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(54) **PEPTIDE FRAGMENTS FOR INDUCING SYNTHESIS OF EXTRACELLULAR MATRIX PROTEINS**

(75) Inventors: **Scott M. Harris**, Seattle, WA (US);  
**Timothy J. Falla**, Woodinville, WA (US);  
**Lijuan Zhang**, Kenmore, WA (US)

(73) Assignee: **Helix Biomedix, Inc.**, Bothell, WA (US)

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*Primary Examiner* — David Lukton

(74) *Attorney, Agent, or Firm* — Novak Druce Connolly Bove + Quigg LLP

(57) **ABSTRACT**

Short biologically active tetrapeptides are disclosed that are comprised of the sequences GxxG and PxxP where G (glycine) and P (proline) are maintained and x is a variable amino acid. The peptides can be used singly or in combination to stimulate production of extracellular matrix proteins in skin. A rapid, low-cost method of producing heterogenous formulations of tetrapeptides is disclosed.

**14 Claims, 3 Drawing Sheets**

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MGPRLSVWLL LLPAALLLHE EHSRAAAKGG CAGSGCGKCD CHGVKGQKGE  
 RGLPGLQGVI GFPGMQGPEG PQGPPGQKGD TGEPGLPGTK GTRGPPGASG  
 YPGNPGLPGI PGQDGPPGPP GIPGCNGTKG ERGPLGPPGL PGFAGNPGPP  
GLPGMKGDPG EILGHVPGML LKGERGFPGI PGTPGPPGLP GLQGPVGGPPG  
 FTGPPGPPGP PGPPGEKQOM GLSFQGPKGD KGDQGVSGPP GVPGOAQVQE  
 KGDFATKGEK GQKGEFQFQ MPGVGEKGEF GKPGPRGKPG KDGDKGEKGS  
PGFPGEFYP GLIGRQGPQG EKGEAGPPGP PGIVIGTGPL GEKGERGYPG  
 TPGPRGEPGP KGFPGLPQOP GPPGLPVPQO AGAPGFPGER GEKDRGFPG  
 TSLPGPSGRD GLPGPPGSPG PPGQPGYTNG IVECQPPGPP DQGPPGIPGO  
PGFIGEIGEK GQKGESCLIC DIDGYRGPPG PQGPPGEIGF PGQPGAAGDR  
GLPGRDGVAG VPGPOGTPGL IGQPGAAGEP GEFYFDLRLK GDKGDPGFPG  
QPGMPGRAGS PGRDGHPGLP GPKGSPGSVG LKGERGPPGG VGFPGSRGDT  
GPPGPPGYGP AGPIGDKGQA GFPGGPGSPG LPGPKGEFPG IVPLPGPPGA  
 EGLPGSPGFP GPQGDRGFPG TPGRPGLPGE KGAVGQPGIG FPGPPGPKGV  
DGLPGDMGPP GTPGRPGFNG LPGNPGVQOQ KGEFVGLPG LKGLPGLPGI  
PGTPGEKCSI GVPGVPEHG AIGPPGLQGI RGEPPGPPGL GSVGSPPVPG  
IGPPGARGPP GGQPPGLSG PPGIKGEKGF PGFPGLDMPG PKGDKGAQGL  
PGITGQSGLP GLPGQQGAPG IPGFPGSKGE MGVMGTPGQP GSPGVPAGP  
LPGEKGDHGF PGSSGPRGDP GLKGDKGDVG LPKPGSMDK VDMGSMKGQK  
 GDQGEKQIG PIGEKGSRGD PGTPGVPGKD GQAGQPGOPG PKGDPGISGT  
PGAPGLPGPK GSVGGMGLPG TPGEKGVPGI PGPQGSPLP GDKGAKGEK  
QAGPPGIGIP GLRGEKGDQO IAGFPSPGE KGEKSGIGIP GMPGSPGLKG  
SPGSVGYPGS PGLPGEKGDK GLPGLDGIPG VKGEAGLPGT PGPTGPAGQK  
GEFGSDGIPG SAGEKGEFGL PGRGFPGFPG AKGDKGSKGE VGFPLAGSP  
GIPGSKGEOG FMGPPGPQOQ PGLPGSPGHA TEGPKGDRGP QGOPGLPGLP  
GPMGPPGLPG IDGVKGDKGN PGWPGAPGVP GPKGDPGFQO MPGIGGSPGI  
 TGSKGDMGPP GVPGFQGPKG LPGLOGIKGD QGDQGVPGAK GLPGPPGPPG  
 PYDIIKGEFG LPGPEGPPGL KGLQGLPGPK GQQGVVTGLVG IPGPPGIPGF  
 DGAPGQKEM GPAGPTGPRG FPGPPGPDGL PGSMGPPGTP SVDHGFLVTR  
 HSQTIDDPQC PSGTKILYHG YSLLYVQNE RAHQDLGTA GSCLRKFSTM  
 PFLFCNINNV CNFASRNDYS YWLSTPEPMP MSMAPITGEN IRPFISRCAV  
 CEAPAMVMAV HSQTIQIPPC PSGWSSLWIG YSFVMHTSAG AEGSGQALAS  
 PGSCLEEFRS APFIECHGRG TCNYYANAYS FWLATIERSE MEKKPTPSTL  
 KAGELRTHVS RCQVCMRRT

FIG. 1

MMSFVQKGSW LLLALLHPTI ILAQQEAVEG GCSHLGQSYA DRDVWKPEPC  
QICVCDSGSV LCDDIICDDQ ELDCPNPEIP FGECCAVCPQ PPTAPTRPPN  
GQGPQGPKGD PGPPGIPGRN GDPGIPGQPG SPGSPGPPGI CESCPTGPQN  
YSPQYDSYDV KSGVAVGGLA GYPGPAGPPG PPGPPGTSGH PGSPGSPGYQ  
GPPGEPGQAG PSGPPGPPGA IGPSGPAGKD GESGRPGRPG ERGLPGPPGI  
KGPAGIPGFP GMKGHRGFDG RNGEKGETGA PGLKGENGLP GENGAPGPMG  
PRGAPGERGR PGLPGAAGAR GNDGARGSDG QPGPPGPPGT AGFPGPSGAK  
GEVGPAGSPG SNGAPGQRGE PGPQGHAGAQ GPPGPPGING SPGGKGEMGP  
AGIPGAPGLM GARGPPGPAG ANGAPGLRGG AGEFGKNGAK GE**PGPR**GERG  
EAGIPGVPGA KGEDGKDGSP GEPGANGLPG AAGERGAPGF RGPAGPNGIP  
GEKGPAGERG APGPAGPRGA AGEFGRDGVP GGPGMRGMPG SPGGPGSDGK  
PGPPGSQGES GRPGPPGPSG PRGQPGVMGF PGPKGNDGAP GKNGERGGPG  
GPGPQGP PGK NGETGPQGPP GPTGPGGDKG DTGPPGPQGL QGLPGTGGPP  
GENGKPGEPG PKGDAGAPGA PGGKGDAGAP GERGPPGLAG APGLR**GAGP**  
PGPEGGKGAA GPPGPPGAAG TPGLQGMPGE RGGLGSPGPK GDKGEPGGPG  
ADGVPKDGDP RGPTGPIGPP GPAGQPGDKG EGGAPGLPGI AGPRGSPGER  
GETGPPGPAG FPGAPQNGE PGGKGERGAP GEKGEPPGPP VAGPPGSSGP  
AGPPGPQGVK GERGSPGGPG AAGFPGARGL PGPPGSNGNP GPPGPSGSPG  
KDGPPGPAGN TGAPGSPGVS GPKGDAGQPG EKGSPPGAQGP PGAPGPLGIA  
GITGARGLAG PPGM**PGPR**GS PGPQGVKGES GKPGANGLSG ERGPPGPQGL  
PGLAGTAGEP GRDGNPGSDG LPGRDGSPGG KGDRGENGSP GAPGAPGHPG  
PPGPVGPAGK SGDRGESGPA GPAGAPGPAG SRGAPGPQGP RGDKGETGER  
GAAGIKGHRG FPGNPGAPGS PGPAGQQGAI GSPGPAGPRG PVGPSGPPGK  
DGTSGHFGPI GP**PGPR**NRG ERGSEGSPGH PGQPGPPGPP GAPGPCCGGV  
GAAAIAGIGG EKAGGFAPYY GDEPMDFKIN TDEIMTSLKS VNGQIESLIS  
PDGSRKNPAR NCRDLKFCHP ELKSGEYWVD PNQGCKLDAI KVFCNMETGE  
TCISANPLNV PRKHWWTDSS AEKKHVWFGE SMDGGFQFSY GNPPELPELVL  
DVQLAFLRLS SSRASQNITY HCKNSIAYMD QASGNVKKAL KLMGSNEGEF  
KAEGNSKFTY TVLEDGCTKH TGEWSKTVFE YRTRKAVRLP IVDIAPYDIG  
GPDQEFVVDV GPVCFL

FIG. 2

MGPRLSVWLL	LLPAALLLHE	EHSRAAAKGG	CAGSGCGKCD	CHGVKGQKGE
RGLPGLQGVI	GFPGMQGP	PQGPPGQKGD	TGEPGLPGTK	GTRGPPGASG
YPGNPGLPGI	PGQDGP <u>PGPP</u>	GIPGCNGTKG	ERGPLGPPGL	PGFAGN <u>PGPP</u>
GLPGMKGDPG	EILGHVPGML	LKGERGFPGI	PGT <u>PGPP</u> GLP	GLQGPVGP
FTGPP <u>PGPP</u> GP	<u>PGPP</u> GEKQOM	GLSFQGP	KGDQGVSGPP	GVPGQAQVQE
KGDFATKGEK	GQKGEPGFQ	MPGVGEKGE	GKPPRGKPG	KDGDKEKGS
PGFPGEPGYP	GLIGRQGP	EKGEAGP <u>PGP</u>	<u>PGP</u> IVIGTG	GEKGERGYP
TPGPRGEPGP	KGFPLPGQP	GPPGLPVP	AGAPGFPER	GEKDRGFPG
TSLPGPSGRD	GL <u>PGPP</u> GSPG	PPGQPGYTNG	IVECQ <u>PGPP</u>	DQGPPGIPGQ
PGFIGEIGEK	GQKGESCLIC	DIDGYRPPG	PQGPPGEIGF	PGQPGA
GLPGRDGVAG	VPGPQGT	IGQPGA	GEFYFDLRLK	GDKGDPGFPG
QPGMPGRAGS	PGRDGHPLP	GPKGSPG	LKGERGPPG	VGFPGSRGDT
G <u>PGPP</u> GYGP	AGPIGDKQA	GFPGGP	LPGPKGEPGK	IVPL <u>PGPP</u> GA
EGLPGSPGFP	GPQDRGFPG	TPGRPGLP	KGAVGQPGIG	<u>FPGPP</u> GPKGV
DGLPGDMGPP	GTPGRP	LPGNPGVQ	KGEPGVGLPG	LKGLPGLPGI
PGTPGEKGS	GVPGV	AIGPPGLQ	RGE <u>PGPP</u> GLP	GSVGS
IGPPGARGPP	GGQPPGLSG	PPGIKGEKGF	PGFPGLDMPG	PKGDKGAQGL
PGITGQSGLP	GLPGQQGAPG	IPGFPGSKGE	MGMGT	GSPGPV
LPGEKGDHGF	PGSSGPRGDP	GLKGDGDVG	LPGKPGSMDK	VDMGSMKGQK
GDQGEKQIG	PIGEKSRGD	PGTPGVPGKD	GQAGQPGQPG	PKGDPGISGT
PGAPGLPGPK	GSVGMGLPG	TPGEKGVPGI	PGPQGS	GDKGAKGEK
QAGPPGIGIP	GLRGEKGDQ	IAGFP	KGEKSGIP	GMPGSPGLKG
SPGSVGYPGS	PGLPGEKGDK	GLPGLDGIPG	VKGEAGLP	PGPTGPAGQK
GEPGSDGIPG	SAGEKGE	PGRGFPGFP	AKGDKGSKGE	VGFPLAGSP
GIPGSKGEQ	FMGPPGPQ	PGLPGSPGHA	TEGPKGDRGP	QGQPGLPGLP
GPMGPPGLPG	IDGVKGDKN	PGWPGAPVP	GPKGDPGFQ	MPGIGGSPGI
TGSKGDMGPP	GVPGFQGP	LPGLQGIKGD	QGDQVPGAK	GL <u>PGPP</u> GPPG
PYDI IKGEPG	LPGPEGP	KGLQGLPGPK	GQQGVTGLVG	I <u>PGPP</u> GIPGF
DGAPGQKEM	GPAGPTGPRG	<u>FPGPP</u> GDGL	PGSMGPPGTP	SVDHGFLVTR
HSQTIDDPQC	PSGTKILYHG	YSLLYVQNE	RAHGQDLGTA	GSCLRKFSTM
PFLFCNINNV	CNFASRNDYS	YWLSTPEPMP	MSMAPITGEN	IRPFISRC
CEAPAMVMAV	HSQTIQIPPC	PSGWSSLWIG	YSFVMHTSAG	AEGSGQALAS
PGSCLEEFRS	APFIECHGRG	TCNYYANAYS	FWLATIERSE	MEKKPTPSTL
KAGELRTHVS	RCQVCMRRT			

FIG. 3

## 1

**PEPTIDE FRAGMENTS FOR INDUCING  
SYNTHESIS OF EXTRACELLULAR MATRIX  
PROTEINS**

This application is a continuation of application Ser. No. 11/811,876 filed Jun. 12, 2007, now U.S. Pat. No. 8,110,658, which claims benefit of priority to U.S. 60/813,284 filed Jun. 13, 2006, each of which is herein incorporated by reference in its entirety.

FIELD OF THE INVENTION

The invention relates to tetrapeptides with the amino acid motif GxxG or PxxP, where G (glycine) and P (proline) are maintained and x is a variable amino acid. The invention also relates to frame shift active tetrapeptides which are tetrapeptide sequences shifted one frame from a GxxG or PxxP tetrapeptide in an ECM protein. In particular, the invention relates to GxxG, PxxP, or frame shift active peptides that stimulate production of extracellular matrix proteins and enhance wound closure of the epithelial cell monolayer of scratch-wounded human skin. The peptide compositions may be used in formulations for repairing damaged skin or maintaining healthy skin.

BACKGROUND OF THE INVENTION

Skin aging is commonly viewed as wrinkle formation and impaired wound healing. A wound is defined as a break in the epithelial integrity of the skin. Normal wound healing involves a complex and dynamic but superbly orchestrated series of events leading to the repair of injured tissues. The largest component of normal skin is the extracellular matrix (ECM), a gel-like matrix produced by the cells that it surrounds. The ECM is composed of two major classes including fibrous structural proteins and proteoglycans. Changes in the composition and crosslinked state of the ECM are known to be associated with aging and a range of acquired and heritable skin disorders. It has been well documented that ECM not only provides structural support, but also influences cellular behavior such as differentiation and proliferation. Also, more and more research suggests that the matrix components may be a source of cell signals to facilitate epithelial cell proliferation and migration and thus enhance wound healing.

The largest class of fibrous ECM molecules is the collagen family, which includes at least 16 different types of collagen. Collagen in the dermal matrix is composed primarily of type I (80-85%) and type III (8-11%) collagens, both of which are fibrillar, or rod-shaped, collagens. The tensile strength of skin is due predominately to these fibrillar collagen molecules, which self-assemble into microfibrils in a head-to-tail and staggered side-to-side lateral arrangement. Collagen molecules become cross-linked to adjacent collagen molecules, creating additional strength and stability in collagen fibers. Damage to the collagen network (e.g. by enzymes or physical destruction), or its total collapse causes healing to take place by repair.

Various bioactive peptides that stimulate production of ECM proteins have been reported in both the scientific literature and in issued patents. Peptides historically have been isolated from natural sources and have recently been the subject of structure-function relationship studies. Natural peptides have also served as starting points for the design of synthetic peptide analogs.

Specific sequences within ECM proteins can stimulate useful elements in skin, such as type I collagen, type III collagen, and fibronectin (Katayama et. al., J. BIOL. CHEM. 288:9941-

## 2

9944 (1983)). Katayama et al. identified the pentapeptide, KTTKS (SEQ ID NO:17), within the carboxy-terminal propeptide (residues 197-241) of type I collagen. The propeptide is cleaved during production of the mature collagen protein. The cleaved propeptide may participate in regulating collagen production via a biosynthesis feedback mechanism, with the KTTKS segment playing an active role. Maquart et al. (J SOC BIOL. 193:423-28 (1999)) reported that the peptides GHK and CNYYSNS also stimulate ECM synthesis. These sequences may be released during ECM turnover, thereby signaling the need for ECM repair. The short peptide sequences liberated by either mechanism are often called "matrikines" (Maquart et al., J. Soc. Biol. 193:423-28 (1999)).

While a number of natural and synthetic peptides exist, there is a need for improved biologically active peptides and methods for their use.

SUMMARY OF THE INVENTION

Tetrapeptides are disclosed that are characterized by the amino acid sequence motif GxxG or PxxP, where G (glycine) and P (proline) residues are maintained and x is a variable amino acid. The tetrapeptides are derived from sequences that occur multiple times throughout the primary sequence of the ECM protein, type IV collagen. The disclosed sequences induce production of all forms of collagen more than previously known peptide sequences, including KTTKS, sold under the trademark MATRIXYL™ by SEDERMA SAS (France). Further, a composition comprising a combination of various multiply-repeating sequences elicits an even greater collagen-producing response. Additional benefits may be expected from peptide combinations present in a variety of ECM proteins.

Producing a specific combination of tetrapeptides for ECM rebuilding can be commercially cost-prohibitive. A relatively simple and cost-effective means of producing a diverse combination of biologically active tetrapeptides is disclosed. By producing a combinatorial library of tetrapeptides with the GxxG or PxxP motif, a variety of biologically active tetrapeptides can be generated in the same manufacturing run (e.g., GEPG, GPEG, GPPG, and GEEG). The combination of tetrapeptides may induce more formation of ECM proteins than single peptides. Compositions comprising the disclosed tetrapeptides, alone or in combination, are useful in skin care markets including, but not limited to, those that address skin wrinkling, toning, firmness, or sagging. The stimulation of collagen by the disclosed tetrapeptides can significantly improve the health and appearance of damaged and aged skin.

BRIEF DESCRIPTION OF THE FIGURES

FIG. 1 is SEQ ID NO:45 which is the Collagen IV amino acid sequence illustrating the occurrences of GxxG tetrapeptides. All bold sequences are underlined and overlapping sequences are double-underlined.

FIG. 2 is SEQ ID NO:46 which is the Collagen III amino acid sequence illustrating the occurrences of the frame shift actives PGPR and GAGP. All frame shift active sequences are bold and underlined and the GxxG sequences occurring one frame shift away are double-underlined.

FIG. 3 is also SEQ ID NO:45, the Collagen IV amino acid sequence, illustrating the occurrences of the tetrapeptide PGPP.

DETAILED DESCRIPTION OF THE INVENTION

The invention is generally directed towards tetrapeptides that stimulate production of ECM proteins and modulate wound healing, and uses of such tetrapeptides.

## Peptides

One embodiment of the invention is directed towards an isolated tetrapeptide comprising the motif GxxG or PxxP. In this embodiment G (glycine) or P (proline) is maintained and x is a variable amino acid. The peptide can generally be any peptide that falls within the above description, and more preferably is SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:8, SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:11, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:14, SEQ ID NO:15, or SEQ ID NO:16.

Another embodiment of the invention is directed towards an isolated tetrapeptide comprising the motif GxPG, where x is P at either variable position, or both. In this embodiment, G (glycine) and P (proline) are maintained and x is a variable amino acid. The peptide can generally be any peptide that falls within the above description, and more preferably is SEQ ID NO:2, SEQ ID NO:3, SEQ ID NO:5, or SEQ ID NO:7.

Another embodiment of the invention is directed towards an isolated tetrapeptide comprising the motif GExG. In this embodiment, G (glycine) and E (glutamic acid) are maintained and x is a variable amino acid. The peptide can generally be any peptide that falls within the above description, and more preferably is SEQ ID NO:5 or SEQ ID NO:8.

Another embodiment of the invention is directed towards an isolated tetrapeptide comprising the motif PGxP. In this embodiment, P (proline) and G (glycine) are maintained and x is a variable amino acid. The peptide can generally be any peptide that falls within the above description, and more preferably is SEQ ID NO:11, SEQ ID NO:12, SEQ ID NO:14, or SEQ ID NO:16.

Another embodiment of the invention is directed towards an isolated tetrapeptide comprising the motif PExP. In this embodiment, P (proline) and E (glutamic acid) are maintained and x is a variable amino acid. The peptide can generally be any peptide that falls within the above description, and more preferably is SEQ ID NO:1 or SEQ ID NO:9.

Another embodiment of the invention is directed towards a frame shift active tetrapeptide. In this embodiment, the tetrapeptide occurs one frame shift from either a GxxG or PxxP tetrapeptide in an ECM protein. The peptide can generally be any peptide that falls within the above description, and more preferably is SEQ ID NO:4 or SEQ ID NO:6.

Each of the above-described peptides can comprise D- or L-amino acids. The peptides can comprise all D-amino acids or L-amino acids. The peptides can have an acid C-terminus ( $-\text{CO}_2\text{H}$ ) or, preferably, an amide C-terminus ( $-\text{CONH}_2$ ,  $-\text{CONHR}$ , or  $-\text{CONR}_2$ ). The peptides may be further augmented or modified, either chemically or enzymatically. For example, the peptides may be amidated ( $-\text{NH}_2$ ) on the C-terminus, which may render the tetrapeptide less susceptible to protease degradation and increase their solubility compared to the free acid forms. The peptides may also be lipidated which may provide for enhanced skin penetration.

The above-described peptides may contain the following amino acids: R (arginine), L (leucine), P (proline), F (phenylalanine), Q (glutamine), E (glutamic acid), I (isoleucine), K (lysine), S (serine), V (valine), A (alanine), N (asparagine), D (aspartic acid), T (threonine), Y (tyrosine) and G (glycine). The above-described peptides do not include the following M (methionine), C (cysteine), H (histidine) or W (tryptophan). Accordingly, in one embodiment, x is not selected from either (methionine), C (cysteine), H (histidine) or W (tryptophan). Methods of Use

An additional embodiment of the invention is directed towards methods of using the above-described peptides. The

methods of use may involve the use of a single peptide, or may involve the use of two or more peptides in combination.

An embodiment of the invention is a method of promoting repair of damaged skin and maintenance of healthy skin using tetrapeptides that stimulate production of ECM proteins. The method generally is directed towards contacting dermal (skin) cells with a composition containing the peptide. The compositions can be an aerosol, emulsion, liquid, lotion, cream, paste, ointment, foam, or other pharmaceutically acceptable formulation. Generally, a pharmaceutically acceptable formulation would include any acceptable carrier suitable for use on human skin, e.g. cosmetically acceptable carrier and dermatological acceptable carrier. The compositions may contain other biologically active agents such as retinoids or other peptides. The compositions may contain pharmaceutically acceptable carriers or adjuvants. The contacting step can be performed in vivo, in situ, in vitro, or by any method known to those of skill in the art. Most preferably, the contacting step is to be performed topically at a concentration sufficient to elicit a stimulatory response. The concentration of the peptide in the composition can be about 0.01  $\mu\text{g}/\text{mL}$  to about 100  $\mu\text{g}/\text{mL}$ , about 0.1  $\mu\text{g}/\text{mL}$  to about 50  $\mu\text{g}/\text{mL}$ , and about 0.1  $\mu\text{g}/\text{mL}$  to about 1  $\mu\text{g}/\text{mL}$ . The contacting step can be performed on a mammal, a cat, a dog, a cow, a horse, a pig, or a human. A preferred composition for promoting ECM protein production comprises SEQ ID NO:8; more preferably, the composition comprises SEQ ID NO:8 in a heterogeneous mixture with at least one other tetrapeptide. In a most preferred embodiment, the individual tetrapeptides in the composition would cause sustained collagen production over a period of at least 48 hours.

An additional embodiment of the invention is directed towards a method for promoting wound healing of skin damaged by normal aging, disease, injury, trauma, or by surgery or other medical procedures. The method can comprise administering to the wound of an animal a composition, wherein the composition comprises any of the above-described peptides, singularly or in combination. The compositions can be a liquid, lotion, cream, paste, ointment, foam, or any other pharmaceutically acceptable formulation. The compositions may contain pharmaceutically acceptable carriers or adjuvants. The compositions may contain other biologically active agents such as antimicrobial agents or growth factors. The compositions may also be used in combination with other therapeutic agents such as tissue grafts, tissue culture products, oxygen or dressings. The concentration of the peptide in the composition can be about 0.01  $\mu\text{g}/\text{mL}$  to about 100  $\mu\text{g}/\text{mL}$ , about 0.1  $\mu\text{g}/\text{mL}$  to about 50  $\mu\text{g}/\text{mL}$ , and about 0.1  $\mu\text{g}/\text{mL}$  to about 1  $\mu\text{g}/\text{mL}$ . The composition can be administered to the wound topically. The animal can generally be any kind of animal, and preferably is a mammal, and more preferably is a human, cow, horse, cat, dog, pig, goat, or sheep. A preferred composition for wound healing applications in which ECM protein production is promoted comprises SEQ ID NO:8; more preferably, the composition comprises SEQ ID NO:8 in a heterogeneous mixture with at least one other tetrapeptide. In a most preferred embodiment, the individual tetrapeptides in the composition would cause sustained collagen production over a period of at least 48 hours.

An additional embodiment of the invention is directed towards a method for reducing scarring of skin damaged by normal aging, disease, injury, trauma, or by surgery or other

medical procedures. The method can comprise administering to the wound of an animal a composition, wherein the composition comprises any of the above-described peptides, singularly or in combination. The compositions can be a liquid, lotion, cream, paste, ointment, foam, or other pharmaceutically acceptable formulation. The compositions may contain pharmaceutically acceptable carriers or adjuvants. The compositions may contain other biologically active agents such as antimicrobial agents or growth factors. The compositions may also be used in combination with other therapeutic agents such as tissue grafts, tissue culture products, oxygen or dressings. The concentration of the peptide in the composition can be about 0.01 µg/mL to about 100 µg/mL, about 0.1 µg/mL to about 50 µg/mL, and about 0.1 µg/mL to about 1 µg/mL. The composition can be administered to the wound topically. The animal can generally be any kind of animal, and preferably is a mammal, and more preferably is a human, cow, horse, cat, dog, pig, goat, or sheep. A preferred composition for wound healing applications in which ECM protein production is promoted comprises SEQ ID NO:8; more preferably, the composition comprises SEQ ID NO:8 in a heterogeneous mixture with at least one other tetrapeptide. In a most preferred embodiment, the individual tetrapeptides in the composition would cause sustained collagen production over a period of at least 48 hours.

A further embodiment of the invention is directed towards a method for producing the disclosed tetrapeptides in combination. The peptides may be produced using any method known to those skilled in the art such as those disclosed in Merrifield, R. B., *Solid Phase Peptide Synthesis I*, J. AM. CHEM. SOC. 85:2149-2154 (1963); Carpino, L. A. et al., [(9-Fluorenylmethyl)Oxy]Carbonyl (Fmoc) Amino Acid Chlorides: Synthesis, Characterization, And Application To The Rapid Synthesis Of Short Peptides, J. ORG. CHEM. 37:51:3732-3734; Merrifield, R. B. et al., *Instrument For Automated Synthesis Of Peptides*, ANAL. CHEM. 38:1905-1914 (1966); or Kent, S. B. H. et al., *High Yield Chemical Synthesis Of Biologically Active Peptides On An Automated Peptide Synthesizer Of Novel Design*, IN: PEPTIDES 1984 (Ragnarsson U., ed.) Almqvist and Wiksell Int., Stockholm (Sweden), pp. 185-188, all of which are incorporated by reference herein in their entirety. Preferably, the peptides will be produced by a machine capable of sequential addition of amino acids to a growing peptide chain. However, the peptides may also be manufactured using standard solution phase methodology.

It has been observed that the addition of a mixture of free amino acids instead of homogenous peptide mixtures during peptide chain synthesis results in varied incorporation of free amino acids such that a combination of peptides results from the synthesis reactions. The relative incorporation frequency of a particular amino acid included in a mixture of two or more amino acids added during synthesis may be adjusted. Adjustment is made possible by modifying the ratio of a free

amino acid made available during the synthesis process relative to the other amino acids in the mixture (this is termed an isokinetic mixture).

The following examples are included to demonstrate preferred embodiments of the invention. It should be appreciated by those of skill in the art that the techniques disclosed in the examples which follow represent techniques discovered by the inventor to function well in the practice of the invention, and thus can be considered to constitute preferred modes for its practice. However, those of skill in the art should, in light of the present disclosure, appreciate that many changes can be made in the specific embodiments which are disclosed and still obtain a like or similar result without departing from the spirit and scope of the invention.

## EXAMPLES

### Example 1

#### Identification of Repeat Tetrapeptide Sequences in Collagen

A relatively high proportion of collagen IV tetrapeptide repeat sequences have the motif GxxG (where x is any amino acid). A number of these are shown in situ as part of the full collagen IV sequence illustrated in FIG. 1 as SEQ ID NO:45. Collagen IV was examined first due to its role of interacting with other specialized ECM components (See Gregory Schultz et al., 2005). There are eleven sequences with the GxxG motif in collagen IV that appear more than ten times (GxxG where xx is represented by: vp, ek, fp, lp, pp, sp, ep, ip, pk, qp and tp). Of these tetrapeptide sequences, eight of eleven sequences contain proline in position 3, two of eleven sequences contain P in position 2, one of eleven sequences contains proline in positions 2 and 3, and one of eleven sequences contains no proline. The disclosed sequences are referred to as REPLIKINES™. "REPLIKINE" is defined as a short sequence within ECM proteins that occurs multiple times (i.e., is replicated). This sequence may be present in one ECM protein (e.g., collagen IV). Preferably, the sequence is present in multiple ECM proteins (e.g., all collagens, elastin, laminin, etc.). The presence of the sequence in multiple ECM proteins increases the likelihood that the fragment may be able to promote ECM synthesis or repair.

The eleven GxxG sequences appearing in collagen IV listed above are highlighted in the human collagen IV sequence illustrated in FIG. 1. In this figure, all bold sequences are underlined and overlapping sequences are double-underlined. All but one of these sequences also appears in collagens I, II, III, and V. This fact contributes to the ability of the disclosed peptides to stimulate the production of all collagen types, particularly when the peptides are used in combination. Table 1 shows the frequency of several tetrapeptide repeats in ECM proteins. Bold sequences in Table 1 are those that appear in collagen IV ten or more times.

TABLE 1

Frequency of tetrapeptides in ECM proteins								
SEQ. ID NO	Sequence	Collagen I	Collagen II	Collagen III	Collagen IV	Collagen V	Elastin	Elastin Precursor
19	GAAG	10	5	7		2	4	5
20	GAKG	3	4	3	5	5		
21	GAPG	13	21	25	6	9		



TABLE 1-continued

Frequency of tetrapeptides in ECM proteins								
SEQ. ID NO	Sequence	Collagen I	Collagen II	Collagen III	Collagen IV	Collagen V	Elastin Elastin	Elastin Precursor
22	GDKG	2	2	4	9	3		
23	GDRG	2	5	2	4	1		
8	<b>GEKG</b>	3	5	4	22	15		
5	<b>GEFG</b>	11	15	10	11	4		
24	GERG	10	11	14	6	7		
2	<b>GFFG</b>	4	8	6	22	5	1	1
25	GIPG	2	2	6	14	6	5	5
26	GKDG	1	4	5	2	2		
27	GKPG	2	3	3	4	1		
28	GLKG	2	1	1	5	4		
29	GLPG	15	10	9	42	15	1	1
30	GMPG	3	5	3	2	1		
31	GPAG	16	20	20	3	6		
32	GPKG	3	11	4	12	9		
7	<b>GPPG</b>	33	40	40	46	43		
33	GPQG	7	11	9	7	5		
34	GPRG	11	13	10	4	7		
35	GPSG	10	11	5	1	5		
36	GPTG	4	3	2	2	6		
37	GPVG	9	3	3	2	5		
38	GQPG	3	4	6	12	7		
39	GRDG	4	2	3	3			
40	GRPG	3	3	4	2	5		
3	<b>GSPG</b>	4	6	21	16	3		
41	GTPG	3	4	2	11	2		
42	GVKG	1	3	2	3	1		
43	GVPG		1	3	10	1	14	15
44	GYPG	1	1	1	4	2		

As also evident from a review of the collagen IV sequence, SEQ ID NO:45, there are also many occurrences of sequences having the PxxP motif. For example, the sequence PGPP occurs no less than fifteen times as illustrated in FIG. 3. Therefore, this disclosed sequence is also referred to as a REPLIKINE™. Preferably, this sequence is present in multiple ECM proteins (e.g., all collagens, elastin, laminin, etc.) as the presence of this sequence in multiple ECM proteins increases the likelihood that the fragment may be able to promote ECM synthesis or repair. The fifteen PGPP sequences appearing in collagen IV listed above are highlighted and underlined in the human collagen IV sequence illustrated in FIG. 3.

#### Example 2

##### Identification of Frame Shift Actives

In addition to the relatively high proportion of collagen IV tetrapeptide repeat sequences with the motif GxxG, other tetrapeptide sequences occurring one amino acid frame shift away from a GxxG or PxxP tetrapeptide sequence have been identified. These sequences may repeat or occur only once within an ECM protein and may be located one amino acid position away from either a GxxG or PxxP tetrapeptide sequence as described herein. These tetrapeptide sequences are referred to as frame shift actives. Such frame shift actives may accordingly contain either a G or a P in either the second

or third position depending on the direction of frame shift. It has been further recognized that frame shift actives may be combined with other tetrapeptide sequences disclosed in this application forming a combikine. An example of such a combikine is H06 and H15.

One example of a frame shift active is GAGP or H12 (SEQ ID NO:6). H12 (GAGP) appears one residue (or frame) shift from the GxxG tetrapeptide GGAG in Collagen III (SEQ ID NO:46) as illustrated in FIG. 2. In this figure, all frame shift active sequences are bold and underlined and the GxxG sequences occurring one frame shift away are double-underlined. Furthermore, as shown in Table 5, this tetrapeptide (GAGP) achieves good results for collagen production at 48 hours. Another example is the sequence PGPR, which is H10 (SEQ ID NO:4) which occurs eleven times in Collagens I-IV. As it appears multiple times in an individual ECM protein, this tetrapeptide would further be considered a REPLIKINE. FIG. 2 (SEQ ID NO:46) illustrates several instances of this tetrapeptide with each occurring one frame shift from the GxxG tetrapeptide GPRG. This particular frame shift active appears in multiple ECM proteins and therefore increases the likelihood that the fragment may be able to promote ECM synthesis or repair.

### Example 3

#### Identification of Repeat Sequences that Stimulate Collagen Production

Several sequences identified in Examples 1 and 2 were synthesized using standard peptide chemistry and assayed for the stimulation of collagen from dermal fibroblasts. The synthesized peptides were amidated at the C-terminus, which rendered the tetrapeptides less susceptible to protease degradation and increased their solubility compared to the free acid forms. Human dermal fibroblasts were incubated in 96-well plates at 37° C. and 5% CO<sub>2</sub> for 24 and 48 hours in 150  $\mu$ L complete cell culture media (Cascade Biologics, Portland, Oreg.; Cat. No. M-106-500), supplemented with Low Serum Growth Supplement (Cascade Biologics, Portland, Oreg.; Cat. No. S-003-10) containing sample peptides at a final peptide concentration of 50  $\mu$ g/mL. Each well was seeded with 10,000 cells. Following the incubation, 100- $\mu$ L medium samples were recovered from each well and assayed for collagen production

The assays were performed by Tebu-bio Laboratories (France) using the SIRCOL™ Collagen Assay Kit (Biocolor Assays, UK) following the manufacturer's protocol. The SIRCOL™ Collagen Assay is a quantitative dye-binding method designed for the analysis of soluble collagens released into culture medium by mammalian cells during in vitro culture. The collagen of the tested samples binds to the anionic SIRCOL™ dye. The collagen-dye complexes precipitate out of solution and are pelleted by centrifugation. The recovered collagen-dye pellet was dissolved in an alkaline solution prior to absorbance measurements. Duplicate measurements were taken at the 24 and 48 hour times from two separate samples. The four measurements for each sample were averaged. The absorbance of reagent blanks, collagen standards, and samples were measured at 560 nm. The reagent blank absorbance was subtracted from the absorbance from each sample at 24 and 48 hours.

Two separate data sets were used to generate two collagen standard calibration curves. The first calibration curve was

generated for purposes of calculating the quantity of collagen in samples H6 (combination of SEQ ID NOs:1-4), H7-H14 (SEQ ID NOs:1-8, respectively) and H15 (combination of SEQ ID NOs:5-8). The second calibration curve was generated for calculating the quantity of collagen in samples H16 (SEQ ID NO:9), H21-23 (SEQ ID NOs:10-12, respectively), H25-26 (SEQ ID NOs:13-14, respectively), or H29-30 (SEQ ID NOs:15-16, respectively), H32 (SEQ ID NO:17), H33 (combination of SEQ ID NOs:9-12), H34 (combination of SEQ ID NOs:11-14), H35 (combination of SEQ ID NOs:13-16), H36 (combination of SEQ ID NOs:1, 6, 5, 8), H37 (SEQ ID NO:17) and H38 (SEQ ID NO:8) from the absorbance measurements was created by plotting the Abs<sub>560 nm</sub> of the known collagen standards versus the respective concentrations of the collagen standards (in micrograms) each time a series of assays were performed. With respect to each data set, the same calibration curve was used for samples taken at the 24 and 48 hour times (Tables 2A and 2B). Accordingly, different standard curves were prepared immediately prior to performing each series of assays.

TABLE 2A

Calibration curve for assaying collagen production by peptides H6-H15

Collagen standards ( $\mu$ g)	A <sub>560 nm</sub> 24 h test	A <sub>560 nm</sub> 48 h test
0	0.00	0.00
5	0.08	0.10
10	0.11	0.15
25	0.32	0.35
50	0.66	0.65

TABLE 2B

Calibration curve for assaying collagen production by peptides H16, H21-23, H25-26, and H29-38

Collagen Standards ( $\mu$ g)	A <sub>560 nm</sub> Assay date 1	A <sub>560 nm</sub> Assay date 2
0	0.00	0.00
5	0.12	0.09
10	0.14	0.15
25	0.48	0.42
50	0.88	0.80

A linear regression was performed from plotting the Abs<sub>560 nm</sub> values versus concentrations of the respective collagen standards using MICROSOFT EXCEL™. The regression resulted in a lines described by the formula  $y=0.013x$  for both incubation times noted in Table 2A. As the results were identical, only the 24-hour time period was used for the second series calibration curves. The formula of the line obtained on assay date 1 and assay date 2 of the second series of samples was  $y=0.0178x$  and  $y=0.0162x$ , respectively. The peptide LL-37 (SEQ ID NO:18) was used as a positive control as it has been widely reported to have an impact upon wound healing in man (Heilborn et al., The Cathelicidin Anti-Microbial Peptide LL-37 Is Involved In The Re-Epithelialization Of Human Skin Wounds And Is Lacking In Chronic Ulcer Epithelium, *J. Invest. Dermatol.* 120:379-89 (2003)). The assay detection limit defined by the manufacturer is 2.5  $\mu$ g.

The total amount of collagen produced in samples containing peptides was calculated from the averaged absorbance values taken at 24 hours (Table 3A) and 48 hours (Table 3B)

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using the linear equation derived from the standard curve. The total amount of collagen produced in samples containing peptides H16 (SEQ ID NO:9), H21-23 (SEQ ID NOs:10-12, respectively), H25-26 (SEQ ID NOs:13-14, respectively), or H29-30 (SEQ ID NOs:15-16, respectively), H32 (SEQ ID NO:17), H33 (combination of SEQ ID NOs:9-12), H34 (combination of SEQ ID NOs:11-14), H35 (combination of SEQ ID NOs:13-16), H36 (combination of SEQ ID NOs:1, 6, 5, 8), H37 (SEQ ID NO:17) and H38 (SEQ ID NO:8) was calculated from the absorbance values taken at 24 hours (Table 4A) and 48 hours (Table 4B) using the linear equation derived from the standard curve. These values were compared with peptide LL37 (SEQ ID NO:18), a peptide known to stimulate collagen. In each table, samples marked by an asterisk (\*) may not be significant as the assay detection limit is 2.5 µg.

TABLE 3A

Absorbance measurements and quantification of collagen in test samples H6-H15 at 24 hours.						
SEQ ID NO	Peptides	$A_{560\text{ nm}}$	Average	Average minus blank	Collagen (µg)	
18	LL37	0.102	0.136	0.12	0.04	3.0
—	H6	0.084	0.140	0.11	0.03	2.5
1	H7	0.098	0.063	0.08	0.00	0.0*
2	H8	0.122	0.078	0.10	0.02	1.5*
3	H9	0.147	0.104	0.13	0.05	3.5
4	H10	0.103	0.146	0.12	0.04	3.4
5	H11	0.110	0.168	0.14	0.06	4.5
6	H12	0.063	0.101	0.08	0.00	0.2*
7	H13	0.114	0.093	0.10	0.02	1.8*
8	H14	0.115	0.122	0.12	0.04	3.0
—	H15	0.132	0.093	0.11	0.03	2.5
—	Blank	0.074	0.076	0.08	0.00	0.0

TABLE 3B

Absorbance measurements and quantification of collagen in test samples H6-H15 at 48 hours.						
SEQ ID NO	Peptides	$A_{560\text{ nm}}$	Average	Average minus blank	Collagen (µg)	
18	LL37	0.262	0.113	0.19	0.07	5.2
—	H6	0.086	0.189	0.14	0.02	1.3*
1	H7	0.192	0.189	0.19	0.07	5.4
2	H8	0.137	0.126	0.13	0.01	0.9*
3	H9	0.117	0.061	0.09	0.00	0.0*
4	H10	0.136	0.085	0.11	0.00	0.0*
5	H11	0.113	0.181	0.15	0.03	2.1*
6	H12	0.106	0.231	0.17	0.05	3.7
7	H13	0.100	0.145	0.12	0.00	0.2*
8	H14	0.132	0.176	0.15	0.03	2.6
—	H15	0.177	0.174	0.18	0.06	4.3
—	Blank	0.120	0.115	0.12	0.00	0.0

TABLE 4A

Absorbance measurements and quantification of collagen in test samples H16, H21-23, H25-26, or H29-38 at 24 hours.						
SEQ ID NO	Peptides	$A_{560\text{ nm}}$	Average	Average minus blank	Collagen (µg)	
9	H16	0.133	0.137	0.14	0.06	3.1
10	H21	0.129	0.119	0.12	0.04	2.5
11	H22	0.192	0.085	0.14	0.06	3.3
12	H23	0.090	0.073	0.08	0.00	0.1*
13	H25	0.129	0.076	0.10	0.02	1.3*

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TABLE 4A-continued

Absorbance measurements and quantification of collagen in test samples H16, H21-23, H25-26, or H29-38 at 24 hours.						
SEQ ID NO	Peptides	$A_{560\text{ nm}}$	Average	Average minus blank	Collagen (µg)	
14	H26	0.114	0.149	0.13	0.05	2.9
15	H29	0.111	0.063	0.09	0.01	0.4*
16	H30	0.099	0.092	0.10	0.02	0.9*
17	H32	0.087	0.055	0.07	-0.01	-0.5*
—	(crystals and cell toxicity)					
—	H33	0.086	0.125	0.11	0.03	1.4*
—	H34	0.117	0.120	0.12	0.04	2.2*
—	H35	0.103	0.090	0.10	0.02	0.9*
—	H36	0.105	0.128	0.12	0.04	2.1*
17	H37	0.099	0.100	0.10	0.02	1.1*
8	H38	0.103	0.159	0.13	0.05	2.9
—	Blank	0.072	0.086	0.08	0.00	0.0

TABLE 4B

Absorbance measurements and quantification of collagen in test samples H16, H21-23, H25-26, or H29-38 at 48 hours.						
SEQ ID NO	Peptides	$A_{560\text{ nm}}$	Average	Average minus blank	Collagen (µg)	
9	H16	0.065	0.064	0.06	0.00	0.3*
10	H21	0.089	0.126	0.11	0.05	2.9
11	H22	0.102	0.087	0.09	0.03	2.1*
12	H23	0.093	0.082	0.09	0.03	1.7*
13	H25	0.059	0.084	0.07	0.01	0.7*
14	H26	0.081	0.153	0.12	0.06	3.5
15	H29	0.086	0.094	0.09	0.03	1.9*
16	H30	0.083	0.101	0.09	0.03	2.0*
17	H32	0.088	0.072	0.08	0.02	1.2*
—	(crystals and cell toxicity)					
—	H33	0.096	0.092	0.09	0.03	2.1*
—	H34	0.076	0.155	0.12	0.06	3.4
—	H35	0.120	0.074	0.10	0.04	2.3*
—	H36	0.154	0.082	0.12	0.06	3.6
17	H37	0.078	0.114	0.10	0.04	2.2*
8	H38	0.123	0.089	0.11	0.05	2.8
—	Blank	0.106	0.0106	0.06	0.00	0.0

Because sample sizes were 100 µL, the concentration of collagen produced in each sample in micrograms per milliliter is determined by multiplying the amount of collagen detected by ten. The results of all samples tested are summarized in Table 5.

TABLE 5

			Collagen synthesis induced by peptides		
SEQ			[Peptide]	Collage produced	
ID NO	Name	Primary sequence	( $\mu\text{g/mL}$ ).	24 hrs	48 hrs
1	H07	PEGP	50	0	54
2	H08	GFPG	50	15	9
3	H09	GSPG	50	35	0
4	H10	PGPR	50	34	0
—	H06	H7, H8, H9, H10 (SEQ ID NOs: 1, 2, 3, 4)	50	25	13
5	H11	GEPG	50	45	21
6	H12	GAGP	50	2	37
7	H13	GPPG	50	18	2
8	H14	GEKG	50	30	26
8	H38	GEKG	0.3	29	28
—	H15	H11, H12, H13, H14 (SEQ ID NOs: 5, 6, 7, 8)	50	25	43
9	H16	PEKP	50	31	3
10	H21	PKGP	50	25	29
11	H22	PGQP	50	33	21
12	H23	PGTP	50	1	17
13	H25	PMGP	50	13	7
14	H26	PGPP	50	29	35
15	H29	PQGP	50	4	19
16	H30	PGNP	50	9	20
17	H32	KTTKS (SEDERMA™ peptide)	50	na	12
17	H37	KTTKS (SEDERMA™ peptide)	0.3	11	22
—	H33	H16, H21, H22, H23 (SEQ ID NOs: 9, 10, 11, 12)	50	14	21
—	H34	H22, H23, H25, H26 (SEQ ID NOs: 11, 12, 13, 14)	50	22	34
—	H35	H25, H26, H29, H30 (SEQ ID NOs: 13, 14, 15, 16)	50	9	23
—	H36	H7, H12, H11, H14 (SEQ ID NOs: 1, 6, 5, 8)	50	21	36
18	LL37	LLGDFFRKSKEKIGKEFKRIVQRIDFLRNLPRTES	50	30	52

All tetrapeptides tested stimulated the production of soluble collagen. Of the sequences tested, GxxG tetrapeptides with a glutamic acid in position 2 best stimulate collagen at both 24 and 48 hour time-points. These sequences are H11 (GEPG; SEQ ID NO:5), H14 (GEKG; SEQ ID NO:8) and H38 (GEKG; SEQ ID NO:8). The peptides were initially screened using a peptide concentration of 50  $\mu\text{g/mL}$ . To survey the concentration effective for stimulating collagen production, H14 (SEQ ID NO:8) was also tested at 0.3  $\mu\text{g/mL}$  as H38. As shown in Table 5, H38-induced collagen stimulation was not diminished at the lower concentration, indicating that the maximal stimulating concentration of SEQ ID NO:8 is at or below 0.3  $\mu\text{g/mL}$ .

To test its efficacy, SEQ ID NO:8 (H14 and H38) was compared to the peptide, LL37, (SEQ ID NO:18) which is known to stimulate collagen production. Based on the amount of collagen released by fibroblasts in response to LL37, 25  $\mu\text{g/mL}$  was considered a significant amount of collagen released due to contact with a tetrapeptide. SEQ ID NO:8 induced about the same amount of collagen as LL37 (SEQ ID NO:18) at 24 hours. Importantly, collagen produced as a result of contact with SEQ ID NO:8 was substantially maintained for at least 48 hours. SEQ ID NO:8 was also compared to a leading skin care peptide known to stimulate collagen production, KTTKS (SEQ ID NO:17) (Katayama et. al., J. BIOL. CHEM. 288:9941-9944 (1983)). KTTKS is an ingredient in the product MATRIXYL™ (SEDERMA SAS, France). SEQ ID NO:8 stimulated more collagen production than the KTTKS (SEQ ID NO:17) peptide (Table 5) at 24 and 48 hours.

#### Example 4

##### Identification of Peptide Combinations that Synergistically Enhance Collagen Stimulation—COMBIKINES

Heterogeneous populations of active tetrapeptides may stimulate collagen production at a higher level than homogeneous samples of tetrapeptides. The components of the heterogeneous composition are called COMBIKINES™. COMBIKINES are a group of REPLIKINES combined to produce a greater or broader effect upon one or more target cell types. The peptides H11 (SEQ ID NO:5), H12 (SEQ ID NO:6), H13 (SEQ ID NO:7), and H14 (SEQ ID NO:8) were combined to a final concentration of 50  $\mu\text{g/mL}$  and assayed using the same protocol as for the individual peptides. As expected, the result obtained at the 24 hour time point equaled the mean of the individual induction scores. The combination of peptides at 48 hours, however, induced collagen to a level of 43  $\mu\text{g/mL}$ . Surprisingly, this amount was far in excess of the anticipated mean (21  $\mu\text{g/mL}$ ) of the four individual peptides (see Table 5). Thus, specific combinations of peptides may stimulate collagen production to a greater degree than the individual peptides at the same concentration. Further, tetrapeptides from a variety of ECM sources such as collagen, laminin, and elastin may produce enhanced induction of a variety of ECM proteins (see Tables 1 and 5).

#### Example 5

##### Cost-Effective COMBIKINE Manufacturing for Enhancing Stimulation of Collagen Production

The high cost of peptide synthesis limits the feasibility of producing of heterogeneous compositions of bioactive peptides. The present invention greatly mitigates this limitation.

Because the presently disclosed sequences have a commonality (e.g., a glycine or proline at both termini), a range of tetrapeptides varied at positions 2 and 3 can be synthesized in a single manufacturing run. The synthetic peptides can be made by any method known in the art. (Benoiton, N., *Chemistry of Peptide Synthesis*, CRC (2005)). During manufacture

such that more collagen is produced by 48 hours than at 24 hours. Although within the scope of the current invention, tetrapeptides that promote production of ECM proteins at 24 hours, but show diminished production at 48 hours, are less favored. In this regard, Table 6 shows the results of the currently disclosed peptides. Preferred peptides are in bold.

TABLE 6

SEQ ID		Released collagen	Released collagen	Significant release	Increase in collagen	Decrease in collagen
NO	Peptides	( $\mu\text{g/mL}$ ) 24 h	( $\mu\text{g/mL}$ ) 48 h	of collagen at 24 h and 48 h	release at 48 h v. 24 h	release at 48 h v. 24 h
18	LL37	30	52	✓	✓	
—	H6	25	13			
1	H7	0	54		✓	
2	H8	15	9			
3	H9	35	0			✓
4	H10	34	0			✓
5	H11	45	21			✓
6	H12	2	37		✓	
7	H13	18	2			
<b>8</b>	<b>H14</b>	<b>30</b>	<b>26</b>	✓		
<b>8</b>	<b>H38</b>	<b>29</b>	<b>28</b>	✓		
—	<b>H15</b>	<b>25</b>	<b>43</b>	✓	✓	
9	H16	31	3			✓
<b>10</b>	<b>H21</b>	<b>25</b>	<b>29</b>	✓		
11	H22	33	21			✓
12	H23	1	17		✓	
13	H25	13	7			✓
<b>14</b>	<b>H26</b>	<b>29</b>	<b>35</b>	✓		
15	H29	4	19		✓	
16	H30	9	20		✓	
17	H32	NA	12			
	(crystals and cell toxicity)					
17	H37	11	22		✓	
—	H33	14	21		✓	
—	H34	22	34		✓	
—	H35	9	23		✓	
—	H36	21	36		✓	

of the peptides, amino acid mixtures are added instead of homogenous samples. The chemistry for determining the correct ratios of amino acid concentrations added at the mixed positions to gain the desired ratio of resulting peptides has been described previously (Greenbaum et al., *Molecular and Cellular Proteomics* 1:60-68, 2002; Krstenansky et al., *Letters in Drug Design and Discovery* 1:6-13, 2004; both of which references are incorporated herein in their entirety). Using this methodology, a library of heterogeneous peptides can be made for nearly the same cost of synthesizing one peptide.

The application of this manufacturing process enables the cost-effective production of bioactive combikines. This is made possible by the unique composition of the disclosed tetrapeptides. The tetrapeptide mixtures are better suited for incorporation into topical use formulations than longer peptides. Because of their length, tetrapeptides have practical and chemical advantages over longer peptides, including the following: easier incorporation and dissolution into formulations, higher skin and pore permeability, and higher production yields with easier methods of manufacturing combinations of peptides. Although not required, the ideal formulations of tetrapeptides, singly or in combination, are formulations that maintain significant collagen production at 24 hours for up to 48 hours. More preferably, the formulations would induce synthesis of ECM for the entire 48 hour period

#### Example 6

##### Collagen Stimulators Also Serve as Multi-Effector Molecules Enhancing Skin Epithelial Cell Wound Closer

Collagens are key components of all phases of wound healing. Stimulation of collagen production reflects that damage has occurred to the collagen network (e.g. by enzymes or physical destruction). Indeed, the total collapse of the collagen network in fact causes healing to take place. Therefore a collagen stimulator may also serve as a multi-effector molecule orchestrating certain matrix remodeling and enhancing wound healing.

Wound healing experiments were performed on monolayers of human skin epithelial cells (CRL-2592) plated onto 12-well plates. Cells were serum-starved for 24 hours before experimentation. Confluent monolayers of CRL-2592 were wounded using a P200 (200- $\mu\text{L}$ ) pipette tip. The wounds were washed and picture-documented prior to peptide treatment. Peptides were added to a final concentration from 20 to 40  $\mu\text{g/ml}$ . Cells were kept in an incubator at 37° C., 5% CO<sub>2</sub>, and 92% humidity, except when images were being captured for a short period at room temperature. Wound closure was followed at 6-hour and 10-hour time points. PBS-treated wounds were used as negative controls for comparison purposes.

TABLE 7

Effect of peptides on human skin epithelial wound closure in vitro					
Compound	0 hr	6 hr		10 hr	
	W-size*	W-size	% closure	W-size	% closure
PBS-1	36	29	19.40%	21	41.70%
PBS-2	52	42	19.20%	30	42.30%
SEQ ID NO: 14	25	12	52%	2.75	89%
SEQ ID NO: 5	48	39	19%	30	37.50%

\*W-size: wound size (arbitrary)

In vitro monolayer wound closure is a result of cell migration, which is important in many biological processes such as embryogenesis, angiogenesis, inflammatory reactions and wound repair. These processes are thought to be regulated by interactions with other cells, cytokines and ECM proteins. As shown in Table 7, SEQ ID NO:14 significantly induces wound closure compared to the effects of PBS alone. Such activity is peptide-specific as well as cell type-specific since SEQ ID NO:14 does not induce wound closure in a human skin fibroblast monolayer (data not shown). SEQ ID NO:5 is also a collagen inducer, but does not enhance wound closure

or epithelial cell migration to any great extent compared to the effects of PBS alone. The fact that SEQ ID NO:14 induced cell migration or wound closure in a manner specific to skin epithelial cells (i.e. does not recruit fibroblasts) may add an advantage to using this peptide for skin care, since it is believed that the recruitment of large numbers of active fibroblasts to a wound site results in excess deposition and contraction of tissue resulting in scarring.

All of the compositions or methods disclosed and claimed herein can be made and executed without undue experimentation in light of the present disclosure. While the compositions and methods of this invention have been described in terms of preferred embodiments, it will be apparent to those of skill in the art that variations may be applied to the compositions and/or methods and in the steps or in the sequence of steps of the methods described herein without departing from the concept, spirit and scope of the invention. More specifically, it will be apparent that certain agents which are both chemically and physiologically related may be substituted for the agents described herein while the same or similar results would be achieved. All such similar substitutes and modifications apparent to those skilled in the art are deemed to be within the spirit, scope and concept of the invention.

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&lt;220&gt; FEATURE:

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Arg Thr Glu Ser  
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Gly Ser Gly Cys Gly Lys Cys Asp Cys His Gly Val Lys Gly Gln Lys  
 35 40 45

Gly Glu Arg Gly Leu Pro Gly Leu Gln Gly Val Ile Gly Phe Pro Gly  
 50 55 60

Met Gln Gly Pro Glu Gly Pro Gln Gly Pro Pro Gly Gln Lys Gly Asp  
 65 70 75 80

Thr Gly Glu Pro Gly Leu Pro Gly Thr Lys Gly Thr Arg Gly Pro Pro  
 85 90 95

Gly Ala Ser Gly Tyr Pro Gly Asn Pro Gly Leu Pro Gly Ile Pro Gly  
 100 105 110

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 Lys Gly Glu Arg Gly Pro Leu Gly Pro Pro Gly Leu Pro Gly Phe Ala  
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 145 150 155 160  
 Glu Ile Leu Gly His Val Pro Gly Met Leu Leu Lys Gly Glu Arg Gly  
 165 170 175  
 Phe Pro Gly Ile Pro Gly Thr Pro Gly Pro Pro Gly Leu Pro Gly Leu  
 180 185 190  
 Gln Gly Pro Val Gly Pro Pro Gly Phe Thr Gly Pro Pro Gly Pro Pro  
 195 200 205  
 Gly Pro Pro Gly Pro Pro Gly Glu Lys Gly Gln Met Gly Leu Ser Phe  
 210 215 220  
 Gln Gly Pro Lys Gly Asp Lys Gly Asp Gln Gly Val Ser Gly Pro Pro  
 225 230 235 240  
 Gly Val Pro Gly Gln Ala Gln Val Gln Glu Lys Gly Asp Phe Ala Thr  
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 Lys Gly Glu Lys Gly Gln Lys Gly Glu Pro Gly Phe Gln Gly Met Pro  
 260 265 270  
 Gly Val Gly Glu Lys Gly Glu Pro Gly Lys Pro Gly Pro Arg Gly Lys  
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 Glu Lys Gly Glu Ala Gly Pro Pro Gly Pro Pro Gly Ile Val Ile Gly  
 325 330 335  
 Thr Gly Pro Leu Gly Glu Lys Gly Glu Arg Gly Tyr Pro Gly Thr Pro  
 340 345 350  
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 355 360 365  
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 405 410 415  
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 420 425 430  
 Glu Cys Gln Pro Gly Pro Pro Gly Asp Gln Gly Pro Pro Gly Ile Pro  
 435 440 445  
 Gly Gln Pro Gly Phe Ile Gly Glu Ile Gly Glu Lys Gly Gln Lys Gly  
 450 455 460  
 Glu Ser Cys Leu Ile Cys Asp Ile Asp Gly Tyr Arg Gly Pro Pro Gly  
 465 470 475 480  
 Pro Gln Gly Pro Pro Gly Glu Ile Gly Phe Pro Gly Gln Pro Gly Ala  
 485 490 495  
 Lys Gly Asp Arg Gly Leu Pro Gly Arg Asp Gly Val Ala Gly Val Pro  
 500 505 510  
 Gly Pro Gln Gly Thr Pro Gly Leu Ile Gly Gln Pro Gly Ala Lys Gly  
 515 520 525  
 Glu Pro Gly Glu Phe Tyr Phe Asp Leu Arg Leu Lys Gly Asp Lys Gly

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Gly	Ser	Val	Gly 580	Leu	Lys	Gly	Glu	Arg 585	Gly	Pro	Pro	Gly	Gly	Val	Gly
Phe	Pro	Gly 595	Ser	Arg	Gly	Asp	Thr 600	Gly	Pro	Pro	Gly	Pro	Pro	Gly	Tyr
Gly 610	Pro	Ala	Gly	Pro	Ile	Gly 615	Asp	Lys	Gly	Gln	Ala	Gly	Phe	Pro	Gly
Gly 625	Pro	Gly	Ser	Pro	Gly 630	Leu	Pro	Gly	Pro	Lys 635	Gly	Glu	Pro	Gly	Lys 640
Ile	Val	Pro	Leu	Pro 645	Gly	Pro	Pro	Gly	Ala	Glu	Gly	Leu	Pro	Gly	Ser 655
Pro	Gly	Phe 660	Pro	Gly	Pro	Gln	Gly	Asp 665	Arg	Gly	Phe	Pro	Gly	Thr	Pro
Gly	Arg	Pro 675	Gly	Leu	Pro	Gly	Glu	Lys 680	Gly	Ala	Val	Gly	Gln	Pro	Gly
Ile	Gly 690	Phe	Pro	Gly	Pro	Pro	Gly	Pro	Lys	Gly	Val	Asp	Gly	Leu	Pro
Gly 705	Asp	Met	Gly	Pro	Pro 710	Gly	Thr	Pro	Gly	Arg 715	Pro	Gly	Phe	Asn	Gly 720
Leu	Pro	Gly	Asn 725	Pro	Gly	Val	Gln	Gly	Gln 730	Lys	Gly	Glu	Pro	Gly	Val 735
Gly	Leu	Pro	Gly 740	Leu	Lys	Gly	Leu	Pro	Gly	Leu	Pro	Gly	Ile	Pro	Gly
Thr	Pro	Gly 755	Glu	Lys	Gly	Ser	Ile 760	Gly	Val	Pro	Gly	Val	Pro	Gly	Glu
His	Gly 770	Ala	Ile	Gly	Pro	Pro	Gly	Leu	Gln	Gly	Ile	Arg	Gly	Glu	Pro
Gly 785	Pro	Pro	Gly	Leu	Pro	Gly	Ser	Val	Gly	Ser 795	Pro	Gly	Val	Pro	Gly 800
Ile	Gly	Pro	Pro	Gly 805	Ala	Arg	Gly	Pro	Pro	Gly	Gly	Gln	Gly	Pro	Pro
Gly	Leu	Ser	Gly 820	Pro	Pro	Gly	Ile	Lys 825	Gly	Glu	Lys	Gly	Phe	Pro	Gly
Phe	Pro	Gly 835	Leu	Asp	Met	Pro	Gly	Pro	Lys	Gly	Asp	Lys	Gly	Ala	Gln
Gly 850	Leu	Pro	Gly	Ile	Thr	Gly	Gln	Ser	Gly	Leu	Pro	Gly	Leu	Pro	Gly
Gln 865	Gln	Gly	Ala	Pro	Gly 870	Ile	Pro	Gly	Phe	Pro	Gly	Ser	Lys	Gly	Glu 880
Met	Gly	Val	Met	Gly 885	Thr	Pro	Gly	Gln	Pro	Gly	Ser	Pro	Gly	Pro	Val 895
Gly	Ala	Pro	Gly 900	Leu	Pro	Gly	Glu	Lys	Gly	Asp	His	Gly	Phe	Pro	Gly
Ser	Ser	Gly 915	Pro	Arg	Gly	Asp	Pro	Gly	Leu	Lys	Gly	Asp	Lys	Gly	Asp
Val	Gly	Leu	Pro	Gly	Lys	Pro	Gly	Ser	Met	Asp	Lys	Val	Asp	Met	Gly
Ser 945	Met	Lys	Gly	Gln	Lys	Gly	Asp	Gln	Gly	Glu	Lys	Gly	Gln	Ile	Gly 960

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Pro Ile Gly Glu Lys Gly Ser Arg Gly Asp Pro Gly Thr Pro Gly Val  
                   965                                  970                                  975

Pro Gly Lys Asp Gly Gln Ala Gly Gln Pro Gly Gln Pro Gly Pro Lys  
                   980                                  985                                  990

Gly Asp Pro Gly Ile Ser Gly Thr Pro Gly Ala Pro Gly Leu Pro Gly  
                   995                                  1000                                  1005

Pro Lys Gly Ser Val Gly Gly Met Gly Leu Pro Gly Thr Pro Gly  
   1010                                  1015                                  1020

Glu Lys Gly Val Pro Gly Ile Pro Gly Pro Gln Gly Ser Pro Gly  
   1025                                  1030                                  1035

Leu Pro Gly Asp Lys Gly Ala Lys Gly Glu Lys Gly Gln Ala Gly  
   1040                                  1045                                  1050

Pro Pro Gly Ile Gly Ile Pro Gly Leu Arg Gly Glu Lys Gly Asp  
   1055                                  1060                                  1065

Gln Gly Ile Ala Gly Phe Pro Gly Ser Pro Gly Glu Lys Gly Glu  
   1070                                  1075                                  1080

Lys Gly Ser Ile Gly Ile Pro Gly Met Pro Gly Ser Pro Gly Leu  
   1085                                  1090                                  1095

Lys Gly Ser Pro Gly Ser Val Gly Tyr Pro Gly Ser Pro Gly Leu  
   1100                                  1105                                  1110

Pro Gly Glu Lys Gly Asp Lys Gly Leu Pro Gly Leu Asp Gly Ile  
   1115                                  1120                                  1125

Pro Gly Val Lys Gly Glu Ala Gly Leu Pro Gly Thr Pro Gly Pro  
   1130                                  1135                                  1140

Thr Gly Pro Ala Gly Gln Lys Gly Glu Pro Gly Ser Asp Gly Ile  
   1145                                  1150                                  1155

Pro Gly Ser Ala Gly Glu Lys Gly Glu Pro Gly Leu Pro Gly Arg  
   1160                                  1165                                  1170

Gly Phe Pro Gly Phe Pro Gly Ala Lys Gly Asp Lys Gly Ser Lys  
   1175                                  1180                                  1185

Gly Glu Val Gly Phe Pro Gly Leu Ala Gly Ser Pro Gly Ile Pro  
   1190                                  1195                                  1200

Gly Ser Lys Gly Glu Gln Gly Phe Met Gly Pro Pro Gly Pro Gln  
   1205                                  1210                                  1215

Gly Gln Pro Gly Leu Pro Gly Ser Pro Gly His Ala Thr Glu Gly  
   1220                                  1225                                  1230

Pro Lys Gly Asp Arg Gly Pro Gln Gly Gln Pro Gly Leu Pro Gly  
   1235                                  1240                                  1245

Leu Pro Gly Pro Met Gly Pro Pro Gly Leu Pro Gly Ile Asp Gly  
   1250                                  1255                                  1260

Val Lys Gly Asp Lys Gly Asn Pro Gly Trp Pro Gly Ala Pro Gly  
   1265                                  1270                                  1275

Val Pro Gly Pro Lys Gly Asp Pro Gly Phe Gln Gly Met Pro Gly  
   1280                                  1285                                  1290

Ile Gly Gly Ser Pro Gly Ile Thr Gly Ser Lys Gly Asp Met Gly  
   1295                                  1300                                  1305

Pro Pro Gly Val Pro Gly Phe Gln Gly Pro Lys Gly Leu Pro Gly  
   1310                                  1315                                  1320

Leu Gln Gly Ile Lys Gly Asp Gln Gly Asp Gln Gly Val Pro Gly  
   1325                                  1330                                  1335

Ala Lys Gly Leu Pro Gly Pro Pro Gly Pro Pro Gly Pro Tyr Asp  
   1340                                  1345                                  1350

Ile Ile Lys Gly Glu Pro Gly Leu Pro Gly Pro Glu Gly Pro Pro  
   1355                                  1360                                  1365



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Gly Leu Lys Gly Leu Gln Gly Leu Pro Gly Pro Lys Gly Gln Gln  
 1370 1375 1380  
 Gly Val Thr Gly Leu Val Gly Ile Pro Gly Pro Pro Gly Ile Pro  
 1385 1390 1395  
 Gly Phe Asp Gly Ala Pro Gly Gln Lys Gly Glu Met Gly Pro Ala  
 1400 1405 1410  
 Gly Pro Thr Gly Pro Arg Gly Phe Pro Gly Pro Pro Gly Pro Asp  
 1415 1420 1425  
 Gly Leu Pro Gly Ser Met Gly Pro Pro Gly Thr Pro Ser Val Asp  
 1430 1435 1440  
 His Gly Phe Leu Val Thr Arg His Ser Gln Thr Ile Asp Asp Pro  
 1445 1450 1455  
 Gln Cys Pro Ser Gly Thr Lys Ile Leu Tyr His Gly Tyr Ser Leu  
 1460 1465 1470  
 Leu Tyr Val Gln Gly Asn Glu Arg Ala His Gly Gln Asp Leu Gly  
 1475 1480 1485  
 Thr Ala Gly Ser Cys Leu Arg Lys Phe Ser Thr Met Pro Phe Leu  
 1490 1495 1500  
 Phe Cys Asn Ile Asn Asn Val Cys Asn Phe Ala Ser Arg Asn Asp  
 1505 1510 1515  
 Tyr Ser Tyr Trp Leu Ser Thr Pro Glu Pro Met Pro Met Ser Met  
 1520 1525 1530  
 Ala Pro Ile Thr Gly Glu Asn Ile Arg Pro Phe Ile Ser Arg Cys  
 1535 1540 1545  
 Ala Val Cys Glu Ala Pro Ala Met Val Met Ala Val His Ser Gln  
 1550 1555 1560  
 Thr Ile Gln Ile Pro Pro Cys Pro Ser Gly Trp Ser Ser Leu Trp  
 1565 1570 1575  
 Ile Gly Tyr Ser Phe Val Met His Thr Ser Ala Gly Ala Glu Gly  
 1580 1585 1590  
 Ser Gly Gln Ala Leu Ala Ser Pro Gly Ser Cys Leu Glu Glu Phe  
 1595 1600 1605  
 Arg Ser Ala Pro Phe Ile Glu Cys His Gly Arg Gly Thr Cys Asn  
 1610 1615 1620  
 Tyr Tyr Ala Asn Ala Tyr Ser Phe Trp Leu Ala Thr Ile Glu Arg  
 1625 1630 1635  
 Ser Glu Met Phe Lys Lys Pro Thr Pro Ser Thr Leu Lys Ala Gly  
 1640 1645 1650  
 Glu Leu Arg Thr His Val Ser Arg Cys Gln Val Cys Met Arg Arg  
 1655 1660 1665

Thr

&lt;210&gt; SEQ ID NO 46

&lt;211&gt; LENGTH: 1466

&lt;212&gt; TYPE: PRT

&lt;213&gt; ORGANISM: Homo sapiens

&lt;400&gt; SEQUENCE: 46

Met Met Ser Phe Val Gln Lys Gly Ser Trp Leu Leu Leu Ala Leu Leu  
 1 5 10 15

His Pro Thr Ile Ile Leu Ala Gln Gln Glu Ala Val Glu Gly Gly Cys  
 20 25 30

Ser His Leu Gly Gln Ser Tyr Ala Asp Arg Asp Val Trp Lys Pro Glu  
 35 40 45

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Pro Cys Gln Ile Cys Val Cys Asp Ser Gly Ser Val Leu Cys Asp Asp  
 50 55 60

Ile Ile Cys Asp Asp Gln Glu Leu Asp Cys Pro Asn Pro Glu Ile Pro  
 65 70 75 80

Phe Gly Glu Cys Cys Ala Val Cys Pro Gln Pro Pro Thr Ala Pro Thr  
 85 90 95

Arg Pro Pro Asn Gly Gln Gly Pro Gln Gly Pro Lys Gly Asp Pro Gly  
 100 105 110

Pro Pro Gly Ile Pro Gly Arg Asn Gly Asp Pro Gly Ile Pro Gly Gln  
 115 120 125

Pro Gly Ser Pro Gly Ser Pro Gly Pro Pro Gly Ile Cys Glu Ser Cys  
 130 135 140

Pro Thr Gly Pro Gln Asn Tyr Ser Pro Gln Tyr Asp Ser Tyr Asp Val  
 145 150 155 160

Lys Ser Gly Val Ala Val Gly Gly Leu Ala Gly Tyr Pro Gly Pro Ala  
 165 170 175

Gly Pro Pro Gly Pro Pro Gly Pro Pro Gly Thr Ser Gly His Pro Gly  
 180 185 190

Ser Pro Gly Ser Pro Gly Tyr Gln Gly Pro Pro Gly Glu Pro Gly Gln  
 195 200 205

Ala Gly Pro Ser Gly Pro Pro Gly Pro Pro Gly Ala Ile Gly Pro Ser  
 210 215 220

Gly Pro Ala Gly Lys Asp Gly Glu Ser Gly Arg Pro Gly Arg Pro Gly  
 225 230 235 240

Glu Arg Gly Leu Pro Gly Pro Pro Gly Ile Lys Gly Pro Ala Gly Ile  
 245 250 255

Pro Gly Phe Pro Gly Met Lys Gly His Arg Gly Phe Asp Gly Arg Asn  
 260 265 270

Gly Glu Lys Gly Glu Thr Gly Ala Pro Gly Leu Lys Gly Glu Asn Gly  
 275 280 285

Leu Pro Gly Glu Asn Gly Ala Pro Gly Pro Met Gly Pro Arg Gly Ala  
 290 295 300

Pro Gly Glu Arg Gly Arg Pro Gly Leu Pro Gly Ala Ala Gly Ala Arg  
 305 310 315 320

Gly Asn Asp Gly Ala Arg Gly Ser Asp Gly Gln Pro Gly Pro Pro Gly  
 325 330 335

Pro Pro Gly Thr Ala Gly Phe Pro Gly Ser Pro Gly Ala Lys Gly Glu  
 340 345 350

Val Gly Pro Ala Gly Ser Pro Gly Ser Asn Gly Ala Pro Gly Gln Arg  
 355 360 365

Gly Glu Pro Gly Pro Gln Gly His Ala Gly Ala Gln Gly Pro Pro Gly  
 370 375 380

Pro Pro Gly Ile Asn Gly Ser Pro Gly Gly Lys Gly Glu Met Gly Pro  
 385 390 395 400

Ala Gly Ile Pro Gly Ala Pro Gly Leu Met Gly Ala Arg Gly Pro Pro  
 405 410 415

Gly Pro Ala Gly Ala Asn Gly Ala Pro Gly Leu Arg Gly Gly Ala Gly  
 420 425 430

Glu Pro Gly Lys Asn Gly Ala Lys Gly Glu Pro Gly Pro Arg Gly Glu  
 435 440 445

Arg Gly Glu Ala Gly Ile Pro Gly Val Pro Gly Ala Lys Gly Glu Asp  
 450 455 460

Gly Lys Asp Gly Ser Pro Gly Glu Pro Gly Ala Asn Gly Leu Pro Gly  
 465 470 475 480

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Ala Ala Gly Glu Arg Gly Ala Pro Gly Phe Arg Gly Pro Ala Gly Pro  
485 490 495

Asn Gly Ile Pro Gly Glu Lys Gly Pro Ala Gly Glu Arg Gly Ala Pro  
500 505 510

Gly Pro Ala Gly Pro Arg Gly Ala Ala Gly Glu Pro Gly Arg Asp Gly  
515 520 525

Val Pro Gly Gly Pro Gly Met Arg Gly Met Pro Gly Ser Pro Gly Gly  
530 535 540

Pro Gly Ser Asp Gly Lys Pro Gly Pro Pro Gly Ser Gln Gly Glu Ser  
545 550 555 560

Gly Arg Pro Gly Pro Pro Gly Pro Ser Gly Pro Arg Gly Gln Pro Gly  
565 570 575

Val Met Gly Phe Pro Gly Pro Lys Gly Asn Asp Gly Ala Pro Gly Lys  
580 585 590

Asn Gly Glu Arg Gly Gly Pro Gly Gly Pro Gly Pro Gln Gly Pro Pro  
595 600 605

Gly Lys Asn Gly Glu Thr Gly Pro Gln Gly Pro Pro Gly Pro Thr Gly  
610 615 620

Pro Gly Gly Asp Lys Gly Asp Thr Gly Pro Pro Gly Pro Gln Gly Leu  
625 630 635 640

Gln Gly Leu Pro Gly Thr Gly Gly Pro Pro Gly Glu Asn Gly Lys Pro  
645 650 655

Gly Glu Pro Gly Pro Lys Gly Asp Ala Gly Ala Pro Gly Ala Pro Gly  
660 665 670

Gly Lys Gly Asp Ala Gly Ala Pro Gly Glu Arg Gly Pro Pro Gly Leu  
675 680 685

Ala Gly Ala Pro Gly Leu Arg Gly Gly Ala Gly Pro Pro Gly Pro Glu  
690 695 700

Gly Gly Lys Gly Ala Ala Gly Pro Pro Gly Pro Pro Gly Ala Ala Gly  
705 710 715 720

Thr Pro Gly Leu Gln Gly Met Pro Gly Glu Arg Gly Gly Leu Gly Ser  
725 730 735

Pro Gly Pro Lys Gly Asp Lys Gly Glu Pro Gly Gly Pro Gly Ala Asp  
740 745 750

Gly Val Pro Gly Lys Asp Gly Pro Arg Gly Pro Thr Gly Pro Ile Gly  
755 760 765

Pro Pro Gly Pro Ala Gly Gln Pro Gly Asp Lys Gly Glu Gly Gly Ala  
770 775 780

Pro Gly Leu Pro Gly Ile Ala Gly Pro Arg Gly Ser Pro Gly Glu Arg  
785 790 795 800

Gly Glu Thr Gly Pro Pro Gly Pro Ala Gly Phe Pro Gly Ala Pro Gly  
805 810 815

Gln Asn Gly Glu Pro Gly Gly Lys Gly Glu Arg Gly Ala Pro Gly Glu  
820 825 830

Lys Gly Glu Gly Gly Pro Pro Gly Val Ala Gly Pro Pro Gly Gly Ser  
835 840 845

Gly Pro Ala Gly Pro Pro Gly Pro Gln Gly Val Lys Gly Glu Arg Gly  
850 855 860

Ser Pro Gly Gly Pro Gly Ala Ala Gly Phe Pro Gly Ala Arg Gly Leu  
865 870 875 880

Pro Gly Pro Pro Gly Ser Asn Gly Asn Pro Gly Pro Pro Gly Pro Ser  
885 890 895

Gly Ser Pro Gly Lys Asp Gly Pro Pro Gly Pro Ala Gly Asn Thr Gly

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900				905				910							
Ala	Pro	Gly	Ser	Pro	Gly	Val	Ser	Gly	Pro	Lys	Gly	Asp	Ala	Gly	Gln
		915					920					925			
Pro	Gly	Glu	Lys	Gly	Ser	Pro	Gly	Ala	Gln	Gly	Pro	Pro	Gly	Ala	Pro
		930				935					940				
Gly	Pro	Leu	Gly	Ile	Ala	Gly	Ile	Thr	Gly	Ala	Arg	Gly	Leu	Ala	Gly
		945			950					955					960
Pro	Pro	Gly	Met	Pro	Gly	Pro	Arg	Gly	Ser	Pro	Gly	Pro	Gln	Gly	Val
			965						970					975	
Lys	Gly	Glu	Ser	Gly	Lys	Pro	Gly	Ala	Asn	Gly	Leu	Ser	Gly	Glu	Arg
			980						985					990	
Gly	Pro	Pro	Gly	Pro	Gln	Gly	Leu	Pro	Gly	Leu	Ala	Gly	Thr	Ala	Gly
		995					1000							1005	
Glu	Pro	Gly	Arg	Asp	Gly	Asn	Pro	Gly	Ser	Asp	Gly	Leu	Pro	Gly	
		1010				1015					1020				
Arg	Asp	Gly	Ser	Pro	Gly	Gly	Lys	Gly	Asp	Arg	Gly	Glu	Asn	Gly	
		1025				1030					1035				
Ser	Pro	Gly	Ala	Pro	Gly	Ala	Pro	Gly	His	Pro	Gly	Pro	Pro	Gly	
		1040				1045					1050				
Pro	Val	Gly	Pro	Ala	Gly	Lys	Ser	Gly	Asp	Arg	Gly	Glu	Ser	Gly	
		1055				1060					1065				
Pro	Ala	Gly	Pro	Ala	Gly	Ala	Pro	Gly	Pro	Ala	Gly	Ser	Arg	Gly	
		1070				1075					1080				
Ala	Pro	Gly	Pro	Gln	Gly	Pro	Arg	Gly	Asp	Lys	Gly	Glu	Thr	Gly	
		1085				1090					1095				
Glu	Arg	Gly	Ala	Ala	Gly	Ile	Lys	Gly	His	Arg	Gly	Phe	Pro	Gly	
		1100				1105					1110				
Asn	Pro	Gly	Ala	Pro	Gly	Ser	Pro	Gly	Pro	Ala	Gly	Gln	Gln	Gly	
		1115				1120					1125				
Ala	Ile	Gly	Ser	Pro	Gly	Pro	Ala	Gly	Pro	Arg	Gly	Pro	Val	Gly	
		1130				1135					1140				
Pro	Ser	Gly	Pro	Pro	Gly	Lys	Asp	Gly	Thr	Ser	Gly	His	Pro	Gly	
		1145				1150					1155				
Pro	Ile	Gly	Pro	Pro	Gly	Pro	Arg	Gly	Asn	Arg	Gly	Glu	Arg	Gly	
		1160				1165					1170				
Ser	Glu	Gly	Ser	Pro	Gly	His	Pro	Gly	Gln	Pro	Gly	Pro	Pro	Gly	
		1175				1180					1185				
Pro	Pro	Gly	Ala	Pro	Gly	Pro	Cys	Cys	Gly	Gly	Val	Gly	Ala	Ala	
		1190				1195					1200				
Ala	Ile	Ala	Gly	Ile	Gly	Gly	Glu	Lys	Ala	Gly	Gly	Phe	Ala	Pro	
		1205				1210					1215				
Tyr	Tyr	Gly	Asp	Glu	Pro	Met	Asp	Phe	Lys	Ile	Asn	Thr	Asp	Glu	
		1220				1225					1230				
Ile	Met	Thr	Ser	Leu	Lys	Ser	Val	Asn	Gly	Gln	Ile	Glu	Ser	Leu	
		1235				1240					1245				
Ile	Ser	Pro	Asp	Gly	Ser	Arg	Lys	Asn	Pro	Ala	Arg	Asn	Cys	Arg	
		1250				1255					1260				
Asp	Leu	Lys	Phe	Cys	His	Pro	Glu	Leu	Lys	Ser	Gly	Glu	Tyr	Trp	
		1265				1270					1275				
Val	Asp	Pro	Asn	Gln	Gly	Cys	Lys	Leu	Asp	Ala	Ile	Lys	Val	Phe	
		1280				1285					1290				
Cys	Asn	Met	Glu	Thr	Gly	Glu	Thr	Cys	Ile	Ser	Ala	Asn	Pro	Leu	
		1295				1300					1305				

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Asn Val	Pro Arg	Lys His	Trp	Trp Thr	Asp Ser	Ser	Ala Glu	Lys
1310			1315			1320		
Lys His	Val Trp	Phe Gly	Glu	Ser Met	Asp Gly	Gly	Phe Gln	Phe
1325			1330			1335		
Ser Tyr	Gly Asn	Pro Glu	Leu	Pro Glu	Asp Val	Leu	Asp Val	Gln
1340			1345			1350		
Leu Ala	Phe Leu	Arg Leu	Leu	Ser Ser	Arg Ala	Ser	Gln Asn	Ile
1355			1360			1365		
Thr Tyr	His Cys	Lys Asn	Ser	Ile Ala	Tyr Met	Asp	Gln Ala	Ser
1370			1375			1380		
Gly Asn	Val Lys	Lys Ala	Leu	Lys Leu	Met Gly	Ser	Asn Glu	Gly
1385			1390			1395		
Glu Phe	Lys Ala	Glu Gly	Asn	Ser Lys	Phe Thr	Tyr	Thr Val	Leu
1400			1405			1410		
Glu Asp	Gly Cys	Thr Lys	His	Thr Gly	Glu Trp	Ser	Lys Thr	Val
1415			1420			1425		
Phe Glu	Tyr Arg	Thr Arg	Lys	Ala Val	Arg Leu	Pro	Ile Val	Asp
1430			1435			1440		
Ile Ala	Pro Tyr	Asp Ile	Gly	Gly Pro	Asp Gln	Glu	Phe Gly	Val
1445			1450			1455		
Asp Val	Gly Pro	Val Cys	Phe	Leu				
1460			1465					

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What is claimed is:

1. A tetrapeptide capable of inducing production of extracellular matrix proteins, wherein the amino acid sequence of the tetrapeptide consists of SEQ ID NO:9, SEQ ID NO:10, SEQ ID NO:11, SEQ ID NO:12, or SEQ ID NO:16 having an acid or amidated carboxy-terminus.

2. A composition comprising at least one tetrapeptide of claim 1 and a pharmaceutically acceptable carrier.

3. The composition of claim 2, wherein the tetrapeptide is present in a concentration ranging from about 0.1 µg/mL to about 50 µg/mL.

4. The composition of claim 2, wherein the composition is in the form of an aerosol, emulsion, liquid, lotion, cream, paste, ointment, or foam.

5. A method for stimulating the production of collagen in a human in need thereof, the method comprising administering to said human a therapeutically effective amount of the composition of claim 2.

6. The method of claim 5, wherein the therapeutically effective concentration is in the range of about 0.1 µg/mL to about 50 µg/mL of tetrapeptide.

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7. The method of claim 5, wherein the administering to said human a therapeutically effective amount of the composition promotes wound healing of damaged skin.

8. The tetrapeptide of claim 1, wherein the tetrapeptide is SEQ ID NO:10.

9. The tetrapeptide of claim 1, wherein the tetrapeptide is SEQ ID NO:9.

10. The method of claim 7, wherein said damaged skin is a result of aging, disease, injury, trauma, or surgery.

11. The composition of claim 2 which further comprises a retinoid.

12. The method of claim 7 wherein the composition is administered topically to a site of damaged skin.

13. A skincare composition comprising at least one tetrapeptide of claim 1 and a dermatological acceptable carrier.

14. The skincare composition of claim 13, wherein the composition is in the form of an aerosol, emulsion, lotion, cream, paste, ointment, or foam.

\* \* \* \* \*