



US 20070172854A1

(19) **United States**

(12) **Patent Application Publication**
Matsumura et al.

(10) **Pub. No.: US 2007/0172854 A1**

(43) **Pub. Date: Jul. 26, 2007**

(54) **GENE EXPRESSION ANALYSIS USING
ARRAY WITH IMMOBILIZED TAGS OF
MORE THAN 25 BP (SUPERSAGE-ARRAY)**

Publication Classification

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(51) **Int. Cl.**
C12Q 1/68 (2006.01)
C12M 1/34 (2006.01)
(52) **U.S. Cl.** **435/6; 435/287.2**

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(57) **ABSTRACT**

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This invention provides a method of gene expression analysis that enables extensive gene expression analysis and simultaneous analysis of multiple samples of organisms for which genomic analysis has not yet been advanced. In this method, tags each comprising an oligonucleotide of more than 25 bp for identifying expressed genes, wherein the 3'-end of the tag is defined by a cleavage site of a type III restriction enzyme and the 5'-end thereof is defined by a cleavage site of another restriction enzyme located closest to the 3'-end of the cDNA of such genes, are immobilized on a solid support, gene-containing samples are hybridized to the solid support, and the signals emitted from the genes hybridized to the tags are detected to analyze the gene expression profiles in the samples.

(21) Appl. No.: **11/635,688**

(22) Filed: **Dec. 8, 2006**

(30) **Foreign Application Priority Data**

Dec. 13, 2005 (JP) 2005-359366
May 18, 2006 (JP) 2006-138515

Fig. 1-1

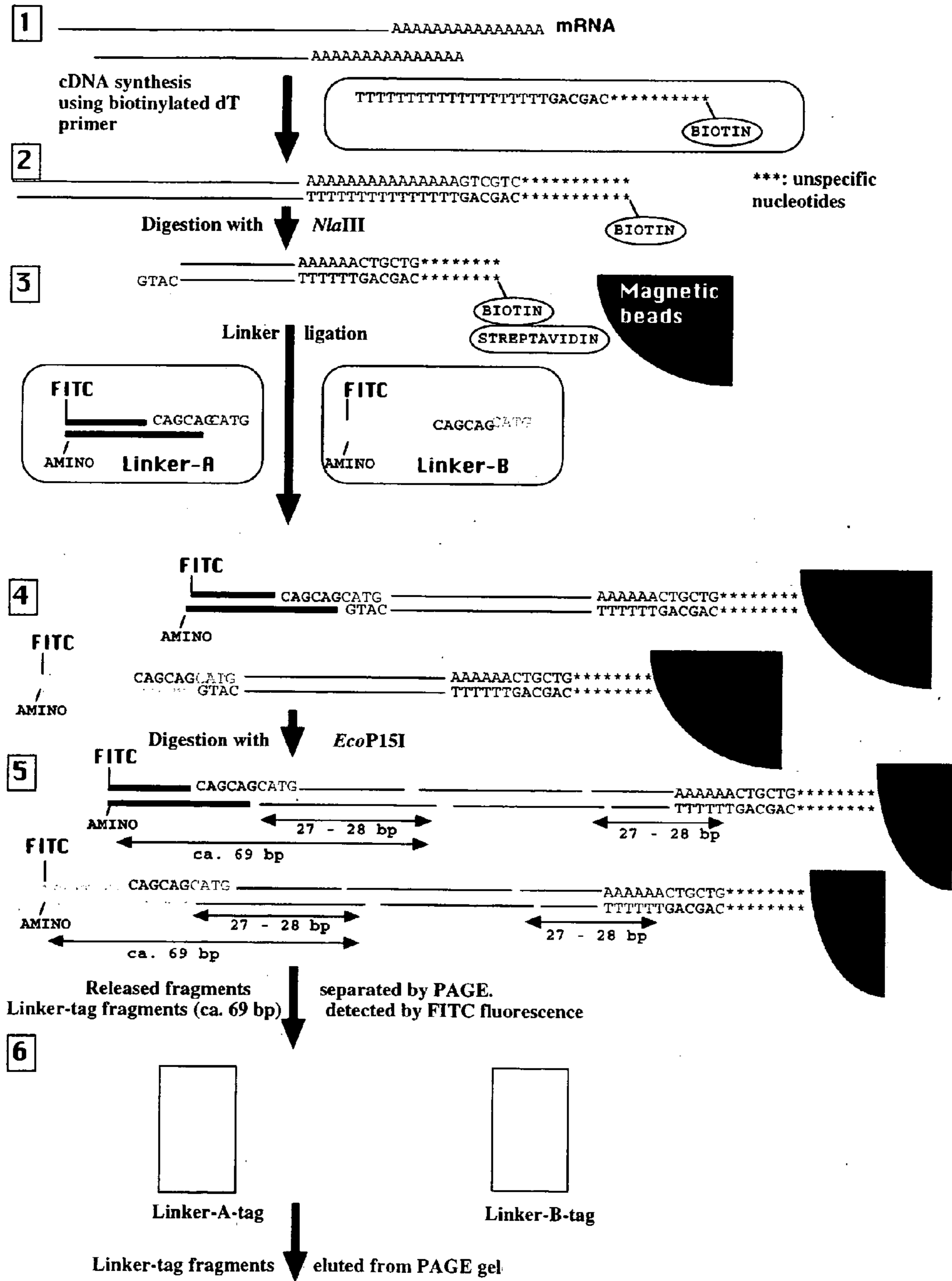


Fig. 1-2

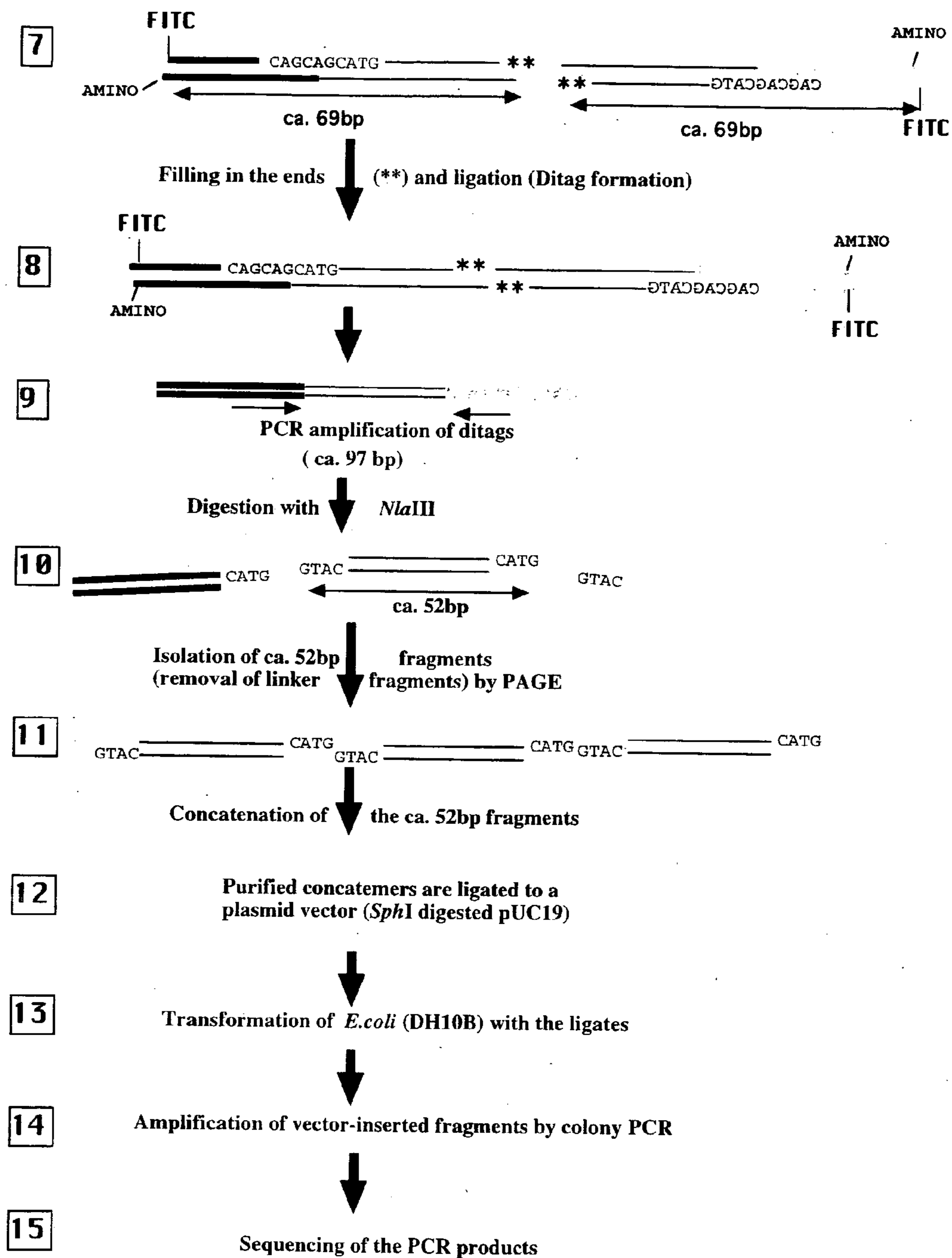


Fig. 2

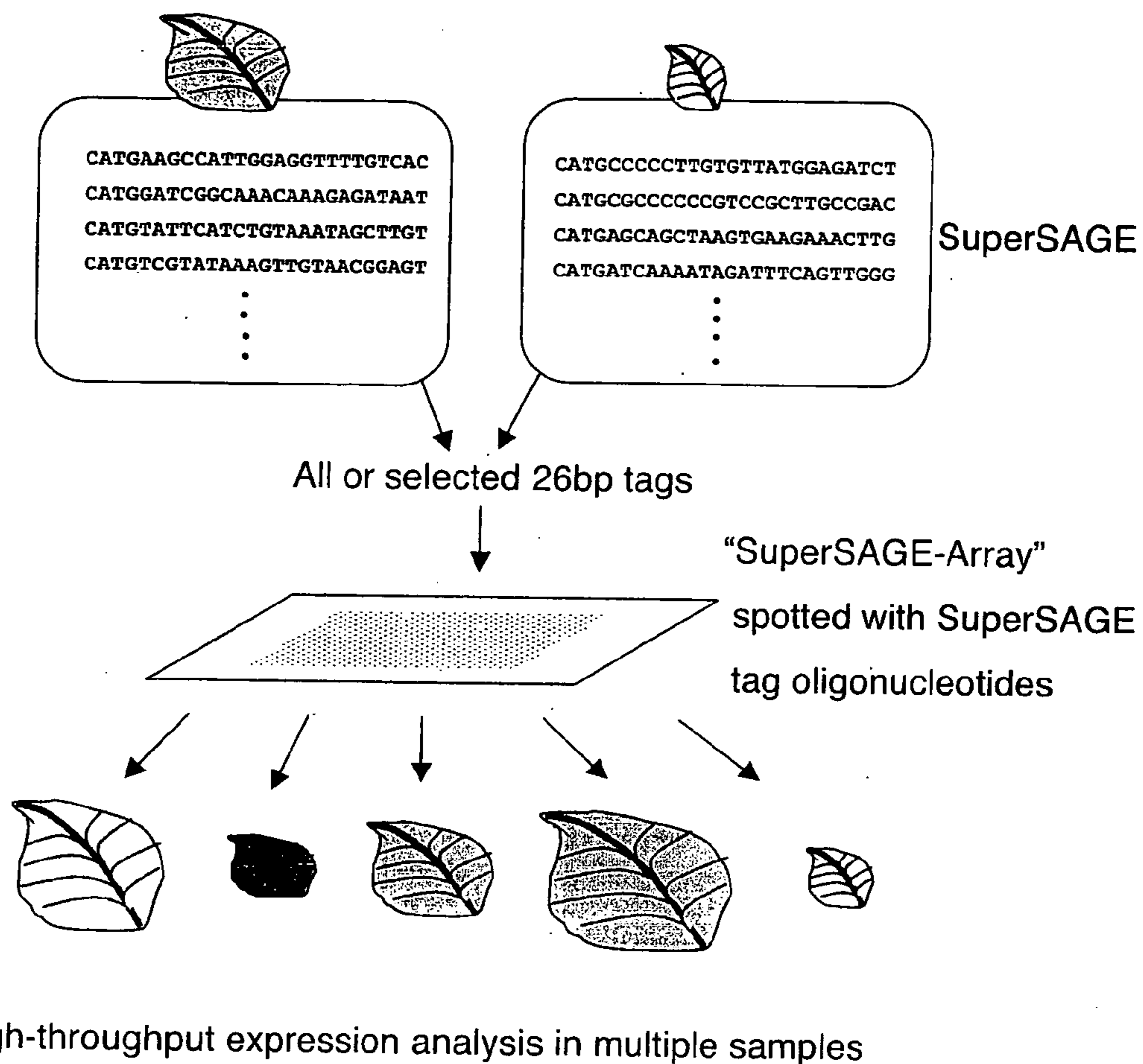


Fig. 3

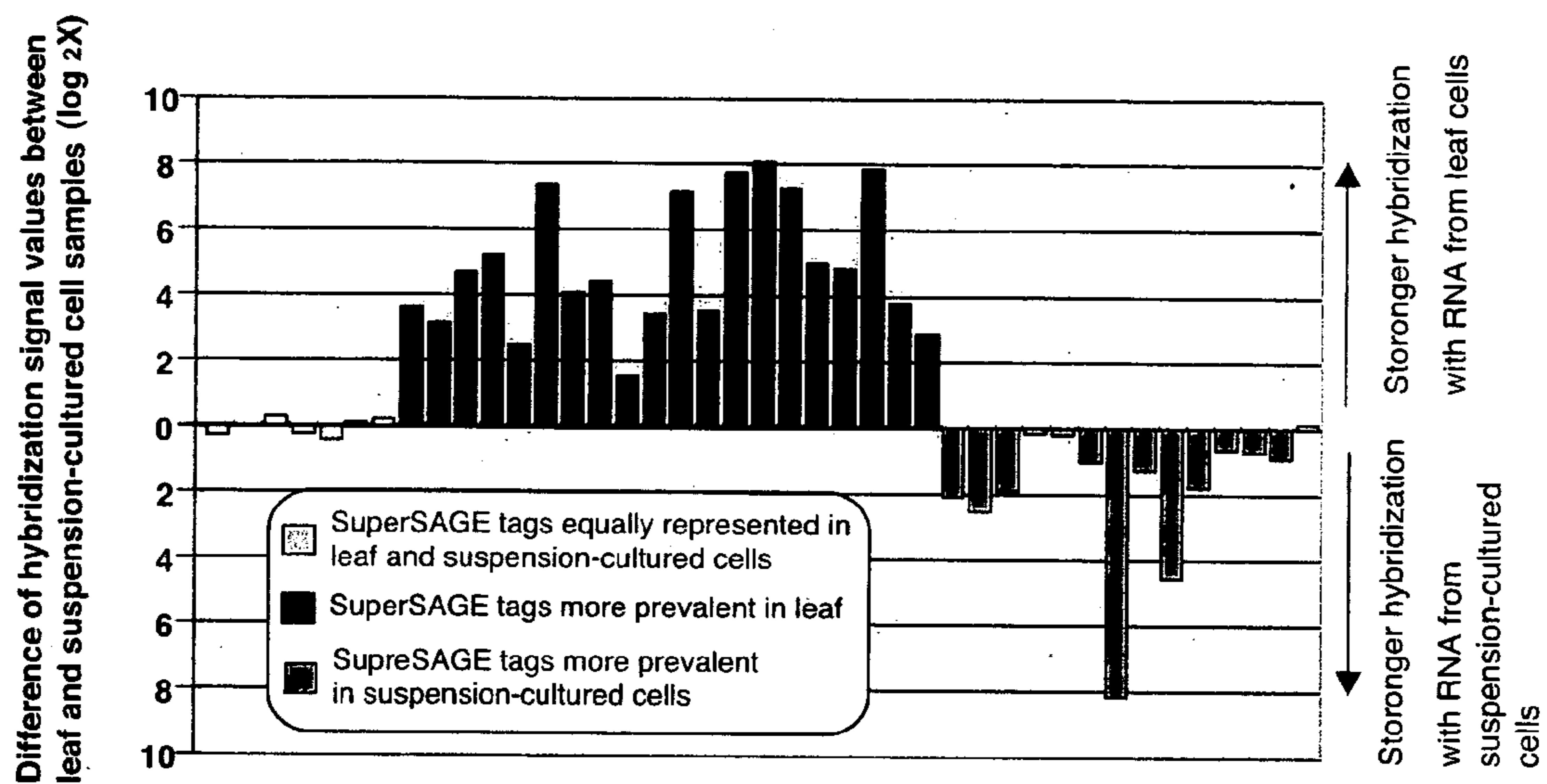


Fig. 4

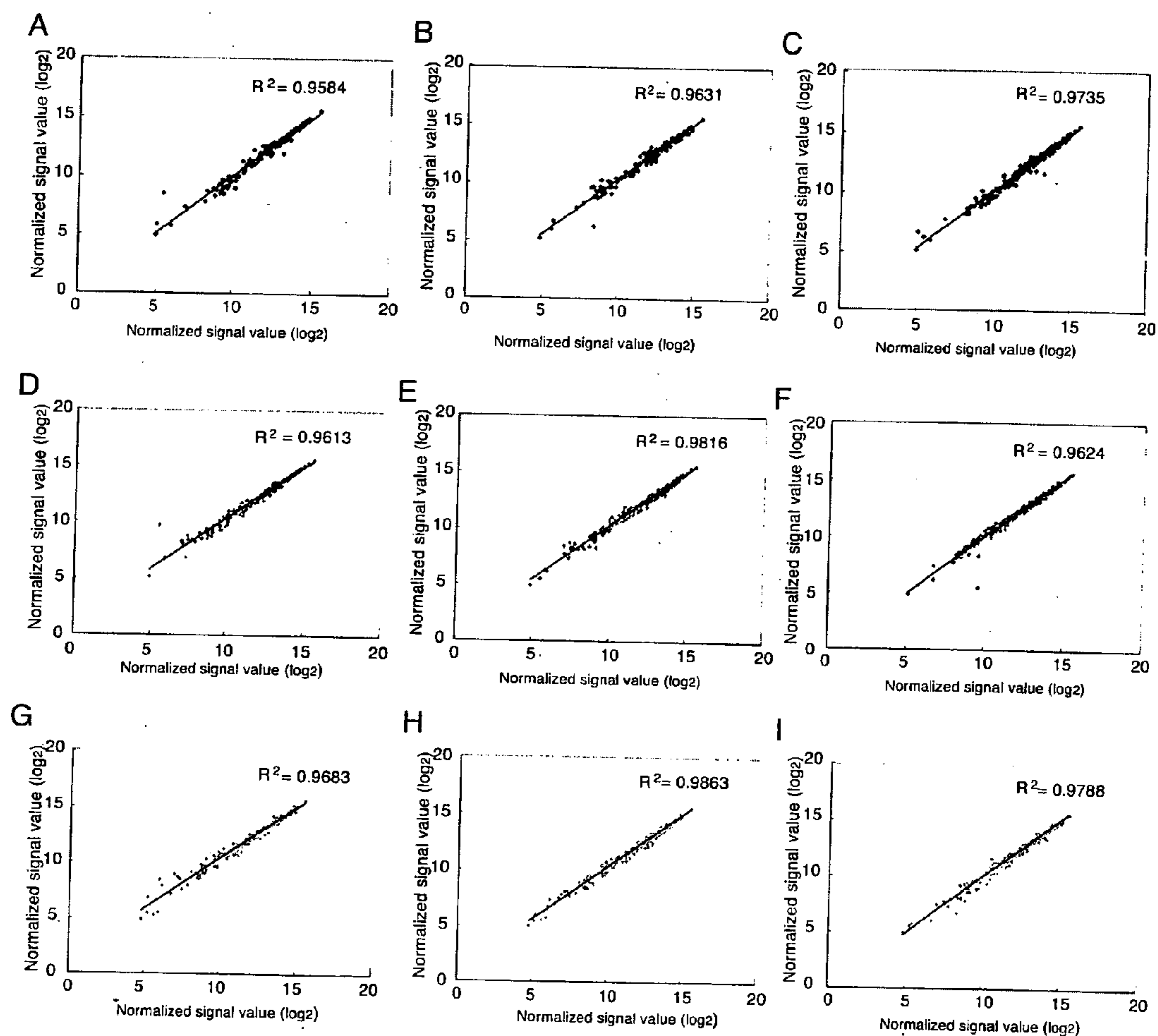


Fig. 5

A	B	C	D
NbCD1U1	NbCD1D1	NbCD3U1	NbCD3D1
NbCD1U2	NbCD1D2	NbCD3U2	NbCD3D2
NbCD1U3	NbCD1D3	NbCD3U3	NbCD3D3
NbCD1U4	NbCD1D4	NbCD3U4	NbCD3D4
NbCD1U5	NbCD1D5	NbCD3U5	NbCD3D5
NbCD1U6	NbCD1D6	NbCD3U6	NbCD3D6
NbCD1U7	NbCD1D7	NbCD3U7	NbCD3D7
NbCD1U8	NbCD1D8	NbCD3U8	NbCD3D8
NbCD1U9	NbCD1D9	NbCD3U9	NbCD3D9
NbCD1U10	NbCD1D10	NbCD3U10	NbCD3D10
NbCD1U11	NbCD1D11	NbCD3U11	NbCD3D11
NbCD1U12	NbCD1D12	NbCD3U12	NbCD3D12
NbCD1U13	NbCD1D13	NbCD3U13	NbCD3D13
NbCD1U14	NbCD1D14	NbCD3U14	NbCD3D14
NbCD1U15	NbCD1D15	NbCD3U15	NbCD3D15
NbCD1U16	NbCD1D16	NbCD3U16	NbCD3D16
NbCD1U17	NbCD1D17	NbCD3U17	NbCD3D17
NbCD1U18	NbCD1D18	NbCD3U18	NbCD3D18
NbCD1U19	NbCD1D19	NbCD3U19	NbCD3D19
NbCD1U20	NbCD1D20	NbCD3U20	NbCD3D20
NbCD1U21	NbCD1D21	NbCD3U21	NbCD3D21
NbCD1U22	NbCD1D22	NbCD3U22	NbCD3D22
NbCD1U23	NbCD1D23	NbCD3U23	NbCD3D23
NbCD1U24	NbCD1D24	NbCD3U24	NbCD3D24
NbCD1U25	NbCD1D25	NbCD3U25	NbCD3D25
NbCD1U26	NbCD1D26	NbCD3U26	NbCD3D26
NbCD1U27	NbCD1D27	NbCD3U27	NbCD3D27
NbCD1U28	NbCD1D28	NbCD3U28	NbCD3D28
NbCD1U29	NbCD1D29	NbCD3U29	NbCD3D28
NbCD1U30	NbCD1D30	NbCD3U30	NbCD3D29
NbCD1U31	NbCD1D31	NbCD3U31	
NbCD1U32	NbCD1D32	NbCD3U32	
NbCD1U33	NbCD1D33	NbCD3U33	
NbCD1U34	NbCD1D34	NbCD3U34	
NbCD1U35	NbCD1D35	NbCD3U35	
NbCD1U36	NbCD1D36	NbCD3U36	
NbCD1U37	NbCD1D37	NbCD3U37	
NbCD1U38	NbCD1D38	NbCD3U38	
NbCD1U39	NbCD1D39	NbCD3U39	
NbCD1U40		NbCD3U40	
NbCD1U41		NbCD3U41	
NbCD1U42		NbCD3U42	
NbCD1U43			
NbCD1U44			

Fig. 6

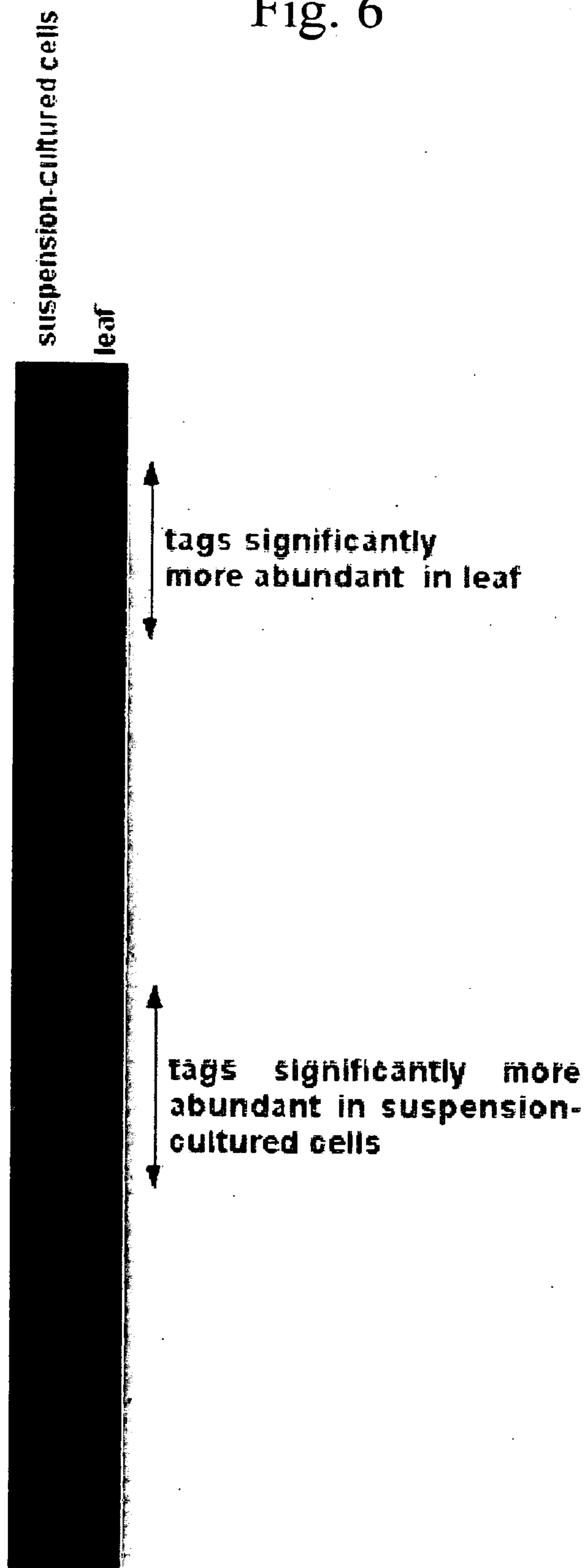
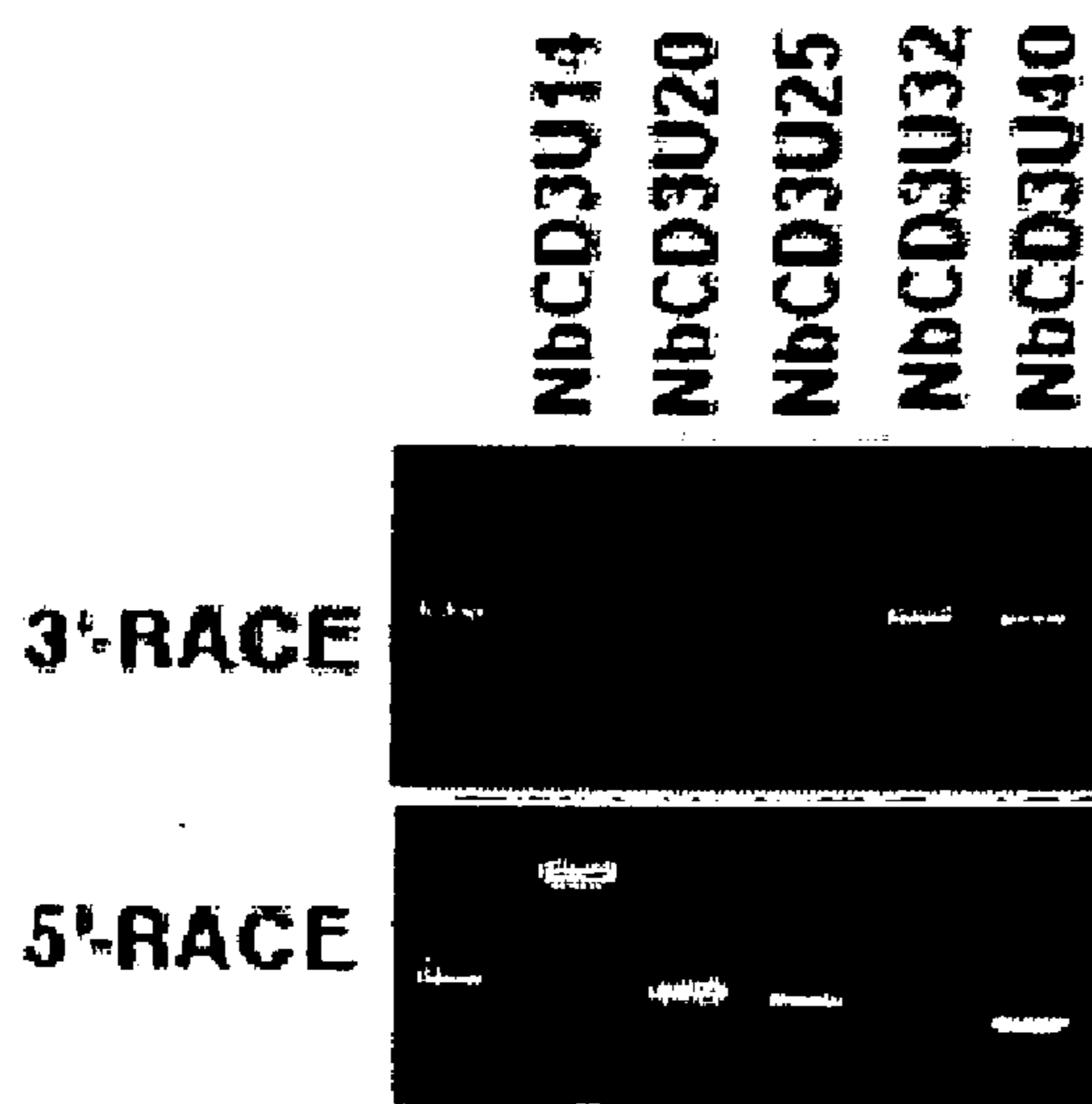
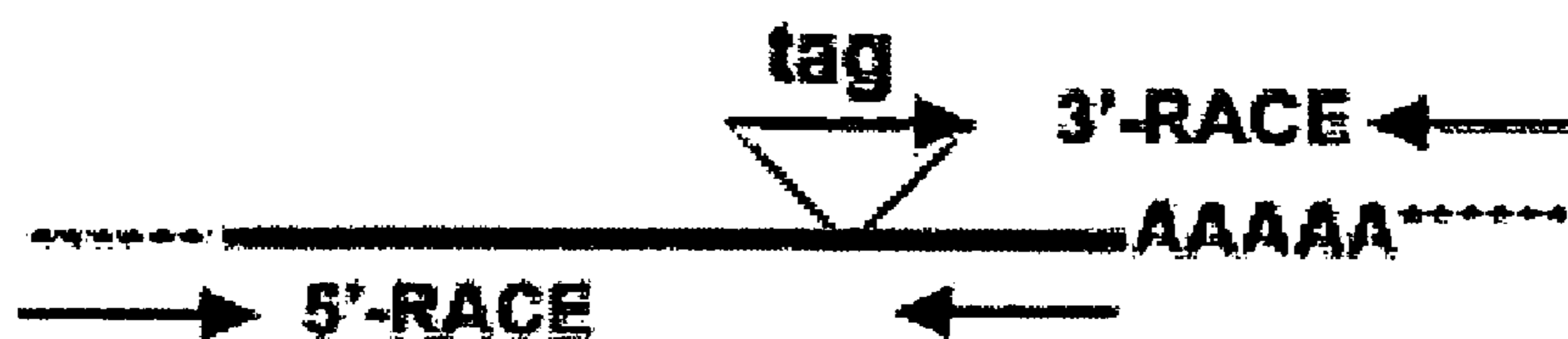


Fig. 7



**GENE EXPRESSION ANALYSIS USING ARRAY
WITH IMMOBILIZED TAGS OF MORE THAN 25
BP (SUPERSAGE-ARRAY)**

CLAIM PRIORITY

[0001] The present application claims priority from Japanese applications JP2005-359366 filed on Dec. 13, 2005 and JP2006-138515 filed on May 18, 2006, the content of which is hereby incorporated by reference into this application.

TECHNICAL FIELD

[0002] The present invention relates to a method of gene expression analysis. More particularly, the present invention relates to a method of gene expression analysis that enables highly reproducible and high-throughput analysis, with the use of a microarray with immobilized improved SAGE™ tags of more than 25 bp.

BACKGROUND ART

[0003] Techniques for transcript analysis, such as microarray analysis and serial analysis of gene expression (SAGE™), are indispensable for various types of biological research. Use of a microarray enables the expression analysis of large quantities of genes at one time and simultaneous analysis of multiple samples. With the use of a microarray, however, expression analysis can only be conducted exclusively for the genes spotted on the array. Accordingly, it is necessary to prepare an array upon which all relevant genes may be spotted, in order to perform extensive analysis. In the case of model organisms, such as rice or *Arabidopsis thaliana*, cDNA arrays or oligonucleotide arrays covering all genes thereof are commercially available and are generally employed in research. Concerning many other organisms, however, researchers are required to independently design arrays from cDNA libraries. This requires large amounts of time and cost.

[0004] In contrast, serial analysis of gene expression (SAGE™) enables the search for novel genes and the quantitative expression analysis thereof (Velculescu et al., Science 270: 484-487, 1995). With this technique, the genes are identified based on a 10- or 11-bp sequence located downstream of the restriction enzyme site (CATG), which is located closest to the 3'-end of the transcript, and the expression levels of such genes are analyzed. Accordingly, sequential reading of the sequences located around the 3'-end with the use of a DNA sequencer enables the extensive expression analysis of genes, including unknown genes. However, SAGE™ is not substantially suitable for simultaneous analysis of multiple samples due to the large number of experimental steps required. In addition, 14-bp SAGE™ tags and 21-bp tags that are employed in LongSAGE™ (Saha et al., Nature Biotechnology 20, 508-512, 2002) are too short to assuredly identify genes. Thus, applications of such tags are restricted to model organisms.

[0005] In recent years, the present inventors had developed the SuperSAGE system, which is an improvement over SAGE™ (WO 2004/099445; Gene expression analysis of plant host-pathogen interactions by SuperSAGE, Matsumura, H., Reich, S., Ito, A., Saitoh, H., Kamoun, S., Winter, P., Kahl, G., Reuter, M., Krueger, D., and Terauchi R., 2003, Proc. Natl. Acad. Sci. U.S.A., 100: 15718-15723; Molekulares Wechselspiel von Wirt und Pathogen: Simul-

tane, genomweite Transkriptprofilierung zweier Organismen mit SuperSAGE, Kahl, G., Winter, P., Matsumura, H., Reuter, M., Krueger, D. and Terauchi R., 2004, Biospektrum 10: 511-513; SuperSAGE, Matsumura, H., Ito, A., Saitoh, H., Winter, P., Kahl, G., Reuter, M., Krueger, D. H. and Terauchi, R., 2005, Cellular Microbiology, 2005, 7: 11-18; and SuperSAGE, a potent transcriptome tool for eukaryotic Organisms, Matsumura, H., Reich, S., Reuter, M., Krueger, D. H., Winter, P., Kahl, G. and Terauchi R., In: S.-M. WANG (ed.) SAGE: Current Technologies and Applications, Horizon Scientific Press, 2004, 77-90). SuperSAGE involves the use of a type III restriction enzyme, EcoP15I, to obtain a 26-bp nucleotide sequence tag. Use of tags each of 26 bp remarkably improves the accuracy of gene identification. Such tags also enable simultaneous analysis of gene expression both in host cells and in pathogen cells, and applications thereof became available with regard to non-model organisms, for which no DNA database is available.

DISCLOSURE OF THE INVENTION

[0006] It is an object of the present invention to provide a method of gene expression analysis that enables extensive gene expression analysis and simultaneous analysis of multiple samples of organisms for which the genomic analysis has not yet been advanced.

[0007] In order to attain the above object, the present inventors had examined whether or not tags each of 26 bp of SuperSAGE (SuperSAGE tags) could be utilized as probes for microarrays. As a result, they discovered that the results of gene expression analysis attained with the use of an array with immobilized SuperSAGE tags would be similar to those attained via conventional SAGE™, and that such results could be attained through a single hybridization step. Further, they also discovered that immobilization of SuperSAGE tags would produce unexpected effects, i.e., preparation of microarrays would be remarkably facilitated in non-model organisms for which no EST, cDNA, or genomic sequences are available.

[0008] More specifically, the present invention relates to a solid support onto which tags each comprising an oligonucleotide of more than 25 bp for identifying the expressed genes are immobilized, wherein the 3'-end of the tag is defined by a cleavage site of a type III restriction enzyme and the 5'-end thereof is defined by a cleavage site of the other restriction enzyme located closest to the 3'-end of the cDNA of such genes. The present invention also relates to a method of gene expression analysis involving the use of such solid support with immobilized tags.

[0009] The sequences of the tags according to the present invention (i.e., SuperSAGE tags) can be determined based on the SuperSAGE system that the present inventors previously developed (WO 2004/099445). Specifically, such tag sequences are determined in accordance with the following steps:

[0010] 1) a cDNA pool is synthesized from mRNAs of expressed genes using a primer comprising a recognition sequence of a type III restriction enzyme and an oligo-dT sequence, and treating the cDNA pool with another restriction enzyme;

[0011] 2) a poly(A)-containing fragment is purified from the cDNA pool, and such fragment is ligated to a linker A or B;

[0012] 3) the fragment is treated with a type III restriction enzyme, and the resulting linker A-containing fragment is ligated to a linker B-containing fragment;

[0013] 4) linker sequences are removed by cleaving the ligated fragments with another restriction enzyme used in step 1) to obtain ditag oligonucleotides;

[0014] 5) ditag oligonucleotides are ligated to each other to prepare polynucleotides; and

[0015] 6) the nucleotide sequences of the above polynucleotides are analyzed to determine the nucleotide sequences of tags contained in such polynucleotides.

[0016] Examples of type III restriction enzymes that can be used in the present invention are disclosed at a web site (<http://rebase.neb.com/cgi-bin/azlist?re3>), and examples thereof include EcoPI and EcoP15I.

[0017] Examples of other restriction enzymes (commercial products) include those shown in the table below.

Recognition sequence	Enzymes (commercial products only)
CATG [^]	NlaIII, Hsp92II,
[^] CATG	FatI
C [^] TAG	BfaI, MaeI, XspI
A [^] CGT	HpyCH4IV, MaeII,
ACGT [^]	TaiI, TscI
AG [^] CT	AluI
T [^] CGA	TaqI
[^] GATC	BfuCI, Bsp143I, BstENII, DpnII, Kzo9I, MboI, NdeII, Sau3AI
GAT [^] C	BstKTI,
G [^] TAC	Csp6I

[0018] As a preferred embodiment of the present invention, use of EcoP15I and N1aIII is described in the Examples below. When EcoP15I and N1aIII are used, the aforementioned linker A and linker B are double-stranded DNAs that are different from each other and that are obtained by annealing the following first strand of DNA (1) and the second strand of DNA (2):

DNA (1):
5'-N₃₀₋₄₀-CAGCAGCATG-3'

DNA (2):
3'-N₃₀₋₄₀-GTCGTC-5'

[0019] wherein "N₃₀₋₄₀" of DNA (1) is complementary to "N₃₀₋₄₀" of DNA (2), each thereof is a sequence comprising 30 to 40 arbitrary nucleotides, the 5'-end of DNA (1) may be labeled, and the 3'-end of DNA (2) may be amino-modified.

[0020] The array according to the present invention may be prepared by synthesizing tag oligonucleotides on the solid support. Alternatively, the array may be prepared by immobilizing pre-synthesized tag oligonucleotides on the solid support.

[0021] The present invention facilitates the extensive gene expression analysis and simultaneous analysis of multiple samples of organisms for which genomic analysis has not yet been advanced. Accordingly, the present invention overcomes the drawbacks of conventional microarray or SAGE™ systems and realizes extensive analysis of known and unknown genes.

BRIEF DESCRIPTION OF THE DRAWINGS

[0022] FIG. 1-1 schematically shows a method for preparing SuperSAGE tags.

[0023] FIG. 1-2 schematically shows a method for preparing SuperSAGE tags.

[0024] FIG. 2 schematically shows the SuperSAGE-Array according to the present invention. SuperSAGE detects a 26-bp tag sequence of a gene with an expression level that varies among 2 or more samples. Microarrays are prepared with oligonucleotides of these tag sequences (SuperSAGE-Array). These arrays enable high-throughput analysis of multiple samples.

[0025] FIG. 3 shows the results of gene expression analysis via SuperSAGE-Array analysis in rice leaves and in cultured cells. The heights of the bar charts represent the differences of the standardized signal values between rice leaves and cultured cells (represented in terms of log values). The more the bar chart extends upward from level 0, the stronger the signals in leaves. The more the bar chart extends downward from level 0, the stronger the signals in cultured cells. The colors of the bars represent the expression patterns observed via SuperSAGE (purple: the same expression levels among samples; red: high expression levels in leaves; orange: high expression levels in cultured cells).

[0026] FIG. 4 shows charts representing the reproducibility of a SuperSAGE-Array of foreign gene-overexpressing *Nicotiana benthamiana* leaves.

[0027] FIG. 5 shows the gene expression profiles in NbCD1- or NbCD3-overexpressing *Nicotiana benthamiana* leaves analyzed via SuperSAGE-Array. Based on the results of SuperSAGE-Array analysis, genes were classified as follows: (A) genes induced upon NbCD1 overexpression; (B) genes repressed upon NbCD1 overexpression; (C) genes induced upon NbCD3 overexpression; and (D) genes repressed upon NbCD3 overexpression. Compared with the control GFP-overexpressing leaves, stronger signals are represented by red tiles and weaker signals are represented by green tiles.

[0028] FIG. 6 shows the results of SuperSAGE-Array analysis of differences in the gene expression profiles between rice leaves and cultured cells. Expression levels that were higher in leaves than in cultured cells are indicated by red tiles, and those that were higher in cultured cells than in leaves are indicated by green tiles.

[0029] FIG. 7 schematically shows the amplification of full-length sequences corresponding to 5 tags derived from *Nicotiana benthamiana* (NbCD3U14, 20, 25, 32, and 40) by the 3'-RACE and 5'-RACE methods (upper portion) and the results of electrophoresis of the PCR products thereof (lower portion).

PREFERRED EMBODIMENTS OF THE INVENTION

[0030] The present invention relates to a method for expression analysis of genes of various organisms with the use of a solid support with immobilized tags of more than 25 bp obtained via SuperSAGE. The solid support (array) with immobilized tags used in the present invention is prepared by SuperSAGE in combination with microarray techniques.

This array is referred to as a “SuperSAGE-Array” herein (FIG. 2). A method for preparing a SuperSAGE-Array and a method of gene expression analysis using the same are hereafter described in detail.

1. Preparation of SuperSAGE Tag

[0031] A method for determining a tag sequence of more than 25 bp (a SuperSAGE tag) for identifying a gene of a target organism via the SuperSAGE method developed by the present inventors is first described with reference to FIG. 1. Details of SuperSAGE as employed herein are described in the following publications: Matsumura, H., et al., 2003, Proc. Natl. Acad. Sci. U.S.A., 100: 15718-15723; Kahl, G. et al., 2004, Biospektrum 10: 511-513; Matsumura, H. et al., 2005, Cellular Microbiology, 2005, 7: 11-18; Matsumura, H. et al., Current Technologies and Applications, Horizon Scientific Press, 2004, 77-90; and WO 2004/099445.

[0032] In accordance with a conventional technique, total RNA or mRNA is prepared from a specimen of a target organism, and cDNA is synthesized with the use of such total RNA or mRNA as a template and a biotinylated oligo-dT primer (a reverse transcription primer). This reverse transcription primer comprises 18 to 25 nucleotides, the 5'-CAGCAG-3' sequence, which is a recognition sequence of the EcoP15I enzyme, and 15- to 25-mer dT subsequent thereto (Step 1, FIG. 1). The type III restriction and modifying enzyme, EcoP15I, recognizes CAGCAG sequences from among the target DNA molecules and cleaves the recognized sequence at a site 25 to 28 nucleotides away from the 3'-end thereof.

[0033] The synthesized cDNA is digested with a CATG-recognizing restriction enzyme, N1aIII, and a digestion fragment containing the biotinylated reverse transcription primer sequence is solely recovered by streptavidin-coated magnetic beads (Steps 2 and 3, FIG. 1).

[0034] Subsequently, two types of linkers, i.e., linker A and linker B (each comprising 46 nucleotides), are ligated to the ends of a cDNA fragment captured by magnetic beads. The linker comprises a CAGCAG sequence, which is a recognition sequence of the EcoP15I enzyme (Step 4, FIG. 1).

[0035] The obtained cDNA pool is divided into two sections, and one of them is ligated to linker A and the other is ligated to linker B. The resultants are designated as “linker A-ligated cDNA” and “linker B-ligated cDNA,” respectively.

[0036] DNA fragments of both “linker A-ligated cDNA” and “linker B-ligated cDNA” that had been bound to magnetic beads are each digested with EcoP15I and magnetic beads are then removed (Step 5, FIG. 1). After the digestion, one of the fragments comprises a linker and the 27- or 28-bp tag sequence (with a total size of 69 or 70 bp), and the other fragment is of a variable size contained in a double-stranded cDNA fragment. A poly(A)-containing fragment remains bound to magnetic beads, and this fragment is not involved in subsequent procedures.

[0037] A 69- or 70-bp fragment comprising a linker sequence and the 27- or 28-bp tag sequence is visualized with FITC under UV irradiation, and such fragment is isolated from polyacrylamide gel via gel excision.

[0038] Fragments of 69 or 70 bp obtained from linker A-ligated cDNA and linker B-ligated cDNA (i.e., tag-linker fragments) are blunted via a 3'-filling reaction, and these fragments are ligated to each other to form “ditags.” The 3'-end of the linker fragment is blocked via amino-modification. Thus, such ligation takes place only at a site between cDNA tag sequences (Steps 6 and 7, FIG. 1).

[0039] The resulting “ditag” molecules are amplified by PCR (Step 8, FIG. 1). PCR primers are constructed from linker sequences and the PCR products are of ca. 97 bp.

[0040] The PCR products of ca. 97 bp are digested with N1aIII (Step 10, FIG. 1) to result in “ditag” fragments of ca. 52 bp. These fragments are isolated from gel and then purified.

[0041] The “ditag” fragments are further ligated (Step 11, FIG. 1) to result in concatemers. The concatemers are subjected to agarose gel electrophoresis, and fragments of 500 or more bp are isolated from gel.

[0042] The size-separated concatemer fragments are cloned into adequate plasmid vectors that have been treated with SphI and CIP (Step 12, FIG. 1). The resulting plasmids are then transformed into *E. coli* (Step 13, FIG. 1).

[0043] Subsequently, fragments that were inserted into plasmids are amplified by PCR (Step 14, FIG. 1).

[0044] A nucleotide sequence is directly read from the PCR product (Step 15, FIG. 1). A ditag of ca. 44 bp is present and is flanked by the recognition sequences of N1aIII, i.e., CATG. Information concerning a ca. 52-bp (44+8) sequence indicates two 26-bp to 28-bp tag sequences isolated from specific regions in each cDNA.

[0045] Via sequencing of several clones of nucleotide sequences, SuperSAGE tag sequences derived from various organisms shown in Tables 1 to 5 are obtained.

2. Tag Immobilization—Preparation of SuperSAGE-Array

[0046] Based on the sequence information concerning the resulting SuperSAGE tag oligonucleotides are synthesized and immobilized on an adequate solid support to prepare the array according to the present invention (e.g., SuperSAGE-Array). The array according to the present invention is not limited to a microarray, and examples thereof include a bead array, a membrane filter, and a capillaries. In the step of synthesizing such tags, desired functional groups may be introduced onto the ends of the tags with the use of primers labeled at their 5'-ends with functional groups such as thiol groups, in order to enhance the efficiency of immobilizing tags upon the substrate.

[0047] Tags may be immobilized upon a substrate by any means without particular limitation. Oligonucleotides may be directly synthesized on a substrate made of glass, metal, silicon, or the like (Affymetrix type). Pre-synthesized oligonucleotides may be spotted to a substrate (standard type). Alternatively, tags may be immobilized in a filter made of nylon, nitrocellulose, or the like. A glass substrate or the like has a small effective area for immobilization and a low charge. Thus, the surface of such substrate is preferably treated with polysilane, silane, polycarbodiimide, aminosilane, or the like, in order to enhance the efficiency of tag immobilization. Use of commercially available substrates,

such as polylysine- or silane-coated glass substrates, the surfaces of which are treated with the aforementioned substances, is also preferable.

[0048] In general, pre-synthesized tags are automatically immobilized on a substrate using a spotter. In order to compare tag information and signals transmitted from tags, it is preferable that the sites of gene immobilization be fully known. Sites of tag immobilization are not particularly limited, provided that such comparison can be made.

3. Gene Expression Analysis Using SuperSAGE-Array

[0049] In accordance with a conventional technique, total RNA is extracted from a sample to prepare mRNA or cDNA. The target is labeled with an adequate fluorescent reagent (e.g., Cy3-UDP or Cy5-UDP) in advance, when mRNA or cDNA is prepared from the sample. The labeled target is hybridized to the aforementioned substrate with an immobilized tag and then washed. Thereafter, fluorescence intensities (signal intensities) at the sites of tag immobilization are assayed. The fluorescence intensities read with a scanner may be subjected to error adjustment or normalization of variations among samples, according to need. Normalization can be carried out based on the genes that are commonly expressed in samples, such as housekeeping genes. Further, a reliability limit may be determined to eliminate the data exhibiting low correlation.

4. Application of SuperSAGE-Array

[0050] Gene expression analysis that is carried out using the solid support according to the present invention (SuperSAGE-Array) can be a useful and fundamental technique for extensive gene expression analysis for any eukaryotic organism. Many of currently available microarray techniques depend on EST, cDNA, or genomic sequences. Non-redundant cDNA sequences are required for the preparation of arrays. To this end, it is necessary to prepare a cDNA library that is normalized or that represents the genes differentially expressed between the two samples. The solid support with immobilized tags (SuperSAGE-Array) according to the present invention becomes a crucial technique for preparing such arrays.

[0051] According to the present invention, information concerning a large number of expressed genes can be attained within a shorter period of time than is possible with large-scale EST analysis. The SuperSAGE tags that are employed in the present invention are obtained from specific sites in the exons. Thus, each tag is unique to each gene. With the utilization of such characteristics, oligoarrays of expressed genes can be easily prepared for any tissues or under any conditions.

[0052] For example, the SuperSAGE-Array obtained from a variety of cancer tissues is useful for clinical testing, and SuperSAGE-Arrays can be used for all eukaryotic organisms. If the tags exhibiting the expression patterns of interest were found, a partial cDNA fragment can be obtained by the 3'-RACE method using a tag sequence of 26 bp. The RACE product may be subjected to BLAST search to identify the genes represented by such tags.

[0053] Microarrays of host organism and pathogen genes can be applied to the host-pathogen interaction analysis. The present inventors can easily prepare such arrays via the SuperSAGE-Array system. By using SuperSAGE for patho-

gen-infected tissues, an array with host and pathogen genes spotted thereon can be easily prepared. Use of such array enables high-throughput analysis of expression changes of genes derived from both host and pathogen, for example.

EXAMPLES

[0054] The present invention is hereafter described in greater detail with reference to the examples, although the present invention is not limited to these examples.

Example 1

1. Material and Method

1) Preparation of RNA

[0055] For rice SuperSAGE and oligoarray systems, rice leaves (variety: Yashiromochi) and suspension-cultured cells (variety: Kakehashi) were prepared. For the oligoarray system, mRNA was extracted from the rice (variety: Yashiromochi) and the cultured cells (variety: Kakehashi) 1 month after sowing using an mRNA Purification Kit (Amersham Pharmacia).

[0056] For *Nicotiana benthamiana* SuperSAGE and oligoarray systems, leaves into which *Agrobacterium* containing the following plasmids had been injected were prepared. Two days after the *Agrobacterium* injection, *Nicotiana benthamiana* leaves were treated with dexamethasone (DEX), and mRNA was extracted using the mRNA Purification Kit (Amersham Pharmacia) 4 hours later.

2) Plasmids

[0057] NbCD1 (JP Patent Publication (Unexamined) No. 2005-278634) and NbCD3 cDNA (JP Patent Publication (Unexamined) No. 2005-245251), which had been isolated by the present inventors in the past, were used. A binary plasmid having these cDNAs and GFP cDNA downstream of a GVG promoter (Aoyama T. and Chua N.-H., 1997, The Plant Journal, 1997, 11: 605-612) capable of specific induction of gene expression via glucocorticoid (dexamethasone) treatment was prepared.

3) SuperSAGE

[0058] mRNAs extracted from rice and *Nicotiana benthamiana* were purified to obtain 3 to 5 μ g of poly(A) RNA. A SuperSAGE library was prepared from this poly(A) RNA and data analysis was carried out in accordance the publications (cited above).

[0059] Specifically, the obtained poly(A) RNA was employed as a template and the oligo-dT primer (reverse transcription primer: CTGATCTAGAGGTACCGGATCCAGCAGTTTTTTTTTTTTTTTTTTTTT (SEQ ID NO: 8)) that had been biotinylated by the "cDNA Synthesis System" (Invitrogen) was used to synthesize cDNA. This reverse transcription primer comprises a 22 bp sequence, the 5'-CAGCAG-3' sequence, which is a recognition sequence of the EcoP15I enzyme, and 19-mer dT subsequent thereto.

[0060] The synthesized cDNA was digested with the N1aIII restriction enzyme, and the digestion fragment containing the biotinylated reverse transcription primer sequence was selectively recovered with the use of streptavidin-coated magnetic beads.

[0061] Subsequently, the following two types of linkers, i.e., linker A and linker B (each comprising 46 nucleotides), were bound to the ends of a cDNA fragment captured by magnetic beads. These linkers comprised a CAGCAG sequence, which is a recognition sequence of the EcoP15I enzyme.

[0062] Linker A is a double-stranded sequence obtained by annealing the following two oligonucleotides.

FITC-5'-TTTGGATTGCTGGTGCAGTACAAC (SEQ ID NO: 197)

AGGCTTAATACAGCAGCATG-3'

5'-CTGCTGTATTAAGCCTAGTTGTACTGCACCA (SEQ ID NO: 198)

GCAAATCCAAA-3'-NH₂.

[0063] Linker B is a double-stranded sequence obtained by annealing the following two oligonucleotides.

FITC-5'-TTTCTGCTCGAATTCAAGCTTCTAAC (SEQ ID NO: 199)

GATGTACGCAGCAGCATG-3'

5'-CTGCTGCGTACATCGTTAGAAAGCTTGAATTC (SEQ ID NO: 200)

GAGCAGAAA-3'-NH₂.

[0064] The obtained cDNA pool was divided into two sections, one of them was ligated to linker A and the other was ligated to linker B. The resultants were designated as "linker A-ligated cDNA" and "linker B-ligated cDNA," respectively. They were digested with EcoP15I, magnetic beads were removed, and a fragment comprising a linker and the 27- or 28-bp tag sequence (with a total size of 69 or 70 bp) and a fragment of a variable size contained in a double-stranded cDNA fragment were obtained. A poly(A)-containing fragment remained bound to magnetic beads, and this fragment was not involved in subsequent procedures.

[0065] A 69- or 70-bp fragment comprising a linker sequence and the 27- or 28-bp tag sequence was visualized with FITC under UV irradiation, and such fragment was isolated from polyacrylamide gel via gel excision.

[0066] Fragments of 69- or 70-bp obtained from linker A-ligated cDNA and linker B-ligated cDNA (i.e., tag-linker fragments) were blunted via a 3'-filling reaction, and these fragments were ligated to each other to form "ditags."

[0067] The resulting "ditag" molecules were amplified by PCR using primers containing linker sequences to obtain

PCR products each of 97 bp. The PCR products of ca. 97 bp were digested with N1aIII to result in "ditag" fragments of ca. 52 bp. These fragments were isolated from gel and then purified.

[0068] The "ditag" fragments were further ligated to result in polynucleotides (concatemers). The concatemers were subjected to agarose gel electrophoresis, and fragments of 500 or more bp were isolated from gel.

[0069] The size-separated concatemer fragments were cloned into adequate plasmid vectors that have been treated with SphI and CIP. The resulting plasmids were then transformed into *E. coli*.

[0070] Subsequently, fragments that were inserted into plasmids were amplified by PCR, and the nucleotide sequences of the PCR products were analyzed with a sequencer. The PCR products each comprised a ditag of ca. 52 bp flanked by the recognition sequences of N1aIII, i.e., CATG. Information concerning a ca. 52 bp sequence indicated two 26-bp tag sequences isolated from specific regions in each cDNA.

[0071] Via sequencing of several clones of nucleotide sequences, SuperSAGE tag sequences derived from various organisms shown in Tables 1 to 5 were obtained.

4) Oligonucleotide Array Analysis

[0072] A SuperSAGE-Array was prepared using the 12-well NimbleGen array system. The design of the SuperSAGE-Array is described below in detail. Total RNA (20 µg each) was prepared from tissues, double-strand cDNAs were synthesized, and biotinylated cDNA probes were prepared via in vitro transcription. These biotinylated probes were labeled with Cy3 fluorescent dye. The labeled probes were hybridized, signals were read with a scanner, and the data of the signal values were standardized by the Robust Multi-chip Analysis (RMA) method. Array preparation and hybridization were entrusted to Gene Frontier.

2. SuperSAGE-Array of Model Rice Species

[0073] In order to actually test the performance of the SuperSAGE-Array, the transcript profiles in rice leaves and in cultured cells were first analyzed by the SuperSAGE method (WO 2004/099445). The tag sequences each of 26 bp (10,968 tags in the case of leaves and 10,044 tags in the case of cultured cells) were compared between two samples, and oligonucleotide arrays were prepared by selecting 7 tags expressed at substantially the same level in the two samples, 20 tags expressed at high levels only in leaves, and 14 tags expressed at high levels only in cultured cells (Table 1).

TABLE 1

SuperSAGE tags from rice spotted onto the SuperSAGE-Array				
Code	Tag sequence (5'→3')	Number of tags		
		Leaf	Suspension-cultured	SEQ ID NO.
RSpCon1	CATGAATTGAGTTCGCTTTGGTTATG	78	98	SEQ ID NO:1
RSpCon2	CATGGTTTGGTTGGATTAGCGGAGT	40	26	SEQ ID NO:2

TABLE 1-continued

<u>SuperSAGE tags from rice spotted onto the SuperSAGE-Array</u>				
Code	Tag sequence (5'→3')	<u>Number of tags</u>		SEQ ID NO.
		Leaf	Suspension-cultured cell	
RSpCon3	CATGGGCTAAAGCCAGCCAAACTGGT	12	21	SEQ ID NO:3
RSpCon4	CATGTCGGTTCAGTTATGTGAACTTG	10	24	SEQ ID NO:4
RSpCon5	CATGTAATGTTTGCTATCGTGAGTTA	10	11	SEQ ID NO:5
RSpCon6	CATGGCTGACCCAGCCTTCCGTCCAC	10	10	SEQ ID NO:6
RSpCon7	CATGGGAGCGACTCCGTGGACAACGG	12	19	SEQ ID NO:7
RSpL1	CATGTTTCGGCTGCACCGATGCCACCC	36	0	SEQ ID NO:9
RSpL2	CATGGGGACGCATCGCCTTCAGCTAA	35	0	SEQ ID NO:10
RSpL3	CATGTAATATGATGCCTAGAGCATAT	30	0	SEQ ID NO:11
RSpL4	CATGTAATGGTACATATCTCCTTGTT	29	0	SEQ ID NO:12
RSpL5	CATGCTCAAGATGATCGAGGACTACC	26	0	SEQ ID NO:13
RSpL6	CATGTATGTATGTCCCTTAATTGTGT	26	0	SEQ ID NO:14
RSpL7	CATGTTGATATTGTATCAGCAAGCAC	26	1	SEQ ID NO:15
RSpL8	CATGGACGAGCGCGACCCAAGATCC	25	0	SEQ ID NO:16
RSpL9	CATGGCGCAGGAGGTGCTTCTCGGCG	24	0	SEQ ID NO:17
RSpL10	CATGTACTACTACCTTGAACTTTT	24	0	SEQ ID NO:18
RSpL11	GATGTTTCGGGTGCACTGACGCCACCC	24	0	SEQ ID NO:19
RSpL12	CATGGGGATTGTGCACGCATCTGGC	21	0	SEQ ID NO:20
RSpL13	CATGTGTGTACGTGGTGTGTTTTGAG	20	0	SEQ ID NO:21
RSpL14	CATGCATAATTGAACGCTTGTCTGTC	18	0	SEQ ID NO:22
RSpL15	CATGTGTAATACTGCCGTGTGTTTC	18	0	SEQ ID NO:23
RSpL16	CATGGATCCGTCTCTCTGGGAGGAAT	120	0	SEQ ID NO:24
RSpL17	CATGTCGGACAAGTGC GGCAACTGCG	111	0	SEQ ID NO:25
RSpL18	CATGTGGTGGCTTAGCTCTACGTGTA	96	0	SEQ ID NO:26
RSpL19	CATGTTGTAATACTCCATCAAAGAGT	81	0	SEQ ID NO:27
RSpL20	CATGCATATGTGAATGCTAGCACCAG	48	0	SEQ ID NO:28
RSpSus1	CATGTGCTGTTGTGGCGTGTCTGCTAG	0	60	SEQ ID NO:29
RSpSus2	CATGCTTCAATATATATCCATCAAAT	0	48	SEQ ID NO:30
RSpSus3	CATGGATTTGCACTGTCTGATCTATC	1	46	SEQ ID NO:31
RSpSus4	CATGCACAACAGCACAAGTGGAGTAG	0	40	SEQ ID NO:32
RSpSus5	CATGGTAATGTACCAAACAGCGATGA	1	36	SEQ ID NO:33
RSpSus6	CATGCGTTTGTGGCAAGAAGACAAT	3	28	SEQ ID NO:34
RSpSus7	CATGCTGGTAGCTCAGCGAATCTCCT	1	20	SEQ ID NO:35
RSpSus8	CATGGCACGGTTACCCGTCATTTCCG	1	21	SEQ ID NO:36
RSpSus9	CATGATGATGGCCGCCACCGCCACCG	0	18	SEQ ID NO:37
RSpSus10	GATGCGAGTTCCCGGGCTCAAGATC	0	19	SEQ ID NO:38

TABLE 1-continued

SuperSAGE tags from rice spotted onto the SuperSAGE-Array				
Code	Tag sequence (5'→3')	Number of tags		
		Leaf	Suspension-cultured	SEQ ID NO.
RSpSus11	CATGGCCACCGCTACCAACGGCAACG	0	26	SEQ ID NO:39
RSpSus12	CATGGTCGCCGCCGCCGTGCCGGAGC	0	32	SEQ ID NO:40
RSpSus13	CATGTGTGTTGTGTGTACGATGAGCT	0	20	SEQ ID NO:41
RSpSus14	CATGTTAAGTTTGAGATATGATATGA	0	15	SEQ ID NO:42

[0074] As described above, the SuperSAGE-Array was prepared using the 12-well array system (NimbleGen Inc. Co.) with direct synthesis of a 26-bp oligonucleotide on a glass substrate. Concerning the tag sequences, oligonucleotides carrying two-base mismatches in positions 7 and 13 of 26-bp SuperSAGE tag sequences were also synthesized and used for a hybridization specificity test.

[0075] Subsequently, total RNAs were extracted from rice leaves and cultured cells, labeled cRNAs were synthesized therefrom, and the resultants were used as SuperSAGE-Array hybridization probes. As shown in FIG. 3, 7 tags more or less expressed at the same levels between two samples exhibited the same level of hybridization signals between the two samples. The 20 tags more prevalent in leaves actually exhibited stronger hybridization signals in leaf RNA. Among the 14 tags more prevalent in cultured cells, 11 thereof exhibited stronger signals in RNA obtained from cultured cells.

[0076] As a result, the results of SuperSAGE analysis of genes and those of SuperSAGE-Array analysis were found to be consistent among 90% or more genes (tags) (38 of 41 tags).

3. SuperSAGE-Array of Non-Model Organism, *Nicotiana benthamiana*

[0077] As described above, SuperSAGE is suitable for searching for novel genes of non-model organisms. In order to demonstrate that SuperSAGE-Array is applicable to non-

model organisms, a microarray was prepared using 26-bp SuperSAGE tag sequences for genes with expression levels that vary upon overexpression of foreign genes in *Nicotiana benthamiana* leaves, for which no genomic information is available.

[0078] The NbCD1 and NbCD3 genes were isolated from *Nicotiana benthamiana* in the past as genes encoding proteins that induce cell death due to overexpression. At the outset, the *Nicotiana benthamiana* genes with expression levels that are increased or decreased upon expression of the NbCD1 and NbCD3 genes were searched for. *Agrobacterium* having plasmids of NbCD1, NbCD3, and GFP genes incorporated into the glucocorticoid-inducible expression cassette GVG were introduced into *Nicotiana benthamiana* leaves. Two days after the introduction, the leaves were treated with dexamethasone (DEX) to induce the transgenes to express. RNAs were extracted from the treated leaves 4 hours after DEX treatment and then subjected to SuperSAGE. By comparing SuperSAGE profile, tags that shows more than four-fold representational differences between NbCD1- or NbCD3- and GFP-overexpressing leaves were selected (Tables 2 to 5).

[0079] A SuperSAGE-Array upon which the selected tags were immobilized was prepared. In total, 154 types of tags with expression levels that were increased or decreased upon NbCD1 and NbCD3 overexpression were selected for the oligonucleotide array (Tables 2 to 5).

TABLE 2

SuperSAGE tags of genes up-regulated by NbCD1-overexpression in <i>N. benthamiana</i> leaves				
Code	Tag sequence (5'→3')	Number of tags		
		NbCD1-overexpressing	GFP-overexpressing	SEQ ID NO.
NbCD1U1	CATGTTGATTATATGACCGGAGGGTA	7	0	SEQ ID NO:43
NbCD1U2	CATGAAGATTATGAGATTGTTTTATC	5	0	SEQ ID NO:44
NbCD1U3	CATGCAAATAAAGTAGTTGTTGCGAAA	5	0	SEQ ID NO:45
NbCD1U4	CATGGGCAGTGAACTGGGAAGAAGA	6	0	SEQ ID NO:46
NbCD1U5	CATGTGGTCTCTCAAATGTTGGAAC	10	0	SEQ ID NO:47

TABLE 2-continued

SuperSAGE tags of genes up-regulated by NbCD1-overexpression in <i>N. benthamiana</i> leaves				
Code	Tag sequence (5'→3')	Number of tags		SEQ ID NO:
		NbCD1- overexpressing	GFP- overexpressing	
NbCD1U6	CATGTACATTGAAAGATGGAGGCGGA	13	0	SEQ ID NO:48
NbCD1U7	CATGTCTATTGGTTGGCAGGCAAATA	8	0	SEQ ID NO:49
NbCD1U8	CATGAATGAAGTTTGTATCCTCTGTG	7	0	SEQ ID NO:50
NbCD1U9	CATGTTTTTACCCTATATCGATAACC	7	0	SEQ ID NO:51
NbCD1U10	CATGAAGCCATTGGAGGTTTTGTAC	6	0	SEQ ID NO:52
NbCD1U11	CATGGATCGGCAAACAAGAGATAAT	6	0	SEQ ID NO:53
NbCD1U12	CATGTATTCATCTGTAAATAGCTTGT	6	0	SEQ ID NO:54
NbCD1U13	CATGTCGTATAAAGTTGTAACGGAGT	6	0	SEQ ID NO:55
NbCD1U14	CATGTTTCATTGCCAAGATCTGGACAT	6	0	SEQ ID NO:56
NbCD1U15	CATGAACCTTGAAACTATGGATATCTG	5	0	SEQ ID NO:57
NbCD1U16	CATGACTCATATATCAAGTTTATGAG	5	0	SEQ ID NO:58
NbCD1U17	CATGATGCTTGCCAAGTGATGACATT	5	0	SEQ ID NO:59
NbCD1U18	CATGCAAAAATTGTACGTGTGGAAGG	5	0	SEQ ID NO:60
NbCD1U19	CATGTTCTTGTATATGTATCATATGT	5	0	SEQ ID NO:61
NbCD1U20	CATGGCTTCTAGATATCCATATGATG	34	1	SEQ ID NO:62
NbCD1U21	CATGTAGTGCTAAGTAATATTGAATA	21	1	SEQ ID NO:63
NbCD1U22	CATGTAATGTTTTTGTGTACAATATA	12	1	SEQ ID NO:64
NbCD1U23	CATGGTACCATCTTGTATATTTGGA	10	1	SEQ ID NO:65
NbCD1U24	CATGGTGGTGGGTACATCGTTAGAAG	9	1	SEQ ID NO:66
NbCD1U25	CATGGGCAGATCAATGGGATCCAGCC	16	2	SEQ ID NO:67
NbCD1U26	CATGCCTTTAGTACTTTGGATTTGGG	8	1	SEQ ID NO:68
NbCD1U27	CATGTTACTTGCAACGGCGATAACCA	8	1	SEQ ID NO:69
NbCD1U28	CATGTACCCTGCTGTATATTCGGGAG	38	5	SEQ ID NO:70
NbCD1U29	CATGACGTATTACAAGTACCAAAAGC	15	2	SEQ ID NO:71
NbCD1U30	CATGGATCATATGATTTTCATATTTGT	22	3	SEQ ID NO:72
NbCD1U31	CATGGGGTGTGACCAAGACGCACTT	7	1	SEQ ID NO:73
NbCD1U32	CATGAGTGCAAGCGTTCGAGGTTCT	7	1	SEQ ID NO:74
NbCD1U33	CATGTCTCATTTTTTGACTGCTGGTT	13	2	SEQ ID NO:75
NbCD1U34	CATGATTACTATTCATCAAGGGACT	6	1	SEQ ID NO:76
NbCD1U35	CATGTTATGTATGTTTCAGTTGAGAT	6	1	SEQ ID NO:77
NbCD1U36	CATGAGGAAGTTTATGTTACCGGAGA	11	2	SEQ ID NO:78
NbCD1U37	CATGTTGAGAGACCACCTATTTGTGG	25	5	SEQ ID NO:79
NbCD1U38	CATGCACTAATAATGCTACTTCAAGT	5	0	SEQ ID NO:80
NbCD1U39	CATGTGGAGTTAGATCCAAATTTCC	4	0	SEQ ID NO:81

TABLE 2-continued

SuperSAGE tags of genes up-regulated by NbCD1-overexpression in <i>N. benthamiana</i> leaves				
Code	Tag sequence (5'→3')	Number of tags		SEQ ID NO:
		NbCD1- overexpressing	GFP- overexpressing	
NbCD1U40	GATGGTACTACTCCTGGAAGATCATT	4	0	SEQ ID NO:82
NbCD1U41	CATGGATTCCAAAAAGAGCAAAAGC	4	0	SEQ ID NO:83
NbCD1U42	GATGGATATTGATGATCAGAATAATG	4	0	SEQ ID NO:84
NbCD1U43	CATGCTAATAAGGAAATTGATGCTGC	4	0	SEQ ID NO:85
NbCD1U44	CATGACTTCTTGGGACTGATGTACAT	4	0	SEQ ID NO:86

[0080]

TABLE 3

SuperSAGE tags of genes down-regulated by NbCD1-overexpression in <i>N. benthamiana</i> leaves				
Code	Tag sequence (5'→3')	Number of tags		SEQ ID NO:
		NbCD1- overexpressing	GFP- overexpressing	
NbCD1D1	CATGACTCAAATACTTGTGCACGAGG	8	33	SEQ ID NO:87
NbCD1D2	CATGGCTAATGCTGGACCTGGAACCA	2	10	SEQ ID NO:88
NbCD1D3	CATGCCCCCTTGTGTTATGGAGATCT	3	16	SEQ ID NO:89
NbCD1D4	CATGGCCCCCGTCCGCTTGCCGAC	3	16	SEQ ID NO:90
NbCD1D5	CATGAGCAGCTAAGTGAAGAACTTG	1	6	SEQ ID NO:91
NbCD1D6	CATGATCAAATAGATTTTCAGTTGGG	1	6	SEQ ID NO:92
NbCD1D7	CATGTAATTTCCCAAATCGAACTGTA	1	6	SEQ ID NO:93
NbCD1D8	CATGGACGCTTCCAGACTACACAGGA	1	6	SEQ ID NO:94
NbCD1D9	CATGTATCTGTTATCAACCTGTGTG	1	6	SEQ ID NO:95
NbCD1D10	CATGGGATTTGGCAGAAGAGGCCCG	1	8	SEQ ID NO:96
NbCD1D11	CATGGCCCCTGCGCAAGGATGACACG	2	21	SEQ ID NO:97
NbCD1D12	CATGATGAGCTTTAAGGGACTAGTCG	42	442	SEQ ID NO:98
NbCD1D13	CATGGCCGACTTGCTGCACGTCAACC	1	13	SEQ ID NO:99
NbCD1D14	CATGATAAGCTTTAAGGGATTAGTCG	1	20	SEQ ID NO:100
NbCD1D15	CATGATGCAGCTGGGTTGTGATGGCG	0	5	SEQ ID NO:101
NbCD1D16	CATGCGCCGTTTTGGCTGTAGAATGG	0	7	SEQ ID NO:102
NbCD1D17	CATGTAATGTATGCAAGTTGTTGCTA	0	7	SEQ ID NO:103
NbCD1D18	CATGGACAATTTGGTTAGGTTTCAGCT	0	5	SEQ ID NO:104
NbCD1D19	CATGACCGTGGAGCCTTGATCATTTT	0	7	SEQ ID NO:105
NbCD1D20	CATGGATAGTCCTTCACATTGGCACG	0	7	SEQ ID NO:106
NbCD1D21	CATGCCAGCTGGGAGAGCTAATCCGC	0	6	SEQ ID NO:107

TABLE 3-continued

SuperSAGE tags of genes down-regulated by NbCD1-overexpression in <i>N. benthamiana</i> leaves				
Code	Tag sequence (5'→3')	Number of tags		SEQ ID NO:
		NbCD1- overexpressing	GFP- overexpressing	
NbCD1D22	CATGGGCGTGACCGTGGGAATGGAGG	0	6	SEQ ID NO:108
NbCD1D23	CATGGGGGTATACCACACTGTCTTTG	0	6	SEQ ID NO:109
NbCD1D24	CATGGGACTTGGTGGATGCATTGCTC	0	5	SEQ ID NO:110
NbCD1D25	CATGGTGACGAAGCCAGATTGGTGGC	0	5	SEQ ID NO:111
NbCD1D26	CATGTGCTGCAGGCAGTGCTTCCGCA	0	5	SEQ ID NO:112
NbCD1D27	CATGTGAAAGAACAGACTGAGCTTGT	0	5	SEQ ID NO:113
NbCD1D28	CATGGATGGTATGTGCCTGCTCCAGT	0	5	SEQ ID NO:114
NbCD1D29	CATGCAAAACACTCTCATCCCCCTA	0	4	SEQ ID NO:115
NbCD1D30	CATGGAGGCATTCCTCCGTACGTCAT	0	4	SEQ ID NO:116
NbCD1D31	CATGTCTACGGAGGCTGTAACCTTTTT	0	4	SEQ ID NO:117
NbCD1D32	CATGGGTAGAGCCAAAGAGTGTGAAC	0	4	SEQ ID NO:118
NbCD1D33	GATGTTCTGCTACTCGACTATGAGAC	0	4	SEQ ID NO:119
NbCD1D34	CATGTGCTTCAAGACGTATCACTTGT	0	4	SEQ ID NO:120
NbCD1D35	CATGTACACTTCAAGAATCCTACTCC	0	4	SEQ ID NO:121
NbCD1D36	CATGGGTAGATGGATGGTTTGCTTAG	0	4	SEQ ID NO:122
NbCD1D37	CATGGCACAGTTAAAGGATTCTCTCT	0	4	SEQ ID NO:123
NbCD1D38	CATGGATGAAGAAGCTGCTGGGTTTT	0	4	SEQ ID NO:124
NbCD1D39	CATGACACGGTCAAGCAAAGATCTGT	0	4	SEQ ID NO:125

[0081]

TABLE 4

SuperSAGE tags of genes up-regulated by NbCD3-overexpression in <i>N. benthamiana</i> leaves				
Code	Tag sequence (5'→3')	Number of tags		SEQ ID NO:
		NbCD3- overexpressing	GFP- overexpressing	
NbCD3U1	GATGTGCAGGACTTTAGATCCTTGCA	54	1	SEQ ID NO:126
NbCD3U2	CATGTTGTATAAAGTTGTAACGAAGC	13	1	SEQ ID NO:127
NbCD3U3	CATGATTTTATGGTAACTTGATTGAT	9	1	SEQ ID NO:128
NbCD3U4	CATGTTTACCCTTTGACGGCCCAAAT	9	1	SEQ ID NO:129
NbCD3U5	CATGCATAACAATACATTTTGGTCAT	8	1	SEQ ID NO:130
NbCD3U6	CATGCCTTCTTTTCTTTGTATTATCA	8	1	SEQ ID NO:131
NbCD3U7	CATGGATTAACATCATTATTCTCTGT	8	0	SEQ ID NO:132

TABLE 4-continued

SuperSAGE tags of genes up-regulated by NbCD3-overexpression in <i>N. benthamiana</i> leaves				
Code	Tag sequence (5'→3')	Number of tags		SEQ ID NO:
		NbCD3- overexpressing	GFP- overexpressing	
NbCD3U8	CATGACACTGATAACTGCCGAGGATT	7	1	SEQ ID NO:133
NbCD3U9	CATGATAACGTTTATCTAAGAAGAGG	7	1	SEQ ID NO:134
NbCD3U10	CATGGATGGAAAACCTTAGTACCAATA	7	1	SEQ ID NO:135
NbCD3U11	CATGTGAAAGAACAGACCGAGCTTGT	7	1	SEQ ID NO:136
NbCD3U12	CATGAAGTCCATCAAAGTCCTAGGCT	7	0	SEQ ID NO:137
NbCD3U13	CATGATCATTCTTTTGTATACCGTGT	7	0	SEQ ID NO:138
NbCD3U14	CATGTTTGGAGTAATTCTCCTTGTAT	7	0	SEQ ID NO:139
NbCD3U15	CATGGCATCTCTTGACAATGTTGGGG	6	1	SEQ ID NO:140
NbCD3U16	CATGGTCCTTCAAGGGGAAGCAGGTG	6	1	SEQ ID NO:141
NbCD3U17	CATGTAAGGAGTGCTACTGAAATGGA	6	1	SEQ ID NO:142
NbCD3U18	CATGTGGTCTCTCAAATGTTGGAAC	6	1	SEQ ID NO:143
NbCD3U19	CATGTTGAACCTCTGTAATTCCGATC	6	1	SEQ ID NO:144
NbCD3U20	CATGAACACAACCTAGAGTGAAGAAGT	6	0	SEQ ID NO:145
NbCD3U21	CATGAAGTTATACGCCGACTAAAGT	6	0	SEQ ID NO:146
NbCD3U22	CATGAATGAATTTAACAGTTCAATAT	6	0	SEQ ID NO:147
NbCD3U23	CATGATAGGATAACAAAGATATATAT	6	0	SEQ ID NO:148
NbCD3U24	CATGCCTTTAGTACTTTGGATTTGGG	6	0	SEQ ID NO:149
NbCD3U25	CATGGATCGGCAAACAAGAGATAAT	6	0	SEQ ID NO:150
NbCD3U26	CATGTTAAAGTGAAACGCAACAATGG	6	0	SEQ ID NO:151
NbCD3U27	CATGTTGATTATATGACCGGAGGGTA	6	0	SEQ ID NO:152
NbCD3U28	CATGACGAAGTTCCAACACGAAGAGA	5	0	SEQ ID NO:153
NbCD3U29	CATGATAAATGGGAGACCCTTGTACT	5	0	SEQ ID NO:154
NbCD3U30	CATGATCATTTCCTGAGATTCGACG	5	0	SEQ ID NO:155
NbCD3U31	CATGCAAACCTAAAATAACCAAAC	5	0	SEQ ID NO:156
NbCD3U32	CATGGAGTTGTATCTTGACTGCTTCT	5	0	SEQ ID NO:157
NbCD3U33	CATGGGATAGCTTTTCATCTTTGGAT	5	0	SEQ ID NO:158
NbCD3U34	CATGTAACCATAACAAGTTGAACCATC	5	0	SEQ ID NO:159
NbCD3U35	CATGTGAATGACGCAAACCTTTCAAGT	5	0	SEQ ID NO:160
NbCD3U36	CATGTTATAGTATGAGATAGAGGAGT	5	0	SEQ ID NO:161
NbCD3U37	CATGTCCCCTGTTTGTATGTTCTGTT	35	7	SEQ ID NO:162
NbCD3U38	CATGGAGCTACAGGACTTCATTCTCC	18	2	SEQ ID NO:163
NbCD3U39	CATGATCTCTTTCCTGAGATTCGTCA	16	2	SEQ ID NO:164

TABLE 4-continued

SuperSAGE tags of genes up-regulated by NbCD3-overexpression in <i>N. benthamiana</i> leaves				
Code	Tag sequence (5'→3')	Number of tags		SEQ ID NO:
		NbCD3- overexpressing	GFP- overexpressing	
NbCD3U40	CATGGGCAGATCAATGGGATCCAGCC	13	3	SEQ ID NO:165
NbCD3U41	CATGGAAGAAGCAACCTTAGTGTGGT	11	2	SEQ ID NO:166
NbCD3U42	CATGTGGAATGTTTCGTATTGTGAATA	9	2	SEQ ID NO:167

[0082]

TABLE 5

SuperSAGE tags of genes down-regulated by NbCD3-overexpression in <i>N. benthamiana</i> leaves				
Code	Tag sequence (5'→3')	Number of tags		SEQ ID NO:
		NbCD3- overexpressing	GFP- overexpressing	
NbCD3D1	CATGTTGCTGAAGTTTTCACGGGTGC	1	12	SEQ ID NO:168
NbCD3D2	CATGCAAGGCCAGTCGGAGAAGAAGG	1	8	SEQ ID NO:169
NbCD3D3	CATGAGGGATGAGCCAGGAGCACGGC	1	7	SEQ ID NO:170
NbCD3D4	CATGTTGCAACTTCTAGTCAATGACT	1	7	SEQ ID NO:171
NbCD3D5	CATGAGCGGAAGCTAACCTGAATCCA	1	6	SEQ ID NO:172
NbCD3D6	CATGCCTGTGAGCCTGCTGTTGGTAA	1	6	SEQ ID NO:173
NbCD3D7	GATGGCACCAGCGTGGAATGTCGCAT	1	6	SEQ ID NO:174
NbCD3D8	CATGGGGATATAGCAAGATCGTGAAT	1	6	SEQ ID NO:175
NbCD3D9	CATGTTATCTTTCCAATAGAGGCGAG	0	6	SEQ ID NO:176
NbCD3D10	CATGAGACTCTAAACAATTTTCGCTTG	0	5	SEQ ID NO:177
NbCD3D11	CATGCAGCAAAGACCAAGAACAGCCC	0	5	SEQ ID NO:178
NbCD3D12	CATGCCGAAGCAAATCCACGAAATCA	0	5	SEQ ID NO:179
NbCD3D13	CATGCTTACAAAGGAATCCAGCTAC	0	5	SEQ ID NO:180
NbCD3D14	CATGGGGTCTCCCGCTGGTAAGGTAT	0	5	SEQ ID NO:181
NbCD3D15	CATGACGCGCTTAACCTACACTCTTG	1	5	SEQ ID NO:182
NbCD3D16	CATGAGGAGGCTAGAAGGAAGAATGT	1	5	SEQ ID NO:183
NbCD3D17	CATGAGGGATGAACCAGGAGCCAGAC	1	5	SEQ ID NO:184
NbCD3D18	CATGATTTGTAAC TATTGGGGATTCT	1	5	SEQ ID NO:185
NbCD3D19	CATGGATATATGGCAATTCGTTTTGT	1	5	SEQ ID NO:186
NbCD3D20	CATGGGTGCTGAGATGGTTTAATGGT	1	5	SEQ ID NO:187
NbCD3D21	CATGTAATTTGGCGGGGAGTAATGTA	1	5	SEQ ID NO:188
NbCD3D22	CATGAATAAATGCTACTCTAATAGCT	0	4	SEQ ID NO:189

TABLE 5-continued

SuperSAGE tags of genes down-regulated by NbCD3-overexpression in <i>N. benthamiana</i> leaves				
Code	Tag sequence (5'→3')	Number of tags		SEQ ID NO:
		NbCD3- overexpressing	GFP- overexpressing	
NbCD3D23	CATGACGGAAAAGCCAATTATCAAGT	0	4	SEQ ID NO:190
NbCD3D24	CATGATTGGGCAATTTGGTGTGGTT	0	4	SEQ ID NO:191
NbCD3D25	CATGATTTTCAAGGACGGAGAGAAGA	0	4	SEQ ID NO:192
NbCD3D26	CATGCCACCGGGTCCACAACGTGCT	0	4	SEQ ID NO:193
NbCD3D27	CATGCTGCCCAACTTTGTGTATTGGC	0	4	SEQ ID NO:194
NbCD3D28	CATGGGTTTCAGCTTGTGTTGATTAAG	0	4	SEQ ID NO:195
NbCD3D29	CATGTATAAATTGTGTAATGTTGTGT	0	4	SEQ ID NO:196

[0083] RNAs were extracted from leaves overexpressing GFP, NbCD1, and NbCD3 for hybridization to the arrays. Gene overexpression and hybridization were independently triplicated. In order to observe reproducibility among hybridization triplicates, the signal values of hybridization of the tags to the oligos were plotted between two arrays. Satisfactory correlations were observed in most signal values among the triplicates ($R^2=0.9584$ to 0.9863). Thus, the array involving the use of the SuperSAGE tags yielded the results of hybridization with very high reproducibility (FIG. 4).

[0084] The average signal values of genes among the triplicates are shown in Tables 6 to 9. Based on the differences in average signal values among the GFP-overexpressing leaves and the NbCD1- or NbCD3-overexpressing leaves, gene expression patterns were analyzed using Cluster and TreeView software and indicated by color tiles (FIG. 5). Based on the results of SuperSAGE-Array analysis, genes were classified as follows: (A) genes induced upon NbCD1 overexpression; (B) genes repressed upon NbCD1 overexpression; (C) genes induced upon NbCD3 overexpression; and (D) genes repressed upon NbCD3 overexpression. Compared with the control GFP-overexpressing leaves, stronger signals are indicated by red tiles and weaker signals are indicated by green tiles. SuperSAGE tags starting from NbCD1U and NbCD3U generally exhibited stronger hybridization signals than the control upon overexpression of the genes. Many tags with lowered expression levels (SuperSAGE tags starting from NbCD1D and NbCD3D) emitted the signals weaker than those of the control. The genes exhibiting statistically significant differences in expression levels were selected. As a result, 115 genes (i.e., 74% of the genes subjected to array analysis) were found to exhibit the same expression patterns between SuperSAGE and SuperSAGE-Array analysis (Table 10).

TABLE 6

Average of normalized hybridization signal values (Log_2) of down-regulated genes in NbCD1-overexpressing <i>N. benthamiana</i> leaves.				
Code	GFP-over- expressing	s.e.(±)*	NbCD1-over- expressing	s.e.(±)*
NbCD1D1	14.445	0.155	14.197	0.045
NbCD1D2	14.684	0.072	14.325	0.010
NbCD1D3	14.025	0.111	13.549	0.050
NbCD1D4	13.212	0.197	12.654	0.214
NbCD1D5	10.779	0.164	9.727	0.131
NbCD1D6	9.406	0.329	8.888	0.113
NbCD1D7	8.019	0.465	7.168	0.294
NbCD1D8	12.916	0.042	12.284	0.085
NbCD1D9	13.187	0.078	12.643	0.029
NbCD1D10	11.379	0.041	10.269	0.229
NbCD1D11	13.119	0.119	11.859	0.208
NbCD1D12	14.943	0.053	6.893	2.392
NbCD1D13	13.945	0.103	13.387	0.094
NbCD1D14	12.539	0.135	4.981	0.138
NbCD1D15	13.637	0.042	12.390	0.040
NbCD1D16	12.631	0.084	11.639	0.138
NbCD1D17	12.881	0.070	11.731	0.133
NbCD1D18	14.231	0.086	12.898	0.059
NbCD1D19	13.159	0.089	12.696	0.050
NbCD1D20	13.157	0.186	8.078	0.105
NbCD1D21	9.424	0.212	10.261	0.106
NbCD1D22	12.751	0.075	11.439	0.133
NbCD1D23	13.481	0.099	12.874	0.088
NbCD1D24	12.077	0.077	11.983	0.099
NbCD1D25	11.270	0.169	10.919	0.244
NbCD1D26	12.649	0.131	11.824	0.113
NbCD1D27	9.903	0.221	10.040	0.350
NbCD1D28	11.677	0.046	11.190	0.100
NbCD1D29	9.702	0.177	9.474	0.176
NbCD1D30	12.901	0.186	12.469	0.074
NbCD1D31	12.757	0.061	12.581	0.068
NbCD1D32	10.435	0.067	10.242	0.075
NbCD1D33	13.756	0.188	13.072	0.067
NbCD1D34	12.270	0.407	11.441	0.204
NbCD1D35	12.549	0.093	12.230	0.031
NbCD1D36	13.547	0.084	12.782	0.141
NbCD1D37	12.171	0.224	11.228	0.098
NbCD1D38	10.725	0.078	10.489	0.193
NbCD1D39	10.679	0.220	10.212	0.219

*s.e.: standard error of hybridization signal values among triplicates

[0085]

TABLE 7

Average of normalized hybridization signal values (Log ₂) of up-regulated genes in NbCD1-overexpressing <i>N. benthamiana</i> leaves.				
Code	GFP-over-expressing	s.e.(±) *	NbCD1-over-expressing	s.e.(±) *
NbCD1U1	11.720	0.211	12.782	0.178
NbCD1U2	10.172	0.250	10.657	0.286
NbCD1U3	9.832	0.264	10.618	0.117
NbCD1U4	7.317	0.424	10.930	0.266
NbCD1U5	12.605	0.239	14.723	0.121
NbCD1U6	5.868	0.134	8.038	0.320
NbCD1U7	12.541	0.471	13.220	0.054
NbCD1U8	10.133	0.214	12.282	0.079
NbCD1U9	13.298	0.031	12.967	0.172
NbCD1U10	11.078	0.359	12.868	0.168
NbCD1U11	9.350	0.321	12.087	0.056
NbCD1U12	11.416	0.164	13.622	0.011
NbCD1U13	10.792	0.632	12.961	0.082
NbCD1U14	10.915	0.657	13.801	0.089
NbCD1U15	11.116	0.196	10.891	0.127
NbCD1U16	9.173	0.631	10.456	0.274
NbCD1U17	13.097	0.043	13.207	0.046
NbCD1U18	9.439	0.308	9.102	0.332
NbCD1U19	12.771	0.139	12.682	0.136
NbCD1U20	6.713	1.561	14.696	0.033
NbCD1U21	12.697	0.219	12.343	0.143
NbCD1U22	11.009	0.400	10.514	0.199
NbCD1U23	13.180	0.106	14.073	0.058
NbCD1U24	8.835	0.073	8.540	0.193
NbCD1U25	11.836	0.073	13.783	0.066
NbCD1U26	13.396	0.192	14.108	0.033
NbCD1U27	14.279	0.046	14.041	0.031
NbCD1U28	14.016	0.138	14.535	0.057
NbCD1U29	12.828	0.118	12.927	0.114
NbCD1U30	12.245	0.486	12.802	0.103
NbCD1U31	12.153	0.102	11.250	0.108
NbCD1U32	12.322	0.320	12.377	0.075
NbCD1U33	13.703	0.126	13.134	0.026
NbCD1U34	12.472	0.081	11.700	0.208
NbCD1U35	13.573	0.103	13.936	0.088
NbCD1U36	11.067	0.377	13.365	0.228
NbCD1U37	13.720	0.193	14.212	0.079
NbCD1U38	12.383	0.232	13.318	0.166
NbCD1U39	11.440	0.257	13.267	0.162
NbCD1U40	12.229	0.295	13.174	0.141
NbCD1U41	5.027	0.102	6.302	0.459
NbCD1U42	11.551	0.165	12.399	0.034
NbCD1U43	8.999	0.227	11.510	0.456
NbCD1U44	11.776	0.403	13.154	0.238

* s.e.: standard error of hybridization signal values among triplicates

[0086]

TABLE 8

Average of normalized hybridization signal values (Log ₂) of up-regulated genes in NbCD3-overexpressing <i>N. benthamiana</i> leaves.				
Code	GFP-over-expressing	s.e.(±) *	NbCD3-over-expressing	s.e.(±) *
NbCD3U1	9.338	0.334	14.490	0.069
NbCD3U2	12.419	0.353	12.565	0.082
NbCD3U3	8.884	0.637	11.439	0.227
NbCD3U4	13.347	0.301	14.981	0.159
NbCD3U5	12.751	0.146	13.141	0.157
NbCD3U6	14.025	0.107	13.136	0.211
NbCD3U7	13.749	0.058	13.902	0.078
NbCD3U8	12.551	0.109	12.843	0.065
NbCD3U9	9.076	0.312	8.941	0.311
NbCD3U10	12.798	0.166	13.198	0.103

TABLE 8-continued

Average of normalized hybridization signal values (Log ₂) of up-regulated genes in NbCD3-overexpressing <i>N. benthamiana</i> leaves.				
Code	GFP-over-expressing	s.e.(±) *	NbCD3-over-expressing	s.e.(±) *
NbCD3U11	10.615	0.165	11.848	0.162
NbCD3U12	10.763	0.071	12.675	0.172
NbCD3U13	12.230	0.161	14.141	0.099
NbCD3U14	8.062	0.241	11.898	0.058
NbCD3U15	12.525	0.114	13.191	0.123
NbCD3U16	12.984	0.105	12.718	0.097
NbCD3U17	12.185	0.325	12.912	0.087
NbCD3U18	12.562	0.276	14.760	0.137
NbCD3U19	13.088	0.183	12.979	0.043
NbCD3U20	9.771	0.720	11.138	0.494
NbCD3U21	14.115	0.124	14.200	0.013
NbCD3U22	9.706	0.178	9.153	0.510
NbCD3U23	5.852	0.775	7.893	0.035
NbCD3U24	13.342	0.240	14.523	0.108
NbCD3U25	9.324	0.303	11.114	0.113
NbCD3U26	9.034	0.373	8.762	0.296
NbCD3U27	11.796	0.431	12.485	0.048
NbCD3U28	9.631	0.044	10.643	0.138
NbCD3U29	9.212	0.278	8.617	0.538
NbCD3U30	12.251	0.863	12.282	0.259
NbCD3U31	9.597	0.476	10.945	0.124
NbCD3U32	13.710	0.091	13.316	0.092
NbCD3U33	14.235	0.022	13.831	0.056
NbCD3U34	11.909	0.037	12.893	0.086
NbCD3U35	9.288	0.371	10.428	0.097
NbCD3U36	8.824	0.350	5.991	0.770
NbCD3U37	14.771	0.071	14.944	0.118
NbCD3U38	14.398	0.048	14.898	0.061
NbCD3U39	13.079	0.142	13.771	0.155
NbCD3U40	11.950	0.183	13.631	0.219
NbCD3U41	12.915	0.061	12.482	0.046
NbCD3U42	13.335	0.075	13.618	0.077

* s.e.: standard error of hybridization signal values among triplicates

[0087]

TABLE 9

Average of normalized hybridization signal values (Log ₂) of down-regulated genes in NbCD3-overexpressing <i>N. benthamiana</i> leaves				
Code	GFP-over-expressing	s.e.(±) *	NbCD3-over-expressing	s.e.(±) *
NbCD3D1	13.757	0.041	13.526	0.078
NbCD3D2	9.681	0.227	9.267	0.354
NbCD3D3	9.764	0.136	9.593	0.108
NbCD3D4	11.050	0.049	7.876	0.974
NbCD3D5	12.010	0.081	11.401	0.146
NbCD3D6	13.409	0.169	13.066	0.103
NbCD3D7	12.111	0.118	11.423	0.111
NbCD3D8	11.656	0.175	11.003	0.198
NbCD3D9	13.842	0.041	13.691	0.051
NbCD3D10	12.116	0.174	10.371	0.490
NbCD3D11	10.711	0.212	9.166	0.426
NbCD3D12	12.084	0.180	11.681	0.123
NbCD3D13	10.922	0.174	10.314	0.223
NbCD3D14	12.254	0.209	12.172	0.076
NbCD3D15	12.308	0.040	9.585	0.642
NbCD3D16	8.563	0.211	8.537	0.189
NbCD3D17	9.126	0.069	8.974	0.121
NbCD3D18	12.063	0.023	10.389	0.333
NbCD3D19	12.362	0.440	10.202	0.520
NbCD3D20	11.170	0.148	11.055	0.152
NbCD3D21	9.823	0.328	7.940	0.545
NbCD3D22	10.923	0.265	10.084	0.371
NbCD3D23	10.983	0.018	10.623	0.074

TABLE 9-continued

Average of normalized hybridization signal values (Log_2) of down-regulated genes in NbCD3-overexpressing <i>N. benthamiana</i> leaves				
Code	GFP-over-expressing	s.e.(\pm) *	NbCD3-over-expressing	s.e.(\pm) *
NbCD3D24	13.841	0.098	13.156	0.120
NbCD3D25	10.456	0.218	7.949	0.719
NbCD3D26	9.331	0.305	11.180	0.285
NbCD3D27	12.287	0.079	11.673	0.023
NbCD3D28	14.425	0.070	14.136	0.061
NbCD3D29	11.301	0.162	10.357	0.118

* s.e.: standard error of hybridization signal values among triplicates

[0088]

TABLE 10

	NbCD1 overexpression		NbCD3 overexpression		Total
	Increased	Decreased	Increased	Decreased	
SuperSAGE	46	39	42	29	156
SuperSAGE-Array	32	33	28	22	115

4. Discussion

[0089] The SuperSAGE-Array system developed by the present inventors directly employed 26-bp tags without optimizing the T_m value. This analysis, however, yielded highly reproducible results. This indicates that a SuperSAGE-Array can be applied to: 1) evaluation of SuperSAGE results; and 2) gene expression analysis of multiple samples detected by SuperSAGE.

[0090] A SuperSAGE-Array does not require the designing of oligo probes. Thus, an array can be prepared 1 to 1.5 months after the SuperSAGE analysis, and extensive gene expression analysis can be carried out. Specifically, the SuperSAGE-Array is an excellent tool that enables precise analysis of genes identified by SuperSAGE.

[0091] Some tags did not yield the results that were consistent between SuperSAGE and SuperSAGE-Array for the following reasons. That is, such tags were located in the vicinity of a poly(A) sequence, hybridization was not satisfactorily carried out, or 26-bp probes hybridized to RNAs of the homologous genes. Such problems may be overcome by performing SuperSAGE analysis using a different anchoring enzyme.

[0092] In this example, specific tags were extracted using a restriction enzyme, N1aIII, which recognizes a 4-nucleotide sequence. Instead of N1aIII, a restriction enzyme, such as Sau3AI, may be used to extract the same tags. By using a SuperSAGE-Array for all the tags of interest in two libraries, hybridization may be satisfactorily carried out with at least 1 oligonucleotide of the gene identified by SuperSAGE.

Example 2

[0093] In Example 1, model rice plants were subjected to expression analysis using an array with 41 SuperSAGE tags immobilized thereon, and non-model *Nicotiana benthami-*

ana plants were subjected to expression analysis using an array with 154 SuperSAGE tags immobilized thereon. In both cases, the results of analysis were very consistent with the results of expression analysis via SuperSAGE. In this example, an array with 1,000 SuperSAGE tags immobilized thereon was prepared for rice, and the results of expression analysis via SuperSAGE-Array were compared with those via SuperSAGE.

[0094] In accordance with the procedure of Example 1, mRNAs were extracted from rice leaves (variety: Yashiro-mochi) and cultured cells of rice (variety: Kakehashi) to prepare SuperSAGE libraries. From these libraries, 1,000 SuperSAGE tag sequences were selected. Among them, 78 tags represented equally expressed genes, 438 tags were more prevalent in leaves and 484 tags were more abundant in suspension-cultured cells. Based on the determined tag sequences, arrays were prepared using the 12-well array system (NimbleScreen 12: NimbleGen Co.). For the tag sequences, mismatch-containing oligonucleotides, wherein nucleotides 7 and 13 were modified (heterogeneous sequences), were synthesized and used for a hybridization specificity test.

[0095] Total RNAs were extracted from rice leaves and cultured cells, Cy3-labeled cRNAs were synthesized, and the resultants were used as hybridization probes. The probes were allowed to hybridize to the prepared array, the signals were read with a scanner, and the data was standardized by the Robust Multi-chip Analysis (RMA) method. Hybridization was independently triplicated. In order to observe reproducibility among hybridization triplicates, the hybridization signal values of the tags were plotted between two arrays. As a result, satisfactory correlations were observed in most signal values among the triplicates ($R^2=0.973$ to 0.992), as with the case of Example 1. Array preparation and hybridization were entrusted to Gene Frontier.

[0096] FIG. 6 shows the expression patterns of genes in rice leaves and in cultured cells observed via SuperSAGE-Array. Table 11 shows a comparison of the results of expression analysis via SuperSAGE-Array and via SuperSAGE.

TABLE 11

	Gene expression in rice leaves and in cultured cells - Comparison of results of gene expression analysis via SuperSAGE-Array and via SuperSAGE -			
	Genes expressed at high level in leaves		Genes expressed at high level in cultured cells	
	With significant difference (p < 0.05)	Whole	With significant difference (p < 0.05)	Whole
SuperSAGE	155	438	167	484
SuperSAGE-Array	136	352	149	421
Consistency (%)*	(87.7)	(80.4)	(89.2)	(87.0)

*Level of consistency between the results of expression analysis via SuperSAGE-Array and via SuperSAGE

[0097] As a result, the gene group with a detected expression level that was higher in leaves than in cultured cells was found to exhibit a consistency of 80.4% (87.7% for those exhibiting statistic significance) between SuperSAGE and

SuperSAGE-Array. The gene group with a detected expression level that was higher in cultured cells than in leaves was found to exhibit a consistency of 87.0% (89.2% for those exhibiting statistic significance) therebetween.

Example 3

[0098] Among the tags that were found to be expressed at high levels in all of the NbCD1- and NbCD3-overexpressing *Nicotiana benthamiana* leaves by the SuperSAGE-Array-based expression analysis in Example 1, 5 tags showing no sequence matches to known cDNA or EST were selected (NbCD3U14, 20, 25, 32, and 40), and identification of the genes corresponding thereto was attempted.

[0099] Full-length sequences of the tags were determined by the 3'-RACE and 5'-RACE methods in the following manner. As a template, RNA was isolated from NbCD3-overexpressing *Nicotiana benthamiana* leaves, and the RNA was flanked by adaptor sequences to synthesize cDNA. Based on the SuperSAGE tag sequences, a gene specific PCR primer and a primer complementary to the adaptor sequence were used to amplify a partial cDNA fragment from template cDNA. A primer was prepared based on the 5'-sequence of the resulting fragment, and the resulting primer and the adaptor primer (i.e., a primer complementary to the adaptor sequence) were used to amplify the upstream region. FIG. 7 schematically shows the amplification of full-length sequences by the 3'-RACE and 5'-RACE methods (upper portion) and the results of electrophoresis of the PCR products thereof (lower portion).

[0100] The results of the 3'-RACE method were combined with those of the 5'-RACE method to determine the full-length sequences of the genes corresponding to the 5 tags. It was confirmed that the SuperSAGE tag sequences were mapped at expected positions in all 5 genes. Functions of

these genes can be deduced from the putative ORFs and the results of BLAST search. The results of SuperSAGE-Array-based analysis demonstrated that full-length sequences of unknown genes could be easily determined and that functions thereof could be easily deduced.

[0101] Thus, the SuperSAGE-Array was found to be unique and effective, providing the advantages of highly quantitative SAGE in combination with those of high-throughput microarray systems.

INDUSTRIAL APPLICABILITY

[0102] According to the present invention, oligoarrays of any tissues or genes expressed under desired conditions can be easily prepared. For example, SuperSAGE-Arrays of various cancer tissues are useful for clinical testing, and SuperSAGE-Arrays can be applied to all eukaryotic organisms. Also, SuperSAGE-Arrays upon which host organism and pathogen genes are spotted can be applied to host-pathogen interaction analysis. Thus, the present invention can be applied to gene expression analysis in any field, including applications ranging from the basic to the clinical.

[0103] All publications, patents, and patent applications cited herein are incorporated herein by reference in their entirety.

SEQUENCE LISTING FREE TEXT

[0104] SEQ ID NO: 8; oligo-dT primer

[0105] SEQ ID NO: 197; linker A sequence

[0106] SEQ ID NO: 198; linker A sequence

[0107] SEQ ID NO: 199; linker B sequence

[0108] SEQ ID NO: 200; linker B sequence

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<400> SEQUENCE: 131

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<210> SEQ ID NO 132
<211> LENGTH: 26
<212> TYPE: DNA
<213> ORGANISM: Nicotiana benthamiana

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<400> SEQUENCE: 132

catggattaa catcattatt ctctgt 26

<210> SEQ ID NO 133

<211> LENGTH: 26

<212> TYPE: DNA

<213> ORGANISM: Nicotiana benthamiana

<400> SEQUENCE: 133

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<210> SEQ ID NO 134

<211> LENGTH: 26

<212> TYPE: DNA

<213> ORGANISM: Nicotiana benthamiana

<400> SEQUENCE: 134

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<210> SEQ ID NO 135

<211> LENGTH: 26

<212> TYPE: DNA

<213> ORGANISM: Nicotiana benthamiana

<400> SEQUENCE: 135

catggatgga aaacttagta ccaata 26

<210> SEQ ID NO 136

<211> LENGTH: 26

<212> TYPE: DNA

<213> ORGANISM: Nicotiana benthamiana

<400> SEQUENCE: 136

catgtgaaag aacagaccga gcttgt 26

<210> SEQ ID NO 137

<211> LENGTH: 26

<212> TYPE: DNA

<213> ORGANISM: Nicotiana benthamiana

<400> SEQUENCE: 137

catgaagtcc atcaaagtcc taggct 26

<210> SEQ ID NO 138

<211> LENGTH: 26

<212> TYPE: DNA

<213> ORGANISM: Nicotiana benthamiana

<400> SEQUENCE: 138

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<210> SEQ ID NO 139

<211> LENGTH: 26

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<213> ORGANISM: Nicotiana benthamiana

<400> SEQUENCE: 139

catgtttggg gtaattctcc ttgtat 26

<210> SEQ ID NO 140

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<400> SEQUENCE: 140
catggcatct cttgacaatg ttgggg 26

<210> SEQ ID NO 141
<211> LENGTH: 26
<212> TYPE: DNA
<213> ORGANISM: Nicotiana benthamiana

<400> SEQUENCE: 141
catggtcctt caaggggaag caggtg 26

<210> SEQ ID NO 142
<211> LENGTH: 26
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<400> SEQUENCE: 142
catgtaagga gtgctactga aatgga 26

<210> SEQ ID NO 143
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<400> SEQUENCE: 143
catgtggtct ctcaaagtgt ggaact 26

<210> SEQ ID NO 144
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<400> SEQUENCE: 144
catggtgaac ctctgtaatt ccgatc 26

<210> SEQ ID NO 145
<211> LENGTH: 26
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<400> SEQUENCE: 145
catgaacaca actagagtga agaagt 26

<210> SEQ ID NO 146
<211> LENGTH: 26
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<400> SEQUENCE: 146
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<210> SEQ ID NO 147
<211> LENGTH: 26
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<400> SEQUENCE: 147

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catgaatgaa tttacagtt caatat 26

<210> SEQ ID NO 148
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<400> SEQUENCE: 148

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<210> SEQ ID NO 149
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<400> SEQUENCE: 149

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<210> SEQ ID NO 150
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<400> SEQUENCE: 150

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<210> SEQ ID NO 151
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<400> SEQUENCE: 151

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<210> SEQ ID NO 152
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<400> SEQUENCE: 152

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<210> SEQ ID NO 153
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<400> SEQUENCE: 153

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<210> SEQ ID NO 154
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<400> SEQUENCE: 154

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<210> SEQ ID NO 155
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<212> TYPE: DNA

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<213> ORGANISM: Nicotiana benthamiana

<400> SEQUENCE: 155

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<210> SEQ ID NO 156

<211> LENGTH: 26

<212> TYPE: DNA

<213> ORGANISM: Nicotiana benthamiana

<400> SEQUENCE: 156

catgcaaac ctaaataac caaact 26

<210> SEQ ID NO 157

<211> LENGTH: 26

<212> TYPE: DNA

<213> ORGANISM: Nicotiana benthamiana

<400> SEQUENCE: 157

catggagttg tatcttgact gcttct 26

<210> SEQ ID NO 158

<211> LENGTH: 26

<212> TYPE: DNA

<213> ORGANISM: Nicotiana benthamiana

<400> SEQUENCE: 158

catgggatag cttttcatct ttggat 26

<210> SEQ ID NO 159

<211> LENGTH: 26

<212> TYPE: DNA

<213> ORGANISM: Nicotiana benthamiana

<400> SEQUENCE: 159

catgtaacca tacaagttga accatc 26

<210> SEQ ID NO 160

<211> LENGTH: 26

<212> TYPE: DNA

<213> ORGANISM: Nicotiana benthamiana

<400> SEQUENCE: 160

catgtgaatg acgcaactt tcaagt 26

<210> SEQ ID NO 161

<211> LENGTH: 26

<212> TYPE: DNA

<213> ORGANISM: Nicotiana benthamiana

<400> SEQUENCE: 161

catggtatag tatgatag aggagt 26

<210> SEQ ID NO 162

<211> LENGTH: 26

<212> TYPE: DNA

<213> ORGANISM: Nicotiana benthamiana

<400> SEQUENCE: 162

catgtcccct gtttgatgt tctggt 26

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<210> SEQ ID NO 163
<211> LENGTH: 26
<212> TYPE: DNA
<213> ORGANISM: Nicotiana benthamiana

<400> SEQUENCE: 163

catggagcta caggacttca ttctcc 26

<210> SEQ ID NO 164
<211> LENGTH: 26
<212> TYPE: DNA
<213> ORGANISM: Nicotiana benthamiana

<400> SEQUENCE: 164

catgatctct ttcctgagat tcgtca 26

<210> SEQ ID NO 165
<211> LENGTH: 26
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<400> SEQUENCE: 165

catgggcaga tcaatgggat ccagcc 26

<210> SEQ ID NO 166
<211> LENGTH: 26
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<400> SEQUENCE: 166

catggaagaa gcaaccttag tgtggt 26

<210> SEQ ID NO 167
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<400> SEQUENCE: 167

catgtggaat gttcgtattg tgaata 26

<210> SEQ ID NO 168
<211> LENGTH: 26
<212> TYPE: DNA
<213> ORGANISM: Nicotiana benthamiana

<400> SEQUENCE: 168

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<210> SEQ ID NO 169
<211> LENGTH: 26
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<400> SEQUENCE: 169

catgcaaggc cagtcggaga agaagg 26

<210> SEQ ID NO 170
<211> LENGTH: 26
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<400> SEQUENCE: 170

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<210> SEQ ID NO 171

<211> LENGTH: 26

<212> TYPE: DNA

<213> ORGANISM: Nicotiana benthamiana

<400> SEQUENCE: 171

catggttcaa cttctagtca atgact 26

<210> SEQ ID NO 172

<211> LENGTH: 26

<212> TYPE: DNA

<213> ORGANISM: Nicotiana benthamiana

<400> SEQUENCE: 172

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<211> LENGTH: 26

<212> TYPE: DNA

<213> ORGANISM: Nicotiana benthamiana

<400> SEQUENCE: 173

catgcctgtg agcctgctgt tggtaa 26

<210> SEQ ID NO 174

<211> LENGTH: 26

<212> TYPE: DNA

<213> ORGANISM: Nicotiana benthamiana

<400> SEQUENCE: 174

catggcacca gcgtggaatg tcgcat 26

<210> SEQ ID NO 175

<211> LENGTH: 26

<212> TYPE: DNA

<213> ORGANISM: Nicotiana benthamiana

<400> SEQUENCE: 175

catggggata tagcaagatc gtgaat 26

<210> SEQ ID NO 176

<211> LENGTH: 26

<212> TYPE: DNA

<213> ORGANISM: Nicotiana benthamiana

<400> SEQUENCE: 176

catgttatct ttccaataga ggcgag 26

<210> SEQ ID NO 177

<211> LENGTH: 26

<212> TYPE: DNA

<213> ORGANISM: Nicotiana benthamiana

<400> SEQUENCE: 177

catgagactc taaacaattt cgcttg 26

<210> SEQ ID NO 178

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<211> LENGTH: 26
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<213> ORGANISM: Nicotiana benthamiana

<400> SEQUENCE: 178

catgcagcaa agaccaagaa cagccc 26

<210> SEQ ID NO 179
<211> LENGTH: 26
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<400> SEQUENCE: 179

catgccgaag caaatccacg aaatca 26

<210> SEQ ID NO 180
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<213> ORGANISM: Nicotiana benthamiana

<400> SEQUENCE: 180

catgcttaca aagggaatcc agctac 26

<210> SEQ ID NO 181
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<400> SEQUENCE: 181

catgggggtct cccgctggta aggtat 26

<210> SEQ ID NO 182
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<400> SEQUENCE: 182

catgacgcgc ttaacctaca ctcttg 26

<210> SEQ ID NO 183
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<400> SEQUENCE: 183

catgaggagg ctagaaggaa gaatgt 26

<210> SEQ ID NO 184
<211> LENGTH: 26
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<400> SEQUENCE: 184

catgagggat gaaccaggag ccagac 26

<210> SEQ ID NO 185
<211> LENGTH: 26
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<400> SEQUENCE: 185

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catgatttgt aactattggg gattct 26

<210> SEQ ID NO 186
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<400> SEQUENCE: 186

catggatata tggcaattgc gtttgt 26

<210> SEQ ID NO 187
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<400> SEQUENCE: 187

catgggtgct gagatggttt aatggt 26

<210> SEQ ID NO 188
<211> LENGTH: 26
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<213> ORGANISM: Nicotiana benthamiana

<400> SEQUENCE: 188

catgtaattt ggcggggagt aatgta 26

<210> SEQ ID NO 189
<211> LENGTH: 26
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<400> SEQUENCE: 189

catgaataaa tgctactcta atagct 26

<210> SEQ ID NO 190
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<400> SEQUENCE: 190

catgacggaa aagccaatta tcaagt 26

<210> SEQ ID NO 191
<211> LENGTH: 26
<212> TYPE: DNA
<213> ORGANISM: Nicotiana benthamiana

<400> SEQUENCE: 191

catgattggg caatttgggtg ttggtt 26

<210> SEQ ID NO 192
<211> LENGTH: 26
<212> TYPE: DNA
<213> ORGANISM: Nicotiana benthamiana

<400> SEQUENCE: 192

catgattttc aaggacggag agaaga 26

<210> SEQ ID NO 193
<211> LENGTH: 26
<212> TYPE: DNA

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<213> ORGANISM: Nicotiana benthamiana

<400> SEQUENCE: 193

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<210> SEQ ID NO 194

<211> LENGTH: 26

<212> TYPE: DNA

<213> ORGANISM: Nicotiana benthamiana

<400> SEQUENCE: 194

catgctgccc aactttgtgt attggc 26

<210> SEQ ID NO 195

<211> LENGTH: 26

<212> TYPE: DNA

<213> ORGANISM: Nicotiana benthamiana

<400> SEQUENCE: 195

catgggtttc agcttgtttg attaag 26

<210> SEQ ID NO 196

<211> LENGTH: 26

<212> TYPE: DNA

<213> ORGANISM: Nicotiana benthamiana

<400> SEQUENCE: 196

catgtataaa ttgtgtaatg ttgtgt 26

<210> SEQ ID NO 197

<211> LENGTH: 46

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic Linker A

<400> SEQUENCE: 197

tttggatttg ctggtgcagt acaactaggc ttaatacagc agcatg 46

<210> SEQ ID NO 198

<211> LENGTH: 42

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic Linker A

<400> SEQUENCE: 198

ctgctgtatt aagcctagtt gtactgcacc agcaaatcca aa 42

<210> SEQ ID NO 199

<211> LENGTH: 44

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic Linker B

<400> SEQUENCE: 199

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<210> SEQ ID NO 200
 <211> LENGTH: 40
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
 Linker B

 <400> SEQUENCE: 200

 ctgctgcgta catcgtaga agcttgaatt cgagcagaaa 40

<210> SEQ ID NO 201
 <211> LENGTH: 50
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
 oligonucleotide
 <220> FEATURE:
 <221> NAME/KEY: misc_feature
 <222> LOCATION: (1)..(40)
 <223> OTHER INFORMATION: n is a, c, g, t, or any arbitrary nucleotide
 and up to 10 of them may be present or absent

 <400> SEQUENCE: 201

 nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn nnnnnnnnnn cagcagcatg 50

<210> SEQ ID NO 202
 <211> LENGTH: 15
 <212> TYPE: RNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
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 <400> SEQUENCE: 202

 aaaaaaaaaa aaaaaa 15

<210> SEQ ID NO 203
 <211> LENGTH: 25
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
 oligonucleotide

 <400> SEQUENCE: 203

 tttttttttt tttttttttg acgac 25

<210> SEQ ID NO 204
 <211> LENGTH: 21
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
 oligonucleotide

 <400> SEQUENCE: 204

 aaaaaaaaaa aaaaagtcgt c 21

<210> SEQ ID NO 205
 <211> LENGTH: 21
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic

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oligonucleotide

<400> SEQUENCE: 205

tttttttttt tttttgacga c 21

<210> SEQ ID NO 206
<211> LENGTH: 12
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligonucleotide

<400> SEQUENCE: 206

aaaaaactgc tg 12

<210> SEQ ID NO 207
<211> LENGTH: 12
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligonucleotide

<400> SEQUENCE: 207

ttttttgacg ac 12

<210> SEQ ID NO 208
<211> LENGTH: 22
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligonucleotide

<400> SEQUENCE: 208

cagcagcatg aaaaaactgc tg 22

<210> SEQ ID NO 209
<211> LENGTH: 16
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligonucleotide

<400> SEQUENCE: 209

gtactttttt gacgac 16

<210> SEQ ID NO 210
<211> LENGTH: 12
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic
oligonucleotide

<400> SEQUENCE: 210

ttttttgacg ac 12

<210> SEQ ID NO 211
<211> LENGTH: 10
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:

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<223> OTHER INFORMATION: Description of Artificial Sequence: Synthetic oligonucleotide

<400> SEQUENCE: 211

cagcagcatg

10

1. A method of gene expression analysis, wherein tags each comprising an oligonucleotide of more than 25 bp for identifying expressed genes, wherein the 3'-end of the tag is defined by a cleavage site of a type III restriction enzyme and the 5'-end thereof is defined by a cleavage site of another restriction enzyme located closest to the 3'-end of the cDNA of the genes, are immobilized on a solid support, gene-containing samples are hybridized to the solid support, and the signals emitted from the genes hybridized to the tags are detected to analyze the gene expression profiles in the samples.

2. The method according to claim 1, wherein the tags comprise nucleotide sequences determined by the following steps:

- 1) a cDNA pool is synthesized from mRNAs of expressed genes using a primer comprising a recognition sequence of a type III restriction enzyme and an oligo-dT sequence, and treating the cDNA pool with another restriction enzyme;
- 2) a poly(A)-containing fragment is purified from the cDNA pool, and the fragment is ligated to a linker A or B;
- 3) the fragment is treated with a type III restriction enzyme, and the resulting linker A-containing fragment is ligated to a linker B-containing fragment;
- 4) linker sequences are removed by cleaving the ligated fragments with another restriction enzyme used in step 1) to obtain ditag oligonucleotides;
- 5) ditag oligonucleotides are ligated to each other to prepare polynucleotides; and
- 6) the nucleotide sequences of the above polynucleotides are analyzed to determine the nucleotide sequences of tags contained in the polynucleotides.

3. The method according to claim 1, wherein the type III restriction enzyme is EcoP15I.

4. The method according to claim 3, wherein the another restriction enzyme is any of N1aIII, Hsp92II, FatI, BfaI, MaeI, XspI, HpyCH4IV, MaeII, Tail, TscI, AluI, TaqI, BfuCI, Bsp143I, BstENII, DpnII, Kzo9I, MboI, NdeII, Sau3AI, BstKTI, or Csp6I.

5. The method according to claim 4, wherein linker A and linker B are double-stranded DNAs different from each other and are obtained by annealing the following first strand of DNA (1) and second strand of DNA (2)

DNA (1):
5'-N₃₀₋₄₀-CAGCAGCATG-3' (SEQ ID NO: 201)

DNA (2):
3'-N₃₀₋₄₀-GTCGTC-5'

wherein "N₃₀₋₄₀" of DNA (1) is complementary to "N₃₀₋₄₀" of DNA (2) and each thereof is a sequence com-

prising 30 to 40 arbitrary nucleotides, the 5'-end of DNA (1) may be labeled, and the 3'-end of DNA (2) may be amino-modified.

6. A solid support on which tags each comprising an oligonucleotide of more than 25 bp for identifying expressed genes are immobilized, wherein the 3'-end of the tag is defined by a cleavage site of EcoP15I and the 5'-end thereof is defined by a cleavage site of N1aIII located closest to the 3'-end of the cDNA of the genes, and

wherein the tags comprises nucleotide sequences determined by the following steps:

- 1) a cDNA pool is synthesized from mRNAs of expressed genes using a primer comprising a recognition sequence of EcoP15I and an oligo-dT sequence, and treating the cDNA pool with N1aIII;
- 2) a poly(A)-containing fragment is purified from the cDNA pool, and such fragment is ligated to a linker A or B;
- 3) the fragment is treated with EcoP15I, and the resulting linker A-containing fragment is ligated to the linker B-containing fragment;
- 4) linker sequences are removed by cleaving the ligated fragments with N1aIII used in step 1) to obtain ditag oligonucleotides;
- 5) the ditag oligonucleotides are ligated to each other to prepare polynucleotides; and
- 6) the nucleotide sequences of the above polynucleotides are analyzed to determine the nucleotide sequences of tags contained in such polynucleotides.

7. The solid support according to claim 6, wherein linker A and linker B are double-stranded DNAs different from each other that are obtained by annealing the following first strand of DNA (1) and second strand of DNA (2):

DNA (1):
5'-N₃₀₋₄₀-CAGCAGCATG-3' (SEQ ID NO: 201)

DNA (2):
3'-N₃₀₋₄₀-GTCGTC-5'

wherein "N₃₀₋₄₀" of DNA (1) is complementary to "N₃₀₋₄₀" of DNA (2) and each thereof is a sequence comprising 30 to 40 arbitrary nucleotides, the 5'-end of DNA (1) may be labeled, and the 3'-end of DNA (2) may be amino-modified.

8. The solid support according to claim 6, which is prepared by synthesizing tag oligonucleotides thereon.

9. The solid support according to claim 6, which is prepared by immobilizing pre-synthesized tag oligonucleotides thereon.

* * * * *