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(54) COMPOSITIONS AND METHODS FOR THE TREATMENT AND PREVENTION OF VASCULAR MALFORMATIONS

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CPC A61K 31/69; A61K 31/145; A61K 31/407; A61K 31/501; A61K 38/06; A61K 38/07; A61K 38/05

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(57) ABSTRACT

Provided herein are methods and compositions for treating and preventing vascular malformations including lymphatic, venous, capillary, arteriovenous, and combinations thereof. Methods of treatment and prevention include the administration of proteasome inhibitors, omipalisib, disulfiram, and agents which target genes in the PI3K/AKT/mTOR or RAS/RAF/MAPK pathways, including but not limited to Pik3ca, Pik3r3, Tsc2, Rasa1, Map2k2, and Glmn.

6 Claims, 20 Drawing Sheets

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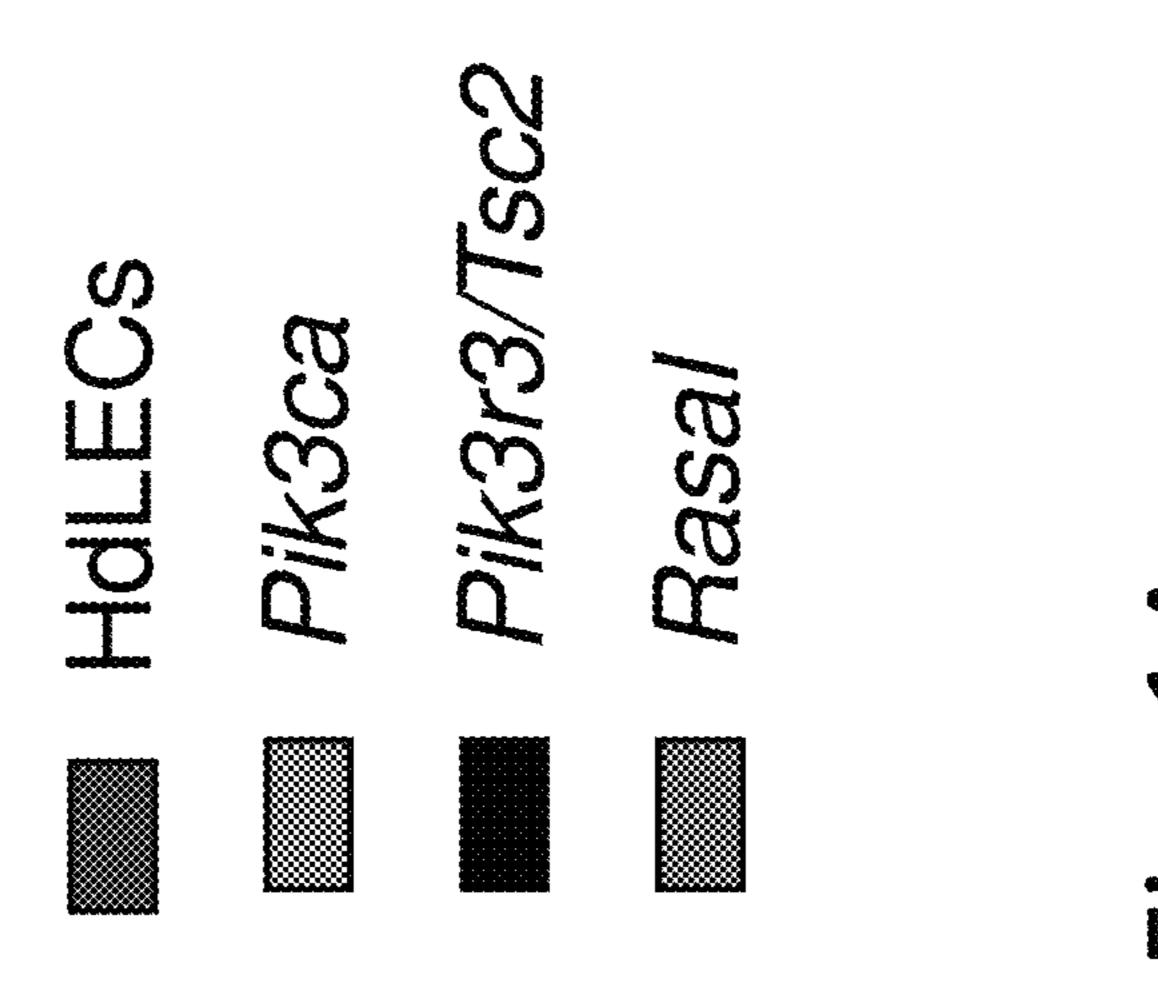
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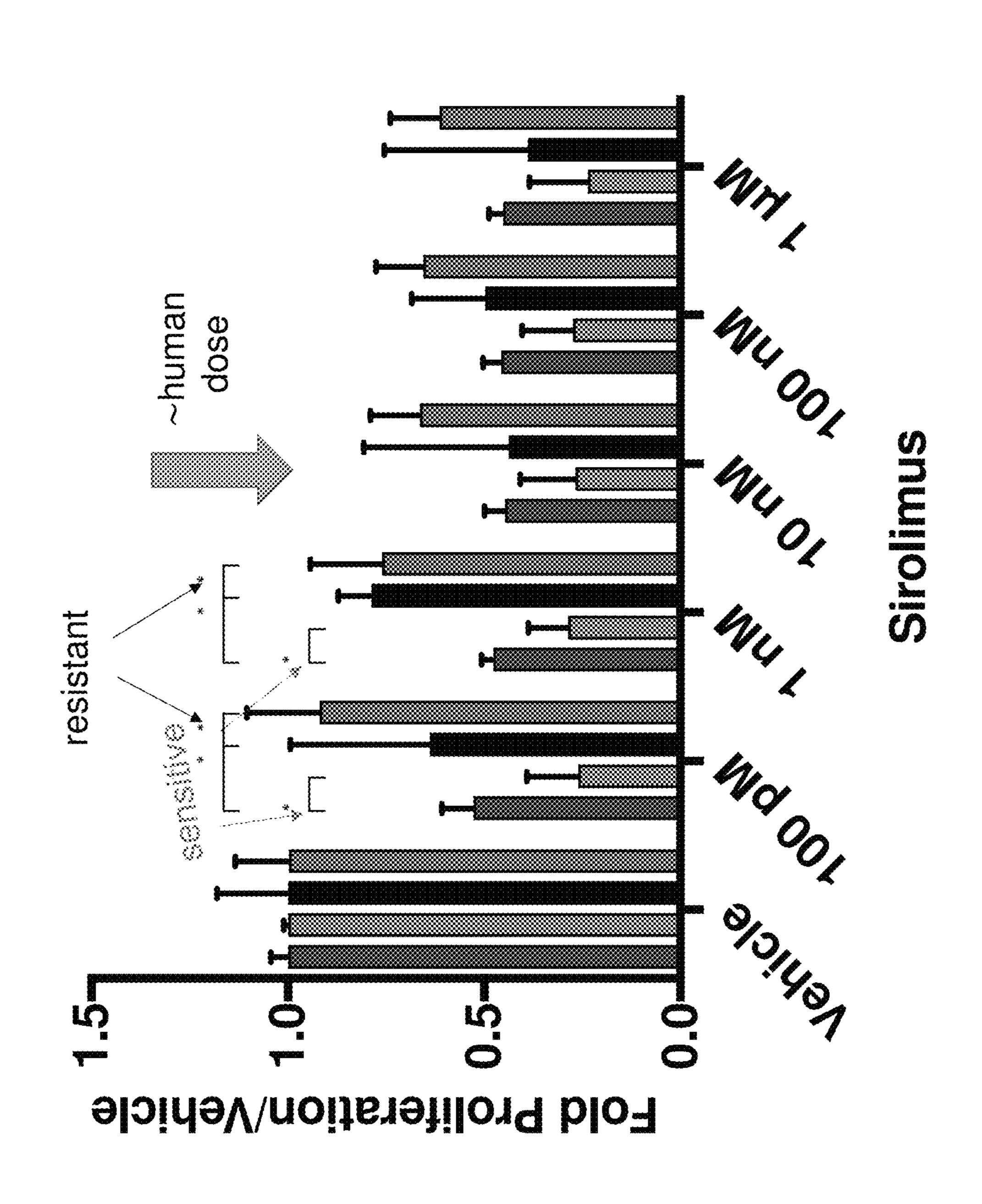
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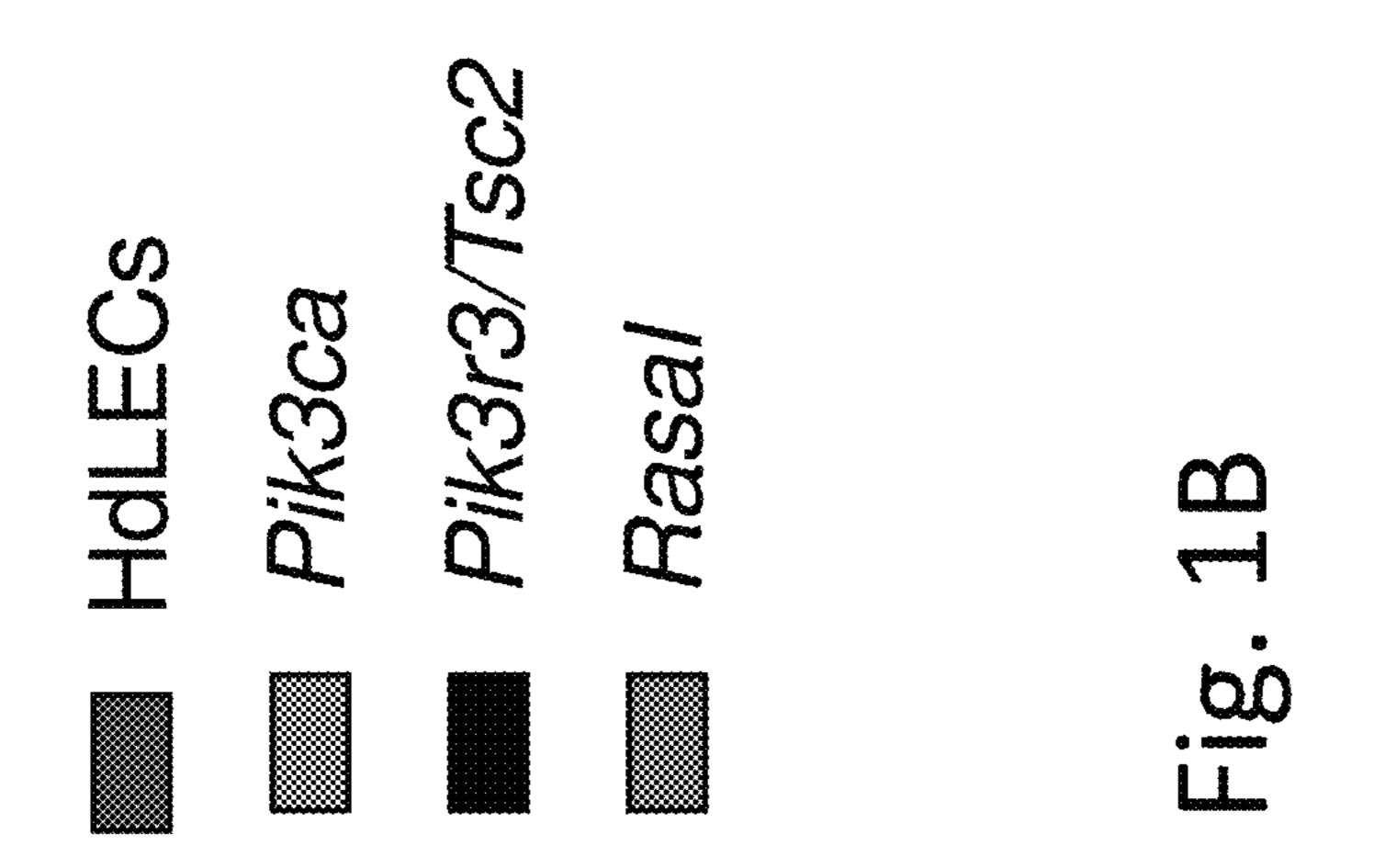
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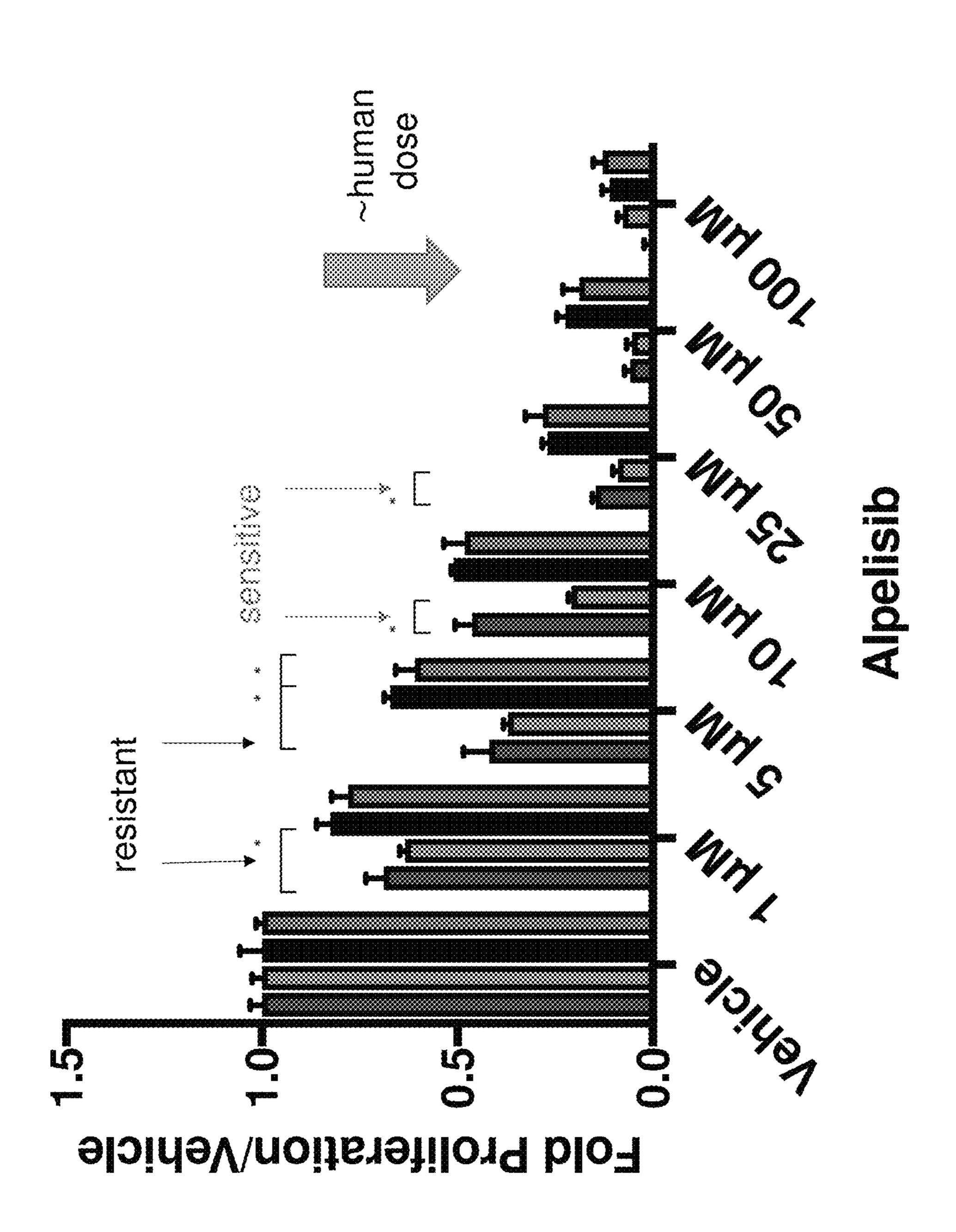
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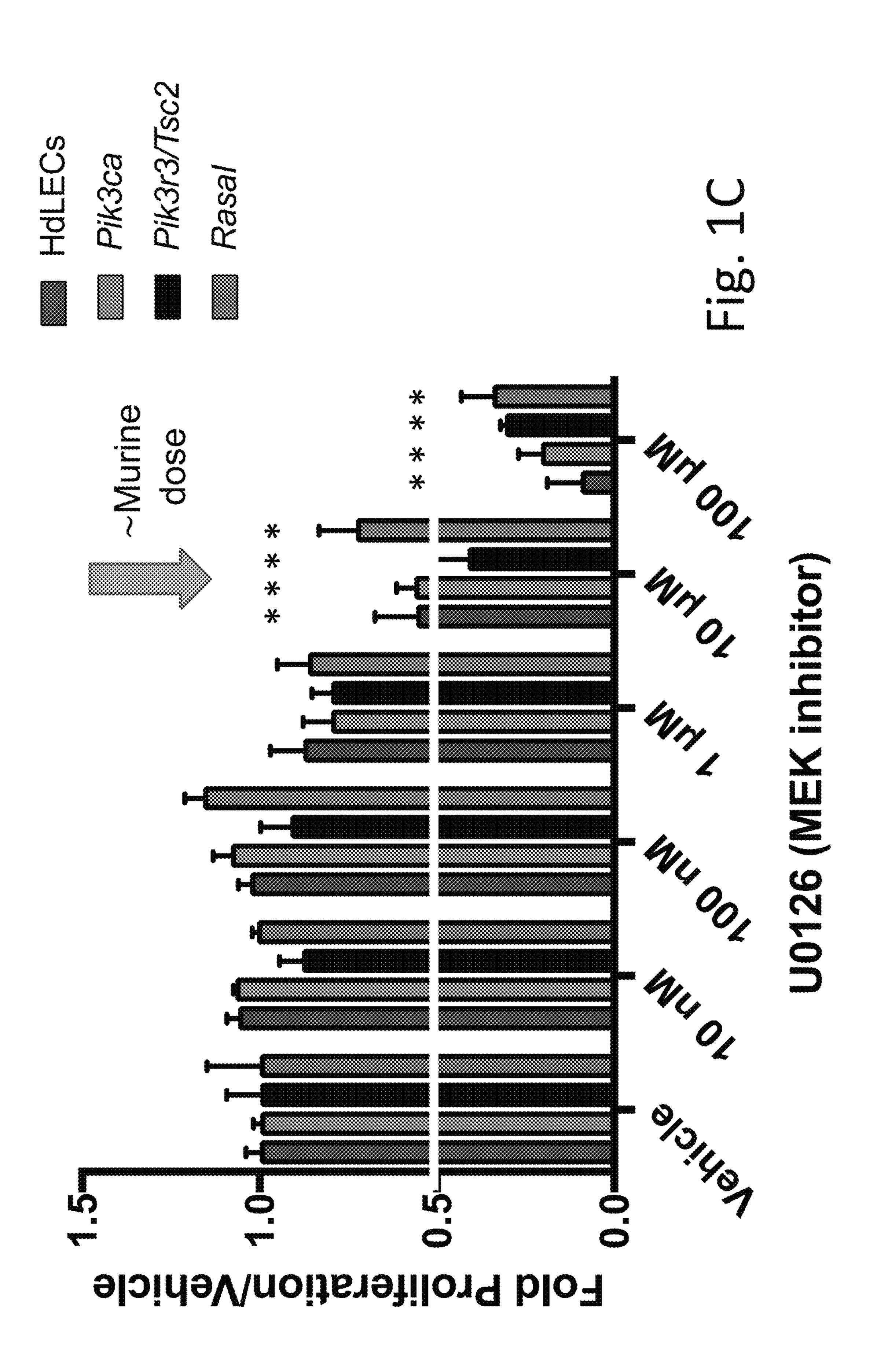
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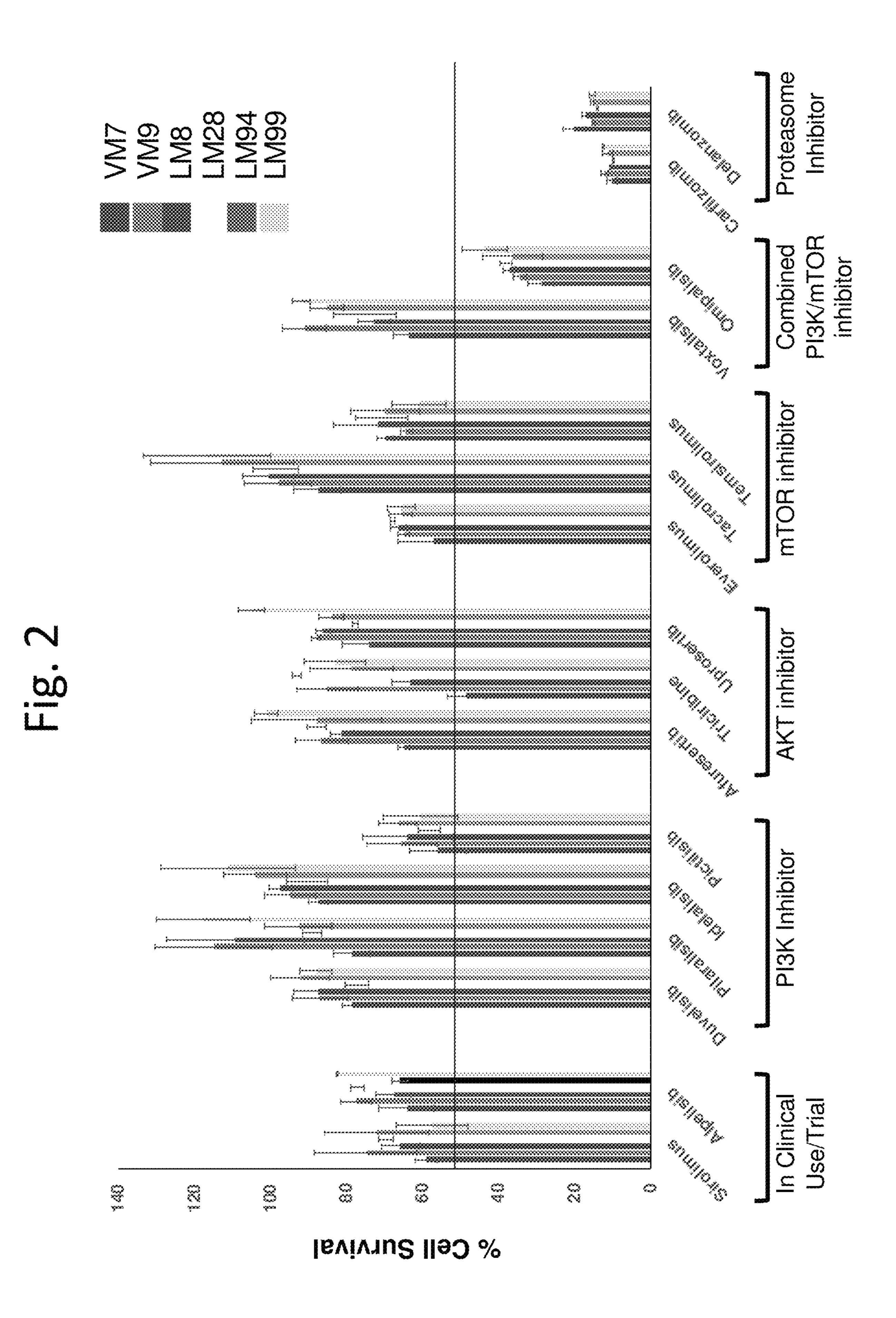


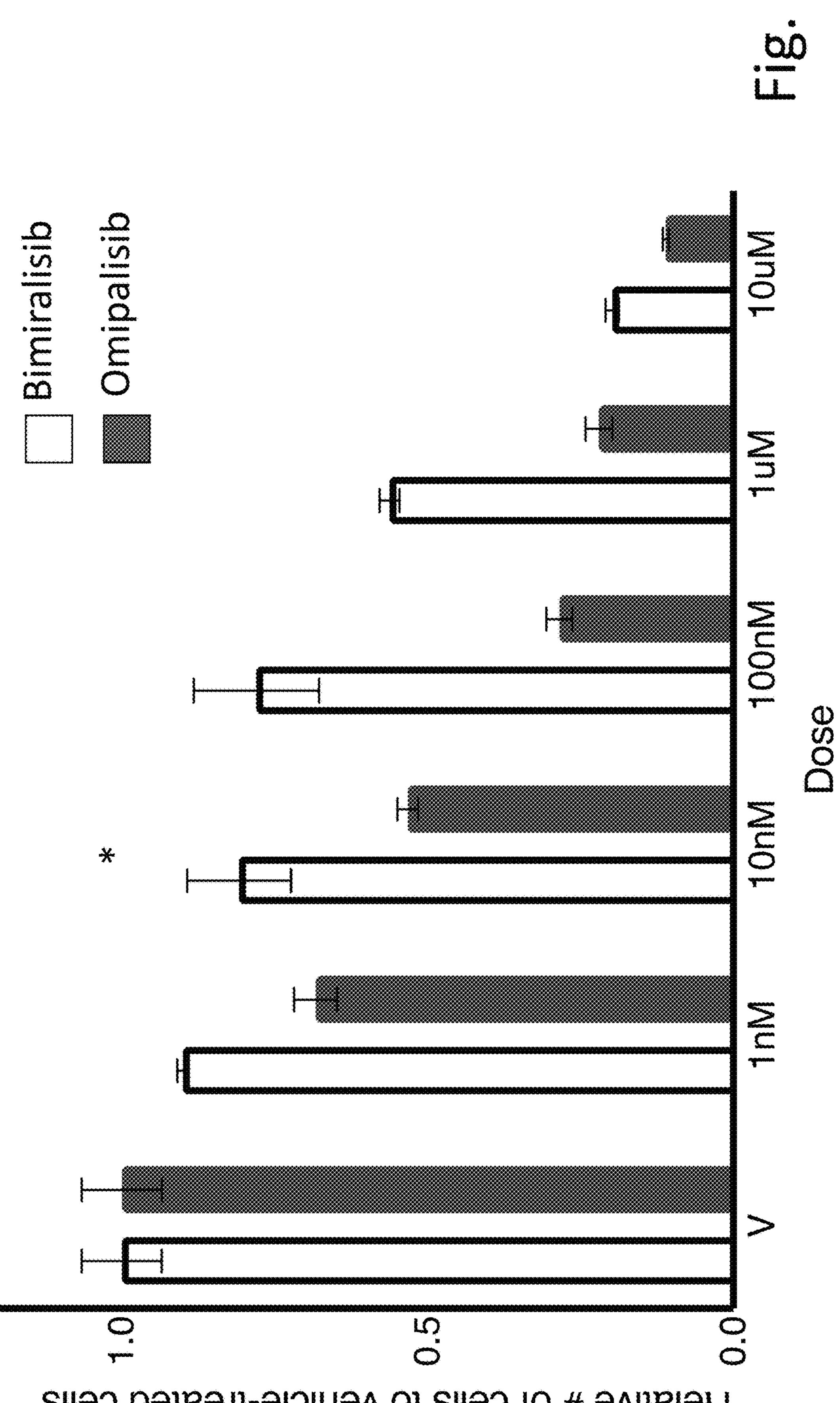




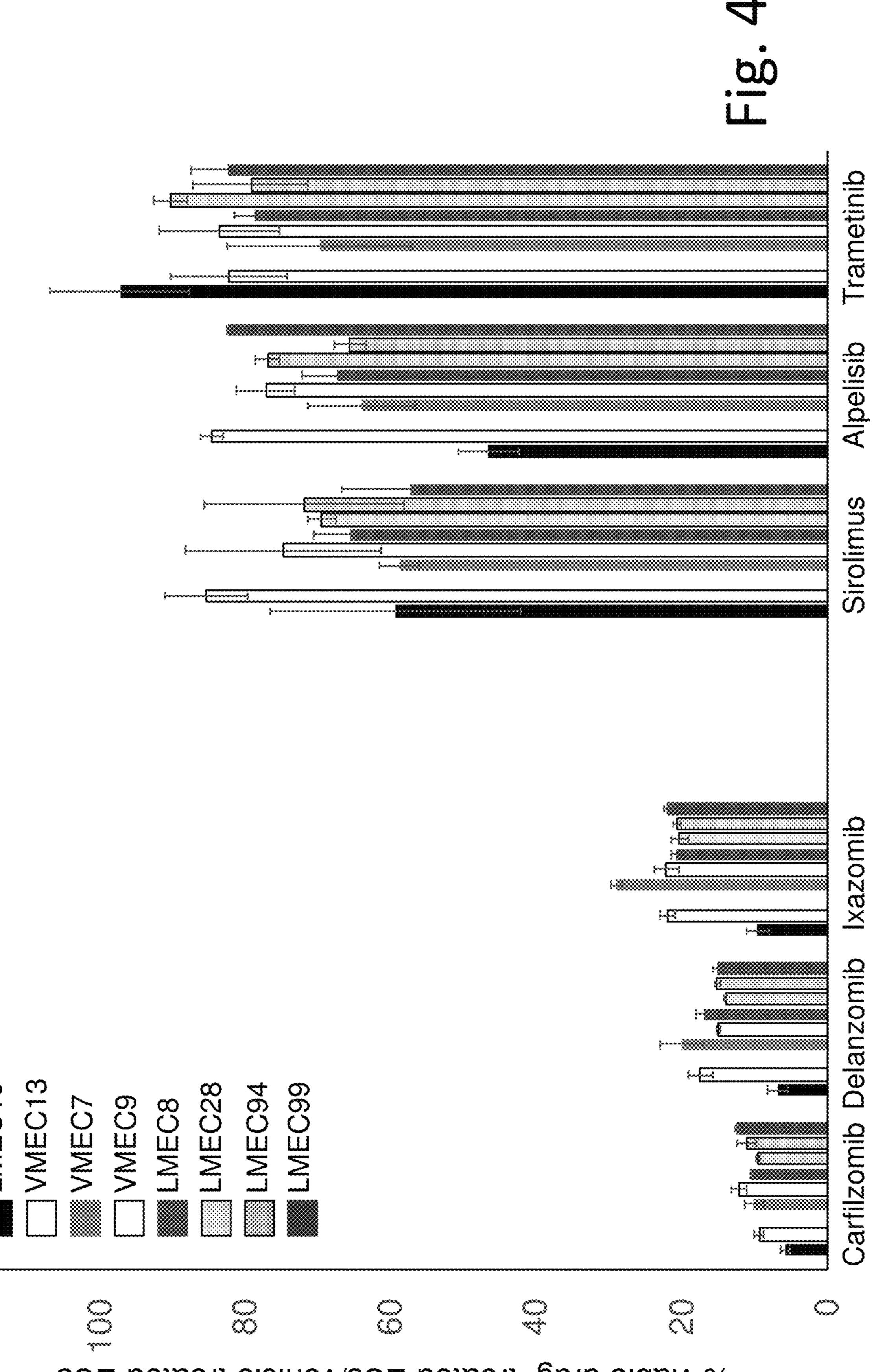




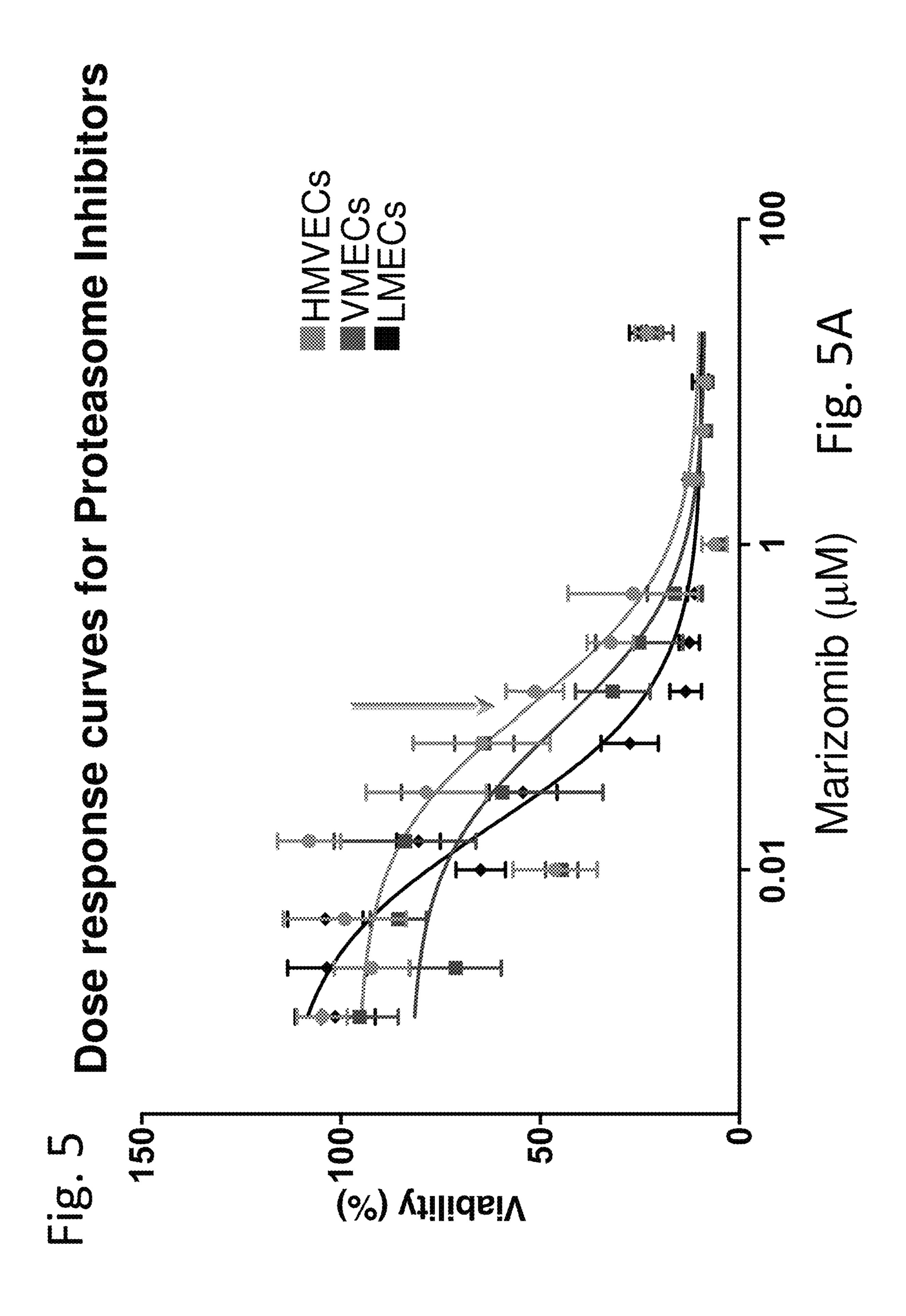


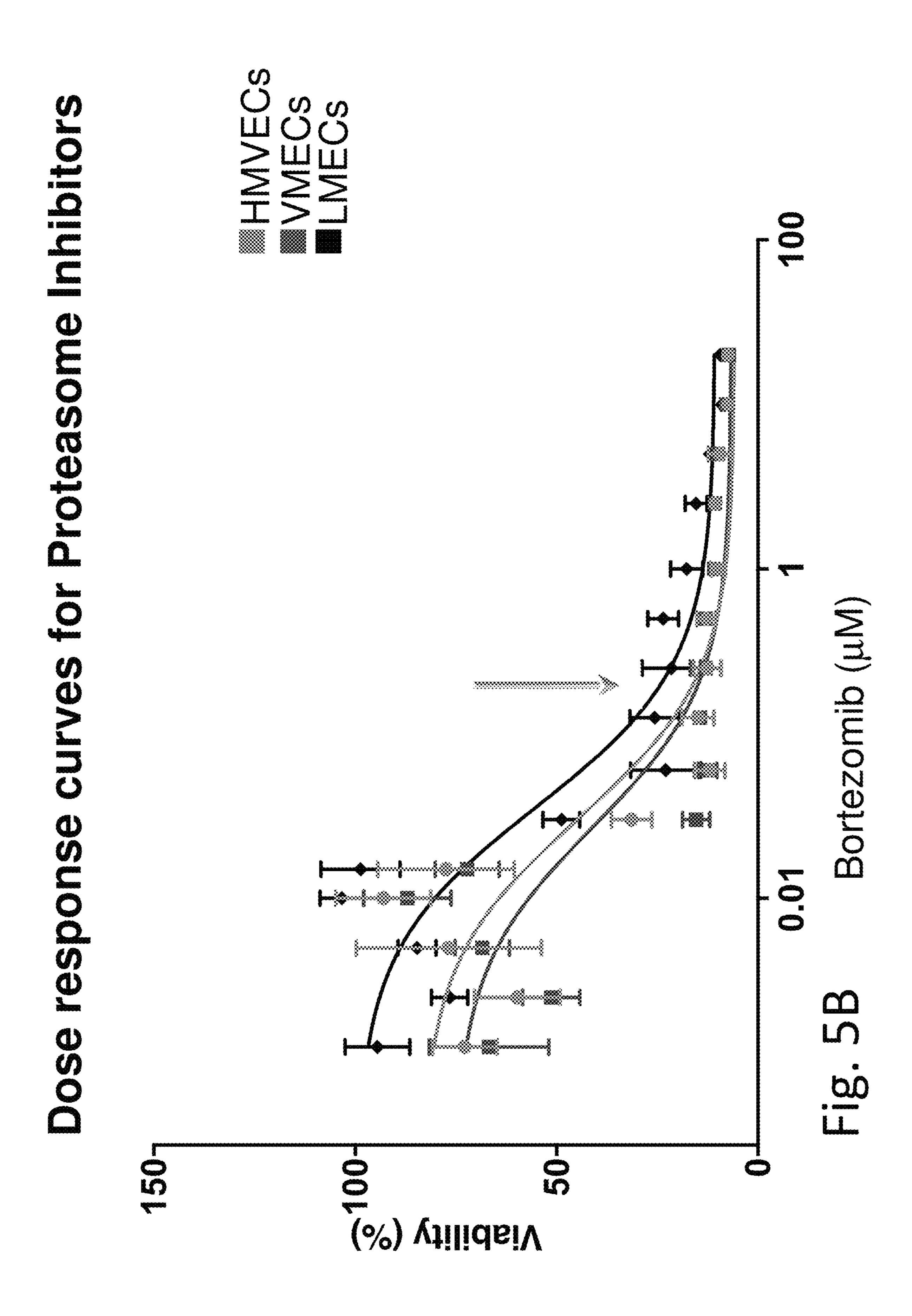


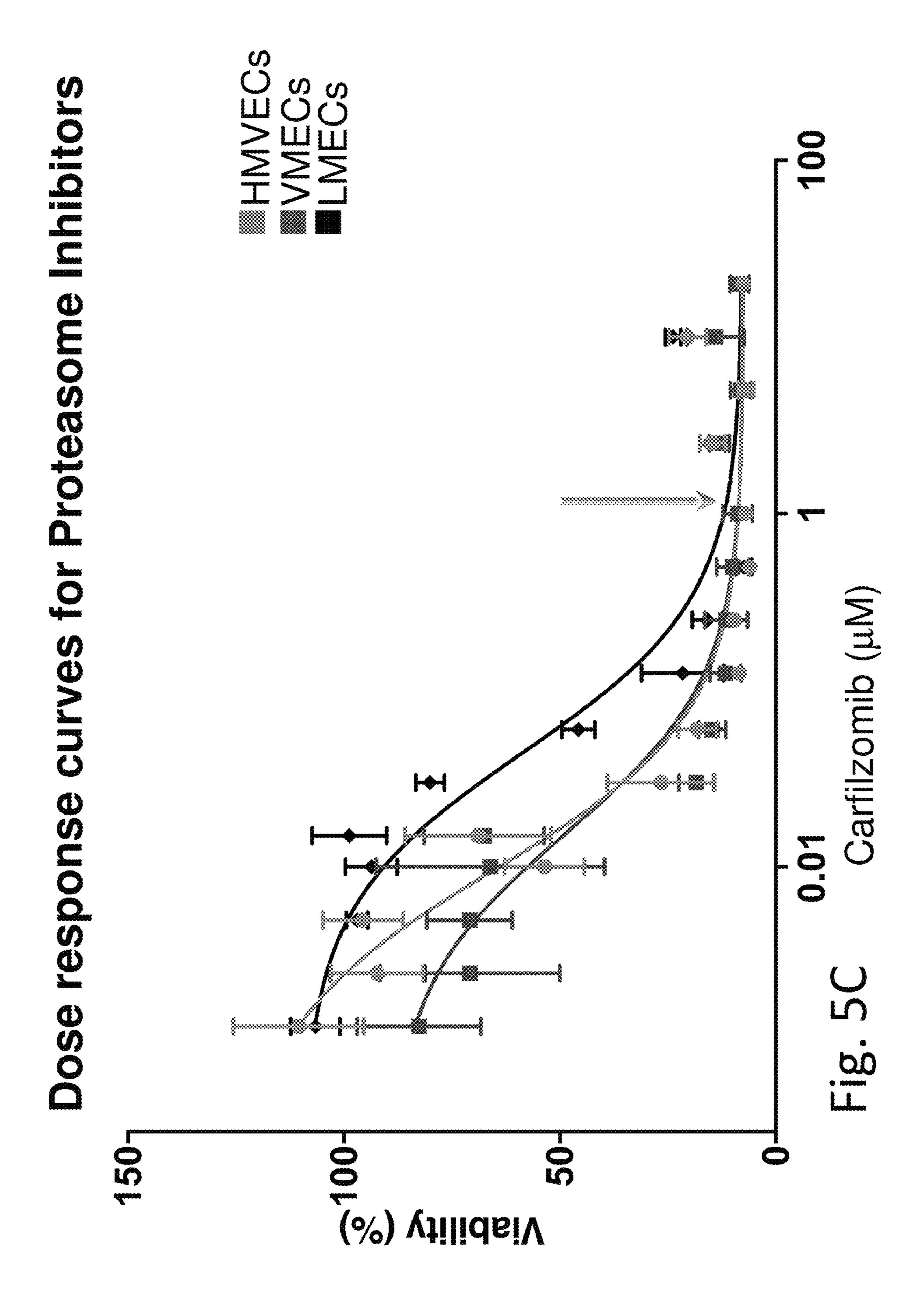
Relative # of cells to vehicle-treated cells

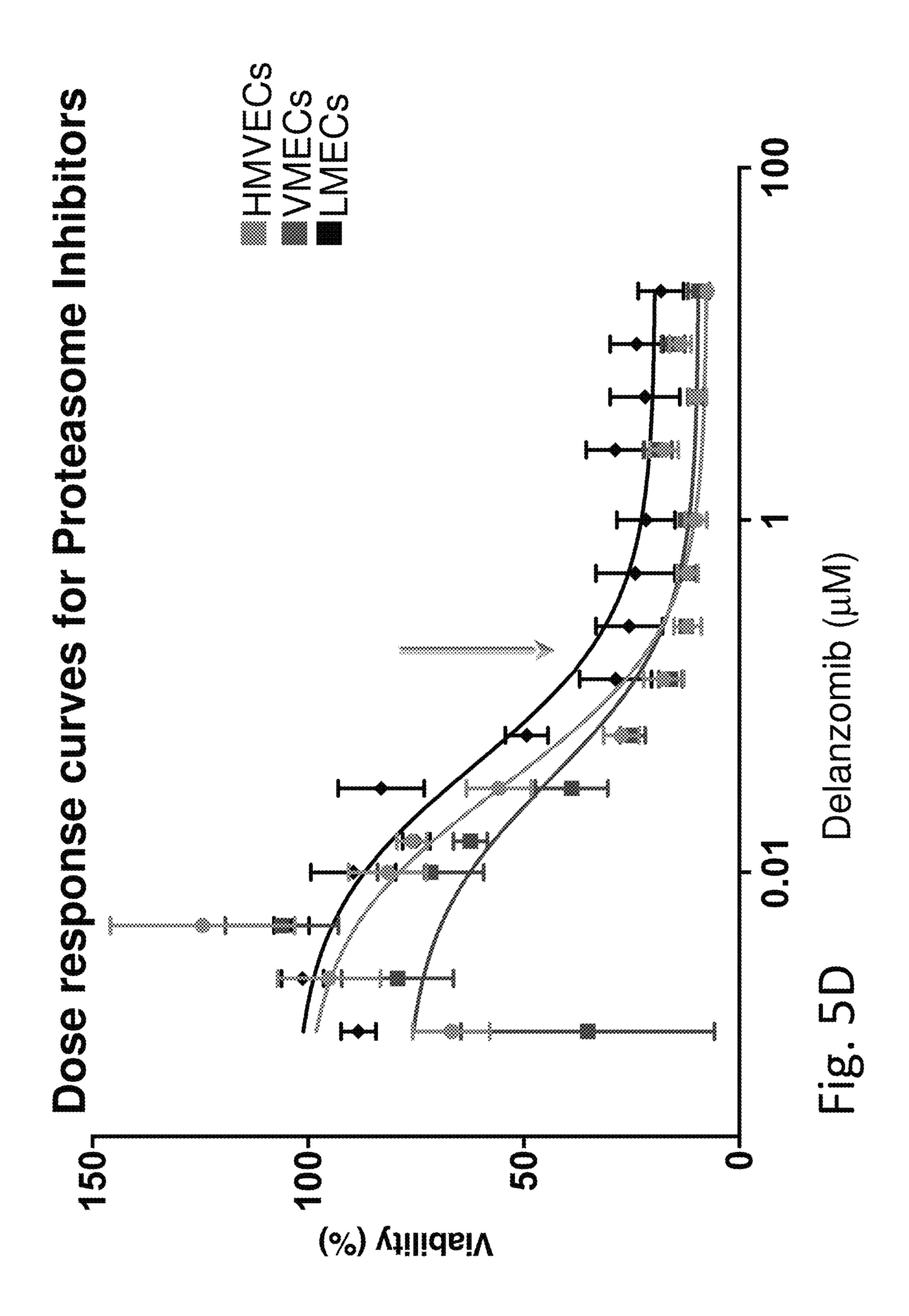


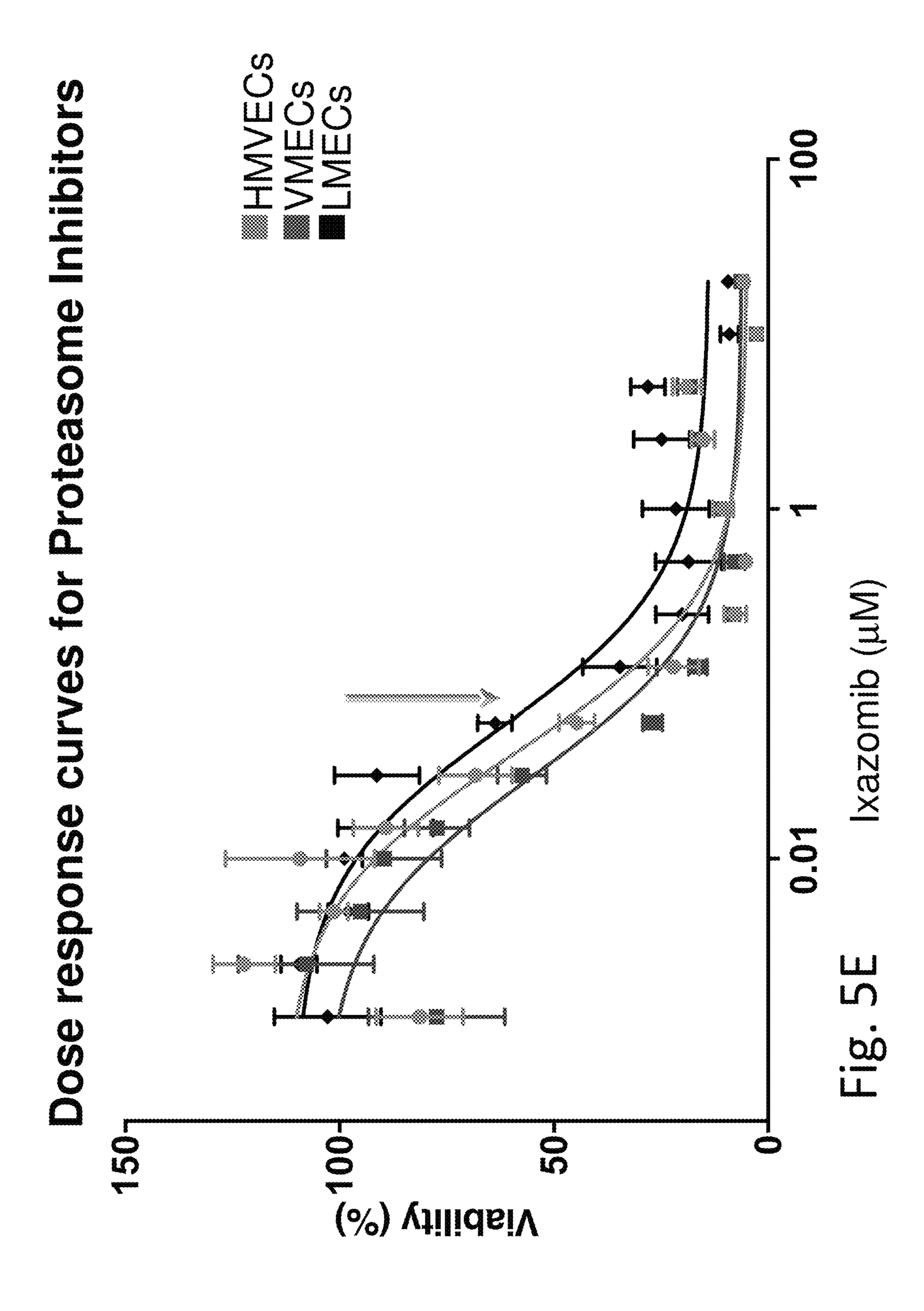
% viable drug treated ECs/vehicle treated ECs

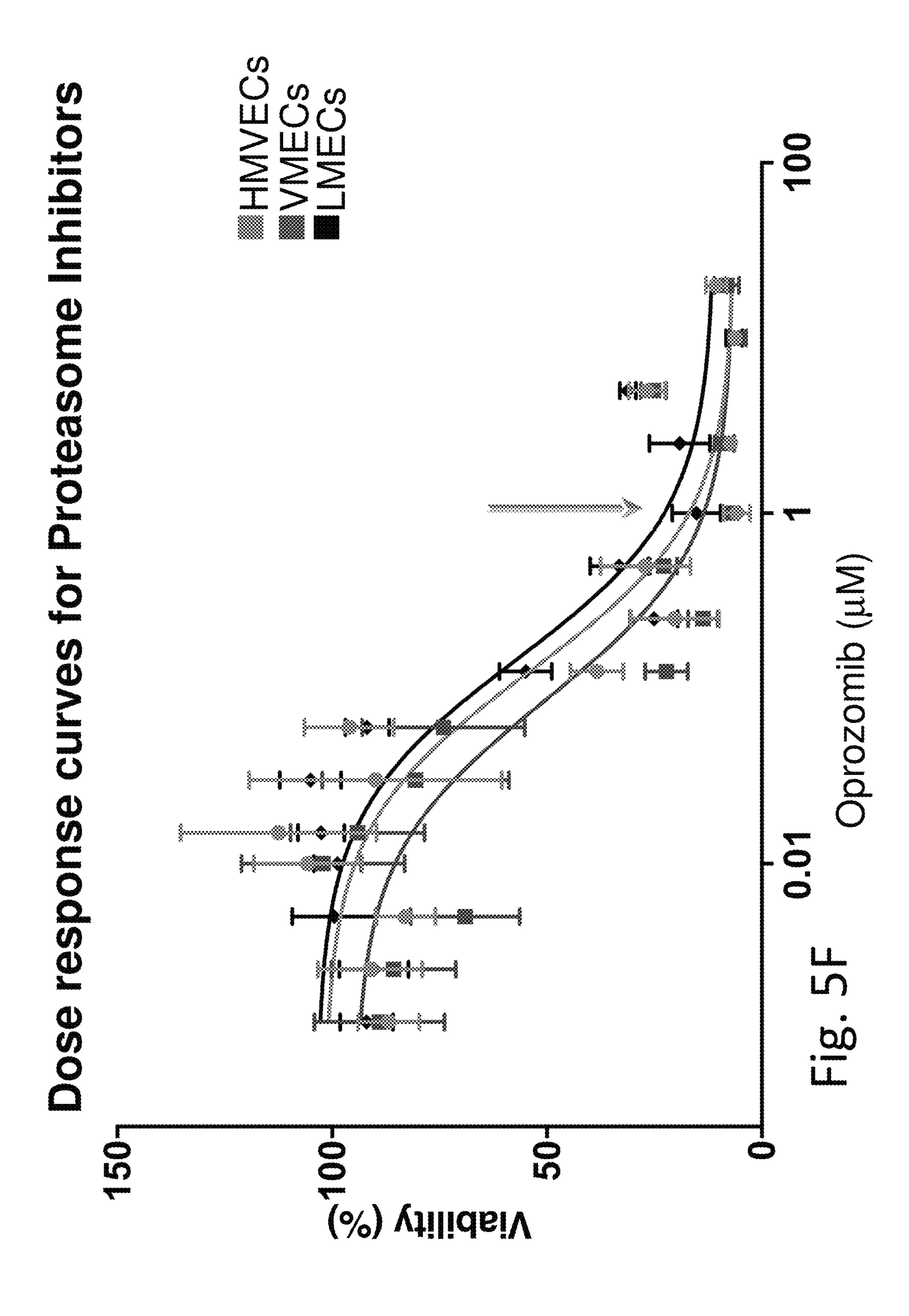


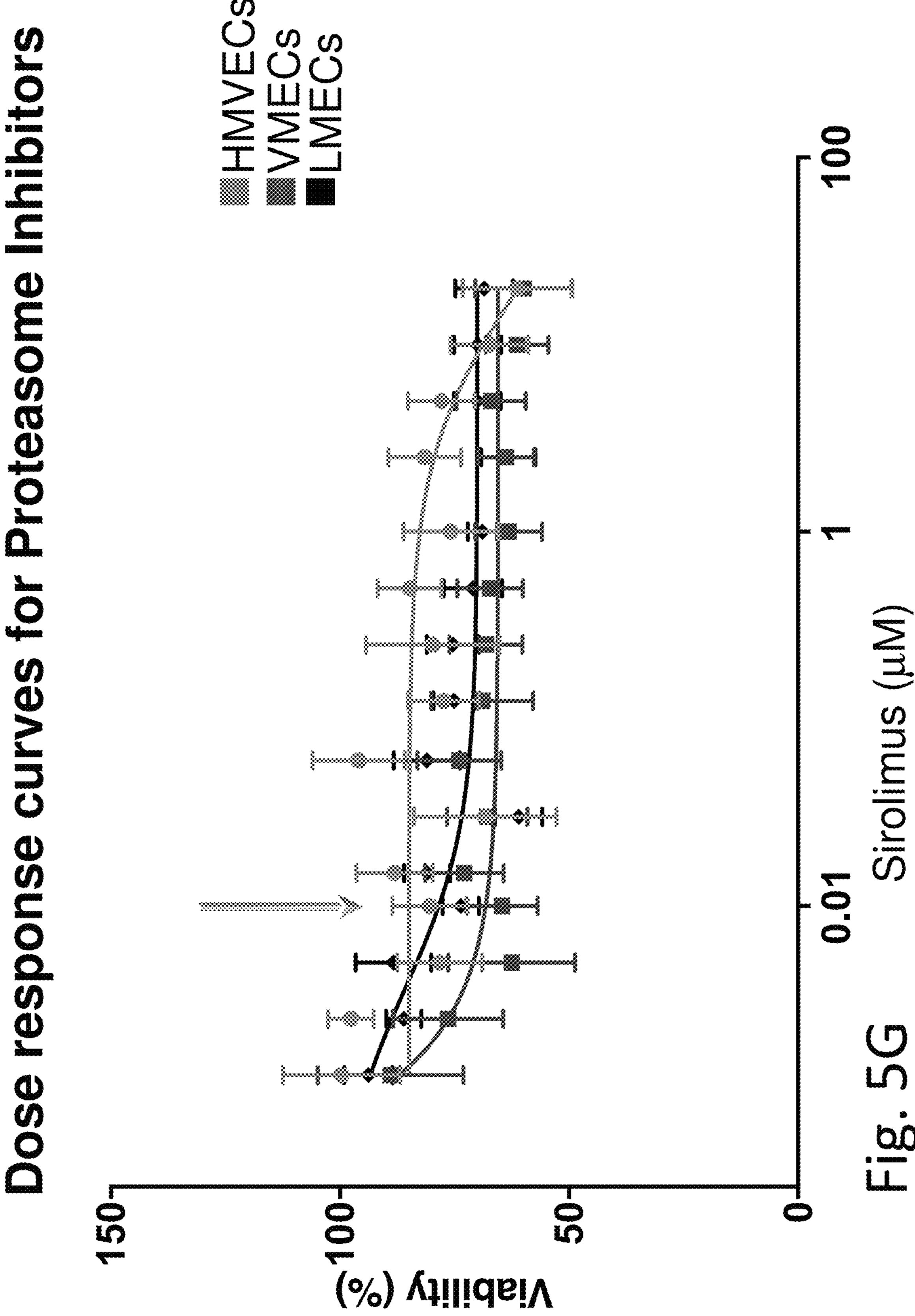


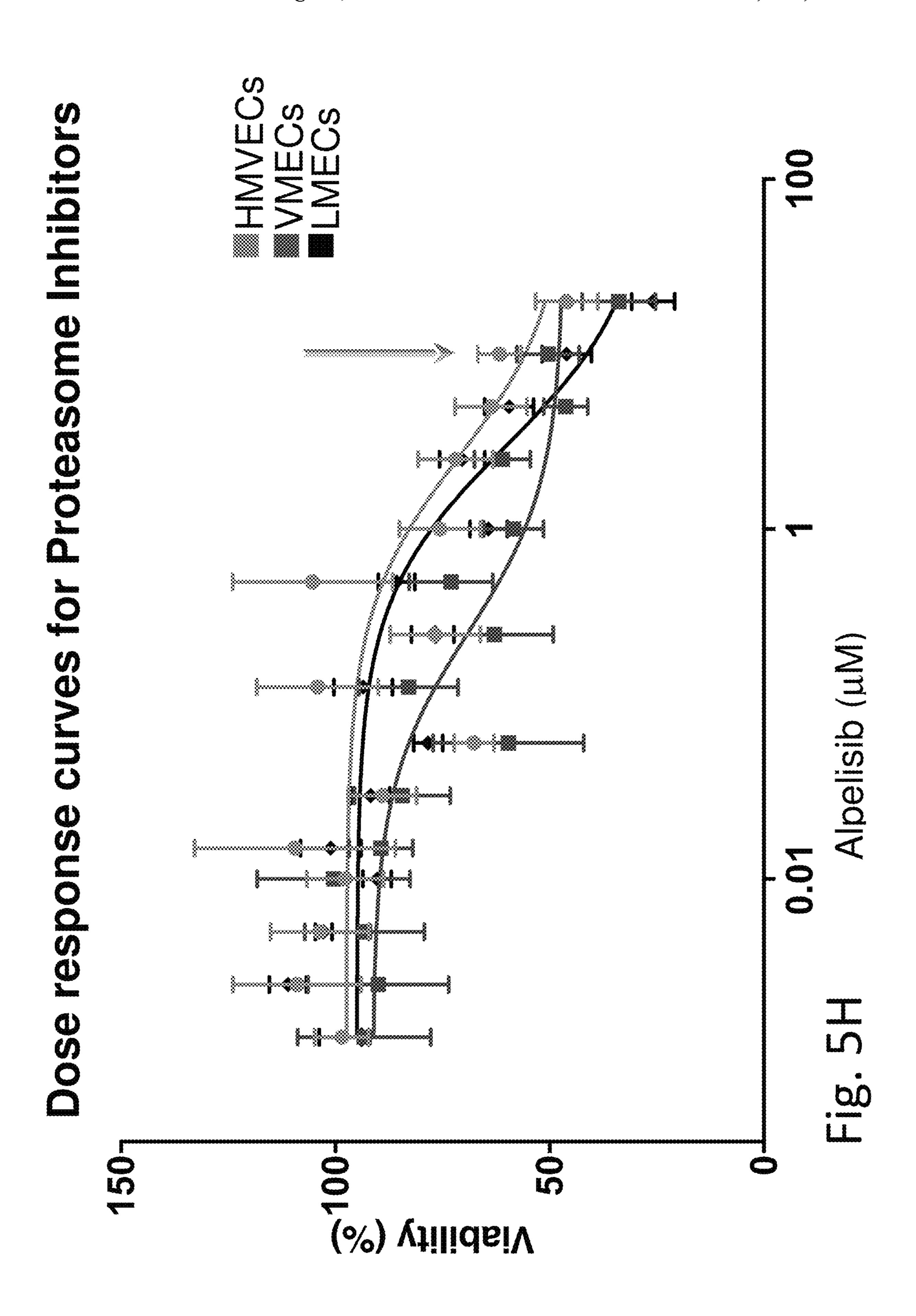


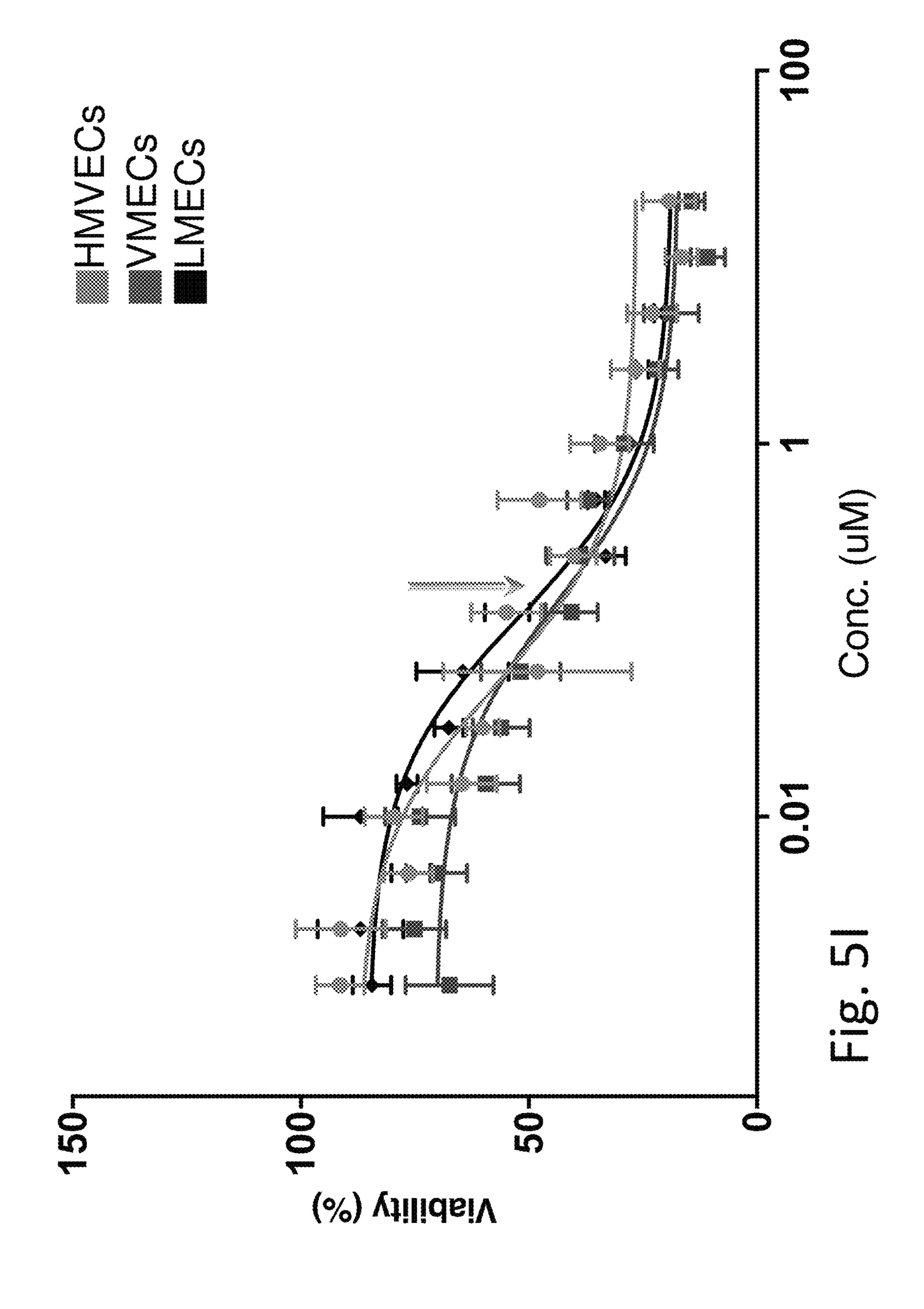




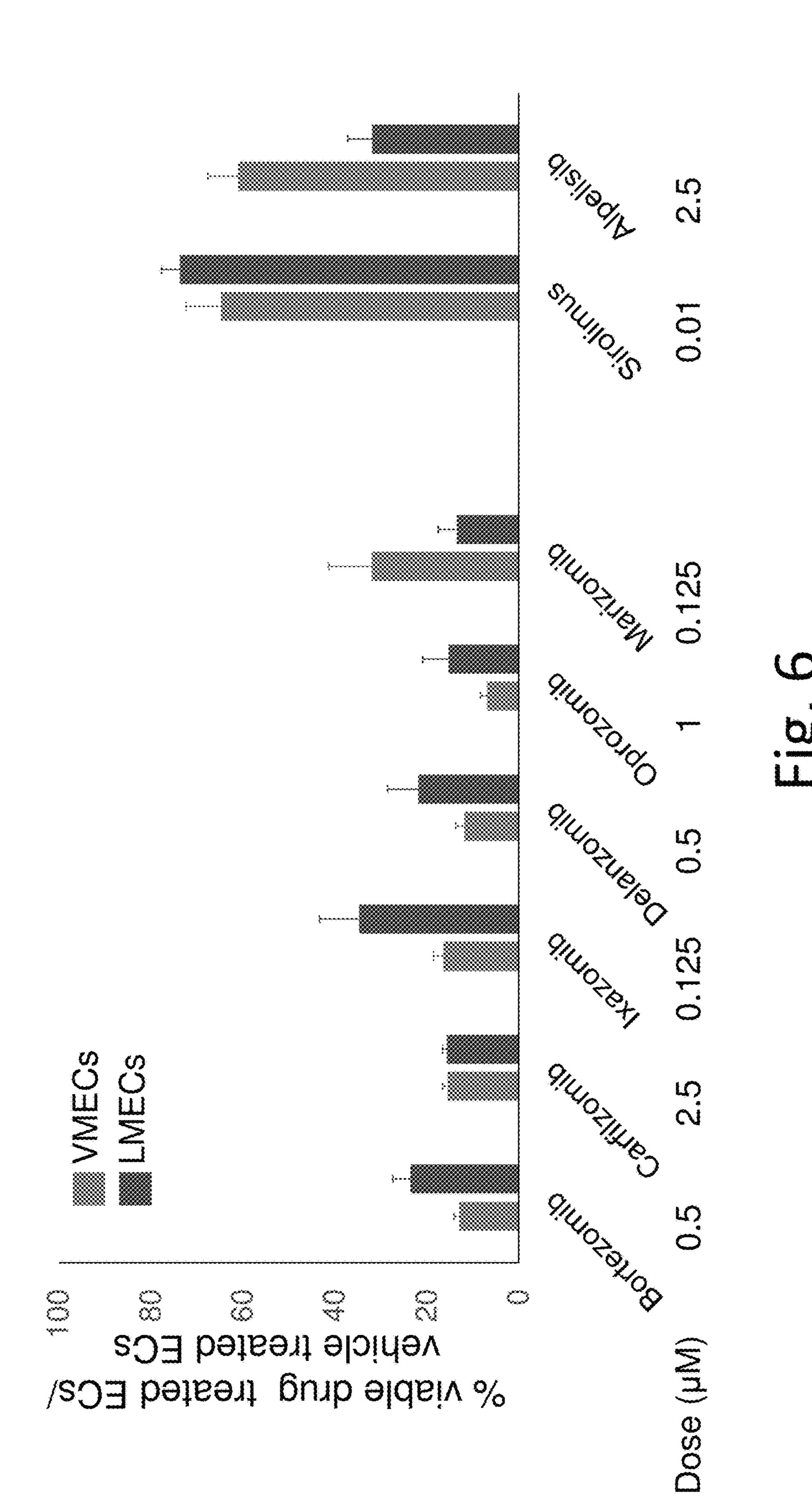


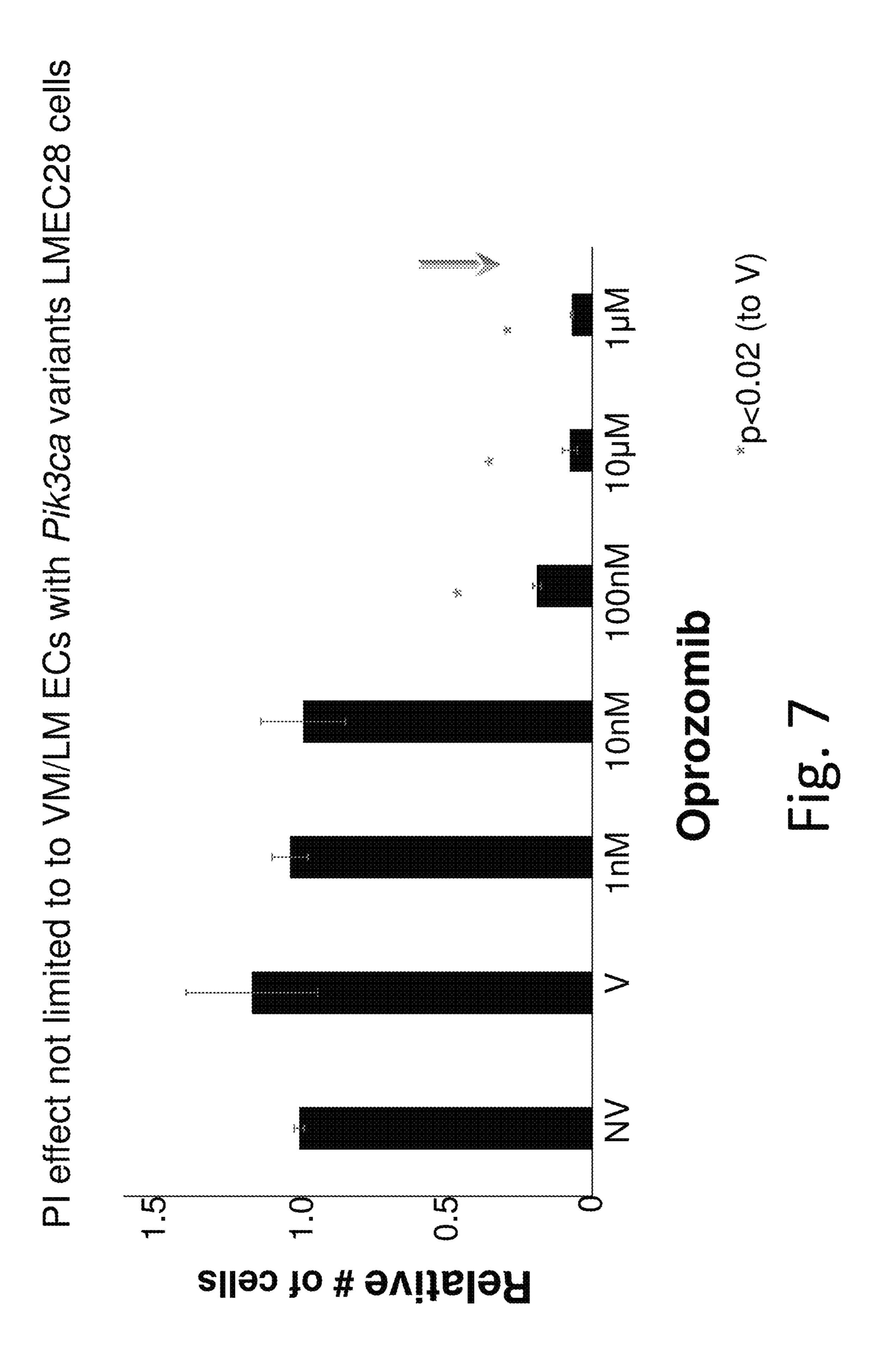


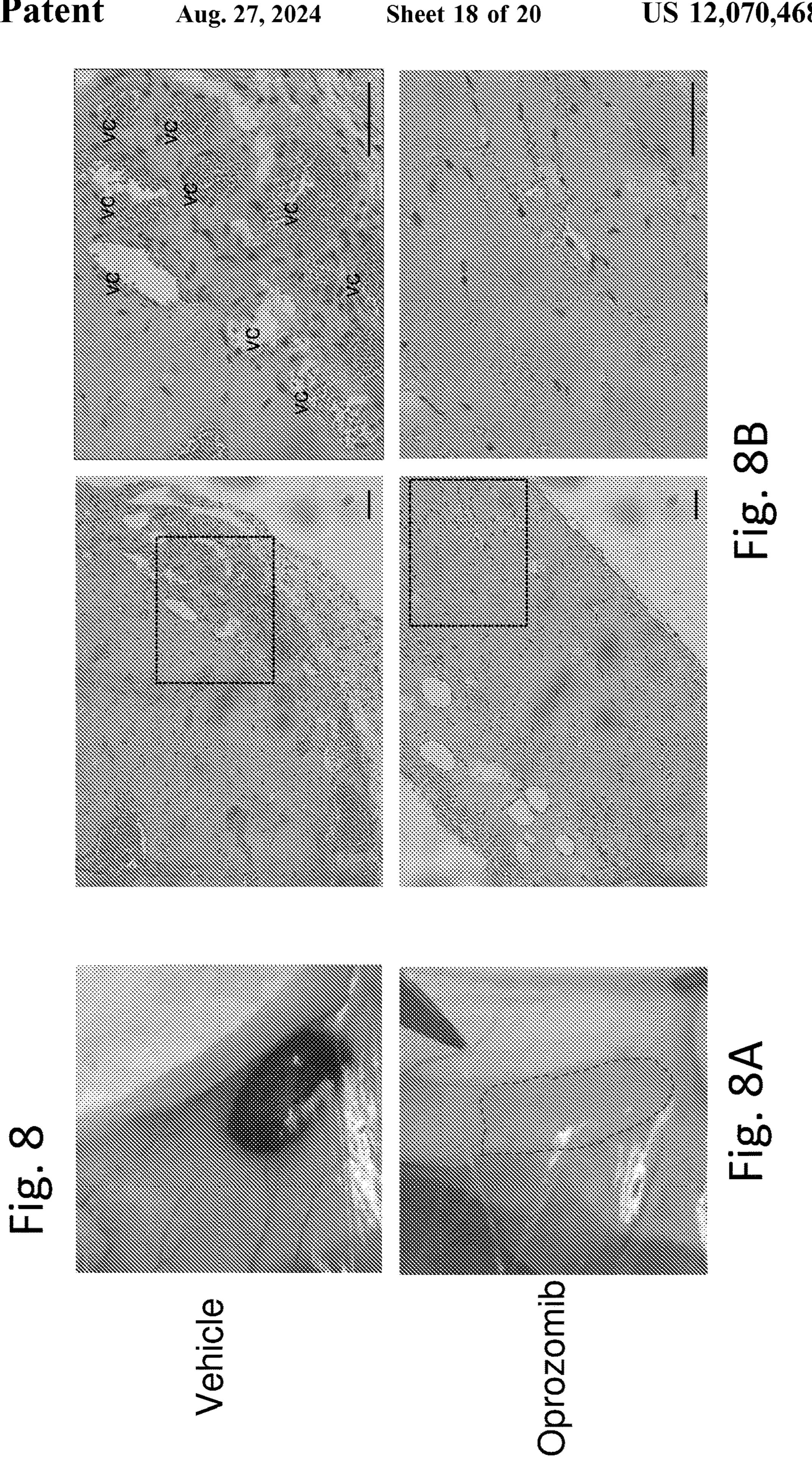




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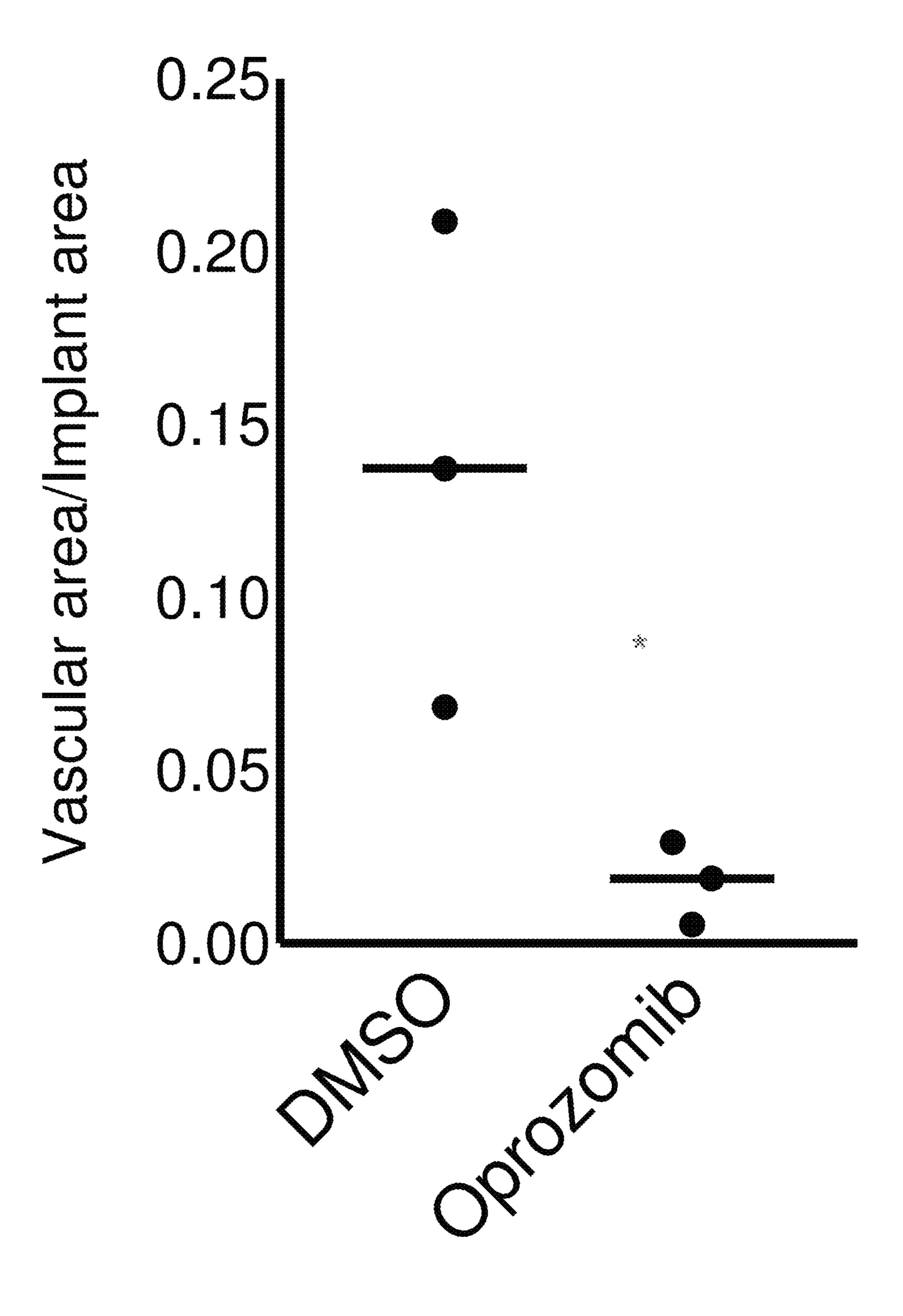
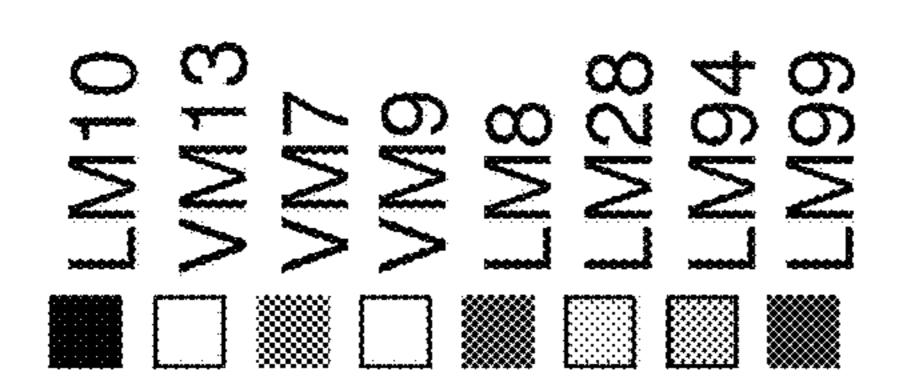
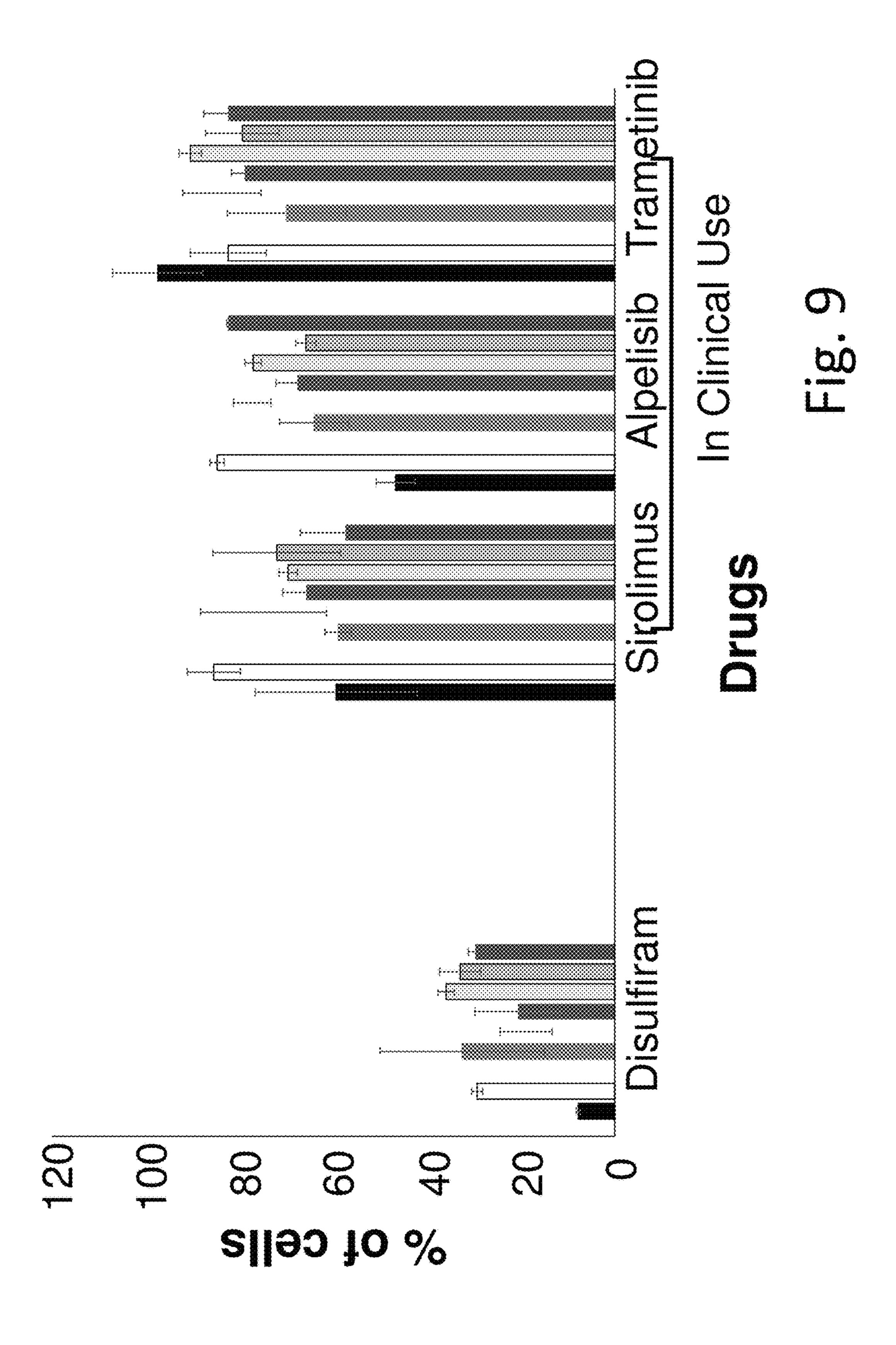


Fig. 8C





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COMPOSITIONS AND METHODS FOR THE TREATMENT AND PREVENTION OF VASCULAR MALFORMATIONS

CROSS REFERENCE TO RELATED APPLICATION

The present application is a U.S. National Phase Application under 35 U.S.C. § 371 of International Patent Application No. PCT/US2021/032374, filed May 14, 2021, which claims priority to U.S. Patent Application Ser. No. 63/024, 670, filed May 14, 2020, each of which is hereby incorporated by reference as if expressly set forth in their respective entirety herein.

STATEMENT OF GOVERNMENTAL INTEREST

This invention was made with government support under W81XWH-19-1-0266, and W81XWH-19-1-0267 awarded by the United States Army Medical Research and Development Command, and HD092662 awarded by the National Institutes of Health. The government has certain rights in the invention.

FIELD

This disclosure is in the field of preventing and treating vascular malformations, as well as in the field of screening for agents which prevent and treat these vascular malformations.

BACKGROUND

Vascular malformations are congenital anomalies of the blood and lymphatic vascular systems with a prevalence 35 ranging from 1:3000 for lymphatic malformations (LMs) to 1% for venous malformations (VMs) and classified by the affected vessel type; capillary (CM), lymphatic (LM), venous (VM), and arteriovenous (AVM) malformations (Greene and Goss 2018; Behravesh et al. 2016). Vascular 40 malformations are often apparent at birth and affected patients have significant morbidities and even mortality. Lymphatic dysfunction in LMs leads to defects in immune surveillance and lipid absorption leading to repeat infections and sepsis to malabsorption (Potente and Makinen 2017). 45 VMs are associated with hematologic anomalies and painful intravascular coagulopathy (Dompmartin et al. 2008). In combined lymphatic and venous malformations (LVMs), deep vein thrombosis can result in life-threatening pulmonary emboli (Mazereeuw-Hautier et al. 2007). AVMs are 50 associated with potentially life-threatening bleeding and heart failure (Uller et al. 2014). Another co-morbidity that affects all types of vascular malformation is hemorrhaging that can require repeated transfusions. In sensitive locations such as the brain, VMs and AVMs can have neurologically 55 devastating consequences that can lead to death (Nikolaev et al. 2018). Despite these severe morbidities, there are no FDA-approved treatment for any vascular malformations.

Currently available treatments (such as surgery and interventional procedures) alleviate symptoms, but are not curative, and recurrence is common often requiring repeat interventions (Greene and Goss 2018; Behravesh et al. 2016). When the malformation is extensive, the effects of these treatments are limited at best. More recently, vascular malformation treatments have been guided by the genetics of the 65 malformation. Studies have shown both germline and postzygotic genetic variants leading to hyperactivation of

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the PI3K/AKT/mTOR and/or the RAS/RAF/MAPK signaling pathways contribute to vascular malformation pathogenesis (Greene and Goss 2018). Based on this knowledge, pharmacotherapies—often cancer drugs—targeting these pathways have been used off-label clinically. Sirolimus, an mTOR inhibitor targeting the PI3K/AKT hyperactivation, has been used most extensively for VMs and LMs, but has generated only partial response (Hammill et al. 2011).

More recently, there have been multiple anecdotal reports using alpelisib, a Pik3ca inhibitor which targets PI3K/AKT/mTOR hyperactivation (Lopez Gutierrez et al. 20191), and trametinib, a MEK inhibitor targeting hyperactivated RAS/RAF/MAPK pathway hyperactivation (Foster et al. 2020). However, no pharmacotherapy, or class of pharmacotherapies, have shown complete response, despite this targeted approach.

Development of effective, biologically targeted therapies is sorely needed, but hampered by our limited understanding of how their genetics influences therapeutic responses.

SUMMARY

Using high throughput screening of endothelial cells from patients with vascular malformations, which had been identified as having specific genetic mutations, pharmacological agents were identified which inhibited the growth and viability of the cells. These pharmacological agents can be used for the treatment and prevention of vascular malformations, as well as to develop therapeutic and preventative agents for vascular malformations.

Thus, one embodiment of the present disclosure is a method of preventing and/or treating vascular malformations, comprising administering to a subject in need thereof a therapeutically effective amount of a proteasome inhibitor.

In some embodiments, the proteasome inhibitor is an inhibitor of the 20s subunit of proteasomes. In some embodiments, the proteasome inhibitor is an inhibitor of the chymotrypsin-like activity (CT-L), trypsin-like activity (T-L) and/or caspase-like (C-L) activity of the 20s subunit. In some embodiments, the proteasome inhibitor is an inhibitor of all three activities. In some embodiments, the proteasome inhibitor is an inhibitor of one of the three activities. In some embodiments, the proteasome inhibitor is an inhibitor of one of the three activities. In some embodiments, the proteasome inhibitor is an inhibitor of chymotrypsin-like activity of the 20s subunit.

In some embodiments, the proteasome inhibitor acts indirectly to inhibit proteolytic activities of proteasome complex.

In some embodiments, the proteasome inhibitor is listed in Tables 1 and 2.

The proteasome inhibitor for use in the method includes but is not limited to carfilzomib, delanzomib, ixazomib, bortezomib, oprozomib, and marizomib. See Table 1.

Additional proteasome inhibitors for use in the method include but are not limited to those listed in Table 2.

A further embodiment of the present disclosure is a method of preventing and/or treating vascular malformations, comprising administering to a subject in need thereof a therapeutically effective amount of disulfiram.

A further embodiment of the present disclosure is a method of preventing and/or treating vascular malformations, comprising administering to a subject in need thereof a therapeutically effective amount of an agent or compound which has proteasome inhibiting ability.

A further embodiment of the present disclosure is a method of preventing and/or treating vascular malforma-

tions, comprising administering to a subject in need thereof a therapeutically effective amount of a copper-binding agent or compound.

A further embodiment of the present disclosure is a method of preventing and/or treating vascular malforma- 5 tions, comprising administering to a subject in need thereof a therapeutically effective amount of omipalisib.

A further embodiment of the present disclosure is a method of preventing and/or treating vascular malformations, comprising administering to a subject in need thereof 10 a therapeutically effective amount of an agent which targets a mutation in a gene chosen from the group consisting of Pik3ca, Pik3r3, Tsc2, Rasa1, Map2k2, Glmn, and combinations thereof.

method of preventing and/or treating vascular malformations, comprising administering to a subject in need thereof a therapeutically effective amount of an agent which targets a mutation in a gene in the PI3K/AKT/mTOR or RAS/RAF/ MAPK pathway. Such genes include but are not limited to 20 Kras and Nras.

Vascular malformations that can be prevented and/or treated by the disclosed methods include but are not limited to lymphatic malformations (LMs), venous malformations (VMs), capillary malformations (CM), arteriovenous mal- 25 formations (AVM), and combinations thereof.

In some embodiments, the agent further comprises a pharmaceutically acceptable carrier and is part of a composition.

In some embodiment, the subject is over 18 years old. In 30 some embodiments, the subject is under 18 years old. In some embodiments, the subject is under 16 years old. In some embodiments, the subject is under 10 years old. In some embodiments, the subject is under 5 years old. In some embodiments, the subject is under 1 year old.

In an additional embodiment, the disclosure provides for methods of obtaining cells from the vascular malformation of the subject and testing the efficacy and/or dosage of any agent prior to administration of the agent. In some embodiments, these cells are endothelial cells. In some embodi- 40 ments, the cells are subject to sequencing or other analysis to determine the mutations and/or defects within the cells.

A further embodiment of the present disclosure are kits comprising compositions and agents for practicing the disclosed methods.

A further embodiment of the present disclosure is a method and/or assay for screening and/or identifying an agent for the treatment and/or prevention of vascular malformations including lymphatic, venous, capillary, and arteriovenous and combinations thereof, comprising contacting 50 or incubating a test agent with a cell derived from a patient with a vascular malformation, wherein if the viability and/or proliferation of the cells is decreased after contact or incubation with the test agent, the test agent is identified as a therapeutic and/or preventative agent for vascular malfor- 55 mations.

In some embodiments, the cells are endothelial cells. In some embodiments, the cells harbor mutations in a gene in the PI3K/AKT/mTOR or RAS/RAF/MAPK pathway. In some embodiments, the cells harbor mutations in a gene 60 chosen from the group consisting of Pik3ca, Pik3r3, Tsc2, Rasa1, Map2k2, Glmn, and combinations thereof. In some embodiments, the cells are chosen from the group of cells listed in Table 3. In some embodiments, high throughput screening is performed. In some embodiments, the cells are 65 subject to sequencing or other analysis to determine the mutations and/or defects within the cells.

The disclosure also includes any agents identified by the screening methods described herein and methods of using the same for the prevention and/or treatment of vascular malformations including lymphatic, venous, capillary, and arteriovenous and combinations thereof.

BRIEF DESCRIPTION OF THE FIGURES

For the purpose of illustrating the invention, there are depicted in drawings certain embodiments of the invention. However, the invention is not limited to the precise arrangements and instrumentalities of the embodiments depicted in the drawings.

FIG. 1—Inhibitors that target the PI3K/AKT and RAS/ A further embodiment of the present disclosure is a 15 MAPK pathways display different efficacies against LMECs. FIG. 1A shows the results of LMECs with different genetic variants (PIK3CA, PIK3CA/TSC2, and RASA1) and control cells (HdLECs) treated with increasing log doses of sirolimus. FIG. 1B shows the results of the same cells treated with increasing log doses of alpelisib. FIG. 1C shows results of the same cells treated with increasing log doses of the MAPK kinase inhibitor U0126. Proliferation rate was determined by WST-8 assay and expressed as a fraction of the number of cells in vehicle controls±SD. *p<0.05. The large arrow denotes the range of human doses (FIGS. 1A and 1B) and murine doses (FIG. 1C) used in clinical setting.

> FIG. 2—PI3K/AKT pathway inhibitors differentially affect cell survival in a HTPS assay. Six endothelial cell populations isolated from venous malformation (VM) and lymphatic malformation (LM) patients (VMECs and LMECs), carrying mutations that hyperactivate PI3K signaling, were subjected HTPS with all inhibitors at 1 μM. Cell survival was determined after 48 hours and data presented as percent cell survival of cells treated with 1 µM 35 compound versus vehicle-treated control±SD. Small molecules presented belonged to 2 classes of inhibitors: PI3K inhibitors and proteasome inhibitors (right). Dark gray line: 50% cell from starting number. Omipalisib and proteasome inhibitors were significantly more efficacious than sirolimus at inhibiting growth/survival of the ECs with PI3K hyperactivation. Sirolimus to PI3K/mTOR inhibitors: ANOVA p=0.0017, post-hoc TTEST sirolimus vs. omipalisib p=0.0005. Sirolimus vs proteasome Inhibitors: ANOVA p=0.0013, post-hoc TTEST sirolimus vs. carfilzomib 45 p<0.0001, sirolimus vs. delanzomib p=0.0093.

FIG. 3—Omipalisib demonstrated as a specific inhibitor with improved efficacy against LMEC28. Cell survival was determined after 48 hours and data presented as percent cell survival of cells treated versus vehicle treated cells. Omipalisib, a PI3K/mTOR inhibitor, at a lower dose (*) was more efficacious than another PI3K/mTOR dual inhibitor, bimiralisib, in inhibiting LMEC proliferation. At 10 nM; TTEST p<0.03.

FIG. 4—Proteasome Inhibitors efficaciously suppressed VMEC and LMEC growth/viability more than that observed for drugs used off label for vascular malformation patients. FIG. 4 is a graph of cell viability of treated cells relative to vehicle treated cells. Using HTPS, VMEC/LMECs carrying variants in genes in the RAS/RAF/MAPK (LMEC10, VMEC13) or PI3K/AKT/mTOR (VMEC7/9, LMEC8/28/ 94/99) pathways (Table 3) were treated with 1 µM of 1344 compounds. Cell viability relative to vehicle treated cells was determined after 48 hours. Data presented for three PIs tested, and sirolimus, alpelisib, and trametinib, all of which have been used off label for vascular malformation patients.

FIG. 5—Dose Response Curves of PIs, sirolimus, alpelisib and omipalisib using HMVEC (n=2) (control/ 5

normal/wild-type cells to be compared to those from vascular malformation patients), VMEC7, VMEC9, LMEC94 and LMEC99 (Table 3). The graphs show the percent viability of the cells at the various dosages in μ M (dose range, 0.00125 μ M to 20 μ M). The large arrows mark the reported plasma concentration in humans. FIG. 5A shows marizomib. FIG. 5B shows bortezomib. FIG. 5C shows carfilzomib. FIG. 5D show delanzomib. FIG. 5E shows ixazomib. FIG. 5F shows oprozomib. FIG. 5G shows sirolimus. FIG. 5H shows alpelisib. FIG. 5I shows omipalisib.

FIG. 6—Proteasome inhibitors (PIs) are more effective at reducing viable cells than current therapies currently used off-label for vascular malformation patients. FIG. 6 is a graph of the results of VMECs (n=2) and LMECs (n=2) carrying Pik3ca mutations subjected to increasing amounts of designated PI (left 6) or sirolimus or alpelisib, and cell viability determined after 48 hours at doses corresponding to clinically reported plasma concentration in μM. Data represent plasma concentration achieved in humans and presented as % viable cells treated with drug relative to vehicle treated cells.

FIG. 7—PI effect not limited to VM/LM ECs with Pik3ca variants. FIG. 7 is a graph of the results of LMEC28 carrying Pik3r3;Tsc2 mutations subjected to increasing amounts of 25 oprozomib and cell viability determined after 48 hours. Yellow arrow marks clinically relevant dose and presented as % viable cells treated with drug relative to vehicle treated cells.

FIG. **8**—In vivo testing of PIs. In a xenograft model, ³⁰ oprozomib inhibited VM vessel growth. FIG. **8**A are images of the gross appearance of implants after treatment with vehicle or oprozomib. The dotted line marks the border of the Matrigel®. FIG. **8**B are representative H&E images of vehicle and oprozomib treated implants. Boxed area is ³⁵ enlarged to the right. Scale bar—50 μm. FIG. **8**C is a graph of the quantification of vascular area normalized to area. *p<0.05. vc—vascular channels.

FIG. 9—Disulfiram efficaciously suppressed VMEC and LMEC growth/viability more than that observed for drugs 40 used off label for vascular malformation patients. Using HTPS, cells were treated with 1 μM of compounds. Cell viability relative to vehicle treated cells was determined after 48 hours. Data presented for disulfiram against sirolimus, alpelisib, and trametinib, all of which have been used 45 off label for vascular malformation patients.

DETAILED DESCRIPTION

Definitions

The terms used in this specification generally have their ordinary meanings in the art, within the context of this invention and the specific context where each term is used. Certain terms are discussed below, or elsewhere in the 55 specification, to provide additional guidance to the practitioner in describing the methods of the invention and how to use them. Moreover, it will be appreciated that the same thing can be said in more than one way. Consequently, alternative language and synonyms may be used for any one 60 or more of the terms discussed herein, nor is any special significance to be placed upon whether or not a term is elaborated or discussed herein. Synonyms for certain terms are provided. A recital of one or more synonyms does not exclude the use of the other synonyms. The use of examples 65 anywhere in the specification, including examples of any terms discussed herein, is illustrative only, and in no way

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limits the scope and meaning of the invention or any exemplified term. Likewise, the invention is not limited to its preferred embodiments.

The term "subject" as used in this application means an animal with an immune system such as avians and mammals. Mammals include canines, felines, rodents, bovine, equines, porcines, ovines, and primates. Avians include, but are not limited to, fowls, songbirds, and raptors. Thus, the invention can be used in veterinary medicine, e.g., to treat companion animals, farm animals, laboratory animals in zoological parks, and animals in the wild. The invention is particularly desirable for human medical applications.

The term "patient" as used in this application means a human subject. In some embodiments of the present invention, the "patient" is suffering with from a vascular malformation, or a disease or condition characterized by a vascular malformation including capillary (CM), lymphatic (LM), venous (VM), and arteriovenous (AVM), and combinations thereof.

The terms "treat", "treatment", and the like refer to a means to slow down, relieve, ameliorate or alleviate at least one of the symptoms of the disease, or reverse the disease after its onset.

The terms "prevent", "prevention", and the like refer to acting prior to overt disease onset, to prevent the disease from developing or minimize the extent of the disease or slow its course of development.

The term "in need thereof" would be a subject known or suspected of having or being at risk of a vascular malformation or a disease or condition characterized by a vascular malformation including capillary (CM), lymphatic (LM), venous (VM), and arteriovenous (AVM), and combinations thereof.

A subject in need of treatment would be one that has already developed the disease or condition. A subject in need of prevention would be one with risk factors of the disease or condition.

The term "agent" as used herein means a substance that produces or is capable of producing an effect and would include, but is not limited to, chemicals, pharmaceuticals, biologics, small organic molecules, antibodies, nucleic acids, peptides, and proteins.

The phrase "therapeutically effective amount" is used herein to mean an amount sufficient to cause an improvement in a clinically significant condition in the subject, or delays or minimizes or mitigates one or more symptoms associated with the disease, or results in a desired beneficial change of physiology in the subject.

The terms "screen" and "screening" and the like as used herein means to test an agent to determine if it has a particular action or efficacy.

The terms "identification", "identify", "identifying" and the like as used herein means to recognize an agent as being effective for a particular use.

The term "about" or "approximately" means within an acceptable error range for the particular value as determined by one of ordinary skill in the art, which will depend in part on how the value is measured or determined, i.e., the limitations of the measurement system, i.e., the degree of precision required for a particular purpose, such as a pharmaceutical formulation. For example, "about" can mean within 1 or more than 1 standard deviations, per the practice in the art. Alternatively, "about" can mean a range of up to 20%, preferably up to 10%, more preferably up to 5%, and more preferably still up to 1% of a given value. Alternatively, particularly with respect to biological systems or processes, the term can mean within an order of magnitude,

preferably within 5-fold, and more preferably within 2-fold, of a value. Where particular values are described in the application and claims, unless otherwise stated, the term "about" meaning within an acceptable error range for the particular value should be assumed.

Abbreviations

LM lymphatic malformation

VM venous malformation

CM capillary malformation

AVM arteriovenous malformation

EC s endothelial cells

LMECs lymphatic malformation endothelial cells

VMECs venous malformation endothelial cells

HMVECs human microvascular endothelial cells (controls)

WES whole exome sequencing

HTPS high throughput screening

PI proteasome inhibitor

Identification of Therapeutic and Prophylactic Agents for Vascular Malformations

Germline mutations have been identified in some vascular malformation subtypes yet screening for germline variants has often failed to identify causative mutations. Recent 25 studies have shown that post-zygotic mutations contribute to a significant portion of AVMs, VMs, and LMs. Somatic mutations are present only in affected tissues, specifically the malformed endothelial cells (ECs).

Endothelial cells from vascular malformation tissues carrying known disease associated pathogenic variants, including those that target the PI3K/AKT/mTOR and RAS/RAF/ MAPK pathways were collected. It was hypothesized that ECs isolated from patients that carried pathogenic somatic, as well as yet to be identified variants, could be used in a 35 drug screen to identify novel therapies for the treatment of vascular malformations. Whole exome sequencing (WES) was used in the patient derived endothelial cells from venous and lymphatic malformation patients to identify mosaic pathogenic variants. These cells were then used in high 40 throughput drug screening (HTPS) to identify new potential therapies for the venous and lymphatic malformation population (Table 3).

Using these endothelial cells carrying variants leading to activation of the PI3K pathway in HTPS, omipalisib, a 45 combined PI3K/mTOR inhibitor, as well as proteasome inhibitors, as a class, and disulfiram, were identified to be efficacious in inhibiting cell survival. In the HTPS and proliferation assays, omipalisib and proteasome inhibitors, as well as disulfiram, had significantly improved efficacy 50 when compared to sirolimus and alpelisib which are the standard of care for patients with vascular malformations. Methods and Compositions for the Prevention and Treatment of Vascular Malformations

disclosure provides for methods of preventing and/or treating vascular malformations including lymphatic, venous, capillary, arteriovenous, and combinations thereof, comprising administering to a subject in need thereof a therapeutically effective amount of an agent which targets a mutation 60 in a gene in the PI3K/AKT/mTOR or RAS/RAF/MAPK pathway.

The current disclosure further provides for methods of preventing and/or treating vascular malformations including lymphatic, venous, capillary, arteriovenous, and combina- 65 tions thereof, comprising administering to a subject in need thereof a therapeutically effective amount of an agent which

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targets a mutation in a gene chosen from the group consisting of Pik3ca, Pik3r3, Tsc2, Rasa1, Map2k2, Glmn, and combinations thereof.

The current disclosure further provides for methods of preventing and/or treating vascular malformations including lymphatic, venous, capillary, arteriovenous, and combinations thereof, comprising administering to a subject in need thereof a therapeutically effective amount of omipalisib.

HTPS determined that omipalisib was the strongest PI3K 10 pathway inhibitor of proliferation/viablity of vascular malformation endothelial cells with PI3K hyperactivation, relative to the 14 tested inhibitors of the PI3K pathway. Omipalisib is a dual PI3K pathway inhibitor which targets the p110 activation domain of PI3K and MTOR, suggesting 15 targeting two protein complexes in the PI3K pathway is more effective than a single target. When another dual PI3K and MTOR inhibitor, bimiralisib, or the clinically used sirolimus were assessed, omipalisib was significantly more efficacious at inhibiting proliferation of LMECs at a lower 20 concentration then either of the other drugs.

Together these data suggest omipalisib is more effective at targeting the pathogenic hyperactivation of PI3K in vascular malformations than current therapies.

Omipalisib has completed Phase I clinical trials for both solid tumors (NCT00972686) and idiopathic pulmonary fibrosis (NCT01725139). It has never been investigated as a candidate for therapy for vascular malformations.

Without being bound by any theory, it is hypothesized that this drug uniquely inhibits the pathological hyperactivation of the PI3K signaling in two different subtypes of vascular malformations, venous and lymphatic malformations. Both vascular malformations have been shown to arise from somatic mutations endothelial cells in genes of the PI3K signaling pathway, including Pik3ca, Pik3r3, Tsc2, and Akt, which lead to increased activity of the PI3K p110 subunit and mTOR proteins targeted by omipalisib. Omipalisib:

formula: $C_{25}H_{17}F_2N_5O_3S$

chemical 2,4-difluoro-N-(2-methoxy-5-(4name: (pyridazin-4-yl)quinolin-6-yl)pyridin yl)benzenesulfonamide.

Dose: 0.5 to 2 mg BID

Effective concentration—74.6 ng/mL (=0.15 μM) (Lukey et al. 2019)

The current disclosure further provides for methods of preventing and/or treating vascular malformations including lymphatic, venous, capillary, arteriovenous, and combinations thereof, comprising administering to a subject in need thereof a therapeutically effective amount of one or more proteasome inhibitors. The disclosure further provides for methods of preventing and/or treating vascular malformations, including lymphatic, venous, capillary, arteriovenous, and combinations thereof, comprising administering to a subject in need thereof a therapeutically effective amount of Based upon the findings set forth herein, the present 55 an agent or compound which has proteasome inhibiting ability.

Proteasome inhibitors that can used in the method include but are not limited to bortezomib, oprozomib, carfilzomib, ixazomib, delanzomib, and marizomib.

Several proteasome inhibitors are already FDA-approved, while several are in different phases of clinical trials. The FDA-approved indication is for the treatment of multiple myeloma, while clinical trials are also investigating their use in solid tumors and non-Hodgkins lymphoma.

Proteasome inhibitors as a class has not been investigated for use in patients with vascular malformations. See Tables 1 and 2.

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Some of the proteasome inhibitors identified target the 20S catalytic subunit in the proteasome complex and in some instances, target all three proteolytic activities of the 20S subunit including the chymotrypsin-like activity (CT-L), trypsin-like activity (T-L) and caspase-like (C-L) activ- 5 ity. In some instance, the PIs targeted the chymotrypsin-like activity. In some instances, the proteasome inhibitors act indirectly to inhibit the proteasome complex.

Mutations in genes that make up the proteasome complex or their involvement in vascular pathologies have not been $_{10}$ reported for vascular malformations.

Without being bound by any theory, proteasomes function in protein degradation via autophagy and ubiquitin-dependent, or ubiquitin-independent proteasome degradation signaling. Thus, proteasome inhibitors may also inhibit proliferation of vascular malformation cells by targeting genetic activation of the PI3K pathway.

Proteasomes have also been shown to modulate angiogenic signaling via regulation of VEGF-A/VEGFR2 signaling, as well as other angiogenic and lymphangiogenic pathways.

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Additionally, disulfiram was also found to inhibit cell proliferation and viability. Disulfiram has shown inhibition of proteasome activity. Thus, the current disclosure further provides for methods of preventing and treating vascular malformations including lymphatic, venous, capillary, arteriovenous, and combinations thereof, comprising administering to a subject in need thereof a therapeutically effective amount of disulfiram.

Copper-binding compounds, such as disulfiram, Clioquinol and diethyldithiocarbamate have also been documented to have proteasome inhibiting ability (Chen et al. 2006; Chen and Dou 2008). Thus, current disclosure further provides for methods of preventing and/or treating vascular pathway and has been implicated as a regulator of PI3K 15 malformations including lymphatic, venous, capillary, arteriovenous, and combinations thereof, comprising administering to a subject in need thereof a therapeutically effective amount of a copper-binding agent or compound. Copperbinding agents or compounds for use in the current methods include but are not limited to NSC-109268, pyrrolidine dithiocarbamate, disulfiram, Clioquinol and diethyldithiocarbamate.

TABLE 1

	FDA Approved Cu	rrently Available Pr	oteasome Inhibitors	
Inhibitor	Mechanism	FDA Approval Status	Dose	Plasma Concentration (ng/mL)
Bortezomib	Selective, reversible inhibitor of chymotrypsin- like activity of the 20S	2005	0.4-1.38 mg/m ²	100-200 (260-520 nM) (Shabaneh et al. 2013)
Carfilzomib	Irreversible epoxyketone inhibitor of the chymotrypsin-like activity of 20S	2012	27-56 mg/m ²	932-2733 (1294-3796 nM) (Brown et al. 2017)
Marizomib	Irreversible inhibitor of all three proteasome protease activities (chymotrypsin-like, trypsin-like and caspase-like activity of 20S)	Orphan drug designation 2015	0.075-0.9 mg/m ²	2.76-57.75 (8.8-184 nM) (Harrison et al. 2016)
Izazomib (oral)	Boronic acid proteasome inhibitor of chymotrypsin-like activity of 20S	2015	1.5-4 mg/d	26.1-61 (72.3-169 nM) (Gupta et al. 2016)
Oprozomib (oral)	Irreversible tripeptide epoxyketone inhibitor of chymotrypsin- like activity of 20S (orally bioavailable analog of	Orphan drug designation 2014	240 mg/day (30-150 mg/d)	218-870 (409-1633 nM) (Infante et al. 2016)
Delanzomib (IV or oral)	Carfilzomib) Inhibitor of the proteasome chymotrypsin-like activity of 20S that decreases NF-kB activity	Clinical trials	0.1-1.8 mg/m ²	88.4-557.3 (214-1348 nM) (Gallerani et al. 2013)

TABLE 2

		Additional Proteason	ne Inhibitors		
Generic Name	Drug Name	Drug Disease	Mechanism of Action	Delivery Route	Notes
4SC-206	4SC-206; SC-68896	Cancer solid, unspecified; brain; myeloma	Proteasome inhibitor	Injectable	Injectable proteasome inhibitor, which was under development for the treatment of solid tumors and inflammation. Indications included multiple myeloma, small cell lung cancer and neurological tumors. Preclinical testing in colon cancer and glioblastoma
ACU-D1	ACCU D1; ACCU-D1; ACCU-D1; ACU-D1; ACU-D1	Acne; dry eye syndrome; psoriasis; rosacea	Proteasome inhibitor Transcription factor NF-kappaB inhibitor	Topical	glioblastoma. Nuclear factor kappa beta (NFkB) inhibitor, which was under development for the treatment of skin and ocular diseases including acne rosacea, psoriasis and acne. Phase II testing for acne rosacea. It was previously under development for dry eye
anticancer therapy, Teva	anticancer therapy, Teva	Cancer, unspecified	Proteasome inhibitor		Teva was developing a series of (2S, 3R)-2-amino- 3-hydroxy- butyric acid derived proteasome inhibitors, for the treatment
AVR-147	AVR-147; CTK- 000147; CTK- 100147	Cancer, unspecified	Proteasome inhibitor		of cancer A novel, natural product small- molecule proteasome inhibitor, which was under development) for the treatment of cancer. It is highly selective for the

	Additional Proteasome Inhibitors					
Generic Name	Drug Name	Drug Disease	Mechanism of Action	Delivery Route	Notes	
BSc-2118	BSc-2118	Cancer, lymphoma; unspecified; myeloma	Proteasome inhibitor; Apoptosis stimulant		chymotrypsin- like activity of the proteasome. A novel tripeptide with inhibitory activity against the 3 proteolytic activities of the 20S proteasome, which was under development for the treatment of multiple myeloma (MM) and	
CEP-28331	CEP-28331	Cancer, myeloma	Proteasome inhibitor	Oral; Oral, swallowed	mantle cell lymphoma (MCL). Oral proteasome inhibitor,	
					which was under development for the treatment of multiple	
CVT-857	CVT-710; CVT-857	Inflammatory disease, unspecified	Proteasome inhibitor		myeloma CVT-857 and CVT-710 are alpha- ketoamide- terminated retropeptides, which were under investigation as potential anti- inflammatories. They selectively inhibit the chymotrypsin- like site of the 20S proteasome complex without inhibiting either the peptidylglutamyl- or trypsin-like proteolytic activities of the 20S complex, suggesting that the active site, threonine, in the 20S proteasome is not undergoing nucleophilic addition to	

		Additional Proteasom	e Inhibitors		
Generic Name	Drug Name	Drug Disease	Mechanism of Action	Delivery Route	Notes
CX13-608	CX13 608; CX13608; CX13- 608	Cancer, myeloma; lymphoma; unspecified	Proteasome inhibitor	Injectable, intravenous	the electrophilic carbonyl of the alphaketomide group. A proteasome inhibitor, under development for the treatment of
CYS-006	anticancers, Cytomics Systems; CYS-006	Cancer, unspecified	Proteasome inhibitor		multiple myeloma. Phase I and Phase II testing. CYS-006 was a series of compounds
					targeting the ubiquitin- proteasome protein degradation pathway, under development as proteasome inhibitors to stabilize the degradation of a protein that has been
disulfiram +	CX 02; CX02; CX-		Chemokine	Oral;	implicated in the pathogenesis of malignancies. Dicopp (CX-
copper, Cantex	02; Dicopp; disulfiram + copper gluconate, Cantex; disulfiram + copper, Cantex	myeloma; sarcoma; unspecified brain; breast; lung, non-small cell; prostate, Poisoning, radiation	receptor antagonist; Proteasome inhibitor	Oral, swallowed	o2) is an oral, single-dose combination of disulfiram + copper, under development for the treatment of pancreatic cancer, pediatric sarcoma and myeloma. It was previously under development for brain, prostate, lung and breast cancers.
ER-807446	ER-804191; ER- 807446	Cancer, unspecified	Proteasome inhibitor		Phase I and Phase II testing of some cancers. A representative compound in a series of epoxyketone derivatives based on the

TABLE 2-continued					
		Additional Proteason	ne Inhibitors		
Generic Name	Drug Name	Drug Disease	Mechanism of Action	Delivery Route	Notes
FV-162	anticancer therapy, Fluorinov Pharma; FV162; FV-162	Cancer, lymphoma, non-Hodgkin's Cancer, myeloma	Proteasome inhibitor	Oral; Oral, swallowed	natural product eponemycin, which was under investigation as proteasome inhibitors for the treatment of cancer. A potent, orally- delivered small molecule proteasome inhibitor, was under development
FV-214	FV214; FV-214	Cancer, myeloma Waldenstrom's hypergamma- globulinaemia	Proteasome inhibitor	Injectable, intravenous	development for the treatment of multiple myeloma. It is a novel fluorine-based therapy. Fluorine atoms are introduced in to drug candidates to modify the specific physical and chemical properties of the parent drug molecules. It works by selectively inhibiting the chymotrypsin-like enzymatic activity of the proteasome. Phase I testing in myeloma. A potent, intravenously (iv) delivered small molecule proteasome inhibitor, which was under development for the
					multiple myeloma, Waldenstrom's macroglobulin- aemia. It works by selectively inhibiting the chymotrypsin- like

enzymatic

		Additional Protease	me Inhibitors		
Generic Name	Drug Name	Drug Disease	Mechanism of Action	Delivery Route	Notes
G4-1	G4 1; G4-1	Cancer, solid; unspecified	Proteasome inhibitor		activity of the proteasome. A novel proteasome inhibitor, under development for the treatment of
GNF-6702	GNF6702; GNF-6702	Infection, leishmaniasis; trypanosomiasis	Proteasome inhibitor		solid tumors. A selective inhibitor of the kinetoplastid proteasome which was under development for the treatment of leishmaniasis, Chagas disease and sleeping sickness.
GSK439			Proteasome inhibitor		bieiniebb.
GSK- 3494245	DDD 1305143; DDD1305143; DDD-1305143; GSK 3494245; GSK3494245; GSK-3494245; GSK3494245/DDD 1305143, DNDi; GSK3494245/DDD 1305143, GlaxoSmithKline	Infection, leishmaniasis	Proteasome inhibitor	Oral; Oral, swallowed	Under development for the treatment of Leishmaniasis. Phase I testing.
immuno- proteasome inhibitors, Principia BioPharma	immunoproteasome inhibitors, AbbVie; immunoproteasome inhibitors, Principia BioPharma	Autoimmune disease, Inflammatory disease	Proteasome inhibitor	Oral; Oral, swallowed	oral immuno- proteasome inhibitors, for the treatment of inflammatory disease and autoimmune disorders such as inflammatory bowel diseases (IBD), lupus and psoriasis. It was previously under development for the treatment of immunological disease. Some are LMP2 and LMP7 inhibitors (Basler et al.
KZR-616	KZR616; KZR-616; KZR616 (IV); KZR- 616 (IV); KZR616 (SC); KZR-616 (SC); ONX 0914; ONX-0914; ONYX0914;		Proteasome inhibitor	Injectable; Injectable, intravenous; Injectable, subcutaneous	2018). Selective small molecule inhibitor of the immuno- proteasome, under

		Additional Proteasom	e Inhibitors		
Generic Name	Drug Name	Drug Disease	Mechanism of Action	Delivery Route	Notes
	ONYX-0914; PR957; PR-957				development for the treatment of autoimmune diseases and inflammatory diseases. Phase I and Phase II testing.
LGP-07154 LMP7 inhibitor, Shouyao Holdings	LGP07154; LGP- 07154; VAL-789- CHUM LMP7 inhibitor, Shouyao Holdings	Cancer, unspecified Immunological disease, unspecified Cancer, hematological; unspecified, Autoimmune disease, unspecified, unspecified, unspecified,	Proteasome inhibitor unspecified proteasome subunit beta type-8 inhibitor	Oral; Oral, swallowed Oral; Oral, swallowed	costing.
LODO-141	TIR199, TIR 199, LODO141, LODO 141,TIR-199	Inflammatory disease Solid Tumors			LODO-141 (TIR-199) is an
LXE-408	LXE 408; LXE408; LXE-408	Infection, leishmaniasistry- panosomiasis, American	Proteasome inhibitor	Oral; Oral, swallowed	irreversible and potent hybrid cyclic peptide proteasome inhibitor from the syrbactin natural product family. Proteasome inhibitor, under development for the treatment of visceral leishmaniasis. It was previously under
M-3258	M 3258; M3258; M-3258	Cancer, myeloma	Proteasome subunit beta type-8 inhibitor	Oral; Oral, swallowed	development for Chagas disease. Phase I and Phase II testing. Immuno-proteasome subunit LMP7 inhibitor
MCIT-375	Anti- CD38/bortezomib/ lenalidomide Dual Drug Pay load ADC; MCIT375; MCIT- 375	Cancer, myeloma	Proteasome inhibitor; Cereblon E3 ubiquitin ligase stimulant; Angiogenesis inhibitor; Immuno-	Injectable	under development for the treatment of multiple myeloma. Phase 1 testing. An anti- CD38 antibody drug conjugate with dual drug payloads bortezomib and lenalidomide,

		Additional Proteason	ne Inhibitors		
Generic Name	Drug Name	Drug Disease	Mechanism of Action	Delivery Route	Notes
			oncology therapy		which was under development for the treatment of multiple
MLN519		Inflammatory Disease and Stroke	Ubiquitin Proteasome		myeloma. Phase I and Phase II
OSH-101	OSH-101	Alopecia	inhibitor Proteasome inhibitor	Topical; Topical, skin	testing A tetrapeptide aldehyde
					proteasome inhibitor, which was under development for the treatment of male and female pattern baldness and chemotherapy- induced alopecia. Other possible indications include alopecia areata, age- related hair thinning and post- transplant hair regrowth. Phase I testing complete, Phase II planned.
peptide epoxyketones, Onyx Pharmaceuticals	Pharmaceuticals	Inflammatory disease, unspecified	Proteasome inhibitor		Subunit selective peptide epoxyketones as proteasome inhibitors, for the treatment of inflammatory
proteasome inhibitor, Flaveome	proteasome inhibitor, Flaveome	Cancer, unspecified	Proteasome inhibitor	Oral; Oral, swallowed	disease. A next generation orally- bioavailable proteasome inhibitor for the treatment of cancer. It also inhibits PI3K/Akt signalling. They are small molecules derived from flavonoid structures.
proteasome inhibitors, EntreMe	proteasome inhibitors, EntreMe	Cancer, unspecified	Proteasome inhibitor		Small- molecule proteasome

Additional Proteasome Inhibitors						
Generic Name	Drug Name	Drug Disease	Mechanism of Action	Delivery Route	Notes	
proteasome inhibitors, Ergon Pharmaceuticals	proteasome inhibitors, Ergon Pharmaceuticals	Cancer, unspecified	Proteasome inhibitor		inhibitors, for the treatment of cancer. Novel 26S proteasome inhibitors, for the treatment	
proteasome inhibitors, Jeil	proteasome inhibitors, Jeil	Cancer, unspecified	Proteasome inhibitor		of cancer.	
proteasome inhibitors, NYU	proteasome inhibitors, NYU	Infection, malaria	Proteasome inhibitor		Inhibitors of enzymes of the ubiquitin-proteasome pathway, different from those of the PS-341 series (qv), for the treatment of malaria and other parasitic diseases. The pathway is involved in proteolysis which occurs during Plasmodium falciparum's lifecycle (Scrip, 1997, 2239, 19).	
proteasome inhibitors, QLi5 Therapeutics	proteasome inhibitors, QLi5 Therapeutics	Autoimmune disease, unspecified Cancer, unspecified Inflammatory disease, unspecified	Proteasome inhibitor		Peptide mimetic proteasome inhibitors for the treatment of cancer and inflammation.	
proteasome inhibitors, Signature	peptide mimetics, Signature; proteasome inhibitors, Signature	Inflammatory disease, unspecified	Proteasome inhibitor			
proteasome inhibitors, Takeda	proteasome inhibitors, Millenium; proteasome inhibitors, Takeda	Cancer, unspecified	Proteasome inhibitor		A series of human 20S proteasome non-covalent inhibitors derived from N-β-neopentyl asparagine, for the treatment of cancer.	
proteasome inhibitors, Telik	proteasome inhibitors, Telik	Cancer, unspecified	Proteasome inhibitor		Amall- molecule, orally-active, non-peptide and non- boron-based proteasome inhibitors, for the treatment of cancer.	
proteasome inhibitors, Novartis	proteasome inhibitors, Novartis	Cancer, unspecified	Proteasome inhibitor; Apoptosis stimulant			
proteasome inihibitors,	proteasome inihibitors,	Cancer, hematological;	Proteasome Inhibitor			

TABLE 2-continued

TABLE 2-continued								
Additional Proteasome Inhibitors								
Generic Name	Drug Name	Drug Disease	Mechanism of Action	Delivery Route	Notes			
IkerChem syrbactins, Pono Pharma	IkerChem syrbactins, Pono Pharma	unspecified; solid Cancer, myeloma; unspecified	Proteasome inhibitor					
Pharma timosaponin AIII	AA-101; AA102; AA-102; BN-108; timosaponin AIII	Cancer, breast	Apoptosis stimulant; Caspase stimulant; Proteasome inhibitor	Oral; Oral, swallowed	Oral plant-based aqueous extract from Anemarrhena asphodeloides bunge, which attenuates mitochondrial membrane potential to cause cytochrome C release and caspase activation, inducing low molecular grade apoptosis, which was under development for the treatment of breast cancer. It induces cancer cell death via activation of the AKT and mTOR pathways in breast cancer cells, but not normal cells. Phase II			
TMC-86A	proteasome inhibs, Tanabe; TMC-86A; TMC-86B; TMC-96	Immunological disease, unspecified Inflammatory disease, unspecified	Proteasome inhibitor		testing. TMC-86A, TMC-86B and TMC-96 are 20S proteasome inhibitors with epoxy- β-aminoketone moieties, which were under investigation for the treatment of inflammatory disease, autoimmune diseases and muscle wasting associated with cancer cachexia, diabetes and sepsis. The compounds were isolated from Streptomyces sp TC-1084 and Saccharothrix			

TABLE 2-continued

Additional Proteasome Inhibitors							
Generic Name	Drug Name	Drug Disease	Mechanism of Action	Delivery Route	Notes		
TMC-95A	TMC-95A; TMC-95B; TMC-95D	Immunological disease, unspecified Inflammatory disease, unspecified Musculoskeletal disease, unspecified	Proteasome inhibitor		sp TC-1094 (J Antibiot, 1999, 52, 1069). TMC-95A and its diastereomers, TMC-95B to D are 20S proteasome inhibitors, which were under investigation for the treatment of inflammatory and autoimmune diseases, and rapid muscle wasting associated with cancer cachexia, diabetes and sepsis. This series of cyclic peptides was isolated from Apiospora montagnei Sacc, TC- 1093.		
TQB-3602	TQB 3602; TQB3602; TQB- 3602	Cancer, myeloma	Proteasome inhibitor		TQB-3602 is a proteasome inhibitor, under development for the treatment of multiple myeloma. Phase I testing.		
Ubiquitin Proteasome Program VL-01	ubiquitin proteasome system	Cancer	Proteasome inhibitor Proteasome Inhibitor that induces apopotosis				
	4SC-206, SC-68896, SC68896	Hepatitis C (HCV); HIV (Antiviral)					
VLX157	VLX1570	Cancer-bone, mantle cell lymphoma, multiple myeloma	Proteasome inhibitor				

All of the agents discussed herein can be in the form of pharmaceutical compositions.

Most preferred methods of administration of the agents and compositions for use in the disclosed methods are oral and parental including intravenous and injection. The pharmacological agent must be in the appropriate form for administration of choice.

Such pharmaceutical compositions comprising one or more pharmacological agents for administration may comprise a therapeutically effective amount of the pharmacological agent and a pharmaceutically acceptable carrier.

The phrase "pharmaceutically acceptable" as used herein refers to molecular entities and compositions that are physi-

ologically tolerable and do not typically produce an allergic or similar untoward reaction, such as gastric upset, dizziness and the like, when administered to a human, and approved by a regulatory agency of the Federal or a state government or listed in the U.S. Pharmacopeia or other generally recognized pharmacopeia for use in animals, and more particularly in humans.

The term "carrier" refers to a diluent, adjuvant, excipient, or vehicle with which the therapeutic is administered. Such pharmaceutical carriers can be sterile liquids, such as saline solutions in water and oils, including those of petroleum, animal, vegetable, or synthetic origin, such as peanut oil,

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soybean oil, mineral oil, sesame oil, and the like. A saline solution is a preferred carrier when the pharmaceutical composition is administered intravenously. Saline solutions and aqueous dextrose and glycerol solutions can also be employed as liquid carriers, particularly for injectable solu- 5 tions. Suitable pharmaceutical excipients include starch, glucose, lactose, sucrose, gelatin, malt, rice, flour, chalk, silica gel, sodium stearate, glycerol monostearate, talc, sodium chloride, dried skim milk, glycerol, propylene, glycol, water, ethanol, and the like. The composition, if desired, 10 can also contain minor amounts of wetting or emulsifying agents, or pH buffering agents.

Pharmaceutical compositions adapted for oral administration may be capsules, tablets, powders, granules, solutions, syrups, suspensions (in non-aqueous or aqueous liq- 15 uids), or emulsions. Tablets or hard gelatin capsules may comprise lactose, starch or derivatives thereof, magnesium stearate, sodium saccharine, cellulose, magnesium carbonate, stearic acid or salts thereof. Soft gelatin capsules may comprise vegetable oils, waxes, fats, semi-solid, or liquid 20 polyols. Solutions and syrups may comprise water, polyols, and sugars. An active agent intended for oral administration may be coated with or admixed with a material that delays disintegration and/or absorption of the active agent in the gastrointestinal tract. Thus, the sustained release may be 25 achieved over many hours and if necessary, the active agent can be protected from degradation within the stomach. Pharmaceutical compositions for oral administration may be formulated to facilitate release of an active agent at a particular gastrointestinal location due to specific pH or 30 enzymatic conditions.

A further preferred form of administration is parenteral including intravenous administration. Pharmaceutical compositions adapted for parenteral administration, including intravenous administration, include aqueous and non-aque- 35 ous sterile injectable solutions or suspensions, which may contain anti-oxidants, buffers, bacteriostats, and solutes that render the compositions substantially isotonic with the blood of the subject. Other components which may be present in such compositions include water, alcohols, poly-40 ols, glycerine, and vegetable oils. Compositions adapted for parental administration may be presented in unit-dose or multi-dose containers, such as sealed ampules and vials, and may be stored in a freeze-dried (lyophilized) condition requiring only the addition of a sterile carrier, immediately 45 prior to use. Extemporaneous injection solutions and suspensions may be prepared from sterile powders, granules, and tablets. Suitable vehicles that can be used to provide parenteral dosage forms of the invention are well known to those skilled in the art. Examples include: Water for Injec- 50 tion USP; aqueous vehicles such as Sodium Chloride Injection, Ringer's Injection, Dextrose Injection, Dextrose and Sodium Chloride Injection, and Lactated Ringer's Injection; water-miscible vehicles such as ethyl alcohol, polyethylene glycol, and polypropylene glycol; and non-aqueous vehicles 55 such as corn oil, cottonseed oil, peanut oil, sesame oil, ethyl oleate, isopropyl myristate, and benzyl benzoate.

Pharmaceutical compositions adapted for nasal and pulmonary administration may comprise solid carriers such as powders, which can be administered by rapid inhalation 60 Kits through the nose. Compositions for nasal administration may comprise liquid carriers, such as sprays or drops. Alternatively, inhalation directly through into the lungs may be accomplished by inhalation deeply or installation through a mouthpiece. These compositions may comprise aqueous or 65 oil solutions of the active ingredient. Compositions for inhalation may be supplied in specially adapted devices

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including, but not limited to, pressurized aerosols, nebulizers or insufflators, which can be constructed so as to provide predetermined dosages of the active ingredient.

Further methods of administration include sublingual, vaginal, buccal, rectal, or transdermal administration to a subject.

In some instances, the agent is in a next generation formulation such as nanoencapsulation, such as bortezomib. Such formulations are under investigation for cancer treatment.

Selection of a therapeutically effective dose will be determined by the skilled artisan considering several factors, which will be known to one of ordinary skill in the art. Such factors include the particular form of the pharmacological agent, and its pharmacokinetic parameters such as bioavailability, metabolism, and half-life, which will have been established during the usual development procedures typically employed in obtaining regulatory approval for a pharmaceutical compound. Further factors in considering the dose include the condition or disease to be treated or the benefit to be achieved in a normal individual, the body mass of the patient, the route of administration, whether the administration is acute or chronic, concomitant medications, and other factors well known to affect the efficacy of administered pharmaceutical agents. Thus, the precise dose should be decided according to the judgment of the person of skill in the art, and each patient's circumstances, and according to standard clinical techniques.

Doses can be adjusted to optimize the effects in the subject. For example, the agent can be administered at a low dose to start and then increased over time to depending upon the subject's response. A subject can be monitored for improvement of their condition prior to changing, i.e., increasing or decreasing, the dosage. A subject can also be monitored for adverse effects prior to changing the dosage, i.e., increasing or decreasing, the dosage.

The FDA approved PIs and dosages are found in Table 1. It will be appreciated by those of skill in the art that the subjects on which the methods of the present disclosure are being practiced are under 10 years of age and in some cases under one year of age. Thus, the therapeutically effective dosages for these subjects may be less than the FDA approved dosages.

The agents may be administered daily, weekly, biweekly, several times daily, semi-weekly, every other day, bi-weekly, quarterly, several times per week, semi-weekly, monthly, or more. The duration and frequency of treatment may depend upon the subject's response to treatment.

The agents described herein can be co-administered with other agents including those disclosed herein and additional ones for the prevention and treatment of vascular malformations. The co-administration of agents can be by any administration described herein. Moreover, it can be in one composition, or in more than one composition. The administration of the agents can be simultaneous, concurrently or sequentially.

Treatment using the present methods can continue as long as needed.

Also within the scope of the present disclosure are kits for practicing the disclosed methods.

In some embodiments, the kit can comprise instructions for use in any of the methods described herein. The included instructions can comprise a description of administration of the agents to a subject to achieve the intended activity in a subject. The kit may further comprise a description of 33

selecting a subject suitable for treatment based on identifying whether the subject is in need of the treatment.

The instructions relating to the use of the agents described herein generally include information as to dosage, dosing schedule, and route of administration for the intended treatment. The containers may be unit doses, bulk packages (e.g., multi-dose packages) or sub-unit doses. Instructions supplied in the kits of the disclosure are typically written instructions on a label or package insert. The label or package insert indicates that the pharmaceutical compositions are used for treating, delaying the onset, and/or alleviating a disease or disorder in a subject.

The kits provided herein are in suitable packaging. Suitable packaging includes, but is not limited to, vials, bottles, jars, flexible packaging, and the like. Also contemplated are packages for use in combination with a specific device, such as an inhaler, nasal administration device, or an infusion device. A kit may have a sterile access port (for example, the container may be an intravenous solution bag or a vial having a stopper pierceable by a hypodermic injection 20 needle). The container may also have a sterile access port.

Kits optionally may provide additional components such as buffers and interpretive information. Normally, the kit comprises a container and a label or package insert(s) on or associated with the container. In some embodiment, the ²⁵ disclosure provides articles of manufacture comprising contents of the kits described above.

EXAMPLES

The present invention may be better understood by reference to the following non-limiting examples, which are presented in order to more fully illustrate the preferred embodiments of the invention. They should in no way be construed to limit the broad scope of the invention.

Example 1—Direct Targeting of Affected Pathways with Known Drugs

LMECs and matching tissue with an activating mutation 40 in PIK3CA or an inactivating mutation in RASA1 had increased AKT and ERK activation when compared to control tissue and cells, respectively. LMECs carry inactivating mutations in PIK3R3/TSC2 activation unexpectedly displayed increased ERK activation as well as AKT activation. Using these cells, it was earlier found that the LMECs with MAPK pathway activation were resistant to sirolimus, a mTOR inhibitor commonly used to treat LMs. See FIG. 1A.

It was then asked whether this finding was specific to 50 sirolimus, or to inhibitors that target the PI3K/AKT/MTOR pathway. The effect of inhibiting the MAPK (RAS/MAPK) pathway on proliferation of these cells was also assessed.

Cells with the same mutations (Pik3ca, Pik3r3/Tsc2, Rasa1; LMEC8, LMEC28, and LMEC10 respectively), 55 (Table 3) were treated with increasing log doses of alpelisib, a PI3K inhibitor, ranging from 1 µM to 100 µM. Proliferation rate was determined by WST-8 assay and expressed as a fraction of the number of cells in vehicle controls.

It was found that alpelisib significantly inhibited both 60 control and the LMECs at concentrations flanking the calculated human sera level of 75 μ M; it was more effective than sirolimus. However, at lower doses, LMEC responses were different.

Cells with only PIK3CA mutation (and only increased 65 AKT signaling) were more sensitive to alpelisib, while the cells with increased ERK signaling were more resistant

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when compared to control HDLECs (FIG. 1B). These results were similar to sirolimus (FIG. 1A).

Cells were also treated with U0126, a MAPK inhibitor, in increasing log doses ranging from 1 μ M to 100 μ M. At the concentration used in animal models (10 μ M), U0126 suppressed the growth of all cells by 50% (FIG. 1C).

Interestingly, cells with the RASA1 mutations were not any more sensitive to MAPK inhibition than those with the PIK3CA mutation or control cells. These data demonstrated that drug responses were affected by genetic variants, and MAPK hyperactivation may contribute to pathogenic proliferation of LMECs.

Example 2—Unbiased High Throughput Screening (HTS) of ECs Identified Omipalisib and Proteasome Inhibitors as Effective

ECs from vascular malformation tissues carrying known disease associated pathogenic variants, including those that target the PI3K/AKT/mTOR and RAS/RAF/MAPK pathways have been collected. It was hypothesized that ECs isolated from patients that carried pathogenic somatic, as well as yet to be identified variants, could be used in a drug screen to identify novel therapies for the treatment of vascular malformations.

High throughput screening of drugs was performed using ECs from either LMs (LMECs) or VM (VMECs) carrying pathogenic variants in genes of the PI3K/AKT/mTOR path-30 way or variants predicted to hyperactivate the RAS/RAF/MAPK pathway (Table 3).

Per optimization studies, about 500 cells were seeded into the wells of a 384-well plate and the VMECs or LMECs were treated with 1 µM of approximately 2500 drugs, which were either in clinical trials or FDA approved for another condition. The effects of the drugs on the proliferation and cell viability of the cells were assessed. Cell viability relative to vehicle treated cells was determined after 48 hours.

HTPS identified several candidate drugs that inhibited the PI3K/AKT signaling pathways (FIG. 2). Interestingly, the inhibitory effects on of the same class of drugs on all VMEC and LMEC proliferation were variable across all cell populations. As an example, omipalisib, a dual PI3K/mTOR inhibitor, was more efficacious in inhibiting vascular malformation cells than voxtalisib, another PI3K/mTOR dual inhibitor. As omipalisib was found as the strongest PI3K/mTOR dual inhibitor of LMEC and VMEC proliferation in the HTPS screen, another dual PI3K/mTOR inhibitor, bimiralisib, was assessed as it was not in the HTPS panel (FIG. 3). Omipalisib was significantly more efficacious than bimiralisib at every dose in inhibiting an LMEC population, suggesting omipalisib may be the optimal drug in its class to target the pathological growth in LMs and VMs.

A novel class of therapeutics, proteasome inhibitors (PIs), were identified in the HTPS that were more efficacious at suppressing cell growth/viability than drug therapies currently used off-label for VM and LM patients, sirolimus and alpelisib (FIG. 4; Table 4). The 3 PIs tested in the HTPS suppressed LMEC (n=5) and VMEC (n=3) cell numbers 75%-90% relative to vehicle treated cells, while sirolimus and alpelisib reduced their growth and viability only 40-50%. The PIs had a much stronger effect on the LMECs and VMECs regardless of the mutation. As can be seen in Table 4, the 3 PIs tested in the HTPS were: number 1 in effectiveness for all cell variants (carfilzomib); number 2 for 5% cell variants and number 3 and 4 for the 3% (defanzomib);

and in the top 9 for all cell variants (ixazomib). In contrast, sirolimus and alpelisib ranged from number 28-383 for effectiveness for all variants.

TABLE 3

LMECs and VMECs Used in HTPS Screening					
Cell Population	Pathogenic Variant				
LMEC8 LMEC10 LMEC28 LMEC94 LMEC99	Pik3ca c.1636 C > A Rasa ^{frameshift} c.617_621delTAAGA Pik3r3 ^{Sp-Donor} 1:46527598 T/TA; Tsc2 ⁵ 'utr 16-2110805-G-A Pik3ca c.3140A > G Pik3ca c.1258T > C				
VMEC7 VMEC9 VMEC13	Pik3ca c.1035T > A Pik3ca c.1624G > A Map2k2c.1112g > A; $Glmn^{Sp-Donor}$ c.1214+2T > C				

Example 5—In Vivo Effectiveness of Proteasome Inhibitors on Treating VMs

To assess the efficacy of PIs in an animal, the effect of oprozomib was assessed in a xenograft model. VMECs carrying Pik3ca variants (n=2 VMECs; VMEC7 and VMEC9—Table 3; 4 implants each for treated and vehicle) were resuspended in Matrigel® (Corning) and subcutaneously implanted in the flanks of immunocompromised mice.

VM vessels were allowed to develop and mice randomized for one week. After 1 week, mice were given either 30 mg/kg oprozomib or vehicle orally two days a week for 4 weeks. Oprozomib, a second generation of carfilzomib, was chosen as it is reported to be more tolerable, and orally available (Teicher and Tomaszewski 2017). After 4 weeks of treatment, mice were sacrificed, and the implants surgically removed. The vehicle-treated implant was vascular and easily distinguished from surrounding soft tissues. The opro-

TABLE 4

	Ranking of Proteasome Inhibitors in HTPS for Suppressing Cell Proliferation and Growth Compared to Drugs used for Off-Label Treatment						
Cell Pop.	Carfitzomib	Delanzomib		Disulfiram		Alpelisib	Trematinib
VMEC 13	1	2	8	8	118	107	79
LMEC 10	1	3	5	3	383	50	1225
VMEC 7	1	4	7	10	32	44	71
VMEC 9	1	2	9	5	52	58	71
LMEC 8	1	3	8	7	38	42	72
LMEC 28	1	2	9	19	63	81	241
LMEC 94	1	2	6	11	45	28	62
LMEC 99	1	2	4	8	40	83	82

Example 3—Dose Response Studies of Proteasome Inhibitors and Omipalisib

Using the plasma concentrations reported in the literature for six commercially available PIs (Table 1) and omipalisib (Lukey et al. 2019), a dose response study was performed using VMEC7, VMEC9, LMEC94 and LMEC99, all carrying Pik3ca variants (Table 3) as well as HMVECs. Cell growth and viability was measured as in Example 2 after 48 hours. The 6 PIs and omipalisib were all more effective at suppressing growth/viability of LMEC and VMEC carrying Pik3ca variants, than either sirolimus or alpelisib at clinically relevant doses (as shown by the arrows in FIG. 5) (FIGS. 5 and 6).

Example 4—Effectiveness of Proteasome Inhibitors is not Limited to VM/LM ECs with Pik3ca Variants

LMEC28 cells which carries the Pik3r3;Tsc2 mutations 60 were subjected to increasing amounts of oprozomib from 1 nM to 1 μ M and cell viability determined after 48 hours as set forth in Example 2.

As shown in FIG. 7, there was a dose dependent inhibition of the LMECs carrying the Pik3r3; Tsc2 showing that the PI 65 was effective in suppressing cell proliferation in cells harboring mutations other than Pik3ca.

zomib-treated implant had no vascularity on gross inspection. The outline of the Matrigel® was visualized as a translucent, oval-shaped foreign body in the subcutaneous space. The implants were then fixed by formalin, paraffin embedded for histological analysis. Five (5) micron sections were stained with Hematoxylin and Eosin and images captured and used to assess implant morphology and vascularity.

Findings were dramatic. The implants in the vehicle treated group were highly vascularized and hemorrhagic. In contrast, the vessels appeared to be regressed or normalized in the implants from the treatment group. See FIG. 8. Additionally, no adverse effects were seen in the treated mice.

Taken together, these results demonstrate that PIs are efficacious against pathological VMECs and LMECs carrying genetic variants in the two most commonly mutated pathways in vascular malformations, PI3K/AKT/mTOR and RAS/RAF/MAPK pathways.

Example 6—Unbiased High Throughput Screening (HTS) of ECs Additionally Identified Disulfiram as Effective

Using the HTPS described in Example 2, disulfiram was also shown to efficaciously suppress VMEC and LMEC

growth/viability more than that observed for drugs used off label for vascular malformation patients. See FIG. 9 and Table 4.

As can be seen in Table 4, the disulfiram tested in the HTPS ranged from number 3 to number 19 for effectiveness in the variants tested. In contrast, sirolimus and alpelisib ranged from number 28-383 for effectiveness for all variants.

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The invention claimed is:

- 1. A method of treating a vascular malformation in a subject in need thereof, the method comprising administering to the subject a therapeutically effective amount of disulfiram, wherein the vascular malformation is chosen from the group consisting of lymphatic, venous, capillary, and combinations thereof.
- 2. The method of claim 1, wherein the disulfiram is administered to the subject by oral administration, parental administration, topical administration, or injection.
- 3. The method of claim 1, wherein the subject is under 10 years old.
- 4. The method of claim 1, wherein the subject is under 5 years old.
- 5. The method of claim 1, wherein the subject is under 1 year old.
- 6. The method of claim 1, further comprising, prior to administration, obtaining endothelial cells from the vascular malformation of the subject and testing the efficacy and/or dosage of the disulfiram on endothelial cells obtained from the subject.

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