Eba	ta et al.		[45]	Date of Patent:	Feb. 5, 1985		
[54]		TOSIDASE INHIBITOR GT-2558 DERIVATIVES	[58] Field of Search				
[75]	Inventors:	Mitsuo Ebata; Yukio Miyake;	[56] References Cited U.S. PATENT DOCUMENTS				
		Yoshimi Kawamura, all of Osaka, Japan		,210 8/1976 Mizuno et al. ,183 11/1976 Celmer et al.			
[73]	Assignee:	Shionogi & Co., Ltd., Osaka, Japan					
[21]	Appl. No.:	393,506	_	Examiner—Jerome D. Go Agent, or Firm—Wender	_		
[22]	Filed:	Jun. 29, 1982	[57]	ABSTRACT			
	Rela	ted U.S. Application Data		osidase inhibitors GT-25:	•		
[63]	63] Continuation-in-part of Ser. No. 311,326, Oct. 14, 1981, abandoned.			GT-2558-B produced by a strain <i>Streptomyces lydic</i> PA-5726, and their derivatives, acetyl-GT-2558-A at acetyl-GT-2558-B, useful as diagnostatic aid in β-gala			
[30]	Foreig	n Application Priority Data	tosidase o	deficiency, used in purific	ation of enzymes, in		
Oc	t. 27, 1980 [J]	P] Japan 55-150927		of new β -galactosidase, is actose-containing foods	_		

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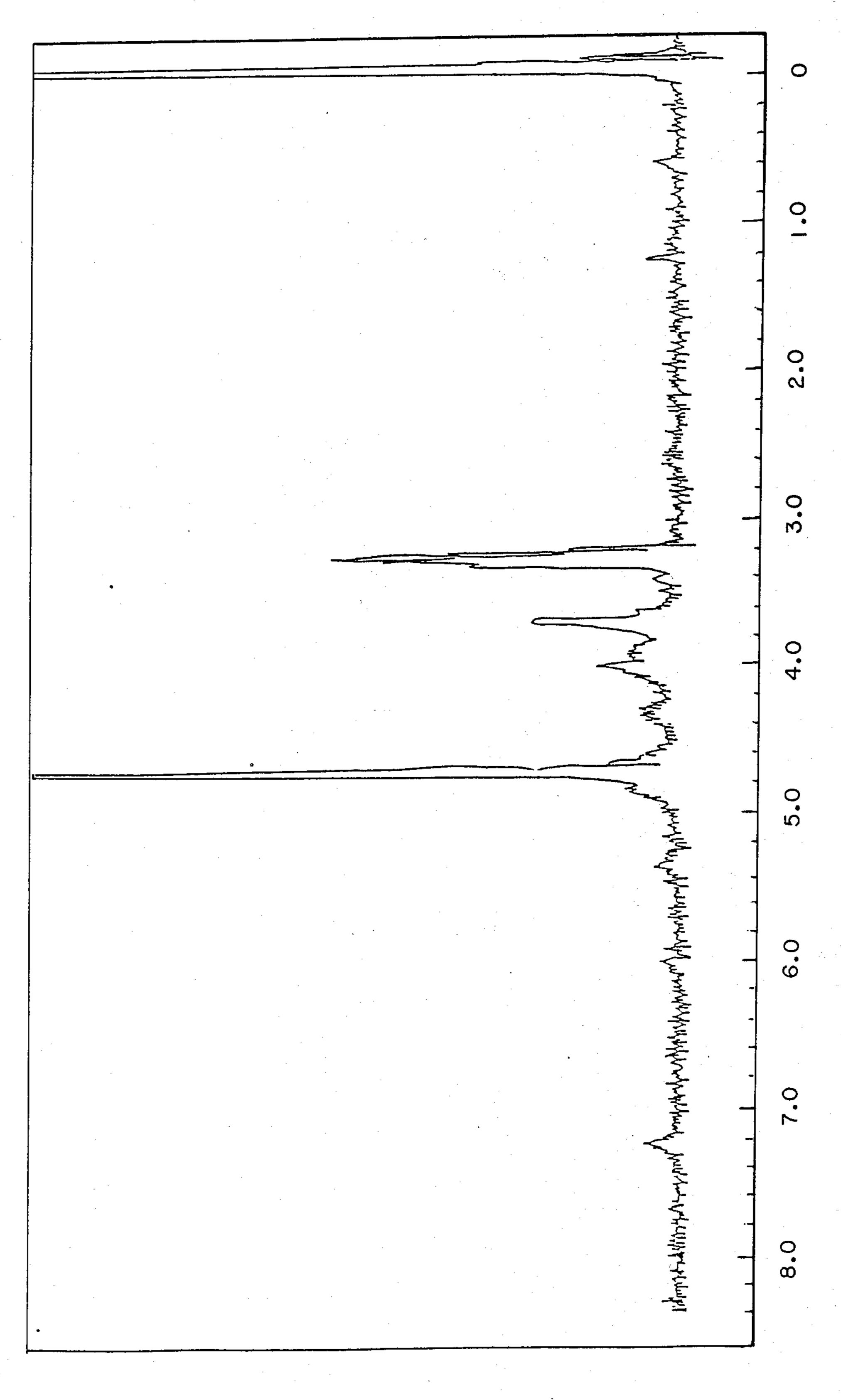
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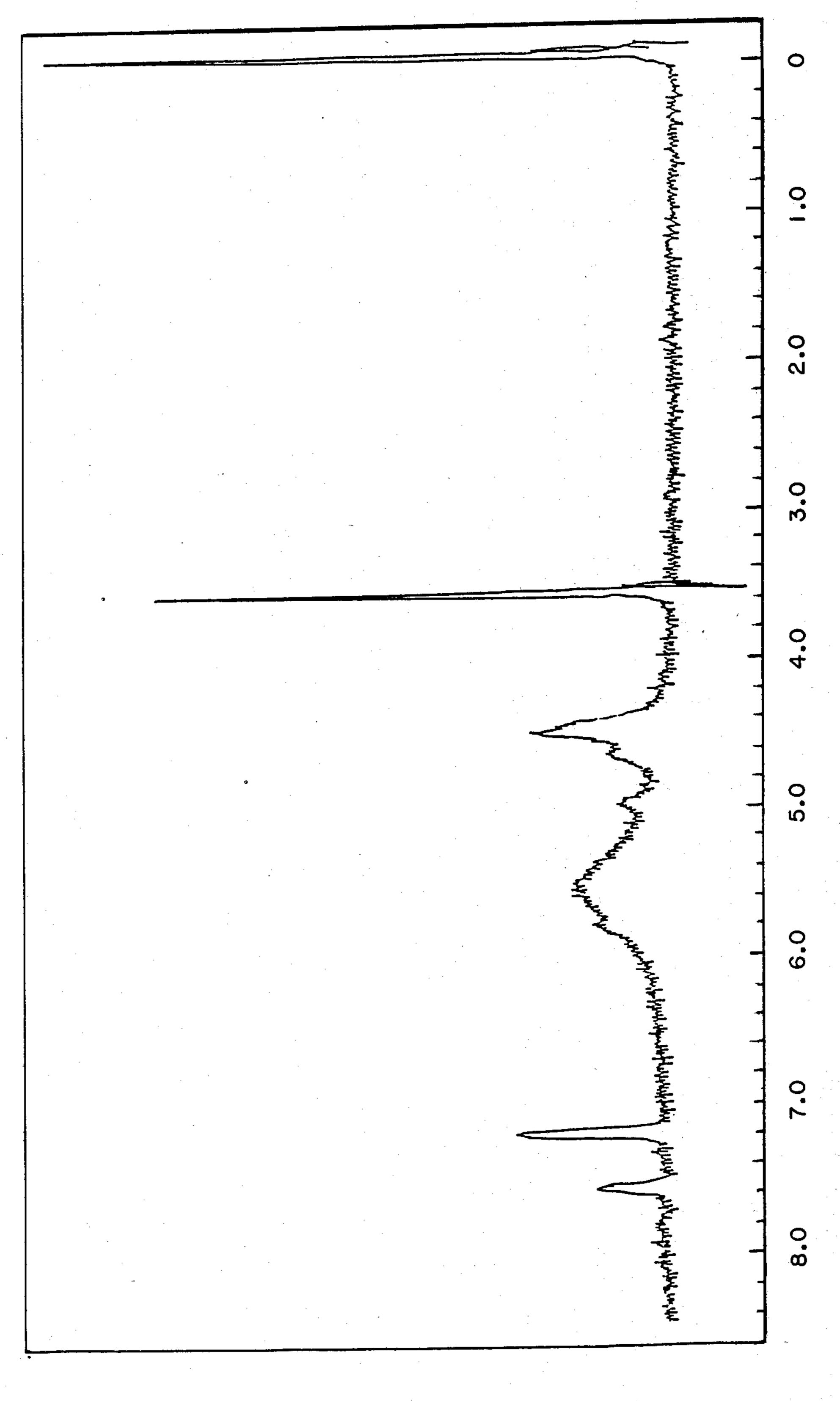
4 Claims, 4 Drawing Figures

United States Patent [19]

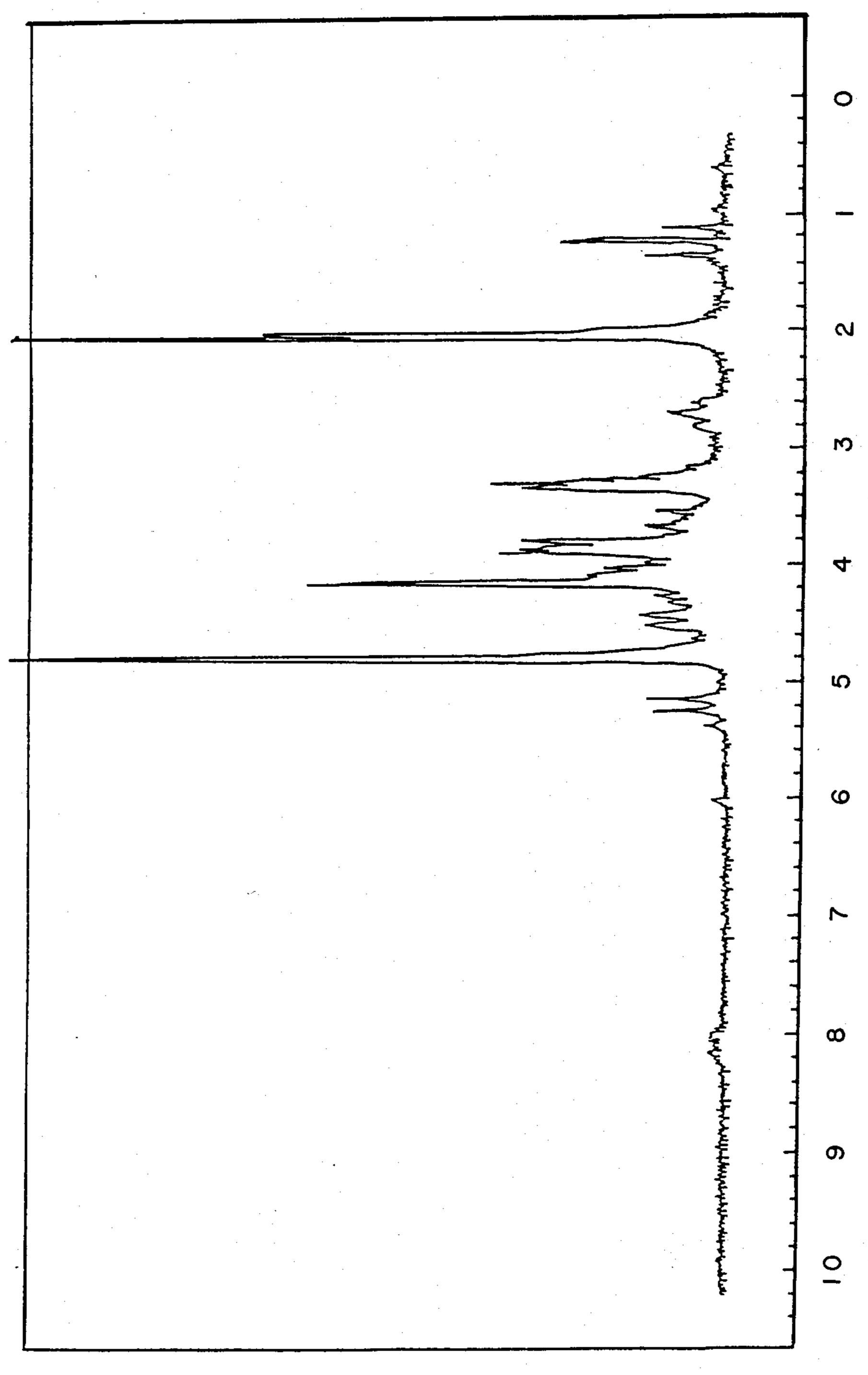
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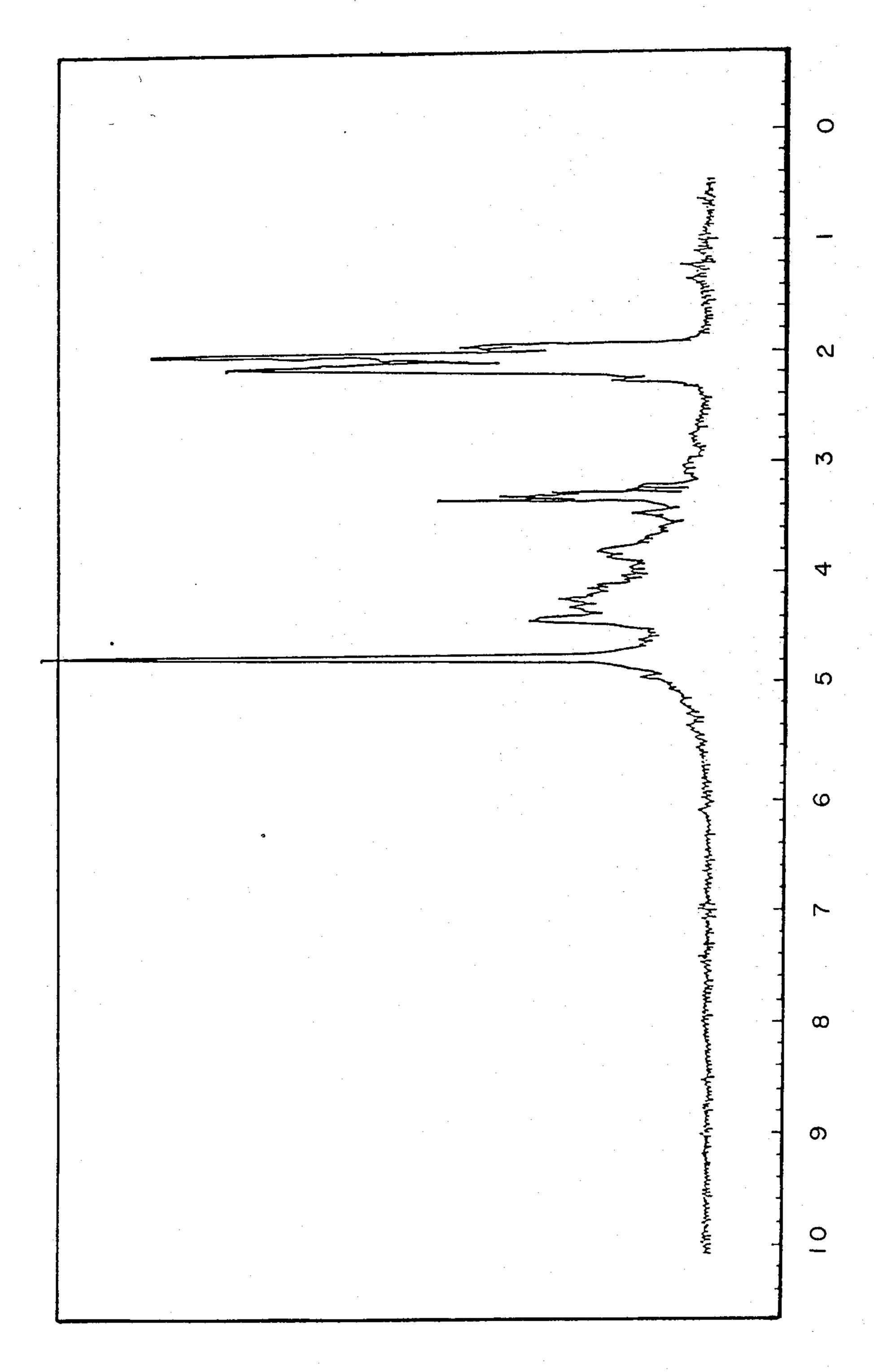
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F16.



=1G. 3



F1G. 4

β-GALACTOSIDASE INHIBITOR GT-2558 AND ITS DERIVATIVES

This application is a continuation-in-part of applica- 5 tion Ser. No. 311,326, filed Oct. 14, 1981 (now abandoned).

BACKGROUND OF THE INVENTION

It has been reported by H. B. Bosmann [Biochem. 10 Biophys. Acta. 264, 339 (1972)] that glucosidase activity, particularly, β -galactosidase activity is elevated in 3T3 fibroflast cells transformed by RNA tumor virus. In this connection, various researches have been made and described in, e.g., Jap. Pat. Publication No. 15 53-31238; The Journal of Antibiotics 28, 1006 (1975); ibid. 32 (3), 212 (1979); and ibid. 32 (3), 217 (1979). The present inventors have also investigated intensively to find the substances having β -galactosidase inhibitory activity, and as the result, they have found novel β - 20 galactosidase inhibitor GT-2558 in the culture broth resulted from a strain of certain microorganisms and accomplished this invention.

SUMMARY OF THE INVENTION

The present invention relates to novel β -galactosidase inhibitor GT-2558 and its derivatives, and a method for producing them.

BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1 and FIG. 2 show the NMR spectrums of the GT-2558-A measured in heavy methanol and d₅-pyridine respectively.

FIG. 3 shows the NMR spectrum of the acetyl-GT-2558-B measured in heavy methanol.

FIG. 4 shows the NMR spectrum of the acetyl-GT-2558-A measured in heavy methanol.

The β-galactosidase inhibitor GT-2558 in the present invention is a mixture of A, B, and other minor components and can be separated into GT-2558-A and GT-40 2558-B in a conventional manner. Additionally these compounds may be stabilized by acetylation. The properties of GT-2558, GT-2558-A and B, and their acetyl derivatives are shown as follows. The present invention includes not only these compounds per se but also the 45 pharmaceutically acceptable salts of these compounds. (1) GT-2558

a. Nature

Weakly basic powder

b. Solubility

Freely soluble in water, soluble in methanol, acetic acid, and pyridine, slightly soluble in ethanol and propanol, and insoluble in acetone, ether, ethyl acetate, chloroform, and benzene.

c. Color reaction

Ninhydrin reaction test positive

Silver mirror test positive

Tollens reaction test (with yellow phloroglucinol and hydrochloric acid)

d. UV

No characteristic absorption

e. TLC on a silica gel plate

 R_f =0.29 (developed with acetonitrile:acetic acid:water=5:1:2)

f. Specific β-galactosidase inhibitory activity 55,000 IU/mg (IC₅₀ 0.0045 mcg)

(2) GT-2558-A

a. m.p.

106°-108° C.

b. Molecular weight (mass spectrum)
Maximum mass m/e=302

c. Elemental analysis

Found. C,41.34; H,6.16; N,7.43; O, 45.06 (%)

d. IR

 ν_{max}^{KBr} 3400, 2900, 1640, 1565, 1505, 1455, 1415, 1360, 1300, 1270, 1160, 1110, 1075, 1035, 955, 930, 890, 855, 795 cm⁻¹.

e. NMR

FIG. 1 and FIG. 2

f. β -Galactosidase inhibitory activity

Enzyme	pH at	Origin	IC ₅₀	
Taka β- galactosidase	5.0	Aspergillus oryzae(Sankyo)	0.0018 п	ncg
Taka β- galactosidase	4.5	Aspergillus oryzae(Sankyo)	0.0045	
Galantase	4.5	Aspergillus oryzae(Tokyo Tanabe)	0.0045	
Bovine β -galactosidase	7.2	Bovine liver (Sigma)	26.6	

g. Specific β-galactosidase inhibitory activity 136,700 IU/mg (IC₅₀ 0.0018 mcg)

(3) Acetyl-GT-2558-A

a. Nature

Colorless fine crystals (recrystallized from benzene-methanol)

b. Solubility

Soluble in methanol, ethanol, chloroform, dioxane, and water, slightly soluble in ethyl acetate, and insoluble or hardly soluble in carbon tetrachloride and benzene.

c. Color reaction

Ninhydrin reaction test positive

Ammoniac silver nitrate reaction test positive

d. m.p. and decom.p.

m.p. 41° C.; decomp.p. 149°-153° C.

e. Specific rotation

 $[\alpha]_D^{24} + 29.6^{\circ}$ (c 1.0 methanol)

f. Molecular weight (mass spectrum)
Maximum mass m/e=491 (M+1)

g. Elemental analysis

Found. C,51.30; H,6.27; N,5.65; O,36.78 (%)

h. UV

No characteristic absorption

i. IR

60

65

 ν_{max}^{KBr} 3430, 2920, 1745, 1630, 1435, 1390, 1370, 1245, 1090, 1040, 820, 760 cm⁻¹.

(4) Acetyl-GT-2558-B

a. Nature

Colorless fine crystals (recrystallized from ethyl acetate-methanol)

b. Solubility

Soluble in methanol, ethanol, and water, slightly soluble in ethyl acetate and chloroform, and insoluble or hardly soluble in carbon tetrachloride and benzene.

c. Color reaction

Ninhydrin reaction test positive

Ammoniac silver nitrate reaction test positive

d. m.p. and decomp.p.

m.p. 76°-78° C.; decomp.p. 161°-163° C.

e. Specific rotation

 $[\alpha]_{D^{24}} + 13.8^{\circ}$ (c 1.0 methanol)

f. Molecular weight (mass spectrum)

Maximum mass m/e=429 (M+1)

g. Elemental analysis

Found. C,46.58; H,6.58; N,6.56; O,38.51 (%)

h. UV

No characteristic absorption

i. IR

 ν_{max}^{KBr} 3410, 2930, 2735, 2360, 1640, 1550, 1435, 1380, 1245, 1195, 1115, 1090, 1045, 875, 815 cm⁻¹.

j. NMR

FIG. 3

k. Specific β-galactosidase inhibitory activity 1,640 IU/mg (IC₅₀ 0.15 mcg)

The β -galactosidase inhibitory activity and the specific β -galactosidase inhibitory activity shown in the 1: above properties are observed by means of colorimetric determination of o-nitrophenol which is liberated on hydrolysis of the substrate o-nitrophenyl β -D-galactopyranoside catalyzed by β -galactosidase. In this connection, to a solution (0.25 ml) of β -galactosidase dis- 20 solved in 0.1M acetic acid buffer (pH 5.0), a solution (0.25 ml) containing test compounds is added, incubated at 30° C. for 10 minutes, and mixed with 0.01M substrate solution (0.5 ml) of o-nitrophenyl β -D-galactopyranoside dissolved in the above buffer. The mixture is incu- 25 bated at 30° C. for 15 minutes, the reaction is stopped by the addition of 1M sodium carbonate (1 ml), the total volume is adjusted to 10 ml by addition of water (8 ml), and then the absorbance (A) at 420 nm is measured. Concurrently the absorbance (B) of the control pre- 30 pared with only the buffer not containing the above test compounds is measured, and the inhibitory rate is calculated from $(B-A)/B \times 100$. Wherein β -galactosidase prepared from Taka-diastase (trade name, Sankyo Co.) is employed. One unit of inhibitory activity (1 IU) is 35 defined as the amount of a test compound required for 50% inhibition (IC₅₀) of the β -galactosidase activity in the above condition.

The GT-2558 seems to be a novel β -galactosidase inhibitor since the properties are different from the 40 known β -galactosidase inhibitors. As an organism which produces the said GT-2558, the strain of PA-5726 which was isolated from the soil sample collected in Nakadori Island, Minami-Matsuura District, Nagasaki Pref., Japan is shown.

This strain was determined to be a strain of Streptomyces lydicus classificationally, and the strain of PA-5726 has been deposited as Streptomyces lydicus PA-5726, FERM-BP61 in Agency of Industrial Science & Technology, Ibaraki, Pref., Japan, and as ATCC No. 31975 50 at American Type Culture Collection, Rockville, Md. U.S.A.

The taxonomical properties of the said strain are shown as follows.

(a) Morphological characteristics (Bennett's 55 agar, cultured at 28° C. for 14 days)

Neither sporangium, flagellated spore, nor sclerotium is observed, and no split by fragmentation in substrate hyphae is observed. On this medium, it forms abundant aerial hyphae which develop into many spirals. The 60 surface structure of the spore is smooth under the electron microscopic observation. In this strain, with the lapse of time the surface of the colony is moistened to become so-called hygroscopic and change into black.

(b) Cultural views on various media (cultured at 28° 65 C. for 14 days)

Expression of color is in accordance with "Guide to Color Standard" (Japan Color Institute ed.).

		Aerial	Hyphae	Color of	
Media	Growth	Form- ation	Color	Substrate Hyphae	Soluble Pigmen
Sucrose. Nitrate Agar	fair	no	<u></u>	pale yel- lowish brown	по
Glucose. Asparagine Agar		fair	light brownish	pale yel- lowish	
Glycerin. Asparagine	good	good	gray light brownish	brown pale yel- lowish	"
Agar Inorganic salt.	. **	,,	gray brownish gray*	brown pale yel- lowish brown	,,
Starch Agar Tyrosine Agar	,,	"	light brownish gray	yellowish brown	"
Nutrient Agar	••	no		yellowish brown	· • • • • • • • • • • • • • • • • • • •
Yeast extract. Malt extract	**	good	brownish gray*	yellowish brown	,,
Agar Oatmeal Agar	,,	,,	brownish gray*	lowish	
Bennett's Agar	• • • • • • • • • • • • • • • • • • • •	"	brownish gray*	brown yellowish brown	***

^{*}The surface of the colony is moistened to become so-called hygroscopic and change into black.

Growth temperature (Bennett's agar, cultured at each temperature for 2 weeks. The growth at 10° C. was observed also 3 weeks after.)

10° C.:

Observed for 2 weeks, scarcely growth.

Observed for 3 weeks, good growth forming aerial hyphae fairly.

28° C.:

Growth, formation of aerial hyphae, and formation of spore each is good.

37° C.: No growth.

45° C.: No growth.

(c) Physiological properties (cultured at 28° C. for 14 days)

	Liquefaction of gelatin	positive
•	Production of melanoid pigment	negative
	Tyrosinase reaction	negative
	Peptonization of milk	positive
	Coagulation of milk	positive
	Hydrolysis of starch	positive

(d) Utilization of carbon source Sugar much required for growth

L-arabinose, D-xylose, D-glucose, D-fructose, su-crose, inositol, raffinose, and D-mannitol

Sugar slightly concerning for growth L-rhamnose

From the above description of various properties, it is obvious that this strain belongs to the genus Streptomyces. In Waksman's "The Actinomycetes" vol. 2 (1961), Shirling and Gottlieb's "International Streptomyces Project" report, ["International Journal of Systematic Bacteriology" vol. 18 p.69 and p.279 (1968), ibid. vol. 19 p.391 (1969), ibid. vol. 22 p.265 (1972)], "Bergey's Manual of Determinative Bacteriology" 8th Edition (1974), and other publications as to new species of actinomy-

cetes, the species closely related to the strain of PA-

5726 were searched, and Streptomyces lydicus [International Journal of Systematic Bacteriology vol. 19, p.448 (1969), Bergey's Manual of Determinative Bacteriology 8th Edition, p.772 & p.777 (1974), and U.S. Pat. No. 3,160,560 (1964)] was shown as the most closely related species. As a result of comparing the various properties of the strain PA-5726 with those of Streptomyces lydicus described in these publications, good agreements were obtained. Therefore the strain PA-5726 was determined to be a strain belonging to the same species as Streptomy- 10 ces lydicus and named Streptomyces lydicus PA-5726.

In the present invention not only the strain PA-5726 and its natural and artificial variants but also all of the strains which belong to Streptomyces lydicus and product GT-2558, can be used and are within the scope of this 15 invention.

Production of GT-2558 comprises culturing a GT-2558 producing strain on a nutrious medium under aerobic conditions and recovering GT-2558 from the culture broth after cultivation. General prodedure for pro- 20 ducing GT-2558 is described in the followings.

Referring to medium components and culture conditions, the conventional ones employed in producing antibiotics may be employed. The medium contains essentially carbon sources, nitrogen sources, inorganic 25 salts, etc. If necessary, vitamines, precursors, etc. may be added. As carbon sources, for example, glucose, starch, dextrin, glycerin, molasses, and organic acid are employed alone or as a mixture. As nitrogen sources, for example, soybean meal, corn steep liquor, meat 30 extract, yeast extract, cotton seed meal, peptone, wheat embryo, ammonium sulfate, and ammonium nitrate are employed alone or as a mixture. As inorganic salts, for example, calcium carbonate, sodium chloride, potassium chloride, magnesium sulfate, cobaltous chloride, 35 and various phosphates may be added to the medium, if necessary.

The cultivation may be conducted in a conventional manner employed in producing antibiotics, preferably in liquid media or under submerged aerial conditions in 40 case of a large scale production. If a change of the medium pH occurs, a buffer such as calcium carbonate may be added to the medium. The cultivation is conducted preferably at 20°-40° C., and particularly at 25°-32° C. The cultivation time is altered fairly accord- 45 ing to the fermentation scale; e.g., about 20–80 hours are required as cultivation time in a large scale production. If vigorous foaming occurs during the cultivation, for example, antifoaming agents such as vegetable oil, lard, and polypropylene glycol may be added suitably before 50 or during the cultivation.

After termination of the cultivation a conventional manner for recovering the fermentation products may be used properly in recovering GT-2558 from the culture broth. For example, filtration, centrifugal separa- 55 tion, adsorption-desorption procedure and chromatography with various ion-exchange resins and other adsorbents, and extraction with various organic solvents may be employed in proper combination preferably. Additionally the acetyl derivative of GT-2558 can be sepa-60 (pH 7.0) containing glycerin (4.0%), meat extract rated as a means for purification. To obtain the acetyl derivative crude powder of GT-2558 is dissolved in acetic acid, the excess amount of acetic anhydride is added thereto, and the mixture may be allowed to react at room temperature under stirring. In this connection, 65 in order to prevent decomposition of GT-2558, if desired, it may be considered to use a suitable stabilizer in the separating step.

Occasionally it is preferred that GT-2558 is changed into the salts for convenience in separating procedure and medical application to human and animals. The acids which can form the salts together with GT-2558 are inorganic acids such as hydrochloric acid, hydrobromic acid, hydroiodic acid, sulfuric acid, nitric acid, phosphoric acid, and carbonic acid, and organic acids such as acetic acid, fumaric acid, malic acid, tartaric acid, maleic acid citric acid, mandelic acid, ascorbic acid, and gallic acid.

As shown in the above, GT-2558 is a mixture of A, B, and other minor components, and so GT-2558 may be separated into each of these components if desired. As a means for separation, e.g., various chromatographic means and gel filtration are employed. Acetylation and formation of salts mentioned above may also be used.

GT-2558, its components and acetyl derivatives, and the pharmaceutically acceptable salts thereof can be used as reagents for determination of β -galactosidase activity, acting as β -galactosidase inhibitors. These compounds together with conventional types of diluents, stabilizers, preservatives, moistening agents, and surface active agents can be administered orally as tablets, capsules, and powders, or parenterally as injections, liniments, and suppositories, and the dosage is changeable according to therapeutic purpose, the age, and conditions of a patient, etc.

GT-2558 and its derivatives can also be used in controlling sweetness of lactose-added foods and drinks in which lactose is used in place of sucrose, for example, dairy products, confectionary, fruit juices. In order to sweeten the foods and drinks by addition of lactose, the lactose added is hydrolyzed with β -galactosidase to give glucose and galactose which afford sweetness to the foods and drinks. It is appropriate, however, to control the sweetness by suppressing hydrolysis of lactose at a pre-fixed stage with addition of a β -galactosidase inhibitor, GT-2558 or its derivatives. Examples 19 and 20 as shown below illustrate more in detail the method of controlling sweetness of milk and dry skim milk.

The following examples serve to illustrate the processes for producing a group of the GT-2558 compounds, but are not intended to limit the scope of the invention.

EXAMPLE 1

Each 100 ml of the medium, (pH 7.0) containing glucose (1.0%), casamino acid (0.2%), yeast extract (0.2%), and meat extract (0.1%) is placed in 500 ml shaking flasks and sterilized at 120° C. for 20 minutes, and the thus obtained media are used for seed culture.

The platinum loop of the slant culture resulting from a strain of Streptomyces lydicus PA-5726, FERM-BP61 is inoculated on the above media for seed culture, and cultured under shaking at 28° C., 140 reciprocations per minute, and 7 cm amplitude for 2 days to give the seed mash.

On the other hand, lard (0.1%) is added to a medium (1.5%), peptone (1.5%), KCl (0.076%), MgCl₂.6H₂O (0.042%), FeCl₃.6H₂O (0.00245%), ZnSO₄.7H₂O (0.00219%), and MnCl₂.4H₂O (0.00181%). This medium (80 ml each) is placed in 500 ml shaking flasks and sterilized at 120° C. for 20 minutes. The thus obtained medium is used as main fermentation medium, and the above seed mash is inoculated on the fermentation medium at 2% content and cultured under shaking at 28°

C., 140 reciprocations per minute, and 7 cm amplitude for 6 days.

In this culture broth the β -galactosidase inhibitor GT-2558 (48,100 IU/mg, IC₅₀=0.0052 mcl) is accumulated.

EXAMPLE 2

In the same manner as in Example 1, main fermentation medium (8 L) is employed in cultivation and immediately after termination of the cultivation the organism 10 is separated by centrifugal separation to yield a clear filtrate (6.3 L).

This filtrate is adjusted at pH 4.5 by addition of hydrochloric acid, and active carbon (1.5%, 94.5 g) is added thereto on which pigments and impurities are 15 absorbed under stirring for 30 minutes, and filtered with assistance of Celite to give a clear filtrate (6.9 L).

This filtrate is passed through 2 L of a strongly basic anion-exchange resin Dowex-1×8 (OH type) (trade name, Dow Chemical Co.) to remove anionic sub- 20 stances. The resultant solution is furthermore passed through a column of 1 L of a strongly acidic cationexchange resin Dowex-50W×8 (H type), on which GT-2558 is adsorbed, washed with water sufficiently, and then eluted with 0.5N hydrochloric acid.

The eluted fractions with the β -galactosidase inhibitory activity are collected and immediately neutralized by removal of excess Cl-ion with a weakly basic anionexchange resin Amberlite IRA-47 (OH type) (trade name, Rohm & Haas Co.) to give a GT-2558 solution 30 (400 ml). The β -galactosidase inhibitory activity of this solution is 454,500 IU/ml (IC₅₀=0.00055 mcl).

EXAMPLE 3

The GT-2558 solution obtained in Example 2 is con- 35 same 70% ethanol as a developer. densed by means of lyophilization to give a yellowish brown oily substance, which is dissolved in a small amount of 80% methanol. The residue is filtered and acetone (10 times in volume) is added to the filtrate to yield the precipitate, from which supernatant solution 40 of acetone is removed by centrifugal separation.

The precipitate is dissolved in a small amount of methanol again and ethyl acetate (5–10 times in volume) is added thereto gradually to yield the precipitate, which is collected by filtration and dried to give pale 45 yellow crude powder of GT-2558 (8,090 mg). The β galactosidase inhibitory activity of this crude powder is 20,000 IU/mg (IC₅₀=0.013 mcg) and in the above procedure this substance is obtained in yield of more than 50%.

EXAMPLE 4

The crude powder (800 mg) obtained in Example 3 is dissolved in a small amount of 80% methanol, applied to a column $(2.60 \times 100 \text{ cm})$ of a weakly basic anion- 55 exchange resin Amberlite IRA-47 (OH type) in 80% methanol, and eluted with the same 80% methanol as a developer. The eluate is collected as 6 g each fractions, the β -galactosidase inhibitory activity is detected in bined and condensed under reduced pressure to give pale yellowish powder (262 mg). The inhibitory activity of this crude powder is 35,600 IU/mg (IC₅₀=0.007 mcg) and the yield is 58.3%.

EXAMPLE 5

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The crude powder (400 mg) obtained in Example 4 is furthermore dissolved in a small amount of 80% metha-

nol, applied to a column $(3.10 \times 130 \text{ cm})$ of Sephadex LH-20 (trade name, Pharmacia Fine Chemical Co.) equilibrated with 80% methanol, and eluted with the same 80% methanol as a developer. The elute is collected as 4 g each fractions, and the β -galactosidase inhibitory activity is detected in Nos. 125-135 fractions. The active fractions are combined and condensed under reduced pressure to give colorless powder (214 mg). The inhibitory activity of this crude powder is 55,000 IU/mg (IC₅₀=0.0045 mcg) and the yield is 82.7%.

EXAMPLE 6

The crude powder (135 mg) obtained in Example 5 is dissolved in a small amount of 0.2N pyridine-acetic acid buffer solution, applied to a column $(20 \times 72 \text{ cm})$ of a strongly acidic cation-exchange resin Dowex-50 W×8 equilibrated sufficiently with the same buffer solution, and eluted with the same buffer solution as a developer. The eluate is collected in every 7 g fractions, and the β -galactosidase inhibitory activity is detected in Nos. 122–138 fractions and in Nos. 141–160 fractions respectively. Named GT-2558-A and B in order of elution, the yields are 37.2% (A) and 18.3% (B). The fractions are condensed under reduced pressure separately to give 25 light yellow powder (37 mg) of the GT-2558-A whose inhibitory activity is 73,500 IU/mg (IC₅₀=0.0034 mcg) and brown powder (54 mg) of the GT-2558-B whose inhibitory activity is 24,500 IU/mg (IC₅₀=0.010 mcg).

EXAMPLE 7

The GT-2558-A powder (57 mg) obtained in Example 6 is dissolved in a small amount of 70% ethanol, applied to a column (2.6 $\emptyset \times$ 95 cm) of Sephadex LH-20 equilibrated with 70% ethanol, and eluted with the

Each fraction (3 g) of the eluate is collected, and the β -galactosidase inhibitory activity is detected in Nos. 80–95 fractions. The active fractions are combined and condensed under reduced pressure to give colorless powder (32.6 mg) whose inhibitory activity as 117,800 IU/mg (IC₅₀=0.0021 mcg). This colorless powder is recrystallized from a small amount of a mixture of solvents (isopropanol-ethanol) repeatedly to give colorless fine crystals (19.4 mg) of the GT-2558-A. The inhibitory activity of this product is 136,700 IU/mg $(IC_{50}=0.0018 \text{ mcg}).$

EXAMPLE 8

The crude powder (500 mg) of GT-2558 (IC₅₀=0.00750 mcg) obtained in Example 4 is dissolved in acetic acid (10 ml) under ice-cooling, allowed to warm up to room temperature under stirring after addition of acetic anhydride (10 ml), and acetylated by further addition of acetic anhydride (10 ml). The reaction is carried out at room temperature for 2 hours and terminated on addition of alcohol. The mixture is condensed under reduced pressure in the presence of toluene to give a yellow oily substance of the actyl-GT-2558. This substance is checked by means of thin layer chromatogra-Nos. 53-90 fractions, and the active fractions are com- 60 phy on a silica gel plate with hexane-chloroformmethanol (30:30:12) as the solvent system, and as the result, the acetyl-GT-2558-A is recognized at 0.40 (R_f value) and the acetyl-GT-2558-B at 0.05 (R_f value).

EXAMPLE 9

The yellow oily substance of the acetyl-GT-2558 obtained in Example 8 is dissolved in a small amount of methanol, applied to a column $(1.70 \times 40 \text{ cm})$ of silica

gel (50 g) equilibrated with hexane-chloroform-methanol (30:30:15) as the solvent system, and eluted with the same solvent system (40 ml) and successively with the solvent system of hexane-chloroform-methanol (30:30:20). Each fraction (5 g) of the eluate is collected, and checked by thin layer chromatography on a silica gel plate, which indicates that the acetyl-GT-2558-A is eluted in Nos. 30-42 fractions and the acetyl-GT-2558-B in Nos. 115-210 fractions. The fractions are combined respectively and condensed under reduced pressure to give the acetyl-GT-2558-A and B as a yellow oily substances respectively.

EXAMPLE 10

The yellow oily substance of the acetyl-GT-2558-A obtained in Example 9 is chromatographed for purification repeatedly on a silica gel column, in which the solvent system of hexane-chloroform-methanol (30:30:6) is employed, to give a transparent colorless 20 oily substance. Thus obtained substance is dissolved in a small amount of methanol and recrystallized from a mixture of benzene and methanol to give colorless fine crystals of the acetyl-GT-2558-A (50.8 mg). The β -galactosidase inhibitory activity of this product is 0.5 IU/mg (IC₅₀=500 mcg).

EXAMPLE 11

The yellow oily substance of the acetyl-GT-2558-B obtained in Example 9 is chromatographed for purification repeatedly on a silica gel column, in which the solvent system of hexane-chloroform-methanol (30:30:20) is employed, and recrystallized from a mixture of ethyl acetate and methanol to give colorless fine 35 crystals (12.7 mg). The β -galactosidase inhibitory activity of this product is 1,640 IU/mg (IC₅₀=0.15 mcg).

Fractional enzyme assay of β -galactosidases (Example 12–18)

EXAMPLE 12

Enzyme solutions are prepared as a mixture of Bovine liver β -galactosidase solution (A) with Aspergillus oryzae β -galactosidase solution (B) at a rate of 4:0, 3:1, 45 2:2, 1:3, and 0:4 (V/V), respectively.

To the substrate 0.01M o-nitrophenyl β -D-galactopyranoside (ONPG) solution (0.5 ml) dissolved in McIlvaine buffer (pH 4.5), the GT-2558-A solution (0.25 ml) dissolved in the said buffer and the above 50 enzyme solution (0.25 ml) are added, and the enzyme reaction is carried out at 37° C. for 15 minutes. Wherein, the concentration of GT-2558-A is 1.9×10^{-4} M. The reaction is stopped by the addition of 1M sodium carbonate (1 ml), the total volume is adjusted to 10 ml by ⁵⁵ addition of water (8 ml), and then the enzyme activity (a) is determined from the measurement of the absorbance at 420 nm. Concurrently the said buffer not containing GT-2558-A is added, the enzyme reaction is 60 conducted, the absorbance is measured, and the enzyme activity (t) is determined. In this connection, the difference between t and a value is the amount of β -galactosidase activity which is inhibited by GT-2558-A (b=t-a). Referring to expression of activity, one unit 65 of activity (1 U) is defined as the activity of the enzyme which hydrolyzes 1 µmole ONPG per minute and liberates o-nitrophenol. In this way the amounts of Bovine

liver β -galactosidase activity (a) and Aspergillus oryzae β -galactosidase activity (b) are determined fractionally.

In Table 1 the results show good accordance between the practical values determined from enzymatic assay and the theoretical ones determined by calculation.

EXAMPLE 13

Enzyme A: Bovine liver β -galactosidase

Enzyme B: Rat liver β -galactosidase

Substrate: 0.01M ONPG

Inhibitor: 1.9×10^{-5} M GT-2558-A

Condition (employed in the reaction): McIlvaine

buffer (pH 4.5), 37° C., 15 minutes

Method and the rest are the same as in Example 12. Results are shown in Table 2.

EXAMPLE 14

Enzyme A: Bovine liver β -galactosidase

Enzyme B: Rat Kidney β-galactosidase

Substrate: 0.01M ONPG

Inhibitor: 1.9×10^{-4} M GT-2558-A

Condition: McIlvaine buffer (pH 4.5), 37° C., 15 min-

utes

Method and the rest are the same as in Example 12. Results are shown in Table 3.

EXAMPLE 15

Enzyme A: Bovine liver β -galactosidase

Enzyme B: Rat spleen β-galactosidase

Substrate: 0.01M ONPG

Inhibitor: 1.9×10^{-5} M GT-2558-A

Condition: McIlvaine buffer (pH 4.5), 37° C., 15 min-

utes

Method and the rest are the same as in Example 12. Results are shown in Table 4.

EXAMPLE 16

Enzyme A: Bovine liver β -galactosidase

Enzyme B: Aspergillus oryzae β-galactosidase

Substrate: 0.01M ONPG

Inhibitor: 1.9×10^{-5} M GT-2558-A

Condition: McIlvaine buffer (pH 7.2), 37° C., 15 min-

utes

Method and the rest are the same as in Example 12. Results are shown in Table 5.

EXAMPLE 17

Enzyme A: Rat Kidney \(\beta\)-galactosidase

Enzyme B: Aspergillus oryzae β-galactosidase Sub-

strate: 0.01M ONPG

Inhibitor: 1.9×10^{-6} M GT-2558-A

Condition: McIlvaine buffer (pH 7.2), 37° C., 15 min-

utes.

Method and the rest are the same as in Example 12. Results are shown in Table 6.

EXAMPLE 18

Enzyme A: Bovine liver β -galactosidase

Enzyme B: E. $coli \beta$ -galactosidase

Substrate: 0.01M ONPG

Inhibitor: 4.8×10^{-5} M GT-2558-A

Condition: McIlvaine buffer (pH 7.2), 37° C., 15 minutes

Method and the rest are the same as in Example 12. Results are shown in Table 7.

TABLE

	Origin of		Concentration				: Practica	al value cal value	·
Table No.	Mixed β- Galactosidase	pH at	of GT-2558-A (M)	Enzyme activity	4/0* (U/ml)	3/1 (U/ml)	2/2 (U/ml)	1/3 (U/ml)	0/4 (U/ml)
1	A Bovine Liver	4.5	1.9×10^{-4}	t	0.32	0.28	0.25	0.22	0.19
	B Aspergillus				0.32	0.28	0.25 0.08	0.22	0.19 0.19
	oryzae			a	0.00 0.00	0.04 0.05	0.08	0.14	0.19
				ь	0.32	0.03	0.17	0.08	0.00
				Ü	0.32	0.23	0.16	0.08	0.00
2	A Bovine Liver	4.5	1.9×10^{-5}	t	0.16	0.18	0.20	0.22	0.24
	B Rat Liver				0.16	0.18	0.20	0.22	0.24
				a	0.01	0.07	0.13	0.19	0.24
				•	0.00	0.06	0.12	0.18	0.24
			•	b	0.15	0.11	0.07	0.04 0.04	0.00
2	A Douine Liver	4.5	1.9×10^{-4}	t	0.16 0.35	0.12 0.32	0.08 0.29	0.04	0.25
3	A Bovine Liver B Rat Kidney	4.5	1.9 × 10	ι	0.35	0.32	0.30	0.27	0.25
	D Rat Ridney			a	0.01	0.08	0.12	0.18	0.24
				-	0.00	0.06	0.12	0.19	0.25
				ь	0.34	0.24	0.17	0.08	0.01
			_		0.35	0.27	0.18	0.08	0.00
4	A Bovine Liver	4.5	1.9×10^{-5}	t	0.39	0.35	0.31	0.26	0.24
	B Rat Spleen				0.39	0.36	0.32	0.28	0.24
				a	0.01	0.07	0.11	0.17	0.24
				ь	0.00 0.38	0.06 0.28	0.12 0.20	0.18 0.09	0.24 0.00
				U	0.39	0.30	0.20	0.10	0.00
5	A Bovine Liver	7.2	1.9×10^{-5}	t	0.30	0.28	0.27	0.26	0.26
5	B Aspergillus	,	1.7 / 10	-	0.30	0.29	0.28	0.27	0.26
	oryzae			a .	0.00	0.06	0.12	0.19	0.25
	-				0.00	0.06	0.13	0.20	0.26
				b	0.30	0.22	0.15	0.07	0.01
_					0.30	0.23	0.15	0.07	0.00
6	A Rat Kidney	7.2	1.9×10^{-6}	t	0.33	0.32	0.32	0.31	0.30
	B Aspergillus				0.33 0.01	0.33 0.09	0.32 0.15	0.31 0.22	0.30 0.29
	oryzae			a	0.00	0.09	0.15	0.22	0.30
				ь	0.32	0.23	0.17	0.09	0.01
				-	0.33	0.25	0.17	0.08	0.00
7	A Bovine ₂ Liver	7.2	4.8×10^{31} 5	t	0.21	0.23	0.25	0.27	0.29
	B E. coli		•		0.21	0.23	0.25	0.27	0.29
	•			a	0.01	0.07	0.14	0.21	0.29
					0.00	0.07	0.15	0.22	0.29
				ь	0.20	0.16	0.11	0.06	0.00
					0.21	0.16	0.10	0.05	0.00

Notes

EXAMPLE 19

Cow's milk (1000 ml) was slowly stirred at 30° C., to which 500 mg of β -galactosidase (produced by Aspergillus oryzae) (lactose hydrolyzing potency: 20 U/mg) was added. A portion (100 ml each) of the mixture was collected at pre-determined intervals, and GT-2558-A 55 (2 mg) was added thereto to stop the enzymatic reaction to give a partially lactose-hydrolyzed milk. Table 8 indicates the rate (%) of hydrolysis of lactose and the result of sweetness test by testers A-D. As seen from the table, the sweetness of milk can freely be controlled 60 by addition of the inhibitor at an appropriate time.

TABLE 8									
Time (min)	·		20	30	60	120	180	240	
Rate (%) of Hydrolysis	15.4	24.8	32.3	48.6	67.0	79.9	90.1	65	
Tester**	A	_	_	±		++	+++	+++	-
	B C	± ±	_	+ ±	+	+ + +	+++++++++++++++++++++++++++++++++++++++	+++	

TABLE 8-continued

D	<u>±</u>	±	+	+	++	+++	+++
Rate (%) of Hydrolysis*	15.4	24.8	32.3	48.6	67.0	79.9	90.1
Time (min)	10	20	30	60	120	180	240

- *The rate of hydrolysis of lactose was determined by conventional quantitative analysis with glucose oxidase on glucose released from lactose.
- **Result of the test was represented by the following 5 degrees compared with the taste of milk.
- -: no change appreciable
- ±: slightly sweet

50

- +: sweet and agreeable ++: sweet but disagreeable
- +++: too sweet and unacceptable

EXAMPLE 20

Preparation of Sweetening Agent from Skim Milk:

Dry skim milk (200 g) was suspended in water so as to give 20% suspension (1L), which was slowly stirred at 30° C. β-Galactosidase (1 g) (produced by Aspergillus oryzae) was added thereto, and a portion of the mixture (100 ml each) was collected at predetermined intervals and treated with 4 mg GT-2558-A. Table 9 indicates the

A: the enzyme which is not completely inhibited GT-2558-A of the shown concentrations

B: the enzyme which is completely inhibited by GT-2558-A of the shown concentrations

t: the β -galactosidase activity measured in the absence of GT-2558-A (total activity)

a: the β -galactosidase activity measured in the presence of GT-2558-A (the enzyme activity of A)

b: b = t - a, the difference between t and a, the amount of the enzyme activity inhibited by GT-2558-A (the enzyme activity of B)

^{*:} B/A, the mixture rate of the enzyme A and the enzyme B

rate (%) of hydrolysis of lactose and the result of sweetness test by tasters A-D. As shown in Table 9, skim milk can be converted into sweetening agents having appropriate sweetness.

TABLE 9									
Time (nr) Rate (76) of		0.5	1	1.5	2	3	4		
Hydrolysis*		27.4	39.2	50.1	59.2	70.6	78.9	·.	
Taster**	A B C D	- + + +	+ + +	+ + ± +	++++	++++	++++	10	

*Determined in the same manner as in Example 19.

- **Represented by the following 4 degrees compared with the taste of 20% skim milk suspension.
- : no change appreciable
- 士: slightly sweet
- + sweet
- --: very sweet

What is claimed is:

- 1. GT-2558-A with the following properties or a ²⁰ pharmaceutically acceptable salt thereof:
 - a. m.p.

106°-108° C.

- b. Molecular weight (mass spectrum)
 Maximum mass m/e=302
- c. Elemental analysis

Found: C,41.34; H,6.16; N,7.43; O,45.06 (%)

d. IR

 ν_{max}^{KBr} 3400, 2900, 1640, 1565, 1505, 1455, 1415, 1360, 1300, 1270, 1160, 1110, 1075, 1035, 955, 30 930, 890, 855, 795 cm⁻¹

e. NMR

FIG. 1 and FIG. 2

f. β -Galactosidase inhibitory activity

Enzyme	pH at	Origin	IC ₅₀	
Taka β- galactosidase	5.0	Aspergillus oryzae(Sankyo)	0.0018 mc	g
Taka β- galactosidase	4.5	Aspergillus oryzae(Sankyo)	0.0045	4
Galantase	4.5	Aspergillus oryzae(Tokyo Tanabe)	0.0045	
Bovine β- galactosidase	7.2	Bovine liver (Sigma)	26.6	4

- g. Specific β-galactosidase inhibitory activity 136,700 IU/mg (IC₅₀ 0.0018 mcg).
- 2. Acetyl-GT-2558-A with the following properties or a pharmaceutically acceptable salt thereof:
 - a. Nature

Colorless fine crystals (recrystallized from benzene-methanol)

b. Solubility

Soluble in methanol, ethanol, chloroform, dioxane, 55 and water, slightly soluble in ethyl acetate, and insoluble or hardly soluble in carbon tetrachloride and benzene

c. Color reaction

Ninhydrin reaction test positive

Ammoniac silver nitrate reaction test positive

d. m.p. and decom.p.

m.p. 41° C.; decomp.p. 149°-153° C.

e. Specific rotation

 $[\alpha]_D^{24} + 29.6^{\circ}$ (c 1.0 methanol)

f. Molecular weight (mass spectrum)
Maximum mass m/e=491 (M+1)

g. Elemental analysis

Found: C,51.30; H,6.27; N,5.65; O,36.78 (%)

h. UV

No characteristic absorption

i. IR

15

35

 ν_{max}^{KBr} 3430, 2920, 1745, 1630, 1435, 1390, 1370, 1245, 1090, 1040, 820, 760 cm⁻¹

j. NMR

FIG. 4

- k. Specific β-galactosidase inhibitory activity 0.5 IU/mg (IC₅₀ 500 mcg).
- 3. Acetyl-GT-2558-B with the following properties or a pharmaceutically acceptable salt thereof:
 - a. Nature

Colorless fine crystals (recrystallized from ethyl acetate-methanol)

b. Solubility

Soluble in methanol, ethanol, and water, slightly soluble in ethyl acetate and chloroform, and insoluble or hardly soluble in carbon tetrachloride and benzene

c. Color reaction

Ninhydrin reaction test positive

Ammoniac silver nitrate reaction test positive

d. m.p. and decom.p.

m.p. 76°-78° C; decomp.p. 161°-163° C.

e. Specific rotation

 $[\alpha]_D^{24} + 13.8^{\circ}$ (c 1.0 methanol)

f. Molecular weight (mass spectrum) Maximum mass m/e=429 (M+1)

g. Elemental analysis

Found: C,46.58; H,60.58; N,6.56;038.51 (%)

h. UV

No characteristic absorption

i. IR

 ν_{max}^{KBr} 3410, 2930, 2735, 2360, 1640, 1550, 1435, 1380, 1245, 1195, 1115, 1090, 1045, 875, 815 cm⁻¹

j. NMR

FIG. 3

- k. Specific β-galactosidase inhibitory activity 1,640 IU/mg (IC₅₀ 0.15 mcg).
- 4. Method for producing GT-2558-A as defined in claim 1, which comprises culturing a GT-2558-A producing strain of *Streptomyces lydicus* PA-5726, FERM BP-61 on a culture medium, under aerobic conditions at a temperature of 20° to 40° C. for about 20 to 80 hours and recovering GT-2558-A from the culture broth.