



US005859208A

United States Patent [19]

Fiddes et al.

[11] Patent Number: **5,859,208**

[45] Date of Patent: **Jan. 12, 1999**

[54] **HUMAN BASIC FIBROBLAST GROWTH FACTOR ANALOG**

[76] Inventors: **John C. Fiddes**, 2320 Bryant St., Palo Alto, Calif. 94301; **Judith A. Abraham**, 655 S. Fair Oaks Ave., Sunnyvale, Calif. 94086; **Andrew A. Protter**, 185 N. California Ave., Palo Alto, Calif. 94301

[21] Appl. No.: **459,739**

[22] PCT Filed: **Jul. 6, 1988**

[86] PCT No.: **PCT/US88/02264**

§ 371 Date: **Feb. 12, 1990**

§ 102(e) Date: **Feb. 12, 1990**

[87] PCT Pub. No.: **WO89/00198**

PCT Pub. Date: **Jan. 12, 1989**

[51] **Int. Cl.⁶** **C07K 14/50**

[52] **U.S. Cl.** **530/399; 530/350**

[58] **Field of Search** **530/399, 350, 530/395**

[56] **References Cited**

U.S. PATENT DOCUMENTS

4,959,314 9/1990 Mark et al. 435/69.1

FOREIGN PATENT DOCUMENTS

0237966 9/1987 European Pat. Off. .
WO 86/07595 12/1986 WIPO .

OTHER PUBLICATIONS

Abraham et al., "Human basic fibroblast growth factor: nucleotide sequence and genomic organization" *EMBO J.* (1986) 5:2523-2528.

Fiddes et al., "Isolation and characterization of clones encoding basic and acidic fibroblast growth factors" *J. Biol. Chem.* (1986) 0:149 (No. 10, Part C, abstract No. L146).

Seno et al. *Biochem Biophys Res Comm* 151:701-708 (1988).

Baird et al. *Rec Prog Horn Res* 42:143-705 (1986).

Primary Examiner—Stephen Walsh

Attorney, Agent, or Firm—Susan K. Lehnhardt

[57] **ABSTRACT**

A human basic fibroblast growth factor (FGF) protein analog wherein the cysteine at positions 78 and 96 is replaced by serine, and said analog exhibits the biological activity of native, human basic FGF.

The DNA sequences encoding analogs of human acidic and basic fibroblast growth factors (FGF) can be recombinantly expressed to obtain practical amounts of proteins useful in effecting both pathologies related to persistent angiogenesis and wound healing and related tissue repair.

1 Claim, 8 Drawing Sheets

FIG. 1-1

Human Basic Fibroblast Growth Factor

10	20	30	40	50	60	70	
AATTCATGCC	TCTTTCTCTC	CTTTTGTTGG	TAGACGACTT	CAGCCTCTGT	CCTTTAATTT	TAAAGTTTAT	
80	90	100	110	120	130	140	
GCCCCACTTG	TACCCCTCGT	CTTTTGGTGA	TTTAGAGATT	TTCAAAGCCT	GCTCTGACAC	AGACTCTTCC	
150	160	170	180	190	200	210	
TTGGATTGCA	ACTTCTCTAC	TTTGGGGTGG	AAACGGCTTC	TCCGTTTTGA	AACGCTAGCG	GGGAAAAAAT	
220	230	240	250	260	270	280	
GGGGGAGAAA	GTTGAGTTTA	AACTTTTAAA	AGTTGAGTCA	CGGCTGGTTG	CGCACGAAAA	GCCCCGCAGT	
290	300	310	320	330	340	350	
GTGGAGAAAG	CCTAAACGTG	GTTTGGGTGG	TGCGGGGGTT	GGGCGGGGGT	GACTTTTGGG	GGATAAGGGG	
360	370	380	390	400	410	420	
CGGTGGAGCC	CAGGGAATGC	CAAAGCCCTG	CCGCGGCCTC	CGACGCGCGC	CCCCCGCCCC	TCGCCTCTCC	
430	440	450	460	470	480	490	
CCCGCCCCCG	ACTGAGGCCG	GGCTCCCCGC	CGGACTGATG	TCGCGCGCTT	GCGTGTTGTG	GCCGAAGCCG	
500	510	520	530	540	550	560	
CCGAACTCAG	AGGCCGGCCC	CAGAAAACCC	GAGCGAGTAG	GGGGCGGCGC	GCAGGAGGGA	GGAGAACTGG	
570	580	590	600	610	620	630	
GGGCGCGGGA	GGCTGGTGGG	TGTGGGGGGT	GGAGATGTAG	AAGATGTGAC	GCCGCGGCCC	GGCGGGTGCC	
640	650	660	670	680	690	700	
AGATTAGCGG	ACGGCTGCCC	GCGGTTGCAA	CGGGATCCCC	GGCGCTGCAG	CTTGGGAGGC	GGCTCTCCCC	
710	720	730	740	750	760	770	
AGGCGGCGTC	CGCGGAGACA	CCCATCTGTG	AACCCAGGT	CCCGGGCCGC	CGGCTCGCCG	CGCACCAGGG	
780	790	800	810	820	830	840	
GCCGGCGGAC	AGAAGAGCGG	CCGAGCGGCT	CGAGGCTGGG	GGACCGCGGG	CGCGGCCCGG	CGCTGCCGGG	
850	860	870	880	890	900	910	
CGGGAGGCTG	GGGGGCCGGG	GCCGGGGCCG	TGCCCCGAGC	GGGTCCGAGG	CCGGGGCCCG	GGCCGGGGGA	
920	930	940	950	960			
CGGCGGCTCC	CCGCGCGGCT	CCAGCGGCTC	GGGGATCCCC	GCCGGGCCCC	GCAGGGACC	ATG GCA GCC	
						Met Ala Ala	
984	999		1014			1029	
GGG AGC ATC ACC ACG CTG CCC GCC TTG CCC GAG GAT GGC GGC AGC GGC GCC TTC CCG							
Gly Ser Ile Thr Thr Leu Pro Ala Leu Pro Glu Asp Gly Gly Ser Gly Ala Phe Pro							
1044	1059		1074			1089	
CCC GGC CAC TTC AAG GAC CCC AAG CGG CTG TAC TGC AAA AAC GGG GGC TTC TTC CTG							
Pro Gly His Phe Lys Asp Pro Lys Arg Leu Tyr Cys Lys Asn Gly Gly Phe Phe Leu							

FIG. 1-2

1104 1119 1134 1149
 CGC ATC CAC CCC GAC GGC CGA GTT GAC GGG GTC CGG GAG AAG AGC GAC CCT CAC ATC
 Arg Ile His Pro Asp Gly Arg Val Asp Gly Val Arg Glu Lys Ser Asp Pro His Ile

1164 1179 1194
 AAG CTA CAA CTT CAA GCA GAA GAG AGA GGA GTT GTG TCT ATC AAA GGA GTG TGT GCT
 Lys Leu Gln Leu Gln Ala Glu Glu Arg Gly Val Val Ser Ile Lys Gly Val Cys Ala

1224 1239 1254
 AAC CGT TAC CTG GCT ATG AAG GAA GAT GGA AGA TTA CTG GCT TCT AAA TGT GTT ACG
 Asn Arg Tyr Leu Ala Met Lys Glu Asp Gly Arg Leu Leu Ala Ser Lys Cys Val Thr

1269 1284 1299 1314
 GAT GAG TGT TTC TTT TTT GAA CGA TTG GAA TCT AAT AAC TAC AAT ACT TAC CGG TCA
 Asp Glu Cys Phe Phe Phe Glu Arg Leu Glu Ser Asn Asn Tyr Asn Thr Tyr Arg Ser

1329 1344 1359 1374
 AGG AAA TAC ACC AGT TGG TAT GTG GCA TTG AAA CGA ACT GGG CAG TAT AAA CTT GGA
 Arg Lys Tyr Thr Ser Trp Tyr Val Ala Leu Lys Arg Thr Gly Gln Tyr Lys Leu Gly

1389 1404 1419 1434
 TCC AAA ACA GGA CCT GGG CAG AAA GCT ATA CTT TTT CTT CCA ATG TCT GCT AAG AGC
 Ser Lys Thr Gly Pro Gly Gln Lys Ala Ile Leu Phe Leu Pro Met Ser Ala Lys Ser

1450 1460 1470 1480 1490 1500
 TGA TTT TAATGGCCAC ATCTAATCTC ATTTACATG AAAGAAGAAG TATATTTTAG AAATTTGTTA

1510 1520 1530 1540 1550 1560 1570
 ATGAGAGTAA AAGAAAATAA ATGTGTATAG CTCAGTTTGG ATAATTGGTC AAACAATTTT TTATCCAGTA

1580 1590 1600 1610 1620 1630 1640
 GTAAAATATG TAACCATGCC CAGTAAAGAA AAATAACAAA AGTTGTAAAA TGTATATTCT CCCTTTTATA

1650 1660 1670 1680 1690 1700 1710
 TTGCATCTGC TGTTACCCAG TGAAGCTTAC CTAGAGCAAT GATCTTTTTC ACGCATTTC TTTATTCGAA

1720 1730 1740 1750 1760 1770 1780
 AAGAGGCTTT TAAAATGTGC ATGTTTAGAA AACAAAATTT CTTCATGGAA ATCATATACA TTAGAAAATC

1790 1800 1810 1820 1830 1840 1850
 ACAGTCAGAT GTTTAATCAA TCCAAAATG TCCACTATTT CTTATGTCAT TCGTTAGTCT ACATGTTTCT

1860 1870 1880 1890 1900 1910 1920
 AACATATAA ATGTGAATTT AATCAATTCC TTTCATAGTT TTATAATTCT CTGGCAGTTC CTTATGATAG

1930 1940 1950 1960 1969
 AGTTTATAAA ACAGTCCTGT GTAAACTGCT GGAAGTTCTT CCGGAATTC

FIG. 2

Human Acidic FGF

TGC	ATT	TTG	TGC	CTT	TGC	TGG	AAG	AAC	CGA	CTA	CAG	GTT	TGT	TCA	ATT	TCT	TAC	27	54
AGT	CTT	GAA	AGC	GCC	ACA	AGC	AGC	AGC	TGC	TGA	GCC	ATG	GCT	GAA	GGG	GAA	ATC	81	108
												MET	Ala	Glu	Gly	Glu	Ile		
																		135	162
ACC	ACC	TTC	ACA	GCC	CTG	ACC	GAG	AAG	TTT	AAT	CTG	CCT	CCA	GGG	AAT	TAC	AAG		
Thr	Thr	Phe	Thr	Ala	Leu	Thr	Glu	Lys	Phe	Asn	Leu	Pro	Pro	Gly	Asn	Tyr	Lys	10	20
																		189	216
AAG	CCC	AAA	CTC	CTC	TAC	TGT	AGC	AAC	GGG	GGC	CAC	TTC	CTG	AGG	ATC	CTT	CCG		
Lys	Pro	Lys	Leu	Leu	Tyr	Cys	Ser	Asn	Gly	Gly	His	Phe	Leu	Arg	Ile	Leu	Pro	30	40
																		243	270
GAT	GGC	ACA	GTG	GAT	GGG	ACA	AGG	GAC	AGG	AGC	GAC	CAG	CAC	ATT	CAG	CTG	CAG		
Asp	Gly	Thr	Val	Asp	Gly	Thr	Arg	Asp	Arg	Ser	Asp	Gln	His	Ile	Gln	Leu	Gln	50	60
																		297	324
CTC	AGT	GCG	GAA	AGC	GTG	GGG	GAG	GTG	TAT	ATA	AAG	AGT	ACC	GAG	ACT	GGC	CAG		
Leu	Ser	Ala	Glu	Ser	Val	Gly	Glu	Val	Tyr	Ile	Lys	Ser	Thr	Glu	Thr	Gly	Gln	70	
																		351	378
TAC	TTG	GCC	ATG	GAC	ACC	GAC	GGG	CTT	TTA	TAC	GGC	TCA	CAG	ACA	CCA	AAT	GAG		
Tyr	Leu	Ala	MET	Asp	Thr	Asp	Gly	Leu	Leu	Tyr	Gly	Ser	Gln	Thr	Pro	Asn	Glu	80	90
																		405	432
GAA	TGT	TTG	TTC	CTG	GAA	AGG	CTG	GAG	GAG	AAC	CAT	TAC	AAC	ACC	TAT	ATA	TCC		
Glu	Cys	Leu	Phe	Leu	Glu	Arg	Leu	Glu	Glu	Asn	His	Tyr	Asn	Thr	Tyr	Ile	Ser	100	110
																		459	486
AAG	AAG	CAT	GCA	GAG	AAG	AAT	TGG	TTT	GTT	GGC	CTC	AAG	AAG	AAT	GGG	AGC	TGC		
Lys	Lys	His	Ala	Glu	Lys	Asn	Trp	Phe	Val	Gly	Leu	Lys	Lys	Asn	Gly	Ser	Cys	120	130
																		513	540
AAA	CGC	GGT	CCT	CGG	ACT	CAC	TAT	GGC	CAG	AAA	GCA	ATC	TTG	TTT	CTC	CCC	CTG		
Lys	Arg	Gly	Pro	Arg	Thr	His	Tyr	Gly	Gln	Lys	Ala	Ile	Leu	Phe	Leu	Pro	Leu	140	150
																		567	594
CCA	GTC	TCT	TCT	GAT	TAA	AGA	GAT	CTG	TTC	TGG	GTG	TTG	ACC	ACT	CCA	GAG	AAG		
Pro	Val	Ser	Ser	Asp														155	
																		621	
TTT	CGA	GGG	GTC	CTC	ACC	TGG	TTG	ACC	CAA	AAA	TGT	TCC	CTT	GA					

Comparison of amino acid sequence of
human basic and acidic FGF

(basic/acidic)

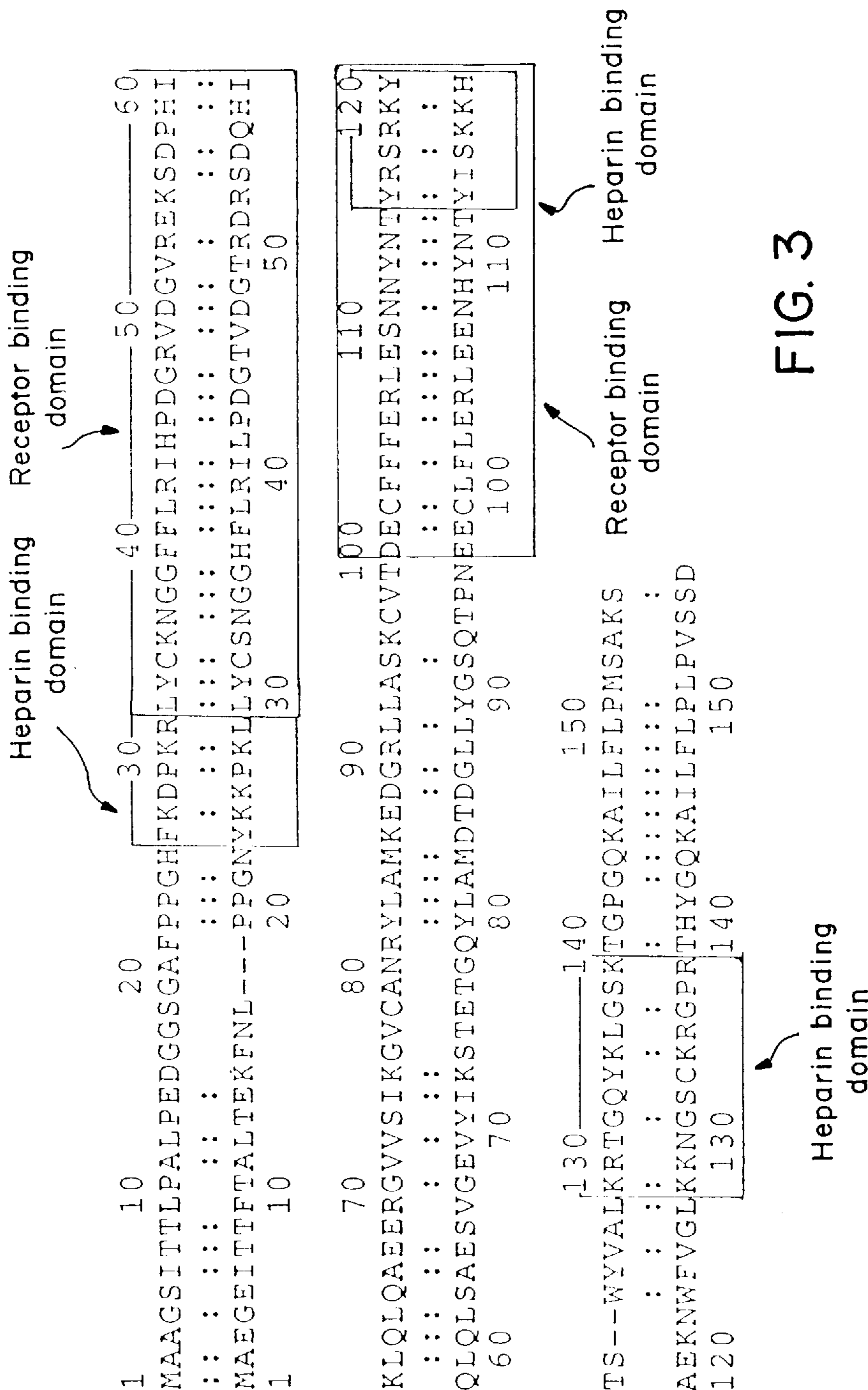


FIG. 3

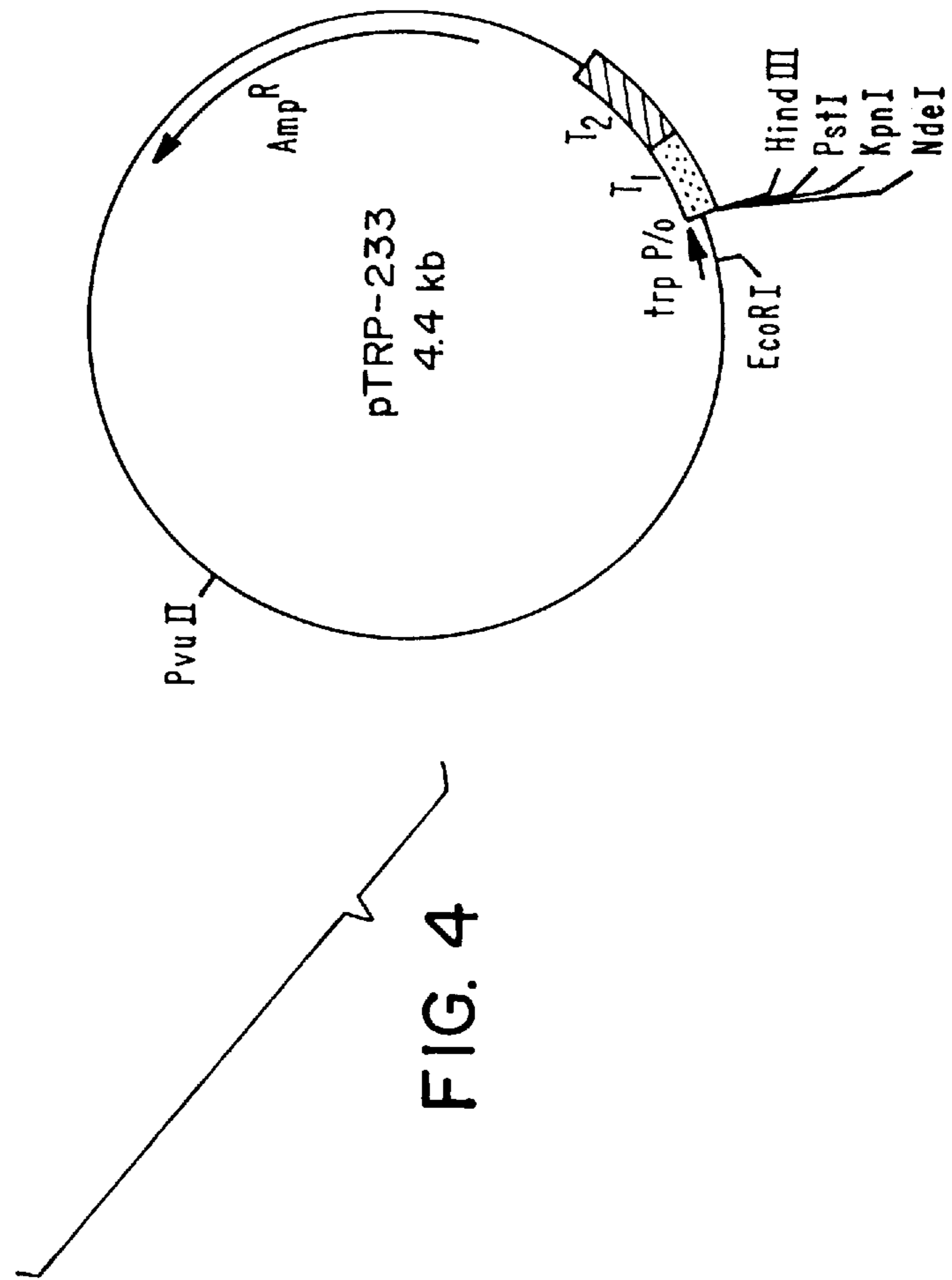
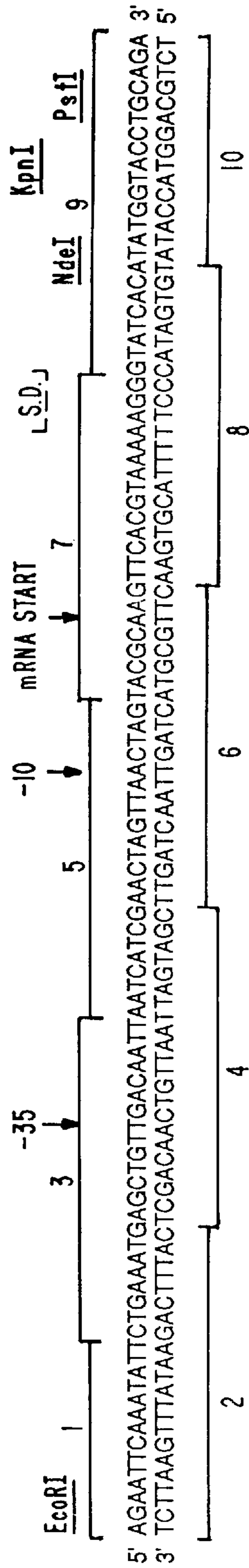


FIG. 4

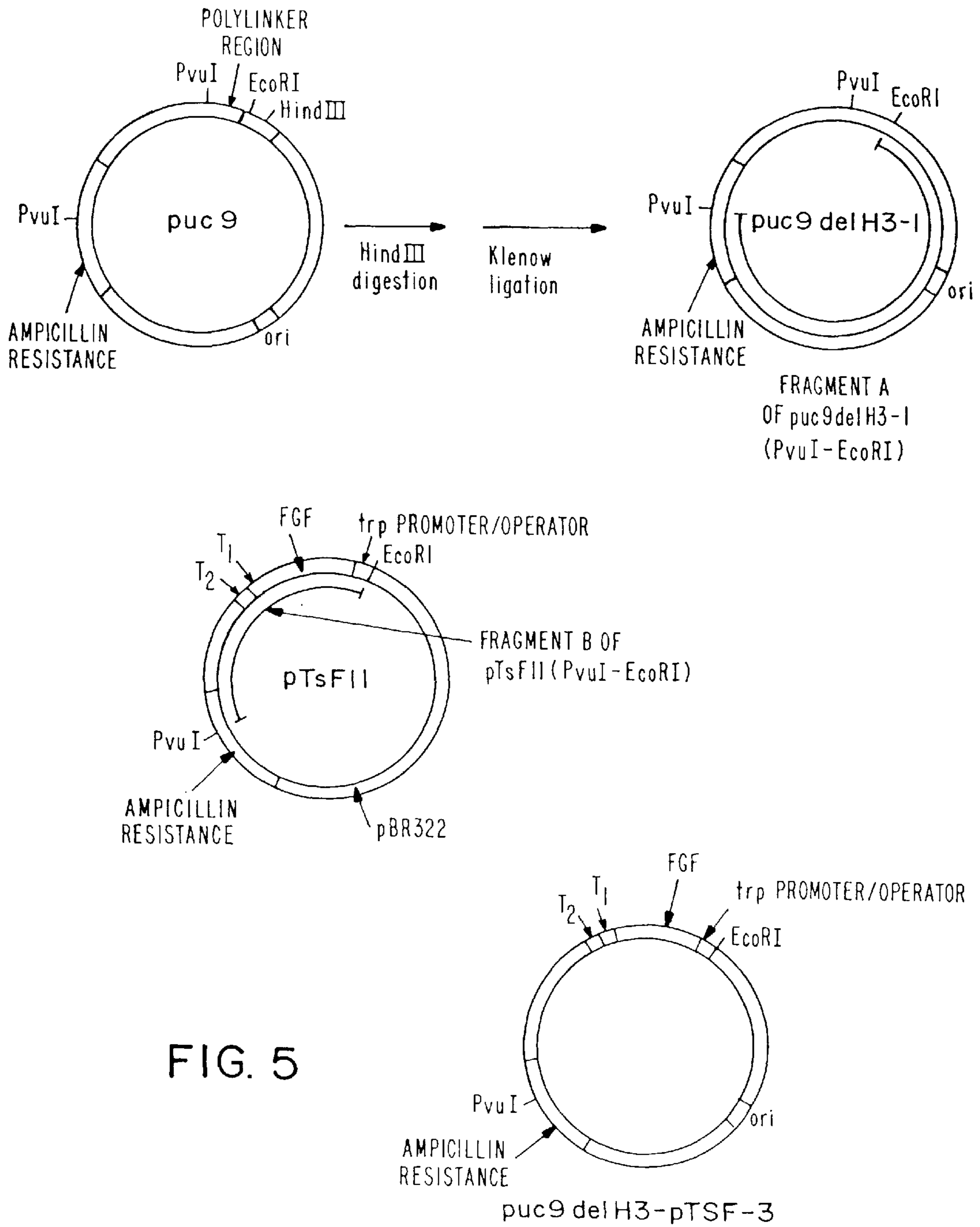


FIG. 5

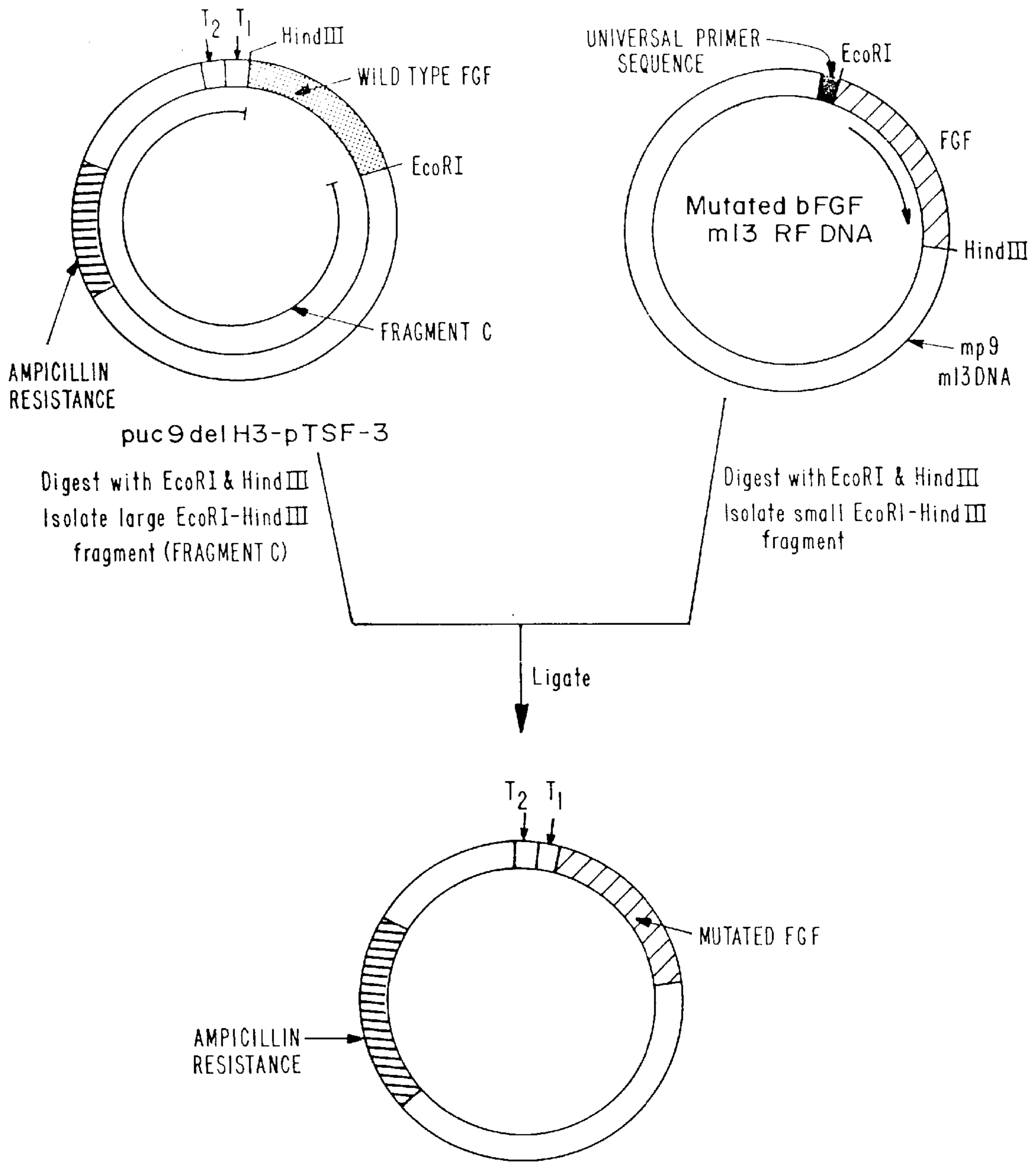


FIG. 6

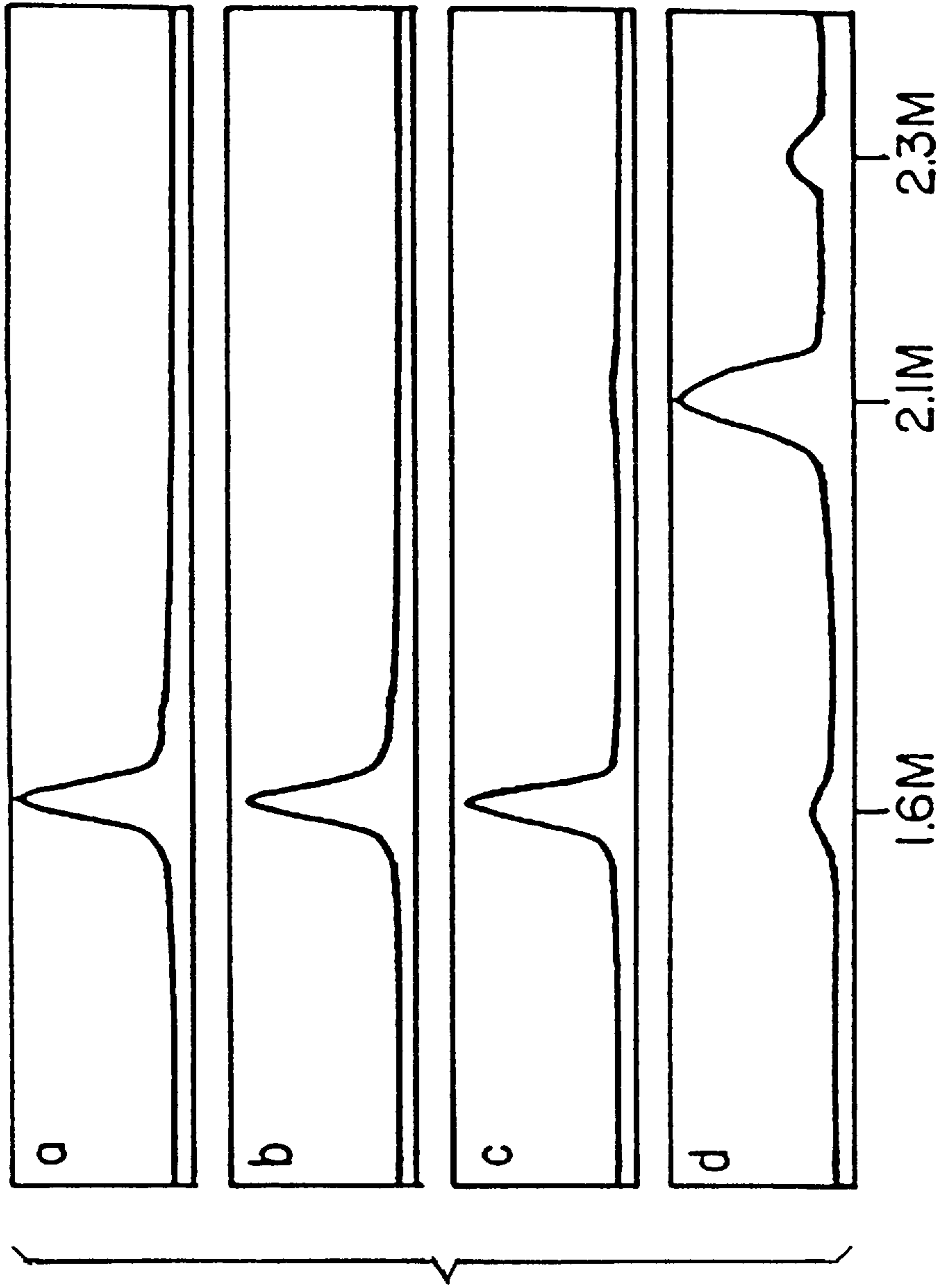


FIG. 7

HUMAN BASIC FIBROBLAST GROWTH FACTOR ANALOG

TECHNICAL FIELD

The invention relates to recombinant production of growth factors important for constructing vascular systems in healing tissues and inhibiting abnormal persistent angiogenesis. In particular, analogs of genes encoding human basic and acidic fibroblast growth factors (FGF) are cloned and expressed.

BACKGROUND ART

The process of healing when tissue is subjected to trauma, such as wounding or burns, is an extremely complex one, but it is known to be mediated by a number of protein factors. These factors are essential to the growth and differentiation of the cells which serve to replace the tissue destroyed. A number of candidate factors have been identified on the basis of the ability of extracts from various tissues, such as brain, pituitary, and hypothalamus, to stimulate the mitosis of cultured cells. Numerous shorthand names have been applied to the active factors in these extracts, including platelet-derived growth factor (PDGF), macrophage-derived growth factor (MDGF), epidermal growth factor (EGF), tumor angiogenesis factor (TAF), endothelial cell growth factor (ECGF), fibroblast growth factor (FGF), hypothalamus-derived growth factor (HDGF), retina-derived growth factor (RDGF), and heparin-binding growth factor (HGF). (See, for example, Hunt, T. K., *J Trauma* (1984) 24:S39-S49; Lobb, R. R., et al, *Biochemistry* (1984) 23:6295-6299).

Folkman, J., et al, *Science* (1983) 221:719-725, reported that one of the processes involved in wound healing, the formation of blood vessels, is profoundly affected in tumors by heparin. From this and other studies, it is clear that heparin specifically binds to protein(s) associated with a number of these growth factor activities, and therefore heparin has been used as a purification tool. It has been shown that the affinity of some growth factors for heparin is independent of overall ionic charge, since both positively and negatively charged factors are bound (Maciag, T., et al, *Science* (1984) 225:932-935; Shing, Y., et al, *Science* (1984) 223:1296-1299; Klagsbrun, M., et al, *Proc Natl Acad Sci (USA)* (1985) 82:805-809). The capacity to bind or not to bind to heparin is one measure of differentiation between the activities in the various extracts. For example, EGF and PDGF do not bind strongly to heparin; in fact, EGF does not bind to heparin at all. The other factors above do show strong heparin binding. However, it is believed that acidic brain FGF, ECGF, RDGF, and HGF-alpha are in fact the same factor. Similarly, it is also believed that pituitary FGF, cationic brain FGF, TAF, and HGF- are the same protein. (Lobb, R. R., et al (supra)). A summary and comparison of thirteen endothelial growth factors which have been purified using heparin affinity is found in Lobb, R., et al, *J Biol Chem* (1986) 261:1924-1928.

Using heparin affinity chromatography, basic fibroblast growth factors exhibiting a potent mitogenic activity for capillary endothelium have been isolated from rat chondrosarcoma (Shing, Y., et al, supra) and from bovine cartilage (Sullivan, R., et al, *J Biol Chem* (1985) 260:2399-2403). Thomas, K. A., et al, *Proc Natl Acad Sci (USA)* (1984) 81:357-361, U.S. Pat. No. 4,444,760, purified two heterogeneous forms of an acidic bovine brain fibroblast growth factor having molecular weights of 16,600 and 16,800 daltons. Gospodarowicz and collaborators have shown the

presence in both bovine brains and pituitaries of basic fibroblast growth factor activities and purified these proteins using heparin-affinity chromatography in combination with other purification techniques (Bohlen, P., et al, *Proc Natl Acad Sci (USA)* (1984) 81:5364-5368; Gospodarowicz, D., et al (ibid) 6963-6967). These factors also have molecular weights of approximately 16 kd, as does a similar factor isolated from human placenta (Gospodarowicz, D., et al, *Biochem Biophys Res Comm* (1985) 128:554-562).

The complete sequence for basic FGF derived from bovine pituitary has been determined (Esch, F., et al, *Proc Natl Acad Sci (USA)* (1985) 82:6507-6511). Homogeneous preparations were obtained and showed potent mitogenic activity in in vitro assays with endothelial cells (basic FGF has an ED₅₀ of 60 pg/ml).

Acidic FGF has an ED₅₀ of about 6,000 pg/ml. An N-terminal sequence for acidic FGF derived from bovine brain tissue was determined by Bohlen, P., et al, *EMBO J* (1985) 4:1951-1956. Gimenez-Gallego, G., et al, determined the N-terminal sequences for both acidic and basic FGF prepared from human brain, and compared them to the bovine sequences (*Biochem Biophys Res Comm* (1986) 135:541-548). Their results are consistent with those disclosed herein. Also, the complete amino acid sequence of bovine brain-derived acidic FGF was determined from the isolated protein (Gimenez-Gallego, G., et al, *Science* (1985) 230:1385-1388; Esch, F., et al, *Biochem Biophys Res Comm* (1985) 133:554-562). These two determinations are in agreement with the exception of a single amino acid. However, Esch et al later reported that their sequence is in agreement with that of Gimenez-Gallego et al. The complete amino acid sequence of human acidic FGF was deduced from the gene (Jaye, M., et al, *Science* (1986) 233:541-545 and the complete human protein sequence was also determined by Gimenez-Gallego, G., et al, *Biochem Biophys Res Comm* (1986) 138:611-617 and Harper, J. W., et al, *Biochem* (1986) 25:4097-4103).

The FGF proteins described above and other growth factors are clearly effective in promoting the healing of tissue subjected to trauma (see, e.g., Sporn, M. B., et al, *Science* (1983) 219:1329-1331; Davidson, J. M., et al, *J.C.B.* (1985) 100:1219-1227; Thomas, K. A., et al, *Proc Natl Acad Sci (USA)* (1985) 82:6409-6413). Davidson, et al, (supra) specifically discloses the efficacy of FGF in wound healing. The basic FGF native proteins have been alleged to be useful in treatment of myocardial infarction (Svet-Moldavsky, G. J., et al, *Lancet* (Apr. 23, 1977) 913; U.S. Pat. Nos. 4,296,100 and 4,378,347). In addition, human basic FGF has been found to increase neuronal survival and neurite extension in fetal rat hippocampal neurons (Walicke, P., et al, *Proc Natl Acad Sci (USA)* (1986) 83:3012-3016), suggesting that this factor may also be useful in the treatment of degenerative neurological disorders, such as Alzheimer's disease and Parkinson's disease.

The FGF proteins described above provide an effective means to promote the repair of traumatized tissue as a result of wounding, surgery, burns, fractures or neurological degeneration. However, data is accumulating regarding certain properties of these growth factors which suggests that agonists of FGF may be more therapeutically effective than the native FGF proteins for tissue repair, and in certain circumstances that FGF antagonists may also be useful therapeutically.

For example, agonists of FGF which have greater biological activity as compared to native FGF would be more desirable for use in the wound healing indications described

above. In contrast, antagonists of FGF would be extremely useful in therapies where neovascularization is a dominant pathology and it would be therapeutically useful to inhibit the process of angiogenesis. Therefore, it would also be desirable to construct FGF analogs which antagonize the effects of native FGF thereby inhibiting angiogenesis.

It is considered desirable to provide modifications to the native FGF DNA sequences reported for these growth factors in order to isolate the regions of the protein responsible for the distinct biological activities or regions important in the interactions of the factor with the cellular environment. Having determined the appropriate region or site of the specific interaction, structural analogs can be created which preserve certain activities, e.g. wound healing activity, while reducing or eliminating undesirable functions, such as the sequestration of FGF in the extracellular matrix.

It would also be desirable to insure the availability of these FGF protein analogs in large quantities and in a form free from any toxic or infectious impurities. The human form of the protein is preferred, and perhaps required, for therapeutic use. Since the DNA sequences encoding the proteins for both human acidic and basic FGF have been cloned and expressed by recombinant DNA techniques, site-directed mutagenesis may be employed to produce a variety of acidic and basic FGF analogs. The invention herein provides the means whereby acidic and basic FGF analogs can be obtained in practical quantities and in pure, uncontaminated form.

DISCLOSURE OF THE INVENTION

The invention provides the tools for synthesis and manipulation of fibroblast growth factor analogs useful in effecting accelerated healing of wounds, bone fractures, burn tissue, damaged myocardial tissue, degenerated neurological tissue, or other trauma. Concurrently, fibroblast growth factor antagonists, such as angiogenesis inhibitors, which would be useful for treatment of diseases common to ophthalmology, dermatology and rheumatology where neovascularization is a dominant pathology, and in certain neoplasms that include, but are not limited to, the most highly angiogenic, such as brain tumors, are also provided. Cloning and expression of the genes encoding these analogs are provided by the methods and materials of the invention.

In one aspect, the invention relates to recombinant DNA sequences encoding analogs of human acidic and basic FGF (human aFGF and human bFGF). In other aspects, the invention relates to recombinant vectors bearing these DNA sequences, to host cells transformed with such vectors and harboring these DNA sequences, and to the recombinant proteins produced by these transformed cells. In yet other aspects, the invention relates to methods of producing these fibroblast growth factor analogs using recombinant techniques.

BRIEF DESCRIPTION OF THE DRAWINGS

FIGS. 1-1, 1-2 and FIG. 2 show the native DNA sequences encoding, and the deduced amino acid sequences of, human basic FGF and acidic FGF, respectively.

FIG. 3 shows a comparison of the amino acid sequences for human acidic and basic FGF and the various regions targeted for alteration, including potential heparin-binding domains and receptor-binding regions.

FIG. 4 shows the construction of a synthetic tryptophan operon promoter and operator regulatory sequence, and a restriction site map of plasmid pTRP-233.

FIG. 5 is a flow chart of the construction of plasmid pUC9deIH3-pTSF-3.

FIG. 6 is an illustration of the procedure used to insert any of the FGF analog gene sequences into the expression vector pUC9deIH3-pTSF-3.

FIG. 7 shows the results of wild type bFGF as compared to the double cysteine substituted FGF analog, bFGF-C78/96S, using a high performance liquid chromatography (HPLC) heparin affinity column. FIG. 7 shows the elution of 10 ug of reduced (FIG. 7a) and nonreduced (FIG. 7b) bFGF-C78/96S from a heparin HPLC column developed with a NaCl gradient (0.6M-3.0M). A similar experiment using purified wild type bFGF under reduced (FIG. 7c) and nonreduced (FIG. 7d) conditions is provided for comparison.

MODES OF CARRYING OUT THE INVENTION

A. The Fibroblast Growth Factors

Two different bovine (and analogous human) fibroblast growth factors have been purified to homogeneity by others and partially or completely sequenced. Both factors are capable of mitogenic activity in in vitro assays using cultured cells, such as bovine brain and adrenal cortex-derived capillary endothelial cells, human umbilical vein endothelial cells, bovine adrenal cortex steroidogenic cells, granulosa cells, and vascular smooth muscle cells. In vitro assays employing these cell cultures have been described by Gospodarowicz, D., et al, *J Cell Physiol* (1985) 122:323-332; and Gospodarowicz, D., et al, *J Cell Biol* (1983) 97:1677-1685. More recently, alternative in vitro assays have been described by Esch et al, *Proc Natl Acad Sci (USA)* (1985) 82:6507-6511; and by Gospodarowicz, D., et al, *J Cell Physiol* (1986) 127:121-136. Purified bovine basic FGF has been shown to be angiogenic in vivo in a chicken chorioallantoic membrane assay. (Gospodarowicz, D. in *Hormonal Proteins and Peptides* XII:205-230 (Academic Press). Purified bovine acidic FGF has been shown to be angiogenic in vivo in the same assay (Thomas, K. A., et al, *Proc Natl Acad Sci* (supra)).

Bovine pituitary basic FGF has been completely sequenced by Esch, *Proc Natl Acad Sci USA* (supra); the human sequence is shown in FIG. 1. The reported primary sequence contains 146 amino acids, beginning with the proline residue numbered "10" in FIG. 1; the N-terminal portion of this sequence is in agreement with the sequence previously reported for the N-terminus of the native bovine protein by Bohlen et al, *Proc Natl Acad Sci USA* (supra). A higher molecular weight human basic FGF has been reported from the human hepatoma cell line, SK-HEP-1, by Sullivan, R. J., et al, *J Cell Biol* (1985) 101:108a; by Klagsbrun, M., et al, *Proc Natl Acad Sci USA* (1986) 83:2448-2452; and by Klagsbrun, M. et al, *Proc Natl Acad Sci USA* (1987) 84:1839-1843. Longer forms of FGF have been reported by Sommer, A., et al, *Biochem Biophys Res Comm* (1987) 144:543 (human placental tissue) as well as from pituitary and human prostatic tissue reported by Uneo, et al, *Biochem Biophys Res Comm* (1986) 138:580-588 and Story, et al, *Biochem Biophys Res Comm* (1987) 142:702-709, respectively. Translation of the upstream sequences of FIG. 1 back to a potential ATG translation start codon in human basic FGF DNA shows that it is likely that an additional form of the protein containing the amino acids upstream of the proline shown as residue 10 in FIG. 1 is also produced. The ATG codon lies nine codons upstream from the codon for the proline residue. It is reasonably certain that if the methionine encoded by this ATG serves as the initiating methionine, then it will be processed off when the gene

is expressed in eucaryotic systems. Such processing may or may not occur when the gene is expressed recombinantly in bacterial systems. Thus, the "long" form of the protein expressed in bacteria contains an additional 8 or 9 amino acid sequence at the N-terminus, for a total of 154 or 155 amino acids. All of the investigative groups have also shown that much of this extended FGF is blocked at the N-terminus.

Proteins having FGF activity in the abovementioned in vitro assays and sharing a similar putative N-terminal sequence with the bovine pituitary basic FGF (the 146 amino acid form) have also been isolated from bovine brain, adrenal gland, retina, and from human placenta. The native protein obtained from certain of these tissues is heterogeneous—a second form missing the putative fifteen N-terminal amino acids retains activity. (Gospodarowicz, D., *Meth Enz* (1987) 147A:106–119.) It is considered, therefore, that bovine and human basic FGFs exist in at least three forms, a mature form starting at amino acid 10 in FIG. 1 (a proline), longer forms containing eight additional amino acids at the N-terminus, and shorter forms lacking fifteen amino acids of the putative mature sequences shown. Thus, there is believed to be natively produced "long" basic FGF containing 154 or 155 amino acids (Abraham, J. A., et al, *EMBO J* (1986) 5:2523–2528), "primary" basic FGF containing 146 amino acids, and "short" basic FGF containing 131 amino acids. It is also possible that forms extending even further upstream exist. These FGFs are designated "basic" FGF, because they contain a high number of basic amino acid residues (lysine, arginine, histidine) and are therefore cations at neutral pH.

A protein is defined herein as basic FGF (also referred to as bFGF) if it shows FGF activity in the foregoing assays, binds to heparin, is a cation at neutral pH, and reacts immunologically with antibodies prepared using a synthetic analog of the amino terminal sequence [tyr¹⁰] FGF (1–10) conjugated to bovine serum albumin (if appropriate) or to other antibodies raised against bovine (or human) FGF or synthetic or native peptides thereof. See Baird, A., et al, *Regulatory Peptides* (1985) 10:309–317.

Acidic FGF has been isolated from bovine and human brain by others, and the complete coding sequence for human acidic FGF was determined and is shown in FIG. 2.

The acidic protein also has three known active forms, one having the 140 amino acid sequence beginning at the phenylalanine residue numbered "16" in the figure, and a second shorter form corresponding to amino acids 22–155, and an N-terminal extended form corresponding to 2–155 (blocked by acetylation) Burgess, et al, *Proc Natl Acad Sci USA* (1986) 83:7216. These proteins contain a disproportionate number of acidic amino acid residues, i.e., glutamic and aspartic acids and the proteins are therefore anions at neutral pH.

A protein is defined herein as acidic FGF (also referred to herein as aFGF) if it shows FGF activity in in vitro assays, binds to heparin, is an anion at neutral pH, and is immunologically reactive with antibodies prepared against human or bovine acidic FGF or against synthetic or native peptides thereof.

Acidic FGF and basic FGF are thus used herein to designate the foregoing proteins or proteins having amino acid sequences represented by those shown in FIGS. 1 and 2. Of course, these definitions are not restricted to the specific sequences shown, but include analog proteins which contain accidentally or deliberately induced alterations, such as deletions, additions, extensions, or exchanges of amino acid residues, so long as the biological activity of the FGF

agonists, as measured by the foregoing in vitro assay and immunological cross-reactivity assay, is retained. Analogs of FGF with antagonist activity will, of course, have altered activity and specificity.

The various FGF analogs described herein contain deliberately induced alterations formed by directed mutagenesis techniques. These analogs retain the general secondary structure of FGF but have been mutated so as to produce various antagonist and agonist forms of FGF.

In designing such analogs, Shing et al (*Science* (1984) 223:1269–1299) have demonstrated in vitro that basic FGF binds tightly to heparin and Maciag, T., et al, *Science* (1984) 225:932 have reported that acidic FGF also binds heparin. Thus it is likely that heparin, heparan sulfate, heparin-like glycosaminoglycans, and heparan-like glycosaminoglycans, which are present in the extracellular environment, including the extracellular matrix, may bind FGF in vivo. Since basic FGF binds in the extracellular matrix produced by vascular and capillary endothelial cells in vitro (Baird and Ling, *Biochem Biophys Res Comm* (1987) 142:428–435), it follows that analogs of basic FGF with reduced heparin binding ability will have enhanced potency, as more FGF will reach its targeted receptor and will not be sequestered by heparin and heparin-like compounds in the extracellular environment. These analogs will be more useful therapeutically as lower dosages of the particular analog will be required per treatment.

Baird et al (*Rec Prog Horm Res* (1986) 42:143–205) have recently speculated on the regions of basic FGF, residues 26–31 and residues 115–120 illustrated in FIG. 3, which might mediate the binding to heparin. The ability of clustered basic residues, possibly in conjunction with aromatic residues, to mediate heparin binding has been described previously with respect to other proteins (Schwarzbauer et al, *Cell* (1983) 35:421–431; Cardin et al, *Biochem Biophys Res Comm* (1986) 134:783–789). Mutations created in bFGF, as described herein, replace positively charged amino acids within those targeted regions with neutral or negatively charged residues, with consideration given towards minimizing change in secondary structure of the molecule (e.g., alpha helix, beta sheet, turn motifs). In contrast to the putative heparin binding domains identified above, which do not appear to be the main functional heparin binding domains in the present studies, a third region of bFGF including residues 128–138 which contains a clustering of basic residues, was targeted as a potential heparin binding domain. Preferred mutations targeting the heparin binding domains include bFGF-K128S, bFGF-K128E, bFGF-R129T, bFGF-K134S, bFGF-K138S, and K128S/R129T. Substitutions of a basic or positively charged residue with a negatively charged residue such as glutamic acid are preferred.

Analogs of bFGF are defined as: bFGF-XYZ where X is the amino acid in the native human bFGF sequence that is being mutated, Y is the position of amino acid X, and Z is the amino acid residue that is being substituted for X at position Y.

Mutations of bFGF which are found to decrease or eliminate heparin binding can also be combined with other mutations found to result in the formation of analogs with either agonist or antagonist activity.

It is also within the skill of the art to create additional FGF analogs following the teaching provided herein, wherein those residues important for heparin binding are changed to other neutral amino acids (e.g., serine, alanine, glycine, etc.), or negatively charged amino acids (e.g., glutamic acid, aspartic acid), or deleted in order to reduce heparin binding

activity as tested by HPLC heparin-affinity analysis as described herein. Analogs of the acidic form of FGF can be constructed as described above by deleting positively charged amino acids or by replacing positively charged amino acids within the corresponding heparin-binding domains (23–27, 115–120, 127–137) with neutral or negatively charged amino acids.

It has been found that bacterially-produced recombinant proteins can be difficult to recover in an active form. For example, it is known that cysteine-containing proteins produced in bacteria often form incorrect intramolecular cysteines which can inhibit biological function (see human interleukin-2; Wang et al, *Science* (1984) 224:1431–1433 and human fibroblast interferon; Mark et al, *Proc Natl Acad Sci (USA)* (1984) 81:5662–5666). Modifications of one or more of the cysteine residues present in the native FGF proteins may minimize incorrect disulfide bridge formation, eliminate the need for use of reducing agents to stabilize the FGF protein, and hence reduce multimerization or incorrect disulfide bonds thereby increasing the recoverable yield of the recombinantly produced analog, increasing the uniformity of the FGF preparation by maintaining it over time in a monomeric form, improving its shelf stability and reducing its half life when applied to wounds. Unexpectedly, these analogs have been shown to have augmented biological activity.

Generally, the above modifications at cysteine residues are conducted by changing a single nucleotide within the codon specifying a particular cysteine, corresponding to an amino acid substitution in the resulting protein. Cysteine residues occur in the basic form of FGF at positions 34, 78, 96, and 101, and occur in the acidic form at positions 31, 98, and 132. Since the disulfide structure of native FGF is not known, both single and multiple cysteine substitutions of FGF are exemplified herein. While these modifications produce a change in the primary structure of the protein analog, preferred analogs will generally retain the ability to effect cellular responses normally induced by FGF, unless the cysteine-substituted analogs are combined with other antagonist changes.

These same cysteine substitutions can be made in combination with other analog substitutions, such as the aforementioned heparin-binding mutants, to produce yet additional illustrative FGF analogs. Correspondingly, any of the aforementioned FGF analogs can be modified to contain one or more of the amino acid substitutions described below to produce a desired analog.

Antagonists of bFGF activity would have clinical applications in a variety of pathologies related to abnormal persistent angiogenesis (Folkman, J. and Klagsbrun, M., *Science* (1987) 235:442–447) including diabetic retinopathy, retrolental fibroplasia, neovascular glaucoma, rheumatoid arthritis, hemangiomas, angiofibromas, psoriasis, atherosclerosis and as contraceptives. In addition, it has been shown that certain solid tumors require neovascularization in order to sustain growth. Given the important role FGF plays in the process of angiogenesis, it is clear that analogs of FGF which are capable of inhibiting its effect would be useful in treating these diseases therapeutically. Thus, analogs which bind the FGF receptor yet do not elicit a biological response or that demonstrate a reduced biological response will exhibit useful antagonist properties.

The ability to elaborate a specific cell surface receptor for basic FGF has been described in a variety of cell types including baby hamster kidney cells (Neufeld and Gospodarowicz, *J Biol Chem* (1985) 260:13860–13868), bovine epithelial lens cells (Moenner, et al. *Proc Natl Acad*

Sci (USA) (1986) 83:5024–5028), Swiss 3T3 and a murine skeletal muscle cell line (Olwin and Hauschka, *Biochemistry* (1986) 25:3487–3492) and Swiss 3T3 and aortic endothelial cells (Huang et al, *J Biol Chem* (1986) 261:11600–11607).

In addition binding studies have suggested that both the basic and acidic forms of FGF can bind to the same high affinity receptor (Olwin and Hauschka, supra, and Neufeld and Gospodarowicz, *J Biol Chem* (1986) 261:5631–5637).

The interaction of a hormone (e.g., bFGF) with its receptor results in a tight, specific molecular association. This association may involve any or all of the known intermolecular attractive forces such as ion pairing or van der Waals forces. The specificity and the stability of the association are due to what may be thought of as “exactness of fit” (the precise three-dimensional molecular conformations of the two proteins, receptor and hormone) and “tightness of fit” (the fact that these molecular structures are composed of precise amino acid sequences which therefore results in specific intermolecular attractions due to energetically favorable juxtaposition of amino acid side chains). Thus, amino acid substitutions, deletions and insertions within receptor binding regions may effect either molecular conformation of the region or amino acid side chain interactions (between hormone and receptor) or both. Changes which stabilize favorable conformation or enhance amino acid side chain interactions will result in increased receptor affinity while those which destabilize favorable conformation or decrease amino acid side chain interactions will result in decreased receptor affinity. The former changes are useful in themselves as their introduction into agonists may result in more potent agonists and their introduction into antagonists may result in more potent antagonists. The latter changes are useful in terms of defining amino acid segments crucial to receptor binding.

Schubert et al (*J Cell Biol* (1987) 104:635–643) have shown that synthetic peptides containing fragments of bFGF (residues 33–77 and 112–129 numbered according to FIG. 3) inhibit binding of bFGF to its receptor. Therefore, these regions appear to contain FGF receptor binding sequences. We have introduced amino acid substitutions into human basic FGF within these putative receptor binding regions and additional regions adjacent to the latter (e.g., amino acids 99–111 which exhibit strong homology to the equivalent amino acid sequence region in acidic FGF). Both charged (positive and negative) and aromatic amino acids were targeted for replacement with neutral residues. These substitutions were made with consideration given towards minimizing changes in the secondary structure of the resultant protein. Accordingly, the analogs D99A and R116T appear to exhibit increased receptor affinity and 3T3 mitogenic activity, respectively, whereas analogs E105S and Y112A exhibit decreased receptor binding (see Table 3 herein).

For purposes of the present invention the following terms are defined below. “Agonist” refers to an FGF analog capable of combining with the FGF receptor and producing a typical biological response. For example, an FGF agonist might be a protein that can bind to the FGF receptor but has reduced ability to bind heparin, thereby creating a more potent therapeutic.

“Antagonist” refers to an FGF analog that opposes the effects of FGF by a competitive mechanism for the same receptor sites. The antagonist has reduced ability to induce secondary biological responses normally associated with FGF.

“Site-specific mutagenesis” or “directed mutagenesis” refers to the use of the oligonucleotide-directed mutagenesis

procedure, which entails using a synthetic oligonucleotide primer that is complementary to the region of the bFGF gene at the specific codon or codons to be altered, but which contains single or multiple base changes in that codon. By this technique, a designer gene may be produced that results in a specific amino acid being replaced with any other amino acid of choice. When deletion is desired the oligonucleotide primer lacks the specific codon. Conversion of, for example, a specific cysteine, to neutral amino acids such as glycine, valine, alanine, leucine, isoleucine, tyrosine, phenylalanine, histidine, tryptophan, serine, threonine or methionine is a preferred approach. Serine and alanine are the most preferred replacements because of their chemical analogy to cysteine. When a cysteine is deleted, the mature analog is one amino acid shorter than the native parent protein or the microbially produced wild type bFGF.

“Purified” or “pure” refers to material which is free from substances which normally accompany it as found in its native state. Thus “pure” acidic human FGF (hFGF), for example, refers to acidic hFGF which does not contain materials normally associated with its in situ environment in human brain or pituitary. Of course, “pure” acidic hFGF may include materials in covalent association with it, such as glycoside residues.

“Operably linked” refers to a juxtaposition wherein the components are configured so as to perform their usual function. Thus, control sequences or promoters operably linked to a coding sequence are capable of effecting the expression of the coding sequence.

“Control sequence” refers to a DNA sequence or sequences which are capable, when properly ligated to a desired coding sequence, of affecting its expression in hosts compatible with such sequences. Such control sequences include at least promoters in both procaryotic and eucaryotic hosts, and optionally, transcription termination signals. Additional factors necessary or helpful in effecting expression may also be identified. As used herein, “control sequences” simply refers to whatever DNA sequence may be required to effect expression in the particular host used.

“Cells” or “cell cultures” or “recombinant host cells” or “host cells” are often used interchangeably as will be clear from the context. These terms include the immediate subject cell, and, of course, the progeny thereof. It is understood that not all progeny are exactly identical to the parental cell, due to chance mutations or differences in environment. However, such altered progeny are included in these terms, so long as the progeny retain the desired characteristics conferred on the originally transformed cell. In the present case, for example, such a characteristic might be the ability to produce recombinant FGF analogs.

B. General Description Utility and Administration

The invention provides DNAs encoding growth factor protein analogs which have two diverse applications. The first application is similar to FGF in that the analogs augment tissue repair by encouraging vascularization and/or cell growth or cell survival. These purified growth factors are generally applied topically to the traumatized or diseased tissue in order to stimulate vascularization, regeneration, and healing. Appropriate substrates are burns, wounds, bone fractures, surgical abrasions such as those of plastic surgery, or others requiring repair. Because application of these factors accelerates healing, they also reduce the risk of infection.

Indications wherein FGF is of value in encouraging neovascularization include musculo-skeletal conditions such as bone fractures, ligament and tendon repair,

tendonitis, and bursitis; skin conditions such as burns, cuts, lacerations, bed sores, and slow-healing ulcers such as those seen in diabetics; and in tissue repair during ischaemia and myocardial infarction.

In addition to analogs which augment wound healing, analogs of FGF can be constructed which inhibit angiogenesis. Analogs of FGF which can antagonize the FGF angiogenesis activity would be clinically useful for treating certain diseases where neovascularization is the dominant pathology, such as retinopathies of the eye including diabetic retinopathy and neovascular glaucoma; skin disorders including psoriasis and retrolental fibroplasia; chronic inflammation; rheumatoid arthritis; atherosclerosis; and certain neoplasms that are highly angiogenic, such as the growth of certain benign and malignant tumors such as hemangiomas and angiofibromas, and solid tumors.

Formulations of the recombinantly produced growth factors using available excipients and carriers are prepared according to standard methods known to those in the art. The proteins can be formulated as eyedrops, lotions, gels, powder, dressing, as part of a controlled release system, or ointments with additional active ingredients, such as antibiotics, if desired.

For topical administration, which is the most appropriate with regard to superficial lesions, standard topical formulations are employed using, for example, 10 ng/ml–100 mg/ml solutions; the preferred range is 10 ug/ml–10 mg/ml. Such solutions would be applied up to 6 times a day to the affected area. In certain applications, such as burns, a single dose would be preferred. In other applications, such as ulcers, multiple doses may be preferred. The concentration of the ointment or other formulation depends, of course, on the severity of the wound or stage of disease and the nature of the subject. In most protocols, the dose is lowered with time to lessen likelihood of scarring. For example, the most severe wounds, such as third degree burns, are typically treated with a 100 ug/ml composition, but as healing begins, the dose is progressively dropped to approximately 10 ug/ml or lower, as the wound heals. A topical formulation for EGF using BSA as carrier was disclosed by Franklin, J. D., et al, *Plastic and Reconstructive Surg* (1979) 64:766–770.

For treatment of pathologies related to persistent angiogenesis wherein FGF inhibitors are to be applied, the concentration of the formulation is generally 10-fold higher, regardless of the mode of administration. The higher dosage assures that the FGF inhibitor is able to compete effectively with endogenously produced FGF. Thus for topical administration of the FGF inhibitor used to treat psoriasis and retrolental fibroplasia, the dosage would be increased 10-fold.

For arthritis and bone and tissue repair, administration is preferred locally by means of subcutaneous implant, staples or slow release formulation implanted directly proximal the target. Surgery may be required for such conditions as bone injuries, thus making implantation directly practical. Slow-release forms can be formulated in polymers, such as Hydron (Langer, R., et al, *Nature* (1976) 263:797–799) or Elvax 40P (Dupont) (Murray, J. B., et al, *In Vitro* (1983) 19:743–747). Other sustained-release systems have been suggested by Hsieh, D. S. T., et al, *J Pharm Sci* (1983) 72:17–22), and a formulation specifically for epidermal growth factor, but not preferred in the present invention, is suggested by Buckley, A., *Proc Natl Acad Sci (USA)* (1985) 82:7340–7344.

As with topical administration, for sustained release delivery, the concentration of FGF in the formulation depends on a number of factors, including the severity of the

condition, the stability of FGF at 37° C., the rate of FGF release from the polymer, and the agonist or antagonist nature of the FGF analog. In general, the formulations are constructed so as to achieve a constant local concentration of about 100 times the serum level of factor or 10 times the tissue concentration, as described by Buckley et al (*Proc Natl Acad Sci (USA)* (supra)). Based on an FGF concentration in tissue of 5–50 ng/g wet weight (comparable to EGF at 60 ng/g wet weight), release of 50–5000 ng FGF per hour is acceptable. The initial concentration, of course, depends on the severity of the wound or advancement of pathology.

For treatment in diseases common to ophthalmology, such as retinopathies and neovascular glaucoma, eyedrop formulation or direct injection into the eye would be two preferred routes of administration. Liquid formulations for these applications are generally known in the art and include formulation in buffer or physiological saline, or other appropriate excipient. Dosage levels may be supplied between 1 ug/ml and 10 mg/ml from two to four times a day.

It is expected that FGF may act in concert, and even synergistically, with other growth factors such as epidermal growth factor (EGF), the transforming growth factors (TGF-alpha or TGF- β), insulin-like growth factors (IGF-1 and IGF-2), and/or platelet-derived growth factor (PDGF). In addition, specifically for bone repair, it may act in synergy with agonists or antagonists of parathyroid hormone or calcitonin, since these compounds promote bone growth and resorption. Therefore, also included within the compositions and administration protocols of the invention are embodiments wherein the FGF of the invention is administered in the same composition with, or in the same protocol with, one or more of the foregoing factors, thus more effectively to achieve the desired tissue repair.

Since FGF is effective in promoting neurite outgrowth, nerve regeneration, and neuronal survival, it may be useful for treatment of certain neurological disorders such as Alzheimer's and Parkinson's diseases, amyotrophic lateral sclerosis, stroke, peripheral neuropathies, and general aging of the nervous system, as well as traumatic injury to the spinal cord and peripheral nerves. Administration of the drug for these indications is preferably by implant in formulations similar to those set forth above in connection with rheumatoid arthritis and bone healing. The drug may also be delivered by means of implants of cell cultures by means of implants of cell cultures which produce FGF. Treatment of neurological disorders may also involve transplantation of new cells or tissues to functionally replace damaged neural tissue (e.g., adrenal and fetal brain tissue transplants in Parkinsonian patients). In such cases, the degree of success of transplantation as well as the degree of function of the transplanted tissue are enhanced by treating the cell cultures or tissue explants with the FGF or analog preparations of the invention prior to transplantation and/or by administration of FGF or FGF analogs of the invention following transplantation.

FGF and analogs thereof may also be injected directly into the spinal fluid or into the brain by means of canulation or by administration using osmotic minipumps or they may be applied systemically. For atherosclerosis peripheral neuropathies and the like, and tumor angiogenesis, systemic administration is preferred, with administration of the drug delivered initially at the time of surgery, where appropriate.

Systemic formulations are generally as are known in the art and include formulation in buffer or physiological saline, or other appropriate excipient. Dosage levels for FGF agonist administration are approximately those of wound healing; however, for tissue culture, explant maintenance, ath-

erosclerosis or tumor angiogenesis, it may be supplied at 1.0–100 ng/ml of serum or culture medium.

In addition, it has been shown that angiogenic stimuli, such as those provided by the FGF proteins discussed herein, result in the release of plasminogen activator (PA) and of collagenase in vitro (Gross, J. L., et al, *Proc Natl Acad Sci (USA)* (1983) 80:2623–2627). Therefore, the FGF proteins of the invention are also useful in treatment of conditions which respond to these enzymes. While it may be necessary in acute situations (such as the presence of a blood clot associated with stroke or heart attack) directly to administer large doses of PA to dissolve the clot, for treatment of chronic propensity to form embolisms, administration of FGF to maintain a suitable level of PA in the blood stream may be desirable. Therefore, for this indication, systemic administration of the drug, especially an analog with reduced heparin-binding ability, using conventional means such as intramuscular or intravenous injection, is preferred.

The invention provides practical quantities of pure FGF analogs for use in connection with the foregoing indications. Specific growth factors are exemplified herein, each of which is apparently active in at least three forms. Both acidic and basic analogs are considered to occur in long, primary, and short forms, as described above. It is considered that the N-terminal methionine of the long forms is processed off when the protein is produced in eucaryotic systems, and that the subsequent amino acid residue is derivatized, probably by acetylation, post-translation.

While FGF in its various forms does not have a recognized signal sequence, it must somehow be secreted or retrieved from the cell, since it acts outside the cells producing it at a membrane-bound receptor. Therefore, while it may not be secreted by the recognized constitutive secretion pathway, its secretion is accomplished by some means, for example by cell lysis or by exocytosis, by association with a glycosaminoglycan, such as heparan sulfate or heparin. For most tissues from which FGF is naturally derived, and for many mammalian expression systems, such release may be achieved by securing exocytosis with a calcium ionophore, such as the commonly employed A23187 (CalBiochem), which, in in vitro conditions, is added to the culture medium at 1–10 uM in the presence of 1 mM CaCl₂. For expression systems derived from macrophages or monocytes, other activation methods have been shown to be effective, such as the addition of lipopolysaccharide (LPS) at 10 ug/ml or the addition of *E. coli* endotoxin (Difco) (300 ng/ml). These stimulators have been shown to release the analogous factor interleukin-1 from macrophages by March, C. J., et al, *Nature* (1985) 315:641–647. These techniques can also be employed in releasing recombinantly produced FGF proteins when produced intracellularly without added signal sequences, as described below. Additional stimulators for release of intracellularly produced proteins include the phorbol esters and the triglycerides.

55 Gene Retrieval

The general strategy whereby the illustrated FGF-encoding sequences were obtained is as described in Abraham, J. A. et al, *EMBO J* (1986) supra, and Abraham, J. A. et al, *Science* (1986) 233:545–548, each of these references are incorporated herein by reference.

60 Expression of FGF Genes

The DNA sequences described herein can be expressed in appropriate expression systems. Of course, for the DNAs disclosed herein, the foregoing protocol for retrieving the genomic or cDNA FGF sequences need not be repeated, but conventional chemical synthesis methods can suitably be employed. Alternatively, the gene encoding basic FGF can

be retrieved from the deposited bacteriophage lambdaBB2 and converted to the human form. Site-directed mutagenesis permits adjustment of the DNA to obtain any desired form of the protein. DNA sequences can be provided with appropriate controls suitable for any host, including bacteria, yeast, or eucaryotic cells. Exemplary control sequence DNAs and hosts are given in paragraph C.1 below.

In particular, complete DNA encoding any of the FGF analogs described herein can be constructed, for example, using a combination of recombinant and synthetic methods to obtain each of the DNA analog sequences of FGF. These gene sequences have been constructed with convenient restriction sites bounding the FGF coding sequence so that the entire gene may be inserted on an ~503 bp NcoI-HindIII restriction fragment for insertion into an appropriately digested host vector such that the FGF coding sequence is operably linked to control sequences present on the vector. Intracellularly produced forms of the FGF protein analogs can be obtained by cell lysis, or their release from the cells can be stimulated by using heterologous signal sequences fused to the gene sequence using the known relationship of the signal sequence to cleavage site to obtain the protein in the desired form. Particularly preferred are bacterial expression systems which utilize control systems compatible with *E. coli* cells, such as plasmids pUC9-TSF11 and pUC9delH3-pTSF-3. These vectors are derived from pUC9 (Messing and Vieira, *Gene* (1982) 19:259–268), which contains parts of pBR322 and M13mp9 and a multiple cloning site polylinker.

The recombinant FGF proteins thus produced are then purified in a manner similar to that utilized for purification of FGF from natural sources, but purification is considerably simpler, as the proteins form a much larger proportion of the starting material.

B. Standard Methods

Most of the techniques which are used to transform cells, construct vectors, construct oligonucleotides, perform site-specific mutagenesis, and the like are widely practiced in the art, and most practitioners are familiar with the standard resource materials which describe specific conditions and procedures. However, for convenience, the following paragraphs may serve as a guideline.

B.1. Hosts and Control Sequences

Both procaryotic and eucaryotic systems may be used to express the FGF analog encoding sequences; procaryotic hosts are, of course, the most convenient for cloning procedures. Procaryotes most frequently are represented by various strains of *E. coli*; however, other microbial strains may also be used. Plasmid vectors which contain replication sites, selectable markers and control sequences derived from a species compatible with the host are used; for example, *E. coli* is typically transformed using derivatives of pBR322, a plasmid derived from an *E. coli* species by Bolivar, et al, *Gene* (1977) 2:95. pBR322 contains genes for ampicillin and tetracycline resistance, and thus provides multiple selectable markers which can be either retained or destroyed in constructing the desired vector. Commonly used procaryotic control sequences which are defined herein to include promoters for transcription initiation, optionally with an operator, along with ribosome binding site sequences, include such commonly used promoters as the -lactamase (penicillinase) and lactose (lac) promoter systems (Chang, et al, *Nature* (1977) 198:1056), the tryptophan (trp) promoter system (Goeddel, et al, *Nucleic Acids Res* (1980) 8:4057), the lambda-derived P_L promoter (Shimatake, et al, *Nature* (1981) 292:128) and N-gene ribosome binding site, and the trp-lac (trc) promoter system (Amann and Brosius, *Gene* (1985) 40:183).

In addition to bacteria, eucaryotic microbes, such as yeast, may also be used as hosts. Laboratory strains of *Saccharomyces cerevisiae*, Baker's yeast, are most used although a number of other strains or species are commonly available. Vectors employing, for example, the 2 u origin of replication of Broach, J. R., *Meth Enz* (1983) 101:307, or other yeast compatible origins of replication (see, for example, Stinchcomb, et al, *Nature* (1979) 282:39, Tschumper, G., et al, *Gene* (1980) 10:157 and Clarke, L., et al, *Meth Enz* (1983) 101:300) may be used. Control sequences for yeast vectors include promoters for the synthesis of glycolytic enzymes (Hess, et al, *J Adv Enzyme Reg* (1968) 7:149; Holland, et al, *Biochemistry* (1978) 17:4900). Additional promoters known in the art include the promoter for 3-phosphoglycerate kinase (Hitzeman, et al, *J Biol Chem* (1980) 255:2073). Other promoters, which have the additional advantage of transcription controlled by growth conditions and/or genetic background are the promoter regions for alcohol dehydrogenase 2, isocytochrome C, acid phosphatase, degradative enzymes associated with nitrogen metabolism, the alpha factor system and enzymes responsible for maltose and galactose utilization. It is also believed terminator sequences are desirable at the 3' end of the coding sequences. Such terminators are found in the 3' untranslated region following the coding sequences in yeast-derived genes.

It is also, of course, possible to express genes encoding polypeptides in eucaryotic host cell cultures derived from multicellular organisms. See, for example, Axel, et al, U.S. Pat. No. 4,399,216. These systems have the additional advantage of the ability to splice out introns and thus can be used directly to express genomic fragments. Useful host cell lines include VERO, HeLa baby hamster kidney (BHK), CV-1, COS, MDCK, NIH 3T3, L, and Chinese hamster ovary (CHO) cells. Expression vectors for such cells ordinarily include promoters and control sequences compatible with mammalian cells such as, for example, the commonly used early and late promoters from Simian Virus 40(SV40) (Fiers, et al, *Nature* (1978) 273:113), or other viral promoters such as those derived from polyoma, Adenovirus 2, bovine papilloma virus, or avian sarcoma viruses. The controllable promoter, hMTII (Karin, M., et al, *Nature* (1982) 299:797–802) may also be used. General aspects of mammalian cell host system transformations have been described by Axel (supra). It now appears, also that "enhancer" regions are important in optimizing expression; these are, generally, sequences found upstream or downstream of the promoter region in noncoding DNA regions. Origins of replication may be obtained, if needed, from viral sources. However, integration into the chromosome is a common mechanism for DNA replication in eucaryotes.

B.2. Transformations

Depending on the host cell used, transformation is done using standard techniques appropriate to such cells. The calcium treatment employing calcium chloride, as described by Cohen, S. N., *Proc Natl Acad Sci (USA)* (1972) 69:2110, or the RbCl₂ method described in Maniatis, et al, *Molecular Cloning: A Laboratory Manual* (1982) Cold Spring Harbor Press, p. 254 and Hanahan, D., *J Mol Biol* (1983) 166:557–580 may be used for procaryotes or other cells which contain substantial cell wall barriers. For mammalian cells without such cell walls, the calcium phosphate precipitation method of Graham and van der Eb, *Virology* (1978) 52:546, optionally as modified by Wigler, M., et al, *Cell* (1979) 16:777–785 may be used. Transformations into yeast may be carried out according to the method of Beggs, J. D., *Nature* (1978) 275:104–109 or of Hinnen, A., et al, *Proc Natl Acad Sci (USA)* (1978) 75:1929.

B.3. Vector Construction

Construction of suitable vectors containing the desired coding and control sequences employs standard ligation and restriction techniques which are well understood in the art. Isolated plasmids, DNA sequences, or synthesized oligonucleotides are cleaved, tailored, and religated in the form desired.

The DNA sequences which form the vectors are available from a number of sources. Backbone vectors and control systems are generally found on available "host" vectors which are used for the bulk of the sequences in construction. Typical sequences have been set forth in §C.1 above. For the pertinent coding sequence, initial construction may be, and usually is, a matter of retrieving the appropriate sequences from cDNA libraries, genomic DNA libraries, or deposited plasmids. However, once the sequence is disclosed it is possible to synthesize the entire gene sequence in vitro starting from the individual nucleoside derivatives. The entire gene sequence for genes of sizeable length, e.g., 500–1000 bp may be prepared by synthesizing individual overlapping complementary oligonucleotides and filling in single stranded nonoverlapping portions using DNA polymerase in the presence of the deoxyribonucleotide triphosphates. This approach has been used successfully in the construction of several genes of known sequence. See, for example, Edge, M. D., *Nature* (1981) 292:756; Nambair, K. P., et al, *Science* (1984) 223:1299; Jay, Ernest, *J Biol Chem* (1984) 259:6311.

Synthetic oligonucleotides are prepared by either the phosphotriester method as described by Edge, et al, *Nature* (supra) and Duckworth, et al, *Nucleic Acids Res* (1981) 9:1691 or the phosphoramidite method as described by Beaucage, S. L., and Caruthers, M. H., *Tet Letts* (1981) 22:1859 and Matteucci, M. D., and Caruthers, M. H., *J Am Chem Soc* (1981) 103:3185 and can be prepared using commercially available automated oligonucleotide synthesizers. Kinasing of single strands prior to annealing or for labeling is achieved using an excess, e.g., approximately 10 units of polynucleotide kinase to 1 nmole substrate in the presence of 50 mM Tris, pH 7.6, 10 mM MgCl₂, 5 mM dithiothreitol, 1–2 mM ATP, 1.7 pmoles [λ -³²P]-ATP (2.9 mCi/mMole), 0.1 mM spermidine, 0.1 mM EDTA.

Once the components of the desired vectors are thus available, they can be excised and ligated using standard restriction and ligation procedures.

Site specific DNA cleavage is performed by treating with the suitable restriction enzyme (or enzymes) under conditions which are generally understood in the art, and the particulars of which are specified by the manufacturer of these commercially available restriction enzymes. See, e.g., New England Biolabs, Product Catalog. In general, about 1 μ g of plasmid or DNA sequence is cleaved by one unit of enzyme in about 20 μ l of buffer solution; in the examples herein, typically, an excess of restriction enzyme is used to insure complete digestion of the DNA substrate. Incubation times of about one hour to two hours at about 37° C. are workable, although variations can be tolerated. After each incubation, protein is removed by extraction with phenol/chloroform, and may be followed by ether extraction, and the nucleic acid recovered from aqueous fractions by precipitation with ethanol. If desired, size separation of the cleaved fragments may be performed by polyacrylamide gel or agarose gel electrophoresis using standard techniques. A general description of size separations is found in *Methods in Enzymology* (1980) 65:499–560.

Restriction cleaved fragments may be blunt ended by treating with the large fragment of *E. coli* DNA polymerase

I (Klenow) in the presence of the four deoxynucleotide triphosphates (dNTPs) using incubation times of about 15 to 25 min at 20° to 25° C. in 50 mM Tris pH 7.6, 50 mM NaCl, 6 mM MgCl₂, 6 mM DTT and 0.1–1.0 mM dNTPs. The Klenow fragment fills in at 5' single-stranded overhangs but chews back protruding 3' single strands, even though the four dNTPs are present. If desired, selective repair can be performed by supplying only one of the, or selected, dNTPs within the limitations dictated by the nature of the overhang. After treatment with Klenow, the mixture is extracted with phenol/chloroform and ethanol precipitated. Treatment under appropriate conditions with Si nuclease or BAL-31 results in hydrolysis of any single-stranded portion.

Ligations are performed in 15–50 μ l volumes under the following standard conditions and temperatures: for example, 20 mM Tris-Cl pH 7.5, 10 mM MgCl₂, 10 mM DTT, 33 μ g/ml BSA, 10 mM–50 mM NaCl, and either 40 μ M ATP, 0.01–0.02 (Weiss) units T4 DNA ligase at 0° C. (for "sticky end" ligation) or 1 mM ATP, 0.3–0.6 (Weiss) units T4 DNA ligase at 14° C. (for "blunt end" ligation). Intermolecular "sticky end" ligations are usually performed at 33–100 μ g/ml total DNA concentrations (5–100 nM total end concentration). Intermolecular blunt end ligations are performed at 1 μ M total ends concentration.

In vector construction employing "vector fragments", the vector fragment is commonly treated with bacterial alkaline phosphatase (BAP) or calf intestinal alkaline phosphatase (CIP) in order to remove the 05' phosphate and prevent self-ligation of the vector. Digestions are conducted at pH 8 in approximately 10 mM Tris-HCl, 1 mM EDTA using about 1 unit of BAP or CIP per μ g of vector at 60° C. for about one hour. In order to recover the nucleic acid fragments, the preparation is extracted with phenol/chloroform and ethanol precipitated. Alternatively, religation can be prevented in vectors which have been double digested by additional restriction enzyme digestion and separation of the unwanted fragments.

For portions of vectors derived from cDNA or genomic DNA which require sequence modifications, site specific mutagenesis may be used (Zoller, M. J., and Smith, M. *Nucleic Acids Res* (1982) 10:6487–6500 and Adelman, J. P., et al, *DNA* (1983) 2:183–193). This is conducted using a primer synthetic oligonucleotide primer complementary to a single stranded phage DNA to be mutagenized except for limited mismatching, representing the desired mutation.

The size of the oligonucleotide primer is determined by the requirement for stable hybridization of the primer to the region of the gene in which the mutation is to be induced, and by the limitations of the currently available methods for synthesizing oligonucleotides. The factors to be considered in designing oligonucleotides for use in oligonucleotide-directed mutagenesis (e.g., overall size, size of portions flanking the mutation site) are described by Smith, M. and Gillam, S. in *Genetic Engineering: Principles and Methods*, Plenum Press (1981) 3:1–32. In general the overall length of the oligonucleotide will be such as to optimize stable, unique hybridization at the mutation site with the 5' and 3' extensions from the mutation site being of sufficient size to avoid editing of the mutation by the exonuclease activity of the DNA polymerase. Oligonucleotides used for mutagenesis in accordance with the present invention usually contain from about 18 to about 45 bases, preferably from about 23 to about 27 bases. They will usually contain at least about three bases 3' of the altered or missing codon.

The method for preparing the modified bFGF genes generally involves inducing a site-specific mutagenesis in the bFGF gene at a specific codon using a synthetic nucle-

otide primer which omits the codon or alters it so that it codes for another amino acid. It must be recognized that when deletions are introduced, the proper reading frame for the DNA sequence must be maintained for expression of the desired protein.

The primer is hybridized to single-stranded phage such as M13, fd, or deltaX174 into which a strand of the bFGF gene has been cloned. It will be appreciated that the phage may carry either the sense strand or antisense strand of the gene. When the phage carries the antisense strand the primer is identical to the region of the sense strand that contains the codon to be mutated except for a mismatch with that codon that defines a deletion of the codon or a triplet that codes for another amino acid. When the phage carries the sense strand the primer is complementary to the region of the sense strand that contains the codon to be mutated except for an appropriate mismatch in the triplet that is paired with the codon to be deleted. Conditions that may be used in the hybridization are described by Smith, M. and Gillam, S., *supra*. The temperature will usually range between about 0° C. and 70° C., more usually about 10° C. to 50° C. After the hybridization, the primer is extended on the phage DNA by reaction with DNA polymerase I, T₄ DNA polymerase, reverse transcriptase, or other suitable DNA polymerase. The resulting dsDNA is converted to closed circular dsDNA by treatment with a DNA ligase such as T₄ DNA ligase. DNA molecules containing single-stranded regions may be destroyed by S1 endonuclease treatment.

The resulting double-stranded DNA is transformed into a phage-supporting host bacterium. Cultures of the transformed bacteria are plated in top agar, permitting plaque formation from single cells which harbor the phage.

Theoretically, 50% of the new plaques will contain the phage having, as a single strand, the mutated form; 50% will have the original sequence. The resulting plaques are washed after hybridization with kinased synthetic primer at a wash temperature which permits binding of an exact match, but at which the mismatches with the original strand are sufficient to prevent binding. Plaques which hybridize with the probe are then picked, cultured, and the DNA recovered.

C.4. Verification of Construction

In the constructions set forth below, correct ligations for plasmid construction are confirmed by first transforming *E. coli* strain MC1061 obtained from Dr. M. Casadaban (Casadaban, M., et al, *J Mol Biol* (1980) 138:179–207) or other suitable host with the ligation mixture. Successful transformants are selected by ampicillin, tetracycline or other antibiotic resistance or using other markers depending on the mode of plasmid construction, as is understood in the art. Plasmids from the transformants are then prepared according to the method of Clewell, D. B., et al, *Proc Natl Acad Sci (USA)* (1969) 62:1159, optionally following chloramphenicol amplification (Clewell, D. B., *J Bacteriol* (1972) 110:667). Several mini DNA preps are commonly used, e.g., Holmes, D. S., et al, *Anal Biochem* (1981) 114:193–197 and Birnboim, H. C., et al, *Nucleic Acids Res* (1979) 7:1513–1523. The isolated DNA is analyzed by dot blot analysis as described by Kafatos, F. C., et al, *Nucl Acid Res* (1977) 7:1541–1552, restriction enzyme analysis, or sequenced by the dideoxy nucleotide method of Sanger, F., et al, *Proc Natl Acad Sci (USA)* (1977) 74:5463, as further described by Messing, et al, *Nucleic Acids Res* (1981) 9:309, or by the method of Maxam, et al, *Methods in Enzymology* (1980) 65:499.

C.5. Hosts Exemplified

Host strains used in cloning and procaryotic expression herein are as follows:

For cloning and sequencing, and for expression of construction under control of most bacterial promoters, *E. coli* strains such as MC1061, DH1, RR1, B, C600hf1, K803, HB101, JA221, and JM101 were used.

5 D. Illustrative Procedure

The following examples are intended to illustrate but not to limit the invention. The DNA encoding the FGF starting material was obtained initially by screening a bovine genomic library and obtaining a pivotal probe, followed by retrieval of additional DNA. However, it would not be necessary to repeat this procedure, as the sequence of the pivotal probe is now known and could thus be constructed chemically in vitro. In addition, bacteriophage harboring bovine aFGF and bFGF and human aFGF and bFGF sequences are deposited at the American Type Culture Collection. Thus, the DNA sequence used as the starting material for the mutagenesis in the following examples is available from a variety of sources.

20 EXAMPLE 1

Construction of pTrp-233 Bacterial Expression Plasmid

1. Construction of the Synthetic Tryptophan Operon Promoter and Operator Regulatory Sequence

The ten oligodeoxynucleotides shown in FIG. 4 were synthesized by the phosphotriester method and purified. 500 pmole of each oligodeoxynucleotide except 1 and 10 were phosphorylated individually in 20 ul containing 60 mM Tris-HCl, pH 8, 15 mM DTT, 10 mM MgCl₂, 20 uCi of [λ -³²P]-ATP and 20 units of polynucleotide kinase (P/L Biochemicals) for 30 min. at 37° C. This was followed by the addition of 10 ul containing 60 mM Tris-HCl, pH 8, 15 mM DTT, 10 mM MgCl₂, 1.5 mM ATP and 20 additional units of polynucleotide kinase followed by another 30 min incubation at 37° C. Following incubation the samples were incubated at 100° C. for 5 min. 500 pmole of oligodeoxynucleotides 1 and 10 were diluted to 30 ul in the above buffer without ATP.

16.7 pmole of each oligodeoxynucleotide constituting a double stranded pair (e.g. oligodeoxynucleotides 1 and 2, 3 and 4 etc. FIG. 4) were mixed and incubated at 90° C. for 2 min followed by slow cooling to room temperature. Each pair was then combined with the others in the construction and extracted with phenol/chloroform followed by ethanol precipitation. The oligodeoxynucleotide pairs were reconstituted in 30 ul containing 5 mM Tris-HCl, pH 8, 10 mM MgCl₂, 20 mM DTT, heated to 50° C. for 10 min and allowed to cool to room temperature followed by the addition of ATP to a final concentration of 0.5 mM. 800 units of T4 DNA ligase were then added and the mixture incubated at 12.5° C. for 12–16 hours.

The ligation mixture was extracted with phenol/chloroform and the DNA ethanol precipitated. The dried DNA was reconstituted in 30 ul and digested with EcoRI and PstI for 1 hour at 37° C. The mixture was extracted with phenol/chloroform and ethanol precipitated followed by separation of the various double stranded DNA segments by electrophoresis on an 8% polyacrylamide gel, according to the method of Laemmli et al, *Nature* (1970) 227:680. The DNA fragments were visualized by wet gel autoradiography and a band corresponding to approximately 100 bp in length was cut out and eluted overnight as described. The excised synthetic DNA fragment was ligated to plasmids M13-mp8 or M13-mp9 (Messing and Vieira, *supra*) similarly digested with EcoRI and PstI, and submitted to dideoxynucleotide

sequence analysis (Sanger et al. supra) to confirm the designed sequence shown in FIG. 4. This designed sequence contains the promoter (-35 and -10 regions) and operator regions of the tryptophan operon (*trp*) as well as the ribosome binding region of the tryptophan operon leader peptide. Analogous sequences to that shown in FIG. 4 have been proven to be useful in the expression of heterologous proteins in *E. coli* (Hallewell, R. A., and Emtage, S., *Gene* (1980) 9:27-47, Ikehara, M., et al. *Proc Natl Acad Sci (USA)* (1984) 81:5956-5960).

2. Construction of the Synthetic *trp* Promoter/Operator Containing Plasmid, pTrp-233

Plasmid pKK233-2 (Amann, E. and Brosius, J., supra) was digested to completion with NdeI followed by the filling in of the termini by the method of Maniatis et al, *Molecular Cloning*, Cold Spring Harbor Laboratories, 1982 at p. 394, with 5 units of *E. coli* polymerase I, Klenow fragment (Boehringer-Mannheim, Inc.) and the addition of dATP, dCTP, dGTP and TTP to 50 uM. This was incubated at 25° C. for 20 min. Following phenol/chloroform extraction and ethanol precipitation, the NdeI-digested DNA was ligated and transformed into *E. coli* (Nakamura, K. et al, *J Mol Appl Genet* (1982) 1:289-299). The resulting plasmid lacking the NdeI site was designated pKK-233-2-Nde.

Twenty nanograms of plasmid pKK-233-2-Nde was digested to completion with EcoRI and PstI followed by calf intestinal phosphatase treatment (Boehringer Mannheim) in accordance with Maniatis et al., supra at pp. 133-134. Fifty nanograms of the synthetic *trp* promoter/operator sequence obtained from M13 RF, (described above) by digesting with EcoRI and PstI, were mixed with ten nanograms of EcoRI-PstI digested pKK-233-2-Nde and ligated with T4 -DNA-ligase as described followed by transformation into *E. coli* JA221 1pp³¹/I¹acI. Transformants were screened for the presence of plasmid DNA containing the 100 bp EcoRI-PstI synthetic *trp* promoter/operator; the correct plasmid was then isolated and designated pTrp-233.

EXAMPLE 2

Construction of Plasmid pTFS11

A. Human Basic Fibroblast Growth Factor

The bovine basic FGF cDNA was used to develop hybridization probes to isolate basic FGF clones from human cDNA and genomic libraries as described in U.S. Ser. No. 869, 382, supra, Abraham, J. A. et al, *Science* (1986) supra, and Abraham, J. A. et al, *The EMBO Journal* (1986) supra, all of which are incorporated herein by reference.

There are only two amino acid differences between basic bovine FGF and human FGF, at position 123, where the bovine protein has Ser and the human protein has Thr, and at position 137, where the bovine protein has Pro and the human has Ser. These differences are the result of a single nucleotide difference in each case; therefore bovine cDNA may conveniently be modified by site directed mutagenesis as described below to encode the human protein, and,

HindIII NdeI

```

          10          20          30          40          50
AGCTTCATATG GCTGCTGGTT CTATCACTAC CCTGCCAGCT CTGCCAGAAG
          AGTATAC CGACGACCAA GATAGTGATG GGACGGTCGA GACGGTCTTC

          60          70          80          90          100
ACGGTGGTTC TGGTGCCTTC CCACCAGGTC ACTTCAAAGA CCCAAAACGT
TGCCACCAAG ACCACGGAAG GGTGGTCCAG TGAAGTTTCT GGGTTTTGCA

```

indeed, standard site-specific mutagenesis techniques were used to alter these codons. The lambda BB2 clone (ATCC. No. 40196) was digested with EcoRI and the 1.4 kb region spanning the bFGF protein-encoding portion was ligated into the EcoRI site of M13mp8 and phage carrying the insert in the correct orientation were recovered. The in vitro mutagenesis was carried out in the presence of three oligonucleotides: the "universal" primer, a 17-mer; the mutagenic 16-mer 5'-GAAATACACCAGTTGG-3'; which alters the coding sequence at codon 123, and the mutagenic 17-mer 5'-ACTTGGATCCAAAACAG-3', which alters the sequence at codon 137. The mutagenized phage was also subjected to a second round of in vitro primer-directed mutagenesis to create a HindIII site 34 bp downstream from the translation termination codon using the mutagenic 25-mer, 5'-TTTTACATGAAGCTTTATATTTTCAG-3'. The resultant mutated DNA was sequenced by dideoxynucleotide sequence analysis (Sanger et al, supra) to confirm that the desired mutagenesis had occurred, and the approximately 630 bp fragment spanning the FGF coding region was excised with HindIII and ligated into HindIII digested pUC13 to obtain the intermediate plasmid pJJ15-1.

B. Construction of Gene with Synthetic Coding Region for N-terminal End of FGF Gene

In order to lower the G+C content of the 5' end (the first 125 base pairs) of the coding region contained in pJJ15-1, a synthetic DNA fragment was constructed with the sequence shown below using the following synthetic oligonucleotides. The oligonucleotides were annealed in pairs, ligated together sequentially, and ligated into HindIII cut m13mp9. The sequence of the synthetic 125 bp insert in mp9 was confirmed by dideoxy sequencing. The NdeI to HhaI subfragment of the synthetic insert was isolated, joined to the 377 base pair, HhaI-to-HindIII DNA fragment from JJ15-1 that spans approximately the carboxyterminal three quarters of the basic FGF coding sequence, and then ligated into the NdeI and HindIII sites of the expression vector pTrp-233 to yield the plasmid pTFS11.

Oligonucleotides:

Number	Sequence
1670	5'-pAGCTTCATATGGCTGCTGGTTCTATCACTACC
1623R	5'-pCTGCCAGCTCTGCCAGAAGACGGTGGTT
1624R	5'-pCTGGTGCCTTCCCACCAGGTCACITCAA
1625R	5'-pAGACCCAAAACGTCTGTACTGCAAAAAC
1680	5'-pGGTGGTTTCTTCTGCGCA
1679	5'-pTAGAACCCAGCAGCCATATGA
1622	5'-pTCTTCTGGCAGAGCTGGCAGGGTAGTGA
1619	5'-pACCTGGTGGGAAGGCACCAGAACCACCG
1626	5'-pAGTACAGACGTTTTGGGTCTTTGAAGTG
1673	5'-pAGCTTGCAGGAAGAAACCACCGTTTTTGC

Construction of Synthetic Gene for the Amino Terminal Region of bFGF:

-continued
HhaI

110	120	130
CTGTACTGCA	AAAACGGTGG	TTTCTTCCTG CGCA
GACATGACGT	TTTTTCGGACC	AAAGAAGGAC GCGTTCGA

A plasmid map of pTSF11 is given in FIG. 5 of the accompanying drawings.

EXAMPLE 3

Preparation of Expression Vector for Mutagenized Gene Inserts

The HindIII site of the polylinker region of plasmid pUC9 was removed so as to facilitate subcloning mutated DNA into the final expression vector illustrated in FIG. 5. Approximately 5 ug of pUC9 (New England Biolabs) was digested with HindIII (20 units; New England Biolabs) according to the manufacturers instructions in 0.05 ml. The reaction was then supplemented with 0.5 mM dNTPs and the Klenow fragment of DNA Polymerase I (5 units; Boehringer Mannheim) and incubated for 30 minutes at 15° C. The reaction was then extracted twice with an equal volume of phenol/chloroform (1/1), twice with chloroform, made 0.2M NaCl, and then precipitated with two and a half volumes of ethanol. The precipitate was collected by centrifugation (15,000 g in a Microfuge at 4° C.), lyophilized, and then incubated in 0.1 ml with 1X kinase ligase buffer, 1 mM ATP, and T4 DNA ligase (20 units; New England Biolabs) for 4 hours at 12° C.

An aliquot of the reaction (0.01 ml) was then used to transfect competent MC1061 cells. The transfected bacteria were grown overnight on L agar plates supplemented with 100 ug/ml ampicillin. DNA was isolated from 6 colonies by the alkaline lysis procedure and tested for the loss of the HindIII site. A bacteria containing the plasmid, pUC9delH3-1, was isolated. The plasmid DNA was prepared and 10 ug was digested in 0.4 ml with PvuI (20 units; New England Biolabs) and EcoRI (50 units; New England Biolabs) for 2 hours according to the manufacturers instructions. The reaction was then extracted twice with an equal volume of phenol/chloroform (1/1) and twice with an equal volume of phenol and then precipitated with isopropanol. The precipitate was collected by centrifugation, washed with 70% ethanol, lyophilized, resuspended in 0.008 ml water and the ~2.07 kb PvuI-EcoRI fragment of pUC9delH3-1 (designated fragment A) containing the origin of replication was isolated by acrylamide gel electrophoresis.

Concurrently pTSF11 DNA (10 ug) was incubated with PvuI (10 units) and EcoRI (10 units) in 0.15 ml for 1 hour at 37° C. according to the manufacturers directions and collected as described above. The ~1.3 kb PvuI-EcoRI fragment of pTSF11 containing the Trp promoter/operator region, FGF coding region and the transcription termination sequences, designated fragment B of pTSF11, was isolated by polyacrylamide gel electrophoresis and ligated to the ~2.07 kb PvuI-EcoRI fragment A of pUC9delH3-1, and used to transfect competent *E. coli* HB101 cells. The bacteria were grown overnight on L agar plates supplemented with 100 ug/ml ampicillin. Plasmid DNA from one recombinant, pUC9delH3-pTSF-3, was isolated and shown to contain the expected restriction map (HindIII cuts the plasmid only once; the sizes of HindIII-EcoRI, HindIII-PvuI and HindIII-PstI fragments are approximately 560 and 2900, 800 and 2700, and 560 and 2900 bp respectively. DNA from the plasmid pUC9delH3-pTSF-3 was isolated and 200 ug incu-

bated in 1.0 ml with 100 units of HindIII, 100 units of EcoRI, 5 mM spermidine for 4 hours at 37° C. according to the manufacturers instructions. The reaction was butanol extracted to reduce the volume to 0.7 ml and then extracted with phenol/chloroform and chloroform as described above. The DNA was collected by ethanol precipitation and the ~2.9 kb HindIII-EcoRI fragment containing the ampicillin resistance gene, the origin of replication and the two transcription stop signals, designated fragment C of pUC9delH3-pTSF-3, was isolated by two sequential runs on polyacrylamide gels. This vector fragment serves as the preferred vector for expressing any of the DNA which has been altered by in vitro mutagenesis. The construction of this vector is illustrated in FIG. 5.

Plasmid pUC9-pTSF11, a vector closely resembling plasmid pUC9delH3-pTSF-3 but containing an intact HindIII site in the multiple site polylinker region, can also be used as a preferred vector for expressing both recombinantly produced FGF (all forms) and any of the analogs of the present invention. This vector was constructed by individually digesting plasmids pUC9 and pTSF11 with PvuI and EcoRI, isolating the ~2.07 kb PvuI-EcoRI vector fragment from pUC9 and the ~1.3 kb PvuI-EcoRI fragment containing the trp promoter/operator region, FGF coding region, and the transcription termination sequences from pTSF11, and ligating the two isolated fragments. This vector can then be used to express the FGF analog gene sequences as taught with pUC9pTSF11 by inserting the HindIII-EcoRI DNA cassettes into the appropriately digested vector and transforming *E. coli* bacterium.

EXAMPLE 4

Generalized Procedure for Production of FGF Mutants

The following protocol can be used to construct all of the DNA sequences encoding the FGF analogs described herein. Plasmid FGFt7910 was constructed by ligating the ~603 bp EcoRI-HindIII DNA fragment of pTSF11 (comprising the Trp promoter region and the DNA encoding human bFGF) into the EcoRI-HindIII sites of an M13mp9 vector. Once the single-stranded DNA of FGFt7910 was isolated, in vitro mutagenesis, as described by Zoller and Smith, supra, may be performed utilizing one or more of the synthetic oligonucleotides designated in any of the tables herein.

The conditions for site specific mutagenesis can be generalized as follows. One ug of the single stranded DNA is hybridized with 5 ng of the phosphorylated mutagenic oligonucleotide(s) (23 mer to 25 mer encoding the appropriate mutation) and 1 ng of the M13 universal sequencing primer (17 mer purchased from P.L. Biochemicals) for 5 to 15 minutes at 55° C. in 0.01 ml solution of 10 mM Tris-HCl, pH 7.5, and 10 mM MgCl₂. The reaction is cooled to room temperature and then added to 0.01 ml of 0.12 mM dXTPs, 5 units Klenow fragment of DNA polymerase I (Boehringer Mannheim), 20 units of T4 DNA ligase (New England Biolabs), and incubated for 4-6 hours at 15° C. An aliquot (0.002 ml) of the reaction is then added to competent JM101 bacteria and plated overnight on L agar plates at 37° C. The DNA of the resulting M13 clones is transferred to each of

two nitrocellulose filters, baked under vacuum at 80° C. for 2 hours and then incubated for 2 hours at 42° C. in pre-hybridization solution: 6×SSC. (1×SSC is 150 mM NaCl, 15 mM sodium citrate, pH 7.0), 0.1% sodium dodecyl sulfate, 2×Denhardt's (0.05% ficoll, 0.05% polyvinylpyrrolidone, 0.05% bovine serum albumin) solution) and 0.4 mg/ml of denatured salmon sperm DNA. The filters are then incubated for 3 hours at 42° C. with fresh pre-hybridization solution containing the appropriate mutagenic oligonucleotide which has been 5'-end labeled with [λ -³²P]-ATP and T4 polynucleotide kinase. The filters are then washed twice with 4×SSC. at room temperature for 15 minutes, once for 15 minutes at 65° C., once at room temperature in TMACL solution (3M tetramethylammonium chloride, 50 mM Tris-HCl, pH 8.0, 2 mM EDTA, 0.1% SDS) and once at 65° C. in TMACL solution and then used to expose X-ray film overnight at room temperature. Clones corresponding to dark positives on the X-ray film are then picked from the original plate, the DNA is isolated and then analyzed for the mutated sequence by dideoxy sequencing. If two oligonucleotides are being used to produce a double mutant then one filter is screened with one oligonucleotide and the other filter is screened with the second oligonucleotide. Double mutants will have a positive signal with both oligonucleotides.

The DNA replicative form of the mutated M13 clone is then prepared, digested with EcoRI and HindIII, and the DNA fragment encoding the mutated FGF is isolated by agarose gel electrophoresis. The DNA fragment is then ligated to Fragment C. of pUC9delH3-pTSF-3 (described in Example 3 and illustrated in FIG. 6), transfected into competent HB101 cells, and grown overnight on L agar plates supplemented with 100 ug/ml ampicillin. Colonies are selected, grown in L broth supplemented with 100 ug/ml ampicillin and then the plasmid DNA is isolated from the bacteria. The DNA is then used to transform competent *E. coli* B cells (Luria and Delbrck, Arch Biochem (1942) 1:111).

EXAMPLE 5

Preparation of Basic FGF Analog bFGF-C34/101S

In this example, cysteine residues at positions 34 and 101 of the human basic FGF protein were changed to serine residues thereby producing a double mutation. Approximately 2 micrograms each of the mutagenic 23-mer 5'-ACGTCTGTACTCCAAAAACGGTG-3' (#2222); which alters the sequence at codon 34, and the mutagenic 23-mer 5'-TACAGACGAGTCTTTCTTTTTTG-3' (#2323); which alters the sequence at codon 101 were incubated in 50 ul of 1×kinase/ligase buffer (7 mM Tris-HCl pH 7.6, 10 mM MgCl₂, 5 mM dithiothreitol) with 1 mM ATP and 5 units T4 polynucleotide kinase for 30 minutes at 37° C. The phosphorylated oligonucleotides were diluted two-fold into 1 mM Tris-HCl, pH 8.0 and 1 mM EDTA.

One ug of the single stranded M13 template FGFt 7910 was incubated with 20 ng each of the phosphorylated oligonucleotides 2222 and 2323 and 1 ng of the universal M13 sequencing primer (New England Biolabs) in 10 ul of 10 mM Tris-HCl pH 7.5 and 10 mM MgCl₂ for 20 minutes at 55° C. and then at room temperature for 10 minutes. The reaction was then supplemented with 0.5 mM dXTPs, 5 units of the Klenow fragment of DNA polymerase I, 1 mM ATP, and 20 units T4 DNA ligase and incubated at 15° C. for 5 hours. 2 ul of the reaction was then used to transform competent JM101 bacteria. The transformed cells were plated overnight at 37° C, and the resulting M13 DNA was

transferred to nitrocellulose filters as described above. One ug of each of the oligonucleotides (#2222 and #2323) was phosphorylated as described above except 1 mCi of [λ -³²P]-ATP (New England Nuclear #NEG035C, approximately 5 mCi/nmole) was substituted for cold ATP. The radioactive probes were then added separately to the duplicate filters and processed as described above. M13 clones corresponding to positive signals from the resulting autoradiographs were isolated and the single stranded M13 DNA prepared by the method of Sanger et al, supra. The resulting M13 DNA template (#8725) was analyzed by dideoxy sequencing and shown to contain the expected changes.

The double stranded replicative form (RF) of the M13 template #8725 was isolated by the method of Birnboim and Doly, *Nucl Acids Res* (1979) 7:1513-1519. In this procedure, fifty ul of M13 phage #8725, isolated from infected JM101, were used to inoculate 50 ml of a JM11 culture (saturated culture, 20×diluted into J broth) which were then grown for 6 hours at 37° C. The bacteria were harvested by centrifugation and the DNA isolated as described by Birnboim and Doly, supra. Approximately 5 ug of the RF DNA was cut in 0.4 ml of 1×HindIII buffer as described by the manufacturer with 40 units each of HindIII and EcoRI for 2 hours at 37° C. The reaction was then extracted twice with equal volumes of phenol/chloroform (1/1) and twice with chloroform and then, ethanol precipitated. The resulting DNA was collected by centrifugation, washed with 70% ethanol, lyophilized, and resuspended in 20 ul of 1 mM Tris-HCl, pH 8.0, and 1 mM EDTA. The resulting EcoRI-HindIII fragment was isolated by agarose gel electrophoresis using GENECLEAN (BI0101 Inc.; La Jolla, Calif.) according to the manufacturer's instructions. Approximately 50 ng of the EcoRI-HindIII insert was ligated to the EcoRI-HindIII vector fragment C of pUC9delH3-pTSF-3 and used to transform competent MC1061 cells. The bacteria were processed as described in Example 7 in order to purify the analog and then the purified analog was tested for its ability to stimulate adrenal cortex endothelial (ACE) cells as described in Example 8.

Other mutants containing cysteine-to-serine substitutions have been constructed and expressed in bacteria. These constructions contain from one to four Cys→Ser substitutions. All of these substitutions result in the recovery of an FGF protein with varying levels of activity. The specific constructions are listed below in Table 1. Each of these FGF analog proteins has been isolated using a heparin affinity column.

TABLE 1

bFGF Analog ⁺	Oligonucleotide [#]	Number*
1) bFGF-C78S	5'-pCAAAGGAGTGTCTGCAAACCGTT	2217
2) bFGF-C96S	5'-pAGCTTCTAAATCTGTTACAGACG	2218
3) bFGF-C78/96S		2218/2217
4) bFGF-C34/78/96/101S		2217/2218/ 2222/2323
	5'-pACGTCTGTACTCCAAAAACGGTG	2222
	5'-pTACAGACGAGTCTTTCTTTTTTG	2323
5) bFGF-C34/78/96S		2222/2218/ 2217
6) bFGF-		2217/2218/

TABLE 1-continued

C78/96/ 101S	2323	
7) bFGF- C34/78/ 101S	2222/2217/ 2323	5
8) bFGF- C34/78S	2222/2217	
9) bFGF- C34/101S	2222/2323	10

⁺Analogues of bFGF are defined as: bFGF-XYZ where X is the amino acid in the native human bFGF sequence that is being mutated, Y is the position of amino acid X, and Z is the amino acid residue that is being substituted for X at position Y. Multiple mutations are indicated. Mutations that involve the deletion of a region of the native bFGF protein are indicated with parenthesis (X-Z) with the deleted region defined by the amino acids included in residues X to Z.

[#]Oligonucleotide used for in vitro mutagenesis.

^{*}Number of the oligonucleotide used for the mutagenesis.

NOTE: This legend is applicable to all of the following tables and analog descriptions. The one-letter code depicting specific amino acids is as follows:

Amino Acid	Three-Letter abbreviation	One-Letter symbol
Alanine	Ala	A
Arginine	Arg	R
Asparagine	Asn	N
Aspartic acid	Asp	D
Asparagine or aspartic acid	Asx	B
Cysteine	Cys	C
Glutamine	Gln	Q
Glutamic acid	Glu	E
Glutamine or glutamic acid	Glx	Z
Glycine	Gly	G
Histidine	His	H
Isoleucine	Ile	I
Leucine	Leu	L
Lysine	Lys	K
Methionine	Met	M
Phenylalanine	Phe	F
Proline	Pro	P
Serine	Ser	S
Threonine	Thr	T
Tryptophan	Trp	W
Tyrosine	Tyr	Y
Valine	Val	V

The bFGF analogs described below in Table 2 were tested for their ability to stimulate bovine adrenal cortex endothelial cell proliferation. As indicated below, the double mutant bFGF-C78/96S has enhanced activity compared to wild type bFGF. Any alteration of cysteines at the conserved positions 34 and 101, that is, positions conserved throughout the family of FGF-related molecules including human and bovine aFGF, bovine and xenopus bFGF, murine int-2, human hst and human KS3, significantly decreased the activity of the resulting analog.

TABLE 2

Activity of bFGF and Various Cysteine Analogs in the ACE Cell Proliferation Assay	
FGF ANALOG	% ACTIVITY
WILD TYPE bFGF	100
C78S*	53
C96S*	95
C78/96S*	159

TABLE 2-continued

Activity of bFGF and Various Cysteine Analogs in the ACE Cell Proliferation Assay	
FGF ANALOG	% ACTIVITY
C34/101S	2
C34/78/101S ⁺	2
C78/96/101S ⁺	23
C34/78/96/101S ⁺	7

^{*}Average of 2 independent assays.

⁺Average of 3 independent assays.

EXAMPLE 6

Heparin Binding Assay

The interaction of bFGF analogs with heparin is characterized by the ionic strength (NaCl concentration) of a Tris-HCl buffered solution required to elute the protein from heparin-Sepharose resin. This analysis determines the NaCl concentration required to remove the bFGF analogs that were bound to the heparin-Sepharose resin.

A heparin-5PW column was prepared by Bio-Rad Laboratories (Richmond, Calif.) by introducing heparin onto Bio-Gel TSK-50 resin. The column (75×7.5 mm I.D.; 4–6 mg/ml of heparin) was used with two Beckman model 110B Solvent Delivery Modules; a Beckman model #421 Controller, and a Kratos Spectraflow absorbance detector model #757. Samples were loaded onto the column in 0.5M NaCl, 20 mM Tris HCl, pH 7.5 and then eluted with a gradient of 0.6–3.0M. Protein was monitored by absorbance at 214 nm. The conductivity of various samples was tested and compared to buffered NaCl standards to determine NaCl concentrations along the gradient. Data from the absorbance detector was collected and analyzed using Access*Chrom (Nelson Analytical, Inc., Cupertino).

The cysteine-substituted FGF analogs of Example 5 were analyzed by heparin HPLC. The analog bFGF-C78/96S elutes as a single species with (FIG. 7a) and without (FIG. 7b) dithiothreitol treatment. This is in contrast to wild type bFGF which elutes as a single species with dithiothreitol treatment (FIG. 7c) but as a heterogeneous species in the absence of a reducing agent (FIG. 7d). In addition, the double mutant does not exhibit any heterogeneity when analyzed by reverse phase-HPLC. or by size exclusion chromatography as is the case for wild type bFGF.

The single cysteine-substituted mutants, C78S and C96S, when analyzed as above, will also reduce the heterogeneity of the resulting product as compared to wild type bFGF.

The same supernatant fraction is tested for mitogenic activity using the endothelial cell proliferation assay or the Balb/c 3T3 thymidine uptake assay (Hauschka et al *J Biol Chem* (1986) 261:12665–12674) described below.

EXAMPLE 7

Isolation of Recombinant Human bFGF and Analogs of bFGF, and Characterization of the In Vitro Activity

This will describe the procedure for isolating approximately 100 ug of recombinant bFGF from bacteria. The method can be scaled up to obtain larger quantities. Bacteria containing the appropriate plasmid are grown overnight in L broth supplemented with 100 ug/ml ampicillin. 0.2 ml of the culture are inoculated into 100 ml of 1×M9 salts (Maniatis

et al, supra) 0.4% glucose, 2 ug/ml thiamine, 200 ug/ml MgSO₄·7H₂O, 0.5% casamino acids, 100 uM CaCl₂ and 100 ug/ml ampicillin and grown on a shaker at 37° C. The culture is supplemented with 20 ug/ml indole acrylic acid upon reaching an optical density of 0.1 at a wavelength of 550 nm. The bacteria are harvested upon reaching an optical density of 1.0 by centrifugation (5000 rpm, 4° C., 15 minutes), quick frozen in a dry ice/ethanol bath and then stored at -80° C. The bacterial pellet is resuspended in 10 ml of 0.02M Tris-HCl, pH 7.5, 0.6M NaCl, 1 mM PMSF, 80 ng/ml aprotinin, and 10 ug lysozyme and incubated at 40° C. for 15 minutes. The mixture is then sonicated 5 times at a setting of 3 using a Sonicator Cell Disruptor (Heat Systems). The reaction is then incubated with DNaseI (100 units) and RNaseA (100 units) for 15 minutes at 4° C. and then centrifuged for 15 minutes at 4° C. at 10,000 rpm. The supernatant is then loaded onto an 8.0 ml heparin-Sepharose column (Pharmacia) which has been prewashed with 3.0M NaCl, 10 mM Tris-HCl, pH 7.5 and equilibrated with 0.6M NaCl, and 0.02M Tris-HCl, pH 7.5. The column is washed with 0.6M NaCl until no detectable protein, as judged by absorbance at 280 nm, is eluting off the column. The column is then washed with 1.0M NaCl and the bound material eluted off with 2.0M NaCl, 20 mM Tris-HCl, pH 7.5. This purification scheme can be performed in buffers in the presence or absence of 5 mM dithiothreitol.

EXAMPLE 8

Mitogenic Assays

FGF analogs are tested for either agonist or antagonist activity with respect to wild type FGF in an adrenal cortex capillary endothelial (ACE) cell proliferation assay as described by Gospodarowicz et al (*J Cell Physiol* (1985) 122:323-332). Individual analogs were tested as follows. Approximately 1×10⁴ cells were plated in 2 ml of DME 16 supplemented with 10% calf serum, 50 units/well of penicillin and 50 units/well of streptomycin in a Falcon 6-well plate. Appropriate dilutions (1 pg/ml to 1 ug/ml final concentration) of each sample, as well as wild type (bovine pituitary basic) FGF were added in 10 ul volumes to the cells. As a negative control, 6 wells without added FGF samples were run simultaneously. The plates were incubated at 37° C. for 48 hours and cell samples were re-added to the appropriate well and incubated for an additional 48 hours at 37° C. Cells were then trypsinized, collected and counted in a Coulter counter.

Balb/c 3T3 cells, obtained from ATCC, were used to test for the ability of bFGF preparations to stimulate DNA synthesis essentially by the method described by Hauschka et al (1986) supra. Cells were seeded onto 96 well plates at a density of about 20,000/well in 0.2 ml Dulbecco's modified Eagle's medium (DME; GIBCO) containing 4.5 g/liter glucose, 2.2 g/liter NaHCO₃, 50 units/ml penicillin, 50 ug/ml streptomycin, and 10% calf serum (HYCLONE) and allowed to grow to confluency (2-3 days) in a 5% CO₂, 95% Air incubator at 37° C. Cultures were switched to serum free medium containing 0.01% bovine serum albumin, after which 0.01 ml of appropriate dilutions of test substance were added. Cultures were incubated at 37° C. for an additional 16 hr, after which the medium was changed to serum free medium containing 0.01% bovine serum albumin plus 50 uCi/ml of [³H] thymidine. Plates were then incubated for 2 hours after which TCA precipitable counts were determined as follows. The 96 well plate was placed on ice and the medium carefully removed, washed twice with cold PBS, followed by incubation with 10% trichloroacetic acid for 20

min at 4° C. Remaining radioactivity was solubilized in 0.1N NaOH, and counted.

These assays were used to test the FGF analogs for their respective agonist or antagonist activity toward wild type basic FGF. The ability of FGF analogs to serve as antagonists to basic FGF is characterized by mixing appropriate quantities, such as 1-1000 ng, of the particular analog with 1 ng of basic FGF and testing the mixture in the above-described assays.

EXAMPLE 9

Construction of Receptor Binding FGF Analogs

A number of oligonucleotides were constructed and tested in an FGF receptor competitive binding assay. The specific mutants are provided below and include single amino acid substitutions, double amino acid substitutions and deletion mutations.

Analog	Oligonucleotide	Number
bFGF-K35S	5'-pGTCTGTACTGCTCAAACGGTGGTT	2553
bFGF-R42L	5'-pTTTCTTCTGCTCATCCACCCCG	2327
bFGF-D46A	5'-pCATCCACCCCGCCGCGGAGTGG	2221
bFGF-R48L	5'-pCCCCGACGGCCTAGTGGACGGGG	2454
bFGF-R48A	5'-pACCCCGACGGCGCAGTGGACGGGG	2555
bFGF-D50A	5'-pCGGCCGGAGTGGCCGGGTCCGCG	2224
bFGF-V52K	5'-pGAGTGGACGGGAAACGCGAGAAGAG	2491
bFGF-R53L	5'-pGGACGGGGTCTCGAGAAGAGCG	2220
bFGF-K55M	5'-pGGTCCGCGAGATGAGCGACCCAC	2223
bFGF-K55I	5'-PGGTCCGCGAGATAAGCGACCCACA	2567
bFGF-D57A	5'-pCGAGAAGAGCGCCCCACACATCA	2225
bFGF-H59N	5'-pGAGCGACCCAAACATCAAACACTAC	2383
bFGF-R90T	5'-pAGAAGATGGAACITTTACTAGCTTC	3088
bFGF-D99A	5'-pATGTGTACAGCAGAGTGTITCT	2381
bFGF-E100A	5'-pGTTACAGACGCCTGTTTCTTTTTTG	2549
bFGF-E100S	5'-pGTGTTACAGACAGTGTITCTTTTTT	2380
bFGF-E105S	5'-pGTTTCTTTTTTTCACGATTGGAGT	2556
bFGF-R106L	5'-pCTTTTTTGAACATTTGGAGTCTA	2494
bFGF-E108A	5'-pTGAACGATTGGCATCTAATAACTA	2554
bFGF-Y112A	5'-pAGTCTAATAACGCAAATACCTACCG	2450
bFGF-N113S	5'-pCTAATAACTACAGTACTTACCGG	2452
bFGF-R116T	5'-pCAATACTTACACTTCAAGGAAATA	3091
bFGF-R118L	5'-pCAATACTTACCTGTCAAGGAAAT	2483
bFGF-K119S	5'-pACCGGTCAAGGTCTTACACCAGTTG	2548
bFGF-(41-43)	5'-pGGTGGTITCTTCCACCCCGACGGC	2336
bFGF-(49-51)	5'-pCCCGACGGCCGAGTCCGCGAGAAG	2335
bFGF-(62-64)	5'-pCCACACATCAAACAAGCAGAAGAG	2334
bFGF-(83-85)	5'-pGCAAACCGTTACAAAGAAGATGGA	2333
bFGF-(105-107)	5'-pTGTTTCTTTTTTGAGTCTAATAAC	2332
bFGF-(112-114)	5'-pGAGTCTAATAACTACCGGTCAAGG	2337

The FGF analogs were produced as described in Example 7 using an appropriate expression vector, such as plasmid pUC9delH3-pTSF11-3 or pUC9pTSF11, and isolated by heparin-Sepharose chromatography.

A competitive binding assay was established to determine the relative affinity of FGF analogs compared to that of recombinant basic FGF for the FGF receptor. Analogs having high affinity for the FGF receptor and reduced mitogenic activity are designated potential FGF antagonists.

The assay involved binding saturating concentrations of [¹²⁵I]-basic recombinant FGF (10 ng/ml) to Balb/c 3T3 cells in the presence of various concentrations of unlabeled FGF or analogs. The binding was conducted at 4° C. for 3-4 hours to establish equilibrium. The cells were then washed 12× with a 0.1% gelatin, 2N NaCl balanced salt solution containing 50 mM Hepes to maintain the pH at 7.5. Cells were solubilized in 1N NaOH and cell-associated radioactivity measured. Following this procedure, the non-specific binding was kept at or below 5%. The affinity of an analog

for the FGF receptor was determined relative to that of bovine pituitary FGF by taking the ratio of the concentration of analog that inhibits specific binding by 50% over the concentration of FGF that inhibits specific binding by 50%. A ratio of less than 1 indicated that the analog has a higher affinity for the FGF receptor than FGF and a ratio of greater than 1 indicated that the analog has a lower affinity for the FGF receptor than FGF.

A number of analogs that had reduced mitogenic activity in the ACE assay had equal or higher affinity for the FGF receptor compared to bovine pituitary FGF. Those mutants that had less than 5% of wild type activity in the ACE assay but had equal or higher affinity for the FGF receptor include: R31S, K35S, D46A, R48L, D50A, V52K, R53L, R90T, E100S, E100A, R106L, R116T, R118L, K119S. These compounds may be useful as antagonists.

These analogs were also tested for mitogenic ability using either one of the previously described assays. The results are provided below in Table 3.

TABLE 3

Activities of FGF Analogs				
Analog	3T3 Mito. Heparin -/+	ACE Mito. Heparin -/+	FGF-Rc Comp. Heparin -/+	Heparin Elution
hFGF (b) (EC ₅₀)	100%/100% (630 pg/ml)	100%/100% (160 pg/ml)	1.0/1.0 (10 ng/ml)	1.58 M
hFGF (b) 25-155	50%/100%			1.62 M
hFGF (a) bba*	5%/25% <0.2%/100%	0.01%/10%	0.5/ 0.3/	1.23 M 1.43 M
K27M			0.3/0.2	
D28K		8.8%/	1.0/1.3	
KKR#	8.5%/8.5%	0.11%/4.2%		~1.5 M
R31S	100%/100%	0.45%/17.8%	1.0/0.8	
K35S		2.6%/55.5%	0.1/0.05	1.23 M
D46A		2.8%/	0.5/	
R48L		1.7%/	0.16/2.0	
R48A		8.9%/84.2%	0.32/0.3	
D50A		0.37%/	2.5/2.5	
V52K		0.72%/>20%	0.5/0.4	
R53L		0.5%/	0.22/0.25	1.56 M
K55I	1%/8%	<0.1%/4.2%	2.0/1.6	
K55M		90%/	0.4/0.4	
D57A		0.27%		
H59N	315%/	42%/120%	0.4/0.4	
R90T	77%/216%	2.68%/95.4%	0.6/0.5	1.58 M
hFGF (b) (EC ₅₀)	100%/100% (630 pg/ml)	100%/100% (160 pg/ml)	1.0/1.0 (10 ng/ml)	1.58 M
K95T	100%/170%	44.9%/		1.58 M
D99A	150%/280%	19.2%/	0.04/0.03	1.53 M
E100S		0.03%/	1.1/0.5	
E100A		0.75%/	0.8/1.0	
E105S	12.6%/100%	0.3	50/40	
R106L	20%/200%	0.27%/	0.3/0.13	1.55 M
R106T	210%/250%	9.1%/62.5%		1.58 M
E108A	180%/	30%/	2.5/2.5	
Y112A	0.2%/0.2%	0.01%/4.3%	100/100	1.49 M
N113S	160%/	7.7%/	1.0/1.0	
R116T	50%/300%	0.64%/	0.2/	1.53 M
R118L		1.8%/	0.5/0.4	
K119S	48%/250%	0.6%/41%	0.35/0.25	1.58 M
K128S	210%/	68%/	0.3/0.25	1.38 M
K128E	200%/140%	13%/		1.04 M
R129T	87%/190%	11.9%/	1.3/0.5	1.45 M
R129L		1.3%/		
KR128, 129ST	185%/	8.4%/	2.0/0.8	1.14 M
K134S				0.6 M
K138S				1.0 M
C78S		53%/123%	0.3/0.25	1.57 M
C96S		95%/96%	0.3/0.8	1.58 M
C78, 96S	118%/160%	159%/151%	0.2/0.1	1.58 M

TABLE 3-continued

Activities of FGF Analogs				
Analog	3T3 Mito. Heparin -/+	ACE Mito. Heparin -/+	FGF-Rc Comp. Heparin -/+	Heparin Elution
C34, 101S		2%/		
C34, 78, 101S	65%/144%	2%/72%	0.45/0.5	1.50 M
C78, 96, 101S		23%/125%		
C34, 78, 96, 101S		7%/119%	1.0/0.5	1.51 M

*bba is hFGF(b) with hFGF(a) substitutions for aa 95-155.

#KKR has neutral substitutions (m, S or T) for K27, K30 & R31.

@C-4-S has 4 S substitutions for each of the 4 Cs.

Legend:

The data indicate the activities of the various FGF analogs relative to the activity of wild type FGF. The first row indicates the actual ED₅₀ value for wild type FGF in each assay (in parentheses). The ED₅₀ is the concentration at which the analog elicits a half-maximal response in the assay and is therefore a measure of potency. For analogs exhibiting activity lower than wild type activity, higher concentrations are required to elicit activity equivalent to wild type FGF. Therefore, the ED₅₀ values of such analogs are higher than wild type (more analog required to elicit half-maximal stimulation). For analogs exhibiting activity higher than wild type activity, less analog is required to elicit activity equivalent to wild type activity. Therefore, the ED₅₀ values of such analogs are lower than wild type (less analog required to elicit half-maximal stimulation). The ED₅₀ values for the various analogs in each assay are indicated as a percentage of wild type activity (the ED₅₀ of wild type FGF divided by the ED₅₀ of the analog times 100%). Therefore, analogs exhibiting values greater than 100% appear to have activity greater than wild type FGF in the assay, while analogs exhibiting values less than 100% appear to have activity less than wild type FGF in the assay.

"3T3 Mito./Heparin -/+":

The data reflect the activity observed in the 3T3/Balb/c cell mitogenic assay (thymidine uptake) relative to the activity observed for wild type FGF in this assay. Activity values obtained in the absence or presence of 1 ug/ml heparin are indicated to the left and right of the slash respectively.

"ACE Mito./Heparin -/+":

The data reflect the activity observed in the Adrenal Cortical Endothelial cell proliferation assay relative to the activity observed for wild type FGF in this assay. Activity values obtained in the absence or presence of 1 ug/ml heparin are indicated to the left and right of the slash respectively.

"FGF-Rc Comp./Heparin -/+":

The data reflect the relative ability of the analog to bind the FGF receptor as measured in the competitive binding assay. The values are the ratio of the ED₅₀ of the analog to the ED₅₀ of wild type FGF in this assay. Therefore, analogs which exhibit values less than 1.0 appear to have an affinity for the receptor which is greater than that of wild type FGF. Analog values greater than 1.0 appear to have an affinity for the receptor which is less than that of wild type FGF. Activity values obtained in the absence or presence of 1 ug/ml heparin are indicated to the left and right of the slash respectively.

Analog values which exhibit near wild type receptor activity and exhibit low relative activity in the mitogenic assays are potential antagonists.

"Heparin/Elution":

The data indicate the approximate salt (NaCl) concentration at which the analog elutes from a heparin-TSK column during high performance liquid chromatography. Analog values less than 1.58 M appear to have reduced heparin binding as judged by this procedure. Analog values app. equal to 1.58 (1.53 to 1.63) appear to have affinity for heparin which is insignificantly changed from that of wild type bFGF as judged by this procedure.

EXAMPLE 10

Reduced Heparin Binding FGF Analogs

FGF analogs were constructed wherein mutagenesis was targeted for the region of the basic FGF molecule which may be involved in binding heparin and heparin-like compounds. The analogs, with the specific oligonucleotide sequences which correspond to the amino acid to be changed, are listed below.

Analog	Oligonucleotide	Number
bFGF-K27M	5'-pAGGTCACITTCATGGACCCAAAACG	2487
bFGF-K30A	5'-pTCAAAGACCCAGCACGTCTGTACT	2566
bFGF-R31S	5'-pAAGACCCAAAATCTCTGTACTGCA	2568
bFGF-D28K	5'-pGTCACTTCAAAAAGCCAAAACGTCT	2480
bFGF-R118L	5'-pCAATACTTACCTGTCAAGGAAAT	2483
bFGF-K35S	5'-pGTCTGTACTGCTCAAACGGTGGTT	2553
bFGF-K128S	5'-pATGTGGCACTGTCTCGAACTGGGCA	2545
bFGF-K128E	5'-pATGTGGCACTGGAGCGAACTGGGCA	3332
bFGF-R129T	5'-pGGCACTGAAAACACTACTGGGCAGT	3087
bFGF-K128E/ R129T		
bFGF-K134S	5'-pCTGGGCAGTATTCTCTTGGATCCAA	3212
bFGF-K138S	5'-pAACTTGGATCCTCTACAGGACCTGG	3215

These gene sequences were inserted into an appropriate expression vector as taught previously and the resulting protein tested for reduced heparin binding activity. The results provided in Table 3 indicate that the region of bFGF encompassing residues 128–138 is a targeted heparin binding region as amino acid substitutions in this region has led to a decrease in heparin binding as measured by elution from the heparin-SPW resin.

EXAMPLE 11

Preparation of an FGF Antagonist by N-terminal Deletion

The blunted NdeI-HindIII FGF fragment from pUC9delH3-PTSF-3 was subcloned into the SmaI-HindIII site of M13mp18. An oligo was used to introduce a new NdeI site in the FGF molecule at amino acid 25 using in vitro mutagenesis. The new NdeI site serves as both a new restriction site for subcloning the FGF fragment and also as a new translational start site for the shortened form of FGF. The mutagenic oligo used has the sequence:

5'-TTG GGT CTT TGA AGT GCA TAT GTG GGA AGG CAC CAG

The shortened FGF was subcloned into pTSF-delbeta-gal for expression as an NdeI-HindIII fragment and the resulting plasmid designated bFGF(25–155). Protein sequence confirmed that the N-terminus of the protein is histidine. pTSF-delbeta-gal was constructed by digesting pTSF11 with PvuII and EcoRI, thereby deleting approximately one-half of the beta-gal promoter operator.

The N-terminal deletion analog, bFGF(25–155) was purified by heparin-Sepharose chromatography as above.

This analog exhibits agonist activity in the 3T3 mitogenic assay with an ED₅₀ similar to that of bFGF. Although stimulation in the 3T3 assay peaks at approximately 1 ng/ml for both wild type and bFGF(25–155) the level of stimulation for the analog determined in the absence of heparin is not as great as observed for wild type bFGF. Thus, bFGF(25–155) displays characteristics of a partial agonist. In addition, concentrations of the bFGF(25–155) analog greater than 1 ng/ml result in apparent inhibition of activity; whereas for wild type bFGF, activity in the 3T3 assay peaks at approximately 1 ng/ml and is not significantly reduced even at 1 ug/ml, the activity of bFGF(25–155) at 10 ng/ml is approximately 15% that for wild type bFGF and at 100 ng/ml, bFGF(25–155) essentially lacks activity.

The FGF analog bFGF(25–155) is an FGF antagonist as determined by its ability, at concentrations of 1 ng/ml or greater, to inhibit the activity of wild type FGF in the

absence of heparin. The activity elicited by 1 ng/ml wild type bFGF is reduced approximately 50% in the presence of 1 ng/ml bFGF(25–155), reduced approximately 75% in the presence of 10 ng/ml bFGF(25–155), and reduced by more than 95% in the presence of 100 ng/ml bFGF(25–155). That this inhibition is competitive is demonstrated by the shift in the ED₅₀ for wild type bFGF observed in the presence of bFGF(25–155). The ED₅₀ for wild type bFGF is less than 1 ng/ml in the absence of the analog bFGF(25–155), approximately 10 ng/ml in the presence of 10 ng/ml bFGF(25–155), and approximately 100 ng/ml in the presence of 100 ng/ml bFGF(25–155). These data suggest that bFGF(25–155) binds the FGF receptor, probably with an affinity similar to that of wild type FGF, but bFGF(25–155) exhibits altered (reduced) activity. Thus, in the absence of heparin, the FGF analog bFGF(25–155) is a competitive inhibitor of wild type FGF and is therefore an antagonist.

While the FGF analog bFGF(25–155) has demonstrated FGF antagonist activity, it retains partial agonist activity. In addition, the agonist activity of this analog is enhanced, and the antagonist activity inhibited, by the presence of heparin. Therefore it is desirable to make additional alterations in the sequence to further reduce the activity of the analog without significantly reducing its affinity for the FGF receptor. The reason for the reduced activity of bFGF(25–155) is not known, however, it is presumed that the integrity of the N-terminal segment of wild type FGF is necessary for full activity. Therefore it is possible that further deletions and/or amino acid substitutions in the N-terminal region exclusive of the receptor binding domain will further diminish activity without reducing receptor binding. For example, deletions in the region of amino acids 25 to 33 may accomplish this end. Another approach would be to delete or alter amino acids in other, non-receptor binding domains such as amino acids 78–98 or 130–155. Finally, since the antagonist activity this analog is inhibited by heparin, introducing substitutions which reduce heparin binding may reduce agonist activity

and increase relative antagonist activity. These approaches may be used in combination and are considered within the scope of the present invention.

EXAMPLE 12

Construction of Expression Vectors and Stable Expression of FGF Analogs in Mammalian Cells

The DNA sequences encoding FGF are most conveniently used to produce the recombinant proteins in a variety of hosts, as set forth in ¶C. 1 above. However, expression in mammalian systems is an alternative to bacterial expression as the mammalian host is capable of post translational processing analogous to that experienced by the natively produced protein.

To construct the vectors, the cloned FGF-encoding analog is excised with EcoRI and HindIII, provided with EcoRI or other appropriate linkers if necessary, and then inserted into an appropriate host vector such as pHS1 or its derivatives as described below.

Construction of Host Vectors

pHS1

The plasmid pHS1 is suitable for expression of inserted DNA in mammalian hosts. It contains 840 bp of the hMT-II sequence from p84H (Karin, M., et al, *Nature* (1982) 299:297–802) which spans from the HindIII site at position

-765 of the hMT-II gene to the BamHI cleavage site at base +70. To construct pHS1, plasmid p84H was digested to completion with BamHI, treated with exonuclease BAL-31 to remove terminal nucleotides, and then digested with HindIII. The desired 840 bp fragment was ligated into pUC8 (Vieira, J., et al, *Gene* (1982) 19: 259-268) which had been opened with HindIII and HincII digestion. The ligation mixture was used to transform *E. coli* HB101 to Amp^r, and one candidate plasmid, designated pHS1, was isolated and sequenced by dideoxy sequencing.

pHS1 contains the hMT-II control sequences upstream of a polylinker containing convenient restriction sites.

The workable host plasmid pHS1 can be further modified to contain additional control elements besides the metallothionein promoter. In particular, the enhancer elements of viral systems, such as SV40, can be included, as well as termination signals associated with the 3' untranslated regions of other proteins such as hGH.

Viral Enhancer

A pair of host expression vectors containing the SV40 enhancer in operable linkage to the MT-II promoter was constructed by inserting an 1120 bp SV40 fragment into the HindIII site preceding the MT-II promoter sequences in pHS1. The SV40 DNA fragment spans the SV40 origin of replication and includes nucleotide 5171 through nucleotide 5243 (at the origin), the duplicated 72 bp repeat from nucleotide 107-250, and continues through nucleotide 1046 on the side of the origin containing the 5' end of late viral mRNAs. This HindIII 1120 bp fragment is obtained from a HindIII digest of SV40 DNA (Buchman, A. R., et al, *DNA Tumor Viruses*, 2d ed (J. Tooze, ed.), Cold Spring Harbor Laboratory, New York (1981), pp. 799-841), and cloned into pBR322 for amplification. The cloning vector was cut with HindIII, and the 1120 bp SV40 DNA fragment isolated by gel electrophoresis and ligated into HindIII-digested, CIP-treated, pHS1. The resulting vectors, designated pHS1-SV (9) and pHS1-SV(10), contain the fragment in opposite orientations preceding the MT-II promoter. In pHS1-SV(9), the enhancer is about 1600 bp from the 5' mRNA start site; in the opposite orientation it is approximately 980 bp from the 5' mRNA start site. Both orientations are operable, but the orientation wherein the enhancer sequences are proximal to the start site provides higher levels of expression. It is believed that deletions which place the enhancer 250-400 bp upstream of the transcription start are optimal.

Additional vectors were constructed which place the SV40 enhancer 3' terminus 190 bp, 250 bp, and 360 bp respectively upstream from the 5' end of the MT promoter TATA box. The constructions were based on the mapping of the upstream regulatory regions of the human MT promoter described by Karin, M., et al, *Nature* (1984) 308:513-519. All constructions retain the sequences containing the duplicated sites for regulation by heavy metals, but the constructions with the 190 bp and 250 bp separations do not retain the sequence for glucocorticoid regulation which is further upstream from these sites.

These vectors, designated pHS'-SV190, pHS'SV250, and pHS'-SV360 are prepared as follows; all constructions are identical except for the length of sequence containing the metallothionein promoter and upstream region which is supplied as a fragment excised from pHS1.

For pHS'-SV190, pHS1 is digested with SacII, blunted, and ligated to KpnI linkers. The DNA is then digested with EcoRI and KpnI to liberate the appropriate portion of the MT-II control sequences. Similarly, for pHS'-SV250, pHS1 is digested with HgaI, blunted, ligated to KpnI linkers and digested with EcoRI and KpnI; for pHS'-SV360, DdeI is used in the initial digestion.

An intermediate vector containing the SV40 enhancer is prepared by inserting the HindIII/KpnI fragment of SV40 (which extends from position 5171 to position 294 and which contains the enhancer element 50 bp from the KpnI site) into KpnI/HindIII digested pUC19 to obtain pUCSV. (pUC19 contains three convenient restriction sites in the polylinker region, in order, HindIII, KpnI, and EcoRI.) The finished vectors are obtained by inserting the KpnI/EcoRI fragments prepared as described above into KpnI/EcoRI digested pUC-SV.

All of the foregoing modified vectors, thus, take advantage of the SV40 enhancer element. Other viral enhancers could, of course, be used in an analogous manner.

Transcription Termination Sequences

To provide transcription termination control sequences, DNA representing the coding sequence and 3' untranslated sequence of human growth hormone was ligated into pHS1. The intermediate vector can provide the hGH 3' untranslated sequence to coding sequences subsequently ligated into the vector in place of the hGH coding sequence.

The genomic sequences encoding hGH were isolated from p2.6-3 (DeNoto, et al, *Nucleic Acids Res* (1981) 19:3719) by digestion with BamHI, which cuts at the 5' end of the first exon, and EcoRI, which cuts 3' of the functional gene, followed by polyacrylamide gel purification. The isolated fragment was ligated into BamHI/EcoRI digested pHS1 and the ligation mixture transformed into *E. coli* MC1061 to Amp^r. Successful transformants were screened by restriction analysis, and a strain containing the desired plasmid, pMT-hGHg, was further propagated to prepare quantities of plasmid DNA.

In a manner similar to that described above for constructing pHS1-SV(9) or pHS1-SV(10), but substituting for pHS1, pMT-hGHg, a pair of vectors containing the hGH gene under the control of the MT promoter, and operably linked to SV40 enhancer, and designated, respectively, phGHg-SV(9) and phGHg-Sv(10), were obtained. The ligation mixtures were used to transform *E. coli* 1061 to Amp^r, and the correct constructions verified.

Construction of Expression Vectors

phGHg-SV(10) is then used as a host vector to accommodate the DNA sequences encoding any of the FGF analogs. phGHg-SV(10) is digested with BamHI and SmaI, blunted with Klenow, and treated with CIP to excise the hGH coding sequence. This opened vector is ligated to an NdeI(blunt)/HindIII(blunt) FGF analog fragment to obtain the desired expression vector pFGF-SV(10).

In addition, other host vectors may be used to obtain expression of these sequences, including pHS1 and pHS1 modified to contain the various configurations of SV enhancer as above described. Insertion is by analogous means, using BamHI/EcoRI digestion of the host vector. Also, DNA modified to encode any of the "long", "primary" or "short" forms of the acidic or basic FGF analogs may be employed.

These vectors are generically designated PMT-FGF for the purposes of the discussion below.

Production of FGF by Mammalian Recombinants

Chinese hamster ovary (CHO)-K1 cells are grown on medium composed of a 1:1 mixture of F12 medium and DME medium with 12% fetal calf serum. The competent cells are co-transformed with PMT-FGF and pSV2:NEO (Southern, P., et al, *J Mol Appl Genet* (1982) 1:327-341). pSV2:NEO contains a functional gene conferring resistance to the neomycin analog G418. In the transformation, 500 ng of pSV2-NEO and 5 ug of PMT-FGF are applied to a 16 mm dish of cells in a calcium phosphate-DNA co-precipitate

according to the protocol of Wigler, M., et al, *Cell* (1979) 16:777-785, with the inclusion of a two minute "shock" with 15% glycerol after four hours of exposure to the DNA. A day later, the cells are subjected to 1 mg/ml G418 to provide a pool of G418-resistant colonies, which are assayed for FGF production and then can be cloned out.

Successful transformants, also having a stable inheritance of pMT-FGF, are plated at low density for purification of clonal isolates. Small amounts of these isolates are grown in multi-well plates after exposure to 10^{-4} M zinc chloride for convenient assay of FGF production. FGF determinations are made by standard ELISA or radio-immunoassays against the antisera prepared against the appropriate FGF protein analog using standard methods. Clonal isolates which produce large amounts of the desired FGF analogs are selected.

The cells, which have been shown to produce FGF analogs under suitable conditions, are seeded at 1/10 confluency in basal medium supplemented with 10% fetal calf serum, incubated overnight, and then induced for FGF production by addition of zinc chloride in the concentration range of 1×10^{-4} M to 3×10^{-4} M. FGF levels rise for 7-10 days, under optimal inducing conditions, 2×10^{-4} M $ZnCl_2$.

If desired, the FGF analog can be obtained from the lysed cells and purified according to the procedures set forth above for the native protein, or by other standard methods known in the art.

In addition, as discussed above, secretion of the FGF protein analogs produced by the foregoing constructs can be achieved by exocytosis initiated by a calcium ionophore or other suitable stimulant. While it is not expected that pro-

teins produced by CHO cells, specifically, would be released by LPS or phorbol ester stimulation, for example, by substituting for CHO cells, cell lines derived from macrophage as recombinant hosts, such secretion can be effected. Also, by altering the construction so as to provide a signal sequence secretion using the normal constitutive pathways could also be effected using CHO or other mammalian cell hosts. Effecting secretion has some advantages, of course, since the protein purification task becomes much simpler.

On or before 9 Sep. 1985, Applicants deposited with the American Type Culture Collection (ATCC), Rockville, Md., USA, the lambda phage lamdaBB2 which was assigned ATCC accession number 40196. On or before 12 Sep. 1986, conditions of deposit for lamdaBB2 (ATCC. 40196) was converted to conform to those specified under the Budapest Treaty on the International Recognition of the Deposit of Microorganisms (Budapest Treaty). Availability of the deposited strain is not to be construed as a license to practice the invention in contravention of the rights granted under the authority of any government in accordance with its patent laws.

The embodiments of the invention in which an exclusive property or privilege is claimed are defined as follows:

1. A human basic fibroblast growth factor (FGF) protein analog wherein the cysteine at positions 78 and 96 is replaced by serine, and said analog exhibits the biological activity of native, human basic FGF, which is bFGF-C78/96S.

* * * * *