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Tuschl et al.

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(54) **RNA-INTERFERENCE BY SINGLE-STRANDED RNA MOLECULES**

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C12N 15/113 (2010.01)
C07K 14/47 (2006.01)
C07K 19/00 (2006.01)
A61K 47/54 (2017.01)

(52) **U.S. Cl.**

CPC **C12N 15/113** (2013.01); **A61K 47/549** (2017.08); **A61K 47/557** (2017.08); **C07K 14/4705** (2013.01); **C07K 19/00** (2013.01); **C12N 15/111** (2013.01); **C12N 2310/14** (2013.01); **C12N 2310/351** (2013.01); **C12N 2310/3513** (2013.01); **C12N 2320/30** (2013.01); **C12N 2320/51** (2013.01); **C12N 2330/30** (2013.01)

(58) **Field of Classification Search**

None
See application file for complete search history.

(56) **References Cited**

U.S. PATENT DOCUMENTS

4,469,863 A 9/1984 Ts'o et al.
6,379,931 B1* 4/2002 Rossi C12N 15/113
435/6.14
2002/0150945 A1 10/2002 Finney et al.
2002/0162126 A1* 10/2002 Beach C12N 15/102
800/8
2003/0125281 A1* 7/2003 Lewis A61K 31/58
514/44 A
2003/0153521 A1 8/2003 McSwiggen
2003/0170891 A1 9/2003 McSwiggen
2004/0014108 A1 1/2004 Eldrup et al.
2004/0203145 A1 10/2004 Zamore et al.

OTHER PUBLICATIONS

Kuramochi-Miyagawa et al., Two mouse piwi-related genes: miwi and mili, 2001, Mechanisms of Development, vol. 108, pp. 121-133.*
Agrawal et al., "Antisense oligonucleotides as antiviral agents", Trends in Biotechnol., vol. 10, May 1992, pp. 152-158.
Chiu et al., RNAi in human cells: Basic structural and functional features of small interfering RNA, 2002, Molecular Cell, vol. 10, pp. 549-561.
Hamada et al., Effects on RNA interference in gene expression (RNAi) in cultured mammalian cells of mismatches and the introduction of chemical modifications at the 3' ends of siRNAs, 2002, Antisense and Nucleic Acid Drug Development, vol. 12, pp. 301-309.
Chiu et al., siRNA function in RNAi: A chemical modification analysis, 2003, RNA, vol. 9, pp. 1034-1048.
Hamilton et al., A species of small antisense RNA in post-transcriptional gene silencing in plants, 1999, Science, vol. 286, pp. 950-952.
Vaucheret et al., Post-transcriptional gene silencing in plants, Journal of Cell Science, vol. 114, pp. 3083-3091, 2001.
Bernstein et al., Role for a bidentate ribonuclease in the initiation step of RNA interference, Jan. 18, 2001, Nature, vol. 409, pp. 363-366.
Elbashir et al., Duplexes of 21-nucleotide RNAs mediate RNA interference in cultured mammalian cells, May 2001, Nature vol. 411, pp. 494-498.
Opalinska et al., Nucleic-acid therapeutics: Basic principles and recent applications, Jul. 2002, Nature Reviews Drug Discovery, vol. 1, pp. 503-514.
Charlie Schmidt, Negotiating the RNAi patent thicket, Mar. 2007, Nature Biotechnology, vol. 25, pp. 273-275.
Bernstein et al., "The rest is silence", RNA, 2001, vol. 7, pp. 1509-1521.

(Continued)

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(57) **ABSTRACT**

The present invention relates to sequence and structural features of single-stranded (ss)RNA molecules required to mediate target-specific nucleic acid modifications by RNA-interference (RNAi), such as target mRNA degradation and/or DNA methylation.

16 Claims, 26 Drawing Sheets

(56)

References Cited

OTHER PUBLICATIONS

Caplen et al., "Rescue of polyglutamine-mediated cytotoxicity by double-stranded RNA-mediated RNA interference", *Human Molecular Genetics*, 2002, vol. 11, No. 2, pp. 175-184.

Canadian Office Action No. 2,489,174 dated May 28, 2010 (5 pages).

Schmitz et al., Effect of s'-0-methyl antisense ORNs on expression of thymidylate synthase in human colon cancer RKO cells, 2001, *Nucleic Acids Research*, vol. 29, pp. 415-422.

Tijsterman Marcel et al., "RNA helicase MUT-14-dependent gene silencing triggered in *C.elegans* by short antisense RNAs", *Science* vol. 295, No. 5555, Jan. 25, 2002, pp. 694-697.

Boutla A et al., "Short 5'-phosphorylated double-stranded RNAs induce RNA interference in *Drosophila*", *Current Biology*, *Current Science*, vol. 11, No. 22, Nov. 13, 2001, pp. 1776-1780.

Elbashir S M et al., "Analysis of gene function in somatic mammalian cells using small interfering RNAs", *Methods: a Companion to Methods in Enzymology*, vol. 26, No. 2, Feb. 2002, pp. 199-213.

Yu Jenn-Yah et al., "RNA interference by expression of short-interfering RNAs and hairpin RNAs in mammalian cells", *Proceedings of the National Academy of Sciences of the United States*, vol. 99, No. 9, Apr. 30, 2002, pp. 6047-6052.

Schwarz Dianne et al., "Evidence that siRNAs function as guides, not primers, in the *Drosophila* and human RNAi pathways", *Molecular Cell*, vol. 10, No. 3, Sep. 2002, pp. 537-548.

Hammond S M et al. "Argonaute2, a link between genetic and biochemical analyses of RNAi", *Science*, Aug. 10, 2001, LNKD-PubMed: 11498593 vol. 293, No. 5532, pp. 1146-1150, XP002183120.

Mourelatos Zissimos et al. "miRNPs: a novel class of ribonucleoproteins containing numerous microRNAs", *Genes & Development*, Mar. 15, 2002, LNKD-PubMed: 11914277, vol. 16, No. 6, pp. 720-728, XP002619532.

Martinez Javler et al. "Single-stranded antisense siRNAs guide target RNA cleavage in RNAi", *Cell*, vol. 110, No. 5, Sep. 6, 2002, pp. 563-574, XP00225781.

Meister Gunter et al. Human Argonaute2 mediates RNA cleavage targeted by miRNAs and siRNAs, *Molecular Cell*, Jul. 23, 2004, LNKD-PubMed: 15260970, vol. 15, No. 2, pp. 185-197, XP002619534.

McManus Michael T et al. "Gene silencing using micro-RNA designed hairpins", *RNA (New York)* Jun. 2002, LNKD PubMed: 12088155, vol. 8, No. 6, pp. 842-850, XP008021481.

Paddison Patrick J et al. "Short hairpin RNAs (shRNAs) induce sequence-specific silencing in mammalian cells", *Genes and Development*, Apr. 15, 2002, LNKD-PubMed: 11959843, vol. 16, No. 8, pp. 948-958, XP002204653.

Van Den Berg Aden et al. "RISC—target interaction: cleavage and translational suppression", *Biochimica et Biophysica Acta*, LNKD-PubMed: 18692607, vol. 1779, No. 11,(2008), pp. 668-677, XP002619533.

Brummelkamp Thijn R et al. "A system for stable expression of short interfering RNAs in mammalian cells", *Science (New York)*, Apr. 19, 2002, LNKD-PubMed: 11910072, pp. 550-553, XP002626048.

Zeng Yan et al. "Both natural and designed micro RNAs can inhibit the expression of cognate mRNAs when expressed in human cells", *Molecular Cell*, Jun. 2002, LNKD-PubMed: 12086629, vol. 9, No. 6, pp. 1327-1333, XP002296481.

Conklin Douglas S. "RNA-interference-based silencing of mammalian gene expression", *ChemBiochem: A European Journal of Chemical Biology*, Oct. 6, 2003, LNKD-PubMed: 14523921, vol. 4, No. 10, pp. 1033-1039, XP002623636.

Yoda et al., ATP-dependent human RISC assembly pathways, 2010, *Nature Structural & Molecular Biology*, vol. 17, pp. 17-24.

Lima et al., Single-stranded siRNAs activate RNAi in animals, 2012, *Cell*, vol. 150, pp. 883-894.

* cited by examiner

Fig. 2 A



Fig. 2 B

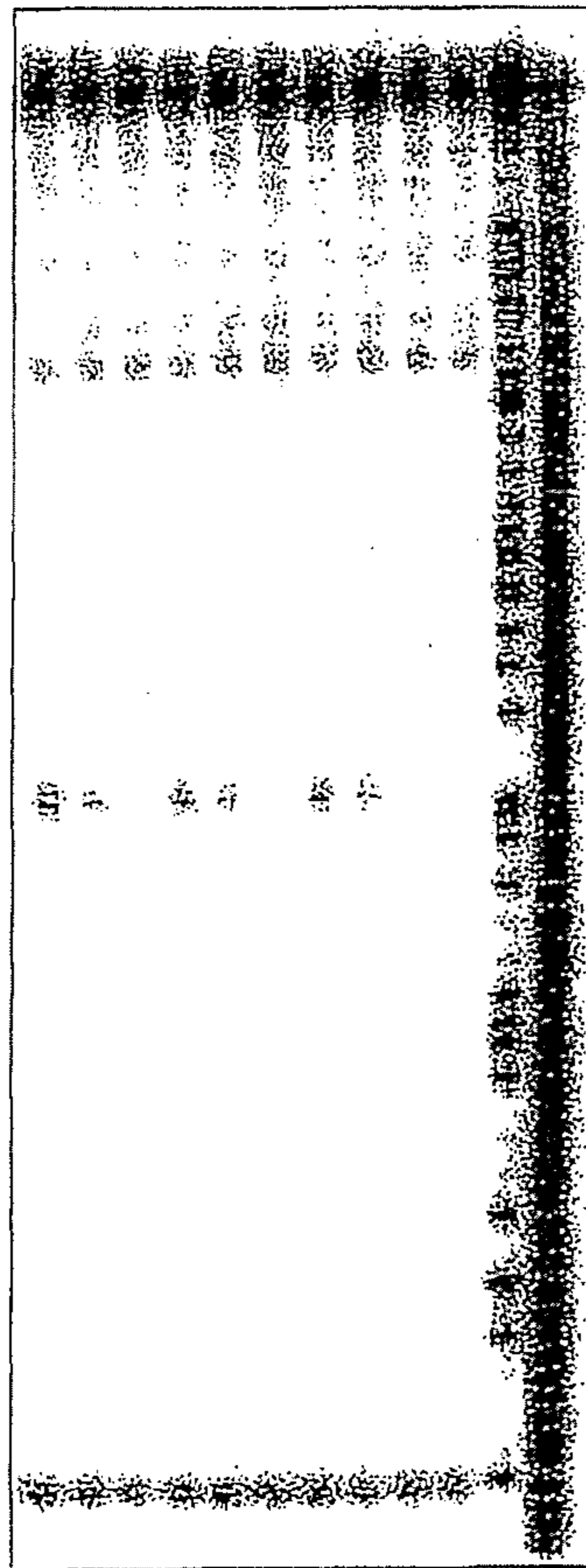
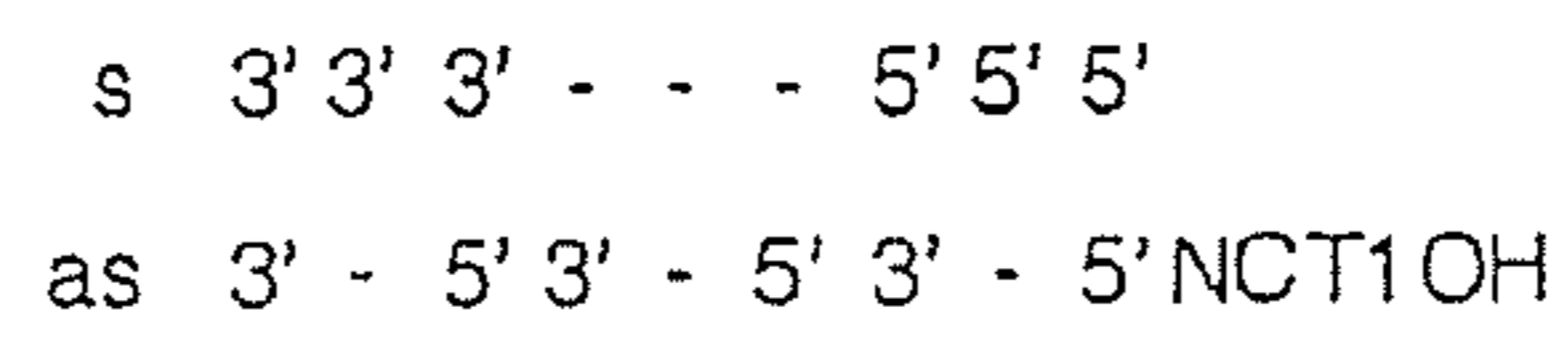
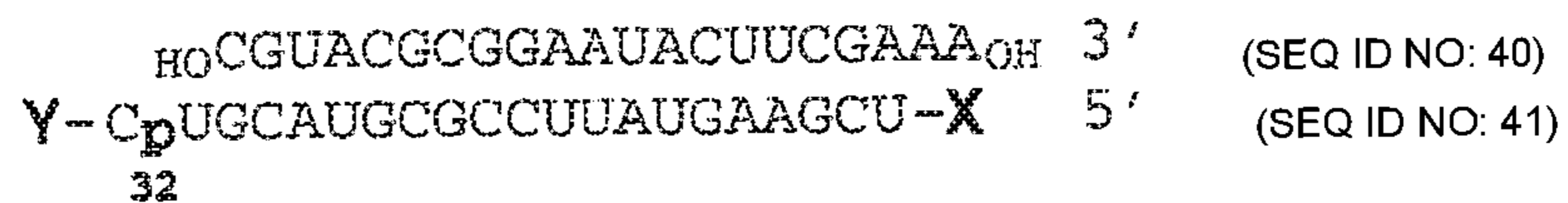


Figure 3

A



B

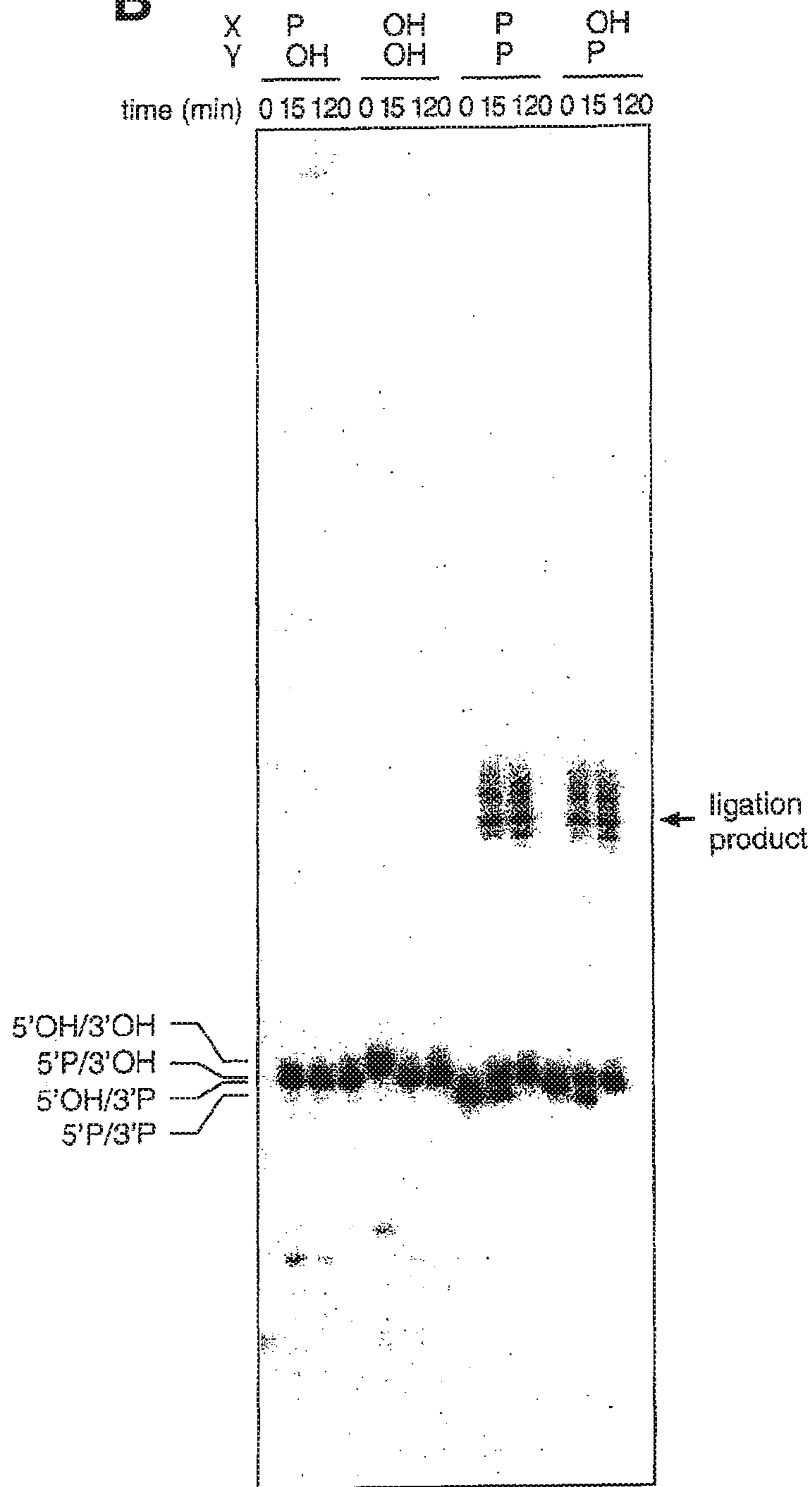


Fig. 4

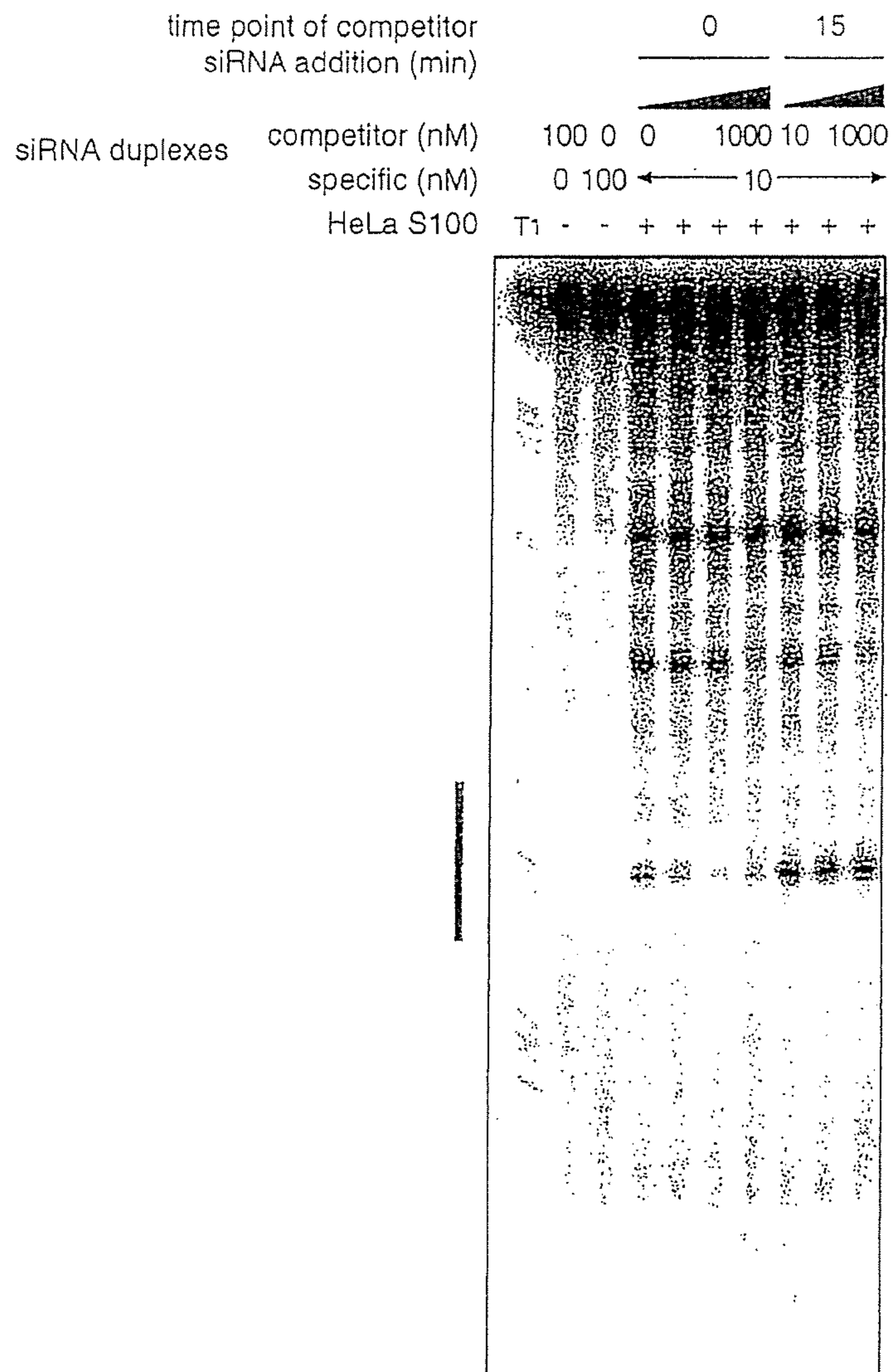


Fig. 5 A

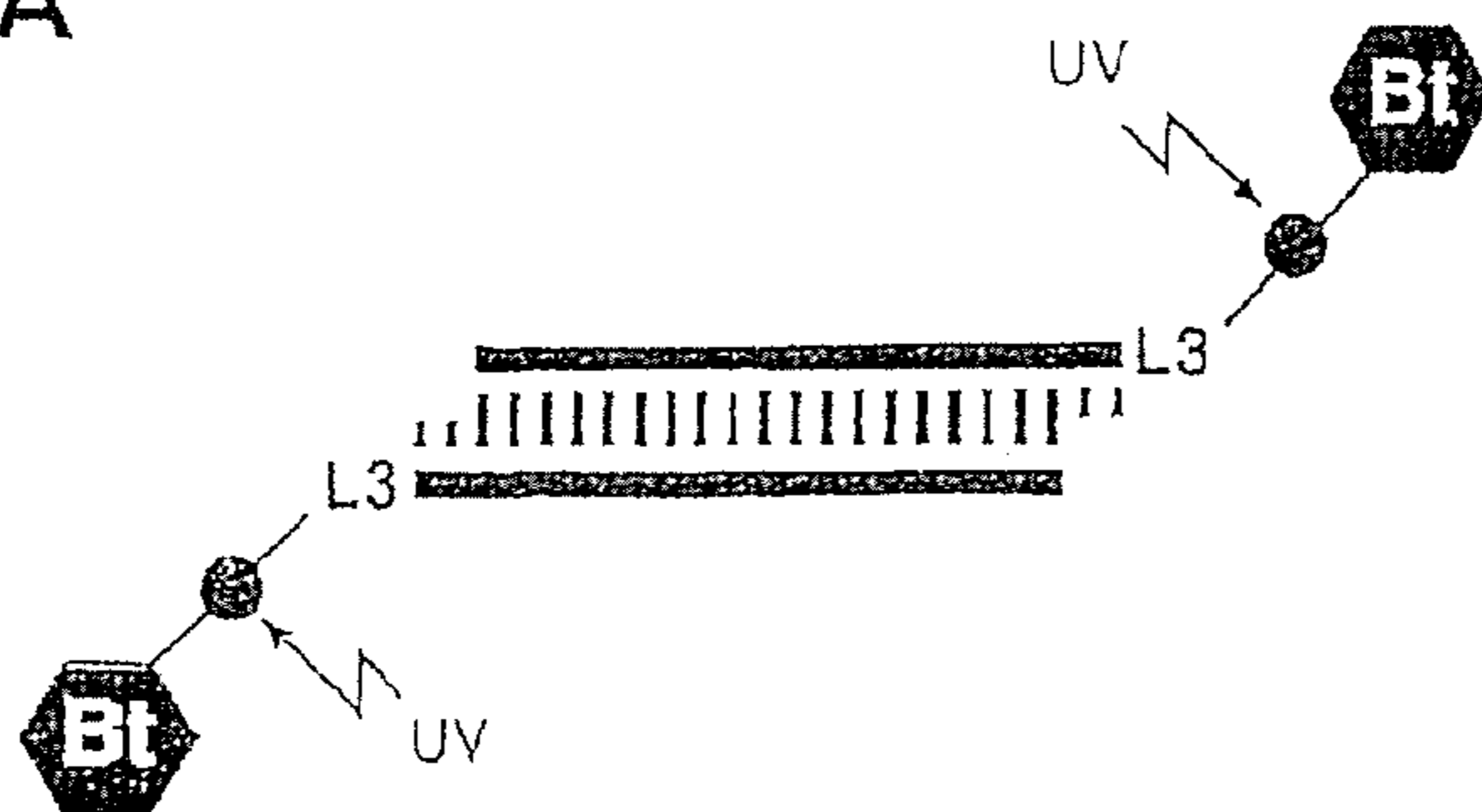


Fig. 5 B

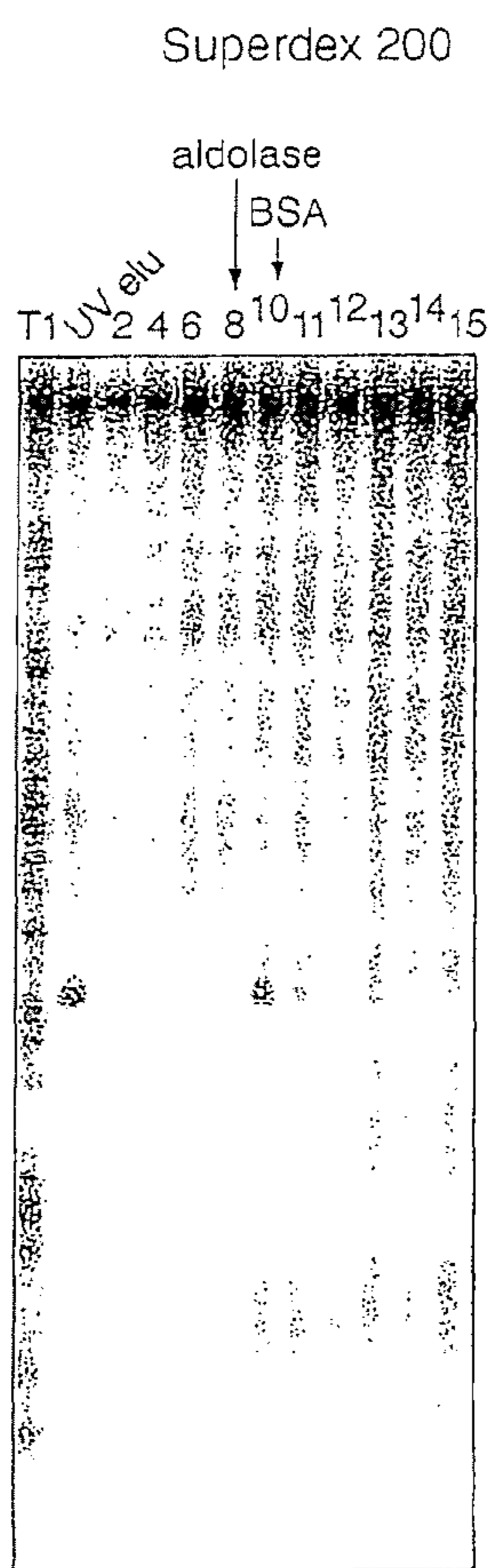


Fig. 5 C

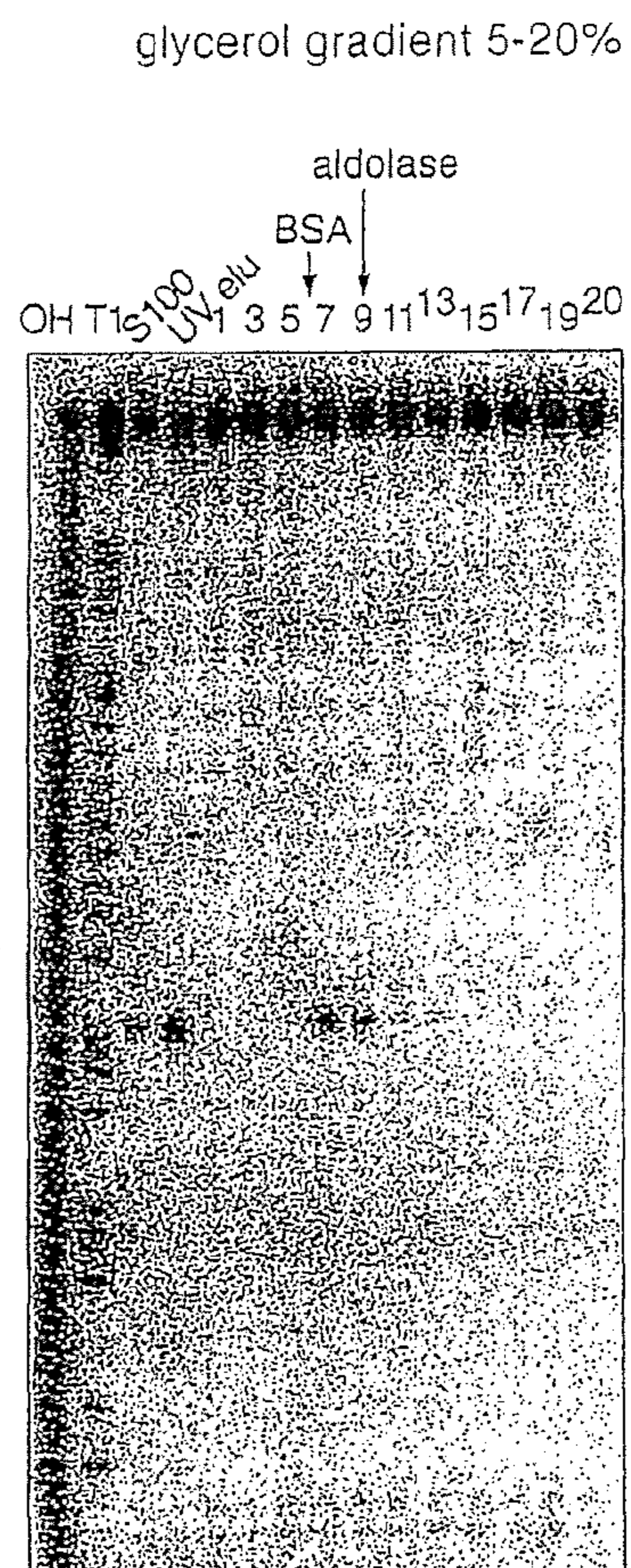


Fig. 6

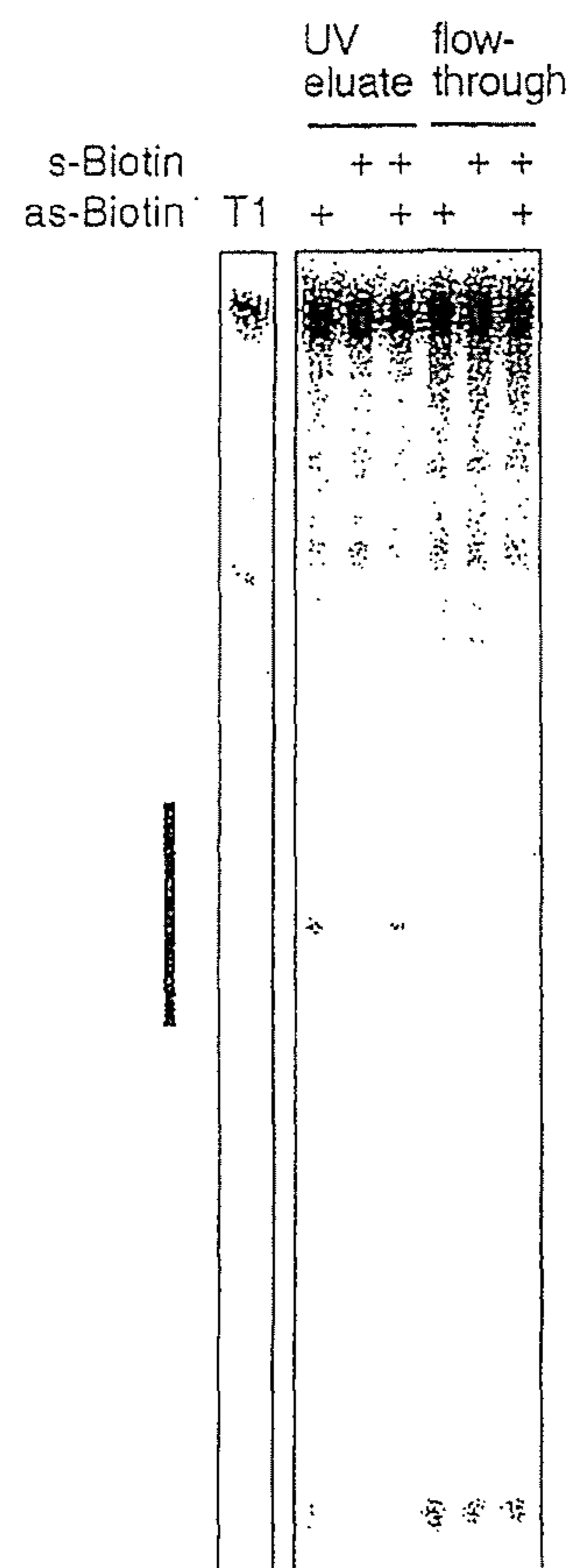


Fig. 7 **A**

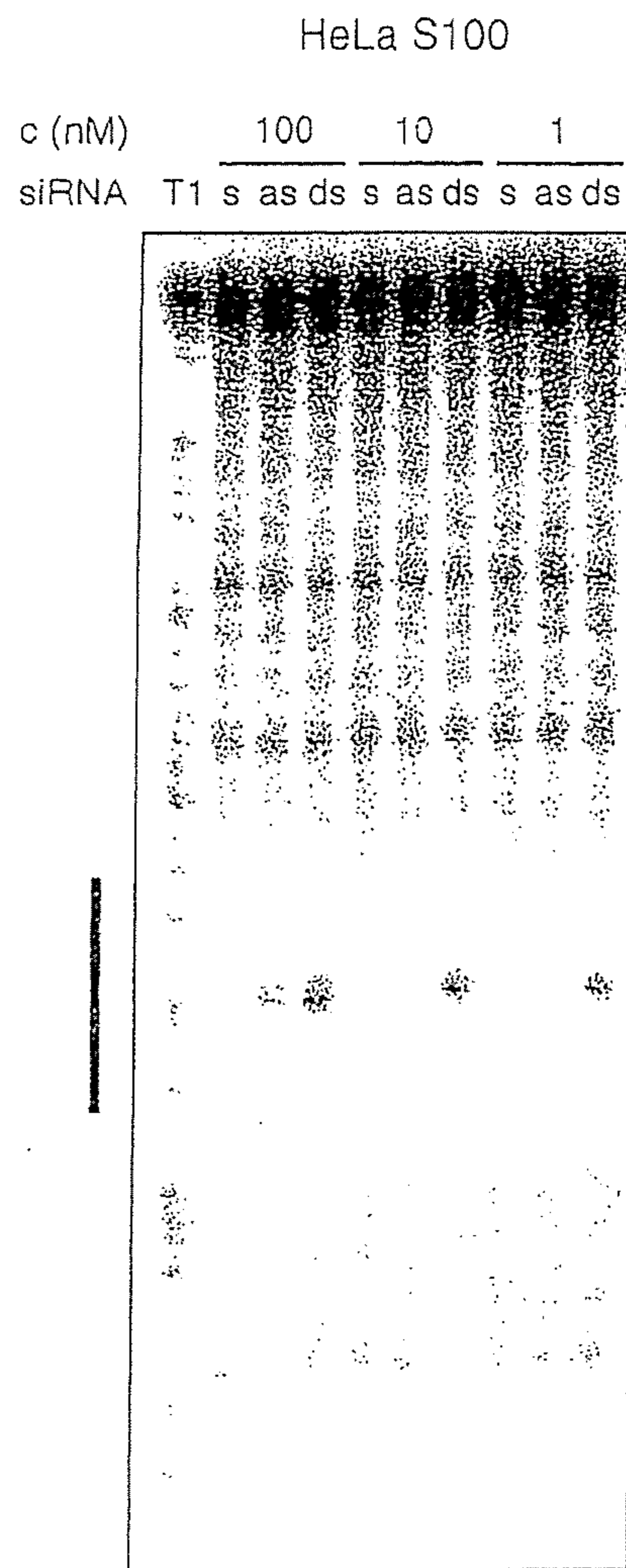


Fig. 7 **B**

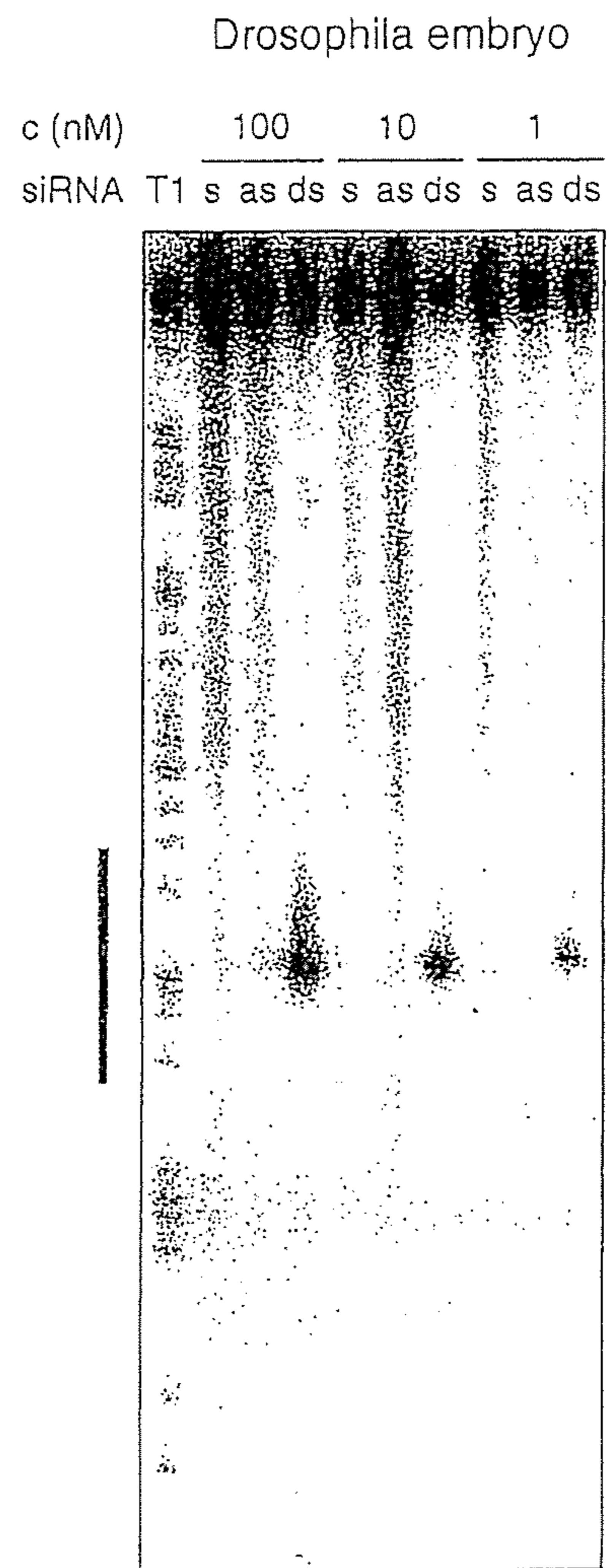


Fig. 8 A

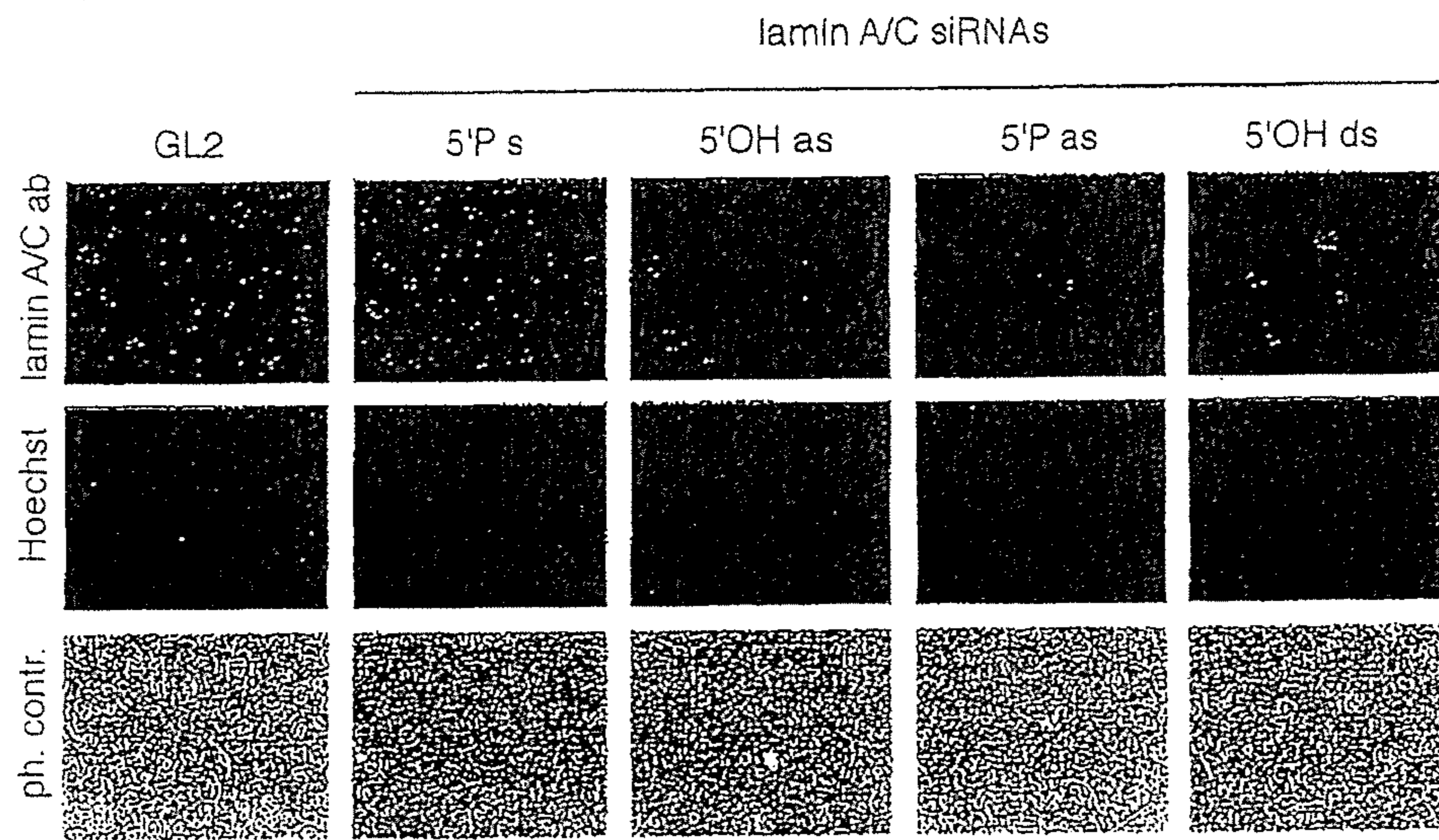


Fig. 8 B

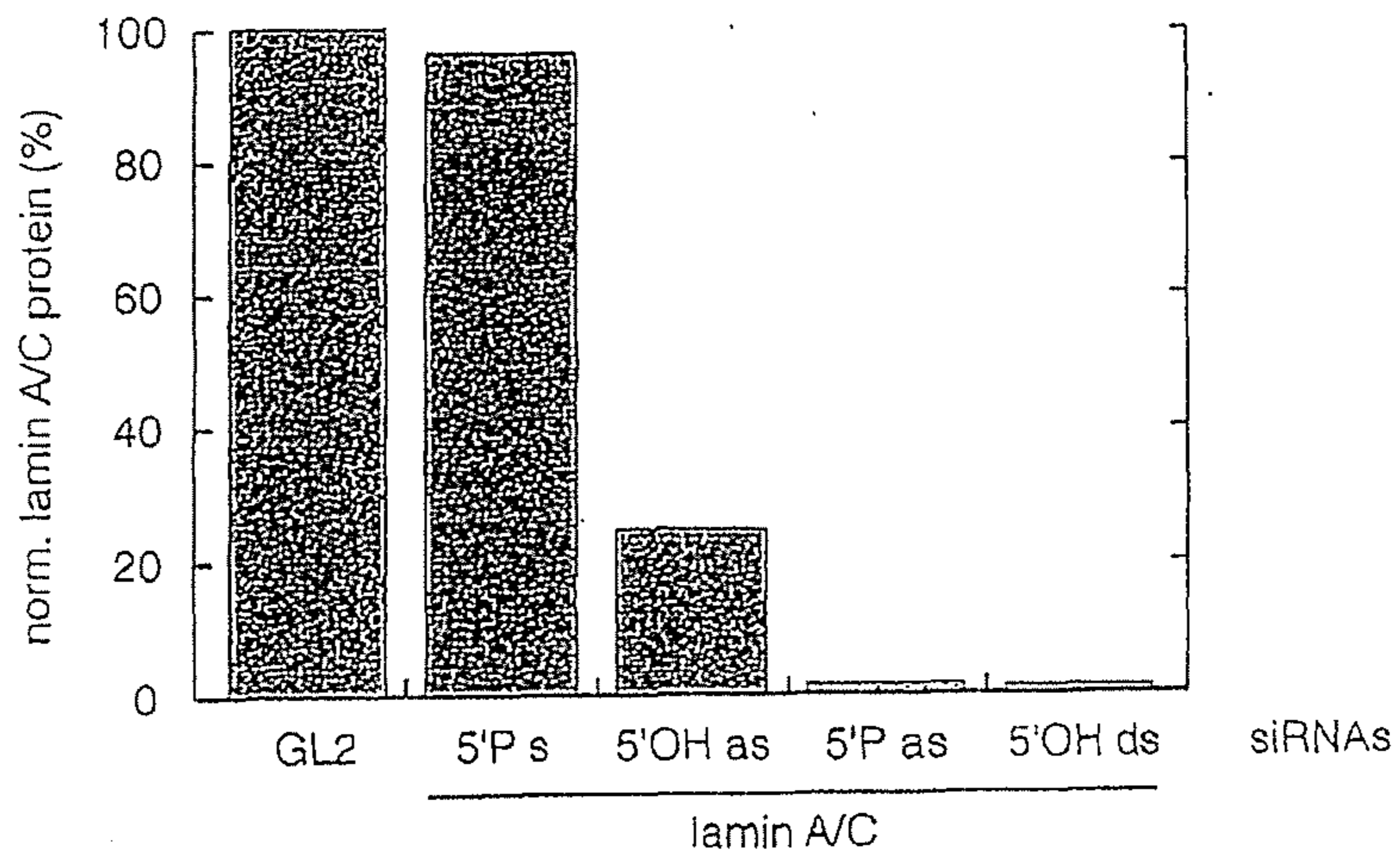


Fig. 9 A

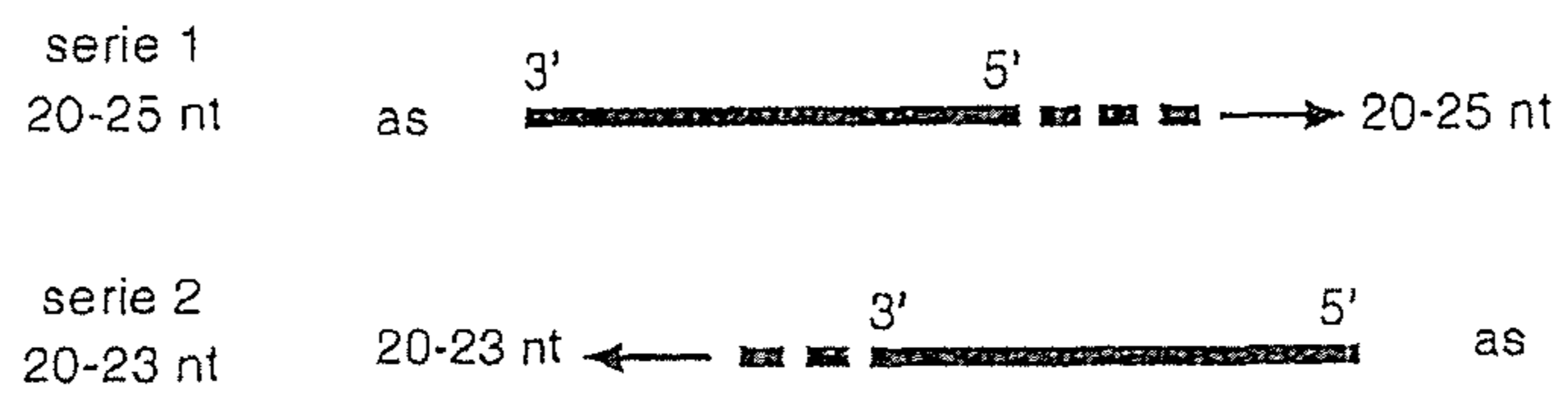


Fig. 9 B

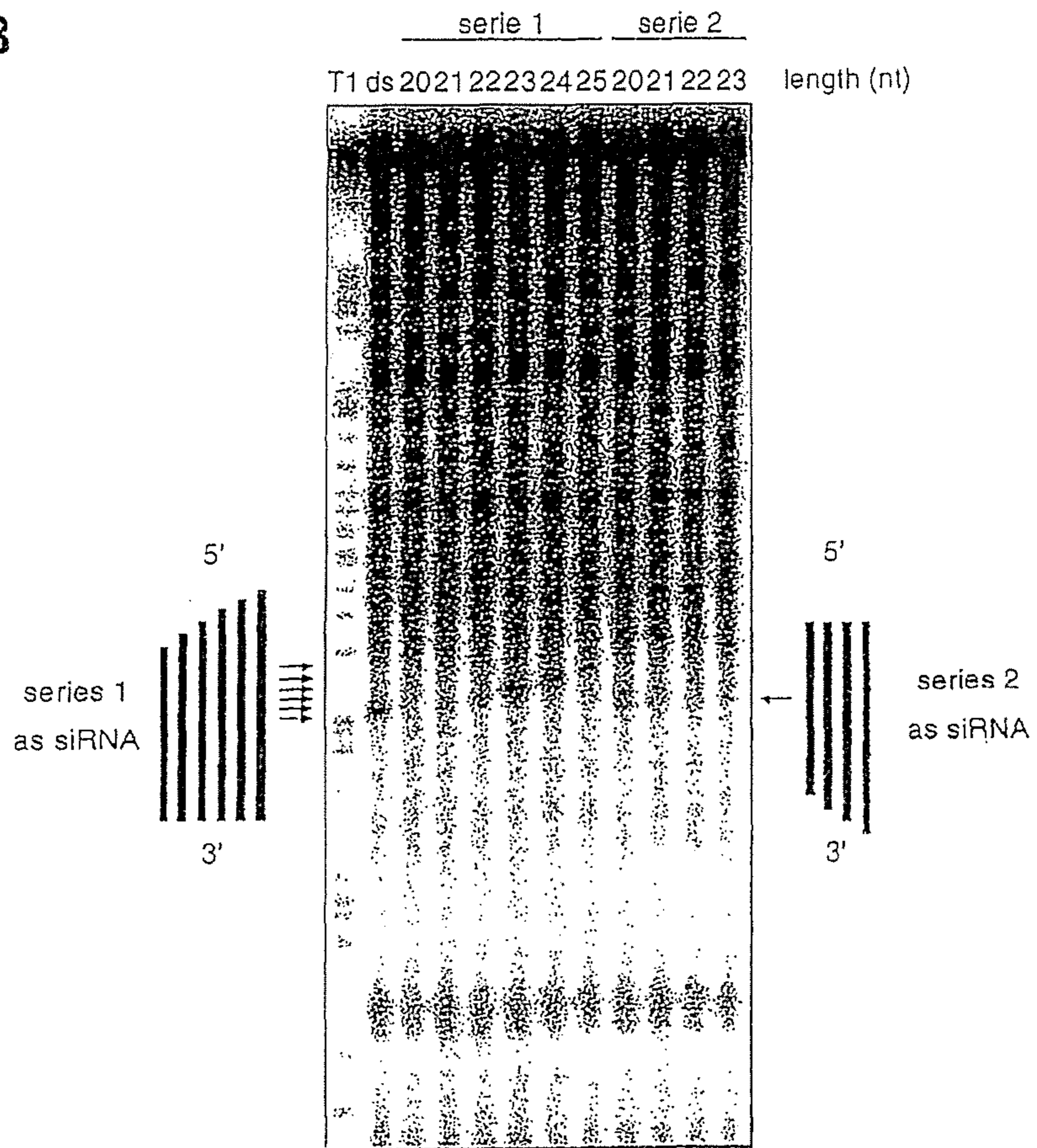


Fig. 10

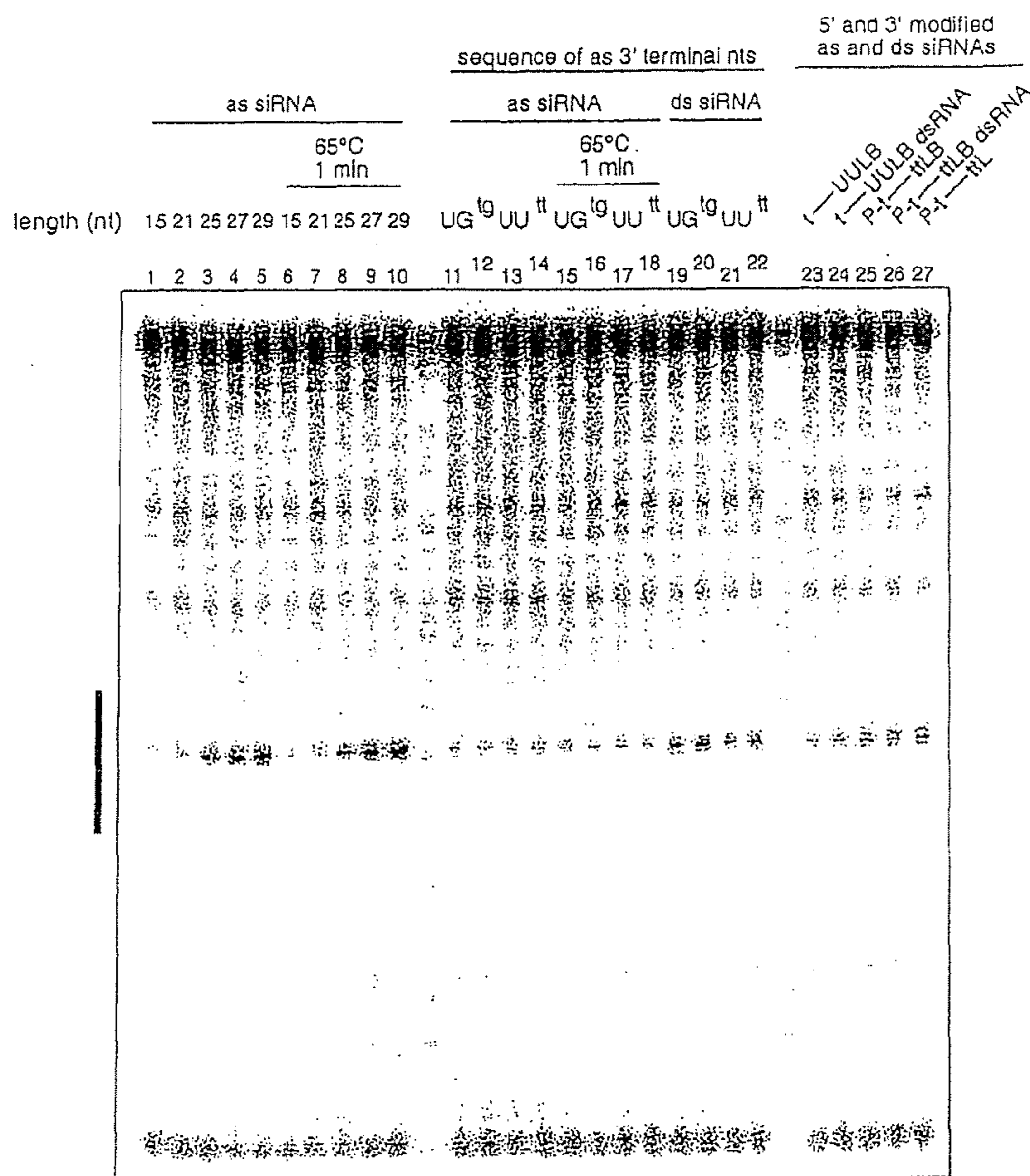


Fig. 11

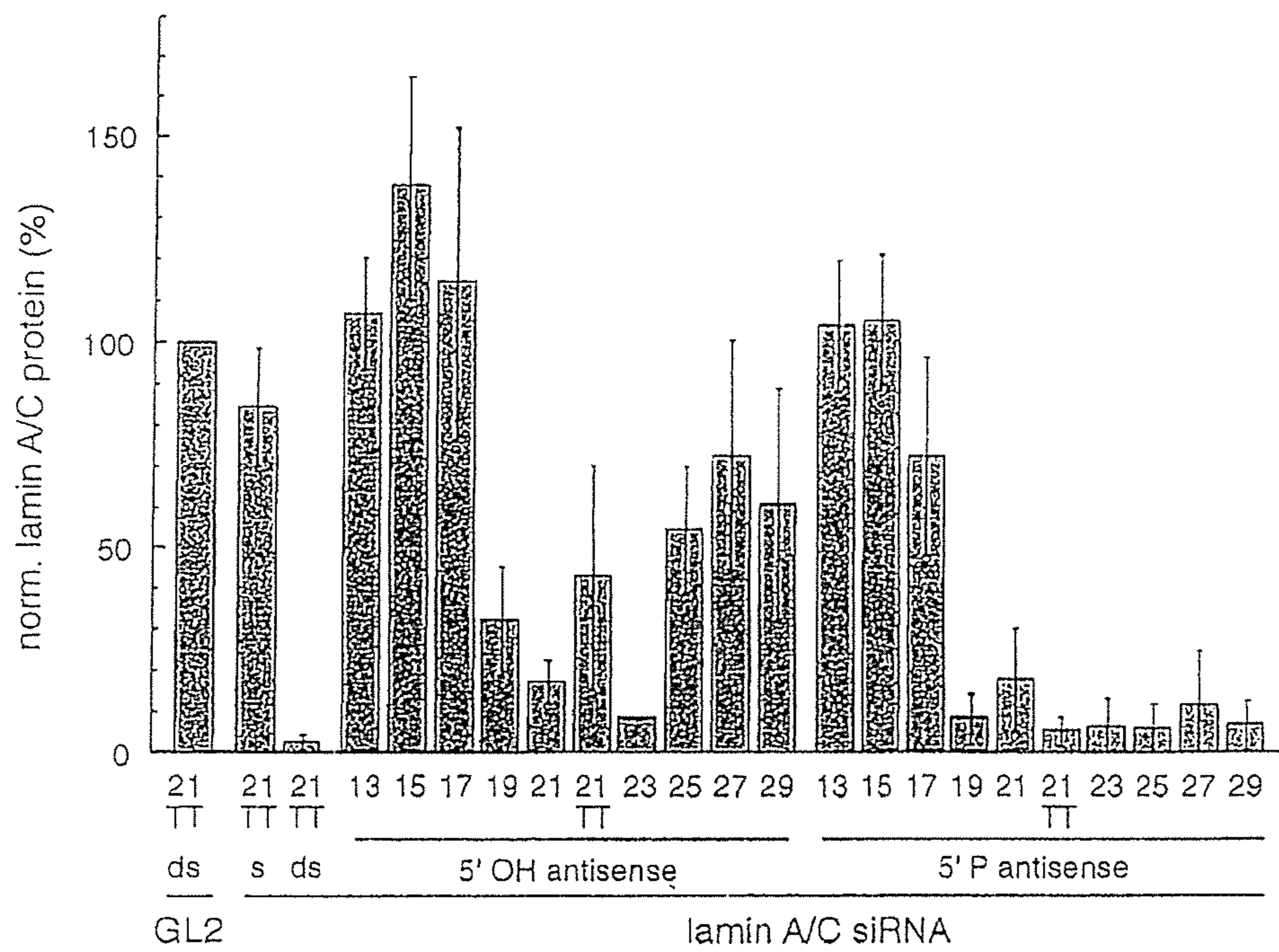


Fig. 12 A

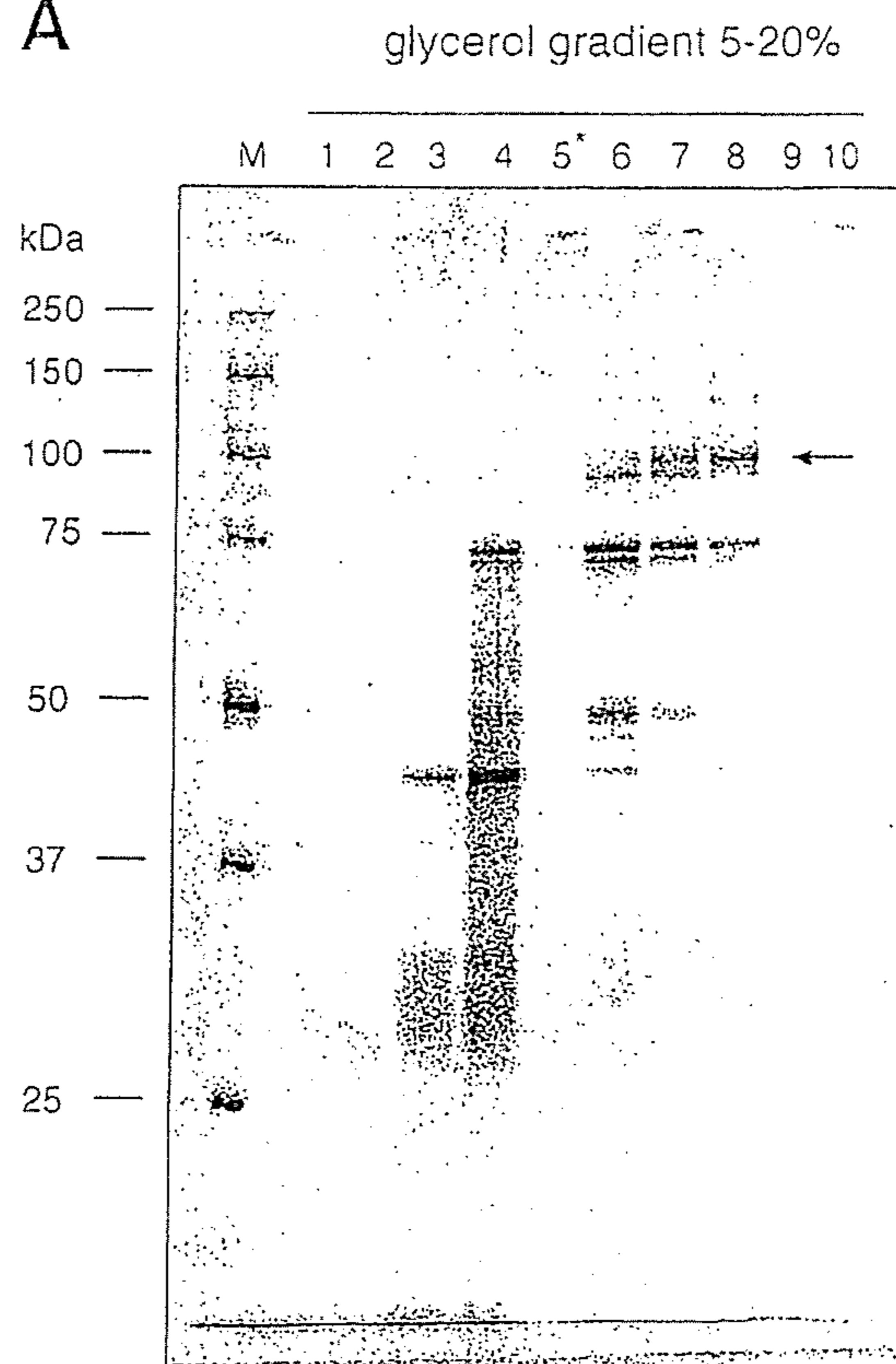


Fig. 12 B

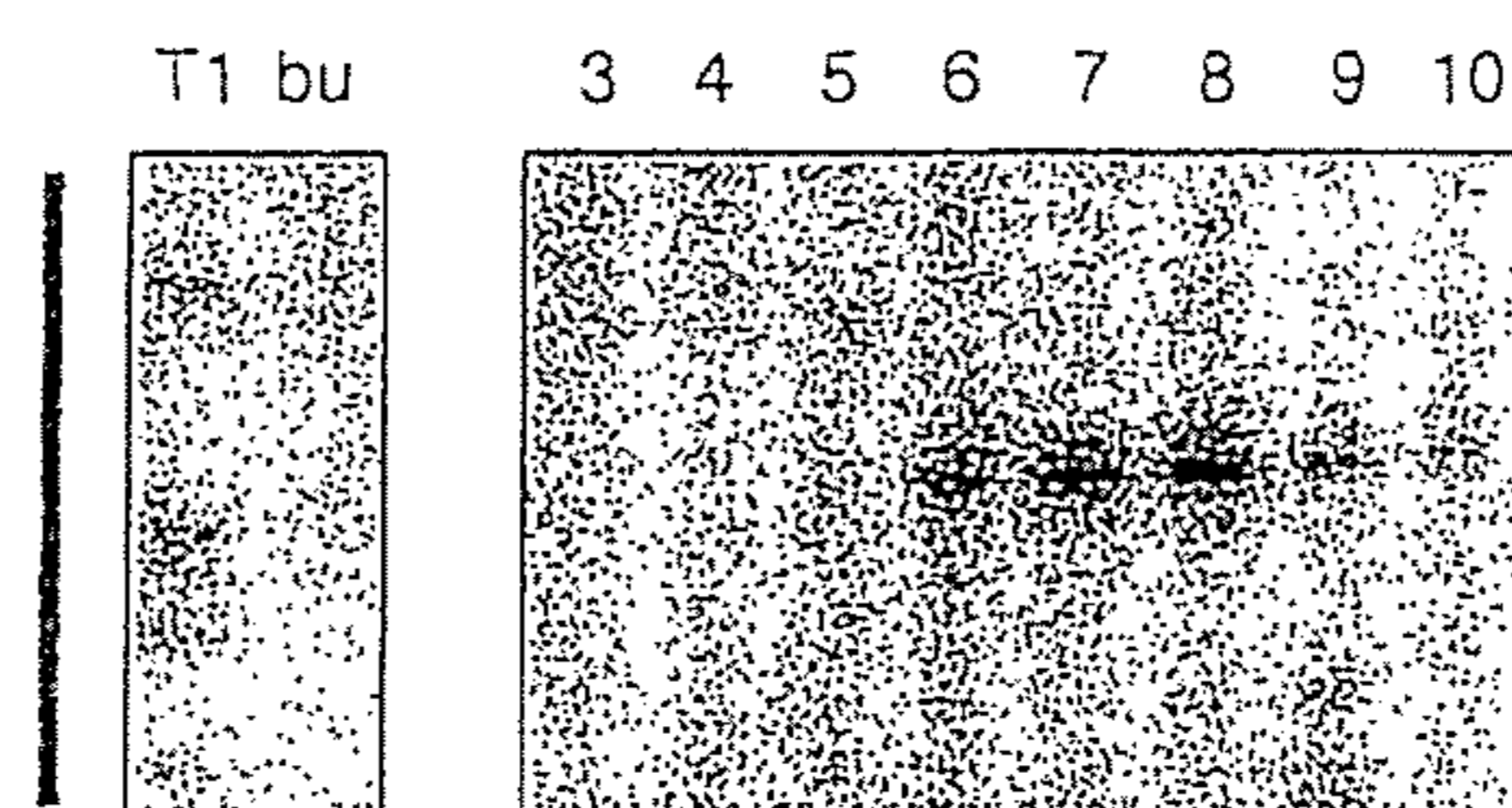
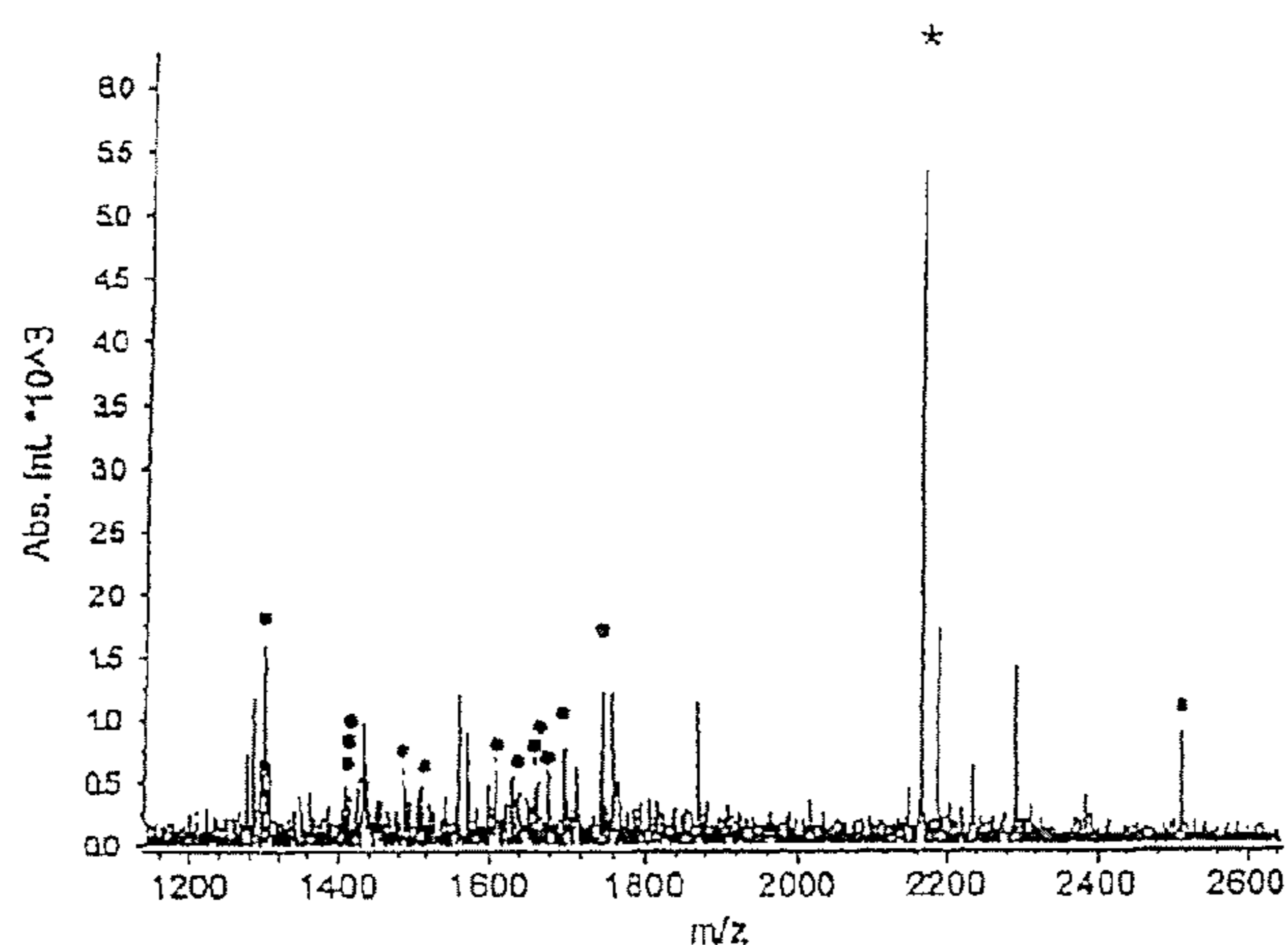


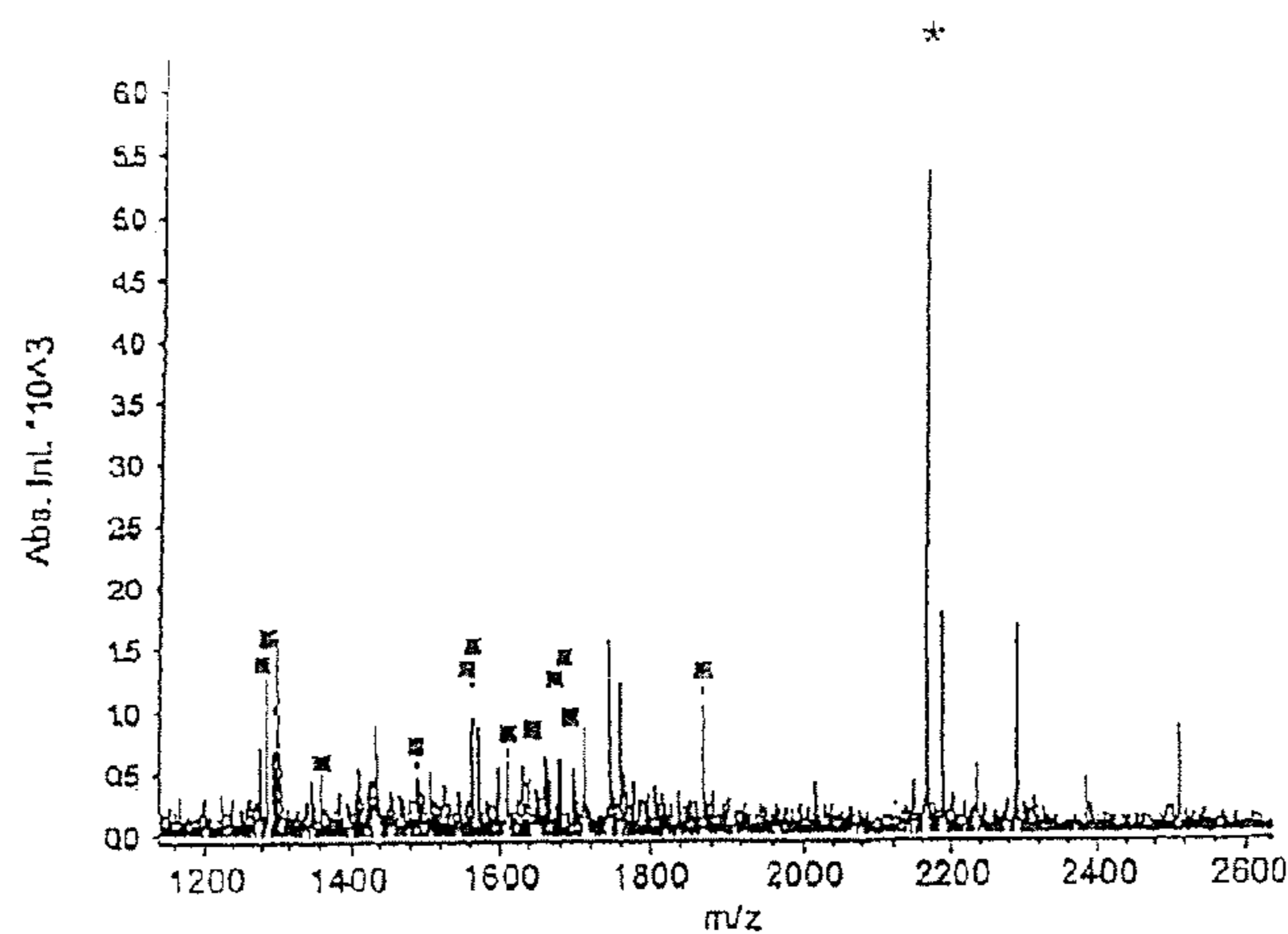
Fig. 13 A



eukaryotic translation initiation factor 2C2

Observed	Mr (expt)	Mr (calc)	Delta	Position	Miss	Peptide	
1299.67	1298.67	1298.73	-0.07	413 - 424	0	VLQPPSILYGGK	[SEQ ID NO: 42]
1402.64	1401.64	1401.74	-0.10	637 - 648	0	QETIQDLAAMVR Oxidation (M)	[SEQ ID NO: 43]
1413.62	1412.61	1412.73	-0.12	159 - 180	1	ELPSMRYTPVGR	[SEQ ID NO: 44]
1423.60	1422.59	1422.71	-0.12	356 - 367	1	KLTDNQTSTAIR Oxidation (M)	[SEQ ID NO: 45]
1486.56	1485.56	1485.66	-0.10	495 - 507	0	YAQGADSVDPHFR Oxidation (M)	[SEQ ID NO: 46]
1513.71	1512.70	1512.80	-0.10	112 - 125	1	DKVELEYTLPGEGK	[SEQ ID NO: 47]
1608.67	1607.66	1607.69	-0.03	481 - 494	0	DRGMPIQQQPCFCK	[SEQ ID NO: 48]
1635.84	1634.83	1634.85	-0.02	85 - 98	1	TQIFGDRKEVFDGR	[SEQ ID NO: 49]
1658.85	1657.85	1657.84	0.01	368 - 382	2	ATRRSAPDROEEISK	[SEQ ID NO: 50]
1663.85	1662.85	1662.91	-0.06	698 - 711	1	DYQPGITFIVVQKR	[SEQ ID NO: 51]
1675.79	1674.78	1674.84	-0.06	372 - 385	2	SAPDRQEEISKLMK Oxidation (M)	[SEQ ID NO: 52]
1696.77	1695.76	1695.84	-0.08	323 - 336	0	YPHLPCLQVGEOK	[SEQ ID NO: 53]
1743.75	1742.74	1742.77	-0.03	181 - 197	0	SFFLASEGCSNPLGGK	[SEQ ID NO: 54]
2511.07	2510.06	2510.12	-0.05	816 - 838	1	YHLVDKERDSEAGSETSGQSNK	[SEQ ID NO: 55]

Fig. 13 B



eukaryotic translation initiation factor 2C1

Observed	Mr (expt)	Mr (calc)	Delta	Position	Miss	Peptide	
1283.66	1282.65	1282.74	-0.09	410 - 421	0	VLPAPILQYGGR	{SEQ ID NO: 56}
1294.65	1293.64	1293.67	-0.03	794 - 805	0	SVSIPAPAYAR	{SEQ ID NO: 57}
1361.61	1360.60	1360.70	-0.10	553 - 564	0	TSPOTLSNLCLK	{SEQ ID NO: 58}
1486.56	1485.56	1485.66	-0.10	492 - 504	0	YAQGASVEPMFR Oxidation (M)	{SEQ ID NO: 59}
1560.76	1559.75	1559.83	-0.08	97 - 110	0	NYTYVALPIGNER	{SEQ ID NO: 60}
1561.76	1560.75	1560.78	-0.02	111 - 124	1	VDFEVTIPGEGKDR	{SEQ ID NO: 61}
1608.67	1607.66	1607.69	-0.03	478 - 491	0	DAGMPIQQPCFCK	{SEQ ID NO: 62}
1640.74	1639.73	1639.82	-0.08	240 - 253	0	NIDEQEKPLDSQK	{SEQ ID NO: 63}
1675.79	1674.78	1674.84	-0.06	369 - 382	2	SAPDROEELISRLMK Oxidation (M)	{SEQ ID NO: 64}
1679.86	1678.85	1678.90	-0.05	695 - 708	1	DYQPGITYTVVQKR	{SEQ ID NO: 65}
1696.77	1695.76	1695.84	-0.08	320 - 333	0	YPRLPCLQVQEQK	{SEQ ID NO: 66}
1867.85	1866.85	1866.87	-0.02	178 - 194	0	SFFSPPEGYYRPLGGR	{SEQ ID NO: 67}

Fig.13C

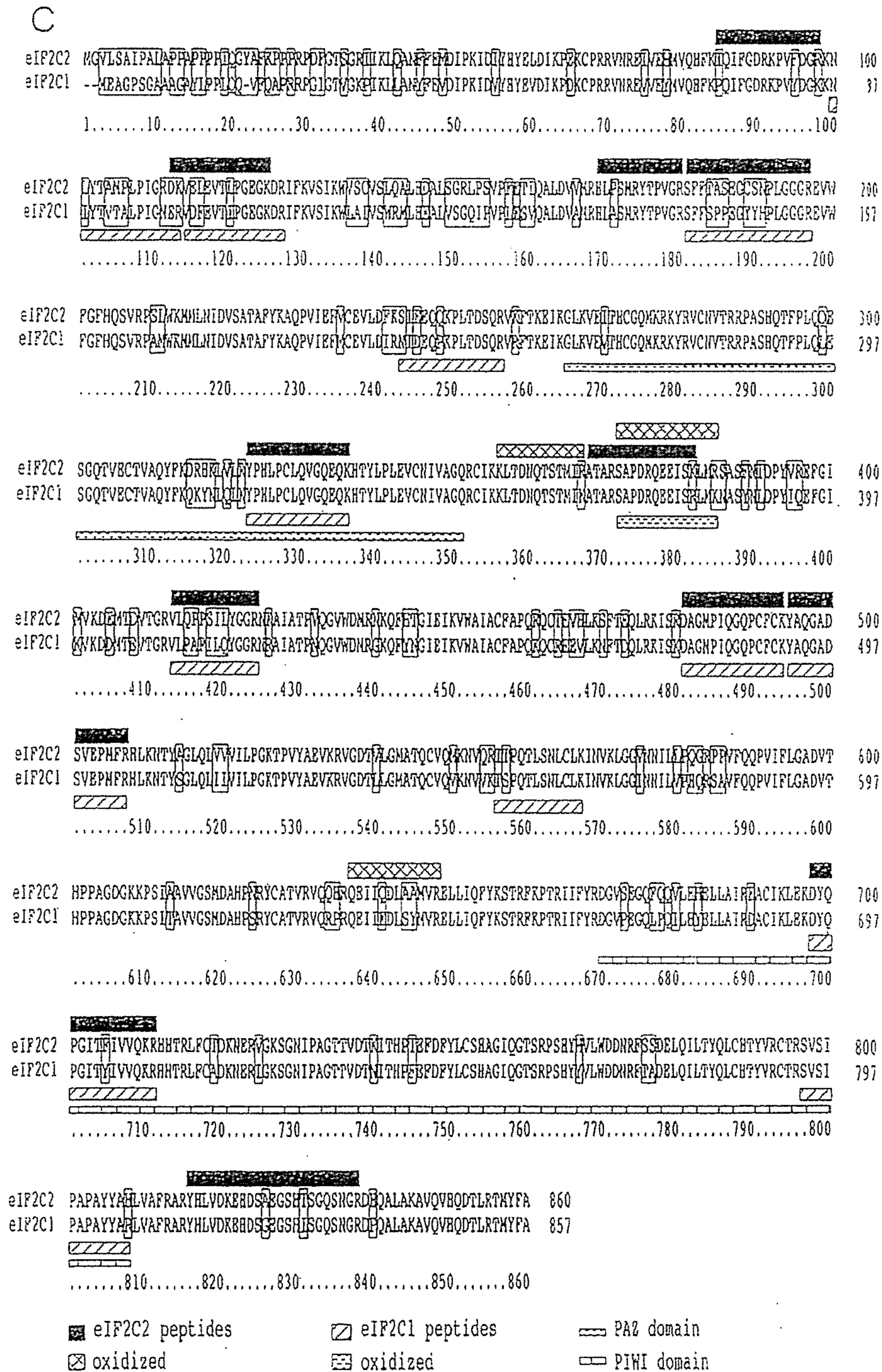


Fig. 14

>eIF2C1, predicted protein sequence

MEAGPSGAAAGAYLPPLQQVFQAPRRPGIGTVGKPIKLLANYFEVDIPKIDVYHYEVDIKPD
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MRGKQFYNGIEIKVWAIACFAPQKQCREEVLKNFTDQLRKISKDAGMPIQGQPCFCKYAQGA
DSVEPMFRHLKNTYSGLQLIIVILPGKTPVYAEVKRVGDTLLGMATQCVQVKNVVKTSPTL
SNLCLKINVKLGGINNILVPHQRSVFPQPVIFLGADVTHPPAGDGKKPSITAVVGSMDAHP
SRYCATVRVQRPRQEIIEDLSYVRELLIQFYKSTRFKPTRIFFYRDGVPEGQLPQILHYEL
LAIRDACIKLEKDYQPGITYIVVQKRHHTRLFCADKNERIGKSGNIPAGTTVDNITHPFEP
DFYLCSHAGIQGTSRPSHYVWLWDDNRFATADELQILTYQLCHTYVRCRTRSVSIPAPAYARL
VAFRARYHLVDKEHDSGEGSHISGQSNGRDPQALAKAVQVHQDTLRTMYFA

>eIF2C2, predicted protein sequence

MGVLSAIPALAPPAPPPIQGYAFKPPRPDFGTSGRTIKLQANFFEMDIPKIDIYHYELDI
KPEKCPRRVNREIVEHMQHFQKQIFGDRKPVFDGRKNLYTAMPLPIGRDKVELEVTLPEEG
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APDRQEEISKLMSASFNTDPYVREFGIMVKDEMTEVTGRVLPQPSILYGGRNKAIATPVQG
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HELLAIREACIKLEKDYQPGITFIVVQKRHHTRLFCADKNERVKGKSGNIPAGTTVDTKITHP
TEFDYLCSHAGIQGTSRPSHYVWLWDDNRFSSDELQILTYQLCHTYVRCRTRSVSIPAPAY
AHLVAFRARYHLVDKEHDSAEGSHTSGQSNGRDHQALAKAVQVHQDTLRTMYFA

>eIF2C3, predicted protein sequence

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AMEALGPGPPASLFQPPRRPGLGTVGKPIRLLANHFQVQIPKIDVYHYDVIDIKPEKRPRRN
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LKYPHLPCLQVQEQKHTYLPLEVCNIVAGQRCIKKLTNDQSTMIKATARSAPDRQEEISR
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Fig. 14 (Con.)

>eIF2C4, predicted protein sequence
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>HILI, predicted protein sequence
ISSGDAGSTFMERGVKNKQDFMDSLICTREKLAHVRNCKTGSSGIPVKLVTNLNFNLD'FPQDW
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TIKMTITLKR'ELPSSSPVCIQVFNIFR'KILKLSMYQIGRNFYNPSEPMEIPQHKLSLWPG
FAISVSYFERKLLFSADVS'YKVLNRNETVLEFMTALCQRTGLSCFTQTCEKQLIGLIVLTRYN
NRTYSIDDDIDWSVKP'THTFQKR'DGTEITYVDYKQYDITVSDLNQ'PMLVSLK'KKRNDNSE
AQLAHLIPELC'FLTGLTDQATSDFQ'LMKAVAEKTRLS'PSGRQQLARLVDNIQRNTNARFEL
ETWGLHFGS'QISLTGRIVPSEKILMQDHICQ'PVSAADWSKD'IRTKILNAQSLNTWLTLCSD
RTEYVAESFLNCLRRVAGSMGFNVMC'ILPSNQKTY'YDSIKKYLSSDCPVPSQCVLARTLNKQ
GMMNSIATKIAMQMTCKLGGELWAVEI'PLKSLMVV'GIDVCKDALSKDVMVVGCVASVNPRI
T'RWFSRCILQRTMTDVADCLKVFMTGALNKWYKYNHDL'PARIIVYRAGVGDGQLKTLIEYEVP
QLLSSVAESSNTSSRLSVIVVR'KKCMRPF'FTENRRTVQNPPLGTVVDS'EA'RN'EWQYDFYL
ISQVACRGT'VSPTYYNVIYDDNGLKPDHM'QRLTFKLC'HL'Y'NWP'GIVSV'PAPCQYAHKLTFL
VAQSIHKEPSLELANHLFYL

>HIWI, predicted protein sequence
MTGRARARARGRARGQETAQLVGSTASQQPGYIQPRPQPPPAEGELFGRGRQRGTAGGTAKS
QQLQISAGFQELSLAERGRRRDFHDLGVNTRQNL'DHVKESKTGSSGII'VRLSTNHFR'LT'SR
PQWALYQYHIDYNPLMEARRLSALLFQ'HEDLIGKCHA'FDG'TILF'LPKRLQ'QKVTEVFSKTR
NGEDV'RITITLTN'ELPPTSPTCLQFYNIIFR'LLKIMNLOQ'IGRNYNPN'DPIDIPSHRLVI
WPGFTT'SILQYENSIMLCTDVSHKVL'RSETVLD'FMFNFYHQ'EEH'KFQE'QVSKELIGLVVLT
KYN'NKTYRVDDIDWDQ'NPKSTFKKADGSEV'SFLEYR'KQYNQ'EI'DLKQ'PVLVSQPKRRRGF
GGTLP'GPAMLIPELCYLTGLTDKMRNDFNVMKDLAVHTR'LTPEQRQ'EVGRLIDYIHKNDNV
QRELRDWGLSFD'SNLLSFSGRILQTEKIHQ'GGKTFDYNPQ'FADWSKETRGAPLISVKPLDNW
LLIYTRRNYEAANS'LIQNL'FKVTPAMGMQMRKAIMI'EVDDRTEA'YL'RVLQ'QKV'TADTQIVVC
LLSSNRKDKYDAIKKYLCTDCPTPSQCVVARTL'GKQQTVM'AIATKIALQMNCKMGGELWRVD
IPLKLVMI'VGIDCYHDMT'PAGRRSIAGFVASIN'EGMTRWF'SRCIFQDRGQELVDGLKVCLQAA
LRAWNSCNEYMPSRIIVYR'DCVGDGQLKTLVNYE'V'PQFLDCLKSIGR'GYNPRLT'VI'VVKR'V
NTRFFAQSGGRLQ'NPLPGTVIDVEVTRPEWYDF'FIVSQAVRSGSVSPTHY'NVIYD'NSGLKPD
HIQRLTYKLC'HIYYNWP'GIVRVPAPCQYAHKLA'FLV'GQSIHREPNLSLSNR'LYYL

Fig. 15

eIF2C3 ¹SRKRVVLPQPGAAARPEEATASPRRHSANIEPKRYA¹⁰AAAAAAGECAGGACDRGSRAPAAANKAL²⁰SETPASLF³⁰PPREGLGIVKPIRLANFCA⁴⁰
 eIF2C4 -----RCPKAV³⁴ELLVPPRFYICVAKPIKLANCFQV
 eIF2C1 -----MRAGPSCAAAGVLPPLQVVDAPPREGLGIVKPIKLANFCEM⁴⁵
 eIF2C2 -----MGMISHTPAIRPPVPPPHDCAAFPPRPEDYDTSQRNKLQAAVFEH⁴⁸
 HILI -----ISSDAESTHFERGVNKKDFDL²⁴
 HIRI -----MTEPARAKAGSAREDETRDLVSTAGQPGYICDPPFPAGEHFCGRORGTAGGTPKSGLDISAGFOELSLERGGQR-RRDFEDL⁸⁵
 ruler 10 20 30 40 50 60 70 80 90 100

eIF2C3 ¹QIPKIDVYHYVDIKPEK¹⁰PRRWREVDIMVRFRIQIFEDRQESHDKRN²⁰MTTAPHELETGNDVWEVILEEG-KDITPKVSVQVYVSW³⁰
 eIF2C4 ¹ETPKIDVYHYVDIKPKC¹⁰PRRWREVDISVDFRILVIFEDRQVYDKRS²⁰LHTAPHELETGNDVWEVILEEG-KDITPKVSVKVFVSRV³⁰
 eIF2C1 ¹DIPKIDVYHYVDIKPEK¹⁰PRRWREVDVYVDFRILVIFEDRQVYDKRN²⁰LHTAPHELETGNDVWEVILEEG-KDITPKVSVKGLAIA³⁰
 eIF2C2 ¹DIPKIDVYHYVDIKPEK¹⁰PRRWREVDVYVDFRILVIFEDRQVYDKRN²⁰LHTAPHELETGNDVWEVILEEG-KDITPKVSVKGLVSW³⁰
 HILI ¹STCTREKLABVRRNCKTGSSTENKLVINEFVLDREDDVQVDFVYTYIEFLASRRIRIALVSHSELNKKAKAFDCMILFSLQMLEKAVILESSSEIQRSE¹²⁴
 HIRI ¹GVNTRONLDEWESKMGSSGIIURLSTVHFVLSRFDVHILVQVHIDVHFLMARRLSALLEHSDLLIKCHAFDQVLLLEKRIQVWTEVSKVTRNCE¹⁸⁹
 ruler 110 120 130 140 150 160 170 180 190 200

eIF2C3 ¹STQLLLEALDQVIA¹⁰MPDSSVQALDVVTR-MVLSRYITVEGSEFSPPEGVMEPLGGCREVWHGTFHOSVPPAMKRMMLNIDNSATAFVRAQF²⁰
 eIF2C4 ¹SMALLHRLVLTETVLEPEL¹⁰YVISTVMAVAVVLA-FLASRYITVEGSEFSPPEGVMEPLGGCREVWHGTFHOSVPPAMKRMMLNIDNSATAFVRAQF²⁵
 eIF2C1 ¹SVRDLHEALVSGDIE¹⁰MPDSSVQALDVVTR-MVLSRYITVEGSEFSPPEGVMEPLGGCREVWHGTFHOSVPPAMKRMMLNIDNSATAFVRAQF²⁷
 eIF2C2 ¹SLQALHDAISVRAPE¹⁰MPDSSVQALDVVTR-MVLSRYITVEGSEFSPPEGVMEPLGGCREVWHGTFHOSVPPAMKRMMLNIDNSATAFVRAQF³⁰
 HILI ¹IIKMTITLKRBLPSS¹⁰SPVCIQVPMIIFRRLKKL²⁰SNVQIGRNFVAPSEP³⁰VIPQHK-LSLWCFMISVSTFBRKLLFSADNSYKVL⁴⁰-NET²¹³
 HIRI ¹DVRITITLQNELPPI¹⁰SEPCIQVPMIIFRRLKKL²⁰INELOIGRNFVAPSEP³⁰VIPQHK-LVWVCFMISVSTFBRKLLFSADNSYKVL⁴⁰-SET²⁷⁸
 ruler 210 220 230 240 250 260 270 280 290 300

eIF2C3 ¹ITTEKCEVLDVINEQKPLTDSQVRRTRRTRCLVETHECOMRRKRVVQVMTRRPASRDTFFLQRMEDMECTHADVFKDRVLSLCKYPHLPCLD³⁰
 eIF2C4 ¹VIQPKCEVLDVINEQKPLTDSQVRRTRRTRCLVETHECOMRRKRVVQVMTRRPASRDTFFLQRMEDMECTHADVFKDRVLSLCKYPHLPCLD³²⁵
 eIF2C1 ¹VIEPKCEVLDVINEQKPLTDSQVRRTRRTRCLVETHECOMRRKRVVQVMTRRPASRDTFFLQRMEDMECTHADVFKDRVLSLCKYPHLPCLD³²⁷
 eIF2C2 ¹VIEFVCEVLDVINEQKPLTDSQVRRTRRTRCLVETHECOMRRKRVVQVMTRRPASRDTFFLQRMEDMECTHADVFKDRVLSLCKYPHLPCLD³³⁰
 HILI ¹VLDFAVLCQRTGLS¹⁰CFVTCRKLICLIMLRVH²⁰HRVYSDIDWVSRPHTFQKR³⁰DSSEITVWYKQDQDITVSDLNQPHL⁴⁰
 HIRI ¹VLDFAVLYHOTZEH¹⁰KFGQVSKELICLIMLRVH²⁰HRVYVDDIDWQVKSFPKKA³⁰DSSEITVWYKQDQDITVSDLNQPHL⁴⁰
 ruler 310 320 330 340 350 360 370 380 390 400

eIF2C3 ¹VEEDKHT¹⁰YLPLEVCHVAGRCIKKLDNOT²⁰STAIKAVARSAPDROBEISLWSSAYET³⁰DPVVECFVWQVRSMAHTEWLPAPVILQ⁴⁰
 eIF2C4 ¹VEEDKHT¹⁰YLPLEVCHVAGRCIKKLDNOT²⁰STAIKAVARSAPDROBEISLWSSAYET³⁰DPVVECFVWQVRSMAHTEWLPAPVILQ⁴⁰
 eIF2C1 ¹VEEDKHT¹⁰YLPLEVCHVAGRCIKKLDNOT²⁰STAIKAVARSAPDROBEISLWSSAYET³⁰DPVVECFVWQVRSMAHTEWLPAPVILQ⁴⁰
 eIF2C2 ¹VEEDKHT¹⁰YLPLEVCHVAGRCIKKLDNOT²⁰STAIKAVARSAPDROBEISLWSSAYET³⁰DPVVECFVWQVRSMAHTEWLPAPVILQ⁴⁰
 HILI ¹SLKRRN-DNSAQLAHLIFELCUTGLTONTSDVQKAVAKRNLSPSEFOYAPLVNDIQRVWVRELVKELHES-QISLIGVWPSERIL¹²⁵
 HIRI ¹SQKRRRCEGGTLPQPAHLIFELCUTGLTONTSDVQKAVAKRNLSPSEFOYAPLVNDIQRVWVRELVKELHES-QISLIGVWPSERIL¹²⁵
 ruler 410 420 430 440 450 460 470 480 490 500

Fig.15 (cont.)

eIF2C3 YGG-RNRIVVAPNQQVYDMPGKQRLAQDAIRVHAVACFAPQKQCHEDLIRSFNDQIRPISKDAQCHYKCCQECFQMAQCADSVAPRRHHRHMTVQIQLL 573
eIF2C4 YGG-RNRIVVAPESHGVDMKGRQFTQVETAWAIACFADPQCPPEILRQFTLQLRKMSKDAQCHYKCCQECFQMAQCADSVAPRRHHRHMTVQIQLL 514
eIF2C1 YGG-RNRIVVAPNQQVYDMPGKQRLAQDAIRVHAVACFAPQKQCHEDLIRSFNDQIRPISKDAQCHYKCCQECFQMAQCADSVAPRRHHRHMTVQIQLL 516
eIF2C2 YGG-RNRIVVAPNQQVYDMPGKQRLAQDAIRVHAVACFAPQKQCHEDLIRSFNDQIRPISKDAQCHYKCCQECFQMAQCADSVAPRRHHRHMTVQIQLL 519
HIL1 QDDHICDPVSRADWSKDIRTCRILNAQSLTWHLLCSDR---TSYVASSFLAKLRRAVCSMGTH-----VM 459
HIM1 QGKTFDYEDPADWSKDIRTCRILNAQSLTWHLLCSDR---HYEAMASLIONRFRVMPVAGLQMRKMHLEVDRTBAVMAVCKVATADQ--INV 557
ruler510.....520.....530.....540.....550.....560.....570.....580.....590.....600

eIF2C3 VVILFGKTEVVAEVRKMAETILLCHATCCVAVRMYK-TSPQTLSEKLRINAKLGGDINVLVHAPPSVFCQVIFLQADMTHPHACCKKPSIAAVV 671
eIF2C4 IVILFGKTEVVAEVRKMAETILLCHATCCVAVRMYK-TSPQTLSEKLRINAKLGGDINVLVHAPPSVFCQVIFLQADMTHPHACCKKPSIAAVV 612
eIF2C1 IVILFGKTEVVAEVRKMAETILLCHATCCVAVRMYK-TSPQTLSEKLRINAKLGGDINVLVHAPPSVFCQVIFLQADMTHPHACCKKPSIAAVV 614
eIF2C2 VVILFGKTEVVAEVRKMAETILLCHATCCVAVRMYK-TSPQTLSEKLRINAKLGGDINVLVHAPPSVFCQVIFLQADMTHPHACCKKPSIAAVV 617
HIL1 CIIIPSNOKTYVDSIKFYSSDCPPSCCVLAVILAKQCHASIPVPIAMQVCKLGG-----ELNAGDIFLKSLEHVVHILVCKDLSK--DMAVGCV 551
HIM1 CLLSSNRKDKYDAIKKYLCTDCPPSCCVVAVFLSKOOTVAIPVPIAMQVCKLGG-----ELNAGDIFLKSLEHVVHILVCKDLSK--RRSIAAGFV 649
ruler610.....620.....630.....640.....650.....660.....670.....680.....690.....700

eIF2C3 SDCHESSYCAIVVAVTSRQISQELLYSCVVIQDITMVRPELLIQVYKSTRFKPTRIIFVRCVSEGVFQVAVPELAIKFACTSLEQVPPSTITV 771
eIF2C4 SDCHESSYCAIVVAVTSRQISQELLYSCVVIQDITMVRPELLIQVYKSTRFKPTRIIFVRCVSEGVFQVAVPELAIKFACTSLEQVPPSTITV 702
eIF2C1 SDCHESSYCAIVVAVTSRQISQELLYSCVVIQDITMVRPELLIQVYKSTRFKPTRIIFVRCVSEGVFQVAVPELAIKFACTSLEQVPPSTITV 704
eIF2C2 SDCHESSYCAIVVAVTSRQISQELLYSCVVIQDITMVRPELLIQVYKSTRFKPTRIIFVRCVSEGVFQVAVPELAIKFACTSLEQVPPSTITV 707
HIL1 SVAERITRVFSCRILQRTWT-----DVADCLVYKFCALNKKVKNHDLPARTIVRAGVGGQLRFLIBEVVQCLSSVASSSSMSSRLSVIV 641
HIM1 STNEQITRVFSCRILQRTWT-----ELVDGLKVCLEALRAVNSCNEYVSRIVVRCVSEGVFQVAVPELAIKFACTSLEQVPPSTITV 739
ruler710.....720.....730.....740.....750.....760.....770.....780.....790.....800

eIF2C3 MQRHHTRLFCADKNEVAGKGNIPAGTIVDNTIIL-HPSEDFYLCSEAGTQCTSPSHYVLDWDDKQTTADELILTYQLCHTAVRQTESVSTHAPAY 870
eIF2C4 MQRHHTRLFCADKNEVAGKGNIPAGTIVDNTIIL-HPSEDFYLCSEAGTQCTSPSHYVLDWDDKQTTADELILTYQLCHTAVRQTESVSTHAPAY 801
eIF2C1 MQRHHTRLFCADKNEVAGKGNIPAGTIVDNTIIL-HPSEDFYLCSEAGTQCTSPSHYVLDWDDKQTTADELILTYQLCHTAVRQTESVSTHAPAY 803
eIF2C2 MQRHHTRLFCADKNEVAGKGNIPAGTIVDNTIIL-HPSEDFYLCSEAGTQCTSPSHYVLDWDDKQTTADELILTYQLCHTAVRQTESVSTHAPAY 806
HIL1 VRKCKPFRFTEHN---RTVCAPELGMVDESEVTRNEVDQKYLISQVACRGVSPHYVLDWDDKQTTADELILTYQLCHTAVRQTESVSTHAPAY 737
HIM1 VRKRVNTRFPQSC---GRLEPILPGLMDESEVTRNEVDQKYLISQVACRGVSPHYVLDWDDKQTTADELILTYQLCHTAVRQTESVSTHAPAY 834
ruler810.....820.....830.....840.....850.....860.....870.....880.....890.....900

eIF2C3 AFLVAFRARIHLVQKESSEVSGSNGRDPALAKAVQIRHDDHTMHPA 924
eIF2C4 AFLVAFRARIHLVQKESSEVSGSNGRDPALAKAVQIRHDDHTMHPA 855
eIF2C1 AFLVAFRARIHLVQKESSEVSGSNGRDPALAKAVQIRHDDHTMHPA 857
eIF2C2 AFLVAFRARIHLVQKESSEVSGSNGRDPALAKAVQIRHDDHTMHPA 860
HIL1 AHHLIT-----LVQSISHEEP-----SLSLNHLNYL 764
HIM1 AHHLAF-----LVQSISHEEP-----NLSLSNRLNYL 861
ruler910.....920.....930.....940.....950.....

Fig. 16

>eIF2C1, cDNA sequence of predicted ORF
ATGGAAGCGGGACCCCTCGGGAGCAGCTGCGGGCGCTTACCTGCCCCCCTGCAGCAGGTGTT
CCAGGCACCTCGCCGGCCTGGCATTGGCACTGTGGGAAACCAATCAAGCTCCTGGCCATT
ACTTTGAGGTGGACATCCCTAAGATCGACGTGTACCCTACGAGGTGGACATCAAGCCGAT
AAGTGTCCCCGTAGAGTCAACCGGGAAGTGGTGAATACATGGTCCAGCATTTCAAGCCTCA
GATCTTTGGTGATCGCAAGCCTGTGTATGATGGAAAGAAGAACATTTACTACTGTCACAGCAC
TGCCCATTTGGCAACGAACGGGTCGACTTTGAGGTGACAATCCCTGGGGAAGGGAAGGATCGA
ATCTTTAAGGTCTCCATCAAGTGGCTAGCCATTGTGAGCTGGCGAATGCTGCATGAGGCCCT
GGTCAGCGGCCAGATCCCTGTCCCTTGGAGTCTGTGCAAGCCCTGGATGTGGCCATGAGGC
ACCTGGCATCCATGAGGTACACCCCTGTGGGCCGCTCCTTCTTCTCACCGCCTGAGGGCTAC
TACCACCCGCTGGGGGGTGGGCGCGAGGTCTGGTTCGGCTTTTACCAGTCTGTGCGCCCTGC
CATGTGGAAGATGATGCTCAACATTGATGTCTCAGCCACTGCCTTTTATAAGGCACAGCCAG
TGATTGAGTTCATGTGTGAGGTGCTGGACATCAGGAACATAGATGAGCAGCCCAAGCCCTC
ACGGACTCTCAGCGCGTTCGCTTACCAAGGAGATCAAGGGCCTGAAGGTGGAAGTCAECCA
CTGTGGACAGATGAAGAGGAAGTACCGCGTGTGTAATGTTACCCGTCGCCCTGCTAGCCATC
AGACATTCCCCTTACAGCTGGAGAGTGGACAGACTGTGGAGTGCACAGTGGCACAGTATTTT
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AAAAGCTGACCGACAACCAGACCTCGACCATGATAAAGGCCACAGCTAGATCCGCTCCAGAC
AGACAGGAGGAGATCAGTCGCTGATGAAGAATGCCAGCTACAACCTAGATCCCTACATCCA
GGAAATTTGGGATCAAAGTGAAGGATGACATGACGGAGGTGACAGGGCGAGTCTGCCGGCGC
CCATCTTGCAGTACGGCGCGCGGAACCGGGCCATTGCCACACCCAATCAGGGTGTCTGGGAC
ATGCCGGGGAAACAGTTCTACAATGGGATTGAGATCAAAGTCTGGGCCATCGCTTGCCTTGC
ACCCCAAAAACAGTGTGAGAGAGGTGCTCAAGAACTTACAGACCAGCTGCGGAAGATTT
CCAAGGATGCGGGGATGCCATCCAGGGTCAACCTTGTCTTCTGCAAAATGACACAGGGGCA
GACAGCGTGGAGCCTATGTTCCGGCATCTCAAGAACACCTACTCAGGGCTGCAGCTCATTAT
TGTTCATCCTGCCAGGGAAGACGCGCGTGTATGCTGAGGTGAAACGTGTCGGAGATACACTCT
TGGGAATGGCTACGCAGTGTGTGACGGTGAAGAACGTGGTCAAGACCTCACCTCAGACTCTG
TCCAACCTCTGCCCTCAAGATCAATGTCAAACCTGGTGGCATTAACAACATCCTAGTCCCACA
CCAGCGCTCTGCCGTTTTTCAACAGCCAGTGATATTCCTGGGAGCAGATGTTACACACCCCC
CAGCAGGGGATGGAAAAACCTTCTATCACAGCAGTGGTAGGCAGTATGGATGCCCCACCC
AGCCGATACTGTGCTACTGTGCGGGTACAGCGACCACGGCAAGAGATCATTGAAGACTTGT
CTACATGGTGCCTGAGCTCCTCATCCAATCTACAAGTCCACCCGTTTCAAGCCTACCCGCA
TCATCTTCTACCGAGATGGGGTGCCTGAAGGCCAGCTACCCAGATACTCCACTATGAGCTA
CTGGCCATTCGTGATGCCCTGCATCAAACCTGGAAGAAGACTACCAGCCTGGGATCACTTATAT
TGTGGTGCAGAAACGCCATCACACCCGCTTTTCTGTGCTGACAAGAATGAGCGAATTGGGA
AGAGTGGTAACATCCCAGCTGGGACCACAGTGGACACCAACATCACCCACCCATTTGAGTTT
GACTTCTATCTGTGCAGCCACGCAGGCATCCAGGGCACCAGCCGACCATCCCATTAATATGT
TCTTTGGGATGACAACCGTTTTACAGCAGATGAGCTCCAGATCCTGACGTACCAGCTGTGCC
ACACTTACGTACGATGCACACGCTCTGTCTATCCCAGCACCTGCCACTATGCCCGCCTG
GTGGCTTTCCGGGCACGATACCACCTGGTGGACAAGGAGCATGACAGTGGAGAGGGGAGCCA
CATATCGGGGCAGAGCAATGGGCGGGACCCCGAGCCCTGGCCAAAGCCGTGCAGGTTCCACC
AGGATACTCTGCGCACCATGTACTTCCGCT

Fig. 16 (con.)

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>eIF2C2, cDNA sequence of predicted ORF
ATGGGTGTTCTCTCTGCCATTCCCGCACTTGCACCTCCTGCGCCGCGCCCCCATCCAAGG
ATATGCCCTTCAAGCCTCCACCTAGACCCGACTTTGGGACCTCCGGGAGAACAATCAAATTAC
AGGCCAATTTCTTCGAAATGGACATCCCCAAAATGACATCTATCATTATGAATTGGATATC
AAGCCAGAGAAGTGCCCGAGGAGAGTTAACAGGGAAATCGTGGAAACACATGGTCCAGCACTT
TAAAACACAGATCTTTGGGGATCGGAAGCCCGTGTGACGGCAGGAAGAATCTATACACAG
CCATGCCCCCTTCCGATTGGGAGGGACAAGGTGGAGCTGGAGGTCACGCTGCCAGGAGAAGGC
AAGGATCGCATCTTCAAGGTGTCCATCAAGTGGGTGTCCTGCGTGAGCTTGCAGGCGTTACA
CGATGCACTTTTCAAGGCGGCTGCCCAGCGTCCCTTTTGAGACGATCCAGGCCCTGGACGTGG
TCATGAGGCACTTGCATCCATGAGGTACACCCCGTGGGCGGCTCCCTTCTTACCAGCGTCC
GAAGGCTGCTCTAACCCCTTTGGCGGGGGCCGAGAAGTGTGGTTTGGCTTCCATCAGTCCGT
CCGGCCTTCTCTCTGAAAATGATGCTGAATATGATGTGTGCAACAGCGTTTTTACAAGG
CACAGCCAGTAATCGAGTTTGTGTTGTGAAGTTTGGATTTTAAAAGTATTGAAGAACAACA
AAACCTCTGACAGATTCCCAAAGGGTAAAGTTTACCAAAGAAATTAAGGTCTAAAGGTGGA
GATAACGCACTGTGGGCAGATGAAGAGGAAGTACCGTGTCTGCAATGTGACCCGGCGGCCG
CCAGTCACCAAACATCCCGCTGCAGCAGGAGAGCGGGCAGACGGTGGAGTGCACGGTGGCC
CAGTATTTCAAGGACAGGCACAAGTTGGTTCTGCGCTACCCCCACCTCCCATGTTTACAAGT
CGGACAGGAGCAGAAACACACCTACCTTCCCCTGGAGGTCTGTAACATGTGGCAGGACAAA
GATGTATTAATAAATAACGGACAATCAGACCTCAACCATGATCAGAGCAACTGCTAGGTGCG
GCGCCCGATCGGCAAGAAGAGATTAGCAAAATGATGCGAAGTGCAAGTTTCAACACAGATCC
ATACGTCCGTGAATTTGGAATCATGGTCAAAGATGAGATGACAGACGTGACTGGGCGGGTGC
TGCAGCCGCCCTCCATCCTCTACGGGGGCAGGAATAAAGCTATTGCGACCCCTGTCCAGGGC
GTCTGGGACATGCGGAACAAGCAGTTCACACGGGCATCGAGATCAAGGTGTGGGCAATGCG
GTGCTTCGCCCCCAGCGCCAGTGCACGGAAAGTCCATCTGAAGTCCCTCACAGAGCAGCTCA
GAAAGATCTCGAGAGACGCTGGCATGCCATCCAGGGCCAGCCGTGCTTCTGCAATAACCGG
CAGGGGCGGACAGCGTGGAGCCCATGTTCCGGCACCTGAAGAACACGTATGCGGGCCTGCA
GCTGGTGGTGGTCATCCTGCCCGGCAAGACGCCCCGTGACGCCGAGGTCAAGCGCGTGGGAG
ACACGGTGTGGGATGGCCACGCAGTGCCTGTCAGATGAAGAACCCTGCAGAGGACCACGCCA
CAGACCCTGTCCAACCTTTGCCTGAAGATCAACGTCAAGCTGGGAGGGCTGAACAACATCCT
GCTGCCCCAGGGCAGGCCGCCGGTGTCCAGCAGCCCGTCACTTCTTCTGGGAGCAGACGTCA
CTCACCCCCCGCGGGGATGGGAAGAAGCCCTCCATTGCCGCCGTGGTGGGCAGCATGGAC
GCCCACCCCAATCGCTACTGCGCCACCCTGCGCGTGCAGCAGCACCCGGCAGGAGATCATAC
AGACCTGGCCGCCATGGTCCGCGAGCTCCTCATCCAGTCTACAAGTCCACGCGCTTCAAGC
CCACCCGCATCATCTTCTACCGCGACGGTGTCTCTGAAGGCCAGTTCAGCAGGTTCTCCAC
CACGAGTTGCTGGCCATCCGTGAGGCCTGTATCAAGCTAGAAAAGACTACCAGCCCGGGAT
CACCTTCATCGTGGTGCAGAAGAGGCACCACACCCGGCTCTTCTGCACTGACAAGAACGAGC
GGGTGGGAAAAGTGGAAACATCCAGCAGGCACGACTGTGGACACGAAAATCACCCACCCC
ACCGAGTTCGACTTCTACCTGTGTAGTCACGCTGGCATCCAGGGACAAAGCAGCCCTTCGCA
CTATCACGTCTCTGGGACGACATCGTTTCTCCTCTGATGAGCTECAGATCCTAACCTACC
AGCTGTGTCACACCTACGTGCGCTGCACACGCTCCGTGTCCATCCAGCGCCAGCACTACTAC
GCTCACCTGGTGGCCTTCCGGGCCAGGTACCACCTGGTGGATAAGGAACATGACAGTGTGA
AGGAAGCCATACCTCTGGGCAGAGTAACGGGCGAGACCACCAAGCACTGGCCAAGGCGGTCC
AGGTTACCAAGACACTCTGCGCACCATGTACTTTGCT
```

Fig. 16 (con.)

>eIF2C3, cDNA sequence of predicted ORF
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GTCTCCGCGGCGCCACCCAGCGCCAATATTCGGGAGATCAAGCGTTACGCGCGGCGGGCGG
CGGCGGCGGCGGGCCCGGAGCGGGAGGCGCCGGGGACCAGGGCGAGGCGGCCCCCGCCGCC
GCCATGGAGGCGCTGGGACCCGGACCTCCGGCTAGCCTGTTTCAGCCACCTCGTCGTCCTGG
CCTTGGAACCTGTTGAAAACCAATTCGACTGTTAGCCAATCATTTCAGGTCAGATTCCTA
AAATAGATGTGTATCACTATGATGTGGATATTAAGCCTGAAAAACGGCCTCGTAGAGTCAAC
AGGGAGGTAGTAGATACAATGGTGCAGGCACTTCAAGATGCAAAATATTTGGTGATCGGCAGCC
TGGGTATGATGGCAAAAGAAACATGTACACAGCACATCCACTACCAATGGACGGGATAGGG
TTGATATGGAGGTGACTCTTCCAGGCGAGGGTAAAGACCAACATTTAAAGTGTCTGTTCAG
TGGGTGTGAGTTGTGAGCCTTCAGTTGCTTTTAGAAGCTTTGGCTGGGCACCTGAATGAAGT
CCCAGATGACTCAGTACAAGCACTTGATGTTATCACAAGACACCTTCCCTCCATGAGGTACA
CCCCAGTGGGCGCTTCCCTTTTCTCACCCCGGAAGGTTACTACCACCTCTGGGAGGGGGC
AGGGAGGTCTGGTTGGTTTTTCATCAGTCTGTGAGACCTGCCATGTGGAATATGATGCTCAA
CATTGATGTATCTGCAACTGCTTTCTACCGGGCTCAGCCTATCATGAGTTCATGTGTGAGG
TTTTAGACATTCAGAACATCAATGAACAGACCAAACTCTAACAGACTCCCAGCGTGTCAA
TTTACCAAAGAAATCAGAGGTCTCAAAGTTGAGGTGACCCACTGTGGACAGATGAAACGAAA
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AAAACGGTCAAGCTATGGAATGTACAGTAGCTCAATATTTAAGCAAAAGTATAGTCTGCAA
CTGAAATACCCCCATCTTCCCTGTCTCCAAGTGGGACAAGAACAAGCATACATACTTGCC
ACTCGAGGTCTGTAATATAGTGGCAGGACAGCGATGTATCAAGAAGCTCACAGACAATCAGA
CTTCCACAATGATCAAAGCTACAGCAAGATCTGCTCCTGACAGACAGGAAGAGATCAGTAGA
CTGGTGAAGAGCAACAGTATGGTGGGTGGACCTGATCCATACTTAAAGAAATTTGGTATTGT
TGTCACAATGAAATGACAGAGCTCACAGGCAAGGTTACTTCCAGCACCAATGCTGCAATATG
GAGGCCGGAATAAAACAGTAGCCACACCCCAAGGCTGTCTGGGACATGCGAGGAAAGCAG
TTTTATGCTGGCATTGAAATTAAGTTTGGGCAAGTTGCTTCTTTTGCACCTCAGAAACAATG
TAGGGAAGATTTACTAAAGAGTTTCACTGACCAGCTGCGTAAATCTCTAAGGATGCAGGAA
TCCCCATCCAGGGTCAGCCATGTTTCTGCAAGTATGCACAAGGTGCAGACAGTGTGGAGCCT
ATGTTTAAACATCTGAAATGACTTATGTGGGCTACAGCTAATAGTGGTTATCCTGCCTGG
AAAGACACCAGTATATGCGGAGGTGAAACGTGTTGGAGATACCCCTTCTAGGTATGGCCACAC
AGTGTGTCAGGTAAAAATGTAGTGAAGACCTCACCTCAAACCTTTCCAATCTTTGCCTG
AAGATAAATGCAAAACTTGGAGGAATTAACAATGTGCTTGTGCCTCATCAAAGGCCCTCGGT
GTCCAGCAGCCTGTCATCTTCTGGGAGCGGATGTCACACACCCCCCAGCAGGGGATGGGA
AGAAACCTTCCATTGCTGCTGTGGTTGGCAGTATGGATGGCCACCCAGCCGGTACTGTGCC
ACCGTTCCGGTGCAGACTTCCCGCAGGAGATCTCCCAAGAGCTCCTCTACAGTCAAGAGGT
CATCCAGGACCTGACTAACATGGTTTCGAGAGCTGCTGATTCAGTTCACAAATCCACACGCT
TCAAACCCACTCGGATCATCTATTACCCTGGAGGGGTATCTGAGGGACAAATGAAACAGGTA
GCTTGGCCAGAACTAATAGCAATTCGAAAGGCATGTATTAGCTTGAAGAAGATTACCGGCC
AGGAATAACTTATATTGTGGTGCAAAAAGACATCACACACGACTCTTCTGTGCAGATAAAA
CAGAAAGGGTAGGGAAAAGTGGCAATGTACCAGCAGGCACTACAGTGGATAGTACCATCACA
CATCCATCTGAGTTTACTTTTACCTCTGTAGTCATGCAGGAATTCAGGGAAACCAGCCGTCC
CTCACATTACCAGGTCTTGTGGGATGACAACCTGCTTCACTGCAGATGAACTCCAGCTACTGA
CTTACCAGCTGTGTACACCTATGTGAGGTGCACTCGCTCAGTCTCTATTCCAGCCCCGTGA
TATTATGCCCGGCTTGTAGCATTTAGGGCAAGGTATCATCTGGTGGATAAAGATCATGACAG
TGCGGAAGGCAGTCATGTGTGAGGACAGAGCAACGGCCGGGATCCTCAGGCCCTGGCTAAGC
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Fig. 16 (con.)

>eIF2C4, cDNA sequence of predicted ORF
GCAGGACCCGCTGGGGCCCAGCCCCTACTCATGGTGGCCAGAAGACCTGGCTATGGCACCAT
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ACCTCTATGAGGTAGATATTAACCAGACAAGTGTCCTAGGAGAGTGAACAGGGAGGTGGTT
GACTCAATGGTTCAGCATTTTAAAGTAACTATATTTGGAGACCGTAGACCAGTTTATGATGG
AAAAAGAAGTCTTTACACCGCCAATCCACTTCCTGTGGCAACTACAGGGGTAGATTTAGACG
TTACTTTACCTGGGGAAGGTGGAAAAGATCGACCTTTCAAGGTGTCAATCAAATTTGTCTCT
CGGGTGAGTTGGCACCTACTGCATGAAGTACTGACAGGACGGACCTTGCCTGAGCCACTGGA
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CCTCCATGAAATACACACCTGTGGGGCGTTTCAATTTTCTCCGCTCCAGAAGGATATGACCAC
CCTCTGGGAGGGGGCAGGGAAGTGTGGTTTGGATTCCATCAGTCTGTTCCGGCCTGCCATGTG
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TCTCATCGGGTAAAATTCACCAAAGAGATAAAAAGGTTTGAAGGTTGAAGTGAAGTCAATTTGTGG
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GGGAAACAATTCACACAGGAGTTGAAATCAAATGTGGGCTATCGCTTGT'TTGGCCACACA
GAGGCAGTGCAGAGAAGAAATATTGAAGGGTTTACAGACCAGCTGCGTAAGATTTCTAAGG
ATGCAGGGATGCCCATCCAGGGCCAGCCATGCTTCTGCAAATATGCACAGGGGGCAGACAGC
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CCTGCCGGGGGAAGACACCAGTGTATCCGGAAGTGAACGTGTAGGAGACACACTTTTGGGTA
TGGCTACACAATGTGTTCAAGTCAAGAATGTAATAAAAACATCTCCTCAAACCTCTGTCAAAC
TTGTGCCTAAAGATAAATGTTAAACTCGGAGGGATCAATAATATTCTTGTACCTCATCAAAG
ACCTTCTGTGTTCCAGCAACCAGTGATCTTTTGGGAGCCGATGTCACTCATCCACCTGCTG
GTGATGGAAAGAAGCCTTCTATTTGCTGCTGTTGTAGGTAGTATGGATGCACACCCAAAGCAGA
TACTGTGCCACAGTAAGAGTTCAGAGACCCCGACAGGAGATCATCCAGGACTTGGCCTCCAT
GGTCCGGGAAC'TCTTATTCAATTTTATAAGTCAACTCGGTTCAAGCCTACTCGTATCATCT
TTTATCGGGATGGTGTTCAGAGGGGCAGTTTAGGCAGGTATTATATTATGAACTACTAGCA
ATTCGAGAAGCCTGCATCAGTTTGGAGAAAGACTATCAACCTGGAATAACCTACATTTGTAGT
TCAGAAGAGACATCACACTCGATTATTTTGTGCTGATAGGACAGAAAGGGTTGGAAGAAGTG
GCAATATCCCAGCTGGAACAACAGTTGATACAGACATTACACACCCATATGAGTTCGATTTT
TACCTCTGTAGCCATGCTGGAATACAGGGTACCAGTCGTCCTTACACTATCATGTTTTATG
GGATGATAACTGCTTTACTGCAGATGAACTTCAGCTGCTAACTTACCAGCTCTGCCACACTT
ACGTACGCTGTACACGATCTGTTTCTATACCTGCACCAGCGTATTATGCTCACCTGGTAGCA
TTTAGAGCCAGATATCATCTTGTGGACAAAGAACATGACAGTGTGAAGGAAGTCACGTTTC
AGGACAAAGCAATGGGCGAGATCCACAAGCTCTTGCCAAGGCTGTACAGATTCACCAAGATA
CCTTACGCACAATGTACTTTCGCTTAA

Fig. 16 (con).

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>HILI, cDNA sequence of predicted ORF
ATATCTTCTGGTGATGCTGGAAGTACCTTCATGGAAAGAGGTGTGAAAAACAAACAGGACTT
TATGGATTTGAGTATCTGTACCAGAGAAAAATTGGCACATGTGAGAAATTGTPAAAACAGGTT
CCAGTGGAATACCTGTGAACTGGTTACAAAACCTCTTTAACTTAGATTTTCCCAAGACTGG
CAGCTATACCAGTACCATGTGACATATATTCAGATTTAGCATCTAGAAGGCTGAGAATTGC
TTTACTTTATAGTCATAGTGAACCTTCCAACAAAAGCAAAAGCATTTCGACGGTCCCATCCTTT
TTCTGTCACAAAAGCTAGAAGAAAAGGTCACAGAGTTGTCAAGTGAACTCAAAGAGGTGAG
ACTATAAAGATGACTATCACCCCTGAAGAGGGAGCTGCCATCAAGTTCTCCCGTGTGCATCCA
GGTCTTCAATATCATCTTCAGAAAGATCCTCAAAAAGTTGTCCATGTACCAAATGGACGGA
ACTTCTATAATCCTTCAGAGCCAATGGAAATTCCCCAGCACAAATATCCCTTGGCCTGGG
TTTGCCATTTCTGTGTCATATTTTGAAGGAAGCTCCTGTTTAGTGCTGATGTGAGTTACAA
AGTCCCTCCGGAATGAGACGGTCTGGAATTCATGACTGCTCTCTGTCAAAGAACTGGCTTGT
CCTGTTTCACCCAGACGTGTGAGAAGCAGCTAATAGGGCTCATTGTCCTTACAAGATACAAT
AACAGAACCCTACTCCATGATGACATGACTGGTCAGTGAAGCCCACACACACCTTTCAGAA
GCGGGATGGCACCGAGATCACCTATGTGGATTAACAAGCAGCAGTATGATATTACTGTAT
CGGACCTGAATCAGCCCATGCTTGTAGTCTGTAAAGAAGAAGAGAAATGACAACAGTGAG
GCTCAGCTCGCCCACCTGATACCTGAGCTCTGCTTCTAACAGGGCTGACTGACCAGGCAAC
ATCTGATTTCCAGCTGATGAAGGCTGTGGCTGAAAAGACACGTCTCAGTCCCTCAGGCCGGC
AGCAGCGCCTGGCCAGGCTTGTGGACAACATCCAGAGGAATACCAATGCTCGCTTGAACATA
GAGACCTGGGGACTGCATTTTGGAAAGCCAGATATCTCTGACTGGCCGGATTGTGCCTTCAGA
AAAAATATTAATGCAAGACCACATATGTCAACCTGTGTCTGCTGCTGACTGGTCCAGGATA
TTCGAACTTGCAAGATTTTAAATGCACAGTCTTGAATACCTGGTTGATTTATGTAGCGAC
AGAACTGAATATGTTGCCGAGAGCTTCTGAACCTGCTTGAGAAGAGTTGCAGGTTCCATGGG
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GGCATGATGATGAGTATCGCCACCAAGATCGCTATGCAGATGACTTGCAAGCTCGGAGGCGA
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TTTCATGACTGGAGCACTCAACAAATGGTACAAGTACAATCATGATTTGCCAGCACGGATAA
TTGTGTACCGTGCTGGTGTAGGGGATGGTCAGCTGAAAACACTTATTGAATATGAAGTCCCA
CAGCTGCTGAGCAGTGTGGCAGAATCCAGCTCAAATACCAGCTCAAGACTGTCCGGTATTGT
GGTCAGGAAGAAGTGCATGCCACGATTCTTTACCAGAAATGAACCGCACTGTACAGAACCCCC
CACTTGGCACTGTTGTGGATTCAGAAGCAACACGTAACGAATGGCAGTATGACTTTTATCTG
ATCAGCCAGGTGGCTTCCCGGGAACTGTTAGTCCACTACTATAATGTCATCTATGATGA
CAACGGCTTGAAGCCCGACCATATGCAGAGACTTACATTCAAATGTGCCACCTGTACTACA
ACTGGCCGGGCATAGTCAGTGTCCCAGCACCATGTCAGTATGCTCACAAGCTGACCTTCTG
GTGGCACAAAGCATTTCATAAAGAACCCAGTCTGGAATTAGCCAACCATCTCTTCTACCTG
```

Fig. 16 (con.)

>HIWI, cDNA sequence of predicted ORF
ATGACTGGGAGAGCCCGAGCCAGAGCCAGAGGAAGGGCCCCGCGGTCAGGAGACAGCGCAGCT
GGTGGGCTCCACTGCCAGTCAGCAACCTGGTTATATTCAGCCTAGGCCTCAGCCGCCACCAG
CAGAGGGGGAATTATTTGGCCGTGGACGGCAGAGAGGAACAGCAGGAGGAACAGCCAAAGTCA
CAAGGACTCCAGATATCTGCTGGATTTTCAGGAGTTATCGTTAGCAGAGAGAGGAGGTCGTCG
TAGAGATTTTCATGATCTTGGTGTGAATACAAGGCAGAACCCTAGACCATGTTAAAGAAATCAA
AAACAGGTTCTTCAGGCATTATAGTAAGGTTAAGCACTAACCATTTCCGGCTGACATCCCCT
CCCCAGTGGGCCTTATATCAGTATCACATTGACTATAACCCACTGATGGAAGCCAGAAGACT
CCGTTTCAGCTCTTCTTTTCAACACGAAGATCTAATTGGAAAGTGCCATGCTTTTGATGGAA
CGATATTATTTTACCTAAAAGACTACAGCAAAAGGTTACTGAAGTTTTAGTAAGACCCGG
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TTGTTTGCAGTTCTATAATATTATTTTCAGGAGGCTTTTGAAAATCATGAATTTGCAACAAA
TTGGACGAAATTTATTATAACCCAAATGACCCAATTGATATTTCCAAGTCACAGGTTGGTGATT
TGGCCTGGCTTCACTACTTCCATCCTTCAGTATGAAAACAGCATCATGCTCTGCACTGACGT
TAGCCATAAAGTCCTTCGAAGTGAGACTGTTTTGGATTTTCATGTTCAACTTTTATCATCAGA
CAGAAGAACATAAATTTCAAGAACAAGTTTTCCAAAGAACTAATAGGTTTAGTTGTTCTTACC
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CTTTAAGAAAGCCGACGGCTCTGAAGTCAGCTTCTTAGAATACTACAGGAAGCAATACAACC
AAGAGATCACCGACTTGAAGCAGCCTGTCTTGGTCAGCCAGCCCAAGAGAAGGCGGGGCCCT
GGGGGACACTGCCAGGGCCTGCCATGCTCATTCCTGAGCTCTGCTATCTTACAGGTTAAC
TGATAAAAATGCGTAATGATTTTAAACGTGATGAAAGACTTAGCCGTTTCATACAAGACTAACTC
CAGAGCAAAGGCAGCGTGAAGTGGGACGACTCATGATTACATTCATAAAAACGATAATGTT
CAAAGGGAGCTTCGAGACTGGGGTTTTGAGCTTTGATTCCAACCTTACTGTCCCTTCTCAGGAAG
AATTTTGCAACAGAAAAGATTCACCAAGGTGGAAAAACATTTGATTACAATCCACAATTTG
CAGATTGGTCCAAAGAAAACAAGAGGTGCACCATTAATTAGTGTTAAGCCACTAGATAACTGG
CTGTTGATCTATACGCGAAGAAATTTATGAAGCAGCCAATTCATTGATACAAAATCTATTTAA
AGTTACACCAGCCATGGGCATGCAATGAGAAAAGCAATAATGATTGAAGTGGATGACAGAA
CTGAAGCCTACTTAAGAGTCTTACAGCAAAAGGTCACAGCAGACACCCAGATAGTTGTCTGT
CTGTTGTCAAGTAATCGGAAGGACAAATACGATGCTATTAAAAAATACCTGTGTACAGATTG
CCCTACCCCAAGTCAGTGTGTGTTGGCCCGAACCTTAGGCAAAACAGCAACTGTTCATGGCCA
TTGCTACAAAGATTGCCCTACAGATGAACTGCAAGATGGGAGGAGAGCTCTGGAGGGTGGAC
ATCCCCCTGAAGCTCGTGATGATCGTTGGCATCGATTGTTACCATGACATGACAGCTGGGCG
GAGGTCAATCGCAGGATTTGTTGCCAGCATCAATGAAGGGATGACCCGCTGGTTCTCACGCT
GCATATTTTCAGGATAGAGGACAGGAGCTGGTAGATGGGCTCAAAGTCTGCCTGCAAGCGGCT
CTGAGGGCTTGGAAATAGCTGCAATGAGTACATGCCAGCCGGATCATCGTGTACCGGATGG
CGTAGGAGACGGCCAGCTGAAAACACTGGTGAACCTACGAAGTGCCACAGTTTTTTGGATTGTC
TAAAATCCATTGGTAGAGGTTACAACCTAGACTAACGGTAATTTGTGGTGAAGAAAAGAGTG
AACACCAGATTTTTTGCTCAGTCTGGAGGAAGACTTCAGAATCCACTTCCTGGAACAGTTAT
TGATGTAGAGGTTACCAGACCAGAATGGTATGACTTTTTTATCGTGAGCCAGGCTGTGAGAA
GTGGTAGTGTCTCTCCACACATTACAATGTCATCTATGACAACAGCGGCCCTGAAGCCAGAC
CACATACAGCGCTTGACCTACAAGCTGTGCCACATCTATTACAACCTGGCCAGGTGTCATTCG
TGTTCCCTGCTCCTTGCCAGTACGCCACAAGCTGGCTTTTCTTGTGGCCAGAGTATTCACA
GAGAGCCAAATCTGTCACTGTCAAACCGCCTTTACTACCTC

Fig. 17 A

Gene name	1 st primer pair (5'-3')	2 nd primer pair (5'-3')	Expected length (bp)
(SEQ ID NO: 80)	GAGGTCTGTAAACATTGTGGC*	GAGGTCTGTAAACATTGTGGC*	287
(SEQ ID NO: 81)	CGGTAGAAGATGATGCCGGT	AAGTTCTTGAGCACCTCTTCTCGA	
(SEQ ID NO: 82)	GAGGTCTGTAAACATTGTGGC	CCACACCAGCGCTCTGCCC	207
(SEQ ID NO: 85)	CGGTAGAAGATGATGCCGGT	CTCACCACCCATGTAGGA	
(SEQ ID NO: 88)	GAGGTCTGTAAACATTGTGGC	ATCCTGCTGCCCCCAAGGG	186
(SEQ ID NO: 89)	CGGTAGAAGATGATGCCGGT	GATCTCTGCCCCCTGCTG	
(SEQ ID NO: 92)	GAGGTCTGTAAACATTGTGGC*	GAGGTCTGTAAACATTGTGGC*	891
(SEQ ID NO: 93)	CGGTAGAAGATGATGCCGGT	GATCTCTGCCCCCTGCTG	
(SEQ ID NO: 96)	AGAGCAKCAATGATGGTGGTGGAC	CCTCTACAGTCAAGAGGT	334
(SEQ ID NO: 97)	TGGATGTGTGATGGTACT*	TGGATGTGTGATGGTACT*	
(SEQ ID NO: 100)	CACTTGAATGAATGCCA	AGAGCAKCAATGATGGTGGTGGAC	808
(SEQ ID NO: 101)	TCCTGGATGACCTCTGACTGTAG*	TCCTGGATGACCTCTGACTGTAG*	
(SEQ ID NO: 104)	TCCGGCATCTCAAGAACACATATTCT	ATCCACGACTTCCCTCC	324
(SEQ ID NO: 105)	GAATCTCATGCGGTGTAAATGTCTG*	GAATCTCATGCGGTGTAAATGTCTG*	
(SEQ ID NO: 108)	CAGCACAAATTAATCCCTT*	CAGCACAAATTAATCCCTT*	264
(SEQ ID NO: 109)	CGGCCTGAAGGACTGAGACGTGT	GTGTGTGGGCTTCACTGA	
(SEQ ID NO: 112)	TCTCTGTCAAAGACTGGCTTGTCTT*	TCTCTGTCAAAGACTGGCTTGTCTT*	393
(SEQ ID NO: 113)	CTGTACAGTCCGGTTCAT	CGGCCTGAAGGACTGAGACGTGT	

* primers used in both reactions (semi-nested PCR)

Fig. 17 B

Gene name	eIF2C1		eIF2C2		eIF2C3		eIF2C4	HILI	
Expected length (bp)	287	207	186	891	808	334	324	264	393
PCR products									

RNA-INTERFERENCE BY SINGLE-STRANDED RNA MOLECULES

This application is a divisional of U.S. Ser. No. 14/337,710, filed Jul. 22, 2014, which is a continuation of U.S. Ser. No. 13/329,710 filed Dec. 19, 2011, which is a divisional of U.S. Ser. No. 10/520,470 filed Jan. 7, 2005, now U.S. Pat. No. 8,101,348 issued Jan. 24, 2012, which is a 35 U.S.C. 371 National Phase Entry Application from PCT/EP2003/007516, filed Jul. 10, 2003, which claims the benefit of European Patent Application Nos. 02015532.1 filed Jul. 10, 2002 and 02018906.4 filed Aug. 23, 2002, the disclosures of which are incorporated herein in their entirety by reference.

DESCRIPTION

The present invention relates to sequence and structural features of single-stranded (ss)RNA molecules required to mediate target-specific nucleic acid modifications by RNA-interference (RNAi), such as target mRNA degradation and/or DNA methylation.

Most eukaryotes possess a cellular defense system protecting their genomes against invading foreign genetic elements. Insertion of foreign elements is believed to be generally accompanied by formation of dsRNA that is interpreted by the cell as a signal for unwanted gene activity (e.g. 5 Ahlquist, *Science* 296 (2002), 1270-1273; Fire et al., *Nature* 391 (1998), 806-811). Dicer RNase III rapidly processes dsRNA to small dsRNA fragments of distinct size and structure (e.g. Bernstein et al., *Nature* 409 (2001), 363-366), the small interfering RNAs (siRNAs) (Elbashir et al., *Genes & Dev.* 15 (2001 b), 188-200), which direct the sequence-specific degradation of the single-stranded mRNAs of the invading genes. siRNA duplexes have 2- to 3-nt 3' overhanging ends and contain 5' phosphate and free 3' hydroxyl termini (WO 02/44321). The process of posttranscriptional dsRNA-dependent gene silencing is commonly referred to as RNA interference (RNAi), and in some instances is also linked to transcriptional silencing.

Experimental introduction of siRNA duplexes into mammalian cells is now widely used to disrupt the activity of cellular genes homologous in sequence to the introduced dsRNA. Used as a reverse genetic approach, siRNA-induced gene silencing accelerates linking of gene sequence to biological function. siRNA duplexes are short enough to bypass general dsRNA-induced unspecific effects in vertebrate animal and mammalian cells. siRNAs may also be expressed intracellularly from introduced expression plasmids or viral vectors providing an alternative to chemical RNA synthesis. Therefore, an understanding of how siRNAs act in mammalian systems is important for refining this gene silencing technology and for producing gene-specific therapeutic agents.

Biochemical studies have begun to unravel the mechanistic details of RNAi. The first cell-free systems were developed using *D. melanogaster* cell or embryo extracts, and were followed by the development of in vitro systems from *C. elegans* embryo and mouse embryonal carcinoma cells. While the *D. melanogaster* lysates support the steps of dsRNA processing and sequence-specific mRNA targeting, the latter two systems only recapitulate the first step.

RNAi in *D. melanogaster* extracts is initiated by ATP-dependent processing of long dsRNA to siRNAs by Dicer RNase III (e.g. Bernstein et al., (2001), supra). Thereafter, siRNA duplexes are assembled into a multi-component

(2001 b), supra). This complex is referred to as RNA-induced silencing complex (RISC) (Hammond et al., *Nature* 404 (2000), 293-296). siRNAs in *D. melanogaster* are predominantly 21- and 22-nt, and when paired in a manner to contain a 2-nt 3' overhanging structure effectively enter RISC (Elbashir et al., *EMBO J.* 20 (2001 c), 6877-6888). Mammalian systems have siRNAs of similar size, and siRNAs of 21- and 22-nt also represent the most effective sizes for silencing genes expressed in mammalian cells (e.g. Elbashir et al., *Nature* 411 (2001 a), 494-498, Elbashir et al., *Methods* 26 (2002), 199-213).

RISC assembled on siRNA duplexes in *D. melanogaster* embryo lysate targets homologous sense as well as antisense single-stranded RNAs for degradation. The cleavage sites for sense and antisense target RNAs are located in the middle of the region spanned by the siRNA duplex. Importantly, the 5'-end, and not the 3'-end, of the guide siRNA sets the ruler for the position of the target RNA cleavage. Furthermore, a 5' phosphate is required at the target-complementary strand of a siRNA duplex for RISC activity, and ATP is used to maintain the 5' phosphates of the siRNAs (Nykanen et al., *Cell* 107 (2001), 309-321). Synthetic siRNA duplexes with free 5' hydroxyls and 2-nt 3' overhangs are so readily phosphorylated in *D. melanogaster* embryo lysate that the RNAi efficiencies of 5'-phosphorylated and non-phosphorylated siRNAs are not significantly different (Elbashir et al. (2001 c), supra).

Unwinding of the siRNA duplex must occur prior to target RNA recognition. Analysis of ATP requirements revealed that the formation of RISC on siRNA duplexes required ATP in lysates of *D. melanogaster*. Once formed, RISC cleaves the target RNA in the absence of ATP. The need for ATP probably reflects the unwinding step and/or other conformational rearrangements. However, it is currently unknown if the unwound strands of an siRNA duplex remain associated with RISC or whether RISC only contains a single-stranded siRNA.

A component associated with RISC was identified as Argonaute2 from *D. melanogaster* Schneider 2 (S2) cells (Hammond et al., *Science* 293 (2001 a), 1146-1150), and is a member of a large family of proteins. The family is referred to as Argonaute or PPD family and is characterized by the presence of a PAZ domain and a C-terminal Piwi domain, both of unknown function (Cerutti et al., *Trends Biochem. Sci.* (2000), 481-482); Schwarz and Zamore, *Genes & Dev.* 16 (2002), 1025-1031). The PAZ domain is also found in Dicer. Because Dicer and Argonaute2 interact in S2 cells, PAZ may function as a protein-protein interaction motif. Possibly, the interaction between Dicer and Argonaute2 facilitates siRNA incorporation into RISC. In *D. melanogaster*, the Argonaute family has five members, most of which were shown to be involved in gene silencing and development. The mammalian members of the Argonaute family are poorly characterized, and some of them have been implicated in translational control, microRNA processing and development. The biochemical function of Argonaute proteins remains to be established and the development of more biochemical systems is crucial.

Here we report on the analysis of human RISC in extracts prepared from HeLa cells. The reconstitution of RISC and the mRNA targeting step revealed that RISC is a ribonucleo-protein complex that is composed of a single-stranded siRNA. Once RISC is formed the incorporated siRNA can no longer exchange with free siRNAs. Surprisingly, RISC can be reconstituted in HeLa S100 extracts providing single-stranded siRNAs. Introducing 5' phosphorylated single-

stranded antisense siRNAs into HeLa cells potently silences an endogenous gene with similar efficiency than duplex siRNA.

The object underlying the present invention is to provide novel agents capable of mediating target-specific RNAi.

The solution of this problem is provided by the use of a single-stranded RNA molecule for the manufacture of an agent for inhibiting the expression of said target transcript. Surprisingly, it was found that single-stranded RNA molecules are capable of inhibiting the expression of target transcripts by RNA-interference (RNAi).

The length of the single-stranded RNA molecules is preferably from 14-50 nt, wherein at least the 14 to 20 5'-most nucleotides are substantially complementary to the target RNA transcript. The RNA oligonucleotides may have a free 5' hydroxyl moiety, or a moiety which is 5' phosphorylated (by means of chemical synthesis or enzymatic reactions) or which is modified by 5'-monophosphate analogues.

The inhibition of target transcript expression may occur in vitro, e.g. in eucaryotic, particularly mammalian cell cultures or cell extracts. On the other hand, the inhibition may also occur in vivo i.e. in eucaryotic, particularly mammalian organisms including human beings.

Preferably, the single-stranded RNA molecule has a length from 15-29 nucleotides. The RNA-strand may have a 3 hydroxyl group. In some cases, however, it may be preferable to modify the 3' end to make it resistant against 3' to 5' exonucleases. Tolerated 3'-modifications are for example terminal 2'-deoxy nucleotides, 3' phosphate, 2',3'-cyclic phosphate, C3 (or C6, C7, C12) aminolinker, thiol linkers, carboxyl linkers, non-nucleotidic spacers (C3, C6, C9, C12, abasic, triethylene glycol, hexaethylene glycol), biotin, fluoresceine, etc.

The 5'-terminus comprises an OH group, a phosphate group or an analogue thereof. Preferred 5' phosphate modifications are 5'-monophosphate $\llcorner(\text{HO})_2(\text{O})\text{P}-\text{O}-5'$), 5'-diphosphate $((\text{HO})_2(\text{O})\text{P}-\text{O}-\text{P}(\text{HO})(\text{O})-\text{O}-5')$, 5'-triphosphate $((\text{HO})_2(\text{O})\text{P}-\text{O}-\text{P}(\text{HO})(\text{O})-\text{O}-\text{P}(\text{HO})(\text{O})-\text{O}-5')$, 5'-guanosine cap (7-methylated or non-methylated) $(7\text{m-G-O-}5'-(\text{HO})(\text{O})\text{P}-\text{O}-\text{P}(\text{HO})(\text{O})-\text{O}-5')$, 5'-adenosine cap (Appp), and any modified or unmodified nucleotide cap structure $(\text{N}-\text{O}-5'-(\text{HO})(\text{O})\text{P}-\text{O}-\text{P}(\text{HO})(\text{O})-\text{O}-5')$, 5'-monothiophosphate (phosphorothioate; $(\text{HO})_2(\text{S})\text{P}-\text{O}-5'$), 5'-monodithiophosphate (phosphorodithioate; $(\text{HO})(\text{HS})(\text{S})\text{P}-\text{O}-5'$), 5'-phosphorothiolate $((\text{HO})_2(\text{O})\text{P}-\text{S}-5')$; any additional combination of oxygen/sulfur replaced monophosphate, diphosphate and triphosphates (e.g. 5'-alpha-thiotriphosphate, 5'-gamma-thiotriphosphate, etc.), 5'-phosphoramidates $((\text{HO})_2(\text{O})\text{P}-\text{NH}-5'-(\text{HO})(\text{NH}_2)(\text{O})\text{P}-\text{O}-5')$, 5'-alkylphosphonates (R=alkyl=methyl, ethyl, isopropyl, propyl, etc., e.g. $\text{RP}(\text{OH})(\text{O})-\text{O}-5'$ -, $(\text{OH})_2(\text{O})\text{P}-5'$ - CH_2-), 5'-alkyletherphosphonates (R=alkylether=methoxymethyl (MeOCH_2-), ethoxymethyl, etc., e.g. $\text{RP}(\text{OH})(\text{O})-\text{O}-5'$ -).

The sequence of the RNA molecule of the present invention has to have a sufficient identity to a nucleic acid target molecule in order to mediate target-specific RNAi. Thus the single-stranded RNA molecule of the present invention is substantially complementary to the target transcript.

The target RNA cleavage reaction guided by the single-stranded RNA molecules of the present invention is highly sequence-specific. However, not all positions of the RNA molecule contribute equally to target recognition. Mismatches, particularly at the 3'-terminus of the single-stranded RNA molecule, more particularly the residues 3' to the first 20 nt of the single-stranded RNA molecule are

tolerated. Especially preferred are single-stranded RNA molecules having at the 5'-terminus at least 15 and preferably at least 20 nucleotides which are completely complementary to a predetermined target transcript or have at only mismatch and optionally up to 35 nucleotides at the 3'-terminus which may contain 1 or several, e.g. 2, 3 or more mismatches.

In order to enhance the stability of the single-stranded RNA molecules, the 3'-ends may be stabilized against degradation, e.g. they may be selected such that they consist of purine nucleotides, particularly adenosine or guanosine nucleotides. Alternatively or additionally, 3'nucleotides may be substituted by modified nucleotide analogues, including backbone modifications of ribose and/or phosphate residues.

In an especially preferred embodiment of the present invention the RNA molecule may contain at least one modified nucleotide analogue. The nucleotide analogues may be located at positions where the target-specific activity, e.g. the RNAi mediating activity is not substantially affected, e.g. in a region at the 5'-end and/or the 3'-end of the RNA molecule. Particularly, the 3'-terminus may be stabilized by incorporating modified nucleotide analogues, such as non-nucleotidic chemical derivatives such as C3 (or C6, C7, C12) aminolinker, thiol linkers, carboxyl linkers, non-nucleotidic spacers (C3, C6, C9, C12, abasic, triethylene glycol, hexaethylene glycol), biotin, fluoresceine, etc. A further modification, by which the nuclease resistance of the RNA molecule may be increased, is by covalent coupling of inverted nucleotides, e.g. 2'-deoxyribonucleotides or ribonucleotides to the 3'-end of the RNA molecule. A preferred RNA molecule structure comprises: 5'-single-stranded siRNA-3'-O—P(O)(OH)—O-3'-N, wherein N is a nucleotide, e.g. a 2'-deoxyribonucleotide or ribonucleotide, typically an inverted thymidine residue, or an inverted oligonucleotide structure, e.g. containing up to 5 nucleotides.

Preferred nucleotide analogues are selected from sugar- or backbone-modified ribonucleotides. It should be noted, however, that also nucleobase-modified ribonucleotides, i.e. ribonucleotides, containing a non-naturally occurring nucleobase instead of a naturally occurring nucleobase such as uridines or cytidines modified at the 5-position, e.g. 5-(2-amino)propyl uridine, 5-bromo uridine; 5-methyl-cytidine; adenosines and guanosines modified at the 8-position, e.g. 8-bromo guanosine; deaza nucleotides, e.g. 7-deaza-adenosine; O- and N-alkylated nucleotides, e.g. N6-methyl adenosine are suitable. In preferred sugar-modified ribonucleotides the 2' OH-group is replaced by a group selected from H, OR, R, halo, SH, SR, NH_2 , NHR, NR_2 or CN, wherein R is C_1 , C_6 alkyl, alkenyl, alkynyl or methoxyethoxy, and halo is F, Cl, Br or I. In preferred backbone-modified ribonucleotides the phosphoester group connecting to adjacent ribonucleotides is replaced by a modified group, e.g. a phosphorothioate, phosphorodithioate, N3'-O5'- and/or N5'-O3' phosphoramidate group. It should be noted that the above modifications may be combined. For example, complementary or non-complementary nucleotides at the 3'-terminus, particularly after at least 15, more particularly after at least 20 5'-terminal nucleotides may be modified without significant loss of activity.

The single-stranded RNA molecule of the invention may be prepared by chemical synthesis. Methods of synthesizing RNA molecules are known in the art.

The single-stranded RNAs can also be prepared by enzymatic transcription from synthetic DNA templates or from DNA plasmids isolated from recombinant bacteria and sub-

sequent 5'-terminal modification. Typically, phage RNA polymerases are used such as T7, T3 or SP6 RNA polymerase.

A further aspect of the present invention relates to a method of mediating RNA interference in a cell or an organism comprising the steps:

(a) contacting the cell or organism with the single-stranded RNA molecule of the invention under conditions wherein target-specific nucleic acid modifications may occur and

(b) mediating a target-specific nucleic acid modification effected by the single-stranded RNA towards a target nucleic acid having a sequence portion substantially complementary to the single-stranded RNA.

Preferably the contacting step (a) comprises introducing the single-stranded RNA molecule into a target cell, e.g. an isolated target cell, e.g. in cell culture, a unicellular microorganism or a target cell or a plurality of target cells within a multicellular organism. More preferably, the introducing step comprises a carrier-mediated delivery, e.g. by liposomal carriers and/or by injection. Further suitable delivery systems include Oligofectamine (Invitrogen) and Transit-TKO siRNA Transfection reagent (Mirus)

The method of the invention may be used for determining the function of a gene in a cell or an organism or even for modulating the function of a gene in a cell or an organism, being capable of mediating RNA interference.

The cell is preferably a eukaryotic cell or a cell line, e.g. a plant cell or an animal cell, such as a mammalian cell, e.g. an embryonic cell, a pluripotent stem cell, a tumor cell, e.g. a teratocarcinoma cell or a virus-infected cell. The organism is preferably a eukaryotic organism, e.g. a plant or an animal, such as a mammal, particularly a human.

The target gene to which the RNA molecule of the invention is directed may be associated with a pathological condition. For example, the gene may be a pathogen-associated gene, e.g. a viral gene, a tumor-associated gene or an autoimmune disease-associated gene. The target gene may also be a heterologous gene expressed in a recombinant cell or a genetically altered organism. By determining or modulating, particularly, inhibiting the function of such a gene valuable information and therapeutic benefits in the agricultural field or in the medicine or veterinary medicine field may be obtained.

The ssRNA is usually administered as a pharmaceutical composition. The administration may be carried out by known methods, wherein a nucleic acid is introduced into a desired target cell in vitro or in vivo. Commonly used gene transfer techniques include calcium phosphate, DEAE-dextran, electroporation and microinjection and viral methods (Graham, F. L. and van der Eb, A. J. (1973) *Virol.* 52, 456; McCutchan, J. H. and Pagano, J. S. (1968), *J. Natl. Cancer Inst.* 41, 351; Chu, G. et al (1987), *Nucl. Acids Res.* 15, 1311; Fraley, R. et al. (1980), *J. Biol. Chem.* 255, 10431; Capecchi, M. R. (1980), *Cell* 22, 479). A recent addition to this arsenal of techniques for the introduction of nucleic acids into cells is the use of cationic liposomes (Feigner, P. L. et al. (1987), *Proc. Natl. Acad. Sci USA* 84, 7413). Commercially available cationic lipid formulations are e.g. Tfx 50 (Promega) or Lipofectamin2000 (Life Technologies). A further preferred method for the introduction of RNA into a target organism, particularly into a mouse, is the high-pressure tail vein injection (Lewis, D. L. et al. (2002), *Nat.Genet.* 29, 29; McCaffrey, A. P. et al. (2002), *Nature* 418, 38-39).

Herein, a buffered solution comprising the single-stranded RNA (e.g. about 2 ml) is injected into the tail vein of the mouse within 10 s.

Thus, the invention also relates to a pharmaceutical composition containing as an active agent at least one single-stranded RNA molecule as described above and a pharmaceutical carrier. The composition may be used for diagnostic and for therapeutic applications in human medicine or in veterinary medicine.

For diagnostic or therapeutic applications, the composition may be in form of a solution, e.g. an injectable solution, a cream, ointment, tablet, suspension or the like. The composition may be administered in any suitable way, e.g. by injection, by oral, topical, nasal, rectal application etc. The carrier may be any suitable pharmaceutical carrier. Preferably, a carrier is used, which is capable of increasing the efficacy of the RNA molecules to enter the target-cells. Suitable examples of such carriers are liposomes, particularly cationic liposomes. A further preferred administration method is injection.

A further preferred application of the RNAi method is a functional analysis of eukaryotic cells, or eukaryotic non-human organisms, preferably mammalian cells or organisms and most preferably human cells, e.g. cell lines such as HeLa or 293 or rodents, e.g. rats and mice. By transfection with suitable single-stranded RNA molecules which are homologous to a prede-termined target gene or DNA molecules encoding a suitable single-stranded RNA molecule a specific knockout phenotype can be obtained in a target cell, e.g. in cell culture or in a target organism. The presence of short single-stranded RNA molecules does not result in an interferon response from the host cell or host organism.

In an especially preferred embodiment, the RNA molecule is administered associated with biodegradable polymers, e.g. polypeptides, poly(d,l-lactic-co-glycolic acid) (PLGA), polylysine or polylysine conjugates, e.g. polylysine-graft-imidazole acetic acid, or poly(beta-amino ester) or microparticles, such as microspheres, nanoparticles or nanospheres. More preferably the RNA molecule is covalently coupled to the polymer or microparticle, wherein the covalent coupling particularly is effected via the 3'-terminus of the RNA molecule.

Further, the invention relates to a pharmaceutical composition for inhibiting the expression of a target transcript by RNAi comprising as an active agent a single-stranded RNA molecule having a length from 14-50, preferably 15-29 nucleotides wherein at least the 14-20 5'most nucleotides are substantially complementary to said target transcript.

Furthermore, the invention relates to a method for the prevention or treatment of a disease associated with overexpression of at least one target gene comprising administering a subject in need thereof a single-stranded RNA molecule having a length from 14-50, preferably 15-29 nucleotides wherein at least the 14-20 5'most nucleotides are substantially complementary to a target transcript in an amount which is therapeutically effective for RNAi.

Still, a further subject matter of the invention is a eukaryotic cell or a eukaryotic non-human organism exhibiting a target gene-specific knockout phenotype comprising an at least partially deficient expression of at least one endogenous target gene wherein said cell or organism is transfected with at least one single-stranded RNA molecule capable of inhibiting the expression of at least one endogenous target gene. It should be noted that the present invention allows the simultaneous delivery of several antisense RNAs of different sequences, which are either cognate to a different or the same target gene.

Gene-specific knockout phenotypes of cells or non-human organisms, particularly of human cells or non-human mammals may be used in analytic procedures, e.g. in the functional and/or phenotypical analysis of complex physiological processes such as analysis of gene expression profiles and/or proteomes. For example, one may prepare the knock-out phenotypes of human genes in cultured cells which are assumed to be regulators of alternative splicing processes. Among these genes are particularly the members of the SR splicing factor family, e.g. ASF/SF2, SC35, SRp20, SRp40 or SRp55. Further, the effect of SR proteins on the mRNA profiles of predetermined alternatively spliced genes such as CD44 may be analysed. Preferably the analysis is carried out by high-throughput methods using oligonucleotide based chips.

Using RNAi based knockout technologies, the expression of an endogenous target gene may be inhibited in a target cell or a target organism. The endogenous gene may be complemented by an exogenous target nucleic acid coding for the target protein or a variant or mutated form of the target protein, e.g. a gene or a cDNA, which may optionally be fused to a further nucleic acid sequence encoding a detectable peptide or poly-peptide, e.g. an affinity tag, particularly a multiple affinity tag. Variants or mutated forms of the target gene differ from the endogenous target gene in that they encode a gene product which differs from the endogenous gene product on the amino acid level by substitutions, insertions and/or deletions of single or multiple amino acids. The variants or mutated forms may have the same biological activity as the endogenous target gene. On the other hand, the variant or mutated target gene may also have a biological activity, which differs from the biological activity of the endogenous target gene, e.g. a partially deleted activity, a completely deleted activity, an enhanced activity etc.

The complementation may be accomplished by co-expressing the polypeptide encoded by the exogenous nucleic acid, e.g. a fusion protein comprising the target protein and the affinity tag and the double stranded RNA molecule for knocking out the endogenous gene in the target cell. This coexpression may be accomplished by using a suitable expression vector expressing both the polypeptide encoded by the exogenous nucleic acid, e.g. the tag-modified target protein and the single-stranded RNA molecule or alternatively by using a combination of expression vectors. Proteins and protein complexes which are synthesized de novo in the target cell will contain the exogenous gene product, e.g. the modified fusion protein. In order to avoid suppression of the exogenous gene product expression by the RNAi molecule, the nucleotide sequence encoding the exogenous nucleic acid may be altered on the DNA level (with or without causing mutations on the amino acid level) in the part of the sequence which is homologous to the single-stranded RNA molecule. Alternatively, the endogenous target gene may be complemented by corresponding nucleotide sequences from other species, e.g. from mouse.

Preferred applications for the cell or organism of the invention is the analysis of gene expression profiles and/or proteomes. In an especially preferred embodiment an analysis of a variant or mutant form of one or several target proteins is carried out, wherein said variant or mutant forms are reintroduced into the cell or organism by an exogenous target nucleic acid as described above. The combination of knockout of an endogenous gene and rescue by using mutated, e.g. partially deleted exogenous target has advantages compared to the use of a knockout cell. Further, this method is particularly suitable for identifying functional

domains of the target protein. In a further preferred embodiment a comparison, e.g. of gene expression profiles and/or proteomes and/or phenotypic characteristics of at least two cells or organisms is carried out. These organisms are selected from:

(i) a control cell or control organism without target gene inhibition, (ii) a cell or organism with target gene inhibition and (iii) a cell or organism with target gene inhibition plus target gene complementation by an exogenous target nucleic acid.

The method and cell of the invention may also be used in a procedure for identifying and/or characterizing pharmacological agents, e.g. identifying new pharmacological agents from a collection of test substances and/or characterizing mechanisms of action and/or side effects of known pharmacological agents.

Thus, the present invention also relates to a system for identifying and/or characterizing pharmacological agents acting on at least one target protein comprising:

(a) a eukaryotic cell or a eukaryotic non-human organism capable of expressing at least one endogenous target gene coding for said target protein,

(b) at least one single-stranded RNA molecule capable of inhibiting the expression of said at least one endogenous target gene by RNAi and

(c) a test substance or a collection of test substances wherein pharmacological properties of said test substance or said collection are to be identified and/or characterized.

Further, the system as described above preferably comprises:

(d) at least one exogenous target nucleic acid coding for the target protein or a variant or mutated form of the target protein wherein said exogenous target nucleic acid differs from the endogenous target gene on the nucleic acid level such that the expression of the exogenous target nucleic acid is substantially less inhibited by the single-stranded RNA molecule than the expression of the endogenous target gene.

Furthermore, the RNA knockout complementation method may be used for preparative purposes, e.g. for the affinity purification of proteins or protein complexes from eukaryotic cells, particularly mammalian cells and more particularly human cells. In this embodiment of the invention, the exogenous target nucleic acid preferably codes for a target protein which is fused to an affinity tag.

The preparative method may be employed for the purification of high molecular weight protein complexes which preferably have a mass of >150 kD and more preferably of >500 kD and which optionally may contain nucleic acids such as RNA. Specific examples are the heterotrimeric protein complex consisting of the 20 kD, 60 kD and 90 kD proteins of the U4/U6 snRNP particle, the splicing factor SF3b from the 17S U2 snRNP consisting of 5 proteins having molecular weights of 14, 49, 120, 145 and 155 kD and the 25S U4/U6/U5 tri-snRNP particle containing the U4, U5 and U6 snRNA molecules and about 30 proteins, which has a molecular weight of about 1.7 MD.

This method is suitable for functional proteome analysis in mammalian cells, particularly human cells.

Finally, the invention relates to a purified and isolated mammalian, particularly human RNA-induced silencing complex (RISC) having an apparent molecular weight of less than about 150-160 kDa, e.g. about 120 to 150-160 kDa. The RISC comprises polypeptide and optionally nucleic acid components, particularly single-stranded RNA molecules as described above. The RISC may be used as a target for diagnosis and/or therapy, as a diagnostic and/or therapeutic

agent itself, as a molecular-biological reagent or as component in a screening procedure for the identification and/or characterization of pharmaceutical agents.

Polypeptide components of RISC preferably comprise members of the Argonaute family of proteins, and contain eIF2C1 and/or eIF2C2, and possibly at least one other expressed eIF2C family member, particularly selected from eIF2C3, eIF2C4, HILI and HIWI.

Expression or overexpression of one or several proteins present in RISC in suitable host cells, e.g. eukaryotic cells, particularly mammalian cells, is useful to assist an RNAi response. These proteins may also be expressed or overexpressed in transgenic animals, e.g. vertebrates, particularly mammals, to produce animals particularly sensitive to injected single-stranded or double-stranded siRNAs. Further, the genes encoding the proteins may be administered for therapeutic purposes, e.g. by viral or non-viral gene delivery vectors.

It is also conceivable to administer a siRNA/eIF2C1 or 2 complex directly by the assistance of protein transfection reagents (e.g. Amphoteric Protein Transfection Reagents, ProVectin protein (Imgenex), or similar products) rather than RNA/DNA transfection. This may have technical advantages over siRNA transfection that are limited to nucleic acid transfection.

Alternatively to the application of siRNAs as synthetic double-stranded or single-stranded siRNAs, it is conceivable to also administer an antisense siRNA precursor molecule in the form of a hairpin stem-loop structure comprising 19 to 29 base pairs in the stem with or without 5' or 3' overhanging ends on one side of the duplex and a nucleotide or non-nucleotide loop on the other end. Preferably, the hairpin structure has a 3' overhang of from 1-5 nucleotides. Further, the precursor may contain modified nucleotides as described above, particularly in the loop and/or in the 3' portion, particularly in the overhang. The siRNA or precursors of siRNAs may also be introduced by viral vectors or RNA expression systems into a RISC compound, e.g. eIF2C1 and/or 2 overexpressing organism or cell line. The siRNA precursors may also be generated by direct expression within an organism or cell line. This may be achieved by transformation with a suitable expression vector carrying a nucleic acid template operatively linked to an expression control sequence to express the siRNA precursor.

Further, the present invention is explained in more detail in the following figures and examples.

BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1A and FIG. 1B. HeLa cytoplasmic S100 extracts show siRNA-dependent target RNA cleavage.

FIG. 1(A) Representation of the 177-nt ³²P-cap-labeled target RNA with the targeting siRNA duplex. Target RNA cleavage site and the length of the expected cleavage products is also shown. The fat black line positioned under the antisense siRNA is used in the following figures as symbol to indicate the region of the target RNA, which is complementary to the antisense siRNA sequence. FIG. 1(B) Comparison of the siRNA mediated target RNA cleavage using the previously established *D. melanogaster* embryo in vitro system and HeLa cell S100 cytoplasmic extract. 10 nM cap-labeled target RNA was incubated with 100 nM siRNA as described in materials. Reaction products were resolved on a 6% sequencing gel. Position markers were generated by partial RNase T1 digestion (T1) and partial alkaline hydro-

lysis (OH) of the cap-labeled target RNA. The arrow indicates the 5' cleavage product, the 3' fragment is unlabeled and therefore invisible.

FIG. 2A and FIG. 2B. Chemical modification of the 5' end of the antisense but not the sense siRNAs prevents sense target RNA cleavage in HeLa S100 extracts. FIG. 2(A) Illustration of the possible 5' and 3' aminolinker modifications of the sense and antisense strands of a siRNA duplex. L5 represents a 6-carbon chain aminolinker connected via a 5'-phosphodiester linkage, L3 represents a 7-carbon aminolinker connected via a phosphodiester bond to the terminal 3' phosphate, s, sense; as, antisense. FIG. 2(B) Target RNA cleavage testing various combinations of 5' and 3' aminolinker-modified siRNA duplexes. NC (negative control) shows an incubation reaction of the target RNA in the absence of siRNA duplex. T1, RNase T1 ladder; OH, partial alkaline hydrolysis ladder.

FIG. 3A and FIG. 3B. siRNA containing 3'-terminal phosphates are subjected to ligation as well as dephosphorylation reactions.

FIG. 3(A) Sequence of the radiolabeled siRNA duplex. The labeled nucleotide was joined to synthetic 20-nt antisense siRNA by T4 RNA ligation of ³²pCp. The various combinations of 5' and 3' hydroxyl/phosphate were prepared as described in materials. X and Y indicate 5' and 3' modifications of the antisense siRNA. FIG. 3(B) Fate of the antisense siRNA during incubation of the modified siRNA duplexes in HeLa S100 extract in the presence of non-radiolabeled target RNA. The different phosphorylated forms of the antisense siRNA were distinguished based on their gel mobility. Identical results were obtained when using 5' phosphorylated sense siRNA or when leaving out the target RNA during incubation. Ligation products are only observed when 3' phosphates were present on the labeled antisense siRNA.

FIG. 4. RISC is a stable complex that does not rapidly exchange bound siRNA. Increasing concentrations of non-specific siRNA compete with target-specific RISC formation when added simultaneously to HeLa S100 extracts (lanes 4 to 7). However, when the unspecific siRNA duplex is added 15 min after pre-incubation with the specific siRNA duplex, no more competition was observed (3 lanes to the right). T1, RNase T1 ladder.

FIG. 5A, FIG. B, FIG. C. Partial purification of human RISC.

FIG. 5(A) Graphical representation of the structure of the biotinylated siRNA duplex used for affinity purification of siRNA-associated factors. L3 indicates a C7-aminolinker that was conjugated to a photo-cleavable biotin N-hydroxysuccinimidyl ester; UV indicates photocleavage of the UV-sensitive linkage to release affinity selected complexes under native conditions. FIG. 5 (B) Superdex-200 gel filtration analysis of siRNA-protein complexes (siRNPs) recovered by UV treatment/elution (UV elu) from the streptavidin affinity column. Fractions were assayed for their ability to sequence-specifically cleave the cap-labeled target RNA. The number of the collected fractions and the relative positions of the aldolase (158 kDa) and BSA (66 kDa) size markers are indicated. FIG. 5 (C) Glycerol gradient (5%-20%) sedimentation of siRNPs recovered by UV treatment/elution from the streptavidin affinity column. For legend, see (B). When monitoring the precise size of target RNA cleavage fragments using internally ³²P-UTP-labeled, capped mRNA, the sum is equal to the full-length transcript, thus indicating that target RNA is indeed only cleaved once in the middle of the region spanned by the siRNA.

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FIG. 6. RISC contains a single-stranded siRNA.

siRNPs were subjected to affinity selection after incubation using siRNA duplexes with one or both strands biotinylated. The eluate recovered after UV treatment or the unbound fraction after streptavidin affinity selection (flow-through) was assayed for target RNA degradation. If the antisense strand was biotinylated, all sense target RNA-cleaving RISC was bound to the streptavidin beads, while sense siRNA biotinylation resulted in RISC activity of the flow-through. The cleavage reaction in the flow-through fraction was less efficient than in the UV eluate, because affinity-selected RISC was more concentrated.

FIG. 7A and FIG. 7B. Single-stranded antisense siRNAs reconstitute RISC in HeLa S100 extracts.

Analysis of RISC reconstitution using single-stranded or duplex siRNAs comparing HeLa S100 extracts FIG. 7(A) and the previously described *D. melanogaster* embryo lysate FIG. 7(B). Different concentrations of single-stranded siRNAs (s, sense; as, antisense) and duplex siRNA (ds) were tested for specific targeting of cap-labeled substrate RNA. 100 nM concentrations of the antisense siRNA reconstituted RISC in HeLa S100 extract, although at reduced levels in comparison to the duplex siRNA. Reconstitution with single-stranded siRNAs was almost undetectable in *D. melanogaster* lysate, presumably because of the higher nuclease activity in this lysate causing rapid degradation of uncapped single-stranded RNAs.

FIG. 8A and FIG. 8B. Single-stranded antisense siRNAs mediate gene silencing in HeLa cells.

FIG. 8(A) Silencing of nuclear envelope protein lamin A/C. Fluorescence staining of cells transfected with lam in A/C-specific siRNAs and GL2 luciferase (control) siRNAs. Top row, staining with lamin A/C specific antibody; middle row, Hoechst staining of nuclear chromatin; bottom row, phase contrast images of fixed cells. FIG. 8(B) Quantification of lamin A/C knockdown after Western blot analysis. The blot was stripped after lamin A/C probing and reprobed with vimentin antibody. Quantification was performed using a Lumi-Imager (Roche) and LumiAnalyst software to quantify the ECL signals (Amersham Biosciences), differences in gel loading were corrected relative to non-targeted vimentin protein levels. The levels of lamin A/C protein were normalized to the non-specific GL2 siRNA duplex.

FIG. 9A and FIG. 9B. Antisense siRNAs of different length direct target RNA cleavage in HeLa S100 extracts.

FIG. 9(A) Graphical representation of the experiment. Antisense siRNAs were extended towards the 5' side (series 1, 20 to 25-nt) or the 3' side (series 2, 20 to 23-nt).

FIG. 9(B) Target RNA cleavage using the antisense siRNAs described in FIG. 9(A). HeLa S100 extract was incubated with 10 nM cap-labeled target RNA and 100 nM antisense siRNAs at 30° C. for 2.5 h. Reaction products were resolved on a 6% sequencing gel. Position markers were generated by partial RNase T1 digestion (T1) and partial alkaline hydrolysis (OH) of the cap-labeled target RNA. Arrows indicate the position of the 5' cleavage products generated by the different antisense siRNAs. The fat black lines on the left (series 1) and the right (series 2) indicate the region of the target RNA, which is complementary to the antisense siRNA sequences.

FIG. 10. Length dependence of antisense siRNAs and effect of terminal modifications for targeting RNA cleavage in HeLa S100 extracts.

HeLa S100 extract was incubated with 10 nM cap-labeled target RNA and 100 nM antisense siRNAs at 30° C. for 2.5 h. Reaction products were resolved on a 6% sequencing gel. Position markers were generated by partial RNase T1 diges-

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tion (T1) of the cap-labeled target RNA. The fat black line on the left indicates the region of the target RNA, which is complementary to the 21-nt antisense siRNA sequence. The siRNA sequences used in each experiment are listed below (sense and antisense siRNAs are listed together, they were pre-annealed to form duplex siRNAs). p, phosphate; t, 2'-deoxythymidine, c, 2'-deoxycytidine, g, 2'-deoxycytidine, g, 2'-deoxyguanosine; L, aminolinker, B, photocleavable biotin; A,C,G,U, ribonucleotides.

Lane	Sense siRNA (5'-3')	Antisense siRNA (5'-3')
1		pUCGAAGUAUCCG CG
2		pUCGAAGUAUCCG CGUACGUG
3		pUCGAAGUAUCCG CGUACGUGAUGU
4		pUCGAAGUAUCCG CGUACGUGAUGUUC
5		pUCGAAGUAUCCG CGUACGUGAUGUUC AC
6		pUCGAAGUAUCCG CG
7		pUCGAAGUAUCCG CGUACGUG
8		pUCGAAGUAUCCG CGUACGUGAUGU
9		pUCGAAGUAUCCG CGUACGUGAUGUUC
10		pUCGAAGUAUCCG CGUACGUGAUGUUC AC
11		pUCGAAGUAUCCG CGUACGUG
12		pUCGAAGUAUCCG CGUACGtg
13		pUCGAAGUAUCCG CGUACGUU
14		pUCGAAGUAUCCG CGUACGtt
15		pUCGAAGUAUCCG CGUACGUG
16		pUCGAAGUAUCCG CGUACGtg
17		pUCGAAGUAUCCG CGUACGUU
18		pUCGAAGUAUCCG CGUACGtt
19	CGUACGCGGAAUACUUCG AAA	pUCGAAGUAUCCG CGUACGUG
20	CGUACGCGGAAUACUUCG AAA	pUCGAAGUAUCCG CGUACGtg
21	CGUACGCGGAAUACUUCG AAA	pUCGAAGUAUCCG CGUACGUU

-continued

Lane	Sense siRNA (5'-3')	Antisense siRNA (5'-3')
22	CGUACGCGGAAUACUUCG AAA	pUCGAAGUAUUCG CGUACGtt
23		tCGAAGUAUUCGCG GUACGUULB
24	cGUACGCGGAAUACUUCG AUULB	tCGAAGUAUUCGCG GUACGUULB
25		ptCGAAGUAUUCGCG GUACGttLB
26	cGUACGCGGAAUACUUCG AttLB	ptCGAAGUAUUCGCG GUACGttLB
27		ptCGAAGUAUUCGCG GUACGttL

FIG. 11: Single-stranded antisense siRNAs mediate gene silencing in HeLa cells. Quantification of lamin A/C knock-down after Western blot analysis. The blot was stripped after lamin A/C probing and re-probed with vimentin antibody. Quantification was performed using a Lumi-Imager (Roche) and LumiAnalyst software to quantitate the ECL signals (Amersham Biosciences), differences in gel loading were corrected relative to non-targeted vimentin protein levels. The levels of lamin A/C protein were normalized to the non-specific GL2 siRNA duplex.

FIG. 12 A and FIG. B. Protein composition of affinity purified RISC.

FIG. 12(A) Silver-stained SDS-PAGE gel of affinity-selected ribonucleoprotein complexes after glycerol gradient (5%-20%) sedimentation. The arrow indicates the band containing eIF2C1 and eIF2C2. Molecular size markers are indicated on the left. The asterisk indicates a fraction for which the protein pellet was lost after precipitation. FIG. 12(B) Target RNA cleavage assay of the collected fractions. RISC activity peaked in fraction 7 and 8; bu, buffer.

FIG. 13A, FIG. 13B and FIG. 13C. Mass spectrometric characterization of eIF2C1 and eIF2C2. The 100 kDa band was analysed by mass spectrometry. Mass spectrum indicating the peptide peaks corresponding to eIF2C2 FIG. 13(A) and eIF2C1 FIG. 13(B). FIG. 13(C) Alignment of eIF2C2 (SEQ ID NO:69) and eIF2C1 (SEQ ID NO:68) amino-acid sequences indicating the position of the identified peptides. Sequence differences are indicated by yellow boxes.

FIG. 14. Predicted amino-acid sequences of the six human Argonaute protein family members, eIF2C1 (SEQ ID NO:68), eIF2C2 (SEQ ID NO:69), eIF2C3 (SEQ ID NO:70), eIF2C4 (SEQ ID NO:71), HILI (SEQ ID NO:72), and HIWI (SEQ ID NO:73).

FIG. 15. Alignment of the sequences of the six human Argonaute protein family members. [eIF2C1; SEQ ID NO: 68; eIF2C2; SEQ ID NO: 69; eIF2C3; SEQ ID NO: 70; eIF2C4; SEQ ID NO: 71; HILI; SEQ ID NO: 72; HIWI; SEQ ID NO: 73].

FIG. 16. Predicted cDNA sequences of the six human Argonaute protein family members [eIF2C1; SEQ ID NO: 74; eIF2C2; SEQ ID NO: 75; eIF2C3; SEQ ID NO: 76; eIF2C4; SEQ ID NO: 77; HILI; SEQ ID NO: 78; HIWI; SEQ ID NO: 79].

FIG. 17A and FIG. 17B. All members of the Argonaute family but HIWI are expressed in HeLa cells.

RT-PCR analysis on polyA RNA from HeLa cells. FIG. 17(A) Primers (forward and reverse) used for nested and semi-nested PCR amplification of the different Argonautes and expected length of the FOR products. FIG. 17(B) Agarose gel electrophoresis of the obtained PCR products, confirming the expected 5 length. Left lanes, 100 bp DNA ladder.

EXAMPLE

1. Material and Methods

5 1.1 siRNA Synthesis and Biotin Conjugation

siRNAs were chemically synthesized using RNA phosphoramidites (Proligo, Hamburg, Germany) and deprotected and gel-purified as described previously. 5' aminolinkers were introduced by coupling MMT-C6-aminolinker phosphoramidite (Proligo, Hamburg), 3' C7-aminolinkers were introduced by assembling the oligoribonucleotide chain on 3'-aminomodifier (TFA) C7 lcaa control pore glass support (Chemgenes, Mass., USA). The sequences for GL2 luciferase siRNAs were as described (Elbashir et al., 2001 a, supra). If 5'-phosphates were to be introduced, 50 to 100 nmoles of synthetic siRNAs were treated with T4 polynucleotide kinase (300 µl reaction, 2.5 mM ATP, 70 mM Tris-HCl, pH 7.6, 10 mM MgCl₂, 5 mM DTT, 30 U T4 PNK, New England Biolabs, 45 min, 37° C.) followed by ethanol precipitation.

3' Terminal ³²pCp labeling (FIG. 3) was performed in a 30 µl reaction (17 µM siRNA, 0.5 µM ³²pCp (110 TBq/mmol), 15% DMSO, 20 U T4 RNA ligase, NEB, and 1xNEB-supplied reaction buffer) for 1.5 h at 37° C., and gel-purified. One half of the pCp-labeled RNA was dephosphorylated (25 µl reaction, 500 U alkaline phosphatase, Roche, and Roche-supplied buffer, 30 min, 50° C.), followed by phenol/chloroform extraction and ethanol precipitation. Half of this reaction was 5' phosphorylated (20 µl reaction, 2 units T4 polynucleotide kinase, NEB, 10 mM ATP, NEB-supplied buffer, 60 min, 37° C.). A quarter of the initial pCp-labeled siRNA was also 5' phosphorylated (10 µl reaction, 10 units 3' phosphatase-free T4 polynucleotide kinase, Roche, 10 mM ATP, Roche-supplied buffer, 3 min, 37° C.).

For conjugation to biotin, 20 to 65 nmoles of fully deprotected aminolinker-modified siRNA were dissolved in 100 µl of 100 mM sodium borate buffer (pH 8.5) and mixed with a solution of 1 mg of EZ-Link NHS-PC-LC-Biotin (Pierce, Ill., USA) in 100 µl of anhydrous dimethylformamide. The solution was incubated for 17 h at 25° C. in the dark. Subsequently, siRNAs were precipitated by the addition of 60 µl 2 M sodium acetate (pH 6.0) and 1 ml ethanol. The RNA pellet was collected by centrifugation and biotin-conjugated siRNA was separated from non-reacted siRNA on a preparative denaturing 18% acrylamide gel (40 cm length) in the dark. The RNA bands were visualized by 254 nm UV shadowing and minimized exposure time. The bands were excised, and the RNA was eluted overnight in 0.3 M NaCl at 4° C. and recovered by ethanol precipitation. siRNA duplexes were formed as previously described (Elbashir et al., Methods 26 (2002), 199-213).

1.2 Preparation of S100 Extracts from HeLa Cells

5 Cytoplasm from HeLa cells adapted to grow at high density was prepared following the Dignam protocol for isolation of HeLa cell nuclei (Dignam et al., Nucleic Acids Res. 11 (1983), 1475-1489). The cytoplasmic fraction was supplemented with KCl, MgCl₂ and glycerol to final concentrations of 100 mM, 2 mM and 10%, respectively. At this stage, the extracts can be stored frozen at -70° C. after quick-freezing in liquid nitrogen without loss of activity.

S100 extracts were prepared by ultracentrifugation at 31 0.500 rpm for 60 minutes at 4° C. using a Sorvall T-865 rotor. The protein concentration of HeLa S100 extract varied between 4 to 5 mg/ml as determined by Bradford assay.

1.3 Affinity Purification of RISC with 3' Biotinylated siRNA Duplexes

For affinity purification of siRNA-associated protein complexes from HeLa S100 extracts, 10 nM of a 3' double-biotinylated siRNA duplex were incubated in 0.2 mM ATP, 0.04 mM GTP, 10 U/ml RNasin, 6 µg/ml creatine kinase, and 5 mM creatine phosphate in 60% S100 extract at 30° C. for 30 to 60 min and gentle rotation. Thereafter, 1 ml slurry of Immobilized Neutravidin Biotin Binding Protein (Pierce, Ill., USA) was added per 50 ml of reaction solution and the incubation was continued for another 60 to 120 min at 30° C. with gentle rotation. The Neutravidin beads were then collected at 2000 rpm for 2 minutes at 4° C. in a Heraeus Megafuge 1.0 R centrifuge using a swinging bucket rotor type 2704. Effective capturing of RISC components after affinity selection was confirmed by assaying the supernatant for residual RISC activity with and without supplementing fresh siRNA duplexes. The collected Neutravidin beads were washed with 10 volumes of buffer A relative to the bead volume (30 mM HEPES, pH 7.4, 100 mM KCl, 2 mM MgCl₂, 0.5 mM DTT, 10% glycerol) followed by washing with 5 volumes of buffer B (same as buffer A with only 3% glycerol content). The beads were transferred to a 0.8x4 cm Poly-Prep chromatography column (BioRad; CA, USA) by resuspending in 3 volumes of buffer B at 4° C., followed by 10 volumes of washing with buffer B. Washing of the beads was continued by 10 volumes of buffer B increased to 300 mM KCl. The column was then reequilibrated with regular buffer B. To recover native siRNA-associated complexes, the column was irradiated in the cold room by placing it at a 2 cm distance surrounded by four 312 nm UV lamps (UV-B tube, 8 W, Herolab, Germany) for 30 minutes. To recover the photocleaved siRNP solution, the column was placed into a 50 ml Falcon tube and centrifuged at 2000 rpm for 1 minute at 4° C. using again the 2704 rotor. For full recovery of siRNPs, the beads were once again resuspended in buffer B followed by a second round of UV treatment for 15 minutes. Both eluates were pooled and assayed for target RNA degradation.

1.4 Target RNA Cleavage Assays

Cap-labeled target RNA of 177 nt was generated as described (Elbashir et al., EMBO J. 20 (2001 c), 6877-6888) except that his-tagged guanylyl transferase was expressed in *E. coli* from a plasmid generously provided by J. Wilusz and purified to homogeneity. If not otherwise indicated, 5' phosphorylated siRNA or siRNA duplex was pre-incubated in supplemented HeLa S100 extract at 30° C. for 15 min prior to addition of cap-labeled target RNA. After addition of all components, final concentrations were 100 nM siRNA, 10 nM target RNA, 1 mM ATP, 0.2 mM GTP, 10 U/ml RNasin, 30 µg/ml creatine kinase, 25 mM creatine phosphate, 50% S100 extract. Incubation was continued for 2.5 h. siRNA-mediated target RNA cleavage in *D. melanogaster* embryo lysate was performed as described (Zamore et al., Cell 101 (2000), 25-33). Affinity-purified RISC in buffer B was assayed for target RNA cleavage without preincubation nor addition of extra siRNA (10 nM target RNA, 1 mM ATP, 0.2 mM GTP, 10 U/ml RNasin, 30 µg/ml creatine kinase, 25 mM creatine phosphate, 50% RISC in buffer B). Cleavage reactions were stopped by the addition of 8 vols of proteinase K buffer (200 mM Tris-HCl pH 7.5, 25 mM EDTA, 300 mM NaCl, 2% w/v SDS). Proteinase K, dissolved in 50 mM Tris-HCl pH 8.0, 5 mM CaCl₂, 50% glycerol, was added to

a final concentration of 0.6 mg/ml and processed as described (Zamore et al. (2000), supra). Samples were separated on 6% sequencing gels.

1.5 Analytical Gel Filtration

UV-eluates in buffer B were fractionated by gel filtration using a Superdex 200 PC 3.2/30 column (Amersham Biosciences) equilibrated with buffer A on a SMART system (Amersham Biosciences). Fractionation was performed by using a flow rate of 40 µl/minute and collecting 100 µl fractions. Fractions were assayed for specific target RNA cleavage. Size calibration was performed using molecular size markers thyroglobulin (669 kDa), ferritin (440 kDa), catalase (232 kDa), aldolase (158 kDa) and BSA (66 kDa) (Amersham Biosciences).

1.6 Glycerol Gradient Sedimentation

UV-eluates were layered on top of 4 ml linear 5% to 20% (w/w) glycerol gradient adjusted to 30 mM HEPES, pH 7.4, 100 mM KCl, 2 mM MgCl₂, 0.5 mM DTT. Centrifugation was performed at 35000 rpm for 14.5 h at 4° C. using a Sorvall SW 60 rotor. Twenty fractions of 0.2 ml volume were removed sequentially from the top and 15 µl aliquots were used to assay for target RNA cleavage.

Results

2.1 A Human Biochemical System for siRNA Functional Analysis

We were interested in assaying siRNA-mediated target RNA degradation in human cell extracts, because siRNAs are powerful reagents to knockdown gene expression in human cells but the action of siRNAs in human cells was uncertain. To investigate whether siRNAs guide target RNA degradation in human cells with a similar mechanism to the one observed in *D. melanogaster* (e.g. Elbashir et al. (2001 b), supra), we prepared substrates for targeted mRNA degradation as described previously (Elbashir et al. (2001 c), supra). A 5'-³²P-cap-labeled, 177-nt RNA transcript, derived from a segment of the firefly luciferase gene, was incubated in HeLa cell S100 or *D. melanogaster* embryo extracts with a 21-nt siRNA duplex in the presence of an ATP regeneration system (FIG. 1 A, B). siRNA cleavage assays were performed at 25° C. in *D. melanogaster* lysate and at 30° C. in HeLa S100 extracts for 2.5 h. After deproteinization using proteinase K, the reaction products were separated on a 6% sequencing gel.

Similar to the previous observation in *D. melanogaster* lysate, we observed the appearance of a cleavage product in HeLa S100 extract at exactly the same position, thus indicating that the siRNA duplex guides target RNA cleavage in the human system with the same specificity and mechanism. The cleavage reaction appeared less efficient when compared to the *D. melanogaster* system, but this could be explained by the 5-fold lower total protein concentration of HeLa S100 extracts (25 mg/ml vs. 5 mg/ml). Similar to *D. melanogaster* lysates, siRNA duplexes without 5' phosphate were rapidly 5' phosphorylated in HeLa S100 extracts (see below) and the ability to cleave the target RNA was independent of the presence of a 5' phosphate on the synthetic siRNA duplexes.

Comparative analysis of the efficiency of siRNA duplexes of different length in *D. melanogaster* lysate and in transfected mammalian cells indicated that the differences in silencing efficiencies between 20- to 25-nt siRNA duplexes were less pronounced in mammalian cells than in *D. melanogaster* (Elbashir et al. (2002), supra). Duplexes of 24- and 25-nt siRNAs were inactive in *D. melanogaster* lysate, whereas the same 30 duplexes were quite effective for silencing when introduced by transfection into HeLa cells. We therefore asked whether siRNA duplexes of 20- to 25-nt

are able to reconstitute RISC also with approximately equal efficiency. Indeed, we observed no large differences in our biochemical assay, and the position of target RNA cleavage was as predicted according to the cleavage guiding rules established in *D. melanogaster* lysate (data not shown). Our biochemical results therefore support the in vivo observations.

2.2 5' Modification of the Guide siRNA Inhibits RISC Activity

Modification of siRNAs at their termini is important for developing siRNA-based affinity purification schemes or for conjugating reporter tags or for biophysical measurements. The most common method for introducing reactive side chains into nucleic acids is by chemical synthesis using aminolinker derivatives (Eckstein (1991), Oligonucleotides and analogues, 2nd Ed., Oxford UK, Oxford University Press). After complete deprotection of the oligonucleotide, the primary amine is typically reacted with the 5 N-hydroxy-succinimidyl ester of the desired compound. We have introduced 5' and 3' aminolinkers with six and seven methylene groups as spacers, respectively. The linker-modified siRNA duplexes were tested for mediating target RNA degradation in HeLa S100 extract (FIG. 2A, B). Modification of the 5'-end of the antisense guide siRNA abolished target RNA cleavage, while modification of neither the sense 5'-end nor of both 3'-ends showed any inhibitory effect. In an identical experiment using *D. melanogaster* embryo lysate, we observed a similar pattern of RISC activity although the duplex carrying the 5' aminolinker-modified antisense siRNA showed some residual activity (data not shown). Presumably, introduction of additional 5 atoms or the change in terminal phosphate electric charge at the 5'-end of the antisense siRNA interfered with its ability to function as guide RNA. The critical function of the guide siRNAs 5' end was previously documented (Elbashir et al. (2001 c), supra).

The ability to modify siRNAs at their 3'-end suggests that siRNAs do not play a major role for priming dsRNA synthesis and do not act as primers for degenerative PCR. The fate of a siRNA in HeLa S100 extracts was followed directly by incubation of an internally ³²P-labeled siRNA duplexes. The radiolabeled antisense siRNA strand was also prepared with different 5' and 3' phosphate modifications (FIG. 3A). All described combinations of siRNA duplexes were fully competent for RISC-dependent target RNA degradation (data not shown). As previously observed for *D. melanogaster* lysates (Nykanen et al. (2001), supra), rapid 5' phosphorylation of siRNA duplexes with free 5' hydroxyl termini was apparent. To our surprise, we noted that a small fraction of the 3' phosphorylated antisense siRNA could be ligated to the opposing 5' hydroxyl of the sense siRNA producing a lower mobility band. The inter-strand ligation was confirmed by changing the length of the unlabeled sense siRNA, which resulted in the expected mobility changes of the ligation product (data not shown). RNA ligase activity was previously observed in HeLa S100 extracts and it is mediated by two enzymatic activities (e.g. Vicente and Filipowicz, Eur. J. Biochem., 176 (1988), 431-439). The 3' terminal phosphate is first converted to a 2',3'-cyclic phosphate requiring ATP and 3' terminal phosphate cyclase. Thereafter, the opposing 5' hydroxyl is ligated to the cyclic phosphate end by an as yet uncharacterized RNA ligase. We chemically synthesized the predicted 5' phosphorylated, 42-nt ligation product and found that it is unable to mediate target RNA cleavage, presumably because it can not form activated RISC. The majority of the 3' phosphorylated duplexes siRNA was gradually dephosphorylated at its 3' end and emerged chemically similar to

naturally generated siRNA. Together, these observations indicate that the cell has a mechanism to preserve the integrity of siRNAs. We were unable to detect a proposed siRNA-primed polymerization product (FIG. 3B), suggesting that siRNAs do not function as primers for template-dependent dsRNA synthesis in our system. However, we acknowledge that a proposed RNA-dependent polymerase activity may have been inactivated during preparation of our extracts.

2.3 siRNAs Incorporated into RISC do not Compete with a Pool of Free siRNAs

In order to analyze RISC assembly and stability, we tested whether target-unspecific siRNA duplexes were able to compete with target-specific siRNA duplexes. When specific and non-specific siRNA duplexes were co-incubated in HeLa S100 extracts, increasing concentrations of unspecific siRNA duplex competed with the formation of target-specific

RISC (FIG. 4, left lanes). However, when target-specific siRNAs were pre-incubated in HeLa S100 extract for 15 min in the absence of competitor siRNA duplex, the assembled siRNA in the target-specific RISC could no longer be competed with the target-unspecific siRNA duplex

(FIG. 4, right lanes). This result suggests that RISC is formed during the first 15 minutes of incubation and that siRNAs were irreversibly associated with the protein components of RISC during the 2.5 h time window of the experiment.

2.4 Purification of Human RISC

After having the 3' termini of siRNAs defined as the most suitable position for chemical modification, a photo-cleavable biotin derivative was conjugated to the 3' aminolinker-modified siRNAs. A photo-cleavable biotin derivative was selected because of the advantage of recovering RISC under non-denaturing conditions after capturing complexes on streptavidin-coated affinity supports. 3' Conjugation of biotin to the sense, antisense or to both of the strands did not affect target RNA cleavage when compared to non-biotinylated siRNAs (data not shown). siRNA duplexes with biotin residues on both 3' ends were therefore used for affinity purification (FIG. 5A). The double biotinylated siRNA duplex was incubated in HeLa S100 extracts in the presence of ATP, GTP, creatine phosphate, and creatine kinase for ATP regeneration. Thereafter, streptavidin-conjugated agarose beads were added to capture the biotinylated siRNA ribonucleoprotein complexes (siRNPs) including RISC. After extensive washing of the collected beads, the siRNPs were released by UV irradiation at 312 nm. The eluate cleaved target RNA sequence-specifically, thus indicating that RISC was recovered in its native state from the resin (FIG. 5B, C, lane UV elu). The flow-through from the affinity selection showed no detectable RISC activity indicating complete binding of RISC by the beads (FIG. 6). The affinity eluate was further analyzed by applying it onto a Superdex 200 gel filtration column (FIG. 5B) as well as a 5%-20% glycerol gradient ultra-centrifugation (FIG. 5C). Individual fractions were collected and assayed for target RNA cleavage without the addition of any further siRNA. RISC activity appeared between the molecular size markers aldolase (158 kDa) and BSA (66 kDa) after gel filtration or glycerol gradient centrifugation (FIG. 5B, C). The molecular size of human RISC is therefore estimated to be between 90 and 160 kDa, significantly smaller than the complex previously analyzed in *D. melanogaster* lysates (Hammond et al. (2000), supra; Nykanen et al. (2001), supra). The small size of RISC suggests that Dicer (210 kDa) is not contained in RISC and that the formation of RISC from synthetic siRNAs

may occur independently of Dicer. While these results do not rule out a role for Dicer during assembly of RISC, they emphasize the absence of Dicer in RISC.

2.5 RISC Contains a Single siRNA Strand and can be Reconstituted Using Single-Stranded siRNAs

Two models are currently discussed concerning the siRNA strand composition of RISC. The first model suggests that both strands of the initially added siRNA duplex are physically present in RISC, but in an unwound conformation. The second model proposes that RISC carries only a single siRNA strand, implying loss of one of the siRNA strands during assembly. The latter model has been favored based on the analogy to miRNA precursor processing, where only one 21-nt strand accumulated from a dsRNA hairpin precursor. The molecular basis for the asymmetry of the miRNA precursor processing reaction is not yet understood. Because siRNAs have symmetric 2-nt 3'-overhangs it is assumed that siRNA duplexes enter RISC with equal probability for both orientations, thus giving rise to distinct sense and antisense targeting RISCs.

To address the constitution of siRNAs in RISC, we affinity selected the assembled complexes with siRNA duplexes that were biotinylated at only one of the two constituting strands or both (FIG. 6). If both strands were present together in RISC, the cleavage activity should be affinity selected on Neutravidin independently of the position of the biotin residue. In contrast, we observed target RNA cleavage from UV eluates after streptavidin selection only for siRNA duplexes with biotin conjugated to the antisense strand, but not the sense strand (FIG. 6). RISC activity, assembled on siRNA duplexes with only the sense siRNA biotinylated, remained in the flow-through. These data suggest that RISC contains only a single-stranded RNA molecule.

To assess whether single-stranded siRNAs may be able to reconstitute RISC, single-stranded 5' phosphorylated siRNAs as well as the siRNA duplex were incubated at concentrations between 1 to 100 nM with cap-labeled target RNA in HeLa S100 extract (FIG. 7A). At 100 nM single-stranded antisense siRNA, we detected RISC-specific target RNA cleavage, thus confirming that single-stranded siRNAs are present in RISC. At lower concentrations of single-stranded siRNAs, RISC formation remained undetectable while duplex siRNAs were effectively forming RISC even at 1 nM concentration. Therefore, a specific pathway exists which converts double-stranded siRNA into single-stranded siRNA containing RISC. Using *D. melanogaster* embryo lysate, we were unable to detect RISC activity from antisense siRNA (FIG. 7B), presumably because of the high load of single-strand specific ribonucleases (Elbashir et al. (2001 b), supra). Furthermore, 5' phosphorylated 20- to 25-nt antisense siRNAs were able to mediate RISC-specific target RNA degradation in HeLa S100 extract producing the same target RNA cleavage sites as duplex siRNAs of this length (data not shown).

Finally, we tested single-stranded and duplex siRNAs for targeting of an endogenous gene in HeLa cells following our standard protocol previously established for silencing of lamin A/C. 200 nM concentrations of single-stranded siRNAs with and without 5' phosphate and 100 nM concentrations of duplex siRNAs were transfected into HeLa cells. Lamin A/C levels were monitored 48 h later using immunofluorescence (FIG. 8A) and quantitative luminescence-based Western blot analysis (FIG. 8B). Non-phosphorylated antisense siRNA caused a substantial knockdown of lamin A/C to about 25% of its normal level while 5' phosphorylated siRNAs reduced the lamin A/C content to less than 5%,

similar to the reduction observed with the lamin A/C 5' phosphorylated (data not shown) or 5 non-phosphorylated duplex siRNA (FIG. 8). Sense siRNA and GL2 unspecific siRNA did not affect lamin A/C levels. The levels of non-targeted vimentin protein were monitored and used for normalizing of the loading of the lanes of the lamin A/C Western blots.

Gene silencing was also observed with phosphorylated as well as non-phosphorylated antisense siRNAs ranging in size between 19 to 29 nt. The phosphorylated antisense siRNAs were consistently better performing than the non-phosphorylated antisense, and their silencing efficiencies were comparable to that of the conventional duplex siRNA (FIG. 11).

2.6 Protein Composition of RISC

In order to identify the protein components of the RNA-induced silencing complex (RISC) in HeLa S100 extract, the specific affinity selection previously outlined was used. UV eluates were fractionated on a 5-20% glycerol gradient, fractions were recovered from the gradient and analysed for protein composition and target RNA endonucleolytic activity.

Two proteins of approximately 100 kDa were identified by mass spectrometry in the peak fraction of the endonucleolytic activity (FIG. 12, fractions 7 and 8), corresponding to eIF2C1 and eIF2C2/GERp95 (FIGS. 13A and B). These proteins are 82% similar and are both members of the Argonaute family (FIG. 13C). The first evidence that Argonaute proteins are part of RISC was provided by classical biochemical fractionation studies using dsRNA-transfected *D. melanogaster* S2 cells (Hammond et al., 2001, supra). The closest relative to *D. melanogaster* Argonaute2, *D. melanogaster* Argonautel, was recently shown to be required for RNAi (Williams and Rubin, PNAS USA 99 (2002), 6889-6894).

Mass spectrometry analysis also revealed the presence of three peptides belonging exclusively to the HILI member of the Argonaute family of proteins. The sequences of those peptides are: NKQDFMDLSICTR, corresponding to positions 17-29 of the protein; TEYVAESFLNCLRR, corresponding to positions 436-449 of the protein, and; YNH-DLPARIIVYR, corresponding to positions 591-603 of the protein. This finding suggests that the protein HILI may also be part of RISC.

In human, the Argonaute family is composed of 6 members, eIF2C1, eIF2C2, eIF2C3, eIF2C4, HILI and HIWI (FIG. 14). The alignment of the six predicted amino-acid sequences show a high conservation, in particular between the eIF2C members, and HILI and HIWI (FIG. 15). Predicted cDNA sequences encoding the Argonaute proteins are also shown (FIG. 16).

The expression of the human Argonaute proteins was also investigated in HeLa cells by RT-PCR analysis using total and poly (A) selected RNA. All members of the family but HIWI were detected (FIG. 17).

3. Discussion

The development of a human biochemical system for analysis of the mechanism of RNAi is important given the recent success of siRNA duplexes for silencing genes expressed in human cultured cells and the potential for becoming a sequence-specific therapeutic agent. Biochemical systems are useful for defining the individual steps of the RNAi process and for evaluating the constitution and molecular requirements of the participating macromolecular complexes. For the analysis of RNAi, several systems were developed, with the *D. melanogaster* systems being the most comprehensive as they enable to reconstitute dsRNA pro-

cessing as well as the mRNA targeting. For mammalian systems, reconstitution of the mRNA targeting reaction has not yet been accomplished. Here, we describe the development and application of a biochemical system prepared from the cytoplasmic fraction of human HeLa cells, which is able to reconstitute the human mRNA-targeting RNA-induced silencing complex (RISC). Formation of RISC was accomplished using either 5' phosphorylated or non-phosphorylated siRNA duplexes; as well as single-stranded antisense siRNAs; non-phosphorylated siRNA duplexes and presumably also single-stranded antisense siRNAs are rapidly 5' phosphorylated in HeLa cell extracts (FIG. 3).

Biochemical Characterization of siRNA Function

Reconstitution of RISC activity was only observed using cytoplasmic HeLa extracts. HeLa nuclear extracts assayed under the same conditions did not support siRNA-specific target RNA cleavage, thus suggesting that RISC components are located predominantly in the cytoplasm (data not shown).

Modifications of the 5' and 3' termini of siRNAs were tested in order to assess the importance of the siRNA termini for the targeting step. It was found that the 5' end modification of the guide siRNA was more inhibitory for target RNA cleavage than 3' end modification. Introduction of the 3' biotin affinity tag into the target-complementary guide siRNA enabled us to affinity select sense-RNA-targeting RISC, whereas 3' biotinylation of the sense siRNA strand resulted in RISC activity in the flowthrough. Furthermore, the single RNA strand composition of RISC was confirmed by reconstituting the sequence-specific endonuclease complex using 5'-phosphorylated single-stranded guide siRNA. The reconstitution of RISC from single-stranded siRNA was however less effective and required 10- to 100-fold higher concentrations compared to duplex siRNA. Reconstitution of RISC from single-stranded siRNA was undetectable using *D. melanogaster* embryo lysate, which is most likely explained by the high content of 5' to 3' exonucleases in embryo lysate.

The size of RISC in HeLa lysate was determined by gel filtration as well as glycerol gradient ultracentrifugation after streptavidin affinity purification with 3' biotinylated siRNA duplexes. Sizes for RISC in *D. melanogaster* systems have been reported within a range of less than 230 to 500 kDa, however size determinations were conducted without having affinity purified RISC. Our affinity-purified RISC sediments in a narrow range between the size makers of 66 and 158 kDa. The differences to the reported sizes for RISC are not species-specific as we observed a similar size for RISC in *D. melanogaster* S2 cell cytoplasmic extracts after affinity purification (data not shown). It has previously been proposed that siRNAs act as primers for target RNA-templated dsRNA synthesis (Lipardi et al., Cell 107 (2001), 297-307) although homologs for such RNA-dependent RNA polymerases known to participate in gene silencing in other systems are not identified in *D. melanogaster* or mammalian genomes. Analysis of the fate of siRNA duplexes in the HeLa cell system did not provide evidence for such a siRNA-primed activity (FIG. 3), but indicates that the predominant pathway for siRNA-mediated gene silencing is sequence-specific endonucleolytic target RNA degradation.

Single-Stranded 5' Phosphorylated Antisense siRNAs as Triggers of Mammalian Gene Silencing

It was previously noted that introduction of sense and antisense RNAs of several hundred nucleotides in length into *C. elegans* was able to sequence-specifically silence homologous genes (Guo and Kemphues, Cell 81 (1995), 611-620). Later, it was suggested that the sense and anti-

sense RNA preparation were contaminated with a small amount of dsRNA, which was responsible for the silencing effect and is a much more potent inducer of gene silencing (Fire et al. (1998), supra). It is however conceivable that antisense RNA directly contributed to initiation of silencing. Indeed, most recently it was shown that antisense RNAs between 22 and 40 nt, but not sense RNAs were able to activate gene silencing in *C. elegans* (Tijsterman et al., Science 295 (2002), 694-697). The authors, however, favored the hypothesis of siRNA-primed dsRNA synthesis.

We have shown that modification of the 3' ends of antisense siRNA did not interfere with reconstitution of RISC in the human system. Together, these observations suggest that the driving forces for gene silencing in *C. elegans* may be predominantly dsRNA synthesis followed by Dicer cleavage, while in human and possibly also in *D. melanogaster* RISC-specific target mRNA degradation predominates.

Targeting of endogenously expressed lamin A/C by transfection of duplex siRNA into HeLa cells was the first reported example of siRNA-induced gene silencing. Lamin A/C protein was drastically reduced by a lamin A/C-specific siRNA duplex within two days post transfection, while unspecific siRNA duplexes showed no effect. At the time, transfection of non-phosphorylated sense or antisense siRNA did not reveal a substantial effect on lamin A/C levels, although more recently a minor reduction upon antisense siRNA transfection was noticed when similar concentrations of antisense siRNA were delivered as described in this study. However, the effect was not interpreted as RISC-specific effect. Assaying 5'-phosphorylated antisense siRNAs revealed a substantial increase in lamin A/C silencing. Probably, 5' phosphorylated siRNAs are more stable or enter RISC more rapidly. Alternatively, the 5' end of transfected single-stranded siRNA may be less rapidly phosphorylated in the cell in comparison to duplex siRNAs.

Finally, it should be noted that HeLa cells are generally poor in nucleases and represent one of the preferred mammalian systems for studying RNA-processing or transcription reactions in vivo and in vitro. However, it can be expected that 5' phosphorylated single-stranded antisense siRNAs are suitable to knockdown gene expression in other cell types or tissues with a different content of nucleases, since chemical strategies to improve nuclease resistance of single stranded RNA are available. The general silencing ability of various cell types may also depend on the relative levels of siRNA/miRNA-free eIF2C1 and eIF2C2 proteins capable of associating with exogenously delivered siRNAs.

In summary, single-stranded 5'-phosphorylated antisense siRNAs of 19- to 29-nt in size broaden the use of RNA molecules for gene silencing because they can enter the mammalian RNAi pathway in vitro as well as in vivo through reconstitution of RISC. Human eIF2C1 and/or eIF2C2 seem to play a critical role in this process. Considering the feasibility of modulating the stability and uptake properties of single-stranded RNAs, 5'-phosphorylated single-stranded antisense siRNAs may further expand the utility of RNAi-based gene silencing technology as tool for functional genomics as well as therapeutic applications.

Argonaute proteins are a distinct class of proteins, containing a PAZ and Piwi domain (Cerutti et al., 2000, supra) and have been implicated in many processes previously linked to post-transcriptional silencing, however only limited biochemical information is available.

Human eIF2C2 is the ortholog of rat GERp95, which was identified as a component of the Golgi complex or the

endoplasmic reticulum and copurified with intracellular membranes (Cikaluk et al., *Mol. Biol. Cell* 10 (1999), 3357-3722). More recently, HeLa cell eIF2C2 was shown to be associated with microRNAs and components of the SMN complex, a regulator of ribonucleoprotein assembly, suggesting that eIF2C2 plays a role in miRNA precursor processing or miRNA function (Mourelatos et al., *Genes & Dev.* 16 (2002), 720-728). A more provocative hypothesis is that miRNAs are also in a RISC-like complex, which could potentially mediate target RNA degradation, if only perfectly matched miRNA target mRNAs existed. Sequence analysis using cloned human and mouse, however, did not reveal the presence of such perfectly complementary sequences in the genomes (Lagos-Quintana et al., *Science* 294 (2001), 853-858). Therefore, miRNPs may only function as translational regulators of partially mismatched target mRNAs, probably by recruiting additional factors that prevent dissociation from mismatched target mRNAs.

Human eIF2C1 has not been linked to gene silencing previously, but it is more than 80% similar in sequence to eIF2C2 (Koesters et al., *Genomics* 61 (1999), 210-218). This similarity may indicate functional redundancy, but it is also conceivable that functional RISC may contain eIF2C1 and eIF2C2 heterodimers. The predicted molecular weight of

this heterodimeric complex would be slightly larger than the observed size of 90-160 kDa, but because size fractionation is based on globular shape, we can not disregard this possibility at this time.

Due to the high conservation between the members of the Argonaute family, it is possible that peptides that derive from regions 100% conserved in the 6 predicted proteins, may belong to members others than eIF2C1 and eIF2C2. In this respect, three peptides were identified with masses corresponding to HILI, meaning that this protein might be also a component of RISC.

To precisely assess the protein composition of RISC, reconstitution of the siRNA-mediated target RNA cleavage must be achieved by using recombinant proteins which may be obtained by cloning and expression in suitable bacterial or eukaryotic systems. We expect that the biochemical characterization or the siRNA-mediated target RNA degradation process will have immediate applications, such as the development of cell lines or transgenic animals overexpressing RISC components. The efficiency in targeting endogenous genes in those lines or organisms will be enhanced. Furthermore, a reconstituted in vitro system for RNAi will allow the design of more potent and specific siRNA to achieve gene silencing.

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antisense siRNA (5'-3')

<400> SEQUENCE: 3

ucgaaguauu ccgcguacgu gaugu

25

<210> SEQ ID NO 4

<211> LENGTH: 27

<212> TYPE: RNA

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<213> ORGANISM: Homo sapiens
 <220> FEATURE:
 <221> NAME/KEY: misc_feature
 <223> OTHER INFORMATION: HeLa S100
 antisense siRNA (5'-3')

 <400> SEQUENCE: 4

 ucgaaguauu ccgcuacgu gauguuc 27

 <210> SEQ ID NO 5
 <211> LENGTH: 29
 <212> TYPE: RNA
 <213> ORGANISM: Homo sapiens
 <220> FEATURE:
 <221> NAME/KEY: misc_feature
 <223> OTHER INFORMATION: HeLa S100
 antisense siRNA (5'-3')

 <400> SEQUENCE: 5

 ucgaaguauu ccgcuacgu gauguuac 29

 <210> SEQ ID NO 6
 <211> LENGTH: 15
 <212> TYPE: RNA
 <213> ORGANISM: Homo sapiens
 <220> FEATURE:
 <221> NAME/KEY: misc_feature
 <223> OTHER INFORMATION: HeLa S100
 antisense siRNA (5'-3')

 <400> SEQUENCE: 6

 ucgaaguauu ccgcuacg 15

 <210> SEQ ID NO 7
 <211> LENGTH: 21
 <212> TYPE: RNA
 <213> ORGANISM: Homo sapiens
 <220> FEATURE:
 <221> NAME/KEY: misc_feature
 <223> OTHER INFORMATION: HeLa S100
 antisense siRNA (5'-3')

 <400> SEQUENCE: 7

 ucgaaguauu ccgcuacgu g 21

 <210> SEQ ID NO 8
 <211> LENGTH: 25
 <212> TYPE: RNA
 <213> ORGANISM: Homo sapiens
 <220> FEATURE:
 <221> NAME/KEY: misc_feature
 <223> OTHER INFORMATION: HeLa S100
 antisense siRNA (5'-3')

 <400> SEQUENCE: 8

 ucgaaguauu ccgcuacgu gaugu 25

 <210> SEQ ID NO 9
 <211> LENGTH: 27
 <212> TYPE: RNA
 <213> ORGANISM: Homo sapiens
 <220> FEATURE:
 <221> NAME/KEY: misc_feature
 <223> OTHER INFORMATION: HeLa S100
 antisense siRNA (5'-3')

 <400> SEQUENCE: 9

 ucgaaguauu ccgcuacgu gauguuc 27

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<210> SEQ ID NO 10
<211> LENGTH: 29
<212> TYPE: RNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: HeLa S100
        antisense siRNA (5'-3')

<400> SEQUENCE: 10

ucgaaguauu ccgcuacgu gauguuacac                29

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<210> SEQ ID NO 11
<211> LENGTH: 21
<212> TYPE: RNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: HeLa S100
        antisense siRNA (5'-3')

<400> SEQUENCE: 11

ucgaaguauu ccgcuacgu g                          21

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<210> SEQ ID NO 12
<211> LENGTH: 21
<212> TYPE: DNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(21)
<223> OTHER INFORMATION: RNA/DNA hybrid
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: HeLa S100
        antisense siRNA (5'-3')
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (20)..(20)
<223> OTHER INFORMATION: n = 2'-deoxythymidine
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (21)..(21)
<223> OTHER INFORMATION: n = 2'-deoxyguanosine

<400> SEQUENCE: 12

ucgaaguauu ccgcuacgn n                          21

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<210> SEQ ID NO 13
<211> LENGTH: 21
<212> TYPE: RNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: HeLa S100
        antisense siRNA (5'-3')

<400> SEQUENCE: 13

ucgaaguauu ccgcuacgu u                          21

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<210> SEQ ID NO 14
<211> LENGTH: 21
<212> TYPE: DNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(21)
<223> OTHER INFORMATION: RNA/DNA hybrid
<220> FEATURE:

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<221> NAME/KEY: misc_feature
 <223> OTHER INFORMATION: HeLa S100
 antisense siRNA (5'-3')
 <220> FEATURE:
 <221> NAME/KEY: misc_feature
 <222> LOCATION: (20)..(21)
 <223> OTHER INFORMATION: n = 2'-deoxythymidine

 <400> SEQUENCE: 14

 ucgaaguauu ccgcuacgn n 21

 <210> SEQ ID NO 15
 <211> LENGTH: 21
 <212> TYPE: RNA
 <213> ORGANISM: Homo sapiens
 <220> FEATURE:
 <221> NAME/KEY: misc_feature
 <223> OTHER INFORMATION: HeLa S100
 antisense siRNA (5'-3')

 <400> SEQUENCE: 15

 ucgaaguauu ccgcuacgu g 21

 <210> SEQ ID NO 16
 <211> LENGTH: 21
 <212> TYPE: DNA
 <213> ORGANISM: Homo sapiens
 <220> FEATURE:
 <221> NAME/KEY: misc_feature
 <222> LOCATION: (1)..(21)
 <223> OTHER INFORMATION: RNA/DNA hybrid
 <220> FEATURE:
 <221> NAME/KEY: misc_feature
 <223> OTHER INFORMATION: HeLa S100 cells
 antisense siRNA (5'-3')
 <220> FEATURE:
 <221> NAME/KEY: misc_feature
 <222> LOCATION: (20)..(20)
 <223> OTHER INFORMATION: n = 2'-deoxythymidine
 <220> FEATURE:
 <221> NAME/KEY: misc_feature
 <222> LOCATION: (21)..(21)
 <223> OTHER INFORMATION: n = 2'-deoxyguanosine

 <400> SEQUENCE: 16

 ucgaaguauu ccgcuacgn n 21

 <210> SEQ ID NO 17
 <211> LENGTH: 21
 <212> TYPE: RNA
 <213> ORGANISM: Homo sapiens
 <220> FEATURE:
 <221> NAME/KEY: misc_feature
 <223> OTHER INFORMATION: HeLa S100
 antisense siRNA (5'-3')

 <400> SEQUENCE: 17

 ucgaaguauu ccgcuacgu u 21

 <210> SEQ ID NO 18
 <211> LENGTH: 21
 <212> TYPE: DNA
 <213> ORGANISM: Homo sapiens
 <220> FEATURE:
 <221> NAME/KEY: misc_feature
 <222> LOCATION: (1)..(21)
 <223> OTHER INFORMATION: RNA/DNA hybrid
 <220> FEATURE:
 <221> NAME/KEY: misc_feature
 <223> OTHER INFORMATION: HeLa S100
 antisense siRNA (5'-3')

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<211> LENGTH: 21
<212> TYPE: RNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: HeLa S100
      sense siRNA (5'-3')

<400> SEQUENCE: 23

cguacgcgga auacuucgaa a                                21

<210> SEQ ID NO 24
<211> LENGTH: 21
<212> TYPE: RNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: HeLa S100
      antisense siRNA (5'-3')

<400> SEQUENCE: 24

ucgaaguauu ccgcguacgu u                                21

<210> SEQ ID NO 25
<211> LENGTH: 21
<212> TYPE: RNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: HeLa S100
      sense siRNA (5'-3')

<400> SEQUENCE: 25

cguacgcgga auacuucgaa a                                21

<210> SEQ ID NO 26
<211> LENGTH: 21
<212> TYPE: DNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(21)
<223> OTHER INFORMATION: RNA/DNA hybrid
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: HeLa S100
      antisense siRNA (5'-3')
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (20)..(21)
<223> OTHER INFORMATION: n = 2'-deoxythymidine

<400> SEQUENCE: 26

ucgaaguauu ccgcguacgn n                                21

<210> SEQ ID NO 27
<211> LENGTH: 21
<212> TYPE: DNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(21)
<223> OTHER INFORMATION: RNA/DNA hybrid
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: HeLa S100
      antisense siRNA (5'-3')
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(1)
<223> OTHER INFORMATION: n = 2'-deoxythymidine

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<400> SEQUENCE: 27

ncgaaguauu ccgcguacgu u

21

<210> SEQ ID NO 28

<211> LENGTH: 21

<212> TYPE: DNA

<213> ORGANISM: Homo sapiens

<220> FEATURE:

<221> NAME/KEY: misc_feature

<222> LOCATION: (1)..(21)

<223> OTHER INFORMATION: RNA/DNA hybrid

<220> FEATURE:

<221> NAME/KEY: misc_feature

<223> OTHER INFORMATION: HeLa S100

sense siRNA (5'-3')

<220> FEATURE:

<221> NAME/KEY: misc_feature

<222> LOCATION: (1)..(1)

<223> OTHER INFORMATION: n = 2'-deoxycytidine

<400> SEQUENCE: 28

nguacgcgga auacuucgau u

21

<210> SEQ ID NO 29

<211> LENGTH: 21

<212> TYPE: DNA

<213> ORGANISM: Homo sapiens

<220> FEATURE:

<221> NAME/KEY: misc_feature

<222> LOCATION: (1)..(21)

<223> OTHER INFORMATION: RNA/DNA hybrid

<220> FEATURE:

<221> NAME/KEY: misc_feature

<223> OTHER INFORMATION: HeLa S100

antisense siRNA (5'-3')

<220> FEATURE:

<221> NAME/KEY: misc_feature

<222> LOCATION: (1)..(1)

<223> OTHER INFORMATION: n = 2'-deoxythymidine

<400> SEQUENCE: 29

ncgaaguauu ccgcguacgu u

21

<210> SEQ ID NO 30

<211> LENGTH: 21

<212> TYPE: DNA

<213> ORGANISM: Homo sapiens

<220> FEATURE:

<221> NAME/KEY: misc_feature

<222> LOCATION: (1)..(21)

<223> OTHER INFORMATION: RNA/DNA hybrid

<220> FEATURE:

<221> NAME/KEY: misc_feature

<223> OTHER INFORMATION: HeLa S100 cells

antisense siRNA (5'-3')

<220> FEATURE:

<221> NAME/KEY: misc_feature

<222> LOCATION: (1)..(21)

<223> OTHER INFORMATION: n = 2'-deoxythymidine

<400> SEQUENCE: 30

ncgaaguauu ccgcguacgn n

21

<210> SEQ ID NO 31

<211> LENGTH: 21

<212> TYPE: DNA

<213> ORGANISM: Homo sapiens

<220> FEATURE:

<221> NAME/KEY: misc_feature

<222> LOCATION: (1)..(21)

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<223> OTHER INFORMATION: RNA/DNA hybrid
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: HeLa S100
      sense siRNA (5'-3')
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(1)
<223> OTHER INFORMATION: n = 2'-deoxycytidine
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (20)..(21)
<223> OTHER INFORMATION: n = 2'-deoxythymidine

<400> SEQUENCE: 31

nguacgcgga auacuucgan n                                     21

<210> SEQ ID NO 32
<211> LENGTH: 21
<212> TYPE: DNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(21)
<223> OTHER INFORMATION: RNA/DNA hybrid
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: HeLa S100
      antisense siRNA (5'-3')
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(21)
<223> OTHER INFORMATION: n = 2'-deoxythymidine

<400> SEQUENCE: 32

ncgaaguauu ccgcguacgn n                                     21

<210> SEQ ID NO 33
<211> LENGTH: 21
<212> TYPE: DNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(21)
<223> OTHER INFORMATION: RNA/DNA hybrid
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: HeLa S100
      antisense siRNA (5'-3')
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (1)..(21)
<223> OTHER INFORMATION: n = 2'-deoxythymidine

<400> SEQUENCE: 33

ncgaaguauu ccgcguacgn n                                     21

<210> SEQ ID NO 34
<211> LENGTH: 13
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: peptide fragment of HILI, corresponding to
      position 17-29 of the protein

<400> SEQUENCE: 34

Asn Lys Gln Asp Phe Met Asp Leu Ser Ile Cys Thr Arg
1           5           10

<210> SEQ ID NO 35

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-continued

<211> LENGTH: 14
 <212> TYPE: PRT
 <213> ORGANISM: Homo sapiens
 <220> FEATURE:
 <221> NAME/KEY: MISC_FEATURE
 <223> OTHER INFORMATION: peptide fragment of HILI, corresponding to
 position 436-449 of the protein

<400> SEQUENCE: 35

Thr Glu Tyr Val Ala Glu Ser Phe Leu Asn Cys Leu Arg Arg
 1 5 10

<210> SEQ ID NO 36
 <211> LENGTH: 13
 <212> TYPE: PRT
 <213> ORGANISM: Homo sapiens
 <220> FEATURE:
 <221> NAME/KEY: MISC_FEATURE
 <223> OTHER INFORMATION: peptide fragment of HILI, corresponding to
 position 591-603 of the protein

<400> SEQUENCE: 36

Tyr Asn His Asp Leu Pro Ala Arg Ile Ile Val Tyr Arg
 1 5 10

<210> SEQ ID NO 37
 <211> LENGTH: 35
 <212> TYPE: RNA
 <213> ORGANISM: Homo sapiens
 <220> FEATURE:
 <221> NAME/KEY: misc_feature
 <223> OTHER INFORMATION: HeLa S100
 target RNA

<400> SEQUENCE: 37

aacaucacgu acgcggaaua cuucgaaaug uccgu 35

<210> SEQ ID NO 38
 <211> LENGTH: 21
 <212> TYPE: RNA
 <213> ORGANISM: Homo sapiens
 <220> FEATURE:
 <221> NAME/KEY: misc_feature
 <223> OTHER INFORMATION: HeLa S100
 strand of siRNA duplex

<400> SEQUENCE: 38

cguacgcgga auacuucgau u 21

<210> SEQ ID NO 39
 <211> LENGTH: 21
 <212> TYPE: RNA
 <213> ORGANISM: Homo sapiens
 <220> FEATURE:
 <221> NAME/KEY: misc_feature
 <223> OTHER INFORMATION: HeLa S100
 strand of siRNA duplex

<400> SEQUENCE: 39

ucgaaguauu ccgcguacgu u 21

<210> SEQ ID NO 40
 <211> LENGTH: 21
 <212> TYPE: RNA
 <213> ORGANISM: Homo sapiens
 <220> FEATURE:
 <221> NAME/KEY: misc_feature
 <223> OTHER INFORMATION: HeLa S100
 strand of siRNA duplex

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<400> SEQUENCE: 40

cguacgcgga auacuucgaa a

21

<210> SEQ ID NO 41

<211> LENGTH: 20

<212> TYPE: RNA

<213> ORGANISM: Homo sapiens

<220> FEATURE:

<221> NAME/KEY: misc_feature

<223> OTHER INFORMATION: HeLa S100
strand of siRNA duplex

<400> SEQUENCE: 41

ucgaaguauu ccgcguacgu

20

<210> SEQ ID NO 42

<211> LENGTH: 12

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<220> FEATURE:

<221> NAME/KEY: MISC_FEATURE

<223> OTHER INFORMATION: peptide fragment of eIF2C2, obtained by mass
spectrometry

<400> SEQUENCE: 42

Val Leu Gln Pro Pro Ser Ile Leu Tyr Gly Gly Arg
1 5 10

<210> SEQ ID NO 43

<211> LENGTH: 12

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<220> FEATURE:

<221> NAME/KEY: MISC_FEATURE

<223> OTHER INFORMATION: peptide fragment of eIF2C2, obtained by mass
spectrometry

<400> SEQUENCE: 43

Gln Glu Ile Ile Gln Asp Leu Ala Ala Met Val Arg
1 5 10

<210> SEQ ID NO 44

<211> LENGTH: 12

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<220> FEATURE:

<221> NAME/KEY: MISC_FEATURE

<223> OTHER INFORMATION: peptide fragment of eIF2C2, obtained by mass
spectrometry

<400> SEQUENCE: 44

His Leu Pro Ser Met Arg Tyr Thr Pro Val Gly Arg
1 5 10

<210> SEQ ID NO 45

<211> LENGTH: 12

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<220> FEATURE:

<221> NAME/KEY: MISC_FEATURE

<223> OTHER INFORMATION: peptide fragment of eIF2C2, obtained by mass
spectrometry

<400> SEQUENCE: 45

Lys Leu Thr Asp Asn Gln Thr Ser Thr Met Ile Arg
1 5 10

-continued

<210> SEQ ID NO 46
 <211> LENGTH: 13
 <212> TYPE: PRT
 <213> ORGANISM: Homo sapiens
 <220> FEATURE:
 <221> NAME/KEY: MISC_FEATURE
 <223> OTHER INFORMATION: peptide fragment of eIF2C2, obtained by mass spectrometry

<400> SEQUENCE: 46

Tyr Ala Gln Gly Ala Asp Ser Val Glu Pro Met Phe Arg
 1 5 10

<210> SEQ ID NO 47
 <211> LENGTH: 14
 <212> TYPE: PRT
 <213> ORGANISM: Homo sapiens
 <220> FEATURE:
 <221> NAME/KEY: MISC_FEATURE
 <223> OTHER INFORMATION: peptide fragment of eIF2C2, obtained by mass spectrometry

<400> SEQUENCE: 47

Asp Lys Val Glu Leu Glu Val Thr Leu Pro Gly Glu Gly Lys
 1 5 10

<210> SEQ ID NO 48
 <211> LENGTH: 14
 <212> TYPE: PRT
 <213> ORGANISM: Homo sapiens
 <220> FEATURE:
 <221> NAME/KEY: MISC_FEATURE
 <223> OTHER INFORMATION: peptide fragment of eIF2C2, obtained by mass spectrometry

<400> SEQUENCE: 48

Asp Ala Gly Met Pro Ile Gln Gly Gln Pro Cys Phe Cys Lys
 1 5 10

<210> SEQ ID NO 49
 <211> LENGTH: 14
 <212> TYPE: PRT
 <213> ORGANISM: Homo sapiens
 <220> FEATURE:
 <221> NAME/KEY: MISC_FEATURE
 <223> OTHER INFORMATION: peptide fragment of eIF2C2, obtained by mass spectrometry

<400> SEQUENCE: 49

Thr Gln Ile Phe Gly Asp Arg Lys Pro Val Phe Asp Gly Arg
 1 5 10

<210> SEQ ID NO 50
 <211> LENGTH: 15
 <212> TYPE: PRT
 <213> ORGANISM: Homo sapiens
 <220> FEATURE:
 <221> NAME/KEY: MISC_FEATURE
 <223> OTHER INFORMATION: peptide fragment of eIF2C2, obtained by mass spectrometry

<400> SEQUENCE: 50

Ala Thr Ala Arg Ser Ala Pro Asp Arg Gln Glu Glu Ile Ser Lys
 1 5 10 15

<210> SEQ ID NO 51
 <211> LENGTH: 14
 <212> TYPE: PRT

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<213> ORGANISM: Homo sapiens
 <220> FEATURE:
 <221> NAME/KEY: MISC_FEATURE
 <223> OTHER INFORMATION: peptide fragment of eIF2C2, obtained by mass spectrometry

<400> SEQUENCE: 51

Asp Tyr Gln Pro Gly Ile Thr Phe Ile Val Val Gln Lys Arg
 1 5 10

<210> SEQ ID NO 52
 <211> LENGTH: 14
 <212> TYPE: PRT
 <213> ORGANISM: Homo sapiens
 <220> FEATURE:
 <221> NAME/KEY: MISC_FEATURE
 <223> OTHER INFORMATION: peptide fragment of eIF2C2, obtained by mass spectrometry

<400> SEQUENCE: 52

Ser Ala Pro Asp Arg Gln Glu Glu Ile Ser Lys Leu Met Arg
 1 5 10

<210> SEQ ID NO 53
 <211> LENGTH: 14
 <212> TYPE: PRT
 <213> ORGANISM: Homo sapiens
 <220> FEATURE:
 <221> NAME/KEY: MISC_FEATURE
 <223> OTHER INFORMATION: peptide fragment of eIF2C2, obtained by mass spectrometry

<400> SEQUENCE: 53

Tyr Pro His Leu Pro Cys Leu Gln Val Gly Gln Glu Gln Lys
 1 5 10

<210> SEQ ID NO 54
 <211> LENGTH: 17
 <212> TYPE: PRT
 <213> ORGANISM: Homo sapiens
 <220> FEATURE:
 <221> NAME/KEY: MISC_FEATURE
 <223> OTHER INFORMATION: peptide fragment of eIF2C2, obtained by mass spectrometry

<400> SEQUENCE: 54

Ser Phe Phe Thr Ala Ser Glu Gly Cys Ser Asn Pro Leu Gly Gly Gly
 1 5 10 15

Arg

<210> SEQ ID NO 55
 <211> LENGTH: 23
 <212> TYPE: PRT
 <213> ORGANISM: Homo sapiens
 <220> FEATURE:
 <221> NAME/KEY: MISC_FEATURE
 <223> OTHER INFORMATION: peptide fragment of eIF2C2, obtained by mass spectrometry

<400> SEQUENCE: 55

Tyr His Leu Val Asp Lys Glu His Asp Ser Ala Glu Gly Ser His Thr
 1 5 10 15

Ser Gly Gln Ser Asn Gly Arg
 20

<210> SEQ ID NO 56
 <211> LENGTH: 12

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<212> TYPE: PRT
 <213> ORGANISM: Homo sapiens
 <220> FEATURE:
 <221> NAME/KEY: MISC_FEATURE
 <223> OTHER INFORMATION: peptide fragment of eIF2C1, obtained by mass spectrometry

<400> SEQUENCE: 56

Val Leu Pro Ala Pro Ile Leu Gln Tyr Gly Gly Arg
 1 5 10

<210> SEQ ID NO 57
 <211> LENGTH: 12
 <212> TYPE: PRT
 <213> ORGANISM: Homo sapiens
 <220> FEATURE:
 <221> NAME/KEY: MISC_FEATURE
 <223> OTHER INFORMATION: peptide fragment of eIF2C1, obtained by mass spectrometry

<400> SEQUENCE: 57

Ser Val Ser Ile Pro Ala Pro Ala Tyr Tyr Ala Arg
 1 5 10

<210> SEQ ID NO 58
 <211> LENGTH: 12
 <212> TYPE: PRT
 <213> ORGANISM: Homo sapiens
 <220> FEATURE:
 <221> NAME/KEY: MISC_FEATURE
 <223> OTHER INFORMATION: peptide fragment of eIF2C1, obtained by mass spectrometry

<400> SEQUENCE: 58

Thr Ser Pro Gln Thr Leu Ser Asn Leu Cys Leu Lys
 1 5 10

<210> SEQ ID NO 59
 <211> LENGTH: 13
 <212> TYPE: PRT
 <213> ORGANISM: Homo sapiens
 <220> FEATURE:
 <221> NAME/KEY: MISC_FEATURE
 <223> OTHER INFORMATION: peptide fragment of eIF2C1, obtained by mass spectrometry

<400> SEQUENCE: 59

Tyr Ala Gln Gly Ala Asp Ser Val Glu Pro Met Phe Arg
 1 5 10

<210> SEQ ID NO 60
 <211> LENGTH: 14
 <212> TYPE: PRT
 <213> ORGANISM: Homo sapiens
 <220> FEATURE:
 <221> NAME/KEY: MISC_FEATURE
 <223> OTHER INFORMATION: peptide fragment of eIF2C1, obtained by mass spectrometry

<400> SEQUENCE: 60

Asn Ile Tyr Thr Val Thr Ala Leu Pro Ile Gly Asn Glu Arg
 1 5 10

<210> SEQ ID NO 61
 <211> LENGTH: 14
 <212> TYPE: PRT
 <213> ORGANISM: Homo sapiens
 <220> FEATURE:
 <221> NAME/KEY: MISC_FEATURE

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<223> OTHER INFORMATION: peptide fragment of eIF2C1, obtained by mass spectrometry

<400> SEQUENCE: 61

Val Asp Phe Glu Val Thr Ile Pro Gly Glu Gly Lys Asp Arg
 1 5 10

<210> SEQ ID NO 62

<211> LENGTH: 14

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<220> FEATURE:

<221> NAME/KEY: MISC_FEATURE

<223> OTHER INFORMATION: HeLa S100 cells
 peptide fragment of eIF2C1 obtained by mass spectrometry

<400> SEQUENCE: 62

Asp Ala Gly Met Pro Ile Gln Gly Gln Pro Cys Phe Cys Lys
 1 5 10

<210> SEQ ID NO 63

<211> LENGTH: 14

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<220> FEATURE:

<221> NAME/KEY: MISC_FEATURE

<223> OTHER INFORMATION: peptide fragment of eIF2C1, obtained by mass spectrometry

<400> SEQUENCE: 63

Asn Ile Asp Glu Gln Pro Lys Pro Leu Thr Asp Ser Gln Arg
 1 5 10

<210> SEQ ID NO 64

<211> LENGTH: 14

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<220> FEATURE:

<221> NAME/KEY: MISC_FEATURE

<223> OTHER INFORMATION: peptide fragment of eIF2C1, obtained by mass spectrometry

<400> SEQUENCE: 64

Ser Ala Pro Asp Arg Gln Glu Glu Ile Ser Arg Leu Met Lys
 1 5 10

<210> SEQ ID NO 65

<211> LENGTH: 14

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<220> FEATURE:

<221> NAME/KEY: MISC_FEATURE

<223> OTHER INFORMATION: peptide fragment of eIF2C1, obtained by mass spectrometry

<400> SEQUENCE: 65

Asp Tyr Gln Pro Gly Ile Thr Tyr Ile Val Val Gln Lys Arg
 1 5 10

<210> SEQ ID NO 66

<211> LENGTH: 14

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<220> FEATURE:

<221> NAME/KEY: MISC_FEATURE

<223> OTHER INFORMATION: peptide fragment of eIF2C1, obtained by mass spectrometry

<400> SEQUENCE: 66

-continued

Tyr Pro His Leu Pro Cys Leu Gln Val Gly Gln Glu Gln Lys
1 5 10

<210> SEQ ID NO 67

<211> LENGTH: 17

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<220> FEATURE:

<221> NAME/KEY: MISC_FEATURE

<223> OTHER INFORMATION: peptide fragment of eIF2C1, obtained by mass spectrometry

<400> SEQUENCE: 67

Ser Phe Phe Ser Pro Pro Glu Gly Tyr Tyr His Pro Leu Gly Gly Gly
1 5 10 15

Arg

<210> SEQ ID NO 68

<211> LENGTH: 857

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<220> FEATURE:

<221> NAME/KEY: MISC_FEATURE

<223> OTHER INFORMATION: eIF2C1, predicted protein sequence

<400> SEQUENCE: 68

Met Glu Ala Gly Pro Ser Gly Ala Ala Ala Gly Ala Tyr Leu Pro Pro
1 5 10 15

Leu Gln Gln Val Phe Gln Ala Pro Arg Arg Pro Gly Ile Gly Thr Val
20 25 30

Gly Lys Pro Ile Lys Leu Leu Ala Asn Tyr Phe Glu Val Asp Ile Pro
35 40 45

Lys Ile Asp Val Tyr His Tyr Glu Val Asp Ile Lys Pro Asp Lys Cys
50 55 60

Pro Arg Arg Val Asn Arg Glu Val Val Glu Tyr Met Val Gln His Phe
65 70 75 80

Lys Pro Gln Ile Phe Gly Asp Arg Lys Pro Val Tyr Asp Gly Lys Lys
85 90 95

Asn Ile Tyr Thr Val Thr Ala Leu Pro Ile Gly Asn Glu Arg Val Asp
100 105 110

Phe Glu Val Thr Ile Pro Gly Glu Gly Lys Asp Arg Ile Phe Lys Val
115 120 125

Ser Ile Lys Trp Leu Ala Ile Val Ser Trp Arg Met Leu His Glu Ala
130 135 140

Leu Val Ser Gly Gln Ile Pro Val Pro Leu Glu Ser Val Gln Ala Leu
145 150 155 160

Asp Val Ala Met Arg His Leu Ala Ser Met Arg Tyr Thr Pro Val Gly
165 170 175

Arg Ser Phe Phe Ser Pro Pro Glu Gly Tyr Tyr His Pro Leu Gly Gly
180 185 190

Gly Arg Glu Val Trp Phe Gly Phe His Gln Ser Val Arg Pro Ala Met
195 200 205

Trp Lys Met Met Leu Asn Ile Asp Val Ser Ala Thr Ala Phe Tyr Lys
210 215 220

Ala Gln Pro Val Ile Glu Phe Met Cys Glu Val Leu Asp Ile Arg Asn
225 230 235 240

Ile Asp Glu Gln Pro Lys Pro Leu Thr Asp Ser Gln Arg Val Arg Phe
245 250 255

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Thr Lys Glu Ile Lys Gly Leu Lys Val Glu Val Thr His Cys Gly Gln
 260 265 270
 Met Lys Arg Lys Tyr Arg Val Cys Asn Val Thr Arg Arg Pro Ala Ser
 275 280 285
 His Gln Thr Phe Pro Leu Gln Leu Glu Ser Gly Gln Thr Val Glu Cys
 290 295 300
 Thr Val Ala Gln Tyr Phe Lys Gln Lys Tyr Asn Leu Gln Leu Lys Tyr
 305 310 315 320
 Pro His Leu Pro Cys Leu Gln Val Gly Gln Glu Gln Lys His Thr Tyr
 325 330 335
 Leu Pro Leu Glu Val Cys Asn Ile Val Ala Gly Gln Arg Cys Ile Lys
 340 345 350
 Lys Leu Thr Asp Asn Gln Thr Ser Thr Met Ile Lys Ala Thr Ala Arg
 355 360 365
 Ser Ala Pro Asp Arg Gln Glu Glu Ile Ser Arg Leu Met Lys Asn Ala
 370 375 380
 Ser Tyr Asn Leu Asp Pro Tyr Ile Gln Glu Phe Gly Ile Lys Val Lys
 385 390 395 400
 Asp Asp Met Thr Glu Val Thr Gly Arg Val Leu Pro Ala Pro Ile Leu
 405 410 415
 Gln Tyr Gly Gly Arg Asn Arg Ala Ile Ala Thr Pro Asn Gln Gly Val
 420 425 430
 Trp Asp Met Arg Gly Lys Gln Phe Tyr Asn Gly Ile Glu Ile Lys Val
 435 440 445
 Trp Ala Ile Ala Cys Phe Ala Pro Gln Lys Gln Cys Arg Glu Glu Val
 450 455 460
 Leu Lys Asn Phe Thr Asp Gln Leu Arg Lys Ile Ser Lys Asp Ala Gly
 465 470 475 480
 Met Pro Ile Gln Gly Gln Pro Cys Phe Cys Lys Tyr Ala Gln Gly Ala
 485 490 495
 Asp Ser Val Glu Pro Met Phe Arg His Leu Lys Asn Thr Tyr Ser Gly
 500 505 510
 Leu Gln Leu Ile Ile Val Ile Leu Pro Gly Lys Thr Pro Val Tyr Ala
 515 520 525
 Glu Val Lys Arg Val Gly Asp Thr Leu Leu Gly Met Ala Thr Gln Cys
 530 535 540
 Val Gln Val Lys Asn Val Val Lys Thr Ser Pro Gln Thr Leu Ser Asn
 545 550 555 560
 Leu Cys Leu Lys Ile Asn Val Lys Leu Gly Gly Ile Asn Asn Ile Leu
 565 570 575
 Val Pro His Gln Arg Ser Ala Val Phe Gln Gln Pro Val Ile Phe Leu
 580 585 590
 Gly Ala Asp Val Thr His Pro Pro Ala Gly Asp Gly Lys Lys Pro Ser
 595 600 605
 Ile Thr Ala Val Val Gly Ser Met Asp Ala His Pro Ser Arg Tyr Cys
 610 615 620
 Ala Thr Val Arg Val Gln Arg Pro Arg Gln Glu Ile Ile Glu Asp Leu
 625 630 635 640
 Ser Tyr Met Val Arg Glu Leu Leu Ile Gln Phe Tyr Lys Ser Thr Arg
 645 650 655
 Phe Lys Pro Thr Arg Ile Ile Phe Tyr Arg Asp Gly Val Pro Glu Gly
 660 665 670

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Gln Leu Pro Gln Ile Leu His Tyr Glu Leu Leu Ala Ile Arg Asp Ala
675 680 685

Cys Ile Lys Leu Glu Lys Asp Tyr Gln Pro Gly Ile Thr Tyr Ile Val
690 695 700

Val Gln Lys Arg His His Thr Arg Leu Phe Cys Ala Asp Lys Asn Glu
705 710 715 720

Arg Ile Gly Lys Ser Gly Asn Ile Pro Ala Gly Thr Thr Val Asp Thr
725 730 735

Asn Ile Thr His Pro Phe Glu Phe Asp Phe Tyr Leu Cys Ser His Ala
740 745 750

Gly Ile Gln Gly Thr Ser Arg Pro Ser His Tyr Tyr Val Leu Trp Asp
755 760 765

Asp Asn Arg Phe Thr Ala Asp Glu Leu Gln Ile Leu Thr Tyr Gln Leu
770 775 780

Cys His Thr Tyr Val Arg Cys Thr Arg Ser Val Ser Ile Pro Ala Pro
785 790 795 800

Ala Tyr Tyr Ala Arg Leu Val Ala Phe Arg Ala Arg Tyr His Leu Val
805 810 815

Asp Lys Glu His Asp Ser Gly Glu Gly Ser His Ile Ser Gly Gln Ser
820 825 830

Asn Gly Arg Asp Pro Gln Ala Leu Ala Lys Ala Val Gln Val His Gln
835 840 845

Asp Thr Leu Arg Thr Met Tyr Phe Ala
850 855

<210> SEQ ID NO 69
 <211> LENGTH: 860
 <212> TYPE: PRT
 <213> ORGANISM: Homo sapiens
 <220> FEATURE:
 <221> NAME/KEY: MISC_FEATURE
 <223> OTHER INFORMATION: eIF2C2, predicted protein sequence

<400> SEQUENCE: 69

Met Gly Val Leu Ser Ala Ile Pro Ala Leu Ala Pro Pro Ala Pro Pro
1 5 10 15

Pro Pro Ile Gln Gly Tyr Ala Phe Lys Pro Pro Pro Arg Pro Asp Phe
20 25 30

Gly Thr Ser Gly Arg Thr Ile Lys Leu Gln Ala Asn Phe Phe Glu Met
35 40 45

Asp Ile Pro Lys Ile Asp Ile Tyr His Tyr Glu Leu Asp Ile Lys Pro
50 55 60

Glu Lys Cys Pro Arg Arg Val Asn Arg Glu Ile Val Glu His Met Val
65 70 75 80

Gln His Phe Lys Thr Gln Ile Phe Gly Asp Arg Lys Pro Val Phe Asp
85 90 95

Gly Arg Lys Asn Leu Tyr Thr Ala Met Pro Leu Pro Ile Gly Arg Asp
100 105 110

Lys Val Glu Leu Glu Val Thr Leu Pro Gly Glu Gly Lys Asp Arg Ile
115 120 125

Phe Lys Val Ser Ile Lys Trp Val Ser Cys Val Ser Leu Gln Ala Leu
130 135 140

His Asp Ala Leu Ser Gly Arg Leu Pro Ser Val Pro Phe Glu Thr Ile
145 150 155 160

Gln Ala Leu Asp Val Val Met Arg His Leu Pro Ser Met Arg Tyr Thr
165 170 175

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Pro Val Gly Arg Ser Phe Phe Thr Ala Ser Glu Gly Cys Ser Asn Pro
 180 185 190

Leu Gly Gly Gly Arg Glu Val Trp Phe Gly Phe His Gln Ser Val Arg
 195 200 205

Pro Ser Leu Trp Lys Met Met Leu Asn Ile Asp Val Ser Ala Thr Ala
 210 215 220

Phe Tyr Lys Ala Gln Pro Val Ile Glu Phe Val Cys Glu Val Leu Asp
 225 230 235 240

Phe Lys Ser Ile Glu Glu Gln Gln Lys Pro Leu Thr Asp Ser Gln Arg
 245 250 255

Val Lys Phe Thr Lys Glu Ile Lys Gly Leu Lys Val Glu Ile Thr His
 260 265 270

Cys Gly Gln Met Lys Arg Lys Tyr Arg Val Cys Asn Val Thr Arg Arg
 275 280 285

Pro Ala Ser His Gln Thr Phe Pro Leu Gln Gln Glu Ser Gly Gln Thr
 290 295 300

Val Glu Cys Thr Val Ala Gln Tyr Phe Lys Asp Arg His Lys Leu Val
 305 310 315 320

Leu Arg Tyr Pro His Leu Pro Cys Leu Gln Val Gly Gln Glu Gln Lys
 325 330 335

His Thr Tyr Leu Pro Leu Glu Val Cys Asn Ile Val Ala Gly Gln Arg
 340 345 350

Cys Ile Lys Lys Leu Thr Asp Asn Gln Thr Ser Thr Met Ile Arg Ala
 355 360 365

Thr Ala Arg Ser Ala Pro Asp Arg Gln Glu Glu Ile Ser Lys Leu Met
 370 375 380

Arg Ser Ala Ser Phe Asn Thr Asp Pro Tyr Val Arg Glu Phe Gly Ile
 385 390 395 400

Met Val Lys Asp Glu Met Thr Asp Val Thr Gly Arg Val Leu Gln Pro
 405 410 415

Pro Ser Ile Leu Tyr Gly Gly Arg Asn Lys Ala Ile Ala Thr Pro Val
 420 425 430

Gln Gly Val Trp Asp Met Arg Asn Lys Gln Phe His Thr Gly Ile Glu
 435 440 445

Ile Lys Val Trp Ala Ile Ala Cys Phe Ala Pro Gln Arg Gln Cys Thr
 450 455 460

Glu Val His Leu Lys Ser Phe Thr Glu Gln Leu Arg Lys Ile Ser Arg
 465 470 475 480

Asp Ala Gly Met Pro Ile Gln Gly Gln Pro Cys Phe Cys Lys Tyr Ala
 485 490 495

Gln Gly Ala Asp Ser Val Glu Pro Met Phe Arg His Leu Lys Asn Thr
 500 505 510

Tyr Ala Gly Leu Gln Leu Val Val Val Ile Leu Pro Gly Lys Thr Pro
 515 520 525

Val Tyr Ala Glu Val Lys Arg Val Gly Asp Thr Val Leu Gly Met Ala
 530 535 540

Thr Gln Cys Val Gln Met Lys Asn Val Gln Arg Thr Thr Pro Gln Thr
 545 550 555 560

Leu Ser Asn Leu Cys Leu Lys Ile Asn Val Lys Leu Gly Gly Val Asn
 565 570 575

Asn Ile Leu Leu Pro Gln Gly Arg Pro Pro Val Phe Gln Gln Pro Val
 580 585 590

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His Phe Gln Val Gln Ile Pro Lys Ile Asp Val Tyr His Tyr Asp Val
 100 105 110

Asp Ile Lys Pro Glu Lys Arg Pro Arg Arg Val Asn Arg Glu Val Val
 115 120 125

Asp Thr Met Val Arg His Phe Lys Met Gln Ile Phe Gly Asp Arg Gln
 130 135 140

Pro Gly Tyr Asp Gly Lys Arg Asn Met Tyr Thr Ala His Pro Leu Pro
 145 150 155 160

Ile Gly Arg Asp Arg Val Asp Met Glu Val Thr Leu Pro Gly Glu Gly
 165 170 175

Lys Asp Gln Thr Phe Lys Val Ser Val Gln Trp Val Ser Val Val Ser
 180 185 190

Leu Gln Leu Leu Leu Glu Ala Leu Ala Gly His Leu Asn Glu Val Pro
 195 200 205

Asp Asp Ser Val Gln Ala Leu Asp Val Ile Thr Arg His Leu Pro Ser
 210 215 220

Met Arg Tyr Thr Pro Val Gly Arg Ser Phe Phe Ser Pro Pro Glu Gly
 225 230 235 240

Tyr Tyr His Pro Leu Gly Gly Gly Arg Glu Val Trp Phe Gly Phe His
 245 250 255

Gln Ser Val Arg Pro Ala Met Trp Asn Met Met Leu Asn Ile Asp Val
 260 265 270

Ser Ala Thr Ala Phe Tyr Arg Ala Gln Pro Ile Ile Glu Phe Met Cys
 275 280 285

Glu Val Leu Asp Ile Gln Asn Ile Asn Glu Gln Thr Lys Pro Leu Thr
 290 295 300

Asp Ser Gln Arg Val Lys Phe Thr Lys Glu Ile Arg Gly Leu Lys Val
 305 310 315 320

Glu Val Thr His Cys Gly Gln Met Lys Arg Lys Tyr Arg Val Cys Asn
 325 330 335

Val Thr Arg Arg Pro Ala Ser His Gln Thr Phe Pro Leu Gln Leu Glu
 340 345 350

Asn Gly Gln Ala Met Glu Cys Thr Val Ala Gln Tyr Phe Lys Gln Lys
 355 360 365

Tyr Ser Leu Gln Leu Lys Tyr Pro His Leu Pro Cys Leu Gln Val Gly
 370 375 380

Gln Glu Gln Lys His Thr Tyr Leu Pro Leu Glu Val Cys Asn Ile Val
 385 390 395 400

Ala Gly Gln Arg Cys Ile Lys Lys Leu Thr Asp Asn Gln Thr Ser Thr
 405 410 415

Met Ile Lys Ala Thr Ala Arg Ser Ala Pro Asp Arg Gln Glu Glu Ile
 420 425 430

Ser Arg Leu Val Lys Ser Asn Ser Met Val Gly Gly Pro Asp Pro Tyr
 435 440 445

Leu Lys Glu Phe Gly Ile Val Val His Asn Glu Met Thr Glu Leu Thr
 450 455 460

Gly Arg Val Leu Pro Ala Pro Met Leu Gln Tyr Gly Gly Arg Asn Lys
 465 470 475 480

Thr Val Ala Thr Pro Asn Gln Gly Val Trp Asp Met Arg Gly Lys Gln
 485 490 495

Phe Tyr Ala Gly Ile Glu Ile Lys Val Trp Ala Val Ala Cys Phe Ala
 500 505 510

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<210> SEQ ID NO 71
<211> LENGTH: 855
<212> TYPE: PRT
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<223> OTHER INFORMATION: eIF2C4, predicted protein sequence

<400> SEQUENCE: 71

Ala Gly Pro Ala Gly Ala Gln Pro Leu Leu Met Val Pro Arg Arg Pro
1          5          10          15
Gly Tyr Gly Thr Met Gly Lys Pro Ile Lys Leu Leu Ala Asn Cys Phe
20          25          30
Gln Val Glu Ile Pro Lys Ile Asp Val Tyr Leu Tyr Glu Val Asp Ile
35          40          45
Lys Pro Asp Lys Cys Pro Arg Arg Val Asn Arg Glu Val Val Asp Ser
50          55          60
Met Val Gln His Phe Lys Val Thr Ile Phe Gly Asp Arg Arg Pro Val
65          70          75          80
Tyr Asp Gly Lys Arg Ser Leu Tyr Thr Ala Asn Pro Leu Pro Val Ala
85          90          95
Thr Thr Gly Val Asp Leu Asp Val Thr Leu Pro Gly Glu Gly Gly Lys
100         105         110
Asp Arg Pro Phe Lys Val Ser Ile Lys Phe Val Ser Arg Val Ser Trp
115         120         125
His Leu Leu His Glu Val Leu Thr Gly Arg Thr Leu Pro Glu Pro Leu
130         135         140
Glu Leu Asp Lys Pro Ile Ser Thr Asn Pro Val His Ala Val Asp Val
145         150         155         160
Val Leu Arg His Leu Pro Ser Met Lys Tyr Thr Pro Val Gly Arg Ser
165         170         175
Phe Phe Ser Ala Pro Glu Gly Tyr Asp His Pro Leu Gly Gly Gly Arg
180         185         190
Glu Val Trp Phe Gly Phe His Gln Ser Val Arg Pro Ala Met Trp Lys
195         200         205
Met Met Leu Asn Ile Asp Val Ser Ala Thr Ala Phe Tyr Lys Ala Gln
210         215         220
Pro Val Ile Gln Phe Met Cys Glu Val Leu Asp Ile His Asn Ile Asp
225         230         235         240
Glu Gln Pro Arg Pro Leu Thr Asp Ser His Arg Val Lys Phe Thr Lys
245         250         255
Glu Ile Lys Gly Leu Lys Val Glu Val Thr His Cys Gly Thr Met Arg
260         265         270
Arg Lys Tyr Arg Val Cys Asn Val Thr Arg Arg Pro Ala Ser His Gln
275         280         285
Thr Phe Pro Leu Gln Leu Glu Asn Gly Gln Thr Val Glu Arg Thr Val
290         295         300
Ala Gln Tyr Phe Arg Glu Lys Tyr Thr Leu Gln Leu Lys Tyr Pro His
305         310         315         320
Leu Pro Cys Leu Gln Val Gly Gln Glu Gln Lys His Thr Tyr Leu Pro
325         330         335
Leu Glu Val Cys Asn Ile Val Ala Gly Gln Arg Cys Ile Lys Lys Leu
340         345         350
Thr Asp Asn Gln Thr Ser Thr Met Ile Lys Ala Thr Ala Arg Ser Ala
355         360         365

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Pro	Asp	Arg	Gln	Glu	Glu	Ile	Ser	Arg	Leu	Val	Arg	Ser	Ala	Asn	Tyr
	370					375					380				
Glu	Thr	Asp	Pro	Phe	Val	Gln	Glu	Phe	Gln	Phe	Lys	Val	Arg	Asp	Glu
385					390				395						400
Met	Ala	His	Val	Thr	Gly	Arg	Val	Leu	Pro	Ala	Pro	Met	Leu	Gln	Tyr
			405					410						415	
Gly	Gly	Arg	Asn	Arg	Thr	Val	Ala	Thr	Pro	Ser	His	Gly	Val	Trp	Asp
			420					425					430		
Met	Arg	Gly	Lys	Gln	Phe	His	Thr	Gly	Val	Glu	Ile	Lys	Met	Trp	Ala
		435					440					445			
Ile	Ala	Cys	Phe	Ala	Thr	Gln	Arg	Gln	Cys	Arg	Glu	Glu	Ile	Leu	Lys
450						455					460				
Gly	Phe	Thr	Asp	Gln	Leu	Arg	Lys	Ile	Ser	Lys	Asp	Ala	Gly	Met	Pro
465					470					475					480
Ile	Gln	Gly	Gln	Pro	Cys	Phe	Cys	Lys	Tyr	Ala	Gln	Gly	Ala	Asp	Ser
				485					490					495	
Val	Glu	Pro	Met	Phe	Arg	His	Leu	Lys	Asn	Thr	Tyr	Ser	Gly	Leu	Gln
			500					505					510		
Leu	Ile	Ile	Val	Ile	Leu	Pro	Gly	Lys	Thr	Pro	Val	Tyr	Ala	Glu	Val
		515					520					525			
Lys	Arg	Val	Gly	Asp	Thr	Leu	Leu	Gly	Met	Ala	Thr	Gln	Cys	Val	Gln
	530					535					540				
Val	Lys	Asn	Val	Ile	Lys	Thr	Ser	Pro	Gln	Thr	Leu	Ser	Asn	Leu	Cys
545					550					555					560
Leu	Lys	Ile	Asn	Val	Lys	Leu	Gly	Gly	Ile	Asn	Asn	Ile	Leu	Val	Pro
			565						570					575	
His	Gln	Arg	Pro	Ser	Val	Phe	Gln	Gln	Pro	Val	Ile	Phe	Leu	Gly	Ala
			580					585					590		
Asp	Val	Thr	His	Pro	Pro	Ala	Gly	Asp	Gly	Lys	Lys	Pro	Ser	Ile	Ala
	595						600					605			
Ala	Val	Val	Gly	Ser	Met	Asp	Ala	His	Pro	Ser	Arg	Tyr	Cys	Ala	Thr
	610					615					620				
Val	Arg	Val	Gln	Arg	Pro	Arg	Gln	Glu	Ile	Ile	Gln	Asp	Leu	Ala	Ser
625					630					635					640
Met	Val	Arg	Glu	Leu	Leu	Ile	Gln	Phe	Tyr	Lys	Ser	Thr	Arg	Phe	Lys
			645						650					655	
Pro	Thr	Arg	Ile	Ile	Phe	Tyr	Arg	Asp	Gly	Val	Ser	Glu	Gly	Gln	Phe
			660					665					670		
Arg	Gln	Val	Leu	Tyr	Tyr	Glu	Leu	Leu	Ala	Ile	Arg	Glu	Ala	Cys	Ile
		675					680					685			
Ser	Leu	Glu	Lys	Asp	Tyr	Gln	Pro	Gly	Ile	Thr	Tyr	Ile	Val	Val	Gln
	690					695					700				
Lys	Arg	His	His	Thr	Arg	Leu	Phe	Cys	Ala	Asp	Arg	Thr	Glu	Arg	Val
705					710					715					720
Gly	Arg	Ser	Gly	Asn	Ile	Pro	Ala	Gly	Thr	Thr	Val	Asp	Thr	Asp	Ile
				725					730					735	
Thr	His	Pro	Tyr	Glu	Phe	Asp	Phe	Tyr	Leu	Cys	Ser	His	Ala	Gly	Ile
			740					745					750		
Gln	Gly	Thr	Ser	Arg	Pro	Ser	His	Tyr	His	Val	Leu	Trp	Asp	Asp	Asn
		755					760					765			
Cys	Phe	Thr	Ala	Asp	Glu	Leu	Gln	Leu	Leu	Thr	Tyr	Gln	Leu	Cys	His
	770					775					780				
Thr	Tyr	Val	Arg	Cys	Thr	Arg	Ser	Val	Ser	Ile	Pro	Ala	Pro	Ala	Tyr

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705	710	715	720
Leu Tyr Tyr Asn Trp Pro Gly Ile Val Ser Val Pro Ala Pro Cys Gln			
	725	730	735
Tyr Ala His Lys Leu Thr Phe Leu Val Ala Gln Ser Ile His Lys Glu			
	740	745	750
Pro Ser Leu Glu Leu Ala Asn His Leu Phe Tyr Leu			
	755	760	

<210> SEQ ID NO 73
 <211> LENGTH: 861
 <212> TYPE: PRT
 <213> ORGANISM: Homo sapiens
 <220> FEATURE:
 <221> NAME/KEY: MISC_FEATURE
 <223> OTHER INFORMATION: HIWI, predicted protein sequence

<400> SEQUENCE: 73

Met Thr Gly Arg Ala Arg Ala Arg Ala Arg Gly Arg Ala Arg Gly Gln			
1	5	10	15
Glu Thr Ala Gln Leu Val Gly Ser Thr Ala Ser Gln Gln Pro Gly Tyr			
	20	25	30
Ile Gln Pro Arg Pro Gln Pro Pro Pro Ala Glu Gly Glu Leu Phe Gly			
	35	40	45
Arg Gly Arg Gln Arg Gly Thr Ala Gly Gly Thr Ala Lys Ser Gln Gly			
	50	55	60
Leu Gln Ile Ser Ala Gly Phe Gln Glu Leu Ser Leu Ala Glu Arg Gly			
65	70	75	80
Gly Arg Arg Arg Asp Phe His Asp Leu Gly Val Asn Thr Arg Gln Asn			
	85	90	95
Leu Asp His Val Lys Glu Ser Lys Thr Gly Ser Ser Gly Ile Ile Val			
	100	105	110
Arg Leu Ser Thr Asn His Phe Arg Leu Thr Ser Arg Pro Gln Trp Ala			
	115	120	125
Leu Tyr Gln Tyr His Ile Asp Tyr Asn Pro Leu Met Glu Ala Arg Arg			
	130	135	140
Leu Arg Ser Ala Leu Leu Phe Gln His Glu Asp Leu Ile Gly Lys Cys			
145	150	155	160
His Ala Phe Asp Gly Thr Ile Leu Phe Leu Pro Lys Arg Leu Gln Gln			
	165	170	175
Lys Val Thr Glu Val Phe Ser Lys Thr Arg Asn Gly Glu Asp Val Arg			
	180	185	190
Ile Thr Ile Thr Leu Thr Asn Glu Leu Pro Pro Thr Ser Pro Thr Cys			
	195	200	205
Leu Gln Phe Tyr Asn Ile Ile Phe Arg Arg Leu Leu Lys Ile Met Asn			
	210	215	220
Leu Gln Gln Ile Gly Arg Asn Tyr Tyr Asn Pro Asn Asp Pro Ile Asp			
225	230	235	240
Ile Pro Ser His Arg Leu Val Ile Trp Pro Gly Phe Thr Thr Ser Ile			
	245	250	255
Leu Gln Tyr Glu Asn Ser Ile Met Leu Cys Thr Asp Val Ser His Lys			
	260	265	270
Val Leu Arg Ser Glu Thr Val Leu Asp Phe Met Phe Asn Phe Tyr His			
	275	280	285
Gln Thr Glu Glu His Lys Phe Gln Glu Gln Val Ser Lys Glu Leu Ile			
	290	295	300

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Gly	Leu	Val	Val	Leu	Thr	Lys	Tyr	Asn	Asn	Lys	Thr	Tyr	Arg	Val	Asp	305	310	315	320
Asp	Ile	Asp	Trp	Asp	Gln	Asn	Pro	Lys	Ser	Thr	Phe	Lys	Lys	Ala	Asp	325	330	335	
Gly	Ser	Glu	Val	Ser	Phe	Leu	Glu	Tyr	Tyr	Arg	Lys	Gln	Tyr	Asn	Gln	340	345	350	
Glu	Ile	Thr	Asp	Leu	Lys	Gln	Pro	Val	Leu	Val	Ser	Gln	Pro	Lys	Arg	355	360	365	
Arg	Arg	Gly	Pro	Gly	Gly	Thr	Leu	Pro	Gly	Pro	Ala	Met	Leu	Ile	Pro	370	375	380	
Glu	Leu	Cys	Tyr	Leu	Thr	Gly	Leu	Thr	Asp	Lys	Met	Arg	Asn	Asp	Phe	385	390	395	400
Asn	Val	Met	Lys	Asp	Leu	Ala	Val	His	Thr	Arg	Leu	Thr	Pro	Glu	Gln	405	410	415	
Arg	Gln	Arg	Glu	Val	Gly	Arg	Leu	Ile	Asp	Tyr	Ile	His	Lys	Asn	Asp	420	425	430	
Asn	Val	Gln	Arg	Glu	Leu	Arg	Asp	Trp	Gly	Leu	Ser	Phe	Asp	Ser	Asn	435	440	445	
Leu	Leu	Ser	Phe	Ser	Gly	Arg	Ile	Leu	Gln	Thr	Glu	Lys	Ile	His	Gln	450	455	460	
Gly	Gly	Lys	Thr	Phe	Asp	Tyr	Asn	Pro	Gln	Phe	Ala	Asp	Trp	Ser	Lys	465	470	475	480
Glu	Thr	Arg	Gly	Ala	Pro	Leu	Ile	Ser	Val	Lys	Pro	Leu	Asp	Asn	Trp	485	490	495	
Leu	Leu	Ile	Tyr	Thr	Arg	Arg	Asn	Tyr	Glu	Ala	Ala	Asn	Ser	Leu	Ile	500	505	510	
Gln	Asn	Leu	Phe	Lys	Val	Thr	Pro	Ala	Met	Gly	Met	Gln	Met	Arg	Lys	515	520	525	
Ala	Ile	Met	Ile	Glu	Val	Asp	Asp	Arg	Thr	Glu	Ala	Tyr	Leu	Arg	Val	530	535	540	
Leu	Gln	Gln	Lys	Val	Thr	Ala	Asp	Thr	Gln	Ile	Val	Val	Cys	Leu	Leu	545	550	555	560
Ser	Ser	Asn	Arg	Lys	Asp	Lys	Tyr	Asp	Ala	Ile	Lys	Lys	Tyr	Leu	Cys	565	570	575	
Thr	Asp	Cys	Pro	Thr	Pro	Ser	Gln	Cys	Val	Val	Ala	Arg	Thr	Leu	Gly	580	585	590	
Lys	Gln	Gln	Thr	Val	Met	Ala	Ile	Ala	Thr	Lys	Ile	Ala	Leu	Gln	Met	595	600	605	
Asn	Cys	Lys	Met	Gly	Gly	Glu	Leu	Trp	Arg	Val	Asp	Ile	Pro	Leu	Lys	610	615	620	
Leu	Val	Met	Ile	Val	Gly	Ile	Asp	Cys	Tyr	His	Asp	Met	Thr	Ala	Gly	625	630	635	640
Arg	Arg	Ser	Ile	Ala	Gly	Phe	Val	Ala	Ser	Ile	Asn	Glu	Gly	Met	Thr	645	650	655	
Arg	Trp	Phe	Ser	Arg	Cys	Ile	Phe	Gln	Asp	Arg	Gly	Gln	Glu	Leu	Val	660	665	670	
Asp	Gly	Leu	Lys	Val	Cys	Leu	Gln	Ala	Ala	Leu	Arg	Ala	Trp	Asn	Ser	675	680	685	
Cys	Asn	Glu	Tyr	Met	Pro	Ser	Arg	Ile	Ile	Val	Tyr	Arg	Asp	Gly	Val	690	695	700	
Gly	Asp	Gly	Gln	Leu	Lys	Thr	Leu	Val	Asn	Tyr	Glu	Val	Pro	Gln	Phe	705	710	715	720
Leu	Asp	Cys	Leu	Lys	Ser	Ile	Gly	Arg	Gly	Tyr	Asn	Pro	Arg	Leu	Thr				

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725				730				735							
Val	Ile	Val	Val	Lys	Lys	Arg	Val	Asn	Thr	Arg	Phe	Phe	Ala	Gln	Ser
			740												750
Gly	Gly	Arg	Leu	Gln	Asn	Pro	Leu	Pro	Gly	Thr	Val	Ile	Asp	Val	Glu
			755												765
Val	Thr	Arg	Pro	Glu	Trp	Tyr	Asp	Phe	Phe	Ile	Val	Ser	Gln	Ala	Val
			770												780
Arg	Ser	Gly	Ser	Val	Ser	Pro	Thr	His	Tyr	Asn	Val	Ile	Tyr	Asp	Asn
															800
Ser	Gly	Leu	Lys	Pro	Asp	His	Ile	Gln	Arg	Leu	Thr	Tyr	Lys	Leu	Cys
															815
His	Ile	Tyr	Tyr	Asn	Trp	Pro	Gly	Val	Ile	Arg	Val	Pro	Ala	Pro	Cys
															830
Gln	Tyr	Ala	His	Lys	Leu	Ala	Phe	Leu	Val	Gly	Gln	Ser	Ile	His	Arg
															845
Glu	Pro	Asn	Leu	Ser	Leu	Ser	Asn	Arg	Leu	Tyr	Tyr	Leu			

<210> SEQ ID NO 74

<211> LENGTH: 2571

<212> TYPE: DNA

<213> ORGANISM: Homo sapiens

<220> FEATURE:

<221> NAME/KEY: misc_feature

<223> OTHER INFORMATION: eIF2C1, cDNA sequence of predicted ORF

<400> SEQUENCE: 74

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atggaagcgg gaccctcggg agcagctgcg ggcgcttacc tgccccccct gcagcaggtg    60
ttccaggcac ctgcgccggc tggcattggc actgtgggga aaccaatcaa gctcctggcc    120
aattactttg aggtggacat ccctaagatc gacgtgtacc actacgaggt ggacatcaag    180
ccggataagt gtccccgtag agtcaaccgg gaagtgggtg aatacatggt ccagcatttc    240
aagcctcaga tctttggtga tcgcaagcct gtgtatgatg gaaagaagaa catttacct    300
gtcacagcac tgcccattgg caacgaacgg gtgcactttg aggtgacaat ccctggggaa    360
gggaaggatc gaatctttaa ggtctccatc aagtggctag ccattgtgag ctggcgaatg    420
ctgcatgagg cctgggtcag cggccagatc cctgttcctt tggagtctgt gcaagccctg    480
gatgtggcca tgaggcacct ggcacccatg aggtacaccc ctgtgggccc ctccctcttc    540
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caccagtctg tgcgccctgc catgtggaag atgatgctca acattgatgt ctcagccact    660
gccttttata aggcacagcc agtgattgag ttcattgtgt aggtgctgga catcaggaac    720
atagatgagc agcccaagcc cctcacggac tctcagcgcg ttcgcttcac caaggagatc    780
aagggcctga aggtggaagt caccactgt ggacagatga agaggaagta ccgctgtgtg    840
aatgttaccg gtcgccctgc tagccatcag acattcccct tacagctgga gagtggacag    900
actgtggagt gcacagtggc acagtatttc aagcagaaat ataacctca gctcaagtat    960
ccccatctgc cctgcctaca agttggccag gaacaaaagc atacctacct tcccctagag   1020
gtctgtaaca ttgtggctgg gcagcgctgt attaaaaagc tgaccgaaa ccagacctcg   1080
accatgataa aggccacagc tagatccgct ccagacagac aggaggagat cagtcgctcg   1140
atgaagaatg ccagctacaa cttagatccc tacatccagg aatttgggat caaagtgaag   1200
gatgacatga cggaggtgac agggcgagtg ctgccggcgc ccatcttgca gtacggcggc   1260

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cgaaccggg ccattgccac acccaatcag ggtgtctggg acatgcgggg gaaacagttc 1320
tacaatggga ttgagatcaa agtctggggc atcgctgct tcgcaccca aaaacagtgt 1380
cgagaagagg tgctcaagaa cttcacagac cagctgcgga agatttccaa ggatgcgggg 1440
atgcctatcc aggggtcaacc ttgtttctgc aaatatgcac agggggcaga cagcgtggag 1500
cctatgttcc ggcatctcaa gaacacctac tcagggtgc agctcattat tgtcatcctg 1560
ccaggaaga cgccggtgta tgctgaggtg aaacgtgtcg gagatacact cttgggaatg 1620
gctacgcagt gtgtgcaggt gaagaacgtg gtcaagacct cacctcagac tctgtccaac 1680
ctctgcctca agatcaatgt caaacttggg ggcattaaca acatcctagt cccacaccag 1740
cgctctgccg ttttcaaca gccagtgata ttctggggag cagatgttac acaccccca 1800
gcaggggatg gaaaaaac ttctatcaca gcagtggtag gcagtatgga tgcccacccc 1860
agccgatact gtgctactgt gcgggtacag cgaccacggc aagagatcat tgaagacttg 1920
tctacatgg tgcgtgagct cctcatccaa ttctacaagt ccaccggtt caagcctacc 1980
cgcatcatct tctaccgaga tgggggtgcct gaaggccagc taccacagat actccactat 2040
gagctactgg ccattcgtga tgctgcac aaactggaaa aggactacca gcctgggatc 2100
acttatattg tgggtgcagaa acgccatcac accgccttt tctgtgctga caagaatgag 2160
cgaattggga agagtggtaa catcccagct gggaccacag tggacaccaa catcacccac 2220
ccatttgagt ttgacttcta tctgtgcagc cacgcaggca tccaggcac cagccgacca 2280
tcccattact atgttctttg ggatgacaac cgtttcacag cagatgagct ccagatcctg 2340
acgtaccagc tgtgccacac ttacgtacga tgacacgct ctgtctctat cccagcacct 2400
gcctactatg cccgctggt ggctttccgg gcacgatacc acctggtgga caaggagcat 2460
gacagtggag aggggagcca catatcgggg cagagcaatg ggcgggaccc ccaggccctg 2520
gcaaagccg tgcaggttca ccaggatact ctgcgcacca tgtacttcgc t 2571

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<210> SEQ ID NO 75

<211> LENGTH: 2580

<212> TYPE: DNA

<213> ORGANISM: Homo sapiens

<220> FEATURE:

<221> NAME/KEY: misc_feature

<223> OTHER INFORMATION: eIF2C2, cDNA sequence of predicted ORF

<400> SEQUENCE: 75

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atgggtgttc tctctgcat tcccgcactt gcacctcctg cgccgccgcc ccccatccaa 60
ggatatgctt tcaagcctcc acctagacct gactttggga cctccgggag aacaatcaaa 120
ttacaggcca atttcttca aatggacatc cccaaaattg acatctatca ttatgaattg 180
gatatcaagc cagagaagtg cccgaggaga gttaacaggg aaatcgtgga acacatggtc 240
cagcacttta aaacacagat ctttggggat cggaagcccg tgtttgacgg caggaagaat 300
ctatacacag ccatgccctt tccgattggg agggacaagg tggagctgga ggtcacgctg 360
ccaggagaag gcaaggatcg catcttcaag gtgtccatca agtgggtgtc ctgcgtgagc 420
ttgcaggcgt tacacgatgc actttcaggg cggtgcca gcgtcccttt tgagacgatc 480
caggccctgg acgtggtcat gaggcacttg ccatccatga ggtacacccc cgtggggccg 540
tccttcttca cccgctccga aggctgctct aacctcttg gcgggggccc agaagtgtgg 600
tttggttcc atcagtcctg ccggccttct ctctggaaaa tgatgctgaa tattgatgtg 660
tcagcaacag cgttttacia ggcacagcca gtaatcgagt ttgtttgtga agttttggat 720

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tttaaaagta ttgaagaaca acaaaaacct ctgacagatt cccaaagggt aaagtttacc 780
aaagaaatta aaggtctaaa ggtggagata acgcactgtg ggcagatgaa gaggaagtac 840
cgtgtctgca atgtgacctg gcgccccgcc agtcaccaa cattccccct gcagcaggag 900
agcgggcaga cgggtggagt cacgggtggc cagtatttca aggacaggca caagttggtt 960
ctgcgctacc cccacctccc atgtttacaa gtcggacagg agcagaaaaca cacctacctt 1020
cccctggagg tctgtaacat tgtggcagga caaagatgta ttaaaaaatt aacggacaat 1080
cagacctcaa ccatgatcag agcaactgct aggtcggcgc cccgatcggca agaagagatt 1140
agcaaattga tgcgaagtgc aagtttcaac acagatccat acgtccgtga atttggaatc 1200
atggtcaaag atgagatgac agacgtgact gggcgggtgc tgcagccgcc ctccatcctc 1260
tacgggggca ggaataaagc tattgcgacc cctgtccagg gcgtctggga catgcggaac 1320
aagcagttcc acacgggcat cgagatcaag gtgtgggcca ttgcgtgctt cccccccag 1380
cgccagtgca cgggaagtcca tctgaagtcc ttcacagagc agctcagaaa gatctcgaga 1440
gacgctggca tgcccatcca gggccagccg tgcttctgca aatacgcgca gggggcggac 1500
agcgtggagc ccatgttccg gcacctgaag aacacgtatg cgggcctgca gctggtggtg 1560
gtcatcctgc ccggcaagac gcccggtgac gccgaggtca agcgcgtggg agacacggtg 1620
ctggggatgg ccacgcagtg cgtgcagatg aagaacgtgc agaggaccac gccacagacc 1680
ctgtccaacc tttgcctgaa gatcaacgtc aagctgggag gcgtgaacaa catcctgctg 1740
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caccccccg ccgggatgg gaagaagccc tccattgccg ccgtggtggg cagcatggac 1860
gccacccca atcgctactg cgccaccgtg cgcgtgcagc agcaccggca ggagatcata 1920
caagacctgg ccgcatggt ccgcgagctc ctcatccagt tctacaagtc cacgcgcttc 1980
aagcccacc gcacatctt ctaccgcgac ggtgtctctg aaggccagtt ccagcaggtt 2040
ctccaccacg agttgctggc catccgtgag gcctgtatca agctagaaaa agactaccag 2100
cccgggatca ccttcatcgt ggtgcagaag aggaccaca cccggctctt ctgactgac 2160
aagaacgagc ggggtgggaa aagtggaaac attccagcag gcacgactgt ggacacgaaa 2220
atcaccacc ccaccgagtt cgacttctac ctgtgtagtc acgctggcat ccaggggaca 2280
agcaggcctt cgcactatca cgtcctctgg gacgacaatc gtttctctc tgatgagctg 2340
cagatcctaa cctaccagct gtgtcacacc tacgtgcgct gcacacgctc cgtgtccatc 2400
ccagcggcag catactacgc tcacctggtg gccttccggg ccaggtacca cctggtggat 2460
aaggaacatg acagtgctga aggaagccat acctctgggc agagtaacgg gcgagaccac 2520
caagcactgg ccaaggcggc ccaggttcac caagacactc tgcgcacat gtactttgct 2580

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<210> SEQ ID NO 76

<211> LENGTH: 2772

<212> TYPE: DNA

<213> ORGANISM: Homo sapiens

<220> FEATURE:

<221> NAME/KEY: misc_feature

<223> OTHER INFORMATION: eIF2C3, cDNA sequence of predicted ORF

<400> SEQUENCE: 76

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agccggagcc gggtcctgt ccccgggccg ggcgcgcgcg ccgccccctg cccagcggcc 60
gcgtctccgc ggcgccacc cagcgccaat attccggaga tcaagegtta cgcggcggcg 120
gcggcggcgg cggcggggcc cggagcggga ggcgcgggg accggggcga ggccggcccc 180

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gccgcccga	tggaggcgt	gggacccgga	cctccggcta	gcctgtttca	gccacctcgt	240
cgctctggcc	ttggaactgt	tggaaaacca	attcgactgt	tagccaatca	ttttcagggt	300
cagattccta	aaatagatgt	gtatcactat	gatgtggata	ttaagcctga	aaaacggcct	360
cgtagagtca	acagggaggt	agtagataca	atggtgcggc	acttcaagat	gcaaatattt	420
ggtgatcggc	agcctgggta	tgatggcaaa	agaaacatgt	acacagcaca	tccactacca	480
attggacggg	atagggttga	tatggagggtg	actcttcag	gcgagggtaa	agaccaaaaca	540
tttaaagtgt	ctgttcagtg	ggtgtcagtt	gtgagccttc	agttgctttt	agaagctttg	600
gctgggcact	tgaatgaagt	cccagatgac	tcagtacaag	cacttgatgt	tatcacaaga	660
caccttccct	ccatgaggta	caccccagtg	ggccgttcc	ttttctcacc	cccggaaggt	720
tactaccacc	ctctgggagg	gggcagggag	gtctggtttg	gttttcatca	gtctgtgaga	780
cctgccatgt	ggaatatgat	gctcaacatt	gatgtatctg	caactgcttt	ctaccgggct	840
cagctatca	ttgagttcat	gtgtgagggt	ttagacattc	agaacatcaa	tgaacagacc	900
aaacctctaa	cagactccca	gcgtgtcaaa	ttaccaaaag	aatcagagg	tctcaaagtt	960
gaggtgacct	actgtggaca	gatgaaacga	aaataccgag	tttgtaatgt	gactagacgg	1020
ccagccagtc	atcaaacttt	tcctttgcag	ctagaaaacg	gtcaagctat	ggaatgtaca	1080
gtagctcaat	attttaagca	aaagtatagt	ctgcaactga	aataccccca	tcttcctgt	1140
ctccaagtgg	gacaagaaca	aaagcataca	tacttgccac	tcgaggctctg	taatatagtg	1200
gcaggacagc	gatgtatcaa	gaagctcaca	gacaatcaga	cttccacaat	gatcaaagct	1260
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atggtgggtg	gacctgatcc	ataccttaaa	gaatttggtg	ttggtgtcca	caatgaaatg	1380
acagagctca	caggcagggt	acttccagca	ccaatgctgc	aatatggagg	ccggaataaa	1440
acagtagcca	cacccaacca	gggtgtctgg	gacatgcgag	gaaagcagtt	ttatgctggc	1500
attgaaatta	aagtttgggc	agttgcttgt	tttgcacctc	agaaacaatg	tagggaagat	1560
ttactaaaga	gtttcactga	ccagctgcgt	aaaatctcta	aggatgcagg	aatgcccatc	1620
cagggtcagc	catgtttctg	caagtatgca	caaggtgcag	acagtgtgga	gcctatgttt	1680
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tgtgtccagg	taaaaaatgt	agtgaagacc	tcacctcaaa	ccctttcaa	tctttgcctg	1860
aagataaatg	caaaacttgg	aggaattaac	aatgtgcttg	tgctcatca	aaggccctcg	1920
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gggaagaaac	cttccattgc	tgctgtgggt	ggcagtatgg	atggccacc	cagccggtac	2040
tgtgccaccg	ttcgggtgca	gacttcccgg	caggagatct	cccaagagct	cctctacagt	2100
caagaggcca	tccaggacct	gactaacatg	gttcgagagc	tgctgattca	gttctacaaa	2160
tccacacgct	tcaaaccac	tcggatcatc	tattaccgtg	gaggggtatc	tgaggggaaa	2220
atgaaacagg	tagcttggcc	agaactaata	gcaattcgaa	aggcatgtat	tagcttggaa	2280
gaagattacc	ggccaggaat	aacttatatt	gtggtgcaaa	aaagacatca	cacacgactc	2340
ttctgtgcag	ataaaacaga	aagggtaggg	aaaagtggca	atgtaccagc	aggcactaca	2400
gtggatagta	ccatcacaca	tccatctgag	tttgactttt	acctctgtag	tcatgcagga	2460
attcagggaa	ccagccgtcc	ctcacattac	caggtcttgt	gggatgacaa	ctgcttact	2520
gcagatgaac	tccagctact	gacttaccag	ctgtgtcaca	cctatgtgag	gtgcactcgc	2580

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tcagtctcta ttccagcccc tgcatattat gcccggcttg tagcatttag ggcaaggtat 2640
catctggtagg ataaagatca tgacagtgcg gaaggcagtc atgtgtcagg acagagcaac 2700
ggccgggatc ctcaggcctt ggctaaggct gtgcaaatcc accatgatac ccagcacacg 2760
atgtattttg cc 2772

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<210> SEQ ID NO 77
<211> LENGTH: 2568
<212> TYPE: DNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: eIF2C4, cDNA sequence of predicted ORF

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<400> SEQUENCE: 77

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gcaggaccgc ctggggccca gcccctactc atggtgcccc gaagacctgg ctatggcacc 60
atgggcaaac ccattaaact gctggctaac tgttttcaag ttgaaatccc aaagattgat 120
gtctacctct atgaggtaga tattaacca gacaagtgtc ctaggagagt gaacagggag 180
gtggttgact caatggttca gcattttaaa gtaactatat ttggagaccg tagaccagtt 240
tatgatggaa aaagaagtct ttacaccgcc aatccacttc ctgtggcaac tacaggggta 300
gatttagacg ttactttacc tggggaaggt ggaaaagatc gacctttcaa ggtgtcaatc 360
aaatttgtct ctcgggtgag ttggcaccta ctgcatgaag tactgacagg acggaccttg 420
cctgagccac tgggaattaga caagccaatc agcactaacc ctgtccatgc cgttgatgtg 480
gtgctacgac atctgcctc catgaaatac acacctgtgg ggcgttcatt tttctccgct 540
ccagaaggat atgaccacc cctgggaggg ggcagggaag tgtggtttg attccatcag 600
tctgttcggc ctgccatgtg gaaaatgatg cttaatatcg atgtttctgc cactgccttc 660
tacaaagcac aacctgtaat tcagttcatg tgtgaagttc ttgatattca taatattgat 720
gagcaaccaa gacctctgac tgatttctcat cgggtaaaat tcaccaaaga gataaaaggt 780
ttgaaggttg aagtgactca ttgtggaaca atgagacgga aataccgtgt ttgtaatgta 840
acaaggaggc ctgccagtca tcaaaccctt cctttacagt tagaaaacgg ccaaactgtg 900
gagagaacag tagcgcagta tttcagagaa aagtatactc ttcagctgaa gtaccgcgac 960
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aatattgtgg cagggcaacg atgtatcaag aagctaacag acaatcagac ttccactatg 1080
atcaaggcaa cagcaagatc tgcaccagat agacaagagg aaattagcag attggtaaga 1140
agtcaaatt atgaaacaga tccatttggt caggagtttc aatttaaagt tcgggatgaa 1200
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cggacagtag caacaccgag ccatggagta tgggacatgc gagggaaaca attccacaca 1320
ggagttgaaa tcaaaatgtg ggctatcgct tgttttgcca cacagaggca gtgcagagaa 1380
gaaatattga agggtttcac agaccagctg cgtaagattt ctaaggatgc agggatgccc 1440
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ttccggcatc tcaagaacac atattctggc ctacagctta ttatcgtcat cctgccgggg 1560
aagacaccag tgtatgcgga agtgaaacgt gtaggagaca cacttttggg tatggctaca 1620
caatgtgttc aagtcaagaa tgtaataaaa acatctctc aaactctgtc aaacttgtgc 1680
ctaaagataa atgttaaac cggagggatc aataatattc ttgtacctca tcaaagacct 1740
tctgtgttcc agcaaccagt gatctttttg ggagccgatg tcaactatcc acctgctggt 1800

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gatggaaga agccttctat tgctgctggt gtaggtagta tggatgcaca cccaagcaga 1860
tactgtgcca cagtaagagt tcagagaccc cgacaggaga tcatccagga cttggcctcc 1920
atggtccggg aacttcttat tcaatthttat aagtcaactc ggttcaagcc tactcgtatc 1980
atctthttatc gggatggtgt ttcagagggg cagthttagge aggtattata ttatgaacta 2040
ctagcaattc gagaagcctg catcagthttg gagaaagact atcaacctgg aataacctac 2100
attgtagthtc agaagagaca tcacactcga thattthttgtg ctgataggac agaaagggtht 2160
ggaagaagtg gcaatatccc agctggaaca acagthtgata cagacattac acacccatat 2220
gagthtcgatt thtacctctg tagccatgct ggaatacagg gtaccagthcg thctthcacac 2280
tatcatgthtt tatgggatga thaactgcttht actgcagatg aactthcagct gcthaactthac 2340
cagctctgcc acactthacgt acgctgtaca cgatctgthtt ctataacctgc accagcgtat 2400
tatgctcacc tggtagcatt tagagccaga tatcatcttg tggacaaaga acatgacagth 2460
gctgaaggaa gthcacgthttc aggacaaagc aatgggctgag atccacaagc thcttgccaag 2520
gctgtacaga thccaaga thacctacgc acaatgthact thgctthaa 2568

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<210> SEQ ID NO 78

<211> LENGTH: 2292

<212> TYPE: DNA

<213> ORGANISM: Homo sapiens

<220> FEATURE:

<221> NAME/KEY: misc_feature

<223> OTHER INFORMATION: HILI, cDNA sequence of predicted ORF

<400> SEQUENCE: 78

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gththccagtg gaatactgt gaaactggtht acaaacctct thhaacttaga thththcccaa 180
gactggcagc thataccagth ccatgtgaca thaththccag aththagcathc thagaaggctg 240
agaathtgctt thctththtag thcatagthga cththccaaca aagcaaaagc aththcgacggt 300
gccathcttht thctgtcaca aaagctagaa gaaaaggthc cagagththgc aagthgaaact 360
caagaggthg agactataaa gatgactathc accctgaaga gggagctgcc athcaagthct 420
cccgtgtgca thccagthctt caathathcathc thcagaaaga thctcaaaaa gththgtccatg 480
thaccaathg gacggaactt ctataathct thcagagccaa thggaaththc ccagcacaaa 540
thathctctth ggctgggtht thgctaththct gththcatath thgaaaggaa gthctctgtht 600
agthctgatg thagththcaa agthctccgg aathgagacgg thctggaathc cathgactgct 660
ctctgtcaaa gaactggctt gthctgththc acccagacgt gthgagaagca gthhaathggg 720
ctcaththgthc thacaagata caathaacaga acctactcca thgatgacath thgactggthc 780
gthgaagccca cacacactt thcagaagcgg gatggcaccg agathcacctha thgtggaththc 840
thacaagcagc agthathgath thactgthctg gactggaathc agccathgct thgttagthctg 900
thaaagaaga agagaaathg caacagthgag gthcagctcg cccactgathc acctgagctc 960
thgctthctha cagggctgac thgaccaggca athathctgath thcagctgath gaaggctgth 1020
gthgaaaaga cacgthctcag thctthcaggc cggcagcagc gcctggccag gctthgtggac 1080
aathathcagha ggaathaccaa thgctcgcttht gaactagaha cctggggact gctthththgga 1140
agccagathc ctctgactgg ccgaththgtg cctthcagaaa aathaththaat gcaagaccac 1200
athaththcaac ctgthgthctg thgctgactgg thccaaggata thcgaactthg caagaththth 1260

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aatgcacagt ctttgaatac ctggttgatt ttatgtagcg acagaactga atatggtgcc 1320
gagagctttc tgaactgctt gagaagagtt gcaggttcca tgggatttaa tgtaatgtgc 1380
attctgcctt ctaatcagaa gacctattat gattccatta aaaaatattt gagctcagac 1440
tgcccagtc caagccaatg tgtgcttgc cggacctga ataacaggg catgatgatg 1500
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<210> SEQ ID NO 79

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<212> TYPE: DNA

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<220> FEATURE:

<221> NAME/KEY: misc_feature

<223> OTHER INFORMATION: HIWI, cDNA sequence of predicted ORF

<400> SEQUENCE: 79

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<220> FEATURE:
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<223> OTHER INFORMATION: eIF2C1, primer (5'-3')

<400> SEQUENCE: 80

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<210> SEQ ID NO 81
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<223> OTHER INFORMATION: Oligodeoxynucleotide with homology to human gene
 <220> FEATURE:
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 <223> OTHER INFORMATION: eIF2C1, primer (5'-3')

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<210> SEQ ID NO 82
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<220> FEATURE:
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 <223> OTHER INFORMATION: eIF2C1, primer (5'-3')

<400> SEQUENCE: 86
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<400> SEQUENCE: 87
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<210> SEQ ID NO 88
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<400> SEQUENCE: 88
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 <223> OTHER INFORMATION: eIF2C2, primer (5'-3')

<400> SEQUENCE: 90
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 <223> OTHER INFORMATION: eIF2C2, primer (5'-3')

 <400> SEQUENCE: 95

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 <210> SEQ ID NO 96
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 <223> OTHER INFORMATION: eIF2C3, primer (5'-3')

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<400> SEQUENCE: 102

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<223> OTHER INFORMATION: eIF2C4, primer (5'-3')

<400> SEQUENCE: 105

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 <223> OTHER INFORMATION: eIF2C4, primer (5'-3')

 <400> SEQUENCE: 106

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<210> SEQ ID NO 107
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 <223> OTHER INFORMATION: eIF2C4, primer (5'-3')

 <400> SEQUENCE: 107

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<210> SEQ ID NO 108
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 <400> SEQUENCE: 108

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 <223> OTHER INFORMATION: HILI, primer (5'-3')

 <400> SEQUENCE: 109

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<210> SEQ ID NO 110
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 <400> SEQUENCE: 110

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<223> OTHER INFORMATION: HILI, primer (5'-3')

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<223> OTHER INFORMATION: HILI, primer (5'-3')

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<210> SEQ ID NO 114
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<221> NAME/KEY: misc_feature
<223> OTHER INFORMATION: HILI, primer (5'-3')

<400> SEQUENCE: 114

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<223> OTHER INFORMATION: Oligodeoxynucleotide with homology to human gene

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<220> FEATURE:
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 <223> OTHER INFORMATION: HILI, primer (5'-3')

<400> SEQUENCE: 115

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The invention claimed is:

1. Purified human RISC, wherein said RISC is a ribonucleoprotein complex containing a single stranded RNA component, and wherein said RNA component comprises at least one modified nucleotide analogue which is selected from sugar backbone and nucleobase-modified ribonucleotides and combinations thereof, wherein said RNA component is a single-stranded RNA molecule wherein the single-stranded RNA molecule has a length from 19-29 nucleotides, and wherein said RISC has a molecular weight of less than about 160 kDa.

2. The RISC of claim 1 comprising at least one member of the Argonaute family of proteins.

3. The RISC of claim 2 wherein said at least one member of the Argonaute family of proteins is eIF2C1 and/or eIFC2.

4. The RISC of claim 3, further comprising at least one of eIFC3, eIFC4, HILI or HIWI.

5. The RISC of claim 1, wherein at least the 14-20 5' most nucleotides are substantially complementary to a target transcript.

6. The RISC of claim 1, wherein said RNA molecule has a free 5' hydroxyl moiety or a moiety selected from phosphate groups or analogues thereof.

7. The RISC of claim 6, wherein said RNA molecule has a 5'-moiety selected from the group consisting of 5'-monophosphate ((HO)₂(O)P—O-5'), 5'-diphosphate ((HO)₂(O)P—O—P(HO)(O)—O-5'), 5'-triphosphate ((HO)₂(O)P—O—(HO)(O)P—O—P(HO)(O)—O-5'), 5'-guanosine cap (7-methylated or non-methylated) (7m-G-O-5'-(HO)(O)P—O—(HO)(O)P—O—P(HO)(O)—O-5'), 5'-adenosine cap (Appp), and any modified or unmodified nucleotide cap structure (N—O-5'-(HO)(O)P—O—(HO)(O)P—O—P(HO)(O)—O-5'), 5'-monothiophosphate (phosphorothioate; (HO)₂(S)P—O-5'), 5'-monodithiophosphate (phosphorodithioate; (HO)(HS)(S)P—O-5'), 5'-phosphorothiolate ((HO)₂

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(O)P—S-5'); any additional combination of oxygen/sulfur replaced monophosphate, diphosphate and triphosphates, 5'-phosphoramidates ((HO)₂(O)P—NH-5', (HO)(NH₂)(O)P—O-5'), 5'-alkylphosphonates, and 5'-alkyletherphosphonates.

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8. The RISC of claim 1, wherein said RNA molecule is completely complementary to said target transcript or completely complementary to said target transcript with the exception of nucleotides that extend beyond position 20 counted from the 5' terminus.

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9. The RISC of claim 1, wherein said covalent coupling is via the 3'-terminus of the RNA molecule.

10. A host cell or non-human host organism capable of overexpressing the RISC according to claim 1.

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11. A method of enhancing RNAi in a cell or an organism comprising causing said cell or organism to overexpress at least one component of RISC according to claim 1.

12. A composition comprising the RISC according to claim 1 in combination with a pharmaceutical carrier.

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13. The RISC according to claim 7, wherein said sulfur replaced monophosphate, diphosphate or triphosphate is 5'-alpha-thiotriphosphate or 5'-gamma-thiotriphosphate, said 5'-alkylphosphonate is RP(OH)(O)—O-5'- or (OH)₂(O)P-5'-CH₂—, and said 5'-alkyletherphosphonate is RP(OH)(O)—O-5'-.

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14. The RISC according to claim 1, wherein the 3' end of said RNA component is stabilized against degradation.

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15. The RISC according to claim 14, wherein the 3' end of said RNA component is stabilized against degradation by incorporating non-nucleotide chemical derivatives selected from the group consisting of aminolinkers, thiol linkers, carboxyl linkers, and non-nucleotidic spacers.

16. The RISC according to claim 1, wherein the 3' end of said RNA component is modified with biotin or fluoresceine.

* * * * *