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Baichwal et al.

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# (54) RIP: NOVEL HUMAN PROTEIN INVOLVED IN TUMOR NECROSIS FACTOR SIGNAL TRANSDUCTION

(75) Inventors: Vijay R. Baichwal, San Mateo; Jianing Huang, San Bruno; Hailing Hsu, Moon Park; David V. Goeddel, Hillsborough,

all of CA (US)

(73) Assignee: Tularik Inc., South San Francisco, CA (US)

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Notice:

(22) Filed: Aug. 11, 1998

# Related U.S. Application Data

- (63) Continuation-in-part of application No. 08/553,727, filed on Oct. 23, 1995, now abandoned.
- (51) Int. Cl.<sup>7</sup> ...... C07K 14/435

- (52) **U.S. Cl.** ...... **530/350**; 435/194; 436/501

# (56) References Cited

# U.S. PATENT DOCUMENTS

\* cited by examiner

Primary Examiner—Lorraine Spector Assistant Examiner—Eliane Lazar-Wesley

(74) Attorney, Agent, or Firm—Richard Aron Osman

# (57) ABSTRACT

The invention relates to a human Receptor Interacting Protein (hRIP), nucleic acids which encode hRIP and methods of using the subject compositions; in particular, methods such as hRIP-based in vitro binding assays and phosphorylation assays for screening chemical libraries for lead compounds for pharmacological agents.

### 24 Claims, No Drawings

# RIP: NOVEL HUMAN PROTEIN INVOLVED IN TUMOR NECROSIS FACTOR SIGNAL TRANSDUCTION

# CROSS-REFERENCE TO RELATED APPLICATION

This is a CIP of claims priority under 35USC120 to U.S. patent application Ser. No. 08/553,727, filed Oct. 23, 1995 now abandoned.

#### INTRODUCTION

#### 1. Field of the Invention

The field of this invention is a novel human kinase involved in tumor necrosis factor signal transduction and its 15 use in drug screening.

# 2. Background

Tumor necrosis factor (TNF) is an important cytokine involved in the signaling of a number of cellular responses  $_{20}$ including cytotoxicity, anti-viral activity, immun.-regulatory activities and the transcriptional regulation of a number of genes. The TNF receptors (TNF-R1 and TNF-R2) are members of the larger TNF receptor superfamily which also includes the Fas antigen, CD27, CD30, CD40, and the low affinity nerve growth factor receptor. Members of this family have been shown to participate in a variety of biological properties, including programmed cell death, antiviral activity and activation of the transcription factor NF-κB in a wide variety of cell types.

Accordingly, it is desired to identify agents which specifically modulate transduction of TNF receptor family signaling. Unfortunately, the components of the signaling pathway remain largely unknown; hence, the reagents necessary for the development of high-throughput screening 35 assays for such therapeutics are unavailable. Elucidation of TNF receptor family signal transduction pathways leading to NF-κB activation would provide valuable insight into mechanisms to alleviate inflammation. In particular, components of this pathway would provide valuable targets for 40 automated, cost-effective, high throughput drug screening and hence would have immediate application in a broad range of domestic and international pharmaceutical and biotechnology drug development programs. Relevant Literature

Stanger et al. (1995) Cell 81, 513–523 report the existence of a Receptor Interacting Protein (RIP) and its functional expression. VanArsdale and Ware (1994) J Immunology 153:3043–3050 describe proteins associated with TNF-R1. The cloning and amino acid sequencing of TNF-R1 is 50 disclosed in Schall et al (1990) Cell 61, 361 and Loetscher et al (1990) Cell 61, 351; the identification of a "death domain" in TNF-R1 is disclosed in Tartaglia et al. (1993) Cell 74:845–853. The cloning and amino acid sequence of a TNF-R associated death domain protein (TRADD) is 55 described by Hsu et al. (1995) Cell 81, 495–504. The cloning and amino acid sequence of the Fas antigen is disclosed in Itoh et al (1991) Cell 66, 233–243. For a recent review, see Smith et al. (1994) Cell 76:959–962 and Vandenabelle et al. (1995) Trends Cell Biol. 5, 392–399.

# SUMMARY OF THE INVENTION

The invention provides methods and compositions relating to a human Receptor Interacting Protein (hRIP). The compositions include nucleic acids which encode hRIP, 65  $\alpha\Delta3$  (SEQ ID NO:2, residues 506–514) hRIP kinase domains, and recombinant proteins made from these nucleic acids. The invention also provides methods for

screening chemical libraries for lead compounds for a pharmacological agent useful in the diagnosis or treatment of disease associated hRIP activity or hRIP-dependent signal transduction. In one embodiment, the methods involve incu-5 bating a mixture of hRIP, a natural intracellular hRIP substrate or binding target and a candidate pharmacological agent and determining if the presence of the agent modulates the ability of hRIP to selectively phosphorylate the substrate or bind the binding target. Specific agents provide lead 10 compounds for pharmacological agents capable of disrupting hRIP function.

# DETAILED DESCRIPTION OF THE INVENTION

A human RIP-encoding nucleic acid sequence is set out in SEQ ID NO: 1. A human RIP kinase domain-encoding nucleic acid sequence is set out in SEQ ID NO: 1, nucleotides 1–900. A human RIP amino acid sequence is set out in SEQ ID NO: 2; and a hRIP kinase domain sequence is set out in SEQ ID NO:2, residues 1–300.

Natural nucleic acids encoding hRIP are readily isolated from cDNA libraries with PCR primers and hybridization probes containing portions of the nucleic acid sequence of SEQ ID NO:1. For example, we used low stringency hybridization at 42° C. (hybridization buffer: 20% formamide, 10%) Denhardt, 0.5% SDS, 5×SSPE; with membrane washes at room temperature with 5×SSPE/0.5% SDS) with a 120 base oligonucleotide probe (SEQ ID NO: 1, nucleotides 1728-1847) to isolate a native human RIP cDNA from a library prepared from human umbilical vein endothelial cells. In addition, synthetic hRIP-encoding nucleic acids may be generated by automated synthesis.

The subject nucleic acids are recombinant, meaning they comprise a sequence joined to a nucleotide other than that to which sequence is naturally joined and isolated from a natural environment. The nucleic acids may be part of hRIP-expression vectors and may be incorporated into cells for expression and screening, transgenic animals for functional studies (e.g. the efficacy of candidate drugs for disease associated with expression of a hRIP), etc. These nucleic acids find a wide variety of applications including use as templates for transcription, hybridization probes, PCR primers, therapeutic nucleic acids, etc.; use in detecting the presence of hRIP genes and gene transcripts, in detecting or amplifying nucleic acids encoding additional hRIP homologs and structural analogs, and in gene therapy applications.

In a particular embodiment, the invention provides RIP-Thr<sup>514</sup> polypeptides, RIP-Thr<sup>514</sup> polypeptide-encoding nucleic acids/polynucleotides, and RIP-Thr<sup>514</sup> polypeptidebased methods (below), which RIP-Thr<sup>514</sup> polypeptides comprise at least 8, preferably at least 10, more preferably at least 12, more preferably at least 16, most preferably at least 24 consecutive amino acid residues of the amino acid sequence set forth as SEQ ID NO:2, which consecutive amino acid residues comprise the amino acid residue 514 (Thr) of SEQ ID NO:2. Exemplary RIP-Thr<sup>514</sup> polypeptides having RIP-Thr<sup>514</sup> binding specificity and immunologically distinguishable from RIP-Ser<sup>514</sup> are shown in Table I.

TABLE I. Exemplary RIP-Thr<sup>514</sup> Polypeptides Having RIP-Thr<sup>514</sup> Binding Specificity

 $\alpha\Delta 1$  (SEQ ID NO:2, residues 509–518)

 $\alpha\Delta 2$  (SEQ ID NO:2, residues 514–521)

 $\alpha \Delta 4$  (SEQ ID NO:2, residues 504–524)

 $\alpha \Delta 5$  (SEQ ID NO:2, residues 498–514)

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αΔ6 (SEQ ID NO:2, residues 514–534) αΔ7 (SEQ ID NO:2, residues 513–520) αΔ8 (SEQ ID NO:2, residues 508–515) αΔ9 (SEQ ID NO:2, residues 512–522) αΔ10 (SEQ ID NO:2, residues 423–514) αΔ11 (SEQ ID NO:2, residues 423–543) αΔ12 (SEQ ID NO:2, residues 423–579) αΔ13 (SEQ ID NO:2, residues 423–633) αΔ14 (SEQ ID NO:2, residues 423–671) αΔ15 (SEQ ID NO:2, residues 514–543) αΔ16 (SEQ ID NO:2, residues 514–543) αΔ16 (SEQ ID NO:2, residues 514–579) αΔ17 (SEQ ID NO:2, residues 514–671)
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In a particular embodiment, the invention provides RIP-ACA<sup>1540-1542</sup> polynucleotides, comprising at least 18, 24, 15 36, 48, 72, 148, 356 or 728 consecutive nucleotides of the nucleotide sequence set forth as SEQ ID NO:1, which consecutive polynucleotides comprise the polynucleotides 1540–1542 (ACA) of SEQ ID NO:1. Exemplary RIP-ACA<sup>1540-1542</sup> polynucleotides and allele specific oligonucleotide probes having RIP-ACA<sup>1540-1542</sup> binding specificity and distinguishable by hybridization assays from RIP-TCT<sup>1540-1542</sup> are shown in Table II.

TABLE II. Exemplary RIP-ACA<sup>1540-1542</sup> Polynucleotides Having RIP-ACA<sup>1540-1542</sup> Binding Specificity  $\alpha\Delta 1$  (SEQ ID NO:1, nucleotides 1540–1557)  $\alpha\Delta 2$  (SEQ ID NO:1, nucleotides 1540–1563)  $\alpha \Delta 3$  (SEQ ID NO:1, nucleotides 1540–1675)  $\alpha\Delta4$  (SEQ ID NO:1, nucleotides 1540–1699)  $\alpha \Delta 5$  (SEQ ID NO:1, nucleotides 1525–1542)  $\alpha\Delta6$  (SEQ ID NO:1, nucleotides 1519–1542)  $\alpha\Delta7$  (SEQ ID NO:1, nucleotides 1507–1542)  $\alpha \Delta 8$  (SEQ ID NO:1, nucleotides 1483–1542)  $\alpha\Delta9$  (SEQ ID NO:1, nucleotides 1537–1545)  $\alpha\Delta 10$  (SEQ ID NO:1, nucleotides 1534–1548)  $\alpha\Delta 11$  (SEQ ID NO:1, nucleotides 1528–1554)  $\alpha\Delta 12$  (SEQ ID NO:1, nucleotides 1516–1566)  $\alpha \Delta 13$  (SEQ ID NO:1, nucleotides 1504–1554)  $\alpha\Delta 14$  (SEQ ID NO:1, nucleotides 1492–1568)

The invention provides efficient methods of identifying 40 pharmacological agents or lead compounds for agents active at the level of a hRIP modulatable cellular function, particularly hRIP mediated TNF receptor or Tumor necrosis factor receptor associated Factor-2 (TRAF2) or TRADDinduced signal transduction. For example, we have found 45 that a binding complex comprising TNF R1, TRADD, and hRIP exists in TNF-stimulated cells. Generally, the screening methods involve assaying for compounds which interfere with a hRIP activity such as kinase activity or TRAF2 or TRADD binding. The methods are amenable to 50 automated, cost-effective high throughput screening of chemical libraries for lead compounds. Identified reagents find use in the pharmaceutical industries for animal and human trials; for example, the reagents may be derivatized and rescreened in in vitro and in vivo assays to optimize 55 activity and minimize toxicity for pharmaceutical development. Target therapeutic indications are limited only in that the target cellular function be subject to modulation, usually inhibition, by disruption of the formation of a complex comprising hRIP and one or more natural hRIP intracellular 60 binding targets including substrates or otherwise modulating hRIP kinase activity. Target indications may include infection, genetic disease, cell growth and regulatory or immunologic dysfunction, such as neoplasia, inflammation, hypersensitivity, etc.

A wide variety of assays for binding agents are provided including labeled in vitro kinase assays, protein-protein

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binding assays, immunoassays, cell based assays, etc. The hRIP compositions used in the methods are recombinantly produced from nucleic acids having the disclosed hRIP nucleotide sequences. The hRIP may be part of a fusion product with another peptide or polypeptide, e.g. a polypeptide that is capable of providing or enhancing protein-protein binding, stability under assay conditions (e.g. a tag for detection or anchoring), etc.

The assay mixtures comprise one or more natural intracellular hRIP binding targets including substrates, such as TRADD, TRAF2, or, in the case of an autophosphorylation assay, the hRIP itself can function as the binding target. In one embodiment, the mixture comprises a complex of hRIP, TRADD and TNFR1. A hRIP derived pseudosubstrate may be used or modified (e.g. A to S/T substitutions) to generate effective substrates for use in the subject kinase assays as can synthetic peptides or other protein substrates. Generally, hRIP-specificity of the binding agent is shown by kinase activity (i.e. the agent demonstrates activity of an hRIP substrate, agonist, antagonist, etc.) or binding equilibrium constants (usually at least about 10<sup>6</sup> M<sup>-1</sup>, preferably at least about  $10^8 \,\mathrm{M}^{-1}$ , more preferably at least about  $10^9 \,\mathrm{M}^{-1}$ . A wide variety of cell-based and cell-free assays may be used to demonstrate hRIP-specific binding; preferred are rapid in vitro, cell-free assays such as mediating or inhibiting hRIPprotein (e.g. hRIP-TRADD) binding, phosphorylation assays, immunoassays, etc.

The assay mixture also comprises a candidate pharmacological agent. Candidate agents encompass numerous chemical classes, though typically they are organic compounds; preferably small organic compounds and are obtained from a wide variety of sources including libraries of synthetic or natural compounds. A variety of other reagents may also be included in the mixture. These include reagents like salts, buffers, neutral proteins, e.g. albumin, detergents, etc. which may be used to facilitate optimal binding and/or reduce non-specific or background interactions, etc. Also, reagents that otherwise improve the efficiency of the assay, such as protease inhibitors, nuclease inhibitors, antimicrobial agents, etc. may be used.

In a preferred in vitro, binding assay, a mixture of at least the kinase domain of hRIP, one or more binding targets or substrates and the candidate agent is incubated under conditions whereby, but for the presence of the candidate pharmacological agent, the hRIP specifically binds the cellular binding target at a first binding affinity or phosphorylates the substrate at a first rate. After incubation, a second binding affinity or rate is detected. Detection may be effected in any convenient way. For cell-free binding assays, one of the components usually comprises or is coupled to a label. The label may provide for direct detection as radioactivity, luminescence, optical or electron density, etc. or indirect detection such as an epitope tag, an enzyme, etc. A variety of methods may be used to detect the label depending on the nature of the label and other assay components. For example, the label may be detected bound to the solid substrate or a portion of the bound complex containing the label may be separated from the solid substrate, and thereafter the label detected.

The following examples are offered by way of illustration and not by way of limitation.

# **EXAMPLES**

- 1. Protocol for hRIP Autophosphorylation Assay A. Reagents:
- Neutralite Avidin: 20  $\mu$ g/ml in PBS.
  - hRIP:  $10^{-8}$ – $10^{-5}$  M biotinylated hRIP kinase domain, residues 1–300 at 20  $\mu$ g/ml in PBS.

Blocking buffer: 5% BSA, 0.5% Tween 20 in PBS; 1 hour at room temperature.

Assay Buffer: 100 mM KCl, 20 mM HEPES pH 7.6, 0.25 mM EDTA, 1% glycerol, 0.5% NP-40, 50 mM BME, 1 mg/ml BSA, cocktail of protease inhibitors.

[ $^{32}$ P] $\gamma$ -ATP 10×stock: 2×10 $^{-5}$  M cold ATP with 100  $\mu$ Ci [ $^{32}$ P] $\gamma$ -ATP. Place in the 4° C. microfridge during screening.

Protease inhibitor cocktail (1000×): 10 mg Trypsin Inhibitor (BMB #109894), 10 mg Aprotinin (BMB #236624), 25 mg Benzamidine (Sigma #B-6506), 25 mg Leupeptin (BMB #1017128), 10 mg APMSF (BMB #917575), and 2 mM NaVo<sub>3</sub> (Sigma #S-6508) in 10 ml PBS.

B. Preparation of assay plates:

Coat with 120  $\mu$ l of stock Neutralite avidin per well overnight at 4° C.

Wash 2 times with 200  $\mu$ l PBS.

Block with 150  $\mu$ l of blocking buffer.

Wash 2 times with 200  $\mu$ l PBS.

C. Assay:

Add 40  $\mu$ l assay buffer/well.

Add 40  $\mu$ l biotinylated hRIP (0.1–10 pmoles/40 ul in assay buffer)

Add 10  $\mu$ l compound or extract.

Add 10  $\mu$ l [<sup>32</sup>P] $\gamma$ -ATP 10×stock.

Shake at 30° C. for 15 minutes.

Incubate additional 45 minutes at 30° C.

Stop the reaction by washing 4 times with 200  $\mu$ l PBS.

Add 150  $\mu$ l scintillation cocktail.

Count in Topcount.

D. Controls for all assays (located on each plate):

a. Non-specific binding (no RIP added)

b. cold ATP to achieve 80% inhibition.

2. Protocol for hRIP—Substrate Phosphorylation Assay

A. Reagents:

Neutralite Avidin: 20  $\mu$ g/ml in PBS.

hRIP:  $10^{-8}$ – $10^{-5}$  M hRIP at 20  $\mu$ g/ml in PBS.

Blocking buffer: 5% BSA, 0.5% Tween 20 in PBS; 1 hour at room temperature.

Assay Buffer: 100 mM KCl, 20 mM HEPES pH 7.6, 0.25 mM EDTA, 1% glycerol, 0.5% NP-40, 50 mM BME, 1 mg/ml BSA, cocktail of protease inhibitors.

[<sup>32</sup>P]γ-ATP 10×stock: 2×10<sup>-5</sup> M cold ATP with 100 μCi [<sup>32</sup>P]γ-ATP. Place in the 4° C. microfridge during screening.

Substrate:  $2 \times 10^{-6}$  M biotinylated synthetic peptide kinase substrate at 20  $\mu$ g/ml in PBS.

Protease inhibitor cocktail (1000×): 10 mg Trypsin Inhibitor (BMB # 109894), 10 mg Aprotinin (BMB #236624), 25 mg Benzamidine (Sigma #B-6506), 25 55 mg Leupeptin (BMB #1017128), 10 mg APMSF (BMB #917575), and 2 mM NaVo<sub>3</sub> (Sigma #S-6508) in 10 ml PBS.

B. Preparation of assay plates:

Coat with 120  $\mu$ l of stock Neutralite avidin per well <sup>60</sup> overnight at 4° C.

Wash 2 times with 200  $\mu$ l PBS.

Block with 150  $\mu$ l of blocking buffer.

Wash 2 times with 200  $\mu$ l PBS.

C. Assay:

Add 40  $\mu$ l assay buffer/well.

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Add 40  $\mu$ l hRIP (0.1–10 pmoles/40 ul in assay buffer)

Add 10  $\mu$ l compound or extract.

Shake at 30° C. for 15 minutes.

Add 10  $\mu$ l [<sup>32</sup>P] $\gamma$ -ATP 10×stock.

Add  $10 \mu l$  substrate.

Shake at 30° C. for 15 minutes.

Incubate additional 45 minutes at 30° C.

Stop the reaction by washing 4 times with 200  $\mu$ l PBS.

Add 150  $\mu$ l scintillation cocktail.

Count in Topcount.

D. Controls for all assays (located on each plate):

a. Non-specific binding (no RIP added)

b. cold ATP to achieve 80% inhibition.

3. Protocol for hRIP—TRADD Binding Assay

A. Reagents:

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Anti-myc antibody: 20  $\mu$ g/ml in PBS.

Blocking buffer: 5% BSA, 0.5% Tween 20 in PBS; 1 hour at room temperature.

Assay Buffer: 100 mM KCl, 20 mM HEPES pH 7.6, 0.25 mM EDTA, 1% glycerol, 0.5% NP-40, 50 mM β-mercaptoethanol, 1 mg/ml BSA, cocktail of protease inhibitors.

<sup>33</sup>P hRIP 10×stock: 10<sup>-8</sup>–10<sup>-6</sup> M "cold" hRIP (full length) supplemented with 200,000–250,000 cpm of labeled hRIP (HMK-tagged) (Beckman counter). Place in the 4° C. microfridge during screening.

Protease inhibitor cocktail (1000×): 10 mg Trypsin Inhibitor (BMB #109894), 10 mg Aprotinin (BMB #236624), 25 mg Benzamidine (Sigma #B-6506), 25 mg Leupeptin (BMB #1017128), 10 mg APMSF (BMB #917575), and 2 mM NaVo<sub>3</sub> (Sigma #S-6508) in 10 ml PBS.

TRADD:  $10^{-8}$ – $10^{-5}$  M myc eptitope-tagged TRADD in PBS.

B. Preparation of assay plates:

Coat with 120  $\mu$ l of stock anti-myc antibody per well overnight at 4° C.

Wash  $2\times$  with 200  $\mu$ l PBS.

Block with 150  $\mu$ l of blocking buffer.

Wash  $2\times$  with 200  $\mu$ l PBS.

C. Assay:

Add 40  $\mu$ l assay buffer/well.

Add 10  $\mu$ l compound or extract.

Add 10  $\mu$ l <sup>33</sup>P-RIP (20,000–25,000 cpm/0.1–10 pmoles/well= $10^{-9}$ – $10^{-7}$  M final concentration).

Shake at 25° C. for 15 minutes.

Incubate additional 45 minutes at 25° C.

Add 40 µl eptitope-tagged TRADD (0.1–10 pmoles/40 ul in assay buffer)

Incubate 1 hour at room temperature.

Stop the reaction by washing 4 times with 200  $\mu$ l PBS. Add 150  $\mu$ l scintillation cocktail.

Count in Topcount.

- D. Controls for all assays (located on each plate):
  - a. Non-specific binding (no hRIP added)
  - b. Soluble (non-tagged TRADD) to achieve 80% inhibition.
- 4. Protocol for hRIP—TRAF2 Binding Assay

A. Reagents:

Anti-myc antibody: 20  $\mu$ g/ml in PBS.

Blocking buffer: 5% BSA, 0.5% Tween 20 in PBS; 1 hour at room temperature.

Assay Buffer: 100 mM KCl, 20 mM HEPES pH 7.6, 0.25 mM EDTA, 1% glycerol, 0.5% NP-40, 50 mM β-mercaptoethanol, 1 mg/ml BSA, cocktail of protease inhibitors.

 $^{33}$ P hRIP 10×stock:  $10^{-8}$ – $10^{-6}$  M "cold" hRIP kinase  $^{5}$ domain, residues 1-300, supplemented with 200, 000-250,000 cpm of labeled hRIP kinase domain (HMK-tagged) (Beckman counter). Place in the 4° C. microfridge during screening.

Protease inhibitor cocktail (1000×): 10 mg Trypsin Inhibitor (BMB #109894), 10 mg Aprotinin (BMB #236624), 25 mg Benzamidine (Sigma #B-6506), 25 mg Leupeptin (BMB #1017128), 10 mg APMSF (BMB #917575), and 2 mM NaVo<sub>3</sub> (Sigma #S-6508) in 10 ml PBS.

TRAF2:  $10^{-8}$ – $10^{-5}$  M myc eptitope-tagged TRAF2 in PBS.

B. Preparation of assay plates:

Coat with 120  $\mu$ l of stock anti-myc antibody per well overnight at 4° C.

Wash  $2\times$  with 200  $\mu$ l PBS.

Block with 150  $\mu$ l of blocking buffer.

Wash  $2 \times$  with 200  $\mu$ l PBS.

C. Assay:

Add 40  $\mu$ l assay buffer/well.

Add 10  $\mu$ l compound or extract.

Add 10  $\mu$ l <sup>33</sup>P-RIP kinase domain (20,000–25,000 cpm/  $0.1-10 \text{ pmoles/well}=10^{-9}-10^{-7} \text{ M}$  final concentration).

Shake at 25° C. for 15 minutes.

Incubate additional 45 minutes at 25° C.

Add 40 µl eptitope-tagged TRAF2 (0.1–10 pmoles/40 ul in assay buffer)

Incubate 1 hour at room temperature.

Stop the reaction by washing 4 times with 200  $\mu$ l PBS.

Add 150  $\mu$ l scintillation cocktail.

Count in Topcount.

- D. Controls for all assays (located on each plate):
  - a. Non-specific binding (no hRIP kinase domain added)
  - b. Soluble (non-tagged TRAF2) to achieve 80% inhibition.

All publications and patent applications cited in this specification are herein incorporated by reference as if each individual publication or patent application were specifically and individually indicated to be incorporated by reference. Although the foregoing invention has been described in some detail by way of illustration and example for purposes of clarity of understanding, it will be readily apparent to those of ordinary skill in the art in light of the teachings of 25 this invention that certain changes and modifications may be made thereto without departing from the spirit or scope of the appended claims.

SEQUENCE LISTING

GENERAL INFORMATION:

(iii) NUMBER OF SEQUENCES: 2

- (2) INFORMATION FOR SEQ ID NO:1:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 2016 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: double (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: cDNA

  - (ix) FEATURE:
    - (A) NAME/KEY: CDS
    - (B) LOCATION: 1..2013

(xi	) SE	QUENC	CE DI	ESCRI	[PTI(	ON: S	SEQ I	ID NO	0:1:			
CAA Gln												48
 CTG Leu										 	 	 96
 TGT Cys												144
GGG Gly 50											 	 192
 ATG Met										 	 	 240

# -continued

											_	con	tin	ıed		
						AAG Lys										288
				85					90					95		
						GTG Val										336
						ATT Ile										384
						ATA Ile 135										432
	_				_	CAC His	_		_	_			_		_	480
						AAA Lys						_				528
						GCT Ala										576
				_		AAT Asn										624
						GCT Ala 215										672
	_			_	Asn	GCT Ala	Ile	Cys	Glu	Gln	Gln	Leu	Ile	Met		720
						CCA Pro										768
		_				CTC Leu										816
						CCT Pro										864
			_		_	GAA Glu 295		_	_	_		_				912
		_				GAA Glu		_	_	_				_		960
	_				_	GCA Ala	_								_	1008
						CTG Leu	_									1056
	_	_	_			TTT Phe	_				_	_		_	_	1104
_		_				CAG Gln 375				_		_	_			1152
_			_			ATG Met			_	_		_	_			1200

# -continued

_		GTG Val					_	_	_				_		1248
		CCT Pro													1296
		AAA Lys 435								 					1344
		CAG Gln													1392
		GGA Gly					_								1440
		ACA Thr													1488
		AGT Ser		_											1536
	_	CCC Pro 515	_			_					_		_		1584
		TAT Tyr													1632
		ATG Met													1680
	_	AAC Asn	_		_	_					_	_	_	_	1728
		ACC Thr													1776
		GGA Gly 595								 					1824
		CAG Gln													1872
		AAG Lys													1920
		GGA Gly										_			1968
		ATC Ile													2013
TAA															2016

# (2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 671 amino acids
  - (B) TYPE: amino acid
  - (D) TOPOLOGY: linear

# -continued

	(ii)	MOI	LECUI	E TI	PE:	prot	ein								
	(xi)	SEÇ	QUENC	CE DE	ESCRI	PTIC	N: S	SEQ I	D NC	2:					
Met 1	Gln	Pro	Asp	Met 5	Ser	Leu	Asn	Val	Ile 10	Lys	Met	Lys	Ser	Ser 15	Asp
Phe	Leu	Glu	Ser 20	Ala	Glu	Leu	Asp	Ser 25	Gly	Gly	Phe	Gly	L <b>y</b> s 30	Val	Ser
Leu	Сув	Phe 35	His	Arg	Thr	Gln	Gl <b>y</b> 40	Leu	Met	Ile	Met	L <b>y</b> s 45	Thr	Val	Tyr
Lys	Gl <b>y</b> 50	Pro	Asn	Cys	Ile	Glu 55	His	Asn	Glu	Ala	Leu 60	Leu	Glu	Glu	Ala
L <b>y</b> s 65	Met	Met	Asn	Arg	Leu 70	Arg	His	Ser	Arg	Val 75	Val	Lys	Leu	Leu	Gl <b>y</b> 80
Val	Ile	Ile	Glu	Glu 85	_	Lys	Tyr	Ser	Leu 90	Val	Met	Glu	Tyr	Met 95	Glu
Lys	Gly	Asn	Leu 100	Met	His	Val	Leu	L <b>y</b> s 105	Ala	Glu	Met	Ser	Thr 110	Pro	Leu
Ser		_	_	Arg								Gl <b>y</b> 125		Сув	Tyr
Leu	His 130	Gly	Lys	Gly	Val	Ile 135	His	Lys	Asp	Leu	L <b>y</b> s 140	Pro	Glu	Asn	Ile
Leu 145	Val	Asp	Asn	Asp	Phe 150	His	Ile	Lys	Ile	Ala 155	Asp	Leu	Gly	Leu	Ala 160
Ser	Phe	Lys	Met	Trp 165		Lys	Leu	Asn	Asn 170	Glu	Glu	His	Asn	Glu 175	Leu
Arg	Glu	Val	Asp 180	Gly	Thr	Ala	Lys	L <b>y</b> s 185	Asn	Gly	Gly	Thr	Leu 190	Tyr	Tyr
Met	Ala	Pro 195	Glu	His	Leu	Asn	<b>Asp</b> 200	Val	Asn	Ala	Lys	Pro 205	Thr	Glu	Lys
Ser	Asp 210	Val	Tyr	Ser	Phe	Ala 215	Val	Val	Leu	Trp	Ala 220	Ile	Phe	Ala	Asn
L <b>y</b> s 225	Glu	Pro	Tyr	Glu	Asn 230	Ala	Ile	Сув	Glu	Gln 235	Gln	Leu	Ile	Met	C <b>y</b> s 240
Ile	Lys	Ser	Gly	Asn 245	Arg	Pro	Asp		<b>Asp</b> 250	Asp	Ile	Thr	Glu	<b>Ty</b> r 255	Cys
Pro	Arg	Glu		Ile				_		_	_		Ala 270	Asn	Pro
Glu	Ala	<b>A</b> rg 275	Pro	Thr	Phe	Pro	Gl <b>y</b> 280	Ile	Glu	Glu	Lys	Phe 285	Arg	Pro	Phe
Tyr	Leu 290	Ser	Gln	Leu	Glu	Glu 295	Ser	Val	Glu	Glu	Asp 300	Val	Lys	Ser	Leu
L <b>y</b> s 305	Lys	Glu	Tyr	Ser	Asn 310	Glu	Asn	Ala	Val	Val 315	Lys	Arg	Met	Gln	Ser 320
Leu	Gln	Leu	Asp	С <b>у</b> в 325	Val	Ala	Val	Pro	Ser 330	Ser	Arg	Ser	Asn	Ser 335	Ala
Thr	Glu	Gln	Pro 340	Gly	Ser	Leu	His	Ser 345	Ser	Gln	Gly	Leu	Gl <b>y</b> 350	Met	Gly
Pro	Val	Glu 355	Glu	Ser	Trp	Phe	Ala 360	Pro	Ser	Leu	Glu	His 365	Pro	Gln	Glu
Glu	Asn 370	Glu	Pro	Ser	Leu	Gln 375	Ser	Lys	Leu	Gln	<b>Asp</b> 380	Glu	Ala	Asn	Tyr
His 385	Leu	Tyr	Gly	Ser	Arg 390	Met	Asp	Arg	Gln	Thr 395	Lys	Gln	Gln	Pro	Arg 400

#### -continued

Gln	Asn	Val	Ala	<b>Ty</b> r 405	Asn	Arg	Glu	Glu	Glu 410	Arg	Arg	Arg	Arg	Val 415	Ser
His	Asp	Pro	Phe 420	Ala	Gln	Gln	Arg	Pro 425	Tyr	Glu	Asn	Phe	Gln 430	Asn	Thr
Glu	Gly	L <b>y</b> s 435	Gly	Thr	Val	Tyr	Ser 440	Ser	Ala	Ala	Ser	His 445	Gly	Asn	Ala
Val	His 450	Gln	Pro	Ser	Gly	Leu 455	Thr	Ser	Gln	Pro	Gln 460	Val	Leu	Tyr	Gln
Asn 465	Asn	Gly	Leu	Tyr	Ser 470		His	Gly	Phe	Gl <b>y</b> 475	Thr	Arg	Pro	Leu	Asp 480
Pro	Gly	Thr	Ala	Gl <b>y</b> 485	Pro	Arg	Val	Trp	<b>Ty</b> r 490	Arg	Pro	Ile	Pro	Ser 495	His
Met	Pro	Ser	Leu 500	His	Asn	Ile	Pro	Val 505	Pro	Glu	Thr	Asn	<b>Ty</b> r 510	Leu	Gly
Asn	Thr	Pro 515	Thr	Met	Pro	Phe	Ser 520	Ser	Leu	Pro	Pro	Thr 525	Asp	Glu	Ser
Ile	<b>Lys</b> 530	Tyr	Thr	Ile	Tyr	Asn 535	Ser	Thr	Gly	Ile	Gln 540	Ile	Gly	Ala	Tyr
Asn 545	Tyr	Met	Glu	Ile	Gl <b>y</b> 550	_	Thr	Ser	Ser	Ser 555	Leu	Leu	Asp	Ser	Thr 560
Asn	Thr	Asn	Phe	L <b>y</b> s 565	Glu	Glu	Pro	Ala	Ala 570	Lys	Tyr	Gln	Ala	Ile 575	Phe
Asp	Asn	Thr	Thr 580	Ser	Leu	Thr	Asp	<b>Ly</b> s 585	His	Leu	Asp	Pro	Ile 590	Arg	Glu
Asn	Leu	Gl <b>y</b> 595	Lys	His	Trp	Lys	Asn 600	Cys	Ala	Arg	Lys	Leu 605	Gly	Phe	Thr
Gln	Ser 610	Gln	Ile	Asp	Glu	Ile 615	Asp	His	Asp	Tyr	Glu 620	Arg	Asp	Gly	Leu
L <b>y</b> s 625	Glu	Lys	Val	Tyr	Gln 630	Met	Leu	Gln	Lys	Trp 635	Val	Met	Arg	Glu	Gl <b>y</b> 640
Ile	Lys	Gly	Ala	Thr 645	Val	Gly	Lys	Leu	Ala 650	Gln	Ala	Leu	His	Gln 655	Суѕ
Ser	Arg	Ile	<b>A</b> sp 660	Leu	Leu	Ser	Ser	Leu 665	Ile	Tyr	Val	Ser	Gln 670	Asn	

What is claimed is:

1. An isolated RIP-Thr<sup>514</sup> polypeptide, comprising at least 10 consecutive amino acid residues of the amino acid sequence set forth as SEQ ID NO:2, which consecutive amino acid residues comprise the amino acid residue 514 50 (Thr) of SEQ ID NO:2.

- 2. An isolated polypeptide according to claim 1, wherein said polypeptide has an activity selected from at least one of: a kinase or kinase inhibitory activity or a RIP-binding or binding inhibitory activity.
- 3. A method of screening for an agent which modulates the interaction of a RIP polypeptide to a binding target, said method comprising the steps of:

incubating a mixture comprising:

- an isolated polypeptide according to claim 1, a binding target of said polypeptide, and
- a candidate agent;
- under conditions whereby, but for the presence of said agent, said polypeptide specifically binds said binding target at a reference affinity;
- detecting the binding affinity of said polypeptide to said binding target to determine an agent-biased affinity,

- wherein a difference between the agent-biased affinity and the reference affinity indicates that said agent modulates the binding of said polypeptide to said binding target.
- 4. A method according to claim 3, wherein said binding target is a natural intracellular substrate of the polypeptide and said agent-biased binding affinity is detected as phosphorylation of said substrate.
- 5. A method according to claim 3, wherein said binding target comprises a Tumor necrosis factor receptor Associated Factor-2 (TRAF2) or a Tumor necrosis factor Receptor-1 Associated Death Domain protein (TRADD).
  - 6. An isolated polypeptide according to claim 1, wherein said consecutive amino acid residues comprise  $\alpha\Delta 1$  (SEQ ID NO:2, residues 509–518).
  - 7. An isolated polypeptide according to claim 1, wherein said consecutive amino acid residues comprise  $\alpha\Delta 2$  (SEQ ID NO:2, residues 514–521).
  - 8. An isolated polypeptide according to claim 1, wherein said consecutive amino acid residues comprise  $\alpha\Delta 3$  (SEQ ID NO:2, residues 506–514).
  - 9. An isolated polypeptide according to claim 1, wherein said consecutive amino acid residues comprise  $\alpha\Delta4$  (SEQ ID NO:2, residues 504–524).

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- 10. An isolated polypeptide according to claim 1, wherein said consecutive amino acid residues comprise  $\alpha\Delta 5$  (SEQ ID NO:2, residues 498–514).
- 11. An isolated polypeptide according to claim 1, wherein said consecutive amino acid residues comprise  $\alpha\Delta6$  (SEQ ID NO:2, residues 514–534).
- 12. An isolated polypeptide according to claim 1, wherein said consecutive amino acid residues comprise  $\alpha\Delta 7$  (SEQ ID NO:2, residues 513–520).
- 13. An isolated polypeptide according to claim 1, wherein 10 said consecutive amino acid residues comprise  $\alpha\Delta 8$  (SEQ ID NO:2, residues 508–515).
- 14. An isolated polypeptide according to claim 1, wherein said consecutive amino acid residues comprise  $\alpha\Delta9$  (SEQ ID NO:2, residues 512–522).
- 15. An isolated polypeptide according to claim 1, wherein said consecutive amino acid residues comprise  $\alpha\Delta 10$  (SEQ ID NO:2, residues 423–514).
- 16. An isolated polypeptide according to claim 1, wherein said consecutive amino acid residues comprise  $\alpha\Delta 11$  (SEQ 20 ID NO:2, residues 423–543).
- 17. An isolated polypeptide according to claim 1, wherein said consecutive amino acid residues comprise  $\alpha\Delta 12$  (SEQ ID NO:2, residues 423–579).

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- 18. An isolated polypeptide according to claim 1, wherein said consecutive amino acid residues comprise  $\alpha\Delta 13$  (SEQ ID NO:2, residues 423–633).
- 19. An isolated polypeptide according to claim 1, wherein said consecutive amino acid residues comprise  $\alpha\Delta 14$  (SEQ ID NO:2, residues 423–671).
- 20. An isolated polypeptide according to claim 1, wherein said consecutive amino acid residues comprise  $\alpha\Delta 15$  (SEQ ID NO:2, residues 514–543).
- 21. An isolated polypeptide according to claim 1, wherein said consecutive amino acid residues comprise  $\alpha\Delta 16$  (SEQ ID NO:2, residues 514–579).
- 22. An isolated polypeptide according to claim 1, wherein said consecutive amino acid residues comprise  $\alpha\Delta 17$  (SEQ ID NO:2, residues 514–633).
- 23. An isolated polypeptide according to claim 1, wherein said consecutive amino acid residues comprise  $\alpha\Delta 18$  (SEQ ID NO:2, residues 514–671).
- 24. An isolated polypeptide according to claim 1, wherein said consecutive amino acid residues comprise SEQ ID NO:2.

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