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[54] **METHOD OF PERFORMING A PROCESS FOR DETERMINING AN ITEM OF INTEREST IN A SAMPLE**

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[58] Field of Search ..... **436/43, 47, 48, 436/49, 50, 174, 180; 422/63, 64, 65, 67, 68.1, 100, 104**

[56] **References Cited**

**U.S. PATENT DOCUMENTS**

Re. 29,169	4/1977	Schuurs et al. ....	195/103.5
Re. 29,880	1/1979	Duff .....	422/64
Re. 30,627	5/1981	Bagshawe et al. ....	23/230
Re. 30,730	9/1981	Duff .....	422/64
D. 278,182	3/1985	Aihara et al. ....	D24/29
D. 283,728	5/1986	Aihara .....	D24/29
D. 288,845	3/1987	Borer et al. ....	D24/29
D. 309,080	7/1990	Buchholz .....	D9/375
D. 330,428	10/1992	Lewis et al. ....	D24/224
D. 344,138	2/1994	Nagata .....	D24/224
D. 347,479	5/1994	Hansen et al. ....	D24/224

D. 358,660	5/1995	Swift .....	D24/224
D. 359,361	6/1995	Swift .....	D24/224
544,767	9/1895	Lewis et al. ....	422/52
893,469	7/1908	Essmuller .	
1,180,665	4/1916	McElroy .....	215/247
2,587,221	2/1952	Richardson et al. ....	23/230
2,837,092	6/1958	Scholler et al. ....	128/218
2,906,423	9/1959	Sandhage .....	215/47
2,948,940	8/1960	Degener .....	24/257
2,955,722	10/1960	Antonious .....	215/100.5
3,019,932	2/1962	Singiser .....	215/37
3,038,340	6/1962	Isreeli .....	73/423

(List continued on next page.)

**FOREIGN PATENT DOCUMENTS**

2058175	12/1990	Canada .
2083424	11/1992	Canada .
0019277	5/1980	European Pat. Off. .
0019871	5/1980	European Pat. Off. .

(List continued on next page.)

**OTHER PUBLICATIONS**

Luninescent Labels for Immunoassay From Concept to Practice; F. McCapra et al., Journal of Bioluminescence and Chemiluminescence vol. 4 51-58 1989.

J. Guesdon, et al Magnetic Solid Phase Enzyme Immunoassay; Immunochemistry 1977, vol. 14 443-447.

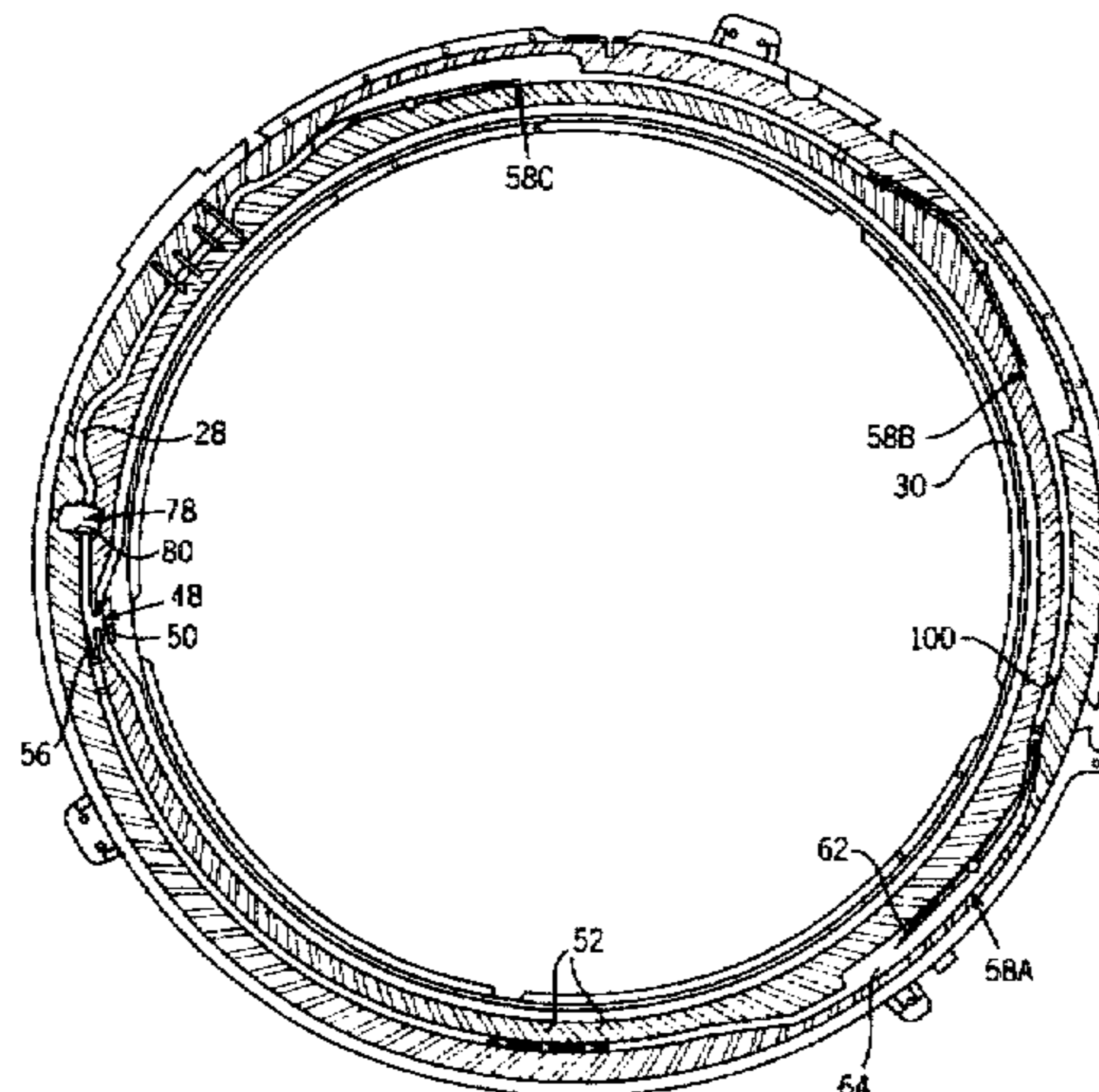
(List continued on next page.)

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[57] **ABSTRACT**

Embodiments described herein provide methods of performing a process for determining an item of interest in a sample. In one embodiment, a container for holding the sample is accepted in a process lane where a process step is selectively automatically performed on the sample in the container. The process step is selectively automatically performed on the sample in the container. An effective length of the process lane is maintained constant while a physical length of the process lane is selectively varied.

**3 Claims, 30 Drawing Sheets**





U.S. PATENT DOCUMENTS		
3,085,705	4/1963	Varney ..... 215/41
3,107,537	10/1963	Isreeli et al. .... 73/423
3,178,266	4/1965	Anthon ..... 23/253
3,190,731	6/1965	Weiskoff ..... 23/292
3,192,968	7/1965	Baruch et al. .... 141/82
3,193,358	7/1965	Baruch ..... 23/253
3,223,269	12/1965	Williams ..... 215/43
3,272,240	9/1966	Roth ..... 141/176
3,302,772	2/1967	Alsop ..... 198/221
3,307,371	3/1967	Andros ..... 62/234
3,317,069	5/1967	Chin ..... 215/47
3,399,116	8/1968	Du Bois et al. .... 196/98
3,457,048	7/1969	Stephens et al. .... 23/272.6
3,489,521	1/1970	Buckle et al. .... 23/253
3,489,525	1/1970	Natelson ..... 23/253
3,533,744	10/1970	Unger ..... 23/230
3,555,143	1/1971	Axen et al. .... 424/1
3,578,291	5/1971	Oberil ..... 259/75
3,594,129	7/1971	Jones ..... 23/253
3,606,074	9/1971	Hayes ..... 220/42
3,607,094	9/1971	Beer ..... 23/253
3,607,098	9/1971	Strande ..... 23/259
3,609,040	9/1971	Kuzel et al. .... 356/36
3,615,230	10/1971	Barnick et al. .... 23/253
3,616,264	10/1971	Ray et al. .... 195/127
3,617,222	11/1971	Matte ..... 23/230
3,621,203	11/1971	Geominy et al. .... 219/528
3,633,768	1/1972	Gulgen ..... 214/8.5
3,634,651	1/1972	Siegel ..... 219/400
3,635,094	1/1972	Oberli ..... 73/423
3,638,507	2/1972	Orner ..... 74/424
3,643,812	2/1972	Mander et al. .... 211/74
3,644,095	2/1972	Netheler et al. .... 23/259
3,652,761	3/1972	Woetail ..... 424/12
3,653,528	4/1972	Wimmer ..... 215/38
3,654,090	4/1972	Wihelmus ..... 195/103
3,655,089	4/1972	Tower ..... 220/42
3,658,478	4/1972	Spergel et al. .... 23/253
3,673,886	7/1972	Tomita et al. .... 74/424
3,676,080	7/1972	Richterich ..... 23/253
3,676,679	7/1972	Waters ..... 250/83
3,687,632	8/1972	Natelson ..... 23/259
3,702,612	11/1972	Schlesinger ..... 128/350
3,708,264	1/1973	Jottier ..... 23/230
3,720,116	3/1973	Better et al. .... 74/459
3,722,312	3/1973	Better et al. .... 74/459
3,723,066	3/1973	Moran ..... 23/253
3,727,029	4/1973	Chrow ..... 219/301
3,728,079	4/1973	Moran ..... 23/253
3,746,514	7/1973	Colvin et al. .... 23/253
3,753,657	8/1973	Downing et al. .... 23/253
3,764,268	10/1973	Kosowsky et al. .... 23/253
3,765,237	10/1973	Blackmer et al. .... 73/190
3,767,364	10/1973	Ritchie et al. .... 23/259
3,770,382	11/1973	Carter et al. .... 23/253
3,784,785	1/1974	Noland ..... 219/301
3,790,346	2/1974	Ritchie ..... 23/253
3,791,537	2/1974	Conklin ..... 214/6.5
3,796,544	3/1974	Zauft et al. .... 23/259
3,806,321	4/1974	Durrum et al. .... 23/253
3,807,457	4/1974	Logsdon ..... 138/89
3,807,955	4/1974	Note, Jr. et al. .... 23/230
3,811,842	5/1974	Diebler et al. .... 23/259
3,813,215	5/1974	Ward ..... 432/226
3,823,840	7/1974	Zackheim ..... 215/37
3,826,397	7/1974	Atkins ..... 215/41
3,826,621	7/1974	Johnson, Jr. et al. .... 23/259
3,830,108	8/1974	Spong ..... 73/425.6
3,843,323	10/1974	Quame ..... 23/230
3,850,174	11/1974	Ayres ..... 128/272
3,850,341	11/1974	Bart ..... 220/212
3,850,580	11/1974	Moore et al. .... 23/259
3,851,541	12/1974	Ploss et al. .... 74/459
3,854,879	12/1974	Figuroa et al. .... 23/230
3,868,493	2/1975	Caroleo ..... 219/318
3,883,305	5/1975	Hoskins et al. .... 23/253
3,883,306	5/1975	Widen ..... 23/253
3,894,706	7/1975	Mizusawa ..... 248/68
3,897,216	7/1975	Jones ..... 23/259
3,900,289	8/1975	Liston ..... 23/230
3,912,456	10/1975	Young ..... 23/253
3,917,455	11/1975	Bak et al. .... 23/253
3,933,997	1/1976	Hersh et al. .... 424/1
3,948,607	4/1976	Atwood et al. .... 23/259
3,949,189	4/1976	Bibro et al. .... 219/301
3,951,605	4/1976	Natelson ..... 23/253
3,970,518	7/1976	Graever ..... 195/1.5
3,977,551	8/1976	Ciarico ..... 215/1
3,980,268	9/1976	Ellis ..... 249/49
3,981,776	9/1976	Saxholm ..... 195/103
3,985,508	10/1976	Williams ..... 23/253
4,000,252	12/1976	Kosak ..... 424/1
4,000,973	1/1977	Petesen ..... 23/230
4,000,974	1/1977	Acord ..... 23/230
4,002,532	1/1977	Weltman et al. .... 195/103
4,004,883	1/1977	Meyer et al. .... 23/259
4,016,043	4/1977	Schuurs et al. .... 195/103
4,018,886	4/1977	Graever ..... 424/12
4,022,579	5/1977	Revillet et al. .... 23/259
4,034,071	7/1977	Strickler ..... 424/1
4,039,287	8/1977	Moran ..... 23/253
4,039,652	8/1977	Adams et al. .... 424/1
4,040,533	8/1977	De Boer et al. .... 214/310
4,041,146	8/1977	Graever ..... 424/1
4,045,179	8/1977	Bunce ..... 23/259
4,046,248	9/1977	Goffredo et al. .... 198/583
4,047,034	9/1977	Auphan ..... 250/354
4,054,416	10/1977	Duff ..... 23/259
4,058,367	11/1977	Gilford ..... 23/253
4,065,358	12/1977	Kawai et al. .... 195/127
4,066,412	1/1978	Johnson et al. .... 23/253
4,067,694	1/1978	Blakely et al. .
4,077,444	3/1978	Gilson et al. .... 141/130
4,077,804	3/1978	Vanzo ..... 96/1
4,078,971	3/1978	Arkles et al. .... 195/63
4,080,833	3/1978	Huber ..... 73/423
4,081,245	3/1978	Polito ..... 23/230
4,081,246	3/1978	Polito et al. .... 23/230
4,094,641	6/1978	Friswell ..... 23/230
4,098,876	7/1978	Plasie et al. .... 424/1
4,104,029	8/1978	Maier, Jr. .... 23/230
4,106,907	8/1978	Charlton et al. .... 23/230
4,108,972	8/1978	Dreyer ..... 424/1
4,112,517	9/1978	Giombini ..... 366/102
4,118,280	10/1978	Charles et al. .... 195/127
4,118,801	10/1978	Kraft et al. .... 366/111
4,119,709	10/1978	Holub ..... 424/1
4,120,662	10/1978	Fosslien ..... 73/425
4,123,121	10/1978	Ernst et al. .... 308/6
4,125,492	11/1978	Cuatrecasas et al. .... 260/9
4,131,788	12/1978	Fulbrook ..... 219/535
4,133,383	1/1979	Burns et al. .... 356/36
4,133,436	1/1979	Werder et al. .... 422/65
4,133,873	1/1979	Noller ..... 424/8
4,134,512	1/1979	Nugent ..... 215/247
4,137,216	1/1979	Lemper et al. .... 260/42
4,139,242	2/1979	Ernst ..... 308/6
4,139,604	2/1979	Gutcho et al. .... 424/1
4,140,020	2/1979	Cook ..... 73/425
4,141,524	2/1979	Corvese, Jr. .... 248/70
4,141,687	2/1979	Forrest et al. .... 23/230
4,147,250	4/1979	Schulz ..... 198/472



4,151,931	5/1979	Scherer et al.	221/226	4,294,799	10/1981	Stephens et al.	422/62
4,152,210	5/1979	Robinson et al.	195/63	4,295,572	10/1981	Percarpio	215/247
4,152,269	5/1979	Babson	210/516	4,297,337	10/1981	Mansfield et al.	424/1
4,152,577	5/1979	Leavines	219/301	4,298,593	11/1981	Ling	424/1
4,155,534	5/1979	Ithakissios	424/1	4,298,687	11/1981	Maes	435/7
4,155,535	5/1979	Giaever	424/1	4,302,534	11/1981	Halmann et al.	435/6
4,155,861	5/1979	Allington	364/497	4,305,668	12/1981	Bilbrey	366/111
4,155,978	5/1979	Naono et al.	422/64	4,305,924	12/1981	Piasio et al.	424/1
4,157,323	6/1979	Yen et al.	260/29	4,307,766	12/1981	Tanokura	150/8
4,160,165	7/1979	McCombs et al.	250/354	4,311,348	1/1982	Olschewski et al.	308/6
4,166,095	8/1979	Kling et al.	422/67	4,311,667	1/1982	Gocho	422/64
4,166,104	8/1979	Wagner et al.	424/1	4,312,835	1/1982	Zoltan et al.	422/70
4,168,955	9/1979	Allington	23/230	4,313,734	2/1982	Leuvering	23/230
4,169,138	9/1979	Jonsson	424/12	4,313,735	2/1982	Yamashita et al.	23/230
4,169,804	10/1979	Yapel et al.	252/62	4,315,891	2/1982	Sakurada	422/64
4,177,253	12/1979	Davies et al.	424/1	4,315,907	2/1982	Fridlender et al.	424/1
4,185,084	1/1980	Mochida et al.	424/1	4,318,707	3/1982	Litman et al.	23/230
4,187,075	2/1980	Noller	23/230	4,318,884	3/1982	Suzuki	422/63
4,191,287	3/1980	Brook et al.	198/472	4,318,980	3/1982	Boguslaski et al.	435/7
4,192,845	3/1980	Kalasek	422/25	4,320,109	3/1982	Wolf et al.	424/1
4,193,866	3/1980	Slusarczyk et al.	210/6	4,322,216	3/1982	Lillig et al.	23/230
4,200,436	4/1980	Mochida et al.	23/230	4,325,909	4/1982	Coulter et al.	422/63
4,202,634	5/1980	Kraft et al.	366/111	4,325,910	4/1982	Jordan	422/64
4,205,885	6/1980	Ernst et al.	308/6	4,330,299	5/1982	Cerami	23/230
4,206,094	6/1980	Yen et al.	260/8	4,332,471	6/1982	Gross	356/246
4,206,951	6/1980	Ernst et al.	308/6	4,332,783	6/1982	Pernice et al.	424/1
4,207,289	6/1980	Weiss	422/104	4,333,356	6/1982	Bartels et al.	73/864
4,208,484	6/1980	Sogi et al.	4253/286	4,335,620	6/1982	Adams	73/863
4,213,999	7/1980	Witak et al.	424/285	4,335,730	6/1982	Griffin	128/760
4,218,539	8/1980	Weltman	435/188	4,341,736	7/1982	Drbal et al.	422/100
4,228,237	10/1980	Hevey et al.	435/7	4,343,766	8/1982	Sisti et al.	422/63
4,229,104	10/1980	Lahme et al.	356/246	4,343,901	8/1982	DeFilippi	435/176
4,230,685	10/1980	Senyei et al.	424/12	4,345,843	8/1982	Berglund et al.	366/219
4,230,797	10/1980	Bogulaski et al.	435/7	4,346,056	8/1982	Sakurada	422/64
4,231,750	11/1980	Dowben et al.	23/230	4,346,742	8/1982	Chase et al.	141/1
4,232,119	11/1980	Carlsson et al.	435/7	4,349,510	9/1982	Kolehmainen et al.	422/66
4,234,538	11/1980	Ginsberg et al.	422/64	4,351,800	9/1982	Kopp et al.	422/70
4,234,540	11/1980	Ginsberg et al.	422/64	4,355,165	10/1982	Boguslaski et al.	544/237
4,235,869	11/1980	Schwarzberg	424/8	4,356,722	11/1982	Bunce et al.	73/53
4,235,960	11/1980	Sasse et al.	435/7	4,356,967	11/1982	Lunick	237/14
4,238,195	12/1980	Boguslaski et al.	23/230	4,357,301	11/1982	Cassaday et al.	422/64
4,239,298	12/1980	Ernst et al.	308/6	4,362,531	12/1982	de Steenwinkel et al.	23/230
4,241,176	12/1980	Avrameas et al.	435/7	4,363,759	12/1982	Boguslaski et al.	260/112
4,244,459	1/1981	Garrett	198/389	4,363,781	12/1982	Akamatsu et al.	422/65
4,244,920	1/1981	Manschot et al.	422/102	4,366,118	12/1982	Bunce et al.	422/57
4,250,400	2/1981	Lee	219/549	4,366,119	12/1982	Takeuchi	422/65
4,254,460	3/1981	Achter et al.	364/104	4,369,226	1/1983	Rembaum	428/334
4,256,106	3/1981	Shoor	128/247	4,372,745	2/1983	Mandle et al.	436/537
4,256,725	3/1981	Rutner et al.	424/1	4,373,925	2/1983	Litman et al.	435/7
4,256,960	3/1981	Snider	250/252	4,373,931	2/1983	Takekawa	436/539
4,257,884	3/1981	Lim	210/656	4,376,110	3/1983	David et al.	436/513
4,259,288	3/1981	Welch	422/63	4,380,580	4/1983	Boguslaski et al.	435/7
4,259,289	3/1981	Curry et al.	422/64	4,383,031	5/1983	Boguslaski et al.	435/7
4,259,291	3/1981	Smythe	422/82	4,385,126	5/1983	Chen et al.	436/518
4,261,893	4/1981	Boguslaski et al.	260/326	4,397,385	8/1983	Booth et al.	198/345
4,265,855	5/1981	Mandle et al.	422/654	4,402,909	9/1983	Solazzi	422/50
4,266,651	5/1981	Strom	198/345	4,403,040	9/1983	Van Aken	436/501
4,267,149	5/1981	Bruckner et al.	422/65	4,407,943	10/1983	Cole et al.	435/7
4,267,234	5/1981	Rembaum	428/403	4,407,964	10/1983	Elings et al.	436/518
4,267,235	5/1981	Rembaum	428/407	4,414,324	11/1983	Stout	435/7
4,268,477	5/1981	Herzstark	422/64	4,415,700	11/1983	Batz et al.	524/548
4,272,482	6/1981	Jessop et al.	422/65	4,418,846	12/1983	Pong et al.	222/189
4,272,506	6/1981	Schwarzberg	424/8	4,419,734	12/1983	Wolfson et al.	365/567
4,272,510	6/1981	Smith et al.	427/47	4,421,860	12/1983	Elings et al.	436/518
4,276,051	6/1981	Ginsberg et al.	23/230	4,433,059	2/1984	Chang et al.	436/512
4,276,258	6/1981	Ginsberg et al.	422/64	4,433,060	2/1984	Frenzel	436/518
4,277,440	7/1981	Jessop et al.	422/100	4,435,509	3/1984	Berthold et al.	436/518
4,278,437	7/1981	Haggar	23/230	4,437,762	3/1984	Glenn et al.	356/326
4,279,992	7/1981	Boguslaski et al.	435/7	4,438,068	3/1984	Forrest	422/61
4,281,387	7/1981	Kraft et al.	364/497	4,438,986	3/1984	Teramachi	308/6
4,292,920	10/1981	Smith et al.	118/425	4,451,433	5/1984	Yamashita et al.	422/63
4,294,078	10/1981	Percarpio et al.	215/247	4,454,226	6/1984	Ali et al.	435/7



4,454,234	6/1984	Czerlinski	436/526	4,634,576	1/1987	Galle et al.	422/102
4,454,939	6/1984	Kampf et al.	198/341	4,637,985	1/1987	Sidki et al.	436/518
4,455,280	6/1984	Shinohara et al.	422/63	4,639,135	1/1987	Borer et al.	356/246
4,456,037	6/1984	Gocho	141/1	4,639,425	1/1987	Baier	436/518
4,459,265	7/1984	Berglund	422/64	4,639,875	1/1987	Abraham et al.	364/479
4,459,359	7/1984	Neurath	436/54	4,643,879	2/1987	Hanaway	422/104
4,472,352	9/1984	Quesneau et al.	422/52	4,645,646	2/1987	Gadow et al.	422/61
4,477,576	10/1984	Deutsch et al.	436/500	4,646,747	3/1987	Cais et al.	436/500
4,477,578	10/1984	Miles et al.	436/518	4,647,432	3/1987	Wakatake	422/64
4,478,817	10/1984	Campbell et al.	424/7	4,649,116	3/1987	Daty et al.	435/287
4,482,636	11/1984	Mochida et al.	436/518	4,651,813	3/1987	Witt et al.	165/30
4,483,823	11/1984	Umetsu et al.	422/63	4,652,519	3/1987	Warszawsky et al.	435/7
4,483,927	11/1984	Takekawa	436/43	4,652,533	3/1987	Jolley	436/518
4,484,061	11/1984	Zelinka et al.	219/301	4,654,299	3/1987	Lentfer	435/7
4,484,815	11/1984	Akiyama	356/325	4,654,300	3/1987	Zuk et al.	435/7
4,488,633	12/1984	Kampf	198/472	4,657,851	4/1987	Feller et al.	435/7
4,491,634	1/1985	Frenzel	436/518	4,659,678	4/1987	Forrest et al.	436/512
4,492,751	1/1985	Boguslaski et al.	435/7	4,661,444	4/1987	Li	435/7
4,495,295	1/1985	Neurath	436/518	4,661,498	4/1987	Lau et al.	428/405
4,495,296	1/1985	Neurath et al.	436/530	4,663,277	5/1987	Wang	435/5
4,497,774	2/1985	Scordato	422/73	4,666,866	5/1987	Krauth	436/518
4,504,585	3/1985	Reynolds	436/518	4,668,617	5/1987	Furuta et al.	435/4
4,506,009	3/1985	Lenhoff et al.	435/7	4,670,219	6/1987	Nelson et al.	422/63
4,506,777	3/1985	Kampf	198/341	4,670,381	6/1987	Frickey et al.	435/7
4,515,752	5/1985	Miramanda	422/99	4,670,383	6/1987	Baier et al.	435/7
4,517,160	5/1985	Galle et al.	422/65	4,672,040	6/1987	Josephson	436/526
4,517,288	5/1985	Giegel et al.	435/7	4,676,656	6/1987	Cook et al.	366/142
4,517,460	5/1985	Meulenbrugge et al.	250/252	4,676,951	6/1987	Armes et al.	422/65
4,517,851	5/1985	Tice	73/864	4,678,752	7/1987	Thorne et al.	435/291
4,522,921	6/1985	Ogawa	436/65	4,680,275	7/1987	Wagner et al.	436/518
4,523,295	6/1985	Zato	364/900	4,681,741	7/1987	Hanaway	422/100
4,523,862	6/1985	Yasui et al.	384/564	4,687,638	8/1987	Benajam	422/73
4,523,864	6/1985	Walter et al.	384/613	4,689,305	8/1987	Stiffey et al.	435/291
4,528,159	7/1985	Liston	422/65	4,693,969	9/1987	Saxena et al.	435/7
4,533,255	8/1985	Gronholz et al.	366/108	4,693,970	9/1987	O'Connell et al.	435/7
4,533,629	8/1985	Litman et al.	435/7	4,695,392	9/1987	Whitehead et al.	252/62
4,534,465	8/1985	Rothermel et al.	206/443	4,695,393	9/1987	Whitehead et al.	252/62
4,536,369	8/1985	Sakurada et al.	422/65	4,698,302	10/1987	Whitehead et al.	435/94
4,537,510	8/1985	Takahasi	356/435	4,699,766	10/1987	Yamashita	422/64
4,537,861	8/1985	Elings et al.	436/518	4,699,767	10/1987	Aihara	422/65
4,540,660	9/1985	Harte et al.	435/7	4,706,736	11/1987	Gyori	165/30
4,541,291	9/1985	Churchill et al.	73/864	4,707,251	11/1987	Jenkins et al.	209/569
4,542,661	9/1985	Teramachi	74/424	4,708,886	11/1987	Nelson	422/72
4,542,833	9/1985	DeVaughn	215/319	4,710,472	12/1987	Saur et al.	435/287
4,545,497	10/1985	Martha, Jr.	215/253	4,711,839	12/1987	Singhal	435/4
4,551,426	11/1985	Freytag et al.	435/7	4,713,974	12/1987	Stone	73/864
4,551,619	11/1985	Lefebvre	219/523	4,731,225	3/1988	Wakatake	422/65
4,552,812	11/1985	Margel et al.	428/407	4,731,337	3/1988	Luotola et al.	436/526
4,552,839	11/1985	Gould et al.	435/7	4,732,811	3/1988	Margel	428/403
4,554,088	11/1985	Whitehead et al.	252/62	4,737,342	4/1988	Herrmann et al.	422/64
4,555,183	11/1985	Thomas	366/208	4,737,453	4/1988	Primus	435/5
4,559,120	12/1985	Royse et al.	204/182	4,738,825	4/1988	Kelln	422/72
4,560,269	12/1985	Baldszun et al.	356/246	4,740,457	4/1988	Parratt	435/7
4,561,820	12/1985	Matheny, III et al.	414/331	4,743,536	5/1988	Evanega et al.	435/7
4,576,912	3/1986	Yaverbaum et al.	435/7	4,743,544	5/1988	Namba et al.	435/7
4,578,244	3/1986	Cosgrove, Jr. et al.	422/65	4,743,560	5/1988	Campbell et al.	436/501
4,581,521	4/1986	Grise	219/535	4,745,077	5/1988	Holian et al.	436/526
4,583,668	4/1986	Maynard, Jr.	222/529	4,747,693	5/1988	Kahl	366/208
4,584,277	4/1986	Ullman	436/501	4,753,775	6/1988	Ebersole et al.	422/81
4,588,697	5/1986	Khanna et al.	436/518	4,754,414	6/1988	Gocho	364/497
4,595,562	6/1986	Liston et al.	422/65	4,755,055	7/1988	Johnson et al.	356/440
4,604,348	8/1986	Neurath	435/7	4,755,356	7/1988	Robbins et al.	422/102
4,610,546	9/1986	Intraub	366/110	4,758,523	7/1988	Harjunmaa	436/531
4,613,567	9/1986	Yasoshima et al.	435/7	4,761,268	8/1988	Andersen et al.	422/72
4,615,360	10/1986	Jacobs	141/18	4,761,379	8/1988	Williams et al.	435/296
4,623,621	11/1986	Pestka	435/7	4,763,803	8/1988	Schneider	215/260
4,623,629	11/1986	Kerschensteiner	436/518	4,764,342	8/1988	Kelln et al.	422/72
4,628,037	12/1986	Chagnon et al.	436/526	4,771,429	9/1988	Davis et al.	371/27
4,629,688	12/1986	Bolguslaski et al.	435/7	4,772,453	9/1988	Lisenbee	422/52
4,629,690	12/1986	Weng et al.	435/7	4,772,550	9/1988	Greenquist	435/7
4,629,703	12/1986	Uffenheimer	436/45	4,774,055	9/1988	Wakatake et al.	422/64
4,634,575	1/1987	Kawakami et al.	422/63	4,774,174	9/1988	Giegel et al.	435/5



4,774,191	9/1988	Khanna et al. ....	436/518	4,931,385	6/1990	Block et al. ....	435/7
4,775,635	10/1988	Ebersole et al. ....	436/501	4,931,402	6/1990	Abplanalp ....	435/291
4,775,636	10/1988	Moeremans et al. ....	436/518	4,933,146	6/1990	Meyer et al. ....	422/63
4,777,145	10/1988	Luotola et al. ....	436/526	4,933,276	6/1990	Baret ....	453/7
4,778,763	10/1988	Makiguchi et al. ....	436/47	4,935,339	6/1990	Zahradnik ....	435/5
4,778,767	10/1988	Hummelen et al. ....	436/531	4,937,048	6/1990	Sakai et al. ....	422/63
4,780,421	10/1988	Kameda et al. ....	436/518	4,939,946	7/1990	Teramschi ....	74/89
4,781,891	11/1988	Galle et al. ....	422/64	4,941,201	7/1990	Davis ....	455/41
4,783,336	11/1988	Margel et al. ....	424/462	4,941,809	7/1990	Pinkerton ....	417/500
4,783,835	11/1988	Satoh ....	382/48	4,942,017	7/1990	Turpen ....	422/64
4,784,213	11/1988	Eager et al. ....	165/2	4,943,164	7/1990	Ohishi et al. ....	366/149
4,785,953	11/1988	Buchholz et al. ....	215/365	4,944,924	7/1990	Mawhirt et al. ....	422/104
4,786,606	11/1988	Giegel et al. ....	436/500	4,946,651	8/1990	Liston et al. ....	422/102
4,788,136	11/1988	Grenier et al. ....	435/7	4,948,726	8/1990	Longoria ....	435/7
4,788,150	11/1988	Nelson et al. ....	436/45	4,950,588	8/1990	Dattagupta ....	435/6
4,791,055	12/1988	Boguslaski et al. ....	435/7	4,951,512	8/1990	Mazza et al. ....	73/861
4,791,056	12/1988	Sizto et al. ....	435/7	4,952,707	8/1990	Edwards et al. ....	549/221
4,793,973	12/1988	Ringrose ....	422/102	4,953,075	8/1990	Nan et al. ....	364/140
4,798,095	1/1989	Itoh ....	73/863	4,954,149	9/1990	Fullermann ....	55/386
4,799,599	1/1989	Herrmann ....	215/307	4,954,319	9/1990	Koizumi et al. ....	422/67
4,806,313	2/1989	Ebersole et al. ....	422/61	4,954,452	9/1990	Yost et al. ....	436/524
4,808,380	2/1989	Minekane ....	422/64	4,954,882	9/1990	Kamemoto ....	358/22
4,808,522	2/1989	Atabekov et al. ....	435/7	4,959,303	9/1990	Milburn et al. ....	435/7
4,816,418	3/1989	Mack et al. ....	436/518	4,961,906	10/1990	Andersen et al. ....	422/102
4,820,497	4/1989	Howell ....	422/63	4,962,023	10/1990	Todd et al. ....	435/7
4,824,778	4/1989	Nagai et al. ....	435/7	4,965,049	10/1990	Lillig et al. ....	422/68
4,827,780	5/1989	Sarrine et al. ....	73/864	4,965,187	10/1990	Tonelli ....	435/5
4,830,832	5/1989	Arpagaus et al. ....	422/65	4,966,839	10/1990	Kaspar ....	435/7
4,837,159	6/1989	Yamada ....	436/45	4,967,159	10/1990	Manes ....	324/650
4,837,395	6/1989	Leeder et al. ....	435/7	4,969,565	11/1990	Justai et al. ....	215/250
4,842,827	6/1989	Graf et al. ....	422/112	4,977,786	12/1990	Davis ....	73/864
4,843,000	6/1989	Litman et al. ....	435/7	4,978,625	12/1990	Wagner et al. ....	436/518
4,843,001	6/1989	Haug et al. ....	435/7	4,980,293	12/1990	Jeffs ....	435/296
4,843,010	6/1989	Nawinski et al. ....	435/7	4,984,628	1/1991	Uchida et al. ....	165/26
4,845,025	7/1989	Lary et al. ....	435/2	4,986,891	1/1991	Sarrine et al. ....	204/299
4,848,917	7/1989	Benin et al. ....	366/208	4,988,618	1/1991	Li et al. ....	435/6
4,849,176	7/1989	Sakagami ....	422/64	4,989,822	2/1991	Fannon ....	248/632
4,849,338	7/1989	Litman et al. ....	435/7	4,992,377	2/1991	Saxholm ....	435/299
4,850,470	7/1989	Ferkany ....	198/345	4,997,768	3/1991	Uffenheimer et al. ....	436/45
4,852,967	8/1989	Cook et al. ....	350/96	5,004,582	4/1991	Miyata et al. ....	422/56
4,855,110	8/1989	Marker et al. ....	422/102	5,004,904	4/1991	Yamakawa et al. ....	250/257
4,855,242	8/1989	Soeldner ....	436/539	5,008,082	4/1991	Shaw ....	422/65
4,859,423	8/1989	Perlman ....	422/102	5,009,942	4/1991	Benin et al. ....	428/36
4,859,583	8/1989	Heller et al. ....	435/7	5,009,998	4/1991	Chow et al. ....	435/7
4,861,553	8/1989	Mawhirt et al. ....	422/65	5,015,157	5/1991	Pinkerton et al. ....	417/500
4,861,554	8/1989	Sakuma ....	422/65	5,017,790	5/1991	Kojima ....	250/455
4,863,690	9/1989	Berthold et al. ....	422/52	5,020,980	6/1991	Pinkerton ....	417/500
4,863,875	9/1989	Bailey et al. ....	436/518	5,024,256	6/1991	Vadher ....	141/329
4,868,130	9/1989	Hargreaves ....	436/526	5,030,418	7/1991	Miyata ....	422/63
4,868,131	9/1989	Hiratsuka ....	436/528	5,037,612	8/1991	Takahashi et al. ....	422/64
4,871,683	10/1989	Harris et al. ....	436/531	5,038,911	8/1991	Doane et al. ....	198/357
4,886,177	12/1989	Foster ....	215/247	5,038,958	8/1991	Dreier ....	220/366
4,890,930	1/1990	Nohso ....	366/208	5,039,860	8/1991	Yrionen et al. ....	250/328
4,891,311	1/1990	Anawis et al. ....	435/7	5,043,141	8/1991	Wilson et al. ....	436/52
4,895,453	1/1990	Devlin et al. ....	366/219	5,043,143	8/1991	Shaw et al. ....	422/65
4,895,650	1/1990	Wang ....	210/222	5,044,889	9/1991	Pinkerton ....	417/53
4,900,513	2/1990	Barker et al. ....	422/64	5,047,210	9/1991	Melet ....	422/64
4,900,686	2/1990	Arnost et al. ....	436/546	5,055,262	10/1991	Sakagami ....	422/64
4,904,583	2/1990	Mapes et al. ....	435/7	5,057,281	10/1991	Torti et al. ....	422/100
4,904,632	2/1990	Pesek et al. ....	502/158	5,061,448	10/1991	Mabe et al. ....	422/99
4,906,432	3/1990	Geiselman ....	422/63	5,061,630	10/1991	Knopf et al. ....	435/290
4,906,433	3/1990	Minekane ....	422/64	5,066,135	11/1991	Meyer et al. ....	366/208
4,908,186	3/1990	Sakamaki ....	422/64	5,066,844	11/1991	Schuster et al. ....	219/85
4,910,148	3/1990	Sorensen et al. ....	435/317	5,068,088	11/1991	Hall et al. ....	422/52
4,911,230	3/1990	Maver et al. ....	165/48	5,071,625	12/1991	Kelln et al. ....	422/72
4,916,080	4/1990	Imai et al. ....	436/518	5,071,766	12/1991	Barr et al. ....	435/284
4,916,081	4/1990	Kamada et al. ....	436/526	5,073,625	12/1991	Derbyshire ....	530/333
4,919,887	4/1990	Wakatake ....	422/67	5,077,013	12/1991	Guigan ....	422/64
4,927,545	5/1990	Roginski ....	210/745	5,077,488	12/1991	Davis ....	307/296
4,927,605	5/1990	Dorn et al. ....	422/102	5,079,424	1/1992	Kobayashi ....	250/369
4,927,769	5/1990	Chang et al. ....	436/518	5,081,872	1/1992	Greter ....	73/864
4,928,539	5/1990	Champecix ....	73/864	5,082,628	1/1992	Andreotti et al. ....	422/82



5,084,242	1/1992	Sakuma et al.	422/100	5,240,674	8/1993	Armor	422/6
5,086,215	2/1992	Carsner et al.	235/462	5,240,679	8/1993	Stettler	422/67
5,086,233	2/1992	Stafford et al.	250/256	5,242,659	9/1993	Wurschum	422/65
5,087,423	2/1992	Ishibashi	422/67	5,242,660	9/1993	Hsei	422/102
5,089,418	2/1992	Shaw et al.	436/46	5,244,633	9/1993	Jakubowicz et al.	422/64
5,091,206	2/1992	Wang et al.	427/2	5,244,663	9/1993	Bruttman et al.	424/400
5,096,670	3/1992	Harris et al.	422/65	5,246,354	9/1993	Pardinas	417/500
5,098,660	3/1992	Devaney, Jr.	422/99	5,246,665	9/1993	Tyranski et al.	422/64
5,098,661	3/1992	Froehlich et al.	422/102	5,250,440	10/1993	Kelln et al.	436/48
5,098,663	3/1992	Berthold et al.	422/104	5,252,485	10/1993	Zlobinsky et al.	435/316
5,102,631	4/1992	Jordan et al.	422/42	5,254,312	10/1993	Staebler et al.	422/100
5,104,231	4/1992	Collier et al.	366/208	5,260,028	11/1993	Astle	422/81
5,104,621	4/1992	Pfost et al.	422/67	5,264,182	11/1993	Sakagami	422/63
5,104,807	4/1992	Mitsumaki et al.	436/47	5,266,272	11/1993	Griner et al.	422/104
5,104,808	4/1992	Laska et al.	436/48	5,270,210	12/1993	Weyrauch et al.	436/43
5,108,175	4/1992	Whitlock	356/218	5,272,092	12/1993	Hamasaki et al.	436/172
5,108,703	4/1992	Pfost et al.	422/65	5,273,715	12/1993	Bridgham et al.	422/63
5,108,928	4/1992	Menard et al.	436/43	5,275,299	1/1994	Konrad et al.	215/341
5,112,646	5/1992	Koshi et al.	422/52	5,277,873	1/1994	Hsei	422/102
5,122,343	6/1992	Ishizaka et al.	422/66	5,279,210	1/1994	Pinkerton	92/170
5,123,477	6/1992	Tyler	165/2	5,282,149	1/1994	Grandone et al.	364/497
5,127,541	7/1992	Wakatake	220/737	5,283,079	2/1994	Wang et al.	427/2
5,128,103	7/1992	Wang et al.	422/64	5,286,652	2/1994	James et al.	436/48
5,128,105	7/1992	Berthold et al.	422/104	5,288,466	2/1994	Burns	422/102
5,130,254	7/1992	Collier et al.	436/54	5,290,513	3/1994	Berthold et al.	422/52
5,133,936	7/1992	Umetsu et al.	422/64	5,290,708	3/1994	Ashihara et al.	436/526
5,137,693	8/1992	Mawhirt	422/104	5,296,191	3/1994	Hall et al.	422/52
5,139,743	8/1992	Ishizaka et al.	422/63	5,296,195	3/1994	Pang et al.	422/82
5,139,744	8/1992	Kowalski	422/67	5,297,599	3/1994	Bucheli	141/329
5,139,951	8/1992	Butz et al.	435/284	5,298,425	3/1994	Kuhn et al.	436/43
5,143,236	9/1992	Gueret	215/311	5,304,347	4/1994	Mann et al.	422/67
5,145,784	9/1992	Cox et al.	436/526	5,304,787	4/1994	Wang	235/462
5,147,529	9/1992	Lee et al.	210/695	5,309,981	5/1994	Binder	265/64
5,158,895	10/1992	Ashihara et al.	436/526	5,312,730	5/1994	Piran et al.	435/7
5,162,236	11/1992	Pang et al.	436/517	5,314,663	5/1994	Mimura	422/67
5,163,360	11/1992	Petz	99/468	5,314,825	5/1994	Weyrauch et al.	436/43
5,163,582	11/1992	Godolphin et al.	222/1	5,315,094	5/1994	Lisy	235/385
5,164,318	11/1992	Sato et al.	435/288	5,315,375	5/1994	Allen	356/417
5,167,929	12/1992	Korf et al.	422/102	5,316,245	5/1994	Ruckwardt	248/68
5,171,979	12/1992	Kwa et al.	250/223	5,316,726	5/1994	Babson et al.	422/65
5,174,960	12/1992	Shaw et al.	422/63	5,316,954	5/1994	Hupe et al.	436/89
5,175,086	12/1992	Takekawa et al.	435/7	5,318,914	6/1994	Matte et al.	436/526
5,176,203	1/1993	Larzul	165/61	5,320,809	6/1994	Dunn et al.	422/64
5,178,019	1/1993	Keiter	73/863	5,322,668	6/1994	Tomasso	422/104
5,178,479	1/1993	Brown et al.	403/13	5,324,025	6/1994	Chadwick et al.	271/184
5,178,834	1/1993	Kagayama et al.	422/65	5,324,480	6/1994	Shumate et al.	422/63
5,180,555	1/1993	Monget	422/102	5,332,679	7/1994	Simons et al.	436/518
5,183,638	2/1993	Wakatake	422/64	5,344,610	9/1994	Shaw	422/100
5,186,339	2/1993	Heissler	211/74	5,346,303	9/1994	Heinonen et al.	366/208
5,186,827	2/1993	Liberti et al.	210/222	5,348,705	9/1994	Koreyasu et al.	422/67
5,187,084	2/1993	Hallsby	435/91	5,350,564	9/1994	Mazza et al.	422/63
5,191,967	3/1993	Woltjer et al.	198/781	5,351,801	10/1994	Markin et al.	198/346
5,200,084	4/1993	Liberti et al.	210/695	5,352,612	10/1994	Huber et al.	436/47
5,200,151	4/1993	Long	422/100	5,355,304	10/1994	DeMoranville et al.	364/413
5,200,975	4/1993	Kato	373/109	5,363,885	11/1994	McConnell et al.	141/1
5,202,093	4/1993	Cloyd	422/102	5,366,062	11/1994	Markin et al.	198/345
5,206,171	4/1993	Dillon et al.	435/293	5,366,697	11/1994	Tomasso et al.	422/64
5,207,986	5/1993	Kadota et al.	422/65	5,370,843	12/1994	Chiodo	422/99
5,208,762	5/1993	Charhut et al.	364/478	5,371,350	12/1994	Motolese	250/207
5,209,903	5/1993	Kanamori et al.	422/65	5,372,782	12/1994	Karkantis et al.	422/63
5,213,761	5/1993	Sakagami	422/63	5,374,395	12/1994	Robinson et al.	422/64
5,215,376	6/1993	Schulte et al.	366/348	5,377,854	1/1995	Cusack	215/364
5,215,714	6/1993	Okada et al.	422/64	5,378,433	1/1995	Duckett et al.	422/100
5,216,926	6/1993	Lipscomb	73/864	5,378,881	1/1995	Adachi	235/462
5,223,218	6/1993	Fukuoka et al.	422/52	5,380,485	1/1995	Takahashi et al.	422/62
5,225,165	7/1993	Perlman	422/102	5,380,487	1/1995	Choperena et al.	422/63
5,229,074	7/1993	Heath et al.	422/64	5,380,488	1/1995	Wakatake	422/65
5,232,664	8/1993	Krawzak et al.	422/64	5,384,096	1/1995	Burns	422/102
5,232,665	8/1993	Burkovich	422/65	5,391,499	2/1995	Karkantis et al.	436/180
5,236,666	8/1993	Hulette	422/65	5,392,949	2/1995	McKenna	220/712
5,236,824	8/1993	Fujiwara et al.	435/5	5,393,965	2/1995	Bravman et al.	235/383
5,238,810	8/1993	Fujiwara et al.	435/5	5,399,836	3/1995	Pavlidis et al.	235/462



5,401,465	3/1995	Smethers et al. ....	422/52	0209290	7/1986	European Pat. Off. .
5,414,251	5/1995	Durbin .....	235/462	0219695	9/1986	European Pat. Off. .
5,415,839	5/1995	Zaun et al. ....	422/64	0221308	9/1986	European Pat. Off. .
5,422,075	6/1995	Saito et al. ....	422/52	0239382B1	3/1987	European Pat. Off. .
5,424,036	6/1995	Ushikubo .....	422/64	0251087	6/1987	European Pat. Off. .
5,426,976	6/1995	McHardy et al. ....	73/299	0252631A2	6/1987	European Pat. Off. .
5,427,243	6/1995	Roshdy .....	206/438	0252631B1	6/1987	European Pat. Off. .
5,428,470	6/1995	Labriola, II .....	359/119	0253519A2	6/1987	European Pat. Off. .
5,429,330	7/1995	Bond et al. ....	248/61	0253519B1	6/1987	European Pat. Off. .
5,430,957	7/1995	Eigen et al. ....	34/423	0273969B1	7/1987	European Pat. Off. .
5,433,120	7/1995	Boyd et al. ....	73/863	0285654	10/1987	European Pat. Off. .
5,434,051	7/1995	Allard et al. ....	435/7	0285654B1	10/1987	European Pat. Off. .
5,434,083	7/1995	Mitsumaki et al. ....	436/48	0269752	12/1987	European Pat. Off. .
5,437,361	8/1995	Ohmori et al. ....	198/465	0281390	3/1988	European Pat. Off. .
5,437,838	8/1995	DeMoranville et al. ....	422/67	0286119B1	4/1988	European Pat. Off. .
5,437,841	8/1995	Balmer .....	422/102	0299659A2	7/1988	European Pat. Off. .
5,439,645	8/1995	Saralegui et al. ....	422/64	0549573A1	7/1988	European Pat. Off. .
5,439,646	8/1995	Tanimizu et al. ....	422/64	0314525	10/1988	European Pat. Off. .
5,441,895	8/1995	Jakubowicz et al. ....	436/518	0353264B1	11/1988	European Pat. Off. .
5,442,164	8/1995	Adachi .....	235/463	0346878B1	1/1989	European Pat. Off. .
5,443,790	8/1995	Coeurveille et al. ....	422/63	0329183A2	2/1989	European Pat. Off. .
5,443,791	8/1995	Cathcart et al. ....	422/65	0329183B1	2/1989	European Pat. Off. .
5,445,936	8/1995	Piran et al. ....	435/6	0355801B1	8/1989	European Pat. Off. .
5,445,970	8/1995	Rohr .....	436/526	0355802B1	8/1989	European Pat. Off. .
5,445,971	8/1995	Rohr .....	436/526	0355823	8/1989	European Pat. Off. .
5,453,610	9/1995	Gibbons .....	250/207	0355823A2	8/1989	European Pat. Off. .
5,455,175	10/1995	Wittwer et al. ....	435/286	0356250B1	8/1989	European Pat. Off. .
5,455,414	10/1995	Wang .....	235/462	0356883	8/1989	European Pat. Off. .
5,456,360	10/1995	Griffin .....	206/443	0358948A2	8/1989	European Pat. Off. .
5,457,530	10/1995	Nagai .....	356/330	0358948B1	9/1989	European Pat. Off. .
5,458,785	10/1995	Howe et al. ....	210/695	0396657B1	9/1989	European Pat. Off. .
5,458,852	10/1995	Buechler .....	422/58	0371265	10/1989	European Pat. Off. .
5,462,715	10/1995	Koch et al. ....	422/64	0371265B1	10/1989	European Pat. Off. .
5,468,453	11/1995	Holt et al. ....	422/100	0658762A2	2/1990	European Pat. Off. .
5,469,749	11/1995	Shimada et al. ....	73/861	0660115A2	2/1990	European Pat. Off. .
5,483,843	1/1996	Miller et al. ....	73/864	0409126A2	7/1990	European Pat. Off. .
5,501,841	3/1996	Lee et al. ....	422/101	0409606	7/1990	European Pat. Off. .
5,503,036	4/1996	Nguyen et al. ....	73/864	0410645	7/1990	European Pat. Off. .
5,507,193	4/1996	Ishihara .....	73/864	0410645A2	7/1990	European Pat. Off. .
5,519,635	5/1996	Miyake et al. ....	364/497	0410645A3	7/1990	European Pat. Off. .
5,543,112	8/1996	Ghead et al. ....	422/52	0416285B1	8/1990	European Pat. Off. .
5,623,415	4/1997	O'Bryan et al. ....	364/478	0417301A1	9/1990	European Pat. Off. .
B1 4,906,432	6/1991	Geiselman .....	422/63	0435481	12/1990	European Pat. Off. .
				0435481A2	12/1990	European Pat. Off. .
				0436995A2	12/1990	European Pat. Off. .
				0438158B1	1/1991	European Pat. Off. .
				0449321	3/1991	European Pat. Off. .
				0452892A2	4/1991	European Pat. Off. .
				0365569B1	5/1991	European Pat. Off. .
				0467302A2	7/1991	European Pat. Off. .
				0492499A2	12/1991	European Pat. Off. .
				0502638	2/1992	European Pat. Off. .
				0502638A2	2/1992	European Pat. Off. .
				0512358A2	4/1992	European Pat. Off. .
				0512368A3	4/1992	European Pat. Off. .
				0523425	6/1992	European Pat. Off. .
				0528708A1	7/1992	European Pat. Off. .
				0596987B1	7/1992	European Pat. Off. .
				0597017B1	7/1992	European Pat. Off. .
				0544578A1	11/1992	European Pat. Off. .
				0571716A1	2/1993	European Pat. Off. .
				0572185A2	5/1993	European Pat. Off. .
				0572185A3	5/1993	European Pat. Off. .
				0572217A1	5/1993	European Pat. Off. .
				0596205A2	8/1993	European Pat. Off. .
				0616208A1	3/1994	European Pat. Off. .
				0637750A2	7/1994	European Pat. Off. .
				0657382A2	7/1994	European Pat. Off. .
				0638806	8/1994	European Pat. Off. .
				0638806A3	8/1994	European Pat. Off. .
				0664501A1	8/1994	European Pat. Off. .
				0643306A2	9/1994	European Pat. Off. .
				0645631A2	9/1994	European Pat. Off. .
				0653720A2	11/1994	European Pat. Off. .
				0661535	12/1994	European Pat. Off. .
				0670483A3	2/1995	European Pat. Off. .

## FOREIGN PATENT DOCUMENTS

0030087	11/1980	European Pat. Off. .
0038181	4/1981	European Pat. Off. .
0106536A2	9/1982	European Pat. Off. .
0078948	10/1982	European Pat. Off. .
0080108	11/1982	European Pat. Off. .
0080109	11/1982	European Pat. Off. .
0087786	2/1983	European Pat. Off. .
0097591	6/1983	European Pat. Off. .
0097591A1	6/1983	European Pat. Off. .
0097591B1	6/1983	European Pat. Off. .
0100663	7/1983	European Pat. Off. .
0101192	7/1983	European Pat. Off. .
0102661	8/1983	European Pat. Off. .
0106662	10/1983	European Pat. Off. .
0125995	5/1984	European Pat. Off. .
0144006B1	11/1984	European Pat. Off. .
0148166	1/1985	European Pat. Off. .
0149565	1/1985	European Pat. Off. .
0149565A2	1/1985	European Pat. Off. .
0149565B1	1/1985	European Pat. Off. .
0152964	2/1985	European Pat. Off. .
0152964A2	2/1985	European Pat. Off. .
0167834	6/1985	European Pat. Off. .
0169434	7/1985	European Pat. Off. .
0169434A2	7/1985	European Pat. Off. .
0169434B1	7/1985	European Pat. Off. .
0189280	1/1986	European Pat. Off. .
0198413	4/1986	European Pat. Off. .



0682258A1	2/1995	European Pat. Off.
0672906A1	3/1995	European Pat. Off.
0692308A2	7/1995	European Pat. Off.
0712000	9/1995	European Pat. Off.
0712000A2	9/1995	European Pat. Off.
1573224	7/1969	France
2309869	4/1976	France
2523320	3/1983	France
2655426	12/1990	France
51272	3/1890	Germany
92212	6/1897	Germany
130053	4/1902	Germany
338227	8/1918	Germany
1428777	3/1973	Germany
2505268	4/1975	Germany
3244508	7/1984	Germany
3621831	1/1988	Germany
3926462A1	2/1991	Germany
1322728	5/1969	Japan
60-188849	9/1985	Japan
62-49070	3/1987	Japan
62-165057	7/1987	Japan
62-194464	8/1987	Japan
2-210266	8/1990	Japan
4047266	2/1992	Japan
123637	8/1964	United Kingdom
1180957	12/1967	United Kingdom
1473042	11/1977	United Kingdom
1566098	4/1980	United Kingdom
1592297	1/1981	United Kingdom
1592299	1/1981	United Kingdom
2202814	5/1988	United Kingdom
2199407	7/1988	United Kingdom
2228730	5/1990	United Kingdom
2239093	6/1991	United Kingdom
WO 80/02280	10/1980	WIPO
WO 86/00139	7/1986	WIPO
WO 86/05518	9/1986	WIPO
WO 87/03966	7/1987	WIPO
WO 87/07727	12/1987	WIPO
WO 88/02866	4/1988	WIPO
WO 89/04373	5/1989	WIPO
WO 91/07662	11/1989	WIPO
WO 90/00252	1/1990	WIPO
WO 90/01168	2/1990	WIPO
WO 90/05903	5/1990	WIPO
WO 90/11511	10/1990	WIPO
WO 90/05411	5/1991	WIPO
WO 91/13335	9/1991	WIPO
WO 91/15768	10/1991	WIPO
WO 92/05448	4/1992	WIPO
WO 92/12255	7/1992	WIPO
WO 92/16841	10/1992	WIPO
WO 92/16844	10/1992	WIPO
WO 92/20449	11/1992	WIPO
WO 92/22201	12/1992	WIPO
WO 83.01308	1/1993	WIPO
WO 93/01308	1/1993	WIPO
WO 93/02364	2/1993	WIPO
WO 93/03383	2/1993	WIPO
WO 93/12430	6/1993	WIPO
WO 93/12431	6/1993	WIPO
WO 93/16801	9/1993	WIPO
WO 93/22686	11/1993	WIPO
WO 94/04929	3/1994	WIPO
WO 94/19451	9/1994	WIPO
WO 95/00829	1/1995	WIPO
WO 95/03548	2/1995	WIPO

## OTHER PUBLICATIONS

Yolken; Enzyme Immunoassays for the Detection of Infectious Antigens in Body Fluids: Current Limitations and Future Prospects; Reviews of Infectious Diseases vol. 4, No. 1 Jan.-Feb. 1982.

Hirschbein, et al; Magnetic separations in chemistry and biochemistry; Chemtech Mar. 1982.

Ollerich; Enzyme-Immunoassay: A Review J. Clin Chem Clin Biochem vol. 22, 1984 pp. 895-904.

Guesdon, et al.; Magnetic enzyme immunoassay for measuring human IgE; J. Allergy Clin Immunol Jan. 78; vol. 61, No. 1, 23-27.

Kamel, et al; Magnetizable Solid-Phase Fluoroimmunoassay of Phenytoin in Disposable Test Tubes; Clin chem 26/9/1281-1284 (1980).

Klibanov; Immobilized Enzymes and Cells as Practical Catalysts; Science vol. 219.

Mosbach, et al; Magnetic ferrofluids for preparation of magnetic polymers and their application in affinity chromatography; Nature vol. 270 Nov. 17, 1977.

Dawes, et al.; Radioimmunoassay of Digoxin Employing Charcoal Entrapped in Magnetic Polyacrylamide Particles; Clinica Chimica Acta 86 1978; 353-356.

Nye, et al., Solide Phase, Magnetic Particle Radioimmunoassay; Clinica Chimica Acta 69(1976) 387-396.

Hersh, et al; Magnetic Solid Phase Radioimmunoassay; Clinica chimica Acta, 63 1975 69-72.

Pourfarzaneh, et al., Cortisol Directly Determined in Serum by Fluoroimmunoassay with Magnetizable Solid Phase; Clin Chem 26/6 730-733.

Kamel, et al., Nove 125I-Labeled Nortriptyline Derivatives and Their Use in Liquid Phase or Magnetizable Solid Phase Second-Antibody Radioimmunoassays; Clin Chem 25/12, 1997-2002 (1979).

Ithakissios, et al., Use of Protein Containing Magnetic Micro-particles in Radioassays; Clin chem 23/11; 2072-1079 (1977).

Robinson; The Properties of Magnetic Supports in Relation to Immobilized Enzyme Reactors; Biotech and Bioeng. vol. XV 1973.

Carter; Preparation of Ligand Free Human Serum for Radioimmunoassay by Adsorption on Activated Charcoal; Clin Chem 24/2, 362-364(1978).

Jacobs; Separation Methods in Immunoassays; The Ligand Quarterly vol. 4, No. 4, 1981.

Rembaum et al., Synthesis and Reactions of Hydrophilic Functional Microspheres; J. Macromol Sci Chem A13(5) 603-632 (1979).

Pourfarzaneh, et al, Production and Use of Magnetizable Particles in Immunoassay, The Ligand Quarterly, vol. 5, No. 1, 1982.

Kaiser, et al., Magnetic Properties of Stable Dispersions of subdomain Magnetite Particles; Journal of Applied Physics; vol. 41, No. 3, Mar. 1970.

Halling et al., Magnetic supports for immobilized enzymes and bioaffinity adsorbents; Enzyme Microb Technol. 1980 vol. 2, Jan.

Allman, et al., Fluoroimmunoassay of Progesterone in Human Serum or Plasma; Clin Chem 27/7 1176-1179-(1981).

Radioimmunoassay and Related Procedures in Medicine 1982.

Klingler, et al., Immunoassay of unconjugated estriol in serum of pregnant women monitored by chemiluminescence; Steroids, vol. 42, No. 2, Aug. 1983.

Wisdom; Enzyme Immunoassay; Clin Chem 22/8/ 1243-1255 (1976).

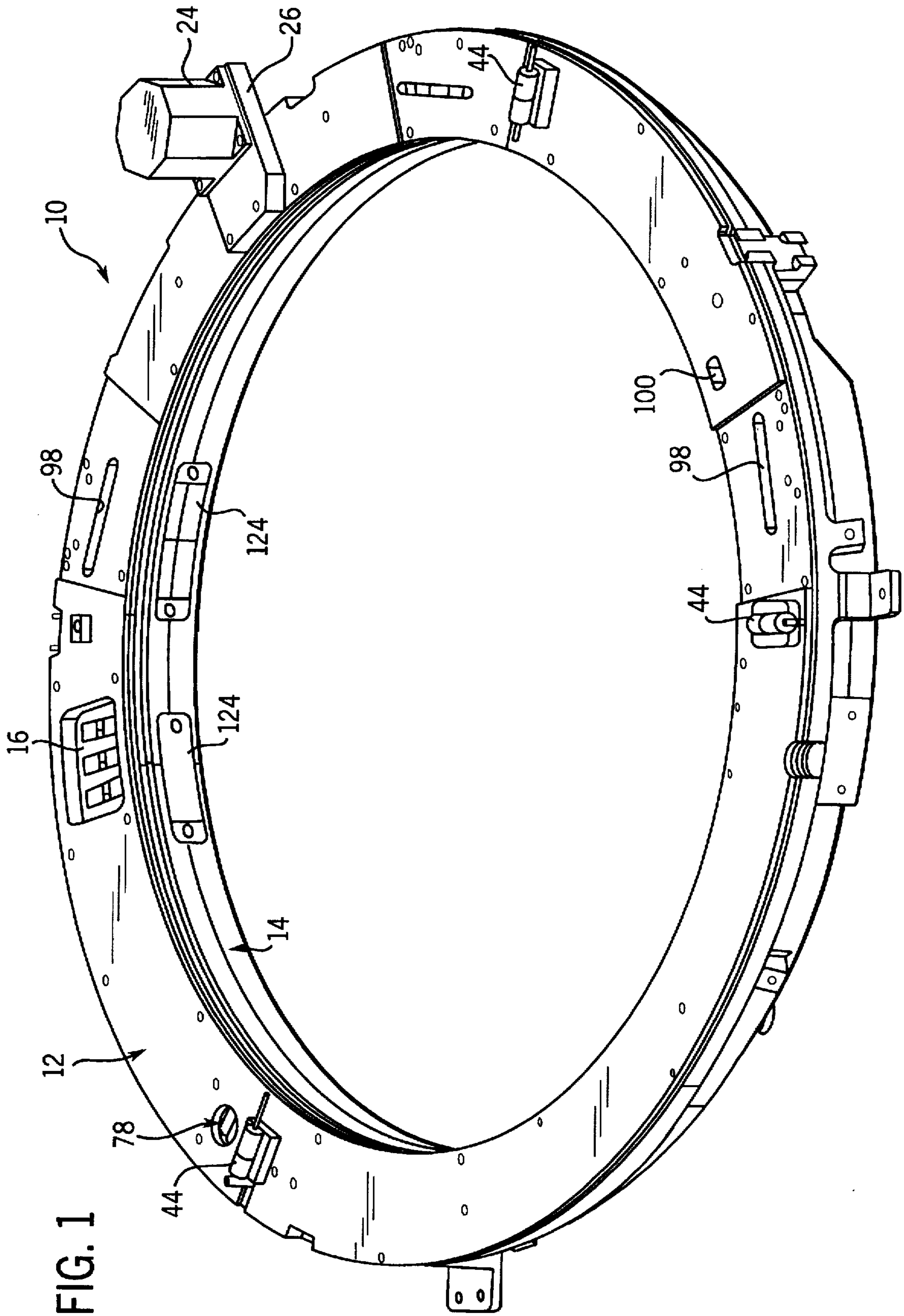
Scholmerich, et al., Bioluminescence and Chemiluminescence; New Perspectives.

Thorpe et al., [29]Enhanced Chemiluminescent Reactions Catalyzed by Horseradish Peroxidase; Method in Enzymology vol. 133.



- Margel et al., Polyglutaraldehyde: A new reagent for coupling proteins to microspheres and for labeling cell surface receptors . . . ; *Journal of Immunological Methods* 28(1979) 341-353.
- Margel; Polyaldehyde Microspheres as Probes for Cell Membranes; *Ind. Eng. Chem. Prod. Res. Dev.* 1982, 21, 343-348.
- Sovetskaia Meditsina No Translation.
- Margel, et al., Polyacrolein Microspheres as a New Tool in Cell Biology, *J. Cell Sci.* 56, 157-175 (1982).
- Margel, et al., Cell Fractionation with Affinity Ligands Conjugated to Agarose-Polyacrolein Microsphere Beads; *J. Cell Sci.* 62, 149-159(1983).
- Kreuter; Evaluation of Nanoparticles as Drug-Delivery Systems I: Preparation Methods. *Pharm Acta Helv.* 58 No. 7, 1983.
- Margel, et al., Chelation of Mercury by Polymercaptal Microspheres: new Potential Antidote for Mercury Poisoning; *Journ of Pharm. Sciences*; vol. 71. No. 9, Sep. 1982.
- Basch, et al., Cell Separation Using Positive Immunoselective Techniques. *Journ. of Immunol. Methods*, 56(1983) 269-280.
- Kaplan, et al., The Selective Detection of Cell Surface Determinants by means of Antibodies and Acetylated Avidin Attached tooHighly Fluorescent Polymer Microspheres; *Biochem and Biophysic Acta* 728(1983)112-120.
- Margel, et al., Novel Effective Immunoabsorbents Based on Agarose-Polyaldehyde Microsphere Beads: Synthesis and Affinity Chromatography, *Analytical Biochem.* 128 342-350(1983).
- Marcus, et al., A New Immunoabsorbent for Hemoperfusion: Agarose-Polyacrolein Microspheres Beads; *Biomat Med. Dev., Art. Org.* 10(3) 157-171 (1982).
- Merivuori, et al., Cell Labelling and separation with polyglutaraldehyde microspheres; *Exp Cell Res* 130 (1980).
- Kempner, et al., Electrophoretic cell separation using polyacrolein microspheres, *Electrophoresis* 1982, 3, 109-113.
- Kumakura et al., Polymeric Microspheres for Immunoresearch; *Immunological communications* 13(2), 119-125(1984).
- Morimoto et al., Dispersion State of Protein stabilized Magnetic Emulsions; *Chem Pharm Bull* 30(8) 3024-3027(1982).
- Boland et al., The Ciba Corning ACS:180 Benchtop Immunoassay Analyzer; *Clin Chem* 36/9, 1598-1602 (1990).
- ACS:180 Automated Chemiluminescence System.
- Rembaum et al., Immunomicrospheres: Reagents for Cell Labeling and Separation, *Science* vol. 208. 25 Apr. 1980.
- Tokes et al., Synthesis of adriamycin coupled polyglutaraldehyde microspheres and evaluation of their cytostatic activity; *Proc Natl. Acad Sci. USA* vol. 79; 2026-2030 Mar. 1982.
- Margel et al., Synthesis and Characterization of Poly(glutaraldehyde). A Potential Reagent for Protein Immobilization and Cell Separation; *Macromolucule* vol. 13, No. 1, Jan. Feb. 1980.
- Margel; Characterization and Chemistry of Polyaldehyde Microspheres; *Journ of Polymer Science* vol. 22, 3521-3522(1984).
- Marcus et al., Extracorporeal removal of specific antibodies by hemoperfusion through the immunosorbent agarose-polyacrolein microsphere beads: Remaoval of Anti-bovine serum albumin in animals; *Journ of Viomed Materials Research* vol. 18 1153-1167(1984).
- Margel; Agarose polyacrolein microsphere beads; *Febs Letter*; vol. 145 No 2; Aug. 1982.
- Hage et al., High performance immunoaffinity chromatography and Chemilumiscent Detection in the Automation of a Parathyroid Hormone Sandwich Immunoassay; *Anal. Chem* 1991, 63, 586-595.
- Bronstein et al., Instrumentation for luminescent assays; *ACL*: 33.
- Campbell, *Chemiluminescence Principles and Applications in Biology and Medicine.*
- The Ideal chemiluminometer. Sec. 2.4.
- Chemiluminescence Immunoassay Sec. 8.2.
- O'Brien et al., The magic lite system and acridinium ester-based immunoassays; *Immunoassay Automation: A practical guide.*
- Dudley; *Chemiluminescence Immunoassay: An Alterman-tive to RIA; Laboratory Medicine* vol. 21, No. 4, Apr. 1990.
- Dyke; *Luminescence Immunoassay and Molecular Applications.*
- Luminescence Immunopassay and Molecular Applications*; pp. 69-75.
- Luminescence Immunopassay and Molecular applications*; pp. 149-156.
- Woodhead et al., Magic Lite Design and Development, *Journ of Bioluminescence* vol. 4, 611-614 (1989).
- Express. New Intellegent random access cheimistry analysis Circle No. 578.
- Shamberger et al., Evaluation of bichromatic random access analyzer; *Analyzer Oct.* 1989, 22.
- The Lancet jan. 14, 1984, vol. 1 1984.
- 550 EXPRESS Shemistry Analyzer.
- T3 Uptake [125I] Radioassay Magic Ciba Corning.
- Quality Value. Customer Satisfaction Ciba Corning Diagnostics Corp.
- Corning Magic Lite II.
- ACS-180.
- Berry et al., A Laboratory and Clinical Evaluation of an Immunochemiluminometric Assay of Thyrotropin in Serum; *Clin Chem.* vol. 34. No. 10, 1988.
- Weeks et al., Chemiluminescence immunoassay: an overview; *Clin Science* 1986 70.403-408.
- Weeks et al., Acridinium Esters as High-Specific-Activity Labels in Immunoassay; *Clin Chem* 29/8, 1474-1479(1983).
- Shridi et al., A direct fluoroimmunoassay for conjugated chenodeoxycholic acid using antibody coupling to magnetisable particles; *Ann Clin Biochem* 1980; 17:188-191.
- Sidki et al., Direct determination of primidone in serum of plasma by a magnetisable solid-phase fluoroimmunoassay; *Ann Clin Biochem* 1983:20:227-232.
- Abdulla et al., Development of a Magnetisable solid-phase fluoroimmunoassay for primaquine and carboxyprimaquine; *Southeast Asian J. Trop Med. Pub Hlth.* vol. 20, No. 3, 1989.
- Ciba Corning Laboratory Diagnostic Reagents and Systems packet.
- Patent Abstracts of Japan vol. 16 No. 225(p. 1360).
- Symbol: A New Communications Medium fo the information Age.
- European Search Report ep 92 30 1483.
- The Future of Bar Coding.
- Symbol Technologies: The Bar Code Data Capture Company.
- Symbol Just What the Doctors Ordered: Met Path Boosts Qualigy, Beats the Competition with PDF41.
- Symbol: A PDF417 Primer: A guide to understanding second generation bar codes and portable data files.







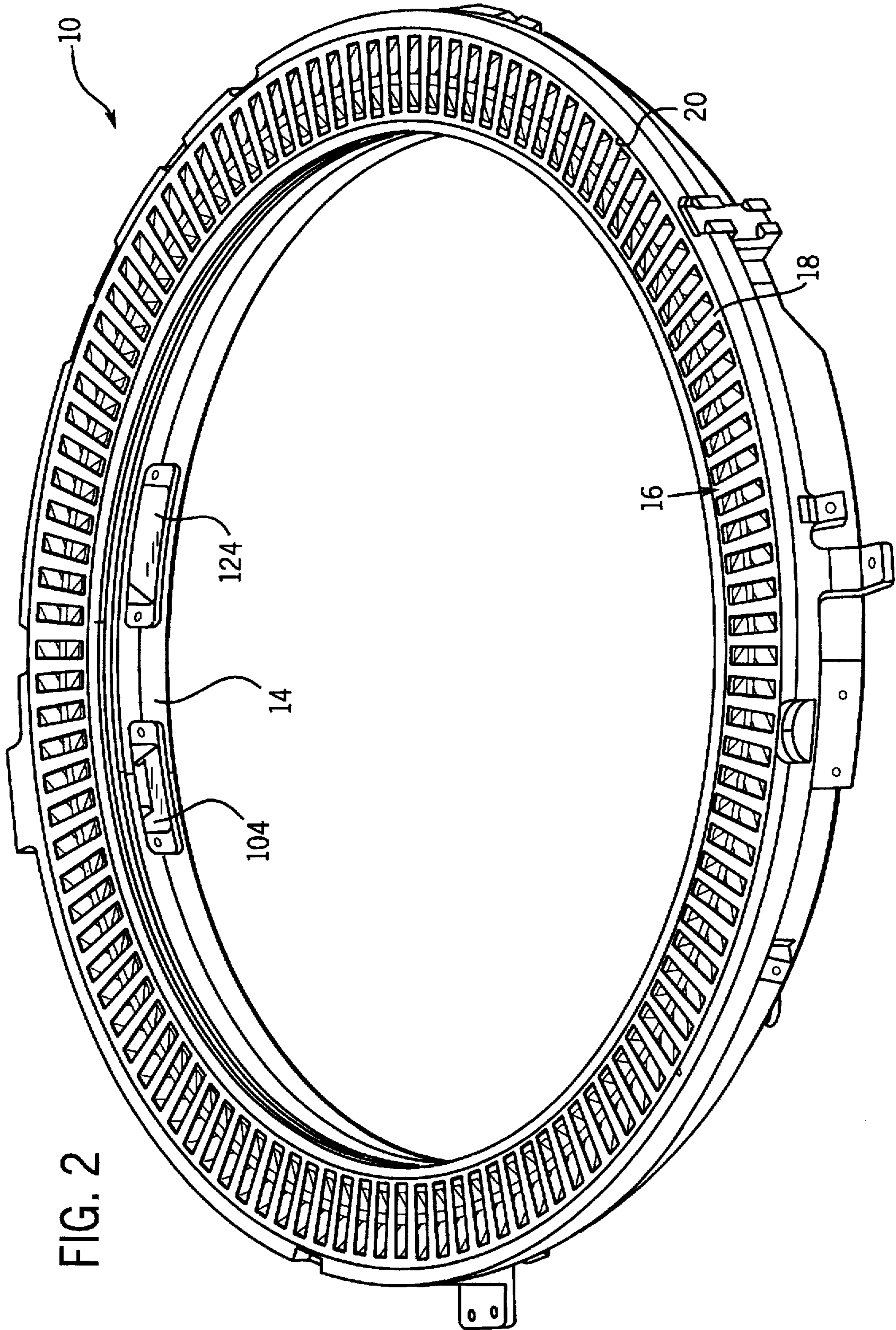


FIG. 2



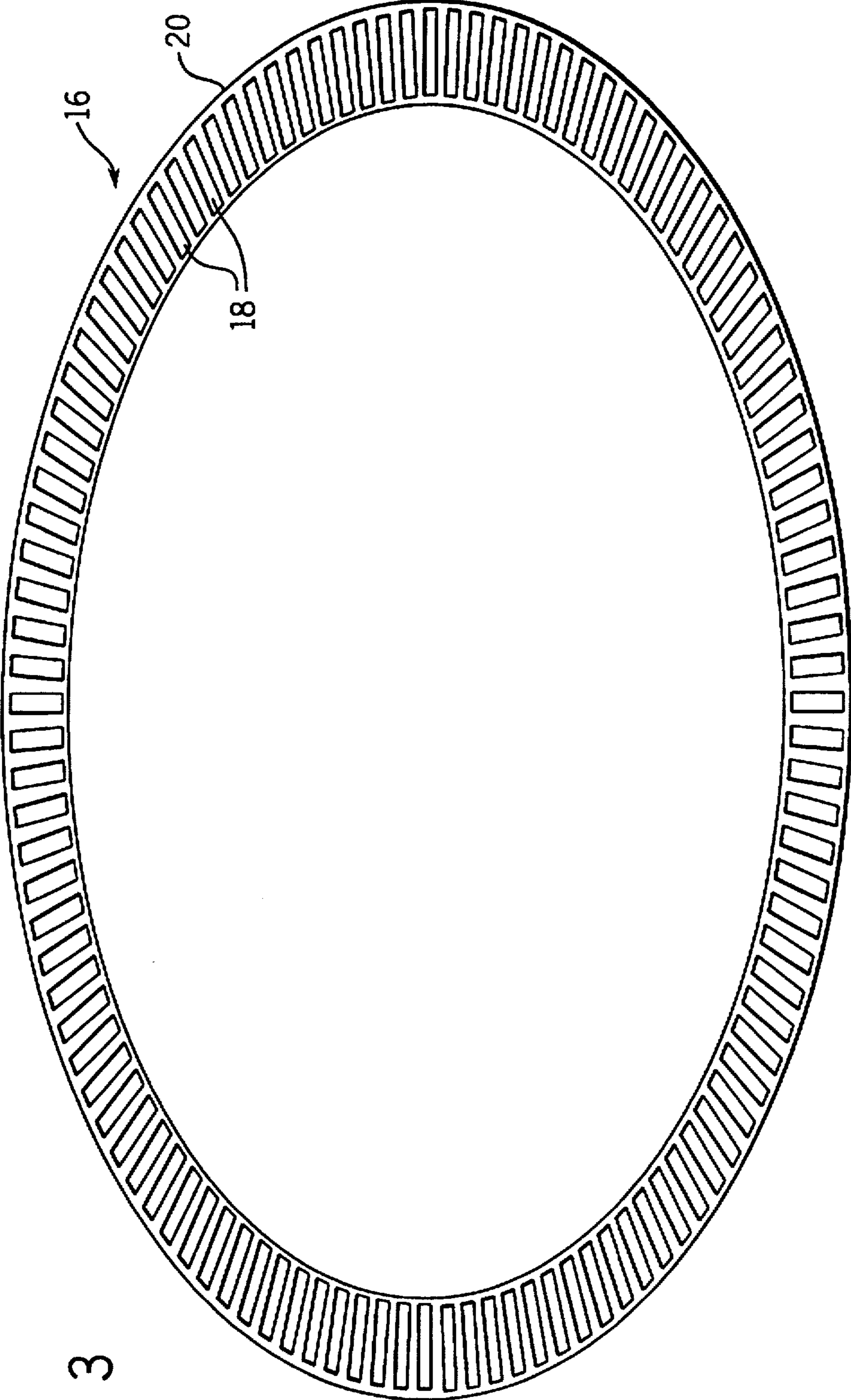


FIG. 3



FIG. 4

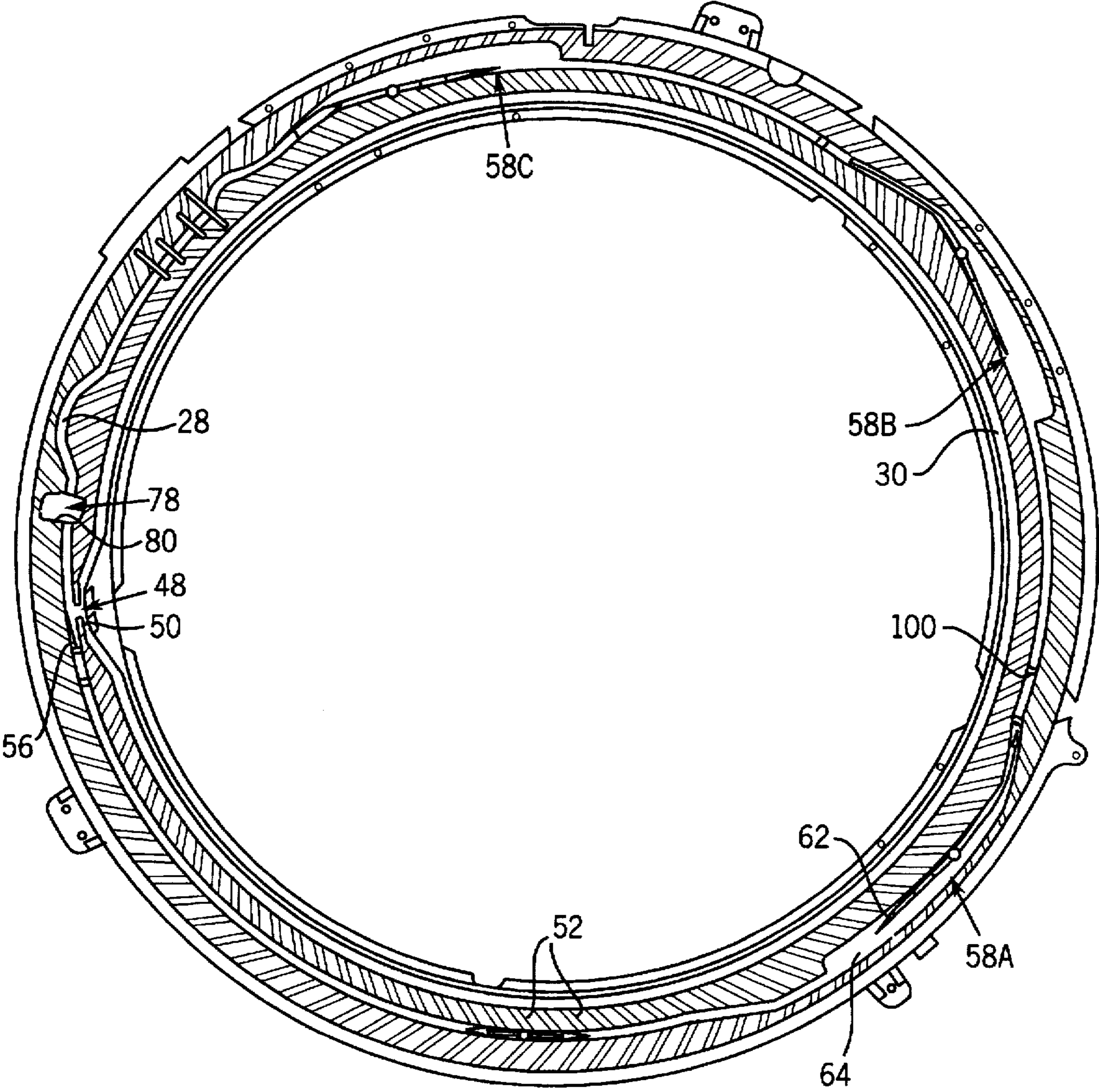




FIG. 5A

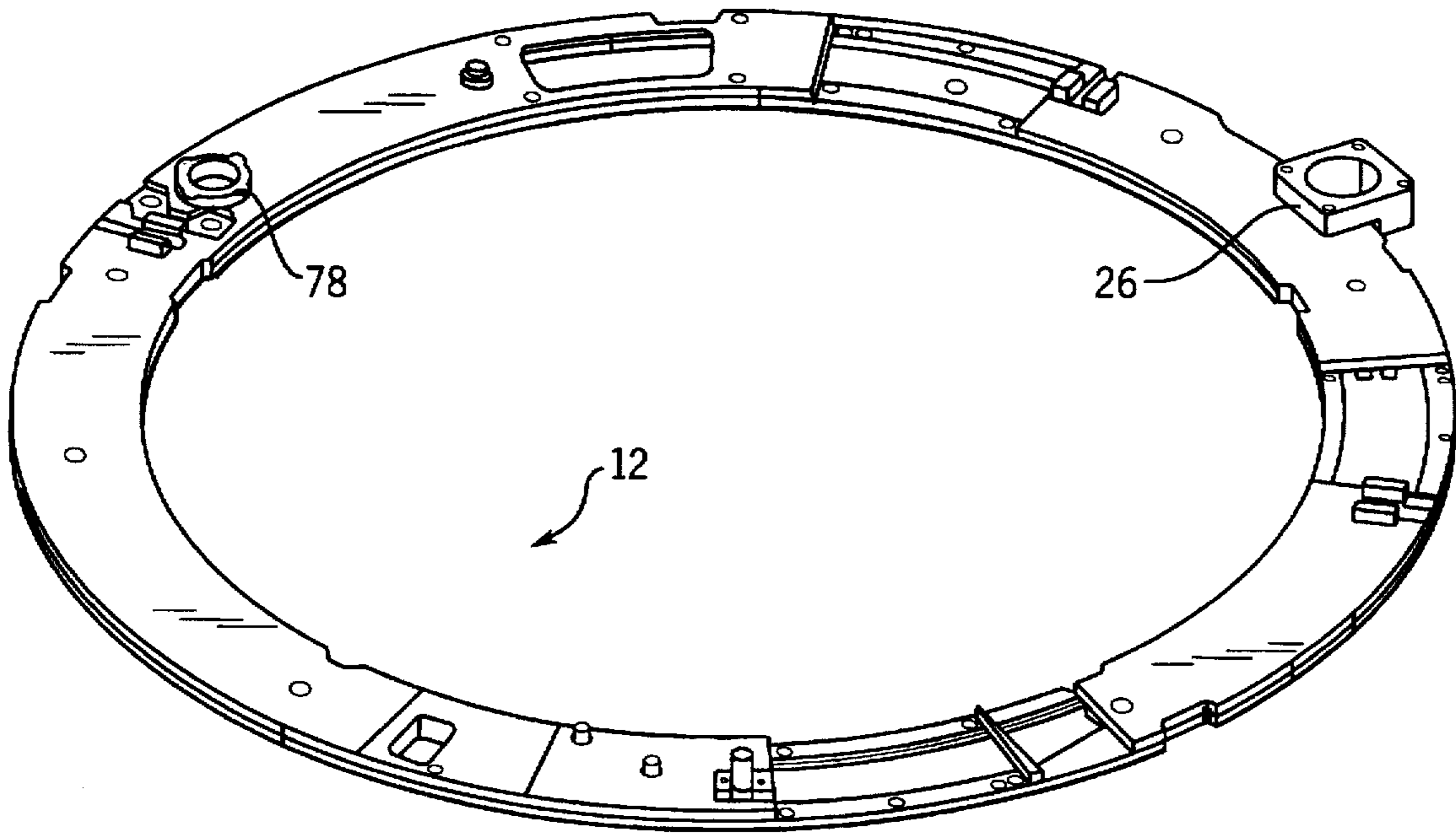
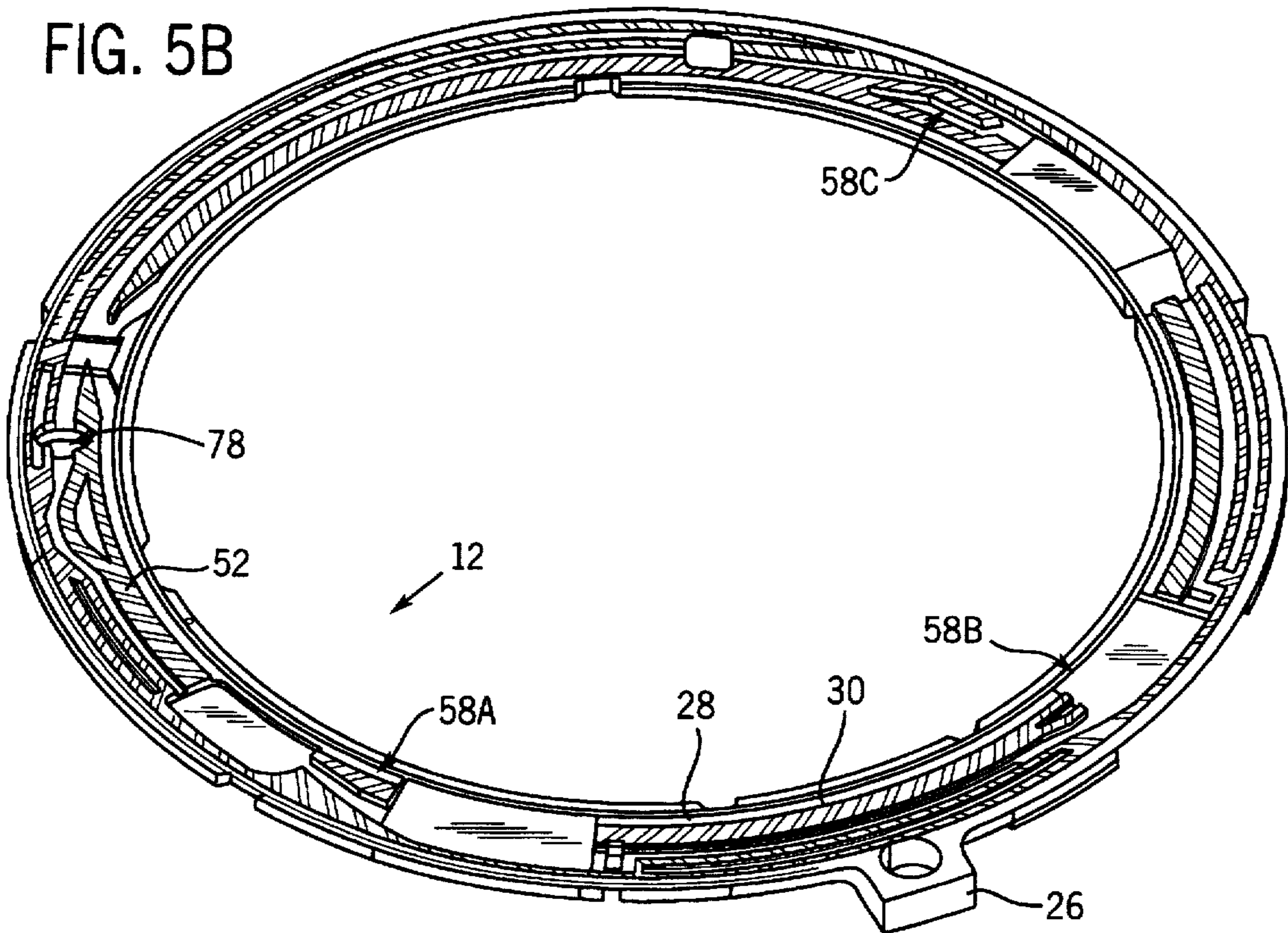


FIG. 5B





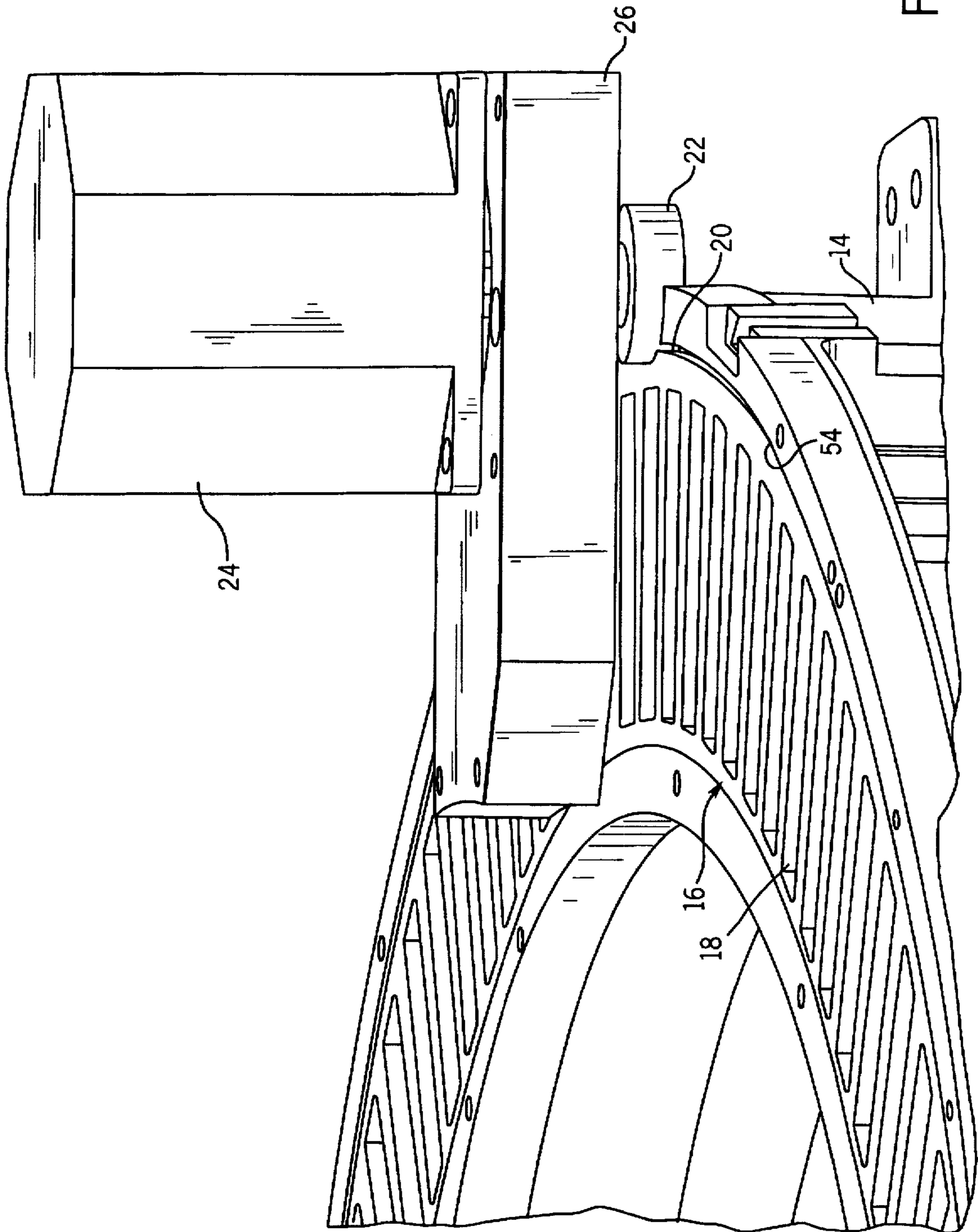
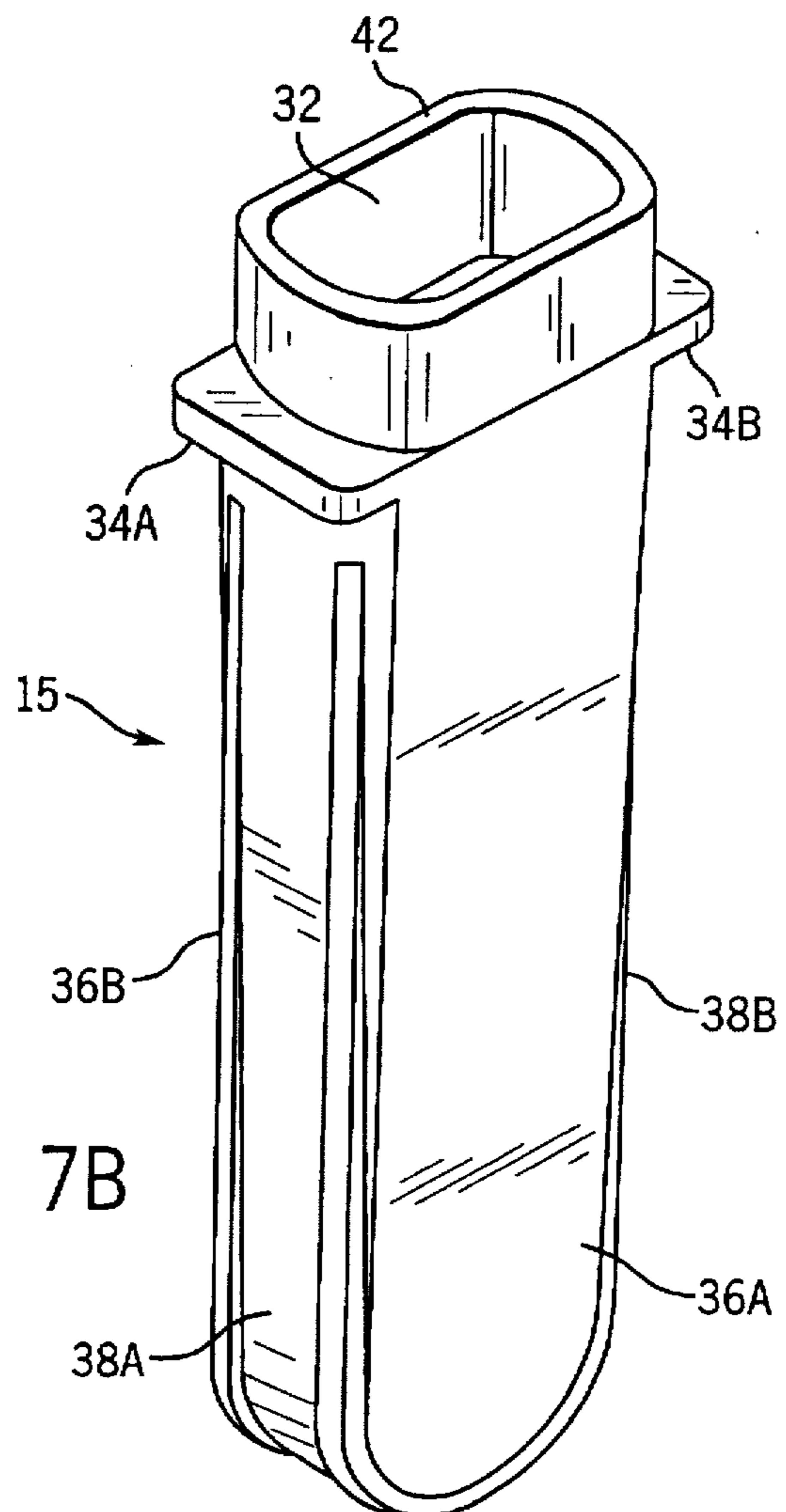
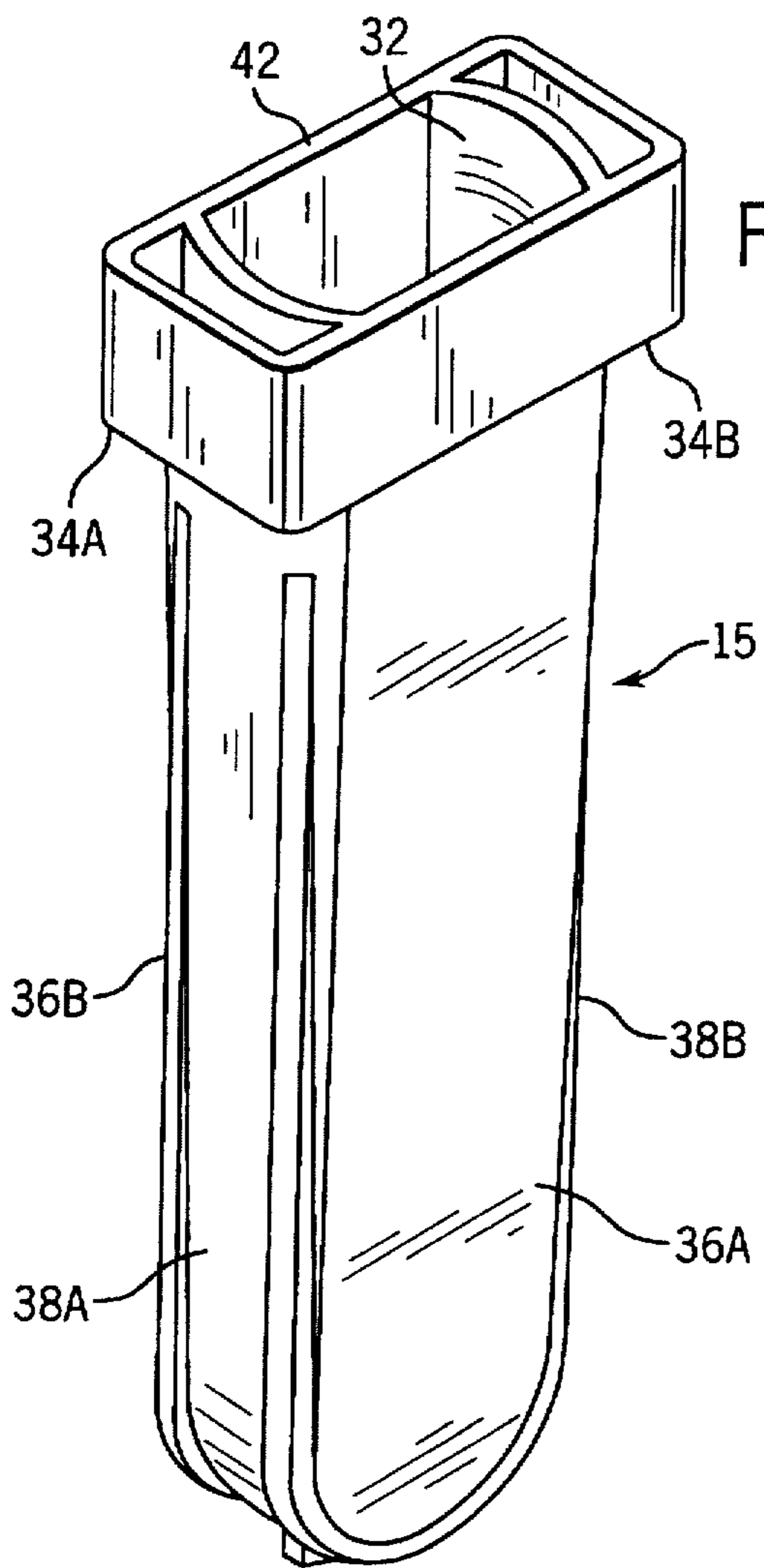


FIG. 6







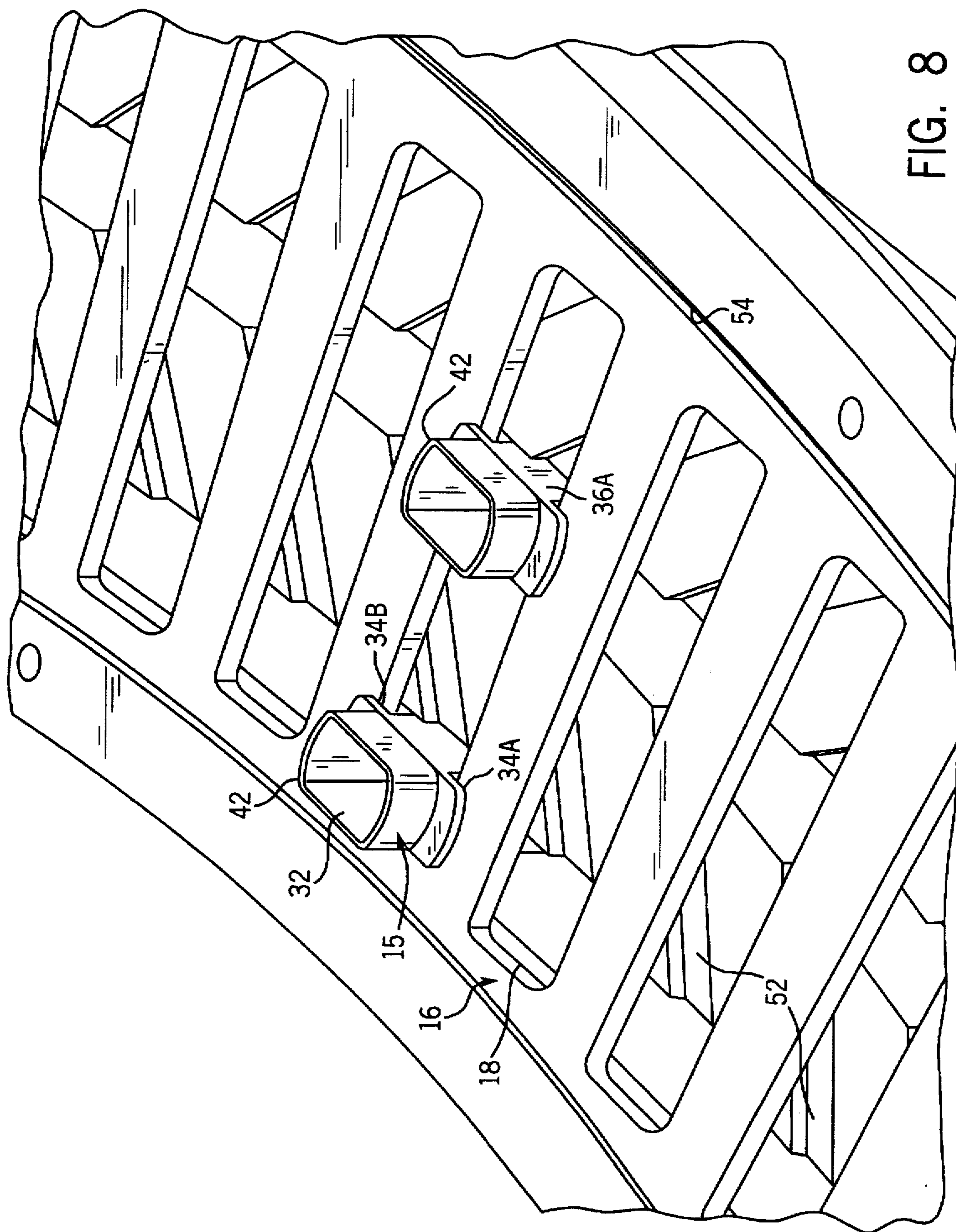


FIG. 8



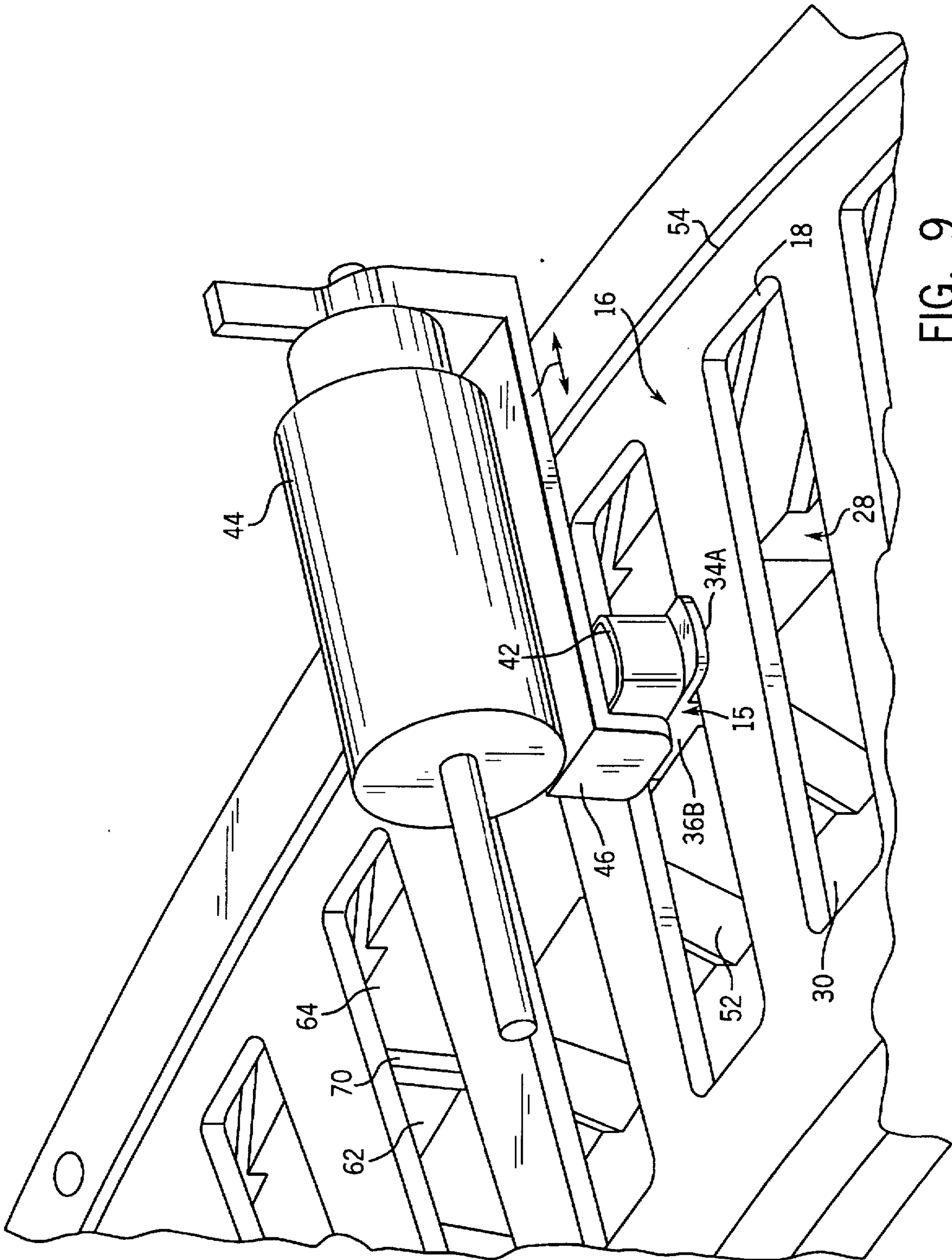


FIG. 9



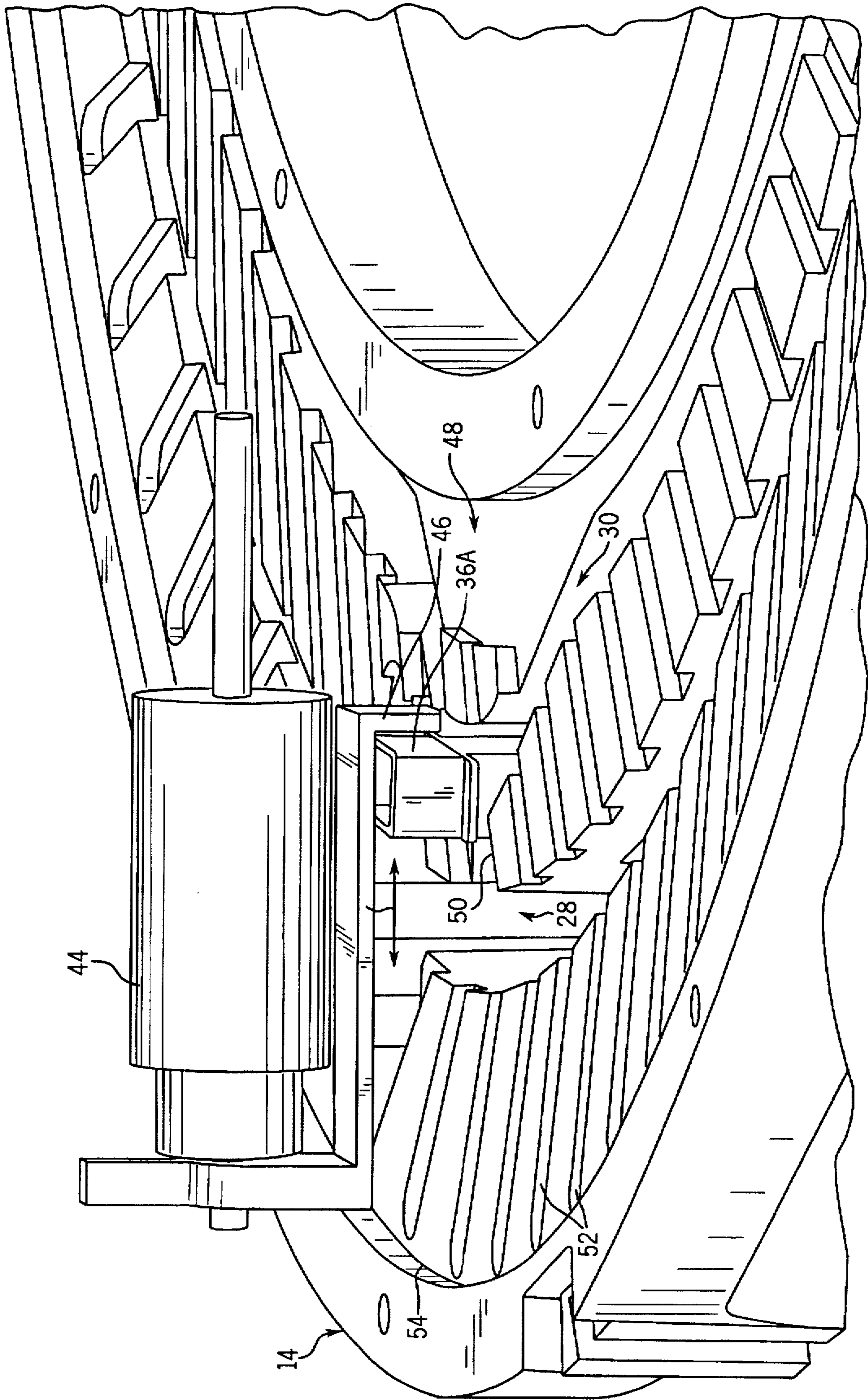


FIG. 10



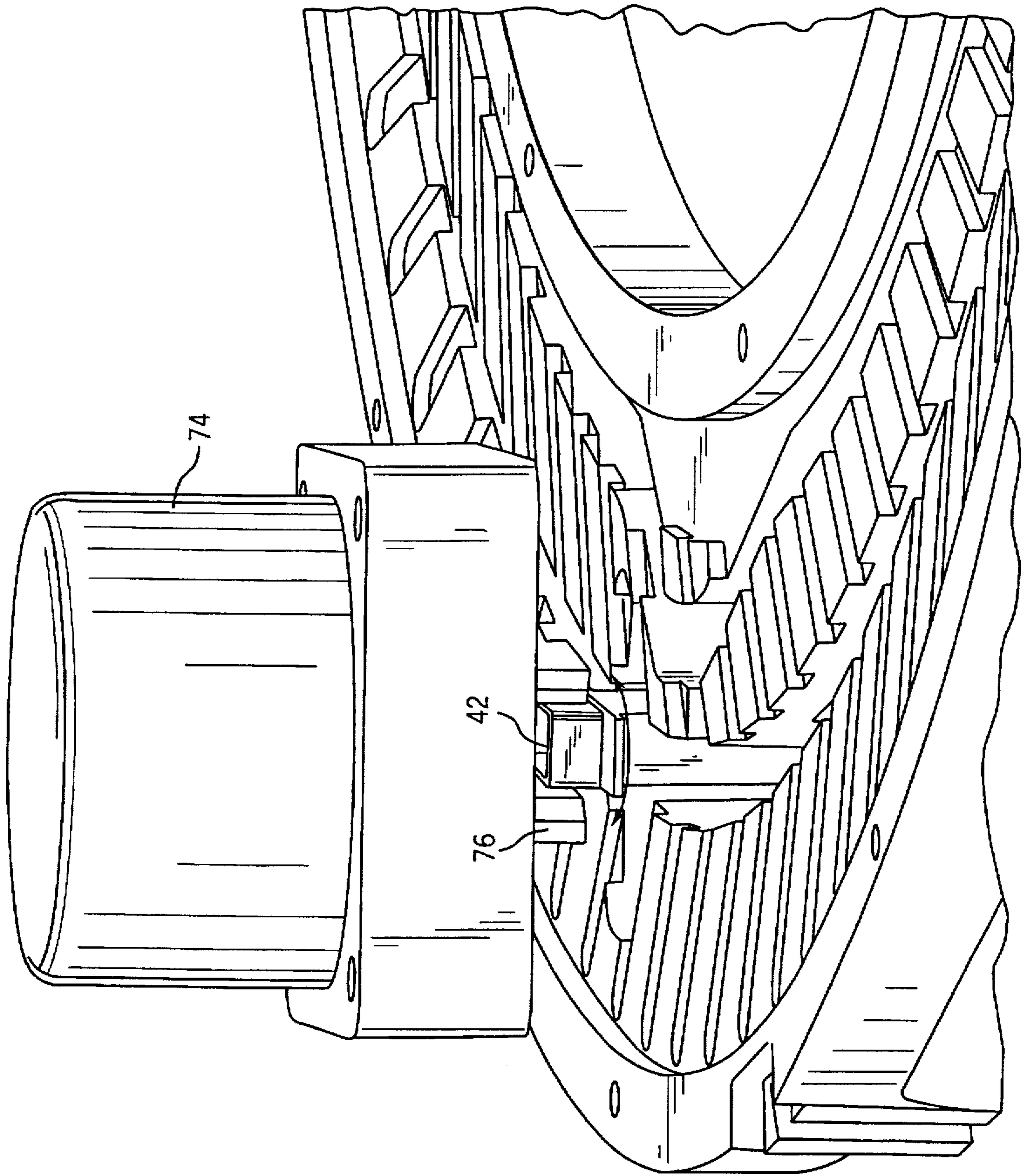


FIG. 11



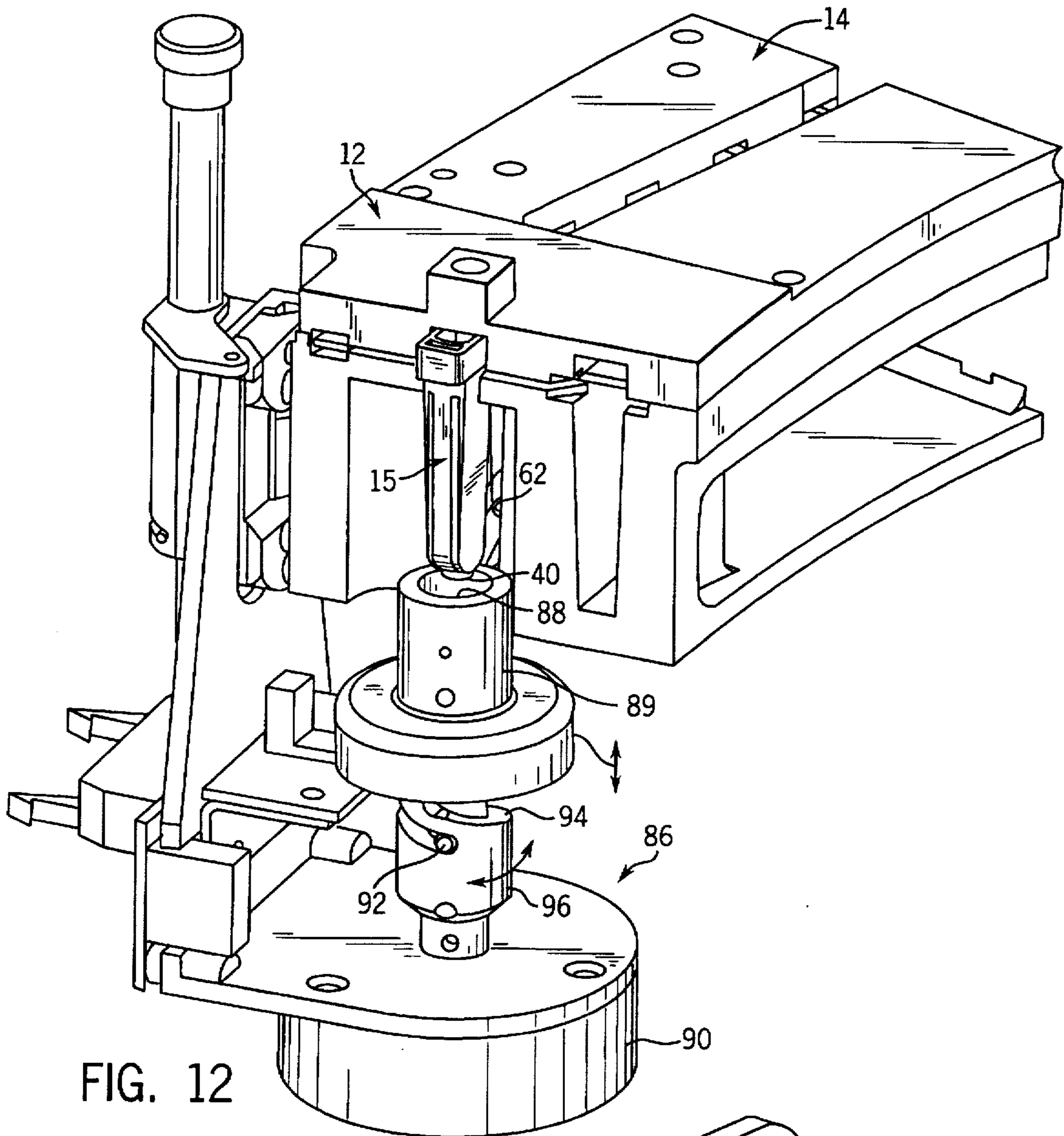


FIG. 12

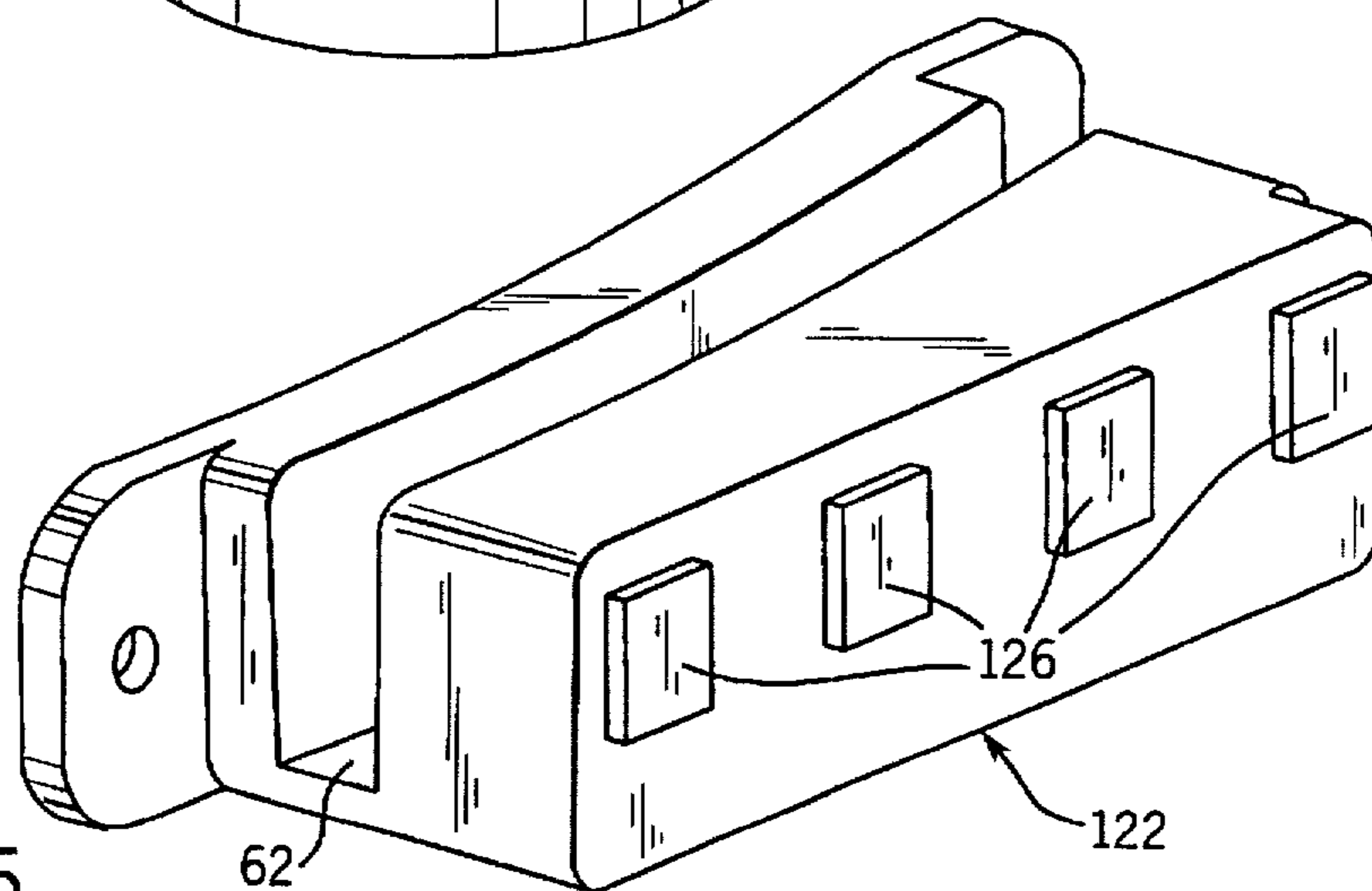


FIG. 15



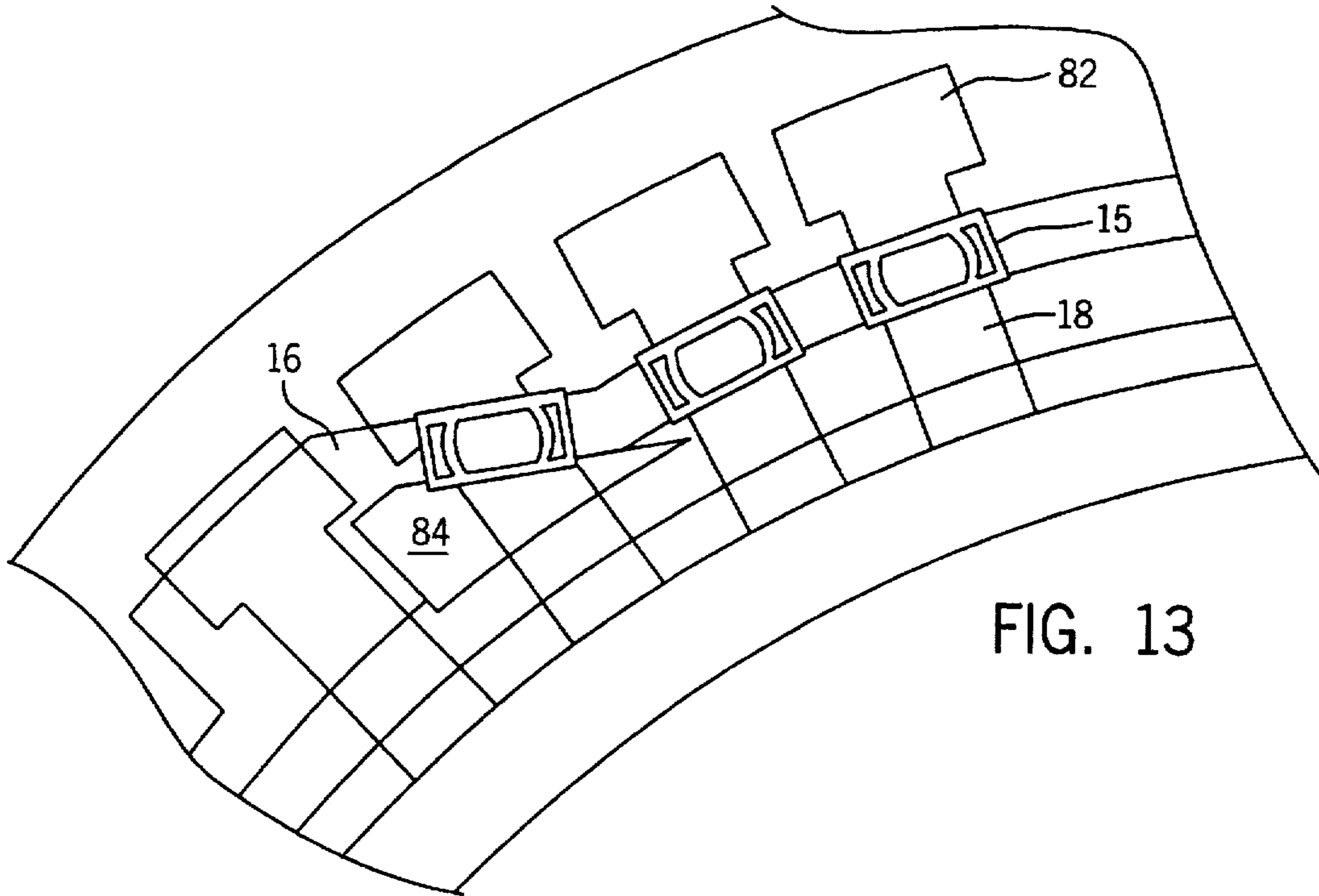


FIG. 13

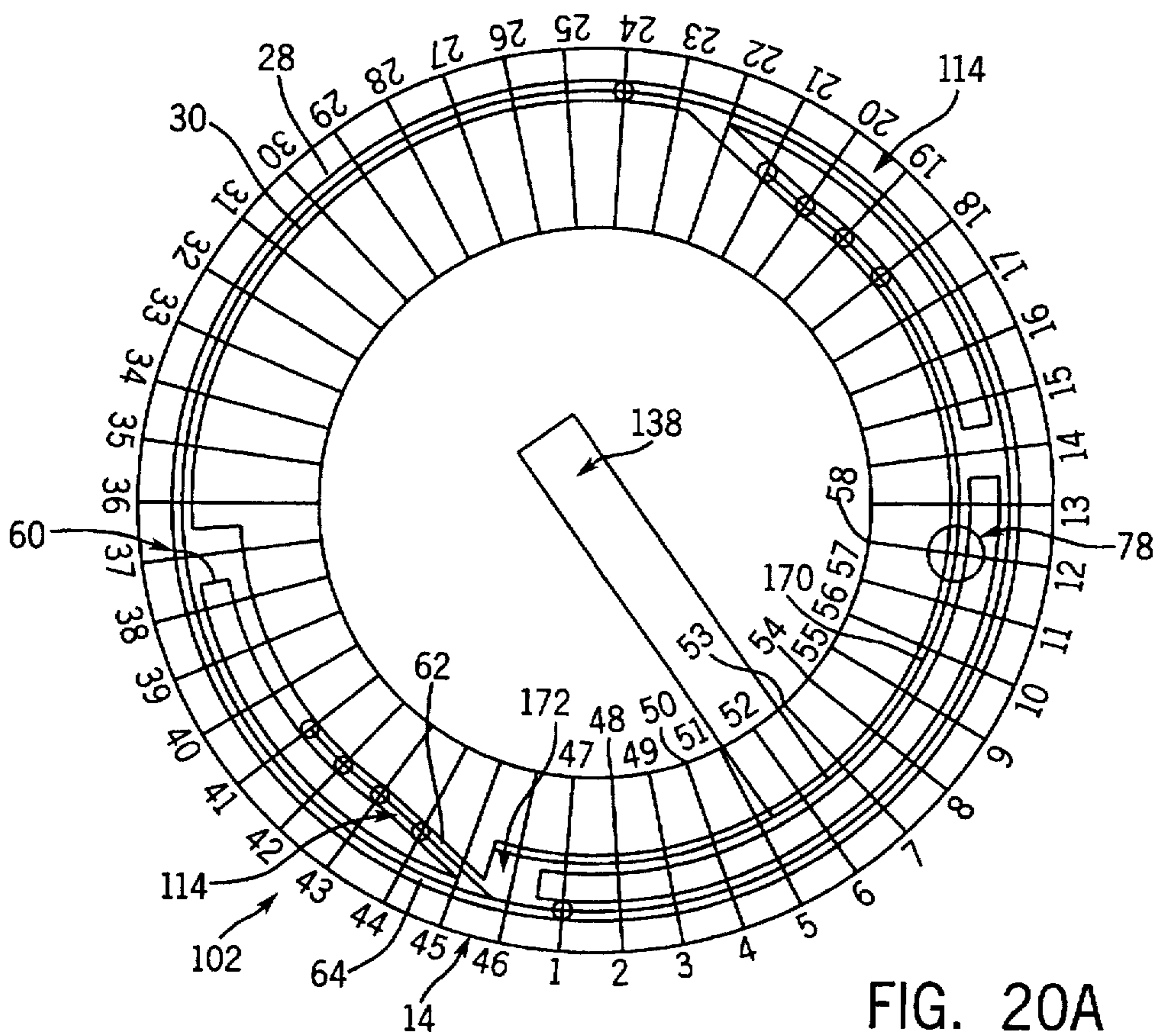
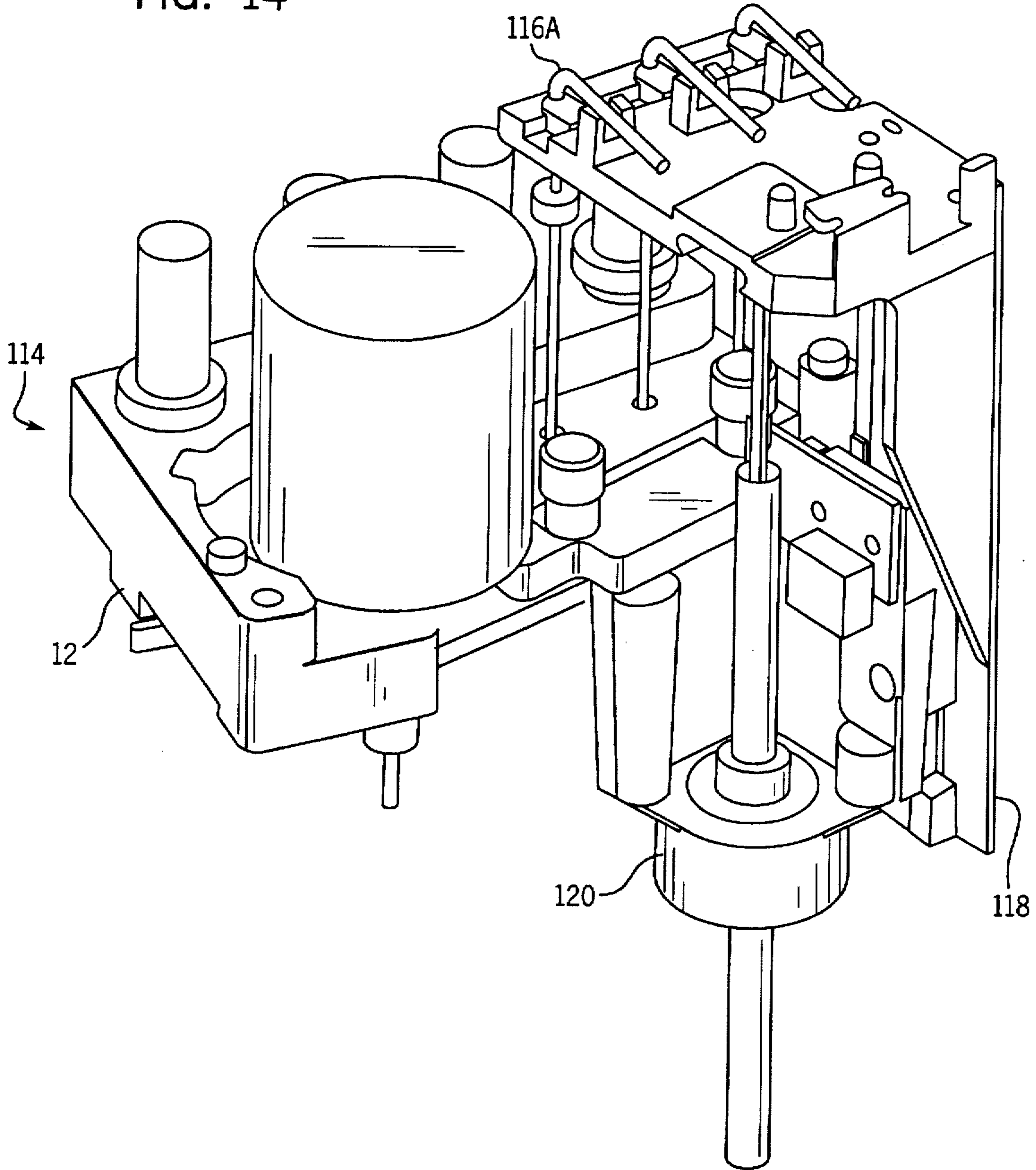


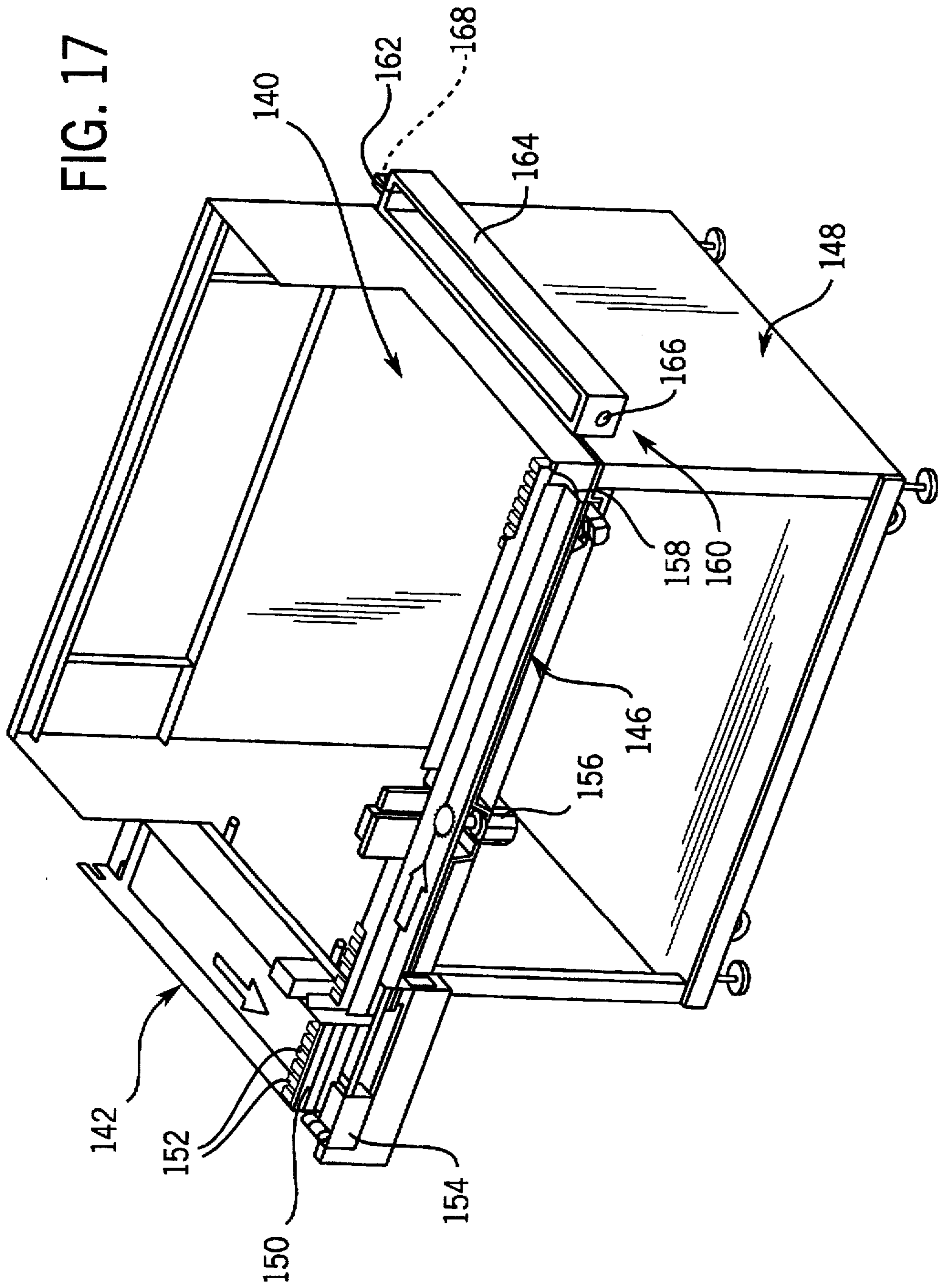
FIG. 20A

FIG. 14

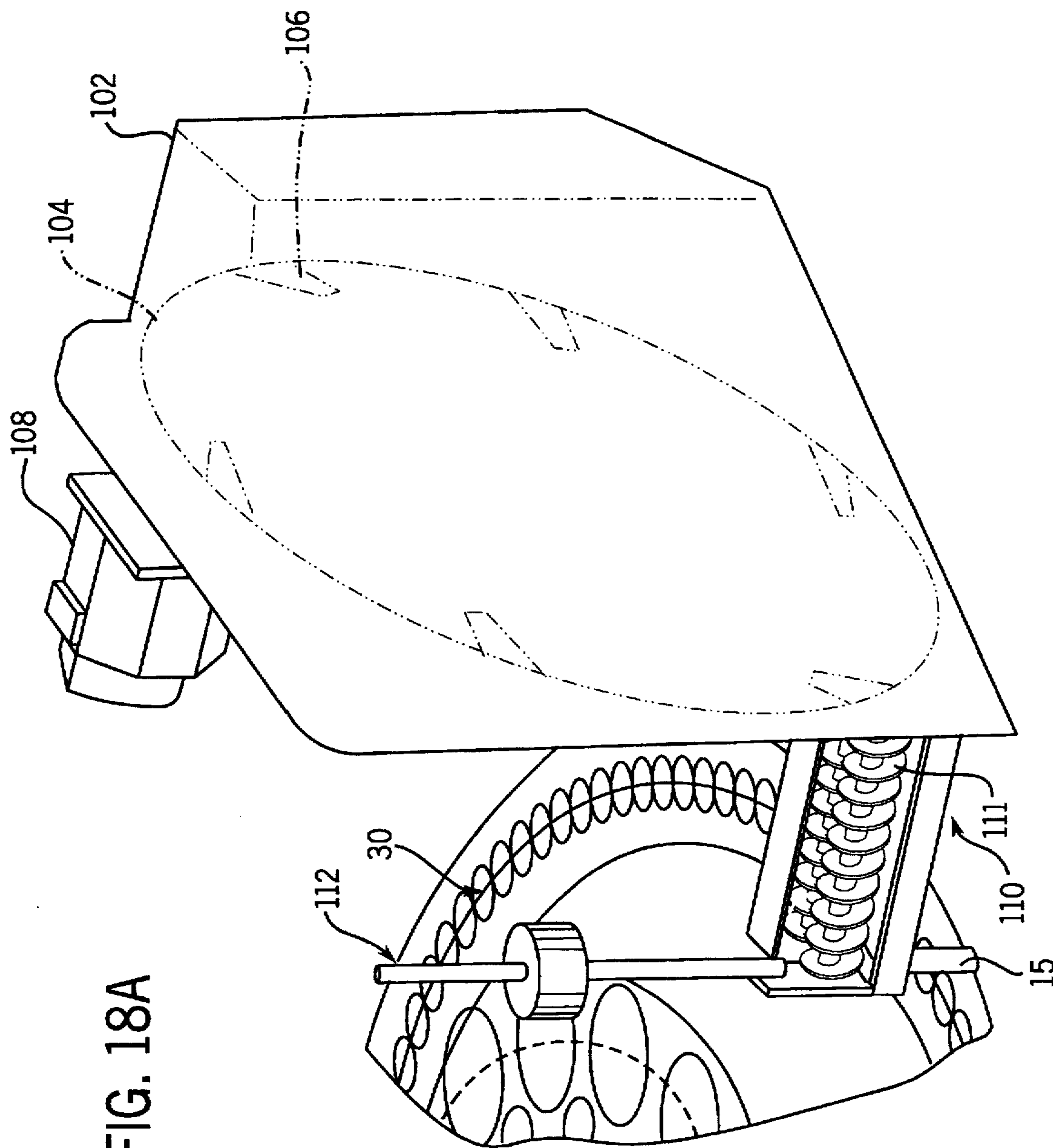


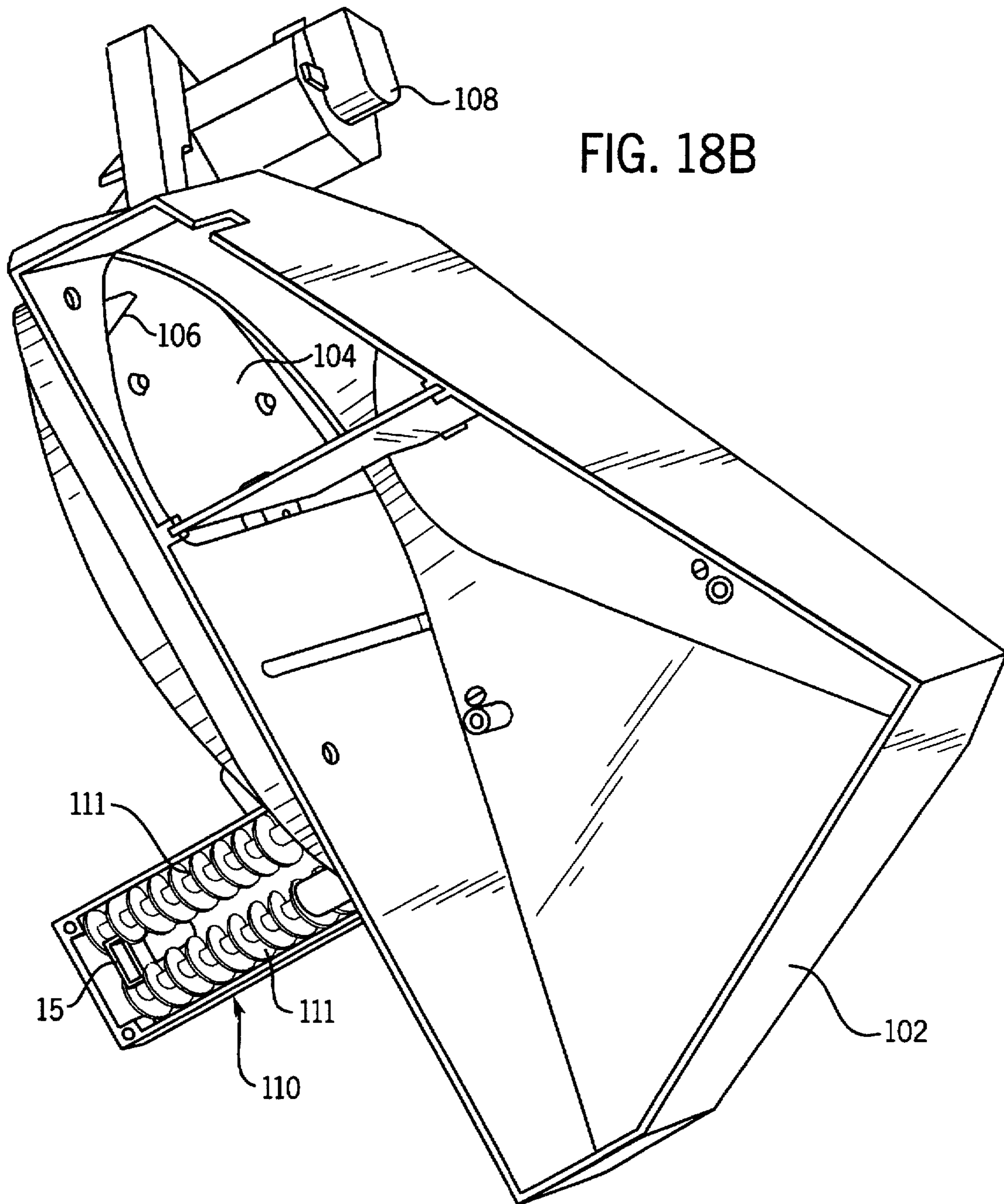














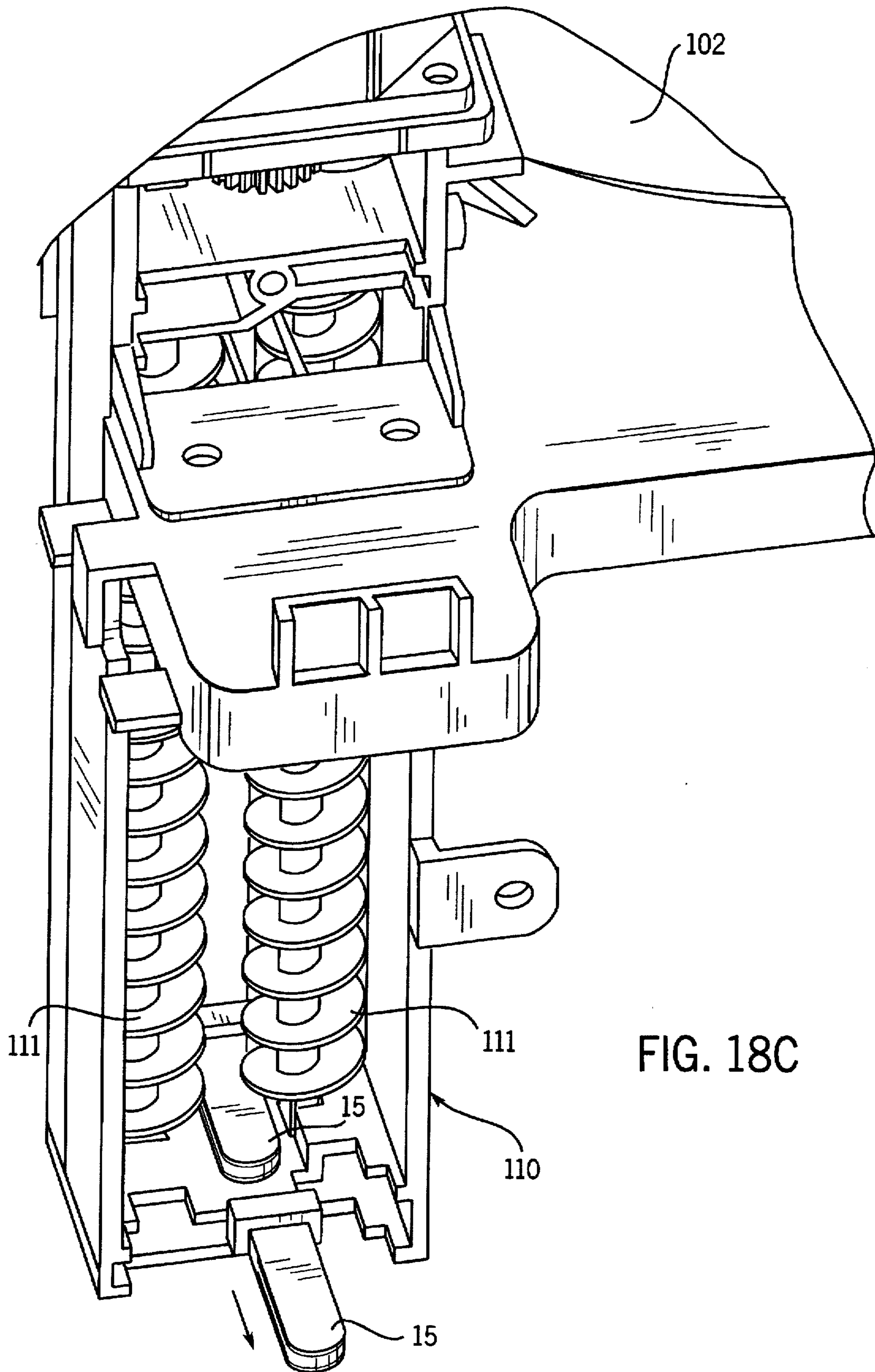


FIG. 18C

FIG. 19

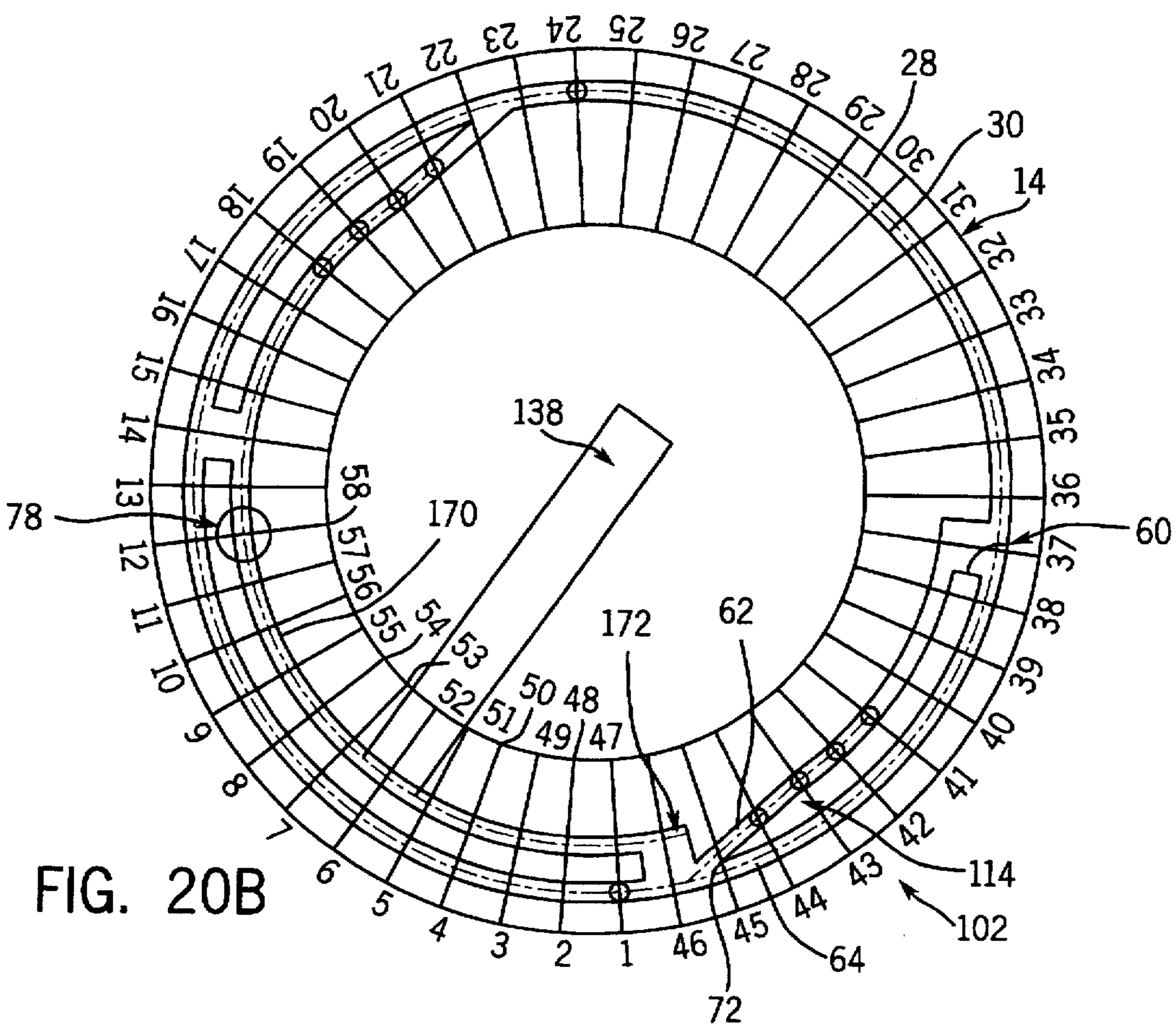
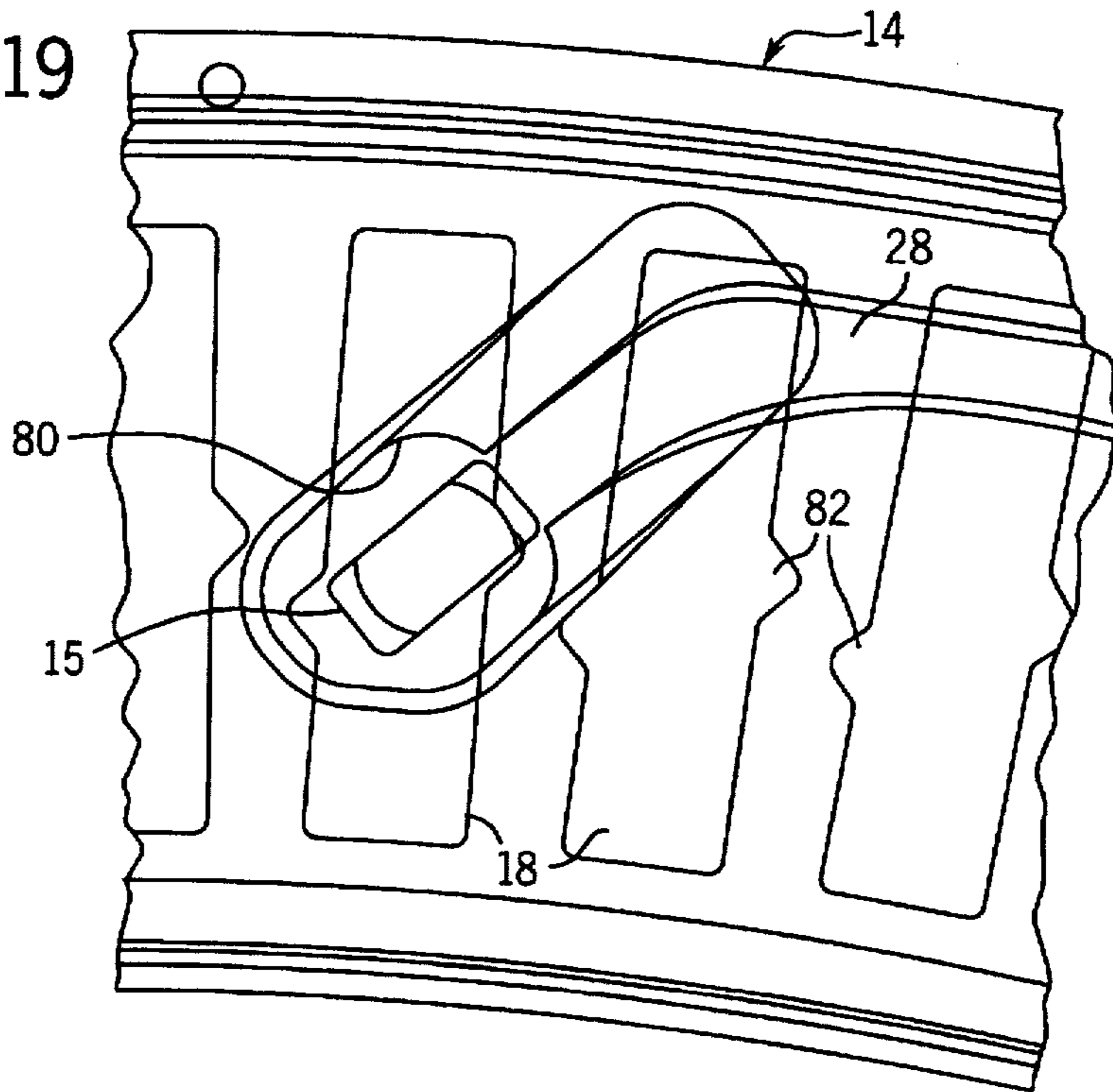


FIG. 20B



FIG. 21A

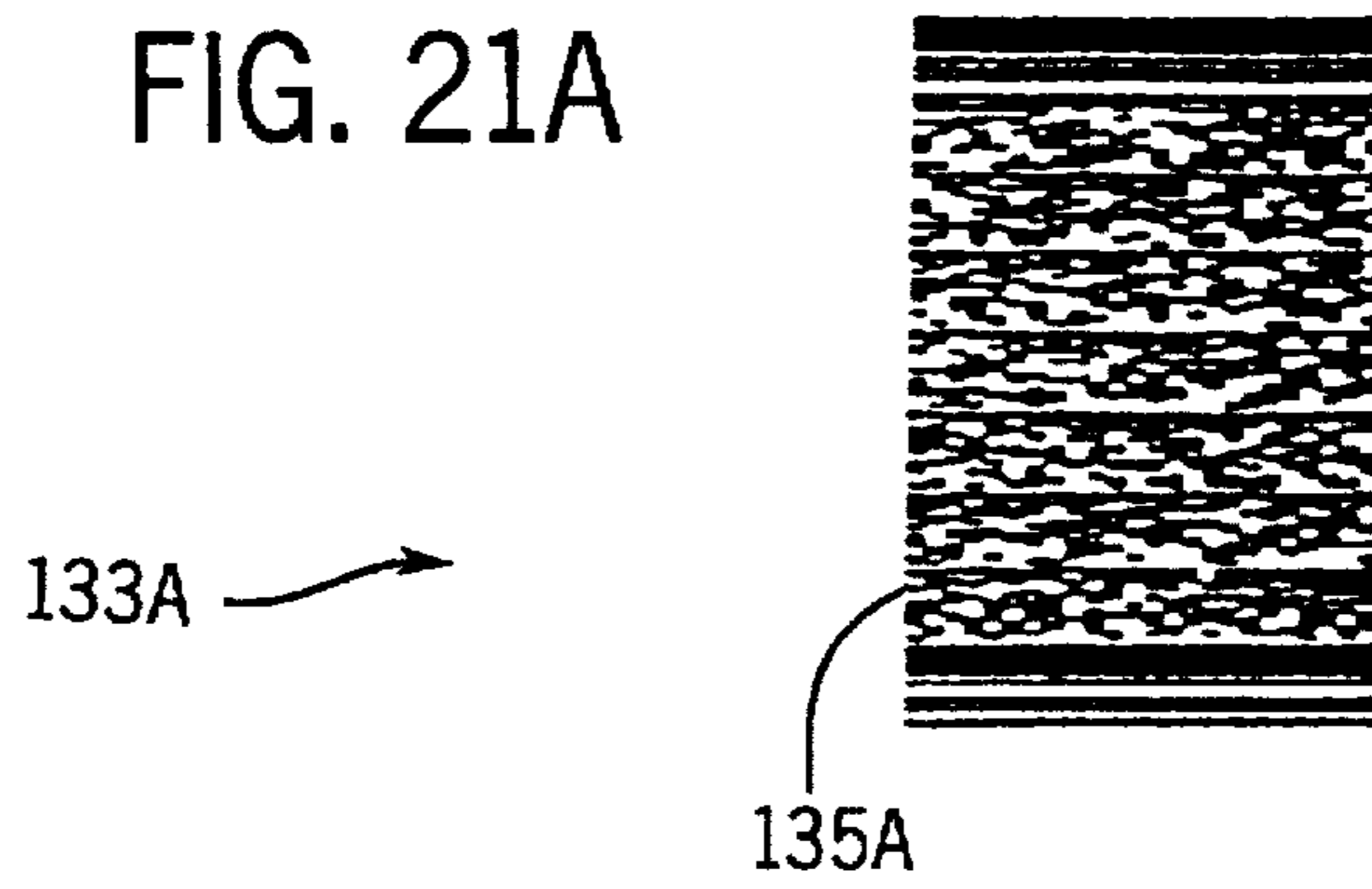


FIG. 21B

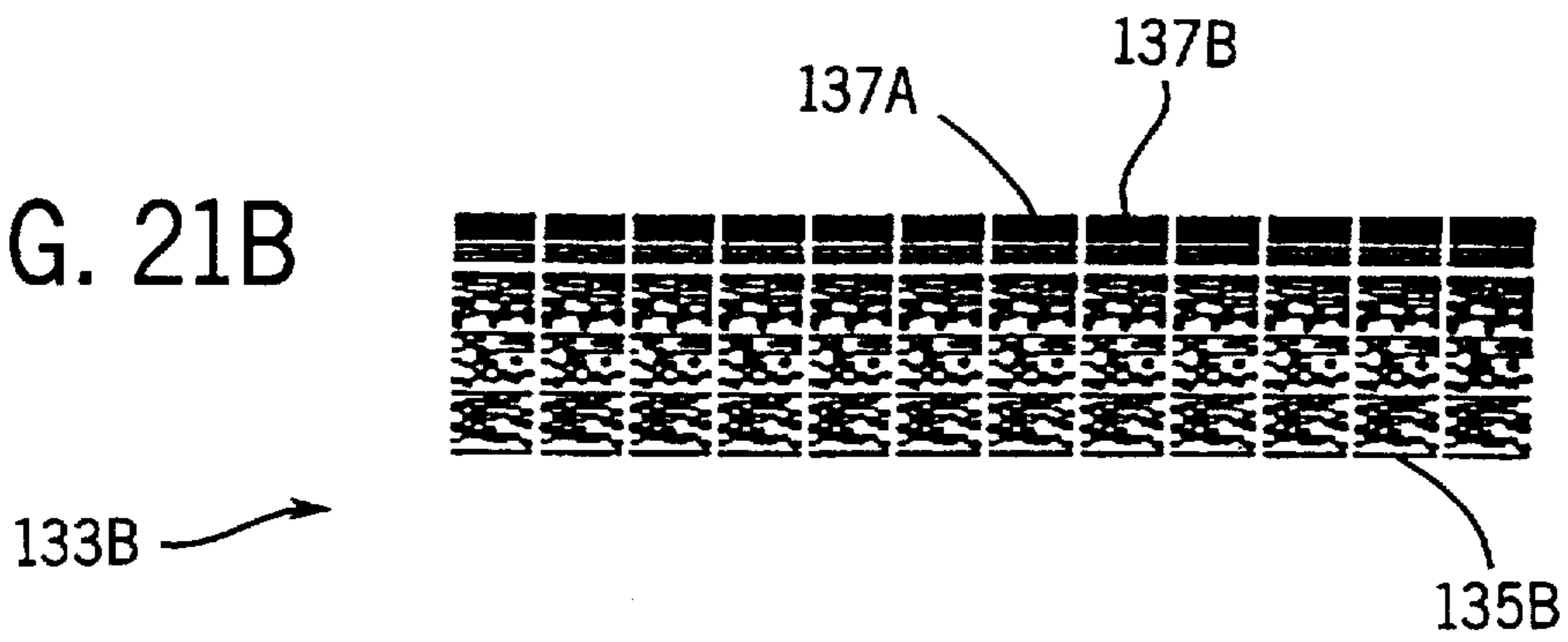


FIG. 21C

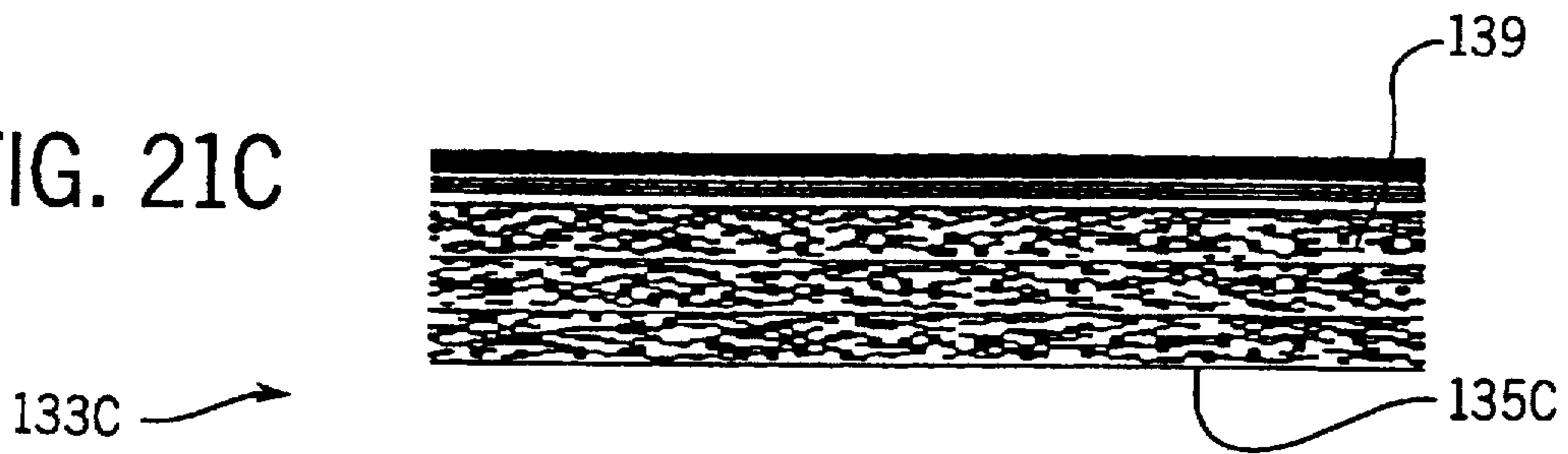


FIG. 22

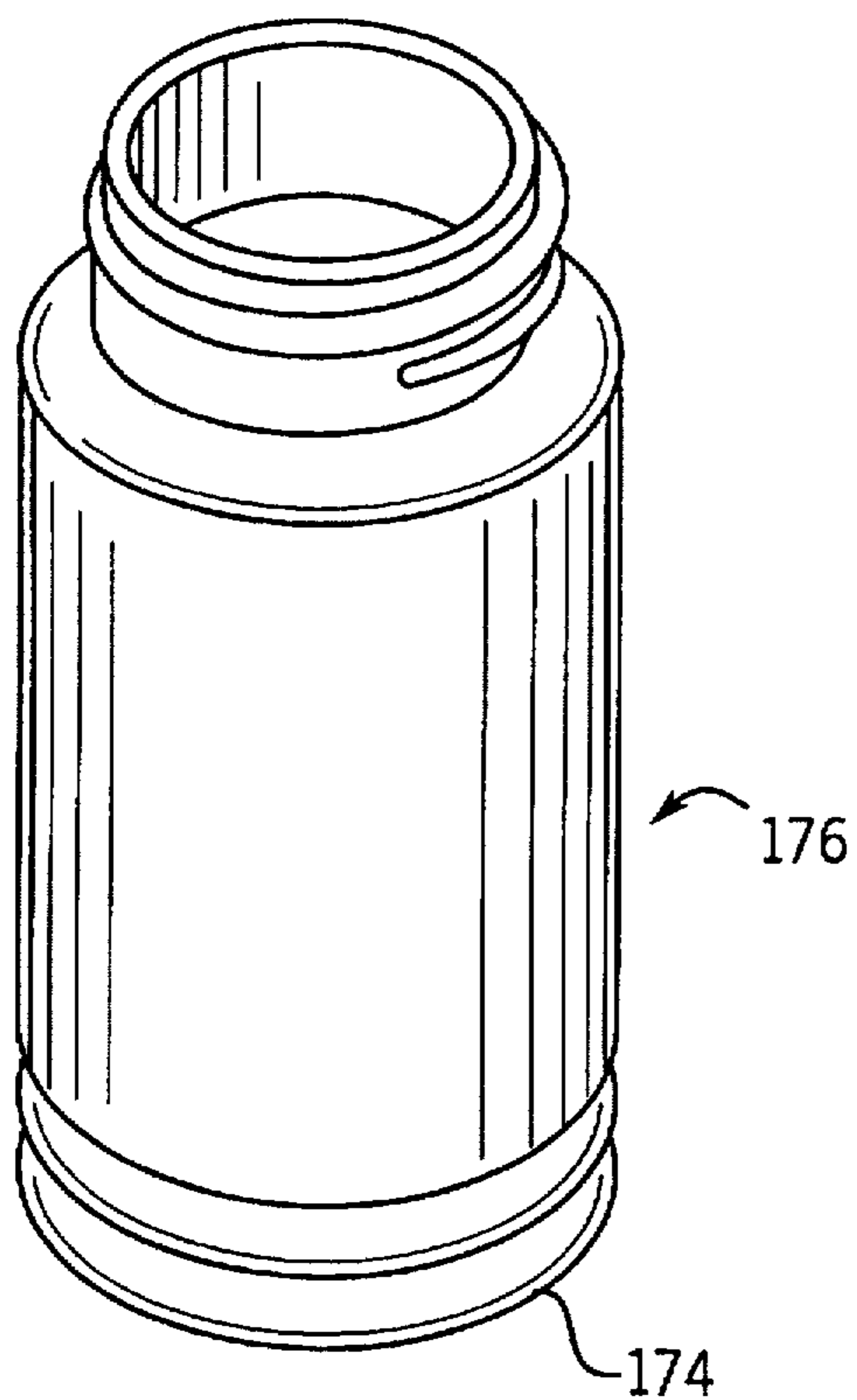


FIG. 23A

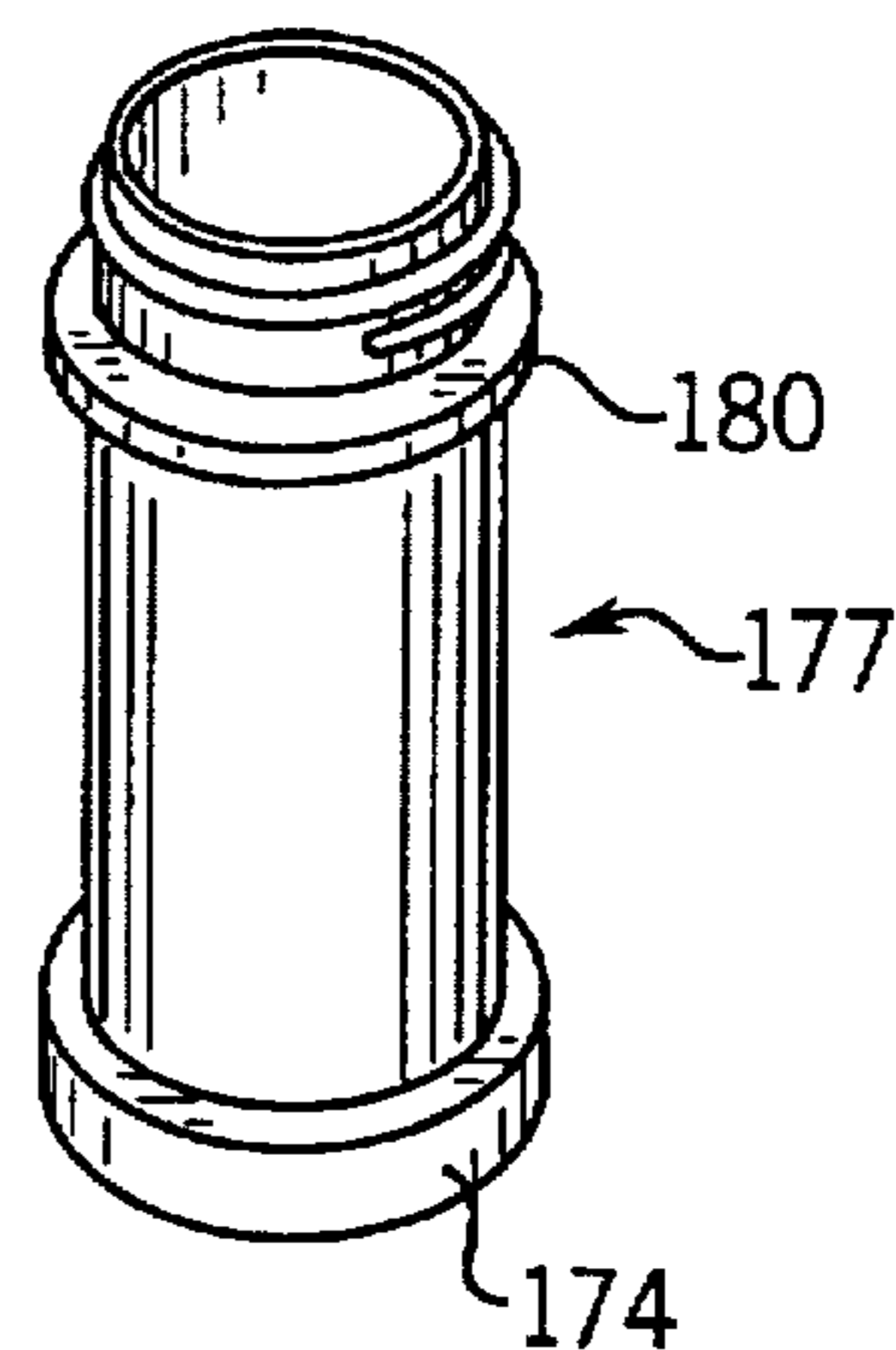


FIG. 23B

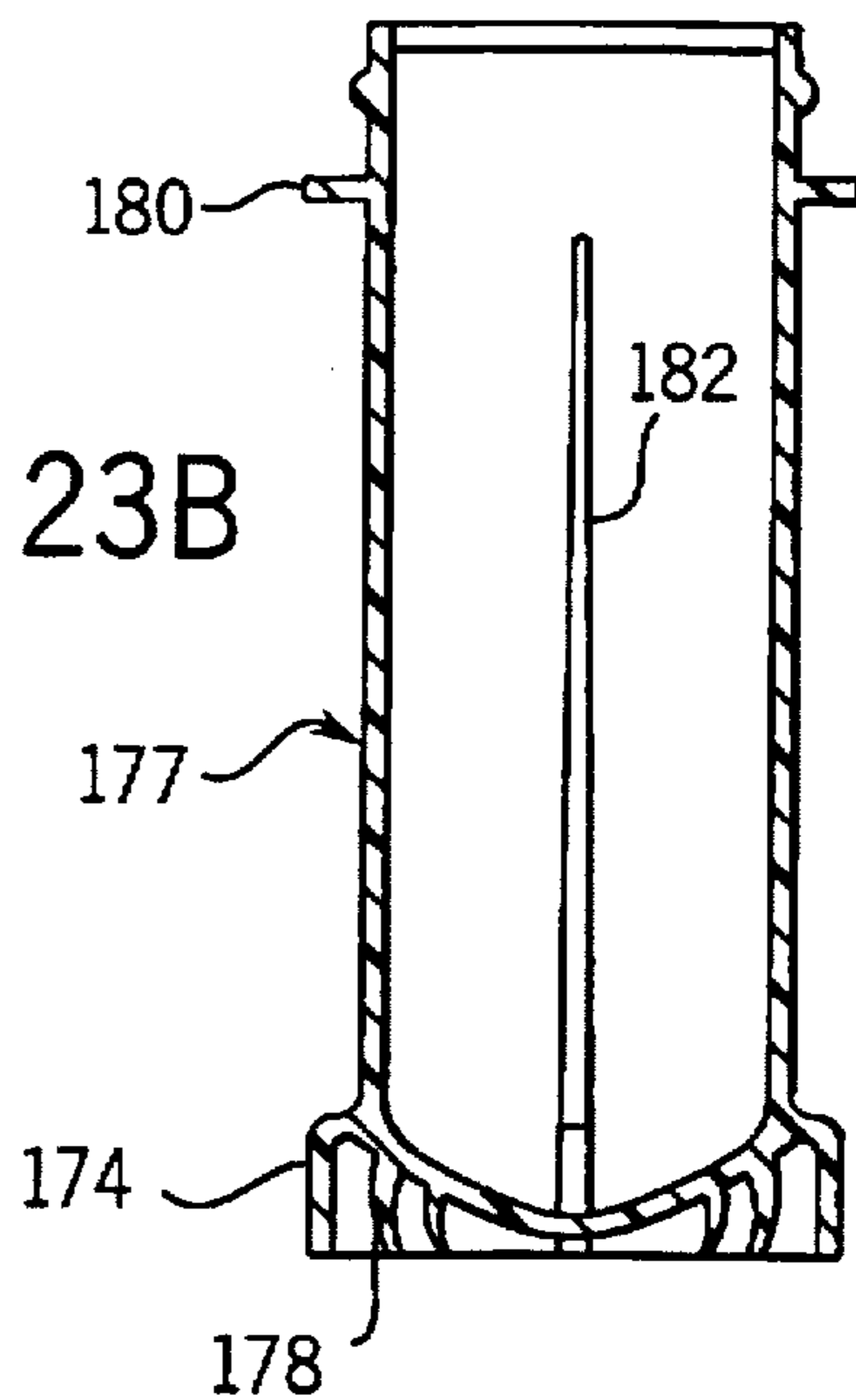


FIG. 23C

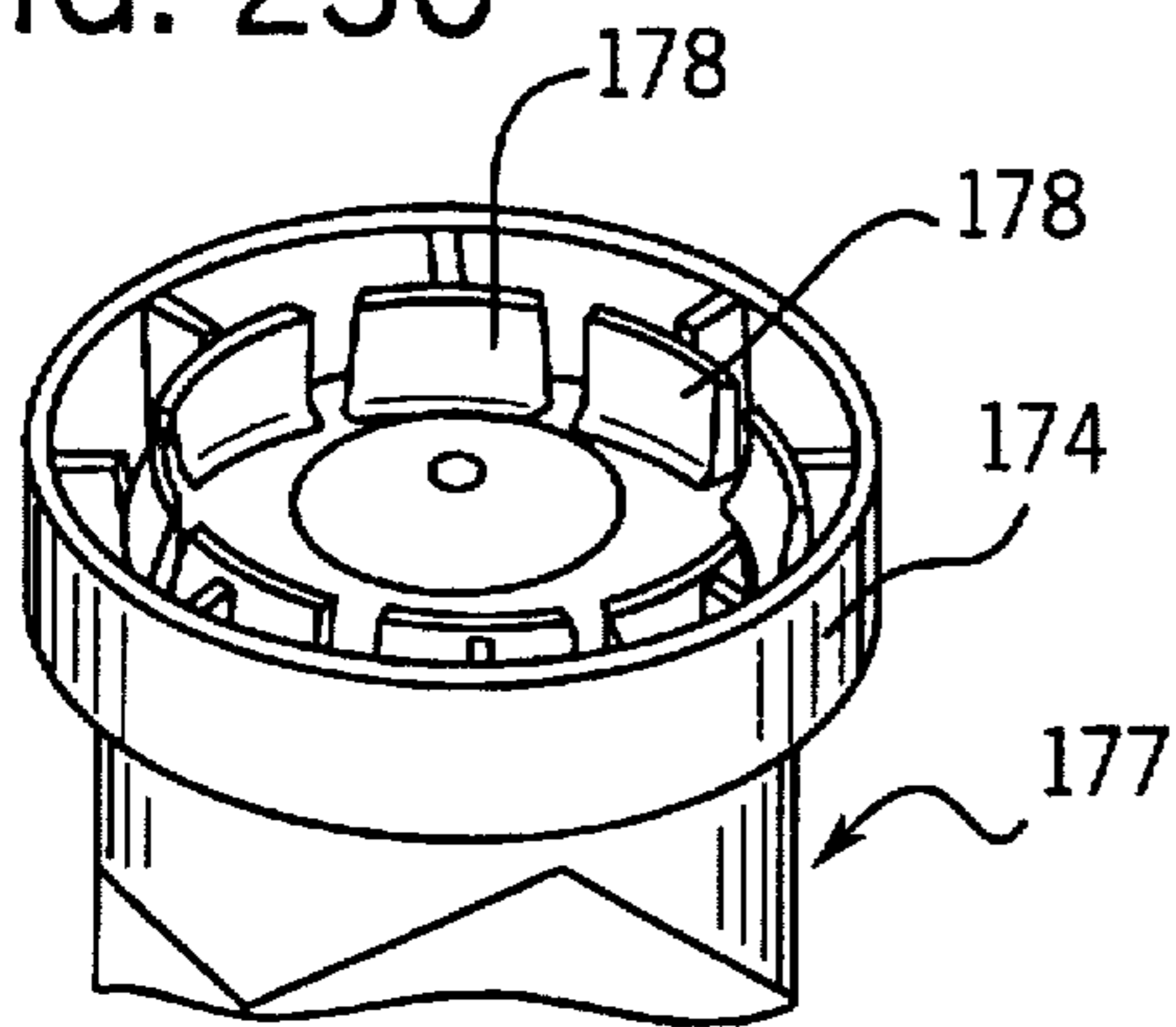


FIG. 25

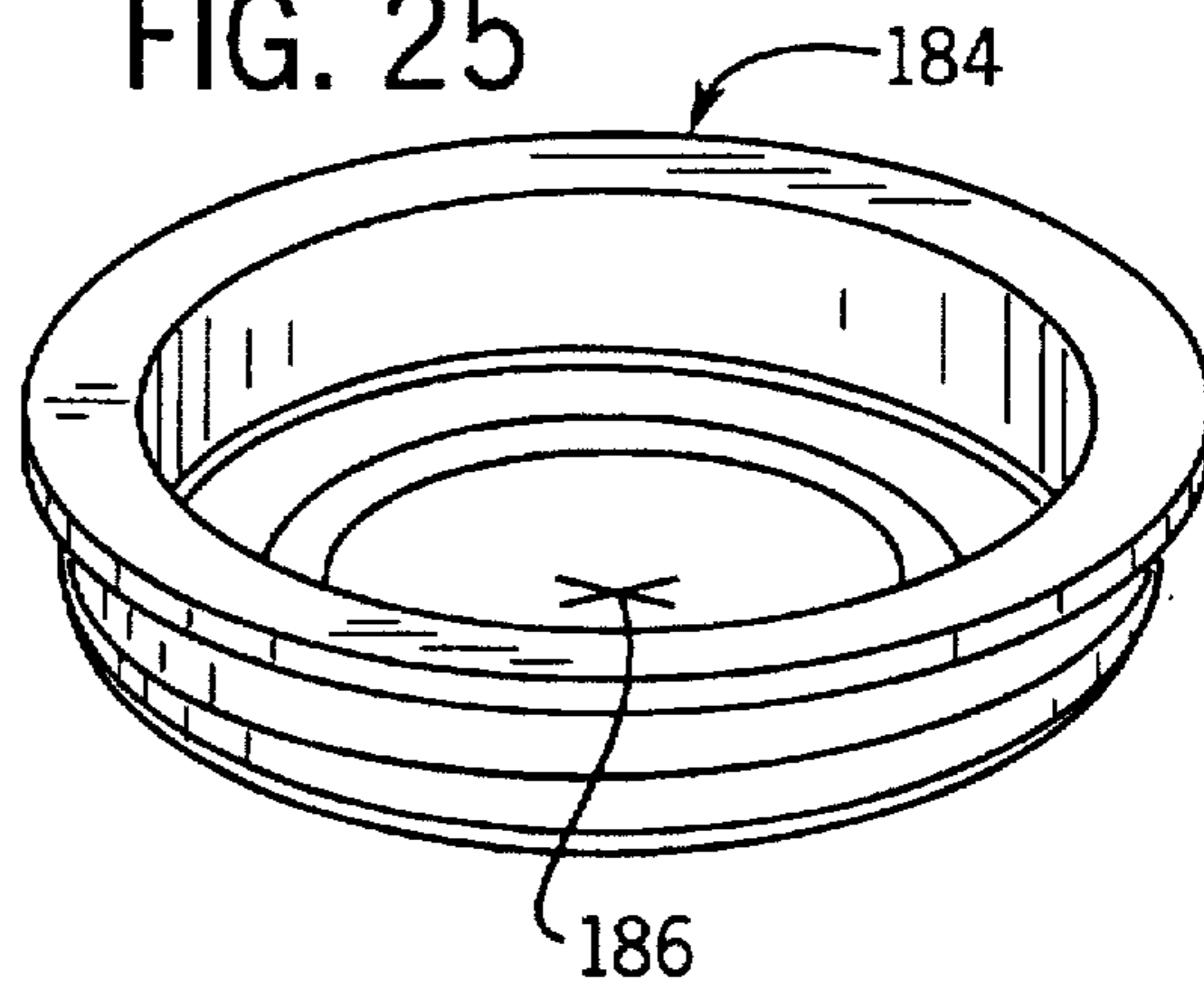
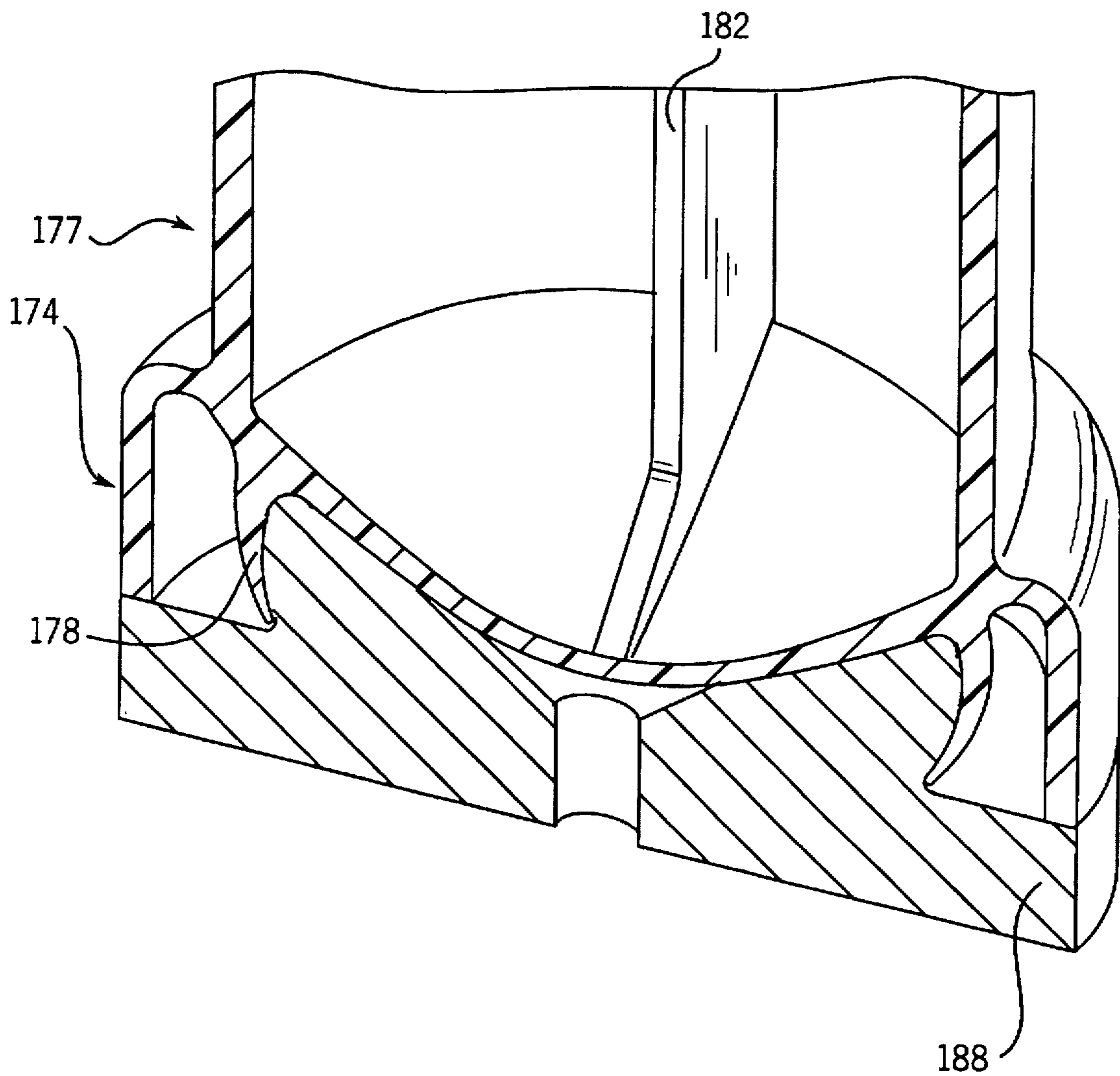
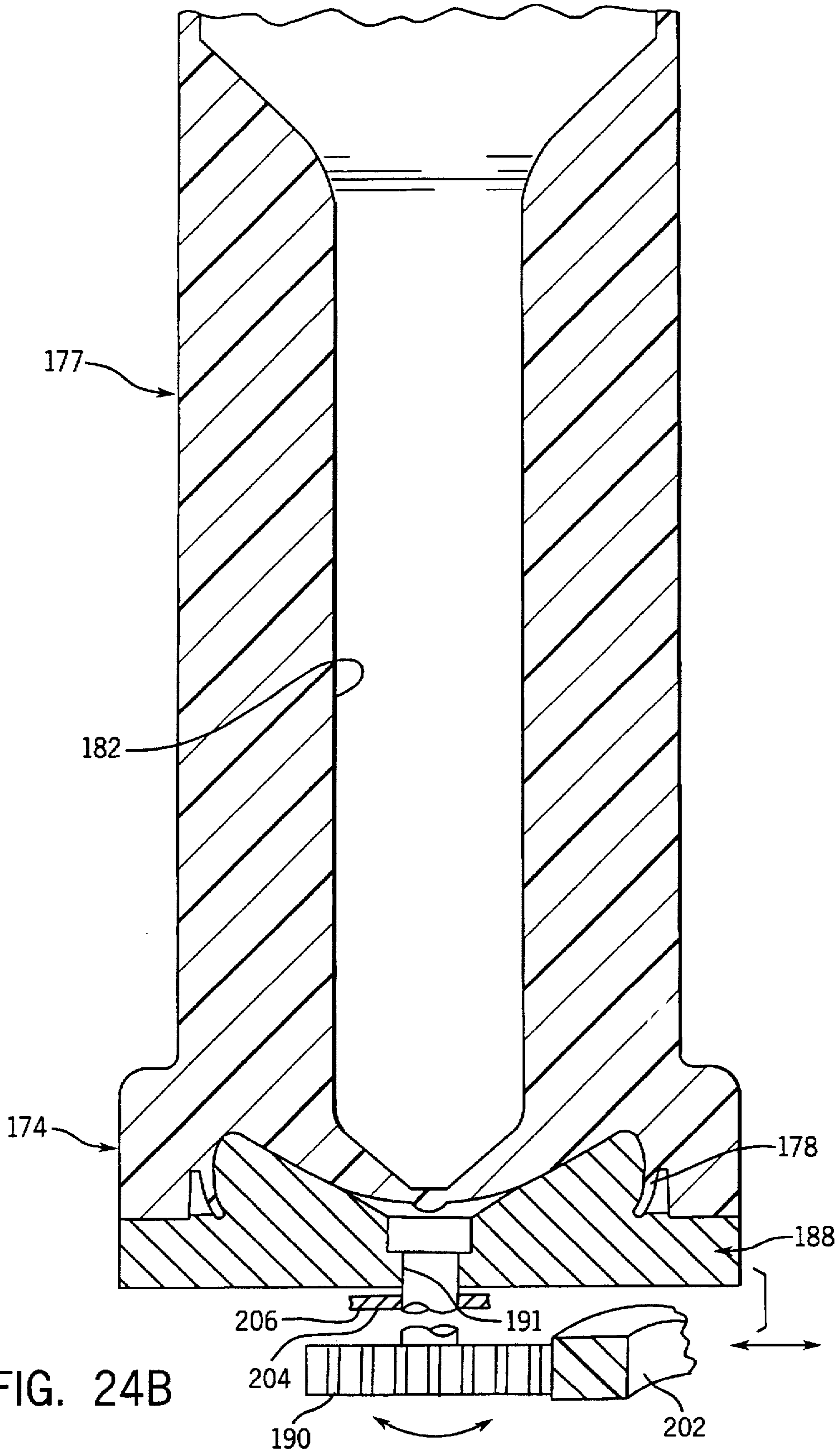


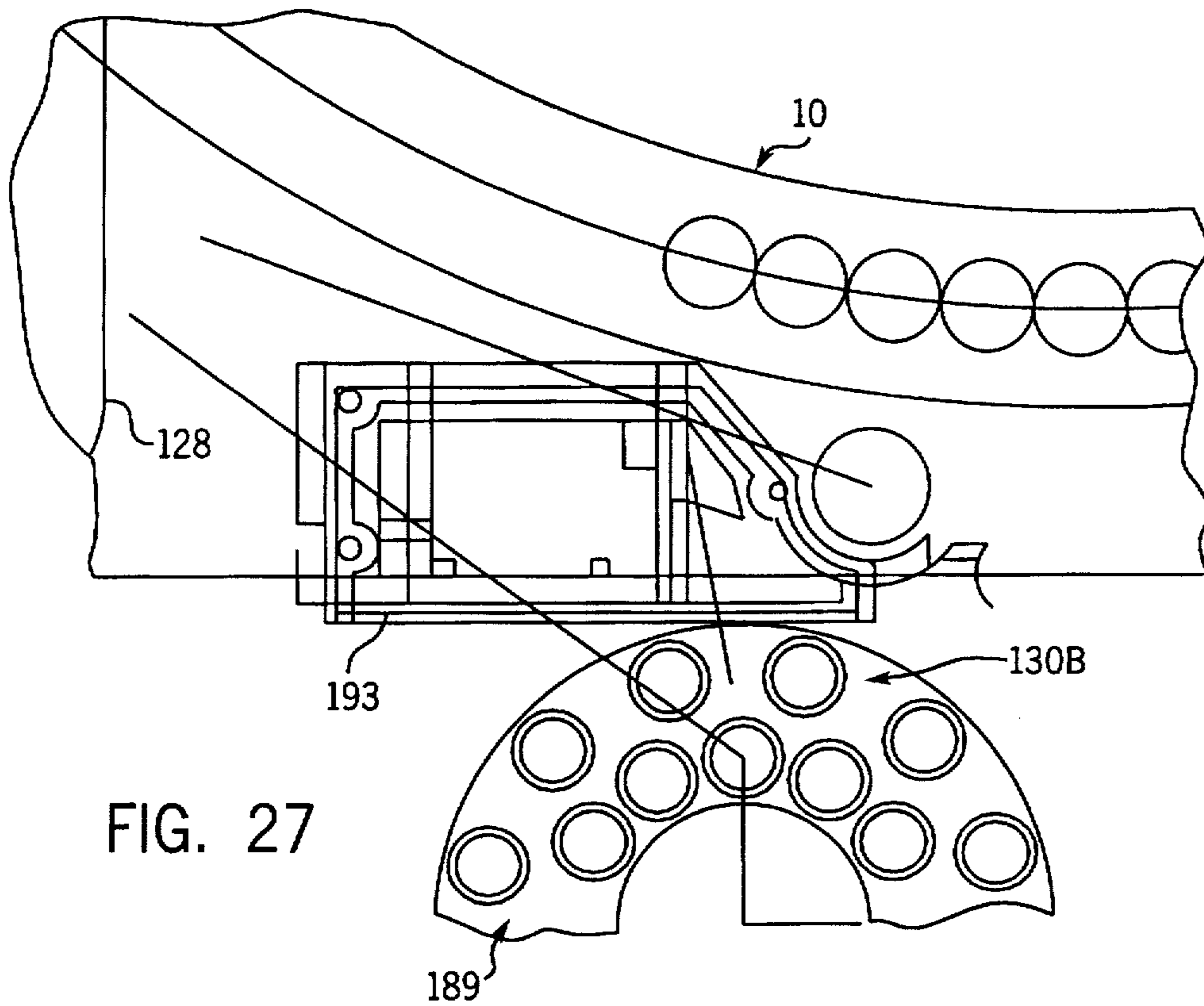
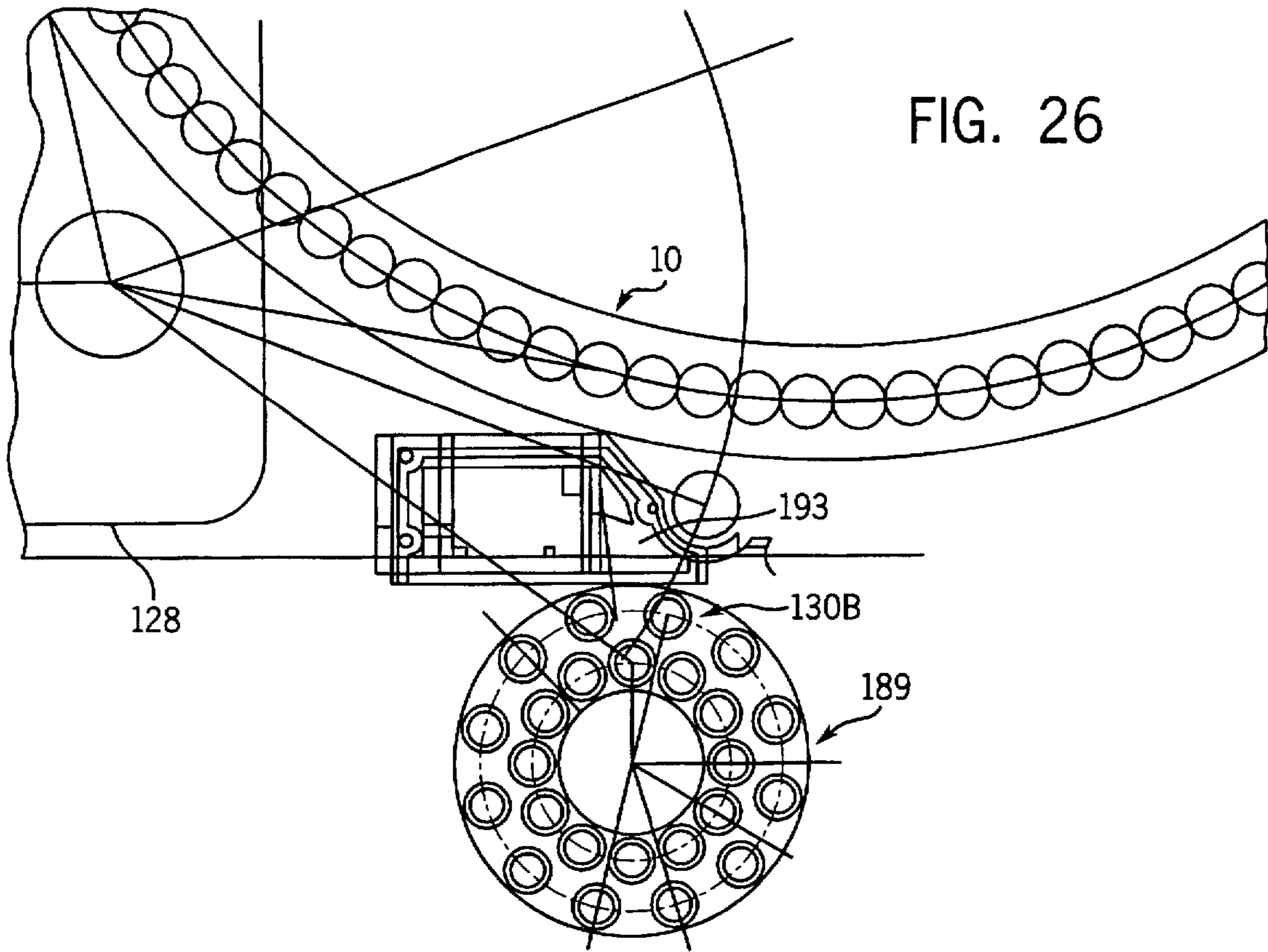


FIG. 24A









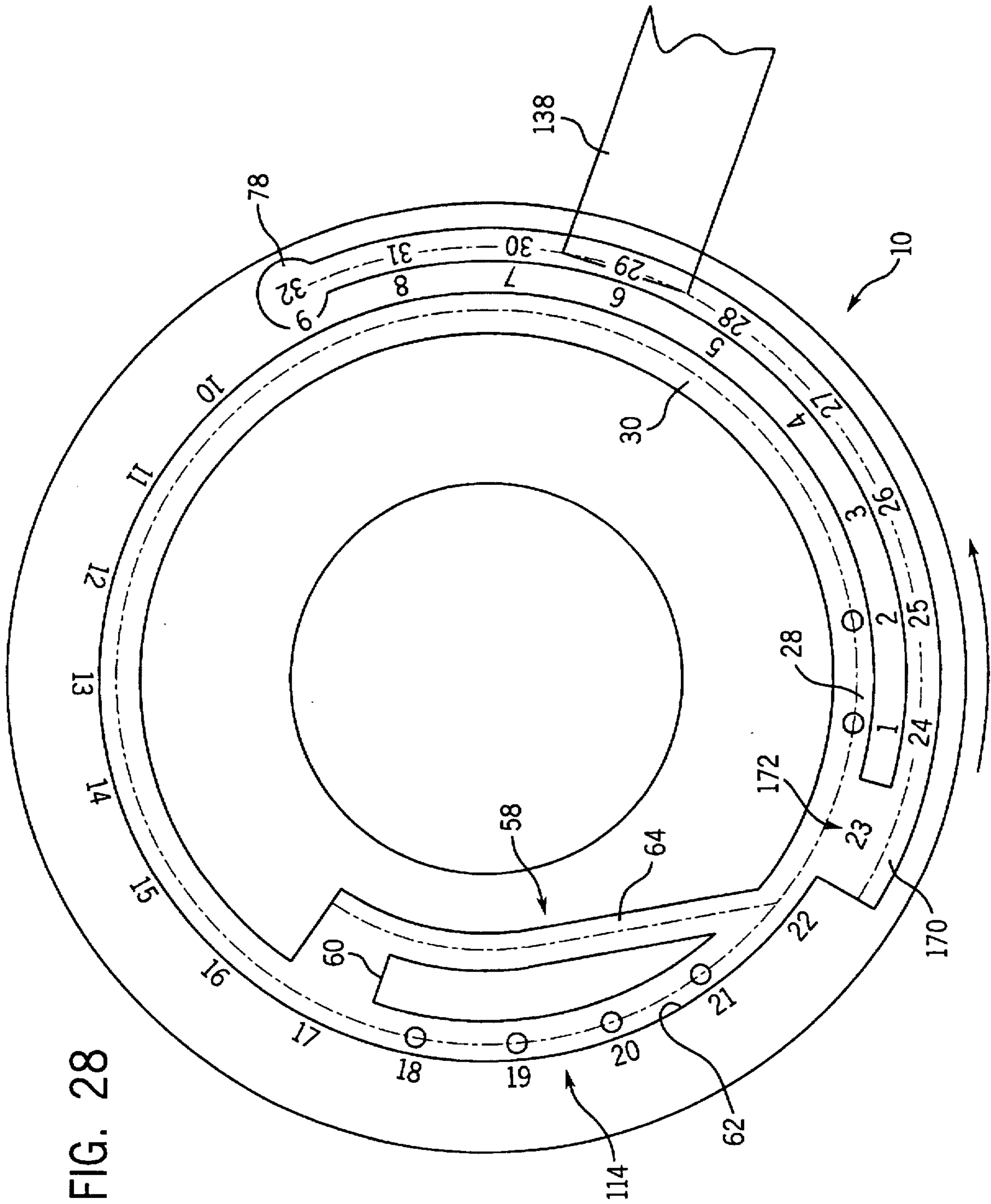
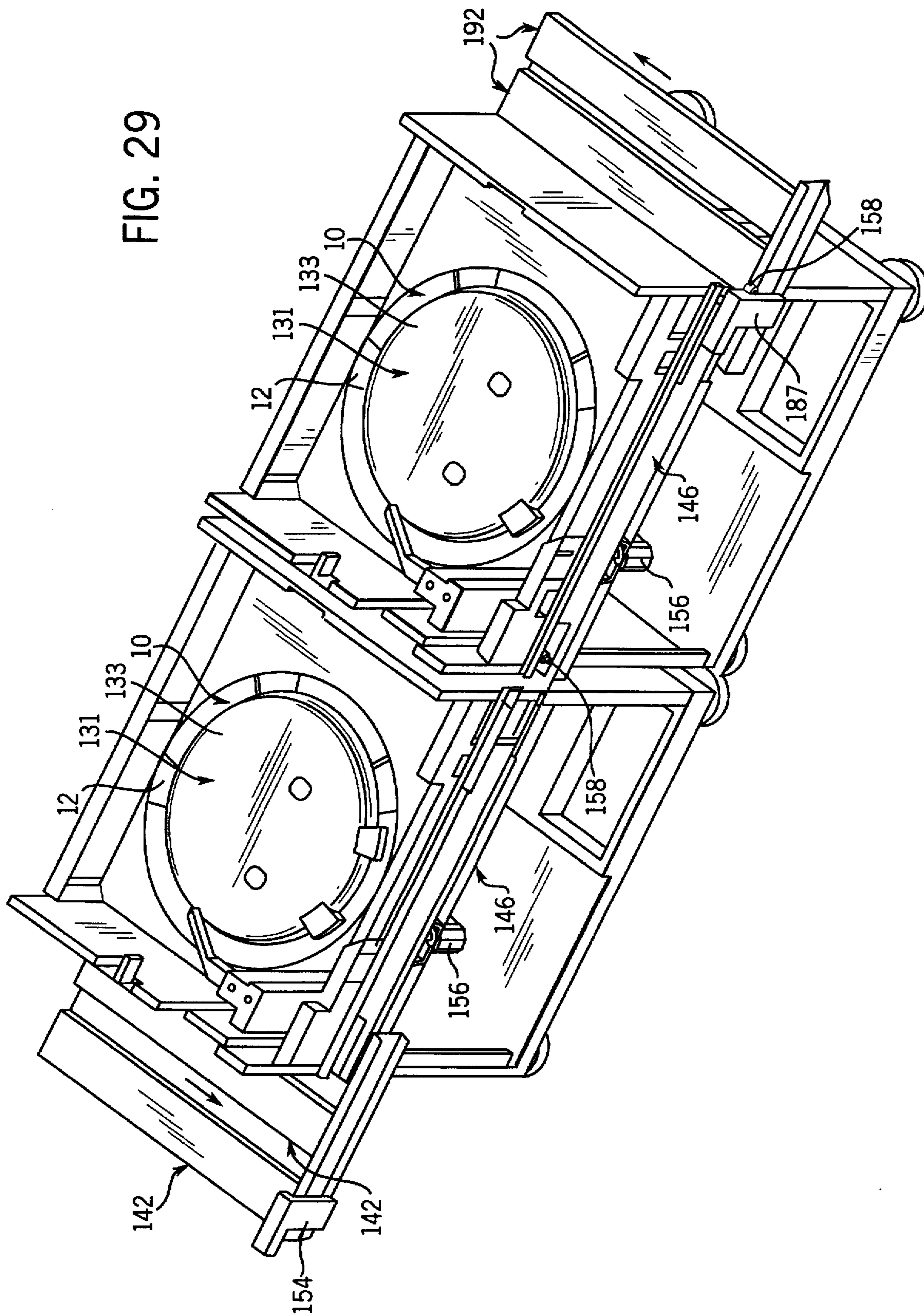


FIG. 28



FIG. 29



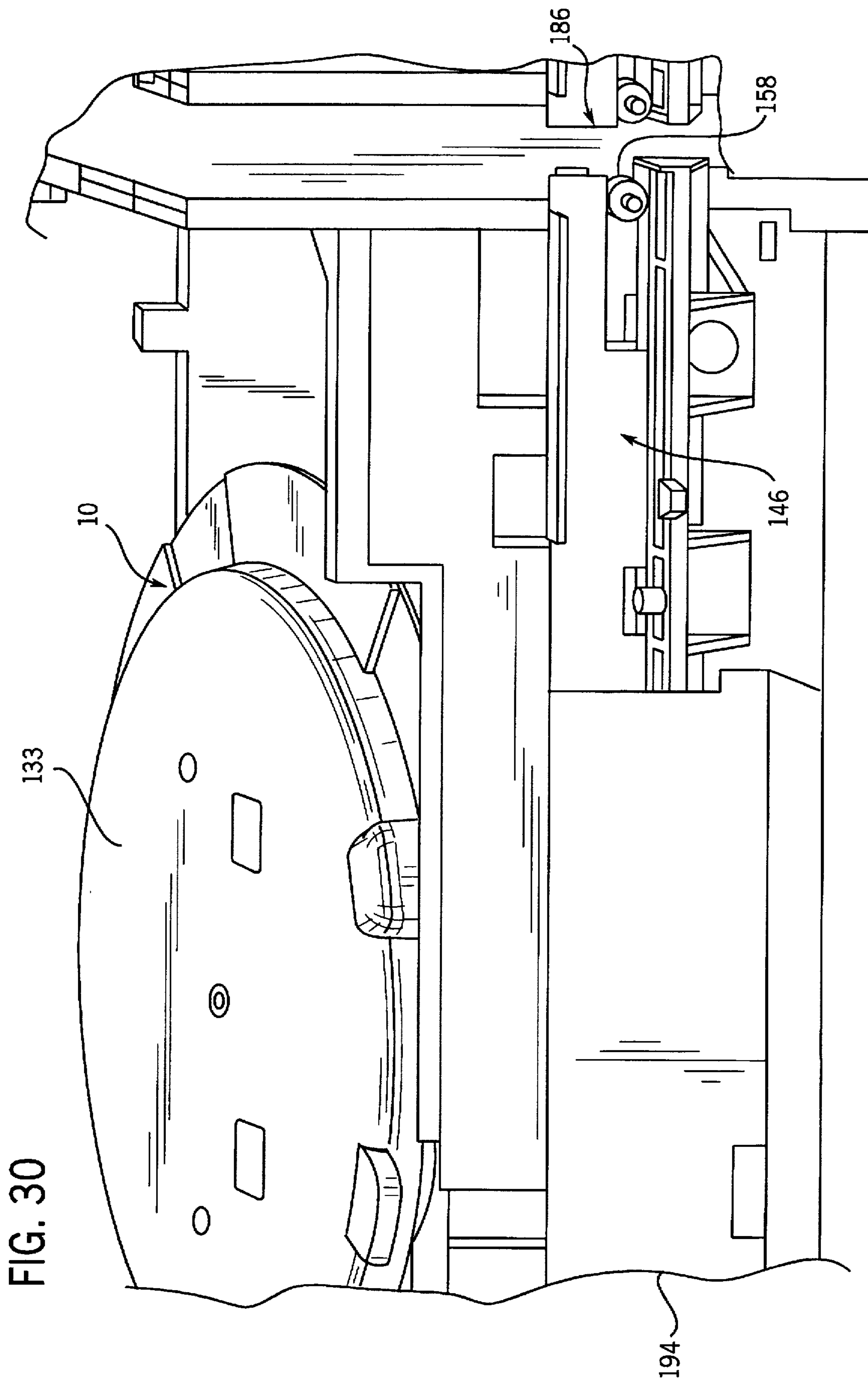
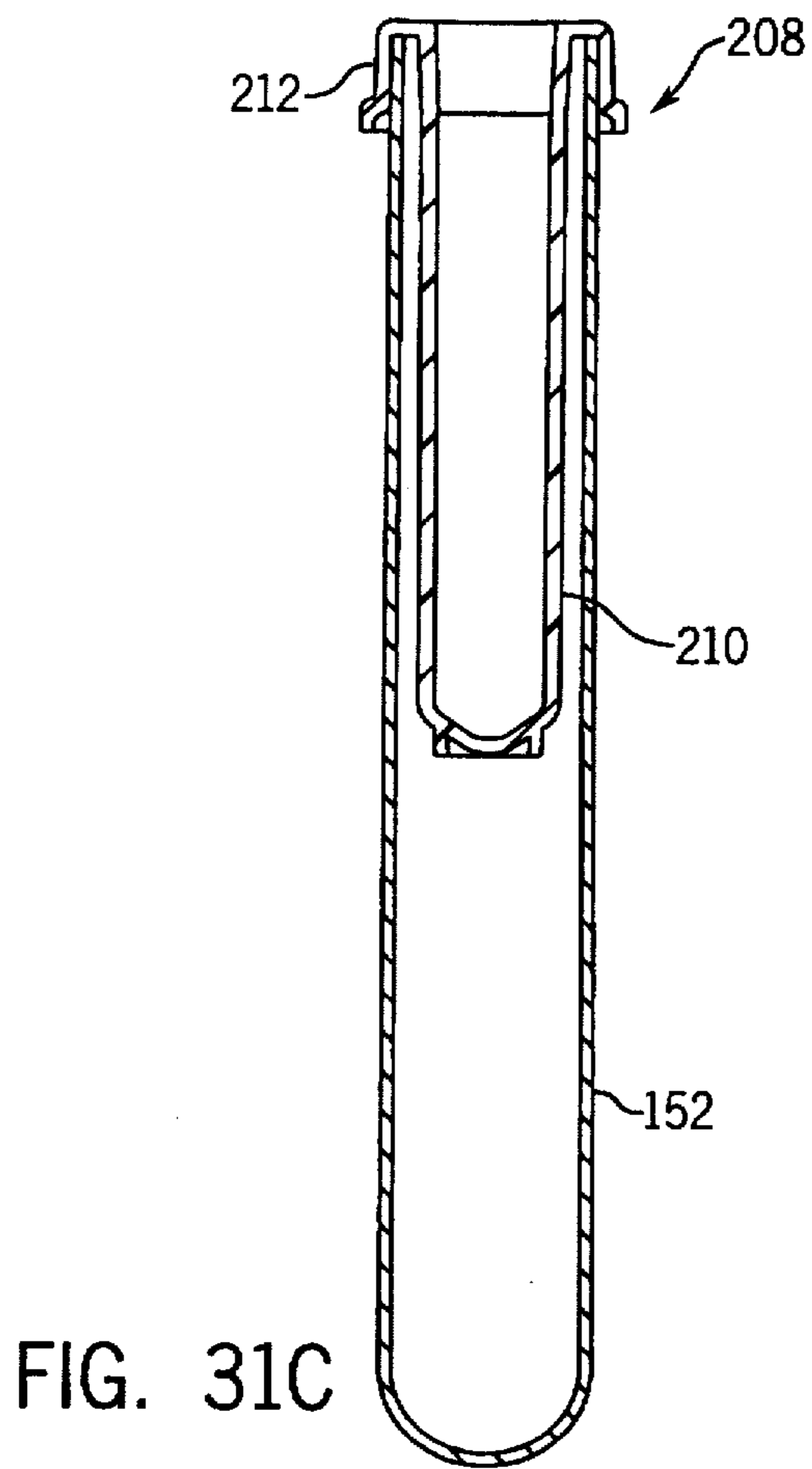
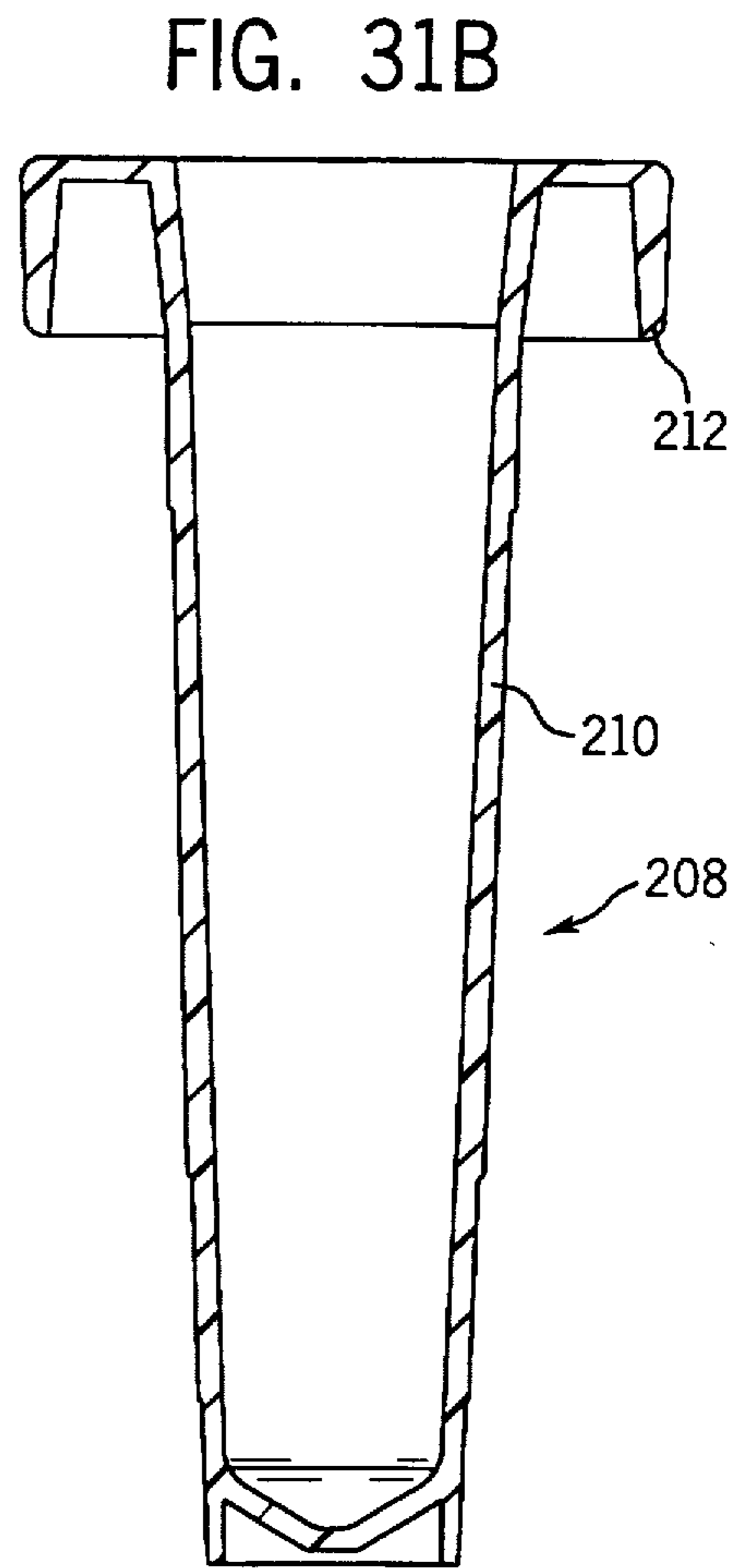
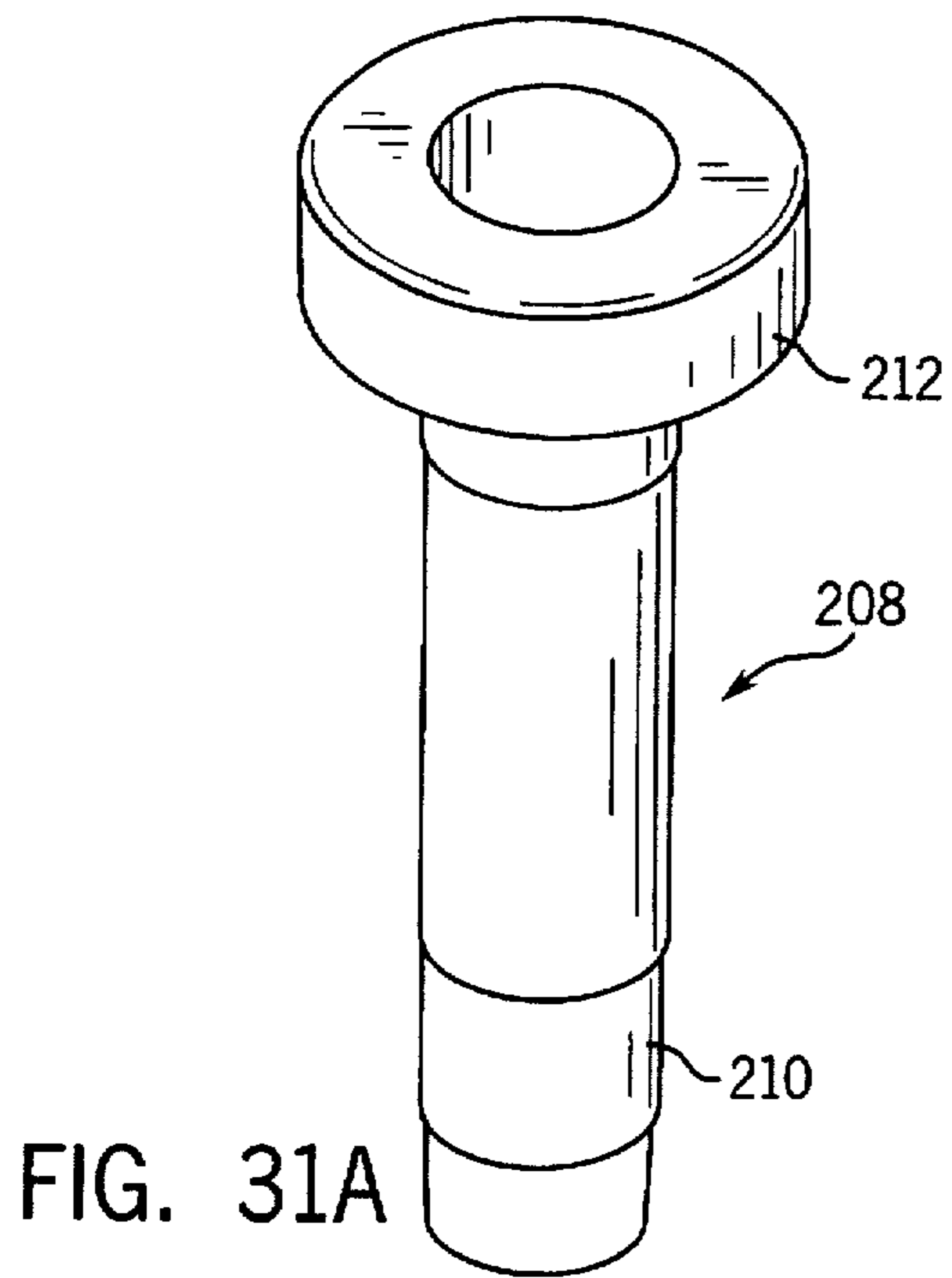


FIG. 30





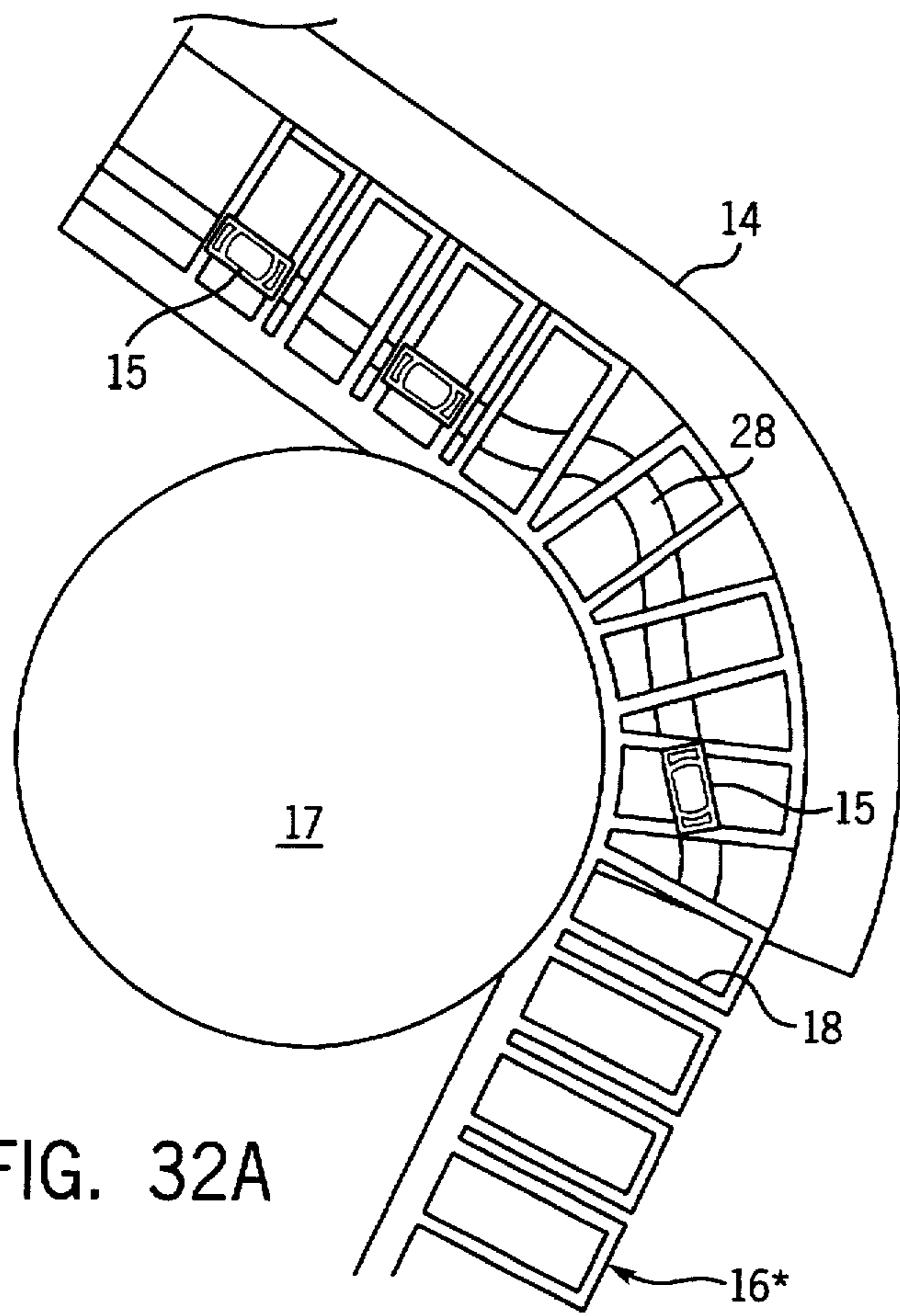


FIG. 32A

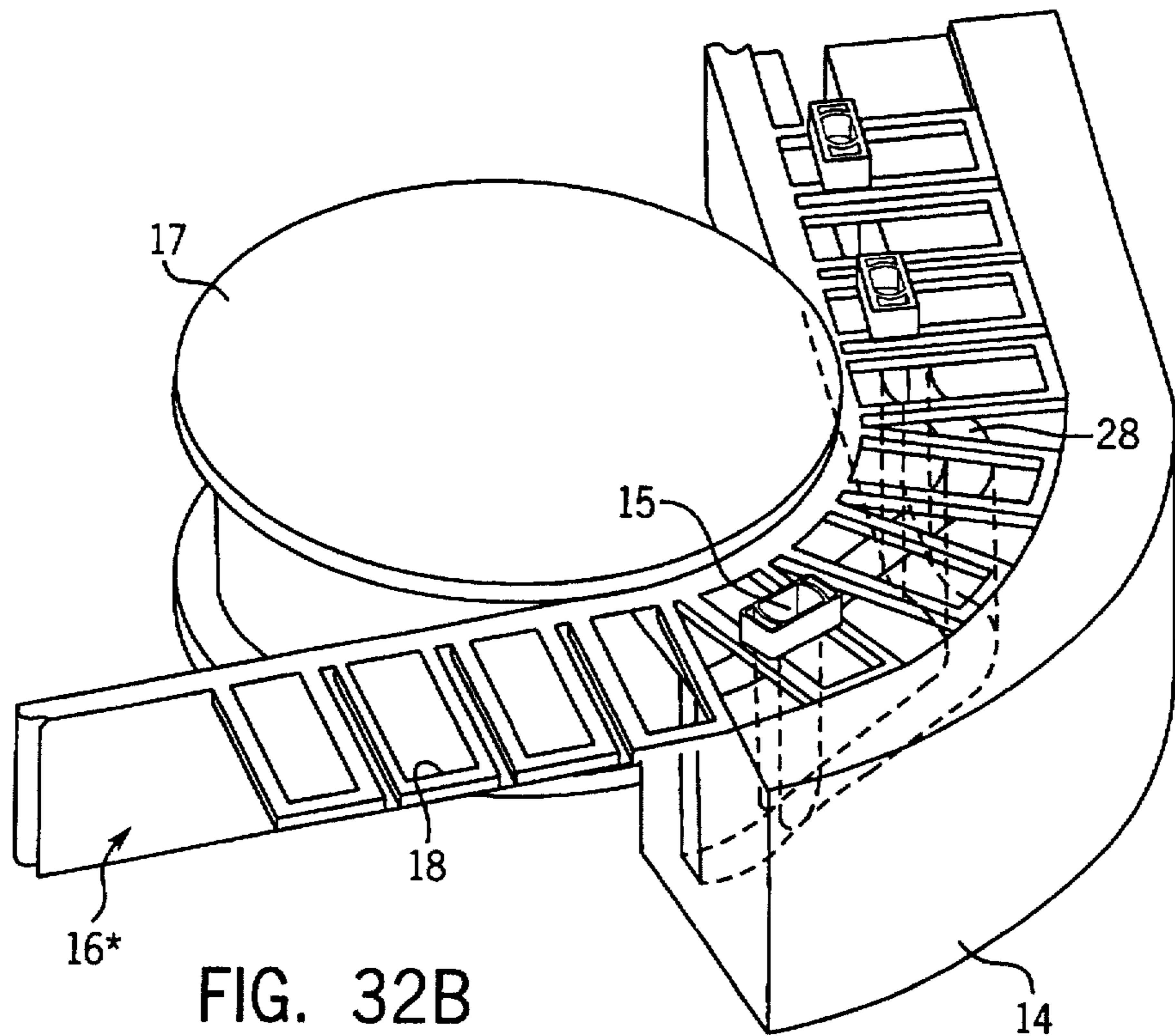


FIG. 32B



## METHOD OF PERFORMING A PROCESS FOR DETERMINING AN ITEM OF INTEREST IN A SAMPLE

### BACKGROUND

Embodiments described herein relate generally to methods and structures which determine an item of interest in a sample.

To provide information about a patient's health, a number of tests can be performed on a patient sample, such as the patient's bodily fluids. These bodily fluids may include blood, urine, etc. The tests performed on the patient's bodily fluids can determine an item of interest in the bodily fluids. Based on the determination of the item of interest in the patient's bodily fluids, information about the patient's health status can be obtained.

### SUMMARY

Embodiments described herein provide methods of performing a process for determining an item of interest in a sample. In one embodiment, a container for holding the sample is accepted in a process lane where a process step is selectively automatically performed on the sample in the container. The process step is selectively automatically performed on the sample in the container. An effective length of the process lane is maintained constant while a physical length of the process lane is selectively varied.

In another embodiment, a container for holding the sample is accepted in a process lane where a process step is selectively automatically performed on the sample in the container. The process step is selectively automatically performed on the sample in the container. A physical length of the process lane is selectively varied.

### BRIEF DESCRIPTION OF DRAWINGS

FIG. 1 is a perspective view of a component of an analyzer;

FIG. 2 shows the component of FIG. 1 with elements thereof removed for clarity;

FIG. 3 is a perspective view of an element of the component shown in FIG. 1;

FIG. 4 is a top view of the component of FIG. 1 with elements thereof removed for clarity;

FIGS. 5A and 5B show another element of the component of FIG. 1 which is connected with the structure shown in FIG. 2;

FIG. 6 is an enlarged sectional view of the component of FIG. 1 with elements removed for clarity;

FIG. 7A is a perspective view of a container for use with the component of FIG. 1;

FIG. 7B is a perspective view of another container for use with the component of FIG. 1;

FIG. 8 is an enlarged sectional view of a portion of the component of FIG. 1 showing interaction with the container of FIG. 7B;

FIG. 9 is an enlarged sectional view, substantially similar to that of FIG. 8, of another portion of the component of FIG. 1;

FIG. 10 is substantially similar to FIG. 9 but shows another portion of the component of FIG. 1;

FIG. 11 is substantially similar to FIG. 10 but shows another portion of the component of FIG. 1;

FIG. 12 is a perspective view of an element of the component of FIG. 1;

FIG. 13 is an enlarged sectional view of a section of another embodiment of the component shown in FIG. 1;

FIG. 14 is a perspective view of an element of the component of FIG. 1;

FIG. 15 is a perspective view of an element of the component of FIG. 1;

FIG. 16 is a generic view of the component of FIG. 1 cooperating with other portions of an analyzer;

FIG. 17 is a perspective view of a frame for the structures shown in FIG. 16;

FIGS. 18A, 18B and 18C illustrate an element of the component shown in FIG. 1;

FIG. 19 is an enlarged sectional view of a section of another embodiment substantially similar to that shown in FIG. 13;

FIGS. 20A and 20B are generic views of other related analyzers having oppositely directed components substantially similar to the component of FIG. 1;

FIGS. 21A, 21B and 21C show an embodiment of a high density data carrier which may be used with the component of FIG. 1;

FIG. 22 is an isometric view of a container for use with the process path of FIG. 1;

FIGS. 23A, 23B and 23C show another container for use with the process path of FIG. 1;

FIGS. 24A and 24B are enlarged sectional views of a portion of the container of FIGS. 23A, 23B and 23C operatively associated with a support;

FIG. 25 is an isometric view of a seal which may be used with the containers of FIGS. 22, 23A, 23B and 23C;

FIG. 26 is an enlarged section of another application of the process path of FIG. 1;

FIG. 27 is an enlargement of a portion of FIG. 27;

FIG. 28 is a generic view of another related analyzer having a component substantially similar to the component of FIG. 1;

FIG. 29 is an illustration of two components of FIG. 1 joined together;

FIG. 30 is an enlarged view of a portion of FIG. 29;

FIGS. 31A, 31B and 31C show another container for use with the process path of FIG. 1; and

FIGS. 32A and 32B illustrate portions of another embodiment of the process path.

### DETAILED DESCRIPTION OF PREFERRED EMBODIMENTS

The embodiments described herein relate to methods and structures for determining an item of interest in a sample. The item of interest may be an antibody, an antigen, concentrations of the former or latter or any other desired element of the sample. In an exemplary embodiment, the item of interest is selected from, but are not limited to, antibodies to HCV, antibodies to HIV 1/HIV 2, antibodies to hepatitis B core antigen (HBcAb), carcinoembryonic antigen (CEA), cancer antigen 19-9 (CA19-9), Hepatitis B Surface Antigen (HBsAg), antibodies to Hepatitis B Surface antigen (HBsAb), alpha-fetoprotein (AFP), Total prostate specific antigen (Total PSA) Free PSA, Thyroid stimulating Hormone (TSH), luteinizing hormone (LH), follicle stimulating hormone (FSH), beta human chorionic gonadotropin (B-hCG), Free Thyroxine (Free T4), Free triiodothyronine (Free T3), Total T4, Total T3, Progesterone, Testosterone, Estradiol, Prolactin, vitamin B12 (B12), Folate, Glycated



Hemoglobin, and Ferritin. The structures and methods may be employed in a number of different configurations.

For the sake of clarity of understanding, the structures and methods will be discussed with respect to their employment in an immunoassay analyzer which performs approximately 200 determinations of items of interest in a sample in an hour. It is to be noted that the structures and methods can be used in other employments, such as analyzers which perform 600, 400, 100, 50, etc. determinations in an hour. A number of analyzers may be joined together or integrated to meet individual needs, such as modifying the number of tests performed in a given time period (throughput), tailoring the items of interest to be determined, etc. For example a number X of analyzers which perform Y determinations in a given hour may be connected such that the connected analyzers perform XY determinations in an hour.

It is to be noted that all such analyzers perform all determinations of items on interest in substantially the same way. For instance, all determination process steps for all items of interest are performed within the same time frame, such as 18 seconds, irrespective of the number or type of determinations to be performed by the given analyzer. These analyzers may include common elements, such as reagents, disposable articles, element, such as fluids and the like, delivery technologies, determination step performance mechanisms, software, etc.

In other applications, the analyzer may be joined, e.g. with a conveyor system and the like, along with supporting hardware and software, such that the analyzer can be used with different analyzers, such as clinical chemistry or hematology analyzers and the like, in the same setting. This conveyor system may move samples among the analyzers such that different determinations can be made with respect to one sample. Also, while operation of the analyzer is described herein with respect to only one analyzer, for the sake of clarity, it is to be remembered that multiple analyzers can operate in the same of in different fashion, either simultaneously or at different times. Furthermore, steps of one method of operation can be combined with steps of another method of operation to arrive at yet more methods of operation.

As illustrated in FIG. 1, the analyzer comprises a process path 10. It is understood that there are other elements (not shown), such as fluid delivery mechanisms, suppliers, and the like, of the analyzer that support operation of the process path 10. While the process path 10 is illustrated as being substantially circular in configuration, the process path 10 may take other configurations, such as linear, serpentine, etc., as desired.

The process path 10 includes a cover 12 and a base 14. The base 14 may be attached to a support frame (FIG. 17) and the cover 12 is attached to the base 14. The cover 12 may be a single piece or may comprise multiple, sometimes 6, pieces. Various elements, some of which are described below, of the process path 10 are connected to at least one of the cover 12 and the base 14. The cover 12 and the base 14 include structures, such as openings and the like, for accommodating some of the elements. In one embodiment, the base 14 has an inner diameter of about 24.58 inches, an outer diameter of about 30.08 inches and a height of about 1.99 inches. The base 14 may be made of any suitable material, such as a metal, a polymer and the like. In one embodiment, the base 14 is made of anodized aluminum, including a reduced friction coating, such as a PTFE-impregnated anodized coating. In a particular embodiment, the base 14 is made from 6061-T6 aluminum with a MIL-

A-63576, Type I finish. The cover 12 may be made of a material which is substantially similar to the material of the base 14.

FIG. 2 shows the process path 10 with the cover 12 removed from the base 14. With the cover 12 removed, a disk 16 is visible. The disk 16 is located between the cover 12 and the base 14 and is movable with respect to both the cover 12 and the base 14.

In some embodiments, the disk 16 may be replaced by a belt 16\*, shown in FIGS. 32A and 32B, driven by a wheel 17. Use of the belt 16\* provides for orientations other than substantially circular, i.e. serpentine and like, of the process path 10. The belt 16\* moves with respect to the cover 12 and the base 14 in substantially the same manner as the disk 16. In other aspects, construction of the process path 10 is substantially similar irrespective of use of the disk 16 or the belt 16\*.

The disk 16, illustrated more clearly in FIG. 3, has, in one embodiment, an inner radius of about 25.2 inches and an outer radius of about 29.3 inches. The disk 16 may have a thickness of about 0.063 inches. The disk 16 may be formed from any suitable material, such as a polymer and the like. In a particular embodiment, the disk 16 is made from polyvinyl chloride. The disk 16 may be machined, molded or the like. In an exemplary embodiment, the material comprising the disk 16 is chosen with respect to the material of the base 14 to reduce friction between the base 14 and the disk 16.

A plurality 112 in the illustrated embodiment, of slots 18 are disposed on the disk 16. As is discussed in greater detail later, the slots 18 cooperate with structures on the base 14 to move containers 15 (FIGS. 7A and 7B) along the process path 10. Each slot 18 has, with respect to the disk 16 in an exemplary embodiment, a radial length of about 1.75 inches and a tangential width of about 0.45 inches with a slot 18 centerline being located at a radius of about 13.614 inches. As is discussed further below, the slot 18 has a longitudinal axis and the container 15 is capable of moving within the slot 18 along the slot's 18 longitudinal axis. To facilitate movement of the container 15 along the longitudinal axis of the slot 18, the process path 10 may include a configuration, such as a surface, a diverter, a prime mover engagable with the container 15, and the like. In another embodiment, one end of the slot 18 may include a latitudinally expanded width (FIG. 13) to facilitate removal of a container 15 from the disk 16. In still a further embodiment, the latitudinally expanded width may be located at another region of the slot 18 (FIG. 19).

The disk 16 is configured to facilitate movement of the disk 16 with respect to the cover 12 and the base 14. In one embodiment, a plurality of teeth 20 are disposed along an outer diameter surface of the disk 16. In an exemplary embodiment, the teeth 20 may be about 938 in number with a diametral pitch of about 32, a pressure angle of about 20 degrees and a pitch diameter of about 29.3125 inches.

As shown in FIG. 6, the teeth 20 mate with a gear 22 which is driven by a prime mover 24 attached to the cover 12 by a bracket 26. In an exemplary embodiment, the gear 22 is made from Estane 58130 natural 92A/50D polyurethane and the motor 24 is a P21 model available from Pacific Scientific of Rockford, Ill. The prime mover 24, the entire process path 10 and its supporting elements, are connected with and are operated by a suitable controller, such as a computer (not shown) running appropriate routine and the like. In this manner, the disk 16 moves responsive to movement of the gear 22 by the prime mover 24. In a particular embodiment, the prime mover 24 is a stepper motor.



Referring to FIG. 4, the base 14 includes structures to facilitate determination of an item of interest in a sample. The base 14 comprises at least one lane 28 for guiding movement of a container 15 along the process path 10 responsive to movement of the disk 16. As the disk 16 moves responsive to activation of the prime mover 24, the container 15 moves along the lane 28 from one processing station to another to complete determination of the item of interest in the sample.

In the illustrated embodiment, there are a first processing lane 28 and a loading lane 30 in the process path 10. Complimentary portions of the lanes 28 and 30 are formed in both the cover 12 and the base 14. Because these two lanes 28 and 30 are substantially concentric, the disk 16, which is adjacent to both lanes 28 and 30, and its slots 18 are dimensioned to accept and to support containers 15 disposed in both the process lane 28 and the loading lane 30 at substantially the same circumferential position, while being radially offset, on the disk 16. In an exemplary embodiment, the lanes 28 and 30 have a width of about 0.279 inches at the top and have a draft angle of about 1.5 degrees.

As shown in FIGS. 18A, 18B and 18C, in one embodiment, the loading lane 30 accepts and orients containers 15 from a container 15 supply or hopper 102. A disk 104 including a projection 106 is moved within the hopper 102 by a prime mover 108. In some embodiments, structures may be included with the hopper 102, such as a baffle for directing container 15 movement within the hopper 102 responsive to disk 104 movement, an "inherent flat spring" actuated by a cam driven mechanism associated with the disk 104 to move containers 15 within the hopper 102, and the like, to facilitate movement of the containers 15. As the disk 104 moves within the hopper 102, the projection 106 is inserted through the top surface 42 of a container 15 in the hopper 102. The projection 106 carries the container 15 toward a loading mechanism 110, which may include a mover 111, such as a barrel cam and the like, for moving a container 15 from the hopper 102 toward the loading lane 30. As the container 15 approaches the loading lane 30, in one embodiment, another mover 112, such as a solenoid-driven rod and the like, moves the container 15 into a slot 18 in the disk 16 at the loading lane 30. Alternatively, the container 15 may move from an end of the mover 111 into a slot 18 in the disk 16 at the loading lane 30 under the influence of gravity.

In an exemplary embodiment, the hopper 102 is made from Lexan WR2210 (GE Plastics of Pittsfield, Mass.) with a black SPI B1 finish and has a volume substantially within the range of about 396 to about 540 cubic inches, thereby allowing the hopper 102 to hold approximately 1000 containers 15. The disk 104 is made from Lexan 500 with a finish of gray SPI B1 and the projection 106 is made from Lexan WR2210 with a finish of black SPI B1. The disk 104 includes four projection 106 mounts spaced equidistantly along a circumference of the disk 104, i.e. every 90 degrees, at a radius of about 4.5 inches from a center of the disk 102. To assist movement of containers 15 within the hopper 102, the disk 102 includes a plurality, such as four, of nubs having a spherical radius of about 0.165 inches spaced equidistantly along a circumference of the disk 104, i.e. every 90 degrees, at a radius of about 3.312 inches from a center of the disk 102. The projection 106 has a nominal thickness of about 0.1 inches and a length of about 0.9 inches. The projection 106 is aligned substantially tangentially to a 4.5 inch radius of the disk 102. The mover 108 may be No. 78431-101 from Pacific Scientific of Elgin, Ill. The mover 111 includes a screw made from Delrin 500 having a black SPI B1 finish.

The screw is about 7.126 inches long and has 18 threads of a diameter measuring about 0.706 inches and of a pitch of about 0.394 inches. The screw is connected to a drive gear made from Celcon M90 having a finish of black SPI B1. The drive gear is an involute gear having 24 teeth with a diametral pitch of about 32, a pressure angle of about 20 degrees and a pitch diameter of about 0.75 inches. The mover 112 may be No. 78851-102 available from Haydon Switch & Instrument of Waterbury, Conn. In other embodiments, No. 78425-101 available from SPM/Portland of Hillsboro, Oreg. may be used for some of the components.

As shown in FIGS. 7A and 7B, the container 15 includes a sample receiving chamber 32 and a pair of support surfaces 34A and 34B connected with the sample receiving chamber 32. As shown in FIG. 8, the support surfaces 34A and 34B rest on portions of the disk 16 which bound the slot 18. The chamber 32 is formed by two sets of side walls 36A, 36B, 38A and 38B and a bottom wall 40. In an exemplary embodiment, the largest external distance between the side walls 36A and 36B, which have a rib width of about 0.020 inches, is about 0.26 inches, the largest external distance between the side walls 38A and 38B is about 0.44 inches, the support surfaces 34A and 34B extend a distance measuring about 0.085 inches from the side walls 38A and 38B, respectively, the maximum length of the container 15 is about 1.445 inches, an open end of the sample receiving chamber 32 measures about 0.391 inches by about 0.219 inches, a nominal thickness of the walls 36A, 36B, 38A and 38B is about 0.030 inches, an inside depth of the sample receiving chamber 32 is about 1.34 inches having a volume of about 1.4 ml and a volume of the sample receiving chamber 32 at a location, from which determination measurements are made, measuring about 0.699 inches from a bottom of the container 15 is about 0.45 ml. A top surface 42 of the container 15 is located a distance measuring about 0.18 inches from the support surfaces 34A and 34B. The container 15 may be made from Escorene 3345-E5 (Exxon, Houston, Tex.) or Montell PD701N (Wilmington, Del.) with an internal finish of polished SPE/SPE 1 B-2.

Returning to FIGS. 4 and 8, cooperation among the container 15, the slots 18 in the disk 16 and the lanes 28 and 30 facilitate movement of the container 15 along the process path 10. Specifically, the dimensions of the container 15, the slots 18 and the lanes 28 and 30 are predetermined such that the support surfaces 34A and 34B of the container 15 radially slidingly engage the disk 16 adjacent to the slot 18 in which the container 15 is disposed while the container 15 itself is restrained from rotation within the slot 18. In one embodiment, the process lane 28 has a radius of about 27.6 inches and a width of about 0.28 inches while the loading lane 30 has a smaller radius but a similar width. The container 15 is disposed such that axes of the side walls 36A and 36B are positioned substantially radially with respect to the process path 10 and the support surfaces 34A and 34B are aligned substantially circumferentially with respect to the process path 10. In this manner, as the disk 16 moves responsive to activation of the prime mover 24, the container 15 within the slot 18 moves substantially tangentially to the process path 10 within the lanes 28 and 30.

As the process path 10 may be used with biological samples, it is desirable to maintain the process path 10, or portions thereof, at a suitable temperature, such as 37 degrees Celsius, to facilitate determination of the item of interest. Thus, a heater (not shown), such as an electric heater and the like, may be thermally associated with the process path 10. In an exemplary embodiment, a plurality of



electric resistive flexible strip heaters may be applied, such as by a suitable adhesive and the like, to the cover 12 and/or the base 14 of the process path 10. These heaters apply sufficient thermal energy to the process path 10 such that the contents of the container 15 is maintained at the desired temperature. Also, because the loading lane 30 is part of the process path 10, it is possible to bring the container 15 to the desired temperature prior to addition of anything to the container 15. For example, if determination of an item of interest in a sample is performed optimally at a given temperature; the container 15 in the loading lane 30 can be brought to that given temperature at a certain time period after introduction of the container 15 from the hopper to the loading lane 30 but before the container 15 is needed to perform the desired determination. Suitable temperature control devices, such as thermistors and the like, are also provided along the process path 10. Additionally, in some embodiments, materials, such as reagents and the like, to be added to the container 15 may be heated prior to addition to the container 15. In some cases, the material delivery apparatus, such as a fluid conduit and the like, may be associated with appropriate heaters and heat sensors.

When a container 15 is needed to perform a given item of interest determination, the container 15 is moved from the loading lane 30 to the process lane 28. This function is performed at location 48 shown at FIG. 4. To move the container 15 from the loading lane 30 toward the process lane 28, as shown in FIG. 10, a prime mover 44, mounted on the process path 10, is operated. A container 15 engaging member 46 operatively connected with the prime mover 44 bears against the side wall 36A of the container 15 and moves the container 15 radially outward with respect to the disk 16 within the slot 18 from the loading lane 30 towards the process lane 28 responsive to activation of the prime mover 44. In an exemplary embodiment, the member 46 is made from 6061-T6 aluminum with a MIL-A-63576, Type I finish. The member 46 may include structures, such as a slot, which mate with complimentary structures, such as a pin, on the prime mover 44 to provide desired alignment of the mover 44 and the arm 46 and to limit undesired movement, such as rotation, of the member 46. Operation of the prime mover 44 causes the member 46 to move a distance of about 0.5 inches with a minimum starting force of about 7.08/25 gm/oz and a minimum ending force of about 56.7/2.0 gm/oz.

To accommodate movement of the container 15, a passageway 50 is formed on the cover 12 and the base 14 connecting the process lane 28 with the loading lane 30. Once the container 15 is in the process lane 28, the prime mover 44 moves the container 15 engaging member 46 away from the container 15 just moved to a waiting position to move another container 15 from the loading lane 30 toward the process lane 28. In an exemplary embodiment, the prime mover 44 is a solenoid, a pneumatically actuated motor, a linear positioner or the like. In a particular embodiment, the prime mover 44 is an electric solenoid with its windings modified such that the solenoid travel occurs without splashing or spilling of container 15 contents.

Now that the container 15 has been moved from the loading lane 30 to the process lane 28, movement of the disk 16 causes the container 15 to move along the process lane 28 for performance of determination of an item of interest in a sample. In some cases, the sample, such as blood or other bodily fluids, added to the container 15 is in liquid form. Also, in some cases, other substances, such as reagents and the like, are added to the sample in the container 15 during determination of an item of interest in the sample. These other substances may also be in liquid form.

As these liquids are added to the container 15 it is possible that some of the liquids may not end up within the container 15 but may be disposed on the disk 16 or other portions of the process path 10. To substantially remove these liquids, drain ducts 52 are provided on the base 14 of the process path 10. These drain ducts 52 are recessed from a groove 54 on the base 14 in which the disk 16 is disposed. In an exemplary embodiment, the drain ducts 52, about 112 in number, are equidistantly spaced along a circumference of the base 14, recess a distance of about 0.125 inches from the groove 54, have an internal angle of about 90 degrees and are about 0.05 inches deep and about 0.1875 inches wide. In some embodiments, the drain ducts 52 may be inclined toward the process lane 28 such that liquid within the drain ducts 52 will move under the influence of gravity toward and into the process lane 28. In the illustrated embodiment, the drain ducts 52 are oriented in an expected direction of disk 16 rotation. In this embodiment, liquid movement within the drain ducts 52 is encouraged by movement of the disk 16. Similar drain ducts 52 may be formed on the cover 12. To facilitate substantial removal of the liquids from the process lane 28, drain holes 56 are provided in the base 14 at various locations along bottom portions of the process lane 28.

The process of determining an item of interest in a sample comprises a number of steps. However, given the specific item of interest to be determined, different steps are to be performed. For instance, for determination of a first item of interest, three process steps are to be performed, whereas for a second item of interest, only two process steps are to be performed. These process steps may include, for example, solid/liquid phase (for example, magnetic) separation, aspiration of container 15 contents, container 15 contents washing, etc. To offer determination of both the first and second items of interest, the process path 10 includes structures for selective automated performance of process steps. However, it is to be noted that the process path 10 includes all structures necessary to perform all process steps for determining a predetermined set of items of interest.

At least one location along the process lane 28, structures or elements for providing selective automated performance of a determination of item of interest process step are disposed. As shown in FIG. 4, in one embodiment, these structures or elements are located in a bypass region of the process path 10. In the illustrated embodiment, the process path 10 includes three bypass regions 58A, 58B and 58C. At the bypass regions 58A, 58B and 58C, the process lane 28 is radially expanded with respect to other portions of the process lane 28. In an exemplary embodiment, the process lane 28 at the bypass regions 58A, 58B and 58C is about 0.65 inches wide radially. The radial expansion of the process lane 28 at the bypass regions 58A, 58B and 58C allows the container 15 to be positioned at multiple places longitudinally along the slot 18 and radially with respect to the disk 16 at the bypass regions 58A, 58B and 58C. Depending on the position of the container 15 within the slot 18 in the disk 16, the container 15 may or may not participate in the item of interest determination process step performed at the bypass regions 58A, 58B and 58C.

In an alternate embodiment, the structures or elements for providing selective automated performance of a determination of item of interest process step may include routines, such as those embodied in software, hardware and the like, for selectively activating or deactivating certain process path 10 elements, such as a wash zone and the like, selectively moving process path 10 elements into and out of a process step performance position with respect to the process path 10, such as moving a magnet and the like, or any appropriate combination of the methods discussed herein.



The cover 12 also includes structures forming the bypass regions 58A, 58B and 58C on the process path 10. As shown in FIGS. 5A and 5B, a wall 60 on the cover 12 separates the process lane 28 on the cover 12 at the bypass regions 58A, 58 and 58C into a process step performance lane 62 and a process step avoidance lane 64 offset radially on the cover 12. The wall 60 engages a portion of the side walls 36A and 36B adjacent of top surface 42 of the container 15 to guide the container 15 through either the process step performance lane 62 or the process path avoidance lane 64.

To encourage a desired container 15 into the desired one of the process step performance lane 62 or the process step avoidance lane 64, a prime mover 44 connected with a container engaging member 46 is provided attached to the process path 10, as shown in FIG. 9. The structure illustrated in FIG. 9 is substantially similar to the construction illustrated in FIG. 10, hence the like reference numbers. Activation of the prime mover 44 enables selective radial positioning of the container 15 at either an inner 66 or outer radial edge 68 (FIGS. 5A and 5B) of the process lane 28. Once so positioned, advancement of the disk 16 with respect to the base 14 moves the container 15 into the preselected one of the process step performance lane 62 or the process step avoidance lane 64.

In some embodiments, the prime mover 44, and/or the wall 60 may be constructed to take advantage of natural movement of the container 15 in the process lane 16. For instance, the container 15 may tend to move radially outwardly along the process lane 28. In this case, the prime mover 44 and/or the wall 60 may be constructed such that a container 15 moved toward the process step avoidance lane 64 moves toward that lane 64 under centrifugal force without any assistance from the prime mover 44. In this case, the prime mover 44 would only act on a container 15 to be moved into the process step performance lane 62.

In the illustrated embodiment, the bypass regions 58A, 58B and 58C are positioned along the process lane 28 dependent upon the anticipated frequency of performance and avoidance of a particular process step. This frequency is, in turn, dependent upon a particular step of determinations of items of interest to be performed with the process path 10. Also, depending upon the determinations to be performed, there may be more or less bypass regions 58A, 58B and 58C provided.

Illustrating further by example, the process lane 28 diverges radially prior to entering the bypass region 58A (FIGS. 5A and 5B). The process lane 28 enters the bypass region 58A along its outer radial edge. Since performance of the process step occurs at the inboard process step performance lane 62 of the bypass region 58A, the prime mover 44 associated with the bypass region 58A moves the container 15 radially inward toward the process step performance lane 62 only if performance of this process step were desired. If performance of this process step were not desired, then the prime mover 44 would not be activated and the container 15 would remain on the outer radius surface of the process lane 28 and move into the process step avoidance lane 64 upon movement of the disk 16. This construction favors performance of a set of determinations where performance of the relevant process step is required for a minority of the determinations to be performed.

If the set of determinations were to change such that performance of the relevant process step is required for a majority of the determinations to be performed, then it may be desirable to construct the bypass region 58A substantially similarly to the bypass regions 58B and 58C. At the bypass

regions 58B and 58C, the process lane 28 enters the bypass regions 58B and 58C at its inner radial edge. Thus, if the prime mover 44 is not activated, then the container 15 would move under the influence of movement of the disk 16 into the process step performance lane 62 and the process step would be performed. The prime mover 44 would be activated only to move those containers 15 that did not require performance of this process step. Of course, this would represent a minority of the determinations to be performed with the process path 10.

Once a container 15 is in one of the bypass regions 58A, 58B or 58C, movement of the container 15 through the bypass region 58A, 58B or 58C is controlled by cooperation among the disk 16, edges of the process step performance and avoidance lanes 62 and 64 and the wall 60. The container 15 moves substantially tangentially through the process path 10 under the influence of rotation of the disk 16. The position of the container 15 radially within the radially inner-most one of the process step performance lane 62 (e.g. bypass region 58A) or the process step avoidance lane 64 (e.g. bypass region 58B) is maintained by an inner radial edge of the wall 60. A radius defining this inner radial edge of the wall 60 gradually increases along the wall 60 from a first end 70 to a second end 72 thereof. The container 15 is moved radially outward as the container 15 moves through the bypass region 58A, 58B or 58C. A radius defining an inner edge of the process step performance lane 62 (e.g. bypass region 58A) and the process step avoidance lane 64 (e.g. bypass region 58B) also increases from one end of the bypass region 58A, 58B or 58C adjacent the first end 70 of the wall 60 to an opposite end of the bypass region 58A, 58B or 58C adjacent the second end 72 of the wall 60. Thus, a portion of the container 15 adjacent its top surface 42 is maintained adjacent the wall 60, thereby maintaining intended positioning of the container 15 within the bypass regions 58A, 58B and 58C.

Once the determination of an item of interest is complete, the relevant container 15 is removed from the process lane 28 and the process path 10 altogether. As shown in FIG. 11, a prime mover 74 is connected with the process path 10. The prime mover 74 drives a container 15 engaging surface 76 which acts on the container 15 adjacent the top surface 42 of the container 15. The prime mover 74, which may be a stepper motor and the like, drives the container engaging surface 76 to rotate the container 15 about 90 degrees with respect to the disk 16. This occurs at location 78 shown in FIG. 4. The process path 10 at the location 78 is configured to allow axial rotation of the container 15 and includes an aperture 80 having dimensions larger than corresponding dimensions of the container 15.

In an exemplary embodiment, the prime mover 74 may be a solenoid such as P/N 197855-001 BTA 2 DV 90° available from Lucas Control Systems Products of Vandalia, Ohio. The surface 76 may be made from 6061-T6 aluminum with a MIL-A-63576, Type I finish and moves approximately 90 degrees responsive to operation of the prime mover 74.

Once the container 15 has been rotated, the support surfaces 34A and 34B of the container 15 are no longer in engagement with the disk 16. Under the influence of gravity, the container 15 falls through the aperture 80 in the process path 10 into a waste receptacle (not shown). In some constructions, a chute may be provided to guide the container 15 from the process path 10 toward the waste container. In other constructions, liquid present in the container 15 may be removed from the container 15 prior to encountering the prime mover 74.

With the container 15 being removed from the process lane 28, another container 15 within the same slot 18 on the



disk 16 can be moved from the loading lane 30 to the process lane 28 as soon as the relevant slot 18 reaches the location 48. In some instances, it may not be desirable to remove a container 15 from the process lane 28 once that container 15 reaches location 78. In this case, the prime mover 74 will not be activated. Also, a container 15 disposed within the same slot 18 on the disk 16 but in the loading lane 30 will not be moved from the loading lane 30 to the process lane 28 when the relevant slot 18 reaches location 48.

In an alternative embodiment shown in FIGS. 13 and 19, the disk 16 is constructed to facilitate removal of a container 15 from the process path 10. In this embodiment, the slots 18 on the disk 16 include an enlarged container 15 removal area 82. Also, a diverter 84 is disposed in the process lane 28 adjacent to the location 78. The diverter 84, along with movement of the disk 16, urges the container 15 radially outward with respect to the disk 16 toward the container removal area 82 of the slot 18. The container removal area 82 is wider than the remainder of the slot 18 such that, when the container 15 reaches the container removal area 82 of the slot 18, gravity causes the container 15 to fall from the disk 16 and the process path 10 through the aperture 80 and into the waste receptacle. However, this embodiment does not allow a container 15 to pass the location 78 and still remain with the disk 16. But, if it were desirable to allow the container 15 to remain with the disk 16 in this embodiment, then the diverter 84 may be replaced with a prime mover, similar to the prime mover 44, to move the container 15 within the slot 18 toward the container removal area 82.

Another construction of the disk 16, the slot 18 and the container removal area 82 is shown in FIG. 19. This construction functions in a manner substantially similar to that of FIG. 13. It is to be noted that, in the embodiment illustrated in FIG. 19, an end of the process lane 28 is defined by the aperture 80.

Additional features may be incorporated into the process path 10 as desired. For example, liquid level sensing devices, such as radio frequency liquid level sense devices and the like, may be incorporated at positions along the process path 10 where liquid movement may occur. Also, any suitable structures, such as any of those disclosed in U.S. Pat. Nos. 5,358,691, 5,536,471 and 5,482,861 may be added, sometimes with appropriate modifications. Those patents are assigned to the assignee of the present case and the disclosures thereof are incorporated herein in their entirety by this reference.

It may also be desirable to construct the process lane 28 to reduce light in portions of the process lane 28. In one embodiment, the process lane 28 is constructed such that there is a radial divergence of that lane 28 prior to and following any position on the process path 10 where light, such as chemiluminescently generated light, measurements are performed. Such a radial divergence of the process lane 28 may increase the sensitivity of the light measurer by reducing introduction of stray or ambient light into the light measuring position of the process lane 28.

The process path automated described above allows sequential automated performance of multiple determination of item of interest process steps. The motion of a container 15 along the process lane 28 may be executed in discrete steps, that is discrete with respect to time and with respect to position along the process lane 28. At regular time intervals, such as about 18 seconds, the disk 16 rotates a distance substantially equal to the angular distance between two adjacent slots 18. This rotation causes each container 15 to move to the position along the process path 10 previously

occupied by the container 15 in the adjacent slot 18. The disk 16 and the container 15 remain stationary for a remainder of the regular time period prior to the next rotation or indexing of the disk 16. The process lane 28 may be considered as having a fixed number of process positions, positions at which a process step comprising the determination of an item of interest in a sample occur, equal to the number of slots 18 in the disk 16.

In the examples described here, there are 112 slots 18 in the disk 16, and consequently the process lane 28 may be considered as having 112 process positions. The total processing time of a container 15 and its contents may be thought of as integral multiples of the index period. For example, if the index period is 18 seconds, a container 15 in the 10th process position has undergone a total of 180 seconds of processing. Similarly, a process step that is performed over 20 process positions takes a total of 360 seconds of process time on an individual container 15.

An example of process steps that may be performed during determination of an item of interest in a sample may be described by specifying the process position at which each process step occurs, as is provided in the following examples. This example may be more easily understood with reference to FIG. 16. The dotted line 129 indicates a boundary of a support on which the process path 10 is mounted.

A reagent carousel 131 is located substantially concentrically with the process path 10 and is rotatable. The reagent carousel 131 may include one or more carousels and may provide for axial rotation of individual containers, i.e. magnetic microparticle containers, disposed thereon. In one embodiment, the reagent carousel 131 may include multiple substantially concentric carousels to provide simultaneous and/or shared access of multiple containers by multiple pipette assemblies, such as assemblies 128 and 134. Such an arrangement may facilitate performance of the Formats discussed later. The reagent carousel 131 may be constructed substantially similarly to the structure disclosed in GB 2,081,118B issued on Sep. 7, 1983, with appropriate, well known bearings and gear trains being provided as and where needed (See FIG. 24B), as disclosed on Page 3, lines 86-91 of that patent. In an exemplary embodiment, the carousel 131 may be No. 77829-101 available from SPM/Portland of Hillsboro, Oreg., with appropriate motors available from Pacific Scientific, gears from Turnamatic of Richardson, Tex. and SPM/Portland and sensors from Aromat of Rolling Meadows, Ill.

The reagent carousel 131 may be maintained within a thermostatically controlled environment. The thermostatically controlled environment may be provided by an air cooling unit which provides forced cooled air to a housing 133 (FIGS. 29 and 30) containing the reagent carousel 131. In an exemplary embodiment, the housing 133 may be similar to No. 76848 available from General Pattern of Blaine, Minn. This may reduce evaporation of fluid from the containers held on the reagent carousel 131. To further reduce evaporation, open mouths of the containers may be fitted with a seal 184 as shown in FIG. 25. The seal 184 may be made of a polymeric material, such as an elastomer and the like, and may include a slit 186 for allowing a pipettor access to the interior of the container.

In one embodiment, the reagent carousel 131 supports a plurality of reagent containers. These containers may be of at least four types, such as microparticle, conjugate, determination specific diluent and pretreatment, dependent upon the type of reagent contained therein. FIGS. 22, 23A, 23B



and 23C give two exemplary configurations of the containers. A bottom portion 174 of the containers 176 (FIG. 22) and 177 (FIGS. 23A, 23B and 23C) is constructed to fit with mating portions of the reagent carousel 131.

As shown more clearly in FIGS. 24A and 24B, the bottom portion 174 of the container 177 bears a projection 178 which engages a complementary portion 188 of the reagent carousel 131. The engagement between the projection 178 and the portion 188 of the reagent carousel 131 provides a user who is placing the container 177 on the reagent carousel 131 with positive feedback, i.e. tactile feel, indicative of proper positioning of the container 177 with respect to the carousel 131.

As shown in FIG. 24B, the portion 188 of the carousel 131 is operatively connected by a shaft 191 with a drive gear 190 which drivingly engages a gear 202 which is connected with a prime mover (not shown). The gear 202 engages all drive gears 190 associated with the carousel 131. Operation of the prime mover moves the gear 202 which, in turn, moves the gear 190. Movement of the gear 190 causes axial rotation, which may be bi-directional, of the portion 188 and the container 177. The shaft 191 also electrically contacts a plate 204 which is electrically connected with a conductor 206. In this manner, the plate 204 and the conductor 206, and possibly the portion 188 of the carousel 131, if it is electrically conductive, comprise a portion of a radio frequency liquid level sense mechanism which determines a fluid level inside the container 177.

To further facilitate manipulation of the container 177, a substantially annular rib 180 (FIGS. 23A, 23B and 23C) may be provided on an outer surface of the container 177. Also, if it were desirable to maintain the container contents in a substantially homogeneous state, i.e. magnetic particles substantially uniformly dispersed in a liquid medium, then at least one fin 182 (FIGS. 24A and 24B) may be provided on an interior, fluid facing surface of the container 177 to agitate container contents upon axial rotation, as discussed above, of the container 177.

Illustrating constructions of the containers and seals with specific examples, the containers may be made from DOW 30460M HDPE or Chevron 90512 HDPE with a finish of SPI A3. The fins 182 may have a finish of SPI C1. The seals may be made from Lexington Medical 3401005 EPDM. The containers may have a neck inner diameter measuring about 1.069 inches. The rib may have a thickness of about 0.025 inches, a width, from an inner wall of the container, measuring about 0.31 inches, a top geometry measuring about 45 degrees, and a bottom geometry tapering to a center at an angle of about 48 degrees. The seal may have a diameter of about 1.094 inches when installed with a container, a maximum thickness of about 0.070 inches at a centerline of the seal, and a reinforced hinge section measuring about 0.025 inches thick by about 0.062 inches deep from an underside of a pipettor contact area on the seal. The slit on the seal may comprise two slits having a length of about 0.5 inches through a center of the seal and offset about 90 degrees from each other.

To facilitate identification of the containers, at least some of the containers may bear a label 133A, 133B, or 133C, substantially similar to those shown in FIGS. 21A, 21B and 21C. The labels 133A, 133B and 133C include a high density data carrier 135A, 135B and 135C, respectively, which includes information to facilitate performance of the determinations.

In a specific embodiment, the high density data carrier 135A, 135B and 135C is a two dimensional bar code

utilizing PDF 417 technology to provide desired data capacity. This technology allows for inclusion of more information than a common one dimensional bar code. Usage of such a high density data carrier 135A, 135B and 135C provides structural flexibility, i.e. individual containers for a given determination do not have to be physically joined together. The data carrier 135A, 135B and 135C contains information desirable for performance of a given determination. This information may include master lot number, container lot number, container contents, i.e. reagent, lot number and expiration date, calibration curve data, container contents type, etc. The information may also contain a serial number specific to the particular container to facilitate tracking of process path 10 resources.

In the illustrated embodiment, the data carrier 135A is used with magnetic microparticle containers and holds approximately 185 characters of information. The data carrier 135A is approximately 1.5 inches tall and about 0.75 inches wide, as viewed by the bar code reader. Because the microparticle container is rotated as discussed above, this rotation may be utilized while reading the data carrier 135A. In this case, the orientation of the data carrier 135A with respect to the bar code reader may not be important.

The data carriers 135B and 135C of the illustrated embodiment comprise two dimensional bar codes containing about 15 characters of information each. The data density of the carrier 135B and 135C is adjusted to allow the carrier 135B and 135C to be about 0.7 inches high. Furthermore, the data carrier 135B and 135C is printed with error correction, X bar, Y bar and a column count that allows the carrier 135B and 135C to be about 3.125 inches wide. In this manner, the data carrier 135B and 135C can be disposed along an outer circumference of a container such that the carrier 135B and 135C is accessible to the bar code reader through approximately 220 to approximately 270 degrees of visibility, depending on container size. Alternatively, instead of the carrier 135B which includes only one bar code, the data carrier 135C includes a plurality of repetitions of a similar, but narrower in form bar code with gaps between adjacent code repetitions. Additionally, various modifications of the data carriers 135A, 135B and 135C are also possible. For instance, one dimensional bar codes could be used, but the surface area of the one dimensional bar code would have to be sufficient for the amount of data contained in the two dimensional bar code.

#### EXAMPLE

##### DETERMINING AN ITEM OF INTEREST IN A SAMPLE

The process path 10 illustrated in FIG. 1 is utilized to perform a sequence of process steps, executed with a index period of about 18 seconds. Each index step comprises about 1 second of rotation of the disk 16 (and consequent motion of the containers 15 disposed within the disk 16) and about 17 seconds during which the containers 15 are stationary at their respective process positions. The process step performed at each process position is as follows:

Process Position	Process Step	Description
1	Container 15 load	Container 15 moved from loading lane 30 to process lane 28 as required
1	Sample Pipettor	Sample deposited into container 15 by pipetting system 128. The sample may be obtained from



## 15

-continued

Process Position	Process Step	Description
		position 130A or 130B which are located on appropriate conveyors, sample handlers or structures associated with a laboratory automation system
2	Reagent Pipettor 1	Reagent obtained from reagent carousel 131 deposited into container 15 by pipetting system 132. Liquid present in the pipetting system 132 may also be added to the container 15.
3	Mixer	Contents of container 15 are mixed by a device 86 imparting motion to the container 15
4-23	Incubation	Contents of container 15 are incubated at a controlled temperature, about 37 degrees Celsius
24	Sample Pipettor	Container contents may be aspirated from container 15 by pipetting system 128 for deposition into a second container 15 at position 1
25-39	Incubation	Contents of container 15 are incubated at a controlled temperature
40	Bypass region 58A start	Container 15 is selectively positioned at entry to performance lane 62 or avoidance lane 64 of bypass region 58A
41	Wash zone 1	Container 15 in performance lane 62 undergoes magnetic separation and fluid addition
42	Wash zone 1	Container 15 in performance lane 62 undergoes magnetic separation, container 15 contents aspiration and fluid addition
43	Wash zone 1	Container 15 in performance lane 62 undergoes magnetic separation, container 15 contents aspiration and fluid addition
44	Wash zone 1	Container 15 in performance lane 62 undergoes magnetic separation and container 15 contents aspiration
45.5	Bypass region 58A end	Performance lane 62 and avoidance lane 64 of bypass region 58A merge (midway between positions 45 and 46)
46	Container 15 load into loading lane 30	New containers 15 are loaded into loading lane 30
48	Reagent Pipettor 2	Reagent selectively deposited into container 15 by pipetting system 134
49	Mixer	Contents of container 15 are mixed by a device 86 imparting motion to the container 15
50-62	Incubation	Contents of container 15 are incubated at a controlled temperature
63	Bypass region 58B	Container 15 is selectively positioned at entry to performance lane 62 or avoidance lane 64 of bypass region 58B
64	Wash zone 2	Container 15 in performance lane 62 undergoes magnetic separation and fluid addition
65	Wash zone 2	Container 15 in performance lane 62 undergoes magnetic separation, container 15 contents aspiration and fluid addition
66	Wash zone 2	Container 15 in performance lane 62 undergoes magnetic separation, container 15 contents aspiration and fluid addition
67	Wash zone 2	Container 15 in performance lane 62 undergoes magnetic separation, and container 15 contents

## 16

-continued

Process Position	Process Step	Description
5		aspiration
68	Bypass region 58B end	Performance and avoidance lanes 62 and 64 of bypass region 58B merge
69-70	Incubation	Contents of container 15 are incubated at a controlled temperature
10	71	Reagent Pipettor 2
	72	Mixer
15	73-86	Incubation
	75	Motor/Encoder
20	77.5	Home Sensor
	86	Bypass region 58C
25	87	Wash zone 3
	88	Wash zone 3
30	89	Wash zone 3
35	90	Wash zone 3
	91	Bypass region 58C end
40	91-93	Incubation
	94	Pre-Trigger and Mixer
45	95-97	Incubation
	98	Shutter, reader, and trigger
50	99	Magnet
	100	Liquid Waste Aspirate
55	109	Container 15 unload
	111	Container 15 unload sensor
60		

Adding more specificity to the example, in a particular embodiment, the determination of an item of interest in a sample is an immunoassay. When the process path 10 is used to perform an immunoassay, the container 15 is moved into the process lane 28 at position 1. Also at position 1, a known



quantity of sample (for example, 50  $\mu$ l of blood) is deposited into the container 15 by a pipetting system 128. The pipetting system 128 comprises a pipettor, which may be substantially similar to the pipettors 116A, 116B and 116C, mounted on an arm for movement up and down and angularly, as shown in FIG. 16.

After indexing the container 15 to position 2, a known quantity of a first reagent, possibly along with an amount of fluid present in the pipetting system 132, is deposited into the container 15 by a second pipetting system 132. The first reagent may contain magnetically responsive microparticles coated with antibodies or other binding substances that specifically bind to the item of interest in the sample. The first reagent may be added with an assay specific diluent. In some cases, the first reagent and a conjugate, possibly along with an amount of fluid present in the pipetting system 132, may be added at position 2.

At position 3, a mechanical device 86 (illustrated in FIG. 12) is provided to mechanically move the container 15 and cause mixing of the contents of the container 15. The mechanical mixing device 86 includes a bore 88 formed within a body 89, which is eccentrically formed in the illustrated embodiment, that moves axially and rotatably under the influence of a prime mover 90 connected with the body 89. When the prime mover 90 is activated, the body 89 rotates clockwise, and a protrusion 92 connected with the prime mover 90 moves within a slot 94 in a second body 96. The second body 96 rotates freely about a drive shaft of the prime mover 90.

As the protrusion 92 moves within the slot 94, the body 89 and the bore 88 move toward the bottom 40 of the receptacle 15 as the body 89 rotates. When the body 89 and the bore 88 move toward the container 15, the bore 88 engages the bottom 40 of the container 15 and imparts an orbital (but not rotational) motion to the bottom 40 of the container 15. The portions of the container 15 adjacent the top surface 42 remain relatively stationary within the slot 18 in the disk 16.

The mechanical motion imparted to the container 15 mixes the sample with the first reagent. After the contents of the container 15 have been mixed for a predetermined time period, the prime mover 90 rotates its drive shaft counterclockwise, causing the protrusion 92 to move in an opposite direction within the slot 94, thereby moving the first body 89, the bore 88 and the second body 96 away from the bottom 40 of the container 15.

Illustrating further with a specific example, in one embodiment, the body 89 is made of PEEK with a black finish, the protrusion 92 is made of AISI 301 stainless steel with a #10 passivated finish, the second body 96 is made of Acetron GP with a white finish and the slot 94 has a #32 finish. The bore 88 in the body 89 is offset from an axis of the body 89 and has a radius of about 0.020 inches. An interface between the body 89 and the container 15 provides a minimum of about 0.05 inches of eccentric rotation of the container 15. The slot 94 provides a rise of about 0.315 inches over a rotation of the second body 96 of about 226.8 degrees. The prime mover 90 is a 3 phase, 8 pole, Y connected DC brushless motor P/N DR538-504 available from Shinano Kenshi of California. The prime mover 90 is supplied with a 36 Volt potential and operates substantially within the range of about 500 to about 5500 rpm's with a torque constant of about 620 g\*cm/A.

The container 15 is freed from the bore 88 and processing of the container 15 contents continues. Subsequently, the container 15 contents is incubated for a predetermined time period.

At position 24, depending upon the particular item of interest in the sample to be determined, the first pipetting system 128 may withdraw a portion of the contents of the container 15 for deposition into another container 15 located at position 1. This may be appropriate when a particular determination requires pretreatment, such as pre-heating, heated incubation with a first reagent prior to second reagent introduction, and the like, prior to introduction of magnetically responsive microparticles comprising the first reagent. At position 37, the process path 10 selectively positions the container 15 for performing or avoiding a series of magnetic separation and wash steps. Structures for performing the wash and separation comprise a wash station 114, shown in FIGS. 14 and 15.

Each wash station 114 includes a plurality, 3 in the illustrated embodiment, of movable pipettors 116A, 116B and 116C and at least one stationary nozzle (not shown) for moving fluids at least one of into and out of the container 15. In some embodiments, the movable pipettors 116A, 116B and 116C may be used to move fluids out of the container 15 while the at least one stationary nozzle moves fluid into the container 15. Sensors, such as thermistors and the like, may be operatively associated with the pipettors 116A, 116B and 116C to verify fluid movements.

The pipettors 116A, 116B and 116C move liquids both into and out of the container 15 whereas the nozzle only moves liquid into the container 15. The movable pipettors 116A, 116B and 116C are connected to a common base plate 118 which moves with respect to the cover 12 under the influence of a prime mover 120, such as a stepper motor and the like. Responsive to the prime mover 120, the pipettors 116A, 116B and 116C move into and out of the container 15. Suitable fluid delivery conduits, not shown, are connected with the pipettors 116A, 116B and 116C and the nozzle. The pipettors 116A, 116B and 116C are spring-loaded to facilitate their replacement and to cushion any contact between the pipettors 116A, 116B and 116C and another surface, such as the bottom 40 of the container 15 and the like.

The pipettors 116A, 116B and 116C are movable to remove fluid from the container 15. Because the item of interest is connected with the magnetic particles, a magnet assembly 122 is also included at the wash station 114. The magnet assembly 122 is disposed in a receptacle 124 in the base 14. The magnet assembly 122 includes a portion of the performance lane 62 and holds a plurality of permanent magnets 126. In an exemplary embodiment, the assembly 122 is made from 6061 T6 aluminum with a finish of MIL-A-63576 Type I and the magnets 126 are neodymium Iron Boron (NdFeB) magnets with a residual flux density (Br) substantially within the range of about 12.1 to about 13.2 KG, a coercive force (Hc) substantially within the range of about 11.0 to about 12.0 KOe, an intrinsic coercive force (Hci) substantially within the range of about 17.0 to about 19.0 KOe and a total energy product (BHmax) substantially within the range of about 34.0 to about 41.0 MGOe. The field intensity of the magnets 126 at a distance of about 0.030 inches from the container 15 is about 4470 Gauss and at a distance of about 0.176 inches from the container 15 is about 1570 Gauss.

At the wash station 126, the magnets 126 hold the magnetic particles, and thereby the item of interest, against a side wall 36A or 36B of the container 15. This allows removal of contents of the container 15 other than the magnetic particles and the item of interest bound to the magnetic particles. In some constructions, the pipettors 116A, 116B and 116C may be positioned such that the pipettors 116A, 116B and 116C move substantially along a



central axis of elongation of the container 15, may be biased away from a side wall 36A or 36B against with the magnetic particles are held, or otherwise constructed to reduce the chances of the pipettors 116A, 116B and 116C removing magnetic particles and the item of interest bound to the magnetic particles from the container 15.

In an exemplary embodiment, the pipettors 116A, 116B and 116C are made from Inconel. The pipettors 116A, 116B and 116C are disposed such that longitudinal center lines of the pipettors 116A, 116B and 116C are offset a distance measuring about 0.029 inches from a center line of the containers 15 into which the pipettors 116A, 116B and 116C are inserted. This offset distances the pipettors 116A, 116B and 116C from magnetic particles within the containers 15. When the pipettors 116A, 116B and 116C dispense fluid into the containers 15, the pipettors 116A, 116B and 116C are located a distance measuring about 0.342 inches from a side wall of the containers 15 adjacent the magnets 126. The pipettors 116A, 116B and 116C are mounted with springs so as to absorb up to 0.1 inches of overdrive. The pipettors 116A, 116B and 116C are fluidly connected with a valve which allows for bubble flushing without use of a container 15. The stationary nozzle is made of 0.031 inch inner diameter PEEK tubing. The base plate 118 is a two piece, thermally bonded assembly of acrylic with a clear Iridite finish on top and an opaque finish on the bottom to allow fluid visibility and light protection for a chemiluminescence reader.

If, for a particular determination, magnetic separation and washing is required at position 37, the container 15 is moved to the performance lane 62. Containers 15 in the performance lane 62 undergo, at each processing position between 41 and 44, magnetic separation (effected by permanent magnets 126 at fixed locations adjacent to the performance lane 62), fluid aspiration, and fluid dispensing, performed by fluid handling devices introduced through an opening 98 (FIG. 1) in the cover 12. In one embodiment, one of these wash stations (position 41) includes only a magnetic separation and fluid dispensing step that introduces a wash buffer to the container 15. In some cases, wash buffer or other fluid is added such that the amount of fluid present within the container 15 facilitates separation (magnetic) of the particles from the fluid in the container 15. At positions 42 and 43, separation, fluid aspiration, and fluid dispensing occur. In position 44, the magnetic particles are separated from the fluid in the container 15 by magnets 126 and fluid is aspirated. In this example, these steps would remove substantially all substances within the container 15 that have not bound to binding conjugate elements on the magnetic particles deposited as the first reagent. Containers 15 within the avoidance lane 64 are undisturbed and continue incubation. The performance and avoidance lanes 62 and 64 merge between positions 45 and 46.

A second reagent may be deposited into the container 15 at location 48 (FIG. 4) by a third pipetting system 134, again followed by a mechanical device 86 at position 49 to mix the container 15 contents. The second reagent may include an indicator substance, such as a chemiluminescent substance, linked to binding elements that also bind to the item of interest (remaining occurrences of which are bound to the magnetic particles of the first reagent). The contents of the container 15 are incubated at positions 50-59.

The second bypass region 58B begins at position 60, where the container 15 may selectively automatically undergo a set of magnetic separations, fluid aspirations, and fluid dispensing steps.

The third pipetting system 134 may deposit a third reagent into the container 15 at position 71, with subsequent mixing at position 72 and incubation between positions 73 and 86.

The third bypass region 58C begins at position 86, where the container 15 may selectively automatically undergo a set of magnetic separations, fluid aspirations and fluid dispensing steps.

In one embodiment, where it is assumed that substantially a majority of the containers 15 will undergo magnetic separation, fluid aspiration, and fluid dispensing at positions 87-90, no bypass region 58C may be provided at these locations. For example, these step would cause the removal of substantially all indicator (chemiluminescent) substances that are not bound to the magnetic particles (via the analyte of interest), yielding a container 15 holding indicator substance in an amount indicative of the amount of the item of interest in the initial sample deposition. However, in some determinations, it is desirable to avoid those process steps.

A pretrigger reagent may be deposited by a fluid dispensing device at position 94.

A fluid dispensing device will deposit a triggering agent at position 98, which causes the indicator reaction to occur. For example, a chemiluminescent substance releasing reagent may be deposited at position 94, which causes the release of the indicator (chemiluminescent) substance from the magnetic particles.

The contents of the container 15 are incubated between positions 95 and 97, inclusive.

Position 98 may also include a magnet, which separates or removes substantially all of the magnetic particles from the fluid within the container 15. The magnet holds substantially all of the magnetic particles against a side wall 36A or 36B of the container 15 prior to reading of light from the chemiluminescent substance. Preferably, all of the magnetic particles are removed from a path of chemiluminescent photons from the chemiluminescent substance, which remains in solution in the fluid in the container 15, to a light detection apparatus 138. This read step is substantially similar to that described in EP 0 371 265 B1 issued Jan. 1, 1994. The introduction of the triggering reagent would initiate a chemiluminescent reaction which would be detected and quantified by an optical detection system (not shown) such as a photomultiplier tube or photon counting system.

In an exemplary embodiment, the apparatus 138 may comprise a reader assembly such as No. 78262 available from Thorn EMI of Rockaway, N.J., a photomultiplier tube such as No. 78252-101 available from Hamamatsu of Middlesex, N.J. and a substantially light-tight shutter operable by a plunger such as No. 78200-101 available from Ironwood Industries of Libertyville, Ill. and a motor such as No. 78851-101 available from Haydon Switch & Instrument of Waterbury, Conn.

The embodiment described in the following examples demonstrates its utility in processing multiple assays of different formats and timing requirements within a common process path 10. In these examples, the embodiment described enables the execution of at least the following four assay formats, the first three of which may be executed simultaneously with no degradation in processing capacity.

Format A	
Step	Position
Sample introduction	1
First reagent introduction and mixing	2-3
First incubation (18 minutes)	4-63



-continued

<u>Format A</u>	
Step	Position
Separation and wash	64-67
Second reagent introduction and mixing	71-72
Second incubation (4 minutes)	73-86
Separation and wash	87-90
Pretrigger introduction and mixing	94
Third incubation (1 minute)	95-97
Trigger and read	98

As an example, Format A may be used to determine at least the following items of interest: antibodies to HCV, antibodies to HIV 1/HIV 2, antibodies to hepatitis B core antigen (HBcAb), carcinoembryonic antigen (CEA), cancer antigen 19-9 (CA19-9), Hepatitis B Surface Antigen (HBsAg), antibodies to Hepatitis B Surface antigen (HBsAb), alpha-fetoprotein (AFP), Total prostate specific antigen (Total PSA), Free PSA, Thyroid stimulating Hormone (TSH), luteinizing hormone (LH), follicle stimulating hormone (FSH), beta human chorionic gonadotropin (B-hCG), Free Thyroxine (Free T4), Free triiodothyronine (Free T3), Total T4, Total T3, Prolactin and Ferritin. It is to be noted that almost any item of interest discussed herein may be determined by properly using this format. For instance, this format may also be used to determine beta human chorionic gonadotropin (B-hCG), prolactin and ferritin.

Format B

Step	Position
Sample introduction	1
First reagent introduction and mixing	2-3
First incubation (11 minutes)	4-40
Separation and wash	41-44
Second reagent introduction and mixing	48-49
Second incubation (11 minutes)	50-86
Separation and wash	87-90
Pretrigger introduction and mixing	94
Third incubation (1 minute)	95-97
Trigger and read	98

As an example, Format B may be used to determine an item of interest in a sample where a relatively increased degree of sensitivity, as compared with some other formats, is desired. It is to be noted that almost any item of interest discussed herein may be determined by properly using this format.

Format C

Step	Position
Sample introduction	1
First reagent introduction and mixing	2-3
First incubation (11 minutes)	4-40
Separation and wash	41-44
Second reagent introduction and mixing	48-49
Second incubation (4 minutes)	50-63
Separation and wash	64-67
Third reagent introduction and mixing	71-72

-continued

Format C

Step	Position
Third incubation (4 minutes)	73-86
Separation and wash	87-90
Pretrigger introduction and mixing	94
Fourth incubation (1 minute)	95-97
Trigger and read	98

As an example, Format C may be used when the item of interest relates to hepatitis, such as determinations for anti-M, HBcAb-M and HAVAb-M.

Format D

Step	Position
Sample introduction	1
First reagent introduction and mixing	2-3
First incubation (7 minutes)	4-23
Transfer to second container 15 in position 1	24
Second reagent introduction and mixing	2-3
Second incubation (11 minutes)	4-40
Separation and wash	41-44
Third reagent introduction and mixing	48-49
Third incubation (4 minutes)	50-63
Separation and wash	64-67
Fourth reagent introduction and mixing	71-72
Fourth incubation (4 minutes)	73-86
Separation and wash	87-90
Pretrigger introduction and mixing	94
Fifth incubation	95-97
Trigger and read	98

Format E

Step	Position
Sample introduction	1
First reagent introduction and mixing	2-3
First incubation (7 minutes)	4-23
Transfer portion of container 15 contents to second container 15, remainder of container 15 continues on process lane 28	24
First container 15 contents continues first incubation (11 minutes)	24-63
Second reagent introduction and mixing with second container 15 contents	2-3
First container 15 passes through bypass region 58B	64-67
Second container 15 first incubation (18 minutes)	4-63
Introduction of fourth reagent into first container 15 and mixing (optional, enhances total Hb chemiluminescent signal)	71-72
Second container separation and wash	64-67
Fourth incubation (4 minutes - optional) of first container 15	73-86
Third reagent	71-72

-continued

Step	Position
introduction into second container 15 and mixing	
First container 15 passes through bypass region 58C	87-90
Third incubation (4 minutes) of second container 15	73-86
Pretrigger introduction into first container 15 and mixing	94
Separation and wash of second container 15	87-90
Trigger and read value 1 (Total Hb) from first container 15	98
Pretrigger introduction into second container 15 and mixing	94
Trigger and read value 2 (GlyHb)	98

$$\text{Reported result} = \frac{\text{value 2}}{\text{value 1}} \times 100$$

For example, in Format E, it is possible to modify the format by disregarding the first container 15 after the portion of the container 15 contents has been transferred (Position 24) to the second container 15. In that case, Format E may be used to determine, for example, folate and vitamin B12.

Format F

Step	Position
Sample introduction	1
First reagent introduction and mixing	2-3
First incubation (7 minutes)	4-23
Transfer portion of container 15 contents to second container 15, remainder of container 15 continues on process lane 28	24
First container 15 contents continues first incubation (11 minutes)	24-63
Second reagent introduction and mixing with second container 15 contents	2-3
First container 15 passes through bypass region 58B	64-67
Second container 15 first incubation (11 minutes)	4-40
Introduction of fourth reagent into first container 15 and mixing (optional, enhances total Hb chemiluminescent signal)	71-72
Second container separation and wash	41-44
Fourth incubation (4 minutes-optional) of first container 15	73-86
Third reagent introduction into second container 15 and mixing	48-49
First container 15 passes through bypass region 58C	87-90
Third incubation (11 minutes) of second	50-86

-continued

Format F

Step	Position
container 15	
Pretrigger introduction into first container 15 and mixing	94
Separation and wash of second container 15	87-90
Trigger and read value 1 (Total Hb) from first container 15	98
Pretrigger introduction into second container 15 and mixing	94
Trigger and read value 2 (GlyHb)	98

$$\text{Reported result} = \frac{\text{value 2}}{\text{value 1}} \times 100$$

This format may be used, for example, to determine at least one of total and glycated hemoglobin. Also, this format may be modified by disregarding the first container 15 as in Format E.

Format G

Step	Position
Sample introduction	1
First reagent introduction and mixing	2-3
First incubation (7 minutes)	4-23
Transfer portion of container 15 contents to second container 15, remainder of container 15 continues on process lane 28	24
First container 15 contents continues first incubation (11 minutes)	24-63
Second reagent introduction and mixing with second container 15 contents	2-3
First container 15 separation and wash	64-67
Second container 15 first incubation (18 minutes)	4-63
Introduction of fourth reagent into first container 15 and mixing (optional, enhances total Hb chemiluminescent signal)	71-72
Second container separation and wash	64-67
Fourth incubation (4 minutes-optional) of first container 15	73-86
Third reagent introduction into second container 15 and mixing	71-72
First container 15 separation and wash	87-90
Third incubation (4 minutes) of second container 15	73-86
Pretrigger introduction into first container 15 and mixing	94
Separation and wash of second container 15	87-90



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-continued

<u>Format G</u>	
Step	Position
Trigger and read value 1 (Total Hb) from first container 15	98
Pretrigger introduction into second container 15 and mixing	94
Trigger and read value 2 (GlyHb)	98

$$\text{Reported result} = \frac{\text{value 2}}{\text{value 1}} \times 100$$

As an example, this format may also be modified as may be done with Format F. With that modification, this Format may be used to determine progesterone, testosterone and estradiol.

<u>Format H</u>	
Step	Position
Sample introduction	1
First reagent introduction and mixing	2-3
First incubation	4-86
Separation and wash	87-90
Pretrigger introduction and mixing	94
Second incubation	95-97
Trigger and read	98

As an example, this format may be used to determine, among other things, beta human chorionic gonadotropin (B-hCG), prolactin, progesterone, testosterone, estradiol and ferritin. It is to be noted that almost any item of interest discussed herein may be determined by properly using this format.

<u>Format I</u>	
Step	Position
Sample introduction into first container 15, possibly with diluent fluid	1
First reagent introduction into first container 15, portion of first container 15 contents moved into pipettor, remainder of container continues on process lane 28, bypassing all wash stations, to Position 71	2
First reagent introduction into second container 15 and mixing	2-3
First incubation of second container 15	4-23
Second container 15 first incubation (18 minutes)	24-63
Introduction of second reagent into first container 15 and mixing (optional, enhances total Hb chemiluminescent signal)	71-72

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-continued

<u>Format I</u>	
Step	Position
Second container separation and wash	64-67
Fourth incubation (4 minutes - optional) of first container 15	73-86
Third reagent introduction into second container 15 and mixing	71-72
First container 15 passes through bypass region 58C	87-90
Third incubation (4 minutes) of second container 15	73-86
Pretrigger introduction into first container 15 and mixing	94
Separation and wash of second container 15	87-90
Trigger and read value 1 (Total Hb) from first container 15	98
Pretrigger introduction into second container 15 and mixing	94
Trigger and read value 2 (GlyHb)	98

$$\text{Reported result} = \frac{\text{value 2}}{\text{value 1}} \times 100$$

As an example, in Format I, it is possible to modify the format by disregarding the first container 15 after the portion of the container 15 contents has been transferred (Position 24) to the second container 15. In that case, Format I may be used to determine, for example, folate and vitamin B12.

<u>Format J</u>	
Step	Position
Sample introduction into container 15, possibly with diluent fluid	1
First reagent introduction and mixing	2-3
First incubation (27 minutes)	4-93
Pretrigger introduction and mixing	94
Second incubation (1 minute)	95-97
Trigger and read	98

As an example, Format J may be used to determine, among other things, total hemoglobin.

The embodiments described herein also allow for sample pretreatment which may be performed in at least two ways, indicated as Formats K and L. During performance of sample pretreatment, fluid present in the containers 15 indicated may be processed, after they are no longer significant in the pretreatment steps, in any appropriate manner, such as any of the Formats discussed above. Also, as will become clear later on, both Formats K and L are substantially similarly applicable to the other embodiments of the process path 10 discussed below.



<u>Format K</u>	
Step	Position
Sample introduction into first container 15, possibly with diluent fluid	1
First reagent introduction and mixing	2-3
First incubation (7 minutes)	4-23
Transfer portion of contents of first container 15 to second container 15 in Position 1	24
Second reagent introduction to second container 15 and mixing (optional)	2-3
Transfer portion of contents of second container 15 to third container 15 in Position 1	24
Third reagent introduction to third container and mixing (optional)	2-3

As an example, the third container 15 may be processed according to at least one of Formats A (to determine, among other things, folate), B, C, H and J.

<u>Format L</u>	
Step	Position
Sample introduction into first container 15, possibly with diluent fluid	1
First reagent introduction and mixing	2-3
First incubation (7 minutes)	4-23
Transfer portion of contents of first container 15 to second container 15 in Position 1	24
Second reagent introduction to second container 15 and mixing (optional)	2-3

As an example, the second container 15 may be processed according to at least one of Formats A (to determine, among other things, folate, vitamin B12, confirm HBsAg), B, C, H and J.

In each of the formats discussed above, it is possible to move contents of a first container 15 at Positions 2 or 24 into a second container 15 at Position 1. Thereafter, the first container 15 may or may not be ignored.

It is to be remembered, as pointed out earlier, that the steps of one format may be mixed with steps of another format to arrive at yet further formats. Also, it is to be remembered that the construction of the process path 10, and its elements and supporting components, allow for selective automated performance (i.e. a particular step may or may not be performed, as desired) of the above-described steps.

These examples demonstrate usefulness of the described embodiments in controlled processing of determinations of items of interest in a sample within a common process path 10.

As discussed earlier, multiple process paths 10 may be connected to meet specific needs. If the process path 10 were to perform approximately 200 determinations per hour, and if an analyzer (FIG. 29) that performed 400 determinations per hour were needed, then two process paths 10 could be connected. One way of doing is described with reference to FIGS. 17, 29 and 30.

As FIG. 17 illustrates, the process path 10 would be located in space 140. To supply samples to the process path 10, a load track 142 and a conveyor 146 are provided connected to a frame 148 defining the space 140. In some embodiments, at least one of the load track 142, the conveyor 146 and an unload track 192 may be provided with a cover 194 (FIG. 30). A carrier 150 supporting multiple sample tubes 152, which may be any suitable tubes, rides along both the load track 142 and the conveyor 146. Both the load track 142 and the conveyor 146 move the carrier 150 as indicated by the arrows. A transfer mechanism 154, such as a solenoid (e.g. linear actuation) driven arm and the like, shifts the carrier 150 from the load track 142 to the conveyor 146.

The carrier 150 moves along the conveyor 146 until the carrier 150 is stopped by a retention member 156, which is, in the illustrated embodiment, a stepper motor driving a star wheel which mates with the carrier 150. The pipetting system 128 accesses sample at the position 130B and supplies that sample to a container 15 on the process path 10. Of course, suitable identification structures, such as bar codes on the sample tubes 152 and a bar code reader, can be provided. When the pipetting system 128, or any of the pipetting systems 128, 132 or 134 access a fluid, pipetting system pressure can be monitored as described in commonly owned U.S. patent application, Ser. No. 08/572,835 filed on Dec. 14, 1995. The disclosure of that application is incorporated herein in its entirety. Appropriate liquid level sense devices, such as radio frequency based devices and the like, may also be located in suitable positions.

In an exemplary embodiment, the load track 142 may be No. 77325-101 and the conveyor 146 may be No. 77425-101 both available from Dorner Manufacturing of Hartland, Wis. The unload track 192 may be No. 77525-101 available from SPM/Portland of Hillsboro, Oreg. The retention member 156 may be No. 77476-101 available from Pacific Scientific of Elgin, Ill. The transfer mechanism 154 may comprise a solenoid such as No. 77952 available from Lucas/Ledex of Vandalia, Ohio, a belt such as No. 6R25-M225090 and a pulley such as No. A 6725-020DF0908 both available from Stock Drive Parts of New Hyde Park, N.Y., and a stepper motor such as No. P21NSXS-LSS-NS07 available from Pacific Scientific of Elgin, Ill.

In some cases, a level of sample in a sample tube 152 may be insufficient for access by the pipetting system 128. In these cases, the sample within the sample tube 152 may be moved by an operator into another container 208 shown in FIGS. 31A, 31B and 31C. The container 208 comprises a barrel 210 and a flange 212. The barrel 210 is dimensioned to fit within the sample tube 152 as shown in FIG. 31C. The flange 212 is offset from an outer diameter surface of the barrel 210 by a distance sufficient to accommodate any suitable sample tubes 152. In this way, sample can be moved from the sample tube 152 into the container 208 and the container 208 can be placed within the sample tube 152. The sample tube 152 bearing the container 208 may then be



placed into the carrier 150. Because the sample is now in the container 208, the level of the sample is elevated with respect to the level of the sample in the sample tube 152, thereby facilitating sample access by the pipetting system 128.

In an exemplary embodiment, the container 208 may be made from Dow 666 polystyrene and is dimensioned to fit within sample tubes having outer diameters substantially within the range of about 0.4 inches through about 0.7 inches. The barrel 210 has an outer diameter measuring about 0.4 inches and a length of about 1.964 inches. The flange 212 has an outer diameter measuring about 0.776 inches, depends from an open end of the container 208 by a distance of about 0.216 inches and is offset from the outer diameter surface of the barrel 210 by a distance of about 0.258 inches.

In some embodiments, the load track 142 is removed and replaced by a sample supply conveyor having a similar retention member 156. If this were done, then the pipetting system 128 would access sample at position 130A. If this case, then, in additional embodiments, a carousel 189 may be operatively connected with the frame 148 by a connection member 193 as shown in FIGS. 26 and 27. The connection member 193 locates the carousel 189 with respect to the process path 10 such that the pipetting system 128 can also access containers on the carousel 189 at position 130B. The carousel 189 may be used, for instance, to house determination calibrators and controls and certain samples, such as emergency samples that need to be processed immediately. In an exemplary embodiment, the carousel 189 may be a 2 or 3 part injection molded polymeric (ABS, GE-Cyclocac or the like) article constructed substantially similar to a TDx@ unit dose carousel, an IMx Select@ carousel (Abbott Laboratories, Abbott Park, Ill.) and the like.

In some instances, the retention member 156 may not retain the carrier 150 for sample access, but may allow the carrier 150 to move toward an end 158 of the conveyor 146 towards another process path 10. In this case, the frame 148 includes a connecting structure 160 for operatively coupling one process path 10 to another, or more specifically, one frame 148 holding one process path 10 to another frame 148 holding another process path 10. In an exemplary embodiment, the connecting structure 160 may be constructed such that two adjacent frames 148 are offset by a distance substantially within the range of about 0.25 inches to about 1.5 inches.

The connecting structure 160 comprises a first bracket 162 and a second bracket 164. The first bracket 162 is connected with one frame 148 and the second bracket 164 is connected with the another frame 148. To connect the frames 148, a fastener, such as a bolt and the like, is placed between aligned apertures 166 in the first and second brackets 162 and 164. Another fastener is inserted into slots 168 disposed on opposite ends of the brackets 162 and 164. The conveyors 146 supported by both frames 148 have sufficient tolerance such that more precise alignment of the frames 148 is not required. As a carrier 150 leaves an end 158 of one conveyor 146, the carrier 150 is supported by an opposing end 196 of an the adjacent conveyor 146. Once the carrier 150 reaches the end 158 of the last conveyor 146, the carrier 150 is moved to an unload track 192 (FIG. 29), constructed and operated substantially similarly to the load track 142, by another transfer mechanism 197, which may be substantially similar to the transfer mechanism 154.

The construction of the process path 10 is also adaptable in other ways to meet other requirements. For example, it may be desirable to provide a process path 10 that performs

100, 50 or any desired number of determination per hour. Viewing this requirement in another way, it may be desirable to provide a process path 10 that fits within a certain physical space, such as a table surface. To meet such requirements, the process path 10 may be scaled, i.e. altered in size or determinations per hour while still including elements discussed above, such as a bypass region, a mixing device, a pipetting system, a wash station and a reader.

Another embodiment is a process path 10~, constructed to perform 100 determinations per hour, and is illustrated in FIGS. 20A and 20B. This embodiment utilizes elements substantially similar to those described above, hence the like reference characters. The same index period and assay formats are used, thereby allowing the same reagents to be used. Because of the reduced number of determinations per hour, it is possible to reduce correspondingly the physical dimensions of the embodiment. For instance, whereas the process path 10 of the previous Figures comprises 112 positions, the process path 10~ comprises about 55 positions. In another embodiment, which performs 50 determinations per hours the corresponding process path comprises approximately 32 positions.

Whereas the determinations performed with the process path 10 are completed without a container 15 passing the same location along the process lane 28 more than once, the containers 15 used with the process path 10~ may pass the same location along the process lane 28 more than once. Depending upon the particular needs to be addressed, the process path may be modified such that a container 15 passes the same location along the process path any appropriate number of times. Of course, depending upon the particular employment, a given container 15 may be positioned in a different one of a performance lane 62 and an avoidance lane 64 of a given bypass region 58 at different times passing through the same bypass region 58 during a given determination.

Illustrating further by example, the following describes the procedures performed at each location along the process path 10~ which performs 100 determinations in an hour. As noted above, a particular container 15 may pass a given location along the process path 10~ more than once. Therefore, Process Position 1 indicates the first time the container 15 encounters Process Position 1, while Process Position 1' indicates the second time the container 15 encounters Process Position 1. Also, in similar fashion, Process Position 1" indicates the third time the container 15 encounters Process Position 1. Furthermore, the process path 10~ is constructed such that once a container 15 reaches Process Position 46 a first time, the next Process Position reached by the container 15 may be Process Position 1', i.e. the container 15 moves from one end of the process path 10~ to an opposite end of the process path 10~.

In the illustrated embodiment of the process path 10~, a second processing lane 170 is included. The second processing lane 170 may be located in any suitable position with respect to the processing lane 28 so that a container 15 can move between the process lane 28 and the process lane 170. In some embodiments, the position of the process lane 170 may be chosen to maintain the process path 10~ within specified physical dimensions.

A prime mover, which may be substantially similar to the prime movers discussed earlier, is located, in an exemplary embodiment, adjacent position 46 along the process lane 28. This prime mover is operable to move a container 15 from the process lane 28 to the process lane 170 when desired, viz. for reading a determination reaction, removal of a container 15 from the process path 10~, etc. The process



lane 28 may be joined to the process lane 170 by suitable connection structures 172, such as those associated with a bypass region. In this manner, a container 15 may be selectively automatically moved between the process lane 28 and the process lane 170. Thus, upon reaching Process Position 46, a container 15 may move to Process Position 1 of the process lane 28, or, alternatively, may move from Process Position 46 of the process lane 28 to Process Positions 47 through 55 of the process lane 170. Once in the process lane 170, process steps detailed in the example below are performed. Of course, structures, similar to those discussed above, that perform those process steps are disposed along the process lane 170 which has sufficient dimensions to accommodate those structures.

Process Position	Process Step	Description
1	Container 15 load	Container 15 moved from loading lane 30, if present, to process lane 28 as required
1	Sample Pipettor	Sample deposited into container 15 by pipetting system 128. The sample may be obtained from position 130A or 130B which are located on appropriate conveyors
2	Reagent Pipettor 1	Reagent obtained from reagent carousel 131 deposited into container 15 by pipetting system 132
3	Mixer	Contents of container 15 are mixed by a device 86 imparting motion to the container 15
4-16	Incubation	Contents of container 15 are incubated at a controlled temperature, about 37 degrees Celsius
17	Bypass region start	Container 15 is selectively positioned at entry to performance lane 62 or avoidance lane 64 of bypass region
18	Wash zone 1	Container 15 in performance lane 62 undergoes magnetic separation and fluid addition
19	Wash zone 1	Container 15 in performance lane 62 undergoes magnetic separation, container 15 contents aspiration and fluid addition
20	Wash zone W	Container 15 in performance lane 62 undergoes magnetic separation, container 15 contents aspiration and fluid addition
21	Wash zone 1	Container 15 in performance lane 62 undergoes magnetic separation, and container 15 contents aspiration
22	Bypass region end	Performance and avoidance lanes 62 and 64 of bypass region merge
23-24	Incubation	Contents of container 15 are incubated at a controlled temperature
24	Sample Pipettor	Sample may be aspirated from container 15 by pipetting system 128 for deposition into a second container 15 at position 1
25	Reagent Pipettor 2	Reagent obtained from reagent carousel 131 may be deposited into container by pipetting system 132
26	Mixer	Contents of container 15 are mixed by a device 36 imparting motion to the container 15
27-39	Incubation	Contents of container 15 are incubated at a controlled temperature
40	Bypass region start	Container 15 is selectively positioned at entry to

-continued

Process Position	Process Step	Description
5		performance lane 62 or avoidance lane 64 of bypass region
41	Wash zone 2	Container 15 in performance lane 62 undergoes magnetic separation and fluid addition
42	Wash zone 2	Container 15 in performance lane 62 undergoes magnetic separation, container 15 contents aspiration and fluid addition
43	Wash zone 2	Container 15 in performance lane 62 undergoes magnetic separation, container 15 contents aspiration and fluid addition
44	Wash zone 2	Container 15 in performance lane 62 undergoes magnetic separation and container 15 contents aspiration
45.5	Bypass region end	Performance lane 62 and avoidance lane 64 of bypass region merge (midway between positions 45 and 46)
46	Process lane transfer	Container moves from Process Position 46' of process lane 28 to Process Position 46 of process lane 170
25 46-47	Incubation	Contents of container 15 are incubated at a controlled temperature
48	Pre-Trigger and Mixer	Reagent added to container 15 and mechanically mixed
49-51	Incubation	Contents of container 15 are incubated at controlled temperature
52	Shutter, reader, and trigger	Indicator reaction (such as chemiluminescent reaction) triggered and read with magnetic particles pulled out of solution with magnet. Shutter blocks light.
54	Liquid Waste Aspirate	Magnetic particles are held at a wall of the container 15 and all liquid in container 15 is aspirated and discarded
55	Container 15 unload	Container 15 removed from process lane 28

Given these modifications, it is possible to utilize determination formats that are substantially similar to those discussed previously. For the sake of clarity, those formats, as performed by the process path 10-, are listed below.

## Format A

Step	Position
Sample introduction	1
First reagent introduction and mixing	2-3
First incubation (18 minutes)	4-17
Container 15 passes through bypass region, first incubation continues	18-21
First incubation continues	22-40
Container 15 passes through bypass region, first incubation continues	41-44
First incubation continues	45-17'
Separation and wash	18'-21'
Second reagent introduction and mixing	25'-26'
Second incubation (4 minutes)	27'-40'
Separation and wash	41'-44'
Container 15 transferred from process lane 28 to process lane	46' of 28 to 46 of



-continued

<u>Format A</u>	
Step	Position
170	170
Pretrigger introduction and mixing	48
Third incubation (1 minute)	49-51
Trigger and read	52
Container 15 evacuate	54
Container 15 removal	55

As an example, Format A may be used to determine at least the following items of interest: antibodies to HCV, antibodies to HIV 1/HIV 2, antibodies to hepatitis B core antigen (HBcAb), carcinoembryonic antigen (CEA), cancer antigen 19-9 (CA19-9), Hepatitis B Surface Antigen (HBsAg) antibodies to Hepatitis B Surface antigen (HBsAb), alpha-fetoprotein (AFP), Total prostate specific antigen (Total PSA), Free PSA, Thyroid stimulating Hormone (TSH), luteinizing hormone (LH), follicle stimulating hormone (FSH), beta human chorionic gonadotropin (B-hCG), Free Thyroxine (Free T4), Free triiodothyronine (Free T3), Total T4, Total T3, Prolactin and Ferritin. It is to be noted that almost any item of interest discussed herein may be determined by properly using this format. For instance, this format may also be used to determine beta human chorionic gonadotropin (B-hCG), prolactin and ferritin.

Format B

Step	Position
Sample introduction	1
First reagent introduction and mixing	2-3
First incubation (11 minutes)	4-40
Separation and wash	41-44
Second reagent introduction and mixing	2'-3'
Second incubation (11 minutes)	4'-40'
Separation and wash	41'-44'
Container 15 transferred from process lane 28 to process lane 170	46' of 28 to 46 of 170
Pretrigger introduction and mixing	48
Third incubation (1 minute)	49-51
Trigger and read	52
Container 15 evacuate	54
Container 15 removal	55

As an example, Format B may be used to determine an item of interest in a sample where a relatively increased degree of sensitivity, as compared with some other formats, is desired. It is to be noted that almost any item of interest discussed herein may be determined by properly using this format.

Format C

Step	Position
Sample introduction	1
First reagent introduction and mixing	2-3
First incubation (11 minutes)	4-40
Separation and wash	41-44
Second reagent introduction and mixing	2'-3'
Second incubation (4 minutes)	4'-17'

-continued

<u>Format C</u>	
Step	Position
Separation and wash	18'-21'
Third reagent introduction and mixing	25'-26'
Third incubation (4 minutes)	27'-40'
Separation and wash	41'-44'
Container 15 transferred from process lane 28 to process lane 170	46' of 28 to 46 of 170
Pretrigger introduction and mixing	48
Fourth incubation (1 minute)	49-51
Trigger and read	52
Container 15 evacuate	54
Container 15 removal	55

As an example, Format C may be used when the item of interest relates to hepatitis, such as determinations for anti-M, HBcAb-M and HAVAb-M.

Format D

Step	Position
Sample introduction	1
First reagent introduction and mixing	2-3
First incubation (6 minutes)	4-17
Transfer to second container 15 in position 1	24
Second reagent introduction and mixing	2-3
Second incubation (11 minutes)	4-40
Separation and wash	41-44
Third reagent introduction and mixing	2'-3'
Third incubation (4 minutes)	4'-17'
Separation and wash	18'-21'
Fourth reagent introduction and Mixing	24'-25'
Fourth incubation (4 minutes)	27'-40'
Separation and wash	41'-44'
Container transferred from process lane 28 to process lane 170	46' of 28 to 46 of 170
Pretrigger introduction and mixing	48
Fifth incubation (1 minute)	49-51
Trigger and read	52
Container 15 evacuate	54
Container 15 removal	55

Format E

Sample introduction	1
First reagent introduction and mixing	2-3
First incubation (7 minutes)	4-23
Transfer portion of container 15 contents to second container 15, remainder of container 15 continues on process lane 28	24
First container 15 contents continues first incubation (11 minutes)	25-17'
Second reagent introduction and mixing with second container 15 contents	2-3
First container 15 passes through bypass region 58 B	18'-21'
Second container 15 first	4-17'

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-continued

Step	Position
incubation (18 minutes) Introduction of fourth reagent into first container 15 and mixing (optional, enhances total Hb chemiluminescent signal)	24'-25'
Second container separation and wash	18'-21'
Fourth incubation (4 minutes-optional) of first container 15	26-40'
Third reagent introduction into second container 15 and mixing	24'-25'
First container 15 passes through bypass region	41'-44'
Third incubation (4 minutes) of second container 15	26-40'
First container 15 transferred from process lane 28 to process lane 170	46' of 28 to 46 of 170
Pretrigger introduction into first container 15 and mixing	48
Separation and wash of second container 15	41'-44'
Second container 15 transferred from process lane 28 to process lane 170	46' of 28 to 46 of 170
Trigger and read value 1 (Total Hb) from first container 15	52
Pretrigger introduction into second container 15 and mixing	48
Trigger and read value 2 (GlyHb)	52

$$\text{Reported result} = \frac{\text{value 2}}{\text{value 1}} \times 100$$

For example, in Format E, it is possible to modify the format by disregarding the first container 15 after the portion of the container 15 contents has been transferred (Position 24) to the second container 15. In that case, Format E may be used to determine, for example, folate and vitamin B12.

Format F

Step	Position
Sample introduction	1
First reagent introduction and mixing	2-3
First incubation (6 minutes)	4-23
Transfer portion of container 15 contents to second container 15, remainder of container 15 continues on process lane 28	24
First container 15 contents continues first incubation (11 minutes)	25-17'
Second reagent introduction and mixing with second container 15 contents	2-3
First container 15 passes	18'-21'

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-continued

<u>Format F</u>	
Step	Position
through bypass region 58 B	
Second container 15 first incubation (11 minutes)	4-40
Introduction of fourth reagent into first container 15 and mixing (optional, enhances total Hb chemiluminescent signal)	24'-25'
Second container separation and wash	41-44
Fourth incubation (4 minutes-optional) of first container 15	26-40'
Third reagent introduction into second container 15 and mixing	2'-3'
First container 15 passes through bypass region	41'-44'
Third incubation (11 minutes) of second container 15	4'-40'
First container 15 transferred from process lane 28 to process lane 170	46' of 28 to 46 of 170
Pretrigger introduction into first container 15 and mixing	48
Separation and wash of second container 15	41'-44'
Trigger and read value 1 (Total Hb) from first container 15	52
Second container 15 transferred from process lane 28 to process lane 170	46' of 28 to 46 of 170
Pretrigger introduction into second container 15 and mixing	48
Trigger and read value 2 (GlyHb)	52

$$\text{Reported result} = \frac{\text{value 2}}{\text{value 1}} \times 100$$

This format may be used, for example, to determine at least one of total and glycated hemoglobin. Also, this format may be modified by disregarding the first container 15 as in Format E.

Format G

Step	Position
Sample introduction	1
First reagent introduction and mixing	2-3
First incubation (6 minutes)	4-23
Transfer portion of container 15 contents to second container 15, remainder of container 15 continues on process lane 28	25
First container 15 contents continues first incubation (12 minutes)	25-17'
Second reagent introduction and mixing with second container 15	2-3



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-continued

Format G	
Step	Position
contents	
First container 15 separation and wash	18'-21'
Second container 15 first incubation (18 minutes)	4-17'
Introduction of fourth reagent into first container 15 and mixing (optional, enhances total Hb chemiluminescent signal)	24"-25'
Second container separation and wash	18'-21'
Fourth incubation (4 minutes-optional) of first container 15	27'-40'
Third reagent introduction into second container 15 and mixing	24-26'
First container 15 separation and wash	41'-44'
Third incubation (4 minutes) of second container 15	26-40'
First container 15 transferred from process lane 28 to process lane 170	46' of 28 to 46 of 170
Pretrigger introduction into first container 15 and mixing	48
Separation and wash of second container 15	41'-44'
Second container 15 transferred from process lane 28 to process lane 170	46' of 28 to 46 of 170
Trigger and read value 1 (Total Hb) from first container 15	52
Pretrigger introduction into second container 15 and mixing	48
Trigger and read value 2 (GlyHb)	52
Reported result = $\frac{\text{value 2}}{\text{value 1}} \times 100$	

As an example, this format may also be modified as may be done with Format F. With that modification, this Format may be used to determine progesterone, testosterone and estradiol.

Format H	
Step	Position
Sample introduction	1
First reagent introduction and mixing	2-3
First incubation	4-41'
Separation and wash	42'-44'
Container 15 transfer from process lane 28 to process lane 170	46' of 28 to 46 of 170
Pretrigger introduction and mixing	48
Second incubation	49-51
Trigger and read	52

As an example, this format may be used to determine, among other things, beta human chorionic gonadotropin

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(B-hCG), prolactin, progesterone, testosterone, estradiol and ferritin. It is to be noted that almost any item of interest discussed herein may be determined by properly using this format.

Format I	
Step	Position
Sample introduction into first container 15, possibly with diluent fluid	1
First reagent introduction into first container 15, portion of first container 15 contents moved into pipettor, remainder of container continues on process lane 28, bypassing all wash stations, to Position 25'	2
First reagent introduction into second container 15 and mixing	2-3
Second container 15 first incubation (18 minutes)	4-17'
Introduction of second reagent into first container 15 and mixing (optional, enhances total Hb chemiluminescent signal)	25'-26'
Second container separation and wash	18'-21'
Fourth incubation (4 minutes-optional) of first container 15	27'-40'
Third reagent introduction into second container 15 and mixing	25'-26'
First container 15 passes through bypass region 58	41'-44'
Third incubation (4 minutes) of second container 15	27'-40'
Pretrigger introduction into first container 15 and mixing	48
Separation and wash of second container 15	41'-44'
Trigger and read value 1 (Total Hb) from first container 15	52
Pretrigger introduction into second container 15 and mixing	48
Trigger and read value 2 (GlyHb)	52
Reported result = $\frac{\text{value 2}}{\text{value 1}} \times 100$	

55 As an example, in Format I, it is possible to modify the format by disregarding the first container 15 after the portion of the container 15 contents has been transferred (Position 24) to the second container 15. In that case, Format I may be used to determine, for example, folate and vitamin B12.

Format J	
Step	Position
Sample introduction into container 15, possibly	1

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-continued

Format J	
Step	Position
with diluent fluid	
First reagent introduction and mixing	2-3
First incubation (27 minutes-two times along process lane 28)	4-47
Pretrigger introduction and mixing	48
Second incubation (1 minute)	49-51
Trigger and read	52

As an example, Format J may be used to determine, among other things, total hemoglobin.

The embodiments described herein also allow for sample pretreatment which may be performed in at least two ways, indicated as Formats K and L. During performance of sample pretreatment, fluid present in the containers 15 indicated may be processed, after they are no longer significant in the pretreatment steps, in any appropriate manner, such as any of the Formats discussed above. Also, as will become clear later on, both Formats K and L are substantially similarly applicable to the other embodiment of the process path 10 discussed below.

Format K	
Step	Position
Sample introduction into first container 15, possibly with diluent fluid	1
First reagent introduction and mixing	2-3
First incubation (6 minutes)	4-23
Transfer portion of contents of first container 15 to second container 15 in Position 1	24
Second reagent introduction to second container 15 and mixing (optional)	2-3
Transfer portion of contents of second container 15 to third container 15 in Position 1	24
Third reagent introduction to third container and mixing (optional)	2-3

As an example, the third container 15 may be processed according to at least one of Formats A (to determine, among other things, folate), B, C, H and J.

Format L	
Step	Position
Sample introduction into first container 15, possibly with diluent fluid	1

-continued

Format L	
Step	Position
First reagent introduction and mixing	2-3
First incubation (7 minutes)	4-23
Transfer portion of contents of first container 15 to second container 15 in Position 1	24
Second reagent introduction to second container 15 and mixing (optional)	2-3

As an example, the second container 15 may be processed according to at least one of Formats A (to determine, among other things, folate, vitamin B12, confirm HBsAg), B, C, H and J.

Another embodiment is a process path 10\*, substantially similar to the previous embodiment, of the process path 10 is constructed to perform 50 determinations per hour. Elements similar to those described earlier, along with the same index period and assay formats are used, thereby allowing use of the same reagents albeit in an embodiment having relatively smaller physical dimensions. Following the examples discussed above, the following examples relate to this process path 10\*. In these examples, it is assumed that only one pipettor is utilized. Also, whereas the previous examples performed determinations while moving a container 15 along the process lane 28 twice, the process path 10\* performs determinations while moving a container 15 along the process lane 28 four times. Thus, the second time Process Position 1 is encounter is indicated as 1', the third time as 1" and the four time as 1". However, it is to be noted that more or less movements along the process lane 28 may be employed. Also, the process lane 28 of this embodiment includes 23 Process Positions with Process Position 23 being located adjacent to Process Position 1.

Process Position	Process Step	Description
1	Container 15 load	Container 15 moved from loading lane 30, if present, to process lane 28 as required
1	Pipettor	Sample deposited into container 15 by pipetting system
2	Pipettor	Reagent obtained from reagent carousel 131 deposited into container 15
3	Mixer	Contents of container 15 are mixed by a device 86 imparting motion to the container 15
4-16	Incubation	Contents of container 15 are incubated at a controlled temperature, about 37 degrees Celsius
17	Bypass region start	Container 15 is selectively positioned at entry to performance lane 62 or avoidance lane 64 of bypass region
18	Wash zone 1	Container 15 in performance lane 62 undergoes magnetic separation and fluid addition
19	Wash zone 1	Container 15 in performance lane 62 undergoes magnetic separation, container 15 contents aspiration and fluid addition



-continued

Process Position	Process Step	Description
20	Wash zone 1	Container 15 in performance lane 62 undergoes magnetic separation, container 15 contents aspiration and fluid addition
21	Wash zone 1	Container 15 in performance lane 62 undergoes magnetic separation, and container 15 contents aspiration
22	Bypass region end	Performance and avoidance lanes 62 and 64 of bypass region merge
23	Process lane transfer	Container 15 selectively transferred from process lane 23 to process lane 170
24	Pre-Trigger and Mixer	Reagent added to container 15 and mechanically mixed
26-28	Incubation	Contents of container 15 are incubated at controlled temperature
29	Shutter, reader, and trigger	Indicator reaction (such as chemiluminescent reaction) triggered and read with magnetic particles pulled out of solution with magnet. Shutter blocks light.
30-31	Liquid Waste Aspirate	Magnetic particles are held at a wall of the container 15 and all liquid in container 15 is aspirated and discarded
32	Container 15 unload	Container 15 removed from process lane 28

Given these modifications, it is possible to utilize determination formats that are substantially similar to those discussed previously. For the sake of clarity, those formats, as performed by a process path that performs 50 determinations per hour, are listed below.

Format A	
Step	Position
Sample introduction	1
First reagent introduction and mixing	2-3
First incubation (18 minutes)	4-17
Container 15 passes through bypass region, first incubation continues	18-21
First incubation continues	22-17"
Separation and wash	18"-21"
Second reagent introduction and mixing	2"-3"
Second incubation (4 minutes)	4"-17"
Separation and wash	18"-21"
Container 15 transferred from process lane 28 to process lane 170	23" of 28 to 23 of 170
Pretrigger introduction and mixing	24
Third incubation (1 minute)	25-28
Trigger and read	29
Container 15 evacuate	31
Container 15 removal	32

As an example, Format A may be used to determine at least the following items of interest: antibodies to HCV, antibodies to HIV1/HIV2, antibodies to hepatitis B core antigen (HBcAb), carcinoembryonic antigen (CEA), cancer antigen 19-9 (CA19-9), Hepatitis B Surface Antigen (HBsAg), antibodies to Hepatitis B Surface antigen (HBsAb), alpha-fetoprotein (AFP), Total prostate specific antigen (Total PSA), Free PSA, Thyroid stimulating Hormone (TSH), luteinizing hormone (LH), follicle stimulating hormone

(FSH), beta human chorionic gonadotropin (B-hCG), Free Thyroxine (Free T4), Free triiodothyronine (Free T3), Total T4, Total T3, Prolactin and Ferritin. It is to be noted that almost any item of interest discussed herein may be determined by properly using this format. For instance, this format may also be used to determine beta human chorionic gonadotropin (B-hCG), prolactin and ferritin.

Format B	
Step	Position
Sample introduction	1
First reagent introduction and mixing	2-3
First incubation (14 minutes)	4-17'
Separation and wash	18'-21'
Second reagent introduction and mixing	2"-3"
Second incubation (14 minutes)	4"-17"
Separation and wash	18"-21"
Container 15 transferred from process lane 28 to process lane 170	23" of 28 to 23 of 170
Pretrigger introduction and mixing	25
Third incubation (1 minute)	26-28
Trigger and read	29
Container 15 evacuate	31
Container 15 removal	32

As an example, Format B may be used to determine an item of interest in a sample where a relatively increased degree of sensitivity, as compared with some other formats, is desired. It is to be noted that almost any item of interest discussed herein may be determined by properly using this format.

Format C	
Step	Position
Sample introduction	1
First reagent introduction and mixing	2-3
First incubation (11 minutes)	4-17'
Separation and wash	18'-21'
Second reagent introduction and mixing	2"-3"
Second incubation (4 minutes)	4"-17"
Separation and wash	18"-21"
Third reagent introduction and mixing	2"-3"
Third incubation (4 minutes)	4"-17"
Separation and wash	18"-21"
Container 15 transferred from process lane 28 to process lane 170	23" of 28 to 23 of 170
Pretrigger introduction and mixing	25
Fourth incubation (1 minute)	26-28
Trigger and read	29
Container 15 evacuate	31
Container 15 removal	32

As an example, Format C may be used when the item of interest relates to hepatitis, such as determinations for anti-M, HBcAb-M and HAVAb-M.

Format D	
Step	Position
Sample introduction	1
First reagent introduction and mixing	2-3

-continued

Step	Position
First incubation (4 minutes)	4-17
Transfer to second container 15 in position 1	24
Second reagent introduction and mixing	2-3
Second incubation (14 minutes)	4-17'
Separation and wash	18'-21'
Third reagent introduction and mixing	2"-3"
Third incubation (4 minutes)	4"-17"
Separation and wash	18"-21"
Fourth reagent introduction and mixing	2"-3"
Fourth incubation (4 minutes)	4"-17"
Separation and wash	18"-21"
Container transferred from process lane 28 to process lane 170	23" of 28 to 23 of 170
Pretrigger introduction and mixing	25
Fifth incubation (1 minute)	26-28
Trigger and read	29
Container 15 evacuate	31
Container 15 removal	32
<u>Format E</u>	
Sample introduction	1
First reagent introduction and mixing	2-3
First incubation (7 minutes)	4-17
Transfer portion of container 15 contents to second container 15, remainder of container 15 continues on process lane 28	24
First container 15 contents continues first incubation (11 minutes)	24-17'
Second reagent introduction and mixing with second container 15 contents	2-3
First container 15 passes through bypass region 58B	18"-21"
Second container 15 first incubation (18 minutes)	4-17"
Introduction of fourth reagent into first container 15 and mixing (optional, enhances total Hb chemiluminescent signal)	2"-3"
Second container separation and wash	18'-21'
Fourth incubation (4 minutes - optional) of first container 15	4"-17"
Third reagent introduction into second container 15 and mixing	24'-25'
First container 15 passes through bypass region	18"-21"
Third incubation (4 minutes) of second container 15	4"-17"
First container 15 transferred from process lane 28 to process lane 170	23" of 28 to 23 of 170
Pretrigger introduction into first container 15 and mixing	25
Separation and wash of second container 15	18"-21"

-continued

Step	Position
5 Second container 15 transferred from process lane 28 to process lane 170	23" of 28 to 23 of 170
Trigger and read value 1 (Total Hb) from first container 15	29
10 Pretrigger introduction into second container 15 and mixing	25
Trigger and read value 2 (GlyHb)	29
15 Reported result = $\frac{\text{value 2}}{\text{value 1}} \times 100$	
For example, in Format E, it is possible to modify the format by disregarding the first container 15 after the portion of the container 15 contents has been transferred (Position 24) to the second container 15. In that case, Format E may be used to determine, for example, folate and vitamin B12.	
<u>Format F</u>	
25	
Step	Position
Sample introduction	1
First reagent introduction and mixing	2-3
30 First incubation (7 minutes)	4-17
Transfer portion of container 15 contents to second container 15, remainder of container 15 continues on process lane 28	23
35 First container 15 contents continues first incubation (11 minutes)	23-17"
Second reagent introduction and mixing with second container 15 contents	2-3
40 First container 15 passes through bypass region 58B	18"-21"
Second container 15 first incubation (11 minutes)	4-17'
45 Introduction of fourth reagent into first container 15 and mixing (optional, enhances total Hb chemiluminescent signal)	2"-3"
50 Second container separation and wash	18'-21'
Fourth incubation (4 minutes - optional) of first container 15	4"-17"
55 Third reagent introduction into second container 15 and mixing	2"-3"
First container 15 passes through bypass region	18"-21"
Third incubation (11 minutes) of second container 15	4"-17"
60 First container 15 transferred from process lane 28 to process lane 170	23" of 28 to 23 of 170
Pretrigger introduction into first container 15 and mixing	25



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-continued

Format F	
Step	Position
Separation and wash of second container 15	18"-21"
Trigger and read value 1 (Total Hb) from first container 15	29
Second container 15 transferred from process lane 28 to process lane 170	23" of 28 to 23 of 170
Pretrigger introduction into second container 15 and mixing	25
Trigger and read value 2 (GlyHb)	29

Reported result =  $\frac{\text{value 2}}{\text{value 1}} \times 100$

This format may be used, for example, to determine at least one of total and glycated hemoglobin. Also, this format may be modified by disregarding the first container 15 as in Format E.

Format G	
Step	Position
Sample introduction	1
First reagent introduction and mixing	2-3
First incubation (6 minutes)	4-23
Transfer portion of container 15 contents to second container 15, remainder of container 15 continues on process lane 28	23
First container 15 contents continues first incubation (12 minutes)	24-17"
Second reagent introduction and mixing with second container 15 contents	2-3
First container 15 separation and wash	18"-21"
Second container 15 first incubation (18 minutes)	4-17"
Introduction of fourth reagent into first container 15 and mixing (optional, enhances total Hb chemiluminescent signal)	2"-3"
Second container separation and wash	18"-21"
Fourth incubation (4 minutes - optional) of first container 15	4"-17"
Third reagent introduction into second container 15 and mixing	2"-3"
First container 15 separation and wash	18"-21"
Third incubation (4 minutes) of second container 15	4"-17"
First container 15 transferred from process lane 28 to process lane 170	23" of 28 to 23 of 170
Pretrigger introduction	25

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-continued

Format G	
Step	Position
into first container 15 and mixing	
Separation and wash of second container 15	18"-21"
Second container 15 transferred from process lane 28 to process lane 170	23" of 28 to 23" of 170
Trigger and read value 1 (Total Hb) from first container 15	29
Pretrigger introduction into second container 15 and mixing	25
Trigger and read value 2 (GlyHb)	29

20 Reported result =  $\frac{\text{value 2}}{\text{value 1}} \times 100$

As an example, this format may also be modified as may be done with Format F. With that modification, this Format may be used to determine progesterone, testosterone and estradiol.

Format H	
Step	Position
Sample introduction	1
First reagent introduction and mixing	2-3
First incubation	4-17"
Separation and wash	18"-21"
Container 15 transfer from process lane 28 to process lane 170	23" of 28 to 23 of 170
Pretrigger introduction and mixing	25
Second incubation	26-28
Trigger and read	29

As an example, this format may be used to determine, among other things, beta human chorionic gonadotropin (B-hCG), prolactin, progesterone, testosterone, estradiol and ferritin. It is to be noted that almost any item of interest discussed herein may be determined by properly using this format.

Format I	
Step	Position
Sample introduction into first container 15, possibly with diluent fluid	1
First reagent introduction into first container 15, portion of first container 15 contents moved into pipettor, remainder of container continues on process lane 28, bypassing all wash stations, to Position 25'	2
First reagent introduction into second	2-3

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-continued

Format I

Step	Position
container 15 and mixing	
Second container 15 first incubation (18 minutes)	4-17"
Introduction of second reagent into first container 15 and mixing (optional, enhances total Hb chemiluminescent signal)	2"-3"
Second container separation and wash	18"-21"
Fourth incubation (4 minutes - optional) of first container 15	4"-17"
Third reagent introduction into second container 15 and mixing	2"-3"
First container 15 passes through bypass region 58	18"-21"
Third incubation (4 minutes) of second container 15	4"-17"
Pretrigger introduction into first container 15 and mixing	25
Separation and wash of second container 15	18"-21"
Trigger and read value 1 (Total Hb) from first container 15	29
Pretrigger introduction into second container 15 and mixing	25
Trigger and read value 2 (GlyHb)	29

$$\text{Reported result} = \frac{\text{value 2}}{\text{value 1}} 100$$

As an example, in Format I, it is possible to modify the format by disregarding the first container 15 after the portion of the container 15 contents has been transferred (Position 24) to the second container 15. In that case, Format I may be used to determine, for example, folate and vitamin B12.

Format J

Step	Position
Sample introduction into container 15, possibly with diluent fluid	1
First reagent introduction and mixing	2-3
First incubation (27 minutes - four times along process lane 28)	4-47"
Pretrigger introduction and mixing	25
Second incubation (1 minute)	26-28
Trigger and read	29

As an example, Format J may be used to determine, among other things, total hemoglobin.

The embodiments described herein also allow for sample pretreatment which may be performed in at least two ways, indicated as Formats K and L. During performance of sample pretreatment, fluid present in the containers 15 indicated may be processed, after they are no longer significant in the pretreatment steps, in any appropriate manner, such as any of the Formats discussed above. Also, as will

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become clear later on, both Formats K and L are substantially similarly applicable to the other embodiment of the process path 10 discussed below.

Format K

Step	Position
Sample introduction into first container 15, possibly with diluent fluid	1
First reagent introduction and mixing	2-3
First incubation (6 minutes)	4-23
Transfer portion of contents of first container 15 to second container 15 in Position 1	24
Second reagent introduction to second container 15 and mixing (optional)	2-3
Transfer portion of contents of second container 15 to third container 15 in Position 1	24
Third reagent introduction to third container and mixing (optional)	2-3

As an example, the third container 15 may be processed according to at least one of Formats A (to determine, among other things, folate), B, C, H and J.

Format L

Step	Position
Sample introduction into first container 15, possibly with diluent fluid	1
First reagent introduction and mixing	2-3
First incubation (7 minutes)	4-23
Transfer portion of contents of first container 15 to second container 15 in Position 1	24
Second reagent introduction to second container 15 and mixing (optional)	2-3

As an example, the second container 15 may be processed according to at least one of Formats A (to determine, among other things, folate, vitamin B12, confirm HBsAg), B, C, H and J.

Given commonality among the various embodiments of the process path discussed and exemplified above, it is to be appreciated that assay formats performed on each of the various embodiments are essentially the same. The time frames are identical. Reagents for a particular assay used on one of the embodiments may also be used on other embodiments.

Upon consideration of all of these examples and their common features, it is to be understood that the process path 10, or in other words the process lane 28, has a variable physical length. However, the effective length of the process path 10 is constant in all embodiments. This effective length represents the total distance traveled by the container 15



along the process path 10 during performance of a certain determination. The physical length, i.e. the physical dimensions of the process path 10, is variable, for instance, to make the process path 10 fit within a given space. The effective length of the process path 10 is maintained constant by moving the container 15 multiple times along the same process path 10 (4 times in the last set of examples). Maintenance of the effective length is achieved with appropriate combination of selective automatic performance of a given determination process step. In all instances, the effective length remains constant even though the physical length of a given process path 10 may be longer or shorter than other process paths 10.

It is to be noted that all of the above discussed embodiments of the process path 10 include and utilize certain common elements, such as reagents, a sample/reagent pipettor, a mixer, a wash zone and a reader. The structural elements are arranged along each embodiment of the process path 10 such that each embodiment is able to perform the same determinations in substantially the same manner by keeping the effective length of the process path 10 constant. Each of the embodiments of the process path executes determinations with approximately the same number, such as 98 in the above examples, "steps" of the container 15 along the process path 10 between sample introduction and reading. Determination of a given item of interest by one of the embodiments of the process path 10 takes substantially the same amount of time as a determination of the same item of interest by another embodiment of the process path 10. Thus, it is possible to construct a structure for performing item of interest determinations which conforms to desired physical dimensions, throughput requirements, etc., while using the common elements discussed herein by maintaining the effective length of the process path constant.

What is claimed is:

1. A method of constructing a process lane for determining an item of interest in a sample, the method comprising the steps of:

- (a) accepting a container containing said sample in a process lane having a first physical length where a plurality of process steps for determining an item of interest in said sample are selectively automatically performed on said sample;
- (b) selectively automatically performing said process steps on the sample in the container on said process lane having said first physical length;
- (c) selectively varying a physical length of said process lane to form a second physical length which differs from said first physical length;
- (d) accepting said container containing said sample in said process lane having said second physical length;
- (e) selectively automatically performing said process steps on said sample in said container on said process lane having said second physical length; and
- (f) maintaining an effective length of said process lane on both first and second physical lengths constant wherein said effective length representing a total traveled distance of said container on said process lane.

wherein said process steps performed on said sample on said first physical length and second physical length being substantially the same by maintaining said effective length of said process lane constant.

2. A method as defined in claim 1 further comprising the step of:

- (d) moving the container along the first physical length of the process lane at least once.

3. A method as defined in claim 1 further comprising the step of:

- (d) moving the container along the second physical length of the process lane at least twice.

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