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**Johansen et al.**

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(54) **SURFACE PLASMON APPARATUS**

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(73) Assignee: **Knut Johansen**, Linköping (SE)

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**G01J 4/00** (2006.01)

(52) **U.S. Cl.** ..... **356/445**; 356/244; 356/246; 356/367

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See application file for complete search history.

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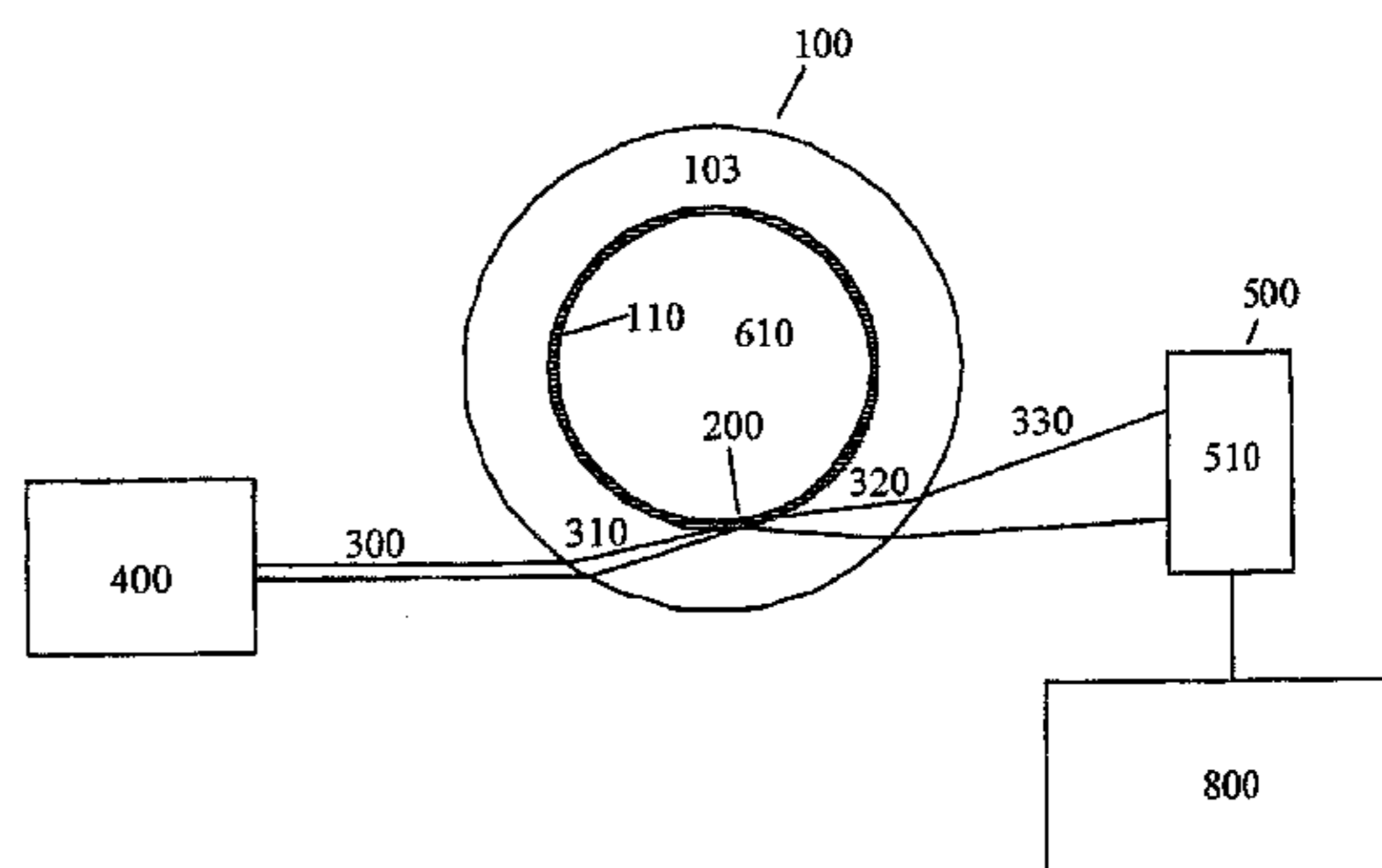
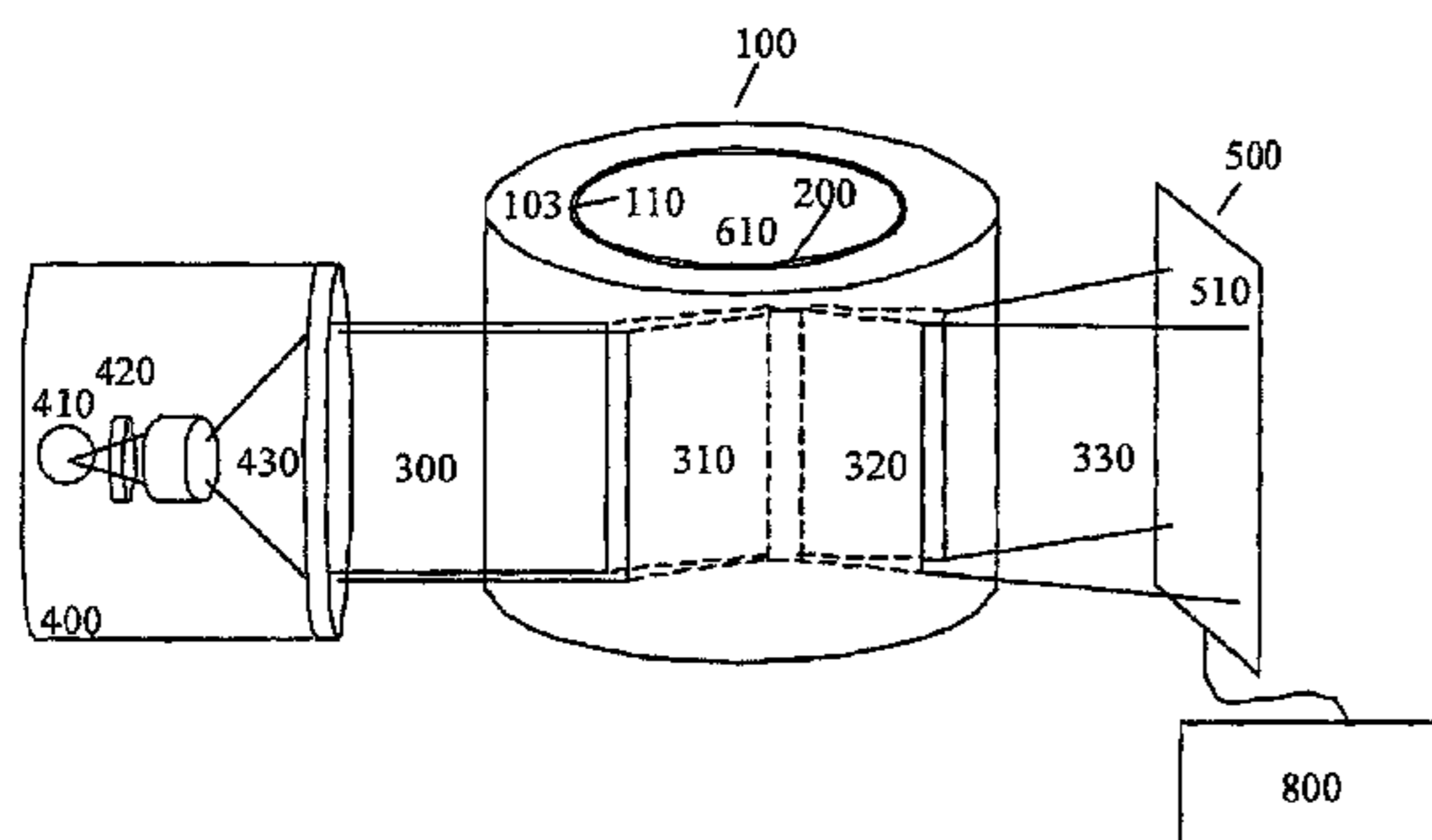
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(57) **ABSTRACT**

A surface plasmon apparatus includes a light source, a sensor unit for Surface Plasmon Resonance (SPR) which includes a transparent sensor structure forming at least one wall of a cavity, the wall being defined by a concave inner surface and a convex outer surface, wherein the inner surface is provided with a layer of a conductive material capable of supporting a surface plasmon, a flow structure in the cavity so as to form at least one compartment for sample between the flow structure and the inner wall of the cavity, a detector for detecting reflected light from the sensor unit, and a processing unit.

**11 Claims, 39 Drawing Sheets**



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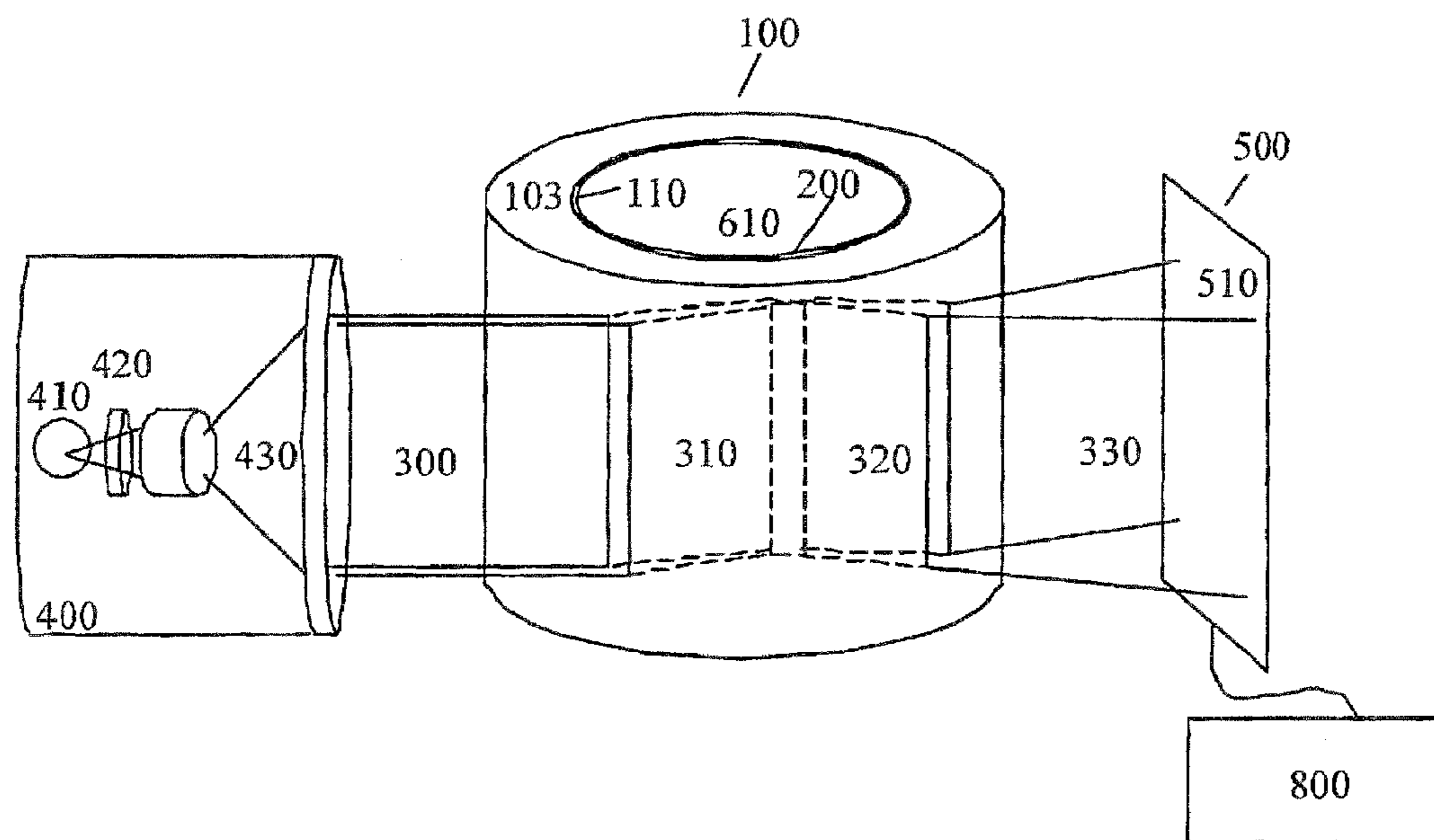


Fig. 1a

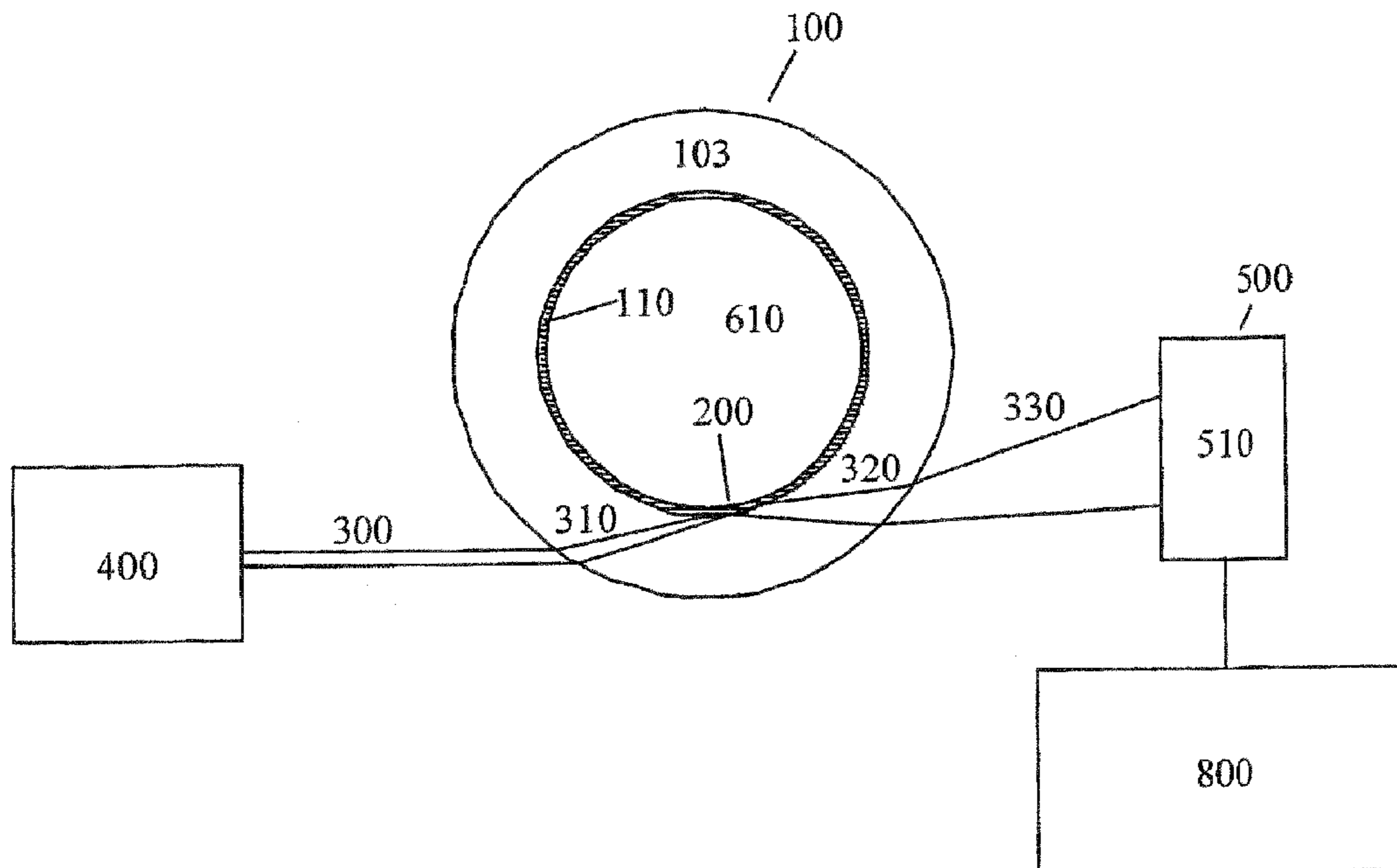


Fig. 1b

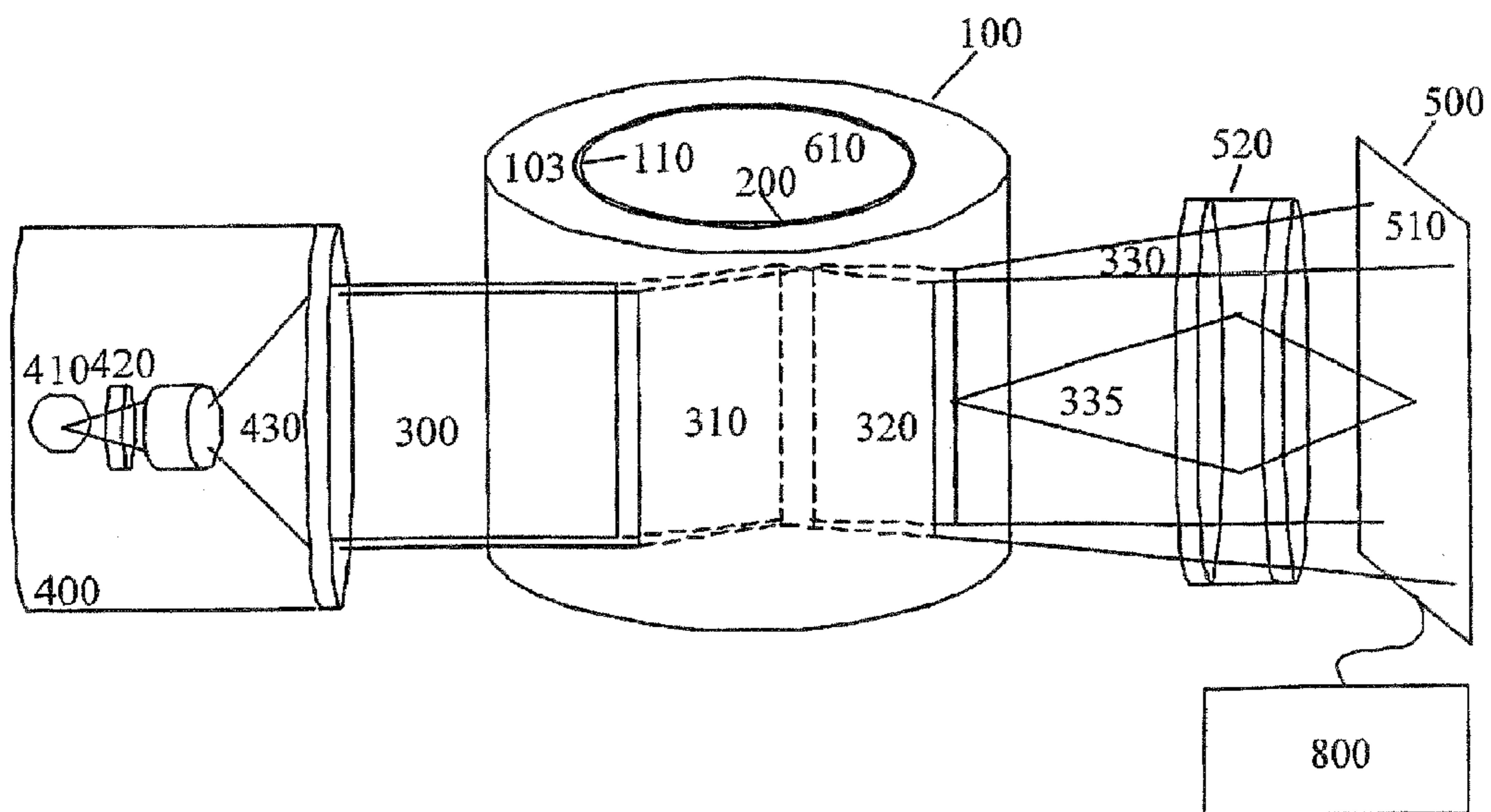


Fig. 1c

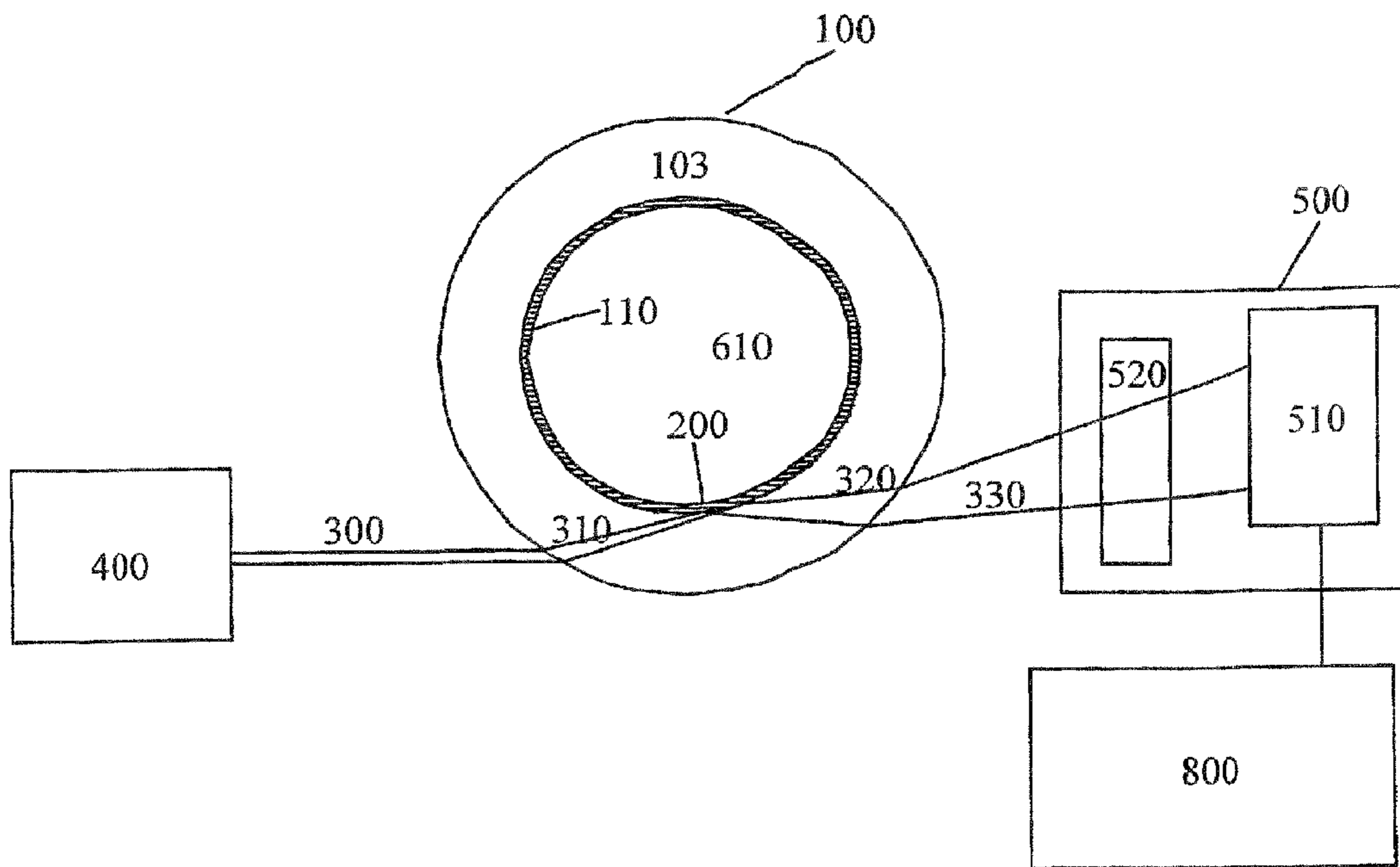


Fig. 1d

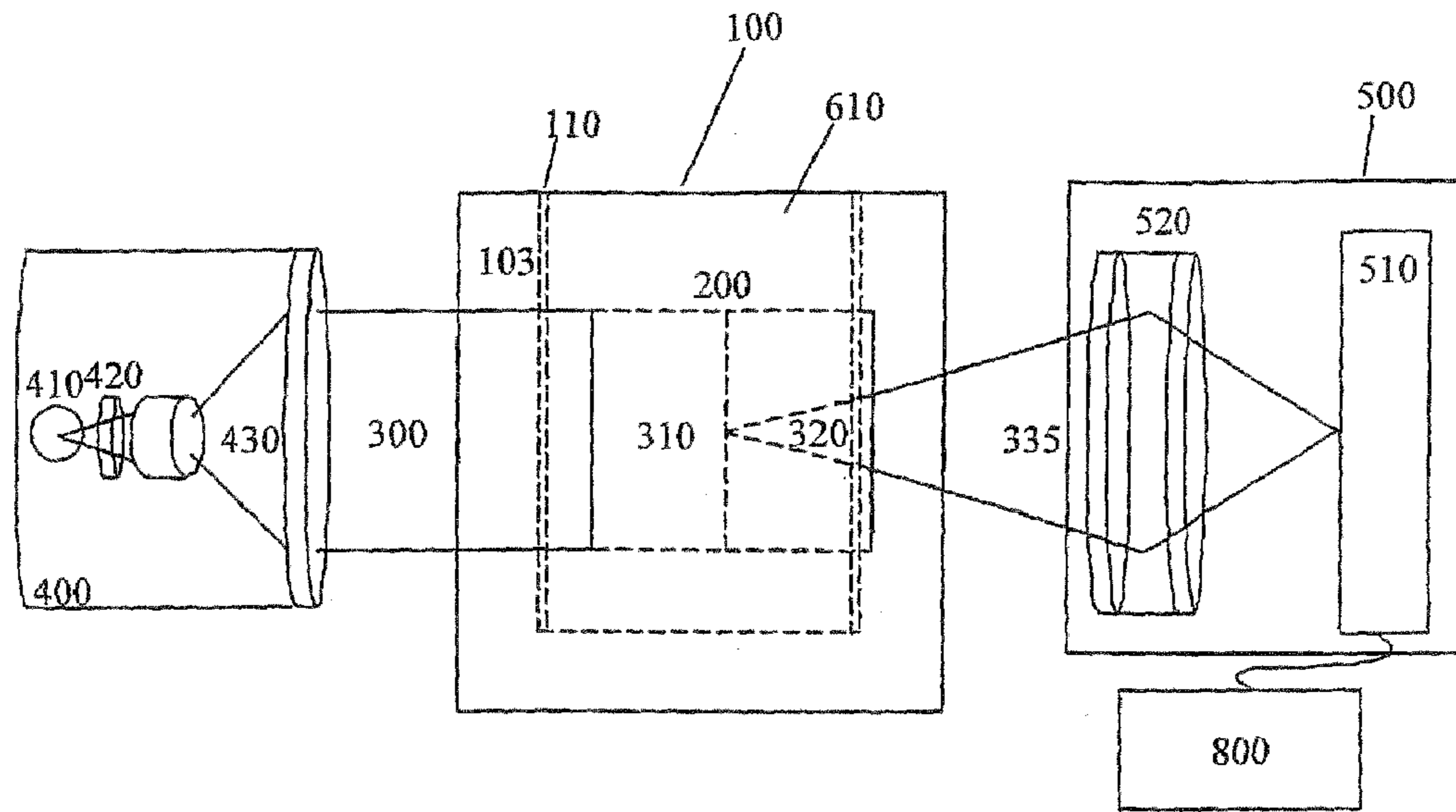


Fig. 1e

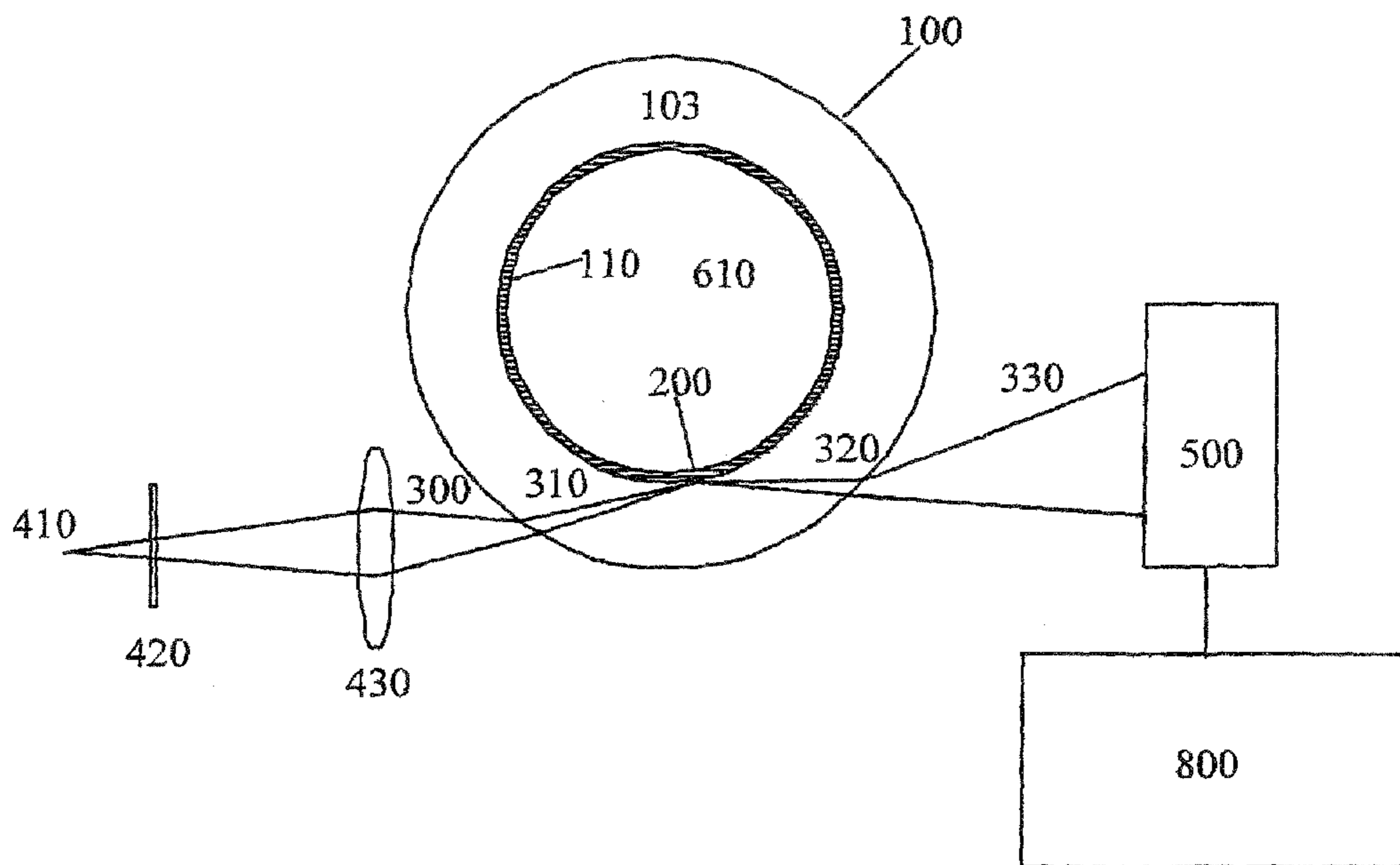


Fig. 2

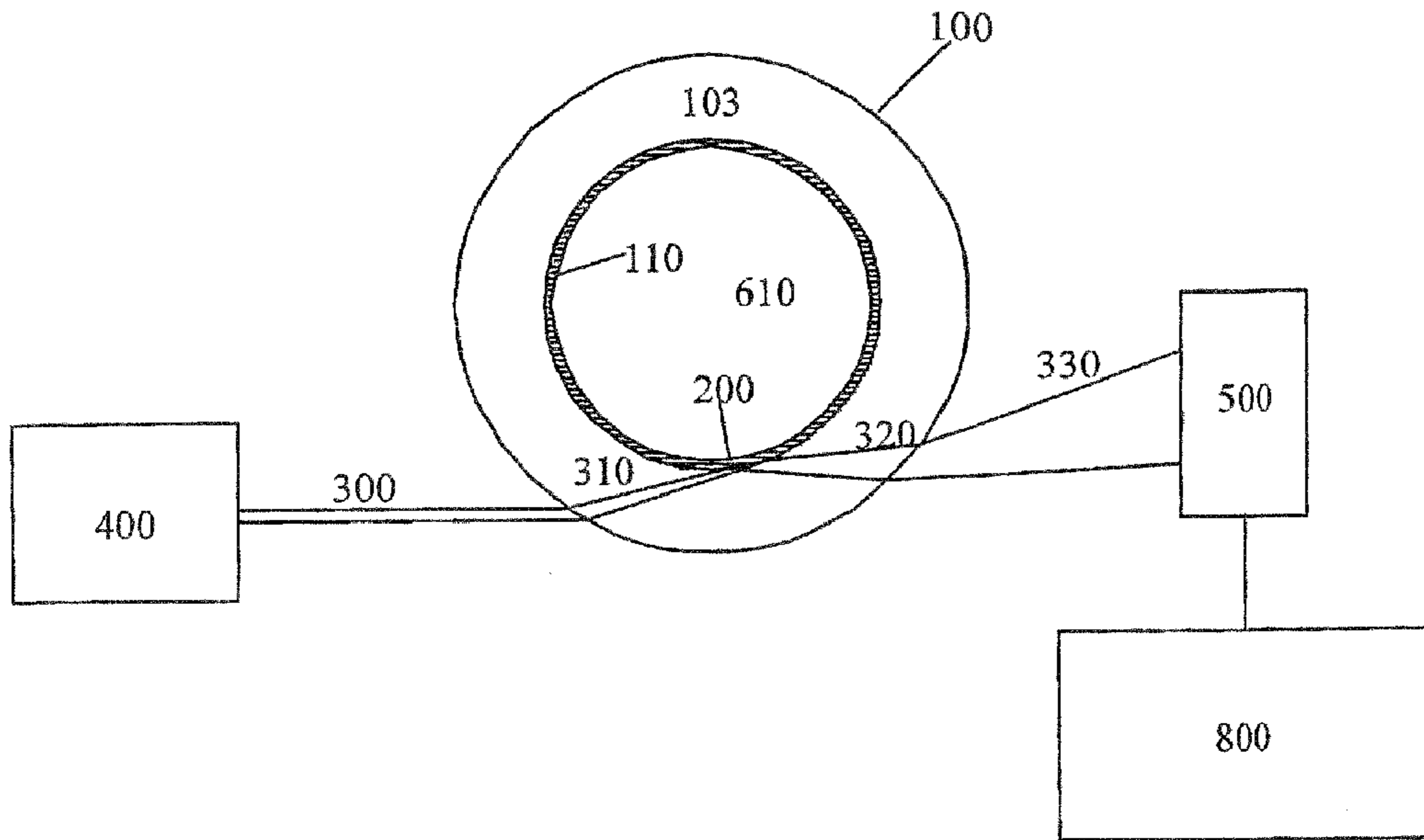


Fig. 3a

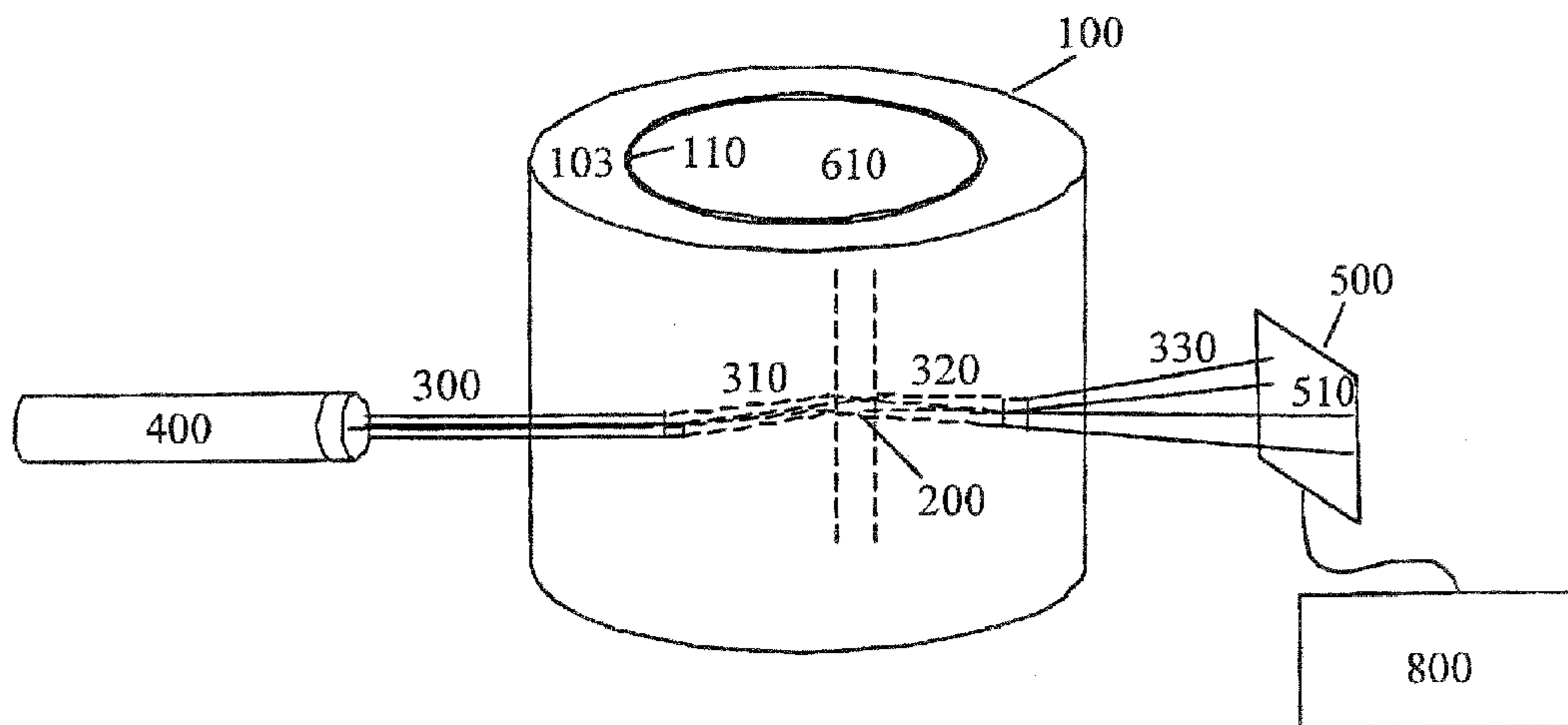


Fig. 3b





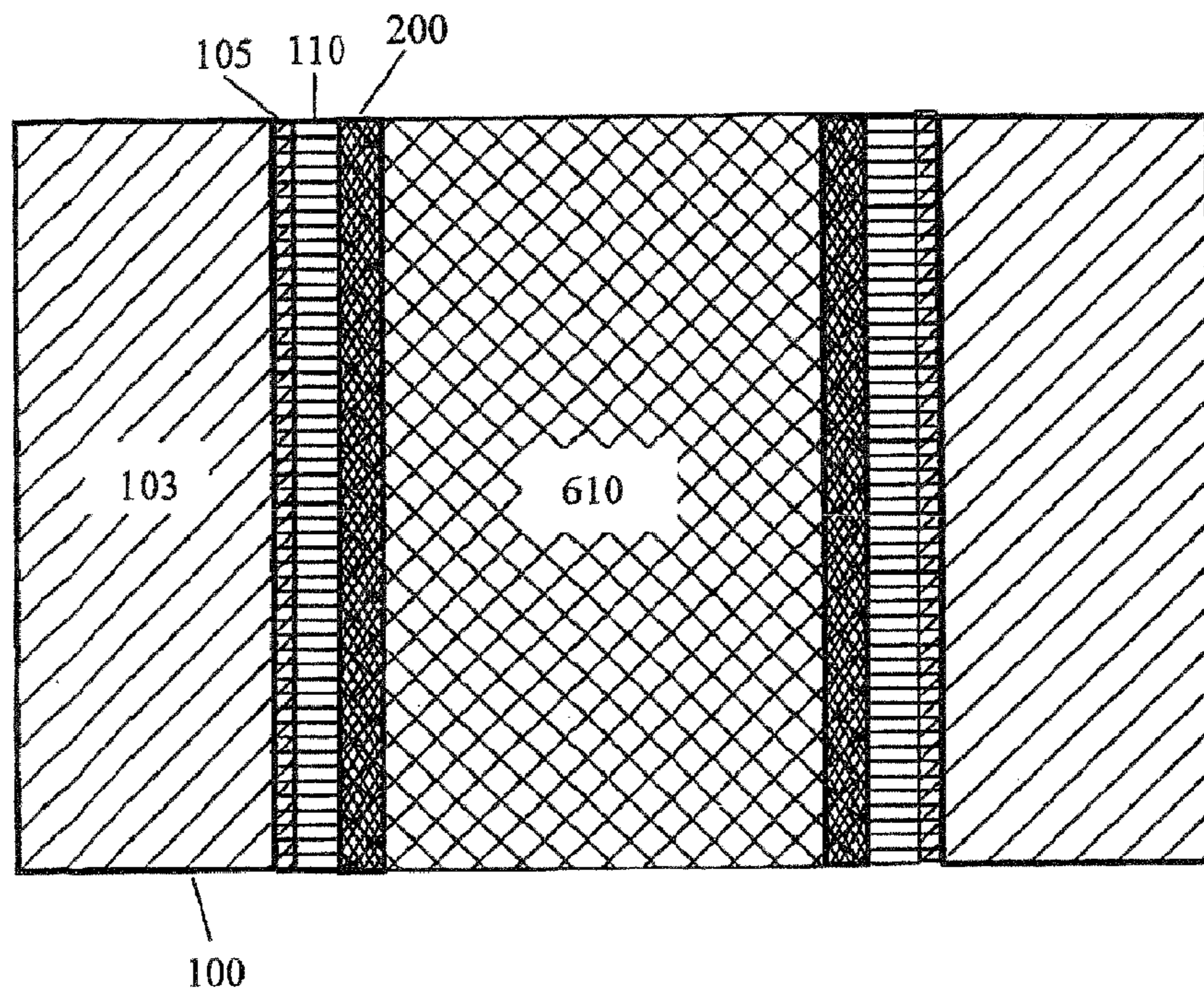


Fig 5

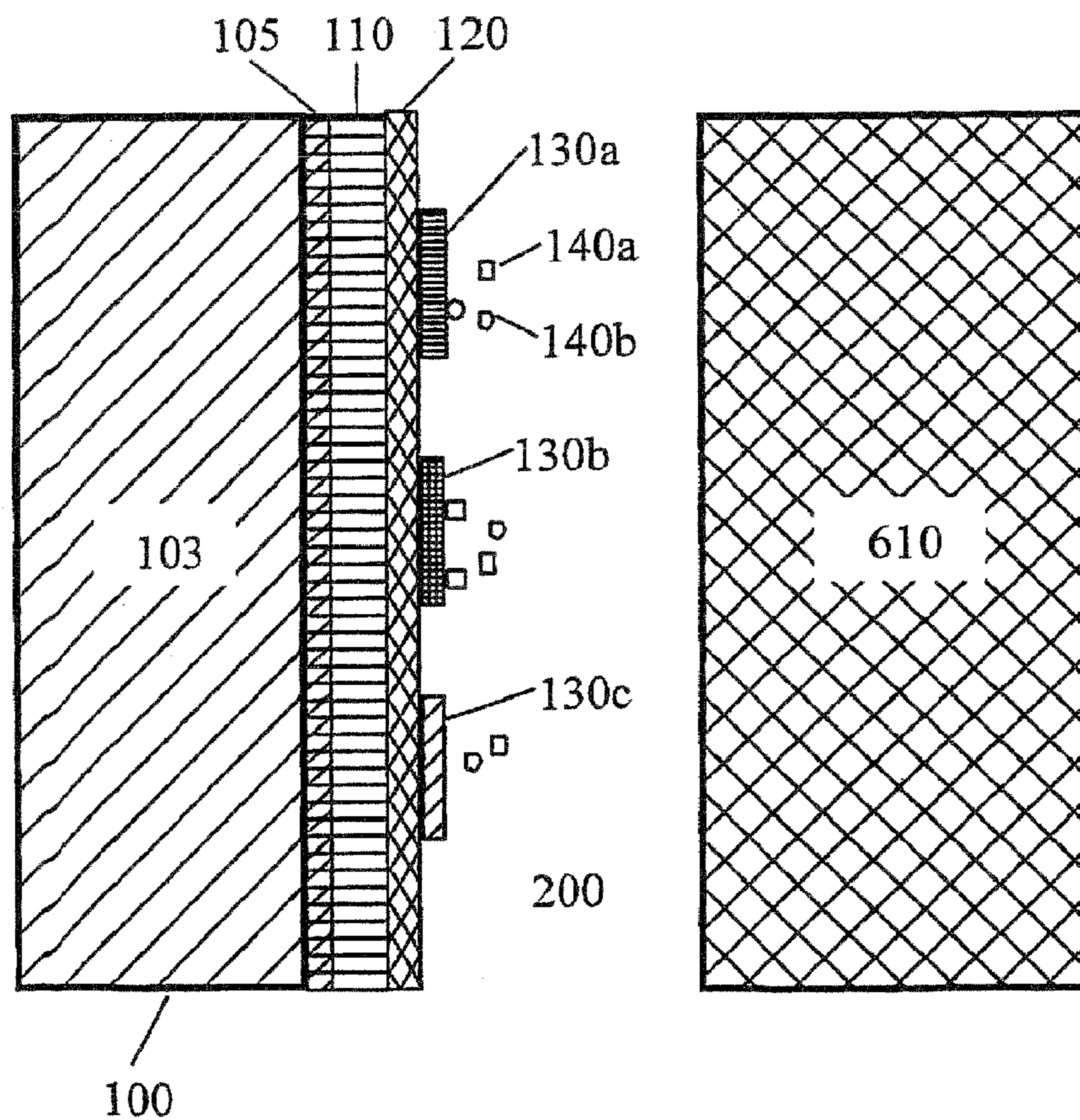


Fig 6

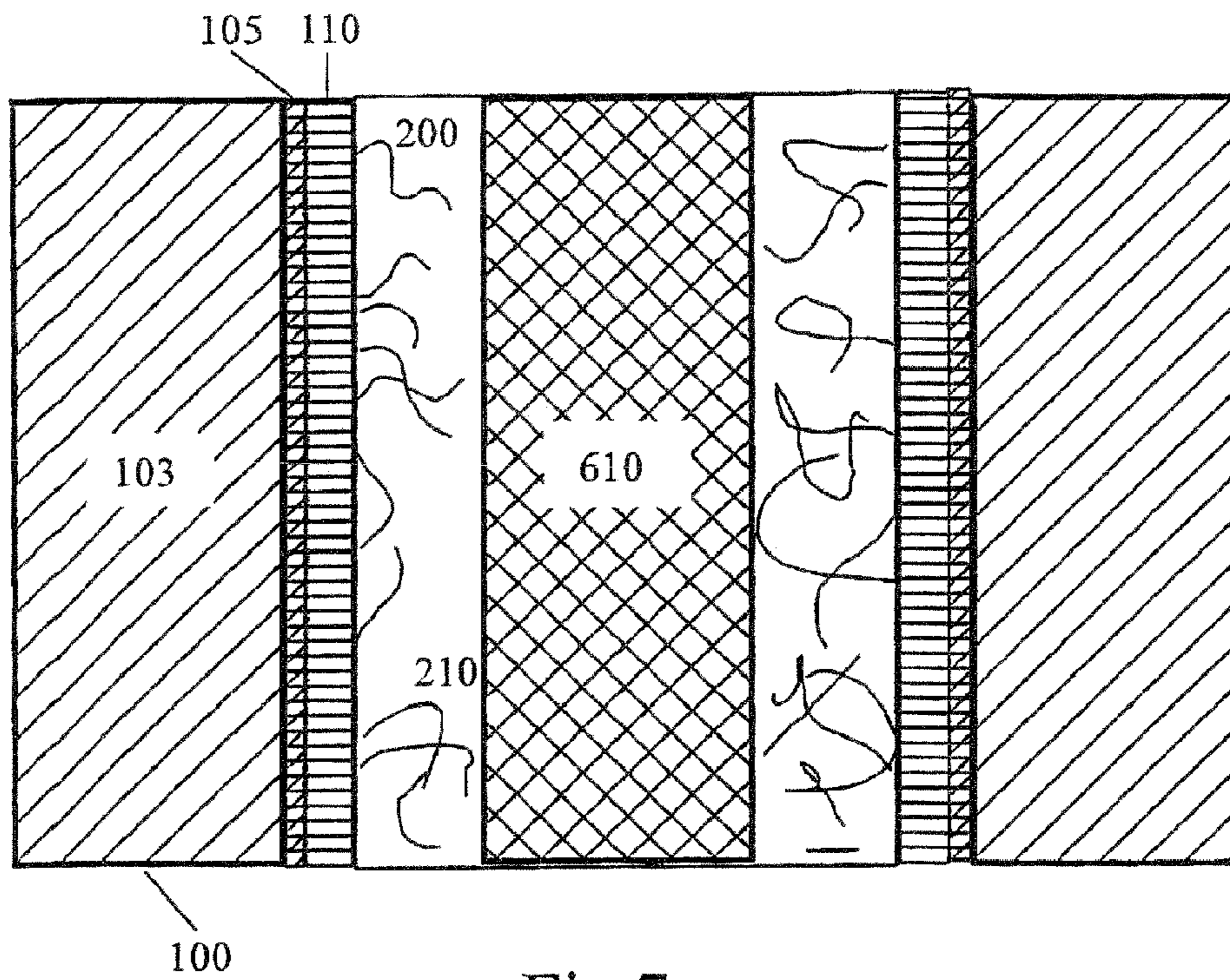


Fig 7

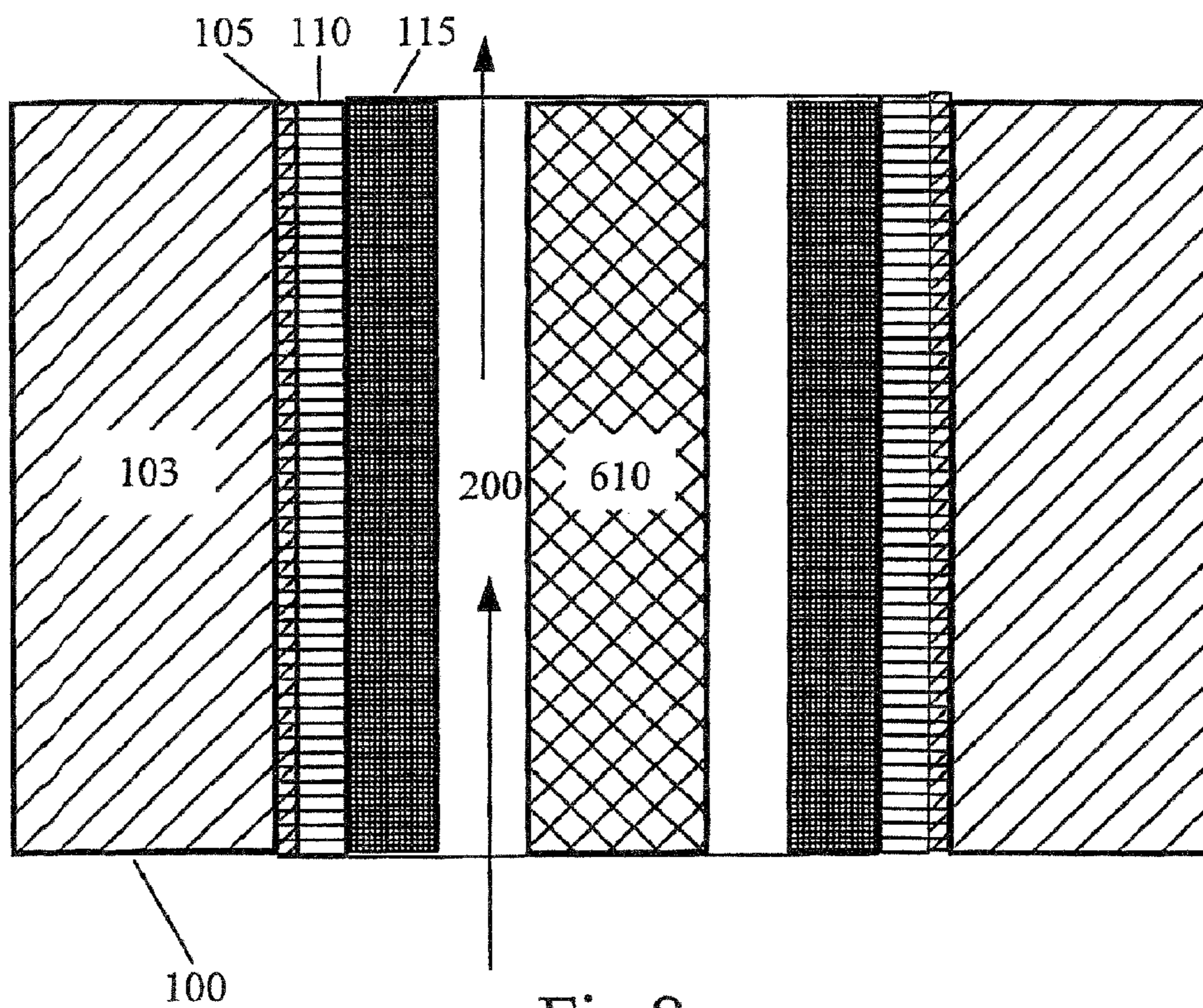


Fig 8

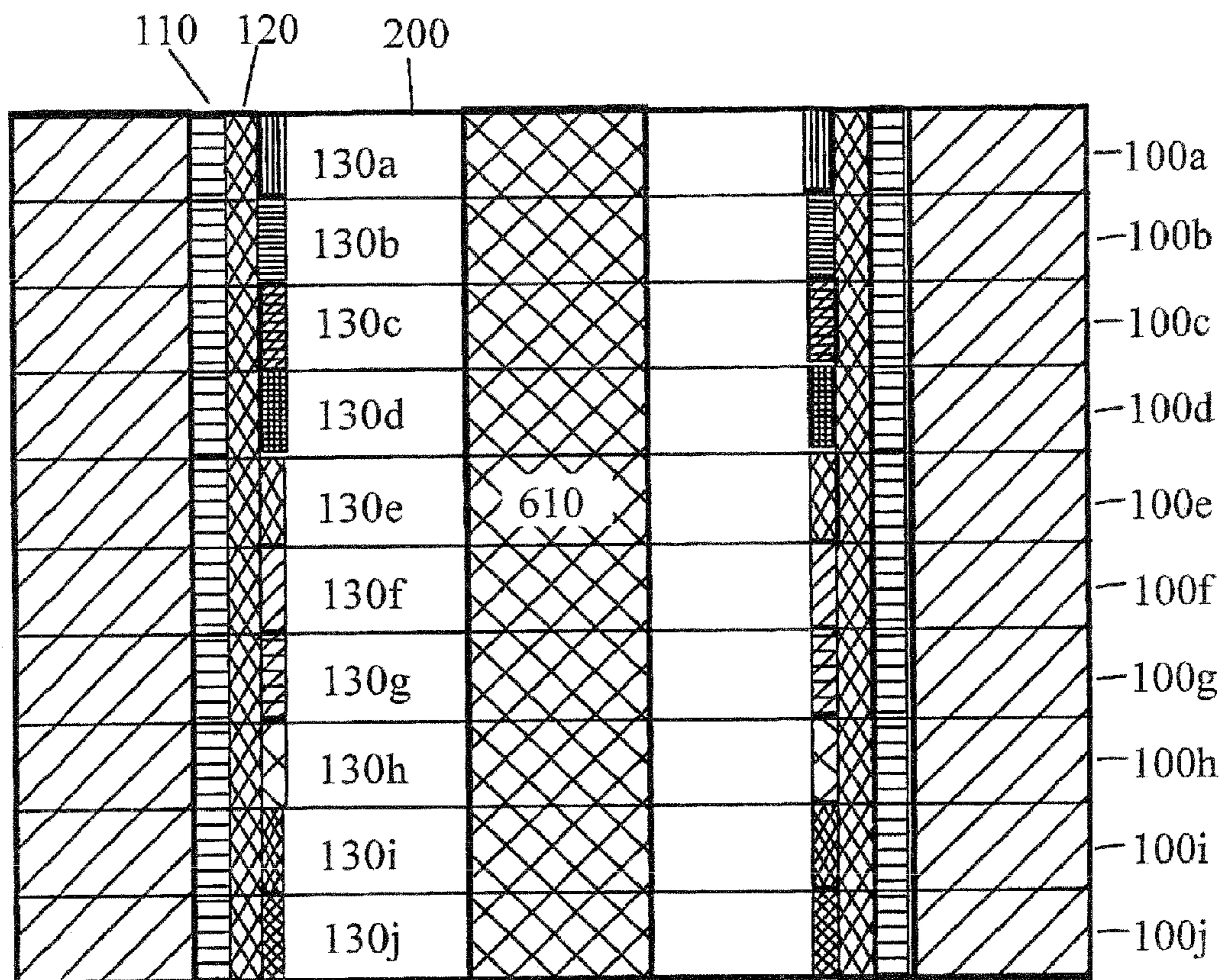


Fig 9a

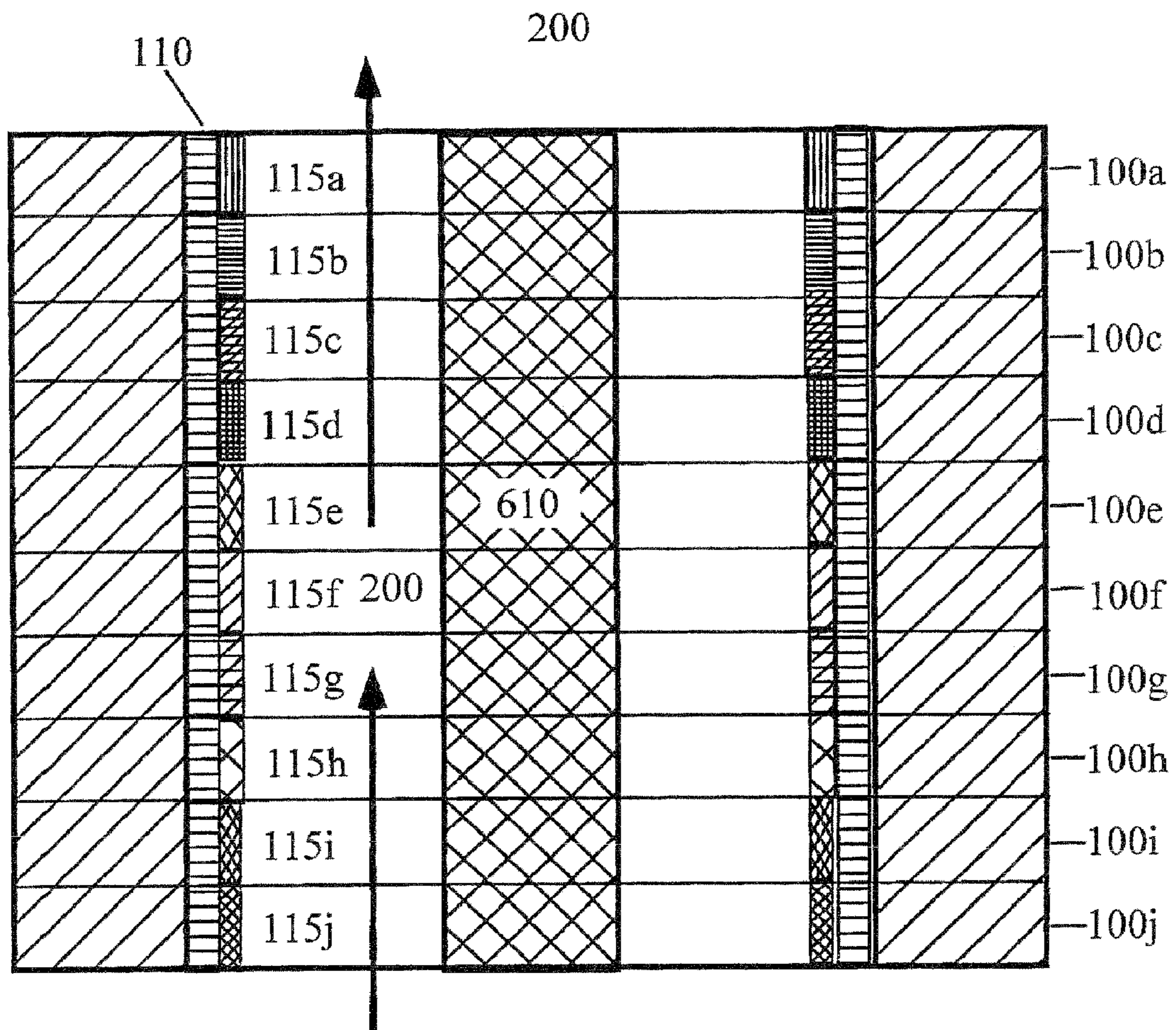


Fig 9b

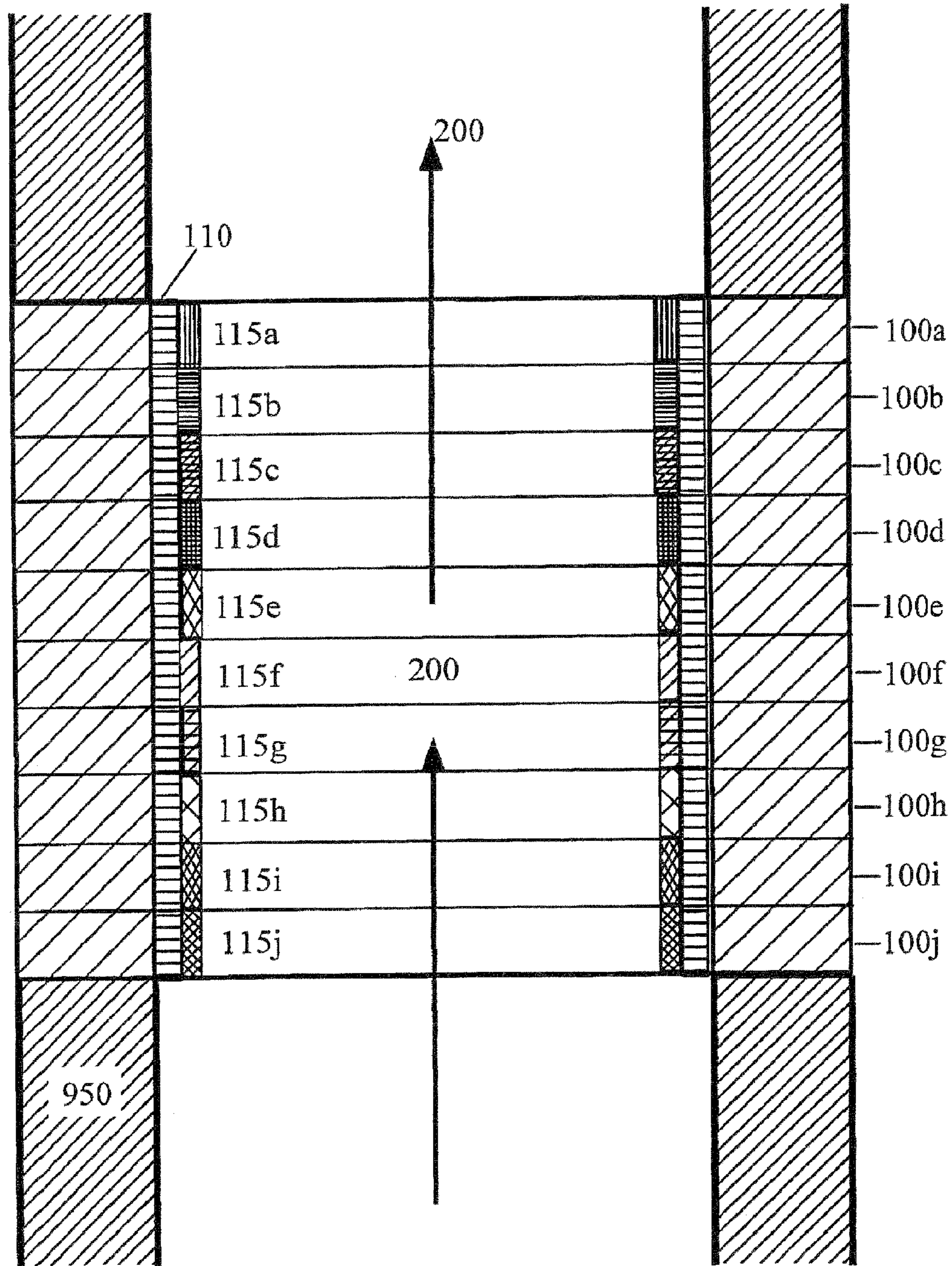


Fig 9c

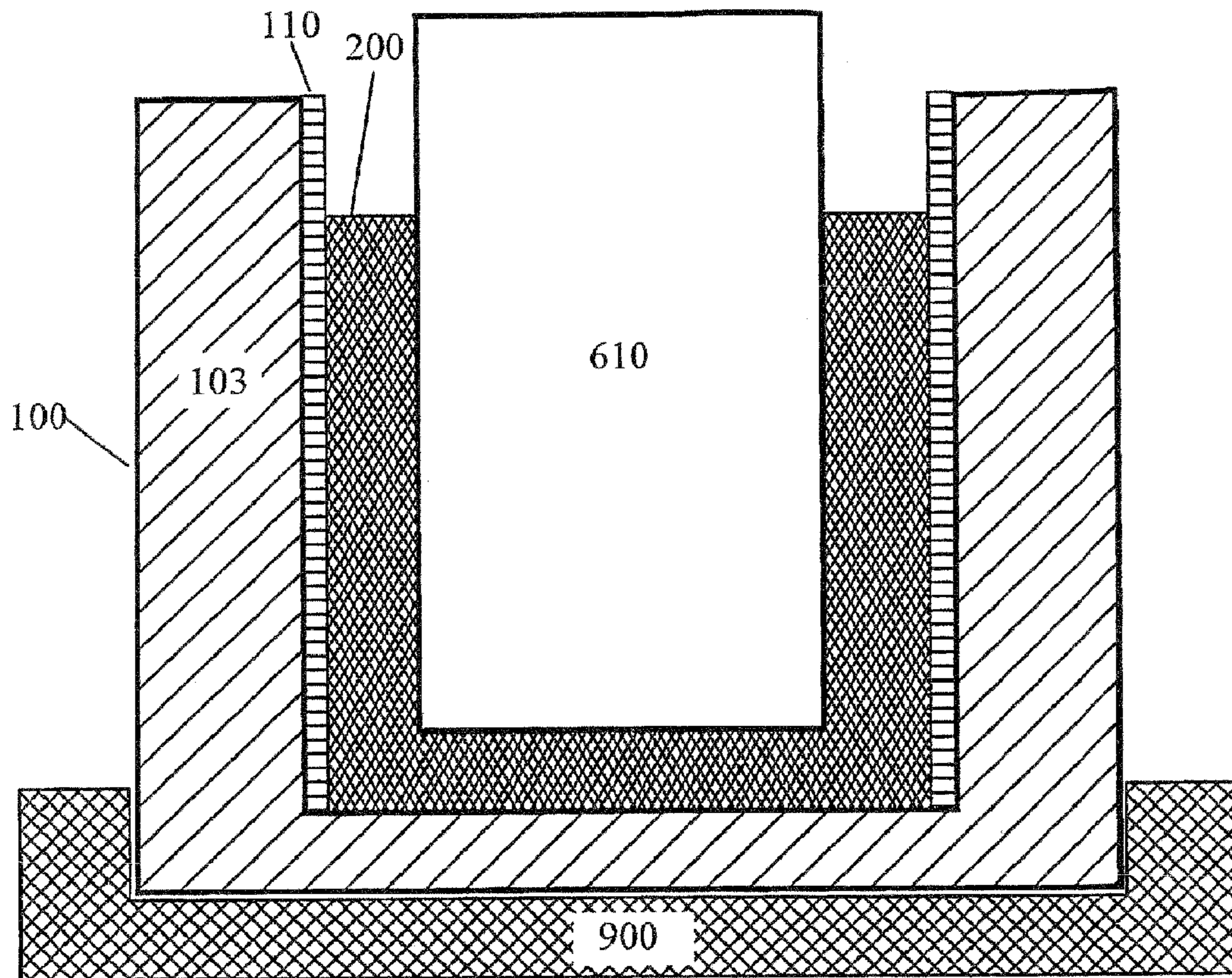


Fig. 10a

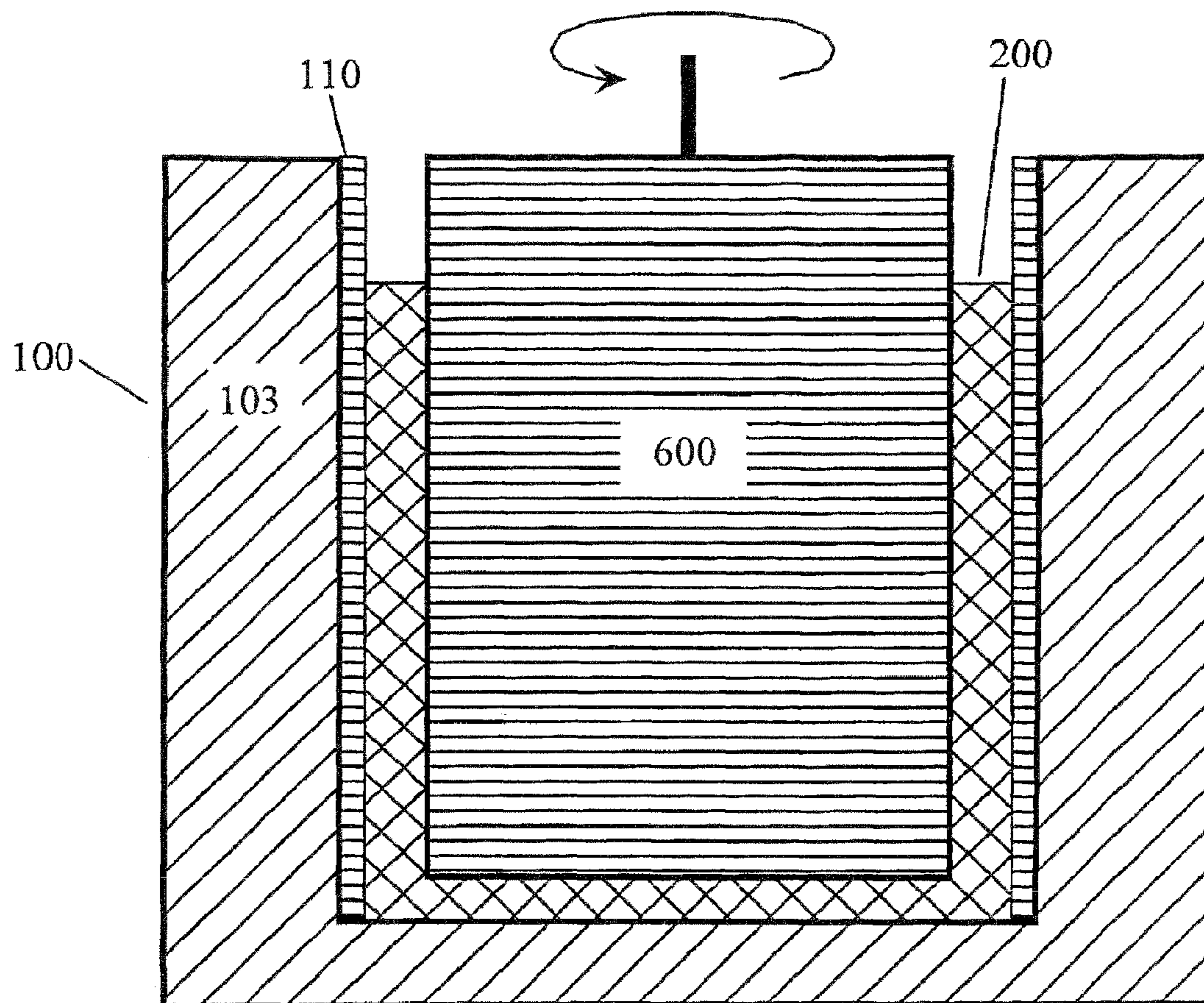


Fig. 10b

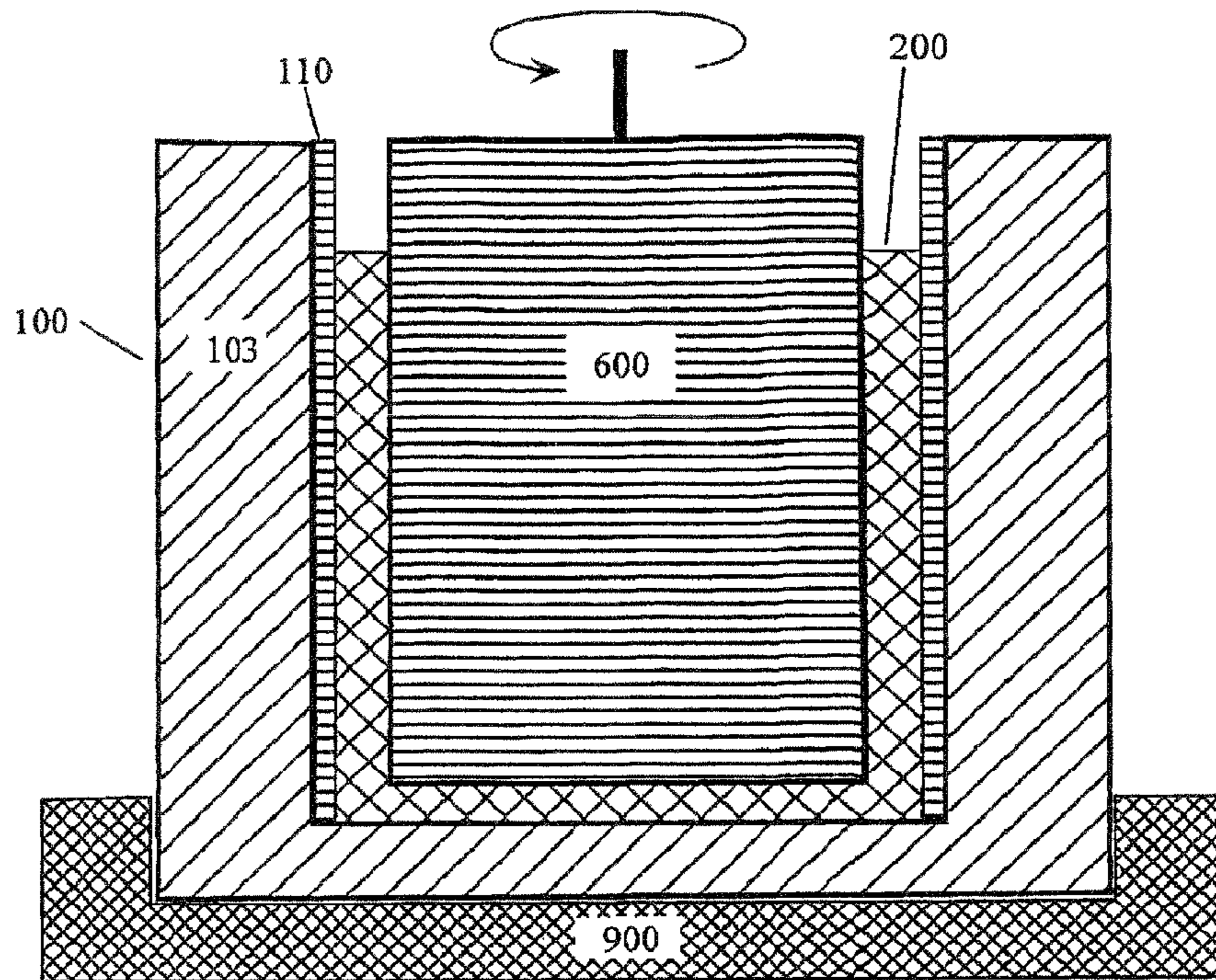


Fig. 10c

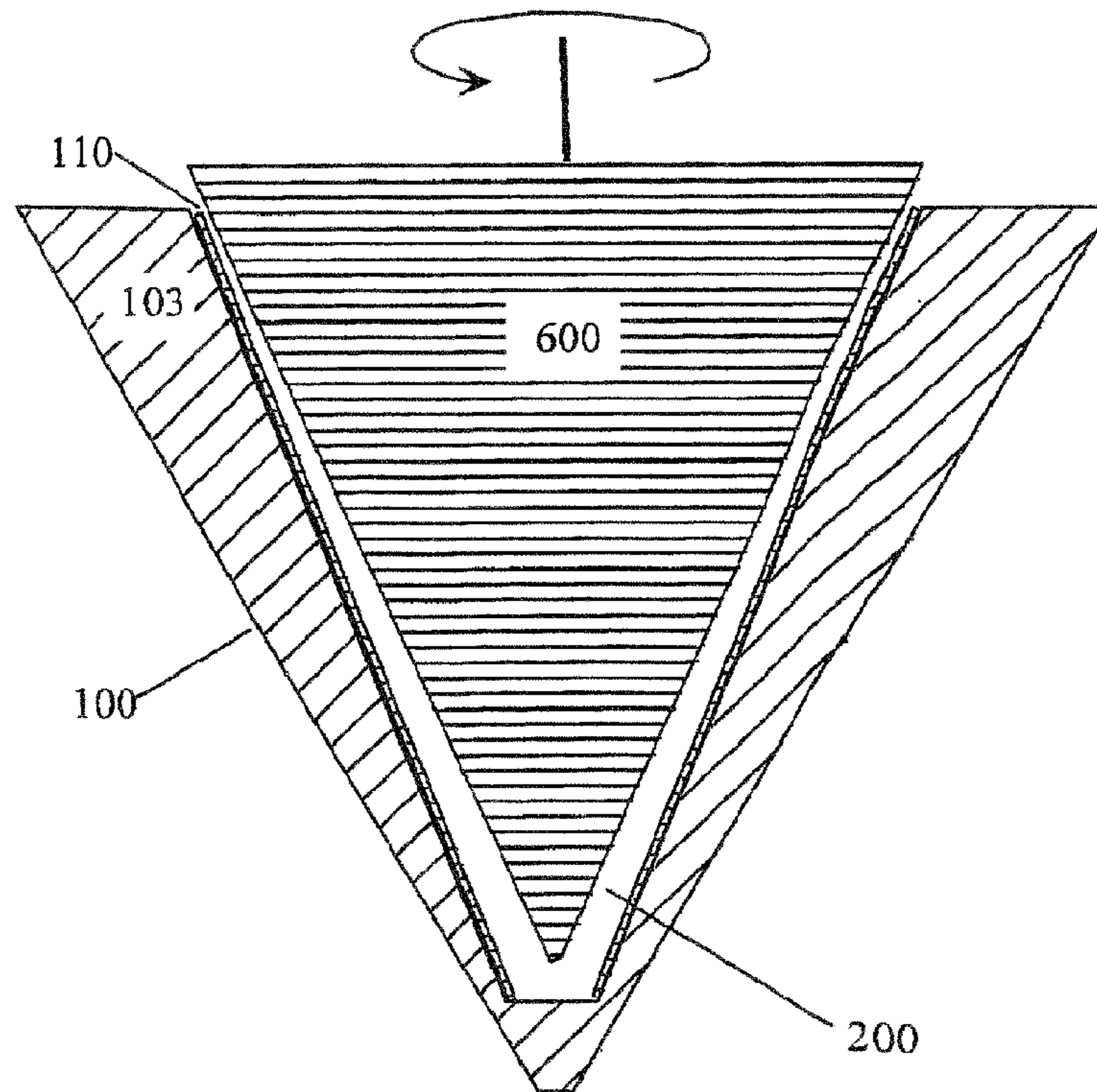
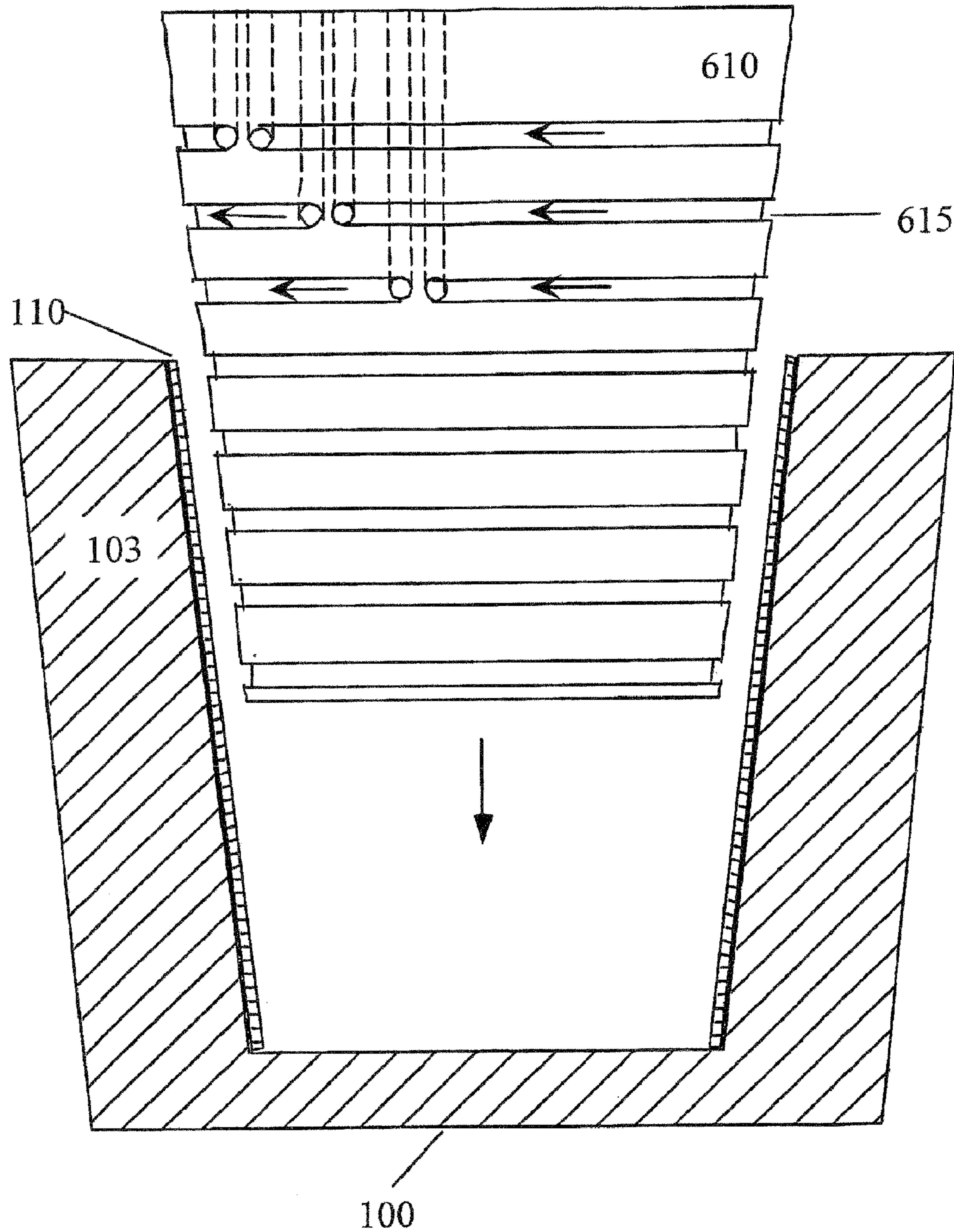


Fig. 11





100  
Fig. 12a

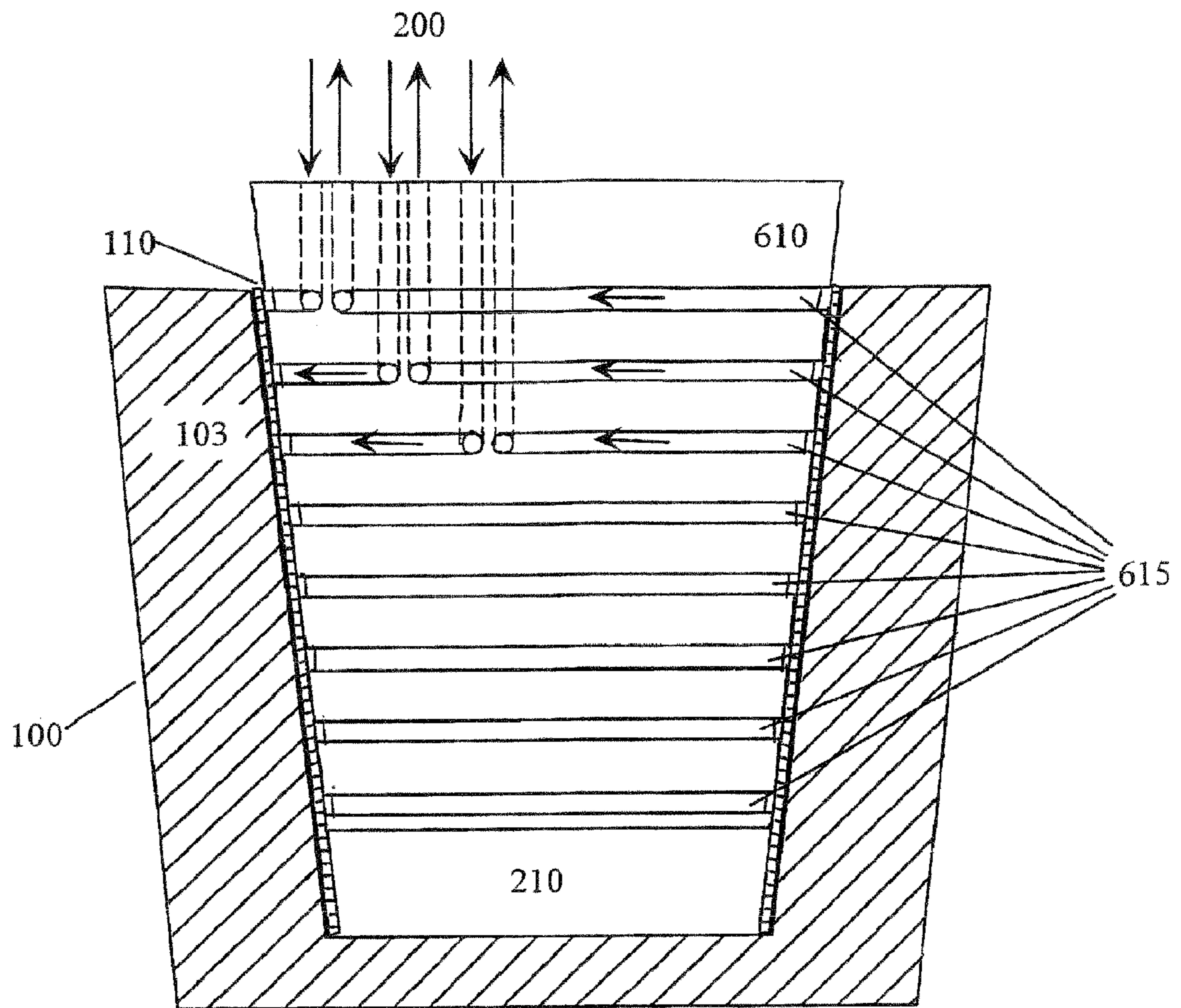


Fig. 12b

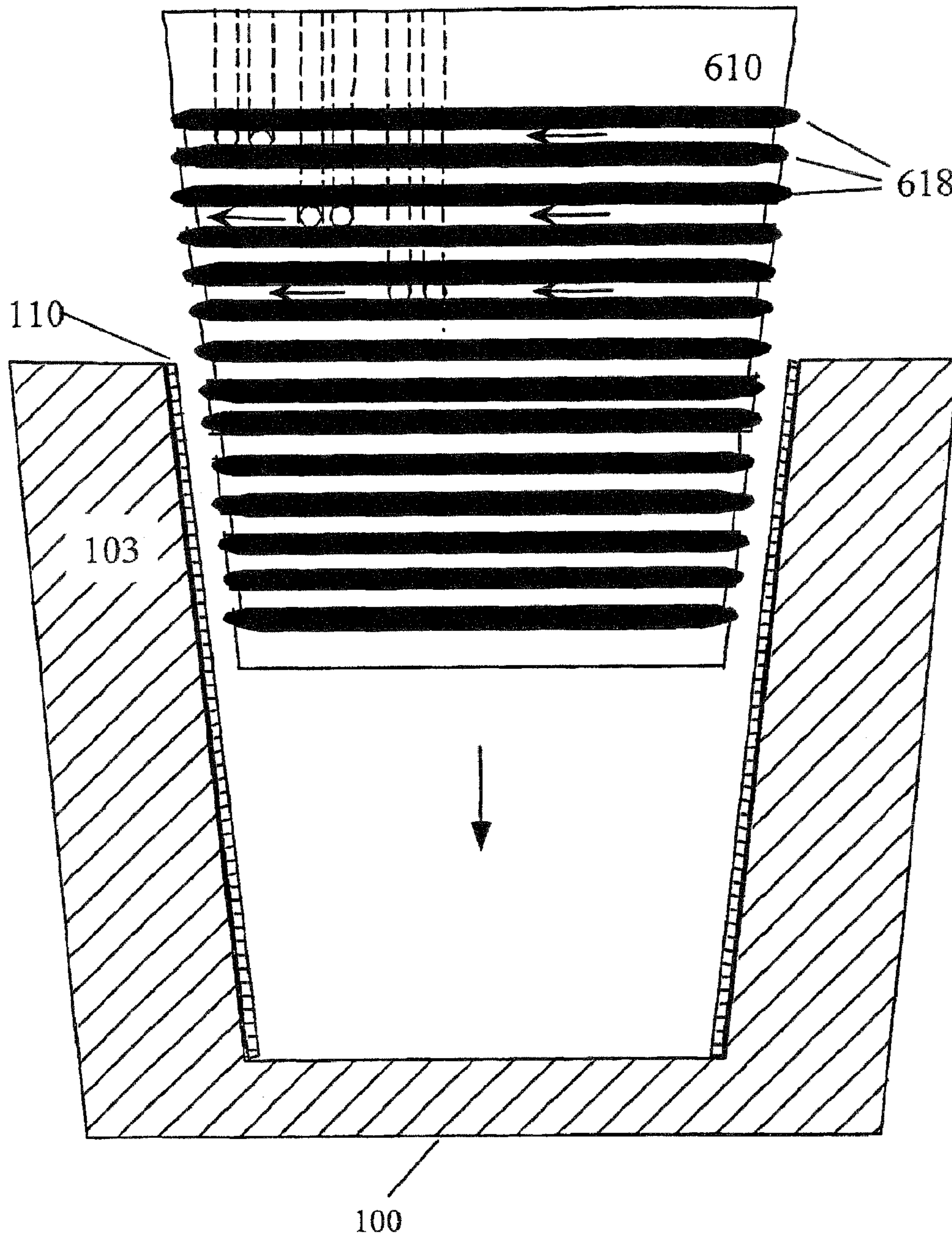


Fig. 12c

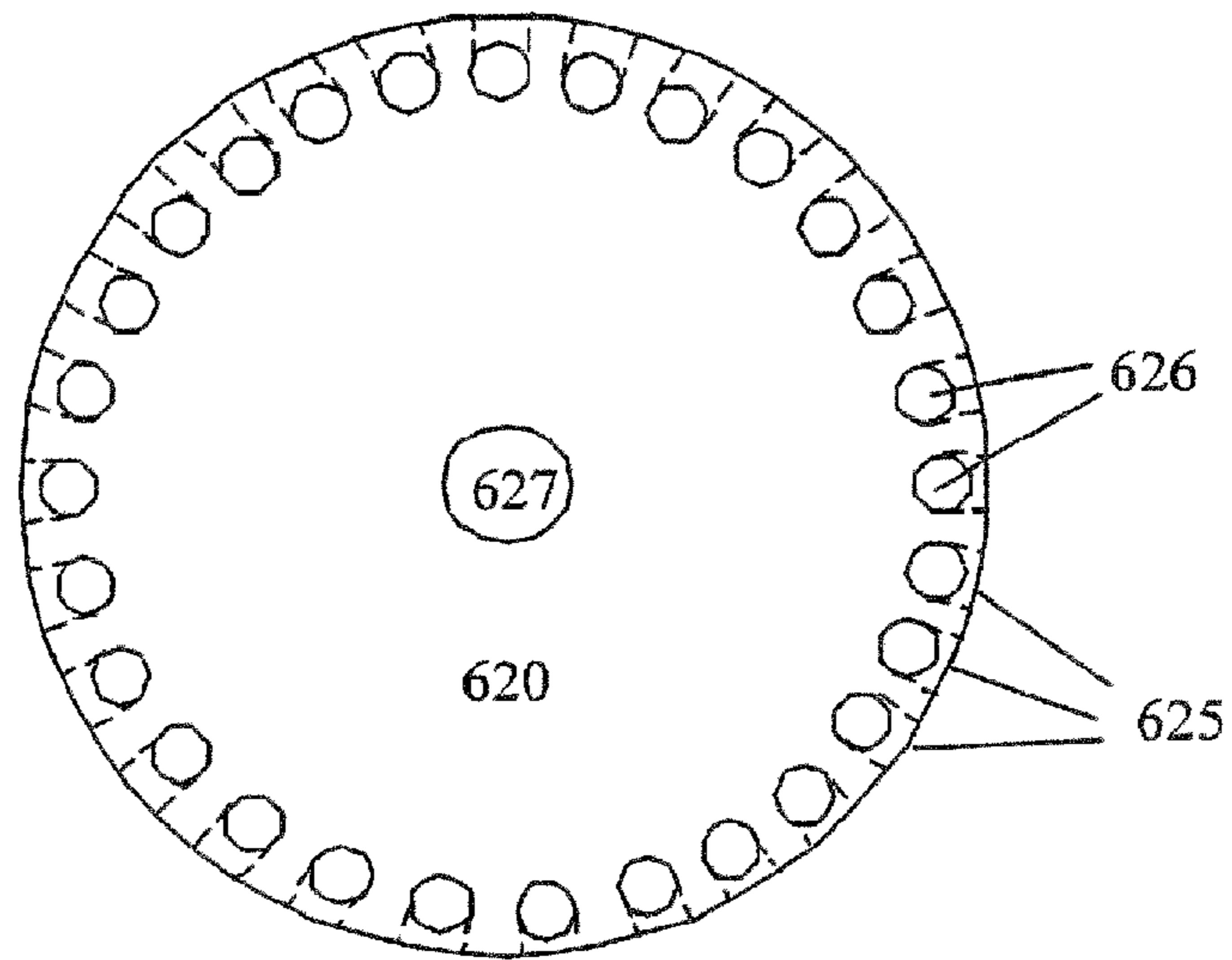


Fig. 13a

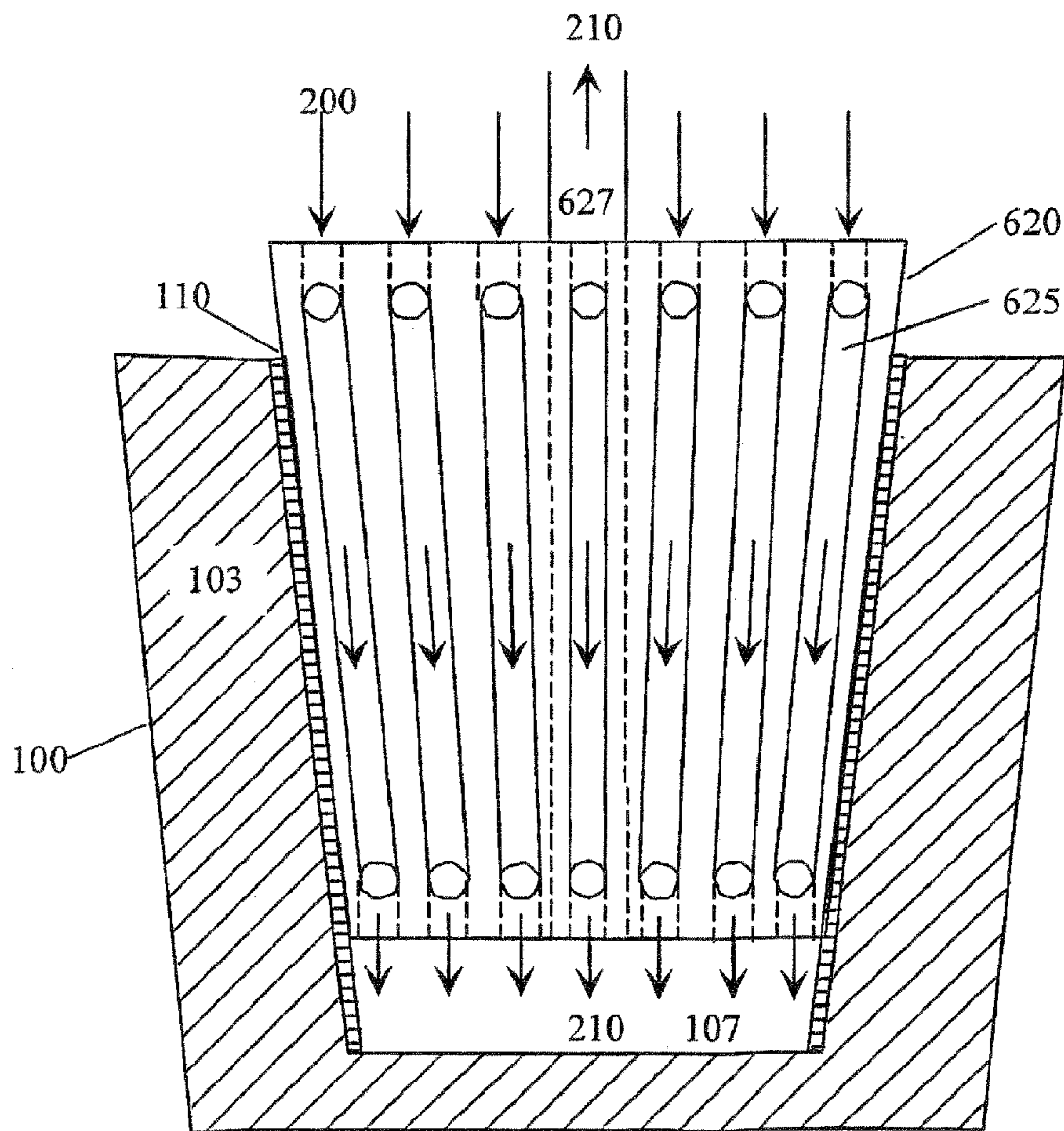


Fig. 13b

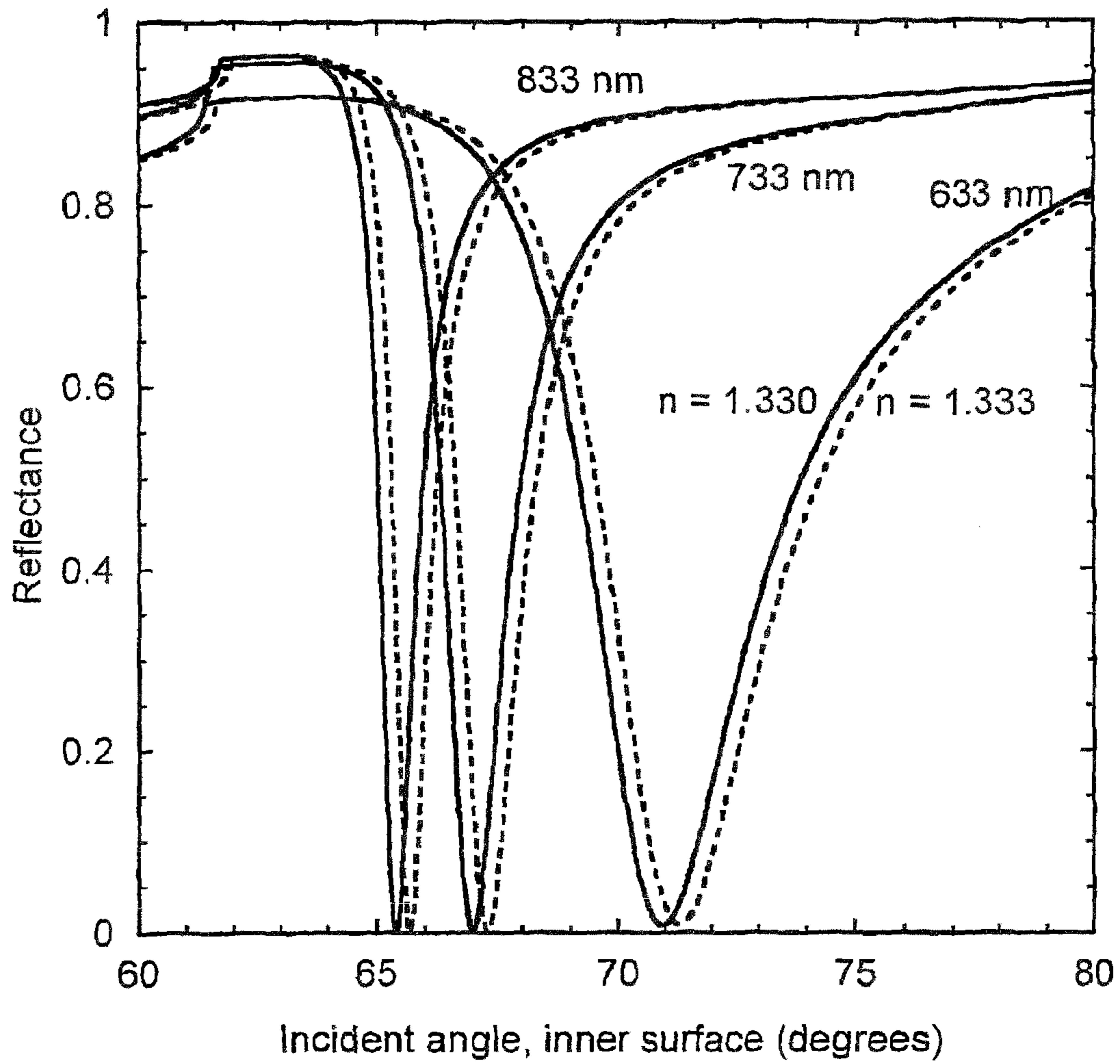


Fig. 14

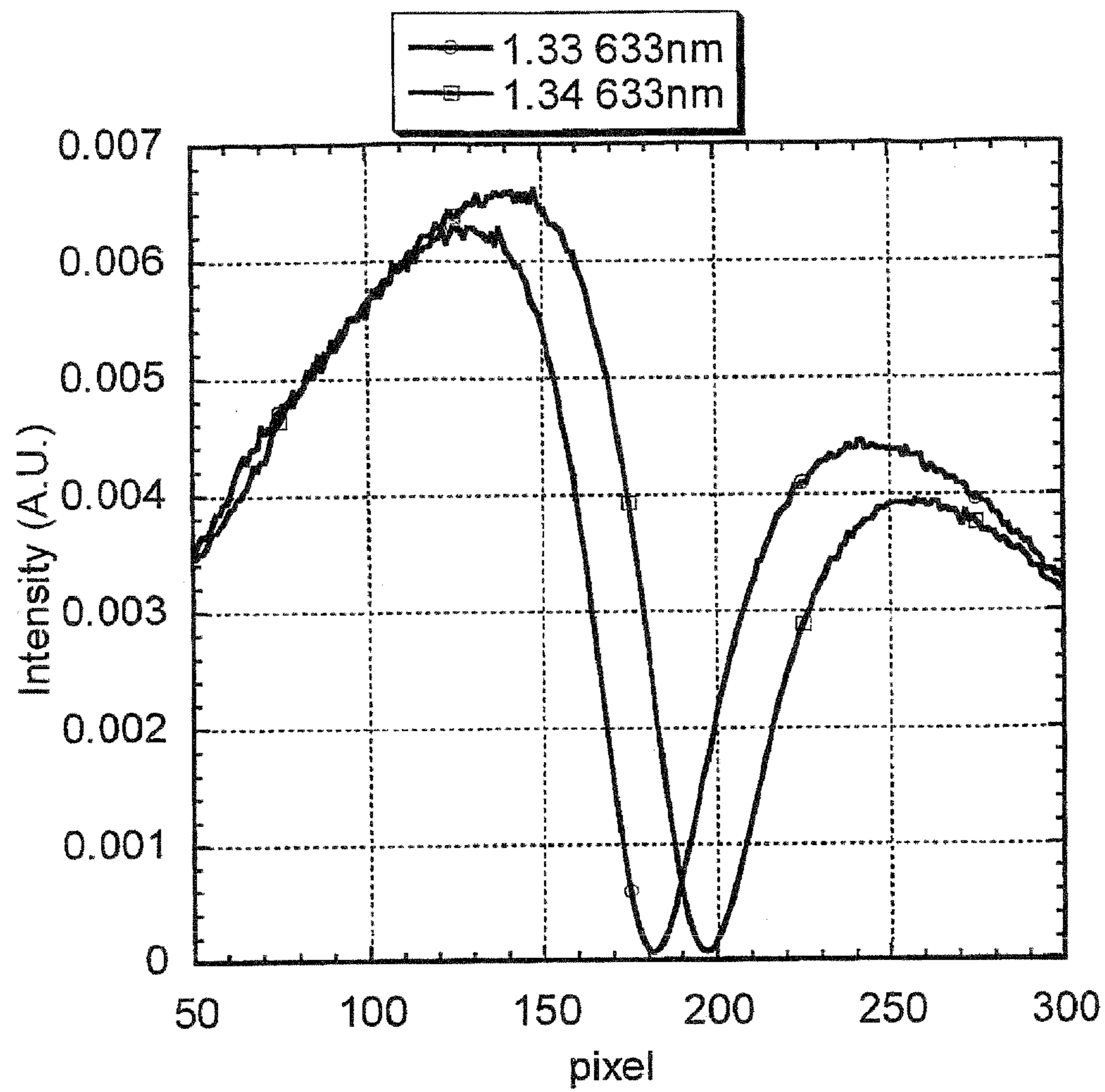


Fig. 15

Duran Glass, Different Thickness ratio: outer/inner diameter

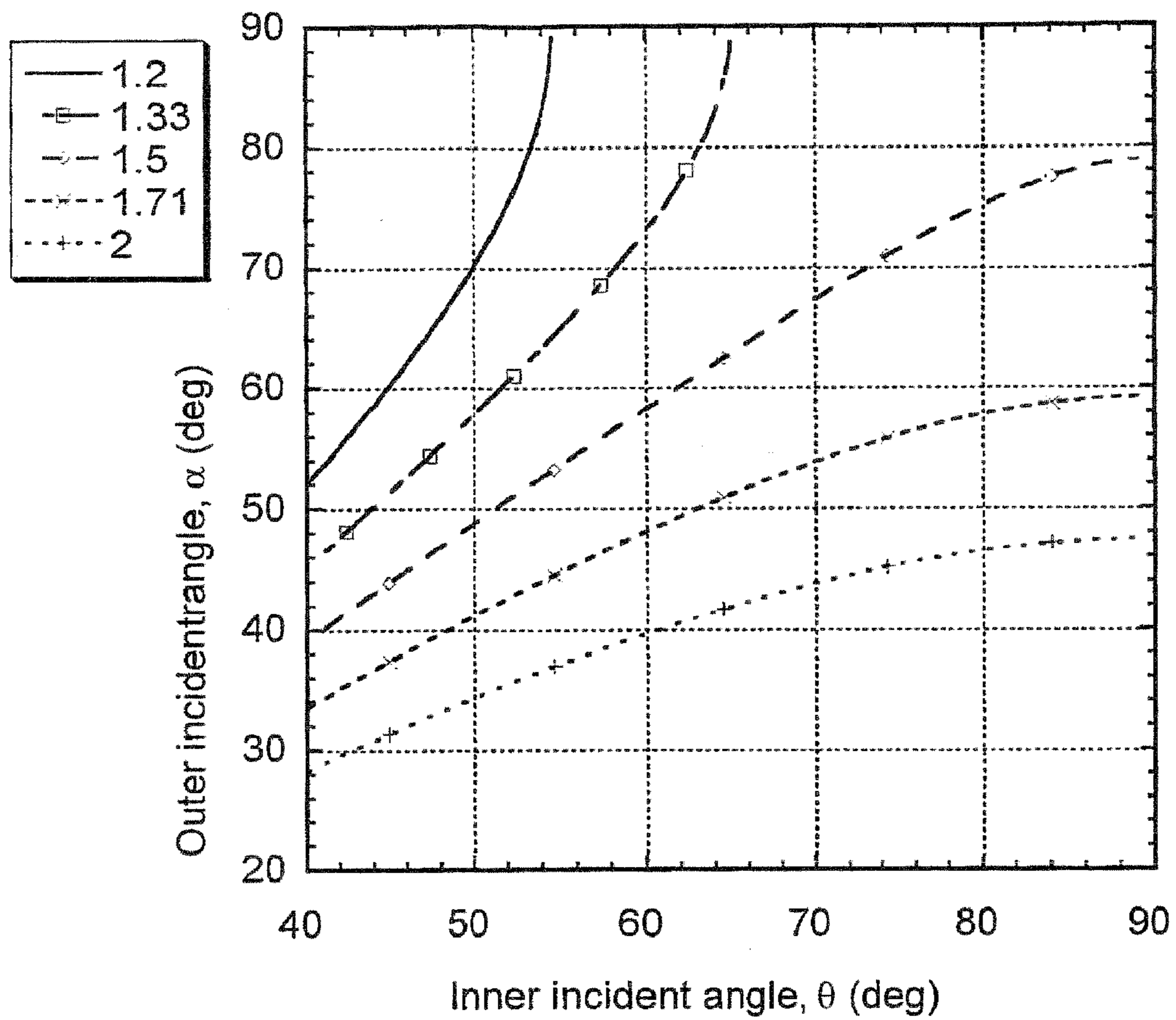


Fig. 16

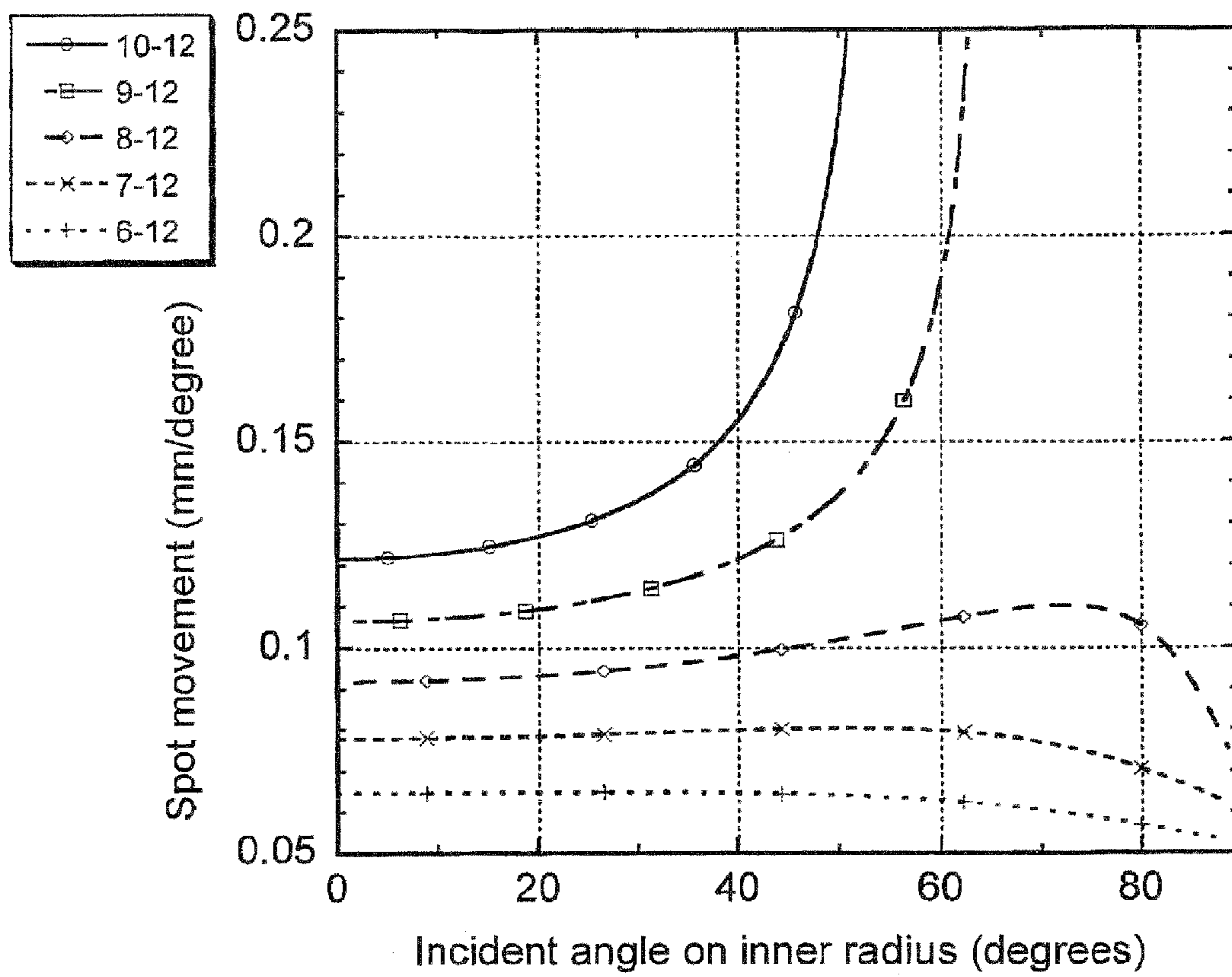


Fig.17



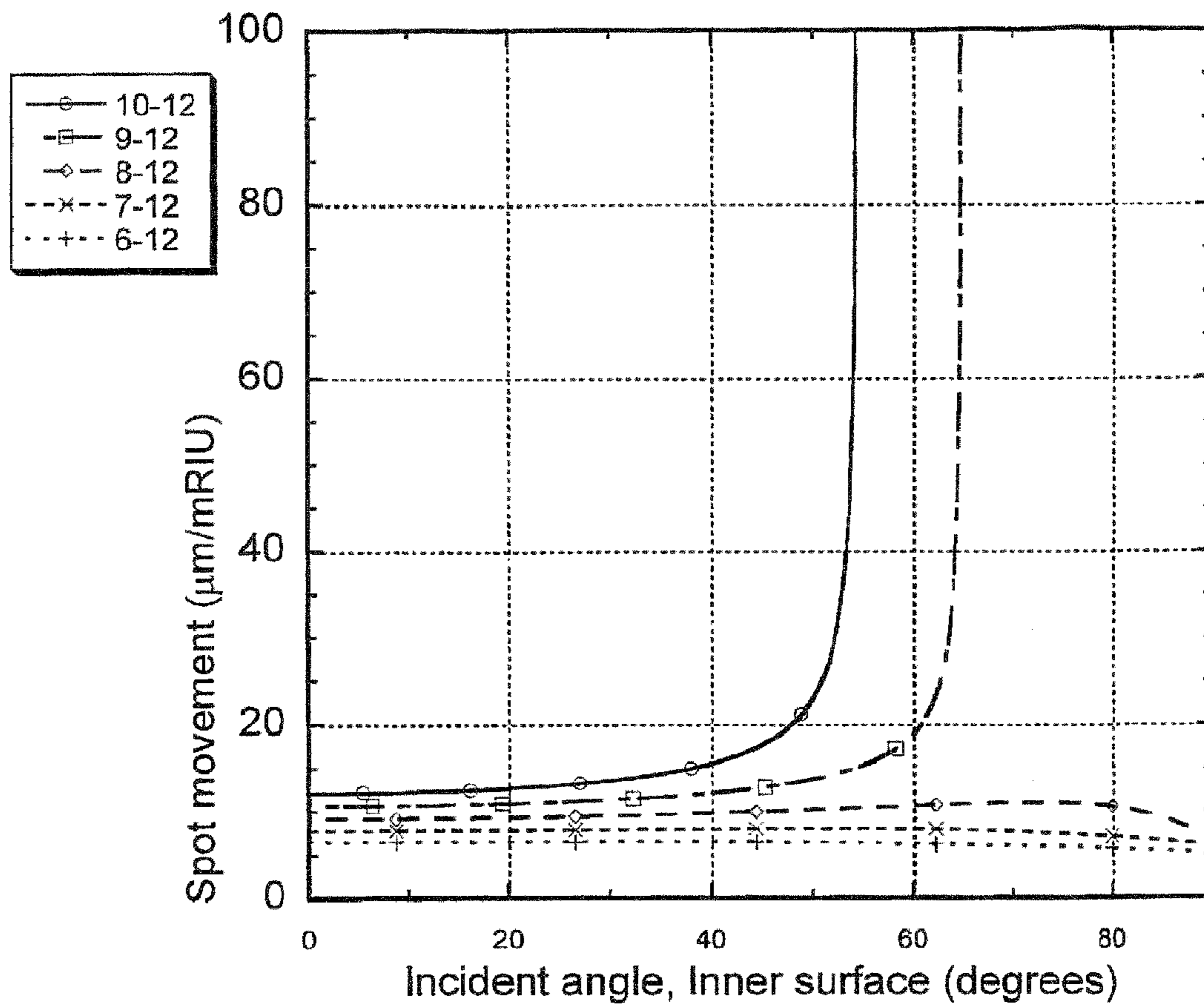


Fig. 18

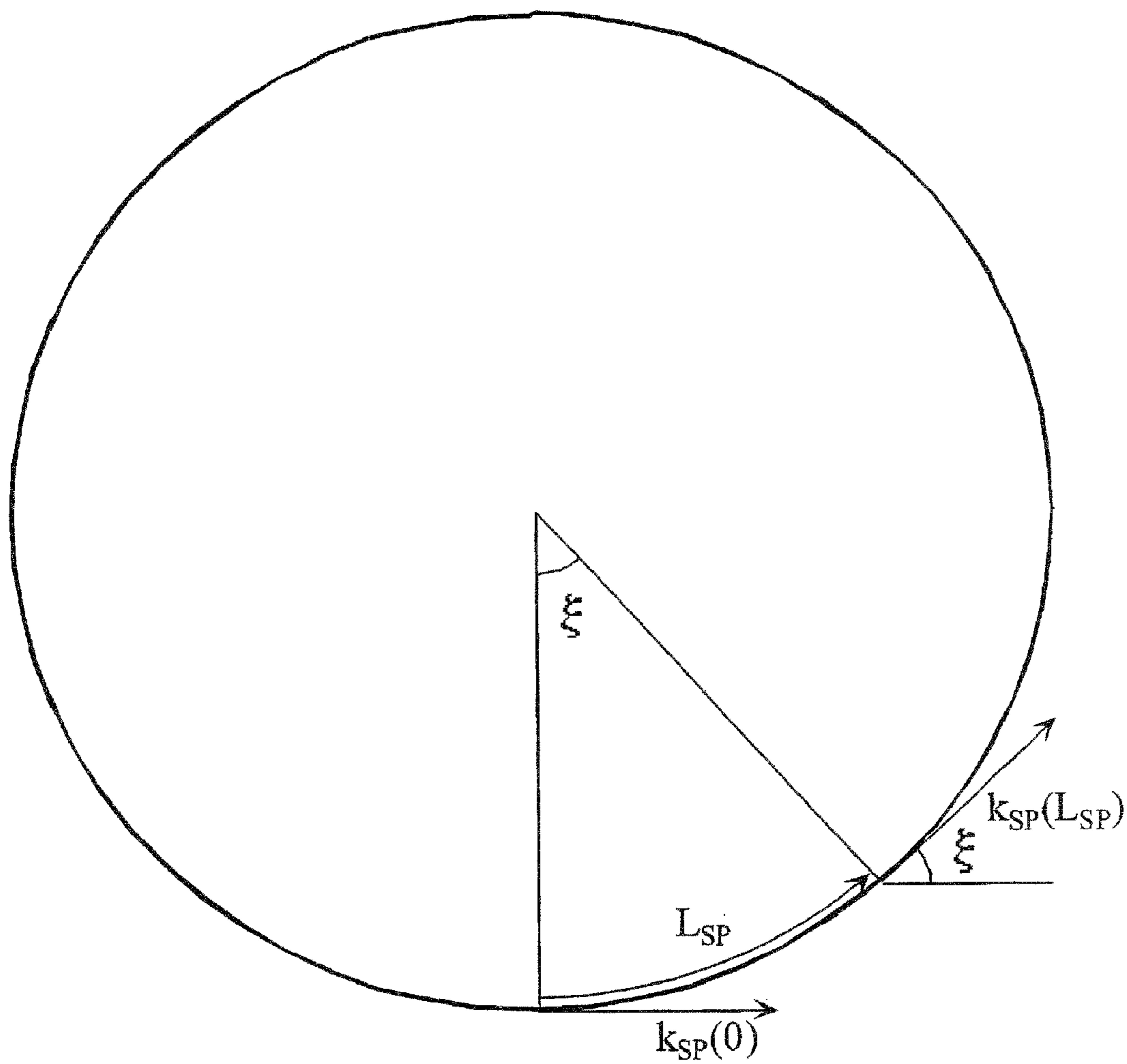


Fig. 19

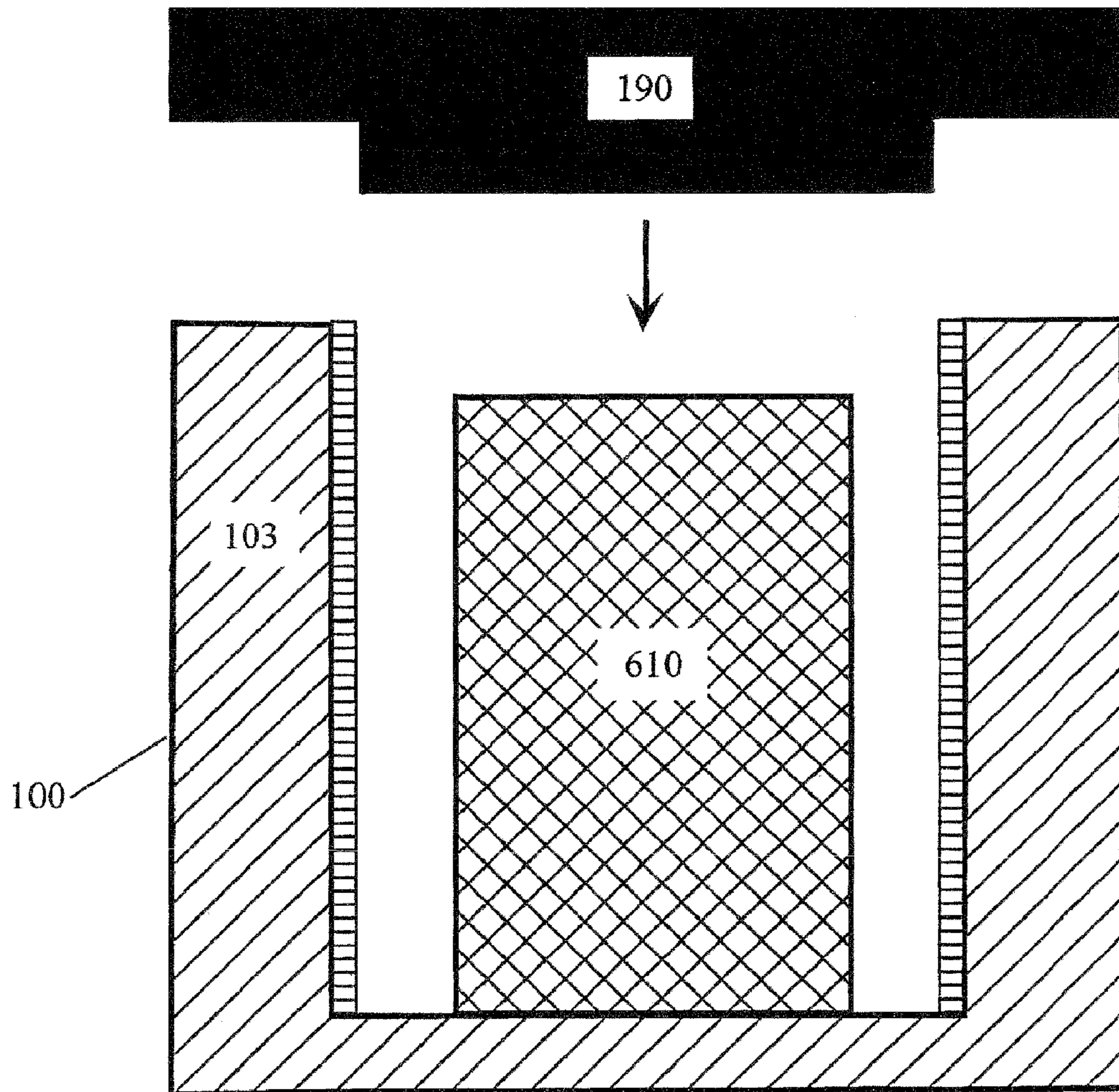


Fig. 20a

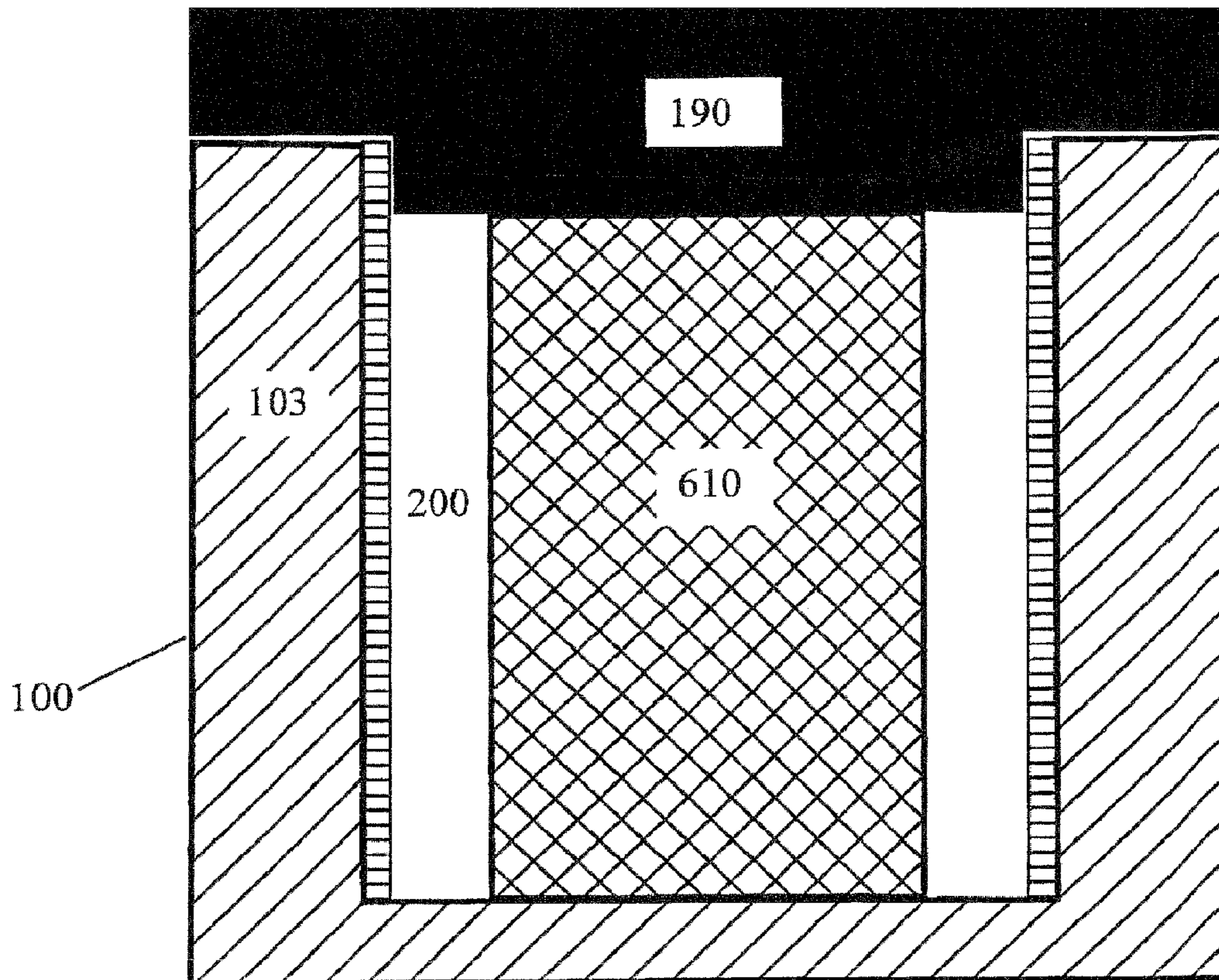


Fig. 20b

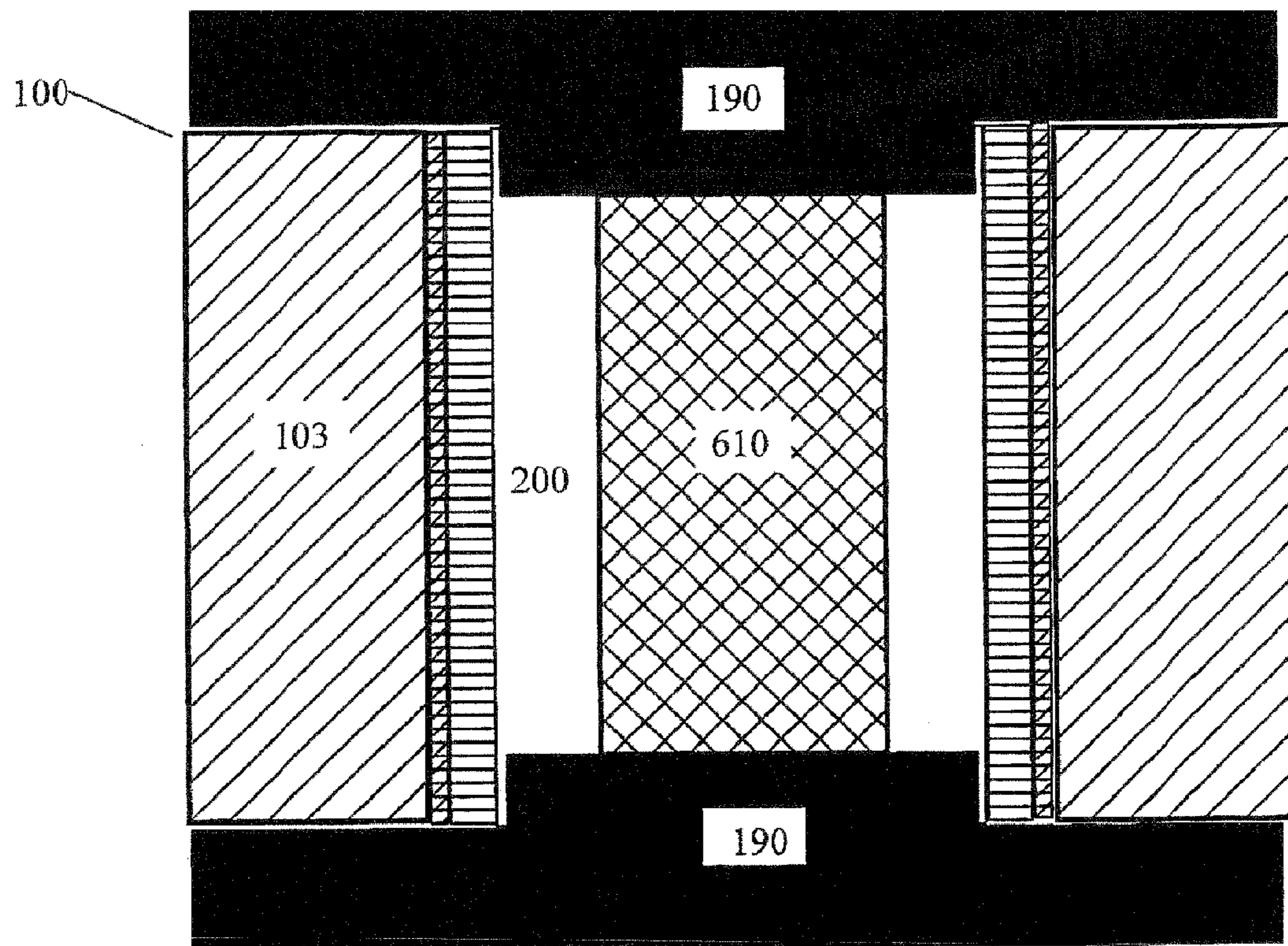


Fig. 20c

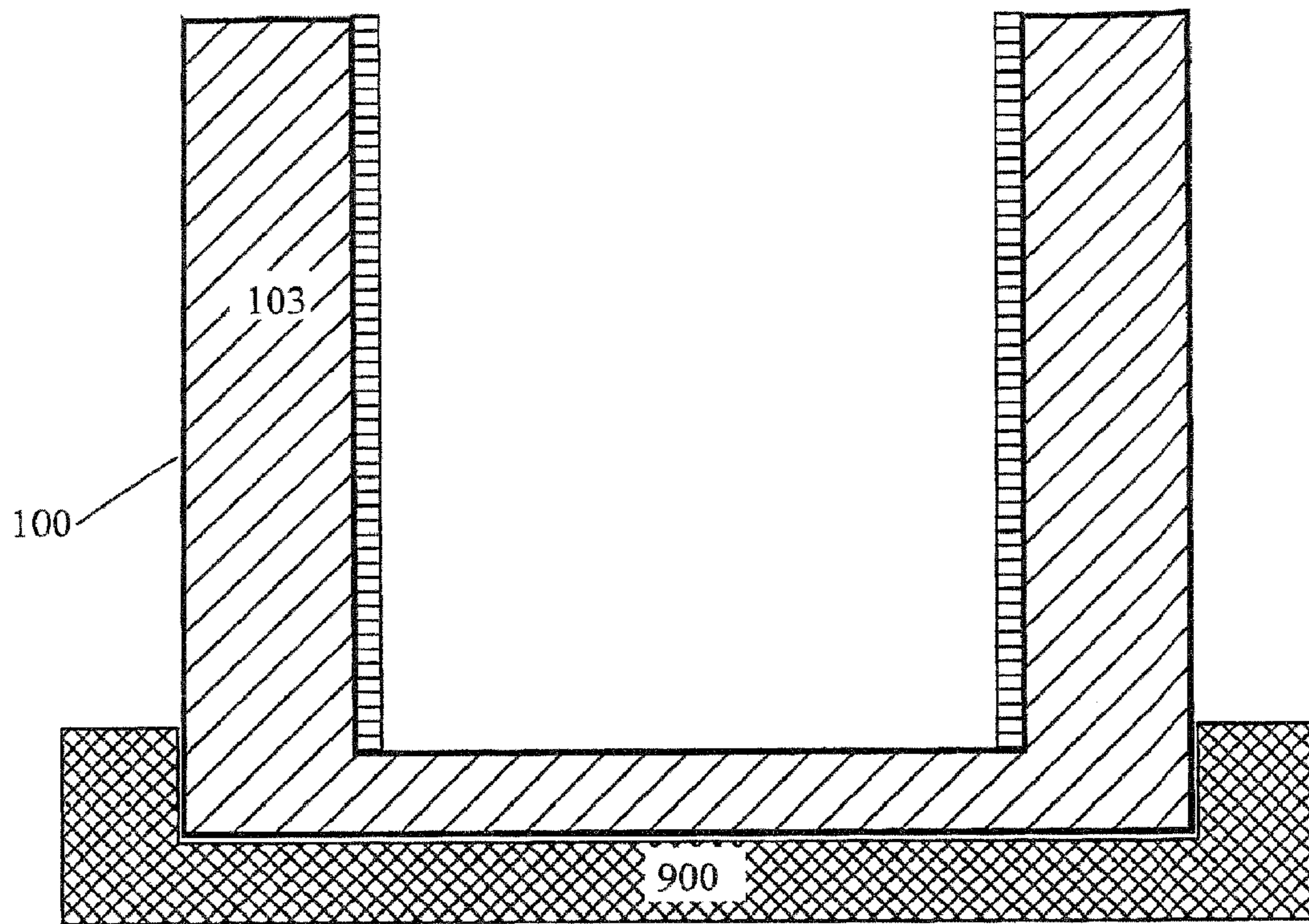


Fig. 21

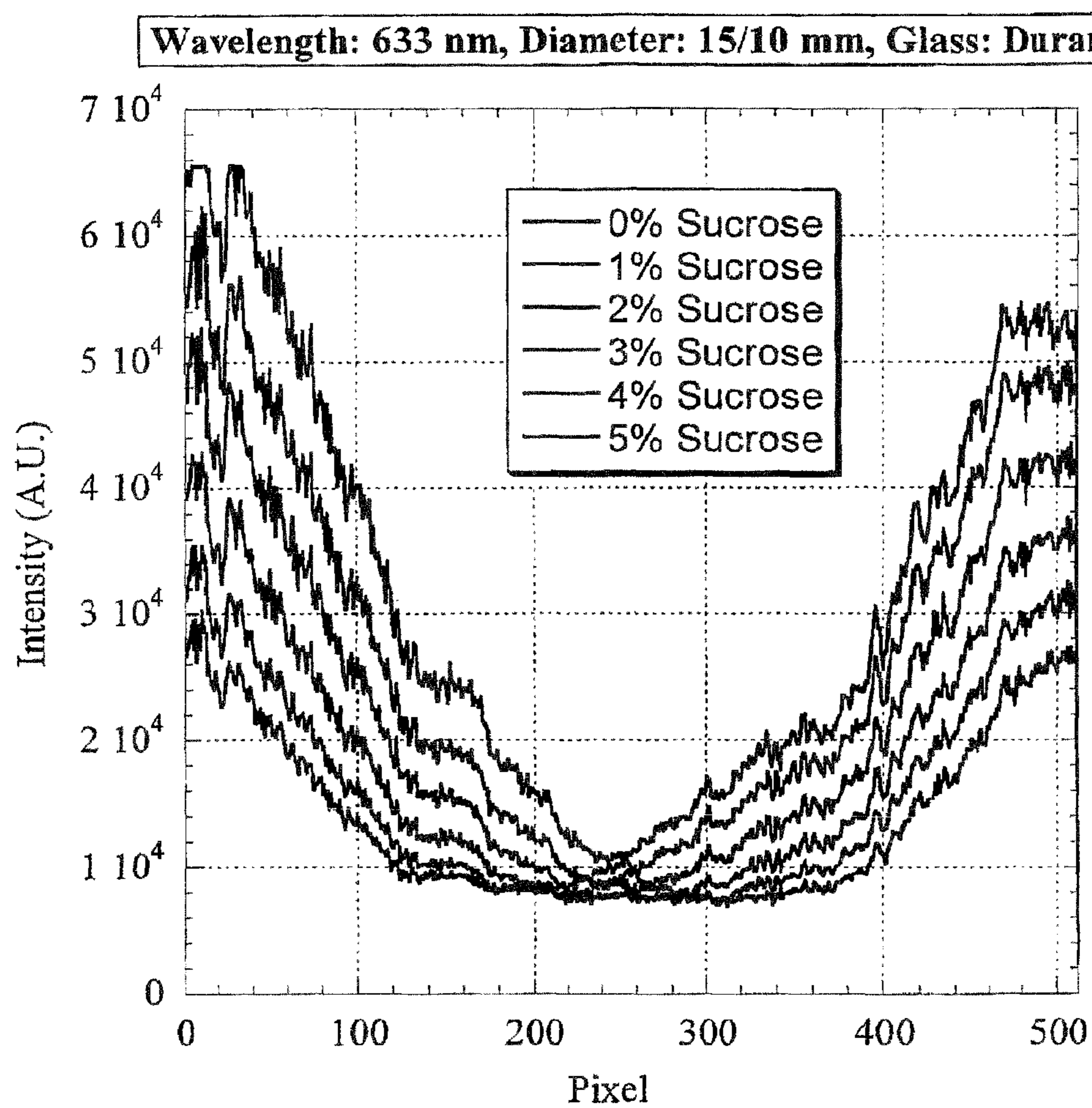


Fig. 22a

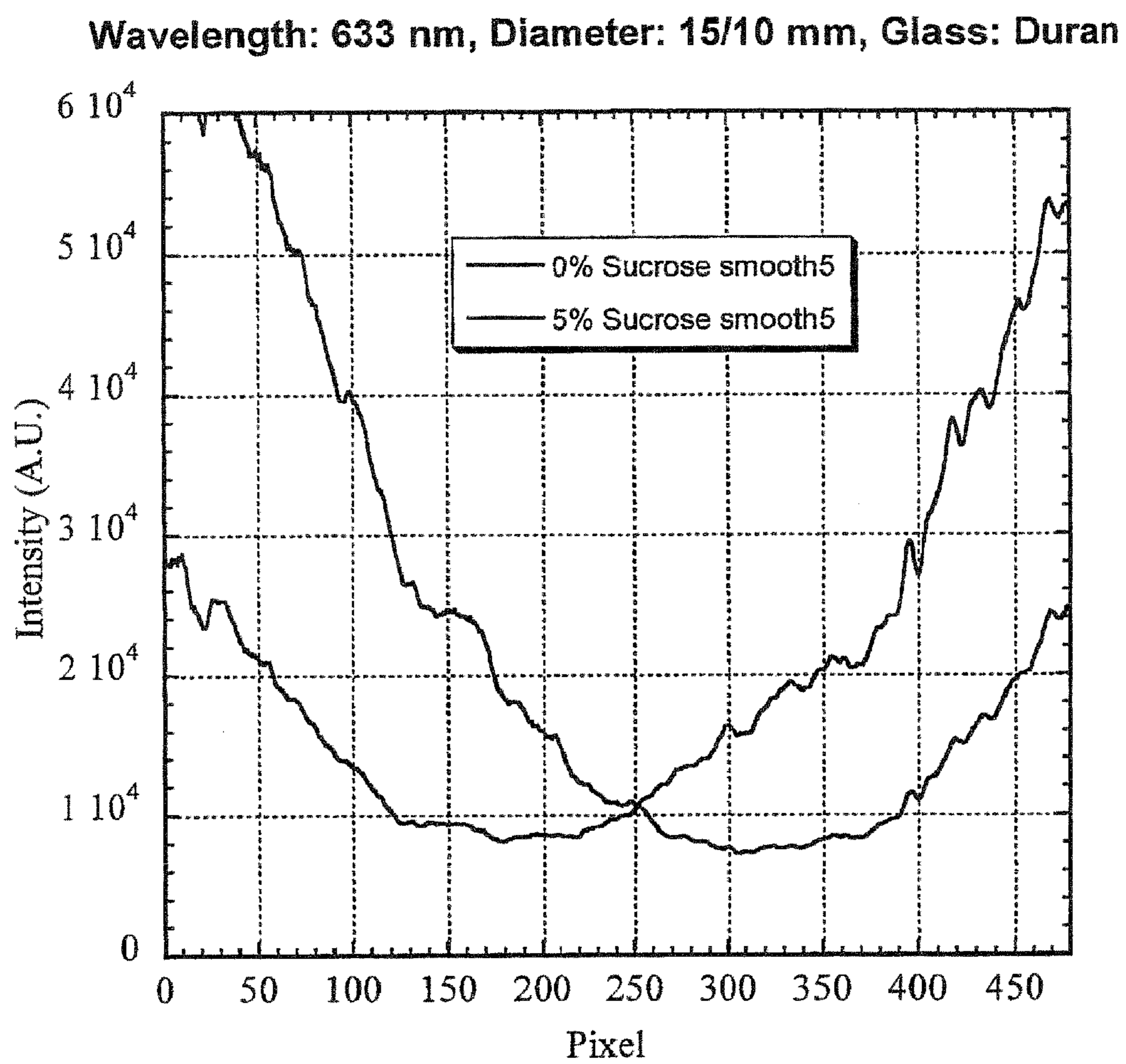


Fig. 22b



100

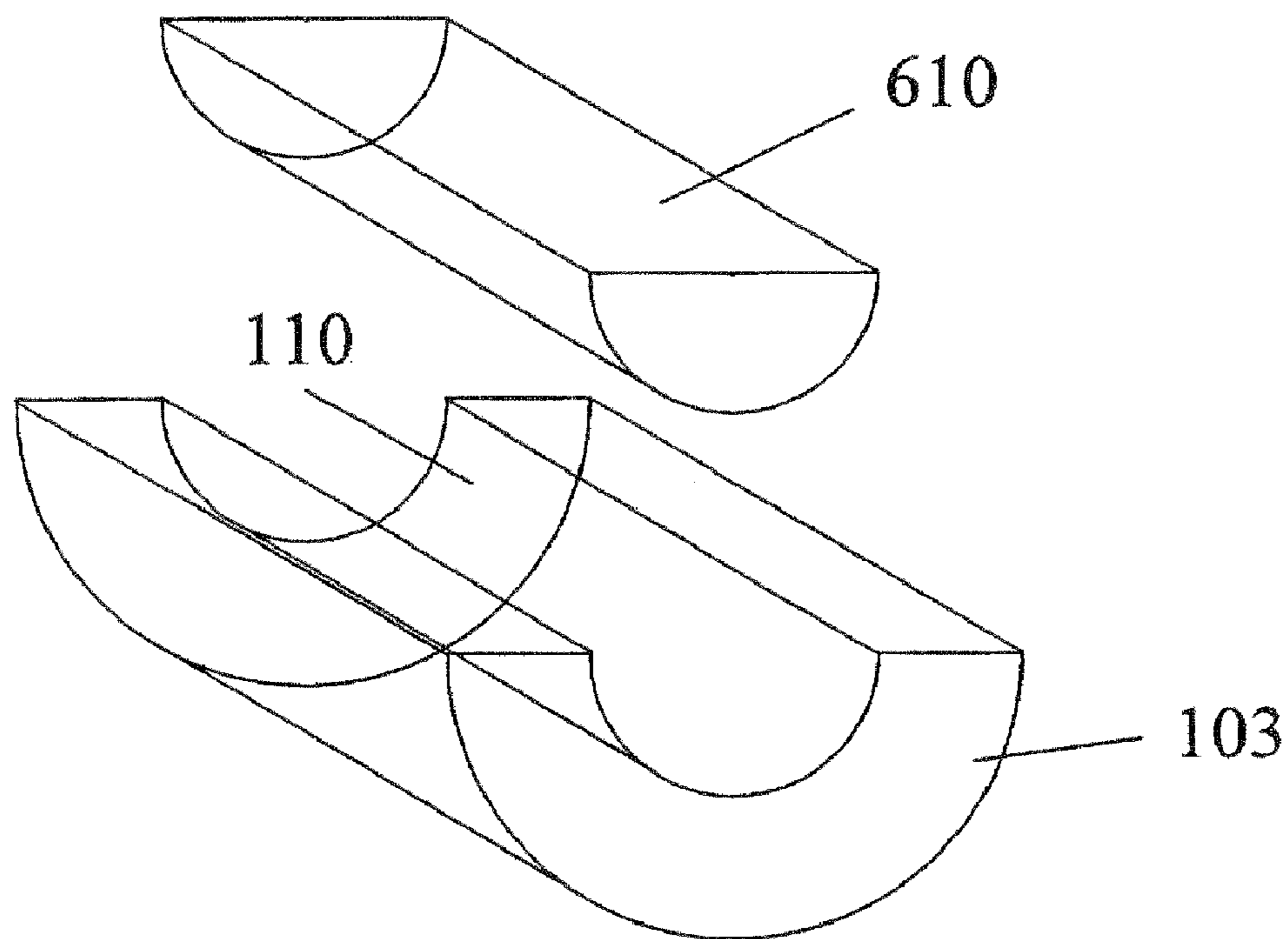


Fig. 23a

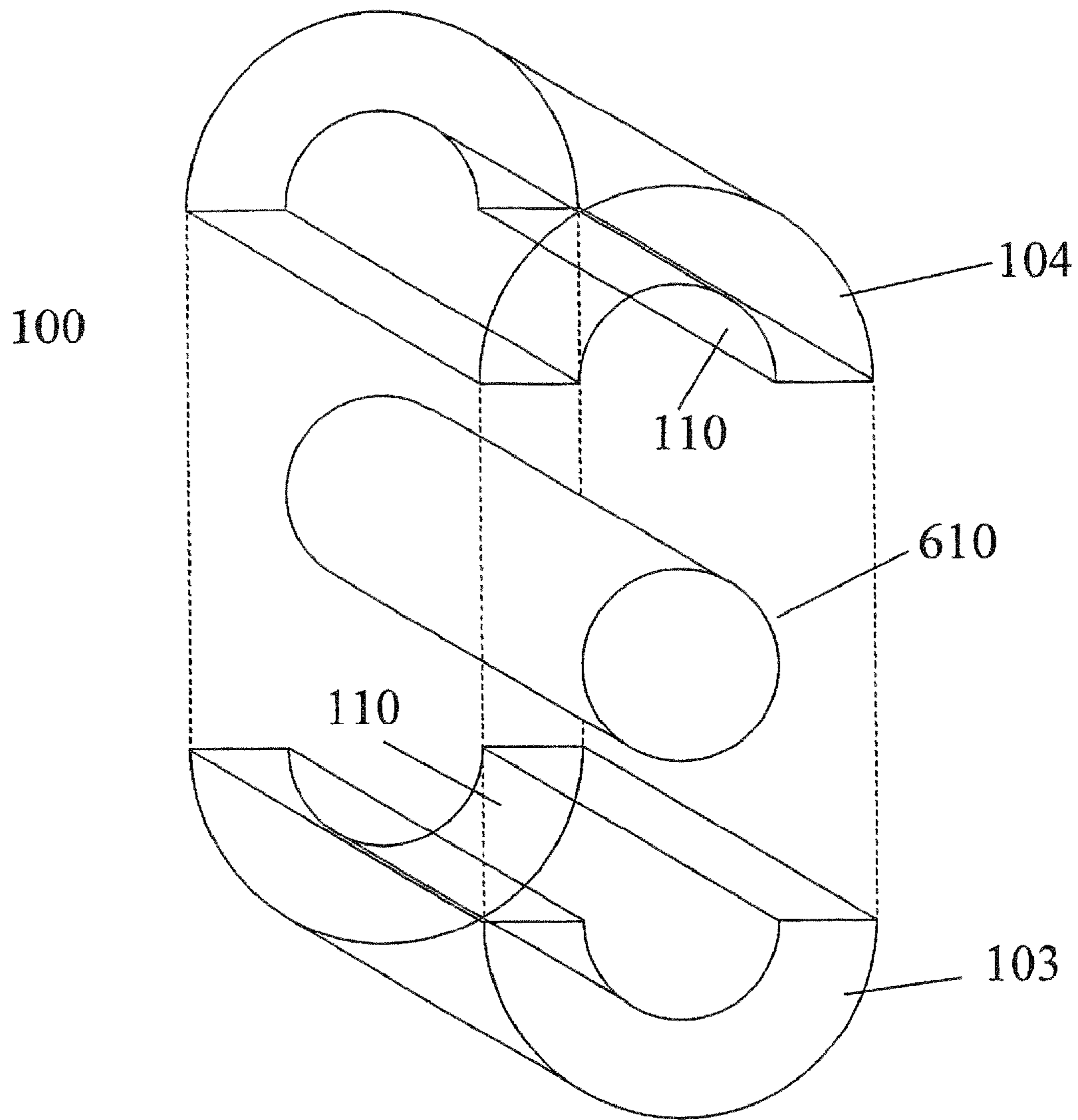


Fig. 23b

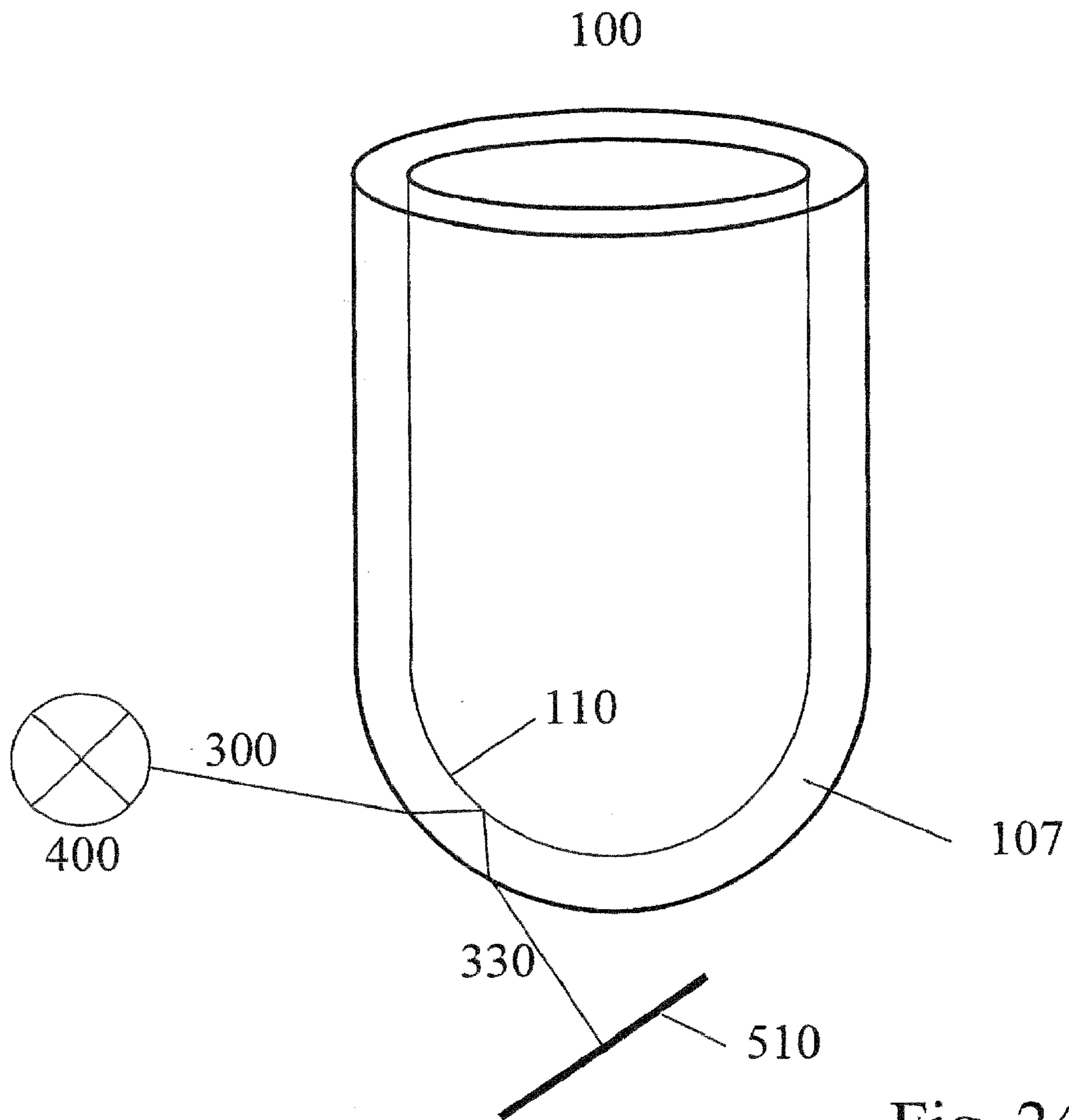


Fig. 24

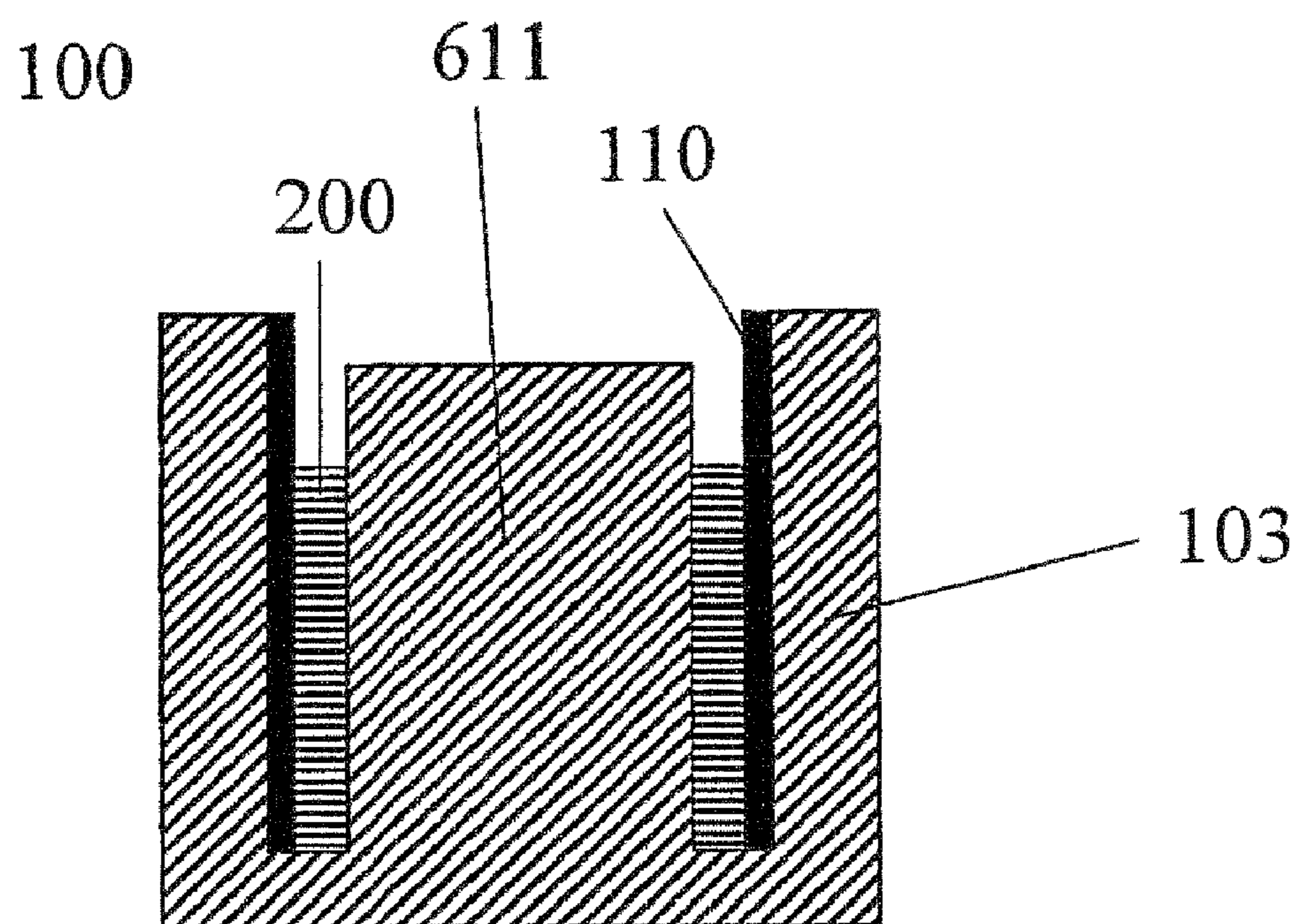


Fig. 25

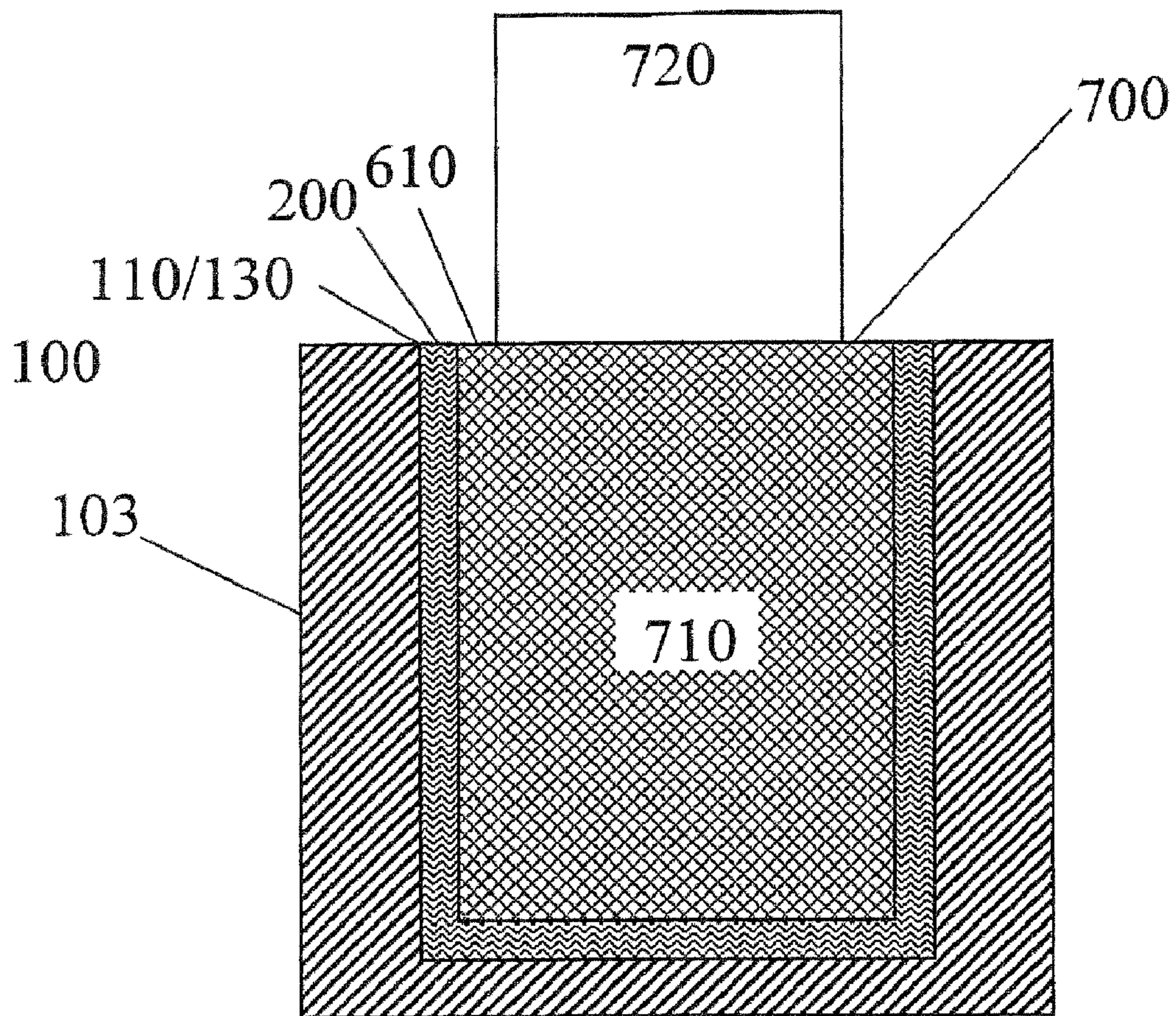


Fig. 26a

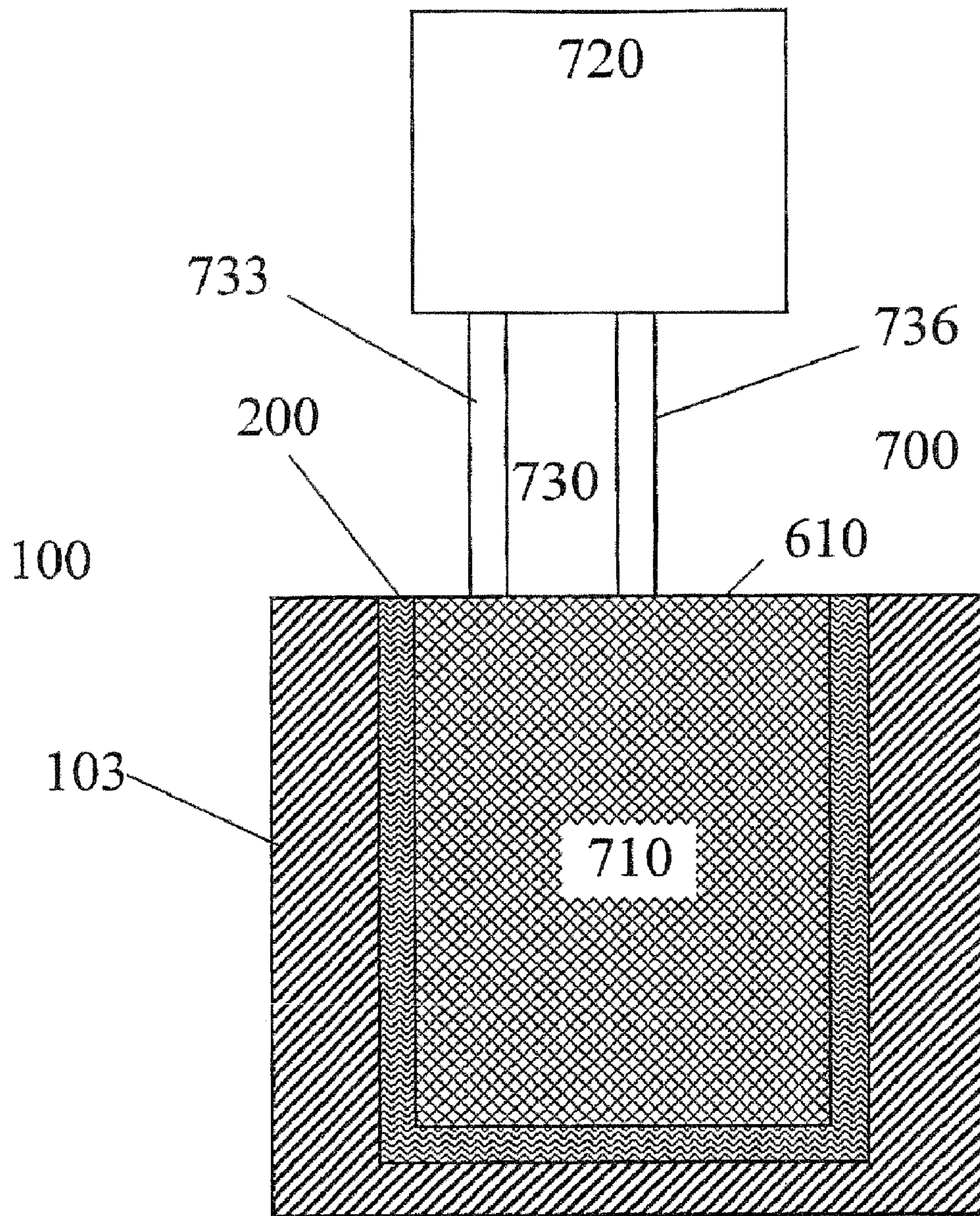


Fig. 26b

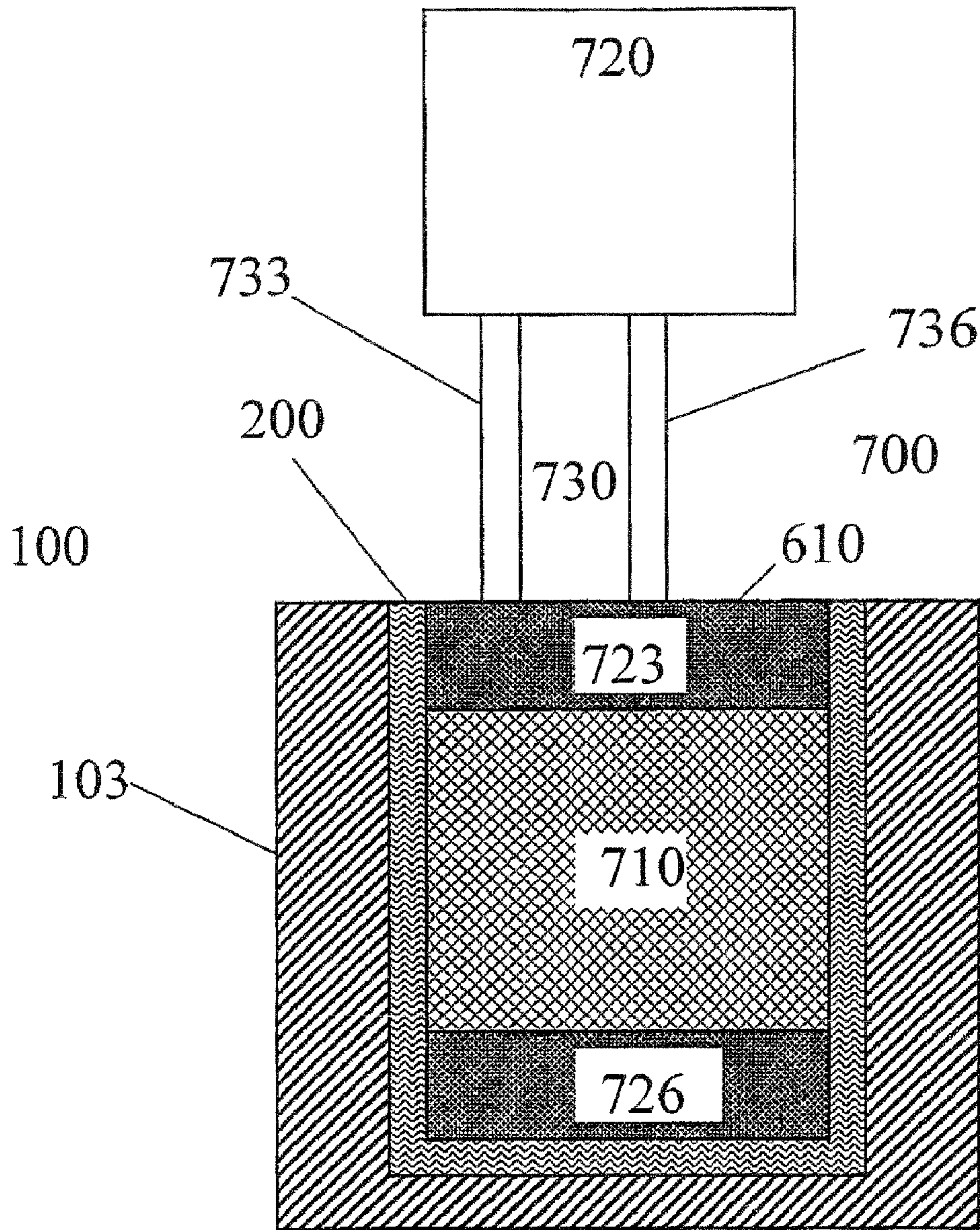


Fig. 26c

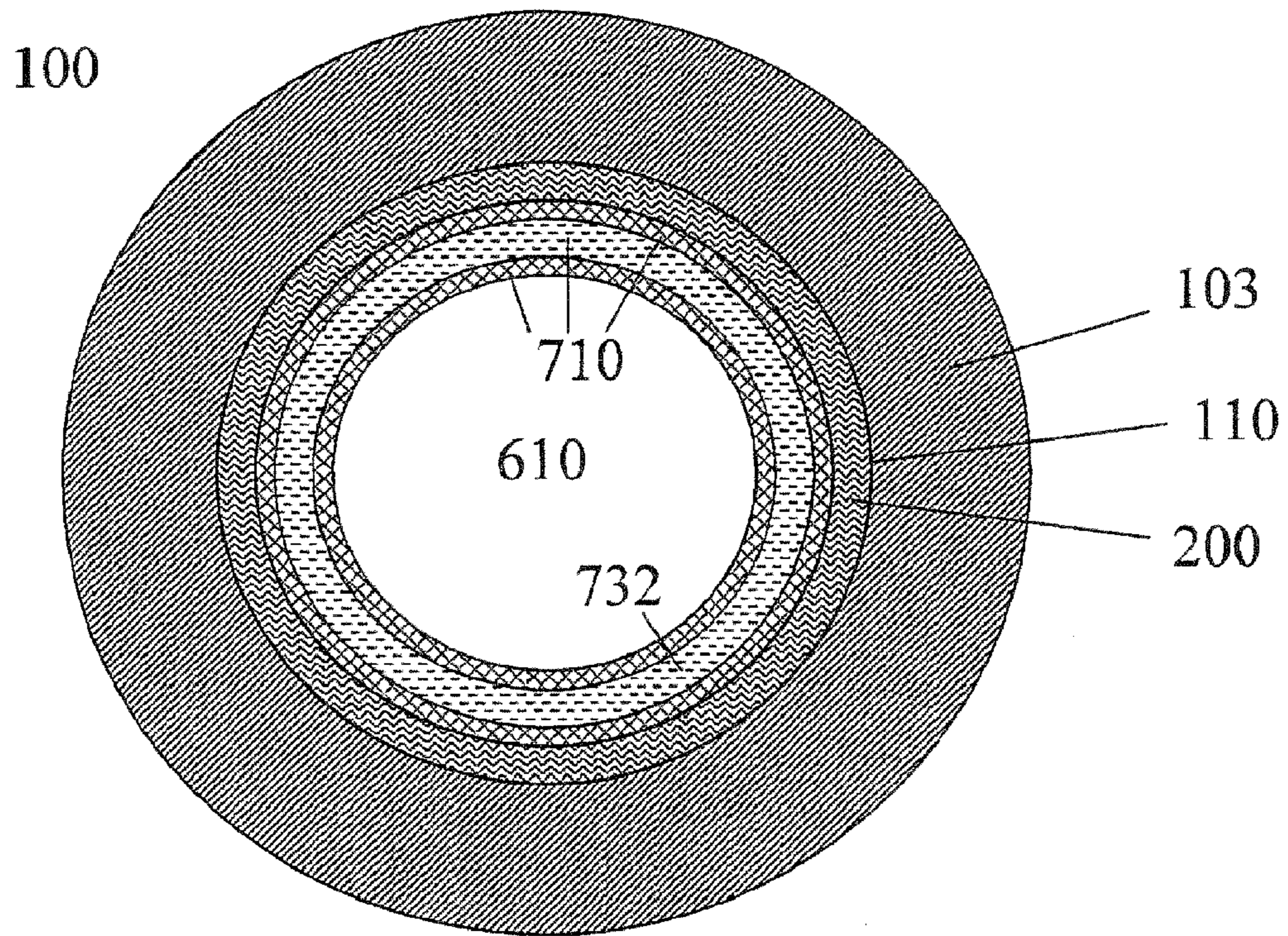


Fig. 27



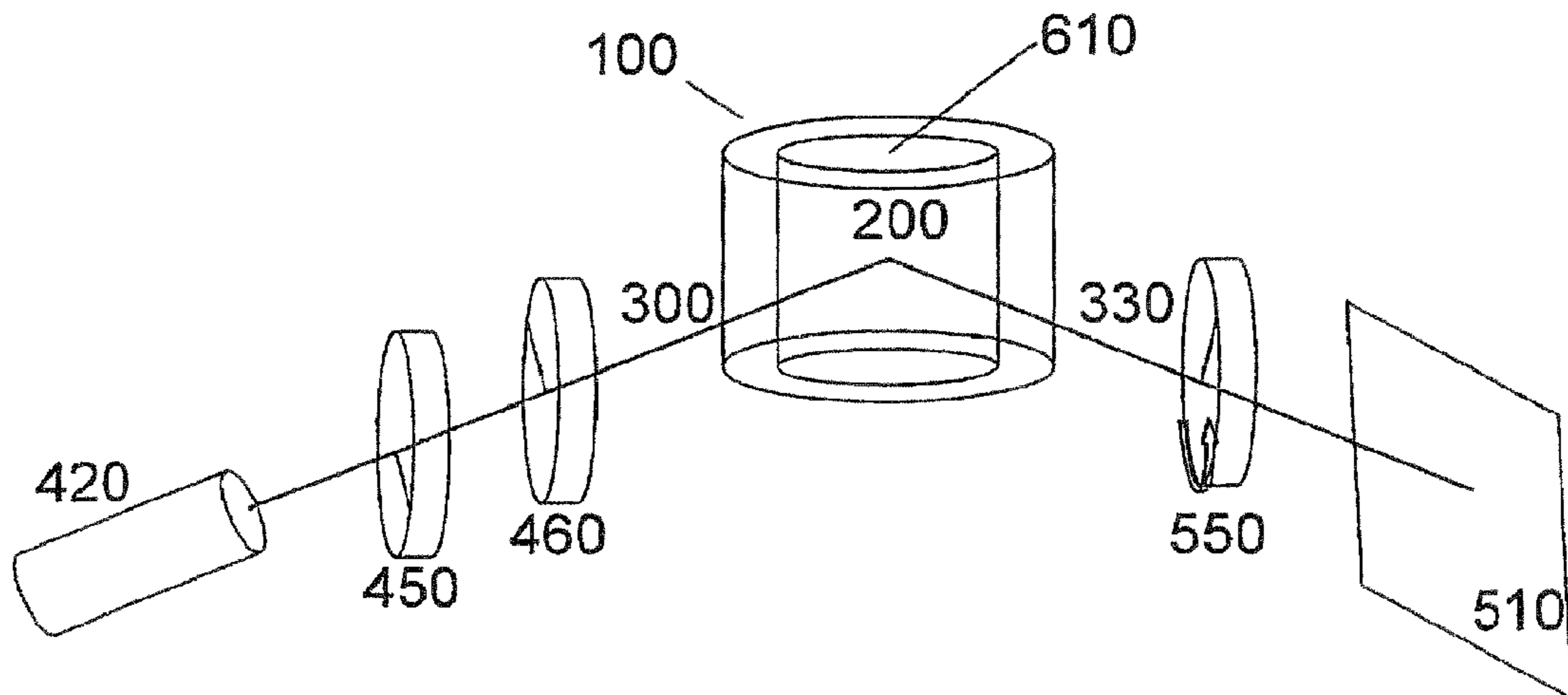


Fig. 28

## 1

## SURFACE PLASMON APPARATUS

## CROSS-REFERENCE TO RELATED APPLICATION

This is a divisional of U.S. patent application Ser. No. 12/447,825, which is a U.S. National Phase Application under 35 USC 371 of International Application PCT/SE2007/000950 filed Oct. 29, 2007, which in turn claims priority from Swedish Application No. SE 0602286-7 filed Oct. 31, 2006, the entire disclosure of each of which is incorporated herein by reference.

## FIELD OF THE INVENTION

The present invention relates to an apparatus for optical surface analysis of a plurality of sample areas on a curved sensor unit using surface plasmon resonance. The invention is particularly concerned with a rotational symmetric container or tube or a device using an angular part of a rotational symmetric container, using surface plasmon resonance (SPR), suitable for use in biological, biochemical, chemical and physical analyzing, as well as gas sensing, where the container or tube may be fixed or movable, e.g. rotating or oscillating.

## BACKGROUND OF THE INVENTION

There is an interest in surface sensitive techniques for analyzing the amount of molecules and larger substances, their chemical and physical properties, and their interactions with other molecules or materials. Properties that are of interest are e.g. the concentration of molecules in a solution or gas, the surface concentration of molecules on a sensor surface, the reaction kinetics of interacting substances, the affinity of the substances, allosteric effects or epitope mappings. Examples of interacting substances are antigen-antibody, protein-protein, receptor-ligand, DNA-DNA, DNA-RNA, protein-DNA, peptides-proteins, carbohydrates-proteins, glycoproteins-proteins, etc. There is also an interest in measuring the concentration of different gases or liquids, which can be performed by measuring the change in the optical density of a sensing material, e.g. polymer films, that are affected by some substance, e.g. fluid or gas [1, 2]. Change of conformation or formation of new materials on a surface, e.g. blood coagulation and fibrinolysis are also of large interest [3] [4] [5] [6]. There are many techniques that are suitable for these tasks, e.g. surface plasmon resonance (SPR), resonant mirror, grating couplers, interferometers, surface acoustic wave (SAW), Quartz Crystal Microbalance (QCM) etc. SPR is a popular technique, which have been proven to be both sensitive and reliable. Areas of application are e.g. measurement of concentration of substances in biological research, biochemistry research, chemical research, clinical diagnosis, food diagnostics, environmental measurements, etc. Kinetic measurements can be used to determine rate constants as ( $k_{on}$ ) and ( $k_{off}$ ). Affinity measurements can be used to determine equilibrium association ( $K_A$ ) or dissociation ( $K_D$ ) constant as well as avidity.

SPR is a well-known phenomenon that is a bound electromagnetic wave, due to oscillations of electrons at the interface between a plasma and a dielectricum. The surface plasmon can only exist at an interface between the plasma (e.g. a metal) and the dielectricum. A change in the optical constants of the dielectricum will change the propagation constant of the surface plasmon. The surface plasmon can be excited by light if the propagation constant of the light parallel to the interface is

## 2

equal to, or close to, the propagation constant of the surface plasmon. Normally, the Kretschmann configuration [7] is used where a thin metallic film is applied on a prism, having a higher refractive index than the measured sample. This is also denoted backside illumination, because no light is propagating in the sample medium. The surface plasmon is then evanescently excited under total internal reflection, i.e. at an incident angle, normal to the surface, larger than the critical angle. At a certain incident angle, the component of the wave vector parallel to the surface meets the real part of the complex wave vector for a surface plasmon, and hence the light will couple into the surface plasmon and propagate at the interface between the plasma and the dielectricum. The surface plasmon will reradiate into the prism, and for a certain thickness of the plasma a destructive interference will occur, leading to zero or close to zero intensity of the reflected light. For a smooth surface of the plasma, coupled light will be absorbed in the plasma and generate heat. When molecules bind close to the interface (within the probe depth of the surface plasmon) the interaction can be detected by a shift in the resonance condition of the surface plasmon. This can be detected as a shift in a reflected light intensity. This is also the case when a layer changes its density due to conformal changes or external interference.

The most common way to design an SPR apparatus is to use a prism (triangular, hemispherical or an arbitrary shape) and apply to the prism a separate planar substrate carrying the SPR-metal. In this case it is necessary to use a refractive index matching material between the prism and substrate to obtain good optical coupling. The material can either be an opto-gel [8] or a refractive index matching fluid. The use of an opto-gel has the disadvantage of wear, optical imperfection and high cost. If a refractive index matching fluid is used, a circumstantial procedure of application and cleaning is needed, besides the extra cost. There are other configurations, e.g. a prism with evaporated metal film [9] and SPR-light-pipe [10], that do not need an optical coupling medium. Yet another configuration that doesn't need to use an optical coupling medium, is a capillary [11]. In another configuration, the fiber optic SPR [12-14], one can refrain from using an optical coupling medium, but for the case of an exchangeable probe, a coupling medium is needed.

The surface plasmon resonance (SPR) phenomenon was already described in 1959 [15] and SPR apparatuses for thin adlayer analysis have been thoroughly described since 1968 [16, 17]. SPR setups for biosensing were used for the first time in 1982 [1] and for imaging applications in 1987 [18, 19]. With imaging SPR, also denoted SPR microscopy, new applications arise, e.g., label free-real-time-multi spot biochemical analyses [20, 21], which can increase the throughput tremendously. The pioneering work on imaging SPR was undertaken by Knoll et al., who investigated surfaces patterned with Langmuir-Blodgett films [22, 23]. They also investigated the physical aspects of the technique, including lateral resolution [24], and proposed different setups, e.g. the rotating grating coupler [25].

Most SPR setups utilize a separate planar sensing substrate, refractive index matching layer, and a coupling element, e.g. a prism. Using a planar sensor surface with multiple sensor areas arranged in a two-dimensional way, means that there are a couple of somewhat cumbersome (and expensive) ways to read the optical output from the surface plasmon resonance device. In principle, there are two methods to perform a readout, by mechanically scan the sensor substrate, or the use of imaging optics. Not only is the read out complex,

but the distribution of samples for investigation is often very complex, with use of valves and channels or expensive autosamplers.

However, there are approaches that do not use planar substrates. Chinowsky et al. are using an approach [1,1], where a capillary tube is used, U.S. Pat. No. 6,480,282. The tube wall itself is the then the coupling medium, and no index matching fluid is necessary. The capillary is useful for multi sensor configuration, where the other techniques can be critical angle detection, fluorescence, chemiluminiscence, adsorption or Raman scattering. It can, with difficulties be used as an axial multispot sensor, and with severe difficulties may be with some sensors spots radially separated. However, this approach has several disadvantages. It is very difficult to cover the inside of the capillary tube with a metal of precise thickness, due to the small diameter and long length. Capillary tubes are not normally high precision optics, which will distort optical images. The capillary device is suitable for one or a few sensing spots, and the small diameter means that there will be difficulties to manufacture device with many sensors spots. The capillary device has typically a relatively large diameter, e.g. 400  $\mu\text{m}$ , compared to a thin flowcell, e.g. 10  $\mu\text{m}$ , making the capillary tube inefficient regarding small sample volumes, and mass transport.

Another structure similar to the capillary device is proposed by Nakaso Nobutaka, Japan patent JP2003-294616. It uses a curved cavity, with a diameter of typically 20  $\mu\text{m}$  e.g. a cylinder, or part of a sphere, which is formed in a transparent block. The block has a typical dimension of 2x2 mm, and is typically cut from a glass wafer of thickness 0.2 mm. The different blocks, having different recognition molecules, can be stacked. The surface plasmon is excited radial inside the cavity.

Yet another structure that uses a curved sensor surface is proposed by Atsushi et al. Japan Patent JP2003-075333. This device uses curved cavities for recognition sites, preferably many cavities are used for a multi spot sensor. The cavity can be cylindrical, spherical or an arbitrary curvature, and the surface plasmon is excited radially, as described by Chinowsky. However, the outer surface, which is hit by the incident light, is planar. The proposed cavity is not intended for multiple sensor areas.

Using a small radius at the surface plasmon carrying surface, will not only lead to mismatch between wavevector for surface plasmon and incident light, but also leads to difficulties to obtain small light beams and smooth reflectance curves.

A SPR-setup utilizing a convex curved SPR-supporting surface is described by Rooney et al., Sensors and Actuators B, 26 Apr. 2006. There is also described a SPR-setup consisting of a tubular cup, where a SPR-supporting layer is present on the planar bottom, EP 1186881, Haya et al., 2002 (Fuji Photo Film Co).

#### SUMMARY OF THE INVENTION

In view of the shortcomings of prior art devices, the object of the invention is to provide an improved SPR device enabling e.g. analysis of multiple samples in a simple manner, and also which is easy and inexpensive to manufacture.

This object is achieved with a device as claimed in claim 1, by a sensor unit comprising a transparent sensor structure forming at least one wall of a cavity, the wall being defined by a concave inner surface and a convex outer surface; wherein the inner surface is provided with a layer of a conductive material capable of supporting a surface plasmon; a flow structure in said cavity so as to form at least one compartment

for sample between the flow structure and the inner wall of the cavity. The flow structure is suitably provided as an insert fitting in the cavity, the insert being configured and structured so as to provide the required channels, compartments or other spaces necessary to accommodate a sample. The configuration of the inset makes it also ideal for precise temperature control of the sample.

Thus, a new configuration of a surface plasmon resonance set up is presented. A sensor unit device with convex curved outer surface and concave inner surface, in one or two dimensions is used (e.g. cylinder or hemisphere). The inner surface is, at least partly, covered with a conductive layer, having the possibility to carry a surface plasmon. The device is a development from the planar structure having the possibility to incorporate multiple recognition sites in one or two dimensions. The curvatures of both the outer and inner surfaces means that the structure itself acts as an optical device eliminating expensive optics for light sources and detectors, e.g. a collimated incident beam will be transformed to a multi angle beam incorporating incident angles of interest by the curved outer and inner surfaces, and further spread to a detector, without the use of other optics. The use of macroscopic dimensions like in the conventional planar configuration, means that a flow structure is easily incorporated close to or in contact with the inner surface. The flow structure can in its simplest form be a wall close to the inner surface, incorporated in the optical structure. Such a wall can decrease sample volumes, without decreasing the advantage of macroscopic size of the sensor unit. With an insert, versatile fluidistics can be formed, both for immobilization of recognition molecules and samples. The curvature makes the device ideal for rotation, whereas different rows of recognition sites can be addressed both optically and by the flow cells in a single operation, eliminating expensive fluidistics. Especially, it is ideal as a diagnostic revolver, where one or more analytes are measured by one or more sensor spots axially, and different samples are analyzed by rotating the device, using new sensor spots, for the same or different analytes. The surface plasmon is excited radially, and one embodiment is a closed structure, another is an open one, where an angular part of the structure is missing, e.g. a tube which is axially cut in half.

In one embodiment a rotational symmetry makes the device ideal for monitoring of substances or the formation of new substances close to the wall. The shape of the device makes it ideal for multi-spot and large area detection. The shape makes it easy to scan the whole inner surface of the device by either rotating the device or by rotating the light source and detector. A rotational symmetry and macroscopic dimensions are ideal for an insert containing a flow system with sample distribution.

Use of macroscopic dimensions leads to simpler optics where beam widths from commercial components can be used with optical output that generates surface plasmon resonance angles of interest, e.g. for effective refractive indices from e.g. 1.33 to 1.45. Yet another advantage due to the relatively large radius if the inner surface is a little bent surface plasmon wave, and hence unwanted optical effects are avoided, e.g. interferences and bad coupling to the surface plasmon.

Not only is this new configuration perfect for measurements using surface plasmon resonance, it also leads to a much simpler instrument than is previously standard. The absence of a refractive index matching substance leads to both lower cost, less maintenance and simpler handling. The physical dimensions of the device mean that is easy to manu-

facture with high precision and at a low cost. Because the device can easily be sealed, it is also perfect for hazardous substances.

The invention can easily be fitted with an effective temperature control. A symmetric configuration of a SPR-tube/vial will reduce temperature gradients, and therefore it is suitable for accurate temperature control and high precision measurements.

The invention can be used for biosensing, e.g., for monitoring of chemical and biological reactions in real time with label-free molecules. A set-up with a collimated light source in a new configuration with gold as the supporting metal is described, both theoretically and experimentally. Simulations of the sensor response based on independently recorded optical (ellipsometric) data of gold show that the proposed optical set up working as a surface sensitive differential refractometer, where the performance is equal or better than contemporary instruments.

A closed configuration of the sample area makes it ideal also for gas sensing.

The main advantages of the invention are: Simpler and cheaper instrument and consumables can be produced.

The elimination of a refractive index matching layer (as used in the Kretschmann configuration) means that the handling and change of sensors surfaces are easily performed.

A rotational or partly rotational symmetry of the sensor surface means that it can be used in conjunction with a rotating bob creating shear rates and a more effective mass transport of the analyte to the sensing surface. The use of a conical bob and/or vial means that a gradient of the immobilized molecules can easily be created. Furthermore, due to different mass transport conditions, a gradient of the free analyte can also be created. These properties make it possible to perform an effective interaction analysis.

The design as a circular container means that a sample and flow handling system can be integrated inside the cavity. Even the waste can be integrated, e.g. for safety reasons.

The SPR tube, cup or vial can have the advantage of utilizing the rotational symmetric configuration, where the wall acts as a coupling medium, which increases the propagation vector parallel to the inner surface of the tube, making it possible to excite a surface plasmon. The use of the wall means that no refractive index matching medium, e.g. fluid, gel or film, is necessary, leading to a simple and cheap set up. Furthermore, the handling and change of sensor surfaces are then very simple. A rotational symmetric structure of the tube/vial, means that the tube/vial can be rotated and scanned over the whole inner surface, making it ideal for multi-spot measurements. A rotational symmetric configuration make it possible to place inserts within the vial/tube which are easily sealed.

If the SPR tube/vial is conical, a correspondingly conical insert will be self-positioned and self-tightened, by virtue of it fitting exactly in the cavity, making it an ideal structure for an integrated flow system, as will be disclosed below in detail. The system can also be made leakage proof.

It is also ideal when a bob (fixed or rotating) is used. The rotation of the bob relative the tube/vial makes it possible to monitor interactions, adlayer formations, and conformational changes at different shear rates. The SPR-vial can efficiently be used in the monitoring of polymerization and depolymerization processes [3], e.g. hemostasis.

Whereas reflectance measurements of p-polarized light is the dominating technique for SPR, the polarization state, i.e. ellipsometric measurements can be performed, using both p- and s-polarized light. Several ellipsometric techniques are well known for planar structures, such as null ellipsometry,

off null ellipsometry, rotating analyzer etc, which all can be used in the proposed structure.

For enhancement of the response signal, a sandwich assay can be used, where an immobilized molecule act as capturing molecule for an analyte molecule, and a third molecule is attached to the analyte either by mixing in the sample or by a further reagent step. The third molecule can be a large molecule, or have high refractive index or using both properties.

Further scope of applicability of the present invention will become apparent from the detailed description given hereinafter and the accompanying drawings which are given by way of illustration only, and thus not to be considered limiting on the present invention.

The invention is now illustrated by description of embodiments with reference to the drawings and experiments, but it should be understood that the invention is not limited to the specifically disclosed embodiments.

#### BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1a shows an apparatus for surface plasmon resonance measurements with a line shaped beam (tilted view) without an imaging lens.

FIG. 1b shows an apparatus for surface plasmon resonance measurements with a line shaped beam (top view) without an imaging lens.

FIG. 1c shows an apparatus for surface plasmon resonance measurements with a line shaped beam (tilted view) and an imaging lens.

FIG. 1d shows an apparatus for surface plasmon resonance measurements with a line shaped beam (top view) and an imaging lens.

FIG. 1e shows an apparatus for surface plasmon resonance measurements with a line shaped beam (side view) and an imaging lens.

FIG. 2 shows an apparatus for surface plasmon resonance measurements with a focused fan-shaped beam.

FIG. 3a shows an apparatus for surface plasmon resonance measurements with a gaussian, circular shaped beam, side view.

FIG. 3b shows an apparatus for surface plasmon resonance measurements with a gaussian, circular shaped beam, perspective view.

FIG. 4 shows the angles of the incident and the reflected beams.

FIG. 5 shows a sensor unit with adhesion layer and metal coating.

FIG. 6 shows a sensor unit with different molecular recognition layers.

FIG. 7 shows a sensor unit measuring a polymerization/depolymerization, with or without oscillation of the cup/vial.

FIG. 8 shows a sensor unit with a polymer sensing film for gas or liquid recognition.

FIG. 9a shows a sensor unit consisting of slices with different molecular recognition layers.

FIG. 9b shows a sensor unit consisting of slices with different molecular recognition layers where the analyzing medium is flowing through the device.

FIG. 9c shows a sensor unit consisting of slices with different molecular recognition layers fitted into a tube.

FIG. 10a shows an apparatus where sensor unit is rotating.

FIG. 10b shows an apparatus where a rotating bob is inserted inside the sensor unit.

FIG. 10c shows an apparatus where a rotating bob is inserted inside a rotating sensor unit.

FIG. 11 shows a sensor unit with a conical shape.

FIG. 12a shows a device with an interchangeable radial fluidic insert.

FIG. 12b shows a device with a radial fluidic insert.

FIG. 12c shows a device with a fluidic insert utilizing seals.

FIG. 13a shows an axial fluidic insert.

FIG. 13b shows a device with an axial fluidic insert.

FIG. 14 shows the relation between reflectance and the inner incident angle for three different wavelengths (633, 733 and 833 nm) for two different effective refractive indices (1.330 and 1.333).

FIG. 15 shows the reflected intensity at the detector for two different refractive indices of the sample ( $n=1.33$  and  $n=1.34$ ), due to the surface plasmon at the inside of the wall of a cylindrical device, when a monochromatic light source at 633 nm is used.

FIG. 16 shows the relation between the outer incident angle (air-glass interface) and the inner incident angle (SPR-angle).

FIG. 17 shows the movement of the spot position (in)mm/ $^{\circ}$  versus the incident angle (SPR-angle) for different inner diameter of the device.

FIG. 18 shows the movement of the spot position (in  $\mu\text{m}/\text{mRIU}$ ) versus the incident angle (SPR-angle) for different inner diameter of the device.

FIG. 19 shows the difference in directions of the surface plasmon propagation vector due to the propagation length and the curvature of the surface of the device

FIG. 20a-c show the device with a cap or two caps.

FIG. 21 shows the device placed in a holder.

FIG. 22a-b, illustrates the detector signal from experiments using the apparatus with different refractive indices of the sample.

FIG. 23a, illustrates exploded view of the sensor unit as an angular cut out, here as a half pipe with a flow insert.

FIG. 23b, illustrates an exploded view the sensor unit built from two or more elements, with a flow insert.

FIG. 24, illustrates a sensor unit where at least one sensor surface is double curved.

FIG. 25, illustrates a cross section of a rotational symmetric sensor unit with an integrating flow structure

FIG. 26a, illustrates an embodiment with thermo controlled sensor unit.

FIG. 26b, illustrates an embodiment where the thermal flux is directed to the sensor unit by tubes.

FIG. 26c, illustrates an embodiment where there are different temperatures on different places on the sensor unit, creating a temperature gradient.

FIG. 27, illustrates an embodiment where heat flux is transferred by a liquid close to the sensor surface.

FIG. 28, illustrates an embodiment for measurement of the polarization state.

## DETAILED DESCRIPTION OF PREFERRED EMBODIMENTS

### SPR-Technique

The surface plasmon is excited by an electromagnetic wave, in the visible region denoted light, which electrical field can be described as:

$$\vec{E} = \vec{E}_0 e^{i(\vec{k}\vec{r} - \omega t)} \quad (1)$$

where  $E$  is the varying electrical field in time and space,  $E_0$  a field strength constant,  $k$  the propagation constant,  $r$  the space vector,  $\omega$  the angular frequency, and  $t$  the time. The surface plasmon is excited by the electrical field component parallel to the plane of incidence, i.e. p-polarized light, also denoted transverse magnetic (TM).

The propagation constant is given by:

$$k = k_0 \cdot N = \frac{\omega}{c} N = \frac{2\pi}{\lambda} N \quad (2)$$

where  $k_0$ , is the free space propagation constant,  $N$  is an effective refractive index,  $c$  the speed of light in free space, and  $\lambda$  the wavelength in free space.

The propagation constant of a plasmon can be calculated from Maxwells Equations, and is for the semi-infinite case given by [26]:

$$k_{SP} = k_0 N_{SP} = k_0 \sqrt{\frac{\epsilon_m(\omega) \cdot \epsilon_a}{\epsilon_m(\omega) + \epsilon_a}} \quad (3)$$

where  $k_{SP}$  is the (complex) surface plasmon propagation constant,  $N_{SP}$  is the effective (complex) refractive index of the surface plasmon,  $\epsilon_m(\omega)$  is the frequency dependent dielectric function (complex) of the metal carrying the plasmon, and  $\epsilon_a$  is the effective dielectric function of the ambient medium (normally real, but can be frequency dependent and complex). The effective dielectrical function is monitored (weighted within the probe depth of the surface plasmon) by the surface plasmon. The function of the weighting is exponential decaying, and given by:

$$e^{-\frac{z}{\delta}} \quad (4)$$

where  $z$  is the space coordinate out from the surface, and  $\delta$  is the characteristic probe depth.

The characteristic probe depth is a function of the wavelength and the polarizability of the material, and is at resonance given by:

$$\delta = \frac{\lambda}{4\pi} \cdot \frac{1}{\text{Im} \left[ \frac{\epsilon_a}{\sqrt{\epsilon_m + \epsilon_a}} \right]} \quad (5)$$

The surface plasmon will propagate along the interface and its intensity will decay exponentially:

$$I \propto e^{-\frac{x}{L_{SP}}} \quad (6)$$

where  $x$  is the coordinate along the interface parallel to the propagation, and  $L_{SP}$ , is a characteristic propagation length, which is given by:

$$L_{SP} = \frac{1}{2\text{Im}[k_{SP}]} = \frac{\lambda}{4\pi} \cdot \frac{1}{\text{Im}[N_{SP}]} = \frac{\lambda}{4\pi} \cdot \frac{1}{\text{Im} \left[ \sqrt{\frac{\epsilon_m(\omega) \cdot \epsilon_a}{\epsilon_m(\omega) + \epsilon_a}} \right]} \quad (7)$$

There are in principal three different ways to measure changes in the SPR-propagation constant. First, by measuring the reflected intensity (reflectance) at a flank of the SPR-dip at a certain wavelength and incident angle. Second, by measuring the intensity of the reflected light versus the angle of the

incident light (angular interrogation). Third, by measuring the intensity of reflected light for different wavelengths at a certain incident angle (wavelength interrogation).

For zero-dimensional SPR (measurement of a single spot) the intensity measurement requires only a spot (one element) detector. A one-dimensional SPR measurement (measurement of a single line) the intensity measurement requires a one-dimensional (line) detector to make an instant measurement of the position of an SPR-dip. If a two dimensional (matrix) detector is used, a two-dimensional sensor surface can be monitored. The intensity measurement has the disadvantage that only a small part of the SPR-dip is measured. This can lead to false interpretations, if e.g. the dip is broadening or influenced by a drifting offset. If the whole dip is measured as in the angular and wavelength interrogation, these drawbacks are eliminated or at least significantly reduced.

If the angular or wavelength interrogation is used, one dimension of the sensor surface is eliminated and replaced by an SPR-dip, i.e. a two-dimensional detector can monitor a line, and a one line-detector can monitor a spot. In this case, one dimension is used for the length scale (real image) and one dimension is used for the dip (either angle or wavelength). However, if the SPR-tube/vial is rotated, a fully two-dimensional surface can be monitored by a two-dimensional detector, measuring the whole SPR-dip, and a line can be monitored by a one-dimensional detector, using this scanning technique.

The preferred detection method for the SPR-tube/vial is the angular interrogation. If a collimated (parallel) or slightly divergent or convergent light is used, the light beam will hit the outer wall of the tube/vial and refract into the inner surface at a continuum of incident angles due to the width of the beam. The total internal reflected beam will be further refracted at the outer surface, creating a divergent beam, where the SPR-dip (angular) are easily monitored by a photodetector or a photographic film. The relation between the incident angle,  $\alpha$ , at the outer surface and the incident angle,  $\theta$ , at the inner surface is given by:

$$\sin \theta = \frac{r_o \cdot n_o}{r_i \cdot n_1} \cdot \sin \alpha \quad (8)$$

where  $r_o$  and  $r_i$  are the radii of the outer and inner surfaces respectively, and  $n_o$  is the refractive index for the ambient medium outside the device (normally air) and  $n_1$  is the refractive index of the device. From equation 8 it follows that there is only a limited range of incident angles,  $\theta$ , at the inner surface, depending on the values of  $r_o$ ,  $r_i$ ,  $n_o$ , and  $n_1$ . For the case that the refractive index of the substrate is higher than the ratio between the outer radius and inner radius, it is not possible to obtain high incident angles at the inner surface (it is assumed that the ambient is air).

If collimated light is used, the width of the beam will lead to a finite measuring spot, where the different positions on the curved surface is associated with a specific incident angle. This means that the position for resonance will change when the resonance condition is changed. The change is dependant on the ratio between the inner and outer radius, but can for most cases be neglected. Typically, for a crown glass tube with 12 mm outer diameter and 8 mm inner radius, the movement of the measuring spot is 100  $\mu\text{m}$  for every degree the resonance condition is shifted. This corresponds approximately to 10  $\mu\text{m}$  for 1000  $\mu\text{RIU}$ , i.e. the movement is of the

same order as the propagation length of the plasmon. This means that for most purposes the movement of the measuring spot is insignificant.

The presence of a curved surface on the tube means that the surface plasmon will “change direction” along the surface, which may lead to a lower coupling between the plasmon and the incident light. The characteristic propagation length of the plasmon is approximately 2  $\mu\text{m}$  and 20  $\mu\text{m}$  for a wavelength of 633 and 900 nm, respectively, and for a 400  $\mu\text{m}$  diameter capillary tube the deviation of the plasmon at the characteristic propagation length is 0.6 and 6 degrees respectively. This means that for the longer wavelength the curvature will probably interfere with the surface plasmon coupling, but for the shorter wavelength the curvature should have little effect. For an 8 mm inner diameter vial the deviation is only 0.03 and 0.3 degrees respectively, and hence the effect from the curvature is negligible. Using other conducting materials with shorter surface plasmon propagation lengths, will lead to lower sensitivity due to broadening of the dip. A preferred inner radius is typically more than 1 mm, and is preferably 3 to 8 mm, and is typically less than 20 mm, preferably less than 10 mm.

Ellipsometric Technique

A useful measurement technique is to perform an ellipsometric readout, i.e. keep track of the changes in the polarisation state. In other words, the relative phase and amplitude between the reflection coefficients of the p-polarized and the s-polarized light is measured. The s-polarized light is the electrical field component perpendicular to the plane of incidence, also denoted transverse electrical (TE). The ellipsometric properties are often expressed as the complex (real and imaginary numbers) reflection coefficient:

$$\rho = \frac{R_p}{R_s} = \left| \frac{R_p}{R_s} \right| e^{i(\delta_p - \delta_s)} = \tan(\Psi) e^{i\Delta} \quad (9)$$

Where

$$\tan(\Psi) = \left| \frac{R_p}{R_s} \right| \quad (10)$$

and

$$\Delta = \delta_p - \delta_s \quad (11)$$

i.e. the polarization state can be expressed in the “ellipsometric angles”  $\Psi$  and  $\Delta$ . The polarization state can be measured by both compensating (null ellipsometry) and non-compensating (photometric) ellipsometric systems. The compensating ellipsometer can use a polarizer-compensator-sample-analyzer (PCSA) setup, where the light source is placed before a rotating polarizer followed by a fixed compensator (usually a  $\frac{1}{4}$  wave plate at  $45^\circ$ ). Between the compensator and the analyzer, is the sample placed, consisting of both SPR-supporting layer and sample under test. The analyzer, is a polarizer, which is also rotating. The photo detector is placed after the analyzer. The polarizer and analyzer are independently rotated until extinction of occurs. The compensator introduces a phase shift between the p- and s-polarized light. The ellipsometric angles can be found from:

$$\Psi = A$$

and

$$\Delta = 2P + 90^\circ$$

Where A and P are the azimuths for the analyzer and polarizer, respectively. The compensating ellipsometer is not limited to this setup.

Non-compensating ellipsometer can be achieved in several ways. One is with fixed azimuths close to extinction and measurement of light intensity at the detector. Another is to use a rotating analyzer RAE, where the polarizer is fixed and the compensator is optional. By measuring both the amplitude and phase (with respect to the azimuth of the analyzer) at the detector, the ellipsometric angles  $\Psi$  and  $\Delta$  can be determined.

The ellipsometric measurements can be performed with imaging optics.

The proposed configuration of the device means that it is possible to measure changes in optical parameters with an extremely high accuracy, without using any optical coupling medium, nor using complex optics, making the system very cost effective. It would also be possible to use a divergent beam as proposed by Chinowsky et al. [11], but that will require additional optics. For this case the measuring spot can be focused and its position will be independent of the effective refractive index of the sample medium.

By using a container or a tube, an integrated flow handling system can be placed inside the container, leading to a compact, precise and cost effective solution. The macroscopic shape of the container means that it has a large sensor area, ideal for multi-spot measurements. The possibility of easy rotation of the tube/vial means that the whole sensor area can be scanned extremely fast. A rotation of the tube/vial gives also the opportunity to use the centrifugal force to move samples. A typical flow cell height is preferably between 10 and 100  $\mu\text{m}$  and more preferably between 20 and 50  $\mu\text{m}$ .

The shape of the insert makes it suitable for temperature control of the sample. There are at least three reasons for temperature control: 1) Reaction kinetics is temperature dependent, 2) Resemble In Vivo conditions, e.g. 37 degrees C., 3) Baseline of SPR-signal is temperature dependent. If a reaction is following the Arrhenius Rule every 7 (or 10) degrees C. increase in temperature will double reaction rate. In case of water based buffer, SPR base line will change approximately -100 micro refraction units for every degree C. An increase in temperature will decrease the refraction unit due to thermal expansion. The configuration of the invention makes it possible to place thermal control close to the sensor surface and hence a good thermal conductivity can be obtained leading to precise thermal control. One embodiment uses a liquid, e.g. water, water/glycol, etc, to transfer heat/cold to the sample area. The sample heat unit in the insert is placed very close (typical within 1 mm) to the roof of the flow cell, making temperature control very fast and precise. Temperature controlling liquid is flowing at the outer part of the sample temperature unit leading to very low thermal resistance to the sample. The temperature controlling liquid is preferably heated or chilled by a thermoelectric element. But, other temperature regulating devices such as compressor-condenser, or just tap water, are alternatives. If only heating is required a resistive element (Joule heating) is preferred. Another embodiment with only heating is using resistive heating close to the sample area, without using circulating liquids, and only use thermal conduction to temperate the sample.

Another embodiment uses two or more temperature controlling liquids, when switched into the sample temperature unit, making fast temperature steps or precise temperature ramps with respect to time.

Yet another embodiment uses different temperatures at different places on the sample temperature unit, which will

create a well defined temperature gradient due to the thermal conductivity of the sample temperature unit.

Because of the rotational symmetry and macroscopic radius and a relatively short height, the tube is very easy to manufacture with high optical standards and high mechanical precision. The dimensions of the tube/vial make it easy to deposit the essential surface plasmon carrying metal film with a high precision at a low cost.

The present invention provides a device and apparatus for measurement of refractive indices close to the inner surface of a tube, cup or vial or a part thereof, based on surface plasmon resonance. The sensing surface may consist of one or a plurality of sensing areas, in axial or radial or diagonal direction or a combination of these directions. The specimen under investigation may be studied in a gaseous, liquid or solid environment. The device can be used for concentration measurement of gases or analytes in a liquid. The device can also be used for analyses of interaction between molecules, e.g. biomolecules. The device can be used for measurement of changes in conformation of a specimen, e.g. coagulation of blood. The device can be used for monitoring of adlayer formations.

## DEFINITIONS

For the purpose of this invention a "flow structure" is taken to mean any structure that cooperates with the inner surface of the sensor unit in such a way as to provide a reduced sample volume in the cavity of the unit.

Non-limiting examples of a such a flow structure are an insert forming an annular space in a cylindrical sensor unit, or an insert provided with grooves or recesses forming flow channels when the insert rests against the inner surface of the sensor unit.

The function of such inserts is to direct the sample, to limit the extension of the sample within the unit, or to adjust the height from sensor surface of e.g. a flow cell, provided by the insert, in order to control the diffusion of sample analyte to the sensor surface, or for controlling the flow rate across the sensor surface.

A "cavity" shall be taken to mean a space that is surrounded by something, although it need not, but can be completely surrounded.

A "compartment" shall be taken to mean the space wherein a sample is located, i.e. the space formed in cooperation between an insert and the inner surface of the sensor unit.

The invention will now be described, but not limited, with reference to the drawings.

FIG. 1a illustrates one embodiment of the apparatus of the invention wherein a collimated input beam **300** emanates from an illumination system **400** onto a sensor unit **100**, containing a specimen under test **200**, such as a liquid sample or a gas, directed by a structure **610**, and the reflected light **330** is measured by a detection system **500**. The sensor unit **100** has a transparent substrate **103**, typically a plastic material or glass, which is rotationally symmetric or partly rotationally symmetric. The inner surface of the substrate **103** is covered with a thin, typically 50 nm, conducting layer **110** of a free electron like material, typically gold or silver, which can conduct a surface plasmon. The incident light **300** is refracted **310** at the outer surface of the substrate **103**, onto the inner surface and the conducting layer **110**, where the light is either reflected **320** or absorbed in the conducting layer **110** by a surface plasmon, or a combination of both. The reflected light **320** is then refracted at the outer surface of the substrate **103**, forming a light-beam **330** which impinges on a detector system **500**. The optical detection system **500** incorporates a

photon sensitive device **510**, that may be a photographic film, but is preferably a photo-detector, e.g. a charge coupled device (CCD), charge injection device (CID), CMOS-device, photo diode etc. For the case of using the photo detector **510**, the optical signal is evaluated by an electronic device **800**, preferably a computer system incorporating an analog to digital converter. The illumination system **400** incorporates at least one light source **410**, but also a plurality is possible, that may be a laser, light emitting diode (LED), gas discharge lamp (e.g. Xenon), tungsten halogen lamp etc., or an array or matrix of the elements. The light source **410** may create a polarized light, e.g. some lasers, or a polarizing element **420** may be inserted anywhere within the light path. The polarization is preferably p-polarised (transverse magnetic mode) with regard to the inner surface of the sensor unit **100** for reflectance measurements. The polarizing element **420** can be a dichroic sheet, Glan-Taylor prism, Glan-Thompson prism, or an equivalent device. However, the light beam does not necessarily have to be polarized, but that will enhance signal to noise ratio. The light beam **300** is in this configuration expanded by an optical element **430**, e.g. lenses or diffractive optical elements (DOE's), or a combination of both, creating a line pattern, making it possible to simultaneously monitor a line (axial) at the sensor surface **110**.

FIG. **1b** illustrates the same configuration as FIG. **1a**, but with a view from above.

FIG. **1c** illustrates an apparatus with imaging optics **520**. The imaging optics **520** will image the sensor surface **110** axially. The imaging optics will collect the angular spectrum **335** from each point on the sensor surface and focus the rays onto the detector **510**. The imaging optics **520** will minimize the effect from diffraction effects due to uneven reflectance distribution along the sensor surface **110**, or by obstruction of the optical wave propagation. The imaging optics **520** preferably contains at least one cylindrical lens or on equivalent DOE.

FIG. **1d**, illustrates the same configuration as FIG. **1c**, but with a view from above.

FIG. **1e**, illustrates the same configuration as FIG. **1c**, but with a cross section of the sensor unit.

FIG. **2**, illustrates an apparatus utilizing a convergent beam (fan shaped beam), **300** and **310**, to focus on a specific spot at the sensor surface **110**. By using a convergent beam the position of the sensor spot can be small and independent of the resonance condition, i.e. all incident angles impinge on the same spot on the sensor surface **110**.

FIG. **3a**, illustrates an apparatus with a collimated or nearly collimated beam **300** from a light source **400**. The light source is preferably a laser or LED. The curvature of the substrate **103** will convert the collimated beam **300** to a convergent beam **310**, with respect to the surface normal of the sensor surface **110**. The collimated beam will, due to general physics laws, have a gaussian intensity profile, which is not a disadvantage, but rather an advantage making it possible to just illuminate the angles where the surface plasmon can be excited. The specimen under test **200** is directed by a structure **610**.

FIG. **3b**, illustrates the configuration of FIG. **3a**, but seen from a perspective view.

FIG. **4**, illustrates the optical path in the sensor unit **100**. The substrate **103** has a refractive index of  $n_1$  and an outer and inner diameter of  $r_o$  and  $r_i$  respectively. The ambient medium, typically air, has a refractive index of  $n_o$ . An incident beam **300** impinges on the substrate **103** at an incident angle,  $\alpha$ , relative the surface normal. The incident angle,  $\alpha$ , is controlled by the height,  $h$ , from a centerline,  $C_L$ , parallel to the beam **300**, by the formula:

$$\alpha = \arcsin\left(\frac{h}{r_o}\right) \quad (9)$$

The beam **300** is refracted into a refracted beam **310** with an angle of  $\beta$  relative the surface normal according to Snells Law:

$$n_o \sin \alpha = n_1 \sin \beta \quad (10)$$

The refracted beam **310** will impinge on the inner surface of the substrate at an incident angle  $\theta$ . The angle  $\theta$  is denoted the resonance angle  $\theta_{SP}$  at a match of the momenta of the incident light **310** and the surface plasmon. According to optical laws the exit angle  $\theta$  of the reflected beam **320** is equal to the incident angle  $\theta$  of the incident beam **310**. Due to symmetry, the incident and refracted angles,  $\beta$  and  $\theta$  respectively, at the substrate-ambient interface for the outgoing beam **320** and **330** will be the same as the refracted and incident angle,  $\beta$  and  $\theta$  respectively for the incident beam **300**.

The substrate **103** is coated with a conducting layer **110** with refractive index  $n_2$ . The specimen under test **200** is usually a gas or liquid with an effective refractive index of  $n_3$ , which is directed towards the conducting layer **110** by a flow controlling structure **610**.

FIG. **5** illustrates the sensor unit **100** with an adhesion layer **105** between the substrate **103** and the conducting layer **110**. The adhesion layer is thin, typically less than 2 nm and preferably 0.5 nm, which means that it is thick enough to assure good adhesion and thin enough, not disturbing the surface plasmon. The adhesion layer **105** is typically titanium or chromium.

Molecular Recognition Layer

FIG. **6** illustrates the sensor unit **100** with one or a plurality of molecular recognition area/areas **130a-c**. The molecular recognition areas **130 a-c** are typically specific to analytes **140a-c** respectively. The molecular recognition layer **130** is typically attached to the conductive layer **110** by the means of a linker layer **120**. The linker layer **120** is typically an alkanethiol with a carbon chain length greater than three and preferably 16, using the thiol groups to bond to the gold surface. The linker layer **120** may incorporate a hydrogel **210**, as shown in FIG. **7**. The linker layer **120** may incorporate avidin/streptavidin biotin molecules. When the analyte molecules **140** binds to the recognition layer **130**, the dipoles at the analyzing frequency within the probe depth of the surface will normally increase and hence increase the effective refractive index  $n_3$  of the specimen under test **200**. The increase of  $n_3$  will increase the propagation constant of the surface plasmon, i.e. the resonance condition will change. The specimen under test **200** is held between the substrate and the structure **610**.

FIG. **7**, illustrates a polymer layer **210**, fibrin etc. The polymer formation can be supervised in conjunction with a rheometer, i.e. the cup or vial is rotating or oscillating, e.g. free oscillating rheometer (FOR).

FIG. **8** illustrates a sensor unit **100** with a polymer layer **115** as the recognition layer. The polymer layer **115** is sensitive to the specimen under test **200**, which is either a gas or liquid. The polymer **115** will either be denser or less dense upon contact with the specimen **200**. A denser polymer will increase the effective refractive index  $n_3$ . The polymer film can be made of polyfluoroalkylsiloxane, and the gases can be halogenated hydrocarbons such as trichloroethylene, carbon tetrachloride, chloroform, methylene chloride, etc.



FIG. **9a** illustrates the sensor unit **100** incorporating slices **100a-j** containing different sensing layers **130a-j** attached preferably to a linker layer **120**, however the linker layer can be omitted.

FIG. **9b** illustrates the sensor unit **100** incorporating slices **100a-j** incorporating different polymer sensing layers **115a-j**.

FIG. **9c** illustrates the sensor unit **100** incorporating slices **100a-j** incorporating different polymer sensing layers **115a-j**, inserted into a tube.

Rotatable Sensor Unit

FIG. **10a** illustrates a sensor unit **100**, which rotates. The rotation of the sensor unit **100** means that the whole sensing surface **110** can be scanned. A rotating sensor unit **100** means that the surface concentration at the inside of the device is monitored fast for many sensing spots or e.g. gradients.

FIG. **10b** illustrates the same set up shown in FIG. **10a**, but with a rotatable bob **600** inserted into the cavity of the sensor unit. The rotating bob introduces shear rates in the specimen under test **200**. The bob will also reduce the sample volume of the specimen under test **200**. The bob **600** can also be fixed to reduce the sample volume **200**.

FIG. **10c** illustrates the same set up shown in FIG. **10b**, but with a rotating or fixed bob **600** and a rotating or fixed sensor unit **100**.

FIG. **11** illustrates a similar setup to FIG. **10b**, but with a conical sensor unit **100** and a conical bob **600**. The conical set up will introduce different shear rates along the axis, which will e.g. affect a polymerization process monitored by surface plasmon resonance. This is particularly interesting for studies of the hemostasis system. The conical sensor unit **100** will obtain a larger dynamic range of shear rates if conical shape of the substrate **103** and the bob are different, as illustrated in FIG. **11**.

Micro Fluidic Insert

FIG. **12a** illustrates a sensor unit **100** with a fluidic insert **610** to be inserted into the substrate **103**.

FIG. **12b** illustrates the sensor unit **100** with the fluidic insert **610**. The fluidic insert **610** has radial channels **615**, which can be used for both immobilized molecules for the recognition layer, or for analyte molecules. The substrate **103** may be conical to assure good sealing to a conical insert **610**, but the substrate **103** can also be cylindrical, utilizing a cylindrical insert **610**. The substrate could be made of an elastic material e.g. silicon rubber, or made of a rigid material with elastic sealings **118**, as illustrated in FIG. **12c**.

FIG. **12c** illustrates the fluidic insert **610** with elastic seals **618**, e.g. o-rings. The substrate **103** and the insert **610** can either have a conical or cylindrical shape.

FIG. **13a-b** illustrates the sensor unit **100** with a fluidic insert **620** incorporating axial flow channels **625**. The insert **620** may create a cavity **107** which can contain the waste **210** after performed analysis. The waste may be drained by a channel **627**. The flow channels **625** may be fed by individual tubes **626**, that may be filled, e.g. by an external tube, needle or syringe tip. The tube **626** may be sealed by a membrane or an ordinary elastic seal. Rotation of either the units **103** and **610** or external tubes may act as valves, changing flow cells.

SPR Curves

FIG. **14** illustrates calculation of the reflectance, i.e. the ratio between the incident light **310** at the sensing surface **110** and the reflected light **320** from the sensing surface **110** versus the incident angle at the inner surface of the substrate **103**, for three different wavelengths, 633 nm, 733 nm, and 833 nm respectively, and two different effective refractive index,  $n_3$ , ( $n_3$  is equal to 1.330 and 1.333 respectively) of the specimen under test **200**. The conducting layer **110** is in this case gold with a thickness of 50 nm. The substrate **103** is in

this case BK7 glass, with a refractive index  $n_1$  of approximately 1.5. The position of the SPR-curves is a function of the effective refractive index  $n_{\text{seen}}$  by the surface plasmon.

FIG. **15** illustrates simulations of the reflected intensity at a detector **510** shown in FIG. **3**, at a wavelength of 633 nm, where the substrate **103** is made of BK7 glass, and a 50 nm thick conduction layer **110** made of gold is applied onto the inside of the substrate **103**. The refractive indices  $n_{\text{of}}$  of the specimen under test **200** are 1.33 and 1.34 respectively. The fall off of the intensity at the border of the detector **510** is an effect of the gaussian shape of the input beam **300**. The irregular shape of the curves emanates from photon shot noise.

Incident Angles at the Outer and Inner Surfaces

The incident angle,  $\theta$ , at the inner surface of the substrate **103** depends on the incident angle,  $\alpha$ , at the outer surface, the refractive indices of the ambient environment and the substrate,  $n_0$  and  $n_1$  respectively, and the ratio between the radii of the inner and outer surfaces,  $r_i$  and  $r_o$  respectively, according to equation 8.

FIG. **16** illustrates the incident angle,  $\alpha$ , at the outer surface of the substrate **103** as a function of the incident angle,  $\theta$ , at the inner surface, at different inner and outer radii ratios, 1.2, 1.33, 1.5, 1.71, and 2, for a substrate **103** made of Schott Duran glass in air. If the ratios are greater than the ratio between the refractive indices of the substrate **103** and the ambient, it is not possible to obtain all incident angles,  $\theta$ , at the inner surface.

For a collimated input beam **300** with a physical width, there are a diversity of incident angles,  $\theta$ , along the inner surface of the substrate **103**, which means that the resonance angle  $\theta_{SP}$  will occur at different places along the inner surface. The movement of the measuring spot as a function of the incident angle (e.g. resonance angle) at the inner surface of the substrate **103** is given in FIG. **17** for different values of the inner diameter,  $r_i$ , (6, 7, 8, 9, and 10 mm) at an outer diameter of 12 mm of the substrate made of Schott Duran glass. Using the approximation of a change of  $10^{-6}$  refractive index units corresponds to  $10^{-4}$  degrees in resonance shift of  $\theta_{SP}$ , leads to a sensing spot movement as illustrated in FIG. **18**. The conditions are the same as for FIG. **17**.

Propagation Vector Mis-Match

FIG. **19** illustrates the mismatch between the surface plasmon wave vector,  $k_{SP}$ , and the parallel component of the exciting light at the start  $k_{SP}(0)$  and at the propagation length  $k_{SP}(L_{SP})$  of the surface plasmon, due to the curvature of the substrate **103**. The angular mismatch (in radians) at the characteristic propagation length is given by  $\xi$ . Where  $\xi$  is given by the formula:

$$\xi = \frac{L_{SP}}{r_i} \quad (11)$$

Closed Cavity

FIG. **20a-c** illustrates how the sensor unit **100** can be closed in one or both ends by means of a cap **190**.

FIG. **21** illustrates a sensor unit **110** placed in a holder **900** ensuring exact positions.

## EXAMPLE 1

### Experiments

FIG. **22a**, illustrates the signal from the detector **500**, seen on the computer **800** from one embodiment of the surface

plasmon resonance apparatus of the invention as illustrated in FIG. 3a-b. The embodiment utilizes a monochromatic light source comprising a 5 mW HeNe laser (Melles-Griot) 400, emitting coherent and p-polarized (with respect to the sensor surface) light 300 at a wavelength of 633 nm. The light 300 impinges on the outer surface of the substrate 103 made of Duran (Schott) glass with a refractive index of 1.5, and with outer and inner diameters of 15 mm and 10 mm respectively. The glass substrate is sputtered with approximately 0.5 nm chromium acting as the adhesion layer 105 for the surface plasmon supporting metal layer 110. The surface plasmon supporting metal layer 110 is sputtered to a thickness of 50 nm. Both metal layers 105 and 110 are deposited using physical vapor deposition techniques (PVD). Sample 200 consists of sucrose solutions ranging from zero to 5% w/w sucrose in deionized water (Millipore) in steps of 1%. The sucrose solutions 200 get the following refractive indices: 1.3330, 1.3337, 1.3359, 1.3379, 1.3388, and 1.3403. The reflected light pattern 330 from the sensor surface impinges on the CCD camera 500 (Orbis 2, Spectra Source Inc.). The computer 800 is an ordinary PC.

FIG. 22b, illustrates the signal from the detector 500, with noise reduction by the computer 800 for two refractive indices (1.33 and 1.34) of the sample 200.

FIG. 23a, illustrates an exploded view of the sensor unit 100 as an "angular cut out" of the substrate 103, here as a half pipe. At least one, but not limited to one, insert 610 is placed in the cavity, either in direct contact with the surface using flow channels in the insert 610, or in the vicinity of the surface with or without a structure to control the flow of gas or fluid used. The use of an "angular cut" substrate means that formation of the adhesion layer 105 as shown in FIG. 5 and conductive layer 110 is easily formed by traditional physical vapor deposition (PVD) techniques, such as evaporation, electronic beam, and sputtering. Moreover, angular break up of the structure means that there will be an open large cavity which will help formation of the molecular recognition layers 130 as shown in FIG. 6, with a plurality of techniques, as soft lithography stamps etc.

FIG. 23b, illustrates an exploded view of the sensor unit 100 as assembled of two, but not limited to that number, substrates 103 and 104 forming a large cavity, in which a flow controlling part 610 is inserted, either in contact with the inner surface or in the vicinity. The flow controlling part has preferably integrated flow channels when in contact with the curved surface. The setup is not limited to that for the case when the insert 610 is in contact with the surface. For the non contacting case the insert 610 will preferably rotate or oscillate in the large cavity formed by the substrates 103 and 104, or vice versa when the substrates 103 and 104 together are moving, or both when both insert 610 and both substrates 103 and 104 is moving.

FIG. 24, illustrates a sensor unit 100 with a double curved surface 107, e.g. the bottom of a vial, with a surface plasmon supporting conductive layer 110, where the surface plasmon can be excited in many directions.

FIG. 25, illustrates a sensor unit 100 with an integrated flow structure 611 into the optical substrate 103. The cavity that is formed between the substrate 103 and the flow structure 611 is filled with a sample 200. The surface plasmon supporting conductive layer 110 is present at the concave surface of the wall.

FIG. 26a, illustrates a sensor unit 100 with a flow structure 610 inserted in the optical substrate 103. The cavity that is formed between the substrate 103 and the flow structure 610 is filled with a sample 200. The molecular recognition sites 130 are placed on the surface plasmon supporting conductive

layer 110 which is present at the concave surface of the wall of the substrate 103. The flow structure incorporates a temperature control unit 700 the purpose of which is to keep sample at sensor surface at a controlled temperature. Temperature can either be held constant or ramped. The thermo control unit consists basically of two parts, one sample temperature unit 710 which performs the local heat exchange at the sensor surface, and a heat/cold supplying unit 720, which supplies the sample temperature unit with heat or cold. The sample temperature unit 710 is in a simple embodiment, typically a piece of metal, and in a more advanced embodiment an insert made of e.g. polymers, ceramics, metal or a combination of these materials with channels for liquids carrying the heat flux. The heat/cold supplying unit is typical a thermoelectrical device (e.g. Peltier unit), but can also be a conventional compressor with heater/condenser or for the case only heating is necessary, a resistive element. Using a resistive element, the unit can be integrated with the sample temperature unit 710, forming a single unit. To obtain fast temperature ramps, different temperatures can be provided simultaneously at the heat/cold supplying unit 720, and these temperature stores can be switched into the sample control unit 710.

FIG. 26b, illustrates a temperature control unit 700, where the sample temperature unit 710 and heat/cold supply unit 720 is separated by tubes, inlet 733 and outlet 736 carrying the liquid for thermal control.

FIG. 26c, illustrates a temperature control unit 700, where the sample temperature unit 710 has two (or more) separated temperature zones 723 and 726, creating a temperature gradient between them.

FIG. 27a, shows the sample temperature unit 710 inserted into the substrate 103, with the temperature controlling liquid 732 close to the sample 200, therefore obtaining fast and precise temperature control of the sample.

Typical fixed temperatures are 25 and 37 degrees centigrade.

FIG. 28, shows an ellipsometric setup, i.e. measurement of the polarisation state. The light source 420, which can be monochromatic or multispectral, provides both p- and s-polarized light. This basic setup can be used for compensated as well as uncompensated ellipsometry. The uncompensated ellipsometer is shown as a PCSA-setup. For this case the compensator 460, which is a quarter wave plate, is set to 45° to introduce a phase shift between the p- and s-polarized light. Both the polarizer 450 and analyzer 550 are rotated until the light at the detector 510 vanishes. The setup can also be used off-null, where both the analyzer 550 and polariser 450 are fixed at azimuths close to zero light at the detector 510. A rotating analyser ellipsometer, RAE, is obtained for a fixed polarizer 450, usually at 45°, and a rotating analyzer 550. For this case the compensator 460 is optional. The light source 410 and detector 510 can be used in imaging.

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The invention claimed is:

1. Surface Plasmon Apparatus, comprising:

a light source;

a sensor unit for Surface Plasmon Resonance (SPR) comprising:

a transparent sensor structure forming at least one wall of a cavity, the wall being defined by a concave inner surface and a convex outer surface; wherein the inner surface is provided with a layer of a conductive material capable of supporting a surface plasmon;

a flow structure in said cavity so as to form at least one compartment for sample between the flow structure and the inner wall of the cavity;

a detector for detecting reflected light from the sensor unit; and

a processing unit.

2. The apparatus according to claim 1, wherein said light source provides collimated light.

3. The apparatus according to claim 1, wherein said sensor unit is rotatable relative to said light source and said detector to enable positioning of different sensor spots on said sensor unit.

4. The apparatus according to claim 1, wherein said light source and said detector are rotatable relative to said sensor unit to enable positioning of different sensor spots on said sensor unit.

5. The apparatus according to claim 1, wherein said light source is movable along a longitudinal axis of the sensor unit.

6. The apparatus according to claim 1, wherein said light source is positionable at a height, h, where it is capable of emitting a beam parallel relative a centerline to a radial direction of said sensor unit, to incorporate a critical angle, of an incident angle  $\theta$ , said incident angle given by

$$\sin \theta = \{(r_o \cdot n_o) / (r_i \cdot n_i)\} \sin \alpha,$$

$$\text{and } \alpha = \arcsin(h/r_o),$$

where  $r_o$  and  $r_i$  are radii of said outer and said inner surface, respectively, and  $n_o$  is a refractive index outside said sensor unit, and  $n_i$  is a refractive index of said transparent sensor structure.

7. The apparatus according to claim 1, further comprising means for measurement of polarization state of a reflected radiation.

8. The apparatus according to claim 1, further comprising means for generating light with a line shape along a longitudinal axis of said sensor unit.

9. The apparatus according to claim 1, wherein the sensor unit is rotatable, whereby said compartments are selectable.

10. The apparatus according to claim 1, wherein the light source and the detector are rotatable whereby said compartments are selectable.

11. The apparatus according to claim 1, further comprising means for measuring a plurality of angles of said reflected light simultaneously.