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(54) **MAPPING CYTOSINE MODIFICATIONS**

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C12Q 1/6827 (2013.01); *C07K 2319/80*
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(58) **Field of Classification Search**

None

See application file for complete search history.

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(22) Filed: **Mar. 14, 2013**

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(65) **Prior Publication Data**

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on Nov. 7, 2012, provisional application No.
61/722,968, filed on Nov. 6, 2012, provisional
application No. 61/611,295, filed on Mar. 15, 2012.

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C07H 21/00 (2006.01)

C12P 19/18 (2006.01)

C12P 19/30 (2006.01)

C12P 19/34 (2006.01)

C12N 9/10 (2006.01)

C12N 9/22 (2006.01)

C07K 14/47 (2006.01)

(52) **U.S. Cl.**

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(2013.01); *C07H 21/00* (2013.01); *C07K 14/47*
(2013.01); *C12N 9/1051* (2013.01); *C12N 9/22*
(2013.01); *C12P 19/18* (2013.01); *C12P 19/30*
(2013.01); *C12P 19/34* (2013.01); *C12Q 1/68*
(2013.01); *C12Q 1/683* (2013.01); *C12Q*

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(57) **ABSTRACT**

Methods, compositions and kits for selectively altering and
detecting modified cytosine residues are provided.

6 Claims, 8 Drawing Sheets

FIG. 1

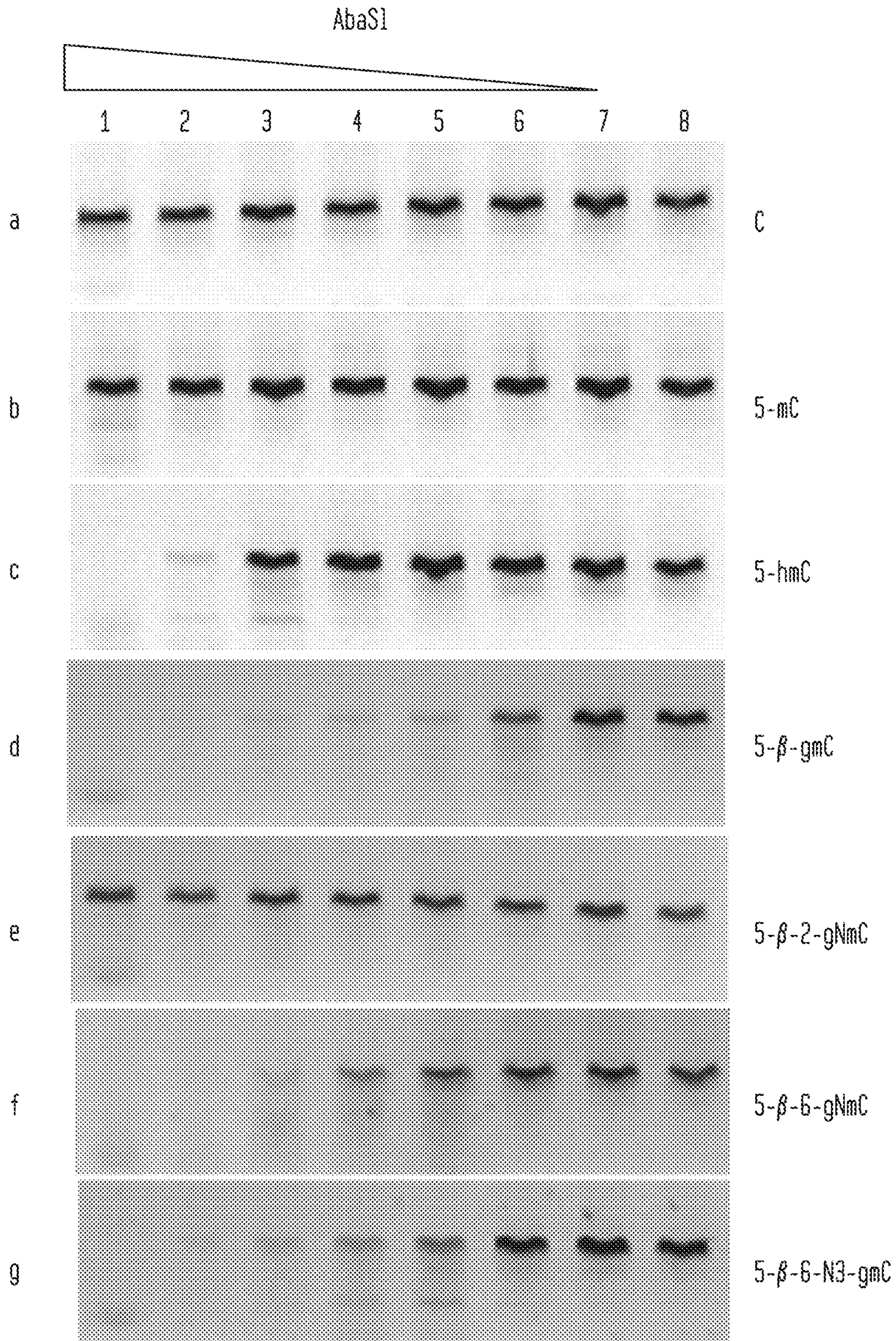
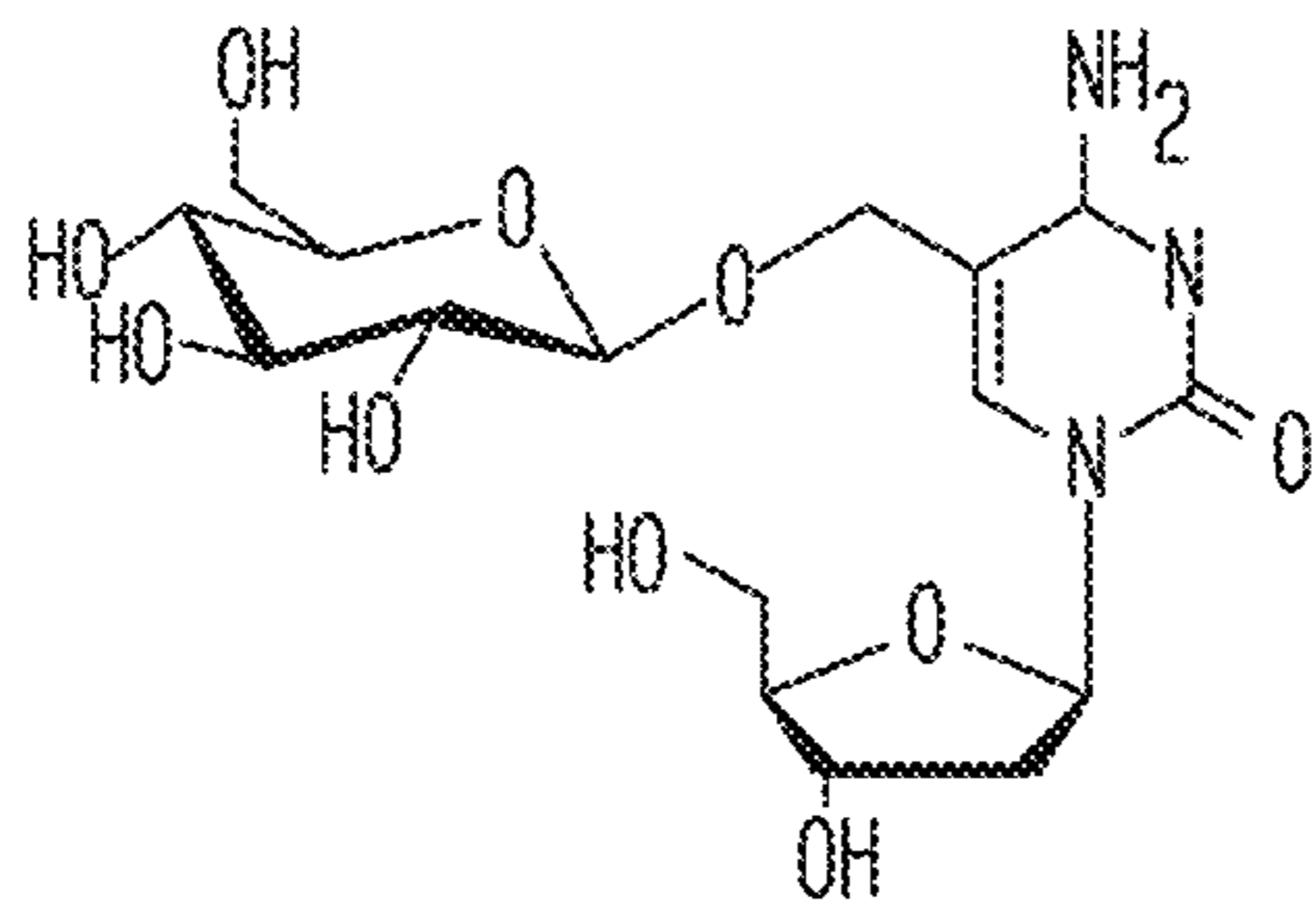
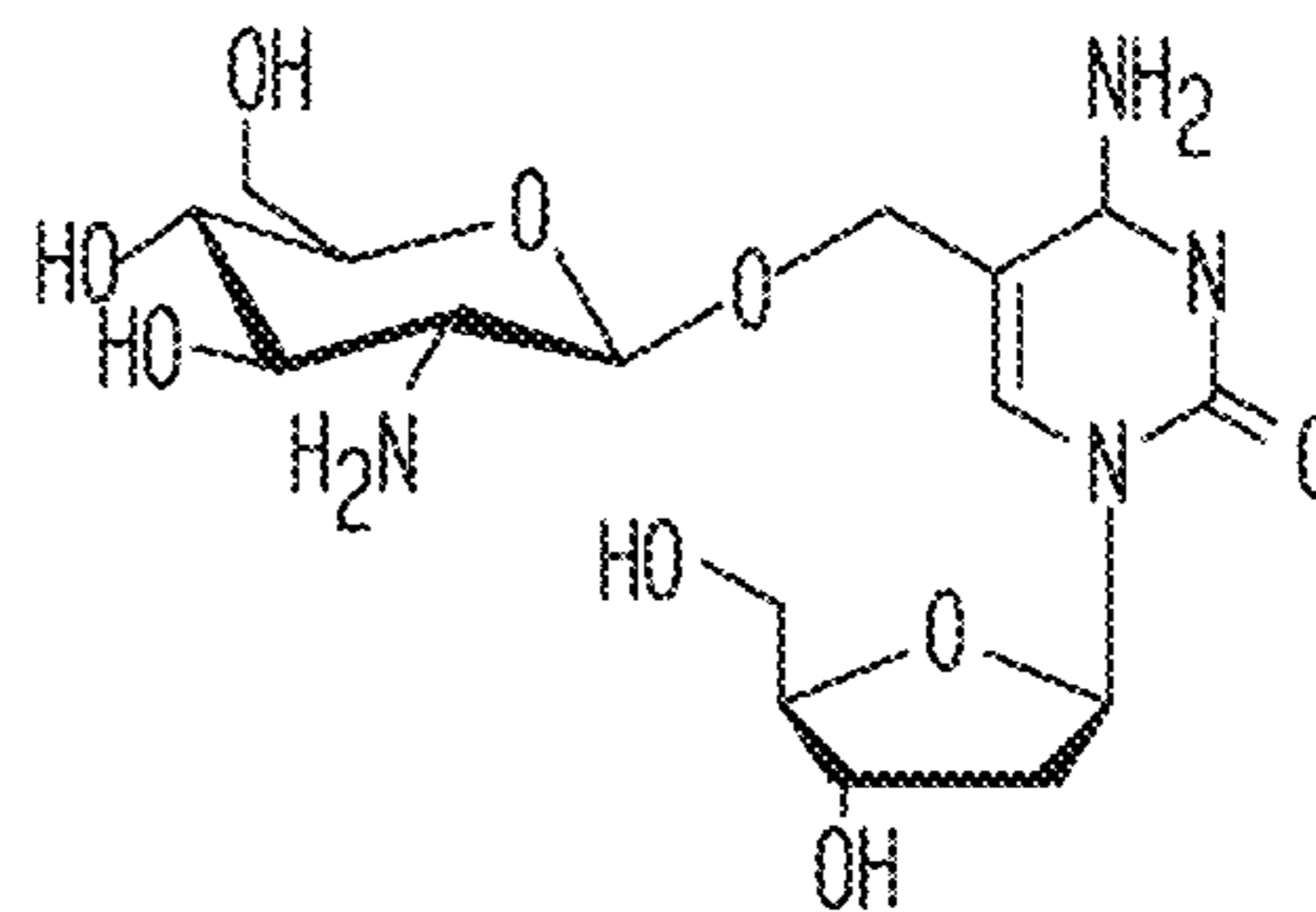


FIG. 3

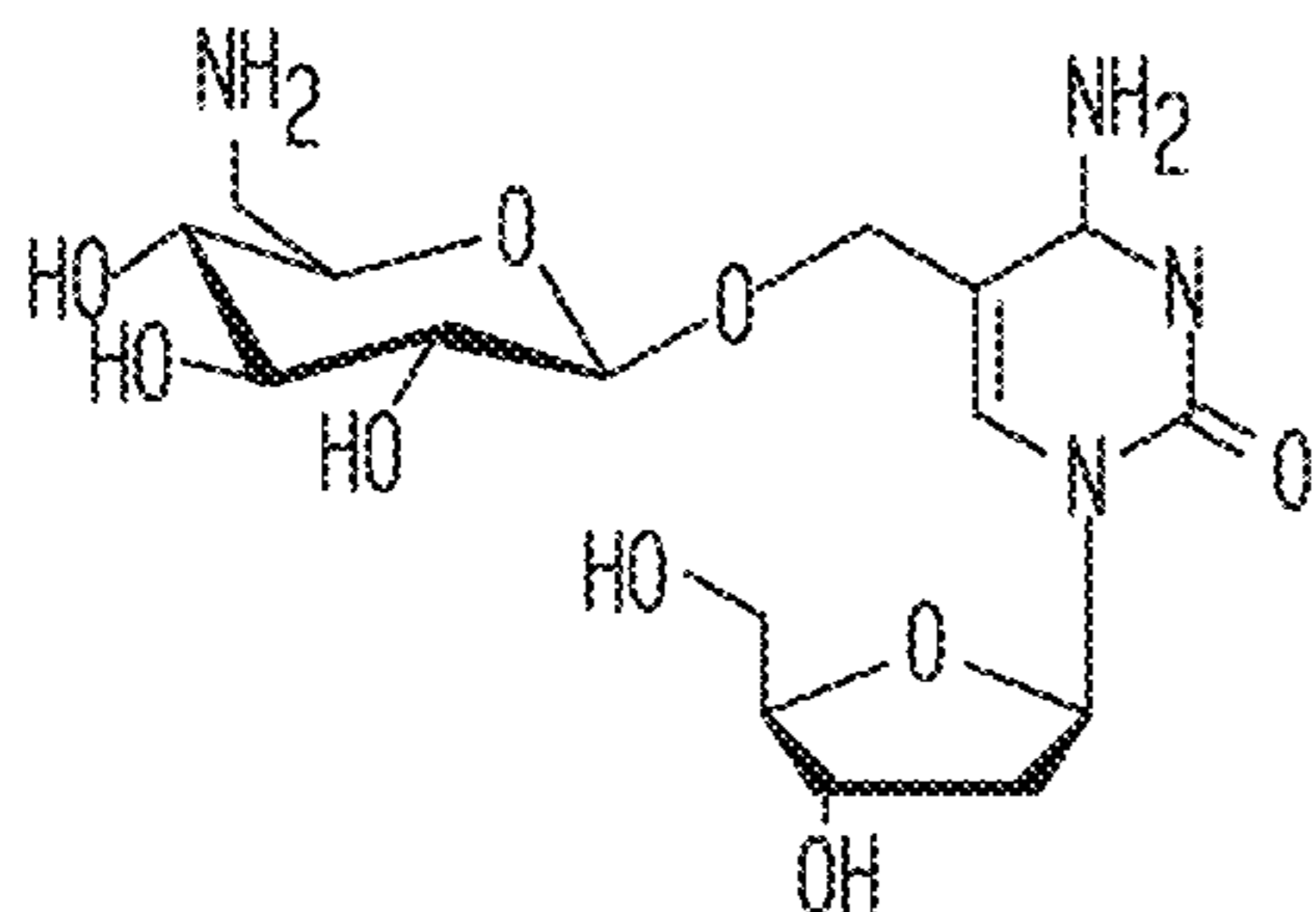
T4-BGT



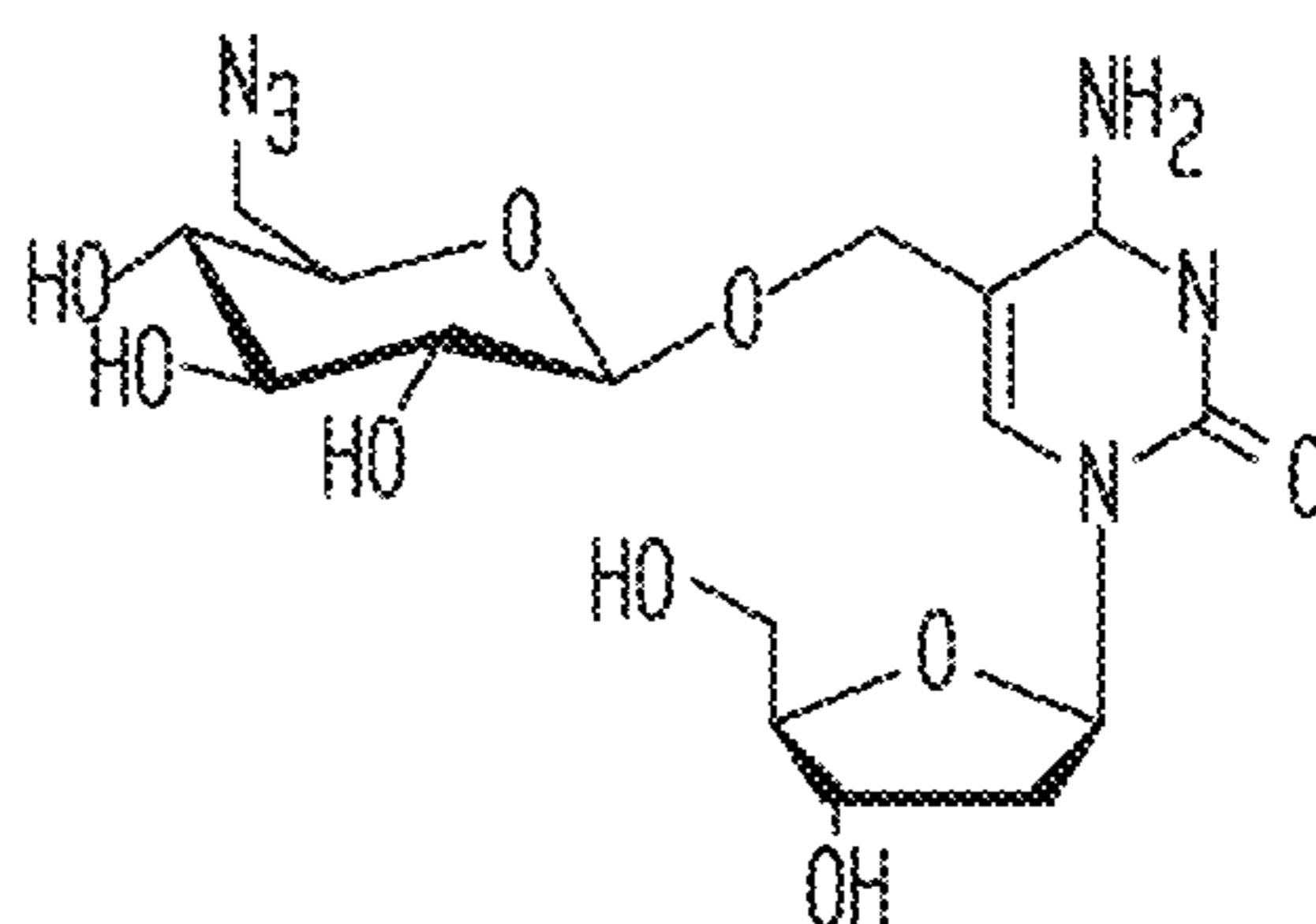
5-β-gmC



5-β-2-gNmC

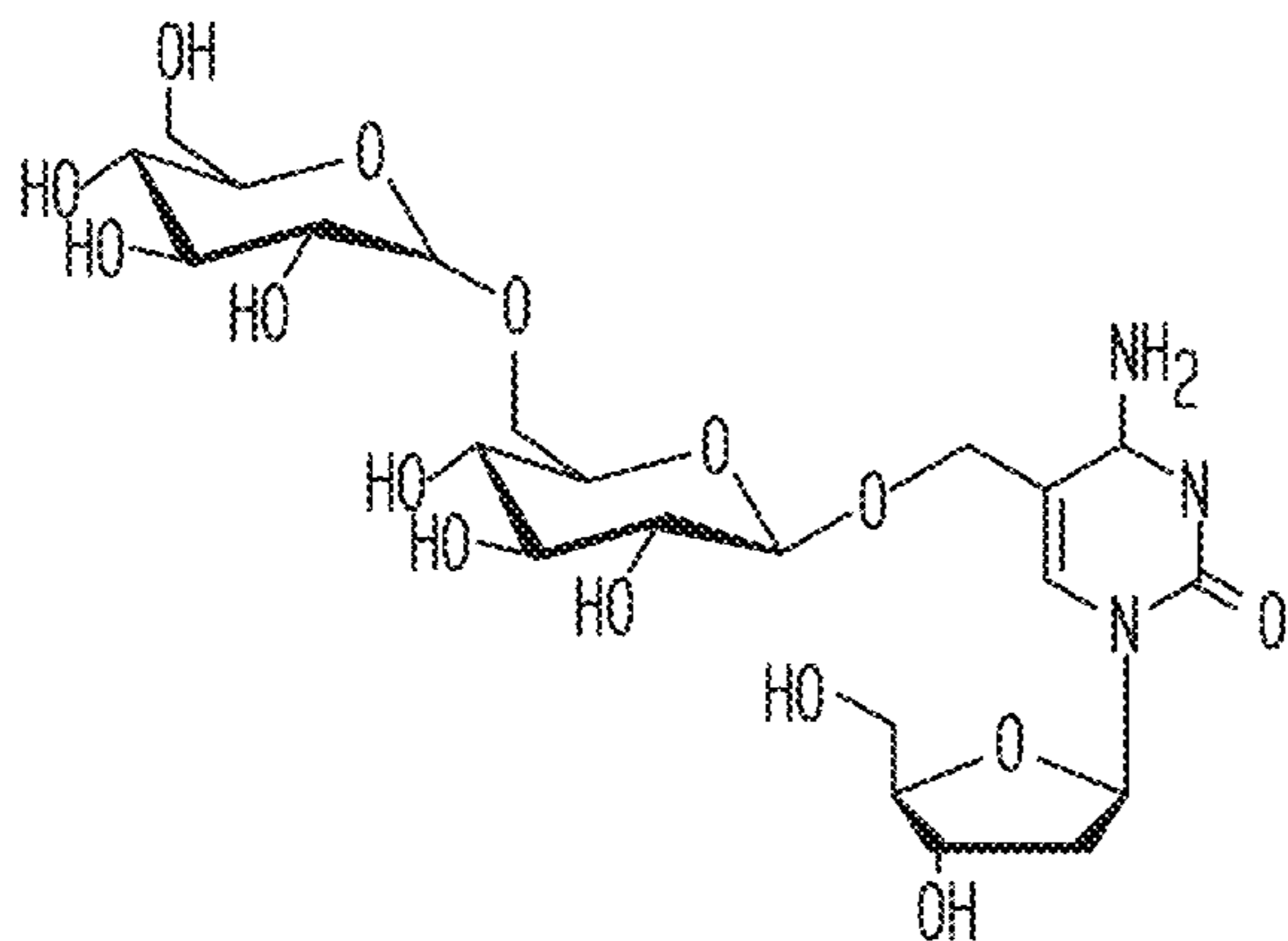


5-β-6-gNmC

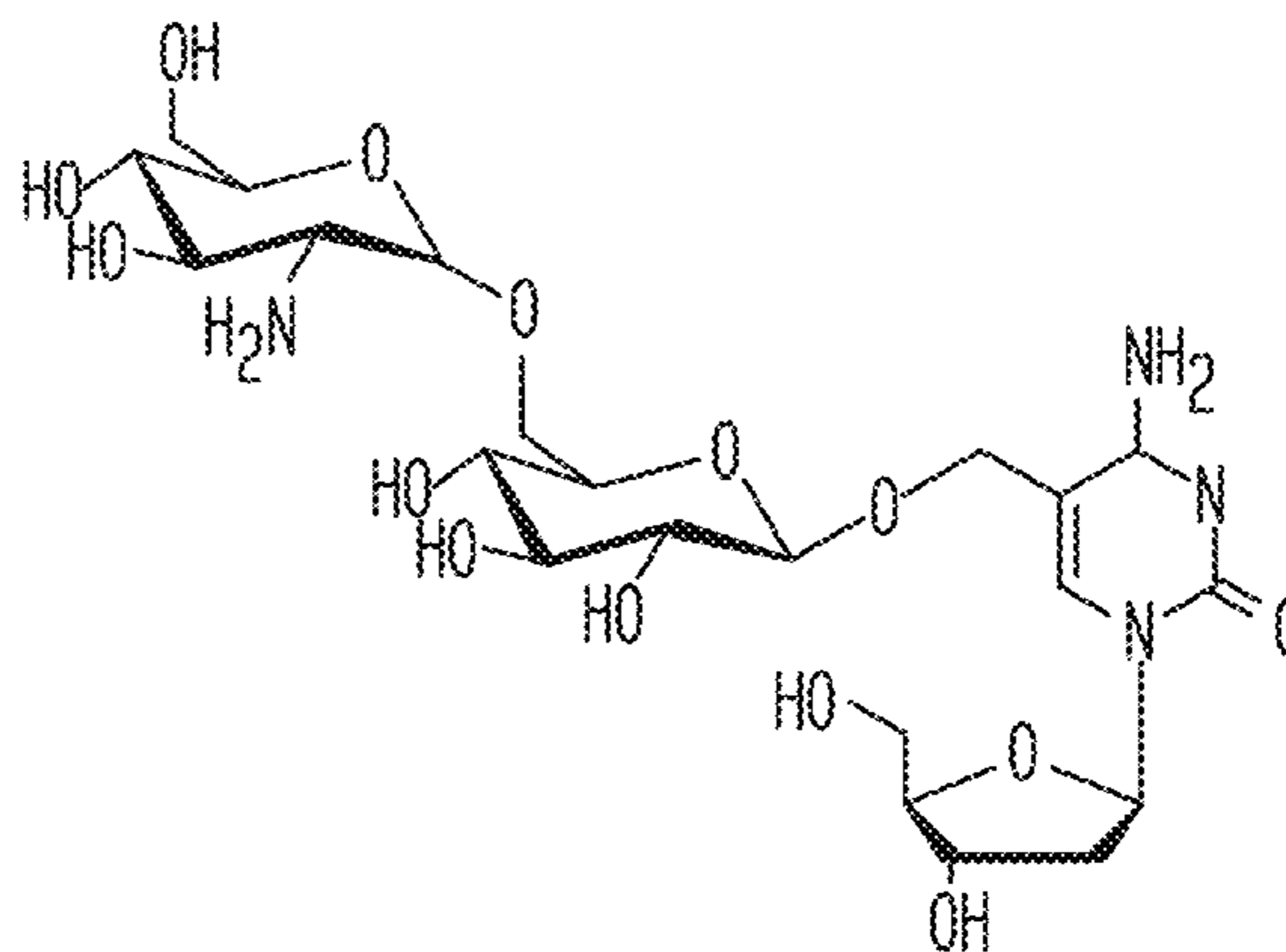


5-β-6-N₃-gmC

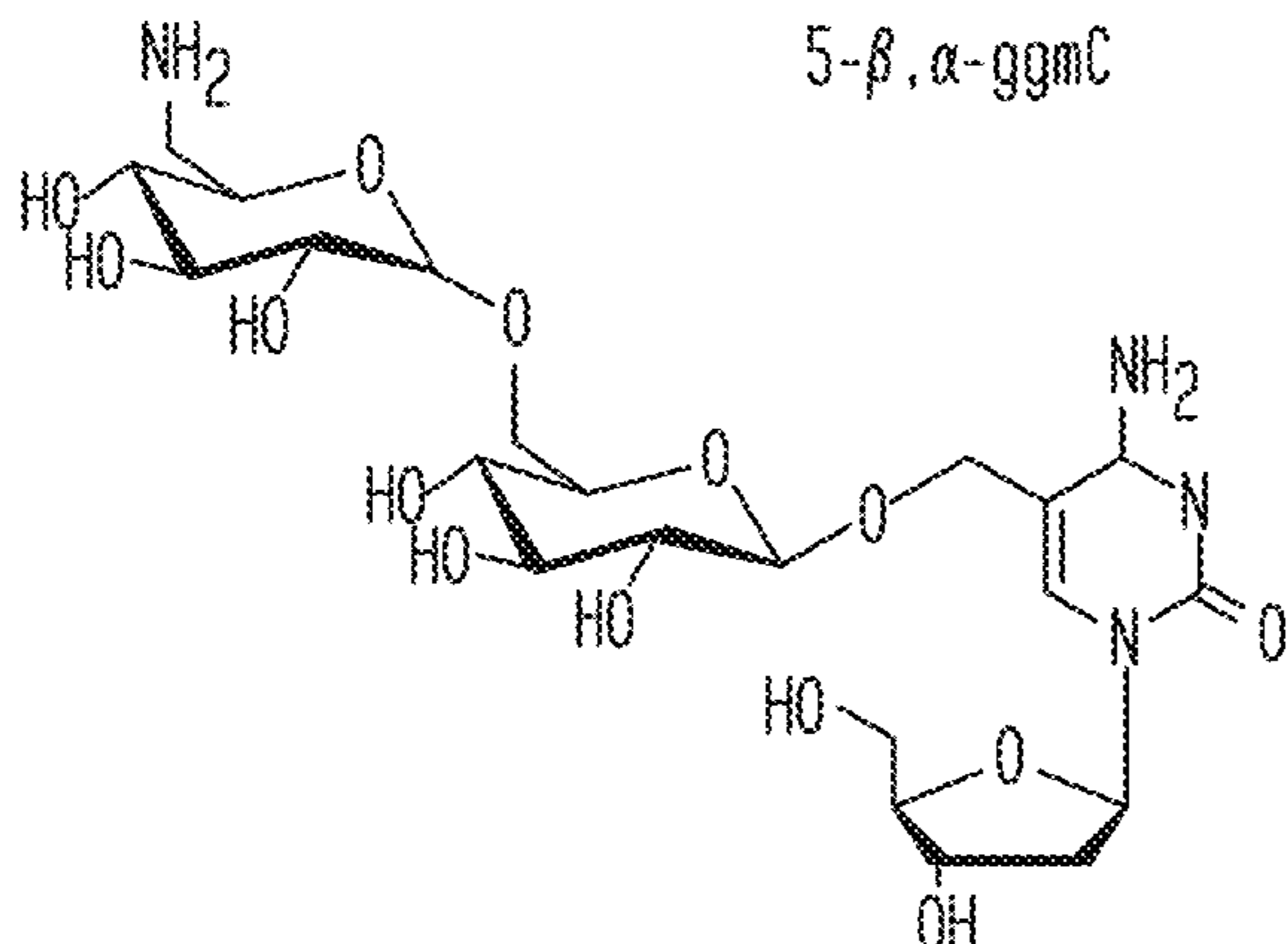
T4-BGT FOLLOWED BY T6-BGAGT



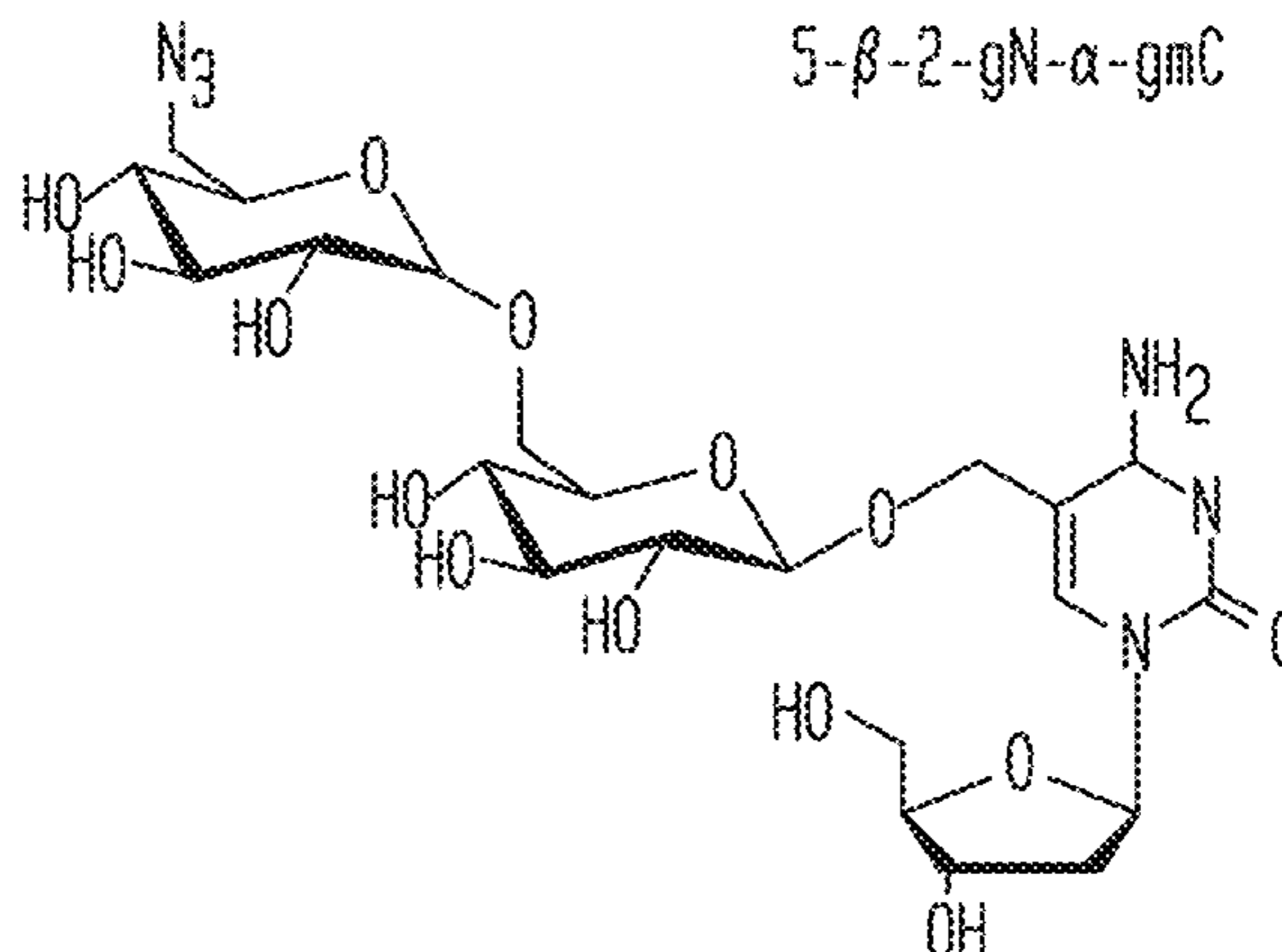
5-β,α-ggmC



5-β-2-gN-α-gmC



5-β-6-gN-α-gmC



5-β-6-N₃-α-gmC

FIG. 4

EXCLUSIVE ^mC MAPPING

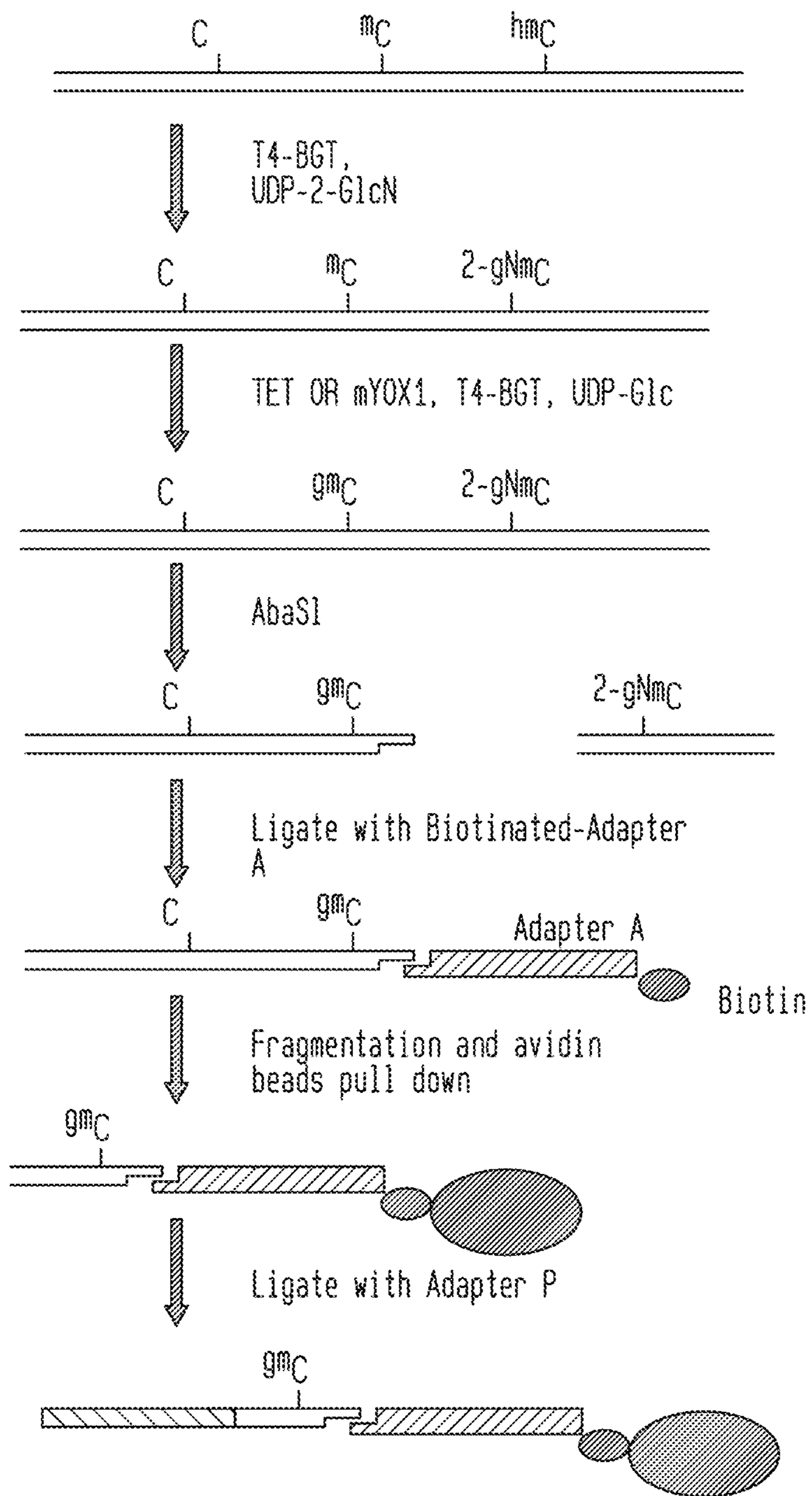


FIG. 5A
EXCLUSIVE fC MAPPING

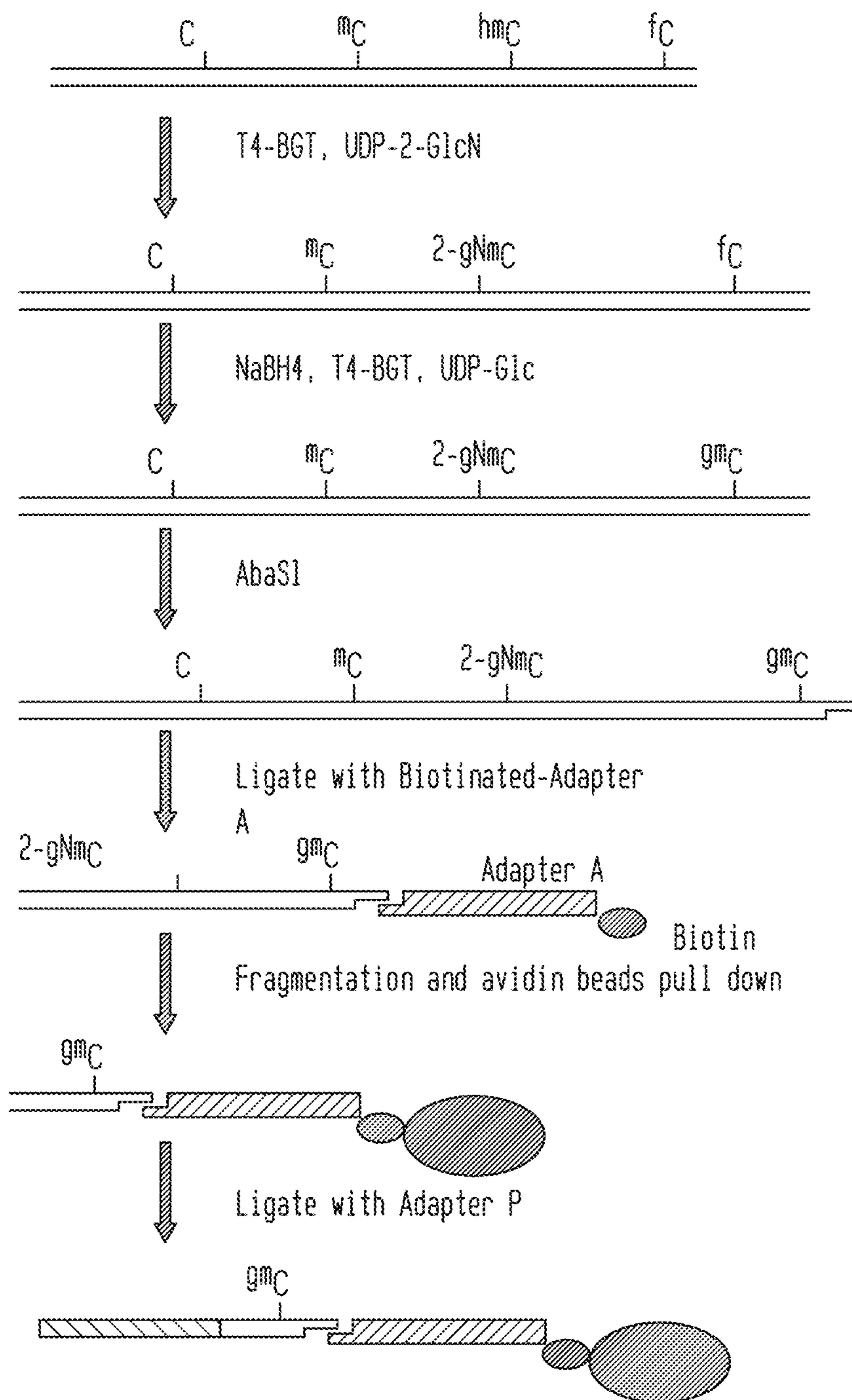


FIG. 5B

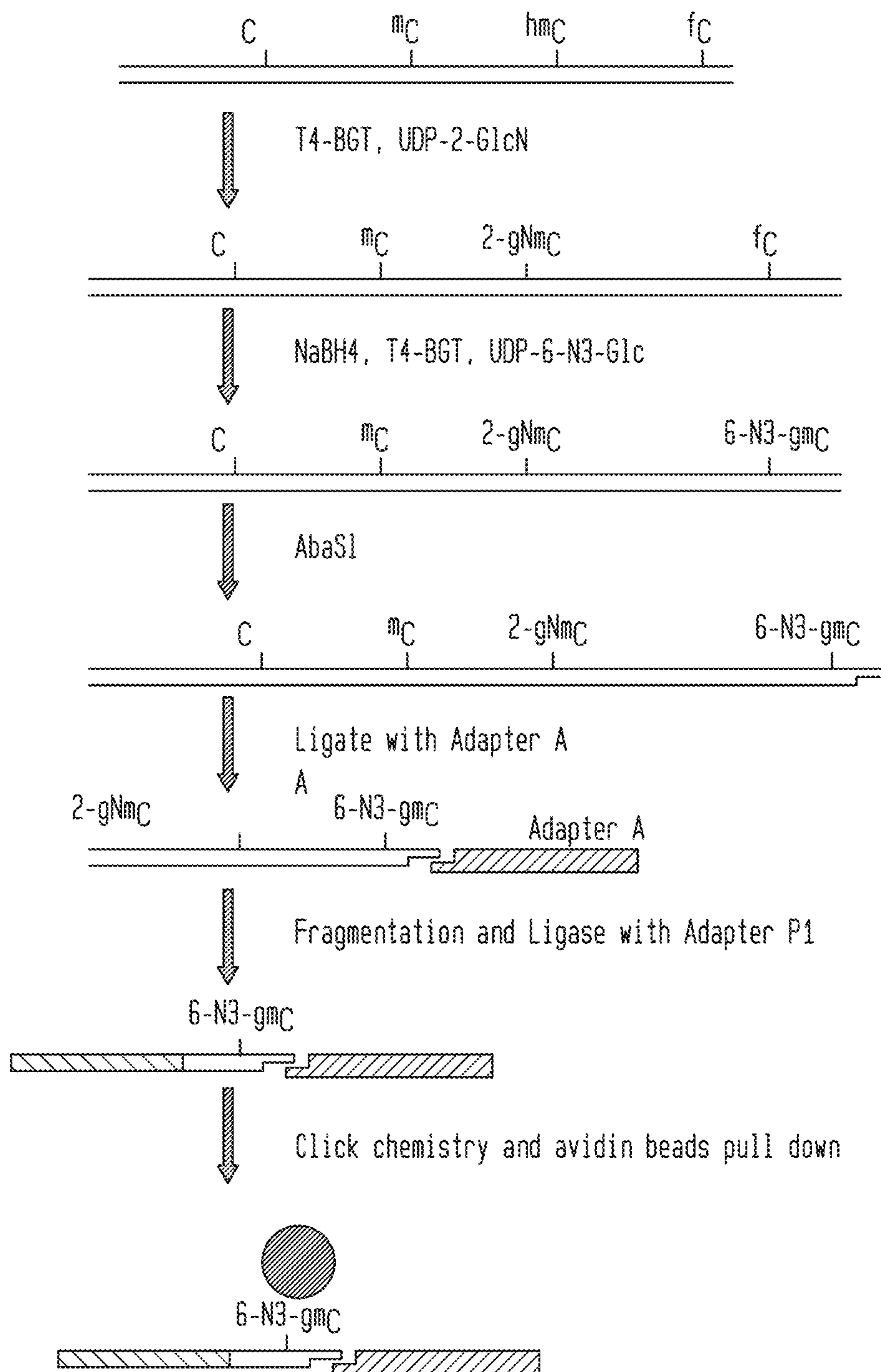


FIG. 6

EXCLUSIVE ^{ca}C MAPPING

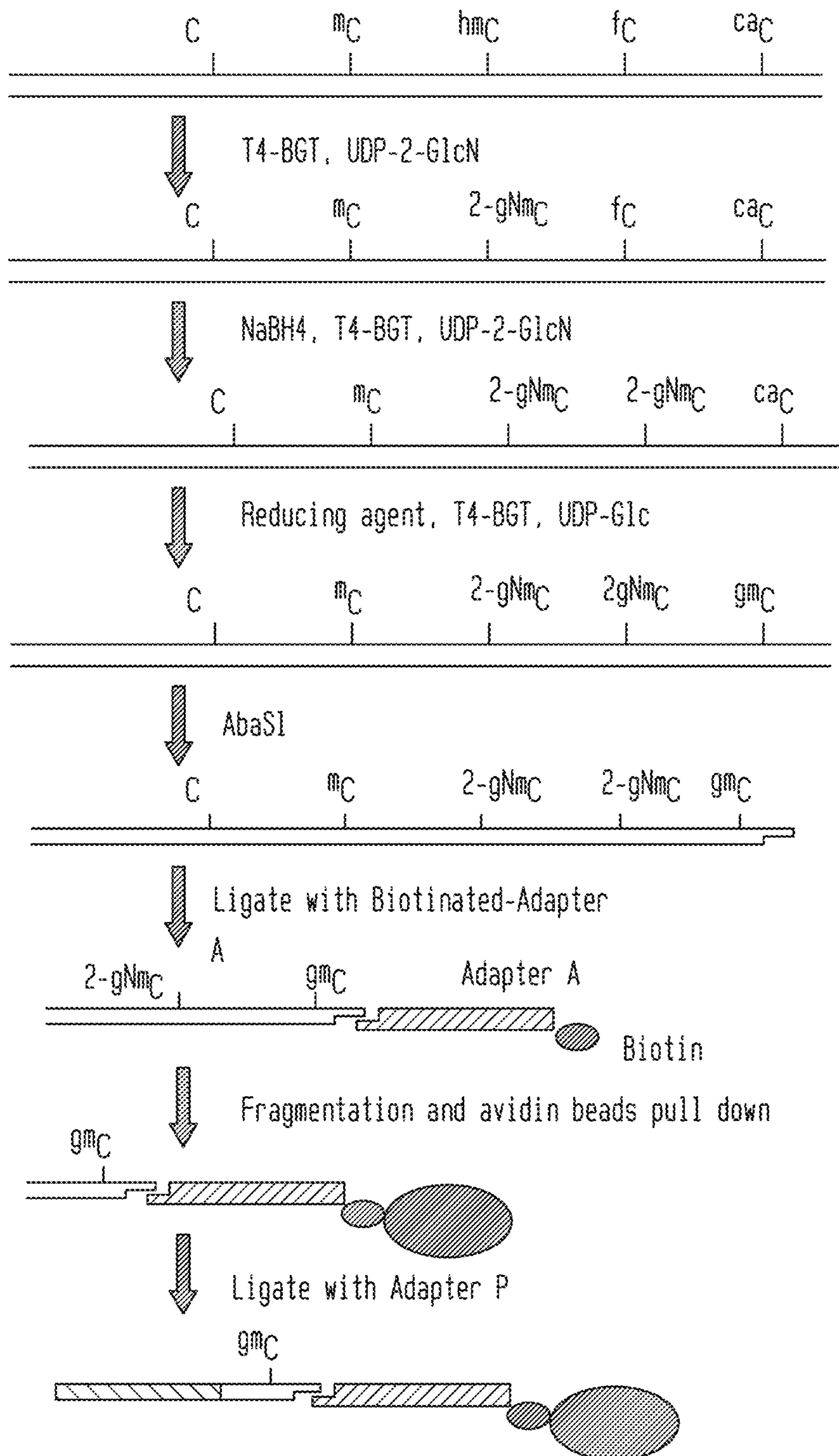
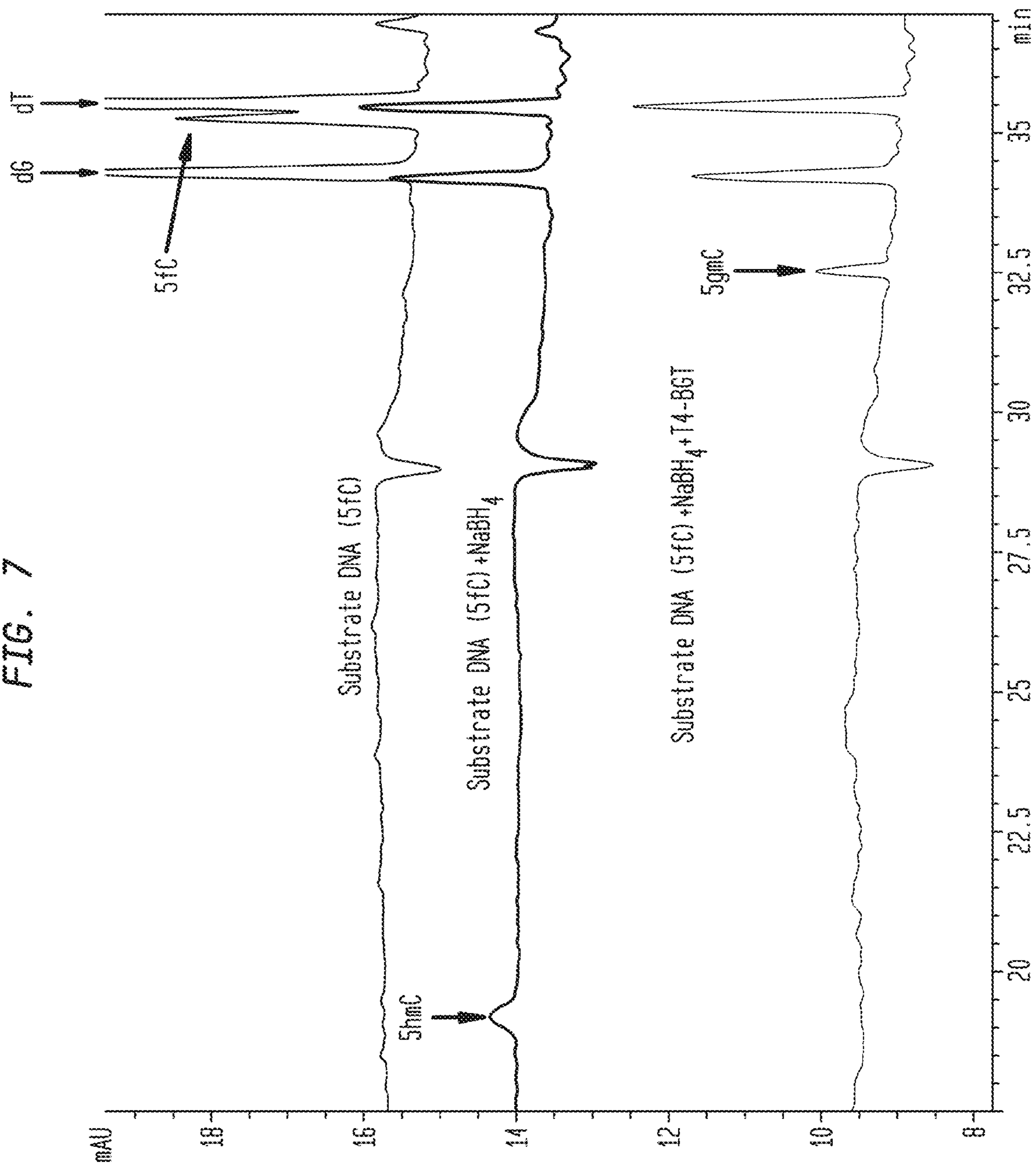


FIG. 7



MAPPING CYTOSINE MODIFICATIONS

REFERENCE TO RELATED APPLICATIONS

This application claims the benefit of each of the following patent applications, each of which are hereby incorporated by reference into the present application: U.S. 61/611,295, filed Mar. 15, 2012; U.S. Application No. 61/722,968, filed Nov. 6, 2012; U.S. Application No. 61/723,427, filed Nov. 7, 2012; U.S. Application No. 61/724,041, filed Nov. 8, 2012; and U.S. application Ser. No. 13/804,804, filed Mar. 14, 2013. Also incorporated by reference in their entireties are the following applications filed on the same day as the present application: Ser. No. 13/827,087, "Compositions and Methods for Oxygenation of Nucleic Acids Containing 5-Methylpyrimidine"; and Ser. No. 13/827,885, "Methods and Compositions for Discrimination Between Cytosine and Modifications Thereof, and for Methylome Analysis."

GOVERNMENT RIGHTS

This invention was made with government support under GM096723 awarded by the National Institutes of Health. The government has certain rights in this invention.

BACKGROUND

5-hydroxymethylcytosine (5-hmC), 5-formylcytosine (5-fC), and 5-carboxycytosine (5-caC) were recently identified in mammalian brain and embryonic stem cells as products of the oxidation of 5-methylcytosine (5-mC) by cytosine oxygenases. The biological functions of 5-hmC, 5-fC, and 5-caC are not completely understood; however, several lines of evidence suggest that 5-hmC is involved in epigenetic regulation and DNA demethylation. Iterative oxidation of 5-hmC by cytosine oxygenase enzymes yields 5-fC and 5-caC which are hypothesized to be intermediates in the DNA demethylation process. Several challenges are associated with the identification these biologically modified nucleobases in genomic DNA samples due to their low abundance and temporal fluctuation. Mapping and quantifying 5-mC, 5-hmC, 5-fC, and 5-caC at the DNA level is, therefore, important for unraveling their role in the dynamics of gene expression and regulation.

SUMMARY OF THE INVENTION

The present invention provides a variety of reagents, kits and methods for selectively altering and identifying modified nucleotides in a nucleic acid such as DNA. The modified nucleotides that can be identified include, for example, 5-mC, 5-hmC, 5-fC and 5-caC. The methods, reagents and kits of the present invention permit not merely the determination that a modified cytosine residue is present, but also permit the discrimination among or between different types of modified nucleotides, such as 5-mC and 5-hmC, or between 5-hmC and 5-fC. In this way, the invention can be used to elucidate the precise state of a modified nucleic acid, which may be a genome or genome fragment, for example.

In some embodiments, the discrimination among or between different oxidation states of a nucleotide is facilitated by alternately glucosylating or glucosaminylating various modified nucleotides. The glucosylated or glucosaminylated forms can be distinguished based on their performance in various assays, such as by their differential sensitivity to certain restriction endonucleases. In this way, alternately reacting the nucleic acid with a UDP-GlcN (Uridine diphos-

phate glucosamine or UDP-Glucosamine) substrate and a UDP-Glc (Uridine diphosphate glucose or UDP-Glucose) substrate, places off and on switches for endonuclease cleavage of a nucleic acid containing modified nucleotides. By detecting the presence of cleavage-sensitive sites, the modified nucleotides can be located and identified.

In the present invention, the term glucosylation (and any form of glucosylation such as "glucosylating" or "glucosylated") refers to the incorporation of a glucosyl moiety from UDP-Glc into the 5-hydroxy position of 5-hmC via the action of a glycosyltransferase to produce 5-gmC (5-glucosyloxymethylcytosine). Other names in common use for 5-gmC include glucosyl-5-hydroxymethyl-cytosine, glucosyl-5-hydroxymethylcytosine, glucosyl-oxy-5-methylcytosine, 5-glucosylhydroxymethylcytosine. In the present invention, the term glucosylation (and any form of glucosylation such as "glucosylating" or "glucosylated") may also refer to the incorporation of an azido modified glucosyl moiety from UDP-Azido-Glc (for example, UDP-6-Azido-Glc or UDP-6-N3-Glc) into the 5-hydroxy position of 5-hmC via the action of a glycosyltransferase to produce a N3-5-gmC (for example, 6-azido-glucosyl-5-hydroxymethylcytosine or 6-N3gmC). In the present invention, the term glucosaminylation (and any form of glucosaminylation such as "glucosaminylating" or "glucosaminylated") refers to the incorporation of a glucosaminyl moiety from UDP-GlcN into the 5-hydroxy position of 5-hmC via the action of a glycosyltransferase to produce 5-gNmC (5-glucosaminylloxymethylcytosine). Other names for 5-gNmC include glucosaminyl-5-hydroxymethyl-cytosine, glucosaminyl-5-hydroxymethylcytosine, 5-glucosylhydroxymethylcytosine, aminoglucosyl-5-hydroxymethyl-cytosine, aminoglucosyl-5-hydroxymethylcytosine, and 5-aminoglucosylhydroxymethylcytosine. If the glycosyltransferase is an inverting glycosyltransferase such as T4 β -glucosyltransferase (BGT or β GT or beta-GT) the product is formed with a beta glycosydic linkage (for example, 5- β -gmC, 5- β -6-N3gmC, 5- β -6-gNmC, and 5- β -2-gNmC). If the glycosyltransferase is a retaining glycosyltransferase such as T4 α -glucosyltransferase (AGT or α GT or alpha-GT) the product is formed with an alpha glycosydic linkage (for example, 5- α -gmC).

Specifically, for example, the invention permits the differentiation of 5- β -glucosyloxymethylcytosine (5- β -gmC) from 5- β -2-glucosaminylloxymethylcytosine (5- β -2-gNmC) in a nucleic acid, by reacting the nucleic acid with an endonuclease capable of cleaving a nucleic acid at a glucosylated nucleotide but not at an glucosaminylated nucleotide. One suitable endonuclease is AbaSI. Other useful endonucleases include AbaAI, AbaCI, AbaDI, AbaTI, AbaUI, AcaPI, and PxyI, or one of the ZZYZ proteins or its variants described in US Patent Application Publication No. 2012/0301881, the complete disclosure of which is hereby incorporated by reference. Accordingly, one of these endonucleases, or a polypeptide at least 70% (e.g. at least 75%, at least 80%, at least 82%, at least 84%, at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, or 100% identical to one of those endonucleases or an active fragment thereof) can be used to selectively cleave a target nucleic acid. By controlling the conversion of glucosylation and glucosaminylation of various forms of naturally occurring modified nucleotides, the modified nucleotides can be located and identified in a manner distinguishing their original forms, such as methylcytosine, hydroxymethylcytosine, formylcytosine, and/or carboxycytosine.

Accordingly, in one aspect the invention provides methods for selectively altering modified nucleotides in a nucleic acid containing hydroxymethylated nucleotides and other modified nucleotides such as, 5-fC or 5-caC. The methods generally include reacting a first subset of the modified nucleotides (e.g. 5-hmC) in the nucleic acid with UDP-GlcN in the presence of a β -glycosyltransferase, such as T4- β -glucosyltransferase, to convert hydroxymethylated nucleotides in the nucleic acid to glucosaminylated nucleotides.

In some embodiments, the methods include the subsequent step of reacting the nucleic acid with a reducing agent, such as NaBH_4 , and a UDP derivative, such as UDP-Glc or UDP-azido-glucose, to convert a second subset of nucleotides in the nucleic acid to glucosylated or azidoglucosylated nucleotides. The reducing agent promotes the reduction of a nucleotide in a higher oxidation state, such as 5-fC or 5-caC, to a hydroxymethylated nucleotide, which is then glucosylated or azidoglucosylated in a reaction typically catalyzed by a glycosyltransferase such as an α -glycosyltransferase or a β -glycosyltransferase. The glycosyltransferase may or may not be the same glycosyltransferase used to catalyze the previous conversion of hydroxymethylated nucleotides to glucosaminylated nucleotides. The method can optionally differentiate pre-existing 5-hmC in a nucleic acid from newly formed as a result of the reduction of 5-fC or 5-caC, for example. When a UDP-azido-glucose such as UDP-6-azido-glucose is used, 5-hmC in the nucleic acid can be converted to a β -6-azide-glucosyl-5-hydroxymethylcytosine in the presence of a β -glycosyltransferase; Further derivatization of the azido moiety via azide-alkyne Huisgen cycloaddition using a copper(I) catalyst ("click chemistry") or via copper-free variants (for example, using strained cyclooctyne derivatives) can then optionally be used to label the nucleotide, such as with a biotin label, permitting the subsequent use of avidin to selectively isolate the labeled nucleotide.

In other embodiments, following the conversion of hydroxymethylated nucleotides in the nucleic acid to glucosaminylated nucleotides, the nucleic acid is reacted with an oxidizing agent, a UDP-Glc derivative, and a glycosyltransferase to convert a second subset of modified nucleotides in the nucleic acid to glucosylated nucleotides. Suitable oxidizing agents include those capable of oxidizing 5-mC to 5-hmC, such as mYOX1, a TET enzyme, or an inorganic oxidizing agent such as KRuO_4 . The reaction with UDP-Glc is typically conducted in the presence of a glycosyltransferase such as an α -glycosyltransferase or a β -glycosyltransferase. These methods can, for example, differentiate pre-existing 5-hmC in a nucleic acid from 5-hmC generated from a 5-mC precursor.

In any of these methods, whether incorporating a reducing agent or an oxidizing agent, an endonuclease can be used to characterize the reaction products. Typically, the endonuclease is specific for the glucosylated (or azidoglucosylated) nucleotides, and not for glucosaminylated nucleotides, i.e., the endonuclease has a higher enzymatic activity for a glucosylated or azidoglucosylated nucleic acid than it has for the same nucleic acid with an glucosaminylated nucleotide at the same position(s). Endonucleases that may be used include, for example, AbaSI, AbaAI, AbaCI, AbaDI, AbaTI, AbaUI, AcaPI, and/or PxyI. An adapter molecule can then be ligated to the cleaved end of the endonuclease reaction product, facilitating the subsequent purification, amplification or sequencing of the nucleic acid.

Accordingly, in one embodiment the invention provides a method for differentiating a 5-mC from 5-hmC in a genome or genome fragment. The genome or genome fragment may, for example, be mammalian in origin, such as a human

genome or genome fragment. The method includes reacting the isolated genome or genome fragment containing 5-mC and 5-hmC with (i) UDP-GlcN in the presence of a glycosyltransferase catalyzing transfer of glucosamine to the 5-hmC; (ii) oxygenating any existing 5-mC residues to 5-hmC by the action of TET or mYOX1; (iii) reacting the newly created 5-hmC sites with UDP-Glc in the presence of a glycosyltransferase catalyzing transfer of glucose to the 5-hmC; (iv) cleaving the glucosylated template with a modification-dependent endonuclease that recognizes at least one of the modified nucleotides; and (v) differentiating the original 5-mC from the 5-hmC by an altered cleavage pattern.

In this embodiment, the oxygenation of any existing 5-mC residues to 5-hmC can be done in the presence of UDP-Glc and a glycosyltransferase to catalyze the transfer of glucose to 5-hmC as it is being formed from 5-mC. Alternatively, reaction conditions such as pH of the oxygenation reaction can be selected to optimize the yield of 5-hmC. See, for example, Ser. No. 13/827,087, "Compositions and Methods for Oxygenation of Nucleic Acids Containing 5-Methylpyrimidine," filed on the same date as the present application and hereby incorporated by reference in its entirety.

In another embodiment, the invention provides a method for differentiating a 5-mC from one or more of its oxidation products in a genome or genome fragment containing 5-mC and 5-hmC. The method includes reacting the isolated genome or genome fragment with UDP-2-glucosamine in the presence of a β -glycosyltransferase (BGT) catalyzing the transfer of 2-glucosamine to the 5-hmC; reacting the isolated genome or genome fragment with mYOX1 or TET or a chemical oxidizing agent and optionally with a reducing agent; cleaving the template with a modification dependent endonuclease that is capable of selectively cleaving a 5-hmC and not a 5-glucosaminated hydroxymethylcytosine; and differentiating the 5-mC from one or more of its oxidation products by an altered cleavage pattern.

The invention also provides preparations useful for converting methylcytosine or an oxidized nucleotide, such as 5-fC or 5-caC, or to a glucosylated nucleotide. The preparations include a reducing agent, such as NaBH_4 , or an oxidizing agent, such as mYOX1, a TET enzyme, or an inorganic oxidizing agent such as KRuO_4 ; a glycosyltransferase, such as an α -glycosyltransferase or a β -glycosyltransferase; a UDP-GlcN or a UDP derivative, such as UDP-Glc or UDP-azido-glucose.

The invention also provides preparations useful for modifying and selectively cleaving nucleic acids. The preparations include a glycosyltransferase and an endonuclease having an amino acid sequence at least 95% identical to an enzyme such as AbaSI, AbaAI, AbaCI, AbaDI, AbaTI, AbaUI, AcaPI, and/or PxyI. The preparations also include (a) UDP-Glc and an oxidizing agent, or (b) UDP-GlcN. Where UDP-GlcN is included, the glycosyltransferase may be a BGT to catalyze the transfer of glucose to hydroxymethylated pyrimidine residues. Where UDP-Glc and an oxidizing agent are included, the oxidizing agent, which may be a methylcytosine oxygenase such as mYOX1 or a TET enzyme or may be an inorganic oxidizing agent such as KRuO_4 , promotes the conversion of methylcytosine residues in a nucleic acid to hydroxymethylcytosine, which can be glucosylated by the combination of UDP-Glc and the glycosyltransferase, and can be recognized by the endonuclease.

The invention also provides kits useful for making these preparations and practicing these methods. For example, kits for modifying formylcytosine or carboxycytosine residues in a nucleic acid can include a reducing agent, such as sodium borohydride, permitting the reduction of formylcytosine or

carboxycytosine residues to hydroxymethylcytosine; a glycosyltransferase (such as a β -glycosyltransferase); and a UDP derivative, such as UDP-Glc and/or UDP-azidoglucose (such as UDP-6-azidoglucose), permitting the transfer of a sugar or modified sugar to the hydroxymethylcytosine. These kits may also include an oxidizing agent (e.g., a methylcytosine oxygenase such as mYOX1 or a TET enzyme or an inorganic oxidizing agent such as KRuO_4) to promote the conversion of methylcytosine residues in a nucleic acid to hydroxymethylcytosine; UDP-GlcN; a restriction endonuclease (e.g., having an amino acid sequence at least 95% identical to AbaSI, AbaAI, AbaCI, AbaDI, AbaTI, AbaUI, AcaPI, and/or PxyI); or any combination of the above.

Kits for selectively modulating the susceptibility of modified nucleic acid residues to cleavage can include UDP-Glc; UDP-GlcN; a β -glycosyltransferase; and a reducing (e.g. sodium borohydride) or oxidizing agent (e.g., a methylcytosine oxygenase such as mYOX1 or a TET enzyme or an inorganic oxidizing agent such as KRuO_4).

Kits useful for modifying and selectively cleaving nucleic acids can include, for example, a glycosyltransferase and an endonuclease having an amino acid sequence at least 95% identical to an enzyme such as AbaSI, AbaAI, AbaCI, AbaDI, AbaTI, AbaUI, AcaPI, and/or PxyI. These kits can also include (a) UDP-Glc and an oxidizing agent, and/or (b) UDP-GlcN.

Some embodiments of the kits or preparations of the present invention include an optional nucleic acid, such as a nucleic acid that is to be modified, that is undergoing modification, and/or a control nucleic acid. Accordingly, a nucleic acid, if present, may include 5-hmC, 5-gmC, 5-gNmC, 5-fC, 5-caC, 5-mC, or any combination of the above.

BRIEF DESCRIPTION OF THE FIGURES

FIG. 1 shows that AbaSI recognizes 5-(β -glucosyloxymethyl)cytosine (5- β -gmC) but not 5-(β -2-glucosaminyloxymethyl)cytosine (5- β -2-gNmC) with high specificity as compared to 5-mC and C. The different forms of cytosine modification (panels a-g) were digested by a 10-fold serial dilution of AbaSI enzyme (lane 1 to lane 7, lane 8 is undigested control).

FIG. 2 depicts the relative selectivity of a variety of endonucleases for nucleic acid substrates containing 5- β -gmC (β -glucosylated hmC), 5- α -gmC (α -glucosylated hmC), 5-hydroxymethylcytosine (hmC), 5-methylcytosine (mC), or unmodified cytosine (C).

FIG. 3 depicts selected cytosine modifications that can be achieved through the addition of glucose, modified glucose, or glucosamine residues by T4-BGT alone or in combination with β -glucosyl- α -glycosyltransferase from bacteriophage T6 (T6-BGAGT).

FIG. 4 depicts one example of an assay for detecting and/or mapping 5-mC.

FIG. 5 depicts two examples of assays for detecting and/or mapping 5-fC. The assay in FIG. 5A includes a step involving glucosylating hydroxymethylcytosine residues newly-formed from the reduction of 5-fC. The assay in FIG. 5B includes a step involving azidoglucosylating hydroxymethylcytosine residues newly-formed from the reduction of 5-fC.

FIG. 6 depicts an example of an assay for detecting and/or mapping 5-caC.

FIG. 7 depicts LC-MS analyses of a nucleic acid containing 5-fC (top trace) and the same nucleic acid after treatment

with 100 mM sodium borohydride (middle trace) or with sodium borohydride, UDP-Glc, and T4-BGT (bottom trace).

DETAILED DESCRIPTION OF EMBODIMENTS

Endonucleases have been identified from bacteria and more will undoubtedly be discovered using routine BLAST searches based on the present disclosure that are capable of preferentially cleaving 5- β -gmC compared to C, 5-mC and 5- β -2-gNmC. In one example, the ZZY family members for example AbaSI (see for example WO 2011/091146), isoschizomers and mutants thereof preferentially cleave 5- β -gmC compared to C, 5-mC and 5- β -2-gNmC. For example, AbaSI has cleavage activity at 5- β -gmC that is 500 fold greater than 5-mC and 5- α -gmC. 5- β -gmC is the product of BGT mediated transfer of glucose from UDP-Glc to 5-hmC.

The specificity of AbaSI is demonstrated in the data shown in FIG. 1. AbaSI cleaves recognizes nucleic acids containing certain cytosine modifications, cleaving them a short distance from those modifications. Ten-fold serial dilutions of AbaSI were combined with a PCR product containing cytosine (C, panel a), 5-methylcytosine (5-mC, panel B), 5-hydroxymethylcytosine (5-hmC, panel C), 5-(β -glucosyloxymethyl)cytosine (5- β -gmC, panel D), 5-(β -2-glucosaminyloxymethyl)cytosine (5-(β -2-gNmC, panel E), 5-(β -6-glucosaminyloxymethyl)cytosine (5- β -6-gNmC, panel F), or 5-(β -6-azidoglucosyloxymethyl)cytosine (5- β -6-N3gmC, panel G). The PCR products containing 5- β -gmC, 5- β -2-gNmC, 5- β -6-gNmC, or 5- β -6-N3gmC were generated by treatment of 5-hmC PCR DNA (panel c) with T4- β -glucosyltransferase (BGT) and the corresponding UDP-sugar (uridine diphospho-glucose (UDP-Glc), UDP-2-glucosamine (UDP-2-GlcN), UDP-6-glucosamine (UDP-6-GlcN), and UDP-6-N3-Glc, respectively). The results show that AbaSI does not digest C and 5-mC even at high concentrations (panels a and b) whereas AbaSI can digest 5- β -gmC even at very low concentrations (panel d). With respect to the three glucosylated or glucosaminated cytosines, AbaSI can digest 5- β -6-gNmC and 5- β -6-N3gmC (panels f and g) more efficiently than it digests 5-hmC (although less efficiently than it digests 5- β -gmC. In contrast, it does not digest 5- β -2-gNmC (panel e).

Other endonucleases that differentiate between various forms of modified cytosine are also available, such as other ZZY family members described in WO 2011/091146 and enzymes described in Borgaro et al. (2013) "Characterization of the 5-hydroxymethylcytosine-specific DNA restriction endonucleases," *Nucleic Acids Research*, doi: 10.1093/nar/gkt102, the entire disclosures of each of which are incorporated herein by reference. Several endonucleases that discriminate among various cytosine modifications are described in FIG. 2. For example, PvuRts1I cleaves nucleic acids containing hydroxymethylcytosine or glucosyloxymethylcytosine far more efficiently than it cleaves nucleic acids containing only methylcytosine or unmodified cytosine. AbaSI, as described in the preceding paragraph, cleaves nucleic acids containing 5- β -gmC more efficiently than it cleaves nucleic acids containing 5-hmC, which are nevertheless cleaved more efficiently than those nucleic acids containing only 5-mC or unmodified cytosine. Like AbaSI, AbaAI, AbaCI, AbaDI, AbaTI, AbaUI, AcaPI, and PxyI all demonstrate increased specificity for nucleic acids containing 5- β -gmC compared to nucleic acids containing only 5-hmC. Accordingly, for any of these enzymes β -glucosylation of hydroxymethylcytosine can be used as an "on switch" to promote cleavage near those positions, whereas β -glucosylation could be used as an "off switch" for enzymes such as

PvuRtsII, AbaBGI, BbiDI, BmeDI, or PatTI. On the other hand, α -glucosylation of 5-hmC generates a more efficient substrate for PvuRtsII, and could be used as an “on switch” to target cleavage events to those locations in a nucleic acid.

It is expected that the use of endonucleases with preferential specificity for a specific modified nucleotide over other modified and unmodified nucleotides can be detected using the method described herein for 5-mC, 5-hmC, 5-fC, and 5-caC. In combination with the cofactor UDP-Glc, this system enables sequencing different epigenetic states of 5-mC and greatly enhances the ability to determine the epigenetic modification at a single base resolution level.

An embodiment of the method relies on modifying the non-target modified base (for example, 5-hmC) chemically and/or enzymatically in such a way that its reactivity to an endonuclease is completely or partially blocked, and then chemically or enzymatically reacting the target modified base to convert it into a newly formed 5-hmC (e.g., with a reducing or oxidant agent) which can be then be reacted with a glycosyltransferase to form 5-gmC which in turn can be cleaved by an endonuclease that recognizes 5-gmC preferentially in a positive identification.

By the appropriate choice of the substrate for modifying the non-target and target modified bases, this invention provides an on-off switch assay which is determined by enzyme specificity. For example, if in the first step UDP-2-GlcN is used to label pre-existing 5-hmC sites (resulting in an off-signal for AbaSI cleavage), and in the second step the sample is treated with a methylpyrimidine oxygenase (mYOX), BGT and UDP-Glc, an “on-signal” for AbaSI cleavage will be generated exclusively for 5-mC sites which underwent oxidation by mYOX. If UDP-Glc is used to label pre-existing 5-hmC sites (on-signal for AbaSI cleavage), and the sample is treated with an mYOX, BGT and UDP-2-GlcN, then an “off-signal” will be generated for all 5-mC sites which were converted into 5-hmC by mYOX-mediated oxidation.

The 5-mC may be chemically or enzymatically converted to 5-hmC by reacting the 5-mC with mYOX1 (see for example, “nMCO1” described in U.S. Provisional Application No. 61/723,427), TET enzymes or chemical oxidizing agents. Similarly, oxidation of 5-mC to 5-hmC to 5-fC to 5-caC can be achieved by chemical or enzymatic oxidation using mYOX1 or TET. Specific chemical oxidation of 5-hmC to 5-fC in synthetic nucleotide oligomer single strand (ss-DNA) containing 5-hmC can be achieved with potassium perruthenate, KRuO₄ (Booth, et al. *Science*, 336:934-937 (2012)). KRuO₄ can oxidize 5-hmC in double-stranded DNA (dsDNA), with an initial denaturing step before the addition of the oxidant, resulting in quantitative conversion of 5-hmC to 5-fC. Other oxidants known in art, such as Osmium (VIII)-based oxidants, Cerium (IV)-based oxidants, and Chromium (VI)-based oxidants may be used for the oxidation of 5-hmC to 5-fC.

A variety of sugars and modified sugars can be used to alter the propensity of a 5-hmC to trigger an endonuclease-mediated cleavage event. Some of these sugars and modified sugars are depicted in FIG. 3. For example, T4-BGT can be used to add glucose, 2-glucosamine, 6-glucosamine, or 6-azido-glucose to 5-hmC. T4-BGT can also be used in combination with a β -glucosyl- α -glycosyltransferase, such as the one from bacteriophage T6, to generate disaccharidyl cytosine modifications as shown in FIG. 3. By controlling the modifications made to a particular nucleobase, the properties of that nucleobase can be changed in a manner facilitating its discrimination, whether through changes in its reactivity with an

endonuclease; changes in the kinetics of synthesis of a complementary nucleic acid; or directly measured changes in size, shape, or charge density.

The reduction of 5-fC and its conversion to 5- β -gmC was demonstrated in the experiment depicted in FIG. 7. Specifically, FIG. 7 is an LC-MS analysis of a nucleic acid originally including 5-fC (top trace). In the presence of sodium borohydride (NaBH₄), the 5-fC is converted to 5-hmC (middle trace). When UDP-Glc and a BGT such as T4-BGT were also provided, the 5-fC was converted all the way to 5- β -gmC, confirming that the modified forms of cytosine can be interconverted, facilitating their subsequent detection and discrimination.

In one embodiment the method comprises one or more of the following steps;

- (a) Genomic DNA is treated with BGT and UDP-2-GlcN, so that all 5-hmC residues are converted to 5- β -2-gNmC.
- (b) (i) The resulting DNA is treated with mYOX1, a Tet enzyme or a chemical oxidant agent (converts 5-mC to 5-hmC), BGT and UDP-Glc, so that all existing 5-mC residues are converted to 5- β -gmC; or
 - (ii) The resulting DNA is treated with a reducing agent (e.g., NaBH₄, converts 5-fC to 5-hmC), BGT and UDP-Glc to generate 5- β -gmC; or
 - (iii) The resulting DNA is treated with a reducing agent (e.g., NaBH₄, converts 5-fC to 5-hmC), BGT and UDP-2-GlcN, so that all 5-fC residues are converted to 5- β -2-gNmC. Then, the resulting DNA is treated with a second reducing agent (a different reducing agent or the same reducing agent but in the presence of certain additives that converts 5-caC to 5-hmC), BGT, and UDP-Glc to generate 5- β -gmC.
- (c) (i) The DNA is digested with a 5- β -gmC-dependent endonuclease such as AbaSI, which cleaved at a fixed distance from 5- β -gmC and left a sticky end (2-base 3'-overhang). Since the endonuclease does not recognize C or 5- β -2-gNmC no cleavage associated with these sites occurs. The only sticky ends created are those resulting from 5- β -gmC residues, which in turn are exclusively associated to 5-mC sites; or
 - (ii) The DNA is digested with endonuclease, which cleaves 5- β -gmC exclusively associated to 5-fCs sites, but not 5- β -2-gNmC,
 - (iii) the DNA is digested with endonuclease, which cleaves 5- β -gmC exclusively associated to CaC sites, but not 5- β -2-gNmC, 5-mC, or C, leaving a sticky end (2-base 3'-overhang).
- (d) A first adaptor (e.g., biotinylated adaptor A) is then ligated onto the cleaved ends.
- (e) The ligated DNA is then subjected to random fragmentation to about 200 bp.
- (f) Beads may be used to pull out the fragments with the ligated adaptor. For example, avidin beads may be used to pull out the biotin labeled adaptor (adaptor A). A person of ordinary skill in the art will recognize other affinity systems and immobilization matrices that can be used in place of biotin/avidin beads.
- (g) After polishing the ends, adaptor P is then ligated onto the DNA fragments on the beads.
- (h) The adaptor-specific PCR and the adapter ligated DNA enters the library preparation pipeline for specific sequencing platform where the end-sequencing is done from the adaptor A.

Reducing agents and conditions can be used herein that can convert the carboxylic acids into alcohols, e.g., NaBH₄, CoCl₂, i-Pr₂NH, EtOH/H₂O (Jagdale, et al. *Synthesis*, 660-664 (2009)); EDC, HOBt, NaBH₄, THF/H₂O (Morales-

Serna, et al. *Synthesis*, 1375-1382 (2011)); and cyanuric chloride, NaBH₄, NMM/H₂O (Falorni, et al. *Tetrahedron Lett.*, 4395-4396 (1999)) as well as other reducing agents known to a person of ordinary skilled in the art.

Indeed, many water-soluble metal or metalloid hydrides are able to reduce aldehydes and/or carboxylic acids to alcohols. Examples of such reducing agents are sodium borohydride and related compounds where from 1 to 3 of the hydrogens are replaced by other moieties, such as cyano and alkoxy containing up to about 5 carbon atoms. Examples of substituted borohydrides, all of which are sodium, potassium, or lithium salts, include cyanoborohydride, dicyanoborohydride, methoxyborohydride, dimethoxyborohydride, trimethoxyborohydride, ethoxyborohydride, diethoxyborohydride, triethoxyborohydride, propoxyborohydride, dipropoxyborohydride, tripropoxyborohydride, butoxyborohydride, dibutoxyborohydride, tributoxyborohydride, and so forth. Examples of other water-soluble metal hydrides include lithium borohydride, potassium borohydride, zinc borohydride, aluminum borohydride, zirconium borohydride, beryllium borohydride, and sodium bis(2-methoxyethoxy)aluminum hydride. Sodium borohydride can also be used in combination with a metal halide, such as cobalt(II), nickel(II), copper(II), zinc(II), cadmium (II), calcium (II), magnesium(II), aluminum(III), titanium (IV), hafnium(IV), or rhodium(III), each of which can be provided as a chloride, bromide, iodide, or fluoride salt. Alternatively, sodium borohydride can be used in combination with iodine, bromine, boron trifluoride diethyl etherate, trifluoroacetic acid, catechol-trifluoroacetic acid, sulfuric acid, or diglyme. Particular reducing strategies include the combination of potassium borohydride with lithium chloride, zinc chloride, magnesium chloride, or hafnium chloride; or the combination of lithium borohydride and chlorotrimethylsilane. Other reducing strategies include the use of borane, borane dimethyl sulfide complex, borane tetrahydrofuran complex, borane-ammonia complex, borane morpholine complex, borane dimethylamine complex, borane trimethylamine complex, borane N,N-diisopropylethylamine complex, borane pyridine complex, 2-picoline borane complex, borane 4-methylmorpholine complex, borane tert-butylamine complex, borane triphenylphosphine complex, borane N,N-diethylaniline complex, borane di(tert-butyl)phosphine complex, borane diphenylphosphine complex, borane ethylenediamine complex, or lithium ammonia borane. Alternative reducing strategies include the reduction of carboxylic acids via the formation of hydroxybenzotriazole esters, carboxy methyleniminium chlorides, carbonates, O-acylisoureas, acyl fluorides, cyanurates, mixed anhydrides, arylboronic anhydrides, acyl imidazolide, acyl azides, or N-acyl benzotriazoles, followed by reaction with sodium borohydride to give the corresponding alcohols.

In one embodiment, 5-hmC sequencing can use BGT and UDP-Glc to generate AbaSI-active 5-β-gmC sites, and T4-α-glucosyltransferase (AGT) and UDP-Glc to generate AbaSI-inactive 5-α-gmC sites as a negative control. In another embodiment, 5-hmC sequencing can use BGT and UDP-Glc to generate the AbaSI-active 5-β-gmC sites, and BGT and UDP-2-GlcN to generate AbaSI-inactive 5-β-2-gNmC as a negative control for 5-hmC.

In one embodiment, newly generated 5-hmC sites can be differentiated from pre-existing 5-hmC sites by sequentially transferring distinct sugar moieties from UDP-2-GlcN or native UDP-Glc using BGT.

In a further embodiment of the invention, UDP-Glc modified by any of keto, thiol, chloro, fluoro, bromo, iodo, nitro, boron, and other substituents may be transferred onto 5-hmC

using BGT and may block AbaSI activity. Keto, thiol, chloro, fluoro, bromo, iodo, nitro, boron, and other substituents modifying glucose containing 5-hmC residues in a nucleic acid may facilitate cytosine modification mapping and inhibit AbaSI cleavage.

All references cited herein are incorporated by reference.

EXAMPLES

Example 1

Mapping of 5-hmC in a Nucleic Acid Sequence

The locations of 5-hmC in a nucleic acid sample can be determined using differential cleavage.

A BGT can transfer glucose ("Glc") from UDP-Glc to 5-hmC to form the glucosylated residue 5-β-2-gmC. Glucosylation enhances the sensitivity of the nucleic acid to a glucosylation-sensitive restriction enzyme such as AbaSI (see FIG. 2). Accordingly, the identification of 5-hmC sites in a sample can be facilitated by glucosylating the nucleic acid, followed by identifying the locations of AbaSI cleavage sites.

Example 2

Exclusive 5-mC Methylome Mapping

An exemplary process for mapping 5-mC residues in a nucleic acid is depicted in FIG. 4.

As shown in FIG. 4, genomic DNA is treated with a BGT and UDP-2-glucosamine, converting 5-hmC residues to 5-β-2-gNmC. The resulting DNA is treated with a methylpyrimidine oxygenase from *Neisseria* (mYOX1), TET or a chemical oxidizing agent, BGT and UDP-Glc, converting existing 5-mC residues to 5-β-gmC. The DNA is digested with a 5-β-gmC-dependent restriction enzyme, such as AbaSI, cleaving at a fixed distance from 5-β-gmC and leaving a sticky end (2-base 3'-overhang). Since AbaSI does not recognize C or 5-β-2-gNmC, the only sticky ends created are those resulting from 5-β-gmC residues, which in turn are exclusively associated to 5-mC sites. A biotinylated adaptor A is then ligated onto the cleaved ends. The ligated DNA is then subjected to random fragmentation to an average size of about 200 bp. Avidin beads are used to pull out the fragments with the ligated adaptor A. After polishing the ends, adaptor P is then ligated onto the DNA fragments on the beads. The adaptor-specific PCR and the adapter ligated DNA enters the library preparation pipeline for specific sequencing platform where the end-sequencing is done from the adaptor A.

Bioinformatic analysis of the sequencing reads is facilitated by the presence of adapter A which marks the enzyme cleavage sites. After mapping the read back to the reference genome, the modified cytosine can be mapped at fixed distance away from the cleavage sites.

Example 3

Exclusive 5-fC and/or 5-caC Mapping

Exemplary processes for identifying 5-fC or 5-caC residues in a nucleic acid are depicted in FIGS. 5A and 5B.

Example 3A

Glucosamination

As shown in FIG. 5A, a BGT can be used to catalyze the addition of 2-glucosamine from UDP-2-GlcN to a 5-hmC

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residue, converting those residues to 5- β -2-gNmC. A reducing agent such as NaBH₄ (optionally in the presence of additives), can be used to reduce 5-fC and/or 5-caC to 5-hmC. The newly formed 5-hmC when combined with BGT and UDP-Glc can be subsequently converted to 5- β -gmC.

As described in Example 2, the DNA is digested with a 5- β -gmC-dependent restriction enzyme, such as AbaSI, cleaving at a fixed distance from 5- β -gmC and leaving a sticky end (2-base 3'-overhang). Since AbaSI does not recognize C or 5- β -2-gNmC, the only sticky ends created are those resulting from 5- β -gmC residues, which in turn are exclusively associated to 5-fC and/or 5-caC sites. A biotinylated adaptor A is then ligated onto the cleaved ends. The ligated DNA is then subjected to random fragmentation to an average size of about 200 bp. Avidin beads are used to pull out the fragments with the ligated adaptor A. After polishing the ends, adaptor P is then ligated onto the DNA fragments on the beads. The adaptor-specific PCR and the adapter ligated DNA enters the library preparation pipeline for specific sequencing platform where the end-sequencing is done from the adaptor A.

Bioinformatic analysis of the sequencing reads is facilitated by the presence of adapter A which marks the enzyme cleavage sites. After mapping the read back to the reference genome, the modified cytosine can be mapped at fixed distance away from the cleavage sites.

Example 3B

6-Azido-Glucose

Another exemplary process for mapping the locations of 5-fC and/or 5-caC is shown in FIG. 5B. The process depicted in FIG. 5B, like the process depicted in FIG. 5A, begins with the addition of 2-glucosamine from UDP-2-GlcN to a 5-hmC residue, converting those residues to 5- β -2-gNmC in a reaction catalyzed by a BGT, and the subsequent reduction of 5-fC and/or 5-caC residues to newly-generated 5-hmC residues. In the method depicted in FIG. 5B, UDP-6-azido-glucose

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(UDP-6-N3-Glc) is added to those newly-generated 5-hmC residues by a BGT. The DNA is digested with a 5- β -gmC-dependent restriction enzyme, such as AbaSI, cleaving at a fixed distance from 5- β -gmC and leaving a sticky end (2-base 3'-overhang). Since AbaSI does not recognize C or 5- β -2-gNmC, the only sticky ends created are those resulting from the azidoglucosylated residues, which in turn are exclusively associated to 5-fC and/or 5-caC sites.

The sticky ends are ligated to an adaptor A. The resulting DNA is fragmented and ligated to a second adaptor P1. The azido moiety can derivatized with a biotin label via azide-alkyne Huisgen cycloaddition using a copper(I) catalyst ("click chemistry") or via copper-free variants (for example, using strained cyclooctyne derivatives) and avidin beads can then be used to selectively purify the fragments containing the azidoglucosylated residues.

Example 4

Exclusive 5-caC Mapping

An exemplary process for mapping the locations of 5-caC in a nucleic acid sample is provided in FIG. 6.

As shown in FIG. 6, genomic DNA is treated with BGT and UDP-2-GlcN, so that substantially all 5-hmC residues are converted to 5- β -2-gNmC. The resulting DNA is treated with a reducing agent (e.g., NaBH₄, converts 5-fC to 5-hmC), BGT and UDP-2-GlcN, so that all 5-fC residues are converted to 5- β -2-gNmC. Then, the resulting DNA is treated with a second reducing agent (a different reducing agent or the same reducing agent but in the presence of certain additives converts 5-caC to 5-hmC), BGT, and UDP-Glc to generate 5- β -gmC. The DNA is digested with a restriction endonuclease such as AbaSI, which cleaves 5- β -gmC exclusively associated to 5-caC sites but not 5- β -2-gNmC, 5-mC or C to leave a sticky end (2-base 3'-overhang). These sites are then identified through ligation to a biotinylated adapter, fragmentation and purification using avidin-associated beads, ligation and analysis as described in Example 2.

SEQUENCE LISTING

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Lys Ser Phe Asp Glu Val Asp Val Asn Asp Tyr Asp Arg Leu Ile Val
50 55 60

Val Asn Ser Ser Ile Asn Phe Phe Gly Gly Lys Pro Asn Leu Ala Ile
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Leu Phe Thr Asp Ile Arg Leu Pro Phe Ser Gln Ser Trp Pro Asn Val

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Pro Thr Lys Lys Thr Leu Asp Val Ile Tyr Gly Gly Ser Phe Arg Ser 180 185 190		
Gly Gln Arg Glu Ser Lys Met Val Glu Phe Leu Phe Asp Thr Gly Leu 195 200 205		
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Lys Tyr Pro Trp Thr Lys Ala Pro Val Phe Thr Gly Lys Ile Pro Met 225 230 235 240		
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Ser His Asp His Lys Ser Phe Ser Ile Pro Val Ile Leu Ala Lys Glu 50 55 60
Tyr Asp Lys Ala Leu Lys Leu Val Asn Asp Cys Asp Ile Leu Ile Ile 65 70 75 80
Asn Ser Val Pro Ala Thr Ser Val Gln Glu Ala Thr Ile Asn Asn Tyr 85 90 95
Lys Lys Leu Leu Asp Asn Ile Lys Pro Ser Ile Arg Val Val Val Tyr 100 105 110
Gln His Asp His Ser Val Leu Ser Leu Arg Arg Asn Leu Gly Leu Glu 115 120 125

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 Glu Ile Asn Met Asn Ile Asn Arg Trp Ile Gly Arg Thr Thr Thr Trp
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 Lys Gly Phe Tyr Gln Met Phe Asp Phe His Glu Lys Phe Leu Lys Pro
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 Ala Gly Lys Ser Thr Val Met Glu Gly Leu Glu Arg Ser Pro Ala Phe
 225 230 235 240
 Ile Ala Ile Lys Glu Lys Gly Ile Pro Tyr Glu Tyr Tyr Gly Asn Arg
 245 250 255
 Glu Ile Asp Lys Met Asn Leu Ala Pro Asn Gln Pro Ala Gln Ile Leu
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 340 345 350
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 Pro Gln Leu Gly Ile His Ile Glu Val Asp Glu Gly His His Phe Leu
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 165 170 175

Tyr Ile Asp Leu Gly Arg Ile Ser Leu Ala Asp Asn Val Val Leu Lys
 180 185 190

Thr Thr Lys Asp Val Cys Asn Cys Phe Gly Tyr Ser Tyr Lys Asn Tyr
 195 200 205

Gln Arg Gly Gly Ala Leu His Pro Tyr Lys Lys Asp Thr Leu Ile Trp
 210 215 220

Phe Pro Arg Leu Tyr Glu Asn Lys Asp Trp Ile Asn Thr Ile Ser Pro
 225 230 235 240

Asp Gly Leu Thr Ile Thr Glu Lys Ser Thr Asp Glu Thr Ile Thr Leu
 245 250 255

Lys Lys Leu Glu Glu Trp Lys Asn Gly Pro Gln Lys Arg Ile Val Phe
 260 265 270

Ala Arg Val Lys Asp Asn Leu Ser Ser Arg Ala Met Tyr Arg Phe Met
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Cys

<210> SEQ ID NO 4
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 <212> TYPE: PRT
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Leu Leu Asn Asp Phe Thr Leu Lys Phe Val Thr Gln Gln Phe Val Arg
 35 40 45

Leu Ser Asn Lys Lys Ile Ala Leu Thr Asp Leu Tyr Phe Pro Gln Leu
 50 55 60

Gly Ile His Ile Glu Val Asp Glu Glu His His Phe Leu Arg Asn Ser
 65 70 75 80

Lys Met Glu Tyr Ser Leu Asn Gln Ile Asp Glu Pro Leu Tyr Ser Ile
 85 90 95

Ser His Thr Glu Ser Asp Ala Met Arg Glu Glu Asp Ile Ile Ser Ile
 100 105 110

Thr Gly His Lys Ile Phe Arg Val Asn Val Phe Lys Asn Gln Glu Gly
 115 120 125

Gln Pro Gln Asn Leu Glu Ser Ile His Gln Gln Ile Asp Lys Ile Ile
 130 135 140

Glu Lys Ile Lys Thr Ala Lys Asn Lys Leu Ile Glu Ala Ser Thr Phe
 145 150 155 160

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Lys Glu Trp Asn Ile Glu Thr Glu Tyr Asn Pro Gln Thr Tyr Ile Asp
 165 170 175

Leu Gly Arg Ile Ser Leu Ala Asp Asn Val Val Leu Lys Thr Thr Lys
 180 185 190

Asp Val Cys Asn Cys Phe Gly Tyr Asn Tyr Lys Asn Tyr Gln Arg Gly
 195 200 205

Gly Ala Leu His Pro Tyr Glu Lys Asp Thr Leu Ile Trp Phe Pro Arg
 210 215 220

Leu Tyr Glu Asn Lys Asp Trp Phe Asn Thr Ile Ser Pro Asp Gly Leu
 225 230 235 240

Thr Ile Thr Glu Lys Ser Thr Asp Glu Ala Ile Thr Leu Lys Lys Leu
 245 250 255

Glu Glu Trp Lys Asn Gly Pro Gln Lys Arg Ile Val Phe Ala Arg Val
 260 265 270

Lys Asp Asn Leu Ser Ser Arg Ala Met Tyr Arg Phe Met Gly Leu Tyr
 275 280 285

Glu Phe Gln Lys Ala Asp Leu Lys Asp Gly Ala Val Trp Lys Arg Val
 290 295 300

Glu Cys Glu Val Gln Thr Tyr Ser Pro Lys Glu Thr Lys Cys
 305 310 315

<210> SEQ ID NO 5
 <211> LENGTH: 318
 <212> TYPE: PRT
 <213> ORGANISM: Acinetobacter baumannii

<400> SEQUENCE: 5

Met Phe Ser Ser Asp Leu Thr Asp Tyr Val Ile Arg Gln Leu Gly Arg
 1 5 10 15

Thr Lys Asn Lys Arg Tyr Glu Ala Tyr Val Val Ser Arg Ile Ile His
 20 25 30

Leu Leu Asn Asp Phe Thr Leu Lys Phe Val Thr Gln Gln Phe Val Arg
 35 40 45

Leu Ser Asn Lys Lys Ile Ala Leu Thr Asp Leu Tyr Phe Pro Gln Leu
 50 55 60

Gly Ile His Ile Glu Val Asp Glu Gly His His Phe Leu Arg Asn Ser
 65 70 75 80

Lys Met Glu Tyr Ser Leu Asn Gln Ile Asp Glu Pro Leu Tyr Ser Ile
 85 90 95

Ser Gln Thr Glu Ser Asp Ala Met Arg Glu Glu Asp Ile Ile Ser Ile
 100 105 110

Thr Gly His Lys Ile Phe Arg Val Asn Val Phe Lys Asn Gln Glu Gly
 115 120 125

Gln Pro Gln Asn Leu Glu Ser Ile His Gln Gln Ile Asp Lys Ile Ile
 130 135 140

Glu Glu Ile Lys Thr Ala Lys Asn Lys Leu Ile Glu Ala Ser Thr Phe
 145 150 155 160

Lys Glu Trp Asn Ile Glu Thr Glu Tyr Asn Pro Gln Thr Tyr Ile Asp
 165 170 175

Leu Gly Arg Ile Ser Leu Ala Asp Asn Val Val Leu Lys Thr Thr Lys
 180 185 190

Asp Val Cys Asn Cys Phe Gly Tyr Asn Tyr Lys Asn Tyr Gln Arg Gly
 195 200 205

Gly Ala Leu His Pro Tyr Glu Lys Asp Thr Leu Ile Trp Phe Pro Arg

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Lys Asp Asn Leu Ser Ser Arg Ala Met Tyr Arg Phe Met Gly Leu Tyr
 275 280 285

Glu Phe Gln Lys Ala Asp Leu Lys Asp Gly Ala Val Trp Lys Arg Val
 290 295 300

Glu Cys Glu Val Gln Thr Tyr Ser Pro Lys Glu Thr Lys Cys
 305 310 315

<210> SEQ ID NO 7
 <211> LENGTH: 268
 <212> TYPE: PRT
 <213> ORGANISM: Acinetobacter baumannii

<400> SEQUENCE: 7

Ile Cys Arg Val Gln Arg Thr Asp Leu Tyr Phe Pro Gln Leu Gly Ile
 1 5 10 15

His Ile Glu Val Asp Glu Gly His His Phe Leu Arg Asn Ser Lys Met
 20 25 30

Glu Tyr Ser Leu Asn Gln Ile Asp Glu Pro Leu Tyr Ser Ile Ser Gln
 35 40 45

Thr Glu Ser Asp Ala Met Arg Glu Glu Asp Ile Ile Ser Ile Thr Gly
 50 55 60

His Lys Ile Phe Arg Val Asn Val Tyr Lys Asn Gln Glu Gly Gln Pro
 65 70 75 80

Gln Asn Leu Glu Ser Ile His Gln Gln Ile Asp Lys Ile Ile Glu Glu
 85 90 95

Ile Lys Thr Ala Lys Asn Lys Leu Ile Glu Ala Ser Thr Phe Lys Glu
 100 105 110

Trp Asn Ile Glu Thr Glu Tyr Asn Pro Gln Thr Tyr Ile Asn Leu Gly
 115 120 125

Arg Ile Ser Leu Ala Asp Asn Val Val Leu Lys Thr Thr Lys Asp Val
 130 135 140

Cys Asn Cys Phe Gly Tyr Asn Tyr Lys Asn Tyr Gln Arg Gly Gly Ala
 145 150 155 160

Ile His Pro Tyr Glu Glu Asp Thr Leu Ile Trp Phe Pro Arg Leu Tyr
 165 170 175

Glu Asn Lys Asp Trp Ile Asn Thr Ile Ser Pro Asp Gly Leu Thr Ile
 180 185 190

Thr Glu Lys Ser Thr Asp Glu Thr Ile Thr Leu Lys Lys Leu Glu Glu
 195 200 205

Trp Lys Asn Gly Pro Gln Lys Arg Ile Val Phe Ala Arg Val Lys Asp
 210 215 220

Asn Leu Asn Ser Arg Ala Met Tyr Arg Phe Met Gly Leu Tyr Lys Phe
 225 230 235 240

Gln Lys Ala Asp Leu Lys Asp Gly Ala Val Trp Lys Arg Val Glu Cys
 245 250 255

Glu Val Gln Thr Tyr Ser Pro Lys Glu Thr Lys Cys
 260 265

<210> SEQ ID NO 8
 <211> LENGTH: 318
 <212> TYPE: PRT
 <213> ORGANISM: Acinetobacter baumannii

<400> SEQUENCE: 8

Met Phe Ser Ser Asp Leu Thr Asp Tyr Val Ile Arg Gln Leu Gly Arg
 1 5 10 15

-continued

Thr Lys Asn Lys Arg Tyr Glu Ala Tyr Val Val Ser Arg Ile Ile His
 20 25 30
 Leu Leu Asn Asp Ile Thr Leu Lys Phe Val Thr Gln Gln Phe Val Arg
 35 40 45
 Leu Ser Asn Lys Lys Ile Ala Leu Thr Asp Leu Tyr Phe Pro Gln Leu
 50 55 60
 Gly Ile His Ile Glu Val Asp Glu Gly His His Phe Leu Arg Asn Ser
 65 70 75 80
 Lys Met Glu Tyr Ser Leu Asn Gln Ile Asp Glu Pro Leu Tyr Ser Ile
 85 90 95
 Ser Gln Thr Glu Ser Asp Ala Met Arg Glu Glu Asp Ile Ile Ser Ile
 100 105 110
 Thr Glu His Lys Ile Phe Arg Val Asn Val Tyr Lys Asn Gln Glu Gly
 115 120 125
 Gln Pro Gln Asn Leu Glu Ser Ile His Gln Gln Ile Asp Lys Ile Ile
 130 135 140
 Glu Glu Ile Lys Thr Ala Lys Asn Lys Leu Val Glu Glu Phe Lys Phe
 145 150 155 160
 Lys Glu Trp Asn Ile Glu Thr Glu Tyr Asn Pro Gln Thr Tyr Ile Asp
 165 170 175
 Leu Gly Arg Ile Ser Leu Ala Asp Asn Val Val Leu Lys Thr Thr Lys
 180 185 190
 Asp Val Cys Asn Cys Phe Gly Tyr Asn Tyr Lys Asn Tyr Gln Arg Gly
 195 200 205
 Gly Ala Leu His Pro Tyr Glu Lys Asp Thr Leu Ile Trp Phe Pro Arg
 210 215 220
 Leu Tyr Glu Asn Lys Asp Trp Ile Asn Thr Ile Ser Pro Asp Gly Leu
 225 230 235 240
 Thr Ile Thr Glu Lys Ser Thr Asp Glu Thr Ile Thr Leu Lys Lys Leu
 245 250 255
 Glu Glu Trp Lys Asn Gly Pro Gln Lys Arg Ile Val Phe Ala Arg Val
 260 265 270
 Lys Asp Asn Leu Ser Ser Arg Ala Met Tyr Arg Phe Met Gly Leu Tyr
 275 280 285
 Glu Phe Gln Lys Ala Asp Leu Lys Asp Gly Ala Val Trp Lys Arg Val
 290 295 300
 Lys Cys Glu Val Gln Thr Tyr Ser Pro Lys Glu Thr Lys Cys
 305 310 315

<210> SEQ ID NO 9

<211> LENGTH: 318

<212> TYPE: PRT

<213> ORGANISM: Acinetobacter calcoaceticus

<400> SEQUENCE: 9

Met Phe Ser Ser Asp Leu Thr Asp Tyr Val Ile Arg Gln Leu Gly Arg
 1 5 10 15
 Thr Lys Asn Lys Arg Tyr Glu Ala Tyr Val Val Ser Arg Ile Ile His
 20 25 30
 Leu Leu Asn Asp Phe Thr Leu Lys Phe Val Thr Gln Gln Phe Val Arg
 35 40 45
 Leu Ser Asn Lys Lys Ile Ala Leu Thr Asp Leu Tyr Phe Pro Gln Leu
 50 55 60
 Asp Ile His Ile Glu Val Asp Glu Gly His His Phe Leu Arg Asn Ser
 65 70 75 80

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Lys Met Glu Tyr Ser Leu Asn Gln Ile Asp Glu Pro Leu Tyr Ser Ile
 85 90 95
 Ser Gln Thr Glu Ser Asp Ala Met Arg Glu Glu Asp Ile Ile Ser Ile
 100 105 110
 Thr Gly His Lys Ile Phe Arg Val Asn Val Tyr Lys Asn Gln Glu Gly
 115 120 125
 Glu Pro Gln Asn Leu Glu Ser Ile His Gln Gln Ile Asp Lys Ile Ile
 130 135 140
 Glu Glu Ile Lys Val Ala Lys Asn Lys Gln Ile Lys Ala Ser Thr Phe
 145 150 155 160
 Lys Glu Trp Asn Ile Glu Thr Glu Tyr Asn Pro Gln Thr Tyr Ile Asp
 165 170 175
 Leu Gly Ser Ile Ser Leu Ala Asp Asn Val Val Leu Lys Thr Thr Lys
 180 185 190
 Asp Val Cys Asn Cys Phe Gly Tyr Asn Tyr Lys Asn Tyr Gln Arg Gly
 195 200 205
 Gly Ala Ile His Pro Tyr Glu Lys Asp Thr Leu Ile Trp Phe Pro Arg
 210 215 220
 Leu Tyr Glu Asn Lys Asp Trp Ile Asn Thr Ile Ser Pro Asp Gly Leu
 225 230 235 240
 Thr Ile Thr Glu Lys Ser Thr Asp Glu Ala Ile Thr Leu Lys Lys Leu
 245 250 255
 Glu Glu Trp Lys Asn Gly Pro Gln Lys Arg Ile Val Phe Ala Arg Val
 260 265 270
 Lys Asp Asn Leu Ser Ser Arg Ala Met Tyr Arg Phe Met Gly Leu Tyr
 275 280 285
 Glu Phe Gln Lys Ala Asp Leu Lys Asp Gly Ala Val Trp Lys Arg Glu
 290 295 300
 Gly Cys Lys Val Gln Thr Tyr Ser Pro Lys Glu Ala Lys Cys
 305 310 315

<210> SEQ ID NO 10

<211> LENGTH: 296

<212> TYPE: PRT

<213> ORGANISM: Paraprevotella xylaniphila

<400> SEQUENCE: 10

Val Lys Tyr Gly Thr Asn Ser Lys Phe Lys Asp Met Asp Tyr Lys Leu
 1 5 10 15
 Asp Tyr Met Glu Arg Leu Phe Ala Lys Ile Ser Lys Lys Lys Thr Glu
 20 25 30
 Ser Tyr Val Ile Ser Arg Ile Trp His Gln Leu Asp Asp Asp Arg Val
 35 40 45
 Lys Phe Val Val Gln Gln Tyr Ile Arg Arg Thr Gln Asp Lys Tyr Ala
 50 55 60
 Leu Ala Asp Leu Tyr Leu Pro Gln Leu Asn Ile Phe Ile Glu Ile Asn
 65 70 75 80
 Glu Pro Phe His Lys Asn Asn Thr Glu Ile Asp Lys Ile Arg Asn Glu
 85 90 95
 Glu Ile Leu Asn Ile Thr Asn Ser Lys Pro Ile Ile Ile Asp Cys Asp
 100 105 110
 Asn Asn Ile Gln Glu Ile His His Gln Ile Thr Asp Val Val Ser Leu
 115 120 125
 Ile Lys Gln Cys Ile Gln Glu Met Gly Asp Asn Phe Gln Pro Trp Asp

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130				135				140							
Asp	Val	Ser	Thr	Leu	Ser	Val	Glu	Tyr	His	Arg	Asn	Lys	Gly	Tyr	Leu
145				150						155					160
Lys	Val	Asp	Asp	Asn	Glu	Cys	Leu	Arg	Thr	Thr	Asp	Asp	Val	Ala	Glu
				165					170					175	
Thr	Phe	Gly	Thr	Lys	Pro	Lys	His	Arg	Gly	Phe	Leu	Arg	Ala	Ser	Gly
			180						185				190		
Ala	Ala	Val	Pro	Asn	Lys	Lys	Asn	Glu	Ile	Ile	Trp	Trp	Pro	Asn	Thr
		195					200						205		
Glu	His	Arg	Leu	Trp	Cys	Asn	Glu	Leu	Ser	Glu	Asp	Gly	Met	Phe	Ile
	210					215					220				
Tyr	Glu	Tyr	Pro	Lys	Ala	Glu	Asn	Lys	Arg	Thr	Ala	His	Leu	Lys	Gln
225					230					235					240
Trp	Leu	Ser	Ala	Pro	Glu	Glu	Thr	Arg	Ile	Thr	Phe	Leu	Arg	Tyr	Lys
				245					250					255	
Asp	Asp	Leu	Gly	Phe	Cys	Phe	Tyr	Arg	Phe	Val	Gly	Val	Phe	Asn	Leu
			260						265				270		
Asn	Lys	Glu	Lys	Ser	Ile	Lys	Glu	Asn	Lys	Cys	Val	Trp	Glu	Arg	Val
		275					280					285			
Ser	Asn	Thr	Tyr	Gln	Leu	Asn	Val								
	290					295									

<210> SEQ ID NO 11

<211> LENGTH: 321

<212> TYPE: PRT

<213> ORGANISM: Naegleria gruberi

<400> SEQUENCE: 11

Met	Thr	Thr	Phe	Lys	Gln	Gln	Thr	Ile	Lys	Glu	Lys	Glu	Thr	Lys	Arg
1				5					10					15	
Lys	Tyr	Cys	Ile	Lys	Gly	Thr	Thr	Ala	Asn	Leu	Thr	Gln	Thr	His	Pro
			20					25					30		
Asn	Gly	Pro	Val	Cys	Val	Asn	Arg	Gly	Glu	Glu	Val	Ala	Asn	Thr	Thr
		35					40					45			
Thr	Leu	Leu	Asp	Ser	Gly	Gly	Gly	Ile	Asn	Lys	Lys	Ser	Leu	Leu	Gln
	50					55					60				
Asn	Leu	Leu	Ser	Lys	Cys	Lys	Thr	Thr	Phe	Gln	Gln	Ser	Phe	Thr	Asn
65					70				75					80	
Ala	Asn	Ile	Thr	Leu	Lys	Asp	Glu	Lys	Trp	Leu	Lys	Asn	Val	Arg	Thr
				85					90					95	
Ala	Tyr	Phe	Val	Cys	Asp	His	Asp	Gly	Ser	Val	Glu	Leu	Ala	Tyr	Leu
			100						105				110		
Pro	Asn	Val	Leu	Pro	Lys	Glu	Leu	Val	Glu	Glu	Phe	Thr	Glu	Lys	Phe
		115					120					125			
Glu	Ser	Ile	Gln	Thr	Gly	Arg	Lys	Lys	Asp	Thr	Gly	Tyr	Ser	Gly	Ile
	130					135					140				
Leu	Asp	Asn	Ser	Met	Pro	Phe	Asn	Tyr	Val	Thr	Ala	Asp	Leu	Ser	Gln
145					150					155					160
Glu	Leu	Gly	Gln	Tyr	Leu	Ser	Glu	Ile	Val	Asn	Pro	Gln	Ile	Asn	Tyr
			165						170					175	
Tyr	Ile	Ser	Lys	Leu	Leu	Thr	Cys	Val	Ser	Ser	Arg	Thr	Ile	Asn	Tyr
			180						185				190		
Leu	Val	Ser	Leu	Asn	Asp	Ser	Tyr	Tyr	Ala	Leu	Asn	Asn	Cys	Leu	Tyr
		195					200				205				

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Pro Ser Thr Ala Phe Asn Ser Leu Lys Pro Ser Asn Asp Gly His Arg
 210                215                220

Ile Arg Lys Pro His Lys Asp Asn Leu Asp Ile Thr Pro Ser Ser Leu
 225                230                235                240

Phe Tyr Phe Gly Asn Phe Gln Asn Thr Glu Gly Tyr Leu Glu Leu Thr
                245                250                255

Asp Lys Asn Cys Lys Val Phe Val Gln Pro Gly Asp Val Leu Phe Phe
                260                265                270

Lys Gly Asn Glu Tyr Lys His Val Val Ala Asn Ile Thr Ser Gly Trp
                275                280                285

Arg Ile Gly Leu Val Tyr Phe Ala His Lys Gly Ser Lys Thr Lys Pro
 290                295                300

Tyr Tyr Glu Asp Thr Gln Lys Asn Ser Leu Lys Ile His Lys Glu Thr
 305                310                315                320

Lys

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<210> SEQ ID NO 12
<211> LENGTH: 2039
<212> TYPE: PRT
<213> ORGANISM: Mus musculus

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<400> SEQUENCE: 12

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Met Ser Arg Ser Arg Pro Ala Lys Pro Ser Lys Ser Val Lys Thr Lys
 1                5                10                15

Leu Gln Lys Lys Lys Asp Ile Gln Met Lys Thr Lys Thr Ser Lys Gln
                20                25                30

Ala Val Arg His Gly Ala Ser Ala Lys Ala Val Asn Pro Gly Lys Pro
 35                40                45

Lys Gln Leu Ile Lys Arg Arg Asp Gly Lys Lys Glu Thr Glu Asp Lys
 50                55                60

Thr Pro Thr Pro Ala Pro Ser Phe Leu Thr Arg Ala Gly Ala Ala Arg
 65                70                75                80

Met Asn Arg Asp Arg Asn Gln Val Leu Phe Gln Asn Pro Asp Ser Leu
                85                90                95

Thr Cys Asn Gly Phe Thr Met Ala Leu Arg Arg Thr Ser Leu Ser Trp
                100                105                110

Arg Leu Ser Gln Arg Pro Val Val Thr Pro Lys Pro Lys Lys Val Pro
 115                120                125

Pro Ser Lys Lys Gln Cys Thr His Asn Ile Gln Asp Glu Pro Gly Val
 130                135                140

Lys His Ser Glu Asn Asp Ser Val Pro Ser Gln His Ala Thr Val Ser
 145                150                155                160

Pro Gly Thr Glu Asn Gly Glu Gln Asn Arg Cys Leu Val Glu Gly Glu
                165                170                175

Ser Gln Glu Ile Thr Gln Ser Cys Pro Val Phe Glu Glu Arg Ile Glu
                180                185                190

Asp Thr Gln Ser Cys Ile Ser Ala Ser Gly Asn Leu Glu Ala Glu Ile
 195                200                205

Ser Trp Pro Leu Glu Gly Thr His Cys Glu Glu Leu Leu Ser His Gln
 210                215                220

Thr Ser Asp Asn Glu Cys Thr Ser Pro Gln Glu Cys Ala Pro Leu Pro
 225                230                235                240

Gln Arg Ser Thr Ser Glu Val Thr Ser Gln Lys Asn Thr Ser Asn Gln
                245                250                255

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675					680					685					
Ile	Glu	Val	Glu	Lys	Trp	Ala	Pro	Asn	Lys	Lys	Ser	His	Leu	Ala	Glu
690					695					700					
Gly	Gln	Val	Lys	Gly	Ser	Cys	Asp	Ala	Asn	Leu	Thr	Gly	Val	Glu	Asn
705					710					715					720
Pro	Gln	Pro	Ser	Glu	Asp	Asp	Lys	Gln	Gln	Thr	Asn	Pro	Ser	Pro	Thr
				725					730					735	
Phe	Ala	Gln	Thr	Ile	Arg	Asn	Gly	Met	Lys	Asn	Val	His	Cys	Leu	Pro
			740					745					750		
Thr	Asp	Thr	His	Leu	Pro	Leu	Asn	Lys	Leu	Asn	His	Glu	Glu	Phe	Ser
			755				760					765			
Lys	Ala	Leu	Gly	Asn	Asn	Ser	Ser	Lys	Leu	Leu	Thr	Asp	Pro	Ser	Asn
770					775					780					
Cys	Lys	Asp	Ala	Met	Ser	Val	Thr	Thr	Ser	Gly	Gly	Glu	Cys	Asp	His
785					790					795					800
Leu	Lys	Gly	Pro	Arg	Asn	Thr	Leu	Leu	Phe	Gln	Lys	Pro	Gly	Leu	Asn
				805					810					815	
Cys	Arg	Ser	Gly	Ala	Glu	Pro	Thr	Ile	Phe	Asn	Asn	His	Pro	Asn	Thr
			820					825					830		
His	Ser	Ala	Gly	Ser	Arg	Pro	His	Pro	Pro	Glu	Lys	Val	Pro	Asn	Lys
		835					840					845			
Glu	Pro	Lys	Asp	Gly	Ser	Pro	Val	Gln	Pro	Ser	Leu	Leu	Ser	Leu	Met
850					855					860					
Lys	Asp	Arg	Arg	Leu	Thr	Leu	Glu	Gln	Val	Val	Ala	Ile	Glu	Ala	Leu
865					870					875					880
Thr	Gln	Leu	Ser	Glu	Ala	Pro	Ser	Glu	Ser	Ser	Ser	Pro	Ser	Lys	Pro
				885					890					895	
Glu	Lys	Asp	Glu	Glu	Ala	His	Gln	Lys	Thr	Ala	Ser	Leu	Leu	Asn	Ser
			900					905					910		
Cys	Lys	Ala	Ile	Leu	His	Ser	Val	Arg	Lys	Asp	Leu	Gln	Asp	Pro	Asn
		915					920					925			
Val	Gln	Gly	Lys	Gly	Leu	His	His	Asp	Thr	Val	Val	Phe	Asn	Gly	Gln
930					935					940					
Asn	Arg	Thr	Phe	Lys	Ser	Pro	Asp	Ser	Phe	Ala	Thr	Asn	Gln	Ala	Leu
945					950					955					960
Ile	Lys	Ser	Gln	Gly	Tyr	Pro	Ser	Ser	Pro	Thr	Ala	Glu	Lys	Lys	Gly
				965					970					975	
Ala	Ala	Gly	Gly	Arg	Ala	Pro	Phe	Asp	Gly	Phe	Glu	Asn	Ser	His	Pro
			980					985					990		
Leu	Pro	Ile	Glu	Ser	His	Asn	Leu	Glu	Asn	Cys	Ser	Gln	Val	Leu	Ser
		995					1000					1005			
Cys	Asp	Gln	Asn	Leu	Ser	Ser	His	Asp	Pro	Ser	Cys	Gln	Asp	Ala	
1010					1015					1020					
Pro	Tyr	Ser	Gln	Ile	Glu	Glu	Asp	Val	Ala	Ala	Gln	Leu	Thr	Gln	
1025					1030					1035					
Leu	Ala	Ser	Thr	Ile	Asn	His	Ile	Asn	Ala	Glu	Val	Arg	Asn	Ala	
1040					1045					1050					
Glu	Ser	Thr	Pro	Glu	Ser	Leu	Val	Ala	Lys	Asn	Thr	Lys	Gln	Lys	
1055					1060					1065					
His	Ser	Gln	Glu	Lys	Arg	Met	Val	His	Gln	Lys	Pro	Pro	Ser	Ser	
1070					1075					1080					
Thr	Gln	Thr	Lys	Pro	Ser	Val	Pro	Ser	Ala	Lys	Pro	Lys	Lys	Ala	
1085					1090					1095					

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Leu	Arg	Ser	Tyr	Ser	Gly	His	Pro	Thr	Asp	Arg	Arg	Cys	Thr	Leu
1490						1495					1500			
Asn	Lys	Lys	Arg	Thr	Cys	Thr	Cys	Gln	Gly	Ile	Asp	Pro	Lys	Thr
1505						1510					1515			
Cys	Gly	Ala	Ser	Phe	Ser	Phe	Gly	Cys	Ser	Trp	Ser	Met	Tyr	Phe
1520						1525					1530			
Asn	Gly	Cys	Lys	Phe	Gly	Arg	Ser	Glu	Asn	Pro	Arg	Lys	Phe	Arg
1535						1540					1545			
Leu	Ala	Pro	Asn	Tyr	Pro	Leu	His	Asn	Tyr	Tyr	Lys	Arg	Ile	Thr
1550						1555					1560			
Gly	Met	Ser	Ser	Glu	Gly	Ser	Asp	Val	Lys	Thr	Gly	Trp	Ile	Ile
1565						1570					1575			
Pro	Asp	Arg	Lys	Thr	Leu	Ile	Ser	Arg	Glu	Glu	Lys	Gln	Leu	Glu
1580						1585					1590			
Lys	Asn	Leu	Gln	Glu	Leu	Ala	Thr	Val	Leu	Ala	Pro	Leu	Tyr	Lys
1595						1600					1605			
Gln	Met	Ala	Pro	Val	Ala	Tyr	Gln	Asn	Gln	Val	Glu	Tyr	Glu	Glu
1610						1615					1620			
Val	Ala	Gly	Asp	Cys	Arg	Leu	Gly	Asn	Glu	Glu	Gly	Arg	Pro	Phe
1625						1630					1635			
Ser	Gly	Val	Thr	Cys	Cys	Met	Asp	Phe	Cys	Ala	His	Ser	His	Lys
1640						1645					1650			
Asp	Ile	His	Asn	Met	His	Asn	Gly	Ser	Thr	Val	Val	Cys	Thr	Leu
1655						1660					1665			
Ile	Arg	Ala	Asp	Gly	Arg	Asp	Thr	Asn	Cys	Pro	Glu	Asp	Glu	Gln
1670						1675					1680			
Leu	His	Val	Leu	Pro	Leu	Tyr	Arg	Leu	Ala	Asp	Thr	Asp	Glu	Phe
1685						1690					1695			
Gly	Ser	Val	Glu	Gly	Met	Lys	Ala	Lys	Ile	Lys	Ser	Gly	Ala	Ile
1700						1705					1710			
Gln	Val	Asn	Gly	Pro	Thr	Arg	Lys	Arg	Arg	Leu	Arg	Phe	Thr	Glu
1715						1720					1725			
Pro	Val	Pro	Arg	Cys	Gly	Lys	Arg	Ala	Lys	Met	Lys	Gln	Asn	His
1730						1735					1740			
Asn	Lys	Ser	Gly	Ser	His	Asn	Thr	Lys	Ser	Phe	Ser	Ser	Ala	Ser
1745						1750					1755			
Ser	Thr	Ser	His	Leu	Val	Lys	Asp	Glu	Ser	Thr	Asp	Phe	Cys	Pro
1760						1765					1770			
Leu	Gln	Ala	Ser	Ser	Ala	Glu	Thr	Ser	Thr	Cys	Thr	Tyr	Ser	Lys
1775						1780					1785			
Thr	Ala	Ser	Gly	Gly	Phe	Ala	Glu	Thr	Ser	Ser	Ile	Leu	His	Cys
1790						1795					1800			
Thr	Met	Pro	Ser	Gly	Ala	His	Ser	Gly	Ala	Asn	Ala	Ala	Ala	Gly
1805						1810					1815			
Glu	Cys	Thr	Gly	Thr	Val	Gln	Pro	Ala	Glu	Val	Ala	Ala	His	Pro
1820						1825					1830			
His	Gln	Ser	Leu	Pro	Thr	Ala	Asp	Ser	Pro	Val	His	Ala	Glu	Pro
1835						1840					1845			
Leu	Thr	Ser	Pro	Ser	Glu	Gln	Leu	Thr	Ser	Asn	Gln	Ser	Asn	Gln
1850						1855					1860			
Gln	Leu	Pro	Leu	Leu	Ser	Asn	Ser	Gln	Lys	Leu	Ala	Ser	Cys	Gln
1865						1870					1875			
Val	Glu	Asp	Glu	Arg	His	Pro	Glu	Ala	Asp	Glu	Pro	Gln	His	Pro

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1880	1885	1890
Glu Asp Asp Asn Leu Pro Gln Leu Asp Glu Phe Trp Ser Asp Ser 1895 1900 1905		
Glu Glu Ile Tyr Ala Asp Pro Ser Phe Gly Gly Val Ala Ile Ala 1910 1915 1920		
Pro Ile His Gly Ser Val Leu Ile Glu Cys Ala Arg Lys Glu Leu 1925 1930 1935		
His Ala Thr Thr Ser Leu Arg Ser Pro Lys Arg Gly Val Pro Phe 1940 1945 1950		
Arg Val Ser Leu Val Phe Tyr Gln His Lys Ser Leu Asn Lys Pro 1955 1960 1965		
Asn His Gly Phe Asp Ile Asn Lys Ile Lys Cys Lys Cys Lys Lys 1970 1975 1980		
Val Thr Lys Lys Lys Pro Ala Asp Arg Glu Cys Pro Asp Val Ser 1985 1990 1995		
Pro Glu Ala Asn Leu Ser His Gln Ile Pro Ser Arg Val Ala Ser 2000 2005 2010		
Thr Leu Thr Arg Asp Asn Val Val Thr Val Ser Pro Tyr Ser Leu 2015 2020 2025		
Thr His Val Ala Gly Pro Tyr Asn Arg Trp Val 2030 2035		

<210> SEQ ID NO 13

<211> LENGTH: 1912

<212> TYPE: PRT

<213> ORGANISM: Mus musculus

<400> SEQUENCE: 13

Met Glu Gln Asp Arg Thr Thr His Ala Glu Gly Thr Arg Leu Ser Pro 1 5 10 15
Phe Leu Ile Ala Pro Pro Ser Pro Ile Ser His Thr Glu Pro Leu Ala 20 25 30
Val Lys Leu Gln Asn Gly Ser Pro Leu Ala Glu Arg Pro His Pro Glu 35 40 45
Val Asn Gly Asp Thr Lys Trp Gln Ser Ser Gln Ser Cys Tyr Gly Ile 50 55 60
Ser His Met Lys Gly Ser Gln Ser Ser His Glu Ser Pro His Glu Asp 65 70 75 80
Arg Gly Tyr Ser Arg Cys Leu Gln Asn Gly Gly Ile Lys Arg Thr Val 85 90 95
Ser Glu Pro Ser Leu Ser Gly Leu His Pro Asn Lys Ile Leu Lys Leu 100 105 110
Asp Gln Lys Ala Lys Gly Glu Ser Asn Ile Phe Glu Glu Ser Gln Glu 115 120 125
Arg Asn His Gly Lys Ser Ser Arg Gln Pro Asn Val Ser Gly Leu Ser 130 135 140
Asp Asn Gly Glu Pro Val Thr Ser Thr Thr Gln Glu Ser Ser Gly Ala 145 150 155 160
Asp Ala Phe Pro Thr Arg Asn Tyr Asn Gly Val Glu Ile Gln Val Leu 165 170 175
Asn Glu Gln Glu Gly Glu Lys Gly Arg Ser Val Thr Leu Leu Lys Asn 180 185 190
Lys Ile Val Leu Met Pro Asn Gly Ala Thr Val Ser Ala His Ser Glu 195 200 205

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Glu	Asn	Thr	Arg	Gly	Glu	Leu	Leu	Glu	Lys	Thr	Gln	Cys	Tyr	Pro	Asp
210						215					220				
Cys	Val	Ser	Ile	Ala	Val	Gln	Ser	Thr	Ala	Ser	His	Val	Asn	Thr	Pro
225					230					235					240
Ser	Ser	Gln	Ala	Ala	Ile	Glu	Leu	Ser	His	Glu	Ile	Pro	Gln	Pro	Ser
			245						250					255	
Leu	Thr	Ser	Ala	Gln	Ile	Asn	Phe	Ser	Gln	Thr	Ser	Ser	Leu	Gln	Leu
			260					265					270		
Pro	Pro	Glu	Pro	Ala	Ala	Met	Val	Thr	Lys	Ala	Cys	Asp	Ala	Asp	Asn
		275					280					285			
Ala	Ser	Lys	Pro	Ala	Ile	Val	Pro	Gly	Thr	Cys	Pro	Phe	Gln	Lys	Ala
290						295					300				
Glu	His	Gln	Gln	Lys	Ser	Ala	Leu	Asp	Ile	Gly	Pro	Ser	Arg	Ala	Glu
305					310					315					320
Asn	Lys	Thr	Ile	Gln	Gly	Ser	Met	Glu	Leu	Phe	Ala	Glu	Glu	Tyr	Tyr
				325					330					335	
Pro	Ser	Ser	Asp	Arg	Asn	Leu	Gln	Ala	Ser	His	Gly	Ser	Ser	Glu	Gln
			340					345						350	
Tyr	Ser	Lys	Gln	Lys	Glu	Thr	Asn	Gly	Ala	Tyr	Phe	Arg	Gln	Ser	Ser
		355					360					365			
Lys	Phe	Pro	Lys	Asp	Ser	Ile	Ser	Pro	Thr	Thr	Val	Thr	Pro	Pro	Ser
	370					375					380				
Gln	Ser	Leu	Leu	Ala	Pro	Arg	Leu	Val	Leu	Gln	Pro	Pro	Leu	Glu	Gly
385					390					395					400
Lys	Gly	Ala	Leu	Asn	Asp	Val	Ala	Leu	Glu	Glu	His	His	Asp	Tyr	Pro
				405					410					415	
Asn	Arg	Ser	Asn	Arg	Thr	Leu	Leu	Arg	Glu	Gly	Lys	Ile	Asp	His	Gln
			420					425					430		
Pro	Lys	Thr	Ser	Ser	Ser	Gln	Ser	Leu	Asn	Pro	Ser	Val	His	Thr	Pro
		435					440					445			
Asn	Pro	Pro	Leu	Met	Leu	Pro	Glu	Gln	His	Gln	Asn	Asp	Cys	Gly	Ser
	450					455					460				
Pro	Ser	Pro	Glu	Lys	Ser	Arg	Lys	Met	Ser	Glu	Tyr	Leu	Met	Tyr	Tyr
465					470					475					480
Leu	Pro	Asn	His	Gly	His	Ser	Gly	Gly	Leu	Gln	Glu	His	Ser	Gln	Tyr
				485					490					495	
Leu	Met	Gly	His	Arg	Glu	Gln	Glu	Ile	Pro	Lys	Asp	Ala	Asn	Gly	Lys
			500					505						510	
Gln	Thr	Gln	Gly	Ser	Val	Gln	Ala	Ala	Pro	Gly	Trp	Ile	Glu	Leu	Lys
		515					520					525			
Ala	Pro	Asn	Leu	His	Glu	Ala	Leu	His	Gln	Thr	Lys	Arg	Lys	Asp	Ile
	530					535					540				
Ser	Leu	His	Ser	Val	Leu	His	Ser	Gln	Thr	Gly	Pro	Val	Asn	Gln	Met
545					550					555					560
Ser	Ser	Lys	Gln	Ser	Thr	Gly	Asn	Val	Asn	Met	Pro	Gly	Gly	Phe	Gln
			565						570					575	
Arg	Leu	Pro	Tyr	Leu	Gln	Lys	Thr	Ala	Gln	Pro	Glu	Gln	Lys	Ala	Gln
			580					585						590	
Met	Tyr	Gln	Val	Gln	Val	Asn	Gln	Gly	Pro	Ser	Pro	Gly	Met	Gly	Asp
		595					600					605			
Gln	His	Leu	Gln	Phe	Gln	Lys	Ala	Leu	Tyr	Gln	Glu	Cys	Ile	Pro	Arg
	610					615					620				
Thr	Asp	Pro	Ser	Ser	Glu	Ala	His	Pro	Gln	Ala	Pro	Ser	Val	Pro	Gln

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625	630	635	640
Tyr His Phe Gln Gln Arg Val Asn Pro Ser Ser Asp Lys His Leu Ser	645	650	655
Gln Gln Ala Thr Glu Thr Gln Arg Leu Ser Gly Phe Leu Gln His Thr	660	665	670
Pro Gln Thr Gln Ala Ser Gln Thr Pro Ala Ser Gln Asn Ser Asn Phe	675	680	685
Pro Gln Ile Cys Gln Gln Gln Gln Gln Gln Leu Gln Arg Lys Asn	690	695	700
Lys Glu Gln Met Pro Gln Thr Phe Ser His Leu Gln Gly Ser Asn Asp	705	710	715
Lys Gln Arg Glu Gly Ser Cys Phe Gly Gln Ile Lys Val Glu Glu Ser	725	730	735
Phe Cys Val Gly Asn Gln Tyr Ser Lys Ser Ser Asn Phe Gln Thr His	740	745	750
Asn Asn Thr Gln Gly Gly Leu Glu Gln Val Gln Asn Ile Asn Lys Asn	755	760	765
Phe Pro Tyr Ser Lys Ile Leu Thr Pro Asn Ser Ser Asn Leu Gln Ile	770	775	780
Leu Pro Ser Asn Asp Thr His Pro Ala Cys Glu Arg Glu Gln Ala Leu	785	790	795
His Pro Val Gly Ser Lys Thr Ser Asn Leu Gln Asn Met Gln Tyr Phe	805	810	815
Pro Asn Asn Val Thr Pro Asn Gln Asp Val His Arg Cys Phe Gln Glu	820	825	830
Gln Ala Gln Lys Pro Gln Gln Ala Ser Ser Leu Gln Gly Leu Lys Asp	835	840	845
Arg Ser Gln Gly Glu Ser Pro Ala Pro Pro Ala Glu Ala Ala Gln Gln	850	855	860
Arg Tyr Leu Val His Asn Glu Ala Lys Ala Leu Pro Val Pro Glu Gln	865	870	875
Gly Gly Ser Gln Thr Gln Thr Pro Pro Gln Lys Asp Thr Gln Lys His	885	890	895
Ala Ala Leu Arg Trp Leu Leu Leu Gln Lys Gln Glu Gln Gln Gln Thr	900	905	910
Gln Gln Ser Gln Pro Gly His Asn Gln Met Leu Arg Pro Ile Lys Thr	915	920	925
Glu Pro Val Ser Lys Pro Ser Ser Tyr Arg Tyr Pro Leu Ser Pro Pro	930	935	940
Gln Glu Asn Met Ser Ser Arg Ile Lys Gln Glu Ile Ser Ser Pro Ser	945	950	955
Arg Asp Asn Gly Gln Pro Lys Ser Ile Ile Glu Thr Met Glu Gln His	965	970	975
Leu Lys Gln Phe Gln Leu Lys Ser Leu Cys Asp Tyr Lys Ala Leu Thr	980	985	990
Leu Lys Ser Gln Lys His Val Lys Val Pro Thr Asp Ile Gln Ala Ala	995	1000	1005
Glu Ser Glu Asn His Ala Arg Ala Ala Glu Pro Gln Ala Thr Lys	1010	1015	1020
Ser Thr Asp Cys Ser Val Leu Asp Asp Val Ser Glu Ser Asp Thr	1025	1030	1035
Pro Gly Glu Gln Ser Gln Asn Gly Lys Cys Glu Gly Cys Asn Pro	1040	1045	1050

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Asp	Lys	Asp	Glu	Ala	Pro	Tyr	Tyr	Thr	His	Leu	Gly	Ala	Gly	Pro
1055						1060					1065			
Asp	Val	Ala	Ala	Ile	Arg	Thr	Leu	Met	Glu	Glu	Arg	Tyr	Gly	Glu
1070						1075					1080			
Lys	Gly	Lys	Ala	Ile	Arg	Ile	Glu	Lys	Val	Ile	Tyr	Thr	Gly	Lys
1085						1090					1095			
Glu	Gly	Lys	Ser	Ser	Gln	Gly	Cys	Pro	Ile	Ala	Lys	Trp	Val	Tyr
1100						1105					1110			
Arg	Arg	Ser	Ser	Glu	Glu	Glu	Lys	Leu	Leu	Cys	Leu	Val	Arg	Val
1115						1120					1125			
Arg	Pro	Asn	His	Thr	Cys	Glu	Thr	Ala	Val	Met	Val	Ile	Ala	Ile
1130						1135					1140			
Met	Leu	Trp	Asp	Gly	Ile	Pro	Lys	Leu	Leu	Ala	Ser	Glu	Leu	Tyr
1145						1150					1155			
Ser	Glu	Leu	Thr	Asp	Ile	Leu	Gly	Lys	Cys	Gly	Ile	Cys	Thr	Asn
1160						1165					1170			
Arg	Arg	Cys	Ser	Gln	Asn	Glu	Thr	Arg	Asn	Cys	Cys	Cys	Gln	Gly
1175						1180					1185			
Glu	Asn	Pro	Glu	Thr	Cys	Gly	Ala	Ser	Phe	Ser	Phe	Gly	Cys	Ser
1190						1195					1200			
Trp	Ser	Met	Tyr	Tyr	Asn	Gly	Cys	Lys	Phe	Ala	Arg	Ser	Lys	Lys
1205						1210					1215			
Pro	Arg	Lys	Phe	Arg	Leu	His	Gly	Ala	Glu	Pro	Lys	Glu	Glu	Glu
1220						1225					1230			
Arg	Leu	Gly	Ser	His	Leu	Gln	Asn	Leu	Ala	Thr	Val	Ile	Ala	Pro
1235						1240					1245			
Ile	Tyr	Lys	Lys	Leu	Ala	Pro	Asp	Ala	Tyr	Asn	Asn	Gln	Val	Glu
1250						1255					1260			
Phe	Glu	His	Gln	Ala	Pro	Asp	Cys	Cys	Leu	Gly	Leu	Lys	Glu	Gly
1265						1270					1275			
Arg	Pro	Phe	Ser	Gly	Val	Thr	Ala	Cys	Leu	Asp	Phe	Ser	Ala	His
1280						1285					1290			
Ser	His	Arg	Asp	Gln	Gln	Asn	Met	Pro	Asn	Gly	Ser	Thr	Val	Val
1295						1300					1305			
Val	Thr	Leu	Asn	Arg	Glu	Asp	Asn	Arg	Glu	Val	Gly	Ala	Lys	Pro
1310						1315					1320			
Glu	Asp	Glu	Gln	Phe	His	Val	Leu	Pro	Met	Tyr	Ile	Ile	Ala	Pro
1325						1330					1335			
Glu	Asp	Glu	Phe	Gly	Ser	Thr	Glu	Gly	Gln	Glu	Lys	Lys	Ile	Arg
1340						1345					1350			
Met	Gly	Ser	Ile	Glu	Val	Leu	Gln	Ser	Phe	Arg	Arg	Arg	Arg	Val
1355						1360					1365			
Ile	Arg	Ile	Gly	Glu	Leu	Pro	Lys	Ser	Cys	Lys	Lys	Lys	Ala	Glu
1370						1375					1380			
Pro	Lys	Lys	Ala	Lys	Thr	Lys	Lys	Ala	Ala	Arg	Lys	Arg	Ser	Ser
1385						1390					1395			
Leu	Glu	Asn	Cys	Ser	Ser	Arg	Thr	Glu	Lys	Gly	Lys	Ser	Ser	Ser
1400						1405					1410			
His	Thr	Lys	Leu	Met	Glu	Asn	Ala	Ser	His	Met	Lys	Gln	Met	Thr
1415						1420					1425			
Ala	Gln	Pro	Gln	Leu	Ser	Gly	Pro	Val	Ile	Arg	Gln	Pro	Pro	Thr
1430						1435					1440			

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Leu	Gln	Arg	His	Leu	Gln	Gln	Gly	Gln	Arg	Pro	Gln	Gln	Pro	Gln
1445						1450					1455			
Pro	Pro	Gln	Pro	Gln	Pro	Gln	Thr	Thr	Pro	Gln	Pro	Gln	Pro	Gln
1460						1465					1470			
Pro	Gln	His	Ile	Met	Pro	Gly	Asn	Ser	Gln	Ser	Val	Gly	Ser	His
1475						1480					1485			
Cys	Ser	Gly	Ser	Thr	Ser	Val	Tyr	Thr	Arg	Gln	Pro	Thr	Pro	His
1490						1495					1500			
Ser	Pro	Tyr	Pro	Ser	Ser	Ala	His	Thr	Ser	Asp	Ile	Tyr	Gly	Asp
1505						1510					1515			
Thr	Asn	His	Val	Asn	Phe	Tyr	Pro	Thr	Ser	Ser	His	Ala	Ser	Gly
1520						1525					1530			
Ser	Tyr	Leu	Asn	Pro	Ser	Asn	Tyr	Met	Asn	Pro	Tyr	Leu	Gly	Leu
1535						1540					1545			
Leu	Asn	Gln	Asn	Asn	Gln	Tyr	Ala	Pro	Phe	Pro	Tyr	Asn	Gly	Ser
1550						1555					1560			
Val	Pro	Val	Asp	Asn	Gly	Ser	Pro	Phe	Leu	Gly	Ser	Tyr	Ser	Pro
1565						1570					1575			
Gln	Ala	Gln	Ser	Arg	Asp	Leu	His	Arg	Tyr	Pro	Asn	Gln	Asp	His
1580						1585					1590			
Leu	Thr	Asn	Gln	Asn	Leu	Pro	Pro	Ile	His	Thr	Leu	His	Gln	Gln
1595						1600					1605			
Thr	Phe	Gly	Asp	Ser	Pro	Ser	Lys	Tyr	Leu	Ser	Tyr	Gly	Asn	Gln
1610						1615					1620			
Asn	Met	Gln	Arg	Asp	Ala	Phe	Thr	Thr	Asn	Ser	Thr	Leu	Lys	Pro
1625						1630					1635			
Asn	Val	His	His	Leu	Ala	Thr	Phe	Ser	Pro	Tyr	Pro	Thr	Pro	Lys
1640						1645					1650			
Met	Asp	Ser	His	Phe	Met	Gly	Ala	Ala	Ser	Arg	Ser	Pro	Tyr	Ser
1655						1660					1665			
His	Pro	His	Thr	Asp	Tyr	Lys	Thr	Ser	Glu	His	His	Leu	Pro	Ser
1670						1675					1680			
His	Thr	Ile	Tyr	Ser	Tyr	Thr	Ala	Ala	Ala	Ser	Gly	Ser	Ser	Ser
1685						1690					1695			
Ser	His	Ala	Phe	His	Asn	Lys	Glu	Asn	Asp	Asn	Ile	Ala	Asn	Gly
1700						1705					1710			
Leu	Ser	Arg	Val	Leu	Pro	Gly	Phe	Asn	His	Asp	Arg	Thr	Ala	Ser
1715						1720					1725			
Ala	Gln	Glu	Leu	Leu	Tyr	Ser	Leu	Thr	Gly	Ser	Ser	Gln	Glu	Lys
1730						1735					1740			
Gln	Pro	Glu	Val	Ser	Gly	Gln	Asp	Ala	Ala	Ala	Val	Gln	Glu	Ile
1745						1750					1755			
Glu	Tyr	Trp	Ser	Asp	Ser	Glu	His	Asn	Phe	Gln	Asp	Pro	Cys	Ile
1760						1765					1770			
Gly	Gly	Val	Ala	Ile	Ala	Pro	Thr	His	Gly	Ser	Ile	Leu	Ile	Glu
1775						1780					1785			
Cys	Ala	Lys	Cys	Glu	Val	His	Ala	Thr	Thr	Lys	Val	Asn	Asp	Pro
1790						1795					1800			
Asp	Arg	Asn	His	Pro	Thr	Arg	Ile	Ser	Leu	Val	Leu	Tyr	Arg	His
1805						1810					1815			
Lys	Asn	Leu	Phe	Leu	Pro	Lys	His	Cys	Leu	Ala	Leu	Trp	Glu	Ala
1820						1825					1830			
Lys	Met	Ala	Glu	Lys	Ala	Arg	Lys	Glu	Glu	Glu	Cys	Gly	Lys	Asn

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1835	1840	1845
Gly Ser Asp His Val Ser Gln Lys Asn His Gly Lys Gln Glu Lys		
1850	1855	1860
Arg Glu Pro Thr Gly Pro Gln Glu Pro Ser Tyr Leu Arg Phe Ile		
1865	1870	1875
Gln Ser Leu Ala Glu Asn Thr Gly Ser Val Thr Thr Asp Ser Thr		
1880	1885	1890
Val Thr Thr Ser Pro Tyr Ala Phe Thr Gln Val Thr Gly Pro Tyr		
1895	1900	1905
Asn Thr Phe Val		
1910		
<p><210> SEQ ID NO 14 <211> LENGTH: 1713 <212> TYPE: PRT <213> ORGANISM: Mus musculus</p>		
<400> SEQUENCE: 14		
Met Phe Leu Pro Glu Thr Pro Gln Gln Tyr Ala Val Glu Ile Asn Ala		
1	5	10 15
Arg Glu Gly Thr Gly Pro Trp Ala Gln Gly Ala Thr Val Lys Thr Gly		
	20	25 30
Ser Glu Leu Ser Pro Val Asp Gly Pro Val Pro Gly Gln Met Asp Ser		
	35	40 45
Gly Pro Val Tyr His Gly Asp Ser Arg Gln Leu Ser Thr Ser Gly Ala		
	50	55 60
Pro Val Asn Gly Ala Arg Glu Pro Ala Gly Pro Gly Leu Leu Gly Ala		
65	70	75 80
Ala Gly Pro Trp Arg Val Asp Gln Lys Pro Asp Trp Glu Ala Ala Ser		
	85	90 95
Gly Pro Thr His Ala Ala Arg Leu Glu Asp Ala His Asp Leu Val Ala		
	100	105 110
Phe Ser Ala Val Ala Glu Ala Val Ser Ser Tyr Gly Ala Leu Ser Thr		
	115	120 125
Arg Leu Tyr Glu Thr Phe Asn Arg Glu Met Ser Arg Glu Ala Gly Ser		
	130	135 140
Asn Gly Arg Gly Pro Arg Pro Glu Ser Cys Ser Glu Gly Ser Glu Asp		
145	150	155 160
Leu Asp Thr Leu Gln Thr Ala Leu Ala Leu Ala Arg His Gly Met Lys		
	165	170 175
Pro Pro Asn Cys Thr Cys Asp Gly Pro Glu Cys Pro Asp Phe Leu Glu		
	180	185 190
Trp Leu Glu Gly Lys Ile Lys Ser Met Ala Met Glu Gly Gly Gln Gly		
	195	200 205
Arg Pro Arg Leu Pro Gly Ala Leu Pro Pro Ser Glu Ala Gly Leu Pro		
	210	215 220
Ala Pro Ser Thr Arg Pro Pro Leu Leu Ser Ser Glu Val Pro Gln Val		
225	230	235 240
Pro Pro Leu Glu Gly Leu Pro Leu Ser Gln Ser Ala Leu Ser Ile Ala		
	245	250 255
Lys Glu Lys Asn Ile Ser Leu Gln Thr Ala Ile Ala Ile Glu Ala Leu		
	260	265 270
Thr Gln Leu Ser Ser Ala Leu Pro Gln Pro Ser His Ser Thr Ser Gln		
	275	280 285

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Ala	Ser	Cys	Pro	Leu	Pro	Glu	Ala	Leu	Ser	Pro	Ser	Ala	Pro	Phe	Arg
290						295					300				
Ser	Pro	Gln	Ser	Tyr	Leu	Arg	Ala	Pro	Ser	Trp	Pro	Val	Val	Pro	Pro
305					310					315					320
Glu	Glu	His	Pro	Ser	Phe	Ala	Pro	Asp	Ser	Pro	Ala	Phe	Pro	Pro	Ala
				325					330					335	
Thr	Pro	Arg	Pro	Glu	Phe	Ser	Glu	Ala	Trp	Gly	Thr	Asp	Thr	Pro	Pro
			340					345					350		
Ala	Thr	Pro	Arg	Asn	Ser	Trp	Pro	Val	Pro	Arg	Pro	Ser	Pro	Asp	Pro
		355					360					365			
Met	Ala	Glu	Leu	Glu	Gln	Leu	Leu	Gly	Ser	Ala	Ser	Asp	Tyr	Ile	Gln
370						375					380				
Ser	Val	Phe	Lys	Arg	Pro	Glu	Ala	Leu	Pro	Thr	Lys	Pro	Lys	Val	Lys
385					390					395					400
Val	Glu	Ala	Pro	Ser	Ser	Ser	Pro	Ala	Pro	Val	Pro	Ser	Pro	Ile	Ser
				405					410					415	
Gln	Arg	Glu	Ala	Pro	Leu	Leu	Ser	Ser	Glu	Pro	Asp	Thr	His	Gln	Lys
			420					425					430		
Ala	Gln	Thr	Ala	Leu	Gln	Gln	His	Leu	His	His	Lys	Arg	Asn	Leu	Phe
		435					440					445			
Leu	Glu	Gln	Ala	Gln	Asp	Ala	Ser	Phe	Pro	Thr	Ser	Thr	Glu	Pro	Gln
	450					455					460				
Ala	Pro	Gly	Trp	Trp	Ala	Pro	Pro	Gly	Ser	Pro	Ala	Pro	Arg	Pro	Pro
465					470					475					480
Asp	Lys	Pro	Pro	Lys	Glu	Lys	Lys	Lys	Lys	Pro	Pro	Thr	Pro	Ala	Gly
				485					490					495	
Gly	Pro	Val	Gly	Ala	Glu	Lys	Thr	Thr	Pro	Gly	Ile	Lys	Thr	Ser	Val
			500					505					510		
Arg	Lys	Pro	Ile	Gln	Ile	Lys	Lys	Ser	Arg	Ser	Arg	Asp	Met	Gln	Pro
		515					520					525			
Leu	Phe	Leu	Pro	Val	Arg	Gln	Ile	Val	Leu	Glu	Gly	Leu	Lys	Pro	Gln
	530					535					540				
Ala	Ser	Glu	Gly	Gln	Ala	Pro	Leu	Pro	Ala	Gln	Leu	Ser	Val	Pro	Pro
545					550					555					560
Pro	Ala	Ser	Gln	Gly	Ala	Ala	Ser	Gln	Ser	Cys	Ala	Thr	Pro	Leu	Thr
				565					570					575	
Pro	Glu	Pro	Ser	Leu	Ala	Leu	Phe	Ala	Pro	Ser	Pro	Ser	Gly	Asp	Ser
			580					585					590		
Leu	Leu	Pro	Pro	Thr	Gln	Glu	Met	Arg	Ser	Pro	Ser	Pro	Met	Val	Ala
		595					600						605		
Leu	Gln	Ser	Gly	Ser	Thr	Gly	Gly	Pro	Leu	Pro	Pro	Ala	Asp	Asp	Lys
	610					615					620				
Leu	Glu	Glu	Leu	Ile	Arg	Gln	Phe	Glu	Ala	Glu	Phe	Gly	Asp	Ser	Phe
625					630					635					640
Gly	Leu	Pro	Gly	Pro	Pro	Ser	Val	Pro	Ile	Gln	Glu	Pro	Glu	Asn	Gln
				645					650					655	
Ser	Thr	Cys	Leu	Pro	Ala	Pro	Glu	Ser	Pro	Phe	Ala	Thr	Arg	Ser	Pro
			660					665					670		
Lys	Lys	Ile	Lys	Ile	Glu	Ser	Ser	Gly	Ala	Val	Thr	Val	Leu	Ser	Thr
		675					680					685			
Thr	Cys	Phe	His	Ser	Glu	Glu	Gly	Gly	Gln	Glu	Ala	Thr	Pro	Thr	Lys
	690					695					700				
Ala	Glu	Asn	Pro	Leu	Thr	Pro	Thr	Leu	Ser	Gly	Phe	Leu	Glu	Ser	Pro

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705	710	715	720
Leu Lys Tyr Leu Asp Thr Pro Thr Lys Ser Leu Leu Asp Thr Pro Ala 725 730 735			
Lys Lys Ala Gln Ser Glu Phe Pro Thr Cys Asp Cys Val Glu Gln Ile 740 745 750			
Val Glu Lys Asp Glu Gly Pro Tyr Tyr Thr His Leu Gly Ser Gly Pro 755 760 765			
Thr Val Ala Ser Ile Arg Glu Leu Met Glu Asp Arg Tyr Gly Glu Lys 770 775 780			
Gly Lys Ala Ile Arg Ile Glu Lys Val Ile Tyr Thr Gly Lys Glu Gly 785 790 795 800			
Lys Ser Ser Arg Gly Cys Pro Ile Ala Lys Trp Val Ile Arg Arg His 805 810 815			
Thr Leu Glu Glu Lys Leu Leu Cys Leu Val Arg His Arg Ala Gly His 820 825 830			
His Cys Gln Asn Ala Val Ile Val Ile Leu Ile Leu Ala Trp Glu Gly 835 840 845			
Ile Pro Arg Ser Leu Gly Asp Thr Leu Tyr Gln Glu Leu Thr Asp Thr 850 855 860			
Leu Arg Lys Tyr Gly Asn Pro Thr Ser Arg Arg Cys Gly Leu Asn Asp 865 870 875 880			
Asp Arg Thr Cys Ala Cys Gln Gly Lys Asp Pro Asn Thr Cys Gly Ala 885 890 895			
Ser Phe Ser Phe Gly Cys Ser Trp Ser Met Tyr Phe Asn Gly Cys Lys 900 905 910			
Tyr Ala Arg Ser Lys Thr Pro Arg Lys Phe Arg Leu Thr Gly Asp Asn 915 920 925			
Pro Lys Glu Glu Glu Val Leu Arg Asn Ser Phe Gln Asp Leu Ala Thr 930 935 940			
Glu Val Ala Pro Leu Tyr Lys Arg Leu Ala Pro Gln Ala Tyr Gln Asn 945 950 955 960			
Gln Val Thr Asn Glu Asp Val Ala Ile Asp Cys Arg Leu Gly Leu Lys 965 970 975			
Glu Gly Arg Pro Phe Ser Gly Val Thr Ala Cys Met Asp Phe Cys Ala 980 985 990			
His Ala His Lys Asp Gln His Asn Leu Tyr Asn Gly Cys Thr Val Val 995 1000 1005			
Cys Thr Leu Thr Lys Glu Asp Asn Arg Cys Val Gly Gln Ile Pro 1010 1015 1020			
Glu Asp Glu Gln Leu His Val Leu Pro Leu Tyr Lys Met Ala Ser 1025 1030 1035			
Thr Asp Glu Phe Gly Ser Glu Glu Asn Gln Asn Ala Lys Val Ser 1040 1045 1050			
Ser Gly Ala Ile Gln Val Leu Thr Ala Phe Pro Arg Glu Val Arg 1055 1060 1065			
Arg Leu Pro Glu Pro Ala Lys Ser Cys Arg Gln Arg Gln Leu Glu 1070 1075 1080			
Ala Arg Lys Ala Ala Ala Glu Lys Lys Lys Leu Gln Lys Glu Lys 1085 1090 1095			
Leu Ser Thr Pro Glu Lys Ile Lys Gln Glu Ala Leu Glu Leu Ala 1100 1105 1110			
Gly Val Thr Thr Asp Pro Gly Leu Ser Leu Lys Gly Gly Leu Ser 1115 1120 1125			

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Gln	Gln	Ser	Leu	Lys	Pro	Ser	Leu	Lys	Val	Glu	Pro	Gln	Asn	His
1130						1135					1140			
Phe	Ser	Ser	Phe	Lys	Tyr	Ser	Gly	Asn	Ala	Val	Val	Glu	Ser	Tyr
1145						1150					1155			
Ser	Val	Leu	Gly	Ser	Cys	Arg	Pro	Ser	Asp	Pro	Tyr	Ser	Met	Ser
1160						1165					1170			
Ser	Val	Tyr	Ser	Tyr	His	Ser	Arg	Tyr	Ala	Gln	Pro	Gly	Leu	Ala
1175						1180					1185			
Ser	Val	Asn	Gly	Phe	His	Ser	Lys	Tyr	Thr	Leu	Pro	Ser	Phe	Gly
1190						1195					1200			
Tyr	Tyr	Gly	Phe	Pro	Ser	Ser	Asn	Pro	Val	Phe	Pro	Ser	Gln	Phe
1205						1210					1215			
Leu	Gly	Pro	Ser	Ala	Trp	Gly	His	Gly	Gly	Ser	Gly	Gly	Ser	Phe
1220						1225					1230			
Glu	Lys	Lys	Pro	Asp	Leu	His	Ala	Leu	His	Asn	Ser	Leu	Asn	Pro
1235						1240					1245			
Ala	Tyr	Gly	Gly	Ala	Glu	Phe	Ala	Glu	Leu	Pro	Gly	Gln	Ala	Val
1250						1255					1260			
Ala	Thr	Asp	Asn	His	His	Pro	Ile	Pro	His	His	Gln	Gln	Pro	Ala
1265						1270					1275			
Tyr	Pro	Gly	Pro	Lys	Glu	Tyr	Leu	Leu	Pro	Lys	Val	Pro	Gln	Leu
1280						1285					1290			
His	Pro	Ala	Ser	Arg	Asp	Pro	Ser	Pro	Phe	Ala	Gln	Ser	Ser	Ser
1295						1300					1305			
Cys	Tyr	Asn	Arg	Ser	Ile	Lys	Gln	Glu	Pro	Ile	Asp	Pro	Leu	Thr
1310						1315					1320			
Gln	Ala	Glu	Ser	Ile	Pro	Arg	Asp	Ser	Ala	Lys	Met	Ser	Arg	Thr
1325						1330					1335			
Pro	Leu	Pro	Glu	Ala	Ser	Gln	Asn	Gly	Gly	Pro	Ser	His	Leu	Trp
1340						1345					1350			
Gly	Gln	Tyr	Ser	Gly	Gly	Pro	Ser	Met	Ser	Pro	Lys	Arg	Thr	Asn
1355						1360					1365			
Ser	Val	Gly	Gly	Asn	Trp	Gly	Val	Phe	Pro	Pro	Gly	Glu	Ser	Pro
1370						1375					1380			
Thr	Ile	Val	Pro	Asp	Lys	Leu	Asn	Ser	Phe	Gly	Ala	Ser	Cys	Leu
1385						1390					1395			
Thr	Pro	Ser	His	Phe	Pro	Glu	Ser	Gln	Trp	Gly	Leu	Phe	Thr	Gly
1400						1405					1410			
Glu	Gly	Gln	Gln	Ser	Ala	Pro	His	Ala	Gly	Ala	Arg	Leu	Arg	Gly
1415						1420					1425			
Lys	Pro	Trp	Ser	Pro	Cys	Lys	Phe	Gly	Asn	Gly	Thr	Ser	Ala	Leu
1430						1435					1440			
Thr	Gly	Pro	Ser	Leu	Thr	Glu	Lys	Pro	Trp	Gly	Met	Gly	Thr	Gly
1445						1450					1455			
Asp	Phe	Asn	Pro	Ala	Leu	Lys	Gly	Gly	Pro	Gly	Phe	Gln	Asp	Lys
1460						1465					1470			
Leu	Trp	Asn	Pro	Val	Lys	Val	Glu	Glu	Gly	Arg	Ile	Pro	Thr	Pro
1475						1480					1485			
Gly	Ala	Asn	Pro	Leu	Asp	Lys	Ala	Trp	Gln	Ala	Phe	Gly	Met	Pro
1490						1495					1500			
Leu	Ser	Ser	Asn	Glu	Lys	Leu	Phe	Gly	Ala	Leu	Lys	Ser	Glu	Glu
1505						1510					1515			

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Lys	Leu	Trp	Asp	Pro	Phe	Ser	Leu	Glu	Glu	Gly	Thr	Ala	Glu	Glu
1520						1525					1530			
Pro	Pro	Ser	Lys	Gly	Val	Val	Lys	Glu	Glu	Lys	Ser	Gly	Pro	Thr
1535						1540					1545			
Val	Glu	Glu	Asp	Glu	Glu	Glu	Leu	Trp	Ser	Asp	Ser	Glu	His	Asn
1550						1555					1560			
Phe	Leu	Asp	Glu	Asn	Ile	Gly	Gly	Val	Ala	Val	Ala	Pro	Ala	His
1565						1570					1575			
Cys	Ser	Ile	Leu	Ile	Glu	Cys	Ala	Arg	Arg	Glu	Leu	His	Ala	Thr
1580						1585					1590			
Thr	Pro	Leu	Lys	Lys	Pro	Asn	Arg	Cys	His	Pro	Thr	Arg	Ile	Ser
1595						1600					1605			
Leu	Val	Phe	Tyr	Gln	His	Lys	Asn	Leu	Asn	Gln	Pro	Asn	His	Gly
1610						1615					1620			
Leu	Ala	Leu	Trp	Glu	Ala	Lys	Met	Lys	Gln	Leu	Ala	Glu	Arg	Ala
1625						1630					1635			
Arg	Gln	Arg	Gln	Glu	Glu	Ala	Ala	Arg	Leu	Gly	Leu	Gly	Gln	Gln
1640						1645					1650			
Glu	Ala	Lys	Leu	Tyr	Gly	Lys	Lys	Arg	Lys	Trp	Gly	Gly	Ala	Met
1655						1660					1665			
Val	Ala	Glu	Pro	Gln	His	Lys	Glu	Lys	Lys	Gly	Ala	Ile	Pro	Thr
1670						1675					1680			
Arg	Gln	Ala	Leu	Ala	Met	Pro	Thr	Asp	Ser	Ala	Val	Thr	Val	Ser
1685						1690					1695			
Ser	Tyr	Ala	Tyr	Thr	Lys	Val	Thr	Gly	Pro	Tyr	Ser	Arg	Trp	Ile
1700						1705					1710			

What is claimed is:

1. A composition comprising a reducing agent capable of converting 5-formylcytosine or 5-carboxycytosine to 5-hydroxymethylcytosine; T4 β -glucosyltransferase (BGT) and a UDP derivative selected from the group consisting of UDP-glucose and UDP-azidoglucose.

2. A composition according to claim 1, further comprising a nucleic acid.

3. A kit comprising a reducing agent capable of converting 5-formylcytosine or 5-carboxycytosine to 5-hydroxymethyl-

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cytosine; T4 β -glucosyltransferase (BGT); and a UDP derivative selected from the group consisting of UDP-glucose and UDP-azidoglucose.

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4. A kit according to claim 3, further comprising UDP-glucosamine.

5. A kit according to claim 3, further comprising a nucleic acid.

6. A kit according to any claim 3, further comprising an oxidizing agent that is capable of oxidizing 5-methylcytosine to 5-hydroxymethylcytosine.

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