used 0.5 µg DNA with Lipofectamine 2000 as per manufacturer's protocol. Cells were harvested 72 hours post transfection. Fluorescence of the two ORFs encoding fluorescent reporters DasherGFP and CayenneRFP was measured at Ex/Em of 488/518 nm for DasherGFP and Ex/Em of 525/580 nm for CayenneRFP.

Gene transfer vectors comprising the two proteins translationally coupled by a CHYSEL sequence (e.g. construct 135171 in Table 7) express the two proteins at an equimolar ratio and can be used to normalize for different fluorescent intensities of the proteins. Tables 7-11 show that different IRES elements can be used to obtain different ratios of expression between two different polynucleotides in a variety of vector configurations. The use of IRES elements is particularly advantageous for expression of polypeptides when the ratio of expression is important at the level of individual cells, for example in the expression of antibodies where the light chain is often thought to perform a chaperonin function for the heavy chain.

We have identified IRES elements that show different levels of activity as seen from the varying expression levels for the two open reading frames (ORFs) linked by an IRES element (Tables 7-11). A choice of IRES elements with varying activities allows the appropriate IRES element to be used for controlling the relative expression levels of two ORFs. This is especially useful for expression of antibodies wherein the ratios of expression of the heavy chain to light chain influences proper assembly of the functional antibody. Use of the identified IRES elements for antibody expression is an important aspect of this invention. We have shown use of one IRES element linking two transcripts operably linked to one promoter, similarly use of two or more IRES elements

linking three or more ORFs is expressly contemplated and is another aspect of the invention. Expression constructs with two or more IRES elements selected such that transcript levels of two or more ORFs is selectively modulated is expressly contemplated and is an important aspect of the invention. The identified IRES elements of the invention work well in both transient and stable integration vectors in the two cell lines tested, Human embryonic kidney (HEK293) cells and Chinese hamster ovary (CHO) cells. Preferred embodiments of a gene transfer vector include all vector configurations shown in Tables 7-11, and all IRES elements shown in these tables.

6.1.7 Expression of Antibodies Using IRES Elements in Transiently Transfected HEK293 Cells

In some embodiments the gene transfer system is used to express an antibody. In some embodiments genes encoding the two antibody chains are operably linked to separate promoters. In some embodiments genes encoding the two chains are operably linked to the same promoter and to each other by a translational coupling element, in some embodiments the translational coupling element is an IRES or a CHYSEL sequence.

Table 12 shows a variety of vector configurations expressing two antibody chains from a single gene transfer vector. In some configurations genes encoding the two chains were each operably linked to separate promoters and polyadenylation signals, in some configurations the genes were operably linked to a single promoter preceding the first gene and a single polyA signal following the second gene, where the two genes are operably linked by an IRES sequence. The number 1 indicates promoters preceding or polyA signals following the first gene, the number 2 indicates promoters preceding or polyA signals following the second gene.

All of these vectors further comprised a viral amplification sequence encoding the SV40T antigen, and the SV40 origin of replication. Antibody expression from these gene transfer vectors was measured by ELISA, and compared with the expression obtained by co-transfection of two gene transfer vectors, one encoding the heavy chain and the other encoding the light chain, transfected at different ratios as indicated in the table.

HEK 293a cells (from ATCC) were grown in EMEM (from ATCC)+10% FBS (from ATCC)+1% Penicillin-streptomycin (from ATCC) at 37° C., 5% CO₂ to 80% confluence, 1E+05 cells were plated in 24-well tissue culture plates and incubated at 37° C., 5% CO₂ for 24 hours prior to transfection, transfections were set up in triplicates. Each transfection used 0.5 µg DNA with Lipofectamine 2000 as per manufacturer's protocol. Cells were harvested 72 hours post transfection. Culture supernatants were harvested and used in an ELISA assay for quantitation of heavy chain (HC) (Table 12) and run on a gel for a western blot for detection of the heavy and light chains (data not shown).

96-well ELISA plates (Cat. No. M9410, Sigma) were coated with 50 µl per well of goat anti-Human IgG (Fc specific) antibody (Cat No. 12136, Sigma) at 1 µg/ml in 1xPBS (Cat. No. P-7059, Sigma) and incubated overnight at 4° C. Plates were washed 4 times with 300 µl per well PBST (Cat. No. P3563, Sigma) and blocked with 300 µl per well ELISA Block solution (PBST+1% BSA (Cat. No. 85040C), Sigma) for 1 hour at room temperature. ELISA Block solution was removed and culture supernatants from the transient transfections above diluted in ELISA Block solution at dilutions ranging from 1:50 to 1:200,000 at 100 µl per well were added to plates and incubated for 1 hour at room temperature. Plates were washed 4 times with 300 µl per well PBST (Sigma) and antibody was detected by incubation

with 100 μ l/well (0.16 μ g/ml) HRP conjugated-goat anti-Human IgG (Fab specific) (Cat. No. 31482, Thermo Scientific) in ELISA Block solution for 1 hour at room temperature. Plates were washed 4 times with 300 μ l per well PBST (Sigma) and bound HRP-goat anti-Human IgG measured by adding 100 μ l/well QuantaBlu Fluorogenic Peroxidase substrate (Cat. No. 15162, Thermo Scientific). Plates were incubated for 5 minutes at room temperature, reaction stopped by adding 100 μ l/well stop solution (Cat. No. 15162, Thermo Scientific) and fluorescence measured using fluorimeter at excitation (Ex) 325 nm and emission (Em) 420 nm. Antibody concentration was calculated by comparing to a standard curve generated using dilutions of purified Human IgG (Thermo Scientific) using a 4-parameter logistic curve fit. Concentrations of IgG calculated shown in Table 12 were in good agreement with quantitation from western blot (data not shown).

Table 12 shows that many gene transfer vector configurations using dual promoters or IRES elements produce comparable or better antibody expression than co-transfection of two separate gene transfer vectors. Preferred embodiments of a gene transfer vector include all vector configurations shown in Table 12, and all IRES elements shown in Tables 7-11.

Using IRES sequences which can produce different levels of expression of the second polypeptide relative to the first has advantages over methods wherein dual-transfections are used. In the case of co-transfection, individual cells take up different numbers of each plasmid. Because it is only possible to control the average number of each plasmid taken up by each cell, many cells do not end up expressing the optimal ratio of heavy and light chain. This problem is amplified in the case of stable cell lines, because there is the further variable of integration location which also affects expression levels. Use of IRES sequences to control expression levels of two or more ORFs, in particular expression of ORFs encoding heavy and light chains in stable integration expression vectors is expressly contemplated and is an important aspect of the invention.

6.1.8 Expression of Two ORFs Linked by IRES Elements in Stably Transfected CHO Cells

In some embodiments a gene transfer system comprises genes encoding two polypeptides encoded on a single polynucleotide to be stably integrated into the genome of a cell. In some embodiments the polynucleotide comprises a transposon. In some embodiments the two polypeptides are the heavy and light chains of an antibody.

Table 13 shows the configurations of a set of transposons comprising transposon ends SEQ ID NO: 9 and 6. Transposons comprised genes encoding DasherGFP and/or CayenneRFP as indicated in the table. In some configurations the two genes were each operably linked to separate promoters and polyadenylation signals, in some configurations the genes were operably linked to a single promoter preceding the first gene and a single polyA signal following the second gene, where the two genes are operably linked by an IRES sequence. The number 1 indicates promoters preceding or polyA signals following the first gene, the number 2 indicates promoters preceding or polyA signals following the second gene. All of these sequences further comprised an SAR sequence and an HPRE sequence following the second gene and preceding polyA2. Transposons were transfected into CHO cells, either alone or together with a gene encoding a transposase (SEQ ID NO: 45) fused to a nuclear localization signal.

CHO-K1 cells (from ATCC) were grown in F12-K (from ATCC)+10% FBS (from ATCC)+1% Penicillin-streptomycin

(from ATCC) at 37° C., 5% CO₂ to 80% confluence. 5E+05 cells were plated in 24-well tissue culture plates and incubated at 37° C., 5% CO₂ for 24 hours prior to transfection, transfections were set up in triplicates. Each transfection used a total of 0.5 g DNA with Roche Extreme Gene 9 reagent (2:1 ratio) as per manufacturer's protocol. Media with 5 μ g/ μ l puromycin was added 72 hours post transfection. Puromycin selection was carried out for 72 hours. Cells were grown for 14 days post puromycin selection with two passages and changes of media. Fluorescence represents expression of the ORFs encoding fluorescent reporter DasherGFP from stably integrated transposons measured at Ex/Em of 488/518 nm and CayenneRFP was measured at Ex/Em of 525/580 nm.

Co-transfection of transposons with the vector encoding the transposase increased expression of both proteins encoded by the transposon between 4-fold and nearly 20-fold relative to transfections with the transposon alone. A transposon comprising genes encoding two polypeptides and a gene encoding a transposase fused to a nuclear localization signal is a preferred embodiment of a gene transfer system. Configurations shown in Table 13 are preferred embodiments.

FIG. 4 shows FACS analysis of populations of cells transfected with the gene transfer systems described in Table 13 rows 3 and 4.

6.1.9 Stable Integration of Transposons in Chinese Hamster Ovary (CHO) Cells

In some embodiments, a gene transfer system comprises a transposon and a transposase. The transposon comprises a heterologous expression polynucleotide that includes expression control elements and a sequence encoding a first expressible polypeptide. Cells into which the transposon and transposase have both been introduced express higher levels of the expressible polypeptide than cells into which only the transposon have been introduced. In some embodiments the cell is a mammalian cell, in some embodiments the cell is a CHO cell or an HEK cell.

Table 14 shows a set of gene transfer vector configurations for expression of a DasherGFP. Each vector comprised transposon ends SEQ ID NO: 9 and 6. All of these sequences except for 192462 further comprised an SAR sequence and an HPRE sequence following the sequence encoding DasherGFP. All of these sequences further comprised a rabbit globin polyA sequence. Some transposons further comprised a pair of HS4 insulator sequences between the transposon ends, as indicated in the table. Transposons were transfected into CHO cells, either alone or together with a gene encoding a transposase (SEQ ID NO: 45) fused to a nuclear localization signal.

Chinese hamster ovary (CHO) cells were grown and transfected as described in Example 6.2 above.

Table 14 shows that expression from the transposon was increased for each configuration tested by co-transfection with a gene encoding the transposase. The expression increases were between 2-fold and 80-fold. Gene transfer systems comprising a transposase and a transposon with a configuration shown in Table 14 are preferred embodiments of the invention.

FIG. 3 shows a FACS analysis of the same stably transfected cell populations as shown in Table 14. FACS shows the expression level of DasherGFP in individual cells in the stably transfected population. FIG. 3 shows that for every gene transfer vector in the set, the transposase causes a shift of a part of the population from the poorly expressing group to the highly expressing group. This may be caused by an increase in the number of copies of the transposon that have

been stably integrated, or it may be caused by the transposon integrating into genomic loci that result in improved expressibility.

We have shown preferred configurations of vector elements including enhancers, promoters, introns, 5' UTRs, RNA export sequences, poly A and insulators that contribute to expression activity seen from integrated transposons. Other vector configurations of control elements shown in Tables 15-18, placed into a transposon context for stable integration, are also preferred embodiments.

In a preferred embodiment for stable integration into a mammalian genome, a gene transfer vector comprises a transposon for expression of a first polynucleotide, wherein the expression of the first polynucleotide is operably linked to a mammalian promoter selected from amongst the EF1a (translation elongation factor 1a) promoter from any mammalian or avian species including but not limited to human, rat, mice, chicken and Chinese hamster; the CMV (cytomegalovirus) promoter, the GAPDH (glyceraldehyde-3-phosphate dehydrogenase) promoter from any mammalian species; the MCI and HSV-TK (Herpes Simplex Virus thymidine kinase) viral promoters; the actin promoter from any mammalian or avian species including but not limited to human, rat, mice, chicken and Chinese hamster; the PGK (phosphoglycerate kinase) promoter from any mammalian or avian species including but not limited to human, rat, mice, chicken and Chinese hamster; the SV40 (Simian virus 40) promoter and the ubiquitin promoter. In preferred embodiments the promoter may be operably linked to an enhancer selected from amongst the CMV immediate early enhancer, the EF1a enhancer (for example but not limited to SEQ ID NO: 116), the adenoviral major late protein enhancer (for example but not limited to SEQ ID NO: 118), the SV40 enhancer (for example but not limited to SEQ ID NO: 117), and a retroviral LTR. In further preferred embodiments expression of the first polynucleotide is operably linked to an intron selected from among a CMV (cytomegalovirus) intron, including CMV introns A, B and C; an EF1a (translation elongation factor 1a) intron from any mammalian or avian species including but not limited to human, rat, mice, chicken and Chinese hamster, the actin intron from any mammalian or avian species including but not limited to human, rat, mice, chicken and Chinese hamster; a GAPDH (glyceraldehyde-3-phosphate dehydrogenase) intron from any mammalian or avian species including but not limited to human, rat, mice, chicken and Chinese hamster; synthetic introns (for example but not limited to SEQ ID NO: 119) and chimeric natural introns (for example SEQ ID NO: 123). In further preferred embodiments expression of the first polynucleotide is operably linked to an RNA export element (for example but not limited to SEQ ID NOS: 104-107), an insulator sequence (for example but not limited to SEQ ID NOS: 112-113), or an expression enhancing element (for example but not limited to SEQ ID NOS: 108-111).

Regardless of their actual mechanism, we have shown incorporation of expression enhancing elements into gene transfer vectors with transposon ends greatly enhances protein expression from these vectors, both in the absence and presence of transposase.

In addition to vector elements within the transposon, use of vector elements located in the vector such that these elements are not transposed by the transposase, for example viral replication elements such as SV40 origin of replication (SV40 ori) with the SV40 large T antigen and EBV (Epstein Barr virus) oriP with EBNA (Epstein-Barr nuclear antigen) is another aspect of the invention.

6.1.10 Creating and Testing Combinations of Vector Elements in Transiently Transfected HEK and CHO Cells

In some embodiments preferred configurations of expression control elements in a gene transfer vector are determined by creating a set of vectors comprising different combinations of elements, and then measuring expression from the set. In preferred embodiments the set comprises between 5 and 200 members, in preferred embodiments the set comprises between 10 and 100 members, in preferred embodiments the set comprises between 15 and 60 members.

48 vectors were designed such that the set of vectors comprised 3 different enhancer elements (CMV, SV40, none); 9 different promoter elements (CMV, Ub-B, SV40, PGK, MCI, HSV-TK, GAPDH, EF1a, chick actin); 10 different intron elements (CMV intron a, CMV intron c, chick actin (partial), GAPDH, chick actin/rabbit, EF1a-v1, EF1a-v2, EF1a-hybrid, synthetic and none); 2 different 5' UTRs (CMV and none); 7 different polyadenylation sequences (SV40 late, SV40 early, bovine growth hormone, synthetic, human beta globin, rabbit beta globin, HSV-TK); 3 different viral replication origins (Epstein Barr virus oriP, SV40 ori and none) and 3 different viral replication proteins (Epstein Barr virus EBNA, SV40 T antigen and none). The specific combinations in the set are shown in Table 15. The total number of possible combinations of these elements is 34,020.

Dasher GFP was cloned into each of the vectors, and each construct was transfected in triplicate into HEK cells and CHO cells. Cells were plated in 24-well plates in DMEM (ATCC)+10% FBS+Penicillin/Streptomycin for HEK293a cells and F12K (ATCC)+10% FBS+Penicillin/Streptomycin for CHO-K1 cells and grown for 24 hours to approximately 70-80% confluence. Cells were transfected in duplicate in 24-well plates using 1 µl Lipofectamine 2000 per 0.5 µg DNA. After 72 hours the cells were lysed using 200 l/well M-PER Mammalian Protein Extraction Reagent (Thermo Scientific Pierce®) and total fluorescence measured on a fluorescence plate reader at excitation/emission wavelengths of 505/525 nm. The average fluorescence readings are shown in Table 19.

Partial Least Squares regression models were constructed from the fluorescence data, and regression weights were calculated for each of the sequence elements. The regression weights indicate the importance of the element for performance of the vector. It is clear that some elements or combinations of elements are more favorable for expression in both HEK and CHO (CMV enhancer, promoter and CMV intron a for example); some elements are more favorable for expression in HEK but less favorable for expression in CHO (SV40 ori plus SV40 T antigen, HSVTK polyadenylation signal, EBV oriP); some elements are more favorable for expression in CHO but less favorable for expression in HEK (GAPDH promoter, human beta globin polyadenylation signal); some elements are less favorable for expression in either HEK or CHO (SV40 late polyadenylation signal, HSV-TK promoter).

Particularly favorable for expression in HEK cells were vectors that contained the SV40 origin of replication and the SV40 T antigen. These were even more effective when combined with the CMV enhancer plus the chick actin promoter plus the chick actin/rabbit intron, or the CMV enhancer with the CMV promoter with CMV intron a. Particularly favorable for expression in CHO cells were vectors that lacked any viral replication sequences. Vector configurations shown in Table 15 are preferred embodiments of the invention.

6.1.11 Creating and Testing a Second Set of Combinations of Vector Elements in Transiently Transfected HEK and CHO Cells

In some embodiments preferred configurations of expression control elements in a gene transfer vector are determined by creating a set of vectors comprising different combinations of elements, measuring expression from the set, and then designing a second set of vectors in which the most favorable elements for expression from the first set are retained and recombined, and, optionally, new elements are added into the new set of vectors. In preferred embodiments the second set comprises between 5 and 200 members, in preferred embodiments the second set comprises between 10 and 100 members, in preferred embodiments the second set comprises between 15 and 60 members.

48 vectors were designed such that the set of vectors comprised 4 different enhancer elements (CMV, synthetic, EF1a, none); 4 different promoter elements (CMV, GAPDH, EF1a, EF1a_LTR-HTLV); 6 different intron elements (CMV intron a, CMV intron c, GAPDH, chick actin/rabbit, EF1a and none); 8 different 5' UTRs (CMV, satellite tobacco necrosis virus (sTNV), human beta globin, polyhedrin, tobacco necrosis virus (TNV), barley yellow dwarf virus (BYDV), *Xenopus* globin, and none); 6 different 3' UTRs (satellite tobacco necrosis virus (sTNV), polyhedrin, tobacco necrosis virus (TNV), barley yellow dwarf virus (BYDV), *Xenopus* globin, and none); 6 different polyadenylation sequences (bovine growth hormone, bovine growth hormone plus gastrin terminator, rabbit beta globin, rabbit beta globin plus gastrin terminator, HSV-TK, HSV-TK plus gastrin terminator); 3 different viral replication origins (Epstein Barr virus oriP, SV40 ori and none) and 3 different viral replication proteins (Epstein Barr virus EBNA, SV40 T antigen and none). The specific combinations in the set are shown in Table 16. The total number of possible combinations of these elements is 248,832.

Dasher GFP was cloned into each of the vectors, and each construct was transfected in triplicate into HEK cells and CHO cells. Cells were plated in 24-well plates in DMEM (ATCC)+10% FBS+Penicillin/Streptomycin for HEK293a cells and F12K (ATCC)+10% FBS+Penicillin/Streptomycin for CHO-K1 cells and grown for 24 hours to approximately 70-80% confluence. Cells were transfected in duplicate in 24-well plates using 1 μ l Lipofectamine 2000 per 0.5 μ g DNA. After 72 hours the cells were lysed using 200 μ l/well M-PER Mammalian Protein Extraction Reagent (Thermo Scientific Pierce®) and total fluorescence measured on a fluorescence plate reader at excitation/emission wavelengths of 505/525 nm. The average fluorescence readings relative to control constructs are shown in Table 20.

Partial Least Squares regression models were constructed from the fluorescence data, and regression weights were again calculated for each of the sequence elements. Again some elements are more favorable for expression in both HEK and CHO, some elements are more favorable for expression in HEK but less favorable in CHO, some elements are more favorable for expression in CHO but less favorable in HEK and some elements are less favorable for expression in both HEK and CHO.

Particularly favorable for expression in HEK cells were vectors that combined the CMV enhancer with the CMV promoter, CMV intron a or intron c, and the polyadenylation signal from rabbit beta globin or the polyadenylation signal from HSV-TK plus the gastrin terminator. Even more favorable combinations also included the SV40 origin of replication and the SV40 T antigen. Vector configurations shown in Table 16 are preferred embodiments of the invention.

6.1.12 Creating and Testing a Set of Combinations of Vector Elements in Transiently Transfected CHO Cells

In some embodiments preferred configurations of expression control elements in a gene transfer vector are determined by creating a set of vectors comprising different combinations of elements, measuring expression from the set, and then designing a second set of vectors in which the most favorable elements for expression from the first set are retained and recombined, and, optionally, new elements are added into the new set of vectors. In some embodiments this process is repeated.

The most favorable elements for CHO were selected to create a third set of vectors as shown in Table 17. Dasher GFP was cloned into each of the vectors, and each construct was transfected in triplicate into CHO cells. Cells were plated in 24-well plates in DMEM (ATCC)+10% FBS+Penicillin/Streptomycin for HEK293a cells and F12K (ATCC)+10% FBS+Penicillin/Streptomycin for CHO-K1 cells and grown for 24 hours to approximately 70-80% confluence. Cells were transfected in duplicate in 24-well plates using 1 μ l Lipofectamine 2000 per 0.5 μ g DNA. After 72 hours the cells were lysed using 200 μ l/well M-PER Mammalian Protein Extraction Reagent (Thermo Scientific Pierce®) and total fluorescence measured on a fluorescence plate reader at excitation/emission wavelengths of 505/525 nm. The average fluorescence readings relative to control constructs are shown in Table 17.

Vector configurations shown in Table 17 are preferred embodiments of the invention. Particularly favorable vector elements for expression in CHO include the CMV enhancer together with the CMV promoter, the GAPDH promoter and the actin promoter. Expression is enhanced with the GAPDH intron, the CMV intron A or CMV intron C or the adenoviral major late protein enhancer.

6.1.13 Creating and Testing a Set of Combinations of Vector Elements in Stably Transfected CHO Cells

In some embodiments a gene transfer vector comprises an RNA export element. Table 18 shows a set of different vector configurations. The vectors further comprised an SAR sequence. A gene encoding DasherGFP was cloned into the vectors, stably transfected into CHO cells, and expression of the Dasher GFP was measured. The transposons were not co-transfected with transposases.

CHO-K1 cells (from ATCC) were grown in F12-K (from ATCC)+10% FBS (from ATCC)+1% Penicillin-streptomycin (from ATCC) at 37° C., 5% CO₂ to 80% confluence. 5E+05 cells were plated in 24-well tissue culture plates and incubated at 37° C., 5% CO₂ for 24 hours prior to transfection, transfections were set up in triplicates. Each transfection used a total of 0.5 μ g DNA with Roche Extreme Gene 9 reagent (2:1 ratio) as per manufacturer's protocol. Media with 5 μ g/ μ l puromycin was added 72 hours post transfection. Puromycin selection was carried out for 72 hours. Cells were grown for 14 days post puromycin selection with two passages and changes of media. Fluorescence represents expression of the ORFs encoding fluorescent reporter DasherGFP from stably integrated transposons measured at Ex/Em of 488/518 nm.

Particularly favorable for expression in stably transfected CHO cells were vectors that combined the SAR plus HPRE RNA export elements. Even more favorable combinations also included the EF1a promoter or HS4 insulators. Particularly favorable for expression in stably transfected HEK cells were vectors that combined the SAR plus HPRE or SAR plus AGS_1 or SAR plus AGS_3 RNA export elements.

6.1.14 Selecting Vector Elements Preferred by Different Expression Systems

In some embodiments a set of gene transfer vectors are tested for an expression property of a first polynucleotide in an expression system. In some embodiments a sequence-activity model is constructed between an expression property and the element configuration in the gene transfer vectors. In some embodiments the expression system is a mammalian cell, in some embodiments the mammalian cell is an human cell or a rodent cell, in some embodiments the mammalian cell is an HEK293 cell or a Chinese hamster ovary cell.

A model relating the elements of gene transfer vectors shown in Table 15, and the expression data shown in Table 19 was constructed using partial least square regression. A comparison between the measured expression property and the predicted expression property is shown in FIG. 5. Table 21 shows the regression weights for each element or combination of elements calculated from the model for HEK and CHO expression systems.

Table 21 shows that different combinations of elements are more favorable in different expression systems. Elements may be selected or rejected for incorporation into new vector configurations, depending on the regression weights from the model. For example the SV40 viral amplification system is highly favorable for expression in an HEK expression system (regression weight 2,679) but unfavorable for a CHO system (regression weight—113). Thus SV40 replication sequences were included in a new set of vectors designed for HEK (Table 16) but not in a new set of vectors designed for CHO (Table 17). The model also indicates that there are some polyA signals that are favorable for both systems (for example the sequence from rabbit beta globin and some that are less favorable (the SV40 late polyadenylation signal).

BRIEF DESCRIPTION OF TABLES

Table 1. Expression from gene transfer systems comprising transposons and transposases. Transposons comprising an expression cassette for DasherGFP, and with transposon end sequences as identified in columns D and E, were transfected into CHO cells, optionally together with a gene encoding a transposase (column G) at a predetermined ratio (column C). Cells were selected and expression of the DasherGFP measured (columns I-K) as described in Example 6.1.1.

Table 2. Expression from gene transfer systems comprising transposons and transposases. Transposons comprising an expression cassette for DasherGFP, and with transposon end sequences as identified in columns D and E, were transfected into CHO cells, optionally at a predetermined ratio (column C) together with a gene encoding a transposase (column G) which was optionally fused to a heterologous nuclear localization sequence (column H). Cells were selected and expression of the DasherGFP measured (columns I and J) as described in Example 6.1.1.

Table 3. Expression from gene transfer systems comprising transposons and transposases. Transposons comprised transposon end sequences (columns B and C), an expression cassette with a promoter (column E) operably linked to a gene encoding DasherGFP, and optionally insulator sequences on either side of the Dasher expression cassette (column F). Transposons were transfected into CHO cells, optionally together with a gene encoding a transposase (column G) fused to a heterologous nuclear localization

signal. Cells were selected and expression of the DasherGFP measured (columns H-J) as described in Example 6.1.2.

Table 4. Expression from gene transfer systems comprising transposons and transposases. Transposons comprised transposon end sequences (columns B and C), an expression cassette with a promoter (column E) and an element to enhance RNA export (column F) operably linked to a gene encoding DasherGFP, and optionally insulator sequences on either side of the Dasher expression cassette (column G). Transposons were transfected into CHO cells, optionally together with a gene encoding a transposase (column H) optionally fused to a heterologous nuclear localization signal (column L) and operably linked to a promoter (column K). In some configurations a single polynucleotide comprised the transposon and the gene encoding the transposase (column J). Cells were selected and expression of the DasherGFP measured (columns M-O) as described in Example 6.1.3.

Table 5. Expression from gene transfer systems comprising transposons and transposases. Transposons comprising an expression cassette for DasherGFP, and with transposon end sequences as identified in columns C and D, were transfected into CHO cells, optionally at a predetermined ratio (column B) together with a gene encoding a transposase (column F) which was optionally fused to a heterologous nuclear localization sequence (column G). Cells were selected and expression of the DasherGFP measured (columns H-J) as described in Example 6.1.4.

Table 6. Expression from gene transfer systems comprising transposons and transposases. Transposons comprised transposon end sequences SEQ ID NO. 9 and SEQ ID NO. 6, and an expression cassette with an enhancer (column C), promoter (column D), intron (column E), element to enhance RNA export (column F) and polyadenylation sequence (column G) operably linked to a gene encoding DasherGFP. Transposons were transfected into CHO cells, optionally together with a gene encoding a transposase (SEQ ID NO. 45) fused to a heterologous nuclear localization signal. Cells were selected and expression of the DasherGFP measured (columns H and I, no transposase, columns J and K plus transposase) as described in Example 6.1.5.

Table 7. Expression from gene transfer systems comprising genes encoding two polypeptides linked by translational coupling elements. Gene transfer vectors comprised an enhancer (column E), promoter (column F), intron (column G) and polyadenylation signal (column I) operably linked to a gene encoding DasherGFP (rows 1 and 3-22) or CayenneRFP (row 2). For rows 3-22, vectors further comprised a gene encoding CayenneRFP operably linked to the expression control elements by a translational-coupling sequence (sequences identified in column H). Vectors were transfected into HEK293 cells (columns J-M) or CHO cells (columns N-Q), and expression of the fluorescent proteins measured as described in Example 6.1.6.

Table 8. Expression from gene transfer systems comprising genes encoding two polypeptides linked by translational coupling elements. Gene transfer vectors comprised an enhancer (column E), promoter (column F), intron (column G) and polyadenylation signal (column I) operably linked to a gene encoding DasherGFP (rows 1-27 and 29) or CayenneRFP (row 28). For rows 1-27, vectors further comprised a gene encoding CayenneRFP operably linked to the expression control elements by a translational-coupling sequence (sequences identified in column H). Vectors were transfected into HEK293 cells (columns J-M) or CHO cells (columns N-Q), and expression of the fluorescent proteins measured as described in Example 6.1.6.

Table 9. Expression from gene transfer systems comprising genes encoding two polypeptides linked by translational coupling elements. Gene transfer vectors comprised an enhancer (column E), promoter (column F), intron (column G) and polyadenylation signal (column I) operably linked to a gene encoding DasherGFP (rows 1-20) or CayenneRFP (row 21). For rows 1-19, vectors further comprised a gene encoding CayenneRFP operably linked to the expression control elements by a translational-coupling sequence (sequences identified in column H). Vectors were transfected into CHO cells and expression of the fluorescent proteins measured (columns J-M) as described in Example 6.1.6.

Table 10. Expression from gene transfer systems comprising genes encoding two polypeptides linked by translational coupling elements. Gene transfer vectors comprised an enhancer (column E), promoter (column F), intron (column G) and polyadenylation signal (column I) operably linked to a gene encoding DasherGFP (rows 1-22) or CayenneRFP (row 23). For rows 1-21, vectors further comprised a gene encoding CayenneRFP operably linked to the expression control elements by a translational-coupling sequence (sequences identified in column H). Vectors were transfected into HEK293 cells (columns J-M) or CHO cells (columns N-Q), and expression of the fluorescent proteins measured as described in Example 6.1.6.

Table 11. Expression from gene transfer systems comprising genes encoding two polypeptides linked by translational coupling elements. Gene transfer vectors comprised an enhancer (column E), promoter (column F), intron (column G) and polyadenylation signal (column I) operably linked to a gene encoding DasherGFP (rows 1-12) or CayenneRFP (row 13). For rows 1-11, vectors further comprised a gene encoding CayenneRFP operably linked to the expression control elements by a translational-coupling sequence (sequences identified in column H). Vectors were transfected into CHO cells and expression of the fluorescent proteins measured (columns J-M) as described in Example 6.1.6.

Table 12. Expression of antibodies from gene transfer systems comprising genes encoding both antibody chains. Gene transfer vectors comprised an enhancer (column C), promoter (column D), intron (column E) and polyadenylation signal (column F) operably linked to a gene encoding Herceptin light chain. For rows 7-12 and 17, vectors further comprised a gene encoding Herceptin heavy chain operably linked to the expression control elements by a translational-coupling sequence (sequences identified in column G). For rows 1-6, vectors further comprised a gene encoding Herceptin heavy chain operably linked to a second enhancer (column I), a second promoter (column J), a second intron (column K) and a second polyadenylation signal (column L). Optionally an insulator sequence was interposed between the first polyadenylation signal and the second enhancer (column H). Vectors were transfected into HEK293 cells, and expression of the assembled secreted antibody proteins measured as described in Example 6.1.7. Vectors encoding the two chains separately were also co-transfected at 3 different ratios (rows 13-15).

Table 13. Expression of fluorescent proteins from a gene transfer system comprising a transposon and a transposase. Transposons comprised transposon end sequences SEQ ID NO. 9 and SEQ ID NO. 6, and an enhancer (column F), promoter (column G), intron (column H) and polyadenylation signal (column I) operably linked to a gene encoding DasherGFP. For rows 3-6, vectors further comprised a gene encoding CayenneRFP operably linked to the expression control elements by a translational-coupling sequence (sequences identified in column J). For rows 7-18, vectors

further comprised a gene encoding Cayenne RFP operably linked to a second enhancer (column L), a second promoter (column M), a second intron (column N) and a second polyadenylation signal (column O). Optionally an insulator sequence was interposed between the first polyadenylation signal and the second enhancer (column K). Transposons were transfected into CHO cells, optionally together with a gene encoding a transposase (SEQ ID NO. 45) fused to a heterologous nuclear localization signal, cells were selected and expression of the fluorescent proteins measured (columns Q-V) as described in Example 6.1.8. Rows 1-2 and 19-20 show the transfection of constructs encoding only GFP (rows 1-2) or RFP (rows 19-20). Rows 21 and 22 shows the co-transfection of the constructs shown in rows 1 and 19.

Table 14. Expression of a fluorescent protein from a gene transfer system comprising a transposon and a transposase. Transposons comprised transposon end sequences SEQ ID NO. 9 and SEQ ID NO. 6, an enhancer (column B), promoter (column C), intron (column D) and rabbit beta globin polyadenylation signal operably linked to a gene encoding DasherGFP, and optionally insulator sequences on either side of the Dasher expression cassette (column E). Transposons were transfected into CHO cells, optionally together with a gene encoding a transposase (column G) fused to a heterologous nuclear localization signal, cells were selected and expression of the fluorescent protein measured (columns H-J) as described in Example 6.1.9.

Table 15. Vector element combinations used in gene transfer vectors. Gene transfer vectors comprised a gene encoding DasherGFP operably linked to an enhancer (column B), a promoter (column C), an intron (column D), a 5'UTR (column E), a sequence to enhance RNA export (column F) and a polyadenylation signal (column G). Some vectors further comprised a viral replication origin (column H) and/or a gene encoding a viral replication protein (column I), as described in Example 6.1.10.

Table 16. Vector element combinations used in gene transfer vectors. Gene transfer vectors comprised a gene encoding DasherGFP operably linked to an enhancer (column B), a promoter (column C), an intron (column D), a 5'UTR (column E), a 3'UTR (column F), a sequence to enhance RNA export (column G) and a polyadenylation signal (column H). Some vectors further comprised a viral replication origin (column I) and/or a gene encoding a viral replication protein (column J), as described in Example 6.1.11.

Table 17. Vector element combinations used in gene transfer vectors. Gene transfer vectors comprised a gene encoding DasherGFP operably linked to an enhancer (column B), a promoter (column C), an intron (column D), a 5'UTR (column E), a sequence to enhance RNA export (column F) and a polyadenylation signal (column G). Vectors were transfected into CHO cells and DasherGFP measured (columns H-J), as described in Example 6.1.12.

Table 18. Vector element combinations used in gene transfer vectors. Transposons comprised transposon end sequences SEQ ID NO. 9 and SEQ ID NO. 6, an enhancer (column B), promoter (column C), intron (column D) and polyadenylation signal (column F) operably linked to a gene encoding DasherGFP, and optionally insulator sequences on either side of the Dasher expression cassette (column G). Transposons were transfected into CHO cells, cells were selected and expression of the fluorescent protein measured (columns H-J), as described in Example 6.1.13.

Table 19. Expression of a fluorescent protein from a gene transfer systems designed to test control element configurations. Gene transfer vectors configured as shown in Table

15 were tested for expression in HEK (D-E) and CHO (B-C) cells as described in Example 6.1.10. Average fluorescent counts from independent triplicate transfections are shown.

Table 20. Expression of a fluorescent protein from a gene transfer systems designed to test control element configurations. Gene transfer vectors configured as shown in Table 16 were tested for expression in HEK (D-E) and CHO (B-C) cells as described in Example 6.1.11. Average fluorescent counts from independent triplicate transfections are shown.

Table 21. Regression weights for vector elements used in transient expression in HEK and CHO cells. A model relating the elements of gene transfer vectors shown in Table 15, and the expression data shown in Table 19 was constructed using partial least square regression. The regression weights for each element or combination of elements calculated from the model for HEK and CHO expression systems is indicated.

TABLES

TABLE 1

Expression from gene transfer vectors comprising transposons and transposase.									
A	B	C	D	E	F	G	H	I	J
	Ratio	Transposon (Tn)			Transposase	TP NLS	GFP Expression		
Row	(Tn:TP)	Left end	Right end	Gene id. (TP)		fusion	1	2	3
1	N/A	SEQ ID NO: 30	SEQ ID NO: 31	133371	none	N/A	549	703	592
2	3:1	SEQ ID NO: 30	SEQ ID NO: 31	133371	SEQ ID NO: 57	no	5,279	4,421	4,528
3	5:1	SEQ ID NO: 30	SEQ ID NO: 31	133371	SEQ ID NO: 57	no	4,992	5,328	5,512
4	1:1	SEQ ID NO: 30	SEQ ID NO: 31	133371	SEQ ID NO: 57	no	3,717	3,910	3,056
5	N/A	SEQ ID NO: 1	SEQ ID NO: 2	133365	none	N/A	709	554	655
6	3:1	SEQ ID NO: 1	SEQ ID NO: 2	133365	SEQ ID NO: 44	no	3,193	3,052	3,981
7	5:1	SEQ ID NO: 1	SEQ ID NO: 2	133365	SEQ ID NO: 44	no	3,043	3,165	2,929
8	1:1	SEQ ID NO: 1	SEQ ID NO: 2	133365	SEQ ID NO: 44	no	3,848	3,641	3,664
9	N/A	SEQ ID NO: 5	SEQ ID NO: 6	134925	none	N/A	316	330	314
10	3:1	SEQ ID NO: 5	SEQ ID NO: 6	134925	SEQ ID NO: 46	yes	535	422	542
11	5:1	SEQ ID NO: 5	SEQ ID NO: 6	134925	SEQ ID NO: 46	yes	498	420	524
12	1:1	SEQ ID NO: 5	SEQ ID NO: 6	134925	SEQ ID NO: 46	yes	509	478	518
13	N/A	SEQ ID NO: 27	SEQ ID NO: 28	134924	none	N/A	582	640	554
14	3:1	SEQ ID NO: 27	SEQ ID NO: 28	134924	SEQ ID NO: 55	no	294	462	545
15	5:1	SEQ ID NO: 27	SEQ ID NO: 28	134924	SEQ ID NO: 55	no	606	489	466
16	1:1	SEQ ID NO: 27	SEQ ID NO: 28	134924	SEQ ID NO: 55	no	698	613	699

TABLE 2

Expression from gene transfer vectors comprising transposons and transposases.									
	C	D	E	F	G	H	I	J	
A	Ratio	Transposon (Tn)			Transposase	TP	GFP		
Row	(Tn:TP)	left end	right end	Gene id (TP)		fusion	(N = 3)	sd	
1	N/A	SEQ ID NO: 5	SEQ ID NO: 6	134925	none	N/A	172	N/A	
2	2:1	SEQ ID NO: 5	SEQ ID NO: 6	134925	SEQ ID NO: 45	no	204	1.19	
3	5:1	SEQ ID NO: 5	SEQ ID NO: 6	134925	SEQ ID NO: 45	no	281	1.63	
4	2:1	SEQ ID NO: 5	SEQ ID NO: 6	134925	SEQ ID NO: 46	no	184	1.07	
5	5:1	SEQ ID NO: 5	SEQ ID NO: 6	134925	SEQ ID NO: 46	no	161	0.94	
6	2:1	SEQ ID NO: 5	SEQ ID NO: 6	134925	SEQ ID NO: 45	yes	577	3.36	
7	5:1	SEQ ID NO: 5	SEQ ID NO: 6	134925	SEQ ID NO: 45	yes	483	2.61	
8	N/A	SEQ ID NO: 30	SEQ ID NO: 31	133371	none	N/A	288	N/A	
9	5:1	SEQ ID NO: 30	SEQ ID NO: 31	133371	SEQ ID NO: 57	no	3674	12.8	
10	N/A	SEQ ID NO: 24	SEQ ID NO: 25	134922	none	N/A	327	N/A	
11	2:1	SEQ ID NO: 24	SEQ ID NO: 25	134922	SEQ ID NO: 53	no	183	0.59	
12	5:1	SEQ ID NO: 24	SEQ ID NO: 25	134922	SEQ ID NO: 53	no	277	0.85	
13	N/A	SEQ ID NO: 10	SEQ ID NO: 11	133366	none	N/A	332	N/A	
14	2:1	SEQ ID NO: 10	SEQ ID NO: 11	133366	SEQ ID NO: 47	no	5	0.02	
15	5:1	SEQ ID NO: 10	SEQ ID NO: 11	133366	SEQ ID NO: 47	no	393	1.18	
16	N/A	SEQ ID NO: 12	SEQ ID NO: 13	133367	none	N/A	505	N/A	
17	2:1	SEQ ID NO: 12	SEQ ID NO: 13	133367	SEQ ID NO: 48	no	185	0.37	
18	5:1	SEQ ID NO: 12	SEQ ID NO: 13	133367	SEQ ID NO: 48	no	179	0.35	
19	N/A	SEQ ID NO: 22	SEQ ID NO: 23	134717	none	N/A	303	N/A	
20	2:1	SEQ ID NO: 22	SEQ ID NO: 23	134717	SEQ ID NO: 52	no	208	0.68	
21	5:1	SEQ ID NO: 22	SEQ ID NO: 23	134717	SEQ ID NO: 52	no	177	0.58	
22	N/A	SEQ ID NO: 14	SEQ ID NO: 15	133368	none	N/A	277	N/A	
23	2:1	SEQ ID NO: 14	SEQ ID NO: 15	133368	SEQ ID NO: 49	no	243	0.88	
24	5:1	SEQ ID NO: 14	SEQ ID NO: 15	133368	SEQ ID NO: 49	no	169	0.61	
25	N/A	SEQ ID NO: 16	SEQ ID NO: 17	133369	none	N/A	256	N/A	
26	2:1	SEQ ID NO: 16	SEQ ID NO: 17	133369	SEQ ID NO: 50	no	161	0.63	
27	5:1	SEQ ID NO: 16	SEQ ID NO: 17	133369	SEQ ID NO: 50	no	184	0.72	

TABLE 2-continued

Expression from gene transfer vectors comprising transposons and transposases.								
A	C	D	E	F	G	H	I	J
	Ratio	Transposon (Tn)			Transposase	TP	GFP	
Row	(Tn:TP)	left end	right end	Gene id (TP)		fusion	(N = 3)	sd
28	N/A	SEQ ID NO: 18	SEQ ID NO: 19	133370	none	N/A	248	N/A
29	2:1	SEQ ID NO: 18	SEQ ID NO: 19	133370	SEQ ID NO: 56	no	148	0.6
30	5:1	SEQ ID NO: 18	SEQ ID NO: 19	133370	SEQ ID NO: 56	no	133	0.54
31	N/A	SEQ ID NO: 20	SEQ ID NO: 21	134716	none	N/A	97	N/A
32	2:1	SEQ ID NO: 20	SEQ ID NO: 21	134716	SEQ ID NO: 51	no	67	0.69
33	5:1	SEQ ID NO: 20	SEQ ID NO: 21	134716	SEQ ID NO: 51	no	92	0.95
34	N/A	SEQ ID NO: 26	SEQ ID NO: 27	134923	none	N/A	319	N/A
35	2:1	SEQ ID NO: 26	SEQ ID NO: 27	134923	SEQ ID NO: 54	no	189	0.59
36	5:1	SEQ ID NO: 26	SEQ ID NO: 27	134923	SEQ ID NO: 54	no	270	0.85

TABLE 3

Expression from gene transfer vectors comprising transposons and transposases.									
A	B	C	D	E	F	G	H	I	J
Row	Transposon					Transposase	GFP Expression		
	left end	right end	Gene id	P_GFP	Insulator		1	2	3
1	SEQ ID NO: 5	SEQ ID NO: 6	147759	EF1a	no	none	203	188	218
2	SEQ ID NO: 5	SEQ ID NO: 6	147759	EF1a	no	SEQ ID NO: 45	640	638	661
3	SEQ ID NO: 5	SEQ ID NO: 6	181650	EF1a	HS4	none	784	865	838
4	SEQ ID NO: 5	SEQ ID NO: 6	181650	EF1a	HS4	SEQ ID NO: 45	3,924	3,643	4,645
5	SEQ ID NO: 1	SEQ ID NO: 2	194091	EF1a	no	none	660	638	746
6	SEQ ID NO: 1	SEQ ID NO: 2	194091	EF1a	no	SEQ ID NO: 44	70	81	111
7	SEQ ID NO: 1	SEQ ID NO: 2	194092	EF1a	HS4 core	none	552	520	534
8	SEQ ID NO: 1	SEQ ID NO: 2	194092	EF1a	HS4 core	SEQ ID NO: 44	3,001	2,830	3,236
9	SEQ ID NO: 1	SEQ ID NO: 2	194093	EF1a	HS4	none	968	1,034	1,238
10	SEQ ID NO: 1	SEQ ID NO: 2	194093	EF1a	HS4	SEQ ID NO: 44	4,601	4,445	5,530
11	SEQ ID NO: 1	SEQ ID NO: 2	194094	CMV	no	none	92	117	95
12	SEQ ID NO: 1	SEQ ID NO: 2	194094	CMV	no	SEQ ID NO: 44	815	931	915
13	SEQ ID NO: 1	SEQ ID NO: 2	194095	CMV	HS4 core	none	57	56	62
14	SEQ ID NO: 1	SEQ ID NO: 2	194095	CMV	HS4 core	SEQ ID NO: 44	684	852	663
15	SEQ ID NO: 1	SEQ ID NO: 2	194096	CMV	HS4	none	97	84	82
16	SEQ ID NO: 1	SEQ ID NO: 2	194096	CMV	HS4	SEQ ID NO: 44	1,327	1,196	1,257

TABLE 4

Expression from gene transfer vectors comprising transposons and transposases.														
A	B	C	D	E	F	G	H	I	J	K	L	M	N	O
	Transposon						Transposase							
	RNA						Gene	All-	GFP Expression					
Row	Left end	Right end	Gene id	P_GFP	element	Insulator	Transposase	id	in-one	P_TP	NLS	1	2	3
1	SEQ ID NO: 9	SEQ ID NO: 6	187151	CMV	HPRE	no	none	N/A	N/A	N/A	N/A	42	50	58
2	SEQ ID NO: 9	SEQ ID NO: 6	187151	CMV	HPRE	no	SEQ ID NO: 45	136651	no	CMV	yes	1,004	903	894
3	SEQ ID NO: 9	SEQ ID NO: 6	198993	CMV	HPRE	no	SEQ ID NO: 45	136651	yes	SV40	yes	511	506	523
4	SEQ ID NO: 9	SEQ ID NO: 6	181650	EF1a	HPRE	HS4	none	N/A	N/A	N/A	N/A	1,649	1,675	1,634
5	SEQ ID NO: 9	SEQ ID NO: 6	181650	EF1a	HPRE	HS4	SEQ ID NO: 45	136651	no	CMV	yes	3,849	3,935	4,226
6	SEQ ID NO: 9	SEQ ID NO: 6	197821	EF1a	HPRE	HS4	SEQ ID NO: 45	136651	yes	CMV	yes	2,287	2,482	2,263
7	SEQ ID NO: 9	SEQ ID NO: 6	198994	EF1a	HPRE	HS4	SEQ ID NO: 45	136651	yes	SV40	yes	2,475	2,340	2,604
8	SEQ ID NO: 30	SEQ ID NO: 31	133371	EF1a	WPRE	no	none	N/A	N/A	N/A	N/A	490	579	603
9	SEQ ID NO: 30	SEQ ID NO: 31	133371	EF1a	WPRE	no	SEQ ID NO: 57	133255	no	CMV	none	3,947	3,069	3,518
10	SEQ ID NO: 30	SEQ ID NO: 31	94097	EF1a	WPRE	no	SEQ ID NO: 57	133255	yes	CMV	none	5,314	4,133	4,830
11	SEQ ID NO: 1	SEQ ID NO: 2	194094	CMV	HPRE	no	none	N/A	N/A	N/A	N/A	57	56	49
12	SEQ ID NO: 1	SEQ ID NO: 2	194094	CMV	HPRE	no	SEQ ID NO: 44	136653	no	CMV	yes	1,461	1,412	1,443
13	SEQ ID NO: 1	SEQ ID NO: 2	198176	CMV	HPRE	no	SEQ ID NO: 44	136653	yes	CMV	yes	1,530	1,574	1,727
14	SEQ ID NO: 1	SEQ ID NO: 2	198997	CMV	HPRE	no	SEQ ID NO: 44	136653	yes	SV40	yes	932	1,058	1,035

TABLE 4-continued

Expression from gene transfer vectors comprising transposons and transposases.														
A	B	C	D	E	F	G	H	I	J	K	L	M	N	O
	Transposon						Transposase							
	RNA						Gene	All-	GFP Expression					
Row	Left end	Right end	Gene id	P_GFP	element	Insulator	Transposase	id	in-one	P_TP	NLS	1	2	3
15	SEQ ID NO: 1	SEQ ID NO: 2	194092	EF1a	HPRE	HS4 core	none	N/A	N/A	N/A	N/A	1,584	1,518	1,215
16	SEQ ID NO: 1	SEQ ID NO: 2	194092	EF1a	HPRE	HS4 core	SEQ ID NO: 44	136653	no	CMV	yes	3,343	3,482	3,427
17	SEQ ID NO: 1	SEQ ID NO: 2	198154	EF1a	HPRE	HS4 core	SEQ ID NO: 44	136653	yes	CMV	yes	4,732	3,481	3,543
18	SEQ ID NO: 1	SEQ ID NO: 2	198996	EF1a	HPRE	HS4 core	SEQ ID NO: 44	136653	yes	SV40	yes	2,056	2,147	2,258
19	SEQ ID NO: 1	SEQ ID NO: 2	194093	EF1a	HPRE	H54	none	N/A	N/A	N/A	N/A	1,185	1,252	1,560
20	SEQ ID NO: 1	SEQ ID NO: 2	194093	EF1a	HPRE	H54	SEQ ID NO: 44	136653	no	CMV	yes	4,348	4,605	6,072
21	SEQ ID NO: 1	SEQ ID NO: 2	197822	EF1a	HPRE	H54	SEQ ID NO: 44	136653	yes	CMV	yes	4,331	3,786	4,459
22	SEQ ID NO: 1	SEQ ID NO: 2	198995	EF1a	HPRE	H54	SEQ ID NO: 44	136653	yes	SV40	yes	864	1,089	844

TABLE 5

Expression from gene transfer vectors comprising transposons and transposases.										
A	B	C	D		E			H	I	J
	Ratio		Transposon (Tn)			F	G	GFP Expression		
Row	(Tn:TP)	left end	right end		Gene	Transposase (Tpase)	NLS	1	2	3
1	N/A	SEQ ID NO: 30	SEQ ID NO: 31		136214	none	N/A	15	13	16
2	5:1	SEQ ID NO: 30	SEQ ID NO: 31		136214	SEQ ID NO: 57	yes	78	76	83
3	N/A	SEQ ID NO: 5	SEQ ID NO: 6		192462	none	N/A	15	16	19
4	3:1	SEQ ID NO: 5	SEQ ID NO: 6		192462	SEQ ID NO: 45	yes	50	49	40
5	N/A	SEQ ID NO: 7	SEQ ID NO: 8		195739	none	N/A	15	18	18
6	3:1	SEQ ID NO: 7	SEQ ID NO: 8		195739	SEQ ID NO: 45	yes	46	56	54
7	N/A	SEQ ID NO: 1	SEQ ID NO: 2		192465	none	N/A	13	16	11
8	3:1	SEQ ID NO: 1	SEQ ID NO: 2		192465	SEQ ID NO: 44	yes	31	25	24
9	N/A	SEQ ID NO: 1	SEQ ID NO: 2		192465	SEQ ID NO: 43	yes	11	11	9
10	3:1	SEQ ID NO: 1	SEQ ID NO: 2		192465	SEQ ID NO: 44	no	36	41	41
11	N/A	SEQ ID NO: 24	SEQ ID NO: 25		192459	none	N/A	9	11	11
12	3:1	SEQ ID NO: 24	SEQ ID NO: 25		192459	SEQ ID NO: 53	yes	8	10	11
13	N/A	SEQ ID NO: 26	SEQ ID NO: 27		192460	none	N/A	8	12	12
14	3:1	SEQ ID NO: 26	SEQ ID NO: 27		192460	SEQ ID NO: 54	yes	12	10	12
15	N/A	SEQ ID NO: 20	SEQ ID NO: 21		192463	none	N/A	7	9	11
16	3:1	SEQ ID NO: 20	SEQ ID NO: 21		192463	SEQ ID NO: 51	yes	4	7	4
17	N/A	SEQ ID NO: 22	SEQ ID NO: 23		192464	none	N/A	15	16	11
18	3:1	SEQ ID NO: 22	SEQ ID NO: 23		192464	SEQ ID NO: 52	yes	8	11	7
19	N/A	SEQ ID NO: 10	SEQ ID NO: 11		192466	none	N/A	10	6	7
20	3:1	SEQ ID NO: 10	SEQ ID NO: 11		192466	SEQ ID NO: 47	yes	9	10	9
21	N/A	SEQ ID NO: 12	SEQ ID NO: 13		192467	none	N/A	8	12	12
22	3:1	SEQ ID NO: 12	SEQ ID NO: 13		192467	SEQ ID NO: 48	yes	9	13	9
23	N/A	SEQ ID NO: 14	SEQ ID NO: 15		192468	none	N/A	11	7	12
24	3:1	SEQ ID NO: 14	SEQ ID NO: 15		192468	SEQ ID NO: 49	yes	9	9	7
25	N/A	SEQ ID NO: 16	SEQ ID NO: 17		192469	none	N/A	13	13	15
26	3:1	SEQ ID NO: 16	SEQ ID NO: 17		192469	SEQ ID NO: 50	yes	9	8	10
27	N/A	SEQ ID NO: 18	SEQ ID NO: 19		192470	none	N/A	15	13	14
28	3:1	SEQ ID NO: 18	SEQ ID NO: 19		192470	SEQ ID NO: 56	yes	6	6	6
29	N/A	SEQ ID NO: 28	SEQ ID NO: 29		192461	none	N/A	16	16	11
30	3:1	SEQ ID NO: 28	SEQ ID NO: 29		192461	SEQ ID NO: 55	yes	5	10	9

TABLE 6

Expression from gene transfer vectors comprising transposons and transposases.										
		C	D		F		H	I	J	K
							No transposase		Plus transposase	
A	B	En-	Pro-	E	RNA	G	GFP Exp.		GFP Exp.	
Row	Construct	hancer	moter	Intron	export	polyA	(Mean)	sd	(Mean)	sd
1	145736	CMV	CMV	none	WPRE	BGH	1,046	63	6,986	737
2	145737	CMV	CMV	CMVa	WPRE	BGH	1,172	133	6,380	206
3	187151	CMV	CMV	none	HPRE	rabbit globin	1,378	80	7,243	382

Expression from gene transfer vectors comprising transposons and transposases.											
		C		D		F		H	I	J	K
								No transposase		Plus transposase	
A	B	En-	Pro-	E		RNA	G	GFP Exp.		GFP Exp.	
Row	Construct	hancer	moter	Intron		export	polyA	(Mean)	sd	(Mean)	sd
4	187152	CMV	CMV	CMVa		HPRE	rabbit globin	1,073	27	4,902	254
5	189858	CMV	CMV	synthetic (eMLP)		HPRE	rabbit globin	1,527	63	6,437	398

Expression from gene transfer vectors comprising genes encoding two polypeptides linked by translational coupling elements.																
									J	K	L	M	N	O	P	Q
									HEK293 cells				CHO cells			
A	B	C	D	E	F	G	H	I	Avg.		Avg.		Avg.		Avg.	
Row	Gene	GFP	RFP	En-hancer	Pro-moter	Intron	IRES/Other	PolyA	GFP (N = 3)	sd	RFP (N = 3)	sd	GFP (N = 3)	sd	RFP (N = 3)	sd
1	143090	yes	no	CMV	CMV	CMVc	none	rabbit	13,585	426	3	1	21,909	1,128	4	1
2	188552	no	yes	CMV	CMV	eMLP	none	rabbit	7	1	1,620	42	5	1	1,316	241
3	135171	yes	yes	CMV	CMV	none	SEQ ID NO: 101	BGH	1,064	54	293	11	548	108	168	13
4	186390	yes	yes	CMV	CMV	CMVc	SEQ ID NO: 75	rabbit	5,252	180	286	11	1,786	140	136	5
5	183439	yes	yes	CMV	CMV	CMVc	SEQ ID NO: 68	rabbit	3,687	329	249	26	12,618	712	170	3
6	186386	yes	yes	CMV	CMV	CMVc	SEQ ID NO: 64	rabbit	3,678	221	11	1	4,428	310	59	5
7	183432	yes	yes	CMV	CMV	CMVc	SEQ ID NO: 98	rabbit	3,515	290	69	9	7,656	1,229	250	28
8	180533	yes	yes	CMV	CMV	CMVc	SEQ ID NO: 61	rabbit	3,275	590	268	49	2,775	193	273	31
9	183431	yes	yes	CMV	CMV	CMVc	SEQ ID NO: 63	rabbit	2,788	250	186	17	1,760	428	163	42
10	180536	yes	yes	CMV	CMV	CMVc	SEQ ID NO: 62	rabbit	2,324	315	159	25	13,203	1,030	251	25
11	183437	yes	yes	CMV	CMV	CMVc	SEQ ID NO: 67	rabbit	1,940	200	195	25	9,170	781	272	20
12	186382	yes	yes	CMV	CMV	CMVc	SEQ ID NO: 74	rabbit	743	30	51	3	5,384	108	85	0
13	180530	yes	yes	CMV	CMV	CMVc	SEQ ID NO: 58	rabbit	1,714	173	72	11	1,660	232	93	12
14	183429	yes	yes	CMV	CMV	CMVc	SEQ ID NO: 59	rabbit	1,581	53	195	6	2,060	353	231	31
15	188326	yes	yes	CMV	CMV	none	SEQ ID NO: 99	SV40	1,425	86	88	14	6,901	484	158	7
16	188220	yes	yes	CMV	CMV	none	SEQ ID NO: 63	SV40	1,380	125	80	9	1,887	142	160	15
17	188697	yes	yes	CMV	CMV	none	SEQ ID NO: 100	SV40	1,363	36	6	1	6,081	1,338	97	29
18	188222	yes	yes	CMV	CMV	none	SEQ ID NO: 67	SV40	1,537	81	143	9	6,558	600	205	14
19	188221	yes	yes	CMV	CMV	none	SEQ ID NO: 59	SV40	2,788	165	326	19	4,217	180	491	25
20	188210	yes	yes	EF1a	EF1a	EF1a	SEQ ID NO: 67	rabbit	362	41	43	6	5,915	542	196	25
21	188208	yes	yes	EF1a	EF1a	EF1a	SEQ ID NO: 63	rabbit	269	44	19	3	2,044	372	154	39
22	188209	yes	yes	EF1a	EF1a	EF1a	SEQ ID NO: 59	rabbit	499	65	61	7	3,356	723	306	51

TABLE 8

Expression from gene transfer vectors comprising genes encoding two polypeptides linked by translational coupling elements.																
A Row	B Gene	C GFP	D RFP	E En- hancer	F Pro- moter	G Intron	H IRES/Other	I PolyA	J	K	L	M	N	O	P	Q
									HEK293 cells				CHO cells			
									Avg GFP (N = 3)	sd	Avg RFP (N = 3)	sd	Avg GFP (N = 3)	sd	Avg RFP (N = 3)	sd
1	188220	yes	no	CMV	CMV	None	SEQ ID NO: 63	SV40	6,259	227	400	14	152	44	13	3
2	188222	yes	yes	CMV	CMV	None	SEQ ID NO: 67	SV40	5,752	864	657	88	1,147	25	66	2
3	188223	yes	yes	CMV	CMV	None	SEQ ID NO: 68	SV40	8,322	1,051	611	48	1,826	385	39	3
4	188224	yes	yes	CMV	CMV	None	SEQ ID NO: 62	SV40	7,542	652	661	50	3,391	563	93	15
5	188225	yes	yes	CMV	CMV	None	SEQ ID NO: 61	SV40	4,551	111	377	10	774	75	63	6
6	188226	yes	yes	CMV	CMV	None	SEQ ID NO: 70	SV40	3,389	314	66	4	1,889	103	13	1
7	188227	yes	yes	CMV	CMV	None	SEQ ID NO: 71	SV40	2,415	180	29	2	1,246	22	4	0
8	188228	yes	yes	CMV	CMV	None	SEQ ID NO: 64	SV40	5,767	1,300	17	3	1,057	88	17	0
9	188326	yes	yes	CMV	CMV	None	SEQ ID NO: 99	SV40	5,227	436	395	25	2,446	311	89	11
10	188697	yes	yes	CMV	CMV	None	SEQ ID NO: 100	SV40	3,687	93	14	1	3,684	64	37	1
11	191353	yes	yes	CMV	CMV	None	SEQ ID NO: 94	SV40	5,681	273	630	48	1,850	131	93	18
12	191354	yes	yes	CMV	CMV	None	SEQ ID NO: 95	SV40	6,248	132	114	2	4,169	737	66	16
13	191355	yes	yes	CMV	CMV	None	SEQ ID NO: 96	SV40	5,315	834	108	18	2,024	27	18	2
14	191356	yes	yes	CMV	CMV	None	SEQ ID NO: 88	SV40	3,659	773	415	124	4,075	674	218	39
15	191357	yes	yes	CMV	CMV	None	SEQ ID NO: 97	SV40	3,428	420	7	1	3,062	223	5	1
16	191435	yes	yes	CMV	CMV	None	SEQ ID NO: 73	SV40	4,648	451	835	54	2,223	21	207	2
17	191436	yes	yes	CMV	CMV	None	SEQ ID NO: 75	SV40	10,885	549	599	42	1,915	103	143	4
18	191437	yes	yes	CMV	CMV	None	SEQ ID NO: 76	SV40	8,745	662	262	19	1,742	86	75	4
19	191438	yes	yes	CMV	CMV	None	SEQ ID NO: 77	SV40	4,764	409	593	61	3,587	149	203	1
20	191439	yes	yes	CMV	CMV	None	SEQ ID NO: 78	SV40	5,242	640	555	77	3,229	338	163	19
21	191440	yes	yes	CMV	CMV	None	SEQ ID NO: 79	SV40	5,381	413	175	16	3,358	246	53	8
22	191441	yes	yes	CMV	CMV	None	SEQ ID NO: 80	SV40	5,277	723	155	21	4,136	1,024	78	18
23	191442	yes	yes	CMV	CMV	None	SEQ ID NO: 74	SV40	4,447	261	321	17	4,219	572	138	21
24	191433	yes	yes	CMV	CMV	None	SEQ ID NO: 58	SV40	8,177	606	406	11	2,082	138	101	6
25	188221	yes	yes	CMV	CMV	None	SEQ ID NO: 59	SV40	8,144	979	1,110	98	228	95	29	12
26	189939	yes	yes	CMV	CMV	None	SEQ ID NO: 101	SV40	3,710	638	1,407	223	1,326	114	466	39
27	189940	yes	yes	CMV	CMV	None	SEQ ID NO: 101	SV40	3,049	111	911	41	962	34	260	14
28	135149	no	yes	CMV	CMV	None	none	BGH	8	2	1,398	50	10	6	133	9
29	136032	yes	no	CMV	CMV	None	none	SV40	16,336	416	3	1	4,879	270	1	1

TABLE 9

Expression from gene transfer vectors comprising genes encoding two polypeptides linked by translational coupling elements.													
A Row	B Gene	C GFP	D RFP	E Enhancer	F Promoter	G Intron	H IRES/Other	I PolyA	CHO cells				
									J Avg GFP (N = 3)	K sd	L Avg RFP (N = 3)	M sd	
1	188220	yes	yes	CMV	CMV	None	SEQ ID NO: 63	SV40	1,971	149	145	8	
2	188222	yes	yes	CMV	CMV	None	SEQ ID NO: 67	SV40	5,448	151	252	36	
3	188223	yes	yes	CMV	CMV	None	SEQ ID NO: 68	SV40	6,338	332	93	7	
4	188224	yes	yes	CMV	CMV	None	SEQ ID NO: 62	SV40	4,817	139	99	3	
5	188225	yes	yes	CMV	CMV	None	SEQ ID NO: 92	SV40	1,360	100	112	18	
6	188326	yes	yes	CMV	CMV	None	SEQ ID NO: 99	SV40	6,534	940	228	29	
7	188697	yes	yes	CMV	CMV	None	SEQ ID NO: 100	SV40	6,221	321	67	4	
8	191353	yes	yes	CMV	CMV	None	SEQ ID NO: 94	SV40	5,785	1,222	303	31	
9	191356	yes	yes	CMV	CMV	None	SEQ ID NO: 88	SV40	6,700	1,054	309	51	
10	191433	yes	yes	CMV	CMV	None	SEQ ID NO: 58	SV40	1,863	162	79	8	
11	191435	yes	yes	CMV	CMV	None	SEQ ID NO: 73	SV40	5,740	459	467	39	
12	191436	yes	yes	CMV	CMV	None	SEQ ID NO: 75	SV40	2,809	363	176	15	
13	191437	yes	yes	CMV	CMV	None	SEQ ID NO: 76	SV40	3,328	643	130	21	
14	191438	yes	yes	CMV	CMV	None	SEQ ID NO: 77	SV40	7,817	847	336	25	
15	191439	yes	yes	CMV	CMV	None	SEQ ID NO: 78	SV40	7,492	87	280	9	
16	191440	yes	yes	CMV	CMV	None	SEQ ID NO: 79	SV40	5,242	135	63	5	
17	191442	yes	yes	CMV	CMV	None	SEQ ID NO: 74	SV40	5,065	432	122	10	
18	188221	yes	yes	CMV	CMV	None	SEQ ID NO: 59	SV40	4,838	275	492	35	
19	189939	yes	yes	CMV	CMV	None	SEQ ID NO: 101	SV40	1,755	70	583	65	
20	136032	yes	no	CMV	CMV	None	none	SV40	8,170	694	2	0	
21	191432	no	yes	CMV	CMV	None	none	SV40	9	1	915	85	

TABLE 10

Expression from gene transfer vectors comprising genes encoding two polypeptides linked by translational coupling elements.																	
A Row	B Gene	C GFP	D RFP	E En- hancer	F Pro- moter	G Intron	H IRES/Other	I PolyA	HEK293 cells				CHO cells				
									J Avg GFP (N = 3)	K sd	L Avg RFP (N = 3)	M sd	N Avg GFP (N = 3)	O stdv	P Avg RFP (N = 3)	Q sd	
1	188220	yes	yes	CMV	CMV	None	SEQ ID NO: 63	SV40	7,076	337	475	12	1,927	143	160	13	
2	188222	yes	yes	CMV	CMV	None	SEQ ID NO: 67	SV40	4,915	872	528	99	5,552	558	182	20	
3	188223	yes	yes	CMV	CMV	None	SEQ ID NO: 68	SV40	8,119	1,600	703	120	4,245	232	109	5	
4	188224	yes	yes	CMV	CMV	None	SEQ ID NO: 62	SV40	5,101	851	539	94	4,406	1,929	119	13	
5	188225	yes	yes	CMV	CMV	None	SEQ ID NO: 92	SV40	5,122	547	422	40	1,084	44	95	5	
6	188326	yes	yes	CMV	CMV	None	SEQ ID NO: 99	SV40	3,916	692	278	56	5,147	699	132	20	
7	191353	yes	yes	CMV	CMV	None	SEQ ID NO: 94	SV40	5,769	76	598	16	6,647	818	209	34	
8	191356	yes	yes	CMV	CMV	None	SEQ ID NO: 88	SV40	4,501	231	447	25	6,001	105	218	12	
9	191435	yes	yes	CMV	CMV	None	SEQ ID NO: 73	SV40	5,105	754	800	126	3,423	214	278	12	
10	191436	yes	yes	CMV	CMV	None	SEQ ID NO: 75	SV40	8,299	2,248	533	153	1,270	122	88	8	
11	101438	yes	yes	CMV	CMV	None	SEQ ID NO: 77	SV40	5,539	462	618	61	3,346	416	119	20	
12	195055	yes	yes	CMV	CMV	None	SEQ ID NO: 81	SV40	4,596	57	493	12	4,730	714	164	33	
13	195056	yes	yes	CMV	CMV	None	SEQ ID NO: 82	SV40	4,752	474	492	40	6,161	395	228	15	
14	195057	yes	yes	CMV	CMV	None	SEQ ID NO: 83	SV40	3,883	821	408	92	4,952	243	185	9	
15	195063	yes	yes	CMV	CMV	None	SEQ ID NO: 84	SV40	9,428	1,292	626	95	2,619	92	198	8	
16	195064	yes	yes	CMV	CMV	None	SEQ ID NO: 85	SV40	6,625	696	569	56	1,809	16	181	3	

TABLE 10-continued

Expression from gene transfer vectors comprising genes encoding two polypeptides linked by translational coupling elements.																
A Row	B Gene	C GFP	D RFP	E En- hancer	F Pro- moter	G Intron	H IRES/Other	I PolyA	J HEK293 cells				M CHO cells			
									Avg GFP (N = 3)	sd	Avg RFP (N = 3)	sd	Avg GFP (N = 3)	stdv	Avg RFP (N = 3)	sd
17	195065	yes	yes	CMV	CMV	None	SEQ ID NO: 86	SV40	7,983	1,660	394	66	3,470	314	217	19
18	195066	yes	yes	CMV	CMV	None	SEQ ID NO: 87	SV40	6,951	863	270	35	2,211	77	92	2
19	191433	yes	yes	CMV	CMV	None	SEQ ID NO: 58	SV40	6,599	827	377	42	2,068	9	109	1
20	188221	yes	yes	CMV	CMV	None	SEQ ID NO: 59	SV40	8,283	1,439	1,082	243	3,045	192	342	27
21	189939	yes	yes	CMV	CMV	None	SEQ ID NO: 101	SV40	3,916	554	1,588	231	1,202	63	410	18
22	136032	yes	no	CMV	CMV	None	none	SV40	12,110	1,502	2	1	7,378	244	2	0
23	191432	no	yes	CMV	CMV	None	none	SV40	5	0	898	72	34	17	890	35

TABLE 11

Expression from gene transfer vectors comprising genes encoding two polypeptides linked by translational coupling elements.													
A Row	B Gene	C GFP	D RFP	E Enhancer	F Promoter	G Intron	H IRES/Other	I PolyA	J Avg GFP (N = 3)	K stdv	L Avg RFP (N = 3)	M stdv	
1	188224	yes	yes	CMV	CMV	None	SEQ ID NO: 62	SV40	4,755	220	88	7	
2	191435	yes	yes	CMV	CMV	None	SEQ ID NO: 73	SV40	3,195	176	219	12	
3	191436	yes	yes	CMV	CMV	None	SEQ ID NO: 93	SV40	1,370	198	76	5	
4	195055	yes	yes	CMV	CMV	None	SEQ ID NO: 81	SV40	3,995	128	105	4	
5	195056	yes	yes	CMV	CMV	None	SEQ ID NO: 82	SV40	3,038	608	83	15	
6	195063	yes	yes	CMV	CMV	None	SEQ ID NO: 84	SV40	1,679	23	105	2	
7	195064	yes	yes	CMV	CMV	None	SEQ ID NO: 85	SV40	1,234	19	96	1	
8	201084	yes	yes	CMV	CMV	None	SEQ ID NO: 89	SV40	3,494	250	234	18	
9	201085	yes	yes	CMV	CMV	None	SEQ ID NO: 90	SV40	3,221	92	204	10	
10	201086	yes	yes	CMV	CMV	None	SEQ ID NO: 91	SV40	3,489	745	260	59	
11	188221	yes	yes	CMV	CMV	None	SEQ ID NO: 59	SV40	2,257	109	236	13	
12	189939	yes	yes	CMV	CMV	None	SEQ ID NO: 101	SV40	1,190	103	355	6	
13	136032	yes	no	CMV	CMV	None	none	SV40	4,789	1,049	2	0	

TABLE 12

Expression of antibodies from gene transfer vectors comprising genes encoding both antibody chains.														
A	B	C Enhancer	D Promoter	E Intron	F	G	H Intergenic	I Enhancer	J Promoter	K Intron	L pA	M	N	O
Row	Gene(s)	1	1	1	polyA1	IRES	insulators	2	2	2	2	1	2	3
1	201223	CMV	Actin	Actin	HSV-TK	N/A	no	CMV	CMV	CMVc	BGH	173	138	133
2	201224	CMV	Actin	Actin	HSV-TK	N/A	no	CMV	CMV	none	BGH	158	91	137
3	201225	CMV	Actin	Actin	HSV-TK	N/A	no	CMV	GAPDH	eMLP	BGH	44	33	36
4	201226	CMV	Actin	Actin	HSV-TK	N/A	2x HS4 core	N/A	EF1a	EF1a	BGH	48	31	28
5	201227	CMV	Actin	Actin	HSV-TK	N/A	2x HS4 core	CMV	GAPDH	GAPDH	BGH	136	118	130
6	201228	CMV	Actin	Actin	HSV-TK	N/A	2x HS4 core	CMV	CMV	none	BGH	138	139	100
7	201238	CMV	Actin	Actin	BGH	SEQ ID NO: 68	no	N/A	N/A	N/A	none	31	30	23
8	201239	CMV	Actin	Actin	BGH	SEQ ID NO: 62	no	N/A	N/A	N/A	none	9	7	11
9	201240	CMV	Actin	Actin	BGH	SEQ ID NO: 73	no	N/A	N/A	N/A	none	129	91	108

TABLE 12-continued

Expression of antibodies from gene transfer vectors comprising genes encoding both antibody chains.														
A	B	C Enhancer	D Promoter	E Intron	F	G	H Intergenic	I Enhancer	J Promoter	K Intron	L pA	M N O Ab (µg/ml)		
Row	Gene(s)	1	1	1	polyA1	IRES	insulators	2	2	2	2	1	2	3
10	201241	CMV	Actin	Actin	BGH	SEQ ID NO: 93	no	N/A	N/A	N/A	none	61	39	54
11	201242	CMV	Actin	Actin	BGH	SEQ ID NO: 78	no	N/A	N/A	N/A	none	57	47	73
12	201243	CMV	Actin	Actin	BGH	SEQ ID NO: 86	no	N/A	N/A	N/A	none	21	19	25
13	145409 & 145443	CMV	Actin	Actin	BGH	N/A	no	N/A	N/A	N/A	N/A	94	131	122
14	145409 & 145443	CMV	Actin	Actin	BGH	N/A	no	N/A	N/A	N/A	N/A	17	19	17
15	145409 & 145443	CMV	Actin	Actin	BGH	N/A	no	N/A	N/A	N/A	N/A	110	98	147
16	150153	CMV	Actin	Actin	glob (rabbit)	N/A	no	CMV	CMV	CMVc	BGH	137	157	189
17	150154	CMV	Actin	Actin	BGH	SEQ ID NO: 59	no	N/A	N/A	N/A	none	191	196	295

TABLE 13

Expression of fluorescent proteins from a gene transfer system comprising a transposon and a transposase.											
Transposon											
A Row	Gene id 1	Gene id 2	D GFP	E RFP	F Enhancer 1	G Promoter 1	H Intron 1	I polyA1	J IRES	K intergenic insulator	L Enhancer 2
1	187151	N/A	yes	no	CMV	CMV	none	(rabbit)	N/A	N/A	N/A
2	187151	N/A	yes	no	CMV	CMV	none	(rabbit)	N/A	N/A	N/A
3	188209	N/A	yes	yes	none	EF1a	EF1a	(rabbit)	SEQ ID NO: 59	N/A	none
4	188209	N/A	yes	yes	none	EF1a	EF1a	(rabbit)	SEQ ID NO: 59	N/A	none
5	188219	N/A	yes	yes	none	EF1a	EF1a	(rabbit)	SEQ ID NO: 73	N/A	none
6	188219	N/A	yes	yes	none	EF1a	EF1a	(rabbit)	SEQ ID NO: 73	N/A	none
7	198833	N/A	yes	yes	CMV	CMV	none	HSV-TK	none	none	CMV
8	198833	N/A	yes	yes	CMV	CMV	none	HSV-TK	none	none	CMV
9	198834	N/A	yes	yes	CMV	CMV	none	HSV-TK	none	none	CMV
10	198834	N/A	yes	yes	CMV	CMV	none	HSV-TK	none	none	CMV
11	198835	N/A	yes	yes	CMV	CMV	none	HSV-TK	none	none	CMV
12	198835	N/A	yes	yes	CMV	CMV	none	HSV-TK	none	none	CMV
13	198836	N/A	yes	yes	CMV	CMV	none	HSV-TK	none	core	none
14	198836	N/A	yes	yes	CMV	CMV	none	HSV-TK	none	core	none
15	198837	N/A	yes	yes	CMV	CMV	none	HSV-TK	none	core	CMV
16	198837	N/A	yes	yes	CMV	CMV	none	HSV-TK	none	core	CMV
17	198838	N/A	yes	yes	CMV	CMV	none	HSV-TK	none	core	CMV
18	198838	N/A	yes	yes	CMV	CMV	none	HSV-TK	none	core	CMV
19	200967	N/A	no	yes	CMV	CMV	none	(rabbit)	none	N/A	N/A
20	200967	N/A	no	yes	CMV	CMV	none	(rabbit)	none	N/A	N/A
21	187151	200967	yes	yes	CMV	CMV	none	(rabbit)	N/A	N/A	N/A
22	187151	200967	yes	yes	CMV	CMV	none	(rabbit)	N/A	N/A	N/A

M N O Transposon			Q R S			T	U	V		
A	Promoter	Intron	P	GFP Expression			RFP Expression			
Row	2	2	pA2	Transposase	1	2	3	1	2	3
1	N/A	N/A	N/A	none	70	66	65	2	2	2
2	N/A	N/A	N/A	45	1,250	1,083	1,330	1	2	1
3	none	none	none	none	706	660	698	62	60	66
4	none	none	none	45	6,764	4,922	5,238	643	467	480
5	none	none	none	none	307	370	375	32	38	36
6	none	none	none	45	3,656	4,019	4,243	407	452	474
7	CMV	CMVc	(rabbit)	none	20	17	17	15	12	17
8	CMV	CMVc	(rabbit)	45	87	94	99	113	120	126
9	CMV	none	(rabbit)	none	19	22	21	9	10	10
10	CMV	none	(rabbit)	45	152	128	141	64	56	62

TABLE 13-continued

Expression of fluorescent proteins from a gene transfer system comprising a transposon and a transposase.											
	11	GAPDH	eMLP	(rabbit)	none	26	32	27	17	17	18
	12	GAPDH	eMLP	(rabbit)	45	272	231	222	306	257	237
	13	EF1a	EF1a	(rabbit)	none	38	39	36	104	94	98
	14	EF1a	EF1a	(rabbit)	45	320	374	449	1,102	1,245	1,471
	15	GAPDH	H	(rabbit)	none	67	55	55	58	45	42
	16	GAPDH	H	(rabbit)	45	396	470	411	418	483	425
	17	CMV	none	(rabbit)	none	25	27	22	11	13	10
	18	CMV	none	(rabbit)	45	280	260	245	122	118	104
	19	N/A	N/A	N/A	none	5	5	4	4	10	11
	20	N/A	N/A	N/A	45	5	6	6	375	389	392
	21	N/A	N/A	N/A	none	34	33	35	7	8	8
	22	N/A	N/A	N/A	45	546	583	628	186	196	197

TABLE 14

Expression of a fluorescent protein from a gene transfer system comprising a transposon and a transposase.									
A	B	C	D	E	F	G	H	I	J
							DasherGFP		
Row	Enhancer 1	Promoter 1	Intron 1	HS4 insulators	Gene	Transposase	1	2	3
1	none	EF1a	EF1a	no	147759	none	283	415	373
2	none	EF1a	EF1a	no	147759	SEQ ID NO: 45	865	846	500
3	none	EF1a	EF1a	yes	181650	none	858	980	944
4	EF1a	EF1a	EF1a	yes	181650	SEQ ID NO: 45	3,147	2,878	2,565
5	CMV	CMV	none	no	187151	none	59	42	42
6	CMV	CMV	none	no	187151	SEQ ID NO: 45	891	868	902
7	CMV	CMV	CMVa	no	187152	none	70	94	86
8	CMV	CMV	CMVa	no	187152	SEQ ID NO: 45	1,157	1,239	946
9	CMV	Actin	Actin	no	187153	none	43	48	46
10	CMV	Actin	Actin	no	187153	SEQ ID NO: 45	845	974	833
11	CMV	EF1a	EF1a	no	187154	none	430	497	385
12	CMV	EF1a	EF1a	no	187154	SEQ ID NO: 45	905	1,046	1,143
13	CMV	GAPDH	GAPDH	yes	189262	none	230	212	215
14	CMV	GAPDH	GAPDH	yes	189262	SEQ ID NO: 45	525	776	699
15	CMV	Actin	Actin	yes	189305	none	232	217	254
16	CMV	Actin	Actin	yes	189305	SEQ ID NO: 45	900	828	822
17	CMV	EF1a	EF1a	yes	189306	none	582	547	594
18	CMV	EF1a	EF1a	yes	189306	SEQ ID NO: 45	1,166	1,239	1,501
19	CMV	GAPDH	eMLP	yes	189855	none	350	273	347
20	CMV	GAPDH	EF1a	yes	189855	SEQ ID NO: 45	890	1,121	513
21	eMLP	EF1a	EF1a + eMLP	no	189856	none	388	221	349
22	eMLP	EF1a	EF1a + eMLP	no	189856	SEQ ID NO: 45	796	661	807
23	CMV	CMV	EF1a	no	189858	none	57	57	55
24	CMV	CMV	EF1a	no	189858	SEQ ID NO: 45	794	1,228	1,493
25	CMV	EF1a	EF1a + eMLP	yes	189859	none	602	329	695
26	CMV	EF1a	EF1a + eMLP	yes	189859	SEQ ID NO: 45	1,578	1,275	1,144
27	CMV	Actin	Actin + eMLP	no	189860	none	183	169	185
28	CMV	Actin	Actin + eMLP	no	189860	SEQ ID NO: 45	460	491	436
29	none	PGK	none	no	192462	none	7	7	6
30	none	PGK	none	no	192462	SEQ ID NO: 45	443	687	484

TABLE 15

Vector element combinations used in gene transfer vectors.								
A	B	C	D	E	F	G	H	I
Construct	Enhancer	Promoter	Intron	5' UTR	RNA export	polyA	viral ori	viral rep prot
128975	CMV	Actin	Chick actin (partial)	none	none	sv40 late	none	none
128986	CMV	CMV	none	none	WPRE	GH-bovine	oriP	EBNA
129966	CMV	CMV	none	none	none	GH-bovine	none	none
128978	CMV	CMV	none	none	none	sv40 late	none	none
128985	CMV	CMV	CMV intron c	none	none	sv40 late	none	none
129091	CMV	CMV	none	none	WPRE	GH-bovine	oriP	none
133139	CMV	GAPDH	GAPDH	none	none	sv40 late	none	none
136024	CMV	CMV	none	none	none	synthetic polyA	SV40	SV40 T
136025	CMV	Chick actin	Chick actin/rabbit	none	none	GH-bovine	SV40	SV40 T
136026	CMV	Chick actin	Chick actin/rabbit	none	none	sv40 early	oriP	none

TABLE 15-continued

Vector element combinations used in gene transfer vectors.								
A Construct	B Enhancer	C Promoter	D Intron	E 5' UTR	F RNA export	G polyA	H viral ori	I viral rep prot
136027	CMV	Chick actin	Chick actin/rabbit	none	none	sv40 late	none	none
136028	CMV	Chick actin	Chick actin (partial)	none	none	beta globin-human	SV40	SV40 T
136029	CMV	Chick actin	Chick actin (partial)	none	none	beta globin-rabbit	none	none
136030	CMV	Chick actin	Chick actin (partial)	none	none	sv40 early	oriP	none
136031	CMV	CMV	none	none	none	beta globin-rabbit	oriP	EBNA
136032	CMV	CMV	none	none	none	sv40 early	none	none
136033	CMV	CMV	CMV intron a	CMV	none	beta globin-rabbit	oriP	none
136034	CMV	CMV	CMV intron a	CMV	none	GH-bovine	oriP	EBNA
136035	CMV	CMV	CMV intron a	CMV	none	sv40 late	SV40	SV40 T
136036	CMV	CMV	CMV intron a	CMV	none	synthetic polyA	none	none
136037	CMV	CMV	CMV intron c	none	none	beta globin-human	none	none
136038	CMV	CMV	CMV intron c	none	none	HSV-TK	oriP	EBNA
136039	CMV	CMV	CMV intron c	none	none	sv40 late	oriP	none
136040	CMV	CMV	CMV intron c	none	none	synthetic polyA	SV40	SV40 T
136041	CMV	EF1a	EF1a_v1	none	none	beta globin-rabbit	SV40	SV40 T
136042	CMV	EF1a	EF1a_v1	none	none	HSV-TK	oriP	none
136043	CMV	EF1a	EF1a_v1	none	none	sv40 early	none	none
136044	CMV	EF1a	EF1a_v1	none	none	sv40 late	oriP	EBNA
136045	CMV	EF1a	EF1a_v2	none	none	beta globin-human	oriP	none
136046	CMV	EF1a	EF1a_v2	none	none	beta globin-rabbit	SV40	SV40 T
136047	CMV	EF1a	EF1a_v2	none	none	GH-bovine	oriP	EBNA
136048	CMV	EF1a	EF1a_v2	none	none	HSV-TK	none	none
136049	CMV	EF1a	EF1a_hybrid	none	none	GH-bovine	oriP	none
136050	CMV	EF1a	EF1a_hybrid	none	none	HSV-TK	SV40	SV40 T
136051	CMV	EF1a	EF1a_hybrid	none	none	synthetic polyA	oriP	EBNA
136052	CMV	GAPDH	GAPDH	none	none	beta globin-human	oriP	none
136053	CMV	GAPDH	GAPDH	none	none	beta globin-rabbit	none	none
136054	CMV	GAPDH	GAPDH	none	none	HSV-TK	SV40	SV40 T
136055	CMV	GAPDH	GAPDH	none	none	sv40 late	oriP	EBNA
136056	none	HSV-TK	none	none	none	beta globin-human	oriP	EBNA
136057	none	HSV-TK	none	none	none	GH-bovine	none	none
136058	none	HSV-TK	none	none	none	sv40 late	oriP	none
136059	none	MC1	none	none	none	GH-bovine	oriP	none
136060	none	MC1	none	none	none	HSV-TK	oriP	EBNA
136061	none	MC1	none	none	none	sv40 early	SV40	SV40 T
136062	none	PGK	none	none	none	sv40 early	oriP	EBNA
136063	none	PGK	none	none	none	sv40 late	none	none
136064	none	PGK	none	none	none	synthetic polyA	oriP	none
136065	E_SV40	SV40	none	none	none	beta globin-human	none	none
136066	E_SV40	SV40	none	none	none	HSV-TK	oriP	none
136067	E_SV40	SV40	none	none	none	sv40 early	SV40	SV40 T
136068	E_SV40	SV40	none	none	none	synthetic polyA	oriP	EBNA
136069	none	Ub-B	none	none	none	beta globin-rabbit	oriP	EBNA
136070	none	Ub-B	none	none	none	GH-bovine	SV40	SV40 T
136071	none	Ub-B	none	none	none	synthetic polyA	none	none
128979	CMV	CMV	none	none	WPRE	sv40 early	none	none
128980	CMV	CMV	none	none	WPRE	beta globin-human	none	none
128986	CMV	CMV	none	none	WPRE	GH-bovine	oriP	EBNA
129091	CMV	CMV	none	none	WPRE	GH-bovine	oriP	none
128977	CMV	CMV	Synthetic	none	none	sv40 early	none	none
133528	CMV	CMV	none	none	none	GH-bovine	SV40	none
134746	CMV	CMV	none	none	WPRE	HSV-TK	SV40	none

TABLE 16

Vector element combinations used in gene transfer vectors.									
A Construct	B Enhancer	C Promoter	D Intron	E 5' UTR	F 3' UTR	G RNA export	H polyA	I viral ori	J viral rep prot
143088	CMV	CMV	none	CMV	none	AGS	HSV-TK	oriP	EBNA
143089		EF1a	EF1a	none	none	WPRE	HSV-TK	oriP	none
143090	CMV	CMV	CMVc	CMV	none	none	Globin-rabbit	none	none
143091	CMV	CMV	CMVc	CMV_sTNV	sTNV	none	GH-bovine	oriP	none
143092	CMV	GAPDH	GAPDH	none	none	AGS	Globin-rabbit	none	none
143093	CMV	CMV	CMVa	bglob-Hs	bglob-XI	AGS	GH-bovine	oriP	EBNA
143094	synthetic	EF1a_LTR-HTLV	none	polyhedrin	polyhedrin	none	HSV-TK	none	none
143095	CMV	CMV	none	none	none	none	GH-bovine_gastrin	none	none
143096	CMV	GAPDH	Chick actin/rabbit	none	none	none	Globin-rabbit_gastrin	none	none

TABLE 16-continued

Vector element combinations used in gene transfer vectors.									
A Construct	B Enhancer	C Promoter	D Intron	E 5' UTR	F 3' UTR	G RNA export	H polyA	I viral ori	J viral rep prot
143097	CMV	CMV	CMVa	CMV_sTNV	sTNV	none	GH-bovine	none	none
143098	CMV	CMV	none	TNV	TNV	WPRE	HSV-TK	none	none
143099	CMV	CMV	none	CMV_TNV	TNV	none	GH-bovine_gastrin	SV40	SV40 T
143100	CMV	GAPDH	Chick actin/rabbit	none	none	none	GH-bovine	oriP	none
143101	CMV	CMV	Chick actin/rabbit	none	none	AGS	HSV-TK	none	none
143102	EF1a	EF1a_LTR-HTLV	none	none	none	none	HSV-TK	oriP	EBNA
143103	none	EF1a_LTR-HTLV	none	none	none	AGS	Globin-rabbit	oriP	EBNA
143104	EF1a	EF1a_LTR-HTLV	none	TNV	TNV	AGS	HSV-TK	oriP	none
143105	CMV	GAPDH	GAPDH	none	none	none	Globin-rabbit	SV40	SV40 T
143106	none	EF1a	EF1a	bglob-Hs	bglob-XI	none	HSV-TK	SV40	SV40 T
143107	CMV	GAPDH	Chick actin/rabbit	none	none	none	HSV-TK	none	none
143108	CMV	GAPDH	GAPDH	BYDV	BYDV	none	HSV-TK	SV40	SV40 T
143109	CMV	GAPDH	Chick actin/rabbit	none	none	WPRE	Globin-rabbit	SV40	SV40 T
143110	CMV	CMV	none	none	none	none	HSV-TK	SV40	SV40 T
143111	none	EF1a	EF1a	none	none	none	Globin-rabbit_gastrin	SV40	SV40 T
143112	none	EF1a_LTR-HTLV	none	BYDV	BYDV	AGS	Globin-rabbit	oriP	EBNA
143113	CMV	CMV	Chick actin/rabbit	bglob-XI	bglob-XI	none	HSV-TK	oriP	none
143114	CMV	GAPDH	GAPDH	none	none	none	HSV-TK	oriP	none
143115	none	EF1a_LTR-HTLV	none	TNV	TNV	WPRE	GH-bovine	oriP	EBNA
143116	none	EF1a_LTR-HTLV	none	bglob-Hs	bglob-XI	none	GH-bovine	none	none
143117	CMV	GAPDH	GAPDH	sTNV	sTNV	none	Globin-rabbit	oriP	EBNA
143118	none	EF1a_LTR-HTLV	none	bglob-XI	bglob-XI	none	GH-bovine	none	none
143119	synthetic	EF1a_LTR-HTLV	none	sTNV	sTNV	AGS	GH-bovine	SV40	SV40 T
143120	none	EF1a_LTR-HTLV	none	none	none	none	Globin-rabbit	oriP	none
143121	none	EF1a	EF1a	TNV	TNV	AGS	GH-bovine	none	none
143122	CMV	CMV	CMVa	CMV	none	none	GH-bovine	oriP	none
143123	none	EF1a_LTR-HTLV	none	polyhedrin	polyhedrin	WPRE	GH-bovine	SV40	SV40 T
143124	CMV	CMV	CMVc	CMV	none	none	GH-bovine	oriP	EBNA
143125	CMV	CMV	CMVc	CMV_bglob-XI	bglob-XI	AGS	HSV-TK	oriP	EBNA
143126	synthetic	EF1a_LTR-HTLV	none	bglob-Hs	bglob-XI	AGS	GH-bovine	SV40	SV40 T
143127	CMV	CMV	none	polyhedrin	polyhedrin	none	HSV-TK	oriP	none
143128	CMV	CMV	CMVa	CMV	none	none	HSV-TK	SV40	SV40 T
143129	none	EF1a_LTR-HTLV	none	none	none	WPRE	Globin-rabbit	none	none
143130	CMV	CMV	Chick actin/rabbit	polyhedrin	polyhedrin	none	Globin-rabbit	SV40	SV40 T
143131	CMV	CMV	CMVa	CMV_bglob-Hs	bglob-XI	none	Globin-rabbit	SV40	SV40 T
143132	CMV	CMV	none	CMV_BYDV	BYDV	none	Globin-rabbit	oriP	none
143133	CMV	CMV	none	CMV	none	AGS	Globin-rabbit	oriP	none
143134	CMV	CMV	CMVa	CMV_bglob-Hs	bglob-XI	none	HSV-TK_gastrin	SV40	SV40 T
143135	CMV	GAPDH	Chick actin/rabbit	none	none	none	GH-bovine	oriP	EBNA

TABLE 17

Vector element combinations used in gene transfer vectors.									
A	B En-	C Pro-	D	E	F	G	H	I	J
Construct	hancer	moter	Intron	5' UTR	RNA export	polyA	GFP Expression		
							1	2	3
136053	CMV	GAPDH	GAPDH	None	None	beta globin-rabbit	14,964	13,968	N/A
143090	CMV	CMV	CMVc	CMV	None	Globin-rabbit	16,825	15,882	12,666
180102	CMV	CMV	MLP	TPL	None	Globin-rabbit (trunc)	14,213	14,600	12,166
184141	CMV	Actin	Chick actin/rabbit	None	HPRE	Globin-rabbit	13,339	14,374	11,753
184142	CMV	CMV	GAPDH	CMV	AGSPRE	Globin-rabbit	5,893	6,734	4,947
184143	CMV	CMV	CMVa	CMV	None	Globin-rabbit	17,123	18,100	13,429
184144	CMV	GAPDH	Chick actin/rabbit	bglob-XI	None	HSV-TK_gastrin	15,608	14,932	12,082
184145	CMV	CMV	MLP	CMV_TPL	HPRE	HSV-TK_gastrin	10,837	8,024	9,687
184146	CMV	EF1a	GAPDH	TPL	None	Globin-rabbit	787	963	957
184147	CMV	GAPDH	GAPDH	TPL	HPRE	Globin-rabbit	993	1,167	713
184148	CMV	EF1a	CMVa	None	HPRE	HSV-TK_gastrin	6,479	6,230	2,848
184149	CMV	GAPDH	GAPDH	bglob-XI	None	HSV-TK_gastrin	18,036	17,007	9,516
184150	CMV	EF1a	None	bglob-XI	None	Globin-rabbit	13,987	14,431	11,176
184151	CMV	EF1a	None	None	None	BGH	13,502	14,312	12,185

TABLE 17-continued

Vector element combinations used in gene transfer vectors.									
A	B En-	C Pro-	D	E	F	G	H GFP Expression	I	J
Construct	hancer	moter	Intron	5' UTR	RNA export	polyA	1	2	3
184152	CMV	EF1a	GAPDH_MLP	TPL	AGSPRE	HSV-TK_gastrin	8,403	9,626	7,187
184153	CMV	Actin	GAPDH	None	None	Globin-rabbit	3,164	3,903	2,821
184154	CMV	Actin	CMVa	TPL	AGSPRE	HSV-TK_gastrin	645	717	639
184155	CMV	CMV	GAPDH	CMV_bglob-XI	HPRE	BGH	9,977	9,493	8,187
184156	CMV	GAPDH	None	None	AGSPRE	BGH	5,069	4,789	2,429
184157	CMV	EF1a	CMVa	bglob-XI	AGSPRE	Globin-rabbit	6,879	6,645	6,182
184158	CMV	Actin	GAPDH_MLP	TPL	None	BGH	9,739	10,908	8,815
184159	CMV	CMV	CMVa	CMV_bglob-XI	None	BGH	11,174	12,611	10,503
184160	CMV	CMV	Chick actin/rabbit_MLP	CMV_TPL	AGSPRE	Globin-rabbit	5,202	4,660	3,895
184162	CMV	EF1a	Chick actin/rabbit	TPL	None	BGH	758	923	779
184163	CMV	GAPDH	CMVa_MLP	TPL	None	Globin-rabbit	13,667	15,531	14,426
184164	CMV	CMV	None	CMV_TPL	None	HSV-TK_gastrin	409	400	424
188000	CMV	Actin	None	bglob-XI	AGSPRE_3	Globin-rabbit	10,467	10,274	10,189
189478	CMV	GAPDH	MLP	None	None	Globin-rabbit	13,341	13,005	11,990
189479	CMV	CMV	MLP	None	None	Globin-rabbit	12,099	12,077	12,621

TABLE 18

Vector element combinations used in gene transfer vectors.									
A	B	C	D	E	F	G	H GFP Expression	I	J
Construct	Enhancer	Promoter	Intron	RNA export	polyA	insulator	1	2	3
178620	CMV	CMV	none	SEQ ID NO: 104	BGH	none	1,513	1,572	1,490
178621	CMV	CMV	none	SEQ ID NO: 106	BGH	none	1,272	1,185	1,109
178622	CMV	CMV	none	SEQ ID NO: 107	BGH	none	865	891	979
178623	none	EF1a	EF1a	SEQ ID NO: 104	BGH	HS4	2,698	2,302	2,388
178624	none	EF1a	EF1a	SEQ ID NO: 106	BGH	HS4	138	122	114
178625	none	EF1a	EF1a	SEQ ID NO: 107	BGH	HS4	2,547	2,303	2,551
178626	CMV	CMV	none	SEQ ID NO: 104	BGH	HS4	245	258	137
178627	CMV	CMV	none	SEQ ID NO: 106	BGH	HS4	950	884	844
negative	none	none	none	none	none	none	29	12	30
145736	CMV	CMV	none	WPRE	BGH	none	908	1,106	952
142628	none	EF1a	EF1a	WPRE	synthetic	HS4	3,377	4,151	3,699
150708	CMV	CMV	none	WPRE	synthetic	HS4	1,074	1,080	1,056
150711	CMV	EF1a	EF1a	WPRE	synthetic	HS4	1,634	1,246	1,798
147759	none	EF1a	EF1a	SEQ ID NO: 104	BGH	none	850	812	1,003

TABLE 19

Expression of a fluorescent protein from a gene transfer systems designed to test control element configurations.				
A Construct	B CHO Avg-neg	C sd	D HEK Avg-neg	E sd
136024	146.1	25.3	4,650.6	245.1
136025	235.9	23.3	13,829.7	673.1
136026	242.2	50.6	4,299.9	199.1
136027	159.1	27.7	2,444.3	109.4
136028	0.4	0.5	51.6	14.8
136029	238.6	22.4	1,356.0	126.4
136030	70.2	12.7	1,651.3	112.8
136031	90.1	4.7	2,021.1	91.0
136032	281.8	16.8	2,560.9	127.2
136033	235.2	46.3	7,308.9	582.5
136034	166.1	22.2	6,412.6	491.3
136035	94.0	12.5	8,838.9	566.1
136036	156.1	10.4	3,557.0	260.5
136037	211.2	14.5	3,237.3	72.4
136038	143.7	4.3	3,662.4	220.0
136039	106.8	7.8	4,368.4	31.9
136040	68.1	13.6	4,669.5	646.8

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TABLE 19-continued

Expression of a fluorescent protein from a gene transfer systems designed to test control element configurations.				
A Construct	B CHO Avg-neg	C sd	D HEK Avg-neg	E sd
136041	199.4	24.0	5,646.5	517.9
136042	155.8	13.1	4,685.6	274.3
136043	225.5	24.0	3,638.9	377.8
136044	153.6	28.2	1,887.9	141.2
136045	187.1	8.0	3,942.0	5.1
136046	106.6	13.6	7,872.8	105.1
136047	121.2	8.3	1,824.1	116.9
136048	196.9	7.5	2,091.8	14.4
136049	112.5	7.5	1,994.0	100.3
136050	78.2	10.0	6,189.9	136.0
136051	74.0	24.9	738.6	19.7
136052	105.4	19.2	1,123.3	68.9
136053	349.3	30.9	1,433.4	59.5
136054	92.9	8.3	4,068.9	396.1
136055	129.8	9.9	522.5	6.4
136056	1.7	0.6	2.2	1.6
136057	6.9	1.0	0.6	1.2

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TABLE 19-continued

Expression of a fluorescent protein from a gene transfer systems designed to test control element configurations.				
A Construct	B CHO Avg-neg	C sd	D HEK Avg-neg	E sd
136058	2.6	0.5	1.2	1.3
136059	7.5	1.0	8.5	2.2
136060	11.4	1.5	6.2	2.0
136061	32.2	0.9	172.0	3.4
136062	17.1	2.9	10.4	1.2
136063	220.9	8.8	129.8	2.8
136064	27.0	1.8	18.5	2.4
136065	295.5	60.6	84.0	3.5
136066	55.9	8.3	76.9	5.1
136067	16.9	1.8	69.7	3.3
136068	33.9	8.9	18.5	5.4
136069	15.1	2.1	108.7	13.8
136070	19.6	3.5	789.6	86.2
136071	41.1	4.6	191.5	16.4
128979	175.4	30.5	2,884.7	160.0
128980	189.2	12.6	2,377.4	63.7
128986	97.5	21.7	5,160.8	32.4
129091	74.7	11.4	4,410.3	156.4
128977	67.9	6.8	2,453.4	205.9
133528	28.0	0.9	2,089.0	56.0
134746	47.1	2.2	4,004.1	345.9

TABLE 20

Expression of a fluorescent protein from a gene transfer systems designed to test control element configurations.				
A Construct	B CHO: Rel to 129091	C CHO: Rel to 136025	D HEK: Rel to 129091	E sd
143088	0.90	0.64	1.16	0.07
143089	0.85	0.61	0.22	0.01
143090	1.49	1.06	2.52	0.22
143091	0.06	0.04	0.09	0.01
143092	1.01	0.72	0.17	0.01
143093	1.13	0.81	1.51	0.12
143094	0.70	0.50	0.06	0.00
143095	0.62	0.44	0.41	0.03

TABLE 20-continued

Expression of a fluorescent protein from a gene transfer systems designed to test control element configurations.				
A Construct	B CHO: Rel to 129091	C CHO: Rel to 136025	D HEK: Rel to 129091	E sd
143096	2.21	1.58	0.57	0.06
143097	0.13	0.10	0.11	0.01
143098	0.44	0.31	0.39	0.04
143099	0.37	0.27	1.72	0.10
143100	0.71	0.51	0.35	0.02
143101	0.99	0.70	1.02	0.06
143102	0.42	0.30	0.06	0.01
143103	0.55	0.39	0.09	0.01
143104	0.03	0.03	0.01	0.00
143105	0.11	0.13	0.74	0.02
143106	0.39	0.48	0.89	0.04
143107	0.15	0.19	0.07	0.01
143108	0.05	0.06	0.49	0.01
143109	0.48	0.58	0.93	0.03
143110	0.32	0.38	1.81	0.09
143111	0.51	0.61	1.23	0.03
143112	0.07	0.08	0.02	0.00
143113	0.73	0.89	1.89	0.29
143114	0.64	0.78	0.24	0.01
143115	0.17	0.21	0.08	0.00
143116	0.96	1.16	0.11	0.01
143117	0.26	0.32	0.06	0.00
143118	0.85	1.03	0.07	0.00
143119	0.06	0.08	0.07	0.00
143120	0.45	0.32	0.00	0.00
143121	0.79	0.57	0.00	0.00
143122	1.25	0.89	0.29	0.03
143123	0.69	0.49	0.04	0.01
143124	1.40	1.00	1.24	0.12
143125	1.81	1.29	1.15	0.11
143126	0.46	0.33	0.01	0.00
143127	0.48	0.34	0.31	0.04
143128	1.56	1.11	3.31	0.31
143129	2.28	1.63	0.14	0.02
143130	1.05	0.75	2.59	0.26
143131	1.48	1.05	4.02	0.38
143132	0.38	0.27	0.95	0.09
143133	0.92	0.65	0.17	0.02
143134	1.45	1.03	3.78	0.35
143135	1.56	1.12	0.38	0.03

TABLE 21

Regression weights for vector elements used in transient expression in HEK and CHO cells.							
Enhancer	Promoter	Intron	polyA	viral ori	viral rep prot	CHO weight	HEK weight
CMV	CMV					51.37	682.70
CMV	Chick actin	Chick actin/rabbit				93.79	3,925.31
CMV	Actin	Chick actin (partial)				-44.92	-1,676.55
CMV	CMV	CMV intron a				52.61	4,103.61
CMV	CMV	CMV intron c				30.18	1,923.01
CMV	EF1a	EF1a_v1				63.03	1,452.29
CMV	EF1a	EF1a_v2				29.90	1,179.46
CMV	EF1a	EF1a_hybrid				6.34	110.31
CMV	GAPDH	GAPDH				44.57	-972.75
none	HSV-TK					-121.04	-1,898.14
none	MC1					-80.29	-2,735.93
none	PGK					-24.59	-1,364.48
E_SV40	SV40					-7.61	-2,138.04
none	Ub-B					-93.35	-2,590.80
			synthetic polyA			-39.44	-554.30
			GH-bovine			7.05	599.58
			sv40 early			0.53	-485.28
			sv40 late			-17.59	-599.00
			beta globin-human			16.44	-402.53
			beta globin-rabbit			43.28	869.96

TABLE 21-continued

Regression weights for vector elements used in transient expression in HEK and CHO cells.							
Enhancer	Promoter	Intron	polyA	viral ori	viral rep prot	CHO weight	HEK weight
			HSV-TK			-10.28	571.57
				SV40	SV40 T	-113.24	2,679.35
				oriP	none	-84.67	593.83
				oriP	EBNA	-106.31	-139.82

7. REFERENCES

All references cited herein are incorporated herein by reference in their entirety and for all purposes to the same extent as if each individual publication or patent or patent application was specifically and individually indicated to be incorporated by reference in its entirety for all purposes. If different versions of a sequence are associated with an accession number at different times, the version associated with the accession number at the effective filing date of this application is meant. The effective filing date means the earlier of the actual filing date or filing date of a priority application referring to the accession number if applicable. Likewise if different versions of a publication, website or the

like are published at different times, the version most recently published at the effective filing date of the application is meant unless otherwise indicated.

15 Many modifications and variations of this invention can be made without departing from its spirit and scope, as will be apparent to those skilled in the art. The specific embodiments described herein are offered by way of example only, and the invention is to be limited only by the terms of the
20 appended claims, along with the full scope of equivalents to which such claims are entitled. Any feature, step, element, embodiment, or aspect of the invention can be used in combination with any other unless specifically indicated otherwise.

SEQUENCE LISTING

<160> NUMBER OF SEQ ID NOS: 130

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gggaggggat agtggcgtga tcgcagtgtg gcacgggaca ccggcgagat attcgtgtgc 180
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caaaataaga tcaacataat tgagtaaata ataataagaa caatattata gtacaacaaa 300
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taaactaaaa gtcccctcga ggaaaggccc ctaaagtga acagtgcacaa acgttcacaa	180	
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taaactaaaa gtcccctcga ggaaaggccc ctaaagtga acagtgc aaa acgttcaaaa	180
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acaacacacg tcacgtcacg tcgttgcacg tcattttgac gtataattgg gctttgtgta	240
acttttgaat ttgtttcaaa ttttttatgt ttgtgattta tttgagttaa tcgtattgtt	300
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tttattactt ttatagttat ttagccaatt gttataaatt ccttggttatt gctgaaaaat	360	
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ttgcacgtca gcttgacgta tatttgggct ttgtgttatt tttgtaaatt ttcaacgtta	240	
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cataagtatg aaatgttatg atttgataca tatgtcattt tattctactg acattaacct	180
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<212> TYPE: DNA	
<213> ORGANISM: Xenopus tropicalis	
<400> SEQUENCE: 39	
ttaaccyttt tactgcca	18
<210> SEQ ID NO 40	
<211> LENGTH: 18	
<212> TYPE: DNA	
<213> ORGANISM: Xenopus tropicalis	
<400> SEQUENCE: 40	
tggcagtaaa agggttaa	18
<210> SEQ ID NO 41	
<211> LENGTH: 18	
<212> TYPE: DNA	
<213> ORGANISM: Xenopus tropicalis	
<400> SEQUENCE: 41	
tggcagtgaag agggttaa	18
<210> SEQ ID NO 42	
<211> LENGTH: 18	
<212> TYPE: DNA	
<213> ORGANISM: Xenopus tropicalis	
<400> SEQUENCE: 42	
ttaaccyttt kmctgcca	18
<210> SEQ ID NO 43	
<211> LENGTH: 610	
<212> TYPE: PRT	
<213> ORGANISM: Bombyx mori	
<400> SEQUENCE: 43	
Met Asp Ile Glu Arg Gln Glu Glu Arg Ile Arg Ala Met Leu Glu Glu	
1 5 10 15	
Glu Leu Ser Asp Tyr Ser Asp Glu Ser Ser Ser Glu Asp Glu Thr Asp	
20 25 30	
His Cys Ser Glu His Glu Val Asn Tyr Asp Thr Glu Glu Glu Arg Ile	
35 40 45	
Asp Ser Val Asp Val Pro Ser Asn Ser Arg Gln Glu Glu Ala Asn Ala	

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50				55				60							
Ile	Ile	Ala	Asn	Glu	Ser	Asp	Ser	Asp	Pro	Asp	Asp	Asp	Leu	Pro	Leu
65					70					75				80	
Ser	Leu	Val	Arg	Gln	Arg	Ala	Ser	Ala	Ser	Arg	Gln	Val	Ser	Gly	Pro
				85					90					95	
Phe	Tyr	Thr	Ser	Lys	Asp	Gly	Thr	Lys	Trp	Tyr	Lys	Asn	Cys	Gln	Arg
			100					105					110		
Pro	Asn	Val	Arg	Leu	Arg	Ser	Glu	Asn	Ile	Val	Thr	Glu	Gln	Ala	Gln
		115					120					125			
Val	Lys	Asn	Ile	Ala	Arg	Asp	Ala	Ser	Thr	Glu	Tyr	Glu	Cys	Trp	Asn
	130					135					140				
Ile	Phe	Val	Thr	Ser	Asp	Met	Leu	Gln	Glu	Ile	Leu	Thr	His	Thr	Asn
145					150					155					160
Ser	Ser	Ile	Arg	His	Arg	Gln	Thr	Lys	Thr	Ala	Ala	Glu	Asn	Ser	Ser
				165					170					175	
Ala	Glu	Thr	Ser	Phe	Tyr	Met	Gln	Glu	Thr	Thr	Leu	Cys	Glu	Leu	Lys
			180					185					190		
Ala	Leu	Ile	Ala	Leu	Leu	Tyr	Leu	Ala	Gly	Leu	Ile	Lys	Ser	Asn	Arg
	195						200					205			
Gln	Ser	Leu	Lys	Asp	Leu	Trp	Arg	Thr	Asp	Gly	Thr	Gly	Val	Asp	Ile
	210					215					220				
Phe	Arg	Thr	Thr	Met	Ser	Leu	Gln	Arg	Phe	Gln	Phe	Leu	Gln	Asn	Asn
225					230					235					240
Ile	Arg	Phe	Asp	Asp	Lys	Ser	Thr	Arg	Asp	Glu	Arg	Lys	Gln	Thr	Asp
			245						250					255	
Asn	Met	Ala	Ala	Phe	Arg	Ser	Ile	Phe	Asp	Gln	Phe	Val	Gln	Cys	Cys
		260						265					270		
Gln	Asn	Ala	Tyr	Ser	Pro	Ser	Glu	Phe	Leu	Thr	Ile	Asp	Glu	Met	Leu
	275						280					285			
Leu	Ser	Phe	Arg	Gly	Arg	Cys	Leu	Phe	Arg	Val	Tyr	Ile	Pro	Asn	Lys
	290					295					300				
Pro	Ala	Lys	Tyr	Gly	Ile	Lys	Ile	Leu	Ala	Leu	Val	Asp	Ala	Lys	Asn
305					310					315					320
Phe	Asp	Val	Val	Asn	Leu	Glu	Val	Tyr	Ala	Gly	Lys	Gln	Pro	Ser	Gly
				325					330					335	
Pro	Tyr	Ala	Val	Ser	Asn	Arg	Pro	Phe	Glu	Val	Val	Glu	Arg	Leu	Ile
		340						345					350		
Gln	Pro	Val	Ala	Arg	Ser	His	Arg	Asn	Val	Thr	Phe	Asp	Asn	Trp	Phe
		355					360					365			
Thr	Gly	Tyr	Glu	Leu	Met	Leu	His	Leu	Leu	Asn	Glu	Tyr	Arg	Leu	Thr
	370					375					380				
Ser	Val	Gly	Thr	Val	Arg	Lys	Asn	Lys	Arg	Gln	Ile	Pro	Glu	Ser	Phe
385					390					395					400
Ile	Arg	Thr	Asp	Arg	Gln	Pro	Asn	Ser	Ser	Val	Phe	Gly	Phe	Gln	Lys
			405						410					415	
Asp	Ile	Thr	Leu	Val	Ser	Tyr	Ala	Pro	Lys	Lys	Asn	Lys	Val	Val	Val
		420						425					430		
Val	Met	Ser	Thr	Met	His	His	Asp	Asn	Ser	Ile	Asp	Glu	Ser	Thr	Gly
	435						440					445			
Glu	Lys	Gln	Lys	Pro	Glu	Met	Ile	Thr	Phe	Tyr	Asn	Ser	Thr	Lys	Ala
	450					455					460				
Gly	Val	Asp	Val	Val	Asp	Glu	Leu	Ser	Ala	Asn	Tyr	Asn	Val	Ser	Arg
465					470					475					480

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Asn	Ser	Lys	Arg	Trp	Pro	Met	Thr	Leu	Phe	Tyr	Gly	Val	Leu	Asn	Met	
				485					490					495		
Ala	Ala	Ile	Asn	Ala	Cys	Ile	Ile	Tyr	Arg	Ala	Asn	Lys	Asn	Val	Thr	
			500					505					510			
Ile	Lys	Arg	Thr	Glu	Phe	Ile	Arg	Ser	Leu	Gly	Leu	Ser	Met	Ile	Tyr	
		515					520					525				
Glu	His	Leu	His	Ser	Arg	Asn	Lys	Lys	Lys	Asn	Ile	Pro	Thr	Tyr	Leu	
	530					535					540					
Arg	Gln	Arg	Ile	Glu	Lys	Gln	Leu	Gly	Glu	Pro	Ser	Pro	Arg	His	Val	
545					550				555						560	
Asn	Val	Pro	Gly	Arg	Tyr	Val	Arg	Cys	Gln	Asp	Cys	Pro	Tyr	Lys	Lys	
				565					570					575		
Asp	Arg	Lys	Thr	Lys	His	Ser	Cys	Asn	Ala	Cys	Ala	Lys	Pro	Ile	Cys	
			580					585					590			
Met	Glu	His	Ala	Lys	Phe	Leu	Cys	Glu	Asn	Cys	Ala	Glu	Leu	Asp	Ser	
		595					600					605				
Ser	Leu															
	610															
<210> SEQ ID NO 44																
<211> LENGTH: 610																
<212> TYPE: PRT																
<213> ORGANISM: Bombyx mori																
<400> SEQUENCE: 44																
Met	Asp	Ile	Glu	Arg	Gln	Glu	Glu	Arg	Ile	Arg	Ala	Met	Leu	Glu	Glu	
1				5					10					15		
Glu	Leu	Ser	Asp	Tyr	Ser	Asp	Glu	Ser	Ser	Ser	Glu	Asp	Glu	Thr	Asp	
			20					25					30			
His	Cys	Ser	Glu	His	Glu	Val	Asn	Tyr	Asp	Thr	Glu	Glu	Glu	Arg	Ile	
		35					40					45				
Asp	Ser	Val	Asp	Val	Pro	Ser	Asn	Ser	Arg	Gln	Glu	Glu	Ala	Asn	Ala	
	50					55				60						
Ile	Ile	Ala	Asn	Glu	Ser	Asp	Ser	Asp	Pro	Asp	Asp	Asp	Leu	Pro	Leu	
65				70					75					80		
Ser	Leu	Val	Arg	Gln	Arg	Ala	Ser	Ala	Ser	Arg	Gln	Val	Ser	Gly	Pro	
			85					90						95		
Phe	Tyr	Thr	Ser	Lys	Asp	Gly	Thr	Lys	Trp	Tyr	Lys	Asn	Cys	Gln	Arg	
			100					105					110			
Pro	Asn	Val	Arg	Leu	Arg	Ser	Glu	Asn	Ile	Val	Thr	Glu	Gln	Ala	Gln	
		115					120					125				
Val	Lys	Asn	Ile	Ala	Arg	Asp	Ala	Ser	Thr	Glu	Tyr	Glu	Cys	Trp	Asn	
		130				135					140					
Ile	Phe	Val	Thr	Ser	Asp	Met	Leu	Gln	Glu	Ile	Leu	Thr	His	Thr	Asn	
145				150					155						160	
Ser	Ser	Ile	Arg	His	Arg	Gln	Thr	Lys	Thr	Ala	Ala	Glu	Asn	Ser	Ser	
			165					170					175			
Ala	Glu	Thr	Ser	Phe	Tyr	Met	Gln	Glu	Thr	Thr	Leu	Cys	Glu	Leu	Lys	
			180					185					190			
Ala	Leu	Ile	Ala	Leu	Leu	Tyr	Leu	Ala	Gly	Leu	Ile	Lys	Ser	Asn	Arg	
		195				200						205				
Gln	Ser	Leu	Lys	Asp	Leu	Trp	Arg	Thr	Asp	Gly	Thr	Gly	Val	Asp	Ile	
	210					215				220						
Phe	Arg	Thr	Thr	Met	Ser	Leu	Gln	Arg	Phe	Gln	Phe	Leu	Gln	Asn	Asn	

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225								230								235								240																							
Ile	Arg	Phe	Asp	Asp	Lys	Ser	Thr	Arg	Asp	Glu	Arg	Lys	Gln	Thr	Asp																																
				245									250									255																									
Asn	Met	Ala	Ala	Phe	Arg	Ser	Ile	Phe	Asp	Gln	Phe	Val	Gln	Cys	Cys																																
				260									265									270																									
Gln	Asn	Ala	Tyr	Ser	Pro	Ser	Glu	Phe	Leu	Thr	Ile	Asp	Glu	Met	Leu																																
				275																	285																										
Leu	Ser	Phe	Arg	Gly	Arg	Cys	Leu	Phe	Arg	Val	Tyr	Ile	Pro	Asn	Lys																																
				290																	300																										
Pro	Ala	Lys	Tyr	Gly	Ile	Lys	Ile	Leu	Ala	Leu	Val	Asp	Ala	Lys	Asn																																
				305																	315					320																					
Phe	Tyr	Val	Val	Asn	Leu	Glu	Val	Tyr	Ala	Gly	Lys	Gln	Pro	Ser	Gly																																
								325													330					335																					
Pro	Tyr	Ala	Val	Ser	Asn	Arg	Pro	Phe	Glu	Val	Val	Glu	Arg	Leu	Ile																																
																				340					345					350																	
Gln	Pro	Val	Ala	Arg	Ser	His	Arg	Asn	Val	Thr	Phe	Asp	Asn	Trp	Phe																																
																				355					360					365																	
Thr	Gly	Tyr	Glu	Leu	Met	Leu	His	Leu	Leu	Asn	Glu	Tyr	Arg	Leu	Thr																																
																				370					375					380																	
Ser	Val	Gly	Thr	Val	Arg	Lys	Asn	Lys	Arg	Gln	Ile	Pro	Glu	Ser	Phe																																
																				385					390					395					400												
Ile	Arg	Thr	Asp	Arg	Gln	Pro	Asn	Ser	Ser	Val	Phe	Gly	Phe	Gln	Lys																																
																								405					410					415													
Asp	Ile	Thr	Leu	Val	Ser	Tyr	Ala	Pro	Lys	Lys	Asn	Lys	Val	Val	Val																																
																								420					425					430													
Val	Met	Ser	Thr	Met	His	His	Asp	Asn	Ser	Ile	Asp	Glu	Ser	Thr	Gly																																
																								435					440					445													
Glu	Lys	Gln	Lys	Pro	Glu	Met	Ile	Thr	Phe	Tyr	Asn	Ser	Thr	Lys	Ala																																
																								450					455					460													
Gly	Val	Asp	Val	Val	Asp	Glu	Leu	Cys	Ala	Asn	Tyr	Asn	Val	Ser	Arg																																
																								465					470					475					480								
Asn	Ser	Lys	Arg	Trp	Pro	Met	Thr	Leu	Phe	Tyr	Gly	Val	Leu	Asn	Met																																
																												485					490					495									
Ala	Ala	Ile	Asn	Ala	Cys	Ile	Ile	Tyr	Arg	Thr	Asn	Lys	Asn	Val	Thr																																
																												500					505					510									
Ile	Lys	Arg	Thr	Glu	Phe	Ile	Arg	Ser	Leu	Gly	Leu	Ser	Met	Ile	Tyr																																
																												515					520					525									
Glu	His	Leu	His	Ser	Arg	Asn	Lys	Lys	Lys	Asn	Ile	Pro	Thr	Tyr	Leu																																
																												530					535					540									
Arg	Gln	Arg	Ile	Glu	Lys	Gln	Leu	Gly	Glu	Pro	Ser	Pro	Arg	His	Val																																
																																545					550					555					560
Asn	Val	Pro	Gly	Arg	Tyr	Val	Arg	Cys	Gln	Asp	Cys	Pro	Tyr	Lys	Lys																																
																																565					570					575					
Asp	Arg	Lys	Thr	Lys	Arg	Ser	Cys	Asn	Ala	Cys	Ala	Lys	Pro	Ile	Cys																																
																																580					585					590					
Met	Glu	His	Ala	Lys	Phe	Leu	Cys	Glu	Asn	Cys	Ala	Glu	Leu	Asp	Ser																																
																																595					600					605					
Ser	Leu																																														
																	610																														

<210> SEQ ID NO 45
<211> LENGTH: 589
<212> TYPE: PRT
<213> ORGANISM: Xenopus tropicalis

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<400> SEQUENCE: 45																	
Met	Ala	Lys	Arg	Phe	Tyr	Ser	Ala	Glu	Glu	Ala	Ala	Ala	His	Cys	Met		
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Ala	Ser	Ser	Ser	Glu	Glu	Phe	Ser	Gly	Ser	Asp	Ser	Glu	Tyr	Val	Pro		
			20					25					30				
Pro	Ala	Ser	Glu	Ser	Asp	Ser	Ser	Thr	Glu	Glu	Ser	Trp	Cys	Ser	Ser		
		35					40					45					
Ser	Thr	Val	Ser	Ala	Leu	Glu	Glu	Pro	Met	Glu	Val	Asp	Glu	Asp	Val		
	50					55					60						
Asp	Asp	Leu	Glu	Asp	Gln	Glu	Ala	Gly	Asp	Arg	Ala	Asp	Ala	Ala	Ala		
65					70					75					80		
Gly	Gly	Glu	Pro	Ala	Trp	Gly	Pro	Pro	Cys	Asn	Phe	Pro	Pro	Glu	Ile		
				85					90					95			
Pro	Pro	Phe	Thr	Thr	Val	Pro	Gly	Val	Lys	Val	Asp	Thr	Ser	Asn	Phe		
			100					105					110				
Glu	Pro	Ile	Asn	Phe	Phe	Gln	Leu	Phe	Met	Thr	Glu	Ala	Ile	Leu	Gln		
		115					120					125					
Asp	Met	Val	Leu	Tyr	Thr	Asn	Val	Tyr	Ala	Glu	Gln	Tyr	Leu	Thr	Gln		
	130					135					140						
Asn	Pro	Leu	Pro	Arg	Tyr	Ala	Arg	Ala	His	Ala	Trp	His	Pro	Thr	Asp		
145					150					155					160		
Ile	Ala	Glu	Met	Lys	Arg	Phe	Val	Gly	Leu	Thr	Leu	Ala	Met	Gly	Leu		
				165					170					175			
Ile	Lys	Ala	Asn	Ser	Leu	Glu	Ser	Tyr	Trp	Asp	Thr	Thr	Thr	Val	Leu		
			180					185					190				
Ser	Ile	Pro	Val	Phe	Ser	Ala	Thr	Met	Ser	Arg	Asn	Arg	Tyr	Gln	Leu		
		195					200					205					
Leu	Leu	Arg	Phe	Leu	His	Phe	Asn	Asn	Asn	Ala	Thr	Ala	Val	Pro	Pro		
		210				215					220						
Asp	Gln	Pro	Gly	His	Asp	Arg	Leu	His	Lys	Leu	Arg	Pro	Leu	Ile	Asp		
225					230					235				240			
Ser	Leu	Ser	Glu	Arg	Phe	Ala	Ala	Val	Tyr	Thr	Pro	Cys	Gln	Asn	Ile		
				245					250					255			
Cys	Ile	Asp	Glu	Ser	Leu	Leu	Leu	Phe	Lys	Gly	Arg	Leu	Gln	Phe	Arg		
		260						265					270				
Gln	Tyr	Ile	Pro	Ser	Lys	Arg	Ala	Arg	Tyr	Gly	Ile	Lys	Phe	Tyr	Lys		
		275					280					285					
Leu	Cys	Glu	Ser	Ser	Ser	Gly	Tyr	Thr	Ser	Tyr	Phe	Leu	Ile	Tyr	Glu		
	290					295					300						
Gly	Lys	Asp	Ser	Lys	Leu	Asp	Pro	Pro	Gly	Cys	Pro	Pro	Asp	Leu	Thr		
305					310					315				320			
Val	Ser	Gly	Lys	Ile	Val	Trp	Glu	Leu	Ile	Ser	Pro	Leu	Leu	Gly	Gln		
				325					330					335			
Gly	Phe	His	Leu	Tyr	Val	Asp	Asn	Phe	Tyr	Ser	Ser	Ile	Pro	Leu	Phe		
			340					345					350				
Thr	Ala	Leu	Tyr	Cys	Leu	Asp	Thr	Pro	Ala	Cys	Gly	Thr	Ile	Asn	Arg		
		355					360					365					
Asn	Arg	Lys	Gly	Leu	Pro	Arg	Ala	Leu	Leu	Asp	Lys	Lys	Leu	Asn	Arg		
		370				375					380						
Gly	Glu	Thr	Tyr	Ala	Leu	Arg	Lys	Asn	Glu	Leu	Leu	Ala	Ile	Lys	Phe		
385					390					395				400			
Phe	Asp	Lys	Lys	Asn	Val	Phe	Met	Leu	Thr	Ser	Ile	His	Asp	Glu	Ser		


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405
Val Ile Arg Glu Gln Arg Val Gly Arg Pro Pro Lys Asn Lys Pro Leu
420 425 430
Cys Ser Lys Glu Tyr Ser Lys Tyr Met Gly Gly Val Asp Arg Thr Asp
435 440 445
Gln Leu Gln His Tyr Tyr Asn Ala Thr Arg Lys Thr Arg Ala Trp Tyr
450 455 460
Lys Lys Val Gly Ile Tyr Leu Ile Gln Met Ala Leu Arg Asn Ser Tyr
465 470 475 480
Ile Val Tyr Lys Ala Ala Val Pro Gly Pro Lys Leu Ser Tyr Tyr Lys
485 490 495
Tyr Gln Leu Gln Ile Leu Pro Ala Leu Leu Phe Gly Gly Val Glu Glu
500 505 510
Gln Thr Val Pro Glu Met Pro Pro Ser Asp Asn Val Ala Arg Leu Ile
515 520 525
Gly Lys His Phe Ile Asp Thr Leu Pro Pro Thr Pro Gly Lys Gln Arg
530 535 540
Pro Gln Lys Gly Cys Lys Val Cys Arg Lys Arg Gly Ile Arg Arg Asp
545 550 555 560
Thr Arg Tyr Tyr Cys Pro Lys Cys Pro Arg Asn Pro Gly Leu Cys Phe
565 570 575
Lys Pro Cys Phe Glu Ile Tyr His Thr Gln Leu His Tyr
580 585
<210> SEQ ID NO 46
<211> LENGTH: 589
<212> TYPE: PRT
<213> ORGANISM: Xenopus tropicalis
<400> SEQUENCE: 46
Met Ala Lys Arg Phe Tyr Ser Ala Glu Glu Ala Ala Ala His Cys Met
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Ala Pro Ser Ser Glu Glu Phe Ser Gly Ser Asp Ser Glu Tyr Val Pro
20 25 30
Pro Ala Ser Glu Ser Asp Ser Ser Thr Glu Glu Ser Trp Cys Ser Ser
35 40 45
Ser Thr Val Ser Ala Leu Glu Glu Pro Met Glu Val Asp Glu Asp Val
50 55 60
Asp Asp Leu Glu Asp Gln Glu Ala Gly Asp Arg Ala Asp Ala Ala Ala
65 70 75 80
Gly Gly Glu Pro Ala Trp Gly Pro Pro Cys Asn Phe Pro Pro Glu Ile
85 90 95
Pro Pro Phe Thr Thr Val Pro Gly Val Lys Val Asp Thr Ser Asn Phe
100 105 110
Glu Pro Ile Asn Phe Phe Gln Leu Phe Met Thr Glu Ala Ile Leu Gln
115 120 125
Asp Met Val Leu Tyr Thr Asn Val Tyr Ala Glu Gln Tyr Leu Thr Gln
130 135 140
Asn Pro Leu Pro Arg Tyr Ala Arg Ala His Ala Trp His Pro Thr Asp
145 150 155 160
Ile Ala Glu Met Lys Arg Phe Val Gly Leu Thr Leu Ala Met Gly Leu
165 170 175
Ile Lys Ala Asn Ser Leu Glu Ser Tyr Trp Asn Thr Thr Thr Val Leu
180 185 190

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Ser	Ile	Pro	Val	Phe	Ser	Ala	Thr	Met	Ser	Arg	Asn	Arg	Tyr	Gln	Leu
		195					200					205			
Leu	Leu	Arg	Phe	Leu	His	Phe	Asn	Asn	Asn	Ala	Thr	Ala	Val	Pro	Pro
	210					215					220				
Asp	Gln	Pro	Asp	His	Asp	Arg	Leu	His	Lys	Leu	Arg	Pro	Leu	Ile	Asp
225					230					235					240
Ser	Leu	Ser	Glu	Arg	Phe	Ala	Ala	Val	Tyr	Thr	Pro	Cys	Gln	Asn	Ile
				245					250					255	
Cys	Ile	Asp	Glu	Ser	Leu	Leu	Leu	Phe	Lys	Gly	Arg	Leu	Arg	Phe	Arg
			260					265					270		
Gln	Tyr	Ile	Pro	Ser	Lys	Arg	Ala	Arg	Tyr	Gly	Ile	Lys	Phe	Tyr	Lys
		275					280					285			
Leu	Cys	Glu	Ser	Ser	Ser	Gly	Tyr	Thr	Ser	Tyr	Phe	Leu	Ile	Tyr	Glu
	290					295					300				
Gly	Lys	Asp	Ser	Lys	Leu	Asp	Pro	Pro	Gly	Cys	Pro	Pro	Asp	Leu	Thr
305					310					315					320
Val	Ser	Gly	Lys	Ile	Val	Trp	Glu	Leu	Ile	Ser	Pro	Leu	Leu	Gly	Gln
				325					330					335	
Gly	Phe	His	Leu	Tyr	Val	Asp	Asn	Phe	Tyr	Ser	Ser	Ile	Pro	Leu	Phe
			340					345					350		
Thr	Ala	Leu	Tyr	Cys	Leu	Asp	Thr	Pro	Ala	Cys	Gly	Thr	Ile	Asn	Arg
		355					360					365			
Thr	Arg	Lys	Gly	Leu	Pro	Arg	Ala	Leu	Leu	Asp	Lys	Lys	Leu	Asn	Arg
	370					375					380				
Gly	Glu	Thr	Tyr	Ala	Leu	Arg	Lys	Asn	Glu	Leu	Leu	Ala	Ile	Lys	Phe
385					390					395					400
Phe	Asp	Lys	Lys	Asn	Val	Phe	Met	Leu	Thr	Ser	Ile	His	Asp	Glu	Ser
				405					410					415	
Val	Ile	Arg	Glu	Gln	Arg	Val	Gly	Arg	Pro	Pro	Lys	Asn	Lys	Pro	Leu
			420					425					430		
Cys	Ser	Lys	Glu	Tyr	Ser	Lys	Tyr	Met	Gly	Gly	Val	Asp	Arg	Thr	Asp
		435					440					445			
Gln	Leu	Gln	His	Tyr	Tyr	Asn	Ala	Thr	Arg	Lys	Thr	Ser	Ala	Trp	Tyr
	450					455					460				
Lys	Lys	Val	Gly	Ile	Tyr	Leu	Ile	Gln	Met	Ala	Leu	Arg	Asn	Ser	Tyr
465					470					475					480
Ile	Val	Tyr	Lys	Ala	Ala	Val	Pro	Gly	Pro	Lys	Leu	Ser	Tyr	Tyr	Lys
				485					490					495	
Tyr	Gln	Leu	Gln	Ile	Leu	Pro	Ala	Leu	Leu	Phe	Gly	Gly	Val	Glu	Glu
			500					505					510		
Gln	Thr	Val	Pro	Glu	Met	Leu	Pro	Ser	Asp	Asn	Val	Ala	Arg	Leu	Ile
		515					520					525			
Gly	Lys	His	Phe	Ile	Asp	Thr	Leu	Pro	Pro	Thr	Pro	Gly	Lys	Gln	Arg
	530					535					540				
Pro	Gln	Lys	Gly	Cys	Lys	Val	Cys	Arg	Lys	Arg	Gly	Ile	Arg	Arg	Asp
545					550					555					560
Thr	Arg	Tyr	Tyr	Cys	Pro	Lys	Cys	Pro	Arg	Asn	Pro	Gly	Leu	Cys	Phe
				565					570					575	
Lys	Pro	Cys	Phe	Glu	Ile	Tyr	His	Thr	Gln	Leu	His	Tyr			
			580					585							

<210> SEQ ID NO 47
<211> LENGTH: 597
<212> TYPE: PRT

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<213> ORGANISM: Helicoverpa armigera															
<400> SEQUENCE: 47															
Met	Ala	Ser	Arg	Gln	Arg	Leu	Asn	His	Asp	Glu	Ile	Ala	Thr	Ile	Leu
1				5					10					15	
Glu	Asn	Asp	Asp	Asp	Tyr	Ser	Pro	Leu	Asp	Ser	Glu	Ser	Glu	Lys	Glu
		20						25					30		
Asp	Cys	Val	Val	Glu	Asp	Asp	Val	Trp	Ser	Asp	Asn	Glu	Asp	Ala	Ile
		35					40					45			
Val	Asp	Phe	Val	Glu	Asp	Thr	Ser	Ala	Gln	Glu	Asp	Pro	Asp	Asn	Asn
	50					55					60				
Ile	Ala	Ser	Arg	Glu	Ser	Pro	Asn	Leu	Glu	Val	Thr	Ser	Leu	Thr	Ser
65					70				75						80
His	Arg	Ile	Ile	Thr	Leu	Pro	Gln	Arg	Ser	Ile	Arg	Gly	Lys	Asn	Asn
				85					90					95	
His	Val	Trp	Ser	Thr	Thr	Lys	Gly	Arg	Thr	Thr	Gly	Arg	Thr	Ser	Ala
			100					105					110		
Ile	Asn	Ile	Ile	Arg	Thr	Asn	Arg	Gly	Pro	Thr	Arg	Met	Cys	Arg	Asn
	115					120					125				
Ile	Val	Asp	Pro	Leu	Leu	Cys	Phe	Gln	Leu	Phe	Ile	Thr	Asp	Glu	Ile
	130					135					140				
Ile	His	Glu	Ile	Val	Lys	Trp	Thr	Asn	Val	Glu	Ile	Ile	Val	Lys	Arg
145					150					155					160
Gln	Asn	Leu	Lys	Asp	Ile	Ser	Ala	Ser	Tyr	Arg	Asp	Thr	Asn	Thr	Met
			165						170					175	
Glu	Ile	Trp	Ala	Leu	Val	Gly	Ile	Leu	Thr	Leu	Thr	Ala	Val	Met	Lys
		180						185					190		
Asp	Asn	His	Leu	Ser	Thr	Asp	Glu	Leu	Phe	Asp	Ala	Thr	Phe	Ser	Gly
		195					200				205				
Thr	Arg	Tyr	Val	Ser	Val	Met	Ser	Arg	Glu	Arg	Phe	Glu	Phe	Leu	Ile
	210					215					220				
Arg	Cys	Ile	Arg	Met	Asp	Asp	Lys	Thr	Leu	Arg	Pro	Thr	Leu	Arg	Ser
225					230					235					240
Asp	Asp	Ala	Phe	Leu	Pro	Val	Arg	Lys	Ile	Trp	Glu	Ile	Phe	Ile	Asn
			245						250					255	
Gln	Cys	Arg	Gln	Asn	His	Val	Pro	Gly	Ser	Asn	Leu	Thr	Val	Asp	Glu
			260					265					270		
Gln	Leu	Leu	Gly	Phe	Arg	Gly	Arg	Cys	Pro	Phe	Arg	Met	Tyr	Ile	Pro
	275						280					285			
Asn	Lys	Pro	Asp	Lys	Tyr	Gly	Ile	Lys	Phe	Pro	Met	Met	Cys	Ala	Ala
	290					295					300				
Ala	Thr	Lys	Tyr	Met	Ile	Asp	Ala	Ile	Pro	Tyr	Leu	Gly	Lys	Ser	Thr
305					310					315					320
Lys	Thr	Asn	Gly	Leu	Pro	Leu	Gly	Glu	Phe	Tyr	Val	Lys	Asp	Leu	Thr
			325					330						335	
Lys	Thr	Val	His	Gly	Thr	Asn	Arg	Asn	Ile	Thr	Cys	Asp	Asn	Trp	Phe
			340					345					350		
Thr	Ser	Ile	Pro	Leu	Ala	Lys	Asn	Met	Leu	Gln	Ala	Pro	Tyr	Asn	Leu
		355					360					365			
Thr	Ile	Val	Gly	Thr	Ile	Arg	Ser	Asn	Lys	Arg	Glu	Met	Pro	Glu	Glu
	370					375					380				
Ile	Lys	Asn	Ser	Arg	Ser	Arg	Pro	Val	Gly	Ser	Ser	Met	Phe	Cys	Phe
385					390					395					400

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Asp	Gly	Pro	Leu	Thr	Leu	Val	Ser	Tyr	Lys	Pro	Lys	Pro	Ser	Lys	Met	
				405					410						415	
Val	Phe	Leu	Leu	Ser	Ser	Cys	Asp	Glu	Asn	Ala	Val	Ile	Asn	Glu	Ser	
			420					425					430			
Asn	Gly	Lys	Pro	Asp	Met	Ile	Leu	Phe	Tyr	Asn	Gln	Thr	Lys	Gly	Gly	
		435					440					445				
Val	Asp	Ser	Phe	Asp	Gln	Met	Cys	Lys	Ser	Met	Ser	Ala	Asn	Arg	Lys	
	450					455					460					
Thr	Asn	Arg	Trp	Pro	Met	Ala	Val	Phe	Tyr	Gly	Met	Leu	Asn	Met	Ala	
465					470					475					480	
Phe	Val	Asn	Ser	Tyr	Ile	Ile	Tyr	Cys	His	Asn	Lys	Ile	Asn	Lys	Gln	
				485					490					495		
Glu	Lys	Pro	Ile	Ser	Arg	Lys	Glu	Phe	Met	Lys	Lys	Leu	Ser	Ile	Gln	
			500					505					510			
Leu	Thr	Thr	Pro	Trp	Met	Gln	Glu	Arg	Leu	Gln	Ala	Pro	Thr	Leu	Lys	
		515					520						525			
Arg	Thr	Leu	Arg	Asp	Asn	Ile	Thr	Asn	Val	Leu	Lys	Asn	Val	Val	Pro	
	530					535					540					
Ala	Ser	Ser	Glu	Asn	Ile	Ser	Asn	Glu	Pro	Glu	Pro	Lys	Lys	Arg	Arg	
545					550					555					560	
Tyr	Cys	Gly	Val	Cys	Ser	Tyr	Lys	Lys	Arg	Arg	Met	Thr	Lys	Ala	Gln	
				565					570					575		
Cys	Cys	Lys	Cys	Lys	Lys	Ala	Ile	Cys	Gly	Glu	His	Asn	Ile	Asp	Val	
			580					585					590			
Cys	Gln	Asp	Cys	Ile												
		595														
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<213> ORGANISM: Pectinophora gossypiella																
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1				5					10					15		
Asp	Ile	Glu	Glu	Asp	Ser	Lys	Gly	Glu	Ser	Asp	Asn	Ser	Ser	Ser	Glu	
			20					25					30			
Thr	Glu	Asp	Ile	Val	Glu	Met	Glu	Val	His	Lys	Asn	Thr	Ser	Ser	Glu	
		35					40					45				
Ser	Glu	Val	Ser	Ser	Glu	Ser	Asp	Tyr	Glu	Pro	Val	Cys	Pro	Ser	Lys	
	50					55					60					
Arg	Gln	Arg	Thr	Gln	Ile	Ile	Glu	Ser	Glu	Glu	Ser	Asp	Asn	Ser	Glu	
65					70					75					80	
Ser	Ile	Arg	Pro	Ser	Arg	Arg	Gln	Thr	Ser	Arg	Val	Ile	Asp	Ser	Asp	
				85					90					95		
Glu	Thr	Asp	Glu	Asp	Val	Met	Ser	Ser	Thr	Pro	Gln	Asn	Ile	Pro	Arg	
			100					105					110			
Asn	Pro	Asn	Val	Ile	Gln	Pro	Ser	Ser	Arg	Phe	Leu	Tyr	Gly	Lys	Asn	
		115					120						125			
Lys	His	Lys	Trp	Ser	Ser	Ala	Ala	Lys	Pro	Ser	Ser	Val	Arg	Thr	Ser	
						135						140				
Arg	Arg	Asn	Ile	Ile	His	Phe	Ile	Pro	Gly	Pro	Lys	Glu	Arg	Ala	Arg	
145					150					155					160	
Glu	Val	Ser	Glu	Pro	Ile	Asp	Ile	Phe	Ser	Leu	Phe	Ile	Ser	Glu	Asp	
				165					170					175		

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Met	Leu	Gln	Gln	Val	Val	Thr	Phe	Thr	Asn	Ala	Glu	Met	Leu	Ile	Arg
		180						185					190		
Lys	Asn	Lys	Tyr	Lys	Thr	Glu	Thr	Phe	Thr	Val	Ser	Pro	Thr	Asn	Leu
		195					200					205			
Glu	Glu	Ile	Arg	Ala	Leu	Leu	Gly	Leu	Leu	Phe	Asn	Ala	Ala	Ala	Met
	210					215					220				
Lys	Ser	Asn	His	Leu	Pro	Thr	Arg	Met	Leu	Phe	Asn	Thr	His	Arg	Ser
225					230					235					240
Gly	Thr	Ile	Phe	Lys	Ala	Cys	Met	Ser	Ala	Glu	Arg	Leu	Asn	Phe	Leu
				245					250					255	
Ile	Lys	Cys	Leu	Arg	Phe	Asp	Asp	Lys	Leu	Thr	Arg	Asn	Val	Arg	Gln
			260					265					270		
Arg	Asp	Asp	Arg	Phe	Ala	Pro	Ile	Arg	Asp	Leu	Trp	Gln	Ala	Leu	Ile
	275						280					285			
Ser	Asn	Phe	Gln	Lys	Trp	Tyr	Thr	Pro	Gly	Ser	Tyr	Ile	Thr	Val	Asp
	290					295					300				
Glu	Gln	Leu	Val	Gly	Phe	Arg	Gly	Arg	Cys	Ser	Phe	Arg	Met	Tyr	Ile
305					310					315					320
Pro	Asn	Lys	Pro	Asn	Lys	Tyr	Gly	Ile	Lys	Leu	Val	Met	Ala	Ala	Asp
			325						330					335	
Val	Asn	Ser	Lys	Tyr	Ile	Val	Asn	Ala	Ile	Pro	Tyr	Leu	Gly	Lys	Gly
			340					345					350		
Thr	Asp	Pro	Gln	Asn	Gln	Pro	Leu	Ala	Thr	Phe	Phe	Ile	Lys	Glu	Ile
	355					360						365			
Thr	Ser	Thr	Leu	His	Gly	Thr	Asn	Arg	Asn	Ile	Thr	Met	Asp	Asn	Trp
	370					375					380				
Phe	Thr	Ser	Val	Pro	Leu	Ala	Asn	Glu	Leu	Leu	Met	Ala	Pro	Tyr	Asn
385					390					395					400
Leu	Thr	Leu	Val	Gly	Thr	Leu	Arg	Ser	Asn	Lys	Arg	Glu	Ile	Pro	Glu
			405						410					415	
Lys	Leu	Lys	Asn	Ser	Lys	Ser	Arg	Ala	Ile	Gly	Thr	Ser	Met	Phe	Cys
			420					425					430		
Tyr	Asp	Gly	Asp	Lys	Thr	Leu	Val	Ser	Tyr	Lys	Ala	Lys	Ser	Asn	Lys
		435					440					445			
Val	Val	Phe	Ile	Leu	Ser	Thr	Ile	His	Asp	Gln	Pro	Asp	Ile	Asn	Gln
	450					455					460				
Glu	Thr	Gly	Lys	Pro	Glu	Met	Ile	His	Phe	Tyr	Asn	Ser	Thr	Lys	Gly
465					470					475					480
Ala	Val	Asp	Thr	Val	Asp	Gln	Met	Cys	Ser	Ser	Ile	Ser	Thr	Asn	Arg
			485						490					495	
Lys	Thr	Gln	Arg	Trp	Pro	Leu	Cys	Val	Phe	Tyr	Asn	Met	Leu	Asn	Leu
		500						505					510		
Ser	Ile	Ile	Asn	Ala	Tyr	Val	Val	Tyr	Val	Tyr	Asn	Asn	Val	Arg	Asn
		515					520					525			
Asn	Lys	Lys	Pro	Met	Ser	Arg	Arg	Asp	Phe	Val	Ile	Lys	Leu	Gly	Asp
	530					535					540				
Gln	Leu	Met	Glu	Pro	Trp	Leu	Arg	Gln	Arg	Leu	Gln	Thr	Val	Thr	Leu
545					550					555					560
Arg	Arg	Asp	Ile	Lys	Val	Met	Ile	Gln	Asp	Ile	Leu	Gly	Glu	Ser	Ser
			565						570					575	
Asp	Leu	Glu	Ala	Pro	Val	Pro	Ser	Val	Ser	Asn	Val	Arg	Lys	Ile	Tyr
			580					585					590		

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Tyr	Leu	Cys	Pro	Ser	Lys	Ala	Arg	Arg	Met	Thr	Lys	His	Arg	Cys	Ile
		595					600					605			
Lys	Cys	Lys	Gln	Ala	Ile	Cys	Gly	Pro	His	Asn	Ile	Asp	Ile	Cys	Ser
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Arg	Cys	Ile	Glu												
625															
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<212> TYPE: PRT															
<213> ORGANISM: Ctenoplusia agnata															
<400> SEQUENCE: 49															
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1				5					10					15	
Glu	Asn	Glu	Asp	Asp	Tyr	Ser	Pro	His	Asp	Thr	Asp	Ser	Glu	Met	Glu
			20					25					30		
Asp	Cys	Val	Thr	Gln	Asp	Asp	Val	Arg	Ser	Asp	Val	Glu	Asp	Glu	Met
		35					40					45			
Val	Asp	Asn	Ile	Gly	Asn	Gly	Thr	Ser	Pro	Ala	Ser	Arg	His	Glu	Asp
	50					55					60				
Pro	Glu	Thr	Pro	Asp	Pro	Ser	Ser	Glu	Ala	Ser	Asn	Leu	Glu	Val	Thr
65					70					75					80
Leu	Ser	Ser	His	Arg	Ile	Ile	Ile	Leu	Pro	Gln	Arg	Ser	Ile	Arg	Glu
			85						90					95	
Lys	Asn	Asn	His	Ile	Trp	Ser	Thr	Thr	Lys	Gly	Gln	Ser	Ser	Gly	Arg
			100					105					110		
Thr	Ala	Ala	Ile	Asn	Ile	Val	Arg	Thr	Asn	Arg	Gly	Pro	Thr	Arg	Met
		115					120					125			
Cys	Arg	Asn	Ile	Val	Asp	Pro	Leu	Leu	Cys	Phe	Gln	Leu	Phe	Ile	Lys
	130					135					140				
Glu	Glu	Ile	Val	Glu	Glu	Ile	Val	Lys	Trp	Thr	Asn	Val	Glu	Met	Val
145					150					155					160
Gln	Lys	Arg	Val	Asn	Leu	Lys	Asp	Ile	Ser	Ala	Ser	Tyr	Arg	Asp	Thr
			165						170					175	
Asn	Glu	Met	Glu	Ile	Trp	Ala	Ile	Ile	Ser	Met	Leu	Thr	Leu	Ser	Ala
			180					185					190		
Val	Met	Lys	Asp	Asn	His	Leu	Ser	Thr	Asp	Glu	Leu	Phe	Asn	Val	Ser
	195						200					205			
Tyr	Gly	Thr	Arg	Tyr	Val	Ser	Val	Met	Ser	Arg	Glu	Arg	Phe	Glu	Phe
	210					215					220				
Leu	Leu	Arg	Leu	Leu	Arg	Met	Gly	Asp	Lys	Leu	Leu	Arg	Pro	Asn	Leu
225					230					235					240
Arg	Gln	Glu	Asp	Ala	Phe	Thr	Pro	Val	Arg	Lys	Ile	Trp	Glu	Ile	Phe
			245						250					255	
Ile	Asn	Gln	Cys	Arg	Leu	Asn	Tyr	Val	Pro	Gly	Thr	Asn	Leu	Thr	Val
		260						265					270		
Asp	Glu	Gln	Leu	Leu	Gly	Phe	Arg	Gly	Arg	Cys	Pro	Phe	Arg	Met	Tyr
		275					280					285			
Ile	Pro	Asn	Lys	Pro	Asp	Lys	Tyr	Gly	Ile	Lys	Phe	Pro	Met	Val	Cys
	290					295					300				
Asp	Ala	Ala	Thr	Lys	Tyr	Met	Val	Asp	Ala	Ile	Pro	Tyr	Leu	Gly	Lys
305					310					315					320
Ser	Thr	Lys	Thr	Gln	Gly	Leu	Pro	Leu	Gly	Glu	Phe	Tyr	Val	Lys	Glu
				325					330					335	

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Leu	Thr	Gln	Thr	Val	His	Gly	Thr	Asn	Arg	Asn	Val	Thr	Cys	Asp	Asn	
			340					345					350			
Trp	Phe	Thr	Ser	Val	Pro	Leu	Ala	Lys	Ser	Leu	Leu	Asn	Ser	Pro	Tyr	
		355					360					365				
Asn	Leu	Thr	Leu	Val	Gly	Thr	Ile	Arg	Ser	Asn	Lys	Arg	Glu	Ile	Pro	
	370					375					380					
Glu	Glu	Val	Lys	Asn	Ser	Arg	Ser	Arg	Gln	Val	Gly	Ser	Ser	Met	Phe	
385					390					395					400	
Cys	Phe	Asp	Gly	Pro	Leu	Thr	Leu	Val	Ser	Tyr	Lys	Pro	Lys	Pro	Ser	
				405					410					415		
Lys	Met	Val	Phe	Leu	Leu	Ser	Ser	Cys	Asn	Glu	Asp	Ala	Val	Val	Asn	
			420					425					430			
Gln	Ser	Asn	Gly	Lys	Pro	Asp	Met	Ile	Leu	Phe	Tyr	Asn	Gln	Thr	Lys	
		435					440					445				
Gly	Gly	Val	Asp	Ser	Phe	Asp	Gln	Met	Cys	Ser	Ser	Met	Ser	Thr	Asn	
	450					455					460					
Arg	Lys	Thr	Asn	Arg	Trp	Pro	Met	Ala	Val	Phe	Tyr	Gly	Met	Leu	Asn	
465					470					475					480	
Met	Ala	Phe	Val	Asn	Ser	Tyr	Ile	Ile	Tyr	Cys	His	Asn	Met	Leu	Ala	
				485					490					495		
Lys	Lys	Glu	Lys	Pro	Leu	Ser	Arg	Lys	Asp	Phe	Met	Lys	Lys	Leu	Ser	
			500					505				510				
Thr	Asp	Leu	Thr	Thr	Pro	Ser	Met	Gln	Lys	Arg	Leu	Glu	Ala	Pro	Thr	
	515						520					525				
Leu	Lys	Arg	Ser	Leu	Arg	Asp	Asn	Ile	Thr	Asn	Val	Leu	Lys	Ile	Val	
	530					535					540					
Pro	Gln	Ala	Ala	Ile	Asp	Thr	Ser	Phe	Asp	Glu	Pro	Glu	Pro	Lys	Lys	
545					550					555					560	
Arg	Arg	Tyr	Cys	Gly	Phe	Cys	Ser	Tyr	Lys	Lys	Lys	Arg	Met	Thr	Lys	
				565					570					575		
Thr	Gln	Cys	Phe	Lys	Cys	Lys	Lys	Pro	Val	Cys	Gly	Glu	His	Asn	Ile	
		580						585					590			
Asp	Val	Cys	Gln	Asp	Cys	Ile										
		595														
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<213> ORGANISM: Agrotis ipsilon																
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Glu	Asn	Asp	Asp	Asp	Tyr	Ser	Pro	Leu	Asp	Ser	Asp	Ser	Glu	Ala	Glu	
		20						25					30			
Asp	Arg	Val	Val	Glu	Asp	Asp	Val	Trp	Ser	Asp	Asn	Glu	Asp	Ala	Met	
		35					40					45				
Ile	Asp	Tyr	Val	Glu	Asp	Thr	Ser	Arg	Gln	Glu	Asp	Pro	Asp	Asn	Asn	
	50					55					60					
Ile	Ala	Ser	Gln	Glu	Ser	Ala	Asn	Leu	Glu	Val	Thr	Ser	Leu	Thr	Ser	
65					70					75					80	
His	Arg	Ile	Ile	Ser	Leu	Pro	Gln	Arg	Ser	Ile	Cys	Gly	Lys	Asn	Asn	
				85				90						95		
His	Val	Trp	Ser	Thr	Thr	Lys	Gly	Arg	Thr	Thr	Gly	Arg	Thr	Ser	Ala	

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100						105						110					
Ile	Asn	Ile	Ile	Arg	Thr	Asn	Arg	Gly	Pro	Thr	Arg	Met	Cys	Arg	Asn		
115						120						125					
Ile	Val	Asp	Pro	Leu	Leu	Cys	Phe	Gln	Leu	Phe	Ile	Thr	Asp	Glu	Ile		
130						135						140					
Ile	His	Glu	Ile	Val	Lys	Trp	Thr	Asn	Val	Glu	Met	Ile	Val	Lys	Arg		
145						150						155					
160						165						170					
Gln	Asn	Leu	Ile	Asp	Ile	Ser	Ala	Ser	Tyr	Arg	Asp	Thr	Asn	Thr	Met		
175						180						185					
Glu	Met	Trp	Ala	Leu	Val	Gly	Ile	Leu	Thr	Leu	Thr	Ala	Val	Met	Lys		
190						195						200					
Asp	Asn	His	Leu	Ser	Thr	Asp	Glu	Leu	Phe	Asp	Ala	Thr	Phe	Ser	Gly		
205						210						215					
Thr	Arg	Tyr	Val	Ser	Val	Met	Ser	Arg	Glu	Arg	Phe	Glu	Phe	Leu	Ile		
220						225						230					
Arg	Cys	Met	Arg	Met	Asp	Asp	Lys	Thr	Leu	Arg	Pro	Thr	Leu	Arg	Ser		
235						240						245					
Asp	Asp	Ala	Phe	Ile	Pro	Val	Arg	Lys	Leu	Trp	Glu	Ile	Phe	Ile	Asn		
250						255						260					
Gln	Cys	Arg	Leu	Asn	Tyr	Val	Pro	Gly	Gly	Asn	Leu	Thr	Val	Asp	Glu		
265						270						275					
Gln	Leu	Leu	Gly	Phe	Arg	Gly	Arg	Cys	Pro	Phe	Arg	Met	Tyr	Ile	Pro		
280						285						290					
Asn	Lys	Pro	Asp	Lys	Tyr	Gly	Ile	Arg	Phe	Pro	Met	Met	Cys	Asp	Ala		
295						300						305					
Ala	Thr	Lys	Tyr	Met	Ile	Asp	Ala	Ile	Pro	Tyr	Leu	Gly	Lys	Ser	Thr		
310						315						320					
Lys	Thr	Asn	Gly	Leu	Pro	Leu	Gly	Glu	Phe	Tyr	Val	Lys	Glu	Leu	Thr		
325						330						335					
Lys	Thr	Val	His	Gly	Thr	Asn	Arg	Asn	Val	Thr	Cys	Asp	Asn	Trp	Phe		
340						345						350					
Thr	Ser	Ile	Pro	Leu	Ala	Lys	Asn	Met	Leu	Gln	Ala	Pro	Tyr	Asn	Leu		
355						360						365					
Thr	Ile	Val	Gly	Thr	Ile	Arg	Ser	Asn	Lys	Arg	Glu	Ile	Pro	Glu	Glu		
370						375						380					
Ile	Lys	Asn	Ser	Arg	Ser	Arg	Pro	Val	Gly	Ser	Ser	Met	Phe	Cys	Phe		
385						390						395					
Asp	Gly	Pro	Leu	Thr	Leu	Val	Ser	Tyr	Lys	Pro	Lys	Pro	Ser	Arg	Met		
400						405						410					
Val	Phe	Leu	Leu	Ser	Ser	Cys	Asp	Glu	Asn	Ala	Val	Ile	Asn	Glu	Ser		
415						420						425					
Asn	Gly	Lys	Pro	Asp	Met	Ile	Leu	Phe	Tyr	Asn	Gln	Thr	Lys	Gly	Gly		
430						435						440					
Val	Asp	Ser	Phe	Asp	Gln	Met	Cys	Lys	Ser	Met	Ser	Ala	Asn	Arg	Lys		
445						450						455					
Thr	Asn	Arg	Trp	Pro	Met	Ala	Val	Phe	Tyr	Gly	Met	Leu	Asn	Met	Ala		
460						465						470					
Phe	Val	Asn	Ser	Tyr	Ile	Ile	Tyr	Cys	His	Asn	Lys	Ile	Asn	Lys	Gln		
475						480						485					
Lys	Lys	Pro	Ile	Asn	Arg	Lys	Glu	Phe	Met	Lys	Asn	Leu	Ser	Thr	Asp		
490						495						500					
Leu	Thr	Thr	Pro	Trp	Met	Gln	Glu	Arg	Leu	Lys	Ala	Pro	Thr	Leu	Lys		
505						510						515					
520						525						530					

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Arg	Thr	Leu	Arg	Asp	Asn	Ile	Thr	Asn	Val	Leu	Lys	Asn	Val	Val	Pro
530						535					540				
Pro	Ser	Pro	Ala	Asn	Asn	Ser	Glu	Glu	Pro	Gly	Pro	Lys	Lys	Arg	Ser
545					550					555					560
Tyr	Cys	Gly	Phe	Cys	Ser	Tyr	Lys	Lys	Arg	Arg	Met	Thr	Lys	Thr	Gln
				565					570					575	
Phe	Tyr	Lys	Cys	Lys	Lys	Ala	Ile	Cys	Gly	Glu	His	Asn	Ile	Asp	Val
			580					585					590		
Cys	Gln	Asp	Cys	Val											
			595												
<210> SEQ ID NO 51															
<211> LENGTH: 520															
<212> TYPE: PRT															
<213> ORGANISM: Megachile rotundata															
<400> SEQUENCE: 51															
Met	Asn	Gly	Lys	Asp	Ser	Leu	Gly	Glu	Phe	Tyr	Leu	Asp	Asp	Leu	Ser
1				5					10					15	
Asp	Cys	Leu	Asp	Cys	Arg	Ser	Ala	Ser	Ser	Thr	Asp	Asp	Glu	Ser	Asp
			20					25					30		
Ser	Ser	Asn	Ile	Ala	Ile	Arg	Lys	Arg	Cys	Pro	Ile	Pro	Leu	Ile	Tyr
		35					40					45			
Ser	Asp	Ser	Glu	Asp	Glu	Asp	Met	Asn	Asn	Asn	Val	Glu	Asp	Asn	Asn
	50					55					60				
His	Phe	Val	Lys	Glu	Ser	Asn	Arg	Tyr	His	Tyr	Gln	Ile	Val	Glu	Lys
65					70					75					80
Tyr	Lys	Ile	Thr	Ser	Lys	Thr	Lys	Lys	Trp	Lys	Asp	Val	Thr	Val	Thr
				85					90					95	
Glu	Met	Lys	Lys	Phe	Leu	Gly	Leu	Ile	Ile	Leu	Met	Gly	Gln	Val	Lys
			100					105					110		
Lys	Asp	Val	Leu	Tyr	Asp	Tyr	Trp	Ser	Thr	Asp	Pro	Ser	Ile	Glu	Thr
		115					120					125			
Pro	Phe	Phe	Ser	Lys	Val	Met	Ser	Arg	Asn	Arg	Phe	Leu	Gln	Ile	Met
	130					135					140				
Gln	Ser	Trp	His	Phe	Tyr	Asn	Asn	Asn	Asp	Ile	Ser	Pro	Asn	Ser	His
145					150					155					160
Arg	Leu	Val	Lys	Ile	Gln	Pro	Val	Ile	Asp	Tyr	Phe	Lys	Glu	Lys	Phe
				165					170					175	
Asn	Asn	Val	Tyr	Lys	Ser	Asp	Gln	Gln	Leu	Ser	Leu	Asp	Glu	Cys	Leu
			180					185					190		
Ile	Pro	Trp	Arg	Gly	Arg	Leu	Ser	Ile	Lys	Thr	Tyr	Asn	Pro	Ala	Lys
		195					200					205			
Ile	Thr	Lys	Tyr	Gly	Ile	Leu	Val	Arg	Val	Leu	Ser	Glu	Ala	Arg	Thr
	210					215						220			
Gly	Tyr	Val	Ser	Asn	Phe	Cys	Val	Tyr	Ala	Ala	Asp	Gly	Lys	Lys	Ile
225					230					235					240
Glu	Glu	Thr	Val	Leu	Ser	Val	Ile	Gly	Pro	Tyr	Lys	Asn	Met	Trp	His
				245					250					255	
His	Val	Tyr	Gln	Asp	Asn	Tyr	Tyr	Asn	Ser	Val	Asn	Ile	Ala	Lys	Ile
			260					265					270		
Phe	Leu	Lys	Asn	Lys	Leu	Arg	Val	Cys	Gly	Thr	Ile	Arg	Lys	Asn	Arg
		275					280					285			
Ser	Leu	Pro	Gln	Ile	Leu	Gln	Thr	Val	Lys	Leu	Ser	Arg	Gly	Gln	His

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290				295				300							
Gln	Phe	Leu	Arg	Asn	Gly	His	Thr	Leu	Leu	Glu	Val	Trp	Asn	Asn	Gly
305					310					315					320
Lys	Arg	Asn	Val	Asn	Met	Ile	Ser	Thr	Ile	His	Ser	Ala	Gln	Met	Ala
				325					330					335	
Glu	Ser	Arg	Asn	Arg	Ser	Arg	Thr	Ser	Asp	Cys	Pro	Ile	Gln	Lys	Pro
			340					345					350		
Ile	Ser	Ile	Ile	Asp	Tyr	Asn	Lys	Tyr	Met	Lys	Gly	Val	Asp	Arg	Ala
		355					360					365			
Asp	Gln	Tyr	Leu	Ser	Tyr	Tyr	Ser	Ile	Phe	Arg	Lys	Thr	Lys	Lys	Trp
	370					375					380				
Thr	Lys	Arg	Val	Val	Met	Phe	Phe	Ile	Asn	Cys	Ala	Leu	Phe	Asn	Ser
385					390					395					400
Phe	Lys	Val	Tyr	Thr	Thr	Leu	Asn	Gly	Gln	Lys	Ile	Thr	Tyr	Lys	Asn
				405					410					415	
Phe	Leu	His	Lys	Ala	Ala	Leu	Ser	Leu	Ile	Glu	Asp	Cys	Gly	Thr	Glu
			420					425					430		
Glu	Gln	Gly	Thr	Asp	Leu	Pro	Asn	Ser	Glu	Pro	Thr	Thr	Thr	Arg	Thr
		435					440					445			
Thr	Ser	Arg	Val	Asp	His	Pro	Gly	Arg	Leu	Glu	Asn	Phe	Gly	Lys	His
	450					455					460				
Lys	Leu	Val	Asn	Ile	Val	Thr	Ser	Gly	Gln	Cys	Lys	Lys	Pro	Leu	Arg
465					470					475					480
Gln	Cys	Arg	Val	Cys	Ala	Ser	Lys	Lys	Lys	Leu	Ser	Arg	Thr	Gly	Phe
				485					490					495	
Ala	Cys	Lys	Tyr	Cys	Asn	Val	Pro	Leu	His	Lys	Gly	Asp	Cys	Phe	Glu
			500					505					510		
Arg	Tyr	His	Ser	Leu	Lys	Lys	Tyr								
		515					520								
<210> SEQ ID NO 52															
<211> LENGTH: 570															
<212> TYPE: PRT															
<213> ORGANISM: Bombus impatiens															
<400> SEQUENCE: 52															
Met	Asn	Glu	Lys	Asn	Gly	Ile	Gly	Glu	Phe	Tyr	Leu	Asp	Asp	Leu	Ser
1				5					10					15	
Asp	Cys	Pro	Asp	Ser	Tyr	Ser	Arg	Ser	Asn	Ser	Gly	Asp	Glu	Ser	Asp
			20					25					30		
Gly	Ser	Asp	Thr	Ile	Ile	Arg	Lys	Arg	Gly	Ser	Val	Leu	Pro	Pro	Arg
		35					40					45			
Tyr	Ser	Asp	Ser	Glu	Asp	Asp	Glu	Ile	Asn	Asn	Val	Glu	Asp	Asn	Ala
	50					55					60				
Asn	Asn	Val	Glu	Asn	Asn	Asp	Asp	Ile	Trp	Ser	Thr	Asn	Asp	Glu	Ala
65					70				75						80
Ile	Ile	Leu	Glu	Pro	Phe	Glu	Gly	Ser	Pro	Gly	Leu	Lys	Ile	Met	Pro
				85					90					95	
Ser	Ser	Ala	Glu	Ser	Val	Thr	Asp	Asn	Val	Asn	Leu	Phe	Phe	Gly	Asp
			100					105					110		
Asp	Phe	Phe	Glu	His	Leu	Val	Arg	Glu	Ser	Asn	Arg	Tyr	His	Tyr	Gln
		115					120					125			
Val	Met	Glu	Lys	Tyr	Lys	Ile	Pro	Ser	Lys	Ala	Lys	Lys	Trp	Thr	Asp
	130					135					140				

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Ile	Thr	Val	Pro	Glu	Met	Lys	Lys	Phe	Leu	Gly	Leu	Ile	Val	Leu	Met
145					150					155					160
Gly	Gln	Ile	Lys	Lys	Asp	Val	Leu	Tyr	Asp	Tyr	Trp	Ser	Thr	Asp	Pro
				165					170					175	
Ser	Ile	Glu	Thr	Pro	Phe	Phe	Ser	Gln	Val	Met	Ser	Arg	Asn	Arg	Phe
			180					185					190		
Val	Gln	Ile	Met	Gln	Ser	Trp	His	Phe	Cys	Asn	Asn	Asp	Asn	Ile	Pro
		195					200					205			
His	Asp	Ser	His	Arg	Leu	Ala	Lys	Ile	Gln	Pro	Val	Ile	Asp	Tyr	Phe
	210					215					220				
Arg	Arg	Lys	Phe	Asn	Asp	Val	Tyr	Lys	Pro	Cys	Gln	Gln	Leu	Ser	Leu
225					230					235					240
Asp	Glu	Ser	Ile	Ile	Pro	Trp	Arg	Gly	Arg	Leu	Ser	Ile	Lys	Thr	Tyr
				245					250					255	
Asn	Pro	Ala	Lys	Ile	Thr	Lys	Tyr	Gly	Ile	Leu	Val	Arg	Val	Leu	Ser
			260					265					270		
Glu	Ala	Val	Thr	Gly	Tyr	Val	Cys	Asn	Phe	Asp	Val	Tyr	Ala	Ala	Asp
		275					280					285			
Gly	Lys	Lys	Leu	Glu	Asp	Thr	Ala	Val	Ile	Glu	Pro	Tyr	Lys	Asn	Ile
	290					295					300				
Trp	His	Gln	Ile	Tyr	Gln	Asp	Asn	Tyr	Tyr	Asn	Ser	Val	Lys	Met	Ala
305					310					315					320
Arg	Ile	Leu	Leu	Lys	Asn	Lys	Val	Arg	Val	Cys	Gly	Thr	Ile	Arg	Lys
				325					330					335	
Asn	Arg	Gly	Leu	Pro	Arg	Ser	Leu	Lys	Thr	Ile	Gln	Leu	Ser	Arg	Gly
			340					345					350		
Gln	Tyr	Glu	Phe	Arg	Arg	Asn	His	Gln	Ile	Leu	Leu	Glu	Val	Trp	Asn
		355					360					365			
Asn	Gly	Arg	Arg	Asn	Val	Asn	Met	Ile	Ser	Thr	Ile	His	Ser	Ala	Gln
	370					375					380				
Leu	Met	Glu	Ser	Arg	Ser	Lys	Ser	Lys	Arg	Ser	Asp	Val	Pro	Ile	Gln
385					390					395					400
Lys	Pro	Asn	Ser	Ile	Ile	Asp	Tyr	Asn	Lys	Tyr	Met	Lys	Gly	Val	Asp
				405					410					415	
Arg	Ala	Asp	Gln	Tyr	Leu	Ala	Tyr	Tyr	Ser	Ile	Phe	Arg	Lys	Thr	Lys
			420					425					430		
Lys	Trp	Thr	Lys	Arg	Val	Val	Met	Phe	Phe	Ile	Asn	Cys	Ala	Leu	Phe
		435					440					445			
Asn	Ser	Phe	Arg	Val	Tyr	Thr	Ile	Leu	Asn	Gly	Lys	Asn	Ile	Thr	Tyr
	450					455					460				
Lys	Asn	Phe	Leu	His	Lys	Val	Ala	Val	Ser	Trp	Ile	Glu	Asp	Gly	Glu
465					470					475					480
Thr	Asn	Cys	Thr	Glu	Gln	Asp	Asp	Asn	Leu	Pro	Asn	Ser	Glu	Pro	Thr
				485					490					495	
Arg	Arg	Ala	Pro	Arg	Leu	Asp	His	Pro	Gly	Arg	Leu	Ser	Asn	Tyr	Gly
			500					505					510		
Lys	His	Lys	Leu	Ile	Asn	Ile	Val	Thr	Ser	Gly	Arg	Ser	Leu	Lys	Pro
		515					520					525			
Gln	Arg	Gln	Cys	Arg	Val	Cys	Ala	Val	Gln	Lys	Lys	Arg	Ser	Arg	Thr
	530					535					540				
Cys	Phe	Val	Cys	Lys	Phe	Cys	Asn	Val	Pro	Leu	His	Lys	Gly	Asp	Cys
545					550					555					560
Phe	Glu	Arg	Tyr	His	Thr	Leu	Lys	Lys	Tyr						

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565	570
<div><210> SEQ ID NO 53</div> <div><211> LENGTH: 345</div> <div><212> TYPE: PRT</div> <div><213> ORGANISM: Mamestra brassicae</div> <div><400> SEQUENCE: 53</div>	
Met Phe Ser Phe Val Pro Asn Lys Glu Gln Thr Arg Thr Val Leu Ile	
1 5 10 15	
Phe Cys Phe His Leu Lys Thr Thr Ala Ala Glu Ser His Arg Pro Leu	
20 25 30	
Val Glu Ala Phe Gly Glu Gln Val Pro Thr Val Lys Thr Cys Glu Arg	
35 40 45	
Trp Phe Gln Arg Phe Lys Ser Gly Asp Phe Asp Val Asp Asp Lys Glu	
50 55 60	
His Gly Lys Pro Pro Lys Arg Tyr Glu Asp Ala Glu Leu Gln Ala Leu	
65 70 75 80	
Leu Asp Glu Asp Asp Ala Gln Thr Gln Lys Gln Leu Ala Glu Gln Leu	
85 90 95	
Glu Val Ser Gln Gln Ala Val Ser Asn Arg Leu Arg Glu Gly Gly Lys	
100 105 110	
Ile Gln Lys Val Gly Arg Trp Val Pro His Glu Leu Asn Glu Arg Gln	
115 120 125	
Arg Glu Arg Arg Lys Asn Thr Cys Glu Ile Leu Leu Ser Arg Tyr Lys	
130 135 140	
Arg Lys Ser Phe Leu His Arg Ile Val Thr Gly Glu Glu Lys Trp Ile	
145 150 155 160	
Phe Phe Val Asn Pro Lys Arg Lys Lys Ser Tyr Val Asp Pro Gly Gln	
165 170 175	
Pro Ala Thr Ser Thr Ala Arg Pro Asn Arg Phe Gly Lys Lys Thr Arg	
180 185 190	
Leu Cys Val Trp Trp Asp Gln Ser Gly Val Ile Tyr Tyr Glu Leu Leu	
195 200 205	
Lys Pro Gly Glu Thr Val Asn Thr Ala Arg Tyr Gln Gln Gln Leu Ile	
210 215 220	
Asn Leu Asn Arg Ala Leu Gln Arg Lys Arg Pro Glu Tyr Gln Lys Arg	
225 230 235 240	
Gln His Arg Val Ile Phe Leu His Asp Asn Ala Pro Ser His Thr Ala	
245 250 255	
Arg Ala Val Arg Asp Thr Leu Glu Thr Leu Asn Trp Glu Val Leu Pro	
260 265 270	
His Ala Ala Tyr Ser Pro Asp Leu Ala Pro Ser Asp Tyr His Leu Phe	
275 280 285	
Ala Ser Met Gly His Ala Leu Ala Glu Gln Arg Phe Asp Ser Tyr Glu	
290 295 300	
Ser Val Glu Glu Trp Leu Asp Glu Trp Phe Ala Ala Lys Asp Asp Glu	
305 310 315 320	
Phe Tyr Trp Arg Gly Ile His Lys Leu Pro Glu Arg Trp Asp Asn Cys	
325 330 335	
Val Ala Ser Asp Gly Lys Tyr Phe Glu	
340 345	
<div><210> SEQ ID NO 54</div> <div><211> LENGTH: 347</div>	

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<212> TYPE: PRT
<213> ORGANISM: Mayetiola destructor

<400> SEQUENCE: 54

Met Glu Asn Phe Glu Asn Trp Arg Lys Arg Arg His Leu Arg Glu Val
1 5 10 15

Leu Leu Gly His Phe Phe Ala Lys Lys Thr Ala Ala Glu Ser His Arg
20 25 30

Leu Leu Val Glu Val Tyr Gly Glu His Ala Leu Ala Lys Thr Gln Cys
35 40 45

Phe Glu Trp Phe Gln Arg Phe Lys Ser Gly Asp Phe Asp Thr Glu Asp
50 55 60

Lys Glu Arg Pro Gly Gln Pro Lys Lys Phe Glu Asp Glu Glu Leu Glu
65 70 75 80

Ala Leu Leu Asp Glu Asp Cys Cys Gln Thr Gln Glu Glu Leu Ala Lys
85 90 95

Ser Leu Gly Val Thr Gln Gln Ala Ile Ser Lys Arg Leu Lys Ala Ala
100 105 110

Gly Tyr Ile Gln Lys Gln Gly Asn Trp Val Pro His Glu Leu Lys Pro
115 120 125

Arg Asp Val Glu Arg Arg Phe Cys Met Ser Glu Met Leu Leu Gln Arg
130 135 140

His Lys Lys Lys Ser Phe Leu Ser Arg Ile Ile Thr Gly Asp Glu Lys
145 150 155 160

Trp Ile His Tyr Asp Asn Ser Lys Arg Lys Lys Ser Tyr Val Lys Arg
165 170 175

Gly Gly Arg Ala Lys Ser Thr Pro Lys Ser Asn Leu His Gly Ala Lys
180 185 190

Val Met Leu Cys Ile Trp Trp Asp Gln Arg Gly Val Leu Tyr Tyr Glu
195 200 205

Leu Leu Glu Pro Gly Gln Thr Ile Thr Gly Asp Leu Tyr Arg Thr Gln
210 215 220

Leu Ile Arg Leu Lys Gln Ala Leu Ala Glu Lys Arg Pro Glu Tyr Ala
225 230 235 240

Lys Arg His Gly Ala Val Ile Phe His His Asp Asn Ala Arg Pro His
245 250 255

Val Ala Leu Pro Val Lys Asn Tyr Leu Glu Asn Ser Gly Trp Glu Val
260 265 270

Leu Pro His Pro Pro Tyr Ser Pro Asp Leu Ala Pro Ser Asp Tyr His
275 280 285

Leu Phe Arg Ser Met Gln Asn Asp Leu Ala Gly Lys Arg Phe Thr Ser
290 295 300

Glu Gln Gly Ile Arg Lys Trp Leu Asp Ser Phe Leu Ala Ala Lys Pro
305 310 315 320

Ala Lys Phe Phe Glu Lys Gly Ile His Glu Leu Ser Glu Arg Trp Glu
325 330 335

Lys Val Ile Ala Ser Asp Gly Gln Tyr Phe Glu
340 345

<210> SEQ ID NO 55
<211> LENGTH: 342
<212> TYPE: PRT
<213> ORGANISM: Apis mellifera

<400> SEQUENCE: 55

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Met	Glu	Asn	Gln	Lys	Glu	His	Tyr	Arg	His	Ile	Leu	Leu	Phe	Tyr	Phe
1				5					10					15	
Arg	Lys	Gly	Lys	Asn	Ala	Ser	Gln	Ala	His	Lys	Lys	Leu	Cys	Ala	Val
			20					25					30		
Tyr	Gly	Asp	Glu	Ala	Leu	Lys	Glu	Arg	Gln	Cys	Gln	Asn	Trp	Phe	Asp
		35					40					45			
Lys	Phe	Arg	Ser	Gly	Asp	Phe	Ser	Leu	Lys	Asp	Glu	Lys	Arg	Ser	Gly
	50					55					60				
Arg	Pro	Val	Glu	Val	Asp	Asp	Asp	Leu	Ile	Lys	Ala	Ile	Ile	Asp	Ser
65					70					75					80
Asp	Arg	His	Ser	Thr	Thr	Arg	Glu	Ile	Ala	Glu	Lys	Leu	His	Val	Ser
				85					90					95	
His	Thr	Cys	Ile	Glu	Asn	His	Leu	Lys	Gln	Leu	Gly	Tyr	Val	Gln	Lys
			100					105					110		
Leu	Asp	Thr	Trp	Val	Pro	His	Glu	Leu	Lys	Glu	Lys	His	Leu	Thr	Gln
		115					120					125			
Arg	Ile	Asn	Ser	Cys	Asp	Leu	Leu	Lys	Lys	Arg	Asn	Glu	Asn	Asp	Pro
	130					135					140				
Phe	Leu	Lys	Arg	Leu	Ile	Thr	Gly	Asp	Glu	Lys	Trp	Val	Val	Tyr	Asn
145					150					155					160
Asn	Ile	Lys	Arg	Lys	Arg	Ser	Trp	Ser	Arg	Pro	Arg	Glu	Pro	Ala	Gln
				165					170					175	
Thr	Thr	Ser	Lys	Ala	Gly	Ile	His	Arg	Lys	Lys	Val	Leu	Leu	Ser	Val
			180					185					190		
Trp	Trp	Asp	Tyr	Lys	Gly	Ile	Val	Tyr	Phe	Glu	Leu	Leu	Pro	Pro	Asn
		195					200					205			
Arg	Thr	Ile	Asn	Ser	Val	Val	Tyr	Ile	Glu	Gln	Leu	Thr	Lys	Leu	Asn
	210					215					220				
Asn	Ala	Val	Glu	Glu	Lys	Arg	Pro	Glu	Leu	Thr	Asn	Arg	Lys	Gly	Val
225					230					235					240
Val	Phe	His	His	Asp	Asn	Ala	Arg	Pro	His	Thr	Ser	Leu	Val	Thr	Arg
				245					250					255	
Gln	Lys	Leu	Leu	Glu	Leu	Gly	Trp	Asp	Val	Leu	Pro	His	Pro	Pro	Tyr
			260					265					270		
Ser	Pro	Asp	Leu	Ala	Pro	Ser	Asp	Tyr	Phe	Leu	Phe	Arg	Ser	Leu	Gln
		275					280					285			
Asn	Ser	Leu	Asn	Gly	Lys	Asn	Phe	Asn	Asn	Asp	Asp	Asp	Ile	Lys	Ser
	290					295					300				
Tyr	Leu	Ile	Gln	Phe	Phe	Ala	Asn	Lys	Asn	Gln	Lys	Phe	Tyr	Glu	Arg
305					310					315					320
Gly	Ile	Met	Met	Leu	Pro	Glu	Arg	Trp	Gln	Lys	Val	Ile	Asp	Gln	Asn
				325					330					335	
Gly	Gln	His	Ile	Thr	Glu										
			340												
<210> SEQ ID NO 56															
<211> LENGTH: 345															
<212> TYPE: PRT															
<213> ORGANISM: Messor bouvieri															
<400> SEQUENCE: 56															
Met	Ser	Ser	Phe	Val	Pro	Glu	Asn	Val	His	Leu	Arg	His	Ala	Leu	Leu
1				5					10					15	
Phe	Leu	Phe	His	Gln	Lys	Lys	Arg	Ala	Ala	Glu	Ser	His	Arg	Leu	Leu
			20					25					30		

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Val	Glu	Thr	Tyr	Gly	Glu	His	Ala	Pro	Thr	Ile	Arg	Thr	Cys	Glu	Thr
		35					40					45			
Trp	Phe	Arg	Gln	Phe	Lys	Cys	Gly	Asp	Phe	Asn	Val	Gln	Asp	Lys	Glu
	50					55				60					
Arg	Pro	Gly	Arg	Pro	Lys	Thr	Phe	Glu	Asp	Ala	Glu	Leu	Gln	Glu	Leu
65					70				75					80	
Leu	Asp	Glu	Asp	Ser	Thr	Gln	Thr	Gln	Lys	Gln	Leu	Ala	Glu	Lys	Leu
				85					90					95	
Asn	Val	Ser	Arg	Val	Ala	Ile	Cys	Glu	Arg	Leu	Gln	Ala	Met	Gly	Lys
			100					105					110		
Ile	Gln	Lys	Met	Gly	Arg	Trp	Val	Pro	His	Glu	Leu	Asn	Asp	Arg	Gln
		115					120					125			
Met	Glu	Asn	Arg	Lys	Ile	Val	Ser	Glu	Met	Leu	Leu	Gln	Arg	Tyr	Glu
	130					135					140				
Arg	Lys	Ser	Phe	Leu	His	Arg	Ile	Val	Thr	Gly	Asp	Glu	Lys	Trp	Ile
145					150					155					160
Tyr	Phe	Glu	Asn	Pro	Lys	Arg	Lys	Lys	Ser	Trp	Leu	Ser	Pro	Gly	Glu
				165					170					175	
Ala	Gly	Pro	Ser	Thr	Ala	Arg	Pro	Asn	Arg	Phe	Gly	Arg	Lys	Thr	Met
			180					185					190		
Leu	Cys	Val	Trp	Trp	Asp	Gln	Ile	Gly	Val	Val	Tyr	Tyr	Glu	Leu	Leu
		195				200						205			
Lys	Pro	Gly	Glu	Thr	Val	Asn	Thr	Asp	Arg	Tyr	Arg	Gln	Gln	Met	Ile
	210					215				220					
Asn	Leu	Asn	Cys	Ala	Leu	Ile	Glu	Lys	Arg	Pro	Gln	Tyr	Ala	Gln	Arg
225					230					235					240
His	Asp	Lys	Val	Ile	Leu	Gln	His	Asp	Asn	Ala	Pro	Ser	His	Thr	Ala
			245						250					255	
Lys	Pro	Val	Lys	Glu	Met	Leu	Lys	Ser	Leu	Gly	Trp	Glu	Val	Leu	Ser
		260						265					270		
His	Pro	Pro	Tyr	Ser	Pro	Asp	Leu	Ala	Pro	Ser	Asp	Tyr	His	Leu	Phe
		275					280					285			
Ala	Ser	Met	Gly	His	Ala	Leu	Ala	Glu	Gln	His	Phe	Ala	Asp	Phe	Glu
		290				295					300				
Glu	Val	Lys	Lys	Trp	Leu	Asp	Glu	Trp	Phe	Ser	Ser	Lys	Glu	Lys	Leu
305					310					315					320
Phe	Phe	Trp	Asn	Gly	Ile	His	Lys	Leu	Ser	Glu	Arg	Trp	Thr	Lys	Cys
			325						330					335	
Ile	Glu	Ser	Asn	Gly	Gln	Tyr	Phe	Glu							
		340						345							
<210> SEQ ID NO 57															
<211> LENGTH: 594															
<212> TYPE: PRT															
<213> ORGANISM: Trichoplusia ni															
<400> SEQUENCE: 57															
Met	Gly	Ser	Ser	Leu	Asp	Asp	Glu	His	Ile	Leu	Ser	Ala	Leu	Leu	Gln
1				5					10					15	
Ser	Asp	Asp	Glu	Leu	Val	Gly	Glu	Asp	Ser	Asp	Ser	Glu	Val	Ser	Asp
			20					25					30		
His	Val	Ser	Glu	Asp	Asp	Val	Gln	Ser	Asp	Thr	Glu	Glu	Ala	Phe	Ile
		35					40					45			
Asp	Glu	Val	His	Glu	Val	Gln	Pro	Thr	Ser	Ser	Gly	Ser	Glu	Ile	Leu

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50				55				60							
Asp 65	Glu	Gln	Asn	Val 70	Ile	Glu	Gln	Pro	Gly	Ser 75	Ser	Leu	Ala	Ser	Asn 80
Arg	Ile	Leu	Thr 85	Leu	Pro	Gln	Arg	Thr	Ile 90	Arg	Gly	Lys	Asn	Lys 95	His
Cys	Trp	Ser	Thr 100	Ser	Lys	Ser	Thr	Arg 105	Arg	Ser	Arg	Val	Ser 110	Ala	Leu
Asn	Ile	Val 115	Arg	Ser	Gln	Arg	Gly 120	Pro	Thr	Arg	Met	Cys 125	Arg	Asn	Ile
Tyr 130	Asp	Pro	Leu	Leu	Cys	Phe 135	Lys	Leu	Phe	Phe	Thr 140	Asp	Glu	Ile	Ile
Ser 145	Glu	Ile	Val	Lys	Trp 150	Thr	Asn	Ala	Glu	Ile 155	Ser	Leu	Lys	Arg	Arg 160
Glu	Ser	Met	Thr 165	Ser	Ala	Thr	Phe	Arg	Asp 170	Thr	Asn	Glu	Asp	Glu 175	Ile
Tyr	Ala	Phe	Phe 180	Gly	Ile	Leu	Val	Met 185	Thr	Ala	Val	Arg	Lys 190	Asp	Asn
His	Met	Ser 195	Thr	Asp	Asp	Leu	Phe 200	Asp	Arg	Ser	Leu	Ser 205	Met	Val	Tyr
Val	Ser 210	Val	Met	Ser	Arg	Asp 215	Arg	Phe	Asp	Phe	Leu	Ile	Arg	Cys	Leu
Arg 225	Met	Asp	Asp	Lys	Ser 230	Ile	Arg	Pro	Thr	Leu 235	Arg	Glu	Asn	Asp	Val 240
Phe	Thr	Pro	Val 245	Arg	Lys	Ile	Trp	Asp	Leu 250	Phe	Ile	His	Gln	Cys 255	Ile
Gln	Asn	Tyr	Thr 260	Pro	Gly	Ala	His 265	Leu	Thr	Ile	Asp	Glu	Gln 270	Leu	Leu
Gly	Phe	Arg 275	Gly	Arg	Cys	Pro	Phe 280	Arg	Val	Tyr	Ile	Pro 285	Asn	Lys	Pro
Ser 290	Lys	Tyr	Gly	Ile	Lys	Ile 295	Leu	Met	Met	Cys	Asp 300	Ser	Gly	Thr	Lys
Tyr 305	Met	Ile	Asn	Gly	Met 310	Pro	Tyr	Leu	Gly 315	Arg	Gly	Thr	Gln	Thr	Asn 320
Gly	Val	Pro	Leu 325	Gly	Glu	Tyr	Tyr	Val	Lys 330	Glu	Leu	Ser	Lys	Pro 335	Val
His	Gly	Ser	Cys 340	Arg	Asn	Ile	Thr	Cys 345	Asp	Asn	Trp	Phe	Thr 350	Ser	Ile
Pro	Leu	Ala 355	Lys	Asn	Leu	Leu 360	Gln	Glu	Pro	Tyr	Lys 365	Leu	Thr	Ile	Val
Gly	Thr 370	Val	Arg	Ser	Asn	Lys 375	Arg	Glu	Ile	Pro	Glu 380	Val	Leu	Lys	Asn
Ser 385	Arg	Ser	Arg	Pro	Val 390	Gly	Thr	Ser	Met	Phe 395	Cys	Phe	Asp	Gly	Pro 400
Leu	Thr	Leu	Val 405	Ser	Tyr	Lys	Pro	Lys	Pro 410	Ala	Lys	Met	Val	Tyr 415	Leu
Leu	Ser	Ser	Cys 420	Asp	Glu	Asp	Ala	Ser 425	Ile	Asn	Glu	Ser	Thr 430	Gly	Lys
Pro	Gln	Met 435	Val	Met	Tyr	Tyr	Asn 440	Gln	Thr	Lys	Gly 445	Gly	Val	Asp	Thr
Leu 450	Asp	Gln	Met	Cys	Ser	Val 455	Met	Thr	Cys	Ser	Arg 460	Lys	Thr	Asn	Arg
Trp 465	Pro	Met	Ala	Leu	Leu	Tyr 470	Gly	Met	Ile	Asn 475	Ile	Ala	Cys	Ile	Asn 480

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Ser	Phe	Ile	Ile	Tyr	Ser	His	Asn	Val	Ser	Ser	Lys	Gly	Glu	Lys	Val	
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Gln	Ser	Arg	Lys	Lys	Phe	Met	Arg	Asn	Leu	Tyr	Met	Ser	Leu	Thr	Ser	
			500					505					510			
Ser	Phe	Met	Arg	Lys	Arg	Leu	Glu	Ala	Pro	Thr	Leu	Lys	Arg	Tyr	Leu	
		515					520					525				
Arg	Asp	Asn	Ile	Ser	Asn	Ile	Leu	Pro	Lys	Glu	Val	Pro	Gly	Thr	Ser	
	530					535					540					
Asp	Asp	Ser	Thr	Glu	Glu	Pro	Val	Met	Lys	Lys	Arg	Thr	Tyr	Cys	Thr	
545					550				555						560	
Tyr	Cys	Pro	Ser	Lys	Ile	Arg	Arg	Lys	Ala	Asn	Ala	Ser	Cys	Lys	Lys	
				565				570						575		
Cys	Lys	Lys	Val	Ile	Cys	Arg	Glu	His	Asn	Ile	Asp	Met	Cys	Gln	Ser	
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Cys	Phe															
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<212> TYPE: DNA																
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tgcg	ttt	gtc	tatat	gttat	tttccac	cat	attgcc	gtct	tttggc	aatg	tgagg	gccc				120
gaaac	ctgg	c	cctgt	cttct	tgacg	agcat	tcctag	gggt	ctttccc	ctc	tcgcc	aaag				180
aatgc	aagg	t	ctgtt	gaatg	tcgtg	aagga	agcag	ttcct	ctgga	agctt	cttga	agaca				240
aaca	acgt	ct	gtagc	gacc	tttgc	aggca	gcgga	acccc	ccac	ctgg	c	acagg	tcct			300
ctgc	ggcc	aa	aagcc	acgtg	tata	agatac	acctg	caaag	gcgg	caca	ac	cccag	tgcc			360
cg	ttgt	gagt	tggat	agttg	tggaa	agagt	caaat	ggctc	tcct	caag	c	tattc	aaca			420
ggg	gctg	aag	gatg	cccaga	aggt	accca	ttgt	atggga	tctg	atct	gg	ggc	ctcg	gta		480
cacat	gcttt	acat	gtgtt	tt	agtc	gaggt	aaaaa	acgtc	tagg	cccc		gaacc	acgg			540
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<211> LENGTH: 589																
<212> TYPE: DNA																
<213> ORGANISM: Encephalomyocarditis virus																
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tgcg	ttt	gtc	tatat	gttat	tttccac	cat	attgcc	gtct	tttggc	aatg	tgagg	gccc				120
gaaac	ctgg	c	cctgt	cttct	tgacg	agcat	tcctag	gggt	ctttccc	ctc	tcgcc	aaag				180
aatgc	aagg	t	ctgtt	gaatg	tcgtg	aagga	agcag	ttcct	ctgga	agctt	cttga	agaca				240
aaca	acgt	ct	gtagc	gacc	tttgc	aggca	gcgga	acccc	ccac	ctgg	c	acagg	tcct			300
ctgc	ggcc	aa	aagcc	acgtg	tata	agatac	acctg	caaag	gcgg	caca	ac	cccag	tgcc			360
cg	ttgt	gagt	tggat	agttg	tggaa	agagt	caaat	ggctc	tcct	caag	c	tattc	aaca			420
ggg	gctg	aag	gatg	cccaga	aggt	accca	ttgt	atggga	tctg	atct	gg	ggc	ctcg	gta		480
cacat	gcttt	acat	gtgtt	tt	agtc	gaggt	aaaaa	acgtc	tagg	cccc		gaacc	acgg			540
gacgt	ggttt	tcct	ttgaa		aacac	gatga	taat	atgg	cc	aca	ac	ctag				589

<210> SEQ ID NO 60
<211> LENGTH: 837
<212> TYPE: DNA
<213> ORGANISM: Cardio virus

<400> SEQUENCE: 60
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acgaaatcag aacgagatgt tctccctccc ttgccgcttg ttcacacca cttgattgat 120
tcggcctttt gtgacaagcc ccttggtgaa aggacctctc tctttctggc gtggttggaa 180
ttaacatcct ttccgacgaa agtgctatca tgcctccccg attatgtgat gttttctgcc 240
ctgctgggcg gagcattctc gggttgagaa atctcaaacc ttttcttttg gaactttggt 300
tcccccggtc taagccgctc ggaatatgac agggttattt tcttaatctt atttctactt 360
ttatgggttc tatccataaa aagggtatgt gctgccccct ccttcttttg agaattcaca 420
cggcggtctt tccgtctctc aacaagtgtg aatgcagcat gccggaaacg gtgaagaaaa 480
cagttttctg cggaagtcta gagtgacat cgaaacagct gtagcgacct cacagtagca 540
gcggactccc ctcttggtga caagagcctc tgcggccaaa agccccgtgg attagatcca 600
ctgctgtgag cggtgcaacc ccagcacctt gattcgatgg tcattctcta cggaatcaga 660
gaatggtttt cctaagccct ccggtagaga agccaagaat gtctgaagg taccgccgt 720
gcgggatctg atcagaagac caattgacag tgctttacac tgccactttg gtttaaaaat 780
tgtcacagct tctccaaacc aagtggctt ggttttccaa ctttattgaa tggcaat 837

<210> SEQ ID NO 61
<211> LENGTH: 1064
<212> TYPE: DNA
<213> ORGANISM: Theiler's murine encephalomyocarditis virus

<400> SEQUENCE: 61
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atcatcccc ggttaccccc ttctgacgcg ggtactgcga tagtgccacc ccagtctttc 120
ctactcccgga ctcccgacct taaccaggt tctctggaac aggaacacca atttactcat 180
cccctggatg ctgactaatc agaggaacgt cagcattttc cggcccaggc taagagaagt 240
agataagtta gaatccaaat tgatttatca tccccttgac gaattcgctg tggaaaaaca 300
cctctcactt gcccctcttc acaccatta atttaattcg gcctctgtgt tgagccccctt 360
gttgaagtgt ttccctccat cgcgacgtgg ttggagatct aagtcaaccg actccgacga 420
aactaccatc atgcctcccc gattatgtga tgctttctgc cctgctgggt ggagcacctt 480
cgggttgaga aaaccttctt cctttttcct tggactccgg tcccccggtc taagccgctc 540
ggaatatgac aggggtattt tcacctctt ttttctactc cacagtgttc tatactgtgg 600
aagggtatgt gttgccccct ccttcttggga gaacgtgcgc ggcggtcttt ccgtctctcg 660
acaagcggcg gtgcaacata cagagtaacg cgaagaaagc agttctcggt ctagctctag 720
tgcccacaag aaaacagctg tagcgaccac acaaaggcag cggaaccccc ctcttggtaa 780
caggagcctc tgcggccaaa agccacgtgg ataagatcca cctttgtgtg cggtgcaacc 840
ccagcacctt ggtttcttgg tgacactcta gtgaaccctt gaatggcaat ctcaagcgcc 900
tctgtaggga agccaagaat gtccaggagg tacccttcc tctcggaagg gatctgacct 960
ggagacacat cacacgtgct ttacacctgt gcttggtgtt aaaaattggt acagcttccc 1020

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gaaaccogta tacaccggac cttttctccc ctccctctcc acttaccttt ttcccctctt	60
cggcatgaaa caaggattat tcaagtggaa acgcgattta atatgcggct ggccaccgcg	120
gaataacggc aattgtgtat ctgctggaag ccaagcctgc ctagccgata gcccttgacc	180
gggtgtgtag gatagcccag gaaccagcaa tacgcgacag gttatggtag agtagatacc	240
tagccagggg caatgggact gcattgcata tccctaatga accattgaga tttctctggt	300
cattaccogg tgatggttac tagagggggg cctctagtag tagatctata ctgcctgata	360
gggtcgcggc tggccgacca tgacctgtat agtcagttga tttgagcaat	410
<div><210> SEQ ID NO 63</div> <div><211> LENGTH: 1052</div> <div><212> TYPE: DNA</div> <div><213> ORGANISM: Sikhote-Alin virus</div> <div><400> SEQUENCE: 63</div>	
ggggatctcc cccgcggcag ctggttacag ctgtcgcgga cggagatcat cccccggcca	60
ccccctttcg acgcgggtac tgcgatagtg ccacccagct ctttctact cccgactccc	120
gactctaacc caggttcctt ggaacaggaa caccaatata ctcatcccct ggatgctgac	180
taatcagagg aacgtcagca ttttcgggcc caggctaaga gaagtagata agttagattc	240
caaattgatt tatcatcccc ttgacgaatt cgcgttgga atgcacctct cacttgccgc	300
tctccacacc cattaacttg attcggcctc tgtgttgagc cccttgttga agtgcttccc	360
tccatcgtga cgtgggttga gatctaagtc aaccgactcc gacgaaacta ccatcatgcc	420
tccccgatta tgtgatgctt tctgcctgc tgggtggagc atcctcgggt tgagaaaacc	480
tttctccttt ttccttggac cccggctccc cgggtctaag cgcttggaat aagacagggt	540
tatcttcacc tcttccttct tctacttcat agtgttctat actatgaaag ggtatgtgtc	600
gccccctcct tctttggaga acacgcgcgg cggctcttcc gtctctcgaa aagcgcgtgt	660
gcgacatgca gagaaccgtg aagaaagcag tttgcggact agcttttagtg cccacaagaa	720
aacagctgta gcgaccacac aaaggcagcg gacccccct cctggcaaca ggagcctctg	780
cggccaaaag ccacgtggat aagatccacc tttgtgtgcg gcacaacccc agtgccctgg	840
tttcttggtg acacttcagt gaaaacgcaa atggcgatct gaagcgctc tgtaggaaag	900
ccaagaatgt ccaggaggtg ccccttcctt cgggaaggga tctgacctgg agacacatca	960
catgtgcttt acacctgtgc ttgtgtttaa aaattgtcac agctttccca aaccaagtgg	1020
tcttggtttt cactctttaa actgatttca ct	1052
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cactcctaac ccgactccca cctcgggtcc aatgaaagga ccaaataatt catcccttga	180	
atgttggcta atcagaggaa agtcagcatt ttccggccca ggctaagaga aacatataag	240	
ttagatttga ttcaaattcc ttgacgaatt cgaaacgaga tgttctccct cccttgccgc	300	
ttgttcacac ccatacattt aattcggcct cctgtgacaa gcccctcggg gaaagaacct	360	
ctctcttttc gacgtggttg gaattaacat catttccgac gaaagtgcta tcatgcctcc	420	
ccgattatgt gatgttttct gccctgctgg gcggagcatt ctcggttga gaaaccttga	480	
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ttttctttat cttatttcta ctttcacggg ttctatccgt gaaaagggtta cgtgttgccc	600	
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aaaagccccg tggataagat ccactgctgt gagcggtgca accccagcac cctgattcga	840	
tgaacgttct ttacggaacc agaggatggg tttcctaagc cctccggtag agaagccaag	900	
aatgtcctga aggtaccccg cgtgcgggat ctgatcaggg gaccaattga ctgtgcttta	960	
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<211> LENGTH: 741		
<212> TYPE: DNA		
<213> ORGANISM: Echovirus		
<400> SEQUENCE: 67		
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cataccgatc aatagtgggc gtggcacacc agccgtgtct agatcaagca ctctgtttc	180	
cccggaccga gtatcaatag actgctcacg cggttgaagg agaaaacgtt cgttatccgg	240	
ctaactactt cgaaaaacct agtaacacca tgaaagttgc ggagtgtttc actcagcact	300	
tccccagtgt agatcaggtc gatgagtcac cgcattcttc acgggcgacc gtggcgggtgg	360	
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aagagtctat tgagctagtt agtagtctc cggccctga atgcggataa tctaactgt	480	
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cgactacttt ggggtgtccgt gtttcctttt attccaaatc tggctgctta tggtgacaat	600	
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<212> TYPE: DNA		
<213> ORGANISM: Picornavirus		
<400> SEQUENCE: 68		

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ccaccacgga ataacggcaa ttgtctacat gtgggaagtg caacctccct agccgataac	180
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tctctgctca ttaccgggtg atgggtgtgt gggggggggc ccatacacta gatccatact	360
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gatttctctc ctcatctacc ggtgtcttgt cactgttggg ggggcccac agtcttagtc	180
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gtgcccactg tgagcgacag gttatggtag agtgatacc tagccagggg caatgggact	180
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 <210> SEQ ID NO 71 <211> LENGTH: 305 <212> TYPE: DNA <213> ORGANISM: Sikhote-Alin virus <400> SEQUENCE: 71	
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ccgataacct ttgaccgggt gtgtaggata gaacaggaa cccactacagg cgacaggtta	120
tggtagagtg gatacctagc caggggcaat gggactgcgt tgcatatccc taatgagcca	180
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aggccacgta ggcccgtaa atattataac tattttatta tattttattc acccccaca	300	
ttaatcccag ttaaagcttt ataactataa gtaagccgtg ccgaaacgtt aatcggtcgc	360	
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agattttagc ttgccttaag cagtctttat atcttctgta tattatttta aagtttatag	480	
gagcaaagtt cgctttactc gcaatagcta ttttatttat tttaggaata ttatcacctc	540	
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<211> LENGTH: 742		
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<213> ORGANISM: Cocksackievirus		
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cggagcacac accctcaagc cagagggcag tgtgtcgtaa cgggcaactc tgcagcggaa	540	
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<212> TYPE: DNA		
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agaaaacgtt cgttatccgg ctaactactt cgaaaaacct agtaacacca tgaaagttgc	180	
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tgagagattg ttgcatata gctattggat tggccatccg gtgagcaata gagcgattat	660
ttactctttt gttggatttg tgccattgga tcacaccaca atcatcacac taaagtatat	720
tttactatta aatacagcaa a	741
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acacaccgat caacagtcag cgtggcacac cagccacgtt ttgatcaagc acttctgtta	180
ccccggactg agtatcaata gactgctcac gcggttgaag gagaaagcgt tcgttatccg	240
gccaactact tcgaaaaacc tagtaacacc gtggaagttg cagagtgttt cgtcagcac	300
tacccagtg tagatcaggt cgatgagtc cgcattccc cacgggcgac cgtggcggtg	360
gctgcgttgg cggcctgcc atggggaaac ccatgggacg ctctaataca gacatggtgc	420
gaagagtcta ttgagctagt tggtagtct cggccccctg aatgcggcta atcctaactg	480
cggagcacac accctcaagc cagagggcag tgtgtcgtaa cgggcaactc tgcagcgga	540
ccgactactt tgggtgtccg tgtttcattt tattcctata ctggctgctt atggtgacaa	600
ttgagagatt gttaccatat agctattgga ttggccatcc ggtgagcaat agagcgatta	660
tttactcttt tgttgatttg gtgccattgg atcacaccac aatcatcaca ctaaagtata	720
ttttactatt aaatacagca aa	742
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<212> TYPE: DNA	
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acacaccgat caacagtcag cgtggcacac cagccacgtt ttgatcaagc acttctgtta	180
ccccggactg agtatcaata gactgctcac gcggttgaag gagaaagcgt tcgttatccg	240
gccaactact tcgaaaaacc tagtaacacc gtggaagttg cagagtgttt cgctcagcac	300
taccccgagt tagatcaggt cgatgagtca ccgcattccc cacgggcgac cgtggcggtg	360
gctgcgttgg cggcctgccc atggggaaac ccatgggacg ctctaataca gacatggtgc	420
gaagagtcta ttgagctagt tggtagtcct ccggccccctg aatgcggcta atcctaactg	480
cggagcacac accctcaagc cagagggcag tgtgtcgtaa cgggcaactc tgcagcggaa	540
ccgactactt tgggtgtccg tgtttcattt tattcctata ctggctgctt atggtgacaa	600
ttgagagatt gttaccatat agctattgga ttggccatcc ggtgaataat agagcgataa	660
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acacaccgat caacagtcag cgtggcacac cagccacgtt ttgatcaagc acttctgtta	180
ccccggactg agtatcaata gactgctcac gcggttgaag gagaaagcgt tcgttatccg	240
gccaactact tcgaaaaacc tagtaacacc gtggaagttg cagagtgttt cgctcagcac	300
taccccgagt tagatcaggt cgatgagtca ccgcattccc cacgggcgac cgtggcggtg	360
gctgcgttgg cggcctgccc atggggaaac ccatgggacg ctctaataca gacatggtgc	420
gaagagtcta ttgagctagt tggtagtcct ccggccccctg aatgcggcta atcctaactg	480
cggagcacac accctcaagc cagagggcag tgtgtcgtaa cgggcaactc tgcagcggaa	540
ccgactactt tgggtgtccg tgtttcattt tattcctata ctggctgctt atggtgacaa	600
ttgagagatt gttaccatat agctattgga ttggccatcc ggtgaccaat agagctatta	660
tatatctctt tgttgagct tgcttgttct ttttcagaa gctcagaata aacgctcaac	720
tttgcccgcc acc	733
<210> SEQ ID NO 92	
<211> LENGTH: 1064	
<212> TYPE: DNA	
<213> ORGANISM: Theiler's murine encephalomyocarditis virus	
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atcatcccc gggtaccccc ttctgacgcg ggtactgcga tagtgccacc ccagtctttc	120
ctactccga ctcccgaccc taaccaggt tcctcggaac aggaacacca atttactcat	180
cccctggatg ctgactaatc agaggaaagt cagcattttc cggcccaggc taagagaagt	240

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agataagtta gaatccaaat tgatttatca tccccttgac gaattcgcgt tggaaaaaca	300
cctctcaactt gcccctcttc acaccatta atttaattcg gcctctgtgt tgagcccctt	360
gttgaagtgt ttccctccat cgcgacgtgg ttggagatct aagtcaaccg actccgacga	420
aactaccatc atgcctcccc gattatgtga tgctttctgc cctgctgggt ggagcaccct	480
cgggttgaga aaaccttctt cctttttcct tggactcggg tcccccggtc taagcgcctc	540
ggaatatgac aggggttattt tcacctcttc ttttctactc cacagtgttc tatactgtgg	600
aagggtatgt gttgcccctt ccttcttgga gaacgtgcgc ggcggtcttt ccgtctctcg	660
acaagcgcgc gtgcaacata cagagtaacg cgaagaaagc agttctcggc ctagctctag	720
tgccacaag aaaacagctg tagcgaccac acaaaggcag cggaaccccc ctcttggtaa	780
caggagcctc tgcggccaaa agccacgtgg ataagatcca cctttgtgtg cggtgcaacc	840
ccagcaccct ggtttcttgg tgacactcta gtgaaccctt gaatggcaat ctcaagcgcc	900
tctgtaggga agccaagaat gtccaggagg tacccttccc tctcggaagg gatctgacct	960
ggagacacat cacacgtgct ttacacctgt gcttgtgttt aaaaattgtt acagcttccc	1020
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<210> SEQ ID NO 93	
<211> LENGTH: 652	
<212> TYPE: DNA	
<213> ORGANISM: Saffold virus	
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atcctcgggt tgagaaaacc ttcttccttt ttcttggaac cccggtcccc cgggtctaagc	120
cgtttggaat aagacagggt tatcttcacc tcttccttct tctacttcat agtgttctat	180
actatgaaag ggtatgtgtc gccccttcct tctttggaga acacgcgcgg cggctcttcc	240
gtctctcgaa aagcgcgtgt gcgacatgca gagaaccgtg aagaaagcag tttgcggact	300
agcttttagtg cccacaagaa aacagctgta gcgaccacac aaaggcagcg gacccccct	360
cctggcaaca ggagcctctg cggccaaaag ccacgtggat aagatccacc tttgtgtgcg	420
gcacaacccc agtgccttgg tttcttgggt acacttcagt gaaaacgcaa atggcgatct	480
gaagcgctc tgtaggaaag ccaagaatgt ccaggaggta ccccttcctt cgggaaggga	540
tctgacctgg agacacatca catgtgcttt acacctgtgc ttgtgtttaa aaattgtcac	600
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<210> SEQ ID NO 94	
<211> LENGTH: 741	
<212> TYPE: DNA	
<213> ORGANISM: Echovirus	
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cataccgac c aatagtggc gtggcacacc agccgtgtct agatcaagca ctctgtttc	180
cccggaccga gtatcaatag actgctcacg cggttgaagg agaaaacgtt cgttatccgg	240
ctaactactt cgaaaaacct agtaacacca tgaaagttgc ggagtgtttc actcagcact	300
tcccagtgat agatcaggtc gatgagtcac cgcattcctc acgggcgacc gtggcggtgg	360

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ctgcgctggc ggctgccta tggggtgacc cataggacgc tctaatacag acatgggtgcg	420
aagagtctat tgagctagtt agtagtcctc cggccctga atgcggataa tcctaactgt	480
ggagcagata cccacgaacc agtgggcagt ctgtcgtaac gggcaactcc gcagcggaac	540
cgactacttt ggggtgtccgt gtttcctttt attccaaatc tggctgctta tggtgacaat	600
tgagagattg ttgccatata gctattggat tggccatccg gtgaataata gagcgataat	660
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cataccgata aatagtgggc gtggcacacc agccgtgtct agatcaagca cttctgtttc	180
cccgggaccg agtatcaata gactgctcac gcggttgaag gagaaagcgt tcgttaccg	240
gccatctact tcgagaagcc tagtaacacc atgaaagttg cagagcgttt cgctcagcac	300
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gctgcgttgg cggcctgcc atgggattac ccatgggacg ctctaataca gacatggtgt	420
gaagaacctt ttgagctagt tggtagtct cggccctcg aatgcggcta atcctaactg	480
cggagcatat accctcaatc cagggggcag tgtgtcgtaa cgggcaactc tgcagcgga	540
ccgactactt tgggtgtccg tgtttccttt tattcttatt atggctgctt atggtgacaa	600
ttgagagatt gttgccatat agctattgga ttggccatcc ggtgagcaat agagcgatta	660
tttactcttt tgttgattt gtgccattgg atcacaccac aatcatcaca ctaaagtata	720
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tcaatagact gctcacgcgg ttgaaggaga aaacgttcgt tatccggcta actacttcga	180
aaaacctagt aacaccatga aagttgcgga gtgtttcact cagcacttcc ccagtgtaga	240
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ctgcctatgg ggtgacccat aggacgctct aatacagaca tggtgcaag agtctattga	360
gctagttagt agtcctccg cccctgaatg cggataatcc taactgtgga gcagataccc	420
acgaaccagt gggcagtctg tcgtaacggg caactccgca gcggaaccga ctactttggg	480
tgtccgtgtt tccttttatt ccaaactctg ctgcttatgg tgacaattga gagattgttg	540
ccatatagct attggattgg ccatccggtg aataatagag cgataatata tttgtttgtt	600

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ggattcgtgc cacttagtct gaaagttttg agaacactca actacgtttt attgctgaat	660
agtgcaag	668
<div><210> SEQ ID NO 97</div> <div><211> LENGTH: 669</div> <div><212> TYPE: DNA</div> <div><213> ORGANISM: Artificial sequence</div> <div><220> FEATURE:</div> <div><223> OTHER INFORMATION: Viral</div> <div><400> SEQUENCE: 97</div>	
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caggcgcgac ggcaccaagt cgtgtctcgg tcaagcactt ctgtttcccc gggaccgagt	120
atcaatagac tgctcacgcy gttgaaggag aaagcgttcg ttacccgcc atctacttcg	180
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cctgcccata ggattacca tgggacgctc taatacagac atgggtgtgaa gaacctattg	360
agctagttag tagtcctccg gcccctgaat gcggctaata ctaactgcgg agcatacacc	420
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gcatatagc tattggattg gccatccggt gaataataga gcgataatat atttgtttgt	600
tggattcgtg ccacttagtc tgaaagtttt gagaacactc aactacgttt tattgctgaa	660
tagtgcaag	669
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tcaccccggg aaacccctt tggacgcggg acctgcgaca gtgccatccc ccgtctctcc	120
tattccaact acccgaccct aaccaggggt ccaggacact ggatcaatac aagtcatccc	180
ctgaatgctg gctaatacaga ggaaagtcag cattttcctg cccaggctaa gagaaacaca	240
ataagttaga atctaaatta atcaccttga cgaattcgca aagataagtc ctccctccct	300
tgccgctcga tcacaccag aactaacaat tcggcctctc gtgacgagcc ccttggtgaa	360
aggacctctt tcaacgcgac gtggttgag attaaaaccg actccgacga aagtgcctatc	420
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gtggatgcag catgctggaa acggtgaaga aagtagttct ctgtggaaac ttagaacaga	720
catcgaaaca gctgtagcga cctcacagta gcagcggaac cccctcctgg cgacaggagc	780
ctctgcggcc aaaagccccg tggataagat cactgctgt gagcggtgca acccagcac	840
cctggttcga tggttgttct ctgtggaatc agagaatggc tttcctaagc cctccagtag	900
agaagccaag aatgtcctga aggtaccccg cgtgcgggat ctgatcagaa gaccaattgc	960

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cagtgcata cactggtact ttggtttaaa aattgtcaca gcttctccaa accaagtgg	1020
cttggttttc tatctttaat aattggttca tgatg	1055
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ggtgatgtcg tatagactgc ttgcgtggtt gaaagcgacg gatccgttat ccgcttatgt	240
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gattgttatc ataaagcgaa ttggattggc catccggtga aagtgagact cattatctat	660
ctgtttgctg gatccgctcc attgagtgtg tttactctaa gtacaatttc aacagttatt	720
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atcaatagac tgctcacgcg gttgaaggag aaagcgttcg ttacccggcc atctacttcg	180
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gccatatagc tattggattg gccatccggt gagcaataga gcgattattt actcttttgt	600
tggatttgtg ccattggatc acaccacaat catcacacta aagtatatat tactattaaa	660
tacagcaaa	669
<div><210> SEQ ID NO 101</div> <div><211> LENGTH: 20</div> <div><212> TYPE: PRT</div> <div><213> ORGANISM: Teschovirus</div> <div><400> SEQUENCE: 101</div>	
Arg Ala Glu Gly Arg Gly Ser Leu Leu Thr Cys Gly Asp Val Glu Glu	

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1	5	10	15
Asn Pro Gly Pro			
20			
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<211> LENGTH: 236			
<212> TYPE: PRT			
<213> ORGANISM: Artificial sequence			
<220> FEATURE:			
<223> OTHER INFORMATION: Mammalian			
<400> SEQUENCE: 102			
Met Thr Ala Leu Thr Glu Gly Ala Lys Leu Phe Glu Lys Glu Ile Pro			
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20 25 30			
Lys Gly Glu Gly Thr Gly Asp Ala Thr Thr Gly Thr Ile Lys Ala Lys			
35 40 45			
Tyr Ile Cys Thr Thr Gly Asp Leu Pro Val Pro Trp Ala Thr Leu Val			
50 55 60			
Ser Thr Leu Ser Tyr Gly Val Gln Cys Phe Ala Lys Tyr Pro Ser His			
65 70 75 80			
Ile Lys Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Thr Gln Glu			
85 90 95			
Arg Thr Ile Ser Phe Glu Gly Asp Gly Val Tyr Lys Thr Arg Ala Met			
100 105 110			
Val Thr Tyr Glu Arg Gly Ser Ile Tyr Asn Arg Val Thr Leu Thr Gly			
115 120 125			
Glu Asn Phe Lys Lys Asp Gly His Ile Leu Arg Lys Asn Val Ala Phe			
130 135 140			
Gln Cys Pro Pro Ser Ile Leu Tyr Ile Leu Pro Asp Thr Val Asn Asn			
145 150 155 160			
Gly Ile Arg Val Glu Phe Asn Gln Ala Tyr Asp Ile Glu Gly Val Thr			
165 170 175			
Glu Lys Leu Val Thr Lys Cys Ser Gln Met Asn Arg Pro Leu Ala Gly			
180 185 190			
Ser Ala Ala Val His Ile Pro Arg Tyr His His Ile Thr Tyr His Thr			
195 200 205			
Lys Leu Ser Lys Asp Arg Asp Glu Arg Arg Asp His Met Cys Leu Val			
210 215 220			
Glu Val Val Lys Ala Val Asp Leu Asp Thr Tyr Gln			
225 230 235			
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<211> LENGTH: 237			
<212> TYPE: PRT			
<213> ORGANISM: Artificial sequence			
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<223> OTHER INFORMATION: Mammalian			
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1 5 10 15			
Leu Tyr Met Glu Gly Thr Val Asn Asn His His Phe Lys Cys Thr Ser			
20 25 30			
Glu Gly Glu Gly Lys Pro Tyr Glu Gly Thr Gln Thr Met Arg Ile Lys			
35 40 45			

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Val	Val	Glu	Gly	Gly	Pro	Leu	Pro	Phe	Ala	Phe	Asp	Ile	Leu	Ala	Thr
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Ser	Phe	Met	Tyr	Gly	Ser	Arg	Thr	Phe	Ile	Lys	Tyr	Pro	Lys	Gly	Ile
65					70					75					80
Pro	Asp	Phe	Phe	Lys	Gln	Ser	Phe	Pro	Glu	Gly	Phe	Thr	Trp	Glu	Arg
				85					90					95	
Val	Thr	Thr	Tyr	Glu	Asp	Gly	Gly	Val	Val	Thr	Val	Met	Gln	Asp	Thr
			100					105					110		
Ser	Leu	Glu	Asp	Gly	Cys	Leu	Val	Tyr	Asn	Val	Lys	Ile	Arg	Gly	Val
		115					120					125			
Asn	Phe	Pro	Ser	Asn	Gly	Pro	Val	Met	Gln	Lys	Lys	Thr	Leu	Gly	Trp
	130					135						140			
Glu	Ala	Asn	Thr	Glu	Met	Leu	Tyr	Pro	Ala	Asp	Gly	Gly	Leu	Glu	Gly
145					150					155					160
Arg	Ser	Asp	Met	Ala	Leu	Lys	Leu	Val	Gly	Gly	Gly	His	Leu	Ser	Cys
				165					170					175	
Ser	Phe	Val	Thr	Thr	Tyr	Arg	Ser	Lys	Lys	Pro	Ala	Lys	Asn	Leu	Lys
		180						185					190		
Met	Pro	Gly	Ile	His	Ala	Val	Asp	His	Arg	Leu	Glu	Arg	Leu	Glu	Glu
		195					200					205			
Ser	Asp	Asn	Glu	Met	Phe	Val	Val	Gln	Arg	Glu	His	Ala	Val	Ala	Arg
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Tyr	Cys	Asp	Leu	Pro	Ser	Lys	Leu	Gly	His	Lys	Leu	Asn			
225					230					235					

<210> SEQ ID NO 104
<211> LENGTH: 721
<212> TYPE: DNA
<213> ORGANISM: Hepatitis B virus

<400> SEQUENCE: 104

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caggctttta ctttctcgcc aacttacaag gcctttctca gtaaacagta tatgaccctt	180
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catctgccgg accgtgtgca ctctgcttca cctctgcacg tcgcatggag gccaccgtga	660
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gtcatc	726
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atgctttgct ctaaatcttg atatttagtg	gaacattctt tccatttttg ttctacaaga	240
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aagctt	246
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Val Gln Cys	
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Val Leu Ser	
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gagaatagca tgcgtgag	258
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- What is claimed is:
1. A polynucleotide encoding a piggyBac-like transposase operably linked to a heterologous promoter, wherein the transposase comprises an amino acid sequence that is at least 90% identical to SEQ ID NO: 44, and wherein the transposase can integrate a transposon comprising a heterologous DNA sequence flanked by a pair of transposon ends into the genomic DNA of a target cell, wherein one transposon end comprises 16 contiguous nucleotides from SEQ ID NO: 1 and the other transposon end comprises 16 contiguous nucleotides from SEQ ID NO: 2.

2. The polynucleotide of claim 1, wherein the transposase comprises the amino acid sequence of SEQ ID NO: 44.

3. The polynucleotide of claim 1, wherein the nucleic acid further encodes a heterologous nuclear localization signal (NLS) expressible fused to the transposase.

4. An mRNA transcribed from the polynucleotide of claim 3 encoding the transposase fused to the heterologous NLS.

5. The polynucleotide of claim 1, wherein the nucleic acid further encodes a DNA binding domain (DBD) expressible as a fusion protein with the transposase.

6. The polynucleotide of claim 1, wherein either (a) the polynucleotide further comprises the transposon or (b) the polynucleotide is a component of a kit further comprising the transposon.

7. A transposase linked to a heterologous NLS expressed from polynucleotide of claim 3.

8. A pharmaceutical composition comprising a transposase comprising an amino acid sequence that is at least 90% identical to SEQ ID NO: 44 and a transposon comprising a heterologous polynucleotide flanked by a pair of transposon ends, wherein one transposon end comprises 16 contiguous nucleotides from SEQ ID NO: 1 and the other transposon end comprises 16 contiguous nucleotides from SEQ ID NO: 2 together with a pharmaceutically acceptable carrier, adjuvant or vehicle.
- * * * * *

UNITED STATES PATENT AND TRADEMARK OFFICE
CERTIFICATE OF CORRECTION

PATENT NO. : 9,580,697 B2
APPLICATION NO. : 15/222830
DATED : February 28, 2017
INVENTOR(S) : Jeremy Minshull et al.

Page 1 of 1

It is certified that error appears in the above-identified patent and that said Letters Patent is hereby corrected as shown below:

In the Claims

In Claim 8, at Column 218, Line 37:

Delete the text:

“SEQ ID NO; 1”

Replace it with the text:

--SEQ ID NO: 1--.

Signed and Sealed this
Twenty-third Day of May, 2017

A handwritten signature in black ink, reading "Michelle K. Lee". The signature is fluid and cursive, with the first letters of each name being capitalized and prominent.

Michelle K. Lee
Director of the United States Patent and Trademark Office



US009580697C1

(12) **EX PARTE REEXAMINATION CERTIFICATE** (11721st)
United States Patent
Minshull et al.

(10) **Number:** **US 9,580,697 C1**(45) **Certificate Issued:** **Sep. 4, 2020**(54) **ENHANCED NUCLEIC ACID CONSTRUCTS
FOR EUKARYOTIC GENE EXPRESSION**(71) Applicant: **DNA2.0, INC.**, Newark, CA (US)(72) Inventors: **Jeremy Minshull**, Los Altos, CA (US);
Mark Welch, Fremont, CA (US);
Sridhar Govindrajan, Los Altos, CA
(US); **Kate Caves**, San Jose, CA (US)(73) Assignee: **DNA2.0, INC.**, Menlo Park, CA (US)**Reexamination Request:**

No. 90/014,319, Jun. 19, 2019

Reexamination Certificate for:Patent No.: **9,580,697**Issued: **Feb. 28, 2017**Appl. No.: **15/222,830**Filed: **Jul. 28, 2016**

Certificate of Correction issued May 23, 2017

Related U.S. Application Data(63) Continuation of application No. 14/683,121, filed on
Apr. 9, 2015, now Pat. No. 9,428,767.(60) Provisional application No. 61/977,474, filed on Apr.
9, 2014, provisional application No. 62/003,397, filed
on May 27, 2014, provisional application No.
62/046,875, filed on Sep. 5, 2014, provisional
application No. 62/046,705, filed on Sep. 5, 2014,
provisional application No. 62/069,656, filed on Oct.
28, 2014, provisional application No. 62/120,522,
filed on Feb. 25, 2015.(51) **Int. Cl.****C07K 16/00** (2006.01)**C12N 15/85** (2006.01)**A61K 31/7105** (2006.01)**A61K 38/00** (2006.01)**C12N 9/12** (2006.01)**C07K 2/00** (2006.01)**A61K 31/00** (2006.01)**C07K 14/435** (2006.01)**C07K 14/46** (2006.01)**C07K 16/32** (2006.01)**C12N 15/90** (2006.01)(52) **U.S. Cl.**CPC **C12N 9/1241** (2013.01); **A61K 31/00**
(2013.01); **A61K 31/7105** (2013.01); **C07K**
2/00 (2013.01); **C07K 14/43586** (2013.01);
C07K 14/463 (2013.01); **C07K 16/00**
(2013.01); **C07K 16/32** (2013.01); **C12N 15/85**
(2013.01); **C12N 15/907** (2013.01); **C12Y**
207/07 (2013.01); **A61K 38/00** (2013.01);
C07K 2317/10 (2013.01); **C07K 2317/24**
(2013.01); **C07K 2317/56** (2013.01); **C07K**
2319/09 (2013.01); **C07K 2319/80** (2013.01);
C12N 2800/90 (2013.01); **C12N 2830/40**
(2013.01); **C12N 2830/60** (2013.01)(58) **Field of Classification Search**

None

See application file for complete search history.

(56)

References CitedTo view the complete listing of prior art documents cited
during the proceeding for Reexamination Control Number
90/014,319, please refer to the USPTO's public Patent
Application Information Retrieval (PAIR) system under the
Display References tab.*Primary Examiner* — Sharon Turner

(57)

ABSTRACTThe present invention provides polynucleotide vectors for
high expression of heterologous genes, and methods for
constructing such vectors. Some vectors further comprise
novel transposons and transposases that further improve
expression. Further disclosed are vectors that can be used in
a gene transfer system for stably introducing nucleic acids
into the DNA of a cell. The gene transfer systems can be
used in methods, for example, but not limited to, gene
expression, gene therapy, insertional mutagenesis, or gene
discovery.

1
EX PARTE
REEXAMINATION CERTIFICATE

2

THE PATENT IS HEREBY AMENDED AS
INDICATED BELOW. 5

AS A RESULT OF REEXAMINATION, IT HAS BEEN
DETERMINED THAT:

Claims **1-2** and **6** are cancelled. 10
Claims **3-5** and **7-8** were not reexamined.

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