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Hamajima et al.

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CYSTEINE PROTEASE DERIVED FROM [54] PARASITIC HELMINTHS

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Appl. No.: 451,409

[56]

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Related U.S. Application Data

Continuation of Ser. No. 246,917, May 20, 1994, abandoned, which is a continuation of Ser. No. 920,092, Jun. 24, 1992, abandoned.

Foreign Application Priority Data [30] Jul. 25, 1991 Japan 3-208546 [**JP**] Japan 4-057189 Feb. 12, 1992 [JP]

[51]	Int. Cl. ⁶	C12N 9/50
[52]	U.S. Cl	/219 ; 435/212
[58]	Field of Search 424	/94.63, 94.65;

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435/219; 514/2, 12

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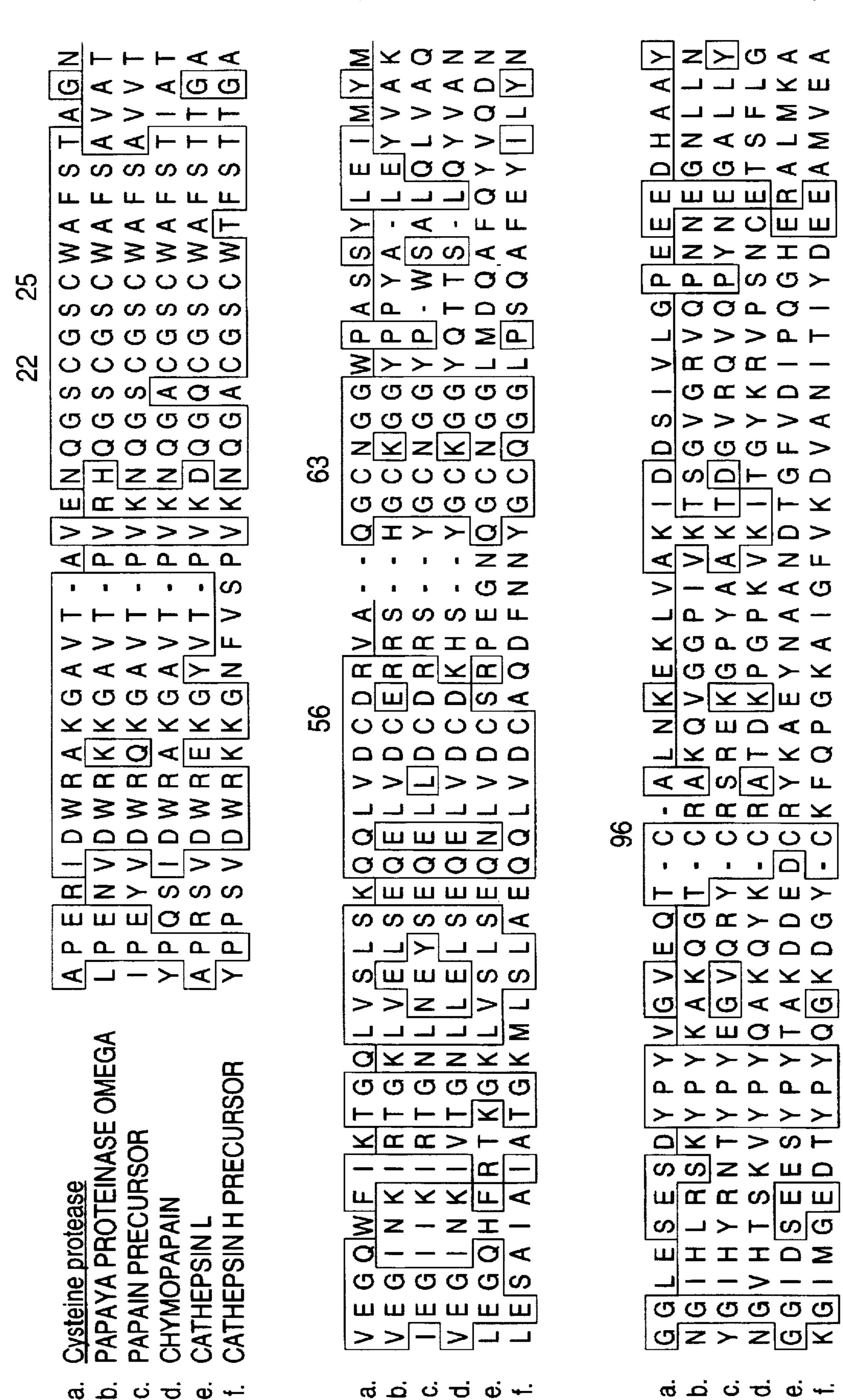
Primary Examiner—Vasu S. Jagannathan Assistant Examiner—Enrique Longton Attorney, Agent, or Firm—Limbach & Limbach L.L.P.

ABSTRACT [57]

The immunosuppressive drug of the present invention can suppress both delayed-type hypersensitivity and antibody production against specific antigens and graft tissues and induce immunological tolerance to them by several administrations, instead of long continuous administration.

Cysteine protease, a secretory protein accumulated in the tissue of parasitic helminths, is extracted and purified. The cysteine protease is administered to a mammal and then tissue is implanted to the mammal. The immune response of the mammal against the tissue implant is suppressed even one year later.

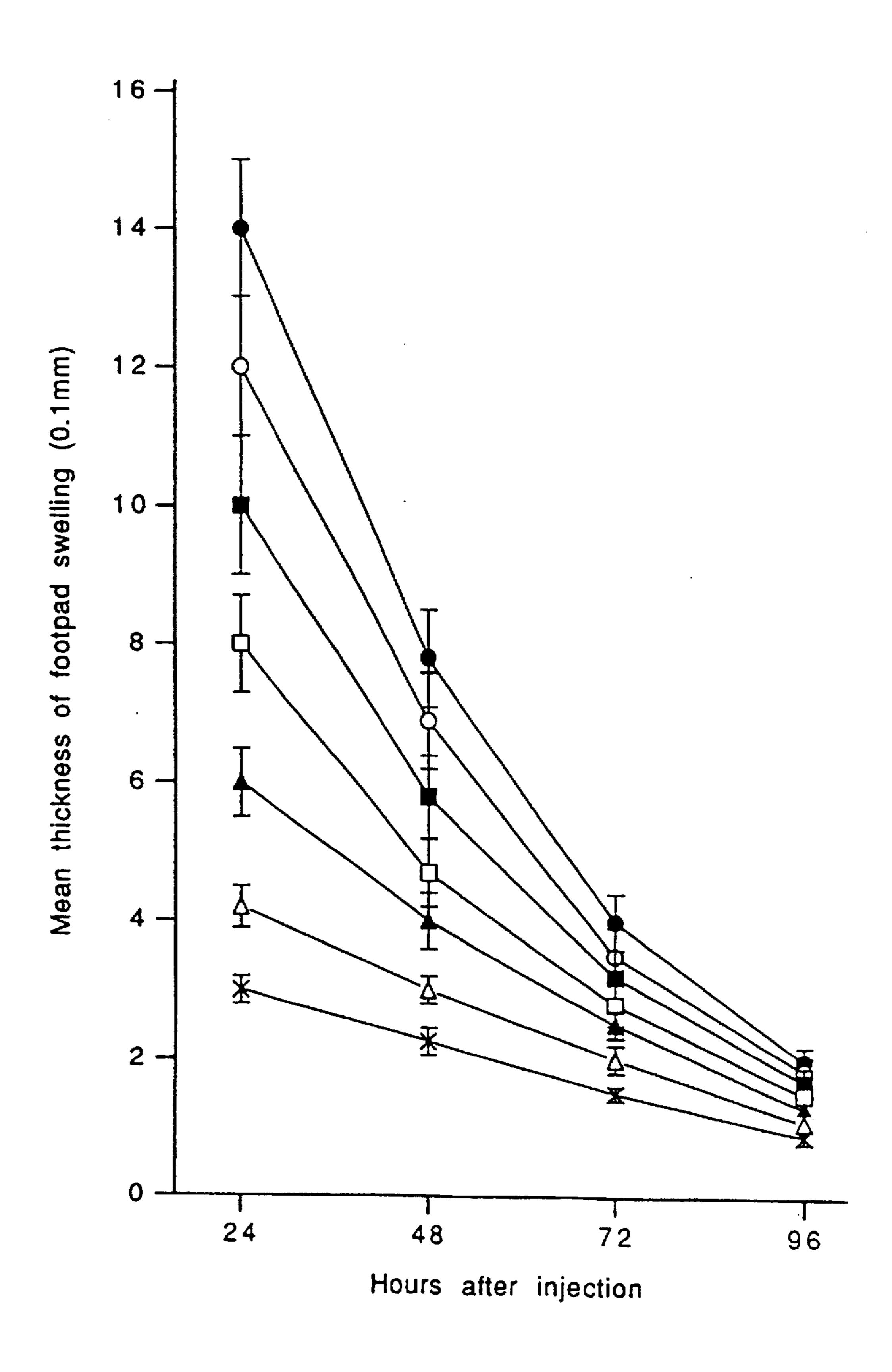
13 Claims, 6 Drawing Sheets

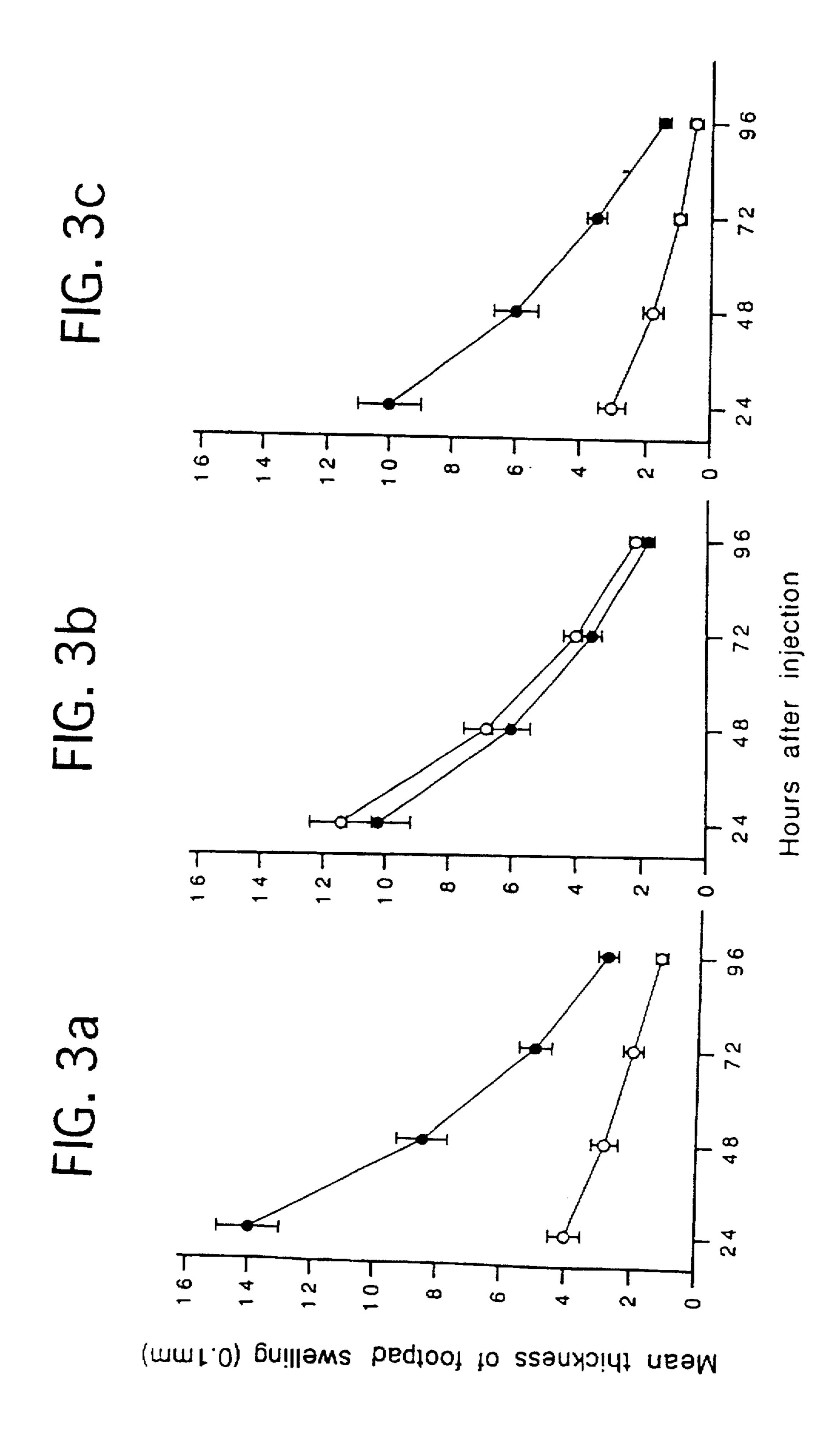


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FIG. 2





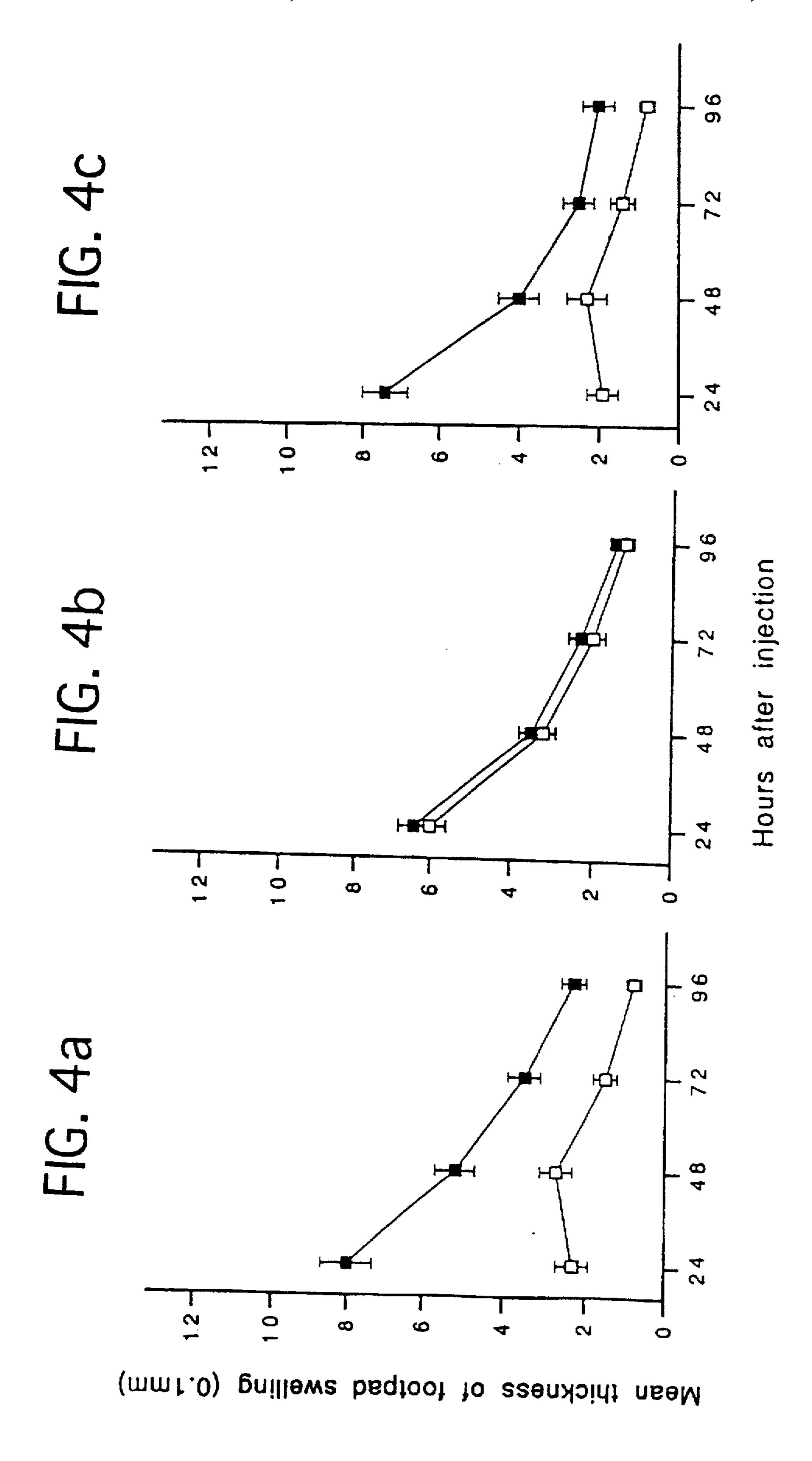
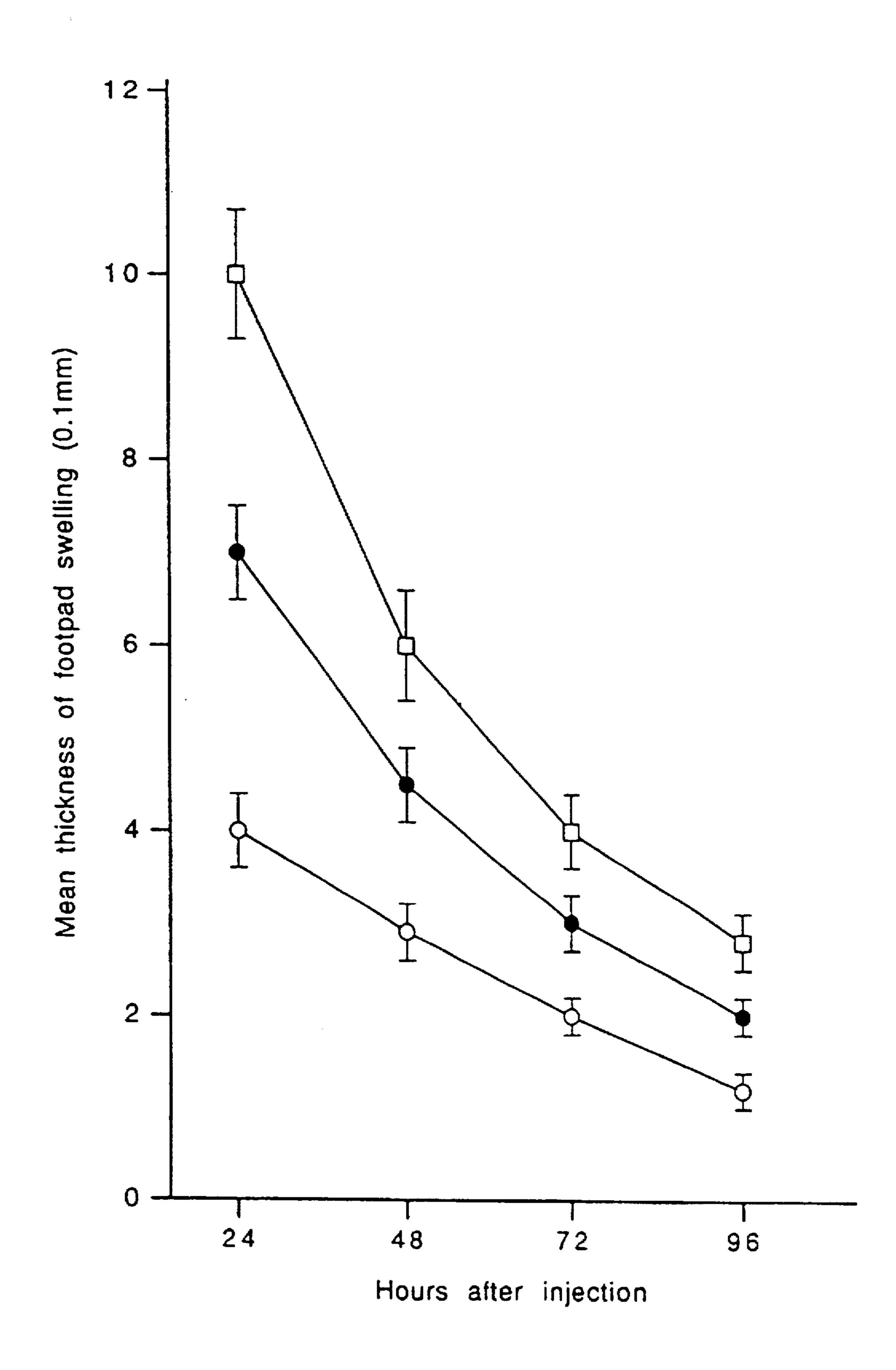


FIG. 5



CYSTEINE PROTEASE DERIVED FROM PARASITIC HELMINTHS

This is a continuation of application Ser. No. 08/246,917 filed on May 20, 1994 which is a File Wrapper Continuation 5 application of Ser. No. 07/920,092 filed on Jun. 24, 1992, both now abandoned.

FIELD OF THE INVENTION

The present invention relates to immunosuppressive drugs used for suppression of graft rejection in organ transplantation or suppression of autoimmune diseases.

BACKGROUND OF THE INVENTION

Currently, azathiopline, cyclosporin A, FK-506, and 15-deoxysperguarine are known as immunosuppressive drugs used for a therapeutic purpose, and have potent suppression effect.

However, the conventional immunosuppressive drugs do 20 not induce immunological tolerance and are therefore required that the drug be continuously administered for a long period of time. Side effects such as cancer, nephrotoxicity, angiitis, hepatotoxicity and disorders in the digestive tract have resulted from the continuous adminis- 25 tration of these doses.

The present inventors have studied agents of human parasites, especially parasitic helminths, which protect them from the immune response of human hosts, and have found that a cysteine protease, a secretory protein accumulated in the tissue of parasitic helminths, suppresses the cell-mediated and humoral immunities of mammalian hosts and induces the hosts to acquire immunological tolerance to the parasite.

SUMMARY OF THE INVENTION

The immunosuppressive drug of the present invention can suppress both delayed-type hypersensitivity and antibody production against specific antigens and graft tissues, and induce immunological tolerance by several administrations. Conventional immunosuppressive drugs are required that they be administered for a long period of time, and cause an adverse effect which is a drawback of the drugs. In contrast, it is not required that the immunosuppressive drug of the present invention it be administered for a long period of time. Therefore, the immunosuppressive drug of the invention overcomes the drawback of conventional immunosuppressive drugs.

DESCRIPTION OF THE FIGURES

FIGS. 1A and 1B show a comparison of the deduced amino acid sequences of a cysteine protease from *Paragonimus westermani* metacercariae and other similar cysteine proteases. Homologous areas are boxed and heterologous amino acids are underlined.

FIG. 2 is a graph showing a footpad response (swelling of footpad) of $C_{57}BL/6$ female mice. The female mice were intraperitoneally injected with sheep red blood cells (SRBC) at various times after intraperitoneal injection of the present cysteine protease. The intervals were 0(X), $1(\Delta)$, $2(\triangle)$, 3 and $4(\square)$, 5 and $6(\square)$, 7 and 8 days (\bigcirc) and untreated control (\bigcirc) . Bars represent standard errors.

FIGS. 3A, 3B and 3C are graphs showing a footpad response of C₅₇BL/6 female mice to sheep red blood cells 65 (SRBC) and adult *Paragonimus westermani* antigens. The mice were immunized with SRBC one day after an injection

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of the present cysteine protease. Group treated with the protease on day-1 (\bigcirc) and control group (\bullet). Bars represent standard errors. (a) The initial SRBC injection one day after the protease treatment. (b) Adult *P. westermani* antigen injection six months after the initial SRBC immunization. (c) SRBC injection one year after the initial SRBC immunization.

FIGS. 4A, 4B and 4C are graphs showing a footpad response of C₅₇BL/6 male mice to adult *Paragonimus* westermani antigens and sheep red blood cells (SRBC). The mice were immunized with adult *Paragonimus* westermani antigens one day after injection of the present cysteine protease. Group treated with the protease on day-1 (□) and control group (●). Bars represent standard errors. (a) The initial adult *P.* westermani antigen injection one day after the protease treatment. (b) SRBC injection six months after the initial adult *P.* westermani antigen immunization. (c) adult *P.* westermani antigen injection one year after the initial adult *P.* westermani antigen immunization.

FIG. 5 is a graph showing a footpad response of $C_{57}BL$ female mice to sheep red blood cells (SRBC). The mice were immunized with SRBC one day after injection of various cysteine proteases. Groups treated with plasmin, trypsin, papain, cathepsin B_1 and untreated control (\square), collagenase (\bullet), and P. westermani cysteine protease (\bigcirc). Bars represent standard errors.

DETAILED DESCRIPTION OF THE INVENTION

It is an object of the present invention to provide immunosuppressive drugs which, without long continuous administration, suppress both delayed-type hypersensitivity and antibody production against specific antigens and graft tissues and induce immunological tolerance to specific antigens and grafted tissues.

The present invention is characterized by the following description.

- (1). The present invention provides an immunosuppressive drug in which cysteine protease comprises its composition.
- (2). The present invention provides the immunosuppressive drug of (1) in which the cysteine protease contains a larger amount of acidic amino acids than basic amino acids.
- 45 (3). The present invention provides the immunosuppressive drug of (1) or (2) in which the cysteine protease is active in neutral hydrogen ion concentration.
 - (4). The present invention provides the immunosuppressive drug of (1), (2), or (3) in which the cysteine protease is extracted from the infected larvae of parasitic helminths.
 - (5). The present invention provides the immunosuppressive drug of (1), (2), (3), or (4) in which the cysteine protease comprises an amino acid sequence described in SEQ ID No. 1.
- 55 (6). The present invention provides a cysteine protease having an amino acid sequence described in SEQ ID No.
 - (7). The present invention provides a cysteine protease gene encoding a polypeptide having an amino acid sequence described in SEQ ID No. 1.
 - (8). The present invention provides a cysteine protease gene having a base sequence described in SEQ ID No. 2.

The term, "cysteine protease" of the present invention is intended to mean an endopeptidase comprising cysteine residues at amino acid positions, 22, 25, 56, 63, 96, 154, and 202, and a histidine residue at amino acid position 161 (the position is numbered from N terminus in the amino acid

sequence) and having an active center at cysteine and histidine residues.

The immunosuppressive drug of the invention comprises cysteine protease directly obtained by extracting the cell and tissue of organisms including invertebrates such as parasitic helminths to vertebrates such as various animals including mammal, or by cultured cells isolated from organisms described above, or by genetic engineering using recombinant DNA containing a cysteine protease gene (recombinant DNA technique), or by chemical synthesis.

When the cysteine protease of the present invention (hereinafter referred to as "present cysteine protease") is injected within a few days and/or several days before an injection of an antigen, the injection of the present cysteine protease suppresses cell-mediated immune response and/or humoral immune response, which is antibody production, against the antigen, for a long period of time. In addition, such a prior injection of the present cysteine protease allow hosts to take a graft for a long period of time and induce the hosts to acquire immunological tolerance to the graft.

Although the detailed mechanism of how cysteine protease works on the immune system is not clearly understood, it is believed that the immunosuppressive drug of the invention administered to a mammalian host is involved with the antigen presentation of immunocytes, receptors or suppressor T cells, thereby suppressing both delayed-type 25 hypersensitivity and antibody production and inducing the immunological tolerance of the mammalian host.

The present invention will be further described in Examples, which are not intended to limit the scope of the invention.

EXAMPLE

1. Extraction and Purification of Cysteine Protease from Parasitic Helminth

The cysteine protease of the invention was obtained by extracting *Paragonimus westermani* metacercaria with 50 35 mM imidazole-HCl/pH7.0, subjecting the extract to centrifugation (at 105,000 g, for 60 minutes), loading the supernatant onto Arginine-Sepharose CL-4B affinity chromatography, filtering the cluate on an Ultrogel AcA-54 gel and purifying the filtrate by DEAE-toyopearl S column 40 chromatography. N-terminal 25 amino acids of the purified native cysteine protease is shown in SEQ ID No. 5.

2. Amino Acid and Base Sequence of Present Cysteine Protease

The purified present cysteine protease was emulsified 45 with Freund's complete adjuvant. The emulsion was intravenously injected in the ears of rabbits for immunization, and antiserum against the present cysteine protease was obtained.

About 2×10^4 Paragonimus westermani metacercariae 50 were used to obtain mRNA. Total RNA (about 100 µg) was extracted from the pooled material by phenol extraction and poly A⁺RNA (about 2 µg) was recovered by passing the total RNA through an oligo (dT)-cellulose column. The mRNA thus obtained was used to construct a cDNA library using the 55 λgt11 vector. The cDNA library was screened by the antiserum and 25 independent positive clones containing about 300-600 bp long insert were obtained. Of 25 clones, two clones having the longest cDNA sequence were designated as λPW6 and λPW24. The two clones were subcloned using 60 pUC118, and the two subclones were named pPW6 and pPW24, respectively. Then, the base sequences of the two clones were determined by the dideoxy nucleotide sequencing method using a T7 sequencing kit (United States Biochemicals, U.S.A.).

E. coli MV1184 was transformed with the subclone pPW6 and the transformant, E. coli MV1184/pPW6, was deposited

with Fermentation Research Institute, Agency of Industrial Science and Technology and was assigned the accession number FERM BP-3937.

The base sequence and deduced amino acid sequence of pPW6 and pPW24 are shown in SEQ ID Nos. 3 and 4, respectively. Primers were synthesized based on the sequence near the ends of the cDNA insert of the clone and were used to screen the cDNA library again by the PCR.

A base sequence including the entire coding region of the gene of one mature cysteine protease of *Paragonimus westermani* metacercaria was determined (SEQ ID No. 2). The mRNA sequence contains an open reading frame with a significant length. The deduced amino acid sequence is shown in SEQ ID No. 1. The base sequence of the other several cDNA clones strongly suggested that each protease has a few different amino acids and belongs to a closely related protein family. The mature cysteine protease of the invention was found to contain 215 amino acids.

3. Characterization of Amino Acid Sequence of Present Cysteine Protease

The comparison of the primary structures of the present cysteine protease and the cysteine protease of other species (hereinafter referred to as "similar cysteine proteases") has revealed the following features: an amino acid sequence (boxed region) in FIG. 1 is homologous in both the present cysteine protease and similar cysteine proteases; cysteine residues are found at amino acid positions, 22, 25, 56, 63, 96, 154, and 202, and a histidine residue is found at amino acid position 161, the position that is numbered from N terminus in the amino acid sequence. This primary structure suggests that the present cysteine protease has a typical feature of cysteine protease. However, the underlined amino acid sequence of the present cysteine protease is different from those of all the other similar cysteine proteases.

In addition, the base sequences of other clones have revealed some substitutions of amino acid residues: in the present cysteine protease, there are substitutions. Ala \rightarrow Pro (amino acid position 15), Ser \rightarrow Glu (21), Argo \rightarrow Met (58), Val \rightarrow Ala (59), Gln \rightarrow Glu (61), Ala \rightarrow Ser (69), and Tyr \rightarrow Asp (77), the position that is numbered from N terminus in the amino acid sequence. This amino acid substitution strongly suggests that the primary structure of the present cysteine protease is at least 90% homologous to the other cysteine protease and that the present cysteine protease has a few different amino acids from other cysteine proteases to form its own family. These findings also coincide with variability found in the serine proteases of nematodes and are believed to be the common feature of parasitic helminths.

Furthermore, there is a difference in the number of amino acid residues between cysteine at amino acid position 56 and the following cysteine: 6 amino acid residues are found in the present cysteine protease, papain, chymopapain and papaya proteinase while 8 amino acid residues are found in cathepsin L and H. Based on the known three-dementional structure of papain, it is believed that the present cysteine protease forms a disulfide bridge at amino acid positions 22-63, 56-96 and 154-202, and its active site residue is cysteine at amino acid position 25 and histidine at 161.

The present cysteine protease is composed of more acidic amino acids(14.89-14.96 mole % acidic amino acid such as glutamic acid and aspartic acid) than basic amino acid (10.17-10.23 mole % basic amino acid such as lysine and arginine). The amino acid composition is similar to that of cathepsin L, but different from those of papain, chymopapain and papaya proteinase, which contains more basic amino acids (13.66-14.29 mole %) than acidic amino acids (7.08-8.37 mole %) (see Table 1). In order that a cysteine

protease works well in the neutral or faint acidic environment of the organism, it appears reasonable that the cysteine protease have a high acidic amino acid content. The high acidic amino acid content of the present cysteine protease is believed to be the reason that the present cysteine protease 5 alone has an immunosuppressive effect while other similar cysteine proteases such as papain and the like do not.

The present cysteine protease contains leucine in a large amount (8.10-8.14 mole %) and asparagine and tyrosine in a small amount -3.86 and 5.95-6.58 mole %, respectively), 10 a prominent feature that distinguishes the present cysteine protease from other similar cysteine proteases (Table 1).

TABLE 1

Amino acid composition of a cysteine protease and

other homologous cysteine proteases

									
									
Amino acid	CP*	PPA*	CPPA*	PP*	CL*	СН*			
Glutamic acid	9.09	5.93	3.73	5.44	9.30	5.23			
Leucine	8.10	5.29	5.71	4.85	5.17	5.13			
Valine	5.96	7.72	6.80	9.09	6.63	6.25			
Glycine	6.00	7.70	7.35	8.04	6.63	6.14			
Aspartic acid	5.80	2.44	3.35	2.46	6.12	4.26			
Alanine	5.83	4.57	4.52	4.61	5.04	6.02			
Lysine	6.37	5.36	11.13	7.56	6.72	7.27			
Glutamine	5.31	5.35	5.83	6.48	4.13	5.19			
Threonine	4.76	3.05	6.05	3.96	3.37	4.23			
Tryptophan	5.19	3.74	2.96	3.02	3.61	3.63			
Asparagine	3.36	5.81	5.27	6.83	5.14	5.64			
Serine	5.35	5.01	6.48	6.99	7.06	4.86			
Cysteine	3.52	3.11	3.51	2.69	3.00	3.45			
Methionine	2.17	0.00	0.54	0.00	3.16	3.71			
Tyrosine	6.58	12.61	9.86	8.70	8.33	9.65			

TABLE 1-continued

Amino acid composition of a cysteine protease and

other homologous cysteine proteases

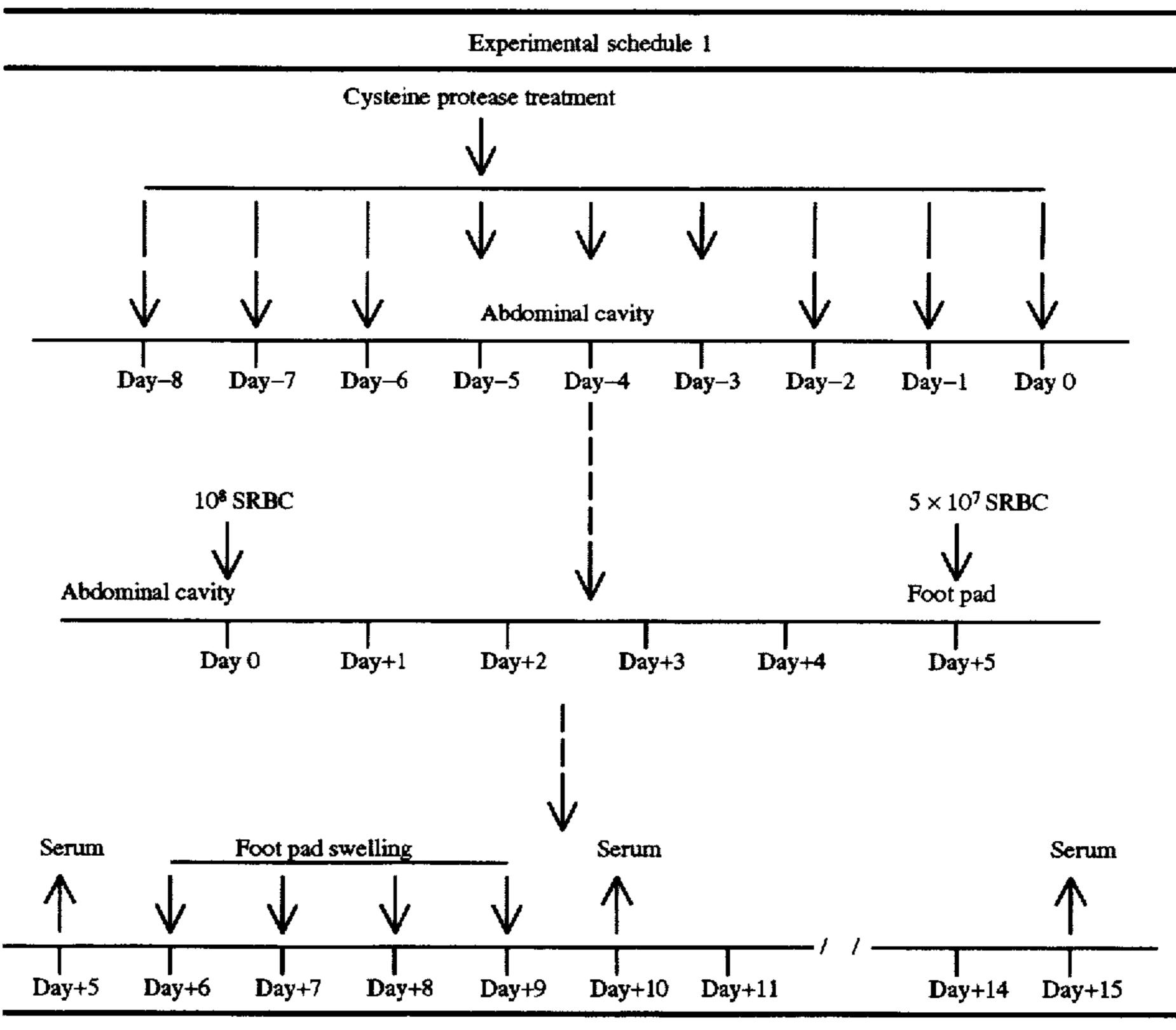
Amino acid	CP*	PPA*	CPPA*	PP*	CL*	CH*					
Arginine	3.80	8.30	3.16	6.44	3.69	1.86					
Proline	3.35	4.22	4.59	3.40	3.66	4.91					
Phenylalanine	3.00	2.42	3.59	1.83	4.67	5.87					
Isoleucine	4.77	6.25	3.80	5.33	2.78	5.60					
Histidine	1.69	1.14	1.68	2.29	1.64	1.10					

*CP: Cysteine protease, PPA: Papain, CPPA: Chymopapain, PP: Papaya proteinase, CL: Cathepsin L, CH: Cathepsin H

4 Suppressive Effects of Present Cysteine Protease on Cellmediated Immune Response and Humoral Immune Response.

12-week old C57BL/6 female mice were divided into 10 groups, each group having 6 mice. The present cysteine protease was intraperitoneally administered (100 ng protein per mouse which is an enough concentration to suppress footpad reaction) to the mice, except for a control group, 1, 25 2, 3, 4, 5, 7, and 8 days before the administration of antigens and on the same day when antigens were administered. The mice were immunized with the intraperitoneal injection of 1×10⁸ sheep red blood cells (SRBC) suspended in 0.1 ml of phosphate buffed saline (PBS) as an initial antigen. Five days later. 5×10⁷ SRBCs suspended in 0.05 ml PBS as booster was injected to the footpad. Then, degree of swelling (thickness) was measured (Experimental schedule 1 shown in Table 2).

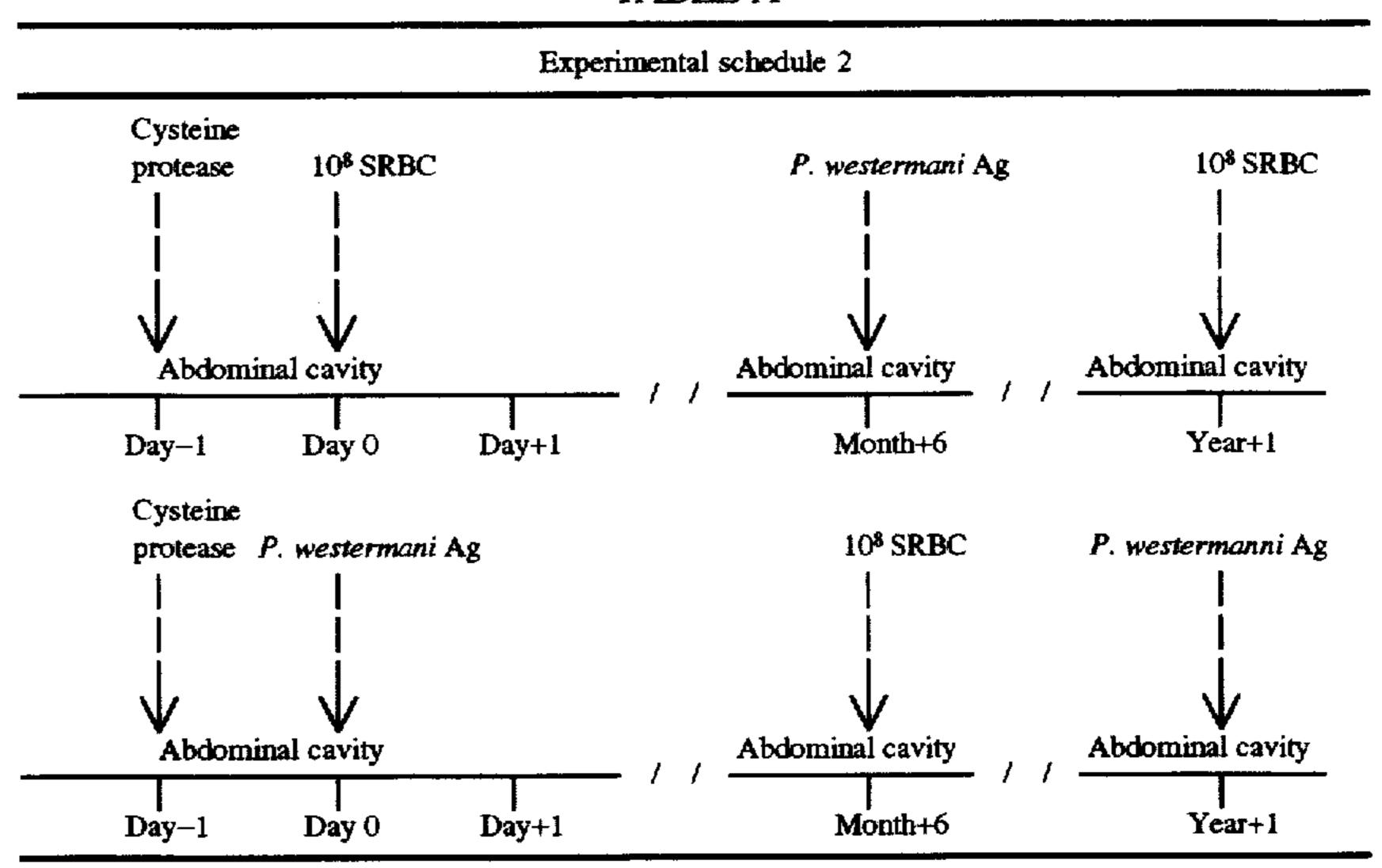
TABLE 2



Compared with control, footpad reaction (swelling) was significantly suppressed in the group (P<0.01) to which present cysteine protease was administered one day before immunization. This result suggests that administration of the

groups, the immunosuppression was specifically occurred to the antigen that was initially injected before one year while no immunosuppression of a footpad reaction was occurred to other antigens (FIGS. 3 and 4).

TABLE A



present cysteine prolease suppresses delayed-type ³⁰ hypersensitivity, against SRBC (FIG. 2).

Each group of mice was tested for the antibody titer (HA) of blood serum using SRBC. Compared with control, antibody production was significantly suppressed in the group of mice to which the present cysteine protease was adminis
stered 4-5 days before the injection of the antigen (Table 3).

TABLE 3

Day of CP	Anti-SRBC (HA) titer (Log ₂) on 15 days after immunization
treatment	Mean ± SE
Day 0	6.0 ± 0.26
Day-1	5.5 ± 0.22
Day-2	5.0 ± 0.31
Day-3	4.2 ± 0.31
Day-4	3.0 ± 0.31
Day-5	2.2 ± 0.17
Day-6	3.3 ± 0.37
Day-7	4.5 ± 0.22
Day-8	5.0 ± 0.30
Control	5.5 ± 0.34

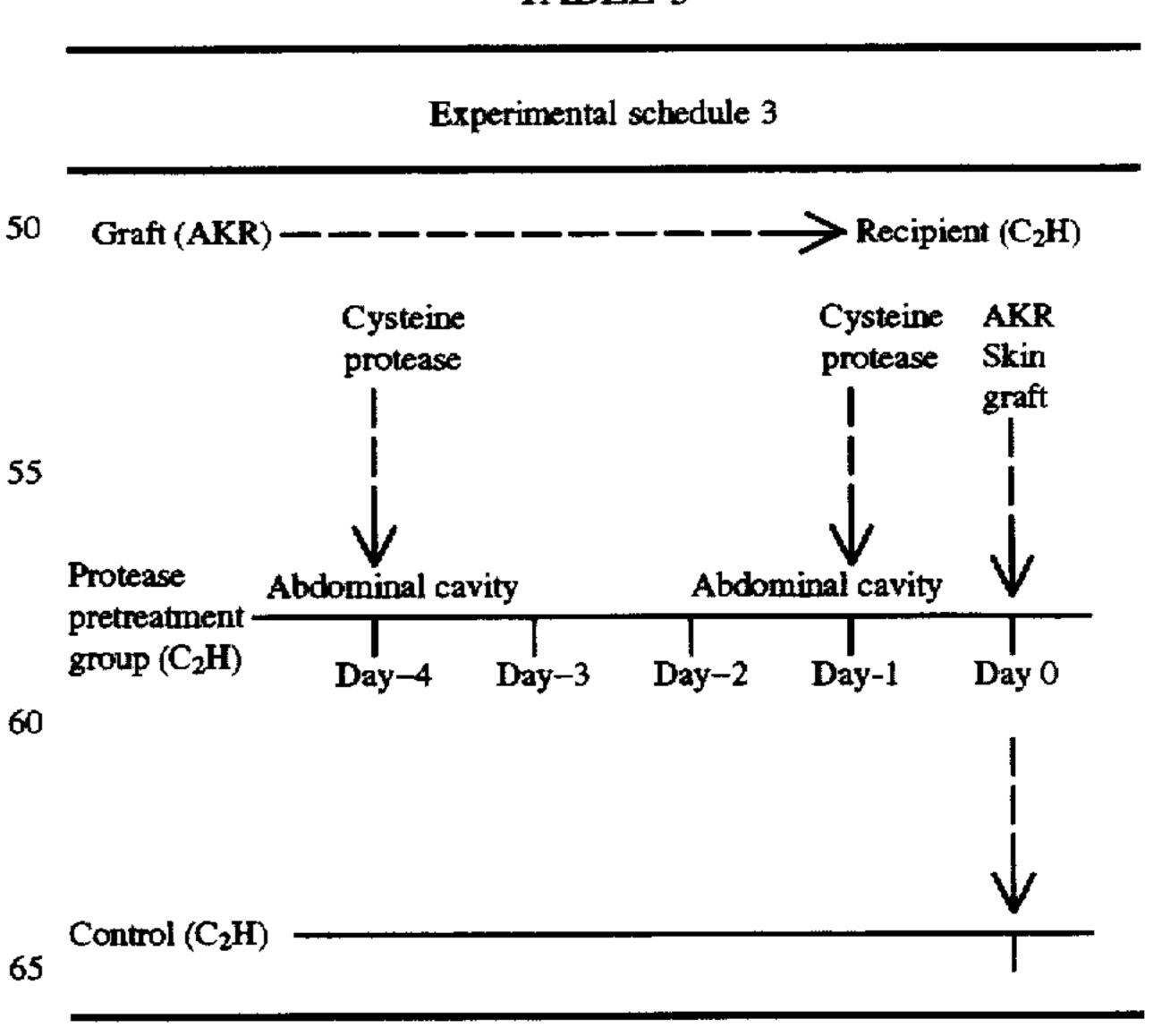
CP (Cysteine protease) and SE (Standard error)

5 Immunological Tolerance Induced by Present Cysteine Protease

The present protease (100 ng protein per mouse) was intraperitoneally administered to 12-week old B57BL/6 female and male mice. One day later, a group was immunized with SRBC or adult *Paragonimus westermani* antigens (Experimental schedule 2 shown in Table 4). The group demonstrated the suppression of a footpad reaction against SRBC or adult *Paragonimus westermani* antigens even one 65 year after the initial immunization (FIGS. 3 and 4). In these

As is shown in Experimental schedule 3 in Table 5, when the skin of 8-week old, AKR female mice was implanted to 10-week old, C3H/He female mice to which the present cysteine protease (total amount, 1.5 µg protein) had been administered one and 4 days before implantation, the administered group took the graft for a significantly longer period of time (P<0.05-0.01; mean survival time: 100±36) than control (mean survival time: 18±0.5 days) (Table 6).

TABLE 5



The abbreviations are mean survival time (MST), standard error (SE) and cysteine protease (CP)

As is evident from the results, the present cysteine protease intraperitoneally administered to mice was found to 15 induce immunological tolerance to the antigen or the tissue which were injected or implanted immediately after the administration of the present cysteine protease.

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6. Immunological Tolerance of Other Protease Species

Most of the different proteases (e.g., 100 ng of Plasmin, 20 Trypsin, Cathepsin B1, Papain, or Collagenase per mouse) intraperitoneally administered to 12-week old, C57BL/6 female mice did not induce immunological tolerance (FIG. 5). Weak suppression of a footpad reaction was observed in the collagenase administered group but no immunosuppression was observed one year later.

As is evident from the above experiment, the administration of the present cysteine protease suppresses delayed-type hypersensitivity and antibody production against specific antigens and implants, and induces immunological tolerance 30 in the mice.

Pharmaceutical agents utilizing the immunosuppressive effects of the present cysteine protease for therapeutic purpose may be in various forms including an injectable solution described in Example, inhalant, ointment, or a lyophilized form. Dosage may depend on how the present cysteine protease is administered and what therapeutic purpose one is in need.

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Although the present cysteine protease used in Example is one that is directly extracted from parasitic helminths, an amount of the cysteine protease that is extracted from one parasitic helminth is a very small. For the isolation and purification of the cysteine protease, it not only takes long time but is also costly so that it is desirable for a commercial scale of the preparation of the cysteine protease to produce it in a large amount by cultured animal cells or by genetic engineering using recombinant DNA containing a gene of interest (recombinant DNA technology).

SEQUENCE LISTING

(1) GENERAL INFORMATION: (i i i) NUMBER OF SEQUENCES: 5 (2) INFORMATION FOR SEQ ID NO:1: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 215 (B) TYPE:amino acid (C) STRANDEDNESS: (D) TOPOLOGY:Linear (i i) MOLECULE TYPE:peptide (vi) ORIGINAL SOURCE: (A) ORGANISM:Paragonimus westermani metacecaria B) CELL TYPE: (C) CELL LINE: (i x) FEATURE: (A) LOCATION: 5 (B) NAME/KEY: Xaa:Met or Ile (i x) FEATURE: (A) LOCATION:15 B) NAME/KEY:Xaa:Ala or Pro (i x) FEATURE: (A) LOCATION:21 (B) NAME/KEY:Xaa:Ser or Glu (i x) FEATURE: (A) LOCATION:58 (B) NAME/KEY:Xaa:Arg or Met

(i x) FEATURE:

-continued

- (A) LOCATION:59
- (B) NAME/KEY:Xaa:Val or Ala
- (i x) FEATURE:
 - (A) LOCATION:61
 - (B) NAME/KEY:Xaa:Gln or Glu
- (i x) FEATURE:
 - (A) LOCATION:69
 - (B) NAME/KEY:Xaa:Ala or Ser
- (i x) FEATURE:
 - (A) LOCATION:77
 - (B) NAME/KEY:Xaa:Tyr or Asp
- (x i) SEQUENCE DESCRIPTION:SEQ ID NO:1:

Ala	Рто	Glu	Агд	X a a 5	A s p	Trp	Arg	Ala	L y s 10
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Xaa	C y s	Gly	Ser		Тгр				T h r 3 0
Ala	G 1 y	A s n	V a 1	G l u 3 5	G 1 y	G 1 n	Ттр	Phe	I l e 4 0
Lys	Thr	Gly	Gin	L e u 4 5	V a 1	Ser	Leu	Ser	L y s 5 0
Gln	Gla	Leu	V a 1		Cys			Хаа	Ala 60
Хаа	G 1 y	C y s	Asn		Gly		Pro	Xaa	S e 1 7 0
Ser	Тут	Leu	G1u		Met		Met	G 1 y	G 1 y 8 0
Leu	Glu	Ser	G 1 u	S e r 8 5	Asp	Туr	Pro	Туг	V a 1 9 0
Gly	V a l	G 1 u	G l n		C y s			Asn	L y s 100
G l u	L y s	Leu	Val		L y s				S e r 1 1 0
Ιlε	V a 1	Leu	Gly		Glu				H i s 1 2 0
A 1 a	Ala	Tyr	Leu	A 1 a 1 2 5	Glu	His	Gly	Pro	L c u 1 3 0
Ser		Leu							G l n 1 4 0
Туг	Туг	Gln	Ser		V a 1		Lys	Pro	Thr 150
Phe	G 1 u	Glu	Сys		Asp		Glu	L¢u	Asn 160
His	Ala	Val	Leu	Thr 165	Val			Asp	Lys 170
Glu	Gly	Asp	M e t		Туг			I 1 e	Lys 180
Asn	Ser	Trp	G 1 y	T h r 1 8 5	Asp	Trp	G 1 y	G 1 u	Lys 190
G l y	Туг	Phe	Arg	Leu 195	P h e	Агд	G 1 y	A s p	C y s 2 0 0
Thr	C y s	G 1 y	I 1 e	A s n 2 0 5	Arg	Met	Ala	Thr	S e r 2 1 0

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Ala lle Ile Lys Lys
                            2 1 5
( 2 ) INFORMATION FOR SEQ ID NO:2:
         ( i ) SEQUENCE CHARACTERISTICS:
                  ( A ) LENGTH: 648
                   (B) TYPE nucleic acid
                  (C) STRANDEDNESS: single
                  (D) TOPOLOGY: linear
       ( i i ) MOLECULE TYPE:cDNA to mRNA
       (vi) ORIGINAL SOURCE:
                  ( A ) ORGANISM:Paragonimus westermani metacecaria
                  (B) CELL TYPE:
                  (C) CELL LINE:
       ( i x ) FEATURE:
       ( i x ) FEATURE:
                  (A) LOCATION:15
                  (B) NAME/KEY:N:T or G
       ( i x ) FEATURE:
                  (A) LOCATION:43
                  (B) NAME/KEY:N:G or C
       ( i x ) FEATURE:
                  ( A ) LOCATION:48
                  (B) NAME/KEY:N:T or G
       ( i x ) FEATURE:
                  (A) LOCATION:61
                  (B) NAME/KEY:N:T or G
       ( i x ) FEATURE:
                  (A) LOCATION:62
                  (B) NAME/KEY:N:C or A
       ( i x ) FEATURE:
                  (A) LOCATION:66
                  (B) NAME/KEY:N:C or T
       ( i x ) FEATURE:
                  ( A ) LOCATION:123
                  (B) NAME/KEY:N:A or G
       ( i x ) FEATURE:
                  (A) LOCATION:173
                  (B) NAME/KEY:N:G or T
       ( i x ) FEATURE:
                  (A) LOCATION:176
                  (B) NAME/KEY:N:T or C
       (ix) FEATURE:
                  (A) LOCATION:181
                  (B) NAME/KEY:N:C or G
       ( i x ) FEATURE:
                  (A) LOCATION:205
                  (B) NAME/KEY:N:G or T
       ( i x ) FEATURE:
                  (A) LOCATION:213
                  (B) NAME/KEY:N:C or A
       ( i x ) FEATURE:
                  (A) LOCATION:229
                  (B) NAME/KEY:N:T or G
       ( i x ) FEATURE:
                  (A) LOCATION:237
                  (B) NAME/KEY:N:C or T
       ( i x ) FEATURE:
                  (A) LOCATION:306
                  (B) NAME/KEY:N:G or A
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1	í	Ŧ) FEATURE:
1	•		A T THE STATE OF STATE OF

- (A) LOCATION:366
- (B) NAME/KEY:N:C or T

(i x) FEATURE:

- (A) LOCATION:370
- (B) NAME/KEY:N:T or C

(i x) FEATURE:

- (A) LOCATION:375
 - (B) NAME/KEY:N:A or T

(i x) FEATURE:

- (A) LOCATION:471
- (B) NAME/KEY:N:T or C

(i x) FEATURE:

- (A) LOCATION:489
- (B) NAME/KEY:N:T or G

(x i) SEQUENCE DESCRIPTION:SEQ ID NO:2:

GCT	ссс	G A A	CGT	ATN	GAC	TGG	CGG	GCT	AAG	3 0
GGC	GCT	GTG	A C A	NCG	GTN	G A A	A A T	CAA	GGC	6 0
NNG	TGN	GGT	TCG	TGT	TGG	GCG	TTC	TCG	ACT	9 0
GCC	GGA	AAC	GTT	GAA	GGT	C A A	TGG	TTC	ATC	1 2 0
AAN	ACC	GGT	C A G	СТТ	GTC	A G T	CTG	AGC	AAA	1 5 0
CAG	C A A	TTG	GTC	GAC	TGT	GAC	ANG	GNG	GCC	180
N A G	GGA	TGC	AAT	GGT	GGA	TGG	C C A	NCC	AGT	2 1 0
TCN	TAC	CTG	G A A	ATC	A T G	NAT	A T G	GGN	GGT	2 4 0
TTG	GAG	TCC	G A A	AGC	GAC	TAT	ссс	TAT	GTT	270
GGT	GTG	G A A	CAA	ACG	TGT	GCC	CTG	AAC	A A G	3 0 0
GAG	AAN	CTG	G T A	GCC	AAA	ATC	GAT	G A T	TCG	3 3 0
ATT	GTT	C T G	GGT	CCG	GAG	GAG	GAG	GAC	CAC	3 6 0
GCC	GCN	ТАТ	NTG	GCN	G A A	CAC	GGA	CCG	TTG	390
A G T	A C G	CTG	CTC	AAT	GCC	GTC	GCT	CTT	CAG	4 2 0
TAC	TAC	C A G	TCC	GGA	G T A	CTC	AAA	CCG	ACC	4 5 0
TTT	GAG	G A G	TGT	CCG	GAT	ACN	G A G	TTG	AAC	480
CAC	GCG	G T N	CTC	ACC	GTC	GGC	TAT	GAC	A A G	5 1 0
G A A	GGC	GAT	ATG	CCA	TAC	TGG	ATC	ATC	AAG	5 4 0
A A T	A G T	TGG	GGT	ACC	GAC	TGG	GGC	GAG	AAA	5 7 0
GGC	TAC	TTC	CGA	CTC	TTC	CGA	GGA	GAT	T G C	600
A C G	TGT	GGA	ATC	AAC	CGC	A T G	GCA	ACA	T C C	6 3 0
GCG	ATC	ATC	A A G	AAA	TGA					6 4 8

(2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 648
 - (B) TYPE mucleic acid
 - (C) STRANDEDNESS:single
 - (D) TOPOLOGY: linear
- (i i) MOLECULE TYPE:cDNA to mRNA
- (v i) ORIGINAL SOURCE:
 - (A) ORGANISM:Paragonimus westermani metacecaria

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							<u> </u>			
		'	B) CELL C) CELL							
	(i x) FEATUR	RE:							
	(x i) SEQUE	NCE DES	CRIPTIO	N:SEQ ID	NO:3:				
G С Т	000	GAA	CGT	АТТ	GAC	TGG	CGG	GCT	AAG	3 0
			Arg						L y s	
				5					1 0	
GGC	GCT	GTG	ACA	GCG	GTT	GAA	A A T	CAA	GGC	6 0
Gly	Ala	Val	Thr	Ala	V a 1	Glu	Asn	Gln	G 1 y 2 0	
				1.5					20	
			TCG							9 0
SEI	Суѕ	Gry	Ser	2 5	ı ı p	Ais	г п с	361	30	
C C C	C C 4		e e e e	C 1 1	007	C 4 4	тсс	T T C	A T C	1 2 0
			GTT Val							
	•			3 5	•		-		4 0	
AAA	ACC	GGT	CAG	СТТ	GTC	AGT	CTG	AGC	AAA	1 5 0
L y s	Thr	Gly	Gln		V a 1	Ser	Leu	Ser		
				4 5					5 0	r
CAG	CAA	TTG	GTC	GAC	TGT	GAC	AGG	G T G	GCC	180
G 1 n	Gln	Leu	Val	Asp 55	Cys	Asp	Arg	V a 1	A.la 60	
				3 3					0 0	
			AAT Asn							2 1 0
GII	Gry	Суѕ	W 2 II	6 5	O i y	119	110	Ала	70	
T C C	T . C	C T C	GAA	4 T.C	A T.G	T A T	ATG	aac	CCT	2 4 0
			Glu							
				7 5					8 0	
TTG	GAG	тсс	GAA	AGC	GAC	TAT	ссс	TAT	GTT	270
Leu	Glu	Ser	Glu		Asp	Туг	Pro	Туг		
				8 5					90	
			CAA							3 0 0
Gly	Val	Glu	Gln	Thr 95	Суѕ	Ala	Leu	Asn	Lys 100	
	_									
			GTA Val							3 3 0
	_ , -			1 0 5	,			•	1 1 0	
АТТ	GTT	СТС	GGT	CCG	GAG	GAG	GAG	GAC	CAC	360
			G 1 y	Pro					H i s	
				1 1 5					120	
GCC	GCC	TAT	TTG	GCA	GAA	CAC	GGA	CCG	TTG	390
Ala	Ala	Туг	Leu	A 1 a 125	Glu	His	G 1 y	Рго	Leu 130	
<u>-</u>			CTC Leu							4 2 0
261	1 11 1	Leu	Leu	1 3 5	Ala	∀ a 1	АГА	Leu	140	
T A C	T A C	CAG	тсс	GGA	GTA	стс		CCG	A C C	4 5 0
			Ser							
				1 4 5					150	
ттт	GAG	GAG	TGT	CCG	GAT	ACT	GAG	TTG	AAC	480
P h e	Glu	Glu	C y s		_	Thr	Glu	Leu		
				1 5 5					160	
			CTC							5 1 0
His	Ala	V a l	Leu	Thr 165		Gly	Туг	Asp	Lys 170	
			ATG Mct							5 4 0
	,	· P	* 1	175		F	- 		180	
ΔΑΤ	ልጤፕ	ፐርር	GGT	ACC	GAC	тсс	GGC	GAG	A A A	5 7 0
AAI	WAT	100	901	ALL	JAC		330	JAU	AAA	

				19						20
								-co	ntinued	
A s n	Ser	Ттр	Gly	Thr 185	Asp	Trp	G 1 y	Glu	Lys 190	
GGC	TAC	TTC	CGA	СТС	TTC	CGA	GGA	GAT	TGC	600
Gly	Туг	P h c	Агд	Leu 195	Phe	Arg	G 1 y	Asp	C y s 2 0 0	
ACG	TGT	GGA	ATC	AAC	CGC	ATG	GCA	ACA	TCC	6 3 0
Thr	Суs	Gly	lle	A s n 2 0 5	Arg	Met	Ala	ТЪг	S e r 2 1 0	
				A A A L y s 2 1 5						648
(2)1	NFORM.	ATION FO	R SEQ II	D NO:4:						
	(i	() ()	A) LENC B) TYPE C) STRA	ARACTER 5TH: 648 Emucicie a ANDEDNI OLOGY:lin	icid ESS:single	•				
	(i i) MOLEC	CULE TY	PE:cDNA	to mRNA	•				
	(vi	()		ANISM:Pa . TYPE:	aragonimu	is western	ani metac	ecaria		
	(i x) FEATU	RE:							
	(x i) SEQUE	NCE DES	CRIPTIO	N:SEQ II	NO:4:				
								GCT		3 0
								CAA		6 0
								T C G S e r		90
								TTC Phe		1 2 0
								AGC Ser		150
L, , s		. ,	0.2	4 5	• • •			J V 1	5 0	
								G C G A l a		180
GAG	GGA	TGC	AAT	сст	GGA	таа	CCA	тсс	AGT	2 1 0
								Ser		
								GGT		2 4 0
Ser	Tyr	Leu	Glu	I 1 e 7 5	Met	Asp	Met	Gly	G 1 y 8 0	
								TAT		270
								AAC Asn		3 0 0
								GAT Asp		3 3 0
ATT	GTT	C T G	GGT	CCG	G A G	G A G	G A G	G A C	CAC	3 6 0

	-continued										
I 1 ¢	Val	Leu	G 1 y	Pro 115	Glu	Glu	Glu	Asp	His 120		
					GAA Glu						390
					GCC Ala						4 2 0
					GTA Val						4 5 0
					GAT Asp						480
					GTC Val						5 1 0
					TAC						5 4 0
		-			GAC Asp						5 7 0
					TTC Phe						600
					CGC Arg		_				630
		ATC Ile									6 4 8

(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25
- (B) TYPE:amino acid
- (C) STRANDEDNESS:
- (D) TOPOLOGY:linear

(i i) MOLECULE TYPE:peptide

(v i) ORIGINAL SOURCE:

- (A) ORGANISM:Paragonimus westermani metacecaria
- (B) CELL TYPE:
- (C) CELL LINE:

(i x) FEATURE:

(A) OTHER INFORMATION:

The 22nd and 25th residues, Xaa, are to be cysteine which could not be detected with this PTH analyser system. The amino acid sequence shown below is N-terminal 25 amino acids of the purified native cysteine protease.

(x i) SEQUENCE DESCRIPTION:SEQ ID NO:5:

Ala Pro Glu Ser lle Asp Trp Arg Glu Lys

5

Gly Ala Val Ala Pro Val Glu Asp Gln Gly 15 20

Ser Xaa Gly Ser Xaa

2 5

What is claimed is:

- 1. A composition comprising a biologically active cysteine protease derived from parasitic helminths wherein the amino acid sequence is at least 90% identical to the amino acid sequence of SEQ ID No. 1.
- 2. The composition of claim 1 wherein the cysteine protease has an amino acid sequence in which the total number of acidic amino acid residues in the sequence exceeds the total number of basic amino acid residues in the sequence.
- 3. The composition of claim 1 wherein the cysteine 10 protease is active at about pH 7.
- 4. The composition of claim 2 wherein the cysteine protease is active at about pH 7.
- 5. The composition of claim 2 wherein the cysteine protease has the amino acid sequence of SEQ ID No. 1.
- 6. The composition of claim 3 wherein the cysteine protease has the amino acid sequence of SEQ ID No. 1.
- 7. The composition of claim 4 wherein the cysteine protease has the amino acid sequence of SEQ ID No. 1.
- 8. A composition comprising a biologically active cysteine protease derived from parasitic helminths wherein the amino acid sequence is at least 90% identical to the amino acid sequence of SEQ ID No. 1 and wherein the amino acid sequence of the cysteine protease has a proline residue at position 15, a glutamate residue at positions 21 and 61, a cysteine residue at positions 22, 25, 56, 63, 96, 154 and 202, a methionine residue at position 58, an alanine residue at position 59, a serine residue at position 69, an aspartate residue at position 77 and a histidine residue at position 161, wherein the amino acid residue positions are relative to the N terminus.

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- 9. The composition of claim 8 wherein the cysteine protease has an amino acid sequence in which the total number of acidic amino acid residues in the sequence exceeds the total number of basic amino acid residues in the sequence.
- 10. The composition of claim 8 wherein the cysteine protease is active at about pH 7.
- 11. The composition of claim 8 wherein the cysteine protease is from infected larvae of parasitic helminths.
- 12. A composition comprising a cysteine protease with an amino acid sequence of SEQ ID number 1.
- 13. A composition comprising a biologically active cysteine protease derived from parasitic helminths wherein the amino acid sequence is at least 90% identical to the amino acid sequence of SEQ ID No. 1 and wherein the amino acid sequence of the cysteine protease has a proline residue at position 15, a glutamate residue at positions 21 and 61, a cysteine residue at positions 22, 25, 56, 63, 96, 154 and 202, a methionine residue at position 58, an alanine residue at position 59, a serine residue at position 69, an aspartate residue at position 77 and a histidine residue at position 161. wherein the amino acid residue positions are relative to the N terminus wherein the cyateine protease has an amino acid sequence in which the total number of acidic amino acid residues in the sequence exceeds the total number of basic amino acid residues in the sequence wherein the cysteine protease is active at about pH 7.

* * * *