SYSTEM, OPTODE AND CAP FOR NEAR-INFRARED DIFFUSE-OPTICAL FUNCTION NEUROIMAGING

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(56) References Cited

U.S. PATENT DOCUMENTS


OTHER PUBLICATIONS


* cited by examiner
Figure 1

Figure 2
PRIOR ART

FIGURE 9

10-20 EEG Electrode Location
SYSTEM, OPTODE AND CAP FOR NEAR-INFRARED DIFFUSE-OPTICAL FUNCTION NEUROIMAGING

RELATED APPLICATIONS

The present application claims priority to U.S. Provisional Patent application No. 61/048,446, filed Apr. 28, 2008.

FIELD

The present application relates to an optode and an electrode-optode cap having multiple optodes for use in diffuse-optical imaging, and in particular in Near-Infrared Diffuse-Optical Tomographic Functional Neuroimaging (NIR-DOTFINI) and correlation of NIR-DOTFINI with electroencephalography.

BACKGROUND

Functional Magnetic Resonance Imaging (fMRI) portrays particular parts of the brain that are active during specific activities of a subject. For example, experiments have illustrated those parts of the brain that are most active while performing mental arithmetic, or while opening and clenching a hand. Some differences have been noted between activity patterns of the mentally ill and those of “normal” subjects. fMRI operates by observing a coupled set of blood flow, volume and oxygenation changes in the brain, which are collectively termed the hemodynamic response. This hemodynamic response correlates with neuronal activity in the brain.

While the spatial resolution of fMRI is good, fMRI requires the subject’s head remain stationary between poles of a magnet in a large, bulky, sometimes noisy, and usually expensive machine throughout a study. The requirement of stable head position renders fMRI impractical as a way to observe patterns of brain activity during many activities of a subject such as— for illustration and not by limitation—walking on a treadmill or, even holding an animated conversation. Further, the expense and immobility of fMRI machines precludes routine clinical use of fMRI on patients of average wealth in diagnosis and treatment monitoring of such psychiatric and neurological disorders as schizophrenia, Parkinsonism, epilepsy, multiple sclerosis, tumors, dementia, stroke rehabilitation and traumatic brain injury where it is expected that brain activity patterns may differ from the norm.

It is well known that light, including near-infrared light, penetrates to a limited extent through many human tissues, including the brain, skull and scalp; although that light is scattered by those tissues and some wavelengths are absorbed more than others. It is also well known that a pattern of absorbed wavelengths (or color) of light transmitted by tissue varies with oxygenation of blood in the tissue. Further, volume and flow of blood in the tissue is known to change scattering properties in the tissue.

Diffuse-optical functional neuroimaging is a technique of determining patterns of brain activity in mammalian or human subjects by projecting light into the subject into selected points on the subject’s head while observing patterns of intensity, phase and color of scattered light emitted from the head at selected points. This may be performed using light having wavelengths in the near-infrared, with tomographic processing to obtain some three-dimensional localization of activity regions. It is expected that brain activity patterns obtained through this near-infrared diffuse-optical functional neuroimaging (NIR-DOTFINI) can be correlated to activity patterns obtained through fMRI, and that these patterns may also correlate with brain activity patterns obtained through electroencephalography (EEG), magnetoencephalogram (MEG), transcranial Doppler sonography (TCD), positron emission tomography (PET), and single-photon emission computed tomography (SPECT).

NIR-DOTFINI is expected to provide a more portable apparatus for functional neuroimaging than possible with fMRI, thereby providing imaging useful for research, as well as diagnosing a variety of psychiatric conditions and identifying lesions including tumors in a subject. A second advantage of NIR-DOTFINI over fMRI is its superior temporal resolution over fMRI, which permits detailed quantitative analysis of the time-course of the hemodynamic response. A third advantage of NIR-DOTFINI over fMRI is its ability to determine changes in blood volume, oxygenation and flow through the use of multiple colors of laser light and spectroscopic analysis and computer modeling.

NIR-DOTFINI apparatus may also prove useful for determining truth and falsity of statements made by a suspect, although this is still a subject of research. Other potential applications of NIR-DOTFINI include, but are not limited to, brain computer interface (BCI), real-time neurofeedback for academic learning, real-time neurofeedback for rehabilitation training, acute patient monitoring in the neuro-intensive care unit, and attention monitoring for pilots and drivers.

An optode is a device for coupling light between optical or optoelectronic components (such as fiber bundles and/or lasers and/or photo-detectors) and a surface (such as skin or mucus membranes) of a subject. An optode may be used to couple light into the surface of the subject, out of the surface, or both. Optodes are typically connected to an end of a flexible light guide such as fiber-optic fibers or bundles or liquid light guides that in turn connect to light emitting devices such as lasers and/or light measuring and detecting apparatus such as photodiodes; some optodes may be used for light emission into the subject, some for light detection, and some for both. The optic fibers or bundles or liquid light guides may be bifurcated, quadfurcated, or further divided to or from the optode for the purpose of, for example, coupling multiple colors of laser light into the surface or coupling the light collected from multiple optodes into a single detector. The light emitting apparatus such as lasers and/or light measuring and detecting apparatus may be worn by the subject, or may be located several meters away.

Some optode designs have proved to be bulky, others have proved to be incompatible with EEG, MEG or fMRI, others have proved time consuming to attach to a subject, overly difficult to attach, or to be too painful to the subject for practical use.

U.S. Pat. No. 5,361,316 to Tanaka, et al., describes a probe for coupling light from an optical fiber into a body cavity for phototherapy, the probe having a ball lens. This device has no prism and does not lend itself to use as a compact optode for functional neuroimaging. A ball lens is also used in the coupling device of Schwarz, et al., Bull Lens Coupled Fiber-Optic Probe For Depth Resolved Spectroscopy Of Epithelial Tissue, Optics Letters, 15 May 2005 1159-1161. The coupling device of Schwarz lacks a prism, and has several stimulus fibers surrounding a central receive fiber in the same coupling device, and is intended to provide for infrared measurements of skin rather than deeper structures.

Hamamatsu’s NIRO-200 system provides a multiple-channel device for generating infrared light, for coupling this light to an optode having a prism at the end of an optical fiber for coupling light into scalp regions without hair, and for receiving and measuring transmitted light. The prism optode
design of Hamamatsu lacks any small optical component to
displace the hair and make direct contact with scalp skin. This
device also does not support simultaneous electroencepha-
lography. The optodes of this device have no lens and is bulky
eough to preclude the high-density optode arrays required
for good resolution and accurate neuroimaging, and is bulky
eough to preclude high density optode arrays combined with
electroencephalographic arrays.

SUMMARY

An optode for a functional near-infrared diffuse optical
neuroimaging system uses a GRIN lens and prism for cou-
pling to an optical fiber at a right-angle to the lens. The lens is
inserted into a grommet for attachment to an elastomeric cap
worn by a subject. In an embodiment, the cap also has an array
of electroencephalographic electrodes, with optodes arranged
such that an optical path exists between a transmitter
and a receiver optode beneath each electroencephalographic
electrode.

BRIEF DESCRIPTION OF THE FIGURES

FIG. 1 is an exploded cross-sectional diagram illustrating a
prior art optode.

FIG. 2 is a cross-sectional diagram illustrating the prior art
optode of FIG. 1 assembled.

FIG. 3 is a cross-sectional diagram illustrating a prior art
optode with a flange replacing the upper washer.

FIG. 4 is an exploded cross-sectional diagram illustrating a
new optode design.

FIG. 5 is a cross-sectional diagram illustrating the optode
of FIG. 3 partially assembled.

FIG. 6 is a cross-sectional diagram illustrating the optode
of FIG. 3 fully assembled into an elastomeric cap.

FIG. 7 is a cross-sectional diagram illustrating an alterna-
tive embodiment of the optode.

FIG. 8 is an illustration of quadrifurcated fibers permitting
illumination of the optodes with multiple wavelengths using
one laser per optode.

FIG. 9 is an illustration of a 21-electrode prior-art 10-20
electroencephalographic pattern.

FIG. 10 is an illustration of a 21-electrode, 24-optode,
optical neuroimaging and electroencephalographic pattern.

FIG. 11 is an illustration of a 75-electrode, 64-optode,
optical neuroimaging and electroencephalographic combina-
tion cap pattern.

FIG. 12 is an illustration of optode placement in the cap for
use with quadrifurcated fiber allowing individual lasers to illu-
minate multiple widely-separated optodes.

FIG. 13 is an illustration of a system for functional neu-
roimaging using the cap of FIG. 11 and the optodes of FIG. 4.

DETAILED DESCRIPTION OF THE EMBODIMENTS

In the prior art optode 100 embodiment of FIGS. 1 and 2, an
optical fiber 102 or fiber bundle as known in the art of fiber
optics and having a jacket 104 are secured within a 90-degree
fitting 106 to form a cable termination assembly 108. Cable
termination assembly 108 is slideably engaged within an
optode body 110. An elastomeric spring 112 secured within
the optode body 110 by an optode cap 114 tends to hold the
cable termination assembly 108 in an extended position
where it may contact a scalp of a mammal (not shown).
Optode body 110 is secured into a polyethylene cap 116 by an
upper washer 118 and a lower washer 120 or snap, which may
be attached to the optode body 110 by glue, a friction fit, or by
a shoulder (not shown) on the optode body 110.

In an alternative prior embodiment, as illustrated in FIG. 3,
upper washer 118 is replaced by a flange of optode body 130.

The prior art optode 100 has been found to be uncom-
fortable for subjects because of the sharp square bottom edge
of the cable termination assembly 108 and because the elastom-
eric spring 112 does not always provide for adequate move-
mement of cable termination assembly 108 in body 110. Further,
the optode has an overall height 122 that is sufficient to
interfere with performance of functional magnetic resonance
neuroimaging while cap 116 is being worn by a subject.

The improved optode 200 of FIGS. 4, 5, and 6 has a fiber
optic fiber or fiber bundle 202 as known in the art of fiber
optics and as suitable for transmitting light of a desired wave-
length. The optic fiber may have a jacket 204 and is inserted
into and attached by glue or a collet into an optode body 206. Optical
fiber 202 is optically coupled to a prism 208 also
mounted within the optode body 206, and prism 208 is
coupled to the high-density central portion of a flat end of a
graded-index (GRIN) lens 210. GRIN lens has a convex
curved end 211 for optical coupling to a scalp of a subject.

The GRIN lens 210 is also attached to the optode body 206,
and has an outer circumferential surface 214.

An elastomeric cap 218, such as a cap formed from a sheet
of neoprene rubber or woven elastic has holes 220. Elasto-
meric grommets 222, such as may be made from neoprene
rubber, are inserted into cap 218. The elastomeric grommet
222 has a central hole 224 having diameter small enough to
grip the circumferential surface 214 of the GRIN lens 210.

In use, GRIN lens 210 of each optode is inserted into hole
224 of the associated elastomeric grommet 222, as shown in FIG.
6; the cap is then secured to the head of a subject such that the
convex surface or curved end 211 of each optode is held
adjacent to the scalp of a subject (not shown), and the optical
fiber 202 is coupled to an appropriate light source or light
measurement device.

An alternative embodiment of the optode is illustrated in
FIG. 7. In this embodiment, the optode body 206 has a tubular
extension 230 adjacent to a circumferential surface of GRIN
lens 210 such that mechanical stresses are passed directly
from body 206 extension 230 into grommet 222.

To perform functional neuroimaging, many optodes, typi-
cally in an array of from 24 to 64 optodes, are provided in a
cap. Each optode is optically coupled to a suitable light
source, such as a pulsed, continuous wave or frequency
modulated laser, and/or a suitable light detection and meas-
urement device such as an avalanche photodiode or a pho-
tomultiplier tube, the light sources periodically transmit light
into the subject’s head and the light detection and measure-
ment devices periodically make measurements of transmitted
light that are then processed by a computer to determine a
pattern of blood oxygenation and volume in the brain of the
subject.

Optodes typically operate in transmitter/receiver optode
pairs. Transmitter optodes pass light from a light source
through a scalp surface through tissue including bone into a
subject’s brain, some of this light is absorbed and some is
scattered back out through bone and scalp back through a
receiver optode to a light detection and measurement appa-
ratus. Each transmitter optode, and each receiver optode,
participates in multiple transmitter/receiver pairs. The detec-
tion and measurement apparatus detects variations in light
absorption by tissue beneath and between the optodes of a
pair. A computer processes these variations and measure-
ments from multiple transmitter/receiver pairs into an image and may also perform spectroscopic analysis if multiple wavelengths of light are used.

It is known that heme changes color with oxygenation. Oxygenated arterial blood (having heme in hemoglobin) tends to be a brighter red while deoxygenated venous blood tends to have a bluer color. This is a consequence of changes in optical absorption of heme with oxygenation that occurs in the near-infrared as well as visible wavelengths.

It is therefore possible to determine both heme concentration and approximate oxygenation along the scattering path between transmitter and receiver optode pairs by measuring differences in scattering and absorption at several wavelengths of red and near-infrared light.

Each time a neuron fires, a small amount of that neuron’s reserves of Adenosine TriPhosphate (ATP) is consumed in repolarizing the neuron; mitochondria of the neuron then recharge the ATP supply through a process that consumes oxygen. Areas of the brain having high activity therefore have higher oxygen consumption than those areas having low activity. This increased oxygen consumption in the active areas is associated with an increase in blood flow in those portions of the brain and results in localized increases in blood volume and oxygen levels—the so-called hemodynamic response to brain activation that is monitored by fMRI. The increased blood flow may also lead to an increase in heme concentration along with changes in oxygen tension in active areas of the brain.

It is therefore desirable to measure differences in optical absorption along the paths between optodes at several wavelengths of light, as well as measuring overall absorption and phase changes in the emitted light; these measurements can give indications of heme concentration and oxygenation and flow and can thereby monitor changes in activity of areas of the brain along optical paths between optodes.

Each individual optode may, and usually does, participate in more than one transmitter/receiver optode pair, with each pair having an optical path through the head between the optodes of the pair. Pairs may be distinguished by time multiplexing or by frequency encoding modulation on the lasers. Pairs and pathways may also be distinguished by operating transmitter optodes at different wavelengths while using frequency-selective light detection and measurement devices. Pairs, including transmit optodes of the same groups, operating at the same time and frequency but positioned sufficiently far apart on the head that the light transmission along paths between them is effectively zero can also be distinguished from each other and attenuation of pathways between such pairs may be measured simultaneously.

In an embodiment, transmit optodes are grouped into transmit optode groups, as illustrated in FIG. 8. In a transmit optode group each transmit optode, such as optode 402 is fed by an optical guide that is an optical fiber bundle that is quadfurcated—or divided into four separate subguides or strands 404, 406, 408, 410, where each strand 404, 406, 408, 410 has at least one optical fiber, the optical fiber of each strand extending into the optode body 206 and coupled to prism 208.

In this embodiment, each strand 404, 406, 408, 410 is coupled to be fed by a separate laser of a laser group, such as laser 412 feeding into strand 404, laser 414 feeding into strand 406, laser 416 feeding into strand 408, and laser 418 feeding into strand 410. Each of the four lasers 412, 414, 416, 418 of a laser group operates at a separate wavelength, in an embodiment laser 412 is at six hundred ninety nanometers, laser 414 operates at seven hundred eighty five nanometers, laser 416 operates at eight hundred eight nanometers, and laser 418 operates at eight hundred thirty nanometers.

Since it would be expensive, albeit functional, to have a four laser laser-group, including lasers 412, 414, 416, 418, for each transmit optode; in an embodiment each laser, such as laser 412, feeds strands, such as strands 404, 420, 422, 424 coupled to multiple optodes, such as optodes 402, 426, 428, 430. This architecture permits construction of the system for significantly lower cost than fMRI machines. Similarly, the remaining lasers 414, 416, and 418 of the laser group are coupled through optical fiber strands of quadfurcated cables to all four optodes 402, 426, 428, 430, of the optode group.

The four optodes of each transmit optode group are placed in the cap at locations such that pathways through the head to each associated receive optode are short enough to produce strong signals from only one transmit optode of each group, while pathways from other optodes of the group are significantly longer. For example, as illustrated in FIG. 12, if optode 610 is in a first group, nearby transmit optodes 602, 612, 630 belong to different groups that have transmit lasers either illuminated at different times, or carrying different frequency modulations. The remaining optodes in the first group are well separated on the head and in the cap, for example and not limitation the optodes of the first group may include optodes 620, 652, and 658 in addition to optode 610. This permits disambiguation of the optode-optode paths.

In a time-division embodiment, one laser selected from lasers 412, 414, 416, 418 of the transmit optode group is illuminated at a time, and that illumination time is distinguished by being illuminated at a time separated from times of illumination of similar wavelengths of other transmit optode groups.

In an alternative embodiment, each laser of lasers 412, 414, 416, 418 is amplitude-modulated at a frequency that is unique within the group, but all four lasers 412, 414, 416, 418 generate light simultaneously. With this embodiment, electronics associated with each receive optode determines at least an intensity of modulation at each laser modulation frequency to disambiguate the lasers and to determine spectral characteristics of absorption along paths from transmit optodes to the receive optode.

Since all four optodes 402, 426, 428, and 430, of the optode group are illuminated simultaneously, at the same wavelength, and with the same modulation, these optodes are preferably placed well apart on the head such that light emitted from each optode, such as optode 402, does not reach receive optodes near, and having significant optical paths to, another optode, such as optode 426 of the group.

In an alternative embodiment, the fibers of each transmit optode are bifurcated instead of quadfurcated, and each is coupled to only two lasers operating on different wavelengths.

A standard arrangement of electroencephalographic electrodes, known as the 10-20 electrode pattern, is illustrated in FIG. 7. The 10-20 international system includes 19 scalp electrodes and 2 reference electrodes on the ears.

In an embodiment where it is desired to correlate electroencephalographic measurements with NIRS-DOTfNIR, the cap has an array of low-contact-area electroencephalographic electrodes interspersed with a pattern of transmitter and receiver optodes for NIRS-DOTfNIR. For example, in the embodiment of FIG. 11, the electroencephalographic electrodes are laid out in a cap in a standard 10-10 electrode pattern, which includes the 10-20 electrodes. In the embodiment of FIG. 11, most of the electroencephalographic electrodes (see FIG. 11 for key), with exception of reference electrodes and some electrodes near periphery of the pattern,
are arranged such that a transmitter and a receiver optode of a sixty-four optode array are on approximately opposite sides of each of the electroencephalographic electrodes; such that the optode array provides for measurement of neurovascular activity beneath at least three fourths of the electroencephalographic electrodes.

The cap of FIG. 10 using the optodes of FIG. 4 may be used without shaving a subject's hair because the GRIN lenses tend to displace hair and to make contact primarily with skin of the scalp.

Similarly, the embodiment of FIG. 10 provides an arrangement for the cap where each electroencephalographic electrode in the standard "10-20" pattern of FIG. 9, with exception of reference electrodes, has a transmitter and a receiver optode of an optode array are on approximately opposite sides of each of the electroencephalographic electrodes; such that the optode array provides for measurement of neurovascular activity beneath at least three fourths of the electroencephalographic electrodes.

In the embodiments of FIGS. 10 and 11, the electroencephalographic electrodes are sensitive to neural activity primarily beneath each electrode, while the optodes are primarily sensitive to activity along an optical path describing an arc between the optodes, the arrangements of FIGS. 10 and 11 therefore provide coincident electroencephalographic and NIR-DOT/NI imaging.

An embodiment according to FIG. 11 has quadruflracted transmit optode groups as illustrated in FIG. 12.

The transmit optodes of each group are coupled to transmit lasers as illustrated in the system diagram of FIG. 13. In this embodiment, each of the thirty-two receive optodes 504 is coupled by a straight-through optical fiber—a fiber that is neither bifurcated nor quadruflracted—to an avalanche photodiode 506 for detecting transmitted light from one or more transmit optodes. In a more-expensive alternative embodiment, each receive optode 504 is coupled by a bifurcated or quadruflracted optical fibers, these fibers couple through filters for selecting specific laser wavelengths to avalanche photodiodes.

The avalanche photodiode 506 is coupled to a data acquisition system that uses signals from the photodiode 506 to measure at least an attenuation at each wavelength from transmit optodes 510 of at least one transmit optode group; this attenuation information is provided to a processor 512 for image construction. The processor 512 also activates transmit lasers 514 of each transmit optode group, providing near-infrared light to the transmit optodes.

It should be noted that the matter contained in the above description or shown in the accompanying drawings should be interpreted as illustrative and not in a limiting sense. The following claims are intended to cover generic and specific features described herein, as well as all statements of the scope of the present method and system, which, as a matter of language, might be said to fall therebetween.

What is claimed is:

1. An optode for interfacing light to biological tissue comprising:
   at least one flexible light guide;
   a prism optically coupled to the flexible light guide;
   a graded incidence (GRIN) refractive lens having a circumferential surface, a convex surface and a flat surface, the flat surface optically coupled to the prism; and
   a body for holding the optical fiber, prism, and GRIN lens, the body configured to maintain the fiber, prism, and GRIN lens in fixed relationship having a right angle between an axis of the fiber and an axis of the GRIN lens; wherein the convex surface of the graded incidence refractive lens is capable of interfacing light to biological tissue.

2. The optode of claim 1 further comprising an elastomeric grommet having a hole, the circumferential surface of the GRIN lens secured in the hole of the elastomeric grommet, the grommet and body retaining the optode in a hole of an optode-positioning device.

3. The optode of claim 1 wherein the body has a tubular extension, the extension having an inner surface in contact with the circumferential surface of the GRIN lens and an outer surface in contact with an elastomeric grommet, the grommet and body retaining the optode in a hole of an optode-positioning device.

4. The optode of claim 1 wherein the GRIN lens and flexible light guide are optimized for transparency in the infrared.

5. The optode of claim 1 wherein the flexible light guide comprises at least one optical fiber.

6. The optode of claim 1 wherein the flexible light guide comprises at least two optical fibers, and wherein the flexible light guide is at least bifurcated into a first and a second subguide.

7. An optode for interfacing light to biological tissue comprising:
   at least one flexible light guide;
   a prism optically coupled to the flexible light guide;
   a GRIN lens having at least a convex surface and a second surface, the second surface optically coupled to the prism;
   and
   a body for holding the flexible light guide, prism, and GRIN lens, the body configured to maintain the guide, prism, and GRIN lens in fixed alignment having right angle between an axis of the fiber and an axis of the GRIN lens;

   wherein the convex surface of the lens is capable of interfacing light to biological tissue; and
   wherein the flexible light guide comprises a plurality of optical fibers and divides into a plurality of subguides, and wherein at least a first optical fiber of the plurality of fibers extends from the prism to an end of a first subguide of the subguides, and wherein at least a second optical fiber of the plurality of fibers extends from the prism to an end of a second subguide of the subguides.

8. The optode of claim 7 wherein the body has a tubular extension, the extension having an inner surface in contact with a circumferential surface of the lens and an outer surface for contacting an elastomeric grommet.

9. The optode of claim 7 wherein the GRIN lens and flexible light guide are optimized for transparency in the infrared.

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