

US008222016B2

(12) **United States Patent**
Okumura et al.

(10) **Patent No.:** **US 8,222,016 B2**
(45) **Date of Patent:** **Jul. 17, 2012**

(54) **RECOMBINANT C-TERMINAL
 α -AMIDATING ENZYME DERIVATIVE**

(75) Inventors: **Takeshi Okumura**, Sano (JP); **Kazuaki Furukawa**, Tatebayashi (JP); **Masayuki Yabuta**, Tatebayashi (JP)

(73) Assignee: **Daiichi Sankyo Company, Limited**,
Chuo-ku, Tokyo (JP)

(*) Notice: Subject to any disclaimer, the term of this
patent is extended or adjusted under 35
U.S.C. 154(b) by 0 days.

(21) Appl. No.: **12/666,032**

(22) PCT Filed: **Jun. 27, 2008**

(86) PCT No.: **PCT/JP2008/062123**

§ 371 (c)(1),
(2), (4) Date: **Dec. 22, 2009**

(87) PCT Pub. No.: **WO2009/005140**

PCT Pub. Date: **Jan. 8, 2009**

(65) **Prior Publication Data**

US 2010/0261249 A1 Oct. 14, 2010

(30) **Foreign Application Priority Data**

Jun. 29, 2007 (JP) 2007-173461

(51) **Int. Cl.**
C12N 9/02 (2006.01)

(52) **U.S. Cl.** **435/189**; 435/7.1; 530/350

(58) **Field of Classification Search** None
See application file for complete search history.

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Primary Examiner — Hope Robinson

(74) *Attorney, Agent, or Firm* — Drinker Biddle & Reath
LLP

(57) **ABSTRACT**

Disclosed are: a recombinant C-terminal α -amidated enzyme
derivative which lacks the formation of at least one disulfide
bond among five disulfide bonds occurring in a C-terminal
 α -amidated enzyme derived from *Xenopus laevis*; DNA
encoding the derivative; an expression vector carrying the
DNA; a bacterium *Escherichia coli* transformed with the
expression vector; and a method for producing the derivative
by using the bacterium *Escherichia coli*.

3 Claims, 7 Drawing Sheets

Fig. 1

UPPER LINE: *Xenopus* Leavis AE- I

LOWER LINE: Rat AE (PHM domein)

-41 9

MASLSSSFLLVLL- - - FQNSCYCFRSPLSVFKRYEESTRSLNSDCLGT
MAGRARSGLLLLLLGLLALQSSCLAFRSPLSVFKRFKETTRSFSNECLGT

59

TRPVMSPGSSDYTLDIRMPGVTPTESDTYLCKSYRLPVDDEAYVVDFRPH
IGPVTPLDASDFALDIRMPGVTPKESDTYFCMSMRLPVDEEAFVIDFKPR

109

ANMDTAHHMLLFGCNIPSSSTDDYWDCSAGTCMDKSSIMYAWAKNAPPTKL
ASMDTVHHMLLFGCNMPSSSTGSYWFCDEGTCTDKANILYAWARNAPPTRL

159

PEGVGFRVGGKSGSRYFVLQVHYGNVKAQDKHKDCTGVTVRVTPEKQPQ
PKGVGFRVGGETGSKYFVLQVHYGDISAFRDNHKDCSGVSVHLTRVPQPL

209

IAGIYLSMSVDTVIPPGEAEVNSDIACLYNRPTIHPFAYRVHTHQLGQVV
IAGMYLMMSVDTVIPPGEKVVNADISQYKMYPMHVFAYRVHTHHLGKV

259

SGFRVRHGKWSLIGRQSPQLPQAFYPVEHPVEISPGDIIATRCLFTGKGR
SGYRVRNGQWTLIGRQNPQLPQAFYPVEHPVDVTFGDILAAARCVFTGEGR

309

TSATYIGGTSNDEMKNLYIMYYMDAAHATSYMTCVQTGEPKLFQNIPEIA
TEATHIGGTSSDEMKNLYIMYYMEAKYALSFMTCTKNVAPDMFRTIPAEA

359

NVPIPVSPDMMMMMGHGHHTTEAE- PEKNTGLQQPKREEEVLDQGLITL
NIPVPKPDMMVMHGH- - - HKEAENKEKSALMQQPKQGEEVLEQGDFYS

GDSAV

LLS- -

Fig.2

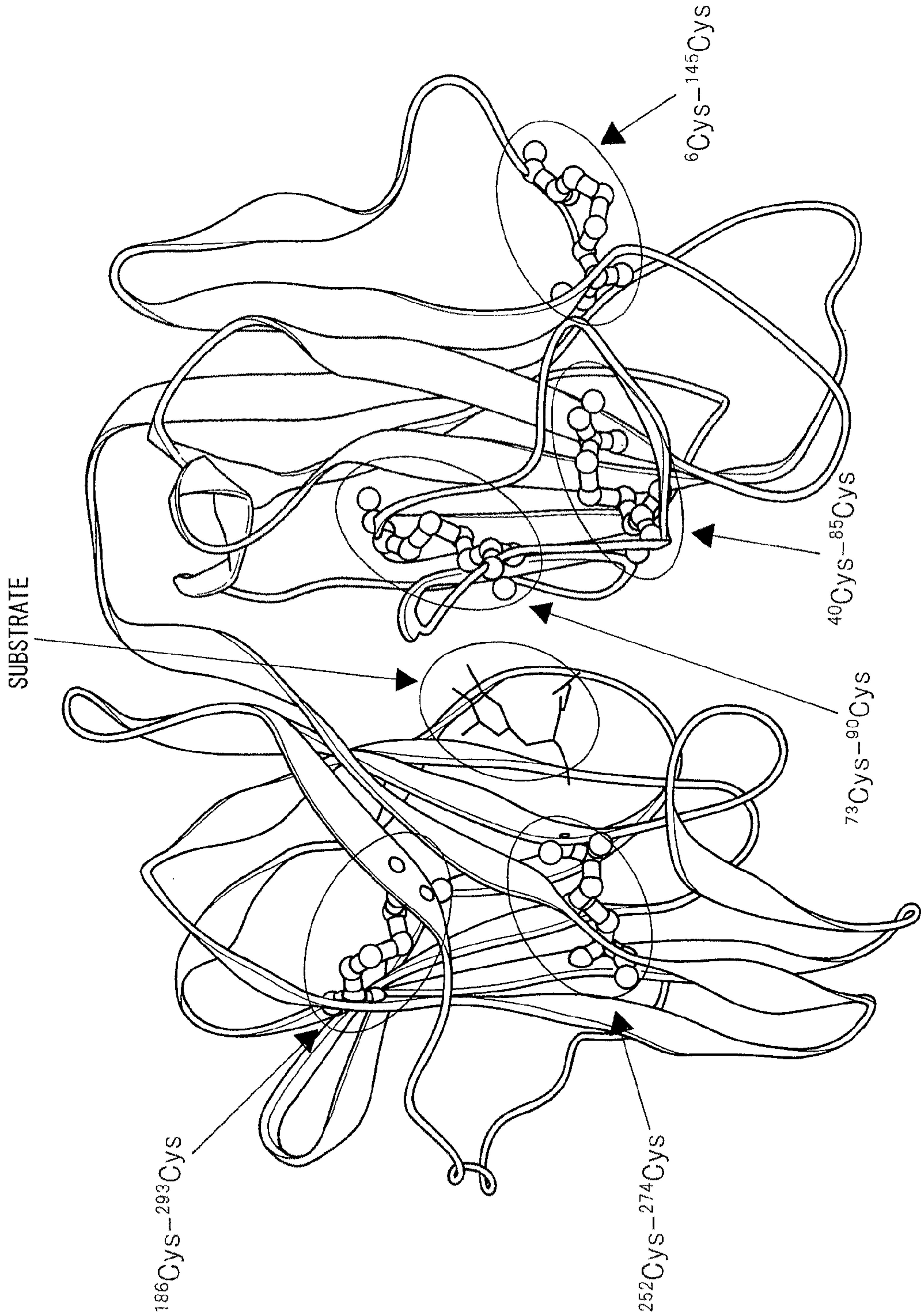


Fig.3

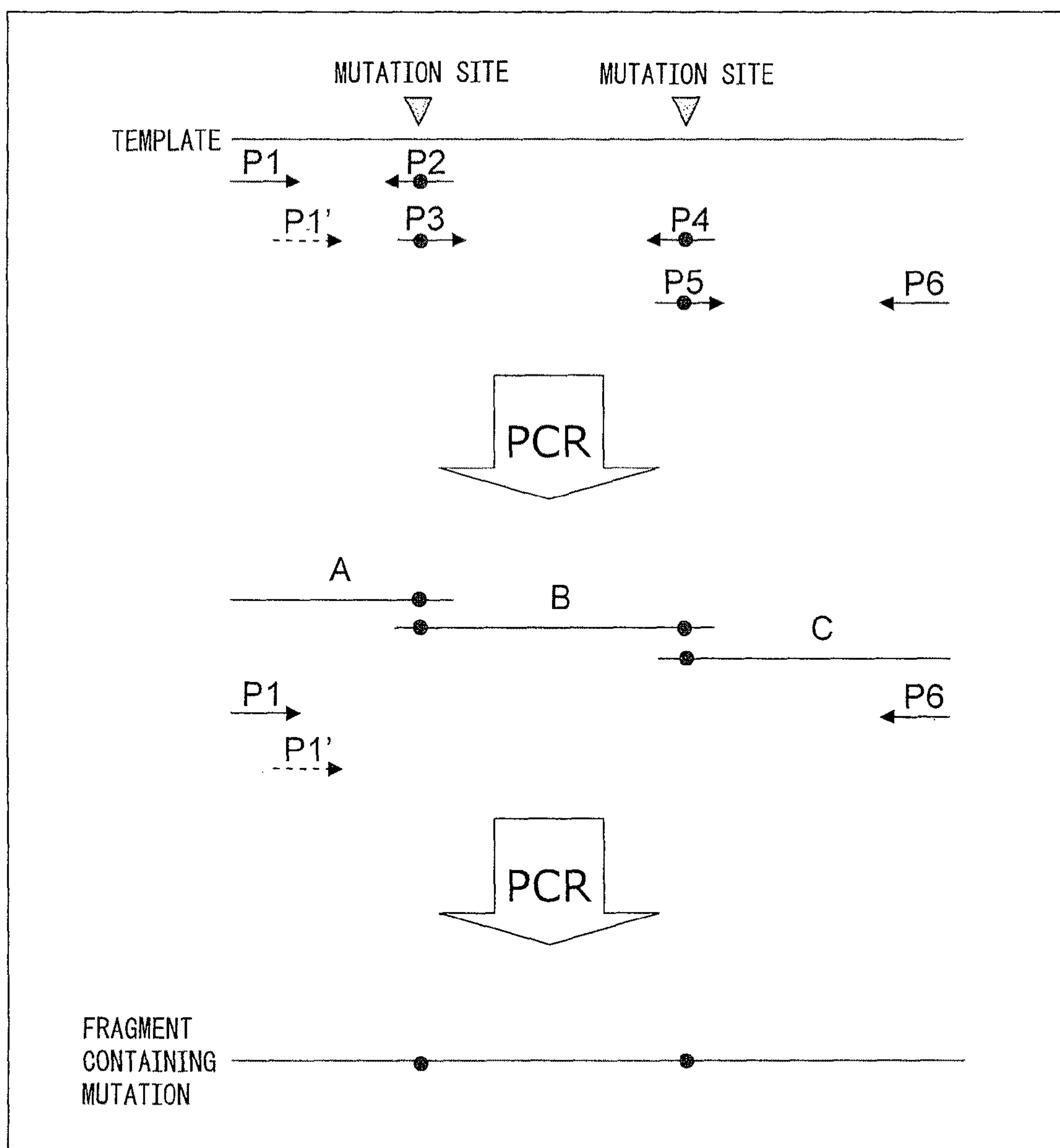


Fig.4

AE-I [8-321] (C145A) (NO P2,P3)

P1' AEp003 SEQ ID NO 3: 5' -ATTGGATCCGGGAACACGCGGCC
P4 AEp004 SEQ ID NO 4: 5' -TCACCCCGTGGCATCTTTATGT
P5 AEp005 SEQ ID NO 5: 5' -ACATAAAGATGCCACGGGGGTGA
P6 AEp006 SEQ ID NO 6: 5' -TGCCTCGAGTTACATCATCATGTCAGGGCT

AE-I [1-321] (C40A/C85A)

P1 AEp007 SEQ ID NO 7: 5' -ATTGGATCCGTCACCTTTCCAATGACTGCTT
P2 AEp008 SEQ ID NO 8: 5' -GGTAAGACTTGGCCAAATATGTG
P3 AEp009 SEQ ID NO 9: 5' -CACATATTTGGCCAAGTCTTACC
P4 AEp010 SEQ ID NO 10: 5' -TTCCCGCACTAGCGTCCCAGTAA
P5 AEp011 SEQ ID NO 11: 5' -TTACTGGGACGCTAGTGCGGGAA
P6 AEp006 SEQ ID NO 6: 5' -TGCCTCGAGTTACATCATCATGTCAGGGCT

AE-I [1-321] (C73A/C90A)

P1 AEp007 SEQ ID NO 7: 5' -ATTGGATCCGTCACCTTTCCAATGACTGCTT
P2 AEp012 SEQ ID NO 12: 5' -AAGGTATATTGGCTCCAAATAGA
P3 AEp013 SEQ ID NO 13: 5' -TCTATTTGGAGCCAATATACCTT
P4 AEp014 SEQ ID NO 14: 5' -ATTTGTCCATGGCAGTTCCCGCA
P5 AEp015 SEQ ID NO 15: 5' -TGCGGGAAGTGGCATGGACAAAT
P6 AEp006 SEQ ID NO 6: 5' -TGCCTCGAGTTACATCATCATGTCAGGGCT

AE-I [1-321] (C186A/C293A)

P1 AEp007 SEQ ID NO 7: 5' -ATTGGATCCGTCACCTTTCCAATGACTGCTT
P2 AEp016 SEQ ID NO 16: 5' -TGTTGTAGAGGGCGGCGATATCA
P3 AEp017 SEQ ID NO 17: 5' -TGATATCGCCGCCCTCTACAACA
P4 AEp018 SEQ ID NO 18: 5' -CCGTCTGTACAGCGGTCATGTAT
P5 AEp019 SEQ ID NO 19: 5' -ATACATGACCGCTGTACAGACGG
P6 AEp006 SEQ ID NO 6: 5' -TGCCTCGAGTTACATCATCATGTCAGGGCT

AE-I [1-321] (C252A/C274A)

P1 AEp007 SEQ ID NO 7: 5' -ATTGGATCCGTCACCTTTCCAATGACTGCTT
P2 AEp020 SEQ ID NO 20: 5' -CAGTGAACAGAGCCCTGGTTGCT
P3 AEp021 SEQ ID NO 21: 5' -AGCAACCAGGGCTCTGTTCCTG
P4 AEp022 SEQ ID NO 22: 5' -TGTATAAATTAGCCATTTTCATCG
P5 AEp023 SEQ ID NO 23: 5' -CGATGAAATGGCTAATTTATACA
P6 AEp006 SEQ ID NO 6: 5' -TGCCTCGAGTTACATCATCATGTCAGGGCT

Fig.5

AE- I [8-321] (C145A,C40A/C85A)

P1' AEp003 SEQ ID NO 3:	5' -ATTGGATCC	LGGGAACACGCGGCC
P2 AEp008 SEQ ID NO 8:	5' -GGTAAGACTT	GGCCAAATATGTG
P3 AEp009 SEQ ID NO 9:	5' -CACATATTT	GGCCAAGTCTTACC
P4 AEp010 SEQ ID NO 10:	5' -TTCCCGCACT	AGCGTCCCAGTAA
P5 AEp011 SEQ ID NO 11:	5' -TTACTGGGAC	GCTAGTGCGGGAA
P6 AEp006 SEQ ID NO 6:	5' -TGCCTCGAG	TTACATCATCATGTCAGGGCT

AE- I [1-321] (C40A/C85A,C252A/C274A)

P1 AEp007 SEQ ID NO 7:	5' -ATTGGATCC	ATCACITCCAATGACTGOTT
P2 AEp020 SEQ ID NO 20:	5' -CAGTGAACAG	AGCCCTGGTTGCT
P3 AEp021 SEQ ID NO 21:	5' -AGCAACCAGG	GCTCTGTTCCTG
P4 AEp022 SEQ ID NO 22:	5' -TGTATAAATT	AGCCATTTTCATCG
P5 AEp023 SEQ ID NO 23:	5' -CGATGAAAT	GGCTAATTTATACA
P6 AEp006 SEQ ID NO 6:	5' -TGCCTCGAG	TTACATCATCATGTCAGGGCT

Fig.6

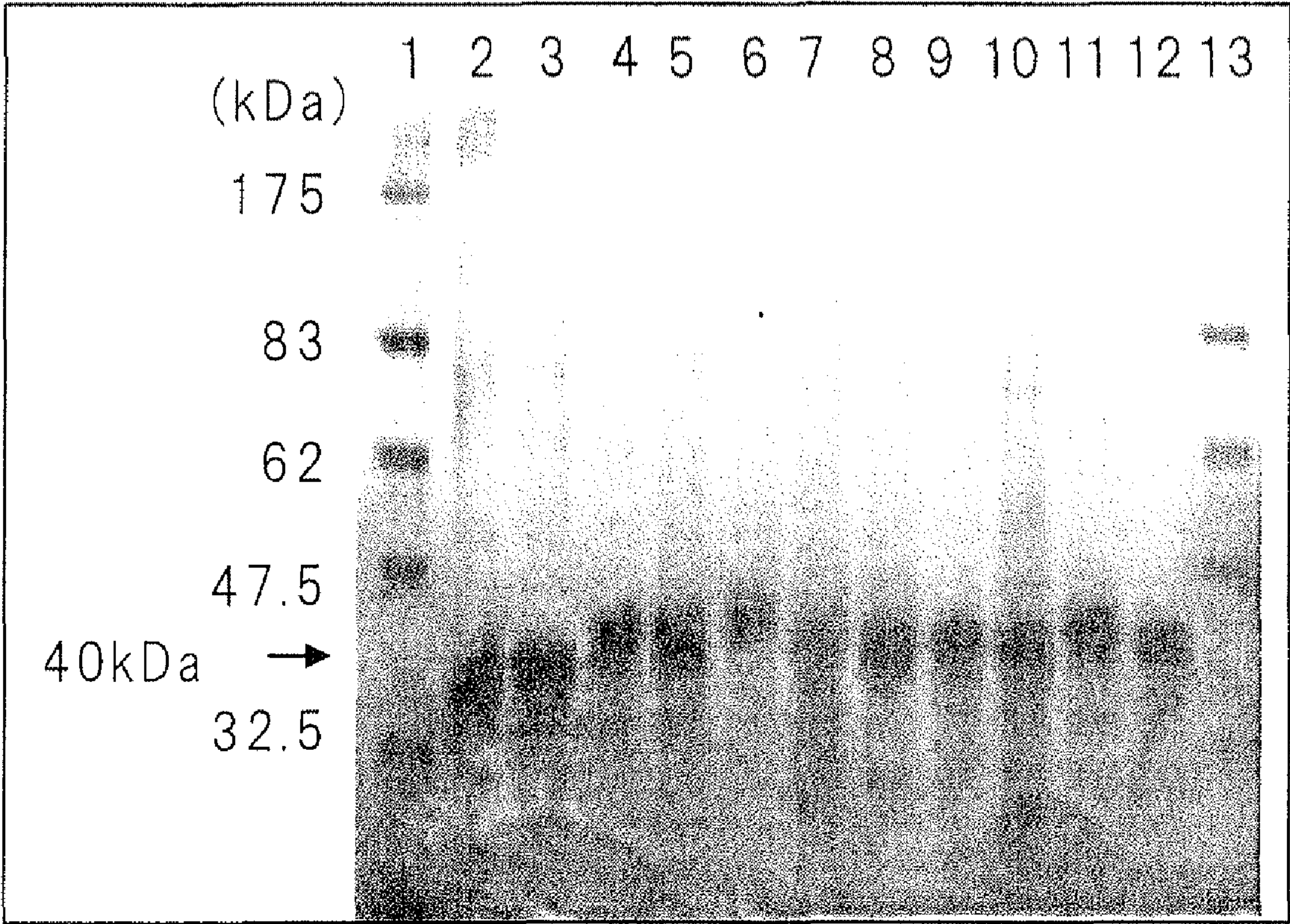


Fig. 7

PROTEIN CONCENTRATION
AFTER DIALYSIS (mg/L)

AMIDATING ENZYME AND ITS DERIVATIVES		ENZYME ACTIVITY (U/mL)	ACTIVITY/PROTEIN CONCENTRATION (U/mg)	ACTIVITY/1mL OF MEDIUM (U/mL)	COMPARED WITH AE-1 [1-321]
AE-1 [1-321]	200	41.1	205.4	26.7	100.0%
AE-1 [8-321] (C145A)	281	236.1	840.3	109.2	409.1%
AE-1 [1-321] (C40A/C85A)	249	447.7	1797.9	233.7	875.3%
AE-1 [1-321] (C73A/C90A)	287	16.1	56.1	7.3	27.3%
AE-1 [1-321] (C186A/C293A)	213	-	-	-	-
AE-1 [1-321] (C252A/C274A)	452	122.4	270.7	35.2	131.8%
AE-1 [8-321] (C145A,C40A/C85A)	273	212.4	778.1	101.1	378.8%
AE-1 [1-321] (C40A/C85A,C252A/C274A)	245	553.6	2259.6	293.7	1100.1%

RECOMBINANT C-TERMINAL α-AMIDATING ENZYME DERIVATIVE

CROSS REFERENCE TO RELATED APPLICATIONS

This application is the National Stage of International Application No. PCT/JP2008/062123 filed Jun. 27, 2008, and claims benefit of Japanese Patent Application No. 2007-173461 filed Jun. 29, 2007, which are herein incorporated by reference in their entirety.

REFERENCE TO A SEQUENCE LISTING

A Sequence Listing containing SEQ ID NOS: 1-37 is incorporated herein by reference.

FIELD OF THE INVENTION

The present invention relates to a recombinant C-terminal α-amidating enzyme derivative in which at least one of the five disulfide bonds to be normally owned by C-terminal α-amidating enzyme derived from *Xenopus laevis* has not been formed, DNA encoding said derivative, an expression vector containing said DNA, an *Escherichia coli* (*E. coli*) transformed with said expression vector, and a method of producing said derivative using said *E. coli*.

Specifically the present invention relates to a recombinant C-terminal α-amidating enzyme derivative of which enzyme activity has been enhanced by inhibiting the formation of at least one specific disulfide bond of the five disulfide bonds capable of being formed during refolding in the production of C-terminal α-amidating enzyme using a gene recombinant technology.

BACKGROUND ART

C-terminal α-amidating enzyme (peptidyl-glycine α-amidating monooxygenase I, EC 1.14.17.3) is present in eukaryotic organisms and forms the C-terminal amide structure of some biologically active peptides (peptide hormones, neuropeptides, peptide toxins etc.) or proteins. The C-terminal amide structure is known to be indispensable for the expression of biological activities of these peptides or proteins. In the case of human calcitonin, for example, it is known when the native C-terminal proline amide residue is converted to a proline residue, the biological activity decreases to as low as 1/1600 of the original activity.

Also, the *Xenopus laevis* C-terminal α-amidating enzyme per se has been disclosed in Japanese Patent No. 2598050 (registered on Jan. 9, 1997) and its coding gene has been disclosed in Japanese Patent No. 2581527 (registered on Nov. 21, 1996), respectively.

From the structural analysis of precursors of peptides and proteins having the C-terminal amide structure, it was found that in substrates for C-terminal α-amidating enzymes there is always glycine (Gly) present at the C-terminal end of the residue to be amidated (conversion of a —COOH group to a —CONH₂ group), which is represented by a general formula R-X-Gly wherein X represents any amino acid residue to be α-amidated at the C-terminus, Gly represents a glycine residue, and R represents the rest of said peptide or protein. On this Gly, a two-stage reaction of oxidation via a copper ion (first stage: hydroxylation of the α-carbon of Gly) and dealkylation (second stage: release of glyoxylic acid) takes place so that the C-terminus of the substrate is amidated. It is reported that in order to obtain the maximum enzyme activity

of this amidating enzyme, ascorbic acid in addition to molecular oxygen and copper ion (Cu²⁺) are required (see Betty A. Eipper, Richard E. Mains, and Christopher C. Glembofski, "Identification in Pituitary Tissue of a Peptide-amidation Activity That Acts on Glycine-Extended Peptides and Requires Molecular Oxygen, Copper and Ascorbic Acid" Proc. Natl. Acad. Sci. U.S.A. 80, 5144-5148, 1983).

Generally since such modifications including amidation, phosphorylation and acylation take place after translation from mRNA, they are called post-translational modifications, phenomena that are only observed in eucaryotic cells. Prokaryotic cells such as *E. coli* that is widely used in the production of recombinant proteins and peptides are incapable of such a post-translational modification. Considering the biosynthetic mechanisms of amidated peptides by eucaryotic cells that have been elucidated to date, amidated peptides can be produced in large quantities by gene recombinant technology using prokaryotic cells such as *E. coli*.

An amidated peptide can be produced in large quantities and at low cost by a method in which an amidated peptide precursor represented by a general formula R-X-Gly is expressed in large quantities as a recombinant in prokaryotic cells such as *E. coli*, a C-terminal α-amidating enzyme derived from eucaryotic cells is secured in large quantities, and said amidated peptide precursor is treated with said C-terminal α-amidating enzyme in vitro in an optimal reaction condition for producing an amidated peptide to produce the amidated peptide. In fact, efforts to produce amidated peptides by such a method has been made up to now, as described below.

Unigene Laboratories, Inc., Fairfield, N.J. 07004, "Production of recombinant salmon calcitonin by in vitro amidation of an *Escherichia coli* produced precursor peptide." Biotechnology (NY), 1993 January; 11(1):64-70 reports a method in which a salmon calcitonin (sCT) recombinantly produced using *Escherichia coli* was fused to part of glutathione S-transferase and expressed, sulfonated, and cleaved with cyanogen bromide, and using a C-terminal α-amidating enzyme expressed separately in CHO cells, the C-terminus of sCT was amidated in vitro.

Kokai (Japanese Unexamined Patent Publication) No. 7-163340 also describes a method of producing a human-derived calcitonin (hCT) using an amidating enzyme that was similarly expressed in CHO cells.

In these methods, the C-terminal α-amidating enzymes used in amidating the C-terminus of a protein of interest were produced by the CHO cell which is an animal cell.

Generally, however, the production of a recombinant protein using an animal cell takes a long culturing time and thus poses problems such as low productivity per unit time. As a method for resolving this problem, a method of using *E. coli* that enables production in a shorter culturing time has been developed as exemplified in Kokai (Japanese Unexamined Patent Publication) No. 7-250691.

This method permits the expression of a *Xenopus laevis* C-terminal α-amidating enzyme (peptidyl-glycine α-amidating monooxygenase I, EC 1.14.17.3) in large quantities by a recombinant technology in *E. coli*. However, most of the C-terminal α-amidating enzyme and derivatives thereof expressed by this method are forming inclusion bodies (a mass of inactive protein having the same amino acid sequence but does not have a higher-order structure, and thus is called insoluble granules) in *E. coli* and do not exhibit the activity of the C-terminal α-amidating enzyme.

Thus, an inert enzyme produced by such a method must be converted by some means (for example, refolding) to an active form. For this purpose, in the invention described in

Kokai (Japanese Unexamined Patent Publication) No. 7-250691, the C-terminal α -amidating enzyme expressed in *E. coli* was treated with a denaturing agent such as urea or guanidine hydrochloride, and then was refolded by lowering the concentration of the denaturing agent. However, the activity of the enzyme obtained by this method was about 10-15 mU per mL of the culture liquid, which was lower than that (2,860 U/mL culture liquid) of the amidating enzyme expressed in CHO cells described in the invention of Kokai (Japanese Unexamined Patent Publication) No. 7-163340.

DISCLOSURE OF THE INVENTION

Thus, it is an object of the present invention to provide a recombinant C-terminal α -amidating enzyme derivative that has a high enzyme activity compared to a conventional enzyme obtained by a gene recombinant technology using *E. coli*.

The present invention provides a recombinant C-terminal α -amidating enzyme derivative that has a high enzyme activity compared to a conventional enzyme obtained by a gene recombinant technology using *E. coli* by a method wherein a recombinant C-terminal α -amidating enzyme derivative of which amino acid sequence has been altered so as to prevent the formation of at least one specific disulfide bond of the five disulfide bonds that can be formed during refolding in the production, using a gene recombinant technology, of a *Xenopus laevis* C-terminal α -amidating enzyme derived having the amino acid sequence set forth in SEQ ID NO: 2 is expressed in *E. coli*, and the inclusion body obtained is solubilized under a non-reducing condition and subjected to a refolding procedure.

Specifically the above problem may be resolved by the following [1] to [7]:

[1] A recombinant C-terminal α -amidating enzyme derivative comprising:

(a) a polypeptide having an amino acid sequence in which at least one cysteine residue selected from the group consisting of cysteine residues at positions 6, 145, 40, 85, 252, and 274 has been altered in the amino acid sequence set forth in SEQ ID NO: 2; or

(b) a polypeptide having an amino acid sequence in which one or a few amino acid residues out of the amino acid residues other than the cysteine residue have been deleted, substituted, or added in the altered amino acid sequence described in the above (a) and having the activity of C-terminal α -amidating enzyme;

wherein at least one disulfide bond has not been formed out of the bonds between the cysteine residues at positions 6 and 145, between the cysteine residues at positions 40 and 85, and between the cysteine residues at positions 252 and 274.

[2] The C-terminal α -amidating enzyme derivative according to the above [1] wherein a disulfide bond has been formed between the cysteine residues at positions 73 and 90 and between the cysteine residues at positions 186 and 293 in the amino acid sequence set forth in SEQ ID NO: 2.

[3] The C-terminal α -amidating enzyme derivative according to the above [1] or [2] wherein said alteration is substitution with another amino acid or deletion of an amino acid.

[4] The C-terminal α -amidating enzyme derivative according to the above [1] to [3] that is AE-I [1-321] (C40A/C85A) (SEQ ID NO: 27), AE-I [1-321] (C252A/C274A) (SEQ ID NO: 33), AE-I [1-321] (C40A/C85A, C252A/C274A) (SEQ ID NO: 37), AE-I [8-321] (C145A) (SEQ ID NO: 25), or AE-I [8-321] (C145A, C40A/C85A) (SEQ ID NO: 35).

[5] DNA encoding the recombinant C-terminal α -amidating enzyme derivative according to any one of the above [1] to [4].

[6] An expression vector containing the DNA according to the above [5].

[7] An *Escherichia coli* transformed with the expression vector according to the above [6].

[8] A method of producing the recombinant C-terminal α -amidating enzyme derivative according to any one of the above [1] to [4], said method comprising the steps of culturing the *Escherichia coli* according to the above [7], allowing the recombinant C-terminal α -amidating enzyme derivative to be expressed, and then recovering the derivative thus obtained.

BRIEF EXPLANATION OF THE DRAWINGS

FIG. 1 shows a comparison of the amino acid sequence of the *Xenopus laevis* C-terminal α -amidating enzyme (SEQ ID NO: 39) and that of a rat C-terminal α -amidating enzyme (SEQ ID NO: 40). The upper row represents the amino acid sequence of the *Xenopus laevis* enzyme and the lower row represents that of the rat enzyme. The sequences shown here have a signal sequence at the N-terminal end and a transmembrane domain at the C-terminal end of the mature protein of the C-terminal α -amidating enzyme. The underlined parts show the amino acid residues conserved in both species, indicating a homology of about 65%. The boxed parts indicate the cysteine residues of both species, indicating that they are highly conserved.

FIG. 2 is a drawing that shows the putative sites of S—S bond in the C-terminal α -amidating enzyme derived from *Xenopus laevis*. As can be seen from FIG. 2, it may be estimated that five pairs of S—S bonds are formed between ⁶Cys-¹⁴⁵Cys, ⁴⁰Cys-⁸⁵Cys, ⁷³Cys-⁹⁰Cys, ¹⁸⁶Cys-²⁹³Cys, and ²⁵²Cys-²⁷⁴Cys.

FIG. 3 is a schematic drawing of the introduction of mutation by the PCR method.

FIG. 4 shows the base sequences of DNA primers for obtaining the gene fragments of the derivatives AE-I [8-321] (C145A) (SEQ ID NO: 25), AE-I [1-321] (C40A/C85A) (SEQ ID NO: 27), AE-I [1-321] (C73A/C90A) (SEQ ID NO: 29), AE-I [1-321] (C186A/C293A) (SEQ ID NO: 31), and AE-I [1-321] (C252A/C274A) (SEQ ID NO: 33). The boxed parts show the restriction enzymes BamHI (GGATCC) and XhoI (CTCGAG). The one base-insertion site of guanine (G) for adjusting the reading frame and the termination codon are underlined. The mutation site where cysteine is substituted with alanine is double underlined.

FIG. 5 shows the base sequences of DNA primers for obtaining the gene fragments of the derivatives AE-I [8-321] (C145A, C40A/C85A) (SEQ ID NO: 35), and AE-I [1-321] (C40A/C85A, C252A/C274A) (SEQ ID NO: 37). The boxed parts show the restriction enzymes BamHI (GGATCC) and XhoI (CTCGAG). The one base-insertion site of guanine (G) for adjusting the reading frame and the termination codon are underlined. The mutation site where cysteine is substituted with alanine is double underlined.

FIG. 6 shows the result of confirming the expression of the amidating enzyme and derivatives thereof by SDS-PAGE. Each lane is as follows:

Lanes 1 and 13: Marker (molecular weight: 175, 83, 62, 47.5, 32.5, 25, 16.5, 6.5 kDa)

Lane 2: AE-I [1-321] (SEQ ID NO: 2)

Lane 3: AE-I [8-321] (C145A) (SEQ ID NO: 25)

Lane 4: AE-I [1-321] (C40A/C85A) (SEQ ID NO: 27)

Lane 5: AE-I [1-321] (C73A/C90A) (SEQ ID NO: 29)

Lane 6: AE-I [1-321] (C186A/C293A) (SEQ ID NO: 31)

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Lane 7: AE-I [1-321] (C252A/C274A) (SEQ ID NO: 33)
Lane 8: AE-I [8-321] (C145A, C40A/C85A) (SEQ ID NO: 35)

Lane 11: AE-I [1-321] (C40A/C85A, C252A/C274A) (SEQ ID NO: 37)

Lanes 9, 10, and 12: -.

FIG. 7 represents a table showing the result of measuring the protein concentration and enzyme activity of the amidating enzymes and derivatives thereof after dialysis. FIG. 7 discloses the amidating enzyme constructs as SEQ ID NOS 2, 25, 27, 29, 31, 33, 35 and 37, respectively, in order of appearance.

BEST MODE FOR CARRYING OUT THE INVENTION

As described above, with regard to a problem that the C-terminal α -amidating enzyme expressed in *E. coli* is expressed as an inclusion body having no activity, the invention described in Kokai (Japanese Unexamined Patent Publication) No. 7-250691 partially resolved this problem by treating the inclusion body expressed in *E. coli* with a denaturing agent such as urea or guanidine hydrochloride and then by refolding it. However, the enzyme activity of the enzyme obtained by this method was as low as about 10-15 mU per mL of the culture liquid.

The present inventors assumed that the reason for such a low enzyme activity is that the plurality of cysteine residues present in the *Xenopus laevis* C-terminal α -amidating enzyme (said enzyme has ten cysteine residues, i.e. five pairs of S—S bonds, in the molecule) having the amino acid sequence set forth in SEQ ID NO: 2 cannot form the same disulfide bonds (S—S bonds) as those of the native form, i.e. erroneous S—S bonding is taking place during refolding.

The positions of S—S bonds in the *Xenopus laevis* C-terminal α -amidating enzyme have not been elucidated yet. Thus the present inventors investigated the homology of the amino acid sequence of a rat C-terminal α -amidating enzyme for which the positions of S—S bonds had been identified with that of a *Xenopus laevis* C-terminal α -amidating enzyme, and demonstrated that they have a high homology of 65.2% and the positions of cysteine residues in the regions corresponding to the amino acid sequence set forth in SEQ ID NO: 2 are completely conserved. And thus the present inventors assumed that the positions of S—S bonds are identical in them.

Thus, in order to prove the assumption that erroneous S—S bonding is taking place during refolding, the present inventors planned to use a *Xenopus laevis* C-terminal α -amidating enzyme and substitute a specific cysteine included in its amino acid sequence with alanine or delete it so as to create a recombinant C-terminal α -amidating enzyme derivative that were modified so that at least one pair of the five pairs of disulfide bonds capable of being formed by said enzyme derivative cannot be formed, and to obtain a recombinant C-terminal α -amidating enzyme derivative having a higher enzyme activity at a high yield by reducing the likelihood of erroneous S—S bonding to a minimum.

In the field of the present invention, with reference to a protein having a S—S bond in the molecule, it is common to newly introduce a S—S bond and to stabilize said protein in order to obtain said protein having a higher enzyme activity, as described in many reports such as Shimizu-Ibuka A. et al., "Effect of disulfide-bond introduction on the activity and stability of the extended-spectrum class A beta-lactamase To ho-1." Biochim. Biophys. Acta. 2006 August; 1764(3):1349-55, Epub 2006 Jun. 27, and Siadat O R et al., "The effect of

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engineered disulfide bonds on the stability of *Drosophila melanogaster* acetylcholinesterase." BMC Biochem. 2006 Apr. 16; 7:12. In Siadat O R et al., by newly introducing a S—S bond to acetylcholinesterase, the stability of the enzyme at 50° C. was successfully raised by about 170-fold compared to the wild type and resistance to denaturing agents, organic solvents, and proteases was successfully conferred.

In contrast, there are no reports to date on a method of achieving enhanced enzyme activity and stability by removing a S—S bond from a protein of interest as described in the present invention. In other words, those skilled in the art had presumed that since the removal of a S—S bond is likely to induce the reduction in the structural stability and activity of a protein of interest, it should be difficult to obtain the protein of interest having the desired activity by removing a S—S bond.

DEFINITION OF TERMS

As used herein the numbers in the amino acid sequence set forth in SEQ ID NO: 2 have been conferred with the serine residue at the N-terminal of the mature protein of C-terminal α -amidating enzyme (peptidyl-glycine alpha-amidating monooxygenase I, EC 1.14.17.3) being set as No. 1. As used herein, the positions of cysteine residues represent amino acid No. 6, 40, 73, 85, 90, 145, 186, 252, 274, and 293, respectively, in SEQ ID NO: 2.

As used herein the term "alteration" with reference to the cysteine residue represents, nonrestrictively, modification such as the deletion of said cysteine residue, substitution with another amino acid residue, the removal of an amino acid sequence having said cysteine residue or the addition of a protecting group to the thiol group of said cysteine residue, and encompasses any of the modifications that avoid the formation of a disulfide bond between the cysteine residues at positions 6 and 145, between the cysteine residues at positions 40 and 85, and between the cysteine residues at positions 252 and 274.

As used herein the term "C-terminal α -amidating enzyme" means an enzyme having an ability of catalyzing oxidation via copper ion (first stage: hydroxylation of a carbon of Gly) in the amidation (conversion of a —COOH group to a —CONH₂ group) of a glycine residue at the C-terminal end of the precursor of a peptide or protein having a C-terminal amide structure, and specifically means an enzyme having the amino acid sequence set forth in SEQ ID NO: 2.

As used herein the term "C-terminal α -amidating enzyme derivative" means an enzyme having an amino acid sequence in which the amino acid sequence of the above C-terminal α -amidating enzyme has been altered.

As used herein the term "C-terminal α -amidating enzyme activity" means an enzyme activity similar to that of the C-terminal α -amidating enzyme (peptidyl-glycine alpha-amidating monooxygenase I, EC 1.14.17.3) derived from *Xenopus laevis*.

As used herein the term "AE-I [1-321] (C40A/C85A)" means a polypeptide having an amino acid sequence (SEQ ID NO: 27) which has the primary sequence (SEQ ID NO: 2) of the amino acids in a region corresponding to from the serine residue at position 1 to the methionine residue at position 321 among the mature protein of the *Xenopus laevis* C-terminal α -amidating enzyme (peptidyl-glycine alpha-amidating monooxygenase I, EC 1.14.17.3), and in which the cysteine residue at position 40 has been substituted with an alanine residue and the cysteine residue at position 85 has been substituted with an alanine residue. The term "AE-I [1-321]", the term "AE-I [1-321] (C252A/C274A)" (SEQ ID NO: 33), the

term “AE-I [1-321] (C40A/C85A, C252A/C274A)” (SEQ ID NO: 37), the term “AE-I [1-321] (C73A/C90A)” (SEQ ID NO: 29), and the term “AE-I [1-321] (C186A/C293A)” (SEQ ID NO: 31) have a similar meaning. Except for “AE-I [1-321] (SEQ ID NO: 2),” they are simply termed as derivatives of C-terminal α -amidating enzyme.

As used herein the term “AE-I [8-321] (C145A)” means a polypeptide having an amino acid sequence (SEQ ID NO: 25) in which a region from the serine residue at position 1 to the leucine residue at position 7 has been deleted, a region from the glycine residue at position 8 to the methionine residue at position 321 is present, and the cysteine residue at position 145 has been substituted with an alanine residue in the primary sequence of amino acids comprising the serine residue at position 1 to the methionine residue at position 321 excluding a signal sequence comprising 37 amino acids present at the N-terminal end. Herein, by deleting a fragment from the serine residue at position 1 to the leucine residue at position 7, the cysteine residue at position 6 has been deleted. The term “AE-I [8-321] (C145A, C40A/C85A)” means a polypeptide having an amino acid sequence (SEQ ID NO: 35) in which a region from the glycine residue at position 8 to the methionine residue at position 321 are present as described above, and the cysteine residue at position 145 has been substituted with an alanine residue, the cysteine residue at position 40 has been substituted with an alanine residue, and the cysteine residue at position 85 has been substituted with an alanine residue. This is simply termed as a derivative of C-terminal α -amidating enzyme.

Method of Measuring Enzyme Activity and Unit

When the enzyme is expressed in *E. coli*, the majority of it may be recovered in the precipitate fraction after cell disruption. Thus for the measurement of the enzyme activity, a sample prepared by solubilizing the precipitate fraction with 6M guanidine hydrochloride and then dialyzing with a guanidine hydrochloride solution is used. Generally, the enzyme activity can be determined by using a substrate represented by R-X-Gly or an amidated peptide expressed and converting it to R-X-CONH₂ (e.g., the conversion of a synthetic substrate [¹²⁵I]-Ac-Tyr-Phe-Gly to [¹²⁵I]-Ac-Tyr-Phe-NH₂). Thus, a labelled substrate (labelled R-X-Gly) is first reacted to a test enzyme solution in a Tris-HCl buffer. The Tris-HCl buffer and ethyl acetate is added thereto, and after mixing, it is separated by centrifugation to the organic solvent phase and the aqueous phase. Here, since the majority of the unreacted labelled substrate (labelled R-X-Gly) migrates to the aqueous phase and the amidated labelled substrate (labelled R-X-CONH₂) to the organic solvent phase, they can be easily separated. The rate of conversion to the C-terminal α -amidated product can be determined from the ratio of radioactivity of the organic solvent phase to the total radioactivity. In the present determination method, the enzyme activity in which 50% of 1 pmol of the labelled R-X-Gly (substrate) is converted to the labelled R-X-CONH₂ per hour is defined as one Unit.

The amidating enzyme activity was assessed by dealkylation the enzyme through the addition of an alkali (sodium hydroxide) after the oxidation reaction.

Thus, 2 μ l, 10 μ l, and 100 μ l each of a sample to be determined was taken and added to distilled water to make a total volume of 100 μ l. Then 10 mM ascorbic acid (25 μ l), 200 μ M copper sulfate (25 μ l), 20 mg/mL catalase (1.25 μ l), 1% Lubrol (20 μ l), [¹²⁵I]-Ac-Tyr-Phe-Gly (2 μ mol), 1M Tris-HCl (pH 7.0, 50 μ l) and distilled water (25 μ l) were added thereto, and reacted at 37° C. for 1 hour. After the reaction, 250 mM NaOH (250 μ l) was added to the reaction mixture, mixed, and after allowing to stand at room temperature for 15 minutes

(dealkylation), 1M Tris-HCl (pH 7.0, 500 μ l) and ethyl acetate (2 mL) were added thereto, mixed, and centrifuged. Then, 1 mL of the ethyl acetate layer was aliquoted, and the radioactivity of it and of the rest of the solution were measured using a gamma counter to determine the ratio of radioactivity that migrated to the ethyl acetate layer. That the C-terminal amidated [¹²⁵I]-Ac-Tyr-Phe-CONH₂ migrates specifically to the ethyl acetate layer in this method has been confirmed by determination with a liquid chromatography or a gamma counter. The enzyme activity in which 50% of 1 pmol of the labelled R-X-Gly (substrate) is converted to the labelled R-X-CONH₂ per hour is defined as one Unit.

FIG. 1 shows an alignment of the amino acid sequence of the *Xenopus laevis* C-terminal α -amidating enzyme (peptidyl-glycine α -amidating monooxygenase I, EC 1.14.17.3) claimed in the present invention and that of a rat enzyme of which crystal structure has already been analyzed in Prigge S T, Kolhekar A S, Eipper B A, Mains R E, Amzel L M. “Amidation of bioactive peptides: the structure of peptidylglycine α -hydroxylating monooxygenase.” Science 1997 Nov. 14; 278(5341):1300-5. As can be seen from FIG. 1, they have a high homology of 65.2%, and the positions of cysteine residues present in the region corresponding to the amino acid sequence set forth in SEQ ID NO: 2 are completely conserved.

FIG. 2 shows a conformational structure of a rat C-terminal α -amidating enzyme of which crystal structure has been analyzed by Prigge S T et al. and in which the positions of S—S bonds have been identified. The present inventors assumed that, based on the positions of S—S bonds in said conformational structure, the conformational structure of the *Xenopus laevis* C-terminal α -amidating enzyme claimed in the present invention can be maintained by forming five pairs of S—S bonds between ⁶Cys-¹⁴⁵Cys, ⁴⁰Cys-⁸⁵Cys, ⁷³Cys-⁹⁰Cys-¹⁸⁶Cys-²⁹³Cys, and ²⁵²Cys-²⁷⁴Cys.

Based on such assumed positions of S—S bonds, recombinant polypeptides in which at least one S—S bond is not formed were created. Thus, based on a plasmid, pPROEXHTa AE-I [1-321] (SEQ ID NO: 2), comprising a sequence encoding the primary sequence of amino acids from No. 1 to 321 of the amino acid sequence of the *Xenopus laevis* C-terminal α -amidating enzyme AE-I as a wild type, plasmids were created in which at least one S—S bond is not formed. These were plasmids designed so as to be expressed in *E. coli* under the control of a trc (a fusion type of lac and trp) promoter.

By site-directed mutagenesis with pPROEXHTa AE-I [1-321] (SEQ ID NO: 2) as a template, paired cysteine residues capable of forming S—S bonds were substituted with alanine residues or deleted to create plasmids pPROEXHTa AE-I [8-321] (C145A) (SEQ ID NO: 25), pPROEXHTa AE-I [1-321] (C40A/C85A) (SEQ ID NO: 27), pPROEXHTa AE-I [1-321] (C73A/C90A) (SEQ ID NO: 29), pPROEXHTa AE-I [1-321] (C186A/C293A) (SEQ ID NO: 31), and pPROEXHTa AE-I [1-321] (C252A/C274A) (SEQ ID NO: 33) having a sequence that has been mutated so that only one of the above five pairs of S—S bonds is not formed. Furthermore, based on these derivative plasmids, plasmids pPROEXHTa AE-I [8-321] (C145A, C40A/C85A) (SEQ ID NO: 351) and pPROEXHTa AE-I [1-321] (C40A/C85A, C252A/C274A) (SEQ ID NO: 37) were created having a sequence that has been mutated so that only two pairs of the above five pairs of S—S bonds are not formed. Though the number of cysteine residues to be altered is two in the above, the number of cysteine residues to be altered is not limited to it, and it is possible that only one S—S bond is not formed by altering one cysteine residue.

By transforming *E. coli* using these plasmids according to a standard method, recombinant *E. coli* cells in which the gene of interest having the above site-directed mutation has been introduced were obtained. These recombinant *E. coli* cells were cultured to express the desired substance as inclusion bodies in the cells. The cells were disrupted and centrifuged to recover the inclusion bodies as the precipitate fractions. After the inclusion bodies obtained were denatured with a denaturing agent, they were subjected to refolding by diluting with a denaturant-free buffer. The amidating enzymes and derivatives thereof obtained by refolding were assessed by determining the amidating enzyme activity using a synthetic substrate.

Finally, five derivatives exhibiting an enzyme activity higher than the wild type AE-I [1-321] (SEQ ID NO: 2) were obtained (i.e., derivatives AE-I [8-321] (C145A) (SEQ ID NO: 25) AE-I [1-321] (C40A/C85A) (SEQ ID NO: 27) and AE-I [1-321] (C252A/C274A) (SEQ ID NO: 33) that cannot form one pair of S—S bond, and derivatives AE-I [8-321] (C145A, C40A/C85A) (SEQ ID NO: 35) and AE-I [1-321] (C40A/C85A, C252A/C274A) (SEQ ID NO: 37) that cannot form two pairs of S—S bonds).

It is generally estimated that the removal of a S—S bond may lead to reduced stability and reduced activity of the protein. As shown in FIG. 7, however, the present inventors succeeded in obtaining a derivative exhibiting an enzyme activity higher than the wild type for a C-terminal α -amidating enzyme by removing at least one pair of the S—S bond therein.

In said derivatives that exhibit an enzyme activity higher than the wild type, a disulfide bond had been formed between the cysteine residues at positions 73 and 90 and between the cysteine residues at positions 186 and 293 in the amino acid sequence set forth in SEQ ID NO: 2.

The present invention will now be explained in more detail with reference to the following examples.

EXAMPLES

Working Example 1

Preparation of a C-Terminal α -Amidating Enzyme and its Derivative (1) Creation of *E. coli* Expression Plasmids pPROEXHTa AE-I [8-321] (C145A) (SEQ ID NO: 25), pPROEXHTa AE-I [1-321] (C40A/C85A) (SEQ ID NO: 27), pPROEXHTa AE-I [1-321] (C73A/C90A) (SEQ ID NO: 29), pPROEXHTa AE-I [1-321] (C186A/C293A) (SEQ ID NO: 31), and pPROEXHTa AE-I [1-321] (C252A/C274A) (SEQ ID NO: 33) of Derivatives

The derivatives of the amidating enzyme were created based on a plasmid pPROEXHTa AE-I [1-321] (SEQ ID NO: 2) encoding the amino acid sequence comprising amino acids 1-321 of the amino acid sequence of the C-terminal α -amidating enzyme derived from *Xenopus laevis*. pPROEXHTa AE-I [1-321] (SEQ ID NO: 2) is a plasmid designed to be expressed in *E. coli* under the control of the trc (a fusion type of lac and trp) promoter. With this plasmid pPROEXHTa AE-I [1-321] (SEQ ID NO: 2) as a template, a derivative incapable of forming a S—S bond was created by replacing a pair of two cysteine residues each with an alanine residue by site-directed mutagenesis on a protein having an amino acid sequence comprising amino acids at positions 1 to 321 or 8 to 321. Since, for the derivative AE-I [8-321] (C145A) (SEQ ID NO: 25), cysteines at positions 6 and 145 of the amino acid sequence pair to form a S—S bond, the amino acid residues at

positions 1 to 7 were removed in stead of substituting the cysteine residue at position 6 with an alanine residue, and the cysteine residue at position 145 was substituted with an alanine residue so that a S—S bond may not be formed between ⁶Cys-¹⁴⁵Cys.

The derivative incapable of forming a pair of S—S bond was created by substituting a cysteine residue with an alanine residue by mutagenesis using a PCR method. FIG. 3 depicts an outline of the method of preparing said derivative.

First, with pPROEXHTa AE-I [1-321] (its structural gene sequence and the corresponding amino acid sequence are shown in SEQ ID NOS 1 and 2, respectively) as a template, primers for respective derivatives were created. FIG. 4 shows the sequences of primers used. In FIG. 4, primers P1 and P1' have, in addition to a restriction enzyme BamHI site (boxed) at the 5'-end, guanine (underlined) inserted therein in order to adjust with the reading frame, and the primer P6 has, in addition to a restriction enzyme XhoI site (boxed) at the 5'-end, a termination codon antisense chain TTA (underlined) inserted therein. Using primers P1 and P2, P3 and P4, and P5 and P6 (or P1' and P4, and P5 and P6) (only P2, P3, P4, and P5 contain mutation) of respective derivatives, DNA fragments were amplified, and subjected to agarose gel electrophoresis and Gel Extraction Kit (Quiagen) to obtain three (or two) purified DNA fragments. Using primers P1 and P6 (or P1' and P6) (P1, P1', and P6 do not contain mutation) with the mixture of all these DNA fragments as the template, DNA fragments were PCR-amplified again to obtain an about 960 bp DNA fragment having mutation introduced therein for each derivative.

The DNA fragment obtained for each derivative was purified by the Gel Extraction Kit (Quiagen). The purified fragment was cleaved with restriction enzymes BamHI and XhoI to obtain BamHI-XhoI-digested DNA fragments, AE-I [8-321] (C145A) (SEQ ID NO: 25), AE-I [1-321] (C40A/C85A) (SEQ ID NO: 27), AE-I [1-321] (C73A/C90A) (SEQ ID NO: 29), AE-I [1-321] (C186A/C293A) (SEQ ID NO: 31), and AE-I [1-321] (C252A/C274A) (SEQ ID NO: 33). Simultaneously, after pPROEXHTa to be used as an expression vector was cleaved with restriction enzymes BamHI and XhoI, an about 4.7 kb DNA fragment containing said expression vector part was separated and purified. This product and a DNA fragment of each derivative obtained earlier were ligated with the DNA Ligation Kit (TaKaRa) to finally obtain plasmids pPROEXHTa AE-I [8-321] (C145A), pPROEXHTa AE-I [1-321] (C40A/C85A), pPROEXHTa AE-I [1-321] (C73A/C90A), pPROEXHTa AE-I [1-321] (C186A/C293A), and pPROEXHTa AE-I [1-321] (C252A/C274A) of each derivative (the structural gene sequence and the corresponding amino acid sequence of each derivative are shown in SEQ ID NO: 25, 27, 29, 31 or 33, respectively). The plasmid pPROEXHTa (Gibco) is an expression vector having the constitution of the trc (the fusion type of lac and trp) promoter, followed by His tag (His \times 6 tag) (SEQ ID NO: 38), a multi cloning site, and β -lactamase.

Working Example 2

Preparation of C-Terminal α -Amidating Enzyme and its Derivative (2) Creation of *E. coli* Expression Plasmids pPROEXHTa AE-I [8-321] (C145A, C40A/C85A) (SEQ ID NO: 35) and pPROEXHTa AE-I [1-321] (C40A/C85A, C252A/C274A) (SEQ ID NO: 37) of Derivatives

Plasmids pPROEXHTa AE-I [8-321] (C145A, C40A/C85A) (SEQ ID NO: 35) and pPROEXHTa AE-I [1-321]

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(C40A/C85A, C252A/C274A) (SEQ ID NO: 37) of derivatives incapable of forming two of the five pairs of S—S bonds owned by AE-I [1-321] (SEQ ID NO: 2) were created with pPROEXHTa AE-I [8-321] (C145A) (SEQ ID NO: 25) and pPROEXHTa AE-I [1-321] (C40A/C85A) (SEQ ID NO: 27) as the template. Methods of introducing mutation and creating expression vectors were similar to those in Working Example 1.

The gene fragments of the above derivatives were created using a PCR method. A method similar to the one in Working Example 1 was used. First, using primers P1 (or P1') and P2, P3 and P4, and P5 and P6 (only P2, P3, P4, and P5 contain mutation) of respective derivatives with pPROEXHTa AE-I [8-321] (C145A) (SEQ ID NO: 25) and pPROEXHTa AE-I [1-321] (C40A/C85A) (SEQ ID NO: 27) as the template, DNA fragments were amplified, and subjected to agarose gel electrophoresis and the Gel Extraction Kit (Quiagen) to obtain three purified DNA fragments. With the mixture of these three DNA fragments as the template, primers P1 (or P1') and P6 (P1, P1' and P6 do not contain mutation) were PCR-amplified again to obtain an about 960 bp DNA fragment having mutation introduced therein for each derivative. The sequences of primers are shown in FIG. 5. In FIG. 5, primers P1 and P1' have, in addition to a restriction enzyme BamHI site (boxed) at the 5'-end, guanine (underlined) inserted therein in order to adjust with the reading frame, and the primer P6 has, in addition to a restriction enzyme XhoI site (boxed) at the 5'-end, a termination codon antisense chain TTA (underlined) inserted therein.

The DNA fragment obtained for each derivative was purified by the Gel Extraction Kit (Quiagen). The purified fragment was cleaved with restriction enzymes BamHI and XhoI to obtain BamHI-XhoI-digested DNA fragments, AE-I [8-321] (C145A, C40A/C85A) (SEQ ID NO: 35) and AE-I [1-321] (C40A/C85A, C252A/C274A) (SEQ ID NO: 37). Simultaneously, after pPROEXHTa to be used as an expression vector was cleaved with restriction enzymes BamHI and XhoI, an about 4.7 kb DNA fragment containing said expression vector part was separated and purified. This product and a DNA fragment of each derivative obtained earlier were ligated with the DNA Ligation Kit (TaKaRa) to finally obtain plasmids pPROEXHTa AE-I [8-321] (C145A, C40A/C85A) and pPROEXHTa AE-I [1-321] (C40A/C85A, C252A/C274A) of each derivative (the structural gene sequence and the amino acid sequence of each derivative are shown in SEQ ID NO: 35 or 37, respectively).

Working Example 3

Introduction of pPROEXHTa AE-I [1-321] (SEQ ID NO: 2), pPROEXHTa AE-I [8-321] (C145A) (SEQ ID NO: 25), pPROEXHTa AE-I [1-321] (C40A/C85A) (SEQ ID NO: 27), pPROEXHTa AE-I [1-321] (C73A/C90A) (SEQ ID NO: 29), pPROEXHTa AE-I [1-321] (C186A/C293A) (SEQ ID NO: 31), pPROEXHTa AE-I [1-321] (C252A/C274A) (SEQ ID NO: 33), pPROEXHTa AE-I [8-321] (C145A, C40A/C85A) (SEQ ID NO: 35), and pPROEXHTa AE-I [1-321] (C40A/C85A, C252A/C274A) (SEQ ID NO: 37) into *E. coli* and their Expression

Using the plasmids of the amidating enzyme and its derivatives, *E. coli* JM109 was transformed. The transformed *E. coli* was cultured in about 1 liter of the LB medium (0.5% (w/v) yeast extract, 1% (w/v) tryptone, 0.5% (w/v) NaCl) under shaking at 37° C., and the expression was induced by the

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addition of isopropyl- β -D-thiogalactoside (IPTG). After the induction of expression, culturing was continued for about 12-16 hours. The cells obtained were disrupted, and after centrifugation the precipitate fraction containing the inclusion body was recovered. By washing the precipitate with a Triton X-100 (detergent)-containing buffer, the proteins and membrane components derived from JM109 were removed to recover the inclusion bodies of the amidating enzyme and its derivatives. The expression and purity of the amidating enzyme and its derivatives were confirmed by SDS-PAGE (see FIG. 6). The expression level was determined by the UV method after solubilizing the inclusion body with a denaturing agent.

To the *E. coli* JM109 that was made competent, each of expression vectors (pPROEXHTa AE-I [1-321] (SEQ ID NO: 2), pPROEXHTa AE-I [8-321] (C145A) (SEQ ID NO: 25), pPROEXHTa AE-I [1-321] (C40A/C85A) (SEQ ID NO: 27), pPROEXHTa AE-I [1-321] (C73A/C90A) (SEQ ID NO: 29), pPROEXHTa AE-I [1-321] (C186A/C293A) (SEQ ID NO: 31), pPROEXHTa AE-I [1-321] (C252A/C274A) (SEQ ID NO: 33), pPROEXHTa AE-I [8-321] (C145A, C40A/C85A) (SEQ ID NO: 35), and pPROEXHTa AE-I [1-321] (C40A/C85A, C252A/C274A) (SEQ ID NO: 37)) created in Working Examples 1 and 2 was added. After incubating on ice for 10 minutes, they were inoculated into a LB-agar medium (0.5% (w/v) yeast extract, 1% (w/v) tryptone, 0.5% (w/v) NaCl, 1.5% (w/v) agar) containing 10 μ g/mL ampicillin (antibiotics), and incubated overnight at 37° C. to obtain the colonies of transformants JM109[pPROEXHTa AE-I [1-321] (SEQ ID NO: 2)], JM109[pPROEXHTa AE-I [8-321] (C145A) (SEQ ID NO: 25)], JM109[pPROEXHTa AE-I [1-321] (C40A/C85A) (SEQ ID NO: 27)], JM109[pPROEXHTa AE-I [1-321] (C73A/C90A) (SEQ ID NO: 29)], JM109[pPROEXHTa AE-I [1-321] (C186A/C293A) (SEQ ID NO: 31)], JM109[pPROEXHTa AE-I [1-321] (C252A/C274A) (SEQ ID NO: 33)], JM109[pPROEXHTa AE-I [8-321] (C145A, C40A/C85A) (SEQ ID NO: 35)], and JM109[pPROEXHTa AE-I [1-321] (C40A/C85A, C252A/C274A) (SEQ ID NO: 37) of respective derivatives.

Colonies of these transformants were each inoculated into a test tube containing 5 mL of the LB medium (0.5% (w/v) yeast extract, 1% (w/v) tryptone, 0.5% (w/v) NaCl) containing 10 μ g/mL ampicillin (antibiotics), and cultured under shaking at 37° C. for about 12-16 hours. The entire volume of the culture liquid was inoculated into 1 liter of the LB medium containing 10 μ g/mL ampicillin and cultured under shaking at 37° C. At 3-6 hours after culturing (when OD₆₆₀ nM reached 0.5-0.8), IPTG was added to a final concentration of 1 mM to induce expression.

Since the C-terminal α -amidating enzyme and its derivatives are intracellularly expressed as insoluble inclusion bodies, the inclusion bodies were recovered in the following manner. The cells were recovered by centrifuging (6000 rpm, 4° C.) 1 liter of the culture liquid for 10 minutes, suspended in 100 mL of water, and then the cells were disrupted by French press (10,000 psi; twice). The cell-disrupted liquid was centrifuged for 15 minutes (6000 rpm, 4° C.), and the inclusion body of interest was transferred to the precipitate fraction. Since this procedure transfers most of the protein derived from the host *E. coli* JM109 migrates to the supernatant, said protein can be removed. Then, the precipitate fraction was suspended in 50 mL of 100 mM Tris-HCl buffer, pH 7.0, containing 1% (w/w) Triton X-100 (detergent), and centrifuged (6000 rpm, 4° C.) for 15 minutes to recover the precipitate (thus, the membrane components etc. derived from JM109 is dissolved in the detergent and migrated to the supernatant, said membrane components etc. can be removed). By

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repeating this procedure twice, the inclusion body of the C-terminal α -amidating enzyme was recovered, which was finally suspended into 1 mL of 100 mM Tris-HCl buffer, pH 7.0, containing 1% (w/w) Triton X-100 (detergent) to obtain an inclusion body suspension.

The inclusion body suspensions (10 μ l) obtained of each derivative was diluted 2-fold with 10 μ l of a sample buffer (2M urea, 375 mM Tris-HCl, pH 6.8, 30% (v/v) glycerol, 7% (w/v) SDS, 15% (v/v) 2-mercaptoethanol, 0.1% (w/v) bromophenol blue) for SDS-PAGE, and a 0.1 μ l aliquot (corresponding to 1-10 μ g portion) was subjected to a 10% SDS-PAGE gel to confirm the expression and purity (see FIG. 6). In the C-terminal α -amidating enzyme and all of its derivatives, a band was detected at a molecular weight of about 40 kDa, and the purity was about 70-90%.

The inclusion body suspensions (10 μ l) of the amidating enzyme and all of its derivatives were solubilized with 10 mL of a denaturing agent (8M urea), and absorbance A at a wavelength of 280 nm was measured by a spectrophotometer to calculate the concentration C based on the Lambert-Beer's law according to the following equation:

$$\text{Concentration } C(\text{mg/mL}) = A \cdot \text{Mw} / \text{kd}$$

wherein, A is absorbance at a wavelength of 280 nm, Mw is molecular weight (about 45,000 Da), and kd is extinction coefficient (41,700 ($\text{M}^{-1} \cdot \text{cm}^{-1}$); The C-terminal α -amidating enzyme and derivatives include 3 and 21 amino acids of tryptophan (extinction coefficient 5500) and tyrosine (extinction coefficient 1200), respectively.

Based on the concentration C thus calculated, the expression levels of the proteins of the amidating enzyme and its derivatives were calculated to be 100-160 mg/liter of the medium.

Working Example 4

Refolding and Assessment of Enzyme Activity of the Amidating Enzyme AE-I [1-321] (SEQ ID NO: 2) and its derivatives AE-I [8-321] (C145A) (SEQ ID NO: 25), AE-I [1-321] (C40A/C85A) (SEQ ID NO: 27), AE-I [1-321] (C73A/C90A) (SEQ ID NO: 29), AE-I [1-321] (C186A/C293A) (SEQ ID NO: 31), AE-I [1-321] (C252A/C274A) (SEQ ID NO: 33), AE-I [8-321] (C145A, C40A/C85A) (SEQ ID NO: 35), and AE-I [1-321] (C40A/C85A, C252A/C274A) (SEQ ID NO: 37)

Using the inclusion bodies of the amidating enzyme and its derivatives obtained in Working Example 3, the refolding procedure was carried out by denaturing them with a 8M urea buffer having a denaturing effect and then by diluting with a denaturant-free buffer. Since the denaturing agent remains in the refolding solution obtained, it was removed by dialysis. Denaturing is known to inhibit the measurement of enzyme activity. For the dialyzed samples obtained, C-terminal α -amidating enzyme activity was assessed.

One mL of the inclusion bodies of the C-terminal α -amidating enzyme and its derivatives obtained in Working Example 3 were solubilized to a final concentration of 2.4 g/L with 8M urea, 50 mM Tris-HCl (pH 10.0 at 15° C.), and 50 mM NaCl. The solution was incubated at 15° C. for 2-4 days to break down S—S bonds.

Then, the denaturant concentration was lowered by diluting 8-fold with 50 mM Tris-HCl (pH 8.0 at 4° C.) and 50 mM NaCl to facilitate regeneration of conformation (refolding procedure). Furthermore, since urea is known to inhibit the measurement of activity, dialysis was performed overnight

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with 500 mL of 50 mM Tris-HCl (pH 8.0 at 4° C.) and 50 mM NaCl at 4° C. The dialysis membrane used was SPECTRUM's SPECTRA/Por 2 MWCO: 12-14,000 Da.

After refolding, the concentration of the protein obtained for the solution after dialysis was determined by absorbance at 280 nm in a manner similar to that in Working Example 3 (see FIG. 7). For the measurement of enzyme activity of the C-terminal α -amidating enzyme or its derivatives, the conversion of a synthetic substrate [^{125}I]-Ac-Tyr-Phe-Gly to [^{25}I]-Ac-Tyr-Phe-NH₂ was utilized. The method of determining C-terminal α -amidating enzyme activity and the definition of Unit are as described above.

FIG. 7 shows the result of determining the enzyme activity of the C-terminal α -amidating enzyme and its derivatives. The enzyme activity U per mg of protein is as follows: AE-I [1-321] (SEQ ID NO: 2): 205 U/mg, AE-I [8-321] (C145A) (SEQ ID NO: 25): 840 U/mg, AE-I [1-321] (C40A/C85A) (SEQ ID NO: 27): 1798 U/mg, AE-I [1-321] (C73A/C90A) (SEQ ID NO: 29): 56 U/mg, AE-I [1-321] (C186A/C293A) (SEQ ID NO: 31): ND U/mg, AE-I [1-321] (C252A/C274A) (SEQ ID NO: 33): 271 U/mg, AE-I [8-321] (C145A, C40A/C85A) (SEQ ID NO: 35): 778 U/mg, and AE-I [1-321] (C40A/C85A, C252A/C274A) (SEQ ID NO: 37): 2260 U/mg. As a result, compared to the wild type AE-I [1-321] (SEQ ID NO: 2) in which no S—S bonds have been removed, a low enzyme activity was exhibited by AE-I [1-321] (C73A/C90A) (SEQ ID NO: 29) and AE-I [1-321] (C186A/C293A) (SEQ ID NO: 31), and in AE-I [1-321] (C186A/C293A) (SEQ ID NO: 31) among others, no enzyme activity was detected. In other words, it was suggested that the S—S bond formed between ¹⁸⁶Cys-²⁹³Cys be important in the activity expression of an amidating enzyme.

On the other hand, compared to the wild type AE-I [1-321] (SEQ ID NO: 2) in which no S—S bonds have been removed, a high enzyme activity was exhibited by AE-I [8-321] (C145A) (SEQ ID NO: 25), AE-I [1-321] (C40A/C85A) (SEQ ID NO: 27) and AE-I [1-321] (C252A/C274A) (SEQ ID NO: 33) in which one pair of S—S bond has been removed, as well as AE-I [8-321] (C145A, C40A/C85A) (SEQ ID NO: 35) and AE-I [1-321] (C40A/C85A, C252A/C274A) (SEQ ID NO: 37) in which two pairs of S—S bonds have been removed. Among them, AE-I [8-321] (C145A) (SEQ ID NO: 25) and AE-I [1-321] (C40A/C85A) (SEQ ID NO: 27) exhibited the enzyme activity about 4-fold and about 9-fold higher, respectively, compared to the wild type, and AE-I [1-321] (C40A/C85A, C252A/C274A) (SEQ ID NO: 37) exhibited the enzyme activity as high as about 11-fold.

The enzyme activity per mL of the culture liquid for these five derivatives was calculated to be (in the calculation, the mean expression level was assumed to be 130 mg/L of the culture medium): AE-I [1-321] (SEQ ID NO: 2): 27 U/mL, AE-I [8-321] (C145A) (SEQ ID NO: 25): 109 U/mL, AE-I [1-321] (C40A/C85A) (SEQ ID NO: 27): 234 U/mL, AE-I [1-321] (C252A/C274A) (SEQ ID NO: 33): 35 U/mL, AE-I [8-321] (C145A, C40A/C85A) (SEQ ID NO: 35): 101 U/mL, and AE-I [1-321] (C40A/C85A, C252A/C293A) (SEQ ID NO: 37): 294 U/mL.

In the invention described in Kokai (Japanese Unexamined Patent Publication) No. 7-250691, the activity of the amidating enzyme recombinantly produced using *E. coli* was about 10-15 mU per mL of the culture liquid. In contrast, the enzyme activity of the derivatives obtained in the above method of the present invention was about 35-300 U/mL as described above. Though simple comparison may be not applicable because of differences in the expression level of the enzyme and the method of refolding, the enzyme activity was enhanced by about 2,000-30,000 fold compared to that of

the amidating enzyme obtained in the invention described in Kokai (Japanese Unexamined Patent Publication) No. 7-250691.

It has been confirmed that with regard to the culturing of the C-terminal α -amidating enzyme derivative of the present invention, a high-density culture can lead to the expression level of said derivative at about 5-10 g/L. In this case, the enzyme activity finally obtained may be calculated to be about 23,000 U per mL of the culture liquid at the maximum, which far exceeds the enzyme activity (an enzyme activity of 2,860 U per mL of the culture liquid) obtained for the enzyme recombinantly produced using CHO cells in the invention described in Kokai (Japanese Unexamined Patent Publication) No. 7-163340.

In accordance with the present invention, a recombinant C-terminal α -amidating enzyme derivative was obtained having a very high enzyme activity compared to the enzyme activity per mL of the culture liquid attained in the conventional technology (see Kokai (Japanese Unexamined Patent Publication) No. 7-250691) in the production of an amidating enzyme using *E. coli*. In accordance with the present invention furthermore, a recombinant C-terminal α -amidating

enzyme derivative was obtained having a high enzyme activity compared to the enzyme activity attained in a gene recombinant technology (see Kokai (Japanese Unexamined Patent Publication) No. 7-163340) using CHO cells. Since the method claimed in the present invention uses *E. coli*, it can produce said amidating enzyme in a short period of time and its productivity is very high compared to the above CHO cell-culturing method.

It has also been confirmed that the C-terminal α -amidating enzyme of the present invention amidates glucagon like peptide-1 (GLP-1) precursor (Gly is added to the C-terminal) in vitro, which indicates that the recombinant C-terminal α -amidating enzyme of the present invention can be fully used in the amidation reaction for the production of C-terminal α -amidated peptides.

INDUSTRIAL APPLICABILITY

The present invention can provide a recombinant C-terminal α -amidating enzyme derivative having an enzyme activity higher than the conventional enzymes recombinantly produced using *E. coli*.

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1 5

acg cgg ccc gtt atg tct cca ggc tca tca gat tat act cta gat atc 159
Thr Arg Pro Val Met Ser Pro Gly Ser Ser Asp Tyr Thr Leu Asp Ile
10 15 20 25

cgc atg cca gga gta act ccg aca gag tcg gac aca tat ttg tgc aag 207
Arg Met Pro Gly Val Thr Pro Thr Glu Ser Asp Thr Tyr Leu Cys Lys
30 35 40

tct tac cgg ctg cca gtg gat gat gaa gcc tat gta gtt gac ttc aga 255
Ser Tyr Arg Leu Pro Val Asp Asp Glu Ala Tyr Val Val Asp Phe Arg
45 50 55

cca cat gcc aat atg gat act gca cat cac atg ctt cta ttt gga tgc 303
Pro His Ala Asn Met Asp Thr Ala His His Met Leu Leu Phe Gly Cys
60 65 70

aat ata cct tct tcc act gat gat tac tgg gac tgt agt gcg gga act 351
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75 80 85

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Cys Met Asp Lys Ser Ser Ile Met Tyr Ala Trp Ala Lys Asn Ala Pro
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ccc acc aaa ctt cca gaa gga gtt ggc ttt cgt gtt gga ggg aaa tca 447
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ggc tca tca gat tat act cta gat atc cgc atg cca gga gta act ccg      159
Gly Ser Ser Asp Tyr Thr Leu Asp Ile Arg Met Pro Gly Val Thr Pro
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Thr Glu Ser Asp Thr Tyr Leu Cys Lys Ser Tyr Arg Leu Pro Val Asp
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Asp Glu Ala Tyr Val Val Asp Phe Arg Pro His Ala Asn Met Asp Thr
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gca cat cac atg ctt cta ttt gga tgc aat ata cct tct tcc act gat      303
Ala His His Met Leu Leu Phe Gly Cys Asn Ile Pro Ser Ser Thr Asp
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Asp Tyr Trp Asp Cys Ser Ala Gly Thr Cys Met Asp Lys Ser Ser Ile
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Met Tyr Ala Trp Ala Lys Asn Ala Pro Pro Thr Lys Leu Pro Glu Gly
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Gln Val His Tyr Gly Asn Val Lys Ala Phe Gln Asp Lys His Lys Asp
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gcc acg ggg gtg aca gta cga gta aca cct gaa aaa caa ccg caa att      543
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gca ggc att tat ctt tca atg tct gtg gac act gtt att cca cct ggg      591
Ala Gly Ile Tyr Leu Ser Met Ser Val Asp Thr Val Ile Pro Pro Gly
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gaa gag gca gtt aat tct gat atc gcc tgc ctc tac aac agg ccg aca      639
Glu Glu Ala Val Asn Ser Asp Ile Ala Cys Leu Tyr Asn Arg Pro Thr
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Ile His Pro Phe Ala Tyr Arg Val His Thr His Gln Leu Gly Gln Val
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gta agt gga ttt aga gtg aga cat ggc aag tgg tct tta att ggt aga      735
Val Ser Gly Phe Arg Val Arg His Gly Lys Trp Ser Leu Ile Gly Arg
              205              210              215

caa agc cca cag ctg cca cag gca ttt tac cct gta gag cat cca gta      783
Gln Ser Pro Gln Leu Pro Gln Ala Phe Tyr Pro Val Glu His Pro Val
              220              225              230

gag att agc cct ggg gat att ata gca acc agg tgt ctg ttc act ggt      831
Glu Ile Ser Pro Gly Asp Ile Ile Ala Thr Arg Cys Leu Phe Thr Gly
              235              240              245

aaa ggc agg acg tca gca aca tat att ggt ggc aca tct aac gat gaa      879
Lys Gly Arg Thr Ser Ala Thr Tyr Ile Gly Gly Thr Ser Asn Asp Glu
250              255              260              265
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atg	tgt	aat	tta	tac	atc	atg	tat	tac	atg	gat	gcg	gcc	cat	gct	acg	927
Met	Cys	Asn	Leu	Tyr	Ile	Met	Tyr	Tyr	Met	Asp	Ala	Ala	His	Ala	Thr	
			270						275					280		
tca	tac	atg	acc	tgt	gta	cag	acg	ggg	gaa	cca	aag	tta	ttt	caa	aac	975
Ser	Tyr	Met	Thr	Cys	Val	Gln	Thr	Gly	Glu	Pro	Lys	Leu	Phe	Gln	Asn	
			285					290					295			
atc	cct	gag	att	gca	aat	gtt	ccc	att	cct	gta	agc	cct	gac	atg	atg	1023
Ile	Pro	Glu	Ile	Ala	Asn	Val	Pro	Ile	Pro	Val	Ser	Pro	Asp	Met	Met	
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atg	taa															1029
Met																
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<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic																
AE-I[8-321]C145A polypeptide																
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1				5					10					15		
Asp	Ile	Arg	Met	Pro	Gly	Val	Thr	Pro	Thr	Glu	Ser	Asp	Thr	Tyr	Leu	
			20					25					30			
Cys	Lys	Ser	Tyr	Arg	Leu	Pro	Val	Asp	Asp	Glu	Ala	Tyr	Val	Val	Asp	
		35					40					45				
Phe	Arg	Pro	His	Ala	Asn	Met	Asp	Thr	Ala	His	His	Met	Leu	Leu	Phe	
	50					55					60					
Gly	Cys	Asn	Ile	Pro	Ser	Ser	Thr	Asp	Asp	Tyr	Trp	Asp	Cys	Ser	Ala	
65				70					75					80		
Gly	Thr	Cys	Met	Asp	Lys	Ser	Ser	Ile	Met	Tyr	Ala	Trp	Ala	Lys	Asn	
			85						90					95		
Ala	Pro	Pro	Thr	Lys	Leu	Pro	Glu	Gly	Val	Gly	Phe	Arg	Val	Gly	Gly	
			100					105					110			
Lys	Ser	Gly	Ser	Arg	Tyr	Phe	Val	Leu	Gln	Val	His	Tyr	Gly	Asn	Val	
		115					120					125				
Lys	Ala	Phe	Gln	Asp	Lys	His	Lys	Asp	Ala	Thr	Gly	Val	Thr	Val	Arg	
	130					135					140					
Val	Thr	Pro	Glu	Lys	Gln	Pro	Gln	Ile	Ala	Gly	Ile	Tyr	Leu	Ser	Met	
145					150					155					160	
Ser	Val	Asp	Thr	Val	Ile	Pro	Pro	Gly	Glu	Glu	Ala	Val	Asn	Ser	Asp	
			165						170					175		
Ile	Ala	Cys	Leu	Tyr	Asn	Arg	Pro	Thr	Ile	His	Pro	Phe	Ala	Tyr	Arg	
		180						185					190			
Val	His	Thr	His	Gln	Leu	Gly	Gln	Val	Val	Ser	Gly	Phe	Arg	Val	Arg	
		195					200					205				
His	Gly	Lys	Trp	Ser	Leu	Ile	Gly	Arg	Gln	Ser	Pro	Gln	Leu	Pro	Gln	
	210					215					220					
Ala	Phe	Tyr	Pro	Val	Glu	His	Pro	Val	Glu	Ile	Ser	Pro	Gly	Asp	Ile	
225					230					235					240	
Ile	Ala	Thr	Arg	Cys	Leu	Phe	Thr	Gly	Lys	Gly	Arg	Thr	Ser	Ala	Thr	
			245						250					255		
Tyr	Ile	Gly	Gly	Thr	Ser	Asn	Asp	Glu	Met	Cys	Asn	Leu	Tyr	Ile	Met	
		260						265					270			
Tyr	Tyr	Met	Asp	Ala	Ala	His	Ala	Thr	Ser	Tyr	Met	Thr	Cys	Val	Gln	
		275					280						285			

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Thr Gly Glu Pro Lys Leu Phe Gln Asn Ile Pro Glu Ile Ala Asn Val	
290 295 300	
Pro Ile Pro Val Ser Pro Asp Met Met Met	
305 310	
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<211> LENGTH: 1050	
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<213> ORGANISM: Artificial Sequence	
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AE-I[1-321]C40A,C85A polynucleotide	
<220> FEATURE:	
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Ser Leu Ser Asn Asp Cys Leu Gly Thr	
1 5	
acg cgg ccc gtt atg tct cca ggc tca tca gat tat act cta gat atc	159
Thr Arg Pro Val Met Ser Pro Gly Ser Ser Asp Tyr Thr Leu Asp Ile	
10 15 20 25	
cgc atg cca gga gta act ccg aca gag tcc gac aca tat ttg gcc aag	207
Arg Met Pro Gly Val Thr Pro Thr Glu Ser Asp Thr Tyr Leu Ala Lys	
30 35 40	
tct tac cgg ctg cca gtg gat gat gaa gcc tat gta gtt gac ttc aga	255
Ser Tyr Arg Leu Pro Val Asp Asp Glu Ala Tyr Val Val Asp Phe Arg	
45 50 55	
cca cat gcc aat atg gat act gca cat cac atg ctt cta ttt gga tgc	303
Pro His Ala Asn Met Asp Thr Ala His His Met Leu Leu Phe Gly Cys	
60 65 70	
aat ata cct tct tcc act gat gat tac tgg gac gct agt gcg gga act	351
Asn Ile Pro Ser Ser Thr Asp Asp Tyr Trp Asp Ala Ser Ala Gly Thr	
75 80 85	
tgc atg gac aaa tcc agt ata atg tat gcc tgg gca aag aat gca cca	399
Cys Met Asp Lys Ser Ser Ile Met Tyr Ala Trp Ala Lys Asn Ala Pro	
90 95 100 105	
ccc acc aaa ctt cca gaa gga gtt ggc ttt cgt gtt gga ggg aaa tca	447
Pro Thr Lys Leu Pro Glu Gly Val Gly Phe Arg Val Gly Gly Lys Ser	
110 115 120	
ggc agt aga tat ttt gtg ctt caa gtt cac tat gga aat gtg aaa gca	495
Gly Ser Arg Tyr Phe Val Leu Gln Val His Tyr Gly Asn Val Lys Ala	
125 130 135	
ttc cag gat aaa cat aaa gat tgc acg ggg gtg aca gta cga gta aca	543
Phe Gln Asp Lys His Lys Asp Cys Thr Gly Val Thr Val Arg Val Thr	
140 145 150	
cct gaa aaa caa ccg caa att gca ggc att tat ctt tca atg tct gtg	591
Pro Glu Lys Gln Pro Gln Ile Ala Gly Ile Tyr Leu Ser Met Ser Val	
155 160 165	
gac act gtt att cca cct ggg gaa gag gca gtt aat tct gat atc gcc	639
Asp Thr Val Ile Pro Pro Gly Glu Glu Ala Val Asn Ser Asp Ile Ala	
170 175 180 185	
tgc ctc tac aac agg ccg aca ata cac cca ttt gcc tac aga gtc cac	687
Cys Leu Tyr Asn Arg Pro Thr Ile His Pro Phe Ala Tyr Arg Val His	
190 195 200	
act cat cag ttg ggg cag gtc gta agt gga ttt aga gtg aga cat ggc	735
Thr His Gln Leu Gly Gln Val Val Ser Gly Phe Arg Val Arg His Gly	
205 210 215	
aag tgg tct tta att ggt aga caa agc cca cag ctg cca cag gca ttt	783
Lys Trp Ser Leu Ile Gly Arg Gln Ser Pro Gln Leu Pro Gln Ala Phe	

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220	225	230	
tac cct gta gag cat cca gta gag att agc cct ggg gat att ata gca			831
Tyr Pro Val Glu His Pro Val Glu Ile Ser Pro Gly Asp Ile Ile Ala			
235	240	245	
acc agg tgt ctg ttc act ggt aaa ggc agg acg tca gca aca tat att			879
Thr Arg Cys Leu Phe Thr Gly Lys Gly Arg Thr Ser Ala Thr Tyr Ile			
250	255	260	265
ggt ggc aca tct aac gat gaa atg tgt aat tta tac atc atg tat tac			927
Gly Gly Thr Ser Asn Asp Glu Met Cys Asn Leu Tyr Ile Met Tyr Tyr			
270	275	280	
atg gat gcg gcc cat gct acg tca tac atg acc tgt gta cag acg ggt			975
Met Asp Ala Ala His Ala Thr Ser Tyr Met Thr Cys Val Gln Thr Gly			
285	290	295	
gaa cca aag tta ttt caa aac atc cct gag att gca aat gtt ccc att			1023
Glu Pro Lys Leu Phe Gln Asn Ile Pro Glu Ile Ala Asn Val Pro Ile			
300	305	310	
cct gta agc cct gac atg atg atg taa			1050
Pro Val Ser Pro Asp Met Met Met			
315	320		
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<211> LENGTH: 321			
<212> TYPE: PRT			
<213> ORGANISM: Artificial Sequence			
<220> FEATURE:			
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic			
AE-I[1-321]C40A,C85A polypeptide			
<400> SEQUENCE: 27			
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1	5	10	15
Gly Ser Ser Asp Tyr Thr Leu Asp Ile Arg Met Pro Gly Val Thr Pro			
20	25	30	
Thr Glu Ser Asp Thr Tyr Leu Ala Lys Ser Tyr Arg Leu Pro Val Asp			
35	40	45	
Asp Glu Ala Tyr Val Val Asp Phe Arg Pro His Ala Asn Met Asp Thr			
50	55	60	
Ala His His Met Leu Leu Phe Gly Cys Asn Ile Pro Ser Ser Thr Asp			
65	70	75	80
Asp Tyr Trp Asp Ala Ser Ala Gly Thr Cys Met Asp Lys Ser Ser Ile			
85	90	95	
Met Tyr Ala Trp Ala Lys Asn Ala Pro Pro Thr Lys Leu Pro Glu Gly			
100	105	110	
Val Gly Phe Arg Val Gly Gly Lys Ser Gly Ser Arg Tyr Phe Val Leu			
115	120	125	
Gln Val His Tyr Gly Asn Val Lys Ala Phe Gln Asp Lys His Lys Asp			
130	135	140	
Cys Thr Gly Val Thr Val Arg Val Thr Pro Glu Lys Gln Pro Gln Ile			
145	150	155	160
Ala Gly Ile Tyr Leu Ser Met Ser Val Asp Thr Val Ile Pro Pro Gly			
165	170	175	
Glu Glu Ala Val Asn Ser Asp Ile Ala Cys Leu Tyr Asn Arg Pro Thr			
180	185	190	
Ile His Pro Phe Ala Tyr Arg Val His Thr His Gln Leu Gly Gln Val			
195	200	205	
Val Ser Gly Phe Arg Val Arg His Gly Lys Trp Ser Leu Ile Gly Arg			
210	215	220	
Gln Ser Pro Gln Leu Pro Gln Ala Phe Tyr Pro Val Glu His Pro Val			

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225	230	235	240
Glu Ile Ser Pro Gly Asp Ile Ile Ala Thr Arg Cys Leu Phe Thr Gly	245	250	255
Lys Gly Arg Thr Ser Ala Thr Tyr Ile Gly Gly Thr Ser Asn Asp Glu	260	265	270
Met Cys Asn Leu Tyr Ile Met Tyr Tyr Met Asp Ala Ala His Ala Thr	275	280	285
Ser Tyr Met Thr Cys Val Gln Thr Gly Glu Pro Lys Leu Phe Gln Asn	290	295	300
Ile Pro Glu Ile Ala Asn Val Pro Ile Pro Val Ser Pro Asp Met Met	305	310	315
320			
Met			
<210> SEQ ID NO 28			
<211> LENGTH: 1050			
<212> TYPE: DNA			
<213> ORGANISM: Artificial Sequence			
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AE-I[1-321]C73A,C90A polynucleotide			
<220> FEATURE:			
<221> NAME/KEY: CDS			
<222> LOCATION: (85)..(1047)			
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tatttttcagg gcgccatgga tccg tca ctt tcc aat gac tgc ttg gga acc			111
Ser Leu Ser Asn Asp Cys Leu Gly Thr	1	5	
acg cgg ccc gtt atg tct cca ggc tca tca gat tat act cta gat atc			159
Thr Arg Pro Val Met Ser Pro Gly Ser Ser Asp Tyr Thr Leu Asp Ile	10	15	20
25			
cgc atg cca gga gta act ccg aca gag tcg gac aca tat ttg tgc aag			207
Arg Met Pro Gly Val Thr Pro Thr Glu Ser Asp Thr Tyr Leu Cys Lys	30	35	40
tct tac cgg ctg cca gtg gat gat gaa gcc tat gta gtt gac ttc aga			255
Ser Tyr Arg Leu Pro Val Asp Asp Glu Ala Tyr Val Val Asp Phe Arg	45	50	55
cca cat gcc aat atg gat act gca cat cac atg ctt cta ttt gga gcc			303
Pro His Ala Asn Met Asp Thr Ala His His Met Leu Leu Phe Gly Ala	60	65	70
aat ata cct tct tcc act gat gat tac tgg gac tgt agt gcg gga act			351
Asn Ile Pro Ser Ser Thr Asp Asp Tyr Trp Asp Cys Ser Ala Gly Thr	75	80	85
gcc atg gac aaa tcc agt ata atg tat gcc tgg gca aag aat gca cca			399
Ala Met Asp Lys Ser Ser Ile Met Tyr Ala Trp Ala Lys Asn Ala Pro	90	95	100
105			
ccc acc aaa ctt cca gaa gga gtt ggc ttt cgt gtt gga ggg aaa tca			447
Pro Thr Lys Leu Pro Glu Gly Val Gly Phe Arg Val Gly Gly Lys Ser	110	115	120
ggc agt aga tat ttt gtg ctt caa gtt cac tat gga aat gtg aaa gca			495
Gly Ser Arg Tyr Phe Val Leu Gln Val His Tyr Gly Asn Val Lys Ala	125	130	135
ttc cag gat aaa cat aaa gat tgc acg ggg gtg aca gta cga gta aca			543
Phe Gln Asp Lys His Lys Asp Cys Thr Gly Val Thr Val Arg Val Thr	140	145	150
cct gaa aaa caa ccg caa att gca ggc att tat ctt tca atg tct gtg			591
Pro Glu Lys Gln Pro Gln Ile Ala Gly Ile Tyr Leu Ser Met Ser Val	155	160	165
gac act gtt att cca cct ggg gaa gag gca gtt aat tct gat atc gcc			639

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Asp Thr Val Ile Pro Pro Gly Glu Glu Ala Val Asn Ser Asp Ile Ala	
170 175 180 185	
tgc ctc tac aac agg ccg aca ata cac cca ttt gcc tac aga gtc cac	687
Cys Leu Tyr Asn Arg Pro Thr Ile His Pro Phe Ala Tyr Arg Val His	
190 195 200	
act cat cag ttg ggg cag gtc gta agt gga ttt aga gtg aga cat ggc	735
Thr His Gln Leu Gly Gln Val Val Ser Gly Phe Arg Val Arg His Gly	
205 210 215	
aag tgg tct tta att ggt aga caa agc cca cag ctg cca cag gca ttt	783
Lys Trp Ser Leu Ile Gly Arg Gln Ser Pro Gln Leu Pro Gln Ala Phe	
220 225 230	
tac cct gta gag cat cca gta gag att agc cct ggg gat att ata gca	831
Tyr Pro Val Glu His Pro Val Glu Ile Ser Pro Gly Asp Ile Ile Ala	
235 240 245	
acc agg tgt ctg ttc act ggt aaa ggc agg acg tca gca aca tat att	879
Thr Arg Cys Leu Phe Thr Gly Lys Gly Arg Thr Ser Ala Thr Tyr Ile	
250 255 260 265	
ggt ggc aca tct aac gat gaa atg tgt aat tta tac atc atg tat tac	927
Gly Gly Thr Ser Asn Asp Glu Met Cys Asn Leu Tyr Ile Met Tyr Tyr	
270 275 280	
atg gat gcg gcc cat gct acg tca tac atg acc tgt gta cag acg ggt	975
Met Asp Ala Ala His Ala Thr Ser Tyr Met Thr Cys Val Gln Thr Gly	
285 290 295	
gaa cca aag tta ttt caa aac atc cct gag att gca aat gtt ccc att	1023
Glu Pro Lys Leu Phe Gln Asn Ile Pro Glu Ile Ala Asn Val Pro Ile	
300 305 310	
cct gta agc cct gac atg atg atg taa	1050
Pro Val Ser Pro Asp Met Met Met	
315 320	
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<211> LENGTH: 321	
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<400> SEQUENCE: 29	
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20 25 30	
Thr Glu Ser Asp Thr Tyr Leu Cys Lys Ser Tyr Arg Leu Pro Val Asp	
35 40 45	
Asp Glu Ala Tyr Val Val Asp Phe Arg Pro His Ala Asn Met Asp Thr	
50 55 60	
Ala His His Met Leu Leu Phe Gly Ala Asn Ile Pro Ser Ser Thr Asp	
65 70 75 80	
Asp Tyr Trp Asp Cys Ser Ala Gly Thr Ala Met Asp Lys Ser Ser Ile	
85 90 95	
Met Tyr Ala Trp Ala Lys Asn Ala Pro Pro Thr Lys Leu Pro Glu Gly	
100 105 110	
Val Gly Phe Arg Val Gly Gly Lys Ser Gly Ser Arg Tyr Phe Val Leu	
115 120 125	
Gln Val His Tyr Gly Asn Val Lys Ala Phe Gln Asp Lys His Lys Asp	
130 135 140	
Cys Thr Gly Val Thr Val Arg Val Thr Pro Glu Lys Gln Pro Gln Ile	
145 150 155 160	

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Ala Gly Ile Tyr	Leu Ser Met Ser Val	Asp Thr Val Ile Pro Pro Gly	
	165	170	175
Glu Glu Ala Val	Asn Ser Asp Ile Ala Cys Leu Tyr	Asn Arg Pro Thr	
	180	185	190
Ile His Pro Phe	Ala Tyr Arg Val His Thr His Gln	Leu Gly Gln Val	
	195	200	205
Val Ser Gly Phe	Arg Val Arg His Gly Lys Trp Ser	Leu Ile Gly Arg	
	210	215	220
Gln Ser Pro Gln	Leu Pro Gln Ala Phe Tyr Pro Val	Glu His Pro Val	
225	230	235	240
Glu Ile Ser Pro	Gly Asp Ile Ile Ala Thr Arg Cys Leu	Phe Thr Gly	
	245	250	255
Lys Gly Arg Thr	Ser Ala Thr Tyr Ile Gly Gly Thr Ser	Asn Asp Glu	
	260	265	270
Met Cys Asn Leu	Tyr Ile Met Tyr Tyr Met Asp Ala Ala	His Ala Thr	
	275	280	285
Ser Tyr Met Thr	Cys Val Gln Thr Gly Glu Pro Lys Leu	Phe Gln Asn	
	290	295	300
Ile Pro Glu Ile	Ala Asn Val Pro Ile Pro Val Ser Pro	Asp Met Met	
305	310	315	320
Met			
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<211> LENGTH: 1050			
<212> TYPE: DNA			
<213> ORGANISM: Artificial Sequence			
<220> FEATURE:			
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic			
AE-I[1-321]C186A,C293A polynucleotide			
<220> FEATURE:			
<221> NAME/KEY: CDS			
<222> LOCATION: (85)..(1047)			
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tatatttcagg gcgccatgga tccg tca ctt tcc aat gac tgc ttg gga acc			111
	Ser Leu Ser Asn Asp Cys Leu Gly Thr		
	1 5		
acg cgg ccc gtt atg tct cca ggc tca tca gat tat act cta gat atc			159
Thr Arg Pro Val Met Ser Pro Gly Ser Ser Asp Tyr Thr Leu Asp Ile			
10 15 20 25			
cgc atg cca gga gta act ccg aca gag tcg gac aca tat ttg tgc aag			207
Arg Met Pro Gly Val Thr Pro Thr Glu Ser Asp Thr Tyr Leu Cys Lys			
30 35 40			
tct tac cgg ctg cca gtg gat gat gaa gcc tat gta gtt gac ttc aga			255
Ser Tyr Arg Leu Pro Val Asp Asp Glu Ala Tyr Val Val Asp Phe Arg			
45 50 55			
cca cat gcc aat atg gat act gca cat cac atg ctt cta ttt gga tgc			303
Pro His Ala Asn Met Asp Thr Ala His His Met Leu Leu Phe Gly Cys			
60 65 70			
aat ata cct tct tcc act gat gat tac tgg gac tgt agt gcg gga act			351
Asn Ile Pro Ser Ser Thr Asp Asp Tyr Trp Asp Cys Ser Ala Gly Thr			
75 80 85			
tgc atg gac aaa tcc agt ata atg tat gcc tgg gca aag aat gca cca			399
Cys Met Asp Lys Ser Ser Ile Met Tyr Ala Trp Ala Lys Asn Ala Pro			
90 95 100 105			
ccc acc aaa ctt cca gaa gga gtt ggc ttt cgt gtt gga ggg aaa tca			447
Pro Thr Lys Leu Pro Glu Gly Val Gly Phe Arg Val Gly Gly Lys Ser			
110 115 120			

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ggc agt aga tat ttt gtg ctt caa gtt cac tat gga aat gtg aaa gca	495
Gly Ser Arg Tyr Phe Val Leu Gln Val His Tyr Gly Asn Val Lys Ala	
125 130 135	
ttc cag gat aaa cat aaa gat tgc acg ggg gtg aca gta cga gta aca	543
Phe Gln Asp Lys His Lys Asp Cys Thr Gly Val Thr Val Arg Val Thr	
140 145 150	
cct gaa aaa caa ccg caa att gca ggc att tat ctt tca atg tct gtg	591
Pro Glu Lys Gln Pro Gln Ile Ala Gly Ile Tyr Leu Ser Met Ser Val	
155 160 165	
gac act gtt att cca cct ggg gaa gag gca gtt aat tct gat atc gcc	639
Asp Thr Val Ile Pro Pro Gly Glu Glu Ala Val Asn Ser Asp Ile Ala	
170 175 180 185	
gcc ctc tac aac agg ccg aca ata cac cca ttt gcc tac aga gtc cac	687
Ala Leu Tyr Asn Arg Pro Thr Ile His Pro Phe Ala Tyr Arg Val His	
190 195 200	
act cat cag ttg ggg cag gtc gta agt gga ttt aga gtg aga cat ggc	735
Thr His Gln Leu Gly Gln Val Val Ser Gly Phe Arg Val Arg His Gly	
205 210 215	
aag tgg tct tta att ggt aga caa agc cca cag ctg cca cag gca ttt	783
Lys Trp Ser Leu Ile Gly Arg Gln Ser Pro Gln Leu Pro Gln Ala Phe	
220 225 230	
tac cct gta gag cat cca gta gag att agc cct ggg gat att ata gca	831
Tyr Pro Val Glu His Pro Val Glu Ile Ser Pro Gly Asp Ile Ile Ala	
235 240 245	
acc agg tgt ctg ttc act ggt aaa ggc agg acg tca gca aca tat att	879
Thr Arg Cys Leu Phe Thr Gly Lys Gly Arg Thr Ser Ala Thr Tyr Ile	
250 255 260 265	
ggt ggc aca tct aac gat gaa atg tgt aat tta tac atc atg tat tac	927
Gly Gly Thr Ser Asn Asp Glu Met Cys Asn Leu Tyr Ile Met Tyr Tyr	
270 275 280	
atg gat gcg gcc cat gct acg tca tac atg acc gct gta cag acg ggt	975
Met Asp Ala Ala His Ala Thr Ser Tyr Met Thr Ala Val Gln Thr Gly	
285 290 295	
gaa cca aag tta ttt caa aac atc cct gag att gca aat gtt ccc att	1023
Glu Pro Lys Leu Phe Gln Asn Ile Pro Glu Ile Ala Asn Val Pro Ile	
300 305 310	
cct gta agc cct gac atg atg atg taa	1050
Pro Val Ser Pro Asp Met Met Met	
315 320	
<210> SEQ ID NO 31	
<211> LENGTH: 321	
<212> TYPE: PRT	
<213> ORGANISM: Artificial Sequence	
<220> FEATURE:	
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic	
AE-I[1-321]C186A,C293A polypeptide	
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Ser Leu Ser Asn Asp Cys Leu Gly Thr Thr Arg Pro Val Met Ser Pro	
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Gly Ser Ser Asp Tyr Thr Leu Asp Ile Arg Met Pro Gly Val Thr Pro	
20 25 30	
Thr Glu Ser Asp Thr Tyr Leu Cys Lys Ser Tyr Arg Leu Pro Val Asp	
35 40 45	
Asp Glu Ala Tyr Val Val Asp Phe Arg Pro His Ala Asn Met Asp Thr	
50 55 60	
Ala His His Met Leu Leu Phe Gly Cys Asn Ile Pro Ser Ser Thr Asp	
65 70 75 80	
Asp Tyr Trp Asp Cys Ser Ala Gly Thr Cys Met Asp Lys Ser Ser Ile	
85 90 95	

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Met	Tyr	Ala	Trp	Ala	Lys	Asn	Ala	Pro	Pro	Thr	Lys	Leu	Pro	Glu	Gly	
			100					105					110			
Val	Gly	Phe	Arg	Val	Gly	Gly	Lys	Ser	Gly	Ser	Arg	Tyr	Phe	Val	Leu	
		115					120					125				
Gln	Val	His	Tyr	Gly	Asn	Val	Lys	Ala	Phe	Gln	Asp	Lys	His	Lys	Asp	
		130					135				140					
Cys	Thr	Gly	Val	Thr	Val	Arg	Val	Thr	Pro	Glu	Lys	Gln	Pro	Gln	Ile	
145						150				155					160	
Ala	Gly	Ile	Tyr	Leu	Ser	Met	Ser	Val	Asp	Thr	Val	Ile	Pro	Pro	Gly	
				165					170					175		
Glu	Glu	Ala	Val	Asn	Ser	Asp	Ile	Ala	Ala	Leu	Tyr	Asn	Arg	Pro	Thr	
			180					185					190			
Ile	His	Pro	Phe	Ala	Tyr	Arg	Val	His	Thr	His	Gln	Leu	Gly	Gln	Val	
		195					200					205				
Val	Ser	Gly	Phe	Arg	Val	Arg	His	Gly	Lys	Trp	Ser	Leu	Ile	Gly	Arg	
		210					215				220					
Gln	Ser	Pro	Gln	Leu	Pro	Gln	Ala	Phe	Tyr	Pro	Val	Glu	His	Pro	Val	
225						230				235					240	
Glu	Ile	Ser	Pro	Gly	Asp	Ile	Ile	Ala	Thr	Arg	Cys	Leu	Phe	Thr	Gly	
				245					250					255		
Lys	Gly	Arg	Thr	Ser	Ala	Thr	Tyr	Ile	Gly	Gly	Thr	Ser	Asn	Asp	Glu	
			260					265					270			
Met	Cys	Asn	Leu	Tyr	Ile	Met	Tyr	Tyr	Met	Asp	Ala	Ala	His	Ala	Thr	
		275					280					285				
Ser	Tyr	Met	Thr	Ala	Val	Gln	Thr	Gly	Glu	Pro	Lys	Leu	Phe	Gln	Asn	
		290				295					300					
Ile	Pro	Glu	Ile	Ala	Asn	Val	Pro	Ile	Pro	Val	Ser	Pro	Asp	Met	Met	
305					310					315					320	
Met																

<210> SEQ ID NO 32
<211> LENGTH: 1050
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
AE-I[1-321]C252A,C274A polynucleotide
<220> FEATURE:
<221> NAME/KEY: CDS
<222> LOCATION: (85)..(1047)

<400> SEQUENCE: 32

atgtcgtact accatcacca tcaccatcac gattacgata tcccaacgac cgaaaacctg 60

tatttttcagg gcgccatgga tccg tca ctt tcc aat gac tgc ttg gga acc 111
Ser Leu Ser Asn Asp Cys Leu Gly Thr
1 5

acg cgg ccc gtt atg tct cca ggc tca tca gat tat act cta gat atc 159
Thr Arg Pro Val Met Ser Pro Gly Ser Ser Asp Tyr Thr Leu Asp Ile
10 15 20 25

cgc atg cca gga gta act ccg aca gag tcg gac aca tat ttg tgc aag 207
Arg Met Pro Gly Val Thr Pro Thr Glu Ser Asp Thr Tyr Leu Cys Lys
30 35 40

tct tac cgg ctg cca gtg gat gat gaa gcc tat gta gtt gac ttc aga 255
Ser Tyr Arg Leu Pro Val Asp Asp Glu Ala Tyr Val Val Asp Phe Arg
45 50 55

cca cat gcc aat atg gat act gca cat cac atg ctt cta ttt gga tgc 303
Pro His Ala Asn Met Asp Thr Ala His His Met Leu Leu Phe Gly Cys
60 65 70

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aat ata cct tct tcc act gat gat tac tgg gac tgt agt gcg gga act	351
Asn Ile Pro Ser Ser Thr Asp Asp Tyr Trp Asp Cys Ser Ala Gly Thr	
75 80 85	
tgc atg gac aaa tcc agt ata atg tat gcc tgg gca aag aat gca cca	399
Cys Met Asp Lys Ser Ser Ile Met Tyr Ala Trp Ala Lys Asn Ala Pro	
90 95 100 105	
ccc acc aaa ctt cca gaa gga gtt ggc ttt cgt gtt gga ggg aaa tca	447
Pro Thr Lys Leu Pro Glu Gly Val Gly Phe Arg Val Gly Gly Lys Ser	
110 115 120	
ggc agt aga tat ttt gtg ctt caa gtt cac tat gga aat gtg aaa gca	495
Gly Ser Arg Tyr Phe Val Leu Gln Val His Tyr Gly Asn Val Lys Ala	
125 130 135	
ttc cag gat aaa cat aaa gat tgc acg ggg gtg aca gta cga gta aca	543
Phe Gln Asp Lys His Lys Asp Cys Thr Gly Val Thr Val Arg Val Thr	
140 145 150	
cct gaa aaa caa ccg caa att gca ggc att tat ctt tca atg tct gtg	591
Pro Glu Lys Gln Pro Gln Ile Ala Gly Ile Tyr Leu Ser Met Ser Val	
155 160 165	
gac act gtt att cca cct ggg gaa gag gca gtt aat tct gat atc gcc	639
Asp Thr Val Ile Pro Pro Gly Glu Glu Ala Val Asn Ser Asp Ile Ala	
170 175 180 185	
tgc ctc tac aac agg ccg aca ata cac cca ttt gcc tac aga gtc cac	687
Cys Leu Tyr Asn Arg Pro Thr Ile His Pro Phe Ala Tyr Arg Val His	
190 195 200	
act cat cag ttg ggg cag gtc gta agt gga ttt aga gtg aga cat ggc	735
Thr His Gln Leu Gly Gln Val Val Ser Gly Phe Arg Val Arg His Gly	
205 210 215	
aag tgg tct tta att ggt aga caa agc cca cag ctg cca cag gca ttt	783
Lys Trp Ser Leu Ile Gly Arg Gln Ser Pro Gln Leu Pro Gln Ala Phe	
220 225 230	
tac cct gta gag cat cca gta gag att agc cct ggg gat att ata gca	831
Tyr Pro Val Glu His Pro Val Glu Ile Ser Pro Gly Asp Ile Ile Ala	
235 240 245	
acc agg gct ctg ttc act ggt aaa ggc agg acg tca gca aca tat att	879
Thr Arg Ala Leu Phe Thr Gly Lys Gly Arg Thr Ser Ala Thr Tyr Ile	
250 255 260 265	
ggt ggc aca tct aac gat gaa atg gct aat tta tac atc atg tat tac	927
Gly Gly Thr Ser Asn Asp Glu Met Ala Asn Leu Tyr Ile Met Tyr Tyr	
270 275 280	
atg gat gcg gcc cat gct acg tca tac atg acc tgt gta cag acg ggt	975
Met Asp Ala Ala His Ala Thr Ser Tyr Met Thr Cys Val Gln Thr Gly	
285 290 295	
gaa cca aag tta ttt caa aac atc cct gag att gca aat gtt ccc att	1023
Glu Pro Lys Leu Phe Gln Asn Ile Pro Glu Ile Ala Asn Val Pro Ile	
300 305 310	
cct gta agc cct gac atg atg atg taa	1050
Pro Val Ser Pro Asp Met Met Met	
315 320	
<210> SEQ ID NO 33	
<211> LENGTH: 321	
<212> TYPE: PRT	
<213> ORGANISM: Artificial Sequence	
<220> FEATURE:	
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic	
AE-I[1-321]C252A,C274A polypeptide	
<400> SEQUENCE: 33	
Ser Leu Ser Asn Asp Cys Leu Gly Thr Thr Arg Pro Val Met Ser Pro	
1 5 10 15	
Gly Ser Ser Asp Tyr Thr Leu Asp Ile Arg Met Pro Gly Val Thr Pro	

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20					25					30					
Thr	Glu	Ser	Asp	Thr	Tyr	Leu	Cys	Lys	Ser	Tyr	Arg	Leu	Pro	Val	Asp
		35					40					45			
Asp	Glu	Ala	Tyr	Val	Val	Asp	Phe	Arg	Pro	His	Ala	Asn	Met	Asp	Thr
	50					55					60				
Ala	His	His	Met	Leu	Leu	Phe	Gly	Cys	Asn	Ile	Pro	Ser	Ser	Thr	Asp
65					70					75					80
Asp	Tyr	Trp	Asp	Cys	Ser	Ala	Gly	Thr	Cys	Met	Asp	Lys	Ser	Ser	Ile
				85					90					95	
Met	Tyr	Ala	Trp	Ala	Lys	Asn	Ala	Pro	Pro	Thr	Lys	Leu	Pro	Glu	Gly
			100					105					110		
Val	Gly	Phe	Arg	Val	Gly	Gly	Lys	Ser	Gly	Ser	Arg	Tyr	Phe	Val	Leu
		115					120					125			
Gln	Val	His	Tyr	Gly	Asn	Val	Lys	Ala	Phe	Gln	Asp	Lys	His	Lys	Asp
	130					135					140				
Cys	Thr	Gly	Val	Thr	Val	Arg	Val	Thr	Pro	Glu	Lys	Gln	Pro	Gln	Ile
145					150					155					160
Ala	Gly	Ile	Tyr	Leu	Ser	Met	Ser	Val	Asp	Thr	Val	Ile	Pro	Pro	Gly
				165					170					175	
Glu	Glu	Ala	Val	Asn	Ser	Asp	Ile	Ala	Cys	Leu	Tyr	Asn	Arg	Pro	Thr
			180					185					190		
Ile	His	Pro	Phe	Ala	Tyr	Arg	Val	His	Thr	His	Gln	Leu	Gly	Gln	Val
		195					200					205			
Val	Ser	Gly	Phe	Arg	Val	Arg	His	Gly	Lys	Trp	Ser	Leu	Ile	Gly	Arg
	210					215					220				
Gln	Ser	Pro	Gln	Leu	Pro	Gln	Ala	Phe	Tyr	Pro	Val	Glu	His	Pro	Val
225					230					235					240
Glu	Ile	Ser	Pro	Gly	Asp	Ile	Ile	Ala	Thr	Arg	Ala	Leu	Phe	Thr	Gly
				245					250					255	
Lys	Gly	Arg	Thr	Ser	Ala	Thr	Tyr	Ile	Gly	Gly	Thr	Ser	Asn	Asp	Glu
			260					265					270		
Met	Ala	Asn	Leu	Tyr	Ile	Met	Tyr	Tyr	Met	Asp	Ala	Ala	His	Ala	Thr
		275					280					285			
Ser	Tyr	Met	Thr	Cys	Val	Gln	Thr	Gly	Glu	Pro	Lys	Leu	Phe	Gln	Asn
	290					295					300				
Ile	Pro	Glu	Ile	Ala	Asn	Val	Pro	Ile	Pro	Val	Ser	Pro	Asp	Met	Met
305					310					315				320	
Met															
<210> SEQ ID NO 34															
<211> LENGTH: 1029															
<212> TYPE: DNA															
<213> ORGANISM: Artificial Sequence															
<220> FEATURE:															
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic															
AE-I[8-321]C145A,C40A,C85A polynucleotide															
<220> FEATURE:															
<221> NAME/KEY: CDS															
<222> LOCATION: (85)..(1026)															
<400> SEQUENCE: 34															
atgtcgtact accatcacca tcaccatcac gattacgata tcccaacgac cgaaaacctg														60	
tatttttcagg gcgccatgga tccg gga acc acg cgg ccc gtt atg tct cca														111	
Gly Thr Thr Arg Pro Val Met Ser Pro															
1 5															
ggc tca tca gat tat act cta gat atc cgc atg cca gga gta act ccg														159	
Gly Ser Ser Asp Tyr Thr Leu Asp Ile Arg Met Pro Gly Val Thr Pro															

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10	15	20	25	
aca gag tcg gac	aca tat ttg gcc	aag tct tac cgg	ctg cca gtg gat	207
Thr Glu Ser Asp	Thr Tyr Leu Ala	Lys Ser Tyr Arg	Leu Pro Val Asp	
	30	35	40	
gat gaa gcc tat	gta gtt gac ttc	aga cca cat gcc	aat atg gat act	255
Asp Glu Ala Tyr	Val Val Asp Phe	Arg Pro His Ala	Asn Met Asp Thr	
	45	50	55	
gca cat cac atg	ctt cta ttt gga	tgc aat ata cct	tct tcc act gat	303
Ala His His Met	Leu Leu Phe Gly	Cys Asn Ile Pro	Ser Ser Thr Asp	
	60	65	70	
gat tac tgg gac	gct agt gcg gga	act tgc atg gac	aaa tcc agt ata	351
Asp Tyr Trp Asp	Ala Ser Ala Gly	Thr Cys Met Asp	Lys Ser Ser Ile	
	75	80	85	
atg tat gcc tgg	gca aag aat gca	cca ccc acc aaa	ctt cca gaa gga	399
Met Tyr Ala Trp	Ala Lys Asn Ala	Pro Pro Thr Lys	Leu Pro Glu Gly	
	95	100	105	
gtt ggc ttt cgt	gtt gga ggg aaa	tca ggc agt aga	tat ttt gtg ctt	447
Val Gly Phe Arg	Val Gly Gly Lys	Ser Gly Ser Arg	Tyr Phe Val Leu	
	110	115	120	
caa gtt cac tat	gga aat gtg aaa	gca ttc cag gat	aaa cat aaa gat	495
Gln Val His Tyr	Gly Asn Val Lys	Ala Phe Gln Asp	Lys His Lys Asp	
	125	130	135	
gcc acg ggg gtg	aca gta cga gta	aca cct gaa aaa	caa ccg caa att	543
Ala Thr Gly Val	Thr Val Arg Val	Thr Pro Glu Lys	Gln Pro Gln Ile	
	140	145	150	
gca ggc att tat	ctt tca atg tct	gtg gac act gtt	att cca cct ggg	591
Ala Gly Ile Tyr	Leu Ser Met Ser	Val Asp Thr Val	Ile Pro Pro Gly	
	155	160	165	
gaa gag gca gtt	aat tct gat atc	gcc tgc ctc tac	aac agg ccg aca	639
Glu Glu Ala Val	Asn Ser Asp Ile	Ala Cys Leu Tyr	Asn Arg Pro Thr	
	175	180	185	
ata cac cca ttt	gcc tac aga gtc	cac act cat cag	ttg ggg cag gtc	687
Ile His Pro Phe	Ala Tyr Arg Val	His Thr His Gln	Leu Gly Gln Val	
	190	195	200	
gta agt gga ttt	aga gtg aga cat	ggc aag tgg tct	tta att ggt aga	735
Val Ser Gly Phe	Arg Val Arg His	Gly Lys Trp Ser	Leu Ile Gly Arg	
	205	210	215	
caa agc cca cag	ctg cca cag gca	ttt tac cct gta	gag cat cca gta	783
Gln Ser Pro Gln	Leu Pro Gln Ala	Phe Tyr Pro Val	Glu His Pro Val	
	220	225	230	
gag att agc cct	ggg gat att ata	gca acc agg tgt	ctg ttc act ggt	831
Glu Ile Ser Pro	Gly Asp Ile Ile	Ala Thr Arg Cys	Leu Phe Thr Gly	
	235	240	245	
aaa ggc agg acg	tca gca aca tat	att ggt ggc aca	tct aac gat gaa	879
Lys Gly Arg Thr	Ser Ala Thr Tyr	Ile Gly Gly Thr	Ser Asn Asp Glu	
	255	260	265	
atg tgt aat tta	tac atc atg tat	tac atg gat gcg	gcc cat gct acg	927
Met Cys Asn Leu	Tyr Ile Met Tyr	Tyr Met Asp Ala	Ala His Ala Thr	
	270	275	280	
tca tac atg acc	tgt gta cag acg	ggt gaa cca aag	tta ttt caa aac	975
Ser Tyr Met Thr	Cys Val Gln Thr	Gly Glu Pro Lys	Leu Phe Gln Asn	
	285	290	295	
atc cct gag att	gca aat gtt ccc	att cct gta agc	cct gac atg atg	1023
Ile Pro Glu Ile	Ala Asn Val Pro	Ile Pro Val Ser	Pro Asp Met Met	
	300	305	310	
atg taa				1029
Met				

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<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
AE-I[8-321]C145A,C40A,C85A polypeptide

<400> SEQUENCE: 35

Gly Thr Thr Arg Pro Val Met Ser Pro Gly Ser Ser Asp Tyr Thr Leu
1 5 10 15
Asp Ile Arg Met Pro Gly Val Thr Pro Thr Glu Ser Asp Thr Tyr Leu
20 25 30
Ala Lys Ser Tyr Arg Leu Pro Val Asp Asp Glu Ala Tyr Val Val Asp
35 40 45
Phe Arg Pro His Ala Asn Met Asp Thr Ala His His Met Leu Leu Phe
50 55 60
Gly Cys Asn Ile Pro Ser Ser Thr Asp Asp Tyr Trp Asp Ala Ser Ala
65 70 75 80
Gly Thr Cys Met Asp Lys Ser Ser Ile Met Tyr Ala Trp Ala Lys Asn
85 90 95
Ala Pro Pro Thr Lys Leu Pro Glu Gly Val Gly Phe Arg Val Gly Gly
100 105 110
Lys Ser Gly Ser Arg Tyr Phe Val Leu Gln Val His Tyr Gly Asn Val
115 120 125
Lys Ala Phe Gln Asp Lys His Lys Asp Ala Thr Gly Val Thr Val Arg
130 135 140
Val Thr Pro Glu Lys Gln Pro Gln Ile Ala Gly Ile Tyr Leu Ser Met
145 150 155 160
Ser Val Asp Thr Val Ile Pro Pro Gly Glu Glu Ala Val Asn Ser Asp
165 170 175
Ile Ala Cys Leu Tyr Asn Arg Pro Thr Ile His Pro Phe Ala Tyr Arg
180 185 190
Val His Thr His Gln Leu Gly Gln Val Val Ser Gly Phe Arg Val Arg
195 200 205
His Gly Lys Trp Ser Leu Ile Gly Arg Gln Ser Pro Gln Leu Pro Gln
210 215 220
Ala Phe Tyr Pro Val Glu His Pro Val Glu Ile Ser Pro Gly Asp Ile
225 230 235 240
Ile Ala Thr Arg Cys Leu Phe Thr Gly Lys Gly Arg Thr Ser Ala Thr
245 250 255
Tyr Ile Gly Gly Thr Ser Asn Asp Glu Met Cys Asn Leu Tyr Ile Met
260 265 270
Tyr Tyr Met Asp Ala Ala His Ala Thr Ser Tyr Met Thr Cys Val Gln
275 280 285
Thr Gly Glu Pro Lys Leu Phe Gln Asn Ile Pro Glu Ile Ala Asn Val
290 295 300
Pro Ile Pro Val Ser Pro Asp Met Met Met
305 310

<210> SEQ ID NO 36
<211> LENGTH: 1050
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
AE-I[1-321]C40A,C85A,C252A,C274A polynucleotide
<220> FEATURE:
<221> NAME/KEY: CDS
<222> LOCATION: (85)..(1047)

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<400> SEQUENCE: 36		
atgtcgtact accatcacca tcaccatcac gattacgata tcccaacgac cgaaaacctg	60	
tatttttcagg gcgccatgga tccg tca ctt tcc aat gac tgc ttg gga acc	111	
Ser Leu Ser Asn Asp Cys Leu Gly Thr		
1 5		
acg cgg ccc gtt atg tct cca ggc tca tca gat tat act cta gat atc	159	
Thr Arg Pro Val Met Ser Pro Gly Ser Ser Asp Tyr Thr Leu Asp Ile		
10 15 20 25		
cgc atg cca gga gta act ccg aca gag tcg gac aca tat ttg gcc aag	207	
Arg Met Pro Gly Val Thr Pro Thr Glu Ser Asp Thr Tyr Leu Ala Lys		
30 35 40		
tct tac cgg ctg cca gtg gat gat gaa gcc tat gta gtt gac ttc aga	255	
Ser Tyr Arg Leu Pro Val Asp Asp Glu Ala Tyr Val Val Asp Phe Arg		
45 50 55		
cca cat gcc aat atg gat act gca cat cac atg ctt cta ttt gga tgc	303	
Pro His Ala Asn Met Asp Thr Ala His His Met Leu Leu Phe Gly Cys		
60 65 70		
aat ata cct tct tcc act gat gat tac tgg gac gct agt gcg gga act	351	
Asn Ile Pro Ser Ser Thr Asp Asp Tyr Trp Asp Ala Ser Ala Gly Thr		
75 80 85		
tgc atg gac aaa tcc agt ata atg tat gcc tgg gca aag aat gca cca	399	
Cys Met Asp Lys Ser Ser Ile Met Tyr Ala Trp Ala Lys Asn Ala Pro		
90 95 100 105		
ccc acc aaa ctt cca gaa gga gtt ggc ttt cgt gtt gga ggg aaa tca	447	
Pro Thr Lys Leu Pro Glu Gly Val Gly Phe Arg Val Gly Gly Lys Ser		
110 115 120		
ggc agt aga tat ttt gtg ctt caa gtt cac tat gga aat gtg aaa gca	495	
Gly Ser Arg Tyr Phe Val Leu Gln Val His Tyr Gly Asn Val Lys Ala		
125 130 135		
ttc cag gat aaa cat aaa gat tgc acg ggg gtg aca gta cga gta aca	543	
Phe Gln Asp Lys His Lys Asp Cys Thr Gly Val Thr Val Arg Val Thr		
140 145 150		
cct gaa aaa caa ccg caa att gca ggc att tat ctt tca atg tct gtg	591	
Pro Glu Lys Gln Pro Gln Ile Ala Gly Ile Tyr Leu Ser Met Ser Val		
155 160 165		
gac act gtt att cca cct ggg gaa gag gca gtt aat tct gat atc gcc	639	
Asp Thr Val Ile Pro Pro Gly Glu Glu Ala Val Asn Ser Asp Ile Ala		
170 175 180 185		
tgc ctc tac aac agg ccg aca ata cac cca ttt gcc tac aga gtc cac	687	
Cys Leu Tyr Asn Arg Pro Thr Ile His Pro Phe Ala Tyr Arg Val His		
190 195 200		
act cat cag ttg ggg cag gtc gta agt gga ttt aga gtg aga cat ggc	735	
Thr His Gln Leu Gly Gln Val Val Ser Gly Phe Arg Val Arg His Gly		
205 210 215		
aag tgg tct tta att ggt aga caa agc cca cag ctg cca cag gca ttt	783	
Lys Trp Ser Leu Ile Gly Arg Gln Ser Pro Gln Leu Pro Gln Ala Phe		
220 225 230		
tac cct gta gag cat cca gta gag att agc cct ggg gat att ata gca	831	
Tyr Pro Val Glu His Pro Val Glu Ile Ser Pro Gly Asp Ile Ile Ala		
235 240 245		
acc agg gct ctg ttc act ggt aaa ggc agg acg tca gca aca tat att	879	
Thr Arg Ala Leu Phe Thr Gly Lys Gly Arg Thr Ser Ala Thr Tyr Ile		
250 255 260 265		
ggt ggc aca tct aac gat gaa atg gct aat tta tac atc atg tat tac	927	
Gly Gly Thr Ser Asn Asp Glu Met Ala Asn Leu Tyr Ile Met Tyr Tyr		
270 275 280		
atg gat gcg gcc cat gct acg tca tac atg acc tgt gta cag acg ggt	975	
Met Asp Ala Ala His Ala Thr Ser Tyr Met Thr Cys Val Gln Thr Gly		
285 290 295		

Met

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<210> SEQ ID NO 38
<211> LENGTH: 6
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
6xHis tag

<400> SEQUENCE: 38

His His His His His His
1 5

<210> SEQ ID NO 39
<211> LENGTH: 400
<212> TYPE: PRT
<213> ORGANISM: Xenopus leavis

<400> SEQUENCE: 39

Met Ala Ser Leu Ser Ser Ser Phe Leu Val Leu Phe Leu Leu Phe Gln
1 5 10 15
Asn Ser Cys Tyr Cys Phe Arg Ser Pro Leu Ser Val Phe Lys Arg Tyr
20 25 30
Glu Glu Ser Thr Arg Ser Leu Ser Asn Asp Cys Leu Gly Thr Thr Arg
35 40 45
Pro Val Met Ser Pro Gly Ser Ser Asp Tyr Thr Leu Asp Ile Arg Met
50 55 60
Pro Gly Val Thr Pro Thr Glu Ser Asp Thr Tyr Leu Cys Lys Ser Tyr
65 70 75 80
Arg Leu Pro Val Asp Asp Glu Ala Tyr Val Val Asp Phe Arg Pro His
85 90 95
Ala Asn Met Asp Thr Ala His His Met Leu Leu Phe Gly Cys Asn Ile
100 105 110
Pro Ser Ser Thr Asp Asp Tyr Trp Asp Cys Ser Ala Gly Thr Cys Met
115 120 125
Asp Lys Ser Ser Ile Met Tyr Ala Trp Ala Lys Asn Ala Pro Pro Thr
130 135 140
Lys Leu Pro Glu Gly Val Gly Phe Arg Val Gly Gly Lys Ser Gly Ser
145 150 155 160
Arg Tyr Phe Val Leu Gln Val His Tyr Gly Asn Val Lys Ala Phe Gln
165 170 175
Asp Lys His Lys Asp Cys Thr Gly Val Thr Val Arg Val Thr Pro Glu
180 185 190
Lys Gln Pro Gln Ile Ala Gly Ile Tyr Leu Ser Met Ser Val Asp Thr
195 200 205
Val Ile Pro Pro Gly Glu Glu Ala Val Asn Ser Asp Ile Ala Cys Leu
210 215 220
Tyr Asn Arg Pro Thr Ile His Pro Phe Ala Tyr Arg Val His Thr His
225 230 235 240
Gln Leu Gly Gln Val Val Ser Gly Phe Arg Val Arg His Gly Lys Trp
245 250 255
Ser Leu Ile Gly Arg Gln Ser Pro Gln Leu Pro Gln Ala Phe Tyr Pro
260 265 270
Val Glu His Pro Val Glu Ile Ser Pro Gly Asp Ile Ile Ala Thr Arg
275 280 285
Cys Leu Phe Thr Gly Lys Gly Arg Thr Ser Ala Thr Tyr Ile Gly Gly
290 295 300

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Thr	Ser	Asn	Asp	Glu	Met	Cys	Asn	Leu	Tyr	Ile	Met	Tyr	Tyr	Met	Asp
305					310					315					320
Ala	Ala	His	Ala	Thr	Ser	Tyr	Met	Thr	Cys	Val	Gln	Thr	Gly	Glu	Pro
				325					330					335	
Lys	Leu	Phe	Gln	Asn	Ile	Pro	Glu	Ile	Ala	Asn	Val	Pro	Ile	Pro	Val
			340					345					350		
Ser	Pro	Asp	Met	Met	Met	Met	Met	Gly	His	Gly	His	His	His	Thr	Glu
		355					360					365			
Ala	Glu	Pro	Glu	Lys	Asn	Thr	Gly	Leu	Gln	Gln	Pro	Lys	Arg	Glu	Glu
	370					375					380				
Glu	Glu	Val	Leu	Asp	Gln	Gly	Leu	Ile	Thr	Leu	Gly	Asp	Ser	Ala	Val
385					390					395					400
<210> SEQ ID NO 40															
<211> LENGTH: 400															
<212> TYPE: PRT															
<213> ORGANISM: Rattus sp.															
<400> SEQUENCE: 40															
Met	Ala	Gly	Arg	Ala	Arg	Ser	Gly	Leu	Leu	Leu	Leu	Leu	Gly	Leu	
1				5				10					15		
Leu	Ala	Leu	Gln	Ser	Ser	Cys	Leu	Ala	Phe	Arg	Ser	Pro	Leu	Ser	Val
			20					25					30		
Phe	Lys	Arg	Phe	Lys	Glu	Thr	Thr	Arg	Ser	Phe	Ser	Asn	Glu	Cys	Leu
		35					40					45			
Gly	Thr	Ile	Gly	Pro	Val	Thr	Pro	Leu	Asp	Ala	Ser	Asp	Phe	Ala	Leu
	50					55					60				
Asp	Ile	Arg	Met	Pro	Gly	Val	Thr	Pro	Lys	Glu	Ser	Asp	Thr	Tyr	Phe
65					70					75					80
Cys	Met	Ser	Met	Arg	Leu	Pro	Val	Asp	Glu	Glu	Ala	Phe	Val	Ile	Asp
				85					90					95	
Phe	Lys	Pro	Arg	Ala	Ser	Met	Asp	Thr	Val	His	His	Met	Leu	Leu	Phe
			100					105					110		
Gly	Cys	Asn	Met	Pro	Ser	Ser	Thr	Gly	Ser	Tyr	Trp	Phe	Cys	Asp	Glu
		115					120					125			
Gly	Thr	Cys	Thr	Asp	Lys	Ala	Asn	Ile	Leu	Tyr	Ala	Trp	Ala	Arg	Asn
	130					135					140				
Ala	Pro	Pro	Thr	Arg	Leu	Pro	Lys	Gly	Val	Gly	Phe	Arg	Val	Gly	Gly
145					150					155					160
Glu	Thr	Gly	Ser	Lys	Tyr	Phe	Val	Leu	Gln	Val	His	Tyr	Gly	Asp	Ile
				165					170					175	
Ser	Ala	Phe	Arg	Asp	Asn	His	Lys	Asp	Cys	Ser	Gly	Val	Ser	Val	His
			180					185					190		
Leu	Thr	Arg	Val	Pro	Gln	Pro	Leu	Ile	Ala	Gly	Met	Tyr	Leu	Met	Met
		195					200					205			
Ser	Val	Asp	Thr	Val	Ile	Pro	Pro	Gly	Glu	Lys	Val	Val	Asn	Ala	Asp
	210					215					220				
Ile	Ser	Cys	Gln	Tyr	Lys	Met	Tyr	Pro	Met	His	Val	Phe	Ala	Tyr	Arg
225					230					235					240
Val	His	Thr	His	His	Leu	Gly	Lys	Val	Val	Ser	Gly	Tyr	Arg	Val	Arg
				245					250					255	
Asn	Gly	Gln	Trp	Thr	Leu	Ile	Gly	Arg	Gln	Asn	Pro	Gln	Leu	Pro	Gln
			260					265					270		
Ala	Phe	Tyr	Pro	Val	Glu	His	Pro	Val	Asp	Val	Thr	Phe	Gly	Asp	Ile
		275					280					285			

-continued

Leu	Ala	Ala	Arg	Cys	Val	Phe	Thr	Gly	Glu	Gly	Arg	Thr	Glu	Ala	Thr
290						295					300				
His	Ile	Gly	Gly	Thr	Ser	Ser	Asp	Glu	Met	Cys	Asn	Leu	Tyr	Ile	Met
305					310					315					320
Tyr	Tyr	Met	Glu	Ala	Lys	Tyr	Ala	Leu	Ser	Phe	Met	Thr	Cys	Thr	Lys
				325					330					335	
Asn	Val	Ala	Pro	Asp	Met	Phe	Arg	Thr	Ile	Pro	Ala	Glu	Ala	Asn	Ile
			340					345					350		
Pro	Ile	Pro	Val	Lys	Pro	Asp	Met	Val	Met	Met	His	Gly	His	His	Lys
		355					360					365			
Glu	Ala	Glu	Asn	Lys	Glu	Lys	Ser	Ala	Leu	Met	Gln	Gln	Pro	Lys	Gln
	370					375					380				
Gly	Glu	Glu	Glu	Val	Leu	Glu	Gln	Gly	Asp	Phe	Tyr	Ser	Leu	Leu	Ser
385					390					395					400

What is claimed is:

1. A recombinant enzyme comprising:
a polypeptide having an amino acid sequence set forth in
SEQ ID NO: 2 wherein at least one cysteine residue at
positions 6, 145, 40, 85, 252, or 274 is substituted to a
non-cysteine amino acid, and
wherein said recombinant enzyme has α -amidating activ-
ity and lacks at least one disulfide bond between posi-
tions 6 and 145, positions 40 and 85, or positions 252 and
274.
2. A recombinant enzyme comprising:
a polypeptide having an amino acid sequence set forth in
SEQ ID NO: 2 wherein at least one cysteine residue at

- positions 6, 145, 40, 85, 252, or 274 is substituted to a
non-cysteine amino acid, and
wherein said recombinant enzyme has α -amidating activ-
ity and lacks at least one disulfide bond between posi-
tions 6 and 145, positions 40 and 85, or positions 252 and
274; and
wherein said non-cysteine amino acid is alanine.
3. The recombinant enzyme of claim 2, wherein said
enzyme is AE-I [1-321] (C40A and C85A) (SEQ ID NO: 27),
AE-I [1-321] (C252A and C274A) (SEQ ID NO: 33), AE-I
[1-321] (C40A and C85A, C252A and C274A) (SEQ ID NO:
37), AE-I [8-321] (C145A) (SEQ ID NO: 25), or AE-I [8-321]
(C145A, C40A, and C85A) (SEQ ID NO: 35).

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