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(54) **SEMI-SYNTHETIC TEREPHTHALIC ACID
VIA MICROORGANISMS THAT PRODUCE
MUCONIC ACID**

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(57) **ABSTRACT**

The invention provides a non-naturally occurring microbial organism having a muconate pathway having at least one exogenous nucleic acid encoding a muconate pathway enzyme expressed in a sufficient amount to produce muconate. The muconate pathway including an enzyme selected from the group consisting of a beta-ketothiolase, a beta-ketoadipyl-CoA hydrolase, a beta-ketoadipyl-CoA transferase, a beta-ketoadipyl-CoA ligase, a 2-fumarylacetate reductase, a 2-fumarylacetate dehydrogenase, a trans-3-hydroxy-4-hexendioate dehydratase, a 2-fumarylacetate aminotransferase, a 2-fumarylacetate aminating oxidoreductase, a trans-3-amino-4-hexenoate deaminase, a beta-ketoadipate enol-lactone hydrolase, a muconolactone isomerase, a muconate cycloisomerase, a beta-ketoadipyl-CoA dehydrogenase, a 3-hydroxyadipyl-CoA dehydratase, a 2,3-dehydroadipyl-CoA transferase, a 2,3-dehydroadipyl-CoA hydrolase, a 2,3-dehydroadipyl-CoA ligase, a muconate reductase, a 2-maleylacetate reductase, a 2-maleylacetate dehydrogenase, a cis-3-hydroxy-4-hexendioate dehydratase, a 2-maleylacetate aminoatransferase, a 2-maleylacetate aminating oxidoreductase, a cis-3-amino-4-hexendioate deaminase, and a muconate cis/trans isomerase. Other muconate pathway enzymes also are provided. Additionally provided are methods of producing muconate.

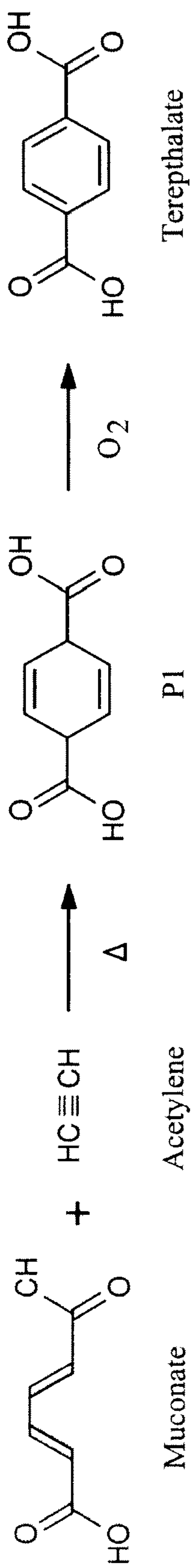


FIG. 1

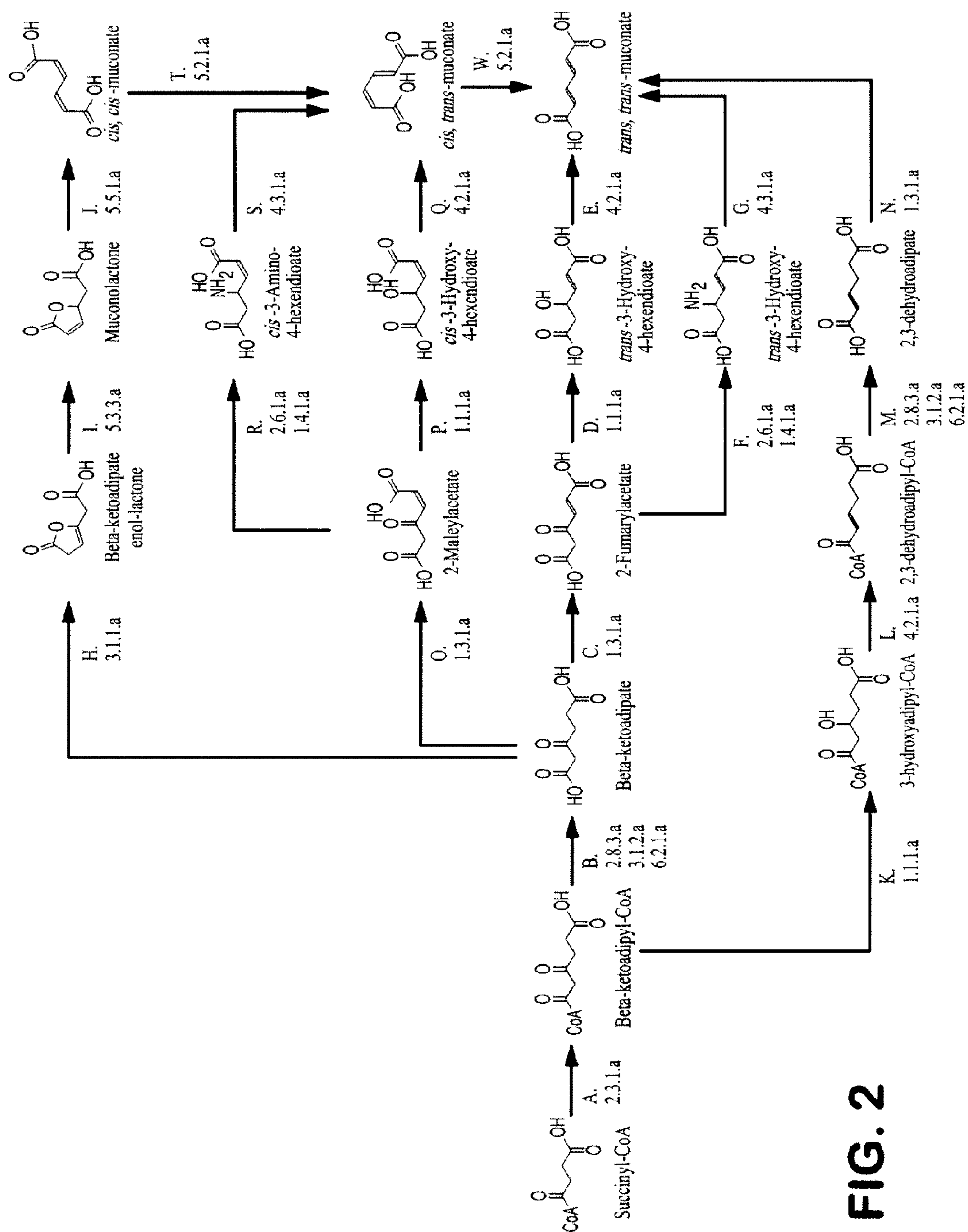


FIG. 2

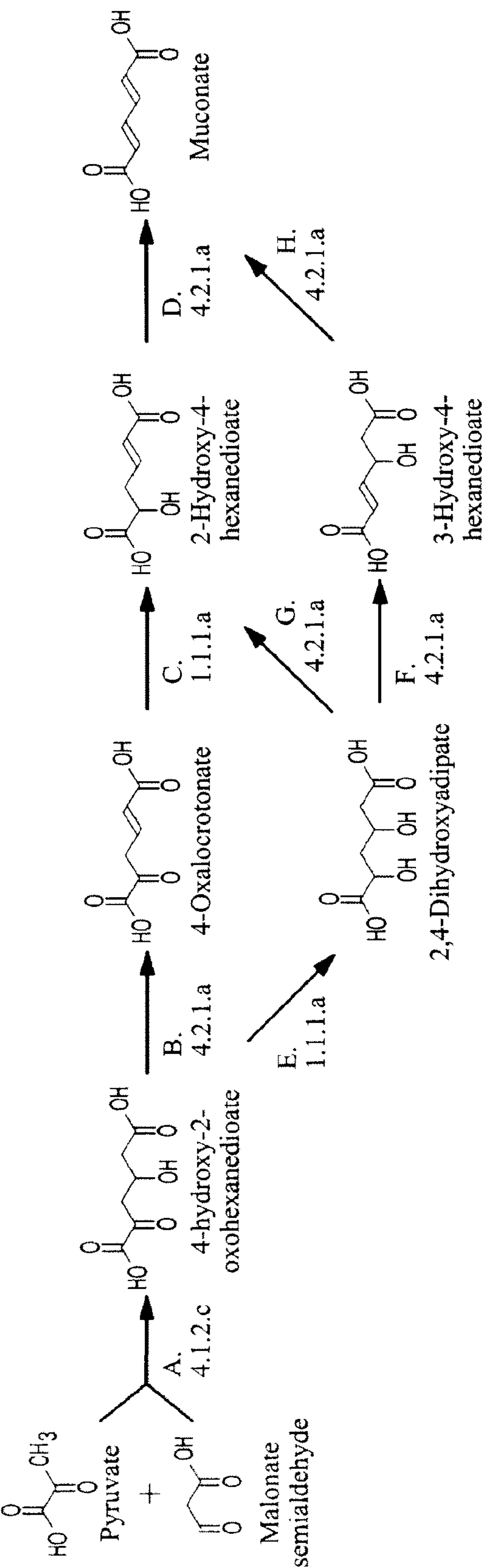


FIG. 3

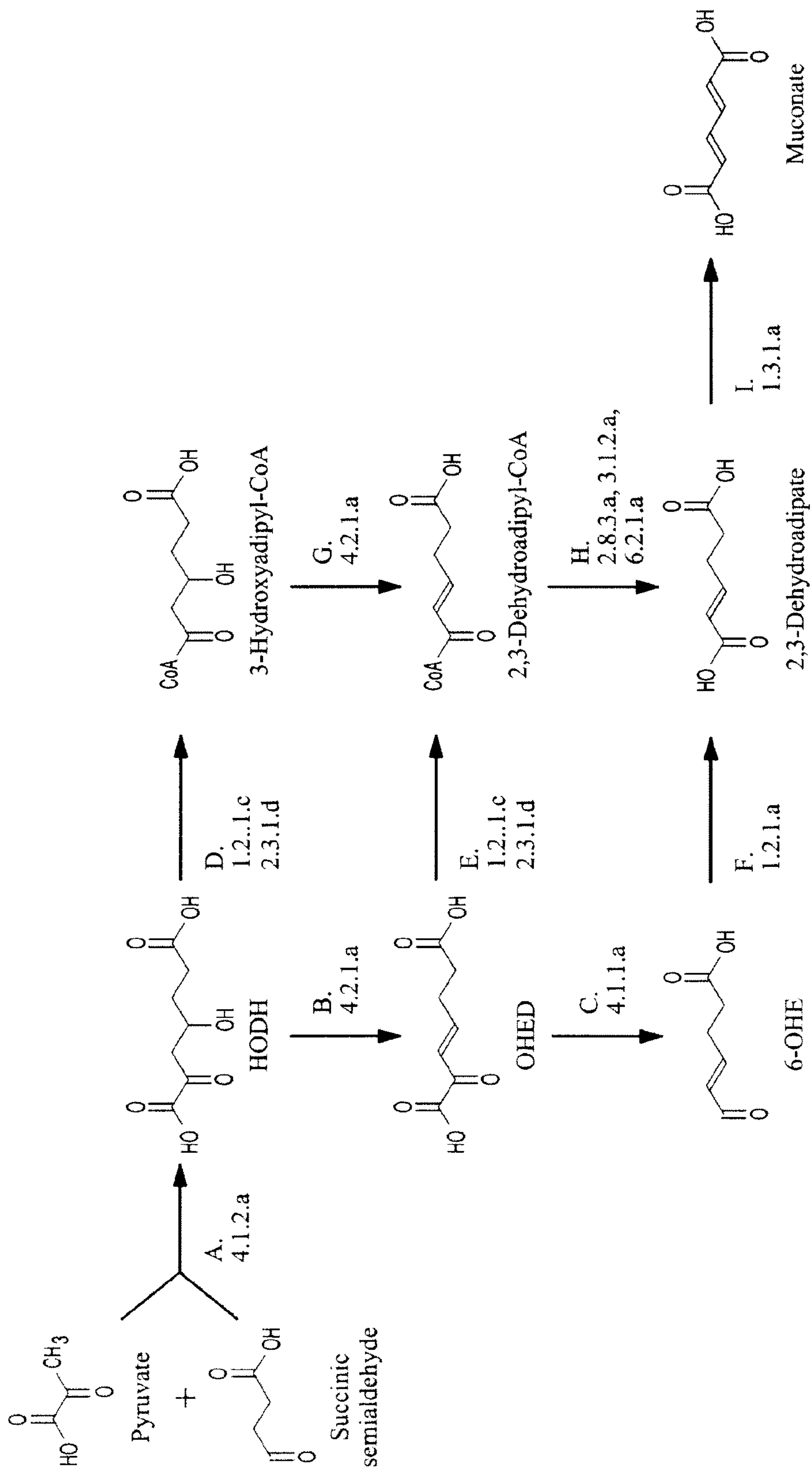


FIG. 4

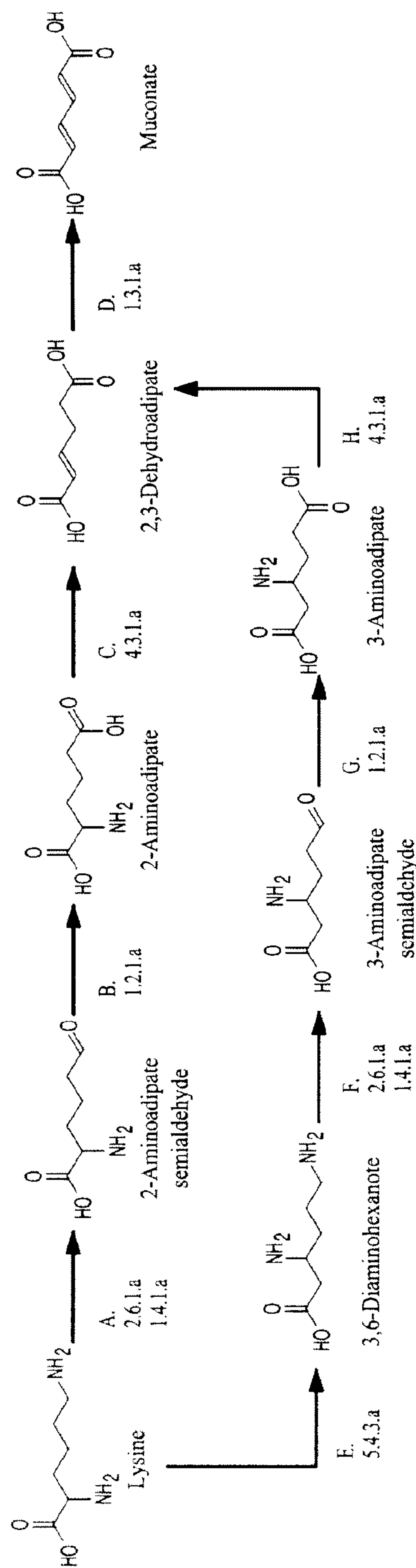


FIG. 5

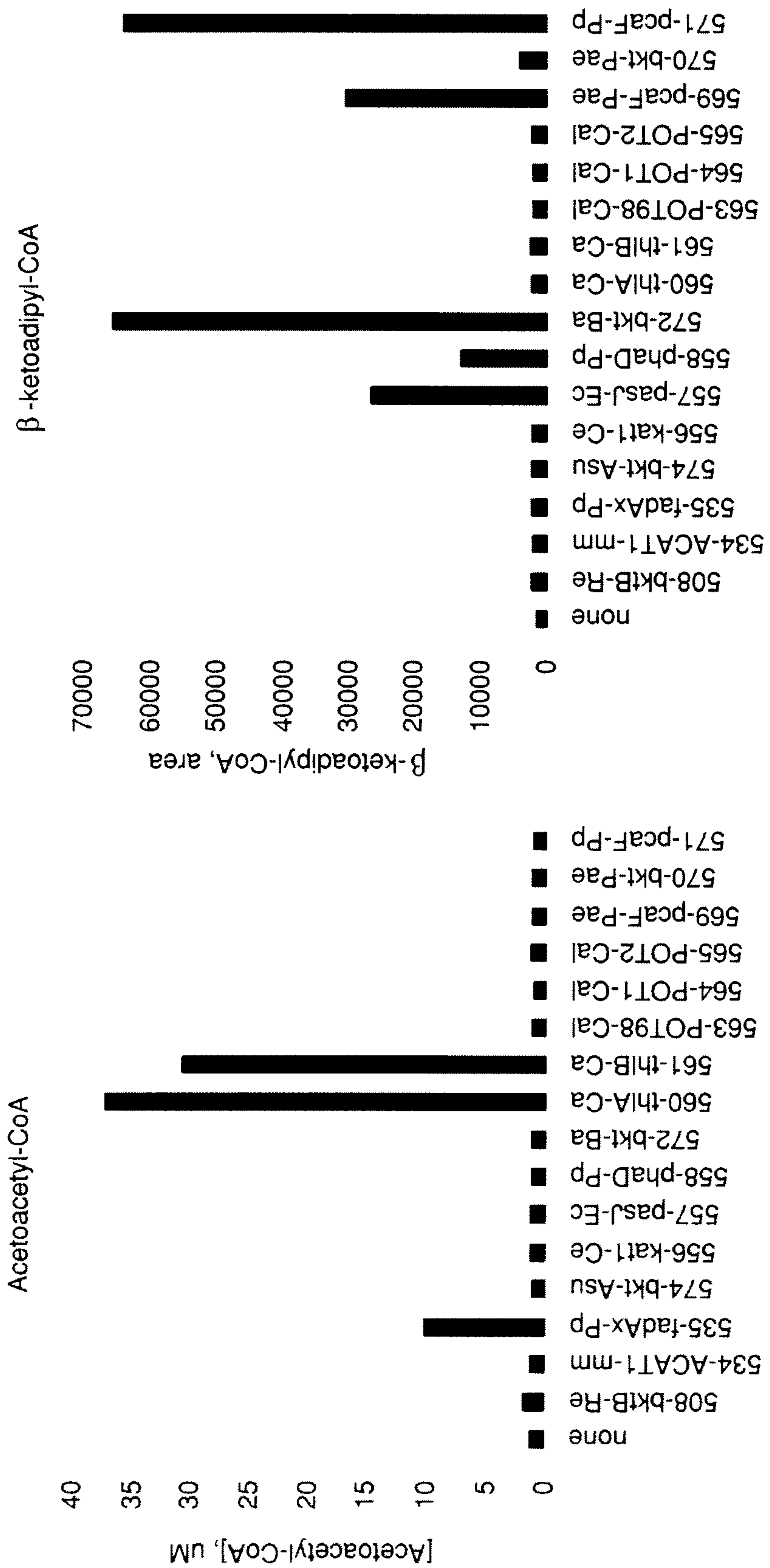


FIG. 6

SEMI-SYNTHETIC TEREPHTHALIC ACID VIA MICROORGANISMS THAT PRODUCE MUCONIC ACID

[0001] This application claims the benefit of priority of U.S. Provisional Application No. 61/231,637, filed Aug. 5, 2009, the entire contents of which are incorporated herein by this reference.

BACKGROUND OF THE INVENTION

[0002] The present disclosure relates generally to the design of engineered organisms and, more specifically to organisms having selected genotypes for the production of muconic acid.

[0003] Terephthalate (also known as terephthalic acid and PTA) is the immediate precursor of polyethylene terephthalate (PET), used to make clothing, resins, plastic bottles and even as a poultry feed additive. Nearly all PTA is produced from para-xylene by the oxidation in air in a process known as the Mid Century Process (Roffia et al., *Ind. Eng. Chem. Prod. Res. Dev.* 23:629-634 (1984)). This oxidation is conducted at high temperature in an acetic acid solvent with a catalyst composed of cobalt and/or manganese salts. Para-xylene is derived from petrochemical sources, and is formed by high severity catalytic reforming of naphtha. Xylene is also obtained from the pyrolysis gasoline stream in a naphtha steam cracker and by toluene disproportionation.

[0004] PTA, toluene and other aromatic precursors are naturally degraded by some bacteria. However, these degradation pathways typically involve monooxygenases that operate irreversibly in the degradative direction. Hence, biosynthetic pathways for PTA are severely limited by the properties of known enzymes to date.

[0005] Muconate (also known as muconic acid, MA) is an unsaturated dicarboxylic acid used as a raw material for resins, pharmaceuticals and agrochemicals. Muconate can be converted to adipic acid, a precursor of Nylon-6,6, via hydrogenation (Draths and Frost, *J. Am. Chem. Soc.* 116: 399-400 (1994)). Alternately, muconate can be hydrogenated using biometallic nanocatalysts (Thomas et al., *Chem. Commun.* 10:1126-1127 (2003)).

[0006] Muconate is a common degradation product of diverse aromatic compounds in microbes. Several biocatalytic strategies for making cis,cis-muconate have been developed. Engineered *E. coli* strains producing muconate from glucose via shikimate pathway enzymes have been developed in the Frost lab (U.S. Pat. No. 5,487,987 (1996); Niu et al., *Biotechnol. Prog.*, 18:201-211 (2002)). These strains are able to produce 36.8 g/L of cis,cis-muconate after 48 hours of culturing under fed-batch fermenter conditions (22% of the maximum theoretical yield from glucose). Muconate has also been produced biocatalytically from aromatic starting materials such as toluene, benzoic acid and catechol. Strains producing muconate from benzoate achieved titers of 13.5 g/L and productivity of 5.5 g/L/hr (Choi et al., *J. Ferment. Bioeng.* 84:70-76 (1997)). Muconate has also been generated from the effluents of a styrene monomer production plant (Wu et al., *Enzyme and Microbiology Technology* 35:598-604 (2004)).

[0007] All biocatalytic pathways identified to date proceed through enzymes in the shikimate pathway, or degradation enzymes from catechol. Consequently, they are limited to producing the cis, cis isomer of muconate, since these pathways involve ring-opening chemistry. The development of

pathways for producing trans,trans-muconate and cis,trans-muconate would be useful because these isomers can serve as direct synthetic intermediates for producing renewable PTA via the inverse electron demand Diels-Alder reaction with acetylene. The present invention satisfies this need and provides related advantages as well.

SUMMARY OF INVENTION

[0008] The invention provides a non-naturally occurring microbial organism having a muconate pathway having at least one exogenous nucleic acid encoding a muconate pathway enzyme expressed in a sufficient amount to produce muconate. The muconate pathway including an enzyme selected from the group consisting of a beta-ketothiolase, a beta-ketoadipyl-CoA hydrolase, a beta-ketoadipyl-CoA transferase, a beta-ketoadipyl-CoA ligase, a 2-fumarylacetate reductase, a 2-fumarylacetate dehydrogenase, a trans-3-hydroxy-4-hexendioate dehydratase, a 2-fumarylacetate aminotransferase, a 2-fumarylacetate aminating oxidoreductase, a trans-3-amino-4-hexenoate deaminase, a beta-ketoadipate enol-lactone hydrolase, a muconolactone isomerase, a muconate cycloisomerase, a beta-ketoadipyl-CoA dehydrogenase, a 3-hydroxyadipyl-CoA dehydratase, a 2,3-dehydroadipyl-CoA transferase, a 2,3-dehydroadipyl-CoA hydrolase, a 2,3-dehydroadipyl-CoA ligase, a muconate reductase, a 2-maleylacetate reductase, a 2-maleylacetate dehydrogenase, a cis-3-hydroxy-4-hexendioate dehydratase, a 2-maleylacetate aminoatransferase, a 2-maleylacetate aminating oxidoreductase, a cis-3-amino-4-hexendioate deaminase, and a muconate cis/trans isomerase. Other muconate pathway enzymes also are provided. Additionally provided are methods of producing muconate.

BRIEF DESCRIPTION OF THE DRAWINGS

[0009] FIG. 1 shows the synthesis of terephthalate from muconate and acetylene via Diels-Alder chemistry. P1 is cyclohexa-2,5-diene-1,4-dicarboxylate.

[0010] FIG. 2 shows pathways to trans-trans muconate from succinyl CoA. Enzymes are A) beta-ketothiolase, B) beta-ketoadipyl-CoA hydrolase, transferase and/or ligase, C) 2-fumarylacetate reductase, D) 2-fumarylacetate dehydrogenase, E) trans-3-hydroxy-4-hexendioate dehydratase, F) 2-fumarylacetate aminotransferase and/or 2-fumarylacetate aminating oxidoreductase, G) trans-3-amino-4-hexenoate deaminase, H) beta-ketoadipate enol-lactone hydrolase, I) muconolactone isomerase, J) muconate cycloisomerase, K) beta-ketoadipyl-CoA dehydrogenase, L) 3-hydroxyadipyl-CoA dehydratase, M) 2,3-dehydroadipyl-CoA transferase, hydrolase or ligase, N) muconate reductase, O) 2-maleylacetate reductase, P) 2-maleylacetate dehydrogenase, Q) cis-3-hydroxy-4-hexendioate dehydratase, R) 2-maleylacetate aminotransferase and/or 2-maleylacetate aminating oxidoreductase, S) cis-3-amino-4-hexendioate deaminase, T) muconate cis/trans isomerase, W) muconate cis/trans isomerase.

[0011] FIG. 3 shows pathways to muconate from pyruvate and malonate semialdehyde. Enzymes are A) 4-hydroxy-2-ketovalerate aldolase, B) 2-oxopentenoate hydratase, C) 4-oxalocrotonate dehydrogenase, D) 2-hydroxy-4-hexendioate dehydratase, E) 4-hydroxy-2-oxohexanedioate oxidoreductase, F) 2,4-dihydroxyadipate dehydratase (acting on

2-hydroxy), G) 2,4-dihydroxyadipate dehydratase (acting on 4-hydroxyl group) and H) 3-hydroxy-4-hexenedioate dehydratase.

[0012] FIG. 4 shows pathways to muconate from pyruvate and succinic semialdehyde. Enzymes are A) HODH aldolase, B) OHED hydratase, C) OHED decarboxylase, D) HODH formate-lyase and/or HODH dehydrogenase, E) OHED formate-lyase and/or OHED dehydrogenase, F) 6-OHE dehydrogenase, G) 3-hydroxyadipyl-CoA dehydratase, H) 2,3-dehydroadipyl-CoA hydrolase, transferase or ligase, I) muconate reductase. Abbreviations are: HODH=4-hydroxy-2-oxoheptane-1,7-dioate, OHED=2-oxohept-4-ene-1,7-dioate, 6-OHE=6-oxo-2,3-dehydrohexanoate.

[0013] FIG. 5 shows pathways to muconate from lysine. Enzymes are A) lysine aminotransferase and/or aminating oxidoreductase, B) 2-aminoadipate semialdehyde dehydrogenase, C) 2-aminoadipate deaminase, D) muconate reductase, E) lysine-2,3-aminomutase, F) 3,6-diaminohexanoate aminotransferase and/or aminating oxidoreductase, G) 3-aminoadipate semialdehyde dehydrogenase, H) 3-aminoadipate deaminase.

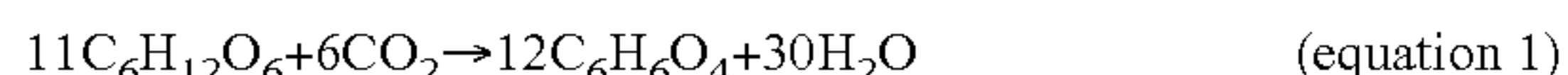
[0014] FIG. 6 shows 3 thiolases with demonstrated thiolase activity resulting in acetoacetyl-CoA formation (left panel). FIG. 6 also shows that several enzymes demonstrated selective condensation of succinyl-CoA and acetyl-CoA to form β -ketoadipyl-CoA as the sole product (right panel).

DETAILED DESCRIPTION OF THE INVENTION

[0015] The present invention is directed, in part, to a biosynthetic pathway for synthesizing muconate from simple carbohydrate feedstocks, which in turn provides a viable synthetic route to PTA. In particular, pathways disclosed herein provide trans,trans-muconate or cis,trans-muconate biocatalytically from simple sugars. The all trans or cis,trans isomer of muconate is then converted to PTA in a two step process via inverse electron demand Diels-Alder reaction with acetylene followed by oxidation in air or oxygen. The Diels-Alder reaction between muconate and acetylene proceeds to form cyclohexa-2,5-diene-1,4-dicarboxylate (P1), as shown in FIG. 1. Subsequent exposure to air or oxygen rapidly converts P1 to PTA.

[0016] This invention is also directed, in part, to non-naturally occurring microorganisms that express genes encoding enzymes that catalyze muconate production. Pathways for the production of muconate disclosed herein are derived from central metabolic precursors. Successfully engineering these pathways entails identifying an appropriate set of enzymes with sufficient activity and specificity, cloning their corresponding genes into a production host, optimizing the expression of these genes in the production host, optimizing fermentation conditions, and assaying for product formation following fermentation.

[0017] The maximum theoretical yield of muconic acid is 1.09 moles per mole glucose utilized. Achieving this yield involves assimilation of CO₂ as shown in equation 1 below:



[0018] As used herein, the term “muconate” is used interchangeably with muconic acid. Muconate is also used to refer to any of the possible isomeric forms: trans,trans, cis,trans, and cis,cis. However, the present invention provides pathways to the useful trans,trans and cis,trans forms, in particular.

[0019] As used herein, the term “non-naturally occurring” when used in reference to a microbial organism or microorganism of the invention is intended to mean that the microbial organism has at least one genetic alteration not normally found in a naturally occurring strain of the referenced species, including wild-type strains of the referenced species. Genetic alterations include, for example, modifications introducing expressible nucleic acids encoding metabolic polypeptides, other nucleic acid additions, nucleic acid deletions and/or other functional disruption of the microbial genetic material. Such modifications include, for example, coding regions and functional fragments thereof, for heterologous, homologous or both heterologous and homologous polypeptides for the referenced species. Additional modifications include, for example, non-coding regulatory regions in which the modifications alter expression of a gene or operon. Exemplary metabolic polypeptides include enzymes or proteins within a muconate biosynthetic pathway.

[0020] A metabolic modification refers to a biochemical reaction that is altered from its naturally occurring state. Therefore, non-naturally occurring microorganisms can have genetic modifications to nucleic acids encoding metabolic polypeptides or, functional fragments thereof. Exemplary metabolic modifications are disclosed herein.

[0021] As used herein, the term “isolated” when used in reference to a microbial organism is intended to mean an organism that is substantially free of at least one component as the referenced microbial organism is found in nature. The term includes a microbial organism that is removed from some or all components as it is found in its natural environment. The term also includes a microbial organism that is removed from some or all components as the microbial organism is found in non-naturally occurring environments. Therefore, an isolated microbial organism is partly or completely separated from other substances as it is found in nature or as it is grown, stored or subsisted in non-naturally occurring environments. Specific examples of isolated microbial organisms include partially pure microbes, substantially pure microbes and microbes cultured in a medium that is non-naturally occurring.

[0022] As used herein, the terms “microbial,” “microbial organism” or “microorganism” is intended to mean any organism that exists as a microscopic cell that is included within the domains of archaea, bacteria or eukarya. Therefore, the term is intended to encompass prokaryotic or eukaryotic cells or organisms having a microscopic size and includes bacteria, archaea and eubacteria of all species as well as eukaryotic microorganisms such as yeast and fungi. The term also includes cell cultures of any species that can be cultured for the production of a biochemical.

[0023] As used herein, the term “CoA” or “coenzyme A” is intended to mean an organic cofactor or prosthetic group (nonprotein portion of an enzyme) whose presence is required for the activity of many enzymes (the apoenzyme) to form an active enzyme system. Coenzyme A functions in certain condensing enzymes, acts in acetyl or other acyl group transfer and in fatty acid synthesis and oxidation, pyruvate oxidation and in other acetylation.

[0024] As used herein, the term “substantially anaerobic” when used in reference to a culture or growth condition is intended to mean that the amount of oxygen is less than about 10% of saturation for dissolved oxygen in liquid media. The

term also is intended to include sealed chambers of liquid or solid medium maintained with an atmosphere of less than about 1% oxygen.

[0025] “Exogenous” as it is used herein is intended to mean that the referenced molecule or the referenced activity is introduced into the host microbial organism. The molecule can be introduced, for example, by introduction of an encoding nucleic acid into the host genetic material such as by integration into a host chromosome or as non-chromosomal genetic material such as a plasmid. Therefore, the term as it is used in reference to expression of an encoding nucleic acid refers to introduction of the encoding nucleic acid in an expressible form into the microbial organism. When used in reference to a biosynthetic activity, the term refers to an activity that is introduced into the host reference organism. The source can be, for example, a homologous or heterologous encoding nucleic acid that expresses the referenced activity following introduction into the host microbial organism. Therefore, the term “endogenous” refers to a referenced molecule or activity that is present in the host. Similarly, the term when used in reference to expression of an encoding nucleic acid refers to expression of an encoding nucleic acid contained within the microbial organism. The term “heterologous” refers to a molecule or activity derived from a source other than the referenced species whereas “homologous” refers to a molecule or activity derived from the host microbial organism. Accordingly, exogenous expression of an encoding nucleic acid of the invention can utilize either or both a heterologous or homologous encoding nucleic acid.

[0026] The non-naturally occurring microbial organisms of the invention can contain stable genetic alterations, which refers to microorganisms that can be cultured for greater than five generations without loss of the alteration. Generally, stable genetic alterations include modifications that persist greater than 10 generations, particularly stable modifications will persist more than about 25 generations, and more particularly, stable genetic modifications will be greater than 50 generations, including indefinitely.

[0027] Those skilled in the art will understand that the genetic alterations, including metabolic modifications exemplified herein, are described with reference to a suitable host organism such as *E. coli* and their corresponding metabolic reactions or a suitable source organism for desired genetic material such as genes for a desired metabolic pathway. However, given the complete genome sequencing of a wide variety of organisms and the high level of skill in the area of genomics, those skilled in the art will readily be able to apply the teachings and guidance provided herein to essentially all other organisms. For example, the *E. coli* metabolic alterations exemplified herein can readily be applied to other species by incorporating the same or analogous encoding nucleic acid from species other than the referenced species. Such genetic alterations include, for example, genetic alterations of species homologs, in general, and in particular, orthologs, paralogs or nonorthologous gene displacements.

[0028] An ortholog is a gene or genes that are related by vertical descent and are responsible for substantially the same or identical functions in different organisms. For example, mouse epoxide hydrolase and human epoxide hydrolase can be considered orthologs for the biological function of hydrolysis of epoxides. Genes are related by vertical descent when, for example, they share sequence similarity of sufficient amount to indicate they are homologous, or related by evolution from a common ancestor. Genes can also be con-

sidered orthologs if they share three-dimensional structure but not necessarily sequence similarity, of a sufficient amount to indicate that they have evolved from a common ancestor to the extent that the primary sequence similarity is not identifiable. Genes that are orthologous can encode proteins with sequence similarity of about 25% to 100% amino acid sequence identity. Genes encoding proteins sharing an amino acid similarity less than 25% can also be considered to have arisen by vertical descent if their three-dimensional structure also shows similarities. Members of the serine protease family of enzymes, including tissue plasminogen activator and elastase, are considered to have arisen by vertical descent from a common ancestor.

[0029] Orthologs include genes or their encoded gene products that through, for example, evolution, have diverged in structure or overall activity. For example, where one species encodes a gene product exhibiting two functions and where such functions have been separated into distinct genes in a second species, the three genes and their corresponding products are considered to be orthologs. For the production of a biochemical product, those skilled in the art will understand that the orthologous gene harboring the metabolic activity to be introduced or disrupted is to be chosen for construction of the non-naturally occurring microorganism. An example of orthologs exhibiting separable activities is where distinct activities have been separated into distinct gene products between two or more species or within a single species. A specific example is the separation of elastase proteolysis and plasminogen proteolysis, two types of serine protease activity, into distinct molecules as plasminogen activator and elastase. A second example is the separation of mycoplasma 5'-3' exonuclease and *Drosophila* DNA polymerase III activity. The DNA polymerase from the first species can be considered an ortholog to either or both of the exonuclease or the polymerase from the second species and vice versa.

[0030] In contrast, paralogs are homologs related by, for example, duplication followed by evolutionary divergence and have similar or common, but not identical functions. Paralogs can originate or derive from, for example, the same species or from a different species. For example, microsomal epoxide hydrolase (epoxide hydrolase I) and soluble epoxide hydrolase (epoxide hydrolase II) can be considered paralogs because they represent two distinct enzymes, co-evolved from a common ancestor, that catalyze distinct reactions and have distinct functions in the same species. Paralogs are proteins from the same species with significant sequence similarity to each other suggesting that they are homologous, or related through co-evolution from a common ancestor. Groups of paralogous protein families include HipA homologs, luciferase genes, peptidases, and others.

[0031] A nonorthologous gene displacement is a non-orthologous gene from one species that can substitute for a referenced gene function in a different species. Substitution includes, for example, being able to perform substantially the same or a similar function in the species of origin compared to the referenced function in the different species. Although generally, a nonorthologous gene displacement will be identifiable as structurally related to a known gene encoding the referenced function, less structurally related but functionally similar genes and their corresponding gene products nevertheless will still fall within the meaning of the term as it is used herein. Functional similarity requires, for example, at least some structural similarity in the active site or binding region of a nonorthologous gene product compared to a gene

encoding the function sought to be substituted. Therefore, a nonorthologous gene includes, for example, a paralog or an unrelated gene.

[0032] Therefore, in identifying and constructing the non-naturally occurring microbial organisms of the invention having muconate biosynthetic capability, those skilled in the art will understand with applying the teaching and guidance provided herein to a particular species that the identification of metabolic modifications can include identification and inclusion or inactivation of orthologs. To the extent that paralogs and/or nonorthologous gene displacements are present in the referenced microorganism that encode an enzyme catalyzing a similar or substantially similar metabolic reaction, those skilled in the art also can utilize these evolutionally related genes.

[0033] Orthologs, paralogs and nonorthologous gene displacements can be determined by methods well known to those skilled in the art. For example, inspection of nucleic acid or amino acid sequences for two polypeptides will reveal sequence identity and similarities between the compared sequences. Based on such similarities, one skilled in the art can determine if the similarity is sufficiently high to indicate the proteins are related through evolution from a common ancestor. Algorithms well known to those skilled in the art, such as Align, BLAST, Clustal W and others compare and determine a raw sequence similarity or identity, and also determine the presence or significance of gaps in the sequence which can be assigned a weight or score. Such algorithms also are known in the art and are similarly applicable for determining nucleotide sequence similarity or identity. Parameters for sufficient similarity to determine relatedness are computed based on well known methods for calculating statistical similarity, or the chance of finding a similar match in a random polypeptide, and the significance of the match determined. A computer comparison of two or more sequences can, if desired, also be optimized visually by those skilled in the art. Related gene products or proteins can be expected to have a high similarity, for example, 25% to 100% sequence identity. Proteins that are unrelated can have an identity which is essentially the same as would be expected to occur by chance, if a database of sufficient size is scanned (about 5%). Sequences between 5% and 24% may or may not represent sufficient homology to conclude that the compared sequences are related. Additional statistical analysis to determine the significance of such matches given the size of the data set can be carried out to determine the relevance of these sequences.

[0034] Exemplary parameters for determining relatedness of two or more sequences using the BLAST algorithm, for example, can be as set forth below. Briefly, amino acid sequence alignments can be performed using BLASTP version 2.0.8 (Jan. 5, 1999) and the following parameters: Matrix: 0 BLOSUM62; gap open: 11; gap extension: 1; x_dropoff: 50; expect: 10.0; wordsize: 3; filter: on. Nucleic acid sequence alignments can be performed using BLASTN version 2.0.6 (Sep. 16, 1998) and the following parameters: Match: 1; mismatch: -2; gap open: 5; gap extension: 2; x_dropoff: 50; expect: 10.0; wordsize: 11; filter: off. Those skilled in the art will know what modifications can be made to the above parameters to either increase or decrease the stringency of the comparison, for example, and determine the relatedness of two or more sequences.

[0035] In some embodiments, the invention provides a non-naturally occurring microbial organism having a muconate pathway that includes at least one exogenous nucleic acid

encoding a muconate pathway enzyme expressed in a sufficient amount to produce muconate. The muconate pathway includes an enzyme selected from the group consisting of a beta-ketothiolase, a beta-ketoadipyl-CoA hydrolase, a beta-ketoadipyl-CoA transferase, a beta-ketoadipyl-CoA ligase, a 2-fumarylacetate reductase, a 2-fumarylacetate dehydrogenase, a trans-3-hydroxy-4-hexendioate dehydratase, a 2-fumarylacetate aminotransferase, a 2-fumarylacetate aminating oxidoreductase, a trans-3-amino-4-hexenoate deaminase, a beta-ketoadipate enol-lactone hydrolase, a muconolactone isomerase, a muconate cycloisomerase, a beta-ketoadipyl-CoA dehydrogenase, a 3-hydroxyadipyl-CoA dehydratase, a 2,3-dehydroadipyl-CoA transferase, a 2,3-dehydroadipyl-CoA hydrolase, a 2,3-dehydroadipyl-CoA ligase, a muconate reductase, a 2-maleylacetate reductase, a 2-maleylacetate dehydrogenase, a cis-3-hydroxy-4-hexendioate dehydratase, a 2-maleylacetate aminotransferase, a 2-maleylacetate aminating oxidoreductase, a cis-3-amino-4-hexendioate deaminase, and a muconate cis/trans isomerase.

[0036] In particular embodiments, the muconate pathway includes a set of muconate pathway enzymes shown in FIG. 2 and selected from the group consisting of:

[0037] A) (1) beta-ketothiolase, (2) an enzyme selected from beta-ketoadipyl-CoA hydrolase, beta-ketoadipyl-CoA transferase, and beta-ketoadipyl-CoA ligase, (3) beta-ketoadipate enol-lactone hydrolase, (4) muconolactone isomerase, (5) muconate cycloisomerase, and (6) a muconate cis/trans isomerase;

[0038] B) (1) beta-ketothiolase, (2) an enzyme selected from beta-ketoadipyl-CoA hydrolase, beta-ketoadipyl-CoA transferase and beta-ketoadipyl-CoA ligase, (3) 2-maleylacetate reductase, (4) 2-maleylacetate dehydrogenase, (5) cis-3-hydroxy-4-hexendioate dehydratase, and (6) muconate cis/trans isomerase;

[0039] C) (1) beta-ketothiolase, (2) an enzyme selected from beta-ketoadipyl-CoA hydrolase, beta-ketoadipyl-CoA transferase and beta-ketoadipyl-CoA ligase, (3) 2-maleylacetate reductase, (4) an enzyme selected from 2-maleylacetate aminotransferase and 2-maleylacetate aminating oxidoreductase, (5) cis-3-amino-4-hexenoate deaminase, and (6) muconate cis/trans isomerase;

[0040] D) (1) beta-ketothiolase, (2) beta-ketoadipyl-CoA dehydrogenase, (3) 3-hydroxyadipyl-CoA dehydratase, (4) an enzyme selected from 2,3-dehydroadipyl-CoA transferase, 2,3-dehydroadipyl-CoA hydrolase and 2,3-dehydroadipyl-CoA ligase, and (5) muconate reductase;

[0041] E) (1) beta-ketothiolase, (2) an enzyme selected from beta-ketoadipyl-CoA hydrolase, beta-ketoadipyl-CoA transferase and beta-ketoadipyl-CoA ligase, (3) 2-fumarylacetate reductase, (4) 2-fumarylacetate dehydrogenase, and (5) trans-3-hydroxy-4-hexendioate dehydratase;

[0042] F) (1) beta-ketothiolase, (2) an enzyme selected from beta-ketoadipyl-CoA hydrolase, beta-ketoadipyl-CoA transferase and beta-ketoadipyl-CoA ligase, (3) 2-fumarylacetate reductase, (4) an enzyme selected from 2-fumarylacetate aminotransferase and 2-fumarylacetate aminating oxidoreductase, and (5) trans-3-amino-4-hexenoate deaminase.

[0043] In some embodiments, a microbial organism having a pathway exemplified by those shown in FIG. 2 can include two or more exogenous nucleic acids each encoding a muconate pathway enzyme, including three, four, five, six, that is up to all of the of enzymes in a muconate pathway. The non-naturally occurring microbial organism having at least one exogenous nucleic acid can include a heterologous nucleic

acid. A non-naturally occurring microbial organism having a pathway exemplified by those shown in FIG. 2 can be cultured in a substantially anaerobic culture medium.

[0044] In some embodiments, the invention provides a non-naturally occurring microbial organism having a muconate pathway that includes at least one exogenous nucleic acid encoding a muconate pathway enzyme expressed in a sufficient amount to produce muconate. The muconate pathway includes an enzyme selected from the group consisting of a 4-hydroxy-2-ketovalerate aldolase, a 2-oxopentenoate hydratase, a 4-oxalocrotonate dehydrogenase, a 2-hydroxy-4-hexenedioate dehydratase, a 4-hydroxy-2-oxohexanedioate oxidoreductase, a 2,4-dihydroxyadipate dehydratase (acting on 2-hydroxy), a 2,4-dihydroxyadipate dehydratase (acting on 4-hydroxyl group) and a 3-hydroxy-4-hexenedioate dehydratase.

[0045] In particular embodiments, the muconate pathway includes a set of muconate pathway enzymes shown in FIG. 3 and selected from the group consisting of:

[0046] A) (1) 4-hydroxy-2-ketovalerate aldolase, (2) 2-oxopentenoate hydratase, (3) 4-oxalocrotonate dehydrogenase, (4) 2-hydroxy-4-hexenedioate dehydratase;

[0047] B) (1) 4-hydroxy-2-ketovalerate aldolase, (2) 4-hydroxy-2-oxohexanedioate oxidoreductase, (3) 2,4-dihydroxyadipate dehydratase (acting on 2-hydroxy), (4) 3-hydroxy-4-hexenedioate dehydratase; and

[0048] C) (1) 4-hydroxy-2-ketovalerate aldolase, (2) 4-hydroxy-2-oxohexanedioate oxidoreductase, (3) 2,4-dihydroxyadipate dehydratase (acting on 4-hydroxyl group), (4) 2-hydroxy-4-hexenedioate dehydratase.

[0049] In some embodiments, a microbial organism having a pathway exemplified by those shown in FIG. 3 can include two or more exogenous nucleic acids each encoding a muconate pathway enzyme, including three, four, that is up to all of the of enzymes in a muconate pathway. The non-naturally occurring microbial organism having at least one exogenous nucleic acid can include a heterologous nucleic acid. A non-naturally occurring microbial organism having a pathway exemplified by those shown in FIG. 3 can be cultured in a substantially anaerobic culture medium.

[0050] In some embodiments, the invention provides a non-naturally occurring microbial organism having a muconate pathway that includes at least one exogenous nucleic acid encoding a muconate pathway enzyme expressed in a sufficient amount to produce muconate. The muconate pathway includes an enzyme selected from the group consisting of an HODH aldolase, an OHED hydratase, an OHED decarboxylase, an HODH formate-lyase, an HODH dehydrogenase, an OHED formate-lyase, an OHED dehydrogenase, a 6-OHE dehydrogenase, a 3-hydroxyadipyl-CoA dehydratase, a 2,3-dehydroadipyl-CoA hydrolase, a 2,3-dehydroadipyl-CoA transferase, a 2,3-dehydroadipyl-CoA ligase, and a muconate reductase.

[0051] In particular embodiments, the muconate pathway includes a set of muconate pathway enzymes shown in FIG. 4 and selected from the group consisting of:

[0052] A) (1) HODH aldolase, (2) OHED hydratase, (3) OHED decarboxylase, (4) 6-OHE dehydrogenase, and (5) muconate reductase;

[0053] B) (1) HODH aldolase, (2) OHED hydratase, (3) an enzyme selected from OHED formate-lyase and OHED dehydrogenase, (4) an enzyme selected from 2,3-dehydroa-

dipyl-CoA hydrolase, 2,3-dehydroadipyl-CoA transferase and 2,3-dehydroadipyl-CoA ligase, and (5) muconate reductase; and

[0054] C) (1) HODH aldolase, (2) an enzyme selected from HODH formate-lyase and HODH dehydrogenase, (3) 3-hydroxyadipyl-CoA dehydratase, (4) an enzyme selected from 2,3-dehydroadipyl-CoA hydrolase, 2,3-dehydroadipyl-CoA transferase and 2,3-dehydroadipyl-CoA ligase, and (5) muconate reductase

[0055] In some embodiments, a microbial organism having a pathway exemplified by those shown in FIG. 4 can include two or more exogenous nucleic acids each encoding a muconate pathway enzyme, including three, four, five, that is up to all of the of enzymes in a muconate pathway. The non-naturally occurring microbial organism having at least one exogenous nucleic acid can include a heterologous nucleic acid. A non-naturally occurring microbial organism having a pathway exemplified by those shown in FIG. 4 can be cultured in a substantially anaerobic culture medium.

[0056] In some embodiments, the invention provides a non-naturally occurring microbial organism having a muconate pathway that includes at least one exogenous nucleic acid encoding a muconate pathway enzyme expressed in a sufficient amount to produce muconate. The muconate pathway includes an enzyme selected from the group consisting of a lysine aminotransferase, a lysine aminating oxidoreductase, a 2-aminoadipate semialdehyde dehydrogenase, a 2-aminoadipate deaminase, a muconate reductase, a lysine-2,3-aminomutase, a 3,6-diaminohexanoate aminotransferase, a 3,6-diaminohexanoate aminating oxidoreductase, a 3-aminoadipate semialdehyde dehydrogenase, and a 3-aminoadipate deaminase.

[0057] In particular embodiments, the muconate pathway includes a set of muconate pathway enzymes shown in FIG. 5 and selected from the group consisting of:

[0058] A) (1) lysine aminotransferase, (2) lysine aminating oxidoreductase, (3) 2-aminoadipate semialdehyde dehydrogenase, (4) 2-aminoadipate deaminase, and (5) muconate reductase

[0059] B) (1) lysine-2,3-aminomutase, (2) 3,6-diaminohexanoate aminotransferase, (3) 3,6-diaminohexanoate aminating oxidoreductase, (4) 3-aminoadipate semialdehyde dehydrogenase, (5) 3-aminoadipate deaminase, and (6) muconate reductase.

[0060] In some embodiments, a microbial organism having a pathway exemplified by those shown in FIG. 5 can include two or more exogenous nucleic acids each encoding a muconate pathway enzyme, including three, four, five, six, that is up to all of the of enzymes in a muconate pathway. The non-naturally occurring microbial organism having at least one exogenous nucleic acid can include a heterologous nucleic acid. A non-naturally occurring microbial organism having a pathway exemplified by those shown in FIG. 2 can be cultured in a substantially anaerobic culture medium.

[0061] In an additional embodiment, the invention provides a non-naturally occurring microbial organism having a muconate pathway, wherein the non-naturally occurring microbial organism comprises at least one exogenous nucleic acid encoding an enzyme or protein that converts a substrate to a product selected from the group consisting of succinyl-CoA to beta-ketoadipyl-CoA, beta-ketoadipyl-CoA to 3-hydroxyadipyl-CoA, 3-hydroxyadipyl-CoA to 2,3-dehydroadipyl-CoA, 2,3-dehydroadipyl-CoA to 2,3-dehydroadipate, and 2,3-dehydroadipate to trans,trans-muconate. Alterna-

tively, the non-naturally occurring microbial organism comprises at least one exogenous nucleic acid encoding an enzyme or protein that converts a substrate to a product selected from the group consisting of succinyl-CoA to beta-ketoadipyl-CoA, beta-ketoadipyl-CoA to beta-ketoadipate, beta-ketoadipate to 2-maleylacetate, 2-maleylacetate to cis-3-hydroxy-4-hexendioate, cis-3-hydroxy-4-hexendioate to cis,trans-muconate, and cis,trans-muconate to trans,trans-muconate. Alternatively, the non-naturally occurring microbial organism comprises at least one exogenous nucleic acid encoding an enzyme or protein that converts a substrate to a product selected from the group consisting of succinyl-CoA to beta-ketoadipyl-CoA, beta-ketoadipyl-CoA to beta-ketoadipate, beta-ketoadipate to 2-maleylacetate, 2-maleylacetate to cis-3-amino-4-hexendioate, cis-3-amino-4-hexendioate to cis,trans-muconate, and cis,trans-muconate to trans,trans-muconate. Alternatively, the non-naturally occurring microbial organism comprises at least one exogenous nucleic acid encoding an enzyme or protein that converts a substrate to a product selected from the group consisting of succinyl-CoA to beta-ketoadipyl-CoA, beta-ketoadipyl-CoA to beta-ketoadipate, beta-ketoadipate to 2-fumarylacetate, 2-fumarylacetate to trans-3-hydroxy-4-hexendioate, and trans-3-hydroxy-4-dienoate to trans,trans-muconate. Alternatively, the non-naturally occurring microbial organism comprises at least one exogenous nucleic acid encoding an enzyme or protein that converts a substrate to a product selected from the group consisting of succinyl-CoA to beta-ketoadipyl-CoA, beta-ketoadipyl-CoA to beta-ketoadipate, beta-ketoadipate to 2-fumarylacetate, 2-fumarylacetate to trans-3-amino-4-hexendioate, trans-3-amino-4-hexendioate to trans,trans-muconate. Alternatively, the non-naturally occurring microbial organism comprises at least one exogenous nucleic acid encoding an enzyme or protein that converts a substrate to a product selected from the group consisting of succinyl-CoA to beta-ketoadipyl-CoA, beta-ketoadipyl-CoA to beta-ketoadipate, beta-ketoadipate to beta-ketoadipate enol-lactone, beta-ketoadipate enol-lactone to muconolactone, muconolactone to cis,cis-muconate, cis,cis-muconate to cis,trans-muconate, and cis,trans muconate to trans,trans-muconate. Thus, the invention provides a non-naturally occurring microbial organism containing at least one exogenous nucleic acid encoding an enzyme or protein, where the enzyme or protein converts the substrates and products of a muconate pathway, such as those shown in FIG. 2.

[0062] In an additional embodiment, the invention provides a non-naturally occurring microbial organism having a muconate pathway, wherein the non-naturally occurring microbial organism comprises at least one exogenous nucleic acid encoding an enzyme or protein that converts a substrate to a product selected from the group consisting of pyruvate and malonate semialdehyde to 4-hydroxy-2-oxohexandioate, 4-hydroxy-2-oxohexandioate to 4-oxalocrotonate, 4-oxalocrotonate to 2-hydroxy-4-hexendioate, and 2-hydroxy-4-hexendioate to muconate. Alternatively, the non-naturally occurring microbial organism comprises at least one exogenous nucleic acid encoding an enzyme or protein that converts a substrate to a product selected from the group consisting of pyruvate and malonate semialdehyde to 4-hydroxy-2-oxohexandioate, 4-hydroxy-2-oxohexandioate, 4-hydroxy-2-oxohexandioate to 2,4-dihydroxyadipate, 2,4-dihydroxyadipate to 2-hydroxy-4-hexendioate, and 2-hydroxy-4-hexendioate to muconate. Alternatively, the non-naturally occurring microbial organism comprises at

least one exogenous nucleic acid encoding an enzyme or protein that converts a substrate to a product selected from the group consisting of pyruvate and malonate semialdehyde to 4-hydroxy-2-oxohexandioate, 4-hydroxy-2-oxohexandioate, 4-hydroxy-2-oxohexandioate to 2,4-dihydroxyadipate, 2,4-dihydroxyadipate to 3-hydroxy-4-hexendioate, and 3-hydroxy-4-hexendioate to muconate. Thus, the invention provides a non-naturally occurring microbial organism containing at least one exogenous nucleic acid encoding an enzyme or protein, where the enzyme or protein converts the substrates and products of a muconate pathway, such as those shown in FIG. 3.

[0063] In an additional embodiment, the invention provides a non-naturally occurring microbial organism having a muconate pathway, wherein the non-naturally occurring microbial organism comprises at least one exogenous nucleic acid encoding an enzyme or protein that converts a substrate to a product selected from the group consisting of pyruvate and succinic semialdehyde to HODH, HODH to 3-hydroxyadipyl-CoA, 3-hydroxy adipyl-CoA to 2,3-dehydroadipyl-CoA, 2,3-dehydroadipyl-CoA to 2,3-dehydroadipate, and 2,3-dehydroadipate to muconate. Alternatively, the non-naturally occurring microbial organism comprises at least one exogenous nucleic acid encoding an enzyme or protein that converts a substrate to a product selected from the group consisting of pyruvate and succinic semialdehyde to HODH, HODH to OHED, OHED to 2,3-dehydroadipyl-CoA, 2,3-dehydroadipyl-CoA to 2,3-dehydroadipate, and 2,3-dehydroadipate to muconate. Alternatively, the non-naturally occurring microbial organism comprises at least one exogenous nucleic acid encoding an enzyme or protein that converts a substrate to a product selected from the group consisting of pyruvate and succinic semialdehyde to HODH, HODH to OHED, OHED to 6-OHE, 6-OHE to 2,3-dehydroadipate, and 2,3-dehydroadipate to muconate. Thus, the invention provides a non-naturally occurring microbial organism containing at least one exogenous nucleic acid encoding an enzyme or protein, where the enzyme or protein converts the substrates and products of a muconate pathway, such as those shown in FIG. 4.

[0064] In an additional embodiment, the invention provides a non-naturally occurring microbial organism having a muconate pathway, wherein the non-naturally occurring microbial organism comprises at least one exogenous nucleic acid encoding an enzyme or protein that converts a substrate to a product selected from the group consisting of lysine to 2-aminoadipate semialdehyde, 2-aminoadipate semialdehyde to 2-aminoadipate, 2-aminoadipate to 2,3-dehydroadipate, and 2,3-dehydroadipate to muconate. Alternatively, the non-naturally occurring microbial organism comprises at least one exogenous nucleic acid encoding an enzyme or protein that converts a substrate to a product selected from the group consisting of lysine to 3,6-diaminohexanoate, 3,6-diaminohexanoate to 3-aminoadipate semialdehyde, 3-aminoadipate semialdehyde to 3-aminoadipate, 3-aminoadipate to 2,3-dehydroadipate, and 2,3-dehydroadipate to muconate. Thus, the invention provides a non-naturally occurring microbial organism containing at least one exogenous nucleic acid encoding an enzyme or protein, where the enzyme or protein converts the substrates and products of a muconate pathway, such as those shown in FIG. 5.

[0065] The invention is described herein with general reference to the metabolic reaction, reactant or product thereof, or with specific reference to one or more nucleic acids or

genes encoding an enzyme associated with or catalyzing, or a protein associated with, the referenced metabolic reaction, reactant or product. Unless otherwise expressly stated herein, those skilled in the art will understand that reference to a reaction also constitutes reference to the reactants and products of the reaction. Similarly, unless otherwise expressly stated herein, reference to a reactant or product also references the reaction, and reference to any of these metabolic constituents also references the gene or genes encoding the enzymes that catalyze or proteins involved in the referenced reaction, reactant or product. Likewise, given the well known fields of metabolic biochemistry, enzymology and genomics, reference herein to a gene or encoding nucleic acid also constitutes a reference to the corresponding encoded enzyme and the reaction it catalyzes or a protein associated with the reaction as well as the reactants and products of the reaction.

[0066] Muconate can be produced from succinyl-CoA via beta-ketoadipate in a minimum of five enzymatic steps, shown in FIG. 2. In the first step of all pathways, succinyl-CoA is joined to acetyl-CoA by a beta-ketothiolase to form beta-ketoadipyl-CoA (Step A). In one embodiment, the beta-keto functional group is reduced and dehydrated to form 2,3-dehydroadipyl-CoA (Steps K and L). The CoA moiety is then removed by a CoA hydrolase, transferase or ligase to form 2,3-dehydroadipate (Step M). Finally, 2,3-dehydroadipate is oxidized to form the conjugated diene muconate by an enoate oxidoreductase (Step N).

[0067] In other embodiments, beta-ketoadipyl-CoA is converted to beta-ketoadipate by a CoA hydrolase, transferase or ligase (Step B). Beta-ketoadipate is then converted to 2-maleylacetate by maleylacetate reductase (Step O). The beta-ketone of 2-maleylacetate is then reduced to form cis-3-hydroxy-4-hexenoate (Step P). This product is further dehydrated to cis,trans-muconate in Step Q. Step W provides a muconate cis/trans-isomerase to provide trans,trans-muconate.

[0068] A similar route entails the conversion of 2-maleylacetate to cis-3-amino-4-hexenoate by an aminotransferase or aminating oxidoreductase (Step R). Deamination of cis-3-amino-4-hexenoate is subsequently carried out to form cis,trans-muconate (Step S).

[0069] Alternatively, beta-ketoadipate can be converted to 2-fumarylacetate by action of a fumarylacetate reductase (Step C). Such a reductase can be engineered by directed evolution, for example, of the corresponding maleylacetate reductase. Reduction of the keto group and dehydration provides trans,trans-muconate (Steps D and E). Alternatively, reductive amination, followed by deamination also affords the trans,trans-muconate product (Steps F and G).

[0070] In yet another route, beta-ketoadipate can be cyclized to an enol-lactone by beta-ketoadipyl enol-lactone hydrolase (Step H). The double bond in the lactone ring is then shifted by muconolactone isomerase (Step I). Finally, muconolactone is converted to cis,cis-muconate by muconate cycloisomerase (Step J). Muconate cycloisomerase may selectively form the cis,cis isomer of muconate. Further addition of a cis/trans isomerase converts the cis,cis isomer to the favored trans, trans or trans, cis configurations (Steps T and W, which can be incorporated into a single isomerization step).

[0071] The pathways detailed in FIG. 2 can achieve a maximum theoretical yield of 1.09 moles muconate per mole glucose utilized under anaerobic and aerobic conditions. With and without aeration, the maximum ATP yield is 1 mole of

ATP per glucose utilized at the maximum muconate yield. The first step of this pathway, the condensation of succinyl-CoA and acetyl-CoA by beta-ketothiolase, has been demonstrated by Applicants is shown below in Example I.

[0072] Another pathway for muconate synthesis involves the condensation of pyruvate and malonate semialdehyde, as shown in FIG. 3. Malonate semialdehyde can be formed in the cell by several different pathways. Two example pathways are: 1) decarboxylation of oxaloacetate, and 2) conversion of 2-phosphoglycerate to glycerol which can then be dehydrated to malonate semialdehyde by a diol dehydratase. In one pathway, malonate semialdehyde and pyruvate are condensed to form 4-hydroxy-2-oxohexanedioate (Step A). This product is dehydrated to form 4-oxalocrotonate (Step B). 4-Oxalocrotonate is converted to muconate by reduction and dehydration of the 2-keto group (Steps C and D).

[0073] Alternately, the 2-keto group of 4-hydroxy-2-oxohexanedioate is reduced by an alcohol-forming oxidoreductase (Step E). The product, 2,4-dihydroxyadipate is then dehydrated at the 2- or 4-hydroxy position to form 2-hydroxy-4-hexenedioate (Step G) or 3-hydroxy-4-hexenedioate (Step F). Subsequent dehydration yields the diene, muconate (Steps D or H). This pathway is energetically favorable and is useful because it does not require carboxylation steps. Also, the pathway is driven by the stability of the muconate end product.

[0074] Several pathways for producing muconate from pyruvate and succinic semialdehyde are detailed in FIG. 4. Such pathways entail aldol condensation of pyruvate with succinic semialdehyde to 4-hydroxy-2-oxoheptane-1,7-dioate (HODH) by HODH aldolase (Step A). In one route, HODH is dehydrated to form 2-oxohept-4-ene-1,7-dioate (OHED) by OHED hydratase (Step B). OHED is then decarboxylated to form 6-oxo-2,3-dehydrohexanoate (6-OHE) (Step C). This product is subsequently oxidized to the diacid and then further oxidized to muconate (Steps F, I).

[0075] Alternately, HODH is converted to 3-hydroxyadipyl-CoA by a formate-lyase or an acylating decarboxylating dehydrogenase (Step D). The 3-hydroxy group of 3-hydroxyadipyl-CoA is then dehydrated to form the enoyl-CoA (Step G). The CoA moiety of 2,3-dehydroadipyl-CoA is removed by a CoA hydrolase, ligase or transferase (Step H). Finally, 2,3-dehydroadipate is oxidized to muconate by muconate reductase (Step 1).

[0076] In yet another route, OHED is converted to 2,3-dehydroadipyl-CoA by a formate-lyase or acylating decarboxylating dehydrogenase (Step E). 2,3-Dehydroadipyl-CoA is then transformed to muconate.

[0077] Pathways for producing muconate from lysine are detailed in FIG. 5. In one embodiment, lysine is converted to 2-aminoadipate semialdehyde by an aminotransferase or aminating oxidoreductase (Step A). 2-Aminoadipate semialdehyde is then oxidized to form 2-aminoadipate (Step B). The 2-amino group is then deaminated by a 2-aminoadipate deaminase (Step C). The product, 2,3-dehydroadipate is further oxidized to muconate by muconate reductase (Step D).

[0078] In an alternate route, the 2-amino group of lysine is shifted to the 3-position by lysine-2,3-aminomutase (Step E). The product, 3,6-diaminohexanoate, is converted to 3-aminoadipate semialdehyde by an aminotransferase or aminating oxidoreductase (Step F). Oxidation of the aldehyde (Step G) and deamination (Step H) yields 2,3-dehydroadipate, which is then converted to muconate (Step D).

[0079] All transformations depicted in FIGS. 2-5 fall into the general categories of transformations shown in Table 1. Below is described a number of biochemically characterized genes in each category. Specifically listed are genes that can be applied to catalyze the appropriate transformations in FIGS. 2-5 when properly cloned and expressed.

[0080] Table 1 shows the enzyme types useful to convert common central metabolic intermediates into muconate. The first three digits of each label correspond to the first three Enzyme Commission number digits which denote the general type of transformation independent of substrate specificity.

TABLE 1

Label	Function
1.1.1.a	Oxidoreductase (oxo to alcohol, and reverse)
1.2.1.a	Oxidoreductase (acid to oxo)
1.2.1.c	Oxidoreductase (2-ketoacid to acyl-CoA)
1.3.1.a	Oxidoreductase (alkene to alkane, and reverse)
1.4.1.a	Oxidoreductase (aminating)
2.3.1.b	Acyltransferase (beta-ketothiolase)
2.3.1.d	Acyltransferase (formate C-acyltransferase)
2.6.1.a	Aminotransferase
2.8.3.a	CoA transferase
3.1.1.a	Enol-lactone hydrolase
3.1.2.a	CoA hydrolase
4.1.1.a	Carboxy-lyase
4.1.2.a	Aldehyde-lyase
4.2.1.a	Hydro-lyase
4.3.1.a	Ammonia-lyase
5.2.1.a	Cis/trans isomerase
5.3.3.a	Lactone isomerase
5.4.3.a	Aminomutase
5.5.1.a	Lactone cycloisomerase
6.2.1.a	CoA synthetase

[0081] Several transformations depicted in FIGS. 2-5 require oxidoreductases that convert a ketone functionality to a hydroxyl group. The conversion of beta-ketoadipyl-CoA to 3-hydroxyadipyl-CoA (FIG. 2, Step K) is catalyzed by a 3-oxoacyl-CoA dehydrogenase. The reduction of 2-fumarylacetate to trans-3-hydroxy-4-hexendioate (FIG. 2, Step D) or 2-maleylacetate to cis-3-hydroxy-4-hexendioate (FIG. 2, Step P), is catalyzed by an oxidoreductase that converts a 3-oxoacid to a 3-hydroxyacid. Reduction of the ketone group of 4-oxalocrotonate and 4-hydroxy-2-oxohexanedioate to their corresponding hydroxyl group is also catalyzed by enzymes in this family (FIG. 3, Steps C and E).

[0082] Exemplary enzymes for converting beta-ketoadipyl-CoA to 3-hydroxyadipyl-CoA (FIG. 2, Step K) include 3-hydroxyacyl-CoA dehydrogenases. Such enzymes convert 3-oxoacyl-CoA molecules into 3-hydroxyacyl-CoA molecules and are often involved in fatty acid beta-oxidation or phenylacetate catabolism. For example, subunits of two fatty acid oxidation complexes in *E. coli*, encoded by fadB and fadJ, function as 3-hydroxyacyl-CoA dehydrogenases (Binstock and Schultz, Methods Enzymol. 71Pt C:403-411 (1981)). Furthermore, the gene products encoded by phaC in *Pseudomonas putida* U (Olivera et al., Proc. Natl. Acad. Sci. U.S.A. 95:6419-6424 (1998)) and paaC in *Pseudomonas fluorescens* ST (Di et al., Arch. Microbiol. 188:117-125 (2007)) catalyze the reverse reaction of step B in FIG. 10, that is, the oxidation of 3-hydroxyadipyl-CoA to form 3-oxoadipyl-CoA, during the catabolism of phenylacetate or styrene. Note that the reactions catalyzed by such enzymes are reversible. In addition, given the proximity in *E. coli* of paaH to other genes in the phenylacetate degradation operon (Nogales et al.,

Microbiology 153:357-365 (2007)) and the fact that paaH mutants cannot grow on phenylacetate (Ismail et al., Eur. J. Biochem. 270:3047-3054 (2003)), it is expected that the *E. coli* paaH gene encodes a 3-hydroxyacyl-CoA dehydrogenase. Genbank information related to these genes is summarized in Table 2 below.

TABLE 2

Gene	GI #	Accession No.	Organism
fadB	119811	P21177.2	<i>Escherichia coli</i>
fadJ	3334437	P77399.1	<i>Escherichia coli</i>
paaH	16129356	NP_415913.1	<i>Escherichia coli</i>
phaC	26990000	NP_745425.1	<i>Pseudomonas putida</i>
paaC	106636095	ABF82235.1	<i>Pseudomonas fluorescens</i>

[0083] Additional exemplary oxidoreductases capable of converting 3-oxoacyl-CoA molecules to their corresponding 3-hydroxyacyl-CoA molecules include 3-hydroxybutyryl-CoA dehydrogenases. The enzyme from *Clostridium acetobutylicum*, encoded by hbd, has been cloned and functionally expressed in *E. coli* (Youngleson et al., J. Bacteriol. 171:6800-6807 (1989)). Additional genes include Hbd1 (C-terminal domain) and Hbd2 (N-terminal domain) in *Clostridium kluyveri* (Hillmer and Gottschalk, FEBS Lett. 21:351-354 (1972)) and HSD17B10 in *Bos taurus* (Wakil et al., J. Biol. Chem. 207:631-638 (1954)). Yet other genes demonstrated to reduce acetoacetyl-CoA to 3-hydroxybutyryl-CoA are phbB from *Zoogloea ramigera* (Ploux et al., Eur. J. Biochem. 174:177-182 (1988)) and phaB from *Rhodobacter sphaeroides* (Alber et al., Mol. Microbiol. 61:297-309 (2006)). The former gene is NADPH-dependent, its nucleotide sequence has been determined (Peoples and Sinskey, Mol. Microbiol. 3:349-357 (1989)) and the gene has been expressed in *E. coli*. Substrate specificity studies on the gene led to the conclusion that it could accept 3-oxopropionyl-CoA as an alternate substrate (Ploux et al., Eur. J. Biochem. 174:177-182 (1988)). Genbank information related to these genes is summarized in Table 3 below.

TABLE 3

Gene	GI #	Accession No.	Organism
hbd	18266893	P52041.2	<i>Clostridium acetobutylicum</i>
Hbd2	146348271	EDK34807.1	<i>Clostridium kluyveri</i>
Hbd1	146345976	EDK32512.1	<i>Clostridium kluyveri</i>
HSD17B10	3183024	O02691.3	<i>Bos taurus</i>
phaB			<i>Rhodobacter sphaeroides</i>
phbB			<i>Zoogloea ramigera</i>

[0084] A number of similar enzymes have been found in other species of *Clostridia* and in *Metallosphaera sedula* (Berg et al., Science 318:1782-1786 (2007)) as shown in Table 4.

TABLE 4

Gene	GI#	Accession No.	Organism
hbd	15895965	NP_349314.1	<i>Clostridium acetobutylicum</i>
hbd	20162442	AAM14586.1	<i>Clostridium beijerinckii</i>
Msed_1423	146304189	YP_001191505	<i>Metallosphaera sedula</i>
Msed_0399	146303184	YP_001190500	<i>Metallosphaera sedula</i>
Msed_0389	146303174	YP_001190490	<i>Metallosphaera sedula</i>
Msed_1993	146304741	YP_001192057	<i>Metallosphaera sedula</i>

[0085] There are various alcohol dehydrogenases for converting 2-maleylacetate to cis-3-hydroxy-4-hexenoate (FIG. 2, Step P), 2-fumarylacetate to trans-3-hydroxy-4-hexenoate (FIG. 2, Step D), 4-oxalocrotonate to 5-hydroxyhex-2-enedioate (FIG. 3, Step C) and 4-hydroxy-2-oxohexanedioate to 2,4-dihydroxyadipate (FIG. 3, Step E). Two enzymes capable of converting an oxoacid to a hydroxyacid are encoded by the malate dehydrogenase (mdh) and lactate dehydrogenase (ldhA) genes in *E. coli*. In addition, lactate dehydrogenase from *Ralstonia eutropha* has been shown to demonstrate high activities on substrates of various chain lengths such as lactate, 2-oxobutyrate, 2-oxopentanoate and 2-oxoglutarate (Steinbuchel and Schlegel, *Eur. J. Biochem.* 130:329-334 (1983)). Conversion of alpha-ketoadipate into alpha-hydroxyadipate can be catalyzed by 2-ketoadipate reductase, an enzyme reported to be found in rat and in human placenta (Suda et al., *Arch. Biochem. Biophys.* 176:610-620 (1976); Suda et al., *Biochem. Biophys. Res. Commun.* 77:586-591 (1977)). An additional gene for these steps is the mitochondrial 3-hydroxybutyrate dehydrogenase (bdh) from the human heart which has been cloned and characterized (Marks et al., *J. Biol. Chem.* 267:15459-15463 (1992)). This enzyme is a dehydrogenase that operates on a 3-hydroxyacid. Another exemplary alcohol dehydrogenase converts acetone to isopropanol as was shown in *C. beijerinckii* (Ismail et al., *Eur. J. Biochem.* 270:3047-3054 (2003)) and *T. Brockii* (Lamed and Zeikus, *Biochem. J.* 195:183-190 (1981); Peretz and Burstein, *Biochemistry* 28:6549-6555 (1989)). Genbank information related to these genes is summarized in Table 5 below.

TABLE 5

Gene	GI #	Accession No.	Organism
mdh	1789632	AAC76268.1	<i>Escherichia coli</i>
ldhA	16129341	NP_415898.1	<i>Escherichia coli</i>
bdh	177198	AAA58352.1	<i>Homo sapiens</i>
adh	60592974	AAA23199.2	<i>Clostridium beijerinckii</i>
adh	113443	P14941.1	<i>Thermoanaerobacter Brockii</i>

[0086] Enzymes in the 1.2.1 family are NAD(P)⁺-dependent oxidoreductases that convert aldehydes to acids. Reactions catalyzed by enzymes in this family include the oxidation of 6-OHE (FIG. 4, Step F), 2-aminoadipate semialdehyde (FIG. 5, Step B) and 3-aminoadipate semialdehyde (FIG. 5, Step G) to their corresponding acids. An exemplary enzyme is the NAD⁺-dependent aldehyde dehydrogenases (EC 1.2.1.3). Two aldehyde dehydrogenases found in human liver, ALDH-1 and ALDH-2, have broad substrate ranges for a variety of aliphatic, aromatic and polycyclic aldehydes (Klyosov, A. A., *Biochemistry* 35:4457-4467 (1996)). Active ALDH-2 has been efficiently expressed in *E. coli* using the GroEL proteins as chaperonins (Lee et al., *Biochem. Biophys. Res. Commun.* 298:216-224 (2002)). The rat mitochondrial aldehyde dehydrogenase also has a broad substrate range that includes the enoyl-aldehyde crotonaldehyde (Siew et al., *Arch. Biochem. Biophys.* 176:638-649 (1976)). The *E. coli* gene astD also encodes an NAD⁺-dependent aldehyde dehydrogenase active on succinic semialdehyde (Kuznetsova et al., *FEMS Microbiol. Rev.* 29:263-279 (2005)). Genbank information related to these genes is summarized in Table 5 below.

TABLE 6

Gene	GI #	Accession No.	Organism
ALDH-2	118504	P05091.2	<i>Homo sapiens</i>
ALDH-2	14192933	NP_115792.1	<i>Rattus norvegicus</i>
astD	3913108	P76217.1	<i>Escherichia coli</i>

[0087] Two transformations in FIG. 4 require conversion of a 2-ketoacid to an acyl-CoA (FIG. 4, Steps D and E) by an enzyme in the EC class 1.2.1. Such reactions are catalyzed by multi-enzyme complexes that catalyze a series of partial reactions which result in acylating oxidative decarboxylation of 2-keto-acids. Exemplary enzymes that can be used include 1) branched-chain 2-keto-acid dehydrogenase, 2) alpha-ketoglutarate dehydrogenase, and 3) the pyruvate dehydrogenase multienzyme complex (PDHC). Each of the 2-keto-acid dehydrogenase complexes occupies positions in intermediary metabolism, and enzyme activity is typically tightly regulated (Fries et al., *Biochemistry* 42:6996-7002 (2003)). The enzymes share a complex but common structure composed of multiple copies of three catalytic components: alpha-ketoacid decarboxylase (E1), dihydrolipoamide acyltransferase (E2) and dihydrolipoamide dehydrogenase (E3). The E3 component is shared among all 2-keto-acid dehydrogenase complexes in an organism, while the E1 and E2 components are encoded by different genes. The enzyme components are present in numerous copies in the complex and utilize multiple cofactors to catalyze a directed sequence of reactions via substrate channeling. The overall size of these dehydrogenase complexes is very large, with molecular masses between 4 and 10 million Da (i.e., larger than a ribosome).

[0088] Activity of enzymes in the 2-keto-acid dehydrogenase family is normally low or limited under anaerobic conditions in *E. coli*. Increased production of NADH (or NADPH) could lead to a redox-imbalance, and NADH itself serves as an inhibitor to enzyme function. Engineering efforts have increased the anaerobic activity of the *E. coli* pyruvate dehydrogenase complex (Kim et al., *Appl. Environ. Microbiol.* 73:1766-1771 (2001); Kim et al., *J. Bacteriol.* 190:3851-3858 (2008); Zhou et al., *Biotechnol. Lett.* 30:335-342 (2008)). For example, the inhibitory effect of NADH can be overcome by engineering an H322Y mutation in the E3 component (Kim et al., *J. Bacteriol.* 190:3851-3858 (2008)). Structural studies of individual components and how they work together in complex provide insight into the catalytic mechanisms and architecture of enzymes in this family (Aevvarsson et al., *Nat. Struct. Biol.* 6:785-792 (1999); Zhou et al., *Proc. Natl. Acad. Sci. U.S.A.* 98:14802-14807 (2001)). The substrate specificity of the dehydrogenase complexes varies in different organisms, but generally branched-chain keto-acid dehydrogenases have the broadest substrate range.

[0089] Alpha-ketoglutarate dehydrogenase (AKGD) converts alpha-ketoglutarate to succinyl-CoA and is the primary site of control of metabolic flux through the TCA cycle (Hansford, R. G., *Curr. Top. Bioenerg.* 10:217-278 (1980)). Encoded by genes sucA, sucB and lpd in *E. coli*, AKGD gene expression is downregulated under anaerobic conditions and during growth on glucose (Park et al., *Mol. Microbiol.* 15:473-482 (1993)). Although the substrate range of AKGD is narrow, structural studies of the catalytic core of the E2 component pinpoint specific residues responsible for substrate specificity (Knapp et al., *J. Mol. Biol.* 280:655-668 (1998)). The *Bacillus subtilis* AKGD, encoded by odhAB (E1

and E2) and pdhD (E3, shared domain), is regulated at the transcriptional level and is dependent on the carbon source and growth phase of the organism (Resnekov et al., *Mol. Gen. Genet.* 234:285-296 (1992)). In yeast, the LPD1 gene encoding the E3 component is regulated at the transcriptional level by glucose (Roy and Dawes, *J. Gen. Microbiol.* 133:925-933 (1987)). The E1 component, encoded by KGD1, is also regulated by glucose and activated by the products of HAP2 and HAP3 (Repetto and Tzagoloff, *Moll. Cell. Biol.* 9:2695-2705 (1989)). The AKGD enzyme complex, inhibited by products NADH and succinyl-CoA, is known in mammalian systems, as impaired function of has been linked to several neurological diseases (Tretter and dam-Vizi, *Philos. Trans. R. Soc. Lond B Biol. Sci.* 360:2335-2345 (2005)). Genbank information related to these genes is summarized in Table 7 below.

TABLE 7

Gene	GI #	Accession No.	Organism
sucA	16128701	NP_415254.1	<i>Escherichia coli</i>
sucB	16128702	NP_415255.1	<i>Escherichia coli</i>
lpd	16128109	NP_414658.1	<i>Escherichia coli</i>
odhA	51704265	P23129.2	<i>Bacillus subtilis</i>
odhB	129041	P16263.1	<i>Bacillus subtilis</i>
pdhD	118672	P21880.1	<i>Bacillus subtilis</i>
KGD1	6322066	NP_012141.1	<i>Saccharomyces cerevisiae</i>
KGD2	6320352	NP_010432.1	<i>Saccharomyces cerevisiae</i>
LPD1	14318501	NP_116635.1	<i>Saccharomyces cerevisiae</i>

[0090] Branched-chain 2-keto-acid dehydrogenase complex (BCKAD), also known as 2-oxoisovalerate dehydrogenase, participates in branched-chain amino acid degradation pathways, converting 2-keto acids derivatives of valine, leucine and isoleucine to their acyl-CoA derivatives and CO₂. The complex has been studied in many organisms including *Bacillus subtilis* (Wang et al., *Eur. J. Biochem.* 213:1091-1099 (1993)), *Rattus norvegicus* (Namba et al., *J. Biol. Chem.* 244:4437-4447 (1969)) and *Pseudomonas putida* (Sokatch et al., *J. Bacteriol.* 148:647-652 (1981)). In *Bacillus subtilis* the enzyme is encoded by genes pdhD (E3 component), bfmBB (E2 component), bfmBAA and bfmBAB (E1 component) (Wang et al., *Eur. J. Biochem.* 213:1091-1099 (1993)). In mammals, the complex is regulated by phosphorylation by specific phosphatases and protein kinases. The complex has been studied in rat hepatocytes (Chicco et al., *J. Biol. Chem.* 269:19427-19434 (1994)) and is encoded by genes Bckdha (E1 alpha), Bckdhh (E1 beta), Dbt (E2), and Dld (E3). The E1 and E3 components of the *Pseudomonas putida* BCKAD complex have been crystallized (Aevansson et al., *Nat. Struct. Biol.* 6:785-792 (1999); Mattevi et al., *Science* 255:1544-1550 (1992)) and the enzyme complex has been studied (Sokatch et al., *J. Bacteriol.* 148:647-652 (1981)). Transcription of the *P. putida* BCKAD genes is activated by the gene product of bkdR (Hesslinger et al., *Mol. Microbiol.* 27:477-492 (1998)). In some organisms including *Rattus norvegicus* (Paxton et al., *Biochem. J.* 234:295-303 (1986)) and *Saccharomyces cerevisiae* (Sinclair et al., *Biochem. Mol. Biol. Int.* 31:911-9122 (1993)), this complex has been shown to have a broad substrate range that includes linear oxo-acids such as 2-oxobutanoate and alpha-ketoglutarate, in addition to the branched-chain amino acid precursors. The active site of the bovine BCKAD was engineered to favor alternate substrate acetyl-CoA (Meng and Chuang, *Biochemistry* 33:12879-12885 (1994)). Genbank information related to these genes is summarized in Table 8 below.

TABLE 8

Gene	GI #	Accession No.	Organism
bfmBB	16079459	NP_390283.1	<i>Bacillus subtilis</i>
bfmBAA	16079461	NP_390285.1	<i>Bacillus subtilis</i>
bfmBAB	16079460	NP_390284.1	<i>Bacillus subtilis</i>
pdhD	118672	P21880.1	<i>Bacillus subtilis</i>
lpdV	118677	P09063.1	<i>Pseudomonas putida</i>
bkdB	129044	P09062.1	<i>Pseudomonas putida</i>
bkdA1	26991090	NP_746515.1	<i>Pseudomonas putida</i>
bkdA2	26991091	NP_746516.1	<i>Pseudomonas putida</i>
Bckdha	77736548	NP_036914.1	<i>Rattus norvegicus</i>
Bckdhh	158749538	NP_062140.1	<i>Rattus norvegicus</i>
Dbt	158749632	NP_445764.1	<i>Rattus norvegicus</i>
Dld	40786469	NP_955417.1	<i>Rattus norvegicus</i>

[0091] The pyruvate dehydrogenase complex, catalyzing the conversion of pyruvate to acetyl-CoA, has also been studied. In the *E. coli* enzyme, specific residues in the E1 component are responsible for substrate specificity (Bisswanger, H., *J. Biol. Chem.* 256:815-822 (1981); Bremer, J., *Eur. J. Biochem.* 8:535-540 (1969); Gong et al., *J. Biol. Chem.* 275:13645-13653 (2000)). As mentioned previously, enzyme engineering efforts have improved the *E. coli* PDH enzyme activity under anaerobic conditions (Kim et al., *Appl. Environ. Microbiol.* 73:1766-1771 (2007); Kim et al., *J. Bacteriol.* 190:3851-3858 (2008); Zhou et al., *Biotechnol. Letter.* 30:335-342 (2008)). In contrast to the *E. coli* PDH, the *B. subtilis* complex is active and required for growth under anaerobic conditions (Nakano et al., *J. Bacteriol.* 179:6749-6755 (1997)). The *Klebsiella pneumoniae* PDH, characterized during growth on glycerol, is also active under anaerobic conditions (Menzel et al., *J. Biotechnol.* 56:135-142 (1997)). Crystal structures of the enzyme complex from bovine kidney (Zhou et al., *Proc. Natl. Acad. Sci. U.S.A.* 98:14802-14807 (2001)) and the E2 catalytic domain from *Azotobacter vinelandii* are available (Mattevi et al., *Science* 255:1544-1550 (1992)). Some mammalian PDH enzymes complexes can react on alternate substrates such as 2-oxobutanoate, although comparative kinetics of *Rattus norvegicus* PDH and BCKAD indicate that BCKAD has higher activity on 2-oxobutanoate as a substrate (Paxton et al., *Biochem. J.* 234:295-303 (1986)). Genbank information related to these genes is summarized in Table 9 below.

TABLE 9

Gene	GI #	Accession No.	Organism
aceE	16128107	NP_414656.1	<i>Escherichia coli</i>
aceF	16128108	NP_414657.1	<i>Escherichia coli</i>
lpd	16128109	NP_414658.1	<i>Escherichia coli</i>
pdhA	3123238	P21881.1	<i>Bacillus subtilis</i>
pdhB	129068	P21882.1	<i>Bacillus subtilis</i>
pdhC	129054	P21883.2	<i>Bacillus subtilis</i>
pdhD	118672	P21880.1	<i>Bacillus subtilis</i>
aceE	152968699	YP_001333808.1	<i>Klebsiella pneumonia</i>
aceF	152968700	YP_001333809.1	<i>Klebsiella pneumonia</i>
lpdA	152968701	YP_001333810.1	<i>Klebsiella pneumonia</i>
Pdha1	124430510	NP_001004072.2	<i>Rattus norvegicus</i>
Pdha2	16758900	NP_446446.1	<i>Rattus norvegicus</i>
Dlat	78365255	NP_112287.1	<i>Rattus norvegicus</i>
Dld	40786469	NP_955417.1	<i>Rattus norvegicus</i>

[0092] As an alternative to the large multienzyme 2-keto-acid dehydrogenase complexes described above, some anaerobic organisms utilize enzymes in the 2-ketoacid oxidoreductase family (OFOR) to catalyze acylating oxidative

decarboxylation of 2-keto-acids. Unlike the dehydrogenase complexes, these enzymes contain iron-sulfur clusters, utilize different cofactors, and use ferredoxin or flavodoxin as electron acceptors in lieu of NAD(P)H. While most enzymes in this family are specific to pyruvate as a substrate (POR) some 2-keto-acid:ferredoxin oxidoreductases have been shown to accept a broad range of 2-ketoacids as substrates including alpha-ketoglutarate and 2-oxobutanoate (Fukuda and Wakagi, *Biochim. Biophys. Acta.* 1597_74-80 (2002); Zhang et al., *J. Biochem.* 120:587-599 (1996)). One such enzyme is the OFOR from the thermoacidophilic archaeon *Sulfolobus tokodaii* 7, which contains an alpha and beta subunit encoded by gene ST2300 (Fukuda and Wakagi, supra; Zhang et al., supra). A plasmid-based expression system has been developed for efficiently expressing this protein in *E. coli* (Fukuda et al., *Eur. J. Biochem.* 268:5639-5646 (2001)) and residues involved in substrate specificity were determined (Fukuda and Wakagi, supra). Two OFORs from *Aeropyrum pernix* str. K1 have also been recently cloned into *E. coli*, characterized, and found to react with a broad range of 2-oxoacids (Nishizawa et al., *FEBS Lett.* 579_2319-2322 (2005)). The gene sequences of these OFOR enzymes are available, although they do not have GenBank identifiers assigned to date. There is bioinformatic evidence that similar enzymes are present in all archaea, some anaerobic bacteria and amitochondrial eukarya (Fukuda and Wakagi, supra). This class of enzyme is also interesting from an energetic standpoint, as reduced ferredoxin could be used to generate NADH by ferredoxin-NAD reductase (Petitdemanche et al., *Biochim. Biophys. Acta* 421:334-337 (1976)). Also, since most of the enzymes are designed to operate under anaerobic conditions, less enzyme engineering may be required relative to enzymes in the 2-keto-acid dehydrogenase complex family for activity in an anaerobic environment. Genbank information related to these genes is summarized in Table 10 below.

TABLE 10

Gene	GI #	Accession No.	Organism
ST2300	15922633	NP_378302.1	<i>Sulfolobus tokodaii</i> 7

[0093] Three transformations fall into the category of oxidoreductases that reduce an alkene to an alkane (EC 1.3.1.-). The conversion of beta-ketoadipate to 2-maleylacetate (FIG. 2, Step O) is also catalyzed by the 2-enoate oxidoreductase maleylacetate reductase (MAR). A similar enzyme converts beta-ketoadipate to 2-fumarylacetate (FIG. 2, Step C). The oxidation of 2,3-dehydroadipate to muconate (FIG. 2, Step N) is catalyzed by a 2-enoate oxidoreductase with muconate reductase functionality.

[0094] 2-Enoate oxidoreductase enzymes are known to catalyze the NAD(P)H-dependent reduction and oxidation of a wide variety of α , β -unsaturated carboxylic acids and aldehydes (Rohdich et al., *J. Biol. Chem.* 276:5779-5787 (2001)). In the recently published genome sequence of *C. kluyveri*, 9 coding sequences for enoate reductases were reported, out of which one has been characterized (Seedorf et al., *Proc. Natl. Acad. Sci. U.S.A.* 105:2128-2133 (2008)). The enr genes from both *C. tyrobutyricum* and *M. thermoacetica* have been cloned and sequenced and show 59% identity to each other. The former gene is also found to have approximately 75% similarity to the characterized gene in *C. kluyveri* (Giesel and Simon, *Arch. Microbiol.* 135:51-57 (1983)). It has been reported based on these sequence results that enr is very

similar to the dienoyl CoA reductase in *E. coli* (fadH) (Rohdich et al., *J. Biol. Chem.* 276:5779-5787 (2001)). The *C. thermoacetica* enr gene has also been expressed in a catalytically active form in *E. coli* (Rohdich et al., supra). Genbank information related to these genes is summarized in Table 11 below.

TABLE 11

Gene	GI #	Accession No.	Organism
enr	169405742	ACA54153.1	<i>Clostridium botulinum</i> A3 str
enr	2765041	CAA71086.1	<i>Clostridium tyrobutyricum</i>
enr	3402834	CAA76083.1	<i>Clostridium kluyveri</i>
enr	83590886	YP_430895.1	<i>Moorella thermoacetica</i>
fadH	16130976	NP_417552.1	<i>Escherichia coli</i>

[0095] MAR is a 2-enoate oxidoreductase catalyzing the reversible reduction of 2-maleylacetate (cis-4-oxohex-2-enedioate) to 3-oxoadipate (FIG. 2, Step O). MAR enzymes naturally participate in aromatic degradation pathways (Camara et al., *J. Bacteriol.* 191:4905-4915 (2009); Huang et al., *Appl. Environ. Microbiol.* 72:7238-7245 (2006); Kaschabek and Reineke, *J. Bacteriol.* 177:320-325 (1995); Kaschabek and Reineke, *J. Bacteriol.* 175:6075-6081 (1993)). The enzyme activity was identified and characterized in *Pseudomonas* sp. strain B13 (Kaschabek and Reineke, (1995) supra; Kaschabek and Reineke, (1993) supra), and the coding gene was cloned and sequenced (Kasberg et al., *J. Bacteriol.* 179:3801-3803 (1997)). Additional MAR genes include clcE gene from *Pseudomonas* sp. strain B13 (Kasberg et al., supra), macA gene from *Rhodococcus opacus* (Seibert et al., *J. Bacteriol.* 175:6745-6754 (1993)), the macA gene from *Ralstonia eutropha* (also known as *Cupriavidus necator*) (Seibert et al., *Microbiology* 150:463-472 (2004)), tfdFII from *Ralstonia eutropha* (Seibert et al., (1993) supra) and NCgl1112 in *Corynebacterium glutamicum* (Huang et al., *Appl. Environ. Microbiol.* 72:7238-7245 (2006)). A MAR in *Pseudomonas reinekei* MT1, encoded by ccaD, was recently identified and the nucleotide sequence is available under the DBJ/EMBL GenBank accession number EF159980 (Camara et al., *J. Bacteriol.* 191:4905-4915 (2009)). Genbank information related to these genes is summarized in Table 12 below.

TABLE 12

Gene	GI #	Accession No.	Organism
clcE	3913241	O30847.1	<i>Pseudomonas</i> sp. strain B13
macA	7387876	O84992.1	<i>Rhodococcus opacus</i>
macA	5916089	AAD55886	<i>Cupriavidus necator</i>
tfdFII	1747424	AC44727.1	<i>Ralstonia eutropha</i> JMP134
NCgl1112	19552383	NP_600385	<i>Corynebacterium glutamicum</i>
ccaD			<i>Pseudomonas reinekei</i> MT1

[0096] In Step R of FIG. 2, 2-maleylacetate is transaminated to form 3-amino-4-hexanoate. The conversion of 2-fumarylacetate to trans-3-amino-4-hexenedioate is a similar transformation (FIG. 2, Step F). These reactions are performed by aminating oxidoreductases in the EC class 1.4.1. Enzymes in this EC class catalyze the oxidative deamination of alpha-amino acids with NAD⁺ or NADP⁺ as acceptor, and the reactions are typically reversible. Exemplary enzymes include glutamate dehydrogenase (deaminating), encoded by gdhA, leucine dehydrogenase (deaminating), encoded by ldh, and aspartate dehydrogenase (deaminating), encoded by nadX. The gdhA gene product from *Escherichia coli* (Korber

et al., *J. Mol. Biol.* 234:1270-1273 (1993); McPherson and Wootton, *Nucleic Acids Res.* 11:5257-5266 (1983)), *gdh* from *Thermotoga maritima* (Kort et al., *Extremophiles* 1:52-60-1997); Lebbink et al., *J. Mol. Biol.* 280:287-296 (1998); Lebbink et al., *J. Mol. Biol.* 289:357-369 (1999)), and *gdhA1* from *Halobacterium salinarum* (Ingoldsby et al., *Gene* 349:237-244 (2005)) catalyze the reversible conversion of glutamate to 2-oxoglutarate and ammonia, while favoring NADP(H), NAD(H), or both, respectively. The *ldh* gene of *Bacillus cereus* encodes the LeuDH protein that has a wide of range of substrates including leucine, isoleucine, valine, and 2-aminobutanoate (Ansorge and Kula, *Biotechnol. Bioeng.* 68:557-562 (2000); Stoyan et al., *J. Biotechnol.* 54:77-80 (1997)). The *nadX* gene from *Thermotoga maritima* encoding for the aspartate dehydrogenase is involved in the biosynthesis of NAD (Yang et al., *J. Biol. Chem.* 278:8804-8808 (2003)). Genbank information related to these genes is summarized in Table 13 below.

TABLE 13

Gene	GI #	Accession No.	Organism
<i>gdhA</i>	118547	P00370	<i>Escherichia coli</i>
<i>gdh</i>	6226595	P96110.4	<i>Thermotoga maritima</i>
<i>gdhA1</i>	15789827	NP_279651.1	<i>Halobacterium salinarum</i>
<i>ldh</i>	61222614	POA393	<i>Bacillus cereus</i>
<i>nadX</i>	15644391	NP_229443.1	<i>Thermotoga maritima</i>

[0097] The conversions of lysine to 2-aminoadipate semialdehyde (FIG. 5, Step A) and 3,6-diaminohexanoate to 3-aminoadipate semialdehyde (FIG. 5, Step F) are catalyzed by aminating oxidoreductases that transform primary amines to their corresponding aldehydes. The lysine 6-dehydrogenase (deaminating), encoded by the *lysDH* genes, catalyze the oxidative deamination of the 6-amino group of L-lysine to form 2-aminoadipate-6-semialdehyde, which can spontaneously and reversibly cyclize to form Δ^1 -piperidine-6-carboxylate (Misono and Nagasaki, *J. Bacteriol.* 150:398-401 (1982)). Exemplary enzymes are found in *Geobacillus stearothermophilus* (Heydari et al., *Appl. Environ. Microbiol.* 70:937-942 (2004)), *Agrobacterium tumefaciens* (Hashimoto et al., *J. Biochem.* 106:76-80 (1989), Misono and Nagasaki, supra), and *Achromobacter denitrificans* (Ruldeekulthamrong et al., *BMB. Rep.* 41:790-795 (2008)). Such enzymes can convert 3,6-diaminohexanoate to 3-aminoadipate semialdehyde given the structural similarity between 3,6-diaminohexanoate and lysine. Genbank information related to these genes is summarized in Table 14 below.

TABLE 14

Gene	GI #	Accession No.	Organism
<i>lysDH</i>	13429872	BAB39707	<i>Geobacillus stearothermophilus</i>
<i>lysDH</i>	15888285	NP_353966	<i>Agrobacterium tumefaciens</i>
<i>lysDH</i>	74026644	AAZ94428	<i>Achromobacter denitrificans</i>

[0098] FIG. 2, step A uses a 3-oxoadipyl-CoA thiolase, or equivalently, succinyl CoA:acetyl CoA acyl transferase (β -ketothiolase). The gene products encoded by *pcaF* in *Pseudomonas* strain B13 (Kaschabek et al., *J. Bacteriol.* 184:207-215 (2002)), *phaD* in *Pseudomonas putida* U (Olivera et al., *Proc. Natl. Acad. Sci. U.S.A.* 95:6419-6424 (1998)), *paaE* in *Pseudomonas fluorescens* ST (Di et al., *Arch. Microbiol.* 188:117-125 (2007)), and *paaJ* from *E. coli* (Nogales et al.,

Microbiology 153:357-365 (2007)) catalyze the conversion of 3-oxoadipyl-CoA into succinyl-CoA and acetyl-CoA during the degradation of aromatic compounds such as phenylacetate or styrene. Since beta-ketothiolase enzymes catalyze reversible transformations, these enzymes can also be employed for the synthesis of 3-oxoadipyl-CoA. Several beta-ketothiolases were shown to have significant and selective activities in the oxoadipyl-CoA forming direction as shown in Example I below including *bkt* from *Pseudomonas putida*, *pcaF* and *bkt* from *Pseudomonas aeruginosa* PAO1, *bkt* from *Burkholderia ambifaria* AMMD, *paaJ* from *E. coli*, and *phaD* from *P. putida*. Genbank information related to these genes is summarized in Table 15 below.

TABLE 15

Gene	GI #	Accession No.	Organism
<i>paaJ</i>	16129358	NP_415915.1	<i>Escherichia coli</i>
<i>pcaF</i>	17736947	AAL02407	<i>Pseudomonas knackmussii</i> (B13)
<i>phaD</i>	3253200	AAC24332.1	<i>Pseudomonas putida</i>
<i>pcaF</i>	506695	AAA85138.1	<i>Pseudomonas putida</i>
<i>paaE</i>	106636097	ABF82237.1	<i>Pseudomonas fluorescens</i>
<i>bkt</i>			<i>Burkholderia ambifaria</i> AMMD
<i>bkt</i>			<i>Pseudomonas aeruginosa</i> PAO1
<i>pcaF</i>			<i>Pseudomonas aeruginosa</i> PAO1

[0099] The acylation of ketoacids HODH and OHED to their corresponding CoA derivatives (FIG. 4, Steps D and E) and concurrent release of formate, is catalyzed by formate C-acyltransferase enzymes in the EC class 2.3.1. Enzymes in this class include pyruvate formate-lyase and ketoacid formate-lyase. Pyruvate formate-lyase (PFL, EC 2.3.1.54), encoded by *pflB* in *E. coli*, converts pyruvate into acetyl-CoA and formate. The active site of PFL contains a catalytically essential glycyl radical that is posttranslationally activated under anaerobic conditions by PFL-activating enzyme (PFL-AE, EC 1.97.1.4) encoded by *pflA* (Knappe et al., *Proc. Natl. Acad. Sci. U.S.A.* 81:1332-1335 (1984); Wong et al., *Biochemistry* 32:14102-14110 (1993)). A pyruvate formate-lyase from *Archaeoglobus fulgidus* encoded by *pflD* has been cloned, expressed in *E. coli* and characterized (Lehtio and Goldman, *Protein Eng Des Sel* 17:545-552 (2004)). The crystal structures of the *A. fulgidus* and *E. coli* enzymes have been resolved (Lehtio et al., *J. Mol. Biol.* 357:221-235 (2006); Leppanen et al., *Structure* 7:733-744 (1999)). Additional PFL and PFL-AE enzymes are found in *Clostridium pasteurianum* (Weidner and Sawyers, *J. Bacteriol.* 178:2440-2444 (1996)) and the eukaryotic alga *Chlamydomonas reinhardtii* (Hem-schemeier et al., *Eukaryot. Cell* 7:518-526 (2008)). Ketoacid formate-lyase (EC 2.3.1.-), also known as 2-ketobutyrate formate-lyase (KFL) and pyruvate formate-lyase 4, is the gene product of *tdcE* in *E. coli*. This enzyme catalyzes the conversion of 2-ketobutyrate to propionyl-CoA and formate during anaerobic threonine degradation, and can also substitute for pyruvate formate-lyase in anaerobic catabolism (Simanshu et al., *J. Biosci.* 32:1195-1206 (2007)). The enzyme is oxygen-sensitive and, like *PflB*, requires post-translational modification by PFL-AE to activate a glycyl radical in the active site (Hesslinger et al., *Mol. Microbiol.* 27:477-492 (1998)). Genbank information related to these genes is summarized in Table 16 below.

TABLE 16

Gene	GI #	Accession No.	Organism
pflB	16128870	NP_415423.1	<i>Escherichia coli</i>
pflA	16128869	NP_415422.1	<i>Escherichia coli</i>
tdcE	48994926	AAT48170.1	<i>Escherichia coli</i>
pflD	11499044	NP_070278.1	<i>Archaeglobus fulgidus</i>
pfl	2500058	Q46266.1	<i>Clostridium pasteurianum</i>
act	1072362	CAA63749.1	<i>Clostridium pasteurianum</i>
pfl1	159462978	XP_001689719.1	<i>Chlamydomonas reinhardtii</i>
pflA1	159485246	XP_001700657.1	<i>Chlamydomonas reinhardtii</i>

[0100] Several reactions in FIGS. 2 and 5 are catalyzed by aminotransferases in the EC class 2.6.1 (FIG. 2, Steps F and R and FIG. 5, Steps A and F). Such enzymes reversibly transfer amino groups from aminated donors to acceptors such as pyruvate and alpha-ketoglutarate. The conversion of lysine to 2-amino adipate (FIG. 5, Step A) is naturally catalyzed by lysine-6-aminotransferase (EC 2.6.1.36). This enzyme function has been demonstrated in yeast and bacteria. Enzymes from *Candida wilesii* (Hammer et al. *J. Basic Microbiol.* 32:21-27 (1992)), *Flavobacterium lutescens* (Fuji et al. *J. Biochem.* 128:391-397 (2000)) and *Streptomyces clavuligenus* (Romero et al. *J. Ind. Microbiol. Biotechnol.* 18:241-246 (1997)) have been characterized. A recombinant lysine-6-aminotransferase from *S. clavuligenus* was functionally expressed in *E. coli* (Tobin et al. *J. Bacteriol.* 173:6223-6229 (1991)). The *F. lutescens* enzyme is specific to alpha-ketoglutarate as the amino acceptor (Soda et al. *Biochemistry* 7:4110-4119 (1968)). Lysine-6-aminotransferase is also an enzyme that can catalyze the transamination of 3,6-diaminohexanoate (FIG. 5, Step F), as this substrate is structurally similar to lysine. Genbank information related to these genes is summarized in Table 17 below.

TABLE 17

Gene	GI #	Accession No.	Organism
lat	10336502	BAB13756.1	<i>Flavobacterium lutescens</i>
lat	153343	AAA26777.1	<i>Streptomyces clavuligenus</i>

[0101] In Steps R and F of FIG. 2 the beta-ketones of 2-maleylacetate and 2-fumarylacetate, respectively, are converted to secondary amines. Beta-alanine/alpha-ketoglutarate aminotransferase (WO08027742) reacts with beta-alanine to form malonic semialdehyde, a 3-oxoacid similar in structure to 2-maleylacetate. The gene product of SkPYD4 in *Saccharomyces kluyveri* was shown to preferentially use beta-alanine as the amino group donor (Andersen and Hansen, *Gene* 124:105-109 (1993)). SkUGA1 encodes a homologue of *Saccharomyces cerevisiae* GABA aminotransferase, UGA1 (Ramos et al., *Eur. J. Biochem.* 149:401-404 (1985)), whereas SkPYD4 encodes an enzyme involved in both -alanine and GABA transamination (Andersen and Hansen, supra). 3-Amino-2-methylpropionate transaminase catalyzes the transformation from methylmalonate semialdehyde to 3-amino-2-methylpropionate. The enzyme has been characterized in *Rattus norvegicus* and *Sus scrofa* and is encoded by Abat (Kakimoto et al., *Biochim. Biophys. Acta* 156:374-380 (1968); Tamaki et al., *Methods Enzymol.* 324:376-389 (2000)). Genbank information related to these genes is summarized in Table 18 below.

TABLE 18

Gene	GI #	Accession No.	Organism
SkyPYD4	98626772	ABF58893.1	<i>Lachancea kluyveri</i>
SkUGA1	98626792	ABF58894.1	<i>Lachancea kluyveri</i>
UGA1	6321456	NP_011533.1	<i>Saccharomyces cerevisiae</i>
Abat	122065191	P50554.3	<i>Rattus norvegicus</i>
Abat	120968	P80147.2	<i>Sus scrofa</i>

[0102] Another enzyme that can catalyze the aminotransferase reactions in FIGS. 2 and 5 is gamma-aminobutyrate transaminase (GABA transaminase), which naturally interconverts succinic semialdehyde and glutamate to 4-aminobutyrate and alpha-ketoglutarate and is known to have a broad substrate range (Liu et al., *Biochemistry* 43:10896-10905 (2004); Shigeoka and Nakano, *Arch. Biochem. Biophys.* 288:22-28 (1991); Schulz et al., *Appl. Environ. Microbiol.* 56:1-6 (1990)). *E. coli* has two GABA transaminases, encoded by gabT (Bartsch and Schulz, *J. Bacteriol.* 172:7035-7042 (1990)) and puuE (Kurihara et al., *J. Biol. Chem.* 280:4602-4608 (2005)). GABA transaminases in *Mus musculus*, *Pseudomonas fluorescens*, and *Sus scrofa* have been shown to react with alternate substrates (Cooper, A. J., *Methods Enzymol.* 113:80-82 (1985); Scott and Jakoby, *J. Biol. Chem.* 234:932-936 (1959)). Genbank information related to these genes is summarized in Table 19 below.

TABLE 19

Gene	GI #	Accession No.	Organism
gabT	16130576	NP_417148.1	<i>Escherichia coli</i>
puuE	16129263	P_415818.1	<i>Escherichia coli</i>
aba	37202121	NP_766549.2	<i>Mus musculus</i>
gabT	70733692	YP_257332.1	<i>Pseudomonas fluorescens</i>
aba	47523600	NP_999428.1	<i>Sus scrofa</i>

[0103] CoA transferases catalyze the reversible transfer of a CoA moiety from one molecule to another. Conversion of beta-ketoacyl-CoA to beta-ketoacyl-CoA (FIG. 2, Step B) is accompanied by the acylation of succinate by beta-ketoacyl-CoA transferase. The de-acylation of 2,3-dehydroacyl-CoA (FIG. 2, Step M and FIG. 4, Step H) can also be catalyzed by an enzyme in the 2.8.3 family.

[0104] Beta-ketoacyl-CoA transferase (EC 2.8.3.6), also known as succinyl-CoA:3-oxoacid-CoA transferase, is encoded by pcaI and pcaJ in *Pseudomonas putida* (Kaschabek et al., *J. Bacteriol.* 184:207-215 (2002)). Similar enzymes based on homology exist in *Acinetobacter* sp. ADP1 (Kowalchuk et al., *Gene* 146:23-30 (1994)). Additional exemplary succinyl-CoA:3-oxoacid-CoA transferases are present in *Helicobacter pylori* (Corthesy-Theulaz et al., *J. Biol. Chem.* 272:25659-25667 (1997)) and *Bacillus subtilis* (Stols et al., *Protein Expr. Purif.* 53:396-403 (2007)). Genbank information related to these genes is summarized in Table 20 below.

TABLE 20

Gene	GI #	Accession No.	Organism
pcaI	24985644	AAN69545.1	<i>Pseudomonas putida</i>
pcaJ	26990657	NP_746082.1	<i>Pseudomonas putida</i>
pcaI	50084858	YP_046368.1	<i>Acinetobacter</i> sp. ADP1
pcaJ	141776	AAC37147.1	<i>Acinetobacter</i> sp. ADP1

TABLE 20-continued

Gene	GI #	Accession No.	Organism
pcaI	21224997	NP_630776.1	<i>Streptomyces coelicolor</i>
pcaJ	21224996	NP_630775.1	<i>Streptomyces coelicolor</i>
HPAG1_0676	108563101	YP_627417	<i>Helicobacter pylori</i>
HPAG1_0677	108563102	YP_627418	<i>Helicobacter pylori</i>
ScoA	16080950	NP_391778	<i>Bacillus subtilis</i>
ScoB	16080949	NP_391777	<i>Bacillus subtilis</i>

[0105] The glutaconyl-CoA-transferase (EC 2.8.3.12) enzyme from anaerobic bacterium *Acidaminococcus fermentans* reacts with glutaconyl-CoA and 3-butenoyl-CoA (Mack et al., *Eur. J. Biochem.* 226:41-51 (1994)), substrates similar in structure to 2,3-dehydroadipyl-CoA. The genes encoding this enzyme are gctA and gctB. This enzyme has reduced but detectable activity with other CoA derivatives including glutaryl-CoA, 2-hydroxyglutaryl-CoA, adipyl-CoA and acrylyl-CoA (Buckel et al., *Eur. J. Biochem.* 118:315-321 (1981)). The enzyme has been cloned and expressed in *E. coli* (Mack, supra). Genbank information related to these genes is summarized in Table 21 below.

TABLE 21

Gene	GI #	Accession No.	Organism
gctA	559392	CAA57199.1	<i>Acidaminococcus fermentans</i>
gctB	559393	CAA57200.1	<i>Acidaminococcus fermentans</i>

[0106] Other exemplary CoA transferases are catalyzed by the gene products of cat1, cat2, and cat3 of *Clostridium kluyveri* which have been shown to exhibit succinyl-CoA, 4-hydroxybutyryl-CoA, and butyryl-CoA transferase activity, respectively (Seedorf et al., *Proc. Natl. Acad. Sci. U.S.A.* 105:2128-2133 (2008); Sohling and Gottschalk, *J. Bacteriol.* 178:871-880 (1996)). Similar CoA transferase activities are also present in *Trichomonas vaginalis* (van Grinsven et al., *J. Biol. Chem.* 283:1411-1418 (2008)) and *Trypanosoma brucei* (Riviere et al., *J. Biol. Chem.* 279:45337-45346 (2004)). Genbank information related to these genes is summarized in Table 22 below.

TABLE 22

Gene	GI #	Accession No.	Organism
cat1	729048	P38946.1	<i>Clostridium kluyveri</i>
cat2	172046066	P38942.2	<i>Clostridium kluyveri</i>
cat3	146349050	EDK35586.1	<i>Clostridium kluyveri</i>
TVAG_395550	123975034	XP_001330176	<i>Trichomonas vaginalis</i> G3
Tb11.02.0290	71754875	XP_828352	<i>Trypanosoma brucei</i>

[0107] A CoA transferase that can utilize acetyl-CoA as the CoA donor is acetoacetyl-CoA transferase, encoded by the *E. coli* atoA (alpha subunit) and atoD (beta subunit) genes (Korolev et al., *Acta Crystallogr. D. Biol. Crystallogr.* 58:2116-2121 (2002); Vanderwinkel et al., *Biochem. Biophys. Res. Commun.* 33:902-908 (1968)). This enzyme has a broad substrate range (Sramek and Frerman, *Arch. Biochem. Biophys.* 171:14-26 (1975)) and has been shown to transfer the CoA moiety to acetate from a variety of branched and linear acyl-CoA substrates, including isobutyrate (Matthies and Schink, *Appl. Environ. Microbiol.* 58:1435-1439 (1992)), valerate (Vanderwinkel et al, supra) and butanoate (Vanderwinkel et

al, supra). This enzyme is induced at the transcriptional level by acetoacetate, so modification of regulatory control may be necessary for engineering this enzyme into a pathway (Pauli and Overath, *Eur. J. Biochem.* 29:553-562 (1972)). Similar enzymes exist in *Corynebacterium glutamicum* ATCC 13032 (Duncan et al., *Appl. Environ. Microbiol.* 68:5186-5190 (2002)), *Clostridium acetobutylicum* (Cary et al., *Appl. Environ. Microbiol.* 56:1576-1583 (1990); Weisenborn et al., *Appl. Environ. Microbiol.* 55:323-329 (1989)), and *Clostridium saccharoperbutylacetonicum* (Kosaka et al., *Biosci. Biotechnol. Biochem.* 71:58-58 (2007)). Genbank information related to these genes is summarized in Table 23 below.

TABLE 23

Gene	GI #	Accession No.	Organism
atoA	2492994	P76459.1	<i>Escherichia coli</i>
atoD	2492990	P76458.1	<i>Escherichia coli</i>
actA	62391407	YP_226809.1	<i>Corynebacterium glutamicum</i>
cg0592	62389399	YP_224801.1	<i>Corynebacterium glutamicum</i>
ctfA	15004866	NP_149326.1	<i>Clostridium acetobutylicum</i>
ctfB	15004867	NP_149327.1	<i>Clostridium acetobutylicum</i>
ctfA	31075384	AAP42564.1	<i>Clostridium saccharoperbutylacetonicum</i>
ctfB	31075385	AAP42565.1	<i>Clostridium saccharoperbutylacetonicum</i>

[0108] In Step H of FIG. 2, the lactonization of beta-keto-adipate to form β -keto-adipate-enol-lactone is catalyzed by the beta-keto-adipate enol-lactonase (EC-3.1.1.24). Beta-keto-adipate enol-lactonase also participates in the catechol branch of the beta-keto-adipate pathway to degrade aromatic compounds, in the reverse direction of that required in Step H of FIG. 2. This enzyme is encoded by the pcaD gene in *Pseudomonas putida* (Hughes et al., *J. Gen. Microbiol.* 134:2877-2887 (1988)), *Rhodococcus opacus* (Eulberg et al., *J. Bacteriol.* 180:1072-1081 (1998)) and *Ralstonia eutropha*. In *Acinetobacter calcoaceticus*, genes encoding two β -keto-adipate enol-lactone hydrolases were identified (Patel et al., *J. Biol. Chem.* 250:6567 (1975)). Genbank information related to these genes is summarized in Table 24 below.

TABLE 24

Gene	GI #	Accession No.	Organism
ELH	6015088	Q59093	<i>Acinetobacter calcoaceticus</i>
ELH2	6166146	P00632	<i>Acinetobacter calcoaceticus</i>
pcaD	24982842	AAN67003	<i>Pseudomonas putida</i>
pcaD	75426718	O67982	<i>Rhodococcus opacus</i>
pcaD	75411823	javascript: Blast2('Q9EV41')Q9EV45	<i>Ralstonia eutropha</i>

[0109] The hydrolysis of acyl-CoA molecules to their corresponding acids is carried out by acyl CoA hydrolase enzymes in the 3.1.2 family, also called thioesterases. Several eukaryotic acetyl-CoA hydrolases (EC 3.1.2.1) have broad substrate specificity and thus represent suitable enzymes for hydrolyzing beta-keto-adipyl-CoA and 2,3-dehydroadipyl-CoA (FIG. 2, Steps B and M and FIG. 4, Step H). For example, the enzyme from *Rattus norvegicus* brain (Robinson et al., *Biochem. Biophys. Res. Commun.* 71:959-965 (1976)) can react with butyryl-CoA, hexanoyl-CoA and malonyl-CoA. The enzyme from the mitochondrion of the

pea leaf also has a broad substrate specificity, with demonstrated activity on acetyl-CoA, propionyl-CoA, butyryl-CoA, palmitoyl-CoA, oleoyl-CoA, succinyl-CoA, and crotonyl-CoA (Zeihner and Randall, *Plant Physiol.* 94:20-27 (1990)). The acetyl-CoA hydrolase, ACH1, from *S. cerevisiae* represents another hydrolase (Buu et al., *J. Biol. Chem.* 278:17203-17209 (2003)). Genbank information related to these genes is summarized in Table 25 below.

TABLE 25

Gene	GI #	Accession No.	Organism
acot12	18543355	NP_570103.1	<i>Rattus norvegicus</i>
ACH1	6319456	NP_009538	<i>Saccharomyces cerevisiae</i>

[0110] Another hydrolase is the human dicarboxylic acid thioesterase, acot8, which exhibits activity on glutaryl-CoA, adipyl-CoA, suberyl-CoA, sebacyl-CoA, and dodecanedioyl-CoA (Westin et al., *J. Biol. Chem.* 280:38125-28132 (2005)) and the closest *E. coli* homolog, tesB, which can also hydrolyze a broad range of CoA thioesters (Naggert et al., *J. Biol. Chem.* 266:11044-11050 (1991)). A similar enzyme has also been characterized in the rat liver (Deana, R., *Biochem. Int.* 26:767-773 (1992)). Genbank information related to these genes is summarized in Table 26 below.

TABLE 26

Gene	GI #	Accession No.	Organism
tesB	16128437	NP_414986	<i>Escherichia coli</i>
acot8	3191970	CAA15502	<i>Homo sapiens</i>
acot8	51036669	NP_570112	<i>Rattus norvegicus</i>

[0111] Other potential *E. coli* thioester hydrolases include the gene products of tesA (Bonner and Bloch, *J. Biol. Chem.* 247:3123-3133 (1972)), ybgC (Kuznetsova et al., *FEBS Microbiol. Rev.* 29:263-279 (2005); Zhuang et al., *FEBS Lett.* 516:161-163 (2002)), paaI (Song et al., *J. Biol. Chem.* 281:11028-11038 (2006)), and ybdB (Leduc et al., *J. Bacteriol.* 1889:7112-7126 (2007)). Genbank information related to these genes is summarized in Table 27 below.

TABLE 27

Gene	GI #	Accession No.	Organism
tesA	16128478	NP_415027	<i>Escherichia coli</i>
ybgC	16128711	NP_415264	<i>Escherichia coli</i>
paaI	16129357	NP_415914	<i>Escherichia coli</i>
ybdB	16128580	NP_415129	<i>Escherichia coli</i>

[0112] Yet another hydrolase is the glutaconate CoA-transferase from *Acidaminococcus fermentans*. This enzyme was transformed by site-directed mutagenesis into an acyl-CoA hydrolase with activity on glutaryl-CoA, acetyl-CoA and 3-butenoyl-CoA (Mack and Buckel, *FEBS Lett.* 405:209-212 (1997)), compounds similar in structure to 2,3-dehydroadipyl-CoA. This indicates that the enzymes encoding succinyl-CoA:3-ketoacid-CoA transferases and acetoacetyl-CoA: acetyl-CoA transferases can also serve as enzymes for this reaction step but would require certain mutations to change their function. Genbank information related to these genes is summarized in Table 28 below.

TABLE 28

Gene	GI #	Accession No.	Organism
gctA	559392	CAA57199	<i>Acidaminococcus fermentans</i>
gctB	559393	CAA57200	<i>Acidaminococcus fermentans</i>

[0113] Step C of FIG. 4 is catalyzed by a 2-ketoacid decarboxylase that generates 6-oxo-2,3-dehydrohexanoate (6-OHE) from 2-oxohept-4-ene-1,7-dioate (OHED). The decarboxylation of keto-acids is catalyzed by a variety of enzymes with varied substrate specificities, including pyruvate decarboxylase (EC 4.1.1.1), benzoylformate decarboxylase (EC 4.1.1.7), alpha-ketoglutarate decarboxylase and branched-chain alpha-ketoacid decarboxylase. Pyruvate decarboxylase (PDC), also termed keto-acid decarboxylase, is a key enzyme in alcoholic fermentation, catalyzing the decarboxylation of pyruvate to acetaldehyde. The enzyme from *Saccharomyces cerevisiae* has a broad substrate range for aliphatic 2-keto acids including 2-ketobutyrate, 2-ketovalerate, 3-hydroxypyruvate and 2-phenylpyruvate (22). This enzyme has been extensively studied, engineered for altered activity, and functionally expressed in *E. coli* (Killenberg-Jabs et al., *Eur. J. Biochem.* 268:1698-1704 (2001); Li and Jordan, *Biochemistry* 38:10004-10012 (1999); ter Schure et al., *Appl. Environ. Microbiol.* 64:1303-1307 (1998)). The PDC from *Zymomonas mobilis*, encoded by pdc, also has a broad substrate range and has been a subject of directed engineering studies to alter the affinity for different substrates (Siegert et al., *Protein Eng Des Sel* 18:345-357 (2005)). The crystal structure of this enzyme is available (Killenberg-Jabs, et al., supra). Other well-characterized PDC enzymes include the enzymes from *Acetobacter pasteurians* (Chandra et al., *Arch. Microbiol.* 176:443-451 (2001)) and *Kluyveromyces lactis* (Krieger et al., *Eur. J. Biochem.* 269:3256-3263 (2002)). Genbank information related to these genes is summarized in Table 29 below.

TABLE 29

Gene	GI #	Accession No.	Organism
pdc	118391	P06672.1	<i>Zymomonas mobilis</i>
pdc1	30923172	P06169	<i>Saccharomyces cerevisiae</i>
pdc	20385191	AM21208	<i>Acetobacter pasteurians</i>
pdc1	52788279	Q12629	<i>Kluyveromyces lactis</i>

[0114] Like PDC, benzoylformate decarboxylase (EC 4.1.1.7) has a broad substrate range and has been the target of enzyme engineering studies. The enzyme from *Pseudomonas putida* has been extensively studied and crystal structures of this enzyme are available (Hasson et al., *Biochemistry* 37:9918-9930 (1998); Polovnikova et al., *Biochemistry* 42:1820-1830 (2003)). Site-directed mutagenesis of two residues in the active site of the *Pseudomonas putida* enzyme altered the affinity (Km) of naturally and non-naturally occurring substrates (Siegert et al., *Protein Eng Des Sel* 18:345-357 (2005)). The properties of this enzyme have been further modified by directed engineering (Lingen et al., *Protein Eng.* 15:585-593 (2002); Lingen et al., *Chembiochem.* 4:721-726 (2003)). The enzyme from *Pseudomonas aeruginosa*, encoded by mdlC, has also been characterized experimentally (Barrowman et al., *FEMS Microbiology Letters* 34:57-60 (1986)). Additional genes from *Pseudomonas stutzeri*, *Pseudomonas fluorescens* and other organisms can be

inferred by sequence homology or identified using a growth selection system developed in *Pseudomonas putida* (Henning et al., *Appl. Environ. Microbiol.* 72:7510-7517 (2006)). Genbank information related to these genes is summarized in Table 30 below.

TABLE 30

Gene	GI #	Accession No.	Organism
mdlC	3915757	P20906.2	<i>Pseudomonas putida</i>
mdlC	81539678	Q9HUR2.1	<i>Pseudomonas aeruginosa</i>
dpgB	126202187	ABN80423.1	<i>Pseudomonas stutzeri</i>
ilvB-1	70730840	YP_260581.1	<i>Pseudomonas fluorescens</i>

[0115] A third enzyme capable of decarboxylating 2-oxoacids is alpha-ketoglutarate decarboxylase (KGD). The substrate range of this class of enzymes has not been studied to date. The KDC from *Mycobacterium tuberculosis* (Tian et al., *Proc. Natl. Acad. Sci. U.S.A.* 102:10670-10675 (2005)) has been cloned and functionally expressed, although it is large (~130 kD) and GC-rich. KDC enzyme activity has been detected in several species of rhizobia including *Bradyrhizobium japonicum* and *Mesorhizobium loti* (Green et al., *J. Bacteriol.* 182:2838-2844 (2000)). Although the KDC-encoding gene(s) have not been isolated in these organisms, the genome sequences are available and several genes in each genome are annotated as putative KDCs. A KDC from *Euglena gracilis* has also been characterized but the gene associated with this activity has not been identified to date (Shigeoka and Nakano, *Arch. Biochem. Biophys.* 288:22-28 (1991)). The first twenty amino acids starting from the N-terminus were sequenced MTYKAPVKDVKFLLDKVFVKV (Shigeoka and Nakano, supra). The gene could be identified by testing genes containing this N-terminal sequence for KDC activity. Genbank information related to these genes is summarized in Table 31 below.

TABLE 31

Gene	GI #	Accession No.	Organism
kgd	160395583	O50463.4	<i>Mycobacterium tuberculosis</i>
kgd	27375563	NP_767092.1	<i>Bradyrhizobium japonicum</i>
kgd	13473636	NP_105204.1	<i>Mesorhizobium loti</i>

[0116] A fourth enzyme for catalyzing this reaction is branched chain alpha-ketoacid decarboxylase (BCKA). This class of enzyme has been shown to act on a variety of compounds varying in chain length from 3 to 6 carbons (Oku and Kaneda, *J. Bio. Chem.* 263:18386-18396 (1988); Smit et al., *App. Environ. Microbiol.* 71:303-311 (2005)). The enzyme in *Lactococcus lactis* has been characterized on a variety of branched and linear substrates including 2-oxobutanoate, 2-oxohexanoate, 2-oxopentanoate, 3-methyl-2-oxobutanoate, 4-methyl-2-oxobutanoate and isocaproate (Smit et al., supra). The enzyme has been structurally characterized (Berg et al., *Science* 318:1782-1786 (2007)). Sequence alignments between the *Lactococcus lactis* enzyme and the pyruvate decarboxylase of *Zymomonas mobilis* indicate that the catalytic and substrate recognition residues are nearly identical (Siegert et al., *Protein Eng Des Sel* 18:345-357 (2005)), so this enzyme can be subjected to directed engineering. Decarboxylation of alpha-ketoglutarate by a BCKA was detected in *Bacillus subtilis*; however, this activity was low (5%) relative to activity on other branched-chain substrates

(Oku and Kaneda, supra) and the gene encoding this enzyme has not been identified to date. Additional BCKA genes can be identified by homology to the *Lactococcus lactis* protein sequence. Many of the high-scoring BLASTp hits to this enzyme are annotated as indolepyruvate decarboxylases (EC 4.1.1.74). Indolepyruvate decarboxylase (IPDA) is an enzyme that catalyzes the decarboxylation of indolepyruvate to indoleacetaldehyde in plants and plant bacteria. Genbank information related to these genes is summarized in Table 32 below.

TABLE 32

Gene	GI #	Accession No.	Organism
kdcA	44921617	AAS49166.1	<i>Lactococcus lactis</i>

[0117] Recombinant branched chain alpha-keto acid decarboxylase enzymes derived from the E1 subunits of the mitochondrial branched-chain keto acid dehydrogenase complex from *Homo sapiens* and *Bos taurus* have been cloned and functionally expressed in *E. coli* (Davie et al., *J. Biol. Chem.* 267:16601-16606 (1992); Wynn et al., *J. Biol. Chem.* 267:1881-1887 (1992); Wynn et al., *J. Biol. Chem.* 267:12400-12403 (1992)). It was indicated that co-expression of chaperonins GroEL and GroES enhanced the specific activity of the decarboxylase by 500-fold (Wynn (1992) supra). These enzymes are composed of two alpha and two beta subunits. Genbank information related to these genes is summarized in Table 33 below.

TABLE 33

Gene	GI #	Accession No.	Organism
BCKDHB	34101272	NP_898871.1	<i>Homo sapiens</i>
BCKDHA	11386135	NP_000700.1	<i>Homo sapiens</i>
BCKDHB	115502434	P21839	<i>Bos taurus</i>
BCKDHA	129030	P11178	<i>Bos taurus</i>

[0118] Aldehyde lyases in EC class 4.1.2 catalyze two key reactions in the disclosed pathways to muconate (FIG. 3, Step A and FIG. 4, Step A). HOHD aldolase, also known as HHED aldolase, catalyzes the conversion of 4-hydroxy-2-oxoheptane-1,7-dioate (HOHD) into pyruvate and succinic semialdehyde (FIG. 4, Step A). HODH aldolase is a divalent metal ion-dependent class II aldolase, catalyzing the final step of 4-hydroxyphenylacetic acid degradation in *E. coli* C, *E. coli* W, and other organisms. In the native context, the enzyme functions in the degradative direction. The reverse (condensation) reaction is thermodynamically unfavorable; however the equilibrium can be shifted through coupling HOHD aldolase with downstream pathway enzymes that work efficiently on reaction products. Such strategies have been effective for shifting the equilibrium of other aldolases in the condensation direction (Nagata et al., *Appl. Microbiol. Biotechnol.* 44:432-438 (1995); Pollard et al., *App. Environ. Microbiol.* 64:4093-4094 (1998)). The *E. coli* C enzyme, encoded by hpcH, has been extensively studied and has recently been crystallized (Rea et al., *J. Mol. Biol.* 373:866-876 (2007); Stringfellow et al., *Gene* 166:73-76 (1995)). The *E. coli* W enzyme is encoded by hpaI (Prieto et al., *J. Bacteriol.* 178:111-120 (1996)). Genbank information related to these genes is summarized in Table 34 below.

TABLE 34

Gene	GI #	Accession No.	Organism
hpcH	633197	CAA87759.1	<i>Escherichia coli</i> C
hpaI	38112625	AAR11360.1	<i>Escherichia coli</i> W

[0119] In Step A of FIG. 3, pyruvate and malonate semialdehyde are joined by an aldehyde lyase to form 4-hydroxy-2-oxohexanedioate. An enzyme catalyzing this exact reaction has not been characterized to date. A similar reaction is catalyzed by 2-dehydro-3-deoxyglucarate aldolase (DDGA, EC 4.1.2.20), a type II aldolase that participates in the catabolic pathway for D-glucarate/galactarate utilization in *E. coli*. Tartronate semialdehyde, the natural substrate of DDGA, is similar in size and structure to malonate semialdehyde. This enzyme has a broad substrate specificity and has been shown to reversibly condense a wide range of aldehydes with pyruvate (Fish and Blumenthal, *Methods Enzymol.* 9:529-534 (1966)). The crystal structure of this enzyme has been determined and a catalytic mechanism indicated (Izard and Blackwell, *EMBO J.* 19:3849-3856 (2000)). Other DDGA enzymes are found in *Leptospira interrogans* (Li et al., *Acta Crystallogr. Sect. F. Struct. Biol. Cryst. Commun.* 62:1269-1270 (2006)) and *Sulfolobus solfataricus* (Buchanan et al., *Biochem. J.* 343 Pt 3:563-570 (1999)). The *S. solfataricus* enzyme is highly thermostable and was cloned and expressed in *E. coli* (Buchanan et al., supra). Genbank information related to these genes is summarized in Table 35 below.

TABLE 35

Gene	GI #	Accession No.	Organism
garL	1176153	P23522.2	<i>Escherichia coli</i>
LA_1624	24195249	AAN48823.1	<i>Leptospira interrogans</i>
AJ224174.1:1...885	2879782	CAA11866.1	<i>Sulfolobus solfataricus</i>

[0120] The pathways in FIGS. 2-4 employ numerous enzymes in the dehydratase class of enzymes (EC 4.1.2). Several reactions in FIGS. 2 and 3 undergo dehydration reactions similar to the dehydration of malate to fumarate, catalyzed by fumarate hydratase (EC 4.2.1.2). These transformations include the dehydration of 3-hydroxy-4-hexenedioate (FIG. 2, Steps E and Q and FIG. 3, Step H), 4-hydroxy-2-oxohexanedioate (FIG. 3, Step B), 2-hydroxy-4-hexenedioate (FIG. 3, Step D) and 2,4-dihydroxyadipate (FIG. 3, Steps F and G). Fumarate hydratase enzymes are exemplary enzymes for catalyzing these reactions. The *E. coli* fumarase encoded by fumC dehydrates a variety of alternate substrates including tartrate and threo-hydroxyaspartate (Teipel et al., *J. Biol. Chem.* 243:5684-5694 (1968)). A wealth of structural information is available for the *E. coli* enzyme and researchers have successfully engineered the enzyme to alter activity, inhibition and localization (Weaver, T., *Acta Crystallogr. D Biol. Crystallogr.* 61:1395-1401 (2005)). Exemplary fumarate hydratase enzymes are found in *Escherichia coli* (Estevez et al., *Protein Sci.* 11:1552-1557 (2002); Hong and Lee, *Biotechnol. Bioprocess Eng.* 9:252-255 (2006); Rose and Weaver, *Proc. Natl. Acad. Sci. U.S.A.* 101:3393-3397 (2004)); Agnihotri and Liu, *Bioorg. Med. Chem.* 11:9-20 (2003)), *Corynebacterium glutamicum* (Genda et al., *Biosci. Biotechnol. Biochem.* 71:1102-1109 (2006)), *Campylobacter*

jejuni (Smith and Gray, *Catalysis Letters* 6:195-199 (1990)), *Therms thermophilus* (Mizobata et al., *Arch. Biochem. Biophys.* 355:49-55 (1998)), and *Rattus norvegicus* (Kobayashi et al., *J. Biochem.* 89:1923-1931 (1981)). Genbank information related to these genes is summarized in Table 36 below.

TABLE 36

Gene	GI #	Accession No.	Organism
fumC	120601	P05042.1	<i>Escherichia coli</i> K12
fumC	39931596	Q8NRN8.1	<i>Corynebacterium glutamicum</i>
fumC	9789756	O69294.1	<i>Campylobacter jejuni</i>
fumC	75427690	P84127	<i>Therms thermophilus</i>
fumH	120605	P14408.1	<i>Rattus norvegicus</i>

[0121] Another enzyme for catalyzing these reactions is citramalate hydrolyase (EC 4.2.1.34), an enzyme that naturally dehydrates 2-methylmalate to mesaconate. This enzyme has been studied in *Methanocaldococcus jannaschii* in the context of the pyruvate pathway to 2-oxobutanoate, where it has been shown to have a broad substrate specificity (Drevland et al., *J. Bacteriol.* 189:4391-4400 (2007)). This enzyme activity was also detected in *Clostridium tetanomorphum*, *Morganella morganii*, *Citrobacter amalonaticus* where it is thought to participate in glutamate degradation (Kato and Asano *Arch. Microbiol.* 168:457-463 (1997)). The *M. jannaschii* protein sequence does not bear significant homology to genes in these organisms. Genbank information related to these genes is summarized in Table 37 below.

TABLE 37

Gene	GI #	Accession No.	Organism
leuD	3122345	Q58673.1	<i>Methanocaldococcus jannaschii</i>

[0122] The enzyme OHED hydratase (FIG. 4, Step B) participates in 4-hydroxyphenylacetic acid degradation, where it converts 2-oxo-hept-4-ene-1,7-dioate (OHED) to 2-oxo-4-hydroxy-hepta-1,7-dioate (HODH) using magnesium as a cofactor (Burks et al., *J. Am. Chem. Soc.* 120 (1998). OHED hydratase enzymes have been identified and characterized in *E. coli* C (Izumi et al., *J. Mol. Biol.* 370:899-911 (2007); Roper et al., *Gene* 156:47-51 (1995)) and *E. coli* W (Prieto et al., *J. Bacteriol.* 178:111-120 (1996)). Sequence comparison reveals homologs in a range of bacteria, plants and animals. Enzymes with highly similar sequences are contained in *Klebsiella pneumonia* (91% identity, eval=2e-138) and *Salmonella enterica* (91% identity, eval=4e-138), among others. Genbank information related to these genes is summarized in Table 38 below.

TABLE 38

Gene	GI #	Accession No.	Organism
hpcG	556840	CAA57202.1	<i>Escherichia coli</i> C
hpaH757830	CAA86044.1	Escherichia coli W	
hpaH	150958100	ABR80130.1	<i>Klebsiella pneumoniae</i>
Sari_01896	160865156	ABX21779.1	<i>Salmonella enterica</i>

[0123] Dehydration of 3-hydroxyadipyl-CoA to 2,3-dehydroadipyl-CoA (FIG. 2, Step L and FIG. 4, Step G) is catalyzed by an enzyme with enoyl-CoA hydratase activity. 3-Hydroxybutyryl-CoA dehydratase (EC 4.2.1.55), also called crotonase, is an enoyl-CoA hydratase that dehydrates

3-hydroxyisobutyryl-CoA to form crotonyl-CoA (FIG. 3, step 2). Crotonase enzymes are required for n-butanol formation in some organisms, particularly *Clostridial* species, and also comprise one step of the 3-hydroxypropionate/4-hydroxybutyrate cycle in thermoacidophilic Archaea of the genera *Sulfolobus*, *Acidianus*, and *Metallosphaera*. Exemplary genes encoding crotonase enzymes can be found in *C. acetobutylicum* (Atsumi et al., *Metab. Eng.* 10:305-211 (2008); Boynton et al., *J. Bacteriol.* 178:3015-3024 (1996)), *C. kluyveri* (Hillmer and Gottschalk, *FEBS Lett.* 21:351-354 (1972)), and *Metallosphaera sedula* (Berg et al., *Science* 318:1782-1786 (2007)) though the sequence of the latter gene is not known. Genbank information related to these genes is summarized in Table 39 below.

TABLE 39

Gene	GI #	Accession No.	Organism
crt	15895969	NP_349318.1	<i>Clostridium acetobutylicum</i>
crt1	153953091	YP_001393856.1	<i>Clostridium kluyveri</i>

[0124] Additional enoyl-CoA hydratases (EC 4.2.1.17) catalyze the dehydration of a range of 3-hydroxyacyl-CoA substrates (Agnihotri and Liu, *Bioorg. Med. Chem.* 11:9-20 (2003); Conrad et al., *J. Bacteriol.* 118:103-111 (1974); Roberts et al., *Arch. Microbiol.* 117:99-108 (1978)). The enoyl-CoA hydratase of *Pseudomonas putida*, encoded by ech, catalyzes the conversion of 3-hydroxybutyryl-CoA to crotonyl-CoA (Roberts et al., supra). Additional enoyl-CoA hydratase enzymes are phaA and phaB, of *P. putida*, and paaA and paaB from *P. fluorescens* (Olivera et al., *Proc. Natl. Acad. Sci. U.S.A.* 95:6419-6424 (1998)). The gene product of pimF in *Rhodopseudomonas palustris* is predicted to encode an enoyl-CoA hydratase that participates in pimeloyl-CoA degradation (Harrison and Harwood, *Microbiology* 151:727-736 (2005)). Lastly, a number of *Escherichia coli* genes have been shown to demonstrate enoyl-CoA hydratase functionality including maoC (Park and Lee, *Appl. Biochem. Biotechnol.* 113-116:335-346 (2004)), paaF (Ismail et al., *Eur. J. Biochem.* 270:3047-3054 (2003); Park and Lee, supra; Park and Yup, *Biotechnol. Bioeng* 86:681-686 (2004)) and paaG (Ismail et al., *Eur. J. Biochem.* 270:3047-3054 (2003); Park and Lee, supra; Park and Yup, *Biotechnol. Bioeng* 86:681-686 (2004)). Genbank information related to these genes is summarized in Table 40 below.

TABLE 40

Gene	GI #	Accession No.	Organism
ech	26990073	NP_745498.1	<i>Pseudomonas putida</i>
paaA	26990002	NP_745427.1	<i>Pseudomonas putida</i>
paaB	26990001	NP_745426.1	<i>Pseudomonas putida</i>
phaA	106636093	ABF82233.1	<i>Pseudomonas fluorescens</i>
phaB	106636094	ABF82234.1	<i>Pseudomonas fluorescens</i>
pimF	39650635	CAE29158	<i>Rhodopseudomonas palustris</i>
maoC	16129348	NP_415905.1	<i>Escherichia coli</i>
paaF	16129354	NP_415911.1	<i>Escherichia coli</i>
paaG	16129355	NP_415912.1	<i>Escherichia coli</i>

[0125] Alternatively, the *E. coli* gene products of fadA and fadB encode a multienzyme complex involved in fatty acid oxidation that exhibits enoyl-CoA hydratase activity (Nakahigashi and Inokuchi, *Nucleic Acids Res.* 18:4937 (1990); Yang, S. Y. *J. Bacteriol.* 173:7405-7406 (1991); Yang et al., *Biochemistry* 30:6788-6795 (1991)). Knocking out a nega-

tive regulator encoded by fadR can be utilized to activate the fadB gene product (Sato et al., *J. Biosci. Bioeng.* 103:38-44 (2007)). The fadI and fadJ genes encode similar functions and are naturally expressed under anaerobic conditions (Campbell et al., *Mol. Microbiol.* 47:793-805 (2003)). Genbank information related to these genes is summarized in Table 41 below.

TABLE 41

Gene	GI #	Accession No.	Organism
fadA	49176430	YP_026272.1	<i>Escherichia coli</i>
fadB	16131692	NP_418288.1	<i>Escherichia coli</i>
fadI	16130275	NP_416844.1	<i>Escherichia coli</i>
fadJ	16130274	NP_416843.1	<i>Escherichia coli</i>
fadR	16129150	NP_415705.1	<i>Escherichia coli</i>

[0126] An enzyme in the ammonia-lyase family is required to deaminate 3-amino-4-hexenedioate (FIG. 2, Steps G and S), 2-aminoadipate (FIG. 5, Step C) and 3-aminoadipate (FIG. 5, Step H). Enzymes catalyzing this exact transformation has not been identified. However the three substrates bear structural similarity to aspartate, the native substrate of aspartase (EC 4.3.1.1.). Aspartase is a widespread enzyme in microorganisms, and has been characterized extensively (Wakil et al., *J. Biol. Chem.* 207:631-638 (1954)). The *E. coli* enzyme has been shown to react with a variety of alternate substrates including aspartatephenylmethylester, asparagine, benzyl-aspartate and malate (Ma et al., *Ann N.Y. Acad. Sci.* 672:60-65 (1992)). In addition, directed evolution was been employed on this enzyme to alter substrate specificity (Asano et al., *Biomol. Eng.* 22:95-101 (2005)). The crystal structure of the *E. coli* aspartase, encoded by aspA, has been solved (Shi et al., *Biochemistry* 36:9136-9144 (1997)). Enzymes with aspartase functionality have also been characterized in *Haemophilus influenzae* (Sjostrom et al., *Biochim. Biophys. Acta* 1324:182-190 (1997)), *Pseudomonas fluorescens* (Takagi and Kisumi, *J. Bacteriol.* 161:1-6 (1985)), *Bacillus subtilis* (Sjostrom et al., supra) and *Serratia marcescens* (Takagi and Kisumi supra). Genbank information related to these genes is summarized in Table 42 below.

TABLE 42

Gene	GI #	Accession No.	Organism
aspA	90111690	NP_418562	<i>Escherichia coli</i>
aspA	1168534	P44324.1	<i>Haemophilus influenzae</i>
aspA	114273	P07346.1	<i>Pseudomonas fluorescens</i>
ansB	251757243	P26899.1	<i>Bacillus subtilis</i>
aspA	416661	P33109.1	<i>Serratia marcescens</i>

[0127] Another deaminase enzyme is 3-methylaspartase (EC 4.3.1.2). This enzyme, also known as beta-methylaspartase and 3-methylaspartate ammonia-lyase, naturally catalyzes the deamination of threo-3-methylaspartate to mesaconate. The 3-methylaspartase from *Clostridium tetanomorphum* has been cloned, functionally expressed in *E. coli*, and crystallized (Asuncion et al., *Acta Crystallogr. D. Biol. Crystallogr.* 57:731-733 (2001); Asuncion et al., *J. Biol. Chem.* 277:8306-8311 (2002); Botting et al. *Biochemistry* 27:2953-2955 (1988); Goda et al., *Biochemistry* 31:10747-10756 (1992)). In *Citrobacter amalonaticus*, this enzyme is encoded by BAA28709 (Kato and Asano, *Arch. Microbiol.* 168:457-463 (1997)). 3-methylaspartase has also been crystallized from *E. coli* YG1002 (Asano and Kato, *FEBS Micro-*

biol. Lett. 118:255-258 (1994)) although the protein sequence is not listed in public databases such as GenBank. Sequence homology can be used to identify additional genes, including CTC_02563 in *C. tetani* and ECs0761 in *Escherichia coli* O157:H7. Genbank information related to these genes is summarized in Table 43 below.

TABLE 43

Gene	GI #	Accession No.	Organism
mal	259429	AAB24070.1	<i>Clostridium tetanomorphum</i>
BAA28709	3184397	BAA28709.1	<i>Citrobacter amalonaticus</i>
CTC_02563	28212141	NP_783085.1	<i>Clostridium tetani</i>
ECs0761	13360220	BAB34184.1	<i>Escherichia coli</i> O157:H7

[0128] In FIG. 2 Step J, muconolactone is converted to muconate by muconate cycloisomerase. However, muconate cycloisomerase usually results in the formation of cis,cis-muconate, which may be difficult for the subsequent Diels-Alder chemistry. The cis, trans- or trans, trans-isomers are preferred. Therefore, the addition of a cis, trans isomerase may help to improve the yield of terephthalic acid. Enzymes for similar isomeric conversions include maleate cis,trans-isomerase (EC 5.2.1.1), maleylacetone cis-trans-isomerase (EC 5.2.1.2), and cis,trans-isomerase of unsaturated fatty acids (Cti).

[0129] Maleate cis, trans-isomerase (EC 5.2.1.1) catalyzes the conversion of maleic acid in cis formation to fumarate in trans formation (Scher and Jakoby, *J. Biol. Chem.* 244:1878-1882 (1969)). The *Alcaligenes faecalis* maiA gene product has been cloned and characterized (Hatakeyama et al., *Biochem. Biophys. Res. Commun.* 239:74-79 (1997)). Other maleate cis,trans-isomerases are available in *Serratia marcescens* (Hatakeyama et al., *Biosci. Biotechnol. Biochem.* 64:1477-1485 (2000)), *Ralstonia eutropha* and *Geobacillus stearothermophilus*. Genbank information related to these genes is summarized in Table 44 below.

TABLE 44

Gene	GI #	Accession No.	Organism
maiA	2575787	BAA23002	<i>Alcaligenes faecalis</i>
maiA	113866948	YP_725437	<i>Ralstonia eutropha</i> H16
maiA	4760466	BAA77296	<i>Geobacillus stearothermophilus</i>
maiA	8570038	BAA96747.1	<i>Serratia marcescens</i>

[0130] Maleylacetone cis,trans-isomerase (EC 5.2.1.2) catalyzes the conversion of 4-maleyl-acetoacetate to 4-fumaryl-acetylacetate, a cis to trans conversion. This enzyme is encoded by maiA in *Pseudomonas aeruginosa* Fernandez-Canon and Penalva, *J. Biol. Chem.* 273:329-337 (1998)) and *Vibrio cholera* (Seltzer, S., *J. Biol. Chem.* 248:215-222 (1973)). A similar enzyme was identified by sequence homology in *E. coli* O157. Genbank information related to these genes is summarized in Table 45 below.

TABLE 45

Gene	GI #	Accession No.	Organism
maiA	15597203	NP_250697	<i>Pseudomonas aeruginosa</i>
maiA	15641359	NP_230991	<i>Vibrio cholerae</i>
maiA	189355347	EDU73766	<i>Escherichia coli</i> O157

[0131] The cti gene product catalyzes the conversion of cis-unsaturated fatty acids (UFA) to trans-UFA. The enzyme has been characterized in *P. putida* (Junker and Ramos, *J. Bacteriol.* 181:5693-5700 (1999)). Similar enzymes are found in *Shewanella* sp. MR-4 and *Vibrio cholerae*. Genbank information related to these genes is summarized in Table 46 below.

TABLE 46

Gene	GI #	Accession No.	Organism
ct	i5257178	AAD41252	<i>Pseudomonas putida</i>
cti	113968844	YP_732637	<i>Shewanella</i> sp. MR-4
cti	229506276	ZP_04395785	<i>Vibrio cholerae</i>

[0132] The endocyclic migration of the double bond in the structure of β -keto adipate-enol-lactone to form muconolactone (FIG. 2, Step 1) is catalyzed by muconolactone isomerase (EC 5.3.3.4). Muconolactone isomerase also participates in the catechol branch of the β -keto adipate pathway to degrade aromatic compounds, at the reverse direction of Step G. Muconolactone isomerase is encoded by the catC gene. The *Pseudomonas putida* muconolactone isomerase was purified and partial amino acid sequences of cyanogen bromide fragments were determined (Meagher, R. B., *Biochim. Biophys. Acta* 494:33-47 (1997)). A DNA fragment carrying the catBCDE genes from *Acinetobacter calcoaceticus* was isolated by complementing *P. putida* mutants and the complemented activities were expressed constitutively in the recombinant *P. putida* strains (Shanley et al., *J. Bacteriol.* 165:557-563 (1986)). The *A. calcoaceticus* catBCDE genes were also expressed at high levels in *Escherichia coli* under the control of a lac promoter (Shanley et al., supra). The aniline-assimilating bacterium *Rhodococcus* sp. AN-22 CatC was purified to homogeneity and characterized as a homo-octamer with a molecular mass of 100 kDa (Matsumura et al., *Biochem. J.* 393:219-226 (2006)). The crystal structure of *P. putida* muconolactone isomerase was solved (Kattie et al., *J. Mol. Biol.* 205:557-571 (1989)). Genbank information related to these genes is summarized in Table 47 below.

TABLE 47

Gene	GI #	Accession No.	Organism
Q3LHT1	122612792	catC	<i>Rhodococcus</i> sp. AN-22
Q43932	5915883	catC	<i>Acinetobacter calcoaceticus</i>
Q9EV41	75464174	catC	<i>Ralstonia eutropha</i>
P00948	5921199	catC	<i>Pseudomonas putida</i>
Q9Z9Y5	75475019	catC	<i>Frateuria</i> species ANA-18

[0133] Lysine 2,3-aminomutase (EC 5.4.3.2) converts lysine to (3S)-3,6-diaminohexanoate (FIG. 5, Step E), shifting an amine group from the 2- to the 3-position. The enzyme is found in bacteria that ferment lysine to acetate and butyrate, including as *Fusobacterium nucleatum* (kamA) (Barker et al., *J. Bacteriol.* 152:201-207 (1982)) and *Clostridium subterminale* (kamA) (Chirpich et al., *J. Biol. Chem.* 245:1778-1789 (1970)). The enzyme from *Clostridium subterminale* has been crystallized (Lepore et al., *Proc. Natl. Acad. Sci. U.S.A.* 102:13819-13824 (2005)). An enzyme encoding this function is also encoded by yodO in *Bacillus subtilis* (Chen et al., *Biochem. J.* 348 Pt 3:539-549 (2000)). The enzyme utilizes pyridoxal 5'-phosphate as a cofactor, requires activation by S-Adenosylmethionine, and is stereoselective, reacting with

the only with L-lysine. Genbank information related to these genes is summarized in Table 48 below.

TABLE 48

Gene	GI #	Accession No.	Organism
yodO	4033499	O34676.1	<i>Bacillus subtilis</i>
kamA	75423266	Q9XBQ8.1	<i>Clostridium subterminale</i>
kamA	81485301	Q8RHX4	<i>Fusobacterium nucleatum</i> subsp. <i>nucleatum</i>

[0134] In Step H of FIG. 2, the ring opening reaction of muconolactone to form muconate is catalyzed by muconate cycloisomerase (EC 5.5.1.1). Muconate cycloisomerase naturally converts cis,cis-muconate to muconolactone in the catechol branch of the β -ketoadipate pathway to degrade aromatic compounds. This enzyme has not been shown to react with the trans,trans isomer. The muconate cycloisomerase reaction is reversible and is encoded by the catB gene. The *Pseudomonas putida* catB gene was cloned and sequenced (Aldrich et al., *Gene* 52:185-195 (1987)), the catB gene product was studied (Neidhart et al., *Nature* 347:692-694 (1990)) and its crystal structures were resolved (Helin et al., *J. Mol. Biol.* 254:918-941 (1995)). A DNA fragment carrying the catBCDE genes from *Acinetobacter calcoaceticus* was isolated by complementing *P. putida* mutants and the complemented activities were expressed constitutively in the recombinant *P. putida* strains (Shanley et al., *J. Bacteriol.* 165:557-563 (1986)). The *A. calcoaceticus* catBCDE genes were also expressed at high levels in *Escherichia coli* under the control of a lac promoter (Shanley et al., supra). The *Rhodococcus* sp. AN-22 CatB was purified to homogeneity and characterized as a monomer with a molecular mass of 44 kDa. The enzyme was activated by Mn^{2+} , Co^{2+} and Mg^{2+} (Matsumura et al., *Biochem. J.* 393:219-226 (2006)). Muconate cycloisomerases from other species, such as *Rhodococcus rhodochrous* N75, *Frateriia* species ANA-18, and *Trichosporon cutaneum* were also purified and studied (Cha and Bruce, *FEMS Microbiol. Lett.* 224:29-34 2003); Mazur et al., *Biochemistry* 33:1961-1970 (1994); Murakami et al., *Biosci Biotechnol. Biochem.* 62:1129-1133 (1998)). Genbank information related to these genes is summarized in Table 49 below.

TABLE 49

Gene	GI #	Accession No.	Organism
catB	P08310	115713	<i>Pseudomonas putida</i>
catB	Q43931	51704317	<i>Acinetobacter calcoaceticus</i>
catB	Q3LHT2	122612793	<i>Rhodococcus</i> sp. AN-22
catB	Q9Z9Y1	75424020	<i>Frateriia</i> species ANA-18
catB	P46057	1170967	<i>Trichosporon cutaneum</i>

[0135] The conversion of beta-ketoadipyl-CoA to beta-ketoadipate (FIG. 2, Step B) and 2,3-dehydroadipyl-CoA to 2,3-dehydroadipate (FIG. 2, Step M and FIG. 4, Step H) can be catalyzed by a CoA acid-thiol ligase or CoA synthetase in the 6.2.1 family of enzymes. Enzymes catalyzing these exact transformations have not been characterized to date; however, several enzymes with broad substrate specificities have been described in the literature. ADP-forming acetyl-CoA synthetase (ACD, EC 6.2.1.13) is an enzyme that couples the conversion of acyl-CoA esters to their corresponding acids with the concomitant synthesis of ATP. ACD I from *Archaeo-*

globus fulgidus, encoded by AF1211, was shown to operate on a variety of linear and branched-chain substrates including isobutyrate, isopentanoate, and fumarate (Musfeldt and Schonheit, *J. Bacteriol.* 184:636-644 (2002)). A second reversible ACD in *Archaeoglobus fulgidus*, encoded by AF1983, was also shown to have a broad substrate range with high activity on cyclic compounds phenylacetate and indoleacetate (Musfeldt and Schonheit, supra). The enzyme from *Haloarcula marismortui* (annotated as a succinyl-CoA synthetase) accepts propionate, butyrate, and branched-chain acids (isovalerate and isobutyrate) as substrates, and was shown to operate in the forward and reverse directions (Brasen and Schonheit, *Arch. Microbiol.* 182:277-287 (2004)). The ACD encoded by PAE3250 from hyperthermophilic crenarchaeon *Pyrobaculum aerophilum* showed the broadest substrate range of all characterized ACDs, reacting with acetyl-CoA, isobutyryl-CoA (preferred substrate) and phenylacetyl-CoA (Brasen and Schonheit, supra). Directed evolution or engineering can be used to modify this enzyme to operate at the physiological temperature of the host organism. The enzymes from *A. fulgidus*, *H. marismortui* and *P. aerophilum* have all been cloned, functionally expressed, and characterized in *E. coli* (Brasen and Schonheit, supra; Musfeldt and Schonheit, supra). An additional enzyme is encoded by sucCD in *E. coli*, which naturally catalyzes the formation of succinyl-CoA from succinate with the concomitant consumption of one ATP, a reaction which is reversible in vivo (Buck et al., *Biochemistry* 24:6245-6252 (1985)). Genbank information related to these genes is summarized in Table 50 below.

TABLE 50

Gene	GI #	Accession No.	Organism
AF1211	11498810	NP_070039.1	<i>Archaeoglobus fulgidus</i> DSM 4304
AF1983	11499565	NP_070807.1	<i>Archaeoglobus fulgidus</i> DSM 4304
scs	55377722	YP_135572.1	<i>Haloarcula marismortui</i>
PAE3250	18313937	NP_560604.1	<i>Pyrobaculum aerophilum</i> str. IM2
sucC	16128703	NP_415256.1	<i>Escherichia coli</i>
sucD	1786949	AAC73823.1	<i>Escherichia coli</i>

[0136] Another enzyme for this step is 6-carboxyhexanoate-CoA ligase, also known as pimeloyl-CoA ligase (EC 6.2.1.14), which naturally activates pimelate to pimeloyl-CoA during biotin biosynthesis in gram-positive bacteria. The enzyme from *Pseudomonas mendocina*, cloned into *E. coli*, was shown to accept the alternate substrates hexanedioate and nonanedioate (Binieda et al., *Biochem. J.* 340 Pt 3:793-801 (1999)). Other enzymes are found in *Bacillus subtilis* (Bower et al., *J. Bacteriol.* 178:4122-4130 (1996)) and *Lysinibacillus sphaericus* (formerly *Bacillus sphaericus*) (Ploux et al., *Biochem. J.* 287 Pt 3:685-690 (1992)). Genbank information related to these genes is summarized in Table 51 below.

TABLE 51

Gene	GI #	Accession No.	Organism
pauA	15596214	NP_249708.1	<i>Pseudomonas mendocina</i>
bioW	50812281	NP_390902.2	<i>Bacillus subtilis</i>
bioW	115012	P22822.1	<i>Lysinibacillus sphaericus</i>

[0137] Additional CoA-ligases include the rat dicarboxylate-CoA ligase for which the sequence is yet uncharacterized (Vamecq et al., *Biochem. J.* 230:683-693 (1985)), either of the two characterized phenylacetate-CoA ligases from *P. chrysogenum* (Lamas-Maceiras et al., *Biochem. J.* 395:147-155 (2006); Wang et al., *Biochem. Biophys. Res. Commun.* 360:453-458 (2007)) and the phenylacetate-CoA ligase from *Pseudomonas putida* (Martinez-Blanco et al., *J. Biol. Chem.* 265:7084-7090 (1990)). Acetoacetyl-CoA synthetases from *Mus musculus* (Hasegawa et al., *Biochim. Biophys. Acta* 1779:414-419 (2008)) and *Homo sapiens* (Ohgami et al., *Biochem. Pharmacol.* 65:989-994 (2003)) naturally catalyze the ATP-dependant conversion of acetoacetate into acetoacetyl-CoA. Genbank information related to these genes is summarized in Table 52 below.

TABLE 52

Gene	GI #	Accession No.	Organism
phl	77019264	CAJ15517.1	<i>Penicillium chrysogenum</i>
phlB	152002983	ABS19624.1	<i>Penicillium chrysogenum</i>
paaF	22711873	AAC24333.2	<i>Pseudomonas putida</i>
AACS	21313520	NP_084486.1	<i>Mus musculus</i>
AACS	31982927	NP_076417.2	<i>Homo sapiens</i>

[0138] In some embodiments, the present invention provides a semi-synthetic method for synthesizing terephthalate (PTA) that includes preparing muconic acid by culturing the above-described organisms, reacting the resultant muconic acid with acetylene to form a cyclohexadiene adduct (P1, FIG. 1), and oxidizing the cyclohexadiene adduct to form PTA. Semi-synthetic methods combine the biosynthetic preparation of advanced intermediates with conventional organic chemical reactions.

[0139] While the culturing of muconic acid is discussed further below, the Diels-Alder reaction conditions are detailed here. Diels-Alder reactions are widespread in the chemical industry and are known to those skilled in the art (Carruthers, W., *Some Modern Methods of Organic Synthesis*, Cambridge University Press (1986); Norton, J., *Chem. Review* 31:319-523 (1942); Sauer, J., *Angewandte Chemie* 6:16-33 (1967)). This class of pericyclic reactions is well-studied for its ability to generate cyclic compounds at low energetic cost. Diels-Alder reactions are thus an attractive and low-cost way of making a variety of pharmaceuticals and natural products.

[0140] In a Diels-Alder reaction, a conjugated diene or heterodiene reacts with an alkene, alkyne, or other unsaturated functional group, known as a dienophile, to form a six-membered ring. One aspect of the Diels-Alder reaction is that the two components usually have complementary electronic character, as determined by the energies of the highest occupied molecular orbital (HOMO) and lowest unoccupied molecular orbital (LUMO) of the diene and dienophile (Carruthers, W., *Some Modern Methods of Organic Synthesis*, Cambridge University Press (1986). In normal mode, the diene is electron-rich and the dienophile is electron-poor, although this is not always the case. The method of the present invention provides the opposite electronic configuration with an electron poor diene and a relatively electron rich dienophile, in what is termed an inverse electron demand Diels-Alder reaction. The main physical constraint for this type of reaction is that the conjugated diene must be able to adopt a

cisoid conformation for the reaction to proceed. A wide variety of substituted conjugated dienes and dienophiles are able to undergo this chemistry.

[0141] In the disclosed reaction of FIG. 1, muconate is the conjugated diene, and is beneficially in the trans,trans or cis,trans isomeric configuration for the reaction to proceed. The cis, cis isomer of muconate, prevalent in biological systems as a degradation product of catechol, is unlikely to adopt the required cisoid conformation due to steric hindrance of the carboxylic acid groups. The trans,trans isomer of muconate (shown in FIG. 1) is able to react in Diels-Alder reactions with a variety of dienes (Deno, N. C., *J. Am. Chem. Soc.* 72:4057-4059 (1950); Sauer, J., *Angewandte Chemie* 6:16-33 (1967)).

[0142] Acetylene serves as the dienophile in the production of PTA. Acetylene and substituted acetylene derivatives are well-known dienophiles ((Carruthers, W., *Some Modern Methods of Organic Synthesis*, Cambridge University Press (1986); U.S. Pat. No. 3,513,209, Clement, R. A.; Dai et al., *J. Am. Chem. Soc.* 129:645-657 (2007)). The addition of electron-withdrawing substituents increases reactivity in normal mode Diels Alder reactions; likewise, in the inverse electron demand, electron donating groups are employed to increase reactivity. At elevated temperatures, unsubstituted acetylene has been shown to react with butadiene and other substituted linear and cyclic dienes (U.S. Pat. No. 3,513,209, Clement, R. A.; Norton, J., *Chem. Review* 31:319-523 (1942); Vijaya et al., *J. Mol. Struct.* 589-590:291-299 (2002)).

[0143] Increased temperature can be used to perform the Diels-Alder reaction in FIG. 1. For example, the Diels-Alder reaction of acetylene with 1,3-butadiene to form 1,4-cyclohexadiene is performed in the range of 80-300° C. (U.S. Pat. No. 3,513,209, Clement, R. A. supra).

[0144] Other reaction conditions that have been shown to enhance the rate of Diels-Alder reactions include elevated pressure, the addition of a Lewis acid, and stoichiometric excess of acetylene. Elevated pressure up to 1000 atmospheres was shown to enhance the rate of 1,4-cyclohexadiene formation from butadiene and acetylene (U.S. Pat. No. 3,513,209, Clement, R. A.). Catalytic amounts of Lewis acids can also improve reaction rate (Nicolaou et al., *Angewandte Chemie* 41:1668-1698 (2002)). Some suitable Lewis acids include magnesium halides such as magnesium chloride, magnesium bromide or magnesium iodide or zinc halides such as zinc chloride, zinc bromide or zinc iodide. Stoichiometric excess of acetylene will aid in reducing formation of homopolymerization byproducts.

[0145] Oxidation of the Diels-Alder product, cyclohexa-2,5-diene-1,4-dicarboxylate (P1), to PTA can be accomplished in the presence or absence of catalyst under mild reaction conditions. The driving force for P1 oxidation is the formation of the aromatic ring of PTA. Precedence for the conversion of P1 to PTA in the absence of catalyst is the conversion of 1,4-cyclohexadiene to benzene in air (U.S. Pat. No. 3,513,209, Clement, R. A.). 1,4-Cyclohexadiene is also converted to benzene by catalysis, for example using transition metal complexes such as bis(arene)molybdenum(0) and bis(arene)chromium(0) (Fochi, G., *Organometallics* 7:225-2256 (1988)) or electroactive binuclear rhodium complexes (Smith and Gray, *Catalysis Letters* 6:195-199 (1990)).

[0146] In some embodiments, the method for synthesizing PTA includes isolating muconic acid from the culture broth prior to reacting with acetylene in the Diels-Alder reaction. This is particularly helpful since the Diels-Alder reaction, is

frequently done in the absence of a solvent, especially under thermal conditions. Isolation of muconic acid can involve various filtration and centrifugation techniques. Cells of the culture and other insoluble materials can be filtered via ultrafiltration and certain salts can be removed by nanofiltration. Because muconic acid is a diacid, standard extraction techniques can be employed that involve adjusting the pH. After removal of substantially all solids and salts, the muconic acid can be separated from water by removal of water with heating in vacuo, or by extraction at low pH. For example, following the addition of sulfuric acid or phosphoric acid to the fermentation broth in sufficient amounts (pH 3 or lower), the free carboxylate acid form of muconic acid precipitates out of solution (U.S. Pat. No. 4,608,338). In this form, muconic acid is readily separated from the aqueous solution by filtration or other conventional means.

[0147] In some embodiments, the muconic acid need not be isolated. Instead, the Diels-Alder reaction between muconic acid and acetylene can be performed in the culture broth. In such a case, the culture broth can be optionally filtered prior to adding acetylene.

[0148] The non-naturally occurring microbial organisms of the invention can be produced by introducing expressible nucleic acids encoding one or more of the enzymes or proteins participating in one or more muconate biosynthetic pathways. Depending on the host microbial organism chosen for biosynthesis, nucleic acids for some or all of a particular muconate biosynthetic pathway can be expressed. For example, if a chosen host is deficient in one or more enzymes or proteins for a desired biosynthetic pathway, then expressible nucleic acids for the deficient enzyme(s) or protein(s) are introduced into the host for subsequent exogenous expression. Alternatively, if the chosen host exhibits endogenous expression of some pathway genes, but is deficient in others, then an encoding nucleic acid is needed for the deficient enzyme(s) or protein(s) to achieve muconate biosynthesis. Thus, a non-naturally occurring microbial organism of the invention can be produced by introducing exogenous enzyme or protein activities to obtain a desired biosynthetic pathway or a desired biosynthetic pathway can be obtained by introducing one or more exogenous enzyme or protein activities that, together with one or more endogenous enzymes or proteins, produces a desired product such as muconate.

[0149] Depending on the muconate biosynthetic pathway constituents of a selected host microbial organism, the non-naturally occurring microbial organisms of the invention will include at least one exogenously expressed muconate pathway-encoding nucleic acid and up to all encoding nucleic acids for one or more muconate biosynthetic pathways. For example, muconate biosynthesis can be established in a host deficient in a pathway enzyme or protein through exogenous expression of the corresponding encoding nucleic acid. In a host deficient in all enzymes or proteins of a muconate pathway, exogenous expression of all enzyme or proteins in the pathway can be included, although it is understood that all enzymes or proteins of a pathway can be expressed even if the host contains at least one of the pathway enzymes or proteins. For example, exogenous expression of all enzymes or proteins in a pathway for production of muconate can be included, such as those shown in FIGS. 2-5.

[0150] Given the teachings and guidance provided herein, those skilled in the art will understand that the number of encoding nucleic acids to introduce in an expressible form will, at least, parallel the muconate pathway deficiencies of

the selected host microbial organism. Therefore, a non-naturally occurring microbial organism of the invention can have one, two, three, four, six, etc. up to all nucleic acids encoding the enzymes or proteins constituting a muconate biosynthetic pathway disclosed herein. In some embodiments, the non-naturally occurring microbial organisms also can include other genetic modifications that facilitate or optimize muconate biosynthesis or that confer other useful functions onto the host microbial organism. One such other functionality can include, for example, augmentation of the synthesis of one or more of the muconate pathway precursors such as succinyl-CoA.

[0151] Generally, a host microbial organism is selected such that it produces the precursor of a muconate pathway, either as a naturally produced molecule or as an engineered product that either provides de novo production of a desired precursor or increased production of a precursor naturally produced by the host microbial organism. For example, succinyl-CoA is produced naturally in a host organism such as *E. coli*. A host organism can be engineered to increase production of a precursor, as disclosed herein. In addition, a microbial organism that has been engineered to produce a desired precursor can be used as a host organism and further engineered to express enzymes or proteins of a muconate pathway.

[0152] In some embodiments, a non-naturally occurring microbial organism of the invention is generated from a host that contains the enzymatic capability to synthesize muconate. In this specific embodiment it can be useful to increase the synthesis or accumulation of a muconate pathway product to, for example, drive muconate pathway reactions toward muconate production. Increased synthesis or accumulation can be accomplished by, for example, overexpression of nucleic acids encoding one or more of the above-described muconate pathway enzymes or proteins. Overexpression of the enzyme or enzymes and/or protein or proteins of the muconate pathway can occur, for example, through exogenous expression of the endogenous gene or genes, or through exogenous expression of the heterologous gene or genes. Therefore, naturally occurring organisms can be readily generated to be non-naturally occurring microbial organisms of the invention, for example, producing muconate, through overexpression of one, two, three, four, five, six, that is, up to all nucleic acids encoding muconate biosynthetic pathway enzymes or proteins. In addition, a non-naturally occurring organism can be generated by mutagenesis of an endogenous gene that results in an increase in activity of an enzyme in the muconate biosynthetic pathway.

[0153] In particularly useful embodiments, exogenous expression of the encoding nucleic acids is employed. Exogenous expression confers the ability to custom tailor the expression and/or regulatory elements to the host and application to achieve a desired expression level that is controlled by the user. However, endogenous expression also can be utilized in other embodiments such as by removing a negative regulatory effector or induction of the gene's promoter when linked to an inducible promoter or other regulatory element. Thus, an endogenous gene having a naturally occurring inducible promoter can be up-regulated by providing the appropriate inducing agent, or the regulatory region of an endogenous gene can be engineered to incorporate an inducible regulatory element, thereby allowing the regulation of increased expression of an endogenous gene at a desired time. Similarly, an inducible promoter can be included as a regula-

tory element for an exogenous gene introduced into a non-naturally occurring microbial organism.

[0154] It is understood that, in methods of the invention, any of the one or more exogenous nucleic acids can be introduced into a microbial organism to produce a non-naturally occurring microbial organism of the invention. The nucleic acids can be introduced so as to confer, for example, a muconate biosynthetic pathway onto the microbial organism. Alternatively, encoding nucleic acids can be introduced to produce an intermediate microbial organism having the biosynthetic capability to catalyze some of the required reactions to confer muconate biosynthetic capability. For example, a non-naturally occurring microbial organism having a muconate biosynthetic pathway can comprise at least two exogenous nucleic acids encoding desired enzymes or proteins. Thus, it is understood that any combination of two or more enzymes or proteins of a biosynthetic pathway can be included in a non-naturally occurring microbial organism of the invention. Similarly, it is understood that any combination of three or more enzymes or proteins of a biosynthetic pathway can be included in a non-naturally occurring microbial organism of the invention and so forth, as desired, so long as the combination of enzymes and/or proteins of the desired biosynthetic pathway results in production of the corresponding desired product. Similarly, any combination of four, or more enzymes or proteins of a biosynthetic pathway as disclosed herein can be included in a non-naturally occurring microbial organism of the invention, as desired, so long as the combination of enzymes and/or proteins of the desired biosynthetic pathway results in production of the corresponding desired product.

[0155] In addition to the biosynthesis of muconate as described herein, the non-naturally occurring microbial organisms and methods of the invention also can be utilized in various combinations with each other and with other microbial organisms and methods well known in the art to achieve product biosynthesis by other routes. For example, one alternative to produce muconate other than use of the muconate producers is through addition of another microbial organism capable of converting a muconate pathway intermediate to muconate. One such procedure includes, for example, the fermentation of a microbial organism that produces a muconate pathway intermediate. The muconate pathway intermediate can then be used as a substrate for a second microbial organism that converts the muconate pathway intermediate to muconate. The muconate pathway intermediate can be added directly to another culture of the second organism or the original culture of the muconate pathway intermediate producers can be depleted of these microbial organisms by, for example, cell separation, and then subsequent addition of the second organism to the fermentation broth can be utilized to produce the final product without intermediate purification steps.

[0156] In other embodiments, the non-naturally occurring microbial organisms and methods of the invention can be assembled in a wide variety of subpathways to achieve biosynthesis of, for example, muconate. In these embodiments, biosynthetic pathways for a desired product of the invention can be segregated into different microbial organisms, and the different microbial organisms can be co-cultured to produce the final product. In such a biosynthetic scheme, the product of one microbial organism is the substrate for a second microbial organism until the final product is synthesized. For example, the biosynthesis of muconate can be accomplished by constructing a microbial organism that contains biosyn-

thetic pathways for conversion of one pathway intermediate to another pathway intermediate or the product. Alternatively, muconate also can be biosynthetically produced from microbial organisms through co-culture or co-fermentation using two organisms in the same vessel, where the first microbial organism produces a muconate intermediate and the second microbial organism converts the intermediate to muconate.

[0157] Given the teachings and guidance provided herein, those skilled in the art will understand that a wide variety of combinations and permutations exist for the non-naturally occurring microbial organisms and methods of the invention together with other microbial organisms, with the co-culture of other non-naturally occurring microbial organisms having subpathways and with combinations of other chemical and/or biochemical procedures well known in the art to produce muconate.

[0158] Sources of encoding nucleic acids for a muconate pathway enzyme or protein can include, for example, any species where the encoded gene product is capable of catalyzing the referenced reaction. Such species include both prokaryotic and eukaryotic organisms including, but not limited to, bacteria, including archaea and eubacteria, and eukaryotes, including yeast, plant, insect, animal, and mammal, including human. Exemplary species for such sources include, for example, *Escherichia coli*, as well as other exemplary species disclosed herein or available as source organisms for corresponding genes. However, with the complete genome sequence available for now more than 550 species (with more than half of these available on public databases such as the NCBI), including 395 microorganism genomes and a variety of yeast, fungi, plant, and mammalian genomes, the identification of genes encoding the requisite muconate biosynthetic activity for one or more genes in related or distant species, including for example, homologues, orthologs, paralogs and nonorthologous gene displacements of known genes, and the interchange of genetic alterations between organisms is routine and well known in the art. Accordingly, the metabolic alterations enabling biosynthesis of muconate described herein with reference to a particular organism such as *E. coli* can be readily applied to other microorganisms, including prokaryotic and eukaryotic organisms alike. Given the teachings and guidance provided herein, those skilled in the art will know that a metabolic alteration exemplified in one organism can be applied equally to other organisms.

[0159] In some instances, such as when an alternative muconate biosynthetic pathway exists in an unrelated species, muconate biosynthesis can be conferred onto the host species by, for example, exogenous expression of a paralog or paralogs from the unrelated species that catalyzes a similar, yet non-identical metabolic reaction to replace the referenced reaction. Because certain differences among metabolic networks exist between different organisms, those skilled in the art will understand that the actual gene usage between different organisms may differ. However, given the teachings and guidance provided herein, those skilled in the art also will understand that the teachings and methods of the invention can be applied to all microbial organisms using the cognate metabolic alterations to those exemplified herein to construct a microbial organism in a species of interest that will synthesize muconate.

[0160] Host microbial organisms can be selected from, and the non-naturally occurring microbial organisms generated in, for example, bacteria, yeast, fungus or any of a variety of

other microorganisms applicable to fermentation processes. Exemplary bacteria include species selected from *Escherichia coli*, *Klebsiella oxytoca*, *Anaerobiospirillum succiniciproducens*, *Actinobacillus succinogenes*, *Mannheimia succiniciproducens*, *Rhizobium etli*, *Bacillus subtilis*, *Corynebacterium glutamicum*, *Gluconobacter oxydans*, *Zymomonas mobilis*, *Lactococcus lactis*, *Lactobacillus plantarum*, *Streptomyces coelicolor*, *Clostridium acetobutylicum*, *Pseudomonas fluorescens*, and *Pseudomonas putida*. Exemplary yeasts or fungi include species selected from *Saccharomyces cerevisiae*, *Schizosaccharomyces pombe*, *Kluyveromyces lactis*, *Kluyveromyces marxianus*, *Aspergillus terreus*, *Aspergillus niger* and *Pichia pastoris*. *E. coli* is a particularly useful host organisms since it is a well characterized microbial organism suitable for genetic engineering. Other particularly useful host organisms include yeast such as *Saccharomyces cerevisiae*.

[0161] Methods for constructing and testing the expression levels of a non-naturally occurring muconate-producing host can be performed, for example, by recombinant and detection methods well known in the art. Such methods can be found described in, for example, Sambrook et al., *Molecular Cloning: A Laboratory Manual*, Third Ed., Cold Spring Harbor Laboratory, New York (2001); and Ausubel et al., *Current Protocols in Molecular Biology*, John Wiley and Sons, Baltimore, Md. (1999).

[0162] Exogenous nucleic acid sequences involved in a pathway for production of muconate can be introduced stably or transiently into a host cell using techniques well known in the art including, but not limited to, conjugation, electroporation, chemical transformation, transduction, transfection, and ultrasound transformation. For exogenous expression in *E. coli* or other prokaryotic cells, some nucleic acid sequences in the genes or cDNAs of eukaryotic nucleic acids can encode targeting signals such as an N-terminal mitochondrial or other targeting signal, which can be removed before transformation into prokaryotic host cells, if desired. For example, removal of a mitochondrial leader sequence led to increased expression in *E. coli* (Hoffmeister et al., *J. Biol. Chem.* 280:4329-4338 (2005)). For exogenous expression in yeast or other eukaryotic cells, genes can be expressed in the cytosol without the addition of leader sequence, or can be targeted to mitochondrion or other organelles, or targeted for secretion, by the addition of a suitable targeting sequence such as a mitochondrial targeting or secretion signal suitable for the host cells. Thus, it is understood that appropriate modifications to a nucleic acid sequence to remove or include a targeting sequence can be incorporated into an exogenous nucleic acid sequence to impart desirable properties. Furthermore, genes can be subjected to codon optimization with techniques well known in the art to achieve optimized expression of the proteins.

[0163] An expression vector or vectors can be constructed to include one or more muconate biosynthetic pathway encoding nucleic acids as exemplified herein operably linked to expression control sequences functional in the host organism. Expression vectors applicable for use in the microbial host organisms of the invention include, for example, plasmids, phage vectors, viral vectors, episomes and artificial chromosomes, including vectors and selection sequences or markers operable for stable integration into a host chromosome. Additionally, the expression vectors can include one or more selectable marker genes and appropriate expression control sequences. Selectable marker genes also can be

included that, for example, provide resistance to antibiotics or toxins, complement auxotrophic deficiencies, or supply critical nutrients not in the culture media. Expression control sequences can include constitutive and inducible promoters, transcription enhancers, transcription terminators, and the like which are well known in the art. When two or more exogenous encoding nucleic acids are to be co-expressed, both nucleic acids can be inserted, for example, into a single expression vector or in separate expression vectors. For single vector expression, the encoding nucleic acids can be operationally linked to one common expression control sequence or linked to different expression control sequences, such as one inducible promoter and one constitutive promoter. The transformation of exogenous nucleic acid sequences involved in a metabolic or synthetic pathway can be confirmed using methods well known in the art. Such methods include, for example, nucleic acid analysis such as Northern blots or polymerase chain reaction (PCR) amplification of mRNA, or immunoblotting for expression of gene products, or other suitable analytical methods to test the expression of an introduced nucleic acid sequence or its corresponding gene product. It is understood by those skilled in the art that the exogenous nucleic acid is expressed in a sufficient amount to produce the desired product, and it is further understood that expression levels can be optimized to obtain sufficient expression using methods well known in the art and as disclosed herein.

[0164] Directed evolution is a powerful approach that involves the introduction of mutations targeted to a specific gene in order to improve and/or alter the properties of an enzyme. Improved and/or altered enzymes can be identified through the development and implementation of sensitive high-throughput screening assays that allow the automated screening of many enzyme variants (e.g., $>10^4$). Iterative rounds of mutagenesis and screening typically are performed to afford an enzyme with optimized properties. Computational algorithms that can help to identify areas of the gene for mutagenesis also have been developed and can significantly reduce the number of enzyme variants that need to be generated and screened.

[0165] Numerous directed evolution technologies have been developed (for reviews, see Hibbert et al., *Biomol. Eng* 22:11-19 (2005); Huisman and Lalonde, In *Biocatalysis in the pharmaceutical and biotechnology industries* pgs. 717-742 (2007), Patel (ed.), CRC Press; Otten and Quax, *Biomol. Eng* 22:1-9 (2005); and Sen et al., *Appl Biochem. Biotechnol* 143:212-223 (2007)) to be effective at creating diverse variant libraries and these methods have been successfully applied to the improvement of a wide range of properties across many enzyme classes.

[0166] Enzyme characteristics that have been improved and/or altered by directed evolution technologies include, for example, selectivity/specificity—for conversion of non-natural substrates; temperature stability—for robust high temperature processing; pH stability—for bioprocessing under lower or higher pH conditions; substrate or product tolerance—so that high product titers can be achieved; binding (K_m)—broadens substrate binding to include non-natural substrates; inhibition (K_i)—to remove inhibition by products, substrates, or key intermediates; activity (kcat)—increases enzymatic reaction rates to achieve desired flux; expression levels—increases protein yields and overall pathway flux; oxygen stability—for operation of air sensitive enzymes

under aerobic conditions; and anaerobic activity—for operation of an aerobic enzyme in the absence of oxygen.

[0167] The following exemplary methods have been developed for the mutagenesis and diversification of genes to target desired properties of specific enzymes. Any of these can be used to alter/optimize activity of a decarboxylase enzyme.

[0168] EpPCR (Pritchard et al., *J Theor. Biol* 234:497-509 (2005)) introduces random point mutations by reducing the fidelity of DNA polymerase in PCR reactions by the addition of Mn^{2+} ions, by biasing dNTP concentrations, or by other conditional variations. The five step cloning process to confine the mutagenesis to the target gene of interest involves: 1) error-prone PCR amplification of the gene of interest; 2) restriction enzyme digestion; 3) gel purification of the desired DNA fragment; 4) ligation into a vector; 5) transformation of the gene variants into a suitable host and screening of the library for improved performance. This method can generate multiple mutations in a single gene simultaneously, which can be useful. A high number of mutants can be generated by EpPCR, so a high-throughput screening assay or a selection method (especially using robotics) is useful to identify those with desirable characteristics.

[0169] Error-prone Rolling Circle Amplification (epRCA) (Fujii et al., *Nucleic Acids Res* 32:e145 (2004); and Fujii et al., *Nat. Protoc.* 1:2493-2497 (2006)) has many of the same elements as epPCR except a whole circular plasmid is used as the template and random 6-mers with exonuclease resistant thiophosphate linkages on the last 2 nucleotides are used to amplify the plasmid followed by transformation into cells in which the plasmid is re-circularized at tandem repeats. Adjusting the Mn^{2+} concentration can vary the mutation rate somewhat. This technique uses a simple error-prone, single-step method to create a full copy of the plasmid with 3-4 mutations/kbp. No restriction enzyme digestion or specific primers are required. Additionally, this method is typically available as a kit.

[0170] DNA or Family Shuffling (Stemmer, *Proc. Natl. Acad. Sci. U.S.A.* 91:10747-10751 (1994); and Stemmer, *Nature* 370:389-391 (1994)) typically involves digestion of two or more variant genes with nucleases such as Dnase I or EndoV to generate a pool of random fragments that are reassembled by cycles of annealing and extension in the presence of DNA polymerase to create a library of chimeric genes. Fragments prime each other and recombination occurs when one copy primes another copy (template switch). This method can be used with >1 kbp DNA sequences. In addition to mutational recombinants created by fragment reassembly, this method introduces point mutations in the extension steps at a rate similar to error-prone PCR. The method can be used to remove deleterious, random and neutral mutations that might confer antigenicity.

[0171] Staggered Extension (StEP) (Zhao et al., *Nat. Biotechnol* 16:258-261 (1998)) entails template priming followed by repeated cycles of 2 step PCR with denaturation and very short duration of annealing/extension (as short as 5 sec). Growing fragments anneal to different templates and extend further, which is repeated until full-length sequences are made. Template switching means most resulting fragments have multiple parents. Combinations of low-fidelity polymerases (Taq and Mutazyme) reduce error-prone biases because of opposite mutational spectra.

[0172] In Random Priming Recombination (RPR) random sequence primers are used to generate many short DNA fragments complementary to different segments of the template.

(Shao et al., *Nucleic Acids Res* 26:681-683 (1998)) Base misincorporation and mispriming via epPCR give point mutations. Short DNA fragments prime one another based on homology and are recombined and reassembled into full-length by repeated thermocycling. Removal of templates prior to this step assures low parental recombinants. This method, like most others, can be performed over multiple iterations to evolve distinct properties. This technology avoids sequence bias, is independent of gene length, and requires very little parent DNA for the application.

[0173] In Heteroduplex Recombination linearized plasmid DNA is used to form heteroduplexes that are repaired by mismatch repair. (Volkov et al., *Nucleic Acids Res* 27:e18 (1999); and Volkov et al., *Methods Enzymol.* 328:456-463 (2000)) The mismatch repair step is at least somewhat mutagenic. Heteroduplexes transform more efficiently than linear homoduplexes. This method is suitable for large genes and whole operons.

[0174] Random Chimeragenesis on Transient Templates (RACHITT) (Coco et al., *Nat. Biotechnol* 19:354-359 (2001)) employs Dnase I fragmentation and size fractionation of ssDNA. Homologous fragments are hybridized in the absence of polymerase to a complementary ssDNA scaffold. Any overlapping unhybridized fragment ends are trimmed down by an exonuclease. Gaps between fragments are filled in, and then ligated to give a pool of full-length diverse strands hybridized to the scaffold (that contains U to preclude amplification). The scaffold then is destroyed and is replaced by a new strand complementary to the diverse strand by PCR amplification. The method involves one strand (scaffold) that is from only one parent while the priming fragments derive from other genes; the parent scaffold is selected against. Thus, no reannealing with parental fragments occurs. Overlapping fragments are trimmed with an exonuclease. Otherwise, this is conceptually similar to DNA shuffling and StEP. Therefore, there should be no siblings, few inactives, and no unshuffled parentals. This technique has advantages in that few or no parental genes are created and many more crossovers can result relative to standard DNA shuffling.

[0175] Recombined Extension on Truncated templates (RETT) entails template switching of unidirectionally growing strands from primers in the presence of unidirectional ssDNA fragments used as a pool of templates. (Lee et al., *J. Molec. Catalysis* 26:119-129 (2003)) No DNA endonucleases are used. Unidirectional ssDNA is made by DNA polymerase with random primers or serial deletion with exonuclease. Unidirectional ssDNA are only templates and not primers. Random priming and exonucleases don't introduce sequence bias as true of enzymatic cleavage of DNA shuffling/RACHITT. RETT can be easier to optimize than StEP because it uses normal PCR conditions instead of very short extensions. Recombination occurs as a component of the PCR steps—no direct shuffling. This method can also be more random than StEP due to the absence of pauses.

[0176] In Degenerate Oligonucleotide Gene Shuffling (DOGS) degenerate primers are used to control recombination between molecules; (Bergquist and Gibbs, *Methods Mol. Biol.* 352:191-204 (2007); Bergquist et al., *Biomol. Eng* 22:63-72 (2005); Gibbs et al., *Gene* 271:13-20 (2001)) this can be used to control the tendency of other methods such as DNA shuffling to regenerate parental genes. This method can be combined with random mutagenesis (epPCR) of selected gene segments. This can be a good method to block the reformation of parental sequences. No endonucleases are

needed. By adjusting input concentrations of segments made, one can bias towards a desired backbone. This method allows DNA shuffling from unrelated parents without restriction enzyme digests and allows a choice of random mutagenesis methods.

[0177] Incremental Truncation for the Creation of Hybrid Enzymes (ITCHY) creates a combinatorial library with 1 base pair deletions of a gene or gene fragment of interest. (Ostermeier et al., *Proc. Natl. Acad. Sci. U.S.A.* 96:3562-3567 (1999); and Ostermeier et al., *Nat. Biotechnol* 17:1205-1209 (1999)) Truncations are introduced in opposite direction on pieces of 2 different genes. These are ligated together and the fusions are cloned. This technique does not require homology between the 2 parental genes. When ITCHY is combined with DNA shuffling, the system is called SCRATCHY (see below). A major advantage of both is no need for homology between parental genes; for example, functional fusions between an *E. coli* and a human gene were created via ITCHY. When ITCHY libraries are made, all possible crossovers are captured.

[0178] Thio-Incremental Truncation for the Creation of Hybrid Enzymes (THIO-ITCHY) is similar to ITCHY except that phosphothioate dNTPs are used to generate truncations. (Lutz et al., *Nucleic Acids Res* 29:E16 (2001)) Relative to ITCHY, THIO-ITCHY can be easier to optimize, provide more reproducibility, and adjustability.

[0179] SCRATCHY combines two methods for recombining genes, ITCHY and DNA shuffling. (Lutz et al., *Proc. Natl. Acad. Sci. U.S.A.* 98:11248-11253 (2001)) SCRATCHY combines the best features of ITCHY and DNA shuffling. First, ITCHY is used to create a comprehensive set of fusions between fragments of genes in a DNA homology-independent fashion. This artificial family is then subjected to a DNA-shuffling step to augment the number of crossovers. Computational predictions can be used in optimization. SCRATCHY is more effective than DNA shuffling when sequence identity is below 80%.

[0180] In Random Drift Mutagenesis (RNDM) mutations made via epPCR followed by screening/selection for those retaining usable activity. (Bergquist et al., *Biomol. Eng* 22:63-72 (2005)) Then, these are used in DOGS to generate recombinants with fusions between multiple active mutants or between active mutants and some other desirable parent. Designed to promote isolation of neutral mutations; its purpose is to screen for retained catalytic activity whether or not this activity is higher or lower than in the original gene. RNDM is usable in high throughput assays when screening is capable of detecting activity above background. RNDM has been used as a front end to DOGS in generating diversity. The technique imposes a requirement for activity prior to shuffling or other subsequent steps; neutral drift libraries are indicated to result in higher/quicker improvements in activity from smaller libraries. Though published using epPCR, this could be applied to other large-scale mutagenesis methods.

[0181] Sequence Saturation Mutagenesis (SeSaM) is a random mutagenesis method that: 1) generates pool of random length fragments using random incorporation of a phosphothioate nucleotide and cleavage; this pool is used as a template to 2) extend in the presence of "universal" bases such as inosine; 3) replication of a inosine-containing complement gives random base incorporation and, consequently, mutagenesis. (Wong et al., *Biotechnol J.* 3:74-82 (2008); Wong et al., *Nucleic Acids Res* 32:e26 (2004); and Wong et al., *Anal. Biochem.* 341:187-189 (2005)) Using this technique

it can be possible to generate a large library of mutants within 2-3 days using simple methods. This technique is non-directed in comparison to the mutational bias of DNA polymerases. Differences in this approach makes this technique complementary (or an alternative) to epPCR.

[0182] In Synthetic Shuffling, overlapping oligonucleotides are designed to encode "all genetic diversity in targets" and allow a very high diversity for the shuffled progeny. (Ness et al., *Nat. Biotechnol* 20:1251-1255 (2002)) In this technique, one can design the fragments to be shuffled. This aids in increasing the resulting diversity of the progeny. One can design sequence/codon biases to make more distantly related sequences recombine at rates approaching those observed with more closely related sequences. Additionally, the technique does not require physically possessing the template genes.

[0183] Nucleotide Exchange and Excision Technology Next exploits a combination of dUTP incorporation followed by treatment with uracil DNA glycosylase and then piperidine to perform endpoint DNA fragmentation. (Muller et al., *Nucleic Acids Res* 33:e117 (2005)) The gene is reassembled using internal PCR primer extension with proofreading polymerase. The sizes for shuffling are directly controllable using varying dUPT::dTTP ratios. This is an end point reaction using simple methods for uracil incorporation and cleavage. Other nucleotide analogs, such as 8-oxo-guanine, can be used with this method. Additionally, the technique works well with very short fragments (86 bp) and has a low error rate. The chemical cleavage of DNA used in this technique results in very few unshuffled clones.

[0184] In Sequence Homology-Independent Protein Recombination (SHIPREC) a linker is used to facilitate fusion between two distantly/unrelated genes. Nuclease treatment is used to generate a range of chimeras between the two genes. These fusions result in libraries of single-crossover hybrids. (Sieber et al., *Nat. Biotechnol* 19:456-460 (2001)) This produces a limited type of shuffling and a separate process is required for mutagenesis. In addition, since no homology is needed this technique can create a library of chimeras with varying fractions of each of the two unrelated parent genes. SHIPREC was tested with a heme-binding domain of a bacterial CP450 fused to N-terminal regions of a mammalian CP450; this produced mammalian activity in a more soluble enzyme.

[0185] In Gene Site Saturation Mutagenesis™ (GSSM™) the starting materials are a supercoiled dsDNA plasmid containing an insert and two primers which are degenerate at the desired site of mutations. (Kretz et al., *Methods Enzymol.* 388:3-11 (2004)) Primers carrying the mutation of interest, anneal to the same sequence on opposite strands of DNA. The mutation is typically in the middle of the primer and flanked on each side by ~20 nucleotides of correct sequence. The sequence in the primer is NNN or NNK (coding) and MNN (noncoding) (N=all 4, K=G, T, M=A, C). After extension, DpnI is used to digest dam-methylated DNA to eliminate the wild-type template. This technique explores all possible amino acid substitutions at a given locus (i.e., one codon). The technique facilitates the generation of all possible replacements at a single-site with no nonsense codons and results in equal to near-equal representation of most possible alleles. This technique does not require prior knowledge of the structure, mechanism, or domains of the target enzyme. If followed by shuffling or Gene Reassembly, this technology creates a diverse library of recombinants containing all possible

combinations of single-site up-mutations. The utility of this technology combination has been demonstrated for the successful evolution of over 50 different enzymes, and also for more than one property in a given enzyme.

[0186] Combinatorial Cassette Mutagenesis (CCM) involves the use of short oligonucleotide cassettes to replace limited regions with a large number of possible amino acid sequence alterations. (Reidhaar-Olson et al. *Methods Enzymol.* 208:564-586 (1991); and Reidhaar-Olson et al. *Science* 241:53-57 (1988)) Simultaneous substitutions at two or three sites are possible using this technique. Additionally, the method tests a large multiplicity of possible sequence changes at a limited range of sites. This technique has been used to explore the information content of the lambda repressor DNA-binding domain.

[0187] Combinatorial Multiple Cassette Mutagenesis (CMCM) is essentially similar to CCM except it is employed as part of a larger program: 1) Use of epPCR at high mutation rate to 2) ID hot spots and hot regions and then 3) extension by CMCM to cover a defined region of protein sequence space. (Reetz et al., *Angew. Chem. Int. Ed Engl.* 40:3589-3591 (2001).) As with CCM, this method can test virtually all possible alterations over a target region. If used along with methods to create random mutations and shuffled genes, it provides an excellent means of generating diverse, shuffled proteins. This approach was successful in increasing, by 51-fold, the enantioselectivity of an enzyme.

[0188] In the Mutator Strains technique conditional ts mutator plasmids allow increases of 20- to 4000-X in random and natural mutation frequency during selection and block accumulation of deleterious mutations when selection is not required. (Selifonova et al., *Appl Environ Microbiol* 67:3645-3649 (2001)) This technology is based on a plasmid-derived mutD5 gene, which encodes a mutant subunit of DNA polymerase III. This subunit binds to endogenous DNA polymerase III and compromises the proofreading ability of polymerase III in any strain that harbors the plasmid. A broad-spectrum of base substitutions and frameshift mutations occur. In order for effective use, the mutator plasmid should be removed once the desired phenotype is achieved; this is accomplished through a temperature sensitive origin of replication, which allows for plasmid curing at 41° C. It should be noted that mutator strains have been explored for quite some time (e.g., see Low et al., *J. Mol. Biol.* 260:359-3680 (1996)). In this technique very high spontaneous mutation rates are observed. The conditional property minimizes non-desired background mutations. This technology could be combined with adaptive evolution to enhance mutagenesis rates and more rapidly achieve desired phenotypes.

[0189] “Look-Through Mutagenesis (LTM) is a multidimensional mutagenesis method that assesses and optimizes combinatorial mutations of selected amino acids.” (Rajpal et al., *Proc Natl Acad Sci U.S.A.* 102:8466-8471 (2005)) Rather than saturating each site with all possible amino acid changes, a set of nine is chosen to cover the range of amino acid R-group chemistry. Fewer changes per site allows multiple sites to be subjected to this type of mutagenesis. A >800-fold increase in binding affinity for an antibody from low nanomolar to picomolar has been achieved through this method. This is a rational approach to minimize the number of random combinations and can increase the ability to find improved traits by greatly decreasing the numbers of clones to be screened. This has been applied to antibody engineering,

specifically to increase the binding affinity and/or reduce dissociation. The technique can be combined with either screens or selections.

[0190] Gene Reassembly is a DNA shuffling method that can be applied to multiple genes at one time or to creating a large library of chimeras (multiple mutations) of a single gene. (Tunable GeneReassembly™ (TGR™) Technology supplied by Verenum Corporation) Typically this technology is used in combination with ultra-high-throughput screening to query the represented sequence space for desired improvements. This technique allows multiple gene recombination independent of homology. The exact number and position of cross-over events can be pre-determined using fragments designed via bioinformatic analysis. This technology leads to a very high level of diversity with virtually no parental gene reformation and a low level of inactive genes. Combined with GSSM™, a large range of mutations can be tested for improved activity. The method allows “blending” and “fine tuning” of DNA shuffling, e.g. codon usage can be optimized.

[0191] In Silico Protein Design Automation (PDA) is an optimization algorithm that anchors the structurally defined protein backbone possessing a particular fold, and searches sequence space for amino acid substitutions that can stabilize the fold and overall protein energetics. (Hayes et al., *Proc Natl Acad Sci U.S.A.* 99:15926-15931 (2002)) This technology uses in silico structure-based entropy predictions in order to search for structural tolerance toward protein amino acid variations. Statistical mechanics is applied to calculate coupling interactions at each position. Structural tolerance toward amino acid substitution is a measure of coupling. Ultimately, this technology is designed to yield desired modifications of protein properties while maintaining the integrity of structural characteristics. The method computationally assesses and allows filtering of a very large number of possible sequence variants (10^{50}). The choice of sequence variants to test is related to predictions based on the most favorable thermodynamics. Ostensibly only stability or properties that are linked to stability can be effectively addressed with this technology. The method has been successfully used in some therapeutic proteins, especially in engineering immunoglobulins. In silico predictions avoid testing extraordinarily large numbers of potential variants. Predictions based on existing three-dimensional structures are more likely to succeed than predictions based on hypothetical structures. This technology can readily predict and allow targeted screening of multiple simultaneous mutations, something not possible with purely experimental technologies due to exponential increases in numbers.

[0192] Iterative Saturation Mutagenesis (ISM) involves: 1) use knowledge of structure/function to choose a likely site for enzyme improvement; 2) saturation mutagenesis at chosen site using Stratagene QuikChange (or other suitable means); 3) screen/select for desired properties; and 4) with improved clone(s), start over at another site and continue repeating. (Reetz et al., *Nat. Protoc.* 2:891-903 (2007); and Reetz et al., *Angew. Chem. Int. Ed Engl.* 45:7745-7751 (2006)) This is a proven methodology, which assures all possible replacements at a given position are made for screening/selection.

[0193] Any of the aforementioned methods for mutagenesis can be used alone or in any combination. Additionally, any one or combination of the directed evolution methods can be used in conjunction with adaptive evolution techniques.

[0194] The present invention provides a method for producing muconate that includes culturing a non-naturally

occurring microbial organism having a muconate pathway. The pathway includes at least one exogenous nucleic acid encoding a muconate pathway enzyme expressed in a sufficient amount to produce muconate, under conditions and for a sufficient period of time to produce muconate. The muconate pathway includes an enzyme selected from the group consisting of a beta-ketothiolase, a beta-ketoadipyl-CoA hydrolase, a beta-ketoadipyl-CoA transferase, a beta-ketoadipyl-CoA ligase, a 2-fumarylacetate reductase, a 2-fumarylacetate dehydrogenase, a trans-3-hydroxy-4-hexendioate dehydratase, a 2-fumarylacetate aminotransferase, a 2-fumarylacetate aminating oxidoreductase, a trans-3-amino-4-hexenoate deaminase, a beta-ketoadipate enol-lactone hydrolase, a muconolactone isomerase, a muconate cycloisomerase, a beta-ketoadipyl-CoA dehydrogenase, a 3-hydroxyadipyl-CoA dehydratase, a 2,3-dehydroadipyl-CoA transferase, a 2,3-dehydroadipyl-CoA hydrolase, a 2,3-dehydroadipyl-CoA ligase, a muconate reductase, a 2-maleylacetate reductase, a 2-maleylacetate dehydrogenase, a cis-3-hydroxy-4-hexendioate dehydratase, a 2-maleylacetate aminoatransferase, a 2-maleylacetate aminating oxidoreductase, a cis-3-amino-4-hexendioate deaminase, and a muconate cis/trans isomerase.

[0195] In some embodiments, the muconate pathway includes, a set of muconate pathway enzymes such as those exemplified in FIG. 2; the set of muconate pathway enzymes are selected from the group consisting of:

[0196] A) (1) beta-ketothiolase, (2) an enzyme selected from beta-ketoadipyl-CoA hydrolase, beta-ketoadipyl-CoA transferase, and beta-ketoadipyl-CoA ligase, (3) beta-ketoadipate enol-lactone hydrolase, (4) muconolactone isomerase, (5) muconate cycloisomerase, and (6) muconate cis/trans isomerase;

[0197] B) (1) beta-ketothiolase, (2) an enzyme selected from beta-ketoadipyl-CoA hydrolase, beta-ketoadipyl-CoA transferase and beta-ketoadipyl-CoA ligase, (3) 2-maleylacetate reductase, (4) 2-maleylacetate dehydrogenase, (5) cis-3-hydroxy-4-hexendioate dehydratase, and (6) muconate cis/trans isomerase;

[0198] C) (1) beta-ketothiolase, (2) an enzyme selected from beta-ketoadipyl-CoA hydrolase, beta-ketoadipyl-CoA transferase and beta-ketoadipyl-CoA ligase, (3) 2-maleylacetate reductase, (4) an enzyme selected from 2-maleylacetate aminotransferase and 2-maleylacetate aminating oxidoreductase, (5) cis-3-amino-4-hexenoate deaminase, and (6) muconate cis/trans isomerase;

[0199] D) (1) beta-ketothiolase, (2) beta-ketoadipyl-CoA dehydrogenase, (3) 3-hydroxyadipyl-CoA dehydratase, (4) an enzyme selected from 2,3-dehydroadipyl-CoA transferase, 2,3-dehydroadipyl-CoA hydrolase and 2,3-dehydroadipyl-CoA ligase, and (5) muconate reductase;

[0200] E) (1) beta-ketothiolase, (2) an enzyme selected from beta-ketoadipyl-CoA hydrolase, beta-ketoadipyl-CoA transferase and beta-ketoadipyl-CoA ligase, (3) 2-fumarylacetate reductase, (4) 2-fumarylacetate dehydrogenase, and (5) trans-3-hydroxy-4-hexendioate dehydratase;

[0201] F) (1) beta-ketothiolase, (2) an enzyme selected from beta-ketoadipyl-CoA hydrolase, beta-ketoadipyl-CoA transferase and beta-ketoadipyl-CoA ligase, (3) 2-fumarylacetate reductase, (4) an enzyme selected from 2-fumarylacetate aminotransferase and 2-fumarylacetate aminating oxidoreductase, and (5) trans-3-amino-4-hexenoate deaminase.

[0202] In some embodiments, the present invention provides a method for producing muconate that includes cultur-

ing a non-naturally occurring microbial organism having a muconate pathway. The pathway comprising at least one exogenous nucleic acid encoding a muconate pathway enzyme expressed in a sufficient amount to produce muconate, under conditions and for a sufficient period of time to produce muconate. The muconate pathway includes a 4-hydroxy-2-ketovalerate aldolase, a 2-oxopentenoate hydratase, a 4-oxalocrotonate dehydrogenase, a 2-hydroxy-4-hexenedioate dehydratase, a 4-hydroxy-2-oxohexanedioate oxidoreductase, a 2,4-dihydroxyadipate dehydratase (acting on 2-hydroxy), a 2,4-dihydroxyadipate dehydratase (acting on 4-hydroxyl group) and a 3-hydroxy-4-hexenedioate dehydratase.

[0203] In some embodiments, the muconate pathway includes, a set of muconate pathway enzymes such as those exemplified in FIG. 3; the set of muconate pathway enzymes are selected from the group consisting of:

[0204] A) (1) 4-hydroxy-2-ketovalerate aldolase, (2) 2-oxopentenoate hydratase, (3) 4-oxalocrotonate dehydrogenase, (4) 2-hydroxy-4-hexenedioate dehydratase;

[0205] B) (1) 4-hydroxy-2-ketovalerate aldolase, (2) 4-hydroxy-2-oxohexanedioate oxidoreductase, (3) 2,4-dihydroxyadipate dehydratase (acting on 2-hydroxy), (4) 3-hydroxy-4-hexenedioate dehydratase; and

[0206] C) (1) 4-hydroxy-2-ketovalerate aldolase, (2) 4-hydroxy-2-oxohexanedioate oxidoreductase, (3) 2,4-dihydroxyadipate dehydratase (acting on 4-hydroxyl group), (4) 2-hydroxy-4-hexenedioate dehydratase.

[0207] In some embodiments, the present invention provides a method for producing muconate that includes culturing a non-naturally occurring microbial organism having a muconate pathway. The pathway includes at least one exogenous nucleic acid encoding a muconate pathway enzyme expressed in a sufficient amount to produce muconate, under conditions and for a sufficient period of time to produce muconate. The muconate pathway includes an enzyme selected from the group consisting of an HODH aldolase, an OHED hydratase, an OHED decarboxylase, an HODH formate-lyase, an HODH dehydrogenase, an OHED formate-lyase, an OHED dehydrogenase, a 6-OHE dehydrogenase, a 3-hydroxyadipyl-CoA dehydratase, a 2,3-dehydroadipyl-CoA hydrolase, a 2,3-dehydroadipyl-CoA transferase, a 2,3-dehydroadipyl-CoA ligase, and a muconate reductase.

[0208] In some embodiments, the muconate pathway includes, a set of muconate pathway enzymes such as those exemplified in FIG. 4; the set of muconate pathway enzymes are selected from the group consisting of:

[0209] A) (1) HODH aldolase, (2) OHED hydratase, (3) OHED decarboxylase, (4) 6-OHE dehydrogenase, and (5) muconate reductase;

[0210] B) (1) HODH aldolase, (2) OHED hydratase, (3) an enzyme selected from OHED formate-lyase and OHED dehydrogenase, (4) an enzyme selected from 2,3-dehydroadipyl-CoA hydrolase, 2,3-dehydroadipyl-CoA transferase and 2,3-dehydroadipyl-CoA ligase, and (5) muconate reductase; and

[0211] C) (1) HODH aldolase, (2) an enzyme selected from HODH formate-lyase and HODH dehydrogenase, (3) 3-hydroxyadipyl-CoA dehydratase, (4) an enzyme selected from 2,3-dehydroadipyl-CoA hydrolase, 2,3-dehydroadipyl-CoA transferase and 2,3-dehydroadipyl-CoA ligase, and (5) muconate reductase.

[0212] In some embodiments, the present invention provides a method for producing muconate that includes cultur-

ing a non-naturally occurring microbial organism having a muconate pathway. The pathway includes at least one exogenous nucleic acid encoding a muconate pathway enzyme expressed in a sufficient amount to produce muconate, under conditions and for a sufficient period of time to produce muconate. The muconate pathway includes an enzyme selected from the group consisting of a lysine aminotransferase, a lysine aminating oxidoreductase, a 2-aminoadipate semialdehyde dehydrogenase, a 2-aminoadipate deaminase, a muconate reductase, a lysine-2,3-aminomutase, a 3,6-diaminohexanoate aminotransferase, a 3,6-diaminohexanoate aminating oxidoreductase, a 3-aminoadipate semialdehyde dehydrogenase, and a 3-aminoadipate deaminase.

[0213] In some embodiments, the muconate pathway includes, a set of muconate pathway enzymes such as those exemplified in FIG. 5; the set of muconate pathway enzymes are selected from the group consisting of:

[0214] A) (1) lysine aminotransferase, (2) lysine aminating oxidoreductase, (3) 2-aminoadipate semialdehyde dehydrogenase, (4) 2-aminoadipate deaminase, and (5) muconate reductase

[0215] B) (1) lysine-2,3-aminomutase, (2) 3,6-diaminohexanoate aminotransferase, (3) 3,6-diaminohexanoate aminating oxidoreductase, (4) 3-aminoadipate semialdehyde dehydrogenase, (5) 3-aminoadipate deaminase, and (6) muconate reductase.

[0216] In some embodiments, the foregoing non-naturally occurring microbial organism can be cultured in a substantially anaerobic culture medium.

[0217] Suitable purification and/or assays to test for the production of muconate can be performed using well known methods. Suitable replicates such as triplicate cultures can be grown for each engineered strain to be tested. For example, product and byproduct formation in the engineered production host can be monitored. The final product and intermediates, and other organic compounds, can be analyzed by methods such as HPLC (High Performance Liquid Chromatography), GC-MS (Gas Chromatography-Mass Spectroscopy) and LC-MS (Liquid Chromatography-Mass Spectroscopy) or other suitable analytical methods using routine procedures well known in the art. The release of product in the fermentation broth can also be tested with the culture supernatant. Byproducts and residual glucose can be quantified by HPLC using, for example, a refractive index detector for glucose and alcohols, and a UV detector for organic acids (Lin et al., *Biotechnol. Bioeng.* 90:775-779 (2005)), or other suitable assay and detection methods well known in the art. The individual enzyme or protein activities from the exogenous DNA sequences can also be assayed using methods well known in the art. For example, a spectrophotometric assay for succinyl-CoA:3-ketoacid-CoA transferase (FIG. 2, Step B) entails measuring the change in the absorbance corresponding to the product CoA molecule (i.e., succinyl-CoA) in the presence of the enzyme extract when supplied with succinate and β -ketoadipyl-CoA (Corthesy-Theulaz et al., *J. Biol. Chem.*, 272(41) (1997)). Succinyl-CoA can alternatively be measured in the presence of excess hydroxylamine by complexing the succinohydroxamic acid formed to ferric salts as referred to in (Corthesy-Theulaz et al., *J. Biol. Chem.* 272(41) (1997)). The specific activity of muconate reductase can be assayed in the reductive direction using a colorimetric assay adapted from the literature (Durre et al., *FEMS Microbiol. Rev.* 17:251-262 (1995); Palosaari et al., *J. Bacteriol.* 170:2971-2976 (1988); Welch et al., *Arch. Biochem. Biophys.*

273:309-318 (1989)). In this assay, the substrates muconate and NADH are added to cell extracts in a buffered solution, and the oxidation of NADH is followed by reading absorbance at 340 nM at regular intervals. The resulting slope of the reduction in absorbance at 340 nM per minute, along with the molar extinction coefficient of NADH at 340 nM (6000) and the protein concentration of the extract, can be used to determine the specific activity of muconate reductase.

[0218] The muconate can be separated from other components in the culture using a variety of methods well known in the art, as briefly described above. Such separation methods include, for example, extraction procedures as well as methods that include continuous liquid-liquid extraction, pervaporation, membrane filtration, membrane separation, reverse osmosis, electrodialysis, distillation, crystallization, centrifugation, extractive filtration, ion exchange chromatography, size exclusion chromatography, adsorption chromatography, and ultrafiltration. All of the above methods are well known in the art.

[0219] Any of the non-naturally occurring microbial organisms described herein can be cultured to produce and/or secrete the biosynthetic products of the invention. For example, the muconate producers can be cultured for the biosynthetic production of muconate.

[0220] For the production of muconate, the recombinant strains are cultured in a medium with carbon source and other essential nutrients. It is highly desirable to maintain anaerobic conditions in the fermenter to reduce the cost of the overall process. Such conditions can be obtained, for example, by first sparging the medium with nitrogen and then sealing the flasks with a septum and crimp-cap. For strains where growth is not observed anaerobically, microaerobic conditions can be applied by perforating the septum with a small hole for limited aeration. Exemplary anaerobic conditions have been described previously and are well-known in the art. Exemplary aerobic and anaerobic conditions are described, for example, in U.S. patent application Ser. No. 11/891,602, filed Aug. 10, 2007. Fermentations can be performed in a batch, fed-batch or continuous manner, as disclosed herein.

[0221] If desired, the pH of the medium can be maintained at a desired pH, in particular neutral pH, such as a pH of around 7 by addition of a base, such as NaOH or other bases, or acid, as needed to maintain the culture medium at a desirable pH. The growth rate can be determined by measuring optical density using a spectrophotometer (600 nm), and the glucose uptake rate by monitoring carbon source depletion over time. NOTE—Ideally this process would operate at low pH using an organisms that tolerates pH levels in the range 2-4.

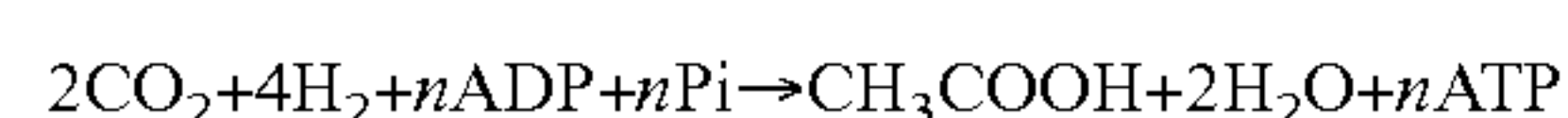
[0222] The growth medium can include, for example, any carbohydrate source which can supply a source of carbon to the non-naturally occurring microorganism. Such sources include, for example, sugars such as glucose, xylose, arabinose, galactose, mannose, fructose, sucrose and starch. Other sources of carbohydrate include, for example, renewable feedstocks and biomass. Exemplary types of biomasses that can be used as feedstocks in the methods of the invention include cellulosic biomass, hemicellulosic biomass and lignin feedstocks or portions of feedstocks. Such biomass feedstocks contain, for example, carbohydrate substrates useful as carbon sources such as glucose, xylose, arabinose, galactose, mannose, fructose and starch. Given the teachings and guidance provided herein, those skilled in the art will understand that renewable feedstocks and biomass other than those

exemplified above also can be used for culturing the microbial organisms of the invention for the production of muconate.

[0223] In addition to renewable feedstocks such as those exemplified above, the muconate microbial organisms of the invention also can be modified for growth on syngas as its source of carbon. In this specific embodiment, one or more proteins or enzymes are expressed in the muconate producing organisms to provide a metabolic pathway for utilization of syngas or other gaseous carbon source.

[0224] Synthesis gas, also known as syngas or producer gas, is the major product of gasification of coal and of carbonaceous materials such as biomass materials, including agricultural crops and residues. Syngas is a mixture primarily of H_2 and CO and can be obtained from the gasification of any organic feedstock, including but not limited to coal, coal oil, natural gas, biomass, and waste organic matter. Gasification is generally carried out under a high fuel to oxygen ratio. Although largely H_2 and CO, syngas can also include CO_2 and other gases in smaller quantities. Thus, synthesis gas provides a cost effective source of gaseous carbon such as CO and, additionally, CO_2 .

[0225] The Wood-Ljungdahl pathway catalyzes the conversion of CO and H_2 to acetyl-CoA and other products such as acetate. Organisms capable of utilizing CO and syngas also generally have the capability of utilizing CO_2 and CO_2/H_2 mixtures through the same basic set of enzymes and transformations encompassed by the Wood-Ljungdahl pathway. H_2 -dependent conversion of CO_2 to acetate by microorganisms was recognized long before it was revealed that CO also could be used by the same organisms and that the same pathways were involved. Many acetogens have been shown to grow in the presence of CO_2 and produce compounds such as acetate as long as hydrogen is present to supply the necessary reducing equivalents (see for example, Drake, *Acetogenesis*, pp. 3-60 Chapman and Hall, New York, (1994)). This can be summarized by the following equation:



[0226] Hence, non-naturally occurring microorganisms possessing the Wood-Ljungdahl pathway can utilize CO_2 and H_2 mixtures as well for the production of acetyl-CoA and other desired products.

[0227] The Wood-Ljungdahl pathway is well known in the art and consists of 12 reactions which can be separated into two branches: (1) methyl branch and (2) carbonyl branch. The methyl branch converts syngas to methyl-tetrahydrofolate (methyl-THF) whereas the carbonyl branch converts methyl-THF to acetyl-CoA. The reactions in the methyl branch are catalyzed in order by the following enzymes or proteins: ferredoxin oxidoreductase, formate dehydrogenase, formyltetrahydrofolate synthetase, methenyltetrahydrofolate cyclodehydratase, methylenetetrahydrofolate dehydrogenase and methylenetetrahydrofolate reductase. The reactions in the carbonyl branch are catalyzed in order by the following enzymes or proteins: methyltetrahydrofolate:corrinoid protein methyltransferase (for example, AcsE), corrinoid iron-sulfur protein, nickel-protein assembly protein (for example, AcsF), ferredoxin, acetyl-CoA synthase, carbon monoxide dehydrogenase and nickel-protein assembly protein (for example, CooC). Following the teachings and guidance provided herein for introducing a sufficient number of encoding nucleic acids to generate a muconate pathway, those skilled in the art will understand that the same engineering design also

can be performed with respect to introducing at least the nucleic acids encoding the Wood-Ljungdahl enzymes or proteins absent in the host organism. Therefore, introduction of one or more encoding nucleic acids into the microbial organisms of the invention such that the modified organism contains the complete Wood-Ljungdahl pathway will confer syngas utilization ability.

[0228] Accordingly, given the teachings and guidance provided herein, those skilled in the art will understand that a non-naturally occurring microbial organism can be produced that secretes the biosynthesized compounds of the invention when grown on a carbon source such as a carbohydrate. Such compounds include, for example, muconate and any of the intermediate metabolites in the muconate pathway. All that is required is to engineer in one or more of the required enzyme or protein activities to achieve biosynthesis of the desired compound or intermediate including, for example, inclusion of some or all of the muconate biosynthetic pathways. Accordingly, the invention provides a non-naturally occurring microbial organism that produces and/or secretes muconate when grown on a carbohydrate or other carbon source and produces and/or secretes any of the intermediate metabolites shown in the muconate pathway when grown on a carbohydrate or other carbon source. The muconate producing microbial organisms of the invention can initiate synthesis from an intermediate, such as any of the intermediates shown in FIGS. 2-5.

[0229] The non-naturally occurring microbial organisms of the invention are constructed using methods well known in the art as exemplified herein to exogenously express at least one nucleic acid encoding a muconate pathway enzyme or protein in sufficient amounts to produce muconate. It is understood that the microbial organisms of the invention are cultured under conditions sufficient to produce muconate. Following the teachings and guidance provided herein, the non-naturally occurring microbial organisms of the invention can achieve biosynthesis of muconate resulting in intracellular concentrations between about 0.1-200 mM or more. Generally, the intracellular concentration of muconate is between about 3-150 mM, particularly between about 5-200 mM and more particularly between about 8-150 mM, including about 10 mM, 50 mM, 75 mM, 100 mM, or more. Intracellular concentrations between and above each of these exemplary ranges also can be achieved from the non-naturally occurring microbial organisms of the invention.

[0230] In some embodiments, culture conditions include anaerobic or substantially anaerobic growth or maintenance conditions. Exemplary anaerobic conditions have been described previously and are well known in the art. Exemplary anaerobic conditions for fermentation processes are described herein and are described, for example, in U.S. patent application Ser. No. 11/891,602, filed Aug. 10, 2007. Any of these conditions can be employed with the non-naturally occurring microbial organisms as well as other anaerobic conditions well known in the art. Under such anaerobic conditions, the muconate producers can synthesize muconate at intracellular concentrations of 5-10 mM or more as well as all other concentrations exemplified herein. It is understood that, even though the above description refers to intracellular concentrations, muconate producing microbial organisms can produce muconate intracellularly and/or secrete the product into the culture medium.

[0231] The culture conditions can include, for example, liquid culture procedures as well as fermentation and other

large scale culture procedures. As described herein, particularly useful yields of the biosynthetic products of the invention can be obtained under anaerobic or substantially anaerobic culture conditions.

[0232] As described herein, one exemplary growth condition for achieving biosynthesis of muconate includes anaerobic culture or fermentation conditions. In certain embodiments, the non-naturally occurring microbial organisms of the invention can be sustained, cultured or fermented under anaerobic or substantially anaerobic conditions. Briefly, anaerobic conditions refers to an environment devoid of oxygen. Substantially anaerobic conditions include, for example, a culture, batch fermentation or continuous fermentation such that the dissolved oxygen concentration in the medium remains between 0 and 10% of saturation. Substantially anaerobic conditions also includes growing or resting cells in liquid medium or on solid agar inside a sealed chamber maintained with an atmosphere of less than 1% oxygen. The percent of oxygen can be maintained by, for example, sparging the culture with an N_2/CO_2 mixture or other suitable non-oxygen gas or gases.

[0233] The culture conditions described herein can be scaled up and grown continuously for manufacturing of muconate. Exemplary growth procedures include, for example, fed-batch fermentation and batch separation; fed-batch fermentation and continuous separation, or continuous fermentation and continuous separation. All of these processes are well known in the art. Fermentation procedures are particularly useful for the biosynthetic production of commercial quantities of muconate. Generally, and as with non-continuous culture procedures, the continuous and/or near-continuous production of muconate will include culturing a non-naturally occurring muconate producing organism of the invention in sufficient nutrients and medium to sustain and/or nearly sustain growth in an exponential phase. Continuous culture under such conditions can be include, for example, 1 day, 2, 3, 4, 5, 6 or 7 days or more. Additionally, continuous culture can include 1 week, 2, 3, 4 or 5 or more weeks and up to several months. Alternatively, organisms of the invention can be cultured for hours, if suitable for a particular application. It is to be understood that the continuous and/or near-continuous culture conditions also can include all time intervals in between these exemplary periods. It is further understood that the time of culturing the microbial organism of the invention is for a sufficient period of time to produce a sufficient amount of product for a desired purpose.

[0234] Fermentation procedures are well known in the art. Briefly, fermentation for the biosynthetic production of muconate can be utilized in, for example, fed-batch fermentation and batch separation; fed-batch fermentation and continuous separation, or continuous fermentation and continuous separation. Examples of batch and continuous fermentation procedures are well known in the art.

[0235] In addition to the above fermentation procedures using the muconate producers of the invention for continuous production of substantial quantities of muconate, the muconate producers also can be, for example, simultaneously subjected to chemical synthesis procedures to convert the product to other compounds or the product can be separated from the fermentation culture and sequentially subjected to chemical conversion to convert the product to other compounds, if desired.

[0236] In addition to the above procedures, growth condition for achieving biosynthesis of muconate can include the

addition of an osmoprotectant to the culturing conditions. In certain embodiments, the non-naturally occurring microbial organisms of the invention can be sustained, cultured or fermented as described above in the presence of an osmoprotectant. Briefly, an osmoprotectant means a compound that acts as an osmolyte and helps a microbial organism as described herein survive osmotic stress. Osmoprotectants include, but are not limited to, betaines, amino acids, and the sugar trehalose. Non-limiting examples of such are glycine betaine, praline betaine, dimethylthetin, dimethylsulfoniopropionate, 3-dimethylsulfonio-2-methylpropionate, pipercolic acid, dimethylsulfonioacetate, choline, L-carnitine and ectoine. In one aspect, the osmoprotectant is glycine betaine. It is understood to one of ordinary skill in the art that the amount and type of osmoprotectant suitable for protecting a microbial organism described herein from osmotic stress will depend on the microbial organism used. The amount of osmoprotectant in the culturing conditions can be, for example, no more than about 0.1 mM, no more than about 0.5 mM, no more than about 1.0 mM, no more than about 1.5 mM, no more than about 2.0 mM, no more than about 2.5 mM, no more than about 3.0 mM, no more than about 5.0 mM, no more than about 7.0 mM, no more than about 10 mM, no more than about 50 mM, no more than about 100 mM or no more than about 500 mM.

[0237] To generate better producers, metabolic modeling can be utilized to optimize growth conditions. Modeling can also be used to design gene knockouts that additionally optimize utilization of the pathway (see, for example, U.S. patent publications US 2002/0012939, US 2003/0224363, US 2004/0029149, US 2004/0072723, US 2003/0059792, US 2002/0168654 and US 2004/0009466, and U.S. Pat. No. 7,127,379). Modeling analysis allows reliable predictions of the effects on cell growth of shifting the metabolism towards more efficient production of muconate.

[0238] One computational method for identifying and designing metabolic alterations favoring biosynthesis of a desired product is the OptKnock computational framework (Burgard et al., *Biotechnol. Bioeng.* 84:647-657 (2003)). OptKnock is a metabolic modeling and simulation program that suggests gene deletion or disruption strategies that result in genetically stable microorganisms which overproduce the target product. Specifically, the framework examines the complete metabolic and/or biochemical network of a microorganism in order to suggest genetic manipulations that force the desired biochemical to become an obligatory byproduct of cell growth. By coupling biochemical production with cell growth through strategically placed gene deletions or other functional gene disruption, the growth selection pressures imposed on the engineered strains after long periods of time in a bioreactor lead to improvements in performance as a result of the compulsory growth-coupled biochemical production. Lastly, when gene deletions are constructed there is a negligible possibility of the designed strains reverting to their wild-type states because the genes selected by OptKnock are to be completely removed from the genome. Therefore, this computational methodology can be used to either identify alternative pathways that lead to biosynthesis of a desired product or used in connection with the non-naturally occurring microbial organisms for further optimization of biosynthesis of a desired product.

[0239] Briefly, OptKnock is a term used herein to refer to a computational method and system for modeling cellular metabolism. The OptKnock program relates to a framework

of models and methods that incorporate particular constraints into flux balance analysis (FBA) models. These constraints include, for example, qualitative kinetic information, qualitative regulatory information, and/or DNA microarray experimental data. OptKnock also computes solutions to various metabolic problems by, for example, tightening the flux boundaries derived through flux balance models and subsequently probing the performance limits of metabolic networks in the presence of gene additions or deletions. OptKnock computational framework allows the construction of model formulations that enable an effective query of the performance limits of metabolic networks and provides methods for solving the resulting mixed-integer linear programming problems. The metabolic modeling and simulation methods referred to herein as OptKnock are described in, for example, U.S. publication 2002/0168654, filed Jan. 10, 2002, in International Patent No. PCT/US02/00660, filed Jan. 10, 2002, and U.S. publication 2009/0047719, filed Aug. 10, 2007.

[0240] Another computational method for identifying and designing metabolic alterations favoring biosynthetic production of a product is a metabolic modeling and simulation system termed SimPheny®. This computational method and system is described in, for example, U.S. publication 2003/0233218, filed Jun. 14, 2002, and in International Patent Application No. PCT/US03/18838, filed Jun. 13, 2003. SimPheny® is a computational system that can be used to produce a network model in silico and to simulate the flux of mass, energy or charge through the chemical reactions of a biological system to define a solution space that contains any and all possible functionalities of the chemical reactions in the system, thereby determining a range of allowed activities for the biological system. This approach is referred to as constraints-based modeling because the solution space is defined by constraints such as the known stoichiometry of the included reactions as well as reaction thermodynamic and capacity constraints associated with maximum fluxes through reactions. The space defined by these constraints can be interrogated to determine the phenotypic capabilities and behavior of the biological system or of its biochemical components.

[0241] These computational approaches are consistent with biological realities because biological systems are flexible and can reach the same result in many different ways. Biological systems are designed through evolutionary mechanisms that have been restricted by fundamental constraints that all living systems must face. Therefore, constraints-based modeling strategy embraces these general realities. Further, the ability to continuously impose further restrictions on a network model via the tightening of constraints results in a reduction in the size of the solution space, thereby enhancing the precision with which physiological performance or phenotype can be predicted.

[0242] Given the teachings and guidance provided herein, those skilled in the art will be able to apply various computational frameworks for metabolic modeling and simulation to design and implement biosynthesis of a desired compound in host microbial organisms. Such metabolic modeling and simulation methods include, for example, the computational systems exemplified above as SimPheny® and OptKnock. For illustration of the invention, some methods are described herein with reference to the OptKnock computation framework for modeling and simulation. Those skilled in the art will know how to apply the identification, design and implementation of the metabolic alterations using OptKnock to any

of such other metabolic modeling and simulation computational frameworks and methods well known in the art.

[0243] The methods described above will provide one set of metabolic reactions to disrupt. Elimination of each reaction within the set or metabolic modification can result in a desired product as an obligatory product during the growth phase of the organism. Because the reactions are known, a solution to the bilevel OptKnock problem also will provide the associated gene or genes encoding one or more enzymes that catalyze each reaction within the set of reactions. Identification of a set of reactions and their corresponding genes encoding the enzymes participating in each reaction is generally an automated process, accomplished through correlation of the reactions with a reaction database having a relationship between enzymes and encoding genes.

[0244] Once identified, the set of reactions that are to be disrupted in order to achieve production of a desired product are implemented in the target cell or organism by functional disruption of at least one gene encoding each metabolic reaction within the set. One particularly useful means to achieve functional disruption of the reaction set is by deletion of each encoding gene. However, in some instances, it can be beneficial to disrupt the reaction by other genetic aberrations including, for example, mutation, deletion of regulatory regions such as promoters or cis binding sites for regulatory factors, or by truncation of the coding sequence at any of a number of locations. These latter aberrations, resulting in less than total deletion of the gene set can be useful, for example, when rapid assessments of the coupling of a product are desired or when genetic reversion is less likely to occur.

[0245] To identify additional productive solutions to the above described bilevel OptKnock problem which lead to further sets of reactions to disrupt or metabolic modifications that can result in the biosynthesis, including growth-coupled biosynthesis of a desired product, an optimization method, termed integer cuts, can be implemented. This method proceeds by iteratively solving the OptKnock problem exemplified above with the incorporation of an additional constraint referred to as an integer cut at each iteration. Integer cut constraints effectively prevent the solution procedure from choosing the exact same set of reactions identified in any previous iteration that obligatorily couples product biosynthesis to growth. For example, if a previously identified growth-coupled metabolic modification specifies reactions 1, 2, and 3 for disruption, then the following constraint prevents the same reactions from being simultaneously considered in subsequent solutions. The integer cut method is well known in the art and can be found described in, for example, Burgard et al., *Biotechnol. Prog.* 17:791-797 (2001). As with all methods described herein with reference to their use in combination with the OptKnock computational framework for metabolic modeling and simulation, the integer cut method of reducing redundancy in iterative computational analysis also can be applied with other computational frameworks well known in the art including, for example, SimPheny®.

[0246] The methods exemplified herein allow the construction of cells and organisms that biosynthetically produce a desired product, including the obligatory coupling of production of a target biochemical product to growth of the cell or organism engineered to harbor the identified genetic alterations. Therefore, the computational methods described herein allow the identification and implementation of metabolic modifications that are identified by an in silico method selected from OptKnock or SimPheny®. The set of metabolic

modifications can include, for example, addition of one or more biosynthetic pathway enzymes and/or functional disruption of one or more metabolic reactions including, for example, disruption by gene deletion.

[0247] As discussed above, the OptKnock methodology was developed on the premise that mutant microbial networks can be evolved towards their computationally predicted maximum-growth phenotypes when subjected to long periods of growth selection. In other words, the approach leverages an organism's ability to self-optimize under selective pressures. The OptKnock framework allows for the exhaustive enumeration of gene deletion combinations that force a coupling between biochemical production and cell growth based on network stoichiometry. The identification of optimal gene/reaction knockouts requires the solution of a bilevel optimization problem that chooses the set of active reactions such that an optimal growth solution for the resulting network overproduces the biochemical of interest (Burgard et al., *Bio-technol. Bioeng.* 84:647-657 (2003)).

[0248] An in silico stoichiometric model of *E. coli* metabolism can be employed to identify essential genes for metabolic pathways as exemplified previously and described in, for example, U.S. patent publications US 2002/0012939, US 2003/0224363, US 2004/0029149, US 2004/0072723, US 2003/0059792, US 2002/0168654 and US 2004/0009466, and in U.S. Pat. No. 7,127,379. As disclosed herein, the OptKnock mathematical framework can be applied to pinpoint gene deletions leading to the growth-coupled production of a desired product. Further, the solution of the bilevel OptKnock problem provides only one set of deletions. To enumerate all meaningful solutions, that is, all sets of knockouts leading to growth-coupled production formation, an optimization technique, termed integer cuts, can be implemented. This entails iteratively solving the OptKnock prob-

lem with the incorporation of an additional constraint referred to as an integer cut at each iteration, as discussed above.

[0249] It is understood that modifications which do not substantially affect the activity of the various embodiments of this invention are also provided within the definition of the invention provided herein. Accordingly, the following examples are intended to illustrate but not limit the present invention.

Example I

Demonstration of Enzyme Activity for Condensing Succinyl-CoA and Acetyl-CoA to Form β -ketoadipyl-CoA

[0250] This Example shows the identification of enzymes for the formation of beta-ketoadipyl-CoA from succinyl-CoA and acetyl-CoA.

[0251] Several β -ketothiolase enzymes have been shown to break β -ketoadipyl-CoA into acetyl-CoA and succinyl-CoA. For example, the gene products encoded by *pcaF* in *Pseudomonas* strain B13 (Kaschabek et al., *J. Bacteriol.* 184 (1): 207-15 (2002)), *phaD* in *Pseudomonas putida* U (Olivera et al., *Proc Natl Acad Sci USA*, 95(11), 6419-24 (1998)), *paaE* in *Pseudomonas fluorescens* ST (Di Gennaro et al., *Arch Microbiol.* 188(2), 117-25 (2007)), and *paaJ* from *E. coli* (Nogales et al., *Microbiology* 153(Pt 2), 357-65 (2007)) catalyze the conversion of 3-oxoadipyl-CoA into succinyl-CoA and acetyl-CoA during the degradation of aromatic compounds such as phenylacetate or styrene. To confirm that β -ketothiolase enzymes exhibit condensation activity, several thiolases (Table 53) were cloned into a derivative of pZE13 (Lutz et al., *Nucleic Acids Res.* 29(18), 3873-81 (2001)), which results in the clones having a carboxy-terminal 6xHis tag.

TABLE 53

Cloned Thiolases					
Enzyme	Species template	Gene Length	5' PRIMER	3' PRIMER	ORF SEQ
beta-ketothiolase	<i>Ralstonia eutropha</i> H16	bktB 1185	ATGACGCGTG AAGTGGTAGT GGTAAG	GATACGCTCGA AGATGGCGG	atgacgcgtgaagtggtagtggtgaagcgggtgtccg taccgcgatcgggaccttggcgccagcctgaagg atgtggcaccggcggagctggcgccactgggtgggtg cgcgaggcgctggcgcgcgcgaggtgtcgggcga cgatgtcgggccacgtggtattcggaacgtgatcc agaccgagccgcgcgacatgtatctgggcgcgcgtc gcgcccggtcaacggcggggtgacgatcaacgcccc cgcgctgaccgtgaaccgcctgtgcggctcgggcc tgcaggccattgtcagcgccgcgcagaccatcctg ctgggcgataccgacgtcgccatcgggcgggcgcg ggaaagcatgagccgcgcacgtacctggcgccgg cagcgcgctggggcgacgcgatggcgacgcgggc ctggtcgacatgatgctgggtgcgctgcacgatcc cttccatcgcatccacatgggcgtgaccgccgaga atgtcgccaaggaatcgcacatctcgcgcgcgag caggacgaggccgcgctggaatcgaccgcgcgcgc ttcggcagcgatcaaggccggctacttcaaggacc agatcgctccgggtggtgagcaaggccgcaagggc gacgtgaccttcgacaccgacgagcagctgcgcca tgacgccaccatcgacgacatgaccaagctcaggc cggctctcgtaaggaaaacggcacgggtcacggcc ggcaatgcctcgggcctgaacgacgcgcgcgcgc gggtggtgatgatggagcgcggaagccgagcgccg cggcctgaagccgctggcccgccctggtgtcgtacg gccatgccggcgtggaccgaaggccatgggcacg ggcccggtgccggcgacgaagatcgcgctggagcg cgccggcctgcagggtgtcgacactggacgtgatcg

TABLE 53-continued

Cloned Thiolases						
Enzyme	Species template	Gene	Length	5' PRIMER	3' PRIMER	ORF SEQ
						aagccaacgaagcctttgccgcacaggcgctgcgcc gtgaccaaggcgctcggctctggaccggccaaggt taaccggaacggctcgggcatctcgctgggccacc cgatcggcgccaaccgggtgccctgatcacgggtgaag gcgctgcatgagctgaaccgcgtgcaggggccgcta cgcgctggtgacgatgtgcatcggcggcgggcagg gcattgccgccatcttcgagcgtatctga
2- Methylacetoa cetyl-CoA Thiolase (branched chain?)	<i>Mus musculus</i>	ACA T1	1215	ATGGAAGTAA GATGCCTGGA ACGAAG	CAGCTTCTCAAT CAGCAGGGC	atggaagtaagatgcctggaacgaagttatgcatc caaaccactttgaatgaagtggttatagtaagtg ctataagaactcccatctggatccttcctgggcagc cttgccctctcagccggccactaaacttgggtactgc tgcaattcaggggagccattgagaaggcagggtatc caaaagaagaagtgaaggaagtctacatgggcaat gtcatccaaggggggtgaaggacaggccccctaccag gcaagcaacactgggcgcagggtttacctatctcca ctccatgcaccacagtaaacagggtttgtgcttca ggaatgaaagccatcatgatggcctctcaaagtct tatgtgtggacatcaggatgtgatgggtggcaggcg ggatggagagcatgtccaatgtcccatacgtaatg agcagaggagcaacaccatattggtgggtaaaact tgaagacctgattgtaaaagacgggctaactgatg tctacaataaaaattcatatgggttaactgtgctgag aatactgcaaagaagatgaatatctcacggcagga acaggatacgtacgctctcagctcttacaccagaa gtaaagaagcgtgggacgcagggaagtttgccagt gagattactcccatcaccatctcagtgaaggtaa accagatgtggtgggtgaaagaagatgaagaataca agcgtgttgactttagtaaagtgccaaagctcaag accgtgttccagaaagaaaatggcacaataacagc tgccaatgccagcacactgaacgatggagcagctg ctctgggttctcatgactgcagaggcagcccagagg ctcaatgttaagccattggcacgaattgcagcatt tgctgatgctgccgtagacccattgattttccac ttgcgcctgcatatgccgtacctaaggttcttaaa tatgcaggactgaaaaaagaagacattgccatgtg ggaagttaatgaagcattcagtgtggttgtgctag ccaacattaaaatgctggagattgacccccaaaaa gtaaatatccacggaggagctgtttctctgggccca tccaattgggatgtctggagcccggttggtgttc atatggctcatgccctgaagccaggagagttcggt ctggctagtatttgcaacggaggaggaggtgcttc cgccctgctgattgagaagctgtag
2- Methylacetoa cetyl-CoA Thiolase (branched chain?)	<i>Pseudomonas putida</i> (KT2440)	fadAx1194		ATGACCCTCG CCAATGACCC	GTACAGGCATTC AACAGCCATGG	atgaccctcgccaatgaccccatcggtatcgtcag cgccgtgcgcacgcccattgggcgggttgaggggcg acctcaagagcctgactgcgcgcgaactgggcagc gccgccattcgtgctgccgtggaacgggcgggcac cgatgccgcgggtgtcgagcaggtactgttcggct gcgtgctgccggccggccagggccaggcacccggca cgccaggccgcgctgggcgcgggctggacaagca caccacctgcaccacctgaacaagatgtgcggct cgggtatgcaagccgcgatcatggcccatgacctg ctgctggccggcacgcgacagtggtagtggcggg tggcatggaaagcatgaccaacgcgcgtacctgc tggacaaagcccgctggcggtaccgcatgggccac ggcaagatcatcgaccacatgttcatggacgggtct cgaagacgcctacgacaaaggccgcctgatgggta cctttgccgaggactgtgcccaggccaatgccttc agccgcgaggcccaggaccagttcgccatcgccctc gctgacccgagcgcaggaagccatcagcagcggcc gttttgccgcgagatcgtgccggtggaagtcacc gagggcaaggaaaagcgcgtcatcaaggatgacga gcagccgcccaggcgcgtctggacaagattgcgc agctcaaaccggcggtttcgtgaaggcggcacccgtg acggcggccaacgccagttcgatttccgacggcgc tgcggcgctggtagtgatgcgcgcgtccgaggccg acaaacgtggcctcaagccattggcgcgtccac ggccacgcgcctttgccgacaccccggcgctgtt cccgaccgccccgatcggcgcgatcgacaaaactga tgaaacgcaccggctggaacctggccgaagtcgac

TABLE 53-continued

Cloned Thiolases					
Enzyme	Species template	Gene Length	5' PRIMER	3' PRIMER	ORF SEQ
					ctgttcgagatcaacgaggccttcgcgcgtgggtcac cctggcggccatgaaacacctcgacctgccacacg acaaggccaatatccacggcggcgccctgcgcctc ggtcacccgatcggcgcttctggcgacgtattct ggtcaccctgttgtcggccttgccgacagaacaatc tgcgtcgggtgtggcgcccatctgcatcggcggt ggcgaggccacggccatggctgttgaaatgcctgta ctga
beta- ketothiolase	<i>Caenorhabditis elegans</i>	kat-11167	ATGAACAAAC ATGCTTTCATC GTCG	TAATTTCTGGAT AACCATTCCACT TGAGC	atgaacaaacatgctttcatcgtcggagccgcgccg tacacctattggatcatttcgttcttctctctctt cggtaactgctccagagctcgccctcggttgccatc aaagcagcattggagcgtggagcagtgaagccgag ttcaattcaggagggtgttccttggtcaagtctgtc aagcaaatgctgggtcaagctcccgcctcgtcaagca gctcttgagaccggactcgatctttcggttgctgt taccaccgtcaataaagtgtgctcttctgggctga aagcaatcattcttgctgcccagcaaattcaaacc ggcatcaagattttgccattggcggagggaatgga gagcatgtcacaaagtaccattttatgttcaaagag gagagatcccatatgggtggatttcaagtgattgat ggaatcgtcaaagacggactgaccgatgcttatga taaagttcacatgggaaactgcggagagaagactt caaaagaaatgggaattacacgtaaagaccaagac gaatatgctatcaacagctacaaaaagtcagctaa agcatgggagaatggaaatatcggaccagaagtgg tgccagtgaacgtcaaatacaagaaggagtcacg attgttgataaagatgaagagttcacaaaagtcaa tttcgacaagttcacctcgctgagaactgttttcc agaaagacggaactatcactgctgctaattgcttca acattgaacgacgggtgcagctgctgtcattgttgc ctcacaggaagcagtttccgagcaaagcttaaagc ctctggcccgaatttttggttatggagatgccgcc acgcacccactcgatttcgctgtagcaccaacttt gatgttcccaaaaattcttgaaagacaggagtga agcaatcagatgttgctcaatgggaagttaatgaa gccttctcatgtgttccccttgcttccatcaaaaa actaggagtcgatccatcccctgtgaacccacatg gaggagctgtttcaattgggtcaccccatcggaatg tccggagcccgcctcatcactcatcttggtgcacac actcaaaagtggccaaatcggagttgctgccattt gcaatggagggtgggtggctcaagtggaatgggtatc cagaaattataa
beta- ketothiolase NP_415915.1	<i>Escherichia coli</i>	paaJ 1206	ATGCGTGAAG CCTTTATTTGT GACG	AACACGCTCCA GAATCATGGCG	atgcgtgaagcctttatattgtgacggaattcgta gccaattgggtcgctacggcggggcattatcaagt ttcgggctgatgatctggctgctatccctttgcgg gaactgctgggtgcgaaacccgcgtctcgatgcgga gtgtatcgatgatgtgatcctcggctgtgctaata aggcgggagaagataaacgtaacgtagcccggtg gcgactttactggcggggctgccgcagagtggttc cggcacacaccattaaccgcttgtgtggttccgggc tggaacgcactgggggttgccgcacgggcgattaaa gcgggcgatggcgatttgctgatcgccggtggcgt ggagtcaatgtcacgggcaccgtttgttatgggca aggcagccagtgcattttctcgtcaggetgagatg ttcgataccactattggctggcgatttgtgaaccc gctcatggctcagcaatttggaactgacagcatgc cgaaacggcagagaatgtagctgaactggttaaaa atctcacgagaagatcaagatagttttgcgctacg cagtcagcaacgtacggcaaaagcgcaatcctcag gcattctggctgaggagattgttccggttggttg aaaaacaagaaagggtgttgtaacagaaatacaaca tgatgagcatctgcgcccgaaacgacgctggaac agttacgtgggttaaaagcaccatttcgtgccaat ggggtgattaccgcaggcaatgcttccggggtgaa tgacggagccgctgcggttgattattgccagtgaac agatggcagcagcgcaaggactgacaccgcgggcg cgtatcgtagccatggcaaccgcgggggtggaacc gcgcctgatggggcttggtccggtgcctgcaactc gccgggtgctggaacgcgcagggtgagttcac

TABLE 53-continued

Cloned Thiolases					
Enzyme	Species template	Gene Length	5' PRIMER	3' PRIMER	ORF SEQ
					gatatggacgtgattgaactgaacgaagcgttcgc ggcccaggcgttgggtgtactacgcgaattggggc tgctgatgatgccccacatgttaaccccaacgga ggcgctatcgcccttaggccatccgttgggaatgag tggtgcccgcctggcactggctgccagccatgagc tgcatcggcgtaacggtcgttacgcatttgtgcacc atgtgcatcggtgtcggtcagggcatcgccatgat tctggagcgtgtttga
beta- ketothiolase AAN68887.1	<i>Pseudomonas putida</i> (KT2440)	phaD 1221	ATGAATGAAC CGACCCACGC C	GAGGCGCTCGA TGATCATGG	atgaatgaaccgacccacgccgatgccttgatcat cgacgccgtgcgcacgcccattggcgcgtatgccg gggccctgagcagcgtgcgcgccgacgacctggcg gccatcccgcctcaaagccttgatccagcgtcacc cgaactggactggaaagccattgatgacgttatct tcggctgtgccaaccaggctggcgaagacaaccgc aacgtggcccatggcgagcctgctggcggggct gccactcgaagtaccagggaccacgatcaaccgcc tgtgcggttcgggtctggatgccatcggtaatgcy gcacgtgccctgcgctgcggtgaagcggggctcat gctggccggtgggtgtggagtccatgtcgcggtgcac cgtttgtgatgggt aagt cggagcaggcattcggg cgtgcggccgagctgttcgacaccaccatcggctg gcgtttcgtcaaccgcgtgatgaaggccgcctacg gcatcgattcgatgccggaaacggctgaaaacgtg gccgaacagttcggcatctcgcgcgccgaccagga tgcccttgccctgcgcagccagcacaaagccgcag cagctcaggcccgcggccgcctggcgcgggaaatc gtgccggtcgaaatcccgcaacgcaaaggccagc caaagtggtcgagcatgacgagcaccgcgcggcg acacgaccctggagcagctggctcggtcgggacg ccgtttcgtgaaggcggcagcgtaacggcgggt aa tgccctccggcgtgaatgacggcgcttgcgccctgc tgctggccagcagcgccgcggcccgcccatggg ttgaaggcccgcggccgcgatcgtcggcattggcgt ggccgggggttgagcccaggctgatgggcattggtc cgggtgcctgcgaccgcgaaggctgctggcgctcacc ggcctggcactggctgacctggatgtcatcgaaact caatgaggcctttgccgcccagggtggccgtgt tgcgcgagctgggcctggccgacgacgaccgcga gtcaaccgcaacggcggcgccatcgccctgggcca tcccctgggcatgagcgggtgcccggttggtgacca ctgccttgcaagcagcttgaagaaacggccggccgc tacgcctgtgcaccatgtgcacggcgtaggcca aggcattgccatgatcatcgagcgccctctga
beta- ketothiolase NP_349476.1	<i>Clostridium acetobutylicum</i> ATCC 824	thiA 1179	ATGAAAGAAG TTGTAATAGCT AGTGCAGTAA GAAC	GCACTTTTCTAG CAATATTGCTGT TCC	atgaaagaagttgtaatagctagtgcagtaagaac agcgattggatcttatggaaagtctcttaaggatg taccagcagtagatttaggagctacagctataaag gaagcagttaaaaagcaggaataaaaccagagga tgtaaatgaagtcattttaggaaatgtctttcaag caggtttaggacagaatccagcaagacaggcatct tttaaagcaggattaccagttgaaattccagctat gactattaataagggtttgtggttcaggacttagaa cagttagcttagcagcacaaattataaaagcagga gatgctgacgtaataatagcaggtgggtatggaaaa tatgtctagagctccttacttagcgaataacgcta gatggggatatagaatgggaaacgctaaatttggt gatgaaatgatcactgacggatttggggatgcatt taatgattaccacatgggaataacagcagaaaaca tagctgagagatggaacatttcaagagaagaacaa gatgagtttgctcttgcatcacaaaaaaagctga agaagctataaaatcagggtcaatttaaagatgaaa tagttcctgtagtaattaaaggcagaaaggagaa actgtagtgtgat acagatgagcaccctagatttg atcaactatagaaggacttgcaaaattaaaacctg ccttcaaaaaagatggaacagttacagctggtaac gcatcaggattaaatgactgtgcagcagtagttgt aatcatgagtgcagaaaaagctaaagagcttgga taaaaccacttgctaagatagtttcttatgggttca gcaggagttgacccagcaataatgggatatggacc tttctatgcaacaaaagcagctattgaaaaagcag

TABLE 53-continued

Cloned Thiolases					
Enzyme	Species template	Gene Length	5' PRIMER	3' PRIMER	ORF SEQ
					gttggacagttgatgaattagatttaatagaatca aatgaagcttttgcagctcaaagtttagcagtagc aaaagatttaaaatttgatatgaataaagtaaag taaattggaggagctattgcccttggtcatccaatt ggagcatcaggtgcaagaatactcgttactcttgt acacgcaatgcaaaaaagagatgcaaaaaaggct tagcaactttatgtataggtggcggacaaggaaca gcaatattgctagaaaagtgctag
beta- ketothiolase NP_149242.1	<i>Clostridium</i> <i>acetobutylicum</i> ATCC 824	thiB 1179	ATGAGAGATG TAGTAATAGT AAGTGCTGTA AGAACTG	GTCTCTTCAAC TACGAGAGCTGT TCCC	atgagagatgtagtaatagtaagtgcgtgaagaac tgcaataggagcatatggaaaaacattaaaggatg tacctgcaacagagttaggagctatagtaataaag gaagctgtaagaagagctaataaaatccaaatga gattaatgaagttattttggaaatgtacttcaag ctggattaggccaaaaccagcaagacaagcagca gtaaaagcaggattacctttagaaacacctgcgtt tacaatcaataaggtttgtgggttcaggtttaagat ctataagtttagcagctcaaattataaaagctgga gatgctgataccattgtagtaggtggtatggaaaa tatgtctagatcaccatatttgattaacaatcaga gatggggtcaaagaatgggagatagtgaattagtt gatgaaatgataaaggatggtttgtgggatgcatt taatggatatcatatgggagtaactgcagaaaata ttgcagaacaatggaatataacaagagaagagcaa gatgaattttcacttatgtcacacaaaaagctga aaaagccattaaaaatggagaatttaaggatgaaa tagttcctgtattaataaagactaaaaaagggtgaa atagtctttgatcaagatgaatttcctagattcgg aaacactattgaagcattaagaaaacttaaaccta ttttcaaggaaaatggtagtactgttacagcaggtaat gcatccggattaaatgatggagctgcagcactagt aataatgagcgtgataaagctaacgctctcggaa taaaaccacttgctaagattacttcttacggatca tatggggtagatccatcaataatgggatatggagc ttttatgcaactaaagctgccttagataaaaatta atttaaaacctgaagacttagatttaattgaagct aacgaggcatatgcttctcaaagtatagcagtaac tagagatttaaaatttagatatgagttaaagttaatg ttaatggtggagctatagcacttggacatccaata ggtgcatctggtgcacgtattttagtaacattact atacgctatgcaaaaaagagattcaaaaaagggtc ttgctactctatgtattgggtggaggtcaggggaaca gctctcgtagttgaaagagactaa
3-oxoadipyl- CoA thiolase	<i>Candida</i> <i>albicans</i> SC5314	POT9 1182 8	ATGTTCAAGA AATCAGCTAA TGATATTGTTG	CTCGTTAGCAAA CAAGGCAGCG	atgttcaagaaatcagctaatagatattgttggtat tgcagcaaagagaactccaatcaccaagtcaatta aagggtgggttgagtagattatttcctgaggaaata ttatatcaagtggttaagggtactgtatcagattc acaagttgatttaaacttgattgatgatgtgttag tcggtacggtcttgcaaactttagggggacagaaa gctagtgccttggccattaaaaagattggattccc aattaagaccacggttaatacgggtcaatcgtcaat gtgctagtctgctcaagcgattacttatcaagca ggtagtttgcgtagtgggggagaatcaatttgctat tgctgctggagtagaaagtatgactcatgattatt ttcctcatcgtgggattcccacaagaatttctgaa tcatttttagctgatgcatccgatgaagctaaaaa cgtcttgatgccaatggggataaccagtgaaaatg ttgccactaaatatggaatttctcgtaaacacaa gatgagtttgcccttaattctcatttgaaagcaga caaggctacaaaactgggtcattttgcaaaagaaa tcattcctattcaaacaacggatgaaaacaaccaa cacgtttcaataaccaagatgatggtataagggg aagttcaacaattgaaaagttgggtggcttaaaac ctgtgttcaaggatgatgggactactactgctggt aattcctcgcaatttcagatggagggctctgctgt gattttaactactcgtcaaatgctgagaaatcgg gagtaaagccaatagctagattttatgggttcgtca gtagctgggtgttccttcgggacttatgggaattgg tccatcggtgctattcctcaattgttgtcgagat taaatgttgacacgaaagacattgatatttttgaa

TABLE 53-continued

Cloned Thiolases					
Enzyme	Species template	Gene Length	5' PRIMER	3' PRIMER	ORF SEQ
					ttgaacgaggcatttgcatcccaactgatttattg tattgaaaaattgggtcttgattatgataaagtca atccatatgggtggagctatagccttgggacatcca ttaggagccactggcgcaagagttacggcaacggt gcttaatggattaaaagatcagaataaagagttgg gtgtcatctcaatgtgcacatccacaggtcaagga tacgctgccttggttgctaacgagtag
3-oxoadipyl-CoA thiolase	<i>Candida albicans</i> SC5314	POT1 1227	ATGGATAGAT TAAATCAATT AAGTGGTCAA TTAAAACC	TTCCTTAATCAA TATGGAGGCAG CAC	atggatagattaaatcaattaagtgggtcaattaaa accaacttcaaaaacaatcccttactcaaaagaacc cagacgatgttgatcatcggtgcagcatacagaact gccatcggtaaagggtttcaaagggtctttcaaatc tgtgcaatctgaattcatcttgactgaattcttgga aagaatttattaaaaagactggagtcgatgcacat ttgattgaagatgttgctatttggtaacgttttgaa ccaagctgctgggtgccaccgaacacagaggtgcta gtttggctgcaggtattccttacactgcagctttc cttgccatcaacagatttgtgttcctcagggttaat ggccatttctgacattgccacaaaatcaaaaccg gtgaaatcgaatgtgggtcttgctgggtggtattgaa tccatgtctaaaaactatggtagtccaaaagttat tccaaagattgaccacacttggctgatgacgaac aatgagtaaatgtttgattccaatgggtatcacc aacgaaaatgttgctaattgaattcaacattccaag agaaaaacaagatgcctttgctgctaaatcttata gtaaagccgaaaaagccatctcctctggagctttc aaagatgaaatcttaccatcagatccattatcag atccccagacgggttctgaaaaagaaatcattgtcg ataccgacgaagggtccaagaaagggtgttgacgct gcttccttgagcaaatgaaaccagcatttggtgg tactaccactgccggtaacgcttctcaaatttcag atgggtgctgctgggtgtttattgatgaagagaagt ttggctgaagccaaagggtacccaattgttgctaa atacattgcttggttcaactgttggtgttcgccag aatcatgggtgttggtccagcttacgccattcca gaagtgtgaagagaactggattgactgtggatga cgttgatgtgtttgaaatcaacgaagcttttgctg ctcaatgtctttactcagctgaacaatgtaatgtt ccagaagaaaaattgaacataaacgggtggtgccat cgcttaggtcatcctcttggttgactgggtgcca gacaatatgccactatcttgagattgttgaaacca ggtgaaattgggtttgacttctatgtgtatcggtag tggtatgggtgctgcctccatattgattaaggaat ag
3-oxoadipyl-CoA thiolase	<i>Candida albicans</i> SC5314	POT2 1233	ATGTCATCCA AACAACAATA CTTGAAGAAG	TTCTCTAACCAA AACAGAAGCAG CACC	atgtcatccaaacaacaatacttgaagaagaatcc tgacgatgtcggtgtcggtgcagcatacagaactg ctttaaccaaagggtggaagaggtggattcaaagat gttggtatctgatttccttttgaaaaaattgactga agaatttggttaaaaaaactgggtgttgacctaaaa tcattcaagatgctgccattggtaatgtcttgaac agaagagctgggtgatttcgaacatagaggtgcatt attatctgctggattaccttattcagttccatttg ttgcccttaacagacaatgttcacatctgggttaatg gccatttctcaagtggccaacaagatcaagactgg tgaaattgaatgtgggttagctgggtggttgaaa gtatgacaaaaaactatgggtccagaagcattgatt gctattgacctgcttatgaaaaagaccagaatt tgtaaaaacgggtattccaatgggtattactaatg aaaatgtttgtgccaaattcaatatttcaagagat gttcaagatcaatttgctgctgaatcttatcaaaa agctgaaaaggcacaaaaagaaggtaaatttgatg atgaaattttaccaattgaagttttccaagaagat gaagatgctgaagatgaagacgaagatgaagatga agatgctgaacaaaaagaaaaattgggtgttatta gtaaagatgaaggatttagaccaggtgttactaaa gaaaaattggctaaaattaaaccagctttcaaatc tgatgggtgatcttcagctggtaactcttcacaag ttccgatgggtgctgccttggtgttattgatgaaa cgttcatttgctgaaaagaatggattcaaaccatt ggctaaatacatcttcttggtgttgctgggtgtcc

TABLE 53-continued

Cloned Thiolases						
Enzyme	Species template	Gene	Length	5' PRIMER	3' PRIMER	ORF SEQ
						caccagaaattatgggtattgggtccagctgttgcc attccaaaagttttgaaacaaactggattatcagt cagtgatattgatatttatgaaatcaatgaagcat ttgccggtcaatgtttgtactcaattgaaagttgt aatattccaagagaaaaagtcaatcttaatggggg tgctattgccttgggtcacccctcttggttgactg gtgctagacaatacgtactattttaagattgtta aaaccaggtgaatttggtgtgacttctatgtgtat tggtactggtatgggtgctgcttctgttttggtta gagaataa
beta- ketoadipyl CoA thiolase pcaF	<i>Pseudomonas aeruginosa</i> PA01	pcaF	1206	ATGAGCCGCG AGGTATTCAT CTG	GACCCGCTCGAT GGCCAG	atgagccgcgaggtattcatctgcgatgccgtgcg cacgccgatcggccgtttcgggcgagctctttccg cggtgcgcgccgacgacctcgcggcggtgccgctg aaggccctggtcgagcgcaaccgggggtcgactg gtcggcgttggacgaggtgttcctcggctgcgcca accaggccggcgaggacaaccgtaacgtggcgcg atggcgctgctgctggccggtttgccggagagcgt gcccggcgtcacctcaaccgctctgcgcctcgg ggatggacgccatcggcacggcgttcgcgcctac gcctgcggcgagatggagctggccatcgccggcgg cgtcgagtcgatgtcgcgcgcgccgtacgtgatgg gcaaggccgatagcgcttcggtcgcggccagaag atcgaggacaccaccatcggctggcgcttcgtcaa tccgctgatgaaggagcagtacggcatcgaccga tgccgcagaccgcccgaacgtcgccgacgactat cgcgtgtcgcgtgccgaccaggatgccttcgcct gcgcagccagcagcgcgccggcaggcgagagg ccggtttcttcgccgaggaaatcgtcccggtgacg attcgcgggcgcaagggcgacaccctggtcgagca cgacgagcatccgcgtcccgacaccacctggagg cgctggcccggtcaagccggtcaacgggcccggag aagaccgtcacccgcggcaacgcgtccggggtaa cgacggcgccgcgcgctggtcctggcctccgccg aggcagtgagagaagcacggcctgactccgcgcgcg cgggtgctgggcatggccagcgccggcgtcgcccc acggatcatgggcatcggcccgtgcccggcggtgc gcaagctgctgcggcgctggacctggcgatcgac gccttcgacgtgatcgaaactcaacgaagccttcgc cagccagggcctggcctgctgcgcgaactgggcg tgcccgacgacagtgagaaggtcaaccgaacggc ggtgccatcgccctcggccacccgctggggatgag cgggtgcgcggtggtcctcacgcgctccatcaac ttgagaagagcggcggccggcgcgccctggcgacc atgtgcgtaggcgtcggccaaggcctggcgctggc catcgagcgggtctga
acyl-CoA thiolase	<i>Pseudomonas aeruginosa</i> PA01	bkt	1206	ATGCTCGATG CCTATATCTAC GCC	TCGGCAGCGCTC GATCAC	atgctcgatgcctatatctacgccggcctgcgtac gcctttcggccggcatgccggtgcactctcgacgg tgcgtccggacgacctggccggcctgctgctggcg cgtctcgcgaaacctccgggttcgccgtcgacga cctggaggatgtgatcctcggttgaccaaccagg ccggcgaagacagccgcaacctggcgcgcaacgcg ctgctcgcagccggcctgccggcgcggtgcccg gcagacggtcaaccgcttggtgtgccagcggactgt cggcggtgatcgacgcggcgcgcgatcagttgc ggtgagggccggctgtacctggccggcgcgccga aagcatgtcccgggcgcgcttcgtcatgggcaagg cggagagcgcttcagccgcacgctggaggtcttc gacagcaccatcggcgcgcgcttcgccaaacccag gctggtcgagcgctatggcaacgacagcatgccgg agaccggcgacaacgtggcccgcgcttcggcatc gcccgcgaagacgcgcgaccttcgcgcgcttcttc ccaggcgcgctaccaggctgcgctggaggagggt ttttcctcggcgagatccttcgggtggagggtgcgt gccggacgcaagggcgagacgcggctggtggagcg cgacgagcatccgcgaccgcaggccgacctggcgg ccctggcgcgcttgccggcggttggtcgccgggtggg gtagtgaccgcggtaaatgcgtctgggatcaacga cggggcgggcggtagtgcgtgctgggcgatcgcgcga tcggcgagcgcgagggcatccggccgttggcgcgg

TABLE 53-continued

Cloned Thiolases						
Enzyme	Species template	Gene	Length	5' PRIMER	3' PRIMER	ORF SEQ
						atcctcgccagcgccagcgctcggcgtcgagccccg gttgatgggcatcgccccgcagcaggcgatcctcc gcgcgctgcaacgcgcggcatcgacctggacgag gtcggcctgatcgagatcaacgaagccttcgcgcc gcaggtcctggcctgcctgaagtgtctcggcctgg actacgaggacccgcgggtcaatccccatggcggc gccattgccctcgccatccgctcggcgcctccgg tgcgcgcctggtgctcaccgcgcgcccggggtgc aacgcacgagcggcgctacgcggtggtcagcctg tgcgtcgggctcggccaggcggtggcgatggtgat cgagcgctgccgatga
3-oxoadipyl- CoA thiolase	<i>Pseudomonas putida</i> (KT2440)	pcaF	1203	ATGCACGACG TATTCATCTGT GACG	AACCCGCTCGAT GGCCAAC	atgcacgacgtattcatctgtgacgccatccgtac cccgatcgcccgcttcggcgggcgccctggccagcg tgcgggcgcgacgacctggccgcccgtgccgctgaag gcgctgatcgagcgcaaccctggcggtgcagtggga ccaggtagacgaagtgttcttcggctgcgccaaacc aggccggtgaagacaaccgcacacgtggcccgcatg gcactgctgctggccggcctgccggaaagcatccc gggcgtcaccctgaaccgtctgtgcgcgtcgggca tggatgccgtcggcaccgcgttcgcgcgccatcgcc agcggcgagatggagctggtgatcgccggtggcggt cgagtcgatgtcgcgcgcgcccggttcgtcatgggca aggctgaaagcgccatttcgcgcgaacatgaagctg gaagacaccaccattggctggcggtttcatcaaccc gctgatgaagagccagtagcgggtgtggattccatgc cggaaaccgcgcgacaacgtggccgacgactatcag gtttcgcgtgctgatcaggacgctttcgccctgcg cagccagcagaaggctgccgctgcgcaggtgccg gcttctttgccgaagaaatcgtgccgggtgcgtatc gctcacaagaagggcgaaatcatcgtcgaacgtga cgaacacctgcgcgccggaaccacgctggaggcgc tgaccaagctcaaaccgggtcaacggcccggacaag acggtcaccgcgcggcaacgcctcgggcgtgaacga cgggtgctgcggcgatgatcctggcctcggccgcag cgggtgaagaaacacggcctgactccgcgtgcccg gttctgggcatggccagcggcggcggttgcgccacg tgtcatgggcattggcccggtgcggcggtgcgca aactgaccgagcgtctggggatagcggtaagtgat ttcgacgtgatcgagcttaacgaagcgtttgccag ccaaggcctggcggtgctgcgtgagctgggtgtgg ctgacgatgcgcgccaggtaaaccctaattggcggt gccattgccctggggcaccctcgggcatgagcgg tgcacgcctggtactgactgcgttgcaccagctgg agaagagtggcggtcgcaagggcctggcgaccatg tgtgtgggtgtcggccaaggctcggcggttggccat cgagcgggtttga
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TABLE 53-continued

Cloned Thiolases						
Enzyme	Species template	Gene	Length	5' PRIMER	3' PRIMER	ORF SEQ
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[0252] The genes were expressed in *E. coli* and the proteins purified using Ni-NTA spin columns and quantified. To assay enzyme activity in vitro, a 5×CoA:DTNB (Ellman’s reagent or 5,5'-dithiobis-(2-nitrobenzoic acid)) mixture was prepared. The mixture consisted of 10 mM succinyl-CoA, 5 mM acetyl-CoA, 30 mM DTNB in 100 mM Tris buffer, pH 7.4. Five μL of the CoA:DTNB mixture was added to 0.5 μM purified thiolase enzyme in 100 mM Tris buffer, pH 7.8 in a final volume of 50 μL. The reaction was incubated at 30° C. for 30 minutes, then quenched with 2.5 μL 10% formic acid and samples frozen at −20° C. until ready for analysis by LC/MS. Because many thiolases can condense two acetyl-CoA molecules into acetoacetyl-CoA, production of acetoacetyl-CoA was examined. FIG. 6 shows that 3 thiolases demonstrated thiolase activity which resulted in acetoacetyl-CoA formation. These were fadAx from *Pseudomonas*

putida, thiA from *Clostridium acetobutylicum* and thiB also from *Clostridium acetobutylicum*. When enzyme assays were examined for condensation of succinyl-CoA and acetyl-CoA into β-ketoadipyl-CoA, several enzymes demonstrated the desired activity; paaJ from *Escherichia coli* (Nogales et al., *Microbiol.* 153:357-365 (2007)), phaD from *Pseudomonas putida* (Olivera et al., *Proc. Natl. Acad. Sci. USA* 95:6419-6424 (1998)), bkt from *Burkholderia ambifaria* AMMD, pcaF from *Pseudomonas putida* KT2440 (Harwood et al., *J. Bacteriol.* 176:6479-6488 (1994)), and pcaF from *Pseudomonas aeruginosa* PAO1. There was excellent specificity between the thiolases. Those that generated significant amounts of β-ketoadipyl-CoA did not produce significant amounts of acetoacetyl-CoA and likewise those that made acetoacetyl-CoA did not make detectable amounts of β-ketoadipyl-CoA.

Example II

Preparation of Terephthalate from Acetylene and Muconate

[0253] This Example provides conditions for the thermal inverse electron demand Diels-Alder reaction for the preparation of PTA from acetylene and muconate.

[0254] A lab-scale Parr reactor is flushed with nitrogen gas, evacuated and charged with (1 equivalent) trans, trans-muconic acid and (10 equivalents) acetylene. The reactor is then heated to 200° C. and held at this temperature for 12 hours. An initial pressure of 500 p.s.i.g. is applied. The reactor is then vented, exposed to air and cooled. The contents of the reactor are distilled at room temperature and pressure to yield volatile and nonvolatile fractions. The contents of each fraction are evaluated qualitatively by gas chromatographic analysis (GC-MS).

[0255] For quantitative analysis, standards of the starting materials and the expected products, cyclohexa-2,5-diene-1,4-dicarboxylate and terephthalate, are prepared. A known amount of cyclohexane is mixed with a known amount of the volatile fraction and the mixture is subjected to gas chromatography. The cyclohexane and terephthalate components are condensed from the effluent of the chromatogram into a single trap, the contents of which are diluted with carbon tetrachloride or CDCl_3 and then examined by NMR spectroscopy. Comparison of the appropriate areas of the NMR spectrum permits calculation of yields.

Example III

Preparation of a Muconate Producing Microbial Organism, in which the Muconate is Derived from succinyl-CoA

[0256] This example describes the generation of a microbial organism that has been engineered to produce muconate from succinyl-CoA and acetyl-CoA via beta-ketoadipate, as shown in FIG. 2. This example also provides a method for engineering a strain that overproduces muconate.

[0257] *Escherichia coli* is used as a target organism to engineer a muconate-producing pathway as shown in FIG. 5. *E. coli* provides a good host for generating a non-naturally occurring microorganism capable of producing muconate. *E. coli* is amenable to genetic manipulation and is known to be capable of producing various products, like ethanol, acetic acid, formic acid, lactic acid, and succinic acid, effectively under anaerobic, microaerobic or aerobic conditions.

[0258] First, an *E. coli* strain is engineered to produce muconate from succinyl-CoA via the route outlined in FIG. 2. For the first stage of pathway construction, genes encoding enzymes to transform central metabolites succinyl-CoA and acetyl-CoA to 2-maleylacetate (FIG. 2, Step A) is assembled onto vectors. In particular, the genes *pcaF* (AAA85138), *pcaI* (AAN69545 and NP_746082) and *clcE* (O30847) genes encoding beta-ketothiolase, beta-ketoadipyl-CoA transferase and 2-maleylacetate reductase, respectively, are cloned into the pZE13 vector (Expressys, Ruelzheim, Germany), under the control of the PA1/lacO promoter. The genes *bdh* (AAA58352.1) and *fumC* (P05042.1), encoding 2-maleylacetate dehydrogenase and 3-hydroxy-4-hexenedioate dehydratase, respectively, are cloned into the pZA33 vector (Expressys, Ruelzheim, Germany) under the PA1/lacO promoter. The two sets of plasmids are transformed into *E.*

coli strain MG1655 to express the proteins and enzymes required for muconate synthesis from succinyl-CoA.

[0259] The resulting genetically engineered organism is cultured in glucose containing medium following procedures well known in the art (see, for example, Sambrook et al., supra, 2001). The expression of muconate pathway genes is corroborated using methods well known in the art for determining polypeptide expression or enzymatic activity, including for example, Northern blots, PCR amplification of mRNA and immunoblotting. Enzymatic activities of the expressed enzymes are confirmed using assays specific for the individually activities. The ability of the engineered *E. coli* strain to produce muconate through this pathway is confirmed using HPLC, gas chromatography-mass spectrometry (GCMS) or liquid chromatography-mass spectrometry (LCMS).

[0260] Microbial strains engineered to have a functional muconate synthesis pathway from succinyl-CoA are further augmented by optimization for efficient utilization of the pathway. Briefly, the engineered strain is assessed to determine whether any of the exogenous genes are expressed at a rate limiting level. Expression is increased for any enzymes expressed at low levels that can limit the flux through the pathway by, for example, introduction of additional gene copy numbers.

[0261] After successful demonstration of enhanced muconate production via the activities of the exogenous enzymes, the genes encoding these enzymes are inserted into the chromosome of a wild type *E. coli* host using methods known in the art. Such methods include, for example, sequential single crossover (Gay et al., *J. Bacteriol.* 153:1424-1431 (1983)) and Red/ET methods from GeneBridges (Zhang et al., Improved RecT or RecET cloning and subcloning method (WO/2003/010322)). Chromosomal insertion provides several advantages over a plasmid-based system, including greater stability and the ability to co-localize expression of pathway genes.

[0262] To generate better producers, metabolic modeling is utilized to optimize growth conditions. Modeling is also used to design gene knockouts that additionally optimize utilization of the pathway (see, for example, U.S. patent publications US 2002/0012939, US 2003/0224363, US 2004/0029149, US 2004/0072723, US 2003/0059792, US 2002/0168654 and US 2004/0009466, and in U.S. Pat. No. 7,127,379). Modeling analysis allows reliable predictions of the effects on cell growth of shifting the metabolism towards more efficient production of muconate. One modeling method is the bilevel optimization approach, OptKnock (Burgard et al., *Biotechnol. Bioengineer.* 84:647-657 (2003)), which is applied to select gene knockouts that collectively result in better production of muconate. Adaptive evolution also can be used to generate better producers of, for example, the 4-acetylbutyrate intermediate or the muconate product. Adaptive evolution is performed to improve both growth and production characteristics (Fong and Palsson, *Nat. Genet.* 36:1056-1058 (2004); Alper et al., *Science* 314:1565-1568 (2006)). Based on the results, subsequent rounds of modeling, genetic engineering and adaptive evolution can be applied to the muconate producer to further increase production.

[0263] For large-scale production of muconate, the above muconate pathway-containing organism is cultured in a fermenter using a medium known in the art to support growth of the organism under anaerobic conditions. Fermentations are performed in either a batch, fed-batch or continuous manner. Anaerobic conditions are maintained by first sparging the

medium with nitrogen and then sealing culture vessel (e.g., flasks can be sealed with a septum and crimp-cap). Microaerobic conditions also can be utilized by providing a small hole for limited aeration. The pH of the medium is maintained at a pH of 7 by addition of an acid, such as H₂SO₄. The growth rate is determined by measuring optical density using a spectrophotometer (600 nm), and the glucose uptake rate by monitoring carbon source depletion over time. Byproducts such as undesirable alcohols, organic acids, and residual glucose can be quantified by HPLC (Shimadzu) with an HPX-087 column (BioRad), using a refractive index detec-

tor for glucose and alcohols, and a UV detector for organic acids, Lin et al., *Biotechnol. Bioeng.*, 775-779 (2005).

[0264] Throughout this application various publications have been referenced. The disclosures of these publications in their entirety are hereby incorporated by reference in this application in order to more fully describe the state of the art to which this invention pertains. Although the invention has been described with reference to the examples provided above, it should be understood that various modifications can be made without departing from the spirit of the invention.

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1. A non-naturally occurring microbial organism, comprising a microbial organism having a muconate pathway comprising at least one exogenous nucleic acid encoding a muconate pathway enzyme expressed in a sufficient amount to produce muconate, said muconate pathway comprising an enzyme selected from the group consisting of a beta-ketothiolase, a beta-ketoadipyl-CoA hydrolase, a beta-ketoadipyl-CoA transferase, a beta-ketoadipyl-CoA ligase, a 2-fumarylacetate reductase, a 2-fumarylacetate dehydrogenase, a trans-3-hydroxy-4-hexendioate dehydratase, a 2-fumarylacetate aminotransferase, a 2-fumarylacetate aminating oxidoreductase, a trans-3-amino-4-hexenoate deaminase, a beta-ketoadipate enol-lactone hydrolase, a muconolactone isomerase, a muconate cycloisomerase, a beta-ketoadipyl-CoA dehydrogenase, a 3-hydroxyadipyl-CoA dehydratase, a 2,3-dehydroadipyl-CoA transferase, a 2,3-dehydroadipyl-CoA hydrolase, a 2,3-dehydroadipyl-CoA ligase, a muconate reductase, a 2-maleylacetate reductase, a 2-maleylacetate dehydrogenase, a cis-3-hydroxy-4-hexendioate dehydratase, a 2-maleylacetate aminoatransferase, a 2-maleylacetate aminating oxidoreductase, a cis-3-amino-4-hexendioate deaminase, and a muconate cis/trans isomerase.

2. The non-naturally occurring microbial organism of claim 1, wherein said muconate pathway comprises a set of muconate pathway enzymes, said set of muconate pathway enzymes selected from the group consisting of:

A) (1) beta-ketothiolase, (2) an enzyme selected from beta-ketoadipyl-CoA hydrolase, beta-ketoadipyl-CoA transferase, and beta-ketoadipyl-CoA ligase, (3) beta-ketoadipate enol-lactone hydrolase, (4) muconolactone isomerase, (5) muconate cycloisomerase, and (6) a muconate cis/trans isomerase;

B) (1) beta-ketothiolase, (2) an enzyme selected from beta-ketoadipyl-CoA hydrolase, beta-ketoadipyl-CoA transferase and beta-ketoadipyl-CoA ligase, (3) 2-maleylacetate reductase, (4) 2-maleylacetate dehydrogenase, (5) cis-3-hydroxy-4-hexendioate dehydratase, and (6) muconate cis/trans isomerase;

C) (1) beta-ketothiolase, (2) an enzyme selected from beta-ketoadipyl-CoA hydrolase, beta-ketoadipyl-CoA transferase and beta-ketoadipyl-CoA ligase, (3) 2-maleylacetate reductase, (4) an enzyme selected from 2-maleylacetate aminotransferase and 2-maleylacetate aminating oxidoreductase, (5) cis-3-amino-4-hexenoate deaminase, and (6) muconate cis/trans isomerase;

D) (1) beta-ketothiolase, (2) beta-ketoadipyl-CoA dehydrogenase, (3) 3-hydroxyadipyl-CoA dehydratase, (4) an enzyme selected from 2,3-dehydroadipyl-CoA transferase, 2,3-dehydroadipyl-CoA hydrolase and 2,3-dehydroadipyl-CoA ligase, and (5) muconate reductase;

E) (1) beta-ketothiolase, (2) an enzyme selected from beta-ketoadipyl-CoA hydrolase, beta-ketoadipyl-CoA transferase and beta-ketoadipyl-CoA ligase, (3) 2-fumarylacetate reductase, (4) 2-fumarylacetate dehydrogenase, and (5) trans-3-hydroxy-4-hexendioate dehydratase;

F) (1) beta-ketothiolase, (2) an enzyme selected from beta-ketoadipyl-CoA hydrolase, beta-ketoadipyl-CoA transferase and beta-ketoadipyl-CoA ligase, (3) 2-fumarylacetate reductase, (4) an enzyme selected from 2-fumarylacetate aminotransferase and 2-fumarylacetate aminating oxidoreductase, and (5) trans-3-amino-4-hexenoate deaminase.

3. The non-naturally occurring microbial organism of claim 2, wherein said microbial organism comprises two exogenous nucleic acids each encoding a muconate pathway enzyme.

4. The non-naturally occurring microbial organism of claim 2, wherein said microbial organism comprises three exogenous nucleic acids each encoding a muconate pathway enzyme.

5. The non-naturally occurring microbial organism of claim 2, wherein said microbial organism comprises four exogenous nucleic acids each encoding a muconate pathway enzyme.

6. The non-naturally occurring microbial organism of claim 2, wherein said microbial organism comprises five exogenous nucleic acids each encoding a muconate pathway enzyme.

7. The non-naturally occurring microbial organism of claim 2, wherein said microbial organism comprises six exogenous nucleic acids each encoding a muconate pathway enzyme.

8. The non-naturally occurring microbial organism of claim 1, wherein said at least one exogenous nucleic acid is a heterologous nucleic acid.

9. The non-naturally occurring microbial organism of claim 1, wherein said non-naturally occurring microbial organism is in a substantially anaerobic culture medium.

10. A method for producing muconate, comprising culturing the non-naturally occurring microbial organism according to claim 1, under conditions and for a sufficient period of time to produce muconate according to claim 1.

11.-18. (canceled)

19. A non-naturally occurring microbial organism, comprising a microbial organism having a muconate pathway comprising at least one exogenous nucleic acid encoding a muconate pathway enzyme expressed in a sufficient amount to produce muconate, said muconate pathway comprising an enzyme selected from the group consisting of a 4-hydroxy-2-ketovalerate aldolase, a 2-oxopentenoate hydratase, a 4-oxalocrotonate dehydrogenase, a 2-hydroxy-4-hexenedioate dehydratase, a 4-hydroxy-2-oxohexanedioate oxidoreductase, a 2,4-dihydroxyadipate dehydratase (acting on 2-hydroxy), a 2,4-dihydroxyadipate dehydratase (acting on 4-hydroxyl group) and a 3-hydroxy-4-hexenedioate dehydratase.

20. The non-naturally occurring microbial organism of claim 19, wherein said muconate pathway comprises a set of muconate pathway enzymes, said set of muconate pathway enzymes selected from the group consisting of:

A) (1) 4-hydroxy-2-ketovalerate aldolase, (2) 2-oxopentenoate hydratase, (3) 4-oxalocrotonate dehydrogenase, (4) 2-hydroxy-4-hexenedioate dehydratase;

B) (1) 4-hydroxy-2-ketovalerate aldolase, (2) 4-hydroxy-2-oxohexanedioate oxidoreductase, (3) 2,4-dihydroxyadipate dehydratase (acting on 2-hydroxy), (4) 3-hydroxy-4-hexenedioate dehydratase; and

C) (1) 4-hydroxy-2-ketovalerate aldolase, (2) 4-hydroxy-2-oxohexanedioate oxidoreductase, (3) 2,4-dihydroxyadipate dehydratase (acting on 4-hydroxyl group), (4) 2-hydroxy-4-hexenedioate dehydratase.

21. The non-naturally occurring microbial organism of claim 20, wherein said microbial organism comprises two exogenous nucleic acids each encoding a muconate pathway enzyme.

22. The non-naturally occurring microbial organism of claim 20, wherein said microbial organism comprises three exogenous nucleic acids each encoding a muconate pathway enzyme.

23. The non-naturally occurring microbial organism of claim 20, wherein said microbial organism comprises four exogenous nucleic acids each encoding a muconate pathway enzyme.

24. The non-naturally occurring microbial organism of claim 19, wherein said at least one exogenous nucleic acid is a heterologous nucleic acid.

25. The non-naturally occurring microbial organism of claim 19, wherein said non-naturally occurring microbial organism is in a substantially anaerobic culture medium.

26. A method for producing muconate, comprising culturing the non-naturally occurring microbial organism according to claim 19, under conditions and for a sufficient period of time to produce muconate.

27.-32. (canceled)

33. A non-naturally occurring microbial organism, comprising a microbial organism having a muconate pathway comprising at least one exogenous nucleic acid encoding a muconate pathway enzyme expressed in a sufficient amount to produce muconate, said muconate pathway comprising an enzyme selected from the group consisting of an HODH aldolase, an OHED hydratase, an OHED decarboxylase, an HODH formate-lyase, an HODH dehydrogenase, an OHED formate-lyase, an OHED dehydrogenase, a 6-OHE dehydrogenase, a 3-hydroxyadipyl-CoA dehydratase, a 2,3-dehydroadipyl-CoA hydrolase, a 2,3-dehydroadipyl-CoA transferase, a 2,3-dehydroadipyl-CoA ligase, and a muconate reductase.

34. The non-naturally occurring microbial organism of claim 33, wherein said muconate pathway comprises a set of muconate pathway enzymes, set of muconate pathway enzymes selected from the group consisting of:

A) (1) HODH aldolase, (2) OHED hydratase, (3) OHED decarboxylase, (4) 6-OHE dehydrogenase, and (5) muconate reductase;

B) (1) HODH aldolase, (2) OHED hydratase, (3) an enzyme selected from OHED formate-lyase and OHED dehydrogenase, (4) an enzyme selected from 2,3-dehydroadipyl-CoA hydrolase, 2,3-dehydroadipyl-CoA transferase and 2,3-dehydroadipyl-CoA ligase, and (5) muconate reductase; and

C) (1) HODH aldolase, (2) an enzyme selected from HODH formate-lyase and HODH dehydrogenase, (3) 3-hydroxyadipyl-CoA dehydratase, (4) an enzyme selected from 2,3-dehydroadipyl-CoA hydrolase, 2,3-dehydroadipyl-CoA transferase and 2,3-dehydroadipyl-CoA ligase, and (5) muconate reductase

35. The non-naturally occurring microbial organism of claim 34, wherein said microbial organism comprises two exogenous nucleic acids each encoding a muconate pathway enzyme.

36. The non-naturally occurring microbial organism of claim 34, wherein said microbial organism comprises three exogenous nucleic acids each encoding a muconate pathway enzyme.

37. The non-naturally occurring microbial organism of claim 34, wherein said microbial organism comprises four exogenous nucleic acids each encoding a muconate pathway enzyme.

38. The non-naturally occurring microbial organism of claim 34, wherein said microbial organism comprises five exogenous nucleic acids each encoding a muconate pathway enzyme.

39. The non-naturally occurring microbial organism of claim **33**, wherein said at least one exogenous nucleic acid is a heterologous nucleic acid.

40. The non-naturally occurring microbial organism of claim **33**, wherein said non-naturally occurring microbial organism is in a substantially anaerobic culture medium.

41. A method for producing muconate, comprising culturing the non-naturally occurring microbial organism according to claim **33**, under conditions and for a sufficient period of time to produce muconate.

42.-47. (canceled)

48. A non-naturally occurring microbial organism, comprising a microbial organism having a muconate pathway comprising at least one exogenous nucleic acid encoding a muconate pathway enzyme expressed in a sufficient amount to produce muconate, said muconate pathway comprising an enzyme selected from the group consisting of a lysine aminotransferase, a lysine aminating oxidoreductase, a 2-aminoadipate semialdehyde dehydrogenase, a 2-aminoadipate deaminase, a muconate reductase, a lysine-2,3-aminomutase, a 3,6-diaminohexanoate aminotransferase, a 3,6-diaminohexanoate aminating oxidoreductase, a 3-aminoadipate semialdehyde dehydrogenase, and a 3-aminoadipate deaminase.

49. The non-naturally occurring microbial organism of claim **48**, wherein said muconate pathway comprises a set of muconate pathway enzymes, set of muconate pathway enzymes selected from the group consisting of:

A) (1) lysine aminotransferase, (2) lysine aminating oxidoreductase, (3) 2-aminoadipate semialdehyde dehydrogenase, (4) 2-aminoadipate deaminase, and (5) muconate reductase

B) (1) lysine-2,3-aminomutase, (2) 3,6-diaminohexanoate aminotransferase, (3) 3,6-diaminohexanoate aminating oxidoreductase, (4) 3-aminoadipate semialdehyde dehydrogenase, (5) 3-aminoadipate deaminase, and (6) muconate reductase.

50. The non-naturally occurring microbial organism of claim **49**, wherein said microbial organism comprises two exogenous nucleic acids each encoding a muconate pathway enzyme.

51. The non-naturally occurring microbial organism of claim **49**, wherein said microbial organism comprises three exogenous nucleic acids each encoding a muconate pathway enzyme.

52. The non-naturally occurring microbial organism of claim **49**, wherein said microbial organism comprises four exogenous nucleic acids each encoding a muconate pathway enzyme.

53. The non-naturally occurring microbial organism of claim **49**, wherein said microbial organism comprises five exogenous nucleic acids each encoding a muconate pathway enzyme.

54. The non-naturally occurring microbial organism of claim **49**, wherein said microbial organism comprises six exogenous nucleic acids each encoding a muconate pathway enzyme.

55. The non-naturally occurring microbial organism of claim **48**, wherein said at least one exogenous nucleic acid is a heterologous nucleic acid.

56. The non-naturally occurring microbial organism of claim **48**, wherein said non-naturally occurring microbial organism is in a substantially anaerobic culture medium.

57. A method for producing muconate, comprising culturing the non-naturally occurring microbial organism according to claim **48**, under conditions and for a sufficient period of time to produce muconate.

58.-65. (canceled)

66. A semi-synthetic method for synthesizing terephthalate (PTA) comprising preparing muconic acid by culturing an organism of any one of claims, **2**, **20**, **34**, or **49**, reacting said muconic acid with acetylene to form a cyclohexadiene adduct, and oxidizing said cyclohexadiene adduct to form PTA.

67. The method of claim **66**, further comprising isolating muconic acid prior to reacting with acetylene.

68. The method of claim **66**, wherein the step of reacting said muconic acid with acetylene comprises adding acetylene to the culture broth.

69. The method of claim **68**, wherein the culture broth is filtered prior to adding acetylene.

70. The method of claim **68**, wherein said step of oxidizing said cyclohexadiene adduct comprises adding air or oxygen to the culture broth.

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