

US007993856B2

(12) **United States Patent**
Flannagan et al.

(10) **Patent No.:** **US 7,993,856 B2**
(45) **Date of Patent:** **Aug. 9, 2011**

(54) **SCREENING METHODS USING NOVEL BT TOXIN RECEPTORS FROM LEPIDOPTERAN INSECTS**

(75) Inventors: **Ronald D. Flannagan**, Grimes, IA (US);
Terry EuClaire Meyer, Urbandale, IA (US)

(73) Assignee: **Pioneer Hi-Bred International, Inc.**,
Des Moines, IA (US)

(*) Notice: Subject to any disclaimer, the term of this patent is extended or adjusted under 35 U.S.C. 154(b) by 213 days.

(21) Appl. No.: **12/413,830**

(22) Filed: **Mar. 30, 2009**

(65) **Prior Publication Data**

US 2009/0203042 A1 Aug. 13, 2009

Related U.S. Application Data

(62) Division of application No. 11/192,967, filed on Jul. 29, 2005, now Pat. No. 7,572,889, which is a division of application No. 09/715,909, filed on Nov. 17, 2000, now Pat. No. 7,060,491.

(60) Provisional application No. 60/234,099, filed on Sep. 21, 2000, provisional application No. 60/166,285, filed on Nov. 18, 1999.

(51) **Int. Cl.**
G01N 33/566 (2006.01)

(52) **U.S. Cl.** **435/7.21**

(58) **Field of Classification Search** None
See application file for complete search history.

(56) **References Cited**

U.S. PATENT DOCUMENTS

5,693,491 A 12/1997 Bulla et al.
5,804,393 A 9/1998 Geiser et al.
6,007,981 A 12/1999 Bulla et al.
6,423,502 B2 7/2002 Bulla et al.
6,660,497 B1 12/2003 Bulla et al.

FOREIGN PATENT DOCUMENTS

WO WO 96/12964 5/1996
WO WO 98/59048 12/1998
WO WO 01/34807 A2 5/2001

OTHER PUBLICATIONS

Dorsch, J., "Isolation and Characterization of the Insecticidal Toxin Binding Site From the Receptor BT-R₁ of *Manduca sexta*," A Dissertation submitted to the Department of Molecular Biology and the Graduate School of the University of Wyoming, 1998.

Estruch, J. et al., "Transgenic Plants: An Emerging Approach to Pest Control," *Nature Biotechnology*, 1997, vol. 15, pp. 137-141.

Francis, B., et al., "Further Characterization of BT-R₁, The Cadherin-Like Receptor for Cry IAB Toxin in Tobacco Hornworm (*Manduca sexta*) Midguts, Insect," *Biochem. Mol. Biol.*, 1997, vol. 27(6), pp. 541-550.

Franklin, S., et al., "Southern Analysis of BT-R₁, The *Manduca sexta* Gene Encoding the Receptor for the CryIAB Toxin of *Bacillus thuringiensis*," *Mol. Gen. Genet.*, 1997, vol. 256, pp. 517-524.

Gahan, L.J., et al., "Identification of a Gene Associated with Bt Resistance in *Heliothis virescens*," *Science* (2001) pp. 857-860, vol. 293.

Garczynski, S., et al., "Identification of Putative Insect Brush Border Membrane-Binding Molecules Specific to *Bacillus thuringiensis* δ-Endotoxin by Protein Blot Analysis, Applied and Environmental Microbiology," 1991, vol. 57(10), pp. 2816-2820.

Gill, S., et al., "Identification, Isolation, and Cloning of a *Bacillus thuringiensis* CryIAC Toxin-Binding Protein from the Midgut of the Lepidopteran Insect *Heliothis virescens*," *The Journal of Biological Chemistry*, 1995, vol. 270(45), pp. 27277-27282.

Hofte, et al., "Insecticidal Crystal Proteins of *Bacillus thuringiensis*," *Microbiological Reviews*, 1989, vol. 53(2), pp. 242-255.

Hua, G., et al., "Binding Analyses of *Bacillus thuringiensis* Cry δ-Endotoxins Using Brush Border Membrane Vesicles of *Ostrinia nubilalis*," *Applied and Environmental Microbiology*, 2001, vol. 67(2), pp. 872-879.

Ihara, H., et al., "Purification and Partial Amino Acid Sequences of the Binding Protein from Bombyx Mori for CryIAa δ-endotoxin of *Bacillus thuringiensis*," *Elsevier Science Inc.*, 1998, pp. 197-204.

Keeton, T., et al., "Effects of Midgut-Protein-Preparative and Ligand Binding Procedures on the Toxin Binding Characteristics of BT-R₁, A Common High-Affinity Receptor in *Manduca sexta* for CryIA *Bacillus thuringiensis* Toxins," *Applied and Environmental Microbiology*, 1998, vol. 64(6), pp. 2158-2165.

Keeton, T., et al., "Ligand Specificity and Affinity of BT-R₁, The *Bacillus thuringiensis* Cry1A Toxin Receptor From *Manduca sexta*, Expressed in Mammalian and Insect Cell Cultures," *Applied and Environmental Microbiology*, 1997, vol. 63(9), pp. 3419-3425.

Knight, P., et al., "The Receptor for *Bacillus thuringiensis* CryIA(c) Delta-Endotoxin in the Brush Border Membrane of the Lepidopteran *Manduca sexta* is Aminopeptidase N," *Molecular Microbiology*, 1994, vol. 11(3), pp. 429-436.

Lee, M., et al., "Aminopeptidase N Purified from Gypsy Moth Brush Border Membrane Vesicles Is a Specific Receptor for *Bacillus thuringiensis* CryIAC Toxin," *Applied and Environmental Microbiology*, 1996, vol. 62(8), pp. 2845-2849.

McGaughey, W., et al., "RT Resistance Management a Plan for Reconciling the Needs of the Many Stakeholders in Bt-Based Products," *Nature Biotechnology*, 1998, vol. 16, pp. 144-146.

Matty, W.S.A., "Identification, Purification and Cloning of a High-Affinity Invertebrate Protocadherin Receptor BT-R₂ From the Pink Bollworm (*Pectinophora gossypiella*) for *Bacillus thuringiensis* CRY1A Toxins," Jul. 1999, Dissertation submitted to the Dept. of Molecular Biology and The Graduate School of the University of Wyoming.

Midboe, E.G., "Characterization of the BT-R₁ Gene and Its Expression in *Manduca sexta*," Jul. 1999, Dissertation submitted to the Dept. of Molecular Biology and The Graduate School of the University of Wyoming.

Nagamatsu, Y., et al., "Cloning, Sequencing, and Expression of the Bombyx Mori Receptor for *Bacillus thuringiensis* Insecticidal CryIA(a) Toxin," *Biosci. Biotechnol. Biochem.*, 1998, vol. 62(4), pp. 727-734.

(Continued)

Primary Examiner — Robert C Hayes

(74) *Attorney, Agent, or Firm* — Alston & Bird LLP

(57) **ABSTRACT**

The invention relates to Bt toxin resistance management. The invention particularly relates to the isolation and characterization of nucleic acid and polypeptides for a novel Bt toxin receptor. The nucleic acid and polypeptides are useful in identifying and designing novel Bt toxin receptor ligands including novel insecticidal toxins.

3 Claims, 1 Drawing Sheet

OTHER PUBLICATIONS

Nagamatsu, Y. et al., "The Cadherin-Like Protein is Essential to Specificity Determination and Cytotoxic Action of the *Bacillus thuringiensis* Insecticidal CryIAa Toxin," *Febs Letters*, 1999, vol. 460, pp. 385-390.

Oddou, P., et al., "Immunologically Unrelated *Heliothis* sp. and *Spodoptera* sp. Midgut Membrane-Proteins Bind *Bacillus thuringiensis* CryIA(b) δ -endotoxin," *Eur. J. Biochem.*, 1993, vol. 212, pp. 145-150.

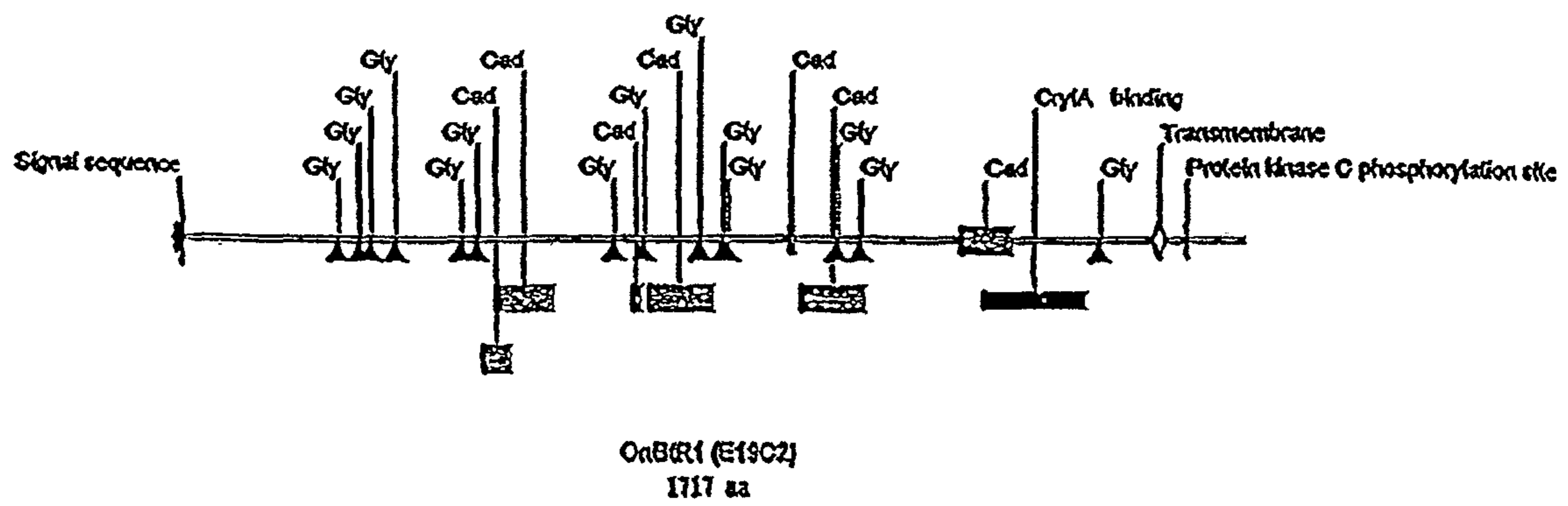
Roush, R., et al., "Assessing the Odds: The Emergence of Resistance to BT Transgenic Plants," *Nature Biotechnology*, 1997, vol. 15, pp. 816-817.

Rudinger, J., "Characteristics of the Amino Acids as Components of a Peptide Hormone Sequence," *J.A. Parsons University*, 1976, Park Press, Baltimore, pp. 1-7.

Skolnick, J. et al., "From Genes to Protein Structure and Function: Novel Applications of Computational Approaches in the Genomic Era," *Trends in Biotechnology*, pp. 34-39, vol. 18, No. 1.

Vadlamudi, R., et al., "Cloning and Expression of a Receptor for an Insecticidal Toxin of *Bacillus thuringiensis*," *The Journal of Biological Chemistry*, 1995, vol. 270(10), pp. 5490-5494.

Vadlamudi, R., et al., "A Specific Binding Protein from *Manduca sexta* for the Insecticidal Toxin of *Bacillus thuringiensis* Subsp. Berliner," *The Journal of Biological Chemistry*, 1993, vol. 268(17), pp. 12334-12340.



Gly = putative glycosilation sites

Cad = cadherin-like domain

1

**SCREENING METHODS USING NOVEL BT
TOXIN RECEPTORS FROM LEPIDOPTERAN
INSECTS**

CROSS-REFERENCE TO RELATED
APPLICATION

This application is a divisional application of U.S. Utility application Ser. No. 11/192,967, filed Jul. 29, 2005, which is a divisional application of U.S. Utility application Ser. No. 09/715,909, filed Nov. 17, 2000, now U.S. Pat. No. 7,060,491, which claims the benefit of U.S. Provisional Application Ser. No. 60/234,099, filed Sep. 21, 2000 and U.S. Provisional Application Ser. No. 60/166,285, filed Nov. 18, 1999, the contents of which are herein incorporated by reference in their entirety.

REFERENCE TO A SEQUENCE LISTING
SUBMITTED AS TEXT FILE VIA EFS-WEB

The official copy of the sequence listing is submitted concurrently with the specification as a text file via EFS-Web, in compliance with the American Standard Code for Information Interchange (ASCII), with a file name of 369837SequenceListing.txt, a creation date of Mar. 24, 2009, and a size of 110 KB. The sequence listing filed via EFS-Web is part of the specification and is hereby incorporated in its entirety by reference herein.

FIELD OF THE INVENTION

The field of the invention is manipulating Bt toxin susceptibility in plant pests. The field of the invention relates to the isolation and characterization of nucleic acid and polypeptides for a novel Bt toxin receptor. The nucleic acid and polypeptides are useful in developing new insecticides.

BACKGROUND OF THE INVENTION

Traditionally, growers used chemical pesticides as a means to control agronomically important pests. The introduction of transgenic plants carrying the delta-endotoxin from *Bacillus thuringiensis* (Bt) afforded a non-chemical method of control. Bt toxins have traditionally been categorized by their specific toxicity towards specific insect categories. For example, the Cry1 group of toxins are toxic to *Lepidoptera*. The Cry1 group includes, but is not limited to, Cry1A(a), Cry1A(b) and Cry1A(c). See Hofte et al (1989) *Microbiol Rev* 53: 242-255.

Lepidopteran insects cause considerable damage to maize crops throughout North America and the world. One of the leading pests is *Ostrinia nubilalis*, commonly called the European Corn Borer (ECB). Genes encoding the crystal proteins Cry1A(b) and Cry1A(c) from Bt have been introduced into maize as a means of ECB control. These transgenic maize hybrids have been effective in control of ECB. However, developed resistance to Bt toxins presents a challenge in pest control. See McGaughey et al. (1998) *Nature Biotechnology* 16: 144-146; Estruch et al. (1997) *Nature Biotechnology* 15: 137-141; Roush et al. (1997) *Nature Biotechnology* 15: 816-817; and Hofte et al (1989) *Microbiol Rev* 53: 242-255.

The primary site of action of Cry1 toxins is in the brush border membranes of the midgut epithelia of susceptible insect larvae such as lepidopteran insects. Cry1A toxin binding polypeptides have been characterized from a variety of *Lepidopteran* species. A Cry1A(c) binding polypeptide with homology to an aminopeptidase N has been reported from

2

Manduca sexta, *Lymantria dispar*, *Helicoverpa zea* and *Heliothis virescens*. See Knight et al (1994) *Mol Micro* 11: 429-436; Lee et al (1996) *Appl Environ Micro* 63: 2845-2849; Gill et al. (1995) *J Biol. Chem* 270: 27277-27282; and Garczynski et al (1991) *Appl Environ Microbiol* 10: 2816-2820.

Another Bt toxin binding polypeptide (BTR1) cloned from *M. sexta* has homology to the cadherin polypeptide superfamily and binds Cry1A(a), Cry1A(b) and Cry1A(c). See Vadlamudi et al. (1995) *J Biol Chem* 270(10):5490-4, Keeton et al. (1998) *Appl Environ Microbiol* 64(6):2158-2165; Keeton et al. (1997) *Appl Environ Microbiol* 63(9):3419-3425 and U.S. Pat. No. 5,693,491.

A subsequently cloned homologue to BTR1 demonstrated binding to Cry1A(a) from *Bombyx mori* as described in Ihara et al. (1998) *Comparative Biochemistry and Physiology, Part B* 120:197-204 and Nagamatsu et al. (1998) *Biosci. Biotechnol. Biochem.* 62(4):727-734.

Identification of the plant pest binding polypeptides for Bt toxins are useful for investigating Bt toxin-Bt toxin receptor interactions, selecting and designing improved toxins, developing novel insecticides, and new Bt toxin resistance management strategies.

SUMMARY OF THE INVENTION

Compositions and methods for modulating susceptibility of a cell to Bt toxins are provided. The compositions include Bt toxin receptor polypeptides, and fragments and variants thereof, from the lepidopteran insects European corn borer (ECB, *Ostrinia nubilalis*), corn earworm (CEW, *Heliothis Zea*), and fall armyworm (FAW, *Spodoptera frugiperda*). The polypeptides bind Cry1A toxins, more particularly Cry1A (b). Nucleic acids encoding the polypeptides, antibodies specific to the polypeptides, as well as nucleic acid constructs for expressing the polypeptides in cells of interest are also provided.

The methods are useful for investigating the structure-function relationships of Bt toxin receptors; investigating the toxin-receptor interactions; elucidating the mode of action of Bt toxins; screening and identifying novel Bt toxin receptor ligands including novel insecticidal toxins; and designing and developing novel Bt toxin receptor ligands.

The methods are useful for managing Bt toxin resistance in plant pests, and protecting plants against damage by plant pests.

BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1 schematically depicts the location of the signal sequence, putative glycosylation sites, cadherin-like domains, transmembrane segment, Cry1A binding region and protein kinase C phosphorylation site of the Bt toxin receptor from *Ostrinia nubilalis*; the nucleotide sequence of the receptor set forth in SEQ ID NO: 1 and the corresponding deduced amino acid sequence in SEQ ID NO: 2.

DETAILED DESCRIPTION OF THE INVENTION

The invention is directed to novel receptor polypeptides that bind Bt toxin, the receptor being derived from the order *lepidoptera*. The receptors of the invention include those receptor polypeptides that bind Bt toxin and are derived from the lepidopteran superfamily Pyraloidea and particularly from the species *Ostrinia*, specifically *Ostrinia nubilalis*; those derived from *Spodoptera frugiperda* (*S. frugiperda*);

and those derived from *Heliothis Zea* (*H. Zea*). The polypeptides have homology to members of the cadherin superfamily of proteins.

Accordingly, compositions of the invention include isolated polypeptides that are involved in Bt toxin binding. In particular, the present invention provides for isolated nucleic acid molecules comprising nucleotide sequences encoding the amino acid sequences shown in SEQ ID NOs: 2, 4, and 6; or the nucleotide sequences having the DNA sequences deposited in a plasmid in a bacterial host as Patent Deposit No. PTA-278, PTA-1760, and PTA-2222. Further provided are polypeptides having an amino acid sequence encoded by a nucleic acid molecule described herein, for example those set forth in SEQ ID NOs: 1, 3, and 5; those deposited in a plasmid in a bacterial host as Patent Deposit Nos. PTA-278, PTA-1760, and PTA-2222; and fragments and variants thereof.

Plasmids containing the nucleotide sequences of the invention were deposited with the Patent Depository of the American Type Culture Collection (ATCC), Manassas, Va. on Jun. 25, 1999; Apr. 25, 2000; and Jul. 11, 2000; and assigned Patent Deposit Nos. PTA-278, PTA-1760, and PTA-2222. These deposits will be maintained under the terms of the Budapest Treaty on the International Recognition of the Deposit of Microorganisms for the Purposes of Patent Procedure. These deposits were made merely as a convenience for those of skill in the art and are not an admission that a deposit is required under 35 U.S.C. §112.

The term "nucleic acid" refers to all forms of DNA such as cDNA or genomic DNA and RNA such as mRNA, as well as analogs of the DNA or RNA generated using nucleotide analogs. The nucleic acid molecules can be single stranded or double stranded. Strands can include the coding or non-coding strand.

The invention encompasses isolated or substantially purified nucleic acid or polypeptide compositions. An "isolated" or "purified" nucleic acid molecule or polypeptide, or biologically active portion thereof, is substantially free of other cellular material, or culture medium when produced by recombinant techniques, or substantially free of chemical precursors or other chemicals when chemically synthesized. Preferably, an "isolated" nucleic acid is free of sequences (preferably polypeptide encoding sequences) that naturally flank the nucleic acid (i.e., sequences located at the 5' and 3' ends of the nucleic acid) in the genomic DNA of the organism from which the nucleic acid is derived. For example, in various embodiments, the isolated nucleic acid molecule can contain less than about 5 kb, 4 kb, 3 kb, 2 kb, 1 kb, 0.5 kb, or 0.1 kb of nucleotide sequences that naturally flank the nucleic acid molecule in genomic DNA of the cell from which the nucleic acid is derived. A polypeptide that is substantially free of cellular material includes preparations of polypeptide having less than about 30%, 20%, 10%, 5%, (by dry weight) of contaminating polypeptide. When the polypeptide of the invention or biologically active portion thereof is recombinantly produced, preferably culture medium represents less than about 30%, 20%, 10%, or 5% (by dry weight) of chemical precursors or non-polypeptide-of-interest chemicals.

It is understood, however, that there are embodiments in which preparations that do not contain the substantially pure polypeptide may also be useful. Thus, less pure preparations can be useful where the contaminating material does not interfere with the specific desired use of the peptide. The compositions of the invention also encompass fragments and variants of the disclosed nucleotide sequences and the polypeptides encoded thereby.

The compositions of the invention are useful for, among other uses, expressing the receptor polypeptides in cells of interest to produce cellular or isolated preparations of the polypeptides for investigating the structure-function relationships of Bt toxin receptors; investigating the toxin-receptor interactions; elucidating the mode of action of Bt toxins; screening and identifying novel Bt toxin receptor ligands including novel insecticidal toxins; and designing and developing novel Bt toxin receptor ligands including novel insecticidal toxins.

The isolated nucleotide sequences encoding the receptor polypeptides of the invention are expressed in a cell of interest; and the Bt toxin receptor polypeptides produced by the expression is utilized in intact cell or in-vitro receptor binding assays, and/or intact cell toxicity assays. Methods and conditions for Bt toxin binding and toxicity assays are known in the art and include but are not limited to those described in U.S. Pat. No. 5,693,491; T. P. Keeton et al. (1998) *Appl. Environ. Microbiol.* 64(6):2158-2165; B. R. Francis et al. (1997) *Insect Biochem. Mol. Biol.* 27(6):541-550; T. P. Keeton et al. (1997) *Appl. Environ. Microbiol.* 63(9):3419-3425; R. K. Vadlamudi et al. (1995) *J. Biol. Chem.* 270(10):5490-5494; Ihara et al. (1998) *Comparative Biochem. Physiol. B* 120:197-204; Nagamatsu et al. (1998) *Biosci. Biotechnol. Biochem.* 62(4):727-734, herein incorporated by reference. Such methods could be modified by one of ordinary skill in the art to develop assays utilizing the polypeptides of the invention.

By "cell of interest" is intended any cell in which expression of the polypeptides of the invention is desired. Cells of interest include, but are not limited to mammalian, avian, insect, plant, bacteria, fungi and yeast cells. Cells of interest include but are not limited to cultured cell lines, primary cell cultures, cells in vivo, and cells of transgenic organisms.

The methods of the invention encompass using the polypeptides encoded by the nucleotide sequences of the invention in receptor binding and/or toxicity assays to screen candidate ligands and identify novel Bt toxin receptor ligands, including receptor agonists and antagonists. Candidate ligands include molecules available from diverse libraries of small molecules created by combinatorial synthetic methods. Candidate ligands also include, but are not limited to antibodies, peptides, and other small molecules designed or deduced to interact with the receptor polypeptides of the invention. Candidate ligands include but are not limited to peptide fragments of the receptor, anti-receptor antibodies, anti-idiotypic antibodies mimicking one or more receptor binding domains of a toxin, fusion proteins produced by combining two or more toxins or fragments thereof, and the like. Ligands identified by the screening methods of the invention include potential novel insecticidal toxins, the insecticidal activity of which can be determined by known methods; for example, as described in U.S. Pat. No. 5,407,454; U.S. application Ser. No. 09/218,942; U.S. application Ser. No. 09/003,217.

The invention provides methods for screening for ligands that bind to the polypeptides described herein. Both the polypeptides and relevant fragments thereof (for example, the toxin binding domain) can be used to screen by assay for compounds that bind to the receptor and exhibit desired binding characteristics. Desired binding characteristics include, but are not limited to binding affinity, binding site specificity, association and dissociation rates, and the like. The screening assays could be intact cell or in vitro assays which include exposing a ligand binding domain to a sample ligand and detecting the formation of a ligand-binding polypeptide com-

plex. The assays could be direct ligand-receptor binding assays or ligand competition assays.

In one embodiment, the methods comprise providing at least one Bt toxin receptor polypeptide of the invention, contacting the polypeptide with a sample and a control ligand under conditions promoting binding; and determining binding characteristics of sample ligands, relative to control ligands. The methods encompass any method known to the skilled artisan which can be used to provide the polypeptides of the invention in a binding assay. For in vitro binding assays, the polypeptide may be provided as isolated, lysed, or homogenized cellular preparations. Isolated polypeptides may be provided in solution, or immobilized to a matrix. Methods for immobilizing polypeptides are well known in the art, and include but are not limited to construction and use of fusion polypeptides with commercially available high affinity ligands. For example, GST fusion proteins can be adsorbed onto glutathione sepharose beads (Sigma Chemical, St. Louis, Mo.) or glutathione derivatized microtitre plates. The polypeptides can also be immobilized utilizing well techniques in the art utilizing conjugation of biotin and streptavidin. The polypeptides can also be immobilized utilizing well known techniques in the art utilizing chemical conjugation (linking) of polypeptides to a matrix. Alternatively, the polypeptides may be provided in intact cell binding assays in which the polypeptides are generally expressed as cell surface Bt toxin receptors.

The invention provides methods utilizing intact cell toxicity assays to screen for ligands that bind to the receptor polypeptides described herein and confer toxicity upon a cell of interest expressing the polypeptide. A ligand selected by this screening is a potential insecticidal toxin to insects expressing the receptor polypeptides, particularly enterally. This deduction is premised on theories that insect specificity of a particular Bt toxin is determined by the presence of the receptor in specific insect species, or that binding of the toxins is specific for the receptor of some insect species and is bind is insignificant or nonspecific for other variant receptors. See, for example Hofte et al (1989) *Microbiol Rev* 53: 242-255. The toxicity assays include exposing, in intact cells expressing a polypeptide of the invention, the toxin binding domain of the polypeptide to a sample ligand and detecting the toxicity effected in the cell expressing the polypeptide. By "toxicity" is intended the decreased viability of a cell. By "viability" is intended the ability of a cell to proliferate and/or differentiate and/or maintain its biological characteristics in a manner characteristic of that cell in the absence of a particular cytotoxic agent.

In one embodiment, the methods of the present invention comprise providing at least one cell surface Bt toxin receptor polypeptide of the invention comprising an extracellular toxin binding domain, contacting the polypeptide with a sample and a control ligand under conditions promoting binding, and determining the viability of the cell expressing the cell surface Bt toxin receptor polypeptide, relative to the control ligand.

By "contacting" is intended that the sample and control agents are presented to the intended ligand binding site of the polypeptides of the invention.

By "conditions promoting binding" is intended any combination of physical and biochemical conditions that enables a ligand of the polypeptides of the invention to determinably bind the intended polypeptide over background levels. Examples of such conditions for binding of Cry1 toxins to Bt toxin receptors, as well as methods for assessing the binding, are known in the art and include but are not limited to those described in Keeton et al. (1998) *Appl Environ Microbiol*

64(6): 2158-2165; Francis et al. (1997) *Insect Biochem Mol Biol* 27(6):541-550; Keeton et al. (1997) *Appl Environ Microbiol* 63(9):3419-3425; Vadlamudi et al. (1995) *J Biol Chem* 270(10):5490-5494; Ihara et al. (1998) *Comparative Biochemistry and Physiology, Part B* 120:197-204; and Nagamatsu et al. (1998) *Biosci. Biotechnol. Biochem.* 62(4):727-734, the contents of which are herein incorporated by reference. In this aspect of the present invention, known and commercially available methods for studying protein-protein interactions, such as yeast and/or bacterial two-hybrid systems could also be used. Two-hybrid systems are available from, for example, CLONTECH (Palo Alto, Calif.) or Display Systems Biotech Inc. (Vista, Ca).

The compositions and screening methods of the invention are useful for designing and developing novel Bt toxin receptor ligands including novel insecticidal toxins. Various candidate ligands; ligands screened and characterized for binding, toxicity, and species specificity; and/or ligands having known characteristics and specificities, could be linked or modified to produce novel ligands having particularly desired characteristics and specificities. The methods described herein for assessing binding, toxicity and insecticidal activity could be used to screen and characterize the novel ligands.

In one embodiment of the present invention, the sequences encoding the receptors of the invention, and variants and fragments thereof, are used with yeast and bacterial two-hybrid systems to screen for Bt toxins of interest (for example, more specific and/or more potent toxins), or for insect molecules that bind the receptor and can be used in developing novel insecticides.

By "linked" is intended that a covalent bond is produced between two or more molecules. Known methods that can be used for modification and/or linking of polypeptide ligands such as toxins, include but are not limited to mutagenic and recombinogenic approaches including but not limited to site-directed mutagenesis, chimeric polypeptide construction and DNA shuffling. Such methods are described in further detail below. Known polypeptide modification methods also include methods for covalent modification of polypeptides. "Operably linked" means that the linked molecules carry out the function intended by the linkage.

The compositions and screening methods of the present invention are useful for targeting ligands to cells expressing the receptor polypeptides of the invention. For targeting, secondary polypeptides, and/or small molecules which do not bind the receptor polypeptides of the invention are linked with one or more primary ligands which bind the receptor polypeptides; including but not limited to Cry1A toxin; more particularly Cry1A(b) toxin or a fragment thereof. By this linkage, any polypeptide and/or small molecule linked to a primary ligand could be targeted to the receptor polypeptide, and thereby to a cell expressing the receptor polypeptide; wherein the ligand binding site is available at the extracellular surface of the cell.

In one embodiment of the invention, at least one secondary polypeptide toxin is linked with a primary Cry1 A toxin capable of binding the receptor polypeptides of the invention to produce a combination toxin which is targeted and toxic to insects expressing the receptor for the primary toxin. Such insects include those of the order *lepidoptera*, superfamily Pyraloidea and particularly from the species *Ostrinia*, specifically *Ostrinia nubilalis*. Such insects include the lepidopterans *S. frugiperda* and *H. Zea*. Such a combination toxin is particularly useful for eradicating or reducing crop damage by insects which have developed resistance to the primary toxin.

For expression of the Bt toxin receptor polypeptides of the invention in a cell of interest, the Bt toxin receptor sequences are provided in expression cassettes. The cassette will include 5' and 3' regulatory sequences operably linked to a Bt toxin receptor sequence of the invention. In this aspect of the present invention, by "operably linked" is intended a functional linkage between a promoter and a second sequence, wherein the promoter sequence initiates and mediates transcription of the DNA sequence corresponding to the second sequence. In reference to nucleic acids, generally, operably linked means that the nucleic acid sequences being linked are contiguous and, where necessary to join two polypeptide coding regions, contiguous and in the same reading frame. The cassette may additionally contain at least one additional gene to be cotransformed into the organism. Alternatively, the additional gene(s) can be provided on multiple expression cassettes.

Such an expression cassette is provided with a plurality of restriction sites for insertion of the Bt toxin receptor sequence to be under the transcriptional regulation of the regulatory regions. The expression cassette may additionally contain selectable marker genes.

The expression cassette will include in the 5'-3' direction of transcription, a transcriptional and translational initiation region, a Bt toxin receptor nucleotide sequence of the invention, and a transcriptional and translational termination region functional in host cells. The transcriptional initiation region, the promoter, may be native or analogous, or foreign or heterologous to the plant host. Additionally, the promoter may be the natural sequence or alternatively a synthetic sequence. By "foreign" is intended that the transcriptional initiation region is not found in the native host cells into which the transcriptional initiation region is introduced. As used herein, a chimeric gene comprises a coding sequence operably linked to a transcription initiation region that is heterologous to the coding sequence.

While it may be preferable to express the sequences using heterologous promoters, the native promoter sequences may be used. Such constructs would change expression levels of Bt toxin receptor in the cell of interest. Thus, the phenotype of the cell is altered.

The termination region may be native with the transcriptional initiation region, may be native with the operably linked DNA sequence of interest, or may be derived from another source.

Where appropriate, the gene(s) may be optimized for increased expression in a particular transformed cell of interest. That is, the genes can be synthesized using host cell-preferred codons for improved expression.

Additional sequence modifications are known to enhance gene expression in a cellular host. These include elimination of sequences encoding spurious polyadenylation signals, exon-intron splice site signals, transposon-like repeats, and other such well-characterized sequences that may be deleterious to gene expression. The G-C content of the sequence may be adjusted to levels average for a given cellular host, as calculated by reference to known genes expressed in the host cell. When possible, the sequence is modified to avoid predicted hairpin secondary mRNA structures.

The expression cassettes may additionally contain 5' leader sequences in the expression cassette construct. Such leader sequences can act to enhance translation. Translation leaders are known in the art and include: picornavirus leaders, for example, EMCV leader (Encephalomyocarditis 5' noncoding region) (Elroy-Stein et al. (1989) *PNAS USA* 86:6126-6130); potyvirus leaders, for example, TEV leader (Tobacco Etch Virus) (Allison et al. (1986); MDMV leader (Maize Dwarf

Mosaic Virus); *Virology* 154:9-20), and human immunoglobulin heavy-chain binding polypeptide (BiP), (Macejak et al. (1991) *Nature* 353:90-94); untranslated leader from the coat polypeptide mRNA of alfalfa mosaic virus (AMV RNA 4) (Jobling et al. (1987) *Nature* 325:622-625); tobacco mosaic virus leader (TMV) (Gallie et al. (1989) in *Molecular Biology of RNA*, ed. Cech (Liss, New York), pp. 237-256); and maize chlorotic mottle virus leader (MCMV) (Lommel et al. (1991) *Virology* 81:382-385). See also, Della-Cioppa et al. (1987) *Plant Physiol.* 84:965-968. Other methods known to enhance translation can also be utilized, for example, introns, and the like.

In preparing the expression cassette, the various DNA fragments may be manipulated, so as to provide for the DNA sequences in the proper orientation and, as appropriate, in the proper reading frame. Toward this end, adapters or linkers may be employed to join the DNA fragments or other manipulations may be involved to provide for convenient restriction sites, removal of superfluous DNA, removal of restriction sites, or the like. For this purpose, in vitro mutagenesis, primer repair, restriction, annealing, resubstitutions, e.g., transitions and transversions, may be involved.

Using the nucleic acids of the present invention, the polypeptides of the invention could be expressed in any cell of interest, the particular choice of the cell depending on factors such as the level of expression and/or receptor activity desired. Cells of interest include, but are not limited to conveniently available mammalian, plant, insect, bacteria, and yeast host cells. The choice of promoter, terminator, and other expression vector components will also depend on the cell chosen. The cells produce the protein in a non-natural condition (e.g., in quantity, composition, location, and/or time), because they have been genetically altered through human intervention to do so.

It is expected that those of skill in the art are knowledgeable in the numerous expression systems available for expression of a nucleic acid encoding a protein of the present invention. No attempt to describe in detail the various methods known for the expression of proteins in prokaryotes or eukaryotes will be made.

In brief summary, the expression of isolated nucleic acids encoding a protein of the present invention will typically be achieved by operably linking, for example, the DNA or cDNA to a promoter, followed by incorporation into an expression vector. The vectors can be suitable for replication and integration in either prokaryotes or eukaryotes. Typical expression vectors contain transcription and translation terminators, initiation sequences, and promoters useful for regulation of the expression of the DNA encoding a protein of the present invention. To obtain high level expression of a cloned gene, it is desirable to construct expression vectors which contain, at the minimum, a strong promoter to direct transcription, a ribosome binding site for translational initiation, and a transcription/translation terminator. One of skill would recognize that modifications can be made to a protein of the present invention without diminishing its biological activity. Some modifications may be made to facilitate the cloning, expression, or incorporation of the targeting molecule into a fusion protein. Such modifications are well known to those of skill in the art and include, for example, a methionine added at the amino terminus to provide an initiation site, or additional amino acids (e.g., poly His) placed on either terminus to create conveniently located restriction sites or termination codons or purification sequences.

Prokaryotic cells may be used as hosts for expression. Prokaryotes most frequently are represented by various strains of *E. coli*; however, other microbial strains may also be

used. Commonly used prokaryotic control sequences which are defined herein to include promoters for transcription initiation, optionally with an operator, along with ribosome binding site sequences, include such commonly used promoters as the beta lactamase (penicillinase) and lactose (lac) promoter systems (Chang et al. (1977) *Nature* 198:1056), the tryptophan (trp) promoter system (Goeddel et al. (1980) *Nucleic Acids Res.* 8:4057) and the lambda-derived P_L promoter and N-gene ribosome binding site (Shimatake et al. (1981) *Nature* 292:128). The inclusion of selection markers in DNA vectors transfected in *E. coli* is also useful. Examples of such markers include genes specifying resistance to ampicillin, tetracycline, or chloramphenicol.

The vector is selected to allow introduction into the appropriate host cell. Bacterial vectors are typically of plasmid or phage origin. Appropriate bacterial cells are infected with phage vector particles or transfected with naked phage vector DNA. If a plasmid vector is used, the bacterial cells are transfected with the plasmid vector DNA. Expression systems for expressing a protein of the present invention are available using *Bacillus* sp. and *Salmonella* (Palva et al. (1983) *Gene* 22:229-235; Mosbach et al. (1983) *Nature* 302:543-545).

A variety of eukaryotic expression systems such as yeast, insect cell lines, plant and mammalian cells, are known to those of skill in the art. The sequences of the present invention can be expressed in these eukaryotic systems. In some embodiments, transformed/transfected plant cells are employed as expression systems for production of the proteins of the instant invention.

Synthesis of heterologous proteins in yeast is well known. Sherman, F. et al. (1982) *Methods in Yeast Genetics*, Cold Spring Harbor Laboratory is a well recognized work describing the various methods available to produce the protein in yeast. Two widely utilized yeast for production of eukaryotic proteins are *Saccharomyces cerevisia* and *Pichia pastoris*. Vectors, strains, and protocols for expression in *Saccharomyces* and *Pichia* are known in the art and available from commercial suppliers (e.g., Invitrogen). Suitable vectors usually have expression control sequences, such as promoters, including 3-phosphoglycerate kinase or alcohol oxidase, and an origin of replication, termination sequences and the like as desired.

A protein of the present invention, once expressed, can be isolated from yeast by lysing the cells and applying standard protein isolation techniques to the lysates. The monitoring of the purification process can be accomplished by using Western blot techniques or radioimmunoassay or other standard immunoassay techniques.

The sequences encoding proteins of the present invention can also be ligated to various expression vectors for use in transfecting cell cultures of, for instance, mammalian, insect, or plant origin. Illustrative of cell cultures useful for the production of the peptides are mammalian cells. Mammalian cell systems often will be in the form of monolayers of cells although mammalian cell suspensions may also be used. A number of suitable host cell lines capable of expressing intact proteins have been developed in the art, and include the COS, HEK293, BHK21, and CHO cell lines. Expression vectors for these cells can include expression control sequences, such as an origin of replication, a promoter (e.g., the CMV promoter, a HSV tk promoter or pgk (phosphoglycerate kinase promoter)), an enhancer (Queen et al. (1986) *Immunol. Rev.* 89:49), and necessary processing information sites, such as ribosome binding sites, RNA splice sites, polyadenylation sites (e.g., an SV40 large T Ag poly A addition site), and transcriptional terminator sequences. Other animal cells use-

ful for production of proteins of the present invention are available, for instance, from the American Type Culture Collection Catalogue of Cell Lines and Hybridomas (7th edition, 1992). A particular example of mammalian cells for expression of a Bt toxin receptor and assessing Bt toxin cytotoxicity mediated by the receptor, includes embryonic 293 cells. See U.S. Pat. No. 5,693,491, herein incorporated by reference.

Appropriate vectors for expressing proteins of the present invention in insect cells are usually derived from the SF9 baculovirus. Suitable insect cell lines include mosquito larvae, silkworm, armyworm, moth and *Drosophila* cell lines such as a Schneider cell line (See Schneider et al. (1987) *J. Embryol. Exp. Morphol.* 27: 353-365).

As with yeast, when higher animal or plant host cells are employed, polyadenylation or transcription terminator sequences are typically incorporated into the vector. An example of a terminator sequence is the polyadenylation sequence from the bovine growth hormone gene. Sequences for accurate splicing of the transcript may also be included. An example of a splicing sequence is the VP1 intron from SV40 (Sprague et al. (1983) *J. Virol.* 45:773-781). Additionally, gene sequences to control replication in the host cell may be incorporated into the vector such as those found in bovine papilloma virus-type vectors. Saveria-Campo, M., Bovine Papilloma Virus DNA a Eukaryotic Cloning Vector in *DNA Cloning Vol. II a Practical Approach*, D. M. Glover, ed., IRL Pres, Arlington, Va. pp. 213-238 (1985).

In a particular embodiment of the invention, it may be desirable to negatively control receptor binding; particularly, when toxicity to a cell is no longer desired or if it is desired to reduce toxicity to a lower level. In this case, ligand-receptor polypeptide binding assays can be used to screen for compounds which bind to the receptor but do not confer toxicity to a cell expressing the receptor. The examples of a molecule that can be used to block ligand binding include an antibody that specifically recognizes the ligand binding domain of the receptor such that ligand binding is decreased or prevented as desired.

In another embodiment, receptor polypeptide expression could be blocked by the use of antisense molecules directed against receptor RNA or ribozymes specifically targeted to this receptor RNA. It is recognized that with the provided nucleotide sequences, antisense constructions, complementary to at least a portion of the messenger RNA (mRNA) for the Bt toxin receptor sequences can be constructed. Antisense nucleotides are constructed to hybridize with the corresponding mRNA. Modifications of the antisense sequences may be made as long as the sequences hybridize to and interfere with expression of the corresponding mRNA. In this manner, antisense constructions having 70%, preferably 80%, more preferably 85% sequence similarity to the corresponding antisensed sequences may be used. Furthermore, portions of the antisense nucleotides may be used to disrupt the expression of the target gene. Generally, sequences of at least 50 nucleotides, 100 nucleotides, 200 nucleotides, or greater may be used.

Fragments and variants of the disclosed nucleotide sequences and polypeptides encoded thereby are encompassed by the present invention. By "fragment" is intended a portion of the nucleotide sequence, or a portion of the amino acid sequence, and hence a portion of the polypeptide encoded thereby. Fragments of a nucleotide sequence may encode polypeptide fragments that retain the biological activity of the native polypeptide and, for example, bind Bt toxins. Alternatively, fragments of a nucleotide sequence that are useful as hybridization probes generally do not encode fragment polypeptides retaining biological activity. Thus, frag-

ments of a nucleotide sequence may range from at least about 20 nucleotides, about 50 nucleotides, about 100 nucleotides, and up to the full-length nucleotide sequence encoding the polypeptides of the invention.

A fragment of a Bt toxin receptor nucleotide sequence that encodes a biologically active portion of a Bt toxin receptor polypeptide of the invention will encode at least 15, 25, 30, 50, 100, 150, 200 or 250 contiguous amino acids, or up to the total number of amino acids present in a full-length Bt toxin receptor polypeptide of the invention (for example, 1717, 1730, and 1734 amino acids for SEQ ID NOs: 2, 4, and 6, respectively. Fragments of a Bt toxin receptor nucleotide sequence that are useful as hybridization probes for PCR primers generally need not encode a biologically active portion of a Bt toxin receptor polypeptide.

Thus, a fragment of a Bt toxin receptor nucleotide sequence may encode a biologically active portion of a Bt toxin receptor polypeptide, or it may be a fragment that can be used as a hybridization probe or PCR primer using methods disclosed below. A biologically active portion of a Bt toxin receptor polypeptide can be prepared by isolating a portion of one of the Bt toxin receptor nucleotide sequences of the invention, expressing the encoded portion of the Bt toxin receptor polypeptide (e.g., by recombinant expression *in vitro*), and assessing the activity of the encoded portion of the Bt toxin receptor polypeptide. Nucleic acid molecules that are fragments of a Bt toxin receptor nucleotide sequence comprise at least 16, 20, 50, 75, 100, 150, 200, 250, 300, 350, 400, 450, 500, 550, 600, 650, 700, 800, 900, 1,000, 1,100, 1,200, 1,300, or 1,400 nucleotides, or up to the number of nucleotides present in a full-length Bt toxin receptor nucleotide sequence disclosed herein (for example, 5498, 5527, and 5614 nucleotides for SEQ ID NOs: 1, 3, and 5, respectively).

By "variants" is intended substantially similar sequences. For nucleotide sequences, conservative variants include those sequences that, because of the degeneracy of the genetic code, encode the amino acid sequence of one of the Bt toxin receptor polypeptides of the invention. Naturally occurring allelic variants such as these can be identified with the use of well-known molecular biology techniques, as, for example, with polymerase chain reaction (PCR) and hybridization techniques as outlined below. Variant nucleotide sequences also include synthetically derived nucleotide sequences, such as those generated, for example, by using site-directed mutagenesis, but which still encode a Bt toxin receptor protein of the invention. Generally, variants of a particular nucleotide sequence of the invention will have at least about 40%, 50%, 60%, 65%, 70%, generally at least about 75%, 80%, 85%, preferably at least about 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, and more preferably at least about 98%, 99% or more sequence identity to that particular nucleotide sequence as determined by sequence alignment programs described elsewhere herein using default parameters.

By "variant" protein is intended a protein derived from the native protein by deletion (so-called truncation) or addition of one or more amino acids to the N-terminal and/or C-terminal end of the native protein; deletion or addition of one or more amino acids at one or more sites in the native protein; or substitution of one or more amino acids at one or more sites in the native protein. Variant proteins encompassed by the present invention are biologically active, that is they continue to possess the desired biological activity of the native protein, that is, activity as described herein (for example, Bt toxin binding activity). Such variants may result from, for example, genetic polymorphism or from human manipulation. Biologically active variants of a native Bt toxin receptor protein of the invention will have at least about 40%, 50%, 60%, 65%, 70%,

generally at least about 75%, 80%, 85%, preferably at least about 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, and more preferably at least about 98%, 99% or more sequence identity to the amino acid sequence for the native protein as determined by sequence alignment programs described elsewhere herein using default parameters. A biologically active variant of a protein of the invention may differ from that protein by as few as 1-15 amino acid residues, as few as 1-10, such as 6-10, as few as 5, as few as 4, 3, 2, or even 1 amino acid residue.

The polypeptides of the invention may be altered in various ways including amino acid substitutions, deletions, truncations, and insertions. Methods for such manipulations are generally known in the art. For example, amino acid sequence variants of the Bt toxin receptor polypeptides can be prepared by mutations in the DNA. Methods for mutagenesis and nucleotide sequence alterations are well known in the art. See, for example, Kunkel (1985) *Proc. Natl. Acad. Sci. USA* 82:488-492; Kunkel et al. (1987) *Methods in Enzymol.* 154:367-382; U.S. Pat. No. 4,873,192; Walker and Gaastra, eds. (1983) *Techniques in Molecular Biology* (MacMillan Publishing Company, New York) and the references cited therein. Guidance as to appropriate amino acid substitutions that do not affect biological activity of the protein of interest may be found in the model of Dayhoff et al. (1978) *Atlas of Protein Sequence and Structure* (Natl. Biomed. Res. Found., Washington, D.C.), herein incorporated by reference. Conservative substitutions, such as exchanging one amino acid with another having similar properties, may be preferable.

Thus, the genes and nucleotide sequences of the invention include both the naturally occurring sequences as well as mutant forms. Likewise, the proteins of the invention encompass both naturally occurring proteins as well as variations and modified forms thereof. Such variants will continue to possess the desired toxin binding activity. Obviously, the mutations that will be made in the DNA encoding the variant must not place the sequence out of reading frame and preferably will not create complementary regions that could produce secondary mRNA structure. See, EP Patent Application Publication No. 75,444.

The deletions, insertions, and substitutions of the protein sequences encompassed herein are not expected to produce radical changes in the characteristics of the protein. For example, it is recognized that at least about 10, 20, 50, 100, 150, 200, 250, 300, 350, 400, 450, 500, 550, 600, 650, 700, 750, 800, 850, 900, 950, and up to 960 amino acids may be deleted from the N-terminus of a polypeptide that has the amino acid sequence set forth in SEQ ID NO: 2, and still retain binding function. It is further recognized that at least about 10, 20, 30, 40, 50, 60, 70, 80, 90, 100, 110, and up to 119 amino acids may be deleted from the C-terminus of a polypeptide that has the amino acid sequence set forth in SEQ ID NO: 2, and still retain binding function. Deletion variants of the invention that encompass polypeptides having these deletions. It is recognized that deletion variants of the invention that retain binding function encompass polypeptides having these N-terminal or C-terminal deletions, or having any deletion combination thereof at both the C- and the N-termini.

However, when it is difficult to predict the exact effect of the substitution, deletion, or insertion in advance of doing so, one skilled in the art will appreciate that the effect will be evaluated by routine screening assays. That is, the activity can be evaluated by receptor binding and/or toxicity assays. See, for example, U.S. Pat. No. 5,693,491; T. P. Keeton et al. (1998) *Appl. Environ. Microbiol.* 64(6):2158-2165; B. R. Francis et al. (1997) *Insect Biochem. Mol. Biol.* 27(6):541-550; T. P. Keeton et al. (1997) *Appl. Environ. Microbiol.*

63(9):3419-3425; R. K. Vadlamudi et al. (1995) *J. Biol. Chem.* 270(10):5490-5494; Ihara et al. (1998) *Comparative Biochem. Physiol. B* 120:197-204; Nagamatsu et al. (1998) *Biosci. Biotechnol. Biochem.* 62(4):727-734, herein incorporated by reference.

Variant nucleotide sequences and polypeptides also encompass sequences and polypeptides derived from a mutagenic and recombinogenic procedure such as DNA shuffling. With such a procedure, one or more different toxin receptor coding sequences can be manipulated to create a new toxin receptor, including but not limited to a new Bt toxin receptor, possessing the desired properties. In this manner, libraries of recombinant polynucleotides are generated from a population of related sequence polynucleotides comprising sequence regions that have substantial sequence identity and can be homologously recombined in vitro or in vivo. For example, using this approach, sequence motifs encoding a domain of interest may be shuffled between the Bt toxin receptor gene of the invention and other known Bt toxin receptor genes to obtain a new gene coding for a polypeptide with an improved property of interest, such as an increased ligand affinity in the case of a receptor. Strategies for such DNA shuffling are known in the art. See, for example, Stemmer (1994) *Proc. Natl. Acad. Sci. USA* 91:10747-10751; Stemmer (1994) *Nature* 370:389-391; Cramer et al. (1997) *Nature Biotech.* 15:436-438; Moore et al. (1997) *J. Mol. Biol.* 272:336-347; Zhang et al. (1997) *Proc. Natl. Acad. Sci. USA* 94:4504-4509; Cramer et al. (1998) *Nature* 391:288-291; and U.S. Pat. Nos. 5,605,793 and 5,837,448.

Where the receptor polypeptides of the invention are expressed in a cell and associated with the cell membrane (for example, by a transmembrane segment), in order for the receptor of the invention to bind a desired ligand, for example a Cry 1 A toxin, the receptor's ligand binding domain must be available to the ligand. In this aspect, it is recognized that the native Bt toxin receptor of the invention is oriented such that the toxin binding site is available extracellularly.

Accordingly, in methods comprising use of intact cells, the invention provides cell surface Bt-toxin receptors. By a "cell surface Bt toxin receptor" is intended a membrane-bound receptor polypeptide comprising at least one extracellular Bt toxin binding site. A cell surface receptor of the invention comprises an appropriate combination of signal sequences and transmembrane segments for guiding and retaining the receptor at the cell membrane such that that toxin binding site is available extracellularly. Where native Bt toxin receptors are used for expression, deduction of the composition and configuration of the signal sequences and transmembrane segments is not necessary to ensure the appropriate topology of the polypeptide for displaying the toxin binding site extracellularly. As an alternative to native signal and transmembrane sequences, heterologous signal and transmembrane sequences could be utilized to produce a cell surface receptor polypeptide of the invention.

It is recognized that it may be of interest to generate Bt toxin receptors that are capable of interacting with the receptor's ligands intracellularly in the cytoplasm, in the nucleus or other organelles, in other subcellular spaces; or in the extracellular space. Accordingly, the invention encompasses variants of the receptors of the invention, wherein one or more of the segments of the receptor polypeptide is modified to target the polypeptide to a desired intra- or extracellular location.

Also encompassed by the invention are receptor fragments and variants that are useful, among other things, as binding antagonists that will compete with a cell surface receptor of the invention. Such a fragment or variant can, for example, bind a toxin but not be able to confer toxicity to a particular

cell. In this aspect, the invention provides secreted receptors, more particularly secreted Bt toxin receptors; or receptors that are not membrane bound. The secreted receptors of the invention can contain a heterologous or homologous signal sequence facilitating its secretion from the cell expressing the receptors; and further comprise a secretion variation in the region corresponding to transmembrane segments. By "secretion variation" is intended that amino acids corresponding to a transmembrane segment of a membrane bound receptor comprise one or more deletions, substitutions, insertions, or any combination thereof, such that the region no longer retains the requisite hydrophobicity to serve as a transmembrane segment. Sequence alterations to create a secretion variation can be tested by confirming secretion of the polypeptide comprising the variation from the cell expressing the polypeptide.

The polypeptides of the invention can be purified from cells that naturally express it, purified from cells that have been altered to express it (i.e. recombinant) or synthesized using polypeptide synthesis techniques that are well known in the art. In one embodiment, the polypeptide is produced by recombinant DNA methods. In such methods a nucleic acid molecule encoding the polypeptide is cloned into an expression vector as described more fully herein and expressed in an appropriate host cell according to known methods in the art. The polypeptide is then isolated from cells using polypeptide purification techniques well known to those of ordinary skill in the art. Alternatively, the polypeptide or fragment can be synthesized using peptide synthesis methods well known to those of ordinary skill in the art.

The invention also encompasses fusion polypeptides in which one or more polypeptides of the invention are fused with at least one polypeptide of interest. In one embodiment, the invention encompasses fusion polypeptides in which a heterologous polypeptide of interest has an amino acid sequence that is not substantially homologous to the polypeptide of the invention. In this embodiment, the polypeptide of the invention and the polypeptide of interest may or may not be operatively linked. An example of operative linkage is fusion in-frame so that a single polypeptide is produced upon translation. Such fusion polypeptides can, for example, facilitate the purification of a recombinant polypeptide.

In another embodiment, the fused polypeptide of interest may contain a heterologous signal sequence at the N-terminus facilitating its secretion from specific host cells. The expression and secretion of the polypeptide can thereby be increased by use of the heterologous signal sequence.

The invention is also directed to polypeptides in which one or more domains in the polypeptide described herein are operatively linked to heterologous domains having homologous functions. Thus, the toxin binding domain can be replaced with a toxin binding domain for other toxins. Thereby, the toxin specificity of the receptor is based on a toxin binding domain other than the domain encoded by Bt toxin receptor but other characteristics of the polypeptide, for example, membrane localization and topology is based on Bt toxin receptor.

Alternatively, the native Bt toxin binding domain may be retained while additional heterologous ligand binding domains, including but not limited to heterologous toxin binding domains are comprised by the receptor. Thus, the invention also encompasses fusion polypeptides in which a polypeptide of interest is a heterologous polypeptide comprising a heterologous toxin binding domains. Examples of heterologous polypeptides comprising Cry1 toxin binding domains include, but are not limited to Knight et al (1994) *Mol Micro* 11: 429-436; Lee et al. (1996) *Appl Environ Micro*

63: 2845-2849; Gill et al (1995) *J Biol Chem* 270: 27277-27282; Garczynski et al (1991) *Appl Environ Microbiol* 10: 2816-2820; Vadlamudi et al. (1995) *J Biol Chem* 270(10): 5490-4, U.S. Pat. No. 5,693,491.

The Bt toxin receptor peptide of the invention may also be fused with other members of the cadherin superfamily. Such fusion polypeptides could provide an important reflection of the binding properties of the members of the superfamily. Such combinations could be further used to extend the range of applicability of these molecules in a wide range of systems or species that might not otherwise be amenable to native or relatively homologous polypeptides. The fusion constructs could be substituted into systems in which a native construct would not be functional because of species specific constraints. Hybrid constructs may further exhibit desirable or unusual characteristics otherwise unavailable with the combinations of native polypeptides.

Polypeptide variants encompassed by the present invention include those that contain mutations that either enhance or decrease one or more domain functions. For example, in the toxin binding domain, a mutation may be introduced that increases or decreases the sensitivity of the domain to a specific toxin.

As an alternative to the introduction of mutations, increase in function may be provided by increasing the copy number of ligand binding domains. Thus, the invention also encompasses receptor polypeptides in which the toxin binding domain is provided in more than one copy.

The invention further encompasses cells containing receptor expression vectors comprising the Bt toxin receptor sequences, and fragments and variants thereof. The expression vector can contain one or more expression cassettes used to transform a cell of interest. Transcription of these genes can be placed under the control of a constitutive or inducible promoter (for example, tissue- or cell cycle-preferred).

Where more than one expression cassette utilized, the cassette that is additional to the cassette comprising at least one receptor sequence of the invention, can comprise either a receptor sequence of the invention or any other desired sequences.

The nucleotide sequences of the invention can be used to isolate homologous sequences in insect species other than *ostrinia*, particularly other lepidopteran species, more particularly other *Pyraloidea* species.

The following terms are used to describe the sequence relationships between two or more nucleic acids or polynucleotides: (a) "reference sequence", (b) "comparison window", (c) "sequence identity", (d) "percentage of sequence identity", and (e) "substantial identity".

(a) As used herein, "reference sequence" is a defined sequence used as a basis for sequence comparison. A reference sequence may be a subset or the entirety of a specified sequence; for example, as a segment of a full-length cDNA or gene sequence, or the complete cDNA or gene sequence.

(b) As used herein, "comparison window" makes reference to a contiguous and specified segment of a polynucleotide sequence, wherein the polynucleotide sequence in the comparison window may comprise additions or deletions (i.e., gaps) compared to the reference sequence (which does not comprise additions or deletions) for optimal alignment of the two sequences. Generally, the comparison window is at least 20 contiguous nucleotides in length, and optionally can be 30, 40, 50, 100, or longer. Those of skill in the art understand that to avoid a high similarity to a reference sequence due to inclusion of gaps in the polynucleotide sequence a gap penalty is typically introduced and is subtracted from the number of matches.

Methods of alignment of sequences for comparison are well known in the art. Thus, the determination of percent identity between any two sequences can be accomplished using a mathematical algorithm. Non-limiting examples of such mathematical algorithms are the algorithm of Myers and Miller (1988) *CABIOS* 4:11-17; the local homology algorithm of Smith et al. (1981) *Adv. Appl. Math.* 2:482; the homology alignment algorithm of Needleman and Wunsch (1970) *J. Mol. Biol.* 48:443-453; the search-for-similarity-method of Pearson and Lipman (1988) *Proc. Natl. Acad. Sci.* 85:2444-2448; the algorithm of Karlin and Altschul (1990) *Proc. Natl. Acad. Sci. USA* 87:2264, modified as in Karlin and Altschul (1993) *Proc. Natl. Acad. Sci. USA* 90:5873-5877.

Computer implementations of these mathematical algorithms can be utilized for comparison of sequences to determine sequence identity. Such implementations include, but are not limited to: CLUSTAL in the PC/Gene program (available from Intelligenetics, Mountain View, Calif.); the ALIGN program (Version 2.0); the ALIGN PLUS program (version 3.0, copyright 1997); and GAP, BESTFIT, BLAST, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Version 8 (available from Genetics Computer Group (GCG), 575 Science Drive, Madison, Wis., USA). Alignments using these programs can be performed using the default parameters. The CLUSTAL program is well described by Higgins et al. (1988) *Gene* 73:237-244 (1988); Higgins et al. (1989) *CABIOS* 5:151-153; Corpet et al. (1988) *Nucleic Acids Res.* 16:10881-90; Huang et al. (1992) *CABIOS* 8:155-65; and Pearson et al. (1994) *Meth. Mol. Biol.* 24:307-331. The ALIGN and the ALIGN PLUS programs are based on the algorithm of Myers and Miller (1988) supra. A PAM120 weight residue table, a gap length penalty of 12, and a gap penalty of 4 can be used with the ALIGN program when comparing amino acid sequences. The BLAST programs of Altschul et al (1990) *J. Mol. Biol.* 215:403 are based on the algorithm of Karlin and Altschul (1990) supra. BLAST nucleotide searches can be performed with the BLASTN program, score=100, wordlength=12, to obtain nucleotide sequences homologous to a nucleotide sequence encoding a protein of the invention. BLAST protein searches can be performed with the BLASTX program, score=50, wordlength=3, to obtain amino acid sequences homologous to a protein or polypeptide of the invention. To obtain gapped alignments for comparison purposes, Gapped BLAST (in BLAST 2.0) can be utilized as described in Altschul et al. (1997) *Nucleic Acids Res.* 25:3389. Alternatively, PSI-BLAST (in BLAST 2.0) can be used to perform an iterated search that detects distant relationships between molecules. See Altschul et al. (1997) supra. When utilizing BLAST, Gapped BLAST, PSI-BLAST, the default parameters of the respective programs (e.g., BLASTN for nucleotide sequences, BLASTX for proteins) can be used. See <http://www.ncbi.nlm.nih.gov>. Alignment may also be performed manually by inspection.

Unless otherwise stated, sequence identity/similarity values provided herein refer to the value obtained using GAP Version 10 using the following parameters: % identity using GAP Weight of 50 and Length Weight of 3; % similarity using Gap Weight of 12 and Length Weight of 4, or any equivalent program. By "equivalent program" is intended any sequence comparison program that, for any two sequences in question, generates an alignment having identical nucleotide or amino acid residue matches and an identical percent sequence identity when compared to the corresponding alignment generated by the preferred program.

GAP uses the algorithm of Needleman and Wunsch (1970) *J. Mol. Biol.* 48: 443-453, to find the alignment of two complete sequences that maximizes the number of matches and

minimizes the number of gaps. GAP considers all possible alignments and gap positions and creates the alignment with the largest number of matched bases and the fewest gaps. It allows for the provision of a gap creation penalty and a gap extension penalty in units of matched bases. GAP must make a profit of gap creation penalty number of matches for each gap it inserts. If a gap extension penalty greater than zero is chosen, GAP must, in addition, make a profit for each gap inserted of the length of the gap times the gap extension penalty. Default gap creation penalty values and gap extension penalty values in Version 10 of the Wisconsin Genetics Software Package for protein sequences are 8 and 2, respectively. For nucleotide sequences the default gap creation penalty is 50 while the default gap extension penalty is 3. The gap creation and gap extension penalties can be expressed as an integer selected from the group of integers consisting of from 0 to 200. Thus, for example, the gap creation and gap extension penalties can be 0, 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25, 30, 35, 40, 45, 50, 55, 60, 65 or greater.

GAP presents one member of the family of best alignments. There may be many members of this family, but no other member has a better quality. GAP displays four figures of merit for alignments: Quality, Ratio, Identity, and Similarity. The Quality is the metric maximized in order to align the sequences. Ratio is the quality divided by the number of bases in the shorter segment. Percent Identity is the percent of the symbols that actually match. Percent Similarity is the percent of the symbols that are similar. Symbols that are across from gaps are ignored. A similarity is scored when the scoring matrix value for a pair of symbols is greater than or equal to 0.50, the similarity threshold. The scoring matrix used in Version 10 of the Wisconsin Genetics Software Package is BLOSUM62 (see Henikoff and Henikoff (1989) *Proc. Natl. Acad. Sci. USA* 89:10915).

(c) As used herein, "sequence identity" or "identity" in the context of two nucleic acid or polypeptide sequences makes reference to the residues in the two sequences that are the same when aligned for maximum correspondence over a specified comparison window. When percentage of sequence identity is used in reference to proteins it is recognized that residue positions which are not identical often differ by conservative amino acid substitutions, where amino acid residues are substituted for other amino acid residues with similar chemical properties (e.g., charge or hydrophobicity) and therefore do not change the functional properties of the molecule. When sequences differ in conservative substitutions, the percent sequence identity may be adjusted upwards to correct for the conservative nature of the substitution. Sequences that differ by such conservative substitutions are said to have "sequence similarity" or "similarity". Means for making this adjustment are well known to those of skill in the art. Typically this involves scoring a conservative substitution as a partial rather than a full mismatch, thereby increasing the percentage sequence identity. Thus, for example, where an identical amino acid is given a score of 1 and a non-conservative substitution is given a score of zero, a conservative substitution is given a score between zero and 1. The scoring of conservative substitutions is calculated, e.g., as implemented in the program PC/GENE (Intelligenetics, Mountain View, Calif.).

(d) As used herein, "percentage of sequence identity" means the value determined by comparing two optimally aligned sequences over a comparison window, wherein the portion of the polynucleotide sequence in the comparison window may comprise additions or deletions (i.e., gaps) as compared to the reference sequence (which does not comprise additions or deletions) for optimal alignment of the two

sequences. The percentage is calculated by determining the number of positions at which the identical nucleic acid base or amino acid residue occurs in both sequences to yield the number of matched positions, dividing the number of matched positions by the total number of positions in the window of comparison, and multiplying the result by 100 to yield the percentage of sequence identity.

(e) (i) The term "substantial identity" of polynucleotide sequences means that a polynucleotide comprises a sequence that has at least 70% sequence identity, preferably at least 80%, more preferably at least 90%, and most preferably at least 95%, compared to a reference sequence using one of the alignment programs described using standard parameters. One of skill in the art will recognize that these values can be appropriately adjusted to determine corresponding identity of proteins encoded by two nucleotide sequences by taking into account codon degeneracy, amino acid similarity, reading frame positioning, and the like. Substantial identity of amino acid sequences for these purposes normally means sequence identity of at least 60%, more preferably at least 70%, 80%, 90%, and most preferably at least 95%.

Another indication that nucleotide sequences are substantially identical is if two molecules hybridize to each other under stringent conditions. Generally, stringent conditions are selected to be about 5° C. lower than the thermal melting point (T_m) for the specific sequence at a defined ionic strength and pH. However, stringent conditions encompass temperatures in the range of about 1° C. to about 20° C. lower than the T_m , depending upon the desired degree of stringency as otherwise qualified herein. Nucleic acids that do not hybridize to each other under stringent conditions are still substantially identical if the polypeptides they encode are substantially identical. This may occur, e.g., when a copy of a nucleic acid is created using the maximum codon degeneracy permitted by the genetic code. One indication that two nucleic acid sequences are substantially identical is when the polypeptide encoded by the first nucleic acid sequence is immunologically cross reactive with the polypeptide encoded by the second nucleic acid sequence.

(e) (ii) The term "substantial identity" in the context of a peptide indicates that a peptide comprises a sequence with at least 70% sequence identity to a reference sequence, preferably 80%, more preferably 85%, most preferably at least 90% or 95% sequence identity to the reference sequence over a specified comparison window. Preferably, optimal alignment is conducted using the homology alignment algorithm of Needleman and Wunsch (1970) *J. Mol. Biol.* 48:443-453. An indication that two peptide sequences are substantially identical is that one peptide is immunologically reactive with antibodies raised against the second peptide. Thus, a peptide is substantially identical to a second peptide, for example, where the two peptides differ only by a conservative substitution. Peptides that are "substantially similar" share sequences as noted above except that residue positions that are not identical may differ by conservative amino acid changes.

The nucleotide sequences of the invention can be used to isolate corresponding sequences from other organisms, particularly other insects, more particularly other lepidopteran species. In this manner, methods such as PCR, hybridization, and the like can be used to identify such sequences based on their sequence homology to the sequences set forth herein. Sequences isolated based on their sequence identity to the entire Bt toxin receptor sequences set forth herein or to fragments thereof are encompassed by the present invention. Such sequences include sequences that are orthologs of the disclosed sequences. By "orthologs" is intended genes

derived from a common ancestral gene and which are found in different species as a result of speciation. Genes found in different species are considered orthologs when their nucleotide sequences and/or their encoded protein sequences share substantial identity as defined elsewhere herein. Functions of orthologs are often highly conserved among species.

In a PCR approach, oligonucleotide primers can be designed for use in PCR reactions to amplify corresponding DNA sequences from cDNA or genomic DNA extracted from any organism of interest. Methods for designing PCR primers and PCR cloning are generally known in the art and are disclosed in Sambrook et al. (1989) *Molecular Cloning: A Laboratory Manual* (2d ed., Cold Spring Harbor Laboratory Press, Plainview, N.Y.). See also Innis et al., eds. (1990) *PCR Protocols: A Guide to Methods and Applications* (Academic Press, New York); Innis and Gelfand, eds. (1995) *PCR Strategies* (Academic Press, New York); and Innis and Gelfand, eds. (1999) *PCR Methods Manual* (Academic Press, New York). Known methods of PCR include, but are not limited to, methods using paired primers, nested primers, single specific primers, degenerate primers, gene-specific primers, vector-specific primers, partially-mismatched primers, and the like.

In hybridization techniques, all or part of a known nucleotide sequence is used as a probe that selectively hybridizes to other corresponding nucleotide sequences present in a population of cloned genomic DNA fragments or cDNA fragments (i.e., genomic or cDNA libraries) from a chosen organism. The hybridization probes may be genomic DNA fragments, cDNA fragments, RNA fragments, or other oligonucleotides, and may be labeled with a detectable group such as ^{32}P , or any other detectable marker. Thus, for example, probes for hybridization can be made by labeling synthetic oligonucleotides based on the Bt toxin receptor sequences of the invention. Methods for preparation of probes for hybridization and for construction of cDNA and genomic libraries are generally known in the art and are disclosed in Sambrook et al. (1989) *Molecular Cloning: A Laboratory Manual* (2d ed., Cold Spring Harbor Laboratory Press, Plainview, N.Y.).

For example, the entire Bt toxin receptor sequence disclosed herein, or one or more portions thereof, may be used as a probe capable of specifically hybridizing to corresponding Bt toxin receptor sequences and messenger RNAs. To achieve specific hybridization under a variety of conditions, such probes include sequences that are unique among Bt toxin receptor sequences and are preferably at least about 10 nucleotides in length, and most preferably at least about 20 nucleotides in length. Such probes may be used to amplify corresponding Bt toxin receptor sequences from a chosen plant organism by PCR. This technique may be used to isolate additional coding sequences from a desired organism or as a diagnostic assay to determine the presence of coding sequences in an organism. Hybridization techniques include hybridization screening of plated DNA libraries (either plaques or colonies; see, for example, Sambrook et al. (1989) *Molecular Cloning: A Laboratory Manual* (2d ed., Cold Spring Harbor Laboratory Press, Plainview, N.Y.).

Hybridization of such sequences may be carried out under stringent conditions. By "stringent conditions" or "stringent hybridization conditions" is intended conditions under which a probe will hybridize to its target sequence to a detectably greater degree than to other sequences (e.g., at least 2-fold over background). Stringent conditions are sequence-dependent and will be different in different circumstances. By controlling the stringency of the hybridization and/or washing conditions, target sequences that are 100% complementary to the probe can be identified (homologous probing). Alternatively, stringency conditions can be adjusted to allow some

mismatching in sequences so that lower degrees of similarity are detected (heterologous probing). Generally, a probe is less than about 1000 nucleotides in length, preferably less than 500 nucleotides in length.

Typically, stringent conditions will be those in which the salt concentration is less than about 1.5 M Na ion, typically about 0.01 to 1.0 M Na ion concentration (or other salts) at pH 7.0 to 8.3 and the temperature is at least about 30° C. for short probes (e.g., 10 to 50 nucleotides) and at least about 60° C. for long probes (e.g., greater than 50 nucleotides). Stringent conditions may also be achieved with the addition of destabilizing agents such as formamide. Exemplary low stringency conditions include hybridization with a buffer solution of 30 to 35% formamide, 1 M NaCl, 1% SDS (sodium dodecyl sulphate) at 37° C., and a wash in 1× to 2×SSC (20×SSC=3.0 M NaCl/0.3 M trisodium citrate) at 50 to 55° C. Exemplary moderate stringency conditions include hybridization in 40 to 45% formamide, 1.0 M NaCl, 1% SDS at 37° C., and a wash in 0.5× to 1×SSC at 55 to 60° C. Exemplary high stringency conditions include hybridization in 50% formamide, 1 M NaCl, 1% SDS at 37° C., and a wash in 0.1×SSC at 60 to 65° C. Duration of hybridization is generally less than about 24 hours, usually about 4 to about 12 hours.

Specificity is typically the function of post-hybridization washes, the critical factors being the ionic strength and temperature of the final wash solution. For DNA-DNA hybrids, the T_m can be approximated from the equation of Meinkoth and Wahl (1984) *Anal. Biochem.* 138:267-284: $T_m = 81.5^\circ \text{C.} + 16.6 (\log M) + 0.41 (\% \text{ GC}) - 0.61 (\% \text{ form}) - 500/L$; where M is the molarity of monovalent cations, % GC is the percentage of guanosine and cytosine nucleotides in the DNA, % form is the percentage of formamide in the hybridization solution, and L is the length of the hybrid in base pairs. The T_m is the temperature (under defined ionic strength and pH) at which 50% of a complementary target sequence hybridizes to a perfectly matched probe. T_m is reduced by about 1° C. for each 1% of mismatching; thus, T_m , hybridization, and/or wash conditions can be adjusted to hybridize to sequences of the desired identity. For example, if sequences with $\geq 90\%$ identity are sought, the T_m can be decreased 10° C. Generally, stringent conditions are selected to be about 5° C. lower than the thermal melting point (T_m) for the specific sequence and its complement at a defined ionic strength and pH. However, severely stringent conditions can utilize a hybridization and/or wash at 1, 2, 3, or 4° C. lower than the thermal melting point (T_m); moderately stringent conditions can utilize a hybridization and/or wash at 6, 7, 8, 9, or 10° C. lower than the thermal melting point (T_m); low stringency conditions can utilize a hybridization and/or wash at 11, 12, 13, 14, 15, or 20° C. lower than the thermal melting point (T_m). Using the equation, hybridization and wash compositions, and desired T_m , those of ordinary skill will understand that variations in the stringency of hybridization and/or wash solutions are inherently described. If the desired degree of mismatching results in a T_m of less than 45° C. (aqueous solution) or 32° C. (formamide solution), it is preferred to increase the SSC concentration so that a higher temperature can be used. An extensive guide to the hybridization of nucleic acids is found in Tijssen (1993) *Laboratory Techniques in Biochemistry and Molecular Biology—Hybridization with Nucleic Acid Probes*, Part I, Chapter 2 (Elsevier, N.Y.); and Ausubel et al., eds. (1995) *Current Protocols in Molecular Biology*, Chapter 2 (Greene Publishing and Wiley-Interscience, New York). See Sambrook et al. (1989) *Molecular Cloning: A Laboratory Manual* (2d ed., Cold Spring Harbor Laboratory Press, Plainview, N.Y.).

Thus, isolated sequences that encode for a Bt toxin receptor protein and which hybridize under stringent conditions to the Bt toxin receptor sequences disclosed herein, or to fragments thereof, are encompassed by the present invention. Such sequences will be at least about 40% to 50% homologous, about 60%, 65%, or 70% homologous, and even at least about 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or more homologous with the disclosed sequences. That is, the sequence identity of sequences may range, sharing at least about 40% to 50%, about 60%, 65%, or 70%, and even at least about 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or more sequence identity.

The compositions and screening methods of the invention are useful for identifying cells expressing the BT toxin receptors of the invention, and variants and homologues thereof. Such identification could utilize detection methods at the protein level, such as ligand-receptor binding; or at the nucleotide level. Detection of the polypeptide could be in situ by means of in situ hybridization of tissue sections but may also be analyzed by bulk polypeptide purification and subsequent analysis by Western blot or immunological assay of a bulk preparation. Alternatively, receptor gene expression can be detected at the nucleic acid level by techniques well known to those of ordinary skill in any art using complimentary polynucleotides to assess the levels of genomic DNA, mRNA, and the like. As an example, PCR primers complimentary to the nucleic acid of interest can be used to identify the level of expression. Tissues and cells identified as expressing the receptor sequences of the invention are determined to be susceptible to toxins which bind the receptor polypeptides.

Where the source of the cells identified to express the receptor polypeptides of the invention is an organism, for example an insect plant pest, the organism is determined to be susceptible to toxins capable of binding the polypeptides. In a particular embodiment, identification is in a lepidopteran plant pest expressing the Bt toxin receptor of the invention.

The invention encompasses antibody preparations with specificity against the polypeptides of the invention. In further embodiments of the invention, the antibodies are used to detect receptor expression in a cell.

In one aspect, the invention is particularly drawn to compositions and methods for modulating susceptibility of plant pests to Bt toxins. However, it is recognized that the methods and compositions could be used for modulating susceptibility of any cell or organism to the toxins. By "modulating" is intended that the susceptibility of a cell or organism to the cytotoxic effects of the toxin is increased or decreased. By "susceptibility" is intended that the viability of a cell contacted with the toxin is decreased. Thus the invention encompasses expressing the cell surface receptor polypeptides of the invention to increase susceptibility of a target cell or organ to Bt toxins. Such increases in toxin susceptibility are useful for medical and veterinary purposes in which eradication or reduction of viability of a group of cells is desired. Such increases in susceptibility are also useful for agricultural applications in which eradication or reduction of population of particular plant pests is desired.

Plant pests of interest include, but are not limited to insects, nematodes, and the like. Nematodes include parasitic nematodes such as root-knot, cyst, lesion, and renniform nematodes, etc.

The following examples are offered by way of illustration and not by way of limitation.

Example 1

Isolation of EC Bt Toxin Receptor

Standard recombinant methods well known to those of ordinary skill in the art were carried out. For library construction, total RNA was isolated from the midgut of European corn borer (ECB), *Ostrinia nubilalis*. Corn borer larvae (for example, a mix of stage 2, 3, and 4, equal weight) can be pulverized in liquid nitrogen, homogenized, and total RNA extracted by standard procedures. PolyA RNA can be isolated from the total RNA with standard PolyA isolation procedures, such as the PolyATact system from Promega Corporation, Madison, Wis. cDNA synthesis can then be performed and, for example, unidirectional cDNA libraries can be constructed according to known and commercial procedures, such as the ZAP Express cDNA synthesis kit from Stratagene, La Jolla, Calif. cDNA can be amplified by PCR, sized and properly digested with restriction fragments to be ligated into a vector. Subcloned cDNA can be sequenced to identify sequences with the proper peptide to identify corresponding to published sequences. These fragments can be used to probe genomic or cDNA libraries corresponding to a specific host, such as *Ostrinia nubilalis*, to obtain a full length coding sequence. Probes can also be made based on Applicants disclosed sequences. The coding sequence can then be ligated into a desired expression cassette and used to transform a host cell according to standard transformation procedures. Such an expression cassette can be part of a commercially available vector and expression system; for example, the pET system from Novagen Inc. (Madison, Wis.). Additional vectors that can be used for expression include pBKCMV, pBKRSV, pPbac and pMbac (Stratagene Inc.), pFASTBac1 (Gibco BRL) and other common bacterial, baculovirus, mammalian, and yeast expression vectors.

All vectors were constructed using standard molecular biology techniques as described for example in Sambrook et al., (1989) *Molecular Cloning: A Laboratory Manual* (2nd ed., Cold Spring Harbor Laboratory: Cold Spring Harbor, N.Y.).

Expression is tested by ligand blotting and testing for Bt toxin binding. Ligand blotting, binding, and toxicity are tested by known methods; for example, as described in Martinez-Ramirez (1994) *Biochem. Biophys. Res. Comm.* 201: 782-787; Vadlamudi et al. (1995) *J Biol Chem* 270(10):5490-4, Keeton et al. (1998) *Appl Environ Microbiol* 64(6):2158-2165; Keeton et al. (1997) *Appl Environ Microbiol* 63(9): 3419-3425; Ihara et al. (1998) *Comparative Biochemistry and Physiology, Part B* 120:197-204; Nagamatsu et al. (1998) *Biosci. Biotechnol. Biochem.* 62(4):718-726 and Nagamatsu et al. (1998) *Biosci. Biotechnol. Biochem.* 62(4):727-734.

Identifying the Cry1A(b) binding polypeptide in ECB was done by ligand blotting brush border membrane vesicle polypeptides and probing those polypeptides for binding with Cry1A(b) toxin. Two polypeptides, approximately 210 and 205 kDa, were found to bind to Cry1A(b). Blotting and binding were done essentially as described in the preceding paragraph.

Degenerate primers for RT-PCR were designed based on known Cry1 toxin binding polypeptide sequences from *Manduca sexta* and *Bombyx mori*. The primers are shown below. cDNA was constructed from total midgut RNA (cDNA synthesis kit GibcoBrL). Degenerate primers were used to amplify products of the expected size. The annealing

23

temperature used was 53° C. in generation of the 280 bp fragment and 55° C. when generating the 1.6 kb fragment.

A 280 bp fragment was obtained from ECB midgut RNA. Upon cloning and sequencing, the fragment was identified as having homology with the Bt toxin receptor 1 polypeptide (BTR1) described in Vadlamudi et al. (1995) *J Biol Chem* 270(10):5490-4.

A similar approach was used to generate a 1.6 kilobase pair clone. The sequence of primers used to generate the 280 base pair fragment were:

Primer BTRD1S: 5'GTTAMYGTGAGAGAGGCA-GAYCC3' (SEQ ID NO: 8), and

Primer BTRD5A: 5'GGATRTTAAGMGTCAGY-ACWCCG3' (SEQ ID NO: 9).

The sequence of primers used to generate the 1.6 kb fragment were:

Primer BTRD6S: 5'TCCGAATTCTTCTTYAACCTCATC-GAYAACTT3' (SEQ ID NO: 10), and

Primer BTRD7A: 5'CGCAAGCTTACTTGGTCGATGT-TRCASGTCAT3' (SEQ ID NO: 11)

The 1.6 kb fragment clone was ligated in an *E. coli* expression vector, pET-28a-c(+), and expressed using the pET system (Novagen Inc., Madison, Wis.). Purified polypeptide encoded by this 1.6 kb fragment demonstrated binding to Cry1A(b) in ligand blots. An ECB midgut cDNA library was generated and screened using this 1.6 kb clone, generating 120 positive plaques. Thirty of these plaques were chosen for secondary screening and fifteen of those plaques were purified and sent for DNA sequencing.

The obtained nucleotide sequence of the selected Bt toxin receptor clone from ECB is set forth in SEQ ID NO: 1. The total length of the clone is 5498 base pairs. The coding sequences are residues 162-5312. The Cry1A binding site is encoded by residues 4038-4547. The predicted transmembrane domain is encoded by residues 4872-4928. The corresponding deduced amino acid sequence for this Bt toxin receptor clone from ECB is set forth in SEQ ID NO: 2.

The purified polypeptide generated from the 1.6 kb fragment set forth in SEQ ID NO: 7 was used to inoculate rabbits for the production of polyclonal antibodies. On zoo western blots prepared from brush border membrane vesicles from various insect species, this set of antibodies specifically recognized ECB Bt toxin receptor polypeptides, in comparison to Bt toxin receptor homologues polypeptides from other insect species. Rabbit polyclonal antibodies were also raised from a purified polypeptide corresponding to amino acids 1293-1462 of SEQ ID NO: 2.

Example 2

Isolation of CEW and FAW Bt Toxin Receptor Orthologues

cDNA encoding a full-length Bt toxin receptor from corn earworm (CEW, *Heliothis Zea*) was isolated. The nucleotide sequence for this cDNA is set forth in SEQ ID NO: 3. Nucleotides 171-5360 correspond to the open reading frame. Nucleotides 4917-4973 correspond to the transmembrane region. Nucleotides 4083-4589 correspond to the Cry1A binding site. The deduced corresponding amino acid sequence for the CEW Bt toxin receptor is set forth in SEQ ID NO: 4.

cDNA encoding a full-length Bt toxin receptor from fall armyworm (FAW, *Spodoptera frugiperda*) was isolated. The nucleotide sequence for this cDNA is set forth in SEQ ID NO: 5. Nucleotides 162-5363 correspond to the open reading frame. Nucleotides 4110-4616 correspond to the Cry1A

24

binding site. Nucleotides 4941-4997 correspond to the transmembrane region. Nucleotides 162-227 correspond to a signal peptide. The deduced corresponding amino acid sequence for the FAW Bt toxin receptor is set forth in SEQ ID NO: 6.

Example 3

Binding and Cell Death in Lepidopteran Insect Cells Expressing the Bt Toxin Receptors of the Invention

An in vitro system is developed to demonstrate the functionality of a Bt toxin receptor of the invention. The results disclosed in this example demonstrate that the ECB Bt toxin receptor of the invention (SEQ ID NOs: 1 and 2) is specifically involved in the binding and killing action of Cry1Ab toxin.

Well known molecular biological methods are used in cloning and expressing the ECB Bt toxin receptor in Sf9 cells. A baculovirus expression system (Gibco BRL Catalogue No. 10359-016) is used according to the manufacturer's provided protocols and as described below. *S. frugiperda* (Sf9) cells obtained from ATCC (ATCC-CRL 1711) are grown at 27° C. in Sf-900 II serum free medium (Gibco BRL, Catalogue No. 10902-088). These cells, which are not susceptible to Cry1Ab toxin, are transfected with an expression construct (pFastBac1 bacmid, Gibco BRL catalogue NO. 10360-014) comprising an operably linked Bt toxin receptor of the invention (SEQ ID NO: 1) downstream of a polyhedrin promoter. Transfected Sf9 cells express the ECB Bt toxin receptor and are lysed in the presence of Cry1Ab toxin. Toxin specificities, binding parameters, such as Kd values, and half maximal doses for cellular death and/or toxicity are also determined.

For generating expression constructs, the ECB Bt toxin receptor cDNA (SEQ ID NO: 1) is subjected to appropriate restriction digestion, and the resulting cDNA comprising the full-length coding region is ligated into the donor plasmid pFastBac1 multiple cloning site. Following transformation and subsequent transposition, recombinant bacmid DNA comprising the ECB Bt toxin receptor (RBECB1) is isolated. As a control, recombinant bacmid DNA comprising the reporter gene β -glucuronidase (RBGUS) is similarly constructed and isolated.

For transfection, 2 μ g each RBECB1 or RBGUS DNA is mixed with 6 μ l of CellFectin (GibcoBRL catalogue NO. 10362-010) in 100 μ l of Sf900 medium, and incubated at room temperature for 30 minutes. The mixture is then diluted with 0.8 ml Sf-900 medium. Sf9 cells (10^6 /ml per 35 mm well) are washed once with Sf-900 medium, mixed with the DNA/CellFectin mixture, added to the well, and incubated at room temperature for 5 hours. The medium is removed and 2 ml of Sf-900 medium containing penicillin and streptomycin is added to the well. 3-5 days after transfection, Western blotting is used to examine protein expression.

For Western blotting, 100 μ l of cell lysis buffer (50 mM Tris, pH7.8, 150 mM NaCl, 1% Nonidet P-40) is added to the well. The cells are scraped and subjected to 16,000 \times g centrifugation. Pellet and supernatant are separated and subjected to Western blotting. An antibody preparation against ECB Bt toxin receptor (Example 1) is used as first antibody. Alkaline phosphatase-labelled anti-rabbit IgG is used as secondary antibody. Western blot results indicate that the full length ECB Bt toxin receptor of the invention (SEQ ID NOs: 1 and 2) is expressed in the cell membrane of these cells.

For determining GUS activity, the medium of the cells transfected with RBGUS is removed. The cells and the medium are separately mixed with GUS substrate and

assayed for the well known enzymatic activity. GUS activity assays indicate that this reporter gene is actively expressed in the transfected cells.

For determining toxin susceptibility, Cry toxins including but not limited to Cry1A, Cry1B, Cry1C, Cry1D, Cry1E, Cry1F, Cry1I, Cry2, Cry3, and Cry9 toxins (Schnepf E. et al. (1998) *Microbiology and Molecular Biology Reviews* 62(3): 775-806) are prepared by methods known in the art. Crystals are dissolved in pH 10.0, 50 mM carbonate buffer and treated with trypsin. Active fragments of Cry proteins are purified by chromatography. Three to five days after transfection, cells are washed with phosphate buffered saline (PBS). Different concentrations of active fragments of Cry toxins are applied to the cells. At different time intervals, the cells are examined under the microscope to readily determine susceptibility to the toxins. Alternatively, cell death, viability and/or toxicity is quantified by methods well known in the art. See, for example, In Situ Cell Death Detection Kits available from Roche Biochemicals (Catalogue Nos. 2 156 792, 1 684 809, and 1 684 817), and LIVE/DEAD® Viability/Cytotoxicity Kit available from Molecular Probes (catalogue No. L-3224).

A dose-dependent response of RBECB1-transfected cells to Cry1Ab is readily observed, with determined Kd values well within the range for many receptors. Control cells, e.g. those transfected with pFastBac1 bacmid without an insert or those transfected with RBGus are not significantly affected by Cry1Ab. Interaction with other Cry toxins are similarly characterized.

This in vitro system is not only be used to verify the functionality of putative Bt-toxin receptors, but also used as a tool to determine the active site(s) and other functional domains of the toxin and the receptor. Furthermore, the system is used as a cell-based high throughput screen. For example, methods for distinguishing live versus dead cells by differential dyes are known in the art. This allows for aliquots of transfected cells to be treated with various toxin samples and to serve as a means for screening the toxin samples for desired specificity or binding characteristics. Since the system is used to identify the specificity of Cry protein receptors, it is a useful tool in insect resistance management.

Example 4

Expression of the ECB Bt Toxin Receptor in Toxin Susceptible Stages of the Insect's Life Cycle

Total RNA was isolated from the eggs, pupae, adults, and the 1st through the 5th instar developmental stages, using TRIzol Reagent (Gibco BRL) essentially as instructed by the manufacturer. (Gibco BRL). The RNA was quantitated and 20 ug of each sample was loaded onto a formaldehyde agarose gel and electrophoresed at constant voltage. The RNA was then transferred to a nylon membrane via neutral capillary transfer and cross-linked to the membrane using ultraviolet light. For hybridization, a 460 base pair ECB Bt toxin receptor DNA probe (bases 3682 to 4141 in SEQ ID NO: 1) was constructed from a 460 base pair fragment prepared according to the manufacturer's protocol for Amersham Rediprime II random prime labeling system. The denatured probe was added to the membrane that had been prehybridized for at least 3 hours at 65° C. and allowed to incubate with gentle agitation for at least 12 hours at 65° C. Following hybridization, the membranes were washed at 65° C. for 1 hour with 1/4x0.5M NaCl, 0.1M NaPO4 (ph 7.0), 6 mM EDTA and 1% SDS solution followed by two 1 hour washes

in the above solution without SDS. The membrane was air dried briefly, wrapped in Saran Wrap and exposed to X-ray film.

An ECB Bt toxin receptor transcript of 5.5 kilobase was expressed strongly in the larval instars with much reduced expression in the pupal stage. The expression levels appeared to be fairly consistent from first to fifth instar, while decreasing markedly in the pupal stage. There were no detectable transcripts in either the egg or adult stages. These results indicate that the ECB Bt toxin transcript is being produced in the susceptible stages of the insects life cycle, while not being produced in stages resistant to the toxic effects of Cry1Ab.

Example 5

Tissue and Subcellular Expression of the ECB Bt Toxin Receptor

Fifth instar ECB were dissected to isolate the following tissues: fat body (FB), malpighian tubules (MT), hind gut (HG), anterior midgut (AM) and posterior midgut (PM). Midguts from fifth instar larvae were also isolated for brush border membrane vesicle (BBMV) preparation using the well known protocol by Wolfersberger et al. (1987) *Comp. Biochem. Physiol.* 86A:301-308. Tissues were homogenized in Tris buffered saline, 0.1% tween-20, centrifuged to pellet insoluble material, and transferred to a fresh tube. 50 ug of protein from each preparation was added to SDS sample buffer and B-mercaptoethanol, heated to 100° C. for 10 minutes and loaded onto a 4-12% Bis-Tris gel (Novex). After electrophoresis, the proteins were transferred to a nitrocellulose membrane using a semi-dry apparatus. The membrane was blocked in 5% nonfat dry milk buffer for 1 hour at room temperature with gentle agitation. The primary antibody (Example 1) was added to a final dilution of 1:5000 and allowed to hybridize for 1 hour. The blot was then washed three times for 20 minutes each in nonfat milk buffer. The blot was then hybridized with the secondary antibody (goat anti-rabbit with alkaline phosphatase conjugate) at a dilution of 1:10000 for 1 hour at room temperature. Washes were performed as before. The bands were visualized by using the standard chemiluminescent protocol (Tropix western light protein detection kit).

The ECB Bt toxin receptor protein was only visible in the BBMV enriched lane, and not detected in any of the other ECB tissues types. This result indicates that the expression of the ECB Bt toxin receptor protein is at very low levels, since the BBMV preparation is a 20-30 fold enriched fraction of the midgut brush border. The result supports propositions that the ECB Bt toxin receptor is an integral membrane protein uniquely associated with the brush border. It also demonstrates that the ECB Bt toxin receptor is expressed in the envisioned target tissue for Cry1Ab toxins. However, the result does not necessarily rule out expression in other tissue types, albeit the expression of this protein in those tissues may be lower than in the BBMV enriched fraction.

All publications and patent applications mentioned in the specification are indicative of the level of those skilled in the art to which this invention pertains. All publications and patent applications are herein incorporated by reference to the same extent as if each individual publication or patent application was specifically and individually indicated to be incorporated by reference.

Although the foregoing invention has been described in some detail by way of illustration and example for purposes of clarity of understanding, it will be obvious that certain changes and modifications may be practiced within the scope of the appended claims.

SEQUENCE LISTING

<160> NUMBER OF SEQ ID NOS: 11

<210> SEQ ID NO 1

<211> LENGTH: 5498

<212> TYPE: DNA

<213> ORGANISM: *Ostrinia nubilalis*

<220> FEATURE:

<221> NAME/KEY: CDS

<222> LOCATION: (162)...(5312)

<400> SEQUENCE: 1

```

cataataaca ataaagagga agtgtgtgtg aaaaacgaag aagttaataa acctggataa      60
ttaaacctga aaaaaccgg tgtttaagtg gaatttttgc tgaaggacaa ccgtgggata      120
gctcaaatat taaaattcta cataactaag gatcatgcaa a atg ggg gtt gag agg      176
                                     Met Gly Val Glu Arg
                                     1           5

ttc ttc cca gca gtg cta ctg gtc tct tta gcc tct gcc gca cta gcc      224
Phe Phe Pro Ala Val Leu Leu Val Ser Leu Ala Ser Ala Ala Leu Ala
                                     10           15           20

aac caa cga tgt tcg tac att atc gca ata cca aga ccg gag act ccg      272
Asn Gln Arg Cys Ser Tyr Ile Ile Ala Ile Pro Arg Pro Glu Thr Pro
                                     25           30           35

gaa ctg ccg cct att gat tac gaa gga aaa tca tgg agt gaa cag cct      320
Glu Leu Pro Pro Ile Asp Tyr Glu Gly Lys Ser Trp Ser Glu Gln Pro
                                     40           45           50

cta ata ccc ggc ccg acc cga gag gaa gta tgt atg gag aac ttc tta      368
Leu Ile Pro Gly Pro Thr Arg Glu Glu Val Cys Met Glu Asn Phe Leu
                                     55           60           65

ccg gat caa atg att cag gtc ata tac atg gag gaa gaa atc gaa gga      416
Pro Asp Gln Met Ile Gln Val Ile Tyr Met Glu Glu Glu Ile Glu Gly
                                     70           75           80           85

gac gtc atc att gcg aag ctt aac tat caa ggg tcc aac acg ccg gtg      464
Asp Val Ile Ile Ala Lys Leu Asn Tyr Gln Gly Ser Asn Thr Pro Val
                                     90           95           100

ctg tcg att atg tca ggc cag ccc aga gcc cag ctg ggc cct gag ttt      512
Leu Ser Ile Met Ser Gly Gln Pro Arg Ala Gln Leu Gly Pro Glu Phe
                                     105           110           115

cga cag aat gaa gca gac ggc caa tgg agc ctt gtt att acg caa aga      560
Arg Gln Asn Glu Ala Asp Gly Gln Trp Ser Leu Val Ile Thr Gln Arg
                                     120           125           130

caa gac tac gag aca gca acc atg cag agc tat gtg ttc tca atc caa      608
Gln Asp Tyr Glu Thr Ala Thr Met Gln Ser Tyr Val Phe Ser Ile Gln
                                     135           140           145

gtg gag ggt gaa tca cag gcc gta ctg gtg gcg ctg gag ata gtc aac      656
Val Glu Gly Glu Ser Gln Ala Val Leu Val Ala Leu Glu Ile Val Asn
                                     150           155           160           165

atc gac gac aat ccg ccc atc ctg caa gtg gtc agc gcc tgc gta att      704
Ile Asp Asp Asn Pro Pro Ile Leu Gln Val Val Ser Ala Cys Val Ile
                                     170           175           180

cca gaa cat ggc gag gct aga ctg acc gac tgc gtg tac caa gtg tca      752
Pro Glu His Gly Glu Ala Arg Leu Thr Asp Cys Val Tyr Gln Val Ser
                                     185           190           195

gac cgc gac ggt gaa atc agc acc cgc ttc atg acg ttc cgt gtc gac      800
Asp Arg Asp Gly Glu Ile Ser Thr Arg Phe Met Thr Phe Arg Val Asp
                                     200           205           210

agc agc agg gct gca gat gaa agc atc ttc tac atg gtt gga gaa tac      848
Ser Ser Arg Ala Ala Asp Glu Ser Ile Phe Tyr Met Val Gly Glu Tyr
                                     215           220           225

gac ccc agc gac tgg ttc aat atg aag atg act gtg ggg atc aat tcg      896

```


-continued

Gly	Asn	Ala	Val	Asp	Tyr	Leu	Phe	Ile	Asp	Glu	Ser	Thr	Gly	Glu	Ile		
550					555					560					565		
ttc	gtg	agc	atg	gac	gat	gcc	ttc	gac	tac	cac	cga	cag	aac	act	cta		1904
Phe	Val	Ser	Met	Asp	Asp	Ala	Phe	Asp	Tyr	His	Arg	Gln	Asn	Thr	Leu		
				570					575					580			
ttt	gtt	cag	gtg	cgc	gct	gac	gat	act	ttg	ggc	gac	ggc	cca	cac	aac		1952
Phe	Val	Gln	Val	Arg	Ala	Asp	Asp	Thr	Leu	Gly	Asp	Gly	Pro	His	Asn		
			585					590					595				
aca	gtg	acc	acc	cag	ctg	gtg	ata	gaa	ctg	gag	gat	gtc	aac	aac	act		2000
Thr	Val	Thr	Thr	Gln	Leu	Val	Ile	Glu	Leu	Glu	Asp	Val	Asn	Asn	Thr		
		600					605					610					
cct	ccc	acc	cta	cgc	ttg	ccc	cgt	tcg	act	cca	agc	gtc	gag	gag	aac		2048
Pro	Pro	Thr	Leu	Arg	Leu	Pro	Arg	Ser	Thr	Pro	Ser	Val	Glu	Glu	Asn		
	615					620					625						
gtt	ccc	gaa	gga	tac	gag	ata	tcc	cgg	gaa	atc	act	gct	acc	gac	ccg		2096
Val	Pro	Glu	Gly	Tyr	Glu	Ile	Ser	Arg	Glu	Ile	Thr	Ala	Thr	Asp	Pro		
630					635					640					645		
gac	acc	agc	gcc	tac	ctg	tgg	ttc	gag	atc	gac	tgg	gac	tcc	acc	tgg		2144
Asp	Thr	Ser	Ala	Tyr	Leu	Trp	Phe	Glu	Ile	Asp	Trp	Asp	Ser	Thr	Trp		
				650						655					660		
gcc	acc	aag	cag	ggc	aga	gag	acc	aac	cct	act	gaa	tac	gtc	ggg	tgt		2192
Ala	Thr	Lys	Gln	Gly	Arg	Glu	Thr	Asn	Pro	Thr	Glu	Tyr	Val	Gly	Cys		
			665					670					675				
ata	gtt	atc	gaa	acg	ata	tac	ccc	acc	gag	ggc	aac	cgg	ggt	tcc	gcc		2240
Ile	Val	Ile	Glu	Thr	Ile	Tyr	Pro	Thr	Glu	Gly	Asn	Arg	Gly	Ser	Ala		
		680					685						690				
atc	ggg	cgc	ctc	gtg	gtg	caa	gag	atc	cgg	gac	aac	gtc	acc	atc	gac		2288
Ile	Gly	Arg	Leu	Val	Val	Gln	Glu	Ile	Arg	Asp	Asn	Val	Thr	Ile	Asp		
	695					700					705						
ttc	gag	gaa	ttc	gag	atg	ctt	tac	ctc	acc	gtc	cgc	gtg	agg	gac	ctc		2336
Phe	Glu	Glu	Phe	Glu	Met	Leu	Tyr	Leu	Thr	Val	Arg	Val	Arg	Asp	Leu		
710					715					720					725		
aac	act	gtc	atc	gga	gat	gac	tac	gat	gag	gcg	acg	ttc	acg	atc	aca		2384
Asn	Thr	Val	Ile	Gly	Asp	Asp	Tyr	Asp	Glu	Ala	Thr	Phe	Thr	Ile	Thr		
				730					735					740			
ata	atc	gac	atg	aac	gac	aac	gcg	ccg	atc	ttc	gcg	aac	ggc	acg	ctg		2432
Ile	Ile	Asp	Met	Asn	Asp	Asn	Ala	Pro	Ile	Phe	Ala	Asn	Gly	Thr	Leu		
			745					750					755				
acg	cag	acg	atg	cgc	gtg	cgc	gag	ctg	gcg	gcc	agc	ggc	acg	ctc	atc		2480
Thr	Gln	Thr	Met	Arg	Val	Arg	Glu	Leu	Ala	Ala	Ser	Gly	Thr	Leu	Ile		
		760					765					770					
ggc	tcc	gtg	ctc	gcc	acc	gac	atc	gac	ggc	ccg	ctc	tac	aac	caa	gtg		2528
Gly	Ser	Val	Leu	Ala	Thr	Asp	Ile	Asp	Gly	Pro	Leu	Tyr	Asn	Gln	Val		
		775				780					785						
cgc	tac	act	ata	caa	cct	aga	aac	aac	act	ccc	gag	gga	tta	gtg	aag		2576
Arg	Tyr	Thr	Ile	Gln	Pro	Arg	Asn	Asn	Thr	Pro	Glu	Gly	Leu	Val	Lys		
790					795					800					805		
att	gac	ttc	aca	act	ggt	caa	att	gag	gtg	gat	gcg	aac	gag	gcg	atc		2624
Ile	Asp	Phe	Thr	Thr	Gly	Gln	Ile	Glu	Val	Asp	Ala	Asn	Glu	Ala	Ile		
				810					815					820			
gat	gca	gac	gaa	ccc	tgg	cgc	ttc	tac	ttg	tac	tac	acc	gtc	atc	gct		2672
Asp	Ala	Asp	Glu	Pro	Trp	Arg	Phe	Tyr	Leu	Tyr	Tyr	Thr	Val	Ile	Ala		
			825					830					835				
agc	gac	gag	tgc	tcc	ctg	gaa	aac	cgc	acg	gaa	tgt	cct	cca	gat	tcc		2720
Ser	Asp	Glu	Cys	Ser	Leu	Glu	Asn	Arg	Thr	Glu	Cys	Pro	Pro	Asp	Ser		
		840					845					850					
aac	tac	ttc	gaa	gtt	cca	ggc	gat	atc	gaa	ata	gaa	atc	atc	gac	aca		2768
Asn	Tyr	Phe	Glu	Val	Pro	Gly	Asp	Ile	Glu	Ile	Glu	Ile	Ile	Asp	Thr		
	855					860					865						
aac	aac	aaa	gtg	cct	gag	ccg	ctc	act	gag	aag	ttc	aac	acg	acg	gtg		2816

-continued

Asn	Asn	Lys	Val	Pro	Glu	Pro	Leu	Thr	Glu	Lys	Phe	Asn	Thr	Thr	Val		
870					875					880					885		
tac	gtc	tgg	gag	aat	gcc	acg	agc	ggc	gac	gag	gtg	gtc	cag	ctg	tac		2864
Tyr	Val	Trp	Glu	Asn	Ala	Thr	Ser	Gly	Asp	Glu	Val	Val	Gln	Leu	Tyr		
				890					895					900			
tcc	cac	gac	cgt	gac	aga	gac	gag	ttg	tac	cac	acg	gta	cga	tac	acg		2912
Ser	His	Asp	Arg	Asp	Arg	Asp	Glu	Leu	Tyr	His	Thr	Val	Arg	Tyr	Thr		
			905					910					915				
atg	aac	ttt	gcg	gtg	aac	ccc	cga	ctg	cgg	gat	ttc	ttc	gag	gtg	gac		2960
Met	Asn	Phe	Ala	Val	Asn	Pro	Arg	Leu	Arg	Asp	Phe	Phe	Glu	Val	Asp		
		920					925					930					
ctg	gac	act	ggg	cgc	ctt	gag	gtg	cat	tac	ccg	ggg	gac	gaa	aaa	ttg		3008
Leu	Asp	Thr	Gly	Arg	Leu	Glu	Val	His	Tyr	Pro	Gly	Asp	Glu	Lys	Leu		
	935					940					945						
gac	cgc	gat	ggg	gat	gag	cct	aca	cat	act	atc	ttt	gta	aat	ttc	atc		3056
Asp	Arg	Asp	Gly	Asp	Glu	Pro	Thr	His	Thr	Ile	Phe	Val	Asn	Phe	Ile		
	950				955					960					965		
gat	aac	ttc	ttt	tct	gat	ggg	gac	ggg	agg	aga	aac	cag	gac	gaa	ggt		3104
Asp	Asn	Phe	Phe	Ser	Asp	Gly	Asp	Gly	Arg	Arg	Asn	Gln	Asp	Glu	Val		
				970					975					980			
gaa	ata	ttt	gtc	ggt	cta	ttg	gat	gtg	aac	gac	aac	gct	cct	gag	atg		3152
Glu	Ile	Phe	Val	Val	Leu	Leu	Asp	Val	Asn	Asp	Asn	Ala	Pro	Glu	Met		
			985					990					995				
cca	ttg	cct	gat	gaa	ctc	cgg	ttt	gat	ggt	tcc	gaa	gga	gca	ggt	gct		3200
Pro	Leu	Pro	Asp	Glu	Leu	Arg	Phe	Asp	Val	Ser	Glu	Gly	Ala	Val	Ala		
		1000					1005					1010					
ggg	gtc	cgt	gta	ctc	cca	gaa	atc	tac	gca	ccg	gac	agg	gat	gaa	cca		3248
Gly	Val	Arg	Val	Leu	Pro	Glu	Ile	Tyr	Ala	Pro	Asp	Arg	Asp	Glu	Pro		
	1015					1020					1025						
gac	acg	gac	aac	tcg	cgt	gtc	ggg	tac	gga	atc	ctg	gac	ctc	acg	atc		3296
Asp	Thr	Asp	Asn	Ser	Arg	Val	Gly	Tyr	Gly	Ile	Leu	Asp	Leu	Thr	Ile		
	1030				1035					1040					1045		
acc	gac	cga	gac	atc	gag	gtg	ccg	gat	ctc	ttc	acc	atg	atc	tcg	att		3344
Thr	Asp	Arg	Asp	Ile	Glu	Val	Pro	Asp	Leu	Phe	Thr	Met	Ile	Ser	Ile		
				1050					1055					1060			
gaa	aac	aaa	act	ggg	gaa	ctt	gag	acc	gct	atg	gac	ttg	agg	ggg	tat		3392
Glu	Asn	Lys	Thr	Gly	Glu	Leu	Glu	Thr	Ala	Met	Asp	Leu	Arg	Gly	Tyr		
			1065					1070					1075				
tgg	ggc	act	tac	gaa	ata	ttc	att	gag	gcc	ttc	gac	cac	ggc	tac	ccg		3440
Trp	Gly	Thr	Tyr	Glu	Ile	Phe	Ile	Glu	Ala	Phe	Asp	His	Gly	Tyr	Pro		
		1080					1085					1090					
cag	cag	agg	tcc	aac	gag	acg	tac	acc	ctg	gtc	atc	cgc	ccc	tac	aac		3488
Gln	Gln	Arg	Ser	Asn	Glu	Thr	Tyr	Thr	Leu	Val	Ile	Arg	Pro	Tyr	Asn		
		1095				1100					1105						
ttc	cac	cac	cct	gtg	ttc	gtg	ttc	ccg	caa	ccc	gac	tcc	gtc	att	cgg		3536
Phe	His	His	Pro	Val	Phe	Val	Phe	Pro	Gln	Pro	Asp	Ser	Val	Ile	Arg		
				1110		1115				1120				1125			
ctt	tct	agg	gag	cgc	gca	aca	gaa	ggc	ggc	ggt	ctg	gcg	acg	gct	gcc		3584
Leu	Ser	Arg	Glu	Arg	Ala	Thr	Glu	Gly	Gly	Val	Leu	Ala	Thr	Ala	Ala		
				1130					1135				1140				
aac	gag	ttc	ctg	gag	ccg	atc	tac	gcc	acc	gac	gag	gac	ggc	ctc	cac		3632
Asn	Glu	Phe	Leu	Glu	Pro	Ile	Tyr	Ala	Thr	Asp	Glu	Asp	Gly	Leu	His		
			1145					1150					1155				
gcg	ggc	agc	gtc	acg	ttc	cac	gtc	cag	gga	aat	gag	gag	gcc	ggt	cag		3680
Ala	Gly	Ser	Val	Thr	Phe	His	Val	Gln	Gly	Asn	Glu	Glu	Ala	Val	Gln		
		1160					1165					1170					
tac	ttt	gat	ata	act	gaa	gtg	gga	gca	gga	gaa	aat	agc	ggg	cag	ctt		3728
Tyr	Phe	Asp	Ile	Thr	Glu	Val	Gly	Ala	Gly	Glu	Asn	Ser	Gly	Gln	Leu		
		1175				1180					1185						
ata	tta	cgc	cag	ctt	ttc	cca	gag	caa	atc	aga	caa	ttc	agg	atc	acg		3776

-continued

Ile Leu Arg Gln Leu Phe Pro Glu Gln Ile Arg Gln Phe Arg Ile Thr	
1190	1195 1200 1205
atc cgg gcc acg gac ggc ggc acg gag ccc ggc ccg ctt tgg acc gac	3824
Ile Arg Ala Thr Asp Gly Gly Thr Glu Pro Gly Pro Leu Trp Thr Asp	
	1210 1215 1220
gtc acg ttt tcg gtg gtc ttc gta ccc aca cag ggc gac cca gtg ttc	3872
Val Thr Phe Ser Val Val Phe Val Pro Thr Gln Gly Asp Pro Val Phe	
	1225 1230 1235
agc gaa aat gca gct act gtc gcc ttc ttc gag ggt gaa gaa ggc ctc	3920
Ser Glu Asn Ala Ala Thr Val Ala Phe Phe Glu Gly Glu Glu Gly Leu	
	1240 1245 1250
cgt gag agt ttt gag ctg ccg caa gca gaa gac ctt aaa aac cac ctc	3968
Arg Glu Ser Phe Glu Leu Pro Gln Ala Glu Asp Leu Lys Asn His Leu	
	1255 1260 1265
tgc gaa gat gac tgc caa gat atc tac tac agg ttt att gac ggc aac	4016
Cys Glu Asp Asp Cys Gln Asp Ile Tyr Tyr Arg Phe Ile Asp Gly Asn	
	1270 1275 1280 1285
aac gag ggt ctt ttc gta ctg gac cag tca agc aac gtc atc tcc ctt	4064
Asn Glu Gly Leu Phe Val Leu Asp Gln Ser Ser Asn Val Ile Ser Leu	
	1290 1295 1300
gcg cag gag ttg gac cgc gag gtg gcc acg tct tac acg ctg cac atc	4112
Ala Gln Glu Leu Asp Arg Glu Val Ala Thr Ser Tyr Thr Leu His Ile	
	1305 1310 1315
gcg gcg agc aac tcg ccc gac gcc act ggg atc cct ctg cag act tcc	4160
Ala Ala Ser Asn Ser Pro Asp Ala Thr Gly Ile Pro Leu Gln Thr Ser	
	1320 1325 1330
atc ctc gtt gtc acg gtc aat gta aga gaa gcg aac ccg cgc cca att	4208
Ile Leu Val Val Thr Val Asn Val Arg Glu Ala Asn Pro Arg Pro Ile	
	1335 1340 1345
ttc gag cag gac ctt tac aca gcg ggc att tcg acg ttg gac agc att	4256
Phe Glu Gln Asp Leu Tyr Thr Ala Gly Ile Ser Thr Leu Asp Ser Ile	
	1350 1355 1360 1365
ggc cgg gaa ttg ctt act gtc agg gcg agc cac aca gaa gac gac acc	4304
Gly Arg Glu Leu Leu Thr Val Arg Ala Ser His Thr Glu Asp Asp Thr	
	1370 1375 1380
atc acg tac acc ata gac cgt gcg agc atg cag ctg gac agc agc cta	4352
Ile Thr Tyr Thr Ile Asp Arg Ala Ser Met Gln Leu Asp Ser Ser Leu	
	1385 1390 1395
gaa gcc gtg cgc gac tcg gcc ttc gcg ctg cat gcg acc acc ggc gtg	4400
Glu Ala Val Arg Asp Ser Ala Phe Ala Leu His Ala Thr Thr Gly Val	
	1400 1405 1410
ctt tcg ctc aat atg cag ccc acc gct tcc atg cac ggc atg ttc gag	4448
Leu Ser Leu Asn Met Gln Pro Thr Ala Ser Met His Gly Met Phe Glu	
	1415 1420 1425
ttc gac gtc atc gct acg gat aca gct tct gca atc gac aca gcc cgt	4496
Phe Asp Val Ile Ala Thr Asp Thr Ala Ser Ala Ile Asp Thr Ala Arg	
	1430 1435 1440 1445
gtg aaa gtc tac ctc atc tca tcg caa aac cgc gtg acc ttc att ttc	4544
Val Lys Val Tyr Leu Ile Ser Ser Gln Asn Arg Val Thr Phe Ile Phe	
	1450 1455 1460
gat aac caa ctt gag acc gtt gag cag aac aga aat ttc ata gcg gcc	4592
Asp Asn Gln Leu Glu Thr Val Glu Gln Asn Arg Asn Phe Ile Ala Ala	
	1465 1470 1475
acg ttc agc acc ggg ttc aac atg acg tgc aac atc gac cag gtg gtg	4640
Thr Phe Ser Thr Gly Phe Asn Met Thr Cys Asn Ile Asp Gln Val Val	
	1480 1485 1490
ccg ttc agc gac agc agc ggc gtg gcg caa gac gac acc acc gag gtg	4688
Pro Phe Ser Asp Ser Ser Gly Val Ala Gln Asp Asp Thr Thr Glu Val	
	1495 1500 1505
cgc gcg cac ttc atc cgg gac aac gtg ccc gtg cag gca caa gag gtc	4736

-continued

Arg Ala His Phe Ile Arg Asp Asn Val Pro Val Gln Ala Gln Glu Val	
1510	1515 1520 1525
gag gcc gtc cgc agc gac acg gtg ctg ctg cgc acc atc cag ctg atg	4784
Glu Ala Val Arg Ser Asp Thr Val Leu Leu Arg Thr Ile Gln Leu Met	
	1530 1535 1540
ctg agc acc aac agc ctg gtg ctg caa gac ctg gtg acg ggt gac act	4832
Leu Ser Thr Asn Ser Leu Val Leu Gln Asp Leu Val Thr Gly Asp Thr	
	1545 1550 1555
ccg acg cta ggc gag gag tca atg cag atc gcc gtc tac gca cta gcc	4880
Pro Thr Leu Gly Glu Glu Ser Met Gln Ile Ala Val Tyr Ala Leu Ala	
	1560 1565 1570
gcg ctc tcc gct gtg cta ggc ttc ctc tgc ctc gta ctg ctt ctc gca	4928
Ala Leu Ser Ala Val Leu Gly Phe Leu Cys Leu Val Leu Leu Ala	
	1575 1580 1585
ttg ttc tgt agg aca aga gca ctg aac cgg cag ctg caa gca ctc tcc	4976
Leu Phe Cys Arg Thr Arg Ala Leu Asn Arg Gln Leu Gln Ala Leu Ser	
	1590 1595 1600 1605
atg acg aag tac ggc tcg gtg gac tcc ggg ctg aac cgc gcc ggg ctg	5024
Met Thr Lys Tyr Gly Ser Val Asp Ser Gly Leu Asn Arg Ala Gly Leu	
	1610 1615 1620
gcg ccg ggc acc aac aag cac gcc gtc gag ggc tcc aac ccc atg tgg	5072
Ala Pro Gly Thr Asn Lys His Ala Val Glu Gly Ser Asn Pro Met Trp	
	1625 1630 1635
aac gag gcc atc cgc gcg ccc gac ttc gac gcc atc agt gac gcg agt	5120
Asn Glu Ala Ile Arg Ala Pro Asp Phe Asp Ala Ile Ser Asp Ala Ser	
	1640 1645 1650
ggc gac tcc gac ctg atc ggc atc gag gac atg ccg caa ttc cgc gac	5168
Gly Asp Ser Asp Leu Ile Gly Ile Glu Asp Met Pro Gln Phe Arg Asp	
	1655 1660 1665
gac tac ttc ccg ccc ggc gac aca gac tca agc agc ggc atc gtc ttg	5216
Asp Tyr Phe Pro Pro Gly Asp Thr Asp Ser Ser Ser Gly Ile Val Leu	
	1670 1675 1680 1685
cac atg ggc gaa gcc acg gac aac aag ccc gtg acc acg cat ggc aac	5264
His Met Gly Glu Ala Thr Asp Asn Lys Pro Val Thr Thr His Gly Asn	
	1690 1695 1700
aac ttc ggg ttc aag tcc acc ccg tac ctg cca cag ccg cac cca aag	5312
Asn Phe Gly Phe Lys Ser Thr Pro Tyr Leu Pro Gln Pro His Pro Lys	
	1705 1710 1715
taactgccag ggtataacct gtccaggggtg cctacgccgc gcgaagtgcg cacacgcggt	5372
tatcatcggg aaacattagc atgaagatac ctatgtacat attgtaaatt gtaacatatac	5432
tatttttata caaatatatt ttatttatat ttgctaaaaa aaaaaaaaaa aaaaaaaaaa	5492
ctcgag	5498

<210> SEQ ID NO 2

<211> LENGTH: 1717

<212> TYPE: PRT

<213> ORGANISM: Ostrinia nubilalis

<400> SEQUENCE: 2

Met Gly Val Glu Arg Phe Phe Pro Ala Val Leu Leu Val Ser Leu Ala	
1	5 10 15
Ser Ala Ala Leu Ala Asn Gln Arg Cys Ser Tyr Ile Ile Ala Ile Pro	
	20 25 30
Arg Pro Glu Thr Pro Glu Leu Pro Pro Ile Asp Tyr Glu Gly Lys Ser	
	35 40 45
Trp Ser Glu Gln Pro Leu Ile Pro Gly Pro Thr Arg Glu Glu Val Cys	
	50 55 60
Met Glu Asn Phe Leu Pro Asp Gln Met Ile Gln Val Ile Tyr Met Glu	

-continued

65	70	75	80
Glu Glu Ile Glu Gly Asp Val Ile Ile Ala Lys Leu Asn Tyr Gln Gly	85	90	95
Ser Asn Thr Pro Val Leu Ser Ile Met Ser Gly Gln Pro Arg Ala Gln	100	105	110
Leu Gly Pro Glu Phe Arg Gln Asn Glu Ala Asp Gly Gln Trp Ser Leu	115	120	125
Val Ile Thr Gln Arg Gln Asp Tyr Glu Thr Ala Thr Met Gln Ser Tyr	130	135	140
Val Phe Ser Ile Gln Val Glu Gly Glu Ser Gln Ala Val Leu Val Ala	145	150	155
Leu Glu Ile Val Asn Ile Asp Asp Asn Pro Pro Ile Leu Gln Val Val	165	170	175
Ser Ala Cys Val Ile Pro Glu His Gly Glu Ala Arg Leu Thr Asp Cys	180	185	190
Val Tyr Gln Val Ser Asp Arg Asp Gly Glu Ile Ser Thr Arg Phe Met	195	200	205
Thr Phe Arg Val Asp Ser Ser Arg Ala Ala Asp Glu Ser Ile Phe Tyr	210	215	220
Met Val Gly Glu Tyr Asp Pro Ser Asp Trp Phe Asn Met Lys Met Thr	225	230	235
Val Gly Ile Asn Ser Pro Leu Asn Phe Glu Thr Thr Gln Leu His Ile	245	250	255
Phe Ser Val Thr Ala Ser Asp Ser Leu Pro Asn Asn His Thr Val Thr	260	265	270
Met Met Val Gln Val Glu Asn Val Glu Ser Arg Pro Pro Arg Trp Val	275	280	285
Glu Ile Phe Ser Val Gln Gln Phe Asp Glu Lys Thr Asn Gln Ser Phe	290	295	300
Ser Leu Arg Ala Ile Asp Gly Asp Thr Gly Ile Asn Arg Ala Ile Asn	305	310	315
Tyr Thr Leu Ile Arg Asp Asp Ala Asp Asp Phe Phe Ser Leu Glu Val	325	330	335
Ile Glu Asp Gly Ala Ile Leu His Val Thr Glu Ile Asp Arg Asp Lys	340	345	350
Leu Glu Arg Glu Leu Phe Asn Leu Thr Ile Val Ala Tyr Lys Ser Thr	355	360	365
Asp Ala Ser Phe Ala Thr Glu Ala His Ile Phe Ile Ile Val Asn Asp	370	375	380
Val Asn Asp Gln Arg Pro Glu Pro Leu His Lys Glu Tyr Ser Ile Asp	385	390	395
Ile Met Glu Glu Thr Pro Met Thr Leu Asn Phe Asn Glu Glu Phe Gly	405	410	415
Phe His Asp Arg Asp Leu Gly Glu Asn Ala Gln Tyr Thr Val Glu Leu	420	425	430
Glu Asp Val Phe Pro Pro Gly Ala Ala Ser Ala Phe Tyr Ile Ala Pro	435	440	445
Gly Ser Gly Tyr Gln Arg Gln Thr Phe Ile Met Gly Thr Ile Asn His	450	455	460
Thr Met Leu Asp Tyr Glu Asp Val Ile Phe Gln Asn Ile Ile Ile Lys	465	470	475
Val Lys Ala Val Asp Met Asn Asn Ala Ser His Val Gly Glu Ala Leu	485	490	495

-continued

Phe Phe Glu Val Asp Leu Asp Thr Gly Arg Leu Glu Val His Tyr Pro
 930 935 940

Gly Asp Glu Lys Leu Asp Arg Asp Gly Asp Glu Pro Thr His Thr Ile
 945 950 955 960

Phe Val Asn Phe Ile Asp Asn Phe Phe Ser Asp Gly Asp Gly Arg Arg
 965 970 975

Asn Gln Asp Glu Val Glu Ile Phe Val Val Leu Leu Asp Val Asn Asp
 980 985 990

Asn Ala Pro Glu Met Pro Leu Pro Asp Glu Leu Arg Phe Asp Val Ser
 995 1000 1005

Glu Gly Ala Val Ala Gly Val Arg Val Leu Pro Glu Ile Tyr Ala Pro
 1010 1015 1020

Asp Arg Asp Glu Pro Asp Thr Asp Asn Ser Arg Val Gly Tyr Gly Ile
 1025 1030 1035 1040

Leu Asp Leu Thr Ile Thr Asp Arg Asp Ile Glu Val Pro Asp Leu Phe
 1045 1050 1055

Thr Met Ile Ser Ile Glu Asn Lys Thr Gly Glu Leu Glu Thr Ala Met
 1060 1065 1070

Asp Leu Arg Gly Tyr Trp Gly Thr Tyr Glu Ile Phe Ile Glu Ala Phe
 1075 1080 1085

Asp His Gly Tyr Pro Gln Gln Arg Ser Asn Glu Thr Tyr Thr Leu Val
 1090 1095 1100

Ile Arg Pro Tyr Asn Phe His His Pro Val Phe Val Phe Pro Gln Pro
 1105 1110 1115 1120

Asp Ser Val Ile Arg Leu Ser Arg Glu Arg Ala Thr Glu Gly Gly Val
 1125 1130 1135

Leu Ala Thr Ala Ala Asn Glu Phe Leu Glu Pro Ile Tyr Ala Thr Asp
 1140 1145 1150

Glu Asp Gly Leu His Ala Gly Ser Val Thr Phe His Val Gln Gly Asn
 1155 1160 1165

Glu Glu Ala Val Gln Tyr Phe Asp Ile Thr Glu Val Gly Ala Gly Glu
 1170 1175 1180

Asn Ser Gly Gln Leu Ile Leu Arg Gln Leu Phe Pro Glu Gln Ile Arg
 1185 1190 1195 1200

Gln Phe Arg Ile Thr Ile Arg Ala Thr Asp Gly Gly Thr Glu Pro Gly
 1205 1210 1215

Pro Leu Trp Thr Asp Val Thr Phe Ser Val Val Phe Val Pro Thr Gln
 1220 1225 1230

Gly Asp Pro Val Phe Ser Glu Asn Ala Ala Thr Val Ala Phe Phe Glu
 1235 1240 1245

Gly Glu Glu Gly Leu Arg Glu Ser Phe Glu Leu Pro Gln Ala Glu Asp
 1250 1255 1260

Leu Lys Asn His Leu Cys Glu Asp Asp Cys Gln Asp Ile Tyr Tyr Arg
 1265 1270 1275 1280

Phe Ile Asp Gly Asn Asn Glu Gly Leu Phe Val Leu Asp Gln Ser Ser
 1285 1290 1295

Asn Val Ile Ser Leu Ala Gln Glu Leu Asp Arg Glu Val Ala Thr Ser
 1300 1305 1310

Tyr Thr Leu His Ile Ala Ala Ser Asn Ser Pro Asp Ala Thr Gly Ile
 1315 1320 1325

Pro Leu Gln Thr Ser Ile Leu Val Val Thr Val Asn Val Arg Glu Ala
 1330 1335 1340

Asn Pro Arg Pro Ile Phe Glu Gln Asp Leu Tyr Thr Ala Gly Ile Ser

-continued

1345	1350	1355	1360
Thr Leu Asp Ser Ile Gly Arg Glu Leu Leu Thr Val Arg Ala Ser His	1365	1370	1375
Thr Glu Asp Asp Thr Ile Thr Tyr Thr Ile Asp Arg Ala Ser Met Gln	1380	1385	1390
Leu Asp Ser Ser Leu Glu Ala Val Arg Asp Ser Ala Phe Ala Leu His	1395	1400	1405
Ala Thr Thr Gly Val Leu Ser Leu Asn Met Gln Pro Thr Ala Ser Met	1410	1415	1420
His Gly Met Phe Glu Phe Asp Val Ile Ala Thr Asp Thr Ala Ser Ala	1425	1430	1435
Ile Asp Thr Ala Arg Val Lys Val Tyr Leu Ile Ser Ser Gln Asn Arg	1445	1450	1455
Val Thr Phe Ile Phe Asp Asn Gln Leu Glu Thr Val Glu Gln Asn Arg	1460	1465	1470
Asn Phe Ile Ala Ala Thr Phe Ser Thr Gly Phe Asn Met Thr Cys Asn	1475	1480	1485
Ile Asp Gln Val Val Pro Phe Ser Asp Ser Ser Gly Val Ala Gln Asp	1490	1495	1500
Asp Thr Thr Glu Val Arg Ala His Phe Ile Arg Asp Asn Val Pro Val	1505	1510	1515
Gln Ala Gln Glu Val Glu Ala Val Arg Ser Asp Thr Val Leu Leu Arg	1525	1530	1535
Thr Ile Gln Leu Met Leu Ser Thr Asn Ser Leu Val Leu Gln Asp Leu	1540	1545	1550
Val Thr Gly Asp Thr Pro Thr Leu Gly Glu Glu Ser Met Gln Ile Ala	1555	1560	1565
Val Tyr Ala Leu Ala Ala Leu Ser Ala Val Leu Gly Phe Leu Cys Leu	1570	1575	1580
Val Leu Leu Leu Ala Leu Phe Cys Arg Thr Arg Ala Leu Asn Arg Gln	1585	1590	1595
Leu Gln Ala Leu Ser Met Thr Lys Tyr Gly Ser Val Asp Ser Gly Leu	1605	1610	1615
Asn Arg Ala Gly Leu Ala Pro Gly Thr Asn Lys His Ala Val Glu Gly	1620	1625	1630
Ser Asn Pro Met Trp Asn Glu Ala Ile Arg Ala Pro Asp Phe Asp Ala	1635	1640	1645
Ile Ser Asp Ala Ser Gly Asp Ser Asp Leu Ile Gly Ile Glu Asp Met	1650	1655	1660
Pro Gln Phe Arg Asp Asp Tyr Phe Pro Pro Gly Asp Thr Asp Ser Ser	1665	1670	1675
Ser Gly Ile Val Leu His Met Gly Glu Ala Thr Asp Asn Lys Pro Val	1685	1690	1695
Thr Thr His Gly Asn Asn Phe Gly Phe Lys Ser Thr Pro Tyr Leu Pro	1700	1705	1710
Gln Pro His Pro Lys	1715		

<210> SEQ ID NO 3
 <211> LENGTH: 5527
 <212> TYPE: DNA
 <213> ORGANISM: Heliothis zea
 <220> FEATURE:
 <221> NAME/KEY: CDS
 <222> LOCATION: (171)...(5360)

-continued

<400> SEQUENCE: 3

gtggattggtt gttctaaaaa cagaaaaaaa acgcagtttg aaaaaagtta tttttgtgat	60
atgtgtgtaa agtgtagtgt taaataattht ggcattgctg taaaggatta aagagtgttc	120
caattgatca cccagaggtg gatcgaccag actagacaca gaactatgag atg gca	176
	Met Ala
	1
gtc gac gtg aga ata ttg acg gca gcg gtt ttc att atc gct gct cac	224
Val Asp Val Arg Ile Leu Thr Ala Ala Val Phe Ile Ile Ala Ala His	
	5 10 15
ttg act ttc gcg caa gat tgt agc tac atg gta gca ata ccc aga cca	272
Leu Thr Phe Ala Gln Asp Cys Ser Tyr Met Val Ala Ile Pro Arg Pro	
	20 25 30
gag cga cca gat ttt cca agt cta aat ttc gat gga ata cca tgg agt	320
Glu Arg Pro Asp Phe Pro Ser Leu Asn Phe Asp Gly Ile Pro Trp Ser	
	35 40 45 50
cgg tat ccc ctg ata cca gtg gag ggt aga gaa gat gtg tgc atg aac	368
Arg Tyr Pro Leu Ile Pro Val Glu Gly Arg Glu Asp Val Cys Met Asn	
	55 60 65
gaa ttc cag cca gat gcc ttg aac cca gtt acc gtc atc ttc atg gag	416
Glu Phe Gln Pro Asp Ala Leu Asn Pro Val Thr Val Ile Phe Met Glu	
	70 75 80
gag gag ata gaa ggg gat gtg gct atc gcg agg ctt aac tac cga ggt	464
Glu Glu Ile Glu Gly Asp Val Ala Ile Ala Arg Leu Asn Tyr Arg Gly	
	85 90 95
acc aat act ccg acc att gta tct cca ttt agc ttt ggt act ttt aac	512
Thr Asn Thr Pro Thr Ile Val Ser Pro Phe Ser Phe Gly Thr Phe Asn	
	100 105 110
atg ttg ggg ccg gtc ata cgt aga ata cct gag aat ggt ggc gac tgg	560
Met Leu Gly Pro Val Ile Arg Arg Ile Pro Glu Asn Gly Gly Asp Trp	
	115 120 125 130
cat ctc gtc att aca cag aga cag gac tac gag acg cca ggt atg cag	608
His Leu Val Ile Thr Gln Arg Gln Asp Tyr Glu Thr Pro Gly Met Gln	
	135 140 145
cag tac atc ttc gac gtg agg gta gac gat gaa ccg cta gtg gcc acg	656
Gln Tyr Ile Phe Asp Val Arg Val Asp Asp Glu Pro Leu Val Ala Thr	
	150 155 160
gtg atg ctg ctc att gtc aac atc gat gac aac gat cct atc ata cag	704
Val Met Leu Leu Ile Val Asn Ile Asp Asp Asn Asp Pro Ile Ile Gln	
	165 170 175
atg ttt gag cct tgt gat att cct gaa cgc ggt gaa aca ggc atc aca	752
Met Phe Glu Pro Cys Asp Ile Pro Glu Arg Gly Glu Thr Gly Ile Thr	
	180 185 190
tca tgc aag tac acc gtg agc gat gct gac ggc gag atc agt aca cgt	800
Ser Cys Lys Tyr Thr Val Ser Asp Ala Asp Gly Glu Ile Ser Thr Arg	
	195 200 205 210
ttc atg agg ttc gaa atc agc agc gat cga gac gat gac gaa tat ttc	848
Phe Met Arg Phe Glu Ile Ser Ser Asp Arg Asp Asp Asp Glu Tyr Phe	
	215 220 225
gaa ctc gtc aga gaa aat ata caa gga caa tgg atg tat gtt cat atg	896
Glu Leu Val Arg Glu Asn Ile Gln Gly Gln Trp Met Tyr Val His Met	
	230 235 240
aga gtt cac gtc aaa aaa cct ctt gat tat gag gaa aac ccg cta cat	944
Arg Val His Val Lys Lys Pro Leu Asp Tyr Glu Glu Asn Pro Leu His	
	245 250 255
ttg ttt aga gtt aca gct tat gat tcc cta cca aac aca cat aca gtg	992
Leu Phe Arg Val Thr Ala Tyr Asp Ser Leu Pro Asn Thr His Thr Val	
	260 265 270
acg atg atg gtg caa gta gag aac gtt gag aac aga ccg ccg cga tgg	1040
Thr Met Met Val Gln Val Glu Asn Val Glu Asn Arg Pro Pro Arg Trp	

-continued

275	280	285	290	
atg gag ata ttt gct gtc cag cag ttc gat gag aag acg gaa caa tcc				1088
Met Glu Ile Phe Ala Val Gln Gln Phe Asp Glu Lys Thr Glu Gln Ser	295	300	305	
ttt agg gtt cga gcc atc gat gga gat acg gga atc gat aaa cct att				1136
Phe Arg Val Arg Ala Ile Asp Gly Asp Thr Gly Ile Asp Lys Pro Ile	310	315	320	
ttc tat agg atc gaa act gaa aaa gga gag gaa gac ttg ttc agc att				1184
Phe Tyr Arg Ile Glu Thr Glu Lys Gly Glu Glu Asp Leu Phe Ser Ile	325	330	335	
caa acg ata gaa ggt ggt cga gaa ggc gct tgg ttt aac gtc gct cca				1232
Gln Thr Ile Glu Gly Gly Arg Glu Gly Ala Trp Phe Asn Val Ala Pro	340	345	350	
ata gac agg gac act cta gag aag gaa gtt ttc cac gtg tcc ata ata				1280
Ile Asp Arg Asp Thr Leu Glu Lys Glu Val Phe His Val Ser Ile Ile	355	360	365	370
gcg tac aaa tat ggc gat aat gac gtg gaa ggc agt tcg tca ttc cag				1328
Ala Tyr Lys Tyr Gly Asp Asn Asp Val Glu Gly Ser Ser Ser Phe Gln	375	380	385	
tcg aaa acc gat gtg gtc atc atc gtg aac gat gtc aat gat cag gcg				1376
Ser Lys Thr Asp Val Val Ile Ile Val Asn Asp Val Asn Asp Gln Ala	390	395	400	
ccg ctt cct ttc cgg gaa gag tac tcc att gaa att atg gag gaa act				1424
Pro Leu Pro Phe Arg Glu Glu Tyr Ser Ile Glu Ile Met Glu Glu Thr	405	410	415	
gcg atg acc ctg aat tta gaa gac ttt ggg ttc cat gat aga gat ctc				1472
Ala Met Thr Leu Asn Leu Glu Asp Phe Gly Phe His Asp Arg Asp Leu	420	425	430	
ggt cct cac gca caa tac aca gta cac tta gag agc atc cat cct ccc				1520
Gly Pro His Ala Gln Tyr Thr Val His Leu Glu Ser Ile His Pro Pro	435	440	445	450
cga gct cac gag gcg ttc tac ata gca ccg gag gtt ggc tac cag cgc				1568
Arg Ala His Glu Ala Phe Tyr Ile Ala Pro Glu Val Gly Tyr Gln Arg	455	460	465	
cag tcc ttc att atg ggc acg cag aac cat cac atg ctg gac ttc gaa				1616
Gln Ser Phe Ile Met Gly Thr Gln Asn His His Met Leu Asp Phe Glu	470	475	480	
gtg cca gag ttc cag aat ata caa ctg agg gcc gta gcg ata gac atg				1664
Val Pro Glu Phe Gln Asn Ile Gln Leu Arg Ala Val Ala Ile Asp Met	485	490	495	
gac gat ccc aaa tgg gtg ggt atc gcg ata atc aac att aaa ctg atc				1712
Asp Asp Pro Lys Trp Val Gly Ile Ala Ile Ile Asn Ile Lys Leu Ile	500	505	510	
aac tgg aac gat gag ctg ccg atg ttc gag agt gac gtg caa act gtc				1760
Asn Trp Asn Asp Glu Leu Pro Met Phe Glu Ser Asp Val Gln Thr Val	515	520	525	530
agc ttc gat gag aca gag ggc gca ggc ttc tat gtg gcc act gtt gtg				1808
Ser Phe Asp Glu Thr Glu Gly Ala Gly Phe Tyr Val Ala Thr Val Val	535	540	545	
gcg aag gac cgg gat gtt ggt gat aaa gtc gaa cac tct cta atg ggt				1856
Ala Lys Asp Arg Asp Val Gly Asp Lys Val Glu His Ser Leu Met Gly	550	555	560	
aac gca gta agc tac ctg agg atc gac aag gaa acc ggc gag ata ttc				1904
Asn Ala Val Ser Tyr Leu Arg Ile Asp Lys Glu Thr Gly Glu Ile Phe	565	570	575	
gtc aca gaa aac gaa gca ttc aac tat cac agg cag aac gaa ctc ttt				1952
Val Thr Glu Asn Glu Ala Phe Asn Tyr His Arg Gln Asn Glu Leu Phe	580	585	590	
gtg cag ata cca gct gac gac acg ctg ggc gag cct tac aac acc aac				2000
Val Gln Ile Pro Ala Asp Asp Thr Leu Gly Glu Pro Tyr Asn Thr Asn				

-continued

595	600	605	610	
act act cag ttg gtg atc aag ctg cgg gac att aac aac acc cct cct				2048
Thr Thr Gln Leu Val Ile Lys Leu Arg Asp Ile Asn Asn Thr Pro Pro	615	620	625	
acg ctc agg ctg cct cgc gcc act cca tca gtg gaa gag aac gtg ccc				2096
Thr Leu Arg Leu Pro Arg Ala Thr Pro Ser Val Glu Glu Asn Val Pro	630	635	640	
gac ggg ttt gtg atc ccc acg cag ctg cac gcc acg gac ccc gac act				2144
Asp Gly Phe Val Ile Pro Thr Gln Leu His Ala Thr Asp Pro Asp Thr	645	650	655	
aca gct gag ctg cgc ttc gag atc gac tgg cag aac tcg tat gct acc				2192
Thr Ala Glu Leu Arg Phe Glu Ile Asp Trp Gln Asn Ser Tyr Ala Thr	660	665	670	
aag cag gga cgg aat act gac tct aag gag tat atc ggt tgt ata gaa				2240
Lys Gln Gly Arg Asn Thr Asp Ser Lys Glu Tyr Ile Gly Cys Ile Glu	675	680	685	690
atc gag acg ata tac ccg aat ata aac cag cga ggc aac gcc atc ggc				2288
Ile Glu Thr Ile Tyr Pro Asn Ile Asn Gln Arg Gly Asn Ala Ile Gly	695	700	705	
cgc gtg gta gtg cga gag atc cgg gac ggc gtc acc ata gac tat gag				2336
Arg Val Val Val Arg Glu Ile Arg Asp Gly Val Thr Ile Asp Tyr Glu	710	715	720	
atg ttt gaa gtt cta tac ctc acc gtc att gtg agg gat ctc aac acc				2384
Met Phe Glu Val Leu Tyr Leu Thr Val Ile Val Arg Asp Leu Asn Thr	725	730	735	
gtt att gga gaa gac cat gat ata tcc aca ttc acg atc acg ata ata				2432
Val Ile Gly Glu Asp His Asp Ile Ser Thr Phe Thr Ile Thr Ile Ile	740	745	750	
gac atg aac gac aac cct ccc ctg tgg gtg gaa ggc acc ctg acg caa				2480
Asp Met Asn Asp Asn Pro Pro Leu Trp Val Glu Gly Thr Leu Thr Gln	755	760	765	770
gag ttc cgt gtg cga gag gtg gca gcc tca gga gtt gtt ata gga tcc				2528
Glu Phe Arg Val Arg Glu Val Ala Ala Ser Gly Val Val Ile Gly Ser	775	780	785	
gta ctg gcc act gat atc gac gga ccg ctg tat aat caa gtg cgg tat				2576
Val Leu Ala Thr Asp Ile Asp Gly Pro Leu Tyr Asn Gln Val Arg Tyr	790	795	800	
act att act ccc aga cta gac act cca gaa gac cta gtg gac ata gac				2624
Thr Ile Thr Pro Arg Leu Asp Thr Pro Glu Asp Leu Val Asp Ile Asp	805	810	815	
ttc aac acg ggt cag atc tcc gta aag tta cac cag gct ata gac gcg				2672
Phe Asn Thr Gly Gln Ile Ser Val Lys Leu His Gln Ala Ile Asp Ala	820	825	830	
gac gag ccg ccg cgt cag aac ctc tac tac acc gtc ata gct agt gac				2720
Asp Glu Pro Pro Arg Gln Asn Leu Tyr Tyr Thr Val Ile Ala Ser Asp	835	840	845	850
aag tgt gac ctc ctt act gtc act gag tgt ccg cct gac cct act tac				2768
Lys Cys Asp Leu Leu Thr Val Thr Glu Cys Pro Pro Asp Pro Thr Tyr	855	860	865	
ttt gag aca ccg gga gag att acc atc cac ata acg gac acg aac aac				2816
Phe Glu Thr Pro Gly Glu Ile Thr Ile His Ile Thr Asp Thr Asn Asn	870	875	880	
aag gtg cct caa gtg gaa gac gac aag ttc gag gcg acg gtg tac atc				2864
Lys Val Pro Gln Val Glu Asp Asp Lys Phe Glu Ala Thr Val Tyr Ile	885	890	895	
tac gag ggc gcg gac gat gga caa cat gtc gtg cag atc tac gcc agc				2912
Tyr Glu Gly Ala Asp Asp Gly Gln His Val Val Gln Ile Tyr Ala Ser	900	905	910	
gat ctg gat aga gat gaa atc tac cac aaa gtg agc tac cag atc aac				2960
Asp Leu Asp Arg Asp Glu Ile Tyr His Lys Val Ser Tyr Gln Ile Asn				

-continued

915	920	925	930	
tac gcg atc aac tct cgt ctc cgc gac ttc ttc gag atg gac ctg gag				3008
Tyr Ala Ile Asn Ser Arg Leu Arg Asp Phe Phe Glu Met Asp Leu Glu				
	935	940	945	
tcc ggc ctc gtg tac gtc aac aac acc gcc ggc gag ctg ctg gac agg				3056
Ser Gly Leu Val Tyr Val Asn Asn Thr Ala Gly Glu Leu Leu Asp Arg				
	950	955	960	
gac ggc gac gag ccc aca cat cgc atc ttc ttc aat gtc atc gat aac				3104
Asp Gly Asp Glu Pro Thr His Arg Ile Phe Phe Asn Val Ile Asp Asn				
	965	970	975	
ttc tat gga gaa gga gat ggc aac cgc aat cag aac gag aca caa gtg				3152
Phe Tyr Gly Glu Gly Asp Gly Asn Arg Asn Gln Asn Glu Thr Gln Val				
	980	985	990	
tta gta gta ttg ctg gac atc aat gac aac tat ccg gaa ctg cct gaa				3200
Leu Val Val Leu Leu Asp Ile Asn Asp Asn Tyr Pro Glu Leu Pro Glu				
	995	1000	1005	1010
act atc cca tgg gct atc tct gag agc tta gag ctg ggt gag cgt gta				3248
Thr Ile Pro Trp Ala Ile Ser Glu Ser Leu Glu Leu Gly Glu Arg Val				
	1015	1020	1025	
cag cca gaa atc ttt gcc cgg gac cgc gac gaa ccc gga aca gac aac				3296
Gln Pro Glu Ile Phe Ala Arg Asp Arg Asp Glu Pro Gly Thr Asp Asn				
	1030	1035	1040	
tcc cgc gtc gcc tat gcc atc aca ggc ctc gcc agc act gac cgg gac				3344
Ser Arg Val Ala Tyr Ala Ile Thr Gly Leu Ala Ser Thr Asp Arg Asp				
	1045	1050	1055	
ata caa gtg cct aat ctc ttc aac atg atc act ata gag agg gac agg				3392
Ile Gln Val Pro Asn Leu Phe Asn Met Ile Thr Ile Glu Arg Asp Arg				
	1060	1065	1070	
gga att gat cag aca gga ata ctt gag gca gct atg gat ttg aga ggc				3440
Gly Ile Asp Gln Thr Gly Ile Leu Glu Ala Ala Met Asp Leu Arg Gly				
	1075	1080	1085	1090
tat tgg ggc acc tat caa ata gat att cag gcg tat gac cat gga ata				3488
Tyr Trp Gly Thr Tyr Gln Ile Asp Ile Gln Ala Tyr Asp His Gly Ile				
	1095	1100	1105	
cct caa agg att tca aat cag aag tac ccg ctg gtg att aga cct tac				3536
Pro Gln Arg Ile Ser Asn Gln Lys Tyr Pro Leu Val Ile Arg Pro Tyr				
	1110	1115	1120	
aac ttc cac gac cca gtg ttc gtg ttc cct caa cct gga tcc act atc				3584
Asn Phe His Asp Pro Val Phe Val Phe Pro Gln Pro Gly Ser Thr Ile				
	1125	1130	1135	
aga ctg gca aag gag cga gca gta gtc aac ggt ata ctg gct aca gta				3632
Arg Leu Ala Lys Glu Arg Ala Val Val Asn Gly Ile Leu Ala Thr Val				
	1140	1145	1150	
gac ggc gaa ttt ctg gac aga atc gtt gcc acc gac gag gat ggt tta				3680
Asp Gly Glu Phe Leu Asp Arg Ile Val Ala Thr Asp Glu Asp Gly Leu				
	1155	1160	1165	1170
gaa gct gga ctt gtc aca ttc tct atc gcc gga gat gat gaa gat gct				3728
Glu Ala Gly Leu Val Thr Phe Ser Ile Ala Gly Asp Asp Glu Asp Ala				
	1175	1180	1185	
cag ttc ttc gac gtg ttg aac gac gga gtg aac tcg ggt gct ctc acc				3776
Gln Phe Phe Asp Val Leu Asn Asp Gly Val Asn Ser Gly Ala Leu Thr				
	1190	1195	1200	
ctc acg cgg ctc ttc cct gaa gag ttc cga gag ttc cag gtg acg att				3824
Leu Thr Arg Leu Phe Pro Glu Glu Phe Arg Glu Phe Gln Val Thr Ile				
	1205	1210	1215	
cgt gct acg gac ggt gga act gag cct ggt cca agg agt acg gac tgc				3872
Arg Ala Thr Asp Gly Gly Thr Glu Pro Gly Pro Arg Ser Thr Asp Cys				
	1220	1225	1230	
ttg gtg acc gta gtg ttt gta ccc acg cag gga gag ccc gtg ttc gag				3920
Leu Val Thr Val Val Phe Val Pro Thr Gln Gly Glu Pro Val Phe Glu				

-continued

1235	1240	1245	1250	
gat agg act tac acg gtt gct ttt gtt gaa aaa gat gag ggt atg tta				3968
Asp Arg Thr Tyr Thr Val Ala Phe Val Glu Lys Asp Glu Gly Met Leu				
	1255	1260	1265	
gag gag gcg gaa cta cct cgc gcc tca gac cca agg aac atc atg tgt				4016
Glu Glu Ala Glu Leu Pro Arg Ala Ser Asp Pro Arg Asn Ile Met Cys				
	1270	1275	1280	
gaa gat gat tgt cac gac acc tat tac agc att gtt gga ggc aat tcg				4064
Glu Asp Asp Cys His Asp Thr Tyr Tyr Ser Ile Val Gly Gly Asn Ser				
	1285	1290	1295	
ggt gaa cac ttc aca gta gac cct cgt acc aac gtg cta tcc ctg gtg				4112
Gly Glu His Phe Thr Val Asp Pro Arg Thr Asn Val Leu Ser Leu Val				
	1300	1305	1310	
aag ccg ctg gac cgc tcc gaa cag gag aca cac acc ctc atc att gga				4160
Lys Pro Leu Asp Arg Ser Glu Gln Glu Thr His Thr Leu Ile Ile Gly				
	1315	1320	1325	1330
gcc agc gac act ccc aac ccg gcc gcc gtc ctg cag gct tct aca ctc				4208
Ala Ser Asp Thr Pro Asn Pro Ala Ala Val Leu Gln Ala Ser Thr Leu				
	1335	1340	1345	
act gtc act gtt aat gtt cga gaa gcg aac ccg cga cca gtg ttc caa				4256
Thr Val Thr Val Asn Val Arg Glu Ala Asn Pro Arg Pro Val Phe Gln				
	1350	1355	1360	
aga gca ctc tac aca gct ggc atc tct gct ggc gat ttc atc gaa aga				4304
Arg Ala Leu Tyr Thr Ala Gly Ile Ser Ala Gly Asp Phe Ile Glu Arg				
	1365	1370	1375	
aat ctg ctg act tta gta gcg aca cat tca gaa gat ctg ccc atc act				4352
Asn Leu Leu Thr Leu Val Ala Thr His Ser Glu Asp Leu Pro Ile Thr				
	1380	1385	1390	
tac act ctg ata caa gag tcc atg gaa gca gac ccc aca ctc gaa gct				4400
Tyr Thr Leu Ile Gln Glu Ser Met Glu Ala Asp Pro Thr Leu Glu Ala				
	1395	1400	1405	1410
gtt cag gag tca gcc ttc atc ctc aac cct gag act gga gtc ctg tcc				4448
Val Gln Glu Ser Ala Phe Ile Leu Asn Pro Glu Thr Gly Val Leu Ser				
	1415	1420	1425	
ctc aac ttc cag cca acc gcc tcc atg cac ggc atg ttc gag ttc gaa				4496
Leu Asn Phe Gln Pro Thr Ala Ser Met His Gly Met Phe Glu Phe Glu				
	1430	1435	1440	
gtc aaa gcc act gat tca agg aca gaa act gcc cgc acg gaa gtg aag				4544
Val Lys Ala Thr Asp Ser Arg Thr Glu Thr Ala Arg Thr Glu Val Lys				
	1445	1450	1455	
gtg tac ctg ata tca gac cgc aac cga gtg ttc ttc acg ttc aat aac				4592
Val Tyr Leu Ile Ser Asp Arg Asn Arg Val Phe Phe Thr Phe Asn Asn				
	1460	1465	1470	
cca ctg cct gaa gtc aca ccc cag gaa gat ttc ata gcg gag acg ttc				4640
Pro Leu Pro Glu Val Thr Pro Gln Glu Asp Phe Ile Ala Glu Thr Phe				
	1475	1480	1485	1490
acg gca ttc ttc ggc atg acg tgc aac atc gac cag tcg tgg tgg gcc				4688
Thr Ala Phe Phe Gly Met Thr Cys Asn Ile Asp Gln Ser Trp Trp Ala				
	1495	1500	1505	
agc gat ccc gtc acc ggc gcc acc aag gac gac cag act gaa gtc agg				4736
Ser Asp Pro Val Thr Gly Ala Thr Lys Asp Asp Gln Thr Glu Val Arg				
	1510	1515	1520	
gct cat ttc atc agg gac gac ctt ccc gtg cct gct gag gag att gaa				4784
Ala His Phe Ile Arg Asp Asp Leu Pro Val Pro Ala Glu Glu Ile Glu				
	1525	1530	1535	
cag tta cgc ggt aac cca act cta gta aat agc atc caa cga gcc ctg				4832
Gln Leu Arg Gly Asn Pro Thr Leu Val Asn Ser Ile Gln Arg Ala Leu				
	1540	1545	1550	
gag gaa cag aac ctg cag cta gcc gac ctg ttc acg ggc gag acg ccc				4880
Glu Glu Gln Asn Leu Gln Leu Ala Asp Leu Phe Thr Gly Glu Thr Pro				

-continued

1555	1560	1565	1570	
atc ctc ggc ggc gac gcg cag gct cga gcc ctg tac gcg ctg gcg gcg				4928
Ile Leu Gly Gly Asp Ala Gln Ala Arg Ala Leu Tyr Ala Leu Ala Ala				
	1575	1580	1585	
gtg gcg gcg gca ctc gcg ctg att gtt gtt gtg ctg ctg att gtg ttc				4976
Val Ala Ala Ala Leu Ala Leu Ile Val Val Val Leu Leu Ile Val Phe				
	1590	1595	1600	
ttt gtt agg act agg act ctg aac cgg cgc ttg caa gct ctg tcc atg				5024
Phe Val Arg Thr Arg Thr Leu Asn Arg Arg Leu Gln Ala Leu Ser Met				
	1605	1610	1615	
acc aag tac agt tcg caa gac tct ggg ttg aac cgc gtg ggt ttg gcg				5072
Thr Lys Tyr Ser Ser Gln Asp Ser Gly Leu Asn Arg Val Gly Leu Ala				
	1620	1625	1630	
gcg ccg ggc acc aat aag cac gct gtc gag ggc tcc aac ccc atc tgg				5120
Ala Pro Gly Thr Asn Lys His Ala Val Glu Gly Ser Asn Pro Ile Trp				
	1635	1640	1645	1650
aat gaa acg ttg aag gct ccg gac ttt gac gct ctt agc gag cag tcg				5168
Asn Glu Thr Leu Lys Ala Pro Asp Phe Asp Ala Leu Ser Glu Gln Ser				
	1655	1660	1665	
tac gac tca gac cta atc ggc atc gaa gac ttg ccg cag ttc agg aac				5216
Tyr Asp Ser Asp Leu Ile Gly Ile Glu Asp Leu Pro Gln Phe Arg Asn				
	1670	1675	1680	
gac tac ttc cca cct gag gag ggc agc tcc atg cga gga gtc gtc aat				5264
Asp Tyr Phe Pro Pro Glu Glu Gly Ser Ser Met Arg Gly Val Val Asn				
	1685	1690	1695	
gaa cac gtg cct gaa tca ata gca aac cat aac aac aac ttc ggg ttt				5312
Glu His Val Pro Glu Ser Ile Ala Asn His Asn Asn Asn Phe Gly Phe				
	1700	1705	1710	
aac tct act ccc ttc agc cca gag ttc gcg aac acg cag ttc aga aga				5360
Asn Ser Thr Pro Phe Ser Pro Glu Phe Ala Asn Thr Gln Phe Arg Arg				
	1715	1720	1725	1730
taaaatatta aagcatttta aattataata ttatgtaccg gtgaaatacc atacttatat				5420
ttacctaatg atatattaa gtgagattaa gtaagatact cgtattaatt aagagcattt				5480
atTTTTTTTaa atacaaaaca attaaactaa aaaaaaaaaa aaaaaaa				5527

<210> SEQ ID NO 4

<211> LENGTH: 1730

<212> TYPE: PRT

<213> ORGANISM: Heliothis zea

<400> SEQUENCE: 4

Met Ala Val Asp Val Arg Ile Leu Thr Ala Ala Val Phe Ile Ile Ala				
1	5	10	15	
Ala His Leu Thr Phe Ala Gln Asp Cys Ser Tyr Met Val Ala Ile Pro				
	20	25	30	
Arg Pro Glu Arg Pro Asp Phe Pro Ser Leu Asn Phe Asp Gly Ile Pro				
	35	40	45	
Trp Ser Arg Tyr Pro Leu Ile Pro Val Glu Gly Arg Glu Asp Val Cys				
	50	55	60	
Met Asn Glu Phe Gln Pro Asp Ala Leu Asn Pro Val Thr Val Ile Phe				
	65	70	75	80
Met Glu Glu Glu Ile Glu Gly Asp Val Ala Ile Ala Arg Leu Asn Tyr				
	85	90	95	
Arg Gly Thr Asn Thr Pro Thr Ile Val Ser Pro Phe Ser Phe Gly Thr				
	100	105	110	
Phe Asn Met Leu Gly Pro Val Ile Arg Arg Ile Pro Glu Asn Gly Gly				
	115	120	125	

-continued

Asp	Trp	His	Leu	Val	Ile	Thr	Gln	Arg	Gln	Asp	Tyr	Glu	Thr	Pro	Gly
130						135					140				
Met	Gln	Gln	Tyr	Ile	Phe	Asp	Val	Arg	Val	Asp	Asp	Glu	Pro	Leu	Val
145					150					155					160
Ala	Thr	Val	Met	Leu	Leu	Ile	Val	Asn	Ile	Asp	Asp	Asn	Asp	Pro	Ile
				165					170					175	
Ile	Gln	Met	Phe	Glu	Pro	Cys	Asp	Ile	Pro	Glu	Arg	Gly	Glu	Thr	Gly
			180					185					190		
Ile	Thr	Ser	Cys	Lys	Tyr	Thr	Val	Ser	Asp	Ala	Asp	Gly	Glu	Ile	Ser
		195					200					205			
Thr	Arg	Phe	Met	Arg	Phe	Glu	Ile	Ser	Ser	Asp	Arg	Asp	Asp	Asp	Glu
		210				215					220				
Tyr	Phe	Glu	Leu	Val	Arg	Glu	Asn	Ile	Gln	Gly	Gln	Trp	Met	Tyr	Val
225					230					235					240
His	Met	Arg	Val	His	Val	Lys	Lys	Pro	Leu	Asp	Tyr	Glu	Glu	Asn	Pro
				245					250					255	
Leu	His	Leu	Phe	Arg	Val	Thr	Ala	Tyr	Asp	Ser	Leu	Pro	Asn	Thr	His
			260					265					270		
Thr	Val	Thr	Met	Met	Val	Gln	Val	Glu	Asn	Val	Glu	Asn	Arg	Pro	Pro
		275					280					285			
Arg	Trp	Met	Glu	Ile	Phe	Ala	Val	Gln	Gln	Phe	Asp	Glu	Lys	Thr	Glu
		290				295					300				
Gln	Ser	Phe	Arg	Val	Arg	Ala	Ile	Asp	Gly	Asp	Thr	Gly	Ile	Asp	Lys
305					310					315					320
Pro	Ile	Phe	Tyr	Arg	Ile	Glu	Thr	Glu	Lys	Gly	Glu	Glu	Asp	Leu	Phe
				325					330					335	
Ser	Ile	Gln	Thr	Ile	Glu	Gly	Gly	Arg	Glu	Gly	Ala	Trp	Phe	Asn	Val
			340					345					350		
Ala	Pro	Ile	Asp	Arg	Asp	Thr	Leu	Glu	Lys	Glu	Val	Phe	His	Val	Ser
			355				360					365			
Ile	Ile	Ala	Tyr	Lys	Tyr	Gly	Asp	Asn	Asp	Val	Glu	Gly	Ser	Ser	Ser
		370				375					380				
Phe	Gln	Ser	Lys	Thr	Asp	Val	Val	Ile	Ile	Val	Asn	Asp	Val	Asn	Asp
385					390					395					400
Gln	Ala	Pro	Leu	Pro	Phe	Arg	Glu	Glu	Tyr	Ser	Ile	Glu	Ile	Met	Glu
				405					410						415
Glu	Thr	Ala	Met	Thr	Leu	Asn	Leu	Glu	Asp	Phe	Gly	Phe	His	Asp	Arg
			420					425					430		
Asp	Leu	Gly	Pro	His	Ala	Gln	Tyr	Thr	Val	His	Leu	Glu	Ser	Ile	His
		435					440					445			
Pro	Pro	Arg	Ala	His	Glu	Ala	Phe	Tyr	Ile	Ala	Pro	Glu	Val	Gly	Tyr
		450				455					460				
Gln	Arg	Gln	Ser	Phe	Ile	Met	Gly	Thr	Gln	Asn	His	His	Met	Leu	Asp
465					470					475					480
Phe	Glu	Val	Pro	Glu	Phe	Gln	Asn	Ile	Gln	Leu	Arg	Ala	Val	Ala	Ile
				485					490					495	
Asp	Met	Asp	Asp	Pro	Lys	Trp	Val	Gly	Ile	Ala	Ile	Ile	Asn	Ile	Lys
				500				505					510		
Leu	Ile	Asn	Trp	Asn	Asp	Glu	Leu	Pro	Met	Phe	Glu	Ser	Asp	Val	Gln
		515					520					525			
Thr	Val	Ser	Phe	Asp	Glu	Thr	Glu	Gly	Ala	Gly	Phe	Tyr	Val	Ala	Thr
		530				535					540				
Val	Val	Ala	Lys	Asp	Arg	Asp	Val	Gly	Asp	Lys	Val	Glu	His	Ser	Leu
545					550					555					560

-continued

980				985				990							
Gln	Val	Leu	Val	Val	Leu	Leu	Asp	Ile	Asn	Asp	Asn	Tyr	Pro	Glu	Leu
		995					1000					1005			
Pro	Glu	Thr	Ile	Pro	Trp	Ala	Ile	Ser	Glu	Ser	Leu	Glu	Leu	Gly	Glu
	1010					1015					1020				
Arg	Val	Gln	Pro	Glu	Ile	Phe	Ala	Arg	Asp	Arg	Asp	Glu	Pro	Gly	Thr
	1025				1030					1035					1040
Asp	Asn	Ser	Arg	Val	Ala	Tyr	Ala	Ile	Thr	Gly	Leu	Ala	Ser	Thr	Asp
				1045					1050					1055	
Arg	Asp	Ile	Gln	Val	Pro	Asn	Leu	Phe	Asn	Met	Ile	Thr	Ile	Glu	Arg
		1060					1065						1070		
Asp	Arg	Gly	Ile	Asp	Gln	Thr	Gly	Ile	Leu	Glu	Ala	Ala	Met	Asp	Leu
		1075					1080					1085			
Arg	Gly	Tyr	Trp	Gly	Thr	Tyr	Gln	Ile	Asp	Ile	Gln	Ala	Tyr	Asp	His
	1090					1095					1100				
Gly	Ile	Pro	Gln	Arg	Ile	Ser	Asn	Gln	Lys	Tyr	Pro	Leu	Val	Ile	Arg
	1105				1110					1115					1120
Pro	Tyr	Asn	Phe	His	Asp	Pro	Val	Phe	Val	Phe	Pro	Gln	Pro	Gly	Ser
				1125						1130				1135	
Thr	Ile	Arg	Leu	Ala	Lys	Glu	Arg	Ala	Val	Val	Asn	Gly	Ile	Leu	Ala
			1140						1145				1150		
Thr	Val	Asp	Gly	Glu	Phe	Leu	Asp	Arg	Ile	Val	Ala	Thr	Asp	Glu	Asp
		1155					1160					1165			
Gly	Leu	Glu	Ala	Gly	Leu	Val	Thr	Phe	Ser	Ile	Ala	Gly	Asp	Asp	Glu
	1170					1175					1180				
Asp	Ala	Gln	Phe	Phe	Asp	Val	Leu	Asn	Asp	Gly	Val	Asn	Ser	Gly	Ala
	1185				1190					1195					1200
Leu	Thr	Leu	Thr	Arg	Leu	Phe	Pro	Glu	Glu	Phe	Arg	Glu	Phe	Gln	Val
				1205					1210					1215	
Thr	Ile	Arg	Ala	Thr	Asp	Gly	Gly	Thr	Glu	Pro	Gly	Pro	Arg	Ser	Thr
			1220						1225					1230	
Asp	Cys	Leu	Val	Thr	Val	Val	Phe	Val	Pro	Thr	Gln	Gly	Glu	Pro	Val
		1235					1240					1245			
Phe	Glu	Asp	Arg	Thr	Tyr	Thr	Val	Ala	Phe	Val	Glu	Lys	Asp	Glu	Gly
	1250					1255					1260				
Met	Leu	Glu	Glu	Ala	Glu	Leu	Pro	Arg	Ala	Ser	Asp	Pro	Arg	Asn	Ile
	1265				1270					1275					1280
Met	Cys	Glu	Asp	Asp	Cys	His	Asp	Thr	Tyr	Tyr	Ser	Ile	Val	Gly	Gly
				1285					1290					1295	
Asn	Ser	Gly	Glu	His	Phe	Thr	Val	Asp	Pro	Arg	Thr	Asn	Val	Leu	Ser
			1300						1305				1310		
Leu	Val	Lys	Pro	Leu	Asp	Arg	Ser	Glu	Gln	Glu	Thr	His	Thr	Leu	Ile
		1315					1320						1325		
Ile	Gly	Ala	Ser	Asp	Thr	Pro	Asn	Pro	Ala	Ala	Val	Leu	Gln	Ala	Ser
	1330						1335				1340				
Thr	Leu	Thr	Val	Thr	Val	Asn	Val	Arg	Glu	Ala	Asn	Pro	Arg	Pro	Val
	1345				1350					1355				1360	
Phe	Gln	Arg	Ala	Leu	Tyr	Thr	Ala	Gly	Ile	Ser	Ala	Gly	Asp	Phe	Ile
				1365					1370					1375	
Glu	Arg	Asn	Leu	Leu	Thr	Leu	Val	Ala	Thr	His	Ser	Glu	Asp	Leu	Pro
			1380						1385				1390		
Ile	Thr	Tyr	Thr	Leu	Ile	Gln	Glu	Ser	Met	Glu	Ala	Asp	Pro	Thr	Leu
			1395				1400					1405			

-continued

Glu Ala Val Gln Glu Ser Ala Phe Ile Leu Asn Pro Glu Thr Gly Val
 1410 1415 1420
 Leu Ser Leu Asn Phe Gln Pro Thr Ala Ser Met His Gly Met Phe Glu
 1425 1430 1435 1440
 Phe Glu Val Lys Ala Thr Asp Ser Arg Thr Glu Thr Ala Arg Thr Glu
 1445 1450 1455
 Val Lys Val Tyr Leu Ile Ser Asp Arg Asn Arg Val Phe Phe Thr Phe
 1460 1465 1470
 Asn Asn Pro Leu Pro Glu Val Thr Pro Gln Glu Asp Phe Ile Ala Glu
 1475 1480 1485
 Thr Phe Thr Ala Phe Phe Gly Met Thr Cys Asn Ile Asp Gln Ser Trp
 1490 1495 1500
 Trp Ala Ser Asp Pro Val Thr Gly Ala Thr Lys Asp Asp Gln Thr Glu
 1505 1510 1515 1520
 Val Arg Ala His Phe Ile Arg Asp Asp Leu Pro Val Pro Ala Glu Glu
 1525 1530 1535
 Ile Glu Gln Leu Arg Gly Asn Pro Thr Leu Val Asn Ser Ile Gln Arg
 1540 1545 1550
 Ala Leu Glu Glu Gln Asn Leu Gln Leu Ala Asp Leu Phe Thr Gly Glu
 1555 1560 1565
 Thr Pro Ile Leu Gly Gly Asp Ala Gln Ala Arg Ala Leu Tyr Ala Leu
 1570 1575 1580
 Ala Ala Val Ala Ala Ala Leu Ala Leu Ile Val Val Val Leu Leu Ile
 1585 1590 1595 1600
 Val Phe Phe Val Arg Thr Arg Thr Leu Asn Arg Arg Leu Gln Ala Leu
 1605 1610 1615
 Ser Met Thr Lys Tyr Ser Ser Gln Asp Ser Gly Leu Asn Arg Val Gly
 1620 1625 1630
 Leu Ala Ala Pro Gly Thr Asn Lys His Ala Val Glu Gly Ser Asn Pro
 1635 1640 1645
 Ile Trp Asn Glu Thr Leu Lys Ala Pro Asp Phe Asp Ala Leu Ser Glu
 1650 1655 1660
 Gln Ser Tyr Asp Ser Asp Leu Ile Gly Ile Glu Asp Leu Pro Gln Phe
 1665 1670 1675 1680
 Arg Asn Asp Tyr Phe Pro Pro Glu Glu Gly Ser Ser Met Arg Gly Val
 1685 1690 1695
 Val Asn Glu His Val Pro Glu Ser Ile Ala Asn His Asn Asn Asn Phe
 1700 1705 1710
 Gly Phe Asn Ser Thr Pro Phe Ser Pro Glu Phe Ala Asn Thr Gln Phe
 1715 1720 1725
 Arg Arg
 1730

<210> SEQ ID NO 5
 <211> LENGTH: 5592
 <212> TYPE: DNA
 <213> ORGANISM: Spodoptera frugiperda
 <220> FEATURE:
 <221> NAME/KEY: CDS
 <222> LOCATION: (162)...(5363)
 <400> SEQUENCE: 5

gacattctgt ggtgaaaca ttttttattt attttttct agtggtttgt gggtagctg 60
 taaacatttt ggaatattgt taaagatttc ggaatattgt taaagtattg acagataaag 120
 ctgtaacatc actagagaag tgagaactgc aagatcatga g atg gcg gtc gat gtg 176
 Met Ala Val Asp Val

-continued

														1	5			
cga	ata	ctg	aca	gca	aca	ttg	ctg	gta	ctc	acc	act	gct	aca	gca	cag	224		
Arg	Ile	Leu	Thr	Ala	Thr	Leu	Leu	Val	Leu	Thr	Thr	Ala	Thr	Ala	Gln			
			10						15					20				
cga	gat	cga	tgt	ggc	tac	atg	gta	gaa	ata	ccc	aga	cca	gac	agg	cct	272		
Arg	Asp	Arg	Cys	Gly	Tyr	Met	Val	Glu	Ile	Pro	Arg	Pro	Asp	Arg	Pro			
		25						30					35					
gac	ttc	cca	cct	caa	aat	ttt	gac	ggg	tta	aca	tgg	gct	cag	cag	cca	320		
Asp	Phe	Pro	Pro	Gln	Asn	Phe	Asp	Gly	Leu	Thr	Trp	Ala	Gln	Gln	Pro			
		40				45						50						
cta	tta	cca	gct	gag	gat	cga	gaa	gag	gtc	tgc	ctc	aat	gac	tat	gaa	368		
Leu	Leu	Pro	Ala	Glu	Asp	Arg	Glu	Glu	Val	Cys	Leu	Asn	Asp	Tyr	Glu			
	55					60					65							
cct	gat	ccc	tgg	agc	aac	aac	cat	ggg	gac	cag	aga	att	tac	atg	gag	416		
Pro	Asp	Pro	Trp	Ser	Asn	Asn	His	Gly	Asp	Gln	Arg	Ile	Tyr	Met	Glu			
	70				75					80					85			
gag	gag	atc	gaa	ggg	ccc	gta	gtc	att	gcg	aaa	att	aac	tac	caa	gga	464		
Glu	Glu	Ile	Glu	Gly	Pro	Val	Val	Ile	Ala	Lys	Ile	Asn	Tyr	Gln	Gly			
			90					95						100				
aac	acc	cct	cct	caa	ata	aga	tta	cct	ttt	cgt	ggt	ggt	gca	gcc	cac	512		
Asn	Thr	Pro	Pro	Gln	Ile	Arg	Leu	Pro	Phe	Arg	Val	Gly	Ala	Ala	His			
		105						110					115					
atg	ctt	gga	gca	gaa	att	cgt	gaa	tat	cct	gac	gca	act	gga	gac	tgg	560		
Met	Leu	Gly	Ala	Glu	Ile	Arg	Glu	Tyr	Pro	Asp	Ala	Thr	Gly	Asp	Trp			
		120					125					130						
tat	ctt	gta	att	act	caa	agg	cag	gac	tat	gaa	act	cct	gat	atg	cag	608		
Tyr	Leu	Val	Ile	Thr	Gln	Arg	Gln	Asp	Tyr	Glu	Thr	Pro	Asp	Met	Gln			
	135					140					145							
aga	tac	acg	ttc	gat	gtg	agt	gtg	gaa	ggc	cag	tcg	ctg	ggt	gta	acg	656		
Arg	Tyr	Thr	Phe	Asp	Val	Ser	Val	Glu	Gly	Gln	Ser	Leu	Val	Val	Thr			
	150				155					160					165			
gtg	agg	ctg	gat	att	gtg	aac	atc	gac	gac	aat	gcg	ccc	atc	att	gag	704		
Val	Arg	Leu	Asp	Ile	Val	Asn	Ile	Asp	Asp	Asn	Ala	Pro	Ile	Ile	Glu			
			170					175						180				
atg	tta	gag	cct	tgc	aac	tta	ccg	gaa	ctt	ggt	gaa	ccc	cat	ggt	aca	752		
Met	Leu	Glu	Pro	Cys	Asn	Leu	Pro	Glu	Leu	Val	Glu	Pro	His	Val	Thr			
		185						190					195					
gaa	tgt	aaa	tat	atc	gtg	tcc	gac	gca	gac	ggg	ctg	atc	agt	aca	agt	800		
Glu	Cys	Lys	Tyr	Ile	Val	Ser	Asp	Ala	Asp	Gly	Leu	Ile	Ser	Thr	Ser			
		200					205					210						
ggt	atg	agt	tat	cat	ata	gac	agc	gag	aga	gga	gac	gaa	aaa	gta	ttc	848		
Val	Met	Ser	Tyr	His	Ile	Asp	Ser	Glu	Arg	Gly	Asp	Glu	Lys	Val	Phe			
	215					220					225							
gaa	ctg	atc	aga	aaa	gat	tat	ccg	ggc	gat	tgg	acg	aag	gtg	tat	atg	896		
Glu	Leu	Ile	Arg	Lys	Asp	Tyr	Pro	Gly	Asp	Trp	Thr	Lys	Val	Tyr	Met			
	230				235				240					245				
ggt	ctt	gaa	ttg	aaa	aaa	tct	ctt	gat	tac	gaa	gag	aat	cct	cta	cac	944		
Val	Leu	Glu	Leu	Lys	Lys	Ser	Leu	Asp	Tyr	Glu	Glu	Asn	Pro	Leu	His			
			250						255					260				
ata	ttc	aga	gtc	acg	gct	tct	gat	tcc	tta	cca	aac	aat	agg	acc	gtg	992		
Ile	Phe	Arg	Val	Thr	Ala	Ser	Asp	Ser	Leu	Pro	Asn	Asn	Arg	Thr	Val			
		265					270						275					
gtc	atg	atg	ggt	gaa	gta	gag	aac	gtg	gaa	cat	aga	aat	cct	cgg	tgg	1040		
Val	Met	Met	Val	Glu	Val	Glu	Asn	Val	Glu	His	Arg	Asn	Pro	Arg	Trp			
		280					285					290						
atg	gag	atc	ttt	gct	gtg	caa	cag	ttt	gat	gaa	aaa	cag	gcg	aaa	tcg	1088		
Met	Glu	Ile	Phe	Ala	Val	Gln	Gln	Phe	Asp	Glu	Lys	Gln	Ala	Lys	Ser			
	295					300					305							
ttc	aca	gtg	cga	gct	att	gat	ggc	gac	acg	gga	atc	aat	aaa	cct	ata	1136		
Phe	Thr	Val	Arg	Ala	Ile	Asp	Gly	Asp	Thr	Gly	Ile	Asn	Lys	Pro	Ile			

-continued

310	315	320	325	
ttc tat cgt ata gaa act gaa gat gaa gac aaa gag ttc ttc agc att				1184
Phe Tyr Arg Ile Glu Thr Glu Asp Glu Asp Lys Glu Phe Phe Ser Ile				
	330	335	340	
gag aac ata ggg gaa ggc aga gac ggt gcc aga ttc cac gtg gct cct				1232
Glu Asn Ile Gly Glu Gly Arg Asp Gly Ala Arg Phe His Val Ala Pro				
	345	350	355	
ata gac aga gac tac ctg aaa agg gat atg ttt cat ata aga ata att				1280
Ile Asp Arg Asp Tyr Leu Lys Arg Asp Met Phe His Ile Arg Ile Ile				
	360	365	370	
gca tat aaa caa ggt gat aat gac aaa gaa ggt gaa tca tcg ttc gag				1328
Ala Tyr Lys Gln Gly Asp Asn Asp Lys Glu Gly Glu Ser Ser Phe Glu				
	375	380	385	
acc tca gca aat gtg acg att ata att aac gat ata aat gat cag agg				1376
Thr Ser Ala Asn Val Thr Ile Ile Ile Asn Asp Ile Asn Asp Gln Arg				
	390	395	400	405
cca gaa ccc ttc cat aaa gaa tac acg atc tcc ata atg gaa gaa act				1424
Pro Glu Pro Phe His Lys Glu Tyr Thr Ile Ser Ile Met Glu Glu Thr				
	410	415	420	
gcg atg acc tta gat ttg caa gag ttt ggt ttc cat gac cgt gac att				1472
Ala Met Thr Leu Asp Leu Gln Glu Phe Gly Phe His Asp Arg Asp Ile				
	425	430	435	
ggt ccc cac gct cag tac gac gtt cac tta gag agt ata cag cca gag				1520
Gly Pro His Ala Gln Tyr Asp Val His Leu Glu Ser Ile Gln Pro Glu				
	440	445	450	
ggg gcc cat acc gct ttc tac atc gcc cct gaa gaa ggt tac cag gcc				1568
Gly Ala His Thr Ala Phe Tyr Ile Ala Pro Glu Glu Gly Tyr Gln Ala				
	455	460	465	
cag tct ttc acc ata ggt act aga atc cat aac atg ttg gat tat gaa				1616
Gln Ser Phe Thr Ile Gly Thr Arg Ile His Asn Met Leu Asp Tyr Glu				
	470	475	480	485
gat gac gac tac aga cca gga ata aag cta aag gca gta gca att gac				1664
Asp Asp Asp Tyr Arg Pro Gly Ile Lys Leu Lys Ala Val Ala Ile Asp				
	490	495	500	
aga cac gat aac aat cac att ggg gaa gca att att aac att aac ctt				1712
Arg His Asp Asn Asn His Ile Gly Glu Ala Ile Ile Asn Ile Asn Leu				
	505	510	515	
atc aat tgg aat gat gag cta cct ata ttc gac gag gac gcc tac aac				1760
Ile Asn Trp Asn Asp Glu Leu Pro Ile Phe Asp Glu Asp Ala Tyr Asn				
	520	525	530	
gtg aca ttt gag gag acg gtc ggt gat ggc ttc cac att ggt aaa tac				1808
Val Thr Phe Glu Glu Thr Val Gly Asp Gly Phe His Ile Gly Lys Tyr				
	535	540	545	
cgg gct aaa gac aga gac atc ggt gac ata gtc gag cac tcg ata ttg				1856
Arg Ala Lys Asp Arg Asp Ile Gly Asp Ile Val Glu His Ser Ile Leu				
	550	555	560	565
ggc aac gct gca aac ttc ctg aga att gac ata gat act gga gat gtg				1904
Gly Asn Ala Ala Asn Phe Leu Arg Ile Asp Ile Asp Thr Gly Asp Val				
	570	575	580	
tac gtg tca cgg gac gat tac ttt gat tat caa aga cag aac gaa atc				1952
Tyr Val Ser Arg Asp Asp Tyr Phe Asp Tyr Gln Arg Gln Asn Glu Ile				
	585	590	595	
ata gtt cag att ctg gct gtt gat aca cta ggt tta cct cag aac agg				2000
Ile Val Gln Ile Leu Ala Val Asp Thr Leu Gly Leu Pro Gln Asn Arg				
	600	605	610	
gct acc aca cag ctc acg ata ttt ttg gaa gac atc aac aac acg cca				2048
Ala Thr Thr Gln Leu Thr Ile Phe Leu Glu Asp Ile Asn Asn Thr Pro				
	615	620	625	
cct ata ctg cga ctg cca cgt tcc agt cca agt gta gaa gag aac gtt				2096
Pro Ile Leu Arg Leu Pro Arg Ser Ser Pro Ser Val Glu Glu Asn Val				

-continued

630	635	640	645	
gaa gtc ggg cac ccg att acc gag ggg cta acg gcg aca gac cca gac Glu Val Gly His Pro Ile Thr Glu Gly Leu Thr Ala Thr Asp Pro Asp 650 655 660				2144
acc aca gcc gat tta cac ttc gag atc gat tgg gac aat tct tac gct Thr Thr Ala Asp Leu His Phe Glu Ile Asp Trp Asp Asn Ser Tyr Ala 665 670 675				2192
acg aag cag ggc acc aat gga ccc aac act gca gac tac cac gga tgc Thr Lys Gln Gly Thr Asn Gly Pro Asn Thr Ala Asp Tyr His Gly Cys 680 685 690				2240
gta gaa atc ctg acg gta tac cca gat cct gac aat cac ggg aga gct Val Glu Ile Leu Thr Val Tyr Pro Asp Pro Asp Asn His Gly Arg Ala 695 700 705				2288
gag ggt cac ttg gtg gca cgt gag gtc agt gat ggc gtg acc atc gat Glu Gly His Leu Val Ala Arg Glu Val Ser Asp Gly Val Thr Ile Asp 710 715 720 725				2336
tac gag aag ttt gag gtg ctg tac ctc gtc gtc agg gtg ata gat cgc Tyr Glu Lys Phe Glu Val Leu Tyr Leu Val Val Arg Val Ile Asp Arg 730 735 740				2384
aac act gtc att ggc cct gat tat gac gaa gca atg ctg acg gtg acg Asn Thr Val Ile Gly Pro Asp Tyr Asp Glu Ala Met Leu Thr Val Thr 745 750 755				2432
ata atc gat atg aac gac aac tgg ccg ata tgg gcc gac aac acg ctg Ile Ile Asp Met Asn Asp Asn Trp Pro Ile Trp Ala Asp Asn Thr Leu 760 765 770				2480
cag cag aca ctg cgc gtg cgc gag atg gcc gac gaa gga gtc atc gtc Gln Gln Thr Leu Arg Val Arg Glu Met Ala Asp Glu Gly Val Ile Val 775 780 785				2528
ggt aca ctg ctc gcc acc gac ttg gat ggc cct ctc tac aac cga gtc Gly Thr Leu Leu Ala Thr Asp Leu Asp Gly Pro Leu Tyr Asn Arg Val 790 795 800 805				2576
cgc tac acc atg gtc ccc atc aag gac act cct gat gac cta ata gcg Arg Tyr Thr Met Val Pro Ile Lys Asp Thr Pro Asp Asp Leu Ile Ala 810 815 820				2624
atc aac tac gtc acc ggt cag ctg act gtg aac aag ggg caa gca att Ile Asn Tyr Val Thr Gly Gln Leu Thr Val Asn Lys Gly Gln Ala Ile 825 830 835				2672
gac gca gat gat cca cct cgc ttc tac ctg tat tac aag gtc act gcc Asp Ala Asp Asp Pro Pro Arg Phe Tyr Leu Tyr Tyr Lys Val Thr Ala 840 845 850				2720
agc gat aag tgc tct ctt gac gag ttc ttc cct gtg tgc cca cct gac Ser Asp Lys Cys Ser Leu Asp Glu Phe Phe Pro Val Cys Pro Pro Asp 855 860 865				2768
ccc act tac tgg aat acc gag gga gag ata gcg atc gcg ata acc gat Pro Thr Tyr Trp Asn Thr Glu Gly Glu Ile Ala Ile Ala Ile Thr Asp 870 875 880 885				2816
acg aac aac aaa att cca cgc gcg gaa aca gat atg ttc cct agt gaa Thr Asn Asn Lys Ile Pro Arg Ala Glu Thr Asp Met Phe Pro Ser Glu 890 895 900				2864
aag cgc atc tat gag aac aca ccc aat ggt acc aag atc acg acg atc Lys Arg Ile Tyr Glu Asn Thr Pro Asn Gly Thr Lys Ile Thr Thr Ile 905 910 915				2912
atc gct agt gac cag gac aga gat cga cca aat aac gcg ctg acg tac Ile Ala Ser Asp Gln Asp Arg Asp Arg Pro Asn Asn Ala Leu Thr Tyr 920 925 930				2960
aga atc aac tac gca ttc aac cac agg ctg gag aac ttc ttc gca gtg Arg Ile Asn Tyr Ala Phe Asn His Arg Leu Glu Asn Phe Phe Ala Val 935 940 945				3008
gac cct gat act ggt gaa ctg ttt gtc cac ttc acc act agc gaa gtg Asp Pro Asp Thr Gly Glu Leu Phe Val His Phe Thr Thr Ser Glu Val				3056

-continued

950	955	960	965	
ttg gac aga gac gga gag gaa ccg gag cat agg atc atc ttc acc atc Leu Asp Arg Asp Gly Glu Glu Pro Glu His Arg Ile Ile Phe Thr Ile 970 975 980				3104
gtc gat aac ttg gaa ggc gct gga gat ggc aat cag aac aca atc tcc Val Asp Asn Leu Glu Gly Ala Gly Asp Gly Asn Gln Asn Thr Ile Ser 985 990 995				3152
acg gag gtg cgt gtt ata ctg ctt gat ata aac gac aat aag ccg gaa Thr Glu Val Arg Val Ile Leu Leu Asp Ile Asn Asp Asn Lys Pro Glu 1000 1005 1010				3200
cta cca att cct gat ggc gaa ttt tgg acc gtt tcc gaa ggt gaa gtg Leu Pro Ile Pro Asp Gly Glu Phe Trp Thr Val Ser Glu Gly Glu Val 1015 1020 1025				3248
gag gga aaa cgc att cca cca gag att cac gca cac gac aga gat gaa Glu Gly Lys Arg Ile Pro Pro Glu Ile His Ala His Asp Arg Asp Glu 1030 1035 1040 1045				3296
cca ttc aac gac aac tct cgc gtg gga tat gaa att cga tcg atc aaa Pro Phe Asn Asp Asn Ser Arg Val Gly Tyr Glu Ile Arg Ser Ile Lys 1050 1055 1060				3344
ttg atc aat aga gac atc gag ctt cct caa gat cca ttc aaa ata ata Leu Ile Asn Arg Asp Ile Glu Leu Pro Gln Asp Pro Phe Lys Ile Ile 1065 1070 1075				3392
acg att gat gat ctc gat acc tgg aaa ttc gtt gga gag ttg gag act Thr Ile Asp Asp Leu Asp Thr Trp Lys Phe Val Gly Glu Leu Glu Thr 1080 1085 1090				3440
acc atg gac ctt aga gga tac tgg gga acc tat gat gtc gag ata cgt Thr Met Asp Leu Arg Gly Tyr Trp Gly Thr Tyr Asp Val Glu Ile Arg 1095 1100 1105				3488
gcg ttt gac cac ggt ttc ccg atg ctg gat tca ttc gag acc tac caa Ala Phe Asp His Gly Phe Pro Met Leu Asp Ser Phe Glu Thr Tyr Gln 1110 1115 1120 1125				3536
cta acc gtc agg cca tac aac ttc cat tca ccg gtg ttt gtg ttc cca Leu Thr Val Arg Pro Tyr Asn Phe His Ser Pro Val Phe Val Phe Pro 1130 1135 1140				3584
act cct ggc tca acc atc agg ctt tct agg gag cgt gct ata gtc aat Thr Pro Gly Ser Thr Ile Arg Leu Ser Arg Glu Arg Ala Ile Val Asn 1145 1150 1155				3632
ggt atg ctg gct ctg gct aat atc gcg agc gga gag ttc ctc gac aga Gly Met Leu Ala Leu Ala Asn Ile Ala Ser Gly Glu Phe Leu Asp Arg 1160 1165 1170				3680
ctc tct gcc act gat gaa gat ggg cta cac gca ggc aga gta act ttc Leu Ser Ala Thr Asp Glu Asp Gly Leu His Ala Gly Arg Val Thr Phe 1175 1180 1185				3728
tcc ata gct gga aac gat gaa gct gcg gaa tat ttc aat gtg ttg aac Ser Ile Ala Gly Asn Asp Glu Ala Ala Glu Tyr Phe Asn Val Leu Asn 1190 1195 1200 1205				3776
gac ggt gac aac tca gca atg ctc acg ctg aag caa gca ttg ccc gct Asp Gly Asp Asn Ser Ala Met Leu Thr Leu Lys Gln Ala Leu Pro Ala 1210 1215 1220				3824
ggc gtc cag cag ttt gag ttg gtt att cgg gcc acg gac ggc ggg acg Gly Val Gln Gln Phe Glu Leu Val Ile Arg Ala Thr Asp Gly Gly Thr 1225 1230 1235				3872
gag ccg gga cct agg agt acc gac tgc tcc gtc act gtg gtg ttt gtg Glu Pro Gly Pro Arg Ser Thr Asp Cys Ser Val Thr Val Val Phe Val 1240 1245 1250				3920
atg acg cag gga gac ccc gtg ttc gac gac aac gca gct tct gtc cgc Met Thr Gln Gly Asp Pro Val Phe Asp Asp Asn Ala Ala Ser Val Arg 1255 1260 1265				3968
ttc gtt gaa aag gaa gct ggt atg tcg gaa aag ttt cag ctg cct cag Phe Val Glu Lys Glu Ala Gly Met Ser Glu Lys Phe Gln Leu Pro Gln				4016

-continued

1270	1275	1280	1285	
gcc gat gac ccc aaa aac tac agg tgt atg gac gac tgc cat acc atc				4064
Ala Asp Asp Pro Lys Asn Tyr Arg Cys Met Asp Asp Cys His Thr Ile				
	1290	1295	1300	
tac tac tct atc gtt gat ggc aac gat ggt gac cac ttc gcc gtg gag				4112
Tyr Tyr Ser Ile Val Asp Gly Asn Asp Gly Asp His Phe Ala Val Glu				
	1305	1310	1315	
ccg gag act aac gtg atc tat ttg ctg aag ccg ctg gac cgc agc caa				4160
Pro Glu Thr Asn Val Ile Tyr Leu Leu Lys Pro Leu Asp Arg Ser Gln				
	1320	1325	1330	
cag gag cag tac agg gtc gtg gtg gcg gct tcc aac acg cct ggc ggc				4208
Gln Glu Gln Tyr Arg Val Val Val Ala Ala Ser Asn Thr Pro Gly Gly				
	1335	1340	1345	
acc tcc acc ttg tcc tcc tca ctc ctc acc gtc acc atc ggc gtt cga				4256
Thr Ser Thr Leu Ser Ser Ser Leu Leu Thr Val Thr Ile Gly Val Arg				
	1350	1355	1360	1365
gaa gca aac cct aga ccg atc ttc gaa agt gaa ttt tac aca gct ggc				4304
Glu Ala Asn Pro Arg Pro Ile Phe Glu Ser Glu Phe Tyr Thr Ala Gly				
	1370	1375	1380	
gtc tta cac acc gat agc ata cac aag gag ctc gtt tac ctg gcg gca				4352
Val Leu His Thr Asp Ser Ile His Lys Glu Leu Val Tyr Leu Ala Ala				
	1385	1390	1395	
aaa cat tca gaa ggg ctt cct atc gtc tac tcg ata gat caa gaa acc				4400
Lys His Ser Glu Gly Leu Pro Ile Val Tyr Ser Ile Asp Gln Glu Thr				
	1400	1405	1410	
atg aaa ata gac gag tcg ttg caa aca gtt gtg gag gac gcc ttc gac				4448
Met Lys Ile Asp Glu Ser Leu Gln Thr Val Val Glu Asp Ala Phe Asp				
	1415	1420	1425	
att aac tct gca acc gga gtc ata tcg ctg aac ttc cag cca aca tct				4496
Ile Asn Ser Ala Thr Gly Val Ile Ser Leu Asn Phe Gln Pro Thr Ser				
	1430	1435	1440	1445
gtc atg cac ggc agt ttc gac ttc gag gtg gtg gct agt gac acg cgt				4544
Val Met His Gly Ser Phe Asp Phe Glu Val Val Ala Ser Asp Thr Arg				
	1450	1455	1460	
gga gcg agt gat cga gca aaa gtg tca att tac atg ata tcg act cgc				4592
Gly Ala Ser Asp Arg Ala Lys Val Ser Ile Tyr Met Ile Ser Thr Arg				
	1465	1470	1475	
gtt aga gta gcc ttc ctg ttc tac aac acg gaa gct gaa gtt aac gag				4640
Val Arg Val Ala Phe Leu Phe Tyr Asn Thr Glu Ala Glu Val Asn Glu				
	1480	1485	1490	
aga aga aat ttc att gca caa acg ttc gcc aac gcg ttt ggt atg aca				4688
Arg Arg Asn Phe Ile Ala Gln Thr Phe Ala Asn Ala Phe Gly Met Thr				
	1495	1500	1505	
tgt aac ata gac agc gtg ctg ccg gct acc gac gcc aac ggc gtg att				4736
Cys Asn Ile Asp Ser Val Leu Pro Ala Thr Asp Ala Asn Gly Val Ile				
	1510	1515	1520	1525
cgc gag ggg tac aca gaa ctc cag gct cac ttc ata cga gac gac cag				4784
Arg Glu Gly Tyr Thr Glu Leu Gln Ala His Phe Ile Arg Asp Asp Gln				
	1530	1535	1540	
ccg gtg cca gcc gac tat att gag gga tta ttt acg gaa ctc aat aca				4832
Pro Val Pro Ala Asp Tyr Ile Glu Gly Leu Phe Thr Glu Leu Asn Thr				
	1545	1550	1555	
ttg cgt gac atc aga gag gta ctg agt act cag caa ttg acg cta ctg				4880
Leu Arg Asp Ile Arg Glu Val Leu Ser Thr Gln Gln Leu Thr Leu Leu				
	1560	1565	1570	
gac ttt gcg gcg gga ggg tcg gca gtg ctg ccc ggc gga gag tac gcg				4928
Asp Phe Ala Ala Gly Gly Ser Ala Val Leu Pro Gly Gly Glu Tyr Ala				
	1575	1580	1585	
cta gcg gtg tac atc ctc gcc ggc atc gca gcg tta ctc gcc gtc atc				4976
Leu Ala Val Tyr Ile Leu Ala Gly Ile Ala Ala Leu Leu Ala Val Ile				

-continued

Ser Leu Val Val Thr Val Arg Leu Asp Ile Val Asn Ile Asp Asp Asn
 165 170 175
 Ala Pro Ile Ile Glu Met Leu Glu Pro Cys Asn Leu Pro Glu Leu Val
 180 185 190
 Glu Pro His Val Thr Glu Cys Lys Tyr Ile Val Ser Asp Ala Asp Gly
 195 200 205
 Leu Ile Ser Thr Ser Val Met Ser Tyr His Ile Asp Ser Glu Arg Gly
 210 215 220
 Asp Glu Lys Val Phe Glu Leu Ile Arg Lys Asp Tyr Pro Gly Asp Trp
 225 230 235 240
 Thr Lys Val Tyr Met Val Leu Glu Leu Lys Lys Ser Leu Asp Tyr Glu
 245 250 255
 Glu Asn Pro Leu His Ile Phe Arg Val Thr Ala Ser Asp Ser Leu Pro
 260 265 270
 Asn Asn Arg Thr Val Val Met Met Val Glu Val Glu Asn Val Glu His
 275 280 285
 Arg Asn Pro Arg Trp Met Glu Ile Phe Ala Val Gln Gln Phe Asp Glu
 290 295 300
 Lys Gln Ala Lys Ser Phe Thr Val Arg Ala Ile Asp Gly Asp Thr Gly
 305 310 315 320
 Ile Asn Lys Pro Ile Phe Tyr Arg Ile Glu Thr Glu Asp Glu Asp Lys
 325 330 335
 Glu Phe Phe Ser Ile Glu Asn Ile Gly Glu Gly Arg Asp Gly Ala Arg
 340 345 350
 Phe His Val Ala Pro Ile Asp Arg Asp Tyr Leu Lys Arg Asp Met Phe
 355 360 365
 His Ile Arg Ile Ile Ala Tyr Lys Gln Gly Asp Asn Asp Lys Glu Gly
 370 375 380
 Glu Ser Ser Phe Glu Thr Ser Ala Asn Val Thr Ile Ile Ile Asn Asp
 385 390 395 400
 Ile Asn Asp Gln Arg Pro Glu Pro Phe His Lys Glu Tyr Thr Ile Ser
 405 410 415
 Ile Met Glu Glu Thr Ala Met Thr Leu Asp Leu Gln Glu Phe Gly Phe
 420 425 430
 His Asp Arg Asp Ile Gly Pro His Ala Gln Tyr Asp Val His Leu Glu
 435 440 445
 Ser Ile Gln Pro Glu Gly Ala His Thr Ala Phe Tyr Ile Ala Pro Glu
 450 455 460
 Glu Gly Tyr Gln Ala Gln Ser Phe Thr Ile Gly Thr Arg Ile His Asn
 465 470 475 480
 Met Leu Asp Tyr Glu Asp Asp Asp Tyr Arg Pro Gly Ile Lys Leu Lys
 485 490 495
 Ala Val Ala Ile Asp Arg His Asp Asn Asn His Ile Gly Glu Ala Ile
 500 505 510
 Ile Asn Ile Asn Leu Ile Asn Trp Asn Asp Glu Leu Pro Ile Phe Asp
 515 520 525
 Glu Asp Ala Tyr Asn Val Thr Phe Glu Glu Thr Val Gly Asp Gly Phe
 530 535 540
 His Ile Gly Lys Tyr Arg Ala Lys Asp Arg Asp Ile Gly Asp Ile Val
 545 550 555 560
 Glu His Ser Ile Leu Gly Asn Ala Ala Asn Phe Leu Arg Ile Asp Ile
 565 570 575
 Asp Thr Gly Asp Val Tyr Val Ser Arg Asp Asp Tyr Phe Asp Tyr Gln

-continued

580					585					590					
Arg	Gln	Asn	Glu	Ile	Ile	Val	Gln	Ile	Leu	Ala	Val	Asp	Thr	Leu	Gly
		595					600					605			
Leu	Pro	Gln	Asn	Arg	Ala	Thr	Thr	Gln	Leu	Thr	Ile	Phe	Leu	Glu	Asp
	610					615					620				
Ile	Asn	Asn	Thr	Pro	Pro	Ile	Leu	Arg	Leu	Pro	Arg	Ser	Ser	Pro	Ser
625					630					635					640
Val	Glu	Glu	Asn	Val	Glu	Val	Gly	His	Pro	Ile	Thr	Glu	Gly	Leu	Thr
				645					650					655	
Ala	Thr	Asp	Pro	Asp	Thr	Thr	Ala	Asp	Leu	His	Phe	Glu	Ile	Asp	Trp
			660					665					670		
Asp	Asn	Ser	Tyr	Ala	Thr	Lys	Gln	Gly	Thr	Asn	Gly	Pro	Asn	Thr	Ala
		675					680					685			
Asp	Tyr	His	Gly	Cys	Val	Glu	Ile	Leu	Thr	Val	Tyr	Pro	Asp	Pro	Asp
	690					695					700				
Asn	His	Gly	Arg	Ala	Glu	Gly	His	Leu	Val	Ala	Arg	Glu	Val	Ser	Asp
705				710						715					720
Gly	Val	Thr	Ile	Asp	Tyr	Glu	Lys	Phe	Glu	Val	Leu	Tyr	Leu	Val	Val
				725					730					735	
Arg	Val	Ile	Asp	Arg	Asn	Thr	Val	Ile	Gly	Pro	Asp	Tyr	Asp	Glu	Ala
			740					745					750		
Met	Leu	Thr	Val	Thr	Ile	Ile	Asp	Met	Asn	Asp	Asn	Trp	Pro	Ile	Trp
		755					760					765			
Ala	Asp	Asn	Thr	Leu	Gln	Gln	Thr	Leu	Arg	Val	Arg	Glu	Met	Ala	Asp
	770					775					780				
Glu	Gly	Val	Ile	Val	Gly	Thr	Leu	Leu	Ala	Thr	Asp	Leu	Asp	Gly	Pro
785				790						795					800
Leu	Tyr	Asn	Arg	Val	Arg	Tyr	Thr	Met	Val	Pro	Ile	Lys	Asp	Thr	Pro
			805						810				815		
Asp	Asp	Leu	Ile	Ala	Ile	Asn	Tyr	Val	Thr	Gly	Gln	Leu	Thr	Val	Asn
			820					825					830		
Lys	Gly	Gln	Ala	Ile	Asp	Ala	Asp	Asp	Pro	Pro	Arg	Phe	Tyr	Leu	Tyr
		835					840					845			
Tyr	Lys	Val	Thr	Ala	Ser	Asp	Lys	Cys	Ser	Leu	Asp	Glu	Phe	Phe	Pro
	850					855					860				
Val	Cys	Pro	Pro	Asp	Pro	Thr	Tyr	Trp	Asn	Thr	Glu	Gly	Glu	Ile	Ala
865				870						875					880
Ile	Ala	Ile	Thr	Asp	Thr	Asn	Asn	Lys	Ile	Pro	Arg	Ala	Glu	Thr	Asp
			885						890					895	
Met	Phe	Pro	Ser	Glu	Lys	Arg	Ile	Tyr	Glu	Asn	Thr	Pro	Asn	Gly	Thr
		900						905					910		
Lys	Ile	Thr	Thr	Ile	Ile	Ala	Ser	Asp	Gln	Asp	Arg	Asp	Arg	Pro	Asn
		915					920					925			
Asn	Ala	Leu	Thr	Tyr	Arg	Ile	Asn	Tyr	Ala	Phe	Asn	His	Arg	Leu	Glu
	930					935					940				
Asn	Phe	Phe	Ala	Val	Asp	Pro	Asp	Thr	Gly	Glu	Leu	Phe	Val	His	Phe
945				950						955					960
Thr	Thr	Ser	Glu	Val	Leu	Asp	Arg	Asp	Gly	Glu	Glu	Pro	Glu	His	Arg
			965						970					975	
Ile	Ile	Phe	Thr	Ile	Val	Asp	Asn	Leu	Glu	Gly	Ala	Gly	Asp	Gly	Asn
			980					985					990		
Gln	Asn	Thr	Ile	Ser	Thr	Glu	Val	Arg	Val	Ile	Leu	Leu	Asp	Ile	Asn
		995					1000						1005		

-continued

Asp Asn Lys Pro Glu Leu Pro Ile Pro Asp Gly Glu Phe Trp Thr Val
 1010 1015 1020

Ser Glu Gly Glu Val Glu Gly Lys Arg Ile Pro Pro Glu Ile His Ala
 1025 1030 1035 1040

His Asp Arg Asp Glu Pro Phe Asn Asp Asn Ser Arg Val Gly Tyr Glu
 1045 1050 1055

Ile Arg Ser Ile Lys Leu Ile Asn Arg Asp Ile Glu Leu Pro Gln Asp
 1060 1065 1070

Pro Phe Lys Ile Ile Thr Ile Asp Asp Leu Asp Thr Trp Lys Phe Val
 1075 1080 1085

Gly Glu Leu Glu Thr Thr Met Asp Leu Arg Gly Tyr Trp Gly Thr Tyr
 1090 1095 1100

Asp Val Glu Ile Arg Ala Phe Asp His Gly Phe Pro Met Leu Asp Ser
 1105 1110 1115 1120

Phe Glu Thr Tyr Gln Leu Thr Val Arg Pro Tyr Asn Phe His Ser Pro
 1125 1130 1135

Val Phe Val Phe Pro Thr Pro Gly Ser Thr Ile Arg Leu Ser Arg Glu
 1140 1145 1150

Arg Ala Ile Val Asn Gly Met Leu Ala Leu Ala Asn Ile Ala Ser Gly
 1155 1160 1165

Glu Phe Leu Asp Arg Leu Ser Ala Thr Asp Glu Asp Gly Leu His Ala
 1170 1175 1180

Gly Arg Val Thr Phe Ser Ile Ala Gly Asn Asp Glu Ala Ala Glu Tyr
 1185 1190 1195 1200

Phe Asn Val Leu Asn Asp Gly Asp Asn Ser Ala Met Leu Thr Leu Lys
 1205 1210 1215

Gln Ala Leu Pro Ala Gly Val Gln Gln Phe Glu Leu Val Ile Arg Ala
 1220 1225 1230

Thr Asp Gly Gly Thr Glu Pro Gly Pro Arg Ser Thr Asp Cys Ser Val
 1235 1240 1245

Thr Val Val Phe Val Met Thr Gln Gly Asp Pro Val Phe Asp Asp Asn
 1250 1255 1260

Ala Ala Ser Val Arg Phe Val Glu Lys Glu Ala Gly Met Ser Glu Lys
 1265 1270 1275 1280

Phe Gln Leu Pro Gln Ala Asp Asp Pro Lys Asn Tyr Arg Cys Met Asp
 1285 1290 1295

Asp Cys His Thr Ile Tyr Tyr Ser Ile Val Asp Gly Asn Asp Gly Asp
 1300 1305 1310

His Phe Ala Val Glu Pro Glu Thr Asn Val Ile Tyr Leu Leu Lys Pro
 1315 1320 1325

Leu Asp Arg Ser Gln Gln Glu Gln Tyr Arg Val Val Val Ala Ala Ser
 1330 1335 1340

Asn Thr Pro Gly Gly Thr Ser Thr Leu Ser Ser Ser Leu Leu Thr Val
 1345 1350 1355 1360

Thr Ile Gly Val Arg Glu Ala Asn Pro Arg Pro Ile Phe Glu Ser Glu
 1365 1370 1375

Phe Tyr Thr Ala Gly Val Leu His Thr Asp Ser Ile His Lys Glu Leu
 1380 1385 1390

Val Tyr Leu Ala Ala Lys His Ser Glu Gly Leu Pro Ile Val Tyr Ser
 1395 1400 1405

Ile Asp Gln Glu Thr Met Lys Ile Asp Glu Ser Leu Gln Thr Val Val
 1410 1415 1420

Glu Asp Ala Phe Asp Ile Asn Ser Ala Thr Gly Val Ile Ser Leu Asn
 1425 1430 1435 1440

-continued

Phe Gln Pro Thr Ser Val Met His Gly Ser Phe Asp Phe Glu Val Val
 1445 1450 1455
 Ala Ser Asp Thr Arg Gly Ala Ser Asp Arg Ala Lys Val Ser Ile Tyr
 1460 1465 1470
 Met Ile Ser Thr Arg Val Arg Val Ala Phe Leu Phe Tyr Asn Thr Glu
 1475 1480 1485
 Ala Glu Val Asn Glu Arg Arg Asn Phe Ile Ala Gln Thr Phe Ala Asn
 1490 1495 1500
 Ala Phe Gly Met Thr Cys Asn Ile Asp Ser Val Leu Pro Ala Thr Asp
 1505 1510 1515 1520
 Ala Asn Gly Val Ile Arg Glu Gly Tyr Thr Glu Leu Gln Ala His Phe
 1525 1530 1535
 Ile Arg Asp Asp Gln Pro Val Pro Ala Asp Tyr Ile Glu Gly Leu Phe
 1540 1545 1550
 Thr Glu Leu Asn Thr Leu Arg Asp Ile Arg Glu Val Leu Ser Thr Gln
 1555 1560 1565
 Gln Leu Thr Leu Leu Asp Phe Ala Ala Gly Gly Ser Ala Val Leu Pro
 1570 1575 1580
 Gly Gly Glu Tyr Ala Leu Ala Val Tyr Ile Leu Ala Gly Ile Ala Ala
 1585 1590 1595 1600
 Leu Leu Ala Val Ile Cys Leu Ala Leu Leu Ile Ala Phe Phe Ile Arg
 1605 1610 1615
 Asn Arg Thr Leu Asn Arg Arg Ile Glu Ala Leu Thr Ile Lys Asp Val
 1620 1625 1630
 Pro Thr Asp Ile Glu Pro Asn His Ala Ser Val Ala Val Leu Asn Ile
 1635 1640 1645
 Asn Lys His Thr Glu Pro Gly Ser Asn Pro Phe Tyr Asn Pro Asp Val
 1650 1655 1660
 Lys Thr Pro Asn Phe Asp Thr Ile Ser Glu Val Ser Asp Asp Leu Leu
 1665 1670 1675 1680
 Asp Val Glu Asp Leu Glu Gln Phe Gly Lys Asp Tyr Phe Pro Pro Glu
 1685 1690 1695
 Asn Glu Ile Glu Ser Leu Asn Phe Ala Arg Asn Pro Ile Ala Thr His
 1700 1705 1710
 Gly Asn Asn Phe Gly Val Asn Ser Ser Pro Ser Asn Pro Glu Phe Ser
 1715 1720 1725
 Asn Ser Gln Phe Arg Ser
 1730

<210> SEQ ID NO 7

<211> LENGTH: 1604

<212> TYPE: DNA

<213> ORGANISM: *Ostrinia nubilalis*

<400> SEQUENCE: 7

tccgaattct tcttcaacct catcgacaac ttcttttctg acggtgacg taggagaaac 60
 caggacgaag ttgaaatatt tgtcgttcta ttggatgtga acgacaacgc tcttgagatg 120
 ccatcgctg atgaactccg gtttgatggt tccgaaggag cagttgctgg tgtccgtgta 180
 ctcccagaaa tctacgcacc tgacagggat gaaccagaca cggacaactc gcgtgtcgg 240
 tacggaatcc tggacctcac gatcaccgac cgagacatcg aggtgccgga tctcttcacc 300
 atgatctcga ttgaaaacaa aactggggaa cttgagaccg ctatggactt gagggggat 360
 tggggcactt acgaaatatt cattgaggcc ttcgaccacg gctaccgca gcagaggtcc 420

-continued

```

aacgggacgt acacactggt cattcgcccc tacaacttcc accaccctgt gttcgtgttc 480
ccgcaacccg actccgcat tgggtctctt agggagcgcg caacagaagg cggggtcctg 540
gcgacggctg ccaacgagtt cctggagccg atctacgcca ccgacgagga cggcctccac 600
gcgggcagcg tcacgttcca cgtccagggg aatgaggagg cgttcagta ctttgatata 660
actgaagtgg gagcaggaga aaatagcggg cagcttatat tacgccagct tttcccagag 720
caaatcagac aattcaggat cacgatccgg gccacagacg gcggcacgga gcccgccccg 780
ctttggaccg acgtcacgtt ttcgggtggtc ttcgtaccca cgcagggcga cccagtgttc 840
agcgaaaatg cagctactgt tgccctcttc gaggggtgaag aaggcctcca tgagagtttt 900
gagctgccgc aagcagaaga ccttaaaaac cacctctgcg aagatgactg ccaagatata 960
tactacaggt ttattgacgg caacaacgag ggtctgttcg tgctggacca gtogagcaac 1020
gtcatctccc ttgctcagga gttggaccgc gaggttgcca cgtcttacac gctgcacatc 1080
gcggcgagca actcgcccga cgccactggg atccctctgc agacttccat cctcgttgtc 1140
acgggtcaatg taagagaagc gaaccgcgcg ccaattttcg agcaggacct ttacacagcg 1200
ggcatttcga cgttgacag cattggccgg gaattgctta ccgtcagggc gagccacaca 1260
gaagacgaca ccatcacgta catcatagac cgtgagagca tgcagctgga cagcagccta 1320
gaagccgtgc gcgactcggc cttcacgctg catgagacca ccggcgtgct ttcgctcaat 1380
atgcagccca ccgcttccat gcacggcatg ttcgagttcg acgtcatcgc tacggataca 1440
gcatctgcaa tcgacacagc tcgtgtgaaa gtctacctca tctcatcga aaaccgctg 1500
tccttcattt tcgataacca acttgagacc gttgagcaga acagaaattt catagcggcc 1560
acgttcagca ccgggttcaa catgacgtgt aacatcgacc aagt 1604

```

```

<210> SEQ ID NO 8
<211> LENGTH: 23
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic oligonucleotide

```

```

<400> SEQUENCE: 8

```

```

gttamygtga gagaggcaga ycc 23

```

```

<210> SEQ ID NO 9
<211> LENGTH: 23
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic oligonucleotide

```

```

<400> SEQUENCE: 9

```

```

ggatrrtaag mgtcagyacw ccg 23

```

```

<210> SEQ ID NO 10
<211> LENGTH: 32
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic oligonucleotide

```

```

<400> SEQUENCE: 10

```

```

tccgaattct tcttyaacct catcgayaac tt 32

```

```

<210> SEQ ID NO 11
<211> LENGTH: 32
<212> TYPE: DNA

```


-continued

<213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Synthetic oligonucleotide

<400> SEQUENCE: 11

cgcaagctta cttggtcgat gttcrasgtc at

32

That which is claimed:

1. A method for screening candidate ligands to identify ligands that bind to an *Ostrinia nubilalis* insect receptor polypeptide, said method comprising:

a) providing at least one *Ostrinia nubilalis* insect receptor polypeptide, wherein the polypeptide is selected from the group consisting of:

i) the amino acid sequence set forth in SEQ ID NO: 2; and,

ii) the amino acid sequence of a sequence variant of the amino acid sequence set forth in SEQ ID NO: 2, wherein said sequence variant has Bt toxin binding activity and has at least 95% sequence identity to the amino acid sequence set forth in SEQ ID NO: 2;

b) contacting said polypeptide with a candidate ligand and a control ligand under conditions promoting binding of the candidate ligand or the control ligand to the polypeptide, wherein the control ligand is a Cry1A toxin; and

c) determining the binding characteristics of said candidate ligand, relative to said control ligand, wherein the binding characteristics are selected from the group consisting of binding affinity, binding site specificity, association rate, and dissociation rate, and thereby identifying a candidate ligand that binds to the *Ostrinia nubilalis* insect receptor polypeptide.

2. A method for screening candidate ligands to identify ligands that bind an *Ostrinia nubilalis* insect receptor polypeptide, said method comprising:

a) providing cells expressing at least one *Ostrinia nubilalis* insect receptor polypeptide wherein said polypeptide comprises a toxin binding domain and is selected from the group consisting of:

i) the amino acid sequence set forth in SEQ ID NO: 2; and,

ii) the amino acid sequence of a sequence variant of the amino acid sequence set forth in SEQ ID NO: 2, wherein said sequence variant has Bt toxin binding activity and has at least 95% sequence identity to the amino acid sequence set forth in SEQ ID NO: 2; and,

b) contacting said cells with a candidate ligand and a control ligand under conditions that promote binding of the candidate ligand or the control ligand to the polypeptide, wherein the control ligand is a Cry1A toxin; and

c) determining the binding characteristics of said candidate ligand, relative to said control ligand, wherein the binding characteristics are selected from the group consisting of binding affinity, binding site specificity, association rate, and dissociation rate, and thereby identifying a candidate ligand that binds to the *Ostrinia nubilalis* insect receptor polypeptide.

3. The method of claim 2, wherein said method further comprises the step of determining the viability of the cells contacted with the candidate ligand relative to the viability of the cells contacted with the control ligand.

* * * * *